## Host Specificity Studies of the Pathogen *Mycovellosiella lantanae* var. *lantanae* for the Biological Control of *Lantana camara* in South Africa

A. DEN BREEŸEN<sup>1</sup>, M. J. MORRIS<sup>2</sup>, and M. SERDANI<sup>3</sup>

<sup>1</sup>ARC-PPRI, Private Bag X5017, Stellenbosch, 7599 South Africa <sup>2</sup>P.O. Box 1105, Howick 3290 South Africa <sup>3</sup>SAPO, Private Bag X5023, Stellenbosch, 7599 South Africa

Lantana camara L. is a poisonous, bushy shrub from South and Central America which has invaded many of the moist, warm sub-tropical parts of South Africa. It rapidly reinvades disturbed areas or areas cleared of other weeds. An isolate (C386) of a leaf pathogen, Mycovellosiella lantanae var. lantanae, which causes extensive defoliation of the plant, was collected in Florida, USA, and subsequently inoculated in quarantine onto South African biotypes of L. camara and a number of closely-related species, i.e. Clerodendrum glabrum, Duranta repens, Lantana rugosa, Lippia javanica, Lippia rehmannii, Lippia scaberrima, Phylla nodiflora, Stachytarpheta sp., Verbena bousariensis, and Verbena brasiliensis, to determine its host specificity. None of the potential alternate hosts showed any symptoms of infection and no signs of hyphal growth were observed. Inoculation of L. camara with the same isolate caused chlorotic, grey lesions (20-60 per leaf), and necrosis of flower buds and stalks, as well as defoliation of some plants after three weeks. This indicates a very restricted host range, making this pathogen a promising control candidate which should reduce the vigour and reproductive potential of the plant.

## Host Specificity of *Algarobius bottimeri* and *Algarobius prosopis* in Australia

## GRAHAM P. DONNELLY

Alan Fletcher Research Station, P.O. Box 36, Sherwood, Queensland 4075, Australia

The mesquite bruchids (Algarobius bottimeri and Algarobius prosopis) were tested for host specificity as biological control agents of mesquite, Prosopis spp., in Australia, following their release in South Africa. In multiple-choice tests in which test-plant pods and mesquite pods were placed close together, both A. bottimeri and A. prosopis oviposited heavily on pods of most non-mesquite test plants as well as on mesquite pods. Both bruchids developed through to adults in low numbers in seeds of Acacia aneura, Petalostylis labicheoides, Neptunia gracilis, and Arachis hypogaea with much longer development times than in mesquite seeds. Algarobius prosopis also developed in

Caesalpinia decapetala. No development occurred in the remaining test-plant pods. When tested in a large cage in which pods of A. aneura, P. labicheoides, N. gracilis, C. decapetala, and A. hypogaea were placed 1.5 metres from mesquite pods, females of both bruchids oviposited only on the mesquite pods. These experimental results indicate that A. bottimeri and A. prosopis females oviposit on non-host pods in very close proximity to mesquite pods but not on non-host pods separated from mesquite pods. Such close proximity in the field could only occur if there was physical overlap between mesquite plants and non-target plants. Mesquite infestations do not occur in areas where the native C. decapetala grows or the introduced A. hypogaea is grown, but do occur in areas where A. aneura, P. labicheoides, and N. gracilis are endemic. It is possible that such overlaps occur or will occur in the future. Indehiscent mesquite pods retain their seeds long after pods drop. Pods of A. aneura and P. labicheoides shed their seeds at maturity. Neptunia gracilis retains its seeds in pods on the plant for some time before it releases seeds. A small proportion of A. aneura and N. gracilis pods may retain some seeds after falling to the ground. Were occasional physical overlap to occur, actual intermingling of A. aneura, N. gracilis, or P. labicheoides pods retaining seeds with mesquite pods would be a rare event and oviposition on no-target pods even rarer. Algarobius bottimeri and A. prosopis females oviposit in cracks in mesquite pods in preference to the smooth surface of mesquite pods lacking cracks. The chance of mistaken oviposition on smooth naked nontarget seeds is low. The pod oviposition habits of the bruchids combined with the different fates of pods and seeds of mesquite and of A. aneura, P. labicheoides, and N. gracilis minimise the possibility of accidental non-target oviposition in the field. Algarobius bottimeri and A. prosopis pose no threat to A. aneura, N. gracilis, or P. labicheoides and are specific to *Prosopis* and safe to release in Australia. Approval for their release was sought and granted. Both species have been released.

## Exploring the Host Range of *Fusarium tumidum*, a Candidate Bioherbicide for Gorse and Broom in New Zealand

J. FRÖHLICH<sup>1</sup>, L. MORIN<sup>2</sup>, and A. GIANOTTI<sup>1</sup>

<sup>1</sup>Landcare Research, Private Bag 92170, Auckland, New Zealand <sup>2</sup>CRC for Weed Management Systems, CSIRO Entomology, GPO Box 1700, Canberra, A.C.T. 2601, Australia

Gorse (*Ulex europaeus*) and broom (*Cytisus scoparius*), both members of the Fabaceae, are important weeds that reduce the productivity of pastures and plantation forests in New Zealand. At Landcare Research, work continues towards developing the fungus *Fusarium tumidum*, which occurs naturally in New Zealand, as a bioherbicide against these weeds. While a bioherbicide capable of affecting both gorse and broom would be commercially attractive, especially to agriculturalists and the forestry industry, it is important to determine whether other plants likely to come into contact with the prototype bioherbicide in the field could also be susceptible to the fungus. Key plants of inter-