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I greatly enjoyed participating in teaching a two week ascidian course at Nagoya University's Sugashima Marine Lab from the end of June to July 10, and then attended the Intl. Tunicata meeting in Aomori, Japan from July 13-17.

This issue is the second for my 40<sup>th</sup> year of compiling Ascidian News. I would greatly appreciate hearing from you whether you still find it useful and interesting. There are **93** New Publications listed at the end of this issue.

### \*Ascidian News is not part of the scientific literature and should not be cited as such.

### NEWS AND VIEWS

**1.** *Ciona intestinalis* now shown to be 2 separate species. Because so many researchers work on *Ciona intestinalis*, and so many papers are published on this species, I draw your attention to 2 new publications showing at last that *Ciona intestinalis* A and B are different species and designating the correct names to be used in all future publications:

Brunetti, R., Gissi, C., Pennati, R., Caicci, F., Gasparini, F. and Manni, L. 2015. Morphological evidence indicates that *Ciona intestinalis* (Tunicata, Ascidiacea) type A and type B are different species. Journal of Zool. Systematics & Evolutionary Research 53 (3): 186–193. [Type A is now designated *C. robusta*; type B retains the name *C. intestinalis*.]

The second new publication describes larval differences between the two species: Pennati, R., Ficetola, G. F., Brunetti, R., Caicci, F., Gasparini, F., Griggio, F., Sato, A., Stach, T., Kaul-Strehlow, S., Gissi, C. and Manni, L. 2015. Morphological differences between larvae of the *Ciona intestinalis* species complex: hints for a valid taxonomic definition of distinct species. PLoS One 10(5): 1-22.

### See the <u>Work in Progress</u> section for an important detailed report of the 8<sup>th</sup> Intl. Tunicata meeting Round Table discussion: Taxonomy of *Ciona* spp. and Suggestions for species designations in publications on *Ciona intestinalis* and/or *Ciona robusta*.

**2.** The **9th Intl. Conference on Marine Bioinvasions** will be held in Sydney, Australia January 19-21, 2016. For more information, go to <u>www.marinebioinvasions.info</u> or contact Dr. Emma Johnston: <u>e.johnston@unsw.edu.au</u>.

**3.** The **8th Intl. Tunicata meeting** was held July 13-17, 2015 in Aomori city, Japan. The full program can be found at <u>http://tunicatemeeting.info/Aomori2015/?q=program</u>. Click on the links for Oral presentations pdf and Poster presentations pdf to view all titles.

4. From Joana Dias (<u>Joana.Dias@fish.wa.gov.au</u>).

I am a researcher at the Western Australia Department of Fisheries working on the population genetics of the marine invasive sea squirt *Didemnum perlucidum*. This species is a tropical/temperate white colonial ascidian and can be found fouling artificial structures in ports, marinas and aquaculture worldwide in warm waters. Its native range is however unknown and finding it would help us better understand its invasive potential. If you work with marine invasive species, particularly in a tropical country, or know someone who does and think you could help finding if it is in your area, please drop me an email. I am happy to send you some pictures.

Thank you, any information will be most appreciated!

Given the widespread low COI diversity of this species, we have developed microsatellite markers to further study its introduction in Australia. The publication has been accepted and is in press:

Dias PJ, Simpson T, Hitchen Y, Lukehurst S, Snow M and Kennington WJ (in press) Isolation and characterization of 17 polymorphic microsatellite loci for the widespread ascidian Didemnum perlucidum (Tunicata, Ascidiacea). Management of Biological Invasions Short Communication.

**5. Anna Di Gregorio** (adg13@nyu.edu) (NYU College of Dentistry) would like to announce that she is celebrating 20 years of publications in ascidian biology (Di Gregorio et al., 1995 to Thompson and Di Gregorio, 2015) and has an upcoming publication from the lab by Diana Jose'-Edwards et al. Congratulations Anna!

6. From Elisabetta Tosti (tosti@szn.it), Stazione Zoologica, Naples.

A new paper by Gallo and Tosti includes the following cartoon she thought would be of interest and amusement to AN readers.

*Ciona intestinalis*: from endangered species to top model

Gallo & Tosti, 2015. (artwork by Fiammetta Formisano) [see New Publications section for the complete citation.]



Elisabetta writes that in March 2016 she will have worked 40 years at the Stazione Zoologica in Naples. Congratulations Elisabetta!

### WORK IN PROGRESS

### 1.8<sup>th</sup> International Tunicata meeting, Aomori, Japan July 13-17, 2015 Round Table Summary: Taxonomy of *Ciona* spp.

At the conclusion of the Tunicata meeting a round table discussion "Taxonomy of *Ciona* spp." was organized by Lucia Manni (University of Padova, Italy) (<u>Imanni@civ.bio.unipd.it</u>) together with Fabio Gasparini (University of Padova, Italy), Carmela Gissi (University of Milan, Italy), Thomas Stach (Humboldt-Universität zu Berlin, Germany) and Ken Hastings (McGill Univ., Canada). The following document was prepared from that discussion. The meeting participants were invited to:

- become aware that *Ciona intestinalis* type A is *Ciona robusta* Hoshino & Tokioka, 1967, and *Ciona intestinalis* type B is *Ciona intestinalis* (Linnaeus, 1767) (see Brunetti et al., 2015; Pennati et al., 2015)
- learn how to distinguish the two species looking at adults and/or larvae;
- verify if information already in public databases belongs to *C.intestinalis* or to *C. robusta*
- discuss the opportunity to update the public tunicate databases with the correct species names.

### Presentation on species nomenclature and diagnostic criteria

### Nomenclature

*Ciona intestinalis* type A is assigned to *Ciona robusta* Hoshino and Tokioka, 1967 because both possess tubercular prominences on the tunic.

*Ciona intestinalis* type B is assigned to *Ciona intestinalis* (Linnaeus, 1767) because:

- 1) *Ciona intestinalis* type B is common on North Atlantic coasts (Suzuki et al. 2005; Caputi et al. 2007; Nydam and Harrison 2007)
- Ciona intestinalis (L.) was first described as Ascidia intestinalis from the Northern European Seas (Linnaeus' "Oceano europaeo" Syst. Nat. 1791. Gmelin edition page 3123)
- 3) *Ciona intestinalis* (L.) is universally recognized to correspond to Millar's description (1953) (referred to animals sampled in British waters).

*Ciona intestinalis sensu* Hoshino and Nishikawa (1985) included both *Ciona intestinalis* type A and type B (see Table 1).

### **Diagnostic criteria**

Researchers were invited to examine adult individuals in order to verify which species they are studying, using a dissection microscope with transmitted and/or reflected light, and isolating the tunic if necessary.

Tubercular prominences on the tunic are papilla-shaped or elongated protuberances. They are distributed along the whole body and often more conspicuous around the siphons, where they prevalently are arranged in longitudinal rows.

Late swimming larva (stage 29 according to the FABA2 database -

http://chordate.bpni.bio.keio.ac.jp/faba2/2.2/top.html - or stage 2 according to Chiba et., 2004; 24 h post-fertilization at 18°C) belonging to the two species can be discriminated. The larvae of *C. intestinalis* have a longer pre-oral lobe, a longer and relatively narrower total body length, and a shorter ocellus-tail distance than larvae of *C. robusta*. Researchers can successfully apply two different discriminant functions based on four or two larval morphometric parameters to discriminate larvae belonging to the two species (Pennati et al., 2015).

### Points arising during round table discussion

The discussion was positive and collaborative although the issue was recognized as a complex one, with different significance for different disciplines

It was stressed that the two taxa are genetically divergent. They display deep molecular divergence (14% of homologous codons encoding identical amino acid residues are distinct synonyms in the two species (Roux et al.2013)). In addition, an extensive population survey showed that only a single F1-hybrid and no backcrossed individual were identified in nature in the only area where the two types are presently known to co-occur (Bouchemousse et al. (2015) and talk by Frederique Viard). While acknowledging that species definition and delimitation is complex because amongst other reasons speciation is a gradual process, these results, combined with significant morphological differences, strongly support the assignment of type A and type B to distinct species following phylogenetic and morphological criteria (Frederique Viard from the Station Biologique de Roscoff; Xavier Turon from CSIC Blanes).

Molecular and developmental biologists attending the meeting were worried that the Tunicate Community may be seen as lacking scientific rigor when they had to admit that the genome of *C. intestinalis* published in 2002 (Dehal et al., 2002) now turned out to be from *C. robusta*. Moreover, they were worried that the split of the "previous" single *C. intestinalis* taxon into two distinct species could produce chaos in the interpretation of the previous literature.

Taxonomists and evolutionary biologists, used to name changes, argued that on the contrary the community could be seen as lax only if it would not follow the latest scientific evidence. Systematics is a dynamic field and species are working hypothesis. Pointing out the 250 years of tradition leading to a set of rules – The International Code of Zoological Nomenclature (ICZN, 4ed) – both parties discovered in their debate that these rules, although formalistic and legalistic, were designed to secure the main common goal: enabling science to progress and restore order in debated issues. Therefore the taxonomically valid name of *C. robusta* has to be used instead of "*C. intestinalis* type A" from now on. This will allow future

scientists to understand which species had been used in the respective molecular or developmental research. In particular, it was advised that the collection site (or resources) and the collection dates of the animals should be stated in the Materials and Methods in all future reports. The community should also consider that the two species are possibly distributed sympatrically even in regions other than the English Channel, and that the distribution of the two species can change during time.

Several proposals were put forward and discussed:

Researchers who deposited their *Ciona* sequences in public databases (GenBank, ENA, Ensembl, JGI Genome Browser, UCSC Genome Browser, etc.) are invited to contact the database managers and to change the species names if they are now seen to be in error. In addition, the community suggested that the *Ciona* species names and taxonomy should be updated in specialized reference databases, such as WoRMS (World Register of Marine Species) (suggestion of Xavier Turon) and possibly also in ANISEED, because it reports not only genomic sequences but also anatomical and gene expression data.

In terms of citations, especially in the imminent future, where there is potential for confusion, several clear and transparent ways of citation are suggested:

- Ciona robusta Hoshino and Tokioka, 1967. Formerly Ciona intestinalis type A (see Brunetti et al., 2015; Pennati et al., 2015) (suggested by Gretchen Lambert from University of Washington Friday Harbor Labs, USA)
- *Ciona robusta* (= former *C. intestinalis* type A *sensu* Nydam and Harrison, 2007) (suggested by Euichi Hirose from University of Ryukyus, Japan)
- Ciona intestinalis (Linnaeus, 1767), according to Brunetti et al. (2015) and Pennati et al. (2015). Formerly Ciona intestinalis type B (suggested by G. Lambert).

Species name	Ciona intestinalis (Linnaeus, 1767)	Ciona robusta Hoshino et Tokioka, 1967	Ciona savignyi Herdman, 1882
Type locality	Oceano Europaeo	Onagawa, Miyagi, Japan	Kobe, Hyogo, Japan
Type specimen	Original type was missing. Neotype is deposited in Natural History Museum in Venice	Syntype specimens were deposited in Seto Marine Biol. Lab., Kyoto Univ.	British Museum of natural Science
Linnaeus 1767	Ciona intestinalis		
Hoshino et Tokioka, 1967	Ciona intestinalis	Ciona robusta	Ciona intestinalis
Hoshina and Nishikawa, 1985	Ciona intestinalis	Ciona intestinalis	Ciona savignyi
Brunetti et al., 2015	Ciona intestinalis	Ciona robusta	Ciona savignyi

Table 1. Summary of changes occurred in the taxonomy of *Ciona* sp. (by E. Hirose).

\*This table does not included synonyms that are invalid at present.

#### Suggestions for species designations in publications on *Ciona intestinalis* and/or *Ciona robusta*

### Overview

The following suggestions regarding Title and Abstract are meant to minimize confusion in the non-specialist scientific reader during the transitional period in which the term "*Ciona robusta*" is first coming into common use. In a few years, when the scientific public has become familiar with the *intestinalis/robusta* split, these suggestions will have lost their importance. However, the suggestions for the Methods may have a more permanent validity.

The suggestions for the Methods section should be considered as "minimal requirements". We suggest to detail as much as possible the methodology used for the morphological and/or the molecular characterization of the specimens, i.e. reporting which genes(s), sequence(s) or restriction fragment(s) were analyzed; which larval characters were measured; if the presence/absence of tubercules was investigated with a dissection microscope with transmitted and/or reflected light; etc.

### Rationale

The rationale is that during the transitional period the new and unfamiliar name *Ciona robusta* will not appear in the Title or Abstract except in the company of the familiar name *Ciona intestinalis*. First mention of *Ciona robusta* in Abstracts will always be linked to "formerly *Ciona intestinalis* type A". For logical coherence and precision, in the Title or Abstract the name *Ciona intestinalis* should be linked to "formerly *Ciona intestinalis* type B". Use of the generic term "*Ciona*" in the Title can in many cases be a useful approach to maintain simplicity when the species complexity is not especially relevant.

### Suggested species designations

Your study of *Ciona intestinalis* and/or *robusta* will likely fall into one of the following 6 classes. A seventh class, at the end, includes studies that also involve an additional *Ciona* species beyond *intestinalis/robusta*, e.g. *Ciona savignyi.* 

# 1) <u>It is probable</u>, based on the species geographic ranges, that all the animals studied were *Ciona intestinalis*

### Title: say "Ciona"

### Abstract: say "Ciona intestinalis (formerly Ciona intestinalis type B)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which only *Ciona intestinalis* is known to occur (range citation). Species-defining morphological or molecular characters were not assessed, but based on current knowledge of distribution we assume that the animals studied were *Ciona intestinalis*.

# 2) <u>It is known</u>, based on morphological or molecular criteria, that all the animals studied were *Ciona intestinalis*

**Title:** say "Ciona" **Abstract:** say "Ciona intestinalis (formerly Ciona intestinalis type B)" **Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which only *Ciona intestinalis* is known to occur (range citation). Species-defining morphological or molecular characters were assessed (name the characters), which confirmed the animals studied were *Ciona intestinalis*.

# 3) <u>It is probable</u>, based on the species geographic ranges, that all the animals studied were *Ciona robusta*.

### Title: say "Ciona"

Abstract: say "Ciona robusta (formerly Ciona intestinalis type A)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which only *Ciona robusta* is known to occur (range citation). Species-defining morphological or molecular characters were not assessed, but based on current knowledge of distribution we assume that the animals studied were *Ciona robusta*.

# 4) <u>It is known</u>, based on morphological or molecular criteria, that all the animals studied were *Ciona robusta*.

### Title: say "Ciona"

Abstract: say "Ciona robusta (formerly Ciona intestinalis type A)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which only *Ciona robusta* is known to occur (range citation). Species-defining morphological or molecular characters were assessed (name the characters), which confirmed the animals studied were *Ciona robusta*.

# 5) <u>It is possible</u>, based on the species geographic ranges, that the animals studied included both *Ciona intestinalis* and *Ciona robusta*.

### Title: say "Ciona"

**Abstract:** say "Ciona intestinalis (formerly Ciona intestinalis type B)" and "Ciona robusta (formerly Ciona intestinalis type A)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which both species are known to occur (range citation). Species-defining morphological or molecular characters were not assessed, but based on current knowledge of distribution we assume that the animals studied may have included both species.

# 6) <u>It is known</u>, based on morphological or molecular criteria, that some of the animals studied were *Ciona intestinalis* and some were *Ciona robusta*.

Within this class of studies there will be two major subclasses.

**6A**. Studies reporting differences between the species, or studies in which possible species differences were looked for and not found.

### Title: say "Ciona intestinalis and "Ciona robusta"

**Abstract:** say "Ciona intestinalis (formerly Ciona intestinalis type B) and "Ciona robusta (formerly Ciona intestinalis type A)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which both species occur (or at distinct sites for each species). Species-defining morphological or molecular characters (name the characters) were assessed.

**6B.** Studies in which it was not an important goal to compare the species, but in which they were both assessed incidentally and no difference was found.

### Title: say "Ciona".

**Abstract:** say "Ciona intestinalis (formerly Ciona intestinalis type B) " and "Ciona robusta (formerly Ciona intestinalis type A)"

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). The animals used in the present study were collected in (collection date) at (name sites) at which both species occur (or at distinct sites for each species). Species-defining morphological or molecular characters (name the characters) were assessed.

[Comment by Carmela Gissi: In my opinion the distinction in two subclasses based on the goal of the study in not necessary: I suggest to use the sentences of case 6A in publications of both subclasses, and to delete in this file all sentences concerning "6B".]

### 7) Studies including an additional *Ciona* species, e.g. *Ciona savignyi*.

**Title:** The suggestion in classes 1 - 5 and 6B to use the generic "*Ciona*" in the title would not be appropriate in class 7. Having to name, e.g., *Ciona savignyi* in the title would force you to supply a species name for your *Ciona intestinalis/robusta* material in the title. In this case say "*Ciona intestinalis* (formerly *Ciona intestinalis* type B)" and/or "*Ciona robusta* (formerly *Ciona intestinalis* type A)", as appropriate.

**Abstract:** say "Ciona intestinalis (formerly Ciona intestinalis type B) and/or "Ciona robusta (formerly Ciona intestinalis type A)" as appropriate.

**Methods (under Animals):** In 2015 it was recognized that *Ciona intestinalis* included two distinct species, *Ciona intestinalis* (formerly *Ciona intestinalis* type B) and *Ciona robusta* (formerly *Ciona intestinalis* type A). Structure the remainder of the Methods along the lines indicated above for one of classes 1 - 6, whichever is most relevant.

**Literature** (\* means reference includes range information, of use in helping to determine which species was actually used)

\*Bouchemousse S, Lévêque L, Dubois G, Viard F (2015) Co-occurrence and reproductive synchrony do not ensure hybridization between an alien tunicate and its interfertile native

congener. Evol Ecol. IN PRESS. First online: 09 August 2015. doi:10.1007/s10682-015-9788-1.

- \*Brunetti R, Gissi C, Pennati R, Caicci F, Gasparini F, Manni L (2015) Morphological evidence that the molecularly determined *Ciona intestinalis* type A and type B are different species: *Ciona robusta* and *Ciona intestinalis*. J Zoolog Syst Evol Res. 53:186-193.
- \*Caputi L, Andreakis N, Mastrototaro F, Cirino P, Vassillo M, Sordino P (2007) Cryptic speciation in a model invertebrate chordate. Proc Natl Acad Sci USA 104:9364-9369.
- Chiba S, Sasaki A, Nakayama A, Takamura K, Satoh N (2004) Development of *Ciona intestinalis* juveniles (through 2nd ascidian stage). Zoolog Sci 21: 285–298.
- Dehal P, Satou Y, Campbell RK, Chapman J, Degnan B, et al. (2002) The draft genome of *Ciona intestinalis*: insights into chordate and vertebrate origins. Science 298:2157-2167.
- \*Hoshino ZI, Tokioka T (1967) An unusually robust *Ciona* from the northeastern coast of Honsyu Island, Japan. Publ Seto Mar Biol Lab 15:275-290.
- ICZN. International Code of Zoological Nomenclature. 4 ed., (The International Trust for Zoological Nomenclature, 1999).
- Linnaeus C (1767) Systema naturae. Editio duodecima. Holmiae, Stockholm.
- Linnaeus C (1791) Systema Naturae. Editio decima tertia ("Gmelin edition"). Lipsiae, Leipzig.
- \*Millar RH (1953) Ciona. Colman, J. S. ed. Liverpool University Press, Liverpool.
- \*Nydam ML, Harrison RG (2007) Genealogical relationships within and among shallow-water *Ciona* species (Ascidiacea). Mar Biol 151:1839-1847.
- \*Nydam ML, Harrison RG (2011) Introgression despite substantial divergence in a broadcast spawning marine invertebrate. Evolution 65: 429-442
- \*Pennati R, Ficetola GF, Brunetti R, Caicci F, Gasparini F, Griggio F, Sato A, Stach T, Kaul-Strehlow S, Gissi C, Manni L (2015) Morphological differences between larvae of the *Ciona intestinalis* species complex: hints for a valid taxonomic definition of distinct species. PLoS ONE 10:e0122879.
- \*Roux C, Tsagkogeorga G, Bierne N, Galtier N (2013) Crossing the species barrier: genomic hotspots of introgression between two highly divergent *Ciona intestinalis* species. Mol Biol Evol 30: 1574-158.
- \*Suzuki MM, Nishikawa T, Bird A (2005) Genomic approaches reveal unexpected genetic divergence within Ciona intestinalis. J Mol Evol 61: 627–635.
- \*Zhan A, Macisaac HJ, Cristescu ME (2010) Invasion genetics of the *Ciona intestinalis* species complex: from regional endemism to global homogeneity. Molecular Ecology 19: 4678-4694.

### 2. From Delphine Dauga (<u>contact@aniseed.cnrs.fr</u>)

After 3 years of refactoring the ANISEED database, its user interfaces, and the data curation system, we are happy to announce the publication in the 2016 Database issue of Nucleic Acid Research the improvement and update of ANISEED. [Brozovic, Martin, Dantec, Dauga et al. NAR 2016 Database issue, In press—see New Publications section].

In this article, we report the development of the system since its initial publication in 2010:

- A new and more adapted database schema
- An improved and enriched formal description of the embryonic development of *Ciona*, *Phallusia*, *Halocynthia* and *Molgula* species, and of budding in *Botryllus*.
- The genomes of nine ascidian species can be explored via dedicated genome browsers, and searched by Blast.

• A full functional gene annotation, anatomical ontologies and some gene expression data for the six species with highest quality genomes are now available.

ANISEED is publicly available at: <u>http://www.aniseed.cnrs.fr</u> .

During the refactoring process we had to stop inserting expression data from published articles, and the last article of the 189 articles currently included in the database was entered in June 2011. Since then, however, many articles dealing with the molecular developmental biology of *Ciona* have been published, a number which vastly exceeds our current biocuration ability. We are therefore looking for volunteers to enter data from their own papers, or from other papers they know well in *Ciona*, but also in *Phallusia*, *Halocynthia* and *Botryllus*.

We are aware that entering data is a time consuming task. Yet, there are many reasons to volunteer:

- At a time when reading (and remembering) all papers in a field is becoming difficult, ANISEED will make your work more visible and more accessible by the community.
- After 3 years of hard work on software improvement, community input in ANISEED is now needed for the database content to be as good as its architecture.
- The curation tools we have developed are nice, logical and friendly to use, even if you are not very keen on computers...
- Community members who have entered a substantial number of experiments will be coauthors of the next ANISEED update paper.

We thank you for your attention and hope to soon welcome some of you among our curation team (please contact Delphine Dauga to create a ANISEED curation account at <u>contact@aniseed.cnrs.fr</u>).

[Ascidian News editor's comment: please specify if your data is for *Ciona intestinalis* or *C. robusta*!]

### **3.** From **Gérard Breton** (<u>gerard-breton@orange.fr</u>), Association Port Vivant, La Havre. **The 2014-2015 new observations on** *Diplosoma listerianum* "balloons" in Le Havre harbor.

In 2014, in the port of Le Havre, the divers of the Association Port Vivant noticed that some ascidians were curiously "ballooned" (see Ascidian news 74). The ascidian, first misidentified as *Didemnum vexillum*, proved, thanks to Françoise Monniot, to be *Diplosoma listerianum*. During summer 2014, the "ballooned" *Diplosoma listerianum* were very abundant in the basins of the port of Le Havre. The phenomenon was not limited to the port of Le Havre, and to 2014! A picture by Thierry Derycke in April 2012 in the port of Le Havre shows a "balloon". Marcos Tatian reports "late August I participated in a campaign on board the vessel Puerto Deseado, along the Argentinean shelf. Surprisingly, the trawls carried a lot of these special formation or "balloons" of *Diplosoma*. They were very big and abundant." Jean-Louis Lenne gives a picture of such "balloons" of *Diplosoma listerianum* from a wreck, - 22 m, North Sea (14/07/2010) and another picture from the port of Boulogne-sur-mer, North Sea (18/10/2014). François-Xavier Huet photographed in the Rance Estuary on 31/10/2015 a typical balloon (Figure 1). Françoise Monniot had observed the phenomenon long ago in the port of La Rochelle.

### The question of the biological meaning of the balloons

In December 2014, Daniel Ingratta from his own pictures (Figure 4), then Gérard Breton from other divers' pictures (Figures 2, 3), remarked that the tunic of ballooned colonies develop groups of short, thin, thread-like expansions (ca 0.3 mm in diameter, several mm in length), better seen on a dark background. Some were attached to the substrate, other ones not. Gretchen Lambert wonders about the meaning of the "balloon" phenomenon and suggests that it could be an original mode of dissemination. The question and its answer are the same from Noa Shenkar and Marcos Tatian. Gretchen adds that the thread-like expansions could be linked to this supposed dispersal strategy, as reattachment structures. But this can be assessed only by an experiment that we tried in January 2015 and in October 2015.

### January and October 2015 experiments

03/01/2015. Dive in the Bassin de la Barre (D.C. and P.A.). The number of *Diplosoma listerianum* – balloons has very much decreased. During one hour, two divers saw and collected only two balloons (1 and 1.3 cm in diameter), plus several which barely began forming vesicles. On the rock-filled side of the basin, *D. listerianum* has thoroughly disappeared.

On 03/01/2015 in the evening, I put in the bottom of a sea water tank with a bubbler some cleaned valves of mussel shells (concavity directed upwards). The two balloons collected by D.C., grey, deflated, were left each in a mussel valve in order to detect their ability to "re-fix" to a substrate. On 04/01/2015, one of the two balloons seemed to begin inflate again, the second one not. The tank was left at the exterior temperature of Le Havre, i.e. 7 to 10 ° C for six days. The two colonies of *D. listerianum* remained grey and deflated and, at the end of the six days, had not at all re-fixed to the support. Even a very soft movement of the mussel shell moved them in relation to the shell.

The scarcity of *D. listerianum* (balloons and ordinary ones), as observed by D.C. and P.A. in a basin in which this species (and its balloon form) was very abundant last fall suggests that its population was declining and in a wintry regression. The two collected balloon colonies were thus seemingly end-of-life colonies, of which the absence of re-fixing to the support is not significant. Thus, a new test was needed.

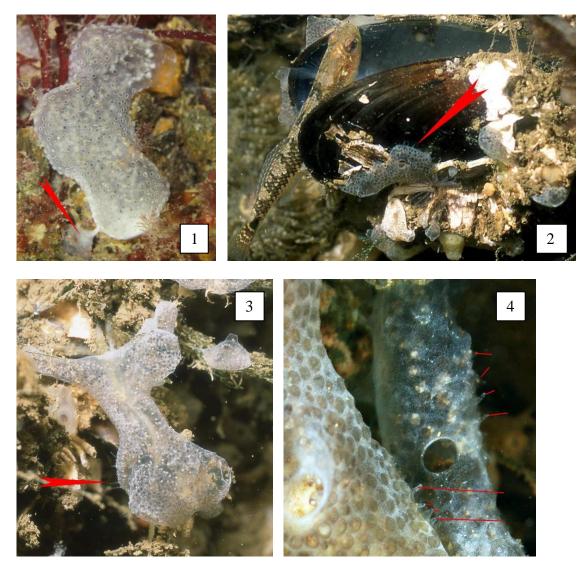
The same experiment was carried on by D.C. between 08 and 17/10/2015, with three balloons collected on 08/10/2015, with an exterior temperature between 10 and 12 °C. At the end of the experiment, the colonies were deflated and in very poor condition, and none was re-fixed to the substrate. The negative results of these trials is not conclusive: the biological signification of the "balloonization" of *Diplosoma listerianum* still remains hypothetical.

Figure 1: Rance Estuary (Brittany), 31/10/2015. © François-Xavier Huet. The arrow indicates a fixation to the substrate by an extension of the tunic.

Figure 2: Port of Le Havre, 07/12/2014. "Ordinary" *Diplosoma listerianum* (i.e. not ballooned), with thread-like extensions attached to a mussel shell. The fish is *Gobiusculus flavescens*. © Paul Leroy.

Figure 3: Port of Le Havre, 06/12/2014. © Marc Lacuisse. The arrow indicates the thread-like extensions of the tunic.

Figure 4: Port of Le Havre, 06/12/2014. © Daniel Ingratta. Enlargement of a close-up picture. The red lines indicate the thread-like extensions of the tunic.



Gérard writes further that *Perophora japonica*, now known from a number of locations in the Atlantic and Pacific, is expanding its population in the port of Le Havre.

### THESIS ABSTRACTS

**Examining genetic diversity and fusion abilities of an invasive colonial ascidian. Darragh Clancy**, MS Thesis (Dec 2015), Romberg Tiburon Center and Dept. of Biology, San Francisco State University, Advisor: C. Sarah Cohen.

<u>Chapter 1:</u> Invasive species cause both ecological and economical problems in their new habitats, either by directly harming or outcompeting native organisms. One such marine invasive species, *D. vexillum*, has recently been found globally and is spreading and altering habitats at an alarming rate. Global diversity of this colonial ascidian has previously been assessed using the barcoding mitochondrial gene, cytochrome c oxidase subunit I (COI), to determine that the native region includes Japan and that four other regions (western North America, eastern North America, Europe and New Zealand) are significantly lower in diversity. However, there have been few comparisons of diversity levels among populations

within a region or uses of higher resolution markers for the species. This study provides such an evaluation with 11 microsatellite markers in addition to COI to determine population structure and diversity of four locations within the western North American region. These locations have differing histories in terms of geographic distances to each other and other locations with *D. vexillum* within the region, boat traffic and aquaculture use. Analyses of the markers used in this study reveal surprising levels of differentiation and suggest that diversity and genetic isolation are affected by more than international shipping and proximity to such ports, and that regional boat traffic and gear movement can be important vectors of *D. vexillum* for both introductions and secondary spread to various points around the world.

Chapter 2: Fusion is a behavior often found in colonial organisms when two conspecific colonies come in contact with one another. However, it presents a puzzling conflict to fused colonies that bear the cost of contributing to shared structures if they are contributing fewer genes to the next generation. Kin selection has been proposed as a possible solution, but it is not evident in all ascidian species. In some species, like the well-studied basal chordate Botryllus schlosseri, fusion is dependent on matching alleles at particular loci, limiting fusion to close relatives, while in other species, like Diplosoma listerianum, there is no specificity when colonies fuse. A previous study inferred that *Didemnum vexillum* may also have genetic specificity for fusion, based on a negative correlation between fusion rate and measured gene diversity. The current study further investigates the specificity of D. *vexillum* by testing whether fusion behavior increases with increased genetic relatedness at three sites within the western North America region. Using microsatellite markers, highresolution multi-locus genotypes of paired fusion assays were assessed. Relatedness values of colony pairs that fused were compared to relatedness values of colony pairs that did not fuse. There is a trend of higher genetic similarity correlating to higher likelihood of fusion. Weak correlation may be due to the use of neutral loci rather than a specific fusion gene, or to a decreased cost associated with fusion compared to Botryllus schlosseri. D. vexillum fusion involves tissue sharing, rather than vascular system integration found in botryllid ascidians. In Botryllus schlosseri, high specificity of fusion is limited to close relatives in theory to reduce the possibility of cell parasitism that occurs when the colonies' vascular systems fuse. This study provides further evidence that *D. vexillum* may also restrict fusion to genetically similar colonies in order to mitigate the inferred costs of fusing with unrelated colonies.

### **NEW PUBLICATIONS**

- Abitua, P. B., Gainous, T. B., Kaczmarczyk, A. N., Winchell, C. J., Hudson, C., Kamata, K., Nakagawa, M., Tsuda, M., Kusakabe, T. G. and Levine, M. 2015. The pre-vertebrate origins of neurogenic placodes. Nature **524**: 462-465.
- Ballarin, L., Du Pasquier, L., Rinkevich, B. and Kurtz, J. 2015. Evolutionary aspects of allorecognition. Invert. Survival J. **12**: 233-236.
- Berná, L. and Alvarez-Valin, F. 2015. Evolutionary volatile cysteines and protein disorder in the fast evolving tunicate *Oikopleura dioica*. Mar. Genomics **epub**:

Bonnet, N. Y. K. and Lotufo, T. M. C. 2015. Description of *Ascidia paulayi* sp. nov. (Phlebobranchia: Ascidiidae) from French Polynesia, with a discussion about the *Ascidia sydneiensis* Stimpson, 1855 group. Zootaxa **3994**: 283–291.

 Bouquet, J. M., Spriet, E., Troedsson, C., Otterå, H., Chourrout, D. and Thompson, E. M.
 2009. Culture optimization for the emerging zooplanktonic model organism *Oikopleura dioica*. J. Plankton Res. **31**: 359-370.

- Brozovic, M., Martin, C., Dantec, C., Dauga, D., Mendez, M. and et.al. 2015. ANISEED 2015: a digital framework for the comparative developmental biology of ascidians. Nucleic Acids Res. **epub**: 1-11.
- Cardenas, C. A. and Montiel, A. 2015. The influence of depth and substrate inclination on sessile assemblages in subantarctic rocky reefs (Magellan region). Polar Biol. **38**: 1631-1644.
- Choi, N.-D., Zeng, J., Choi, B.-D. and Ryu, H.-S. 2014. Shelf life of bottled sea squirt *Halocynthia roretzi* meat packed in vegetable oil (BSMO). Fish. Aquat. Sci. **17**: 37-46.
- Colautti, R. I., Bailey, S. A., van Overdijk, C. D. A., Amundsen, K. and MacIsaac, H. J. 2006. Characterised and projected costs of nonindigenous species in Canada. Biol. Invasions 8: 45-59.
- Cota, C. D. and Davidson, B. 2015. Mitotic membrane turnover coordinates differential induction of the heart progenitor lineage. Dev. Cell **34**: 505–519.
- Crean, A. J. and Marshall, D. J. 2015. Eggs with larger accessory structures are more likely to be fertilized in both low and high sperm concentrations in *Styela plicata* (Ascidiaceae). Mar. Biol. **162**: 2251-2256.
- Davidson, J. D. P., Landry, T., Johnson, G. R., Ramsay, A. and Quijón, P. A. 2015. A field trial to determine the optimal treatment regime for *Ciona intestinalis* on mussel socks. Manag. Biolog. Invasions 6: in press.
- Dias, P. J., Simpson, T., Hitchen, Y., Lukehurst, S., Snow, M. and Kennington, W. J. 2016. Isolation and characterization of 17 polymorphic microsatellite loci for the widespread ascidian *Didemnum perlucidum* (Tunicata, Ascidiacea). Manag. Biolog. Invasions **7**: 1-3.
- Evans, C. G. 2015. Trabectedin: of sea squirts and the deep blue sea. Targeted Oncol. epub:
- Farley, E. K., Olson, K. M., Zhang, W., Brandt, A. J., Rokhsar, D. S. and Levine, M. S. 2015. Suboptimization of developmental enhancers. Science **350**: 325-328.
- Gallo, A. and Tosti, E. 2015. The ascidian *Ciona Intestinalis* as model organism for ecotoxicological bioassays. J. Marine Sci. Res. Dev. **5**: 1-2.
- Gasparini, F., Skobo, T., Benato, F., Gioacchini, G., Voskoboynik, A., Carnevali, O., Manni, L. and Valle, L. D. 2015. Characterization of Ambra1 in asexual cycle of a non-vertebrate chordate, the colonial tunicate *Botryllus schlosseri*, and phylogenetic analysis of the protein group in Bilateria. Molec. Phylogen. & Evol. **epub**:
- Gillum, J. E., Jimenez, L., White, D. J., Goldstien, S. J. and Gemmell, N. J. 2014. Development and application of a quantitative real-time PCR assay for the globally invasive tunicate *Styela clava*. Manag. Biolog. Invasions **5**: 133–142.
- Gupta, R. S. 2016. Molecular signatures that are distinctive characteristics of the vertebrates and chordates and supporting a grouping of vertebrates with the tunicates. Molec. Phylogen. & Evol. 94: 383-391.
- Hamada, M., Goricki, S., Byerly, M. S., Satoh, N. and Jeffery, W. R. 2015. Evolution of the chordate regeneration blastema: Differential gene expression and conserved role of notch signaling during siphon regeneration in the ascidian *Ciona*. Dev. Biol. **405**: 304-315.
- Harunari, E., Hamada, M., Shibata, C., Tamura, T., Komaki, H., Imada, C. and Igarashi, Y.
  2015. *Streptomyces hyaluromycini* sp. nov., isolated from a tunicate (*Molgula manhattensis*). J. Antibiot. (Tokyo) epub:
- Heenan, P., Zondag, L. and Wilson, M. J. 2016. Evolution of the Sox gene family within the chordate phylum. Gene **575**: 385-392.
- Heering, J., Jonker, H. R., Lohr, F., Schwalbe, H. and Dotsch, V. 2015. Structural investigations of the p53/p73 homologs from the tunicate species *Ciona intestinalis* reveal the sequence requirements for the formation of a tetramerization domain. Protein Sci. **epub**:

- Hopwood, N. 2015. The cult of amphioxus in German Darwinism; or, our gelatinous ancestors in Naples' blue and balmy bay. Hist. Philos. Life Sci. **36**: 371-393.
- Hozumi, A., Horie, T. and Sasakura, Y. 2015. Neuronal map reveals the highly regionalized pattern of the juvenile central nervous system of the ascidian *Ciona intestinalis*. Dev. Dyn. **244**: 1375-1393.
- Huang, X., Gao, Y., Jiang, B., Zhou, Z. and Zhan, A. 2015. Reference gene selection for quantitative gene expression studies during biological invasions: A test on multiple genes and tissues in a model ascidian *Ciona savignyi*. Gene **576**: 79-87.
- Iyappan, K., Ananthan, G. and Sathishkumar, R. 2015. Molecular identification of ascidians from the Palk Bay Region, Southeast coast of India. Mitochondrial DNA **epub**: 1-4.
- Jaffar Ali, H. A. and Ahmed, N. S. 2015. DNA barcoding of two solitary ascidians, *Herdmania momus* Savigny, 1816 and *Microcosmus squamiger* Michaelsen, 1927 from Thoothukudi coast, India. Mitochondrial DNA **epub**: 1-3.
- Jaffar Ali, H. A., Tamilselvi, M., Akram, A. S., Kaleem Arshan, M. L. and Sivakumar, V. 2015. Comparative study on bioremediation of heavy metals by solitary ascidian, *Phallusia nigra*, between Thoothukudi and Vizhinjam ports of India. Ecotoxicol. & Envir. Safety **121**: 93-99.
- Januario, S. M., Estay, S. A., Labra, F. A. and Lima, M. 2015. Combining environmental suitability and population abundances to evaluate the invasive potential of the tunicate *Ciona intestinalis* along the temperate South American coast. PeerJ **3**:
- Jeffery, W. R. 2015. Regeneration, stem cells, and aging in the tunicate *Ciona*: insights from the oral siphon. Intl. Rev. Cell Mol. Biol. **319**: 255-282.
- Jo, J.-E., Kim, K.-H., Yoon, M.-H., Kim, N.-Y., Lee, C. and Yook, H.-S. 2010. Quality characteristics and antioxidant activity research of *Halocynthia roretzi* and *Halocynthia aurantium*. J. Korean Soc. Food Sci. & Nutrition **39**: 1481-1486.
- Kassmer, S. H., Rodriguez, D., Langenbacher, A. D., Bui, C. and De Tomaso, A. W. 2015. Migration of germline progenitor cells is directed by sphingosine-1-phosphate signalling in a basal chordate. Nature Commun. **6**:
- Kehr, J. C. and Dittmann, E. 2015. Protective tunicate endosymbiont with extreme genome reduction. Env. Microbiol. **17**: 3430-3432.
- Kimura, S., Nakayama, K., Wada, M., kim, U.-J., Azumi, K., Ojima, T., Nozawa, A., Kitamura, S.-I. and Hirose, E. 2015. Cellulose is not degraded in the tunic of the edible ascidian *Halocynthia roretzi* contracting soft tunic syndrome. Dis. Aquat. Organ. **116**: 143–148.
- Koplovitz, G., Shmuel, Y. and Shenkar, N. 2016. Floating docks in tropical environments a reservoir for the opportunistic ascidian *Herdmania momus*. Manag. Biolog. Invasions **76**: 1-8.
- Kuo, C.-Y., Fan, T.-Y., Li, H.-H., Lin, C.-W., Liu, L.-L. and Kuo, F.-W. 2015. An unusual bloom of the tunicate, *Pyrosoma atlanticum*, in southern Taiwan. Bull. Mar. Sci. **91**: 363-364.
- Kuplik, Z., Kerem, D. and Angel, D. L. 2015. Regulation of *Cyanea capillata* populations by predation on their planulae. J. Plankton Res. **37**: 1068-1073.
- Kwon, T.-H., Kim, J.-K., Kim, T.-W., Lee, J.-W., Kim, J.-T., Seo, H.-J., Kim, M.-J., Kim, C.-G., Jeon, D.-S. and Park, N.-H. 2011. Antioxidant and anti-lipase activity in *Halocynthia roretzi* extracts. Kor. J Food Sci. & Technol. **43**: 464-468.
- Le Goff, E., Martinand-Mari, C., Martin, M., Feuillard, J., Boublik, Y., Godefroy, N., Mangeat, P., Baghdiguian, S. and Cavalli, G. 2015. Enhancer of zeste acts as a major

developmental regulator of *Ciona intestinalis* embryogenesis. Biol. Open **4**: 1109-1121. Lemaire, P. and Piette, J. 2015. Tunicates: exploring the sea shores and roaming the open

ocean. A tribute to Thomas Huxley. Open Biol. 5:

- Lesser, M. P. and Slattery, M. 2015. Picoplankton consumption supports the ascidian *Cnemidocarpa verrucosa* in McMurdo Sound, Antarctica. Mar. Ecol. Prog. Ser. **525**: 117-126.
- Lin, Y., Chen, Y., Xiong, W. and Zhan, A. 2015. Genome-wide gene-associated microsatellite markers for the model invasive ascidian, *Ciona intestinalis* species complex. Molec. Ecol. Resources **epub**:
- López-Legentil, S., Erwin, P. M., Turon, M. and Yarden, O. 2015. Diversity of fungi isolated from three temperate ascidians. Symbiosis **66**: 99-106.
- López-Legentil, S., Turon, X., Espluga, R. and Erwin, P. M. 2015. Temporal stability of bacterial symbionts in a temperate ascidian. Frontiers in Microbiol. **6**: 1-11.
- Lord, J. and Whitlatch, R. 2015. Predicting competitive shifts and responses to climate change based on latitudinal distributions of species assemblages. Ecology **96**: 1264-1274.
- Lowen, J. B., Deibel, D., McKenzie, C. H., Couturier, C. and DiBacco, C. 2015. Tolerance of early life-stages in *Ciona intestinalis* to bubble streams and suspended particles. Manag. Biolog. Invasions **7**: 1-9.
- Matsunobu, S. and Sasakura, Y. 2015. Time course for tail regression during metamorphosis of the ascidian *Ciona intestinalis*. Dev. Biol. **405**: 71-81.
- McDougall, A., Chenevert, J., Pruliere, G., Costache, V., Hebras, C., Salez, G. and Dumollard, R. 2015. Centrosomes and spindles in ascidian embryos and eggs. Methods Cell Biol. **129**: 317-339.
- Miller, R. J., Page, H. M. and Reed, D. C. 2015. Trophic versus structural effects of a marine foundation species, giant kelp (*Macrocystis pyrifera*). Oecologia **179**: 1199-1209.
- Mizotani, Y., Itoh, S., Hotta, K., Tashiro, E., Oka, K. and Imoto, M. 2015. Evaluation of drug toxicity profiles based on the phenotypes of ascidian *Ciona intestinalis*. Biochem. Biophys. Res. Comm. **463**: 656-660.
- Moles, J., Nunez-Pons, L., Taboada, S., Figuerola, B., Cristobo, J. and Avila, C. 2015. Antipredatory chemical defences in Antarctic benthic fauna. Mar. Biol. **162**: 1813-1821.
- Morales Diaz, H., Mejares, E., Newman-Smith, E. and Smith, W. C. 2015. ACAM, a novel member of the neural IgCAM family, mediates anterior neural tube closure in a primitive chordate. Dev. Biol. **epub**:
- Nakazawa, S., Shirae-Kurabayashi, M., Otsuka, K. and Sawada, H. 2015. Proteomics of ionomycin-induced ascidian sperm reaction: Released and exposed sperm proteins in the ascidian *Ciona intestinalis*. Proteomics **epub**: 1-16.
- Nam, K. W., Shin, Y. K. and Park, K. I. 2015. Seasonal variation in *Azumiobodo hoyamushi* infection among benthic organisms in the southern coast of Korea. Parasites & Vectors 8: 1-7.
- Nawata, A., Hirose, E., Kitamura, S. and Kumagai, A. 2015. Encystment and excystment of kinetoplastid *Azumiobodo hoyamushi*, causal agent of soft tunic syndrome in ascidian aquaculture. Dis. Aquat. Organ. **115**: 253-262.
- Negishi, T. and Yasuo, H. 2015. Distinct modes of mitotic spindle orientation align cells in the dorsal midline of ascidian embryos. Dev. Biol. **epub**:
- Omotezako, T. and al., e. 2015. DNA interference: DNA-induced gene silencing in the appendicularian *Oikopleura dioica*. **282**:
- Orito, W., Ohhira, F. and Ogasawara, M. 2015. Gene expression profiles of FABP genes in protochordates, *Ciona intestinalis* and *Branchiostoma belcheri*. Cell Tiss. Res. **362**: 331-345.
- Piette, J. and Lemaire, P. 2015. Thaliaceans, the neglected pelagic relatives of ascidians: a developmental and evolutionary enigma. Quart. Rev. Biol. **90**: 117-145.

- Piperigkou, Z., Karamanou, K., Afratis, N. A., Bouris, P., Gialeli, C., Belmiro, C. L., Pavao, M. S., Vynios, D. H. and Tsatsakis, A. M. 2015. Biochemical and toxicological evaluation of nano-heparins in cell functional properties, proteasome activation and expression of key matrix molecules. Toxicol. Lett. **240**: 32-42.
- Pochon, X., Zaiko, A., Hopkins, G. A., Banks, J. C. and Wood, S. A. 2015. Early detection of eukaryotic communities from marine biofilm using high-throughput sequencing: an assessment of different sampling devices. Biofouling **31**: 241-251.
- Pomin, V. H. 2015. NMR structural determination of unique invertebrate glycosaminoglycans endowed with medical properties. Carbohydr. Res. **413**: 41-50.
- Qi, Z., Han, T., Zhang, J., Huang, H., Mao, Y., Jiang, Z. and Fang, J. 2015. First report on in situ biodeposition rates of ascidians (*Ciona intestinalis* and *Styela clava*) during summer in Sanggou Bay, northern China. Aquaculture Environment Interactions **6**: 233-239.
- Reid, V., McKenzie, C., Matheson, K., Wells, T. and Couturier, C. 2016. Post-metamorphic attachment by solitary ascidian *Ciona intestinalis* (Linnaeus, 1767) juveniles from Newfoundland and Labrador, Canada. **7**:
- Rinkevich, B. 2015. Conserved histocompatible machinery in marine invertebrates? ISJ **12**: 70-172.
- Rosner, A., Alfassi, G., Moiseeva, E., Paz, G., Rabinowitz, C., Lapidot, Z., Douek, J., Haim, A. and Rinkevich, B. 2014. The involvement of three signal transduction pathways in botryllid ascidian astogeny, as revealed by expression patterns of representative genes. Int. J. Dev. Biol. **58**: 677-692.
- Roure, A. and Darras, S. 2015. Msxb is a core component of the genetic circuitry specifying the dorsal and ventral neurogenic midlines in the ascidian embryo. Dev. Biol. **epub**:
- Sahade, R., Lagger, C., Torre, L., Momo, F., Monien, P., Schloss, I., Barnes, D. K. A., Servetto, N., Tarantelli, S., Tatián, M., Zamboni, N. and Abele, D. 2015. Climate change and glacier retreat drive shifts in an Antarctic benthic ecosystem. Science Advances **epub**: 1-9.
- Sato, A., Kawashima, T., Fujie, M., Hughes, S., Satoh, N. and Shimeld, S. M. 2015. Molecular basis of canalization in an ascidian species complex adapted to different thermal conditions. Sci. Rep. 5:
- Schofield, M. M., Jain, S., Porat, D., Dick, G. J. and Sherman, D. H. 2015. Identification and analysis of the bacterial endosymbiont specialized for production of the chemotherapeutic natural product ET-743. Env. Microbiol. **17**: 3964-3975.
- Seaward, K., Acosta, H., Inglis, G. J., Wood, B., Riding, T. A. C., Wilkens, S. and Gould, B.
   2015. The Marine Biosecurity Porthole a web-based information system on nonindigenous marine species in New Zealand. Manag. Biolog. Invasions 6: 177–184.
- Stabili, L., Licciano, M., Longo, C., Lezzi, M. and Giangrande, A. 2015. The Mediterranean non-indigenous ascidian Polyandrocarpa zorritensis: Microbiological accumulation capability and environmental implications. Mar. Pollution Bull. **epub**:
- Stach, T. 2015. What's in a name? J. Zool. Syst. & Evol. Res. 53: 185.
- Stefaniak, L. M. and Heupel, J. 2016. Alternative menthol sources for ascidian relaxation. 7: 1-4.
- Stolfi, A., Ryan, K., Meinertzhagen, I. A. and Christiaen, L. 2015. Migratory neuronal progenitors arise from the neural plate borders in tunicates. Nature **527**: 371-374.
- Sukhachev, A. N., Dyachkov, I. S., Kudryavtsev, I. V., Kumeiko, V. V., Tsybulskiy, A. V. and Polevshchikov, A. V. 2015. Application of flow cytometry for the analysis of circulating hemocyte populations in the ascidian *Halocynthia aurantium* (Pallas, 1787). J. Evol. Biochem. & Physiol. **51**: 246-253.

- Takatori, N., Oonuma, K., Nishida, H. and Saiga, H. 2015. Polarization of PI3K activity initiated by ooplasmic segregation guides nuclear migration in the mesendoderm. Dev. Cell **35**: 333-343.
- Taketa, D. A., Nydam, M. L., Langenbacher, A. D., Rodriguez, D., Sanders, E. and De Tomaso, A. W. 2015. Molecular evolution and in vitro characterization of *Botryllus* histocompatibility factor. Immunogenetics **67**: 605-623.
- Thomas, J. 2015. *Leucothoe eltoni* sp. n., a new species of commensal leucothoid amphipod from coral reefs in Raja Ampat, Indonesia (Crustacea, Amphipoda). Zookeys **518**: 51-66.
- Thompson, H. and Shimeld, S. M. 2015. Transmission and scanning electron microscopy of the accessory cells and chorion during development of *Ciona intestinalis* Type B embryos and the Impact of their removal on cell morphology. Zool. Sci. **32**: 217-222.
- Tran, T. D., Pham, N. B., Ekins, M., Hooper, J. N. A. and Quinn, R. J. 2015. Isolation andtotal synthesis of stolonines A-C, unique taurine amides from the Australian marine tunicate *Cnemidocarpa stolonifera*. Mar. Drugs **13**: 4556-4575.
- Uribe, E. and Etchepare, I. 2002. Effects of biofouling by *Ciona intestinalis* on suspended culture of *Argopecten purpuratus* in Bahía Inglesa, Chile. Bull. Aquaculture Assoc. Canada **102**: 93-95.
- Vercaemer, B., Sephton, D., Clément, P., Harman, A., Stewart-Clark, S. and DiBacco, C. 2015. Distribution of the non-indigenous colonial ascidian *Didemnum vexillum* (Kott, 2002) in the Bay of Fundy and on offshore banks, eastern Canada. Manag. Biolog. Invasions **6**: 385–394.
- Vizzini, A., Bonura, A., Longo, V., Sanfratello, M. A., Parrinello, D., Cammarata, M. and Colombo, P. 2015. Isolation of a novel LPS-induced component of the ML superfamily in *Ciona intestinalis*. Dev. Comp. Immunol. **53**: 70-78.
- Vizzini, A., Di Falco, F., Parrinello, D., Sanfratello, M. A. and Cammarata, M. 2015. Transforming growth factor beta (CiTGF-beta) gene expression is induced in the inflammatory reaction of *Ciona intestinalis*. Dev. Comp. Immunol. **55**: 102-110.
- Waki, K., Imai, K. S. and Satou, Y. 2015. Genetic pathways for differentiation of the peripheral nervous system in ascidians. Nature Commun. 6:
- Waldrop, L. D. and Miller, L. A. 2015. The role of the pericardium in the valveless, tubular heart of the tunicate *Ciona savignyi*. J. Exp. Biol. **218**: 2753-2763.
- Wang, K. and Nishida, H. 2015. REGULATOR: a database of metazoan transcription factors and maternal factors for developmental studies. BMC Bioinformatics **16**: 114.
- Wang, K., Omotezako, T., Kishi, K., Nishida, H. and Onuma, T. A. 2015. Maternal and zygotic transcriptomes in the appendicularian, *Oikopleura dioica*: novel protein-encoding genes, intra-species sequence variations, and trans-spliced RNA leader. Dev. Genes Evol. 225: 149–159.
- Youssef, D. T. A., Mohamed, G. A., Shaala, L. A., Badr, J. M., Bamanie, F. H. and Ibrahim, S. R. M. 2015. New purine alkaloids from the Red Sea marine tunicate *Symplegma rubra*. Phytochemistry Letters **13**: 212-217.
- Zabin, C. J., Ashton, G. V., Brown, C. W., Davidson, I. C., Sytsma, M. D. and Ruiz, G. M. 2014. Small boats provide connectivity for nonindigenous marine species between a highly invaded international port and nearby coastal harbors. Manag. Biolog. Invasions 5: 97– 112.
- Zhan, A., Briski, E., Bock, D. G., Ghabooli, S. and Macisaac, H. J. 2015. Ascidians as models for studying invasion success. Mar. Biol. **epub**: 1-22.