# Cooperative National Park Resources Studies Unit

# ARIZONA

**TECHNICAL REPORT NO. 23** 

A PRELIMINARY INVESTIGATION OF THE ARTHROPOD FAUNA OF QUITOBAQUITO SPRINGS AREA, ORGAN PIPE CACTUS NATIONAL MONUMENT, ARIZONA

KENNETH J. KINGSLEY, RICHARD A. BAILOWITZ, and ROBERT L. SMITH

University of Arizona Tucson, Arizona 85721

Western Region National Park Service Department of the Interior San Francisco, Ca. 94102

#### COOPERATIVE NATIONAL PARK RESOURCES STUDIES UNIT University of Arizona/Tucson - National Park Service

The Cooperative National Park Resources Studies Unit/University of Arizona (CPSU/UA) was established August 16, 1973. The unit is funded by the National Park Service and reports to the Western Regional Office, San Francisco; it is located on the campus of the University of Arizona and reports also to the Office of the Vice-President for Research. Administrative assistance is provided by the Western Archeological and Conservation Center, the School of Renewable Natural Resources, and the Department of Ecology and Evolutionary Biology. The unit's professional personnel hold adjunct faculty and/or research associate appointments with the University. The Materials and Ecological Testing Laboratory is maintained at the Western Archeological and Conservation Center, 1415 N. 6th Ave., Tucson, Arizona 85705.

The CPSU/UA provides a multidisciplinary approach to studies in the natural and cultural sciences. Funded projects identified by park management are investigated by National Park Service and university researchers under the coordination of the Unit Leader. Unit members also cooperate with researchers involved in projects funded by non-National Park Service sources in order to obtain scientific information on Park Service lands.

**NOTICE:** This document contains information of a preliminary nature and was prepared primarily for internal use in the National Park Service. This information is <u>NOT</u> intended for use in open literature prior to publication by the investigators' names unless permission is obtained in writing from the investigators named and from the Unit Leader.

# COOPERATIVE NATIONAL PARK RESOURCES STUDIES UNIT UNIVERSITY OF ARIZONA 125 Biological Sciences (East) Bldg. 43 Tucson, Arizona 85721

R. Roy Johnson, Unit Leader National Park Senior Research Scientist

# **TECHNICAL REPORT NO. 23**

# A PRELIMINARY INVESTIGATION OF THE ARTHROPOD FAUNA OF QUITOBAQUITO SPRINGS AREA, ORGAN PIPE CACTUS NATIONAL MONUMENT, ARIZONA

# KENNETH J. KINGSLEY, RICHARD A. BAILOWITZ, and ROBERT L. SMITH

July 1987

NATIONAL PARK SERVICE/UNIVERSITY OF ARIZONA National Park Service Project Funds CONTRIBUTION NUMBER CPSU/UA 057/01

#### A PRELIMINARY INVESTIGATION OF THE ARTHROPOD FAUNA OF THE QUITOBAQUITO SPRINGS AREA ORGAN PIPE CACTUS NATIONAL MONUMENT, ARIZONA

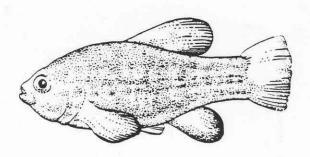
by

KENNETH J. KINGSLEY, Ph.D. 2643-1/2 N. Calle de Romy Tucson, Arizona 85712

RICHARD A. BAILOWITZ, M.S. 2774 W. Calle Morado Tucson, Arizona 85745

and

ROBERT L. SMITH, Ph.D. Department of Entomology University of Arizona Tucson, Arizona 85721



Quitobaquito Science Series, No. 7 -- July 1987 Technical Editors: Peter S. Bennett, Research Scientist and R. Roy Johnson, Sr. Research Scientist

# **TABLE OF CONTENTS**

| Introduction     | 1  |
|------------------|----|
| Methods          | 1  |
| Results          | 2  |
| Discussion       | 20 |
| Literature Cited | 22 |
| Acknowledgements | 23 |

# LIST OF TABLES

| Table 1. | Insects Collected at Quitobaquito Springs                    | 3 |
|----------|--|---|
| Table 2. | Biogeographic Affinities of the Insect Fauna of Quitobaquito | 0 |

#### ABSTRACT

Insects were collected during seven field trips to Quitobaquito Springs. Several different techniques for collecting were used, including blacklight trapping, aerial and sweep netting, and hand capture. The collection reflects the interests of the investigators and is not a thorough sampling of all taxa that might be found or all microhabitats that are available. At total of 559 species-level taxa in 134 families representing 12 orders were collected. Many of these are identified to species level.

The study area is probably the only United States breeding habitat of the butterfly <u>Ascia</u> <u>howarthi</u> (commonly named Giant White) which is dependent upon the plant <u>Atamisquea</u> <u>emarginata</u> (no common name) which grows at no other location in the U.S. Protection of this rare plant is called for if the butterfly is to remain part of the fauna. The insect fauna is probably typical of Sonoran desert habitats where permanent water is present. No significant threats to insect populations were apparent or are in need of management consideration at this time.

#### **INTRODUCTION**

Quitobaquito Springs in Organ Pipe Cactus National Monument has been intensively studied over a period of many years as the most biologically divers in the monument. Cole and Whiteside (1965) gave a brief description of the habitat, including some of the arthropod fauna. Johnson et al. (1983) discussed avian use of the area. Brown and Warren (1986) described the woody riparian vegetation at the springs. Bowers (1980) included references to the flora at Quitobaquito. Crieghton (1952) surveyed the ants of Organ Pipe Cactus National Monument, including Quitobaquito Springs. Increasing understanding and proper management of the resources calls for continued research into some of the lesser known components of the community.

Despite the facts that most of the animals in the world are insects and that the arthropod fauna plays a major part in the functioning of any ecosystem, basic inventories of arthropods are scarce to nonexistent for National Park Service areas. This preliminary survey of the arthropods of the Quitobaquito Springs area is part of an ongoing ecological study being conducted by the National Park Service.

Insects are part of the diet of the Quitobaquito pupfish (<u>Cyprinodon macularius eremus</u>). Insects and spiders are an important food source for many of the birds and bats that visit or reside at Quitobaquito. Native insects may be the sole pollinators of some of the unique native plants found in the area. The relative abundance and species composition of arthropods may be an index of pesticide drift from the agricultural area around Sonoyta, Mexico, which is a problem of increasing concern. This study is a first step toward gathering baseline data on these important animals, and should be of assistance in the development of a management plan for Quitobaquito.

## **METHODS**

This study was limited to the collection and identification of members of the class Insecta. Seven field trips were taken to the area between April 23, 1983 and December 31, 1984. On each trip, insects observed were collected by standard methods such as aerial and sweep netting and hand capture. Efforts were made to sample each of the habitats present in the area, focusing primarily on the examination of individual plants that were in bloom. Blacklight trapping was done at night on each trip. Adjacent areas, including the hills north of the springs and Aguajita Was were examined also. Little effort was made to sample for certain groups that were difficult to find or require specialized techniques, such as soil arthropods, aquatic benthos arthropods in the pond, wood borers, leaf miners, etc. The resulting collection reflects the interests of the investigators and the limitations of time and techniques, and should not be considered a truly exhaustive or complete inventory of the fauna present in the area. Identifications were made by means of use of keys in the literature, comparison with labeled specimens in the University of Arizona insect collection, and consultation with local taxonomists, Dr. Floyd Werner and Mr. Carl Olson of the University of Arizona Department of Entomology.

Specimens collected were deposited with the permanent collection at the University of Arizona Department of Entomology and with Organ Pipe Cactus National Monument.

#### RESULTS

Table 1 lists the insect fauna collected. The order and nomenclature of the higher groups (orders, families, subfamilies) generally follows that of Boror et al. (1981). Within the list, orders are capital letters in BOLDFACE; families are in capitals; subfamilies are indented once and in capitals and lowercase; and genus and species are indented twice and <u>underlined</u>. Example:

#### ORDER

#### FAMILY

Subfamily

#### Genus species Author/(Author)

In general, order names end with "-optera," family names end with "-idae," subfamily names end with "-inae," tribe names end with "-ini." Where possible, specimens were identified to the lowest taxonomic level (species). Species names are identified, all other levels of classification are not. The species names are followed by the name (or names) of the author or first describer of the species in the scientific literature. When an author's name appears in parenthesis, this means that the original description had the species placed in another genus. Advances are continuously being made in insect taxonomy and identification, so names change with some frequency,.

Many specimens were not identified to the genus and species level. This means only that we were unable to identify them, for any one of a number of reasons. Insects identification is a complex science, even an art form; and, even a specialist in a particular group may not be able to readily identify all specimens in that group. Lack of a name in this list does not mean that the specimen represents an undescribed species, although such may be possible. Where we have identified a specimen to the lowest taxonomic level we could, but not to species, the lowest level name is followed by the abbreviation lisp." In some cases, it was evident that we had specimens representing more than one species of a genus or higher taxonomic level, but we could not positively identify them to lower taxonomic levels. Then the lowest level to which we could identify the specimens is given, followed by "sp.," and a number (1, 2, 3, etc.). In the submitted collection, specimens were grouped by taxonomic groupings, and labeled with the lowest level we could ascertain with surety. The numbers used as identifiers in this list are not part of the specimen labels, but only serve to indicate how many apparently different species were collected.

# TABLE 1. Insects Collected at Quitobaquito Springs, Organ Pipe Cactus National Monument, Arizona.

# **ODONATA**

#### GOMPHIDAE

<u>Progomphus borealis</u> MacLachlan <u>Erpetogomphus compositus</u> Hagen

#### AESHNIDAE

<u>Anax junius (Drury)</u>

#### LIBELLULIDAE

<u>Libellulla saturata</u> Uhler <u>Erythemis simplicicollis</u> Say <u>Sympetrum madidum</u> (Hagen) <u>Brachymesia furcata</u> Hagen <u>Perithemis domitia</u> Drury <u>Tramea sp.\*</u> <u>Pachydiplax</u> longipennis (Burmeister) <u>Orthemis ferruginea</u> (Fabricius) <u>Pontala hymenaea</u> (Say)

# COENAGRIONIDAE

<u>Chromagrion</u> sp. <u>Argia</u> sp. \* <u>Telebasis</u> <u>salva</u> (Hagen) <u>Neoneura</u> <u>sp</u>. \* <u>Tellealagma</u> sp. \* Other species: 1

# **NEUROPTERA**

# CHRYSOPIDAE

<u>Chrysopa</u> sp. 1 <u>Chrysopa</u> sp. 2 <u>Chrysopa</u> sp. 3 <u>Chrysoya</u> sp. 4 Eremochrysa punctinervis McLachlan

# MYRMELEONIDAE

<u>Hesperoleon</u> sp. 1 <u>Hesperoleon</u> sp. 2 <u>Hesperoleon</u> sp. 3 <u>Eremoleon nigribasis</u> Banks <u>Vella hesperus</u> Banks

# HEMEROBIIDAE Species: 1

\* denotes species throughout table were also collected by Cole and Whiteside, 1965.

\*\*denotes species collected by W.S. Creighton in 1952 (note found only on page 18).

#### DERMAPTERA

LABIDURIDAE

Labidura riparia (Pallas)

# CARCINOPHORIDAE

Euborellia sp.

#### TRICHOPTERA

#### LEPTOCERIDAE

<u>Oecetis</u> sp.

#### THYSANOPTERA

#### AELOTHRIPIDAE

Species: 1

#### ORTHOPTERA

# ACRIDIDAE

Acridinae <u>Ligurotettix coquilletti</u> McNeill <u>Opeia obscura</u> (Thomas) <u>Horesidotes cinereus</u> Scudder <u>Cibolacris parviceps</u> (Walker) Oedipodinae <u>Trimeritropis pallidipennis</u> (Burmeister) <u>Lactista aztecus</u> (Saussure) <u>Anconia integra</u> (Scudder) <u>Heliastus benjamini</u> Caudell Cyrtacanthacridinae <u>Schistocerca vaga</u> (Scudder) <u>Melanoplus sp.</u> <u>Aeoloplides tenuipennis</u> (Scudder) <u>Hesperotettix viridis</u> (Thomas) <u>Leptysma hebardi</u> Rehn and Eades

# TETTIGONIIDAE

<u>Neoconocephalus triops</u> (L.) <u>Scudderia mexicana</u> (Saussure) <u>Ateloplus schwarzi</u> Caudell <u>Eremopedes bilineatus</u> (Thomas) <u>Insara elegans</u> (Scudder) <u>Insara covilleae</u> Rehn and Hebard

# GRYLLIDAE

<u>Gryllus</u> sp. <u>Nemobius</u> sp.

## MANTIDAE

<u>Litaneutria minor</u> Scudder <u>Stagmomantis californica</u> Rehn and Hebard

BLATTELLIDAE Blattella vaga Hebard **HOMOPTERA** MEMBRACIDAE Spissistilus festinus (Say) CICADELLIDAE Gyponinae Species: 1 Iassinae Stragania sp. Hecalinae Species: 1 Agalliinae <u>Aceratagallia sp.</u> Typhlocybinae <u>Émpoasca</u> sp. 1 <u>Empoasca sp</u>. 2 Deltocephalinae Scaphytopius nigricollis (Ball) Scaphytopius sp. Opsius stactogalus Fieber Other species: 5 Cicadellinae Homalodisca sp. Carneocephala sp. Other species: 1 DELPHACIDAE Species: 1 CIXIIDAE Species: 2 **FULGORIDAE** Cyrpoptus sp. FLATIDAE Ormenis sp. PSYLLIDAE Species: 1 CERCOPIDAE Clastoptera sp.

# ISSIDAE

Species: 1

# HEMIPTERA

# PENTATOMIDAE

<u>Chlorochroa sayi</u> Stal <u>Chlorochroa ligata</u> (Say) <u>Thyanta pallidovirens</u> Stal <u>Mecidea minor</u> Sailer

# CYDNIDAE

<u>Melanaethus</u> sp. Tominotus conformis communis (Uhler)

#### COREIDAE

Leptoalossus brevirostris Barber

#### RHOPALIDAE

<u>Liorhyssus hyalinus</u> (F.) <u>Harmostes reflexulus</u> (Say) <u>Aufeius imnressicollis</u> Stal

# LYGAEIDAE

<u>Lygaeus lateralis</u> Dallas <u>Nysius raphanus</u> Howard

# LARGIDAE

Largus cinctus Herrich-Schaeffer

#### BERYTIDAE

Pronotacantha annulata Uhler

# TINGIDAE

Species: 1

#### REDUVIIDAE

<u>Zelus renardii</u> Kolenati <u>Zelus socius</u> Uhler <u>Rasahus biguttatus</u> (Say) <u>tenolemoides arizonensis</u> (Banks) <u>Triatoma rubida</u> (Uhler)

#### MIRIDAE

<u>Phytocoris</u> sp. <u>Oncerometopus nigriclavus</u> Reuter Other species: 4

# VELIIDAE

Microvelia sp.

#### NOTONECTIDAE

<u>Notonecta indica</u> L. <u>Buenoa arizonis</u> Bare

# NAUCORIDAE

Ambrysus californicus Montandon

# LEPIDOPTERA

#### HESPERIIDAE

Lerodea eufala (Edwards) Lerodea arabus (Edwards) Nyctelius nyctelius (Latreille) Copaeodes aurantiaca (Hewitson) Hylephila Rhyleus (Drury) Pholisora libya (Scudder) Pyrgus albescens Plotz Prygus philetas Edwards Prygus scriptura (Boisduval) Heliopetes domicella (Erichson) Erynnis funeralis (Scudder and Burgess) Systasea zampa Edwards

# PAPILIONIDAE

Battus philenor (L.)

# PIERIDAE

<u>Pieris protodice (L.)</u> <u>Colias eurytheme</u> Boisduval <u>Colias cesonia Stoll</u> <u>Phoebis sennae L.</u> <u>Kricogonia lyside</u> Godart <u>Ascia howarthi (Dixey)</u> <u>Eurema nicippe</u> Cramer <u>Euema mexicana</u> Boisduval <u>Anthocaris pima</u> Edwards <u>Nathalis iole</u> Boisduval

# LYCAENIDAE

<u>Calephelis nemesis</u> Edwards <u>Apodemia mormo</u> Felder and Felder <u>Apodemia palmerii</u> Edwards <u>Minstrymon leda</u> Edwards <u>Chlorostrymon simaethis</u> Drury <u>Atlides halesus</u> Cramer <u>Strymon melinus</u> Hubner <u>Strymon columella</u> Fabricius <u>Leptotes marina</u> Reakirk <u>Hemiargus isola</u> Reakirk <u>Hemiargus ceraunus</u> Fabricius <u>Brephidium exilis</u> Boisduval

# LIBYTHEIDAE

Libytheana bachmanii (Kirtland)

#### HELICONIIDAE

<u>Agraulis</u> vanillae (L.)

#### NYMPHALIDAE

Asterocampa leilia Edwards <u>Marpesia petreus</u> Cramer <u>Nymphalis antiona</u> L. <u>Precis coenia</u> Hubner <u>Precis nigrosuffusa</u> (Barnes and McDunnough) <u>Vanessa cardui</u> L. <u>Vanessa virginiensis</u> Drury <u>Vanessa annabella</u> (Field) <u>Vanessa atalanta</u> L. <u>Anthanassa texana</u> (Edwards) <u>Chlosyne californica</u> (Wright) <u>Chlosyne lacinia</u> (Geyer) <u>Euptoieta claudia</u> (Cramer)

# DANAIDAE

Danaus gilippus (Cramer) Danaus plexippus (L.)

#### TINEIDAE

Acrolophus sp. Other species: 1

**GRACILLARIIDAE Species: 1** 

#### GELECHIIDAE

Specimens: 8

#### PLUTELLIDAE

<u>Plutella</u> sp.

#### YPONOMEUTI DAE

Atteva punctella (Cramer)

#### SESIIDAE

<u>Hypopta palmata</u> Barnes and McDunn <u>Comadia intrusa</u> Barnes and McDunn

#### TORTRICIDAE

Species: 1

# PYRALIDAE

Evergestinae <u>Evergestis notentis</u> Phycitinae <u>Cactobrosis fernaldialis</u> (Hulst) <u>Alberada parabates</u> (Dyar) Other specimens: 8 Crambinae Diatraea grandiosella (Dyar) PYRALIDAE (Contd.)

Pyraustinae

<u>Hymenia perspectalis</u> (Hubner) <u>Loxostege albiceralis</u> Grote <u>Loxostege</u> sp. <u>Achyra</u> sp. <u>Hahncappsia</u> sp. <u>Nomophila</u> sp. Other species: 3 Nyphulinae <u>Petrophila jaliscalis</u>

## GEOMETRIDAE

Synchlora rubrifrontaria Packard Chlorochlamys ghyllinaria Zeller Metasiopsis peralbata Packard Cosymbia serrulata Packard Eubarnesia ritaria (Grossbeck) Glaucina sp. Archihoe sp. Tornos sp. Anacamptodes obliguaria Grote Anacamptodes dataria Grote Semiothisa irrorata Packard Semiothisa hypaethrata Grote Semiothisa s-signata Packard Semiothisa sp. 1 Semiothisa sp. 2 Apicia sp. Other species: 2

# SPHINGIDAE

<u>Hyles lineata</u> (F.) <u>Manduca sexta</u> (L.) Manduca guinquemaculata (Haworth)

# ARCTIIDAE

<u>Cisthene angelus</u> Dyar <u>Ctenucha venosa</u> Walker

# NOCTUIDAE

<u>Pseudaletia unipuncta (</u>Haworth) <u>Bulia deducta</u> (Morrison) <u>Heteranassa mimes</u> Harvey <u>Schinia intrabilis</u> Smith <u>Schinia balba</u> Grote <u>Forsebia perlaeta</u> Edwards <u>Meliothis acontioides</u> Guenee <u>Matigramma rubrosuffusa</u> Grote <u>Xylomyges curialis</u> Grote NOCTUIDAE (Contd.)

Heliothis zea (Boddie) Autographa sp. Erebus odora (L.) Catocacala junctura Grote Hemeroplanis subflavidalis Grote Oncocnemis occata Grote Spragueia magnifica Grote Spodoptera exigua (Hubner) Grotella binda Barnes Azenia implora Grote Timora toralis Grote Acontia sp. 1 Acontia sp. 2 Conochares acuta Smith Conochares catalina no author Conochares arizonae Edwards Conochares sp. Pseudohadena vulnerea Grote Lepipolys perscriptura Guenee Cyathissa pallida Smith Aseptis catalina Smith Lacinipolia sp. 1 Lacinipolia sn. 2 Chorizagrotis auxiliaris Grote Chorizagrotis sp. Agrotis malefida (Guenee) Other specimens: 21

# **COLEOPTERA**

CICINDELIDAE

<u>Cicindela lemniscata</u> LeConte <u>Cincindela</u> sp.

# CARABIDAE

<u>Omophron</u> sp. <u>Lebia viridis</u> Say <u>Lebia</u> sp. 1 Lebia sp. 2 <u>Bembidion</u> sp. 1 <u>Bembidion</u> sp. 2 <u>Agonoderus</u> sp. <u>Calosoma peregrinator</u> Guerin <u>Calosoma</u> sp. <u>Poecilus subchordatus</u> LeConte <u>Bradycellus</u> sp. <u>Brachinus</u> sp. <u>Scizogenuis</u> op. CARABIDAE (Contd.) <u>Tachys</u> sp. <u>Pterostichini</u> sp.

### DYTISCIDAE

<u>Rhantus gutticollis</u> (Say) <u>Eretes sticticus</u> (L.) <u>Laccophilus sonorenis Zimmerman</u> <u>Laccophilus fasciatus</u> Aube \* <u>Laccophilus pictus</u> Castelman <u>Deronectes striatellus</u> (LeConte) <u>Deronectes roffi nebulosus</u> \* <u>Dytiscus habilis</u> Say \* <u>Copelatus chevrolati</u> Guignot <u>Cybister</u> sp. \*

# HYDROPHILIDAE

<u>Hydrophilus triangularis</u> say <u>Helophorus</u> sp. <u>Tropisternus lateralis (F.)</u> \* <u>Tropisternus ellipticus (LeConte)</u> \* <u>Berosus ruqulosus Horne</u> <u>Chaetarthria pallida (LeConte)</u> Enochrus pygmaeus (LeConte)

# STAPHYLINIDAE

Tachyporinae

Species: 4

Paederinae

Species: 3

Staphylininae

Species: 2

# SCARABAEIDAE

Melolonthinae <u>Diplotaxis</u> sp. Other species: 4 Aphodiinae <u>Psammobius guinqueplicatus</u> Horn Other species: 1 Geotrupinae <u>Species: 1</u> <u>Dynastinae</u> <u>Ligyrus</u> sp. Cetoniinae <u>Cremastocheilus</u> sp.

#### HETEROCERIDAE

Dampfius sp. Other species: 1

#### DRYOPIDAE

Helichus sp.

## BUPRESTIDAE

Acmaeodera flavomarginata Gory Acmaeodera gibbula LeConte

# ELATERIDAE

<u>Diplostethus opacicollis</u> Schaeffer <u>Discrepidius corvinus</u> Candeze <u>Eniconyx</u> sp. <u>Aeolus mellilus</u> (Say) <u>Conoderus</u> sp. Other species: 2

# ANOBIIDAE

Tricorynus sp.

#### BOSTRICHIDAE

Bostrichinae Species: 1

#### CLERIDAE

Cymatodera oblita Horn Cymatodera sp. 1 Cymatodera sp. 2

## DASYTIDAE

Species: 1

#### MELOIDAE

<u>Lytta magister</u> Horn <u>Pyrota palpalis</u> Champion <u>Epicauta wheeleri</u> Horn <u>Epicauta lauta</u> (Horn) Epicauta tenella (LeConte)

#### MORDELLIDAE

Mordellistena sp. Other species: 1

# TENEBRIONIDAE

Tenebrioninae <u>Eleodes armata</u> LeConte <u>Eleodes caudata</u> Le Conte <u>Ulus crassus</u> (LeConte) <u>Ammodonus granosus</u> Fall TENEBRIONIDAE (Contd.) Other species: 1 Tentyriinae Cryptoglossa verrucosa LeConte Other species: 2 Asidinae

Asidina confluens Asidinae Asidina confluens (LeConte) Other species:

## LAGRIIDAE

Statira sp.

# ALLECULIDAE

Hymenorus sp. Other species: 4

#### **OEDEMERIDAE**

Xanthochroina sp. Oxacis sp.

# ANTHICIDAE

Notoxus sp. Anthicus sp. 1 Anthicus sp. 2

# NITIDULIDAE

Carpophilus sp.

# COCCINELLIDAE

Hippodamia convergens Guerin chilocorus cacti L. Olla obdominalis (Say)

# CERAMBYCIDAE

Taranomis bivittata (Dupont) Aneflus pratensis Leconte Crossidius suturalis Leconte Osmidius guttatus Leconte Peropleum sp. Cerambycinae Species: 1

## CHRYSOMELIDAE

- Cryptocephalinae
  - Pachybrachys sp. 1 Pachybrachys sp. 2 Pachybrachys sp. 3
  - Galerucinae

Species: 1

Alticinae

Species: 2

# BRUCHIDAE

<u>Algarobius prosopis</u> (Leconte) <u>Mimosestes amicus (</u>Horn)

# CURCULIONIDAE

Anthonominae Species: 1 Ceutorhynchinae Species: 1

# MALACHIIDAE

Collops sp. 1 Collops sp. 2

# DIPTERA

# TIPULIDAE

<u>Tipula</u> sp. 1 Other species: 2

# CULICIDAE

Anopheles franciscanus McCracken

CHIRONOMIDAE Tanypodinae <u>Tanypus</u> sp. Diamesinae Species: 1 Chironominae Species: 2 Orthocladiinae Species: 2 Other species: 1

# CERATOPOGONIDAE

Ceratopogoninae Species: 1 Dasyheleinae Species: 1

#### BIBIONIDAE

Bibiodes sp.

#### CECIDOMYIIDAE

Neolasioptera sp. 1 Asphondyliini Species: 2 Cecidomyiini Species: 1 Lestremiinae Species: 2 Other species: 1

# TABANIDAE

<u>Chrysops</u> sp. <u>Apatolestes aitkeni</u> Philip

#### ASILIDAE

Saropogon sp. <u>Efferia</u> sp. 1 <u>Efferia</u> sp. 2 <u>Efferia</u> sp. 3 Mallophorina sp.

#### BOMBYLIIDAE

<u>Thyridanthrax</u> sp. <u>Geron</u> sp. 1 <u>Geron</u> sp. 2 <u>Paravilla</u> sp. <u>Heterostylum robustum</u> (Osten Sacken) <u>Lordotus</u> sp. 1 <u>Lordotus</u> sp. 2 <u>Poecilanthrax</u> sp. 1 <u>Poecilanthrax</u> sp. 2 <u>Bombylius major</u> L. <u>Anthrax</u> sp. <u>Oligodranes</u> sp. <u>Lepidanthrax</u> sp. 1 <u>Lenidanthrax</u> sp. 2

# DOLICHOPODIDAE

<u>Tachytrechus angustipennis</u> Loew <u>Chrysotus</u> sp. 1 <u>Chrysotus</u> sp. 2 <u>Medetera</u> sp. <u>Hydrophorus</u> sp. <u>Condylostylus inornatus</u> (Aldrich) Others species: 3

#### STRATIOMYIDAE <u>Odontomyia</u> sp. <u>Hedriodiscus</u> <u>currani</u> James

#### SYRPHIDAE

<u>Scaeva pyrastri</u> (L.) <u>Eupeodes volucris</u> Osten Sacken <u>Volucella apicifera</u> Townsend <u>Volucella sp. 1</u> <u>Eristalis aeneus</u> (Scopoli) <u>Eristalis latifrons</u> Loew <u>Mesograpta sp.</u> <u>Baccha clavata</u> (F.)

#### NERIIDAE

Odontoloxozus longicornis (Coquillett)

#### OTITIDAE

Diacrita costalis Gerstaecker

# TEPHRITIDAE

Euaresta bellula Snow Other species: 1

# LAUXANIIDAE

Camptoprosopella sp.

#### SPHAEROCERIDAE

<u>Leptocera</u> sp. 1 <u>Leptocera</u> sp. 2 Scatophora sp.

#### HELIOMYZIDAE

Pseudoleria sp.

# DROSOPHILIDAE

Drosophila sp. Other species: 1

#### EPHYDRIDAE

Species: 1

#### CHLOROPIDAE

<u>Pseudogaurax</u> sp. Lasiopleura sp.

# ANTHOMYIIDAE

Scatophora sp.

### MUSCIDAE

Species: 3

#### CALLIPHORIDAE

<u>Cochliomyia</u> sp. <u>Lucilia</u> sp. <u>Phaenicia</u> sp.

# SARCOPHAGIDAE

Sarcophaga sp. 1 Sarcophaa sp. 2

### TACHINIDAE

Euphaseopteryx ochracea Bigot Others species: 4

#### CUTEREBRIDAE

Species: 1

# HYMENOPTERA

BRACONIDAE Agathidinae Zelomorpha sp. Other species: 1 Braconinae Species: 1

### ICHNEUMONIDAE

<u>Compsocryptus calipterus</u> (Say) Tryphoninae <u>Netelia</u> sp. Ophioninae <u>Enicospilus</u> sp. Tersolochini Species: 1 Other species: 3

# FORMICIDAE

<u>Pogonomyrmex pima</u> Wheeler \*\* <u>Novomessor cockerelli</u> (Andre)\*\* <u>Veromessor pergandei</u> (Mayr)\*\* <u>Pheidole vasliti arizonica</u> Santschi\*\* <u>Iridomyrmex pruinosum analis</u> (Andre)\*\* <u>Solenopsis xyloni</u> (McCook)

#### CHALICIDOIDEA

Species: 1

# EULOPHIDAE

Species: 1

# CYNIPIDAE

Species: 1

#### **SCELIONIDAE**

Species: 2

# TIPHIIDAE

Brachycistis triangularis Fox Brachycistis chinensis Bradley Brachycistis arenivaga Bradley Brachycistis sp. Paratiphia sp. Myzinum sp.

# MUTILLIDAE

Odontophotopsis sp. Sphaerophthalma sp. Acanthophotopsis sp. Other species: 6

# SCOLIIDAE

<u>Campsoscolia flammicoma</u> (Bradley) <u>Scolia otomita</u> Saussure <u>Scolia ardens</u> Smith <u>Campsomeris tolteca (Saussure)</u>

# POMPILIDAE

Hemipepsis ustulata Dahlbom <u>Pepsis mexicana</u> Lucas <u>Pepsis chrysothemis</u> Lucas <u>Pepsis mildei</u> Stal <u>Pepsis pallidolimbata</u> Lucas <u>Episyron posterus</u> (Fox) <u>Anoplius</u> sp. <u>Pompilus</u> expulsus Schulz Other species: 1

# EUMENIDAE

<u>Stenodynerus</u> sp. 1 <u>Stenodynerus</u> sp. 2 Other species: 2

#### SPHECIDAE

<u>Ammophila breviceps</u> Smith <u>Ammophila</u> sp. <u>Ammophila aberti</u> Haldeman Ammophilini <u>Podalonia mexicana</u> (Saussure) <u>Aphilanthops hispida</u> Fox <u>Trypoxylon</u> sp. <u>Glenostictia</u> sp. <u>Bembix sayi</u> Cresson <u>Bembix sp.</u> <u>Steniola duplicata</u> Provancher <u>Liris sp..</u> <u>Prionyx sp.</u> <u>Sphex sp.</u> <u>Eucerceris sp.</u> Other species: 1

# HALICTIDAE

<u>Agapostemon melliventris</u> Cresson <u>Nomia nevadensis</u> (Cockerell) <u>Nomia tetrazonata</u> Cockerell) <u>Augochlorella pomoniella</u> Cockerell <u>Lasioglossum</u> sp.

# ANDRENIDAE

Andrena sp. Other species: 1

# MELITTIDAE

Ashmeadiella sp.

# ANTHOPHORIDAE

<u>Diadasia</u> sp. <u>melissodes</u> sp. <u>Hemisia</u> sp. <u>Anthophora</u> sp. Other species: 1

# APIDAE

Apis mellifera L.

# MEGACHILIDAE

Lithurginae Species: 1 Megachilinae <u>Chalicodoma</u> sp. Other species: 3

#### DISCUSSION

Biogeographic affinities of the insect fauna of Quitobaquito are difficult to evaluate meaningfully. Usually known species ranges are limited to those areas that have received adequate surveillance. For many species, ranges vary form year to year, depending upon weather conditions. However, we have attempted to group the distributions of 129 species found at Quitobaquito Springs into areas in which they might be expected to occur with greatest frequency. These and the number of species and percent of the total 129 species included in this analysis are:

| AREA                    | NUMBER OF<br>SPECIES | PERCENT<br>OF TOTAL |
|-------------------------|----------------------|---------------------|
| Cosmopolitan            | 4                    | 3.1                 |
| Introduced              | 2                    | 1.6                 |
| North & South America   | 18                   | 13.9                |
| North & Central America | 11                   | 8.5                 |
| North America           | 10                   | 7.8                 |
| United States           | ar 10 Sadola abi i   | 3.9                 |
| Western U.S. & Mexico   | 26                   | 20.2                |
| Western U.S.            | 18                   | 13.9                |
| Southern U.S.           | 6                    | 4.7                 |
| Southwestern U.S.       | 16                   | 12.3                |
| Arizona-California      | 1.                   | 0.8                 |
| Arizona-Sonora          | 11                   | 8.5                 |
| Arizona                 | hlan 1 Third         | 0.8                 |
| TOTALS                  | 129                  | 100.0               |

TABLE 2. Biogeographic Affinities of the Insect Fauna of Quitobaquito.

The data presented in Table 2 indicates that the insect fauna of Quitobaquito is representative to that found in any similar area of the Southwestern U.S. or Northern Mexico, than includes permanent water. There is a predominance of widespread species that are adaptable to a variety of situations.

The only really noteworthy distribution record is that of the butterfly, <u>Ascia howarthi</u> (Dixey), which is primarily a Mexican species, with a few U.S. records. This species is associated with, and completely dependent upon the plant <u>Atamisguea emarginata</u>, which is found in the U.S. only near Quitobaquito Springs. Bailowitz (1985) present a detailed discussion

of the distribution and biology of this species. This species was formerly considered to be conspecific with <u>A</u>. josephina (Godart), but has been evaluated to species level based on behavioral, ecological, and morphological evidence (Bailowitz 1987).

Two species are introduced and have become naturalized throughout the United States. The Honeybees, <u>Apis mellifera</u>, have become naturalized from domestic colonies. Bee specimens collected at Quitobaquito may have been members of domestic colonies kept at the site by the National Park Service for observation of pesticide drift, or may have been from wild colonies. The other introduced species is the striped earwig, <u>Labidura riparia (Pallas)</u>. It was first found in Arizona t Yuma in 1952, and has become widespread in the southern part of the state. It is a predator on other insects, especially maggots. Occasionally, it may invade houses because it is attracted to lights, and may become a nuisance (Ebeling 1978). At Quitobaquito, it probably lives in soil, under leaves and other debris, and is uncommon and harmless creature.

Management implications of this research are not clear-cut. The ways in which present management techniques impact the arthropod fauna are not evident, save that the present condition of the habitat diversity provides a variety of resources for insects, including both stream and pond habitats for aquatic insects. Most, if not all, of the insect species present would likely remain present as long as this habitat diversity is maintained. Because of the very small extent of the distribution of <u>Atamiscguea emarginata</u>, the population of <u>Ascia howarthi</u> is at some risk of extirpation in the U.S., should some misfortune befall the host plant. It is possible that insecticide use in the neighboring agricultural land in Mexico may adversely impact some components of the Quitobaquito fauna, particularly in the extent of extreme drift of aerially applied insecticides (NPS data available). However, since most of the fauna is widespread and highly mobile, eventual reestablishment following local catastrophe is likely.

It cannot be overemphasized that this study was based on a very small number of field trips, and many taxa may have been overlooked. Because insect populations in an area may be ephemeral, or cryptic or migratory, a much more extensive survey is needed to develop a clear understanding of the insect fauna. This study may, however, provide a good basis on which to begin more extensive work.

#### LITERATURE CITED

- Bailowitz, R.A. 1987. Systematics of <u>Ascia</u> (<u>Ganyra</u>) (Pieridae) populations in the Sonoran Desert. J. Res. Lepid. (In press).
  - Bailowitz, R.A. 1985. Systematics and biology of <u>Ascia (Ganyra)</u> populations in the Sonoran Desert. M.S. thesis. University of Arizona.
  - Boror, D.J., D.M. De Long, and C.A. Tripplehorn. 1981. An Introduction to the Study of Insects. Philadelphia. Saunders. 827 pp.
- Bowers, J.E. 1980. Flora of Organ Pipe Cactus National Monument. J. Ariz.-Nev. Acad. Sci. 15:1-i1 and 33-47.
- Brown, B.T. and P.L. Warren. 1986. A descriptive analysis of woody riparian vegetation at Quitobaquito springs oasis, Organ Pipe Cactus National Monument, Arizona. CPSU/UA Tech. Rpt. 19. University of Arizona, Tucson. 16 pp.
- Cole, G.A. and M.C. Whiteside. 1965. An ecological reconnaissance of Quitobaquito Spring, Arizona. J. Arizona Acad. Sci. 3:159-163.
- Creighton, W.S. 1952. Notes on the ants of Organ Pipe Cactus National Monument, Arizona. Unpub. notes in the files of Organ Pipe Cactus N.M. 6 pp.
- Ebeling, W. 1978. Urban Entomology. Div. of Agric. Sciences. University of Calif. 695 pp.
- Johnson, R.R., B.T. Brown, and S. Goldwasser. 1983. Avian use of Quitobaquito Springs oasis, organ Pipe Cactus National Monument, Arizona. CPSU/UA Tech. Rpt. 13. University of Arizona, Tucson. 16 pp.

# ACKNOWLEDGEMENTS

Thanks are due several people for assisting with the technical report. CPSU/UA staff, Peter Bennett, R. Roy Johnson, Michael Kunzmann, and Susan Shaw, and Steven Carothers critically reviewed the manuscript. Susan Shaw typed it into final form for publication. Partial funding for the study was provided through NPS Project Funds.