PESTS OF THE COCONUT PALM

Pests and diseases are particularly important among the factors which limit agricultural production and destroy stored products. This publication concerns the pests of the ecconut palm and is intended for the use of research workers, personnel of plant protection services and growers

Descriptions are given for each pest (adult or early stages), with information on the economic importance of the species the type of damage caused, and the control measures which have been applied (with or without success) up to the present time. The text deals with 110 species of insects which attack the palm in the field; in addition there are those that attack copts in storage, as well as the various pests that are not insects

This is the first of a series of publications on the pests and diseases of economically important plants and plant products, and is intended primarily to fill a gap in currently available entomological and phytopathological literature and so to assist developing countries.

PESTS OF THE COCONUT PALM

PESTS OF THE COCONUT PALM

by

R.J.A.W. LEVER

FOOD AND AGRICULTURE ORGANIZATION OF THE UNITED NATIONS
Rome 1969

This One

First printing 1969 Second printing 1979

> P-14 ISBN 92-5-100857-4

> > © FAO 1969 Printed in Italy

FOREWORD

Shortage of food is still one of the most pressing problems in many countries. Among the factors which limit agricultural production and destroy stored products, pests and diseases are particularly important. When left unchecked, they are liable to cause such serious losses as to discourage further attempts to grow certain crops. The application of rational control measures requires thorough knowledge of the biology and behavior of the pests and diseases, but only too often the necessary basic knowledge is lacking in the very countries in which it is most needed.

The Food and Agriculture Organization of the United Nations (rAo), fully aware of this problem, is producing a series of publications on the pests and diseases of economically important plants and plant products, primarily to fill a gap in currently available entomological and phytopathological literature and so to assist developing countries.

The first of these publications concerns the pests of the coconut palm (Coost mutifra) and is intended for the use of research workers, personnel of plant protection services and growers. Its objective is to present an account of the principal species — both invertebrate and vertebrate — that are harmful to the econout palm, and to do so for the whole of the tropical belt. To this end, the description of each pest (adults or early stages) has been made as brief as possible, to allow space for information on the economic importance of the species, the type of damage caused, and the control measures which have been applied, with or without success, up to the present time. It is hoped that the work thus planned will satisfy the requirements of those for whom it is intended; readers who are less familiar with the entomological problems of the coconut palm will find in the Introduction definitions of the principal technical and scientific terms used in the text.

In the course of the preparation of this work it has become apparent that numerous insect species recorded over many years in the literature as coconut pests are merely casual feeders on coconut or cause only minor and occasional damage. It has therefore seemed desirable to omit such species, in order to avoid overloading the text with names whose value in the present context is doubtful. The monograph of Lepesme (1947) mentions no less than 750 species of insectes associated with the coconut

palm, and indicates that over one fifth of these are specific to it. In the present work, 110 species of insects which attack the palm in the field are dealt with; in addition, there are those that attack copra in storage; as well as the various pests that are not insects.

Most of the pests of the coconut palm will readily attack other species of palm. Many of the insects known as coconut pests in various parts of the world have also attacked the oil palm of West Africa which is now cultivated in Asia, tropical America and elsewhere. The alternative host plants of coconut pests are, in general, still very inadequately known.

The bibliographical references incorporated in the text are a selection of the very large number of published works concerning pests of the coconut palm. To illustrate the wealth of the literature on this subject, it suffices to mention that the study by Gressitt (1953) of the single species Orgetes thinserens, in one relatively small area of the Pacific, contains no less than 310 bibliographical references.

ACKNOWLEDGMENTS

The author is indebted to many persons for help in the preparation of this work. Special thanks are due to J.P. Doncaster, Keeper of Entomology, British Museum (Natural History), London, and to various members of his staff to whom problems of nomenclature have been referred; to E.O. Pearson, Director of the Commonwealth Institute of Entomology, London, for identification of species by his staff and for kindly permitting the reproduction of illustrations already published in the Bulletin of Entomological Research; and to the Anti-Locust Research Centre, London, especially to Sir Boris Uvarov, Fellow of the Royal Society, for information on the Orthoptera.

The author also wishes to record his gratitude to the following specialists: W.E. Bain (Imperial Chemical Industries), J.A. Freeman (Ministry of Agriculture, United Kingdom), A.H. Green (Unilever, Ltd.), R.W. Paine (former) renomologist in Fiji), J.J.H. Szent-Ivany (formerly entomologist, Department of Agriculture, Papua and New Guinea), G.E. Tidhury (Commonwealth Bureau of Horticulture and Plantation Crops), J. Walton (Lever's Pacific Plantations Psy, Ltd.), A. Wattanapongsiri (University of Missouri, United States) and F.N. Wright (Department of Technical Co-operation, United Kingdom).

The Verlag für Recht und Gesellschaft A.G., Baske, has permitted the citation of certain passages from Petts of crops in warm climates and their control, 1962, by R. Wyniger; Longmans, Green and Go. have allowed the reproduction of a skeeth of a coconut from Plant life in Malaya, 1984, by RE. Holtrum, the n.v. Uitgeverij W. van Hovee, The Hague, the Netherlands, and P. Lechevalier, Paris, have authorized the reproduction of photographs and the redraving of illustrations altendy published in L.G.E. Kalshoven, De plagar um de cultum genutien in Industrii, 1995-15, and in P. Lepeime, Let sinistic ske publiners, 1947, respectively. Photographs of co-conut palm pests and their damage have been received for publication from the Institut de recherche pour les huiles et les olégiques (IRsto), Paris, France; from the Royal Tropical Institute, Amsterdam, the Netherlands; from the Institut actional de la recherche agronomique (IRska), Station de recherches de lutte biologique et de bioccnotique, La Minière, Versailles, France; from P. Grisson and B. Hurpin, of the above-mentioned INsa.

station; from C. Kurian, Central Coconut Research Station, Kayangulam, Kerala, India; from M.J. Way, Imperial College of Science and Technology, Sunninghill, Ascot, Berks., England; from K.J. Marschall, insect pathologist, at present specialist for the United Nations Development Program/Special Fund-Fao-South Pacific Commis sion project, Research on the control of the coconut palm rhinoceros beetle, Apia, Western Samoa; and from F.Q. Oznaes, National Research Council, Manila, the Philippines. Several photographs of locusts and of the rhinoceros beetle have been provided by Fao.

The author also expresses his thanks to his wife, a former student of the Slade School of Fine Arts, London, for certain sketches included in the text. He also records that it is a pleasure to acknowledge his indebtedness to that unknown band of abstractors who, over many yeas, have provided in the Review of Applied Eutomology, Harticultural Abstracts and Trapical Abstracts: Invaluable aummaries of entomological papers and indexes of pests. He is similarly grateful to the staffs of the libraries in London of the Commonwealth Institute of Entomology, the British Museum (Natural History), the India Office, the Commonwealth Relations Office, the Department of Technical Co-operation, the Ministry of Agriculture, and the Tropical Products Institute.

NOMENCLATURE

Recent changes in nomenclature

In recent years, the scientific names of certain of the insects and other animals associated with the coconut palm have been changed in the light of revisions of the genera concerned. These changes, so far as they have become known to the author, are indicated in the appropriate places in the text; most of them involve the generic name only. Further reference to such changes will be found on page 5.

Similarly, the names of certain of the plant species which are alternative hosts of some of the pests of the coconut palm have been changed; the royal palm, for example, known in the past as Orrodoxa regia, must now be called Roystoma stata.

Popular and scientific names of palms of economic importance

In order to avoid unnecessary repetition in referring to palms of economic importance which are alternative host plants of coconut pests, only the generic names of these palms are used in the text. The popular and the full scientific names of the more important of the palms in question are listed below:

Areca palm								Areca catechu
Date palm .								Phoenix dactylifera
Nibong								Oncosperma tigillaria
Oil palm								Elaeis guineensis
Palmyra								Borassus flabellifera
Rattan palm								Calamus spp.
Royal palm								Roystonea elata
Sago palm .								Metroxylon spp.
Sugar palm								Arenga pinnata
Swamp palm								Nypa fruticans
Talipot								Corypha umbraculifera
Toddy palm								Carvota urens

CONTENTS

	Poreword	V
	Acknowledgments	VII
	Nomenclature	_IX
	Introduction	_1
1.	Invertebrate pests.	14
••	intercolate peace.	4.4
	Insects	14
	Orthoptera	14
	Acrididae	
	Tettigoniidae	23
	Gryllidae	
	Phasmida	27
	Phasmidae	
	Isoptera	31
	Rhinotermitidae	31
	Termitidae	
	Kalotermitidae	
	Rhynchota or Hemiptera	36
	Heteroptera	
	Coreidae	36
	Pentatomidae	43
	Tingidae	44
	Homoptera	45
	Aphididae	45
	Coccidae	46
	Aleurodidae (Alevrodidae)	52

Lepidoptera	54
Rhopalocera	54
Amathusiidae	54
Brassolidae	
Satyridae	57
Hesperiidae	58
Heterocera	
Cossidae	61
Cosmopterygidae	63
Cryptophasiidae	66
Agonoxenidae	69
Galleriidae	71
Pyraustidae	74
Phycitidae	76
Psychidae	77
Limacodidae	78
Castniidae	84
Zygaenidae	86
Noctuidae	96
Coleoptera	97
Lymexylonidae	97
Hispidae	98
Curculionidae	113
Scolytidae	124
Dynastidae	125
Lucanidae	135
Melolonthidae	136
Rutelidae	136
Hymenoptera	138
Formicidae	138
Diptera	141
Scholastidae	
Mrtes	
Tenuipalpidae (Phytoptipalpidae)	
Pyemotidae	142
Tetranychidae	143
Nematodes	143
m 1 111	

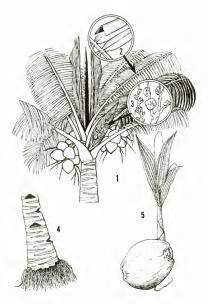
	CRUSTACEANS	144
	Paguridae	144
2.	Vertebrate pests	146
	Birds	146
	Mammals	147
	Muridae	147
	Sciuridae	150
	Pteropidae	150
3.	Insect pests of copra	152
	LEPIDOPTERA	152
	Phycitidae	152
	COLEOPTERA	153
	Cleridae	153
	Cucujidae	154
	Nitidulidae	155
	Tenebrionidae	155
	Dermestidae	155
	Avoidance of insect infestation of copra	155
4.	Practical aspects of pesticidal application	158
	References	162
	Index	174
	Addendum	181
	Supplementary references	189

INTRODUCTION

Description of the coconut palm

There is considerable variation in both the size and the period of development of the palm in the wide tropical belt where it thrives. The leaves, also called fronds (Figs. 1 and 2), reach a length of 4-6.5 meters; each has 200-250 leaflets or pinnae, which are about 1 meter long. On average, the tree produces 12-16 new leaves per year and carries 25-35 open leaves at any one time. The upper surface of the leaflets is smooth and is covered with a waxy layer; the lower surface also is smooth but is without wax, and it is on this surface that most of the insect pests, whether biting or sucking, live. At the center of the crown of the palm, the young developing leaves are tightly compacted into an elongate terminal shoot, called the spear or cabbage; severe injury to this growing point usually results in the death of the palm. Each leaf has a very strong midrib (rachis), flat above and convex below, which is attached to the trunk by a broadened base. At the junction with the trunk there are sheets of brown fibrous tissue. The positions of the bases of the old leaves which have fallen off are marked by prominent scars which are permanent characteristic features of the trunk (Figs. 4 and 6).

The coconut palm is monoecious, i.e., both male and female flowers are present on the same inflorescence (Fig. 7). The latter, which is called a spadix, develops within 2 stout sheaths or spathes, of which the inner continues growing after the outer has ceased and therefore splits is and emerges from it. A new spadix is formed every 20 to 25 days, so that there are about 16 per year. The spadix is branched; it is composed of a central axis with the branches, or spikes, disposed spirally, and each branch carries female flowers on its basal part and male flowers apically. Of the 15-20 female flowers produced on each spadix, only 4 or 5 set; these are initially small mushroom-like objects, commonly called buttons, and they rapidly grow to become pointed acorn-like nutlets (Figs. 3 and 7). Each spadix bears 250-300 male flowers, small in comparison with the females, and within about 2 weeks these fall off, 3-6 days before the female flowers on the same spadix become receptive to pollen. The receptive period lasts 4-6 days. Thus, crossferilization from a spadix opening later is



FIGURES 1-5. - Cacas musifera Linnaeus, the coconut palm. Crown (1) with details of the leaflets or pinnae (2) and of male and female flowers (3), base of the trunk and roughron (4), and a germinating coconut (5). According to Leong Hong Tim, redrawn.



FIGURE 6. - Cocos nucifera Lin-naeus. Scars on the trunk. (Photo P. Grison)



FIGURE 7. - Newly opened spadix, showing female flowers (nutlets) and many male flowers on spikes.

(Photo M.J. Way)

ensured. A single palm produces, on average, some 400 female flowers annually.

The trunk of the tree attains a height of 18-21 meters. As in all monocotyledons, there is no cambium and therefore no secondary thickening. During the first few years of growth, the trunk increases in girth but not in height, so that in young palms the bole looks disproportionately large. Upward growth does not begin until some five years have elapsed.

The wiry, fibrous roots penetrate rather shallowly into the soil but nevertheless anchor the tree firmly (Fig. 4). There is no taproot and there are no true root hairs, but the main roots are finely branched. Prolonged waterlogging is inimical but considerable salinity in the soil is tolerated.

The fruit is a drupe, with a thin epicarp overlying a thick, fibrous mesocarp. The fibers of the mesocarp, which are the commercial product called coir, are used to make rope, matting, baskets, etc. Inside the mesocarp is the hard shell-like endocarp which in turn encloses the endosperm; this provides, as the fruit ripens, the white meat or copra, rich in oil, and the watery milk, of the central cavity. About 5,000 coconuts yield 1 ton of copra, and a single ripe nut weighs 2.5-3 kg and attains a length of about 25 cm. The fruit is fully grown in 6 months but requires 3-7 months more to become mature. The coconut palm begins to bear fruit in or about the sixth year and comes into full bearing in about the twelfth year, with a mean annual production on good plantations of some 80 nuts and on particularly well-maintained ones of 150 nuts, but only about 30 in neglected groves. The embryo which develops within the nut fills the central cavity and emerges through one of three pores (eyes) in the shell; the leaves and roots penetrate the outer layers of the fruit in opposite directions (Fig. 5). The cotyledon, unlike a mature leaf, is undivided.

Classification of insect pests of the coconut palm 1

The body of all insects consists of three parts: head, thorax and abdomen, and adult insects have three pairs of legs and usually one or two pairs of wings, all these appendages arising from the thorax. Spiders, mites and ticks, which are not insects, have a different basic structure. In many insect species the early stages are legless. To reach the adult stage every insect undergoes a series of transformations which are referred to collectively as its metamorphosis. In some kinds of insects, the metamorphosis is said to be complete, because it involves three very different stages after the egg— the larva (also called caterpillar, grub, maggot,

¹ The Orders and Families represented among the pests of the coconut palm dealt with in this work are listed in Table 1, together with notes on selected characteristic features of the Orders (or other major groupings).

Table 1. - Orders and families to which the insect pests of the coconut palm belong

Order	Family	Characteristics
Orthoptera	Acrididae Tettigoniidae Gryllidae	Hind legs saltatorial; forewings tough, at rest covering membranous hindwings. Eggs laid in the soil; early stages similar to adults in form and mode of life. (Grass- hoppers, locusts, eriekets.)
Phasmida	Phasmidae	Body very clongate, stick-like; wings smal or absent. Eggs seed-like, laid freely ir trees, often dropping to the ground Early stages similar to adults. (Stick insects.)
Isoptera	Rhinotermitidae Termitidae Kalotermitidae	Social insects living in communities com- posed of well-marked eastes, such as re- productives, workers, soldiers and queens, structure and arrangement of nests vary greatly according to family and species. (Termites or white ants.)
Rhynehota or Hemiptera	Coreidae Pentatumidae Tingidae Aphididae Coceidae Aleurodidae	This strate is divided into: Heteropter (including Corcides, Pentatomidae and didae, Corcides and Aleusodiae); in the Heteroptera basal zone of forewings thick-ened, differing from distal zone, wings when at rest flat on back, crossed distally in the Homoptera, forewings and hind-upon the control of the c
Lepidoptera	Amathusiidae Brassolidae Satyridae Hesperiidae Cossidae Corprophasiidae Agonoxenidae Galeriidae Phystitidae Pyyhiidae Limacodidae Castriidae Zygaenidae Nocuidae	Two pairs of wings, covered with seales, usually forming compleseous pattern and/or color. Larve monly leef-acters and free-many leef-acters and free-many leef-acters and robbers tunnel in trunk, must or spaced on the sunnel in trunk, must or spaced by silken gridle or freely. Early stages entirely-different from adults. (Buterfiles and no-ths.)

Table 1. - Orders and families to which the insect pests of the coconut palm belong (concluded)

Order	Family	Characteristics
Coleoptera	Lymexylonidae Hispidae Curculionidae Scolytidae Dynastidae Lucanidae Melolonthidae	Forewings (elytra) tough, opaque, at rest usually covering the membranous hind- wings. Larvae with three pairs of legs or without legs. Early stages entirely different from adults. (Beetles.)
Hymenoptera	Formicidae	The ants live in communities organized in castes, in general males, workers and queens. Certain species are beneficial predators. Early stages entirely different from adults. (The order Hymenoptera side contains the bees and wasps, including several families of persatic wasps, many species of which are beneficial as enemies of economu persa.)
Diptera	Scholastidae (Ortalididae)	Only one pair of wings. Early stages en- tirely different from adults. (The order Diptera contains a vast assemblage of less with only two wings; some of them, notably in the family Tachinidae, are beneficial as enemies of eoconut pests.)

nymph, according to the kind of insect), the pupa (also called the chrysalis in the case of butterflies and moths) and then the adult; other examples of complete metamorphosis are provided by the flies, wasps and the beetles. Incomplete metamorphosis, on the other hand, is so called because the early stages after the egg resemble the adult in general form and way of life, except that they are wingless and therefore, unlike most adult insects, cannot fly; examples of types of insects having this kind of metamorphosis are the locusts, grasshoppers and the plant bugs.

The principle of classification which is used throughout the animal and plant kingdoms involves the placing of any particular specimen into its appropriate positions in a series of assemblages of descending status. The insects constitute the Class Insecta, which is divided into several Orders, and each order is divided (often via Suborders and Superfamilies) into Families, each family into Genera, and each genus into Species and, frequently, each species into Subspecies. In practice, every organism is placed directly into the appropriate genus, which is a group of species morpho-

logically and hiologically similar to one another. Thus, the so-called rhinoceros beetles, widely distributed in the tropics, can be classified as follows: Class, Insecta; Order, Coleoptera; Superfamily, Lamellicornia; Family, Dynastidae; Genus, Oryster; Species, rhinoeros (Asia), monoceros (Africa), centeurus (New Guinea).

Many species are so variable that well-marked differences warrant division of them into subspecies or into geographical races. An example is provided by a beetle of the genus Brontipa, the classification of which is as follows: Order, Coleoptera; Superfamily, Phytophaga; Family, Hispidae; Genus, Brantipa; Species, longitima; Subspecies, ulebraiti (Celbes), januna (Java), immendii (Binarack Archipelago), Irogatuli (Melanesia).

Mention is made in this work not only of the pests but also, in the more important cases, of their insect enemies, both parasites and predators. Since the pests are arranged under the names of the families and orders to which they belong, there is no difficulty in classifying them; the natural enemies, however, are named in the accounts of the pests which they attack, and in order that interested readers may know what sorts of innects they are, the family or superfamily name is given in most cases, usually with an indication of a higher group (order) to which the family or superfamily belongs. The names of the families or superfamiles are usually given in full (e.g., Tachinida, Prototrophida, Chalidadda), but the names of the higher groups are abhreviated in a form which will be readily interpreted by entomologists (Hym. for Hymnosphera; Dipt. for Diptera). Frequently, however, the family or the superfamily is sufficiently indicated by adjectival terms such as tachinid, coccinellid, braconid and chalcidid, and the higher group similarly by terms such as dipterous, hymnospherous, wasp, fix and beetle.

The scientific names given to animals and plants are not static. Taxonomic studies sometimes show that a species placed in a certain genus has greater affinity with species of another genus, to which it is therefore transferred with consequent change of its generic name. Thus, a tachinid fly (a beneficial parasitie species) known for some 40 years as Psylobing aremad is now considered by specialists to belong to the genus Bettas. Similarly, a common moth pest of copra known for more than half a century as Expetitia caustile has recently been transferred to the genus Cadra. Changes of specific names must also be made on occasion according to the rules of nomenclature; when it is discovered that another name has previously been used for a related species, the name first used and the first allocation of a name take priority. An example in this category is the most Triathaba templexas, which was for many years referred to in the literature as Tirathaba trinkogramma. Some of the names used in this work may well require changing in the future.

For purposes of accurate taxonomic identification, it is customary, and indeed necessary, to append to a species name the name (often abbreviated)

of the specialist who described that species under that name. Thus, the moth, already mentioned, whose specific name is caustella, was Ephesia cautella Walker until it was placed in the genus Cadra, when it became Cadra cautella Walker. In this volume, the name of the specialist is given at the first mention of each invertebrate species (after the Introduction), but not subsequently. The convention of putting in brackets the names of the authors of those species whose generic allocations have been changed is ignored as being superfluous in a work of this nature, as opposed to a primarily taxonomic work. In certain special instances, however, in which the name now considered correct differs from one by which the species is probably known to many readers (as in the case of Cadra cautella already cited) a special note of warning is given.

Insect damage in relation to the environment of the palm

The degree of damage caused by coconut pests can be related to some extent to the conditions of the environment, particularly to the composition of the vegetation associated with the palm and to the microclimate which it engenders, and also to soil and to weather factors (Lever, 1964a).

Effects of neighboring vegetation and of the condition of the food supply

A striking example of the effect of other plants on insect damage is provided by the long-horned grashopper fream ambile on coonut extates in the Talaud Islands near Celebes. Fransen (1954) showed that where there are areas of dense coverage of the ground by Centrotema pubezens, Sexusualays its eggs in such areas rather than in patches of bare soil or in the coconut palms; this facilitates the discovery of the eggs by the egg parasite Laffsmantis bicher, with the result that heavy parasitization occurs, sometimes attaining 95 percent, and the Sexusa population in the palms is much lower than it would otherwise be.

In New Britain it has been shown (Hoyt, 1963b) that the beetle Opylate centamru is less harmful to coconut palms in places where the palms have been interplanted with occoa than elsewhere. The same author has also demonstrated (Hoyt, 1963a) that in West Africa burning of the jungle has a more adverse effect on the predators that destroy species of Oryclus occurring there than on the beetlest themselves.

In 1928, it was observed in the Solomon Islands that attacks on the palms by the beetle Brontispa longitima fragatifi, which feeds between the unopened leaflets, were invariably less severe in the nursery on young local palms than on corresponding palms of Malayan or Samoan origin (Tothill, 1929).

Later observations showed that this same beetle in New Guinea caused more damage on young palms on which wild vines had been allowed to climb than on those properly tended (O'Connor, 1940).

Since the planting up and maintenance of coconut estates provide artificial pure stands of the one plant species, of which palm-feeding insects in the surrounding jungle can avail themselves, care must be taken to ensure that pests do not establish themselves along the boundaries and spread inward before being noticed. In the Malay Peninsula, the moth Stotas airuss, whose caterpillars eat the coconut leaves, mainly on young palms, is most troublesome in recently cleared and planted areas adjoining jungle (Corbett, 1932.) Similar observations have been made in the Solomon Islands concerning the bug Ambhybeata scaphaga, which is considered to be a relatively recent invader of the plantations, where it thrives on the extensive and readily accessible food supply (Lever, 1935).

An interesting instance of the effect of diet on a caterpillar pest of coconut palms has been given by van der Vecht (1950), who observed that the life cycle of the moth Brachartone consonable in the areas in which he worked in Java was consistently longer by 8 days than the 36 days normally recorded, and there was a reduction in the reproductive capacity of the moths and in the proportion of females to males. These effects were attributed to a physiological decline in the health of the trees. Similar observations have been made in the Sangi (Sangihe) Islands, to the north of Celebes, where Reyne (1948) recorded that deterioration of the food supply of a subspecies of the coconut scale insect Asphätons destructor resulted in a reduction of the fecundity of the female insects and heavy mortality in the early stages.

EFFECTS OF CLIMATE AND WEATHER

The seasonal changes in outbreaks of Brachartons canxuatha have been studied in several countries. In the Malay Peninsula and the Philippines, the combined effects of wind and rain on the insects tend to terminate the outbreaks which always begin in the dry season (Gater, 1925, 1926; Merino, 1938), but wind also transports the moths from their normal sites in jungle areas to distant coconut plantations (Gater, 1926).

The effects of relative humidity and rainfall on the development of the beetle Brentign aurainus have been studied on Saipan Island in the Mariana group, in Micronesia, where Lange (1950) calculated that the number of generations per year can vary from 3 to 9 according to meteorological conditions, the mean being 5 to 6. In the case of one of the true leaf-mining species of the same family (Hispidae), Promentheas serulipmini, which had become a serious pest of coonut palms in the relatively for Lus Islands of Fiji, it was concluded by Taylor (1937) that, while the low rainfall was a dominant factor in the outbreaks, its effect was nevertheless indirect.

The precipitation in Lau (about half that of other parts of Fiji) was not sufficient in the dry season to limit, as it did elsewhere, the multiplication of an unintentionally introduced parasitic mite, Pymotus veniriosus, with the result that the mite completely destroyed the larvae and pupae of Promatshera, and their indigenous parasites, in the mines of the leaves, leaving only the adult beetles. The onset of the rainy season then caused a sudden reduction in the mite population to a very low level, thus enabling the beetle to multiply without hindrans.

In India and Ceylon, it has been observed that outbreaks of the moth Nephasitis irripos are occasionally terminated by ahonormally heavy rains causing the death of the leaf-eating caterpillars in their silken galleries and favoring the development of pathogenic fungi and bacteria. The heaviest infestations occur during the hot, dry months, and it is the onset of the monsoon season that results in the reduction of the caterpillar population to a low level (Nivula vt al., 1951b).

In the Fiji Islands, coconut palms growing in swampy ground were particularly prone to severe infestation by the scale insect Asphilosus distructor. This led to a belief that ill-nourished palms were more favorable to the insect than healthy ones, but it was later concluded that the factor favoring it in such sites was that they were sheltered from strong winds (Taylor and Paine, 1935).

Hurricanes can give rise indirectly to outbreaks of pests. An increase in the population of the beetle Strategut that occurred in Puerto Rico after a particularly severe hurricane was due to the abundance of food provided by the fallen palms. In many of the islands of the Pacific, hurricanes have had the same effect on infestation by Oryster bimoerous.

EFFECTS OF SOIL COMPOSITION AND MANURING

Soil factors also can be responsible for damage to coconut palms by insects. A notable example of this is afforded by the bestle Militosama insular in the Seychelles, which is reported as occurring only on islands of granitic origin, not on those derived from coral (Vescy-Firzgerald, 1941b). This bettle-became a pest at the time when coconut palms, which are not indigenous to the islands, were first cultivated there on a commercial scale; previously its host plants there were indigenous palms of the genera Sevensonia and Nephraspermu. The same group of islands provides another instance of the nature of the soil as a factor in insect infestation, in that the damage caused by the scale insect Islanapit Inagiratiri is more severe on palms growing in alluvial soils than on those in coral sand or eroded ridges. A somewhat similar case of limitation of habitat occurs in the extreme southwest of India, where the beetle Lunapholis comophora favors sandy soils, especially those in which root crops such as supica, van, sweet potato and Cehokania are grown

(Nirula et al., 1952). This beetle is one of the few primary pests of the roots of coconut palms; it did not become serious until 1952.

In India and Ceylon, the termites Odontotermes obesus and O. redemanni attack seedling palms commonly in lateritic soils, rarely in sandy soils (Nivula et al., 1953).

Like all trees, the coconut palm responds to manuting and, in general, is less severely damaged by pests when healthy than when unhealthy. In Fiji, outbreaks of the moth Agonoxona argonala were checked by spraying with malathion to kill the caterpillars, but it was observed that the adverse effects of the insects on the health of the trees persisted longer in areas where no fertilizer had been applied to the soil and no draining or weeding had been practiced (Hinckley, 1961). In Ceylon, the caterpillars of Nophomits serinopa often cause serious damage, but only those palms growing on poor and ill-drained soils are actually killed (Kinta et al., 1956b).

It is most important to give seedling palms a good start by avoiding harsh treatment during transplanting (Barrett, 1928). Care should be taken not to damage the roots, either mechanically or by exposing them too long; otherwise the plants will be severely checked in growth and weakened, in which condition they are highly vulnerable to termite attack and to the ravages of other pests.

The shedding of immature nuts

This phenomenon, commonly referred to as "immature nutfall," must be briefly mentioned, since the primary purpose of coconut growing is the production of ripe nuts. This is a complex problem involving the agronomist, soil chemist, geneticist and entomologist.

The term "nutfall" is used rather loosely to include: firstly, the fall of young nuts, usually less than four months old, which results from attack by plant bugs, caterpillars, weevils or rats, and which is discussed under the appropriate headings: Ambippelia, Pseudubdrespia, Axingastina, Tirathaba, Dissalandra and rodents; and secondly, natural or "button nutfall," which occurs within two months from the opening of the spathe and is due to factors other than insects or rodents. Natural nutfall can be due to the shedding which must inevitably occur merely because the tree cannot nourish all the fruits which are set, as in the case of the shedding of young apples and young cotton bolls; temporarily adverse physiological factors, such as drought or flooding; or permanently adverse environmental factors, such as waterlogged soil, or hardpan preventing adequate root development, or shortage of salts (usually potash). In addition, the feranke flowers may fall off without setting, owing to abortion of the stigma, or to inviability of the pollen or lack of pollinating insects. There are almost certainly cause

of premature shedding which are still unknown. Various authors have estimated that, of the flowers which set, only 25-30 percent yield ripe nuts.

Recently, in India, the application of hormones to the trees by spraying has given encouraging results. An aqueous solution of 24-dichlorophenoxyacetic acid at 30 parts per million, plus the same volume of coconut milk or cow's urine, has led to retention of nuts and an increase of 35-160 percent in the weight of copra produced.

For many years the problem of nutfall has been studied on an ad he basis, with or without its entomological aspects, and it is encouraging to note that it is now receiving comprehensive attention as a full-time investigation. Details on this subject can be found in the literature by the following authors: Dwyer (1937); Gadd (1923); Gangolly et al. (1956); Simmonds, H.W. (1938); Tammes (1937); Taylor (1930).

Current needs and research trends

The uncontrolled utilization of modern, highly toxic insecticides of long residual effect, in place of the older products such as arsenicals, nicotine and kerosine emulsion, constitutes a very serious threat to the beneficial parasitie and predatory insects which contribute greatly to the suppression of many insect pests. The destruction of these natural enemies of harmful insects enables the latter to multiply. It is to be hoped that coconut growers will not have to face the same difficulties in this respect as those which have afflicted fruit growers in Europe and North America. It is essential to avoid the destruction of beneficial insects, and this can be achieved by careful selection of the insecticides to be used, proper timing of application and, in general, by controlled and knowledgeable use.

The recent establishment (in 1961) in Tahiti of the beetle Brantipp longitime, which attacks the young unopened coconut leaves; illustrates the necessity for strict quarantine measures. The arrival of this pest to an island situated some 5,000 km from its probable source, New Caledonia, constitutes a grave menace to the production of coconuts in a vast region of Polynesia, but fortunately there are good prospects of its control by chemical or biological means. It appears that there was also a movement in the opposite direction of another pest, the very harmful scale insect Appliation detrinative, which arrived at about the same time (probably from Polynesia) in New Caledonia and the New Hebrides. The appearance of Orystet rhimeron in Maurituis in 1962 and in the Tolekala Islands, north of Samoa, in 1963 affords further examples of recent accidental introductions of dangerous inspects.

With expanding air services and the use of ever faster aircraft, it can be expected that pests such as scale insects and leaf miners will be transported

increasingly, in coconut leaf baskets that are in common use wherever the palms are grown, unless rigid quarantine is enforced at all places where planes land. A classical example of disastrous transportation of a highly undesirable insect is that of the rhinoceros beetle to Samoa from Ceylon in packing cases of rubber budwood in 1911, prior to the establishment of international phytosanitary regulations.

The mere enactment of quarantine measures cannot attain its objective unless it is supported by means of enforcement. In Ceylon, Orytest rhimeters its still a serious pest more than half a century after the laws designed to control it were promulgated, since they have never been adequately implemented, and similar failure in Samoa to control this beetle has resulted from failure to enforce regulations enacted in 1914.

A recent instance of legislation designed to exclude insect pests is to be found in the U.S. Trust Territory of the Pacific Islands (Anon., 1959), where the introduction of coconur plants and of the nuts themselves (excluding husked nuts) is prohibited unless they have been fumigated with methyl bromide; this particular fumigant is lethal to all insects. Similar measures should be adopted in other countries to ensure mutual benefit.

It is interesting to note that current work on the control of the rhinoceros beetle in Samoa includes a return to the 50-year-old method of using fungal, bacterial and other organisms which cause disease in insects. The program also involves the utilization of parasitic and predatory insects, as well as attractant and sterilant substances and the technique of sterilization of males by irradiation.

Considerable progress has been achieved in research on the coconut palm at the permanent stations that already exist for this purpose in India and Ceylon, where teams of specialists of various disciplines concentrate their efforts on the one plant species.

1. INVERTEBRATE PESTS

INSECTS

Orthoptera

ACRIDIDAE

In comparaison with rice, millet, sugarcane and banana, the coconut palm is not much favored as a food plant by the insects (locusts and grass-hoppers) which comprise this family. Certain species have, however, caused serious damage. In general, attacks on palms have occurred only at intervals of several years and in relation to meceorological conditions, such as drought, which may cause swarms to settle in the moister areas where palms are grown.

Many species of this family, though liable to occur in large numbers, are habitually solitary-living, while others (which are called locusts) pass intermittently, and in particular circumstances, from the solitary (solitaria) phase to a gregarious (gregaria) phase via intermediate (transiens) forms. These phases have corresponding morphological and color differences which, in some species, are very striking. The transformation to the gregarious phase takes place in certain areas which, for some species, are clearly delimited, so that they can be referred to as outbreak areas. In other species, the areas where gregarious behavior develops are much less clearly defined and it is more appropriate to speak of outbreak conditions than outbreak areas. In either case, the areas where the initial swarming occurs may be very far from the coconut plantations reached by the migrating swarms. The eggs are laid in patches of bare soil, often in grassy plains, and each female deposits a succession of oothecae (egg pods), each of which contains many eggs and is buried at a depth of several centimeters in the soil (Fig. 8). From the eggs, wingless hoppers emerge, and in certain species and favorable circumstances these form themselves into yast coherent bands which advance on a front that may be more than a kilometer in length, feeding on vegetation as they go. Migratory flights of adult locusts occur while the ovaries of the females are still immature (Figs. 9 and 10).

Control

In general, satisfactory control of acridids has been obtained by applications of dieldrin, aldrin, abec or malathion. Diazinon has given good results on the experimental scale, As the use of insecticides such as dieldrin and aldrin entails a certain danger for man and domestic animals, and as acridid infestations are liable to occur in remote areas and on a large scale, particular care is necessary to ensure proper supervision in the conduct of the control campaigns. However, the availability of chlorinated hydrocarbon insecticides and the development of effective techniques for their application by aircraft have made antiacridid operations much easier, quicker and more effective than the measures previously employed, such as the distribution of poisoned bait transported in bulk by trucks and the digging of trenches and erection of barriers to stop the advance of the bands of hoppers.

In Thailand, in 1961, maize, rice, sugarcane and banana cultivations suffered serious damage caused by an outbreak of Patangs suctinta Linnaeus (Fig. 14) (Bhenchltra, 1964). The insect was combated by aerial spraying of 2.5 percent dieldrin in oil at about 140 grams dieldrin per hecture. Although cocount palms were not attacked on this occasion, it is worth noting that at this relatively low dosage the same treatment would be appropriate in the eventuality of an attack by Patangs on coconut estates. In India, the application of 100-200 grams of 10 percent patro per patron

is recommended for control of acridids (Kurian, 1963).

The natural enemies of actidids (Diptera, Hymensphra, Coleaphra, birds, bacteria and fungi) have been considerably investigated and the conclusions have been reached that, while they sometimes destroy a high percentage of an actidid population locally, and in particular circumstances, on the whole they are ineffective as controlling agents.

Locusta migratoria manilensis Meyen

Economic aspect

Outbreaks of this locust in Borneo are associated with extensive clearing of forest (Uvarov, 1928).

Several instances of damage to fruits and leaves of the coconut palm through breakage by the weight of locusts settled on them have been recorded in the Philippines (Fig. 13). Since 1919, it has been noticed that all the swarms of this locust have originated in the ince- and maize-growing areas of Cotabota or in the Masbue grasslands, in which areas efforts are made to prevent their migration to the fertile zones of Luzon and Visayas (Oranes, 1956). In 1940, by utilizing the knowledge gained from studies of the intermediate (transient) phase, it was possible to predict an outbreak which occurred in 1941-44 (Otanes, 1941). Sometimes, locusts which have originated in the Philippines have migrated in the gregarious phase to the Malay Peninsula, 1,600 km away, but outbreaks also arise there by local gregariation from the solitary phase (Corbett, 1932).

In addition to coconut, rice and maize, plants attacked are sugarcane, pineapple, bamboo, and various other gramineous species, both wild and cultivated.

Geographic distribution

The Philippines, Borneo, the Malay Peninsula, Indonesia, southern China and Taiwan.

Description | Biology

In the gregarious phase, the hoppers are blackish and the adults are graysis green and constricted in the thorax; the body of the male is 40-50 mm long, that of the female 42-55 mm. In the solitary phase, the hoppers are greenish and the adults green to blackish, without constriction in the thorax; the body length is 29-35 mm in the male and 37-60 mm in the female (Corbett, 1932).

In the Malay Peninsula, the egg stage lasts about 2 weeks. The hopper stage, comprising five or six instars, covers 37-51 days.

Control

In aerial spraying, DDT, which was formerly employed, has been replaced by dieldrin and aldrin. Outbreaks, which have lasted 7-11 years in the past and have caused serious damage to cocont palms, should in the future be brought under control much more rapidly (Otanes, 1956).

Locusta migratoria migratorioides Reiche and Fairmaire

Economic aspect

Damage to occonut palms by the African migratory locust, L. m. migratoriidet, was reported in Dahomey and Ghara during 1930-34 (Bullen, 1964), the swarms concerned having originated in the permanent breeding grounds of this locust in the floodplains of the Niger in Mali. Elasis and rice also are attacked.

Geographic distribution

This locust is found in the solitary phase in the Near East and many African countries. During the last plague period (1928-41), serious infestations of the gregarious phase occurred in nearly all the countries of Africa south of the Sahara.

Schistocerca gregaria Forskål

Economic aspect

This is the well-known desert locust (Figs. 8 to 12). It has damaged date palms more often than coconuts, but the latter have been seriously defoliated by it in coastal Kenya, in Tanzania, and in parts of India (Bullen, 1964). It eats a great range of plants and does enormous damage to many different food and cash crops.

Geographic distribution

Northern, western and eastern Africa, the Near East, Pakistan and India.

Biology

The number of generations per year varies from 2 to 4, which may occur in different areas connected by migrations. The female lays 3 egg pods on average, each pod containing about 70 eggs.

Tropidacris cristata Linnaeus

Economic aspect

This large grasshopper is one of many species that occasionally cause damage to occonut palms in South America; it was particularly severe in Brazil in 1914/15.

Biology

The egg pods, which are relatively large, are deposited at a depth of about 5 cm in the soil. The egg stage lasts 47 days, the hopper stage 166 days, and the adult stage 240 days (Azevedo, 1922).

Control

The egg pods can be exposed by plowing or digging; this results in the death of the embryos. Arsenical insecticides were formerly used against the active stages but presumably dieldrin sprays would be effective if applied suitably in relation to the biology of the insect.

Tropidacris dux Drury

Economic aspect

This species occurs in Trinidad and Venezuela. In 1954, 400 hectares of coconut plantations were damaged in Trinidad. Quassia, Meliocea, Erythrina and banana are also attacked, and are preferred to coconut.



FIGURES 8-9. - Schisteerea gregaria Forskål. Shovel sample from an egg site (8, above) and a marching band of first-instar hoppers early in the morning (9, below).





Figures 10-11. - Schistoerra gregaria Forskål. Thinly populated marching band of fourth-instar hoppers (10, above) and migrating adults (11, below).





FIGURE 12. - Schistocerca gregaria Forskål. Adult.

Control

In 1955, spraying with BHC in Venezuela failed to give satisfactory control (Hall, 1955), but in 1957 low-dosage application of dieldrin over some 35 hectares was effective (Hall, 1958).

Tropidacris latreillei Perty

Economic aspect

This grasshopper attained pest status in Guyana in 1957 and damaged coconuts; it had not previously been a serious pest for 40 years. Its main host plant is *Quassia*.

Control

Spraying with 0.5 percent dieldrin at about 200 liters per hectare is claimed to be effective; aldrin and BHC are less so.

Aularches miliaris Linnaeus

Economic aspect

This is an Asiatic species which appears to be confined to coastal areas. It attacks coconut, coffee, Areca, Erythrina, Casuarina and banana.

Geographic distribution

India, Ceylon, the Malay Peninsula, Thailand, Java.

Description | Biology

A rather large and conspicuously colored insect, with black head, yellow tuberculate thorax, greenish elytra (forewings) with yellow spots and black abdomen. The incubation period is long, about 5 months, and the hopper period is of similar duration. Each egg pod contains 45-115 eggs, and the pods are deposited in a range of soil types, including loose sand, stiff clay and loam (Uvarov. 1928).

Control

Poisoned bait can be used, but spraying with parathion is recommended against the young hoppers, as is scorching with flame guns when they are on low weeds. In Thailand, destruction of the eggs is achieved by plowing to a depth of 10-12 cm (Meksongsee, 1963).

Nomadacris septemfasciata Serville

Economic aspect

Damage to young coconut palms by this locust, which is known as the red locust, has occurred in Tanzania and in Kenya. In 1934/35, when severe infestation by swarms and hopper bands occurred, 40 hectares of young palms were defoliated by large hoppers. This species mainly frequents grasses and is therefore more frequently a pest of gramineous crops, notably sugarcane, than of occounts.

Geographic distribution

The red locust has recognized outbreak areas, mainly in southern Tanzania and Zambia, from which swarms would escape if swarming were not controlled as it is. During the last severe plague period (1930-44) almost all countries of Africa south of the equator were invaded. The solitary phase occurs in several African countries, and also in Madagascar and Mauritius.

Valanga nigricornis Burmeister

Economic aspect

The damage done by this acridid may be severe but is very localized and is normally confined to the tops of trees. It has been recorded as a pest of coconuts in the Malay Peninsula and Java, and is said to be most







Figure 13 (above). - Defoliation of palms by adults of Locusta migratoria manilensis Meyen. Arrows indicate densest groups of locusts causing sagging of leaves. The Philippines.

(Courtesy Philippine Bureau of Plant Industry)

FIGURE 14. - Individuals of Patanga succucta Linnaeus,

harmful when there is an undergrowth of Mimosa (Corbett, 1932). It occasionally occus in very large numbers over small areas. It has a wide range of food plants, including Area, Elaits, tea, coftee, rubber, Fina, Citrus, Castilloa and Artearpus; wild sugarcane and Imperata grass are said to be immune from attack.

Geographic distribution

V. nigricornis extends from India through Southeast Asia to Australia.

Description | Biology

The adult insect is greenish yellow or brown, with two black bands or patches on the femur of the hind leg. The number of eggs per egg pod is 55-115.

Oxya chinensis Thunberg

Economic aspect

This insect, known as the tice grasshopper, since it is common in areas of wet soil, particularly rice fields, occasionally attacks coconut palms. When the level of the water is high, it lays its eggs in the clumps of rice instead of in the soil. In addition to rice and coconuts, it attacks sugarcane, cotton, coffee and kapok.

Geographic distribution

Southeast Asia and Melanesia.

Other species of Acrididae

The following species have occasionally damaged coconut palms in Asia: Patanga succineta Linnaeus (the Bombay locust), Catantops splendens Thunberg, C. humilis Serville and Atractomorpha psittacina Serville.

TETTIGONIIDAE

This family comprises the long-horned grasshoppers. Only a few of them attack coconut palms, but these include species of the genus Sexaya which are very harmful:

Sexava nubila Stål

- S. coriacea Linnaeus
- S. novae-guineae Brancsik

S. karnyi Leefmans

Economic aspect

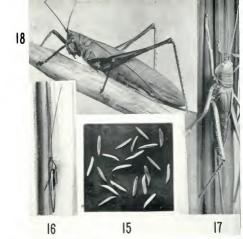
The early stages and the adults damage coconut palms and sometimes cause serious defoliation. When the leaves are reduced to the midribs the insects attack the inflorescences and the young fruits (Fig. 22). The destruction of the leaves results in immature nutfall, the tree being no longer able to nourish the normal crop (Dwyer, 1937). Other plants attacked are Arras, Metrasvilan, Nyab. banana, sugarcane, Ruemalis, Hillionis and Paudanus.

Geographic distribution

S. nubila occurs in Ceram, Kei, Aru and New Guinea; S. coriacea (Figs. 15 to 18, and 22) in the islands east of Celebes as far as Ceram and New Guinea; S. novae-guineae in New Guinea and the Bismarck Archipelago; and S. karnyi in Poat (in the Gulf of Celebes).

Description | Biology

The filiform antennae, being much longer than the body, distinguish these insects from the Acrididae (short-horned grasshoppers and locusts). In the females, the ovipositor is relatively large and scimitar-like. The males are 48-57 mm long and the females 57-63 mm, excluding the antennae which themselves measure up to 200 mm. The eggs are slender, slightly curved and 9-13 mm long. Each female produces 20-40 eggs, which are usually deposited in the soil at a depth of 10-15 mm but may also be laid in the epiphytic plants of the crown of the palm. The incubation period varies from 6 to 14 weeks, averaging 8½. The nymphs (nonadult stages) molt six times in the course of their development and become adult in 20-22 weeks in the male and a slightly longer period in the female. Thus, the developmental cycle from oviposition to the attainment of the adult stage covers 30-40 weeks (Froggatt, 1935). The rate of development is similar among the four species considered here. The insects descend to the ground at night and climb again to the crowns of the palms in the morning. They are said to be particularly harmful during or after periods of heavy rainfall.



FIGURES 15-18, - Sexuru species. Eggs (15); first-instar (16) and last-instar (17) nymphs; and adult (18) of S. coriaces Linnaeus.

FIGURES 19-21. - Eggs of Sexura species before and after hatching of nymphs (19, left), parasitized by Let/maria binder Waterston (20, center), and by Dairania let/marii Waterston (21, right), with typical emergence hole of the parasites. Photos 15-21 S. Leefmans, in L.G.E. Kalshoven, 1990-51.

(Courtesy N.V. Uitgeverij W. van Hoere, The Hague)







19

20

21

Control

BHC is recommended, either as a dust or as an oil emulsion, for application to the soil as an ovicide or to the crowns of the trees as a residual insecticide (Dun, 1955). Dieldrin, as a wettable powder, has also given satisfactory results.

As regards biological control, two parasites of the eggs, Lusfmania biolow Waterston (Hym., Chalithoidea, Emprildae) and Deirania lefimanii Waterston (Hym., Chalithoidea, Emprildae) introduced into the Talaud Islands (north of Celebes) and New Guinea from Amboina (Moluccas), have proved effective (Figs. 20 and 21). A considerable increase in the degree of parasitization in Talaud resulted from the replacement of wild host plants of Sexami in the ecocontu plantations by cocoa, cloves and Centronum pubsreasi (grown as a green manure), the latter forming a mat about 40 cm thick. The Sexama females laid their eggs in these plants, and the percentage parasitization by Lefimania reached 85-95 percent (Franssen, 1954). In New Guinea, another parasitic insect, Stitotrema dallatorrama Hoffer, of the order Strepsiptera, having a very high reproductive rate, has given good control (O'Connor, 1959b). Lizards and birds contribute to the destruction of Sexama in the crowns of the trees.

Other tettigoniids

Certain species of the genera Einnousula, Segstildae and Pseudoniszara, attributed in the past to the genus Sexona, are pests of the oconout planin New Guinea and neighboring islands (Szent-Ivany, 1956). Also in this region, Segetts decoratus Redtenbacher damages the fronds, and in the Palau Islands (Micronesia) S. mielofor Redtenbacher is similarly harmful.

All these tettigoniids, including Sexana, do much less damage on palms growing in the jungle than in coconut plantations (Leefmans, 1927). Like Sexana, they can be controlled by suitable application of BHC and dieldrin.

GRYLLIDAE

Cardiolactylus nowar-gainear Haan is the only member of this family recorded as harmful to the ecoonut palm. Damage by it is most severe on seedling palms; it attacks Pandams and Cadiausm also. It occurs in New Guinea and in the Solomon Islands. The adults are recognizable by the coloration of their wings, which are smoky black with white spots; the legs are reddish brown.



FIGURE 22. - Sexura toriates Linnaeus. Feeding traces on coconuts. Photo S. Leefmans, in L.G.E. Kalshoven, 1950-51. Amboina, Indonesia.

(Courtesy N.V. Uitgeverij W. van Hoeve, The Hague)

Phasmida

PHASMIDAE

Graeffea crouani Le Guillou (formerly known as G. cocophaga Newport)

Economic aspect

This stick insect is a serious pest of coconut palms in Polynesia and a large part of Melanesia, where it is responsible for defoliation and sometimes even death of the trees. In Fiji, the outbreaks are limited in extent to about 20 hectares. The damage by both young and adult insects takes the form of gaps cut out of the edges of the leaflets as if by scissors. In addition to coconut, Pandamus testorius, Mitenathus japanicus and Hibitous tiliansus are attacked; this last is said to be preferred to coconut in the New Hebrides (Risbec, 1937).

Geographic distribution

This species is common in the New Hebrides, New Caledonia, Fiji, Samoa, Tonga, Wallis and Futuna. A much wider distribution, including Australia and the Society, Marquessa and Gambier Islands in eastern Polynesia, has recently been recorded for it (Nakata, 1962). It has been suggested that the eggs may be carried by masses of vecetation which float on the sea.

Description | Biology

The male is 65-70 mm long, the female 105-116 mm. The wings are vestigial in both sexes but are particularly reduced in the female; the hindwing is slightly less reduced than the forewing, and is rose-pink posteriorly. The nymphs measure 20 mm at hatching, and are very fragile. The egg resembles a small grain and has a nipple-like projection. About 120 eggs are laid per female. The incubation period lasts about 100 days. The nymph molts five times if male, and six if female, in the course of its development, which occupies 92 days in the former case and 111 in the latter (O'Connor, 1954b). The total cycle therefore covers 6-7 months. The eggs are laid separately and unattached; most of them drop to the ground but some lodge in the axils of the leaves. The nymphs that hatch on the ground climb up to the crowns of the palms, where they live. Feeding occurs at night; the insects rest by day beneath the leaves, the female often carrying the male on its back. When disturbed, they eject an acrid white fluid from thoracic pores. A normal population is about 150 individuals per palm. Heavy infestations are often associated with the presence of Buchloë dactyloides (buffalo grass) under the trees (Simmonds, H.W., 1938); their direct cause is probably ineffectiveness of their natural parasitic enemies due to temporarily adverse conditions.

Control

Control is difficult, since the insect lives in the crown of the tree. Recently, however, satisfactory results have been obtained in Fiji by application from the air of a spray of 0.5 percent gamma BHC in oil at about 128 liters per hectare; alternatively, a 50 percent BHC dispersible powder can be applied at about 118 per hectare. Treatment should be repeated every four months (O'Connor, 1959a). Putting sticky bands round the trunks to prevent the insects from ascending also gives some control; the sticky material contains 2-3 parts resin mixed with 1 part oil (linseed, coconut or castor) in which is incorporated DDT at 60 grams per kilogram of the mixture. About 50 kg of this mixture suffice for applying bands 7-8 cm wide to

300 palms; the rate of treatment is about 27 palms per man-hour (O'Connor, 1954a).

The old method of causing the insects to drop to the ground by lighting fires is liable to injure the trees.

In addition to two indigenous parasites of the eggs, both belonging to the genus Paranastatus (Hym., Chalcidoidea, Eupelmidae), the ant Tapinoma melanosophalum Fabricius contributes to the destruction of the eggs. An attempt to establish in Fiji an egg parasite, Myrmesomimesis rubrifinum Mickel, (Hym., Claphiae), was unsuccessful.

Graeffea seychellensis Ferrari and G. lifuensis Sharp

These species damage the coconut palm in the same way as G. crount, the first in the Seychelles and the second in the Loyalty Islands, near New Caledonia. Outbreaks of them are often recorded in the smaller islands, where indigenous parasites may become scarce from time to time.

Ophicrania leveri Günther

Economic aspect

This insect (Figs. 23 and 24) causes damage intermittently in Savo, an island of the Solomons, where it defoliates coconut palms in coastal plantations, but not inland (Pagden and Lever, 1935). Area, Calamus and Metrosyslon are also attacked.

Geographic distribution

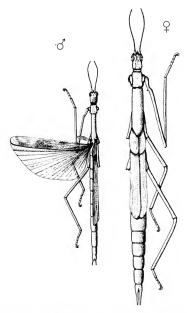
O. literi is present in all the more important of the Solomon Islands, including the isolated Santa Cruz group, but it has never been recorded as a serious pest except in Savo.

Description | Biology

The adults are rather larger than those of the preceding species, the males measuring 72-90 mm and the females 102-120 mm. The body is green and brown, and the hindwings are pale pink. Outbreaks, like those of Graffus, seem to be particularly associated with a dense ground cover of grasses and weeds under the nalms.

Control

In Guadalcanal and New Georgia, this insect is parasitized by a tachinid fly (Dipt.), which has also been recorded in Kolombangara Island and is attaining 35 percent parasitization. The parasite seems to be absent from Savo (Paine, R.W.).



FIGURES 23-24. - Ophierania leveri Günther. Male (23, left) and female (24, right).

Megacrania phelaus Westwood

This is another large phasmatid found in the Solomon Islands. It damages coconut palms on the Island of Ugi only; elsewhere it attacks Pandanur only. It would be interesting to ascertain why this and some other phasmatids attain relatively much larger numbers in small islands than in large ones, this phenomenon having been recorded for many years. According to recent investigations of R.W. Paine on this species and O. lawri in the Solomons, it seems that the inter-island variation noted in the intensity of infestation is related to the degree of parasitization by tuchind files.

Isoptera

RHINOTERMITIDAE

The species of termites belonging to this family live in subterranean nests. The individuals of the "soldier" caste have an enormous gland which opens by a pore at the front of the head and secretes a sticky fluid.

Coptotermes curvignathus Holmgren

Economic aspect

The coconut palm is liable to be attacked by this termite at any stage of its growth, but damage by it is most severe in trees 2-4 years old. Seedling palms are attacked through the nut or at the base of the young shoot, older palms through the crown or trunk (Corbett, 1932). Other plants attacked include mango, rubber, Educia and Oncoppense.

Geographic distribution

New Guinea, Borneo, the Malay Peninsula, Indonesia.

Description | Biology

The soldiers (Fig. 25) are 6-8 mm long and have a pale, brown, pearshaped head. The workers are smaller and have a pale yellow head. The nest is composed of horizontal plates of hardened evacuated food material and is normally situated in hardwood logs or stumps or in hollowed-out boles of biving trees; this species is not a mound builder. From the nest the termites tunnel through the soil searching for food, and their galleries may extend for several hundred meters at a depth in the ground varying from only a few centimeters to more than half a meter. An enormous quantity of wood is required to feed the colony. It is not possible to control this pest simply by removing the current queen because the workers can, by special feeding, create fertile queens from certain individuals called supplementary queens (Corbett, 1932).

Other species of Coptotermes

Another species of the same genus, C. eylonius Holmgren, which occurs in Ceylon, causes similar damage to palms by hollowing out the trunks. A third species, C. trunatus Wasmann, in the Seychelles, increases the damage done by the beetle Mellitumma by driving tunnels further in the sound wood of the trunk.

Schedorbinotermes marjoriae Snyder

Economic aspect

This termite excavates large compact nests among the roots of the coconut palm and also in accumulated dead vegetation. Occasionally it is found nesting in epiphytic plants on the trees.

Geographic distribution

Solomon Islands.

Description | Biology

There are two kinds of soldiers (Fig. 26) in this species, one larger than the other. The larger has a median groove at the front of the head which canalizes the viscous liquid to the mandibles. The soldiers have a characteristic yellow color.

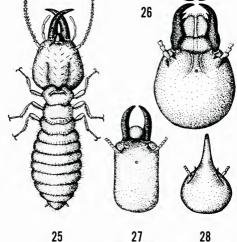
TERMITIDAE

Most of the species of termites belong to this family, which includes species that are entirely subterranean and others that build mounds. There is great variation both in types of the soldier caste and in form of nest.

Microcerotermes biroi Desneux (formerly referred to as M. piliceps Snyder)

Economic aspect

M. biroi is the commonest species of termite in the Solomon Islands. It constructs carton nests on the trunks of coconut trees about a meter above the ground, and also nests among the aerial roots at the base of the trunk and in fallen nuts (Fig. 29). It is not very harmful, although its nests are a



FIGURES 25-28. - Termites. Heads of soldiers of different species, i.e., Copto-terms currigatust Holmgren (25, according to Corbett, 1932), Schadwinsisterms: marginal soldier (26), Mirrocentermes birantherins (27), and Naustiterms: mosurams behrularum Holmgren (28, according to Harris, 1961).

FIGURE 29. - Termitarium of Microcerotermes biroi Desneux on a jungle tree trunk. Solomon Islands.

(Photo R.J.A.W. Lever)



characteristic feature of coconut plantations in the islands. Its workers have been found in the leaf stalks of young palms.

Geographic distribution

Solomon Islands and New Guinea,

Description | Biology

The winged adults probably occur throughout the year. They are small, with blackish wings. The inner edge of the mandibles of the soldiers is finely serrated (Fig. 27). The queen is only 18 mm long. Nests on the ground are smoother than those on the trunk, the latter nests usually being covered with protuberances (Harris, 1958). Strong carton runways radiate out from the nest, both upward and downward; they are constructed at the rate of about 3 cm per day.

Control

Because of the very wide range of nesting places of this termite, the practice of proper estate hygiene is particularly important in its control.

Other species of Microcerotermes

M. bagainsi Holmgren and M. perafinis Silvestri also occur in coconut plantations, in Ceylon and Samoa, respectively, the former nesting deep down among the roots (Harris, 1961) and the latter on the trunks about 2 meters above the ground; abandoned galleries of M. perafinis allow rain to enter the trunk, which rots in consequence (Hopkins, 1927).

Nasutitermes ceylonicus Holmgren

N. ephrates Holmgren

N. laticeps Wasmann

N. novarum-bebridarum Holmgren

These four species have all been recorded as pests of the coconut palm. Their soldiers have a pointed snout and minute mandibles; these features are common to all species of the genus Nasutierms (Fig. 28).

N. eylonicus, in Ceylon, nests in the roots and constructs long runways on the trunks; N. eybrates, in Panama, lives between the leaf sheath and the petiole and damages the living tissues; N. laitep, in Madagasear, is considered to be harmful mainly because it exposes the tissues of the palm to subsequent attack by boring insects of other kinds (Harris, 1961); and N. meanman.

hebridarum (Fig. 28) (formerly known as Euternes jundiniensis Hill), in the Bismarck Archipelago, New Guinea, the Solomons and the New Hebrides, attacks palms which have been struck by lightning or otherwise grossly damaged (Szent-Ivany, 1956).

Other species of Termitidae

The following species damage seedling coconut palms, particularly in nurseries:

Allodontotermes morogoroensis Harris, in eastern Africa.

Odontstremes obesse Rambur and O. radomasmi Wasmann, in India and Ceylon. These termites are more severe (causing up to 20 percent loss of seedlings) on lateritic than on sandy soils (Nirula et al., 1953); the willing of the central shoot is the first sign of their attack, which usually occurs at the base of the collar.

Macrotermes nigeriensis Sjöstedt, in western Africa; M. belliensu Smeath in Zanzibar, where 50 percent losses have occurred; and M. gilvus Hagen in New Guinea and the Malay Peninsula.

KALOTERMITIDAE

The species of this family, collectively known as dry wood termites, are mainly pests of seasoned timber and woodwork rather than living trees; nevertheless, living coconut palms have been severely attacked on occasion.

Neotermes rainbowi Hill

Economic aspect

This is a serious pest in the northern Cook Islands. The infestation starts at or near ground level and may extend up the trunk as far as the crown, even on tall palms. According to Hopkins (1927) the damage so weakened the trees that the trunks broke in hurricanes. Given (1964) records that young palms may have their fruiting capacity seriously restricted by infestations extending only 1.5 meters on the trunk. The potential danger from this termite is locally considered to be at least as great as that from Orytes rhimetern. Pandamu is frequently attacked, as are all other woody plants, except Cordia tukor-data.

Geographic distribution

Cook Islands and Ellice Islands.

THE CONTROL OF TERMITES

A high standard of plantation management and hygiene is the basis of all preventive action against termites. Fallen leaves, useless nuts and felled or broken palms must be collected and burned.

For the chemical control of termites, a wide range of insecticides is now available, and considerable success has been achieved. Harris (1961) recommends the use of BNC, dieldrin, aldrin or chlordane, and Child (1964) advises application of 10 percent BNC dust or 2 percent dieldrin dust at about 56 kg and 112 kg powder, respectively, per hectare. Nursery beds and young palms, being particularly susceptible, can be sprayed with water-miscible oil emulsions, and it is specially recommended that insecticidal dust should be incorporated in the potting soil. Other chemicals used to combat termites are pentachlorphend, sodium pentachlorphendar, parathion and heptachlor. Covering the germinating nuts with a layer of sand has sometimes proved useful.

Since the most serious damage occurs in the nursery and during the first four years of growth, it is particularly important that treatment should be applied whenever necessary during these years, in order to ensure that the palms have a good start. Young palms when first planted out are highly susceptible to damage, and this operation should therefore not be undertaken too early in areas where termites are prevalent.

On older palms termite injury appears to encourage injury by borets, and to aggravate existing damage due to them and to man—made or other wounds on the trunks. The practice of cutting steps in the trunks to facilitate the apping of toddy or the collecting of nuts for drinking should be made an offense in plantations and villages. Child has pointed out that termite attacks on established palms are largely confined to weak, moribund and even dead palms.

Rhynchota or Hemiptera (Heteroptera)

COREIDAE

Amblypelta cocophaga China

Economic aspect

A single feeding puncture of this insect in the inflorescence of the coconut palm can cause a deep fissure to develop in the tissues because the salivary glands produce a highly toxos escretion. The wound provokes a gummy exudation, and if it is on or associated with the young nuts it causes them to fall off prematurely (Figs. 30 and 31). For many years Amblypelia was not incriminated as a causative factor in nutfall because it is usually very

scarce in the trees, but its guilt was eventually established by Lever (1935, 1937) and was confirmed and amplified by Phillips (1940, 1956, O'Connor (1950) and Brown (1956, 1959). Since each feeding puncture destroys a relatively large amount of tissue, a single insect (young or adult) can do great damage in the course of its life to the inflorescences that it visits. In areas where premature nutfall was regarded as only moderate, the loss of crop due to Amblyple was about 10 perecnt (Brown, 1959).

A. cocophaga also attacks cocoa, mango, kapok, cassava, sugarcane, papaya and many other tropical plants.

Geographic distribution

Solomon Islands; A. cocopbage extends from Bougainville through most of the islands of the southern chain; in the northern chain of islands it is present on Malaita.

Description | Biology

The adult male is 13-14 mm long, the female 14.5-16 mm. The antennea are four-segmented, and as long as the body. The thorax is green in front, brown behind and furnished with two small basal spines; the forewing is reddish brown but transparent distally, revealing the brown abdomen. The legs and the ventral surface of the abdomen are yellow-brown (Fig. 32).

The eggs are 2 mm long and change from golden green to wine red as they develop; they are laid on the underside of the bases of the older leaves (Brown, 1959). The egg stage lasts 6-8 days and is followed by five nymphal instars which together cover 4½-5 weeks, the complete development thus requiring 5½-6 weeks (Lever, 1933c; Phillips, 1940). The adults take flight readily. The mean density of populations is very low, on the order of one adult only per tree, and the nymphal stage is clusive and rarely seen.

Cantral

The best results have been obtained with dieldrin, though endrin in the same conditions had the most rapid initial action. Emulsions of suc and chlordane were unsatisfactory. Aerial spraying was considered uneconomic (Fenemore, 1988) and the use of ground-operated equipment is not economic or practical since the height of the trees on long-established plantations is 20-25 meters and the manipulation of the necessary high-power mist blowers on rough ground is difficult.

With regard to control by natural enemies, neither an indigenous parasite of the eggs, Anastans axiagasti Fertière (Hym., Chalcidaidae, Espelmidae), nor indigenous predatory reduviid bugs exert sufficient pressure on this sparsely distributed insect (Lever, 1935). Two egg parasites, Oanserthus

malayensis Fertière (Hym., Chalcidoidea, Encyriidae) and Hadronotus bomooceri Nixon (Hym., Prestatrapoidea, Serlionidae), were introduced into the Solomons from Indonesis but failed to become established (Phillips, 1941). Similarly, two tachinid parasites, Pentatomophogo bicineta de Meijere from Queensland and Tritoppada penniper Fabricius from Florida, were introduced, but neither was ever recovered in the Solomons (Phillips, 1956).

A fauna rather rich in ants in the coconut plantations of the Solomons has been found to be closely related to the degree of abundance of Ambhpella and therefore to the amount of damage done by it. The predatory and Octophylla imaragdina Fabricius, which nests on the palms, is a very effective natural limiting factor when undisturbed, but in many localities it is itself attacked and rendered ineffective by two other species of ants, notably Pheliable magnetable Fabricius, which thus allow Ambhpella to maintain itself in such numbers that much damage is done to the young nuts (O'Connor, 1950; Fhellips, 1956; Brown, 1959; Lever, 1961; Greenslade, 1964). This complex of insect species, in which several other ant species in addition to those mentioned are involved, has been the subject of several intensive investigations, as a result of which attempts have been made to control Ambhpella by adjusting the conditions in the plantations so as to favor Oceophylla at the expense of Phelidels and other ant species.

Amblypelta lutescens Distant

This species, of which two forms are recognized, occurs in New Guinea and Queensland (Australia). It resembles A: ecophage but has a much wider body. It has been known in the past as a pest of banana and beans but has recently been found to attack coconut and also rubber, cassava and Ipomaes carna (Szent-Vany and Catley, 1960). The eggs are parasitized by Anatatau and Opomyrus (Hym., Chalidibida) which, however, are negligible as limiting agents.

Other species of Amblypelta occur in the Solomons on coconut but they are not yet of any economic importance (Brown, 1959).

Pseudotheraptus wayi Brown (formerly placed in the genus Theraptus)

Economic aspect

This insect (Fig. 36) causes the same damage to coconut palms in eastern Africa and Zanzibar as Amblypelas in Melanesia. A single puncture by it, resulting in necrotic lesion and gummosis, can cause the shedding of nuts even in the third month of their development (Figs. 33 to 35), but from the

FIGURE 30. - Amblypelta cocopbaga China. Injury to young nut (with perianth removed) . . .



FIGURE 31. - . . , and to older nut by feeding. According to R. Leach, in B.A. O'Connor, 1950.



FIGURE 32. - Adult.

(Photo E.S. Brown)

age of 4 months onward nuts are able to continue their development even if severely punctured by the bugs. The annual crop loss in Zanzibar due to Pundahrerapta has been valued at the equivalent of U.S. 514,000,00-55,001,000 (Vanderplank, 1960b). The first investigations of this insect were conducted by Way (1951, 1953b), who incriminated it as the cause of gummosis; previous investigators of nutfall had attributed it to unfavorable soil conditions.

Other plants attacked are: cocoa, guava, mango and cinnamon (Tait, 1954).

Geographic distribution

Coastal zones of Tanzania and Kenya, and the islands of Zanzibar, Pemba and Mafia.

Description | Biology

The adult is reddish brown above; beneath it is paler, with pink spots. The male is 12-14 mm long, the female 13-15.5 mm. The period of development (egg plus nymphal instars) is 26-40 days, of which 6-9 days is the incubation period. The adult life of the male is 84 days, and that of the female 73. The female lays, on average, 74 eggs. The first-instar nymphs have not been found on the coconut palm; the damage done to the tissues begins to be serious when the nymphs are about half-grown. Adults of both sexes take wing readily when disturbed. Each individual makes about 200 punctures in the course of its existence. The insect lives in the crown of the palm throughout its life and the nymphs are found only on or near the spadix. Few can be caught by day because they shun direct sunshine and heavy rain. Feeding takes place in late afternoon and early morning, and the insects are most active on cloudy but not heavily overcast days (Vanderplank, 1958b). The density of population varies greatly from tree to tree, but it is always low and the average per tree is only about one individual (as in the case of Amblypelta in the Solomons) (Vanderplank, 1960a.b: 1961).

Control

Dusting with 0.4 percent gamma-snc at about 22.5 kg per hectare gave good results (Way, 1953b), but it destroyed the beneficial predatory ant Occophylla longineda Latreille and should therefore never be used where this ant occurs. If additional control is needed in such areas, a selective insecticide, composed of 120 kg of 80 percent pp-0sr and 12 kg of coumarin resin in a mixture of 500 liters of kerosine and 500 liters of dieselene, can be applied round the trunks; the "bloom" of crystals which forms subsequently does not harm the relatively large Occophyla but destroys the small ants

(mentioned below) which prey upon it. The resinous mixture should be applied on paper bands to avoid scorching the trunks. These bands can prevent the passage of the small, harmful ans for 6-9 months (Vanderplank, 1958a). The effectiveness of this insecticide, which is called 10 dadk, was not improved by the addition of pyrethrum, dieldrin, endrin or rotenone.

It was later calculated that, to cover the cost of this treatment (of which 80 percent is the cost of the insecticide), an increase in yield of nuts of 45 percent was necessary. Unfortunately, the increase was only 40 percent in Tanzania and 20 percent in Zanzibar, and in the following year treated palms yielded less than others. It was therefore concluded that, despite the promising results obtained experimentally for a limited period, it would be more economical to resort to clean cultivation (Hyde-Wyatt and McKinley, 1959), together with spraying where and when necessary. The best single residual insecticide was found to be dieldrin (Wheatley, 1961).

The problem of the natural control of Ambhyelus in the Solomons is remarkably closely paralleled by that of Pawahsteraptas in castern Africa and Zanzibar. In both cases, a complex of ant species is a dominant factor, involving not only a species which is highly beneficial because it destroys the bug but also other species which prey on the beneficial one and are therefore harmful; moreover, in both cases the beneficial ant belongs to the genus Noshphila and the undesirable ones include a species of the genus Phishdo. A further similarity lies in the fact that careful ecological studies of the two complexes of insects have revealed that it is extremely difficult so to arrange the ecological conditions that the potentially highly beneficial Ocoophylla is enabled permanently to achieve an adequate degree of control of the bug.

Workers of the ant Phiriahe pourulates Mayr, which is one of those that attack Oerophylla, destroy eggs of Prandotheraptus. The later stages of the latter are parasitized by Halincaphagus zangibrane Bohart of the order Strepsiptera. An unidentified lizard devours Prandotheraptus in palms that do not contain Overphylla (Vanderplank, 1958b).

It has been observed that cashew, mango and citrus trees growing among the coconut palms on plantations favor Occopbylla and so check Preadatheraptus. Interplanting with such trees is therefore a standard recommendation in Kenya.¹



¹ In the publication Host list of finegi and insects recorded on escenat in South East Asia and the Pasific (Anon., 1962) the bug Physimerus gratifue Fabricius is mentioned as a pest of ecocontu palms, apparently in error; it is, in fact, a pest of beans and sweet potators.



FIGURE 33. - Pseudotheraptus wayi Brown. Female flowers and 6 to 10-week-old nuts showing feeding punctures. Zanzibar.



FIGURE 34. - 5 to 6-month-old nut damaged by feeding (right), compared with an undamaged nut of same age (left).



FIGURE 35. - 6-month-old nut, showing gum oozing from base two weeks after puncturing.



FIGURE 36. - Nymph on young nut. (Figures 33-36 M.J. Way)

PENTATOMIDAE

Axiagastus cambelli Distant

Economic aspect

This bug often occurs in abundance on the newly opened spadices, feeding on both male and female flowers. It can cause some fall of immature nuss, but its economic status is difficult to assess, since the damage done by it appears to be much less than the number of insects present would suggest (Fig. 37). The nutfall is probably due to loss of sap, not to injection of toxic fluid (cf. Amblypelta and Pamalaberaptar). A. cambelli is reported as causing severe nutfall in parts of the Bismarck Archipelago (Dwyer, 1937) but is rarely a serious pest in the Solomons. It attacks Arteu as well as occonut.

Geographic distribution

Solomon Islands, New Hebrides, New Guinea, Bismarck Archipelago.

Description | Biology

The head, thorax and abdomen of the adult are dark brown with yellow marks, and its length is 14-15 mm. The eggs are barrel-shaped and pearly; they are laid in clusters of about 14 on the fibrous sheath which envelops the leaf base and on the spadix, and only rately on the leaves (f. Tothill, 1929). The nymphs are whitish at first, then yellow, and finally orange with black markings. The incubation period is 7 days and the five nymphal instars together last about 45 days, so that the total developmental period is about 7½ weeks.

Control

Spraying with DDT or with a wettable powder of 50 percent dieldrin is recommended.

Two parasites of the eggs are known: Anastatu szciagati Ferrière (Hym., Chalcidaida, Euplenitale) and Mirophanusa paini Ferrière (Hym., Pradictrupaida, Stelionidae). The species of parasite which has attacked an egg can be recognized after the adult parasite has emerged from it, because each species leaves a characteristic pattern on the egg shell.

Agapophyta bipunctata Boisduval

This bug damages the leaves of young coconut palms and of *Metroxylon* in New Guinea (Szent-Ivany and Catley, 1960). Its biology has not yet been studied.



FIGURE 37. - Axiagastus cambelli Distant. Typical deformation of young nuts damaged by feeding. New Hebrides. (Photo V.I., Deluchi)

(Photo V.L. Delucchi)

TINGIDAE

Stephanitis typicus Distant

Economic aspect

Until recently this bug was regarded, mainly as a pest of banana which did only slight damage to corount, but investigations which are still in progress suggest that it may be important as a vector of virus root wilt and other diseaser, including kadang-kadang in the Philippines (Mathen, 1960) and bronze leaf wilt. In addition to coconut and banana, its host plants include Manila hemp, camphor, cardamom, Alpinia, Hedyshim, Annua and Articarpus. Lepseme (1947) cites pineapple also as a host plant. In the laboratory, a wilt has been transmitted by this insect to cowpea (Vigna inuntis) (Shanta and Menon, 1960).

Geographic distribution

Southern China, Taiwan, Korea, the Philippines, south India, Pakistan, Ceylon, the Malay Peninsula, Indonesia, New Guinea.

Description | Biolog y

The body is black and 2.4 mm long. The wings are hyaline, and the forewings are ornamented with a raised brown network characteristic of Tingidae.

Control

Application of malathion at 0.1 percent in an emulsifiable solution, or of diazinon or parathion at the same concentration, is recommended.

(Homoptera)

APHIDIDAE

Astegopteryx nipae van der Goot (formerly placed in the genus Oregma)

Economic aspect

This parthenogenetic aphid causes yellow spotting on the lower surface of the coconut leaves; it is usually of little importance but occasionally multiplies sufficiently to be harmful. It also attacks Nypa, Elasis, Zalacca and Languas.

Geographic distribution

The Malay Peninsula, China (Taiwan), New Hebrides, New Guinea.

Description

The margin of the body produces numerous white waxy filaments which mask its shape.

Control

Treatment with malathion at about 0.1 percent is recommended. The ant Oecophylla smaragdina protects the nymphs.

Cerataphis lataniae Boisduval

Economic aspect

The damage done by this aphid is seldom severe. It feeds on the leaves and sometimes on the nuts. On the rare occasions when dense infestations

occur, its presence results in the development of sooty mold (Capnodium) on the leaflets. Other host plants are Areca, Calamus, Elaeis, Kentia, Roystoma and orchids.

Geographic distribution

C. lataniae originated in tropical America but now occurs in eastern Africa and in some islands of the Pacific; it is especially serious in the Hawaiian Islands.

Description

The young individuals are green. The adults are black with a conspicuous, uniform, white waxy fringe round the margin, which gives them the appearance of scale insects.

Control

Except when beneficial coccinellid beetles are present, malathion, diazinon or parathion should be applied as a spray.

COCCIDAE

Aspidiotus destructor Signoret

Economic aspect

This scale insect is one of the most serious pests of the coconut palm. It is commonly known as the Bourbon scale, the transparent scale, or the coconut scale. In severe outbreaks, it forms a continuous crust (Fig. 40) over the lower surface of all the leaves, which first become yellow owing to heavy loss of sap and blocking of the stomata, and finally die. The leaf petioles, flower spikes and young nuts are also liable to be infested.

The range of host plants is very wide, including, in addition to coconut: Elatis, Phoenix, avocado, banana, cocoa, citrus, ginger, guava, Artocarpus, Pandanus, papaya, rubber, sugarcane, yam and many wild plants.

Geographic distribution

A. destructor is almost tropicopolitan, and is or has been a pest of coconut palms almost wherever they are grown. Recent instances of its spread are from French Polynesia to New Caledonia shortly before 1961 (Cohic, 1961) and to the New Hebrides in 1962.

Description | Biology

The body of the female is bright yellow and nearly circular in outline, and is covered with a flimsy semitransparent, only slightly convex scale, of which the diameter is 1.5-2 mm. The male scale is much smaller and oval in outline, and the insect beneath it is reddish. The adult male, unlike the female, has a pair of wings and leaves the scale when it is fully developed.

The eggs, which are yellow, are laid under the scale around the body of the female. On hatching, the minute active larvae leave the maternal scale and each takes up a position on the leaf suitable for its feeding needs; it remains in this position throughout its development and, if it is a female, throughout its adult life also. The male shed its skin three times in the course of its growth and the female twice. The incubation period is 7-8 days in both sexes, and the subsequent development to the adult stage occupies 24 days in the male and a little longer in the female, the total period of development being 31-35 days. These data on the duration of the cycle were derived from investigations in Fiji (Taylor and Paine, 1935) and Mauritius (Moutia and Mamett, 1946); in Java, the duration is about three days shorter, corresponding to a higher mean temperature. There are about ten generations per year.

Infestation is most severe in areas where the rainfall is high and the palms are close together; neglected plantations and palms growing in dense semi-wild vegetation are particularly susceptible. In general, palms growing in well-managed plantations exposed to the prevailing winds are not severely infested.

In Indonesia, a form of A. destructor, called A. destructor rigidus Reyne, occurs on several of the islands and often predominates over the typical form, from which it differs in several biological respects (Reyne, 1948). Very severe outbreaks of this form of A. destructor came to an end through natural causes, which were apparently reduction of fecundity of the females and heavy mortality in the young insects. In the Philippines also the form rigidus occurs and is often more harmful than the typical form.

Control

The waxy scale which covers the insect makes control by insecticides difficult. However, good results have been obtained in Fiji on young palms sprayed with malathion at 0.1 percent or diazinon at 0.25 percent (Hinckley, 1961). DDT is not recommended. In Trinidad, similarly satisfactory results were obtained with parathion, malathion and dieldrin.

In at least five groups of islands, successful biological control of A. detirutar has been achieved by the introduction of predacious ladybird beetles (Coccinellidae). Since several parasitic enemies of the scale insect (mainly chalcids) which were brought to Fiji from Java in 1926 made no impression on the infestations and probably failed to become established, it was decided in 1928 to introduce certain species of coccinellid beetles from Trinidad. These included Cryptogantha modicept Marshall, which was immediately highly successful, consuming all stages of the pest so voraciously and multiplying and spreading so rapidly that within nine months control was achieved and after a further nine months A. destructor was virtually eradicated (Taylor and Paine, 1935).

Similarly successful results were obtained in Mauritius in 1937 by the introduction of two other species of ladybird beetles, Chilocorus nigritus Fabricius from Ceylon and Chilocorus politus Mulsant from Java. Ten years earlier, 50 percent of the palms in Mauritius were dying in consequence of severe infestation (Moutia and Mamet, 1946). In the Chagos Archipelago, however, C. nigritus has been less effective than in Mauritius (Orian, 1959). Cryptognatha nodiceps gave successful control within two years in Principe Island, in the Gulf of Guinea, where it was released in 1955, and after a further year copra production, which had been seriously affected, was back to normal (Simmonds, F. J., 1960). In Efate Island, in the New Hebrides, where A. destructor became established in 1962 and subsequently became very injurious to the coconut palms, C, nodiceps and two other coccinellid beetles which were introduced failed to check it sufficiently, probably because relatively low night temperatures in the cool season adversely affected oviposition. Fortunately, another species of coccinellid beetle, Lindorus lophanthae Blaisdell was already known to be present elsewhere in the New Hebrides (and also in New Caledonia) and this was collected in large numbers (80,000 adults) and released in the outbreak area of A. destructor in Efate with immediate success; the outbreak was brought to an end within five months, and there is good reason to believe that a lasting biological equilibrium has been established there such that the scale insect is no longer of economic importance (Cochereau, 1965).

There are many other predatory and parasitic insects that attack A. destructor, often attaining a high degree of destruction but failing to suppress it satisfactorily. These include predacious beetles of the family Nitidulidae in the Nalay Peninsula (Lever, 1964b) and in Mauritius (Moutia and Mamet, 1946), and minute parasitic "wasps" of the superfamily Chalcidoidea, notably the aphelinids Aspidiusliphagua citrium Crawford and Aphylis chrymopholi Mercet, both of which are very widely distributed, and the encyrtid Competitla unificativata Ishii in Indonesia, which has sometimes destroyed no less than 90 percent of the scale insects without, however, bringing the outbreaks under control (Reyne, 1948).

Aonidiella aurantii Maskell

This species, commonly known as the citrus red scale, is very widely distributed in the tropics and subtropics. Particularly serious damage to

coconut palms has occurred in the New Hebrides. Metroxylon, Nybo and Phoenix are also attacked. The adult female, with its covering scale, is circular, and is yellow-orange; the kidney shape of this and related species is characteristic. Control of A. aurantii is usually maintained by chalcid parasites, of which 20 species are recorded (Lepsem, 1947).

Chrysomphalus ficus Ashmead

This is often called the Florida red scale. In addition to coconut, many other species of palms are attacked, including Areca, Howen, Kentin, Latania, Linistona, Phenix, Psylosperma and Reystona; among other plants on which it feeds are cinnamon, citrus and mango. In the Seychelles, damage to coconut palms was reported to be particularly severe on palms growing in good soil (Vesey-Fitzgerald, 1941a).

C. ficus is of Central American origin, but is now very widely distributed through the tropics and subtropics.

The female scale is circular and brownish violet; it has a reddish brown "boss" near the center.

Chalcid parasites, of which some 15 species have been recorded, usually check this scale insect (Lepseme, 1947). In the Seychelles, where further control was needed, coccinellid beetles of the genera Chilocorus and Exochomus, which were introduced from eastern Africa, gave good results.

Ischnaspis longirostris Signoret

This is commonly known as the black thread scale.

Economic aspect

On coconut, this species is found mainly on the leaves, where it forms dense colonies, but it occurs on the leaf stalk and nuts also. In severe cases, the vigor of the tree and the yield of nuts are reduced. In the Seychelles, it is more harmful on palms growing in alluvial soils than in coral sand or on croded ridges. Other plants commonly attacked are Strybinss cinnabarins, Annona, Areca, Elasis, Rafia, Reystona, Pritcharlia, Sabal, Washingtonia, Zalatata, Jamboo, Janana, coffee, cittus, cinnamon, litchia and mango.

Geographic distribution

Tropicopolitan.

Description

The female scale is long and slender, black and shiny, and wider posteriorly; the shed skin of the first instar remains attached to the scale conspicuously at the anterior end. The eggs are orange.

Control

Coccinellid beetles of the genera Chilectorus and Exochomus, introduced into the Seychelles from eastern Africa to control this and other scale insects, have greatly reduced the damage done by it.

Pinnaspis buxi Bouché

This species is sometimes called the coconut scale, but this name is applied to Aspidiotus destructor also.

Economic aspect

Outbreaks of P. luxi are sometimes so severe that the yellow-brown spots made on the coconut leaflest coalesce (as in the case of A. destrutor) and the whole leaf becomes yellow. In extreme cases, the yield of the palms is reduced to nil. The insect lives also on Aracomia, Arrica, Calimus, Dicyoperma, Eliatis, Kenlia and Tradyparpus, as well as on Pandamus, which was its first recorded host plant in the Seychelles.

Geographic distribution

Tropical America, Puerto Rico, Ghana, Sierra Leone, Seychelles, Hawaii, Fiji.

Description

The scale is brown, thin, translucent and mussel-shaped, and the shed skin of the first instar is conspicuous at the narrow (anterior) end, as in Ixchnaspii longirostrii. Males are unknown, reproduction being parthenogenetic.

Control

In Hawaii, where occonut leaves are used for the manufacture of baskets and mats, the palms were formerly subject to serious damage by P. busci, but a coccinellid beetle, Telimin initial Chapin, which was introduced from Guam in 1936, has exercised satisfactory control since then. In the Sey-chelles, severe outbreaks of P. busci caused the leaflest to crul up in such a

way as to provide shelter for an ant, *Ichnompymex* allipse Smith, which prevented the coccinellids of the genera Childron's and *Exchonsus' (already introduced for the control of scale insects) from attaining the desired degree of control in the trees in which it was present. Investigations conducted some 14 years later showed that P. how'owas still causing damage despite the destruction of a considerable proportion of the scale insects by the coccinellids (Vesey-Fitzgerald, 1953).

Eucalymnatus tessellatus Signoret

Economic aspect

This species is usually found on the lower surface of the leaves but it also feeds on the immature nuts. Its secretion of honeydew results in the development of sooty mold (Cepmulium) on the leaves, which may inhibit photosynthesis. It attacks mainfy the distal leaflests of each leaf and chiefly the distal leaflests of each leaf and chiefly the listal part of each leaflet, and only the newly opened leaflets of the youngest leaf are free from attack.

Geographic distribution

Generally distributed in the tropics and subtropics.

Description

The scale of this species is characterized by its tessellated appearance, its surface being divided by slender furrows into a pattern of polygons.

Control

As in the case of Pinnasphi, the effectiveness of predatory coccinellids is limited in the presence of the ant Technasprawc, though this effect of the latter has been somewhat offset, particularly in the Seychelles, by a fungus disease, Criphalapprinin Iecanii, which kills the scale insects; in 1953 is was observed in the Seychelles that there was much less E. testilents and correspondingly less sooty mold than there had been in 1939 (Vesey-Fitzgerald, 1953).

Vinsonia stellifera Westwood

Economic aspect

This is yet another scale insect which adversely affects the vigor of the palm by mass-feeding on the leaves. Other host plants are *Elavis, Phoenix*, mango and orchids.

Geographic distribution

Tropical America, eastern Africa, Sevchelles, Cevlon, Fiji.

Description

V. stellifera is characteristically shaped like a seven- or eight-rayed star.

Control

Where no effective parasites or predators are present, treatment with parathion, malathion, Rogor or white oil is recommended (Wyniger, 1962). These insecticles are suitable for use against all the species of scale insects which are serious pests of the coconut palm.

Aleurodidae (Aleyrodidae)

Aleurodicus destructor Mackie

Economic aspect

This insect (like all aleurodids, which are collectively known as white-flies because the wings of the adults are dusted with white powdery waxy lives in the same way as scale insects, feeding by sucking leaf or other tissues, and tending to form colonies which cause the leaves to become yellow in the areas attacked and later to shrivel. The colonies of this whitefly sometimes become very dense and cover the lower surface of the leaf with a thick white layer of tangled waxen threads (Figs. 38 and 39). The inmature stages produce honeydew in abundance, which encourages development of sooty mold, particularly in the dry season and on palms about 6-8 years old. Damage to coconuts has been recorded as slight in the Malay Peninsula but severe in the Philippines (Dammerman, 1929). Other plants attacked are Announ and Wishinghain.

Geographic distribution

Southeast Asia, the Philippines, Melanesia.

Description | Control

The waxy threads produced by the penultimate instar (usually referred to as the pupa) may be ten times as long as its body. Except in localities where parasites and predators normally exercise sufficient control, the insecticides recommended for spraying scale insects can be used to control white-flies also.



FIGURES 38-39. - Aleurodicus destructor Mackie on coconut palm(a) and on Annona leaf (b). In L.G.E. Kalshoven, 1950-51.

(Caurtery N.V. Uitgeverij W. van Hoere, The Hague)

Figure 40. - Aspidiotus destructor Signoret on coconut seedlings. Photo D. Mariau.

(Courtey Institut de recherches pour les builes et les oléagineux, Paris, France)



Dialeurodes elongatus Dumbleton

This whitefly is a pest in New Caledonia. Control measures recommended for young palms are spraying with parathion or with nicotine sulfate in oil emulsion; old palms badly infested should be felled and burned (Cohic, 1959).

Lepidoptera

(Rhopalocera)

AMATHUSIIDAE

Amathusia phidippus Linnaeus

Economic aspect

The caterpillars of this large butterfly, which eat the leaves of palms, are less harmful to coconut palms than to oil palms (Elarit). Area, Reptonea and Cyriotiachys are also attacked, and caterpillars are occasionally found on banana.

Geographic distribution

Malaysia.

Description | Biology

The distinctive hairy caterpillar (Figs. 42 and 43) is greenish gray and has a pair of spiny horns on the head, a pair of "tails" on the posterior segment and a tuft of longer hair on the second and third segments. It attains a length of 90 mm. The light green chrysalts hange from a leaf by its posterior extremity; the duration of this stage is 10-12 days. The adult, which has a wing span of 85-105 mm, is manify brown with various pale and dark streaks on the wings, and the hindwing bears on its underside two large eye-like spots and a pair of smaller dark spots on its blunt tail; the latter spots are on the upper side also (Fig. 41). The butterfly is crepuscular in habit, resting by day under the leaves (Corbett, 1932). Development from egg to adult occupies about 2 months (Kalshoven, 1950-51).

Control

Two species of tachinid flies belonging to the genera Tricholyga and Sturmia, parasitize the caterpillars. Being more susceptible to the action of



FIGURE 41, - Amathusia phidippus Linnaeus. Imago just after emergence. Photo S. Leefmans, in L.G.E. Kalshoven, 1950-51.



FIGURE 42 (left). - Young, and last-instar (Figure 43, right) caterpillars. Photos S. Leefmans, in L.G.E. Kalshoven, 1950-51.

(Figures 41-43 courtesy N.V. Uitgeverij W. van Hoeve, The Hagne) insecticides than their hosts, they would be destroyed by chemical treatments directed against the caterpillars, but when the parasites are not effective DDT, as a wetsable powder, should be applied.

BRASSOLIDAE .

Brassolis sopborae Linnaeus

Economic aspect

The caterpillars of this butterfly are gregarious. The extensive defoliation which they cause on coconut palms during the wet season can result in the premature shedding of nuts and a reduction of yield for 12-18 months and, in extreme cases, even the death of the trees. They feed also on Archontopboenix, Attalaa, Bactris, Copernicia, Detimonaus, Euterpa, Listisma, Orbigna and Reststones, and occasionally on banana (Lepseme, 1947).

Geographic distribution

Tropical South America, Trinidad.

Description | Biology

The adult butterfly is large and brown, and has a diagonal orange band on the forewing. Its wing span is 70-105 mm. The eggs are laid in masses of 100 or more on the trunks of the trees and hatch after an incubation period of 20-25 days. The caterpillars are brownish red, with longitudinal yellow stripes. They live by day in a large silken nest which they construct on the palms by spinning leaflets together, and come out to feed at night; the nest is the size of a football. The pupal stage lasts 10-14 days.

Control

The eggs of B. uphorae are parasitized by species of Telusomus (Hym., Prestortapsides, Zetionidae) and Assatatus (Hym., Chalciduda, Euphomidae); the caterpillars by Winthemia pinguis Fabricius (Dipl., Tachinidae); and the pupae by two species of Brachymeria and by Spilochalis morityi Ashmead (Hym., Chalciduda). The combined effect of these parasites is usually not great, and the entomopathogenous fungus Metarrhizium amispilius is also ineffective as a rule.

In Guyana, good results have been obtained by spraying with 0.5 percent dieldrin, and in Brazil the application of 2 percent BBC dust every 20-25 days in tall trees is recommended (Marconi, 1952). Treatment with DDT as either dust or spray proved satisfactory in Surinam (Dinther, 1955).

The nests of caterpillars should be cut down and burned whenever this is practicable. Hand-collection of insects as a method of controlling them is seldom practiced nowadays, but in the case of this insect it can be done effectively.

Brassolis astyra Godart

This is another large butterfly whose caterpillars live in nests and feed on the leaves of coconut palms. They also attack Areautrum, date palm, banana and sugarcane. Unlike B. sophorue, it is restricted to the mainland of South America. Xauthozona melanopyza Wiedemann (Dipt., Tachinidae) parasitizes the caterpillars. The control measures used against B. sophorue apply equally to this species.

SATYRIDAE

Elymnias bypermnestra Linnaeus

Economic aspect

This is one of several species of the genus *l:lymnias* whose caterpillars feed on palm leaves and occasionally cause damage. In addition to coconut, *Aresa* and *Cyrtostaelyy* are attacked.

Geographic distribution

Malaysia.

Description

The caterpillar is green with longitudinal yellow stripes, and has a pair of horns on the head and a pair of "tails" on the posterior segment. It is about 40 mm long when fully grown. The chrysalis is green, and is suspended from a leaf by its hind end. The colors of the wings differ between the sexes; the male is dark brown with metallic blue markings along the outer margin, while the female is lighter in color, orange-brown, and with ill-defined, pure white spots along the margin. The wing expanse is 55-75 mm, the male being smaller than the female.

Control

DDT at 50 percent in the form of a wettable powder is used for control.

HESPERIIDAE

Hidari irava Moore and Horsfield

Economic aspect

Severe damage by this insect is not frequent, but occasionally the tissue of leaflets is entirely eaten away by the caterpillars so that only the midribs remain and groups of trees are almost completely defoliated (Fig. 46). The leaves of old palms are said to be attacked more severely than those of young ones. Arnga, Elaiti, Areca, Metroxylon, Livitiona and bamboo are also attacked.

Geographic distribution

The Malay Peninsula, Indonesia.

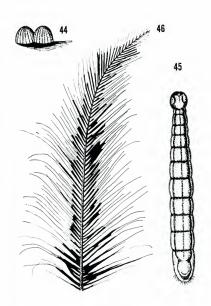
Description | Biology

The butterfly measures 50 mm in wing expanse, and is brown with four prominent golden spors and one to three small translucent spots on each forewing. The eggs (Fig. 44) are laid in chains or masses covered with the scales of the female; the incubation period is 7 days. The greenish yellow caterpillar, which is 45-50 mm long when fully grown, has a large, dark brown head and a violet stripe along each side, and its body widens posteriorly (Fig. 45). The caterpillar stage, which comprises five instars, lasts about 5 weeks. The chrysalis is attached to a leaf at its hind end and is supported by a silken girdle round the middle; it is pinkish brown and covered with a bloom of powdery waxy material. The chrysalis period lasts 10 days, so that the whole life cycle, to the emergence of the adult from the chrysalis, occupies about 25 days.

The caterpillars are more or less gregarious. They spin the two edges of leaflers together to form tubes, in which they live and in which the chrysalis is formed.

Control

Hand-collecting and destruction by burning of the rolled-up leaflets, which are conspicuous, should be undertaken. On tall trees, heavily infested leaves should be cut off at the base. Dipereous and hymenopterous parasites of the caterpillar or chrysalis, as the case may be, are known to exert some controlling influence on this pest locally. Where they are absent or obviously ineffective, a ppr snray can be used.



FIGURES 44-46. - Hideri irana Moore and Horsfield. Eggs (44), caterpillar (45), and damage to ecconut palm (46). According to P. Lepesme, 1947, redrawn. (Courtey P. Lebesalire, Paris, France)

Telicota palmarum Moore (formerly placed in the genus Corone)

Economic aspect

The damage done by this insect is similar to that already described for Hidari irana. The royal palm (Roystoma) seems to be preferred, but coconut palms are commonly attacked and young wild palms aged 2-4 years are particularly susceptible. Artea, Calamus, Elatis and Phoenix are also damaged.

Geographic distribution

India, Southeast Asia, the Philippines.

Description | Biology

The butterfly is brown and yellow, the male with much more extensive and brighter yellow areas than the female. The female, which is much larger than the male, has a wing expanse of 40 mm. The caterpillar, 40 mm long when fully grown, is dull green without a lateral stripe. The chrysalis is light green, with a dusting of white wax. The caterpillar and chrysalis stages are passed within rolled leaflets, of which the edges are spun together with silk by the caterpillars, as in the case of Hidmi roans.

Control

Numerous parasitic insects destroy T. palmarum. Wherever further control is required, the measures advocated for H. irava should be applied.^a

Gangara thyrsis Fabricius

Economic aspect

This is another species whose caterpillars feed on a wide range of palms including, in addition to coconut, Livitiona, Roytonea, Nypa, Arenga and Metroxylon; they have also been recorded on banana. Damage to coconut palms is said to be more severe in India than elsewhere.

Geographic distribution

India, Thailand, Malaysia, the Philippines.

² A very closely related species, *Telicota bambusae* Moore, in New Guinea, the Solomons and Queensland (Australia), damages coconut palms in exactly the same manner.

Description | Biolog y

The butterfly is chocolate brown with, on the forewing, two large, quadrangular, hyaline golden spots in the center and four smaller golden spots nearer the apex. The wing expanse of the female is about 80 mm. The caterpillar is covered with dense rows of long, white, waxy filaments which hidd the body. The incubation period is 7 days, and the caterpillar and chryalis stages last about 35 and 10 days, respectively, so that the total cycle to emergence of the adult covers 7.8 weeks. Both the caterpillar and the chrysalis stages are spent inside tubes made by spinning together the edges of leaflets, and the chrysalis makes an audible noise by vibrating inside the tube.

Control

Measures recommended in India are application of DDT or BHC, supplemented by hand-picking and burning of the rolled leaflets with the caterpillars inside them, or of whole infested leaves.

Other species

Three other species of Hesperiidae occasionally become numerous enough locally to cause damage on coconut palms. These are: Erionata three Linnaeus (which is mainly a pest of barana) in Southeast Asia, Suatna gemine Fabricius, mainly in India; and Zaphoptets dynmphila Trimen in Senegal and South Africa. The caterpillars of all these species live in rolled leaves and can therefore be destroyed in the manner already indicated for other species of this family.

Heterocera

COSSIDAE

Acritocera negligens Butler

Economic aspect

The caterpillar of this moth, which is commonly known as the spathe borer, bores into the unopened flower spike and tunnels through the developing male flowers inside, feeding and growing as it goes. In this behavior, the species differs from *Tirathaba*, of which the caterpillars attack the inflorescence after the spathes have spilt. The coronut palm is the only known food plant of A. negligus. Although in some plantations 50 percent

of the trees may be attacked, the damage done in terms of crop loss is usually not very serious, but on rare occasions inflorescences are almost wholly destroyed (Simmonds, H.W., 1951).

Geographic distribution

Fiji only. (This species has been recorded in error from the Malay Peninsula [Nirula et al., 1955b]).

Description | Biology

The relatively large, flat, oval eggs are laid in the fibrous sheaths which invest the bases of the leaves and spathes. The newly hatched caterpillar wanders in search of an unopened flower spike and, on finding a suitable spike, usually the oldest one, bores into it near the apical extremity. The point of penetration is marked by a gummy exudation, and there are often more than a dozen of such holes in a single spathe. The caterpillar then burrows through the tightly apposed immature male flowers, leaving moist brown excrement behind it which, after the spathe has burst open, dries to form a sawdust-like mass. It has been observed that the young caterpillars are capable of boring into the leaf stalk. After feeding for about four weeks inside the unopened spadix, the caterpillar is fully grown. It then bores through the spathes, leaving a neat round hole which is very characteristic, resembling a bullet hole, and proceeds to drop from the crown of the tree to the ground. The caterpillar then burrows into the soil, sometimes to a depth of about 30 cm, and there surrounds itself with a tough ovoid cocoon, in which it becomes a chrysalis. This stage lasts about 61/2 weeks, and when the adult (moth) is ready to emerge, the chrysalis thrusts its way upward through the cocoon and through the soil so as to project from the surface. The moth then emerges; pairing occurs within a few hours and the female moth lives only about 3 days. The incubation period is not definitely known but the complete cycle of development probably covers 3 months (Taylor, 1930).

Because the emergence holes of the caterpillars in the spathes are large and conspieuous and the damage done to the spatices is often seen when the spathes open to be extensive, there is a tendency to assume that this insect must cause considerable reduction of yield. The reason why the effect on the yield is not commensurate with the scale of the visible injury is that the caterpillar, feeding almost entirely in the distal two thirds of the flower spike, rarly encounters the female flower buds.

Control

Wyniger (1962) suggests that dieldrin at 50 percent in the form of a wettable powder, at 2-3 grams per liter of water, should be used to control this pest if chemical measures are ever considered necessary. Simmonds, H.W. (1951) suggests that cultivation beneath the palms might kill the insects, since the chrysalis period is spent in the ground.

No parasites of A. negligens are known.

COSMOPTERYGIDAE

Batrachedra arenosella Walker

Economic aspect

This small moth (Fig. 47) has been studied in several tropical countries; its caterpillar attacks the young flowers of the coconut pain (Fig. 48), but its economic importance is still in doubt. The consensus of opinion has been that the net damage done is very slight, even where the insect is abundant; this applies in the Solomons (Pagden and Lever, 1935), New Hebridge (Risbec, 1935, 1937) and Fiji. However, although Corbert and Gater considered the injury in the Malay Peninsula to be negligible in 1924, Corbett (1932), while indicating that there was still no definite evidence to the contrary, expressed the view that when the factors concerned in premature nutfall were understood, B. arenstella might well prove to be of considerable importance. Similarly, Simmonds, H.W. (1938) thought there was need in Fiji for investigation of the extent to which the loss of male flowers reduces pollination and whether any damage that may be done to the female flowers affects the yield of nuts.

Other plant species attacked are *Elasis* and *Arecastrum*; the caterpillars have also been found in fallen oranges, grains of *Juneus* and the fungus *Protubera*.

Geographic distribution

The Guianas, Congo, India, the Malay Peninsula, Indonesia, New Guinea, Melanesia, Australia, Tasmania. Nirula et al., (1955b), however, record it as occurring in India only.

Description | Biology

The moth measures only 13-14 mm in wing expanse. The forewing is straw-colored, speckled with dark gray, and has two relatively conspicuous black spots; the hindwing is fawn. Both wings are narrow and have long fringes of hair-like scales. The egg is flat and translucent, its surface

hexagonally reticulate. The caterpillar is brownish yellow, with a dark brown head, and has a brown plate divided by a pale median line on the first segment; it is 8.5 mm long when fully grown. The chrysalis is enclosed in an oval white cocoon which is about 9 mm long (Taylor, 1930).

The eggs are laid singly, in the grooves of the unopened spathes, shortly before they are due to split. The newly hatched caterpillars, according to Corbett and Gater (1924), working in the Malay Peninsula, immediately bore through the spathe and begin feeding on the developing male and female flowers, injuring up to 65 percent of the latter but without seriously affecting the crop. On the other hand, Taylor, working in Fiji, described how the young caterpillars remain in the grooves on the spathe for several days if necessary, under filmsy webs which they spin across the grooves, and wair thus without feeding until the spathe bursts open; he stresses that they do not bore through the spathe and also that they do not attack female flowers. The cocono constructed by the fully fed caterpillar is situated on the branches of the spadis or among the dead flowers that accumulate at the bases of the leaves (Fig. 48).

With regard to the duration of the life cycle, Corbett (1932) records, for the Malay Peninsula, a period of 2-3 days for incubation, 5-8 days for the caterpillar stage, and 6-8 days for the chrysalis, giving a total of 13-19 days. The corresponding periods in Fiji, according to Taylor, are 4, 10-14, and 9-11 days, with a total varying from 24 to 28 days. These differences are probably due to differences in temperature.

Control

Efficient protection can be obtained, if needed, by spraying with DDT wettable powder, either alone or mixed with wettable diazinon.

Various parasites are known in Melanesia but they seem to be of little economic importance. The eggs are parasitized by a species of Trichogramma (Hym., Chalcidoidea, Trichogrammatidae) and the caterpillars by species of Meteorus and Abanteles (Hym., Bratonidae).

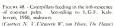
Batrachedra peroptusa Meyrick

The caterpillars of this species of *Batrachedra* devour the pollen of coconut palms and therefore may reduce pollination. Other palms attacked are *Syagrus* and *Attalea* (Bondar, 1940a). This insect occurs in Brazil and the Guianas. The duration of its cycle of development is 15-18 days.³

³ There are numerous other microlepidopterous insects which feed on the male flowers of coconut palms, but there is no reason to suspect that they adversely affect the crop.



FIGURE 47. - Batrachedra arenosella Walker, Adult. After P. Lepesme, 1947, redrawn. (Courtesy P. Lechevalier, Paris, France)





CRYPTOPHASIIDAE

Nephantis serinopa Meyrick

Economic aspect

This insect is considered to be the most important lepidopterous pest on coconut palms in Ceylon, where its caterpillars feed on the lower surface of the older leaves (Figs. 49 and 50). Its importance was recognized before 1905, but particularly serious infestations developed in 1919 (Child, 1964) and have occurred since then at intervals of about four years. In severe outbreaks, thousands of palms have all the older leaves reduced to dead brown tissue and only the three or four youngest leaves at the center of the crown remain green (Javaratnam, 1941b). In the year following a severe outbreak the crop may be reduced to half, or even more where the palms are overcrowded or growing in poor soil. In India, where the insect was comparatively rare prior to 1920, damage is most severe during the hot months (March-May); indigenous parasites are very active, but the degree of parasitization varies greatly in the course of the year and is not economically satisfactory. During the season of the southwest monsoon, the galleries (described below) in which the caterpillars live become flooded, and this favors the development of fungi (Nirula, 1956a,b). The severe damage done to the leaves results in a reduction in the rate of production of flower spikes, an increase in the premature nutfall, constriction of the trunk and retardation of growth.

Other palms attacked are Borassus, Corypba, Hypbaene, Phoenix and Roystonea.

Geographic distribution

Ceylon, southern India, Burma.

Description | Biology

The eggs are laid disally on the lower surface of the older leaves. The female lays on average 140 eggs, and the egg stage lasts 3-5 days. The caterpillar is pink, with a black head and dark thoracic plates. The galleries or runways in which the caterpillars live are constructed of silk, in which excrement is incorporated, on the underside of the leaflest. The caterpillar stage, which comprises five instars, lasts 5-8 weeks, and the chrysalls stage 9-14 days. The male moth lives about 7 days, the female 5 only. Thus, the duration of the whole cycle is 2-2½ months. The wings are gray, the forewing pale with dark specks; the wing expanse is 20-25 mm.



Figure 49. - Damage by Nephantis serinopa Meyrick. Close-up of attacked leaflets. (Photo C. Kurian)

Control

Sprays of DDT at 0.2 percent gave a reduction of caterpillars of 80 percent in two weeks (Nirula *et al.*, 1951b), but the persistence of the insecticide, of about two weeks, was prejudicial to the beneficial parasite *Trichospillar papirora* Ferrière (Hfym., Chalicháidea, Eulophidea) which attacks the chrysalis stage. But a 0.2 percent, on the other hand, lost its toxicity in 6-10 days. Experimental treatments with dieldrin at 0.05 percent destroyed 77 percent of the caterpillars; increasing the dieldrin concentration to 0.2 percent gave only 5 percent increase in the kill (Pillia and Kurian, 1960).



Figure 50. - Damage by Nephantis serinopa Meyrick. Infested tree. (Photo C. Kurian)

In India and Ceylon, a rich insect fauna provides a wide range of natural enemies of Nephantis. Hymenopterous and dipterous enemies are:

Parasitizing the egg:

Trichogramma sp. (Hym., Chalcidoidea, Trichogrammatidae)

Parasitizing the caterpillar:

Apanteles taragnae Viereck (Hym., Ichneumonoidea, Braconidae) Microbracon brevicornis Wesmael (Hym., Ichneumonoidea, Braconidae) Perisierola nephantidis Muesebeck (Hym., Bethyloidea, Bethylidae) Elasmus nephantidis Rohwet (Hym., Chalcidoidea, Elasmidae) Winthemia sp. (Dipt., Tachinidae) Stomatomyia bezziana Baranov (Dipt., Tachinidae)

Parasitizing the chrysalis:

Trisbapilus papinora Fertière (Hym., Chaiciàeidea, Eulophidae) Coryphus nervi Cameron (Hym., Iebneumonoidea, Iebneumonidea) Xauthapimpla puncitate Fabricius (Hym., Ichneumonoidea, Ichneumonidea) Brachymeria nephantidi: Gaixan (Hym., Chaicidoidea, Chaicididae) Stomateoras: unistitusellum Girault (Hym., Chaicidoidea, Chaicididae)

The efficacy of most of these parasites is reduced by secondary parasites. Predators that attack Nephanii include: canabid beetles of the genera Parena and Phlaedrowins, which devour the caterpillars in their galleries; an anthocorid bug of the genus Triphelps and a caterpillars and the mite Promets ventriesus Newport which attacks the caterpillars in southern India but without much effect on the Nebabuti infestations.

In the course of chemical control operations, efforts have been made to avoid neutralizing the beneficial effect of this complex of natural enemies. Persistent insecticides should be either not used, or applied with careful timing in relation to the stage of the outbreak and the biology of the insects concerned. The adult parasites are highly susceptible to toxic residues on the trees.

The parasite Trichapilia papinora has been reared en masse in the laboratory and released in Nephantis-infested plantations at the rate of more than a million per month (Jayaratnam, 1941b). The parasite Perisirala nephantish has also been mass-reared for this purpose. These experimental operations did not give satisfactory control.

Attempts have also been made recently to control this pest by means of the bacterium Serratia maretxent, which causes a septicemia in the caterpillars.

AGONOXENIDAE

Agonoxena argaula Meyrick

Economic aspect

The caterpillars of this small moth scarify the lower surface of the leafters and can cause serious damage, especially on young palms. This is the most common pest of coconut palms in Fiji (Hinckley, 1963). It also attacks Metroxylon and Clinastigma.

Geographic distribution

New Hebrides, Fiji, Tonga, Samoa, Wallis and Futuna, Ellice Islands, Guam, Hawaii, Palmyra.

Description | Biology

The egg is usually laid near an old feeding scar, under the web made by a larva. Incubation occupies 7-8 days. The caterpillar, which is at first yellow and later becomes green, feeds entirely on the underside of a leaflet, removing all the leaf tissue except the upper epidermis between adjacent parallel veins and thus forming a short, mainly parallel-sided sear. There are usually three or four such sears in a group. In severe cases, the sears are so numerous that they reduce the functioning leaf area of the trees by 8-20 percent (Hinckley, 1961) and the leaflets are so weakened that they bend and break in the wind, allowing the entry of fungl. The caterpillar live under a protective web which they spin over the leaf surface. The chrysalis is formed in a whitish cocoon which is formed on the leaflet or, if the caterpillar drops from the tree, on the undergrowth (Simmonds, H.W., 1938). The caterpillar stage lasts 16-22 days and the period spent in the cocoon is 9-10 days, making a total period of development to the emergence of the moth of about 5 weeks (Singh, 1951).

Control

The young palms should be treated with an aqueous preparation of malathion at about 0.3-0.4 percent. Fertilizers, drainage and control of weeds all help to reduce the harm done to the trees.

There are several indigenous insect parasites of Agoustone in Fiji; those atracking the caterpillar belong to the genera Agouative, Broson and Agolithic (Hym., Braomidae) and Tongsumpia (Dipt., Tachinidae), and a parasite attacking the chrystalis belongs to Brachymeria (Hym., Chalcidaidae). These have been reinforced by the introduction into Fiji of Elacherius agoustones Kerrich (Hym., Chalcidaidae), Esulphidae) from New Guinea and a tachinid, Actio painei Crosskey, from New Bratian, both of which parasitize another species of Agomostus, A. programma Meyrick, in their own countries, and also by a braconid, Marmeaturus, from Java. All these introduced parasites attack the caterpillars, and the average total level of parasitrization by them and the indigenous species is 71 percent, of which about half is due to Agantaets. Brathymeria destroys 35 percent of the chrysalides, the total mortality of which is 51 percent.

GALLERIIDAE

Tirathaba complexa Butler (formerly known as T. trichogramma Meyrick)

Economic aspect

The caterpillars of this moth feed on the flowers of recently opened inflorescences of the ecoconut palm. They usually inflict serious damage on the female flowers as well as on the male, and cause much shedding of young nuts. The loss of crop due to this insacti is very difficult to estimate; many of the potential nuts attacked by it would be shed by the tree in any case, since the number of female flowers produced greatly exceeds the number of nuts which the tree can nourish and for which there is room. An estimate made in Fiji indicated that, if Triathbaic could be eliminated or thoroughly suppressed, the premature shedding of nuts would be reduced by 9 percent and the yield increased by 32 percent (Taylor, 1930). The damage to the male flowers, although very great, is of no consequence (Hincklex, 1964). An unusual type of damage was reported in the New Hebrides by Risbec (1935); the caterpillars fed at the heart of the crown of palms which were 20 years old, but the trees were growing in a swampy area and were already in poor health?

Other palms attacked are Roystonea and Pritchardia.

Geographic distribution

New Hebrides, Fiji, Tonga, Samoa, Wallis, Ellice Islands.4

Description | Biology

The eggs are laid singly or in small irregular batches in the fibrous sheaths at the bases of the leaves and flower spikes, and also among the male flowers and buds, particularly on inflorescences that have failed to open fully because the spathes did not split normally. The female deposits at least 250 eggs over a period of adult life of about 8 days. The incubation period is 5.5-6 days. The young caterpillars feed on the male flowers, in and among which they spin silken galleries; the female flowers or the young must are seldom attacked until the caterpillars are about half-grown. As the caterpillars grow they construct larger and longer runways which are conspicuous among the flowers and which, since excrement is incorporated in the silk, give affected inflorescences a ditry appearance. The caterpillar stage last about 20 days, when the food consists of fresh flowers, but sometimes this stage is spent in the mass of dead, rotting flowers which accumilate at the

⁴ Records of it elsewhere than in the Pacific Islands are incorrect.

leaf bases and, in such circumstances, development is much slower. The full-grown caterpillar is 30 mm long, almost hairless, smooth, shiny and state gray and it has a dark brown head. It constructs a tough silken cocoon, reddish brown in color, with a pointed anterior end, and the period spent therein, mainly as a chrysalis, is about 14 days. The total cycle thus covers about 40 days on average. The moth is uniformly grayish brown, with a wing span of 2-35 mm, which is greater than that of the male. The sees are very different, the female being readily distinguishable by its snout-like palsp projecting forward from the head and its long abdomen tapering to a point; there is also much difference in the shape of the forewing, and the color of the male is palet, less gray and less uniform (Taylor, 1930).

Control

Wyniger (1962) suggests treatment with a dieldrin wettable powder as soon as the first damage appears on the spadix.

In Fiji, indigenous parasites are: a trichogrammatid (Hym., Chalidaida) attacking the egg, a species of Merearu (Hym., Bravonidae) and one of Editornous pha (Hym., Libouromoidae) the caterpillar, and Autrocephalus renails Waterston (Hym., Chalidaidae) the chrysalis. Since these were economically ineffective, four more species were introduced in 1930-34 from Java, where they parasitized other, closely related species of Tristbaba (Paine, 1935). These additional parasities were: Argyrophysic (Eriyai) bailquia Bezzi (Dipt., Tachinidae), Denvejilla (Nemeritii) palmaris Wilkinson (Hym., Ichmunomidae), Apantelis tiratbabae Wilkinson (Hym., Proteinropidea, Sectionidae). All these introduced parasites became established in Fiji and have jointly increased to a valuable extent the natural control of Tiratbabae.

Tirathaba rufivena Walker

Economic aspect

The caterpillars of this species cause the same damage to the inflorescences as T. emphs.c, but they are particularly destructive to those that are unusually small and those that do not open properly. Male flowers are preferred but female flowers are also attacked; one caterpillar usually destroys only one female flower. A detailed study of the part played by this insect in premature nutfall was made in the Malay Peninsula by Corbert (1932). This author concluded that the palm sheds those female flowers (and very young nuts) damaged by Tirathuha rather than healthy ones and that, since a great excess of female flowers is normally produced and must be eliminated, the insect is harmful only if it causes 58 percent or more of shedding. Other plants attacked are Nypa, Elusis, Arrea, Reystonea, Musa, Phaseolus and Coix (Lepesme, 1947).

Geographic distribution

Ceylon, the Malay Peninsula, Borneo, Indonesia, New Guinea, Queensland, Solomon Islands, New Hebrides, New Caledonia.

Description | Biology

The forewing of the female has a green reflection and bright red veins. The wing span of the female is 25-30 mm (or rather more, according to Lepesme, 1947) and that of the male 22-26 mm. The egg and the caterpillar are very similar to those of T. complexa. In the Solomons, the caterpillar is a pale plant color when feeding on healthy flowers on the spadis but darker when its food is fallen flowers at the base of the spathe (Lever, 1933b).

The duration of the egg stage is 4-5 days, the caterpillar stage lasts 14-15 days, and the period spent in the ecocon 11-12 days, so that the whole cycle covers about 4½ weeks. These data apply to individuals reared on healthy flowers; when they are fed on old fallen male flowers, the duration of the cycle is greatly extended, to about 7 weeks (Paine, R.W.).

Control

Since the newly split spathe shelters the partially released inflorescence, and in so doing provides feeding sites particularly favorable for the caterpillar, Corbett (1932) suggested that spathes that are about to burst open should be removed so that the flowering branches are fully exposed; this operation is practicable on young palms but would be difficult on tall ones.

Wyniger (1962) suggests the same insecticidal treatment as for T. complexa. The most important parasites of T. roplinea in Malaysia are Devergilla palmaris and Apantiles tirathabus, both of which have been introduced into Fiji to combat T. complexa. A. tirathabus has also been found in the Solomons, where it parasities Tirathabus on Nypa palmas swell as coconut (Lever, 1933b). Two other parasites of T. roplinea in Malaysia are Trichaplate aphieva (which also attacks Niphantii in Ceylon) and Argynphylax basilphus, which is another of the parasites introduced into Fiji. *

Tirathaha mundella Walker

This is yet another species of *Tirathaba*, which occurs in coconut palms. Its distribution extends from Thailand and Indonesia to the Solomon Islands.

 $^{^5}$ T. ignerens Hampson of New Guinea is now regarded as synonymous with T. rufirens Walker, and the latter name has priority.

Its habits and the damage caused by it closely resemble those of the species already described.

PYRAUSTIDAE

Hedylepta blackburni Butler (formerly assigned to the genera Phostria and Omiodes)

Economic aspect

The caterpillar of this moth eats the leaves of the coconut palm, giving them a very ragged appearance and considerably reducing the effective leaf surface when the attack is continuous (Fullavary and Krauss, 1945; Zimmerman, 1958). The damage is often especially severe in exposed situations, probably because the activities of the natural enemies are hindered by the strong winds. Other plants attacked are Pritchardia palms (P. parifica was the endemic host plant) and banana.

Geographic distribution

Hawaiian Islands only.

Description | Biology

The eggs are laid in rows of about 40 along the midribs of the leaflets, overlapping like tiles. They are flattened, and broadly oval in outline. On hatching, the young cateroillars feed gregariously on the underside of the leaflet, protected by a flimsy web of silk. At first they do not eat right through the leaflet but leave the upper epidermis intact. Soon they scatter and make tubes, in which they shelter, by spinning the edges of each leaflet together. From this stage onward they feed at the edge of the leaflet and eat the tissues away completely; when only a small part of their shelter remains on the midrib, they move to another leaflet and repeat the process. They become full-grown in about 4 weeks, and are then dull greenish, with two dorsal whitish lines and a whitish line along each side, and are 32-35 mm long. The chrysalis is formed in a slight cocoon constructed in the rolled leaflet; this stage lasts 11-13 days (Zimmerman, 1958). The adult measures 30 mm in wing span; the forewing is brownish ocher with two cream-colored crosslines sharply angled outward and the veins conspicuously pale, and the paler hindwing has a single, similarly angled crossline.

Control

Apparently no chemical control measures are applied

H. blackburii is attacked by a wide range of hymenopterous and dipterous parasites, some native and some introduced. The most important is Zaleptapgua flavo-orbitalis Cameron (Hym., Ichausunoidae), which sometimes attains 90 percent parasitization, without, however, preventing outbreaks. The others include species of Colebana, Microbravan and Brason (Hym., Brasonidae), Extendidae, Exchibrosorpha and Nespimpla (Hym., Ichausunoidae), Bradymeria and Trichogramma (Hym., Chalcithichia) and Frontina and Chentogida (Dipt., Tachitidae), The two tachinids were introduced into Hawaii from America and the Brachymeria and the Brachymeria and the Brachymeria and the gas are destroyed by the predatory and Philips magaerphala Fabricius and the eage are destroyed by the predatory and Philips (Messae).

This complex of natural enemies undoubtedly exerts a considerable degree of control on *Hedylepta*.

Pimelephila ghesquierei Tams

Economic aspect

The caterpillars damage the spike of young unexpanded leaves at the center of the crown of the coconut palm by burrowing into it. Coconut, though sometimes severely damaged, is not a preferred host plant; Elasis is the principal host plant. Amistrephyllum is also attacked.

Geographic distribution

Throughout the forest zone of central and western Africa, wherever the oil palm (Elavis) is native.

Description | Biology

The adult insect is a small moth, 25-34 mm in wing expanse. The fore-wing is olive-green, with dark brown morthing and diffuse cinamon spots; the hindwing is olive-gray, and paler. The eggs are characteristic, being much flattened and reminiscent of minute fatyl droplets or tiny lenses. At first milky yellow, they become bloodred as development of the embryo proceeds, this being the color of the newly hatched caterpillar. Later, the color of the caterpillar becomes lighter and changes to a yellowish pink, except the head which is light hestnut brown and the first segment which is yellowish. The body is slightly hairy. When fully grown, the caterpillar is about 35 mm long. The mahogany brown chrysalis is enclosed in a cocon composed of fibrous debris spun together with fine silk, and generally situated on the inwarf face of the stalk of an old leaf.

Control

The following procedures are recommended as preventive measures: maintaining the young palms in good condition in the nursery beds, with suitable manuring as required; ensuring, when the young palms are planted out, that the spacing adopted suffices to allow maximum light and aeration; avoiding planting in poorly darined soil; removing neglected palms, young or old, from the proximity of nurseries, since they constitute potential sources of infestation. Hand-collection of the caterpillars and occoons on young trees is recommended, subject to supervision to ensure that the trees are not damaged. Where chemical treatment is necessary, spraying with DDT at 1 percent, parathion at 0.02 percent, or endrin at 0.1 percent can be expected to give good results, especially if the spray reaches the bases of the central leaves.

PHYCITIDAE

Hyalospila ptychis Dyar

Economic aspect

Insufficient attention has been paid to this insect, which may be the most important pest of the ecoconut palm in Bahia State of Brazil. The caterpilar feeds on the female flowers and bores into the young nuts which, in consequence, are either deformed or caused to fall off prematurely. Gum issues from the tunnels made in the nuts. This is one of the very few lep-idopterous insects which bore into nuts when they are at a rather advanced stage of development. Other plants attacked are Althou and Syagrus.

Geographic distribution

Brazil.

Description | Biology

The caterpillar is white with a brown head, and is 15-16 mm long when fully grown. The adult measures 14 mm in wing span; the forewing is brown at the base and whitish distally, with reddish violet marks and black spots, and the hindwing is transparent except for a brown border (Lepesme, 1947). The development cycle lasts 25-30 days (Bondar, 1940a).

Cadra cautella Walker (formerly assigned to the genus Ephestia)

This is a very well-known pest of certain stored products, including copra, and is therefore dealt with in Chapter 3 of this study. The caterpillars

have, however, been recorded in Brazil as attacking coconut flowers, both male and female, and also ripe nuts on the trees. The duration of the cycle of development in this role is one month (Bondar, 1940b).

PSYCHIDAE

Mahasena corbetti Tams

Economic aspect

The caterpillar of this moth is easily recognized among coconut peats because it constructs and lives in a conical silken case in which fragments of leaf are incorporated (Figs. 51 and 52). It is typical, in this respect, of the family Psythidae to which the name "bagworms" is commonly and collectively applied. The caterpillar of M. entertif feed on the leaflest, first scarifying the tissues on the lower surface, then eating holes right through the leaflest and finally feeding at the edges. The damage done to the trees, though sometimes severe, is usually localized. Numerous other plant species are attacked, including Arraa, Arraga, Ellatis, Albaritat, Currans, citrus, kapok, deries and species which yield eutat-percha.

Geographic distribution

From Malaysia and Thailand to New Guinea.

Description | Biology

The male adult is a normally winged moth, brown in color and 25:30 mm in wing expanse. The female is wingless, yellowish brown and 17 mm long; its life is passed entirely within the case which it constructed around itself as a caterpillar. In fact, the female is a very degenerate insect, little more than a bag of eggs. The caterpillar scretes a long thread of silk with which it is transported by the wind, particularly from tall trees to shorter ones. Movement to trees in new areas, however, is more often effected by crawling along the ground and up the trunks. The case constructed by the caterpillar is about 30 mm long when full growth is attained. The duration of the life cycle is probably about 3 months (Corbett, 1932).

Control

Sticky bands of resin and castor oil are applied to the trunks to prevent the caterpillars from climbing to the crowns of the trees. Dusts or sprays of DDT sufficiently protect attacked leaves. In the Malay Peninsula, this pest is parasitized by a chalcid and a tachinid; the latter is Tribbigga aberrans Strobl, 41 individuals of which emerged from a single caterpillar. In Thailand, this bagworm is attacked by the tachinid Exoritia quadrimanulata Baranov (Meksongsee, 1963).

Acanthopsyche bypoleuca Hampson and A. cana Hampson

While Mahasena corbetti, with its wide geographical range, is the most likely psychid to be encountered on occonut palms, there are, in Ceylon, two species of Acanthopyrche which are similar in appearance and life history to it and which are liable to occur on them.⁶

LIMACODIDAE

Setora nitens Walker

Economic aspect

There are several species of this family which attack and damage coconut palms, and Setora nitent is probably the most harmful of them. The caterpillars eat the leaves and in severe outbreaks they are so abundant that the leaves become brown and the crowns of the trees appear scorched. Young trees suffer more severely than older ones. The body of the caterpillar is armed with urticating spines which can cause great discomfort to laborers employed on the plantations. Other plants attacked are Elastis, Nyjae, banana, cocoa, coffee, tea, citrus and tobacco.

Geographic distribution

Malaysia, Viet-Nam, Laos and through Indonesia to New Guinea.

Description | Biology

The egg is somewhat flattened and is 3 mm long and broadly oval in outline; it is semitransparent and for this reason is difficult to detect on the underside of the leaflet where it is deposited. The period of incubation is 6-7 days. The caterpillar moves in a smooth gliding manner reminiscent of slugs (hence the popular name of "slug caterpillars" which is applied to the familt, though many of the species have no other similarity to slugs).

⁶ Other species which occur in South Viet-Nam on Nypa should be regarded as potential coconut pests,



FIGURES 51-52, - Mahasma corbetti Tams. Caterpillar (51, above) and chrysalis (52, below) in their typical case. According to P. Lepesme, 1947, redrawn. (Courtesy P. Lechevalier, Paris, France)

and is fully grown in 18-32 days. The color of the caterpillar varies from green to orange-red, with a median purple band ornamented with spots. At each end there are two pairs of prominent spine-bearing processes, of which the first and the last pairs are largest. The length of the caterpillar varies greatly, up to 35 mm, mainly according to its sex. It constructs, usually near the ground, to which it drops from the tree, a smooth, spherical, parchment-like cocoon, which somewhat resembles an "oak apple" gall, and transforms to a chrysalis therein; the cocoon varies in diameter up to 15 mm (Kalshoven, 1950-51). The period spent in the cocoon, like the previous stage, varies greatly in duration, 17-31 days. The whole cycle covers 6 to 10 weeks, and exceptionally much longer, and the variation in rate of development results in marked overlapping of generations. The moth is brown, with darker reddish brown bands across the wings; the female measures 30-35 mm in wing expanse but the male is much smaller.

Control

Spraying infested trees with DDT at 0.1-0.2 percent is recommended as a general measure (Kurian, 1963), but in Sabah good results have been obtained by spraying the undergrowth with BHC at times of large-scale movement of the caterpillars to the ground; this reduces the risk of destroying the useful parasitic insects, which mainly frequent the crowns of the trees.

The tachinid fly, Chaeteoarista januma Brauer and Bergenstamm, is the most effective controlling agent of the parasite complex; the duration of its life cycle is only 2-3 weeks, less than half that of 5. nitrat. Hymenopercous parasites include species of Bruchymeria, Eurytoma (Chalcidoidea), Spinaria (Bravonida) and Xunthopipulo, (Ichammonida).

Macroplectra nararia Moore (often assigned to the genera Natada and Parasa)

Economic aspect

In India, this species, in the caterpillar stage, damages the leaves of the coconut palm, particularly during the dry season. In Ceylon, it is injurious to coffee, tea, Zizyphus and Pithetolobium, in addition to coconut.

Geographic distribution

Eastern India and Ceylon.

Description Biology

The eggs are laid on both the upper and lower surfaces of leaves. Their period of incubation is 4-5 days. The caterpillar is yellow-green above, pink below, is 8-11 mm long at the completion of its growth, and bears tubercles with short spines. It is fully fed after 31 days, and then proceeds to construct a spherical, brown cocoon, 6 mm in diameter. The moth emerges from the cocoon 16 days later (Menon and Pandalai, 1958). The cycle from egg to moth covers 7 weeks. The moth is brownish ocher; the basal two thirds of the forewing is red, bordered with brown.

Control

Among the numerous chalcids and braconids which parasitize M. nararia, the eulophid Nepfetrus maedatus Fertire (Hyn., Chalcidade) is the most effective. The natural enemies also include a wasp of the family Chryidida. During the rainy season in India, entomogenous fungi of the genus Alprejillur contribute to the natural control (Kurian, 1963), and in Ceylon a suspension of a granulosis virus has been applied in an attempt to suppress this pest.

Parasa lepida Cramer

Economic aspect

The caterpillars of this species are injurious to the leaves of coconut palms, and are sometimes so abundant that large trees are completely defoliated by them. Other plants attacked are: Bosassus, Nypa, banana, Alavinia, Artsearpus, Ricinus, tea, coffee, Comandia, mango, Fagraes, cocoa, pepper, pomegranate and cauliflower (Corbett, 1932).

Geographic distribution

Western Africa, Mozambique, India, and eastward through South Viet-Nam and Malaysia to New Guinea.

Description | Biology

The eggs are laid in masses on the underside of the leaflets; the incubation period is 7 days. The caterpillars are gregatious. When fully grown, they are green with a greenish blue dorsal stripe bordered with yellow, and they are armed with spine-bearing tubercles but not with longer processes like those of Sciena nitens; the length of the larger (female) caterpillars is about 25 mm, and the period from hatching to construction of the cocoon is about 6 weeks. The nearly spherical cocoons are often formed on the trunks of trees, massed together in large numbers (Kalshoven, 1950-51); the moths emerge 3-5 weeks later, the total cycle covering some 10-12 weeks. The adult is mainly green but the outer margin of the forewing is reddish brown. The wing expanse of the male is 30 mm, of the female 45 mm.

Control

Against this pest, an aqueous preparation of DDT at 0.2 percent can be applied as a spray; alternatively, BHC is effective.

Many parasitic enemies of Parata Ispida are known. Among these are: Euryloma monemar Ruschka, Eupelmus catoxonthus Ferrière, a species of Pedibibius (Hym., Chalichidida), Apanteles paratase Rohwer (Hym., Bravonidae), Gorphus oxymorus Tosquince (Hym., Ichneumonidae), Silbum sphanklum Fabricius and Chrysti shankhaimis Smith (Hym., Chrysidado), Chatevoritus januna Brauer and Bergenstamm (Dipt., Tachinidae) and Sarophaga antilope Bottcher (Dipt., Calliphoridae). The caterpillar of the most Physita dentilismalla is predacious on the chrysalis of P. Ispida in its cocoon, and the chrysalis is also susceptible to attack by an entomogenous fungus of the genus Cordycepe (Icepsene, 1947).

Contheyla rotunda Hampson

Economic aspect

This species, like some other members of the family Limacodidat (gf. Macrophiera narraio), is liable to occur in great numbers locally during the dry season. When the caterpillars have virtually defoliated the coconut palms, they turn their attention to the flower spikes and the developing nuts. Tall trees are most severely attacked, but when there is little foliage left on them the pest attacks even seedlings. Banana and tea are also damaged.

Geographic distribution

India, principally the western coastal zone.

Description | Biology

The eggs are deposited in groups of 4-15 on the lower surface of the leaves; the number produced per female averages 215. Incubation requires 3-6 days, development of the caterpillar about 5 weeks, and the subsequent development in the cocoon 10-14 days, so that the cycle covers about 52 days (Nirula, 1954). The moths are grayish brown, with a wing span of 16 mm in the male and 20 mm in the female.

Control

Spraying with DDT at 0.1 percent is recommended. The known parasites of C. rotunda are all hymenopterous: a species of Chrysis (Chrysididae) which may attain 20 percent parasitization, one of Roga: (Brasosidae), one of Gorphus (Ichunumoulae) and one of Antrosephalus (Chalcididae). During the monsoon season the caterpillars and chrysalides succumb to bacterial and fungous disease.

Chalcocelis albiguttata Snellen (formerly assigned to the genus Altha)

Economic aspect

The caterpillar feeds on the lower surface of the coconut leaflet and, in doing so, facilitates the entry of spores of the leaf-spot Inguss Patallop-liapiii palmarnm, which is more harmful to the leaves than the feeding of the insect itself. Other plants attacked are Atada, Alamites, coffee, tea, Erythrina, Enguina, Gardnia and Hydnacarput.

Geographic distribution

From Malaysia eastward to New Guinea.

Description | Biology

The flattened eggs are laid on the coconut leaflets, singly or in small groups of not more than four (Kalshoven, 1950-51). They develop rapidly and hatch in only 2-3 days. The further development is relatively slow, the caterpillar stage lasting 60-70 days and the chrysalis 21-28 days, making a total period of development of 12-14 weeks. The smooth, translucent, ovoid caterpillar has the appearance of a gelatinous mass. The cocoon is nearly spherical, about 10 mm in greatest diameter, and whitish, as if whitewashed (Dammerman, 1929). The adult is grayish brown, with a reddish spot on the forewing; the male is darker and smaller than the female, which has a wing span of about 32 mm.

Control

The treatment recommended against other limacodids is applicable in this case also but a fungicide should be added. The parasite complex of C. albiguttata includes Brachymria enplexar Westwood (Flym., Chalidadida), Fornitis thelaustelidii Wilkinson (Flym., Brauntidu), a species of Corphus (Flym., Itahumantidu), and Exertitae cervindes van der Wulp (Dipt., Takhindan).

Orthocraspeda catenatus Snellen (assigned by some authors to the genus Darna)

Economic aspect

This species is most harmful in relatively dry areas and seasons. In 1950, no less than 100,000 ecotout palms were seriously damaged by it in north-west Celebes; the defoliation reduced the yield of copra over a period of 18-24 months (Ponto and Mo, T.T., 1950). Metrosylun, Elasti and other palms are also attacked.

Geographic distribution

Celebes (and adjacent small islands) and New Guinea.

Description | Biology

Incubation of the egg requires 4 days. The caterpillar is brown above and green beneath, and is armed with two rows of spiny tubercles on each side. It attains full growth in about 24 days and then constructs a brown, nearly spherical cocoon which is about 7.5 mm in greatest diameter; the

moth emerges from the cocoon 10-14 days later, and the whole cycle from egg to adult covers about 6 weeks. The forewing is grayish ocher and has a black central lunule, and the head, thonax and hindwing are gray. The wing expanse is 15-19 mm in the male, 19-23 mm in the female. The female lays about 300 eggs (Expense, 1947; Kalshoven, 1950-51).

Control

When the number of caterpillars per leaflet reaches three on most of the leaves, all the leaves except the three or four youngest should be cut off and burned. It is important to initiate control measures while the areas of severe infestation are still small and well separated.

O. catnatus is parasitized by species of Euphetrus and Syntomosphyrum (Eulophidae), as well as by other hymenopterous insects of the genera Apartitis (Bracouldae) and Chrysis (Chrysididae), and by species of the dipterous genera Chantexorista and Bessa (Tachisidae). Bacterial disease sometimes destross up to 80 percent of the pest.

Trogocrada deleter Tams

A severe outbreak of this limacodid on coconur palms occurred in Mozambique in 1952 (Child, 1964). It is mentioned here mainly as an example of the sudden appearance in great numbers of an insect previously unknown as a pest or (as in this case) previously not believed to exist. The cause of the outbreak in Mozambique is not known, but it was suggested that exceptional flooding which preceded the outbreak may have eliminated, at least temporarily, the normal host plants of the insect and so caused it to attack the palms.

Castniidae

Castnia daedalus Cramer

Economic aspect

The large caterpillars of this moth burrow between the leaf bases and the trunk of the palm, causing the older leaves to hang down the trunk and the nuts to fall off through lack of adequate support. The continuous feeding of the caterpillars causes longitudinal furrows on the trunk which sometimes attain a length of a meter or more (Figs. 55 and 56) (Bodkin, 1913). The most serious damage occurs in plantations on clay soils. Roytones, Mauriti and banana are also liable to be attacked.

Geographic distribution

The Guianas and the Amazon basin.

Description | Biology

The caterpillars are whitish, and 100 mm long when fully grown. The clongate cocoon is made of coarse fibers from the leaf bases. The moth is dark brown, with a whitish bar; it is crescent on the forewing and has two rows of spots on the hindwing. Its wing span is 140-180 mm (Figs. 53 and 54).

Control

Spraying with a 5 percent solution of dieldrin at 4-5 liters per tree is advocated. Kennard (1960) used dieldrin at only 1 percent, however, applying it directly to the center of the crown of the tree by means of stirrup pumps and long hoses, the latter attached to bambou poles.

Castnia licus Drury

Liconomic aspect

This is a somewhat smaller species of Castnia but it does the same type of damage to the palms as C. daedalus. Young palms are the worst affected; their leaves become yellow (Wolcott, 1933). Banana and sugarcane are also attacked.

Geographic distribution

Central and South America, and Trinidad.

Description

The adult is similar to C. deedulus, but the whitish markings on the forewing are relatively larger and wide bands replace the spots on the hindwing. The wing expanse is 90-110 mm.

Control

For this species, dieldrin at 1 percent in solution applied at 2.5 liters per tree is claimed to suffice.

ZYGAENIDAE

Brachartona catoxantha Hampson (often referred to as Artona catoxantha)

Economic aspect

The destruction of leaf tissue of the coconut palm by the caterpillars of this small moth can be so severe as to result in the complete loss of a crop for about 18 months (van der Vecht, 1947). Between outbreaks this insect becomes extremely rare or nonexistent in the coconut planatsions, and it is assumed to be confined at such times to localized areas in the jungle from which it is intermittently transported by the wind. All investigators agree that severe infestations usually become obvious in dry weather and end soon after the onset of the monsoon season with its strong winds and torrential rains; this applies in the Philippines (Merino, 1938), Indonesia (van der Vecht, 1950) and the Malay Peninsula (Gater, 1925; Lever, 1953), and the outbreaks develop and terminate relatively suddenly. In an examination of the dates of 48 Malayan outbreaks (Lever, 1953) it was shown that 36 occurred in the drier half of the year (January-June) and only 12 in the wetter half. Figures 57 and 58 illustrate typical damage due to this interest.

B. catosamtha attacks other palms besides coconut: Nypa, Elanis, Areas, Merosylan, Nonosperan, Zalacas, Calamus. It also feeds on banana. It should be noted, however, that whereas Metrocylon, like coconut, appears to be a normal food plant and outbreaks sometimes originate on it, other plants are, in general, not attacked unless the normal food (Metrocylon and coconut) becomes scarce through heavy infestation (van der Vecht, 1950).

Geographic distribution

Malaysia, the Philippines, New Guinea.7

Description Biology

The eggs are yellow, somewhat flattened and broadly oval in outline. They are laid singly or, more frequently, in irregular groups of 3 to 12, on the underside of the leaflets, and the usual number produced per female is 40-50. They hatch after 3-5 days. The caterpillar is pale in color with a conspicuous, dark purplish, middorsal line and less dark sidelines. There are three rows of tubercles along each side, each row being represented by one tubercle on each segment; each tubercle bears numerous short bris-

⁷ A report of the occurrence of this species in Fiji (Anon., 1962) is erroneous.





FIGURE 53. - Castnia daedalus Cramer. Female with some eggs. Surinam.

FIGURE 54. - Caterpillar, chrysalis (right), empty chrysalis and empty cocoon (left).

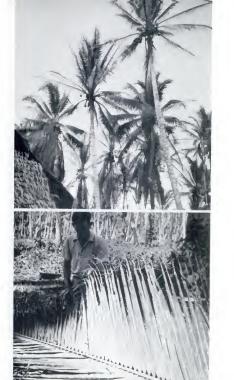
(Courtey Royal Tropical Institute, Amsterdam)





FIGURES 55-56. - Cattnia duedalus Cramer. Trunk of ecconut tree damaged by caterpillars (55, left) and halved trunk, showing rotting precess at top (56, right). Surinant (Contray Reyal Tropical Institute, Amsterdam)

FIGURES 57-58. - Brachartona catassantha Hampson. Injury to ecocoust palms by larval feeding (57, above) and detail of damage to leaves (58, below). Malaysia. (Photos R.J.-t. W. Lerer)



tles. The first segment, into which the head is retracted and which hides it completely during feeding and resting, is large and dorsally brown. About 3 weeks after hatching the caterpillar is fully grown and about 10 mm long. The feeding scars on the underside of the leaflets are very characteristic, being short, parallel-sided furrows bounded by principal veins of the leaflet; the upper epidermis and the larger crossveins are not eaten at this stage. and the scars resemble ladders, with the crossveins as rungs. When the caterpillar reaches the last of its five instars, however, it feeds at the edge of the leaflet and eats right through it. The chrysalis is formed in an oval, somewhat flattened whitish cocoon, 12-14 mm long and of a papery texture. The cocoons - often many clustered together - are formed mainly at the bases of the leaflets but also at the bases of the leaves, or, in the case of caterpillars which fall or make their way to the ground. on undergrowth or debris. The period spent in the cocoon is 9 days, and the complete cycle from egg to emergence of the moth is about 51/4 weeks, from which it follows that there can be about 9 generations a year in suitable conditions. The moths, which are 13-16 mm in wing expanse, have dark brown wings and body; the wings are narrowly yellowbordered but without markings, and are vellow on the underside. When the moth is at rest, the front of the body is held away from the surface on which it is resting at an angle of about 60°. Both sexes are active in the early morning and the afternoon, and in outbreaks can be seen in great numbers in and around the crowns of the trees and feeding on the coconut flowers (Kalshoven, 1950-51).

Control

Insecticidal treatment against Brabartons was first undertaken in Java, with pyrethrum and later derris as the basis of the formulations used. Experience resulted in the recommendation of a derris-talc suspension containing 10 percent of derris (which gave 0.005 percent rotenone in the suspension) (Lecfmans and Awibowo, 1935). Treatments recommended in the Malay Peninsula much later included spraying with lead arsenate at 0.4 percent, gamma-ance at 0.025 percent, or Dar at 0.1 percent, and the application of DDT at 12.5 percent as a fog at about 17 liters per hectare. Emphasis was always laid in the Malay Peninsula, however, on the necessity to refrain from insecticidal treatment unless the outbreak was still confined to a small area and the degree of parasitization by the tachinid hy Bestas remota was below 70 percent in caterpillars of the relevant age.

Early action, before the infestation becomes unmanageable, is important, and this depends on proper inspection of the trees, by means of field glasses where the trees are tall. The use of an insecticide less persistent than BHC, such as malathion, has the advantage of having a correspondingly less adverse effect on the parasites.

The parasite complex of Brachartona has been intensively studied, partly in connection with a need for biological control of other harmful species of the family Zygaenidae and of B. catoxantha itself when it becomes established in islands or areas from which it has previously been absent, and also to gain an understanding of the factors concerned in the causation of outbreaks and the population dynamics of the pest. In this last respect, particularly noteworthy investigations were carried out in Java (van der Vecht, 1947 and subsequent papers).

The principal parasites of the complex are the following:

Besta remedia Aldrich (formerly Pythomyia remedia). This is a tachinid which lays one or several eggs on the thoracis esgements of the half-grown caterpillar. Its developmental cycle lasts about 25 days and is shorter by 12-15 days than that of its host. In areas of the Malay Peninsula, where parasitization by B. remed reaches its maximum, virtually all Brathertona caterpillars of suitable age are eliminated by it at approximately the same time, with the result that an interval is established between generations, whereas otherwise there would be complete overlapping. This tachinid is more effective in the Malay Peninsula than in Indonesia. It was introduced into Fiji to control Levama iridexens. A secondary parasite, Pulcibibia paranta Ferrière (Flym, Chalcibiado) often attacks.

Cadarcia lefmansi Baranov (formerly Degeria albierps Macquart). This is another tachinid. It is particularly effective in Java toward the end of outbreaks and is almost absent from the coconut plantations when Brachartoma is scarce.

Apantetes artonae Wilkinson. This braconid plays an important part in the natural limitation of populations of Brachartone in Java. By parasitizing almost all the caterpillars at approximately the same time (just as Betsa does in the Malay Peninsula) it creates conditions unfavorable for its own survival because complete overlapping of generations no longer occurs. It is often heavily attacked by a secondary parasite of the genus Ceraphron (Hym., Practhrapsida).

Other parasites that attack the caterpillars commonly are species of Euphetromorphs and Eupherms (Hym., Chalitabides), while the chrysalides are
destroyed by two species of Goryphus (Hym., Ichnomonidus). Euphetromorpha
attains about 40 percent parasitration when Brachariona is race but (unlike
Cadurcia) almont disappears when it is common (van der Vecht, 1950).

A beetle, Callimerus arcufer Chapin (Cleridae), is a predatory enemy of Brachartona, which certainly contributes to its control. It too was introduced into Fiji for the control of Levuana.

A fungus, Beameria bassiana (formerly called Batyris necast), infects and kills the caterpillars. An outbreak in Singapore was brought to an end by it, but the combination of weather conditions that favor it seems to be unusual. Attempts to utilize it for control purposes have not been successful.

Levuana iridescens Bethune-Baker

Economic aspect

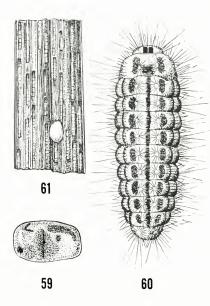
The caterpillars of this species do the same type of damage to the coconut palm as those of Brachartona, which they resemble in appearance as well as behavior. This insect was formerly a very severe pest in Fiji, to which it was confined, and did in fact constitute a serious threat to the coconut industry as a whole. It is, however, no longer a pest, having been brought under permanent control in 1925 by the introduction of a parasitic fly from the Malay Peninsula. Small local outbreaks still occur occasionally but they are quickly suppressed and are almost negligible. Nevertheless, there is always the risk of unintentional transportation of the insect to other countries; a possibility that the control achieved in Fiji may yet have to be applied elsewhere must therefore be recognized. Figures 63 and 64 illustrate typical damage done by Levuana on coconut. Other palms attacked by it are Metroxylon vitiensis and Veitchia joannis, which are indigenous to Fiji, and various introduced species of the genera Roystonea, Elaeis, Livistona, Areca and others. In severe outbreaks, caterpillars were found feeding on banana and sugarcane, but these plants are not to be regarded as true host plants.

Geographic distribution

Fiji Islands only. Originally confined to the largest island, Viti Levu, it had spread to certain other islands before it was brought under control.

Description | Biology

The eggs (Fig. 59) are opalescent and whitish when first laid, and are variable in shape but in general barrel-shaped. They are deposited on the lower surface of the leaflets in clusters of 10 to 40, and hatch in 7.8 days. The caterpillar (Fig. 60) is whitish with two dark purplish, longitudinal bands on the back, one on either side of the middle line. The first segment is large and covers the head, as in *Dirachartoma, and has a blackish area dorsally divided by a pale median line. The body is armed with three rows of tubercles, each bearing numerous short bristles along each side; every row is represented on each segment by a single tubercle. The length of the fully grown caterpillar is 13 mm, and the duration of the caterpillar stage is 26-30 days. A noteworthy difference between this caterpillar and *Dirachartoma* is that whereas the latter, when nearly full-grown, feeds at the edge of the leaflet and east right through it, *Lamana* feeds throughout the caterpillar stage by making parallel-sided furrows in the leaflet from below in the manner already described for all but the last instant of *Birachartoma* instant or *Birachartoma* instant



Figures 59-61. - Leviuma iridescens Bethune-Baker. Egg (59); adult caterpillar (60); larval damage and a cocoon on leaf (61). According to Tothill, Taylor and Paine, 1930, redrawn.

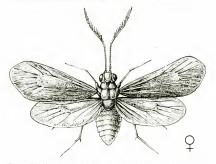


FIGURE 62. - Levnana iridescens Bethune-Baker. Female moth. According to Tothill, Taylor and Paine, 1930, redrawn.

chartona. When feeding is completed, a whitish cocoon is formed, just as in Bradhertona, and it is similar in shape and texture to that of the latter, but whereas in Bradhertona the majority of cocoons are constructed at the bases of the leaflets, in Lermone the majority of them are situated in the crevice between the base of the perioks of the leaf and the fibrous sheath of the succeeding leaf (Fig. 61). The duration of the cocoon stage is 10-13 days. The whole developmental cycle from egg to moto occupies about 43 and 51 days in the hot season and cool season, respectively. The moth is a dark metallic purple color, without markings; the metallic sheen is particularly noticeable on the head and thorax. The male measures 13-15 mm in wing expanse, the female 15-17 mm (Fig. 62) (Tothill, Taylor and Panie, 1930).

Control

In 1925, the tachinid fly Bessa remota Aldrich was introduced into Fiji from the Malay Peninsula and within only six months it had multiplied



FIGURES 63-64. - Fiji. Lenuma iridusens Bethune-Baker. General view of a coconut palm tree, showing larval damage (63, above), and detail of damage to fronds (64, below).

(Plate R J & W J. Legy)



to so great an extent that the Lemma populations had been reduced to negligible dimensions. This is regarded as one of the classic examples of successful biological control. It resulted from the breeding in captivity and subsequent liberation of 32,600 flies in 38 localities over a period of four months. Proof was obtained that the fly could traverse a distance of 12 miles (19 kilometers) over open sea, and there seems little doubt that it could go much further. A predactious beetle, Calliment arrayfer Chaptic (Cheridae), which attacks Brashartona, was introduced into Fiji from the Malay Peninsula with Betas remats but failed to become established.

Chalconycles catori Jordan (sometimes assigned to the genus Homophylotis)

Economic aspect

This West African species is injurious to coconut palms less than 15 years of age, especially near the sea. However, *Elaeis* is preferred.

Geographic distribution

Ivory Coast and Ghana.

Biology | Control

Incubation occupies 6-13 days, the caterpillar period 22-33 days, the chrysalis 10-14; the mean total period from egg to adult is about 7 weeks. For control, aerial spraying of DDT at 0.037 percent is recommended. This should be done after rains, at which time the caterpillar population increases (Cachan, 1959).

NOCTUIDAE

Cyclodes omma van der Hoeven

The caterpillars bore deeply into the young nuts and cause them to fall off, The species is not very common and has never been studied adequately, but it is clearly potentially dangerous. It attacks other species of palms also (Lepesme, 1947), and has been recorded from India, Ceylon, Indonesia, Cambodia, Laos and Viet-Nam. Spraying with dieldrin is recommended as a control measure.

Coleoptera

LYMEXYLONIDAE

Melittomma insulare Fairmaire

Economic aspect

This wood-boring beetle, belonging to a little-known family, is the principal limiting factor in coconut growing in the Seychelles (Vesey-Fitzgerald, 1941b). The larvae excavate tunnels in the central portion of the trunk near the ground, and the trunks of attacked reces may be so weakened that they fall down. Wild palms of the genera Stevensia's and Nephrapperma, and more rarely Deckenis and Reacheria, are also attacked; they were the host plants of M. insularo before the coconut palm was introduced into the islands. In this connection it is interesting to note that two other local palms, Loshies and Vershulgilinia, are not attacked (Brown, 1954). Vigorous, well-manured trees are said to be more liable to attack than less healthy ones (Nye, 1961), and trees 7 to 20 years old are the most severely infeated.

Geographic distribution

Seychelles and Madagascar.

Description | Biology

The white eggs are deposited in masses of 100 or more in crevices in the trunk. They hack in 11 days. The larva is white and, when fully grown, 20 mm long; the first segment is large and strongly chitinized and the last segment has a hoof-shaped chitinized plate in which there are 18 or more small pits (Figs. 65 and 66). The young larvae enter the bole at the points where roots arise, and many larvae (up to about 200) may be found together in the same trunk. The soft wood at the heart of the trunk is preferred to the hard outer layers, and the larvae tunnel upward in it. A severely attacked bole is reduced to a black pulpy mass. The wood itself is not eaten; the juice is extracted from it and characteristic yellowish browr pelletis issue from the boreholes. The pupal stage is passed in a cell excavated in the wood. The pupa is white, 17 mm long, and provided with thorny spines. The adult (Fig. 67), which is cinnamon brown in color varies greatly in size, from 6 to 13 mm long in the male and 9 to 18 mr in the female.

The adults do not feed; the males, which are more active, live 6 days and the females only 3 or 4 days.

This insect occurs on islands of granitic origin, not on those composed

 $^{^{8}}$ The beetle illustrated by Lepesme (1947) as M. incidere is actually another species the same family.

of coral, and the most severe infestations are in localities where the trunks of the trees have a high water content (Brown, 1954). Palm trunks lying on the ground, having been caused to fall by the ravages of Melitimmun, provide scope for the breeding of other pests, including Orgetes, termites and a weevil, and should therefore be destroyed by burning.

Control

Funigation of the infested boles by inserting paradichlorobenzene into the cavities and covering their entrances gave fairly good results (Brown, 1954), but it was eventually decided that the resultant mortality of the larvae was not high enough. The best method is to excise the necrotic wood and expose the entrances of the tunnels, care being taken not to remove more of the hard outer wood than is necessary, and then to apply a mixture of tar cresoste (60 percent), coke residue (9 percent), water (3 percent) and an inter, nonsoluble material. If excrement of the larvae is sull appearing after a week, a second application is necessary. By this method, the loss of trees has been reduced to 1 percent and infestation of trees from 80 to 18 percent (Nye, 1961).

Much consideration has been given to the possibility of biological control, but the conclusion has been reached that this method is unlikely to succeed in this case. A predatory beetle of the family Nitidulidae was introduced but has not given satisfactory results (Simmonds, F.J., 1956).

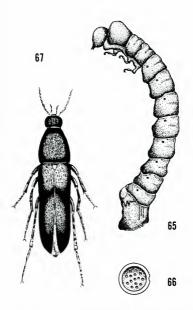
HISPIDAE

In an important analysis of the damage done by beetles of this family, Kalshoven (1957) has shown that they have an association with monocotyledonous plants, particularly palms, zingiberaceous plants, bamboos and orchids. These produce an almost continuous succession of terminal shoots which provide the various hispid species with the leaves, stalks or unopened "hearts" that they need.

Promecotheca coeruleipennis Blanchard (This insect, as a coconut pest, has long been referred to as P. reichei Baly).

Economic aspect

The species of the genus Promeosthera, the adults of most of which are brightly colored, cause very serious damage of two quite distinct kinds to the leaves of coconur palms: the larvae feed by mining the tissues of the basal half of the leaftes (Figs. 68 and 69), whereas the adults feed by making narrow grooves in the lower surface of the distal half of the leaflets. Thus, in severe infestations, the larvae and adults jointly make the greater part of each leafte and therefore of each leaf nonfunctional, and



FIGURES 65-67. - Melistomma inculare Fairmaire. Larva (65) with anal plate (66), and adult (67).

(Courtery Institut de recherches pour les builes et les oliegineux, Paris, France)

since all the leaves are damaged in the same way, affected trees are greatly weakened and the crop is drastically reduced. The damaged leaflets shrivel and readily tear and break in the wind, and the trees acquire a brownish gray color which makes severe outbreaks recognizable at a considerable distance. Alternative food plants are palms of the genera Pritchardia, Livitions and Metraxylon.

Geographic distribution

Fiji, Tonga, Samoa, Futuna and Wallis (Taylor, 1937).9

Description | Biology

The eggs are laid singly on the lower surface of the leaflet. Each is covered by a close-fitting brown capsule composed of semidigested leaf fragments cemented together. The number of eggs laid per female is, on average, only 20. Incubation occupies 15-19 days, according to season. The newly hatched larva penetrates directly into the leaf tissues beneath the egg capsule and proceeds to excavate a mine toward the apex of the leaflet, between the upper and lower epidermes (Fig. 68). In the course of its development, the larva molts three times, and it widens the mine as it grows in the first two instars, but not in the third. The typical mine is straight, about 150 mm long and about 10 mm wide over the greater part of its length; thus each larva destroys about 12 cm2 of leaf. The fully grown larva is 9.5 mm long, vellow, markedly flattened and legless, and the first segment is very large and bears a shiny, brown, strongly chitinized triangular plate, both above and below. The mean duration of the larval stage is 43 days in the hot season and 52 in the cool season. The pupa is formed in the mine and the pupal stage lasts about 12 and 18 days in the two seasons, respectively. On emergence from the pupa, the adult beetle rests for about 3 days inside the mine and then escapes by biting a conspicuous oval hole in the upper wall of the mine. The total duration of the developmental period, from egg to emergence of the adult from the mine, is about 10 and 13 weeks in the hot and cool seasons, respectively. A further period of about 13-16 days elapses before the female begins egg laying, and the total duration of adult life in both sexes is about a month. The adult is 8 mm long; its anterior half is orangeyellow and its posterior half metallic blue. The beetles fly by day and when abundant they can be seen on sunny days flying slowly around the crowns of the palms. The population of adults per tree in outbreak conditions is approximately 4,000 (Taylor, 1937).



⁹ Reports of the occurrence of this insect in Tahiti and Hawaii (Wyniger, 1962) and in the Solomons and the Philippines (Kalshoven, 1957) are erroneous.

Control

Chemical control of this insect has never been attempted, biological control having been successfully achieved in 1933.

Taylor (1937) studied the natural enemies already present in Fiji prior to 1933. The more important species were: a parasite of the egg, Oligosita utilis Kowalski (Hym., Chalcidoidea, Trichogrammatidae); a parasite of the larva, Elasmus hispidarum Ferrière (Hym., Chalcidoidea, Elasmidae); and a predacious mite, Pyemotes ventricosus Newport 10 (Acarina, Pyemotidae), which attacks larvae of all ages and pupae. The first two of these, which are undoubtedly indigenous to Fiji, seem to have kept. P. coeruleipennis well under control until the unintentional butin evitable introduction into Fiji of the mite P. ventricosus, which is well known as a nearly cosmopolitan predator of many different kinds of soft-bodied insects that live in sheltered situations, especially insects that are pests of stored products. This mite multiplies extremely rapidly in favorable circumstances, which evidently include those prevailing in the mines of Promecotheca during the dry season, and it was found to be destroying virtually all the larvae and pupae in their mines during that season, leaving only the adult beetles and eggs. This established a "one-stage condition" in the population of Promecotheca. such that the indigenous parasites were unable to find larvae and pupae on which to breed, and consequently almost died out. When the next wet season began, the mite was no longer able to multiply, since the wet leaves, often-flooded mines and high humidity were highly unfavorable to it; thus, the beetle bred virtually without any natural controlling agencies throughout the wet season and attained once more a very high and harmful level of population. The mite, in destroying over a limited period all the individuals of the preadult stages - an activity which, if sustained, would be regarded as very satisfactory biological control, did in fact cause Promecotheca to become a much more serious pest than it would otherwise have been.

It became clear that to restore biological control a parasite fulfilling certain specifiable requirements was needed. Such a parasite was found in Java, in the form of Pediabina parulus Ferrière (Hym., Chalcidolidea, Eulaphidae) (at that time placed in the genus Phunorophi); this insect, which was parasitizing Promeethies cumingii Baly in Java, was introduced into Fiji, and there proved able immediately to bring Promeethies under control, mainly through its ability to survive, unlike the indigenous parasites, from one Promeethica generation to the next, even in the one-stage condition established by the mite. In the course of about a year, 26,000 individuals of P. parulus were erared in the laboratory and were liberated in

¹⁰ Formerly assigned to the genus Pediculoides.

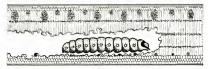


FIGURE 68. - Promecotheca species. Larva in mine (leaf cut lengthwise).

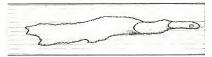


Figure 69. - Leaf mine of P, coernleipennis Blanchard, showing egg capsule and subsequent development.

about 50 different localities; this proved more than sufficient (Taylor, 1937).

This became a classic example of planned biological control of a serious pest by a parasite specially chosen to fulfill the conditions prevailing in its new home.

Promecotheca papuana Csiki (also recorded as P. antiqua Weise)

Economic aspect

This is another species which, as a larva, mines in the leaves of the coconut palm. It is a pest in New Britain and Manus only, though it occurs over a much wider area. It is most abundant on mature palms. As in P. coevulit-pennit, the adults feed on and kill the distal parts of the leaflets and they and the larvae are supplementary in causing serious defoliation and consequent loss of crop. The central shoot of the palm may be killed by constant attack by the adults as the new leaves unfur!. Constrictions in the trunks of old palms of known age indicate the dates of past outbreaks; it is judged that they have occurred at intervals of 10-15 years. Metrocylan, Nam. Area and Elaki are also attacked.



FIGURE 70. - Promeesthera species. Apical frilling of fronds by feeding of larvae and adult beetles of P. aumingi Baly. Singapore.

(Photo R, I, A, W. Lever)

Geographic distribution

New Guinea, Manus (Admiralty Islands) and the Bismarck Archipelago.

Description | Biology

This species differs from *P. coeruleipnomis* in that it lays 3-5 eggs together and covers them with a single capsule, but the capsules of the two species are similar in that they are composed of partially digested leaf fragments cemented together. The mean incubation period is 13½ days. The larvae

from each capsule mine together in the leaf tissues, making a large communal mine by feeding and advancing side by side. A typical completed mine is about 39 cm long. The larval stage lasts 17-30 days. When full-grown, the larva, being 12 mm long, is appreciably larger than that of P. coeruleipenmis, but otherwise it is similar. Pupation takes place in the mine and the adults emerge from it 11-12 days later. The total development period, to emergence from the mine, including incubation, is thus about 45-48 days. which is considerably shorter than the corresponding period (70-91 days) for P. coeruleipennis; however, the following period, to commencement of egg laving, is much longer in P. papuana (4 weeks, as compared with about 2 weeks), and whereas the total duration of adult life in the Fiji species is about a month, that of P. papuana is about 5 months. The female lays 90-100 eggs in about 30 capsules. The coloration of the adult is black on the head and thorax, reddish on the anterior three fifths of the elytra (forewings) and greenish, purplish or bluish black on the remainder; in length the adult measures 7.5-9 mm. A peak population of 3,000 beetles per leaf and 35,000 per tree has been estimated (Froggatt, 1939a; O'Connor, 1940; Gressitt, 1959).

Control

As a result of a particularly severe outbreak of this pest in New Britain, Pudishist paramiac (the parasite used for the control of Promeethra in Fiji) was introduced from Fiji to Rabaul in 1938. The parasite became established quickly and was entirely successful. The coconut crop, which had been greatly reduced, was restored to normal.

Several indigenous parasites of *P. papmana* exist in New Britain, the most important being an egg parasite, *Clositrocerus splendens* Kowalski, and the larval parasites *Apleuroropis* lalori Girault and *Eurytoma promecolhecat* Ferrière (*Hym.*, *Chalidolata*).

Promecotheca opacicollis Gestro

Economic aspect

This species is considered the most important pest of coconut palms in the New Hebrides. The adults and larvae do the same type of damage as those of the other species of Promeoline already described, and the damage to the tissues allows the entry of the fungus Pettalophispit palmarum, which causes further injury. The greatest damage is done in the dry season. Risbec . (1937) expressed the opinion that the injury due to this insect in the New Hebrides is much less than had previously been attributed to it. In the Santa Cruz Islands, so the north, it is certainly serious on occasion (Lever,

1933a); legislation promulgated in 1938 forbids the transportation of coconut leaves to the main Solomon Islands to the west. Other food plants of *Ps. opaciollis* are *Areca, Phoenix, Ravenda* and *Phytelephas*.

Geographic distribution

The New Hebrides and the Santa Cruz Islands of the Solomons.11

Description | Biology

The eggs are laid on the lower surface of the leaflets, under capsules, each of which usually contains 4 eggs. They harch in 13 days. The larvae from each capsule feed communally in a single mine, and there are four larval instars in this species (unlike P. cearuli-promis and P. papuana, which have three only). Risbec (1935) estimated that the ratio of quantities of leaf tissue consumed by the larva in the four instars is 14-1100-6,000. The larva is 8 mm long when fully grown, and the duration of this stage is 25 days and that of the pupal stage 10 days. The total period from egg to adult is thus about 48 days. The adult female is 10.5 mm long, the male a little smaller; the thorax is dark in color as in P. papuana and the coloration of the elyra is similar to that of P. cearuli-pionii, except that the anterior border of the posterior purplish part bends forward in the middle like a circumfex accent instead of being straight.

Control

Chemical control should be used only in the absence of parasites.

The parasite Oligotia utilit Kowalski (Hym., Chalidaida, Trichogrammatika) destroys up to 40 percent of the eggs; according to Risbee (1937) the lowest egg in the capsule cannot be reached by the ovipositor of the parasite. Clestracerus splendent Kowalski (Hym., Chalidaida, Eulophidae) destroys the larvae in the mines but parasitization by it reaches only 6 percent.

Promecotheca cumingi Baly (also referred to as P. nuciferae Maulik)

Economic aspect

This is one of the most important pests (Fig. 70) of coconut palms in the Philippines. Damage has also occurred in the Malay Peninsula (Burkhill, 1918) and Singapore (Lever, 1951). The damage, again of two kinds, is the greater because it allows the entry of spores of the fungus Pestaloptiopsis.

 $^{^{11}\,\}mathrm{The}$ statement by Kurian (1963) that this species occurs in the main islands of the Solomons group is erroneous.

The insect is also harmful to other palms, including Areca, Elaeis, Nypa, Metroxylon and Roystonea.

Geographic distribution

The Philippines, the Malay Peninsula, Singapore, Borneo, south Celebes, Java.

Description | Biology

Morphologically and biologically, P. coming is very similar to the other species of the genus already described. The incubation, larval and pupal periods are 13-15 days, 3-4 weeks and 8-12 days, and the total cycle from egg to adult emergence is 7-8 weeks. The length of the mine is about 20 cm, The bettles are uniformly reddish brown; they are 7-8 mm long.

Control

Chemical control is rarely undertaken, but Wyniger (1962) recommended a wettable powder of DDT or dieldrin at, respectively, 0.3 and 0.2 percent; alternatively, diazinon, guthion or parathion can be employed.

In Singapore, an outbreak which followed the inadvertent introduction of this pest in 1951/52 was rapidly brought under control by the parasite Peduhin paradus Ferrière (Hym., Chakitakida, Eulophida) which was evidently already present although previously unknown there. Other parasites which attacked it in Singapore were Diamackia justuitus Ferrière and Arbiyuscharis promenshesse Ferrière (both Eulophidas), the former parasitizing the larvae and the latter the eggs.

At the time of the outbreaks in the Philippines, some 35 years ago, it was recommended that ten old leaves (the 7th to the 16th) should be cut off from each affected tree; this drastic treatment was considered to be no more severe than the damage done by P. cumingi (Aldaba, 1931). This is mentioned here to illustrate the tolerance of the coconut palm to such measures. The trees recovered completely in two years.

Coelaenomenodera elaeidis Maulik

Certain species of the genus Coslamonenodora occasionally mine as larvae in the leaves of the occount palm in western Africa and in the Congo. The most important is C. slatistic which, however, is more a pest of oil palms (Elasis) than of coconut palms. It resembles Pronucativas both in behavior and in the complex of parasites which attack it; species of Clustrectura and Athrymbarist destroy up to 60 percent of the eggs, and other culophids,

of the genera Dimmockia and Cotterellia, parasitize the larvae (Cotterell, 1925).

Dusting from the air with BHC and spraying from the ground with dieldrin, together with cutting and burning infested leaves, are effective.

Himatidium spp.

At least five species of *Himatidium* occur in Brazil, where they attack the central leaf bud, the young leaves and the fruits of the coconut palm. The fruits are particularly susceptible when they are in contact with one another. Dwarf palms are injured most severely, and may suffer a very heavy loss of croy (Bondar, 1940b).

Brontispa longissima Gestro (formerly called B. froggatti Sharp)

The genus Broutipe comprises at least a dozen species which have been recorded as actual or potential pests of the coconut palm. Some of these are probably merely local forms of more widely distributed species. All of thems are similar in habits and natural enemies and the three species described below can be regarded as trypical.

Economic aspect

B. longissima is a small, elongate, flattened beetle well adapted to live, as it does, as both adult and larva between adjacent leaflets in the central leaf bud (cabbage) before the leaves separate and the leaflets unfold. Thus, the species of this genus are not true miners even as larvae, and in this respect differ fundamentally from the species of Promecotheca. The adults and the larvae feed on both surfaces of the closely appressed leaflets, and both stages gnaw long incisions in the tissues, parallel to one another and to the veins of the leaflets. When the insects are numerous, these incisions are so close to one another that the whole of the attacked part of the leaflet dies; all the leaflets are similarly injured, and photosynthesis is reduced almost to zero (Figs. 75 and 76). Young palms less than about 8 years old suffer most. B. longissima has been recognized as a pest in Melanesia for more than half a century and, until the advent of persistent insecticides, was responsible for severe damage and sometimes total losses in most palm nurseries. It breeds on many other kinds of palms, including Areca, Elaeis, Caryota, Latania, Metroxylon, Phoenix, Ptychosperma, Roystonea and Washingtonia.

Geographic distribution

Indonesia (including the Moluccas), New Guinea, the Solomons, New Hebrides, New Caledonia, and (since 1961) Tahiti. The arrival of this insect in Tahiti constitutes a serious threat to all the coconut plantations of Polynesia (Cohic, 1961).¹²

Description | Biology

The eggs are brown and flat, and are laid in short chains of up to 4, and sometimes singly, on the unexpanded leaflets in the bud. They are not protected by a capsule but are loosely surrounded by leaf debris and excrement (Figs. 71 and 72). They hatch in 3-4 days. The whitish larva has a pair of prominent caliper-like hooks at the hind end (Fig. 73). It becomes fully fed in about 5 weeks, though the actual period varies greatly in different countries or seasons (probably with temperature), as is indicated by the following figures: 33-54 days in the New Hebrides (Risbec, 1935); 30-40 days in New Guinea (Froggatt and O'Connor, 1941); 30 days on average in the Solomons (Lever, 1935); and 23-43 days in Celebes (Awibowo, 1935). The pupa moves freely, by wriggling, between the apposed surfaces of the developing leaflets, and the adult (Fig. 74) emerges from it in 4-6 days. The whole cycle from egg to adult occupies about 5-7 weeks in Indonesia (Kalshoven, 1950-51, referring to observations made in Java and Celebes), but can extend to 9 weeks in some other (presumably cooler) places. The female matures sexually in about 2 more weeks, and then begins egg laying and produces on average 120 eggs in the course of several weeks (Kalshoven, 1950-51). The adult beetle is 8-12 mm long; its color varies geographically from reddish brown in Java to more or less black in the Solomons and New Guinea. The more extreme color forms were long regarded as distinct species: selebensis mainly in Celebes and the Moluccas, froggatti mainly in the Solomons, New Hebrides and New Caledonia, and simmondsi in the Bismarcks; but there is considerable overlapping in the distribution of these forms.

Control

Thirty years ago, it was usual to combat this pest by applying by hand a concoction of lead arsenate and bordeaux mixture or of tobacco and soap, but the results obtained were poor (Pagden and Lever, 1935). Nowadays, infested trees are sprayed every 4-6 weeks with a solution of 0.15 percent dieldrin by means of low-volume knapsack sprayers. Good results have also been obtained with nor at 0.2 percent and chlordane at 0.16 percent, but

¹² The inclusion of Fiji in its range (Kalshoven, 1957) is incorrect.

these products are less persistent than dieldrin. Experience in the Solomon Islands has indicated that the annual cost of modern control procedure is the equivalent of U.S.59 per hectare, of which 80 percent is the cost of labor; small palms 3-4 years old are sprayed once every two months with dieldrin at 0.15 percent by means of a pressurized sprayer and aluminum lances 4 meters long.

The parasite comples of B. longitima comprises three egg parasites, Hackeliana brontipar Fetrière, Trichogrammatoida nana Zehntner (both Hym., Chalidoida, Trichogrammatoida) and a species of Osençrina (Hym., Chalidoida, Eincyridad), and a parasite of the larvae and pupe, Tetratichus brontipa Fetrière (Hym., Chalidoidae, Endophidae). The Tetratichus is the most effective; control of the beetle was achieved in Celebes by introducing this parasite from Java (Awibowo, 1935), but an attempt to introduce it into the Solomons in 1936 failed (Lever, 1936). The developmental cycle of Tetratichus occupies 16-23 days (half the period occupied by that of its hoat) and many Tenatichus develop in each attacked individual of Brontipa. In Java there are two strains of Brontipa, of which only one permits the complete development of Tetratichus (Mo. T.T., 1965).

Brontispa mariana Spaeth

Economic aspect

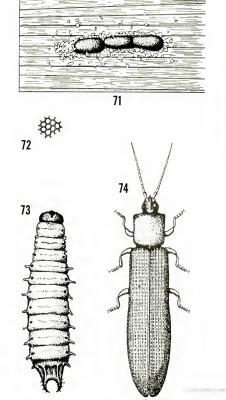
The species mariana is the representative of the genus Brontipa in Micronesia and causes serious damage to occonut palms in Saipan and Rota in the Mariana Islands, and in Truk, Yap and elsewhere in the Carolines. Between 1931 and 1936, it almost eliminated the copra and coconut oil industry of the Islands. Coconut is the only known host plant in the field but it has been reared on Area and Euprithardia (Lange, 1950).

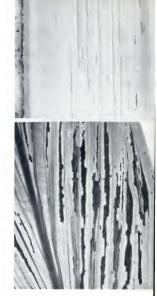
Geographic distribution

The Mariana and Caroline Islands.

Biology

The eggs are laid in groups of 2 to 6 between the unopened leafets. They hatch in 6 days. The larval development occupies on average 22 days and the pupal 5 days, giving a total from egg to adult of about 33 days, which is less than that of B. lengistima. The adults live for several months and the female lays over 100 eggs in the course of its life.





FIGURES 75-76, - Brontipa longistima Gestro. Fresh feeding scars on leaflet (75, above) and opened-out frond, showing damage to leaflet (76, below). Solomon Islands.

(Photos E.S. Brown and H.A. Green, 1958)

FIGURES 71-74. - Brontipa longissima Gestro. Eggs and frass on leaflet (71); detail of pattern on chorton (72); and larva (73). According to E.S. Brown and H.A. Green, 1958, redrawn. Adult beetle (74).

Control

No chemical control measures have been recorded but spraying with dieldrin at 0.15 percent should be effective. For biological control, Titratithus Irontitipus and Hawkilman Irontipus, both already mentioned, were introduced into Saipan in 1948 from the Malay Peninsula and Java with successful results. Within a few months the overall parasitization reached 60 percent and this proved sufficient to check the outbreak.

Brontispa limbata Waterhouse (sometimes called B. gleadowi Weise)

Economic aspect

This species damages coconut palms in the same way as those already described. It also attacks Dictyosperma and Acanthophoenix.

Geographic distribution

Mauritius.

Biolog y

The egg stage lasts 6-8 days, the larval about 2 months, and the pupal stage 7 days. The complete cycle from egg to adult is 60-80 days.

Control

Spraying twice with phosdrin with an interval of a week between treatments is recommended. DDT at 0.1 percent ensures protection for 4-6 weeks, and dieldrin at 0.025 percent for 6-8 weeks. These two insecticides are more effective than chlordane or toxaphene (Orian, 1961).

Plesispa reichei Chapuis

Economic aspect

The larvae and adults of this insect are very harmful to young coconut palms, of which they gnaw the young unopened leaflest in a manner similar to Brantispa, so that when the leaves open much of the tissue dies and shrivels and the leaflest are torn by the wind. The larvae, like those of Brantispa and unlike those of Prantesibera, are not miners. The growth of infested young palms is delayed, and in prolonged outbreaks they may be killed. Less damage occurr in nursery beds under rubber or other shady

trees than under tall coconut palms. The insect breeds on Nypa and Roystonea also.

Geographic distribution

The Malay Peninsula, Indonesia.

Description | Biology

The developmental periods are: egg, 10 days; larva, 27-43 days; pupa, 6-11 days. The whole cycle from egg to adult has been recorded as 40-64 days (Corbett, 1923) and 6-7 weeks (Kalshoven, 1950-51). The female can live 6-8 months and lay 50-95 eggs (Kalshoven). The larva and pupa have a pair of calipper-like processes at the hind end, similar to those of Brouting. The male adult is 6.5 mm long, the female 7.5 mm; the head and thorax are brownish orange, the elytra black.

Control

If chemical control should prove necessary, dieldrin, ppr or chlordane would be appropriate, at the concentrations indicated for use against Brontispa. The eggs of P. reidni are parasitized by Obentyrin podoniae Gahan (Hym., Chalitidalea, Entyrtida) and Haeckellama brontispar Fetrière (Hym., Chalitidalea, Tirkymamatida).

CURCULIONIDAE

This vast family of insects, the weevils, is one of the most easily recognized, because of the characteristic rostrum or "snout" and the elbowed, clubbed antennae.

Rhynchophorus ferrugineus Olivier

Economic aspect

This weevil is one of the most harmful and notorious pests of the coconut palm. The damage that it does is due entirely to the larvae, which feed in the trunk and also in the heart of the crown of the tree. Not infrequently the larvae damage the growing point seriously and the tree dies in consequence. This pest is often associated with Oryteix, whose activities provide it with niches suitable for egg laying. In severely infested plantations, many palms lose their crowns completely, so that only the dead trunks remain. On the Malabar coast of India, 5 percent of the young palms up to 10 years old are killed annually. Other palms attacked by this weevil

ate: Areca, Arenga, Caryota, Coelococcus, Corypha, Elaeis, Livistona, Metroxylon, Nypa, Oncosperma and Phoenix.

Geographic distribution

India, Ceylon, Thailand, New Guinea, the Philippines.

Description | Biology

The eggs (Fig. 77) are laid in existing wounds in the crown or trunk of the palm, in holes made in the tissues with the rostrum. The duration of adult life is 3-4 months, in the course of which the female lays 200-500 eggs. The incubation period is 3 days. The larva (Figs. 78 and 82) lives entirely in the tissues of the tree, where it excavates galleries; the damage it does is therefore not apparent at the time and is not suspected until the consequent drooping of the leaves is recognized as a symptom. When fully grown, the larva is 65 mm long, and is stout, legless and yellowish, with the head and the first segment brown. The larval period varies in duration from 2 to 4 months. Pupation takes place under the bark, in a cylindrical cocoon (Fig. 83) coarsely constructed of tangled fibers and measuring up to 80 mm in length, and this stage lasts about 14 days. The adult (Fig. 79) remains in the cocoon to harden for 11-18 days after shedding the pupal skin. The whole cycle, from the egg to the emergence of the adult from the cocoon, occupies 3-6 months, varying with the amount of water contained in the tissues eaten. The adult is capable of flying strongly and is diurnal in its habits; its size varies greatly, from 25-50 mm. Its color is chestnut brown, with a few black spots on the thorax (Dammerman, 1929). The female is strongly attracted to fermenting sap oozing from wounds in the trunk or the leaf bases.

Control

Dieldrin at 0.1 percent should be applied to the damaged trunks or, in the case of new plantings, sprayed every three months on the crowns and bases of the trees (Vestal, 1956). This is advocated in Thailand where R. Jerngimus, often associated with R. Jenden, in the crowns of the plans is very difficult to combat when it gains entry into the bases of the trunks. As a prophylactic measure, the application of a mixture of 5 percent succordiorane and sand to the axils of the leaves is recommended. Another method consists in pouring diluted Metasystox into a hole about 5 cm deep drilled in the trunk just above the injury.

Since most of the damage done by *Rhynchophorus* is secondary to that of *Oryctes*, control of the latter is an important aspect of control of the former. This requires strict attention to the destruction of logs and stumps by burning and to preventing the accumulation of dead vegetable matter which would decompose and provide breeding places for Oryste. In Thailand, it is recommended that trees heavily infeated in the crown with Rhyndophorus should be felled, the top 50-75 cm of the trunk cut off and split to expose the insects, and this infested part covered with diesel oil and then burned (Vestal, 1956). It is also necessary to prohibit the cutting of footbolds in the trunks for climbing, the cutting of leaves for feeding to cattle, and any other injurious activities which provide sites for egg laying by the weevil (Child, 1964).

There is some parasitization of the larvae in Java by the large wasp Solia erratice Smith and in India by the calliphord fly Sarrophaga fusicuated Bottcher (Lepesme, 1947). A mite, Tetrapolphus thymchophari Ewing, of the family Pymotidus, has been recorded as a predator of Rymchopharu (Kurian, 1963). However, since these natural enemies do not play a significant part in the limitation of the weevil populations, it is necessary to rely on chemical treatment combined with efficient seates assistation.

Rhynchophorus schach Olivier

Economic aspect

This species is often regarded as a form of ferraginus but is here treated as a distinct species. The main behavioral difference between the two is that ferraginus commonly attacks the trunk of the palm, while schath most frequently damages the terminal bud. Otherwise they are very similar and there is an association with Oryster in both. Alternative host plants among the palms include Sahal, Lieuala and Verschaffeltia, nonindigenous species, all of which were killed by R. schach in the Botanic Gardens at Singapore.

Geographic distribution

Sumatra, the Malay Peninsula, Borneo, southern Thailand, New Guinea.

Description | Biology

The beetle (Fig. 80) is shining black, except for a median stripe on the thorax and the "club" of the antenna, which are orange-red; its length is 45 mm. The duration of the developmental cycle, from egg to adult, is about 4 months.

Control

The measures recommended in Thailand against R. ferrugineus are applied against R. sebaeb equally. In Sabah also, spraying with dieldrin is advocated,

in this case at 0.2 percent, and another treatment is injection into the galleries of paradichlorbenzene in ethyl acetate. These measures, however, are not sufficient in themselves; proper plantation management to ensure the maximum possible reduction of the breeding places of Orycles, which attract Rhundsborus, is a basic requirement.

Rhynchophorus papuanus Kirsch

This is another species (if indeed it is a distinct species) in the Southeast Asia-New Guinea complex. It occurs in eastern Indonesia and New Guinea (Kalshoven, 1950-51), and is particularly associated with the indigenous sago palm, Metroxylous sagus, but attacks occonut also. It is distinguishable from R. ferraginus and R. schach in that it is entirely black; Kalshoven regards it as merely a form of ferraginus;

Rhynchophorus palmarum Linnaeus

Economic aspect

In South America, this species has the same importance as the preceding species in Asia. It does the same type of damage, and is, in addition, the vector of a nematode of the genus Rhadinaphelenchus, which is responsible for red ring disease of coconut palms.

Geographic distribution

Tropical South America, Mexico and the West Indies. 18

Description | Biology

The eggs are laid in cracks and wounds in the bark, and the maximum number recorded for a single female in the course of its life is 718. The period of incubation is 3 days. The larva is fully grown in 52 days (Hagley, 1965b). According to Wilson (1962, 1963), larvae reared on coconut "cabbage" (the central leaf bud) are fully developed in 24-41 days, whereas in harder, woody tissues the corresponding period is 47-62 days. When the larva tunnels in the trunk it fills its galleries with a moist, sour-smelling mass of fragmented fibers, through which it moves with a gurgling noise (Wolcott, 1933). The cocoon is similar to that of the other species already described. The pupal period (including a prepupal stage in the cocoon)

¹³ A record of this species from Hawaii (Lenesme, 1947) is erroneous,

has been variously estimated as 11-55 and 27 days, and the whole developmental cycle from egg to emergence of the adult from the pupa as 79 days in the laboratory and 72 in the field (Hagley), and 78 in the field (Maharai, 1962). The adult (Fig. 81) does not commence to feed until 8 days after emergence from the pupa. It is attracted to sap oozing from wounds or burns on the palms. The local people eat the larvae; this applies also in Burma, the Malay Peninsula, the Philippines and New Guinea in respect of the congeneric species occurring there.

Control

Spraying with aldrin is reported to have proved effective in Guyana; dieldrin at 1 percent can be expected to be equally satisfactory. The problem of control of this species is less difficult because Orpicts is absent from the New World, but proper estate sanitation and avoidance of slashing the trunks and leaves are nevertheless important.

A tachinid parasite, Parabillana rhymchophorus Blanchard, has been discovered which attacks the pupae of R. palmarum in the palm Attala princept in Bolivia (Candia and Simmonds, F.J., 1965). The young larvae of the fly seek out the Rhymchophorus pupae in the trunk.

Rhynchophorus phoenicis Fabricius

This is the African representative of the genus. It tunnels in the trunks of coconut palms, stops growth and even kills them (Saraiva, 1939). It occurs in tropical Africa and probably in Iraq (Lepesme, 1947). The cycle from egg to adult covers about 60 days, of which 26 are spent as a larva.

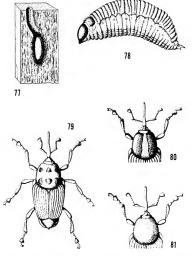
Rhabdoscelus obscurus Boisduval (formerly assigned to the genera Rhabdocnemis and Sphenophorus)

Economic aspect

This weevil is perhaps best known as a pest of sugarcane, but also does serious damage, of the same type as that done by Rhynchophorus, to coconut palms, and damages many other cultivated plants, including Areca, Metroxylon, banana and papaya.

Geographic distribution

Moluccas, New Guinea, the Pacific Islands (including Hawaii) and Queensland (Australia).



FIGURES 77-81, - Rhymchophorus species, Egg (77); larva (78); and adult beetle (79) of R. ferragineus Fabricius; heads and prothorax of R. schuch Olivier (80) and of R. palmarun Linnaeus (81).



FIGURES 82-83. - Rhymbophorus ferragineus Fabricius. Larvae and pupae of different developmental stage in a portion of damaged crown of coconut palm (82, above), and isolated cocoon (83, below).

(Photos C. Kurian)

Description | Biology

The adult weevil is reddish brown with the head darker and a vague longitudinal darker mark on the thorax and often on each elytron. It is about 15 mm long. The duration of the egg, larval and pupal stages are 6, 80 and 8-14 days, and the whole cycle occupies about 13 weeks. The adults are strongly attracted to the fermenting sap which exudes from wounds on the tree.

Control

In New Guinea, this pest is kept in check to a considerable extent by a dipterous parasite, Ceromasia sphenophori Villeneuve (Tuchinidae), which has been introduced into Hawaii and other countries for control of the weevil, primarily as a pest of sugarcane, with useful results. As in the case of the other weevils already mentioned, it is necessary, in order to reduce damage to coconut palms, to avoid wounding the trunks as far as possible.

Rhinostomus barbirostris Fabricius (formerly called Rhina barbirostris)

Economic aspect

The female lays its eggs in the trunk of healthy, mature palms but also attacks rotting palms. It is suspected of being a vector of the nematode Rhadinaphelmun, which is the causative agent of red ring disease, but its importance in this connection is small compared with that of Rhynchophorus palmarum. The foliage of trees infested by it turns yellow, their growth ceases and their trunks become liable to break in the wind (Wyniger, 1962). The weevil breeds also on Arreatirum, Attalea, Diphethemium, Elastis and Syaguu (Lepsene, 1947).

Geographic distribution

Mexico and South America, including Trinidad.

Description | Biology

The eggs (Figs. 85 and 86) are laid in holes 2 mm wide made in the trunk by the female with its rostrum. The resulting larvae (Fig. 87) tunnel horizontally in the trunk. They are whitish with a yellow head and spots on the body. Pupation takes place in a cocoon of tangled fibers. The adult is blackish brown and 30-04 mm long, excluding the rostrum, which is itself 10-12 mm long (Wolcott, 1933). The rostrum has a bushy growth of brown hair; hence the name of the species.

Control

Young larvae can be detected by the presence of their excrement on the trunk; their further penetration into it, and the entry of other larvae nearby, can be prevented by the application of tar (Wolcott, 1933). Treatment of the trees with a solution of dieldrin at 5 percent, or with pyrethrum and piperonylbuxoyl at a similar concentration, is also recommended.

Rhinostomus afzelii Fahraeus

This species (Fig. 84) occurs in tropical Africa and Zanzibar, where it does the same damage to coconut palms as *R. barbirostris*: in South America (Saraiva, 1939). It attacks *Elasis* and *Rappia* also.

Sparganobasis subcruciatus Marshall

Economic aspect

The larva tunnels in the trunk of the coconut palm, often at the point of junction of a leaf petiole. Infested trees are liable to be so severely weak-ened that they fall to the ground (Simmonds, H.W., 1938). Discoloration of the leaves and exudation of sap are characteristic symptoms.

Geographic distribution

New Guinea.

Description | Control

The adult is black and 18 mm long. It is readily attracted to damaged coconut trunks and trapping has therefore been recommended as a control measure, but unless this operation is carefully supervised and the trunks used to attract the weevils are inspected daily it can do more harm than good.

Diocalandra taitensis Guérin

Economic aspect

This weevil attacks all parts of the coconut palm (Lever, 1941b) but opinions as to its status as a pest vary greatly. Some entomologists assert that damage by it is purely secondary but others claim that it does serious primary damage to roots, leaves and fruit stalks, and that it is a cause of

premature shedding of nuts. Particularly severe injury has been reported from the Line Islands (Herms, 1926), Tahiti (Doane, 1909) and elsewhere in the Pacific (Zacher, 1913), where the larvax till the young nuts and feed in the leaf bases, tunneling in the latter from the base toward the leaflets. In Madagascar, the larvace burrow in the trunk at all heights (Frappa, 1947). *D. taitenii* is not known to attack palms other than the coconut.

Geographic distribution

New Guinea, the Solomons, Melanesia, Polynesia; the species was inavertently introduced into Hawaii in 1919 and Madagascar in 1943. It was one of the first coconut pests to be recorded in Tahiti in 1840.15

Description | Biology

The eggs are laid in crevices at the bases of the adventitious roots at the foot of the tunk, or in the inflortescences, or at the base of the peduncle or of the petiole. Incubation requires 4-8½ days. The larva penetrates into the tissues and causes gum to appear at the opening of the gallery. Larval development covers 8-10 weeks, and pupal 10-12 days, (Herms, 1926). No ecocon is formed. The whole cycle from egg to adult occupies 2½-3 months. The adult is usually black with dark reddish markings on the thorax and clytra. It is 6-7 mm long.

Control

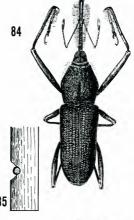
Chemical mesures are not usually employed but Wyniger (1962) recommends application of dieldrin at about 0.5 percent. The treatments generally adopted are painting with tar the wounds made by the insects and earthing up the bases of the trunks.

Diocalandra frumenti Fabricius

Economic aspect

D. frumenti does the same type of damage as D. laitensis, and there is similar diversity of opinion as to whether its attack is primary or secondary. It breeds on Area, Elaiti, Nypa and Borassus as well as coconut, and has been recorded as attacking sorghum.

¹⁴ Records of it from India, Ceylon, the Malay Peninsula and Indonesia are now known to refer to D. frumenti (also called D. stigmaticollis Gyllenhal)..



FIGURES 84-87. Rhimatomus species. Adult beetle of R. affeili Fähraeus (84); egg of R. barbirostris Fabricius laid in the coconut palm trunk (85, cross-section; 86 front view); and larva (87). According to P. Lepesme, 1947, in part redrawn.

(Courtesy P. Lechevalier, Paris, France)



86

Geographic distribution

Zanzibar, Seychelles, India, Ceylon, the Malay Peninsula, Thailand, Indonesia, New Guinea, Solomons, Guam, Carolines, Gilbert and Ellis Islands.

Description | Biology

The habits of *D. frumenti* are almost identical with those of *D. taitensis*. The adults are often found at the base of a nut which is about to fall, and also in the accumulation of fallen male flowers at the base of a leaf (Corbett, 1932).

This beetle is not so dark in color as D. taitensis and has, on each elytron, four spots, two blackish brown and two yellowish brown.

Control

Spraying with dieldrin at about 0.5 percent is recommended. The application of tar to the base of the trunk is also useful. Efforts should be made to prevent slashing of the trunks with knives and to ensure that the adventitious roots are covered by earthing up.

The following natural enemies have been reported:

The parasite Spathius apicalis Westwood (Hym., Braconidat) which attacks the larvae and destroys up to 40 percent of them; the predatory beetle Placisis javanus Erichson (Col., Histeridae), and the predatory fly Chrysophilus ferranjavanus Wiedemann (Dipt., Rhagionidae).

Homalinotus coriaceus Gyllenhal

The adults of this species, which are about 20 mm long, attack the spathe of the coconut palm and the larvae tunnel in the petiole and midrib of the leaf and even in the trunk (Bondar, 1940a). It is found in Brazil and Argentina.

Suitable control measures are spraying with BHC at 0.06 percent in water and, for the petioles, with a mixture of tar in petrol at 1 kg in 8 liters (Fonseca, 1962).

SCOLYTIDAE

Xyleborus perforans Wollaston

The presence of this pest (or other related species) in coconut plantations is indicated by numerous small circular holes, all of the same size, in the

trunks of the trees. These are the ends of galleries made by the adult beetles, which lay eggs in them. The larvae excavate further galleries at right angles, and they and the adults feed on "ambrosis" fungus which grows on the walls. The duration of the life cycle is 2-3 weeks. The exact relation between this small beetle (only 2 mm long) and the host plant is still inadequately understood. In the Seychelles and Jamaica it is recorded as being harmful only to trees which are already unhealthy, while in Fiji it is considered to be a primary pest (Paine, 1934). X. perforas is very widely distributed, almost throughout the tropics and subtropics. It has a wide range of host plants, including Eliati, Ladaina, sugarcane and rubber. For control, bor, dieldrin, endrin or thiodan, as a wettable powder or emulsifiable solution, are ercommended (Wnyinger, 1962).

DYNASTIDAE

Oryctes rhinoceros Linnaeus

Economic aspect

This is the notorious rhinoceros beetle, certainly one of the most serious pests of the cocount palm. The damage is done solely by the adults, which bite through the tightly packed unopened leaves in the central bud, with the result that when the leaves open they are seen to have great triangular gashes in them, as if the component leaflest had been cut with scissors. The midribs of the leaves are often severely injured, and the spathes and inflorescences less often so (Figs. 89 to 91). Much additional damage, and often the death of the trees, result from the attraction of the weevil Rhynchophorus to the wounds inflicted by Opytent. This insuces it is best known as a pest of mature palms, but palms of all ages are attacked and, in 1957, one third of the seedling palms in the Chagos Islands were destroyed by it (Orian, 1959). It attacks numerous other plants, mainly but not only palms: Area, Arnag, Borasun, Corpho, Elunis, Linitiona, Mirescylon, Nypa, Omesperma and Phomix, and also Pandaman, pincapple, Colonain, banan and suggracema.

Geographic distribution

O. rhineceres is found throughout Southeast Asia and also in the Philippines and southern China. It reached Mauritius in or shortly before 1962, but it does not yet occur in Africa. In the Pacific, it was discovered in

¹⁵ All reports of it from Africa have proved erroneous.

Samoa in 1909; in Niuataputapu (Keppel Island) in 1924 and eradicated there in 1931; in Wallis in 1931; in New Britain and the Palau Islands in 1942; in Tonga in 1951; in Fiji and New Guinea in 1953; and in the Tokehau Islands in 1963 (Fig. 88). The date of discovery is likely to be a year or two later than the date of arrival of the insect.

It seems to have arrived in Samoa with rubber seedlings from Ceylon before the institution of phytosanitary legislation. The larvae are known to be capable of surviving in floating logs transported by ocean currents. Wartime shipping aided its spread, and the great number of palms felled or broken in the course of military operations provided it with unprecedented opportunities for breeding.

Description | Biology

The eggs are laid in moist, decomposing vegetable matter. Cow dung is said to be a usual breeding medium in India and Cevlon. The egg is whitish brown, tough, and 3-4 mm long. The incubation period is 7-18 days, the mean being 12 days. The larva (Figs. 95 and 96) develops entirely in the rotting vegetation. It is 7.5 mm long on hatching and attains a length of 60-105 mm when fully grown. Its color is whitish, but the head is brown and the hind part of the abdomen is gravish blue owing to the color of the rectal contents showing vaguely through the outer tissues; the body is permanently curved. The larval period is extremely variable but the mean is 130 days in India (Nirula et al., 1955c) and in Ceylon (Goonewardene, 1958), and the range is 80 to more than 130 in the Palau Islands (Gressitt, 1953); the minimum duration is 72 days. There is a period of 6 days between cessation of feeding and pupation, and the pupal stage lasts 14-29 days, averaging 20 days. Pupation usually takes place in the soil but may occur in rotting vegetation or even in a rotting coconut trunk which is still standing (Gressitt, 1953). The pupa lies in a cocoon or cell composed of soil and agglomerated debris or, when inside the trunk, of fibers; it is 40-52 mm long, yellow-brown and, if male, with the "rhinoceros" horn already conspicuous on the head.

The adults (Figs. 22 to 94) do not leave the pupal cell until 11-20 days after emergence from the pupa. The total cycle, from egg to emergence from the cocoon, lasts from 4 to 9 months, the mean period being about 6 months; thus, at least two generations are possible per year. The beetles live mainly in the breedings sites, spending relatively short periods in the crowns of the palms, for feedings. The duration of adult life is at least 3 months. As the specific name indicates, they bear on the head a median horn curved backward; the horn is much longer in the male than in the female. The protonx has a large concave area situated antercodorsally. The beetle is black,

with a fine reddish pubescence on the ventral surface, and is 30-57 mm long. It is believed to fly only short distances; the longest recorded flight is about 700 meters, in Samoa (O'Connor, 1953). The maximum number of eggs recorded from a single captive female is 70 (in Java) and 140 (in southern India) (Kalshoven, 1950-51).

Control

As a fundamental prophylactic measure, all possible breeding places must be eliminated and every effort made to ensure that no new ones are created. Dead plant material which is rotting or which will rot if accumulated must be burned or buried, and fallen trunks must be split and burned.

Where plant material is used for making compost, chemical treatment of it is necessary to prevent the breeding of the beefle in it. Work in India in 1951 showed that application of 0.001 percent gamma-anc was effective, and subsequently this became normal practice there (Nirula et al., 1951b); recently 0.01 percent aldrin has been used successfully for the treatment of compost (Kurian and Pillai, 1964). In Fiji, good results have been obtained by applying gamma-suce at 0.075 percent or a mixture of 9 parts of damp sawdust and 1 part of gamma-suce at 6.5, 10 or 13 percent active ingredient (O'Connor, 1954b). Diazinon at 32 percent, though more costly, was later advocated because of its more rapid action and longer persistence (O'Connor, 1957b).

The first attempt to control the rhinoceros beetle by biological means was made in 1913 in Samoa by Friedrichs, who introduced a culture of the green muscardine fungus Metarrhizium anisopliae (Fig. 97) and spread cultures of it in compost heaps used at that time to serve as traps for Orycles. The attempt was not successful, and no greater success attended similar attempts made later in Cevlon, Mauritius and the Malay Peninsula. However, further attempts on the same lines but in the light of modern experience have been made recently and are still in progress; results are awaited with much interest. The effect of the fungus is to make the larva crawl out of the breeding medium, with the result that it dies and dries, and is mummified by the development within it of a white fungous mass which soon forms a crust outside the body and gives rise to green conidia from which the spores are distributed by the wind. The pupa and adult are also liable to be attacked by the fungus (Nirula et al., 1955a). Conditions favorable for the development of the fungus include fairly high relative humidity (over 70 percent), a temperature of about 27°C and overcast skies. These conditions occur in India during the season of the southwest monsoon, and it is hoped that it will prove possible during that season to infect Oracles with the fungus on a scale sufficient to result in an epidemic (Nirula et al., 1955c). Bacteria from the United States (Cumber, 1957) and other micro-organisms from Sumatra, Ceylon, Pemba and Africa (Surany, 1960) have been used to infect larvae and have sometimes caused a high degree of mortality. Intensive investigations on these lines are continuing in Western Samoa.

Certain insect parasites of Optitu larvae have been used in attempts at biological control. These are wasps of the family Soliidat indigenous to eastern Africa and Madagascar. These insects are strong fliers, and they feed on the nectar of numerous flowers, including those of Citrus, Urena, Cordia and Pointitia (Simonods, H.W., 1941). The female wasp butrows deep into rotting vegetation to find Orytiti larvae and to lay eggs on them. Among the scoliid wasps utilized in the control of Orytiti, the following are the most important:

Scolla orystophaga Coquillet, introduced from Madagascar to Mauritius in 1917, to Western Samoa in 1939, and to New Britain and Fiji in 1954, and from Mauritius to Indonesia in 1934 and 1936. The adult insect is reddish brown, has yellow antennae and measures up to 40 mm in length.

Scolia rufuconis Fabricius, introduced from Madagascar or Zanzibar to Mauritius and Western Samoa in 1945, to the Chagos Islands, New Britain and the Palaus in 1949, and to Fiji in 1958. This species is blue-black, and smaller than opytophaga.

Elis romandi Saussure, introduced from Madagascar to Western Samoa in 1939. This wasp is blackish brown.

In general, the results of these introductions of scoliid wasps have been disappointing, and in some cases the species have failed to become established. It is yet too early, however, to attempt a final assessment of them, since their increase is very slow and investigations of them are continuing as part of the much wider campaign against Orytes currently being conducted.

These wasps are very susceptible to most insecticides; consequently, chemical control must not be attempted in areas where they have been released and have not yet been evaluated.

Numerous species of predatory beetles have also been introduced into various countries in the hope that they will contribute to the control of Orycles. These include species of the genera Catascopus, Newbryopus and Mecopus (Carabidad), Psymborus, Landater and Alaus (Elateridad), and Holsieptis (Redunidad) has been imported from Zanzibar into India, Fiji and New Caledonia. This large bug, black with prominent orange-red parthes, inserts its proboscis between the head and the thorax of Orycter beetles and kills them; each individual Platymeris can destroy one beetle per day, and the duration of adult life is up to 4 months. Since the egg stage lasts about a month, this species can easily be transported by air (Vanderplank, 1958a).

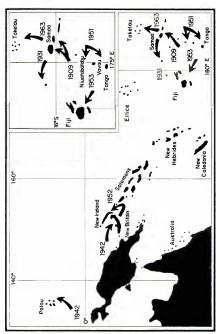


FIGURE 88. - Oryster rhinocerus Linnaeus. Spread in the Pacific area with years when first noted.



FIGURE 89. - Orystes rhinoseros Linnaeus. Typical damage eaused by adult beetles.

FIGURE 90. - Breeding site in a plantation of Western Samoa which has been almost completely destroyed by the pest.

(Photos V. L., Deluchi)





Figure 91. - Orystes rhinoceros Linnaeus. Adult penetrating into the crown center of a young eoconut palm. Western Samoa. (Photo K.J. Marishall)



FIGURES 92-94. - Orystes rhinoteros Linnaeus. Male (92, above); female (93, center); and mating (94, below).

FIGURES 95-97. - Orgetes rhimseeru: Linnaeus. Larvae of first, second and third (last) instar (95, above); adult larvae temoved from a rotted stump of esconst palm (96, center); and adult larvae infected by the fungus Metartheigum antispiam Metachnikoff (77, below).

(Figures 92-97 courtesy Institut national de la recherche agronomique, Station de recherches de lutte biologique et de biocanotique, La Minière, Versailles, France)



Orycles centaurus Sternberg

This large species, 50-60 mm long, inhabits New Guinea and the New Hebrides. It is more frequent in swampy areas where the sago palm (Metrosylon ramphius) grows than in coconut plantations. The eggs are laid in the inflorescences of Metrosylon after flowering is finished; since this palm dies and decays after flowering, it provides a large quantity of breeding material for Orystes at this stage (Hoyt, 1963b).

Oryctes gnu Mohner (also called O. trituberculatus Lansb.)

This is another close relative of Orgets thineerss. It is even larget, being 60 mm long, and has three small teeth at the posterior margin of the thorax, whereas O. rhineerss has only two. It occurs in Indonesia, the Malay Peninsula, Borneo and the Philippines. The life cycle and habits of the two species are very similar, and therefore the same control measures are required.

African species of Oryctes

There are several species of Oryster in tropical Africa. O. monectors Olivier (Fig. 98) is found in eastern Africa, Zanzibar, Madagascar and the Seychelles. Its eggs batch in 7-9 days, the larval stage lasts 100-200 days and the pupal stage 14-21 days; the complete cycle from egg to adult occupies about 6 months. It breeds in much the same situations as O. relineaters, and the same control measures are required, especially a high standard of estate santation and supervision (Dry, 1922). In addition to coconut, it attacks Hybbane and Borasin palms; in the Seychelles, coconut trunks previously damaged by Mellinuma provide breeding places for it, and palms may even be killed by it. In the small coralline Farquibar Islands, northeast of Madagascar, O. monoscerus has been exterminated (Piggott, 1961), as has O. relineaters in Niuataputapu in the Tonga group. It is worth noting that in Nigeria the deliberate destruction of oil palms by means of a herbicide, poured into holes in the trunks, unintentionally provided suitable breeding places for O. monoscerus (Shelldrick, 1963).

Orystes bear Fabricius (Figs. 99 and 100) of tropical Africa and Madagascar, and O. sjöstedii Kolbe and O. gigat Laporte de Castelneau of West Africa, are injurious to coconut and to oil palm, Raphia and Hyphanne. Investigations by Hoyt (195a) have shown that the practice of trapping by means of split logs, as in Samoa, is not satisfactory in Africa.

Strategus anachoreta Burmeister

S. quadrifoveatus Palisot de Beauvois

S. aloeus Linnaeus

These three species attack coconut (and other) palms in, respectively: Cubas and Trinidact Haiti, Puetro Rico and the Virgin Islands; and the Guianas, Venezuela and Brazil. They differ from the Old World species mentioned above in having no horn on the head. The adults (Figs. 101 and 102) burrow into the ground close to young palms and penetrate into the trunks below ground level, usually killing the palms, especially those about 2 years old. Good estate sanitation (burning accumulations of oconut husks, old palm trunks and other dead vegetable matter) is again essential as a control measure, since the larvae live in such situations. Development, from egg to adult, occupies 14 months in 5. quadriforestax (Plank, 1948).

Other dynastid beetles

There are several other genera of dynastid beetles which contain species that occasionally attack occonut palm without, in general, doing much damage. These genera are Sapanes and Trichogenphan (in the Solomons and New Guinea), Aljobrappei (from the Malay Peninsula, Cambodia, Laos and Viet-Nam, through Indonesia to the Solomons and New Hebrides), Papanem (from Burna to the Moluccas). The coconut palm cannot be regarded as the principal food plant of any of these species; when they do attack it, it is again the adults that do so, the larvae (so far as they are known) living in dead, decaying vegetation. The principle of efficient plantation sanitation is therefore important in the control of dynastid beetles in general, and also for the control of Rybushoptorus, since the damage done to various parts of the palms by the dynastids creates favorable breeding sites for the weevil.

LUCANIDAE

Several species of the genus Eurytrabelus are pests of coconut palms in New Guinea and the Solomons; the adults bore into the trunks near the leaf axils, and the larvae live in humus and rotting wood.

MELOLONTHIDAE

Leucopholis coneophora Burmeister

Economic aspect

This species was first recorded as a coconut pest in 1952 in southern India. The larvae damage the roots and reduce the yield of the palms, but severe damage is very localized and occurs principally on sandy soils and in association with areas in which various root crops are grown. Other plants attacked include cassava, sweet potact, yam and Cohansia.

Geographic distribution

Kerala in southern India.

Description | Biology

The eggs are laid, mainly in light soils, at a depth of 8-16 cm. The larvae come to the surface when the soil is moist and cool; heavy rain and a high water table are unfavorable to them. They attain a length of 60 mm. The beetles, which are chestunt brown and 30 mm long, appear suddenly in large numbers two weeks after the onset of the southwest monsoon (Nirula et al., 1952).

Control

Dusting the soil with about 66 kg of 10 percent nate or 33 kg of 5 percent chlordane per hectare and then plowing to a depth of 15 cm is recommended for destruction of the early stages (Menon and Pandelai, 1958). Aldrin, dieldrin, heptachlor and malathion are also effective and, for protecting the germinating nuts in unsery beds, covering the ground with a layer of sand is useful, as for avoidance of damage by termites.

Apogonia cribricollis Burmeister

This is mainly a pest of oil palms and cocoa, in the Malay Peninsula, but it attacks a very wide range of plants (Kalshoven, 1950-51), including the coconut palm occasionally (Lever, 1964a). The adults, which are dark brown and 8-10 mm long, are found on the leaves, and the larvae are in the soil, where they are general root feeders.

RUTELIDAE

Several species of this family, principally in the genus Adoretus, have occasionally been reported in Southeast Asia as being injurious to coconut



FIGURES 98-100. Orycles species.
Male (left) and female (right) of O.
monocero Olivier
(98, above); male
(99, center); and
female (100, below) of O. boas
Fabricius.

(Courtey Institut national de la recherche agronorvique, Station de recherches de lutte biologique et de biocamotique, La Minière, Versailles, France) palms, and to a wide range of other cultivated plants. It appears, however, that the damage done by them is almost always slight.

Hymenoptera

FORMICIDAE

The role of ants in coconut palms varies greatly. Some species are beneficial because they prey on harmful insects, while others are noxious either because they destroy useful insects (including ants) or because they foster injurious scale insects. In rare instances, ants damage the tissues of the palm directly. Many species of ants occur in the palms but the part played by them in the balance of the palm fauna is, in most cases, very inadequately known.

Oecophylla smaragdina Fabricius

(O. longinoda Latreille)

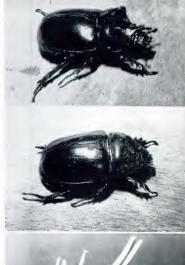
Economic aspect

These aggressive ants, which build their nests in trees, are well known over a large region of the tropics. They are greatly disliked, because of their fierce bite, by the laborers working on plantations, and they encourage scale insects and aphids on whose honeydew they feed. At the same time they are useful, since they prey on various other harmful insects and prevent or limit the breeding of them in the palms. They live in many different cultivated trees, including, in addition to palms, citrus, coffee, cocoa, clove, Finus and Casuarina, as well as numerous wild trees in the jungle. They are reputed not to atrack pests of the coconur palm in the Malay Peninsula (Corbett, 1932), whereas in the Solomon Islands and Zanzibar they are very important as predators of the coreid bugs Ambhyelia and Pruduhirenpina, respectively. Their precise conomic status in each locality must therefore be assessed independently of the palms, in relation to the fauna as a whole, including other ant species.

Geographic distribution

In association with coconut palms, O. Imanaglina occurs in Southeast Asia, New Guiuea, the Bismarck Archipelago, the Solomon Islands and Queensland. O. Imaginada is present in Africa, from the west through the Congo to eastern Africa and Zanzibar, and up to an altitude of 1,200 meters.

¹⁶ A record of India as the only Asian country in which this ant is found (Anon., 1962)



FIGURES 101-103.
Stratagus a lo eus
Linnaeus. M al e
(101, above); female (102, center); and larva infected
by the fungus (Cordyreps (103, below).
(Courtey Institut national de la recherche agronomique,
Station der echerches
de lutte biologique
et de biocamitique,
La Munitre, Versaillit, Frame)



Description | Biology

The nests are constructed in trees, of silk, in which living leaves are incorporated. Only the larvae can produce silk; ants of the worker caste hold larvae in their mandibles and use them as living silk-producing shuttles to spin together the edges of leaves held in place by other workers (Ridley, 1890). The leaves chosen have scale insects (e.g., Appliable deturturly or aphids (e.g., Atteppheryx nipar) on them, which are fostered in the nest by the ants in order to serve as sources of honeydew for food. Bees, termites and other ants are also devoured.

The rate of development is highly susceptible to temperature level. The incubation period is 12½ 6½ and 5½ adys, at 24, 28 and 30%C, respectively; the corresponding larval and pupal periods at the same temperatures are 25, 14 and 9 days, and 12, 5 and 3 days, and the total period from egg to adult is 49½, 25½ and 17½ days (Corbert 24, 1937). The workers live for 68-140 days, and a colony may exist as long as 5 years. The female lays up to 930 eggs in a day. The nests are usually built in exposed positions, where they get ample sunshine (Vanderplank, 1960a).

Control

Where control of Ocophylla is considered desirable, spraying dieldrin at 0.4 percent around the base of the trunks and on the leaves can be expected to destroy it and also to keep the trees free from other species of ants for eight weeks; this treatment was applied in Zanzibar, where other tests, with gamma-suce at 0.4 percent and 2.2.5 kg per hectare, gave similar results (Way, 1953a), DDT, dieldrin, malathion, diazinon and parathion are also recommended (Wwinger, 1963a).

Occopbylla is a relatively slow-moving ant and is prone to attack by several species of smaller, more active ants, among which Auspholely loagipts Jerdon is dominant in many of the countries concerned, including both Zanzibar in the west and the Solomon Islands in the east. The destruction of Occopbylla by these other ants permits the coreid bugs, Paunhalberaphus and Ambhypila, which are well controlled by Occopbylla in the absence of the others, to multiply to injurious proportions; the relations between these various ants have been studied by Way (1951, 1953a), Brown (1956, 1959) and Greenslade (1964).

Azteca cartifex Forel

This brownish black ant occurs in coconut palms in the Guianas and Trinidad, where it builds carton nests consisting of earth and wood fibers in the crowns of the trees. Its most injurious habit is that it fosters the scale insect Asphilotus destructor. Application of a 30-cm band of 15-20 percent dieldrin round the trunk about a meter above ground level, using about 0.3 liter per tree is said to prevent the ants from having access to the tree for one to two months.

Technomyrmex detorquens Walker (formerly often recorded as T. albipes Smith)

In the Seychelles, this ant fosters several scale insects, notably Pineaspii basei, on cocontup plams, specially in dry weather, and in so doing prevents the introduced coccincilid beetles of the genera Chilacorus and Escohomus from destroying them (Vessy-Fitzgerald, 1941a, 1953). Treatment with chlordane, aldrin or dieldrin can be expected to be effective as a control measure.

Dorylus orientalis Westwood

This is the only ant recorded as doing direct damage to coconut palms. It excavates galleries in the leaf stalks and nuts in the nurseries in India and Ceylon. In this case also, it is suggested that control could be achieved by the application of chlordane, aldrin or dieldrin.

Diptera

SCHOLASTIDAE

Species of Scholastes

Several species of flies of this genus have been recorded as damaging the meat of the ripe nuts, while it is still in situ (i.e., before it is converted into copra), in the Malay Peninsula, Melanesia and Polynesia including Hawaii. The flies, whose wings are heavily mottled with black, lay their eggs in the meat of the nuts, which the resulting maggots soon transform to an evilsmelling mass.

These flies are not pests unless the ripe nuts are left on the ground too long, before being collected for the extraction of the meat for the preparation of copra. Prompt and efficient gathering and splitting of the nuts is the remedy.

MITES

TENUIPALPIDAE (PHYTOPTIPALPIDAE)

Raoiella indica Hirst

Economic aspect

This small mite, which may occur in great numbers and sucks say from the leaves, has only recently been recognized as a coconut pest. It is harmful mainly to palms less than 5 years old, but older palms are susceptible if growing in soil that is ill-drained or lacking in nutrients. Dietystperma and Phoenix are also attacked.

Geographic distribution

India, United Arab Republic, Mauritius.

Description | Biology

The eggs are red and are laid on the lower surface of the leaflets. They hatch in 5 days. The subsequent developmental stages occupy 17 days, so that the cycle from egg to adult lasts about 22 days; development is, however, much slower in cool weather (Moutia, 1958).

Control

Destruction of about 80 percent of the mites (but not the eggs) has been achieved by sprays of Trithion, Rogor 40 or parathion, at 0.3 percent (Shashi Kanta et al., 1963).

A predacious mite, Typhlodromus candatus (Berlese) feeds on the eggs of Raoiella; the use of persistent chemicals which would kill this beneficial mite must therefore be avoided in procedures designed to control Raoiella.

PYEMOTIDAE

Pyemotes ventricosus Newport (recorded until recently as Pediculoides ventricosus)

This mite has a unique status in the fauna of the coconut palm, in that while it can destroy almost 100 percent of the larvae and pupes but not adults of certain pests in dry weather, in so doing it may cause extremely harmful one-stage outbreaks. Its role in the initiation of outbreaks of Promocother in Fiji has already been outlined, and this is the only continuous

investigation yet conducted of its effect as a component of the ecology of a pest over a period. It has also been recorded attacking Lennau in Fiji and Nephantis in India. It is nowadays almost cosmopolitan in its distribution; being commonly associated with insect pests of stored products, it is readily transported in ships. The male mite is 0.15 mm long, the female 0.22 mm, but when the female is gravid, part of its abdomen swells enormously and becomes a relatively huge sphere more than 500 times the volume of the whole body before fertilization. The female is viviparous and its progeny are adult at birth; about 90 offspring are produced by each female and almost all are themselves female. In view of this and the fact that the life cycle is very brief — only 6-8 days — the rate of multiplication when food is plentiful is phenomenal.

TETRANYCHIDAE

Tetranychus fijiensis Hirst17

In Fiji, this mite is found on both seedling and mature coconut palms (Simmonds, H.W., 1988). It lives in great numbers under a web on the underside of the leaflets and, as a result of its mass feeding, the leaves are discolored. The adult mite is red, like its allies which are commonly known as "red spiders." It is devoured in Fiji by a cocinellid beetle of the genus Symmus. For control purposes, straying with malathion, Rogor or parathion at about 0.4, 0.8, and 0.8 percent, respectively, is effective.

NEMATODES

TYLENCHIDAE

Rhadinaphelenchus cocophilus Cobb (formerly placed in the genus Aphelenchoides)

Economic aspect/Description

In tropical America, this minute celworm invades the woody tissues and causes the fatal red ring disease, especially in young palms. It is transmitted from one palm to another by the palm weevil, Rôpuchophorus palmanum, and is worst in neglected areas where there are many dead and dying trees in which the weevil is breeding. Extensive new planting after wholesale

¹⁷ A record of the occurrence of this mite in India (Anon., 1962) is erroneous.

felling of palms, practiced since 1925, has led to the increase of the nematode over a wide area in trees of the same age (Fenwick, 1959). Infected trees invariably die.

The symptoms of infection appear in the trunk in the form of a red band 2.5-4 cm wide and situated 2.5-5 cm from the outer surface, and starting about 1.2 meters above ground level. Red streaks appear in the leaf petioles also, and the roots become dry, flaky and discolored. There is always a yellowing and browning of the older leaves, starting at the tips, followed by shedding of leaves and nuts and secondary bacterial rot. Disintegration of the tissues is caused by a thermostable toxin associated with the nematodes but not directly caused by them (Goberdhan, 1963). In Trinidad, only 9.8 percent of the weevils are infected with living worms; the main sources of infection are fragments of infected tissue and, to a lesser extent, the excrement of Rbymkophorus in the larval galleries (Hagley, 1963, 1965a). It has been estimated that an infected trunk contains some 66,000 nematodes per meter of its length. The worm is only 1 mm long. Roystomea, Ellais and Phonis rea also statesche.

Geographic distribution

Honduras, Panama, Brazil, Venezuela, Colombia, Trinidad, Tobago, Grenada, Saint Vincent; and probably elsewhere in Central America.

Control

The best results have been obtained by the application of endrin at 19.5 percent, with coumarone; this reduced the incidence of infection by one third. Dieldrin and parathion were less effective. In Venezuela, po is applied to the soil and implements are treated to ensure disinfection (Median Duno, 1959). Strict estate hygiene, particularly splitting and burning infected trunks, is essential to check the breeding of the vector Rhymbaphorus. A promising method of control is the injection of Nemafos (Ex 18133) at 0.1 percent into the roots; this treatment, applied daily for six consecutive days, gives protection to the tissues for 1.5 meters from the point of injection (Goberdhan, 1964).

CRUSTACEANS

PAGURIDAE

Birgus latro Linnaeus

Economic aspect

This is the so-called coconut or robber crab. The notion that it climbs coconut palms to obtain the nuts is largely due to an account by Darwin

based on information received indirectly from the Cocos Islands in 1836. It seems, however, that no factual evidence exists of this behavior, and that the crab attacks only nuts that have fallen to the ground and are already cracked. In captivity, it does not open nuts and does not even appear to be particularly attracted to them for food. Its status as a pest is probably not high, therefore, and it is included here mainly because of its notoriety. Most of the damage attributed to it is caused, in reality, by rats. The published descriptions of its habits mostly attribute to it an ability to tear away the strong, fibrous husk of the nut in order to reach the meat via the germination "eve," but Revne (1939) and Gibson-Hill (1948), in Indonesia and Christmas Island (in the Indian Ocean), respectively, both concluded, after careful investigations, that the crab neither exposes the nuts in this way nor climbs tall trees to get them; nevertheless, both claims continue to be made, the latest being that of Eden (1963). It is to be hoped that the critical review of this problem by both Child (1964) and Tinker (1965) will dispel belief in this myth. The seeds of trees of the following genera are said to be eaten also: Barringtonia, Calophyllum, Pandanus and Terminalia.

Geographic distribution

The distribution of B. latro extends from the coast of eastern Africa to the Andamans and Nicobars, and also from Celebes, the Philippines and China (Taiway) eastward via the north coast of New Guinea to Melanesia and Polynesia as far as the Gambier Islands in the Tuamotu Archipelago; in the region between these two great zones, that is, in western Indonesia and the Malay Peninsula, the crab is absent (Reyne, 1939).

Description | Biology

The eggs and young larvae (which hatch into the zoca stage) are marine, the later stage terrestrial. The first nine months of the terrestrial period are spent in the shell of a univalve mollusk (Harms, 1932). The adults are nocturnal in habit; by day they rest under boulders or roots of trees, where they feed on nots, fruit and carrion, including corpses of their own species.

The color of the crab varies, not only according to age but also between individuals at the same stage of development. It is normally earthy yellow, with crimson and purple blotches. The adult weighs 2-2½ kg, and its length, including the pincers, may be as much as 45 cm. In inhabited islands it is scarce and seems to be decreasing in numbers, being coveted and captured for human consumption.

2. VERTEBRATE PESTS

BIRDS

Damage is caused in the Solomon Islands by a white cockatoo, Cacatua dwarptii. This bird rips out large strips of the husk of young nuts, which consequently cease growing and fall off prematurely. The percentage of nuts thus caused to fall in a four-month period varied, among trees studied, from 2 to 40, and the maximum number that fell from one tree was 14 (Brown, 1957). This latter figure is seldom reached, and the damage in general is not serious. In New Guinea, another, congeneric species, C. gadrita trion, causes the same damage (Simon Thomas, 1963).

In the Islands of \$30 Tomé and Principe, off West Africa, the control of rats by kites depends on the degree of scarcity of parrots, which are very aggressive. In \$30 Tomé, the parrot Pailtana riilhanus is rare and the kite Milwas migrans can hunt the rats freely and thus limits the damage done by them to coconuts; in Principe, on the other hand, the kite is mobbed by the parrot and driven off (Simmonds, F.I., 1960).

In Asia, crows pierce the immature nuts, especially in periods of drought, to drink the contained liquid. In Africa, weaver birds (of the family *Ploetidae*) tear the leaflets of the coconut palm to make their nests (Fig. 104).

In general, damage by birds is only sporadic and is probably outweighed by the benefit which results from their eating harmful insects (Child, 1964).

The Indian mynah, Aeridultures tristis, was introduced into Mauritus in 1763, primarily for the control of the locust Nomadaris uptumfasiata, which, with other insects, was eaten by it. It was introduced into Fiji, in the hope that it would attack Levuana iriduseus, but its effect was negligible and there are few people today, even on the plantations, who regard it as an asset.

In addition to the direct damage done in some countries by birds, there is also the indirect harm done by them in transporting minute insects in their plumage; it has recently been shown (Cocherau, 1965) that the very harmful scale insect Aspidions destructor is transported in this way in the New Hebrides.



FIGURE 104, - Damage caused by weaver birds (Ploceidae) which tear the leaflets of the coconut palm to make their nests.

MAMMALS

MURIDAE

Rattus rattus rattus

R. r. alexandrinus

R. r. frugivorus

Economic aspect

Rats can breed every two months and produce literes of 8 to 12. They therefore multiply rapidly and cause grave damage to the numerous crops, including coconuts, which they attack. They invade the crowns of coconut palms, including very tall, old trees, primarily to reach the maturing nuts, which they gnaw at the base in order to extract the liquid. The holes thus formed are rather large and usually ragged (Fig. 105) (Taylor, 1930; Paine, 1934; Laird, 1943). Rats also damage the spathes and inflorescences, and eat the copra before it has been stored. It has been noticed that damage by rats is less severe where there are streams, swamps or other sources of moisture (such as sweet potatoes or cassava) nearby. In the Cocos Islands, it is now recognized that damage to coconuts which was attributed in the past to the crab *Bitigue latine* was, in fact, done by rats.

The damage by rats is not easy to assess, and there is wide divergence in the available data. Two observers on the same island in Fiji but in different years (Taylor, 1930; Paine, 1934) estimated the loss of crop due to them as 6.8 percent and 28.7 percent, respectively. On the same island but much later, Simmonds, H.W. (1951) assessed the premature nutfall due to rats as 50 percent of the total premature nutfall. In the Philippines, the crop loss, calculated for a period of five years and for trees 45 years old, was 14 percent (Montenegro, 1962), while in French Polynesia it was 50 to 100 percent (Lassalle-Sere, 1955). The damage is greater in Polynesia and Micronesia than in Melanesia. In each archipelago, injury is more severe on the small flat coral islands than on the lofty volcanic ones.

Geographic distribution

This species is tropicopolitan, and occurs almost wherever the coconut palm grows.

Control

The various poisoned baits usually employed for the control of rodents in general cannot be used effectively in coconut plantations because of the arboreal habits of the rats. They can live almost entirely in the crowns of the trees high above the ground, leaping from one to another or making use of overlapping leaves, and seldom descending. However, in the lower islands of the Society group, a bait of 1 part of Warfarin (a blood anticoagulant) and 19 parts (by weight) of desiccated coconut, the latter lightly roasted to prevent mold formation, gave good results. The bait was placed in metal containers fixed to the trunks of the palms, at a spacing of only 3-5 baiting points per hectare and a rate of about 5 kg per hectare. The supply of bait in the containers should be maintained for a period of 10-14 days on each occasion (Dumbleton, 1955). Zinc phosphide can be used instead of Warfarin, but in this case unpoisoned bait should be used for several nights before the poison is incorporated in it. In the mountainous volcanic islands of the Society group, aluminum bands 40 cm wide, which last for 10-12 years, are fixed round the trunks about 4 meters above the ground to prevent rats from ascending. Zinc bands are less durable because of the corrosive effect of salt spray on the metal; they can, however, be treated with antirust paints. Plastic materials can be used but are more expensive. In Tahiti, the yield of copra has increased from 7-8 kg to 10-11 kg per tree as a result of banding (Lasalle-Sere, 1955). In Jamaica, banding has been practiced since 1884.

Special treatment is necessary to protect germinating nuts in nurseries. It is suggested in India that, since rats do not burrow into sand, a trench 15 cm deep should be dug round the beds and filled with sand (Naidu, 1960).



FIGURES 105-106. - Holes in young coconuts made by rats (105, above) and in older nuts by squirrels (106, below). Photos van Heum, in L.G.E. Kalshoven, 1950-51.

(Country N.V. Uligerei W. pan Hoer, The Hague)

Rattus exulans exulans

R. e. micronesiensis

These two subspecies of Ratina exchana also damage the nuts in the crown of the tree, in at least some of the islands where they occur, but elsewhere they are said to be less harmful to coconuts than Ratina reatus because they spend more time on the ground. A full account of their biology has been given by Strecker et al. (1962). In the Tokeku Islands, according to Laird (1963), R. exchans is active in the crowns of the trees and makes characteristic holes 2-3 cm in diameter in the nuts. Besides causing loss of copra, the rat is also responsible, though indirectly, for the spread of filariasis, since the mosquito vector (Acdet [Stegonyie] palymeitenis Marks) breeds in the liquid contained in the holed nuts. In those islands of the Tokelaus where the rat is absent, breeding of the mosquitoes is limited to holes and niches in trees with, in consequence, a lower incidence of filariasis.

Geographic distribution

R. exulans occurs in Southeast Asia and also throughout the Pacific from the Marshall Islands to Easter Island. The subspecies micronetiensis is in the Caroline Islands.

Control

As for Rattus rattus, bands of aluminum are fixed to the trunks of the trees; it is necessary to ensure that the branches of other trees do not come into contact with the trunks above the bands and so provide bridges for the rats,

The lizard, Varanus indicus, has been introduced into various Micronesian islands, and has checked the rats to some extent. Other vertebrate enemies of rats include snakes, birds (owls, kites) and mammals (civets) (Dammerman, 1929).

SCIURIDAE

Squirrels cause similar damage to rats (Fig. 106).

PTEROPIDAE

Pteropus species

These are the fruit bats or flying foxes. Several species of fruit bats occur in the coconut-growing lands of the tropics and some have been reported to cause damage to the palms. Well-known species include Puropur chawardii in south India, P. rompprus (not the vampire) and P. roluir throughout Malaysia, P. tepulatur in Melanesia and P. tongomo in Polynesia. These bats are about 30 cm long (i.e., head and body) and have a wing span of 1/2 meters. They roost by day, hanging from the trees in great numbers together, and fly in the evening in search of food, often traveling considerable distances and in large numbers. When they feed in coconut palms, they eat the male flowers and gnaw holes in the immature nuts to extract the liquid. All the species mentioned cause the same type of damage, and they attack the fruits of a great variety of trees, including, besides eccount, kapok, papaya, mango, jackfruit, Lanium and Spondias (Dammerman, 1929). Attacked coconuts have either a ragged hole 5 cm in diameter bitten out or an aperture extending some 8-10 cm round the shell, partly concealed by fraved ends of the chewel fibers of the husk (Brown, 1957).

The damage done is, in general, not great, but it may be severe locally; at one plantation in the Solomons the percentage of nuts caused to fail to develop varied from 16 to 46 among the trees studied, but this amount or damage is exceptionally high. It is interesting to note that 50 years earliey no damage due to bats was seen in native occount groves, and it was at that time assumed that they would not invade the estates that were then being planted (Proggatt, 1911). In Fiji, independent observers have reported that the damage done to female flowers and nuts there is almost negligible.

In the New Hebrides, it has been shown that bats are responsible for spreading the scale insect Aspidiotus destructor, which is transported in their fur as it is in the feathers of birds (Cochereau, 1965).

OTHER MAMMALS

Many different kinds of mammals, in addition to rats and bats, are injurious intermittently and locally to coconut palms. Seedling palms are particularly susceptible, the animals being attracted by the considerable amount of food available in a small area. Wild pigs are the most troublesome; they are killed by batis of cassava poisoned with zinc phosphide. Other harmful mammals are monkeys, elephant, deer, bears, squirrels, porcupines and bandicoots. In Principe Island, a monkey, Creopitheam mone, picks the young nuts and is very difficult to control. Most of the larger mammals can be excluded from the plantations by fences of wire or wood or by stone walls.

Domestic animals also damage coconut palms occasionally, particularly by pulling down and breaking the leaves from the smaller trees (Child, 1964), and man himself, with his bad habits of cutting steps in the trunks to facilitate climbing and indiscriminately cutting the leaves and slashing the trunks, is also sometimes in the category of an animal injurious to the coconut palm.

3. INSECT PESTS OF COPRA

Lepidoptera

PHYCITIDAE

Cadra cautella Walker (widely known as Ephestia cautella)

Economic aspect

The caterpillar of this moth is very well known as a pest of oily vegetable products, such as cocoae beans, cottonseed, nuts and copra, and also of dried fruits, especially figs. As a pest of copra, it is injurious mainly when this product has not been properly dried and suitably stored; well-prepared copra is too hard for the caterpillars to attack (Corbett et al., 1937). The insect is said to be strongly attracted to the molds which thrive on poorquality copra, and crossinfestation occurs in the holds of ships between such consignments and cocoa. According to Bondar (1940b), in Brazil the caterpillars are commonly found in the crowns of the trees, feeding on both male and female flowers and on the ripe fruit; this seems to be the only record of this notroious pest of stored products feeding on growing palms.

It should be noted that there has been some confusion in the past between this and certain other similar and related species, through misidentification; thus a statement from the Malay Peninsula to the effect that C. cautella prefer rice debris to copra probably refers to Careyra ephalantica Stainton (Galleridae), while C. cautella was probably responsible for damage to bagged copra in New Guinea, attributed at the time to Ephastia elutella Hubner (Phytildae).

Geographic distribution (cosmopolitan)

C. cautella survives shipment from the tropics to much colder regions in Europe and North America, but since there is no diapause in its development it does not thrive at lower temperatures.

Description | Biology

The egg is pearly white, with a minute protuberance at one end. It hatches in 3-4 days. The caterpillar is pinkish, and is 12 mm long when fully grown. The chrysalis is enclosed in a whitish silken cocoon. The

duration of the caterpillar stage varies greatly from 22 to 49 days, according to the nature and condition of the food as well as to the temperature and humidity, and the chrysalis stage lasts 6-7 days; the cycle from egg to emergence of adult averages 45 days. All these figures refer to the rate of development in tropical conditions. The adult is pale grayish with a vague dark crossband on the forewing; the wing span is 16 mm.

Control

The prevention of serious infestation by C. cauthla requires application of the measures outlined at the end of this chapter for the avoidance of damage by copra pests in general. However, the following recommendations made by Corbett et al. in 1937 with special reference to this pest are still applicable and worth stressing: the copra should not be bagged on the same day as it is removed from the direr; it should not be stored in bags that have been used for rice; copor athat has been wetted by sea spray or rain should be well dried; and storage sheds should he disinfested by cleaning and fumigation.

The caterpillars of C. cantella are parasitized by Bracon hobetor Say (Hym., Bracondae). The degree of parasitization is low, however, and the parasite does not, in any case, become active until the population of C. cantella is already large and dense.

Colcoptera

CLERIDAE

Necrobia rufipes de Geer

Economic aspect

This beelle is common, especially in the wet senson, in copra stores (but not in direis); in many contrities of Southeast Asia and the Pacific, it is popularly known as "the copra bug." In addition to copra, it feeds and breeds on both animal and vegetable commodities, including salded meats, dried fish, prawns, bones, skins, cheese, cocoa beans and nutmegs; it also devours larvae of the fit; Phiphila cani Linnaeus (Phiphilaidai) in cheese, and of the beetle Carpaphili dimitatur Fabricius (Vilhidididai) in copra, as well as molds. It is a serious pest of badly prepared, moldy copra but not of high-quality, properly prepared copra, on which its laval mortality is high. This insect can serve as an indicator of the degree of inefficiency and uncleanliness of copra stores; since the bettle fib by day in the sheds they are easily observed, and their presence in large numbers indicates inferior management in both the preparation of the copra and the hyglene of the stores.

Geographic distribution

N. rufipes occurs throughout the tropics, and is virtually cosmopolitan.1

Description | Biology

The beetle is iridescent blue-green, with reddish brown legs and antennac. The life cycle occupies about 66 days in tropical temperatures when the insects are fed solely on copra, but the addition of fish to their diet reduced the duration of the cycle to about 43 days and when larvae of Carpophilas were included, it was only 38 days (Ashman, 1963); these supplemented diets increased not only the rate of development but also the length of life and fecundity of the adults.

CUCUJIDAE

Oryzaephilus mercator Fauvel

This insect is primarily a pest of oilseeds, spices and dried fruit. There has been confusion in the past between this species and the very closely related O. narinamenti Linnaeus, but it is now generally agreed that nearly all the past records of the latter on copra actually refer to merculor. The life cycle varies very greatly in duration in tropical conditions, from 17 to 97 days, according to the nature of the food. This is another species which does not seriously infest well-dried copra.

Abasverus advena Waltl.

Like Nerobia rufsper, this small beetle frequents the copra stores but not the driers. It does not feed on high-quality copra but only on copra on which the mold Penicillium is growing; it also consumes coconut meat (i.e., the product before it is converted into copra) on which the mold Aspergillus is present (Corbett et al., 1937). On good-quality copra the beetles, after eating what little mold there is, leave it for inferior samples. A. advena occurs in all tropical regions and attacks cocoa beans, ginger, nutmegs, bulbs and yams, as well as copra (Richards and Herford, 1930). In the Malay Peninsula the life cycle can be completed in less than 3 weeks but may require as long as 55 days.

¹ A statement indicating that in Asia and the Pacific it is confined to the Philippines, the Malay Peninsula and the British Solomons (Anon., 1962) is erroneous.

NITIDULIDAE

Carpophilus dim diatus Fabricius

This is a rather flattened beetle with short wing cases (elytra) which do not cover the last two or three abdominal segments. It frequents the copra driers rather than the storage sheds. In the tropics, it attacks, besides copra, cocoa beans, ginger, nutmegs, groundnuts and palm kernels. It is seldom found on copra of good quality. It develops through all its early stages in about 5 weeks, with a range of 26-44 days. This is another cosmopolitan species; in the cooler countries it is established in flour mills, particularly in rice mills in North America.²

TENERRIONIDAE

Tribolium castaneum Herbst

This is a very well-known pest of stored products. Though regarded primarily as a pest of cereal products, including flour and bran, it is also destructive, in circumstances favorable to it, to oilseeds and copra. Like all the other insects that damage copra commonly, it shows a marked preference for poor-quality material with an abundance of the mold Penidlium (Corbert α el., 1937). Its distribution is tropicopolitan. When it is provided with suitable food, its life cycle at 30°C is 26 days on average.

DERMESTIDAE

Dermestes ater de Geer

This is another widespread beetle that is liable to infest copra, though it is probably best known in association with animal products, such as hides, horns, hooves and bones. The length of the adult is 8 mm; it is much larger than any of the other beetles mentioned here as pests of copra. The life evele covers about 6 weeks at 30°C.

Avoidance of insect infestation of copra

SUITABLE PREPARATION OF COPRA

The insect pests of copra are attracted by the molds which grow on poorquality copra rather than by the copra itself. Consequently, severe infesta-

² A statement (Anon., 1962) indicating that in Asia and the Pacific it is confined to the Malay Peninsula and the British Solomon Islands is erroneous.

tion can best be avoided by producing copra of high quality. This is mainly a question of uniform drying to reduce the water content to or below a specified level which, following much experience and experiment, has been stated to be 6 percent. Efficient drying to this specification is most easily achieved by utilizing kilns constructed for this purpose; a good deal of copra, however, is still prepared by drying on racks in the sun, and in this case much effort and supervision are necessary, particularly in unfavorable weather, if the required standard is to be attained. A water content of not less than 7 percent is conductive to the growth of molds and results in the creation of a medium ideal for the feeding and breeding of the whole gamut of insect species. The partial decomposition and increased acidity of moldy copra are in themselves highly undesirable; the insect infestations which accompany them further depreciate the product by contaminating it with silk, excrement and larval skins and reducing much of it to powder (Child, 1964).

PROPER STORAGE HYGIENE

To ensure that serious infestation does not develop in copra during storage, the following recommendations should always be put into practice:

- Clean, noninfested sacks should always be employed; if the sacks have been used they should be treated with steam to ensure disinfestation.
- The floors, corners, ledges and as much of the walls as can be reached should be brushed regularly and thoroughly, and all the dust and debris thus displaced should be collected and burned.
- The bags should be piled neatly and systematically, and a space left between the piles and the walls to increase ventilation and facilitate inspection and cleaning.
- The copra must be removed from the stores for shipment in the order in which it was put in, the oldest batch always being removed first.
- The store must be maintained in a sound structural condition, so that the floor is reasonably free from rising ground moisture and the roof, walls and eaves are waterproof.

CHEMICAL TREATMENT

For the disinfestation of storage sheds and to ensure that the insect populations in them are maintained at a very low and virtually harmless level, regular chemical treatment of the floor and walls is recommended. To every 100 m³, 5 liters of a mixture of 170 grams of 25 percent lindane (suc) dispersible powder and 225 grams of 50 percent DDT wettable powder in water should be applied by spraying. If Cadra caustila is absent, the lindane can be omitted. A malathion spray can be used as an alternative, at 450 grams of 25 percent malathion dispersible powder in 5 liters of water per 100 m² of floor and walls, but this must be applied monthly. In severe cases, additional treatment with lindane smoke as a fumigant or pyrethrum and piperonyl butoxide as a fog may be necessary (Lloyd and Hewitt, 1958).

In India, good results have been obtained in the disinfestation of stored grain by the utilization of calcium phosphate at 2 or 3 percent (Majumder and Bano, 1964). This material is harmless to man but toxic to numerous stored product insects, notably Tribolium castanum and Cadra cantella; the success obtained with it against pests of grain suggests that it should be tested in copra stores also

The degree of severity of the problem of the control of copra pests in driers, small stores and large warehouses is, however, largely determined by those responsible for preparing and storing the commodity. Good-quality copra, stored in sheds that are well managed and maintained, will remain virtually free from pests for an indefinite period, certainly long enough to ensure that it is still in good condition when removed from storage for shipment.

4. PRACTICAL ASPECTS OF PESTICIDAL APPLICATION

To obtain efficient pest control by chemical treatments it is necessary not only to apply a well-selected pesticide at the most convenient doses, but also to determine the correct formulation, the time of application, and the type of spraying or dusting machine.

Formulation

An insecticide is normally manufactured in several different formulations, i.e., dusts, wettable powders, emulsifiable concentrates and aerosols. Care is required in selecting the suitable formulation type for the pest being treated. In the case of Oryster rhimeters, for example, O'Connor (1957) observed that a miscible concentrate of 50 percent вис had a better repellent action against the larvae than the wettable powder of the same concentration.

An insecticide treatment requires one or several applications, the timing of which is very important.

Applications

It is important to follow the manufacturer's stated concentration and doses. The increase of the concentration does not always give expected better results. Pillai and Kurian (1960) indicate, for example, that spraying with dieldrin at 0.05 percent against Nephantis trimopa in India gave a 77 percent mortality of caterpillars, whereas the same insecticide at four times higher concentration caused only a 5 percent increase in mortality (82 percent).

The commercial products contain rather high percentages of active ingredients. It is therefore necessary to dilute them to obtain solutions ready for application. The quantity of commercial product to be diluted is calculated by the "rule of three." If a 0.05 percent malathion water solution has to be prepared by using a commercial product with 45 percent active ingredient, the quantity of commercial product needed for 100 liters of water will be:

$$\frac{50 \times 100}{45} = 110 \text{ cm}^3$$

50 representing the quantity of malathion, in cubic centimeters, that should be found in 100 liters of the prepared solution.

As wettable powders contain a filler and/or a sticker, it is best to make up the amount of insecticide in a passe and then add the balance of the water, in order to avoid lumping. Spray nozzles should always be checked to ensure that their aperture is not too small for the powder in suspension. The container should be shaken, or its contents often agitated, to prevent the mixture from settling in the tank.

Owing to the necessity of getting an adequate cover of insecticide in the crown of tall palms, it is usually necessary to employ power sparyers or dusters. Airplanes have been successfully used to deal both with the Phasmid Graeffea creami in Fiji and with Locusta migratoria manilensis in the Philippines. The more general use of aluminum now permits the choice of long telescopic lances and light extension ladders. Furthermore, in view of the reasonable price and great convenience of portable fogging machines, their use seems worthy of trial in nurseries where a fine, persistent deposit on the leaves is easy to obtain.

The choice of the best sprayer or duster is important. In Zanzibar, three types of sprayer were used in control operations against *Pseudotheraptus*: a micron atomizer, a pressure sprayer and a swing fog machine. The results obtained were as follows:

Type of sprayer	Average number of insects (10 palms)	Average number of insects killed per liter of pyrethrum emulsion applied
Micron atomizer	628	200
Pressure sprayer	343	4.5
Swing fog machine	57	5.3

The higher mortality caused by the micron atomizer was due to a large number of droplets of uniform size (50 to 80 microns) carried on a strong blast of air into the crown of the 10-meter-tall palms. The atomizer was further equipped with a multipurpose nozzle enabling emulsions, suspensions and wettable powders to be used without any changeover (Vanderplank, 1954).

Similar care in the selection of spray machinery was used in the control of Breatispa longistima in the Solomon Islands. By choosing small hand sprayers with an efficient trigger grip, it was found possible to apply only 6 cm³ of insecticide per young palm and to obtain excellent results in the

control of larvae and adults, the center of the unfolded leaves having been conveniently wetted (Brown and Green, 1958).

Effects of insecticides on parasites

With the exception of the scollids parasitic upon dynastid beetles, most entomophagous insects are of small size. They belong mainly to the orders Hyssensphera, Diptera and Coloptera and are very susceptible to pesticides. The residual effect of 0.2 percent DDT applications against Niphantis serimopa, for instance, lasts more that two months, even with 450 mm rain, and the caterpillar parasite Trielospilar is also killed (Nirolla, 1958; Pilait and Kurian, 1960). In Malaysia, recommendation is made to avoid restrement against Brachartona catavantha where parasitism by the tachinid Bissa remota exceeds 70 percent, unless pesticides can be applied against the very early stages of development of the pest (Anon., 1951). In one instance where part of the infested area was sprayed with hist, the rate of parasitism by the fly was reduced from 80 to about 45 percent, thus enabling the pest to get a good hold.

These examples show that while modern pesticides, by virtue of their high toxicity and long persistence, are a considerable improvement on the prewar ones, they can be a two-edged weapon in their effect on the beneficial insects which, by themselves, often give a greater reduction of the pest.

Safety precautions in connection with pesticide application

Concentrated pesticides are toxic to man; therefore, certain precautions have to be adopted in handling them. The most elementary one concerns the keeping of pesticides in a room to which only one authorized person has access. Accidents are more likely in opening tins of the "press-down" type of lids than in field spraying with diluted chemicals. In case of contact with the pesticide by accident, the part of the body involved has to be immediately washed with water and soap, and the clothing removed and also cleaned. The empty tins or packages have to be destroyed or buried. Spraying or dusting must be carried out across-wind or downwind, so that the pesticide doesn't blow back on the laborer. The lightness of the sprayers' containers has to be checked. Protective clothing, gumboots, gauntlets, goggles, etc., have to be worn any time it is recommended by the pesticide manufacturer. Smoking and eating must be strictly forbidden during spray operations; laborers must wash their hands with soap and remove their clothing as soon as the treatment is completed. Care is also needed to avoid spraying ponds or streams which may contaminate drinking water for men and stock, besides killing fish which are very susceptible to chlorinated hydrocarbons such as DDT, BHC or dieldrin.

First-aid measures

A full account of this subject, with medical treatment, is included in Wyniger (1962). When poisoning is provoked by the ingestion of chlorinated hydrocarbons (norr, nict, dieldrin, aldrin, etc.), the stomach must be emptied and a saline purgative should be given; oily purgatives and milk must be avoided. In the case of poisoning with arsenicals and fluorides, the patient must have his clothing removed, lie on his side and be kept warm, quiet, and receive mentics. If intoxication is due to nicotine, the same care has to be given to the patient, except that he should receive coffee or alcohol instead of emetics, and fomentations on the abdomec.

REFERENCES

- ALDABA, V.C. Effect of leaf-miner attack and of cutting off the leaves. Philipp. J. Agric., 1931 2(1): 51-65.
- Anonymous. The coconut leaf moth (Artona catoxantha Hampson). Department of Agri-1951 culture, Malaya. Leaflet No. 29.
- Anonymous. Plant quarantine announcement: United States Trust Territory of the Pa-1959 cific Islands. FAO Plant Prot. Bull., 7(11): 149.
- Anonymous. Host list of fungi and insects on coconut in South East Asia and the 1962 Pacific. Indian Coconut J., 15(3 and 4): 164-178.
- Anonymous. New crop pest seares in the South Seas. Pacific Islands Monthly, 13(6): 13. 1964
- ASHMAN, F. Factors affecting the abundance of the copra beetle (Necrobia rufipes). Bull. 1963 ent. Res., 58(4): 67-180.
- Амивоwo, R. Der Klapperbladkever en zijn biologische bestrijding op Celebes [The 1935 ecoconut leaf beetle Brontipa frogatti var. telebentii and its biological control in Celebes.] Landboar, 10: 76-92.
- BARRETT, O.W. The tropical crops. New York, Macmillan.
- BHENCHITRA, P. Thailand's first locust outbreak. Span, 7(2): 97-98.
 - BODKIN, G.E. A new pest of coconut palms in British Guiana. J. Board Agric. Brit. Guiana, 1913 7(2): 87-90.
 - BONDAR, G. Notas entomologicas da Bahia. V. Rev. Ent. Rio do Janeiro, 11(1-2): 199-1940a 214.
 - BONDAR, G. Notas entomologicas da Bahia. VI. Rev. Ent. Rio de Janeiro, 11(3): 842-1940b 861.
- BRIONES, M.L. & SILL, W.H. Habitat, gross morphology and geographical distribution of four new species of Eriophyid mites from econouts in the Philippines. FAO Plant Prost. Bull., 11(2): 25-30.
- BRITISH SOLOMON ISLANDS. Annual report, Joint Coconut Research Scheme, Lever's Pacific 1962 Plantations Pty. Ltd. Yandina.
- BROWN, E.S. Biology of the coconut pest (Melittomma insulare) and its control in the 1954 Scychelles. Bull. ent. Res., 45: 1-66.
- Brown, E.S. Insecticide experiments. British Solomon Islands. Entomological Progress 1956 Report No. 3.
- Brown, E.S. Survey of distribution and status of premature nutfall in the British Solomon 1957 Islands. British Solomon Islands. Entomological Progress Report No. 4.

- Brown, E.S. Injury to cacao caused by Amblypelta with a summary of food-plants of species of this genus. Bull. ent. Res., 49: 543-544.
- Brown, E.S. Immature nutfall of coconuts in the Solomon Islands. Bull. ent. Res., 50: 1959 97-133, 523-566.
- Brown, E.S. & Green, A.H. Control by insecticides of *Brontispa longissima* in the Brit-1958 ish Solomon Islands. *Bull. ent. Ret.*, 49: 239-272.
- BULLEN, F.I. Unpublished record, Anti-Locust Research Centre, London.
- BURKHILL, I.H. Promecotheca cumingi Baly. Gardens' Bull., Singapore, 2(1): 3-5.
- CACHAN, P. Etude épidémiologique de la zygène en Côte-d'Ivoire. J. Agrit. trop. Bot. 1959 appl., 6(12): 653-674.
- CANDIA, J.D. & SIMMONDS, F.J. A Tachinid parasite of the palm weevil Rhymchophorus 1965 palmarum L. in Bolivia. Commonwealth Institute of Biological Control. Technical Bulletin No. 5, p. 127-128.
- CHILD, R. Coconuts. London, Longmans.

1964

- CLAUSEN, C.P. The time factor in biological control. J. econ. Ent., 44(1): 1-9.
- COCHEREAU, P. Contrôle biologique d'Aspidiotus destructor Sign. dans l'île Vaté, Nou-1965 velles-Hebrides, au moyen de Lindorus lopbantbas Blaisd. C.R. Atad. Agric. France, 51(5): 318-321.
- Соніс, F. Aleurode parasite du cocotier en Nouvelle-Calédonie. Agron. trop., 14: 232-1959 238.
- COHIC, F. Spread of two coconut pests: French Polynesia. FAO Plant Prot. Bull., 1961 9(6): 110-111.
- CORBETT, G.H. The two-coloured cocount leaf-heetle. Department of Agriculture, Straits 1923 Settlements and Federated Malay States. Bulletin No. 34.
- CORBETT, G.H. Insects of toconut in Malaya. Department of Agriculture, Straits Settlements 1932 and Federated Malay States. Bulletin No. 10 (General Series).
- CORBETT, G.H. & GATER, B.A.R. Batrachedra in relation to the nutfall of coconuts. Ma-1924 layan agric. J., 12(5): 115-122.
- CORBETT, G.H. & PONNIAH, D. The red-stripe weevil of coconuts. Department of Agri-1924 culture, Straits Settlements and Federated Malay States. Bulletin No. 26.
- CORBETT, G.H. et al. Fingi and batteria associated with copra in Malaya. Department of Agriculture, Straits Settlements and Federated Malay States. Bulletin No. 28 (Science Series).
- COTTERELL, G.S. The hispid leaf-miner (Coelaenomenodera elacidis Mlk.) of oil palms (Elaeis 1925 guimensis) on the Gold Coast. Bull. ent. Res., 16: 77-83.
- CUMBER, R.A. Ecological studies of rhinoceros beetle. Noumea, South Pacific Commission. 1957 Technical Report No. 107.
- DAMMERMAN, K.W. Agricultural zoology of the Malay Archipelago. Amsterdam, de Bussy. 1929
- DARWIN, C. Journal of researches into the natural history and geology of the countries visited during 1845 the voyage of H.M.S. Beagle round the world. London, Murray.

1947 302.

Dennis, M. Sevin insecticide provides outstanding insect control in Africa. Agrit. 1961 Chem. Digest, 3(3): 4-5.

DINTHER, J.B.M. De cocos bladrups [The coconut canker worm.] Surinam Landbouw, 1955 3(3): 209-216.

DOANE, R.W. Notes on insects affecting trees in the Society Islands. J. econ. Ent., 2(3): 1909 220.

DRY, F.W. Notes on the coconut beetle (Orycles monoceros) in Kenya. Bull. ent. Res., 1922 13: 103-107.

DUMBLETON, L.J. A list of insect pests recorded in South Pacific territories. Noumea, South 1954 Pacific Commission. Technical Paper No. 79.

DUMBLETON, L.J. Rat poisoning in French Oceania. Q. Bull. South Pacific Comm., 5(2): 1955 15-16.

Dun, G.S. Economic entomology in Papua and New Guinea, 1948-1954. Papua New 1955 Guinea agric. J., 9(3): 109-113.

Dwyer, R.E.P. Diseases of coconut in New Guinea. New Guinea agric. Gaz., 3(1): 1937 28-91.

EDEN, D.R.A. Quest for the home of the coconut. Q. Bull. South Pacific Comm., 13(3): 1963 39-42.

FENEMORE, P.G. Further instituted experiments. British Solomon Islands. Entomolo-1958 gist's Progress Report No. 6. FENWICK, D.W. Red ring of coconut, a problem for the nematologist. Indian Coconut J.,

1959 12(3 and 4): 82-86.
FONSECA, J.P. Broca dos pedunculos dos Coqueiros. Biologio, Brazil, 28(1): 20-25.

FORSYTH, J. In Report of the seventh Commonwealth Entomological Conference, Commonwealth

1960 Institute of Entomology, London.
Franssen, C.J.H. Biological control of Sexara nubila in Talaud Is. Ent. Ber., 15(4):

Frappa, G. Un nouveau parasite du cocotier à Madagascar. Agron. trop., 2(5-6): 299-

FRIEDRICHS, K. Ueber der Bekampfung des Nashornkäfers (Oryctes rhimoceros) in Samon.

1913 Tropenpflanzer, 17(10): 538.

FROGGATT, J.L. Life history of the tree hopper of eoconuts. New Guinea agric. Gaz., 1935 1(1): 16-27.

FROGGATT, J.L. Coconut leaf miner, Promecotheca papuana, New Guinea agric. Gaz., 5(1): 1939a 3-10.

FROGGATT, J.L. Copra infestation by Ephestia cautella. New Guinea agric. Gaz., 5(2): 2-5. 1939b

FROGGATT, J.L. & O'CONNOR, B.A. Insects associated with the eccount palm. Pt. II. 1941 New Guinea agric. Gaz., 7(2): 125-133.

FROGGATT, W.W. Petts and disease of the coconut palm. Sydney, Department of Agri-1911 culture of New South Wales. Science Bulletin No. 2.

FULLAWAY, D.T. & KRAUSS, N.L.H. Common insects of Hawaii. Honolulu, Tongg. 1945

- GADD, G.H. A possible physiological cause of nutfall of coconut. Trop. Agric., Ceylon
- 1923 60: 112-114.
 GANCOLLY, S.R. et al. Investigations in the shedding of buttons in the coconut: role
- GANGOLLY, S.R. et al. Investigations in the shedding of buttons in the coconut: ro 1956 of synthetic hormones. *Indian Coconut J.*, 9: 135-160.
- GATER, B.A.R. Some observations on the Malayan coconut Zygaenid. Malayan agric. 1925 J., 13(4): 92.
- GATER, B.A.R. Further observations on the Malayan eoconut Zygaenid. *Malayan agrit*. 1926 J., 14(4): 304-350.
- GIBSON-HILL, C.A. The robber crab. Malay Nature J., 3(1): 10-14.
- GIVEN, R.B. The coconut termite in the Cook Islands. Q. Bull. South Pacific Comm., 1964 14(3): 25-26.
- GOBERDHAN, L.C. Possible presence of a phytotoxin in red-ring infested trees. Nature, 1963 Lond., 197: 619-620.
- GOBERDHAN, L.C. The protection of econour roots against infection by Rhadinaphelen-1964 chus comphilins. Nematologia, 10: 353-360.
- GOONEWARDENE, H.F. Rhinoceros bectle in Ceylon. Pt. 1. Trop. Agric., Ceylon, 104: 1958 39-60.
- GREEN, E.E. McIolonthid Iarvae (Apagonia rauca) attacking ecocount palms in Ceylon, 1903 In First report of economic goology, British Museum (Nat. Hist.), p. 144. (Quoted by Theobald, F.V.)
- GREENSLADE, P.J.M. Studies in the ecology of ants. British Solomon Islands. Entomol-1964 ogist's Progress Report No. 7.
- GRESSITT, J.L. The cocount rhimserus bestle (Oryctes thinocerco), with particular reference 1953 to the Palan Itlands. Honolulu, Bernice P. Bishop Museum, Bulletin No. 212. GRESSITT, J.L. The coconut leal-mining beetle Promocathesa papuama. Papua New Guines 1959 agric. J., 12 (2 and 3): 119-148.
- HAGLEY E.A.C. The role of the palm weevil as vector of red-ring disease of coconuts. 1963 J. eeon. Ent., 56: 375-380.
- HAGLEY, E.A.C. The mechanism of transmission of Rhadinaphelenchus cocophilus by the 1965a palm weevil, Rhynchophorus palmarum. Phytopathology, 55(1): 117-118.
- HAGLEY, E.A.C. On the life history and habits of the palm weevil, Rhynchophorus pal-1965b marum. Ann. ent. Soc. Amer., 58(1): 22-28.
- HALL, W.J. Insect pests in British colonial dependencies. FAO Plant Prot. Bull., 3(12): 1955 178.
- HALL, W.J. Insect pests in British colonial dependencies. FAO Plant Prot. Bull., 6(7): 1958 102.
- HARMS, J.W. Birgus latro. Z. wiss. Zool., 140: 167-290.
- HARRIS, W.V. Termites of the Solomon Islands. Bull. ent. Ret., 49: 737-750.
- HARRIS, W.V. Termites: their recognition and control. London, Longmans. 1961
- HERMS, W.B. Diocalandra taitensis... of Fanning and Washington Islands. Philipp. J. 1926 Sci., 30(2): 243.
- Hinckley, A.D. Insects on young eoconut palms in Fiji. Agric. J., Fiji, 31: 39-41.

HINCKLEY, A.D. Parasitisation of Approxima argula Mey, (Lep. Approximate). Proc. Hawaiian ent. Soc., 1962, 18(2): 267-272.

HINCKLEY, A.D. Ecological notes on the larvae of some Pyraloid moths in Fiji. Pacific Insects, 6(2): 234-241.

HOPKINS, G.H.E. Pests of economic plants in Samoa and other island groups, Bull. ent. Res., 18: 18-32.

HOYT, C.P. Investigations of rhinoceros beetles in West Africa, Pacific Sci., 17(4); 1963a 444-451.

HOYT, C.P. Rhinoceros beetle investigations in Papua and New Guinea. O. Bull, South 1963b Pacific Comm., 13(2): 20-21.

HOYF, C.P. A parasite of the rhinoceros beetle found in Malaya. Q. Bull. South Pacific Comm., 14(1): 44, 64.

HUTSON, J.C. The coconut caterpillar (Nephantis serinopa). Trop. Agric., Ceylon, 59: 1922 21-24.

HYOE-WYATT, B.H. & MCKINLEY, K.G. Miscellameous Report No. 257. Tanganyika, Colonial Pesticides Research Unit.

IMMS, A.D. A general text book of entomology, Revised by O.W. Richards and R.G. Davies. 9th ed. London, Methuen.

JAYARATNAM, T. J. The Bethylid parasite (Perisierala) of the coconut caterpillar Nephantis. Trop. Agric., Ceylon, 97(1): 115-125.

JAYARATNAM, T.J. A study of the control of the coconut caterpillar in Ceylon and its 1941b Eulophid parasite. Trop. Agric., Ceylon, 96(1): 3-21. JOHNSON, C.G. Physiological factors in insect migration by flight. Nature, Lond., 198

1963 (4879); 423-427. JOHNSTON, A. New records of pests and diseases in S.E. Asia and Pacific region. Bangkok, 1964 FAO. FAO Technical Document No. 41.

Kalshoven, L.G.E. De plagen van de cultuurgewassen in Indonesië [Crop pests in Indonesia.]

1950-51 The Hague, van Hoeve. 2v. KALSHOVEN, L.G.E. An analysis of ethological, ecological and taxonomic data on Oriental Hispinae. Tijds. Ent., 100: 5-24.

KALSHOVEN, L.G.E. Habits and host associations of Indomalayan Rhynchophorinae, Beaufortia, 9 (96): 49-73.

KENNARD, G.H. In Report of the seventh Commonwealth Entomological Conference, Commonwealth wealth Institute of Entomology, London, p. 214-215.

KURIAN, C. Destructive pests of coconut other than Oryctes rhinoceros. Document, FAO 1963 Technical Working Party on Coconut, 1961, Bangkok.

KURIAN, C. & PILLAI, G.B. The rhinoceros beetle, Orycles rhinoceros, a menace to coconut cultivation. World Crops, 16(3): 20-24.

LAIRO, M. Rats, coconuts, mosquitoes and filariasis. In Pacific basin biogeography. Tenth Pacific Science Congress, 1961, Honolulu, Bishop Museum Press, p. 534-542.

Lange, W.H. Biology of the Marianas ecconut beetle and the introduction of its parasite. Proc. Hawaii. ent. Soc., 14(1): 143-162.

LASSALLE-SERE, R. Rat control scheme for Tahiti planters. Q. Bull, South Pacific Comm. 1955 5(2): 5-6.

- LEEFMANS, S. Gegerent over sabeltprinkbanenat cocovijanden in Nederlandsteb-Indië en bunnen 1927 paraitee [Locustidae as eoconut pests in the Dutch East Indies, and their parasites.] Meded, Inst. Plantenziekten No. 72.
- LEEFMANS, S. & AWIBOWO, R. Bestrijding van Brathartona met Dusturan [Combatting 1935 Brachartona with "dusturan."] Landbosw, 11: 1-19.
- LEPESME, P. Les insertes des palmiers. Paris, Leehevalier. 1947
- Lever, R.J.A.W. Coconut beetle of Santa Cruz group (Promesothesa opacitollis). British 1933a Solomon Islands agris. Gaz., 1(4): 11.
- LEVER, R.J.A.W. Greater spike moth (Tirathaba rufivena) and its parasites. British Sol-1933b omon Islands agric. Gaz., 1(3): 7-8.
- LEVER, R.J.A.W. Hemipterous pests of coconuts. British Solomon Islands agric. Gaz., 1933c 1(3): 2-6.
- LEVER, R.J.A.W. The green eoconut bug, Amblypelta cotophaga. British Solomon Islands 1935 agrit. Gaz., 3(2): 6-7.
- LEVER, R.J.A.W. Control of Brontispa in Celebes by Tetrastichodes. British Solomon Islands
- 1936 agric. Gaz., 3(4), Supplement: 6.
 LEVER, R.J.A.W. Economic insects and biological control in the British Solomon Islands.
- 1937 Bull. ent. Res., 28: 325-331.
 LEVER, R.J.A.W. Javan parasite of the coconut leaf-miner. Agric, I. Fiii, 9(4): 12-14.
- 1938

 LEVER, R. J.A.W. Fig and cacao moths (Ethestia) in Melanesia and Polyncsia. Agric.
- 1941a J. Fiji, 12(1): 20-21.

 LEVER, R.J.A.W. The small coconut weevil (Diocalandra). Agric. J. Fiji, 12(4): 3-4.
- 1941b

 LEVER, R.J.A.W. A new coconut pest in Singapore. Malayan agric, L. 34(2): 78-82.
- LEVER, R.J.A.W. Notes on outbreaks, parasites, and habits of the coconut moth (Artona 1953 tatoxantha). Malayan arris. I.. 36(1): 20-27.
- Lever, R.J.A.W. Immature nutfall of coconuts. World Crops, 13(2): 60-62.
- LEVER, R.J.A.W. Inseet damage to coconut palms. World Crops, 16(4): 64-68.
- LEVER, R.J.A.W. Notes on some parasites, hyperparasites and predators of coconut 1964b insects in Malaya. FAO Plant Prot. Bull., 12(2): 1-2.
- LLOYD, D.C. Significance of type of host plant-crop in successful biological control. 1960 Nature, Lond., 187(4735): 430-431.
- LLOYD, C.J. & HEWITT, P.S. Relative susceptibility to pyrethrin in oil of Colcoptera and 1958 Lepidoptera infesting stored products. *Bull. ent. Ret.*, 49: 177-185.
- MAHARAJ, S. Rhymchophorus palmarum: field studies on the life history. f. agric. Soc. Trin. 1962 Tob_{s_1} 62: 8.
- MAHARAJ, S. The development of the palm weevil in felled coconut trees. J. agric. 1964 Soc. Trin. Tob., 64: 67-74.
- MAJUMDER, S.K. & BANO, A. Toxicity of calcium phosphate to some pests of stored 1964 grain. Nature, Lond., 202 (4939): 1359-1360.

MARCONI, F.A.M. As gagartas das palmeiras. Biologico, Brasil, 18(6): 103-107.

MARQUES DE AZEVEDO, L.A. Gafanboto nocivo a palmeira (Cocos nucifera), Tropidacris. 1922 Rio de Janeiro.

MARSCHALL, K.J. The history of normal and diseased reserve tissue in rhinoceros beetle 1963 (Orystes spp.). J. Insect Path., 5(1): 39-55.

Marshall, G.A.K. Insets of Samoa. Pt. 9, Fasc. 5. Curculionidae. London, British 1931 Museum (Nat. Hist.).

MATHEN, K. Observations on Stephanitis typicus - a pest of eoconut palms. Indian Co-1960 const J., 14(1): 8-27.

MEDINA DUNO, J. Cómo evitar la propagación del anillo rojo del coco. Agricultar 1959 Venezolano, 23 (214): 21.

MEXEONGER, B. Coconut patts of Thailand. Document, FAO Technical Working Party 1963 on Coconut, 1961, Bangkok.

MENON, K.P.V. & PANDALAI, K.M. The coconut palm: a monograph. Etnakulam. Indian

1958 Central Coconut Committee.
Meano, G. Report on ecoconut Zygaena in province of Palawan. Philipp. J. Agric.

Mexico, G. Report on eoconut Zygoena in province of Palawan. Politips. J. Agric., 1938 9(1): 31-35.

Mo, T.T. The occurrence of two strains of Brontispa longistima (Gestro) based on resis-

1965 tance and non-resistance to the parasite Tetratilibodes brontlipae (Ferr.) in Java.

Bull. ent. Res., 55: 609-614.

MONTENECRO, A. How to prevent rat damage in a coconut plantation. Coffee/Cocoa J., 1962 5: 192-197.

MOUTIA, L.A. Phytophagous Acarina and their predators in Mauritius. Bull. ent. Res.,

MOUTIA, L.A. Phytophagous Acarina and their predators in Mauritius. Buil. etc. Res. 1958 49: 59-75.

MOUTIA, L.A. & MAMET, R. Review of twenty-five years of economic entomology 1946 in Mauritius. Bull. ent. Res., 36: 439-472.

MUNRO, J.W. Report on a survey of infestation of grain by insects. London, HMSO.

NAIDU, G.V.B. A note on rat control in eoconut nurseries. Mysore agris. J., 35: 86-88. 1960

NAKATA, S. Some notes on the Phasmatodea in Oceania. Pacific Islands Monograph, 1962 (2): 107-121.

NIRULA, K.K. et al. Comparative studies with different insecticides for the control of 1951a grubs. Orystes rbinoceros. Indian Cosonut J., 4(2): 49-56.
NIRULA, K.K. et al. Control of coconut cateroillas (Nebantis serinoba) by DDT sprays.

1951b Indian Coconul J., 4(4): 225-234.

NIBULA K K et al. A new pest of excount palms in India (Leucotholit concolhora). Indian

NIRULA, K.K. et al. A new pest of eoconut palms in India (Lowopholis concephora). Indian 1952 Coconut J., 5(3): 137-140.
NIRULA, K.K. et al. Some investigations on the control of termites. Indian Coconut J.,

1953 7(1): 26-34.

Nirula, K.K. et al. The green muscardine disease (Metarrhizium anisopliae). (Investiga-

1955a tions, Orystes rhinoseros). Indian Coconut J., 9(1): 1-10.
NIRULA, K.K. et al. Important pests of the coconut palm: check list. Indian Coconut J.,

1955b 8(3): 1-13.

NIRULA, K.K. et al. Oryctes rhinoceros: life history. Indian Coconut J., 8(4): 1-20. 1955c

- NIRULA, K.K. et al. Parasites and predators of Oryctes rbinoceros. Indian Coconut J., 1955d 9(1): 30-79.
- NIRULA, K.K. et al. Investigation on the pests of coconut palm: Nephantis serinopa. 1956a Indian Coconut J., 9(2): 101-131.
- NIRULA, K.K. et al. Investigation on the pests of coconut palm: Nephantis serinopa. 1956b Indian Coconut J., 9(3): 174-201.
- NIRULA, K.K. et al. Investigations on the pests of eccount palm; Rhynchophorus ferru-1956c gineus. Indian Coconut J., 9(4): 229-247; 10(1): 28-44.
- NIRULA, K.K. & MENON, K.P.V. Insect pests of the coconut palm in India. Pt. 1. 1960 World Crops, 12(6): 213-214.
- Nye, I.W.B. Control of the coconut pest Melittomma insulare (Coleopt., Lymexylidae) 1961 in the Seychelles. Bull. mt. Res., 52(3): 489-499.
- O'CONNOR, B.A. The coconut leaf-miner (Promecotheca). New Guinea agric. Gaz., 6(2): 1940 20-30.
- O'CONNOR, B.A. Some insect pests of Tonga. Agric. J. Fiji, 20(2): 47-57.
- O'CONNOR, B.A. Premature nutfall of coconuts in the British Solomon Islands Protector-1950 ate. Appendix by Leach, R. Agric. J. Fiji, 21(1-2): 21-42.
- O'CONNOR, B.A. The rhinoceros beetle (Oryetes rhinoceros) in Fiji: history. Agric. J. 1953 Fiji, 24(1-2): 35-46.
- O'CONNOR, B.A. Notes on the coconut stick insect, Graeffea crowni. Agric. J. Fiji, 1954a 25(3-4): 89-92.
- O'CONNOR, B.A. The thinoceros beetle (Orystes rhimoceros) in Fiji: notes on the control 1954b campaign in Fiji. Agric. J. Fiji, 25(3-4): 84-88.
- O'CONNOR, B.A. Control of Orycles rhimoerres by the use of insecticides. Agric. J. Fiji, 1957 28(1): 15-18.
- O'CONNOR, B.A. Aerial spraying to control Graeffea crossani. Agric. J. Fiji, 29(4): 138-1959a 141.
- O'CONNOR, B.A. The coconut tree-hopper Sexara and its parasites. Papua New Guima 1959b agric. Gaz., 11(4): 121-125.
- ORIAN, A.J.E. Report on a visit to Diego Garcia, Chagos. Rev. agric. sucr. Maurice, 1959 38(3): 127-143.
- ORIAN, A.J.E. Notes on the Hispine beetle Brontispa limbata (Waterh.). Rev. agric. sucr. 1961 Maurice, 40(4): 193-195.
- OTANES, F.Q. Locust outbreaks probable between 1941 and 1944. National Rice and 1941. Corn Co-operative, 1(4): 281-286.
- OTANES, F.Q. Noteworthy campaigns against... lossusts during a quarter of a century, 1930-54.
 1956 Manila, Department of Agriculture.
- OWEN, R.P. Inuest pests of the Trust Territory of the Pacific Islands. Bangkok, FAO. FAO 1964 Technical Document No. 40.
- PAGDEN, H.T. & LEVER, R.J.A.W. Insects of the coconut palm in the British Solomon 1935 Islands. British Solomon Islands agric. Gaz., 3(1): 2-22.
- Paine, R.W. Entomological notes (Xyleborus). Agric. J. Fiji, 7(1): 39. 1934
- PAINE, R.W. The control of the coconut spike moth (Tirathaba trichogramma). Department 1935 of Agriculture, Fiji. Bulletin No. 18.

PHILLIPS, J.S. Immature nutfall of coconuts in the Solomon Islands. Bull. ent. Res., 1940 31: 295-316.

PHILLIPS, J.S. A search for parasites of Dasynine bugs in the Netherlands Indies. Trans. 1941 R. ent. Soc., Lond., 91(5): 119-144.

PHILLIPS, J.S. Immature nutfall of coconuts in the British Solomon Islands Protectorate, 1956 Bull. ent. Res., 47: 875-895.

Piggorr, C.J. The problems of coconut production on Seychelles "Outer Islands". 1961 East Afric. agric. for. J., 27(2): 127-130.

PILLAI, G. & KURIAN, C. Effect of dieldrin concentrations on Nephantis serinopa larvae. 1960 Indian Coconut J., 13(4): 154-162.

PLANK, H.K. Life bistory, babits and control of the coconut rhinoceros beelle in Puerto Rico. Puerto 1948 Rico, Federal Experiment Station. Bulletin No. 45.

Posvo, S.A.S. & Mo, T.T. De bestrijding van een slakrups, Darna (Orthorratpedo) catenatus (Sn.), op klapper in het Palu-dal [The control of Darna catenatus (Sn.) on coconut trees in the Palu valley, central Celebes.] Landbouw, 22: 69-81.

PRADHAN, S. Pestieide hazards to man under present day conditions in India. *Indian* 1965 J. Ent., 27(1): 1-20.

PRATT, F.C. Note on the rat as a coconut pest. Agric. Bull. F.M.S., 2(7): 192.

REYNE, A. On the food habits of the coconut crab with notes on its distribution. Arch. 1939 neerl. Zool., 3: 283-320.

REYNE, A. Studies on a serious outbreak of Aspidiotus destructor rigidus in the coconut-1948 palms of Sangi (North Celebes). Tijdschr. Ens., 89: 83-120.

RICHAROS, O.W. & HERFORD, G.V.B. Insects found associated with caeao, spices and 1930 dried fruits in London warehouses. Ann. appl. Biol., 17(2): 367-394.

RIDLEY, H.N. On the habits of the earinga (Formica gracilipes Gray). J. (Straits Bramb) 1890 R. Asiatic Soc., 1890.

RISBEC, J. Note préliminaire sur les principaux parasites du cocotier aux Nouvelles-1935 Hébrides. Ann. Soc. ent. France, 104: 159-173.
RISBEC, J. Observations sur les parasites des plantes cultivées des Nouvelles-Hébrides.

1937 Fanne Colon. franc., Parir. 6(1): 214.
SARAIVA, A.C. A preliminary list of the insect pests of crops and fruit trees in Portu-

1939 guese East Africa. J. ent. Soc. S. Africa, 11: 101-114.
SAYEED, P.M. et al. Some observations on the shedding of buttons in the coconut. Indian

1955 Caosnut J., 8(2): 49-57.
SHANTA, P. & MENON, K.P.V. The cowpea as an indicator plant for coconut wilt disease.
1960 Virology, 12(2): 309-310.

SHASHI KANTA et al. Laboratory evaluation of toxicity of pesticides to palm mite Raoiella 1963 indica Hirst. Indian Coronut J., 16(2): 63-66.

SHELLDRICK, R.P. A note on recent investigations into palm poisoning. J. W. Afr. 1963 Inst. Oil Palm Res., 4(3): 101-103.

SIMMONDS, F.J. Investigations of possible biological control of Melittomma insulare in 1956 the Seychelles. Bull. ent. Res., 47: 685-702.

SIMMONDS, F.J. Biological control of coconut scale (Aspidiotus destructor) in Principé, 1960 Portuguese West Africa. Bull. ent. Res., 51: 223-237.

- SIMMONDS, F.J. Biological control of pests in the tropies. Trop. Sci., 5(4): 200-207, 1963
- SIMMONDS, H.W. Report on mission to New Guinea, Bismarcks, Solomons and New Hebrides. 1924 Suva. Legislative Council Paper, Fiji.
- Simmonds, H.W. Problems in biological control. Trop. Agric., Trin., 7(8): 215-219.
- SIMMONDS, H.W. Coconut petts and diseases. Department of Agriculture, Fiji. Bulletin 1938 No. 20.
- SIMMONDS, H.W. Biological control of the rhimocros beetle (Oryetes thinoceros). Department 1941 of Agriculture, Fiji. Bulletin No. 21.
- SIMMONDS, H.W. On the introduction of Scolia ruftiornis F. into Western Samoa for the 1949 control of Orycles rbinaceras. Bull. ent. Res., 40: 445-446.
- Simmonds, H.W. Premature nutfall in Taveuni. Agric. J. Fiji, 22(1): 22-24. 1951
- SIMON THOMAS, R.T. Check list of pests on some crops in West Irian. Landbouw Ser. No. 1. 1963
- SINGH, S.R. Notes on life history of eoconut leaf moth (Agonoxona arguala). Agric. J. 1951 Fiji, 23(3 and 4): 106-107.
- STRECKER, L. et al. Pacific island rat ecology. Honolulu, Bernice P. Bishop Museum. 1962 Bulletin No. 225, 274 p.
- SURANY, P. Diseases and biological control of rhinoceros beetle. Nouméa, South Paeifie Com-1960 mission. Technical Report No. 128.
- SZENT-IVANY, J.J.H. New inseet pest and host plant records in... Papua and New Guinea, 1956 Papua New Guinea agric, I., 11(3): 82-87.
- SZENT-IVANY, J.J.H. & CATLEY, A. Notes on Amblypelta lutescens papaensis. Papua New 1960 Guinea agric., J., 13(2): 59-65.
- TAIT, E.M. Some notes on the life history and habits of Theraptus. Bull. ent. Res., 1954 45: 429-432.
- TAMMES, P.M.L. Over de factoren welke de vruchtdracht van den klapper bepalen [Fac-1937 tors determining yield of eoconut palms.] Landbow, 13: 260-269.
- TAYLOR, T.H.C. Early nutfall from eccount palms in Fiji. Department of Agriculture, 1930 Fiji. Bulletin No. 17.
- TAYLOB, T.H.C. The biological control of an insect in Fiji. London, Imperial Institute of 1937 Entomology.
- TAYLOR, T.H.C. The biological control of insect pests. Ann. appl. Biol., 42(1): 190-1955 196.
- TAYLOR, T.H.C. & PAINE, R.W. The eampaign against A. destructor in Fiji. Bull. ent. 1935 Res., 26: 1-102.
- THEOBALD, F.V. See Green, E.E. 1903
- TINKER, S.W. Pacific crustacea. Rutland, Vermont, and Tokyo, C. Turtle Co. 1965
- TOTHILL, J.D. A reconnaissance survey of agricultural conditions in the British Solomon Islands 1929 Protectorate, Suva.

TOTHILL, J.D., TAYLOR, T.H.C. & PAINE, R.W. The coconut moth in Fiji. London, Im-1930 perial Institute of Entomology.

UVAROV, B.P. Locusts and grasshoppers. London, Imperial Institute of Entomology. 1928

VANDERPLANK, F.L. Field experiments with spray nozzles in the tropies. World Crops, 1954 6: 488-490.

VANDERPLANK, F.L. The assassin bug, Platymerus rhadamanthus Gerst., a useful predator 1958a of Oryctes boas and O. rhinoceros. J. ent. Soc. S. Africa, II, 21(2): 309-314.

VANDERPLANK, F.L. Studies on the eoconut bug *Pseudotheraptus wayi* Brown in Zanzibar. 1958b Pt. I. A method of assessing the damage caused by the insect. *Bull. ent. Res.*, 49: 559-584.

VANDERFLANK, F.L. Bionomics and ecology of the red tree ant (Oerophylla smaragdina). 1960a J. Anim. Evol., 29(1): 15-53.

VANDERPLANK, F.L. Studies on the coconut bug Psudstberaptus wayi Brown in Zanzibar. 1960b Pt. II. Some data on yields of coconuts in relation to damage caused by the insect. Bull. ent. Res., 50: 135-149.

VANDERPLANK, F.L. Studies on the coconut bug. Pseudatheraptur wayi Brown in Zanzi-1960c bar. Pt. III. A selective residual insecticidal formulation and its effect on the ecology of the insect. Bull. ent. Res., 50: 151-164.

Vanderplank, F.L. Availability of the eoconut bug Pseudotheraptus wayi (Coreidae). 1961 Bull. ent. Res., 51: 57-60.

VAN DER VECHT, J. Bestrijding van Brachartona met Dursturan [The eontrol of Brachar-1939 tona with "dusturan."] Landbouw, 15.

VAN DER VECHT, J. Het verband tusschen populatie-dichheid van gastheer en parasiet 1947 bij sommige tropische insecten [Relation between population density of host and parasite among some tropieal insects, *Tijdatir. Eur.*, 88: 427-434.

VAN DER VECHT, J. The coconnu leaf moth (Artona eatoxantha Hampt.), Part. J. Life 1950 bitroy and habits of Artona catoxantha, its paraites and hyperparites. Bogor, Agricultural Research Station. General Contribution No. 110.

VESEY-FITZGERALD, D. The control of Coccidae on coconut palms in the Scychelles. 1941a Bull. ent. Ret., 31: 253-282.

VESEY-FITZGERALD, D. Melistomma insulare, a coconut pest in the Seychelles. Bull. ent. 1941b Ret., 32: 383-402.

VESEY-FITZERALD, D. Review of biological control of Coccids on coconut palms in the 1953 Seychelles. Bull. ent. Ret., 44: 253-282.

VESTAL, E. Control of coconut beetles and weevils in Thailand. FAO Plant Prot. 1956 Bull., 5(3): 37-44.

WAY, M.J. An insect pest of coconuts and its relationship to certain ant species. Nature, 1951 Lond., 168: 302-303.

WAY, M.J. The relationship between certain ant species with particular reference to bio-1953a logical control of the Coreid Theraptus. Bull. eat. Res., 44: 669-691.

WAY, M.J. Studies of *Theraptus* sp. (Coreidae), the cause of the gumming disease of co-1953b conuts in East Africa. *Bull. ent. Res.*, 44: 657-667.

WAY, M.J. Studies of the life history and ecology of the ant Oecophylla longinoda (Latreille).

1954 Bull. ent. Res., 45: 93-112.

- WHEATLEY, P.E. Rearing Pseudotheraptics ways, a pest of coconuts in E. Africa, and an 1961 evaluation of its susceptibility to various insecticides. Bull. ent. Rev., 51: 723-729.
- Wilson, M.E. Laboratory studies on the life history of the palm weevil Rhynchophorus 1962 palmarum. J. agric. Soc. Trin. Tob., 61: 1-9.
- WILSON, M.E. Investigations and development of the palm weevil (Rhynchophorus pal-1963 marson). Trop. Agric., Trin., 40(3): 185-196.
- WOLCOTT, G.N. An economic entomology of the West Indies. San Juan, Entomological 1933 Society of Puerto Rico.
- Wood, B.J. Severe outbreaks of eaterpillar on oil palm estates in Malaya induced by the use of residual contact insecticides. Proceedings of the Twelfth International Congress of Entomology, 1964, p. 574-575. London.
- WYNIGER, R. Pests of crops in warm climates and their control, with Appendix V. Control 1962 measures. Basle, Verlag für Recht und Gesellschaft A.G.
- ZACHER, F.H. Die Schädlinge der Kokospalmen auf der Südseeinseln. Arb. Kaiser
- 1913 Biol. Land-u. Forstw., 9(1): 73-119.
- ZIMMERMAN, E.C. Insects of Hawaii. 8. Lepidoptera: Pyraloidea. Honolulu, University 1958 of Hawaii.

INDEX

aberrans Strobl, Tricholyga	78	Apanteles tiratbabae Wilkinson 72, 73	
Acanthopsyche cana Hampson	78	Aphelencoides 143	
Acanthopsyche hypoleuca Hampson	78	Aphididae 45	
Achresocharis	106	Aphytis chrysomphali Mercet 48	
Achrysocharis promecothecae Ferrière	106	apicalis Westwood, Spathius 124	
Acrididae	1.4	Apleurotropis lalori Girault 104	
Acridotheres tristis	146	Apogonia cribricollis Burmeister 136	
Acritocera negligens Butler	61	arcufer Chapin, Callimerus 91, 23	
Actia painei Grosskey	70	arenosella Walker, Batrachedra 63	
Adoretus	136	argaula Meyrick, Agonoxena 69, 181	
advena Waltl, Abasverus	154	Argyrophylax (Erycia) basifulva Bezzi 72, 73	
Aedes (Stegomyia) polynesiensis Marks	150	Artona catoxantha Hampson 86	
afzelii Fahracus, Rhinostomus	121	artonae Wilkinson, Apanteles 91	
Agapophyta hiptoretata Boisduval	43	.Aspergillus 80, 154	
Agathis	70	Aspidiotiphagus citrinus Crawford 48	
Agonoxena arganla Meyrick	69, 181	Aspidiotus destructor Signoret	
Agonoxena pyrogramma Meyrick	70	46, 50, 140, 141, 146, 151, 181	
agonoxenae Kerrich, Elachertus	70	Aspidiotus destructor rigidus Revne 48	
Agonoxenidae	62	Attegopterys: nipae van der Goot 45, 140	
Abasperus advena Waltl.	154	astyra Godart, Brassolis 57	
Alaus	128	ater de Geer, Dermestes 155	
albiceps Macquart, Degeeria	91	Atractomorpha psittacina Serville 23	
albiguttata Snellen, Chalcocelis	82	Atractomorpha psittacina Serville 23 Aularches miliaris Linnaeus 20	
albipes Smith, Technomyrmes:	51, 141	aurantii Maskell, Aonidiella 48	
Aleurodicus destructor Mackie	52	axiagasti Ferrière, Anastatus 37, 43	
Aleurodidae (Aleyrodidae)	5.2	Axiagastus cambelli Distant 43, 182	
Allodontotermes morogoroensis Harris	35	Aztrea cartifex Forel 140, 182	
	35, 187		
Amathusia phidippus Linnaeus	54	bambusae Moore, Telicota 60	
Amathusiidae	54	barbirostris Fabricius, Rhina 120	
Amblypelta 38, 40, 41, 1		barbirostris Fabricius, Rhinostomus 120, 186	
Amblypelta cocopbaga China 36,		basifulva Bezzi, Argyrophylan (Eryeia) 72, 73	
Amblypelta lutescens Distant	38	hassiana, Beauveria 91	
anachoreta Burmeister, Strategus	135	Batrachedra arenosella Walker 63	
Anastatus	38, 56	Batrachedra peroptusa Meyrick 64	
Anastatus axiagasti Ferriere	37, 43	Beauveria hassiana 91	
	56, 127	bellicosus Smeath, Macrotermes 35	
Anoplolepis longipes Jerdon	140	Bessa 84	
antilope Bottcher, Sarcophaga	81	Bessa remota (Tachinidae) 90	
antiqua Weise, Promecotheca	102	Bessa (Ptychomyia) remota Aldrich 91, 93, 160	
Antrocephalus	82	bezziana Baranov, Stomatomyia 69	
Antrocephalus renalis Waterston	72	bicinota de Meijere, Pentatomophaga 38	
Aonidiella aurantii Maskell	48	bicolor Waterston, Leefmansia 26	
	70, 84	hipunctata Boisduval, Agapophyta 43	
Apanteles artonae Wilkinson	91	Birgus latro Linnaeus 144, 147	
Apanteles parasae Rohwer	81	biroi Desneux, Microcerotermes 32	
Apanteles taragnae Vicrock	68	blackburni Butler, Hedylepta 74, 75	

to Estadaine Ometre	134	Chartennists imme Basses and
boas Fabricius, Orycles Brachartona catoxantha Hampson	86, 21	Chaetexorista javana Brauer and Bergenstamm 80, 81
Brachymeria 56, 70,		Chaetogidia 75
Brachymeria euploeae Westwood	83	Chalcocelis albiguttata Snellen 82
Brachymeria nephantidis Gahan	69	chalcostelidis Wilkinson, Fornicia 83
Botyris necans	91	Chalconycles catori Jordan 93
Bracon	70, 75	Chalcosoma 135
Bracon bebetor Say	153	Chelonus 25
Brassolidae	56	Chilocorus 49, 50, 141
Brassolis astyra Godart	57	Chilocorus nigritus Fabricius 48
Brassolis sophorae Linnaeus	56, 182	Chilocorus politus Mulsant 48
brevicornis Wesmael, Microbracon	68	chinensis Thunberg, Oxya 23
Brontispa froggatti Sharp	107	Chrysomphalus ficus Ashmead 49
Brontispa gleadowi Weise	112	Chrysididae 80
brontispae Ferrière, Haeckeliana 109,		Chrysophilus ferruginosus Wiedemann 124
Brontispa limbata Waterhouse	112	Chrysis 82, 84
Brontispa longissima Gestro 107, 1	12, 182	Chrysis shanghaiensis Smith 81
Brontispa mariana Spaeth	109	chrysomphali Mercet, Aphytis 48
	09, 112 34	citrinus Crawford, Aspidiotiphagus 48 Cleridae 153
hugnioni Holmgren, Microcerotermes		
buxi Bouché, Pinnaspis	50, 141	
Cacatua ducorpsii	146	Closterocerus splendens Kowalski 104, 105 Coccidae
Catatha aucorpsis Catatha galerita triton	146	
Cadra cantella Walker 76, 1		cocopbaga China, Amblypelta 36, 38, 181 cocopbaga Newport, Graeffea 27, 159
Cadarcia leefmansi Baranov	91	cocophilus Cobb, Rhadinaphelenchus 143, 186
Callimerus arcufer Chapin	91, 23	Coelaenomenodera 106
cambelli Distant, Axiagastus	43, 182	Coelaenomenodera elaeidis Maulik 106
cana Hampson, Acanthopsyche	78	coeruleipennis Blanchard, Promecotheca
Cardiodactylus novae-guineae Haan	26	98, 101, 103, 105
Carpophilus dimidiatus Fabricius 1	53, 154	Coleoptera 94, 153
	40, 182	Comperiella smifasciata Ishii 48
casei Linnaeus, Piophila	153	complexa Butler, Tirathaba 71, 72, 73
castaneum Herbst, Tribolium	155	concophora Burmeister, Leucopholis 136, 183
Castnia daedalus Cramer	84, 182	Contheyla rottenda Hampson 82
Castnia licus Drury	85, 183	Coptatermes ceylonicus Holmgren 32
Castniidae	84	Coptotermes curvignathus Holmgren 31
Catantops biomilis Serville	23 23 128	Coptotermes truncatics Wasmann 32
Catantops splendens Thunberg	23	corbetti Tams, Mabasena 77, 78
Catastopus	128	Corcyra cephalonica Stainton 152
catenatus Snellen, Orthocraspeda	83 93	Cordsceps 81 Coreidae 36
catori Jordan, Chalconycles	23	Coreidae 36 coriacea Linnaeus, Sexara 23, 24
(Artona) Reachartona	86, 90	
catoxanthae Ferrière, Eupelmus	81	Corone Gyllenhal, Homalinotus 124
caudatus Betlese, Typblodromus	142	Cosmopteryzidae 63
cautella Walker, Cadra 76, 1		Cossidae 61
cautella, Ephestia	152	Cotterellia 107
centaurus Sternberg, Oryctes	134	cribricollis Burmeister, Apogonia 136
cephalonica Stainton, Corcyra	152	cristata Linnaeus, Tropidacris 17
Cephalosporium lecanii	51	cronani Le Guillou, Graeffea 27, 29, 159
Ceraphron	91	Cryptognatha nodiceps Marshall 48
Cerataphis lataniae Boisduval	45	Cryptophasiidae 66
Cercopithecus mona	151	Crysomphalus ficus Ashmead 49
Ceromasia sphenophori Villeneuve	120	Cucujidae 154
cervinodes van der Wulp, Exorista	83	cumingi Baly, Promecotheca 105, 106
ceylonicus Holmgren, Coptotermes	32	Curculionidae 113
ceylonicus Holmgren, Nasutitermes	34	currignathus Holmgren, Coptotermes 31
Chaetevarista		Cyclodes amma van der Hoeven 94

daedalus Cramer, Castnia	84. 182	Eutermesvandiniensis Hill 35
dallatorreana Hoffer, Stictotrema	26	Exochomus 49, 50, 141
decoratus Redtenbacher, Segestes	26	Exerista cervinodes van der Wulp 83
Degerria albiceps Macquart	91	Exorista quadrimaculata Baranov 78
	84	exulans exidans, Rattus 150
deleter Tams, Trogocrada	81	
dentilinella, Phycita	155	
Dermestes ater de Geer		exulans, Rattus 150
Dermestidae	155 52	6 1 OF 1 PL 1-1
destructor Mackie, Aleurodicus	52	ferrugineus Olivier, Rhynchophorus
destructor Signoret, Aspidiotus		113, 115, 186
46, 140, 141,	, 151, 181	ferruginosus Wiedemann, Chrysophilus 124
detorquens Walker, Technomyrmex	141	ficus Ashmead, Chrysomphalus 49
devastans (Dist.), Pseudotheraptus	185	fijiensis Hirst, Tetranychus 143
Devorgilla (Nemeritis) palmaris		flavo-orbitalis Cameron, Zaleptopygus 25
Wilkinson	72, 73	Formicidae 138
Dialestrodes elongatus Dumbleton	54	Fornicia chalcoscelidis Wilkinson 83 froggatti Sharp, Brontispa 107
dimidiatus Fabricius, Carpophilus	153, 154	froggatti Sharp, Brontispa 107
Dimmockia	107	Frontina 75 frumenti Fabricius, Diocalandra 122
Dimmockia javanica Ferrière	106	
Diocalandra frumenti Fabricius	122	fuscicanda Botteher, Sarcopbaga 115
Diocalandra stigmaticollis Gyllenha	d! 183	
Diocalandra taitensis Guérin	121, 122	galerita triton, Cacatua 146
Diptera	141	Galleriidae 71
Doirania leefmansi Waterston	26	Gangara thyrsis Fabricius 60
Dorylus orientalis Westwood	141	ghesquierei Tams, Pimelephila 25
dworpsii, Casatua	146	gigas Laporte de Castelneau, Oryctes 134
dux Drury, Tropidacris	17	gilvus Hagen, Macrotermes 35
Dynastidae	125	gleadowi Weise, Brontispa 112
dysmephila Trimen, Zophopetes	61	gnu Mohner, Orycles 134
		Goryphus 82, 83, 91
Echthromorpha	72, 75 151	Goryphus nursei Cameron 69
edidis, Pteropus	151	Goryphus oxymorus Tosquinet 81
edwardsii, Pteropus	151	Graeffea cocophaga Newport 27
Elachertus agonoxenae Kerrich	70	Graeffea crouani Le Guillou 27, 29, 183
elaeidis Maulik, Coelaenomenodera	106	Graeffea lifuensis Sharp 29 Graeffea seysbellensis Ferrari 29 granaria Forskal, Schistocerca 17
Elasmus hispidarum Ferrière	101	Graeffea seychellensis Ferrari 29
Elasmus nephantidis Rohwer	69	gregaria Forskål, Schistocerca 17
Elis romandi Saussure	128	gremius Fabricius, Suastus 61
elongatus Dumbleton, Dialeurodes	54	grossipes Fabricius, Physomerus 41
elutella Hubner, Ephestia	152	Gryllidae 26
Elymnias hypermnestra Linnaeus	57	
Ephestia	76, 152	Hadronotus bomoeoteri Nixon 38
Ephestia cantella	152	Haeckeliana brontispae Ferrière 109, 112, 113
Ephestia elutella Hubner	152	Halictophagu zanziharae Bohart 41
ephrates Holmgren, Nasntitermes	34	helietor Say, Bracon 153
erebus Burm., Orycles	183	Hedylepta blackburni Butler 74, 75
Erionota thrax Linnaeus	61	Hemiptera or Rhynchota 36
erithaeus, Psittaeus	146	Hesperiidae 58, 61
	115	Heterocera 61
erratica Smith, Scolia	51	
Eucalymnatus tessellatus Signoret	26	Heteroptera 36 Hidari irara Moore and Horsfield 58, 60
Eumossula	81	Hinatidium 107
Eupelmus catoxantbae Ferrière	81 91	
Euplectromorpha		
Euplectrus	84, 91	hispidarum Ferrière, Elasmin 101
euplocae Westwood, Brachymeria	83	Hololeptis 128
Eurytoma	80	
Eurytoma monemae Ruschka	81	bomoeoceri Nixon, Hadrometus 38
Eurytoma promecothecae Ferriere	104 135	Homophylotis 93 Homophera 45
Eurytrachehu		Homoptera 45

	INL	DEX	1//
bumilis Serville, Catantops	23	Macrotermes gilvus Hagen	36
hyalineata branneri Ckll., Trigona	188	Macrotermes nigeriensis Sjöstedt	35
Hyalospila ptychis Dyar	79	maculatus Ferrière, Neoplectrus	80
Hymenoptera	138	Mahasena corbetti Tams	77, 7
hypermnestra Linnacus, Elymnias	57		37-38
	78	malayensis Ferrière, Ocencyrtus	65
hypolenca Hampson, Acanthopsyche	7.0	marcescens, Serratia	109
ignevena Hampson, Tirathaba	73	mariana Spaeth, Brontispa	105
indica Hirst, Raoiella	142	marjoriae Snyder, Schedorbinotermes	32
indicus, Varanus	150	Mecopus	121
insulare Fairmaire, Melittomma	94	megacephala Fabricius, Pheidole	38, 7
irana Moore and Horsfield, Hidari	58, 60	Megacrania phelaus Westwood	31
		melanocephalum Fabricius, Tapinoma	
iridescens Bethune-Baker, Lermana 92,		melanopyga Wiedemann, Xanthozono	
Ischnaspis longirostris Signoret	49, 50 31	Melittomma	<u>32, 134</u>
Isoptera	31	Melittomma insulare Fairmaire	24
iarana Brauer and Bergenstamm,		Melolonthidae	136
Chartexorista	80, 81	mercator Fauvel, Oryzaephilus	154
iavanica Ferrière, Dimmockia	106	Metarrhizium anisopliae	56, 127
iaranus Erichson, Plaesius	124	Meteorus	64, 72
aramo ratetto-sti, i taksimi		Microbracon	75
Kalotermitidae	35	Microbracon brevicornis Wesmael	64, 72 75 68 32 34 34
karnyi Leefmans, Sexara	24	Microcerotermes biroi Desneux	32
		Microcerotermes bugnioni Holmgren	34
lalori Girault, Apleurotropis	104	Microcerotermes peraffinis Silvestri	34
Lanelater	128	Microcerotermes piliceps Snyder	32 150
lataniae Boisduval, Cerataphis	45	micronesiensis, Rattus exulans	150
laticeps Wasmann, Nasutitermes	34	Microphanurus painei Ferrière	43
latreillei Perty, Tropidacris	20	migrans, Milrus	146
latro Linnaeus, Birgus	44, 147	migratoria manilensis Meyen, Locusto	
lecanii, Cephalosporium	51	migratoria migratorioides Reiche a	
Leefmansia bicolor Waterston	26	Fairmaire, Locuta	16
leefmansi Baranov, Cadurcia	21	miliaris Linnaeus, Aularches	20
leefmansi Waterston, Doirania	26	Milvus migrans	146
lepida Cramer, Parasa	81	mona, Cercopithecus,	151
Lepidoptera	54, 152	monemae Ruschka, Eurytonia	81
Leucopholis concophora Burmeister	36, 183	monoceros Olivier, Oryctes	134
Levuana	92, 143	morleyi Ashmead, Spilochalcis	56
Leonana iridescens Bethune-Baker 22,	93, 146	morogoroensis Harris, Allodontermes	35
leveri Günther, Ophicrania	29, 183	mindella Walker, Tirathalia	7.3
lieus Drury, Castnia	85,. 183	Muridae	147
lifuensis Sharp, Graeffea	29	Mycteromyiella lactifica (Mesnil)	183
Limacodidae	78, 82	Myrmecominesis rubrifemur Mickel	29
limbata Waterhouse, Brontispa	112		
Lindorus Iophanthar Blaisdell	48	nana Zehntner, Trichogrammatoidea	109
Locusta migratoria manilensis Meye	n 15	nararia Moore, Macroplectra	80, 82
Locusta migratoria migratorioides Rei		Nantitermes ceylonicus Holmgren	34
and Fairmaire	16	Nasatitermes ephrates Holmgren	3.4
longinoda Latreille, Occopbylla 40, 1		Nasutitermes laticeps Wasmann	34
longipes Jerdon, Anoplolepis	140	Nasutitermes morarum-behridarum Holi	ngren 34
ongirostris Signoret, Ischnaspis	49, 50	Natada	80
longissima Gestro, Brontispa 107, 1	09, 182	necans, Botyris	91
ophanthae Blaisdell, Lindorus	48		153, 154
Lucanidae	135	Necrobia rufipes de Geer	153, 154
utescens Distant, Amblypelta	38	negligens Butler, Acritocera	61
Lymexylonidae	94	Neochryopus	128
i i i i i i i i i i i i i i i i i i i	24	Neoplectrus maculatus Ferrière	80
	70	Nestermes rainbowi Hill	35
Macrocontrue			
Macrocentrus Macroplectra nararia Moote 80	81, 82	Nephantis 68,	

INDEX

nephantidis Gahan, Brachymeria	69	painti Crosskey, Actia 70
nephantidis Rohwer, Elasmus	69	painei Ferrière, Microphanurus 43
nephantidis Muesebeek, Perisierola	69	Paguridae 144
Nesopimpla	75	palmaris Wilkinson, Dervorgilla
nigeriensis Sjöstedt, Macrotermes	35	(Nemeritis) 72, 73
nigricornis Burmeister, Valanga	21	palmarum, Pestaloptiopsis 82, 104, 105
nigritus Fabrieius, Chilocorus	48	palmarson Linnaeus, Rhynchophorus
nipae van der Goot, Astegopteryx 45,	140	116, 120, 143, 186
nitens Walker, Setora 71	3, 81	palmarum Moore, Telicota 60
nitida Chapin, Telsimia	50	Papuana 135
Nitidulidae 48, 98, 153,	154	papuana Csiki, Promecotheca 102, 104, 185
Noctuidae	94	papuanus Kirsch, Rhynchophorus 116, 187
nodiceps Marshall, Cryptognatha	48	Parabillaea rhynchophorae Blanchard 117
Nomadacris septemfasciata Serville 21,	146	Paranastatus 29
novarum-behridarum Holmgren,		Parasa 80
Nasutitermes	34	parasae Rohwer, Apanteles 81
novae-guineae Haan, Cardiodactylus	26	Parasa lepida Cramer 81
novae-guineae Brancsik, Sexara	24	Parena 69
nubila Stål, Sexava	24	parvulus Ferrière, Pediobius 91, 101, 104, 106
mersei Cameron, Goryphus	69	Patanga succineta Linnaeus 15, 23, 184
	_	Pediculoides ventricosus Newport 142
obesus Rambur, Odontotermes	35	Pediobius 81
	186	Pediobius parvulus Ferrière 91, 101, 104, 106
Odontotermes obesus Rambur	35	Penicillium 154, 155
Odontotermes redemanni Wasmann	35	pennipes Fabrieius, Trichopoda 38
Oecophylla 40, 41, 138,		Pentatomidae 43
Oecophylla longinoda Latreille 40,	138	Pentatomophaga bicincta de Meijere 38
Oecopbylla smaragdina Fabricius 38, 45,		peraffinis Silvestri, Microcerotermes 34
Oligosita utilis Kowalski 101	105	perforans Wollaston, Xylehorus 124
Omiodes	74	Perisierola nephantidis Muesebeek 69
omma van der Hoeven, Cyclodes	94	peroptusa Meyrick, Batrachedra 64
Ovencyrtus 37,	109	Pestaloptiopsis palmarum 82, 104, 105
	7 - 38	Phasmida 27
Ovencyrtus podontiae Gahan	113	Phasmidae 27
	185	Pheidole 38, 41
Ophicrania leveri Günther 22,	183	Pheidole megacephala Fabricius 38, 75
Oregma	45	Pheidole punctulata Mayr 41
orientalis Westwood, Dorylus	141	phelaus Westwood, Megacrania 31
Orthocraspeda catenatus Snellen	83	phidippus Linnaeus, Amathusia 54
Orthoptera	14	Phlaedromius 69
Orycles 98, 114, 115, 127,	134	phoenicis Fabricius, Rhynchophorus 117, 187
Oryctes owariensis P, de B.	183	Phostria 74
Orycles boas Fabricius	134	Phycita dentilinella 81
Orycles centaurus Sternberg	134	Phycitidae 76, 152
Oryctes erebus Burm.	183	Physomerus grossipes Fabricius 41
Oryctes gigas Laporte de Castelneau	134	Phytoptipalpidae 142
Oryctes gnu Mohner	134	piliceps Snyder, Microcerotermes 32
Orycles monoceros Olivier	134	Pimelephila ghesquierei Tams 75
Oryctes owariensis P. de B.	183	pinguis Fabrieius, Winthemia 56
Oryctes rhinoceros Linnaeus 35, 125, 134	184	Pinnaspis buxi Bouché 50, 141
Oryctes sjöstedti. Kolbe	134	Piopbila casei Linnaeus 153
Oryctes trituberculatus Lansb.	134	Plassius jaranus Erichson 124
oryctophaga Coquillet, Scolia	128	Platymeris 128
Oryzaephilus	154	Plesispa reichei Chapuis 112
Oryzaephilus mercator Fauvel	154	Pleurotropis 101
Oryzaephilus surinamensis Linnaeus	154	Ploceidae 146
owariensis P. de B., Oryctes	183	politus Mulsant, Chilocorus 48
oxymorus Tosquinet, Goryphus	81	polynesiensis Marks, Aedes (Stegoneyia) 150
Oxya chinensis Thunberg	23	Promecotheca 98, 142

INDEX

179

	400	DI - I	
promecothecae Ferrière, Achrysochari.	106 102	Rhopalocera	117
Promecotheca antiqua Weise		rhynchophorae Blanchard, Parabillaca	iii
Promecotheca coeruleipennis Blanchas		rhynchophori Ewing, Tetrapolypus Rhynchophorus 114, 117.	13
28, 101,	104, 105 101, 106		
Promecotheca cumingi Baly	104	Rhynchophorus ferrugineus Olivict 113, 116.	ım
promecothecae Ferrière, Eurytoma	104, 185	Rhynchophorus palmarum Linnaeus	10
Promecotheca opacicollis Gestro		116, 117, 120,	18
Promecotheca papuana Csiki 102, Pseudoniscara	105, 185 26	Rhynchopherus papuanus Kirsch 116, Rhynchopherus phoenicis Fabricius 117,	18
Pseudotheraptus 40, 41,	138, 140		18
Pseudotheraptus devastans (Dist.)	185	Rhynchophorus schach Olivier 115, 116, Rhynchota	30
Pseudotheraptus wayi Brown	38, 185	rigidus Reyne, A. destructor	4
psittasina Serville, Atrastomorpha	23	Rosas	82
Psittaeus erithaeus	146	romandi Saussutc, Elis	128
Psychidae	77	retunda Hampson, Contheyla	8
Pteropidae	150	rubrifemur Mickel, Myrmecomimesis	25
Pteropus	150	ruficornis Fabricius, Scolia	128
Pteropus edulis	151	rufipes de Gcct, Necrobia 153,	154
Pteropus edwardsii	151	rufisena Walker, Tirathaha 72, 73.	187
Pteropus scapulatus	151	Rutelidae	136
Pteropus tonganus	151	10,000	_
Pteropus vampyrus	151	Sarcophaga antilope Bottcher	81
ptychis Dyar, Hyalospila	76	Sarcophaga fuscicanda Bottcher	115
punctata Fabricius, Xanthopimpla	69	Satyridae	51
punctulata Mayr, Pheidole	41	Scapanes	135
	7, 62,73	scapulatus, Pteropus	151
Pyemotes ventricosus Newport 69,	101, 142	schach Olivier, Rhynchophorus 115,	
Pvemotidae	115, 142	Schedorhinstermes marjoriae Stryder	32
Pyraustidae	74	Schistocerca gregaria Forskål	12
pyrogramma Meyrick, Agonoxena	70	Scholastes	141
Pyrophorus	128	Scholastidae	141
		Sciuridae	150
quadrifoveatus Palisot de Beauvois,		Scolia erratica Smith	115
Strategus	135	Scolia oryctophaga Coquillet	128
quadrimaculata Baranov, Exerista	78	Scolia ruficornis Fabricius	128
		Scoliidae	128
Raoiella indica Hirst	142	Scolytidae	124
rainbowi Hill, Neotermes	35	Scymnus	143
rattus alexandrinus, Rattus	147, 185	Segestes decorativs Redtenhacher	26
Rattus exulans exulans	150, 185	Segestes unicolor Redtenbacher	26
Rattus exulans micronesiensis	150, 185	Segestidea	26
rattus frugivorus, Rattus	147, 185	selebensis	108
Rattus rattus alexandrinus	147, 185	septemfasciata Serville, Nomadacris 21,	140
Rattus rattus frugisorus	147, 185		183 69
Rattus rattus rattus	147, 185	Serratia marcescens	02
redemanni Wasmann, Odontotermes	35 112	Setora nitens Walker 78, 80.	. 24
reichei Chapuis, Plesispa	112	Sexura coriacea Linnacus	26
remota Aldrich, Bessa (Ptychomyia)	93, 160	Sexara karnyi Lecfmans	24
renalis Waterston, Antrocephalus	72	Sexana mone-guineae Brancsik	24
Rhahdocnemis	117	Sexara nubila Stàl	24
Rhabdoscelus obseserus Boisduval	117, 186	seychellensis Ferrari, Graeffea	29
Rhadinaphelenchus	116, 117	shanghaiensis Smith, Chrysis	81
Rhadinaphelenchus cocophilus Cobb	143, 186	simmondsi	108
Rhina barbirostris Fabricius	120	sjöstedti Kolbe, Oryctes	134
rbinoteras Linnacus, Oryctes 35, 125,		smaragdina Fabricius, Oecophylla 38, 45,	138
Rhinostomus afzelii Fähraeus	121	sopherae Linnacus, Brassolis 56, 57,	182
	120, 186	Sparganobasis subcruciatus Marshall	121
Rhinotermitidae	31	Spathius apicalis Westwood	124

Spokember Vilicincux, Ceraminia phempher Evicing phempher Vilicincux, Ceraminia phempher Evicing phempher Vincex, Ceraminia Cerami		
Spoteshis morbej Ashmead Spoteshis morbej Ashmead Spoteshis morbej Ashmead Spoteshis Trumberg, Catamape the Spoteshis Catamape the Stranguage manufericus, Sillimon the Stratugua manufericus Barmeite Stratug		
Spiechtis worder Ashmead Spiechtis Spie		
Spinetia Tumberg, Catestopp plandeur Catest		
pinedine Vendelis, Cattenerum per pinedine Wasticus, Stillium pinedine Patricius Stratigus quantivate Burmister Stratigus quantiv		
r findame Kovalski, Columeraru 194, 105 prindame Parkicus, Sillium 154 prindame Macroso, Vinamia 157 prindame rationame Herbas 158 prindame prindame Newswood, Vinamia 158 prindame prindame Parkicus 158 prindame prindame Parkicus 158 prindame Julianame Barmeitser 158 prindame Julianame Julianame Juli		
phendikeer Fabricius, Sillium tiellijem Westwood, Viannia 13 Trielogamphu 13 Trielogamphu 13 Trielogamphu 13 Trielogamphu 13 13 Trielogamphu 13 13 Trielogamphu 13 13 Trielogamphu 13 13 Trielogamma (Agricie, Traithain anna Zehniner 14 13 14 Trielogamma (Agricie, Traithain anna Zehniner 14 15 15 Trielogamphu 14 15 Trielogamphu 24 15		
Saturi greiner Fabricius Saturius delter Trainerius Homes verbenger Victorius Disant Street Victorius delinerius Homes Victorius Street Victorius delinerius Homes Victorius Street Victorius Str	splendens Kowalski, Closterocerus 104, 105	
Stephenitis typious Distant Strikturea deliatureasa Hoffee Silhum pholidum Fabricius Strikturea deliatureasa Hoffee Silhum pholidum Fabricius Strategu audurinea Brancius		
Scienterma dellaterrease Hoffer Simenterma dellaterrease Hoffer Simenterma dellaterrease Hoffer Simenterma radalitaterillem Grittult Siratury amatherita Burmiciste Siratury genediferatur Paliste de Stratury quadefineatur Paliste de Stratury genediferatur Palister de Stratury genediferatur Pal		
Stillium ophendidum Febricius Manutaerus audistruitulium Gista Manutaerus audistruitulium Gista Manutaerus audistruitulium Gista Manutaerus audistruitulium Gista Strategu audistruitulium Burmeitus Strategu opingeriba Febricius Strategu opingeriba Burmeitus Strategu opingeriba	Stephanitis typicus Distant 44, 187	
Standarien andationalium Girault Somatonyin bezgana Baranov Stratguy along Linnacus Stratguy along Linnacus Stratguy along barno Stratguy along barno Stratguy andatificatus Palises de Beavois Beavois Beavois Stratguy andatificatus Palises de Beavois Stratguy andatificatus Palises de Stratguy andatificatus Carry Stratguy and Stratguy Stra		
Sometimpsip bergaines Bearmov 150 Trichnofold pomispre Febricius 151 Trichnofold pomispre Ferricke 152 Trichnofold pomispre Ferricke 153 Trichnofold pomispre Ferricke 153 Trigone Spalinands Iromovi Ckll, 154 Trigone Spalinands Iromovi Ckll, 155 Trigone		
Stratigge anhors Linnacus 15, 187 Trikhopfulu pulpriva Ferrice 69, 71 Strategy and water Burmeitter 15, 17 signes pulputude the more Cild. 12, 18 Strategy pulpriva the Patrice Cild. 12, 18 Strategy and the Strategy pulprish the St		
Stratigus ausabortas Burmeistes Stratigus quandiplinatur Paliste de Stratigus Pariste de Stratigus Pariste de Stratigus Vierce, Apanetar S		Trichopoda pennipes l'abricius 38
Strategy with private I Palice de Beauvois Straing Williams I Palice I P		
Stratigue quantifuratur Palisoc de Beauvois Stratigue quantifuratur Palisoc de Sommis Stratigue quantifuratur Palisoc de Sommis Stratigue quantifuratur Palisoc Strati		
Beauvois Starmin Selection Starmin Febricius Santui grimin Febricius Syntamo lipram Santui Grimin Selection Santui		
Starmin Fabricius 54 Tragarada dibetar Tama 84 Santau graniur Fabricius 152 Tragarada dibetar Tama 84 Santau graniur Fabricius 152 Tragadarii rititatu Linaucus 152 Tragadarii rititatu Linaucus 152 Tragadarii rititatu Linaucus 152 Santau filiaucus 152 Santau fil		
Sautu graniu Fabricius Gil Tropidarii riritata Linaucus 12 Iropidarii riritata Linaucus 13 Iropidarii riritata Linaucus 13 Iropidarii riritata Linaucus 13 Iropidarii riritata Linaucus 13 Iropidarii riritata Linaucus 14 Iropidarii riritata Linaucus 15		
nderwinism Marshall, Spergondusti meirita Lineaux, Petanga 15, 23, 23, 24, 24, 24, 25, 24, 24, 24, 25, 24, 24, 24, 24, 24, 24, 25, 24, 24, 24, 24, 24, 24, 24, 24, 24, 24		
meinte Linnacus, Petange 15, 23, 184 Trépidenti Intrittil Verty 20 malatimenthe Givaint, 8, montront 15, 187, 187, 187, 187, 187, 187, 187, 187		
mkatiturithem Girsult, Ismanterear minimenti Linuxu, Orzaphilus Systiem plyram 154 Tjehndemur anduitre Reches 154 Tjehndemur anduitre Reches 155 Tjehndemur Anduitre 155 Tjehndemur Menore		Tropidacris diex Drury 1/
nrimmensi Linnacus, Oryzaphilus Systemolybrum Systemolybru		Tropidacris latrettei Perty 20
Systems phyrome September September Systems phyrome September Sept		
tattenii Gortin, Diocalondra Typinov Distant, Stephaniii + 44, 182 Taginom andensciphalme Fisheirus angune Viercek, Japanteri Canaguar Viercek, Gantine III. Lichamani Tralabaka Wilkinson Telenamai Warrecke Viercek, Canaguar III. Lichamani Tralabaka (Dapin Leadyna Viercek) Lichamani Morce Canaguar Viercek, Perabaka III. Lichamani Marce Canaguar Viercek, Perabaka III. Lichamani Morce Canaguar Viercek, Perabaka III. Lichamani Morce Canaguar Viercek, Perabaka III. Lichamani Morce Canaguar Viercek, Perabakaka III. Lichamani Morce Canaguar Viercek, Perabakaka III. Lichamani Morce Canaguar Viercek, Perabakaka Westewood Lichamani Morce Canaguar Viercek, Perabakakan III. Lichamani Morce Canaguar Viercek, Perabakan III. Lichamani Morce Canaguar Viercek, Perabak		
sateenii Goetin, Diealandeni 12,1 (22 amilioire Redeembasher, Segrits 20 amilioire Ibhl), Comprehi 22 amilioire Ibhl), Comprehi 23 amilioire Ibhl), Comprehi 24 and State 12 amilioire Ibhl), Comprehi 24 and State 12 amilioire Ibhl), Comprehi 24 and State 12 amilioire Ibhl), Comprehi 25 and State 12 amilioire Ibhl), Comprehi 24 and State 12 amilioire Ibhl), Comprehi 25 amilioire Ibhl), Comprehi 25 amilioire Ibhl), Comprehi 25 amilioire Ibhl), Comprehi 25 amilioire Ibhl), Comprehi 26 amilioire Ibhl), Comprehi 27 amilioire Ibhl), Compre	Syntomosphyrum 84	
Tapiones melance/plalow Fishricius terapus Vierces, /pantiri 51, 102 aufjuritus labii, /campreilla 48 terapus Vierces, /pantiri 51, 103 aufjuritus labirus literatus labirus literatus labirus literatus labirus Moore Telionis palamen Moore Telionis palamen Moore Ordinis o		typicus Distant, Mephanilis 44, 18/
toraguse Viercek, Apantoles for a vierce process of the property with the company of the compan	Taitensis Guerin, Diocalanara 121, 122	
Technonymers dalipér Smith Technonymers deripent Wilker Telhomer de l'action d		
Technomyrows deringous Wilker Telemania Tridemania Harriciter Telemania Tridemania Mosce Telisias palaman Mosce Telisias palaman Mosce Telisias midde Chapin Temopialpidae (Dynpifipalpida) Tetropidae		idili Kowaiski, Oligotia 101, 105
Telemenus Tralesholas Wilkinson Telisata kambusa Moore 20 varmasu nidasu 151 Telemenus Iratalbaka Wilkinson Telisata kambusa Moore 20 varmasu nidasu 152 Varmasu nidasu 152 Varmasu nidasu 152 Varmasu Newport, Pendisalah 152 Telisata kambusa Newport, Pendisalah 152 Termitaka Phytopisfopidab 152 Termitaka Phytopisfopidab 152 Termitaka Phytopisfopidab 152 Termitaka Phytopisfopidab 152 Termitaka Phytopisfopida 152 Termitaka Iratalba Ira		17 f and the state of the state
Telement strathabar Wilkinson Tellista koobusen Moore Tellista kapimam Noore Tellista Kapim		
Telista kumbuan Moore 60 vertrieaus Newport, Pedendudat 152 Telista palmarum Moore 60 vertrieaus Newport, Pedendudat 61 Telista palmarum Moore 61 vertrieaus Newport, Pedendudat 62 to 11 Telista initiad Chapin 50 Virunnia italiifare Westwood 51 Tempeldudat (Pappripsipsida) 23 25 Winnia italiifare Westwood 51 Tempeldudat (Pappripsipsida) 23 25 Winnia italiifare Westwood 51 Tempeldudat (Pappripsipsida) 24 20 22 Winnia italiifare Westwood 51 Tetrapsidate Telistation 52 23 24 25 Winnia iniquis Fabricius 60 Tetrapsida principsi Ferriker 61 Tetrapsia 6		
Telisia palmenum Moore 60 retrisians Newport, Pyenestri 69, 101, 142 Telisian eitide (Chaprin 1		
Telinian inida Chapin Tembrinidae Tembrini		
Tendrindiak Tendiada (Phynopipalpida) Tendiada (Phynopipalpida) Tendiada (Phynopipalpida) Tendiada (Phynopipalpida) Tendiada (Phynopipalpida) Tendiada (Phynopipalpida) Tetratida (Phynopipala (Phynopipala) Tetratida (Phynopipala (Phynopipala) Tetratida (Phynopipala (Phynopipala) Tetratida (Phyn		
Temajahdar (Phynophiphiphida) 1.12 wayi Berwan, Pranduheraptan 28, 185 Termitidas University Signores, Eusalymantus 1.53 Willindersia pingui Exbricius 5.51 Willindersia pingui Exbricius 5.52 Willindersia pingui Exbricius 5.53 Willindersia pingui Exbricius 5.54 Willindersia pingui Exbricius 5.55 Willindersia pingui Exbricius 5.55 Willindersia pingui Exbricius 5.57 Willindersia pingui Wilcelmann 3.57 Willindersia Pingui		v msonia stetiljera Westwood 21
Termitake Termit		mani Bernan Demokshireston 30 105
trasilatus Signocet, Euralymanta 1 Tetraspidale 1 Tetraspidale 1 Tetraspidale 1 Tetraspidale 1 Tetraspidale 1 Tetraspidale 1 Tetraspida pia ribundopheri Evinig 1 Tetraspida pia ribundopheri Evinig 1 Tetraspida pia ribundopheri Evinig 1 Tetraspida 1 Tet	Temaparpuae (Phytopreparpuae)	
Tetrasylvida Tetr	1 trminate 52, 33	
Tetragyaba fjiratii Hirst 143 Xanlopinpla 150		w minemia pagini ratificias
Tetrophopor Temohopor Existing Tetrophopor Temohopor Texting Tetrophopor Temohopor Texting Tetrophopor Temohopor Texting Tetrophopor Temohopor Texting Tetrophopor Texting Tetrophopor Texting		Vanthanimeta 90
Terratikus irmulipar Ferriker 109, 112 Xmilogoni midanyipa Wiederman 52 Tettiganida 12 Xmilogoni midanyipa Wiederman 52 Tettiganida 13 Xmilogoni midanyipa Wilaman 135 Milogoni midanyipa 135 Milogoni midanyipa Wilaman 135 Milogoni		
Tettgenidae 23 Xyldorup perforael Wollaston 124 Theraptus 33 Xyldorup perforael Wollaston 125 Theraptus 43 Xyldorup perforael Wollaston 125 Theraptus 40 Syndimients Hill, Enternet 35 Tingidae 7 Syndimients Hill, Enternet 35 Tirathabe completes Hill, 22 Zaleptogyas floro-orbitalis Cameron 25 Tirathabe completes Buller 1, 22, 23 Zaphoptay dynaphile Tritinen 24 Tirathabe completes Buller 2, 12, 23 Zaphoptay dynaphile Tritinen 24		
Therapia 38 Xylatrapei 135 these Lineaues, Erianda 61 yandinlenii Hill, Euternet 137 Tragida 15 Tragida 15 12 Zulpappyi flavorbilatii Cameron 25 Trakhala emplore Bulert 12 22 Zulpappyi flavorbilatii Cameron 25 Trakhala emplore Bulert 12 22 21 Zulpappyi flavorbilatii Cameron 26 12 22 21 Zulpappyi flavorbilatii Cameron 26 12 22 21 Zulpappyi flavorbilatii Cameron 26 12 22 21 Zulpappyi flavorbilatii Cameron 27 22 23 24 Zulpappyi flavorbilatii Cameron 27 22 23 24 Zulpappyi flavorbilatii Cameron 28 24 25 24 Zulpappyi flavorbilatii Cameron 28 24 25 24 Zulpappyi flavorbilatii Cameron 28 24 25 24 Zulpappyi flavorbilatii Cameron 28 25 25 25 25 25 25 25 25 25 25 25 25 25		
thrax Linnacus, Erimons 61 thyrii Fabricius, Gaugera 99 Tingidae Tingidae Tingidae Tirathaka Wilkinson, Apantiets 72, 23 zambihara Bohart, Helitaphagus 41 Tirathaka completos Bulter 72, 23, 23 zambihara Bohart, Helitaphagus 41 Tirathaka completos Bulter 72, 23, 23 zambihara Bohart, Helitaphagus 41		
htyrii Fabricius, Gangara (9 yandiniensii Hill, Eutermes 35 Tingidae Tiraidaba Wilkinson, Apanteles 72, 23 zahiphaya Bohart, Halitophayau 41 Tiraidaba complexo Butler 12, 23 zahiphaya Bohart, Halitophayau 41 Tiraidaba complexo Butler 12, 23 zahiphaya Bohart, Halitophayau 41 Tiraidaba complexo Butler 12, 23 zahiphaya Bohart, Halitophayau 41		. Cyton reput
Tiralbale		vandinioneie Hill Futermee 35
Tiratbaba Wilkinson, Apanteles 72, 73 Zalephopysus flavo-orbitalis Cameron 75 Tiratbaba complexa Butler 71, 72, 73 zanbibarae Bohart, Halistophogus 41 Tiratbaba complexa Butler 71, 72, 73 Zophopyete dynrephila Trimen 61		yannininin IIII, Lintonio
Tirathaba complexa Butler 71, 72, 73 Zophopetes dysmephila Trimen 61	Tinetheles 61 71 72	Zalantonova Ramarhitalia Camerun 75
Tirathaba complexa Butler 71, 72, 73 Zophopetes dysmephila Trimen 61	tirathabas Wilkinson Anastales 72 73	
	Tirothelia compleya Burler 71 72 73	
2 / Summer Guerran Liniquetti 22 / Summer 200 22		
	The state of the s	->0

ADDENDUM 1

Agonoxena argaula Meyrick (p. 69)

In Fiji, damage by larvae may cause a 5 percent tissue loss in young fronds, equal to 10 percent in mature fronds. The total number of larvae present during the life of the frond varies from 400 to 1000. Damage is serious in palms up to two years of age, less at seven, and negligible in mature palms (O'Connor, 1967).

Amblypelta cocophaga China (p. 36)

The final progress report by Greenslade (1968) confirms previous work in the Solomon Islands, that chemical control, either aerial or from the ground, seems unlikely to be effective. The problems of locating the most serious untilal rares, and the high cost of moving spraying equipment through the plantations, are serious. Unless systemic insecticides prove effective, the most promising results seem to be by "brushing" ground vegetation, the use of combined herbicides and insecticides, aided by new planting of palms under old stands. The last is contingent on the eradication and Bimitation, respectively, of the harmful ants Philadis megaciphale (F.) and Iridingresses cardiars F. Smith and their replacement by the predacous ant Oxephylia sumargilina (F.). Similal conclusions have been made in New Guinea by Smec (1965), who also favours encouragement of the last-named beneficial ant (see text, p. 38).

Aspidiotus destructor Signoret (p. 46)

Dinther (1956) in Surinam states that heavy rains can check development of the scale, which is controlled with oil emulsion from a high-power pressure sprayer directed especially on the lower surface of the fronds. Arkotine oil with either 0.2 percent DDT or chlordane were used.

Attention is drawn to a paper by Fennah (1954), showing that large populations of this scale in the West Indies are usually associated with faulty agronomy and unfavourable weather. The combination of "these two factors,

¹ The present addendum, submitted by the author, brings the data on Pests of the coconut palm up to 1968.

when severe, may make chemical control an uneconomic proposition. Basic research is needed to study the association between mass attacks of the scale and the cropping and growth of the palms. For distribution, reference is made to Map No. 218 of the Commonwealth Institute of Entomology, 1960.

Axiagastus cambelli Distant (p. 43)

In New Guinea, Smee (1965) reports that crop losses follow a drain of sap resulting from heavy attacks. See the peculiar, elongate shape of young nuts damaged by feeding in the New Hebrides (Fig. 37, p. 44).

Azteca cartifex Forel (p. 140)

Besides its injury by propagating the coconut scale, Dinther (1956) states that, in Surinam, this ant may be persistent enough in the crowns to prevent the nuts from being harvested.

Brassolis sophorae Linnaeus (p. 56)

The larval stage lasts from 73 to 90 days.

Brontispa longissima Gestro (p. 107)

Life history studies in New Guinea reveal that the preoviposition period lasts 74 days, thus giving a total cycle of 4 months (Smee, 1955). Chemical control with 0.1 percent dieldrin should aim at application of 28 millilitres of fluid to palms up to three years of age. Similar results in the New Hebrides (Manciot, 1965) with 0.15 percent dieldrin were obtained using 2 litres per hectare, enough for 143 palms. For distribution, see Map No. 227 of the Commonwealth Institute of Entomology, 1966. In Tahiti and Moorea, the egg parasite Tatraitibodes broutipus Ferr. has not yet been recovered in the field. The failure of the introduction is attributed to attacks by the ant Philidde megarephala (F.), coupled with dry weather at the period of release (Cochereau, 1965).

Castnia daedalus Cramer (p. 84)

Additional palms attacked are date, oil and sabal palms, Pritchardia, Livitiona and Maximiliana. The insect also occurs in Panama (Dinther, 1956). The pink, spindle-shaped eggs measure 4.3 by 4.7 millimetres. The larval stage occupies one year. The best, but not very effective, control is 0.5 percent DDT spray. Castnia licus Drury (p. 85)

Dinther (1956) reports that the life cycle takes four months.

Diocalandra stigmaticollis Gyllenhall (see footnote, p. 122)

Synonym of D. frumenti (p. 122).

Graeffea crouani Le Guillou (p. 27)

Detailed studies of this phasmid have been made by Paine (1968) who forwarded 960 puparia of a tachinid from the Solomons to Fiji. The fly, Myctromyitla lastifica (Meanil), is a parasite of the larger phasmid Ophiramia lurvi Günther. However, there has been no recovery of the parasite to date.

Leucopholis concophora Burmeister (p. 136)

Later tests by Mathen, Kurian and Mathew (1964) show that BHC, dieldrin and aldrin, respectively at 90, 30 and 288 grammes per hectare, are preferable to malathion (at the first rate) or heptachlor.

Nephantis serinopa Meyrick (p. 66)

Large populations of the mite Pyemotes sentriosus have been found on larvae in south India by Mathen, Sathiamma and Kurian (1968). There is small hope of economic control, so that mass breeding for field release is not planned.

For distribution of N. serinopa, see Map No. 211 of the Commonwealth Institute of Entomology, 1966.

Ophicrania leveri Günther (p. 29)

A paper by Paine (1968) deals in detail with bionomics, and shows that the life cycle occupies five months, two of which are spent in the egg stage. Nymphs feeding on coconut produce larger adults than those on betel palms. A number of jungle plants are also eaten.

African species of Oryctes (p. 134).

Oryctes owariensis P. de B. and Oryctes erebus Burm. are two additional west African species. The latter is a serious pest in Sierra Leone.

The bionomics of these pests have been studied by Mariau (1966, 1968a), especially on the Ivory Coast. As usual with these dynastids, damage is most serious to palms up to four years of age, which are frequently.

killed. Besides routine clean cultivation, it is important to establish as large blocks of palma as possible, so as to have the smallest perimeter against which protective measures are facilitated. Dealing in particular with O. monocrus and O. boat, Mariau shows that the former attack higher up in the palms, of which 75 percent may be invaded. The adults tunnel into the central whorl of unexpanded leaves toward the terminal bud, which in two-year-old palms can be reached by tunnels 45 centimetres long. Nursery seedlings are often killed but nut-bearing palms are little affected.

Oryctes rhinoceros Linnaeus (p. 125)

Control in New Guinea was obtained with 0.01 percent auc spray or 5 percent DDT dust. With the latter, a mixture of equal parts of sand or sawdust is placed in the leaf axils (Smec, 1965). Similar treatment was recommended in India by Mathen and Kurian (1966) for this dynastid as well as for the red palm weevil.

Biological control with the reduvidi Platymeris rhadamanthus Gerst. is being maintained in Fiji (Swaine, 1966), although recovery of this Zanzibar bug has not occurred. Research on control by viruses is reported by Huger (1966a and 1966b), who finds that in Western Malaysia, Sabah, Fiji and Samoa Rhabhlasurisan opties may account for the death of 60 to 88 percent of the larvae, compared with only 1 to 3 percent due to the fungus Metarrhizim anisoplias. The virus may exist in the form of spherical particles 160 mg in diameter, or as rods 195 mg in length. These are in the body fat and prove very toxic when injected into the body exity of larvae. Artificial culture and release in the field may become standard control; best results are obtained with third-instar larvae. Similar work in the Philippiners showed that two conditions exist known as histolytic disease, in which the larvae develops a translucent body, and blue disease in which it changes from steel blue to black, followed by a faccid and then dry state (Fandialan and Ibafe, 1966).

Further details of the spread of the rhinoceros beetle are given in Map No. 54 (revised), published by the Commonwealth Institute of Entomology in 1967. Details of its spread in Fiji are given by Swaine (1966).

Patanga succineta Linnaeus (p. 23)

Severe damage by the Bombay locust was reported in the Laccadive Islands by Menon (1967), who stated that 25 percent of the fronds were injured. Young leaves in the crown as well as ground vegetation were attacked. Control was effected by spraying with 50 percent dieldrin (1 kilorgamme per 50 litres of water) or 20 percent endrin (3 millilitres per litre), or folidol (1.5 millilitres per litre); a power sprayer was used. Against hoppers, dusting with 10 percent BHC at 25 kilogrammes per hectare was effective.

Promecotheca obacicollis Gestro (p. 104)

An outbreak in 1962 in the isolated Tikopia Island of the Santa Cruz Group is assumed to have arisen through transport of adults by air currents from a distance of 240 kilometres (Greenslade, 1965). Unlike the 1933 occurrence, which was controlled by an egg parasite (p. 105), the later outbreak was checked by the transport (from the main Solomon Islands lying to the west) of the predaceous tree ant Oesophylla *maragdina (F.). It is noted, however, that there had been severe competition by the beetle for oviposition sites and a shortage of larval food.

Promecotheca papuana Csiki (p. 102)

The "one-stage" condition has arisen in New Guinea from an incorrect application of insecticides. This development is, therefore, quite apart from the usual external factors such as weather of failure of parasites. That injudicious chemical "control" can bring about an increase in a hispid pest is most interesting, as the same result was reported in the zygaenid moth Brandstranea acknowledge in the displaying seep.

Pseudotheraptus devastans (Dist.) (to be added to section on Rhynchota)

This species, previously known from rubber, cocoa and cassava, occurs throughout tropical Africa and has recently been reported from coconut. It was formerly placed in the genus *Pendulinus*.

Pseudotberaptus wayi Brown (p. 38)

The somewhat complex relationship between insect feeding and crop yield has been studied in Tanzania by McKinlay (1965), who draws attention to the compensating mechanism by which the palm sheds excess nuts when a figure is reached beyond which they can be carried to maturity. The critical age for natural shedding of nuts is just under three months, when they are about 7 centimetres in length. Plots sprayed with a mixture of 0.1 percent pury and 0.1 percent malathion showed an increased crop for the first two years but not for the third.

Rats (р. 147)

A practical method of control is described by Smith (1967) in Jamaica, in which 8 parts of melted paraffin wax are mixed with 11 parts of maize meal, 2 parts of sugar and 1.25 parts of 0.5 percent warfarin. After setting, the blocks are cut into pieces of 225-gramme weight, one placed between

every five or six palms. When this method was used on the Ivory Coast, the damaged nuts fell from 18.4 to 8.2 percent in one month, 1.9 percent in three months, to nil after six months (Mariau, 1967).

Rhabdoscelus obscurus Boisduval (p. 117)

The range should include Celebes, Java, Sumatra, Borneo and many Polynesian and Micronesian records. The larger R. asperipennis Frm. is a coconut pest in the Caroline Islands.

Rhadinaphelenchus cocophilus Cobb (p. 143)

Experiments in Trinidad with sevin (carbaryl) show that this chemical did not actually kill the eelworms but only discoordinated their movements. The Lt 50 was about 20 parts per million. However, both solutions and dusts prevented the petiolar tissue of the palm from being penetrated by the mouth parts (Fenvick, 1968).

Rhinostomus barbirostris Fabricius (p. 120)

Contrary to some earlier workers, Dinther (1956) found that, in Surinam, healthy palms were not attacked.

Rhynchophorus ferrugineus Olivier (p. 113)

The publication of a monograph on this genus by Wattanapongsiri (1966) has resolved the synonymy, a task long overdue. However, his distributional record for Victoria (Australia), when followed up by the present writer showed, from records in the British Museum (Natural History), a solitary record dating back to 1862: the temperature in winter would rule out the survival of this tropical weevil.

In south India, Mathen and Kurian (1966) have obtained prophylactic control by placing 225 grammes of either 5 percent chlordane or BHC powder mixed with an equal volume of sand in the leaf axils. A reduction in numbers from 80 to 90 percent was secured.

Rhynchophorus palmarum Linnaeus (Montr.) (p. 116)

Good results have been obtained in Trinidad by the use of chemical lures of 1 percent skatole and 1 percent teramyl acetate (Hagley, 1965). Another lure was 100 millilitres of 6.25 percent skatole, 100 millilitres of 0.5 percent iso-amyl acetate and 300 millilitres of 5.5 percent liquid malt extract. This concoction, mixed with about 1 kilogramme of maize meal into a paste, attracted three times as many weevils as cocount sap. Also

in Trinidad, Maharaj (1965) described an efficient trap made from polished aluminium with side apertures of 18-gauge galvanized wire set, 2.5 centimetres apart, which prevented the escape of the weevils that entered the trap. The latter contained a bait of coconut tissue.

Rhynchophorus papuanus Kirsch (p. 116) is now R. pilineatus,

Rhynchophorus phoenicis Fabricius (p. 117)

Recent studies by Mariau (1968b) stress the difficulty of dealing with attacks in the crown, as the damage is done internally before it is noticed; in the trunk, however, frass is apparent. Application of 0.08 percent dimethoate from 50 to 200 millilitres is recommended, or brushing trunk wounds with wood tar. It is important to avoid or reduce injury to trunks as this attracts ovijooiting weevils (from 100 to 200 eggs are laid per adult). Attacks on the terminal bud result in the death of palms a few weeks later.

Rynchophorus schach Olivier (p. 115)

According to Wattanapongsiri (1966), this species should be R. vulneratus (Pantz.).

Stephanitis typicus Distant (banana lacewing bug) (p. 44)

Results in south India (Mathen, Mathew and Kurian [1968]) indicate that adult and nymph numbers are positively correlated with temperature and sunshine and negatively with relative humidity and rainfall. There is a maximum peak in April, with minima in June and December.

Strategus aloeus Linnaeus (p. 135)

This species measures from 40 to 58 millimetres. It also extends into Surinam, where its larval period lasts one year. Dinther (1956) suggests control by applying a ring of chlordane or aldrin at a distance of 0.5 metre from the base of the trunk. Vayssière (1965) in Colombia suggests 2.5 percent heptachlor at 300 grammes per palm in the first year, increased to 300-600 grammes subsequently.

5. jugartha Burm. is the Brazilian species against which the soil at the base of palms is drenched with 20 percent endrin at 2 litres per palm at a 1 percent concentration.

Tirathaba rufivena Walker (p. 73)

It is considered uneconomic to use chemical control in New Guinea, as any reduction of the larvae would result in only a small crop increase

(Smee, 1965). This is due to the fact that many nuts attacked by larvae would be shed normally, even in the absence of the insect.

Trigona hyalineata branneri Ckll. (to be added to section on Hymenoptera)

This is a stingless social bee, black in colour. It is 6 millimetres in length. The deep holes bored in the soft tissues of the inflorescence may cause enough deary to prevent nust from setting. Spraying with 0.2 percent DDT is not effective, as the bee nests in the nearby jungle (Dinther, 1956). This is the first known case of a nonmyrmecine hymenopterous pest of coconut.

SUPPLEMENTARY REFERENCES

COCHEREAU, P. Notes sur un essai d'acclimatation en Nouvelle-Calédonie de Tetrattichodes 1965 brontispae Fetr, C.R. Acad. Agric, France, 51(9): 661-667.

DINTHER, J.B.M. VAN. Insects of the coconut palm in Suriname. Paramaribo, Surinam 1956 Landbouw-proefstation. Bulletin No. 69.

FANDIALAN, V.O. & IBANEZ, E. A preliminary report on the study of the biological 1966 control of the rhinoceros beetle. *Philipp. J. Plant Ind.*, 51(2): 125-136.

FENNAH, R.G. The role of insecticides in the agricultural development of the British 1954 Caribbean territories. In Wallace, T. and Martin, J.T., eds. Insecticides and colonial agricultural development. London, Butterworth.

FENWICK, D.W. A note on the nematicidal properties of Sevin. Trop. Agric., Trin., 1968 25(2): 125-126.

GREENSLADE, P.J.M. Promesothesa opacicallis Gestro (Coleoptera: Chrysomelidae) on the 1965 Island of Tikopia. Pacific Invests, 7(4): 661-664.

GREENSLADE, P.J.M. Summary of work carried out, 1961-67. Entomologist's Progress 1968 Report No. 8. British Solomon Islands.

HAGLEY, E.A.C. Tests of attractants for the palm weevil. J. ccon. Ent., 58(5): 1002-1003.

HUGER, A.M. Untersuchungen über mikrobielle Begrenzungsfaktoren von Populationen 1966a des Indischen Nashornkäfers. Z. angew. Eint., 58(1): 89-95.

HUGER, A.M. A virus disease of the Indian rhinoceros beetle, Oryster rhinoceros (L). 1966b caused by a new type of insect virus. J. Invert. Path., 8(1): 38-51.
MAHARAL S. A new design of trap for collecting Rhymchophorus Jahnarum, Trop. Agric.

MAHARAJ, S. A new design of trap for collecting Rhymbophorus palmarum. 1 rop. Agric., 1965 Trin., 42(4): 373-375.
MANICOT, R. Lutte chimique contre le Brontispa du cocotier. Olégimus, 20: 369-371.

1965

MARIAU, D. Aspects du problème de l'Oryctes en Côte-d'Ivoire. Oléagimenx, 21(3): 1966

159-162

MARIAU, D. Lutte chimique contre le rat noir (Rattus rattus) dans les cocoteraies de Côte-1967 d'Ivoire, Oliagimene, 22(3): 161.

MARIAU, D. Biologie du comportement alimentaire de l'Oryctes. Oléagineux, 23(6): 377-1968a 380.

MARIAU, D. Méthode de lutte contre le rhynchophore, Rhymbophorus phoenicis F. 1968b Oléagineux, 23(7): 443-446.

MATHEN, K. & KURIAN, C. Prophylaetic control of Rhynchopborus ferrugineus. Indian 1967 J. agric. Sci., 36(6): 265-286.

MATHEN, K., KURIAN, C. & MATHEW, J. Field studies on insecticidal control of the 1964 occonut white grub Lewopholis convolptor Burm. Indian Coconut J., 17: 101-108.
MATHEN, K., MATHEW, J. & KURIAN, C. Seasonal abundance of the population of 1968 Stephonistis typicus Dist. Indian J. agric. 5xi., 38(4): 644-653.

Menon, K.S. A note on the occurrence of Patanga succinata L. serious pest on coconut 1967 trees. Madrar agric. J., 54(4): 40.

O'CONNOR, B.A. Report of Senior Entomologist for the year 1962. Department of 1967? Agriculture, Fiji. Bulletin No. 47, p. 79.

OSPINA LEON, A. La guelpa enemigo mortal de eocotero. Agric. Trop., 19(4): 12-15. 1963

PAINE, R.W. Investigations for the biological control in Fiji of the coconut stick-insect 1968 Graeffea crouani (Le Guill.). Bull. ent. Rer., 57(4): 567-604.

SMEE, L. Insect pests of Cocoa nuclifera in the Territory of Papua and New Guinea.
1965 Papua New Guinea agric. J., 17(2): 51-64.

SMITH, R.W. The control of rats in ecconuts using rat blocks. Oliaginus, 22(3): 159-1967 180.

SWAINE, G. Fiji's campaign against the eoconut rhinoceros beetle. Int. Pett Control, 1966 8(1): 6-9.

1900 8(1): 6-9. Sur quelques insectes des palmiers en Amérique du Sud. Zerentiende 1965 Internat. Symp. over Frytofarn. Frytatrie (Ghent.). Meded. Landi Hoogesch. Opzockst., Gent., 30(3): 1321-2026.

WATTANAPONGSIRI, A. Dept. Agric. Sci. Bull. Thailand, 1(1).

Note: The contents of this publication are essentially a review of the published literature. They do not constitute official FAO recommendations for pesticide use.

FAO SALES AGENTS AND BOOKSELLERS

Antilles, Netherlands Argentina Australie

Austria

Belgium

Brazil

Canade

El Salvedor

Elpland

France Germeny, F.R.

Ghana

Greece

Guyana

Honduras Hong Kong

Iceland

Indonesia

India

Iran

ireq

Ireland

Israel

Halti

Guatemala

Guinea-Bissau

Bangladesh

Société nationale d'édition et de diffusion, 3, boulevard Zirout-Youcef, Algiers. St. Augustinus Boekhandel, Abraham de Veerstraat 12, Willemstad, Curação,

Editorial Hemisferio Sur S.R.L., Libreria Agropecuaria, Pasteur 743, Buenos Aires.

Hunter Publications, 58A Gipps Street, Collingwood, Vic. 3066, The Assistant Director, Sales and Distribution, Australian Government Publishing Service, P.O. Box 84, Canberra, A.C.T. 2600, and Australian Government Publications and Inquiry Centres in Canberra, Melbourne, Sydney, Perth, Adelaide and Hobart.

Gerold & Co., Buchhandlung und Verlag, Graben 31, 1011 Vienna Agricultural Development Agencies in Bangladesh, P.O. Box 504S, Dacca 5.

Service des publications de la FAO, M.J. De Lannoy, 202, avenue du Roi, 1060 Brussels. CCP 000-0608993-13.

The Belize Bookshop, P.O. Box 147, Belize. Belize

Bolivia Los Amigos del Libro, Perú 3712, Casilla 450, Cochabamba: Mercado 1315, La Paa; René Moreno 26, Santa Crua; Junin esq. 6 de Octubre, Oruro.

Livraria Mestre Jou, Rua Guaipá S18, São Paulo 10; Rua Senador Dantas 19-S20S/206, Rio de Janeiro; PRODIL, Promoçlio e Dist. de Livros Ltda., Av. Venáncio Aires 196, Caixa Postal 400S, Porto Alegre, RS; Livraria Dom Bosco, Rua 14 de Julho 281B, Caixa Postal 962, Campo Grande, MT; A NOSSA LIVRARIA, CLS 103, Bloco C, Lojas 2/6, Brasilia, D.F.; Distribuldora de Livros Ltda., Rua de Bahia 478, Loja 10, Belo Horizonte, ME; METRO CUBICO, Livros e Revistas Técnicas Ltda., Praça São Sebastião, Rua 10 de Julho 613, Calxa Postal 199, Manaus, Amazonas; Distribuidora Luso Mercantil, Rua 13 de Maio 524. Caixa Postal 1124. Beiém, Pará: G. Lisbóa Livros Ltda., Rua Princesa Isabel 129,

Recife, PE; Livraria Cometa Distribuidora Ltda., Rua da Independencia 46, Salvador, Bahla. Renoul Publishing Co. Ltd., 2182 Catherine St. West, Montreal, Que. H3H 1M7.

Chile Tecnolibro S.A., Merced 753, entrepiso 15, Santiago. Chine China National Publications Import Corporation, P.O. Box 88, Peking. Colombia Litexsa Colombiana Ltda., Calle SS, Nº 16-44, Apartado Aéreo S1340, Boeotá.

Costa Rice Libreria, Imprenta y Litografia Lehmann S.A., Apartado 10011, San José, Cuba Empresa de Comercio Exterior de Publicaciones, O'Reiley 407, Havana. Cyprus

MAM, P.O. Box 1722, Nicosia. Denmark Elnar Munksgaard, Norregade 6, Copenhagen S.

Dominican Rep. Fundación Dominicana de Desarrollo, Casa de las Gárgolas, Mercedes 4, Santo Domingo, Ecuador Su Libreria Cla. Ltda., Garcia Moreno 1172, Apartado 2556, Quito; Calle Chimborazo 416, Gua-

Libreria Cultural Salvadoreña S.A., Calle Arce 423, Apartado Postal 2296, San Salvador,

Akateeminen Kirjakauppa, 1 Keskuskatu, Helsinkl, Editions A. Pedone, 13, rue Soufflot, 7500S Paris,

Alexander Horn Internationale Buchhandlung, Spiegelgasse 9, Postfach 3340, Wiesbaden. Fides Enterprises, P.O. Box 1628, Accra; Ghana Publishing Corporation, P.O. Box 3632, Accra. " Eleftheroudakis ", 4 Nikis Street, Athens,

Distribuciones Culturales y Técnicas « Artemis», Quinta Avenida 12-11, Zona 1, Guatemala City.

Conselho Nacional da Cultura, Avenida da Unidade Africana, C.P. 294, Bissau. Guyana National Trading Corporation Ltd., 45-47 Water Street, Georgetown.

Max Bouchereau, Librairie « A la Caravelle », B.P. 111, Port-au-Prince. The Bookstore, Apartado Postal 167-C, Tegucigalpo.

Swindon Book Co., 13-15 Lock Road, Kowloon. Snaebjörn Jónsson and Co. h.f., Hafnarstraeti 9, P.O. Box 1131, Reykjavík.

Oxford Book and Stationery Co., Scindia House, New Delhi: 17 Park Street, Calcutta. P.T. Gunung Agung, 6 Kwitang, Djakarta.

Iran Book Co. Ltd., 127 Nadershah Avenue, P.O. Box 14-1532, Tehran.

National House for Publishing, Distributing and Advertising, Rashid Street, Baghdad. The Controller, Stationery Office, Dublin.

Emanuel Brown, P.O. Box 4101, 35 Allenby Road and Nachlat Benyamin Street, Tel Aviv; 9 Shlomaion Hamalka Street, Jerusalem.

Distribution and Sales Section, Food and Agriculture Organization of the United Nations, Via delle Terme di Caracalla, 00100 Rome; Libreria Scientifica Dott, L. De Biasio " Aeiou ", Via Meravigli 16, 20123 Milan; Libreria Commissionaria Sansoni "Licosa", Via Lamarmora 45, C.P. SS2, 50121 Florence

Teacher Book Centre Ltd., 95 Church Street, Kingston.

Italy Jemaica

FAO SALES AGENTS AND BOOKSELLERS

Maruzen Company Ltd., P.O. Box 5050, Tokyo Central 100-31. Japan

Kenya Text Book Centre Ltd., P.O. Box 47540, Nairobi. Korea, Rep. of The Eul-Yoo Publishing Co. Ltd., S 2-Ka. Chong-ro. Seoul.

Kuwait Saeed & Samir Bookstore Co. Ltd., P.O. Box 5445, Kuwait.

Luxemboura Service des publications de la FAO, M.J. De Lannoy, 202, avenue du Roi, 1060 Brussels (Belgium).

Nalanda Company Limited, 30 Bourbon Street, Port Louis. Mauritiue Mexico Dilitsa S.A., Puebia 182-D, Apartado 24-448, Mexico City 7, D.F.

Librairie « Aux Belles Images », 281, avenue Mohammed V, Rabat. Morocco N.V. Martinus Nijhoff, Lange Voorhout 9. The Hague **Hatherlands**

Government Printing Office: Government Bookshops at Rutland Street, P.O. Box S344, Auckland; Alma Street, P.O. Box 857. Hamilton; Mulgrave Street, Private Bag, Wellington; 130 Ox-ford Terrace, P.O. Box 1721, Christohurch; Princes Street, P.O. Box 1104, Dunedin. New Zealand

Nicaragua Libreria Interamericana Nicaragüense S.A., Apartado 2206, Managua. Nigeria University Bookshop (Nigeria) Ltd., University of Ibadan, Ibadan.

Norway Johan Grundt Tanum Bokhandei, Karl Johansgt. GT 41-43, Osio 1. Pakistan Mirza Book Agency, 65 The Mail, Lahore 3.

Distribuidora Lewis S.A., Edificio Dorasol, Caile 25 y Avenida Baiboa, Apartado 1634, Panama 1. Panama Agencia de Librerias Nizza S.A., Paraguari 144, Asunción. Paraguay

Libreria Distribuidora Santa Rosa, Jirón Apurimac 375, Casilla 4937, Lima. Peru

Philippines The Modern Book Company, 928 Rizal Avenue, Manila.

Poland Ars Polona-Ruch, Krakowskie Przedmiescie 7, Warsaw. Livraria Bertrand, S.A.R.L., Apartado 37, Amadora; Livraria Portugal, Dias y Andrade Ltds **Portugal** Apartado 2681, Rua do Carmo 70-74, Lisbon-2; Edições ITAU, Avda. República 46A c/v-E, Lisbon-1.

Hexim, Cales Grivitei Nº 64-66, B.P. 2001, Bucarest. Saudi Arabia University Bookshop, Airport Road, P.O. Box 394, Riyadh. Librairie Africa, 58, avenue Georges Pompidou, B.P. 1240, Dakar. Senegai

Somalia " Samater's ", P.O. Box 936, Mogadishu. Mundi Prensa Libros S.A., Castelió 37, Madrid-1; Libreria Agricola, Fernando VI 2, Madrid-4. Spain

M.D. Gunasena and Co. Ltd., 217 Norris Road, Colombo 11. Sri Lanka Switzerland Librairie Pavot S.A., Lausanne et Genève: Buchhandlung und Antiquariat, Heinimann & Co.,

Kirchgasse 17, 8001 Zurich. Suriname VACO nv in Suriname, P.O. Box 1841, Domineenstraat 26/32, Paramaribo. Sweden C.E. Fritzes Kungi, Hoybokhandel, Fredsgatan 2, 103 27 Stockholm 16.

Tenvenie Dar es-Salaam Bookshop, P.O. Box 9030, Dar es-Salaam. Theiland Suksapan Panit, Mansion 9, Rajadamnern Avenue, Bangkok. Librairie du Bon Pasteur, B.P. 1164, Lomé

Togo Trinidad and Tobago The Book Shop, 111 Frederick Street, Port of Spain. Tuniala Société tunisienne de diffusion, S, avenue de Carthage, Tunis.

United States

of Amarica

Other countries

Uruquay

Venezuela Yugoslavia

Turkey Güven Bookstores, Güven Bidg., P.O. Box 14S, Müdafaa Cad. 12/S, Kizilay-Ankara; Güven Ari Bookstores, Ankara Cad. No. 45, Cağaloğlu-İstanbui; Güvan Bookstore, S.S.K. Konak Tesisleri P-18, Konak-izmir.

United Kingdom Her Majesty's Stationery Office, 49 High Holborn, London WC1V 6HB (callers only); P.O. Box 569; London SE1 9NH (trade and London area mail orders); 13a Castle Street, Edinburgh EH2 3AR; 41 The Hayes, Cardiff CF1 1JW; 80 Chichester Street, Belfast BT1 4JY;

Brazennose Street, Manchester M60 8AS; 258 Broad Street, Birmingham B1 2HE; Southey House, Wine Street, Bristoi BS1 2BQ. UNIPUB, 34S Park Avenua South, New York, N.Y. 10010; mailing address: P.O. Box 433,

Murray Hill Station, New York, N.Y. 10016. Libreria Editorial Juan Angel Peri, Alzaibar 1328, Casilla de Correos 1755, Montevideo.

Biume Distribuidora S.A., Av. Rómuio Gallegos esq. 2a. Avenida, Centro Residencial « Los Almendros », Torre 3, Mezzanina, Ofc. 6, Urbanización Montecristo, Caracas. Jugoslovenska Knjiga, Terazije 27/11, Belgrade; Cankarjeva Zalozba, P.O. Box 201-IV, Lju-

bijana; Prosveta Terazije 16, P.O. Box SSS, 11001 Belgrade. Requests from countries where sales agents have not yet been appointed may be sent to: Distribution and Sales Section, Food and Agriculture Organization of the United Nations, Via delle Terme di Caracalla, O'DO Rome, Italy.