

FISSIDENS SUBGENUS *ALOMA* (BRYOPHYTA) IN TROPICAL AFRICA I. THE LARGE-CELLED COSTATE AND ECOSTATE SPECIES

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Abstract. A revision of the large-celled costate and ecostate species of *Fissidens* subgenus *Aloma* Kindb. from tropical Africa is presented. *Fissidens flaccidus* var. *mammillosus* Brugg.-Nann. var. *nov.* is described and the Neotropical *F. mollis* Mitt. is reinstated. Most African specimens previously identified as *F. enervis* Sim are referred to *Fissidens dealbatus* Hook. f. & Wilson, a new indigene for Africa. *Fissidens molliculus* P. de la Varde, *F. hedbergii* P. de la Varde and *F. hookerioides* Bizot & Onr. in Bizot are subsumed under *F. dealbatus*. *Fissidens enervis* s.str. appeared to be a rare species. The distributions in O’Shea (2006) are revised, updated and, when necessary, amended. *Fissidens flaccidus* Mitt. is for the first time reported from South Africa and Guinea, *F. zollingeri* Mont. from Ghana, *F. grandifolius* Broth. & P. de la Varde from Cameroon and *F. usambaricus* Broth. from Malawi and Gabon. Pending further research, *F. palmatus* Hedw. is excluded from the African bryoflora. It is stressed that, since vegetative and perichaetial stems are different, perichaetial stems (and leaves of perichaetial stems) are to be compared with perichaetial stems (and leaves of perichaetial stems), whereas vegetative stems (and leaves of vegetative stems) should be compared with other vegetative stems (and leaves of vegetative stems).

Key words: distribution, *Fissidens*, large-celled *Aloma*, taxonomic revision, tropical Africa

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INTRODUCTION

Fissidens subgenus *Aloma* Kindb. embraces a wide range of gametophytic types (Pursell & Bruggeman-Nannenga 2004). Five not sharply distinct groups can be recognized: one without costa, one with large (over 30 μm long) cells, one with pluripapillose cells, one with unipapillose cells and one with smooth cells. Because of the existence of transitional species, these groups are no longer recognized as taxa (Pursell & Bruggeman-Nannenga 2004). *Fissidens* subgenus *Aloma* includes elimbate, semilimbate and a few completely limbate species, and the laminal cells are small, medium-sized or large. Ecostate species as well as species with large (30 μm long or more) laminal cells are restricted to this subgenus. A notable exception is the African *F.* subgenus *Fissidens* Hedw. species *F. magnicellulatus* Brugg.-Nann., which also has cells 30 μm long or more. This species is therefore included in the key below. All ecostate species have large cells but not all large-celled species are ecostate (Pursell & Bruggeman-Nannenga

2004). The present paper is a revision of those tropical African species of subgenus *Aloma* that have large (longer than 30 μm), smooth laminal cells. It includes both costate and ecostate species. In one species, *F. zollingeri* Mont., large cells are restricted to the vaginant lamina. With the exception of *F. usambaricus* Broth., all included species are limbate. Apart from their large cells, limbate, costate species bear a strong resemblance to subgenus *Fissidens*, which is characterized by limbate leaves and smooth laminal cells but has small to medium-sized cells and a different sporophyte.

This paper follows the classification by Pursell and Bruggeman-Nannenga (2004), who emphasized sporophytic characters. They considered large-celled limbate species to belong in subgenus *Aloma*, as they have *Aloma* sporophytes,¹ and classified limbate species with small, smooth cells

¹ Capsule with \pm 32 columns of exothecial cells and a scarious-type peristome.

in subgenus *Fissidens* because of their subgenus *Fissidens* sporophytes.² Suzuki and Iwatsuki (2007, 2012), on the other hand, emphasized gametophytic characters. They classified ecostate species in subgenus *Aneurion* Kindb., and limbate, costate species with smooth cells, regardless of cell size and sporophytic characters, in subgenus *Fissidens*. It will be interesting to learn what fresh characters such as DNA will contribute to this question.

During this revision some challenging differences were found between the data in the monograph on Neotropical Fissidentaceae (Pursell 2007) and what is seen in Africa. Some of these have resulted in taxonomic decisions. A problem concerning *F. palmatus* Hedw. and *F. grandifolius* Broth. & P. de la Varde is discussed, but remains unsolved; hopefully this discussion will stimulate further research.

PRACTICAL REMARKS

It is self-evident that morphological taxonomic research needs to be based on comparisons of equivalent characters. In *Fissidens* the vegetative and perichaetial stems are different and have different leaves. Therefore, perichaetial stems (and leaves of perichaetial stems) are to be compared with perichaetial stems (and leaves of perichaetial stems), whereas vegetative stems (and leaves of vegetative stems) should be compared with other vegetative stems (and leaves of vegetative stems). In medium-sized and long stems all mid leaves have more or less the same shape and size. This makes such leaves most suitable for comparison. In short stems the leaf size increases rapidly up the stem, so comparison of these with leaves of longer stems will be problematic. One also needs to be cautious when comparing laminal cells. Nearness to the costa, the stem and the limbidium induces differentiation (cells become larger, more oblong and less ornamented). Moreover, whereas inner cells of the dorsal and apical lamina are similar, inner vaginant laminal cells tend to differ.

The ‘distributions’ in the present paper are supported by specimens examined for this paper or

else their source is indicated. Distributions of some species are extended and/or had to be amended. No differentiation is made between the terms ‘mammillose’ and ‘unipapillose’. Trans-sections of these large-celled species are best examined in 5% KOH, which makes them swell beautifully.

KEY

1. Leaves (±) ecostate 2
- 1* Leaves with distinct costa 5
 2. Leaves elimbate 4. *F. usambaricus*
 - 2* Leaves limbate 3
3. Limbidial cells 5–12 µm wide, limbidium ill-defined 3. *F. metzgeria*
- 3* Limbidial cells narrower, ± 1 µm wide, limbidium distinct 4
 4. Capsule with 32 columns of exothecial cells, limbidia 1–2-stratose 2. *F. enervis*
 - 4* Capsule with 40 or more columns of exothecial cells, limbidia 1–7-stratose 1. *F. dealbatus*
5. Mid dorsal laminal cells small, (6.5–)11.0–27.0 × 4.5–14.5(–19.5) µm 9. *F. zollingeri*
- 5* At least some mid dorsal laminal cells 30 µm or longer 6
 6. Costa ending well below apex 7
 - 6* Costa reaching apex 9
7. Laminal cells mammillose
 - 5b. *F. flaccidus* var. *mammillosus*
- 7* Laminal cells smooth 8
 8. Stems mostly short, palmate; capsules inclined; antheridia axillary and on tiny stems at base of perichaetial stems; limbidia weakly 1(–2)-stratose on mid dorsal lamina 6. *F. grandifolius*
 - 8* Stems longer, with pinnately arranged leaves; capsules rare, erect; antheridia rare, terminal; limbidia stronger, (1–)2-stratose on mid dorsal lamina
 - 5a. *F. flaccidus* var. *flaccidus*
9. Limbidia not reaching leaf apex, intramarginal on all laminae 8. *F. nigerianus*
- 9* Limbidia reaching leaf apex, marginal on dorsal and apical lamina 10
 10. Leaves distant, peristome *bryoides*-type, ± 40 columns of exothecial cells on theca
 - F. magnicellulatus*³
 - 10* Leaves crowded, peristome *scariosus*-type, ± 32 columns of exothecial cells on theca
 7. *F. minutifolius*

² Capsule with 40 or more columns of exothecial cells and a *bryoides*-type peristome.

³ Not included in this paper. Description and figures in Bruggeman-Nannenga (2013a, b).

I. ECOSTATE SPECIES

Ecostate species represent a remarkable gametophytic group within subgenus *Aloma*, with large laminal cells and with central strands lacking or weakly developed. These species appear ecostate, yet many have or can have vestigial costae (Salmon 1899; Pursell & Bruggeman-Nannenga 2004) which occasionally can reach as far as the length of the vaginant lamina (e.g., in Müller Z215, *F. usambaricus* and the lectotype *F. enervis* Sim). Such weak costae have the same arrangement of cells as the *bryoides*-type costa with two large adaxial and one large central cell, but differ from well-developed *bryoides*-type costae in having thinner cell walls and absent or poorly developed lateral bands. Thus the ecostate species are not sharply distinct from the costate species. Ecostate species resemble costate subgenus *Aloma* species with large inflated cells. Many of these have costae that are short for the genus (Pursell & Bruggeman-Nannenga 2004). Like other subgenus *Aloma* species, ecostate species typically have capsules with ± 32 columns of exothecial cells and scariosus-type peristomes (Pursell & Bruggeman-Nannenga 2004).

It is interesting that not only the laminal cells but all or most gametophytic cells (e.g., cortical cells of the stem and cells of the calyptra) are inflated, whereas sporophytic cells are not. Limbidial cells are inflated in some species (e.g., *F. metzgeria*), but not in, for example, *F. dealbatus*, *F. enervis* and *F. hyalinus* Wilson & Hook.

Ecostate species can form dense mats but they frequently grow scattered among other mosses.

1. *Fissidens dealbatus* Hook. f. & Wilson

Fig. 1

in Wilson in Hook. f., Fl. Nov.-Zel. 2: 63, pl. LXXXIV–2. 1854; *Schistophyllum dealbatum* (Hook. f. & Wilson) Lindb., Utkast Eur. Bladmoss.: 16. 1878 – TYPE (PROTOLOGUE): NEW ZEALAND, Northern Island, Bay of Islands, Hooker [J. D. H.] – HOLOTYPE as Wilson 318 (BM!).

Fissidens molliculus P. de la Varde, Rev. Bryol. Lichenol. 7: 227, f. 1. 1935 – PROTOLOGUE: CENTRAL AFRICAN REPUBLIC [Oubangui], Ravin de la savanne à l'Ouest de Bozoum, sur terre sabloneuse,

III 1933, *Tisserant* – TYPE labels: Oubangui, ravin en savanne, Bozoum ouest, III 1933. *Tisserant 649* (HOLOTYPE: PC-PV!; ISOTYPE BM, same label but *s.n.*!), **syn. nov.**

Fissidens hedbergii P. de la Varde, Ark. Bot., n.s. 3: 129, f. 1: a–c. 1955 [1957]; *F. enervis* subsp. *hedbergii* (P. de la Varde) Bizot in Bizot & Pócs, Acta Bot. Acad. Sci. Hung. 25: 239. 1979 – PROTOLOGUE: TANZANIA, Mt. Kilimanjaro, montane rainforest above Marangu, 2100 m, on the ground at the path towards Bismarck Hut, Hedberg 1144, p.p. – label: Tanzania, [Tanganyika Territory], Kilimandjaro mountain, rainforest above Marangu, on the ground at the path towards Bismarck Hut, 2000 m, Hedberg 1144A, p.p. (HOLOTYPE: PC-PV!; ISOTYPE: BM!), **syn. nov.**

Fissidens hookerioides Bizot & Onr. in Bizot, Rev. Bryol. Lichénol. 42: 843. 1976 – PROTOLOGUE: MADAGASCAR, Tananarive, domaine des Frères de Soavimbahoaka, sur latérite ombragée, 1400 m, 6 VII 1971, Onraedt 71/M/3137 (orthographic error) – label: as protologue except for the collection number which is 71/M/5137 (HOLOTYPE: PC-0096394!; ISOTYPE: BR!), **syn. nov.**

Stem in cross section without or with weakly developed central strand and 20–50 μm wide cortical cells, unbranched, with pinnately or, infrequently, elongated palmately arranged leaves, not heterocaulous, (1.5–)3.0–6.5 \times 1.0–3.0 mm, browning in KOH; *rhizoids* basal, colorless to pale brown, less often orange-brown, smooth; *axillary nodules* not differentiated; *leaves* typically distant, 6–10 leaf pairs, somewhat crispate when dry, elliptical with acute-acuminate or less often acute apex that ends in a narrow pointed cell, 0.9–2.3 \times 0.2–0.6 mm, 2.5–5.0 times as long as wide, limbate; *limbidium* on all laminae, confluent at leaf apex, confluent at apex of vaginant lamina or not, reaching insertion of dorsal lamina, almost to completely reaching insertion of vaginant lamina, in mid dorsal lamina (4.5–)6.0–19.0 μm wide, 1–7-stratose (see remark below), marginal; on vaginant laminae 6.5–11.0 μm wide, 1–5-stratose, marginal; *vaginant lamina* 1/3–3/5 of leaf length, at insertion up to half as wide as stem, closed to \pm closed, unistratose; *dorsal lamina* wide, tapering and straight to slightly rounded towards insertion, typically reaching insertion, not decurrent; *dorsal* and *apical lamina* unistratose, rarely with isolated,

unicellular bistratose dots; *costa* ± lacking, in surface view sometimes visible as differentiated band alongside vaginant lamina; *mid dorsal laminal cells* hexagonal, 22.5–77.0 × 16.0–33.5(–48.0) µm, smooth, plane to inconspicuously bulging; *mid vaginant laminal cells* narrower, 45.0–95.0 × 12.0–30.5 µm; no *gemmae* observed.

Fertile parts, *perigonia* and *perichaetia* terminal on separate plants in same collection. *Perigonia* terminal on 1.5–3.0 mm long perigonial plants; *antheridia* 210 µm long; *perichaetia* terminal, perichaetial leaves 1.5–2.6 mm long; *archegonia* 200–290 µm long; *calyptra* tightly clasping operculum, scabrose, 0.6–0.8 mm long. *Sporophyte*. *Seta* 4–6 mm long, smooth, 1 per perichaetium; *capsule* symmetrical, 0.90–1.00 × 0.40–0.55 mm, 40–64 columns of oblong exothecial cells; *peristome* scariosus-type (basal part and filaments observed), teeth 56–60 µm wide at base; *operculum* rostrate, 0.8–0.9 mm long; *spores* 9.5–13.5 µm diam., smooth.

Fissidens dealbatus is characterized by its thecae with 40–64 columns of exothecial cells (exceptionally high for subgenus *Aloma*), distinct, narrow, 1–7-stratose limbidia of narrow (lumina ± 1 µm wide) limbidial cells, ecostate leaves that are limbate on all laminae, and large laminal cells. The disparity between the large inflated laminal cells and the narrow limbidial cells in this species is striking. This is particularly clear in trans-sections (Fig. 1J, K–L). There is considerable variation in leaf shape and the length and width of the mid laminal cells. The type of *F. hookerioides* is a small expression with short, relatively wide leaves and ± isodiametric cells. *Fissidens dealbatus* is most likely to be confused with *F. hyalinus* (not known from Africa) and *F. enervis*; like *F. dealbatus*, these species have large cells, ecostate, limbate leaves, and marked contrast between the large inflated laminal cells and narrow limbidial cells.

Fissidens dealbatus differs from both in the number of columns of exothecial cells (40 or more versus ± 32) and thicker limbidia (unistratose in *F. hyalinus* – *fide* Iwatsuki & Suzuki 1995 and 1–2-stratose in *F. enervis*). Furthermore, *F. dealbatus* and *F. hyalinus* have acute to acute acuminate leaf apices, whereas *F. enervis* has cuspidate ones. Many collections of this species are too small to make trans-sections of them, and capsules are often lacking. In such cases one has to resort to estimating the thickness of the limbidium, which is unreliable. The third ecostate, limbate African species, *F. metzgeria*, is easily distinguished by its ill-defined limbidium of wide cells and the frequent occurrence of characteristic, multicellular, epilaminal gemmae.

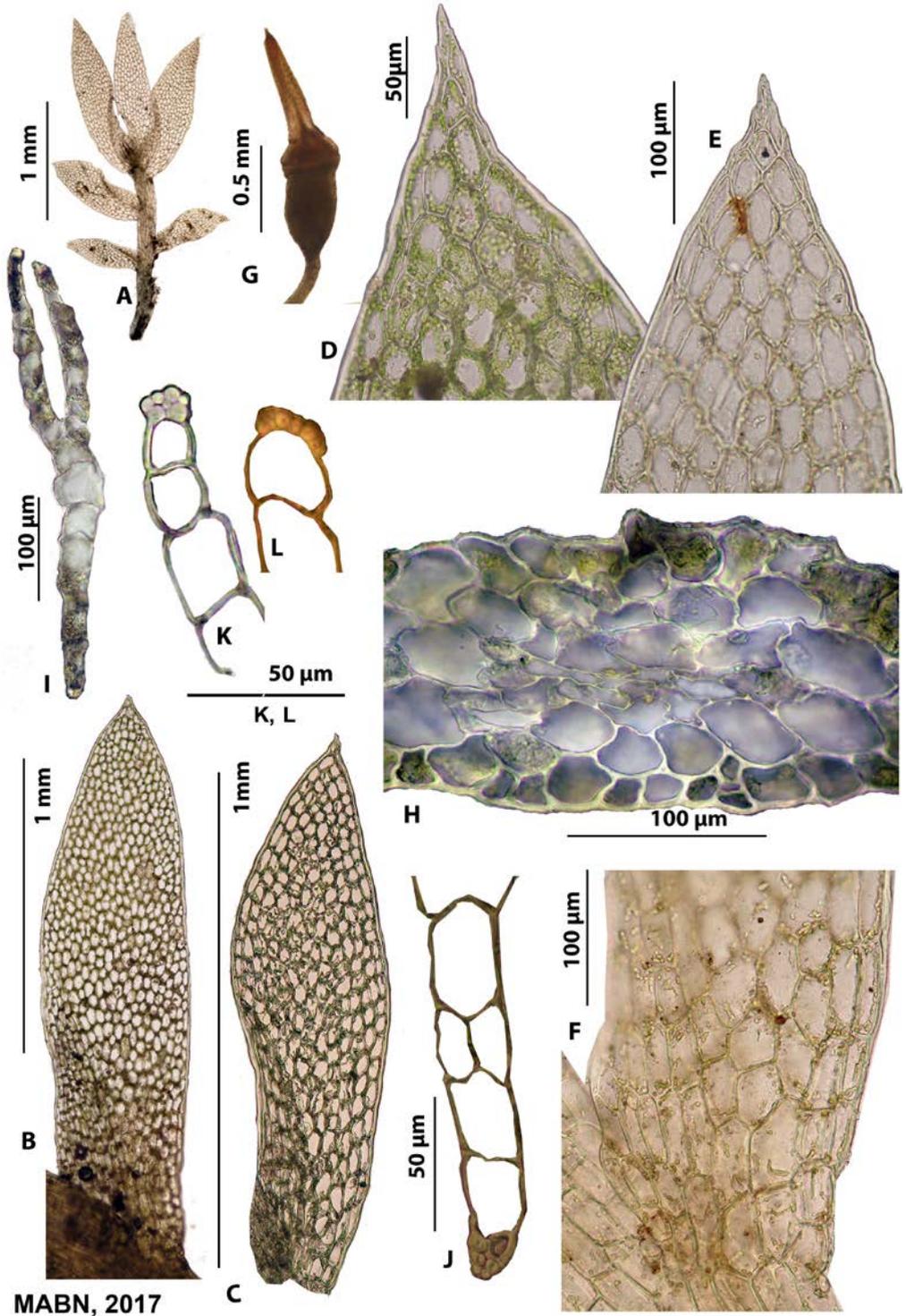
REMARKS. Previously, *F. molliculus* (Bizot & Pócs 1979) and *F. hedbergii* (Bruggeman-Nannenga 1997) were considered to be *F. enervis*. Most specimens were re-examined during the preparation of the present paper. Many specimens earlier identified as *F. enervis* appeared to differ from that species by having limbidia thicker than 1–2 layers and in having 40 or more columns of exothecial cells (versus ± 32 in *F. enervis*). Of the African material studied, only the type specimen could be retained in *F. enervis*. Comparison of tropical African collections with acute-acuminate apices, 40 or more columns of exothecial cells and pluristratose limbidia with *F. dealbatus* from New Zealand and Guadalcanal showed the African collections to be conspecific with *F. dealbatus* Hook. f. & Wils.

No trans-sections were made of the type of *F. hookerioides*. Its limbidia were estimated to be pluristratose. Sporophytes were lacking.

In *F. dealbatus* the limbidia often ‘fold’ around the laminal cells and then are not truly pluristratose (e.g., Fig. 1L, which in the present paper has been interpreted as 7-stratose).



Fig. 1. *Fissidens dealbatus* Hook. f. & Wilson. A – perichaetial plant, B & C – leaves, D & E – leaf apices, F – insertion of leaf, G – capsule and calyptra, H – trans-section of stem, I – trans-section of leaf with unistratose laminae, J – trans-section of dorsal lamina with bistratose dot, K & L – trans-sections of vaginant lamina limbidia (K with 1–2-cells wide, 2–4-stratose limbidium; L with one-cell-wide, 7-stratose limbidium) – A, F, J from Pócs 9104/P; B, D, G–I, K, L from Müller B98; C from Pócs 8430A; E from Townsend 75/330.



SUBSTRATE AND HABIT. On soil, rarely on rocks, once on soil-covered wood. Growing scattered among other mosses or less often in pure mats.

HABITAT. In all kinds of forests (montane rainforest, montane cloud forest, montane evergreen forest, mesic montane forest, submontane rainforest, bamboo forest); also collected in *Philippia* heath with remnants of montane evergreen forest and in a *Eucalyptus* plantation.

ELEVATION. In Africa known from 950–2500 m a.s.l.; in New Zealand from 0–900 m a.s.l. (Beever *et al.* 2002).

DISTRIBUTION. *Fissidens dealbatus* is widespread in tropical Africa. West tropical Africa (Bioko), west central tropical Africa (Central African Republic), east tropical Africa (Kenya, Rwanda, Tanzania, Uganda), south tropical Africa (Malawi) and western Indian Ocean (Madagascar). Outside Africa it is known from New Zealand, New Caledonia, Australia (Beever *et al.* 2002) and the New Hebrides (Iwatsuki & Suzuki 1995).

Also recorded (as *F. enervis*) from the Democratic Republic of the Congo (O’Shea 2006), São Tomé and Príncipe (Shevock *et al.* 2013) and Zambia (O’Shea 2006). These are not included in the distribution since they may refer to *F. dealbatus*. A specimen from South Africa [Transvaal, Wolf-spruit, Ofcolaco, *Brenan M 3280A* (E)] seems to be this species, but the material is too poor for certainty.

SELECTED SPECIMENS EXAMINED: CENTRAL AFRICAN REPUBLIC, type *F. molliculus*. – EQUATORIAL GUINEA, Bioko, 10 km S of Luba, Berg Caldera, 3°22’N, 8°32’E, *Müller B98, p.p.* (DR, L). – MADAGASCAR, type *F. hookerioides*. – KENYA, Rift Valley Province, Trans Nzoia District, Cherangani, *R.A. Maas Geesteranus 10313a* (L, a few stems). – MALAWI, S Escarpment of Zomba Plateau on the S slope of Kuchawe summit, *Pócs 9171/BD* (EGR, L, a few mixed with *F. rotareaui* P de la Varde). – RWANDA, Lac Bulera (au nord-est de Ruhengiri), rive nord du Lac, *De Sloover 13.565, p.p.*, a few mixed with several other *Fissidens* species (L, a few stems). – TANZANIA,

Nguru Mountains in Morogoro District, watershed between Chogowale and Divue headwaters, 6 km SSE of Maskati village, *Kis & Pócs 9129/Q* (EGR, L); West Usambara Mts, E of Magila village, SW edge of Ndelemai F.R., *Pócs 8430/A* (EGR, L); Arusha National Park, E slopes of Meru Crater, along trails between Kitoto and Cabin at Njeku, 3°14’S, 34°47’E, *Crosby & Crosby 11231* (MO, L); Mt. Meru, SW slope of Mt. Meru above Laikinoi, *Pócs & Ochyra 88147/H* (EGR, L). – UGANDA, Kibale NP, Research Forest, MUBFS, Kanyawara. 0°33’N, 30°21’E, *Wigginton U 5285A* (E, L, a few mixed with *Fissidens intramarginatus* s.l. and a few *F. zollingeri*); Kisoro, *Fitzgerald s.n.* (PC). – NEW ZEALAND, North Island, South Auckland Land District, Te Kauri Scenic Reserve, 38°4’0’’S, 174°59’0’’E, *Spragg 93* (AK, L). – GUADALCANAL, near camp at base of Mt. Popemanaseu, *Van Zanten 682582* (GRO, L).

ILLUSTRATIONS: Beever *et al.* (2002: 28; 2014: fig. 10), Bruggeman-Nannenga (2006a: fig. 7a–c, as *F. enervis*), Chuah-Petiot (2003: fig. 30, as *F. enervis*, but the shape of the apex indicates *F. dealbatus* rather than *F. enervis*), Salmon (1899: figs 13–15), Stone (1986: fig. 3), Seppelt & Stone (2016: *F. dealbatus*).

2. *Fissidens enervis* Sim

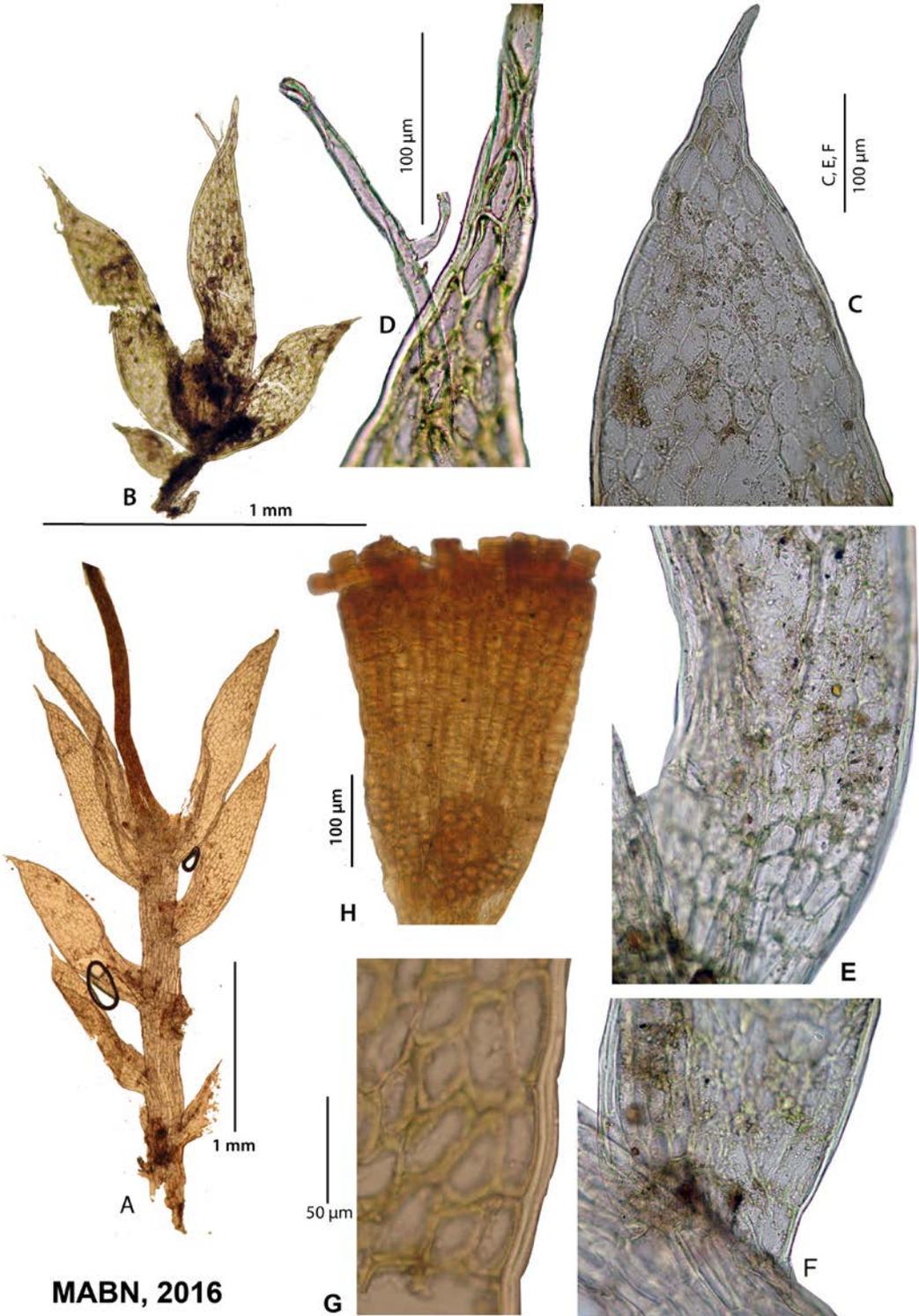
Fig. 2

Trans. Roy. Soc. South Africa **15**: 187, fig. 1926 – PROTOLOGUE: [SOUTH AFRICA], Natal, Town Bush Valley, Pietermaritzburg, 3000 ft, 1917, *Sim 9899*, Nottingham Road, *Sim 8648* and Burgerstreet, Pietermaritzburg, *Sim 9900* – LECTOTYPE (designated by Magill 1981) labeled: Distr. Pietermaritzburg, Town bush valley, 2500 feet, IV 1917, *Sim 9899* (PRE!, ISOLECTOTYPE BM000871763!).

Stem in cross section without central strand (based on Indian specimen), unbranched, with pinnately arranged leaves (only fertile stems seen), $\pm 3.0 \times 1.0$ –2.5 mm; *rhizoids* basal, from lower stem cortex and from apical part of lamina, hyaline or brown, smooth; *leaves* distant, to 7 pairs, crispate when dry, elliptical with cuspidate tip, 0.95 – 1.70×0.25 – 0.40 mm, 3.5–5.0 as long as wide, limbate; *limbidium* \pm confluent at leaf apex, confluent at apex of vaginant laminae, reaching insertion of dorsal lamina, in upper leaves ending well above insertion of vaginant lamina, in lower



Fig. 2. *Fissidens enervis* Sim. A – sporophytic stem, B – perigonal stem, C – leaf apex, D – detail of leaf apex with rhizoid growing from lamina, E – mid leaf, F – insertion of leaf, G – limbidium on mid dorsal lamina, H – capsule. All from type specimen.



leaves sometimes reaching insertion, in mid dorsal lamina 7.5–14.5 μm wide, more than unistratose (estimated); on vaginant lamina, 20 μm wide, 1–3-stratose (based on Indian specimen), marginal throughout; *vaginant lamina* ca 1/2 of leaf length, at base less wide than stem, slightly rounded at insertion, not decurrent, unistratose, subequal; *dorsal lamina* \pm straight towards insertion, reaching insertion, not decurrent, *dorsal* and *apical lamina* unistratose; *costa* lacking or vestigial, composed of narrow cells, reaching to half the length of vaginant lamina, lacking in lower leaves (no cross sections seen); *mid dorsal laminal cells* plane, 32–67 \times 15–30 μm ; *mid vaginant laminal cells* large, plane, 51.5–54.0 \times 18.5–25.5 μm . *Gemmae*, some leaves produce rhizoids from apical part (Fig. 2 D).

Fertile parts. *Perigonia* terminal on small, ca 1.5 mm long, stems; *antheridia* 230–250 μm long; *perichaetia* terminal, *perichaetial leaves* \pm 1.3 mm long; *archegonia* not seen; *calyptra* not seen. *Sporophyte*, *seta* \pm 4 mm long, smooth, 1 per perichaetium; *capsule* symmetrical, 0.5–0.7 \times 0.25–0.40 mm, \pm 32 columns of quadrate-oblong exothecial cells; *peristome* scarious-type, tooth base \pm 33.5–43.0 μm wide; *operculum* and *spores* not seen.

Fissidens enervis is a rare species known from a few small collections. It is characterized by its cuspidate leaf apex, (\pm) ecostate leaves and large laminal cells. Rhizoids (hyaline, green or brown) growing from laminal cells are frequently present (seen in the type specimen as well as in Indian specimens). It strongly resembles *F. hyalinus* (not known from Africa). These two species may eventually prove to be conspecific. In this paper I take a conservative stand and consider specimens with unistratose limbidia and acute to acute-acuminate leaf apices to be *F. hyalinus*, and specimens with 1–2-stratose limbidia and cuspidate apices to be *F. enervis*. In Africa *Fissidens enervis* has been confused with *F. dealbatus* (see under this species). Sporophytic specimens are clearly distinct. *Fissidens enervis* has a smaller number of columns of exothecial cells (32 versus 40 or more in *F. dealbatus*). Gametophytic stems differ in the shape of the leaf apex (cuspidate in *F. enervis* and

acute to acute-acuminate in *F. dealbatus*) and in having thinner limbidia (limbidia of *F. dealbatus* are 1–7-stratose). Rhizoids growing from laminal cells are present in all known collections of *F. enervis* but have not been observed in *F. dealbatus*. Unfortunately, capsules are frequently lacking and the collections are often too small to make trans-sections. In such cases the thickness of the limbidium is estimated from a surface view, which is unreliable.

REMARKS. The lectotype of *F. enervis* consists of a few poor stems, including one sporophyte with \pm 32 columns of exothecial cells on the theca.

Fissidens splachnoides Broth. is very similar. Its type (Queensland, Indooroopilly, *F.M. Bailey 256*, p.p. (H-BR) also has a cuspidate leaf apex (Stone 1986, fig. 2) and 1–2-stratose limbidia. However, in this species the walls of the lateral bands of the vestigial costae are thickened [observed in the type specimen *Bailey 256* (S!), in *Norkett 16173* (BM!) and in New South Wales, *Watts 5624* (H-Broth!)], which is not the case in an Indian specimen of *F. enervis*.

SUBSTRATE & HABIT. On soil, growing scattered among other mosses.

ELEVATION. South Africa: 900 m a.s.l.; India: 900–1200 m a.s.l. (Bruggeman-Nannenga *et al.* 2016).

DISTRIBUTION (amended). Southern Africa (South Africa), Russia? (Ignatov *et al.* 2007), India (Western Ghats). Rare.

To the extent that it could be checked, the African distribution of *F. enervis* in O'Shea (2006) refers, with the exception of the type specimen, to *F. dealbatus*. The *F. hyalinus* with 1–2-stratose limbidia and cuspidate apices in Ignatov *et al.* (2007) seems to refer to *F. enervis*.

SPECIMENS EXAMINED. SOUTH AFRICA, LECTOTYPE (PRE); Natal, Nottingham Road, 900 m, IV 191, Van de Bijl (*Sim 8648*) (PRE). – INDIA, Kerala, Palakkad District, Nelliampathi; on land cuttings, 1200 m, 13.07.2015, *K.M. Manjula 1138B* (CALI, L).

ILLUSTRATIONS. Bruggeman-Nannenga *et al.* (2016: figs 1 & 2), Magill [1981: fig. 8(1–7)], Sim (1926: 18.900).

3. *Fissidens metzgeria* (Müll. Hal.) Broth.

Fig. 3

Oefvers. Förh. Finska Vetensk.- Soc. **35**: 38, 1893 – *Conomitrium metzgeria* Müll. Hal., *Linnaea* **39**: 358, 1875 – PROTOLOGUE: [border area of DEMOCRATIC REPUBLIC OF THE CONGO and CENTRAL AFRICAN REPUBLIC], Monbuttu-regiones, prope Bongua inter *Plagiothecium monbuttoviae*, ut ex Diatomaceis elucet, in locis aquosis, 14. April 1870. *Schweinfurth* – LECTOTYPE (designated by Bruggeman-Nannenga 1997) with label: Monbuttu Land, inter *Plagiothecium monbuttoviae* vereinzelt, 14 IV 1870. Collector not indicated (NY!); ISOLECTOTYPE: BM (slide 3619).

Fissidens bryum Müll. Hal. ex Dusén, Kongl. Svenska Vetensk. Acad. Handl. n.s. **28**(2): 14, figs.: a, b & tab. II, f. 8. 1895 – PROTOLOGUE: CAMEROON [Camerunia], supra Bomanam pagum, c. 1100 m, VII 1892, *Dusén s.n.* – HOLOTYPE label: Africa Occ., Camerunia in monte Camerun, ca. 1100 m, VII 1892, *Dusén s.n.* (S!); ISOTYPE label: Kamerun, Bomana, 1100 m, auf Steinen, VII 1892, *Dusén 834* (PC-PV!).

Fissidens bryum Müll. Hal. ex Dusén var. *terrestris* P. de la Varde, Rev. Bryol. n.s. **4**: 57, 1931 – PROTOLOGUE: GABON, Tsengué (Monzo), sur terre argileuse, *Le Testu 4880* – HOLOTYPE label: as protologue with added collection date: 6 I 1931 (PC-PV!).

Stem without or with weak central strand, not heterocaulous, with pinnately arranged leaves, unbranched, 5.0–21.0 × 1.5–5.0 mm, browning in KOH or not; *rhizoids* basal and from cortex of basal part of stem, brown, smooth; *axillary* cells not differentiated; *leaves* shiny, distant to close, up to 15 pairs, slightly crispate when dry, flattening when moistened, elliptical to elliptico-lanceolate with acute to acute-acuminate apex, 1.5–4.0 × 0.3–0.7 mm, 3.0–5.5 times as long as wide, margin entire to undulate-entire, limbate; *limbidium* on all laminae of all leaves, ill-defined, lax, ± reaching apices of leaf and vaginant lamina, reaching insertions of vaginant and dorsal laminae, in mid dorsal lamina 10.5–33.5 µm wide, unistratose, marginal; on vaginant laminae weak, up to 56 µm wide, unistratose, marginal to weakly intramarginal; *vaginant lamina* 1/2 (–3/5) of total leaf length, about half as wide as stem at insertion, slightly open, unistratose; *dorsal lamina* tapering towards base, straight, reaching insertion, not to inconspicuously decurrent; *dorsal* and *apical lamina*

unistratose, infrequently with scattered, unicellular bistratose dots; *costa* lacking, rarely weakly developed; *mid dorsal laminal cells* oblong-hexagonal, 55.0–120.0 × 15.0–53.5 µm, plane to inconspicuously convex, smooth; *limbidial cells* in mid dorsal lamina 130–150 × 5–12 µm with slightly thicker walls than inner cells; *mid vaginant laminal cells* oblong-rectangular 75–152 × 5–24 µm. Clusters of *gemmae* frequently present on lamina, particularly near apex, less often growing from stem cortex; *gemmae* brown, unbranched to heavily branched, multi-cellular.

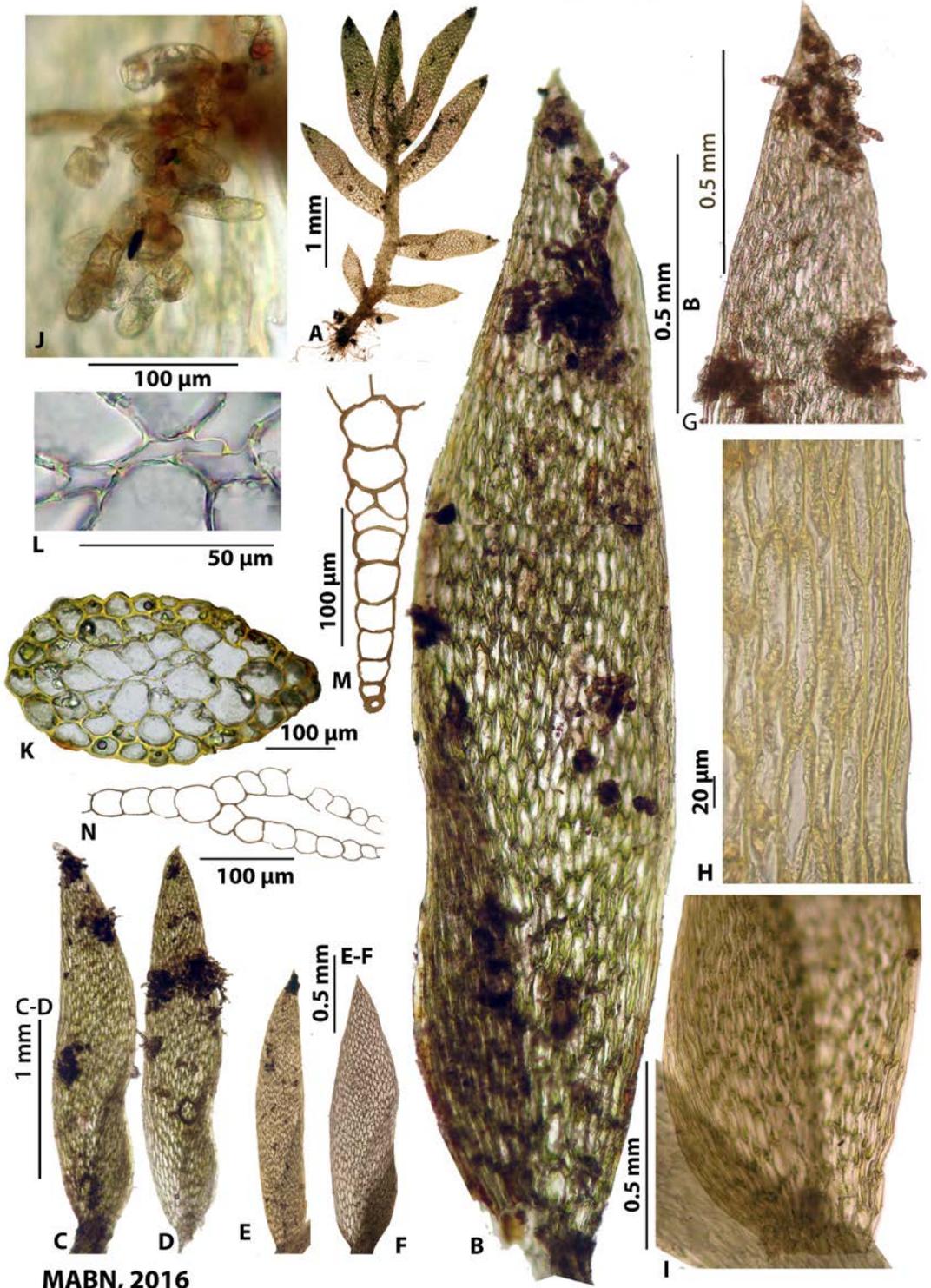
Fertile plants rare, perigonial and perichaetial stems hardly differ from vegetative ones, easily overlooked. *Perigonia*, *perichaetia* and *synoicia* terminal; perigonial plants up to 6 mm long, *antheridia* 210–500 µm long; *perichaetia* terminal, *perichaetial leaves* hardly longer than stem leaves, 1.5–4.0 mm long, *archegonia* 200–450 µm long; *calyptra* tightly clasping operculum, 0.65–0.70 mm long, smooth to scabrose. *Sporophytes* (only 2 very young ones seen), 2 per perichaetium.

Fissidens metzgeria is characterized by its ecostate, limbate leaves, ill-defined limbidia, 5–12 µm wide limbidial cells and large inflated cells (55.0–120.0 × 15.0–53.5 µm in mid dorsal lamina). Epiphyllous clusters of multicellular *gemmae* are frequently present on the apical part of the leaf. The other two African ecostate, limbate species, *F. dealbatus* and *F. enervis*, have distinct, narrower limbidia of narrow limbidial cells and lack epiphyllous clusters of *gemmae*. *Fissidens ecuadorensis* Pursell & Brugg.-Nann. is a smaller (stems up to 4 mm long) Neotropical ecostate species with a similar ill-defined limbidium.

SUBSTRATE AND HABIT. On soil, less often on rocks, living or dead wood. Often scattered among other mosses, rarely in mats or tufts.

HABITAT. On banks of paths, roads and streams in humid, shady to swampy sites and also near waterfalls. In all kinds of forest (montane and submontane rainforest, montane cloud forest, dry semi-deciduous forest, swamp forest, both pristine and secondary).

ELEVATION. 360–2300 m a.s.l.



MABN, 2016

DISTRIBUTION. Endemic to Africa. Known from west tropical Africa (Guinea; Schultze-Motel 1975, Nigeria), west central tropical Africa [Cameroon, Central African Republic, Democratic Republic of the Congo, Equatorial Guinea (Bioko), Gabon, São Tomé], northeast tropical Africa (Ethiopia; Hylander *et al.* 2017), east tropical Africa (Kenya, Rwanda, Tanzania, Uganda) and south tropical Africa (Angola; Müller 2015).

SELECTED SPECIMENS EXAMINED: CAMEROON, [British Cameroons], S. side Cameroon Mountain, Div. Victoria, Mann's Spring below Liwonge, (*Brenan*) *Richards* 4320 (PC-PV); Bamenda, Bafut-Ngamba Forest Reserve, *Richards* 5255 (PC-PV, BM). – CENTRAL AFRICAN REPUBLIC, Bangui, *Assel* 120 (PC, L). – DEMOCRATIC REPUBLIC OF THE CONGO [Congo belgica], Mwenda, affluent de la Biangolo, *Demaret* 4741 (PC-PV); prov. Kivu, Irangi Forest Station 110 km W of Bukavu, Bryotrop Exp. 1991, *Pócs* 6766 (EGR). – EQUATORIAL GUINEA, Insel Bioko, Berg Caldera 10 km S of Luba, 3°23'N, 8°32'E, *F. Müller* B 104 (DR, L); Hochland von Moca, 3°20'N, 8°40'E, *F. Müller* B 410, *p.p.* (DR, L). – GABON, entre Ipoungou et Dibandi (or Dibaudi?), *Le Testu* 6120 (PC-PV). – KENYA, North Kavirondo District, S.E. of road between Rondo and Kakamega Sawmills, *Townsend* 77/258, *p.p.* (E, L, a few mixed with *F. intramarginatus* s.l.). – NIGERIA, Prov. Benin, Distr. Benin, valley between Idanre and Olofin, *Brenan & Jones* R3838 (E, L, *p.p.* mixed with *F. zollingeri*); S.E. State, Obudu, Cattle Range, *McFarlane* 5339A Mitt. (L, *p.p.* with *F. porrectus*). – RWANDA, Pref. de Cyangugu, Forêt de Nyungwe, Bryotrop Exp. 1991, *Pócs* 6473 (L); Kamiranzovu, 2°29'39"S; 29°8'32"E, Bryotrop Exp. 1991, *Frahm* 6330 (L, with *F. porrectus*). – SÃO TOMÉ AND PRÍNCIPE, SÃO TOMÉ, Picada para a Lagoa Amélia, *Sérgio* ST100 (L, LISU). – TANZANIA, SW from W Usambara Mts, on the plateau of Mafi Hill, near the headwaters of Kwalukonge stream, *Pócs* 8522/AN (EGR, L – one stem in with *F. weirii*); Mbulu Highlands, Guay Hill, *Pócs* 9009/AA, *p.p.* mixed with *F. usambaricus* (EGR, L). – UGANDA, Bushenyi District: Kasyoha-Kitomi FR, 0°19'S, 30°7'E, *Hodgetts* U 4584A (E, L); Rukungiri District, Bwindi forest, Buhoma. 0°59'S, 29°36'E, *Wigginton* U8347 A (E, L).

ILLUSTRATIONS: Potier de la Varde [1929: 3, and (same figure) 1936: 3; both as *F. bryum*]; Bruggeman-Nannenga (2006a: fig. 7d–f).

4. *Fissidens usambaricus* Broth. Fig. 3

Bot. Jahrb. Syst. **20**: 182. 1894 – PROTOLOGUE: TANZANIA, Usambara, Lutindi, am Boden in Hochwalde, spärlich, *Holst* 3472, *p.p.* – HOLOTYPE: mica-slide without collection number H-BR!; ISOTYPES: BM!, SI!

Fissidens chevalieri Corb., Bull. Mus. Natl. Hist. Nat. **18**: 108. 1912 – PROTOLOGUE: CENTRAL AFRICAN REPUBLIC, [Haut-Oubangui], Krébedjé (Fort-Sibut), 11 Oct. 1902, *Auguste Chevalier s.n.* – HOLOTYPE in Corbière's herbarium in PC (not seen); ISOTYPES: PC!, PC-PV! [both with label: Oubangui, Krébedjé, *Chevalier* 44A, 11–12 Oct. 1903 (orthographic error for 1902?)] and H-BR! (label: Oubangui, Krébedjé. *Chevalier s.n.* 11–12 X 1902!).

Fissidens usambaricus Broth. var. *acutifolius* P. de la Varde, Bull. Soc. Bot. France **73**: 377, 29. 1926 – PROTOLOGUE: [CENTRAL AFRICAN REPUBLIC] près Ngono, 20 km. N.-O. Bambari, *R.P. Tisserant s.n.* – HOLOTYPE label: Oubangui, 20 km N O Bambari, près Ngono, mêlé à *Trichosteleum tisserantii* sur tronc d'arbre mort. 1925, *Tisserant s.n.* (PV 2145B) (PC-PV!); ISOTYPE: près Ngono, 20 km N-O. Bambari, *R.P. Tisserant* (PV 2145B) (BM-000871903!).

Stem without or with weak central strand, not heterocaulous, mostly unbranched, 3.0–9.0(–15.0) × 1.5–3.0 mm, with pinnately arranged leaves; *rhizoids* basal, infrequently from mid stem cortex, brown to hyaline; *axillary cells* not differentiated; *leaves* green, distant, 5–15 pairs, hardly altered to crispate when dry, broadly elliptical to broadly elliptico-oblongate or oblong with widely acute to obtuse or rounded-obtuse apex, 0.9–1.5(–2.1) × 0.4–0.6(–0.7) mm, 2–3 times as long as wide, elimbate, margin subentire, near apex sometimes serrulate; *vaginant lamina* 2/5–3/5 of total leaf length, narrow, at insertion narrower than stem, slightly unequal, unistratose, infrequently with scattered unicellular bistratose dots; *dorsal lamina*

Fig. 3. *Fissidens metzgeria* (Müll. Hal.) Broth. A – stem, B–F – leaves (black areas are clusters of gemmae), G – leaf apex with 3 clusters of gemmae, H – limbidium mid dorsal lamina, I – insertion of leaf, J – cluster of gemmae, K – trans-section of stem, L – detail of K showing weak central strand, M & N – trans-sections of leaf (M showing isolated bistratose dot in dorsal lamina). A from *Pócs* 9009/AA; B–D from *Hodgetts* U4254A; E, G–H, K–N from *Müller* B104; F, J from *Müller* B410; I from *O'Shea* U 5532A.

± straight towards insertion, reaching insertion, occasionally decurrent, unistratose, rarely bistratose near insertion; *apical lamina* unistratose; *costa* ± absent; *mid dorsal laminal cells* hexagonal, variable in size (see remark below), 22.5–93.0 × 14.0–40.0 µm, smooth, plane to inconspicuously convex; *marginal cells* smaller, isodiametric to oblong, on the mid dorsal lamina 15.0–33.5 × 6.5–15.9 µm; *mid vaginant laminal cells* hardly differentiated to larger, 45.0–136.0 × 7.5–32.0 µm; *basal vaginant laminal cells* hardly differentiated to oblong, 45.0–60.0 × 7.5–14.5 µm. No *gemmae* seen.

Fertile parts. *Gametangia* (perigonia, perichaetia and synoicia) terminal; *perigonial stems* 0.5–2.3 mm long; *antheridia* 180–230 µm tall; *perichaetial* and *synoicial stems* 3–5 mm tall; *archegonia* ± 300 µm long; *perichaetial leaves* 1.85–2.20 mm long; *calyptra* tightly covering rostrum, smooth to slightly bulging. *Sporophytes* rare. *Seta* 1.5–3.0 mm long, 1–2(–3) per perichaetium, smooth or slightly scabrose in basal half; *capsule* symmetrical, rarely slightly inclined, 0.40–0.65 × 0.20–0.45 mm, with ± 32 columns of oblong to quadrate exothecial cells with thickened corners; *peristome* scariosus-type, tooth base 45–48 µm wide; *operculum* rostrate, 0.55 mm long; *spores* subglobose, 8.0–13.0 µm diam., thick-walled or not, smooth to indistinctly papillose.

This species is easily recognized by its elimbate, (±) ecostate leaves and large laminal cells with a border of 1–2 rows of smaller, isodiametric to oblong cells. The variation of cell size in this species is remarkable. Some collections have relatively small, 22.5–45.0 × 14.0–21.0 µm, mid dorsal laminal cells, whereas others have much larger, 56–80 × 30–40 µm, mid dorsal laminal cells (e.g., *O'Shea 2628A* and *Müller Z 215*); mid vaginant laminal cells can be 45–60 × 7.5–14.5 µm in some collections or as large as 88–136 × 17.5–32 µm in others. The presence of transitional forms makes it

impossible to distinguish distinct groups based on this character. *Fissidens usambaricus* is the only elimbate, ecostate species known from Africa. The ecostate *F. hylogenes* Dixon from New Zealand similarly has elimbate leaves but has smaller, ± 12 × 9 µm (versus 15.0–24.0 × 7.5–15.0 µm) marginal cells and crenulate instead of subentire margins. [Specimen seen: *Sainsbury et al., Musci exsicc. Novae-Zelandiae 10* (S)].

SUBSTRATE AND HABIT. Mostly on soil, less often on rock, termite mounds or wood. Growing scattered among other mosses, rarely in loose or dense mats or tufts. Easily overlooked.

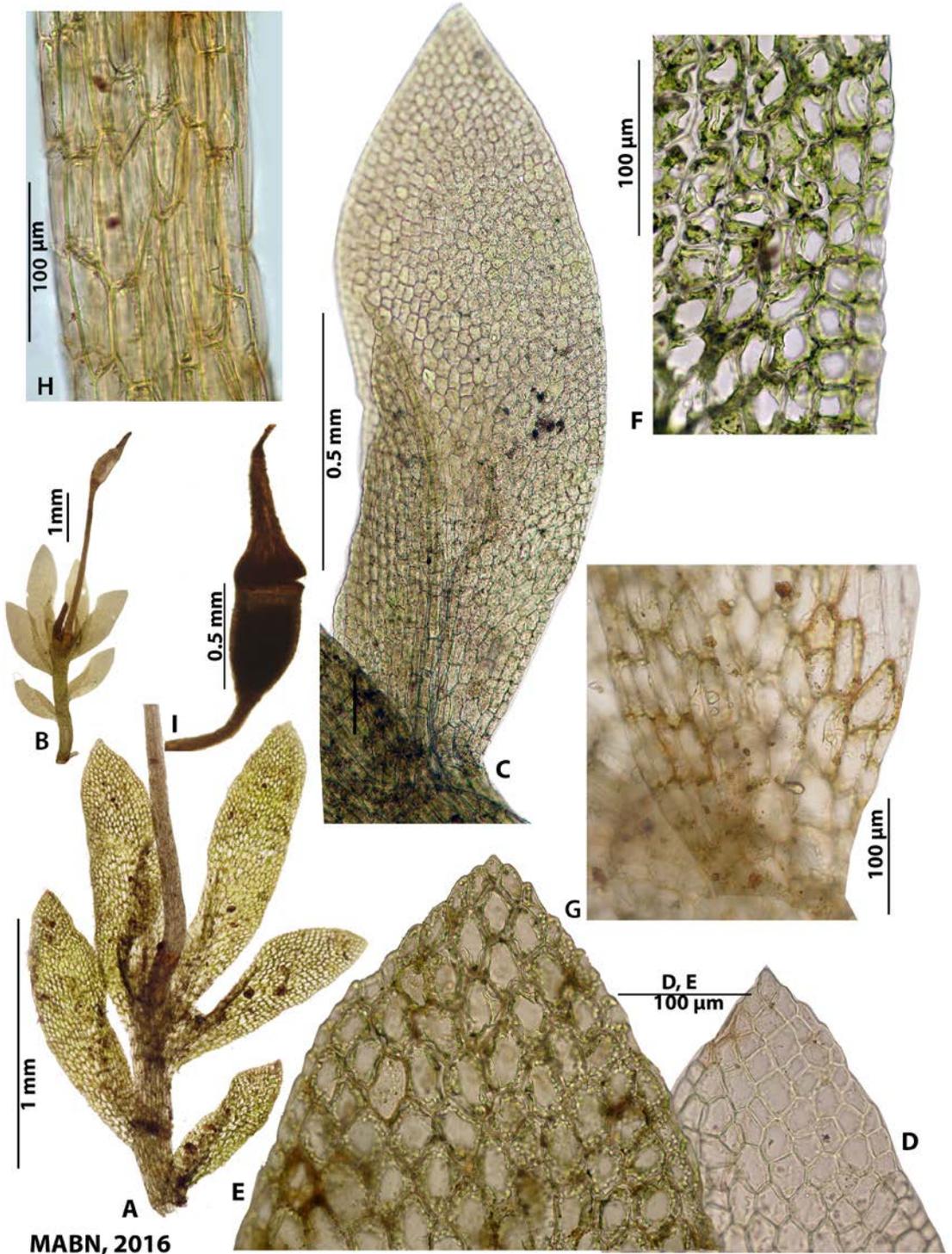
HABITAT. Usually collected on shady banks and road cuts, also found near a waterfall, on rock in streamlet and bank of stream. Known from dry semi-deciduous forests, mesic montane forests and rainforest (lowland, submontane and montane), also from an Afro-montane forest, woodbush, and from *Pinus* plantations. Many labels give no details.

ELEVATION. 25–2400 m a.s.l.

DISTRIBUTION. A widespread African endemic. Known from Macaronesia: Cape Verde Islands (O'Shea 2006), west tropical Africa (Ivory Coast; O'Shea 2006), Nigeria (O'Shea 2006), west central tropical Africa (Cameroon; O'Shea 2006), Central African Republic, Democratic Republic of the Congo, Gabon, Rwanda, São Tomé (Shevock *et al.* 2013), northeast tropical Africa (Ethiopia), east tropical Africa (Kenya; O'Shea 2006), Tanzania, Uganda, south tropical Africa (Malawi) and southern Africa (South Africa, Swaziland; O'Shea 2006). New for Malawi and Gabon.

SELECTED SPECIMENS EXAMINED: CENTRAL AFRICAN REPUBLIC, types of *Fissidens chevalieri*. – DEMOCRATIC REPUBLIC OF THE CONGO, prov. Kivu: Ruwenzori, Lanuri, *Bequaert 4447* (PC, PC-PV, BM); Kahuzi-Biega Nat. Park. Surroundings of Camp Biega 30 km W of Bukavu. Loc. 128, Bryotrop Exp. 1991, *Pócs 7356*, *p.p.*, 1 stem (L); Pinga, 96 km NW

Fig. 4. *Fissidens usambaricus* Broth. A & B – sporophytic plants, C – mid leaf of perichaetial stem, D & E – leaf apices, F – margin mid dorsal lamina (marginal cells at right), G – insertion of leaf, H – part of seta showing large cortical cells, I – capsule with calyptra. A from *Pócs 90096/AA*; B, C from *Porley U45A*; D, H from *Hylander 4527*; E, G from *Porley 410A*; F from *Porley U636A*; I from *Lübenau SA309*.



Goma, Flusstal 4 km NNO Ort; 1°0'S. Br; 28°43'Ö. L., *F. Müller* Z 215 (DR, L). – ETHIOPIA, Kaffa, Gimbo, Bonga. 1.5 km SE of Central Bonga, *Hylander* 4527 (S, L); Kaffa, Gimbo, Ufudo, Chechuwuta forest, 6 km S of Gimbo, *Hylander* 4532 (S, L). – GABON, Ogooué-Ivindo, Makokou, Réserve Intégrale d'Ipassa, 0°30'43.0"N 12°48'12.1"E, *Vanderpoorten* GAB 1399 [a single stem mixed with *F. glaucissimus* (L)]. – MALAWI, S. Escarpment of Zomba Plateau on the S slope of Kuchawe summit, *Pócs* 9171/BB (EGR, L mixed with a few *F. dealbatus* and *F. rotareui*). – SOUTH AFRICA, Mariepskop, *Vorster* 1449B (L); Transvaal, Drakenberge, *Lübenau* SA309 (L). – RWANDA, Rwanda. Pref. de Cyanguu, Forêt de Nyungu, Uwinka Station along road from Butare to Cyanguu, Bryotrop Exp. 1991, *Frahm* 6494 (BONN, L). – TANZANIA, Mbulu Highlands, Quay Hill at the NNE end of Nou Forest Reserve, 6 km SSW of Mbulu town, *Pócs* 90096/AA, *p.p.* mixed with *F. metzgeria* (EGR, L); Mbeya District, Forest Reserve by the Mbeya/Chunya road ca 12 km from Mbeya. *Townsend* 75/330 (E, *p.p.*, with *F. dealbatus*). – UGANDA, Kabale District, Bwindi National Park, Ruhija, *Porley* U45a, *p.p.*, mixed with *F. intramarginatus* s.l. (E, L); Kabarole, Rwenzori National Park, lower Mubuku River; 0°21'N, 30°1'E, *Hodgetts* U 4534A, *p.p.* mixed with *F. dealbatus* (E, L); Rukungiri Distr., Bwindi NP, *Porley* U 636A c. sp. (E, L).

ILLUSTRATIONS: Bruggeman-Nannenga (2006a: fig. 7 g–i), Chuah Petiot (2003: fig. 35), Magill (1981: fig. 8–13), Salmon (1899: fig. 16).

II. COSTATE SPECIES

All species included in the present paper have large (at least 30 µm long) laminal cells, limbidia on all laminae of all or most leaves, and a subgenus *Aloma* sporophyte. Costae are frequently short for the genus (e.g., *F. flaccidus*, *F. grandifolius*) but can be excurrent, as in *F. minutifolius* Broth. & P de la Varde and *F. zollingeri* Mont. Worldwide, costate species with large cells include limbate, partly limbate and ± elimbate species. Most have

smooth cells but a few have mammillose cells (e.g., *F. flaccidus* var. *mammillosus* Brugg.-Nann., *F. lindbergii* A. Jaeg., *F. yucatanensis* Steere, *F. bififormis* Mitt.). In *F. zollingeri* (smooth cells) and *F. bififormis* Mitt. (with mammillose cells, not included in this paper), large inflated cells are restricted to the vaginant laminae.

5. *Fissidens flaccidus* Mitt.

Fig. 5

Trans. Linn. Soc. London **23**: 56, t. 6, f. 18, 1–4. 1860 – PROTOLOGUE: NIGER, on the earth, *Vogel* – HOLOTYPE NY!; ISOTYPES BM as *Vogel* 2!, *Vogel* 3! and *Vogel* 4!

Fissidens atroviridis Besch., Ann. Sci. Nat. Bot. sér. 7, **2**: 86. 1885 – PROTOLOGUE: MAYOTTE, sur la terre à M'Sapéré, *M. Marie* 10 – HOLOTYPE: BM, ISOTYPES PC! (as Mt. Sapéré, 19 II 1881), NY! (*sine dato*), H-BR! (as Comore-insula, Mayotte, *Marie s.n.*, *sine dato*).

Fissidens cellulusus Mitt., J. Linn. Soc. Bot. **22**: 319. 1886 – PROTOLOGUE/label: [TANZANIA], Usagara Mountains, on the earth. *Bishop Hannington*; HOLOTYPE: NY!, ISOTYPE: H!

Fissidens luridus Renaud & Cardot, Bull. Soc. Roy. Bot. Belgique **29**: 172. 1890 – PROTOLOGUE: MADAGASCAR, Imerina, Mahamasina, alt. 1210 m (*Camboué*) – LECTOTYPE (designated by Bruggeman-Nannenga 2009) labelled: Madagascar, Mahamasina, Ouest de Imerina, alt. 1210 m, *Camboué s.n.* (PC! stamped herb. Cardot). The specimen with the same label in Renaud's herbarium (PC) is *F. leucocinctus* Hampe.

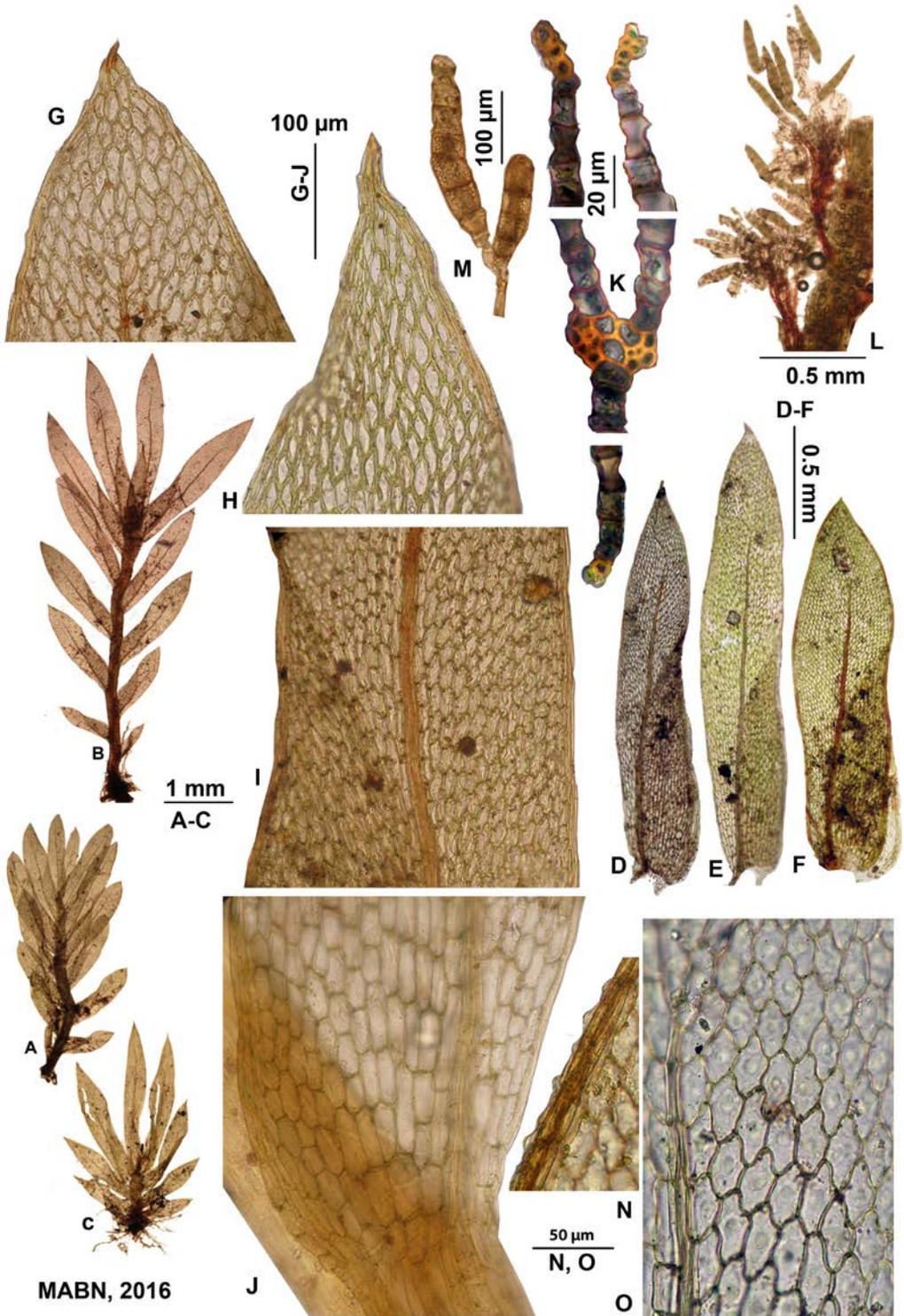
Fissidens bocarangensis P. de la Varde, Rev. Bryol. Lichénol. **11**: 166. 1939; new name for *F. ensifolius* P. de la Varde, Rev. Bryol. **9**: 195, f. 1. 1937 (*hom. illeg.*, non *F. ensifolius* Broth.) – PROTOLOGUE: CENTRAL AFRICAN REPUBLIC [Oubangui], Bocaranga, sur l'humus, *Eckendorff s.n.* – HOLOTYPE: PC-PV! (labeled: Oubangui, Bocaranga, 31 VII 1934, *Eckendorff s.n.* (PV 5971)).

Non-African synonyms:

Fissidens maceratus Mitt., Trans. & Proc. Roy. Soc. Victoria **19**: 91. 1882 – PROTOLOGUE: AUSTRALIA, Queensland, Brisbane River, *Bailey* – HOLOTYPE: NY.



Fig. 5. *Fissidens flaccidus* Mitt. var. *flaccidus*. A – vegetative stem, B & C – perichaetial stems, D–F – leaves, G–H – leaf apices, I – mid dorsal lamina, J – insertion of leaf, K – trans-section of leaf showing *bryoides*-type costa, L – clusters of axillary gemmae, M – 2 gemmae. – *F. flaccidus* var. *mammillosus* Brugg.-Nann., var. nov. N – mammillae in side view, O – mid dorsal laminal cells. A from *Pócs* 9168/O; B from *Pócs* 9168/Q; C, H from type *F. ensifolius*; D, M from *Pobeguín* 306; E, K from *Müller* E194; F from *Dorr* 2752; G, I, from *Pócs* 9168/P; J from *MacFarlane* 811; L from *McFarlane* 168; N, O from type specimen of *F. flaccidus* var. *mammillosus*.



Fissidens splachnobryoides Broth. in K. Schum. & Lauterb., Fl. Schutzgeb. Südsee: **81**. 1900 – PROTOLOGUE: NEW GUINEA, Butaueng, Kaernbach (HOLOTYPE: H-BR).

REMARK. O'Shea (2006) incorrectly listed *F. minutus* var. *minutus* Thwaites & Mitt. in Mitt. (= *F. pallidinervis* Mitt.) as synonym of *F. flaccidus*.

5a. *F. flaccidus* var. *flaccidus* Fig. 5A–M

Stems without or with weak central strand, browning in KOH, hardly heterocaulous, mostly unbranched; vegetative stems 1.4–13.0 × 0.7–4.0 mm (sporophytic/perichaetial stems 5–10 × 2–3 mm); rhizoids basal and from stem cortex, pale to dark brown, smooth; axillary nodules not differentiated; leaves pinnately, less often frondosely arranged, distant, less often close, 5–15(–32) pairs, strongly crispate to hardly altered when dry, usually flattening in 5% KOH, elliptical, elliptico-lanceolate and elliptico-oblongate with acute to acute-acuminate apex ending in pointed colored cell, (0.6–)0.8–2.8 × 0.10–0.70 mm, 3.5–7 times as long as wide; limbidium on all laminae of all well-developed leaves, reaching leaf apex, often becoming weak towards leaf apex (Fig. 5G), occasionally confluent into ecostate mucro (Fig. 5H), confluent at apex of vaginant laminae, mostly reaching insertion of dorsal lamina, on vaginant lamina often becoming lax and indistinct proximally and reaching insertion or not, on mid dorsal laminae 8–21(–27) μm wide, 1–2-stratose, marginal; on mid vaginant lamina (11.0–) 14.5–27.0 (–50.0) μm, 1–2-stratose, near insertion marginal or weakly intramarginal; vaginant lamina 1/2–3/5 of leaf length, at base less wide than stem, frequently slightly decurrent (best visible with KOH; Fig. 5J), unistratose, ± closed; dorsal lamina narrow, straight to slightly rounded at insertion, reaching insertion, not decurrent; dorsal and apical lamina unistratose; costa ending 7–21 cells below leaf apex, in cross section of bryoides-type; mid dorsal laminal cells (18.5–)23.0–50.0(–65.0) × 8.0–25.5(–28.0) μm, smooth, plane to lowly convex; mid vaginant laminal cells oblong, 26.5–76.5(–80.0) × 7.0–24.0(–30.0) μm; juxta-costal cells of vaginant lamina 44.0–58.0(–72.0)

× 9.5–18.0 μm; axillary fascicles of short rhizoids bearing multicellular, uniseriate, hyaline to brown gemmae (Fig. 5L & M) often present.

Fertile parts, perigonia and perichaetia on separate plants, perigonial plants 2–3 mm long, hardly differentiated and easily overlooked, perigonia terminal, antheridia ± 250 μm long; perichaetia terminal on plants or, less often, on branches; perichaetial leaves longer than stem leaves (1.3–)1.7–3.0 mm long; archegonia 200–380 μm long; calyptra not seen. Sporophyte, seta 2–5 mm long, smooth, 1 per perichaetium; capsule erect, 0.50–0.65 × 0.35–0.45 mm with ca 32 columns of quadratic-oblong, exothecial cells with thickened corners; peristome scariousus-type (only basal part seen), tooth base 54.5–56.5 μm wide; operculum long-rostrate, 0.5 mm long; spores 12.5–18.0 μm diam., finely papillose.

Fissidens flaccidus var. *flaccidus* is characterized by its completely limbate leaves, (1–)2-stratose limbidia, smooth, large cells (in mid dorsal lamina, at least some 30 μm or more long), short costae ending 7–21 cells below the leaf apex, and (0.6–)0.8–2.8 mm long leaves. Multicellular gemmae on short, axillary rhizoids are often present. Archegonia are frequently present but sporophytes are rare. Variability. Typically the laminae are unistratose and the limbidia 1–2-stratose. Müller E 201 is a robust expression in which all laminae have unicellular bistratose dots and the limbidia are 2(–3)-stratose near the insertion of vaginant laminae. *Fissidens flaccidus* var. *flaccidus* is most likely to be confused with *F. grandifolius*, which differs by having inclined capsules, 1–4 setae per perichaetium, axillary perigonial buds and dwarf male plants at the base of perichaetial stems (terminal in *F. flaccidus*). The few sporophytic plants of *F. flaccidus* observed [type-specimen of *F. bocarangensis*; Stone (1988) as *F. maceratus*; Eddy (1988) as *F. atroviridis*] have 1 seta per perichaetium and erect capsules. Whereas sporophytic collections of the two species are easily distinguished, this is not true for vegetative collections. However, though both species have 1–2-stratose limbidia (in one and the same leaf), limbidia of *F. flaccidus* are essentially bistratose, whereas those of *F. grandifolius*

are more often unistratose. Furthermore, though their cell sizes overlap, cells of *F. grandifolius* are typically somewhat larger (mid dorsal laminal cells (28.0–)34.5–71.0(–77.0) × 12.5–37.0 μm versus (18.5–)23.0–50.0(–65.0) × 8.0–25.5(–28.0) μm in *F. flaccidus*); moreover, gemmae are common in *F. flaccidus* but have not been observed in *F. grandifolius*. The Namibian *F. capriviensis* Magill (not seen) is corticolous, has weaker limbidia and is smaller in all respects (Magill, 1981). The minute *F. minutifolius* has longer costae which reach the leaf apex. The South African species *F. wageri* Dixon and *F. splachnifolius* Hornsch. similarly have large cells and short costae, but are easily distinguished by their elimbate leaves (Magill 1981). Outside Africa, *F. flaccidus* resembles the Neotropical *F. dissitifolius* Sull. that differs by having shorter limbidia which end below the leaf apex and having smaller laminal cells [(13.9–)18.0–29.0(–36.0) × 9.0–16.0(–22.0) μm; Pursell 2007].

FISSIDENS FLACCIDUS, *F. MOLLIS* AND *F. MACERATUS*. *Fissidens mollis*, a species from the Neotropics, was considered conspecific with *F. flaccidus* by Bruggeman-Nannenga and Pursell (1995). During the present study it appeared that the African specimens, including the type specimen of *F. flaccidus*, differ essentially from Pursell's (2007) diagnosis by having thinner limbidia [(1–)2-stratose rather than 2–4-stratose] and shorter leaves (up to 2.8 mm versus up to 5.0 mm long). Moreover, whereas in the Neotropics the costae vary between ending as many as 18 cells below the leaf apex to percurrent (Pursell 2007), the costae of African collections always end well below the apex. Neotropical specimens agreeing with Pursell's diagnosis (long leaves and thick limbidia confluent at the apex) include the type specimen of *F. mollis*. Therefore this species is here reinstated. Both species have limbate leaves and large cells, and frequently have axillary, multicellular gemmae. *Fissidens flaccidus* s.str. is also known from the Neotropics (for an example see examined specimens). Its distribution in the Neotropics is unknown. *Fissidens maceratus* Mitt. was correctly subsumed under *F. flaccidus* (Pursell 1997).

SUBSTRATE AND HABIT. On rock (unspecified, limestone, travertine, walls) and soil. Once

collected on a termite mound. Growing in mats, tufts or scattered, often sparsely intermixed with other species and then easily overlooked.

HABITAT. mostly in forests [lowland rainforest (7×), riverine forest (6×), also in moist semi-deciduous rainforest, submontane rainforest, evergreen forest, mixed low-elevation hardwood forest, secondary low-elevation tropical hardwood forest with palms, and degraded, dry forest], also collected in Sudano-Guinean savanna, plantations and gardens. Frequently growing near water [stream banks, near waterfalls, irrigated limestone (travertine bank), man-made walls of ditches, swimming pools, etc.], occasionally submerged.

ELEVATION. 0–1800 m a.s.l. (based on African collections).

DISTRIBUTION. Widespread and common in Africa. Macaronesia (Cape Verde Islands); west tropical Africa (Ghana, Guinea, Ivory Coast, Niger, Nigeria, São Tomé, Sierra Leone); west-central tropical Africa (Central African Republic); east tropical Africa (Tanzania, Uganda); south tropical Africa [Angola (Müller 2015), Zambia, Zimbabwe (O'Shea 2006)]; southern Africa (South Africa) and western Indian Ocean (Comoro islands, Madagascar, Mauritius, Réunion, Rodriguez, Seychelles, Socotra; O'Shea 2006). Outside Africa known from Brazil, Asia, Australia and Oceania. New for Guinea and South Africa.

SPECIMENS EXAMINED: CAPE VERDE ISLANDS, Brava, an der Strasse zum Flughafen, *Lindlar 95* (BONN). – CENTRAL AFRICAN REPUBLIC, Bamingui-Bangoran, Parc de Manovo-Gounda-Saint Floris. Campement de Koumbala, 8°30'N, 21°13'E, *Prendergast 91* (PC, L); Bocaranga, sur la route Bocaranga-Kounnang près village Zölé, *Eckendorff s.n.* (L). – COMOROS, Ndzuani (Anjouan) Island, NW coast near Ouani, near Hotel Guinguette, *Pócs 9168/K, M, O, Q* (all EGR, L); collapsed Crater of Mt Pamouni on the S end of the island, E of M'rijou village, 12°20'21"S, 44°29'57"E, *Pócs, Magill & Rupf 9281/G* (EGR, L); Ngazidja (Grande Comore) Island, Ikoni Town on the West Coast below the volcanic cliffs at the fort hill, *Pócs 9148/B, p.p.* a few stems (EGR, L). – GHANA, Ashanti Region, Bobiri Forest reserve; 6.41°N 1.20°W, *Jouko 040001-64* with *F. ramulosus* (H, L). – GUINEA, Djallon-Timbo, *Pobeguïn 306* a few stems (L). – IVORY

COAST, Man, Cascade de Man 4 km W of Man; 7°25'N, 7°35'W. *F. Müller E 201* (L, DR); – MADAGASCAR, prov. Antananarivo, Antananarivo, Parc de Tsimbazaza, 18°55'S, 47°31'E, *L.J. Dorr 2752* (MO, L a few stems among *F. androgynus*); Mahamasina, ouest de Tanararive, Imerina, *Camboué s.n.* (PC). – MAURITIUS, Cave near Bassin Anglais, Curepipe. *Een M 443* (S, PC). – NIGER (type *F. flaccidus* (NY, BM)). – NIGERIA, W-Nigeria, Ijebu Prov., Ijebu-Ode, between Emuren and Imagbon, *McFarlane 168* (L); Lagos State, Epe, *McFarlane 278* (L). – RÉUNION, Bras de Cilaos, route N5 vers Cilaos, entre Ilet Furcy et Le Petit Serré, *Arts RÉU 10/16* (BR, L); la Côte aux Vents, Anse des Cascades 2 km NNE of Bois Blanc on E coast, 21°10'S, 55°50'E, *Bruggeman-Nannenga 11785* (L, REU). – SÃO TOME & PRINCIPE, São Tome Island, 6 km above Guadalupe near the village of Baoesperança, Lat/Long NAD 83: 00°20'19.5"N, 06°37'36.5"E., *Shevock et al. 50025* with *F. ramulosus* Mitt. (L); Island of Principe, at roça Belo Monte Resort, Lat/Long NAD 83: 01°41'07.2"N, 07°26'38.0"E., *Shevock et al. 49760* (L). – SEYCHELLES, Mahé, La Poudrière Road, *Norkett 16123* (BM, L); Path to le Niol road, *Norkett 18657* (BM), Beau Vallon, *Norkett 18677* (BM); Praslin, old track, *Norkett 18385* (BM). – SIERRA LEONE, Kangari Hills, Masati dry path in thick jungle (serpentine ground). 14 IV 1955. *V. Marmo s.n.* (H, L). – SOUTH AFRICA, Transvaal, Wolfspruit, Ofcolaco, *Brenan BM 3280, p.p.*, a few among *F. cf dealbatus* (E, L 1 stem). – RODRIGUEZ, Rivière Cascade Victoire bei Port Sud-Est, *F. Müller R743* (DR, L); Port Mathurin S, Cascade Pigeon, *F. Müller R 751* (DR, L). – TANZANIA, S. Foothills of Mt Meru, 17 km E of Arusha, along Malala Stream, *Pócs & Van Zanten 86125/G, H, p.p.* (EGR, L); E. Usambara Mts, Mtai Forest Reserve, *Pócs & Farkas 86246/H* (EGR, L); Mts Chemka, *Jones & Pócs 6382/F, p.p.* with *F. zollingeri* (EGR, PC); West Usambara Mts, above Mazinde village, *Pócs 8436/AE, p.p.* (EGR, L); Ulanga District, Mahenge Plateau, Mselezi Forest Reserve, *Pócs, Minja & Towo 87202/B* (EGR, L); West slopes of Nguru Mts, between Magunga and Maskati villages, *Kis & Pócs 9103/A* (EGR, L a few mixed with *F. megalotis* subsp. *helictocaulos*); Morogoro, on slopes behind University campus, *Pócs 8663/P, p.p.* (EGR, L mixed with other species); Southern Highlands, Mt Rungwe E of Rungwe village, *Pócs 6777/A, AE, AF, AJ* (PC, EGR, L). – UGANDA, Kabarole, Kibale Forest NP. Between Isunga and Dura River. 0°29'N, 30°21'E, *J.W. Bates U8630A, p.p.*, with *F. dealbatus* and *F. zollingeri* (E, L). – ZAMBIA, 13 km East of Lusaka, *Pócs & Kornás 6609/G, p.p.* (EGR).

BRAZIL, Rio de Janeiro State, Rio de Janeiro, Quinta da Boa Vista, *Vital 5847* (SP, L).

SELECTED ILLUSTRATIONS: Bruggeman-Nannenga (2006a: fig. 8), Bruggeman-Nannenga & Arts (2010: fig. 23), Eddy (1988: fig. 82, as *F. atroviridis*), Iwatsuki & Suzuki (1982: pl. V, as *F. splachnobryoides*), Li (1985: fig. 2a–j, as *F. splachnobryoides*), Potier de la Varde (1937: fig. 1, as *F. ensifolius*); Stone (1888: figs 1 & 2, as *F. maceratus*; type specimens of *F. maceratus* and *F. splachnobryoides*).

5b. *F. flaccidus* var. *mammillosus* Brugg.-Nann., var. nov. Fig. 5N & O

TYPE: SIERRA LEONE, Kambui Hills, Bambano, disused chrome mine, on wall of culvert in deep shade, 11 VIII 1966, *Harrington 588* (HOLOTYPE: BM, ISOTYPE: L).

DIAGNOSIS. The new variety differs from var. *flaccidus* by its mammillose (low-conical with inconspicuously thickened apical area) laminal cells. The mammillae are best observed in leaf folds or in surface view. In cross section they are hard to observe. The new variety resembles the smaller *F. lindbergii* Mitt. from Central and northern South America. Stone (1988) described and figured a similar-looking *Fissidens* from Queensland without naming it, but the mammillae in her figure do not have a thickened apex.

SUBSTRATE, HABIT AND HABITAT. In tuft on wall of culvert in deep shade; semiaquatic (covered by diatoms).

DISTRIBUTION. African endemic. Sierra Leone, known from the type locality only.

6. *Fissidens grandifolius* Broth. & P. de la Varde Fig. 6

in P. de la Varde, Bull. Soc. Bot. France 73: 57, f. 25, 1. 1926 – PROTOLOGUE: [GABON], sur terre argileuse. Rive gauche de la Ngounyé, entre Udendé et Labo, XII 1924, *Le Testu 5346* – LECTOTYPE (*nov.*) with label: Rive gauche de la Ngounyé entre Udendé et Labo, en forêt, sur argile, 29 10-bri 1924, *Le Testu 5346* (PV-PC!; ISOLECTOTYPE: H-BR).

Stem without or with weak central strand, mostly unbranched; vegetative stems, frondose to elongate-frondose, variable in size, (1.5–)2.5–5.0 × 1.0–4.0 mm, unbranched; sporophytic stems taller,

4–8 × 3–5 mm, with frondosely to pinnately arranged leaves; *rhizoids* basal and from stem cortex, brown to hyaline, smooth; *axillary nodules* not developed; *leaves* close, less often distant, 4–9 pairs, shriveled and crispate when dry, not well flattening in water, flattening in 5% KOH, elliptical, elliptico-lanceolate to elliptico-ob lanceolate with acute to acute-acuminate apex, ending in large pointed cell, limbate, in vegetative stems (1.0–)1.6–3.2 × 0.3–0.7 mm, 3.5–6.5(–7.5) times as long as wide; in sporophytic stems 3.3–3.9 × 0.5–0.6 mm, 5.5–7.8 times as long as wide; *limbidium* on all laminae of all well-developed leaves, confluent at leaf apex, often becoming weak towards leaf apex, confluent at apex of vaginant laminae, becoming weak towards insertion of vaginant lamina, reaching or ± reaching insertion of dorsal lamina; in mid dorsal lamina 11–32 µm wide, 1(–2)-stratose, marginal, on vaginant laminae (13–)16–43 µm wide, 1(–2)-stratose, marginal to weakly intramarginal; *vaginant lamina* 1/2–3/5 of leaf length, at base narrower than stem, closed to ± closed, unistratose; *dorsal lamina* narrow, straight to slightly rounded at base, reaching insertion, not decurrent, *dorsal* and *apical lamina* unistratose; *costa* ending 10–26 cells below leaf apex, weak in upper part, in lower leaves not always exceeding length of vaginant laminae, in cross section *bryoides*-type; *mid dorsal laminal cells* hexagonal, large, variable (28.0–)34.5–71.0(–77.0) × 12.5–37.0 µm, smooth, plane to inconspicuously convex; *mid vaginant laminal cells* (30.0–)40.0–110.0 × 10.0–32.0 µm, smooth, plane; *basal vaginant laminal cells* oblong, 69.0–114.0 × 12.0–29.0 µm; *gemmae* not observed.

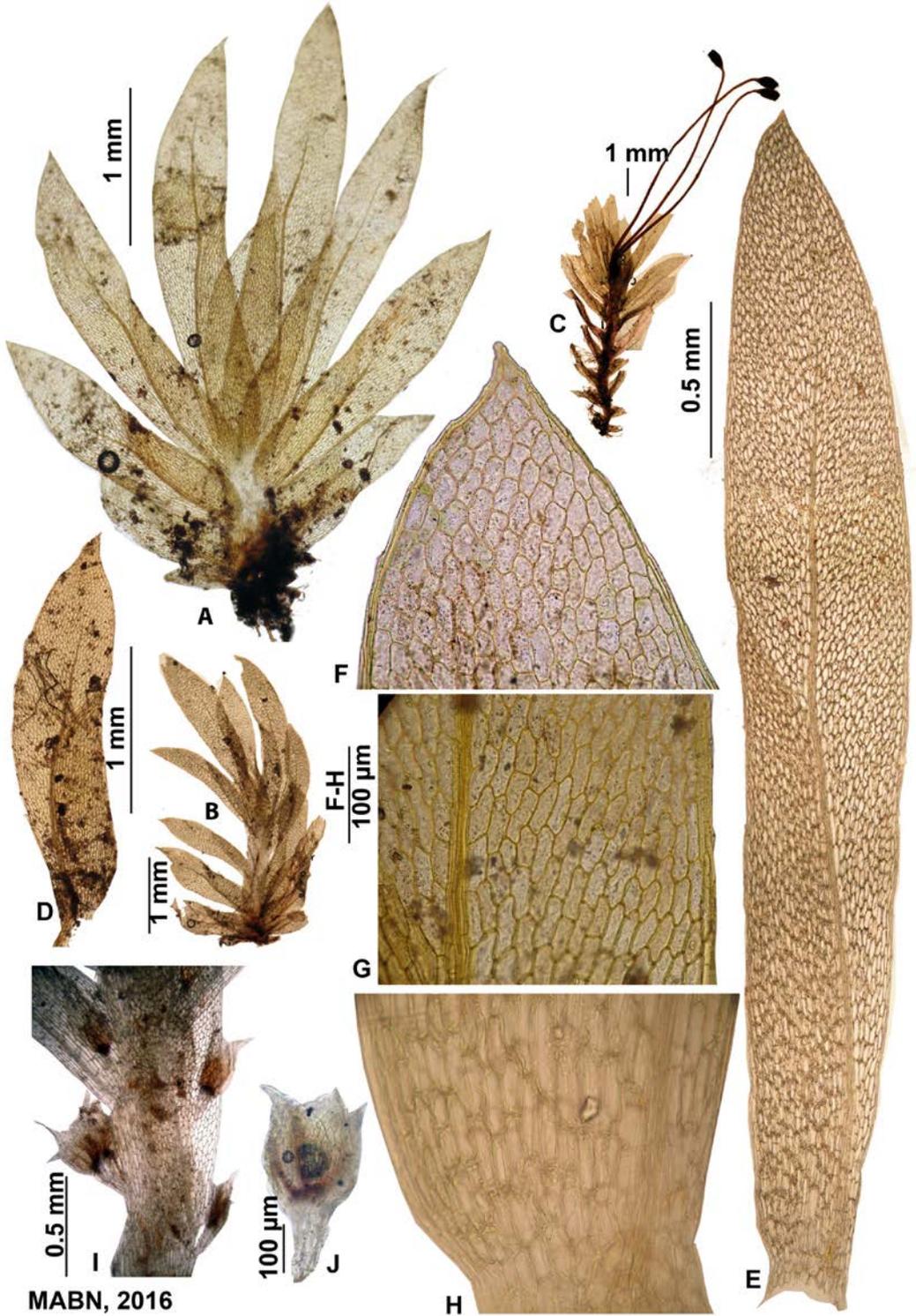
Fertile parts, *perigonia* and *perichaetia* on same stem, *perigonia* frequent, terminal on axillary, stalked (Fig. 6 I, J), budlike, 0.3–0.7 mm long (including stalk) branches and on tiny plants at base of perichaetial plants; *antheridia* 150–180 µm long; *perichaetia* terminal, *perichaetial leaves* (1.8–)3.3–4.0 mm long, *archegonia* 150–300 µm long; *calyptra* 0.7 mm long, smooth. *Sporophyte*, *seta* 5–10 mm long, smooth, 1–4 per perichaetium; *capsule* oblique, small, 0.4–0.6 × 0.2–0.35 mm, with ca 32 columns of exothecial cells with strongly thickened corners; *peristome* scarious-type, tooth

base 50–54 µm wide; *operculum* 0.5 mm long (immature); *spores* subglobose, 9.0–14.5 µm long, faintly papillose.

Fissidens grandifolius is characterized by its limbate leaves, large cells, and 1–4 setae per perichaetium, inclined capsules, stalked axillary perigonial buds and dwarf male plants at the base of perichaetial stems. The limbidia are uni- to bi-stratose. There are typically ± 32 columns of exothecial cells, but sometimes more, due to extra cell divisions on one side of the capsule. The species is highly variable in size and cell size. Sporophytic stems tend to be taller and have longer leaves than vegetative ones, and more often have pinnately arranged leaves. A small expression was identified by Shevock *et al.* (2013) as *F. palmatus*. Based on the available material it seemed impossible to demonstrate a sharp distinction between small and large expressions. *Fissidens grandifolius* can also be confused with *F. flaccidus*. For differences see under the latter.

FISSIDENS PALMATUS AND *F. GRANDIFOLIUS*.

Both *F. palmatus* and the African *F. grandifolius* display two expressions. As indicated above, the African *F. grandifolius* has large and small expressions which are not sharply separated. Pursell's (2007) concept of Neotropical *F. palmatus* is broad and includes *F. palmatus* Hedw. *s.str.* with percurrent costae as well as expressions with costae that end well below the apex. The latter used to be known as *F. reticulosus* (Müll. Hal.) A. Jaeger in the Neotropics. Small non-sporophytic expressions of the African *F. grandifolius* resemble the *F. reticulosus* expression; indeed, one specimen was reported as *F. palmatus* by Shevock *et al.* (2013). The two species may prove to be conspecific in future but the situation is complex. It seems significant that *F. palmatus s.str.* is unknown from Africa, whereas the large expression of *F. grandifolius* is restricted to West Africa. In contrast to *F. palmatus* in the Neotropics, *F. grandifolius* has sporophytic plants that tend to be taller and to have longer leaves, (3.3–3.9 mm versus up to 2.8 mm long) and more setae (1–4 per perichaetium, versus 1–2). There may be differences in cell size as well, but this could not be established as it is not clear



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which cells were measured for the Neotropical monograph. More material from both continents is needed for taxonomic decisions. Pending this, the collection from São Tomé earlier reported as *F. palmatus* (Shevock *et al.* 2013) is here tentatively included in *F. grandifolius*.

SUBSTRATE AND HABIT. On soil and wood. Growing in mats or scattered among other mosses.

HABITAT. Often not indicated, forest including a riverine forest.

ELEVATION. Sea level to 230 m a.s.l.

DISTRIBUTION. Endemic to West Africa. Known from west tropical Africa (Guinea; O'Shea 2006; Liberia, Nigeria, Sierra Leone, Togo; O'Shea 2006) and west central tropical Africa (Cameroon, Central African Republic, Gabon, São Tomé). Not common. New for Cameroon.

Also reported from Ivory Coast (Müller & Schäfer-Verwimp 1999) but this specimen appeared to be *F. flaccidus*.

SELECTED SPECIMENS EXAMINED: CAMEROON, left bank of the Kèlè River, 29 km N of Eséka, *Leeuwenberg 6019 B* (L). – CENTRAL AFRICAN REPUBLIC [Oubangui], route Riv. Ngurro, près Wamiré, 60 km N of Bambari, 27 7-bri 1927, *Tisserant 400* (PC-PV); Mbaïki, Station de la Maboké, *Faurel 10* (PC, L). – GABON, type specimen. – LIBERIA, Bakratown, *Linder 853* (PC). – NIGERIA, Western Nigeria, Ijebu Prov., Omo (Shasha) Forest Reserve, *Richards 3361* (L); Ijebu Prov., Ilugun/Alaro, *McFarlane 513* (L); Ondo Prov., Oluwa Forest Reserve, *McFarlane 250* (L); Benin Prov., Okomu Forest Reserve, *Brenan (R 3886)* (PC-PV). – SÃO TOMÉ, Island of Principe, Along trail towards the SW base of Pico Pagagaio from Rosa Santa Trinitade towards Rio Banzu, 01°36'45.6"N, 07°23'14.7"E, *Shevock 34650* (L). – SIERRA LEONE, Freetown, Mount Aureol, Fourah Bay College Botany Garden, *Harrington 700* (BM, with *F. submarginatus*).

ILLUSTRATIONS: Potier de la Varde (1926: f. 25, 1; 1929: fig. I, 1 & II, 1a, b; 1936: f. 4, 1 – same figure as 1926).



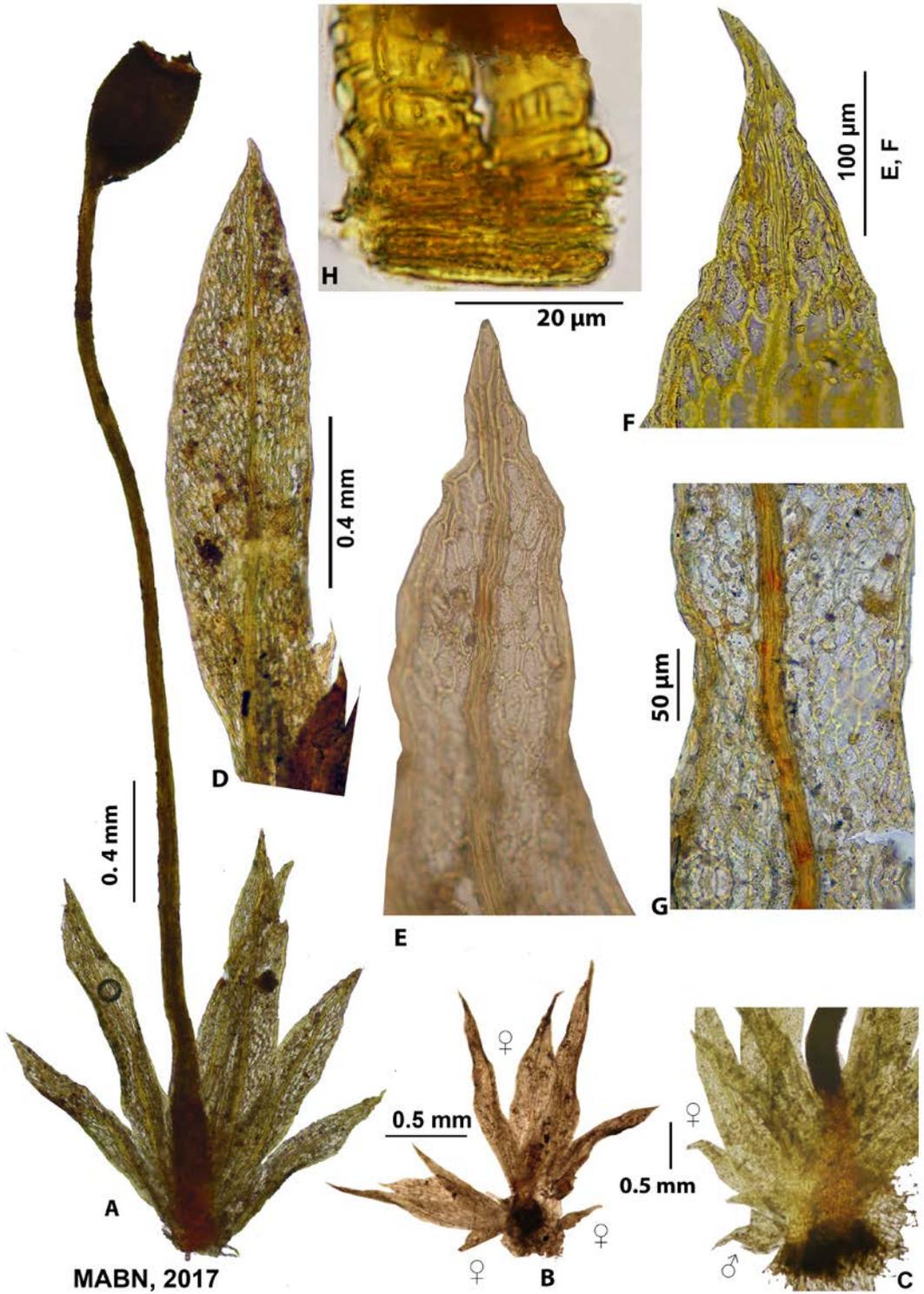
Fig. 6. *Fissidens grandifolius* Broth. & P. de la Varde. A – vegetative stem with frondosely arranged leaves, B – stem with pinnately arranged leaves, C – sporophytic stem, D & E – leaves, F – leaf apex, G – mid dorsal lamina, H – insertion of leaf, I – stalked axillary perigonia (leaves removed), J – stalked axillary perigonium (detail). A, B, D from *McFarlane 513*; C, G, J from *Tisserant 400*; F from type specimen; E, H, I from *Faurel 10*.

7. *Fissidens minutifolius* Broth. & P. de la Varde Fig. 7

in P. de la Varde. Bull. Soc. Bot. France **73**: 59, f. 25–2. 1926 – PROTOLOGUE: [GABON], sur terre argileuse. Rive gauche de la Nounyé, entre Udendé et Labo, intimement mêlé au precedent [*F. grandifolius*], mais beaucoup plus rare, XII 1924, *Le Testu 5347* – LECTOTYPE (*nov.*) with label: sur argile, en forêt, rive g. de la Nyounyé, entre Undendé et Labo (intimement mêlé au precedent), 29 X-bri 1914, *Le Testu 5347* (PC-PV PC0106510!, ISOLECTOTYPES: H-BR!; PC PC106509!).

Stem frondose, unbranched, 1.0–1.7 × 1.5 mm (only fertile plants seen); *rhizoids* numerous, basal and from stem cortex, hyaline, ± smooth; *leaves* close, 3–8 pairs, crispate when dry (impossible to revive without KOH), elliptico-lanceolate to oblanceolate, acute-acuminate ending in pointed cell, 1.0–1.2 × 0.2–0.3 mm, 4–6 times as long as wide, limbate; *limbidium* on all laminae of all well-developed leaves, reaching leaf apex and fusing with costa, confluent at apex of vaginant laminae, becoming weak towards insertion of vaginant laminae, ± reaching insertion of dorsal lamina, in mid dorsal laminae 18 µm wide, 1–2-stratose (estimated), marginal; on vaginant lamina 22.5 µm wide, lax, marginal; *vaginant lamina* 1/3–2/3 of leaf length, at base about as wide as stem, almost closed; *dorsal lamina* narrowing towards insertion, reaching insertion, not decurrent; *costa* ex- to percurrent; *mid dorsal laminal cells* hexagonal, 30.0–52.5 × 15.0–18.0 µm, smooth; *mid vaginant laminal cells* up to 60 µm long. No *gemmae* seen.

Fertile parts, *perigonia* numerous, terminal on short, ± 0.35 mm long, bud-shaped plants at base of female plants, *antheridia* not seen; *perichaetia* terminal, *perichaetial leaves* 1.5–1.7 × 1.5 mm, *archegonia* not seen; *calyptra* not seen. *Sporophyte*, *seta* 3.5–5.0 mm long, smooth, 1 per perichaetium; *capsule* erect to slightly inclined, 0.5 × 0.45 mm, with ca 32 columns of oblong exothelial cells with strongly thickened corners; *peristome* scarious-type, *teeth* at base 33.0–40.5 µm wide; *operculum*



long-rostrate, 0.6 mm long; spores 9–12 μm long, smooth.

This tiny species is characterized by its frondose stems, over 30 μm long laminal cells, completely limbate leaves, per- to excurrent costae, a scariosus-type peristome and \pm 32 columns of exothecial cells on the capsule. It can be confused with *F. flaccidus*, which is larger and has shorter costae, and with *F. zollingeri*, which has half as large mid dorsal laminal cells. *Fissidens minutifolius* further resembles the subgenus *Fissidens* species *F. aciphyllus* Dixon and *F. magnicellulatus* Brugg.-Nann., which both differ in having a higher number of exothecial cells (40 columns or more) and a *bryoides*-type peristome. *F. aciphyllus* Dixon and *F. magnicellulatus* Brugg.-Nann. are very similar; and it cannot be ruled out that *F. magnicellulatus* will prove to be a perichaetial expression, with relatively large cells, of *F. aciphyllus*.

REMARKS. Due to the paucity of the available material, no cross sections were made. Features that can only be observed when leaves are removed (e.g., axillary nodules) are also excluded from the description.

HABITAT, SUBSTRATE AND HABIT. In forest on soil. Sparsely scattered among *F. grandifolius* plants, covered with dirt.

ELEVATION. Not indicated.

DISTRIBUTION. A rare species, known from west-central tropical Africa (Gabon). Further reported from west tropical Africa (Togo and Nigeria O'Shea 2006).

SPECIMENS EXAMINED: GABON, types (PC-PV, H-BR); Pétsyalango, *Le Testu s.n.* (PC-0705948, PC-PV).

ILLUSTRATION: Potier de la Varde (1926: f. 25, 2; 1936: f. 4, 2 – same figure as 1926).

8. *Fissidens nigerianus* Bizot ex Brugg.-Nann.

Fig. 8

J. Hattori Bot. Lab. **81**: 170, 1997; Bizot, Rev. Bryol. Lichénol. **40**: 101, pl. 1A, 1–2, 1974 – PROTOLOGUE/

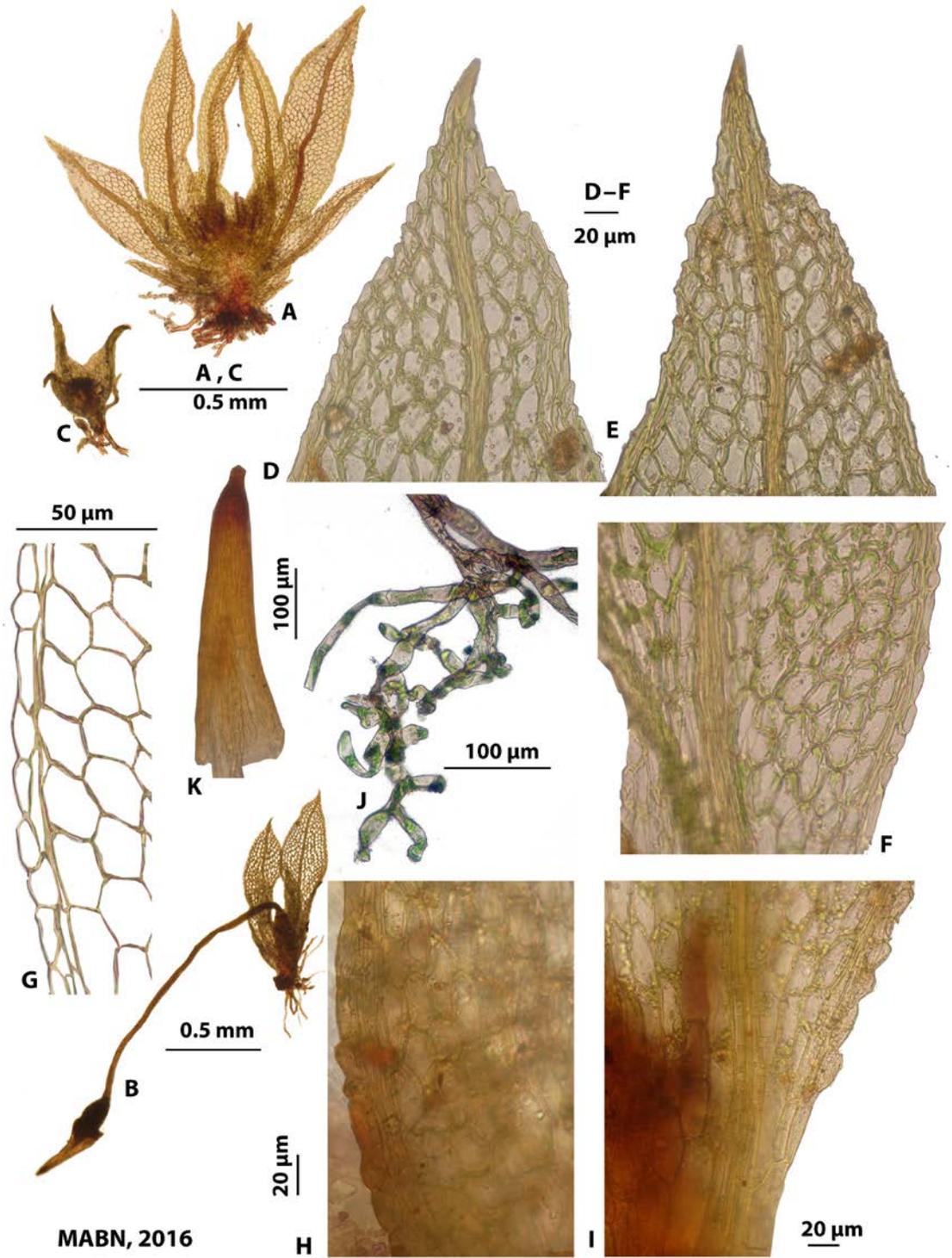
HOLOTYPE label: NIGERIA, on worm casts in deep shade in forest, 1700 ft, Garki Kurmi Abuja Prov., Minna, Jones 886 (PC!).

No vegetative stems observed, probably heterocaulous, perichaetial stems 0.60–1.50 \times 0.45–1.50 mm, unbranched, frondose; persistent filamentous protonema well developed; rhizoids basal, hyaline to pale brown, smooth; axillary nodules weakly differentiated; leaves imbricate, 2–5 pairs, slightly crispate when dry, lanceolate, acuminate-mucronate (perichaetial leaves), 0.7–1.6 \times 0.15–0.3 mm, 3.5 times as long as wide, margins limbate, serrate where elimbate, limbidia frequently intramarginal on all laminae, extension and occurrence variable, typically present on all laminae, in perichaetial leaves not or \pm confluent at apex of vaginant laminae and \pm reaching leaf apex or ending well below it, in lower leaves less developed; often becoming ill-defined towards insertion of vaginant lamina; reaching insertion of dorsal lamina or ending well above it, in mid dorsal lamina 19 μm wide, unistratose (estimated), marginal or intramarginal; on vaginant laminae 16–24 μm wide, unistratose (estimated), marginal or intramarginal by one row of cells; vaginant lamina 3/5 of leaf length, at base narrower than stem, unistratose, slightly open; dorsal lamina tapering towards insertion, in perichaetial leaves reaching insertion to ending well above, not decurrent; dorsal and apical lamina unistratose; costa long excurrent in perichaetial leaves, per- to long excurrent in lower leaves; mid dorsal laminal cells hexagonal, large, 25.0–38.5 \times 13.0–21.5 μm , smooth, plane; mid vaginant laminal cells oblong-hexagonal, 25.5–70.5 \times 11.5–24.0 μm , smooth, plane. Gemmae 150 \times 25 μm , monoseriate, growing from persistent protonema.

Fertile parts, perigonia terminal on small, \pm 0.4 mm tall plants at base of perichaetial plants; antheridia \pm 100 μm long; perichaetia terminal; perichaetial leaves 0.7–1.2 mm long; archegonia \pm 180 μm long; calyptra 0.4 mm long, smooth.



Fig. 7. *Fissidens minutifolius* Broth. & P. de la Varde. A – sporophytic stem, B – 3 young perichaetial stems, C – base of sporophytic stem with attached dwarf perichaetial and perigonial stems, D – leaf, E & F – leaf apices, G – mid leaf, H – bifurcation of peristome tooth showing upper part of basal undivided part and basal part of two filaments. All from lectotype.



Sporophyte, seta 3.0–4.5 mm long, smooth, 1–2 per perichaetium; *capsule* ± erect, 0.40–0.50 × 0.25–0.35 mm, with *ca* 32 columns of oblong exothecial cells; *peristome* details not observed, tooth base 41.5 µm wide; *operculum* rostrate, 0.45 mm long; *spores* subglobose, 11–13 µm long, smooth.

This species is known from the type collection only. It is characterized by its limbate leaves with limbidia that are often intramarginal on all laminae, per- to excurrent costae, large cells (in mid dorsal lamina 25.0–38.5 × 13.0–21.5 µm), and persistent protonemata with gemmae. Though intramarginal limbidia on vaginant lamina are fairly common in the Fissidentaceae, intramarginal limbidia on the dorsal and apical lamina are much rarer and thus have diagnostic value. *Fissidens minutifolius* resembles this species in having frondose stems and per- to excurrent costae, but is larger and has better-developed marginal limbidia. *Fissidens nigerianus* further resembles the Asian-Polynesian *F. bogoriensis* M. Fleisch. Both share short, frondose plants, limbidia that are often intramarginal on all laminae, large laminal cells, short perigonal plants at the base of perichaetial plants, and small spores [8–14(–16) µm in *F. bogoriensis*]. *Fissidens nigerianus* can be separated from this species by its per- to long excurrent costae and well developed persistent protonemata. It also resembles the Neotropical *F. ornatus* Herzog but that species is elimbate.

REMARK. Due to the paucity of the available material, no cross sections were made.

SUBSTRATE AND HABITAT. Scattered on worm casts in deep shade in forest.

ELEVATION. 520 m a.s.l.

DISTRIBUTION. Nigeria, known from the type locality only.

SPECIMEN EXAMINED: type specimen.

ILLUSTRATION: Bizot (1974: pl. 1A, 1–2).

9. *Fissidens zollingeri* Mont.

Fig. 9

Ann. Sci. Nat. Bot., sér. 3, 4: 114. 1845 – PROTOLOGUE: INDONESIA, Java, in terra turfosa, *Zollinger 1604* (HOLOTYPE PC; ISOTYPE PC).

Fissidens vogelianus Mitt., Trans. Linn. Soc. London 23: 54, t. 5, f. 10. 1860 – PROTOLOGUE: River Niger, *Vogel* – type label: Africa, Niger-expedition, *Vogel s.n.* (HOLOTYPE: NY, ISOTYPE PC); ISOTYPE: BM as *Vogel 3*.

Fissidens dubyanus A. Jaeger, Ber. Thätigk. St. Gallischen Naturwiss. Ges. 1874–1875: 107. 1876, new name for *F. macrophyllus* Welw. & Duby in Duby, Mém. Soc. Phys. Genève 21: 223, t. 3, f. 3 a–e. 1870, *hom. illeg.* – PROTOLOGUE: ANGOLA, ad latera viarum cavarum in locis valde umbrosis inter Bango-Aquitamba et Bamba praesidii Golungo alto 1000–2000 ped. altit. in regno Angolensi, *Welwitsch s.n.* – LECTOTYPE (designated by Bruggeman-Nannenga 1997) with label: *Welwitsch, Iter Angolense 136* (BM!); ISOLECTOTYPE (PC!).

Fissidens obsoletidens Müll. Hal. in Besch., Ann. Sci. Nat. Bot., sér. 6, 9: 332. 1880 – PROTOLOGUE: [NOS-SI-BE], talus couverts sur bords de la mer, au-dessus du plateau de Helville, janvier 1850, *Boivin* (herb. Mus. Par.) and: Nossi-Bé, Nossi-Comba et Antourtour, sur les pierres, VIII 1879, *Marie* – LECTOTYPE (designated by Bruggeman-Nannenga 1997) with label: Nossi-bé, talus couverts des bords de la mer au-dessus du plateau de Helville, I 1850, *Boivin s.n.* (PC!).

Fissidens platybryoides Müll. Hal., Flora 69: 505. 1886 – PROTOLOGUE: [NIGERIA], Africa occid. Tropica, Old Calabar, in territorio fluminis Niger, in terra, 10 XI 1884, *Mönkemeyer s.n.* – LECTOTYPE (designated by Bruggeman-Nannenga 1997) with label: Niger Gebiet, Alt Calabar, in terra, 11 (sic!) X 1885 (sic!), *Mönkemeyer s.n.* (H-BR); ISOLECTOTYPE labeled: Afr. Occ. Old Calabar, in terra, 10 1885, *Moenkemeyer s.n.* (PC).

Fissidens tenuisetus Cardot, Rev. Bryol. 35: 64, 1908 – PROTOLOGUE: DEMOCRATIC REPUBLIC OF THE CONGO, [Congo belge], Kisantu, sur la terre et le bois pourri, *H. Vanderyst* (sic!), 1906 – TYPE label: Congo belge, Kisantu, 1906, *Vanderijst* (sic!) *s.n.* (HOLOTYPE: PC!; ISOTYPE: H-BR).

Fissidens brachycaulon Broth. in Mildbr., Wiss. Ergebn. Deut. Zentr.-Afr. Exped., Bot. 2: 143, t. 11, f. 7. 1910 – PROTOLOGUE/TYP label: [TANZANIA], Bukoba-

←
Fig. 8. *Fissidens nigerianus* Bizot ex Brugg.-Nann. A – archegonial stem, B – sporophytic stem, C – perigonal stem, D – apex of stem leaf, E – apex of perichaetial leaf, F – mid leaf, G – mid dorsal lamina with intramarginal limbidium, H – margin of vaginant lamina, I – insertion of dorsal lamina, J – persistent protonema, K – calyptra. All from type specimen.

Bezirk, Buddu-Wald, auf lehmigen Waldboden, VI 1907, *Mildbraed 243* (HOLOTYPE: H-BR).

Fissidens herpetineuron Broth. & P. de la Varde in P. de la Varde, Arch. Bot. Bull. Mens. 3: 2, f. I, 2 and f. II, 2. 1929 – PROTOLOGUE: CENTRAL AFRICAN REPUBLIC [Oubangui], sur terre, marais près d'Ippy, *Tisserant s.n.* – LECTOTYPE (designated by Bruggeman-Nannenga 1997) with label: Oubangui, sur terre, marais près d'Ippy, 19 VI 1928, *Tisserant 473* (PV 3073) (PC-PV).

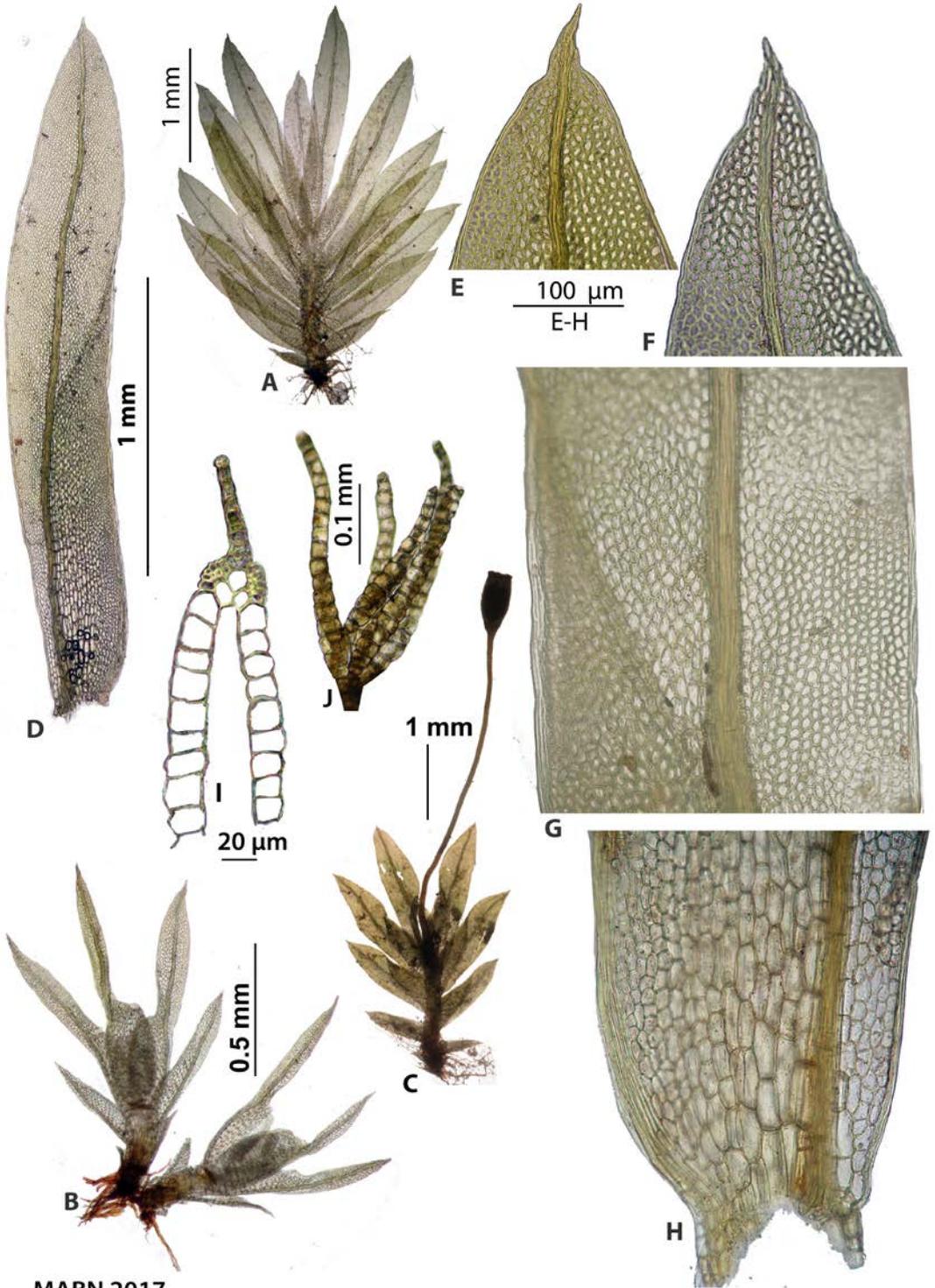
Stem without or with weak central strand, hardly heterocaulous, unbranched to frequently branched, $2.4\text{--}5.0 \times 1.7\text{--}3.5$ mm; *rhizoids* basal and from basal stem cortex, brown, smooth; *axillary nodules* differentiated or not; *leaves* frondosely to pinnately arranged, crowded to distant, 3–12 pairs, crispate when dry, elliptical to elliptic-lanceolate or obovate-lanceolate with acute to acute-acuminate to mucronate apex, $(1.1\text{--})1.5\text{--}2.5\text{--}(3.0) \times 0.2\text{--}0.5$ mm, 3.0–6.0(–7.5) times as long as wide, limbate on all laminae of all leaves; *limbidia* reaching leaf apex, often becoming indistinct at apex or sometimes ending below it, confluent at junction of vaginant laminae, reaching insertion of vaginant lamina, reaching insertion of dorsal lamina or ending slightly above, in mid dorsal lamina $6\text{--}19$ μm wide, (1–)2-stratose; on vaginant laminae $15\text{--}40$ μm wide, 1–2(–3)-stratose, marginal to weakly intramarginal near insertion; *vaginant laminae* $1/5\text{--}2/3$ of leaf length, at base narrower than stem, closed, unistratose; *dorsal laminae* tapering below, straight to slightly rounded at base, reaching insertion or not, not decurrent; *dorsal* and *apical lamina* unistratose; *costa* percurrent to long-excurrent, *bryoides*-type; *mid dorsal laminal cells* hexagonal, variable in size $(6.5\text{--})11.0\text{--}27.0 \times 4.5\text{--}14.5\text{--}(19.5)$ μm , smooth, plane; *mid vaginant laminal cells* hexagonal to oblong, $(10.0\text{--})14.0\text{--}36.5 \times (5.0\text{--})9.0\text{--}16.0\text{--}(18.5)$ μm ; *basal vaginant laminal cells* oblong, $(18\text{--})34.5\text{--}81\text{--}(94.5) \times (6.5\text{--})10.5\text{--}24.5\text{--}(31)$ μm . *Gemmae* on short axillary rhizoids, uniseriate multicellular, once in terminal cluster on short stalks, $0.6\text{--}0.7 \times 0.1$ mm, rare.

Fertile parts. *Perigonia* and *perichaetia* on separate stems or on same stem, *synoicia* rare; *perigonia* terminal on 0.85–3.00 mm long stems, also on budlike, 0.6–1.1 mm long branches; *antheridia* 120–300 μm long; *perichaetia* terminal on stems and/or branches; *perichaetial leaves* slightly longer than stem leaves, 1.1–3.0 mm long; *archegonia* 200–400 μm long; *calyptra* 0.6–0.8 mm long, smooth. *Sporophyte*, *seta* 3.5–6.5 mm long, smooth, 1–2 per perichaetium; *capsule* erect to suberect, $0.40\text{--}0.90 \times 0.20\text{--}0.45$ mm, with *ca* 32, oblong or quadrate-oblong exothecial cells with thin walls and thickened corners; *peristome* scariousus-type, tooth base 33–53 μm wide; *operculum* rostrate, 0.55–0.80 mm long; *spores* $(7.0\text{--})9.0\text{--}14.5 \times (5.0\text{--})8.0\text{--}12.5$ μm , papillose to smooth.

Fissidens zollingeri is a heterogeneous species. Features common to all specimens are the limbidia on all laminae of all leaves, per- to excurrent costa, smooth, medium-sized laminal cells, and when sporophytes are present \pm 32 columns of exothecial cells around the capsule. The combination of this last character with limbate leaves and smooth, medium-sized cells is rare in the *Fissidentaceae*. Stems can be short and frondose with imbricate leaves, but collections with longer stems and pinnately arranged leaves are not rare. The juxta-costal cells of the vaginant laminae are typically large and clear (usually easily visible at $5\times$ magnification) and conspicuously inflated (Fig. 9D, H, I); hyaline axillary nodules are present or not. In the majority of specimens the limbidia reach the leaf apex and the insertion of the dorsal lamina, but in a fair number the limbidia become weak towards the leaf apex or end below it and/or end above the insertion of the dorsal lamina. Mid dorsal laminal cells vary in size at $(6.5\text{--})11.0\text{--}27.0 \times 4.5\text{--}14.5\text{--}(19.5)$ μm . Cell size differs greatly between specimens. In most specimens the mid dorsal laminal cells are \pm 16 μm long, but collections with smaller or larger cells (though always less than 30 μm) are regularly found. Specimens with small cells



Fig. 9. *Fissidens zollingeri* Mont. A – vegetative stem, B – 2 perigonial stems, C – sporophytic stem, D – leaf, E & F – leaf apices, G – mid leaf, H – leaf insertion, I – cross section of vaginant lamina with inflated cells, J – cluster of gemmae. A, F, H from *Richards 3840*; B, D–E, G, I from *Pócs 97120*; C from *Müller E 209*; J from *Kovas s.n.*



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include, for example, *Pócs 8436/AA* (mid dorsal laminal cells $6.5\text{--}11.5 \times 5.0\text{--}8.0 \mu\text{m}$) and *Dilg 1-3* [mid dorsal laminal cells $(6.0\text{--})9.0\text{--}11.5\text{--}(12.5) \times (5.0\text{--})6.0\text{--}8.5\text{--}(10.0) \mu\text{m}$]. Larger plants tend to have larger cells and – rather surprisingly, because typically large plants have better-developed limbata – more often have elimbate leaf apices, but this correlation is weak. The African *F. minutifolius* resembles frondose specimens of *F. zollingeri* in having limbate leaves, per- to excurrent costae and smooth laminal cells, but it has larger, $30.0\text{--}52.5 \times 15.0\text{--}18.0 \mu\text{m}$, mid dorsal laminal cells. *Fissidens zollingeri* can further be confused with *F. bififormis* Mitt. (= *F. angustifolius* Sull.), which is distinct with its mammillose laminal cells, and with species of subgenus *Fissidens*. With the latter it shares limbate leaves and smooth laminal cells, but species of subgenus *Fissidens* have sporophytes with 40 or more columns of exothecial cells around the theca, and also *bryoides*-type peristomes. In tropical Africa the subgenus *Fissidens* species with which *F. zollingeri* is most likely to be confused is *F. curvatus* Hornsch. Apart from the differences in sporophytic characters, this species is distinct for being heterocaulous (that is, having vegetative stems that differ markedly from shorter perichaetial ones).

SUBSTRATE AND HABIT. Most often on soil, frequently on rocks, rarely on wood (twigs, wood and bark) or termite mounds. Growing in mats, small tufts, or scattered.

HABITAT. On shady road edges and stream banks in forests (lowland and montane rainforests, once in semi-deciduous forest, once in riverine forest), less often in savannas or cultivated areas (plantations, gardens, etc.). Frequently found in wet places (near rivers, springs and cascades), occasionally submerged.

ELEVATION. In Africa 0–1750(–2100) m a.s.l.; in the Neotropics up to 3360 m a.s.l. (Pursell 2007).

DISTRIBUTION. Common and widespread. African distribution: west tropical Africa (Benin, Gambia, Ghana, Guinea, Ivory Coast, Nigeria, São Tomé and Príncipe); west-central tropical Africa (Cameroon, Democratic Republic of the Congo,

Central African Republic, Gabon); east tropical Africa (Kenya; Kis 1985; Tanzania, Uganda); south tropical Africa (Angola, Zambia; Kis 1985) and western Indian Ocean (Comoro islands, Madagascar, Réunion, Rodriguez, Seychelles). New for Ghana. Outside Africa known from the Neotropics, Asia and Oceania.

SELECTED SPECIMENS EXAMINED: ANGOLA, type of *Fissidens dubyanus* (BM, PC). – BENIN, Forêt classée de la Lama, $6^{\circ}55'\text{--}7^{\circ}\text{N}$, $2^{\circ}04'\text{--}2^{\circ}12'\text{E}$., *Dilg 1.3* (L). – CAMEROON, Southwest, slopes of Mount Koupe, $4^{\circ}50'\text{N}$, $0^{\circ}43'\text{E}$, *Magill & Crosby 9063* (MO); Johan Albrecht Höhe, *Standt 698* (H-BR); Bipinde, Urwaldgebiet, *Zenker 2614* (H-BR, L); Yaoundé, Université-Yaoundé, *Frahm MD100* (L, BONN); Div. Kumba, Bopo, S. Bakundu Forest Reserve, *Richards 4034* (PC-PV, L). – CENTRAL AFRICAN REPUBLIC, Berberati, *Tisserant M 826* (PC-PV); type of *F. herpetineuron* (PC-PV); – COMORES, Ndzuani (Anjouan), NW coast near Quani, *Pócs 9168/Q*, p.p., with *F. flaccidus* (EGR, L). – DEMOCRATIC REPUBLIC OF THE CONGO, Prov. Kivu, Pinga, $1^{\circ}0'\text{S}$. Br; $28^{\circ}43'\text{Ö}$. L, *F. Müller Z 211* (L, DR); $1^{\circ}0'\text{S}$. Br; $28^{\circ}43'\text{Ö}$. L., *F. Müller Z 212* and *Z.222* (both DR, L); $1^{\circ}0'\text{S}$. Br; $28^{\circ}43'\text{Ö}$. L, *F. Müller Z 228* (DRE, L); Congo belge, *LeRoy 36A* (PC-PV). – EQUATORIAL GUINEA, BIKO, $3^{\circ}21'\text{N}$, $8^{\circ}42'\text{E}$, *F. Müller B 744*, p.p., with *F. crispulus*, *asplenioides* and *androgynus* (DR, L). – GABON, Mougoumou Poubi, *Le Testu 3991* (PC-PV, L); Moyandi, *Le Testu 2220*, p.p. (H-BR). – THE GAMBIA, Kombo East District, Pirang, ca 1 km SE of Bonto. $13^{\circ}16'\text{N}$, $16^{\circ}3'\text{W}$, *Perry 9858* (NMW, L); Kombo North District, Abuko Nature Forest Reserve, $13^{\circ}23'\text{N}$, $16^{\circ}38'\text{W}$, *Perry 9852* (NMW, L). – GHANA, Kumasi, *Kovács s.n.* (PC, a few stems in L). – GUINEA [Guinea gall.], Pita, *Pobeguín 1823* (H-BR); Djallon-Timbo, *Pobeguín 306*, p.p. mixed with *F. flaccidus* (L); 2 km westlich der Grenze zur République Côte d'Ivoire, *Porembski 202* (BONN, L). – IVORY COAST, Umgebung der Ortschaft Gouedie (= Gouele), 24 km NW of Man. $7^{\circ}32'\text{N}$, $7^{\circ}45'\text{W}$, *F. Müller E193* (DR, L); 30 km südlich von Séguéta, Granitinselberg (Nr IB22), *Porembski 297* (BONN, L); Ferkessedougou, im Stadtbereich beim Hotel Auberge de la Reserve, $9^{\circ}35'\text{N}$, $5^{\circ}12'\text{W}$, *F. Müller E 209* (DR, L). – MADAGASCAR, Nossi-bé, lectotype *F. obsoletidens* (PC); Nossi-bé, *Marie s.n.* (PC, H-BR); Nossi-Comba, *Marie s.n.* (PC). – NIGERIA, Oyo State, Ibadan, *McFarlane 140*, 227, 465 (all: L, private herbarium McFarlane); Ogun [Ijebu] State, Ijebu-Ode, *McFarlane 94*, 99, 136, 171, 174, 195, 238, 242, 272, 280, 281, 506 (all: L, private herbarium McFarlane); Ondo Prov., Oluwa Forest Reserve, *McFar-*

lane 254 (L, private herbarium McFarlane); Div. Akure, Richards 3808 (PC-PV). – RÉUNION, Anse des Cascades, 21°10'S, 55°50'E, Bruggeman-Nannenga 11014 (L, REU). – RODRIGUEZ, Balfour s.n. (NY). – SÃO TOME & PRINCIPE, Island of Príncipe, below roça Belo Monte Resort, Lat/Long NAD 83: 01°41'13.1"N, 07°26'34.8"E, Shevock et al. 49767 (L, mixed between *F. grandifolius*). – SEYCHELLES, MAHÉ, path near Botanical Gardens, Norkett 16880 (BM); banks of La Misère Road, Norkett 17025 (BM, L); PRASLIN, path near Grande Anse, Norkett 18349 (BM). – TANZANIA, Dar es Salaam University campus by Mulakuwa stream course, above University Road. Wingfield 2776A (MO); Morogoro Mts, N valley of Lupanga, behind Kiguru-nyembe Missin and TTC, 6°50.4'S, 37°42'E, Pócs 97220/A, p.p. mixed with *Fissidens* specs. (EGR, L); Nguru Mts in Morogoro District, Mt Kanga N of Turiani, Pócs 90110/L (EGR a few mixed with *F. crispulus* Brid.); East Usambara Mts, around Amani Medical Research Center, Pócs 87115/N (EGR, L); West Usambara Mts, forest in entrance of gorge of the SW escarpment, above Mazinde village, Pócs 8436/AA, AG (EGR, L). – UGANDA, Hoima District, Itohya Forest, 1.5 km SW of Muntene village, 25 km WSW of Hoima Town. 1°18.2'N, 31°3.4'E., Lye & Pócs 97120/O (EGR); Mpanga Forest Reserve, Kampala, Jones 600 (E, BM, L, cum spor.); Mpigi, Mpanga Forest Nature Reserve, 0°12'N, 32°18'E, Wigginton U 5249A (E, L); Masindi District, Budongo Forest, near Sonso, 1°43'N, 31°31'E, Wigginton U 3161c (E, L); Kabarole, Kibale Forest, Kanyawara (S), 0°33'N, 30°21'E, Bates U 8644A (E, L).

SELECTED ILLUSTRATIONS: Bruggeman-Nannenga (2006b: fig. 26), Bruggeman-Nannenga & Arts (2010: fig. 38), Bruggeman-Nannenga (2011: fig. 44), Iwatsuki & Suzuki (1982: pl. XIII), Pursell (1994: fig. 51; 2007: fig. 123, l–u).

TYPE MATERIAL NOT AVAILABLE FOR STUDY

Fissidens pechueli (Müll. Hal.) Paris, Index Bryol.: 480. 1896 – *Conomitrium pechueli* Müll. Hal., Flora 69: 503. 1886 – PROTOLOGUE/TYPE: Africa occid. Tropica [Democratic Republic of the Congo], in rivulo Loa inter Vivi et cataracta “Yélala” fluminis Congo, 1 Junio 1884: Dr. Pechuël-Lösche cum *Marchantia* sterili collegit in arena quartzitica.

The protologue is of a species with large cells, limbate leaves, and costae ending far below the apex. It is excluded since the type could not be located.

ACKNOWLEDGEMENTS. I am grateful to Dr. Jessica E. Beever for discussions and for the gift of a specimen of *F. dealbatus*, to the curators of BM, H, L, M, NY and PC for the loan of specimens, to Len Ellis (BM) who, as always, has been helpful in many ways, and to the reviewers, Dr. Jessica E. Beever and Dr. Frank Müller, for helpful and stimulating remarks and suggestions on the manuscript.

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