

19. A PRELIMINARY REPORT OF EARLY CRETACEOUS DINO CYST FLORAS FROM SITE 765, ARGO ABYSSAL PLAIN, NORTHWEST AUSTRALIA¹

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ABSTRACT

Results of a preliminary study of Early Cretaceous dinocyst assemblages from Site 765 on the Argo Abyssal Plain, off northwestern Australia, are presented. The palynological sequence is interpreted in terms of Australian zones and is, in descending order, the late Aptian *Diconodinium davidii* Zone (Cores 123-765C-33R to -39R), the middle to early Aptian *Odontochitina operculata* Zone (Cores 123-765C-40R to -49R), the Barremian *Muderongia australis* Zone (Cores 123-765C-50R to -54R), and the Berriasian lower *Batioladinium reticulatum* Zone (Core 123-765C-59R). The dating of the sequence as late Aptian to Berriasian on the basis of dinocysts is supported, in part, by data concerning associated foraminiferal, radiolarian, and calcareous nannofossil suites.

INTRODUCTION

Well-preserved, variably diverse, Early Cretaceous dinoflagellate cyst (dinocyst) assemblages were recovered from Site 765 of Leg 123. This site was located on marine magnetic anomaly M26 on the Argo Abyssal Plain, approximately 400 km north of Port Hedland, at 15°58.541'S, 117°34.495'E (Fig. 1), in a water depth of 5728.5 m. The sedimentary sequence at Site 765 is presented in Figure 2. Although Neogene dinocysts were recorded at higher levels (McMinn, this volume), attempts to recover dinocysts from intervening samples were unsuccessful. The dinocyst associations have been interpreted in terms of the zonation outlined by Helby et al. (1987).

Wiseman and Williams (1974), in the only published report of dinocysts from the Argo Abyssal Plain, recorded Barremian and Aptian assemblages at Site 261, referable to the interval between the lower *Muderongia australis* and lower *Odontochitina operculata* zones. Helby et al. (1987) summarized relevant Australian dinoflagellate studies prior to 1987. Backhouse (1988) published a major study of Late Jurassic to Early Cretaceous palynology of the Perth Basin, Western Australia, while Ingram and Morgan (1988) reviewed the development and status of Mesozoic palynostratigraphy of the Australian Northwest Shelf, and Islam (1988) discussed aspects of a Rhaetian to Barremian sequence in the Beagle and Dampier sub-basins, off Western Australia.

MATERIALS AND METHODS

Twenty-six samples were selected for study on the basis of relative palynomorph productivity and position, although some specimens from unreported samples are illustrated. Location details are given in Figure 2. The samples were processed by conventional techniques on board the Resolution. Preliminary interpretation of the samples was undertaken during the cruise by McMinn. Quantitative data are based on rapid counts of 200 specimens (or lesser available number; by McMinn). Helby recorded assemblage data. The strew mounts are deposited in the

collections of the Geological Survey of New South Wales. Single-grain mounts have been retained (by Helby) pending further study.

BIOSTRATIGRAPHY

Diconodinium davidii Zone

Samples from Cores 123-765C-33R-1 to -39R-CC yielded extremely low to low numbers of palynomorphs. In Core 123-765C-34R, spores and pollen dominated the palynomorph suites, while microplankton dominated Cores 123-765C-33R, -765C-35R, and -765C-39R. Dinocyst diversity was variable and ranged from 9 to 48 species (Table 1). Samples were characterized by the common to abundant occurrence of *Diconodinium davidii*, with relatively common *Dingodinium cerviculum* and *Rhombodella paucispina*. Significant accessory species included *Ascodinium* sp. A, which occurs throughout and commonly constitutes a distinctive lower acme subzone of the *D. davidii* Zone in sequences elsewhere on the Australian Northwest Shelf. *Canninginopsis intermedia* has not been recorded previously at this level. *Spiniferites* content ranged from 2.5% to 4.5%. An extremely meager dinocyst association in Core-123-765C-39R-CC contained several specimens of *Diconodinium davidii* and more numerous *Ascodinium* sp. A. In view of the extremely low palynomorph recovery, the assignment to the *D. davidii* Zone was tentative. Jurassic reworking was evident in samples from Cores 123-765C-37R and -39R.

Odontochitina operculata Zone

Samples from Cores 123-765C-40R-4 to -49R-CC yielded low-to-moderate numbers of palynomorphs, dominated by microplankton (up to 99%). Diversity of the dinocyst suites was variable (18-73 species). Samples were characterized by the relatively consistent occurrence of the eponymous species and the apparent absence of *Ascodinium* sp. A, *Diconodinium davidii*, and *Pseudoceratium turneri*. *Muderongia macwhaei* formed a distinctive subzone in the lower part of the interval, similar to those from the Carnarvon Basin. A *Batiacasphaera subtilis* plexus is intermittently dominant, while a *Leptodinium* acme occurred in the middle-to-lower part of the zone. *Epitricysta vinckensis* and *Herendeenia postprojecta* occurred in the lower part of the zone. Late Jurassic reworking was prominent in Cores 123-765C-45R and -49R.

The *Ovoidinium cinctum* Zone (Helby et al., 1987) was not sampled.

¹ Gradstein, F. M., Ludden, J. N., et al., 1992. *Proc. ODP, Sci. Results*, 123: College Station, TX (Ocean Drilling Program).

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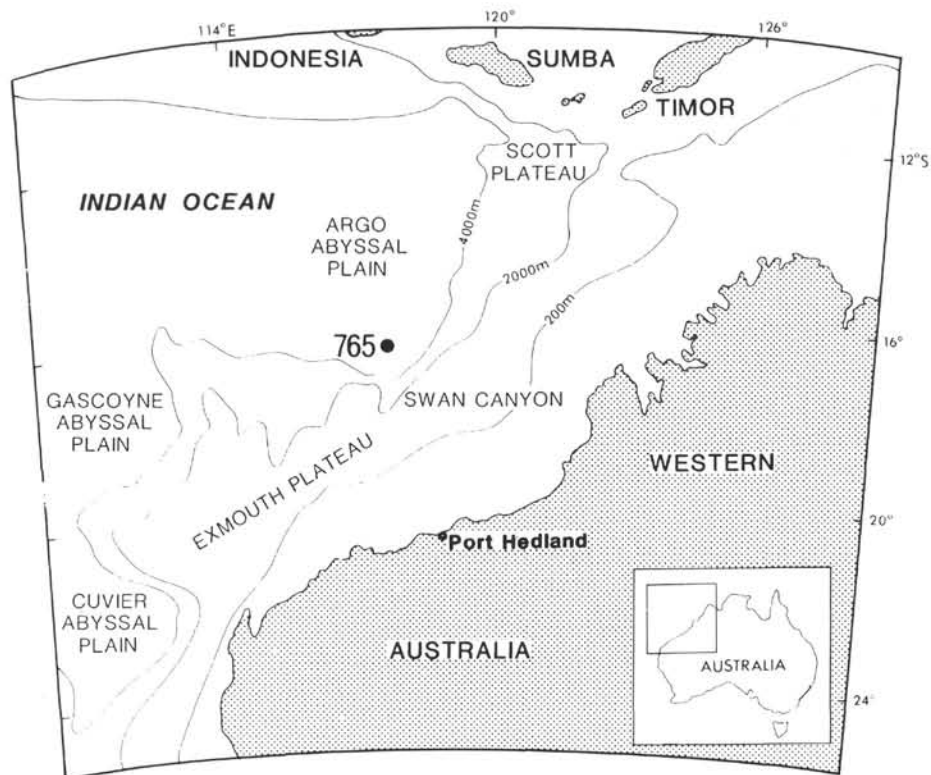


Figure 1. Location of Site 765.

Muderongia australis Zone

Samples from Cores 123-765C-50R-CC to -54R-CC yielded low-to-moderate palynomorph numbers. The palynomorph suites were completely dominated by relatively diverse microplankton associations (Table 1). The dinocyst assemblages were characterized by the occurrence of the eponymous species, with relatively consistent *Belodinium dysculum*, *Circulodinium deflandrei*, *Cribrerodinium aparsium*, *Dingodinium cerviculum*, *Dissiliodinium* sp. A, *Herendeenia postprojecta*, and *Impagidinium phlyctaena*. A *Batiacasphaera imperfecta* plexus acme and a *Cribrerodinium*(?) sp. cf. *C. asarotum* acme occurred in the upper part of the zone, while *Dissiliodinium* sp. A was abundant in the middle part of the zone. The occurrence of *Phoberocysta neocomica* in Core 123-765C-54R indicated that the associations are at least as old as the lower part of the zone. However, the apparent absence of twin-horned muderongoids, *Gardodinium lowii* and *Senoniasphaera tabulata*, suggested that these lower samples were not as old as the Hauterivian *Muderongia testudinaria* Zone. Jurassic reworking was noted in residues from Core 123-765C-54R. Core 123-765C-58R-2 yielded an extremely meager, almost monospecific, palynomorph residue of *Kaiwaradinium scrutillinum*, suggesting that it is no older than mid-Valanginian.

Lower *Batioladinium reticulatum* Zone

Samples from Cores 123-765C-59R-4 and -59R-5 yielded meager palynomorph residues. High-diversity microplankton suites dominated the residues (Table 1). The dinocyst associations were assigned to the lower zonule of the *B. reticulatum* Zone on the basis of the occurrence of the eponymous species with *Dissimulidinium lobispinosum*. Significant accessory species included *Aldorfia*(?) sp. A, *Egmontodinium torynum*, and *Kleithrisphaeridium fasciatum*. The occurrence of *Wanaea* species is evidence of reworking of Kimmeridgian to Callovian sediments,

while the occurrence of *Dingodinium cerviculum* and *Gardodinium attenuatum* indicated downhole contamination of the samples.

DISCUSSION

Summary details are presented in Table 1. Dating of the dinoflagellate zones (adapted from Helby et al., 1987) is outlined together with data from calcareous nannofossil, radiolarian, and foraminiferal studies. The distribution of selected taxa in the samples is presented in Table 2. The distal position of Site 765 in relation to the Early Cretaceous basins of northwestern Australia provides an opportunity for testing quantitative variations in the zones.

The late Aptian *Diconodinium davidii* Zone occurred in lithologic Unit IV (von Rad et al., 1989, Fig. 9), which was predominantly claystone with turbiditic carbonates, nannofossil chalk, and zeolitic claystone; the samples were taken from medium to dark gray claystone beds. The palynomorph suites were characterized by variable ratios of microplankton to spores and pollen (1:5 to 4:1). Reworking was evident throughout this interval. In the Timor Sea region, the major part of the Aptian is supercondensed, sedimentation recommencing with deposition of radiolarian-rich sediments characterized by the *D. davidii* Assemblage (lower Jamieson Formation of Pattillo and Nicholls, 1990). In the Carnarvon Basin, the zone is associated with the *Wandalia* Radiolarite (Hocking et al., 1987, p. 151).

The middle to early Aptian *Odontochitina operculata* Zone occupies the upper part of lithologic Unit V, which was predominantly claystone with rhodochrosite and radiolarite; samples were collected from medium to dark gray and black claystone horizons. All samples were dominated by microplankton (Table 1). The associations were characterized by major abundances of taxa not encountered in these numbers in more proximal depositional locations (*Batiacasphaera* and *Leptodinium*). An apparent inverse relationship exists between the numbers of *Spiniferites* and the microplankton to spore/pollen ratio, which may reflect an

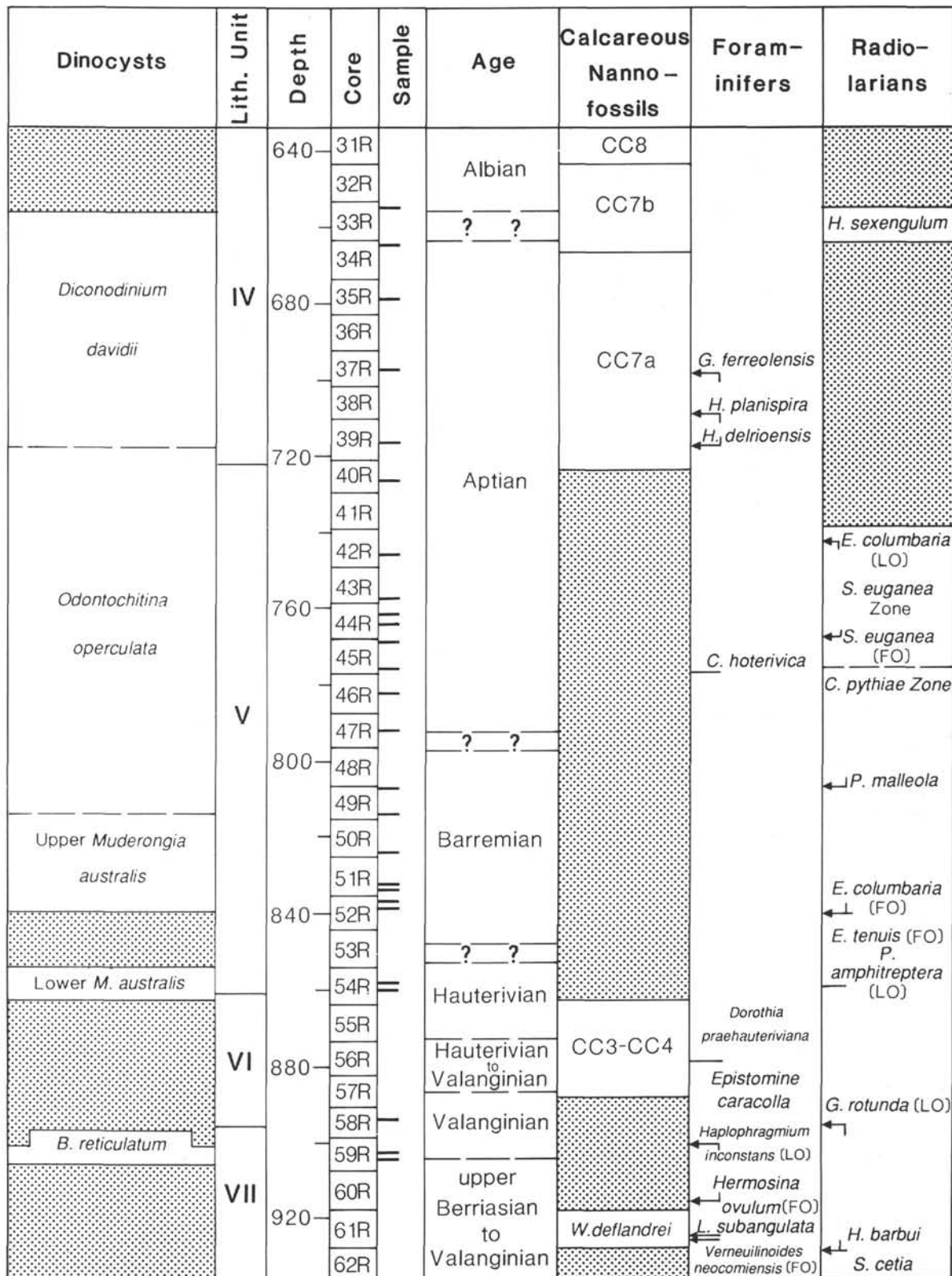


Figure 2. Early Cretaceous biostratigraphy at Site 765.

Table 1. Palynological summary.

Core, section, interval (cm)	Palynomorph yield	Counts			Zone
		(1)	(2)	(3)	
123-765C-33R-1, 38-40	Very low	3:1	38	11%	<i>Diconodinium davidii</i>
34R-1, 130-132	Low	1:5	37	2.5%	<i>Diconodinium davidii</i>
^a 35R-CC	Moderate	4:1	48	4.5%	<i>Diconodinium davidii</i>
37R-CC	Ext. low	1:1	10	2%	Indeterminate
39R-CC1	Ext. low	2:1	9	—	<i>Diconodinium davidii</i>
^a 40R-4, 138-140	Moderate	19:1	37	1.5%	<i>Odontochitina operculata</i>
42R-2, 117-119	Low	3:1	21	19%	<i>Odontochitina operculata</i>
43R-1, 111-112	Low	19:1	25	5%	<i>Odontochitina operculata</i>
43R-CC	Low	4:1	35	7%	<i>Odontochitina operculata</i>
44R-1, 100-102	Moderate	49:1	42	1%	<i>Odontochitina operculata</i>
44R-CC	Low	9:1	20	1%	<i>Odontochitina operculata</i>
^a 45R-1, 12-14	Moderate	19:1	31	3%	<i>Odontochitina operculata</i>
45R-CC	Very low	16:1	31	1%	<i>Odontochitina operculata</i>
47R-3, 39-40	Low	99:1	18	0.5%	<i>Odontochitina operculata</i>
49R-1, 125-126	Low	4:1	73	10.5%	<i>Odontochitina operculata</i>
49R-CC	Very low	9:1	32	2.5%	<i>Odontochitina operculata</i>
^a 50R-CC	Low	99:1	28	0.5%	<i>Muderongia australis</i>
^a 51R-5, 106-107	Low	19:1	39	2%	<i>Muderongia australis</i>
51R-CC	Moderate	99:1	23	0.5%	<i>Muderongia australis</i>
52R-1, 150-151	Moderate	6:1	37	1%	<i>Muderongia australis</i>
52R-2, 36-40	Low	9:1	43	7.5%	<i>Muderongia australis</i>
^a 54R-4, 49-50	Low	99:1	35	Tr	Lower <i>M. australis</i>
54R-CC	Low	49:1	18	—	Lower <i>M. australis</i>
58R-2, 14-19	Ext. low	NR	3	—	Indeterminate
59R-4, 105-112	Low	9:1	76	Tr	Lower <i>Batioladinium reticulatum</i>
123-765C-59R-5, 8-9	Low	4:1	47	Tr	<i>Batioladinium reticulatum</i> or older

^a Single species mounts prepared. Counts: (1) microplankton to spores/pollen ratio; (2) microplankton diversity (s.l.); (3) *Spiniferites* as proportion of microplankton. NR = not recorded; Tr = trace.

increase in the number of taxa introduced from more proximal locations by turbidity flow. The *O. operculata* Zone is commonly obscured by a supercondensed section in the Timor Sea region. The zone occurs in the Muderong Shale in the Carnarvon Basin, where it exhibits much higher spore/pollen to microplankton ratios and is associated with substantial vascular plant debris, reflecting more proximal depositional locations.

The Barremian *Muderongia australis* Zone occupies the lower part of lithologic Unit V (Fig. 2, see also von Rad et al., 1989, Fig. 9). The lithology is predominantly claystone with minor radiolarite; samples were collected from green-gray, medium to dark gray and black claystone intervals. The palynomorph suites were totally dominated by moderately diverse microplankton associations (Table 1). Acmes of *Batiacasphaera* and *Cribroperidinium* sp. cf. *C. asarotum* occurred in the upper part of the zone. The *Muderongia australis* Zone occurs in the upper part of the lower member of the Darwin Formation (Hughes, 1978) and in the Echuca Shoals Formation (Pattillo and Nicholls, 1990) in the Timor Sea region, as well as the lower part of the Muderong Shale in the Carnarvon Basin.

The Berriasian lower part of the *Batioladinium reticulatum* Zone was encountered in Cores 123-765C-59R-4 and -59R-5, which are located in the upper part of lithologic Unit VII. The lithology is predominantly brown-red silty claystone, and samples were taken from thin intercalations of dark gray claystone. The palynomorph residues were dominated by diverse microplankton suites, with evidence of Jurassic reworking. Rare specimens of taxa, not known to extend below the middle to late Valanginian *Senoniasphaera tabulata* Zone (not recorded in Hole 765C), were encountered in the samples. These are considered to represent downhole contamination. The alternative proposition, that the relatively abundant representation from the *B. reticulatum* Zone is reworked, is considered unlikely. The *B. reticulatum* Zone occurs only rarely in the Timor Sea region, where it was not deposited, has been removed, or is obscured by a supercondensed

sequence. This zone has been identified recently in a well in the Arafura Sea. Over much of the Carnarvon Basin, the zone occurs in the upper Barrow Group and exceeds 650 m in some wells.

CONCLUSIONS

The Early Cretaceous palynological sequence in Hole 765C resembles sequences in the coastal margin basins of the Australian Northwest Shelf. The sequence is, in descending order, the late Aptian *Diconodinium davidii* Zone (Cores 123-765C-33R to -39R), the middle to early Aptian *Odontochitina operculata* Zone (Cores 123-765C-40R to -49R), the Barremian *Muderongia australis* Zone (Cores 123-765C-50R to -54R), and the Berriasian lower *Batioladinium reticulatum* Zone (Core 123-765C-59R). Only a few dinocyst species have not been recorded previously on the Northwest Shelf. The quantitative distribution of taxa in Hole 765C, particularly below the late Aptian, was not encountered in the marginal basins and possibly reflects the extreme distal locus of deposition. The ages assigned to the dinocyst zones by Helby et al. (1987) are essentially supported by dating from other fossil groups (Fig. 2).

ACKNOWLEDGMENTS

A. McMinn publishes with permission of the Director of the Geological Survey of New South Wales.

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APPENDIX Species List

- Achomosphaera? neptuni* (Eisenack, 1954) Davey and Williams, 1966
- Adnatosphaeridium tutulosum* (Cookson and Eisenack, 1960) Morgan, 1980
- Aldorfia deflandrei* (Clark and Verdier, 1967) Stover and Evitt, 1978
- Angustidinium acribes* (Davey and Verdier, 1971) Goodman and Evitt, 1981
- Aprobolocysta galeata* Backhouse, 1987
- Apteodinium granulatum* Eisenack, 1958
- Apteodinium maculatum* Eisenack and Cookson, 1960
- Batiacasphaera imperfecta* Stover and Helby, 1987
- Batiacasphaera scrobiculata* (Deflandre and Cookson, 1955) Burger, 1980
- Batiacasphaera subtilis* Stover and Helby, 1987
- Batioladinium jaegeri* (Alberti, 1961) Brideaux, 1975
- Batioladinium longicornutum* (Alberti, 1961) Brideaux, 1975
- Batioladinium micropodium* (Eisenack and Cookson, 1960) Brideaux, 1975
- Batioladinium reticulatum* Stover and Helby, 1987
- Belodinium dysculum* Cookson and Eisenack, 1960
- Broomea simplex* Cookson and Eisenack, 1958
- Callaiosphaeridium asymmetricum* Davey and Williams, 1966
- Canningia reticulata* Cookson and Eisenack, 1960 emend. Helby, 1987
- Canninginopsis intermedia* Morgan, 1980
- Canninginopsis tabulata* (Davey and Verdier, 1974) Duxbury, 1977
- Carpodinium granulatum* Cookson and Eisenack, 1962
- Cassiculosphaeridia magna* Davey, 1974
- Cassiculosphaeridia reticulata* Davey, 1969
- Cernicysta helbyi* (Morgan, 1980) Stover and Helby, 1987
- Chichaouadinium boydii* (Morgan, 1980) Bujak and Davies, 1983
- Chlamydothorella nyei* Cookson and Eisenack, 1958
- Chytroisphaeridia chytrooides* (Sarjeant, 1962) Downie and Sarjeant, 1965 emend. Davey, 1979
- Circulodinium colliveri* (Cookson and Eisenack, 1960) Helby, 1987
- Circulodinium deflandrei* Alberti, 1961
- Circulodinium hirtellum* Alberti, 1961
- Cleistosphaeridium ancoriferum* (Cookson and Eisenack, 1960) Davey et al., 1966 emend. Cookson and Eisenack, 1968
- Codoniella campanulata* (Cookson and Eisenack, 1960) Downie and Sarjeant, 1965
- Cometodinium? whitei* (Deflandre and Courteville, 1939) Stover and Evitt, 1978
- Coronifera oceanica* Cookson and Eisenack, 1958
- Cribroperidinium aparsium* Stover and Helby, 1987
- Cribroperidinium asarotum* Stevens, 1987
- Cribroperidinium edwardsii* (Cookson and Eisenack, 1958) Davey, 1969
- Cribroperidinium leedervilleense* Backhouse, 1988
- Cribroperidinium muderongense* (Cookson and Eisenack, 1958) Davey, 1969
- Dapsilidinium warrenii* (Habib, 1976) Lentin and Williams, 1981
- Diconodinium davidii* Morgan, 1975
- Diconodinium pusillum* Singh, 1971
- Dingodinium cerviculum* Cookson and Eisenack, 1958
- Dingodinium jurassicum* Cookson and Eisenack, 1958
- Dissimulidinium lobispinosum* May et al., 1987
- Dollidinium sinuosum* (Cookson and Eisenack, 1960) Helby and Stover, 1987
- Egmontodinium torynum* (Cookson and Eisenack, 1960) Davey, 1979
- Elytrocysta circulata* (Clarke and Verdier, 1967) Stover and Helby, 1987
- Epelidosphaeridia colligata* (Morgan, 1980) Lentin and Williams, 1985
- Epicricysta vinckensis* Stover and Helby, 1987
- Exiguosphaera phragma* Duxbury, 1979
- Exochosphaeridium arnace* Davey and Verdier, 1973
- Exochosphaeridium phragmites* Davey et al., 1966
- Exochosphaeridium robustum* Backhouse, 1988
- Eyrea nebulosa* Cookson and Eisenack, 1971
- Fibradinium variculum* Stover and Helby, 1987
- Florentinia mantellii* (Davey and Williams, 1966) Davey and Verdier, 1973
- Fromea amphora* Cookson and Eisenack, 1958
- Fromea monilifera* Backhouse, 1987
- Gardodinium attenuatum* Stover and Helby, 1987
- Gardodinium lowii* Backhouse, 1987
- Gonyaulacysta helicoidea* (Eisenack and Cookson, 1960) Sarjeant, 1960
- Gonyaulacysta jurassica* (Deflandre, 1938) Norris and Sarjeant, 1965
- Hapsocysta peridictya* (Eisenack and Cookson, 1960) Davey, 1979
- Herendeenia postprojecta* Stover and Helby, 1987
- Heslertonia striata* (Cookson and Eisenack, 1960) Norvick, 1976
- Hystrichosphaeridium pachydermum* Cookson and Eisenack, 1960
- Hystrichosphaeridium petilum* Gitmez, 1970
- Impagidinium phlyctaena* Stover and Helby, 1987
- Kaiwaradinium scrutillinum* Backhouse, 1987
- Kallosphaeridium norvickii* Burger, 1980
- Kiokansium polypes* (Cookson and Eisenack, 1962) Below, 1982
- Kleithriasphaeridium fasciatum* (Davey and Williams, 1966) Davey, 1974
- Kleithriasphaeridium simplicispinum* (Davey and Williams, 1966) Davey, 1974
- Leptodinium asymmetricum* Morgan, 1980
- Leptodinium eumorphum* (Cookson and Eisenack, 1960) Sarjeant in Davey et al., 1969
- Leptodinium pinnosum* Davey, 1988
- Leptodinium simplex* Burger, 1980
- Leptodinium tunellum* Stover and Helby, 1987
- Meiourogonyaulax maculata* Backhouse, 1988
- Meiourogonyaulax stoveri* Millioud, 1969
- Meiourogonyaulax sp. B* - Helby et al., 1987
- Muderongia australis* Helby, 1987
- Muderongia macwhaei* Cookson and Eisenack, 1958
- Muderongia tetracantha* (Gocht, 1957) Alberti, 1961
- Occiscysta tenuiceras* (Eisenack, 1958) Below, 1981
- Odontochitina operculata* (Wetzel, 1933) Deflandre and Cookson, 1955
- Oligosphaeridium anthophorum* (Cookson and Eisenack, 1958) Davey, 1969
- Oligosphaeridium asterigerum* (Gocht, 1959) Davey and Williams, 1966
- Oligosphaeridium complex* (White, 1842) Davey and Williams, 1966
- Oligosphaeridium poculum* Jain, 1977
- Oligosphaeridium pulcherrimum* (Deflandre and Cookson, 1955) Davey and Williams, 1966
- Palaeoperidinium cretaceum* Pocock, 1962
- Phoberocysta neocomica* (Gocht, 1957) Millioud, 1969 emend. Helby, 1987
- Platycystidia eisenackii* (Mehrotra and Sarjeant, 1984) Backhouse, 1988
- Prolixosphaeridium inequioratum* Stover and Helby, 1987
- Prolixosphaeridium parvispinum* (Deflandre, 1937) Davey et al., 1969
- Protoellipsodinium densispinum* Morgan, 1980
- Pseudoceratium turneri* Cookson and Eisenack, 1958
- Pterospermella aureolata* (Cookson and Eisenack, 1958) Eisenack, 1972
- Pterospermella australiensis* (Deflandre and Cookson, 1955) Eisenack and Cramer, 1973
- Pxydiella tumida* Stover and Helby, 1987

Pyxidopsis challengerensis Habib, 1976
Rhombodella paucispina (Alberti, 1961) Duxbury, 1980
Rigaudella aemula (Deflandre, 1938) Below, 1982
Schizocystia laevigata Cookson and Eisenack, 1962
Scriniodinium attadalense (Cookson and Eisenack, 1958) Eisenack, 1967
Scriniodinium? campanula Gocht, 1959
Senoniasphaera ptomatis Helby, May and Partridge in Helby, 1987
Senoniasphaera tabulata Backhouse and Helby in Helby, 1987
Sirmiodinium grossii Alberti, 1961
Systematophora areolata Klement, 1960

Systematophora palmula Davey, 1982
Tanyosphaeridium boletus Davey, 1974
Trichodinium castanea (Deflandre, 1935) Clarke and Verdier, 1967
Tubotuberella vlamingii Backhouse, 1987
Walloodinium krutzschii (Alberti, 1961) Habib, 1972
Walloodinium luna (Cookson and Eisenack, 1960) Lentin and Williams, 1973
Yalkalpodinium scutum Morgan, 1980

Full details of the citations can be found in Lentin and Williams (1989).

Table 2. Distribution of selected taxa at Hole 765C/D.

Core no.:	3 3 3 3 3					4 4 4 4 4					4 4 4 4 4					4 4 5 5 5					5 5 5 5 5					5 5						
	3	4	5	7	9	0	2	2	3	3	4	4	5	5	7	4	4	5	5	7	4	4	5	5	5	2	2	4	4	8	5	9
Section:	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R
Species	1	1	C	C	C	4	2	C	1	C	1	C	1	C	3	1	C	C	5	C	1	C	C	5	C	1	2	4	C	2	4	5
<i>Achomosphaera(?) neptuni</i>								X			X					X												X	X			
<i>Adnatosphaeridium tutulosum</i>			X																													
<i>Aldorfia deflandrei</i>						X										X												X	X			
<i>Aldorfia(?) sp. A</i>																X			X									F	X			
<i>Angustidinium acribes</i>		X					X																									
<i>Aprobolocysta(?) galeata</i>																X	X		X			X	X					X				
<i>Apteodinium granulatum</i>	X	X	R			F		R	X	X		X			F		X		X				X	X				A	A			
<i>Apteodinium maculatum</i>			X												R	X	X							X	X							
<i>Ascodinium sp. A</i>	X	R	X		F																											
<i>Ascodinium(?) sp. A</i>			X																													
<i>Avellodinium spp.</i>																																
<i>Batiacasphaera imperfecta plexus</i>						R	R	R	X	X	F	F	R	F	X	X	A	A	R	F	F	R	X	C				F	R			
<i>Batiacasphaera scrobiculata</i>								R																								
<i>Batiacasphaera subtilis plexus</i>						A	C	A	F	X	C	C	R	R	A	X	C	F		X	R	R	C	C				F	R			
<i>Batiacasphaera sp. A</i>																												X				
<i>Batiacasphaera spp.</i>		X		F	R	X	X	X	X	X			X	X	X	X	X	X	X	X	X	X	X	X				X	X			
<i>Batioladinium jaegeri</i>						X	X	X	X	X	X																					
<i>Batioladinium longicornutum</i>						X					X																					
<i>Batioladinium micropodum</i>			X			X					X																	X				
<i>Batioladinium reticulatum</i>																X	X											X				
<i>Belodinium dysculum</i>																X		X	X				R	X				X				
<i>Broomea simplex</i>																												X				
<i>Callaoisphaeridium asymmetricum</i>	X		R																													
<i>Canningia reticulata</i>																								X				X				
<i>Canninginopsis intermedia</i>	C		F																													
<i>Canninginopsis tabulata</i>																																
<i>Carpodinium granulatum</i>	X		X								R	X	X	R					X	X	X	F	R	C	F			F				
<i>Cassiculosphaeridia magna</i>						R	F	R	F	X	X	X		F	R	X	X	R		X	X	C	X	R								
<i>Cassiculosphaeridia reticulata</i>			R		R	R	X	X	X		X					X																
<i>Cassiculosphaeridia sp. A</i>						R	X	X	X		X				X																	
<i>Cernicysta helbyi</i>			X										X																			
<i>Chichaouadinium boydii</i>	X	R	R																													
<i>Chlamydothorella nyei</i>	X	F	R		X	C			R	X	R	X			X				X	X		R	X			R	X					
<i>Chytroisphaeridia chytroides</i>																												X	X			
<i>Circulodinium colliveri</i>	X		C	R		F	R	X	R	X					X								X	R				R	X			
<i>Circulodinium deflandrei</i>	X			C	F				R	X				F							X	F	R	R	R	X	R	A	A			
<i>Circulodinium hirtellum</i>																X																
<i>Cleistosphaeridium ancoriferum</i>				X				X			X																					
<i>Cleistosphaeridium spp.</i>	X	F	X	X	X			X	X		X					X	X	X		X	R							X	X			
<i>Codoniella campanulata</i>	X	X	X																													
<i>Cometodinium(?) whitei</i>	X	R	X			R	X	R		R	X				X				X			X						X	X			
<i>Coronifera oceanica</i>		X	X							X			X																			
<i>Cribrerodinium aparsium</i>								X																								
<i>Cribrerodinium sp. cf. C. asarotum</i>																																
<i>Cribrerodinium edwardsii</i>	X		X																													
<i>Cribrerodinium leedervillense</i>	X		R					R	X				X	X														X				
<i>Cribrerodinium muderongense</i>						X										X	X							X				R				
<i>Cribrerodinium sp. A</i>																								X								
<i>Cribrerodinium sp. B</i>										R	X																					
<i>Cymatiosphaera spp.</i>																																
<i>Dapsilidinium warrenii</i>						X	X	X		X	X					X													X			
<i>Diconodinium davidii</i>	F	A	A		X																											
<i>Diconodinium pusillum</i>	X																															
<i>Dingodinium cerviculum</i>	X	C	R		C	X	F		F	X	R		X		X		X	X	X	X	X	X										
<i>Dingodinium jurassicum</i>																																
<i>Dissiliodinium spp.</i>		X	X	R		F			X	X					X	X	R	R	F	A	F							X	R			
<i>Dissimulidinium lobispinosum</i>																												X				
<i>Dollidinium sinuosum</i>																												X				
<i>Druggidium spp.</i>													X							X	X	?					X					
<i>Egmontodinium torynum</i>																												X	R			
<i>Elytrocysta circulata</i>						X																										
<i>Epelidosphaeridia colligata</i>																												X				
<i>Epitricysta vinckensis</i>												X			X			X	X	X		X					R	X				
<i>Epitricysta sp. A</i>																																
<i>Escharisphaeridia sp. A</i>						X				X	A	F	X																			
<i>Exiguosphaera phragma</i>																X		X														
<i>Exochosphaeridium arnace</i>																											X					
<i>Exochosphaeridium phragmites</i>	X		X			R		R	X	R		X			X	X	X	X	X	X	X											
<i>Exochosphaeridium robustum</i>																																
<i>Exochosphaeridium sp. A</i>						X	X	X	X					X					X										R			
<i>Eyrea nebulosa</i>	R		X					R	X						X	X			X										X			
<i>Fibradinium variculum</i>			X							X		X	X						X													
<i>Florentinia mantellii</i>	X	X	X			X																										

Table 2 (continued).

Core no.:	3	3	3	3	3	4	4	4	4	4	4	4	4	4	4	4	4	5	5	5	5	5	5	5	5	5	5
Section:	3	4	5	7	9	0	2	2	3	3	4	4	5	5	7	9	9	0	1	1	2	2	4	4	8	9	9
Species	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R
Section:	1	1	C	C	C	4	2	C	1	C	1	C	1	C	3	1	C	C	5	C	1	2	4	C	2	4	5
Species	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C
<i>Fromea amphora</i>	X	X	X			X			X							X											
<i>Fromea monilifera</i>																X											
<i>Gardodinium</i> sp. cf. <i>G. attenuatum</i>						X																					
Gen. et sp. indet. A																											
Gen. et sp. indet. B																											
Gen. et sp. indet. C																											
Gen. et sp. nov. A																											
<i>Gonyaulacysta helicoidea</i>		R	R			F	R																				
<i>Gonyaulacysta jurassica</i>																											
<i>Hapsocysta peridictya</i>			X																								
<i>Herendeenia postprojecta</i>																X	R	R	R	X	R	R					
<i>Heslertonia striata</i>	X															X					X	X					
<i>Hystrichodinium</i> spp.																X					X	X					
<i>Hystrichosphaeridium pachydermum</i>																X										X	F
<i>Hystrichosphaeridium petilum</i>																X							R			X	
<i>Hystrichosphaeropsis</i> sp. A			X																								
<i>Impagidinium phlyctaena</i>																			X		X	X	X				
<i>Kaiwaradinium scrutillinum</i>																							F	C	F		
<i>Kallosphaeridium norvickii</i>	X	X	X	X	X	X	R			X						X	X		X		X					X	X
<i>Kallosphaeridium</i> sp. A																											
<i>Kiokansium polypes</i>	X	R	X								X		R	X		X	X				X						
<i>Kleithriasphaeridium fasciatum</i>																X	X									X	X
<i>Kleithriasphaeridium simplicispinum</i>																X	X										
<i>Lanterna(?)</i> sp. A						R				X	X	X	R														
<i>Leptodinium asymmetricum</i>						X					X							X									
<i>Leptodinium eumorphum</i>																											
<i>Leptodinium pinnosum</i>																											
<i>Leptodinium simplex</i>																X	R			R	F	R					
<i>Leptodinium tunellum</i>											X	X				X	X	X	X	X	X	X	R			X	X
<i>Leptodinium</i> spp.	F	R	R		X	X	R	C	C	A	A	A	F	R	R	X	X	X	X	X	X	X	R			X	X
<i>Leptodinium(?)</i> sp. A						X																					
<i>Leptodinium(?)</i> sp. B																											
<i>Meiourogonyaulax maculata</i>																											
<i>Meiourogonyaulax stoveri</i>						X		X	X	X				X		X							X			X	X
<i>Meiourogonyaulax</i> sp. B																											
<i>Meiourogonyaulax(?)</i> sp. A								X	X	A	R		X	X	X					X	X	X					
<i>Micrhystridium</i> spp.		X																									
<i>Muderongia australis</i>														X		X	X	X	X	X	X	R	R	X			
<i>Muderongia macwhaei</i>										X	X		X	X	X	X	X	X	X	X	X	R	R	X			
<i>Muderongia tetracantha</i>			X			F	X	X					X	X		X	R	F	R	X	R	X	X	X			
<i>Nummus</i> spp.		X								X						X											
<i>Occisucysta tenuiceras</i>			X			R		X	X	A	X	R	R	X	R	X		X	X	X	X	X				X	
<i>Odontochitina operculata</i>	R		R			F	R	F	X		F	R	R	X	X	X	X										
<i>Oligosphaeridium anthophorum</i>						X				X						X	X	R	R				X	R	X	F	F
<i>Oligosphaeridium</i> cf. <i>O. asterigerum</i>																X	X	R	R								
<i>Oligosphaeridium complex</i>	X		R	F		R	R	X						R	R												
<i>Oligosphaeridium</i> sp. cf. <i>O. poculum</i>			X																				X				
<i>Oligosphaeridium pulcherrimum</i>			R			X								X													
<i>Palaeoperidinium cretaceum</i>			X																								
<i>Pareodinia</i> spp.																X										X	R
<i>Phoberocysta neocomica</i>																											
<i>Platycystidia eisenackii</i>											X					X											
<i>Prolixosphaeridium inequior natum</i>						X	X	X	X																		
<i>Prolixosphaeridium parvispinum</i>	X	X				X	F	F	X		R		X	X	X	X	R		X	X	X	X	X	X		X	X
<i>Prolixosphaeridium</i> spp.		X		X																							
<i>Protoellipsoidinium densispinum</i>																											
<i>Pseudoceratium turneri</i>	X		X																								
<i>Pterospermella aureolata</i>																X											
<i>Pterospermella australiensis</i>																								X			
<i>Pyxidiella tumida</i>																											
<i>Pyxidopsis challengerensis</i>						X				X																	
Reworking —Jurassic				X										X		X							X	X		X	
—Permian		X	X																								
<i>Rhombodella paucispina</i>		C	R													X											
<i>Rigaudella aemula</i>																										X	X
<i>Schizocystia laevigata</i>		X																									
<i>Scriniodinium attadalense</i>										X					X											R	R
<i>Scriniodinium(?) campanula</i>																X										X	R
<i>Senoniasphaera</i> sp. cf. <i>S. ptomatis</i>																										X	X
<i>Sentusidinium</i> spp.	X	F	R	C	F	R	C		X	R	R	R	R	R	X	X	R		X	R	R					A	F

Table 2 (continued).

Species	Core no.: 3 3 3 3 3					4 4 4 4 4					4 4 4 4 4					4 4 5 5 5					5 5 5 5 5					5 5	
	3	4	5	7	9	0	2	2	3	3	4	4	5	5	7	9	9	0	1	1	2	2	4	4	8	9	9
Section:	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R
Section:	1	1	C	C	C	4	2	C	1	C	1	C	1	C	3	1	C	C	5	C	1	2	4	C	2	4	5
Section:			C	C	C			C		C		C		C			C	C		C					C		
<i>Sirmiodinium grossii</i>																X							X	X			
<i>Spiniferites</i> spp.	F	R	F			R	A	F	X	F	R	R	R	X	X	X	R	X	R	X	R	F	X			X	X
<i>Systematophora areolata</i>				X										X												X	R
<i>Systematophora palmula</i>																										R	F
<i>Tanyosphaeridium boletus</i>						X	X	F	X							X	X					X				X	X
<i>Tanyosphaeridium</i> sp. A																										X	X
<i>Trichodinium castanea</i>			R																							X	X
<i>Tubotuberella vlamingii</i>																							X				X
<i>Veryhachium</i> spp.		F	X			X		X	X		X					X											
<i>Wallogodinium krutzschii</i>		X																									
<i>Wallogodinium luna</i>		X														X											
<i>Yalkalpodinium scutum</i>			X			X	X	X	X		X		X	X		X		X	R		X	X	X	X		X	

A = >30%; C = >10%–<30%; F = >3.5%–<10%; R = >1%–<3.5%; X = <1%

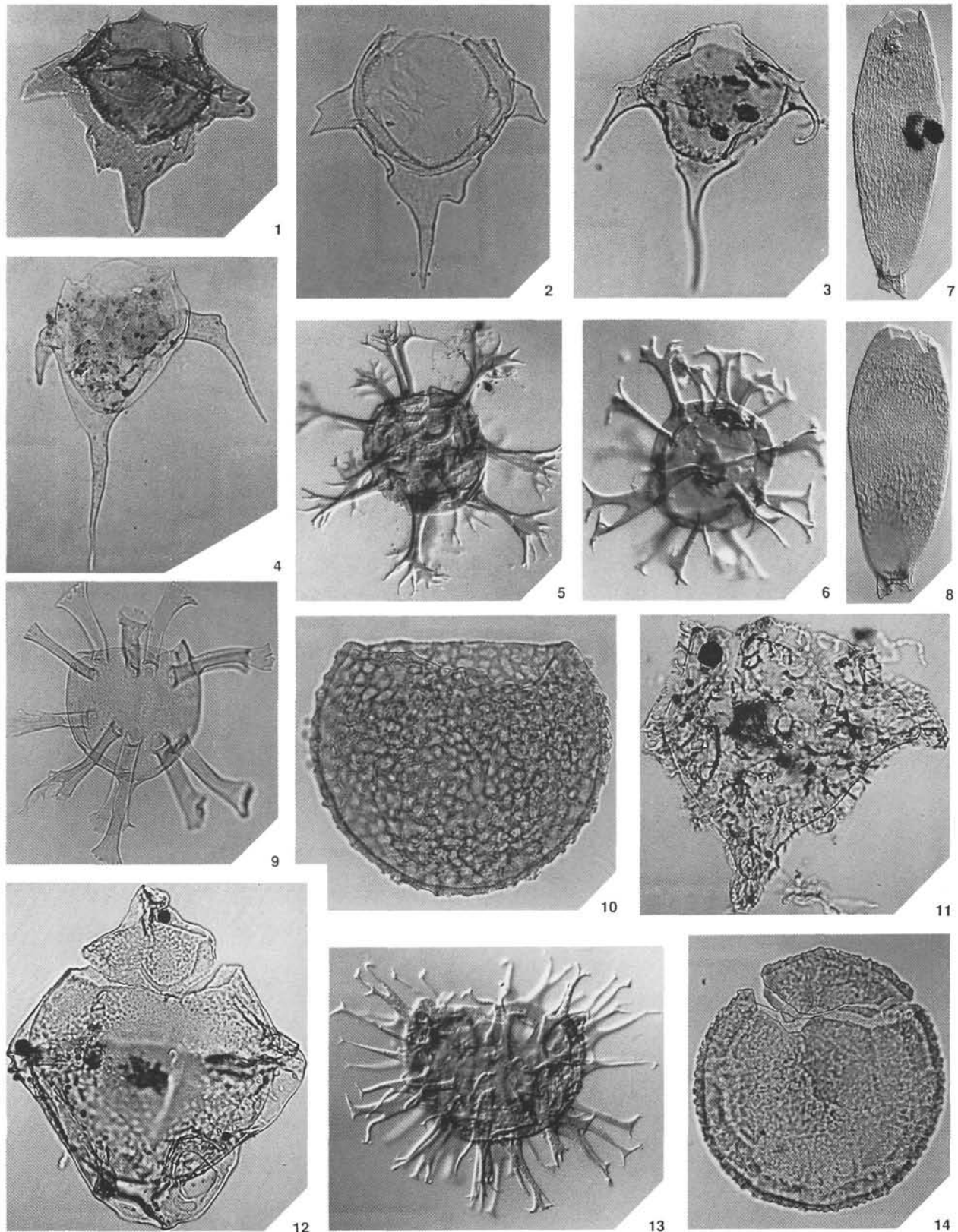


Plate 1. (all x500) **1.** *Phoberocysta neocomica* (Core 123-765C-54R-4 SSM6). **2.** *Muderongia australis* (Core 123-765C-51R-5 SSM12). **3.** *Muderongia* sp. cf. *M. tetracantha* (Core 123-765C-35R-CC SSM35). **4.** *Muderongia tetracantha* (Core 123-765C-50R-CC SSM11). **5.** *Oligosphaeridium asterigerum* (Core 123-765C-59R-4 SSM26). **6.** *Systematophora palmula* (Core 123-765C-59R-4 SSM20). **7.** Gen. et sp. nov. A (Core 123-765C-59R-4 SSM5). **8.** Gen. et sp. nov. A (Core 123-765C-59R-4 SSM6). **9.** *Kaiwaradinium scrutillinum* (Core 123-765C-54R-4 SSM1). **10.** *Cassiculosphaeridia magna* (Core 123-765C-54R-4 SSM8). **11.** *Pseudoceratium turneri* (Core 123-765C-35R-CC SSM8). **12.** *Yalkalpodinium scutum* (Core 123-765C-35R-CC SSM10). **13.** *Systematophora areolata* (Core 123-765C-59R-4 SSM15). **14.** *Meiourogonyaulax?* sp. A (Core 123-765C-40R4 SSM1).

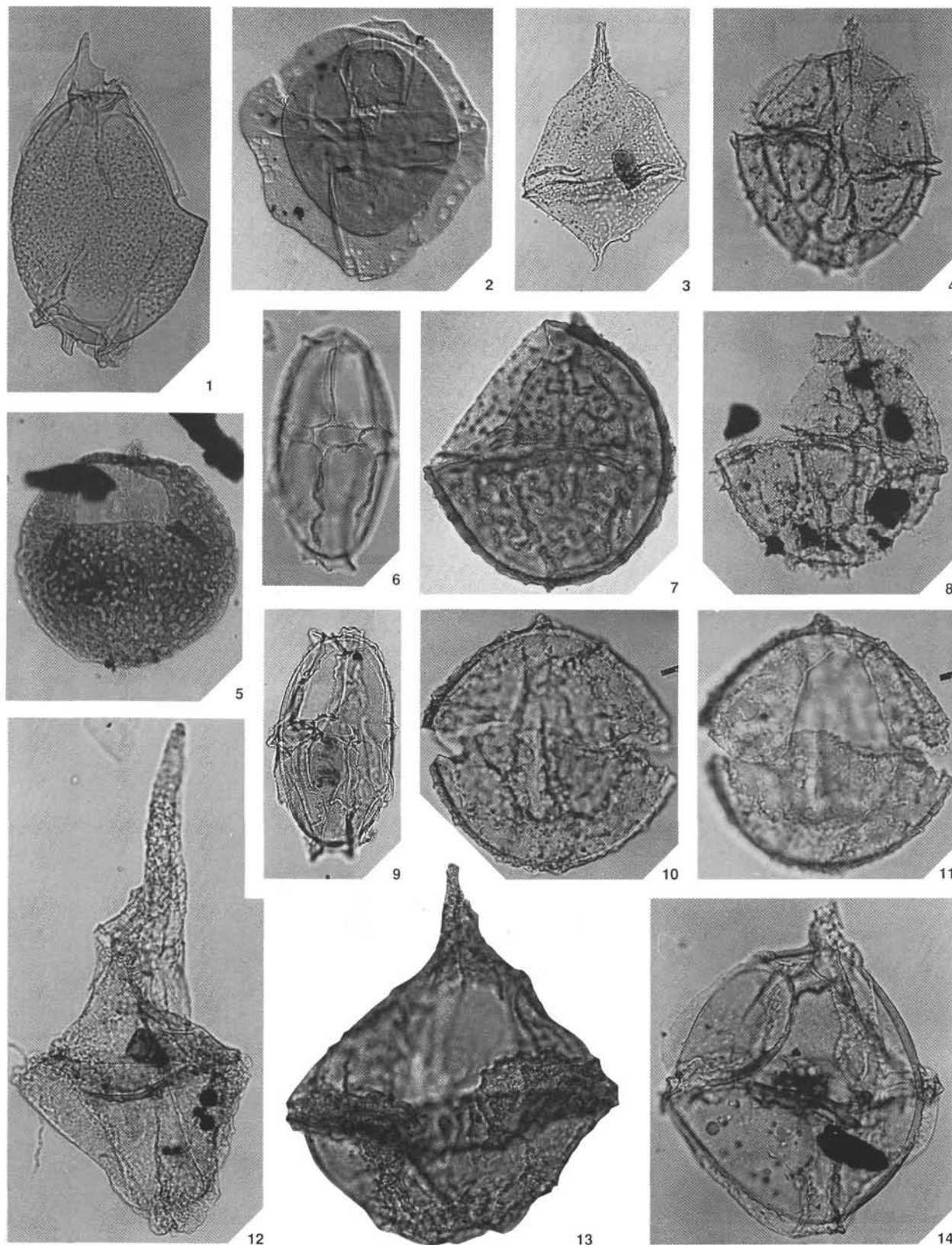


Plate 2. (all x500) **1.** *Herendeenia postprojecta* (Core 123-765C-51R-5 SSM1). **2.** *Scriniodinium attadalense* (Core 123-765C-59R-4 SSM31). **3.** *Diconodinium davidii* (Core 123-765C-35R-CC SSM2). **4.** *Occisucysta tenuiceras* (Core 123-765C-40R-4 SSM2). **5.** *Aldorfia?* sp. A (Core 123-765C-59R-4 2/922 199). **6.** *Carpodinium granulatum* (Core 123-765C-51R-5 SSM4). **7.** *Cribroperidinium* sp. cf. *C. asarotum* (Core 123-765C-54R-4 SSM25). **8.** *Occisucysta tenuiceras* (Core 123-765C-40R-4 SSM2). **9.** *Carpodinium granulatum* (Core 123-765C-50R-CC SSM9). **10,11.** *Cribroperidinium?* sp. cf. *C. asarotum* (Core 123-765C-51R-5 SSM9). **12.** *Cribroperidinium* sp. cf. *C. leedervillense* (Core 123-765C-35R-CC SSM25). **13.** *Cribroperidinium* sp. A (Core 123-765C-54R-4 SSM27). **14.** *Cribroperidium* sp. B (Core 123-765C-45R-1 SSM15).

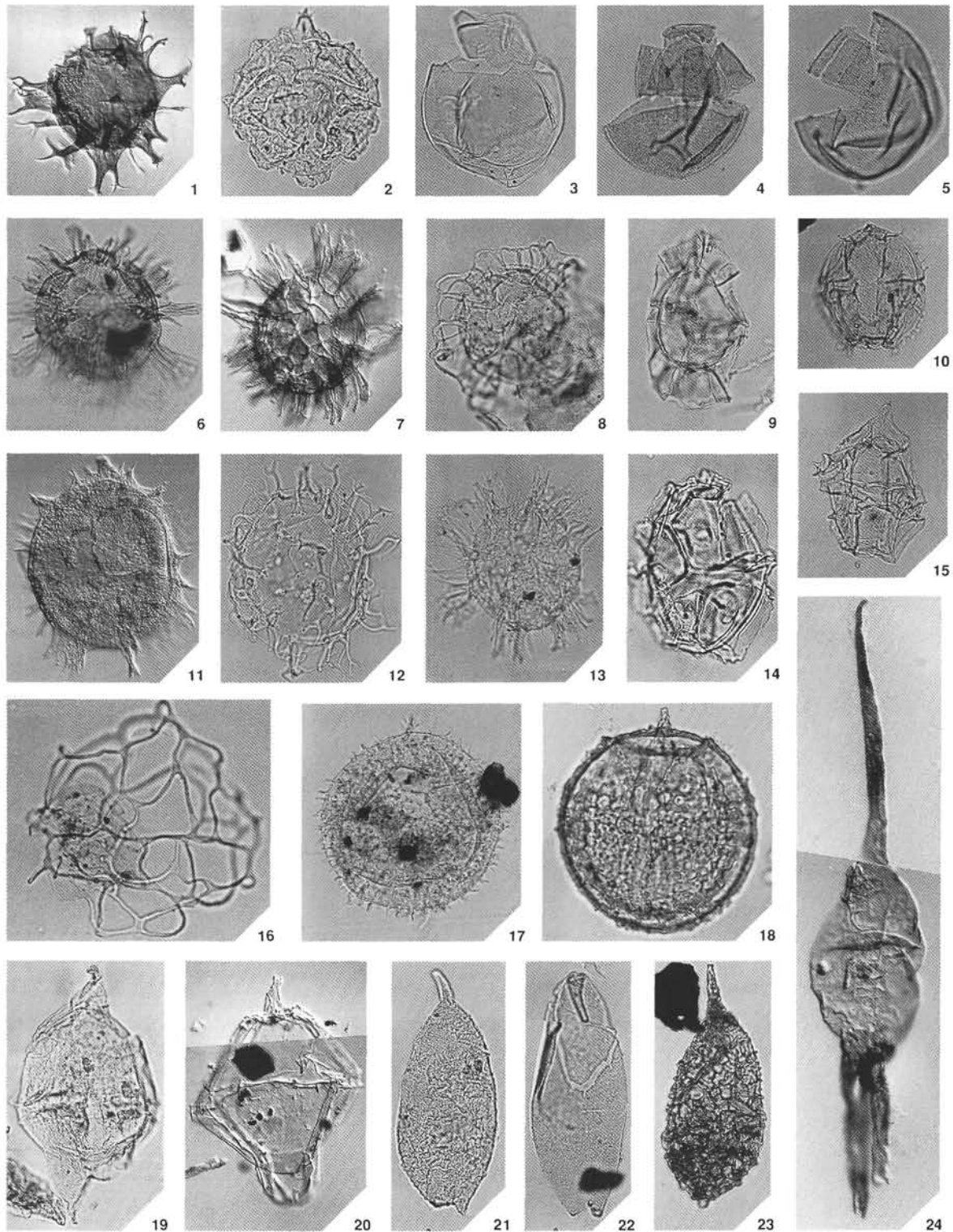


Plate 3. (all x500) **1.** *Achomospaera?* *neptuni* (Core 123-765C-59R-4 SSM36). **2.** *Leptodinium?* sp. (Core 123-765C-40R-4 SSM7). **3.** *Ascodinium?* sp. A (Core 123-765C-35R-CC SSM33). **4.** *Dissiliodinium* sp. A (Core 123-765C-40R-4 SSM19). **5.** *Dissiliodinium* sp. A (Core 123-765C-40R-4 SSM20). **6.** *Kleithriasphaeridium fasciatum* (Core 123-765C-59R- SSM34). **7.** *Kleithriasphaeridium fasciatum* (Core 123-765C-59R-4 1/1100 110). **8.** *Adnatosphaeridium tutulosum* (Core 123-765C-35R-CC SSM31). **9.** *Hystrichosphaeropsis* sp. A (Core 123-765C-35R-CC SSM22). **10.** *Leptodinium?* sp. (Core 123-765C-59R-4/956 040). **11.** *Exochosphaeridium arnace* (Core 123-765C-54R SSM25). **12.** *Avellodinium* sp. A (Core 123765C-51R-5 SSM6). **13.** *Exochosphaeridium* sp. A. (Core 123-765C-40R-4 SSM18). **14.** *Impagidinium phlyctaena* (Core 123-765C-51R-4 SSM4). **15.** *Gonyaulacysta* sp. cf. *G. helicoidea* (Core 123-765C-40R-4 SSM17). **16.** *Hapsocysta perdictya* (Core 123-765C-35R-CC SSM30). **17.** *Trinchodinium castanea* (Core 123-765C-35R-CC SSM24). **18.** *Cribroperidinium* sp. cf. *C. asarotum* (Core 123-765C-50R-CC SSM4). **19.** *Ascodinium* sp. A (Core 123765C-35R-CC 1/1160 180). **20.** *Tubotuberella vlamingii* (Core 123-765C-59R-4 SSM33). **21.** *Batioladinium micropodum* (Core 123-765C-35R-CC SSM14). **22.** *Batioladinium micropodum* (Core 123-765C-40R-4 SSM30). **23.** *Batioladinium reticulatum* (Core 123-765C-59R-4 1/1092 218). **24.** *Broomea simplex* - reworked (Core 123-765C-59R-4 SSM28).

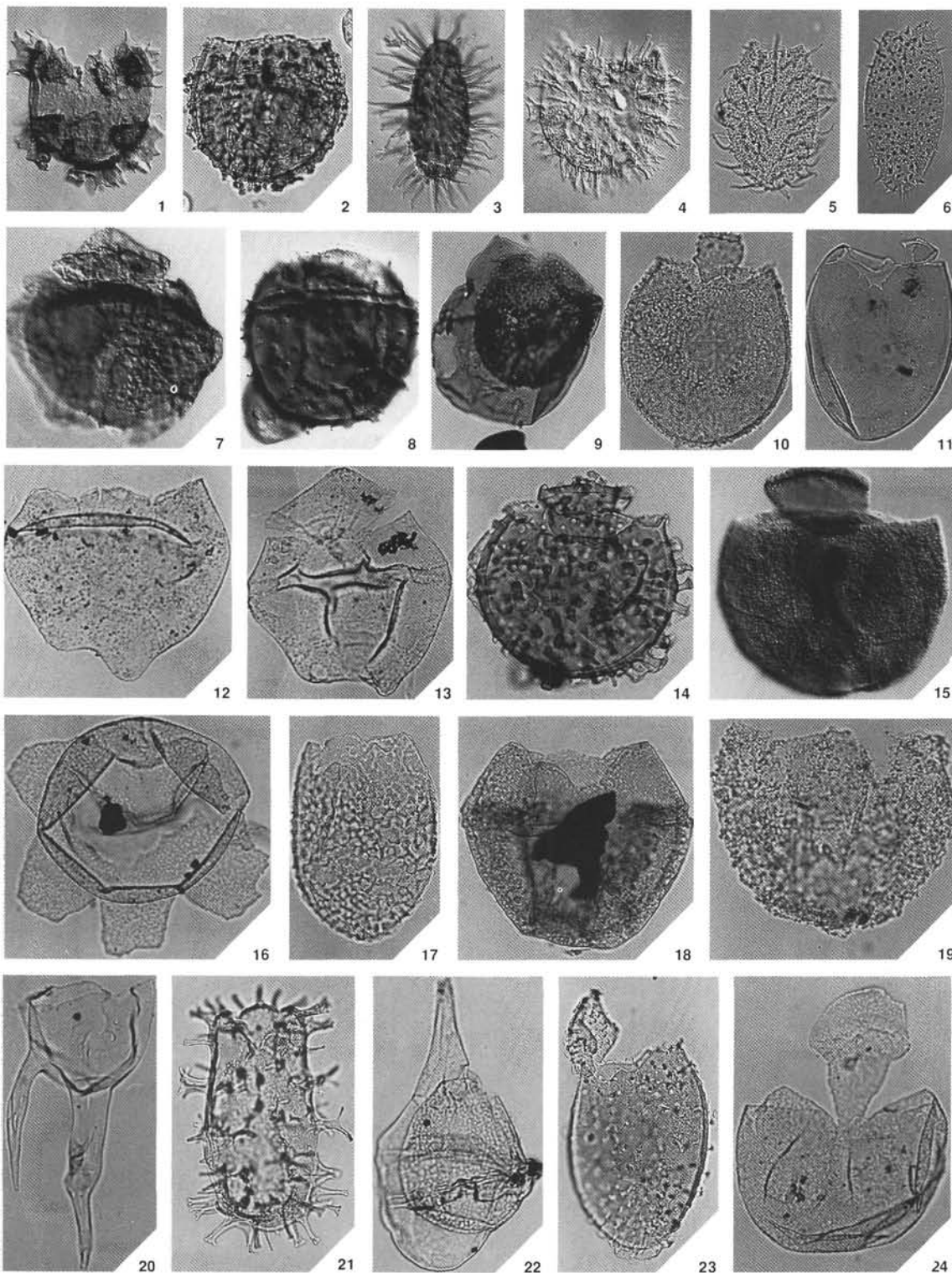


Plate 4. (all x500) **1.** *Oligosphaeridium* sp. cf. *O. poculum* (Core 123-765C-59R-4 SSM24). **2.** *Meiourogonyaux*? sp. A (Core 123-765C-50R-CC SSM5). **3.** *Tanyosphaeridium* sp. A (Core 123-765C-59R-4 SSM16). **4.** *Epelidosphaeridia colligata* (Core 123-765C-54R SS13). **5.** *Prolixosphaeridium parvispinum* (Core 123-765C-40R-4 SSM46). **6.** *Prolixosphaeridium inequornatum* (Core 123-765C-54R-5 SSM11). **7.** *Senoniasphaera* sp. cf. *S. ptomatis* (Core 123-765C-59R-4 SSM23). **8.** *Meiourogonyaux* sp. B (Core 123-765C-59R-4 SSM10). **9.** *Dingodinium jurassicum* (Core 123-765C-59R-4 2/911 130). **10.** *Batiacasphaera* sp. (Core 123-765C-40R-4 SSM46). **11.** *Batiacasphaera*? sp. (Core 123-765C-40R-4 SSM51). **12.** *Canninginopsis intermedia* (Core 123-765C-35R-CC SSM6). **13.** *Canninginopsis intermedia* (Core 123-765C-35R-CC SSM6). **14.** *Meiourogonyaux*? sp. A (Core 123-765C-45R-1 SSM10). **15.** *Batiacasphaera* sp. A-*imperfecta* plexus (Core 123-765C-59R-4 1/995 210). **16.** *Escharisphaeridia* sp. A (Core 123-765C-45R-1 SSM4). **17.** *Batiacasphaera* sp.-*imperfecta* plexus (Core 123-765C-51R-5 SSM6). **18.** *Meiourogonyaux stoveri* (Core 123-765C-40R-1 SSM33). **19.** Genet sp. indet. A (Core 123-765C-40R-4 SSM37). **20.** *Odontochitina operculata* (Core 123-765C-40R-4 SSM31). **21.** *Egmontodinium torynum* (Core 123-765C-59R-4 SSM12). **22.** *Dingodinium cerviculum* (Core 123-765C-35R-CC SSM17). **23.** *Batiacasphaera*? sp. *imperfecta* plexus (Core 123-765C-45R-1 SSM8). **24.** *Kallosphaeridium* sp. A (Core 123-765C-45R-1 SSM14).

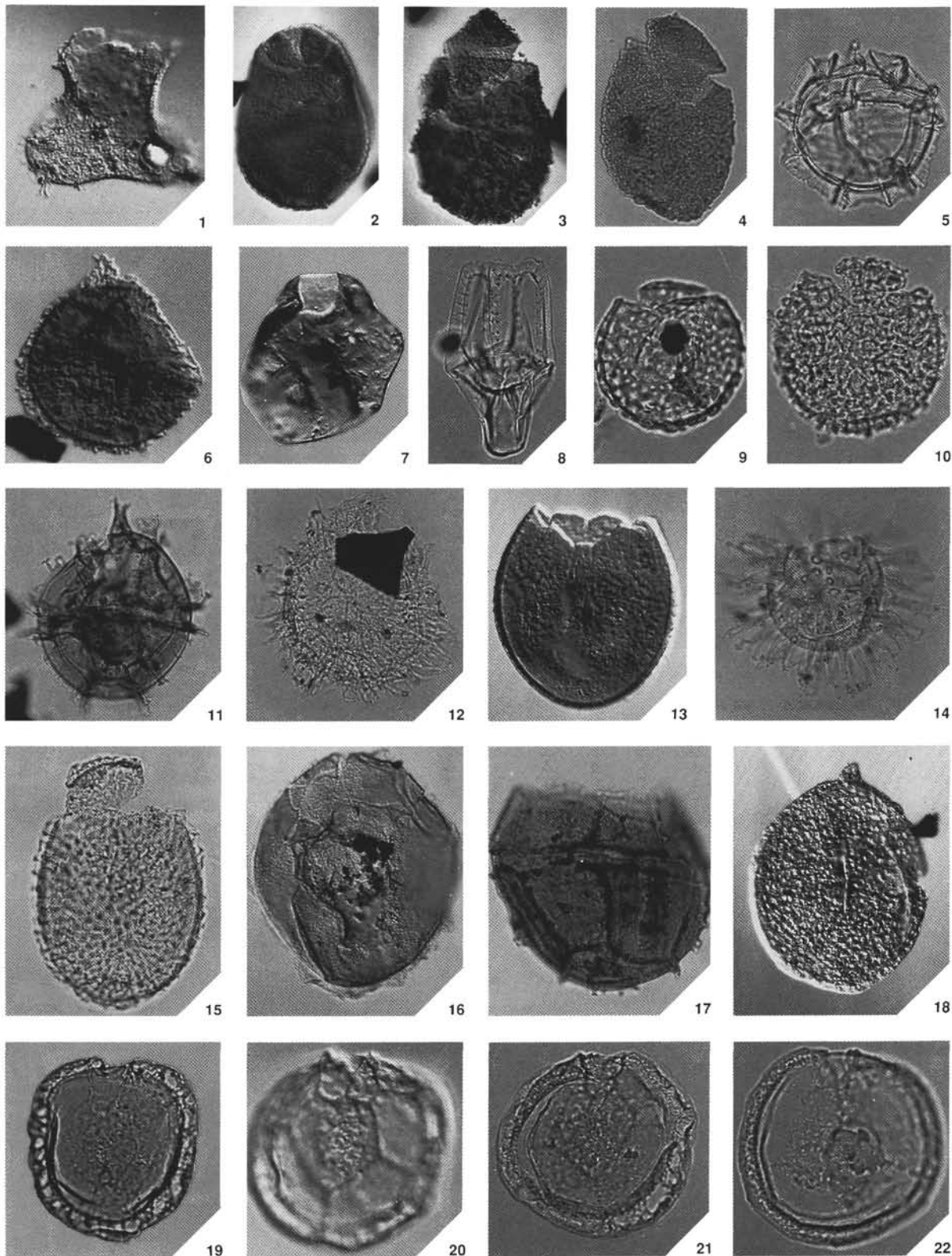


Plate 5. (all x750) 1. *Dissimulidinium lobispinosum* (Core 123-765C-59R-4 SSM17). 2. Gen. et sp. indet. B (Core 123-765C-59R-4 1/1084 190). 3. *Aprobolocysta? galeata* (Core 123-765C-59R-4 2/1030 150). 4. *Aprobolocysta? galeata* (Core 123-765C-59R-4 2/955 225). 5. *Leptodinium* sp. (Core 123-765C-35R-CC SSM15). 6. *Aldorfia deflandrei* (Core 123-765C-59R-4 SSM20). 7. *Dollidinium sinuosum* (Core 123-765C-59R-4 SSM41). 8. *Belodinium dysculum* (Core 123-765C-51R-6 SSM3). 9. *Cassiculosphaeridia* sp. A (Core 123-765C-40R-4 SSM50). 10. *Batiacasphaera* sp.-*imperfecta* plexus (Core 123-765C-40R-4 SSM48). 11. *Spiniferites? sp A* (Core 123-765C-59R-4 1/1113 040). 12. *Cometodinium? whitei* (Core 123-765C-40R-4 SSM28). 13. *Batiacasphaera* sp.-*subtilis* plexus (Core 123-765C-54R-4 SSM13). 14. *Dapsilidinium warrenii* (Core 123-765C-40R-4 SSM44). 15. *Batiacasphaera* sp.-*imperfecta* plexus (Core 123-765C-50R-CC SSM5). 16. Gen. et sp. indet. C (Core 123-765C-59R-4 1/900 145). 17. *Meiourogonyauxalax* sp. B (Core 123-765C-59R-4 1090 210). 18. *Aldorfia? sp. A* (Core 123-765C-59R-4 SSM29). 19–22. *Epitricysta* sp. A (all Core 123-765C-54R-4 SSM19).