

49. MIDDLE EOCENE TO OLIGOCENE PLANKTONIC DIATOM STRATIGRAPHY FROM DEEP SEA DRILLING SITES IN THE SOUTH ATLANTIC, EQUATORIAL PACIFIC, AND INDIAN OCEANS^{1,2}

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ABSTRACT

The relative abundance of Paleogene diatom species was determined at DSDP Sites 356 and 357 (southwest Atlantic), 64, 65, 69, 71A, 72, 73, 77B, 161, 163, 167, 289, and 292 in the equatorial Pacific, and 216, 217, and 220 in the Indian Ocean. Biostratigraphy and paleoceanographic implications of the differences in assemblage compositions are discussed.

The assemblage analysis for the middle and late Eocene at Sites 356 and 357 in the mid-latitude southwest Atlantic provides insight into the paleolatitudinal distribution of diatom species.

In the middle Eocene the diatom assemblages at these sites are composed of low-latitude and cosmopolitan species, whereas during the late Oligocene high-latitude species are present, reflecting an extension toward the equator of the circum-Antarctic assemblage that can be related to climatic cooling and probably intensified meridional wind and surface water circulation.

In addition, the Eocene-Oligocene planktonic diatom zonation developed in the low-latitude Atlantic and the Caribbean (Fenner, in press) is tested against the moderately to well-preserved early Oligocene diatom assemblages of the Pacific sites (except for Site 292, at which preservation of diatoms was too poor), and those of middle and late Eocene age at DSDP Site 167. The age assignments obtained by applying this stratigraphic zonation are in accordance with those obtained using other planktonic microfossil groups, thus proving the applicability of this zonation.

The wide occurrence of the same species through the equatorial Atlantic and Pacific oceans and the identical sequence of first appearances have both to be seen as a result of the homogenizing effect of unobstructed circum-equatorial circulation in Tethys.

In the diatomaceous Eocene-Oligocene section of Site 167 in the equatorial Pacific, the change in assemblage composition toward an assemblage dominated by *Cestodiscus* spp. in the early Oligocene occurs at about the Eocene/Oligocene boundary, as it also does at DSDP Site 366 in the equatorial Atlantic. The value of this change in assemblage composition as a paleoclimatic indicator and stratigraphic marker is suggested.

The only Indian Ocean sites (Sites 216, 217, 220) which contain Eocene-Oligocene diatoms were located just south of the equator during the Eocene. But preservation of diatoms is too poor to allow stratigraphic assignments or to discuss assemblage compositions in relation to other known sections.

INTRODUCTION

Stratigraphic zonation of the Paleogene using planktonic diatoms has been substantially developed during the last ten years. Continuing a long tradition of Russian work on Paleogene diatomaceous deposits, Jousé (1973, 1979) compiled information on the stratigraphic occurrences of Paleogene planktonic diatom species from land outcrops and deep sea cores. Much new information has also been obtained, especially from diatomaceous sections drilled by *Glomar Challenger*. Dzineridze et al. (1976, 1979) and Fenner, in Schrader and Fenner (1976), developed a stratigraphic scheme for the Paleogene of the Norwegian Sea. For the low latitudes, Fenner (1982; in press) described ranges of planktonic diatom species through the Eocene and Oligocene from DSDP Sites 13, 366, 366A, and 369A off Northwest Africa and Site 149 in the Venezuela Basin; planktonic diatom zones were also established. For the late Eocene and Oligocene of the circum-Antarctic, Fenner (in press)

defined diatom zones, revising older zonal definitions proposed by McCollum (1975), Schrader (1976), Gombos (1977), Weaver and Gombos (1981), and Gombos and Ciesielski (1983).

All the results presented here are from DSDP sites not previously analyzed for Paleogene diatoms; they provide basic new stratigraphic data and an opportunity to test zonations proposed earlier. Site locations appear in Table 1 and Figure 1.

The planktonic diatom zonation used in this study is that defined by Fenner (in press). Zones are defined and correlated to the calcareous microfossil zones in Figure 2.

This study presents part of the data analyzed for a study of Eocene-Oligocene paleoceanography being undertaken jointly with other authors (B. H. Corliss, L. D. Keigwin, Jr., G. Keller, M.-P. Aubry, and W. A. Berggren). The sites were selected to give information on the paleogeographic distribution of species and changes in assemblage composition from the middle Eocene through early Oligocene.

METHODS

All samples were prepared by boiling the sediment for 10 min. in an equal mixture of 10% HCl and 30% H₂O₂ in order to dissolve the calcium carbonate and oxidize the organic matter. Clay minerals were

¹ Hay, W. W., Sibuet, J.-C., et al., *Init. Repts. DSDP*, 75: Washington (U.S. Govt. Printing Office).

² Moore, T. C., Rabinowitz, P. D., et al., *Init. Repts. DSDP*, 74: Washington (U.S. Govt. Printing Office).

Table 1. Present geographic location of the DSDP sites examined.

Site	Geographic Location	Latitude	Longitude	Water Depth (m)
64, 64.1	Ontong-Java Plateau	01°44, 56'N	158°36, 51'E	2052
65, 65.1		04°21, 21'N	176°59, 16'E	6130
69, 69A		06°00, 00'N	152°51, 93'W	4978
71A		04°28, 28'N	140°18, 91'W	4419
72		00°26, 49'N	138°52, 02'W	4326
73	Central equatorial Pacific	01°54, 58'S	137°28, 12'W	4387
77B		00°28, 90'N	133°13, 70'W	4291
161A		10°14, 25'N	139°57, 21'W	4939
163		11°14, 66'N	150°17, 52'W	5230
167		07°04, 10'N	176°49, 05'W	3176
216	Ninetyeast Ridge	01°27, 73'N	90°12, 48'E	2262
217		08°55, 57'N	90°32, 33'E	3030
220	Southeast Arabian Basin	06°30, 97'N	70°59, 02'E	4036
289	Ontong-Java Plateau	00°29, 92'S	158°30, 69'E	2206
292	Benham Rise	15°49, 11'N	124°39, 05'E	2943
356	São Paulo Plateau	28°17, 22'S	41°05, 28'W	3203
357	Rio Grande Rise	30°00, 25'S	35°33, 59'W	2086

separated from the insoluble residue by using their slower gravitational settling times: starting with a well-mixed and suspended sample residue, the upper 9 cm of the water column in a 400 ml beaker were siphoned off after 8 hr. of settling time. This process was repeated until, after 8 hours of settling time, the water column was clear. After the acid was washed out, a 0.5% sodium pyrophosphate solution in water was used in one washing to improve the suspension of clay minerals. For more details see Fenner (1982). This method effectively concentrates the equivalent quartz grain sizes larger than 2 μm .

Aroclor 4465 (n.d. = 1.66, solvent: Xylene; Göke, 1958) was used as the mounting medium.

Depending on the abundance of diatoms in a sample, all or half of a 20 \times 20 mm slide was scanned under the light microscope at a magnification of 1000 \times to determine the relative abundances of the diatom species.

MATERIAL

During the late Eocene to early Oligocene, Pacific DSDP Sites 65, 69, 71, 72, 73, 77, 161, 163, and 167 were located in the equatorial belt of high accumulation

rates of biogenic silica (e.g., Boström et al., 1963, Leinen, 1981). Site 292 was located to the west of this zone of high productivity, possibly representing its western extension (Ingle et al., 1975). DSDP Sites 64 and 289, on the Ontong-Java Plateau, were ~10° of latitude further south than their present locations (Hammond et al., 1975), and DSDP Sites 216, 217, and 220, in the Indian Ocean, were also about 10° of latitude further south, just south of the equator (McElhinny, 1970; McKenzie and Sclater, 1971). At DSDP Site 357, on the Rio Grande Rise, which was always located outside of the equatorial and circum-Antarctic upwelling regions, only short intervals of diatom-bearing sediments occur, one in the lower middle Eocene, the other in the upper Oligocene. A similar pattern of restricted occurrence and preservation of siliceous microfossils in the lower middle Eocene and upper Oligocene also occurs at Site 356 on the São Paulo Plateau, at nearby DSDP Sites 22 and 516F on the western Rio Grande Rise, and in the southwestern Brazil Basin at Site 515 (Maxwell et al., 1970; Shipboard Scientific Party, Leg 73).

DSDP Sites 356 and 357 (Southwest Atlantic)

In the Eocene of Sites 356 and 357 on the São Paulo Plateau and western Rio Grande Rise, diatoms are encountered only during a short interval within the upper part of Zone NP15 to the lower part of Zone NP16. At Site 357 diatoms are dissolved in the Eocene sediments above and below a short interval in Core 24 (Fenner, unpublished data). At Site 356, 75 m of diatomaceous sediments of middle Eocene age are unconformably overlain by early Miocene and underlain by middle and early Eocene sediments barren of diatoms. This short interval of increased productivity and biogenic silica preservation is related to a short phase of volcanic activity in this region, combined with uplift and the presence of volcanic islands

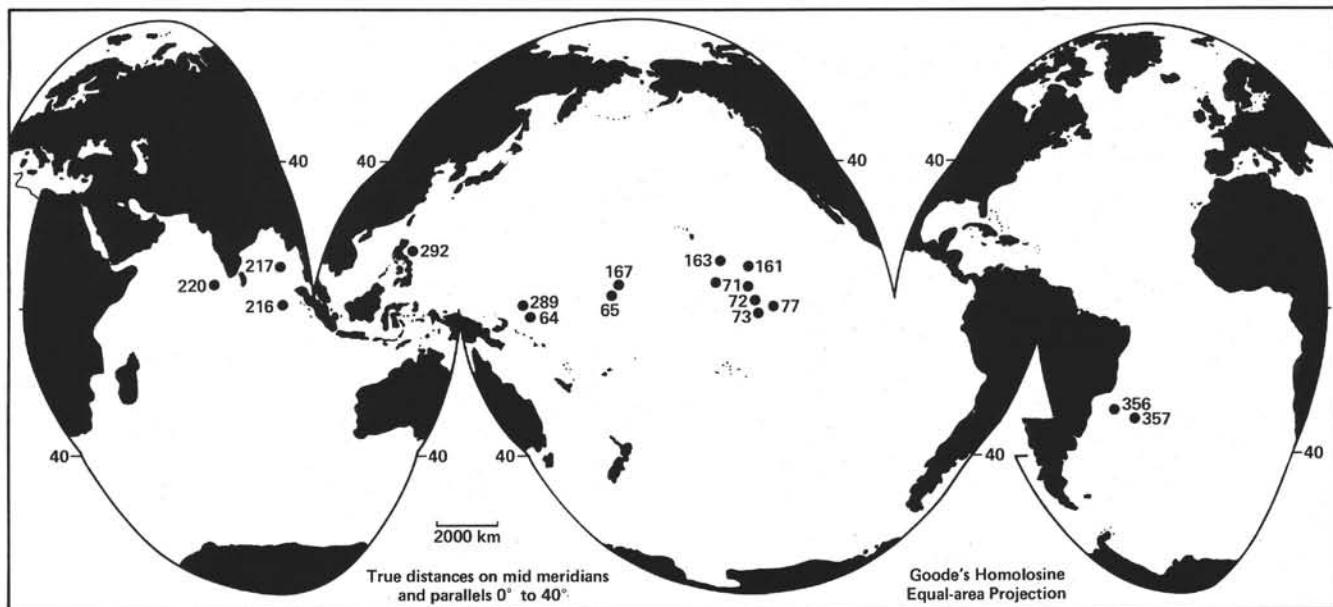


Figure 1. Geographic location of DSDP sites examined in this chapter.

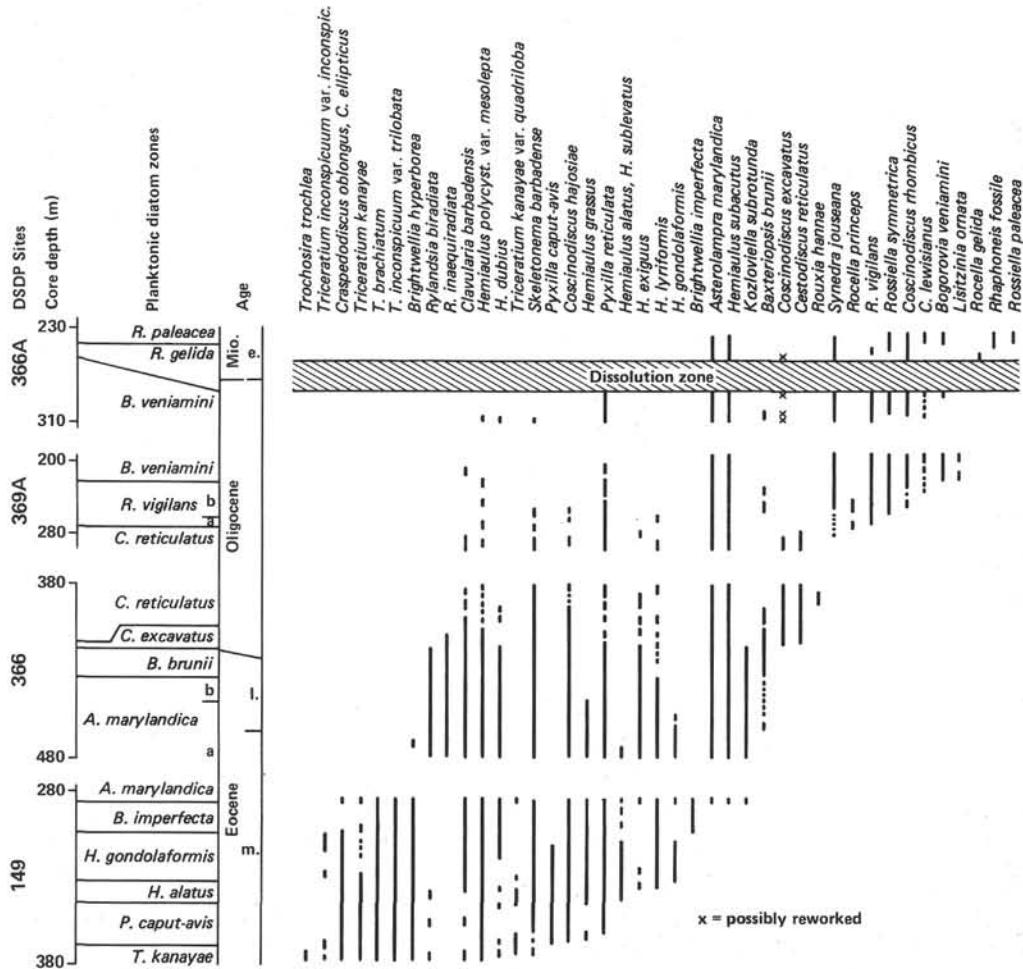


Figure 2. Eocene-Oligocene planktonic diatom zonation of Fenner (in press), as established for the low-latitude Atlantic and the Caribbean. Determination of other planktonic microfossil zones: Holes 366, 366A, 369A by Krasheninnikov and Pflaumann (1978), Cepek (1978), Johnson (1978); and Hole 149 by Hay and Beaudry (1973), Bukry (1973), and Riedel and Sanfilippo (1973). I = *T. bromia* and *P. goetheana* zones undifferentiated. Eocene/Oligocene boundary following Bukry (1978), who based it upon the LAD of *Discoaster barbadiensis*.

(Fodor and Thiede, 1977; Barker et al., 1981; Tjalsma, in press); thus it might be due in part to an "island effect."

The preservation of diatoms is much better at Site 356 (Cores 6-9) than at Site 357 (Core 24; Table 2). The dominant and characteristic species at Site 356 (Fig. 3) are (1) cosmopolitan species such as *Asterolampra vulgaris*, *Brightwellia* spp., *Craspedodiscus oblongus*, *Hemiaulus polycystinorum* var. *mesolepta*, *Pyxilla reticulata*, and tri- and quadrilobate varieties of *Triceratium inconspicuum*; and (2) low-latitude species as *Clavularia barbadensis*, *H. alatus*, *H. altar*, *H. exiguis*, *H. polycystinorum* var. *dissona*, *P. caput-avis*, *P. gracilis*, *Skeletonema barbadense*, and *T. kanayae*. *H. grassus* and *H. gondolaformis* among the low-latitude stratigraphic marker species were not found. As both species are very robust and dissolution-resistant, their absence could either indicate that they were geographically restricted to the low latitudes or could be due to the greater age of the sediments. The composition of the diatom assem-

blages suggests that the first is the case. The succession of first and last occurrences of diatom species at Site 356 is the same as at Site 149 in the Caribbean, except for *C. barbadensis*, which seems to appear later at Site 356. Thus the planktonic diatom zonation established in the low-latitude Atlantic and Caribbean (Fenner, in press) applies at Site 356.

The first consistent occurrence of *Hemiaulus alatus* in Core 7 and the presence of *Pyxilla caput-avis*, common *Triceratium kanayae* var. *kanayae* and *T. kanayae* var. *quadrilobata* in Cores 8 and 9 place Core 7 to 9 in the *H. alatus* Zone and Cores 8-9 below in the *P. caput-avis* Zone. Because the low-latitude marker species *H. gondolaformis* is absent, Core 6 cannot be clearly placed into one of the low-latitude diatom zones. But the consistent presence of *Clavularia barbadensis* and the decrease in abundance of *T. kanayae* (both varieties) suggest that Core 6 has an age equivalent to that of the *H. gondolaformis* Zone of the low latitudes. The corre-

Table 2. Diatom species occurrence in Eocene and Oligocene samples from Site 357, Rio Grande Rise.

	Planktonic diatom stratigraphy	Planktonic foraminiferal zones (Boersma, 1977)	Calcareous nannoplankton zones (Perch-Nielsen, 1977)	Core-Section (level in cm)		<i>Actinopychus senarius</i>	<i>A. stella</i>	<i>Arachnoidiscus</i> spp.	<i>Asterolampra grevillei</i>	<i>A. marylandica</i>	<i>A. tela</i>	<i>A. vulgaris</i>	<i>Auliscus</i> spp.	<i>Biddulphia tridens</i>	<i>Brightwellia</i> spp.	<i>Bogorovia veniamini</i>	<i>Cestodiscus pulchellus</i>	<i>C. strokesianus</i>	<i>C. sp. 1</i> Fenner, 1982	<i>Cladogramma conicum</i> var. <i>campanulatum</i>	<i>Coscinodiscus descrescens</i>	<i>C. extravagans</i>	<i>C. lewisianus</i>	<i>C. marginatus</i>	<i>C. payeri</i>	<i>C. rhombicus</i>	<i>C. symbolophorus</i>	<i>C. sp.</i>	<i>Craspedodiscus elegans</i>	<i>C. oblongus</i>	<i>Diploneis</i> spp.	<i>Endictya robustus</i>
				late Oligo.	R. <i>gelida</i>																											
middle Eocene	P11	NP 15	N4 NN1 P22 NP 25	14	1	90.0	R	F								R P	P		R	F P R F										P		
				14	2	90.0																										
				15	2	29.0																										
				15	2	119.0																										
				15	CC	0.0																										
				16	1	90.0	R	P								R A	P		R	F P C R										P		
				16	2	90.0										C C	P		R C	R R C C										F		
				17	1	78.0	P									C C	P		R R	R R C C												
				17	2	90.0										R C	P		F R R	F R R												
				17	3	88.0										R R	P		P	P											P	
late Oligocene	P22	NP 25	N4 NN1 P22 NP 25	17	4	88.0																										
				17	5	88.0																										
				17	6	96.0	F	P																								
				18	1	40.0																										
				24	4	54.0		P																								P
				24	5	101.0																										P
middle Eocene	P11	NP 15	N4 NN1 P22 NP 25	24	5	8.0		P																								F
				24	5	84.0		P																								R
				24	5	104.0		R																								R
				24	6	60.0	F																									R

Note: The relative abundances are given as P = <0.1%, F = 0.1–1%, R = 1–10%, C = 10–30%, A = >30%. Core 357-19 to Section 357-24-4 was barren of diatoms as were Core 25 and below. In addition to the species listed, the following occurrences were noted: *Spermatogonia* spp., Sample 357-15, CC (P); *Macraea barbadensis* (silicoflagellate), Samples 357-24-5, 8–9 cm (P), and 357-24-6, 60–61 cm (P); freshwater diatoms (*A. granulata*), Samples 357-16-1, 90–91 cm (F), 357-16-2, 90–91 cm (P), 357-17-6, 96–97 cm (P), and 357-18-1, 40–41 cm (P).

lation of the ranges of stratigraphically important diatom species and the planktonic diatom zones to the calcareous microfossil zones at this site is given in Figure 3.

At DSDP Site 357, further east on the western Rio Grande Rise, diatom preservation in the middle Eocene diatomaceous section is relatively poor and most of the low-latitude species such as *T. kanayae*, *Clavularia barbadensis*, and *Skeletonema barbadense*, which are still present at DSDP Site 356, are not found. Only the very robust *P. caput-avis* is present. But as the also very robust *T. kanayae* is missing from the assemblage, the absence of these low-latitude species at this site during this interval is interpreted as being due to paleogeographic restriction (as Boersma [1977] reported also for the planktic foraminifers), rather than as being due to selective dissolution. Whether *H. alatus* is absent for stratigraphic or paleogeographic reasons cannot be decided. Further south, in the middle Eocene section of DSDP Site 512 on the Falkland Plateau (Gombos, 1983), all of the low-latitude species are definitely no longer present. Because most of the low-latitude marker species are absent, the occurrence of *P. caput-avis*, *Craspedodiscus oblongus*, and *T. inconspicuum* var. *inconspicuum* with *P. reticulata* allows us to place Core 24 of Site 357 into the middle Eocene only, with an age equivalent to some-

where within the *P. caput avis*—*H. alatus*—*H. gondoliformis* low-latitude zones. This age determination is in agreement with the dating by calcareous nannofossils—upper part of Zone NP15 (*Discoaster bifax* Subzone) according to Perch-Nielsen (1977) and Bukry (1977)—and the planktonic foraminifers—Zone P11 (Boersma, 1977).

The second diatomaceous interval in the Paleogene of Site 357 (Cores 15–18) occurs in the upper Oligocene (Table 2, Fig. 4). The diatom assemblages are characterized by the frequent occurrence of *Synedra jouseana* and, in addition, *Asterolampra tela*, *Bogorovia veniamini*, *Coscinodiscus extravagans*, *C. lewisianus*, *C. rhombicus*, *Hemiaulus subacutus*, *Lisitzinia ornata*, *Rocella gelida*, *R. princeps*, *R. vigilans*, and *Rossiella symmetrica* species indicating a late Oligocene age. The first occurrence of *Rocella gelida* is used in the circum-Antarctic and in the low latitudes to define the base of the *R. gelida* Zone (Fenner, in press). The diatomaceous section does not extend beyond the earliest Oligocene, as is indicated by the presence of *Hemiaulus taurus* and *Rocella schraderi*. Neither species known to occur just above the Oligocene/Miocene boundary (e.g., *Raphoneis fossile* and *Rossiella paleacea*, typical in low-latitude sites; Fenner 1982) nor the cosmopolitan species

Table 2. (Continued).

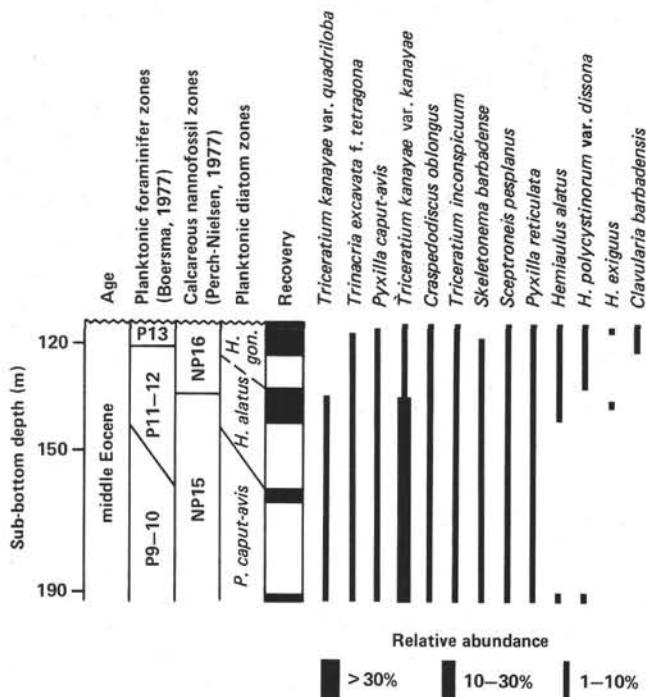


Figure 3. Ranges of stratigraphically important diatom species in middle Eocene cores of DSDP Site 356, São Paulo Plateau.

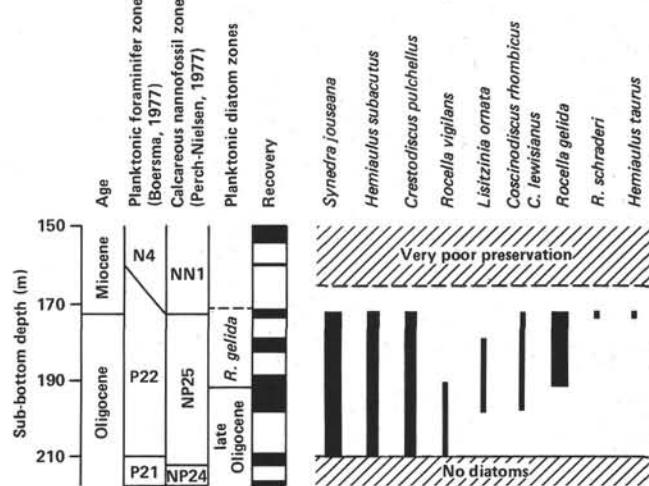


Figure 4. Ranges of stratigraphically important diatom species in late Oligocene core of DSDP Site 357, Rio Grande Rise. Relative abundance as in Figure 3.

Rhaphidodiscus marylandicus and *Thalassiosira fraga*, were found.

Most of the late Oligocene species occurring at Site 357 (*C. rhombicus*, *B. veniamini*, *L. ornata*, *Rossiella symmetrica* and *S. jouseana*) are known to be cosmopolitan (Fenner, 1982). Only one species, which seems

to be restricted to the low and middle latitudes (*Hemiaulus subacutus*) is present at this site. Other species, occurring at this site in the latest Oligocene, such as *H. taurus*, are typical high southern latitude species, thus making application of the circum-Antarctic planktonic diatom zonation more appropriate and reflecting an extension of "cool," high-latitude species toward the equator. A comparison with the Eocene diatom assemblages described from the Falkland Plateau (Gombos, 1976; Gombos and Ciesielski, 1983; Gombos, 1983; Fenner, in press) shows that on the Falkland Plateau in the middle Eocene (upper Lutetian; Gombos, 1983) the diatom assemblages were characterized by the absence of typical low-latitude species, and at least one indigenous circum-Antarctic species had already developed (*H. vitreus*). In the late Eocene and early Oligocene, the number of indigenous circum-Antarctic diatom species increases with the first appearances of *H. characteristicus*, *H. pacificus*, and *H. rectus* var. *twista*, which are common elements of the assemblages. But these species remained restricted to the circum-Antarctic region and were not found at the São Paulo Plateau or Rio Grande Rise. Typical high southern latitude species did not extend toward the equator before the latest Oligocene.

The good preservation of diatoms in the upper Oligocene sediments (Site 357) on the western Rio Grande Rise is correlated to increased sediment accumulation rates, from 5–7 m/m.y. in the diatom-barren sediments below to ~30 m/m.y. in the diatomaceous section (Perch-Nielsen et al., 1977). The calcium carbonate content of the sediment is constant at about 80% (Fenner, unpubl. data). In the diatomaceous interval traces of volcanic ash are found (Perch-Nielsen et al., 1977). Opal phytoliths, which are siliceous fillings of the epidermal cells of Gramineae and Equisetales, and thickly silicified valves of the freshwater diatom, *Aulacosira granulata*, occur consistently during this interval, whereas in the interval of increased preservation of siliceous skeletons during the middle Eocene no freshwater diatoms were found and only rare, single, opal phytoliths (Fenner, unpubl. data). The biosiliceous terrigenous components could be derived from nearby volcanic islands (for which there is no evidence in the late Oligocene) or from the South American hinterland. Their presence in marine sediments might be correlated to increased aridity and/or wind intensity around the time of the Oligocene/Miocene boundary. On the Sierra Leone Rise off Northwest Africa, an identical increase in the abundance of land-derived biosiliceous particles is observed (Fenner, 1982).

Equatorial Pacific Sites (Table 3)

In the equatorial Pacific sites, except for Site 167, preservation of diatoms in the Eocene is very poor because of dissolution and silica diagenesis, but in the early Oligocene it is moderate to good. This offers us the opportunity to study the paleogeographic distribution of the diatom species found to be characteristic of the low-latitude Atlantic Ocean (Fenner, 1982) and to test the planktonic diatom zonation established there (Fenner, in press).

At all sites, the same *Cestodiscus* spp.-dominated assemblage was found in the early Oligocene as is present in the equatorial Atlantic. Characteristic species were *C. gemmifer*, *C. convexus*, *C. reticulatus*, *C. trochus*, *C. parvula*, *Coscinodiscus excavatus*, *C. oligocenicus*. Furthermore, the late Eocene diatom assemblages found at Site 167 are characterized by the same species that dominated the low-latitude Atlantic diatom assemblages.

Site 64

This site on the Ontong-Java Plateau was spot-cored. Sample 64-10-1, 70–74 cm, at 847 m depth, has a typical early Oligocene diatom assemblage dominated by *Cestodiscus gemmifer* and *C. parvula*. Because the two stratigraphic guide species, *C. reticulatus* and *Coscinodiscus excavatus*, co-occur, this sample was assigned to the *Cestodiscus reticulatus* Zone. This age assignment is in accordance with the biostratigraphic results of Bukry (1971a), Martini and Worsley (1971), and Gartner (1971), who place Section 64-10-1 in Zone NP24 (*Helicosphaera reticulata* Zone). Deeper samples belong in the Eocene. Diatom preservation is poor.

Site 65 (Table 3)

This site in the central basin of the central equatorial Pacific was continuously cored. Thin chert layers and thin layers of turbidite sediments occur in the lower Oligocene and upper Eocene (Heath and Moberly, 1971a, b). The turbidites explain the presence of occasional well-preserved calcite tests, which must have originated above the contemporary calcite compensation depth. Throughout the sequence, reworked middle Eocene radiolaria are reported by Riedel and Sanfilippo (1971). However, reworking did not seem to be a major factor in the samples examined for this study. Only one evidently reworked *Hemiaulus* was found, in Sample 65-16-2, 82–86 cm. The presence of *Coscinodiscus excavatus* in Sample 65-14-1, 70–74 cm, together with the absence of *Cestodiscus reticulatus*, places this sample in the *C. excavatus* Zone, which is in accordance with the age determination by calcareous nannofossils (Bukry, 1971a) as *Helicosphaera reticulata* to *Discoaster barbadiensis* zones. The poor preservation of diatoms in samples from Cores 15 and 16, and from Hole 65.1 does not allow a more detailed age assignment than late Eocene, so that a test of the completeness of the upper Eocene section is impossible.

Site 69A

Planktonic diatoms place Sample 69A-8-1, 70–74 cm in the *Cestodiscus reticulatus* Zone, agreeing with the age given by calcareous nannofossils and planktonic foraminifers (*Helicopontosphaera reticulata* Zone or *Discoaster tani ornatus* Zone, respectively [Bukry, 1971b; Haq and Lipps, 1971] and Zone P20 to P18 [Beckmann, 1971]). There is a sharp change in lithology from the calcareous nannofossil ooze in Core 69A-8 to the radiolarian ooze below. Cores 69A-9 and 69A-10 are placed in the *Thyrsocyrtis bromia* Zone (Moore, 1971). The diatoms suggest an earliest Oligocene age in the *Coscinodiscus excavatus* Zone, because *Cestodiscus reticulatus*

latus is absent and *Coscinodiscus excavatus* and *Thalassiosira bukryi* are present. There is no reason to believe that the occurrence of these species is due to downhole contamination, as was suggested for the radiolarians by Moore (1971). In Core 69A-11 only one species, *Riedelia pacifica*, was found. Radiolarians place this core in the middle Eocene *Thyrocyclitis triacantha* Zone (Moore, 1971). Sample 69-6-1, 72-76 cm is also within the middle Eocene, based on the presence of *Hemiaulus polycystinorum* var. *asymmetrica* and *Odontella hastata*. Radiolarian stratigraphy assigns this sample to the middle Eocene *Podocyclitis chalara* Zone (Moore, 1971). This suggests that the upper Eocene section is missing at this site.

Site 71A

The three spot cores studied at this site did not recover diatoms across the Eocene/Oligocene boundary. Core 71A-1 is assigned to the early Oligocene *Cestodiscus reticulatus* Zone, which is in agreement with the age determinations made on the basis of planktonic foraminifers (Zone P18-19), calcareous nannofossils, and radiolarians (*Thyrsocyrtis tuberosa* Zone) by Beckmann (1971), Haq and Lipps (1971), Bukry (1971b), Hay (1971), and Moore (1971). In Cores 71A-2 and 71A-3, which are probably of late Eocene age (*T. bromia* Zone; Moore, 1971), no diatoms were present.

Site 72

The lower Oligocene and upper Eocene were continuously cored, but the lower Oligocene section is incomplete and the Eocene section badly contaminated by younger material. Only one sample was examined from this site (Sample 72-9-1, 70-74 cm); it is placed in the earliest Oligocene.

DSDP Site 73 (Table 4)

At this continuously cored site, a short hiatus at the Eocene/Oligocene boundary is indicated by calcareous nannofossils (Bukry, 1971b). High abundances of down-mixed, well-preserved Neogene diatoms are seen in the uppermost section of each core. Eocene-Oligocene diatoms, by contrast, are poorly preserved and only the most dissolution-resistant species are present. The interval from Core 73-15 to Section 73-19-3 is assigned to the early Oligocene *Cestodiscus reticulatus* Zone based on the presence of *C. reticulatus*. *Coscinodiscus excavatus*, which is known from other equatorial sites to range below *Cestodiscus reticulatus*, is found only as far down as Section 73-18-5, although it is a very robust and also relatively dissolution-resistant species. There are two possibilities. *C. excavatus* may be absent from Core 73-19 because of dissolution. This point is supported by the high abundance of *Cestodiscus* spp. down to Section 73-19-3 that gives this section a typical early Oligocene character. Alternatively, the presence of *C. reticulatus* in Core 73-19 might be due to downhole contamination into a barely recognizable late Eocene assemblage. Such contamination is definitely evident in the presence of down-worked Neogene diatom species. Age determinations using planktonic foraminifers and calcareous nan-

nofossils (Bukry, 1971b; Haq and Lipps, 1971; Hay, 1971; Beckmann, 1971) place Core 73-19 in the late Eocene or later middle Eocene. Below Section 73-19-3, preservation of diatoms becomes very poor.

Site 77B

Diatom preservation in this continuously cored early Oligocene site is moderate to good, allowing determination of the *Cestodiscus reticulatus* and *Coscinodiscus excavatus* zones. The assemblages are dominated by species of the genus *Cestodiscus*. Diatoms are absent from Cores 52 and 53 (both Eocene).

Site 161A

The lithology changes from nannofossil chalk in the lower Oligocene (Cores 161A-2 to 161A-9) to radiolarian ooze in the upper and middle Eocene below. Cores 161A-2 to 161A-9 are assigned to the early Oligocene *Cestodiscus reticulatus* Zone because *C. reticulatus* co-occurs with *Coscinodiscus excavatus*. Preservation of diatoms is moderate. Below, in Cores 161A-10 to 161A-14 preservation of diatoms is extremely poor. The absence of the early Oligocene marker species and the presence of *Cestodiscus trochus* and *Kozloviella meniscosa* suggest a late Eocene age for Cores 161A-10 and 161A-11.

Site 163

The early Oligocene and Eocene at this site is almost barren of calcareous microfossils. The clayey diatomaceous ooze of Sample 163-4-1, 70-74 cm is placed in the *Cestodiscus reticulatus* Zone, which is in agreement with the age determination using radiolarians (*Thyrsocyrtis tuberosa* Zone, Dinkelman, 1973). In the Eocene radiolarian ooze below, thin chert layers are found and preservation of diatoms is poor. The assemblages are dominated by *Riedelia pacifica* and *Coscinodiscus marginatus*.

Site 167 (Fig. 5)

At this site on the Magellan Rise, the zonations of the planktonic calcareous microfossils (Douglas, 1973; Roth, 1973) show no hiatus at the Eocene/Oligocene boundary. Although diatom valves show definite dissolution, their preservation is moderate—enough to apply the planktonic diatom zonation of Fenner (in press). The Eocene/Oligocene boundary is not associated with a strong change in preservation. At this site the sequence of first appearances just below and just above the Eocene/Oligocene boundary is the same as in DSDP Site 366, in the equatorial Atlantic (Fenner, 1982). *Thalassiosira bukryi* appears first, followed by *Coscinodiscus excavatus* and then *Cestodiscus reticulatus*, placing the Eocene/Oligocene boundary between Samples 167-24-1, 48-53 cm and 167-25, CC. Large specimens of *C. pulchellus* var. *novazealandica* are common during the interval of transition from the Eocene to the Oligocene. *Kozloviella subrotunda*, *Asterolampra punctifera*, *Coscinodiscus descrescens*, and *Hemiaulus polycystinorum* var. *mesolepta* last occur below the Eocene/Oligocene boundary. The decrease in abundance of valves of *Hemiaulus* spp.

Table 3. Diatom species occurrences in Eocene and Oligocene samples from DSDP sites in the central equatorial Pacific and Ontong-Java Plateau.

is not so drastic as at Site 366, but the increase in valves of *Cestodiscus* spp. is as dramatic. Samples 167-25,CC to 167-27,CC are placed in the late Eocene *Baxteriopsis brunii* Zone, using the first appearance of *B. brunii*. Samples 167-28-1, 59-63 cm to 167-32-2, 59-63 cm belong in the *A. marylandica* Zone.

In Cores 33-to 35, chert is common and no diatoms are preserved.

Site 289 (Fig. 6)

At this site on the Ontong-Java Plateau, the preservation of the diatom valves is moderate to poor throughout the lower Oligocene and upper Eocene. Only in Core 106 (just above the chert) does preservation become very poor. The first occurrences of *Coscinodiscus excavatus* and *Cestodiscus reticulatus* place the Eocene/Oligocene boundary between Cores 101 and 102. The change in assemblage composition to an assemblage dominated by *Cestodiscus* spp. occurs slightly below, between Cores 102 and 103. The late Eocene preservation of diatoms is

poor. The identification of the planktonic diatom zones therefore has to be viewed with caution. The calcareous nannofossil indicate a short hiatus at the Eocene/Oligocene boundary (Shafik, 1975).

Site 292

At this site on the Benham Rise, a small hiatus at the Eocene/Oligocene boundary is indicated by calcareous nannofossils (Ellis, 1975). The preservation of diatom valves at this site is poor, with an interval of especially intense biogenic silica dissolution during the earliest Oligocene (Cores 33-35) that reaches its maximum in Sample 292-33-1, 10-14 cm, which is completely barren of diatoms. In the late Eocene, *Grammatophora* spp. are common in the diatom assemblages, indicating a shallow-water environment. The last occurrence of *Incertae Sedis: Genus et species indet.* (3) correlates with the nannofossil-determined Eocene/Oligocene boundary. Samples from Core 39, Section 2 are barren of diatoms.

Table 3. (Continued).

Indian Ocean Sites (Table 5)

Sites 216 and 217 (Table 5)

During the Late Cretaceous and Paleocene, both sites, located on the Ninetyeast Ridge (Table 1), experienced biosiliceous accumulation that continued at the more southern Site 216 into the middle Eocene. With continuing northward plate movement (McElhinny, 1970; McKenzie and Sclater, 1971, Whitemarsh et al., 1974) these two sites gradually moved out of the belt of high primary productivity. At both sites late Eocene and Oligocene nannofossil chalks with rare and poorly preserved diatoms were deposited at the northern fringe of the high-productivity belt. The poor preservation of diatoms at both sites allows no exact determination of the Eocene/Oligocene boundary.

Site 216

The Eocene/Oligocene boundary must be placed below Sample 216-13-1, 144-149 cm, using the first occur-

rence of *Cestodiscus reticulatus* as definite early Oligocene. Using calcareous nannofossils and planktonic foraminifers, Gartner (1974), Bukry (1974), and Berggren et al. (1974) place the Eocene/Oligocene boundary at a deeper level, within Core 15.

DSDP Site 217

The last abundant occurrence of *Hemiaulax* spp. is in Sample 217-9-5, 59-64 cm. This, and the presence of *Kozloviella subrotunda* in Sample 217-9-6, 77-82 cm, place the Eocene/Oligocene boundary between Samples 217-9-4, 41-46 cm and 217-9-6, 77-82 cm. Calcareous microfossils (Gartner, 1974; Bukry, 1974; Berggren et al., 1974) place the Eocene/Oligocene boundary between Sections 217-9-5 and 217-9-6.

Site 220 (Table 6)

At this site, at the southeastern tip of the Arabian Basin, diatom abundance, diversity, and preservation decrease from the middle Eocene toward the late Eocene.

Table 3. (Continued).

Age	Plankt. diatom stratigraphy	Core-Section (level in cm)	<i>Actinocyclus octonarius</i> <i>Actinopychus intermedius</i> <i>A. senarius</i> <i>Arachnoptychus</i> sp.	<i>Asterolampra affinis</i> var. <i>cellulosa</i>	<i>A. crenata</i> <i>A. grevillei</i> <i>A. aff. grevillei</i> <i>A. marylandica</i>	<i>A. punctifera</i>	<i>A. telia</i> <i>A. uraster</i> <i>A. vulgaris</i>	<i>Asteronphaulus oligocenicus</i>	<i>Aulacodiscus</i> sp.	<i>Baxteriopsis brunii</i>	<i>Cestodiscus convexus</i> <i>C. gemmifer</i> <i>C. parvula</i>	<i>C. pulchellus</i>	<i>C. pulchellus</i> var. <i>novazealandica</i>	<i>C. reticulatus</i>	<i>C. stokesianus</i> <i>C. trochus</i>	<i>C. sp. + C. SD. 3</i>	<i>Chaetoceros</i> sp. (bristles)	<i>C. sp. 3</i> Fenner, 1982	<i>Coscinodiscus argus</i>	<i>C. asteromphalus</i>	<i>Thalassiothrix bukryi</i>	<i>C. demergitius</i>	<i>C. marginatus</i> <i>C. obscurus</i> <i>C. oligocenicus</i> <i>C. radiatus</i> <i>C. sellatus</i>	<i>C. senarius</i> var. <i>parvula</i> <i>C. hajosiae</i> <i>C. symbolophorus</i> <i>C. tuberculatus</i>	<i>C. vigilans</i>	
early Oligocene	<i>C. reticulatus</i>	778 49 5 32.0			F						R R C R C C C C R C C R F F A	F F F C F F C R F C F F R	P	P	R	R R C R C R	R R C R C R	R R C R C R	R R C R C R	P						
		778 49 6 28.0			F																					
		778 50 1 28.0			F																					
		778 50 3 34.0			F																					
		778 50 4 34.0			F																					
		778 50 5 34.0			F																					
		778 50 6 34.0			F																					
		778 51 1 34.0			F																					
		778 51 2 34.0			F																					
		778 51 3 39.0			F																					
middle Eocene	<i>C. reticulatus</i>	778 51 4 38.0																								
		778 51 5 40.0																								
		778 51 6 36.0																								
		161A 2 1 67.0			P																					
		161A 3 2 30.0			P P																					
		161A 4 1 70.0			F P R																					
		161A 5 1 66.0			R R R																					
		161A 6 1 70.0			F F																					
		161A 7 1 73.0																								
		161A 8 1 70.0																								
e. Olig.	<i>C. ret.</i>	161A 9 1 169.0																								
		161A 10 2 70.0																								
middle Eocene	<i>C. ret.</i>	161A 11 2 35.0																								
		161A 12 1 70.0																								
		161A 13 1 90.0																								
		161A 14 1 82.0																								
		163 4 1 70.0																								
		163 5 1 93.0																								
		163 6 1 70.0																								
		163 7 1 70.0																								
		163 9 1 70.0																								
		163 10 1 74.0																								
		163 11 1 74.0																								
		163 12 1 74.0																								

Note: For explanation of the above symbols, see Table 2. In addition to these species, the following occurrences were observed: *Spermatogonia* spp.: Samples 64-10-1, 70-74 cm (P), 72-76 cm (F), 163-9-1, 70-78 cm (F), 167-24-1, 48-53 cm (F), 167-32-2, 59-63 cm (P), 289-94-2, 55-60 cm (P), 289-97-1, 120-125 cm (P), 289-97-4, 122-127 cm (F), 289-98-2, (P), 65-13-1, 68-72 cm (P), 77B-46-1, 32-38 cm (P), 161A-5-1, 66-70 cm (P), 167-22-1, 59-64 cm (P), 289-94-2, 55-60 cm (P), 289-94-3, 55-60 cm (P), 289-97-4, 122-127 cm (P), 292-36-3, 129-133 cm (R), 292-36, CC (F), 292-37-1, 129-134 cm (F).

No Oligocene samples were available for study. Because *Triceratium kanayae* is present, Sections 220-12-4 to 220-15-1 are considered middle Eocene.

DISCUSSION

The assemblage analysis of the early Oligocene at the equatorial Pacific sites shows that the same new species of the genera *Cestodiscus* (*C. gemmifer*, *C. reticulatus*, *C. convexus*, and *C. parvula*) and *Coscinodiscus* (*C. excavatus*), which dominate the assemblages in the equatorial Atlantic are also dominant in the equatorial Pacific. Data from Site 167 show that the same is true for the latest middle and late Eocene. The characteristic and dominant species are *Baxteriopsis brunii*, *Clavularia barbadensis*, *Hemiaulus polycystinorum* var. *mesolepta*, *H. longicornis*, *H. subacutus*, and *Skeletonema barbadense*. Ages determined by the ranges of stratigraphic marker species from the equatorial Atlantic and Caribbean (Sites 149, 366, 369A; Fig. 2) agree with those de-

termined by calcareous microfossils. Such a homogeneous circum-equatorial species distribution and equal sequence of datum planes is a result of the open, low-latitude seaway between the Atlantic and Pacific and reflects a circum-equatorial circulation in Tethys.

The mid-latitude position of Sites 356 and 357, on the São Paulo Plateau and Rio Grande Rise, makes it possible to recognize latitudinal shifts in assemblage distributions. In the middle Eocene of Site 357, the assemblage is composed of low-latitude species and cosmopolitans; only two of the typical low-latitude stratigraphic marker species, *Hemiaulus grassus* and *H. gondolaformis*, are missing. Further south, on the Falkland Plateau, (Gombos, 1983) all the low-latitude species are absent; one indigenous circum-Antarctic species (*Hemiaulus vitreus*) is already present. In the late Eocene and early Oligocene, the number of indigenous circum-Antarctic planktonic diatom species increases to at least three—*Hemiaulus characteristicus*, *H. pacificus*, and *H. rectus* var.

Table 3. (Continued).

twista, which remain restricted to the circum-Antarctic belt. Only during the latest Oligocene do high-latitude diatom species spread as far toward the equator as the Rio Grande Rise. This extension of high-latitude "cool" species in the latest Oligocene seems to be correlated to a period of increased sediment accumulation, productivity, and wind intensity at Sites 357 and 516. Increased productivity and wind intensity were also found for the same time interval at Hole 366A (Fenner, 1982) and thus seem to be an event of more than local importance.

At Site 167 diatom preservation was moderate in the late Eocene, thus allowing us to test the extent of the assemblage change found in equatorial Atlantic Site 366 (Fenner, 1982), which is correlated there to the strong shift in oxygen isotopes near the Eocene/Oligocene boundary (Vergnaud Grazzini and Rabussier-Lointier, 1980). No mass extinctions of planktonic diatom species occurred at the Eocene/Oligocene boundary at Sites 366 or 167, but a decrease in abundance of *Hemiaulus* spp. and a strong increase in abundance of *Cestodiscus* spp.

at both sites occurred at the base of the *Coscinodiscus excavatus* Zone.

The percentage of *Cestodiscus* spp. in the late Eocene is about 10–20%. Their abundance strongly increases at the base of the *C. excavatus* Zone to 40–60%. As both genera *Hemiaulus* and *Cestodiscus* produce very robust and relatively dissolution resistant species, dissolution, unless it is very strong, should not mask this assemblage change. Indeed, in many other equatorial sites with more poorly preserved diatom assemblages (Sites 64, 65, 161, and 163) the strong increase in abundance of *Cestodiscus* spp. occurs at the base of the *Coscinodiscus excavatus* Zone. But this assemblage composition change is not always correlated with the base of the *C. excavatus* Zone. In other equatorial sites (Sites 69, 292, 216, 217) it occurs in the earliest Oligocene and at Sites 73 and 289 in the late Eocene.

A strong change in assemblage composition occurring so widely in the equatorial regions is most easily interpreted in terms of response to changing ecological pa-

Table 3. (Continued).

early Oligocene		Oligocene	m.	Eocene	late	early	Oligocene	Age
		A.	B.	C.	C.	C.	C. reticulatus	
		maryland.	brunii	exc.	reticulatus			Planktonic diatom zones
								Core-Section (level in cm)
								<i>Actinocyclus octonarius</i>
								<i>Actinoptychus intermedius</i>
								<i>A. senarius</i>
								<i>Arachnoidiscus</i> spp.
								<i>Asterolampra affinis</i> var. <i>cellulosa</i>
								<i>A. crenata</i>
								<i>A. grevillei</i>
								<i>A. insignis</i>
								<i>A. marylandica</i>
								<i>A. punctifera</i>
								<i>A. tala</i>
								<i>A. uraster</i>
								<i>A. vulgaris</i>
								<i>Asteromphalus oligocenicus</i>
								<i>Bacteriopsis brunii</i>
								<i>Castodiiscus convexus</i>
								<i>C. gemmifer</i>
								<i>C. parvula</i>
								<i>C. pulchellus</i>
								<i>C. pulchellus</i> var. <i>novaeseelandica</i>
								<i>C. reticulatus</i>
								<i>C. stokesianus</i>
								<i>C. trochus</i>
								<i>C. SD. + C. SD. 3</i>
								<i>Chaetoceros</i> sp. (bristles)
								<i>C. sp. 3</i> Fennier, 1982
								<i>Coscinodiscus argus</i>
								<i>C. asteromphalus</i>
								<i>Thalassiosira buckryi</i>
								<i>C. denegritus</i>
								<i>C. marginatus</i>
								<i>C. obscurus</i>
								<i>C. oligocenicus</i>
								<i>C. radiatus</i>
								<i>C. sellatus</i>
								<i>C. senarius</i> var. <i>parvula</i>
								<i>C. hajosiae</i>
								<i>C. symbolophorus</i>
								<i>C. symbolophorus</i> group
								<i>C. tuberculatus</i>
								<i>C. vigilans</i>

rameters and suggests that the tectonic, climatic, and oceanographic changes that occurred close to the Eocene/Oligocene boundary were not only a high-latitude and bottom-water event, but must also have had a strong impact on surface waters in low latitudes. The factors or combination of factors important in causing the change in assemblage composition cannot be determined from available data. Oxygen isotope data (Douglas and Savin, 1975, Keigwin, 1980) suggest that the temperature decrease in the surface water of the low latitudes was small, but that the water column became stratified by cold bottom-water formation. Changes in stratification in the water column and vertical and horizontal circulation are major influences in nutrient recycling. Indications for such changes are discussed in Johnson (*in press*). Unfortunately, no oxygen isotope determinations yet exist for the equatorial Pacific sites in which diatoms are relatively well preserved through the critical

interval, so that one cannot test whether the change in composition of the diatom assemblage is correlated to the isotopic shift. If this proves to be the case, the compositional change could be used as an easy and fast indicator of the global cooling and stratification event near the Eocene/Oligocene boundary, and as such it might help in stratigraphic correlation.

TAXONOMIC LIST

All genera and species found in the sections investigated are listed in alphabetical order. For a discussion of the stratigraphic range and stratigraphic value of each species see Fenner, 1982. References given are for a description of the figure. Synonyms are mentioned only when they are still used. For a comprehensive list of synonyms see Mills (1933-1935) and VanLandingham (1967-1979).

For comparison, slides of samples from Moron and Nankoori, Nicobar Islands, generously provided by the British Museum (Natural History), were examined. From the same samples, *Cestodiscus pulchellus* and *C. stokesianus* were described by Greville. These species are illustrated in Plates 1 and 2.

Table 3. (Continued).

Genus *ACTINOCYCLUS* Ehrenberg, 1837

Actinocyclus ellipticus Grunow, in Van Heurck, 1883. Synonym: *Actinocyclus ellipticus* var. *elongatus* (Grunow) Kolbe, 1954, p. 20, pl. 3, figs. 28, 31. Hustedt, 1930, p. 533, fig. 303; Van Heurck, 1883, pl. 124, fig. 10.

Actinocyclus ingens Rattray, 1890. Kanaya, 1959, pp. 97-99, pl. 8, figs. 1-4; Kanaya, 1971, p. 554, pl. 40, figs. 1-8; Schrader, 1973, p. 701, pl. 18, figs. 1-4, 7.

Actinocyclus ehrenbergii var. *tenella* (Breb.) Hustedt, 1930 (Plate 2, Figs. 1-3). Hustedt, 1930, pp. 530-533, fig. 302.

Remarks. The valve outline is circular. The valve face is flat. The areolae are arranged in 4 to 6 sectors, which are separated from each other by radial rows of areolae running from the center to the margin. Within each sector the areolae are arranged in straight rows parallel to the middle row, which, like the sector-dividing rows of areolae, reach from the center of the valve to the margin. Each of the sector-dividing rows ends at the margin with a labiate process. In one of the sectors a pseudocellus is present at the border between the valve face and the margin. The size of the areolae on the valve face is almost constant, but varies with the valve size, there being 9-11 areolae in $10\text{ }\mu\text{m}$. On the margin, there are 15 areolae in $10\text{ }\mu\text{m}$.

Actinocyclus octonarius Ehrenberg, 1837. Hustedt, 1930, pp. 525-528, fig. 289, as *A. ehrenbergii* Ralfs, in Pritchard.

Genus *ACTINOPTYCHUS* Ehrenberg, 1841

Actinoptychus intermedius A. Schmidt, 1886, in A. Schmidt, et al.

1874—. A. Schmidt et al., 1874—, pl. 91, fig. 3, pl. 109, fig. 12.
Actinoptychus senarius Ehrenberg, 1837. Hustedt, 1930, pp. 475–478,

Remarks. There was no attempt to determine accurately all "senarius-like" species of *Actinoptychus*, so that this name comprises all *Actinoptychus* species that have six alternatively raised or depressed sectors and no special structures as, for instance, hyaline fields, lines, or a hyaline marginal ring.

Actinoptychus stellata Schmidt, 1886, in A. Schmidt et al., (1874—).

Genus *ARACHNOIDISCUS* Deane, ex Pritchard, 1852

Arachnoidiscus spp.

Remarks. Single specimens belonging to this group occur only very sporadically. Recent species of this genus are known to be littoral (Hustedt, 1930).

Table 3. (Continued).

Age	Planktonic diatom zones	Core-Section (level in cm)				<i>Actinocyclus octonarius</i>	<i>Actinopithicus intermedius</i>	<i>A. senaria</i>	<i>Astrachnoidiscus</i> spp.	<i>Asterolampra affinis</i> var. <i>cellulosa</i>	<i>A. crenata</i>	<i>A. gracilis</i>	<i>A. insignis</i>	<i>A. marylandica</i>	<i>A. punctifera</i>	<i>A. tela</i>	<i>A. uraster</i>	<i>A. vulgaris</i>	<i>Asteromphalus oligocenicus</i>	<i>Aulacodiscus</i> spp.	<i>Bacteriopsis brunii</i>	<i>Cestodiscus convexus</i>	<i>C. gemmifer</i>	<i>C. parvula</i>	<i>C. pulchellus</i>	<i>C. pulchellus</i> var. <i>novazealandica</i>	<i>C. reticulatus</i>	<i>C. stokesianus</i>	<i>C. troctus</i>	<i>C. sp. + C. sp. 3</i>	<i>Chaetoceros</i> sp. (bristles)	<i>C. sp. 3</i> Fenner, 1982	<i>C. r. C. argus</i>	<i>C. asteromphalus</i>	<i>Thalassiosira bukryi</i>	<i>C. demergitius</i>	<i>C. descrescens</i>	<i>C. elegantulus</i>	<i>C. excavatus</i>	<i>C. grossheimii</i>	<i>C. marginatus</i>	<i>C. obscurus</i>	<i>C. oligocenicus</i>	<i>C. radiatus</i>	<i>C. sellatus</i>	<i>C. senaria</i> var. <i>parvula</i>	<i>C. hajosiae</i>	<i>C. symbolophorus</i>	<i>C. symbolophorus</i> group	<i>C. tuberculatus</i>	<i>C. vigilans</i>
		Planktonic diatom zones	Core-Section (level in cm)	Core-Section (level in cm)	Core-Section (level in cm)																																														
late Eocene	<i>B. brun.</i> <i>A. marylandica</i>	289 102 1 68.0																		R	C	A																													
		289 102 1 125.0																		F	C	C																													
		289 103 1 62.0																		R	C	R																													
		289 103 1 123.0																		R	P	R																													
		289 105 CC 0.0																		R	P	R																													
		289 106 1 55.0																		P	R	R																													
		289 106 2 55.0																		P	R	R																													
		289 106 3 122.0																		P	R	R																													
		289 106 4 50.0																		P	R	R																													
		289 106 5 55.0																		P	R	R																													
early Oligocene		292 30 1 113.0																	P	R	P																														
		292 30 2 58.0																	P	R	C																														
		292 30 CC 0.0																	F	R	R																														
		292 31 2 8.0																	P	R	R																														
		292 31 2 111.0																	P	R	C																														
		292 31 CC 0.0																	P	R	R																														
		292 32 1 23.0																	P	R	R																														
		292 32 1 46.0																	R	F	F																														
		292 32 1 69.0																	P	R	R																														
		292 33 2 108.0																	P	R	P																														
?		292 33 CC 0.0																P	R	F																															
		292 34 1 10.0																P	R	R																															
		292 34 1 117.0																P	R	R																															
		292 35 CC 0.0																P	R	R																															
		292 36 1 139.0																P	R	P																															
		292 36 2 38.0																P	R	F																															
		292 36 2 94.0																P	R	R																															
		292 36 3 40.0																P	R	R																															
		292 36 3 129.0																P	R	R																															
		292 36 4 70.0																P	R	R																															
?		292 36 5 53.0																P	R	C																															
		292 36 CC 0.0																P	R	R																															
		292 37 1 129.0																P	R	R																															
		292 37 2 90.0																P	R	R																															
		292 37 3 40.0																P	R	R																															
		292 37 CC 0.0																P	R	R																															
		292 38 2 35.0																P	R	R																															

Genus *ASTEROLAMPRA* Ehrenberg, 1844*Asterolampra affinis* Greville var. *cellulosa* Forti, 1912. Synonyms:*Asterolampra schmidii* Hajos, 1976, p. 827, pl. 21, fig. 6; *Asterolampra decora* Greville sensu Jouse, 1977, pl. 49, fig. 7. Forti, 1913, p. 1566, pl. 3, figs. 3, 11.*Asterolampra crenata* Greville, 1862. Greville, 1862d, p. 47, pl. 8, figs. 4–16.*Asterolampra grevillei* (Wallich) Greville, 1860. Hustedt, 1930, p. 489, fig. 274.*Asterolampra aff. grevillei* (Wallich) Greville, 1860. This species differs from *Asterolampra grevillei* by having central mashes.*Asterolampra insignis* A. Schmidt, 1889, in A. Schmidt et al. (1874–).

A. Schmidt et al. (1874–), pl. 137, figs. 1–3.

Asterolampra marylandica Ehrenberg, 1844. Hustedt, 1930, pp. 485–487, fig. 271.*Asterolampra punctifera* (Grove) Hanna, 1927. Synonym: *Asterolampra affinis* Greville var. *punctifera* Grove, 1896, in A. Schmidt et al. (1874–), pl. 202, fig. 18. Hanna, 1927, p. 109, pl. 17, fig. 3.*Asterolampra tela* Gombos, in Gombos and Ciesielski, 1983. Gombos and Ciesielski, 1983, pl. 3, figs. 1–4.*Asterolampra uraster* Grove and Sturt, 1889. Grove and Sturt, 1889, p. 143, pl. 13, fig. 42; Gombos, 1980, p. 239, fig. 19.Genus *ASTEROMPHALUS* Ehrenberg, 1845*Asteromphalus oligocenicus* Schrader and Fenner, 1976. Synonym:*Asterolampra rotula* Greville sensu Jousé, 1977, pl. 55, fig. 14. Schrader and Fenner, 1976, p. 965, pl. 21, figs. 13–14; pl. 28, fig. 1.*Asteromphalus imbricatus* Wallich, 1860. Ralfs, in Pritchard, 1861, p. 837.Genus *AULACODISCUS* Ehrenberg, 1844*Aulacodiscus* spp.Remarks. Exact determination to the species level was not attempted. All observed specimens of this predominantly littoral genus are listed under *Aulacodiscus* spp., and are used as environmental indicators.Genus *AULACOSIRA (AULACOSEIRA)* Thwaites, 1848

Table 3. (Continued).

Remarks. This planktonic freshwater species is displaced in marine sediments.

Genus *AULISCUS* Ehrenberg, 1843

Auliscus spp.

Remarks. Only a single specimen belonging to this genus was found in the middle Eocene of Site 357.

Genus *BAXTERIOPSIS* Karsten, 1928

Baxteriopsis brunii (Van Heurck) Karsten, 1928. Van Heurck, 1896, p. 460, fig. 190.

Genus *BIDDULPHIA* Gray, 1821

Biddulphia tridens Ehrenberg, 1840. Synonym: *Biddulphia tuomeyi* (Bailey) Roper, 1859, p. 8, pl. 1, figs. 1-2. Hustedt, 1930, pp. 834-836, fig. 491.

Genus *BOGOROVIA* Jousé, 1973

Bogorovia venlamini Jousé, 1973. Jousé, 1973, p. 351, pl. 4, figs. 1-3.

Genus BRIGHTWELLIA Ralfs, in Pritchard, 1861

Brightwellia spp.

Remarks. Only fragments of species belonging to this genus were found in the middle Eocene of Site 357; these did not allow further determination.

Genus *CESTODISCUS* Greville, 1865

Cestodiscus convexus Castracane, 1886. Synonym: **Cestodiscus pulchellus** Greville sensu Jousé, 1973, pp. 345, 348, pl. 1, figs. 12-13. Castracane, 1886, p. 123, pl. 7, fig. 6; Fenner, 1982, p. 86, pl. 13, figs. 1-3.

Cestodiscus gemmifer Castracane, 1886. Synonym: *Cestodiscus* aff. *superbus* sensu Jousé, 1973, pl. 1, fig. 11. Castracane, 1886, p. 124, pl. 2, fig. 7; pl. 7, fig. 7; Fenner (1982), p. 86, pl. 12, figs. 1-4.

Cestodiscus parvula Castracane, 1886. Synonym: *Cestodiscus* sp. 2 Fenner, 1982, p. 89, pl. 14, figs. 3-4. Castracane, 1886, p. 125, pl. 7., fig. 5.

Cestodiscus pulchellus Greville, 1866. (Plate 1, Figs. 1-4.) Synonym: ?*Coscinodiscus superbus* Hardmann (ms) in Rattray, 1889, p. 458. Greville, 1866, p. 123, pl. 11, fig. 5.

Table 4. Diatom species occurrences in Eocene and Oligocene samples from 73, central equatorial Pacific.

Note: For explanation of symbols, see Table 2.

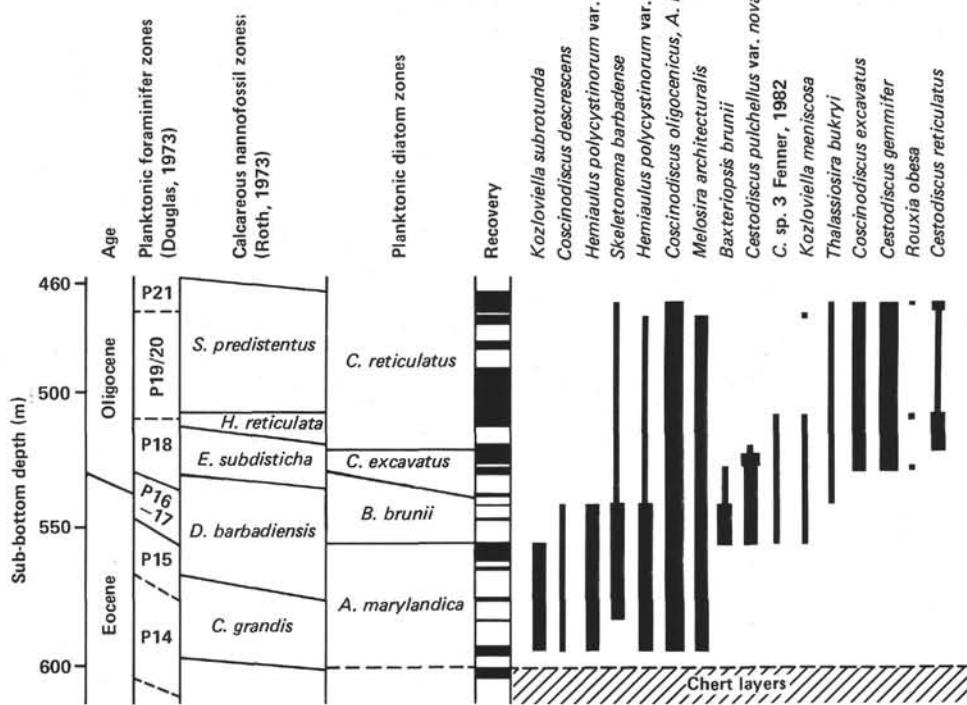


Figure 5. Ranges of stratigraphically important diatom species in middle Eocene to middle Oligocene cores of DSDP Site 167, central equatorial Pacific. Relative abundance as in Figure 3.

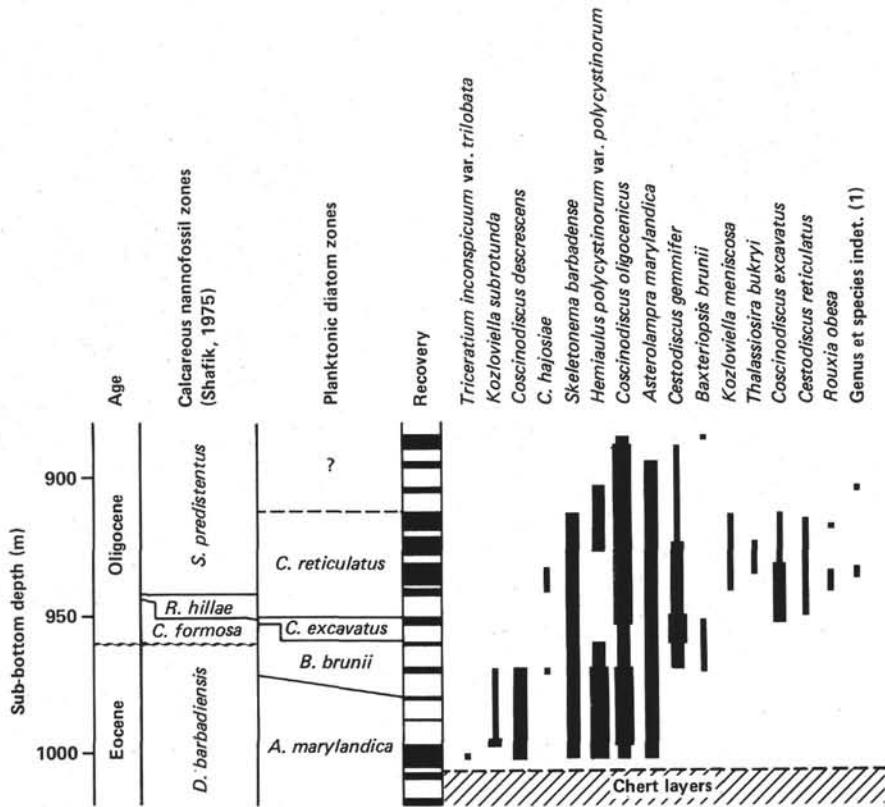


Figure 6. Ranges of stratigraphically important diatom species in late Eocene to early Oligocene cores of DSDP Site 289, Ontong-Java Plateau. Relative abundance as in Figure 3.

Remarks. Greville (1866) gave an accurate description of this species, but later misuse of the species name led to confusion. For a detailed description see Fenner, 1982, p. 86, pl. 15, figs. 1-4. Here the paratypes from slides of Greville's collection are illustrated.

Cestodiscus pulchellus Greville var. *novazealandica* Grove, 1890, in A. Schmidt et al., 1874—. Synonyms: *Coscinodiscus novazealandicus* Hanna, 1932, pp. 182-183, pl. 9, fig. 3; *Cestodiscus spinifer* Grove and Sturt sensu Fenner, 1982, p. 88, pl. 15, fig. 5. A. Schmidt et al. (1874—), pl. 148, fig. 7; pl. 163, figs. 8, 10?; Fenner, 1982, p. 87, pl. 12, figs. 5-7.

Remarks. Very large specimens with valve face slightly concentrically undulated are common around the Eocene/Oligocene boundary. *Cestodiscus reticulatus* Fenner, in press. Synonym: *Coscinodiscus superbus* Hardmann sensu Fenner, 1977, p. 516, pl. 14, figs. 2-3, not figs. 1, 4. Fenner, 1982, p. 87, pl. 14, figs. 1-2.

Cestodiscus stokesianus Greville, 1866. (Plate 1, Figs. 6-8.) Greville, 1866, p. 123, pl. 11, fig. 4; Fenner, 1982, p. 88, pl. 14, figs. 7-8.

Description. The valve outline is circular. The valve face is convex and densely areolated, with the areolae decreasing in size from the center (11 areolae in 10 µm) to the margin (~20 areolae in 10 µm). The areolae are arranged in 4-6 sectors, within which rows of areolae run parallel to the longest row of areolae in the fascicle, which is the middle row of each fascicle. The fascicles are separated from each other by a long radial row of areolae running from the centre of the valve face to the margin, where they end in a labiate process.

Stratigraphic occurrence. This species is found from the late Eocene to the middle Miocene.

Cestodiscus trochus Castracane, 1886. (Plate 1, Fig. 3-4.) Synonym: *Cestodiscus mukhinae* Jousé, 1973, pp. 344-345, pl. 1, figs. 1-5. Castracane, 1886, p. 123, pl. 7, figs. 1, 3; Fenner, 1982, p. 88, pl. 14, fig. 5.

Cestodiscus sp. 1 Fenner, 1982. Synonym: *Cestodiscus* aff. *pulchellus* Jousé, 1973, pl. 1, figs. 9-10, not fig. 11. Fenner, 1982, p. 88, pl. 14, fig. 6.

Cestodiscus sp. 3 Fenner, 1982. Fenner, 1982, p. 89, pl. 11, figs. 3-5.

Genus *CHAETOCEROS* Ehrenberg, 1844

Chaetoceros sp. (1) Fenner, 1982. Fenner, 1982, p. 89, pl. 32, figs. 15-16.

Chaetoceros sp. (3) Fenner, 1982. Fenner, 1982, p. 89, pl. 32, fig. 18. *Chaetoceros* bristles.

Remarks. Under this category undifferentiated types of *Chaetoceros* bristles are listed.

Genus *CLADOGRAMMA* Ehrenberg, 1854

Cladogramma conicum Greville var. *campanulatum* Forti, in Tempère and Peragallo, 1907-1915. Forti, 1913, p. 1563, pl. 3, figs. 2, 4, 7.

Genus *CLAVULARIA* Greville, 1865

Clavularia barbadensis Greville, 1865. Synonyms: *Clavularia catenata*

Brun, 1896, p. 236, pl. 20, figs. 12-13; *Cymatosira* sp. 1 Fenner, 1977, p. 518, pl. 32, figs. 2-5. Greville, 1865a, p. 24, pl. 3, figs. 1-3; Fenner, 1982, p. 90, pl. 28, figs. 9-12.

Genus *COCCONEIS* Ehrenberg, 1838

Cocconeis spp.

Genus *COSCINODISCUS* Ehrenberg, 1838

Coscinodiscus argus Ehrenberg, 1838. Hustedt, 1930, pp. 422-424, fig. 226.

Coscinodiscus asteromphalus Ehrenberg, 1844. Hustedt, 1930, pp. 452-454, fig. 250.

Coscinodiscus demergitus Fenner, 1977. Fenner, 1977, p. 514, pl. 4, figs. 1-3; Fenner, 1982, p. 92.

Coscinodiscus descrevens Grunow, 1878, in A. Schmidt et al., 1874—. A Schmidt et al. (1874—), pl. 61, figs. 7-9; Fenner, 1982, p. 92, pl. 9, fig. 7.

Coscinodiscus elegantulus Greville, 1861. Greville, 1861a, p. 42, pl. 4, fig. 8; Fenner, 1982, p. 92, pl. 9, fig. 4.

Table 5. Diatom species occurrences in Eocene and Oligocene samples from Sites 216 and 217, Ninetyeast Ridge.

Note: For explanation of symbols, see Table 2. *Chaetoceros* sp. in Hole 217 assigned to *Chaetoceros* sp. 1 Fenner, 1982. In addition to the above species, *Macrocystis barbadensis* was identified in Sample 216-16-1, 135–140 cm (F), *M. stella* in Sample 217-9-6, 77–82 cm (F). Genus et species indet. 1 was identified in Sample 217-9-6, 77–82 cm (F); and Gen. et spec. indet. 3 in Samples 217-9-5, 59–64 cm (C) and 217-9-6, 77–82 cm (R).

Table 6. Diatom species occurrences in Eocene samples from Site 220, Arabian Basin.

Eocene	Age	Core-Section (level in cm)										
	middle	1 ?	11 1 60.0	R	<i>Arachnoidiscus</i> spp.							
			12 2 106.0	R	<i>Cestodiscus convexus</i>							
			12 4 117.0	P	<i>Cocconeis</i> spp.							
			13 2 117.0	F	<i>Coscinodiscus</i> spp.							
			13 4 115.0	F	<i>Ethmodiscus</i> spp.							
			14 1 121.0	F	Genus et species indet. 2							
			14 3 121.0	F	<i>Grammatophora</i> spp.							
			15 1 121.0	F	<i>Hemiaulus capitatus</i>							
				R	<i>H. klijushnikovi</i>							
				R	<i>H. longicornis</i> a							
				R	<i>H. polycystinorum</i> var. <i>poly cystinorum</i>							
				C	<i>Listostephania</i> stage of <i>Asterolampra</i> spp.							
				C	<i>Rhizosolenia prokostkajae</i>							
				A	<i>Triceratium inconspicuum</i> var. <i>trilobata</i>							
				F	<i>T. kanayae</i>							
				F	<i>T. sp. 1</i>							
				A	<i>Trinacria subcapitata</i>							

Note: For explanation of symbols, see Table 2. Undifferentiated benthic diatoms were found in Samples 220-11-1, 60-65 cm (C), 220-12-2, 106-111 cm (C), 220-

^a Sample 220-15-1, 131 cm is *H. aff.* /*encinarca*.

Coscinodiscus excavatus Greville, 1861. Synonym: *Coscinodiscus diophthalmus* Castracane, 1886, p. 163, pl. 16, fig. 4. Greville ms., Ralfs in Pritchard, 1861, p. 829, pl. 8, fig. 26; Grunow, 1884, p. 73; Fenner, 1982, p. 93.

Coscinodiscus extravagans A. Schmidt, 1878, in A. Schmidt et al., 1874—. Synonym: *?Tripodiscus beringensis* Mann, 1907, p. 278, pl. 50, fig. 6. A. Schmidt et al., 1874—, pl. 58, fig. 33; Fenner, 1982, p. 93, pl. 16, fig. 6.

Coscinodiscus grossheimii Gleser, in Sheshukova-Poretzkaya and Gleser, 1962. Sheshukova-Poretzkaya and Gleser, 1962, p. 178, pl. 1, figs. 5a–b.

Coscinodiscus hajosiae Fenner, in press. Synonyms: *Podosira aff. maxima* Dzinoridze et al., 1978, pl. 4, figs. 6–7; *Hyalodiscus ambiguus* Grun sensu Paramonova, 1964, pl. 2, fig. 2; *Coscinodiscus spiralis* Karsten, 1905, p. 81, pl. 5, figs. 5a–b. Fenner, in press, pl. 2, fig. 1; Fenner, 1982, p. 96, pl. 8, fig. 3.

Coscinodiscus lewisiensis Greville, 1866. Greville, 1866b, p. 78, pl. 8, figs. 8–10.

Coscinodiscus marginatus Ehrenberg, 1841. Hustedt, 1930, p. 416, fig. 223.

Coscinodiscus nodulifer A. Schmidt, 1878, in A. Schmidt et al., 1874—. Hustedt, 1930, pp. 426–427, fig. 229.

Coscinodiscus obscurus A. Schmidt, 1878, in A. Schmidt et al., 1874—. A. Schmidt et al., 1874—, pl. 61, figs. 16–17.

Coscinodiscus oligocenicus Jousé, 1973. Jousé, 1973, p. 348, pl. 1, figs. 6–8, 16; Fenner, 1977, pp. 515–516, pl. 4, figs. 5–10; Fenner, 1982, p. 95, pl. 9, fig. 8.

Coscinodiscus payeri Grunow, 1884. Grunow, 1884, p. 80, pl. 3(C), figs. 12–13.

Coscinodiscus radiatus Ehrenberg, 1839. Hustedt, 1930, pp. 420–421, fig. 225.

Coscinodiscus rhombicus Castracane, 1886. Synonyms: *Coscinodiscus lanceolatus* Castracane, 1886, p. 164, pl. 17, fig. 19; *Coscinodiscus? naviculoides* Truan y Luard and Witt, 1888, p. 14, pl. 2, fig. 10; *Coscinodiscus punctatus* Ehrenberg var. *rhombica* Ratnay, 1889, p. 547. Castracane, 1886, p. 164, pl. 22, fig. 11.

Coscinodiscus sellatus Jousé, 1973. Jousé, 1973, pp. 348–349, pl. 2, figs. 2–4.

Remarks. Although I consider this “species” to be a variety of *C. excavatus*, it was treated separately to investigate possible differences in its stratigraphic range.

Coscinodiscus senarius var. *parvula* A. Schmidt, in Tempère and Peragallo, 1912. Synonym: *Coscinodiscus senarius* A. Schmidt sensu Dzinoridze et al., 1976, pp. 42–43, figs. 57–61. Tempère and Peragallo, 1912, p. 344, no. 701–726; Fenner, 1982, p. 96, pl. 5, fig. 3.

Coscinodiscus symbolophorus Group. Fenner, 1982, p. 96, pl. 4, fig. 2.

Coscinodiscus tabularis Grunow, 1884. Synonyms: *Coscinodiscus endoi* Kanaya, 1959, pp. 76–77, pl. 3, figs. 8–11; *Coscinodiscus nodulifer* sensu McCollum, 1975, p. 527, pl. 5, figs. 7–8. Hustedt, 1958, pp. 119–120, figs. 48–56; Fenner et al., 1976, p. 774, pl. 7, figs. 10–13; Fenner, 1977, p. 516.

Coscinodiscus tuberculatus Greville, 1861. Greville, 1861a, p. 42, pl. 4, fig. 6; Fenner, 1982, p. 96, pl. 9, figs. 1–3.

Coscinodiscus vigilans A. Schmidt, 1888, in A. Schmidt et al., 1874—. A. Schmidt et al., 1874—, pl. 114, fig. 11.

Genus CRASPEDODISCUS Ehrenberg, 1844

Craspedodiscus elegans Ehrenberg, 1844. Synonym: *Craspedodiscus coscinodiscus* Ehrenberg, 1844, p. 266. Ehrenberg, 1844, p. 266, fig. 12; Fenner, 1982, p. 97.

Craspedodiscus oblongus (Greville) A. Schmidt, 1886, in A. Schmidt et al., 1874—. Synonym: *Porodiscus oblongatus* Greville, 1863b, p. 65, pl. 4, fig. 5. Greville, 1866a, p. 4, pl. 1, figs. 9–10.

Genus CYMATOSIRA Grunow, 1862

Cymatosira aff. cornuta Schrader and Fenner, 1976. Schrader and Fenner, 1976, p. 976, pl. 8, fig. 6; pl. 22, fig. 14; Fenner, 1982, p. 99, pl. 31, fig. 8.

Genus DENTICULOPSIS Simonsen, 1979

Denticulopsis hustedtii (Simonsen and Kanaya) Simonsen, 1979. Synonym: *Denticula hustedtii* Simonsen and Kanaya, 1961, p. 501, pl.

1, figs. 19–25; pl. 2, figs. 36–47. Simonsen and Kanaya, 1961, p. 501, pl. 1, figs. 19–25; pl. 2, figs. 36–47.

Genus DIPLONEIS Ehrenberg, 1840

Diploneis spp.

Remarks. Species of this genus are of shallow-water origin.

Genus ENDICTYA Ehrenberg, 1845

Endictya robustus (Greville) Hanna and Grant, 1926. Hanna and Grant, 1926, p. 144, pl. 16, figs. 2–3.

Genus ETHMODISCUS Castracane, 1886

Ethmodiscus spp.

Remarks. Only fragments of specimens interpreted as belonging to the genus *Ethmodiscus* have been found.

Genus et species indet. 1. (Plate 2, Fig. 8.)

Description. The valve outline is quadrangular, with robust horns at each corner. The valve length is 50–55 μm . One horn is always thinner and generally broken, the other three are of equal width (8–10 μm) from base to top and end in a stout, triangular spine. Thin marginal ridges run along the valve margin and connect the horns. The valve face, mantle, and horns are equally coarsely areolated (4–5 areolae in 10 μm).

Stratigraphic occurrence. This species is found in the middle Oligocene (*S. distentus* Zone) of Sites 292 and 289.

Genus et species indet. 2. (Plate 2, Fig. 5.)

Description. The valve outline is circular. The central part of the valve face is depressed and sharply offset from the outer part of the valve face by a ridge. The areolae are arranged in radial rows. Between the areolae on the outer and inner part of the valve face small nodules are common. The areolation of the inner part is finer (5 areolae in 10 μm) than on the outer part of the valve face (2–3 areolae in 10 μm).

Stratigraphic occurrence. This species was found in the middle Eocene of Site 220.

Genus GRAMMATOPHORA Ehrenberg, 1839

Grammatophora spp.

Remarks. Intercalary bands belonging to species of this genus were found sporadically. These intercalary bands are especially dissolution-resistant.

Genus HEMIAULUS Ehrenberg, 1844

Hemiaulus affinis Grunow, in Van Heurck, 1883. Van Heurck, 1883, pl. 106, figs. 10–11.

Hemiaulus alatus Greville, 1865. Greville, 1865b, p. 31, pl. 3, fig. 14; Fenner, 1977, p. 529, pl. 26, figs. 1–2.

Hemiaulus altar Brun, 1896. Brun, 1896, p. 238, pl. 20, fig. 19–20.

Remarks. In cores with poor preservation, so that only broken arms but no complete valves could be found, differentiation between *H. altar* and *H. longicornis* was not possible.

Hemiaulus barbadiensis Grunow, 1884. Grunow, 1884, p. 63; Fenner, 1982, p. 103, pl. 23, fig. 4.

Hemiaulus capitatus Greville, 1865. Greville, 1865, p. 54, pl. 6, fig. 24.

Hemiaulus dubius Grunow, 1884. Synonym: *Hemiaulus biharensis*

Pantocsek, 1892, p. 57, pl. 38, figs. 528–529. Grunow, 1884, p. 61, pl. 5 (E), fig. 54; Fenner, 1982, pp. 103, 104, pl. 26, figs. 1–5.

Hemiaulus exiguis Greville, 1865. Greville, 1865a, p. 29, pl. 4, fig. 20.

Hemiaulus incisus? Hajós, 1976. Hajós, 1976, p. 829, pl. 23, figs. 4–9.

Remarks. Only fragments—parts of horns—were found. Thus the assignments of these fragments to this species stands on very weak grounds.

Hemiaulus kljushnikovii Gleser, in Sheshukova-Poretzkaya and Gleser, 1964. Sheshukova-Poretzkaya and Gleser, 1964, p. 87, pl. 3, fig. 8; Fenner, 1977, p. 521, pl. 25, figs. 12, 14–15.

Hemiaulus longicornis Greville, 1865. Synonym: *Riedelia longicornis* (Greville) Schrader and Fenner, 1976, p. 997. Greville, 1865a, p. 31, pl. 3, fig. 13.

Hemiaulus aff. longicornis Greville, 1865

Remarks. This species differs from *H. longicornis* by its smaller size, relatively wider horns, and less dense areolation. It is found in the lower Eocene and lower middle Eocene at Sites 149 and 220.

Hemiaulus lyriformis Greville, 1865. Synonym: *Riedelia lyriformis* (Greville) Schrader and Fenner, 1976, p. 997. Greville, 1865a, p. 30, pl. 3, fig. 11.

Hemiaulus polycystinorum Ehrenberg var. *polycystinorum* Grunow, 1884. Grunow, 1884, p. 65; Cleve-Euler, 1951, p. 125; A. Schmidt et al., 1874—, pl. 143, figs. 24?–28; Fenner, 1982, p. 106, pl. 20, figs. 2–5.

Hemiaulus polycystinorum Ehrenberg var. *dissona* Fenner, 1977. Fenner, 1977, p. 521, pl. 22, figs. 14–16.

Hemiaulus polycystinorum Ehrenberg var. *mesolepta* Grunow, 1884. Grunow, 1884, p. 65, pl. 2 (B), fig. 43.

Hemiaulus subacutus Grunow, 1884. Grunow, 1884, p. 61, pl. 5 (E), fig. 55; Fenner, 1982, p. 107, pl. 18, figs. 2–10.

Hemiaulus taurus Gombos, in Gombos and Ciesielski, 1983. Gombos and Ciesielski, 1983, pl. 19, figs. 1–8.

Hemiaulus sp. 5 Fenner, 1982. Fenner, 1982, p. 108, pl. 23, fig. 5.

Genus *HEMIDISCUS* Wallich, 1860

Hemidiscus cuneiformis Wallich, 1860. Hustedt, 1930, p. 904, fig. 542; Fenner, 1977, p. 523, pl. 1, fig. 1.

Genus *HYALODISCUS* Ehrenberg, 1845

Hyalodiscus spp.

Remarks. Specimens belonging in this genus were found extremely rarely, and there was no attempt to determine the species.

Genus *KOZLOVIELLA* Jousé, 1973

Kozloviella meniscosa Fenner, 1982. Fenner, 1982, p. 112, pl. 17, figs. 3–4.

Kozloviella subrotunda Fenner, in press. Fenner, 1982, pp. 112–113, pl. 17, figs. 1–2.

Liosstephania stage of *Asterolampra* spp. Hanna and Brigger, 1970, pp. 89–100.

Genus *LISITZINIA* Jousé, 1978

Lisitzinia ornata Jousé, 1978. Synonym: *Triceratium* aff. *zonulatum* (Greville), Sheshukova-Poretskaya et al., 1967, pl. 37, fig. 2; *Triceratium cruciforme* (A.S.) sensu Schrader and Fenner, 1976, p. 1002, pl. 27, fig. 2. Jouse, 1978, pp. 47–48, pl. 10, figs. 1–6.

Genus *MELOSIRA* Agardh, 1824

Melosira architecturalis Brun, 1892. Synonym: *Cyclotella hannaë* Kanna, 1957, pp. 82–84, pl. 3, figs. 10–14. Brun, in Schmidt et al., 1874—, pl. 177, fig. 45–50.

Genus *NAVICULA* Bory, 1822

Navicula spp.

Remarks. No species were determined for the single specimens belonging to this genus.

Genus *NITZSCHIA* Hassall, 1845

Nitzschia jouseae Burckle, 1972. Burckle, 1972, p. 240, pl. 2, figs. 17–21.

Nitzschia marina Grunow, in Cleve and Grunow, 1880. Cleve and Grunow, 1880, p. 70.

Nitzschia porteri Frenguelli, 1949. Frenguelli, 1949, p. 116, pl. 1, figs. 33–34.

Genus *ODONTELLA* Agardh, 1832

Odontella hastata (Greville) Fenner, 1982. Greville, 1865a, p. 31, pl. 3, fig. 15; Fenner, 1982, p. 115, pl. 23, figs. 6–7.

Genus *PARALLIA* Heilberg, 1863

Paralia sulcata (Ehrenberg) Cleve, 1873. Synonym: *Melosira sulcata* (Ehrenberg) Kützing, 1844, p. 55. Hustedt, 1930, pp. 276–278, figs. 118–120.

Genus *PSEUDOEUNOTIA* Grunow, in Van Heurck, 1880

Pseudoeunotia doliolus (Wallich) Grunow, in Van Heurck, 1880. Hustedt, 1959, pp. 258–260, fig. 737.

Genus *PSEUDOPODOSIRA* Jousé, in Proshkina-Lavrenko, 1949

Pseudopodosira spp.

Remarks. No determination to the species level was possible in the few, poorly preserved specimens found.

Genus *PYXILLA* Greville, 1865

Pyxilla caput-avis Brun, 1896. Brun, 1896, p. 242, pl. 19, fig. 11.

Pyxilla gracilis Tempère and Forti, in Forti, 1909. Synonym: *Pyrgopyxis gracilis* (Tempère and Forti) Hendey, 1969, p. 5. Forti, 1909, p. 26, pl. 2, fig. 5.

Pyxilla reticulata Grove and Sturt, 1887. Synonyms: *Pyxilla gracilis* var. *saratoviana* Tempère and Forti, in Forti, 1909, pl. 2, figs. 7–8; not *Hemiaulus ?sarativianus* Pantocsek, 1889, pl. 13, fig. 235; *Pyxilla johnsoniana* Forti, 1909, pl. 2, fig. 2; not *P. johnsoniana* Greville, 1865a, p. 2, pl. 1, fig. 6; *P. johnsoniana* Greville var. *corniculum* Brun, 1893–1896, p. 243, pl. 19, figs. 12–13; *P. prolongata* Brun, 1893, p. 176, pl. 24, fig. 7; *Pyrgopyxis prolongata* (Brun) Hendey, 1969, p. 5. Fenner, 1982, p. 119.

Genus *RHAPHONEIS* Ehrenberg, 1844

Rhaphoneis amphiceros Ehrenberg, 1844. Andrews, 1975, pp. 204–205, pl. 1, figs. 9–12.

Genus *RHIZOSOLENIA* Ehrenberg, 1841

Rhizosolenia hebetata Group. Grunow, 1884, p. 44, 96, pl. 5, figs. 48–50.

Rhizosolenia interposita Hajós, 1976. Synonym: *Rhizosolenia praebarboi* Schrader sensu Schrader and Fenner, 1976, p. 997, pl. 7, fig. 10, not pl. 5, fig. 16. Hajós, 1976, p. 827, pl. 21, fig. 8.

Rhizosolenia prokovskajae (Jousé) Strelnikova, 1974. Strelnikova, 1974, p. 80, pl. 28, figs. 1–9.

Rhizosolenia paebergonii Muchina var. *robusta* Burckle and Trainer, 1979. Burckle and Trainer, 1979, Appendix, pl. 1, figs. 1–7.

Rhizosolenia sp. 1 Fenner, 1982. Fenner, 1982, p. 122, pl. 3, figs. 10–12.

Genus *RIEDELIA* Jousé and Sheshukova-Poretskaya, 1971

Riedelia pacifica Jousé, in Jousé and Sheshukova-Poretskaya, 1971. Jousé and Sheshukova-Poretskaya, 1971, p. 22, pl. 2, figs. 3–4; Gleser and Jousé, 1974, p. 60, pl. 3, fig. 11.

Genus *ROCELLA* Hanna, 1930

Rocella gelida (Mann) Bukry, 1978. Synonyms: *Stictodiscus gelidus* Mann, 1907, p. 268, pl. 50, fig. 5; *Rocella gemma* Hanna, 1930, in part, p. 415, pl. 40, figs. 1–2, 4–9. Bukry, 1978, p. 788, pl. 5, figs. 1–13.

Rocella princeps (Jousé) Fenner, 1982. Jousé, 1973, p. 350, pl. 3, figs. 8–12; Fenner, 1982, p. 123.

Rocella schraderi Bukry, 1978. Bukry, 1978, p. 788, pl. 6; figs. 1–10, pl. 7, fig. 1.

Rocella vigilans Fenner, in press. Fenner, 1982, p. 123, pl. 4, figs. 6–7; pl. 6, fig. 8.

Genus *ROSSIELLA* Desikachary and Maheshwari, 1958

Rossiella paleacea (Grunow) Desikachary and Maheshwari, 1958. Synonyms: *Stoschia(?) paleacea* Grunow, in Van Heurck, 1883, pl. 128, fig. 6; *Coscinodiscus paleaceus* (Grunow) Rattray, 1889, p. 597; *Cussia paleacea* (Grunow) Schrader, 1974, p. 543, pl. 1, figs. 11–14. Desikachary and Maheshwari, 1958, p. 28, fig. 1; Fenner, 1982, p. 124, pl. 31, figs. 1–2.

Rossiella symmetrica Fenner, in press. Synonyms: *Bogorovia veniana* Jousé sensu Gombos, 1976, p. 593, pl. 1, figs. 6–7, pl. 12, figs. 1, 2, 4; *Rossiella* sp. Gombos and Ciesielski, 1983, pl. 24, figs. 1–2. Fenner, in press, pl. 1, figs. 2–4; Fenner, 1982, p. 124, pl. 31, figs. 3–7 (as *Rossiella* sp. A Gombos (in prep.).

Genus *ROUXIA* Brun and Héribaud, in Héribaud, 1893

Rouxia hannaë Jousé, 1973. Jousé, 1973, pp. 349–350, pl. 2, fig. 13.

Rouxia obesa Schrader, in Schrader and Fenner, 1976. Schrader and Fenner, 1976, p. 997, pl. 24, figs. 5–6.

Genus RUTILARIA Greville, 1863*Rutilaria* sp. (Plate 2, Fig. 6.)

Remarks. Only the specimen illustrated on Plate 2 was found. More observations are necessary to decide whether its asymmetrical shape is an abnormality or of taxonomic value.

Genus SCEPTRONEIS Ehrenberg, 1844

Sceptroneis pesplanus Fenner and Schrader, in Schrader and Fenner, 1976. Schrader and Fenner, 1976, p. 998, pl. 22, figs. 30–31, pl. 25, figs. 10–11.

Sceptroneis pupa Schrader and Fenner, 1976. Schrader and Fenner, 1976, p. 999, pl. 22, figs. 17–21, pl. 24, figs. 11–13.

Genus SKELETONEMA Greville, 1865

Skeletonema barbadense Greville, 1865. Greville, 1865c, p. 43, pl. 5, fig. 1.

Skeletonema utriculosum Brun, 1891. Brun, 1891, p. 44, pl. 11, fig. 1a–b.

Genus STEPHANOPYXIS Ehrenberg, 1844

Stephanopyxis aculeata (Ehrenberg) Grunow var. Fenner, 1982, p. 128, pl. 1, figs. 1–4.

Stephanopyxis grunowii Grove and Sturt, 1888, in A. Schmidt et al., 1874—. A. Schmidt et al., 1874—, pl. 130, figs. 1–6.

Stephanopyxis minuta Greville, 1865. Synonyms: *Creswellia minuta* Greville, 1865a, p. 4, pl. 1, fig. 13, not *Stephanopyxis minuta* (Grunow) A. Cleve, in Cleve-Euler, 1941, pl. 1, fig. 12f. Greville, 1865a, p. 4, pl. 1, fig. 13.

Stephanopyxis superba (Greville) Grunow, 1884. Greville, 1861d, p. 68, pl. 8, figs. 3–5.

Stephanopyxis turris (Greville and Arnott) Ralfs, in Pritchard, 1861. Hustedt, 1930, p. 304, fig. 140; Grunow, 1884, p. 87.

Stephanopyxis turris (Greville and Arnott) Ralfs, in Pritchard var. *cylindrus* Grunow, 1884. Grunow, 1884, p. 87, pl. 5(E), fig. 7–14.

Genus STICTODISCUS Greville, 1861*Stictodiscus* spp.

Remarks. Single specimens belonging to this genus were found only very sporadically, and species-level determinations were not attempted. Species of this genus seem to be predominantly restricted to the littoral zone.

Genus SYNEDRA Ehrenberg, 1830

Synedra jouseana Sheshukova-Poretskaya, 1962. Sheshukova-Poretskaya, 1962, p. 208, fig. 4; Schrader, 1973, p. 710, pl. 23, figs. 21–23, 25, 38.

Synedra aff. *miocenica* Schrader, 1976. Schrader, 1976, p. 94, pl. 1, fig. 1.

Remarks. This species differs from the species described by Schrader (1976) in that it does not have the narrowed ends. In the shorter specimens the sides are not parallel, but the valve outline is narrowly elliptical.

Genus THALASSIONEMA Grunow, in Van Heurck, 1881

Thalassionema nitzschioides Grunow, in Van Heurck, 1881. Hustedt, 1959, pp. 244–246, fig. 725; Hasle and Mendiola, 1967, p. 111, figs. 5, 27–34, 39–44.

Thalassionema nitzschioides Grunow var. *parva* Heiden, in Heiden and Kolbe, 1928. Heiden and Kolbe, 1928, p. 564, fig. 118.

Genus THALASSIOSIRA Cleve, 1873

Thalassiosira bukryi Barron, 1983. Synonyms: *Coscinodiscus* sp. 1 Fenner, 1977, p. 517, pl. 6, figs. 7–8; *Coscinodiscus* sp. 5 Fenner, 1977, p. 517, pl. 6, fig. 6. Barron, 1983, p. 511, pl. 4, figs. 1–2, pl. 6, fig. 9.

Thalassiosira eccentrica (group) (Ehrenberg) Cleve, 1904. Hustedt, 1930, p. 388, fig. 201. Simonsen, 1974, p. 9, pl. 2, figs. 1–3.

Thalassiosira fraga Schrader, in Schrader and Fenner, 1976. Schrader and Fenner, 1976, p. 1001, pl. 16, figs. 9–12.

Thalassiosira leptopus (Grunow) Hasle and Fryxell, 1977. Hasle and Fryxell, 1977, pp. 15–18, pl. 1–4.

Thalassiosira mediaconvexa Schrader, in Schrader and Fenner, 1976.

Synonyms: *Thalassiosira dubiosa* Schrader, in Schrader and Fenner, 1976, p. 1001, pl. 35, fig. 6, not figs. 4–5. *Thalassiosira?* sp. 1 Fenner, 1977, p. 533, pl. 3, figs. 7–8. Schrader and Fenner, 1976, p. 1002, pl. 36, fig. 1.

Genus THALASSIOTHRIX Cleve and Grunow, 1880

Thalassiothrix longissima Cleve and Grunow, 1880. Hasle and Mendiola, 1967, p. 114, fig. 20.

Remarks. As generally only fragments of *T. longissima* and *Thalassionema nitzschioides* (*T. longissima* being heteropolar, *T. nitzschioides* being isopolar) were found, distinction between them was impossible in most cases, and they are listed together.

Genus TRICERATIUM Ehrenberg, 1839

Triceratium americanum Ralfs, in Pritchard, 1861. A. Schmidt et al., 1874—, pl. 76, fig. 27 (pl. 76, fig. 28 is in error, see Boyer, 1900, p. 721).

Triceratium cinnamomeum Greville, 1863. Van Heurck, 1880, pl. 126, fig. 1; Kolbe, 1954, p. 47, pl. 2, fig. 18.

Triceratium inconspicuum Greville, 1861 var. *inconspicuum*. Greville, 1861a, p. 45, pl. 8, fig. 10.

Triceratium inconspicuum Greville var. *trilobata* Fenner, 1977. Synonym: *Triceratium barbadense* Greville sensu Kanaya, 1957, pp. 100–101, pl. 7, figs. 1–4. Fenner, 1982, p. 132, pl. 17, figs. 5–6; pl. 27, figs. 5–6.

Triceratium aff. *zonulatum* Greville, 1865. Fenner, 1982, p. 132, pl. 17, fig. 9.

Triceratium kanayae Fenner, in press. Kanaya (1957), pp. 102–103, pl. 7, figs. 5–7 (as *Triceratium* sp. A); Fenner, in press, pl. 1, figs. 5–6.

Remarks. Three- and four-angled varieties were found to co-occur in samples from DSDP Site 220.

Triceratium kanayae var. *quadriloba* Fenner, in press. Fenner, in press, pl. 2.

Triceratium sp. 1. (Plate 2, Fig. 7.)

Remarks. This species, very similar to *Lisitzinia ornata* but differing in the lack of the deep folds between the central part and the angles, was found in the middle Eocene (P10) of DSDP Site 220.

Genus TRINACRIA Heiberg, 1863

Trinacria excavata Heiberg f. *tetragona* A. Schmidt, 1888, in A. Schmidt et al., 1874—. Illustration: A. Schmidt et al., 1874—, pl. 152, figs. 26–28; Fenner, 1977, pp. 535–536, pl. 27, figs. 9–11.

Trinacria praetenuis (Greville) Grunow, 1884. Synonym: *Triceratium permunitum* Greville, 1864b, p. 89, pl. 13, fig. 18. Greville, 1864b, p. 89, pl. 13, fig. 16; Fenner, 1982, p. 133, pl. 28, figs. 1–2.

Trinacria subcapitata (Greville) Grunow, 1884. Greville, 1863a, p. 244, pl. 10, fig. 20; Grunow, 1884, p. 69; Fenner, 1982, p. 133, pl. 28, figs. 3–8.

Genus TROCHOSIRA Kitton, 1871

Trochosira spinosa Kitton, 1871. Kitton, 1871, p. 170, pl. 14, figs. 6–7; A. Schmidt et al., 1874—, pl. 180, fig. 49; pl. 176, fig. 56.

Genus XANTHIOPYXIS Ehrenberg, 1844

Xanthiopyxis acrolopha Forti, 1912. Forti, 1912, p. 84; Forti, 1913, pp. 15–16, pl. 2, figs. 22, 24, 27–28, 30–37.

Xanthiopyxis oblonga Ehrenberg, 1844. Hanna, 1927, p. 124; Hanna, 1932, p. 226.

Xanthiopyxis ovalis Lohmann, 1938. Synonym: *Liradiscus rugulosus* Forti, 1913, p. 1559, pl. 12, fig. 23. Lohmann, 1938, p. 91, pl. 20, fig. 2; pl. 22, fig. 12; Lohmann, 1974, p. 25, pl. 5, fig. 11.

Genera Incertae Sedis

Genus et Species indet. (1) Fenner, 1982. Illustration: Fenner, 1982, pl. 8, figs. 1–2.

Genus et species indet. (3)

Remarks. Only fragments of this species were found. The valves or skeletons are heavily silicified and consist of a central thickening extending laterally at each end into a narrow prolongation. Both the central part and the prolongations are coarsely areolated, the areolae being arranged in longitudinal rows with ribs between the rows.

Stratigraphic occurrence: This species is found in the late Eocene of DSDP Sites 292 and 217.

Genus MACRORA Hanna, 1932

- Macrora barbadiensis* (Deflandre) Bukry, 1978. Loeblich et al., 1968, p. 207.
Macrora stella (Aspeitia) Hanna, 1932. Hanna, 1932, p. 196, pl. 12, fig. 7.

Genus SPERMATOGONIA Leuduger-Fortmorel, 1892

Spermatogonia spp.

Remarks. As only fragments of species belonging to this genus have been found, no species determination was attempted.

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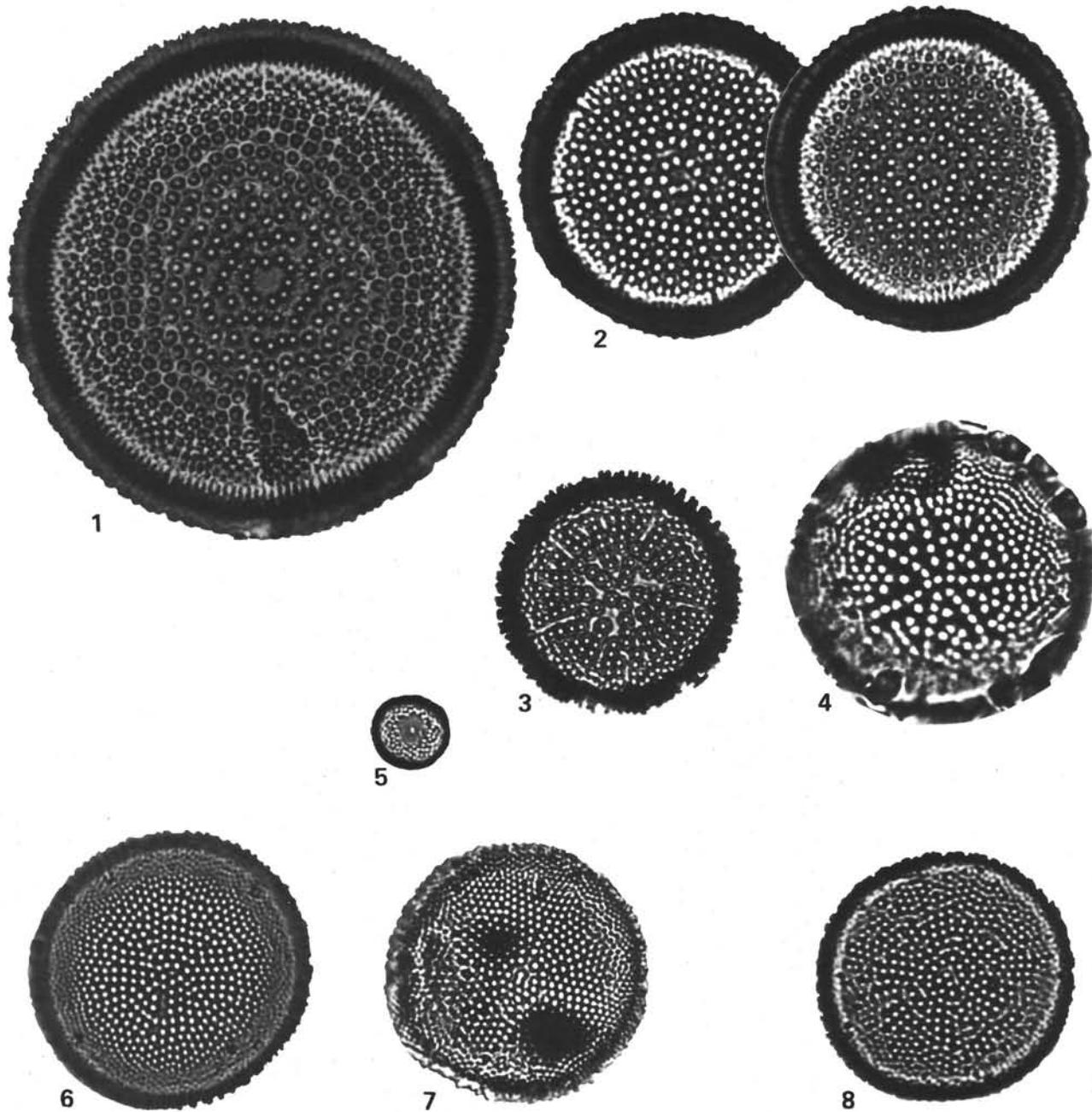


Plate 1. (All figures magnified 1500 \times). 1-2. *Cestodiscus pulchellus* Greville (paratype), Nankoori, Nicobar Islands, Coll. DEBY, B.M. slide 9896. 3. *Cestodiscus parvula* Castracane, DSDP Sample 357-17-6, 96-97 cm. 4. *Cestodiscus* aff. *trochus* Castracane, DSDP Sample 357-15-2, 29-30 cm. 5. *Trochosira?* sp., DSDP Sample 357-24-5, 84-85 cm. 6-8. *Cestodiscus stokesianus* Greville, (6) Paratype, Moron deposit, Coll. Greville (slide obtained from Ref. Stokes), B.M. slide 3235, (7) DSDP Sample 289-97-4, 122-126 cm, (8) Moron deposit, Coll. Tulk, B.M. slide 53516.

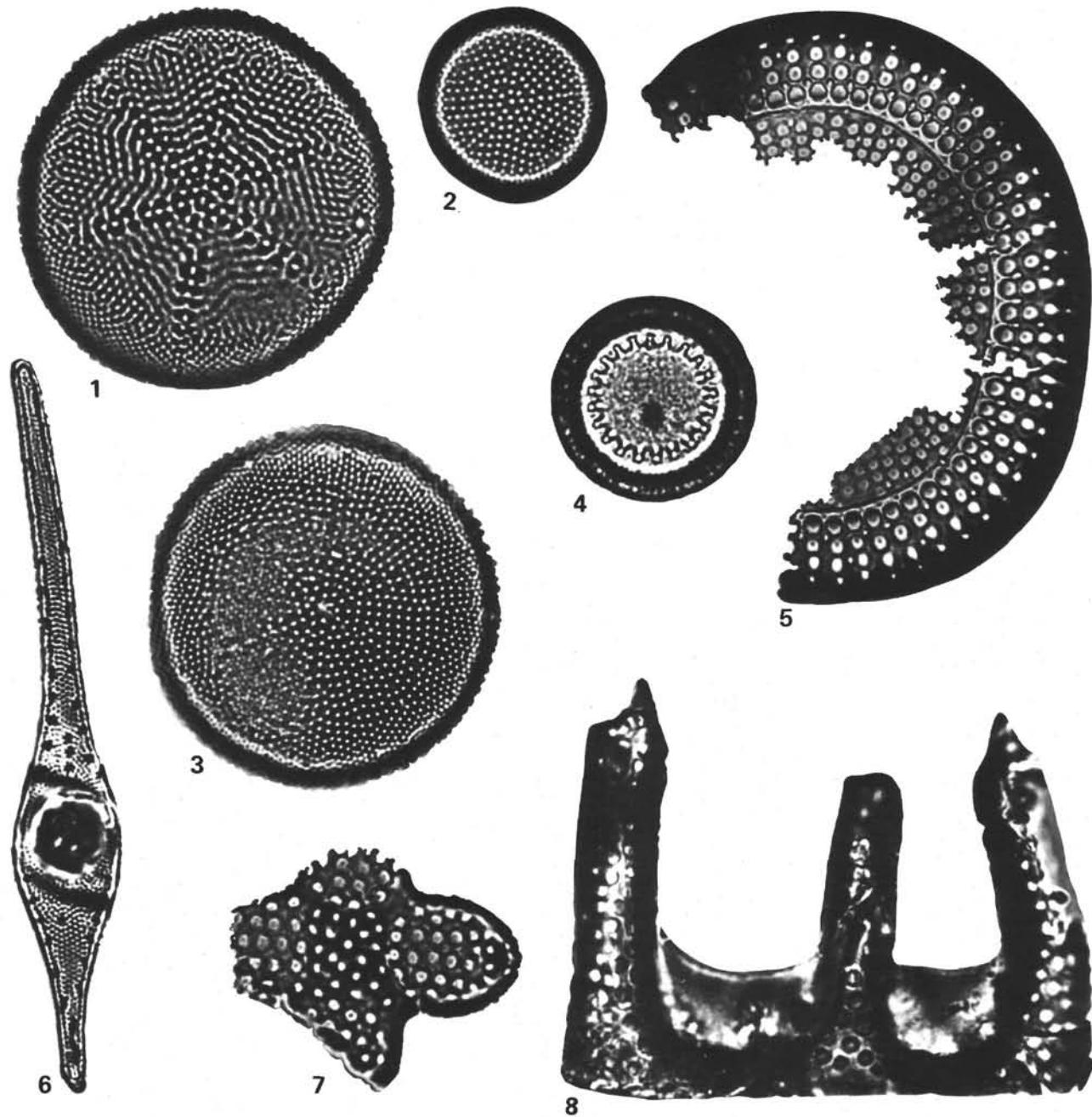


Plate 2. (All figures magnified 1500 \times .) 1-3. *Actinocyclus ehrenbergii* var. *tenella* (Bréb.) Hustedt, (1) Moron deposit, Coll. Greville, B.M. slide 3235, (2) Coll. Greville, B.M. slide 3396, (3) Coll. Tulk, B.M. slide 53516. 4. *Paralia* sp., DSDP Sample 112-9-2, 70-71 cm. 5. Genus et species indet. 2, DSDP Sample 220-14-3, 121-126 cm. 6. *Rutilaria* sp., DSDP 357-24-5, 84-85 cm. 7. *Triceratium* sp. 1, DSDP Sample 220-14-3, 121-126 cm. 8. Genus et species indet. 1, DSDP Sample 289-99-2, 62-67 cm.