FRUITS OF *KOELREUTERIA* (SAPINDACEAE) FROM THE CENOZOIC THROUGHOUT THE NORTHERN HEMISPHERE: THEIR ECOLOGICAL, EVOLUTIONARY, AND BIOGEOGRAPHIC IMPLICATIONS¹

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- *Premise of the study: Koelreuteria* (Sapindaceae) has four extant deciduous tree species, disjunctly distributed in eastern Asia and the Fiji Islands. While *K. paniculata* is widely cultivated, the biogeographic origin and evolutionary history of *Koelreuteria* remain unclear.
- *Methods:* Fruits, pollen, wood, and leaves of closely related extant taxa were examined in comparison with fossil remains to evaluate the fossil record and biogeographic history of *Koelreuteria*.
- Key results: Overall, characters of capsular fruits are more diagnostic than other organs for this genus. We describe two new species of fruit remains from the Eocene, K. taoana sp. nov. (northeastern China and far eastern Russia) and K. dilcheri sp. nov. (western United States), and give emended descriptions of three species: K. allenii (Lesq.) W. N. Edwards (early–late Eocene of the United States), K. macroptera (Kováts) W. N. Edwards (late Oligocene–early Pliocene of Europe), and K. miointegrifoliola Hu et R. W. Chaney (Miocene of eastern Asia).
- Conclusions: Reliable fossil records of capsules and ring-porous wood indicate that Koelreuteria may have originated in North
 Pacific-Rim area of the northern hemisphere by the early Eocene, representing an early temperate lineage in Sapindaceae
 adapted for wind dispersal. The fossils herein place a minimum age (ca. 52 Ma) for the divergence of Koelreuteria from tropical genera that appear more basal in the molecular phylogeny of Sapindaceae. Regional extinctions after the Eocene in North
 America and the Pliocene in Europe, reduced the range of Koelreuteria to eastern Asia, where three species occur today. The
 present distribution of another species in the Southern Pacific may be explained by long-distance dispersal.

Key words: biogeography; Cenozoic; ecology; Eocene; evolution; fruits; Koelreuteria; Miocene; paleobotany; Sapindaceae.

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Koelreuteria Laxm. of the soapberry family (Sapindaceae Juss.) comprises four extant deciduous tree species, disjunctly distributed in eastern Asia (China, Korea, and Japan) and on the Fiji Islands in the South Pacific (Smith, 1985; Wu et al., 2007; Acevedo-Rodríguez et al., 2011; Ying and Chen, 2011) (Table 1). In eastern Asia, distributions of K. paniculata Laxm. to the north and K. bipinnata Franch. to the south converge at approximately 32° north latitude (Xia and Luo, 1995). In the Pacific, two species K. elegans A. C. Sm. (Smith, 1951, 1952, 1955, 1985) and K. henryi Dümmer (= K. formosana Hayata) (Dümmer, 1912; Hayata, 1913; Kanehira, 1936; Li, 1963, 1977; Chen, 1993; Ying and Chen, 2011) are distributed in Taiwan (China) and Fiji, respectively, with a discontinuity of ca. 7000 km. Alternatively, they are treated as two subspecies, K. elegans subsp. formosana (Hayata) F. G. Mey. and K. elegans subsp. elegans A. C. Sm. (Meyer, 1976; Luo and Chen, 1985; Xia and Luo, 1995; Xia and Gadek, 2007). Considering their morphological variation (e.g., pollen and petiolules, see Table 1) and longdistance disjunct distributions, we here did not adopt such subspecific ranks of K. elegans.

Koelreuteria has been used as an official memorial tree cultivated on the tombs of scholars since the Chou Dynasty (1122–240 BC) and was introduced into Europe in 1763 (Li, 1996; Dosmann et al., 2006). Based upon the cultivated tree at St. Petersburg, Finnish naturalist Erik Laxmann (1772) named *Koelreuteria* after a German botanist Joseph Gottlieb Kölreuter (1733–1806) at Karlsruhe, with the type species *K. paniculata*

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TABLE 1.	

		Polle	n			Cap	sule			Leaf			
Taxa	Form	Size	Exine	Ektexine	Form	Valve form	Valve size	Valve Septum	Form	Leaflet margin	Leaflet apex	Petiolule	Distribution and ecology
K. paniculata Laxm. ^{1, 8-9, 15-16}	Subprolate to spheroidal	24–28.2 × 21–33.8 µm	1.5-2 µm thick	More or less long itudinally striate	Trigonous- conoidal	Ovate, tapering to apex, acute	2.2-7.2 × 2-4 cm	1/3 of valve length	Pinnate, rarely bipinnate	Coarsely crenate-serrate to pinnatisect and lobulate	Abruptly to narrowly acuminate, often apiculate	l-2 mm long long	Vorth, Northeast, East, and Southwest China ^{8, 11, 15} ; Korea and Japan ^{12, 14} ; (0)–300–3,800 m. elevations in river courses, open forest areas, rocky mountain slopes,
K. bipinnata Franch. (= K. integrifoliola Merr.) ^{4, 8-9, 15-16}	Oblate- spheroidal	22–28.2 × 23–31 µm	2–2.4 µm	Finely striate	Ellipsoidal, ovoidal- globose	Elliptic to rotund, apex short acuminate, mucronate	3.7–6.6× 3–5 cm	Over 1/3 to 1/2 of valve length, or occasionally complete when suture being straigh	Bipinnate	Entire or uniformly serrate, lower leaflets never pinnatisect	Acute to short acuminate	3 mm 3 mm long	outh and Southwest China ^{8, 11, 15} ; (100)–250– (1,500)–2,600 m. elevations in open fields, hillside forests, light woods, thickets, and
K. henryi Dümmer [= K. formosana Hayata; K. elegans subsp. formosana (Hayata) F. G. Mey.] 2.3.5.7.8.10. 13.15-16	Mostly spheroidal	20–23 × 21–23 μm	1.5 µm	Striate	Subglobose	Suborbicular, apex short acuminate, mucronate	4-5 × 3-4.5 cm	valve length	Bipinnate	Entire or irregularly crenate-serrate, lower leaflets sometimes pinnatisect	Long acuminate, sometimes caudate	long	aroustores at low elevations (less than 1,000 m.) in broad-leaved forests and light woods
K. elegans (Seem.) A. C. Sm. $[= K.$ elegans subsp. elegans A. C. Sm.] 6.8,15-16	Mostly spheroidal	24-26 × 24-26 μm	1.5 µm	Irregularly striate- reticulate	Subglobose	Suborbicular, apex short acuminate, mucronate	3.4-5 × 3.1-4.6 cm	valve valve length	Bipinnate	Entire or irregularly crenate-serrate sparsely, lower leaflets sometimes pinnatisect	Long acuminate, sometimes caudate	4–5 mm 7 long	/anua Levu and Viti Levu of Fiji; 50–(350)– 825–(900) m. 825–(900) m. elevations in dense or open forest or on its edges
References: 1 5	1772 TX	² Dimmer 1	1912: ³ Hav	ata, 1913; ⁴	Merrill, 192	2.5 Kanehira	1936: ⁶ Smit	h 1952 1985	71 i 1963	1977. ⁸ Mever 197	76: ⁹ Wang and	Chien 19	56 ⁻¹⁰ Chen 1993:

reletences: Laxmann, 1772; Dummer, 1912; Hayata, 1912; Merrul, 1922; Merrul, 1922, 1903; LJ, 1903, 1977; Meyer, 1976; Wang and Cmen, 1996; Cnen, 1995; LJ Xia and Luo, 1995; ¹² Dosmann et al., 2006; ¹³ Luo and Chen, 1985; ¹⁴ Sasaki and Ohashi, 2007; ¹⁵ Xia and Gadek, 2007; ¹⁶ Direct observations on the cultivated plants and the herbaria kept at the Chinese National Herbarium (PE), the Missouri Botanical Garden Herbarium (MO), and Amold Arboretum Herbarium, Harvard University (A).

being better known as the golden-rain tree. *Koelreuteria henryi*, native in Taiwan, was introduced to the United States in 1915 and Australia around 2000, and subsequently became naturalized in southern Florida and Hawaii in the United States (Meyer, 1976; Frohlich and Lau, 2010) and in New South Wales (Hosking et al., 2003). The wood of *Koelreuteria* can be used for making furniture. *Koelreuteria* is popularly cultivated as an ornamental around the world because of the aesthetic appeal of its spreading canopy, imparipinnate and bipinnate leaves, large yellow thyrses, and colorful, membranous, inflated capsules. However, the early evolution and biogeographic spread of *Koelreuteria* through the geological ages remain unclear.

Koelreuteria, along with Stocksia Benth. in Afghanistan, Iran, and Pakistan (Abdulia, 1973) and Erythrophysa E. Mey. ex Arn. from Ethiopia, South Africa, and Madagascar (Verdcourt, 1962; Capuron, 1969; Palmer and Pitman, 1972), were traditionally classified into the tribe Koelreuterieae of the subfamily Dodonaeoideae in Sapindaceae on the basis of their imparipinnate leaves, convolute cotyledons, zygomorphic flowers and inflated capsules (Radlkofer, 1931) (Figs. 1-7, 9-12), and simply structured tricolporate pollen (i.e., type-A pollen, sensu Muller and Leenhouts, 1976) (Fig. 8). Koelreuterieae was thought to be derived from the most basal sapindaceous tribe Dodonaeeae, represented by Dodonaea L. mainly from Australia and New Caledonia (Xia and Gadek, 2007; Harrington, 2008; Acevedo-Rodríguez et al., 2011). Recent molecular phylogenetic studies have suggested that Koelreuteria is closely related to some subtropical and tropical genera from Asia, Australasia, and Africa, e.g., Stadmania Lam. (fruits not membranous), Allophylus L. (fruits not capsular), Smelophyllum Radlk. (fruits chartaceous or fleshy), Cupaniopsis Radlk. (fruits fleshy), and Harpullia Roxb. (fruits chartaceous to woody) (Harrington et al., 2005; Muellner et al., 2007; Buerki et al., 2009, 2010). Hence, the evolutionary origin and phylogenetic relationships of Koelreuteria are likely to be rooted in subtropical and tropical lineages. The fossil record of Koelreuteria may provide temporal and paleogeographic data relevant to the evolutionary and biogeographic history of these genera.

The main objectives of this paper are to (1) review the morphology and taxonomy of living *Koelreuteria* with particular reference to fruit morphology and some subsidiary references to pollen, leaf, and wood anatomical characters, all of which can be used to identify the fossil remains of the genus; (2) reinvestigate the previously reported fossil species in *Koelreuteria* and provide a reliable fruit fossil record of the genus; and (3) consider their ecological, evolutionary, and biogeographic implications.

MATERIALS AND METHODS

Nomenclature—Fossils of *Koelreuteria* are preserved as isolated parts (e.g., dispersed leafy shoots, leaves, leaflets, wood, capsules, capsular valves, seeds, and pollen). Some of the previously reported fossil species were established on the basis of the hypothesized combination of both leaves and capsular valves occurring together at the same localities, but there was no direct proof in the form of organic connections (e.g., Massalongo, 1852; Hu and Chaney, 1938; Brown, 1946; Becker, 1961). Such taxonomic practice may be problematic because two or more unrelated elements were prone to be involved in the original description. On the basis of reliable fruit remains, we here reconsider the nomenclature and taxonomy of fossil *Koelreuteria*.

Macrofossils—The macrofossils used in this study (see Appendix 1) were collected from the following localities in approximate order according to geological age. (1) Green River Formation, USA. Capsules from the Green River Formation of Colorado, Utah, and Wyoming are deposited at USNM (National Museum

of Natural History, Smithsonian Institution, Washington, D.C., USA), UCM (University of Colorado Museum of Natural History, Boulder, CO, USA), UCMP (University of California Museum of Paleontology, Berkeley, CA, USA), FMNH (Field Museum, Chicago, IL, USA), and YPM (Peabody Museum of Natural History, Yale University, New Haven, CT, USA). The geological age of these Green River Formation localities is early Eocene (ca. 52 Ma) to middle Eocene on the basis of radiometric dating as well as mammalian biostratigraphy (Smith et al., 2008). (2) Republic, Washington, USA. A few capsular valves were collected from Republic, Washington, northwestern USA. The specimens (prefixed "UWBM") are deposited at the Burke Museum of Natural History, the University of Washington, Seattle. The age of the Republic flora is early-middle Eocene (Wolfe et al., 1998; Greenwood et al., 2005; DeVore and Pigg, 2010). (3) Independence Hill, California, USA. One capsular valve was collected from Independence Hill near Colfax, California and deposited at USNM. The geological age of the Chalk Bluffs flora of Independence Hill is early-middle Eocene (Brown, 1946; Edwards, 2004). (4) Huadian, Jilin Province, China. One capsular valve (part and counterpart) was collected from siltstone beds of the Huadian Formation at the Gonglangtou Mine No. 3, Huadian County, Jilin Province in northeastern China (42°59'54"N, 126°51'58"E). The specimens (prefixed "PE") are deposited at the Chinese National Herbarium, Institute of Botany, Chinese Academy of Sciences, Beijing. The geological age of the Huadian flora has been accepted as middle Eocene on the basis of co-occurring vertebrate fossils (Smith et al., 2011). (5) Teater Road, Oregon, USA. Capsular valves were collected from the John Day Formation of Teater Road, Oregon and deposited at UF (Florida Museum of Natural History, Gainesville). The geological age of this John Day Formation locality is late Eocene (Manchester and McIntosh, 2007). (6) Florissant, Colorado, USA. Numerous capsular valves were collected from the Florissant lake beds of Florissant, Colorado. The specimens are deposited at YPM and USNM. The geological age of the Florissant flora is latest Eocene on the basis of radiometric dating and mammal correlations (Meyer and Smith, 2008). (7) Randecker Maar, Germany. Ten capsular valves were observed from Randecker Maar in collections at the Staatliche Museum für Naturkunde (Stuttgart) (specimens prefixed "SMNS") and the Nature Museum Augsburg (prefixed NMA). The age of the Randecker Maar flora is early Miocene (Gregor, 1982; Krautter and Schweigert, 1991). (8) Shanwang, Shandong Province, China, Twelve capsular ovules were collected from Shanwang (36°54'N, 118°20'E) at Lingu County, Shandong Province, eastern China. The specimens are deposited at UF, PE, and Nanjing Institute of Geology and Palaeontology, Chinese Academy of Sciences, Nanjing (prefixed "PB"). The geological age for plant-bearing diatomaceous shales of the Shanwang Formation is Middle Miocene (Li, 1981; Tao et al., 1999; Wang et al., 2010) or late Early Miocene to early Middle Miocene (Liu and Leopold, 1992; Deng, 2006; Wang, 2012). (9) Tállya, Hungary. One capsular valve was examined from Gomboska of Tállya, Hungary. The specimen (prefixed "BP") is deposited at the Hungarian Natural History Museum, Budapest. The age of the Tállya flora is Middle Miocene (Hably, 2006). (10) Abaújszántó, Hungary. One capsular valve was collected from Abaújszántó, Hungary, which is deposited at BP. The age of the Abaújszántó flora is Middle Miocene (Hably, 2006). (11) Bavaria, Germany. Capsular valves were collected from Derching, Burtenbach, Derndorf, and Geisenhausen, sediments of the Upper Freshwater Molasse of Bavaria, southern Germany. The specimens are deposited at NMA. The ages of these Bavarian floras are from the Lower Middle Miocene of Burtenbach to the middle part of the Middle Miocene of Derching as well as to the Middle-Upper Miocene of Geisenhausen and Derndorf (Gregor, 1982; Gregor et al., 1992; Seehuber, 2009).

Herbaria—The extant voucher specimens examined for this study (see Appendix 1) are housed at the following herbaria: Chinese National Herbarium (PE), Missouri Botanical Garden Herbarium (MO), and Arnold Arboretum Herbarium, Harvard University (A). Capsules and leaves of *K. paniculata*, *K. bipinnata* and *K. henryi* cultivated at the Institute of Botany, Chinese Academy of Sciences (Beijing), Yunnan Academy of Forestry (Kunming), and the University of Florida (Gainesville) were collected to compare with the fossils presented here.

Cleared leaflets and capsular valves of *Koelreuteria* were made by soaking fresh samples in 40% sodium hydroxide (NaOH) solution for about 3 d, then neutralizing in running water, and mounting with Canada balsam in glass slides. Data regarding fossil and extant capsules, pollen, leaflets, and wood of *Koelreuteria* were tabulated (Tables 1, 2), and comparisons of *Koelreuteria* with other taxa bearing similar fruits were made (Table 3). Terminology used in the specimen descriptions for seeds, pollen, fruits, and leaves follows Corner (1976), Müller and Leenhouts (1976), and Ellis et al. (2009). The author citation of plant names is applied by Brummitt and Powell (1992).

Online databases were consulted including those of (1) National Museum of Natural History, Smithsonian Institution, Washington, D.C. (http://collections. nmnh.si.edu/search/paleo/), (2) University of California Museum of Paleontology,

Berkeley (http://bscit.berkeley.edu/cgi/ucmp/), (3) Peabody Museum of Natural History, Yale University, New Haven (http://peabody.yale.edu/collections/ paleobotany), (4) Museum of Natural History, the University of Colorado, Boulder (http://cumuseum.colorado.edu/Exhibits/StoneLace/specimens2.html), (5) Burke Museum of Natural History and Culture, the University of Washington (http://www.burkemuseum.org/paleontology/collections), (6) The Staatliche Museum für Naturkunde (Stuttgart) (http://www.dbsmns.naturkundemuseum-bw.de/), (7) The Inside Wood (http://insidewood.lib.ncsu.edu/search), and (8) eFloras (http://www.efloras.org).

Figures—Photographs of hand specimens were taken with digital cameras (Panasonic DMC-FZ30, Nikon D90, CoolPix S230, and Canon Power-Shot A630). Images of capsular valve and leaflet clearings were made on a flatbed scanner (Epson Perfection 2580 Photo). Micrographs of pollen grains were taken with a Scanning Electron Microscope (SEM) (Hitachi S-4800). Line drawings of previously published figures of fossil *Koelreuteria* were drawn when the original specimens were not available for photography. The International Stratigraphic Chart (Gradstein et al., 2004) was used for time calibration. Figures were prepared with Adobe Photoshop 6.0 (San Jose, California, USA) and CoreIDRAW 10.0 (Corel, Ottawa, Ontario, Canada) programs.

RESULTS

The taxonomy of living *Koelreuteria* is primarily based upon growth habit, vegetative shoot, leaf, inflorescence, flower, fruit, seed and seedling, pollen, and chromosome characters (Meyer, 1976; Xia and Gadek, 2007) (Table 1), but paleobotanists usually have to identify *Koelreuteria* fossils in the light of dispersed leaves, leaflets, wood, capsules, capsular valves, seeds, and pollen (Table 2).

Fruits of extant Koelreuteria—Fruits of Koelreuteria are three-valved, loculicidal capsules, which open completely along the three dorsal sutures; each valve consists of two halves of adjoining carpels (Figs. 1-7). Overall, the capsules of Koelreuteria are trigonous-conoidal, ellipsoidal, ovoidal-globose or subglobose and measure 2.2-7.2 cm long by 2-5 cm wide (Table 1); the carpels are membranous and bladder-like, and each valve bears two seeds oppositely or suboppositely along the two sides of the lower or middle part of the placental suture (Fig. 2); the seed is globose, about 5-9 mm in diameter, with sclerenchymatous testa, and contains a large embryo, with two coiled or convoluted cotyledons and a radicle that protrudes into a pocket of the seed coat (Corner, 1976); the placentation (or axile partition) above the suture where seeds are attached is usually incomplete, or occasionally complete when the placental suture is straight, forming a septum accounting for 1/3 to all of the valve length from the base (Figs. 5, 6). The distally incomplete septum is developed from the septal slits occurring within the style (Ronse Decraene et al., 2000). Although K. bipinnata occasionally has a nearly complete septum in capsules, the distal, slim septal slits above the seed-bearing placental suture are still visible.

The capsules of *Koelreuteria* can be easily subdivided into two types: *K. paniculata*-type (Fig. 1) and *K. bipinnata*-type (Figs. 3, 4). *Koelreuteria paniculata*-type represented only by the type-species *K. paniculata* bears ovate capsular valves with an acute apex and a basal septum approximately 1/3 of the valve length (Figs. 2, 5). In contrast, *K. bipinnata*-type, including the other three species *K. bipinnata*, *K. henryi*, and *K. elegans*, typically possesses elliptical, ovate-elliptical or suborbicular capsular valves with an obtuse apex and a longer septum accounting for more than 1/3 to all of the valve length. Overall, the capsular valves of *K. bipinnata* are the most variable, ranging from elliptic, ovate-elliptic to suborbicular.

Comparisons have been made between *Koelreuteria* and other genera of the Sapindaceae including Arfeuillea Pierre ex Radlk., Boniodendron Gagnep. (= Sinoradlkofera F. G. Mey.), Stocksia Benth., Erythrophysa E. Mey. ex Arn. (Meyer, 1977; Bůžek et al., 1989; Manchester, 1999; Buijsen et al., 2003; Manchester and O'Leary, 2010). Koelreuteria fruits also superficially resemble genera with membranous or papery fruits in several other families, including tilioid Malvaceae Juss. [Craigia A. C. Sm. et W. E. Evans (= Pteleaecarpum Weyland)], Rutaceae Juss. (Ptelea L.), and the Nyctaginaceae Juss. (Abronia Juss.) (Kvaček et al., 1991, 2005; Manchester and O'Leary, 2010) (Table 3). However, Koelreuteria is distinguished from these other genera by several characters: incomplete septum with two seeds at a single level within each ovate, elliptic to suborbicular, polygonally elongated reticulate, membranous valve that is greater than 2 cm long (ca. 4 cm long on average). Occasionally, a nearly complete septum occurs in K. bipinnata when the placental suture appears straight, in which case the distinction of Koelreuteria from other genera may be less secure.

The morphology of capsular valves is helpful in distinguishing taxa. Some small-sized (around 2 cm long) valve fossils with a "central body" (i.e., locular area) formerly attributed to Koelreuteria belong to either a living tilioid malvaceous genus Craigia or probably another extant sapindaceous genus Boniodendron (Kvaček et al., 1991, 2005; Manchester et al., 2009) (Table 2). These misidentified species should be excluded from the fossil record of Koelreuteria, including K. eointegrifolia S. Endô, K. eocenica S. Endô, K. oregonensis C. A. Arnold, and K. microcarpa H. M. Li (Endô, 1942, 1968; Arnold, 1952; WGCPC, 1978; Zhang, 1980). Fossils belonging to Craigia usually have serial seed attachments and/or complete septa within the valves, forming a fusiform or elliptical central body. The small capsular valves called Koelreuteria arnoldii H. F. Becker from the early middle Eocene to Oligocene of the United States and Canada (Becker, 1961, 1969; Wolfe and Wehr, 1987; Dillhoff et al., 2005) may represent Boniodendron, whose capsules are about half the size of those in living Koelreuteria (Manchester et al., 2009) (Table 3). Furthermore, pedicels of Boniodendron, when preserved at the base of such smaller valves, have no persistent sepals (Table 3); K. annosa Brown was described for the capsular valve and leaflet from the Paleocene of the United States (Brown, 1956, 1962), but its valve appears to bear an elongated, central locular area and a long pedicel. Numerous specimens of this species in the UF collections all lack seeds, so the placentation remains unclear. Hence, this species is also rejected from *Koelreuteria*.

Although the trilocular capsules with incomplete septa are also produced sporadically in two other living sapindaceous species *Euphorianthus euneurus* (Miq.) Leenh. (velvety and fleshy capsules), possibly *Erythrophysa paniculata* Capuron (pilose and membranous-papery capsules) and the fossil species *Sapindaceacarpum koelreuterioides* Vaudois-Miéja et Lejal-Nicol (glabrous and membranous capsules), their capsular valves are not reticulate (Capuron, 1969; Vaudois-Miéja and Lejal-Nicol, 1988; Acevedo-Rodríguez et al., 2011) (Table 3). Hence, wellpreserved capsular valves are easily recognizable in the fossil record and can provide a reliable proxy for deciphering the biogeographic history of *Koelreuteria*.

Pollen, leaves, and woods of Koelreuteria—Pollen grains of extant *Koelreuteria* are primarily spheroidal, oblate-spheroidal or rarely prolate-spheroidal, $20-28.2 \times 21-33.8 \mu m$ in size (Table 1). The polar amb is triangular and the grains are tricolporate.

TABLE 2. Previously desc	cribed fossils assignable or sim	ilar to <i>Koelreuteria</i> in a	pproximately stratigraphic order.			
Published name	Locality	Organ	Cited figure	Geological age	Age reference	Identification
Talisipites megorites Z. C. Song et M. Y. Li	Gongjue County of Changdou, Qiabulin and Cuojjangding of Zhongba County, Xizang (Tibet), China	Pollen (simply structured tricolporate pollen, i.e., type-A pollen)	Song and Li, 1982, p. 20, pl. 4, figs. 8, 9; Song et al., 1999, p. 591, pl. 149, figs. 22, 23, 27; Li et al., 2008, p. 297, fig. 3. G, fig. 4, H	Santonian- Maastrichtian, Late Cretaceous to the Eocene	Song et al., 1999, 2004; Li et al., 2008	Similar to living <i>K. bipimata</i> Franch.
Senipites striatus Z. C. Song	Subei Basin, Taizhou Formation, Jianssu Province, China	Pollen (type-A)	Song et al., 1999, p. 590, pl. 152, figs. 9–12, 19	Senonian (Coniacian– Maastrichtian), Late Cretaceous	Song et al., 1999, 2004	Similar to living <i>K. paniculata</i> Laxm. and <i>K. binimata</i>
K. prenigricans W.A. Bell	Vancouver Island, British Columbia. Canada	Leaves and leaflets	Bell, 1957, p. 61, pl. 51, fig. 1, nl 14 fig. 2, nl 15 figs 1, 4, 7	Late Cretaceous	Bell, 1957	Doubtful
K. annosa R. W. Br.	Little Powder River, near Biddle, Montana and Point of Rocks, Wyoming, USA	Capsular valve $(2.7 \times 1.8 \text{ cm}, \text{elliptic})$ and leaflets	Brown, 1956, p. 107, fig. 3; Brown, 1962, p. 76, pl. 59, figs. 8, 9	Paleocene	Brown, 1962	Capsules bear a complete septum, so it was rejected from <i>Koelreuteria</i> herein
K. eakinii Hollick Palaeoallophylus	Yukon River, Alaska, USA Sheppey, southern London,	Leaf Seeds (11–13 mm in diamatar	Hollick, 1936, p. 137, pl. 81, fig. 1 Reid and Chandler, 1933,	Paleocene Early Eocene	Wolfe, 1972 Collinson and	Doubtful Showing the pattern of
roumaans E. Kelu et M. Chandler	4	in danneter, embryo with large coiled cotyledons; globose)	p. 902, pl. 17, ilg. 0-12, text fig. 9d		Clear, 2001	convoluted conjuedons, tongue-like radicle, and thin seed coat characteristic of seeds of living Koelreuteria and Allophytus L.
K viridifluminis (Hollick) R. W. Br.	Green River, Colorado, Wyoming and Utah, USA	Capsular valve (4.2 × 2.1 cm, obovoid-elliptic) and leaflets	Lesquereux, 1878b, p. 132, pl. 17, figs. 9–12; Brown, 1934, p. 62, pl. 10, fig. 10; Brown, 1943, p. 863, fig. 1N; MacGinitie, 1969, p. 120, pl. 21, fig. 3	Early to middle Eocene	Smith et al., 2008	Reliable for the capsular valves, which were reclassified as <i>K. allenii</i> herein; some of the leaflets doubtful
K. mixta (Lesq.) R. W. Br.	You Bet and Independence Hill, California;Republic, Washington, USA	Capsular valve (3.5–5.8×2.8–3.6 cm, elliptic, oblong or oblong-elliptic) and leaflet	Lesquereux, 1878a, p. 30, pl. 9, fig. 13; Brown, 1946, p. 351, figs. 1, 2; this paper, Figs. 17–21	Early to middle Bocene	Brown, 1946; Wolfe et al., 1998; Edwards, 2004; Greenwood et al., 2005; DeVore and Pigg, 2010	Reliable for the capsular valve, which was reclassified as <i>K. dilcheri</i> sp. nov. in this paper; leaflets doubtful
Koelreuteria sp.	Gonglangtou Mine No. 3, Huadian, Jilin Province, China	Capsular valves $(2.5 \times 1.6 \text{ cm}, \text{elliptic})$	Manchester et al., 2005, p. 10, pl. 4, figs. 8–10; this paper, Figs. 13–15	Middle Eocene	Smith et al., 2011	Reliable; reclassified as <i>K. taoana</i> sp. nov. herein
Koelreuteria sp. or Koelreuteria cf. miointegrifolia Hu et R. W. Chanev	Uglovsky (Uglovskaya) Basin, South Primorye, Russia	Capsular valve $(2.4 \times 1.7 \text{ cm}, \text{ovoid-elliptic})$	Ablaev, 1978, p. 170, pl. 10, fig. 5; Ablaev, 2000, p. 53, pl. 15, figs. 6, 7; this paper, Fig. 16	Middle Eocene	Pavlyutkin and Petrenko, 2010	Reliable; reclassified as <i>K. taoana</i> sp. nov. in this paper
Palaeoallophylus globosa Manchester	Clarno, north-central Oregon, USA	Seeds (3.6-6.0)× 3.7-5.2 mm, globose)	Manchester, 1994, p. 86, pl. 38, figs. 9–10	Middle Eocene	Manchester, 1994	Similar to seeds of <i>Koelreuteria</i> and <i>Allophylus</i> ; smaller than seeds of <i>Palaeoallophylus</i> <i>rotundatus</i> E. Reid et M. Chandler from early Eocene London Clay flora
K. kamtschatica Budantsev	Kingy Cape and Napana River, western Kamchatka, far eastern Russia	Leaf and leaflet	Budantsev, 1997, p. 86, pl. 54, figs. 1, 2; Budantsev, 2006, p. 118, pl. 133, fig. 5	Middle to late Eocene	Budantsev, 1997, 2006	Doubtful; superficially resembling the leaves of living K. paniculata
Cf. Koelreuteria sp. Koelreuteria sp.	Florissant, Colorado, USA Florissant, Colorado, USA	Wood (ring porous) Pollen (type-A)	Wheeler, 2001, p. 193, pl. 3, A-F Leopold and Clay-Poole, 2001, p. 35, pl. 7, fig. 1; Leopold et al., 2008, p. 56, tab.1	Late Eocene Late Eocene	Meyer and Smith, 2008 Meyer and Smith, 2008	Reliable

TABLE 2. Continued.						
Published name	Locality	Organ	Cited figure	Geological age	Age reference	Identification
K. allenii (Lesq.) W. N. Edwards	Florissant, Colorado, USA	Leaves, leaflets, and capsular valves (2.3–3.8 × 1.6–2.2 cm, obovate, obovoid-elliptic)	Lesquereux, 1878b, p. 65, pl. 5, fig. 11; Hollick, 1923, p. 207, pl. 10, figs. 1, 3, 6–11, pl. 11, figs. 1–3; Hollick, 1929, pl. 2, fig. 3; MacGinite, 1953, p. 145, pl. 55, figs. 1–4, 7, pl. 75, fig. 1; this namer Fiss 27–29	Late Eocene	Meyer and Smith, 2008	Reliable for capsular valves, which were classified into <i>K. allenii</i> herein; resembling those of living <i>K. paniculata</i>
K. eocenica S. Endô	Ishikari Coal-field, Hokkaido, Japan	Capsular valves (2.4 × 1.2 cm, elliptic, with a central body)	Endő, 1968, p. 437, pl. 14, figs. 4–6, pl. 25, fig. 6	Late Eocene	Endô, 1968	Pteleaecarpum bronnii (Unger) Weyland by Bůžek et al., 1989; reassigned to the genus <i>Craigia</i> A. C. Sm. et W. E. Evans as <i>C. bronnii</i> (Unger) Kvaček
K. borealis Heer	Renardodden, Spitzbergen, Norway	Leaflet	Heer, 1876, p. 89, pl. 25, fig. 5; Budantsev and Golovneva, 2009, p. 40, pl. 94, figs. 4, 5	Late Eocene	Kvaček et al., 1994; Budantsev and Golovneva, 2009	Rejected by Tralau, 1963
Koelreuteria sp.	West Primorye, far eastern Russia	Capsular valve $(1.5 \times 1.1 \text{ cm}, \text{elliptic, with a central body})$	Tashchi et al., 1996, p. 131, pl. 15, figs. 3, 4	Eocene	Tashchi et al., 1996	Reassigned to <i>Craigia</i> bronnii by Kvaček et al., 2005 and herein
K. maffeiana A. Massal.	Monte Bolca, Italy	Leaf	Massalongo, 1852, p. 19, pl. 4, fig. 14	Eocene	Edwards and Wonnacott, 1928	Doubtful or unidentifiable
K. eointegrifolia S. Endô	Fushun Coal-field, Liaoning Province, China	Capsular valve $(1.5 \times 1.1 \text{ cm}, \text{elliptic, with a central body})$	Endô, 1942, p. 42, pl. 17, fig. 13	Eocene	Endô, 1942; WGCPC, 1978	Reassigned to <i>Craigia bronnii</i> herein
K. arnoldii H. F. Becker	Upper Ruby River and Beaverhead Basins, southwestern Montana; Okanagan Highlands, Republic and McAbee, northeastern Washington (USA) and southern British Columbia (Canada)	Leaves and capsular valves(1.4–1.8 × 1.2–1.4 cm, elliptic or suborbicular)	Becker, 1961, p. 83, pl. 27, figs. 1–6; Becker, 1969, p. 115, pl. 36, figs. 20, 21; Wolfe and Wehr, 1987, p. 21, pl. 12, figs. 8, 10; Wehr, 1995, p. 9, pl. 3, fig. 9, pll. 3, fig. F p. 160, pl. 3, fig. F	Early to middle Eocene-Oligocene	Becker, 1969; Wolfe and Wehr, 1987; Greenwood et al., 2005; Manchester et al., 2009	Probably belonging to Boniodendron Gagnep. (= Sinoradlkofera F. G. Mey.); leaves resembling those of living species K. paniculata
K. microcarpa H. M. Li	Fushun, Liaoning Province, China	Leaflets and capsular valves (1.5–2.3 × 0.9–1.5 cm, oblong or suborbicular)	WGCPC, 1978, p. 131, pl. 111, fig. 4, pl. 115, fig. 4, pl. 117, figs. 3–6; Zhang, 1980, p. 333, pl. 209, figs. 2, 6, pl. 210, fig. 6; Guo. 1990, p. 33	Late Eocene to early Oligocene	Bůžek et al., 1989; Guo, 1990	Doubtful for leaflet identification. Capsular valves reclassified as <i>Craigia bronnii</i> by Kvaček et al., 1991, 2005
K. oregonensis C. A. Arnold	Wheeler County, Oregon, USA	Capsular valves (1.5–3 × 1.2–3.3 cm, elliptic or obovate)	Amöld, 1952, p. 77, pl. 1, figs. 1–3, text-fig. 1; Bůžek et al., 1989, n. 485, fiss. 36–44	Early Oligocene	Bůžek et al., 1989	Reclassified as <i>Craigia</i> <i>oregonensis</i> (C. A. Arnold) Kvaček et al. by Kvaček et al. 1991. 2005
Phyllites sp.	Indzhachaj River, Azarbajdžan	Leaf (ca. 8–10 cm long, brochidodromous)	Kasumova, 1966, p. 51, pl. 12, fig. 5	Middle Oligocene	Kasumova, 1966	Identified as a capsular valve of <i>Koelreuteria sp.</i> by Bůžek. 1971: reject herein
Koelreuteria sp.	Rott, Germany	Capsular valve (ca. 4.2 × 2.4 cm, ovate)	Weyland, 1937, p. 88, text-fig. 17, pl. 11, fig. 2; Kirchheimer, 1957, p. 206; Winterscheid, 2006, p. 159, pl. 16, fig. 4: in this paper. Fig. 38	Late Oligocene	Kirchheimer, 1957; Winterscheid, 2006	Reliable; reclassified as Koelreuteria macroptera (Kováts) W. N. Edwards herein
K. reticulata (Ettingsh.) W. N. Edwards	Holedeč, Most Basin, the Czech Republic	Capsular valves $(2-3.2 \times 1.2-2 \text{ cm}, \text{oblong-ovate})$	Brabenec, 1904, p. 2, fig. 1; Teodoridis, 2002, p. 122, pl. 7, fig. 9	Late Oligocene	Teodoridis, 2002	Reliable; synonymous with <i>K. macroptera</i> (Kováts) W. N. Edwards herein

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Puhlished name	I ocality	Organ	Cited fioure	Geological age	A ae reference	Identification
Sapindaceacarpun koelreuterioides Vaudois-Miéja et Lejal-Nicol	South-West of Khartoum, Sudan	Capsules (incomplete septum, but no veins)	Vaudois-Miéja and Lejal-Nicol, 1988, p. 855, pl. 1, figs. 1–6, pl. 2, figs. 1–6	Most possibly Oligocene	Prasad et al., 1986	Similar to extant <i>Koelreuteria</i> Laxm., but lacking reticulate venation in valves. Probably belonging to <i>Erythrophysa</i>
K. miointegrifoliola	Sakito, northwestern Kyushu	Leaflets	Matsuo, 1970, p. 33, pl. 9,	Oligocene	Matsuo, 1970	E. Mey. ex Am. Doubtful
Hu et R. W. Chaney K. jilinensis S. X. Guo (nom. nud.)	Prefecture, Japan Sanhe, Longjing County, Jilin Province, China	Capsular valve $(1.4 \times 1.3 \text{ cm})$	figs. 3–5, pl. 11, fig. 1 Guo, 1990, p. 35; Guo and Zhang, 2002,	Oligocene	Guo, 1990; Guo and Yang, 1997;	Reclassified as <i>Craigia</i> oregonensis by
K. rhombifolia S. X. Guo	Sanhe, Yanji County, Jilin Province, China	suborbicular) Leaflet and capsular valve (1.4 × 1.3 cm, suborbicular)	p. 203, pl. 3, figs. 8, 11 Guo, 1992, p. 578, pl. 269, figs. 1, 1a, 6	Oligocene	Guo and Zhang, 2002 Guo and Yang, 1997; Guo and Zhang, 2002	Guo and Zhang, 2002 Capsular valve reclassified as <i>Craigia oregonensis</i> herein; leaflet doubtful
Koelreuteria sp.	Sanhe, Longjing County, Jilin Province, China	Leaflets	Guo and Zhang, 2002, p. 202, pl. 2, figs. 7, 9	Oligocene	Guo and Yang, 1997; Guo and Zhang, 2002	Doubtful
K. <i>prisca</i> A. Massal. K. <i>prisca</i> A. Massal.	Chiavon, Italy Chiavon, Salcedo, Italy	Leafiet Leaf, leafiet and capsular valve	Massalongo, 1852, p. 18, pl. 4, ng. 13, Principi, 1926, p. 76, pl. 6, fig. 22 Massalongo, 1852, p. 17, pl. 3, figs. 10–12; Principi, 1926, p. 76, pl. 8, fig. 16	Oligocene Oligocene	Principi, 1926; Edwards and Wonnacott, 1928 Principi, 1926; Edwards and Wonnacott, 1928	Doubtful; resembling the leaflet of <i>Rhus</i> L. Doubtful; assigned to <i>Planera ungeri</i> by Ettingshausen, 1867; resembling the leaflet of Zallonic Succh
K. bipinnatoides H. F. Becker	Metzel Ranch, Upper Ruby River Basin, southwestern Montana, USA	Leaflets	Becker, 1972, p. 45, pl. 12, figs. 3–10	Oligocene	Becker, 1972	Doubtful; resembling the leaflet of living <i>K binimata</i>
K. reticulata (Ettingsh.) W. N. Edwards	Tállya and Tokay, Hungary: St. Galler Findlinge (Kumkler), Germany: Pětipsy Area and Blíina (Most Basin), North Bohemia, the Czech Republic	Capsular valves (3.5-4.0×2.5-2.8 cm, oblong-ovate or ovate)	Ettingshausen, 1854, p. 813, pl. 4, fig. 5; Heer, 1857, p. 156, pl. 145, fig. 16; Hollick, 1923, p. 212, pl. 10, figs. 4, 5; Rásky, 1958, p. 184, pl. 17, fig. 9; Bužek, 1971, p. 84, pl. 31, figs. 22, 23; Kvaček et al., 2004b, p. 94, fiest, d. e.	Early to middle Miocene	Kirchheimer, 1957; Rásky, 1958; Bůžek, 1971; Kvaček et al., 2004a, b; Hably, 2006	Reliable, synonymous with <i>K. macroptera</i> (Kováts) W. N. Edwards herein
Sapindaceacarpum lunulatum Andreánszky	Déllő, Hungary	Capsular valve $(3.8 \times 3.2 \text{ cm, ovate})$	zooro, p. 7.1, ng., u. Andreánszky, 1959, p. 156, pl. 47, fig. 5, text-fig. 183;	Early Miocene	Andreánszky, 1959; Hably, 2006	Reclassified as <i>Koelreuteria</i> macroptera (Kováts) W. N. Edwards by Gregor, 1087 and harain
Koelreuteria sp.	Pingzhuang Basin, Chifeng, Inner Mongolia, China	Unknown	Shang et al., 2001,	Early Miocene	Shang et al., 2001	Doubtful
K. macroptera (Kováts) W. N. Edwards	Gomboska of Tällya, Abaújszántó and Szántó, Hungary; Randecker Maar, Bavaria and Kreuzau, Germany; Bohemia, the Czech Republic; Chiuzbaia, Romania; Klepariv of L'viv City, Ukraine	Capsular valves (3.4-5 × 2.5-4 cm, ovate or oblong-ovate, with two seeds)	 p. 7.02, not guest Unger, 1876, p. 51, pl. 1, fig. 2; Unger, 1870, p. 14, pl.5, fig. 1; Kirchheimer, 1943, p. 404, text-fig. 11; Weyland, 1948, p. 133, pl. 21, fig. 6; Kirchheimer, p. 135, p. 206, pl. 33, fig. 143a, b; Rásky, 1958, p. 184, pl. 17, figs. 10, 11; Rüffle, 1963, p. 237, pl. 12, figs. 28–31, pl. 13, figs. 1–5, pl. 27, fig. 9; Gregor, 1982, p. 113, pl. 7, fig. 14; Shvareva, 1993, p. 135, fig. 14; Shvareva, 1993, p. 135, fig. 14; Shvareva, 1993, p. 135, fig. 2; Hably et al., 2001, p. 73, s3, 106, pl. 101, fig. 1, pl. 122, fig. 3; 	Early to middle Miocene	Gregor, 1982; Krautter and Schweigert, 1991; Gregor et al., 1992; Hably, 2006; Seehuber, 2009	Reliable. Basionym: <i>Ptelea macroptera</i> Kováts, 1856; synonym: <i>Koelreuteria</i> sp. in Weyland, 1948

TABLE 2. Continued.

TABLE 2. Continued.						
Published name	Locality	Organ	Cited figure	Geological age	Age reference	Identification
K. macrocarpa J. R. Tao	Shanwang Basin, Linqu County, Shandong Province, China	Leaflet and capsular valve (3.8 × 2.9 cm, elliptic)	WGCPC, 1978, p. 130, pl. 116, fig. 8, text-fig. 65; Tao et al., 1999, p. 46, 75, pl 43, fig. 2	Early to middle Miocene	Li, 1981; Liu and Leopold, 1992; Deng, 2006; Wang et al., 2010; Wang, 2012	Capsular valve referred to <i>K. miointegrifoliola</i> Herein; leaflet similar to <i>K. binimuta</i>
Koelreuteria sp.	Shanwang Basin, Linqu County, Shandong Province China	Capsular valve $(4.5 \times 3.5, \text{ ovate})$	Sze, 1954, p. 80, pl. 68, fig. 5	Early to middle Miocene	Li, 1981; Liu and Leopold, 1992; Deng, 2006; Wang et al. 2010: Wang. 2012	Reliable; referred to K. miointegrifoliola
Koelreuteria sp.	Shandong Hovines, Cuina Shanwang Basin, Linqu County, Shandong Province, China	Leaflet	Tao et al., 1999, pl. 14, fig. 4	Early to middle Miocene	Li, 1981; Liu and Leopold, 1992; Deng, 2006; Wano et al 2010	Doubtful
K. crenulata Pavlyutkin	Novokachalinsk, Khanka Area, Primorve, far eastern Russia	Leaflets	Pavlyutkin, 2005, p. 124, pl. 36. fig. 9. pl. 37. fig. 1	Middle Miocene	Pavlyutkin, 2005	Doubtful
Koelreuteria sp.	Shimo-tokusan, Tatsunokuchi-machi, Kaga, Ishikawa Prefecture. Janan	Leaflets	Fuji and Kitanaka, 1988, p. 138, fig. 3. 4; Kitanaka and Fuji, 1988, p. 112, fig. 17.	Middle Miocene	Fuji and Kitanaka, 1988; Kitanaka and Fuji, 1988	Doubtful
K. oeningensis Heer	Oeningen, Baden, southern Germany	Leaflets	Heer, 1877, p. 63, pl. 121, figs. 18–20, pl. 127, fig. 393, Engelhardt, 1873, p. 31, pl. 12, fig. 12, Schimper and Schenk, 1890, p. 53, fig. 314, 3	Middle Miocene	Edwards and Wonnacott, 1928; Rüffle, 1963	Doubtful
K. vetusta Heer	Oeningen, southern Germany	Leaflet	Heer, 1857, p. 63, pl. 127, fig. 39a	Middle Miocene	Edwards and Wonnacott, 1928; Rüffle, 1963	Assigned to a rosaceous species <i>Crataegus</i> longepetiolata Heer by Hantke. 1954
K. miointegrifoliola Hu et R. W. Chaney	Shanwang Basin, Linqu County Shandong Province, China; Ōsudo, Niigata Prefecture, Oguni-machi, Chõjabaru, Iki Island, Shimonoseki, Tottori Prefecture, Southwest Honshu, Kabutoiwa, border of Gunma and Nagano prefectures, Central Honshu, Japan	. Leaflets and capsular valves (3.2–5.2 × 3.0–4.2, ovate, oblong, elliptic or suborbicular)	Hu and Chaney, 1938, p. 64, pl. 38, figs. 1, 3, pl. 39, fig. 3; Takhashi and Naito, 1952, p. 71, fig. 2; Huzioka and Takahashi, 1973, p. 141, pl. 4, figs. 2, 3; Onoe, 1974, p. 52, pl. 12, figs. 2, 4; Hayashi, 1975, p. 26, pl. 21, fig. 3, pl. 22, figs. 2, p. 26, pl. 21, fig. 3, pl. 22, figs. 2, p. 26, pl. 21, fig. 3, pl. 22, figs. 2, p. 26, pl. 17, fig. 9; Tao et al, 1991, p. 159, pl. 17, fig. 9; Tao et al, 1999, p. 46, 76, pl. 35, fig. 1, pl. 43, fig. 3; this paper, Figs. 39–47	Miocene	Ishida et al., 1970; Ozaki, 1980; Li, 1981; Liu and Leopold, 1992; Deng, 2006; Wang et al., 2010; Wang, 2012; this paper	Reliable for the capsule valves; some leaflets belonging to <i>Juglans</i> L., the others excluded from <i>K. miointegrifoliola</i> by Brown, 1946 and herein
K. caucasica Palib.	Leninakan, Armenia	Leaf	Palibin, 1939, p. 623, not figured; Kolakovsky, 1973, p.135	Miocene	Kolakovsky, 1973	Rejected by Iljinskaja, 1963
K. ulmifolia A. Massal.	Sinigaglia, Italy	Leaflets	Massalongo, 1858, p. 80, pl. 1, pl. 17, pl. 36, fig. 8	Miocene	Meschinelli and Squinabol, 1892	Doubtful; combined into <i>Cissus ulmifolia</i> by Massaloneo 1858
K. ? serrata Heer	Sakhalin, far eastern Russia	Leaflet	Heer, 1878, p. 52, pl. 14, fig. 10	Miocene	Edwards and Wonnacott, 1928	Unidentifiable
Koelreuteria sp.	Khanka, West Primorye, far eastern Russia	Leaflet	Ablaev et al., 1994, p. 129, pl. 15, figs. 4, 5	Miocene	Ablaev et al., 1994	Doubtful
K. cf. reticulata (Ettingsh.) W. N. Edwards) Chiuzbaia, Romania	Capsular valves (2.3 × 2.1 cm, elliptic, with a central body)	Givulescu, 1984, p. 83, pl. 1, figs. 1–2; Givulescu, 1990, p. 120, pl. 34, figs. 4-5	Late Miocene	Givulescu, 1990	Reclassified as Craigia bronnii herein
Koelreuteria sp.	Bangmai and Mengtuo, Lincang, Yunnan Province, China	Leaflet and valves (3.1–3.7 × 2.3–2.6 cm, elliptic)	Tao and Chen, 1983, p. 88, pl. 19, figs. 6, 13; Tao et al., 2000, p. 275, pl. 71, fig. 8; Guo, 2011, p. 359, pl. 9, figs. 6, 7, 11	Late Miocene	Ge and Li, 1999; Tao et al., 2000; Guo, 2011	Doubtful; leaflet incomplete; valves were reclassified as <i>Salvinia paralleloneura</i> S. X. Guo by Guo, 2011

TABLE 2. Continued.						
Published name	Locality	Organ	Cited figure	Geological age	Age reference	Identification
K. margaritifera (R. Ludw.) Mai	Rhön Mountains, Salzhausen, Lettengraben, Salzhausen, Wackersdorf, Germany; Styria, Austria: Wieliczka, Poland; Tanaro Basin, Ca' Viettone and San Miniato, Italy	Seeds (3.4–6.5 mm in diameter, originally globose. Surface very smooth, without furrows or ridges. Hilum scar larger)	Kirchheimer, 1936, p. 89, pl. 9, fig. 1a–f; Gregor, 1982, p. 32, pl. 10, figs. 1–6, 8; Mai and Walther, 1991, p. 108, pl. 13, fig. 17; Cavallc and Martinetto, 1996, p. 22, pl. 6, fig. 3; Łańcucka-Środoniowa and Zastawniak, 1997, p. 18, pl. 3, fig. 3; Martinetto, 2001, p. 309, pl. 1, figs. 26a, b; Kovar-Eder and Meller, 2001, p. 86, pl. 6, fig. 1; Gümbel and Mai, 2006, p. 21, pl. 4, figs. 4–6	Early to middle Miocene-early to middle Pliocene	Gregor, 1982; Cavallo and Martinetto, 1996; Eańcucka-Środomiowa and Zastawniak, 1997; Martinetto, 2001; Kovar-Eder and Meller, 2001; Gümbel and Mai, 2006	Generally smaller than seeds of Koelreuteria (5–9 mm in diameter), with a very smooth surface and larger hilum scar that further differs from Koelreuteria; classified into Sapindoidea margaritifera (R. Ludw.) Kirchh. by Gregor, 1978, 1980, 1982; Cavallo and Martinetto, 1996; Martinetto, 2001 and this proper
Koelreuteria cf. oeningensis Heer	Imola, Italy	Leaflet	Sangiorgi, 1916, p. 293, pl. 15. fig. 19	Miocene-Pliocene	Sangiorgi, 1916	Doubtful
Phyllites cf. K. bipinnata Franch.	Toutang, Mengzi County, Yunnan Province, China	Leaflets	Colani, 1920, p. 227, pl. 9, figs. 4, 8	Miocene-Pliocene	Edwards and Wonnacott, 1928; WGCPC, 1978	Doubtful
Koelreuteria sp. aff. K. integrifolia Merr.	Brusturilor-Bach and Valea Neagră Bezirk Crisana, Romania	Capsular valve $(3.4 \times 2.6 \text{ cm, ovate})$	Givulescu, 1961, p. 103, fig. 8; Givulescu, 1962, p. 160, pl. 37, fig. 263	Early Pliocene	Givulescu, 1967	Reliable; reclassified as <i>Koelreuteria macroptera</i> herein
Koelreuteria cf. integrifolia Merr.	Zhangcun, Yushe Basin, Shanxi Province, China	Unknown	Cao and Cui, 1989, p. 371. not figured	Late Pliocene	Shi et al., 1993	Doubtful
K. yuamouensis Cheng, Yin, Mehrotra et Li	Hutiaotan Earth Forest (Lower Yuanmou Formation), Yuanmou Basin, Yunnan Province, China	Wood (ring porous)	Cheng et al., 2012, p. 302, fig. 1	Pliocene	Qian, 1991; Cheng et al., 2012	Reliable

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Figs. 1–12. Morphology of extant species in *Koelreuteria* Laxm. **1.** Capsule and **2.** capsular valve of *K. paniculata* Laxm. from cultivated tree at the Institute of Botany, Chinese Academy of Sciences, Beijing. **3.** Capsule of *K. bipinnata* Franch. from cultivated tree at Yunnan Academy of Forestry, Kunming. **4.** Capsule of *K. elegans* (Seem.) A. C. Sm. from Tailevu, Viti Levu of Fiji. Voucher: A 1213, B. E. Parham. **5.** Cleared capsular valve of *K. paniculata*, showing a basal septum equivalent to ca.1/3 of valve length. **6.** Cleared capsular valve of *K. bipinnata*, showing a straight placental suture. **7.** Cleared capsular valve of *K. henryi* Dümmer from cultivated tree at the University of Florida, Gainesville, showing polygonal, transversely elongated nets. UF Slide no.: 20100203. **8.** Polar view of pollen grain of *K. paniculata* under SEM, showing a type of longitudinally striated ektexine. **9, 10.** Partial leaf and cleared leaflet of *K. bipinnata*, showing almost entire margins. Bars = 2 cm, except in Fig. 8, which is 10 μm.

The exine is about $1.5-2.4 \,\mu\text{m}$ thick, and the ornamentation of the extexine is, more or less, longitudinally striate, finely striate, or irregularly striate-reticulate (Wang and Chien, 1956; Meyer, 1976; Acevedo-Rodríguez et al., 2011) (Fig. 8). On the basis of the ectoaperture and pollen grain shape, Muller and Leenhouts (1976) recognized 12 pollen types in Sapindaceae and placed Koelreuteria in their type A, characterized by colporate spheroidal pollen grains of the kind found in most genera of the family, which they considered to be basic (Buerki et al., 2009) and possibly plesiomorphic. Pollen fossil records, such as the recently reported Talisiipites megorites Z. C. Song et M. Y. Li, Senipites striatus Z. C. Song (Song and Li, 1982; Song et al., 1999, 2004; Li et al., 2008), and Koelreuteria sp. (Leopold and Clay-Poole, 2001; Leopold et al., 2008) closely resemble this extant genus and date from the Late Cretaceous to the Eocene of southern China and western United States. The paleogeographic pattern of sapindaceous pollen may be important for correlation with the Koelreuteria fruit fossils.

Leaves of Koelreuteria are alternate, imparipinnate, bipinnate, and estipulate (Figs. 9-12; Table 1). The leaflets are sessile or shortly petiolulate. The blades are ovate, obliquely ovate, wide ovate to oblong-ovate or ovate-lanceolate. The bases are slightly or extremely oblique, wide cuneate, rounded or subtruncate. The apxes are obtuse, acute, short or long acuminate to caudate. The margins vary among different species, from entire (e.g., K. bipinnata, see Figs. 11, 12) to variously lobed, with irregularly obtuse or incurved serrations (e.g., K. paniculata, see Figs. 9, 10), belonging to the rosoid tooth type (sensu Hickey and Wolfe, 1975). The rosoid tooth type is thought to stem from the base of core eudicots (Doyle, 2007), but appears to be apomorphic in Koelreuteria and closely related taxa. Koelreuteria bears polygonal leaf epidermal cells and anomocytic stomatal apparatus, both of which also occur in other sapindaceous genera such as Allophylus L., Boniodendron Gagnep., Erioglossum Blume, Delavaya Franch., Eurycorymbus Hand.-Mazz., Handeliodendron Rehder, Paranephelium Miq., and Xanthoceras Bunge (Cao and Xia, 2008). The bipinnate leaves bearing the leaflets with rosoid teeth assigned to Koelreuteria allenii (Lesq.) W. N. Edwards from the late Eocene Florissant Formation of Colorado (see MacGinitie, 1953: pl. 55, figs. 1-4; pl. 75, fig. 1) appear to be a reliable fossil representative of *Koelreuteria*. The name *K. allenii* is originally based on fruits; the hypothesis that associated leaves may represent the same species is reasonable, but not proven by specimens showing these organs in attachment. Overall, the leaflet morphology of *Koelreuteria* shows considerable similarities to that of other sapindalean genera, which often places fossil investigators in a predicament. The majority of species of the previously published leaf and leaflet fossils of Koelreuteria are questionable (Table 2) because they present suites of characters that are not unique to extant Koelreuteria. Some of them are too poorly preserved to be amenable to investigation of higher order venation and epidermal anatomy.

The woods of *Koelreuteria* have distinct growth rings, marked by radial flattened latewood fibers and much larger vessels in the earlywood and marginal parenchyma (Li et al., 1995; Klaassen, 1999). The wood is ring porous. Vessels in the earlywood are mainly solitary (*K. bipinnata* and *K. elegans*) or in irregular clusters of 2 or 3 (*K. paniculata*). More anatomical details of *Koelreuteria* are available (http://insidewood.lib.ncsu.edu/search). Fossil woods of *Koelreuteria* have been described from the latest Eocene Florissant Formation of Colorado, western United States (Wheeler, 2001) and the Pliocene Lower Yuanmou Formation of Yunnan, southwestern China (Cheng et al., 2012). Remarkably, ring to semiring porosity in Sapindaceae is known only in three genera, *Sapindus* L., *Xanthoceras*, and *Koelreuteria* (including *K. elegans* in Fiji as well as the temperate species, see Li et al., 1995; Klaassen, 1999), whereas the usual condition is diffuse, or semiring porous. Hence, the paleogeographic pattern of sapindaceous wood fossils is helpful to correlate with that of *Koelreuteria* fruit fossils.

Systematics of capsular fossils in Koelreuteria—About 30 fossil species under the name *Koelreuteria* have been documented from the Late Cretaceous to the Cenozoic of Eurasia and North America (Table 2). Overall, the fruit capsule and capsular valves are more reliably diagnostic than the pollen, leaflet, and wood for secure identification of the genus. The fruit taxa can be distinguished by the following taxonomic key:

Key to species—

1. Capsular valves obovate or obovoid-elliptic, with a cune-
ate of decurrent base K allenii (Lesa)
W. N. Edwards
1. Capsular valves ovate, elliptic, oblong, ovoid-elliptic, ob-
long-elliptic, or suborbicular, without a cuneate or de-
current base
2. Consular values elliptic shlare evoid elliptic shlare
2. Capsular valves elliptic, oblong, ovoid-elliptic, oblong-
rounded apex
2. Capsular valves ovate, oblong-ovate, oblong, elliptic or
suborbicular, with a rounded or slightly cordate base
and a convex, acute, short acuminate, or slightly
emarginated apex4
3. Capsular valves smaller, 2.4–2.5 cm long
2 Consular values larger 3.5.5.8 cm long
<i>K</i> dilcheri sp. nov
4. Capsular valves ovate, or oblong-ovate, with a septum ac-
counting for approximately 1/3 or slightly more than 1/3
of the valve length from the base
K. macroptera (Kováts) W. N. Edwards
4. Capsular valves ovate, oblong, elliptic or suborbicular, with
a septum accounting for approximately half of the valve
length from the base
K. miointegrifoliola Hu et R. W. Chaney

SYSTEMATIC PALEOBOTANY

Family-

Sapindaceae Juss.

Subfamily—

Dodonaeoideae Hook.

Tribe—

Koelreuterieae Radlk.

Genus-

Koelreuteria Laxm.

Species—Koelreuteria taoana Qi Wang, Manchester, H.-J. Gregor, S. Shen et Z. Y. Li, sp. nov. (Figs. 13–16).

Seed	No./valve (Shape)	1 (Oblong)	1 (Ohovoid)	(Ovoid-oblong)	1, 2 or 4	(Obovoid) (Obovoid)	2–4 (Obovoid)	1 (Reniform)	1 (Globose)	1 (Globose)	1 (Globose)	1 (Globose)	1 (Globose)	2 (Globose)	1 (Oblong)	1–2 (Lenslike)	2 (Globose)	2 (Globose)
	Position	One level	One level	Biserial	Biserial	Biserial	Biserial	One level	One level	One level	One level	One level	One level	One level	One level	One level	One level	One level
	Sepal	Caducous	Caducous	Caducous	Caducous	Caducous	Caducous	Persistent	Caducous	Caducous	Caducous	Caducous	Persistent	Persistent	Persistent	Caducous	Caducous	Persistent
	Septum	No	No	Complete	Complete	Complete	Complete	No	Complete	Complete	Complete	Complete	Complete	Complete	Complete	Complete	Complete	Possibly incomplete
Valve	Shape	Triangular or rhombic	Triangular	Elliptic to oblong	Suborbicular	Elliptic, obovate or suborbicular	Elliptic or obovate	Suborbicular	Wide elliptic or suborbicular	Suborbicular or obcordate	Wide elliptic	Wide elliptic	Obtriangular or elliptic	Elliptic	Suborbicular	Suborbicular or wide obovate	Ovate	Obovate
	Central body	Absent	Absent	Present	Present	Present	Present	present	Absent	Absent	Absent	Absent	Absent	Absent	Absent	present	Absent	Absent
	Dehiscence	No	No	Loculicidal or septicidal	Loculicidal or	septicidal or septicidal	Loculicidal or septicidal	No	Loculicidal	Loculicidal	Loculicidal	Loculicidal	Septifragal or marginicidal	Loculicidal	Loculicidal	Septifragal or septicidal	Loculicidal	Loculicidal
	Texture	Papery or coriaceous	Papery, scarious	Membranous	Membranous	Membranous	Membranous	Papery	Papery	Membranous	Membranous	Papery	Membranous or papery	Membranous or papery	Woody, crustaceous	Membranous or papery	Membranous	Membranous- papery (pilose)
	Length (cm)	0.5-1.2	0.8–1.5	2.5–3	Ca. 3.5	0.9–2.4	1.5–3	1.6–1.8	2-4	1.8–2.3	1.5-1.6	Ca. 1.5	1.5-3	2.5-6.0	2–2.3	1.5-2.2	4-7	3.5-4
Fruit	Shape	4-5-winged or not winged, fusiform,	arrowneau suapeu 5-winged, broadly furhinate or cordate	5-locular, winged, 5-angled	5-locular, winged,	5-locular, winged	5-locular, winged	2-3-Locular, winged, flattened,	3-Locular, winged, inflated	3-locular, winged, subglobose	At least 2-locular, winged, inflated	3-Locular, winged, inflated	3-Locular, inflated, turbinate, obtriangular or ellipsoid	3-Locular, more or less compressed	3-Locular, 3-4-sided, pyriform or	2-6-Locular, winged, obcordate or	oumpressed grouse 3-Locular, inflated, ovoid	3-Locular, inflated, trigonous, obovoid
	Species (Family)	Abronia fragrans Nutt. ex Hook. 5, 14 Muctorinococo Tuce 1	(Inyctagmaccae Juss.) Abronia macrocarpa L. A. Gallowav ⁵ (N)	Craigia kwangsiensis H. H. H. Sue (Maluaceae Evec)	C. yunnanensis A. C. Sm.	C. bronnii (Unger) Kvaček et al. [= Preleaecarpum bronnii (Unger)	Weyland) ^{1,1,0} (M.) C. oregonensis (C. A. Amold) Kvaček et al. [= <i>P.</i> oregonensis (C. A. Arnold) Monchestral 16 (M.)	Ptelea enervosa H. V. Sm. ^{14,22} Dutorosas Luce)	Arfeuillea arborescens Pierre ^{l, 19}	() computaceae 1 uss.) Boniodendron minus (Hems!.) T. C. Chen [= Sinoradlkofera minor (Hems!.) F. G. Mev. 11-4, 19, 23 (S.)	Bridgesia bovayensis Manchester	<i>B. incisifolia</i> Bertero ex Cambess. ^{3, 14, 23} (S.)	Cardiospermum halicacabum L.2.3.15.16 (S.)	Conchopetalum brachysepalum Cannen3, 5, 16, 19 (S)	Cupaniopsis anacardioides (A. Rich.) Radlk. ^{3,5} (S.)	Dodonaea viscosa (L.) Jacq. ^{2,3,5,7,14,16,21,23} (S.)	<i>Erythrophysa alata</i> (Eckl. et Zeyher) Hutch ^{3,8,21} (S)	E. paniculata Capuron ^{3, 16} (S.)

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TABLE 3. Comparisons between Koelreuteria species and other taxa with similar fruits.

WANG ET AL.—*KOELREUTERIA* FRUITS FROM THE CENOZOIC

	Fruit					Valv	0			eed
Species (Family)	Shape	Length (cm)	Texture	Dehiscence	Central body	Shape	Septum	Sepal	Position	No./valve (Shape)
E. septentrionalis	3-Locular, inflated,	3.5	Membranous	Loculicidal	Absent	Suborbicular	Complete	Caducous	One level	2
Verdc. ^{, o} (S.) <i>E. transvaalensis</i> I. Verd. ^{3, 8, 21} (S.)	subglobose 3-Locular, 3-ridged, inflated, obovoid or subalobose	×	Membranous	Loculicidal	Absent	Obovate or suborbicular	Complete	Caducous	One level	(Globose) 1–2 (Globose)
Euphorianthus euneurus	3-Locular, inflated,	0.8 - 1	Velvety or flachy	Loculicidal	Absent	Suborbicular	Incomplete	Persistent	One level	1 (Obovoid)
(wuq.) Lecum (3.) Koelreuteria paniculata Laxm. ^{1-5, 10, 23} (S.)	3-Locular, inflated, not winged, trivonous-conoidal	2.2-7.2	Membranous (glabrous)	Loculicidal	Absent	Ovate	Incomplete	Persistent	One level	(Globose) (Globose)
K. bipinnata Franch. ^{1–5, 10, 14, 23} (S.)	3-Locular, inflated, 3-ridged, not winged, ellipsoid or subglobose	4-7	Membranous (glabrous)	Loculicidal	Absent	Elliptic, suborbicular	Incomplete or rarely complete with septal slirs	Persistent	One level	2 (Globose)
K. henryi Dümmer (= K. formosana Havata) $^{1-5, 10-12, 23}$ (S.)	3-Locular, inflated,3-ridged, not winged,subglobose	4-5	Membranous (glabrous)	Loculicidal	Absent	Suborbicular	Incomplete	Persistent	One level	2 (Globose)
K. elegans (Seem.) A. C. Sm. ^{1–5, 13, 23} (S.)	3-Locular, inflated, 3-ridged, not winged, subelobose	3.4-5	Membranous (glabrous)	Loculicidal	Absent	Suborbicular	Incomplete	Persistent	One level	2 (Globose)
<i>Majidea zanguebarica</i> J. Kirk ex Oliv. ^{3, 5, 16, 19} (S.)	3-Locular, inflated, not winged, subglobose	Ca. 3	Crustaceous	Loculicidal, deeply lobed	Absent	Suborbicular	Complete	Persistent	One level	2 (Obovoid- oblong)
Paullinia hispida Jacq. ^{3,14} (S.)	3-Locular, winged, obovoid	1.2–1.5	Coriaceous	Septifragal	Absent	Obovate	Complete	Caducous	One level	1 (Ovoid)
P. dasystachya Radlk. ^{3,15} (S.)	3-Locular, winged, obovoid	1.8–2.2	Crustaceous	Septifragal	Absent	Obovate	Complete	Persistent	One level	(Ellipsoid)
Sapindaceacarpum koelreuterioides Vaudois-Miéja et Lejal- Nicol ¹⁷ (S.)	3-Locular, inflated, not winged, subglobose	2.0-2.2	Membranous (glabrous, not reticulate)	Loculicidal	Absent	Suborbicular	Incomplete	Persistent	One level	(Globose)
<i>Serjania altissima</i> (Poepp.) Radlk. ^{3,15} (S.)	3-Locular, winged, inflated, ovoid to oblong	Up to 3	Membranous or papery	Septifragal	Absent	Ovate	Complete	Persistent	One level	1 (Ellipsoid)
<i>S. polyphylla</i> (L.) Radlk. ³ (S.)	3-Locular, winged, inflated	3.3	Membranous	Septifragal	Absent	Ovate	Complete	Persistent	One level	1 (Globose)
Smelophyllum capense (Sond.) Radlk. ^{3, 20, 21} (S.)	2–3-Locular, inflated, globose	1.3	Chartaceous or fleshy	Loculicidal, usually two-lobed	Absent	Orbicular	Complete	Caducous	One level	1 (Globose)
Stocksia brahuica Benth. ^{3, 14, 18} (S.)	3-Locular, winged, inflated, subglobose	2.5-3	Membranous	Loculicidal	Absent	Suborbicular or wide ovate	Complete	Caducous	One level	1–2 (Subglobose)
Urvillea filipes Radlk ^{3,14} (S)	3-Locular, inflated, marginally winged	Ca. 2.0	Membranous	Septifragal	Absent	Suborbicular	Complete	Caducous	One level	(Globose)
U. chacoensis Hunz. ^{3, 14, 23} (S.)	3-Locular, inflated, marginally winged	Ca. 2.2	Membranous	Septifragal	Absent	Obovate	Complete	Persistent	One level	(Globose)
U. ulmacea Kunth ^{3, 15} (S.)	3-Locular, inflated, marginally winged, ellinsoid	3.2-4	Membranous or papery	Septifragal	Absent	Elliptic to obovate	Complete	Persistent	One level	(Globose)
Reference notes: ¹ Bůžek ⁸ Verdcourt, 1962; ⁹ Leenhc ¹⁶ Capuron, 1969; ¹⁷ Vaudois on the cultivated plants and t	et al., 1989; ² Xia and Gad uts, 1988; ¹⁰ Luo and Che -Miéja and Lejal-Nicol, 19 he herbaria kept at the Chir	ek, 2007; ³ Ace ⁻ m, 1985; 11 Li, 88; ¹⁸ Abdulia, nese National H	vedo-Rodríguez e 1963, 1977; ¹² C 1973; ¹⁹ Buijsen e ⁱ rbarium (PE), the	t al., 2011; ⁴ Me ⁻ 'hen, 1993; ¹³ Sr t al., 2003; ²⁰ Sch : Missouri Botan	yer, 1976, 1977; mith, 1952, 1985 nönland, 1913; ²¹ ical Garden Herl	⁵ see http://www.d 5; 14 Manchester Palmer and Pitme Darium (MO), and	efforas.org; ⁶ Kva and O'Leary, 20 m, 1972; ²² Call a Arnold Arboretu	tček et al., 2(10; ¹⁵ Weck and Dilcher, m Herbarium	205; ⁷ Paoli a erle and Ruti 1995; ²³ Dire 1, Harvard Un	nd Sarti, 2008; shauser, 2005; st observations iversity (A).

TABLE 3. Continued.

Synonym—Koelreuteria cf. *miointegrifolia* Hu et R. W. Chaney in Ablaev (1978), 187, pl. 10, fig. 5; *Koelreuteria* sp. in Ablaev (2000), 53, pl. 15, figs. 6, 7 (Fig. 16 herein); *Koelreuteria* sp. in Manchester et al. (2005), 10, pl. 4, figs. 8–10 (Figs. 13–15 herein).

Diagnosis—Capsular valves, entire-margined, elliptic or ovoid-elliptic with a longitudinal, middle carpellary suture, over 2 cm long. Apex rounded. Base rounded or slightly truncate. Septum present along the lower part of the carpellary suture. Lateral veins on both sides of the suture sinuous, reaching margin. Thinner veins between lateral veins consisting of irregular, transversely elongated reticulum.

Etymology—The specific epithet is dedicated to Prof. Tao Junrong (Institute of Botany, CAS, Beijing) for her important contributions to Cenozoic paleobotany.

Holotype—PE 054018a, b, c (Figs. 13, 14) (part and counterpart specimens, designated here; Huadian County, Jilin Province, Middle Eocene Huadian Formation of northeastern China; first illustrated by Manchester et al. (2005: pl. 4, figs. 8–10]).

Description-Fossils are preserved as individual capsular valve impressions. Capsular valves are entire-margined, elliptic (Figs. 13-15) or ovoid-elliptic (Fig. 16), 2.4-2.5 cm long and 1.6-1.7 cm wide. The valve apex is rounded, and the base is rounded or slightly truncate. A longitudinal, middle carpellary suture is prominent and thicker in the lower part than the upper one. Along the lower part of the carpellary suture, the valve is involutely fused, forming a septum (Figs. 14, 15). The septum extends approximately half of the valve length from the base and is about 3 mm high, terminating in a prominent placental suture. Seeds are not preserved. On both sides of the carpellary suture, lateral veins diverge at 30°–90° (or even 120° at the base) and arrive at the margin. Numerous thinner veins between lateral veins anastomose, forming a polygonal reticulum with areoles that are mostly irregular and transversely elongated in outline (Figs. 15, 16). The texture of valves is membranous.

Comments—In comparison with the length range observed in extant *Koelreuteria* capsules (2.2–7.2 cm long; see Table 1), the valves of this fossil species are relatively small. The small number of fossil capsular valves from the localities may limit our knowledge of the full range of variation, and larger capsular valves might be found from these Asian localities in the future. *K. taoana* was distributed along the North-West Pacific-Rim areas in the middle Eocene of northeastern China and far eastern Russia (Fig. 48).

Species—Koelreuteria dilcheri Qi Wang, Manchester, H.-J. Gregor, S. Shen et Z. Y. Li, sp. nov. (Figs. 17–21).

Synonym—Koelreuteria mixta (Lesq.) R. W. Br., Brown (1946), pro parte, 351, fig. 1 (non fig. 2) (Figs. 17, 18 herein).

Diagnosis—Capsular valves, entire-margined, oblong or oblong-elliptic with a longitudinal, middle carpellary suture, more than 3 cm long. Apex rounded. Base rounded. Septum ocurring along the lower part of the carpellary suture. Lateral veins on both sides of the suture sinuous, reaching the margin. *Etymology*—The specific epithet is dedicated to Prof. David Dilcher (Indiana University), who has made important contributions to Cenozoic paleobotany.

Holotype—USNM P42363 (Figs. 17, 18) (designated here; Independence Hill near Colfax, California, United States, middle Eocene Chalk Bluffs flora; first illustrated by Brown [1946: fig. 1]).

Paratypes—UWBM PB94575 (Fig. 19), 1695 and 1696 (Figs. 20, 21) (part and counterpart specimens; Republic, Washington, United States, early to middle Eocene Republic flora).

Description—Fossils are preserved as individual capsular valve impressions. Capsular valves are entire-margined, elliptic (Figs. 17, 18), oblong or oblong-elliptic (Figs. 19–21), 3.5-5.8 cm long and 2.8-3.6 cm wide. Both apex and base are rounded. A longitudinal, middle carpellary suture is prominent and thicker in the lower half-part than the upper one (Fig. 19), implying an incomplete septum in the valves. Seeds are not preserved. On both sides of the carpellary suture, lateral veins diverge at $30^{\circ}-110^{\circ}$ and arrive at the margin. Between lateral veins, thinner veins anastomose, forming a polygonal reticulum with areoles that are mostly irregular and transversely elongated in outline (Figs. 18, 20–21). The texture of valves is membranous.

Comments—Brown (1946) first described this capsular valve and associated leaflet fossils under the name *Koelreuteria mixta* from middle Eocene Chalk Bluffs flora of Independence Hill near Colfax, California, but its basionym *Rhus mixta* Lesq. (Lesquereux, 1878a) was only used for leaflet fossils. Hence, we separate the capsular valve fossil from *K. mixta* and reclassify it into *K. dilcheri* sp. nov. This new species is very similar to the above described *K. taoana* from northeastern Asia, but these two species show a certain degree of discontinuity in valve size. *Koelreuteria dilcheri* was distributed along the North-East Pacific-Rim areas in the early to middle Eocene of western United States (Fig. 48).

Species—Koelreuteria allenii (Lesq.) W. N. Edwards (Figs. 22–29).

Koelreuteria alleni (Lesq.) W. N. Edwards (1927), 110; MacGinitie (1953), pro parte, 145, pl. 55, fig. 7 (non pl. 55, figs. 1–4 nec pl. 75, fig. 1).

Basionym—Ophioglossum alleni Lesq., Lesquereux (1873) in Lignitic Formation and Fossil Flora, 371.

Synonym—*Salvinia alleni* (Lesq.) Lesq., Lesquereux (1878b), 65, pl. 5, fig. 11; Lesquereux (1883), 136, pl. 21, figs. 10–11; *Tmesipteris alleni* (Lesq.) Hollick (1894), 256, pl. 205, fig. 12; *Carpolithes alleni* (Lesq.) Cockerell (1913), 500; Hollick (1923), 211, pl. 10, figs. 1–3, 6–11, pl. 11, figs. 1–3; *Phyllites alleni* (Lesq.) Florin (1919), 254; *Staphylea viridifluminis* Hollick (1929), 96, pl. 2, fig. 3; *Koelreuteria viridifluminis* (Hollick) R. W. Br., 62, pl. 10, fig. 10. 1934; *K. nigricans* (Lesq.) R. W. Br. in Manchester (1999), 493, fig. 11A.

Holotype—USNM P1617 (Florissant, Colorado, United States; latest Eocene Florissant Formation; collected by Mr. S. A. Allen and first illustrated by Lesquereux [1878b: pl. 5, fig. 11]).

Other specimens examined—USNM P36692 (Fig. 22), P36691 (Fig. 23), P36695 (Fig. 24), P36688, P36689, P36690, P36693, P36694, and P36695a (Florissant Formation, Florissant,

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Figs. 13–21. *Koelreuteria taoana* sp. nov. and *K. dilcheri* sp. nov. from the Eocene of far eastern Russia, northeastern China, and western United States. Figs. 13–16. *K. taoana*. **13–15**. An elliptic capsular valve from the middle Eocene Huadian Formation of Jilin Province in northeastern China. Arrows refer to a septum only in the lower part. The line drawing shows irregularly polygonal and transversely elongated nets. Arrow indicates a proximal septum. Holotype: PE 054018a, b, c. **16**. Line drawing of ovoid-elliptic capsular valve from the middle Eocene Uglovskaya Formation of South Primorye, Russia (based on Ablaev, 1978: pl. 10, fig. 5). Figs. 17–21. *K. dilcheri*. **17–18**. Elliptic capsular valve (and its line drawing, based on Brown, 1946: fig. 1 and republished with permission of the Washington Academy of Sciences) from the middle Eocene of Independence Hill near Colfax, California. Holotype: USNM P42363. **19–21**. Larger, oblong capsular valves from the early to middle Eocene of Republic, Washington. UWBM PB94575, PB1695, and PB1696. Arrow refers to a lower, thicker carpellary suture. Bars = 2 cm.

Colorado); UF 256-20740 (Fig. 25), 256-20745 (Fig. 26) (John Day Formation, Teater Road, Oregon); FMNH PP55094 (Fig. 27) (Green River Formation, Wyoming); YPM PB158849 (Fig. 28) (Florissant Formation, Florissant, Colorado) and PB020681 (Fig. 29) (Green River Formation, De Beque, Colorado).

Emended description-Fossils are preserved as individual (Figs. 22-26), two connected capsular valves (Fig. 28), or complete capsules (Figs. 27, 29), about 3-5.5 cm long and 1.8-2.4 cm wide. Capsules appear to be inflated in contour, with a short pedicel and an acuminate style (fused when the capsule is indehiscent) (Fig. 29). Pedicels are ca. 3-4 mm long and bear perianth scars (Figs. 27–29), implying that the flower of its parent plant is hypogynous. Placentation is axile, with 2-4 seeds at a single level of placental sutures in each capsule (Figs. 27, 29). Seeds are globose (Figs. 23-25, 27-29), ca. 4-6 mm in diameter. The seed surface appears to be smooth (Fig. 25), irregularly furrowed or ridged (Figs. 23, 24, 27-29). Capsules are threevalved and loculicidal, with a septum in the proximal position of each valve (Fig. 29). The septum extends approximately half or slightly more than 1/2 of the valve length from the base, about 3 mm high (Figs. 22, 29). Individual capsular valves are

slightly asymmetrical, obovate or obovoid-elliptic, with a prominent, entire margin. The apex is rounded or slightly emarginate, or mucronate, representing a partial style. The base is narrowly cuneate or decurrent, and slightly asymmetrical (Figs. 22–26). A longitudinal, middle carpellary suture is prominent and appears to be thicker in the lower part than the upper one (Fig. 28). Lateral veins are sinuous and diverge at about 30° – 90° and extend directly to join a fimbrial vein at the margin. The veins are branched and connected into polygonal areoles that are irregular and transversely elongated in outline (Figs. 22, 23, 25, 26, 28, 29). The texture of valves is membranous.

Comments—Koelreuteria allenii was originally instituted for an isolated capsular valve fossil under the name *Ophioglossum alleni* by Lesquereux (1873) from the latest Eocene Florissant Formation, Colorado, but was subsequently used for detached leafy shoots, leaves, leaflets, and capsular valves (MacGinitie, 1953). These leafy shoot fossils are preserved fairly well, which may allow us to make a detailed comparison with those of living *K. paniculata*, both of which bear extremely similar odd-pinnate leaves and dissected leaflets. However, these leaves and capsular valves do not show any organic



Figs. 22–29. *Koelreuteria allenii* (Lesq.) W. N. Edwards from the Eocene of western USA. 22–24. Obovoid-elliptic capsular valves from the late Eocene Florissant Formation of Colorado. Black arrow in Fig. 22 refers to a lower-half septum. White arrows in Fig. 24 refer to two globose seeds, showing some irregular ridges on the surface. USNM P36692, P36691 and P36695. 25–26. Obovoid-elliptic capsular valves from late Eocene John Day Formation of Teater Road, Oregon. UF 256-20740 and 256-20745. 27. Complete capsule from early Eocene Fossil Butte Member of the Green River Formation of Wyoming. FMNH PP55094. 28. Two united capsular valves from the late Eocene Florissant Formation of Colorado. YPM PB158849. 29. Complete capsule from the middle Eocene Green River Formation near De Beque, Colorado. Arrows refer to proximal septa. YPM PB020681. Bars = 2 cm.

connections, so we here use *K. allenii* only for reliable fruit remains. Associated leaves and leaflets may require another name, perhaps using the epithet *hilliae* from Lesquereux (1883) who described the foliage as *Rhus hilliae* Lesq. *Koelreuteria allenii* occurred in the Rocky Mountain region, ranging from the Early Eocene (ca. 52 Ma) of the Fossil Butte Member, through the Middle Eocene Parachute Creek Member of the Green River Formation, Utah and Colorado, to the latest Eocene (ca. 34 Ma) Florissant Formation, Colorado. It was also discovered in the late Eocene John Day Formation of Teater Road, Oregon. Overall, *K. allenii* appears to have existed during an interval of about 18 Ma in the Eocene of western USA (Fig. 48).

Species—Koelreuteria macroptera (Kováts) W. N. Edwards (Figs. 30–38).

Koelreuteria macroptera (Kováts) W. N. Edwards (1927), 112; Kirchheimer (1943), 404, text-fig. 11; Kirchheimer (1957), 206, pl. 33, fig. 143a, b; Rásky (1958), 184, pl. 17, figs. 10, 11; Rüffle (1963), 237, pl. 12, figs. 28–31, pl. 13, figs. 1–5, pl. 27, fig. 9; Schmidt (1976), 54; Schmidt (1980), 15; Shvareva (1983), 135, pl. 79, fig. 6, text-fig. 33.3; Csaki and Urlichs (1985), 31; Gregor (1982), 113, pl. 7, fig. 14; Gregor (1986), 11; Gregor et al. (1992), 21, pl. 3, fig. 2; Seehuber (2009), 239.

Basionym—*Ptelea macroptera* Kováts (1856) in Arbeiten der Geologischen Gesellschaft für Ungarn 1: 51, pl. 1, fig. 2.

Synonym—Dalbergia reticulata Ettingsh., Ettingshausen (1854), pro parte, 813, pl. 4, fig. 5 (non fig. 6); Salvinia reticulata (Ettingsh.) Heer (1857), 156, pl. 145, fig. 16; Brabenec (1904), 2, fig.1, a-b; Phyllites reticulata (Ettingsh.) Florin (1919), 255; Koelreuteria ? reticulata (Ettingsh.) W. N. Edwards in Edwards and Wonnacott (1928), 39; Abronia bronni (Unger) L. Laurent in Weyland (1937), pro parte, 88, pl. 11, fig. 2, textfig. 17 (non fig. 3, text-figs. 18, 19); K. reticulata (Ettingsh.) R. W. Br., Brown (1946) 350; Koelreuteria sp. in Weyland (1948), 133, pl. 21, fig. 6; Kirchheimer (1957), 206; Sapondaceacarpum lunulatum Andreánszky (1959), 156, pl. 47, fig. 5, text-fig. 183; K. reticulata (Ettingsh.) W. N. Edwards in Rásky (1958), 184, pl. 17, fig. 9; Bůžek (1971), 84, pl. 31, figs. 22, 23; Teodoridis (2002), 122, pl. 7, fig. 9; Kvaček et al. (2004b), 94, figs. d, e (non figs. b, c); Winterscheid (2006), 159, pl. 16, fig. 4; Koelreuteria sp. aff. K. integrifolia Franch. in Givulescu (1961), 103, fig. 8; Givulescu (1962), 160, pl. 37, fig. 263.

Holotype—BP 58.2.1 (Fig. 30) (Gomboska of Tállya, Hungary; middle Miocene; part and counterpart specimens first illustrated by Kováts [1856: pl. 1, fig. 2] and Rásky [1958: pl. 17, fig. 11], respectively).

Other specimens examined—BP 60.815.1 (Fig. 31) (Abaújszántó, Hungary); SMNS P1224/509 (Fig. 32), P1224/506 (Fig. 33), P1224/511 (Fig. 34), P1224/515 (Fig. 35), P1224/508 (Fig. 36), P1224/507, P1224/510 and P1224/512 (Randecker Maar, Germany); NMA 92-15/413, 92-17/413 (Fig. 37) (Randecker Maar, Germany), 91-2048/800 (Burtenbach, Germany), and 91-2518/403 (Geisenhausen, Germany).

Emended description—Fossils are preserved as individual capsular valve impressions. Capsular valves are entire-margined, oblong-ovate (Figs. 30, 31, 33), wide or narrow ovate (Figs. 32, 34–38), about 3.4–5 cm long and 2.5–4 cm wide. The base is rounded, slightly truncate, cordate or concave-convex

(Figs. 33–37). The apex is rounded, acute, slightly emarginate, or mucronate, representing a partial style. A longitudinal, middle carpellary suture is prominent, with a septum in the proximal position of valves (Figs. 32, 34, 36). The septum extends approximately 1/3 or slightly more than 1/3 of the valve length from the base, about 5 mm high. There are two seeds attached at a single level near the upper part of the placental suture on the septum (Figs. 30, 31, 33). The seeds are globose, ca. 5–7 mm in diameter. The seed surface appears to have some irregular furrows (Fig. 30). Lateral veins are straight or sinuous and diverge at about 30°–90° (or even 110° at the base) and arrive at the margin. The veins are branched and connected into polygonal areoles that are irregular and transversely elongated in outline (Figs. 36, 38). The texture of valves is membranous.

Comments-Some of the specimens presented here from Central Europe were previously described as Koelreuteria reticulata (Ettingsh.) W. N. Edwards (Rásky, 1958; Bůžek, 1971; Teodoridis, 2002; Kvaček et al., 2004b), but its basionym Dalbergia reticulata Ettingsh. (Ettingshausen, 1854) was based on two heterogeneous specimens. One was actually a legume fruit fossil (Kováts, 1856; Bůžek, 1971), and the other was a capsular valve fossil, both of which are missing. Also, Dalbergia reticulata Ettingsh. is a senior homonym of an extant legume species name Dalbergia reticulata Merr. (Merrill, 1915), so D. reticulata Merr., unless conserved, is illegitimate (McNeill et al., 2012; Wang, 2013). Considering similar characters (in illustrations and specimens) and geographic distribution, we here treat the name K. reticulata, previously used for those isolated capsular valves, as a synonym of K. macroptera (Kováts) W. N. Edwards. Moreover, we exclude one specimen formerly placed under the name Phyllites sp. (Kasumova, 1966: pl. 12, fig. 5) from the middle Oligocene of Indžačaj River, Azarbajdžan, which was assigned to Koelreuteria by Bůžek (1971) because of its larger size (ca. 8-10 cm long) and brochidodromous venation, which are inconsistent with Koelreuteria. Overall, K. macroptera was distributed in the late Oligocene to early Pliocene of Germany, Hungary, the Czech Republic, Romania, and Ukraine (Fig. 48).

Species—Koelreuteria miointegrifoliola Hu et R. W. Chaney emend. (Figs. 39–47).

Koelreuteria miointegrifoliola Hu et R. W. Chaney, Hu and Chaney (1938), pro parte, 64, pl. 38, fig. 1 (non pl. 38, fig. 3 nec pl. 39, fig. 3) (the epithet *'miointegrifolia'* corrected by Hu and Chaney, who suggested this fossil species similar to a living species *K. integrifoliola* Merr., 1922); Huzioka and Takahashi (1973), 141, pl. 4, figs. 2, 3; Onoe (1974), 52, pl. 12, fig. 2 (non fig. 4); Hayashi (1975), 26, pl. 21, fig. 3, pl. 22, figs. 2, 4 (non pl. 22, figs. 6, 8); WGCPC (1978), 131, pl. 116, figs. 5, 6 (non pl. 117. fig. 2); Ozaki (1980), 24, pl. 4, fig. 1 (non pl. 4, figs. 2, 7); Tao et al. (1999), 46, 76, non pl. 35, fig. 1 nec pl. 43, fig. 3.

Synonym—*Koelreuteria sp.* in Sze (1954), 80, pl. 68, fig. 5; *K. macrocarpa* J. R. Tao in Tao et al. (1999), 46, 75, pl. 43, fig. 2.

Lectotype—PB No. 58 (H) (Figs. 45, 46) [designated here; Shanwang, Shandong Province, the Miocene Shanwang Formation of eastern China; first illustrated by Hu and Chaney (1938: pl. 38, fig. 1)].

Other specimens examined—UF 15839-56774, 15839-56774' (part and counterpart specimens, Figs. 39–42), and 15839-56775 (Fig. 44) (Shanwang, China); PE 50384 (Fig. 43),



Figs. 30–38. *Koelreuteria macroptera* (Kováts) W. N. Edwards from the Oligocene–Miocene of Hungary and Germany. **30.** Incomplete capsule valve from the Miocene of Gomboska at Tállya, Hungary. Arrows indicate two seeds attached on either side of the suture at a single level. Holotype: BP 58.2.1. **31.** Ovate capsular valve from the Miocene of Abaújszántó, Hungary. Arrows indicate two seeds, with some irregular furrows on the surface. BP 60.815.1. **32.** Ovate capsular valve from the Miocene of Randecker Maar, Germany. Arrow refers to a proximal septum. SMNS P1224/509. **33.** Wide ovate capsular valve from the Miocene of Randecker Maar, Germany. Arrow refers to a proximal septum. SMNS P1224/509. **33.** Wide ovate capsular valve from the Miocene of Randecker Maar, Showing two globose seeds. SMNS P1224/506. **34–37.** Ovate capsular valves from the Miocene of Randecker Maar, showing a proximal septum (arrows). SMNS P1224/515, P1224/508 and NMA 92-17/413. **38.** Line drawing of ovate capsular valve from the late Oligocene of Rott, Germany (based on Weyland, 1937: pl. 11, fig. 2). Bars = 2 cm.

50567, 52440, 50455 (Fig. 47), 20111117, 20111118A, B, and 20080916 (Shanwang, China).

Emended description—Fossils are preserved as individual capsular valve impressions. Capsular valves are entire-margined, wide ovate, oblong, elliptic or suborbicular (Figs. 39, 40, 43-47), 3.2-5.2 cm long and 3.0-4.2 cm wide. The base is rounded, slightly truncate or cordate, with a short pedicel (Figs. 39, 43), about 2–3 mm long. The apex is rounded, slightly emarginated, or mucronate, representing a partial style. A longitudinal, middle carpellary suture is prominent and appears to be thicker in the lower part than in the upper part. The septum is proximal and incomplete above the placental suture attached seeds, extending from the base to approximately half of the valve length, about 4 mm high (Figs. 39-42). Two seeds are suboppositely attached at a single level on both sides of approximately the middle part of the longitudinal central placental suture in each valve. Seeds are grossly globose, ca. 4-9 mm in diameter, with a short funiculus (Fig. 41), ca. 2 mm long. The seed surface appears to have some irregular furrows (Figs. 41, 44). The placental suture is curved slightly and diverges into numerous, lateral sinuous veins at 30°-150° through the carpellary suture, arriving at the margin (Figs. 42-47). The lateral veins are branched and connected into polygonal areoles, which are irregular and transversely elongated in outline. Areoles are moderately developed, with branched freely ending veinlets (Fig. 42). The texture of valves is membranous.

Comments-Koelreuteria miointegrifoliola Hu et R. W. Chaney, as a sapindaceous fossil, was originally based on three specimens, which Hu and Chaney (1938) designated as "cotypes" (i.e., syntypes) (PB No. 58 [H] and No. 56 [H], referring to a capsular valve and a leaflet, respectively) and the "paratype" (PB No. 57 [H], referring to a leaflet), from the Miocene Shanwang Formation of Shandong Province, eastern China. The two leaflets were subsequently reidentified as the species of Juglans L. in the Juglandaceae A. Rich. ex Kunth (Brown, 1946; Sze, 1951; Rüffle, 1963; WGCPC, 1978). Although additional leaflets from Shanwang and some Japanese localities (Table 2) continued to be described as K. miointegrifoliola (Onoe, 1974; Hayashi, 1975; WGCPC, 1978; Ozaki, 1980, 1991) or K. macrocarpa J. R. Tao (Tao et al., 1999), there is no evidence of an organic connection between leaflets and capsular valves, nor is there a detailed leaf architectural analysis to reliably assign the leaflets to *Koelreuteria*. Thus, we here use *K. miointegrifoliola* only for the sapindaceous capsular valves, which were distributed in the Miocene of eastern China and Japan (Fig. 48).

DISCUSSION

Hypotheses on the origins of the Sapindaceae—The Sapindaceae is a pantropical angiosperm family of trees, shrubs, lianas, and rarely herbaceous climbers comprising 142 genera and ca. 1900 species (Buerki et al., 2009, 2010; Acevedo-Rodríguez et al., 2011). This family, as currently defined by molecular systematics, encompasses both Aceraceae Juss. and Hippocastanaceae Rich. and is widely distributed in tropical and subtropical regions and is especially well represented in tropical Southeast Asia, but a few genera (e.g., *Acer L., Dodonaea*, and *Koelreuteria*) have species adapted to temperate regions (Xia and Luo, 1995; Harrington et al., 2005; Xia and Gadek, 2007; Buerki et al., 2009, 2010). Recent biogeographic analysis inferred that Sapindaceae originated in Eurasia around the Early Cretaceous (Buerki et al., 2011). However, paleopalynological evidence seems to support that Sapindaceae made their debut in the Late Cretaceous (Coniacian–Santonian) of the paleotropics (i.e., an African–Indian origin, see Muller, 1981; Taylor, 1990; Song et al., 1999, 2004). The earliest fossils bearing finely wrinkled or longitudinally striated extexine ornamentations similar to the simply structured tricolporate pollen (i.e., type-A pollen, sensu Muller and Leenhouts 1976) (Fig. 8) of extant *Koelreuteria* have been reported from the Late Cretaceous to the Eocene of southern Laurasian margin (Tibet) (Song and Li, 1982; Li et al., 2008), South China (Song et al., 1999, 2004) (Table 2; Fig. 48). Therefore, microfossil evidence seems to support a southern Laurasian origin for *Koelreuteria*.

Early differentiation of sapindaceous wood with ring porosity—The wood anatomical literature indicates that the earliest fossil record of Sapindaceae as Sapindoxylon Kräusel is reported from the Late Cretaceous Deccan Intertrappean Beds, India (Dayal, 1965; Chitaley and Shallom, 1969; Friis et al., 2011). Sapindoxylon lacks growth rings and has been reported from numerous tropical localities in the Tertiary of Africa, India, and Southeast Asia (Kräusel, 1922; Bande and Prakash, 1986; Dupéron-Laudoueneix and Dupéron, 1995) (Fig. 48) and in the Eocene of southern England (Wilkinson, 1988) and western United States (Wheeler and Manchester, 2002). By the latest Eocene, ring porous woods of Koelreuteria have been described from the Florissant Formation of Colorado, western United States (Wheeler, 2001). Ring porosity in woody angiosperms is a derived character in response to the climatic conditions characteristic of the North Temperate Zone (Gilbert, 1940) and may be considered a reliable ecological marker after the early Tertiary (Wheeler and Baas, 1993; Baas et al., 2003). Hence, anatomical evidence supports the suggestion that the sapindaceous plants bearing Koelreuteria-type wood with ring porosity appear to have begun to differentiate in the northern hemisphere since the latest Eocene.

Ecological implications for the fruits and seeds of Koelreuteria-Many sapindaceous genera bear schizocarpous, winged fruits such as Acer and Dipteronia Oliv., which are ecological adaptations for wind dispersal in the North Temperate Zone (Manchester and O'Leary, 2010), and are thus functionally similar to the bladder-like capsules of Koelreuteria. Acer and Dipteronia have been reported from North America since the Paleocene (Manchester, 1999; McClain and Manchester, 2001), reflecting ecologically adaptive radiations and deep phylogenetic splits between the temperate (e.g., mixed mesophytic forest) and tropical (e.g., evergreen broad-leaved forest) biomes or lineages that occupy temperate or tropical regions. In contrast with the more ovoid seeds found in some of the other sapindaceous genera (Table 3), the perfectly globose seeds of Koelreuteria are blown from the high trees and still contained in their light, membranous, bladder-like capsules, so they are dispersed by wind or sometimes by water. Furthermore, such small, shiny, nutrition-rich seeds of Koelreuteria have attracted animal dispersants including birds (Lin et al., 2011). Hence, birds may also play a key role in long-distance seed dispersal of Koelreuteria.

Evolutionary and biogeographic implications for the fruits of Koelreuteria—The reliable fossil distribution of *Koelreuteria*



Figs. 39–47. *Koelreuteria miointegrifoliola* Hu et R. W. Chaney from the Miocene of Shanwang, Shandong Province, eastern China. **39**, **40**. An oblong capsular valve (part and counterpart), showing a detached seed (a), a placental suture (b) and a carpellary suture (c). UF 15839-56774 and 15839-56774'. **41**, **42**. Partial enlargements of specimens in Figs. 39 and 40, indicating a detached globose seed, with some irregular ridges on the surface (a), two short funiculi (b and c). **43**. A suborbicular capsular valve. PE 50384. **44**. Wide ovate capsular valve. Arrow indicates a detached seed with some irregular ridges or furrows on the surface. UF 15839-56775. **45–46**. Elliptic capsular valve and its line drawing. Lectotype: PB No. 58 (H). **47**. Elliptic capsular valve. PE 50455. Bars = 2 cm.



Fig. 48. Map and stratigraphic chart showing the diversity and distribution of *Koelreuteria* Laxm., with references to Tables 1 and 2. Red dots refer to *Sapindoxylon*-type wood fossil localities (Kräusel, 1922; Dayal, 1965; Chitaley and Shallom, 1969; Bande and Prakash, 1986; Wilkinson, 1988; Poole and Wilkinson, 1992; Dupéron-Laudoueneix and Dupéron, 1995; Wheeler and Manchester, 2002). Orange dots refer to *Koelreuteria*-type wood fossil localities (Wheeler, 2001; Cheng et al., 2012). Pink dots refer to *Koelreuteria*-like pollen fossil localities (Song and Li, 1982; Song et al., 1999; Leopold and Clay-Poole, 2001; Li et al., 2008).

capsules shows three areas of concentration: eastern Asia, Central Europe, and western North America (Fig. 48). The oldest known specimen of a capsular valve of *Koelreuteria* is *K. allenii* from the early Eocene Fossil Butte Member of the Green River Formation, Wyoming, which places a minimum age (ca. 52 Ma) for the divergence of *Koelreuteria* from presumably tropical genera that occupy more basal positions in molecular phylogenetic trees of Sapindaceae. *Koelreuteria allenii* persisted from the middle Eocene Parachute Creek Member of Green River Formation, Utah and Colorado to the late Eocene (ca. 34 Ma) Florissant Formation, Colorado in the Rocky Mountain region and the John Day Formation of Teater Road, Oregon.

It is remarkable that K. allenii, K. taoana, and K. dilcheri have occurred in Eocene mid-latitudes (ca. 40°-45°) of far eastern Russia, northeastern China, and western United States. Except for valve size, both K. taoana and K. dilcheri are typically elliptic and have occurred around the North Pacific Rim areas-an indication of Eocene floral links between eastern Asia and western North America via the Bering land bridge (Tiffney and Manchester, 2001; Milne and Abbott, 2002) and/ or Aleutian land bridge (Lindroth, 1963). Koelreuteria, as well as the Aceraceae, Betulaceae Gray, Menispermaceae Juss., Vitaceae Juss., and Pinaceae Lindl., which are common in western North American Eocene localities, are noticeably absent from the coeval localities in southeastern North America. Such floristic differences may have resulted from the dispersal barrier represented by the Cannonball Epeiric Sea, that traversed some parts of the Midcontinent in the Paleocene (Leopold et al., 2008) and environmental heterogeneity (Manchester, 1999). Moreover, the Eocene biotas of East Asia show closer relationships to those of North America than to Europe (Budantsev, 1992; Manchester et al., 2005). On the basis of the differences in capsular valve morphology, either there may be an exchange and rapid diversification into different types or there seems to have been no exchange between Koelreuteria species in western North America and Europe via the North Atlantic land bridge (Tiffney and Manchester, 2001; Milne and Abbott, 2002). Koelreuteria may have become regionally extinct in western North America after the Eocene or the early Oligocene. The fossil record of *Koelreuteria* in the late Oligocene of Germany (Weyland, 1937; Kirchheimer, 1957; Winterscheid, 2006) (Fig. 38) and the Czech Republic (Teodoridis, 2002) implies that populations in North America before extinctions invaded Europe via the North Atlantic land bridge, while a direct floristic exchange between the northern central regions of Asia and Europe may have been impeded by the West Siberian Sea and Turgai Strait (Akhmetiev and Beniamovski, 2009) during the early Paleogene. Alternatively, populations of Koelreuteria in northeastern Asia migrated westward into Europe with the closure of the Turgai Strait during the Oligocene. By the Miocene, Koelreuteria had been restricted gradually to the middle latitudes of Eurasia.

Among the four extant species of *Koelreuteria* now restricted to eastern Asia and Fiji, fruit characters and pollen morphology as well as karyotype analyses (Urdampilleta et al., 2005; Zhang and Wang, 2009) suggest that *K. bipinnata* and *K. henryi* are more closely related to each other than either is to *K. paniculata*. The capsular valve fossils (*K. allenii*, *K. taoana*, *K. dilcheri*, and *K. miointegrifoliola*) from the Eocene to Miocene of eastern Asia and western North America appear to belong to the *K. bipinnata*-type, which usually bears a septum extending 1/3 to half of the valve length from the base. In contrast, the

capsular valve fossils (K. macroptera) from the late Oligocene to early Pliocene of Europe are the *K. paniculata*-type, bearing a proximal septum only about 1/3 of the valve length. This distribution suggests that a derivative of *K. macroptera* may have migrated eastward into eastern Asia in the Miocene, evolving into the extant K. paniculata. Extant K. bipinnata and *K. henryi* may stem from a vicariance event involving the separation of Taiwan from eastern Asian continent after the Miocene. Overall, the fruit fossil record of Koelreuteria indicates that this genus may have had a biogeographic origin and rapid spread in the northern hemisphere and around the North Pacific Rim areas during the Eocene. With the regional extirpations after the Eocene in North America and the Pliocene in Europe, Koelreuteria became restricted gradually to eastern Asia. Therefore, eastern Asia is a living museum for Koelreuteria.

On the basis of Koelreuteria shared by eastern Asia and Fiji, Wu et al. (2007) suggested that Fiji was linked with Laurasia. Proximal to the Australian-Pacific Plate boundary, the Fiji Platform has been active since at least the late Eocene, during which the oldest island-arc volcanic rocks were formed in Fiji (Stratford and Rodda, 2000; Neall and Trewick, 2008) and the earliest populations of Koelreuteria were developed around the North Pacific Rim areas. So far, neither fossil nor extant wild populations of Koelreuteria have been discovered on the tropical islands between eastern Asia and the Fiji Islands (Smith, 1978, 1985; Bande and Prakash, 1986). Hence, the present distribution of K. elegans in South Pacific Oceania might have been derived from an ancient chance dissemination ("waif introduction" from the west) as some authors have hypothesized (Smith, 1978; Heads, 2006) or resulted from a historical long-distance dispersal by migrating birds (e.g., Carlquist, 1967; Proctor, 1968; Higgins et al., 2003) between eastern Asia and the Fiji Islands, which appears to have failed to succeed in dispersing to intervening land areas.

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APPENDIX 1. Information on the specimens used in this study.

Taxon	Number	Preservation	Locality	Age	Repository
Boniodendron minus (Hemsl.) T. C. Chen	1111426, 0923579	Herbarium	Ruyuan, Guangdong, China	Recent	PE
	1000957	Herbarium	Douan, Guangxi, China	Recent	PE
Bridgesia incisifolia Bertero ex Cambess.	909993	Herbarium	Coqumbo, Chile	Recent	MO
Craigia yunnanensis A. C. Sm. et W. E. Evans	02157045	Herbarium	Tengchong, Yunnan, China	Recent	PE
	02145497	Herbarium	Malipo, Yunnan, China	Recent	PE
Dodonaea viscosa (L.) Jacq.	1886617, 1234058	Herbarium	Yongsheng, Yunnan, China	Recent	PE
	02174471, 02174472	Herbarium	Haikang, Guangdong, China	Recent	PE
	Grierson & Long 4187	Herbarium	Tongsa District, Bhutan	Recent	А
Koelreuteria allenii (Lesq.) W. N. Edwards	P1617, P36692, P36691, P36695, P36688, P36689, P36690, P36693, P36694, P36695a	Impression	Florissant, Colorado, USA	Latest Eocene	USNM
	256-20740, 256-20745	Impression	Teater Road, Oregon, USA	Late Eocene	UF
	PP55094	Impression	Wyoming, USA	Early Eocene	FMNH
	PB158849	Impression	Florissant, Colorado, USA	Latest Eocene	YPM
	PB020681	Impression	De Beque, Colorado, USA	Middle Eocene	YPM
Koelreuteria bipinnata Franch.	0991406, 0991407	Herbarium	Shangyou, Jiangxi, China	Recent	PE
	02174429, 297889	Herbarium	Hengshan, Hunan, China	Recent	PE
Koelreuteria dilcheri sp. nov.	P42363	Impression	Independence Hill near Colfax, California, USA	Middle Eocene	USNM
	PB94575, 1695, 1696	Impression	Republic, Washington, USA	Early to middle Eocene	UWBM
Koelreuteria elegans (Seem.) A. C. Sm. Koelreuteria henryi Dümmer	1213, B. E. Parham 1516597, 1583132	Herbarium Herbarium	Tailevu, Viti Levu, Fiji Hualian and Taoyuan, Taiwan, China	Recent Recent	A PE
	20100203	Slide	University of Florida, Gainesville, USA	Recent	UF
Koelreuteria macroptera (Kováts) W. N. Edwards	58.2.1	Impression	Gomboska of Tállya, Hungary	Middle Miocene	BP
	60.815.1	Impression	Abaújszántó, Hungary	Middle Miocene	BP
	P1224/509, P1224/506, P1224/511, P1224/515, P1224/508, P1224/507, P1224/510, P1224/512	Impression	Randecker Maar, Germany	Early Miocene	SMNS
	92-15/413, 92-17/413	Impression	Randecker Maar, Germany	Early Miocene	NMA
	91-2048/800, 91-2518/403	Impression	Burtenbach and Geisenhausen, Bavaria, Germany	Middle to late Miocene	NMA
Koelreuteria miointegrifoliola Hu et R. W. Chaney	No. 58 (H)	Impression	Shanwang, Shandong, China	Early to middle Miocene	РВ
	50384, 50567, 52440, 50455, 20111117, 20111118A, 20111118B, 20080916	Impression	Shanwang, Linqu, Shandong, China	Early to middle Miocene	PE
	15839-56774, 15839-56774´, 15839-56775	Impression	Shanwang, Linqu, Shandong, China	Early to middle Miocene	UF
Koelreuteria paniculata Laxm.	0894897	Herbarium	Yuxian, Henan, China	Recent	PE
	0872783, 02234144	Herbarium	Taian, Shandong, China	Recent	PE
Koelreuteria taoana sp. nov.	054018a, b, c	Impression	Huadian, Jilin, China	Middle Eocene	PE
Urvillea chacoensis Hunz.	T. Meyer 8829	Herbarium	Argentina	Recent	А

Notes: A: Arnold Arboretum Herbarium, Harvard University; BP: Hungarian Natural History Museum, Budapest; FMNH: Field Museum, Chicago; MO: Missouri Botanical Garden Herbarium; NMA: Nature Museum Augsburg, Augsburg; PB: Nanjing Institute of Geology and Palaeontology, Chinese Academy of Sciences, Nanjing; PE: Chinese National Herbarium, Beijing; SMNS: Staatliche Museum für Naturkunde, Stuttgart; UF: Florida Museum of Natural History, Gainesville; USNM: National Museum of Natural History, Smithsonian Institution, Washington, D.C.; UWBM: Burke Museum of Natural History, the University of Washington, Seattle; YPM: Peabody Museum of Natural History, Yale University, New Haven.