

Classification of Aphid on The Basis of Their Taxonomic Features

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Abstract: The experiment was conducted at Quetta to classification of different aphids and their host in Balochistan. Aphids were collected randomly from different localities of Quetta such as Baleli, Shaikhmanda, Sariab and Brewrey, etc., during year 2013-2014 and identified them up to the specific level by running the keys of R.L. Blackman and V.F. Eastop. In the whole 5 aphids were identified on the basis of their antennal tubercles, length of third antennal segment, the basal and terminal process of last antennal segment, length of hind tarsus & number of hairs on it, shape or size of cauda and siphunculi. Several host plants were found being attacked by these identified aphids. In view of the greater variation in characters of the specimens in this area, non-availability of detailed descriptions, the keys have been constructed in order to make the future identification easier.

Keywords: classification of aphids.

INTRODUCTION

Aphids (Hemiptera: Aphididae) are an important group of insects with worldwide distribution. They are one of the important vegetable pests (Capinera; 2002). Aphids have small, soft-bodies mostly in range between 1.5 to 3.5mm in length (Blackman and Eastop; 2000). Aphids are feeding insects causing direct or indirect mechanical and economical damage as they feed on plant's sap with piercing sucking mouthparts; aphids also serve as the largest group of plant virus's vector (Blackman & Eastop; 2007, Chan et al.; 1991). Although they have extraordinary horticultural damaging effects, some of society's early interest involved aphid's beneficial or positive attributes. Galls of certain aphid species have been used for centuries in medicine, tanning, and dyeing (Fagan; 1918). Successful use of Aphidiinae as biocontrol agents is affected by the knowledge about their taxonomy, host selection behavior, and ecology (Powell; 1994, Rehman and Powell; 2010), although they are more common in temperate zones (Blackman & Eastop; 2000). Some of the aphid species are significant invasive pests, threatening the agricultural ecosystems worldwide (Capinera; 2002, Blackman & Eastop; 2006).

Aphids have experienced some adaptations in relation to host plants so that many aphid taxa have biologically complex life cycles (Martin & Brown; 2008). Most aphid species comprise a set of closely related populations which may have diverged genetically so that they could be considered as host races, developing or sibling species (or subspecies) (Blackman & Eastop; 2007). The genus *Aphis* is one of the most difficult genera of aphids to identify because of morphological conservation and this problem is complicated by the fact that several species may live on a single host plant. Several authors in various countries have

studied the taxonomy and morphology of *Aphis* species and in some cases have described several new species (Nieto Nafria et al.; 1999, Heie; 2000, Mier Durante et al.; 2003, Perez Hidalgo & Nieto Nafria; 2004, Voegtlin et al.; 2004). Aphid classification at various levels has been attempted using morphological characters, host-plants associations and life cycles (Heie; 1987, Blackman & Eastop; 1994, Shaposhnikov et al.; 1998, Kim et al.; 2010, 2011). However, morphological studies very often tend to be uncertain. Many aphids' characters are difficult to understand; in many cases, it is hard to define whether they are ancestral (plesiomorphes) or derived (apomorphies) (Heie; 2009).

Morphotaxonomy can be regarded as a reliable and powerful tool among applied entomologists for discrimination of aphid taxa (Poulios et al.; 2007). According to certain morphological characteristics, e.g, the 3-faceted eyes observed in aptera, the reduced antennal segments, and vestigial siphunculi (Zhang et al.; 1999). Different factors contribute to morphological diversity among aphids, including effects of environmental and biological factors (Margaritopoulos et al.; 2000). Aphids associated with cereals are destructive pests and are a limiting factor in cereal production worldwide (Dixon; 1987, Blackman and Eastop; 2000).

The green peach aphid (*Myzus persicae*) may at times do considerable damage to the foliage of the peach. The foliage of the cultivated cherry is deformed into gall-like masses by the feeding of the black cherry aphid (*Myzus cerasi*). There is a perception that certain insect pests of crops are well-studied biologically and that their taxonomy is in good order. This perception might lead to the

impression by those who are unfamiliar with the intricacies of acquiring taxonomic understanding of biological diversity that we know all there is to know. In fact, science is a discipline that continually builds on its previous discoveries and technologies as it advances our knowledge. Much research is needed even in economically important insect groups whose taxonomy might be regarded as advanced. Using the Aphidoidea as an example and, in particular, using the early works of North American aphidology as background, we explore various dimensions of taxonomic knowledge in this pest group.

REVIEW OF LITERATURE

Aphids are small, soft-bodied insects generally of 1.5 to 3.5mm in length (Blackman and Eastop; 2000), they feed on plants with piercing sucking mouthparts. Additionally the mechanical damage they cause by this act, aphids as well serves as the prevailing group of vectors of plant viruses (Eastop; 1977, Chan et al.; 1991). Noted as long ago as in (R'eaumur's; 1737) work, honeydew is excreted from the anus and is high in plant sugars and other compounds. Furthermore having an influence on predators (Glen; 1973) and parasitoids (Faria; 2005), it serves as substrates for the growth of fungal complexes that cause sooty mold (Westcott ; 1971).

More than 250 species of the Aphidoidea (in the families Adelgidae, Phylloxeridae, and Aphididae) feed on agricultural or horticultural crops (Blackman and Eastop; 2000). Aphids are also reported as important pests of cereals worldwide (Vickerman and Wratten; 1979, Carter et al.; 1980). The genus *Aphis* L. is certainly the largest worldwide taxon of viviparous aphids (Rhynchota Aphidoidea), being it represented by nearly 600 valid species (Remaudière; 1997).

The earliest North American aphid descriptions (Rafinesque; 1817, 1818, Haldemann; 1844) were based almost entirely on coloration, general appearance, and host association. Subsequent workers (Walsh; 1863, Riley; 1879, and Miller et al.; 2006) relied on descriptions of general appearance but also routinely included measurements of body length in their species descriptions. (Walsh's; 1863) generic descriptions added comparisons to other morphological structures, e.g. relative length of siphunculi in comparison to tarsal length, antennal segment length, such as 'about half as long as the earlier (Monell ; 1879) or 'sub equal' (Oestlund; 1886).

Galls of certain aphid species (*Baizongia pistacia* [Linnaeus] and *Schlechtendalia chinensis* [Bell]) have been used for centuries in medicine, tanning, and dyeing (Fagan; 1918, Das; 1918) studied the Aphididae of Lahore. (Ghulam-ullah; 1940) described a few aphid species from Dehli. (Raychaudhuri; 1956) reviewed the Greenidea and related genera from India. (Eastop; 1961) studied the Aphididae (Homoptera) of West Africa.

A comprehensive work on aphid identification is still needed to be done especially in areas of Quetta of Balochistan province of Pakistan. Therefore, it was planned to identify aphids on the basis of their taxonomic features in Quetta areas. For this purpose, Aphids were collected randomly from different localities of Quetta during 2013-2014 to explore the aphid. In view of the greater variation in characters of the specimens in this area, non-availability of detailed descriptions, the keys have been constructed in order to make the future identification easier.

(Chakrabarti and Raychaudhuri; 1978) provided a comprehensive account of 34 Callipterian aphid species occurring in North-East India and Bhutan. (David; 1975) gave taxonomic revision of *Macrosiphum* of India. (Eastop and Hille-Ris-Lambers; 1976) surveyed fewer than 12 plant groups from India.

(Blackman and Eastop; 1984) reviewed the aphids on the world's crops and gave illustrate key for their identification. (Hodjat; 1984) worked on aphids of Salicaceae of Iran and keyed 27 species. (Eastop and Hodjat; 1981) keyed 38 genera of common aphids from Iran. (Raha and Raychaudhuri; 1983) recorded 59 species from Negaland. (Raychaudhuri et al.; 1981) listed 43 species from South India. (Khaliq; 1965) studied the aphididae of Peshawar District. (Awan; 1973) identified 24 species of aphids from Layallpur.

(Shah; 1988) worked on the taxonomy of the aphids of the summer vegetables in Peshawar region. (Agarwala and Ghosh; 1984) gave a check list of Aphidoidea of India and listed 700 species. (Mondal et al.; 1978) gave aphid fauna of Sikkim and listed 17 species of genera *Aphis*, *Rhopalosiphum*, *Hyseroneura*, *Melanaphis* and *Toxoptea*. (Raychaudhuri et al.; 1978) worked on the root-infesting aphids from North-East India and gave information about 14 species of the subfamily Pemphiginae.

(Eastop; 1979) keyed 24 genera of the worldwide subtribe Aphidina. (Ghosh; 1979) listed 311 aphid species from India, Pakistan, Nepal, Sikkim and Bhutan, with notes on their food plants and distribution. (Raychaudhuri et al.; 1980) studied 78 aphid species from North and North-West India. (Nasir; 1989) keyed 34 species from the Punjab province and gave a check list of Aphidoidea of the Punjab. (Blackman and Eastop; 1994) reviewed the aphids on the world's trees and gave illustrated keys for their identification.

MATERIALS AND METHODS

Aphids were collected either by removing them with an ordinary camel-hair brush or by jarring the plants on white paper sheet from a wide range of habitats including crop field, gardens, grasses, weeds, trees etc.

Specimens were collected from Quetta, Baleli, Shaikhmanda, Sariab and Brewery in the province of Balochistan during 2013-2014.



A few aphid colonies along with the attacked plant parts were also brought to the laboratory and then reared for 2-3 days by keeping them in small polythene bags with a small piece of cotton dipped in water in

order to obtain well-developed adults. The large sized adults were then killed, preserved in 70% alcohol in vials and properly labeled.



The specimens were examined using binocular and the specimens were identified up to specific level by using the keys of Blackman and Eastop (1984).



The morphological terminology is illustrated for generalized aphids. The length of CAUDA is not included in body length of aphids. The following abbreviations are used:

BL	Body Length
ANT I, ANT II, etc	Antennal Segment I, II, etc
ANT PT	Terminal Process of Last Antennal Segment
R IV+V	Last Rostral Segment, (In Fact The Complex Formed By R Iv and R V, Which are Sometimes
HT I, HT II	First and Second Segment of Hind Tarsus
ABD TERG 1, 2, etc	Abdominal Tergites 1, 2, etc
SIPH	Siphinculus
CAUDA	CAUDA

Morphology and key characters help full while identifying the specimens:

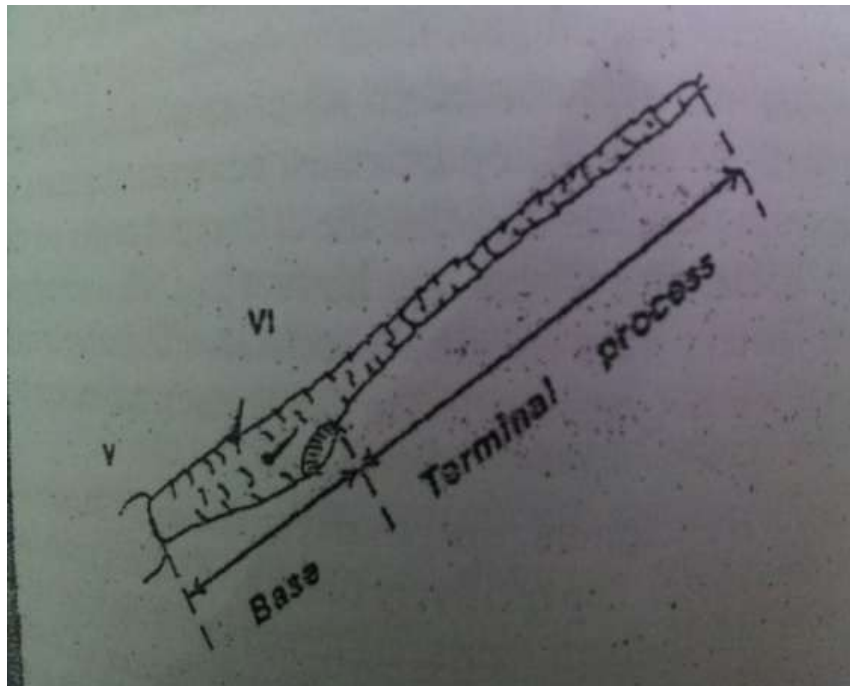


Fig 3.1: The base and terminal process of last antennal segment

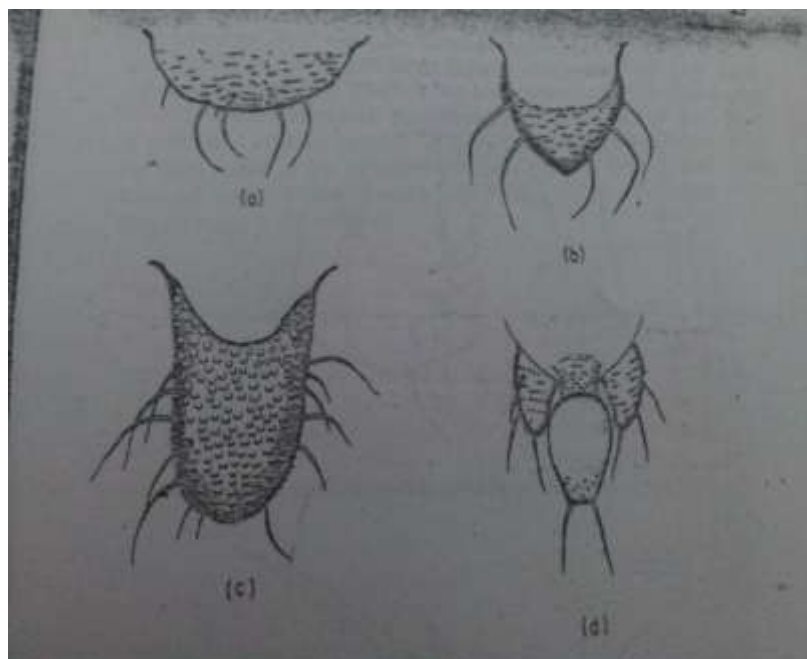


Fig 3.2: CAUDA of various aphids: a) broadly rounded; b) helmet-shaped; c) tongue-like; d) knobbed

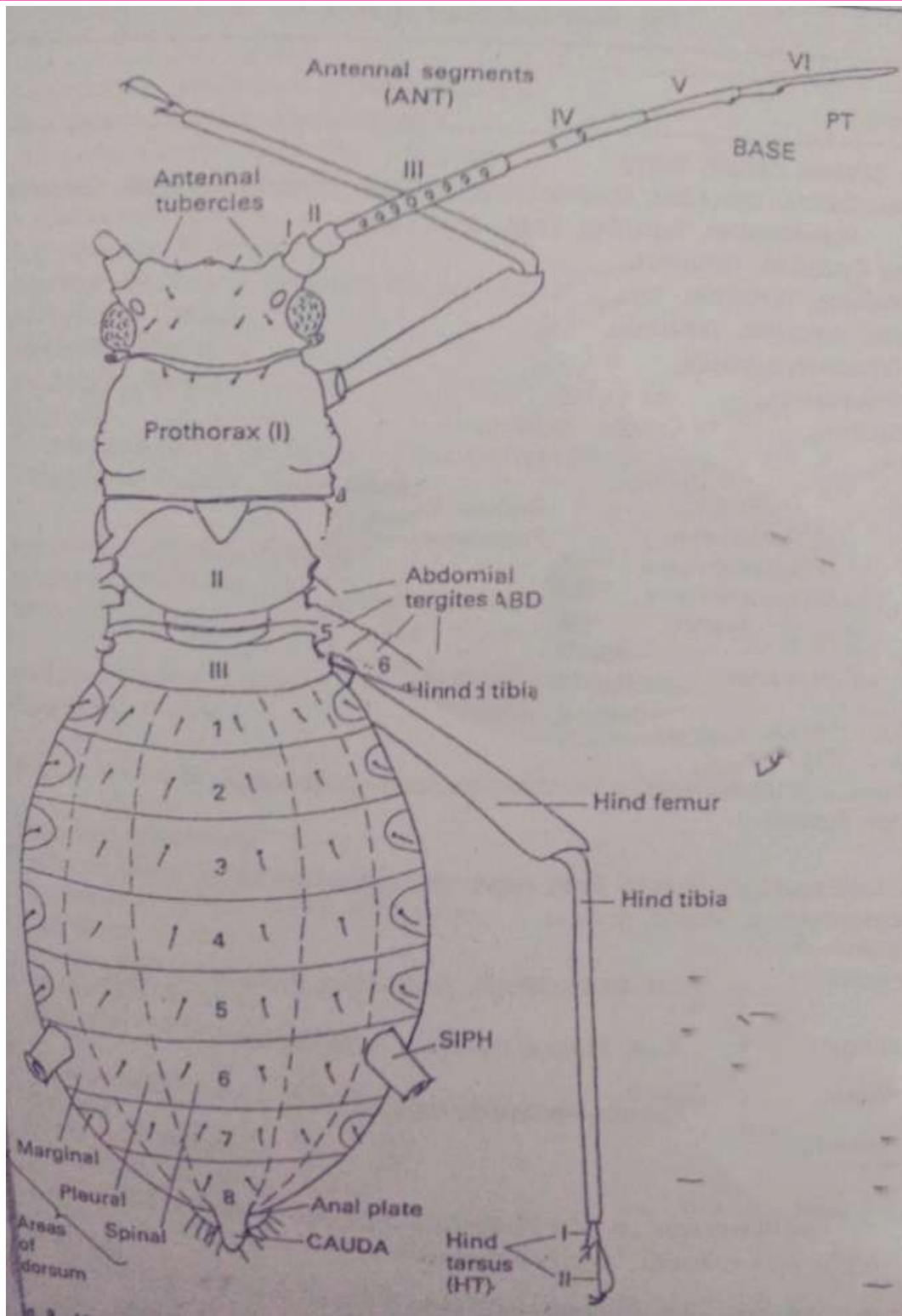


Fig 3.3: Morphology of a generalized aphid, with parts names in accordance with terminology used in keys

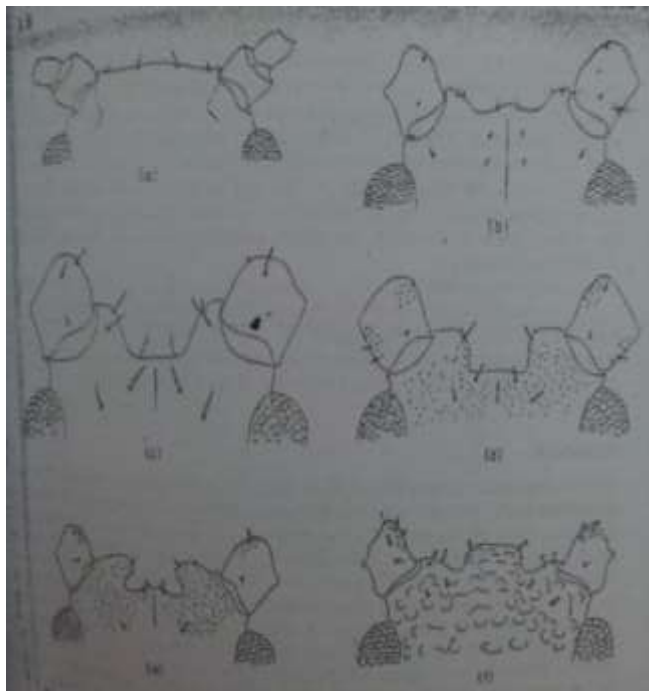


Fig 3.4: Dorsal view of heads of various species shows the degree of the antennal tubercles: a) undeveloped; b) diverging, with median frontal prominence; c) diverging, well developed; d) parallel; e) convergent; f) well developed median frontal projection

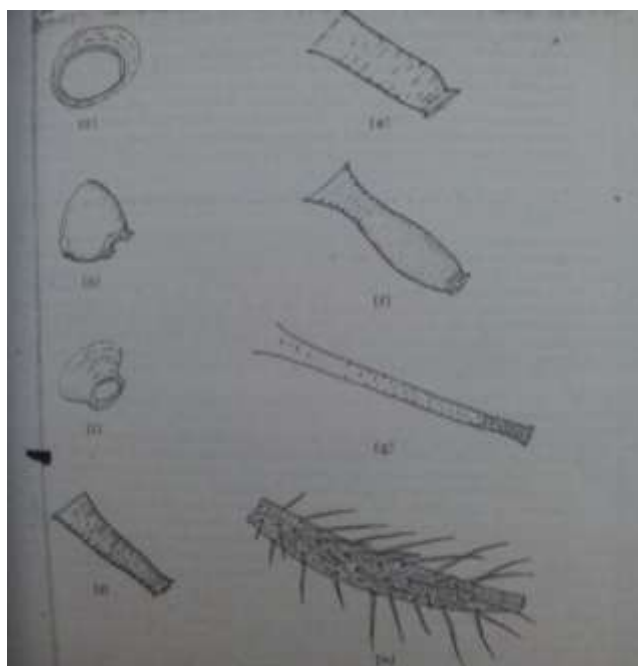


Fig 3.5: Various aphid siphunculi: a) pore-like; b) mammariform; c) truncate; d) tapering; e) swollen proximal to flange; f) clavate; g) with subapical zone of polygonal reticulation; h) greenideine

RESULTS:

Hosts and identified aphids:

HOSTS	APHIDS
Cabbage	<i>Bravicornyne brassicae</i>
Chrysanthemum	<i>Aphis gossypii</i>
Lettuce	<i>Aphis fabae</i>
Rose	<i>Macrosiphum euphrobiae</i>

1. Cabbage: (*Brevicoryne brassicae*)



- 1a. Terminal process longer than base of last antennal segment 2b.
- 1b. Terminal process shorter much than base of last antennal segment **2a.**
- 2a. Siphunculi absent, pale and more than 1.5 times longer than CAUDA **3a.**
- 2b. Siphunculi present, dusky or dark, less than 1.5 times longer than CAUDA 3b.
- 3a. CAUDA broadly triangular in dorsal view **Brevicoryne brassicae.**
- 3b. CAUDA tongue shaped *Lipaphis erysimi.*

Brevicoryne brassicae:

Brevicoryne is small genus. *Brevicoryne brassicae* (L.) is also known as cabbage aphid, Mealy cabbage aphid. They are medium sized, grayish green or dull mid green have dark head and dark dorsal thoracic and abdominal markings. They mostly feed on cabbage, cauliflower, Brussels sprout, radish, and mustard, makes large colonies under the leaves, on the flower heads which reduces the setting of seeds.

Chrysanthemum: (*Aphis gossypii*)



- 1a. CAUDA helmet shape 2b.
- 1b. CAUDA tongue-shaped **2a.**
- 2a. Antennal tubercles absent or weakly developed **3a.**
- 2b. Antennal tubercles well developed 3b.
- 3a. Terminal process of rostrum more than twice as long as last antennal segment **Aphis gossypii.**
- 3b. Terminal process of rostrum less than twice as long as base of last antennal segment *Coloradoa rufomaculata.*

Aphis gossypii:

Aphis gossypii are found variable in color. Large specimens are dark green or almost black. Adults observed in yellow to almost white in color produced in colonies, having pale or dusky cauda and dark siphunculi. Aphis gossypii are extremely polyphagous attacks commonly on cotton, cucurbits, eggplant, pepper, potato, hibiscus, rose and chrysanthemum.

1. Lettuce: (Aphis fabae)



- 1a. Antennal tubercles little developed **2a.**
- 1b. Antennal tubercles well developed **2b.**
- 2a. Siphunculi usually darker than general body **3a.**
- 2b. Siphunculi pale at least on basal half, sometimes darker distally **3b.**
- 3a. Lateral tubercles present at least on abdominal segment 1 and 7 **Aphis fabae.**
- 3b. Lateral tubercles absent from abdominal segment 1 and 7 **Macrosiphum spp.**

Aphis fabae:

It is very polyphagous on the secondary hosts including many crops in addition to which it is a virus vector in sugar beet. Young colonies are of matt black aphids on newly grown or young shoots. Aphis fabae have longer siphunculi, shorter lateral abdominal hair, and shorter cauda. They are highly attracted by ants, and attacks mostly on tomato, tobacco, lettuce, potato, crucifers, beets, peas and beans.

2. Rose: (Macrosiphum euphorbiae)



- 1a. Terminal process very short **2a.**
- 1b. Terminal process longer than base of last antennal segment **2b.**
- 2a. Siphunculi merely pores on flat dark discs, as short cones, much shorter than CAUDA **3b.**
- 2b. Siphunculi tubular, similar in length to or longer than CAUDA **3a.**
- 3a. Antennal tubercles moderately to well developed **4b.**
- 3b. Antennal tubercles weakly developed or absent **4a.**

- 4a. CAUDA short, about as long as its basal width in dorsal view *Pseudocercidis rosae*.
- 4b. CAUDA tongue-shaped, clearly longer than its basal width in dorsal view **5b.**
- 5a. Siphunculi distinctly clavate *Wahalgreniella nervata*.
- 5b. Siphunculi cylindrical or tapering on distal half **6a.**
- 6a. Siphunculi usually dark or at least dusky on distal part **7a.**
- 6b. Siphunculi usually pale *Myzus japonensis*.
- 7a. Siphunculi pale at least over basal half, sometimes dusky towards apex **Macrosiphum euphorbiae.**
- 7b. Siphunculi wholly darker than body color, except sometimes at extreme base *macrosiphum rosae*.

Macrosiphum euphorbiae:

They have pear shaped body. Usually they are shiny green but sometimes are yellow or pink. Legs, siphunculi, and antenna are mainly same in color as body but siphunculi and antennae darker towards apices, sometimes antennae are entirely darker. Primary host for *Macrosiphum euphorbiae* is rose spp.

3. Tomato: (*Aphis craccivora*)



- 1a. Siphunculi wholly dark **2a.**
- 1b. Siphunculi pale, at least basally **2b.**
- 2a. Antennal tubercles weakly developed **3a.**
- 2b. Antennal tubercles well developed *Aulacorthum solani*.
- 3a. dorsal abdomen with an extensive black patch centered on tergites 4-5 **4a.**
- 3b. dorsal abdomen with scattered black markings **4b.**
- 4a. CAUDA black, tongue shaped, rather pointed and with 4-7 hair **Aphis craccivora.**
- 4b. CAUDA pale or dark, if black it usually bears more than 7 hair *Aphis gossypii*.

Aphis craccivora:

Aphis craccivora is also known as black legume aphid, cowpea aphids or groundnut aphid. Adults are shiny black small aphids; young ones are found focused on the growing points of the host and are lightly dusted with wax. Ants are highly attracted to *Aphis craccivora*. It attacks mostly on crucifers, tomato, beet, beans, and cucurbits.

Conclusion:

Overall, only a small fraction of the total variance in ecological traits (<10%) was explained by different degrees of ant attendance. This might indicate that many of the ecological variables that have been suggested to affect

the expression of ant-herbivore interactions are of little importance or act only at small scales and therefore do not show up in a broad comparative analysis.

SUMMARY

Aphids (Hemiptera: Aphididae) are small, soft bodied insects comprising of 1.5 to 3.5mm in length. It is the important group of insects with worldwide distribution.

They are one of the important vegetable pests. Aphids are feeding insects, causing direct or indirect mechanical and economical damage as they feed on plant's sap with

piercing sucking mouthparts; aphids also serve as the largest group of plant virus's vector. Aphids have experienced some adaptations in relation to host plants so that many aphid taxa have biologically complex life

LITERATURE CITED

Agarwala, B.K. and Ghosh, A.K. (1984). A Checklist of Aphidoidea of India. Rec. Zool. Surv. Paper No. 50,69p.

Blackman, R.L. and Eastop, V.F. (1994). Aphids on the World's Trees: An Identification Guide. C. A. B. International, Wallingford, 987p.

Blackman, R.L. Eastop, V.F. (2000). Aphids on the world's crops: An identification and information guide. 2nd Edition. John Wiley & Sons, London, UK.

Blackman R.L. and Eastop, V.F. (2006). Aphids on the World's Herbaceous Plants and Shrubs. (Volume 1: Host Lists and Keys, Volume 2: The Aphids). John Wiley & Sons, Chichester.

Blackman, R.L. and Eastop, V.F. (2007). Taxonomic issues. Pp.1-29. In: van Emden H.F., Harrington, R. (eds.), Aphids as crop pests. CAB International, UK.

Capinera, J.L. (2002). North American vegetable pests: the pattern of invasion. American Entomologist, 48: 20-39.

Chakrabarti, S. (1977). Chaitophorinae (Homoptera: Aphididae) of India with Descriptions of Three New Species from North-West India. Orient. Ins., 11(2): 205-224.

Chakrabarti, S. and Raychaudhuri, D.N. (1978). Callipterine Aphids (Homoptera: Aphididae) of North-East India and Bhutan. Proc. Zool. Soc., Calcutta, India, 28: 71-101.

Chan, C.K., Forbes, A.R. and Raworth, D.A. (1991). Aphid-transmitted viruses and their vectors of the world. Agriculture Canada Technical Bulletin 1991-3E: 1-216.

David, S.K. (1975). A Taxonomic Review of Macrosiphum (Homoptera: Aphididae) In India. Orient. Ins., 9(4): 461-493.

Dixon, A.F.G. (1987). Cereal aphids as an applied problem. Agric. Zool. Rev. 2: 1Ð57.

Eastop, V.F. (1961). A Study of the Aphididae (Homoptera) of West Africa. Brit. Mus. (Nat. Hist.), Lond.,93p.

Eastop, V.F. (1979). Key to the Genera of Subtribe Aphidian (Homoptera). Syst. Ent.,4:379-388.

Eastop, V.F. and D. Hille-Ris-Lambers, (1976). Survey of the World's Aphids. The Hague, the Netherlands, Dr. W. Junk, 573 P.

Eastop, V.F. and Hodjat, H. (1981). Key to the Genera of Common Aphids of Iran. Ent. Phyt. Appliq., 49(1): 45-46.

Fagan, M.M. (1918).Theuses of Insect Galls. American Naturalist 52: 155-176.

Faria, C.A. (2005). The Nutritional Value of Aphid Honeydew for Parasitoids of Lepidopteran Pests. Doctorate of Natural Sciences Dissertation. University of Neuch^Atel 124 Pp.

cycles. The genus *Aphis* is one of the most difficult genera of aphids to identify because of morphological conservation and this problem is complicated by the fact that several species may live on a single host plant.

Ghosh, A.K. (1979). Addition to the List of Aphids (Homoptera: Aphididae) from India and Adjacent Countries. J. Bombay Nat. Hist. Soc., 74(1): 29-44

Glen, D.M. (1973). The Food Requirements of *Blepharidopterus Angularus* Heteroptera Miridae Asapredator of the Limeaphid *Eucallipterus Tiliae*. Entomologia Experimentalis Et Applicata 16(2): 255-267.

Haldemann, S.S. (1844). Descriptions of Several Species of *Aphis* Inhabiting Pennsylvania. Proceedings of The Boston Society of Natural History 1: 168-169.

Heie, O.E. (1987). Paleontology and phylogeny. In: Minks AH, Harrewijn, P. eds. Aphids: Their Biology, Natural Enemies and Control. Elsevier Science Publishers, Amsterdam: 367-391.

Heie, O.E. (2000): Three *Aphis* spp. New to the Danish aphid fauna (Hemiptera: Aphidoidea). Entomologiske Meddelelser 68: 61-62.

Heie, O.E. (2009). Aphid mysteries not yet solved (Hemiptera: Aphidomorpha). Aphids and other Hemipterous Insects.15:31 48.

Hodjat, H. (1984). Aphids of Salicaeae In Iran and Keys To Their Species. Ent. Phyt. Appliq, 52(1): 13-14.

Howard, L.O. (1898). Danger of Importing Insect Pests. Yearbook of the United States Department of Agriculture 1897, Pp. 529-52. Government Printing Office, Washington, Dc.

Kim et al. (2010). Morphometric relationship, phylogenetic correlation, and character evolution in the species-rich genus *Aphis* (Hemiptera: Aphididae). Plosone, 5: e11608.

Kim et al. (2011). Macroevolutionary patterns in the Aphidini aphids (Hemiptera: Aphididae): diversification, host association, and biogeographic origins. Plosone, 6: e24749.

Margaritopoulos, J.T., Tsitsipis, J.A., Zintzaras, E., Blackman, R.L. (2000). Host-correlated morphological variation of *Myzus persicae* (Hemiptera: Aphididae) populations in Greece. Bulletin of Entomological Research 90: 233-244.

Martin, J.H. (1983). The Identification of Common Aphid Pests of Tropical Agriculture Tree Pest Manag., 29(4): 395-411.

Martin, J.H., Brown, P.A. (2008): Global aphids. Systematic Entomology 33: 214-215.

Mier Durante, M.P., et al. (2003): Aphidini (Hemiptera: Aphididae) living on *Senecio* (Asteraceae), with descriptions of a new genus and three new species. The Canadian Entomologist 135: 187-212.

Miller, G.L., Kane, E.C., Eibl, J. and Carlson, R.W. (2006). Resurrecting Asa Fitch's Aphid Notes: Historical Entomology for Application Today.

Mondal, P.K., B.K. Agarwala and Raychaudhuri, D. N. (1978). Aphid (Homoptera: Aphididae) Fauna of Sikkim-I. Sci. Cult., 44(2): 89-92.

Mondal, P.K., Basu, R.C. and Raychaudhuri, D.N. (1976). Studies on Aphids (Homoptera: Aphididae) from Eastern India. Genus Toxoptera. Orient. Ins. 10(4): 533-40.

Monell, J. (1879). Notes on the Aphididae of the United States, With Descriptions of Species Occurring West of the Mississippi. Part ii. Notes on Aphidinae, With Descriptions of New Species. Bulletin of the United States Geological and Geographical Survey of the Territories 5: 18–32.

Nieto Nafria, J.M., et al. (1999): Three new species of Aphis (Hemiptera: Aphididae) living on Mulinum (Umbelliferae) in South America. The Canadian Entomologist 131: 283–292.

Oestlund, O.W. (1886). List of the Aphididae of Minnesota, with Descriptions of Some New Species. Annual Report of the Geological Natural History Survey of Minnesota 14: 17–56.

Perez Hidalgo, N., Nieto Nafria, J.M. (2004). A new species of Aphis (Hemiptera: Aphididae) living on roots of Thymus mastichina (Lamiaceae) from Spain. Annales de la Société Entomologique de France 40: 193–198.

Poulios, K.D. et al. (2007). Morphological separation of host adapted taxa within the Hyalopterus pruni complex (Hemiptera: Aphididae). European Journal of Entomology 104: 235–242.

Powell, W. (1994). Némec and Stary's "Population diversity centre" hypothesis for aphid parasitoids re-visited. Norwegian Journal of Agricultural Sciences Supplement 16: 163–169.

R'Eaumur, R.P. (1737). M'Emoires Pour Servir A l'histoire Des Insectes. Tome Troisieme. De L' Imprimerie Royale Paris. 532 Pp. + Plates

Rafinesque, C.S. (1817). Specimens of Several New American Species of the Genus Aphis. American Monthly Magazine 1:360–361.

Rafinesque, C.S. (1818). Second Memoir on the Genus Aphis, Containing the Descriptions of 24 New American Species. American Monthly Magazine 3: 15–18.

Raha, S.K. and Raychaudhuri, D.N. (1981). Studies on the Aphid (Homoptera: Aphididae) of Nagaland. Entomon, 6(4): 317-323.

Raychaudhuri, D.N. (1956). Revision of Greenidea and Related Genera (Homoptera: Aphididae). Zool. Verh., 31: 1-106.

Raychaudhuri, D.N., Ghosh, L.K. and Das, S.K. (1980). Studies on Aphids (Homoptera: Aphididae) from North and North-West India-I. Matsum., 20:1-42.

Raychaudhuri, D.N., Pal, P.K. and Ghoshi, M.R. (1978). Root-Infesting Aphids (Homoptera: Aphididae: Pemphiginae) from North-East India. Entomon., 3(2): 239-264.

Rehman, A. and Powell, W. (2010). Host selection behavior of aphid parasitoids (Aphidiidae:Hymenoptera). Journal of Plant Breeding and Crop Science 2: 299–311.

Remaudière, G. and Remaudière, M. (1997). Catalogue of the World's Aphididae Homoptera Aphidoidea - Inra, Paris, France.

Riley, C.V. (1879). Notes on the Aphididae of the United States, With Descriptions of Species Occurring West of the Mississippi. Part I. Notes on the Pemphiginae, with Descriptions of New Species. Bulletin of the United States Geological and Geographical Survey of the Territories 5(1): 1–17.

Shaposhnikov, et al. (1998). Evolutionary tendencies and system of Aphididae. In: Nieto JM, Dixon AFG, eds. Proceedings of the Aphids in Natural and Managed Ecosystems. Universidad de Leon, Leon: 481-487.

Stroyan, H.L.G. (1977). Handbook for the Identification of British Insects, Homoptera:Aphidoidea, Chaitophoridae and Callaphidida. R. Ent. Soc., Lond., 130p.

Vickerman, G.P. and Wratten, S.D. (1979). The Biology and Pest Status of Cereal Aphids (Hemiptera: Aphididae) In Europe: A Review. Bull. Entomol. Res. 69: 1832.

Voegtlin, D.J., Halbert, S.E., Qiao, G. (2004). A guide to separating Aphis glycines Matsumura and morphologically similar species that share its hosts. Annals of the Entomological Society of America 97: 227–232.

Walsh, B.D. (1863). On the Genera of Aphidae Found in the United States. Proceedings of the Entomological Society of Philadelphia 1: 294–311.

Westcott, C. (1971). Plant Disease Handbook. Van Nostrand Reinhold Co., New York. 843 Pp.

Worf, G.L., Heimann, M.F. and Pellitteri, P.J. (1995). Sooty Mold. University of Wisconsin Cooperative Extension Publication Madis on Wisconsin A2637. 2 Pp.

Zhang, G.X. and Chen, X.L. (1999). Study on the phylogeny of Pemphigidae (Homoptera: Aphidinea). Acta Entomol Sin 42: 176–183.