

The 3rd International Brachypodium Conference



Beijing ◉ China

July 30-31, 2017

The 3rd International Brachypodium Conference

Beijing, CHINA

July 30-31, 2017

International Brachypodium Steering Committee

Pilar Catalan (University of Zaragoza, Spain)

Mhemmed Gandour (Faculty of Sciences and Technology of Sidi Bouzid, Tunisia)

Samuel Hazen, (Biology Department, University of Massachusetts, USA)

Zhiyong Liu (Institute of Genetics & Developmental Biology, Chinese Academy of Sciences, China)

Keiichi Mocida (RIKEN Center for Sustainable Resource Science, Japan)

Richard Sibout (INRA, France)

John Vogel (Plant Functional Genomics, DOE Joint Genome Institute, USA)

Local Organizing Committee

Zhiyong Liu, Chair (Institute of Genetics & Developmental Biology, Chinese Academy of Sciences, China)

Long Mao, co-Chair (Institute of Crop Sciences, Chinese Academy of Agriculture Sciences, China)

Xiaoquan Qi (Institute of Botany, Chinese Academy of Sciences, China)

Dawei Li (College of Life Science, China Agricultural University, China)

Yueming Yan (College of Life Science, Capital Normal University, China)

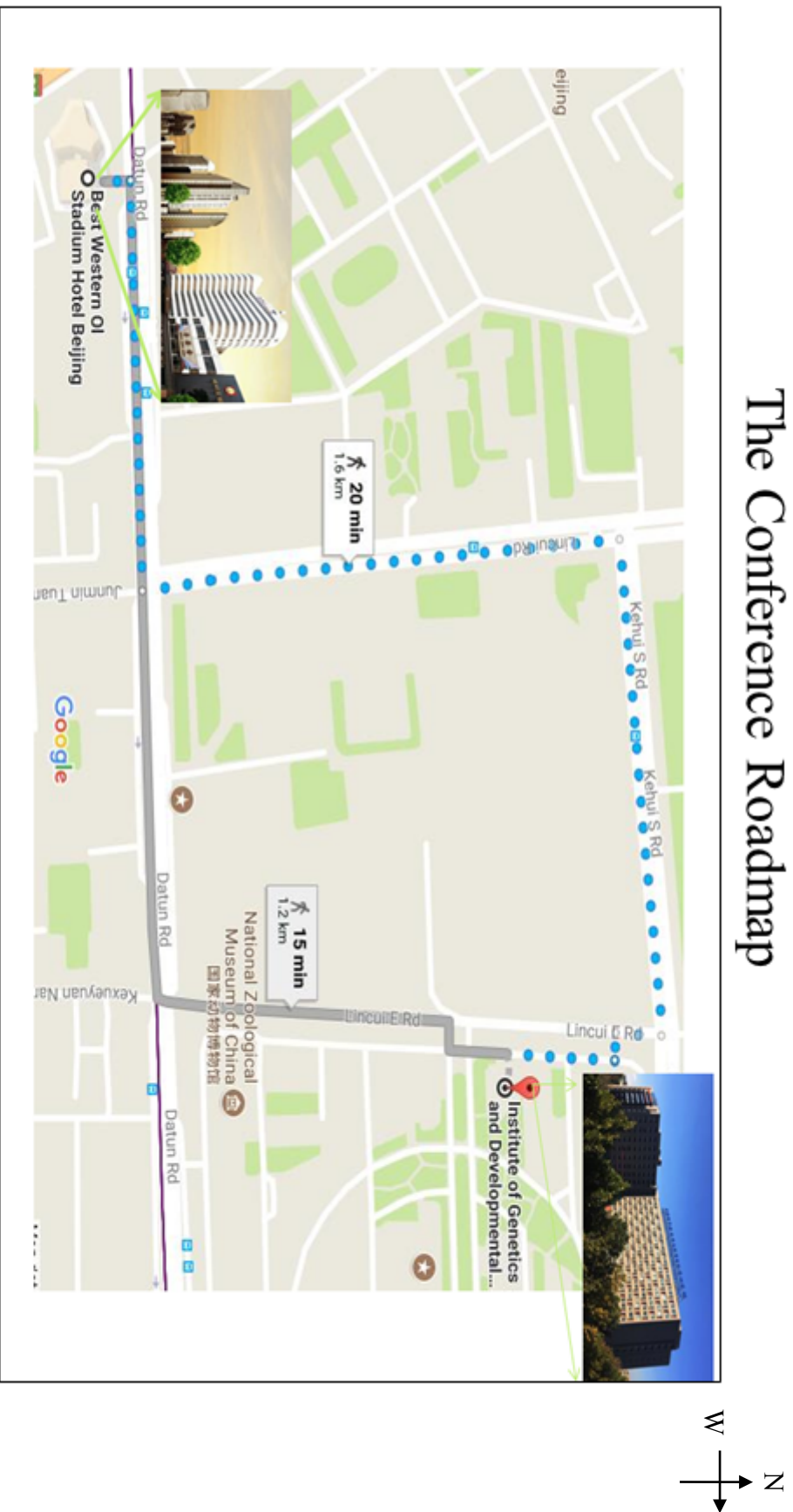
Hailong An (College of Life Science, Shandong Agricultural University, China)

Yuling Jiao (Institute of Genetics & Developmental Biology, Chinese Academy of Sciences, China)

Zhaoqing Chu (Shanghai Chenshan Plant Science Research Center, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences)

Liang Wu (Zhejiang University, China)

The Conference Roadmap



- When you arrive at the Institute of Genetics & Developmental Biology, you may follow the signs in the hall to the meeting room.
- If you have any problems, you can contact our faculty:
Guanghao Guo (15600912996) Qihong Wu (15600910387)

Conference Agenda

July 30, 2017

Conference attendees entry to the IGDB meeting room (Room 215-216)

8:20-8:30 Opening remarks

Session 1: Novel tools and resources. Chair: Zhiyong Liu, IGDB, CAS

Time	Speaker	Institute	Title
8:30-9:00	John Vogel	DOE Joint Genome Institute, USA	What can you learn from 1,019 genomes? Pan-genomics, polyploidy, epigenetics and a cast of mutants
9:00-9:20	Robert Hasterok	University of Silesia in Katowice, Poland	Dissecting grass genome organisation at the cytomolecular level using the model genus <i>Brachypodium</i>
9:20-9:40	Pilar Catalan	Universidad de Zaragoza, Huesca, Spain	Phylogenomics and gene evolution in model annual and perennial <i>Brachypodium</i> species
9:40-10:00	Hailong An	Shandong Agricultural University, China	Ds tagging: a gateway for gene discovery in <i>Brachypodium distachyon</i>

10:00-10:30 Coffee break & Poster

Session 1: Novel tools and resources. Chair: Yongqiang Gu, USDA-ARS

10:30-10:50	Ming Cheng Luo	University of California, Davis, USA	Revisit polyploidization of <i>Brachypodium</i> taxa, evidence from analyses of whole-genome optical maps
10:50-11:10	Cecilie S. L. Christensen	University of Copenhagen, Danmark	Altering lignin composition in <i>Brachypodium</i> using CRISPR/Cas9
11:10-11:30	Yoshihiko Onda	RIKEN Center for Sustainable Resource Science, Japan	A simple and versatile genome-wide SNP genotyping by multiplex PCR targeted amplicon sequencing in <i>Brachypodium distachyon</i>
11:30-11:50	Xiaoquan Qi	Institute of Botany, Chinese Academy of Sciences, China	Generation of <i>Brachypodium distachyon</i> T-DNA mutant population for studying nonhost resistance to wheat stripe rust

12:00-14:00 Lunch

13:00-14:00 International Brachypodium Steering Committee Meeting

Session 2: Development, epigenetics and growth. Chair: Yuling Jiao, IGDB, CAS

14:00-14:20	Sam Hazen	University of Massachusetts, USA	Transcriptional regulation of biomass accumulation in <i>Brachypodium distachyon</i>
14:20-14:40	Karen A. Sanguinet	Washington State University, USA	Identification of the BUZZ kinase involved in root and root hair development
14:40-15:00	Liang Wu	Zhejiang University, China	Gene silencing by endogenous and exogenous miRNAs in flowering-time control in <i>Brachypodium distachyon</i>
15:00-15:20	Alexander Betekhtin	University of Silesia in Katowice	Brachypodium tissue culture as a model system to reveal the functions of the components of the cell wall
15:20-15:40	Koen Geuten	KU Leuven, Belgium	A FLOWERING LOCUS C homolog is a vernalization regulated repressor in <i>Brachypodium</i> and is cold-regulated in wheat
15:40-16:10	Coffee break & Poster		

Session 2: Development, epigenetics and growth. Chair: Karen Sanguinet, Washington State University

16:10-16:30	Natalia Borowska-Zuchowska	University of Silesia in Katowice, Poland	The preferential silencing of <i>B. stacei</i> -inherited rRNA genes in <i>Brachypodium hybridum</i> - an epigenetic point of view
16:30-16:50	Zhongjuan Zhang	Max Planck Institute for Plant Breeding Research, Germany	How does CUC2 regulate leaf serration development in Arabidopsis?

Session 3: Natural variation and evolution

16:50-17:10	Justin Borevitz	Australian National University, Australia	Population structure of the <i>Brachypodium</i> species complex and genome wide dissection of agronomic traits in response to climate
17:10-17:30	Weining Song	North West Agriculture and Forestry University	<i>Brachypodium</i> SPP in Israel likely a hexaploid and containing low genetic diversity
17:30-17:50	Zujun Yang	University of Electronic Science and Technology of China	Diversity of <i>Brachypodium</i> samples in Israel revealed by molecular and cytogenetic methods
18:00-20:30	Dinner		

July 31, 2017

8:00-8:20 Conference attendees entry to the meeting room

Session 4: Plant-Biotic & Abiotic interactions. Chair: Dawei Li, China Agricultural University

8:20-8:40	Kemal Kazan	Queensland Bioscience Precinct, Australia	Brachypodium: A useful model host for cereal-fungal pathogen interactions
8:40-9:00	Pubudu P. Handakumbura	Pacific Northwest National Laboratory, USA	Linking phenotype to genotype: A metabolomics approach to build trait association network models for Brachypodium Expression profiling of marker genes for
9:00-9:20	Yusuke Kouzai	RIKEN Center for Sustainable Resource Science, Japan	defense-associated phytohormones in <i>Brachypodium distachyon</i> highlights its similar immune systems to rice
9:20-9:40	Qihong Wu	Institute of Genetics & Developmental Biology, CAS, China	Interaction of Bsr1 and TGB1 confers Barley Stripe Mosaic Virus resistance in Brachypodium, barley and wheat

9:40-10:10 Coffee break & Poster

Session 5: Plant-Abiotic Stress Interactions. Chair: Justin Borevitz, Australian National University

10:10-10:30	YongqiangGu	USDA-ARS, Western Regional Research Center Albany	Using the JGI Brachypodium T-DNA collection to reveal novel transcription factor roles in abiotic stress responses
10:30-10:50	Zhaoqing Chu	Shanghai Chenshan Plant Science Research Center, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences	Cool season turf grass heat tolerance study through genomic and genetic analyses with <i>Brachypodium distachyon</i>
10:50-11:10	Keiichi Mochida	RIKEN Center for Sustainable Resource Science, Japan	Homoeolog-specific activation for heat acclimation in the allopolyploid grass <i>Brachypodium hybridum</i>
11:10-11:30	AritaKus	University of Silesia in Katowice, Poland	Establishing <i>Brachypodium distachyon</i> as a model in analyses of plant genome stability after mutagenic treatment
11:30-11:50	CAI Jinshu	ShezhenWenke Landscape Corp., Ltd, China	Influence of Supplemental Lighting with different light quality on the turf growth of <i>Festuca Arundinacea</i>
11:50-12:05	Pilar Catalan	Universidad de Zaragoza, Huesca, Spain	Brachypodium Conference 2019

12:05 Meeting Conclusion Remarks: Long Mao, Institute of Crop Sciences, CAAS

Table of contents

Session 1: Novel tools and resources	1
What can you learn from 1,019 genomes? Pan-genomics, polyploidy, epigenetics and a cast of mutants.....	1
Dissecting grass genome organisation at the cytomolecular level using the model genus <i>Brachypodium</i>	3
Phylogenomics and gene evolution in model annual and perennial <i>Brachypodium</i> species...	5
<i>Brachypodium</i> grain transcriptome: a new tool for the identification of potential regulators of key developmental transitions in cereals	7
Revisit polyploidization of <i>Brachypodium</i> taxa, evidence from analyses of whole-genome optical maps	9
Ds tagging: a gateway for gene discovery in <i>Brachypodium distachyon</i>	11
Altering lignin composition in <i>Brachypodium</i> using CRISPR/Cas9.....	13
A simple and versatile genome-wide SNP genotyping by multiplex PCR targeted amplicon sequencing in <i>Brachypodium distachyon</i>	15
Session 2: Development, epigenetics and growth	17
Transcriptional regulation of biomass accumulation in <i>Brachypodium distachyon</i>	17
Identification of the BUZZ kinase involved in root and root hair development.....	19
Gene silencing by endogenous and exogenous miRNAs in flowering-time control in <i>Brachypodium distachyon</i>	21
<i>Brachypodium</i> tissue culture as a model system to reveal the functions of the components of the cell wall	23
A FLOWERING LOCUS C homolog is a vernalization regulated repressor in <i>Brachypodium</i> and is cold-regulated in wheat.....	25
The preferential silencing of <i>B. stacei</i> -inherited rRNA genes in <i>Brachypodium hybridum</i> - an epigenetic point of view	27
Session 3: Natural variation and evolution	29
Population structure of the <i>Brachypodium</i> species complex and genome wide dissection of agronomic traits in response to climate.....	29
Insertion/deletion markers for assessing the genetic variation and the spatial genetic structure of Tunisian <i>Brachypodium hybridum</i> populations.....	31
Diversity of <i>Brachypodium</i> samples in Israel revealed by molecular and cytogenetic methods	33
Environmental isolation explains Iberian genetic diversity in the highly homozygous model grass <i>Brachypodium distachyon</i>	35
Session 4: Plant-biotic and abiotic interactions	37

Generation of <i>Brachypodium distachyon</i> T-DNA mutant population for studying nonhost resistance to wheat stripe rust	37
Brachypodium: A useful model host for cereal-fungal pathogen interactions	39
Linking phenotype to genotype: A metabolomics approach to build trait association network models for Brachypodium.....	41
Expression profiling of marker genes for defense-associated phytohormones in <i>Brachypodium distachyon</i> highlights its similar immune systems to rice	43
Interaction of Bsr1 and TGB1 confers Barley Stripe Mosaic Virus resistance in Brachypodium, barley and wheat.....	45
Using the JGI Brachypodium T-DNA collection to reveal novel transcription factor roles in abiotic stress responses	47
Cool season turf grass heat tolerance study through genomic and genetic analyses with <i>Brachypodium distachyon</i>	49
Homoeolog-specific activation for heat acclimation in the allopolyploid grass <i>Brachypodium hybridum</i>	51
Influence of Supplemental Lighting with Different Light Quality on the Turf Growth of <i>Festuca Arundinacea</i>	53
Establishing <i>Brachypodium distachyon</i> as a model in analyses of plant genome stability after mutagenic treatment.....	55
Phenotypic and metabolomic variation in the model annual grasses <i>Brachypodium distachyon</i> , <i>B. stacei</i> , and <i>B. hybridum</i>	57
How does CUC2 regulate leaf serration development in Arabidopsis?.....	59

Session 1: Novel tools and resources

What can you learn from 1,019 genomes? Pan-genomics, polyploidy, epigenetics and a cast of mutants

John Vogel

Abstract

The sequencing of a single reference genome played a pivotal role in establishing *Brachypodium distachyon* as a model system. Since the production of that initial genome assembly, technological innovations in DNA sequencing have decreased costs to the point where it is now feasible to sequence many *Brachypodium* genomes. The DOE Joint Genome Institute has sequenced over 1,000 genomes from four *Brachypodium* species. An overview of the lessons learned from these sequences will be presented including: A comparison of 54 *B. distachyon* genomes that revealed a pan-genome is considerably larger than the genome of any individual plant. A comparison of the genomes of several *B. hybridum* lines with their diploid progenitors *B. distachyon* and *B. stacei* that revealed multiple origins of *B. hybridum*. An exploration of epigenetic dynamics in *B. distachyon* in response to cold. And, finally, the cataloging of hundreds of thousands of mutations to create a new resource for functional genomic studies.

Session 1: Novel tools and resources

Dissecting grass genome organisation at the cytomolecular level using the model genus *Brachypodium*

Robert Hasterok¹, Alexander Betekhtin¹, Natalia Borowska-Zuchowska¹, Agnieszka Braszewska-Zalewska¹, Karolina Chwialkowska², Marta Hosiawa-Baranska¹, Dominika Idziak-Helmcke¹, Arita Kus¹, Jolanta Kwasniewska¹, Mirosław Kwasniewski², Joanna Lusinska¹, Ewa Robaszkiewicz¹, Magdalena Rojek¹, Rakesh Sinha¹, Aleksandra Skalska¹, Marta Sowa¹, Elzbieta Wolny¹, Karolina Zubrzycka¹

¹Department of Plant Anatomy and Cytology, ²Department of Genetics, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 28 Jagiellonska Street, 40-032 Katowice, Poland

robert.hasterok@us.edu.pl

Abstract

Modern molecular cytogenetics combines various methodological approaches of cytology, molecular genetics and advanced digital image analysis. It focuses on the study of nuclear genomes at the microscopic level. The cytomolecular organisation of plant genomes is still rather poorly investigated, compared to that of animals. Most plant genomes, including those of economically and ecologically crucial cereals and forage grasses, are usually large and saturated with repetitive DNA, which hampers detailed molecular cytogenetic analyses.

Model organisms possess a combination of features, which makes them more amenable to scientific investigation than others. One of the most recent and rapidly developing model systems are representatives of the *Brachypodium* genus, particularly *B. distachyon*. They possess small, and in some cases, already sequenced genomes with a low repeat content, diverse basic chromosome numbers and ploidy levels. They also have an interesting phylogeny, short life cycles and simple growth requirements, complemented by a rapidly and continuously growing repertoire of various experimental tools.

This presentation outlines and discusses our current projects and their future prospects, using *Brachypodium* species for research on various aspects of grass genome organisation, e.g. (i) karyotype structure and evolution, (ii) distribution of

Session 1: Novel tools and resources

Phylogenomics and gene evolution in model annual and perennial *Brachypodium* species

Rubén Sancho¹, Bruno Contreras-Moreira², David Des Marais³, Sean Gordon⁴, John Vogel⁴, Pilar Catalan¹

¹Universidad de Zaragoza, Huesca, Spain

²Estación Experimental de Aula Dei (EEAD-CSIC), Zaragoza, Spain

³Harvard University, Cambridge, MA, USA

⁴DOE Joint Genome Institute, Walnut Creek, CA, USA

Presenting author: pcatalan@unizar.es

Abstract

We have reconstructed a historical scenario for the diverging and merging genomes inherited by diploid and allopolyploid species of *Brachypodium*, using a set of selected loci, Genotyping-By-Sequencing and RNA-seq data. We built a comprehensive phylogeny of *Brachypodium* from five neutral genes using Species-Tree Minimum-Evolution and species-network analyses, and Maximum Likelihood reconstructions of genomic and transcriptomic reads mapped to the three available 2x *Brachypodium* reference genomes. Gene content and dosage in annual-*vs*-perennial, and in diploid-*vs*-polyploid species was estimated using GET_HOMOLOGUES-EST. Our 5-gene data support Mid-Miocene splits of ancestral genomes that preceded Late-Miocene to Quaternary origins of extant diploid species' genomes (*B.stacei*, *B.distachyon*, core perennials). Ancestral Mediterranean genomes presumably merged with recent perennial genomes generating the West-Palaeartic perennial allopolyploids (*B.boissieri*, *B.retusum*, *B.phoenicoides*). Close homeologous American genomes plausibly evolved *in situ*, originating *B.mexicanum*. Quaternary *B.hybridum* resulted from reciprocal *B.stacei* x *B.distachyon* WGD crosses. Core perennial diploids (*B.arbuscula*, *B.sylvaticum*, *B.rupestre*, *B.pinnatum*) evolved in Eurasia from Upper Pleistocene genomes. Karyologically unknown African, Malagasy and Taiwanese species were reconstructed as polyploids. GBS/RNAseq-based phylogenies supported this scenario and further detected ancestral and recent subgenomes in *B. retusum*. Gene content

Session 1: Novel tools and resources

Brachypodium grain transcriptome: a new tool for the identification of potential regulators of key developmental transitions in cereals

Sofia Kourmpetli¹, Syabira Yusoff², Philip Hands³, Sinéad Drea²

¹Cranfield University, School of Water, Energy and Environment, Cranfield, UK, ²University of Leicester, Department of Genetics, Leicester, UK, ³CSIRO, Urrbrae, Australia
s.kourmpetli@cranfield.ac.uk

Abstract

The caryopsis of temperate cereals is a unique type of fruit with great economical value. From a developmental point of view though, very few genetic regulators have been identified to date, and most of the relevant research is undertaken in systems such as maize and rice - which are considerably different in many aspects from the grains of the temperate cereals.

Brachypodium, as a sister to the core pooids that include wheat, barley and rye, represents a good model for the study of grain development. We have selected eight stages of grain development that encompass key transition points in *Brachypodium distachyon* and conducted a comprehensive transcriptomic analysis. We have generated a new valuable resource for the investigation of gene expression patterns throughout grain development and germination that could also be used for comparison purposes with other important crop species in a gene function and evolutionary context. We particularly focused on transcription factors, as they often act as master regulators of developmental processes and we have therefore suggested candidate regulators of developmental transitions and distinctive biological processes they are involved in.

Keywords: Brachypodium, transcriptomics, grain development, transcription factors

Session 1: Novel tools and resources

Revisit polyploidization of Brachypodium taxa, evidence from analyses of whole-genome optical maps

Tingting Zhu¹, Zhaorong Hu^{1,2}, Juan C. Rodriguez¹, Karin R. Deal¹, Jan Dvorak¹, John P. Vogel³, Zhiyong Liu⁴, and Ming-Cheng Luo¹

¹ Department of Plant Sciences, University of California, Davis, CA 95616, USA

² State Key Laboratory for Agrobiotechnology, Key Laboratory of Crop Heterosis Utilization (MOE), China Agricultural University, Beijing, 100193, China.

³ DOE Joint Genome Institute, 2800 Mitchell Dr, Walnut Creek, CA 94598, USA

⁴ State Key Lab of Plant Cell and Chromosome Engineering, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, 100101, China
mcluo@ucdavis.edu

Abstract

There are three recorded Brachypodium taxa, the diploid *B. distachyon* ($2n=10$) and *B. stacei* ($2n=20$), and the allotetraploid *B. hybridum* ($2n=30$). *Brachypodium* has been widely used as genomic and genetic model; it is of importance to shed light on the structure and evolutionary relationships among genomes of the three taxa of Brachypodium. We applied BioNano genome (BNG) mapping technology to construct whole-genome optical maps for the three taxa of Brachypodium, and performed multiple comparisons. Our results shows that *B. stacei* ($2n=20$) is indeed diploid and diverged greatly from the other diploid *B. distachyon* ($2n=10$), while *B. hybridum* ($2n=30$) is an allotetraploid that originated via hybridization of the two diploids. Structural variations between the polyploid *B. hybridum* and its two diploid progenitors indicated the in-del events distributed unevenly across the chromosomes but agrees with the pattern of chromosomal distribution of retro transposons. We demonstrated a great utility of BNG maps for polyploid genome analysis and confirmed the origin of *B. hybridum* via hybridization between *B. distachyon* and *B. stacei*, however, little divergence between the genome of *B. hybridum* and those of the diploids progenitors has taken place.

Keywords: Optical mapping; polyploidization; structural variation

Session 1: Novel tools and resources

Ds tagging: a gateway for gene discovery in *Brachypodium distachyon*

Hongyu Wu¹, Caihua Qin¹, Xiaodong Xue¹, Yi Xu¹, Qinxia Li¹,
Xiaoquan Qi^{2*} and Hailong An^{1*}

¹State Key Laboratory of Crop Biology, College of life Sciences, Shandong Agricultural University, Tai'an 271018, Shandong, China

²The Key Laboratory of Plant Molecular Physiology, Institute of Botany, Chinese Academy of Sciences, 20 Nanxincun, Xiangshan Road, Beijing 100093, China

*for correspondence: hlan@sdau.edu.cn

Abstract

Transposon tagging is a powerful tool for gene discovery in various species. Recently, *Brachypodium distachyon* L. is presented as a model plant for cereals especially the temperate cereals like wheat and barley. So far there is no research work published against successful transposon tagging in Brachypodium. Here we described an efficient Ds transposon tagging system in *Brachypodium distachyon* and its potential for gene discovery in Brachypodium and wheat. The Ac transposase driven by a 1X 35S promoter and Ds element with a *ZmUbi1sGFP* cassette inside were constructed within the same T-DNA, then transformed into Brachypodium via Agrobacterium-mediated transformation. It's easy to detect the somatic transposition events from the leaves of the T0 plants. After selfing, Ds insertional lines without the T-DNA insert were identified from the T1 progeny at a highest frequency about 10%. Using this system, more than 3,000 Ds insertional lines were generated and near 600 Ds flanking sequences were isolated. From the population hundreds of mutants with visible phenotypes were identified. So the system is efficient enough to produce Ds insertional lines in a large scale and to identify genes with new functions. Due to the fact that Brachypodium is close to wheat, so we have good chance to imagine that those genes play similar roles in temperate cereals such as wheat and barley.

Keywords: *B.distachyon*, *Ac/Ds* tagging system, transposants, insertion sites, mutants

Session 1: Novel tools and resources

Altering lignin composition in Brachypodium using CRISPR/Cas9

Cecilie S. L. Christensen, Jeppe O. Husum, Bodil Jørgensen & Søren K. Rasmussen

Department of Plant and Environmental Science, SCIENCE, University of Copenhagen, Frederiksberg, Denmark
cslc@plen.ku.dk

Abstract

The aim of this project is to reduce recalcitrance of bioconversion of the ligno-cellulosic material to increase bioethanol production yield by altering the composition of lignin in Brachypodium. CRISPR/Cas9 genome editing method will be used to target two of the genes coding for cinnamyl alcohol dehydrogenase (CAD4 & CAD5). This project will investigate the function of these two genes coding (CAD4 & CAD5) controlling the last step in the lignin biosynthesis. This will be done by knocking-out or alter the function of the genes.

Waste materials from cereals are a great source for bioethanol production. However lignin is highly recalcitrance to degradation and reduces the hydrolysis of cellulose to fermentable sugars. Cinnamyl alcohol dehydrogenase (CAD) catalyses the last step in the monolignol biosynthesis and mutants with reduced CAD activity results in higher bioethanol production without a growth penalty. Seven CAD genes have been identified in Brachypodium and BdCAD5 was identified as *bona fide* with highest expression rate in all tissue. All BdCAD genes are cytosolic except BdCAD4, which is located in the chloroplasts and the function of this gene is still unclear. In this study the function of BdCAD4 and BdCAD5 will be investigated by using the genome editing tool CRISPR/Cas9 to alter the reading frame by induced mutations. Three individual sgRNA targets for each gene were selected and the activities were tested in protoplast, followed by callus transformation.

Keywords: Brachypodium, CRISPR/Cas9, cinnamyl alcohol dehydrogenase (CAD), lignin, bioethanol

Session 1: Novel tools and resources

A simple and versatile genome-wide SNP genotyping by multiplex PCR targeted amplicon sequencing in *Brachypodium distachyon*

Yoshihiko Onda^{1,2}, Kotaro Takahagi^{1,2,3}, Minami Shimizu¹, Komaki Inoue¹, Keiichi Mochida^{1,2,4,5}

Affiliations:

¹Cellulose Production Research Team, RIKEN Center for Sustainable Resource Science, Yokohama, Japan

²Kihara Institute for Biological Research, Yokohama City University, Yokohama, Japan

³Graduate School of Nanobioscience, Yokohama City University, Yokohama, Japan

⁴Gene Discovery Research Group, RIKEN Center for Sustainable Resource Science, Yokohama, Japan

⁵Institute of Plant Science and Resource, Okayama University, Kurashiki, Japan

Presenting author email address:

yoshihiko.onda@riken.jp

Abstract:

Next-generation sequencing technologies have enabled genome re-sequencing for exploring genome-wide polymorphisms among individuals as well as targeted re-sequencing for rapid and simultaneous detection of polymorphisms in genes associated with various biological functions. Therefore, a simple and robust method for targeted re-sequencing should facilitate genotyping in a wide range of biological fields. In this study, we developed a simple, custom targeted re-sequencing method, designated ‘multiplex PCR targeted amplicon sequencing’ (‘MTA-seq’), and applied it to genotyping of the *Brachypodium distachyon*. To assess the practical usability of MTA-seq, we applied it to genotyping of genome-wide single nucleotide polymorphisms (SNPs) identified in natural accessions by comparing the re-sequencing data to the reference accession Bd21. Examination of the SNP genotyping accuracy from eight parental accessions and an F₁ progeny revealed that approximately 95% of the SNPs were correctly called. The assessment suggested that the method provides an efficient framework for accurate and robust SNP genotyping. The method described here enables easy design of custom target SNP-marker panels in various organisms, facilitating a wide range of high-throughput genetic applications such as genetic mapping, population analysis, and molecular breeding.

Session 2: Development, epigenetics and growth

**Transcriptional regulation of biomass accumulation in
*Brachypodium distachyon***

Sam Hazen .

University of Massachusetts, USA

Session 2: Development, epigenetics and growth

Identification of the BUZZ kinase involved in root and root hair development

Rachel Dannay¹, Thiel A. Lehman¹, Ying Wu^{1,2}, Rhoda Brew-Appiah¹, Tobias I. Basin³, Karen A. Sanguinet¹

1. Department of Crop and Soil Sciences, Washington State University, Pullman, WA 99164
2. Northeast Normal University, Changchun, China
3. Department of Biology, University of Massachusetts, Amherst, MA 01003

Abstract:

Root development and architecture is crucial for plant adaptation and reproductive success in diverse environments. To gain insight into the molecular and genetic cues involved in modulating root development and architecture in temperate grasses, we identified a root hairless mutant in *Brachypodium distachyon* termed *buzz*. The *buzz* mutant displays a root hairless phenotype with a dramatic increase in root growth rate. We used an NGS approach to identify SNPs associated with the *Bdbuzz* mutant phenotype. We identified a SNP in a previously uncharacterized kinase, which leads to an amino acid substitution in a highly conserved *Gly* residue in the kinase domain. We describe characterization of the *Bdbuzz* mutant. *Bdbuzz* is root-specific and expression analysis is consistent with the phenotype as expression is mainly localized to the root tip. We identified a second *Bdbuzz* allele in with a root hairless phenotype suggesting the original EMS allele is a functional null. We then identified the putative BUZZ ortholog in *A. thaliana*. We describe the function of the *A. thaliana* BUZZ kinase by characterization of two independent T-DNA lines. Together these data show that the function of the BUZZ kinase has diverged in grasses as compared to the model dicot *A. thaliana*.

Keywords : root development, kinase

Session 2: Development, epigenetics and growth

Gene silencing by endogenous and exogenous miRNAs in flowering-time control in *Brachypodium distachyon*

Zhengrui Qin, Liang Wu

Zhejiang University, Hangzhou 310058, China

Abstract

The switch from vegetative to reproductive growth is a critical developmental event in plants. Florigen, a small globular protein encoded by *FLOWERING LOCUS T (FT)*, associates with FD, abZIP transcription factor, and 14-3-3 family proteins to form flowering initiation complex, functioning at the core node in multiple flowering pathways in plants. microRNAs (miRNA), a class of small RNAs with a stem-loop precursor structure, play versatile parts in plant development and adaptation to environments. In *Brachypodium distachyon*, on the one hand, we identified a *Pooideae*-specific miRNA, miR5200, that targets two *FT* orthologs, *FT1* and *FT2*, for mRNA cleavage. miR5200 is highly induced under short-day (SD) conditions, but dramatically repressed in long-day (LD) environments. Our over-producing miR5200 transgenic *B. distachyon* exhibits a significant delay of flowering, whereas interfering its activity by a target mimicry strategy accelerates flowering under SDs, indicating an important role of this endogenous miRNA in photoperiod-mediated flowering-time regulation. On the other hand, we identified two alternative splicing variants of *FT2*, namely *FT2 α* and *FT2 β* . Through introducing artificial miRNAs, we obtained transgenic *B. distachyon* specific silencing *FT2 α* and *FT2 β* , respectively. We found an early flowering transition in *FT2 β* silencing plants, in contrast with a severe delay of flowering onset in *FT2 α* repressing plants, suggesting a negative role of *FT2 β* while a positive role of *FT2 α* in flowering control. Since gene editing at the genome level cannot get a loss-of-function mutant of specific splicing variant, our approach by introducing an exogenous miRNA provides a useful tool to inhibit a certain splicing variant activity in transgenic plants to explore its biological relevance. Taken together, we characterize an endogenous miRNA that modulates two *FT* gene expressions, and take advantage of an artificial miRNA to degrade a specific *FT* splicing variant transcript in flowering-time control in *B. distachyon*.

Session 2: Development, epigenetics and growth

***Brachypodium* tissue culture as a model system to reveal the functions of the components of the cell wall**

Alexander Betekhtin¹, Magdalena Rojek¹, Anna Milewska-Hendel², Robert Gawecki², Jagna Karcz³, Ewa Kurczynska², Robert Hasterok¹

¹ Department of Plant Anatomy and Cytology, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 28 Jagiellonska Street, 40-032, Katowice, Poland,

² Department of Cell Biology, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 28 Jagiellonska Street, 40-032 Katowice, Poland,

³ Scanning Electron Microscopy Laboratory, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 28 Jagiellonska Street, 40-032 Katowice, Poland

abetekht@us.edu.pl

Abstract:

Brachypodium Beauv. is believed to be one of the oldest genera within the Poaceae. This genus is predominantly distributed in Europe and Asia with disjunctions occurring in Central America and Southern Africa. *Brachypodium distachyon* (*Brachypodium*) is a model system for functional genomics in grasses. Although there are some studies of invitro *Brachypodium* cultures including somatic embryogenesis, detailed knowledge of the composition of the components of the main cell wall in the embryogenic callus in this species is lacking. Therefore, we used histological, scanning electron microscopy as well as the immunocytochemical approach against the arabinogalactan proteins (AGP), extensins and hemicelluloses to understand the localisation and possible functions of these cell wall components during the embryogenic mass that appears in *Brachypodium* callus. We found that the distribution of pectins, AGPs and hemicelluloses can be used as molecular markers of the embryogenic cells. Furthermore, we showed that AGPs and pectins are components of the extracellular matrix. The presented data extends our knowledge about the chemical composition of the embryogenic cells in the *Brachypodium* callus. This work was supported by the National Science Centre, Poland [grant no. DEC-2014/14/M/NZ2/00519].

Session 2: Development, epigenetics and growth

A FLOWERING LOCUS C homolog is a vernalization regulated repressor in Brachypodium and is cold-regulated in wheat

Neha Sharma¹, Philip Ruelens¹ Mariëlla D'hauw², Thomas Maggen², Niklas Dochy¹, Sanne Torfs¹, Kerstin Kaufmann³, Antje Rohde², and Koen Geuten^{1*}

1. Department of Biology, KU Leuven, B-3001 Leuven, Belgium (N.S., P.R., N.D., S.T., K.G.);

2. Bayer Crop Science, B-9052 Gent, Belgium (M.D., T.M., A.R.); and

3. Institute for Biochemistry and Biology, Potsdam University, 14476 Potsdam-Golm, Germany (K.K.)

Koen Geuten

Abstract:

Winter cereals require prolonged cold to transition from vegetative to reproductive development. This process, referred to as vernalization, has been extensively studied in *Arabidopsis* (*Arabidopsis thaliana*). In *Arabidopsis*, a key flowering repressor called *FLOWERING LOCUS C* (*FLC*) quantitatively controls the vernalization requirement. By contrast, in cereals, the vernalization response is mainly regulated by the *VERNALIZATION* genes, *VRN1* and *VRN2*. Here, we characterize *ODDSOC2*, a recently identified *FLC* ortholog in monocots, knowing that it belongs to the *FLC* lineage. By studying its expression in a diverse set of *Brachypodium* accessions, we find that it is a good predictor of the vernalization requirement. Analyses of transgenics demonstrated that *BdODDSOC2* functions as a vernalization-regulated flowering repressor. In most *Brachypodium* accessions *BdODDSOC2* is down-regulated by cold, and in one of the winter accessions in which this down-regulation was evident, *BdODDSOC2* responded to cold before *BdVRN1*. When stably down-regulated, the mechanism is associated with spreading H3K27me3 modifications at the *BdODDSOC2* chromatin. Finally, homoeolog-specific gene expression analyses identify *TaAGL33* and its splice variant *TaAGL22* as the *FLC* orthologs in wheat (*Triticum aestivum*) behaving most similar to *Brachypodium ODDSOC2*. Overall, our study suggests that *ODDSOC2* is not only phylogenetically related to *FLC* in eudicots but also functions as a flowering repressor in the vernalization pathway of *Brachypodium* and likely other temperate grasses. These

Session 2: Development, epigenetics and growth

The preferential silencing of *B. stacei*-inherited rRNA genes in *Brachypodium hybridum* - an epigenetic point of view

Natalia Borowska-Zuchowska¹, Ewa Robaszkiewicz¹, Mirosław Kwasniewski², Alexander Betekhtin¹, Robert Hasterok¹

¹ Department of Plant Anatomy and Cytology, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 40-032 Katowice, Poland.

² Department of Genetics, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, 40-032 Katowice, Poland.

Presenting author email address: natalia.borowska@us.edu.pl

Abstract:

Nucleolar dominance (ND) was among the first epigenetic phenomena that had been described. It was found in plant and animal allopolyploids and/or hybrids and consisted of reversible silencing of 35/45S rRNA gene loci inherited from one of the two or more ancestral species. Recent studies revealed that the large-scale silencing of rRNA genes via ND is of an epigenetic origin. However, the mechanisms according to which one parental rDNA set is chosen to be silenced still remain unclear.

Brachypodium hybridum is a natural allotetraploid (2n=30) with putative parental genomes originating from two diploid species: *B. distachyon* (2n=10) and *B. stacei* (2n=20). Selective silencing of *B. stacei*-like rDNA loci was observed in this allopolyploid. This presentation outlines the studies on ND mechanisms in several *B. hybridum* genotypes originated from distinct geographic locations. The distribution of 35S rDNA loci inherited from ancestral species was determined in both mitotic and meiotic cells. Moreover, we aimed to investigate the epigenetic status of 35 rRNA gene loci in *B. hybridum* and its putative ancestral species by the determination of DNA methylation and selected histone modification (e.g. H4K5ac, H4K16ac, H3K9ac, H3K9me2) immune patterns. We also show the results of molecular characterisation of intergenic spacers (IGS) between 25S rDNA and 18S rDNA in *B. hybridum* and its progenitors as well as their physical localisation in metaphase chromosomes and interphase nuclei. In all IGS sequences we identified putative transcription initiation sites and spacer promoters followed by subrepeats.

Session 3: Natural variation and evolution

Population structure of the Brachypodium species complex and genome wide dissection of agronomic traits in response to climate

Pip Wilson, Jared Streich, Steve Eichten, Riyan Cheng, Kevin Murray, Niccy Aitkin, Kurt Spokas, Norman Warthmann, Accession Contributors*, Justin Borevitz

Abstract

The development of model systems requires a detailed assessment of standing genetic variation across natural populations. The Brachypodium species complex has been promoted as a new plant model for grass genomics with translational to small grain and energy crops. To capture the global genetic diversity within this species complex, thousands of Brachypodium accessions from around the globe were collected and sequenced using genotyping by sequencing (GBS). Samples were initially separated into two diploid or allopolyploid species defining overlapping and invasive ranges and climate niches. A core set of high diversity *B. distachyon* diploid lines were selected for whole genome sequencing and high resolution phenotyping. Genome-wide association studies was used to identify candidate genes and pathways tied to key fitness and agronomic related traits. A total of 9, 22 and 47 QTLs were identified for flowering time, early vigour and energy traits, respectively. Overall, the results highlight the genomic structure of the species complex and allow powerful complex trait dissection within an emerging model species.

Keywords: GWAS, hapmap, cryptic species

Accession Contributors

ShuangShuang, LiuKent Bradford PI, SmadarEzratiPI, HikmetBudak PI, Diana Lopez, Pilar Catalan PI, David Garvin PI, John Vogel PI, Sean Gordon, Sam Hazen PI, Luis Mur PI

Session 3: Natural variation and evolution

Insertion/deletion markers for assessing the genetic variation and the spatial genetic structure of Tunisian *Brachypodium hybridum* populations

Mhemmed Gandour^{1,4*}, Mohamed Neji^{1,2}, Yosra Ibrahim¹, Wael Taamalli¹, Sean P. Gordon³, John P. Vogel³, Chedly Abdelly¹, Filippo Geuna²

¹Laboratory of Extremophile Plants, Centre of Biotechnology of Borj-Cédria, BP 901 HammamLif 2050 Tunisia,

²Department of Agricultural and Environmental Sciences DISAA, Laboratory of Molecular Genetics, University of Milan, Via Celoria 2, 20133 Milan, Italy,

³USDA-ARS Western Regional Research Center, 800 Buchanan St., Albany, CA 94710, USA,

⁴Faculty of Sciences of SidiBouزيد, University of Kairouan, Tunisia

Corresponding author: gandourmed@yahoo.fr

Abstract

The wild annual grass *Brachypodium distachyon* has been widely investigated across the world as a model plant for the temperate cereals and biofuel grasses. This annual plant shows three cytotypes that have been recently recognized as three independent species, the diploid species *Brachypodium distachyon* ($2n = 10$) and *Brachypodium stacei* ($2n = 20$) and their derived allotetraploid *B. hybridum* ($2n = 30$). The last species appear to be the most relevant in Tunisia. In order to analyze the genetic structure and the ecogeographical adaptation of this species, it is necessary to increase the number of polymorphic markers currently available for the species. In this work, the possibility of using syntenic *Brachypodium* indels as a new source of markers for this purpose has been explored. From 24 *B. distachyon* indels tested for transferability and polymorphism in the *B. hybridum*, 11 primer pairs (45%) gave cross-species transferability and 8 primer pairs (33%) showed polymorphism. The latter were used to examine the spatial distribution of genetic variation of *B. hybridum* across its entire range in Tunisia and to test underlying factors that shaped its genetic variation. Population genetic analyses were conducted on 145 individuals from 9 populations. Indels markers showed a total of 20 alleles overall all loci and a high level of genetic diversity overall populations (average of polymorphism rate

Session 3: Natural variation and evolution

Diversity of Brachypodium samples in Israel revealed by molecular and cytogenetic methods

Tao Lang¹, Guangrong Li¹, Bin Li¹, Zhihui Yu¹, Hongjin Wang¹, Eviatar Nevo², Zujun Yang^{1*}

¹School of Life Science and Technology, University of Electronic and Technology of China, Chengdu 610054, China

²Institute of Evolution, University of Haifa, Mount Carmel, Haifa 31905, Israel

Presenting author email address: yangzujun@uestc.edu.cn

Abstract

As a temperate wild grass species, *Brachypodium distachyon*(2n=10) is becoming a promising model organism for studies of grass genomics and grain crop improvement. The annual species of genus Braypodium with 2n=20 and 2n=30 were described as species of *B. stacei* and *B. hybridum*, respectively. The characterization of diversity of natural populations of Brachypodiumspecies is important for genetic and evolutionary researches. In the present study, we investigated the phenotypical and cytological variation of nine extensively collected natural populations of Braypodium species with different macrogeographic scales originating from Israel. Total 174 genotypes of Braypodium samples were developed. Extensive phenotypical variation including plant height and tiller variation among genotypes were observed. The observation of chromosome number indicated 66 and 108 samples were *B. stacei* and *B. hybridum*, respectively, while no *B.distachyon* species was identified. The genetic diversity of *B. stacei* and *B. hybridum* samples was larger among population (62.1%) than within population (27.9%). Based on the values of Nei's genetic diversity (He) and Shannon's information index (I) correlated with the ecological factors, we found that the distribution of two species was significantly correlated to the environmental ecological factors. The results indicated that the*B. Hybridum* is largely positively correlated with higher environmental climatic stresses of temperature and drought. Identifying chromosomal mechanisms of *Brachypodium* species associated with population genetics and adaptation to climatic variation are needed to advance in

Session 3: Natural variation and evolution

Environmental isolation explains Iberian genetic diversity in the highly homozygous model grass *Brachypodium distachyon*

Authors: Isabel Marques¹, Valeriia Shiposha^{1,2}, Diana López-Alvarez^{1,3}, Antonio J. Manzaneda⁴, Pilar Hernandez⁵, Marina Olonova², Pilar Catalán^{1,2}

1. Departamento de Ciencias Agrarias y del Medio Natural, Escuela Politécnica Superior de Huesca, Universidad de Zaragoza, Ctra. Cuarte km 1, 22071 Huesca, Spain.

2. Department of Botany, Institute of Biology, Tomsk State University, Lenin Av. 36, Tomsk 634050, Russia.

3. Current address: Centro de Bioinformática y Biología Computacional de Colombia, BIOS, Parque los Yarumos, Manizales, Colombia

4. Departamento de Biología Animal, Biología Vegetal y Ecología, Universidad de Jaén, Paraje Las Lagunillas s/n, 23071-Jaén, Spain

5. Instituto de Agricultura Sostenible (IAS-CSIC), Alameda del Obispo s/n, 14004 Córdoba, Spain

Presenting author: lera.forester@mail.ru

Abstract

Brachypodium distachyon, an annual Mediterranean Aluminum (Al)-sensitive grass, has been increasingly investigated as a model plant for temperate cereals, forage grasses and biofuel grass crops. However, despite being a model plant, we still know very little about its genetic diversity. We used nuclear Simple Sequence Repeats (nSSR) to study the patterns of genetic diversity and population structure of *B. distachyon* in 14 populations collected across the Iberian Peninsula. We detected very low levels of genetic diversity, allelic number and heterozygosity in *B. distachyon*, congruent with a highly selfing system. Our results indicate the existence of at least three genetic clusters; populations growing on basic soils (NE and S Spain) were significantly more diverse than those growing in acidic soils (NW Spain). A partial Mantel test confirmed a statistically significant Isolation-By-Distance (IBD) among all studied populations, as well as a statistically significant Isolation-By-Environment (IBE), revealing the presence of environmental-driven isolation as one explanation for the genetic patterns found in the Iberian Peninsula. Despite the low values of allelic and genetic diversity and the low levels of heterozygosity detected in *B.*

Session 4: Plant-biotic and abiotic interactions

Generation of *Brachypodium distachyon* T-DNA mutant population for studying nonhost resistance to wheat stripe rust

Suzhen Zhao, TianyueAn, Guoan Shen, Yingchun Zhang, Xiaoquan Qi

Key Laboratory of Plant Molecular Physiology, Institute of Botany, Chinese Academy of Sciences, Nanxincun 20, Fragrant Hill, Beijing 100093, China. xqi@ibcas.ac.cn

Abstract:

Brachypodium distachyon has become a model system for temperate grasses' functional genomics research. Establishing a large insertion mutant population was very important for functional genomics. Here we reported the generation of about 7,000 T-DNA insertion lines based on a highly efficient *Agrobacterium*-mediated transformation system. Then a very powerful method for isolating flanking sequences of T-DNA insertion site was developed from the previous inverse PCR. Meanwhile, a serial of Perl scripts were utilized to rapidly process sequence data and identify insertion sites combining with network resources. A total of 794 flanking sequences was isolated and analyzed in detail using this method.

The resource of the T-DNA mutants provided us the convenience to study the molecular resistance mechanism against wheat stripe rust, caused by *Puccinia striiformis* f. sp. *tritici*(PST). The phenotypic evaluation showed that the response of *Brachypodium distachyon* to PST was nonhost resistance (NHR), providing a good plant-pathogen system for study of the immune responses and the molecular mechanism underlying wheat-PST interactions. More than 200 mutant lines that were more susceptible or resistant to the wheat stripe rust were identified from our T-DNA insertion population. The infection type was assessed according to the pathological phenotype. One highly susceptible mutant line, T1415, was isolated and studied thoroughly.

Keywords: *Brachypodium distachyon*, transformation, T-DNA insertion, flanking sequence, wheat stripe rust.

Session 4: Plant-biotic and abiotic interactions

Brachypodium: A useful model host for cereal-fungal pathogen interactions

¹Jonathan J. Powell, ¹Timothy L. Fitzgerald, ¹Mark Turner, ¹Jason Carrere, ¹Anuj Rana, ¹Donald M. Gardiner, ²John P. Vogel, ¹Kemal Kazan

¹CSIRO Agriculture and Food, Queensland Bioscience Precinct, St Lucia, QLD, Australia

²Joint Genome Institute, United States Department of Energy, Walnut Creek, CA, USA

Kemal.Kazan@csiro.au

Abstract:

Fusarium pathogens cause serious yield and quality losses on cereal crops such as wheat and barley. There is no complete resistance against these pathogens and available resistance acts quantitatively, implying that many genes in the host, each with small effects, are involved in disease resistance. We have recently adopted Brachypodium as a model host to dissect cereal-*Fusarium* pathogen interactions. We have first developed a *Fusarium* infection assay for Brachypodium. We have then comparatively analysed the molecular responses of wheat and Brachypodium to *Fusarium* infection by RNA-seq and metabolite analyses. These analyses have revealed significant overlapping responses between Brachypodium and wheat. In addition, to understand the importance of salicylic acid (SA) in defence against *Fusarium* pathogens, we have generated a Brachypodium resource by over-expressing the bacterial *nahG* gene encoding a SA degrading enzyme. *nahG* over-expressing Brachypodium plants with very low basal SA levels were then exposed to pathogen infection and differential gene expression patterns of mock- and pathogen-inoculated plants were analysed by RNA-seq. This enabled us to identify Brachypodium genes that require SA for their induction by *Fusarium* pathogens. Together, our results suggest that Brachypodium is an excellent model host to dissect cereal-fungal pathogen interactions.

Keywords: Fungal defense, *Fusarium*, Biotic stress, Transcriptomics, *nahG*

Session 4: Plant-biotic and abiotic interactions

Linking phenotype to genotype: A metabolomics approach to build trait association network models for Brachypodium

Pubudu P. Handakumbura¹, Albert Rivas-Ubach¹, Bryan A. Stanfill², Daniel C. Fortin², Ljiljana Paša-Tolić¹, John P. Vogel³, Christer Jansson¹.

¹ Environmental Molecular Sciences Laboratory, Pacific Northwest National Laboratory, 3335, Innovation Blvd, Richland, WA, 99354, USA

² Pacific Northwest National Laboratory, 3335, Innovation Blvd, Richland, WA, 99354, USA

³ Joint Genome Institute, 2800 Mitchell Drive, Walnut Creek, CA 94598, USA

Pubudu.handakumbura@pnnl.gov

Abstract:

Prognostic understanding of the terrestrial carbon cycle in the face of global change is necessary to secure sustainable energy, water, and food for our nation and the world. To meet this challenge, we urgently need robust predictive models of net carbon flux in terrestrial ecosystems. A step in this direction is to explore the power of multi-scale plant modeling, where the phenotypic expression at one level is prognostic of emergent properties at the next integrative levels. Plant phenotype is shaped as a function of its genotype and interactions with the environment. Thus, for example, how does the genotype of a plant inform phenotypic expression at the level of metabolite and/or protein profiles, and how do these molecular-level phenotypes inform phenotypic expression at the cellular and organismal levels? Plants produce a wide variety of metabolites/ small molecules that are crucial for its growth and development. Metabolomics, the study of these molecules is becoming an increasingly important and powerful tool in predicting the effects of various aspects of plant physiology and biology, enabling our understanding and knowledge of plant growth, development and stress responses. In this study we have used above ground and below ground dry biomass as phenotypic markers to study carbon cycling under well watered and drought conditions in thirty different *Brachypodium distachyon* accessions coupled with metabolomics to build predictive metabolite-trait models. Here we demonstrate the power of metabolomics in defining the plant genotype and predicting the metabolite drivers of drought tolerance.

Keywords: Metabolomics, Biomass, Drought, Carbon Cycling, Statistical Modeling.

Session 4: Plant-biotic and abiotic interactions

Expression profiling of marker genes for defense-associated phytohormones in *Brachypodium distachyon* highlights its similar immune systems to rice

Authors:

Yusuke Kouzai^{1,2,3}, Yoshihiko Onda^{1,3}, Keiichi Mochida^{2,3,4}, Yoshiteru Noutoshi²

Affiliations:

1 RIKEN Center for Sustainable Resource Science, Yokohama, Japan

2 Graduate School of Environmental and Life Science, Okayama University, Okayama, Japan

3 Kihara Institute of Biological Research, Yokohama City University, Yokohama, Japan

4 Institute of Plant Science and Resources, Okayama University, Okayama, Japan

Presenting author email address:

Yusuke.kouzai@riken.jp

Abstract:

Brachypodium distachyon is a model plant closely related to economically important cereals and biomass crops. It has been started to be used as a platform to study plant disease resistance and can be a counterpart of *Arabidopsis* and rice to illustrate the specificity and commonality of immune systems among plant species. To obtain marker genes to evaluate responses of *B. distachyon* to defense-related phytohormones, 34 candidate marker genes for salicylic acid (SA), jasmonic acid (JA), and ethylene (ET) were selected based on the similarities of protein sequences to the known marker genes of *Arabidopsis* and rice, and analyzed their responsiveness to each phytohormone at 24 and 48 h after treatments. Two genes for SA, 7 for JA, and 2 for ET were significantly induced at either or both time points. Next, we compared phylogenetic relationships and expression profiles of *PR1* family genes among *Arabidopsis*, rice and *B. distachyon*. The constitution and phytohormone responsiveness of *BdPR1* genes were shown to be similar to rice but not *Arabidopsis*, suggesting that monocots share a characteristic immune system, defined as the common defense system in contrast to dicots. Since we recently established a model pathosystem for sheath blight disease using *B. distachyon*, the infection process of *Rhizoctonia solani* on *B. distachyon* was evaluated by using these

Session 4: Plant-biotic and abiotic interactions

Interaction of Bsr1 and TGB1 confers Barley Stripe Mosaic Virus resistance in Brachypodium, barley and wheat

Qihong Wu¹, Yu Cui², Lijie Yan², Guoxin Wang², Xuejiao Jin², Meihua Yu², Ling Wang², Hao Wang², Chen Dang², Panpan Zhang¹, Andrew Jackson³, Dawei Li^{2,*} and Zhiyong Liu^{1,*}

¹ Institute of Genetics and Developmental Biology, Chinese Academy of Science, Beijing 100101, China; ² China Agricultural University, Beijing 100193, China; ³University of California-Berkeley, USA

qhwu@genetics.ac.cn

Abstract:

Barley stripe mosaic virus (BSMV) is a model for studies of viral pathogenesis and movement. *Brachypodium distachyon* is a member of the Poaceae subfamily Pooideae and has emerged as a model species for the study of cool season cereal crops. By applying map-based cloning approach, the first BSMV resistance gene *Bsr1*, encoding a typical CC-NBS-LRR protein, was isolated from *Brachypodium* inbred line Bd3-1. Domain switch of the Bsr1 and bsr1 alleles indicated that the LRR domain and the C terminal are critical region for the BSMV resistance of Bsr1. Transgenic experiments showed that *Bsr1* overexpression transgenic Bd21-3 plants performed BSMV resistance against ND18 strain, indicating that *Bsr1* is the causal gene that confers resistance to BSMV ND18 in Bd3-1. Furthermore, we transformed the complimentary construction pCBsr1 into barley cultivar “Golden promise” and wheat cultivar “Kenong199” (KN199), and the results imply that the foreign gene *Bsr1* has function in barley and wheat and the resistance genes from *Brachypodium* could be an assistant to crop breeding. The allelic variations of *Bsr1* in *Brachypodium distachyon* accessions collected mainly from Turkey-Iraq and *Bsr1* in *B. stacei* and *B. hybridum* accessions collected mainly from Israel were explored. The results conformed that *B. distachyon* and *B. stacei* were the genome donors of *B. hybridum*. Further studies demonstrate that the TGB1_{ND} interacts Bsr1 to stimulate *Bsr1*

Session 4: Plant-biotic and abiotic interactions

Using the JGI Brachypodium T-DNA collection to reveal novel transcription factor roles in abiotic stress responses

Toni Mohr¹, Naxin Huo^{1,2}, Prisca Cheng¹, Hongxia Li¹, Zhiyong Liu³, John Vogel⁴, Yongqiang Gu⁴

¹United States Department of Agriculture-Agricultural Research Service, Western Regional Research Center Albany, California 94710, USA.

²Department of Plant Sciences, University of California, Davis, California 95616, USA.

³Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, China

⁴DOE Joint Genome Institute, Walnut Creek, CA 94598, USA

Presenting author: yong.gu@ars.usda.gov

Abstract:

Abiotic stresses such as drought and heat negatively impact the growth, development, yield, and seed quality of crops and other plants. To understand the molecular basis of plant responses to these adverse stresses, we utilized the JGI Brachypodium T-DNA collection to study the specific functions of transcription factors (TFs) under different environmental conditions. The JGI collection contains 23,649 tagged T-DNA lines in *Brachypodium distachyon* Bd21-3; the T-DNA insertion sites in each mutant line have been mapped and are publically accessible. We identified and obtained 348 lines that contain the activation tag T-DNA construct pJJ2LBA in or near transcription factors. We first screened these lines over two generations to obtain homozygous mutant lines, including target gene expression using Q-PCR. The expression of these target genes was also examined in wild type plants using publically available Brachypodium RNA-seq data generated under different experimental conditions such as cold, drought, salt, and heat. At present, we have a set of 43 lines with different gene expression patterns: 5 are knockouts, 11 are downregulated, and 27 are upregulated compared to Bd21-3. These genes are from several different TF families; most are transcription factors that are poorly understood in temperate grasses. We are intensely screening these lines for phenotypes during heat, drought, and salt stresses. Several promising lines involved in heat sensitivity and salt resistance have been identified, along with various morphological phenotypes. This screening method will allow us to uncover novel roles for genes that may be difficult to detect in classic genetic screens.

Keywords: Brachypodium, T-DNA mutant, Abiotic stresses, Gene expression.

Session 4: Plant-biotic and abiotic interactions

Cool season turf grass heat tolerance study through genomic and genetic analyses with *Brachypodium distachyon*

Zhaoqing Chu

1. ShanghaiChenshan Plant Science Research Center,
2. Shanghai Institutes for Biological Sciences,
3. Chinese Academy of Sciences, China

Session 4: Plant-biotic and abiotic interactions

Homoeolog-specific activation for heat acclimation in the allopolyploid grass *Brachypodium hybridum*

Kotaro Takahagi^{1,2,3}, Komaki Inoue³, Minami Shimizu^{2,3}, Yukiko Uehara-Yamaguchi³, Yoshihiko Onda^{2,3} and Keiichi Mochida^{1,2,3,4}

¹Graduate School of Nanobioscience, Yokohama City University, 22-2 Seto, Kanazawa-ku, Yokohama, Kanagawa 236-0027, Japan.

²Kihara Institute for Biological Research, Yokohama City University, 641-12 Maioka-cho, Totsuka-ku, Yokohama, Kanagawa 244-0813, Japan.

³Cellulose Production Research Team, Biomass Engineering Research Division, RIKEN Center for Sustainable Resource Science, 1-7-22 Suehiro-cho, Tsurumi-ku, Yokohama, Kanagawa 230-0045, Japan.

⁴Institute of Plant Science and Resources, Okayama University, 2-20-1 Chuo, Kurashiki, Okayama 710-0046, Japan.

Presenting author email address: keiichi.mochida@riken.jp

Abstract:

Allopolyploid plants often show wider environmental tolerances than their ancestors; this difference would be expected due to the merger of multiple distinct genomes with a fixed heterozygosity. The allopolyploid grass *Brachypodium hybridum* and its ancestor *Brachypodium stacei* show long-term heat stress tolerance, unlike its another ancestor *Brachypodium distachyon*. To understand physiological differences between these species, we compared the transcriptome of the allopolyploids and its ancestors grown under normal and heat stress conditions. We found that *B. distachyon* was transcriptionally insensitive, whereas *B. hybridum* and *B. stacei* were sensitive to heat at 3 days after stress exposure, and at 15 days after heat exposure, *B. hybridum* and *B. stacei* maintained transcriptional states similar to those under normal conditions. These results suggested an earlier response to heat that was specific to homoeologs originating from *B. stacei* and that contributed to cellular homeostasis under long-term heat stress. Our results provide insights into different regulatory events of the homoeo-transcriptome that are associated with stress acclimation in allopolyploid plants that evolved through all opolyploidization.

Keywords: all opolyploidization, *Brachypodium hybridum*, abiotic stress, transcriptome

Session 4: Plant-biotic and abiotic interactions

Influence of Supplemental Lighting with Different Light Quality on the Turf Growth of *Festuca Arundinacea*

CAI Jinshu, ZHU Jiangli, XU Nuo, XIE Yunjun, CAO Huaying, LIU Qi
(*ShezhenWenke Landscape Corp.,Ltd. Shenzhen, Guangdong518048, China*)

Author for correspondence (E-mail: sg_cai@163.com; Tel: 18973165482)

Abstract:

Plant factory was widely used in developed countries, which provided a new model for factory production of soil-free turf. As light was an indispensable factor for plant growth, artificial supplemental lighting was the most effective way to improve the light deficiency in plant factory. In order to study the effects of supplemental lighting with different light quality on turf grass growth, four kinds of light with red and blue 3: 1 (3R/B), red and blue 4: 1 (4R/B), red and blue 5: 1 (5R/B) and full spectrum (control) were used in the experiment as supplemental lights for 10 hours each day. The soil-free substrate was mixed by the same volume of coir dust and straw ash. The seeds of *Festuca Arundinacea* were sown by 30 grams per square meter on December 9, 2016. The laboratory temperature was 22°C up to 25°C, the relative humidity 70% up to 80%. The LED lamps were about 50cm above the canopy of the turf. The experiment was ended on January 16, 2017 and the average stem diameter, leaf width, underground biomass, coverage and turf color index was tested.

Results showed that the average stem diameter decreased gradually with the increase of red light proportion. The stem diameter of the control plant was the thickest (2.04mm), and that treated with 5R/B was the weakest. The average leaf-breadth of the plant treated with 3R/B was 3.47mm, about the same of the control. Leaf width of 3R/B treatment was significantly higher than those of 4R/B and 5R/B treatments.

Results also showed that underground biomass reduced with the increase of red light proportion. The underground biomass of the control (687.00mg) was significantly higher than other treatments, 498.87mg more than 5R/B treatment (188.13mg). And the turf color index decreased with the increase of red light proportion. The turf color index of 3R/B

Session 4: Plant-biotic and abiotic interactions

Establishing *Brachypodium distachyon* as a model in analyses of plant genome stability after mutagenic treatment

Arita Kus^{1*}, Jolanta Kwasniewska¹, Robert Hasterok¹

¹: Department of Plant Anatomy and Cytology, Faculty of Biology and Environmental Protection, University of Silesia in Katowice, Poland

*: arkus@us.edu.pl

Abstract:

Due to their high sensitivity higher plants are widely used for screening and monitoring environmental genotoxicity. *Brachypodium distachyon*, an internationally accepted model grass species, would be a convenient system in mutagenesis to analyse “hot spots” of DNA damage in nuclear genome and consequently could find practical application in the environmental monitoring. The chromosome rearrangements are commonly identified using classical cytogenetic techniques. Physical mapping technology together with the availability of BAC libraries of *B. distachyon* nuclear DNA, allow comprehensive analyses of mutagenic effects at the chromosomal level and extend our understanding of the mechanisms of chromosomal aberrations. The visualisation of mutagen-induced genome changes, including micronuclei formation and alterations of chromosome territories in interphase nuclei using fluorescence *in situ* hybridisation (FISH) with selected chromosome-specific BAC clones, as well as ribosomal DNA and chromosome region-specific, i.e. centromeric and telomeric probes are presented. This work was supported by the Polish National Science Centre (grant no. 2012/04/A/NZ3/00572).

Keywords: *Brachypodium distachyon*, FISH, micronuclei, mutagenesis, model grass

Phenotypic and metabolomic variation in the model annual grasses *Brachypodium distachyon*, *B. stacei*, and *B. hybridum*

Diana López-Álvarez^{1,2}, Hassan Zubair³, Manfred Beckmann³, John Draper³, Luis Villar⁴, Pilar Catalan¹

1. Departamento de Ciencias Agrarias y del Medio Natural, Escuela Politécnica Superior de Huesca, Universidad de Zaragoza, Ctra. Cuarte km 1, 22071 Huesca, Spain.

2. Current address: Centro de Bioinformática y Biología Computacional de Colombia, BIOS, Parques Yarumos, Manizales, Colombia

3. Institute of Biological, Environmental and Rural Sciences, Aberystwyth University, Aberystwyth, UK

4. Instituto Pirenaico de Ecología (CSIC), Jaca, Spain

Presenting author: pcatalan@unizar.es

Abstract:

Morphological traits and metabolite fingerprinting were used to investigate inter- and intra species variation within the model annual grasses *B. distachyon*, *B. stacei* and *B. hybridum*. Phenotypic variation of 15 quantitative and 5 qualitative morphological characters and 2219 nominal mass (m/z) signals generated using Flow Infusion Electrospray ionisation - Mass Spectrometry were analysed in individuals from 174 natural populations and 6 inbred lines, and 12 Iberian lines of the three species, respectively. Basic statistics and multivariate Principal Component Analysis (PCA) and Discriminant Analysis (DA) were used to differentiate inter- and intraspecific variability of phenotypic and metabolomic variables, and their association was assayed with *rcorr*. Eight quantitative [(stomata) leaf guard cell length, pollen grain length, (plant) height, second leaf width, inflorescence length, number of spikelets per inflorescence, lemma length, awn length] and five qualitative (leaf color, softness, shape, and hairiness, and presence of short rhizomes) phenotypic characters, and 434 tentatively annotated metabolite signals significantly discriminated the three species. The three species showed different metabolomic profiles. DA significantly discriminated the three taxa with both, morphometric and metabolome traits and the intraspecific phenotypic diversity within *B. distachyon* and *B. stacei*. The populations of *B. hybridum* were considerably less differentiated. Highly explanatory metabolite signals together with morphological characters revealed concordant patterns of differentiation of the three taxa. Significant association was found for pollen grain

How does *CUC2* regulate leaf serration development in *Arabidopsis*?

Zhongjuan Zhang, Sonja Ravanelli, Gemma Bilsborough, Miltos Tsiantis

Department of Comparative Development and Genetics, Max Planck Institute for Plant Breeding Research, Cologne, Germany

Abstract:

A key question in developmental biology is how developmental patterning generates the final form of an organ. *Arabidopsis thaliana* produces simple leaves bearing repeated marginal outgrowths termed serrations, providing a good opportunity to study this question. The NAC domain transcription factor *CUC2* regulates serration formation by promoting the formation of PIN1 convergence points and auxin activity maxima along the leaf margin during leaf development (Bilsborough et al., 2011). However, how *CUC2* regulates repeated formation of auxin maxima and serrations remains enigmatic. To address this question, we performed an EMS-mutagenesis screen in a *cuc2-3* mutant background to identify novel components in regulating serration formation, specifically looking for suppressors that restore serration development. Among the suppressors identified, we primarily investigate #51 because it displays the strongest suppression effect and no other developmental defects. We found that #51 could restore auxin maxima and PIN1 convergence points along the *cuc2* leaf margin. Currently we are trying to identify the molecular basis for the #51 mutation and understand how the gene defined by this mutation regulates PIN1 convergence point and auxin maxima formation.

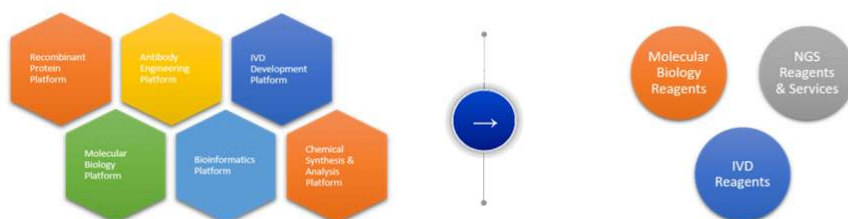
Keywords: Leaf serration, CUC2



Vazyme Biotech is a supplier of best-quality reagents for life science research at very competitive prices. The elite R&D team of Vazyme, lead by a group of Ph.D.s, has devoted to enzyme innovation for over a decade. With years of product development experience, our manufacturing team is tirelessly devoted to producing the highest product quality. In addition, Vazyme has an experienced sales and marketing team to best serve your research needs.

Vazyme now manufactures various products, including molecular biology and next generation sequencing (NGS) reagents. The efficiency and quality of all these products are guaranteed by numerous validation tests before they reach the market. Most of Vazyme products have been cited by top-notch academic journals.

Vazyme's mission is to better serve the globe research community of life science by providing best-in-class and cost-effective research tools.



Pics: The Headquarters Building of Vazyme in Nanjing, China



Innovation in Enzyme Technology

Vazyme Biotech Co.,Ltd

www.vazyme.com

E-mail: sales@vazyme.com

Tel: 400-600-9335

Add: NanjingState Economy & Technology Development Zone, Red Maple Technology Industrial Park, building C 1-2

