

Figure 3. Lycosa sp.

Poecilochroa, Bavia, and a male Athamas which I took back alive to be photographed. That night I succeeded in getting a lycosid.

In the few days after arriving back on Rarotonga we did some additional collecting, but did not get much that was new. We did find, in curled dead leaves hanging in trees, a number of salticids of a species we did not find in 1987. This may be a recent import, or our previous visit may have been at a time when its population was very low.

On one of my stays on Rarotonga I had noticed a salticid on the plants around the motel, but had failed to catch it. After I had made several attempts it simply disappeared. Some time later, while waiting to be served dinner at the Club Raro, I found it walking on my clothing, and this time caught it. It seems to be *Avarua satchelli*, originally described from here by Marples, and one of the few species we had missed in 1987. This species has since been transferred by Jerzy Proszynski to the genus *Trite*, but it may not belong to that rather heterogeneous genus either.

I left Rarotonga about a week earlier than originally planned. Many of the things that still need to be done will have to be done at home, where I have more resources. I changed my airline ticket to go to Hawaii for a week, where a graduate student had asked for my help with spiders he collected there. I spent a week working at the Bishop Museum and returned home.

When I examined the collection in Carbondale I realised that I had been more successful than I had thought, having collected about 50 of the Cook Island species previously known, and added four species and two families not recorded before. Except for some of the small litter spiders, much of what I had missed consisted of very uncommon species.

Two things I noticed that may be useful to anyone making a similar trip. At least at my age, working alone was not much fun. Either having another person accompanying you or making better contact with the local people who know the terrain would be very useful. Better access to transport is also very desirable. On Rarotonga and Aitutaki rental cars and motor scooters are common, but on the other islands they are few. Arrangements ahead of time to be sure that transport is available would be very helpful. While it is possible in some of these places to rent bicycles, our experience in 1987 indicated that in such a climate one is rather worn out before getting to a collecting place unless it is very near.

Department of Zoology, Southern Illinois University, Mailcode 6501, CARBONDALE, Illinois, USA

# Preliminary Survey of the Spiders, Harvestmen and False-scorpions of Alderley Edge, Cheshire

# by Dmitri Logunov

In late 2001, I was invited by some colleagues in the Manchester Museum to survey the spider fauna of Alderley Edge, a wooded sandstone escarpment rising some 200 m above the Cheshire plain. About 250 acres of the Edge are now owned by the National Trust and the site was recognised as an important candidate for review under the Monument Protection Programme. It is now covered with secondary closed woods, which were planted in the middle of the eighteenth century and since then have replaced the former heathland. Further information about the peculiar history of the Edge, its legends and the Alderley Edge Landscape Project can be found on two main sites: http://www.derbyscc.org.uk/alderley/index.html.

This short paper outlines the results of a preliminary survey of the arachnid fauna of the Edge carried out by the author assisted by his daughter, Anna Logunova. We made 16 one-day trips to the site in 2002 (late March to mid-October) and used four collecting methods: handpicking, sieving the litter, sweeping, and shaking of twigs and branches of shrubs and low trees (e.g. gorse, hawthorn and others). Pitfall traps were not used. Altogether, 1,283 adult specimens belonging to 114 spider, 11 harvestmen and one false-scorpion species were collected and identified.

The arachnid fauna of UK can be considered well studied; nevertheless, local lists and records remain important for recording schemes (e.g. Harvey *et al.*, 2002). Although most of the species listed below are quite common and widespread, at least two records seem to be worthy of some attention: one of the northernmost UK records of *Clubiona corticalis;* also one of the northernmost records of *Entelecara congenera* in the UK (see Harvey *et al.*, 2002, p. 82).

An interesting case of araneophagy by a theridiid spider was observed in Alderley Edge. On 13th July 2002, I found a female *Theridion impressum* eating an adult female *Larinioides cornutus*. Both species built up their nests on the same stem of the Marsh Thistle (*Cistium palustre*) and occasionally the former species predated the latter one.

All the material was identified by the author, apart from some linyphiids represented by single females, which were treated by Dr Andrei Tanasevitch (Moscow); I am most grateful for his help. The spiders and harvestmen collected during this study have been divided between the Manchester Museum and the Zoological Museum of Moscow State University.

# References

Harvey, P. R., Nellist, D. R. & Telfer, M. G. (eds). (2002) Provisional Atlas of British Spiders (Arachnida, Araneae). Volumes 1 & 2. Biological Records Centre, Huntingdon.

Platnick, N. I. (2002) The World Spider Catalog, Version 3.0. American Museum of Natural History, New York. http:// research.amnh.org/entomology/spiders/catalog81-87/index.html.

The Manchester Museum, The University, MANCHESTER, M13 9PL; e-mail: dmitri.v.logunov@man.ac.uk

In the **species list** (right), numbers in square brackets refer to habitats: 1- in crevices of sandy gorges; 2- under bark of fallen trees; 3- rocks (under dry ferns and grass clumps, in litter, moss, lichen); 4- secondary forb meadow; 5- gorse shrubs (shaking off from twigs); 6- damp meadow with rushes; 7- heather on stony slope; 8- broad-leaved (holly-oak-beech) wood (in litter and on ferns); 9- moss-heather-bilberry heath (in litter); 10- open sandy heath (with sparse grassy vegetation); 11- pine forest (in litter and on ferns); 12- damp birch forest (under rotten logs and in litter).

# **SPIDERS**

#### Family AGELENIDAE

- Tegenaria duellica Simon, 1875: 49, 6.04–14.06, [1, 2]
- Family AMAUROBIIDAE
- Amaurobius fenestralis (Stroem, 1768): 50 339, 30.03–16.10, [2, 3] Amaurobius similis (Blackwall, 1861): 29, 17.04–14.06, [2]
- Family ARANEIDAE

- Family AKANELDAE Araneus diadematus Clerck, 1757: 2Q, 24.09, [4, 5] Araneus marmoreus Clerck, 1757: 3Q, 24.09, [4] Araneus quadratus Clerck, 1757: 2Q, 24.09–16.10, [4] Cyclosa conica (Pallas, 1772): 1ơ, 19.05, [6] Larinioides cornutus (Clerck, 1757): 3Q, 13.07–24.09, [4] Nuctenea umbratica (Clerck, 1757): 1 juv, Q, 17.04, [2] Zygiella atrica (C. L. Koch, 1845): 2ơ 1Q, 24.09–16.10, [5] Zygiella atrica (Clerck, 1757): 1Q, 24.09, [6] Examity CULIBLONIDAE

### Family CLUBIONIDAE

- Clubiona comta C. L. Koch, 1839: 33 39, 17.04–13.07, [7, 8] Clubiona corticalis (Walckenaer, 1802): 13 19, 14.06, [2] Clubiona diversa O. Pickard-Cambridge, 1862: 13 29, 19.05–7.07, [9, 10]
- Clubiona lutescens Westring, 1851: 10 69, 24.04–24.09, [3, 4, 5, 6, 8]
- Clubiona reclusa O. Pickard-Cambridge, 1863: 20 39, 19.05-14.06, [6] Family HAHNIIDAE
- Antistea elegans (Blackwall, 1841): 199, 6.04–16.10, [6] Cryphoeca silvicola (C. L. Koch, 1834): 39, 30.03-14.06, [3, 8, 11] (I follow Platnick (2002) and consider this species a member of Hahniidae.)
- Family LINYPHIIDAE
- Agyneta conigera (O. Pickard-Cambridge, 1863): 19, 3.06, [8] Agyneta rurestris (C. L. Koch, 1836): 10, 7.07, [9] Bathyphantes approximatus (O. Pickard-Cambridge, 1871): 130 199, 6.04-24.09, [6]
- Bathyphantes gracilis (Blackwall, 1841): 40 189, 24.04–16.10, [3, 4, 5, 6, 8, 9, 10]
- 4, 5, 6, 8, 9, 10] Bathyphantes nigrinus (Westring, 1851): 2\$\alpha\$ 6\$\overline{Q}\$, 6.04–14.06, [6, 12] Centromerus dilutus (O. Pickard-Cambridge, 1875): 1\$\overline{Q}\$, 7.07, [9] Centromerus sylvaticus (Blackwall, 1841): 2\$\overline{Q}\$, 10–24.04, [6, 9] Ceratinella brevipes (Westring, 1851): 9\$\overline{Q}\$, 17.04–7.07, [7, 9] Ceratinella brevis (Wider, 1834): 3\$\vee 4\$\overline{Q}\$, 10.04–3.06, [6, 7, 8] Ceratinella scabrosa (O. Pickard-Cambridge, 1871): 1\$\vee\$, 17.04, [8] Cnephalocotes obscurus (Blackwall, 1834): 1\$\vee\$ 1\$\overline{Q}\$, 607, [4] (Com-Collingia inerrons (O. Pickard-Cambridge, 1851): 2\$\vee\$ 0, 607, [4] (Com-

- Collinsia inerrans (O. Pickard-Cambridge, 1885): 29, 6.07, [4] (Combination follows Platnick (2002).
- Dicymbium nigrum (Blackwall, 1834): 10 49, 24.04, [6]
- Diplocephalus cristatus (Blackwall, 1833): 10, 3.06, [8]
- Diplocephalus latifrons (O. Pickard-Cambridge, 1863): 70 119, 17.04-14.06, [6, 8, 12]

- Diplocephalus picinus (Blackwall, 1841): 30 59, 17.04–14.06, [8] Diplostyla concolor (Wider, 1834): 20 49, 6.04–3.06, [8, 11] Dismodicus bifrons (Blackwall, 1841): 50 69, 24.04–6.07, [4, 6] Entelecara congenera (O. Pickard-Cambridge, 1879): 20 109, 13.07– 24.09, [5]
- Erigone atra Blackwall, 1833: 8° 229, 6.04–16.10, [3, 4, 5, 6, 8, 9, 10] Erigone dentipalpis (Wider, 1834): 3° 109, 6.04–16.10, [3, 4, 5, 8, 9, 10]
- 9, 10] Erigone promiscua (O. Pickard-Cambridge, 1872): 19, 19.05, [10] Erigonella hiemalis (Blackwall, 1841): 4& 89, 17.04–7.07, [7, 9, 12] Floronia bucculenta (Clerck, 1757): 1& 29, 24.09–16.10, [6] Gnathonarium dentatum (Wider, 1834): 3& 79, 19.05–16.10, [6] Gonatium rubens (Blackwall, 1833): 19, 17.04, [7] Gongylidiellum vivum (O. Pickard-Cambridge, 1875): 1& 29, 5.05– 7.06, [4, 9, 12] Gongulidium rubines (Lippacus, 1758): 10, 7.07, [9]

- 7.06, [4, 9, 12] Gongylidium rufipes (Linnaeus, 1758): 19, 7.07, [9] Helophora insignis (Blackwall, 1841): 20 69, 24.09–16.10, [8] Hilaira excisa (O. Pickard-Cambridge, 1871): 19, 14.06, [11] Hypomma bituberculatum (Wider, 1834): 40 19, 10.04–19.05, [6] Kaestneria pullata (O. Pickard-Cambridge, 1863): 19, 6.07, [4] Labulla thoracica (Wider, 1834): 79, 30.03–24.09, [3] Lepthyphantes minutus (Blackwall, 1833): 39, 14.06–24.09, [3, 8] Linyphia hortensis Sundevall, 1829: 10 29, 24.04–7.07, [9, 11] Linyphia triangularis (Clerck, 1757): 329, 24.09–16.10, [4, 5, 6] Lophomma punctatum (Blackwall, 1841): 20 29, 19.05–24.09, [6] Maso sundevalli (Westring, 1851): 230 349, 3.06–7.07, [8, 9] Meioneta saxatilis (Blackwall, 1844): 20 19, 19.05–13.07, [5, 6, 9] Micrargus herbigradus (Blackwall, 1854): 40 79, 17.04–6.07, [4, 5, 7, 11]
- 7. 111
- Microlinyphia pusilla (Sundevall, 1830): 50 69, 19.05–7.07, [4, 9, 10] Microneta viaria (Blackwall, 1841): 20 249, 30.03–14.06, [8] Minyriolus pusillus (Wider, 1834): 19, 4.06, [7] Monocephalus fuscipes (Blackwall, 1836): 30 349, 6.04–24.09, [6,

- Monocephatus Juscipes (Elacivali, 1000), 00 011, 120 7, 8, 9, 11, 12] Neriene clathrata (Sundevall, 1830): 50 119, 6.04–24.09, [4, 6, 9] Neriene peltata (Wider, 1834): 10 39, 5.05–7.06, [8, 9] Obscuriphantes obscurus (Blackwall, 1841): 10 39, 4.07–7.07, [5, 9] Oedothorax fuscus (Blackwall, 1834): 150 89, 4.07–13.07, [4, 5, 6, 9.10
- Oedothorax gibbosus (Blackwall, 1841) (including O. g. form tuberosus): 133 329, 19.05–14.06, [6]
- Oedothorax retusus (Westring, 1851): 20 19, 19.0, [6, 10]

5

- Saaristoa abiorinis (blackwali, 1841): 10, 7.07, [9] Saaristoa firma (O. Pickard-Cambridge, 1905): 12, 3.06, [9] Savignia frontata Blackwall, 1833: 10 19, 4.07–16.10, [3, 8] Silometopus elegans (O. Pickard-Cambridge, 1872): 10, 19.05, [10] Tapinocyba pallens (O. Pickard-Cambridge, 1872): 30, 17–24.04, [7, 11]
- [7, 11]
  Tenuiphantes alacris (Blackwall, 1853): 20 39, 10–24.04, [8, 9, 11]
  Tenuiphantes cristatus (Menge, 1866): 10 69, 10.04–24.09, [6, 8, 9]
  Tenuiphantes flavipes (Blackwall, 1854): 40 39, 24.04–24.09, [3, 6, 8]
  Tenuiphantes mengei (Kulczynski, 1887): 19, 24.04, [6]
  Tenuiphantes tenebricola (Wider, 1834): 40 69, 6.04–14.06, [8, 9]
  Tenuiphantes tenuis (Blackwall, 1852): 150 139, 6.04–7.07, [4, 5,
- 8, 9, 10]
- Tenuiphantes zimmermanni (Bertkau, 1890): 240 229, 6.04-24.09, [3, 6, 8, 9, 11, 12]

- [3, 6, 7, 9, 11, 12] Thyreosthenius parasiticus (Westring, 1851): 19, 5.05, [12] Tiso vagans (Blackwall, 1834): 19, 24.04, [6] Walckenaeria acuminata Blackwall, 1833: 19, 16.10, [9] Walckenaeria cucullata (C. L. Koch, 1836): 29, 17–24.04, [8, 11]
- Walckenaeria cuspidata Blackwall, 1833: 10, 30.03, [3] Walckenaeria nudipalpis (Westring, 1851): 10, 4.07, [9] Walckenaeria vigilax (Blackwall, 1853): 10, 19.05, [10]

# Family LIOCRANIDAE

- Scotina celans (Blackwall, 1841): 19, 17.04, [7] Family LYCOSIDAE
- Alopecosa pulverulenta (Clerck, 1757): 10 19, 10.04, [6] Arctosa perita (Latreille, 1799): 19 4 juv., 13.07, [10] Pardosa amentata (Clerck, 1757): 60 19, 6.04–14.06, [6, 10] Pardosa palustris (Linnaeus, 1758): 30 69, 19.05, [6, 10] Pardosa pullata (Clerck, 1757): 70 179, 10.04–7.07, [4, 8, 9] Pirata piraticus (Clerck, 1757): 30 29, 19.05–14.06, [6]

- Trochosa terricola Thorell, 1856: 29, 6.04-7.06, [8, 9]

#### Family PISAURIDAE

- Pisaura mirabilis (Clerck, 1757): 39, 6-13.07, [4] Family PHILODROMIDAE
- Philodromus aureolus (Clerck, 1757): 10 59, 4.07-13.07, [5, 9] Philodromus cespitum (Walckenaer, 1802): 29, 13.07, [5] Family SALTICIDAE
- Euophrys frontalis (Walckenaer, 1802): 30 109, 10.04-13.07, [3, 4, 9.101
- Neon reticulatus (Blackwall, 1853): 20 39, 3.06-7.07, [9] Family TETRAGNATHIDAE
- Metellina mengei (Blackwall, 1869): 250 239, 10.04-16.10, [3, 5, 6, 8, 10, 12]
- Metellina merianae (Scopoli, 1763): 30 89, 30.03-14.06, [2, 3, in mines (close to entrances)]
- Metellina segmentata (Clerck, 1757): 370 319, 24.09-16.10, [4, 5, 6, 8]
- Pachygnatha clercki Sundevall, 1823: 22, 24.04–24.09, [4, 6] Pachygnatha degeeri Sundevall, 1830: 10 69, 19.05–14.06 [6, 10] Tetragnatha extensa (Linnaeus, 1758): 60 89, 19.05–13.07 [4, 6]
- Tetragnatha montana Simon, 1874: 20 19, 7.0, [9]

# Family THERIDIIDAE

- Anelosimus vittatus (C. L. Koch, 1836): 19, 13.07, [4] Enoplognatha ovata (Clerck, 1757): 130 519, 14.06–13.07, [4, 5, Enoplognatha ovata (Clerck, 1/5/): 130 514, 14.00–13.07, 14, 5 6, 8, 9] Neottiura bimaculata (Linnaeus, 1767): 19, 14.06, [6] Paidiscura pallens (Blackwall, 1834): 40, 17.04–7.07, [6, 7, 8, 9] Theridion impressum L. Koch, 1881: 40, 39, 6–13.07, [4] Theridion mystaceum L. Koch, 1870: 19, 14.06, [3] Theridion sisyphium (Clerck, 1757): 199, 6–13.07, [4, 5, 9] Theridion tinctum (Walckenaer, 1802): 30, 119, 6.07–24.09, [5] Theridion varians Hahn, 1833: 10, 6.07, [5]

Zora spinimana (Sundevall, 1833): 30 99, 10.04-7.07, [9]

- Xysticus cristatus (Clerck, 1757): 20 99, 6.04-24.09, [4, 5, 10]

Leiobunum rotundum (Latreille, 1798): 3° 29, 13.07–24.09 [3, 5] Mitostoma chrysomelas (Hermann, 1804): 1°, 17.04, [7] Nemastoma bimaculatum (Fabricius, 1885): 16° 169, 6.04–7.07,

[4, 6, 8, 9, 12] Lacinius ephippiatus (C. L. Koch, 1835): 1♂, 6.07, [4] Megabunus diadema (Fabricius, 1779): 1♀ 7 juv., 10–30.04, [1, 3] Mitopus morio (Fabricius, 1799): 12♂ 14♀, 4.07–24.09, [3, 4, 5, 8, 9] Oligolophus hanseni (Kraepelin, 1896): 1♂ 2♀, 24.09, [5, 8] Oligolophus tridens (L. Koch, 1836): 1♂, 24.09, [6] Paroligolophus agrestis (Meade, 1855): 2♂ 11♀, 24.09 [3, 5, 8] Phalangium opilio Linnaeus, 1758: 1♀, 13.07 [5] Rilaena triangularis (Herbst, 1799): 13♂ 11♀, 6.04–4.07, [3, 6, 8, 9, 11]

Neobisium (Neobisium) carcinoides (Hermann, 1804): 99, 17.04–3.06,

# Family THOMISIDAE

**PSEUDOSCORPIONS** 

Family ZORIDAE

HARVESTMEN

[4, 6, 8, 9, 12]

[8, 9]