

Isolation and Identification of the Main Compounds of *Satureja sahendica* Bornm.

¹S. Saeidnia, ²M.S. Nourbakhsh, ¹A.R. Gohari, ²A. Davood

¹Medicinal Plants Research Center, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, PO Box 14155-6451, Iran.

²Department of Medicinal Chemistry, Faculty of Pharmacy, Islamic Azad University, Tehran, Iran

Abstract: *Satureja sahendica* Bornm is generally called Marzeh in the Persian language and belongs to Lamiaceae family which comprises 13 species in Iran. In this study, the plant material (aerial parts of *S. sahendica*) was collected from North-East of Iran (Azerbaijan province). Luteolin (**1**), together with oleanolic acid (**2**), beta-sitosterol (**3**) and diosmetin (**4**) were isolated from the ethyl acetate and methanol extracts of *S. sahendica* for the first time. Different chromatographic methods were carried out on the silica gel and sephadex LH₂₀ in order to separate of compounds. The structures of the isolated compounds were determined using the ¹H, ¹³C-NMR and MS spectra in comparison of those reported in the literatures. Diosmetin is an important flavone which converts to luteolin in the human body and affects on the breast cancer via binding to the estrogen receptors.

Key words: Lamiaceae, *Satureja sahendica*, Flavonoids

INTRODUCTION

The genus *Satureja* (Lamiaceae) comprises around 13 species in Iran. Among them *Satureja sahendica* grows widely in north-west of Iran (Hedge 1986; Mozaffarian, 1996). *Satureja* genus consists of the fragrance shrubs which are growing on the rocky mounts and used traditionally for anti-diarrhea, antispasmodic and pesticide activities (Gohari *et al.*, 2009). Literature reviews show that there are a few reports only on chemical compositions of the volatile oil of *S. sahendica*. Recently, composition of the essential oil of *S. sahendica* has been investigated by GLC and GC-MS. Thirty-nine components were identified in the oils. The main constituents of the essential oils were thymol (19.6–41.7%), *p*-cymene (32.5–54.9%) and γ -terpinene (1.0–12.8%) (Hassanpouraghdam *et al.*, 2009, Sefidkon *et al.*, 2004).

We have previously reported the presence of flavones (5,6,3'-trihydroxy-7,8,4'-trimethoxyflavone, 5-desmethoxynobiletin, thymonin and luteolin) from *S. atropatana* using chromatographic methods followed by ¹H and ¹³C-NMR and MS spectra (Gohari *et al.*, 2009).

In this paper, we aimed to report the separation and structural elucidation of the main phytochemical constituents from the aerial parts of *S. sahendica* which has not been reported in advance.

MATERIAL AND METHODS

Experimental:

Aerial parts of *S. sahendica* Bornm, at the full flowering stage, were gathered around Tabriz in East Azerbaijan Province (September, 2008). A voucher specimen of the plant deposited at the Herbarium of the Institute of Medicinal Plants, ACECR, Tehran. Plant specimen was identified by Mr. Yousef Ajani from the mentioned institute.

General Procedure:

The ¹H and ¹³C-NMR spectra were measured on a Bruker Avance TM 500 DRX (500 MHz for ¹H and 125 MHz for ¹³C) spectrometer with tetramethylsilane as an internal standard and chemical shifts are given in δ (ppm). The MS data were recorded on an Agilent Technology (HP TM) instrument with 5973 Network Mass Selective Detector (MS model). The silica gel 60F₂₅₄ pre-coated plates (Merck TM) were used for TLC. The spots were detected by spraying anisaldehyde-H₂SO₄ reagent followed by heating (120 °C for 5 min).

Corresponding Author: A.R. Gohari, Medicinal Plants Research Center, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, PO Box 14155-6451, Iran.
Tel & Fax: +98-21-64122330.
E-mail: goharii_a@sina.tums.ac.ir

Isolation Process:

The flowered aerial parts of *S. sahendica* (2150 g) was cut into small pieces and extracted with ethyl acetate and methanol, consequently, at room temperature to obtain ethyl acetate (25 g) and methanol (62 g) extracts. The ethyl acetate extract was subjected to silica gel column chromatography (CC) with hexane: AcOEt (9:1, 3:2, 0:1), AcOEt: MeOH (1:1) and MeOH as eluent to give nine fractions (A-I). The fraction D (200 mg) was submitted to sephadex LH₂₀ CC with CHCl₃: MeOH (3:7) as an eluent to obtain four fractions D₁-D₄. The fraction C₄ (20 mg) was the pure compound **1**. The fraction E (1.460 g) was subjected to silica gel CC with hexane: AcOEt (8:2, 6:4, 1:1 and 0:1) to gain eight fractions (E₁-E₈). The fractions E₅ chromatographed again on sephadex LH₂₀ to result in compound **2** (77 mg).

The MeOH extract (30 g) was successively subjected to silica gel column chromatography and washed with CHCl₃: AcOEt (1:0, 1:1, 0:1) and MeOH as eluents to result in eight fractions M₁- M₈. Fraction M₂ (292 mg) was fractionated on silica gel CC with CHCl₃: AcOEt (19:1, 6:4, 0:1) to obtain three fractions M₂₁ -M₂₃. The fraction M₂₂ (6 mg) was the pure compound **3**. The fraction M₄ (216 mg) was chromatographed on sephadex LH₂₀ with MeOH to afford Compound **4** (3 mg).

RESULTS AND DISCUSSION

Isolated compounds (Fig.1) from the ethyl acetate and MeOH extracts of *S. sahendica* identified as luteolin (**1**), together with oleanolic acid (**2**), beta-sitosterol (**3**) and diosmetin (**4**) by comparison of their NMR and MS spectral data with those reported in literature (7-11). NMR data of luteolin (**1**) and β-sitosterol (**3**) were described by Saeidnia *et al.* (2009) and Gohari *et al* (2009; 2005).

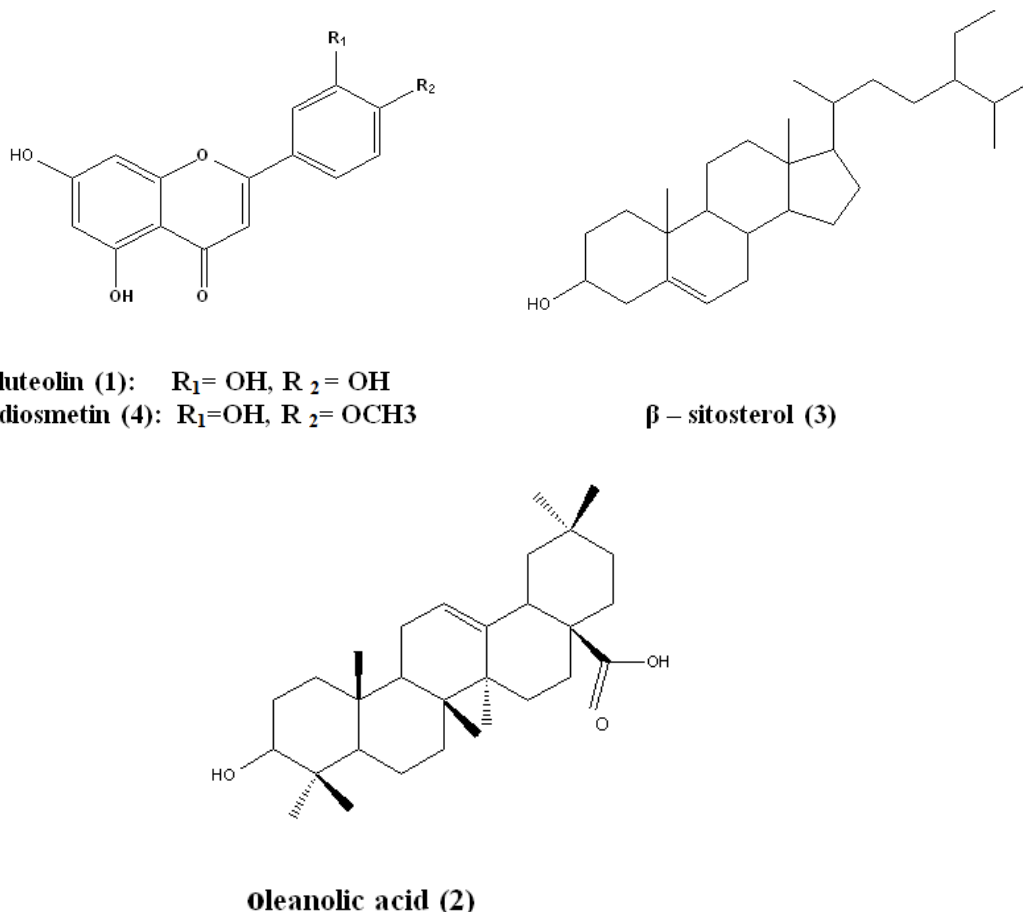


Fig. 1: Structures of the isolated compounds from *Saturejasahendica*.

Oleanolic Acid (2):

White amorphous powder. m.p.: 271-273° C. ¹H-NMR (500 MHz, CDCl₃): 0.75, 0.77, 0.90, 0.91, 0.93, 0.98 (each 3H, s, CH₃ ×6), 1.13 (3H, s, H-27), 2.82 (1H, dd, J= 3.6, 13.2 Hz, H-18), 3.23 (1H, dd, J= 11.2, 4.4 Hz, H-3), 5.27 (1H, t, J=3.5 Hz, H-12). ¹³C-NMR (125 MHz, Pyridine-*d*₅): δC (from C-1 to C-30) 39.0, 28.2, 78.1, 39.4, 55.8, 18.8, 33.3, 39.8, 48.2, 37.4, 23.7, 122.6, 144.8, 42.2, 28.4, 23.8, 46.7, 42.0, 46.5, 31.0, 34.3, 33.2, 28.8, 16.6, 15.6, 17.5, 26.2, 180.2, 33.3, 23.8.

Diosmetin (4):

Yellow needle crystal. ¹H NMR (500 MHz, acetone-*d*₆): δ (ppm), 12.99 (1H, s, 5-OH), 7.57 (1H, dd, J = 2.2, 8.5 Hz, H-6'), 7.50 (1H, d, J = 2.3 Hz, H-2'), 7.13 (1H, d, J = 8.5 Hz, H-5'), 6.64 (1H, s, 3-H), 6.56(1H, d, J = 2.1 Hz, H-8), 6.20 (1H, d, J = 2.1 Hz, H-6) and 3.95 (3H, s, 4 -OCH₃). ¹³C NMR (125 MHz, acetone-*d*₆): δ (ppm) 164.8 (C-2), 104.6 (C-3), 182.9 (C-4), 163.3 (C-5), 99.6 (C-6), 164.9 (C-7), 94.7 (C-8), 158.7 (C-9), 105.3 (C-10), 124.8 (C-1'), 113.6 (C-2'), 147.8 (C-3'), 151.6 (C-4'), 112.4 (C-5'), 119.6 (C-6'), 56.3 (OMe).

Diosmetin, the methyl ether aglycon of diosmin, is the most important flavone in *S. sahendica* which reported only from *S. obovata* among the *Stureja* species (Atta-ur- Rahman, 2005). Anticancer effects of diosmetin on cell cycle progression and proliferation of MDA-MB 468 breast cancer cells, has been reported (Androutsopoulos *et al.*, 2009). The studies of flavonoids on the breast cancer showed that the flavonoid diosmetin is metabolised to the more active molecule luteolin by CYP1 family enzymes (Androutsopoulos *et al.*, 2009). The cytoprotective effect of diosmetin, was investigated on iron-loaded hepatocyte cultures. Thus, *S. sahendica* which consists of both luteolin and diosmetin as the main anticancer agents, seems to be a good cytotoxic medicinal plant. The cytoprotective activity of diosmetin, which previously reported, could be ascribed to its antiradical property but also to iron-chelating effectiveness (Morel *et al.*, 1993).

Oleanolic acid is another bioactive component of *S. sahendica*. As we can see in the literatures, oleanolic acid, a petacyclic triterpene, showed antibacterial, antifungal, trypanocidal and anti-inflammatory effects, antitumor and immunomodulatory activity (Gohari *et al.*, 2009; Saeidnia *et al.*, 2009; Gohari, *et al.*, 2005). Beta-sitosterol is a common sterol in the plants which decreases the symptom of benign prostatic hyperplasia and anti-inflammatory activities (Wilt *et al.*, 1999).

Conclusion:

In conclusion, *S. sahendica*, the medicinal plant of Lamiaceae family, contains luteolin and diosmetin as the bioactive flavones and, oleanolic acid and beta-sitosterol as the bioactive triterpene and sterol. These components have not been reported from *S. sahendica* so far.

ACKNOWLEDGEMENT

This research has been supported by Tehran University of Medical Sciences and Health Services grant (No.10188).

REFERENCES

- Hedge, I.C., 1986. Labiatae. In: Flora Iranica, Ed., Rechinger, K.H., Akademische Druck Verlagsantalt, pp: 495-504.
- Mozaffarian, V., 1996. A Dictionary of Iranian Plant Names. Farhang Moaser, Tehran, pp: 483-484.
- Gohari, A.R., S. Saeidnia, A. Hadjiakhoondi, M. Abdoullahi and M. Nezafati, 2009. Isolation and quantitative analysis of oleanolic acid from *Satureja mutica* Fisch & C.A.Mey. Journal of Medicinal Plants, 8(5): 65-69.
- Hassanpouraghdam, M.B., M. Safi Shalamzari, M.A., Aazami and A. Mohajjer Shoja, 2009. γ-Terpinene and carvacrol rich volatile oil of *Satureja sahendica* Bornm. from Maragheh district in Northwest Iran. Chemija, 20(3): 186-189.
- Sefidkon, F., Z. Jamzad and M. Mirza, 2004. Chemical variation in the essential oil of *Satureja sahendica* from Iran. Food Chemistry, 88(3): 325-328.
- Gohari, A.R., S. Saeidnia, M.R. Gohari, F. Moradi-Afrapoli, M. Malmir and A. Hadjiakhoondi, 2009. Bioactive flavonoids from *Satureja atropatana* Bonge. Natural Product Research, 23(17): 1609- 1614.
- Saeidnia, S., N. Yassa, R. Rezaei-poor, A. Shafiee, A.R. Gohari, M. Kamalinejad and S. Goodarzi, 2009. Immunosuppressive principles from *Achillea talagonica*, an endemic species of Iran. DARU, 17(1): 37-41.

Gohari, A.R., A. Hadjiakhoondi, S.E. Sadat-Ebrahimi, S. Saeidnia and A. Shafiee, 2005. Cytotoxic triterpenoids from *Satureja macrantha* C.A. Mey. Daru, 13: 177-181.

Gohari, A.R., S. Saeidnia, A.R. Shahverdi, N. Yassa, Malmir, M., K. Mollazade and A.R. Naghinejad, 2009c. Phytochemistry and antimicrobial compounds of *Hymenocrater calycinus*. EurAsian Journal of BioSciences, 3(9): 64-68.

Vasilis, A., W. Nicola, R.J. Randolph and A.P. Gerry, 2009. Bioactivation of the phytoestrogen diosmetin by CYP1 cytochromes P450. Cancer Letter, 274: 54-60.

Agrawal, P.K., 1989. Carbon-13 NMR of Flavonoids. Elsevier Science.

Atta-ur- Rahman, 2005. Studies in Natural Products Chemistry. Elsevier Science, pp: 292.

Androutsopoulos, V.P., S. Mahale, R.R. Arroo and G. Potter, 2009. Anticancer effects of the flavonoid diosmetin on cell cycle progression and proliferation of MDA-MB 468 breast cancer cells due to CYP1 activation. Oncology Report, 21(6): 1525-1528.

Androutsopoulos, V., N. Wilsher, R.R. Arroo and G.A. Potter, 2009. Bioactivation of the phytoestrogen diosmetin by CYP1 cytochromes P450. Cancer Letter, 274(1): 54-60.

Morel, I., G. Lescoat, P. Cogrel, O. Sergent, N. Padeloup, P. Brissot, P. Cillard and J. Cillard, 1993. Antioxidant and iron-chelating activities of the flavonoids catechin, quercetin and diosmetin on iron-loaded rat hepatocyte cultures. Biochemical Pharmacology, 45(1): 13-19.

Wilt, T.J., R. Macdonald and A. Ishani, 1999. Beta sitosterol for the treatment of benign prostatic hyperplasia. British Journal of Urology International, 83: 976-983.