37. NEOGENE BENTHIC FORAMINIFER BIOSTRATIGRAPHY AND DEEP-WATER HISTORY OF SITES 645, 646, AND 647, BAFFIN BAY AND LABRADOR SEA¹

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ABSTRACT

Benthic foraminifers were examined from Neogene sediments of Ocean Drilling Program (ODP) Sites 645, 646, and 647 to determine their biostratigraphy and to place constraints on the paleoceanographic history of Baffin Bay, Eirik Ridge, and the Gloria Drift.

At Site 645 in Baffin Bay, a Pleistocene *Stetsonia* assemblage is similar to the modern Baffin Bay assemblage, but an underlying *Epistominella takayanagii* assemblage has no modern analog. Miocene assemblages below a barren interval display low diversity and consist mainly of agglutinated species.

At Site 646 in the Labrador Sea, benthic faunal turnovers occur near important seismic horizons. A Miocene Nuttallides umbonifera assemblage similar to assemblages at other North Atlantic sites occurs below reflector R3. Above reflector R3, a coarse agglutinated assemblage containing more diversified calcareous benthic foraminifers was found that displays affinity to assemblages in the Norwegian-Greenland Sea. The faunal turnover near reflector R3 was interpreted as reflecting the onset (or renewal) of significant Denmark Straits Overflow Water at Site 646 at \sim 7.5 Ma. Agglutinated species disappear between reflector R2 and the base of the sediment drift, indicating a change in deep-water properties that occurred at \sim 4.7 Ma. This turnover ultimately may be linked to the reopening of the Mediterranean. The beginning of drift sedimentation at the Eirik Ridge is dated at \sim 4.5 Ma. Drift formation ceased at \sim 2.5 Ma, comcomitant with the appearance of ice-rafted sediments. Pleistocene assemblages containing *Stetsonia horvathi* display affinity to deep assemblages in high-latitude ocean basins.

Upper Pliocene and Pleistocene benthic assemblages at Site 647 contain N. umbonifera, which indicates a continued influence of corrosive deep water at the Gloria Drift.

INTRODUCTION

One of the main objectives of Leg 105 drilling in Baffin Bay and the Labrador Sea was to recover a continuous record of Neogene sediments, which are required to investigate the nature of the faunal response to circulation changes and to climatic deterioration leading to the onset of Northern Hemisphere glaciation. Baffin Bay and the Labrador Sea serve as conduits for the exchange of water and heat between the Arctic and North Atlantic oceans. Another important goal of Leg 105 was to provide insight into the nature of surface- and deep-water connections between the Atlantic and Arctic via the Baffin Bay corridor. Since the distribution of microfossils in pelagic marine sediments can be related to water-mass characteristics, the history of past circulation patterns can be interpreted from the micropaleontologic record of these basins. Thick sequences of Neogene sediments were recovered by means of hydraulic piston coring and rotary drilling at three sites: Site 645 in Baffin Bay and Sites 646 and 647 in the Labrador Sea (Fig. 1).

Pleistocene sediments from the Baffin Bay area are known only from shallow piston cores (Aksu, 1983) and exploration wells off Baffin Island (Klose et al., 1982) and from the Davis Straits area drilled by the Greenland Geological Survey (Henderson et al., 1982). Micropaleontological data from pre-Pleistocene sediments of Baffin Bay were unavailable, and it was not known whether foraminifer assemblages display closer affinities to the North Atlantic or to the Arctic.

The deep Labrador Sea was drilled during DSDP Leg 12 (Laughton, Berggren, et al., 1972), and the foraminifer data

from this area were summarized by Berggren and Schnitker (1983). Drilling during ODP Leg 105 was undertaken near two prominent sediment drifts in the Labrador Sea. The major paleontologic objectives at Sites 646 and 647 were (1) to provide a biostratigraphic framework for dating important seismic reflectors and (2) to study the nature of deep circulation over the Eirik Ridge and Gloria Drift.

For comparison, we also present the foraminifer record determined in cutting samples from the Texaco Blue-H28 well on the Labrador margin (Fig. 1). This well is one of the few commercial wells along the Canadian Atlantic Margin that has a deep marine upper Miocene to Pliocene stratigraphic record.

SAMPLE LOCALITIES AND LITHOSTRATIGRAPHY

The following summary of lithologic units and seismic stratigraphy derives from Srivastava et al. (1987). Site 645 lies on the continental slope off southern Baffin Island at a water depth of 2020 m. Drilling at Site 645 penetrated 1147.1 m of sediment to provide a continuous lower Miocene to Pleistocene stratigraphic record. The entire lithologic sequence in Holes 645B, 645D, and 645E has a pronounced terrigenous character and was subdivided into three lithologic units. (Fig. 2)

Lithologic Unit I consists of alternating beds of light tan to gray calcareous gravel-bearing muddy sand and dark gray silty mud having scattered dropstones. This unit occurs from 0 to 168.1 meters below sea floor (mbsf), and its age was determined as early Pleistocene to Holocene, based on dinoflagellates.

Lithologic Unit II extends from 168.1 to 330.7 mbsf and consists of noncalcareous silty mud with dropstones. Its age was determined as late Pliocene to early Pleistocene, based on dino-flagellates.

Lithologic Unit III extends from 330.7 to 1147.1 mbsf and is early Miocene to late Pliocene in age, based on dinoflagellates and scattered calcareous planktonic microfossils. Subunit IIIA

¹ Srivastava, S. P., Arthur, M., Clement, B., et al., 1989. Proc. ODP, Final Repts. (Pt. B),105: College Station, TX (Ocean Drilling Program).

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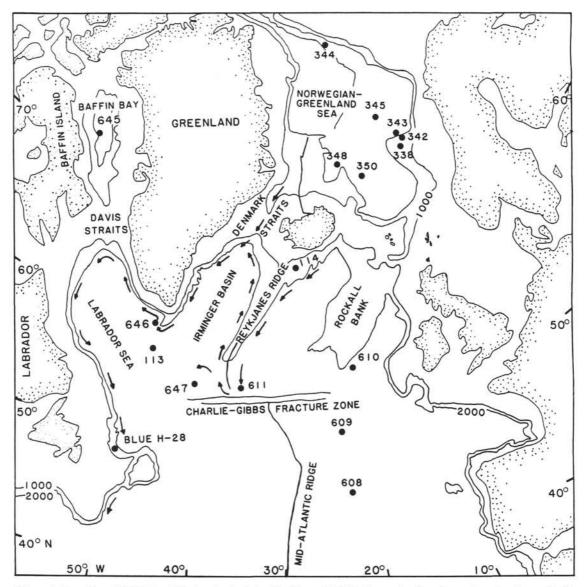


Figure 1. Location of ODP Leg 105 sites in the Labrador Sea and Baffin Bay and other sites and exploration wells discussed in the text. Arrows indicate inferred direction of deep circulation in the western North Atlantic. Base map is after Srivastava and Tapscott (1986), bathymetry in meters.

consists mainly of poorly sorted olive-gray silty mudstone and muddy sandstone. Scattered pebbles occur to a depth of 605 mbsf. Subunit IIIB contains calcareous silty claystones that have an organic carbon content of about 3% interbedded with muddy sandstones. Subunit IIIC consists of fine- to mediumgrained muddy sandstone and silty mudstones.

Site 646 was drilled in 3450 m of water on the northwest flank of the Eirik Ridge, a prominent sediment drift located off the southern tip of Greenland. The Eirik Ridge is a current-generated feature that occurs near the present pathway of Denmark Straits Overflow Water (Fig. 1). Drilling in Hole 646B recovered 766.7 m of terrigenous silty clays and silts having varying amounts of biogenic components. Dropstones are present to a depth of 236.4 mbsf and define the base of lithologic Unit I. Biogenic silica is present to a depth of 330 mbsf. Lithologic Unit II extends from 236.4 mbsf to the base of Hole 646B and consists of silty claystones and clayey siltstones having layers that contain up to 50% or 60% nannofossils.

Prominent seismic reflectors subdivide the sedimentary record of Hole 646B into four major seismic units (Srivastava, Ar-

thur, et al., 1987). Seismic reflector R1 defines the base of seismic Unit 1 and corresponds to a lithologic break between icerafted sediments and pre-glacial Pliocene sediments below. Thus, seismic Unit 1 coincides with lithologic Unit I. Seismic Unit 2 encompasses the drift sequence between 236.4 and 377 mbsf. The base of seismic Unit 2 in Hole 646B lies conformably on seismic Unit 3 and delineates the base of the sediment drift sequence visible in seismic profiles across the Eirik Ridge. Locally, the base of seismic Unit 2 is expressed as an erosional unconformity, but there is no evidence of a hiatus in Hole 646B. Reflector R2 corresponds to a change in carbonate content at ~ 500 mbsf, about 0.05 s below the top of seismic Unit 3. Seismic Unit 3 extends from 377 to 680 mbsf, and its base is delineated by seismic reflector R3. Reflector R3 is actually a couple of closely spaced reflectors that correspond to changes in the carbonate content of the sediments at 680 and 710 mbsf. Below 710 mbsf, the sediments of seismic Unit 4 are composed of dark gray silty claystones having carbonate contents below 10%.

Site 647, the southernmost site drilled during Leg 105, is located about 200 km south of the Gloria Drift at a water depth

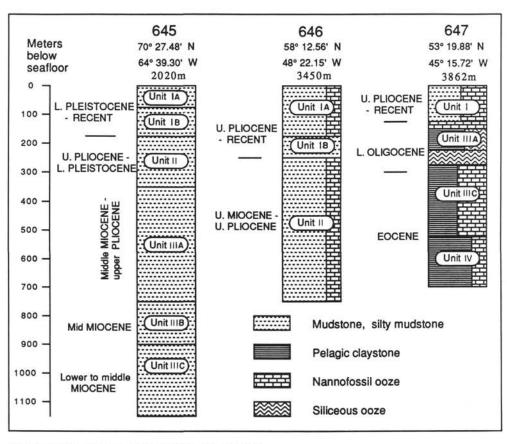


Figure 2. Lithostratigraphy at Sites 645, 646, and 647.

of 3862 m. Drilling at this site recovered a 116-m section of upper Pliocene to Holocene silty nannofossil clays and interbedded gravel-bearing silts that represent depositional cycles related to glacial-interglacial cycles. These sediments overlie an upper Miocene to upper Pliocene hiatus that corresponds to a regional seismic reflector.

The Texaco Blue H-28 well was drilled in 1486 m of water on the northeastern Grand Banks near Orphan Knoll. The well is situated near the present axis of the Western Boundary Undercurrent, a site of rapid sedimentation in the late Neogene. The Cenozoic microfossil record reflects deposition in midbathyal water depths, which sediments from this well has probably occupied since the Late Cretaceous.

METHODS

The chronostratigraphic framework for benthic foraminifer biostratigraphy in Hole 646B is based on magnetic reversal, nannofossil and diatom stratigraphy, (Shipboard Scientific Party, 1987a) and planktonic foraminifer datums determined by Aksu and Kaminski (this volume). The age model for Hole 646B used here is an updated version of the model reported in the Site 646 chapter (Shipboard Scientific Party, 1987a) and is given in Table 1. The ages of magnetostratigraphic units and epoch boundaries used throughout this study are after Berggren et al. (1985), with one exception: the age of the Miocene/Pliocene boundary (4.83 Ma) follows new estimates by Zijderveld et al. (1986).

Core-catcher samples collected aboard ship were processed according to standard laboratory techniques. For Pleistocene samples, foraminifers were picked from the >63- μ m fraction, owing to the presence of small calcareous species. Below the barren zone of Site 645, foraminifers were picked from the >125- μ m fraction because of the abundance of very fine sand in the coarse fraction in most samples. If calcareous benthic foraminifers or *Bolboforma* were found in the >125- μ m fraction, the fine fraction was also examined. However, in most samples from Hole 645E only agglutinated species were present, and the <125- μ m fraction was barren or contained only unidentifiable fragments. All pre-Pleistocene samples from Site 646 were picked from the >125- μ m fraction.

Pleistocene samples were processed in uniform, 10-cm³, wet sediment volumes. Samples with excessive amounts of fine sand were dried and sprinkled into carbon tetrachloride to float off the foraminifers. Pleistocene samples suspended in denatured ethanol and distilled water were split using a wet splitter (Thomas, 1985). Samples having large numbers of foraminifers were split into fractions that contained 300– 500 specimens for quantitative counts. Pre-Pleistocene samples processed onshore were first dried overnight at 60°C and then weighed before sieving to provide data about the absolute abundance of benthic foraminifers. Faunal counts from Holes 645B, 646A, and 647B are given in Tables 2 through 4, and data from Hole 646B are listed in the Appendix. Foraminifer data from Hole 646B also are available from the senior author in LOTUS 1-2-3 (.WK1) format on 360k MS-DOS floppy disk.

The stratigraphy and paleoecological interpretations of the hemipelagic sequence in the Blue H-28 well are based on analysis of 103 picked slides of foraminifers and associated microfossils from ditch cutting samples. Ages of the sediments in the Blue H-28 well are based on planktonic foraminifer biostratigraphy.

RESULTS

Site 645 Biostratigraphy

Fifty-five samples from lithologic Units I and II were examined quantitatively for benthic foraminifers (Table 2). The upper 220 m of Hole 645B contained benthic foraminifer assemblages dominated by calcareous species (Fig. 3). The numerically dominant species in most samples are *Stetsonia horvathi*, *Islandiella teretis*, *Epistominella takayanagii*, *Globocassidulina subglobosa*, *Cassidulina reniforme*, *Elphidium excavatum forma clavata*, *Buliminella hensoni*, and *Bolivina arctica*.

The mudline sample at this site contains an assemblage dominated by large, coarsely agglutinated species of *Rhabdammina*, Table 1. Depths below seafloor of biostratigraphic and magnetostratigraphic ages selected to construct the agedepth model for Hole 646B (after Shipboard Scientific Party, 1987).

Name of Datum	Age (Ma)	Depth (mbsf)
Name of Datum	(Wid)	(mosi)
FAD Emiliania huxleyi	0.247	17.8-19.3
LAD Rhizosolenia curvirostris	0.26	24.1-33.8
LAD Pseudoemiliania lacunosa	0.474	33.8-37.5
FAD Globorotalia inflata	a 2.1	^b 159.1-159.5
LCO Neogloboquadrina atlantica	^a 2.3	^b 183.3-188.2
LAD Discoaster surculis	>2.4	207.5-208.2
LCO Reticulofenestra pseudoumbilica	3.5	271.0-272.5
FAD Globorotalia puncticulata	^a 4.16	383.0-384.5
FAD Ceratolithus cristatus	<4.5	390.0-401.1
FAD Ceretolithus spp.	< 5.0	468.5-478.1
FAD Globorotalia margaritae	5.6	^b 506.9-516.5
LAD Discoaster quinqueramus	5.6	497.1-506.9
FAD Amaurolithus primus	6.6	584.2-585.7
FAD Discoaster quinqueramus	<8.2	714.0-715.5
Brunhes/Matuyama	0.73	52.10-73.86
Top Jaramillo	0.91	76.5-78.10
Matuyama/Gauss	2.47	190.40-192.60
Gauss/Gilbert	3.4	292.85-292.95
Top Nunivak	4.10	347.05-352.85
Bot Nunivak	4.24	359.45-359.55
Top C1 event	4.57	367.84-382.85
Bot C1 event	4.77	404.45-429.85
Top N1 event, Chron 6	6.37	577.35-577.44
Bot N1 event, Chron 6	6.50	578.44-578.55
Top N1 event, Chron 7	6.85	637.61-637.63
Bot N3 event, Chron 7	7.28	661.98-670.25
Top N1 event, Chron 9	7.90	680.14-680.24
Bot N1 event, Chron 9	8.21	710.51-710.53
Top N2 event, Chron 9	8.41	718.53-718.56
Bot N1 event, Chron 10	8.80	733.06-733.08
Top N1 event, Chron 11	8.92	733.88-740.33

^a Ages of planktonic foraminifer datums are after Weaver and Clement (1986).

^b Depths are from Aksu and Kaminski (this volume).

Cribrostomoides, Recurvoides, and Saccammina. The calcareous species accompanying this fauna include Islandiella teretis and Stetsonia horvathi.

The upper 40 m of Hole 645B (Cores 105-645B-1X to -4X) contains an assemblage that consists mainly of *I. teretis*, *S. horvathi*, *E. excavatum*, *C. reniforme*, and *B. hensoni*. Miliolid species are locally abundant at about 40 mbsf (Samples 105-645B-5X-3, 59-61 cm, and -5X, CC). Floods of *Stetsonia horvathi* were found between 83 and 121 mbsf (Samples 105-645B-10X-2, 30-32 cm, and -14X-2, 20-22 cm).

Below ~148 mbsf (Section 105-645B-16X, CC), the assemblage is dominated by *Epistominella takayanagii*. The interval corresponding to the base of lithologic Unit I and the upper part of Unit II (Cores 105-645B-24X to -32X) is barren of foraminifers, except for minor occurrences of *Cassidulina* spp. in Cores 105-654B-28X and -29X and *E. takayanagii* in Core 105-645B-31X.

Sediments recovered from Hole 645D belong to lithologic Units II and IIIA. Twenty core-catcher samples were examined from this hole; only two of which (Sections 105-645D-6R, CC and -13R, CC) contain rare specmens of *I. teretis, E. excavatum*, and *G. subglobosa*. As the first occurrence (FO) of *I. teretis* was reported in the mid-Pliocene in the North Atlantic (Murray, 1984), apparently, this Baffin Bay site does not contain a condensed stratigraphy, as suggested by earlier work (Aksu, 1983).

Drilling in Hole 645E penetrated lithologic Units II and III between 301 and 1147 mbsf. A total of 180 samples was examined for foraminifers and associated microfossils. Samples from Unit II and the upper part of lithologic Unit III are barren. Microfossils again were encountered in Subunit IIIB below ~ 750 mbsf. These consist mainly of fragments of agglutinated foraminifers. Less commonly, calcareous benthic foraminifers, pyritized diatoms, and specimens of *Bolboforma* are present. Planktonic foraminifers were found in only two samples. Because of the sparse and fragmentary nature of the assemblages, the microfossil data from this hole were not treated quantitatively.

The presence of *Bolboforma metzmacheri* at 753.4 mbsf (Section 105-645E-36R, CC) constrains the age of this sample as no younger than nannofossil Zone NN16 (early Pliocene). This species has been reported from the North Atlantic (Murray, 1987b) and the North Sea (King, 1983). In the North Sea, *B. metzmacheri* occurs together with the dextral form of *Neogloboquadrina atlantica* and defines King's late Miocene Zone NSP14.

The highest stratigraphic occurrence of coarse agglutinated species occurs at ~770 mbsf (between Samples 105-645E-38R-3, 44-47cm and -38R-6, 127-129 cm). This event occurs in the lower Pliocene in Hole 646B (see below) and at some DSDP sites in the Norwegian-Greenland Sea (Talwani, Udintsev, et al., 1976; Berggren and Schnitker, 1983). Agglutinated foraminifers are locally abundant to the base of Hole 645E. The assemblages consist mainly of coarse species of Rhizammina, Hyperammina, Cyclammina, Reophax, Lagenammina, Ammodiscus, Haplophragmoides, Glomospira, and Trochammina. Less frequently, specimens of Glomospirella, Ammosphaeroidina, and Martinotiella were found, but many samples contain little more than unidentifiable fragments. Miocene assemblages of this type have been reported previously only from DSDP Site 348 in the Norwegian-Greenland Sea (Talwani, Udintsev et al., 1976; Verdenius and Van Hinte, 1983).

The agglutinated assemblages between 792.0 and 844.0 mbsf (from Section 105-645E-40R, CC to Sample 105-645E-46R-3, 95-97 cm) are dominated by coarse astrorhizids and hormosinids, mainly Rhizammina, Lagenammina, and Reophax. Species with finely agglutinated tests become common in Section 105-645E-49R, CC. Samples between 878.2 and 885.3 mbsf (Section 105-645E-49R, CC to Sample 105-645E-51R-1, 41-43 cm) are dominated by finely agglutinated Bathysiphon fragments. Below this interval, the assemblages contain common specimens of Cyclammina cancellata, Recurvoides spp., and Haplophragmoides carinata. Also present is a compressed species of Reticulophragmium similar to a variety described from the Beaufort Sea (Young and McNeil, 1984) that we tentatively identified as R. cf. arctica. Near the base of Hole 645E, a change in the composition of the agglutinated assemblage was observed. The assemblage in Section 105-645E-78R, CC is dominated by small species of Ammodiscus and Trochammina.

Calcareous benthic foraminifers also were found between 772.2 and 844.0 mbsf (Samples 105-645E-38R, CC to -46R-3, 95-97 cm). The dominant calcareous species in this interval is *Melonis barleeanum*. Unilocular species (*Lagena*, *Oolina*, and *Fissurina*) are common, and accessory forms include *Globocassidulina subglobosa* and species of *Nodosaria*, *Quinqueloculina*, and *Uvigerina*. However, in most samples the preservation of calcareous benthic foraminifers is poor, and only relatively robust specimens are present. Smaller specimens presumably are dissolved. Samples between 861.7 and 974.9 mbsf (105-645E-48R-2, 84-86 cm to -60R, CC) are barren of calcareous foraminifers.

Calcareous foraminifers again were encountered in isolated samples below ~975 mbsf (Section 105-645E-60R, CC to Sample 105-645E-77R-1, 107-109 cm). In this interval *B. metzmacheri* is absent, and the most common genus is *Cibicidoides*. The most diverse assemblage occurs in Sample 105-645E-67R-2, 89-91 cm, which contains *Oridorsalis umbonatus*, *Pullenia bulloides*, *Cibicidoides pachyderma*, *C. lobatulus*, *Uvigerina* sp.

Depth (m)	4	7	14	16	23	32	36	42	43	52	62	63	72	75	81	83	91	95	101	110	113	116	120	122
Core/Section no	1	2-2	2	3-2	3	4	5-3	5	6-1	6	7	8-1	8	9-3	9	10-2	10	11-3	11	12	13-2	13-4	13	14-
Depth in Section(cm)	CC	121	CC	101	CC	CC	59	CC	51	CC	CC	66	CC	64	CC	30	CC	59	CC	cc	128	111	CC	1
Total number of Species	19	20	19	19	20	12	18	18	24	29	28	24	27	13	16	17	15	11	13	23	3	14	18	14
Total number of Specimensx10	50	18	25	26	27	41	54	25	40	95	152	77	136	13	25	117	7	6	5	83	,5	12	60 1	394
Astrononion gallowayi					X	X													X			X		
Bolivina arctica			X				1	2	X	2	2	1	1	8	36	2	16		40	9	50		X	_
Buccella frigida	1	-	-	X	1	_	X		X	X	X	X	X	-	X		1	3		X	-	1	X	_
Bulimina aculeata		X	-			-							-			1011	317	- 10		-			1.6	_
Buliminella elegantissima	4	2	3	5	2	59	11	1	16	2	1	1	X	8	14	1	28	7	4	7	38	4	X	_
Buliminoides spp.				_	-		X									_			_					
Cassidulina reniforme	13	3	11	3	9	2	13	11	20	X	4	8	11	18	5	2	13	5		3		1	1	_
C, subqlobosum	11	7	5	3	20	1	13	11	26	12	7	6	10	12	2	1	3	5	8	13	13	5	3	1
Cibicides lobatulus					X	_				X	X		X		-		1			X		3	-	_
Cibicides spp.		_	_						_		X		1							_		_		
Dentalina spp.	X			_			X		(X							1	_
Discorbis spp.		-	-		X		_		X	X								-			_			_
Elphidium bartletti	X	-		X		-				X		X	X	1		-	-			X		-		_
E. excavatum forma clavata	11	9	12	34	12	10		14	9	9	4	6	6	31	4	2	9	33	4	10		39	2	_
E. groenlundicum		X	_	5				-											-					_
E. subarcticum						-	X		X		X		X				1	3	4	X			X	_
Eoeponidiella pulchella	X						X	X	1	X	χ		X		_				4	_		1	-	_
Epistominella exigua	_	X	X							X									-					_
E. takayanagii	1	4	1	2	3	X	1	1	X	13	13	17	24	12	7	3	12	8	10	33		19	1	_
Eponides bradyi	1	3	3							X	X	X	X											_
Fissurina spp.	X	1	X	X	1		X	X	2	X	X	X	1		2	X			4	1		1	_	
Fursenkoina fusiformis	1	5	8	8	3	4	8	5	4	3	2	2	X		2	X	4	5	2	1		1	X	_
Glabratella wrightii			-	_	X					_							_		_				_	_
Glandulina spp.			_							_	1					_								_
Globobulimina spp.								-																_
Gutulina spp.																							_	_
Gyroidina soldanii		_							X				_	_	_	_	_					_	_	_
Haynesina orbiculare	X	X	X		1	X	1	4	1	X	X	X	1			X		8		X		8	X	_
Islandiella islandica		X		X	X			_																_
I. teretis	16	26	30	29	32	X	5	1	2	27	39	37	32	1	1	9		21		15		13	1	_
Laqena spp.			X	X	X															X				_
Lenticulina spp.																								_
Miliolina spp.							24	30	6	X	X		X			X	1	_		X				
Nonion barleeanum	X			X	2				X	1	X	1	X							2				_
Nonionella labradorica			1	1						8		X		1								2		
Oolina spp.					X	X		X		X	X	X						_			_			
Oridorsalis umbonatus																					_			_
Patellina corrugata									X										2					_
Planulina wuellerstorfi		X	1	X					¥		X	X	X											
Pullenia subcarinata	1	5	1							1														_
Quinqueloculina seminulum	X	2	1				1	2	3	X	X	1	X		14	X	6				_			
Robertinoides charlottensis		X							X															
Siphotextularia rolshauseni										X													_	
Stetsonia horvathi	38	29	17	5	12	22		15		17	17	10	4	1	6	69		2	13	1		2	81	7
Trifarina fluens									102.07		X	X							2					_
Triloculina arctica		4	5	1			12	X	2	X	X	X	X	1	7	X	1		2	1			7	
T. trihedra	X	X	1			X	9	3	3	X	X	X	X		1	X	1			2			3	
Trochammina ochracea										X														
Valvulinera arctica	-						-	X	X	4	8	6	5	7	X	10								
V. laevigata			_	X							X		X							X				

Table 2. Abundances of benthic foraminifers identified in samples from Hole 645B.

Table 2 (continued).

130		_	_		168	173	177	187	193	197	202	207	208	216	223	226	235	245	254	255	258	265	268	274	283	284	291	293	294
14	15	16	17-1	17	18	19-4	19	20	21-4	21	22-4	22	23-1	23	24-25	24	26-6	26	27-6	27	28-3	28	29-3	29cc	30-6	30cc	31-5	31cc	32-1
cc	CC	CC	29	CC	CC	31	CC	CC	100	22	37	CC	93	CC	118	CC	121	CC	83	CC	50	CC	89		89		88		38
15	9	13	25	15	15	16	2	6	5	5	17	10	14	2	1	1	0	2	2	0	3	0	7	0	0	0	12	10	1
16	26	29	159	303	129	218	3	4	37	1	321	99	7	.6	.1	.1	0	.2	.2	0	2	0	6	0	0	0	11	5	.1
														_									2						
1	X	12	1 X	X	1	44		6			X X	X	11		100							_					3	2	
4	1	3	1	X	_	1				_	X							-											
2	3	6	X	1			_			8	X	5	2						50		13		7				2	2	
<u>11</u> 1		13		X		X					X	70	15								78	_	72				1	<u>6</u> 1	
			X			X			_		X																		
1	_		_	X	_	X			X			X	3													_			
6	5	4	4	1	1	X		3	X		X	X	26	_					_		_		12				11	6	
4			X			X							6					_					2			_	3	3	
7	6	17	68	86	69	49	96	83	98		95	2	15	83		100		50	50			_	-				20	68	100
	_	_	X	X	1	X X		3		_	X		2					_	_							_	24	6	
		-	2		X	X		_		_																_		-	
	-					_	_	_															3					_	
		X			_			_			X															_	1		
_1		1	X	X		X				8	X	X	6		_								2				6	6	
	1	4	19 X	10	22	4	4	3	1 X	42	X	21	3					_			9					_		2	
					_			_		_	X							_											_
			X	X	X	X				23	2			17			_										2	-	
		_		X	X X	X		_			1		2											_			11	2	_
			X			_		_	_		X															_			
		X	X	X	1			_					_														16		
57	80	79	X	X	3							X															-	_	_
	wv		X		1							X	2	_							_								
	2	2	X	X	X							_	_					_											_
1	X		X		X	X														_								_	_
	n		-																						1000	101.57		-	

Table 3. Abundances of benthic foraminifers iden	tified in samples from Hole 646B.
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Core/Section no	1H	1H	1	2H	2H	2	3H	3H	3	4H	4H	4	5H	5H	5	6H	6	7H	7H	7	84	8H	8	98	9H	9	10H	10H	10H	10	11H	11H
	02	02	CC	02	05	CC	01	05	CC	02	06	CC	02	06	CC	02	CC	02	07	CC	03	05	CC	01	05	cc	02	02	05	CC	01	04
Depth in Section (cm)	33	132		16	114		34	63		14	130		13	131		13	-	106	38	_	108	108		113	138	_	35	76	35		91	79
Total number of Species	24	1	0	15	30	23	37	17	11	21	18	20	25	27	22	26	0	2	0	5	18	0	1	3	31	0	0	1	0	0	1	0
Total number of Specimens	219	1	0	600	701	107	758	115	34	2160	106	704	2968	297	286	322	0	4	0	12	74	0	1	8	1517	0	0	1	0	ų.	1	0
Astronomion gallowayi				_	_						1			X							1	X			X							_
Bolivina arctica	X		_	_			X		_					X	1	3				17	1			_	23	_		_		_	_	
B. pseudoplactata B. pseudoplicata		_			X	_	5 X					_			X	_		_					_				_			_		_
B. pseudopunctata				-	X	-				X	_		X	X		1		_	_				_								-	
Brazilina spathulata														2																		
Brizalina pseudopunctata			_				_									X																
Buccella frigida	_	_			X	5	X	3		_		1										_			X				_			
Bulimina elegantissima					1	3	2						X		X			1.00			1				X							
Cassidulina reniforme	X				X	2	X	1		4	21	10	5		2	5				17					2							_
C. subglobosum				5	X	2	X	1			12	8	1			1									1							
Cibicides. lobatulus	1			3	1	3	X	3		X	1		1		X													100		_		_
Dentalina spp.	1				1	5	X	2		1		1	X	1	1	1									X				_		_	
Elphidium bartletti									_				_	_							1			_	X		_			_	_	
E. excavatum	_	_	_	_	_	_										X								-		_	_	_	_	_		_
E. excavatum formae clavata			_		_	_	X			X	5	1	X	X		2	_				1		100	_	X			_				_
E. subarcticum	1								_			_			_	_					1			_	X							
Eceponidiella pulchella		_					1						X	_	_							_	_	-	X	_		_		_	_	
Epistominella exigua	2		_	1	5	5	1	9	21		8	6	3	5	4	2				17	7				X				_		_	_
Eponides bradyi	6	_	_	8	22	11	11	_		X		2	2	7	1	4	_	-			_				13			-				_
E. tumidulus					1.20		X	1			-				_		-					_	_	_			_	_	-			_
Fissurina spp.	X		_		3	2	X	3	3	-	4	2	X	3	1	1				_	1		_	_	1	_	_	-		_		_
Fursenkoina fusiformis	16		-	20	22	13	12	4		7	3	9	9	14	37	3		50			9				33			_				
Guttilina spp.			-	_			1				-		-					_								-					-	_
Gyroidina soldani			_	-	X	1	_ 1	1				2		_						-		_	-					-	_			-
Haynesina orbiculare	X					1									X										X		-				-	-
Islandiella teretis Karreriella bradyi	X			-	X	1	-	_	_			_		_		4								25								-
	1		-	-	1			3	_	v	1		2	7		1			_			_	-	25	X			-	-			-
Lagena spp.	X			-	Y	1	Y	2	-	X	1		2	3	1	4				-	1		-	-			-	-			-	-
Lenticulina spp. Nonion barleeanum	17			7	4	2	5	8	9		4	9	2	16	2	2	-		-		14				2					_	100	-
Nonionella labradorica	X		-	/	Y	4	J	0	1	- 1	4	7	2	10	4	4	-				14		_	-	4		-				100	-
N. turgida	<u>A</u>		-				Y		-	-														-	X			-		-		-
Nuttalides umbonifera			-		_	1	Λ.	10	18	X	2	_	1	X		X				17			-		Δ.			-	-			
Oolina spp.	X				X		X	1	6		1	-	-	1	1					11	1			25	X	-					_	-
Ophthalmidium acutimargo	-				<u>n</u>		1		0	1	1	-	-	2	-		1		-	-	-		-	2.0			-					_
Oridorsalis umbonatus				1	¥		•	1		Y	-	2	1	2	2	1		50			31		-		2							
Pattelina corrugata			_	•				•	_		_		· ·	-	-									_	ĩ		-					-
Planulina wuellerstorfi	1		-	5	1		1		6	1		1	-	2						-	4			50		-	_	-				
Pullenia bulloides				7		5		13			23	5	1		4	10	10-5	_			5				1					0.000		
P. subcarinata	26			11	6	16	5	20	12	4		2	4	2							12				2							
Pyrgo williamsoni							X					5		X		X				_						_	-					
Quinqueloculina cultrata					2		2	_		X			1	100		_													_	_		
Q. seminulum														X																		_
Rosalina spp.				_	X	_															_				_							
Siphotextularia rolshauseni				1	X	2	1		3	2	2		X	X	X									_	X		_		_	-		
Stetsonia horvathi	2	100		20	21	15	33	12		69	7	27	56	15	22	39				33					6		_			_	_	_
Tosaia hanzawai		1.441		3		1				X	5	2	2	2		7				_	3				X	_	_					
Triloculina arctica				3			X									1							_		X					_		
T. trihedra	X			4	X		1				1	1	2	3	1	2									3		_		-			
Uvigerina asperula	4					_		_		X					1.1									_		_		_		_		_
U. peregrina	16						X																								_	
Vavulineria arctica	X			1	3	3	8			2		1	4	7	1						2				6		-	_		_	_	
V. levigata	1				X		X							X																		

(smooth), and four specimens of *Globigerina praebulloides*. Although the total range of *G. praebulloides* is from P22 to N18 (Kennett and Srinivasan, 1983), this species is restricted to lower to lower middle Miocene sediments in the North Sea (Zone NSP11 of King, 1983) and lower Miocene sediments on the Labrador Margin (Gradstein and Agterberg, 1982).

Site 646 Biostratigraphy

A total of 140 samples from Holes 646A and 646B was examined for benthic foraminifers. The abundance data for samples from Hole 646A are given in Table 3, and data from Hole 646B are listed in the Appendix. The accumulation rates of ben-

M. A. KAMINSKI, F. M. GRADSTEIN, D. B. SCOTT, K. D. MACKINNON

Table 4. Abundances of benthic foraminifers identified	in samples from Hole 647B.
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Core Section	1-6	1-6	1cc	2-6	2-6	2cc	3cc	4-2	4-3	4cc	5cc	6-2	6-6	6cc	7-2	7cc	8-2	8-3	8cc	9-2	9-3	9cc	10-2	10-4	10cc	11-2	11-4	licc
Depth in section (cm)	8	109	CC	5	139	CC	CC	5	61	CC	CC	20	113	CC	21	cc	12	75	CC	2	54	CC	25	66	CC	83	12	CC
Total number of Species	27	25	20	3	26	26	23	4	20	25	29	18	23	27	25	21	25	21	25	22	26	18	27	24	25	8	19	2
Total number of Specimens/20cc	1324	268	432	3	491	692	2824	6	968	508	460	305	475	529	856	522	1392	463	978	1144	620	1160	1696	472	367	25	544	2

Astrononion gallowayi													1														
Bolivina arctica	_			-		_		_	_	_			_	X	X	1		_	_	_	X	1	2	_	X		
B. pseudoplicata							X									_	_			1			_				
Brizalina pseudopunctata	X		_	_	-				X	2	1	_	_	_			_	_					_	_		_	1
Buccella frigida			4			1		_									_			_					_		1
Buliminella elegantissima	X					_	_			1	_					_		_		_	_	_					
Cassidulina laevigata				_																	X						
C. reniforme		X			2	3		_	2	3	2	X	2	15	1	3	2	3	2	2	4	2	3				
C. subglobosum	2	4	20	33	1	1	2			1	2					X	1				3	4	16	1	20		1
Chilostemolla oolina													1	X													
Dentalina spp.		1			1					_	1	X		1			1	X	X	1	X		1	2	1	8	1
Discorbis nitida			_	_	1	_																					
Eggerella bradyi												X				X	1	1	1	1	X			3.	1		
Elphidium excavatum	1	X	4		5	1	1		X	1	2	1	2	2	1		1	X									
E. groenlandicum					1					· ·			-	-											_		
E. subarcticum																					_				X		
Eceponidiella pulchella	X			_	_		X			1	_	_	_		X												
Eponides bradyi	3	4	37		2	9	1		1	3	7	17	14	17	6	25	10	9	5	4	8	11	12	10	25		22 50
E, tumidulus	0	1	31		2	1	1		2	5	1	1/	14	1/	0	20	2	Y	0	1	0 Y	11	12 X	10	44		44 J
Epistominella exigua	2	1	6		5	4	29	17	4	7	12	X	8	3	2	15	3	49	24	3	3	10	2	18	1	_	7 5
E, takayanagii	2	1	0		3	4	21	1/			12	Ă	6	2	2	13	.9	47	21	2	3	10	2	10	1		/ 24
		7	15		F	*	7	_		2	,	0	3	2	2		2	¥.			i.	7		0	0		4
Fissurina spp.	6	6	15		5	6	3		2	10	6	2	3	2	2	4	2	X	1	1	1	3	4	2	2	,	1
Frondiculina sp.		-								~	-	1.4		-	-					~						4	v
Fursenkoina fusiformis	3	5	13		5	6	2	17	1	2	1	1	2	3	2	1	1	1	X	1	2	_	1	2	1		X
F. pauciloculata		_					_				_				_				_		_		1	1			
Guttulina sp.				_			X		_	1	X	X												_			
Gyroidina soldanii		X			_				2	1	3		2	2	2	2	3	1	X		1		2	2	1	_	X
Haynesina orbiculare	1	1			2					_				X									X				
Hoeglandina elegans		_															_					_	X	_		_	
Islandiella teretis									X																		
Karriella bradyi	X										X		_						X		X						
Lagena spp.	1	1				1	1		X		1			2	2	1		X	X		2	1	1	2	2		1
Lenticulina spp.	X		2											X						1	1		X				
Miliolinella sp.	3		2		_	1	_																¥				
Nodosaria sp.						-			_																	4	
Nonion barleeanum	1		2		1	1	2	33	2	3	6	6	3	6	1	7	1	1	¥	1	6	3	5	4	4	4	4
Nonionella bradyi			-		-		-	~~	-	×		<u>v</u>			-	1.1.	2	Ŷ	X	5	¥	1	2	1	1	1	
Nuttaloides umbonifera	4	12	13		16	3	2		1	1	6	43	14	X	38	3	30	1	2	49	23	14	6	14	X	56	16
Dolina melo	1	12	10		10	<u>×</u>	4		-	1	X	40	1	.0	30	<u>,</u>	- 10	1	4	11	20	14	0	14	^	00	10
		-					-			-	. 19		-		10		0			0	10		7	7	2	4	8
Oridorsalis umbonatus	8	7	41	33	4	2	3		2	5	9	X	5	4	12	6	8	11		8	12	14	F.	1		•	2
Planulina wuellerstorfi	2	5	26	33	15	12	2		X	1	5	X	6	2	1	2	4	1	1	2	5	3	8	1	2		2
Pseudopolymorphine novangliae		X	_			1	X			-	-		1	-	X			-	-								-
Pullenia bulloides	X	3	12.0		2	4	1		-	2	3	6	1	3	X	X	1	7	3	22	2	00	4	4	4	10	3
P. subcarinata	16	21	56	_	17	13	10	33	7	20	17	22	23	32	11	23	13	8	5	11	17	22	13	18	16	12	24
Pyrgo williamsoni	2	4	4		3	2	1	_	4		2	_	1	1	1	2	1		X	1	1		3		2	_	
Pyrulina augusta			_	_	_	1				_				_			_	_		_		_		-			
P. cylindrica																			X			_		_		_	_
Quingueloculina cultrata	2	3	6		2	2	1		X	1	3		_	X						1			_				
Q. seminulum	X	1							_	2										1		_	_	1	X		
Reophax bacillaris													1		1		1		X					1			
Sigmolinopsis schlumbergi		_																					X	1			
Siphotextularia rolshauseri	1	2	9		3	2	1				2	X		2	2	X	3	2	1		2	5	1	1	3		1
Spiroloculina sp.					Ť	1	-	_			X			-	-	X	1			1	-						
Stetsonia horvathi	29	7	19		5		11		67	27	2		1	1	6	1	2			1		1		1	1		4
Tosaia hanzawai	4	2	4		1	0	1		9/	1	1	X	2	2	2	2	4	7	2	1	4	1	2	4	6		1
	4	4	4		1		1			1	1	X	4	4	4	2		\$	4	1	4	1	-	1	0		
Trifarina fluens			50			17		_	F	-		Å						-	y			1	-		¥		
Triloculina trihedra	8	6	50		1	13	2		5	3	1		1	X	4		4	2	X	~	1		-		Å	P.	
Uviqerina peregrina					1		_		1	2	2		8	6	1			X	X	2	2	1		1		8	
Vaqinulinopsis sp.					_		_	_									_	-	X			_		_	_		
Valvulinera arctica		X	_		1						_				1	_	_										
PLANKTONICS	*	×	- 22	279	*			355		÷		305	*	ŧ	*	ŧ	÷	¥	*	+	+	ŧ	÷		1000	140	736 174

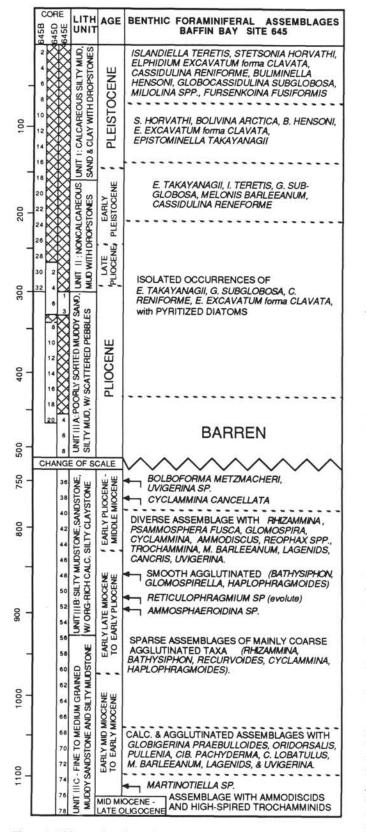


Figure 3. Lithostratigraphy and summary of the benthic foraminifer assemblages and associated microfossils at Site 645, Baffin Bay. The age interpretation of the composite section of Holes 645B, 645D, and 645E is based on the multiple microfossil and paleomagnetic records given in the Site 645 chapter (Shipboard Scientific Party, 1987c).

thic foraminifers and the carbonate record from Hole 646B are compared with the regional seismic reflectors in Figure 4. In Hole 646B, major changes in the benthic foraminifer assemblages occur near several of the seismic reflectors identified from shipboard seismic and downhole logging studies. The record of important first and last occurrences in Hole 646B is shown in Figure 5. The composition of each assemblage is discussed in sequence from youngest to oldest.

The upper interval of seismic Unit 1 (Samples 1Q5-646B-lH-2, 33-35 cm to -6H-2, 13-15 cm) contains abundant benthic foraminifers dominated by *Stetsonia horvathi*, with common occurrences of *F. fusiformis* and *Pullenia subcarinata*. The species *Epistominella exigua*, *Melonis barleeanum*, *Planulina wuellerstorfi*, *Cassidulina* spp., and *Pullenia bulloides* are present throughout this interval in lower but significant percentages. The uppermost sample (105-646B-1H-2, 33-35 cm) is co-dominated by *Pullenia subcarinata*, *M. barleeanum*, *F. fusiformis*, and *Uvigerina* sp. The assemblages in the upper part of seismic Unit 1 generally display good preservation.

Between Cores 105-646B-6X and -11X, the sediments are essentially barren of foraminifers, except for Sample 105-646B-9H-5, 138-140 cm, which contains abundant Fursenkoina fusiformis and Bolivina arctica, and Sample 105-646B-8H-3, 108-110 cm, which contains a few specimens of Oridorsalis (Table 3). A major turnover in the benthic assemblage occurs near the base of seismic Unit 1, which overlies the sediment drift. Between 206 and 236 mbsf (Samples 105-646B-23X-5, 122-124 cm to -25X, CC), the last occurrence (LO) of Laticarinina pauperata, Hoeglundina elegans, and Sigmoilopsis schlumbergeri was encountered. In the Norwegian-Greenland Sea, Laticarinina and Hoeglundina also disappear before the onset of glacial conditions (Talwani, Udintsev, et al., 1976). At Rockall Margin Site 552A, the LO of Laticarinina was recorded in the late Pliocene by Murray (1984). The lower part of seismic Unit 1 contains a low abundance Melonis barleeanum assemblage. Assemblages containing common M. barleeanum are typical of glacial sediments in lower bathyal DSDP sites in the northern North Atlantic (Murray, 1984) and in the Norwegian-Greenland Sea (Talwani, Udintsev, et al., 1976).

Within the sediment drift (seismic Unit 2), the abundance of benthic foraminifers is still low, and the assemblage is numerically dominated by *Melonis barleeanum*, *Pullenia bulloides*, *Cibicidoides* spp., and unilocular forms (*Lagena*, *Oolina*, and *Fissurina*) and entosolenians. This assemblage also contains a greater proportion of species that are typical of modern North Atlantic Deep Water (NADW) (*Oridorsalis* spp., *Planulina wuellerstorfi*, *Globocassidulina subglobosa*) than assemblages in other seismic units. The first occurrence of *Islandiella teretis* was found in Sample 105-646B-39X-2, 15-17 cm. This species first appears in the upper Pliocene of Rockall Margin sites (Murray, 1984).

The base of the drift sequence was placed at 377 mbsf from the seismic record. A major benthic faunal break occurs between 387.4 and 391.0 mbsf (between Samples 105-646B-41X-5, 10-12 cm and -41X, CC). Below the seismic reflector delineating the base of the drift, the upper Miocene to basal Pliocene benthic foraminifer assemblages of seismic Unit 3 are numerically dominated by coarse Rhizammina and associated agglutinated taxa (Cyclammina pusilla, Haplophragmoides, Psammosphaera, Recurvoides, Reophax, Trochammina, Lagenammina, and Ammobaculites). Calcareous benthic foraminifers are numerically subordinate and are represented by Oridorsalis, Melonis barleeanum, M. pompilioides, Pullenia bulloides, and Cibicidoides spp. Near the middle of seismic Unit 3 (between Sections 105-646B-47X, CC and -53, CC), Nuttallides umbonifera occurs sporadically and in low numbers. There is a smaller percentage of unilocular species in seismic Unit 3 than in the overlying unit.

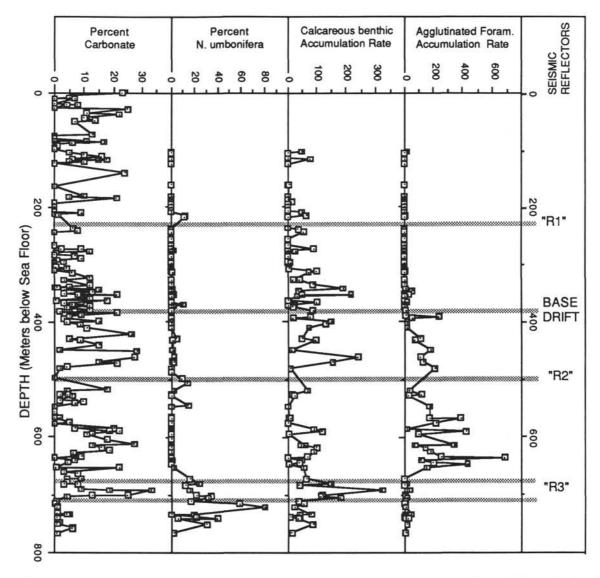


Figure 4. Accumulation rates of benthic foraminifers, calcium carbonate content, and percentages of *Nuttallides umbonifera* in Hole 646B. Carbonate data are from the Site 646 chapter, supplemented by additional analyses of core-catcher samples. Also shown are the positions of important regional seismic reflectors.

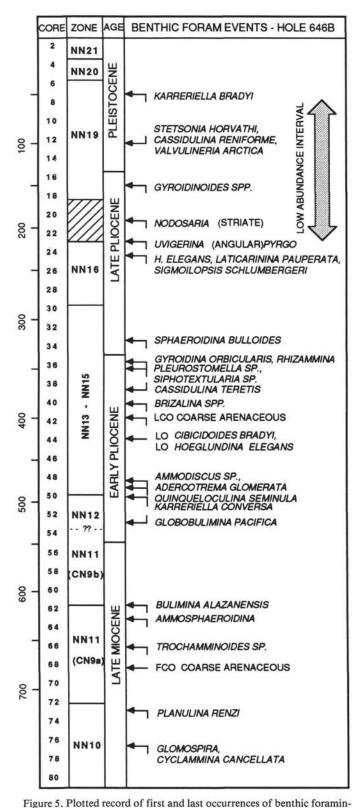
The upper reflector of the R3 couplet originally was placed at 680 mbsf (Shipboard Scientific Party, 1987a). Between 671 and 651 mbsf, another distinct change in the benthic foraminifer assemblage occurs: coarse agglutinated taxa become sporadic and rare downhole. The calcareous assemblages within the seismic couplet (Cores 105-646B-72X and -73X) are transitional between assemblages from seismic Units 3 and 4. In the six samples examined from Cores 105-646B-72X and -73X, the proportion of *N. umbonifera* is approximately equal to the proportion of NADW species. The calcareous species *Planulina renzi* was found in Samples 646B-73X-2, 58-60 cm and -73X, CC. In the Gulf of Mexico, the LO of this species was reported in Zone N17 (Van Morkhoven et al., 1986).

In seismic Unit 4 (710-766 mbsf), the benthic foraminifer assemblage is dominated by *Nuttallides umbonifera* (Fig. 4). The abundance of this species was correlated positively with the degree of undersaturation of bottom waters with respect to calcium carbonate (Bremer and Lohmann, 1982). Characteristic accessory species in this assemblage are agglutinated foraminifers having smooth, finely finished tests, such as *Bathysiphon*, Ammodiscus, and Glomospira. Isolated specimens of Cyclammina cancellata also occur near the base of Hole 646B, but its LO is probably not isochronous with its LO on the Labrador Margin or in Hole 645E.

Site 647 Biostratigraphy

A total of 28 samples from lithologic Unit 1 of Hole 647A was examined for benthic foraminifers, and total numbers of specimens range from 3 to 2824 per 10 cm³. Assemblages in a given sample in Cores 105-647A-1H to -5H are highly variable and may be dominated by any of the following species: *S. horvathi, P. subcarinata, N. umbonifera, Oridorsalis, or E. exigua.* Common accessory species include *M. barleeanum, P. wuellerstorfi, Fissurina* spp., *Triloculina trihedra,* and *Eponides bradyi* (Table 4). The high variability of assemblages may be due to low density of sampling and to Milankovitch-forcing functions in this data set.

Samples 105-647A-6H-2, 20-22 cm and -6H-6, 113-115 cm are dominated by *N. umbonifera* and *P. subcarinata*, with commom *E. bradyi*. The FO of *Uvigerina* was observed in Sample



lation using the age-depth model for this site (Shipboard Scien-

tific Party, 1987b). Cores 105-647A-8H to -10H contain planktonic oozes having high numbers of benthic foraminifers. These assemblages are mostly dominated by N. umbonifera and P. subcarinata, with subdominant G. subglobosa and Oridorsalis. One sample (105-647A-8H-3, 75-77 cm) is dominated by E. exigua.

NEOGENE BENTHIC FORAMINIFER STRATIGRAPHY

Two of the three samples examined from Core 105-647A-11H were almost barren, but Sample 105-647A-11H-4, 12-14 cm, contained an abundant assemblage dominated by P. subcarinata and E. bradyi. The base of lithologic Unit I of Site 647 has an estimated age of 2.5 Ma, based on the planktonic microfossil and magnetostratigraphic evidence. A hiatus of at least 3 m.y. duration separates the upper Pliocene from the underlying upper Miocene.

Texaco Blue H-28 Well

The Neogene biostratigraphic record of the Texaco Blue-H28 well displays a fourfold division, based on planktonic foraminifer stratigraphy (Fig. 6). The foraminifer assemblages are discussed from from oldest to youngest.

Middle Miocene sediments containing Sphaeroidinella seminulina and Catapsydrax dissimilis occur between 3810 and 3640 mbsf and immediately overlie lower Oligocene and older sediments. The middle Miocene benthic assemblage is rich in agglutinated species, including Bathysiphon, Rhizammina, Saccammina, Cystammina, Cyclammina placenta, Cyclammina pusilla, Cyclammina orbicularis, Cribrostomoides subglobosus, Karreriella conversa, and Sigmoilopsis schlumbergeri. This assemblage is remarkably similar to the assemblage from the upper Miocene to lower Pliocene seismic Unit 3 of Hole 646B. Calcareous foraminifers that testify to a deep (at least middle to lower bathyal) environment are Laticarinina, Melonis pompilioides, and Cibicidoides wuellerstorfi.

The upper Miocene (N16-N18) occurs between 3640 and 3160 mbsf. This interval contains the last occurrences of Sigmoilopsis schlumbergeri, Neogloboquadrina humerosa-atlantica, N. acostaensis, N. continuosa, Globoquadrina dehiscens, and the G. menardii group. A diverse agglutinated assemblage similar to the one mentioned above occurs between 3200 and 3500 mbsf.

The upper Pliocene/lower Pliocene boundary in this well was originally determined by the last occurrences of Neogloboquadrina atlantica and Globorotalia hirsuta near 2550 mbsf. However, Aksu and Kaminski (this volume) report the last appearance datum (LAD) of N. atlantica at ~2.4 Ma in the Labrador Sea. Therefore, the LO of this species in exploration wells along the Labrador Margin may be a useful indicator of the preglacial Pliocene. Another useful LO in this well is G. puncticulata, which has a LAD at 3.8 Ma in Hole 646B (Aksu and Kaminski, this volume). Through much of the Pliocene, the benthic foraminifers are represented by calcareous taxa such as Elphidium, Gyroidina, Cassidulina, Uvigerina, and Laticarinina. However, an assemblage rich in coarse agglutinated species similar to those mentioned above occurs at 2600 mbsf.

DISCUSSION

Paleoecology of Benthic Foraminifers

Agglutinated Assemblages

Neogene agglutinated foraminifer assemblages are known from silled high-latitude basins, such as the Norwegian-Greenland Sea and Baffin Bay, but until now, Neogene agglutinated assemblages have not been reported from any DSDP site in the deep North Atlantic. Agglutinated assemblages are present in Paleogene sediments of the Labrador Sea (Gradstein and Berggren, 1981) as well as in the modern North Atlantic (Schroder,

ifers in Site 646 vs. shipboard nannofossil stratigraphy (Shipboard Scientific Party, 1987a).

105-647A-6H-6, 20-22 cm. Section 105-647A-6H, CC marks the FO of B. arctica and is the only sample containing high percentages of Cassidulina reniforme.

The Pliocene/Pleistocene boundary was placed at approximately 72 mbsf, or within Core 105-647A-8H, based on interpo-



Figure 6. Biostratigraphic record of the Texaco Blue H-28 well, Labrador Margin.

1986), but are usually not preserved in Neogene sediments. Until now, the only link in time between these occurrences has been the Pliocene agglutinated assemblages in the Texaco Blue H-28 well on the Labrador Margin. Therefore, the microfossil record from Site 646 is unique in containing well-preserved agglutinated foraminifers.

The presence of agglutinated foraminifers in Cenozoic sediments is usually associated with sediment parameters that allow the preservation of organic matter. The ecology of early Cenozoic "flysch-type" agglutinated assemblages was discussed by Gradstein and Berggren (1981) and Miller et al. (1982). These authors noted the occurrence of these assemblages in areas of rapid sedimentation and in sedimentary basins containing corrosive bottom water. The set of environmental conditions associated with flysch-type assemblages has become known as the "old bottom-water model." The disappearance of these assemblages from the deep Labrador Sea near the Eocene/Oligocene boundary was correlated with the onset of vigorous deep circulation and more oxygenated conditions (Miller et al., 1982; Miller and Tucholke, 1983). After this event in the North Atlantic, agglutinated assemblages persisted only in the high-latitude basins. In accordance with the old bottom-water model, for agglutinated assemblages to be preserved in the upper Miocene to lower Pliocene of the Labrador Sea, sediment parameters or bottom-water properties must have surpassed a certain threshhold. But which parameter or combination of parameters is unique to Site 646?

A compilation of sediment properties (Table 5) reveals that Site 646 has little in common with most Atlantic sites. With the exception of Site 114 on the east flank of the Reykjanes Ridge, lower Pliocene sediments in the eastern Altlantic are nannofossil oozes having high calcium carbonate contents. Drilling at DSDP Hole 114 penetrated drift sediments having a carbonate content of about 26% and a higher sedimentation rate than in Hole 646B, but no agglutinated assemblages were recovered.

The Miocene to early Pliocene deep water in the Norwegian-Greenland Sea and Arctic Ocean apparently was corrosive, as deep sites have carbonate contents of less than 1% (Talwani, Udintsev, et al., 1976). Agglutinated foraminifer assemblages are present in pre-glacial Pliocene sediments of DSDP Hole 344 on the Knipovich Ridge and Hole 345 in the Lofoten Basin. In Cores 26 through 33 of Hole 344, the generic composition of the foraminifer assemblage is remarkably similar to the assemblages in the Eirik Ridge. The assemblage consists mainly of Bathysiphon, Haplophragmoides, Reophax, Hormosina, and Saccammina, but at some levels Melonis barleeanum and I. teretis occur together with the planktonic species Neogloboquadrina atlantica. The LO of coarse agglutinated species is associated with the onset of ice rafting in Hole 345. The pre-glacial assemblage at this site consists of Cyclammina, Haplophragmoides, and Martinotiella, and is more diverse than the underlying Miocene assemblage (Talwani, Udintsev, et al., 1976).

We speculate that the coarse agglutinated assemblages at Site 646 indicate the presence of corrosive bottom water derived from the Norwegian-Greenland Sea. Drilling at Hole 113 in the Labrador Sea recovered a thick sequence of turbidite sediments in a local topographic depression. Calcium carbonate content is

Table 5. Compilation of late Miocene environmental parameters from DSDP holes in the North Atlantic.

Location	Site	Cores	Latitude	Depth (mbsf)	WBD	TOC	Carbon (%)	Clay (%)	Sand (%)	Silica?	Drift?	Sed rate (m/m.y.)	Aggl
Labrador Sea	646	26-53	58°13'N	3455	2	0.4	0-20	90	5	No	Yes	90	Yes
	113	7-12	56°47'N	3615	2	0.4	1-12	62	37	No	No	180	No
	112A	5	54°47'N	3615	1.7	NA	NA	NA	NA	No	No	20	No
	111A	5-6	50°25'N	1797	1.7	0.1	72	21	54	No	No	0.4	No
East North Atlantic	114	4-6	59°56'N	1927	1.6	0.1	26	23	20	Yes	Yes	120	No
	116	2	57°30'N	1151	1.7	0.1	90	44	5	Yes	No	30	No
	611	22-33	52°50'N	3200	1.8	NA	70	NA	NA	No	Yes	58	No
	610	8-10	53°13'N	2417	1.8	NA	95	NA	NA	No	Yes	51	No
	609	26-31	49°52'N	3883	1.8	NA	90	NA	NA	No	No	89	No
	608	13-15	42°50'N	3541	1.7	NA	95	NA	NA	No	No	19	No
	552A	14-23	56°03'N	2311	1.7	0.1	93	80	5	Yes	Yes	25	No
	553A	3	56°06'N	2339	1.7	0.1	92	90	5	Yes	Yes	30	No
	554	5-6	56°18'N	2584	1.7	0.1	97	85	10	Yes	No	7	No
	555	3	56°34'N	1669	1.8	0.1	94	70	15	Yes	No	16	No
Greenland Sea Ridge	407	8-14	63°56'N	2492	1.6	NA	85	NA	NA	No	No	45	No
	408	11-13	63°23'N	1634	1.6	NA	85	NA	NA	Yes	No	35	No
Norwegian-Greenland Sea	337	4-5	64°53'N	2657	1.5	0.2	NA	40	8	No	No	2	No
	344	26-33	76°09'N	2201	2.1	0.8	1	45	5	No	No	76	Yes
	345	5, CC-6	69°50'N	3216	1.7	0.4	1	NA	NA	No	No	NA	Yes
	348	6-7	68°30'N	1777	1.4	0.3	1	50	5	Yes	No	22	No

WBD = wet-bulk density; TOC = total organic carbon; Silica? = presence of biogeneic silica; Drift? = site location on sediment drift; Aggl? = presence of agglutinated foraminifers.

low, and sedimentation rate is higher than in Hole 646B. However, Hole 113 is not in the pathway of Denmark Straits Overflow Water.

The trophic structure of the agglutinated foraminifer assemblage is another clue to the nature of the depositional environment at Site 646. The assemblage in seismic Unit 3 is strongly dominated by coarsely agglutinated specimens of Rhizammina, which is an epibenthic suspension-feeder (Jones and Charnock, 1985). Benthic communities dominated by suspension-feeders are rare in the deep sea (Jumars and Gallagher, 1982) because the ratio of suspension-feeders to deposit-feeders normally declines with depth and distance from shore (Levinton, 1982). Suspension-feeding communities in the deep sea have been found only near hydrothermal vents (Lonsdale, 1977) or in areas where there is a lateral advection of food particles (L. Mullineaux, pers. comm., 1987). In Alvin dives, H. Sanders (pers. comm., 1987) observed concentrations of suspension-feeding animals on top of local topographic elevations, whereas local depressions were occupied by deposit-feeders. In the modern North Atlantic, there is an increased proportion of the coarse tubular agglutinated species beneath the Western Boundary Undercurrent (WBUC) (Schroder, 1986). Thus, a Rhizammina community presumably requires moving bottom currents to supply it with food. However, the genus is sessile and probably cannot escape the effects of strong bottom currents. The change from agglutinated assemblages below the sediment drift to calcareous assemblages in the drift may reflect higher bottom current intensities, which may have inhibited Rhizammina and resulted in increased ventilation of the deep environment.

In contrast, the assemblage in seismic Unit 4 of Site 646 contains a greater proportion of species with finely agglutinated tests (Table 6). Species restricted to this assemblage, *Ammodiscus* and *Glomospira*, were identified as epifaunal detrital feeders (Jones and Charnock, 1985) and presumably have a low tolerance for disturbed environments. In the modern North Atlantic, these organisms are present in greater proportion in environments that are relatively undisturbed by bottom currents (Kaminski, 1985).

The agglutinated assemblages from the Texaco Blue H-28 well are remarkably similar to those from seismic Unit 3 in Hole 646B. Each species we found in the Pliocene of the Blue H-28 well also occurs in Hole 646B, and specimens of *Rhizammina*

Table 6. Summary of important benthic foraminifer species and species groups in Hole 646B.

Seismic Unit 2	Seismic Unit 3	Seismic Unit 3/4*	Seismic Unit 4
18.1%	5.2%	12.1%	3.2%
0	0	0.7	7.0
7.0	65.6	8.0	13.9
0.9	0.9	18.4	25.0
10.3	4.9	19.4	11.5
63.7	23.3	41.3	39.4
32	40	6	12
	Unit 2 18.1% 0 7.0 0.9 10.3 63.7	Unit 2 Unit 3 18.1% 5.2% 0 0 7.0 65.6 0.9 0.9 10.3 4.9 63.7 23.3	Unit 2 Unit 3 Unit 3/4* 18.1% 5.2% 12.1% 0 0 0.7 7.0 65.6 8.0 0.9 0.9 18.4 10.3 4.9 19.4 63.7 23.3 41.3

Seismic Unit 3/4 samples are from the calcareous interval within the reflector R3 couplet.

^a Values for NADW species are derived from the combined abundances of *P. wuellerstorfi, Oridorsalis, G. subglobsa,* and *E. exigua.*

Note: Numbers represent the mean values of census data from each seismic unit.

are about the same size. This well site is located under the current axis of the Western Boundary Undercurrent. The predominant lithology is hemipelagic clay, and sedimentation rates increase dramatically from 100 m/m.y. in the late Miocene to ~ 300 m/m.y. in the Pliocene and Pleistocene. In the Miocene interval of the well, the occurrence of agglutinated taxa is fairly continuous, but in the Pliocene, agglutinated taxa cluster at discrete levels. We tentatively interpreted the preservation of this assemblage as an indication of periods of slower deposition. We postulate that the highest level with agglutinated taxa at 2610 m may coincide with an increase in local WBUC circulation near the onset of ice rafting at 2.5 Ma. However, the exact chronology cannot be resolved.

One of the key environmental factors controlling the presence of Neogene flysch-type faunas in the high-latitude North Atlantic basins appears to be the local formation of bottom waters. The change from a predominantly agglutinated assemblage to a calcareous assemblage in the Pliocene is associated with the onset of drift sedimentation at Site 646. This change may reflect a change in deep-water properties and current intensities, which ultimately may be linked to the initiation of convective overturn of deep waters and the onset of vigorous circulation in the Norwegian-Greenland Sea. Agglutinated assemblages are present in

the Miocene to lower Pliocene of Norwegian-Greenland Sea sites, but the modern microfossil chronology of the area does not provide the resolution to determine if the LO of flysch-type taxa is isochronous in both regions. However, we speculate that the events are closely linked. Agglutinated assemblages persisted to younger stratigraphic levels in the deep Arctic Ocean. In some of the longer cores obtained by the Arctic Ocean CESAR Expedition, the Pliocene/Pleistocene boundary is marked by the last common occurrence (LCO) of agglutinated foraminifers. This event appears to be correlated with the onset of perennial sea-ice formation in the Arctic Ocean (Scott et al., 1987). The formation of dense waters through brine release from seaice formation apparently ventilates the deep basins. Therefore, the observed pattern of the LCO of flysch-type assemblages in the high-latitude North Atlantic basins appears to be consistent with the old bottom-water model of Gradstein and Berggren. We believe that the initiation of significant bottom-water formation by thermohaline processes resulted in the loss of agglutinated foraminifers from the high-latitude fossil record.

Pre-Glacial Pliocene Assemblage

The Pliocene assemblage above the LCO of agglutinated species in Hole 646B is unlike assemblages from the eastern Atlantic described by Murray (1984). The dominant species of the drift sediments are *Melonis barleeanum*, *Pullenia bulloides*, *Cibicidoides* spp., and unilocular species. *Islandiella teretis* and *Uvigerina peregrina* are sometimes common. The proportion of species that are typical of modern NADW, such as *E. exigua*, *P. wuellerstorfi*, and *G. subglobosa*, is small in comparison with pre-glacial Piocene assemblages in eastern Atlantic sites.

Melonis barleeanum (= M. zaandamae) is most common in high latitudes. It is the dominant calcareous species in the Miocene of Baffin Bay and in the rare intervals of calcareous sediments in the Norwegian-Greenland Sea. In the pre-glacial Pliocene of DSDP Site 343, this species occurs with *P. bulloides* (Talwani, Udintsev, et al., 1976).

The change from a pre-glacial Pliocene assemblage to a glacial assemblage is usually marked by a clustering of first and last occurrences, and species that are typical of modern low latitudes are replaced by boreal forms. This turnover was diachronous with latitude in the North Atlantic, and not suprisingly, began earlier at Site 646 than in the eastern Atlantic. For example, *Pleurostomella* disappeared in the early Pliocene at Site 646, but persisted until the Pliocene/Pleistocene boundary at Feni Ridge Site 610 (Thomas, 1987). In the eastern Atlantic, Pliocene peak benthic foraminifer turnover occured between 3.0 and 2.5 Ma at King's Trough Site 608, and between 3.3 and 2.4 Ma at shallower Site 610 (Thomas, 1987). At 1.5 Ma, *N. umbonifera* decreased in abundance in Site 608, signaling a retreat of corrosive water. In Hole 647A, this event occurs at about 47 mbsf, or 1.0 Ma.

Glacial Assemblages

At the sampling resolution undertaken here, biostratigraphic correlation of Pleistocene age assemblages among Leg 105 sites was not possible. Baffin Bay is an isolated basin with assemblages unlike those at any depth in the Atlantic. None of the Pleistocene biostratigraphic events recorded in our data have chronostratigraphic utility. However, the paleoecology of benthic assemblages determined at each site can be interpreted, based on correlations with site-survey piston cores (Scott et al., this volume).

The Baffin Bay piston core recovered a 12-m section of sediment covering a time span of about 30–50 k.y. Low numbers of foraminifers are associated with carbonate-rich turbidite layers, and higher abundances of foraminifers occur in the interbedded clays. In the piston core, *Stetsonia horvathi* is the dominant taxon throughout, with Buliminella hensoni as a secondary species. Elphidium, Cassidulina, and miliolid species are present in low numbers. The Stetsonia assemblage in the piston core is almost identical to that present in the today's Arctic Ocean below a depth of ~900 m (Lagoe, 1977). However, one should be careful when interpreting assemblages with abundant Stetsonia as indicators of Arctic deep-water infuence. The shallow sill depth between Baffin Bay and the Arctic Ocean makes the mixing of deep Arctic and Baffin Bay waters unlikely. In Hole 645B, Stetsonia was found in Cores 105-645B-1X to -8X, which suggests similar hydrographic conditions represented by the upper 80-90 m of Hole 645B to the present day. Epistominella takayanagii becomes the dominant taxon in samples from Cores 105-645B-8X to -23X, which suggests that paleoenvironmental conditions must have been different below Core 105-645B-8X. Although E. takayanagii is present in the piston core, it never dominates. The dominance of this taxon between Cores 105-645B-8X and -23X is unusual, as there are no known recent assemblages dominated by E. takayanagii. This species has been observed in low numbers in today's cold Gaspe Current in Chaleur Bay, New Brunswick, but not in deeper water. The only other reported occurrence of an E. takayanagii assemblage is in the Pleistocene of Sites 612 and 613 off New Jersey (Scott, 1987). In this area the species occurred over a wide depth range (2200-4400 m) in an otherwise glacial assemblage.

At Site 646, we recovered a short (5 m) piston core that only penetrated to the top of isotope stage 5 (see Scott et al., this volume). Interglacial assemblages in this core contain *E. exigua* and *E. bradyi* as dominant forms, and glacial assemblages consist of *P. subcarinata*, *Triloculina* spp., *Fursenkoina fusiformis*, and *S. horvathi*. Species of *Uvigerina* occur rarely in glacial sections. Both of these assemblages were observed in Cores 105-646A-1H to -6H. In particular, the presence of *E. exigua* suggests that the WBUC was present in interglacials during much of the Pleistocene at this water depth.

At Site 647, the site-survey core provided us with a high-resolution record back to isotope stage 7c. The faunal diversity at Site 647 was the highest of the three sites. Interglacial assemblages are characterized by *N. umbonifera* and *E. exigua*. These occur together with *Triloculina* sp. and *S. horvathi* in the upper part of stage 5. Glacial assemblages contain lower numbers of individuals and high percentages of *Uvigerina* spp., *F. fusiformis*, *Triloculina* spp., and *P. subcarinata*. This type of assemblage also was observed in low-resolution samples from Hole 647A in Cores 105-647A-1H to -7H. Although we did not observe *Uvigerina* in the upper five cores of Hole 647A, this form probably may have occurred with closer-spaced sampling, as it is present in the site-survey core. Site 647 contains the most complete Pleistocene stratigraphy, as well as assemblages that will allow detailed comparisons with other North Atlantic sites.

Deep-Water History of the Eirik Ridge

Modern Hydrographic Setting

At present, the deep-water masses in the northern Atlantic are derived from both northern and southern sources. Dilute Antarctic Bottom Water (AABW) is present below 2500 m in the Rockall Trough and Iceland basins and is presumed to flow along the eastern Atlantic margin in a weak cyclonic gyre (Mc-Cave and Tucholke, 1986). In the Rockall region, this water is joined by Norwegian Sea Deep Water (NSDW), which flows southward through the Faeroe-Shetland Channel and across the Wyville-Thompson Ridge and then turns northwestward to follow bathymetric contours around Rockall Bank. Additional amounts of NSDW flow across the Iceland-Scotland Ridge south of Iceland (Worthington and Volkmann, 1965). This flow travels along the eastern flank of the Reykjanes Ridge at a depth of 1300-2200 m and passes over the Garder Drift. The flow then descends and passes through the Charlie-Gibbs Fracture Zone into the Labrador Sea.

In today's Labrador Sea and Irminger Basin, the deep-water mass consists entirely of northern-source water. The Iceland-Scotland Overflow Water (ISOW) flows in a cyclonic loop along the western flank of the Reykjanes Ridge at a depth of around 3200 m (Worthington, 1976). Southwest of Iceland, ISOW is joined by cooler, denser Denmark Straits Overflow Water (DSOW), which descends beneath it. The DSOW is derived mainly from Arctic Intermediate Water, which forms by winter convection north of Iceland and overlies NSDW (Swift, 1986). This combined flow of ISOW and DSOW forms the Western Boundary Undercurrent, which flows southwestward along the eastern margin of Greenland and around the Eirik Ridge (Mc-Cave and Tucholke, 1986).

History of Deep Circulation

The dating of regional seismic reflectors at Site 646 reveals a much younger age for the Eirik Ridge than previously predicted. The Eirik Ridge was originally thought to be coeval with the Gloria Drift in the southern Labrador Sea, which is of Oligocene age (Tucholke and Mountain, 1986). Instead, the major period of drift sedimentation at Site 646 occurred in the Pliocene, although there is evidence of weaker currents occurring earlier. The seismic horizon marking the onset of drift deposition at this site is dated at approximately 4.5 Ma, based on the age-depth model for Hole 646B. Although there is no obvious hiatus in Hole 646B, a prominent seismic horizon observed within the drift sequence to the east of Site 646 was interpreted as indicating erosion. This horizon occurs between reflector RI and the base of the drift, and by correlation with the reflectors in Hole 646B we interpreted the age of this erosional pulse as occurring between 4.5 and 2.5 Ma. This compares well with an age of 3-2 Ma for the "Blue" erosional event (see Arthur et al., this volume) in the western North Atlantic reported by Tucholke and Mountain (1986).

The history of the deep circulation over the Eirik Ridge can be reconstructed by comparing benthic foraminifer assemblages at Site 646 with those recovered during DSDP Legs 38 (Norwegian-Greenland Sea) and 94 (eastern Atlantic). Benthic assemblages in Hole 646B display turnovers that may be interpreted as reflecting changes in water-mass properties. The occurrence of N. umbonifera with smooth agglutinated taxa in Hole 646B and N. umbonifera assemblages in nannofossil Zones NN10 to NN11 and in Holes 608, 609, and 611C in the eastern Atlantic (Murray, 1987a; Thomas, 1987) point to the presence of a sluggish. corrosive deep water mass in the northern Atlantic basins in the Tortonian. This finding is supported by benthic carbon-isotope studies of Miller et al., (1987), which indicate that at ~ 8 Ma, deep water in the North Atlantic did not differ isotopically from deep water in the Pacific. Murray (1987) interpreted the presence of N. umbonifera assemblages in DSDP Leg 94 holes as an indicator of "AABW-type" water in the North Atlantic in the late Miocene. The influence of corrosive deep water persisted into the early Pliocene at Site 611 on the Gardar Drift, where assemblages containing both N. umbonifera and modern NADW species were recovered. If ISOW were the predominant water mass over the Eirik Ridge in the early Pliocene, we might expect to find evidence of this entrained corrosive water reflected by assemblages in Hole 646B. However, the lack of any assemblages containing abundant N. umbonifera above seismic Unit 4 (Fig. 3, Table 6) argues against the influence of this water mass. The assemblages of seismic Unit 3 are composed mainly of coarse agglutinated species and Melonis and are unlike any other North Atlantic Miocene assemblages. Instead, the assemblages from seismic Unit 3 more closely resemble pre-glacial assemblages from intermediate and deep depths in the Norwegian-Greenland Sea. A likely cause of this pattern is that DSOW has been the dominant deep water mass over the Eirik Ridge since the late Miocene, displacing the corrosive bottom water in the Irminger Basin at approximately 7.5 Ma (Fig. 7). This interpretation is supported by both carbon-isotopic evidence and the presence of displaced microfossils. Miller et al. (1987) noted that the benthic carbon-isotope record at Site 608 diverged from the Pacific record between 8 and 6 Ma. At ~6 Ma, deep water in the North Atlantic was isotopically enriched with respect to Pacific water, indicating the production of "younger" deep waters analogous to modern NADW. Also, the reworked palynomorph Aquilapollenites was found above reflector R3 in Hole 646B, but not below (M. Head, pers. comm., 1987). Aquilapollenites is a high-latitude form known from the Cretaceous sediments of the northern North Sea, Greenland, and Svalbard (Batten, 1984). Sedimentologic evidence at Site 646 indicates increased bottom currents and increased sediment accumulation rates above the R3 seismic reflector (Srivastava, Arthur, et al., 1987).

The benthic foraminifer and seismic data indicate the period of maximum DSOW current velocities occurred between ~ 4.5 and 2.5 Ma. The disappearance of *Rhizammina* and the coarse grain size of sediments within the drift sequence is evidence of

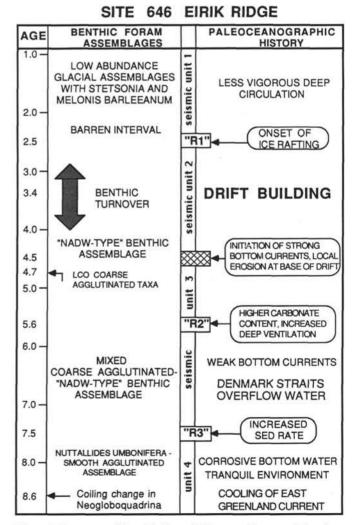


Figure 7. Summary of benthic foraminifer assemblages and the deep water history of the Eirik Ridge.

deposition of drift sediments under conditions of increased current velocities. This contrasts with the model of Miller and Tucholke (1983), who interpreted the onset of drift sedimentation to reflect a decrease and stabilization of abyssal circulation. The approximately coeval occurrence of hiatuses in the western North Atlantic and Norwegian-Greenland Sea also suggests increased current velocities at this time. At Sites 338, 342, 348, 350, and 647, glacial Pliocene-Pleistocene sediments rest on the Miocene, indicating a period of strong current activity possibly associated with the breakdown of a stable, stratified water column. The change from a coarse agglutinated assemblage in seismic Unit 3 of Hole 646B to a calcareous assemblage in seismic Unit 2 probably reflects the increased ventilation of deep water associated with this current activity.

Our observation of DSOW (since ~ 7.5) at Site 646 agrees with findings of Keigwin et al. (1987), who presented isotopic evidence for the continued production of NADW during the Messinian Salinity Crisis. Their results contradict earlier suggestions that the cutoff of Mediterranean Outflow Water during the Messinian should result in a cessation of NADW production in the Norwegian-Greenland Sea (Blanc and Duplessy, 1982). In the basal Pliocene to Messinian sediments beneath the sediment drift, Nuttallides umbonifera is present, but in lower proportions than before 7.5 Ma. The assemblage is still dominated by agglutinated taxa and species typical of modern NADW. The R2 reflector reflects a change from carbonate-poor sediments (below) to more carbonate-rich sediments (above). We interpreted this change as reflecting increased ventilation of the deep water at ~5.6 Ma. At Site 609 in the eastern Atlantic, an analogous change from poor carbonate preservation (below) to increased carbonate preservation (above) began at ~ 5.8 Ma and reached a maximum at ~5.3 Ma (Hooper and Weaver, 1987), which suggests that this may be a basinwide event. Hooper and Weaver interpreted the latest Miocene peak in carbonate preservation as indicating increased contribution of northern-source deep water to the basin at this time.

Increased production of NADW may have been caused by a number of oceanographic and climatic factors. Cooler late Miocene sea-surface temperatures at high latitudes, which culminated in the proposed 5.2 to 4.8 Ma glacial events reported by Keigwin et al. (1987) and Keigwin (in press), may have contributed to greater NADW production. Indeed, the appearance of DSOW at Site 646 is preceded by a coiling change in Neogloboquadrina atlantica (at ~8.6 Ma), which indicates cooling of the surface waters in the East Greenland Current (Aksu and Kaminski, this volume). With the reopening of the Mediterranean at 4.83 Ma (Zijderveld et al., 1986), the northward advection of saline water to the Norwegian-Greenland Sea, combined with cool sea-surface temperatures, may have resulted in the production of denser, more oxygenated intermediate and deep waters. This raises the possibility that Mediterranean water may have been a contributing factor to the presence of hiatuses, the onset of drift sedimentation at Site 646, and the LCO of agglutinated taxa. The period of drift formation also correlates with the maximum northward penetration of AABW observed in the eastern Atlantic in the early Pliocene by Murray (1987). This correlation suggests that a feedback mechanism or "teleconnection" between NADW and AABW may have operated in the early Pliocene analogous to today (see Johnson, 1982).

CONCLUSIONS

Throughout much of the stratigraphic record recovered at Site 645, benthic foraminifer assemblages are endemic and cannot be correlated with any site in the Atlantic. The Miocene foraminifer assemblage consists mainly of agglutinated foraminifers that have poorly constrained stratigraphic ranges. Scattered calcareous benthic species can be seen in the mid-Miocene, but these are boreal in nature. Above the barren interval in Site 645, a Pliocene to Pleistocene calcareous benthic assemblage dominated by *Epistominella takayanagii* was recovered. This type of assemblage has no modern analog. The Pleistocene *Stetsonia* assemblage in the upper 90 m of Site 645 indicates a deep environment similar to today's Baffin Bay.

Drilling at Site 646 recovered benthic foraminifer assemblages that provide information for interpreting the depositional environment and water mass history of the Eirik Ridge (Fig. 7). Before ~7.5 Ma, an assemblage dominated by Nuttallides umbonifera and smooth agglutinated species points to a tranguil environment with corrosive deep water. After ~7.5 Ma, Denmark Straits Overflow Water was present over the Eirik Ridge. Above the R3 seismic reflector, the benthic assemblage consists mainly of coarse agglutinated taxa with affinities to the Norwegian-Greenland Sea. The onset of drift sedimentation at ~ 4.5 Ma was accompanied by a change to a predominantly calcareous assemblage. Another faunal turnover took place between 3 and 4 Ma, as low-latitude species disappeared. This turnover appears to be diachronous from west to east, beginning in the early Pliocene at Site 646 and in the late Pliocene at DSDP sites in the eastern Atlantic. Drift formation ceased at ~2.5 Ma, concomitant with the first occurrence of ice-rafted sediments. Benthic foraminifer species that have arctic affinities first appeared in the early Pleistocene.

Pliocene and Pleistocene benthic foraminifer assemblages at Site 647 differ from those recovered at the other Leg 105 sites in containing N. *umbonifera*. These assemblages provide the closest analog to other North Atlantic sites, with glacial and interglacial benthic assemblages most similar to those reported in other parts of the North Atlantic (e.g., Schnitker, 1974). Site 647 also contains an adequate planktonic microfossil and magnetic reversal record, which allowed more accurate age determinations than at the other two sites.

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Abbreviations of species names: Ebrd = Eggerella bradyi (Cushman), SIPH = Siphotextularia spp., MART = Martinotiella spp., Kbrd = Karreriella bradyi (Cushman), Ssch = Sigmiolopsis schlumbergeri (Silvestri), LAGM = Lagenammina sp., RHIZ = Rhizammina spp., PSAM = Psammosphaera fusca Schultze, AMMB = Ammobaculites spp., ASCH = Aschemonella spp., TROC = Trochammina spp., RECV = Recurvoides spp., Cpus = Cyclammina pusilla (Brady), HAPL = Haplophragmoides sp., REOP = Reophax pilulifer Brady, Khor = Karreriella horrida Mjatliuk, AMMS = Ammosphaeroidina sp., TROI = Trochamminoides sp., ADER = Adercotrema, glomerata, (Brady), AMMD = Ammodiscus spp., GLOM = Glomospira gordialis (Jones and Parker), Ccan = Cyclammina cancellata Brady, PYRG = Pyrgo murthina (Schwager), Qsem = Quinquiloculina seminula (Linne), Qang = Quinqueloculina sp. (angular), Qrnd = Quinqueloculina sp. (rounded), Tarc = Triloculina arctica (Cushman), Ttri = Triloculina trihedra Loeblich and Tappan, OPTH = Opthalmidium sp., ORID = Oridorsalis umbonatus (Reuss), Mpom = Melonis pompilioides (Fichtel and Moll), Psub = Pullenia subcarinata (d'Orbigny), CIB = Cibicidoides sp. (transitional between C. bradyi and P. wuellerstorfi), STIL = Stilostomella spp., DENT = Dentalina spp., FISS = Fissurina spp., OOLN = Oolina spp., LAGN = Lagena spp., Gaur = Globobulimina auriculata (Bailey), Gpac = Globobulimina pacifica Cushman, Pwue = Planulina wuellerstorfi (Schwager), ELPH = Elphidium spp., ?ENT = entosolenian, possibly Glandulina sp., Mbar = Melonis barleeanum (Williamson), GOID = Gyroidinoides spp., NODO = Nodosaria spp., Nstr = Nodosaria sp. (striate), SARA = Saracenaria sp., UVIG = Uvigerina peregrina Cushman, Usma = Uvigerina sp. (small), Numb = Nuttallides umbonifera (Cushman), PSPL = Pseudopolymorphina sp., Pbul = Pullenia bulloides (d'Orbigny), LENT = Lenticulina spp., POLY = polymorphinids, Hele = Hoeglundina elegans (d'Orbigny), LATI = Laticarinina pauperata (Parker and Hones), Eexi = Epistominella exigua (Brady), Gsub = Globocassidulina subglobosa (Brady), EPON = Eponides spp., TOSA = Tosaia hanzawai Takayanagi, FURS = Fursenkoina fusiformis (Williamson), Iter = Islandiella teretis (Tappan), Bfri = Buccella frigida (Cushman), GYRO = Gyroidina orbicularis d'Orbigny, Clob = Cibicidoides lobatulus (Walter and Jacob), PLEU = Pleurostomella spp., SPHA = Sphaeroidina bulloides d'Orbigny, FRON = Frondicularia spp., BRIZ = Brizalina spp., Npyr = Nodosaria pyrula d'Orbigny, Cbrd = Cibicidoides bradyi (Trauth), CIBs = Cibicidoides spp. ind., Bala = Bulimina alazanensis Cushman, Prnz = Planulina renzi Cushman and Stainforth, NONI = Nonion spp., $H^1 =$ Shannon-Wiener Diversity.

APPENDIX Benthic Foraminifers in Hole 646B

11.1 11.3 12.4 11.3 12.4 <th< th=""><th>ADER AMM</th><th>TROI</th><th>AMMS</th><th>Khor</th><th>REOP</th><th>HAPL</th><th>Cpus</th><th>RECV</th><th>TROC</th><th>ASCH</th><th>AMMB</th><th>PSAM</th><th>RHIZ</th><th>LAGM</th><th>Ssch</th><th>Kbrd</th><th>MART</th><th>SIPH</th><th>Ebrd</th><th>Depth (mbsf)</th><th>Interval (cm)</th><th>Core</th></th<>	ADER AMM	TROI	AMMS	Khor	REOP	HAPL	Cpus	RECV	TROC	ASCH	AMMB	PSAM	RHIZ	LAGM	Ssch	Kbrd	MART	SIPH	Ebrd	Depth (mbsf)	Interval (cm)	Core
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3h.3 MS-110 MS-29 0 <	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	161.04	44-46	18-2
11-4 11-12 19-18 0 </td <td>0 0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>õ</td> <td>0</td> <td>0</td> <td>ō</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>185.59</td> <td>108-110</td> <td>20-5</td>	0 0	0	0	0	0	0	0	0	0	õ	0	0	ō	0	0	0	0	0	0	185.59	108-110	20-5
1236 M-46 XX:14 0 0 0 0	0 0	ō	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	193.89	118-120	21-4
134 12-12 14.72 2 0 <th< td=""><td>0 0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>206.14</td><td>84-86</td><td>22-6</td></th<>	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	206.14	84-86	22-6
51.CC 236.00 0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	0	0	0	2	214.72		23-5
asks 15-12 24.35 0 </td <td>0 0</td> <td>0</td> <td>236.10</td> <td></td> <td>25, CC</td>	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	236.10		25, CC
Sh2 H-16 26.0 0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	243.35		26-5
Shore Table 0 0 <th< td=""><td>0 0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>266.94</td><td></td><td>29-2</td></th<>	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	266.94		29-2
385 6:-65 281.43 0 <t< td=""><td>0 0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>1</td><td>0</td><td>0</td><td>0</td><td>3</td><td>275.00</td><td></td><td>29, CC</td></t<>	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	3	275.00		29, CC
11.6 76-78 202.77 0 <	0 0	0	0	0	0	0	0	0	0	0	0	0	õ	0	0	0	0	0	0	281.63	63-65	30-5
312 384-60 386-64 396-64 0	0 0	0	0	0	0	0	0	0	0	ō	õ	0	0	0	0	0	0	0	0	292.97		31-6
313-2 4-6 35.4 0	0 0	0	0	0					1.00					0	0	0	0	0	0	296.48		32-2
33, CC 33, 8 0	0 0 0	0											1.2.1					100		305.64	4-6	33-2
35-3 8-10 33-46 0 0 1 0 <th< td=""><td>0 0</td><td>0</td><td></td><td></td><td></td><td></td><td></td><td></td><td>100</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>0</td><td>313.8</td><td></td><td>33, CC</td></th<>	0 0	0							100										0	313.8		33, CC
36-6 18-140 341.99 1 0 0 1 0	0 0	0					0.72						1.77					100				
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0									27		-									
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0 0	0						0											100			
38-7 28-30 36.49 0 <t< td=""><td>0 0</td><td>0</td><td></td><td>0</td><td></td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td></td><td>1</td><td>0</td><td>6</td><td></td><td></td><td>4</td><td></td><td>352.40</td><td></td><td>37, CC</td></t<>	0 0	0		0		0	0	0	0	0	0		1	0	6			4		352.40		37, CC
39-5 11-13 368.12 2 0 0 1 0 <	0 0	0			0	0	0	0	0	0	0	0		0	0	0						38-7
	0 0	0	0		0	0	0	0	0	0	0		- T. I.	0	1		0		1224	368.12		39-5
	0 0	0	0	0	0	0	0	0	Ō	ō	0	0	0	0	0	0	0	0	2	373.35		40-2
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	3	387.41		41-5
	0 0	0	0	0	0	0	0	1	0	0	0	1	30	0	0	0	2	0	0	391.65		42-1
	0 0	0	0	0	0	0	0	0	0	0	0	0	2	0		0	2	1	8	404.24		43-3
	0 0	0	0	0	0	0	2	3	2	0	0	15	95	2	0	0	0	0	9	429.80	12.14	45, CC
	4 1 0 0	ō	0	0	1	0	13	4	3	0	3	27	90	4	0	0	ō	0	0	449.10		47, CC
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	2	0	0	4	2	ĩ	0	2		151	2	1	0	3	0	3	468.50		49, CC
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	õ	Õ	0	0	0	3	1	0	0	0		126		0	0	0	0	0	480.21		51-2
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	õ	õ	0	0	0	0	0	0	0	0	0	1		3	0	0	1	5	497.10		52, CC
56-1 40-42 527.61 0 0 0 0 0 0 0 0 0	0 0	0	0	ō	0	0	0	1	0	0	0	1	28	0	0	0	0	0	0	519.84	33-35	55-3
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	Ō	0	1	1	0	1	0	0	0	1	51	0	0	0	0	0	0	526.61		56-1
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	0	0	0	0	0		0	0	4	0	0	0	0	1	0	0	545.50		57, CC
	0 0	0	0	0	0	0	3	0	100	0	0	2	121	1	0	0	0	0	0	564.40		59, CC
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	0	0	0			1.2		0		184	3	0	0	0	0	0	566.50	59-61	60-2
	0 0	0	0	0	õ	o					0		24	õ	0	0	2	0		583.60		61, CC
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	0	0	0	4	7	1	0	0	10	300	1	1	0	1	4	24	590.15	54-56	62-5
	0 0	0	3	0	0	0	1	4	0	0	0	2	60	0	0	0	0	0	0	602.90	14-16	63, CC
	2 1 0 0	0	0	0	1	0	0	0	0		0	0	34	0		1	0	0	1	614.04		65-2
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0 0 0	0	0	0	0	0	0	0					90	0	0	0	1	0	3	624.05		66-2
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1 0 0 0	0 4	0	4	2	0	20	5	14	2	3		290	7	0	0	0	0	0	634.42		67-2
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0	0	0	0	1	0	0	0	0		0	1 2		1	0	0	0	1	4	642.92	11-13	68-2
71-2 9-11 671.80 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	1 0 0 0	1	0	1	2		5	0		0	2	20		6	0	2	0	0		651.24	33-35	69-1
72-1 60-62 680.31 8 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0	0	0	0	0	0	0	0	0		0	0	0	0	0	0	0	1	8	680.31	9-11 60-62	72-1
72-2 127-129 682.48 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 1 0 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	682.48	127-129	72-2
73, CC 699.00 9 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 2 0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	9	699.00		73, CC
74-4 10-12 703.61 4 2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 1 0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	4	703.61	10-12	74-4
75-4 87-89 713.98 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	713.98	87-89	75-4
77-4 37-39 732.78 4 0 0 0 0 0 15 1 0 0 0 2 4 0 0 0 0 0	0 2 0 5 0 7	0	0	0	0	0	4	2	0	0	0	1	15	0	0	0	0	0	4	732.78	37-39	77-4
77, CC 737,70 0 0 0 0 0 0 8 0 0 0 3 3 0 0 0 0 0	0 0 0 5	0	0	0	0	0	3	3	0	0	0	0	8	0	0	0	0	0	0	737.70		77, CC
78-2 40-42 739.61 6 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 6 0 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6	739.61	40-42	78-2
	0 4	ŏ																			140.142	

Appendix (continued).

Core	Interval (cm)	Depth (mbsf)	GLOM	Ccan	PYRG	Qsem	Qang	Qrnd	Tarc	Ttri	OPTH	ORID	Mpom	Psub	CIB	STIL	DENT	FISS	OOLN	LAGN	Gaur	Gpac	Pwue
12-2	11-13	103.06	0	0	0	0	0	0	0	0	0	7	1	1	2	1	0	0	0	0	0	0	0
12-5 13-2	22-24 61-63	104.62 113.21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13-5	88-80 81-83	114.98 124.61	0	0	0	0	0	0	0	0	0	3	2	4	4	0	1	1	1	0	1	1	1
18-1 18-2	144-146 44-46	160.45 161.04	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20-2 20-5	7-9 108-110	180.08 185.59	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21-2 21-4	118-120 118-120	190.92 193.89	0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22-2	7-9	199.38	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22-6 23-2	84-86 8-10	206.14 209.08	ō	0	1	0	0	0	0	0	0	0	3	0 2	0	0	0	1	2	0	0	0	0
23-5 23, CC	122-124	214.72 217.10	0	0	0	0	0	0	0	0	0	0 3	2 0	0 5	1 4	02	1	3 2	1	4	0	0	0
25, CC 26-2	20-22	236.10 237.80	0	0	0	0	0	0	0	0	0	0	0	0	03	0	0	0	0	0	0	02	0
26-5 27, CC	125-127	243.35 255.70	0	0	0	0	0	0	0	0	0	0	2	0	0	0	0	0	0	20	0	0	0
29-2 29-5	14-16 88-90	266.94 272.18	0	0	0	0	0	0	0	0	0	0 2	0	0	0	03	0	0	0	02	0	02	0
29, CC 30-2	73-75	275.00 277.23	0	0	0	0	0	0	0	0	0	0	0	5	1	0	1	1	0	4 2	2	3	1
30-5 31-2	63-65 10-12	281.63 286.30	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31-6	76-78	292.97	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0 3	0	0	0	0	0
31, CC 32-2	58-60	294.40 296.48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
32-6 33-2	98-100 4-6	302.88 305.64	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
33-5 33, CC	129-131	311.39 313.8	0	0	3	0	0	0	0	0	2 0	0 5	0	4 0	0 3	0	0	1 2	1	1	0	0	2
35-2 35-3	9-11 8-10	324.99 326.48	0	0	0	0	0	0	0	0	0	0	1	0	1 2	0	0	0	0	1 2	0	0	0
36-2 36-6	5-7 138-140	334.66 341.99	0	0	0	0	0	0	0	0	0	0	5	1	0 5	0	1 2	0 5	0	0 3	0	0	1 2
37-2 37-7	16-18 16-18	344.47 351.96	0	0	2	0	0	0	0	0	0	0	0	0	0 2	0	1	1	0	23	0	0	0
37, CC 38-2	14-16	352.40 354.05	0	0	6 8	0	8	1	0	0	0	9	10	4	35	0	6 0	5	13	7	0	0	8
38-7	28-30	361.69	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
39-2 39-5	15-17 11-13	363.66 368.12	0	0	0	0	0	0	0	0	0	2	5	1	75	0	0	0	3	3 2	0	0	o
39, CC 40-2	14-16	371.70 373.35	0	0	0	0	0	0	0	0	0	3	3	2	3	0	0	4	3	3 0	0	0	4
40-5 41-5	127-129 10-12	378.98 387.41	0	0	0 5	0	0	0	0	0	1	3 4	2	4	777	0	2 2	0	3	0	0	0	0
41, CC 42-1	64-66	391.00 391.65	0	0	1	0	2	0	0	0	0	13 0	8	0	- 40	0	2	12 0	4	3	0	0	2
42-5 43-3	89-91 53-55	397.90 404.24	0	0	0	0	0	0	0	0	0	0	14	3	17 9	0	0	1	4	3	0	0	5
43, CC		410.40	0	0	2	0	0	1	0	0	0	14	14	0	6	0	2	2	2	1	0	0	5
45, CC 46-2	42-44	429.80 431.73	0	0	2	0	1	0	0	0	1	12	6	2	0	0	2	1	2	Ó	0	0	1
47, CC 49-2	54-56	449.10 460.85	0	0	0	0	0	0	0	0	0	0 25	47	1	02	0	1	04	3	0	0	21	9
49, CC 50-2	10-12	468.50 470.11	0	0	1 2	0 2	2 0	2	0	0	2	73	4 2	1	1	0	4 0	0	2 4	23	0	4 44	8
51-2 51, CC	60-62	480.21 487.40	0	0	0	0	03	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0
52, CC 53, CC		497.10 506.90	0	0	0	0	0	6	0	0	4	7	16 5	0	8	0	6 0	0	2	5	0	3	8
55-3 55, CC	33-35	519.84 526.20	0	0	5 0	0	5 0	5	0	0	0	3 2	7	0	0 2	0	4	1	0	0	0	0	0
56-1 56-2	40-42 4-6	526.61 527.75	0	0 0	0	0	0	0	0	0	0	1	0	0 2	0 2	0	0	0	5	0	0	0	0
57, CC		\$45.50	0	0	0	0	0	0	0	0	0	0	2	0 0	00	0	0	0	0	0	0	0	0
58-1 59, CC	59-61	546.10 564.40	0	0	0	1	0	0 2	0	0	0	0	3	0	4	0	0	0	1	0	0	0	1
60-1 60-2	77-79 59-61	565.18 566.50	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
61-2 61, CC	16-18	575.67 583.60	0	0	0	0	0	0 2	0	0	0	03	0 8	0	0 5	2	0	0	0	1	0	0	2
62-2 62-5	2-4 54-56	585.13 590.15	0	0	03	0	0	0	0	0	0	4 27	6	3 0	02	0	1	4	5	9 0	0	0	02
63-2 63, CC	14-16	594.85 602.90	0	0	0	0	0	0	0	0	0	0 2	0	0	0	0	0	0	1	1	0	0	0
64, CC 65-2	3-5	612.5 614.04	0	0	0	0	2 0	03	0	0	1	6 2	5 2	1	2	2	2	1	3	0	0	0	0
65-4 66-2	103-105 34-36	618.04 624.05	0	0	0	0	0	0	0	0	0	3	5 23	0 2	0	0	0	1	4	2 0	0	0	1
66, CC 67-2		631.8	0	0	0	0	1	0	0	0	1	6	3	0	0	0	0	0	2	0	0	0	3 0
67-5	111-113 106-108	634.42 638.87	0	0	0	0	0 2	0 7	0	0	0	3	1	1	1	0	0	1	3	1	0	0	03
68-2 68-3	11-13 47-49	642.92 644.78	0	0	00	0	2 0	3	0	0	0	1	7	0	0	0	0	0	1	0	0	0	0
69-1 71-2	33-35 9-11	651.24 671.80	0	0	1	0	0	0	0	0	0	2 4	3	1	0	0	0	1 2	1	2	0	0	0
72-1 72-2	60-62 127-129	680.31 682.48	0	0	0	0	1 0	0	0	0	0	5	7	0	6	0	0	3 0	1	1	0	0	1
73-2 73, CC	58-60	691.39 699.00	0	0	2	0	1 2	0	0	0	0	40 26	4 3	14 1	8 4	0	0 1	6 2	4 2	5 3	0	0	9 5
74-2 74-4	0-2 10-12	700.51 703.61	0	0	1	1	9	2	0	0	0	3	1	3 15	0	0	1	3	4	2 4	0	0	5 0
75-2	88-90	710.99	0	0	0	0	0 3	0	0	0	0	0	0	0	0	0	0	1 0	3	0	0	0	0
75-4	87-89 17-19	713.98	0	0	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
77-4 77-5	37-39 25-27	732.78 734.16	3	4	0 4	0	1 3	0 4	0	0	3	2	9	0	05	0	0	4	4 3	0	0	0	2
77, CC 78-2	8-10	737.70 739.29	0	0	0	0	1 0	0	0	0	0	3 1	2 3	0	0	1	0	0	03	0	0	0	1
78-2 79-2	40-42 140-142	739.61 750.31	1	3 0	1	0 1	0 2	4 0	0	0	0	0 7	1 4	0 0	0	0	2	1	0	0	0	0	0
80, CC	1997-1992	766.1	9	0	0	0	4	0	0	0	0	2	2	1	5	0	t.	0	3	0	0	0	0

Appendix (continued).

Core	Interval (cm)	Depth (mbsf)	ELPH	?ENT	Mbar	GOID	NODO	Nstr	SARA	UVIG	Usma	Numb	PSPL	Pbul	LENT	POLY	Hele	LATI	Eexi	Gsub	EPON	TOSA	FURS
1-2	11-13	103.06	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2-5 3-2	22-24 61-63	104.62	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
8-5	88-80	114.98	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	ő	0	0	0	0	0
1-5 1-1	81-83 144-146	124.61 160.45	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
8-2	44-46	161.04	0	0	0	1	0	0	0	0	0	0	0	0	0	ő	0	0	0	ő	0	o	ő
0-2 0-5	7-9 108-110	180.08 185.59	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1-2	118-120	190.92	ŏ	ő	0	õ	3	0	0	0	0	0	0	0	0	0	0	0	0	o	0	0	o
2-2	118-120 7-9	193.89 199.38	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2-6	84-86	206.14	ő	ŏ	ő	ő	0	0	0	0	0	0	0	0	0	0	0	0	õ	ŏ	0	o	õ
3-2 3-5	8-10 122-124	209.08 214.72	0	0	1 2	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0
, CC		217.10	11	o	6	ŏ	o	õ	0	õ	0	7	ò	5	1	3	1	2	2	0	õ	o	ő
5, CC 5-2	20-22	236.10 237.80	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
5-5	125-127	243.35	ő	2	ò	õ	0	õ	0	õ	õ	0	ő	0	õ	0	0	0	õ	10	o	o	õ
7, CC	14-16	255.70 266.94	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
9-5	88-90	272.18	õ	1	4	õ	0	ŏ	ő	ŏ	õ	ŏ	0	3	0	0	0	0	0	7	õ	õ	0
9, CC)-2	73-75	275.00 277.23	2	1	15	0	0	0	0	0	0	1	0	10 0	2	0	0	0	0	0	1	1	1
)-5	63-65	281.63	0	ò	0	o	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	õ
-2 -6	10-12 76-78	286.30 292.97	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
, CC		294.40	ö	1	3	ő	0	0	0	0	0	0	0	2	õ	o	o	õ	õ	ō	õ	ö	ŏ
2-2	58-60 98-100	296.48 302.88	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
3-2	4-6	305.64	0	ő	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	o
3-5 3, CC	129-131	311.39 313.8	0	2	0 21	0	0	0	0	0	0	0	0	0 8	0	0	8	0	0	6 8	0 2	0	0
-2	9-11	324.99	ò	õ	0	õ	0	0	0	0	0	1	0	1	0	0	1	ő	0	ô	õ	o	ő
-3	8-10 5-7	326.48 334.66	0	0	7	0	0	0	0	1	0	0	0	2	0	0	0	0	0	0	0	1	0
i-6	138-140	341.99	0	0	15	0	03	0	0	20	02	0	0	8	0	0	0	0	0	1	0	1	0
-2 -7	16-18 16-18	344.47 351.96	1	0	3	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
CC	10-10	352.40	0	0	0 71	0	0	0	0	0	03	0	0	2 9	0	0	0 4	2 8	0	0	0	0	0
-2 -7	14-16	354.05	0	0	5	0	0	0	0	0	1	0	0	1	0	0	0	0	0	1	0	0	0
-2	28-30 15-17	361.69 363.66	0	0	0 23	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
-5	11-13	368.12	0	0	4	0	0	0	1	õ	0	2	0	0	0	0	0	0	0	0	0	0	0
, CC	14-16	371.70 373.35	0	0	10 2	0	0	0	0	1	0	1	0	5	0	0	0	2	0	0	0	0	0
-5	127-129	378.98	0	0	21	1	2	0	0	2	0	0	0	1	0	0	0	0	0	0	0	0	0
. CC	10-12	387.41 391.00	0	2	12 43	0 2	1	4	0	0	0	0	0	2 22	0	0	1	2	0	0	0	0	0
2-1	64-66	391.65	0	0	6	õ	0	0	0	0	õ	õ	0	0	ō	0	0	0	0	0	0	0	0
1-5 1-3	89-91 53-55	397.90 404.24	0	0	21 27	0	0	0	0	0	1	0	0	4	1	0	15 16	0	4	0 4	0	0	0
, CC	240750	410.40	8	0	14	0	0	0	o	2	4	o	0	13	0	õ	9	i	2	0	0	0	0
5, CC	42-44	429.80 431.73	1	1	4	2	0	0	0	2	0	9	0	4	0	1	0	2	0	2	0	0	0
, CC		449.10	õ	õ	0	õ	õ	0	0	3	ő	1	0	0	ő	õ	ő	ò	õ	õ	ŏ	õ	0
0-2 0, CC	54-56	460.85 468.50	9	2	30 38	2	0	0	0	0	1	6	0	27	0	0	0	9	1	1	0	0	0
-2	10-12	470.11	3	ò	11	ò	0	0	0	0	0	3 4	0	10 14	0	0	0	2	0	4	õ	ō	õ
-2 , CC	60-62	480.21 487.40	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	2	0	0	0
, CC		497.10	0	ŏ	25	0	0	0	0	3 80	0	0 23	0	16	o	õ	0	õ	0	6	õ	õ	o
. CC -3	33-35	506.90 519.84	0	0	9	0	03	0	0	9	0	6	0	4	0	0	0	0	0	1	0	0	0
, CC		526.20	0	0	2	0	0	0	0	0	4	2	0	4	ò	0	0	õ	0	3	õ	o	0
-1	40-42 4-6	526.61 527.75	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
, CC		545.50	ő	o	4	1	o	0	0	0	õ	2	0	1	0	0	0	0	î.	0	0	o	0
-1 , CC	59-61	546.10 564.40	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
-1	77-79	565.18	1	0	õ	0	0	0	0	0	õ	0	0	1	0	0	0	0	0	0	0	0	0
-2	59-61 16-18	566.50 575.67	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
CC		583.60	0	0	6	õ	0	0	0	õ	0	0	0	8	0	0	0	1	0	1	0	2	0
2 5	2-4 54-56	585.13 590.15	0	0	14 17	1	0	0	0	3 10	03	0	0	17 15	1	2	0	2	0	2	0	1	0
2	14-16	594.85	0	0	0	0	0	0	õ	0	0	0	0	2	0	0	0	0	0	0	0	0	0
CC		602.90 612.5	0	0	11	0	0	0	0	2	0	0	0	04	0	0	0	0	0	0	0 2	0	0
2	3-5	614.04	0	0	2	1	2	0	0	1	0	0	0	3	0	0	0	0	0	0	0	0	0
4	103-105 34-36	618.04 624.05	1	0	9	0	0	0	0	0	1	0	0	6	0	2	0	0	1	0	0	0	0
CC		631.8	0	0	17	1	17	0	1	1	0	1	0	5	2	0	0	0	0	2	0	0	0
2	111-113 106-108	634.42 638.87	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	11-13	642.92	0	0	1	ő	0	1	0	0	õ	ò	0	4	0	0	0	1	1	0	0	0	0
	47-49 33-35	644.78 651.24	1	0	0	0	0	0	0	0	0	1	0	0 4	0	0	0	0	0	0	0	0	0
2	9-11	671.80	0	0	13 4	0	0	0	0	0	0	3	0	6	0	0	0	1	0	1	0	0	0
	60-62 127-129	680.31 682.48	0	0	5	0	0	0	0	0	0	19	1	97	0	0	0	3 0	0	6 2	0	0	0
2	58-60	691.39	0	0	2 39	0	10 0	0	0	0	0	45	0	35	0	2	0	3	0	4	5	2	0
,CC	0-2	699.00 700.51	0	0	23	1	0	0	0	14	0	37	0	21	0	0	0	3 0	0	10 2	1	0	0
2 4	10-12	700.51 703.61	0	0	23	0	0	0	0	4	02	33 30	0	16 9	0	0	0	1	0	3	0	0	0
2	88-90	710.99	0	0	5	0	0	0	0	0	0	3	0	2	0	0	0	0	0	0	0	0	0
4	87-89 17-19	713.98 719.98	0	0	0	0	1	0	0	0	0	14 12	0	0	0	0	0	3 0	0	0	0	0	0
4	37-39	732.78	1	0	6	0	6	0	0	1	0	21	0	6	0	1	0	0	2	0	0	0	0
5 CC	25-27	734.16 737.70	0	0	0	0	0	0	0	0	2	0	0	2 4	0	0	0	0	03	0	0	0	0
		739.29	1	0	0	0	0	0	0	0	0	10 13	0	0	0	0	0	0	0	3	0	0	0
2	8-10																						
2 2 2 2	8-10 40-42 140-142	739.61 750.31	0	0	3 0	0	0	0	0	0	1	2 17	0	1	0	0	0	2 4	0	0 4	0	0	0

Appendix (continued).

Core	(cm)	Depth (mbsf)	Iter	Bfri	GYRO	Clob	PLEU	SPHA	FRON	BRIZ	Npyr	Cbrd	CIBs	Bala	Prnz	NONI	Sum	H
2-2	11-13	103.06	0	0	0	0	0	0	0	0	0	0	0	0	0	0	16	1.
2-5	22-24 61-63	104.62 113.21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
-5	88-80	114.98	ő	õ	0	ő	o	ő	o	õ	õ	ŏ	õ	o	ŏ	ŏ	23	2.
-5	81-83	124.61	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
-1	144-146	160.45	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	44-46 7-9	161.04 180.08	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
5	108-110	185.59	ő	0	0	0	0	ő	ő	0	o	0	0	õ	õ	õ	0	0
2	118-120	190.92	0	õ	õ	õ	0	o	õ	õ	õ	õ	õ	0	õ	0	5	0.
4	118-120	193.89	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	7-9	199.38	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
6	84-86 8-10	206.14 209.08	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
5	122-124	214.72	õ	ő	0	0	0	õ	õ	õ	õ	õ	õ	õ	õ	õ	25	2
CC		217.10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	63	2
CC	21272220	236.10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	20-22	237.80	0	0	0	0	0	0	0	0	0	0	0	0	0	0	16	1
5 CC	125-127	243.35 255.70	0	0	0	0	0	0	0	0	o	õ	0	0	õ	0	16	1
2	14-16	266.94	õ	õ	õ	õ	o	0	õ	ŏ	õ	õ	õ	0	õ	0	Ő	Ő
5	88-90	272.18	0	0	0	0	0	0	0	0	0	0	0	0	0	0	27	2
CC	1.000	275.00	0	0	0	0	0	0	0	0	0	0	0	0	0	0	56	2
2	73-75	277.23	0	0	0	0	0	0	0	0	0	0	0	0	0	0	11	1
5	63-65 10-12	281.63 286.30	0	0	0	0	0	0	0	0	0	0	0	ő	0	0	0	0
6	76-78	292.97	õ	õ	0	0	o	0	õ	0	ő	õ	0	ŏ	ŏ	o	0	ŏ
CC	374,554	294.40	7	0	0	0	0	0	0	0	0	0	0	0	0	0	20	1
2	58-60	296.48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	3	1
6	98-100	302.88	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
2	4-6 129-131	305.64 311.39	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2 43	02
cc	129-131	311.39	10 17	0	0	0	0	0	0	0	0	0	0	0	0	0	43	2
2	9-11	324.99	ï	0	o	o	õ	ò	õ	0	õ	0	0	0	0	0	7	1
3	8-10	326.48	3	0	0	0	0	0	0	0	0	0	0	0	0	0	24	2
2	5-7	334.66	5	0	0	0	0	0	0	0	0	0	0	0	0	0	21	1
5	138-140 16-18	341.99 344.47	0	2	1	1	0	0	0	0	0	0	0	0	0	0	86 26	2
7	16-18	344.47	0	0	0	0	1	0	0	ő	0	0	0	ő	õ	0	14	2
CC		352.40	õ	õ	ő	1	7	2	0	0	1	0	0	0	0	0	251	2
2	14-16	354.05	0	0	0	0	0	1	0	0	0	0	0	0	0	0	28	2
7	28-30	361.69	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
2	15-17	363.66	1	0	0	1	1	0	1	0	0	0	0	0	0	0	70 20	2 2
CC	11-13	368.12 371.70	0	0	0	0	0	0	0	0	0	ő	0	õ	0	õ	49	2
2	14-16	373.35	0	ő	0	0	0	o	ŏ	ĩ	ŏ	ŏ	ŏ	õ	ŏ	õ	11	2
5	127-129	378.98	0	0	õ	õ	0	0	0	0	0	0	0	0	0	0	53	2
5	10-12	387.41	0	0	0	0	3	0	1	0	7	0	0	0	0	0	64	2
CC	1000000	391.00	0	0	0	0	0	3	0	0	1	0	4	0	0	0	538	1
1	64-66 89-91	391.65 397.90	0	0	0	0	03	0	0	0	0	0	0	0	0	0	45	1 2
3	53-55	404.24	0	0	0	0	2	0	0	0	1	0	õ	õ	õ	õ	113	2
CC	33-32	410.40	ő	0	7	ő	4	1	õ	1	ò	5	0	0	0	õ	147	3
CC		429.80	0	o	0	õ	0	0	0	0	0	0	0	0	0	0	183	2
2	42-44	431.73	0	0	5	0	0	0	0	0	2	1	0	0	0	0	114	2
CC		449.10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	160 263	1
2 CC	54-56	460.85 468.50	0	0	3	0	1	3	0	0	0	5 2	3	ő	0	0	289	2 2
2	10-12	470.11	ő	0	3	õ	0	õ	1	õ	õ	õ	1	õ	õ	õ	228	2
2	60-62	480.21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	139	0
CC		487.40	0	0	0	0	0	0	0	0	0	0	0	0	0	0	138	1
CC		497.10	0	0	0	0	1	0	0	0	0	9	0	0	0	0	241 45	22
CC	33-35	506.90 519.84	0	0	1	0	1	0	0	0	0	3	0	0	õ	ő	89	2
CC	33-33	526.20	0	õ	0	0	õ	õ	0	õ	õ	ō	õ	õ	0	0	40	1
1	40-42	526.61	0	0	0	õ	0	0	0	0	0	0	0	0	0	0	62	0
2	4-6	527.75	0	0	0	0	0	0	0	0	0	0	0	0	0	0	26	1
CC	10 11	545.50	0	0	0	0	0	0	0	0	0	1	0	0	0	0	14 87	20
cc	59-61	546.10 564.40	0	0	0	0	0	0	0	0	0	0	0	0	0	0	153	1
LCC I	77-79	564.40 565.18	0	0	0	0	0	0	0	0	õ	0	ő	0	0	õ	166	0
2	59-61	566.50	õ	0	0	ŏ	ŏ	õ	0	õ	õ	õ	o	õ	õ	õ	192	0
2	16-18	575.67	0	0	õ	õ	0	0	0	0	0	0	0	0	0	0	108	0
CC		583.60	0	0	0	0	0	0	0	0	0	0	0	0	0	0	77 94	2
5	2-4	585.13	0	0	0	0	2	0	0	0	0	0	0	1	0	0	94 453	2
	54-56 14-16	590.15 594.85	0	0	0	0	0	0	0	0	0	0	0	0	0	0	433	1
cc	14-10	602.90	0	0	0	0	0	0	o	0	õ	õ	ő	0	0	ŏ	79	i
cc		612.5	õ	õ	0	o	ĩ	1	0	0	0	0	0	0	0	0	433	1
2	3-5	614.04	0	0	0	0	1	0	0	0	0	0	0	0	0	0	61	1
	103-105	618.04	0	0	0	0	2	0	0	0	0	1	0	0	0	0	97	1
CC	34-36	624.05	0	0	0	0	0	0	0	0	0	0	0	0	0	0	143 296	1
CC	111-113	631.8 634.42	0	0	0	0	0	0	0	0	0	0	0	0	0	0	366	0
5	106-108	638.87	ő	0	2	0	ő	ő	õ	õ	o	õ	õ	õ	0	0	120	1
	11-13	642.92	õ	o	ĩ	o	0	õ	0	0	0	0	0	0	0	0	156	1
	47-49	644.78	0	0	0	0	0	0	0	0	0	0	0	0	0	0	266	1
	33-35	651.24	0	0	0	0	2	0	0	0	1	2	0	0	0	0	143	1
	9-11	671.80	0	0	0	0	1	1	0	0	0	0	0	0	0	0	38 79	2 2
	60-62 127-129	680.31 682.48	0	0	1	0	0	0	0	0	0	0	0	0	0	0	33	2
	58-60	691.39	0	0	4	0	0	0	0	0	0	2	õ	2	1	1	277	2
cc	50 50	699.00	õ	0	2	0	o	0	õ	0	õ	õ	õ	õ	î	ò	175	2
1	0-2	700.51	0	õ	õ	õ	0	õ	0	0	0	0	0	0	0	0	99	2
£	10-12	703.61	0	0	2	1	0	0	0	0	0	0	0	0	0	0	95	2
2	88-90	710.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0	18	1
	87-89	713.98	0	0	1	0	0	0	0	0	0	0	0	0	0	0	24	1
2	17-19	719.98	0	0	0	0	0	0	0	0	0	0	0	0	0	0	15	02
	37-39	732.78	0	0	0	0	0	0	0	0	0	2	0	2	0	0	46	2
CC	25-27	734.16 737.70	0	0	2	0	1	0	0	0	0	0	0	0	0	1	40	2
2	8-10	739.29	0	0	0	0	0	0	0	0	0	o	õ	õ	0	ò	33	1
	40-42	739.61	0	õ	0	ŏ	0	0	õ	õ	0	õ	o	õ	0	0	34	2
£		750.31	ŏ	õ	1	õ	0	0	0	0	0	0	0	0	0	0	57	2
2 2 CC	140-142	766.1	õ	o		0	0	0	0	0	0	0	0	0	0	0	51	2.

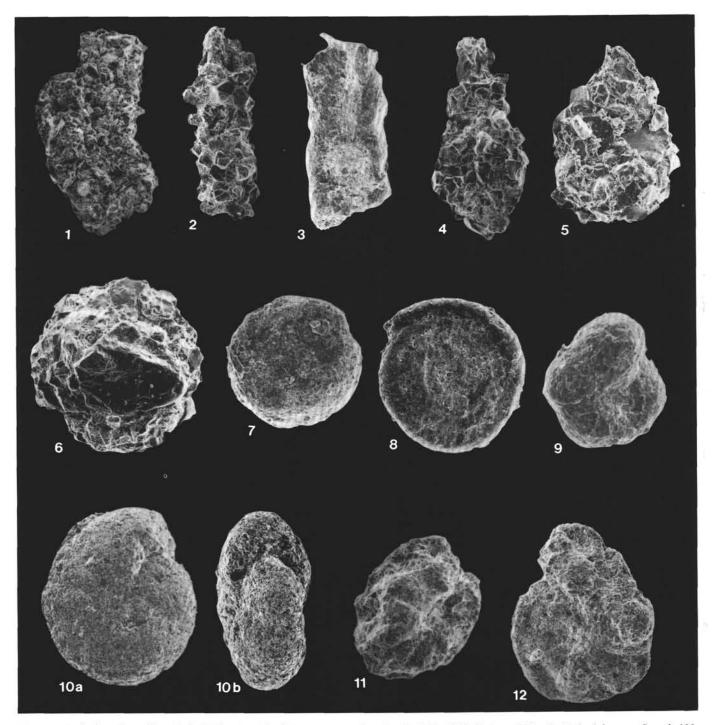


Plate 1. Agglutinated taxa from Hole 645E. 1 and 2. *Rhizammina* sp., Sample 105-645E-43X-1, 3-6 cm, 33X. 3. *Bathysiphon* sp., Sample 105-645E-53X-5, 98-100 cm, 33X. 4 and 5. *Lagenammina* sp., Sample 05-645E-43X-1, 3-6 cm, 55X. 6. *Psammosphaera fusca* Schultze, Sample 105-645E-54X-1, 90-92 cm, 66X. 7. *Psammosphaera* sp., Sample 105-645E-50X-2, 84-86 cm, 110X. 8. *Ammodiscus* sp., Sample 105-645E-76X-6, 38-40 cm, 125X. 9. *Trochammina* sp 5, Sample 105-645E-77X-1, 107-109 cm, 150X. 10a,b. *Cyclammina cancellata* Brady, Sample 105-645E-43X-1, 3-6 cm, 33X. 11 and 12. *Haplophragmoides* sp., Sample 105-645A-76R-6, 38-40 cm, 110X.

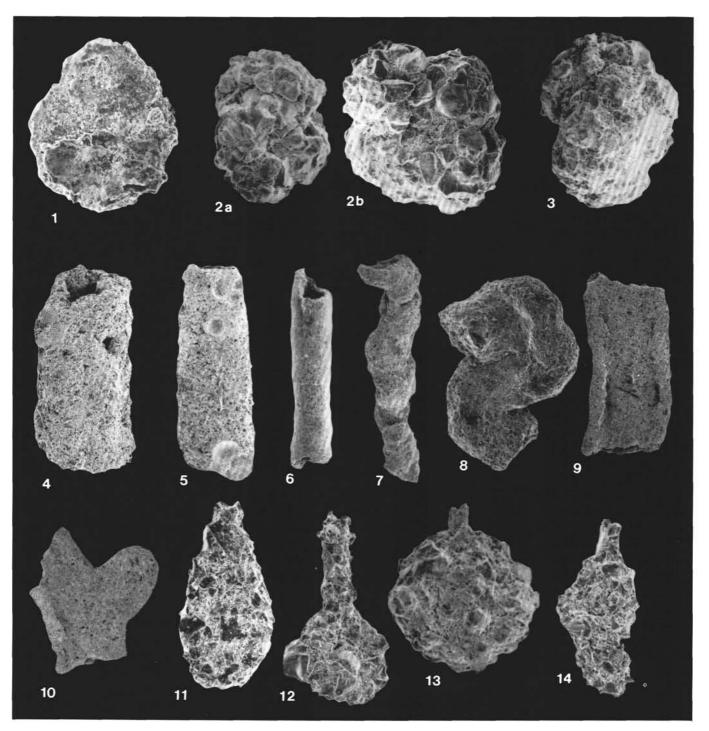


Plate 2. Agglutinated taxa from Sites 645 and 646. 1. *Trochammina* sp. (flat), Sample 105-645E-77R-1, 107-109 cm, 150X. 2. *Haplophragmoides* sp. (coarse), Sample 105-645E-56R-4, 137-140 cm, 66X. 3. *Recurvoides* sp. Sample 105-645E-46R-3, 95-97 cm, 83X. 4 and 5. *Rhizammina* sp., Sample 105-646B-62X-5, 54-56 cm. 6. *Bathysiphon* sp., Sample 105-646B-78X-2, 8-10 cm. 7. *Rhizammina indivisa* Brady, Sample 105-646B-68X-3, 47-49 cm, 66X. 8. *Rhizammina indivisa* Brady, Sample 105-646B-68X-3, 47-49 cm, 55X. 9 and 10. *Dendrophrya* sp., Sample 105-646B-58X-1, 59-61 cm, 28X. 11. *Lagenammina* sp., Section 105-645E-47X, CC, 83X. 12 and 13. *Saccammina* sp., Section 105-645E-47X, CC, 83X. 14. *Reophax subfusiformis* Earland emend. Hoglund, Section 105-645E-47X, CC, 96X.

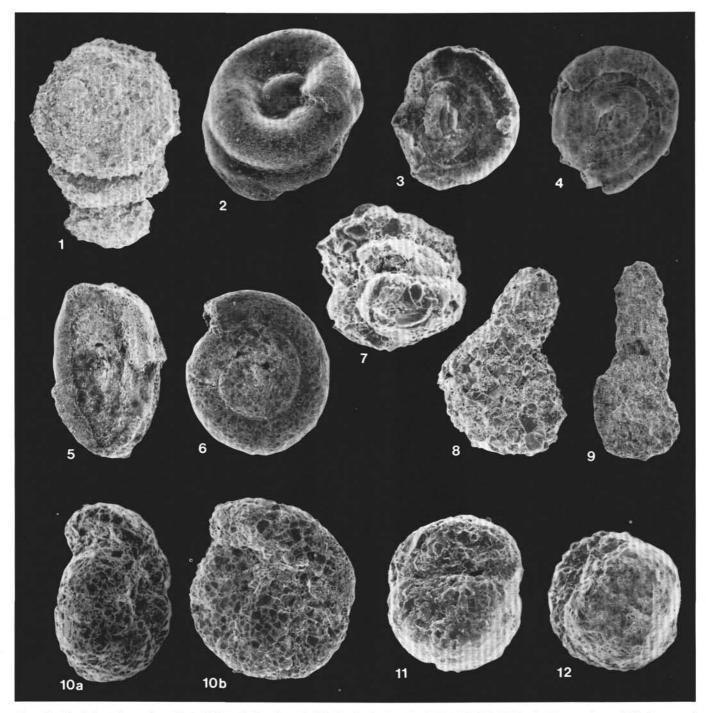


Plate 3. Agglutinated taxa from Hole 646B. 1. Reophax pilulifer Brady, Section 105-646B-47X, CC, 38X. 2. Glomospira gordialis (Jones and Parker), Sample 105-646B-78X-2, 40-42 cm 83X. 3 and 4. Glomospirella sp., Sample 105-646B-77X-5, 25-27 cm, 150X. 5. Ammodiscus sp., Sample 105-646B-78X-2, 40-42 cm, 110X. 6. Ammodiscus incertus (d'Orbigny), Sample 105-646B-77X-4, 37-39 cm, 60X. 7. Adercotrema glomerata (Brady), Section 105-646B-66X, CC, 137X. 8. Ammobaculites sp., Sample 105-646B-69X-1, 33-35 cm, 60X. 9. Ammomarginulina sp., Section 105-646B-47X, CC, 50X. 10a,b. Recurvoides turbinatus (Brady), Section 105-646B-66X, CC, 55X. 11 and 12. Recurvoides contortus Earland, Section 105-646B-51X, CC, 110X.

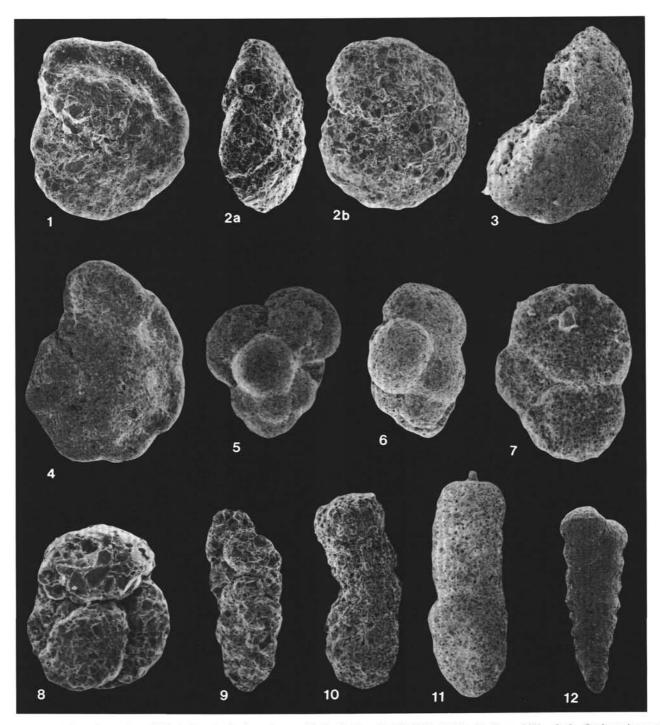


Plate 4. Agglutinated taxa from Hole 646B. 1. Cyclammina pusilla Brady, Sample 105-646B-68X-3, 47-49 cm, 83X. 2a,b. Cyclammina pusilla Brady, Sample 105-646B-69X-1, 33-35 cm, 83X. 3. Cyclammina cancellata Brady, Sample 105-646B-78X-2, 40-42 cm, 28X. 4. Haplophragmoides sp., Section 105-646B-47X, CC, 47X. 5. Trochamminoides sp., Sample 105-646B-68X-3, 47-49 cm, 83X. 6. Trochamminoides sp., Sample 105-646B-67X-2, 111-113 cm, 83X. 7. Ammosphaeroidina sp., Section 105-646B-67X, CC, 50X. 8. Trochamminoides sp., Sample 105-646B-67X-2, 111-113 cm, 125X. 9. Karreriella conversa (Grzybowski), Sample 105-646B-67X-2, 111-113 cm, 83X. 10. Martinotiella sp., Section 105-646B-49X, CC, 83X. 11. Martinotiella sp., Sample 105-646A-62X-5, 54-56 cm, 33X. 12. Siphotextularia sp., Sample 105-646B-73X-2, 58-60 cm, 83X.