
THERMOPHILIC MICROORGANISM SURVEY

YELLOWSTONE NATIONAL PARK

Yellowstone Center for Resources
P.O. Box 168
Yellowstone National Park, Wyoming 82190
August, 1997

YCR-NR-97-6

Acknowledgments: Prepared with editorial assistance from Jessica Boukevekas and Mary Ann Franke

Cover photo: Photomicrograph of the pink filament community, Octopus Springs, Yellowstone National Park. Courtesy of Anna-Louise Reysenbach, Indiana University.

YELLOWSTONE NATIONAL PARK

THERMOPHILIC MICROORGANISM SURVEY

This survey contains a summary of each thermophilic species known to be found in Yellowstone National Park, including a description of the microorganism, its habitat, where it can be found, and any known biotechnological application.

Species Included

Achanthes exigua
Acidothermus cellulolyticus
Alicyclobacillus acidocaldarius
Anaerobranca horikoshii
Archaeoglobus spp.
Bacillus sp.
Bacillus coagulans
Calothrix sp.
Chloroflexus aurantiacus
Chromatium tepidum
Clostridium thermoautotrophicum
EM3, EM17, and EM19 (pink filament community)
Heliobacterium modesticaldum
Mastigocladus laminosus
Methanobacterium thermoautotrophicum
pJp27
Rhodopila globiformis
Synechococcus sp.
Sulfolobus acidocaldarius
Thermoanaerobacter brockii
Thermoanaerobacter ethanolicus
Thermoanaerobacterium thermosulfurigenes
Thermoanaerobacterium xylanolyticum
Thermobacteroides acetoethylicus
Thermodesulfobacterium commune
Thermofilum spp.
Thermoleophilum album
Thermoleophilum minutum
Thermomicrobium roseum
Thermoplasma acidophilium
Thermoplasma volcanium
Thermosphaera aggregans
Thermothrix thiopara
Thermus aquaticus
Thiobacillus thiooxidans

The above list does not represent a complete inventory of the thermophiles that may be found in Yellowstone National Park. It includes only those species brought to our attention whose description in a scientific publication documents their presence in the Park. If you know of a Yellowstone thermophile that is not included in this publication, we encourage you to contribute to our database. See the next page for a sample reporting form. A bibliography of publications related to the Yellowstone geothermal ecosystem appears at the the end of this document.

**YELLOWSTONE NATIONAL PARK
THERMOPHILIC MICROORGANISMS**

Reporting Form

Scientific Name:

Description:

Habitat Type:

YNP Location:

USGS Quadrangle:

Directions:

Date First Observed:

Other Comments:

Citation: [Please include a reprint of the publication describing the organism.]

Photographs:

Specimens:

Person to Contact:

If you know of a thermophilic organism not included in this document, please contact the Yellowstone Center for Resources, P.O. Box 168, Yellowstone National Park, WY 82190, or call Bob Lindstrom at (307) 344-2234, or e-mail Bob_Lindstrom@NPS.GOV. Thank you for your cooperation.

Collection of any material from a U.S. National Park requires a research/collection permit. Information about permits is available at the preceding address and phone number.

Scientific Name:	<i>Achnanthes exigua</i>
Description:	This unialgal diatom was found mixed with flocculent sediments on the surface of a blue-green algal mat which was covered by a shallow layer of water. It reproduces at 12-hour intervals. The maximum metabolism, photosynthetic rate and respiration in this organism occur at thermal temperatures. It is a shade-adapted, phototrophic umbrophile which can also grow in highly intense lighting.
Habitat Type:	Calcium carbonate and neutral chloride Temperature range for growth: 10-44°C Optimum temperature for respiration: 40°C Optimum temperature for photosynthesis: 42°C pH: 8.5
YNP Location:	Alkaline hot spring near Old Faithful; directions not provided.
Abundance:	The population is marginal since it can only grow in low-velocity currents and shallow depressions on bacterial mats.
First Observed:	11/1/72
Other Locations:	Specimens of this diatom have also been collected from Alhambra North Hot Spring in Jefferson County, Montana and from Ohanapecosh Hot Spring No. 6 in Mount Rainier National Park.
Biotech Use:	No known application
Citation:	Fairchild, Eugene and Richard P. Sheridan. 1974. A physiological investigation of the hot spring diatom, <i>Achnanthes exigua</i> Grün. <i>Journal of Physiology</i> . 10:1-4.
Photographs:	None
Specimens:	None preserved

- Scientific Name: *Acidothermus cellulolyticus*
- Description: Acidophilic, cellulolytic, phototrophic, thermophilic bacteria. Obligate aerobe that forms gram-variable, non-motile, non-pigmented, non-sporing rods with long slender filaments. It shares several important features with *Thermus* strains, i.e., aerobic, heterotrophic, and thermophilic mode of growth; morphological features; sensitivity to lysozyme; and presence of catalase. It differs in the carbon sources used for growth, pH and temperature profiles for growth, sensitivity to anti-biotics, resistance to penicillin G, the composition of amino acids in the cell walls, and the structure of the cell walls. It produces the enzyme cellulase, which breaks down cellulose by hydrolysis, an important and unusual characteristic, especially for an acidophilic microorganism.
- Habitat Type: Acid-sulfate
Temperature range for growth: 37-65°C
Optimum temperature: 55°C
pH range: 3.5-7.0
Optimum pH: 5.0
- YNP Location: Upper Norris Geyser Basin; exact locations not provided.
- Abundance: The population appears to be abundant since 12 strains were isolated from various hot springs in the Norris Geyser Basin.
- First Observed: 7/1/86
- Biotech Use: This study was supported by the Office of Alcohol Fuels, U.S. Department of Energy, as part of a research program on the production of fuel alcohol from cellulosic biomass materials. It was hypothesized that an acidophilic, cellulolytic, thermophilic bacterium could be grown in culture with other thermo-tolerant microorganisms to produce ethanol at a high rate from cellulosic wastes; or the cellulolytic organism could be used to produce cellulase, which could be added to a cellulose-containing culture of an organism which ferments glucose to alcohol.
- Citation: D.M. Updegraff et al. 1986. Isolation and characterization of *Acidothermus cellulolyticus* gen. nov. species novum, a new genus of thermophilic, acidophilic, cellulolytic bacteria. International Journal of Systematic Bacteriology. 36(3):435-443.
- Photographs: Electron micrograph in citation
- Specimens: Deposited in American Type Culture Collection as ATCC 43068; type strain 11B
- YNP Comments: Acidic hot springs are common in the Norris Geyser Basin and throughout YNP. This bacterium digests cellulose, indicating the presence of wood fiber in the thermal feature, also common in YNP's rapidly changing geothermal ecosystem.

Scientific Name:	<i>Alicyclobacillus acidocaldarius</i>
Description:	Acidophilic, heterotrophic, thermophilic. Aerobic, gram-variable, spore-forming rods. It was originally called <i>Bacillus acidocaldarius</i> . It is similar to <i>Bacillus coagulans</i> , but is considerably more acidophilic.
Habitat Type:	Acid-sulfate, aqueous and terrestrial thermal features Temperature range for growth: 45-70°C Optimum temperature: 60-65°C pH range: 2.0-6.0 Optimum pH: 3.0-4.0
YNP Location:	1) Nymph Creek 2) Unnamed hot springs near Recess Spring, Realgar Spring and Pearl Geyser, all in Norris Geyser Basin
Abundance:	This population appears to be abundant since this is a common habitat type in YNP and 14 cultures of this organism were isolated from a variety of sources.
First Observed:	4/1/71
Other Locations:	This organism also been isolated in soil from an acid fumarole in Hawaii Volcano National Park.
Biotech Use:	None known
Citation:	T.D. Brock and G. Darland. 1971. <i>Bacillus acidocaldarius</i> species novum, an acidophilic thermophilic spore-forming bacterium. <i>Journal of General Microbiology</i> . 67:9-15.
Photographs:	Photomicrographs in citation
Specimens:	Deposited in American Type Culture Collection as ATCC 43033, 43034, 43035

Scientific Name: *Anaerobranca horikoshii*

Description: Moderately alkali-tolerant, obligately anaerobic, chemoorganotrophic, gram-positive, heterotrophic, mainly proteolytic, thermophilic bacteria with peritrichous rod-shaped cells that form one to three branches and exhibit only slight tumbling motility. Spores have not been detected.

Habitat Type: Temperature range for growth: 30-66°C
Optimum temperature: 57°C
pH range for growth: 6.5-10.3
Optimum pH: 8.5

YNP Location: Unnamed hot spring pools behind the Old Faithful Hotel and Ranger Station

First Observed: 5/90

Other Locations: No similar isolates have been obtained from other hot springs in YNP or other parts of the U.S., or from hot springs in Iceland, New Zealand, or Italy, or from man-made thermobiotic environments such as sewage sludge, compost, and manure piles.

Biotech Use: Potential industrial applications that require enzymes stable at high pH values (9.5 and above) and at temperatures above 50°C, including the use of proteases and xylanases for a non-chlorine bleaching process in the pulp and paper industry.

Citation: Engle, Marcella et al. 1995. Isolation and characterization of a novel alkali-tolerant thermophile, *Anaerobranca horikoshii* gen. nov., species novum Journal of Systematic Bacteriology. 45(3):454-461.

Photographs: Electron micrographs in citation

Specimens: NA

Scientific Name:	<i>Archaeoglobus spp.</i>
Description:	Using in situ 16S rRNA analysis, and specific cell enrichments and cloning with a laser microscope (optical tweezers), this novel hyperthermophilic archaea was isolated from a solfataric hot spring in Yellowstone's Hayden Valley. Originally identified as S10L, it grows anaerobically by fermentation of complex organic material and gains energy by reduction of sulfate. In contrast to other known members of this genus, S10L grows only at low salt concentrations up to 0.7% NaCl and represents the first hyperthermophilic sulfate-reducer from a non-marine environment.
Habitat Type:	Mineral spring; temperature range for growth: up to 90°C
YNP Location:	Obsidian Pool, about one mile SW of the Mud Volcano Parking area. Exit Board Walk at the Sulfur Caldron, follow trail to Moose Pool. Cross country through woods to an extension of Hayden Valley. The pool is SW of a small muddy lake, and the only black pool among predominantly muddy brown/gray acid features.
First Observed:	1993
Abundance:	This is the only known population of these organisms in YNP.
Other Locations:	None known.
Biotech Use:	No known applications.
Citation:	Barns, S.M., Fundyga, R.E., Jeffries, M.W., Pace, N.R. 1964. Remarkable archaeal diversity detected in a Yellowstone National Park hot spring environment. Proceedings of the National Academy of Science. 91:1609-1631.
Specimens:	N/A
YNP Comments:	This organism was discovered by Sue Barns and Norm Pace using PCR-based DNA fingerprinting. It was subsequently cultured and named by Stetter, et al of Regensburg, Germany. S10L represents a new archaeoglobus species within the Chrenarchaeota branch of the archaea.

Scientific Name: *Bacillus sp.*

Description: Acidophilic, gram-positive, microaerophilic, non-motile, terminal spore-forming thermophilic bacterium growing on hot spring sediments using pullulan as a carbon source. The strain reduced nitrate to nitrite both aerobically and anaerobically. It produced extracellular thermostable pullulanase and saccharidase activities which degraded pullulan and starch into maltotriose, maltose, and glucose. Gram-positive, non-motile rods with terminal oval spores occur in single cells, pairs or chains of 3 to 4 cells.

Habitat Type: Acid-sulfate, sediment
Temperature range for growth: 40-65°C
Optimum temperature: 62°C
pH range: 4-6
Optimum pH: 5.5

YNP Location: Unnamed hot spring

Abundance: This population is considered marginal since only one isolate was obtained.

First Observed: 10/1/89

Biotech use: This *Bacillus* produces three thermostable enzymes: Pullulanase, amylase and saccharidase. These enzymes are capable of hydrolyzing pullulan and starch into products with industrial utility. They can be used to increase starch saccharification and to produce high glucose and high maltose syrups.

Citation: Shen, Gwo-Jenn et al. 1990. Physiological and enzymatic characterization of a novel pullulan-degrading thermophilic *Bacillus* strain 3183. *Applied Microbiology Biotechnology*. 33:340-344.

Photographs: None

Specimens: Deposited in American Type Culture Collection as ATCC 49341 and 49342

Scientific Name:	<i>Bacillus coagulans</i>
Description:	Acidophilic, chemoorganotrophic, spore-forming thermophilic bacillus with gram-variable rods that grows in hot spring effluents. It is found in algal mats consisting of the eukaryotic brown alga, <i>Cyanidium caldarium</i> and lives off the algae's extracellular products.
Habitat Type:	Acid-sulfate Temperature range for growth: 30-60°C Optimum temperature: 37-45°C pH range: 2.0-6.0 Optimum pH: 3.0-4.0
Location:	Unnamed hot springs
Abundance:	The organism is probably common in <i>Cyanidium caldarium</i> mats since 17 isolates were cultured.
First Observed:	8/1/73
Other Locations:	This organism is abundant in acid hot springs in Yellowstone as well as around the world.
Biotech Use:	Because of its ability to produce acid from certain sugars and to grow at temperatures of 60°C, <i>B. coagulans</i> may have relevance as a possible causal organism of flat sour spoilage in acidic food products.
Citation:	Brock, Thomas D. and R.T. Belly. 1974. Widespread occurrence of acidophilic strains of <i>Bacillus coagulans</i> in hot springs. J Appl Bact. 37:175-177.
Specimens:	Deposited in American Type Culture Collection as ATCC 7050

Scientific Name: *Calothrix sp.*

Description: Filaments are solitary, in tufts or clumps, or in extended and definite strata, often more or less parallel, mostly erect, unbranched or with false branching, branches usually single, rarely in pairs; filaments and trichomes frequently swollen at bases and tapering into more or less well-developed hairs at apices. Sheaths are mostly firm, never confluent, often lamellate, often pigmented, or homogeneous and colorless. Heterocysts are basal and sometimes also intercalary. Resting cells in some species; basal, single or in rows of a few. Hormogonia often in long rows. Capable of nitrogen fixation.

They are often found associated with *Phormidium*, but in cooler regions. They seem to be of considerable importance in the formation of hot spring terraces and cones. They are the most important components of the Calothrix-Scytonema-Schizothrix formations that cover large areas in the Lower Geyser Basin, and the Calothrix-Diatom associations generally found in the basic, neutral and slightly acid springs. The accurate determination of several species is not easy and many characters are variable and unreliable. Additional study is likely to reveal several other species in the region.

Habitat Type: Neutral chloride and calcium carbonate

YNP Location: Firehole region, Mammoth Hot Springs

Abundance: 12 species have been found in YNP and comprise a very important group. They are characteristic of cooler waters (below 55°C), and are most abundant from 20-40°C. They include active rock-depositing species.

First Observed: 1898 (Tilden)

Biotech Use: None known

Citation: Copeland, Joseph J. Yellowstone Thermal Myxophyceae. Annals of the New York Academy of Sciences. 35:108.

Specimens: N/A

YNP Comments: Common cyanobacteria is responsible for the gray to dark brown mats associated with many thermal features. Possibly second in abundance only to *Phormidia* in Yellowstone thermal features.

Scientific Name: *Chloroflexus aurantiacus*

Description: Gliding, filamentous, photosynthetic, phototrophic, thermophilic bacterium that forms a mat along the floor and walls of hot spring effluents. The mat which grows from the top, leaving the unhealthy cells in the lower layers to die from light limitation. It can grow either aerobically or anaerobically in association with other organisms, from which organic products are obtained.

- When grown anaerobically: the organism produces bacteriochlorophylls *a* and *c*. It oxidizes sulfide under photoautotrophic or photoheterotrophic growth conditions and deposits elemental sulfur outside the cell.
- When grown aerobically: the organism acts as a chemoheterotroph. Aerobic chemoheterotrophic growth occurs in darkness or light. Bacteriochlorophyll syntheses cease, but some types of carotenoids continue to be made.

Filament coloration is orange except under anaerobic conditions in low light intensity, where it is dull green. At higher temperatures (above 50°C), it grows in association with the unicellular blue-green alga *Synechococcus lividus*. At lower temperatures (30-40°C), it grows in association with *Phormidium tenue*. In springs rich in sulfide, this organism can grow separately from blue-green algae.

Chloroflexus aurantiacus has characteristics of both green bacteria (light harvesting bacteriochlorophyll and location in chlorosomes) and purple nonsulphur bacteria (photosynthetic reaction, electron transport components, facultative metabolism, and nutritional versatility), but unlike either group, it is not diazotrophic. And unlike the strictly anaerobic, photoautotrophic nature of green bacteria, *Chloroflexus* is capable of growing as an aerobic heterotroph or an anaerobic photoheterotroph or photoautotroph. It thereby extends the taxonomic and phylogenetic limits of the "green line" of phototrophic bacteria; it is unique in that there have been no previous reports of filamentous or gliding phototrophic bacteria.

Habitat Type: Neutral chloride
 Temperature range: 50-70°C
 Optimum temperature range: 52-60°C
 Optimum pH range: 7.6-8.4

YNP Location:

- 1) Mushroom Spring, on the Howard Eaton trail 0.25km north of Great Fountain Geyser
- 2) Five Sisters Spring, near Octopus Spring in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
- 3) Twin Butte Vista Pool, on a rise 0.10km southeast of Great Fountain Geyser

First Observed: 6/1/71

- Other Locations: This population is found in several hot springs in YNP and around the world including Oregon, Guatemala, Iceland, Japan and New Zealand.
- Biotech Use: No known applications. However, its unique metabolic activity have made it the focus of considerable research in prokaryotic photosynthesis.
- Citation: Brock, Thomas D. and John Bauld. 1973. Ecological studies of *Chloroflexus*, a gliding photosynthetic bacterium. Arch. Mikrobiol. 92:267-284.
- Brock, Thomas D. and Michael T. Madigan. 1975. Photosynthetic sulfide oxidation by *Chloroflexus aurantiacus*, a filamentous, photosynthetic, gliding bacterium. Journal of Bacteriology. 122(2):782-784.
- Pierson, Beverly K. and richard W. Castenholz. 1974. A phototrophic gliding filamentous bacterium of hot springs, *Chloroflexus aurentiacus*, gen. and species novum Arch Microbiol. 100:5-24.
- Madigan, Michael T. and Ghanshyam D. Heda. 1986. Utilization of amino acids and lack of diazotrophy in the thermophilic anoxygenic phototroph *Chloroflexus aurantiacus*. Journal of General Microbiology. 132:2469-2473.
- Photographs: Electron micrographs in Pierson and Castenholz
- Specimens: Deposited in American Type Culture Collection as ATCC 29362, 29363, 29364; numerous university tissue culture collections
- YNP Comments: This is a very well-studied organism. The 55°C mat growing at the outflow channel of Octopus Spring has a 2.5cm layer of *Chloroflexus* which has been studied by Dr. Dave Ward of Montana State University. Using phylogenetic analysis, Dr. Ward and his students have found a guild structure of possibly a hundred organisms comprising this mat. Other researchers working on this organism have altered the site by overharvesting, which has caused concern. The mat vitality is being monitored closely.

Scientific Name:	<i>Chromatium tepidum</i>
Description:	Obligately phototrophic purple bacterium with gram-negative, motile, rod-shaped cells. It produces bacteriochlorophyll <i>a</i> and grows photoautotrophically with sulfide as an electron donor. It is also autotrophic in that it can oxidize sulfide to elemental sulfur, which is stored as globules inside the cell. It is not diazotrophic. It forms a reddish bacterial mat in the carbonate sinter of thermal springs containing a high amount of sulfide with neutral to alkaline pH. Its photosynthesis is anoxygenic.
Habitat Type:	Calcium-carbonate Temperature range: 34-57°C Optimum temperature: 48-50°C Optimum pH: 7.0
YNP Location:	Stygian Springs, located on the top of a ridge in the southwest corner of the Upper Terraces, Mammoth Hot Springs
Abundance:	The population of this organism is marginal since only one isolate was obtained in Yellowstone, and one in a hot spring in New Mexico.
First Observed:	2/1/84
Biotech Use:	No known application
Citation:	Madigan, Michael T. 1984. A novel photosynthetic purple bacterium isolated from a Yellowstone hot spring. <i>Science</i> . 225:313-315. Madigan, Michael T. 1986. <i>Chromatium tepidum</i> species novum a thermophilic photosynthetic bacterium of the family <i>Chromatiaceae</i> . <i>International Journal of Systematic Bacteriology</i> . 36(2):222-227.
Photographs:	Photomicrograph in <i>Science</i>
Specimens:	Deposited in American Type Culture Collection as ATCC 43061 MC

- Scientific Name: *Clostridium thermoautotrophicum*
- Description: Thermophilic strict anaerobe with round to slightly oval spores formed in terminal positions. Gram-variable, rod-shaped, vegetative cells that are slightly motile by peritrichously inserted flagella. It grows hemolithotrophically with hydrogen and carbon dioxide, and chemoorganotrophically with glucose, fructose, glycerate, or methanol. In both cases, acetate was the only organic fermentation product formed in significant amounts.
- Habitat Type: Neutral chloride and acid-sulfate
Temperature range: 36-70°C
Optimum temperature: 56-60°C
pH range: 4.5-7.6
Optimum pH: 5.7
- YNP Location: 1) White Creek hot spring (White Creek crosses the Firehole Loop Road, near White Dome Geyser)
2) Dragon's Mouth, which is off the boardwalk in the Mud Volcano developed thermal area near Canyon Village
- Abundance: The organism is widespread but low in numbers; fourteen isolates were obtained in Yellowstone from two hot springs.
- First Observed: 1/1/81
- Other Locations: Isolates of this organism have also been found in mud and wet soils from Hawaii, Georgia, Africa, Germany and Italy. It is not restricted to locations with elevated temperature.
- Biotech Use: This organism is unique in being able to carry out homoacetate fermentation, in which elemental hydrogen and carbon dioxide are utilized to produce acetate. This ability could lead to biotechnical applications, including the production of road-deicer.
- Citation: Wiegel, Jurgen et al. 1981. *Clostridium thermoautotrophicum* species novum, a thermophile producing acetate from molecular hydrogen and carbon dioxide. *Current Microbiology*. 5:255-260. 1982.

Validation of the publication of new names and new combinations previously effectively published outside the IJSB. *International Journal of Systematic Bacteriology*. 32(3):384-385.
- Photographs: Photomicrographs in *Current Microbiology*
- Specimens: Deposited in American Type Culture Collection as ATCC 33924 (DSM 1974); type strain JW 701/3

Scientific Name:	EM3, EM17, EM19 (Pink Filament Community)
Description:	Hyperthermophilic chemotrophic bacteria found in pink filaments attached to rock surfaces in a rapidly flowing hot water. Phylogenetic analysis identified three species: EM3, a unique lineage within the Thermotogales group; EM17, which predominates (26 out of 35 clones) and is closely related to <i>Aquifex pyrophilus</i> ; and EM19, which is also affiliated with the Aquificales group and is a close relative of another hydrogen-oxidizing bacterium, <i>Hydrogenobacter thermophilus</i> .
Habitat Type:	Neutral chloride Temperature range for growth: 84-88°C pH: 7.0
YNP Location:	Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
First Observed:	6/6/66
Abundance:	The population is marginal, since it has been found in the outflow channel of only one hot spring in Yellowstone.
Other Locations:	None known.
Biotech Use:	These extreme thermophiles could not be cultured using traditional culture enrichment techniques. Analysis of the sequences of phylogenetically informative molecules, such as the 16S short sub-unit rRNA, and comparison with a known database of characterized organisms identified the species within a phylogenetic framework.
Citation:	Reysenbach, Anna-Louise. 1994. Phylogenetic Analysis of the Hyperthermophilic "Pink Filament" Community in Octopus Spring, Yellowstone National Park. Applied and Environmental Microbiology. 60(6):2113-2119.
Photographs:	Photomicrographs in citation
Specimens:	Anna-Louise Reysenbach/Norm Pace Lab, Institute for Molecular and Cellular Biology.
YNP Comments:	Octopus Spring, called "Pool A" by early researchers, is an octopus-shaped pool approximately 15m in size which contains near boiling water at 91°C. It was the site of intense research by T.D. Brock in the 1960s and, due to its thermophilic biodiversity, is the best-studied microbial habitat in YNP. The pink filaments are probably similar to those organisms described by Setchell in 1899 as "filamentous Schizomycetes". The pink filament community, consisting of less than one square meter of habitat, is found no where else in YNP.

- Scientific Name: *Heliobacterium modesticaldum*
- Description: Diazotrophic, gram-negative, thermophilic obligate anaerobe found in hot spring microbial mats, motile by flagella or non-motile. The organism fixes nitrogen up to its growth temperature limit and forms heat-resistant cylindrical, subterminal endospores. It can grow phototrophically by photo-assimilating pyruvate, lactate, and acetate; or chemotrophically on pyruvate. Sugars and other organic acids are not utilized. The organism contains bacteriochlorophyll *g*, and lacks chlorosomes and intracytoplasmic membranes. Neurosporene is probably the only carotenoid.
- Habitat Type: Neutral to alkaline hot springs or volcanic soils
Temperature range: 25-56°C
Optimum temperature: 52°C
Optimum pH: 6.0-7.0
- YNP Location: Unnamed thermal springs near the Firehole River and Octopus Spring
- First Observed: 1/1/81
- Other Locations: Isolates of this organism have also been obtained in from volcanic soils in Iceland.
- Biotech Use: None known.
- Citation: Kimble, Linda K. et al. 1995. *Heliobacterium modesticaldum*, species novum, a thermophilic heliobacterium of hot springs and volcanic soils. Archives of Microbiology. 163:259-267.
- Photographs: Photomicrographs in citation
- Specimens: Deposited in American Type Culture Collection as ATCC 51577 (YS6)

Scientific Name:	<i>Mastigocladus laminosus</i>
Description:	Thermophilic cyanobacteria capable of nitrogen fixation and adaptation to grow at temperatures lower than its normal range; motile by gliding and produces branching filaments. It forms a gelatinous, cartilaginous, or spongy dull blue-green or olive-green mat, doubling about 1.5 times per day. It has little tolerance for hydrogen sulfide and produces spores that are resistant to freezing and drying.
Habitat Type:	Calcium-carbonate and neutral chloride Temperature range for growth: 60-70°C Optimum temperature: 63-64°C pH range: 6.2-7.0
YNP Location:	Widespread throughout YNP including: 1) Upper Geyser Basin: Bijah Geyser, Mastiff Geyser, Firehole River, Chromatic Spring, and elsewhere 2) Lower Geyser Basin: Jelly Spring 3) Twin Buttes Region: Spray Geyser; West Thumb and Mammoth
Abundance:	This cosmopolitan organism can be found in many Yellowstone hot springs, and in alkaline thermal waters throughout the world.
First Observed:	1/1/51
Biotech Use:	None known
Citation:	Castenholz, Richard W. 1969. Thermophilic blue-green algae and the thermal environment. <i>Bacteriological Reviews</i> . 33:476-504. Copeland, Joseph J. 1936. Yellowstone Thermal Myxophyceae. 1936 <i>Annals of the New York Academy of Sciences</i> . 36:1-232. Fogg, G.E. 1951. Studies on nitrogen fixation by blue-green algae. II. Nitrogen fixation by <i>Mastigocladus laminosus</i> . <i>Journal of Experimental Botany</i> . 2:117-120.
Photographs:	Drawings in Copeland, p. 88
Specimens:	Three strains, none preserved

- Scientific Name: *Methanobacterium thermoautotrophicum*
- Description: Anaerobic, asporogenous, gram-positive bacterium which forms long, irregular filaments during growth. It is a non-motile, non-sporing, thermophilic chemolithotroph. Its capable of producing methane and hydrogen during the decomposition of bacterial-algal mat biomass. The 55°C algal-bacterial mat at Octopus Spring is the biomass used in methanogenesis by this organism. Microbial methanogenesis is generally associated with the anaerobic decomposition of organic matter in the absence of electron acceptors, such as oxygen.
- Habitat Type: Acid sulfate and calcium carbonate
Temperature range: 42-72°C
Optimum temperature: 65°C
pH range: 6.4-8.4
- YNP Location: 1) Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
2) Washburn Pools A and B, several meters NW of Devils Ink Pot in the Washburn Springs area, SW of Mount Washburn
3) Firehole Pool A, located at Midway Geyser Basin next to the Firehole River, halfway between Ojo Caliente Spring and Fountain Freight Road Bridge
- Abundance: The population is common since it is found in many springs in Yellowstone. It has also been isolated in sewage sludge.
- First Observed: 7/1/80
- Biotech Use: This organism, due to the production of ethanol and methanol due to biomass decomposition, could have utility in converting biomass to fuels or chemical feedstock.
- Citation: Zeikus, J. G. et al. 1980. Microbiology of methanogenesis in thermal, volcanic environments. Journal of Bacteriology. 143:432-440.
- Specimens: Deposited in American Type Culture Collection as ATCC 29183 and 35610

Scientific Name:	<i>Phormidium laminosum</i>
Description:	Stratum is thin or less often thick, up to 1cm, membranous to grisly, bright blue-green, golden or reddish. Filaments are flexuous to contorted, entangled. Sheaths are colorless, close, of variable thickness, often papery, usually completely or almost completely confluent into an amorphous matrix. Trichomes are 1-2 microns in diameter, not constricted at the cross walls; apices straight, cylindrical except for the apical cell. Cells are longer than broad, 2-5 microns in length, mostly with 2-4 refringent granules on the often inconspicuous cross walls; cell content is pale blue-green. End cells are sharp and pointed, without calyptra. It forms an orange-tan and brownish strata in full sunlight, a blue-green layer when partially shaded, and reddish layers when occurring beneath the surface.
Habitat type:	Calcium carbonate and neutral chloride Temperature: up to 66°C pH range: 6.0 to 9.0
YNP Location:	Numerous hot springs throughout Yellowstone, including the Firehole region and Mammoth Terraces
Abundance:	<i>Phormidium</i> is a common cyanobacteria found in most non-acidic hot springs throughout Yellowstone and the world. In many springs, <i>P. laminosum</i> is the dominant alga over extended areas. Its orange-tan strata have given the name to Orange Mound Spring, on which it is abundant.
First Observed:	1898 (Tilden)
Biotech Use:	No known applications
Citation:	Copeland, Joseph J. 1936. Yellowstone Thermal Myxophyceae. 1936 Annals of the New York Academy of Sciences. 36:1-232. Mann, James E. and Harold E. Schlichting, Jr. Benthic algae of selected thermal springs in Yellowstone National Park. Trans Amer Microsc Soc. 86(1):2-9.
Photographs:	Photograph in Copeland, page 169.
Specimens:	N/A
YNP Comments:	This organism, and possibly many other strains of similar phenotype, is abundant in YNP. It is responsible for many of the beautiful colors associated with hot springs. An organism found in the Firehole region in 1963 and identified as <i>Phormidium bijahensis</i> is now considered to be <i>P. laminosum</i> .

Scientific Name: *pJP27*

Description: Using in situ 16S rRNA analysis, and specific cell enrichments and cloning with a laser microscope (optical tweezers), this novel hyperthermophilic archaea was isolated from a solfataric hot spring in Yellowstone's Hayden Valley. The organism is rod-shaped, slightly curved, variable in length (between 5 and 10 micrometers) and about 0.5 micrometers in diameter. It grows at 85°C, and is therefore considered a hyperthermophile.

Habitat Type: Mineral spring

YNP Location: Obsidian Pool, about one mile SW of the Mud Volcano Parking area. Exit Board Walk at the Sulfur Caldron, follow trail to Moose Pool. Cross country through woods to an extension of Hayden Valley. The pool is SW of a small muddy lake, and the only black pool among predominantly muddy brown/gray acid features.

First Observed: 1993

Abundance: This is the only known population of these organisms in YNP.

Other Locations: None known.

Biotech Use: No known applications.

Citation: Barns, S.M., Fundyga, R.E., Jeffries, M.W., Pace, N.R. 1964. Remarkable archaeal diversity detected in a Yellowstone National Park hot spring environment. *Proceedings of the National Academy of Science*. 91:1609-1631.

Specimens: N/A

YNP Comments: This organism was discovered by Sue Barns and Norm Pace using PCR-based DNA fingerprinting. It was subsequently grown in a mixed culture by Stetter, et al of Regensburg, Germany. PJP27 represents a pivotal new group within the phylogenetic tree that has been named Korarchaeota by Norm Pace. As determined by phylogenetic analysis (evolutionary history), pJP27 is considered the most primitive organism known and the closest known extant relation to the origin of life.

Scientific Name:	<i>Rhodopila globiformis</i>
Description:	Gram-negative cells are spherical to ovoid, and motile by means of polar flagella, have vesicular intracytoplasmic membranes, and grow only at low pH values. Photoorganotroph that forms purplish-red colonies when grown anaerobically with a light source, and pink colonies when grown microaerophilically without light. Discovered by Thomas Brock as a weak red layer of non-mobile, spherical cells from a 3.0 pH spring near the Gibbon River, sample #LIX, 197. The pigments possessed include bacteriochlorophyll a-p and aliphatic methoxylated ketocarotenoids. This organism must have a reduced sulfur source to live because it lacks an assimilatory sulfate reduction system.
Habitat Type:	Acid-sulfate Optimum temperature for growth: 30-35°C pH range: 4.2-6.5 Optimum pH: 4.8-5.6, depending on carbon source
YNP Location:	Gibbon River acid spring, Norris Junction
Abundance:	The population is considered marginal since it was only found in one spring in YNP.
First Observed:	9/1/74
Other Locations:	None known
Biotech Use:	No known application
Citation:	Imhoff, J.F. et al. 1984. Rearrangement of the species and genera of the phototrophic "purple nonsulfur bacteria". <i>International Journal of Systematic Bacteriology</i> . 34(3):340-343. Pfennig, Norbert. 1974. <i>Rhodopila globiformis</i> , species novum, a new species of the Rhodospirillaceae. <i>Archives of Microbiology</i> . 100:197-206.
Photographs:	Photomicrograph in Pfennig
Specimens:	Deposited in American Type Culture Collection as ATCC# 35887; DSM 161
YNP Comment:	This organism was previously included in the genus <i>Rhodopseudomonas</i> .

- Scientific Name: *Sulfolobus acidocaldarius*
- Description: Sulfur-oxidizing, facultative autotroph with spherical cells that occurs in the hot acid environments of solfataras. It produces a citrate synthesis resembling those found in gram-positive eubacteria and eukaryotes. A thermostable type II restriction enzyme, *SauI*, has been isolated from this organism that produces blunt-ends when it cleaves and is an isoschizomer of *BspRI*.
- Habitat Type: Acid-sulfate
Temperature range for growth: 56-92°C
Optimum temperature: 68-80° depending on strain
pH range for growth: 0.9-5.8
Optimum pH: 2-3
- YNP Location: 1) Sulfur Caldron, Moose Pool, Mud Geyser in the Mud Volcano area
2) Evening Primrose in the Sylvan Springs area
3) Vermillion and an unnamed spring (24-2) 450 feet west of Sieve Lake, in the Norris Geyser Basin
- Abundance: Common to acidic hot springs and solfataras in the Yellowstone region.
- First Observed: 1/1/72
- Other Locations: This organism has been found in a variety of aquatic and terrestrial acid thermal areas, including hot springs in Italy, Dominica, and El Salvador.
- Biotech Use: This organism is an important geochemical agent because it contributes to high acidity by oxidizing elemental sulfur to sulfuric acid, dissolving rock matrix. It has been used as a bioleaching agent during recovery of gold. Its enzymes are used in molecular biology and genetic engineering.
- Citation: Brock Thomas D. et al. 1972. *Sulfolobus*: A new genus of sulfur-oxidizing bacteria living at low pH and high temperature. *Arch Mikrobiol.* 84:54-68.

Bohlool, Ben B. and Thomas D. Brock. 1974. Population ecology of *Sulfolobus acidocaldarius*. I. Temperature strains; II. Immunoecological Studies. *Archives of Microbiology.* 97:169-179; 181-194.

Grossebüter, Wilhem and Helmut Görish. 1985. Partial purification and properties of citrate synthases from the thermoacidophilic archaeobacteria *Thermoplasma acidophilum* and *Sulfolobus acidocaldarius*. *System Appl Microbiology.* 6:119-124.

Prangishvili, D.A. et al. 1985. A restriction endonuclease *SuaI* from the thermoacidophilic archaeobacterium *Sulfolobus acidocaldarius*. *Federation of European Biochemical Societies.* 192(1):57-60.
- Photographs: Photomicrographs in Brock, 1972
- Specimens: Deposited in American Type Culture Collection as ATCC 33909

Scientific Name:	<i>Synechococcus sp.</i>
Description:	These organisms can be thermophilic or mesophilic. They are usually obligate photoautotrophs which undergo aerobic photosynthesis. The small cells can be cylindrical, ovoid, or rod-shaped and grow singly, in pairs or in short chains and have no sheaths; they are irregular aggregates united by a slime layer and undergo repeated binary fission in one plane to reproduce. Most of the species are nonmotile; the motile species glide and exhibit a positive phototactic response. 28 species of <i>Synechococcus</i> are known.
Habitat Type:	Acid-sulfate Temperature range: 35-53°C, with thermophiles growing above 53°C.
YNP Location:	Hot springs throughout YNP
Abundance:	These organisms can be found in acidic thermal features throughout Yellowstone National Park.
First Observed:	1/1/71
Other Locations:	These organisms have also been isolated from both thermal and non-thermal sources throughout the U.S. and the world.
Biotech Use:	No known applications
Citation:	Stanier, R.Y. et al. 1971. Purification and properties of unicellular blue-green algae (Order <i>Chroococcales</i>). <i>Bacteriological Review</i> . 35(2)171-205. Rippka, Rosemarie et al. 1979. Generic assignments, strain histories and properties of pure culture of cyanobacteria. <i>Journal of General Microbiology</i> . 111:1-61.
Photographs:	Photomicrographs in Rippka
Specimens:	Deposited in American Type Culture Collection as ATCC 27149 and 27180

- Scientific Name: *Thermoanaerobacter brockii*
- Description: Obligate anaerobic chemoorganotroph with gram-positive, non-motile rods that are frequently uneven in length and occur in chains, pairs or filaments. Forms round terminal endospores. Colonies are uniformly round, mucoid, non-pigmented and flat. The population doubles in about one hour at optimum temperature. It reduces thiosulfate to hydrogen sulfide. It uses a variety of saccharides as energy sources, including starch, maltose, glucose, lactose, sucrose and cellobiose. It does not ferment cellulose. Previously known as *Thermoanaerobium brockii*.
- Habitat Type: Acid-sulfate and neutral chloride
Temperature range for growth: 35-85°C
Optimum temperature: 65-70°C
pH range: 5.5-9.5
Optimum pH: 7.5
- YNP Location: 1) Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
2) Washburn Pools A and B, several meters NW of Devils Ink Pot in the Washburn Springs area, SW of Mount Washburn
3) Firehole Pool A is located at Midway Geyser Basin next to the Firehole River, halfway between Ojo Caliente Spring and Fountain Freight Road Bridge
- Abundance: This population is moderate since it is found in anaerobic decomposing photosynthetic biomass from hot springs throughout Yellowstone.
- First Observed: 7/1/80
- Other Locations: None known
- Biotech Use: The catabolic activity of *Thermoanaerobacter* strains is of industrial interest. It can grow on a variety of sugars, including insoluble starch, and produces largely ethanol and lactic acid as fermentation end products, and therefore may be of use in chemical feedstock production via thermophilic fermentation of biomass. It is used in the perfume industry.
- Citation: Zeikus, J.G. et al. 1979. *Thermoanaerobium brockii* genus novum, species novum; a new chemoorganotrophic, caldoactive, anaerobic bacterium. Archives of Microbiology. 122:41-48.

Lee, Yong-Eok et al. 1993. Taxonomic distinction of saccharolytic thermophilic anaerobes. International Journal of Systematic Bacteriology. 43(1):41-51.
- Photographs: Photomicrographs in Zeikus
- Specimens: Deposited in American Type Culture Collection as ATCC 33075 (DSM 1457)

Scientific Name:	<i>Thermoanaerobacter ethanolicus</i>
Description:	Extremely thermophilic, gram-variable, non-sporing anaerobic chemo-organotroph found in both alkaline (pH 8.8 and temperature 45-50°C) and slightly acidic (pH 5.5, temperature 55-60°C) hot springs. Peritrichously-flagellated rods in early growth phase; later cell shape varies from chains of coccoid cells one micron in diameter to long filamentous cells which often divide by constriction, forming chains of bacteria with differing cells lengths. Uneven division is common. This bacterium possesses two different alcohol dehydrogenases which enable it to metabolize many alcohols. It ferments glucose into ethanol and carbon dioxide. It reduces thiosulfate to hydrogen sulfide; it does not ferment cellulose.
Habitat Type:	Acid-sulfate and calcium-carbonate Temperature range for growth: 37-78°C Optimum temperature: 69°C pH range: 4.4-9.9 Optimum pH: 5.8-8.5
YNP Location:	1) White Creek, where Firehole Loop Road crosses it, on opposite side of road from Great Fountain Geyser 2) Dragons Mouth, in the Sulfur Caldron/Mud Volcano Area, Canyon Village
Abundance:	The organism has been found in only two YNP hot springs.
First Observed:	1/1/81
Other Locations:	None known outside of YNP.
Biotech Use:	This organism releases ethanol and carbon dioxide as fermentation products and has been used in the industrial production of ethanol. U.S. Patent #4,292,406 has been granted for its use in converting cellulose into ethanol.
Citation:	Jurgen Wiegel and Lars G. Ljungdahl. 1981. <i>Thermoanaerobacter ethanolicus</i> genus novum, species novum; a new extreme thermophilic anaerobic bacterium. Archives of Microbiology. 128:343-348. Wiegel, Jurgen et al. 1988. Purification and properties of primary and secondary alcohol dehydrogenases from <i>Thermoanaerobacter ethanolicus</i> . Applied and Environmental Microbiology. 54(2):460-465.
Photographs:	Photomicrographs in Wiegel
Specimens:	Deposited in American Type Culture Collection as ATCC 31550

- Scientific Name: *Thermoanaerobacterium thermosulfurigenes*
- Description: Obligately anaerobic, phototrophic, and thermophilic chemoorganotroph that grows in the algal-bacterial mat associated with thermal volcanic springs. Gram-negative, motile, peritrichously-flagellated straight rods that form long filaments and swollen, white, refractile, spherical endospores. It reduces thiosulfate to elemental sulfur; it does not ferment cellulose. It produces both ethanol and methanol as fermentation products, and β -amylase, which is extracellular, stable and active at high temperatures and a wide range of pH. Previously known as *Clostridium thermosulfurogenes*.
- Habitat Type: Acid sulfate and neutral chloride
Temperature range for growth: 55-75°C
Optimum temperature: 60°C
pH range: 4.0-7.6
Optimum pH: 5.5-6.5
- YNP Location: Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
- Abundance: This organism is considered marginal since it has only been found in one Yellowstone hot spring.
- First Observed: 5/1/82
- Other Locations: None known.
- Biotech Use: Due to unique properties of β -amylase, this organism could be used industrially in brewing and starch processing. Food and beverage industries use β -amylase to convert starch into maltose solutions. High value is placed on extreme thermostability and thermoactivity of amylases for use in the bioprocessing of starch.
- Citation: Hyun, H.H. and J.G. Zeikus. 1985. General biochemical characterization of thermostable extracellular β -amylase from *Clostridium thermosulfurogenes*. *Applied and Environmental Microbiology*. 49(5):1162-1167.
- Lee, Yong-Eok et al. 1993. Taxonomic distinction of saccharolytic thermophilic anaerobes. *International Journal of Systematic Bacteriology*. 43(1):41-51.
- Schink, Bernhard and J.G. Zeikus. 1983. *Clostridium thermosulfurogenes* species novum, a new thermophile that produces elemental sulfur from thiosulfate. *Journal of General Microbiology*. 129:1149-1158
- Photographs: Photomicrographs in Schink and Zeikus, 1983
- Specimens: Deposited in American Type Culture Collection as ATCC 33743 (DSM 2229)

Scientific Name:	<i>Thermoanaerobacterium xylanolyticum</i>
Description:	Anaerobic, thermophilic bacterium growing chemoorganotrophically in hot springs on xylan. Gram-negative, motile, short rod-shaped cells with spherical terminal spores. Surface colonies are circular with smooth edges cloudy to white. In addition to degrading xylan, it reduces thiosulfate to elemental sulfur, which it deposits on its cells. It does not ferment cellulose.
Habitat Type:	Acid-sulfate Temperature range for growth: 45-70°C Optimum temperature: 60°C pH range: 5.0-7.5 Optimum ph: 6.0
YNP Location:	Frying Pan Springs, which are small, shallow hydrothermal areas located in Sylvan Springs just off the Mammoth-Norris Road on the north side
First Observed:	1992
Abundance:	The population is marginal since it has only been found in Frying-Pan Springs.
Biotech Use:	The xylanolytic enzymes produced by this organism could have applications in the pulp and paper industry.
Citation:	Lee, Yong-Eok et al. 1993. Taxonomic distinction of saccharolytic thermophilic anaerobes. <i>International Journal of Systematic Bacteriology</i> . 43(1):41-51.
Photographs:	Photomicrographs in citation.
Specimens:	Deposited in American Type Culture Collection as ATCC 49914 (DSM 7097)

- Scientific Name: *Thermobacteroides acetoethylicus*
- Description: Caldoactive, thermophilic chemoorganotroph with gram-negative, non-sporing, obligately anaerobic rods that exist singly or in pairs and are motile by peritrichous flagellation. Colonies are uniformly round, mucoid, non-pigmented, flat, and grow to a diameter of 3mm in 48 hours. It has a distinctive multiple-layered cell wall architecture without an outer wall membrane. It uses a variety of saccharides as energy sources, including starch, maltose, glucose, lactose, sucrose and cellobiose.
- Habitat Type: Acid sulfate and neutral chloride
Temperature range for growth: 40-80°C
Optimum temperature: 65°C
pH range: 5.5-8.5
- YNP Location: 1) Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
2) Washburn Springs, SW of Mount Washburn along the trail from the Glacial Erratic in the Canyon area
- Abundance: Prevalent in volcanic features where organic matter is being decomposed.
- First Observed: 1/1/81
- Other Locations: None identified outside of YNP.
- Biotech Use: No biotechnical use has been found for this organism. Since it is known to produce ethanol and acetic acid as fermentation products, it is a candidate for future biotechnology research.
- Citation: Zeikus, J.G. et al. 1980. Microbiology of methanogenesis in thermal volcanic environments. *Journal of Bacteriology*. 143:432-440.

Ben-Bassat, Arie, and J.G. Zeikus. 1981. *Thermobacteroides acetoethylicus* genus novum, species novum; a new chemoorganotrophic anaerobic thermophilic bacterium. *Archives of Microbiology*. 128:365-370.
- Photographs: Photomicrographs in Ben-Bassat
- Specimens: Deposited in American Type Culture Collection as ATCC 33265

Scientific Name:	<i>Thermodesulfobacterium commune</i>
Description:	Thermophilic chemoorganotroph with a small gram-negative, obligate anaerobic, non-motile, non-sporing, straight rod which can reduce sulphate. It uses pyruvate, lactate or hydrogen as electron donors, and sulphate or thio-sulphate as electron acceptors for growth and sulphide formation. An outer wall membrane has been found on this organism, as have mesosomes in its interior.
Habitat Type:	Acid-sulfate and neutral chloride Temperature range for growth: 45-85°C Optimum temperature: 70°C pH range: 6.0-8.0
YNP Location:	1) Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek 2) Washburn Pool A, in the Washburn Springs area, SW of Mount Washburn, along the trail from the Glacial Erratic in the Canyon Area 3) Inkpot Spring in Firehole Pool A, located at Midway Geyser Basin next to the Firehole River, halfway between Ojo Caliente Spring and Fountain Freight Road Bridge
Abundance:	This population is considered common since it was found in decomposing biomass from four different locations in Yellowstone.
First Observed:	6/1/82
Other Locations:	None known outside of YNP
Biotech Use:	No known applications
Citation:	Zeikus, J.G. et al. 1983. Microbial ecology of volcanic sulphidogenesis: Isolation and characterization of <i>Thermodesulfobacterium commune</i> genus novum and species novum. Journal of General Microbiology. 129:1159-1169.
Photographs:	Photomicrographs in Zeikus
Specimens:	Deposited in American Type Culture Collection as ATCC 33708

- Scientific Name: *Thermofilum spp.*
- Description: Using in situ 16S rRNA analysis, and specific cell enrichments and cloning with a laser microscope (optical tweezers), this novel hyperthermophilic archaea was isolated from a solfataric hot spring in Yellowstone's Hayden Valley. Originally identified as S10TFL, it is a very thin, filamentous rod that grows anaerobically by fermentation of complex organic material at low salinity.
- Habitat Type: Mineral spring
Temperature range for growth: 70-90°C
pH: 5.5
- YNP Location: Obsidian Pool, about one mile SW of the Mud Volcano Parking area. Exit Board Walk at the Sulfur Caldron, follow trail to Moose Pool. Cross country through woods to an extension of Hayden Valley. The pool is SW of a small muddy lake, and the only black pool among predominantly muddy brown/gray acid features.
- First Observed: 1993
- Abundance: This is the only known population of these organisms in YNP.
- Other Locations: None known.
- Biotech Use: No known applications.
- Citation: Barns, S.M., Fundyga, R.E., Jeffries, M.W., Pace, N.R. 1964. Remarkable archaeal diversity detected in a Yellowstone National Park hot spring environment. *Proceedings of the National Academy of Science*. 91:1609-1631.
- Specimens: N/A
- YNP Comments: This organism was discovered by Sue Barns and Norm Pace using PCR-based DNA fingerprinting. It was subsequently cultured and named by Stetter, et al of Regensburg, Germany.

Scientific Name:	<i>Thermoleophilum album</i>
Description:	Obligately thermophilic aerobe which grows on hydrocarbons at a narrow range of <i>n</i> -alkanes (from 13-20 carbons in length) in a defined mineral salts medium. Small gram-negative, non-motile, non-pigmented non-sporing, rod-shaped cells.
Habitat Type:	Neutral chloride Temperature range for growth: 45-70°C Optimum temperature: 60°C pH range: 6.5-7.5 Optimum pH: 7.0
YNP Location:	Octopus Spring, in Lower Geyser Basin, 300m upstream from where the Firehole Loop Road crosses White Creek
First Observed:	6/15/77
Other Locations:	This organism has also been found in mud and water samples in primarily thermal areas in Arkansas, New Mexico and North Carolina.
Biotech Use:	No known applications. However, its ability to consume hydrocarbons has made it a candidate for further study.
Citation:	Perry, J.J. et al. 1978. Isolation of Thermophilic bacteria capable of growth solely in long-chain hydrocarbons. FEMS Microbiology Letters. 3:81-83. Zarilla, K.A. and J.J. Perry. 1984. <i>Thermoleophilum album</i> genus novum and species novum, a bacterium obligate for thermophily and <i>n</i> -alkane substrates. Archives of Microbiology. 137:286-290.
Photographs:	SEM photomicrographs in Zarilla
Specimens:	Deposited in American Type Culture Collection as ATCC #35264

Scientific Name: *Thermoleophilum minutum*

Description: Obligately thermophilic aerobe which grows on hydrocarbons at a narrow range of *n*-alkanes (from 13-20 carbons in length) in a defined mineral salts medium. Small gram-negative, non-motile, non-pigmented non-sporing, rod-shaped cells.

Habitat Type: Neutral chloride
Temperature range for growth: 45-70°C
Optimum temperature: 60°C
pH range is 6.0-7.0
Optimum pH: 6.8

YNP Location: Unidentified hot spring in YNP

Abundance: It was recovered from only one YNP hot spring.

First Observed: 6/15/85

Biotech Use: No known application. However, its ability to consume hydrocarbons makes it a candidate for bioremediation processes.

Citation: Zarilla, K.A. and J.J. Perry. 1986. Deoxyribonucleic acid homology and other comparisons among obligately thermophilic hydrocarbonoclastic bacteria, with a proposal for *Thermoleophilum minutum* species novum. International Journal of Systematic Bacteriology. 36(1):13-16

Specimens: Deposited in American Type Culture Collection as ATCC #35265

Scientific Name:	<i>Thermomicrobium roseum</i>
Description:	Obligately thermophilic and aerobic bacterium with gram-negative, pleomorphic, non-sporing irregularly-shaped rods, occurring singly or in pairs in compact pink colonies. It has an atypical cell wall composed predominantly of a protein with high concentrations of proline, glutamic acid, glycine, and alanine. The organism grows in an orangish-red bacterial mat.
Habitat Type:	Calcium-carbonate Optimum temperature for growth: 70-75°C Maximum temperature: 85°C pH range: 6.0-9.4 Optimum pH: 8.2-8.5
YNP Location:	Toadstool Spring outflow, two miles north of Mushroom Spring in the Lower Geysir Basin
Abundance:	This organism has been found in only a few hot springs in YNP.
First Observed:	6/15/72
Other Locations:	None known outside of YNP
Biotech Use:	No known applications
Citation:	Ramaley, Robert F. et al. 1973. <i>Thermomicrobium</i> , a new genus of extremely thermophilic bacteria. <i>International Journal of Systematic Bacteriology</i> . 23(1):28-36. Perry, G.J. et al. 1980. The atypical cell wall composition of <i>Thermomicrobium roseum</i> . <i>Can J Microbiol</i> . 25:556-559.
Photographs:	Photomicrographs in Ramaley
Specimens:	Deposited in American Type Culture Collection as ATCC 27502

Scientific Name:	<i>Thermoplasma acidophilum</i>
Description:	Saprophytic, obligately heterotrophic and thermoacidophilic archaebacterium found in acidic solfatara sediments. Facultatively anaerobic organotroph which gains energy by sulfur respiration. Gram-negative, motile, flagellate, irregularly-shaped cells lack cell wall and envelope; surrounded by only a cytoplasmic membrane. Closely related to <i>Thermoplasma volcanium</i> .
Habitat Type:	Acid-sulfate Temperature range for growth: 45-63°C Optimum temperature: 59°C pH range: 0.5-4.0 Optimum pH: 1.0-2.0
YNP Location:	Nymph Lake, Frying Pan Springs, West Thumb, and Mud Volcano
Abundance:	This organism is abundant; isolates were obtained throughout the YNP acid-sulfate habitat type. (Until this discovery, this organism was believed to exist only in self-heated coal refuse piles; Brock et al.)
First Observed:	11/1/87
Other Locations:	In addition to the sites chosen in YNP, this organism has been found in the Azores, Iceland, Indonesia, Italy, and Java.
Biotech Use:	No known application
Citation:	Seegerer, Andreas et al. 1988. <i>Thermoplasma acidophilum</i> and <i>Thermoplasma volcanium</i> species novum from solfatara fields. Systematic and Applied Microbiology. 10:161-171.
Photographs:	Photomicrographs in citation
Specimens:	Deposited in American Type Culture Collection as ATCC 25905 (DSM 1728)

Scientific Name:	<i>Thermoplasma volcanium</i>
Description:	Saprophytic, obligately heterotrophic and thermoacidophilic archaeobacterium found in acidic solfatara sediments. Facultatively anaerobic organotroph which gains energy by sulfur respiration. Gram-negative, motile, flagellate, irregularly-shaped cells lack cell wall and envelope; surrounded by only a cytoplasmic membrane. Closely related to <i>Thermoplasma acidophilum</i> .
Habitat Type:	Acid-sulfate Temperature for growth: 33-67°C. Optimum temperature: 60°C pH range: 1.0-4.0 Optimum pH: 2.0
YNP Location:	Sulfur Caldron and Mud Volcano in the developed thermal area between Canyon Village and Yellowstone Lake
Abundance:	This organism is abundant; isolates were obtained throughout the YNP acid-sulfate habitat type.
First Observed:	11/1/87
Other Locations:	In addition to the sites chosen in YNP, this organism is found in acid-thermal habitats around the world.
Biotech Use:	No known application
Citation:	Segerer, Andreas et al. 1988. <i>Thermoplasma acidophilum</i> and <i>Thermoplasma volcanium</i> species novum from solfatara fields. Systematic and Applied Microbiology. 10:161-171.
Photographs:	Photomicrographs in citation
Specimens:	DSM 4299

- Scientific Name: *Thermosphaera aggregans*
- Description: Using in situ 16S rRNA analysis, and specific cell enrichments and cloning with a laser microscope (optical tweezers), this novel hyperthermophilic archaea was isolated from a solfataric hot spring in Yellowstone's Hayden Valley. Originally identified as M11TL, it grows anaerobically by fermentation of complex organic material at up to 90°C. Hydrogen, carbon dioxide, acetate, and iso-valerate are formed as metabolic products. It is a coccoid archaeum that forms grape-like aggregates and represents a new genus.
- Habitat Type: Mineral spring; temperature for growth: up to 90°C
- YNP Location: Obsidian Pool, about one mile SW of the Mud Volcano Parking area. Exit Board Walk at the Sulfur Caldron, follow trail to Moose Pool. Cross country through woods to an extension of Hayden Valley. The pool is SW of a small muddy lake, and the only black pool among predominantly muddy brown/gray acid features.
- First Observed: 1993
- Abundance: This is the only known population of these organisms in YNP.
- Other Locations: None known.
- Biotech Use: No known applications.
- Citation: Barns, S.M., Fundyga, R.E., Jeffries, M.W., Pace, N.R. 1964. Remarkable archaeal diversity detected in a Yellowstone National Park hot spring environment. *Proceedings of the National Academy of Science*. 91:1609-1631.
- Specimens: N/A
- YNP Comments: This organism was discovered by Sue Barns and Norm Pace using PCR-based DNA fingerprinting. It was subsequently cultured and named by Stetter, et al of Regensburg, Germany. M11TL represents a new genus within the Chrenarchaeota branch of the archaea.

Scientific Name:	<i>Thermothrix thiopara</i>
Description:	Extremely thermophilic chemosynthetic sulfur autotroph which occurs where geothermal ground waters mix with the oxidizing atmosphere and covers tufa microenvironments with microscopic mats of gram-negative cells. It has a complex life cycle, establishing itself at the sulfide-oxygen interface and forming long (2.5 cm) macroscopic streamers of filamentous cells which grow until they extend to the oxidizing side of the interface, where they fragment into flagellated unicells. This strategy apparently optimizes reproductive success despite the rapid washout of planktonic unicells due to their low specific growth rate relative to the dilution rate of the spring.
Habitat Type:	Calcium-carbonate Temperature at site: 74°C Optimum temperature for growth in lab: 72°C pH: 7.0
YNP Location:	Mammoth Hot Springs
Abundance:	This organism is very abundant, occurring in most calcium-carbonate hot springs in YNP and every spring in Mammoth; it deposits the mineral travertine and is the principal fossil/mineral component of the Mammoth Terraces.
Biotech Use:	No known application. However, it is important in understanding the biogeochemical cycles of Earth's development.
First Observed:	6/15/76
Other Locations:	The organism has also been found in the Jemez hot springs and at Soda Dam in New Mexico.
Citation:	Caldwell, D.E. et al. 1982. <i>Thermothrix thiopara</i> : Selection and adaptation of a filamentous sulfur-oxidizing bacterium colonizing hot spring tufa at pH 7.0 and 74°C. <i>Environmental Biogeochemistry</i> . 35:1-6.
Photographs:	Photomicrographs in citation
Specimens:	None preserved

- Scientific Name: *Thermus aquaticus* (Taq)
- Description: Thermophilic, plagic, obligate aerobic eubacterium. Gram-negative, non-motile non-sporing rods which produce a yellow cellular pigment, probably a carotenoid. Under certain cultural conditions, it forms long filaments and large spherical structures, probably related to spheroplasts. Generation time at optimum temperature is about 50 minutes. Taq was the first hyperthermophile discovered and led to the science of life at high temperature, eventually yielding many other bacteria growing at temperatures above 55°C.
- Habitat Type: Neutral chloride
Temperature range for growth: 40-79°C
Optimum temperature: 70°C
pH range: 7.0-8.0
Optimum pH: 7.5-7.8
- YNP Location: Mushroom Pool, located in the White Creek area of the Lower Geyser Basin, off the Firehole Loop Road, 300m east of White Dome Geyser, just inside the tree line. It is a circular pool approximately 20m in diameter with an L-shaped outflow channel and covered with a dense green/brown bacterial mat.
- First Observed: 6/1/65
- Other Locations: Taq is a common thermophilic microorganism with several strains found in hot springs around the world. It is also sometimes found in hot tap water in geographical locations distant from thermal springs.
- Biotech Use: Taq DNA polymerase is the primary enzyme used in polymerase chain reaction (PCR), the enzymatic amplification of DNA for industrial use, which has revolutionized DNA science. The gene producing Taq polymerase has been cloned onto an *E. coli* host for mass production of this enzyme, a process patented by Cetus, Inc. using the original culture collected from Mushroom Pool.
- Citation: Brock, Thomas D. and Hudson Freeze. 1969. *Thermus aquaticus* genus novum and species novum; a non-sporulating extreme thermophile. *Journal of Bacteriology*. 98(1)289-297.
- Photographs: Photomicrographs in citation
- Specimens: Deposited in American Type Culture Collection as ATCC YT-1 25104

Scientific Name:	<i>Thiobacillus thiooxidans</i>
Description:	Chemoautotrophic, rod-shaped, thermophilic, obligately acidophilic bacterium found in solfatara soils. It is responsible for the sulfuric acid production that takes place in solfatara soils. It oxidizes sulfur to sulfuric acid most rapidly at lower temperatures. The habitat generally has little or no vegetation, consists of hydrothermally altered reddish or white soil, with patches of sulfur crystals in the rocks and soil.
Habitat Type:	Acid-sulfate Temperature range: 20-55°C pH range: 0.9-4.5 Optimum pH: 2.5
YNP Location:	A level plain at the southwestern base of Roaring Mountain, a barren white ridge of highly eroded acidic steam vents and solfatara, located north of Norris Junction on the Grand Loop Road.
Abundance:	These organisms are abundant in solfatara habitat types and have also been isolated from other locations within YNP, including Amphitheater Springs.
First Observed:	8/1/72
Other Locations:	This organism is common in solfatara habitats elsewhere in the world.
Biotech Use:	It is used in mine bioleaching operations, and is associated with acid mine pollution.
Citation:	Brock, Thomas D. and Carl Fliermans. 1972. Ecology of sulfur-oxidizing bacteria in hot acid soils. <i>Journal of Bacteriology</i> . 111(2):343-350.
Photographs:	N/A
Specimen:	N/A

BIBLIOGRAPHY

(As of September 1995)

- Aldhous, Peter. 1992. Roche Gets Tough on Illicit Sales of PCR Reagent. *Science*. 258:1572.
- Aldhous, Peter. 1993. PCR Enzyme Patent Challenged. *Science*. 260:486.
- Allen, E.T. 1934. The Agency of Algae in the Deposition of Travertine and Silica from Thermal Waters. *American Journal of Science*. 28:373-389.
- Amann, Rudolf I. et al. 1992. Diversity Among Fibrobacter Isolates: Towards a Phylogenetic Classification. *System Appl Microbiol*. 15:23-31.
- Anderson, Christopher. 1992. Roche Cuts Controversial PCR Fees, Testing Limits. *Nature* 355:379.
- Appenzeller, Tim. 1992. Deep-Living Microbes Mount a Relentless Attack on Rock. *Science*. 258:9.
- Arnhelm, Norman and Corey H. Levenson. 1990. Polymerase Chain Reaction. *C&En*. October 1:36-47.
- Arnold. 1926. A Short Discussion of the Algae of Yellowstone National Park. 8.
- Baggley, Herma Geneva Albertson. 1929. Some Preliminary Studies of the Algae of Mammoth Hot Springs Pertaining to the Coloring of the Terraces. 24.
- Baker, Dean. 1985. Giardia! *National Wildlife*. 23(5):19-21.
- Barany, Francis. 1991. Genetic Disease Detection and DNA Amplification Using Cloned Thermostable Ligase. *Proceedings of The National Academy of Science*. 88:189-193.
- Barany, Francis and David H. Gelfand. 1991. Cloning, Overexpression and Nucleotide Sequence of a Thermostable DNA Ligase-Encoding Gene. *Gene*. 109:1-11.
- Barany, Francis. 1991. The Ligase Chain Reaction in a PCR World. *PCR Methods and Applications*. Cold Spring Harbor Laboratory Press. 1:5-16.
- Barany, Francis et al. 1992. The Corrected Nucleotide Sequences of the Taqi Restriction and Modification Enzymes Reveal a Thirteen-Codon Overlap. *Gene*. 112:91-95.
- Barany, Francis and John Zebala. 1992. Correlation Between Insertion Mutant Activities and Amino Acid Sequence Identities of the Taqi and Tthhb8 Restriction Endonucleases. *Gene*. 112:13-20.
- Barany, Francis et al. 1992. Cloning and Sequencing of Genes Encoding the Tthhb8i Restriction and Modification Enzymes: Comparison with the Isoschizomeric Taqi Enzymes. *Gene*. 112:3-12.

- Barany, Francis et al. 1992. DNA Recognition of Base Analogue and Chemically Modified Substrates by the TaqI Restriction Endonuclease. *Journal of Biological Chemistry*. 267(12):8106-8116.
- Barany, Francis et al. 1992. Characterization of Steady State, Single-Turnover, and Binding Kinetics of the Taq Restriction Endonuclease. *Journal of Biological Chemistry*. 267(12):8097-8105.
- Barinaga, Marcia. 1991. Biotech Nightmare: Does Cetus Own PCR? *News & Comment*. February:739.
- Baskin, Yvonne. 1990. DNA Unlimited. *Discover*. July:77-79.
- Bateson, Mary M. and D.M. Ward. 1988. Photoexcretion and Fate of Glycolate in a Hot Spring Cyanobacterial Mat. *Applied and Environmental Microbiology*. 54(7):1738-1743.
- Bauld, J. 1974. Algal Extraction and Bacterial Assimilation in Hot Spring Algal Mats. *Journal of Phycology*. 10(1):101-106.
- Belly, Robert T. et al. 1973. Algal Excretion of ¹⁴C-Labeled Compounds and Microbial Interactions with *Cyanidium Caldarium*. *Journal of Phycology*. 9(2):123-127.
- Bohlool, Ben B. 1976. Occurrence of *Sulfolobus-Acidocaldarius* an Extremely Thermophilic Acidophilic Bacterium in New Zealand Hot Springs — Isolation and Immunofluorescence Characterization. *Archives of Microbiology*. 106(3):171-174.
- Bott, Thomas and Thomas Brock. 1969. Bacterial Growth Rates above 90°C in Yellowstone Hot Springs. *Science*. 164:1411-1412.
- Bott, Thomas. 1970. Growth and Metabolism of Periphytic Bacteria: Methodology. *Limnology and Oceanography*. 15(3):333-342.
- Brierley, J.A. 1963. Dipicolinic Acid Content and Heat Resistance of Spores of *Bacillus Stearothermophilus* and Thermophilic Bacteria from Yellowstone National Park. M.S. Thesis, Montana State University, Bozeman.
- Brierley, J.A. 1966. Contribution of Chemoautotrophic Bacteria to the Acid Thermal Waters of the Geyser Springs Group in Yellowstone National Park. Ph.d. Dissertation, Montana State University, Bozeman. 104
- Brock, Thomas D. and M. Louise Brock. 1966. Temperature Optima for Algal Development in Yellowstone and Iceland Hot Springs. *Nature*. 209(5024):733-734.
- Brock, Thomas D. 1967. Microorganisms Adapted to High Temperatures. *Nature*. 214(5091):882-885.
- Brock, Thomas D. and M. Louise Brock. 1967. The Measurement of Chlorophyll, Primary Productivity, Photophosphorylation and Macromolecules in Benthic Algal Mats. *Limnology and Oceanography*. 12(4):600-605.
- Brock, Thomas D. 1967. Relationship Between Standing Crop and Primary Productivity Along a Hot Spring Thermal Gradient. *Ecology*. 48(4):566-571.

- Brock, Thomas D. 1968. Taxonomic Confusion Concerning Certain Filamentous Blue-Green Algae. *Journal of Phycology*. 4(3):178-179.
- Brock, Thomas D. and M. Louise Brock. 1968. Measurement of Steady-State Growth Rates of a Thermophilic Alga Directly in Nature. *Journal of Bacteriology*. 95(3):811-815.
- Brock, Thomas D. and M. Louise Brock. 1968. Life in a Hot-Water Basin. *Natural History*. 45-54.
- Brock, Thomas D. 1969. Microbial Growth under Extreme Conditions. *Symposia of the Society for General Microbiology*. No. 19. Great Britain.
- Brock, Thomas D. and M. Louise Brock. 1969. Effect of Light Intensity on Photosynthesis by Thermal Algae Adapted to Natural and Reduced Sunlight. *Limnology and Oceanography*. 14(3):334-341.
- Brock, Thomas D. and M. Louise Brock. 1969. Recovery of a Hot Spring Community from a Catastrophe. *Journal of Phycology*. 5(1):75-77.
- Brock, Thomas D. 1970. High Temperature Systems. *Annual Review of Ecology and Systematics*. 1:191-200.
- Brock, Thomas D. and Gary K. Darland. 1970. Limits on Microbial Existence: Temperature and pH. *Science*. 169:1316-1318.
- Brock, Thomas D. and M. Louise Brock. 1970. The Algae of Waimangu Cauldron (New Zealand): Distribution in Relation to pH. *Journal of Phycology*. 6(4):371-375.
- Brock, Thomas D. and William N. Doemel. 1970. The Upper Temperature Limit of Cyanidium Caldarium. *Archives of Microbiology*. 72:326-332.
- Brock, Thomas D. 1971. Bimodal Distribution of pH Values of Thermal Springs of the World. *Geological Society of American Bulletin*. 82:1393-1394.
- Brock, Thomas D. et al. 1971. Effect of Temperature on the Fatty Acid Composition of *Thermus Aquaticus*. *Journal of Bacteriology* 106(1):25-30.
- Brock, Thomas D. and M. Louise Brock. 1971. Temperature Optimum of Non-Sulfur Bacteria from a Spring at 90°C. *Nature*. 233(20):494-495.
- Brock, Thomas D. and M. Louise Brock. 1971. Life in the Geyser Basins. *Yellowstone Library and Museum Association*. No. 17
- Brock, Thomas D. 1971. Microbial Growth Rates in Nature. *Bacteriological Reviews*. 35(1):39-58.
- Brock, Thomas D. and Gary K. Darland. 1971. *Bacillus Acidocaldarius* Species Novum, an Acidophilic Thermophilic Spore-Forming Bacterium. *Journal of General Microbiology*. 67:9-15.

- Brock, Thomas D. and M. Louise Brock. 1971. Microbiological Studies of Thermal Habitats of the Central Volcanic Region, North Island, New Zealand. *New Zealand Journal of Marine and Freshwater Research*. 5(2):233-258.
- Brock, Thomas D. and Jerry L. Mosser. 1971. Effect of Wide Temperature Fluctuation on the Blue-Green Algae of Bead Geyser, Yellowstone National Park. *Limnology and Oceanography*. 16(4):640-645.
- Brock, Thomas D. and W.N. Doemel. 1971. The Physiological Ecology of *Cyanidium Caldarium*. *Journal of General Microbiology*. 67:17-32.
- Brock, Thomas D. et al. 1972. *Sulfolobus*: a New Genus of Sulfur-Oxidizing Bacteria Living at Low pH and High Temperature. *Archives of Microbiology*. 84:54-68.
- Brock, Thomas D. and J. Gregory Zeikus. 1972. Effects of Thermal Additions from the Yellowstone Geyser Basins on the Bacteriology of the Firehole River. *Ecology*. 53(2):283-290.
- Brock Thomas D. and Carl B. Fliermans. 1972. Ecology of Sulfur-Oxidizing Bacteria in Hot Acid Soils. *Journal of Bacteriology*. 3(2):343-350.
- Brock, Thomas D. and R.T. Belly. 1972. Cellular Stability of a Thermophilic, Acidophilic *Mycoplasma*. *Journal of General Microbiology*. 73:465-469.
- Brock, Thomas D. et al. 1972. Technique for Measuring $^{14}\text{CO}_2$ Uptake By Soil Microorganisms in Situ. *Applied Microbiology*. 23(3):595-600.
- Brock, Thomas D. et al. 1972. Microbiology and Morphogenesis of Columnar Stromatolites (Concophyton, *Vacerrilla*) from Hot Springs in Yellowstone National Park. Elsevier Scientific Publishing Company. Amsterdam. 273-310.
- Brock, Thomas D. et al. 1972. Siliceous Algal and Bacterial Stromatolites in Hot Spring and Geyser Effluents of Yellowstone National Park. *Science*. 178(4059):402-405.
- Brock, Thomas D. and Kathryn L. Boylen. 1973. Presence of Thermophilic Bacteria in Laundry and Domestic Hot-Water Heater. *Applied Microbiology*. 25(1):72-76.
- Brock, Thomas D. 1973. Bacterial Origin of Sulfuric Acid in Geothermal Habitats. *Science*. 179:1323-1324.
- Brock, Thomas D. and Carl B. Fliermans. 1973. Assay of Elemental Sulfur in Soil. *Soil Science*. 115(2):120-122.
- Brock, Thomas D. and Tansey. 1973. *Dactylaria Gallopara*, a Cause of Avian Encephalitis, in Hot Spring Effluents, Thermal Soils and Self-Heated Coal Waste Piles. *Nature*. 242(5394):202-203.
- Brock, Thomas D. and Charles W. Boylen. 1973. Bacterial Decomposition Processes in Lake Wingra Sediments During Winter. *Limnology and Oceanography*. Brock, Thomas D. and J. Bauld. 1973. Ecological Studies of *Chloroflexis*, A Gliding Photosynthetic Bacterium. *Archives of Microbiology*. 92:267-284.

- Brock, Thomas D. and Charles W. Boylen. 1973. Effects of Thermal Additions from the Yellowstone Geyser Basins on the Benthic Algae of the Firehole River. *Ecology*. 54(6):1282-1291.
- Brock, Thomas D. 1973. Primary Colonization of Surtsey with Special Reference to the Blue-Green Algae. *Oikos*. 24:239-243.
- Brock, Thomas D. and David W. Smith. 1973. The Water Relations of the Alga *Cyanidium Caldarium* in Soil. *Journal of General Microbiology*. 79:219-231.
- Brock, Thomas D. and David W. Smith. 1973. Water Status and the Distribution of *Cyanidium Caldarium* in Soil. *Journal of Phycology*. 9(3):330-332.
- Brock, Thomas D. 1973. Lower pH Limits for the Existence of Blue-Green Algae: Evolutionary and Ecological Implications. *Science*. 179:480-483.
- Brock, Thomas D. and R.T. Belly. 1974. Ecology of Iron-Oxidizing Bacteria in Pyritic Materials Associated with Coal. *Journal of Bacteriology*. 117(2):726-732.
- Brock, Thomas D. and B. B. Bohlool. 1974. Immunodiffusion Analysis of Membranes of *Thermoplasma Acidophilum*. *Infection and Immunity*. 10(1):280-281.
- Brock, Thomas D. and Ben B. Bohlool. 1974. Population Ecology of *Sulfolobus Acidocaldarius*. *Immunoecological Studies*. *Archives of Microbiology*. 97:181-194.
- Brock, Thomas D. and Robert T. Belly. 1974. Widespread Occurrence of Acidophilic Strains of *Bacillus Coagulans* in Hot Springs. *Journal of Applied Bacteriology*. 37:175-177.
- Brock, Thomas D. et al. 1974. Growth Rates of *Sulfolobus Acidocaldarius* in Nature. *Journal of Bacteriology*. 118(3):1075-1081.
- Brock, Thomas D. and Ben B. Bohlool. 1974. Immunofluorescence Approach to the Study of the Ecology of the *Thermoplasma Acidophilum* in Coal Refuse Material. *Applied Microbiology* 28(1):11-16
- Brock, Thomas D. and William N. Doemel. 1974. Bacterial Stromatolites: Origin of Laminations. *Science*. 184:1083-1085
- Brock, Thomas D. et al. 1974. Population Ecology of *Sulfolobus Acidocaldarius*. *Archives of Microbiology*. 97:169-179.
- Brock, Thomas D. 1974. Nutritional Studies on *Chloroflexus*, a Filamentous Photosynthetic, Gliding Bacterium. *Archives of Microbiology*. 100:97-103.
- Brock, Thomas D. and John Bauld. 1974. Algal Excretion and Bacterial Assimilation in Hot Spring Algal Mats. *Journal of Phycology*. 10(1):101-106.
- Brock, Thomas D. and Jerry L. Mosser. 1975. Rate of Sulfuric-Acid Production in Yellowstone National Park. *Geological Society of America Bulletin*. 86:194-198.

- Brock, Thomas D. 1975. Effect of Water Potential on a Microcoleus (Cyanophyceae) from a Desert Crust. *Journal of Phycology*. 2(3):316-320.
- Brock, Thomas D. 1975. Salinity and the Ecology of *Dunaliella* from Great Salt Lake. *Journal of General Microbiology*. 89:285-292.
- Brock, Thomas D. and Michael T. Madigan. 1975. Photosynthetic Sulfide Oxidation by *Chloroflexus Aurantiacus*, a Filamentous, Photosynthetic, Gliding Bacterium. *Journal of Bacteriology*. 122(2):782-784.
- Brock, Thomas D. 1975. Predicting the Ecological Consequences of Thermal Pollution from Observations on Geothermal Habitats. International Atomic Energy Agency, Vienna. 599-622.
- Brock, Thomas D. 1975. Effect of Water Potential on Growth and Iron Oxidation by *Thiobacillus Ferrooxidans*. *Applied Microbiology*. 29(4):495-501.
- Brock, Thomas D. et al. 1975. Photochemical Activity of Single Blue-Green Algal Cells Recorded by the Use of Nuclear Tracking Emulsion. *Photosynthetica*. 9(3):331-332.
- Brock, Thomas D. and John Gustafson. 1976. Ferric Iron Reduction By Sulfur- and Iron-Oxidizing Bacteria. *Applied and Environmental Microbiology*. 32(4):567-571.
- Brock, Thomas D. et al. 1976. Biogeochemistry and Bacteriology of Ferrous Iron Oxidation in Geothermal Habitats. *Geochimica et Cosmochimica Acta*. 40:493-500.
- Brock, Thomas D. and William N. Doemel. 1976. Vertical Distribution of Sulfur Species in Benthic Algal Mats. *Limnology and Oceanography*. 21(2):237-244.
- Brock, Thomas D. 1977. Adaptation by Hot Spring Phototrophs to Reduced Light Intensities. *Archives of Microbiology*. 113:111-120.
- Brock, Thomas D. 1977. Co₂ Fixation in Photosynthetically-Grown *Chloroflexus Aurantiacus*.
- Brock, Thomas D. and W.N. Doemel. 1977. Structure, Growth, and Decomposition of Laminated Algal-Bacteria Mats in Alkaline Hot Springs. *Applied and Environmental Microbiology*. 34(1):433.
- Brock, Thomas D. 1992. Business and Science. *Science*. 259:441.
- Brock, Thomas D. and M.P. Starr. 1978. *Thermophilic Microorganisms and Life at High Temperatures*. Springer-Verlag, New York, New York. 449.
- Brock, Thomas D. 1971. Microbial Adaptation to Extremes of Temperature and pH Biochemical Responses to Environmental Stress. Plenum Press, New York. 32-37.
- Brock, Thomas D. 1972. Microbiological Observation on Surtsey, 1970. Surtsey Progress Report 6. 11-13.

- Brock, Thomas D. 1972. One Hundred Years of Algal Research in Yellowstone National Park; in Taxonomy and Biology of Blue-Green Algae. Desikachary, T.V.(ed). Center for Advanced Study in Botany, Madras, India.
- Brock, Thomas D. 1973. Evolutionary and Ecological Aspects of the Cyanophytes; in The Biology of Blue-Green Algae. N.G. Carr and B.A. Whitton (Eds.). Chapter 24:487-500.
- Brock, Thomas D. 1974. Environmental Microbiology of Living Stromatolites. Developments in Sedimentology, 20. Stromatolites. M.R. Walter (Ed.). Chapter 4.3:141-148.
- Brock, Thomas D. 1974. Biological Techniques for the Study of Microbial Mats and Living Stromatolites. Developments in Sedimentology, 20. Stromatolites. M.R. Walter (Ed.). Chapter 2.3: 21-29.
- Brock, Thomas D. 1991. Basic Research in Yellowstone Thermal Environments Leads to a \$300 Million Industry. University of Wisconsin, Madison, Wisconsin. No 11.
- Brown, Douglas et al. 1957. The Effect of Growth Temperature on the Heat Stability of Bacterial Pyrophosphatase. Archives of Biochemistry and Biophysics. 70(1):248-256.
- Burns, Louise and Walter Miltzeretal. 1951. On the Existence of a Cell Granule in a Thermophilic Bacterium. Proceedings of the Society for Experimental Biology and Medicine. 76:598-601.
- Burns, Louise and Walter Miltzeretal. 1953. Organelle Nature of a Cell Granule from a Thermophilic Bacterium. Proceedings of the Society for Experimental Biology and Medicine. 82:411-413.
- Caldwell, D. E. 1982. Thermothrix Thiopara; Selection and Adaptation of a Filamentous Sulfur-Oxidizing Bacterium Colonizing Hot Spring Tufa at pH 7.0 and 74°C. Environmental Biogeochemistry. 35:1-6.
- Carballeira, N. et al. 1990. Purification of a Thermostable DNA Polymerase from Thermus Thermophilus Hb8, Useful in the Polymerase Chain Reacton. Biotechniques. 9(3):276-281.
- Castenholz, Richard W. and Jack A. Peary. 1964. Temperature Strains of a Thermophilic Blue-Green Algae. Nature. 202(4933):720-721.
- Castenholz, Richard W. and Richard P. Sheridan. 1968. Production of Hydrogen Sulphide by a Thermophilic Blue-Green Alga. Nature. 217(5133):1063-1064.
- Castenholz, Richard W. 1969. The Thermophilic Cyanophytes of Iceland and the Upper Temperature Limit. Journal of Phycology. 5(4):360-368.
- Castenholz, Richard W. 1969. Thermophilic Blue-Green Algae and the Thermal Environment. Bacteriological Reviews. 33(4):476-504.
- Castenholz, Richard W. 1970. Laboratory Culture of Thermophilic Cyanophytes. Schweizerische Zeitschrift Fur Hydrologie. 32(2):538-551.

- Castenholz, Richard W. and Lawrence N. Halfen. 1970. Gliding in a Blue-Green Algae: A Possible Mechanism. *Nature*. 225(5238):1163-1165.
- Castenholz, Richard W. and John C. Meeks. 1971. Growth and Photosynthesis in an Extreme Thermophile, *Synechococcus Lividus* (Cyanophyta). *Arch. Mikrobiol.* 78:25-41.
- Castenholz, Richard W. and Beverly K. Pierson. 1971. Bacteriochlorophylls in Gliding Filamentous Prokaryotes from Hot Springs. *Nature New Biology*. 233(35):25-27.
- Castenholz, Richard W. and L.N. Halfen. 1971. Energy Expenditure for Gliding Motility in a Blue-Green Alga. *Journal of Phycology*. 7(3):258-260.
- Castenholz, Richard W. and L.. Halfen. 1971. Gliding Motility in the Blue-Green Alga *Oscillatoria Princeps*. *Journal of Phycology*. 7(2):133-145.
- Castenholz, Richard W. 1972. Hot Spring Microbial Communities Recreated in Modified "Winogradski Columns". *Limnology and Oceanography*. 17(5):767-772.
- Castenholz, Richard W. 1973. The Possible Photosynthetic Use of Sulfide by the Filamentous Phototrophic Bacteria of Hot Springs. *Limnology and Oceanography*. 18(6):863-876.
- Castenholz, Richard W. 1974. Studies of Pigments and Growth in *Chloroflexus Aurantiacus*, a Phototrophic Filamentous Bacterium. *Archives of Microbiology*. 100:283-305.
- Castenholz, Richard W. and Beverly K. Pierson. 1974. A Phototrophic Gliding Filamentous Bacterium of Hot Springs, *Chloroflexus Aurantiacus*, gen. and species novum *Archives of Microbiology*. 100:5-24.
- Castenholz, Richard W. and John E. Jackson. 1975. Fidelity of Thermophilic Blue-Green Algae to Hot Spring Habitats. *Limnology and Oceanography*. 20(3):305-322.
- Castenholz, Richard W. 1976. The Effect of Sulfide on the Blue Green Algae of Hot Springs. I. New Zealand and Iceland. *Journal of Phycology*. 12(1):54-68.
- Castenholz, Richard W. 1977. The Effect of Sulfide on the Blue-Green Algae of Hot Springs. II. Yellowstone National Park. *Microbial Ecology*. 3:79-105.
- Castenholz, Richard W. and Conrad E. Wickstrom. 1978. Association of *Pleurocaps* and *Calothrix* (Cyanophyta) in a Thermal Stream. *Journal of Phycology*. 14:84-88.
- Castenholz, Richard W. 1966. Environmental Requirements of Thermophilic Blue-Green Algae. Symposium Sponsored by University of Washington and Non-springal Water Pollution Control Administration, Pacific Northwest Water Laboratory. 55-79.
- Castenholz, Richard W. 1973. Ecology of Blue-Green Algae in Hot Springs. *The Biology of Blue-Green Algae*. N.G. Carr and B.A. Whitton (eds.). 379-414.
- Delong, Edward et al. 1989. Phylogenetic Stains: Ribosomal RNA-Based Probes for the Identification of Single Cells. *Science*. 243:1360-1363.

- Dickson, David. 1993. European Patent for PCR Enzyme Clouded by Russian Claim. *Nature*. 364:2.
- Distel, D.L. 1988. Sulfur-Oxidizing Bacterial Endosymbionts: Analysis of Phylogeny and Specificity by 16s rRNA Sequences. *Journal of Bacteriology*. 170(6):2506-2510.
- Dobson, G. et al. 1988. Biogeochemistry of Hot Spring Environments: Extractable Lipids of a Cyanobacterial Mat. 68:155-179.
- Dyer, Denzel L. and Robert D. Gafford. 1961. Some Characteristics of a Thermophilic Blue-Green Algae. *Science*. 134(3479):616-617.
- Ehrlich, G.G. and R. Schoen. 1968. Bacterial Origin of Sulfuric Acid in Sulfurous Hot Springs. U.S. Geological Survey, Menlo Park, California.
- Estep, M.L.F. 1984. Carbon and Hydrogen Isotopic Compositions of Algae and Bacteria from Hydrothermal Environments, Yellowstone National Park. *Geochimica et Cosmochimica Acta*. 48(3):591-599.
- Estep, M.L.F. 1982. Stable Isotopic Composition of Algae and Bacteria That Inhabit Hydrothermal Environments in Yellowstone National Park. Annual Report of the Director Geophysical Laboratory, Washington, D.C. 402-410.
- Farmer, J.D. and D.J. Des Marsia. 1992. Biological Versus Inorganic Processes in Stromatolite Morphogenesis: Observations from Mineralizing Systems. *Microbial Mats: Structure, Development and Environmental Significance*. L.J. Stal and P. Caumette (Eds.) Nato Asi Series in Ecological Sciences. Springer Verlag. 1-10.
- Ferris, F.G. et al. 1986. Iron-Silica Crystallite Nucleation by Bacteria in a Geothermal Sediment. *Nature*. 320(6063):609-611.
- Foerster, Harold F. 1983. Activation and Germination Characteristics Observed in Endospores of Thermophilic Strains of *Bacillus*. *Archives of Microbiology*. 134:175-181.
- Fogg, G.E. 1951. Studies on Nitrogen Fixation by Blue-Green Algae. II. Nitrogen Fixation by *Mastigocladus Laminosus* Cohn. *Journal of Experimental Botany*. 2(1):117-120.
- Foos, K. Michael and Judith A. Royer. 1989. A Survey of *Pilobolus* from Yellowstone National Park. *Mycotaxon*. 34(2):395-397.
- Foos, K. Michael and Judith A. Royer. 1989. Isolation of *Pilobolus* spp. From the Northern Elk Herd in Yellowstone National Park. *Journal of Wildlife Diseases*. 25(2):302-304.
- Foos, K. Michael. 1989. *Pilobolus*, a Caprophilous Fungus with Epizootiologic Implications. Montana State University, Veterinary Research Laboratory Seminar. No. 14.
- Fredrick, Jerome F. and Joseph Seckbach. 1981. Search for Primitive Life Forms — *Cyanidium Caldarium* May Be a Missing Link. *Explorers Journal*. 59(1):22-25.
- Fedrick, Jerome F. 1983. Yellowstone: Nature's Laboratory. *Explorer's Journal*. 61(2):64-67.

- Fyfe, W.S. et al. 1986. Manganese Oxide Deposition in a Hot Spring Microbial Mat. *Geomicrobiology Journal*. 5(1):33-41.
- Garcia-Pichal, Ferran and Richard W. Castenholz. 1991. Characterization and Biological Implications of Scytonemin, A Cyanobacterial Sheath Pigment. *Journal of Phycology*. 27:395-409.
- Gelfand, David H. and Thomas J. White. *Thermostable DNA Polymerases. PCR Protocols: a Guide to Methods and Applications*. Academic Press, Inc. 129-139.
- Gelfand, David H. 1992. Taq DNA Polymerase. *PCR Technology: Principles and Applications for DNA Amplification*. Henry A. Erlich (ed.). W.H. Freeman Company, New York. 17-22.
- Georgi, Carl et al. 1955. The Organelle Nature of a Particle Isolated from *Bacillus Stearothermophilus*. *Journal of Bacteriology*. 70(6):716-725.
- Giovannoni, S. J. et al. 1987. Obligately Phototrophic Chloroflexus: Primary Production in Anaerobic Hot Spring Microbial Mats. *Archives of Microbiology*. 147:80-87.
- Gold, Thomas. 1992. The Deep, Hot Biosphere. *Proceedings of the National Academy of Science*. 89:6045-6049.
- Gorden, R.W. 1972. Bacterial Succession of a Thermal Stream Ecosystem in Yellowstone National Park. Annual Meeting of the American Society for Microbiology. Washington, D.C.
- Grohmann, K.A. et al. 1986. Isolation and Characterization of *Acidothermus Cellulolyticus* genus novum species novum; a New Genus of Thermophilic, Acidophilic, Cellulolytic Bacteria. *International Journal of Systematic Bacteriology*. 36(3):435-443.
- Grossebuter, Wilhelm and Helmut Gorisch. 1985. Partial Purification and Properties of Citrate Synthases from the Thermoacidophilic Archaeobacteria *Thermoplasma Acidophilum* and *Sulfolobus Acidocaldarius*. *Systematic and Applied Microbiology*. 6:119-124.
- Hawkes, Nigel. 1993. Going to Extremes. *The Times Magazine*. August:34.
- Heda, Ghanshyam D. and Michael T. Madigan. 1988. Thermal Properties and Oxygenase Activity of Ribulose-1,5-Bisphosphate Carboxylase from the Thermophilic Purple Bacterium, *Chromatium Tepidum*. *Federation of European Microbiological Societies*. 51:45-50.
- Heda, Ghanshyam D. and Michael T. Madigan. 1989. Purification and Characterization of the Thermostable Ribulose-1,5-Bisphosphate Carboxylase/Oxygenase from the Thermophilic Purple Bacterium *Chromatium Tepidum*. *European Journal of Biochemistry*. 184:313-319.
- Himmel, Michael E. et al. 1989. Ultra-Thermostable Cellulases from *Acidothermus Cellulolyticus*: Comparison of Temperature Optima with Previously Reported Cellulases. *Biotechnology*. 7:817-820.
- Huber, Robert and Karl O. Stetter. 1992. The Order Thermoproteales. *The Prokaryotes*. Springer. Berlin, Heidelberg, New York. Chapter 28:677-683.

- Imhoff, J.F. et al. 1984. Rearrangement of the Species and Genera of the Phototrophic "Purple Nonsulfur Bacteria". *International Journal of Systematic Bacteriology*. 34(3):340-343.
- Inman, O.I. 1940. Studies on the Chlorophylls and Photosynthesis of Thermal Algae from Yellowstone National Park, California, and Nevada. *Journal of General Physiology*. 23(6):661-666.
- International Journal of Systematic Bacteriology*. 1982. Validation of the Publication of New Names and New Combinations Previously Effectively Published Outside the IJSB. 32(3):384-385.
- International Journal of Systematic Bacteriology*. 1983. Validation of the Publication of New Names and New Combinations Previously Effectively Published Outside the IJSB. 33(3):672-674; 33(4):896-897.
- Jackson, T. and R.F. Ramaley. 1973. Physiological Properties of Two New Pink Thermophilic Gram Negative Bacteria. Annual Meeting of the American Society for Microbiology. Washington, D.C.
- Jones, William. 1965. Algae Distribution as Related to Water Temperature in Selected Portions of the Firehole River, Yellowstone National Park. National Park Service No. 36.
- Juranek, Dennis D. 1986. Giardiasis: Transmission and Control. *Water Research Updates*. 2(1):1-6.
- Karl, David M. 1988. A Microbiological Study of Guaymas Basin High Temperature Hydrothermal Vents. *Deep Sea Research*. 35(5):777-791.
- Keeler, Robert. 1991. Uses for PCR Are Multiplying in Gene-Related Research. *R&D Magazine*. August:30-34.
- Kempner, Ellis S. 1963. Upper Temperature Limit of Life. *Science*. 142(3597):1318-1319.
- Kleinschmidt, M.G. and Vern A. McMahon. 1970. Effect of Growth Temperature on the Lipid Composition of *Cyanidium Caldarium*. *Plant Physiology*. 46:290-293.
- Kullberg, Russell G. 1977. The Effects of Some Ecological Factors on Cell Size of the Hot Spring Alga *Synechococcus Lividus* (Cyanophyta). *Journal of Phycology*. 13(2):111-115.
- Kwoh, Deborah Y. and T. Jesse Kwoh. 1990. Target Amplification Systems in Nucleic Acid-Based Diagnostic Approaches. *Biotechniques*. 14-24.
- Lauwers, A.M. and W. Heinen. 1981. Growth of Bacteria at 100°C and Beyond. *Archives of Microbiology*. 129(2):127-128.
- Lawyer, Frances C. 1989. Isolation, Characterization, and Expression in *Escherichia Coli* of the DNA Polymerase Gene from *Thermus Aquaticus*. *The Journal of Biological Chemistry*. 264(11):6427-6437.
- Ljungdahl, Lars G. 1979. Ethanol Fermentation Using Anaerobic Thermophilic Bacteria. 27th IUPAC Congress. Helsinki. 546.

- Lukeroth, C. and H.S. Ginoza. 1973. Protein and DNA Synthesis in a Thermophilic Bacterium. Annual Meeting of the American Society for Microbiology. Washington, D.C.
- Macaskie, Lynne E. 1991. The Application of Biotechnology to the Treatment of Wastes Produces from the Nuclear Fuel Cycle: Biodegradation and Bioaccumulation as a Means of Treating Radionuclide-Containing Streams. *Critical Reviews in Biotechnology*. 11(1):41-112.
- Macaskie, Lynne E. et al. 1992. Uranium Bioaccumulation by a *Citrobacter* Sp. as a Result of Enzymically Mediated Growth of Polycrystalline HUO_2PO_4 . *Science*. 257:782-784.
- Madigan, Micheal T. et al. 1974. Nutritional Studies on *Chloroflexus*, a Filamentous Photosynthetic, Gliding Bacterium. *Archives of Microbiology*. 100:97-103.
- Madigan, Michael T. and Thomas Brock. 1976. Quantitative Estimations of Bacteriochlorophyll *c* in the Presence of Chlorophyll *a* in Aquatic Environments. *Limnology and Oceanography*. 21(3):462-467.
- Madigan, Michael T. and Thomas D. Brock. 1977. CO_2 Fixation in Photosynthetically Grown *Chloroflexus Aurantiacus*. *FEMS Microbiology Letters*. 1:301-304.
- Madigan, Michael T. and Thomas D. Brock. 1977. Adaptation by Hot Spring Phototrophs to Reduced Light Intensities. *Archives of Microbiology*. 113:111-120.
- Madigan, Michael T. 1984. A Novel Photosynthetic Purple Bacterium Isolated from a Yellowstone Hot Spring. *Science*. 225:313-315.
- Madigan, Michael T. 1986. *Chromatium Tepidum* Species Novum, a Thermophilic Photosynthetic Bacterium of the Family Chromatiaceae. *International Journal of Systematic Bacteriology*. 36(2):222- 227.
- Madigan, Michael T. et al. 1986. Organization of Intracytoplasmic Membranes in a Novel Thermophilic Purple Photosynthetic Bacterium as Revealed by Absorption, Circular Dichroism and Emission Spectra. *Biochimica et Biophysica Acta*. 852:191-197.
- Madigan, Michael T. et al. 1986. The Light-Harvesting Complexes of a Thermophilic Purple Sulfur Photosynthetic Bacterium *Chromatium Tepidum*. *Biochimica et Biophysica Acta*. 850:390-395.
- Madigan, Michael T. and Ghanshyam D. Heda. 1986. Utilization of Amino Acids and Lack of Diazotrophy in the Thermophilic Anoxygenic Phototroph *Chloroflexus Aurantiacus*. *Journal of General Microbiology*. 132:2469-2473.
- Madigan, Michael T. et al. 1989. Carbon Isotope Fractionation by Thermophilic Sulfur Bacteria: Evidence for Autotrophic Growth in Natural Populations. *Applied and Environmental Microbiology*. 55(3):639-644.
- Madigan, Michael T. and Sol M. Resnick. Isolation and Characterization of a Mildly Thermophilic Nonsulfur Purple Bacterium Containing Bacteriochlorophyll *b*. *FEMS Microbiology Letters*. 65:165-170.

- Mann, James E. and H. Schichting. 1967. Benthic Algae of Selected Thermal Springs in Yellowstone National Park. *Trans Amer Microsc Soc.* 86(1):2-9.
- Mann, Stephen et al. 1990. Magnetotactic Bacteria: Microbiology, Biomineralization, Palaeomagnetism and Biotechnology. *Advances in Microbial Physiology.* 31:125-181.
- Mann, H. et al. 1985. Biological Accumulation of Different Chemical Elements by Microorganisms from Yellowstone National Park, USA. National Park Service, Yellowstone National Park, Wyoming.
- Marsh, Connell and Don Larsen. 1953. Characterization of Some Thermophilic Bacteria from the Hot Springs of Yellowstone National Park. *Journal of Bacteriology.* 65(2):193-197.
- Merkel, G.J. et al. 1978. Isolation of Thermophilic Bacteria Capable of Growth Solely in Long-Chain Hydrocarbons. *FEMS Microbiology Letters.* 3:81-83.
- Meyer, Dr. Richard L. 1981. A Qualitative Evaluation of the Algae Associated with the Springs and Formations in Hot Springs. National Park No. 27.
- Ming, Shiang et al. 1991. Regulation of Cellulase Synthesis in *Acidothermus Cellulolyticus*. *Biotechnology Progress.* 7:315-322.
- Mullis, Kary B. 1988. Primer-Directed Enzymatic Amplification of DNA with a Thermostable DNA Polymerase. *Science.* 239:487-491.
- Munster, Michael J. et al. 1986. Isolation and Preliminary Taxonomic Studies of *Thermus* Strains Isolated from Yellowstone National Park. *Journal of General Microbiology.* 132:1677-1683.
- Murray, R.G. E. and K.H. Schleifer. 1994. Taxonomic Notes: a Proposal for Recording the Properties of Putative Taxa of Prokaryotes. *International Journal of Systematic Bacteriology.* 44(1):174-176.
- National Park Service, Water Resources Division. 1985. Field Survey of *Giardia* in Streams and Wildlife of the Glacier Gorge and Loch Vale Basins. Natural Resources Report 85-3. Fort Collins, Colorado. 65.
- O'Sullivan, Dermot A. 1979. Biotechnology Gains Brighten Resource Outlook. *C&En.* 27-29.
- Olson, G.J. and D.M. Ward. 1980. Terminal Processes in the Anaerobic Degradation of an Algal-Bacteria Mat in a High-Sulfate Hot Spring. *Applied and Environmental Microbiology.* 40(1):67.
- Pace, Norman R. et al. 1985. Analyzing Natural Microbial Populations by rRNA Sequences. *American Society for Microbiology News.* 51(1):4-12.
- Pace, Norman R. et al. 1986. Ribosomal Rna Phylogeny and the Primary Lines of Evolutionary Descent. *Cell.* 45:325-326.
- Pace, Norman R. et al. 1990. Ribonuclease P: Function and Variation. *The Journal of Biological Chemistry.* 265(7):3587-3590.

- Pace, Norman R. et al. 1991. Origin of Life — Facing up to the Physical Setting. *Cell*. 65:531-533.
- Pace, Norman R. et al. 1993. Characterization of the RNase P RNA of *Sulfolobus Acidocaldarius*. *Journal of Bacteriology*. 175(16):5043-5048.
- Pace, Norman R. et al. 1994. Remarkable Archaeal Diversity Detected in a Yellowstone National Park Hot Spring Environment. *Proceedings of the National Academy of Science, U.S.A.* 91:1609-1613.
- Pace, Norman R. et al. 1994. Phylogenetic Analysis of the Hyperthermophilic "Pink Filament" Community in Octopus Spring, Yellowstone National Park. *Applied and Environmental Microbiology*. 60(6):2113-2119.
- Parkes, John and James Maxwell. 1993. Some like it Hot (and Oily). *Nature*. 365:694-695.
- Pentecost, Allan. 1990. The Formation of Travertine Shrubs: Mammoth Hot Springs, Wyoming. *Geology Magazine*. 127(2):159-168.
- Perry, J.J. et al. 1978. Isolation of Thermophilic Bacteria Capable of Growth Solely in Long-Chain Hydrocarbons. *FEMS Microbiology Letters*. 3(2):81-83.
- Perry, J.J. et al. 1980. The Atypical Cell Wall Composition of *Thermomicrobium Roseum*. *Canadian Journal of Microbiology*. 26:556-559.
- Pfennig, Norbert. 1974. *Rhodopseudomonas Globiformis*, species novum; a New Species of the Rhodospirillaceae. *Archives of Microbiology*. 100:197-206.
- Prangishvili, D.A. et al. 1985. A Restriction Endonuclease *SuaI* From the Thermoacidophilic Archaeobacterium *Sulfolobus Acidocaldarius*. *Federation of European Biochemical Societies*. 192(1):57-60.
- Ramaley, Robert F. et al. 1973. *Thermomicrobium*, a New Genus of Extremely Thermophilic Bacteria. *International Journal of Systemic Bacteriology*. 23(1):28-36.
- Ramaley, Robert F. and Keith Bitzinger. 1975. Types and Distribution of Obligate Thermophilic Bacteria in Man-Made and Natural Thermal Gradients. *Applied Microbiology*. 30(1):152-155.
- Ramaley, Robert F. et al. 1975. Isolation of a New Pink Obligately Gram-Negative Bacterium (K-2 Isolate). *International Journal of Systematic Bacteriology*. 25(4):357-364.
- Ramaley, Robert F. et al. 1978. The Morphology and Surface Structure of Some of the Extremely Thermophilic Bacteria Found In Slightly Alkaline Hot Springs. National Science Foundation Grant #7602707.
- Reysenbach, Anna-Louise et al. 1994. Phylogenetic Analysis of the Hyperthermophilic "Pink Filament" Community in Octopus Spring, Yellowstone National Park. Department of Biology and Institute for Molecular and Cellular Biology. Indiana University. Bloomington, Indiana. No. 25.

- Rose, Stanley D. 1990. RNA PCR: an Application Kit. *Biotechniques*. 31-32.
- Roylen, C.W. 1973. Bacterial Decomposition Processes in Lake Wingra Sediment During Winter. Reprinted from *Limnology and Oceanography*. Grafton, Wisconsin. July.
- Runnion, Kenneth et al. 1992. Thermally Stable Urease from Thermophilic Bacteria. *Biocatalysis at Extreme Temperatures: Enzyme Systems near and above 100°C*. American Chemical Society Symposium Series. Atlanta, Georgia. 42-58. Chapter 4:42-58.
- Sandbeck, Kenneth A. and David M. Ward. 1981. Fate of Immediate Methane Precursors in Low-Sulfate, Hot Spring Algal-Bacterial Mats. *Applied and Environmental Microbiology*. 41(3):775- 782.
- Saul, D.J. et al. 1993. Phylogeny of Twenty *Thermus* Isolates Constructed from 16s rRNA Gene Sequence Data. *International Journal of Systematic Bacteriology*. 43(4):754-760.
- Schink, Bernhard. 1983. *Clostridium Thermosulfurogenes* Species Novum, a New Thermophile That Produces Elemental Sulfur from Thiosulphate. *Journal of General Microbiology*. 129:1149-1158.
- Schink, Bernhard et al. 1983. Radioassay for Hydrogenase Activity in Viable Cells and Documentation of Aerobic Hydrogen-Consuming Bacteria Living in Extreme Environments. *Applied and Environmental Microbiology*. 45(5):1491-1501.
- Schmidt, Thomas M. et al. 1991. Analysis of a Marine Picoplankton Community by 16s rRNA Gene Cloning and Sequencing. *Journal of Bacteriology*. 173(14):4371-4378.
- Sears, L.E. et al. 1992. Circumvent Thermal Cycle Sequencing and Alternative Manual and Automated DNA Sequencing Protocols Using the Highly Thermostable Vent (Exo) DNA Polymerase. *Biotechniques*. 13(4):626-633.
- Sederoff, Ronald. 1993. Availability of Taq Polymerase. *Science*. 259:1521.
- Seegerer, Andreas et al. 1988. *Thermoplasma Acidophilum* and *Thermoplasma Volcanium* species novum from Solfataria Fields. *Systemic and Applied Microbiology*. 10:161-171.
- Seegerer, Andreas H. and Karl O. Stetter. 1992. The Order Sulfolobales. *The Prokaryotes*. Springer. Berlin, Heidelberg, New York. Chapter 29:684-701.
- Seegerer, Andreas H. and Karl O. Stetter. 1992. The Genus *Thermoplasma*. *The Prokaryotes*. Springer, Berlin, Heidelberg, New York. Chapter 32:712-718.
- Shiea, Jentaie et al. 1991. Comparative Analysis of Extractable Lipids in Hot Spring Microbial Mats and Their Component Photosynthetic Bacteria. *Organic Geochemistry*. 17(3):309-319.
- Shiea, Jentaie et al. 1989. Mid-Chain Branched Mono- and Dimethyl-Alkanes in Hot Spring Cyanobacterial Mats: a Direct Biogenic Source for Branched Alkanes in Ancient Sediments? 223-231.
- Skerman, V.B.D. et al (eds.). 1980. Approved Lists of Bacterial Names. *International Journal of Systematic Bacteriology*. 30:225-420.

- Stahl, David et al. 1985. Characterization of a Yellowstone Hot Spring Microbial Community by 5s rRNA Sequences. *Applied and Environmental Microbiology*. 49(6):1379-1384.
- Stanier, R.Y. et al. 1971. Purification and Properties of Unicellular Blue-Green Algae (Order Chroococcales). *Bacteriological Reviews*. 35(2):171-205.
- Stanier, Roger Y. et al. 1979. Generic Assignments, Strain Histories and Properties of Pure Cultures of Cyanobacteria. *Journal of General Microbiology*. 111:1-61.
- Stetter, K. O. et al. 1986. *Acidianus Infernus* Genus Novum, Species Novum, and *Acidianus Brierleyi* Comb. Nov.: Facultatively, Aerobic, Extremely Acidophilic Thermophilic Sulfur-Metabolizing Archaeobacteria. *International Journal of Systematic Bacteriology*. 35(4):559-564.
- Stetter, K. O. et al. 1993. Hyperthermophilic Archaea Are Thriving in Deep North Sea and Alaskan Oil Reservoirs. *Nature*. 365:743-745.
- Szybalski, Waclaw. 1992. Reasons and Risks to Study Restriction/modification Enzymes from Extreme Thermophile: Chilly Coldrooms, 13th Sample, and 3-Codon Overlap. *Gene*. 112:1-2.
- Theriot, Edward and Eugene Stoermer. 1984. Principal Component Analysis of *Stephanodiscus*: Observations on the *Stephanodiscus Niagarae* Complex. *Bacillaria*. 7:37-58.
- Theriot, Edward. 1987. Taxonomic Interpretation of Environmentally Related Variation in Silicification in *Stephanodiscus* (Bacillariophyceae). *British Phycological Society Journal*. 22:359-373.
- Theriot, Edward. 1992. Clusters, Species Concepts, and Morphological Evolution of Diatoms. *Systematic Biology*. 41(2):141-157.
- Ulrich, J.T. et al. 1972. Induction and Characterization of B-Galactosidase in an Extreme Thermophile. *Journal of Bacteriology*. 110(2):691-698.
- Ulrich, J.T. 1972. Effect of Temperature on the Respiration and Cytochromes of an Extreme Thermophile. *Journal of Bacteriology*. 110(2):777-779.
- Updegraff, D. et al. 1984. Thermophilic Cellulolytic Bacteria Isolated from Acidic Hot Springs at Yellowstone National Park, Wyoming. Solar Energy Research Institute. Biotechnology Branch. Golden, Colorado.
- Van Niel, C.B. and L.A. Thayer. 1929. Report on Preliminary Observations on the Microflora in and near the Thot Spring in Yellowstone National Park and Their Importance for the Geological Formations. Non-Circulating Book in Yellowstone National Park Research Library.
- Wahlund, Thomas M. and Michael Madigan. 1993. Nitrogen Fixation by the Thermophilic Green Sulfur Bacterium *Chlorobium Tepidum*. *Journal of Bacteriology*. 175(2):474-478.
- Wahlund, Thomas M. et al. 1991. A Thermophilic Green Sulfur Bacterium from New Zealand Hot Springs, *Chlorobium Tepidum* species novum. *Archives of Microbiology*. 156:81-90.

- Walter, William. 1952. The Identity of Aerobic Spore Forming Bacteria Isolated from Hot Springs. *Proceedings of the Montana Academy of Sciences*. 11:9-12.
- Walter, Malcolm R. 1975. Hot Spring Sediments in Yellowstone National Park. Elsevier Scientific Publishing Company, Amsterdam. Chapter 8.8:489-498.
- Walter, Malcolm R. 1993. Preservation of Biological Information in Thermal Spring Deposits: Developing a Strategy for the Search for Fossil Life on Mars. *Icarus* 101:129-143.
- Ward, David M. 1978. Thermophilic Methanogenesis in a Hot Spring Algal-Bacterial Mat (71 to 30°C). *Applied and Environmental Microbiology*. 35(6):1019-1026.
- Ward, David M. and Kenneth A. Sandbeck. 1981. Fate of Immediate Methane Precursors in Low-Sulfate Hot Spring Algal-Bacterial Mats. *Applied and Environment Microbiology*. 41(3):775-782.
- Ward, David M. 1982. Decomposition of Hot Spring Microbial Mats. *Proceedings of the Algal Mat Symposium*. Woodshale, Mass.
- Ward, David M. and Niels Peter Revsbech. 1983. Oxygen Microelectrode that is Insensitive to Medium Chemical Composition: Use in an Acid Microbial Mat Dominated by Cyanidium Caldarium. *Applied and Environmental Microbiology*. 45(3):755-759.
- Ward, David M. et al. 1985. Archaeobacterial Lipids in Hot-Spring Microbial Mats. *Nature*. 318(6047):656-659.
- Ward, David M. et al. 1987. Community Structure and Interactions Among Community Members in Hot Spring Cyanobacterial Mats. *Ecology of Microbial Communities*. Cambridge University Press. 179-210.
- Ward, David M. et al. 1987. Formation and Fate of Fermentation Products in Hot Spring Cyanobacterial Mats. *Applied and Environmental Microbiology*. 53(10):2343-2352.
- Ward, David M. et al. 1987. Use of Chloroflexus-Specific Antiserum to Evaluate Filamentous Bacteria of a Hot Spring Microbial Mat. *Applied and Environmental Microbiology*. 53(8):1962-1964.
- Ward, David M. et al. 1989. Hot Spring Microbial Mats: Anoxygenic and Oxygenic Mats of Possible Evolutionary Significance. *Microbial Mats Physiological Ecology of Benthic Microbial Communities*. Y. Cohen and E. Rosenberg (eds.). American Society for Microbiology, Washington, D.C. Chapter 1: 3-15.
- Ward, David M. 1989. Molecular Probes for Analysis of Microbial Communities. *Structure and Function of Biofilms*. W.G. Characklis and P.A. Wilderer (eds.). 145-163.
- Ward, David M. et al. 1989. Comparative Analysis of 16s Ribosomal RNA Sequences of Thermophilic Fermentative Bacteria Isolated from Hot Spring Cyanobacterial Mats. *Systemic Applied Microbiology*. 12:1-7.

- Ward, David M. and Roland Weller. 1989. Selective Recovery of 16s rRNA Sequences from Natural Microbial Communities in the Form of cDNA. *Applied and Environmental Microbiology*. 5(7):1818-1822.
- Ward, David M. et al. 1990. 16s rRNA Sequences Reveal Uncultured Inhabitants of a Well-Studied Thermal Community. *Microbiology Reviews*. 75:105-116.
- Ward, David M. et al. 1992. Ribosomal RNA Analysis of Microorganisms as They Occur in Nature. *Advances in Microbial Ecology*. 12:219-286.
- Ward, David M. and K.A. Sandbeck. 1981. Temperature Relations of Methanogenesis in Low Sulfate Hot Spring Algal-Bacterial Mats. Department of Microbiology, Montana State University, Bozeman.
- Ward, David M. 1981. Summary of Work Completed on Anaerobic Decomposition of Hot Spring Algal-Bacterial Mats. Department of Microbiology. Montana State University, Bozeman. 20.
- Ward, David M. et al. 1989. Lipid Biochemical Markers and the Composition of Microbial Mats. *American Society of Microbiology*. Washington, D.C. Chapter 38.
- Ward, David M. et al. 1990. 16s rRNA Sequences Reveal Numerous Uncultured Microorganisms in a Natural Community. Department of Microbiology. Montana State University, Bozeman.
- Ward, David D. et al. 1991. Ecology of Hot Spring Microbial Communities. *Investigators Annual Report*. Yellowstone National Park. 7.
- Ward, David M. et al. 1992. Modern Phototrophic Microbial Mats: Anoxygenic, Intermittently Oxygenic/Anoxygenic, Thermal, Eukaryotic, and Terrestrial in the Proterozoic Biosphere, a Multidisciplinary Study. Cambridge University Press. J.W. Schopf and C. Klem (eds). Chapter 6.9.
- Ward, David M. et al. 1992. Species Diversity in Hot Spring Microbial Mats as Revealed by Both Molecular and Enrichment Culture Approaches — Relationship Between Biodiversity and Community Structure. Department of Microbiology. Montana State University, Bozeman. 12.
- Weed, Walter H. 1889. The Diatom Marshes and Diatom Beds of Yellowstone National Park. *Botanical Gazette*. 14:117-120.
- Weed, Walter H. 1889. The Vegetation of Hot Springs. *American Naturalist*. 23:394-400.
- Weiss, Rick. 1991. Hot Prospect for New Gene Amplifier. *Science*. 254:1292-1293.
- Weller, Roland et al. 1991. 16s rRNA Sequences of Uncultivated Hot Spring Cyanobacterial Mat Inhabitants Retrieved as Randomly Primed cDNA. *Applied and Environmental Microbiology*. 57(4):1146-1151.
- White, Roberta et al. 1954. Heat Studies on a Thermophilic Bacteriophage. *Proceedings of the Society for Experimental Biology and Medicine*. 85:137-139.

- White, Roberta et al. 1955. Characteristics of a Thermophilic Bacteriophage. *Proceedings of the Society for Experimental Biology and Medicine*. 88:373-377.
- Wickstrom, Conrad E. 1980. Distribution and Physiological Determinants of Blue-Green Algal Nitrogen Fixation Along a Thermogradient. *Journal of Phycology*. 16:436-443.
- Wickstrom, Conrad E. and Richard G. Wiegert. 1980. Response of Thermal Algal-Bacterial Mat to Grazing by Brine Flies. *Microbial Ecology*. 6:303-315.
- Wickstrom, Conrad E. 1984. Depression of *Mastigocladus Laminosus* (Cyanophyta) Nitrogenase Activity under Normal Sunlight Intensities. *Journal of Phycology*. 20:137-141.
- Wickstrom, Conrad E. 1984. Discovery and Evidence of Nitrogen Fixation by Thermophilic Heterotrophs in Hot Springs. *Current Microbiology*. 10:275-280.
- Wiegel, Jurgen. 1979. Isolation from Soil and Properties of the Extreme Thermophilic *Clostridium Thermohydrosulfuricum*. *Journal of Bacteriology*. 139(3):800-810.
- Wiegel, Jurgen. 1981. *Thermoanaerobacter Ethanolicus* genus novum, species novum; a New Extreme Thermophilic, Anaerobic Bacterium. *Archives of Microbiology*. 128:343-348.
- Wiegel, Jurgen et al. 1981. *Clostridium Thermoautotrophicum* Species Novum, a Thermophile Producing Acetate from Molecular Hydrogen and Carbon Dioxide. *Current Microbiology*. 5:255-260.
- Wiegel, Juergen. 1988. Purification and Properties of Primary and Secondary Alcohol Dehydrogenases from *Thermoanaerobacter Ethanolicus*. *Applied and Environmental Microbiology*. 54(2):460-465.
- Wiegel, Jurgen. 1979. Ethanol as Fermentation Product of Extreme Thermophilic, Anaerobic Bacteria. *Symposium Technische Mikrobiologie*. Berlin.
- Wiegel, Jurgen. 1979. Isolation and Characterization of a New Extreme Thermophilic Anaerobic Bacterium. *Annual Meeting of the American Society for Microbiology*. Abstract 163. Los Angeles, California.
- Wiegert, Richard G. and Peter C. Fraleigh. 1975. A Model Explaining Successional Change in Standing Crop of Thermal Blue-Green Algae. *Ecology*. 56(3):656-664.
- Zarilla, K.A. and J.J. Perry. 1984. *Thermoleophilum Album* genus novum and species novum; a Bacterium Obligate for Thermophily and N-Alkane Substrates. *Archives of Microbiology*. 137:286-290.
- Zarilla, K.A. and J.J. Perry. 1986. Deoxyribonucleic Acid Homology and Other Comparisons among Obligately Thermophilic Hydrocarbonoclastic Bacteria with a Proposal for *Thermoleophilum Minutum* species novum. *International Journal of Systematic Bacteriology*. 36(1):13-16.
- Zedan, Hamdallah. 1992. The Economic Value of Microbial Diversity. *Biodiversity and Biotechnology*. 43(5):178-185.

- Zeikus, J. Gregory. 1977. The Biology of Methanogenic Bacteria. *Bacteriological Reviews*. 41(2):514-541.
- Zeikus, J. Gregory et al. 1979. *Thermoanaerobium Brockii* Genus Novum and Species Novum: A New Chemoorgananothrophic, Caldoactive, Anaerobic Bacterium. *Archives of Microbiology*. 122:41-48.
- Zeikus, J. Gregory. 1979. Thermophilic Bacteria: Ecology, Physiology and Technology. *Enzyme Microbiology Technology*. 1:243-252.
- Zeikus, J. Gregory et al. 1980. Microbiology of Methanogenesis in Thermal, Volcanic Environments. *Journal of Bacteriology*. 143:432-440.
- Zeikus, J. Gregory and Arie Ben-Bassat. 1981. *Thermobacteroides Acetoethylicus* genus novum and species novum; a New Chemoorgananothrophic, Anaerobic, Thermophilic Bacterium. *Archives of Microbiology*. 128:365-370.
- Zeikus, J. Gregory. 1983. Metabolic Communication Between Biodegrative Populations in Nature. Department of Bacteriology, University of Wisconsin, Madison.
- Zeikus, J. Gregory. 1983. Microbial Ecology of Volcanic Sulphidogenesis: Isolation and Characterization of *Thermodesulfobacterium Commune* genus novum and species novum. *Journal of General Microbiology*. 129:1159-1169.
- Zeikus, J. Gregory and H.H. Hyun. 1985. Simultaneous and Enhanced Production of Thermostable Amylases and Ethanol from Starch by Cocultures of *Clostridium Thermosulfurogenes* and *Clostridium Thermohydrosulfuricum*. *Applied and Environmental Microbiology*. 49(5):1174-1181.
- Zeikus, J. Gregory and H.H. Hyun. 1985. General Biochemical Characterization of Thermostable Pullulanase and Glucoamylase from *Clostridium Thermohydrosulfuricum*. *Applied and Environmental Microbiology*. 49(5):1168-1173.
- Zeikus, J. Gregory and H.H. Hyun. 1985. Regulation and Genetic Enhancement of β -Amylase Production in *Clostridium Thermosulfurogenes*. *Journal of Bacteriology*. 164(3):1162-1170.
- Zeikus, J. Gregory et al. 1985. Differential Amylosaccharide Metabolism of *Clostridium Thermosulfurogenes* and *Clostridium Thermohydrosulfuricum*. *Journal of Bacteriology*. 164(3):1153-1161.
- Zeikus, J. Gregory et al. 1990. Physiological and Enzymatic Characterization of a Novel Pullulan-Degrading Thermophilic *Bacillus* Strain 3183. *Appl Microbiol Biotechnol*. 33:340-344.
- Zeikus, J. Gregory et al. 1993. Regulation and Characterization of Xylanolytic Enzymes of *Thermoanaerobacterium Saccharolyticum* B6A-RI. *Applied and Environmental Microbiology*. 59(3):763-771.

- Zeikus, J. Gregory et al. 1993. Taxonomic Distinction of Saccharolytic Thermophilic Anaerobes: Description of *Thermoanaerobacterium Xylanolyticum* genus novum, species novum, and *Thermoanaerobacterium Saccharolyticum* genus novum, species novum; Reclassification of *Thermoanaerobium Brockii*, *Clostridium Thermosulfurogenes*, and *Clostridium Thermohydrosulfuricum* E100-69 as *Thermoanaerobacter Brockii* comb novum, *Thermoanaerobacterium Thermosulfurigenes* comb. novum, and *Thermoanaerobacter Thermohydrosulfuricus* comb. novum, Respectively; and Transfer of *Clostridium Thermohydrosulfuricum* 39E to *Thermoanaerobacter Ethanolicus*. *International Journal of Systematic Bacteriology*. 43(1):41-51.
- Zeikus, J. Gregory et al. 1993. Cloning, Sequencing and Biochemical Characterization of Xylose Isomerase from *Thermoanaerobacterium Saccharolyticum* Strain B6A-RI. *Journal of General Microbiology*. 139:1227-1234.
- Zeikus, J. Gregory et al. 1993. Biology, Ecology, and Biotechnological Applications of Anaerobic Bacteria Adapted to Environmental Stresses in Temperature, pH, Salinity, or Substrates. *Microbiological Reviews*. 57(2):451-509.
- Zeng, Yi Dong B. 1992. Biogeochemistry of Hot Spring Environments 3. Apolar and Polar Lipids in the Biologically Active Layers of a Cyanobacterial Mat. *Chemical Geology*. 95:347-360.