

Characterization and cross
inoculation studies of *rhizobia*
isolated from crop wild
relatives of *Vigna*

P.Saravana Kumar

Department of Botany

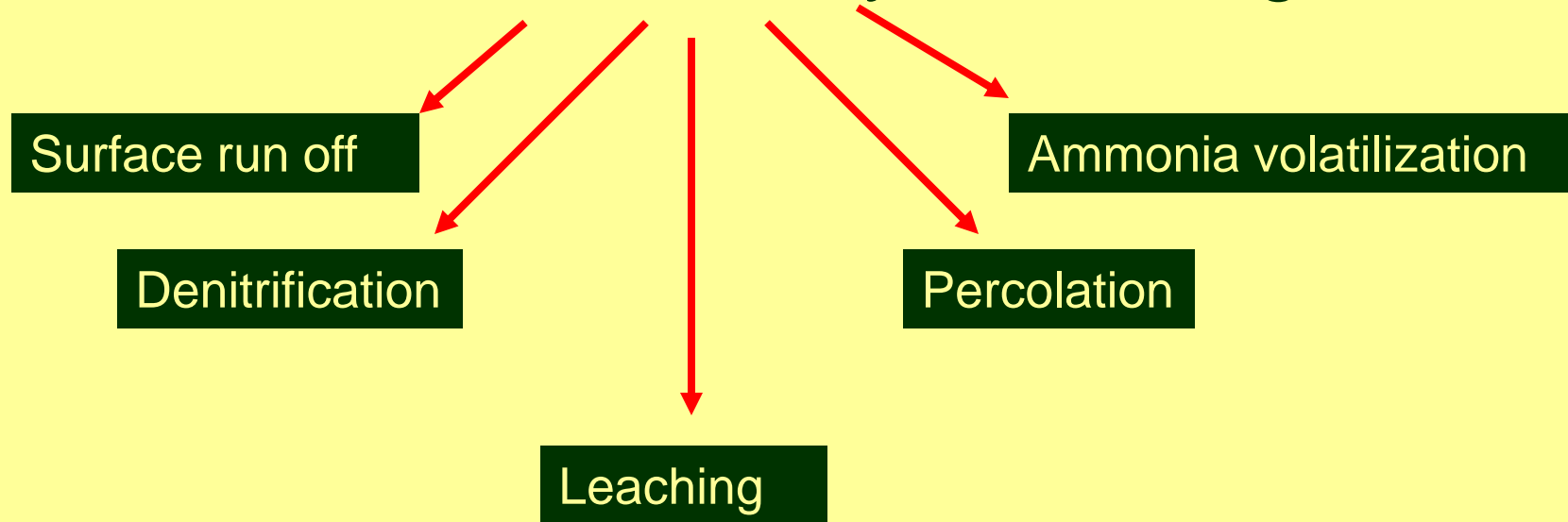
Faculty of Science

University of Pradeniya, Sri Lanka

INTRODUCTION

❖ Importance of **N**

- N - a limiting nutrient for crop production because it easily lost through



**•By adding nitrogen fertilizer
(nitrates, ammonia, urea etc.)**

25%

**soil environment
may be enriched
in N**

**•Non biological N fixation
(storms bring 10 to 15
kg of nitrogen per ha to
the soil every year) - 15%**

**•Through biological
fixation
(by free fixing agents ,
symbiotic fixing agents,
cyanobacteria) - 60%**

BIOLOGICAL N₂ FIXATION

Major contribution – Symbiotic Systems

Rhizobium – Legume associations
50% - all N fixed on Earth

Rhizobia

- Facultative microsymbionts
- Rod shaped
- 1.2-3.0 μm in length 0.5 - 0.9
- Gram negative
- Non spore forming
- Fix N_2 only when symbiotic
- Aerobic
- Optimal growth conditions : T^0 25⁰ - 30⁰C ; pH 6 - 7



Important of rhizobia in wild relatives of *Vigna*

- The successful exploitation of the *rhizobium*-legume symbiosis requires the presence of appropriate strains in the soil.
- *Rhizobia* from the wild relatives of crop legumes are a natural, that has a tremendous potential to offset the use of expensive N-fertilizers

Important of the rhizobial inoculants

Unlike chemical N-fertilizers, rhizobial Inoculants :

- are cheap**
- improve soil fertility by recycling of additional nitrogen obtained from the fixation**
- improve crop productivity**
- replace the mineral N-fertilizer use for grain legumes**
- do not promote weed growth- reduce labour cost and/or use of herbicides for weed control**
- do not promote pollution of soil and water with nitrate.**

CWR Project

A project was aimed to study and explore *rhizobia* from wild relatives of *Vigna* for the crop improvement of *Vigna* group of crops species - (*Vigna radiata* [green gram], *Vigna mungo* [black gram] and *Vigna unguiculata* [cowpea] for crop improvement programmes.

Research Plan

- 1. Field Survey**
- 2. Nodule Collection**
- 3. Nodule Characterization**
- 4. Isolation and Purification of *rhizobia***
- 5. Characterization of stock cultures**
- 6. Authentication**
- 7. Pot Experiments**
- 8. Field Experiments**

Wild relatives of *Vigna* and their distribution in Sri Lanka

Species	Location
<i>Vigna aridicola</i>	Batalow, Mhiyangana, Polonnaruwa, Yala
<i>Vigna dalzelliana</i>	Badulla, Ramboda, Walimada
<i>Vigna radiata</i> , var. <i>sublobata</i>	Mahiyangana
<i>Vigna stipulacea</i>	Humbanthota, Puttalama, Yala
<i>Vigna trilobata</i>	Ampara, Challow, Mountlawaniya, Puttalama Yala
<i>Vigna trinervia</i>	Hakgala, Haliela, Pusallawa, Ramboda, Badulla, Udapussallawa
<i>Vigna marina</i>	Hikkaduwa

Germplasm of Rhizobial isolates from wild relatives of Vigna

Host plant	Code for Isolates	Location
<i>Vigna aridicola</i>	Vr1	Polonnaruwa
<i>Vigna dalzelliana</i>	Vd1	24/7Paradeka/Gampola- Nuwaraeliya
<i>Vigna dalzelliana</i>	Vd2	24/7Paradeka/Gampola- Nuwaraeliya
<i>Vigna dalzelliana</i>	Vd3	34/2Helboda/Gampola- Nuwaraeliya
<i>Vigna dalzelliana</i>	Vd4	Kotmale-Talawakelle
<i>Vigna dalzelliana</i>	Vd5	Hunnasgiriya
<i>Vigna marina</i>	Vma1	Hikkaduwa
<i>Vigna minima</i>	Vmi	Kirinda
<i>Vigna radiata var sublobata</i>	Vrs1	Kitulhitiyaya
<i>Vigna radiata var sublobata</i>	Vrs2	Mhiyangana
<i>Vigna stipulacea</i>	Vst1	Tissa-kirinda
<i>Vigna stipulacea</i>	Vst2	Tissa-kirinda
<i>Vigna stipulacea</i>	Vst3	Kirinda
<i>Vigna radiata var sublobata</i>	Vsu1	Nalanda
<i>Vigna trilobata</i>	Vtril1	34/2Helboda/Gampola- nuwaraeliya
<i>Vigna trinervia</i>	Vtrin1	Ramboda
<i>Vigna trinervia</i>	Vtrin2	Mhiyangana
<i>Vigna trinervia</i>	Vtrin3	Mhiyangana
<i>Vigna trinervia</i>	Vtrin4	24/7Paradeka/Gampola- nuwaraeliya
<i>Vigna wild 1</i>	Vsp1	Putlum -krunagala
<i>Vigna wild 2</i>	Vsp2	Laksapihilla
<i>Vigna wild 3</i>	Vsp3	Kotmale-Talawakelle

1. Field Survey







Vigna marina at Hikkaduwa



Vigna dalzelliana at Hunnasgiriya



Vigna stipulacea at Thissamaharama



Vigna trinervia at mahiyangana



V.stipulaceae and *V. (minima)* growing together in Kirinda



V.aridicola



Arachis pintoi



Pueraria sp.



***Sesbania* sp.**



***Phaseolus* sp.**

2. Nodule collection



3.Characterization of nodules



Vigna unguiculata (Semiglobose)



Vigna aridicola



Vigna mungo



Vigna radiata



Vigna dalzelliana

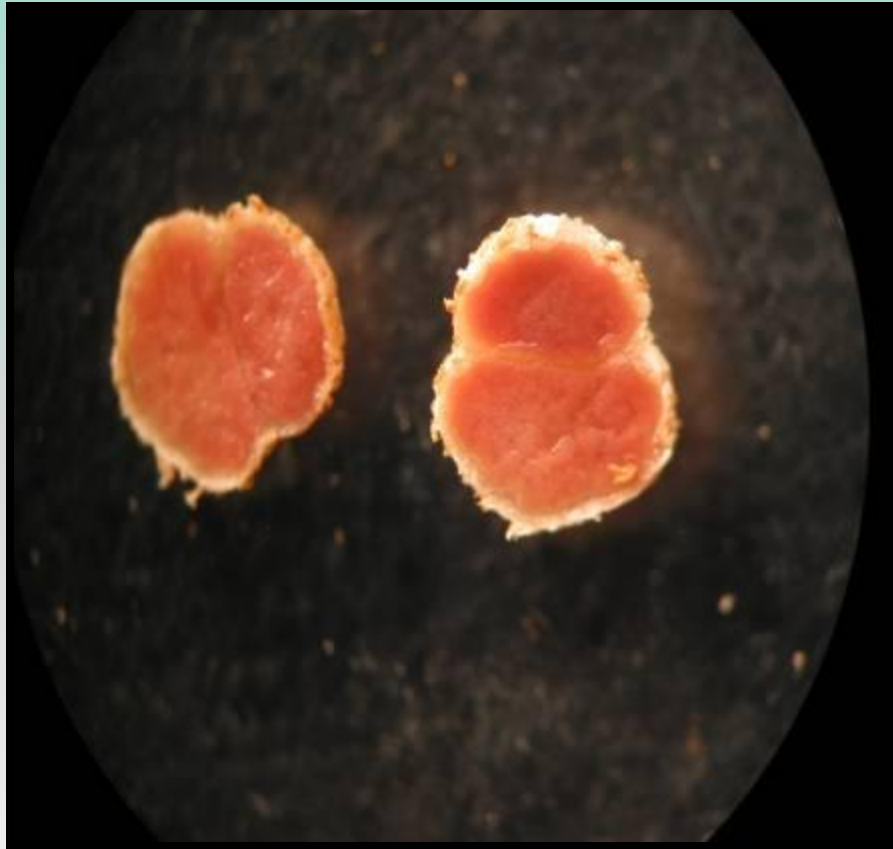


Vigna trinervia





Internal appearance of the nodules



T.S. of a active nodules

4. Isolation and Purification of *rhizobia*

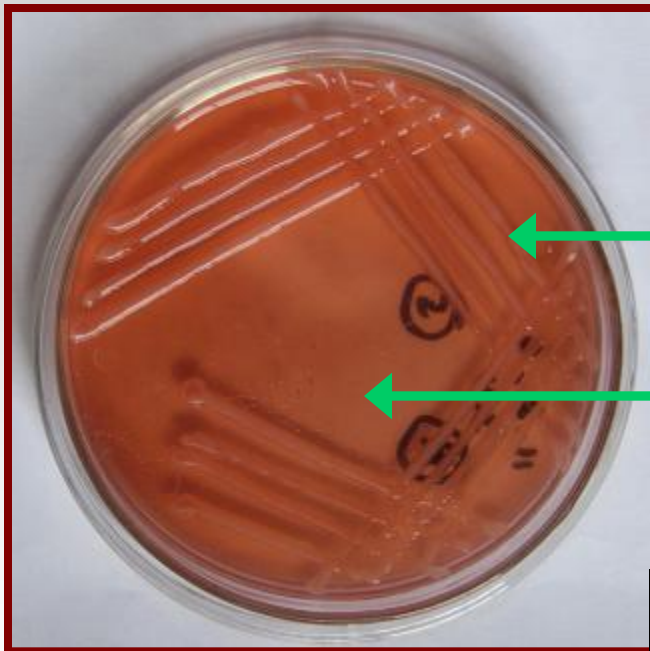


5.Characterization of Stock Culture



Streaking on CRYMA

- All were well grown in to white gummy colonies with in 3 days.



Streaked lines

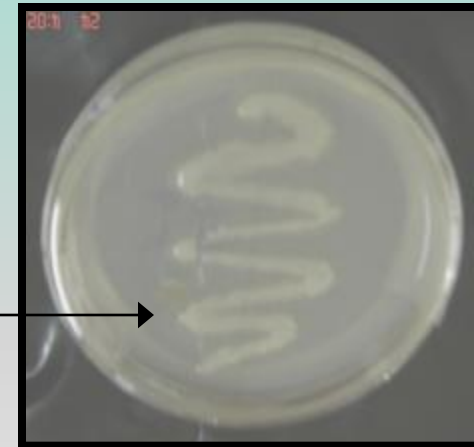
**Red coloured CRYMA
medium**

Ex. A CRYMA plate of V S

Multiplication and Storage

Grown on YMA

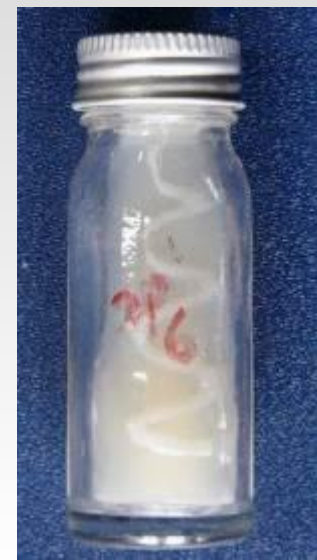
White gummy colonies on **YMA**



Stored on Agar slants

(YMA + CaCo₃)

Cold Storage (long time)



Stock cultures of different isolates





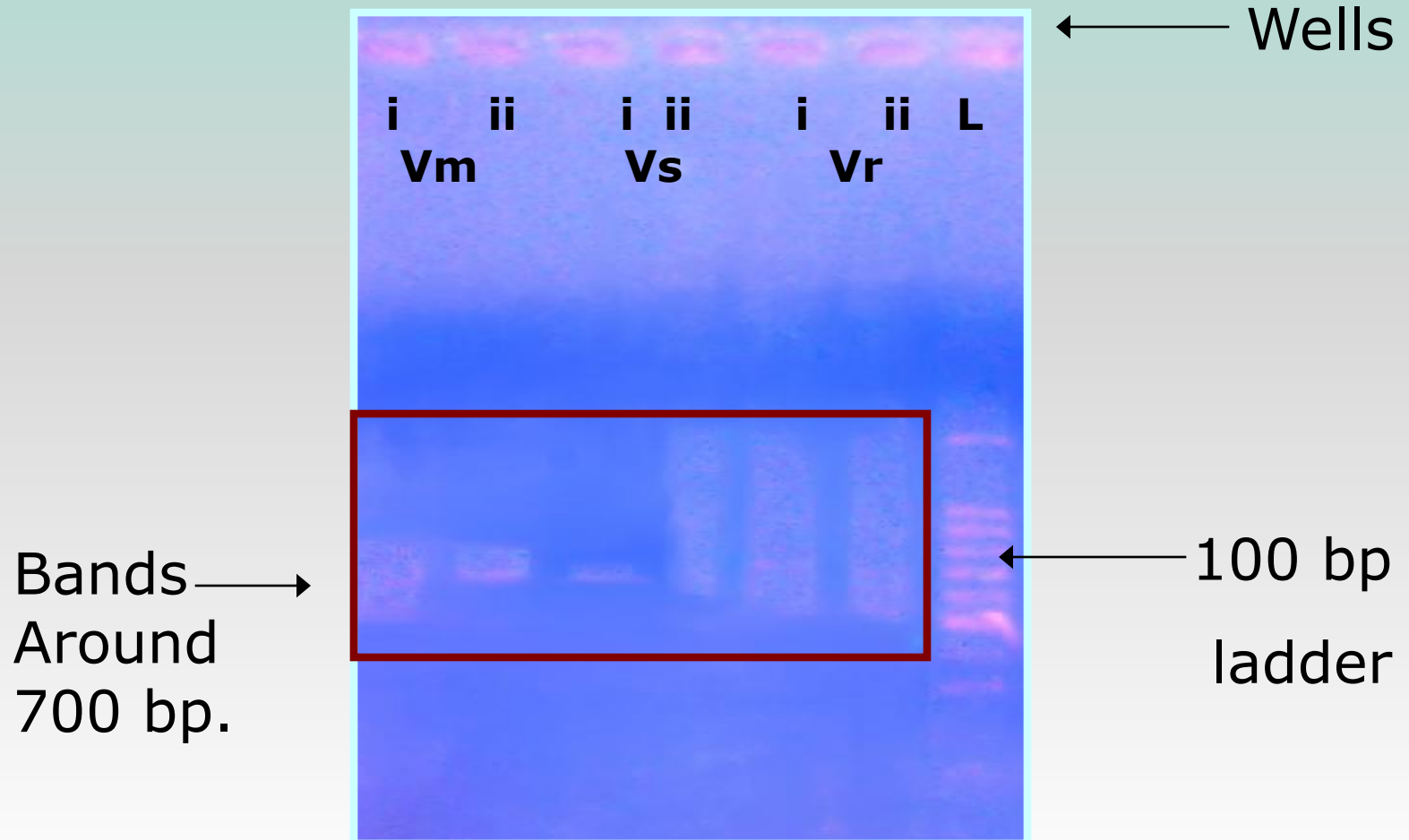
YMB medium, Different strains



Rotary shaker.

Molecular Characterization

Conformation of the Presence of 16 s rDNA IGS Sequence



6. Authentication of the *Rhizobium* isolates

Experimental set-up:

- Free draining pot method
- CRD was used.

Phase I: Pot experiments

- Pot experiments were conducted to test the infectivity of isolated Rhizobia strains from the wild *Vigna* species.
- It was highly promising that the isolates performed well and produced several healthy nodules



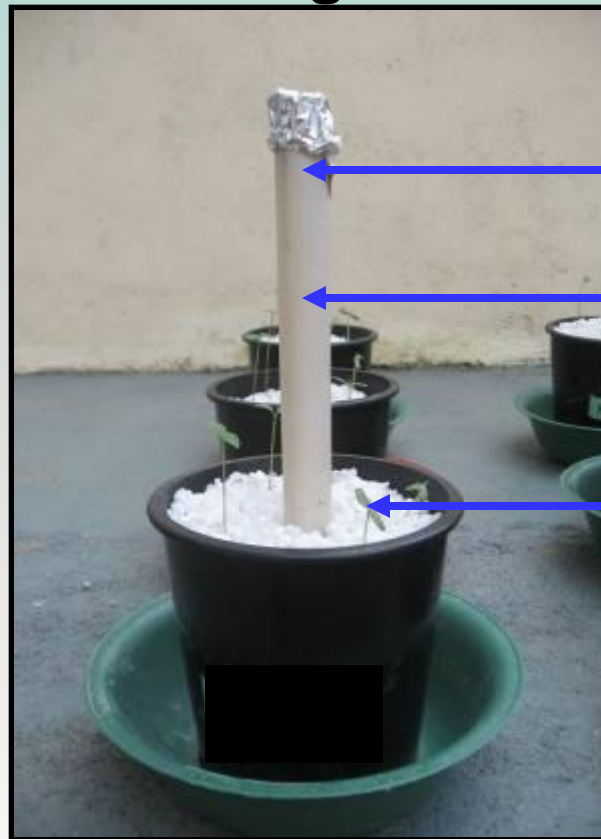
Pot setup just after sowing the seeds



Root inoculation (after 3 days)

Inoculation

- Colonies were washed with 1% sucrose solution
- Inoculated with 1ml of the inoculum
- Covered with sterilized gravel



Aluminium foil

Plastic tube

Sterilized Gravel

Pot after inoculation



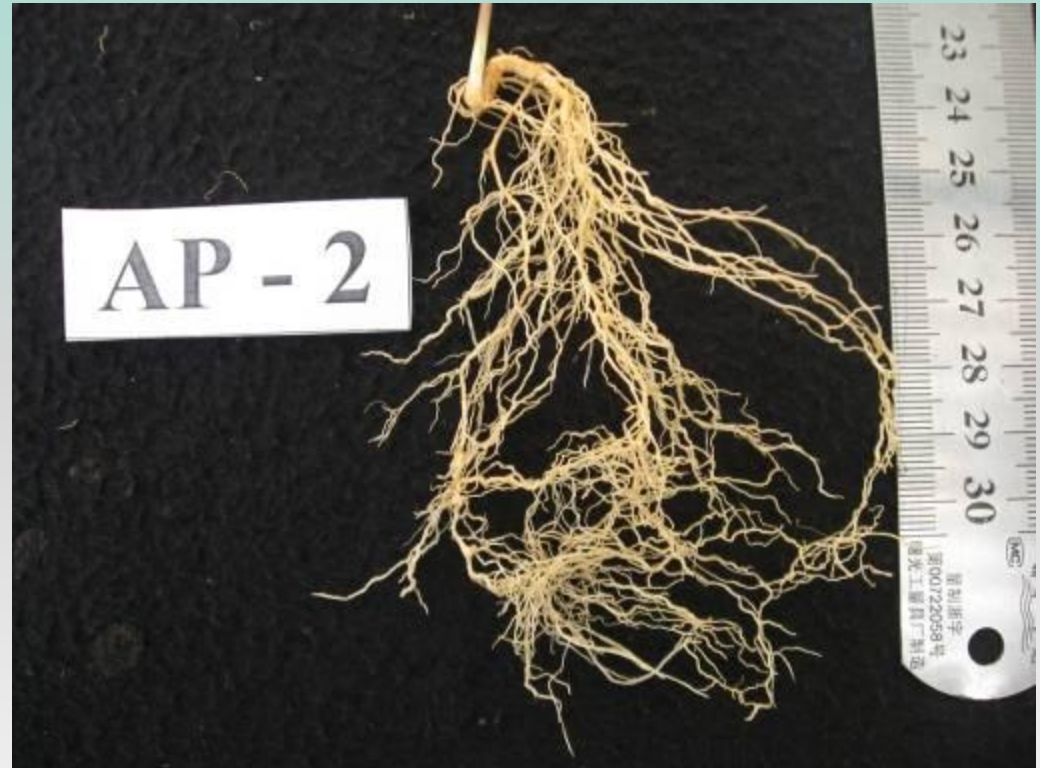
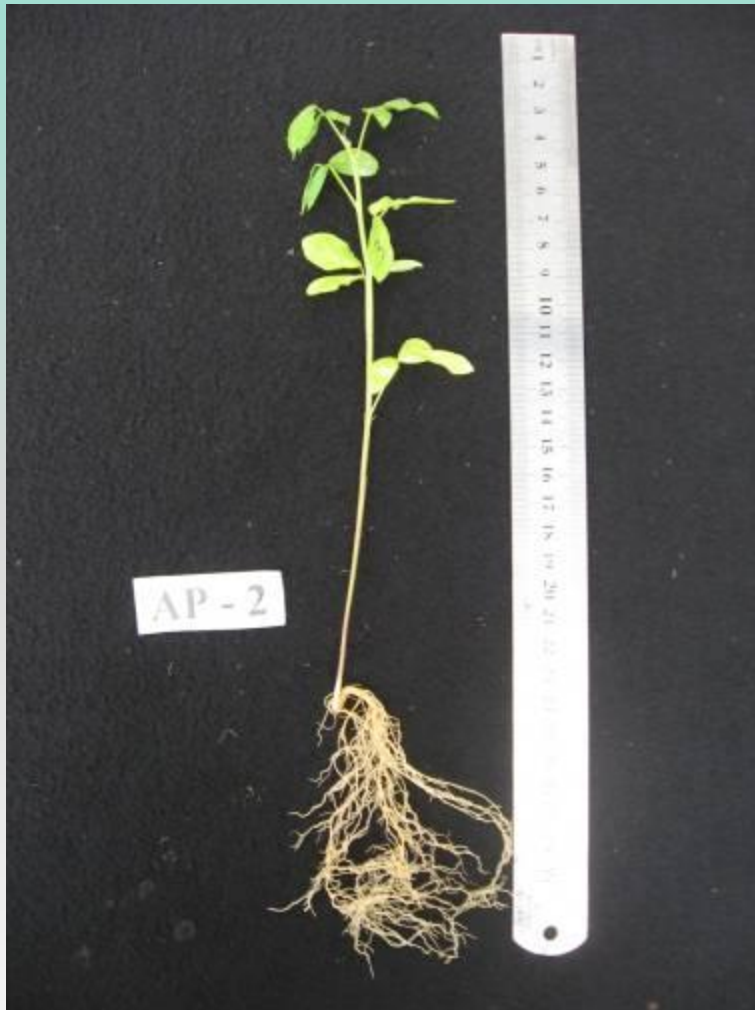


N+ control / Effective / Ineffective / N- control

Effective strain



Ineffective strain



Phase II: Field experiments



Preparation of field for Cross Inoculation Studies



RCBD plot design for field experiments



Greengram and Blackgram plots inoculated with different rhizobial isolates



Greengram inoculated with isolate GG8 showed luxurious vegetative growth compared to adjoining plots



Fig.10 Blackgram inoculated with isolate GG8 showed luxurious vegetative growth compared to adjoining plots



Greengram plants inoculated with BG7 and GG3 showing large and healthy nodules – vital character for effective N fixation



Blackgram plants inoculated with GG3 and GG8 showing nodules



mature plants filled with healthy pods



Harvesting

Note: the white ribbons hanged over the filed to threaten birds and protect yield. White fence for protecting plants from wild animals.

Effective rhizobial isolates obtained from CWR

Code number	Host plant	Collected Location
Group1		
CWR1(GG3)	<i>Vigna radiata</i> var. <i>sublobata</i>	Kakirawa
CWR2(GG8)	<i>V.radiata</i> var. <i>sublobata</i>	Nalanda
CWR3(GG12)	<i>V.trilobata</i>	Puttalm
Group2		
CWR4 (BG3)	<i>Macropetelium</i> sp.	Meegalawa
CWR5(BG4)	<i>Vigna dalzelliana</i>	Peradeniya
CWR6(BG5)	<i>V.radiata</i> var. <i>sublobata</i>	Kekirawa
CWR7(BG7)	<i>V.trilobata</i>	Puttalm
CWR8(BG8)	<i>V.trilobata</i>	<u>Puttalm</u>
CWR9(BG11)	<i>V. dalzelliana</i>	Peradeniya

Discussion

- Rhizobial isolates CWR1, CWR2 and CWR3 (group 1) showed better nodulation on both green gram and black gram plants.
- Although the nodule number was significantly lower with CWR1 at 6th week on both crops, it has significantly increased at 10th week.

(This result has realized that slow growth rate of this isolate in the colonization after the inoculation and later infection has improved the nodule number.)

- All the isolates (CWR1, CWR2 and CWR3) showed high plant dry matter production than that of the N fertilizer application in both crops but it was significantly higher in black gram (Fig 1).

Plant dry matter (group I)

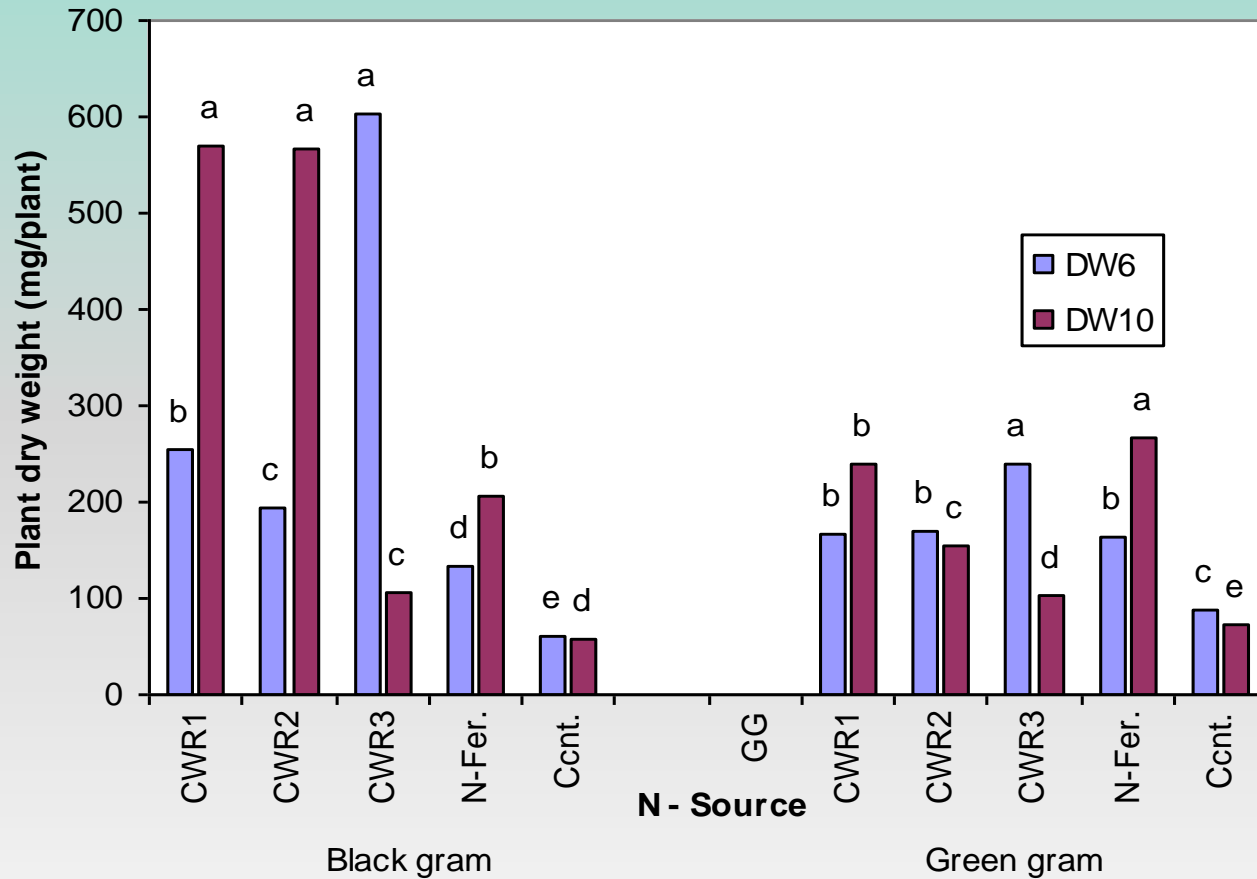


Figure 1. Plant dry matter production of Black gram and Green gram with the inoculation of the isolates of group I, N fertilizer application and the control

- In group 2, nodule formation was observed with the strains CWR7, CWR8 and CWR9.
- Out of these isolates CWR7 showed significantly highest nodulation and increased the plant dry matter production with both Green gram and Black gram at 6 and 10 weeks after the planting (Fig 2).
- According to the results CWR7 is an effective isolate and has the potential to use as rhizobial inoculants both in Green gram and Black gram.

Plant dry matter (group II)

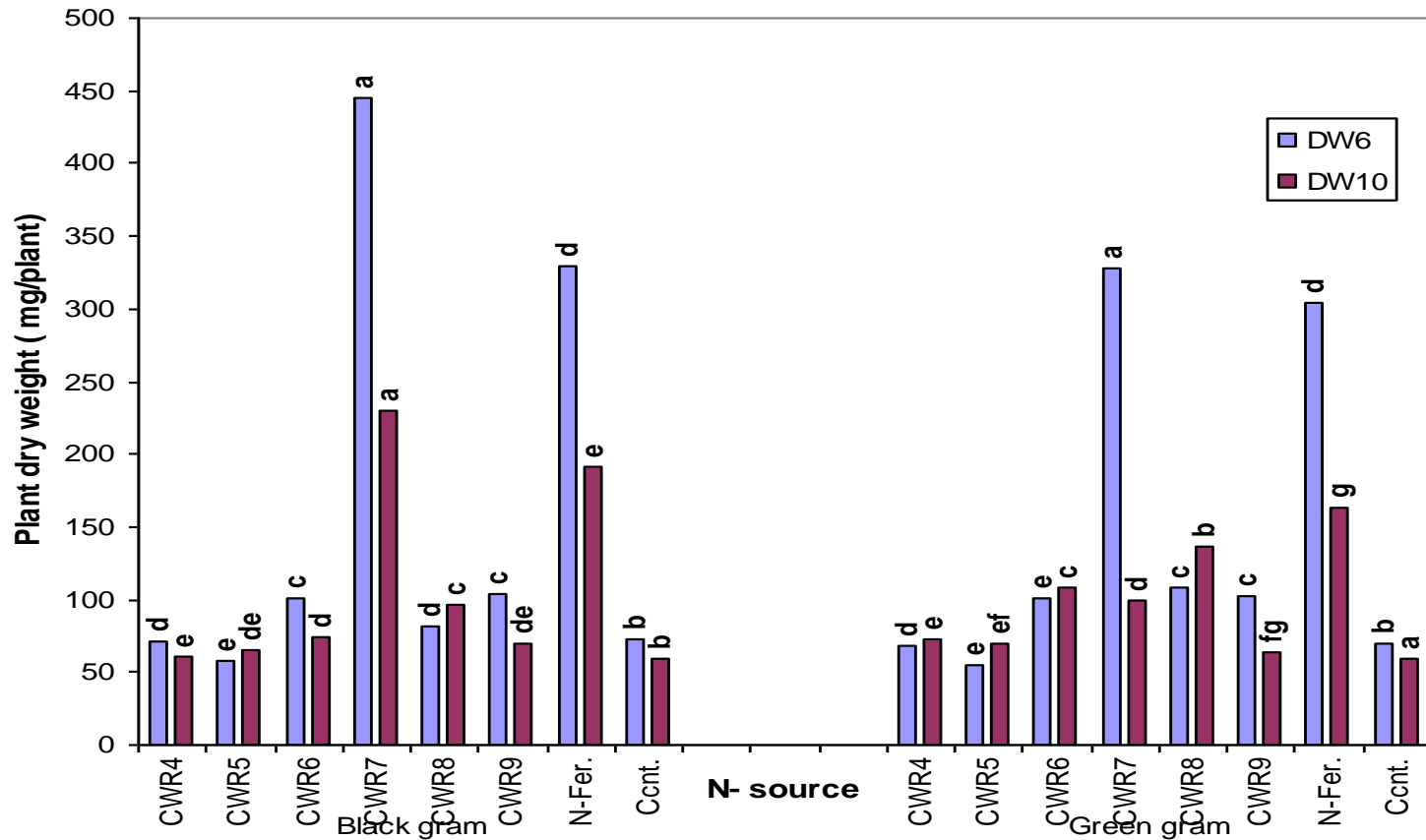
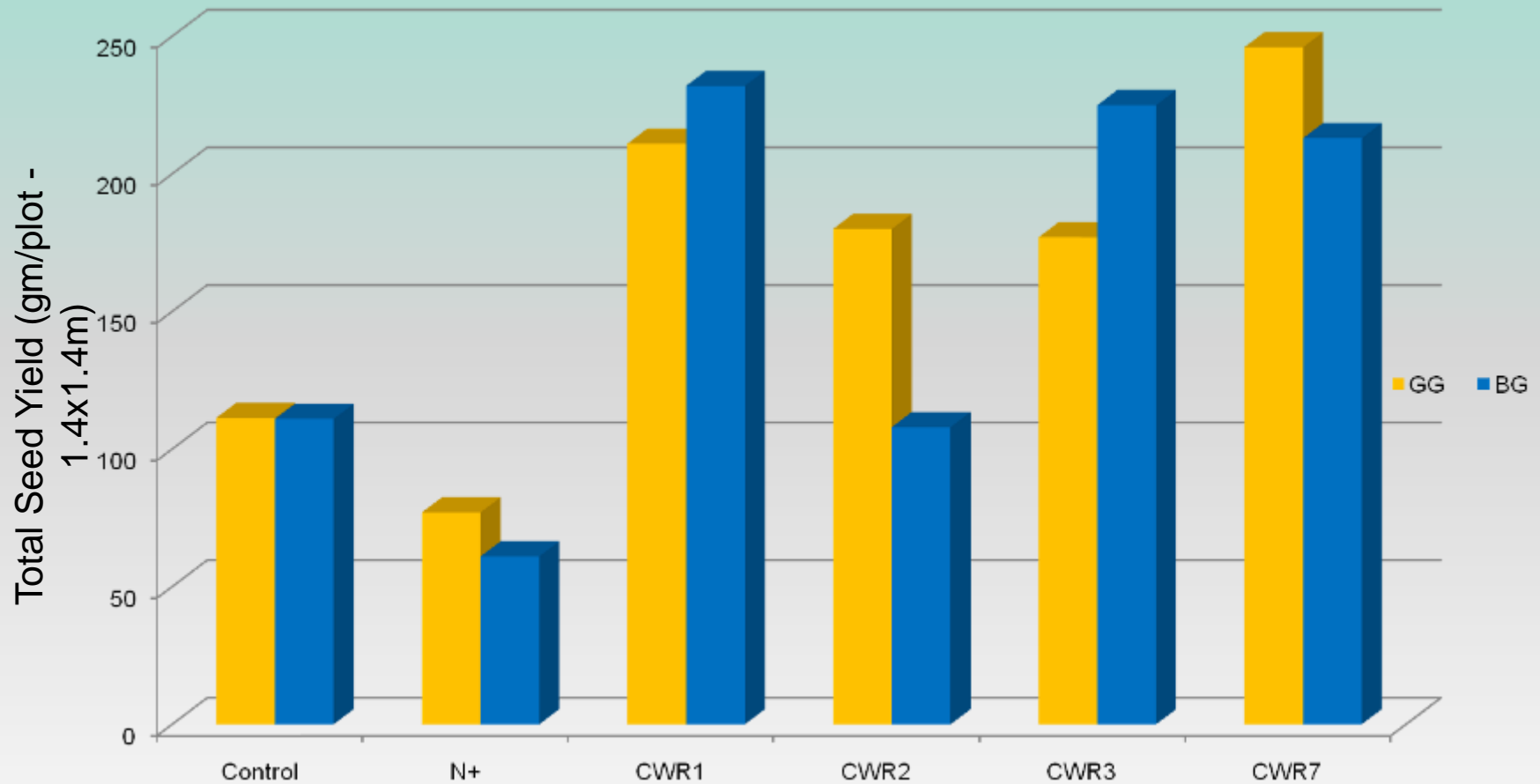


Figure 2. Plant dry matter production of Black gram and Green gram with the inoculation of the isolates of group 2, N fertilizer application and the control

Seed yield comparison in Black and Green gram crops inoculated with CWR rizobial isolates



Conclusion

- The results revealed that the isolates have high infectivity and the effectivity which are important parameters to use as inoculants for crop legumes.
- The results has proven that the *rhizobia* isolated from crop wild relatives of *Vigna* has an ability to form nodule on their cross inoculating group of edible grain legumes such as green gram and black gram.

Conclusion (cont.)

- Isolates CWR1, CWR2, CWR3 and CWR7 are well adapted and successfully overcome the competition among native rhizobia in the experimental conditions.
- Finally, these strains have the potential to replace the recommended N-fertilizer by using inoculants produced by these isolates.
- **Future studies: Adaptive trials at Farmers field in different localities.**

Collaboration



Centre for Rhizobium Studies



HOME

CONFERENCE

CONTACT

Last updated: 1 October 2009

Welcome to the Centre for Rhizobium Studies



The Centre for Rhizobium Studies was formed at Murdoch University in 1997 in response to a decline in expertise in the disciplines of rhizobiology that confronted Australia from the late 1980s. It was particularly relevant at that time when Rural Industry, represented by the GRDC (Grains Research and Development Corporation) and AWI (Australian Wool Innovation), were major voices in the establishment and funding of the CRS. The other substantial partner in the CRS remains the WA Department of Agriculture.

In its first eight years of operation, the CRS has released six strains of root-nodule bacteria to commerce. These strains have been widely sown across southern Australia and these fix nitrogen that forms a substantial portion of this \$2 billion asset. The CRS has been very influential in the improvement of inoculant carrier technologies that deliver these elite strains in good condition to their end users. The CRS is currently strongly involved in selecting and breeding new perennial legumes that are adapted to acidic and infertile soils, as well as developing appropriate rhizobia for them. The CRS has a very strong molecular group that assists in understanding the response of rhizobia to stress, which is very relevant to our agriculture. CRS also facilitates educational workshops for National and International students, such as the Crawford Fund sponsored Master Classes in modern rhizobium technologies.

WELCOME



CENTRE FOR RHIZOBIUM STUDIES

Our vision: To provide a world focus in integrated research and education in the science of root-nodule bacteria.

Dissemination



Acknowledgement

**This Project is fully funded by the CWR
Project**

Thank You!