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## Glossary

This glossary is intended to trace the roots of the often-confusing terms used in placental pathology and perinatal development. The accents are placed for pronunciation.

**Gk.** = Greek; **L.** = Latin; **Fr.** = French; **OE** = Old English; **ME** = Middle English.

**Abrúptio (placéntae):** detachment of placenta [L. *abrumperē* = to break away]

**Acárdius:** malformed twin without heart, invariably one of monozygotic twins [Gk. *a* = without, not + *kardia* = heart]

**Adventitía:** outer layer of vessel wall [L. *advenire* = to add]

**Allántoís (allantóic):** designation of one type of placenta, because of its roots in other mammals; thin membrane between amnion and chorion [Gk. *allantos* = sausage]

**Ámnion:** thin membrane surrounding the fetus; lamb's caul [Gk. *amnos* = lamb] **Note:** Because we use chorionic and not choriotic, it is here preferred to speak of amniotic, rather than amniotic. Hyrtl (1880) explored the various terminology used to describe placental structures. He concluded that amnios and amnion were both correct. It was first used by Galen, referring to skin. The reference to lamb comes from Vesalius.

**Androgénesis:** development of male gender [Gk. *andros* = man + *gennan* = produce]

**Anídian monster:** a hideous fetus with peculiar features; a form of acardiac twin [Gk. *an* = not + *idios* = peculiar + L. *monstrum*]

**Artiodáctyla:** order of mammals, the even-hoofed animals, e.g., cow, deer [Gk. *artios* = even + *daktylos* = toe]

**Báttledore (placenta):** marginal insertion of cord [OE = flat, wooden paddle used in the game of battledore to hit the shuttlecock]

**Blástocyst:** early germinative vesicle [Gk. *blaste* = germ + *kystis* = vesicle]

**Bosselátion:** surface granulation of placenta [Fr. *bosseler* = to ornament with bosses; from *bosse* = knob]

**Capillary:** hair-fine blood vessel [L. *capilla* = hair]

**Céllular:** belonging to the cell [L. *cellula* = small cabin, cell]

**Extracellular:** outside the cell

**Intercellular:** between cells

**Intracellular:** within cells

**Transcellular:** across cells

**Chiméra (Chímerism):** the composite of several genotypes [Gk. *chimaira* = a monstrous beast. In Greek mythology a monster made of the head of a lion, body of a goat, and tail of a dragon.]

**Chirality:** the quality of being chiral [Gk. *chiro* = the hand; a three-dimensional form, as a molecule, that cannot be superimposed on its mirror image; used in designating the twist of the umbilical cord; its first usage was discussed by McManus (2002)]

**Chorioangiópagus parasíticus:** a fetus, connected by blood vessels to another fetus [Gk. *chorion* = "little gut" = outer membranes around embryo + *angeios* = vessel + *pagos* = something set or fixed; *para* = next to + *sitos* = food]

**Chórion:** outer membrane around embryo [Gk. *chorion* = "little gut"; according to Hyrtl (1880) the term was also used by Galen as the outer shell of the membranes]

**frondósium:** the placenta proper [L. *frondosus* = richly covered with leaves, as in tree]

**laéve:** the membranous portion of the chorionic sac [L. *levis* = smooth, without villi]

**Chorioneopitheliómá:** malignant tumor of trophoblast = chorioepithelioma or choriocarcinoma [Gk. *chorion* = "little gut" (outer membrane enclosing an embryo) + *epi* = on + *thele* = nipple + *oma* = tumor]

**Circumvállate (placenta):** an abnormal form of placenta with circumferential, old hemorrhages [L. *circum* + *vallare* = to wall around]

**Cirsoid:** aneurysmal dilatation of vessel [Gk. *kirisos* = enlarged vein]

**Cotylédon:** originally name for the single spots of placental tissue in the ruminants; lobe of the human placenta [Gk. *kotyle* = cup]

**Cytotrophoblast:** cellular type of the trophoblast [Gk. *kytos* = cell + *trephein* = to nourish + *blaste* = germ]

**Decídua:** the endometrium at end of the luteal phase [L. *decidere* = fall, die]

**basalis:** basal portion of placenta [Gk. *basis* = base]

- capsularis:** outer portion of membranes [L. *capsula* = little box]
- parietalis:** endometrium of pregnancy, covering wall portion of uterus [L. *paries* = wall; *parietalis* = pertaining to wall]
- véra:** uterine decidua, contrasting it to pseudodecidua, outside of uterus, as in endometriosis [L. *verus* = true]
- Désmosome:** intercellular junction [Gk. *desmos* = Ligament + *soma* = body]
- Dizygótico:** twins of two ova, “fraternal twins” [Gk. *dis* = twice, two + *zygon* = yoke]
- Eclámpsi:** coma and convulsive seizure in pregnancy [Gk. *eklampsia* = shining forth; or *ek* = out + *lampein* = to shine]
- Émbryoblast:** embryo-forming cells of the blastocyst [Gk. *embryon* = unborn child + *blaste* = germ]
- Endométrium:** innermost layer of the uterus [Gk. *endon* = inside + *metra* = uterus]
- Endoplásmic reticulum:** a net-like, membrane-lined cell organelle [Gk. *endon* = inside + *plasma* = juice; L. *reticulum* = small net]
- Endothélium:** innermost layer of blood vessels [Gk. *endon* = inside + *thele* = mamilla]
- Epígnathus:** tumorous mass in mouth, affixed to jaw and possibly a twin [Gk. *epi* = upon, at, over + *gnathos* = jaw]
- Epitelium:** superficial cellular layer [Gk. *epi* = upon + *thele* = mamilla]
- Fétus papyráceus:** paper-like, macerated, compressed (“compressus”) fetus [L. *papyrus* = plant from which paper is made]
- Fibrín:** blood clot product [L. *fibra* = fiber]
- Fibrinoid:** a substance similar to but not identical with fibrin [Gk: . . . *eides* = looking like]
- Fibroblast:** connective tissue cell [Gk. *blaste* = germ]
- Freemártin:** the female of fraternal cattle twins, sterilized in utero by the male co-twin with whose placental vessels she is joined; the term presumably comes from the St. Martin’s feast in England, when these animals were consumed
- Funiculópagos (twins):** twins joined at umbilical cord [L. *funis* = rope, cord + Gk. *pagos* = fixed]
- Fúnis (funículus):** umbilical cord [L. *funis* = rope, cord]
- Funisítis:** inflammation of umbilical cord [L. *funis* = rope]
- Fúrcate (cord) (“insértio funículi furcata”):** forked insertion of the little rope [L. *furca* = fork]
- Glycocáliz:** superficial layer of polysaccharides, covering the cell surface [Gk. *glycos* = sweet + *kalyx* = goblet]
- Granulomatosis infantiséptica:** neonatal disseminated listeriosis [L. *granulum* = little grain + Gk. *oma* = tumor]
- Gynogénesis:** female development [Gk. *gyne* = woman + *gennan* = produce]
- Hemiacárdius:** acardiac monster in which remnants of heart may be found [L. *hemi* = a half]
- Holoacárdius:** completely heartless monster (twin) [Gk. *holos* = whole]
- Hydatídiform (mole):** severely hydropic placenta with bulbous, villous enlargement [Gk. *hydatis* = watery vesicle + L. *forma* = shape; L. *mola* = false conception; mass]
- Hydrámnios:** excessive amount of amniotic fluid [Gk. *hydror* = water (*hydros* = sweat; some early authors believed that amniotic fluid was fetal sweat) + *amnion* = lamb’s caul]
- Implantación:** establishing of intimate fetomaternal contact in the uterus [L. *implantare* = to embed]
- Intervillous:** between the placental villi, i.e., in the maternal blood space [L. *inter* = between]
- Intravillous:** within a villus [L. *intra* = in]
- Lacúna:** [L. = hole, gap]
- Lithopédion:** stone-like fetus [Gk. *lithos* = stone + *paidion* = child]
- Lóchia (pl.):** uterine discharges after birth [Gk. *lochia*]
- Mácrofage:** phagocytotic and paracrine cell type [Gk. *macros* = large + *phagein* = to eat]
- Márgma reticuláre:** jelly-like fluid in original embryonic sac [Gk. *magma* = suspension of finely divided material in small amount of water + L. *reticulum* = little net (network)]
- Mármoset:** family of small South American primates that always produce fraternal twins [Old Fr. *marmouset* = grotesque figure]
- Mecónium:** fetal intestinal content [Gk. *mekonion* = poppy juice]
- Mésenchyme:** undifferentiated connective tissue [Gk. *mesos* = in the middle of + *chein* = to pour (something poured in between)]
- Mésoderm:** the middle germ layer [Gk. *derma* = skin]
- Mésothelium:** connective tissue-derived epithelial layer [Gk. *thele* = mamilla]
- Microvillus:** finger-like extension of the cell surface
- Mitochondrion:** rod-shaped cell organelle [Gk. *mitos* = thread + *chondros* = grain]
- Mole (hydatidiform, Breus’, etc.):** vesicular mass of placenta [L. *mola* = false conception]
- Monozygótico:** single-egg-derived twins (“identical twins”) [Gk. *monos* = single + *zygotos* = yoked]
- Nódus spúrius vasculósus (gelatínus):** false knot in umbilical cord of vascular genesis [L. *nodus* = knot + *spurius* = not genuine + *gelatina* = gelatin]
- Núchal (cord):** umbilical cord entwined around neck [L. *nucha* = nape of the neck]
- Óctoploid:** having eight sets of chromosomes [Gk. *okto* = eight + *ploos* = fold + *eidos* = form]
- Oligohydrámnios:** too little, or no amniotic fluid [Gk. *oligos* = little + *hydror* = water (*hydros* = Sweat) + *amnion* = lamb caul]
- Ómphalo-(mesentérico):** umbilicus [Gk. *omphalos* = Navel]
- Páraplacénta:** those parts of the chorionic sac, not belonging to the placenta, e.g., membranes [Gk. *para* = beside]
- Périvillous:** around the placental villi [Gk. *peri* = around]
- Placénta:** [L. flat cake] According to Hyrtl (1880) this term, with an originally Greek root, was introduced in 1559 by Realdus Columbus; others referred to it as “secundines”
- accréta:** unusually adherent placenta that fails to detach [L. *accrescere* = to grow together, adhere]

- incréta:** placenta that has grown into the myometrium [L. *increscere* = to grow into]
- membranácea:** very thin placental membrane, the entire outside of which is covered with villi [L. *membrana* = from parchment, membrane]
- percréta:** placenta that has grown through the uterine wall [L. *percrebescere* = to crowd everywhere]
- prévia:** placenta that is located in the lower uterus and is in the way before the fetus can be delivered [L. *praevius* = in front of, before, leading the way]
- Plasmódium:** multinucleate mass of protoplasm [Gk. *plasma* = a thing formed (juice) + *eidos* = to form]
- Pólipoide:** having many sets of chromosomes [Gk. *polys* = much, many + *ploos* = fold + *eidos* = form]
- Pólipoide:** protrusion of tissue, polyp [Gk. *polys* = many + *pous* = foot + *oid* = like]
- Pycnósis (pyknosis):** degenerative condensation of cells or nuclei [Gk. *pyknos* = dense]
- Schistocytes:** broken red cells in disseminated intravascular coagulation [Gk. *schistos* = split]
- Secúndines:** synonymous with afterbirth [L. *secundus* = following]
- Sínusoid:** enlarged capillary [L. *sinus* = bight + Gk. *eides* = looking like]
- Siréniform (fétus):** malformed infant with fused legs [Gk. *seiren* = mermaid]
- Sirenomélia:** malformed infant with fused legs [Gk. *seiren* = mermaid + *melos* = limb]
- Stróma:** connective tissue core of an organ [L. *stroma* = cushion]
- Subchoríal:** under the chorionic plate [L. *sub* = below + Gk. *chorion* = leather, embryonic membrane]
- Succentúriate (lobe):** accessory lobe of placenta [L. *succenturiare* = to substitute]
- Superfecundación:** fertilization of two or more ova at different times during the same menstrual period [L. *super* = beyond, excessively + *fecundus* = fertile]
- Superfetación:** pregnancy on top of already existing pregnancy [L. *superfetare* = to bring forth while already pregnant; L. *fetus* = fruit, offspring]
- Sympus:** fetus with fused legs [Gk. *syn* = together + *pous* = foot (same as siren)]
- Syncytiotrophoblast:** syncytial type of trophoblast
- Syncytium:** multinuclear mass, derived from cell fusion [Gk. *syn* = together + *kytos* = cell]
- Synéchia (pl. synéchia):** adhesion of parts, here in the uterus [Gk. *synecheia* = continuity]
- Tessellátion:** irregular surface of placenta [L. *tessella* = little square stone (From Gk. *tessares* = four)]
- Tétraploid:** having four sets of chromosomes [Gk. *tettares* = four + *ploos* = fold]
- Thalassémias:** hemoglobin disorders [from Gk. *thalassa* = the sea; generally referring to the Mediterranean]
- Thixotrópic (Thyxo-) gel:** gel that liquefies when shaken, generally the extraembryonic fluid [Gk. *thixis* = touching + *trope* = turn]
- Thoracopágus:** twins, conjoined at chest [Gk. *thorax* = chest + *pagos* = fix]
- Trabécula:** small septum [L. small beam]
- Tróphoblast:** the epithelium that covers the placenta [Gk. *trophe* = nourishment + *blastos* = germ]
- Trophótropism:** the “wandering” of the placenta to the site of best nourishment [Gk. *trophe* = nourishment + *trope* = turn]
- Urachus:** connection of bladder to allantoic sac [Gk. *ourachos* (“that which has a tail”) = cord that extends from bladder to navel]
- Vas prévium (pl. Vásá prévia):** blood vessels within membranes that present before fetal parts during delivery [L. *vas* = vessel(s) that is (are) ahead (“previous”) of fetal part]
- Vas vasórum (Vásá vasórum):** blood vessels that nourish the vessels [L. *vas* = vessel]
- Velaméntous (cord insertion):** membranous insertion of umbilical cord [L. *velamen* = veil + *velamentum* = cover]
- Vérmix caseosa:** sebum, hair and other skin secretions from fetus [L. *vernix* = varnish + *caseus* = cheese]
- Víllus (pl. vílli):** ramifications of placenta with fetal vessels that are the “business end” of the placenta, covered with trophoblast [L. *villus* = tuft of hair]
- Vitelline:** belonging to the yolk sac [L. *vitellus* = yolk of an egg]

## References

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# 28

## Normative Values and Tables

Some quantitative structural and biochemical data concerning pregnancy, placental development, and composition of the term placenta are given in Tables 28.1 to 28.10.

When examining the tables on placental morphometry and comparing the results from different authors, it is important to note that quantitative structural data are heavily influenced by the mode of sampling and by the preparation of the material. Because of the high degree of maternal and fetal vascularization, the placenta reacts immediately to changes in intravascular pressure. Thus, the mode of birth, the time elapsing from cessation of maternal and fetal blood flows to tissue fixation (see Tables 28.6 to 28.10), and the nature of cord clamping (see Table 28.8) directly influence the volumetric relations of villi and intervillous space. In particular, parameters such as the width of fetal vessels, degree of fetal vascularization, maternal-fetal diffusion distance, and trophoblastic thickness are easily affected. Moreover, the composition of the fixative and its osmolarity (see Table 28.10), as well as the mode of fixation (immersion versus perfusion fixation) are of importance. Normally, immersion fixation of the entire placenta or of small pieces is used. The more advanced methods, such as perfusion fixation (Burton et al., 1987) or puncture biopsy of the still maternally perfused placenta during cesarean section (Schweikhart & Kaufmann, 1977; Voigt et al., 1978; Sen et al., 1979), are very time-consuming. When studying immersion fixed material, however, one should keep in mind that this material differs quantitatively and qualitatively from the *in vivo* conditions (see Tables 28.6 and 28.9). It is

impossible to include the results of numerous other valuable contributions to placental morphometry in these tables. For further information on special issues, we refer to the following publications: placental growth development in relation to birth weight (Bouw et al., 1978; Molteni et al., 1978); relationship of placental weight to body size at 7 years of age, and to abnormalities in children (Naeye, 1987); fetal and placental weights in relation to maternal weight (Auinger & Bauer, 1974); ultrasonographic measurements of volumetric growth of the placenta (Bleker et al., 1977); weight development of placenta and membranes in early pregnancy (Abramovich, 1969); ratio of gestational sac volume to crown-rump length in early pregnancy (Goldstein et al., 1986); villous surface area and villous volume densities in various placental regions and along different levels of the chorial basal axis (Teasdale, 1978; Boyd et al., 1980; Cabezon et al., 1985; Bacon et al., 1986); local variations of villous surface, fetal vascularization, and amount of vasculosyncytial membranes in the placentone (maternal-fetal circulatory unit) (Schuhmann et al., 1986); total villous surface in relation to fetal weight, in normal and various pathologic cases (Clavero-Nunez & Botella-Llusia, 1961, 1963); morphometric data affecting placental oxygen diffusion (Mayhew et al., 1984, 1986); computer measurement of the mass of syncytiotrophoblast (Boyd et al., 1983); ultrastructural morphometric analysis of the villous syncytiotrophoblast (Sala et al., 1983); microvillous surface enlargement of the villous surface (Teasdale & Jean-Jacques, 1985); comparison of villous structure following immersion and perfusion fixation (Burton et al., 1987).

TABLE 28.1. Length and weight data for placenta and fetus; pregnancies and deliveries at sea level. (1) Data extrapolated from Boyd & Hamilton (1970) and O’Rahilly (1973); (2) data from Johannigmann et al. (1972); (3) data from Winckel (1893), Branca (1922), Naeye & Tafari (1983), and Fujinaga et al. (1990); (4) data from Weissman et al. (1994). Placental weight data published by Molteni et al. (1978) are 5% to 10% lower because fetal blood was drained from chorionic vessels before weighing. Compare with data on influence of cord clamping given in Table 28.8. Umbilical cord length as measured by ultrasound throughout the first trimester (Hill et al., 1994) amounts to only about 50% of the length measured after abortion.

Pregnancy week (postconception)	Pregnancy week (postmenstruation)	Pregnancy month (postmenstruation)	Crown-rump length (mm) <sup>1</sup>	Embryonic/fetal weight (g) <sup>1</sup>	Diameter of the chorionic sac (mm) <sup>1</sup>	Placental diameter (mm)	Placental weight (g) <sup>1</sup>	Placental thickness postpartum (mm) <sup>1</sup>	Placental thickness including uterine wall measured by ultrasound <i>in vivo</i> (mm) <sup>2</sup>	Length of the umbilical cord (mm) <sup>3</sup>	Diameter of the umbilical cord measured by ultrasound (mm) <sup>4</sup>	Diameter of the umbilical artery measured by ultrasound (mm) <sup>4</sup>	Diameter of the umbilical vein measured by ultrasound (mm) <sup>4</sup>	
1														
2		1												
1		3												
2		4									2			
3	5		2.5		5–11					4				
4	6	2	5		12–19					7				
5	7		9		20–25					12				
6	8		14	1.1	26–33		6			20	2.5			
7	9		20	2	34–41		8			33				
8	10	3	26	5	42–48		13			55	3.3			
9	11		33	11	49–56		19			92				
10	12		40	17	57–65		26			126	4.4			
11	13		48	23	66–73	50	32			158				
12	14	4	56	30	74–81	56	41	10		188	6.1	1.2	2.0	
13	15		65	40	82–89	62	50	11		215				
14	16		75	60	90–99	69	60	12		240	7.0	1.1	2.4	
15	17		88	90		75	70	12		264				
16	18	5	99	130		81	80	13		287	10.1	1.9	3.6	
17	19		112	180		87	101	14		309				
18	20		125	250		94	112	15	28	330	11.1	2.0	4.1	
19	21		137	320		100	126	15	29	350				
20	22	6	150	400		106	144	16	30	369	12.8	2.4	4.7	
21	23		163	480		112	162	17	32	387				
22	24		176	560		119	180	18	34	404	13.9	2.6	5.4	
23	25		188	650		125	198	18	35	420				
24	26	7	200	750		131	216	19	36	435	15.2	2.8	6.0	
25	27		213	870		137	234	19	37	450				
26	28		226	1,000		144	252	20	38	464	15.9	3.1	6.6	
27	29		236	1,130		150	270	20	39	477				
28	30	8	250	1,260		156	288	21	40	490	16.3	3.4	7.3	
29	31		263	1,400		162	306	21	41	502				
30	32		276	1,550		169	324	22	42	520	17.6	3.6	7.7	
31	33		289	1,700		175	342	22	42	530				
32	34	9	302	1,900		181	360	23	43	540	17.4	3.3	7.4	
33	35		315	2,100		187	378	23	43	549				
34	36		328	2,300		194	396	24	44	557	17.4	3.7	7.6	
35	37		341	2,500		200	414	24	44	565				
36	38	10	354	2,750		206	432	24	45	572	18.0	4.2	8.2	
37	39		367	3,000		213	451	25	45	579				
38	40		380	3,400		220	470	25	45	585	17.0	3.9	7.8	

TABLE 28.2. Quantitative data of various placental tissues as related to fetal weight and stage of pregnancy. (1) Data calculated from Table 28.1; (2) Data from Kloos & Vogel (1974); (3) data from Kaufmann (1981); (4) data from Knopp (1960). Fetoplacental weight ratios calculated by Molteni et al. (1978) are generally higher because these authors had drained fetal blood from chorionic vessels before weighing.

Pregnancy week (postconception)	Pregnancy week (postmenstruation)	Pregnancy month (postmenstruation)	Fetal weight (g) per 1 g of placental weight <sup>1</sup>	Placental weight index: placental weight (g) per 1 g of fetal weight <sup>2</sup>	Villous weight (g) per placenta <sup>1</sup> (percentage of total placental weight) placental diameter (mm) <sup>1</sup>	Villous surface (cm <sup>2</sup> ) per placenta <sup>1</sup>	Fetal weight (g) per 10,000 cm <sup>2</sup> villous surface <sup>1</sup>	Villous surface (cm <sup>2</sup> ) per 1 g of placental villi <sup>1</sup>	Number of villous cross sections per mm <sup>2</sup> of paraffin sections <sup>3</sup>	Number of villous cross sections per mm <sup>2</sup> of paraffin sections <sup>4</sup>	Villous diameter (mean of all peripheral villous types) (μm) <sup>3</sup>
1	1	1									
2	2	1									
1	3										
2	4										
3	5										
4	6	2									
5	7										
6	8		0.18	10.0–9.3	5 (83%)	830		166	15	47	204
7	9		0.25								
8	10	3	0.38								
9	11		0.58								
10	12		0.65	3.9–3.3	18 (69%)	3,020	29.8	168	20		
11	13		0.72								
12	14	4	0.73								
13	15		0.80								
14	16		1.00	2.5–1.0	28 (43%)	5,440	64.3	194	25	104	158
15	17		1.29								
16	18	5	1.63								
17	19		1.78								
18	20		2.23	0.97–0.57	63 (55%)	14,800	104.7	235	30	131	108
19	21		2.54								
20	22	6	2.78								
21	23		2.96								
22	24		3.11	0.49–0.38	102 (55%)	28,100	156.6	275	40	202	61
23	25		3.28								
24	26	7	3.47								
25	27		3.72								
26	28		3.97	0.31–0.26	135 (54%)	42,200	191.9	313	60		
27	29		4.19	0.26						205	53
28	30	8	4.38	0.242							
29	31		4.58	0.208							
30	32		4.78	0.203	191 (61%)	72,000	184.5	377	90		
32	33		4.97	0.180							
32	34	9	5.28	0.168							
33	35		5.56	0.163							
34	36		5.81	0.157	234 (60%)	101,000	198.0	432	140		
35	37		6.04	0.145						264	52
36	38	10	6.37	0.138							
37	39		6.65	0.132							
38	40		7.23	0.130	273 (58%)	125,000	230.0	458	150	321	48

TABLE 28.3. Histomorphometrical evaluation of placental villi. The data presented here are mean data for the complete villous tree including all villous types. (1) Data from Glöde (1984); (2) data from Kaufmann (1972); (3) calculated from data from Kaufmann (1981); (4) data from Kaufmann & Scheffen (1992).

Pregnancy week (postconception)	Pregnancy week (postmenstruation)	Pregnancy month (postmenstruation)	Trophoblastic volume as a percentage of the villous volume (%) <sup>(1)</sup>	Mean trophoblastic thickness ( $\mu\text{m}$ ) <sup>(1)</sup>	Villous cytotrophoblast volume as a percentage of villous trophoblast (%) <sup>(2)</sup>	Villous stromal volume as a percentage of villous volume (%) <sup>(3)</sup>	Villous connective tissue volume as a percentage of villous volume (%) <sup>(3)</sup>	Volume of fetal vessel lumina as a percentage of villous volume (%) <sup>(3)</sup>	Percentage of the villous surface that is characterized by a double-layered trophoblast (syncytium and Langhans cells) (%) <sup>(2)</sup>	Maternofetal diffusion distance ( $\mu\text{m}$ ) <sup>(4)</sup>
1	1	1								
2	2									
3	5									
4	6	2								
5	7		31.7	18.9		68.3	65.6	2.7		55.9
6	8				30					
7	9		33.5	19.1		66.5	63.5	3.0	85	
8	10	3				65.8	61.8	4.0		
9	11		34.5	21.6						
10	12				35				80	
11	13									
12	14	4								
13	15									
14	16				43			6.0	80	40.2
15	17									
16	18	5	24.8	11.6		75.2	68.9	6.3		
17	19					35				
18	20							6.6	60	22.4
19	21									
20	22	6								
21	23									
22	24				30				55	21.6
23	25									
24	26	7								
25	27									
26	28		30.1	9.7	25	69.9	60.8	9.1	45	
27	29									
28	30	8								
29	31									
30	32				20				35	20.6
31	33									
32	34	9								
33	35									
34	36		30.8	5.2	15	69.2	47.9	21.3	25	11.7
35	37									
36	38	10								
37	39									
38	40		32.9	4.1	14	67.1	38.7	28.4	23	4.8

TABLE 28.4. Fetal and placental weights as related to the number of pregnancies.

Pregnancy no.	Male neonates				Female neonates			
	Mat. age (years)	Neo. length (cm)	Neo. wt. (g)	Plac. wt. (g)	Mat. age (years)	Neo. length (cm)	Neo. wt. (g)	Plac. wt. (g)
First ( <i>n</i> = 101)	22.37	51.04	3,141.3	548.7	22.42	50.96	2,974.0	560.7
Second ( <i>n</i> = 71)	25.24	51.95	3,442.8	607.9	25.15	51.29	3,133.6	565.5
Third ( <i>n</i> = 27)	30.21	50.29	3,495.0	603.2	26.69	50.29	3,228.1	553.9
Fourth ( <i>n</i> = 18)	30.00	53.83	3,677.5	599.6	29.00	50.83	3,095.0	568.3

Mat. = maternal; Neo. = neonatal; Plac. = placental.

All results are given as the mean value.

From Lips (1891).

TABLE 28.5. Qualitative data for the human placenta at term. The data were taken or calculated from (1) Knopp (1960); (2) Aherne &amp; Dunnill (1966); (3) Baur (1970); (4) Wördehoff (1971); (5) Laga et al. (1973); (6) Ehrhardt &amp; Gerl (1970); (7) Bacon et al. (1986); (8) Mayhew et al. (1986); (9) Bouw et al. (1976); (10) Feneley &amp; Burton (1991); (11) Voigt et al. (1978); (12) Burton &amp; Feneley (1992).

Mean placental volume (cm <sup>3</sup> ) (without cord and membranes)	448 (2)	448 (5)	540 (9)	408 (10)
Percentage of villous volume per placenta (%)	57.5 (1)	57.9 ± 5.7 (2)	45.6 (5)	62.9 ± 2.1 (7)
Mean villous volume per placenta (g)	273 (1)	214 (5)	224 (2)	239 (9)
Percentage of intervillous space per placenta (postpartum) (%)	35.8 ± 3.2 (2)	23.29 (5)	29.3 ± 2.0 (7)	37.9 (8)
Volume of intervillous space (cm <sup>3</sup> )	144 (2)	110 (5)	210 (9)	173 (8)
Percentage of intervillous fibrinoid per placenta (%)	4.3 ± 2.1 (2)	2.58 (5)		
Villous surface covered with fibrinoid (%)	0.364 (6)			
Percentage of villous tissues per placenta (chorionic plate, basal plate, septa, cell islands, infarctions) (%)	27.6 (5)			
Villous surface per placenta (m <sup>2</sup> )	13.3 ± 0.47 (4)	12.0 (3)	11.8 (6)	11.0 ± 1.3 (2)
Villous surface per 1 cm <sup>3</sup> of villous volume (cm <sup>2</sup> )	330 (1)	248 (3)	301.2 (4)	
Inner fetal capillary surface per placenta (m <sup>2</sup> )	12.2 ± 1.5 (2)	12.0 (5)		
Total length of all villi per placenta (km)	90 (1)			
Arithmetic mean thickness of villous membrane (mean maternofetal diffusion distance) (μm)	3.5 (2)	4.5 (10)	5.0 (11)	
Harmonic mean thickness of villous membrane (μm)	10.0 (5)	4.9 (8)		

TABLE 28.6. Quantitative composition of peripheral villous types of diameter of less than 80 μm following resin embedding.

Parameter	All villous types after spon. del.	After in situ puncture of placenta						
		All villous types	SV	IIV	MIV	TV, neck	TV, no neck	
Villous surface showing cytrophoblast below the syncytium (%)	27.5	22.8	21.4	23.4	24.6	20.0	23.5	
Cytrophoblastic volume as a percentage of total villous trophoblast (%)	13.3	14.0	11.7	14.3	13.8	11.6	15.7	
Percent of total villous volume	Trophoblast	41.7	37.7	30.0	33.8	38.0	46.2	29.6
	Villous stroma	58.3	62.3	70.0	66.2	62.0	53.8	70.4
	Fetal vessel lumens	20.0	28.4	26.0	15.7	21.0	20.2	45.2
	Connective tissue including vessel walls	38.4	33.9	44.0	50.5	41.0	33.6	25.2
Mean trophoblastic thickness	4.7	4.1	4.7	5.2	4.4	4.0	3.3	
Distribution of maternofetal diffusion distances in % of villous surface	-0-2 μm	10.4	21.0	9.2	10.9	13.1	21.8	37.1
	2.1-5 μm	29.9	40.2	27.3	27.3	37.5	41.4	36.5
	5.1-10 μm	40.3	32.4	50.5	37.7	37.8	32.6	25.1
	10.1-∞ μm	19.5	6.4	13.0	24.1	11.6	4.4	1.3
Mean maternofetal diffusion distance (μm)	7.1	5.0	6.8	7.5	6.0	4.8	3.7	
Macrophages (no.)/mm <sup>2</sup> of histologic section		24	730	241	83		192	
Mast cells (no.)/mm <sup>2</sup> of histologic section		18	10	5	<1		<1	

Spon. del. = spontaneous delivery; SV = stem villi; IIV = immature intermediate villi; MIV = mature intermediate villi; TV, neck = neck regions of terminal villi; TV, no neck = terminal villi, excluding neck regions.

The data obtained from spontaneous deliveries (fixation about 10 minutes after cord clamping, ischemic period far above 10 minutes), are compared with such obtained by puncture aspiration out of the still maternally perfused in situ placenta (ischemic period: 0 minutes) (Voigt et al., 1978). In addition, data have been separately measured and calculated for different villous types (Sen et al., 1979).

TABLE 28.7. Number of villous cross sections (%), percentages of villous surface, and percentages of villous volume in different groups of villi.

Parameter	Villous cross sections (% of total no.)	Surface of villi (% of total)	Volume of villi (% of total)	Type of villus	Mean caliber (μm)	Mean length of cross sections (μm)
<b>Diameter of villi (μm)</b>						
>1,500	0.003	0.1	1.2			
1,201–1,500	0.002	0.1	0.8	Trunci chorii		
901–1,200	0.007	0.1	1.4			
601–900	0.02	0.5	2.4	Rami chorii		
301–600	0.2	1.9	9.0			
226–300	0.3	1.7	6.4	Ramuli chorii, mature and immature intermediate villi, terminal villi <sup>a</sup>		
151–225	0.6	3.2	7.7			
76–150	7.1	15.1	20.8			
0–75	91.8	77.2	50.4			
<b>Type of villus</b>						
Ramuli	5.1	12.4	22.7		120.5	340.1
Immature intermediate	5.1	8.4	10.1		76.6	233.5
Mature intermediate	29.1	31.0	27.8		60.6	153.5
Terminal	55.1	46.3	38.7		50.8	118.4
Neck regions	5.5	2.0	0.8		34.3	46.4

<sup>a</sup>The villi are classified according to diameter (using data from I. Hansen & V. König, personal communication, 1978) and compared according to structure.

Modified from Sen et al. (1979).

TABLE 28.8. Influence of cord clamping on placental structure.

Parameter	Early clamped (n = 9)	Late clamped (n = 7)
Placental weight (g)	567.4 ± 114.5	407.1 ± 74.0
Birth weight (g)	3,593.5 ± 380.0	3,404.3 ± 313.5
Placental weight index	0.156 ± 0.021	0.116 ± 0.088
Villous volume (cm <sup>3</sup> )	239.1 ± 51.9	185.3 ± 39.5
Villous vessel volume (cm <sup>3</sup> )	74.9 ± 28.2	33.4 ± 18.3
Villous vessel volume (%)	30.4 ± 5.8	17.4 ± 6.9
Villous vessel surface (mm <sup>2</sup> ) per mm <sup>3</sup> of villous volume	66.9 ± 10.0	46.3 ± 10.5
Total villous surface (m <sup>2</sup> )	13.3 ± 2.6	9.3 ± 2.1
Volume of intervillous space (cm <sup>3</sup> )	210.3 ± 50.0	116.0 ± 21.7

Results are given as the mean ± SD. A group of spontaneously delivered placentas of which the cord was clamped as early as possible as compared with a group of which the cord was clamped after cessation of the arterial pulsation.

Data from Bouw et al. (1976).

TABLE 28.9. Influence of total ischemia of varying length on the structure of terminal villi.

Parameter	Ischemic period				
	0 min	2 min	5 min	10 min	20 min
Fetal capillary volume (%)	45	32	19	13	17
Connective tissue volume (%)	25	42	49	54	47
Trophoblastic volume (%)	23	26	32	33	36
Mean maternofetal diffusion distance (μm)	3.2	3.7	5.2	7.1	7.4
Epithelial plates (percentage of villous surface) (%)	40.4	—	—	—	14.7
Mean mitochondrial diameter (syncytiotrophoblast) (μm)	0.30	0.34	0.51	0.63	0.61

Data from Kaufmann (1985).

TABLE 28.10. Influence of total osmolarity of the fixative (2.2% phosphate-buffered glutaraldehyde) on the volumes of syncytiotrophoblast and fetal capillary lumina in the guinea pig placenta.

Parameter	Osmolarity of the fixative				
	235	290	340	390	600
Trophoblastic volume (%)	39.5	39.3	33.3	32.7	24.6
Fetal capillary volume (%)	12.1	14.7	25.5	16.9	19.3
Extracellular space (%)	<1.0	<1.0	<1.0	2.3	11.7
Mean mitochondrial diameter (syncytiotrophoblast) (μm)	0.68	0.56	0.30	0.26	0.25

Data from Kaufmann (1980) can be transferred to the human.

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