

Different cytotypes in fishes of the genus *Hypostomus* Lcépède, 1803, (Siluriformes: Loricariidae) from Xingu river (Amazon region, Brazil)

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Abstract. We analyzed the karyotypes of three specimens of fishes of the genus *Hypostomus* Lacépède, 1803 (Loricariidae) from Xingu River (Amazon region). We used conventional staining techniques, including C-banding, Ag-NOR staining, CMA₃- and DAPI-banding. Each specimen had a different cytotype: *Hypostomus* sp. Xingu-1 had 2n=64 (32M/SM, 32ST/A); *Hypostomus* sp. Xingu-2 has 2n=66 (32M/SM, 34ST/A), and *Hypostomus* sp. Xingu-3 had 2n=65 (38M/SM, 26ST/A + 1 B). The three cytotypes showed similar C-, CMA₃- and DAPI-banding patterns. The nucleolus organizing regions were located in the short arm of chromosome pair 25 of *Hypostomus* sp. Xingu-1 and pair 29 of *Hypostomus* sp. Xingu-2, and in the long arm of pair 30 of *Hypostomus* sp. Xingu-3, probably because of a pericentric inversion. A fusion/fission rearrangement explains the difference in the diploid number and number of M/SM and ST/A chromosomes between the 2n= 64 and 2n=66 cytotypes. The B chromosome most probably explains the difference between the 2n= 64 and 2n=65 cytotypes. The cytotype with 2n=65 had a significantly larger number of M/SM chromosomes, probably because of pericentric inversions. These three cytotypes may represent different species.

Key words: Siluriformes, *Hypostomus*, cytogenetics, heterochromatin, variability, C-banding, Ag-NOR, CMA₃ and DAPI.

INTRODUCTION

The Loricariidae is one of the most diversified families within the order Siluriformes. It includes more than 600 species that may be divided into five subfamilies: Delturinae, Neoplecostominae, Hypoptopomatinae, Loricariinae and Hypostominae. These subfamilies

are endemic to the Neotropical region with distributions extending from Panama to Uruguay (Isbrücker, 1980; Reis et al., 2003, 2006; Armbruster, 2004).

Using morpho-osteological analysis, Schaefer (1987) suggested that the subfamily Hypostominae is the most derived among the

Loricariidae. The same conclusion was made by Artoni, Bertollo (1996) using cytogenetic information. The subfamily Hypostominae includes numerous species that are broadly distributed throughout almost all rivers and lakes in South America (Armbruster, 2004; Alves et al., 2006). According to Armbruster (2004), this subfamily can be divided into five tribes: Corymbophanini, Rhinelepini, Hypostomini, Pterygoplichthini and Ancistrini. The tribe Hypostomini contains a single genus, *Hypostomus* Lacépède, 1803, which comprises 116 species and is found in most South American waters (Isbrücker, 1980).

The subfamily Hypostominae includes the largest number of karyotyped species within the Loricariidae (Table 1). The species of the Hypostominae show significant variations in chromosome number and morphology. The diploid number (2n) ranges from 52 to 80 chromosomes (Artoni, Bertollo, 1996, 2001). An XX/XY sex chromosome system has been described for some species (Michele et al., 1977), whereas others have a ZZ/ZW system (Artoni, 1996). The nucleolus organizer region

(NOR) can be found as a single (Artoni, Bertollo, 1996, 2001; Artoni et al., 1998; Alves et al., 2006) or multiple pairs system (Artoni, 1996; Artoni, Bertollo, 1996; Alves et al., 2006). The composition of constitutive heterochromatin (CH) may be species-specific in *Hypostomus* with some blocks rich in A-T base pairs and others rich in G-C base pairs (Artoni, Bertollo, 1999).

In the present work, we describe the karyotypes of three specimens of *Hypostomus* from the Xingu River in the Amazon region of Brazil.

MATERIAL AND METHODS

The specimens were collected in the Amazon region, from the Xingu River in the town of Altamira (Para, Brazil; 03°12'48.0"S/52°12'41.7"W). Two females and a male were analyzed.

Because members of *Hypostomus* are very difficult to identify at the species level, we were unable to determine the species of our samples. Therefore, we called them

Table 1. Cytogenetic data of fishes of the genus *Hypostomus* in Brazil. 2n - diploid number, M - metacentric, SM - submetacentric, ST - subtelocentric, A - acrocentric, p - short arm, q - long arm, (*) - species with more than one karyotype, N - number of NOR bearing chromosomes.

Species	Locality	2n	Classification			Sex System	NOR		References
			M	SM	ST/A		N°	Position	
<i>Hypostomus</i> sp. A	Rincão stream, Sao Paulo	70	18	14	38	–	4	q/A; p/M	Artoni, Bertollo, 1996
<i>Hypostomus</i> sp. B	Mogi-Guaçu river, Sao Paulo	72	12	18	42	–	2	q/ST-A	Artoni, Bertollo, 1996
<i>Hypostomus</i> sp. B (*)	Mogi-Guaçu river, Sao Paulo	72	13	18	41	–	2	–	Artoni, Bertollo, 1999
<i>Hypostomus</i> sp. C	Mogi-Guaçu river, Sao Paulo	72	10	18	44	–	3	q/A	Artoni, Bertollo, 1996
<i>Hypostomus</i> sp. D ₁	Mogi-Guaçu river, Sao Paulo	72	10	26	36	–	2	q/ST	Artoni, Bertollo, 1996
<i>Hypostomus</i> sp. D ₂	Mogi-Guaçu river, Sao Paulo	72	14	20	38	–	2	q/A	Artoni, Bertollo, 1996
<i>Hypostomus</i> sp. E	Mogi-Guaçu river, Sao Paulo	80	8	16	56	–	4	p/SM; p/ST	Artoni, Bertollo, 1996

Table 1. (Continuation).

Species	Locality	2n	Classification			Sex System	NOR		References
			M	SM	ST/A		Nº	Position	
<i>Hypostomus</i> sp. F	São Francisco river, Minas Gerais	76	10	16	50	–	4	q/SM; p/ST	Artoni, 1996
<i>Hypostomus</i> sp. F (*)	São Francisco river, Minas Gerais	75	10	17	48	–	–	–	Artoni, Bertollo, 1999
<i>Hypostomus</i> sp. G	Fundo brook, Araguaia river, Mato Grosso	64	14	24	26	ZZ/ZW	2	p/SM	Artoni et al., 1998
<i>Hypostomus albopunctatus</i>	Mogi-Guaçu river, Sao Paulo	74	10	20	44	–	4	q/A; p/M	Artoni, Bertollo, 1996
<i>Hypostomus ancistroides</i>	–	68	10	28	30	–	–	–	Michele et al., 1977
<i>Hypostomus ancistroides</i>	Mogi-Guaçu river, Sao Paulo; Monjolinho brook, Sao Paulo	68	16	18	34	–	6	p/M; p/SM; p/A	Artoni, Bertollo, 1996
<i>Hypostomus ancistroides</i>	Araquá river, Sao Paulo	68	18	10	40	–	6	p/M; p/SM; p/SM	Alves et al., 2006
<i>Hypostomus affinis</i>	Jacuí brook, Sao Paulo	66	14	14	38	–	–	–	Kavalco et al., 2005
<i>Hypostomus prope auroguttatus</i>	Mogi-Guaçu river, Sao Paulo	76	8	30	38	–	2	q/A	Artoni, Bertollo, 1996
<i>Hypostomus emarginatus</i>	Araguaia river, Mato Grosso	52	16	30	6	–	2	q/M	Artoni, Bertollo, 2001
<i>Hypostomus goyazensis</i>	Vermelho river, Goiás	72	10	16	46	–	2	p/ST	Alves et al., 2006
<i>Hypostomus macrops</i>	–	68	10	14	44	XX/XY	–	–	Michele et al., 1977
<i>Hypostomus nigromaculatus</i>	Mogi-Guaçu river, Sao Paulo	76	8	20	48	–	2	p/ST; q/A	Rubert et al., 2008
<i>Hypostomus paulinus</i>	–	74	10	20	44	–	–	–	Michele et al., 1977
<i>Hypostomus plecostomus</i>	–	54	36		18	–	–	–	Muramoto et al., 1968
<i>Hypostomus regani</i>	Mogi-Guaçu river, Sao Paulo	72	10	20	42	–	4	q/A; p/ST	Artoni, Bertollo (1996)
<i>Hypostomus regani</i>	Araquá river, Sao Paulo	72	12	18	42	–	4	q/A; q/A	Alves et al., 2006
<i>Hypostomus strigaticeps</i>	–	74	8	4	62	–	–	–	Michele et al., 1977
<i>Hypostomus</i> sp. 1	Quinta brook, Sao Paulo	72	–	–	–	–	–	–	Fenerich, 1998
<i>Hypostomus</i> sp. 2	Alambari brook, Sao Paulo	68	–	–	–	–	–	–	Fenerich, 1998
<i>Hypostomus</i> sp. 3	Paranapanema river, Sao Paulo	66	–	–	–	–	–	–	Fenerich, 1998
<i>Hypostomus</i> sp. 4	Hortelã brook, Sao Paulo	76	–	–	–	–	–	–	Fenerich, 1998
<i>Hypostomus</i> sp. Xingu-1	Xingu river, Para	64	16	16	32	–	2	p/A	Present study
<i>Hypostomus</i> sp. Xingu-2	Xingu river, Para	66	18	14	34	–	2	p/A	Present study
<i>Hypostomus</i> sp. Xingu-3	Xingu river, Para	65	15	23	26+1B	–	2	q/A	Present study

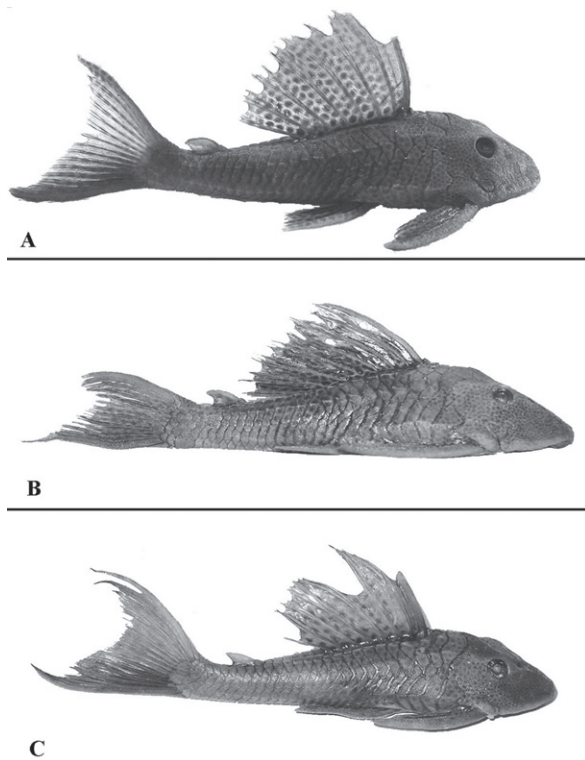


Fig. 1, a-c. a - *Hypostomus* sp. Xingu-1. b - *Hypostomus* sp. Xingu-2. c - *Hypostomus* sp. Xingu-3.

Hypostomus sp. Xingu-1, *Hypostomus* sp. Xingu-2 and *Hypostomus* sp. Xingu-3.

Chromosome preparation was performed as described in Bertollo et al. (1978). The chromosomes were classified as described in Guerra (1986). We used conventional staining techniques, including C-banding (Sumner, 1972), Ag-NOR staining (Howell, Black, 1980), and the fluorochromes chromomycin A₃ (CMA₃) (Schweizer, 1980) and 4'6'-diamidino-2-phenylindole (DAPI) (Piecarka et al., 2006).

The fish specimens studied were vouchered at the Museu Paraense Emilio Goeldi (MPEG) under the following numbers: *Hypostomus* sp. Xingu-1, MPEG 13431; *Hypostomus* sp.

Xingu-2, MPEG 13430; and *Hypostomus* sp. Xingu-3, MPEG 13429.

RESULTS

Hypostomus sp. Xingu-1, a female (Fig. 1, a), shows $2n=64$ and a karyotype formula (KF) with $32M/SM+32ST/A$ (Fig. 2, a). *Hypostomus* sp. Xingu-2, a male (Fig. 1, b), shows $2n=66$ and KF with $32M/SM+34ST/A$ (Fig. 3, a). *Hypostomus* sp. Xingu-3, a female (Fig. 1, c), shows $2n=65$, KF with $38M/SM+26ST/A + 1 B$ microchromosome (Fig. 4, a).

Hypostomus sp. Xingu-1 has blocks of CH in the distal region of both arms of metacentric pair 14, in the proximal region of the long arm and the distal region of the short arm of submetacentric pair 6, and at the distal region of the long arm of pair 25 (Fig. 2, B). *Hypostomus* sp. Xingu-2 has evident heterochromatic blocks found in the distal regions of both arms of metacentric pair 13 and in the distal region of the long arm of pair 29. There is a size heteromorphism of the CH block in the proximal region of the long arm of pair 4 (Fig. 3, B). In *Hypostomus* sp. Xingu-3 chromosome pair 9 is heteromorphic: one of the homologues is metacentric and the other is submetacentric, due to a heteromorphism of the heterochromatin in the proximal region of the long arm: there is an interstitial heterochromatic block in only one of the homologues (Fig. 4, b). Chromosome pair 5 has small blocks of heterochromatin in the distal regions of both arms (Fig. 4, b).

In *Hypostomus* sp. Xingu-1, the NOR is of the simple type, and it is located on the distal short arm of pair 25 (Fig. 5, a). Heteromorphism in the NOR size was found, and the NOR is C-band negative. In *Hypostomus* sp. Xingu-2, the NOR is present in the short arm of pair 29 (Fig. 5, b). In *Hypostomus* sp. Xingu-3 the NOR is found

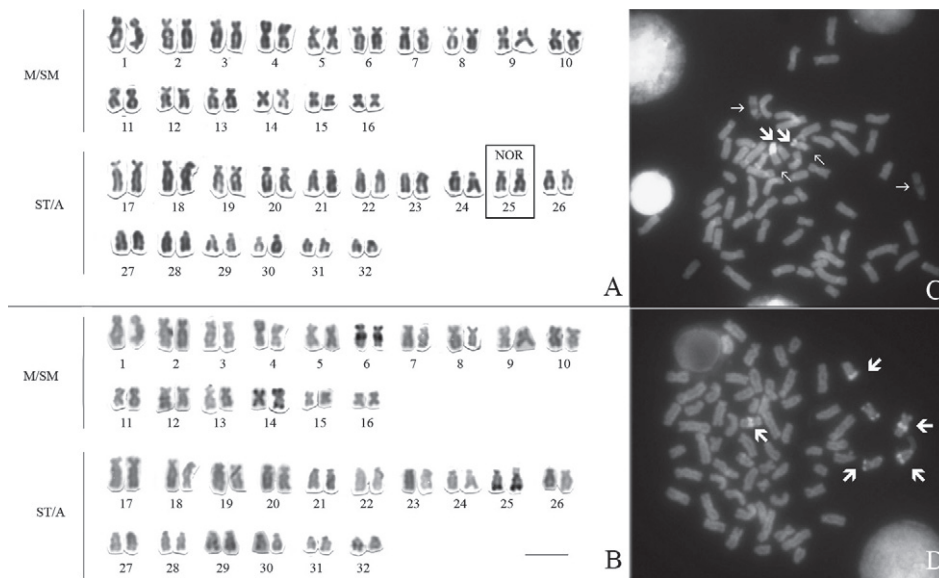


Fig. 2, a-d. Karyotype of *Hypostomus* sp. Xingu-1. **a** - Giemsa staining, showing the NOR-bearing pair. **b** - C-banded karyotype. **c** - metaphase spread stained with CMA₃ (larger arrows show the NOR region, smaller arrows show the CMA₃-quenched regions). **d** - DAPI-banding (arrows show the positive C-banded regions that are A-T rich). Bar = 10 μm.

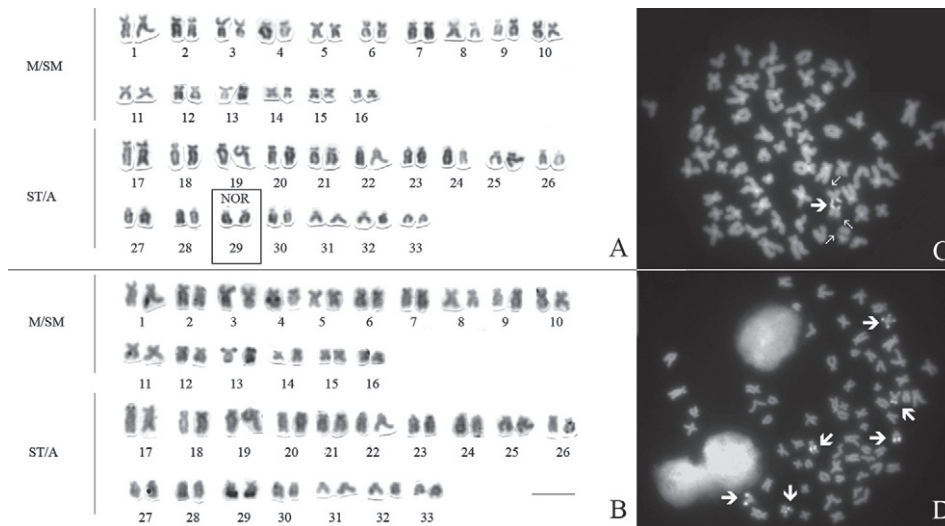


Fig. 3, a-d. Karyotype of *Hypostomus* sp. Xingu-2. **a** - Giemsa staining, showing the NOR-bearing pair. **b** - C-banded karyotype. **c** - metaphase spread stained with CMA₃ (larger arrows show the NOR region, smaller arrows show the CMA₃-quenched regions). **d** - DAPI-banding (arrows show the positive C-banded regions that are A-T rich). Bar = 10 μm.

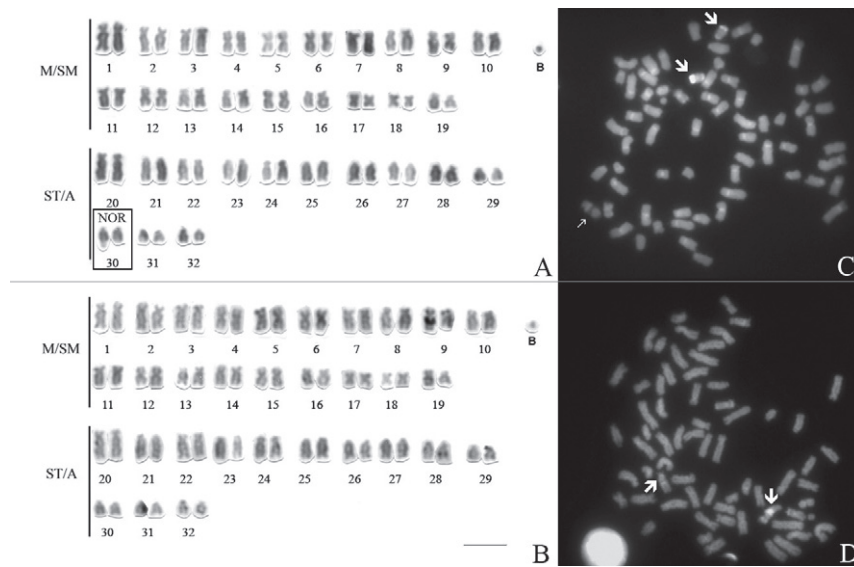


Fig. 4, a-d. Karyotype of *Hypostomus* sp. Xingu-3. **a** - Giemsa staining, showing the NOR-bearing pair (B represents the minichromosome). **b** - C-banded karyotype. **c** - metaphase spread stained with CMA₃ (larger arrows show the NOR region, smaller arrows show the CMA₃-quenched regions). **d** - DAPI-banding (arrows show the positive C-banded regions that are A-T rich). Bar = 10 μ m.

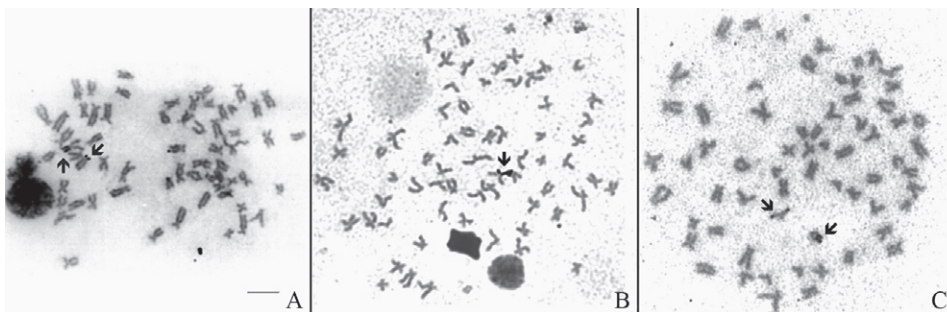


Fig. 5, a-c. Metaphases submitted to Ag-NOR technique. The arrows show the NOR-bearing chromosomes. **a** - *Hypostomus* sp. Xingu-1. **b** - *Hypostomus* sp. Xingu-2. **c** - *Hypostomus* sp. Xingu-3. Bar = 10 μ m.

at the distal region of the long arm of pair 30 and shows size heteromorphism (Fig. 5, c).

The heterochromatic blocks of pairs 6, 14 and 25 produce bright DAPI fluorescence (Fig. 2, d) in *Hypostomus* sp. Xingu-1, whereas the heterochromatic blocks of pairs 14 and 25 have quenched CMA₃ fluorescence. As they also are DAPI bright, these regions are likely

to be rich in A-T base pairs. In *Hypostomus* sp. Xingu-2 the heterochromatic blocks of pairs 4, 13 and 29 are DAPI-bright (Fig. 3, d). CMA₃ staining is bright at the centromeres and at the NOR (Fig. 3, c), while the CH blocks of pairs 4 and 29 are CMA₃-quenched (Fig. 3, c). *Hypostomus* sp. Xingu-3 has DAPI staining bright in the heterochromatin blocks of pairs

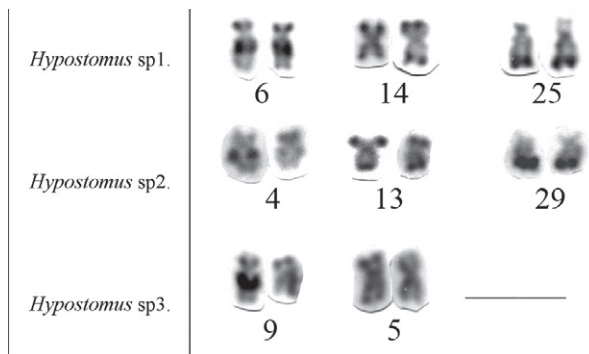


Fig. 6. Probable homologies among the specimens studied herein, as suggested by C-banding. Bar = 10 μ m.

5 and 9 (Fig. 4, d). CMA₃ is bright in most of the centromeres and in the NOR (Fig. 4, c), whereas the above mentioned heterochromatic block in pair 9 is CMA₃-quenched (Fig. 4, c). The additional small chromosome (a “B” or supernumerary chromosome) is neither heterochromatic nor bright with DAPI or CMA₃ staining.

DISCUSSION

A comparative analysis of the three karyotypes suggests that a fusion/fission-type rearrangement could explain the differences in the 2n and in the number of M/SM and ST/A chromosomes between the cytotypes with 2n=64 and 2n=66. The presence of a B chromosome explains the difference in the 2n of the cytotypes with 2n=65 and 2n=64. In addition, the cytotype with 2n=65 has more M/SM chromosomes than the other cytotypes, probably because of pericentric inversions.

When comparing our results with prior descriptions in the literature, we have found that *Hymostomus* sp. Xingu-1 (2n=64; 32M/SM+32ST/A) has the same diploid number as *Hymostomus* sp. G (2n=64; 14M+24SM+26ST/A, Artoni et al., 1998), while *Hymostomus* sp. Xingu-2 (2n=66; 32M/

SM+34ST/) has the same diploid number as *H. affinis* Steindachner, 1877 (2n=66; 14M+14SM+38ST/A, Kavalco et al., 2005). However, the karyotypic formulae differ in both cases, probably because of pericentric inversions, which can change chromosome morphology without changing the diploid number.

The third cytotype, *Hymostomus* sp. Xingu-3 (2n=65; 38M/SM+26ST/A + 1 B) has a karyotype similar to that of *Hymostomus* sp. G, described above. However, *Hymostomus* sp. Xingu-3 differs from *Hymostomus* sp. G in the heteromorphism of pair 9 and the presence of a B chromosome. Notably, this is the first report of a B chromosome in *Hypostomus*. This comparison suggests that *Hymostomus* sp. Xingu-3 is more similar to *Hymostomus* sp. G from the Araguaia River (which is linked to the Amazon basin), than to the other two cytotypes obtained from the Xingu River.

All the three specimens have NORs of a simple type located respectively on the short arms of pairs 25 and 29, and the long arm of pair 30. The difference in the NOR position in *Hymostomus* sp. Xingu-3 can be explained by a pericentric inversion. In fishes, NORs are often found on a single pair of chromosomes, as seen in *Hypostomus* (Artoni, Bertollo 1996, 2001; Artoni et al., 1998, Alves et al., 2006), although there are many inter- and intraspecific deviations from this trend (Oliveira, 1987; Andreatta, 1991; Fenocchio, 1993; Artoni, 1996; Souza, 2003). Artoni (1996) claimed that most of the Hypostominae species with a single pair of NORs have heteromorphism in NOR size. This is consistent with the pattern we found in *Hymostomus* sp. Xingu-1 and *Hymostomus* sp. Xingu-3 (Fig. 5).

The NORs showed bright CMA₃ banding, indicating that these sequences are G-C rich. This fluorochrome stains regions independently of the transcriptional activity in the previous interphase (Howell, 1977; Almeida-Toledo, 1998).

Hypostomus has relatively little constitutive heterochromatin (Artoni, 1996). The cytotypes studied by us have very similar patterns of heterochromatin distribution, with only few chromosome pairs showing small heterochromatic blocks. It is possible to compare the homologies among the pairs based on their heterochromatin patterns (Fig. 6). Our CMA₃ and DAPI staining results suggest that the three cytotypes have a heterogeneous composition of constitutive heterochromatin, with some blocks rich in A-T base pairs (DAPI bright) and other rich in G-C base pairs (CMA₃ bright). The same result was found in some species of *Hypostomus* by Artoni and Bertollo (1999). However, these results differ from that obtained by Kavalco et al. (2005), who demonstrated that *H. affinis* has its CH exclusively rich in GC pairs.

According to Camacho et al. (2000) the B chromosomes are typically heterochromatic. Andreato et al. (1993) described two large heterochromatic B chromosomes in *Microlepidogaster leucofrenatus* (Miranda Ribeiro, 1908) (Siluriformes: Loricariidae). In our study the B-chromosome was neither heterochromatic nor especially rich in AT or GC-bp. More detailed molecular studies may help to understand its nature.

Artoni and Bertollo (2001) proposed that the karyotypes with 2n=54 should be an ancestral condition in *Hypostomus* karyotypes, since this diploid number is found in different genera and even families. As a consequence, centric fissions must be important rearrangements for the evolution of karyotypes of *Hypostomus*, in which the diploid numbers range from 2n=52 to 2n=80 (Table 1). Also, the number of ST/A chromosomes increases in karyotypes with higher diploid numbers, with a consequent reduction in the number of M/SM chromosomes. Kavalco et al. (2005) suggest that Robertsonian rearrangements, as well as

pericentric inversions, were the main changes related with the karyotypic diversification of the Hypostominae. Our data are in agreement with this suggestion, since the differences among the three karyotypes studied can be a consequence of these rearrangements.

Despite the differences in diploid numbers and karyotypic formulae, the three cytotypes share similar patterns of heterochromatin distribution and the presence of a single NOR. These results suggest that: a) the three specimens could be members of a single species that maintains the observed chromosome differences as polymorphisms; b) the population could be undergoing a chromosome-based speciation event through which individuals are diverging from a common ancestor; or c) the specimens may represent members of different species that arose from a common ancestor. Additional studies will be required to distinguish among these possibilities, but at the moment the option "c" seems to be the most reasonable. The lack of cytogenetic information and the lack of knowledge of most of the species morphology (see Table 1, where many *Hypostomus* are "sp.") turns the *Hypostomus* genus one of the most interesting for any kind of study. New information is always helpful, like that described here.

ACKNOWLEDGEMENTS

This research was supported by CNPq, CAPES and UFFa. The sample collection was authorized by IBAMA (Instituto Brasileiro do Meio Ambiente) permission 020/2005 (IBAMA registration: 207419).

REFERENCES

- Almeida-Toledo L. F. 1998.** Cytogenetic markers in neotropical freshwater fishes, (pp. 583-588) // Malabarba L.R., Reis R.E., Vari R.P., Lucena Z.M.S., Lucena C.A.S. (Eds). Phylogeny and Classification

- of Neotropical Fishes. Porto Alegre. 540 p.
- Alves L.A., Oliveira C., Nirchio M., Granado A., Foresti F. 2006.** Karyotypic relationships among the tribes of Hypostominae (Siluriformes: Loricariidae) with description of XO sex chromosome system in a Neotropical fish species // *Genetica*. 128: 1–9.
- Andreato A.A. 1991.** Estudos citogenéticos na subfamília Hypoptopomatinae (Pisces, Siluriformes, Loricariidae). Unpublished Mastership Dissertation. São Paulo, Universidade de São Paulo. 171 p.
- Andreato A.A., Almeida-Toledo L.F., Oliveira C., Almeida-Toledo Filho S. 1993.** Chromosome studies in Hypoptopomatinae (Pisces, Siluriformes, Loricariidae) // *Cytogenet. Cell Genet.* 63: 215–220.
- Armbruster J.W. 2004.** Phylogenetic relationships of the suckermouth armoured catfishes (Loricariidae) with emphasis on the Hypostominae and the Ancistrinae // *Zool. J. Linnean Soc.* 141: 1–80.
- Artoni R.F. 1996.** Estudos Citogenéticos na Família Loricariidae, com ênfase no gênero *Hypostomus* Lacépède (1803) (Pisces, Siluriformes). Unpublished Mastership Dissertation, São Carlos, SP. 162 p.
- Artoni R.F., Bertollo L.A.C. 1996.** Cytogenetic studies on Hypostominae (Pisces, Siluriformes, Loricariidae): Considerations on karyotype evolution in the genus *Hypostomus* // *Caryologia*. 49: 81–90.
- Artoni R.F., Bertollo L.A.C. 1999.** Nature and distribution of constitutive heterochromatin in fishes, genus *Hypostomus* (Loricariidae) // *Genetica*. 106: 209–214.
- Artoni R.F., Bertollo L.A.C. 2001.** Trends in the karyotype evolution of Loricariidae Fish (Siluriformes) // *Hereditas*. 134: 201–210.
- Artoni R.F., Vênere P.C., Bertollo L.A.C. 1998.** A heteromorphic ZZ/ZW sex chromosome system in fish, genus *Hypostomus* (Loricariidae) // *Cytologia*. 63: 421–425.
- Bertollo L.A.C., Takashi C.S., Moreira-Filho O. 1978.** Cytotaxonomic considerations on *Hoplias lacerdae* (Pisces, Erythrinidae) // *Brazilian J. Genet.* 1: 103–120.
- Camacho J.P.M., Sharbel T.F., Beukeboom L.W. 2000.** B-chromosome evolution. *Phil. Trans. Soc. Lond. B.* 355: 163–178.
- Fenerich P.C. 1998.** Estudo de conteúdo de DNA nuclear em células de peixes da ordem Siluriformes (Pisces, Ostariophysi), da região de Botucatu, SP. MSc Thesis, Universidade Estadual Paulista. 127 p.
- Fenocchio A.S. 1993.** Cromossomos supranumerários no gênero *Rhamdia* (Pisces). Caracterização cromossômica e considerações sobre evolução cariotípica em Siluroidei. Unpublished Ph.D Thesis, Universidade de São Paulo, Ribeirão Preto, SP. 122 p.
- Guerra M.S. 1986.** Reviewing the chromosome nomenclature of Levan et al. // *Rev. Brazilian Genet.* 4: 741–743.
- Howell W.M. 1977.** Visualization of ribosomal gene activity: Silver stains proteins associated with rRNA transcribed from oocyte chromosomes // *Chromosoma*. 62: 361–367.
- Howell W.M., Black D.A. 1980.** Controlled silver-staining of nucleolus organizer regions with a protective colloidal developer: a 1-step method // *Experientia*. 36: 1014–1015.
- Isbrücker I.J.H. 1980.** Classification and catalogue of the mailed Loricariidae (Pisces, Siluriformes). Verslagen en Technische Gegevens, Universiteit van Amsterdam, 22. 188 p.
- Kavalco K.F., Pazza R., Bertollo L.A.C., Moreira-Filho O. 2005.** Karyotypic diversity and evolution of Loricariidae (Pisces, Siluriformes) // *Heredity*. 94: 180–186.
- Michele J.L., Takahashi C.S., Ferrari I. 1977.** Karyotypic study of some species of the family Loricariidae // *Cytologia*. 42: 539–546.
- Muramoto I., Ohno S., Atkin N.B. 1968.** On the diploid state of the fish order Ostariophysi // *Chromosoma*. 24: 59–66.
- Oliveira C. 1987.** Estudos citogenéticos no gênero *Corydoras* (Pisces, Siluriformes, Callichthyidae). Unpublished Mastership Dissertation, Universidade de São Paulo, São Paulo, SP. 154 p.
- Pieczarka J.C., Nagamachi C.Y., Souza A.C.P., Milhomem S.S.R., Castro R.R., Nascimento A.L. 2006.** An adaptation to DAPI-banding to fishes chromosomes // *Caryologia*. 59: 43–46.
- Reis R.E., Kullander S.O., Ferraris Jr. C.J. 2003.** Check List of the Freshwater Fishes of South America. Edipucrs, Porto Alegre. 729 p.
- Reis R.E., Pereira E.L., Armbruster J.W. 2006.** Delturinae, a new loricariid catfish subfamily (Teleostei, Siluriformes), with revisions of *Delturus* and *Hemipsilichthys* // *Zool. J. Linnean Soc.* 147: 277–299.
- Rubert M., Zawadzki C.H., Giuliano-Caetano L. 2008.** Cytogenetic characterization of *Hypostomus nigromaculatus* (Siluriformes: Loricariidae) // *Neotropical Ichthyol.* 6: 93–100.
- Schaefer S.A. 1987.** Osteology of *Hypostomus plecostomus* (Linnaeus), with a phylogenetic analysis of the loricariid subfamilies (Pisces Siluroidei) // *Natur. Hist. Mus. Los Angeles County, Contrib. Sci.* 394: 1–31.
- Schweizer D. 1980.** Simultaneous fluorescent staining of R bands and specific heterochromatic regions

(DA/DAPI Bands) in human chromosomes // *Cytogenet. Cell Genet.* 27: 190-193.

- Souza A.C.P. 2003.** Descrição cariotípica de peixes dos gêneros *Baryancistrus*, *Parancistrus*, *Peckoltia* e *Ancistrus* (Ancistrinae, Loricariidae) da bacia Amazônica. Belém, PA. Unpublished Mastership Dissertation, Museu Paraense Emílio Goeldi, Universidade Federal do Pará. 130 p.

Sumner A.T. 1972. A simple technique for demonstrating centromeric heterochromatin // *Exp. Cell Res.* 75: 304-306.

Received December 18, 2009.

Accepted by V.A. Lukhtanov, March 16, 2010.

Published July 09, 2010.