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Two new diterpenoid alkaloids from *Aconitum cochleare*

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Two new diterpenoid alkaloids, cochleareine (1) and acoleareine (2) together with the known alkaloids 14-acetylalatisamine (3) and talatisamine (4) have been isolated from the aerial parts of *Aconitum cochleare* Woroschin growing wild in Eastern Turkey (Van).

1. Introduction

There are four *Aconitum* species (Ranunculaceae) growing wild in Turkey, *A. orientale* Mill., *A. nasutum* Fisch. et Reichb., *A. anthora* L. and *A. cochleare* Woroschin. The first three plants together with a plant from Pakistan *A. leave* Royle have been studied by our group (Ulubelen et al. 1996; Meriçli et al. 1996a, 1996b; Meriçli et al. 2000a, 2000b; Ulubelen et al. 2002). In the present study we investigated the last Turkish species namely *A. cochleare*. The plant was collected from Eastern Turkey (Van). A literature survey has revealed that *Aconitum* preparations have been used as cardiotonics, febrifugics, sedatives and anodynes for many centuries (Benn et al. 1983; Bisset 1981). However, in Turkey they are not included in folk medicine and are used only as pain relievers under physicians' control (Baytop 1984).

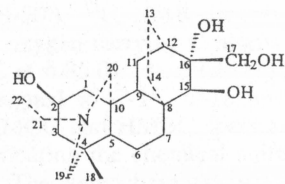
2. Investigations, results and discussion

The powdered plant material (526 g) was macerated with EtOH, filtered and evaporated to dryness. The crude extract was processed to obtain a crude alkaloidal mixture. This mixture was separated by centrifugally accelerated radial TLC (Chromatotron) using Al₂O₃ plates (Merck Art.1092) (Desai et al. 1985). Here we report the isolation and identification of three norditerpene and one diterpene alkaloids in the order 14-acetylalatisamine (3), talatisamine (4) (Konno et al. 1982), acoleareine (2) and cochleareine (1).

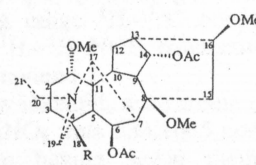
The structures of the compounds were deduced by extensive ¹H and ¹³C NMR studies. The unambiguous chemical shift assignments of 1 and 2 were achieved through a study of their ¹H detected by 2D NMR experiments ¹H-¹H COSY, HMQC, HMBC, and NOESY.

The first new compound, cochleareine (1), has a rather unusual structure. Its HRMS indicated the molecular formula C₂₂H₃₇NO₄ m/z 379.2658. The lack of methoxy group(s) suggested a C₂₀ diterpenoid alkaloid, but the absence of typical exo-methylene signals at C-16 in its ¹H NMR and the presence of an ethyl moiety attached to the nitrogen atom were at first confusing.

The ¹³C NMR (DEPT, APT) findings showed the presence of two methyl quartets, eleven methylene triplets, five methine doublets and four quaternary singlets correlated with the presence of 22 C atoms in the molecule. The ¹H NMR spectrum was quite indicative of the structure of 1, as δ 0.79 (3H, s, Me-18), 1.19 (3H, t, J = 7.0 Hz, Me-22), 4.02 (1H, d, J = 11.5 Hz, H-17), 3.63 (1H, d, J = 11.5 Hz, H-17') indicated the presence of a hydroxymethylene group (CH₂OH) instead of the typical exomethylene group at C-16, and the ¹³C NMR signal at δ 66.56 corresponds to the C atom of CH₂OH group. There are only a few examples having a CH₂OH group at C-16, in the compounds dictysine (Joshi et al. 1987) and

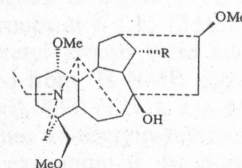


1



2

R
2 CH₂OMe
5 CH₃



R
3 OAc
4 OH

Table 1: NMR data of cochleareine (1) (in CD₃OD, J values (Hz) in parentheses)

	¹³ C	¹ H	COSY	HMBC	NOESY
1	33.82 t	1.70 m	H-2α		
2α	67.38 d	3.89 dd (6.6, 11.1)	H-1α, H-1β	C-10, C-4, C-5	H-22α, H-3α
3	41.88 t	1.95 m	H-2α		H-22α, H-6α
4	37.74 s	—			
5β	35.42 d	1.70 m	H-6α, H-6β	C-18, C-6, C-9	H-11β, H-12β
6	30.70 t	1.30 m	H-5β		
7	27.27 t	1.25 m	H-6α, H-6β		
8	41.76 d	—			
9	42.98 d	2.20 d (4.9)	H-11α, H-11β		
10	52.00 s	—			
11	21.30 t	1.35 dd (8.0, 17.3)	H-9, H-12		H-21β
12	41.67 d	2.06 m	H-11α, H-11β		
13	23.19 t	1.22 m 1.34 m	H-14		
14	23.79 t	2.30 m	H-13	C-12	
15α	85.01 d	3.93 s			
16	79.40 s	—			
17	66.56 t	4.02 d (11.5)			H-17'
17'		3.63 d (11.5)			H-17
18	24.96 q	0.79 s			
19	51.32 t	1.3 m			
20	57.05 t	2.50 d (12.0)		C-2, C-6	H-20'
20'		2.80 d (12.0)			H-20
21	51.94 t	2.75 dd (7.5, 13.8)			H-22, H-11α
21'		2.96 dd (7.5, 13.8)			
22	11.42 q	1.19 t (7.0)			

Table 2: NMR data of acoleareine (2) (in CDCl₃, J values (Hz) in parentheses)

	¹³ C	¹ H	COSY	HMBC
1β	84.0 d	3.20 dd (10.0, 7.0)	H-2α, H-2β	C-6, C-17
2	27.3 t		H-1β, H-3α, H-3β	
3	37.5 t		H-2α, H-2β	
4	34.1 s			
5β	56.4 d	1.40 brs	H-6α	
6α	73.4 d	5.27 t (7.0)	H-5β, H-7β	
7β	42.0 d	2.62 d (7.2)	H-6α	
8	78.5 s			
9	40.9 d			C-10
10	45.9 d			C-9, C-12
11	48.4 s			
12	27.9 t			C-9, C-13
13	39.0 d			C-12
14β	76.4 d	4.80 t (4.5)	H-13	C-16, C-10
15	35.6 t			C-16
16α	83.9 d			C-15
17	63.8 d	2.92 d (2.0)		
18	80.1 t	3.65 d (10.0)		
18'		3.50 d (10.0)		
19	57.4 t	3.04 d (12.7)		
19'		2.59 d (12.7)		
20	48.5 t	2.25 m		
21	13.4 q	0.83 t (7.0)		
OMe-1	56.0 q	3.15 s		
OMe-8	47.9 q	3.29 s		
OMe-16	56.4 q	3.19 s		
OMe-18	59.2 q	3.29 s		
OAc-6	171.0 s	1.99 s		
	22.0 q			
OAc-14	172.1 s	2.10 s		
	21.5 q			

macrocentrine (Benn et al. 1987). Other ¹H NMR signals were at δ 3.93 (1H, s, H-15α), 3.89 (1H, dd, J = 6.6 and 11.1 Hz, H-2α) indicating the presence of hydroxy groups at C-15 and C-2, and at δ 2.75 (1H, dd, J = 7.5 and 13.8 Hz, H-21), 2.96 (1H, dd, J = 7.5 and 13.8 Hz, H-21'), 2.50 (1H, d, J = 12.0 Hz, H-20) and 2.80 (1H, d, J = 12.0 Hz, H-20'). ¹³C NMR signals showed the presence of three oxygen carrying C atoms in addition to the CH₂OH group, at δ 85.01 (d), 79.40 (s) and 67.38 (d) and these were assigned to C-15, C-16 and C-2 respectively by studying HMQC and HMBC spectral data (Table 1) as well as by comparing the chemical shifts of dictysine and macrocentrine. The stereochemistry at C-2, C-15 and C-16 was decided by NOESY studies. ¹H-¹H primary connectivities in the COSY spectrum and the ¹H-¹³C correlations in the HMQC spectrum gave the structure of compound 1. Long range ¹H-¹³C coupling in the HMBC spectrum and ¹H-¹H NOE connectivities confirmed the structure of compound 1.

The second new alkaloid, acoleareine (2), had a molecular formula C₂₉H₄₅NO₈ m/z 535.1045 correlated with the ¹³C NMR spectrum having seven methyl quartets, seven methylene triplets, ten methine doublets and five quaternary carbon singlets. The ¹H NMR spectrum showed a methyl triplet at δ 0.83 (3H, t, J = 7.0 Hz, Me-21), two acetyl methyl signals at δ 1.99 (3H, s), 2.10 (3H, s) and four methoxy groups at δ 3.15 (3H, s), 3.19 (3H, s), 3.29 (6H, s). The acetyl groups were situated at C-6 and at C-14 as followed from ¹H NMR signals at δ 5.27 (1H, t, J = 7.0 Hz, H-6α), 4.80 (1H, t, J = 4.5 Hz, H-14β). Compound 2 resembles 14-acetylperegrine (De la Fuente et al. 1995) with the exception of the presence of a methoxymethylene group instead of a methyl group attached to C-4. The ¹H and ¹³C NMR findings were quite similar to those of 14-acetylperegrine (5), except for the C-18 signals (see Table 2). MS degradation also showed the pre-

sence of two acetyl groups in the molecule. HMBC experiments indicated the placement of the acetyl groups at C-6 and C-14, and the ^1H NMR shifts were also indicative of these positions as observed in a number of examples in the literature (Kulanthaivel et al. 1986; Aiyar et al. 1986).

3. Experimental

3.1. Equipment

IR spectra were recorded on a Perkin-Elmer Model 983 spectrometer. Optical rotations were determined in an Opt. Act. Ltd. AA-5 polarimeter. ^1H NMR (400 MHz) and ^{13}C NMR (100 MHz) were recorded on a Varian Mercury-Vx instrument. EIMS and HRMS were recorded on a JMS 600H spectrometer.

3.2. Plant material

The aerial parts of *A. cochleare* Woroschin were collected from Eastern Turkey (Van, Güzeldere Pass) at an elevation of 2100 m in July 2002 and identified by one of us (F.Ö.). A voucher specimen is deposited in the Herbarium of Faculty of Science and Letter, Balikesir University FS 10947.

3.3. Extraction and isolation

The powdered plant material (526 g) was macerated with EtOH, left to stand for 48 h and filtered, this procedure being repeated 3 times. The extract was evaporated under vacuum and processed as given by Pelletier et al. (1985). A crude alkaloidal mixture (0.9 g) was obtained. Using a Chromatotron apparatus this mixture was fractionated on Al_2O_3 60 GF₂₅₄ neutral (Typ E) radial plates (Merck Art. 1092) (Desai et al. 1985) and eluted with a gradient of petroleum ether, chloroform and methanol. By using TLC plates (Merck Art. 5554) cochleareine (1, 12 mg), acoleareine (2, 7 mg), 14-acetyltalatisamine (3, 25 mg) and talatisamine (4, 20 mg) were isolated. The known compounds were identified by comparing their ^1H and ^{13}C NMR data to those of authentic samples and by co-TLC behavior with standards.

3.4. Cochleareine (1)

$[\alpha]_{\text{D}} = -25^\circ$ (c 0.4, MeOH); IR ν_{max} : 3412, 2928, 2355, 1643, 1573, 1453, 1409, 1261, 1218, 1051, 802, 763. ^1H NMR (CD_3OD) (see Table 1), ^{13}C NMR (CD_3OD) (see Table 1), EIMS (rel. int.) at m/z 379 $[\text{M}]^+$ (3), 348 $[\text{M}-\text{CH}_2\text{OH}]^+$ (70), 319 $[\text{348}-\text{C}_2\text{H}_5]^+$ (68), 286 (72), 258 (23), 228 (17), 186 (100), 158 (12), 122 (16). HRMS: 379.2658 $\text{C}_{22}\text{H}_{37}\text{NO}_4$ (calc. 379.2654).

3.5. Acoleareine (2)

$[\alpha]_{\text{D}} = -42^\circ$ (c 0.2, MeOH); IR ν_{max} : 2930, 2854, 1730, 1461, 1377, 1259, 1160, 1110, 1093, 992, 979, 870. ^1H NMR (CDCl_3) (see Table 2), ^{13}C NMR (CDCl_3) (see Table 2), EIMS (rel. int.) at m/z 535 $[\text{M}]^+$ (4), 491 $[\text{M}-\text{CH}_2\text{OCH}_3 + \text{H}]^+$ (5), 462 $[\text{491}-\text{C}_2\text{H}_5]^+$ (4), 404 $[\text{462}-\text{OAc} + \text{H}]^+$ (75), 345

$[\text{404}-\text{OAc}]^+$ (10), 314 $[\text{345}-\text{OCH}_3]^+$ (15), 283 $[\text{314}-\text{OCH}_3]^+$ (11), 251 $[\text{283}-\text{OCH}_3-\text{H}]^+$ (19), 233 (21), 207 (24), 167 (28), 141 (35), 129 (51), 115 (67), 91 (100), 77 (70), 67 (65). HRMS: 535.1045 $\text{C}_{29}\text{H}_{45}\text{NO}_8$ (calc. 535.1044)

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References

- Aiyar VN, Kulanthaivel P, Benn M (1986) The C_{19} -diterpenoid alkaloids of *Aconitum delphinifolium*. *Phytochemistry* 25: 973–975.
- Baytop T (1984) Therapy with medicinal plants in Turkey (Past and present) No 3255, Istanbul University Press, Istanbul, p. 312.
- Benn MH, Jacyno JM (1983) The toxicology and pharmacology of diterpenoid alkaloids. In: Pelletier SW (ed.) *Alkaloids: Chemical and biological perspectives*, Vol. 1, John Wiley and Sons, New York, p. 153.
- Benn MH, Okanga F, Richardson JF, Munavu RM (1987) Macrocentrine: an unusual diterpenoid alkaloid. *Heterocycles* 26: 2331–2334.
- Bisset NG (1981) Arrow poisons in China. Part II. *Aconitum*-botany, chemistry and pharmacology. *J Ethnopharmacol* 4: 247–336.
- Desai HK, Joshi BS, Panu AM, Pelletier SW (1985) Separation of diterpenoid alkaloid mixtures using Chromatotron. *J Chromatogr* 322: 223–227.
- De la Fuente G, Meriçli AH, Ruiz-Mesia L, Ulubelen A, Meriçli F, Ilarslan R (1995) Norditerpenoid alkaloids of *Delphinium munzianum*. *Phytochemistry* 39: 1467–1473.
- Joshi BS, Wunderlich JK, Pelletier SW (1987) Carbon-13 NMR spectroscopy in the elucidation of structures of diterpenoid alkaloids. *Can J Chem* 65: 99–103.
- Konno C, Shirasaka M, Hikino H (1982) Structure of senbusine A, B and C. Diterpene alkaloids of *Aconitum carmichaeli* roots from China. *J Nat Prod* 45: 128–133.
- Kulanthaivel P, Benn M, Majak W (1986) The C_{19} -diterpenoid alkaloids of *Delphinium bicolor*. *Phytochemistry* 25: 1511–1513.
- Meriçli AH, Meriçli F, Becker H, Ilarslan R, Ulubelen A (1996a) 3-Hydroxytalatisamine from *Aconitum nasutum*. *Phytochemistry* 42: 909–911.
- Meriçli AH, Meriçli F, Becker H, Ulubelen A (1996b) A new prodelphinine type alkaloid from *Aconitum nasutum*. *Tr J Chemistry* 20: 164–167.
- Meriçli AH, Meriçli F, Desai HK, Joshi BS, Teng Q, Bhattacharyya K, Melikoğlu G, Küçükislamoğlu M, Ulubelen A (2000a) Norditerpenoid and diterpenoid alkaloids from the roots of *Aconitum nasutum* Fisch ex Reichb. *Heterocycles* 53: 1987–1996.
- Meriçli AH, Meriçli F, Ulubelen A, Bahar M, Ilarslan R, Algül G, Desai HK, Teng Q, Pelletier SW (2000b) Diterpenoid alkaloids from the aerial parts of *Aconitum anthora* L. *Pharmazie* 55: 696–698.
- Pelletier SW, Joshi BS, Desai HK, Vlietinck AJ, Dommisse RA (1985) in: *In Advances in Medicinal Plant Research*, Wissenschaftliche Verlagsgesellschaft mbH, Stuttgart, p. 153.
- Ulubelen A, Meriçli AH, Meriçli F, Kolak U, Arfan M, Ahmad M, Ahmad H (2002) Norditerpenoid alkaloids from the roots of *Aconitum leave* Royle. *Pharmazie* 57: 427–429.
- Ulubelen A, Meriçli AH, Meriçli F, Yılmaz F (1996) Diterpenoid alkaloids from *Aconitum orientale*. *Phytochemistry* 41: 957–961.