A Report On Coleopteran Species Composition In Rono-Hills, Arunachal Pradesh With Perspective On Ecological And Economic Aspects

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Abstracts: The primary objective of the present study was to focus on the species composition of the coleopteran species in Rono-Hills, Papumpare district, Arunachal Pradesh with a perspective to know the economic and ecological value of the coleopteran insect community in the region. For this, field based survey was conducted from the month January to December during the year 2015. Insect monitoring was based on Pollard walk during morning hours starting from 7 am to 9 am. During the survey undertaken, 30 species of Coleoptera belonging to 12 families were recorded. Cerambycidae was the dominant family representing 7 species followed by family Scarabaeidae (6), Carabidae (4), Lucanidae (3), Curculionidae (2), Coccinellidae (2), Passalidae (1), Elateridae (1), Buprestidae (1), Chrysomelidae (1), Erotylidae (1) and Trictenotomidae (1). Eleven species of Coleoptera were documented in the grassland, 9 species in mixed forest, 5 species each in open land and cropland habitat. The cultivated crops conceded were the *Luffa acutangula* and *Abelmoschus esculentus*. Among the coleopteran insects collected, 6 species viz. *Popillia japonica, Lissorhoptrus oryzophilus, Xylotrupes gideon, Coccinella sp., Triplex collaris* and *Apriona germarii* were identified as pests of crops and plantations; 1 species, *Aulacophora palliata* as crop pollinator of the crop *Luffa acutangula* along with 2 species of dung beetle viz. *Onthophagus taurus* and *Scarabaeus sp.* of ecological importance. Thus, the survey indicated the need of management practice of these insects to gain more economic benefit and ecological service in the region.

Key words: Coleoptera, Ecological service, Habitat, Pest, Pollinator

Introduction

Insects are the most diverse group of organisms in the animal kingdom. Out of these, about 3,87,100 species of Coleoptera belonging to 160 families were found to be recorded and is known to be the most speciose order till date (Chandra, 2011). The coleopterans, commonly known as beetle are characterised by hard fore wings known as elytra which meet at the mid dorsal plane without overlapping. The antennae within this group of insects are known to vary and seven different types of antennae viz. filiform (e.g. Carabidae), Lamellate (e.g. Scarabaeidae), clavate (e.g. Siphidae), monoliform (e.g. Chrysomelidae), serrate (e.g. Elateridae, Buprestidae), pectinate (e.g. Elateridae) and geniculate (e.g. Lucanidae and Curculionidae) are known from the order. Mouthparts are of chewing type. Legs are composed of coxa, trochanter, femur, tibia, tarsus and claw. However, the legs are modified to perform specific functions such as jumping, swimming, running and digging as adaptation (Gillot, 2005). Like other insects, coleopterans also exhibit ecological and economic significance. Certain coleopteran insects like Dried fruit beetle, *Carpophilus sp.* (Nitidulidae); Snout beetles, *Endaenidius sp.* (Curculionidae); Ant like flower beetle, *Formicomus braminus* La Ferte-Senectere (Anthicidae) act

as crop pollinators (Corlett, 2004) while others damage human valued plant and plant products being a pest e.g. Ambrosia beetle, Xyleborus formicates Eichhoff; Cucumber beetles, Diabrotica sp.; Crucifer flea beetle, Phyllotreta cruciferae (Goeze); Coconut leaf beetle, Brontispa longissima (Gestro); Spotted pine sawyer, Monochamus clamator (LeConte); Flat faced longhorn beetle, Acanthocinus obliquus (LeConte); Sculptured pine borer, Chalcophora virginiensis (Drury); Flat headed pine borer, Phaenops gentilis (LeConte, 1863) (Costello et al., 2008; Hazarika et al., 2009; Jin et al., 2014; Tangtrakulwanich et al., 2014; Hong et al., 2016) of major or minor importance in agriculture and forestry (Syed et al., 1982; Kevan et al., 1986; FAO, 2009). Apart from these, certain coleopterans are beneficial to agricultural practices as they act as the natural enemies e.g. Vedalia beetle, Rodolia cardinalis (Mulsant); Lady bird beetle, Cryptognatha nodiceps Marshall; Kuwana's lady beetle, Chilocorus kuwanae Silvestri; Lady bird beetle, Hyperapsis pantherina Fursch; Singular black lady beetle, Rhymzobius lophanithae (Blaisdell); Lady beetle, Chilocorus nigritus (Fabricus, 1798) and Heather lady bird, Chilocorus bipustulatus (Linnaeus) (Obrycki and Kring, 1998) of a number of crop pests (Horgan and Myers, 2004; Snyder et al., 2004). A number of insects are also saprophagous and thus they along with the predatory beetles play a key role in trophic chain (Safranyik and Carroll, 2006; Lindgren and Raffa, 2013).

Around 17,455 species of Coleoptera has been recorded from India. However, in Arunachal Pradesh the works on Coleoptera are very scarce. Only around 135 species of Coleoptera were known to be recorded from the region (Sengupta and Sengupta, 1981; Singh *et al.*, 2010; Kumawat *et al.*, 2015). Therefore, the present study was emphasized to document the species composition of the Coleoptera in Rono-Hills, Papumpare district, Arunachal Pradesh as a preliminary work with perspective to know the economic and ecological value of the coleopteran insect community in the region.

Materials and methods

The field based survey was conducted from the month January to December, 2015 in Rono-Hills, Papumpare district, Arunachal Pradesh. Rono-Hills is located geographically at latitude 7°08′50′′ N, longitude 93°46′01′′ E and altitude 300 m above mean sea level. The area was covered with tropical semi-evergreen forest, grassland, cropland and open land. Insect monitoring was based on Pollard walks (Pollard, 1977; Pollard and Yates, 1993). For this, four individual points were selected in each habitat and four transects of 100 m length and 1 m width were planned across each of the habitats viz. grassland, mixed forest, open land and cropland. Each of the transects was walked in one direction at slow and even pace (~ 0.2 km/h) covering the length. Monitoring was done in the four habitats in consecutive days per month during morning hours starting from 7 to 9 am. Insect samples were collected using sweep net and forceps and were identified using standard keys (Crowson, 1956; Klimaszewski and Watt, 1997; Choate, 1999). Lowest average rainfall was approximately 41.80 mm in the month of January and highest was in the month of July with about 1433.20 mm. Average minimum and maximum ambient temperature were 11.6 °C and 27.6 °C respectively. Humidity varies with rainfall with minimum 33% and maximum 85% RH.

The grassland habitat was dominated by Dog's tooth grass, *Cynodon dactylon* (L) Pers.; Lesser spear grass, *Chrysopogon aciculatus* (Retz.) Trin; Crown grass, *Paspalum longifolium* Roxb; Family grass, *Panicum notatum* Roxb. and was scattered by plants like Phakphet, *Spilanthus paniculata* Wall. ex DC; Billygoat-weed, *Ageratum conyzoides* L.; Milea-minute vine, *Micania micrantha* Kunth; Gotu kola, *Centella asiatica* (L.); Sleepy plant, *Mimosa pudica* L. and Thumbai, *Leucas aspara* L. etc. The mixed forest area was chosen inside the Botanic Garden of Rajiv Gandhi University, Arunachal Pradesh. Some of the forest trees were Jackfruit, *Artocarpus heterophyllus* Lam; Guava, *Psidium guajava* L; Fig tree, *Ficus abutilifolia* (Miq.) Miq; Mango, *Mangifera indica* L; Bay leaf, *Cinnamomum tamala* (Buch.-Ham.) T. Nees & C. H. Eberm; Soalu, *Litsea sp*; Bar-thekera, *Garcinia pedunculata* Roxb. ex Buch.-Ham; Hollong, *Dipterocarpus retusus* Blume; Dhuna, *Canarium bengalese* Roxb; Rupohi-thekera, *Garcinia lanceifloia* Roxb. The open land selected was the barren area resulted from the earth cutting activities. The *Luffa acutangula* (L.) Roxb. and *Abelmoschus esculentus* (L.) Moench. cultivated area was conceded as the cropland habitat for the present study.

Results

A total of 30 Coleoptera species belonging to 12 families were recorded during the present survey (Table 1, Fig. 1-5). Out of these, highest number of species belonged to family

Table 1. Coleopterans in Rono-Hills, Arunachal Pradesh and their habitats.

Cerambycidae (7). It was followed by family Scarabaeidae (6), Carabidae (4), Lucanidae (3), Curculionidae (2), Coccinellidae (2), Passalidae (1), Elateridae (1), Buprestidae (1), Chrysomelidae (1), Erotylidae (1) and Trictenotomidae (1).

In the habitats studied, 11 species inhabited in grassland, 9 in mixed forest, 5 species each in open land and cropland. Two species of Coleoptera viz. *Aulacophora palliata* and *Coccinella sp.* were recorded from the fruit of *Luffa acutangula* and 1 species Coleoptera viz. *Popillia japonica* from the flower of *Abelmoschus esculentus*. Other two species of Coleoptera viz. *Lissorhoptrus oryzophilus* and *Epilachna sp.*

Sl. No.	Scientific Name	Vernacular name	Family	Habitat	Month of Collection
1.	Apriona germarii Hope, 1831	Longhorn beetle	Cerambycidae	Mixed forest	July
2.	Aristobia reticulator (Fabricius, 1781)	Longhorn beetle	Cerambycidae	Mixed forest	August
3.	Xylorhiza adusta Wiedemann, 1819	Longhorn beetle	Cerambycidae	Mixed forest	June
4.	Derobrachus hovorei Leconte, 1853	Palo verde beetle	Cerambycidae	Mixed forest	July
5.	Coptops leucostictica White, 1858	Leopard Spot Longhorn beetle	Cerambycidae	Mixed forest	June
6.	Baralipton maculosum Thomson, 1857	Longhorn beetle	Cerambycidae	Mixed forest	April
7.	Epepeotes uncinatus Gahan, 1888	Flat faced longhorn beetle	Cerambycidae	Mixed forest	April
8.	Lissorhoptrus oryzophilus Kuschel, 1952	Rice Water weevil	Curculionidae	Cropland, Flower of	October
				Fagopyrum esculentum	
9.	Scyphophorus acupunctatus Gyllenhaal, 1838	Agave weevil	Curculionidae	Grassland	October
10.	Passalus unicornis LePeletier & Serville, 1825	Horned passalus beetle	Passalidae	Grassland	August
11.	Lucanus cervus Linnaeus, 1758	Stag beetle	Lucanidae	Grassland	August
12.	Prosopocoilus blanchardi Parry, 1873	Stag beetle	Lucanidae	Grassland	September
13.	Prosopocoilus giraffe Olivier, 1789	Giraffe stag beetle	Lucanidae	Grassland	August
14.	Xylotrupes gideon (Linnaeus, 1767)	Siamese rhinoceros beetle	Scarabeidae	Grassland	September
15.	Cetonia aurata Linnaeus, 1758	Rose chafer	Scarabaeidae	Grassland	August
16.	Popillia japonica Newman, 1841	Japanese beetle	Scarabaeidae	Cropland, Flower of	August
				Abelmoschus esculentus	
17.	Onthophagus taurus Schreber, 1759	Dung beetle	Scarabaeidae	Open land	August
18.	Anoplognathus pallidicollis Blanchard, 1851	Christmas beetle	Scarabaeidae	Grassland	October
19.	Scarabaeus sp.	Dung beetle	Scarabaeidae	Open land	August
20.	Agonum cupripenne Say, 1823	Ground beetle	Carabidae	Grassland	October
21.	Cicindela japonica Thunberg, 1781	Japanese Tiger beetle	Carabidae	Grassland	August
22.	Pheropsophus sp. (i)	Ground beetle	Carabidae	Open land	February
23.	Pheropsophus sp. (ii)	Ground beetle	Carabidae	Open land	February
24.	Cryptalaus berus Rigout, 1987	Click beetle	Elateridae	Grassland	September
25.	Megaloxantha bicolor (Fabricius, 1775)	Jewel beetle	Buprestidae	Mixed forest	August
26.	Aulacophora palliata Schaller, 1783	Leaf beetle	Chrysomelidae	Cropland, Flower of	April
				Luffa acutangula	
27.	Triplax collaris Schaller, 1783	-	Erotylidae	Mixed forest, (Oyster	May
				Mushroom)	
28.	Coccinella sp.	Ladybug	Coccinellidae	Cropland (Fruit of	August/October
				L.acutangula)	
29.	Epilachna sp.	Lady beetle	Coccinellidae	Cropland	June
30.	Trictenotoma formosana Kreische, 1920	-	Trictenotomidae	Open land	May

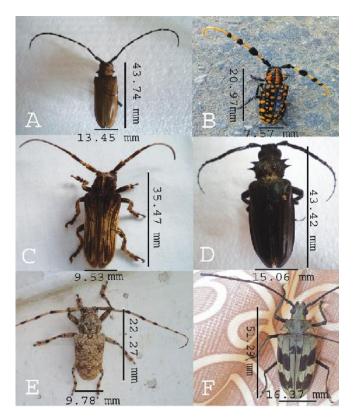


Fig. 1. A- Apriona germarii, B- Aristobia reticulator, C- Xylorhiza adusta, D- Derobrachus hovorei, E- Coptops leucostictica, F- Baralipton maculosum

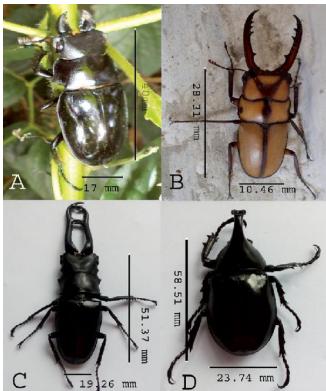


Fig. 3. A- Lucanus cervus, B- Prosopocoilus blanchardi, C- Prosopocoilus giraffe, D- Xylotrupes gideon



Fig. 2. A- Epepeotes uncinatus, B- Lissorhoptrus oryzophilus, C- Scyphophorus acupunctatus, D- Passalus unicornis

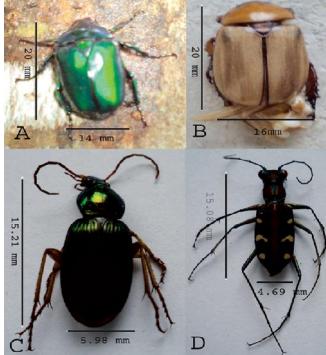


Fig. 4. A- Cetonia aurata, B- Anoplognathus pallidicollis, C- Agonum cupripenne, D- Cicindela japonica

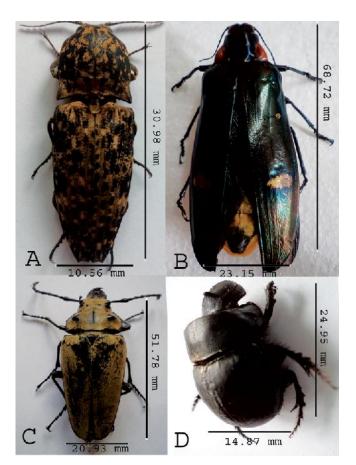


Fig. 5. A- Cryptalaus berus, B- Megaloxantha bicolor, C- Trictenotoma formosana, D-Onthophagus taurus

collected from the cropland area were not found to be associated with the crops during the survey period. *Popillia japonica, Coccinella sp.* and *Triplex collaris* were seen feeding the flowers of *A. esculentus,* fruits of *L. acutangula* and gills of *Pleurotus ostreatus* respectively. *Aulacophora palliata* was recorded pollinating the flowers of *L. acutangula*. Apart from these, 2 species of dung beetle viz. *Onthophagus taurus* and *Scarabaeus sp.* of ecological importance were also documented during this study.

Discussion

Although the order Coleoptera is taxonomically classified into 211 families, during the present study, only 30 Coleoptera species belonging to 12 families were recorded (Bouchard *et al.*, 2011). The comparatively less number of species recorded may be due to the coverage of smaller area, availability of suitable habitat and other resources as the distribution of insects are known to be highly influenced and determined by these factors. Apart from the natural factors, it may also be due to the method of sampling adopted which was conducted during the day time and morning hours. As many coleopterans are known to be nocturnal and inhabit in burrows under soil surface or inside trees, therefore, many of the coleopterans may not be covered while sampled during day time using transect method (Yi *et al.*, 2004). Probably, sampling simultaneously with the use of different methods will be more valid to document the coleopterans that exhibit diverse behavioural patterns from group to group.

Coleoptera possess both negative and positive aspects in terms of economics. Some species of this order are known to act as important crop pollinator whereas some others as pests and predators of the pests. The insects that cause a minimum of 5% economic loss of the crops are considered as crop pest (Paul, 2007). Out of the collected specimens, Coccinella sp., Lissorhoptrus oryzophilus and Popillia japonica are known to cause damage to a number of crops sometimes reaching the pest status (Potter et al., 1966; Klein, 1981; Hummel et al., 2014). Xylotrupes gideon is known to eat foliage of coconut palm (Cocos nucifera L., African oil palm (Elaeis guineensis Jacq.) and Cashew (Anacardium occidentale L.) (Hill, 1987; Howard, 2001). Many species of Triplex destroy Oyster mushroom *Pleurotus ostreatus* (Jacq. ex Fr.) P. Kumm and reside inside them (Gnaneswaran et al., 1999). Certain species of the genus Aulacophora are known as pest of commercial crops, mainly in the Cucurbitaceae family (Rahman and Prodhan, 2007). In this study, A. palliata was recorded as anthophiles of *L. acutangula*. However, literature on A. palliata as pest was lacking. P. japonica feeds on leaves and floral parts of a number of plants. During the present study, P. japonica was collected from the flower of Abelmoschus esculentus. They temporarily reside inside the flower of A. esculentus causing damage to the plant. P. japonica adults are polyphagous in nature and feed on foliage, fruits or flowers of more than 300 species of wild and cultivated plants in 79 families including Malvaceae causing economic loss to the farmers (Fleming, 1972; Ladd, 1987). Apriona

germarii was not recorded to cause damage to the crops covered during the present survey. However, it is known to be a major pest of wide range of economically important plants including Mulberry (*Morus spp.*), Poplar (*Populus spp.*), Willow (*Salix spp.*), Apple (*Malus spp.*), Fig (*Ficus carica* L.), Paper mulberry (*Broussonetia papyrifera* (L.) Vent.), Jackfruit (*Artocarpus heterophyllus* Lam.) and pagoda tree (*Sophora japonica* L.) (EPPO, 2014). Both adults and larvae of *A. germarii* cause damage to the trees eating tender tree bark. Larvae usually lead to serious damage boring emerging tunnels from the site of oviposition (Shimei and Rongwu, 1992).

During the present study, *Aulacophora palliata* was recorded pollinating the flowers of *L. acutangula* in which the male and female flowers borne separately on the same plant but in different internodes (Seshadri, 1999).

Some coleopterans like carabid beetles, ladybug beetles are natural predators of a number of insect pests. On this ground, Coccinellids are used as biocontrol agents against agricultural and forest pests (Obrycki and Kring, 1998; Roy and Migeon, 2010). The adults of *Coccinella sp.* feed on important agricultural pests like aphids, mealy bugs etc. controlling the pest population below economic injury level. Similarly, *Cicindela japonica*, which is a voracious feeder also survive on small insects notorious to agricultural crops (Heinrichs and Barrion, 2004).

From the above results and discussions it was observed that though the present investigation was a preliminary survey using only transect method and confining to a small geographical area, it indicated the presence of some important agricultural and forest pests, crop pollinators and detritus feeder of ecological importance. Therefore, the study urges the management of these insects to gain more economic benefit and ecological service in the region.

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Bouchard, P., Bousquet, Y., Davies, E.A., Miguel, A.A.Z., Lawrence, J.F., Lyal, C. H.C., Newton, A.F., Reid, C.A.M., Schmitt, M., Slipinski, S.A. and Smith, A.B.T. 2011. Family-group names in Coleoptera (Insecta). Zoo Keys. 88: 1-972.

Chandra, K. 2011. Insect fauna of states and union territories in India. ENVIS Bulletin, Arthropods and Their Conservation in India (Insects & Spiders). 14(1): 189-218.

Choate, P.M. 1999. Introduction to the identification of beetles (Coleoptera). In: Dichotomous keys to some families of Florida Coleoptera. Pp: 23-33.

Corlett, R.T. 2004. Flower visitors and pollination in the Oriental (Indomalayan) Region. Bio Rev. 79(3): 497-532.

Costello, S.L., Negron, J.F. and Jacobi W.R. 2008. Traps and attractants for wood-boring insects in Ponderosa Pine stands in the Black Hills, South Dakota. J Econ Entomol. 101(2): 409-420.

Crowson, R.A. 1956. Handbooks for the identification of British insects: Coleoptera - Introduction and keys to families. Royal Entomological Society of London. Pp: 1-65.

EPPO. 2014. Data sheets on quarantine pests. OEPP/EPPO Bulletin. 44 (2): 155-158.

FAO. 2009. Global review of forest pests and diseases. Food and Agriculture Organization of the United Nations, Rome. Pp: 1-235.

Fleming, W.E. 1972. Biology of the Japanese beetle. USDA Tech Bull. 1449: 129.

Gillot, C. 2005. Entomology, 3rd edition. Springer, The Netherlands. Pp: 443-451.

Gnaneswaran, G. and Wijayagunasekara, H.N.P. 1999. Survey and identification of insect pests of oyster mushroom (*Pleurotus ostreatus*) cultures in central province of Sri Lanka. Tropical Agricultural Research and Extension. 2(1): 21-23.

Hazarika, L.K., Bhuyan. M. and Hazarika, B.N. 2009. Insect pests of tea and their management. Annu Rev Entomol. 54: 267-84. Heinrichs, E.A. and Barrion, A.T. 2004. Rice feeding insects and selected natural enemies in West Africa. IRRI. Pp: 1-223.

Hill, D.S. 1987. Agricultural insect pests of the tropics and their control. Cambridge University Press, Cambridge. Pp: 1-647.

Hong, B., Nowatzki, T.M., Sult, T.S., Owens, E.D. and Pilcher, C.D. 2016. Sequential sampling plan for assessing corn rootworm (Coleoptera: Chrysomelidae) larval injury to Bt maize. Crop Protection. 82: 36-44.

Horgan, F.G. and Myers, J.H. 2004. Interactions between predatory ground beetles, the winter moth and an introduced parasitoid on the lower mainland of British Columbia. Pedobiologia. 48: 23-35.

Howard, F.W., Moore, D., Giblin-Davis, R. and Abad, R. 2001. Insects on Palms. CAB International, UK. Pp: 104.

Hummel, N.A., Meszaros, A., Ring, D.R., Beuzelin, J.M. and Stout, M.J. 2014. Evaluation of seed treatment insecticides for management of the rice water weevil, *Lissorhoptrus oryzophilus* Kuschel (Coleoptera: Curculionidae), in commercial rice fields in Louisiana. Crop Protection. 65: 37-42.

Jin, T., Lin, Y., Jin, Q., Wen, H. and Peng, Z. 2014. Sublethal effect of avermectin and acetamiprid on the mortality of different life stages of *Brontispa longissima* (Gestro) (Coleoptera: Hispidae) and its larvae parasitoid *Asecodes hispinarum* Bouèek (Hymenoptera: Eulophidae). Crop Protection. 58: 55-60.

Kevan, P.G. 1986. Pollinating and flower visiting insects and the management of beneficial and harmful insects and plants. In: Biological Control in the Tropics. Eds. M.Y. Hussein and A.G. Ibrahim. Proc.1st Reg. Symp, Biological Control, Universiti Pertanian Malaysia, Serdan. Pp: 439-452.

Klein, M.G. 1981. Mass trapping for suppression of Japanese beetles. In: Management of insect pests with semiochemicals. Ed. E.R. Mithchell. Pp: 183-190.

Klimaszewski, J. and Watt, J.C. 1997. Coleoptera: Family-group review and keys to identification: Fauna of New Zealand. 37: 199.

Kumawat, M.M., Singh, K.M. and Ramamurthy, V.V. 2015. A checklist of the long-horned beetles (Coleoptera: Cerambycidae) of Arunachal Pradesh, northeastern India with several new reports. J Threatened Taxa. 7(12): 7879-7901.

Ladd, T.L. Jr. 1987. Japanese beetle (Coleoptera: Scarabaeidae): influence of favoured food plants on feeding response. J Econ Entomol. 80: 1014-1017.

Obrycki, J.J. and Kring, T.J. 1998. Predaceous Coccinellidae in biological control. Annu Rev Entomol. 43: 295-321.

Paul, A.V.N. 2007. Insect pests and their management. ICAR, New Delhi. Pp: 1-68.

Pollard, E. and Yates, T.J. 1993. Monitoring butterflies for ecology and conservation. Chapman and Hall, UK. Pp: 292.

Pollard, E. 1977. Method for assessing changes in abundance of butterflies. Biol Conserv. 12: 115-134.

Potter, D.A., Loughrin, J.H., Rowe II, W.J. and Hamilton-Kemp, T.R. 1996. Why do Japanese beetles defoliate trees from the top down? Entomologia Experimentalis et Applicata. 80: 209-212.

Lindgren, B.S. and Raffa, K.F. 2013. Evolution of tree killing in bark beetles (Coleoptera: Curculionidae): Tradeoffs between the maddening crowds and a sticky situation. Can Entomol. 145: 471-495.

Rahman, M.A. and Prodhan, M.D.H. 2007. Effects of net barrier and synthetic pesticides on red pumpkin beetle and yield of Cucumber. Int J Sustain Crop Prod. 2: 30-34.

Roy, H. and Migeon, A. 2010. Ladybeetles (Coccinellidae). In: Alien terrestrial arthropods of Europe. Eds. A. Roques, M. Kenis, D. Lees, C. Lopez-Vaamonde, W. Rabitsch, J. Rasplus and D.B. Roy. BioRisk. 4(1): 293-313.

Safranyik, L. and Carroll, A.L. 2006. The biology and epidemiology of the mountain pine beetle in lodgepole pine forests. In: The mountain pine beetle: A synthesis of biology, management and impacts on Lodgepole Pine. Eds. L. Safranyik and B. Wilson, Pacific Forestry Centre, Victoria, Canada. Pp: 3-66. Sengupta, C.K. and Sengupta, T. 1981. Cerambicydae (Coleoptera) of Arunachal Pradesh. Rec Zool Surv India. 78: 133-154.

Seshadri, V.S. 1999 Cucurbits. In: Vegetable crops, vol I. Ed. T.K. Bose, M.G. Som M.G. Naya Prokash, Kolkata, India. Pp: 91-164

Shimei, Z. and Rongwu, S. 1992. *Apriona germarii*. In: Chinese Forest Insects, (2nd edn). Ed. X. Gangrou. China Forestry Press, Beijing. Pp: 461-463.

Singh, O.T., Chakravorty, J. and Varatharajan, R. 2010. Entomofauna of Kane Wildlife Sanctuary, Arunachal Pradesh, northeastern India. J Threatened Taxa. 2(13): 1392-1400.

Snyder, W.E., Ballard, S.N., Yang, S., Clevenger, G.M., Miller, T.D., Ahn, J.J., Hatten, T.D. and Berryman, A.A. 2004. Complementary biocontrol of aphids by the ladybird beetle *Harmonia axyridis* and the parasitoid *Aphelinus asychis* on greenhouse roses. Biological Control. 30: 229-235. Syed, R.A., Law, I.H. and Corley, R.H.V. 1982. Insect pollination of oil palm: Introduction, establishment and pollinating efficiency of *Elaeidobius kamerunicus* in Malaysia. Planter. 58 (681): 547-561.

Tangtrakulwanich, K., Reddy, G.V.P., Wu, S. Miller, J.H., Ophus, V.L. and Prewett, J. 2014. Developing nominal threshold levels for *Phyllotreta cruciferae* (Coleoptera: Chrysomelidae) damage on canola in Montana, USA. Crop Protection. 66: 8-13.

Yi, Z., Jinchao, F., Dayuan, X., Weiguo, S. and Axmacher, J.C. 2012. A comparison of terrestrial arthropod sampling methods. J Resour Ecol. 3(2): 174-182.