3.3.2.1 Subtribe Erysiphinae

3.3.2.1.1 Erysiphe

Erysiphe DC., Fl. Fr. II, p. 272 (1805) (:Fr., Syst. myc. 3, p. 234, 1829) emend. Léveillé (1851)

•Synonyms: Alphitomorpha auct. p.p. Tigria Trev., Spighe e Pagli, Sor. Bot, varj. 1, p. 22 (1853). Erysiphella Peck, 28th Rep. N.Y. Stat. Mus. 2, p. 63 (1874). Erysiphopsis Halst., Bull. Torrey bot. Club 26, p. 594 (1899). Ortochaeta Sawada, Taiwan Agr. Exp. Stat. 85, p. 22 (1943). Linkomyces Golovin (1958, p. 127) nom. nud. Salmonomyces Chidd., Sydowia 13, p. 55 (1959). Ischnochaeta Sawada, Descr. Cat. Taiwan Fungi XI, p. 16 (1959). Golovinomyces (U. Braun) Geljuta, Biol. Z. Arm. 41(5), p. 357 (1988). Uncinula auct. p.p. *Type species: Erysiphe polygoni DC. (selected by Clements & Shear 1931).

Mycelium external, superficial, haustoria in the epidermal cells, appressoria present, unlobed, \pm nipple-shaped to lobed, hyphac ca. 2-12 μ m wide, branched, seplate, anamorphic states belonging to *Oidium*, conidiophores arising from the superficial mycelium, erect, conidia in chains or formed singly, shape and size variable, fibrosin bodies absent. Erysiphacean eleistothecia globose to depressed globose, non-dorsiventral, peridium multilayered, coloured, dark, not transparent, appendages usually mycelioid, simple to irregularly branched, rarely setiform, mostly arising from the lower half of the ascocarp, sometimes from base to top, asei numerous, 2-8-spored, spores developed in the current season or after overwintering.

•Notes: Erysiphe is usually considered as the most ancient genus of the Erysiphaceae. There are some well-established morphological groups within this genus. Braun (1978, 1981) dealt with this problem, and the following classification was proposed: (1) Erysiphe sect. Erysiphe (type species - E. polygoni)

Conidia formed singly (*Pseudoidium* (ypc), appressoria mostly ± lobed, asci (2-) 3-8-spored.

(a) subsect. Erysiphe Asci (2-) 3-6-spored.

(b) subsect. Polysparae U. Braun (1981, p. 693) Asci 5-8-spored, rarely 5-spored, often 8-spored (type species - E. ulmariae).

(2) Erysiphe sect. Golovinomyces U. Braun (1978, p. 659)
 (type species - E. cichoracearum)

Conidia in chains (*Euoidium* type), appressoria \pm nipple-shaped, sometimes indistinct, asci 2 (-4)-spored.

(a) subsect. Golovinomyces

Foot-cells of the conidiophores cylindric, straight to curved, mostly shorter than 100 µm, conidia ellipsoid-ovoid to doliiform, without constricted areas.

(b) subsect. Depressa U. Braun (1981, p. 695) Foot-cells of the conidiophores long, often longer than 100 μ m, width increasing from base to top, conidia broad, often with constricted areas at the ends (type species - *E. depressa*).

(3) Erysiphe sect. Galeopsidis U. Braun (1981, p. 690) (type species - E. galeopsidis)

Conidia in chains (*Euoldium* type), but appressoria lobed, ascospores not developed before overwintering, asci always immature in the current season.

The three sections are well-characterized. Sect. Galeopsidis takes an intermediate position between sect. Erysiphe and sect. Golovinomyces, The asci are (2-) 3-6 (-8)-spored and the appressoria are lobed (agreeing with sect. Erysiphe), but the conidia are produced in chains (as in sect. Golovinomyces). There are also some additional intermediate species (e.g. E. galii var. galii in sect. Golovinomyces - ascospores only developed after overwintering; some forms of E. cichoracearam with moderately lobed appressoria or with 2-4-spored asci; some species in sect. Erysiphe with unlobed appressoria). Sawada (1949, 1951, 1959) was the first author to propose the separation between sect. Erysiphe and sect. Golovinomyces on generic level. He established the genus *lschnochaeta* for the species with *Pseudoidium* anamorphs. Golovin (1958) introduced a similar proposal. He established the new genus Linkomyces. Geljuta (1988) raised Erysiphe sect. Golovinomyces to generic rank. However, the separation of Erysiphe into two distinct genera is not tenable because of the mentioned intermediate species as well as the existence of sect. Galeopsidis.

Key to the species

1. Conidia in chains (Euoidium type), appressoria usually nipple-shaped, asei 2-spored, very rarely with 3 or 4 spores, ascospores developed before overwintering (sect. Golovinomyces). 30 1. Conidia formed singly (Pseudoidium type), appressoria usually \pm lobed, asei (2-) 3-8-spored, spores developed before overwintering (sect. Erysiphe), or conidia in chains, but appressoria lobed and asei without spores in the current season, spores only developed after overwintering (sect. Galeopsidis). 2

2. Conidia formed singly, appressoria lobed, asciwith spores before overwintering (seet. *Erysiphe*).

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Conidia in chains, appressoria ± lobed, asci always without spores before overwintering.
 Appressoria nipple-shaped to slightly lobed, asci of overwintered ascocarps 2-4-spored, mostly
 2-spored, on Galium aparine and related taxa.

(59) E. galii vat. galii

3. Appressoria usually lobed, asci of overwintered ascocarps 2-8-spored, mostly 3-6-spored, on host species of the Lamiaceae. (58) B. galeopsidis 4(2). Asci usually 6-8-, frequently 8-spored (subsect. Polysporae). 5

4. Asci 2-7-, usually 3-6-spored, occasionally 5-7-spored, but never 8-spored (subsect, Erysiphe).

Cleistothecia large, ca.120-160 μm diam., appendages 2-5 times as long as the cleistothecial diam., asci 8-15, on Alnus. (37) E. vernalis
 Cleistothecia smaller, ca. 75-140 μm diam., on other hosts. 6

6. Appendages very short, mostly shorter than the cleistothecial diam., frequently irregularly branched, peridium dark, on *Cirsium*.

(34) E. mayorii vat, mayorii

6. Appendages long and narrow, 0.5-6 times as long as the cleistothecial diam., the asci are easily to be seen within the intacl ascocarps, peridium not very dark, on *Filipendula*. (35) *E. ulmariae* 7(4). Infections often confined to stems, cleistothecia large, (100-) 110-185 (-210) µm diam., appendages fairly thick-walled throughout, strongly flexuous, undulate-geniculate, on Astragalus.

(36) E. caulicola

(if ascocarps ca.110-180 μ m diam., appendages narrow, ca. 3.5-8.5 μ m wide, \pm thick-walled, strongly undulategeniculate, on *Hedysorum*. Anthyllis, cf. immature samples of *Microsphaera hedysari*)

7. Cleistothecia either smaller and/or appendages thin-walled, not confined to stems. 8

8. Appendages frequently irregularly branched. 9 8. Appendages simple (or only a few appendages rarely irregularly branched). 16 (if appendages attached to the equatorial zone or somewhat in the upper half, septate or aseptate, horizontally spread or with a tendency to turn towards one direction, often somewhat thick-walled at the hase, cf. immature samples of Microsphaera species - on Astragalus, cf. M. astragali; on Vicia, cf. M. baeumleri; on various other legumes, cf. M. trifolit; on Hypericum, cf. M. hypericacearum; on Cornus, cf. M. tortilis)

9. Associarps with 6-16 asci, frequently more than 10. appendages constantly short, mostly about as long as the cleistothecial diam. or shorter, on Cicerbita. (34) E. mayoril var. cicerbitae 9. Ascocarps with fewer asci, usually less than 10, and/or appendages longer. 10 10. Appendages long, 0.5-5 times the cleistothecial diam., conidia \pm cylindric, (25-) 40-55 (-65) x 14-22 µm, asci (2-) 3-4 (-6)-spored, on Convolvulus (= var. convolvuli) or asci (3-) 5-6-spored, on Calystegia (= var. calystegiae). (56) E. convolvuli 10. Appendages shorter (and/or conidia smaller).

11

11. Appendages very frequently branched, branchings coral-like, hyaline or only faintly coloured, yellowish, aseptate or few inconspicuous septa, verrucese throughout, on *Paeonia*.

(57) E. paeoniae

11. Branchings moderate, appendages brown when mature, at least below, with many conspicuous septa. 12

12. Foot-cells of the conidiophores long, sometimes slightly flexuous, (35-) 40-75 (-100) x 6.5-8.5 (-10) μ m, on Caryophyllaceae. (55) *E. buhril* 12. Conidiophores shorter, straight, on other hosts. 13

13. Conidia \pm ellipsoid, on hosts of the Fabaceae, especially *Ononis* and *Lathyrus*.

(38) E. pisi var. cruchetiana 13. Conidia cyliadric, on other hosts. 14 14. Foot-cells of the conidiophores usually followed by 1-2 shorter cells or cells of about the same length, on Polygonaceae. (41) E. polygoni 14. Foot-cells frequently followed by a longer second cell and a shorter third cell (dominant arrangement), on other hosts. 15

 On Beta and Chenopodium (Chenopodiaceae), branches moderately developed. (52) E. betae
 On Apiaceae, branches strongly developed.

(51) E. heraclei

16(8). Ascocarps small, ca. 60-115 μ m diam., mostly 80-105 μ m in diam., average mostly below 100 μ m.

 Ascocarps larger, ca. 80-150 μm diam., average larger than 100 μm.

17. Appendages very long, often setiform, ca. 3-12 times as long as the eleistothecial diam., on Ranunculaceae. (47) *E. aquilegiae* var. aquilegiae
17. Appendages shorter, ca. 0.25-4 times the eleistothecial diam., mycelioid. 18

18. Appendages brown throughout when mature, on *Circaea* or host species of the Ranunculaccae, 19. 18. Appendages coloured in the lower half, yellow-

ish to brown, paler or hyaline in the upper part, on other hosts. 20

19. Foot-cells of the conidiophores frequently curved or floxuous, on Circaea. (46) E. circaeae 19. Foot-cells straight, erect, not curved, not flexuous, on Ranunculaceous hosts.

(47) E. aquilegiae var. ranunculi
 20(18), Conidia 32-46 x (13-) 15-20 (-25) μm, ascospores (18-) 20-30.5 x (8.5-) 11-17 μm, on Dipsacaceous hosts.
 (45) E. knautiae
 20. Conidia and ascospores smaller on Counting

Conidia and ascospores smaller, on Geranium.
 (49) E. geraniacearum

21(16). Appendages not very numerous, setiform, stiff and straight, 0.5-1.5 times as long as the cleistothecial diam., aseptate or with few septa, on *Catalpa*. (48) *B. catalpae*

 Appendages not setiform, but flexuous, mycelioid, usually richly and conspicuously septate.
 Appendages short, 0.5-1.5 times as long as the cleistothecial diam., byaline or only faintly coloured, on *Limonium* or Urtica,
 23

22. Appendages longer, 0.5-4 times as long as the cleistothecial diam., often brown, on other hosts.

Conidia ellipsoid-cylindric, wide, ca. (25-)
 30-40 x (12-) 15-20 μm, on Urtica. (50) E. urticae

23. Conidia \pm cylindric, narrow, ca. 25-47 x 11-16 (-19) μ m, on *Limonium*. (54) E. limonii 24(22). Conidia large, \pm cylindric, ca. 35-45 x 13-19 μ m (appendages often irregularly branched), on Polygonaceae, (41) E. polygoni 24. Conidia \pm ellipsoid (- ovoid or doliiform), and/or appendages simple, on other bosts. 25

Ascospores broadly ovoid to subglobose, (18-)
 20-26 (-30) x (11-) 12.5-18 (-20) μm, on Sedum.
 (40) E. sedi

25. Ascospores narrower, not subglobose, on other hosts. 26

26. Conidia cylindric (- doliiform), (27-) 30-48 (-55) x 12-21 μm, on hosts of the Capparales (Capparidaceae, Cleomaceae, Brassicaceae, Reseduceae) and Papaverales (Fumariaceae, Papaveraceae).

(43) B. cruciferarum 26. Conidia ± ellipsoid (- ovoid, dolliform), on other hosts. 27

27. Appendages not very numerous, often only
5-15, fairly long, ca.1-4 times the cleistothecial diam., asci 3-6-spored, on *Thesium*. (39) *E. thesii*27. Appendages numerous, and/or appendages shorter, on other bosts. 28

28. Foot-cells of the conidiophores subcylindric or often somewhat decreasing in width from base to top, (15-) 20-50 (-70) x 6-10 μ m, on Fabaceae.

(33) E. pisi var. pisi
28. Foot-cells cylindric, on other hosts.
29. Conidia (20-) 30-40 (-45) x (12.5-) 16-22.5
(-25) μm, on Anchusa.
(53) E. lycopsidis
29. Conidia narrower, ca.13-17.5 μm wide, on Lythrun.
(42) B. lythri

30(1). Conidiophores very long, foot-cells or sometimes secondary cells very long, ca. 80-250 μ m, width conspicuously increasing from base to top, conidia large, especially wide, 25-50 (-70) x 18-30 μ m, mostly wider than 20 μ m, l/w ratio helow 2, broadly ellipsoid-ovoid to doliiform, often with somewhat constricted ends, on *Verbascum* or hosts of the Asteraceae (subsect. *Depressa*). 31

30. Conidiophores shorter, foot-cells about 40-140 μ m long, mostly 40-80 μ m, cylindric, conidia narrower, ca. 14-22 μ m, rarely wider (subsect. Golovinomyces). 34

31. Ascocarps ca. 95-150 μ m diam., appendages very short, shorter than the eleistothecial diam., often rudimentary, 3.5-9 μ m wide, hyaline to faintly coloured, on *Verbascum*. (72) *E. verbasci* 31. Cleistothecia often larger, more than 150 μ m diam., or appendages well-developed, wider, brown when mature. 32

32. Cleistothecia 80-145 µm diam., appendages

well-developed, brown when mature, ascospores 20-32 x 13-20 µm, on Arctium, Centaurea, Onopordum. (71) E. depressa

Cleistothecia larger, often more than 150 μm diam., appendages hyaline or faintly coloured, and/ or ascospores very large, 24-43 x 16-25 μm.
 Cleistothecia ca.165-210 μm diam., ascospores 20-23 x 16-18 μm, conidiophores with a long footcell, followed by some shorter cells, on *Helichrysum*.
 Cleistothecia 120-180 μm diam., ascospores 33. Cleistothecia 120-180 μm diam., ascospores 33.

24-43 x 16-25 μ m, one or two short or moderately long basal cells of the conidiophores are followed by a very long cell, on *Echinops*. (70) *E. echinopis* 34(30). Cleistothecial appendages short, usually shorter than the cleistothecial diam., often rudimentary, narrow, ca. 2.5-8 μ m wide, hyaline or only faintly coloured, conidia ellipsoid-ovoid to doliiform, short and broad, on *Artemisia*, very rarely on *Achillea millefolium*. (66) *E. artemisiae* 34. Appendages longer, and/or wider, brown when mature, conidia \pm ellipsoid, cylindric (- ovoid), slender. 35

Appendages short, 0.25-1.5 times as long as the cleistothecial diam., usually shorter than the cleistothecial diam., on *Hyoscyamus*. (60) *E. hyoscyami* Appendages longer, on other hosts. 36
 Infections characteristic, dense, white, persistent patches, often confluent, outer peridium cells of the ascocarps large, 10-40 μm diam., on *Phlox* and *Polemonium*.

 (61) E. magnicellulata var. magnicellulata
 36. Appearance of the infections different, and/or peridium cells smaller, on other hosts. 37
 37. Appendages of mature ascocarp usually obviously vertucose, on Valeriana, Centranthus.

(69) E. valerianae 37. Appendages smooth to faintly rough, on other hosts. 38 38. Conidiophores frequently constricted at the basal septum, mature asci filled with numerous oil

drops, on Lamiaceae. (63) *E. biocellata* 38. Conidiophores usually not constricted at the basal septum, mature asci without numerous oil drops. 39

39. Conidia 28-45 x 18-27 μm, broadly ellipsoid-ovoid to doliiform, l/w ratio 1.3-2, mostly 1.4-1.6, on Ambrosia, Iva, Helianthus, Rudbeckia.

(64) *E. cichoracearum* var. *latispora* 39. Conidia narrower, often slender, l/w ratio around 2 or larger. 40

40. Mycelium usually forming persistent, dense, limited patches, germ tubes of the conidia short, often twisted, on Boraginaceae. (62) E. cynoglossi 40. Appearance of the mycelium different, and/or germ tubes longer, not twisted, on other hosts. 41 41. Foot-cells of the conidiophores frequently curved, on *Plantago* or *Senecio sylvaticus*, S. viscosus, S. vulgaris, occasionally on other hosts. 42 41. Foot-cells straight, cylindric. 44

Ascocarps large, ca. 120-160 μm diam., on Senecio. (64) E. cichoracearum var. fischeri
 Ascocarps smaller, on other hosts. 43
 On Plantago (Plantaginaceae). (67) E. sordida
 On other hosts, cf. E. orontii

44(41). Outer peridium cells of the ascocarps 10-30 (-40) μm diam., appendages sometimes in the upper

half of the ascocarp, on Galium. (59) E. galii var. riedliana 44. Peridial cells ca. 8-25 μm diam., appendages in 45 the lower half, on other hosts. 45. Conidiophores erect, conidia in chains, spore units conspicuously swollen, chains often long, appressoria conspicuously nipple-shaped (occasionally slightly lobed), germ tubes moderately long, simple, ascocarps usually developed, asci mostly constantly 2-spored, on various hosts of the Astera-(64) E. cichoracearum var. cichoracearum ceae. 45. Conidiophores erect, foot-cells straight to curved, conidia in chains, chains mostly short, spore units unswollen, appressoria nipple-shaped, but often poorly developed, germ tubes short, often somewhat twisted, seldom forked, ascocarps rarely developed, asci often 3- or 4-spored, on host plants of various families (also on cultivated species of the Asteraceae, e.g. Aster, Chrysanthemum, Dahlia, Helianthus, Solidago). (65) E. orontii

(34) Erysiphe mayorii Blumer, Beitr. Krypt.-Fl. Schweiz 7(1), p. 174 (1933)

var. mayorii

 Synonyms: E. polygoni auct. p.p. E. communis auct. p.p. E. communis f. cirsii Antipova, in Vasjagina et al. (1961, p. 231). Golovinomyces mayorii (Blumer) Geljuta, Flora gribov Ukrainy, mučnistorosjanye griby, p. 135, Kiev 1989.

•Illustrations: Salmon (1900, pl. 7, fig. 134), Blumer (1933, p. 176, fig. 54; 1967, p. 211, fig. 67), Vasjagina et al. (1961, p. 233, fig. 79), Sandu-Ville (1967, p. 203, fig. 32), Bunkina (1979, p. 130, fig. 6), Salata (1985, p. 118, fig. 49), Braun