



DISOMIA UNIPARENTALE:

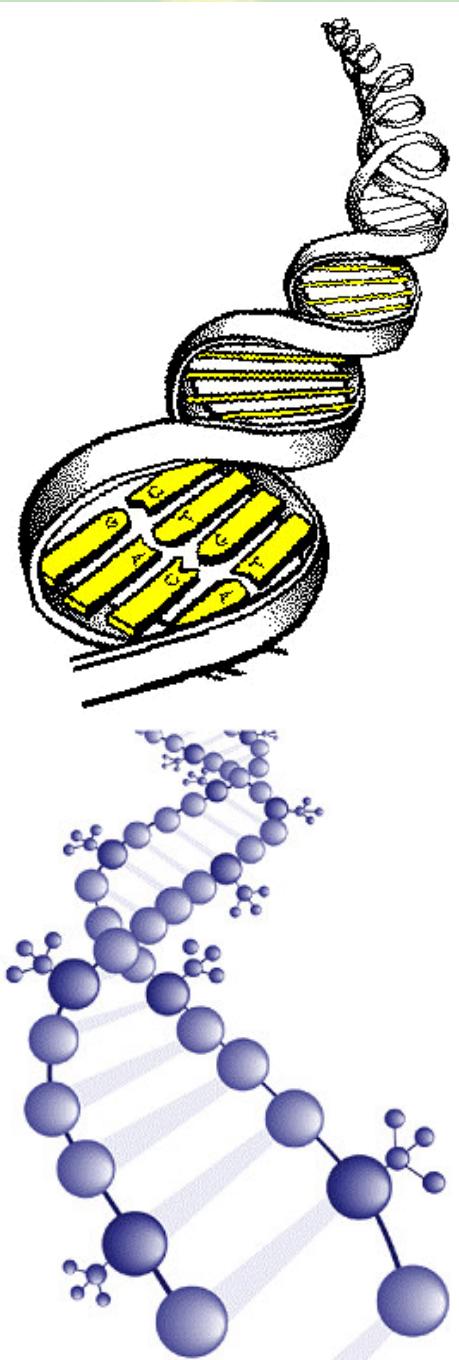
Patogenesi e diagnosi di laboratorio

Francesca Romana GRATI, Ph.D.

TOMA, Advanced Biomedical Assays, S.p.A.

**Controversie e nuove tecnologie nella diagnosi prenatale del primo
trimestre: dal laboratorio alle procedure strumentali**

Bologna, 5 Aprile 2008



INFORMAZIONE DEL DNA



Informazione genetica

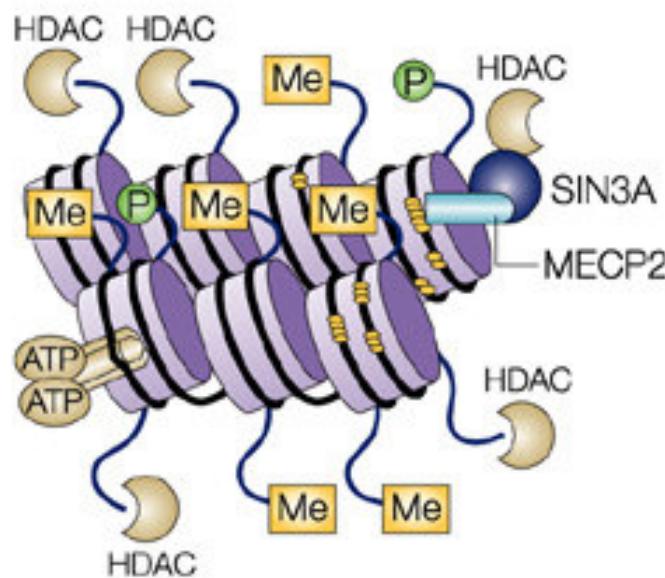
Istruzioni per la sintesi di proteine e RNA.
E’ “STABILE”

Informazione epigenetica

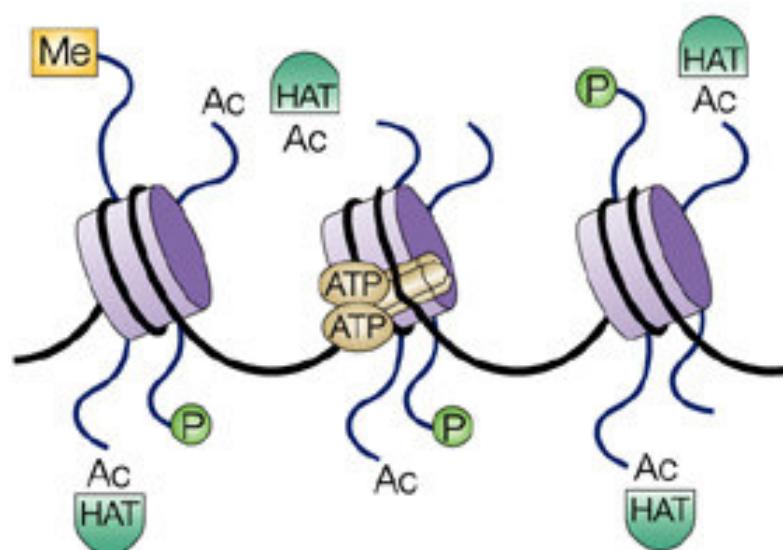
Istruzioni su come, quando e dove deve essere
usata l’informazione genetica.
E’ “DINAMICA”

IMPRINTING

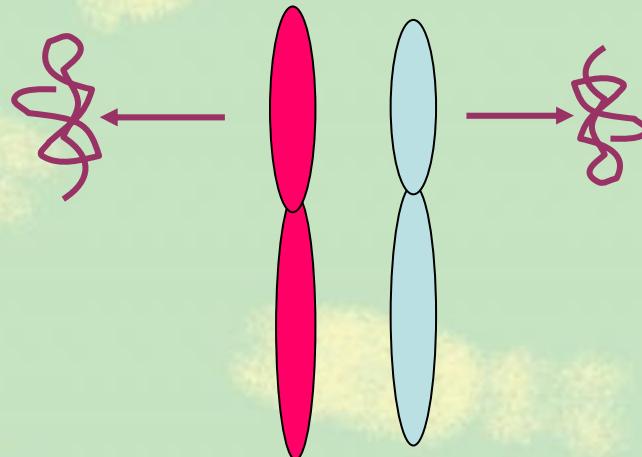
a Closed chromatin: transcriptional repression



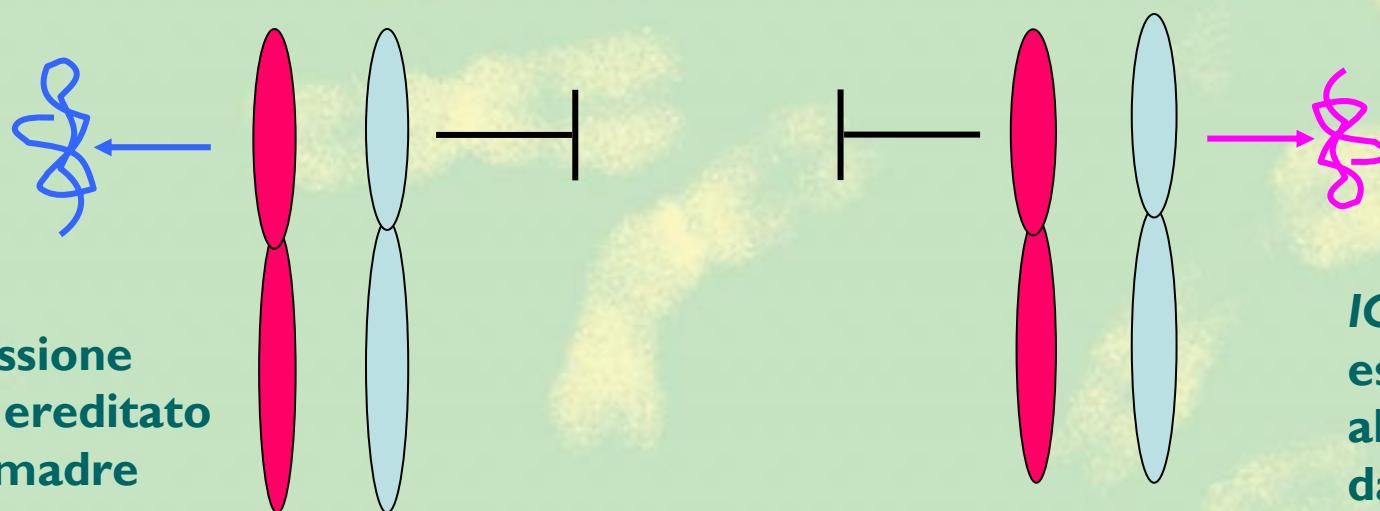
b Open chromatin: transcriptional activation



Gene non “imprinted”(98%): espressione biallelica



Gene “imprinted”(2%): espressione monoallelica



H19:
espressione
allele ereditato
dalla madre

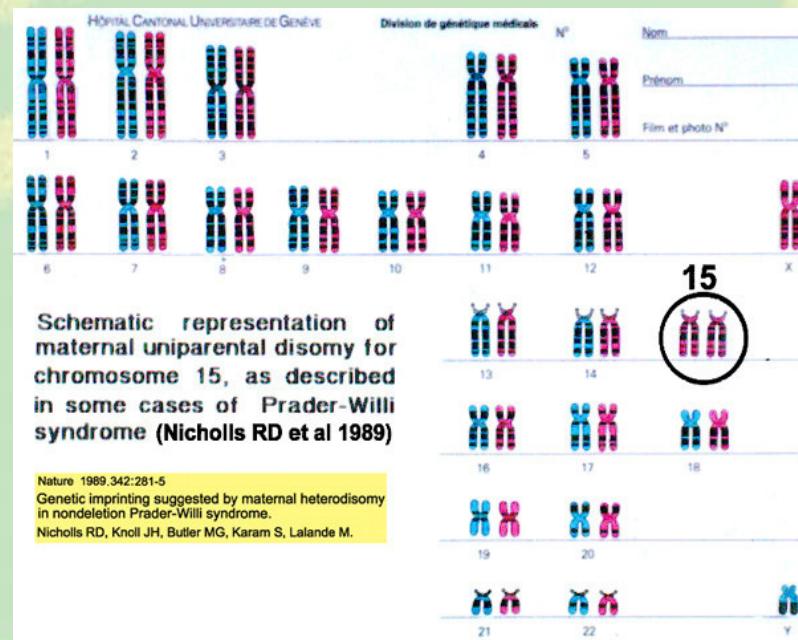
IGF2:
espressione
allele ereditato
dal padre

Uniparental Disomy (UPD)



“A new genetic concept: the uniparental disomy and its potential effect, the isodisomy.” ENGEL E., Am J Med Genet, 1980

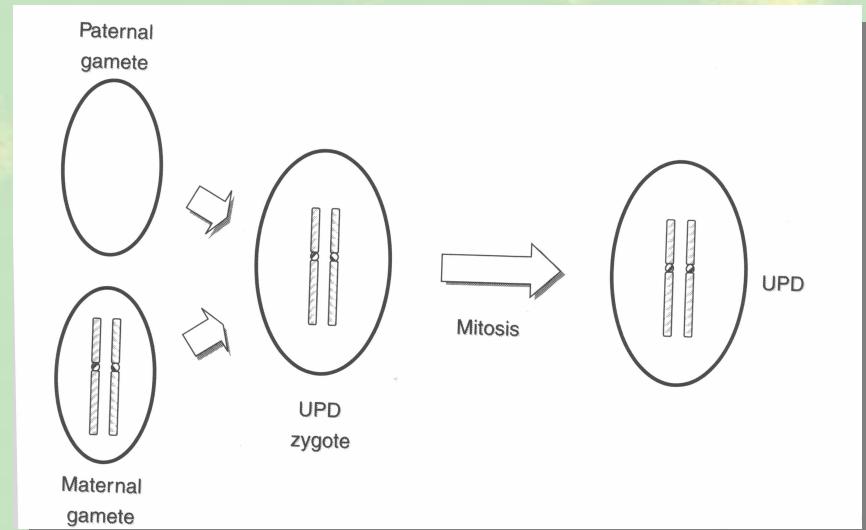
Condizione in cui il numero cromosomico e’ normale, ma una coppia di cromosomi omologhi e’ ereditata da un solo genitore



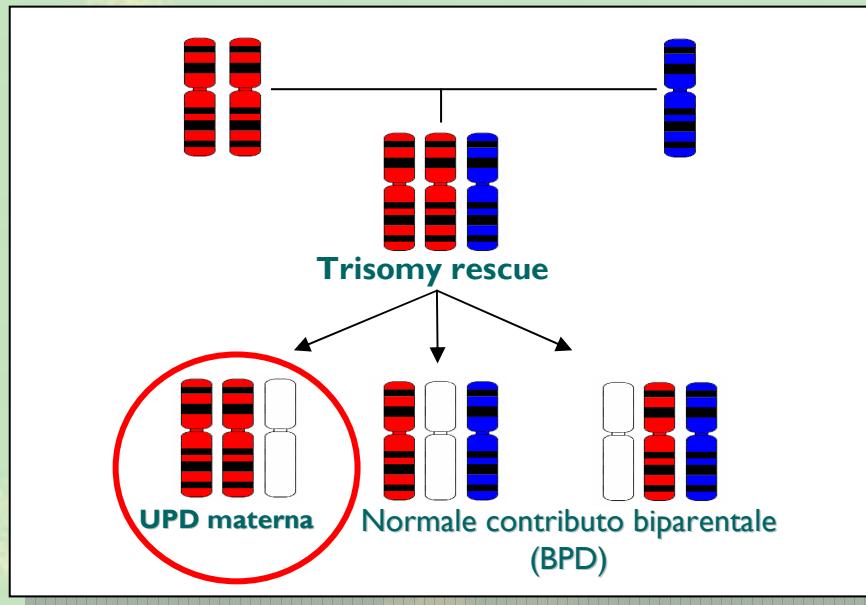
CALCULS DES FREQUENCES POTENTIELLES DE LA DISOMIE UNIPARENTALE

TYPE DE LA DISOMIE (CHROMOSOMES IMPLIQUES)	POUR 20 % D'AVORTEMENTS	POUR 50 % D'AVORTEMENTS
	FREQUENCES / 10.000	FREQUENCES / 10.000
46, (16)	$(0,8 \%)^2 = 0,64$	$(2 \%)^2 = 4,00$
46, (21)	$(0,3 \%)^2 = 0,09$	$(0,75 \%)^2 = 0,56$
46,(15)	$(0,25 \%)^2 = 0,06$	$(0,60 \%)^2 = 0,36$
46,(22)	$(0,25 \%)^2 = 0,06$	$(0,60 \%)^2 = 0,36$
46, (XX OU XY)	$(1,3 \%)^2 = 1,70$	$(3,2 \%)^2 = 10,20$
TOTAL	2.51	15,48

GAMETIC COMPLEMENTATION (fusione di un gamete nullisomico e uno disomico)



UPD and Trisomy rescue



CONFINED PLACENTAL MOSAICISM

G. Simoni and SM Sirchia

Prenatal Diagnosis, 14:1185-1189 (1994)

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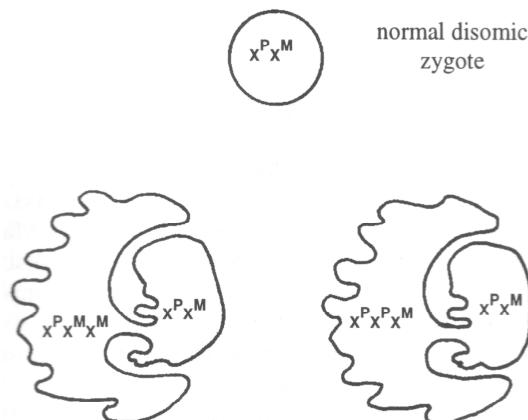


Fig. 1—Possible mechanism by which a confined placental mosaicism could originate from a normal zygote. Chromosomal anaphase lag occurs at a late gestational stage, after fetal and placental compartments have separated. We can assume a different imprinting effect according to the maternal or paternal origin of the supernumerary chromosome. M=Maternal; P=Paternal

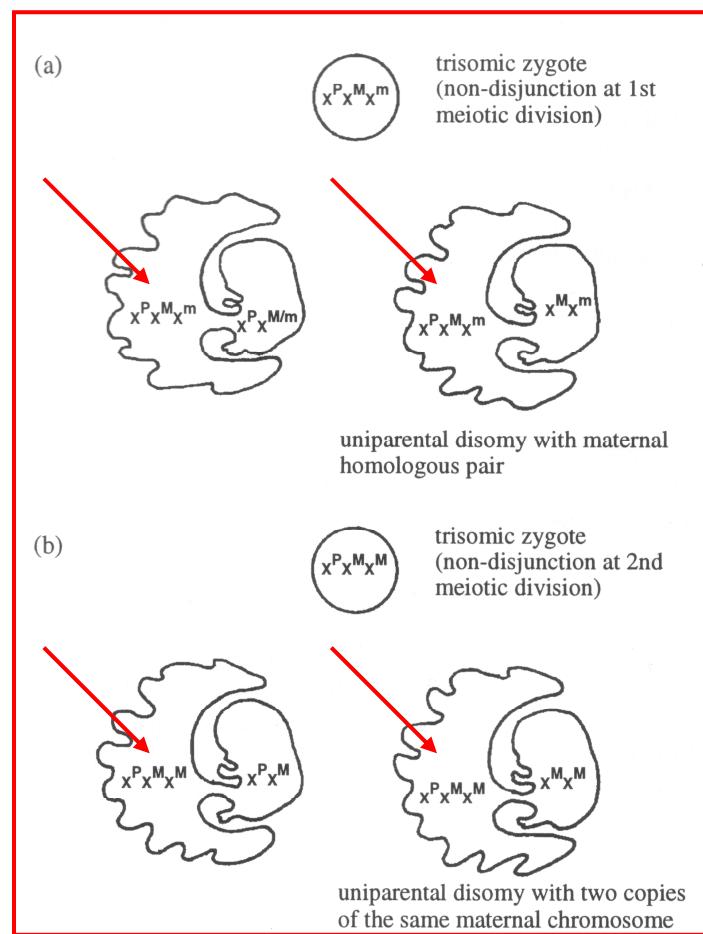


Fig. 2—Possible mechanism by which confined placental mosaicism could originate from a trisomic zygote. Fetal karyotype could be ‘normalized’ by a chromosome loss. We can assume different events would lead to uniparental heterodisomy (a); or isodisomy (b)

ARTICLE

Confirmation of mosaicism and uniparental disomy in amniocytes, after detection of mosaic chromosome abnormalities in chorionic villi

Francesca R Grati^{*1}, Beatrice Grimi¹, Giuditta Frascoli¹, Anna Maria Di Meco¹, Rosaria Liuti¹, Silvia Milani¹, Anna Trotta¹, Francesca Dulcetti¹, Enrico Grosso², Monica Miozzo³, Federico Maggi¹ and Giuseppe Simoni^{1,3}

¹Units of Cytogenetics and Molecular Biology, TOMA Laboratory, Busto Arsizio, Varese, Italy; ²SCDU Genetica Medica, Azienda Ospedaliera San Giovanni Battista, Turin, Italy; ³Cattedra di Genetica Medica, Dipartimento di Medicina, Chirurgia e Odontoiatria, University of Milan, Milan, Italy



UPD: 1 case out of 51 (1.96%)

Table 3 Distribution of specific chromosomal alterations in the different types of mosaisms (CPM and TFM)

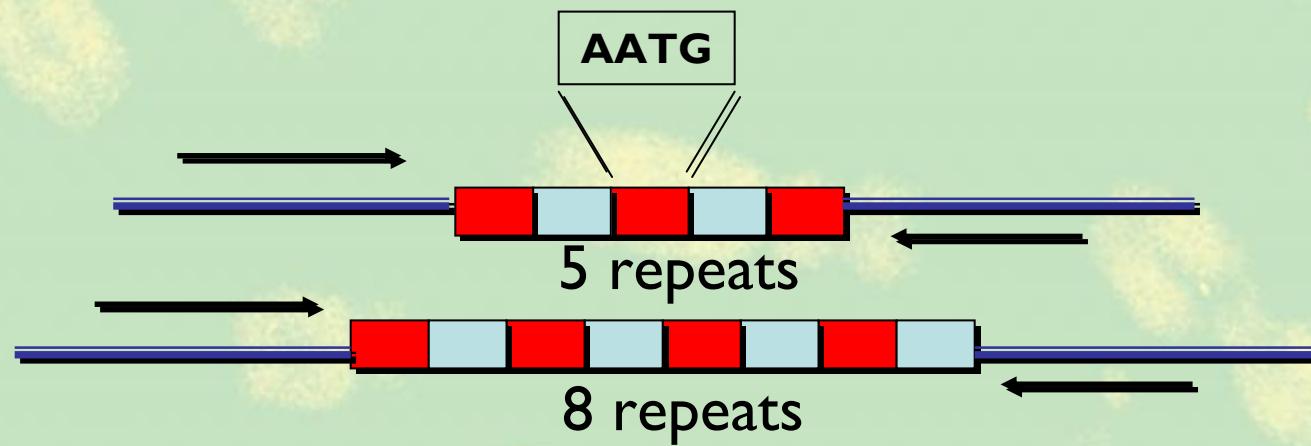
Chr abnormality	CPM number of cases			TFM number of cases			Total (203)
	Type I	Type II	Type III	Type IV	Type V	Type VI	
+1		1					1
+2	1	8+2 [^]					11
+3	6		1				7
+4	1						1
+6		1	1 ^c				2
+7	5+1 [^]	9+1 [^]	1				17
+8	1+2 [^]	1	1				5
+9	2	3					5
+10	1	2					3
+11	2						2
+12	1						1
+13	3	3	1+2 ^c				9
+14	1+1 [*]						2
+15	2+1 [^]	1	1 ^c				5
+16	1+1 [^]	1					3
+17		1					1
+18	4	3		(3 [^])	(1 ^M)		11
+20	1	2	2 ^c				5
+21	1	6+1 [^]		1+(2 [^])	(1 ^c)		12
+22			1 ^{CM}				1
-22		1					1
+X	3 XY+1 XX+1 [^] XX	1 XY		1 XY	1 XX+(1 ^M XY)		9
+Y					1 XY		1
-X	6+2 [^]	2		2+1 [^]	1 ^c		14
-Y	2	3	1	1 [^]	1+(1 ^M)		9
X/XX/XXX					2+1 ^M		3
47,+mar	4+1 [^]	2+1 [^]		1 [^] +1	1		11
46,rearr	7+5 [^]	14+4 [^]			(1 [^])		31
Tetraploidy	4+4 [^]	2+2 [^]	2 ^c				14
Multiple trisomy	2	4					6

* = UPD case.

() = nonmosaic abnormal cell line in the fetus.

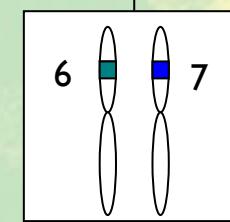
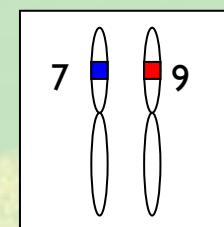
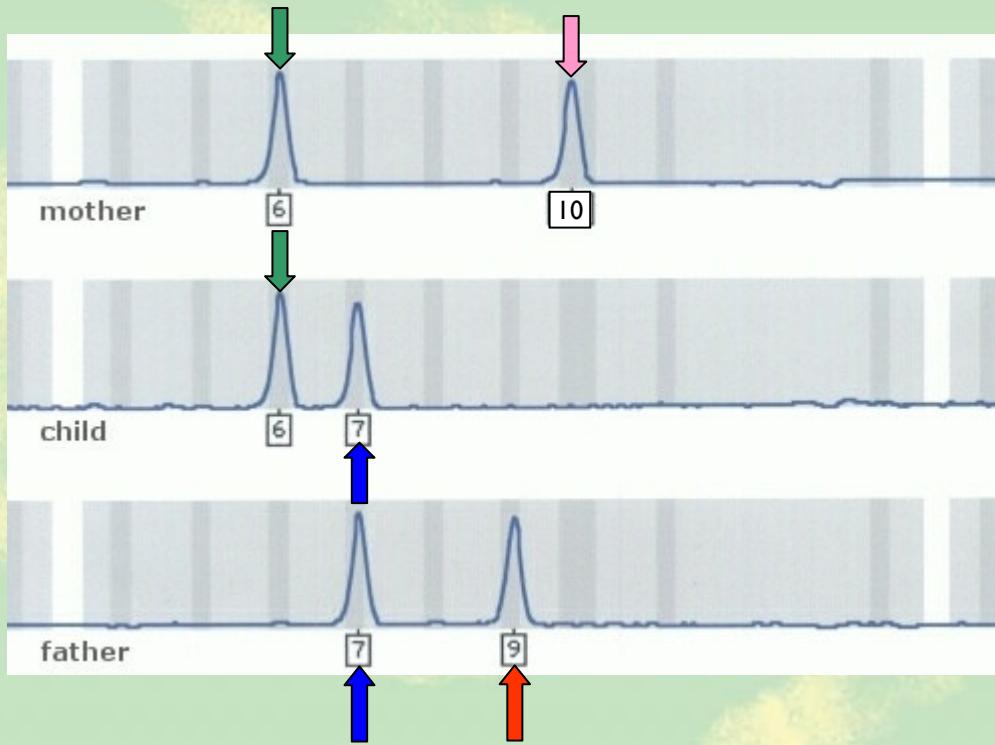
[^] = nonmosaic abnormal cell line in placenta (in cytotrophoblast (^c) or in mesenchyme (^M)).

XY/XX = normal karyotype associated with the +X/+Y cell line.

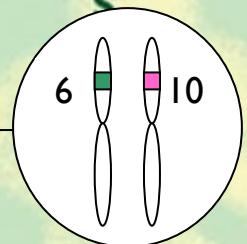


Heterozygote = alleles differ and can be resolved from one another

Homozygote = both alleles are the same length



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STR	Map Localization	M	F	P	Result
D14S122*	q11.2	1,2	1,1	3,4	ISOUPD
D14S283*	q11.2	2,3	2,2	1,1	ISOUPD
D14S990*	q11.2	2,3	3,3	1,2	ISOUPD
D14S52	q21	1,1	1,1	2,3	UPD
D14S58	q21	1,1	1,1	2,2	UPD
D14S258	q23-q23.3	1,1	1,1	2,2	UPD
D14S77	q23-q24	2,3	2,3	1,3	NI
D14S43	q24.3	2,3	2,3	1,3	NI
D14S68	q24.3	2,4	2,4	1,3	HETEROUPD
D14S59*	q24.3	1,1	1,1	2,3	UPD
D14S983	q24.3	1,1	1,1	1,1	NI
D14S53	q24.3	1,1	1,1	1,2	NI
D14S42	q24.3	1,2	1,2	1,2	NI
D14S81	q31-q32	1,1	1,1	1,2	NI
D14S61	q32.2-q32.3	1,2	1,2	2,2	NI

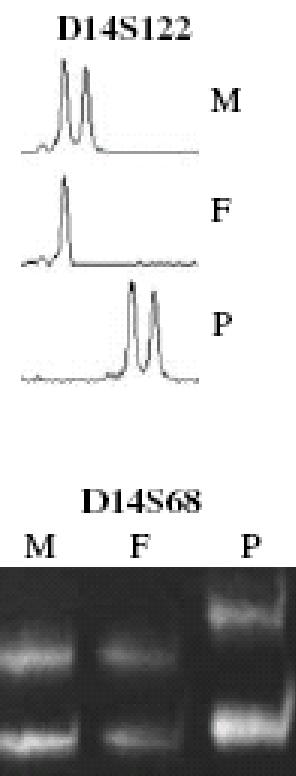


Figure 1 Molecular results of matUPD14 case. For each informative marker, maternal (M), fetal (F) and paternal (P) alleles are indicated. The fetus inherited alleles only from the mother and failed to inherit a paternal allele, consistent with a maternal disomy 14. * Indicate STRs analysed by fluorescent capillary system.

UPD and Robertsonian translocations

PRENATAL DIAGNOSIS

Prenat Diagn 2004; 24: 997–1000.

Published online in Wiley InterScience (www.interscience.wiley.com). DOI: 10.1002/pd.961

Prenatal search for UPD 14 and UPD 15 in 83 cases of familial and *de novo* heterologous Robertsonian translocations

Anna Ruggeri^{1,*}, Francesca Dulcetti¹, Monica Miozzo², Francesca R. Grati², Beatrice Grimi¹, Silvano Bellato³, Federica Natacci⁴, Federico Maggi¹ and Giuseppe Simoni^{1,2}

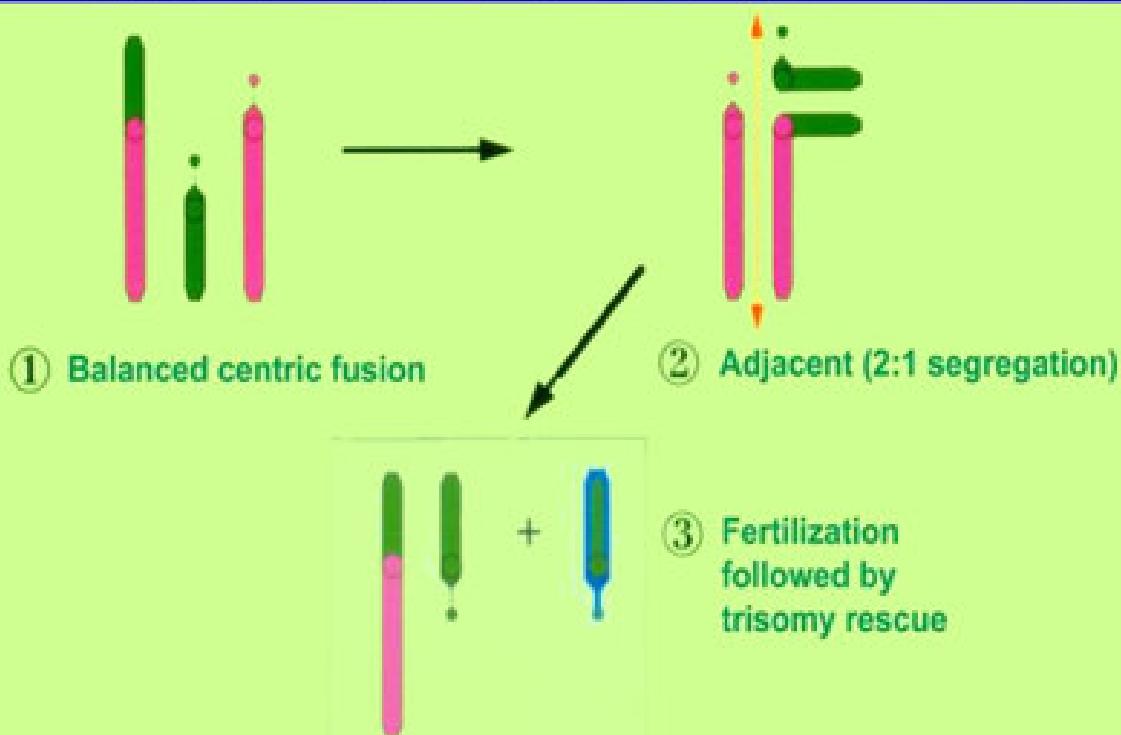
¹Units of Cytogenetics and Molecular Biology, TOMA Laboratory, Busto Arsizio, Varese, Italy

²Laboratory of Medical Genetics, Department of Medicine, Surgery and Dentistry, San Paolo School of Medicine, University of Milan, Milan, Italy

³Division of Pediatrics, L. Cazzavillan Hospital, Arzignano, Vicenza, Italy

⁴Servizio di Genetica Medica, Istituti Clinici di Perfezionamento, Milano, Italy

UPD favored by balanced Robertsonian translocation 2:1 segregation followed by trisomy rescue



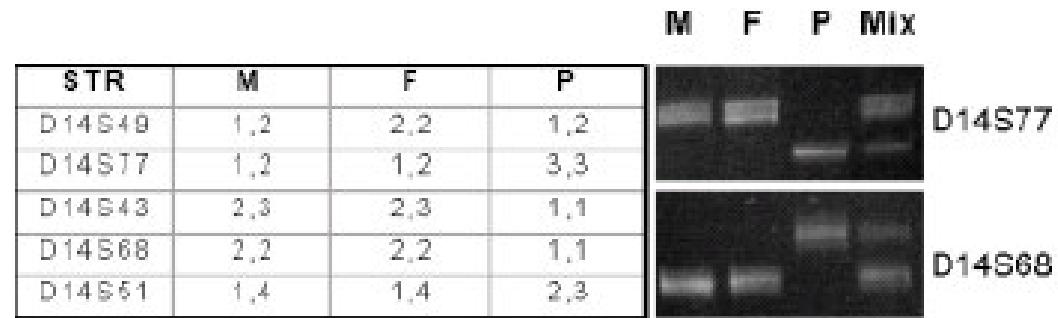


Figure 1—Molecular results of matUPD14 case. For each informative marker, maternal (M), fetal (F) and paternal (P) alleles are indicated (Mix: sample containing M and P alleles). The fetus inherited alleles only from the mother and failed to inherit a paternal allele, consistent with a maternal UPD 14



ROBs	n° cases
13;14	**224
13;15	14
14;15	24
14;21	*42
14;22	13
15;21	3
15;22	5
Subtotal	425 (0.7%)
NotDefined	181
TOT	506 (0.6%)

Papenhausen et al, 1996

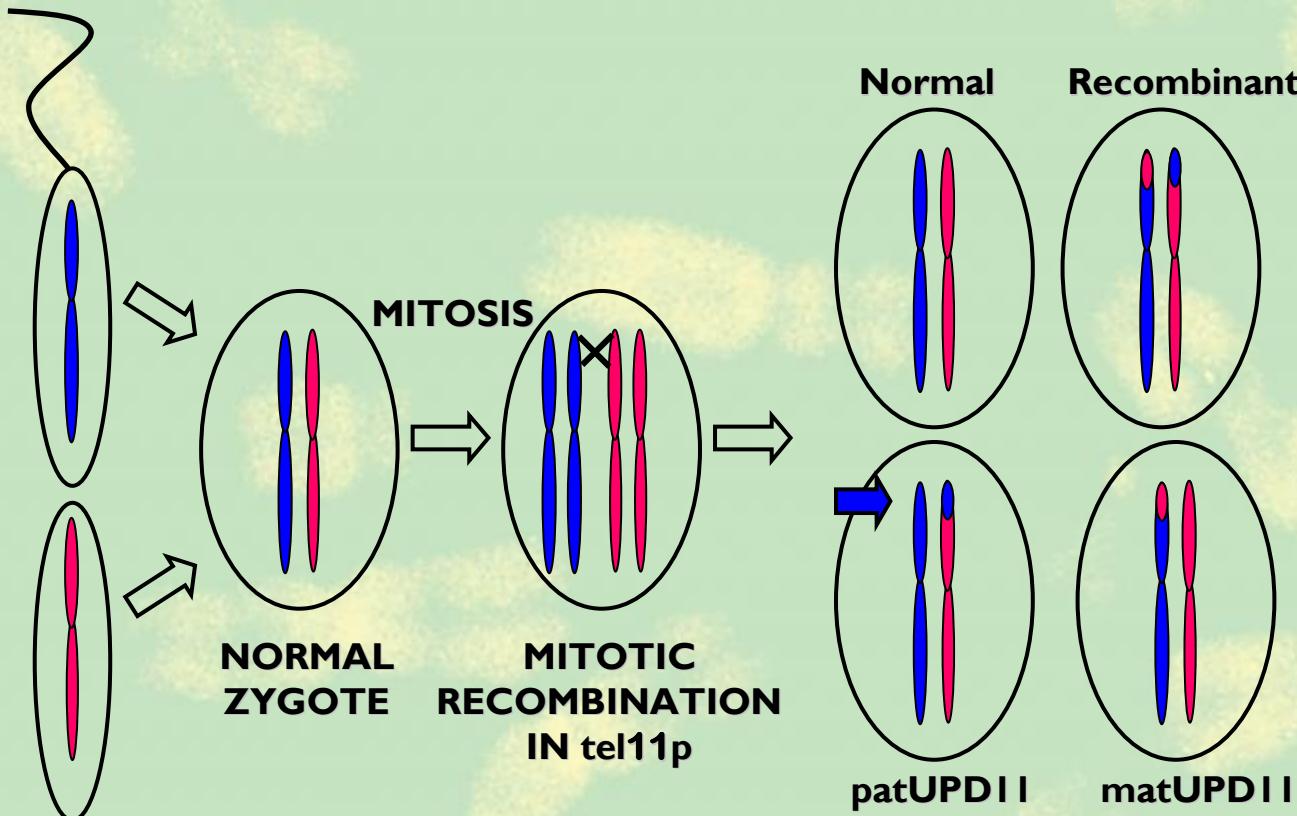
Silverstein et al, 2002

Ruggeri et al, 2004

*=UPD14 *de novo*

patUPD11 and mitotic crossing-over

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A MOSAICO!



CORRELAZIONE GENOTIPO - FENOTIPO

ONFALOCELE:

- MUTAZIONI INATTIVANTI/IPERMETILAZIONE DI CDKNIC
- MAI DESCRITTO IN ASSOCIAZIONE CON patUPD11

J Med Genet 2000;37:921–926

Epigenotype-phenotype correlations in Beckwith-Wiedemann syndrome

Jacqueline R Engel, Alan Smallwood, Antonita Harper, Michael J Higgins, Mitsuo Oshimura, Wolf Reik, Paul N Schofield, Eamonn R Maher

Am. J. Hum. Genet. 70:604–611, 2002

Epigenetic Alterations of H19 and LIT1 Distinguish Patients with Beckwith-Wiedemann Syndrome with Cancer and Birth Defects

Michael R. DeBaun,^{1,2} Emily L. Niemitz,^{3,4} D. Elizabeth McNeil,² Sheri A. Brandenburg,³ Maxwell P. Lee,³ and Andrew P. Feinberg^{3,5}

¹Division of Pediatric Hematology-Oncology, Department of Pediatrics, Washington University School of Medicine, Saint Louis; ²Genetic Epidemiology Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD; and ³Institute of Genetic Medicine, ⁴Predoctoral Program in Human Genetics, and ⁵Departments of Medicine, Oncology, and Molecular Biology and Genetics, Johns Hopkins University School of Medicine, Baltimore

ORIGINAL ARTICLE

Chromosome 11 segmental paternal isodisomy in amniocytes from two fetuses with omphalocele: new highlights on phenotype–genotype correlations in Beckwith–Wiedemann syndrome

F R Grati, L Turolla, P D'Ajello, A Ruggeri, M Miozzo, G Bracalente, D Baldo, L Laurino, R Boldorini, E Frate, N Surico, L Larizza, F Maggi, G Simoni

J Med Genet 2007;44:257–263. doi: 10.1136/jmg.2006.046854

RICERCA DELLA LINEA CELLULARE CON CONTRIBUTO BIPARENTALE NEI TESSUTI FETALI

M: MADRE
 AF: AMNIOCITI
 CNS: SISTEMA NERVOSO CENTRALE
 Li/C/K: FEGATO, CORDONE OMBELICALE, RENE
 H/Lu: CUORE, POLMONE
 T: TIMO
 P: PADRE

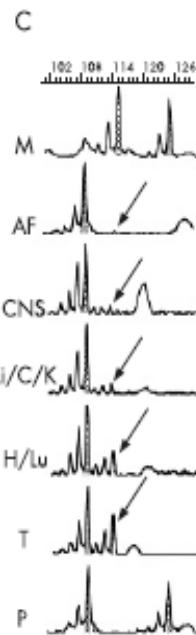


Figure 4 Schematic representation of short tandem repeat (STR) genotyping of case 1. (A) Allelic segregation of the tested polymorphic markers. (B) Electropherograms of D11S1923 and D11S995 STRs included in the uniparental region and of D11S911 mapped in 11q13 showing a normal biparental contribution. (C) Allelic segregation of D11S988 and quantification of the paternal uniparental disomy (pUPD) cell line in different fetal tissues and in alleles from amniotic fluid (AF). The biparental component is represented by the maternal allele (arrow). C, umbilical cord; CNS, central nervous system; H, heart; IsoUPDpat, paternal isodisomy; K, kidney; Li, liver; Lu, lung; M/P, maternal and paternal alleles; NI, not informative; T, thymus.

M: MADRE
 AF: AMNIOCITI
 S/To/C: SURRENE, LINGUA, CORDONE
 B/Pa: INTESTINO, PANCREAS
 U/Lu/C/H: UTERO, POLMONE, CORDONE, CUORE
 CNS: SISTEMA NERVOSO CENTRALE
 P: PADRE

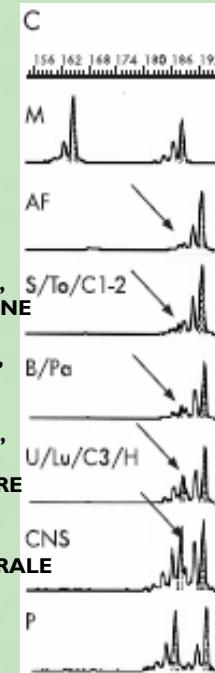


Figure 5 Schematic representation of case 2 short tandem repeat (STR) analysis. (A) Allelic segregation of the tested polymorphic markers. (B) Electropherograms of D11S1923 and D11S986 STRs included in the uniparental region and of D11S4191 mapped in 11q12 showing a normal biparental contribution. (C) Allelic segregation of D11S2071 and quantification of the biparental cell line in different fetal tissues and in alleles from amniotic fluid (AF). The paternal uniparental disomy (pUPD) component is represented by the maternal allele (arrow). B, bowel; C, umbilical cord; CNS, central nervous system; H, heart; isoUPDpat, paternal isodisomy; K, kidney; Lu, lung; M/P, maternal and paternal alleles; NI, not informative; Pa, pancreas; S, adrenal gland; To, tongue; U, uterus.

CONCLUSIONS:

1. the distribution of pUPD cell line is super imposable to that of the organs mainly involved in the omphalocele; the presence at high level of the paternal isodisomic cell line in the abdominal organs could have mimicked the effect of CDKN1C/p57 inactivating mutations or hypermethylation causing the omphalocele.
2. **amniotic fluid cell cultures is the preferential prenatal sample for BWSUPDII testing: during the AF culture the pUPDII cell line might have a proliferative advantage due to the autocrine effect of IGF2 getting the upper hand over the normal biparental cell line, leading to the unmasking of the UPD cell line.**

UPD TYPE	SYNDROME	PHENOTYPE
<i>Certain</i>		
Pat 6	Transient Neonatal Diabetes Mellitus (TNMD)(MIM:601410)	IUGR, diabete neonatale
Mat 7/Mat 11	Silver Russell Syndrome (SRS) (MIM:180860) Beckwith-Wiedemann Syndrome (BWS) (MIM:130650)	IUGR/PNGR, dismorfismi Overgrowth, dismorfismi, tumori
Pat 11		
Mat 14		IUGR, dismorfismi
Pat 14		Nanismo, dismorfismi
Mat 15	Prader Willi Syndrome (PWS) (MIM:176270)	Obesità, dimorfismi,
Mat/Pat 20	Pre- and postnatal growth retardation, dysmorphisms	IUGR/PNGR
<i>Probable</i>		
Mat 2	Growth failure, bronchopulmonary dysplasia	IUGR e displasia broncopolmonare
Mat 16	Growth failure, abnormalities	IUGR e dismorfismi

TOMA LAB

**Prof. Giuseppe Simoni
dott. Federico Maggi**

CYTOGENETICS

dott.ssa Beatrice Grimi
dott.ssa Giuditta Frascoli
dott.ssa Anna Maria Di Meco
dott.ssa Rosaria Liuti
dott.ssa Silvia Milani
dott.ssa Anna Trotta



R&D + BIOLOGIA MOLECOLARE



Simona De Toffol

**Francesca Dulcetti
Barbara Malvestiti**

Anna Ruggeri

Chromosome 1

ARH1/NOEY2
p73

Chromosome 5

U2AF1RS1

Chromosome 6

AIR

HYMA1

M6P/IGF2R

SLC22A2

SLC22A3

ZAC/PLAGL1

Chromosome 7

ASB4

COPG2

GRB10

MIT1/LB9

PEG1-AS

PEG1/MEST

PEG10

SGCE

Chromosome 11

- > ASCL2/HASH2
- > AWT1
- > BWRT/KCNQ1DN
- > CD81
- > H19
- > IGF2
- > IGF2-AS
- > INS
- > INS2
- > KCNQ1
- > KCNQ1QT1
- > MSUIT1
- > NAPIL4
- > OBPH1
- > SDHD
- > SLC22A1L/ITM
- > TRPM5
- > TSSC3/IPL
- > TSSC4
- > WT1
- > ZNF215
- > p57kip2/CDKN1C

Chromosome 12

ATA3
DCN

Chromosome 13

HTR2A

Chromosome 14

DIO3
DLK1/PREF-1
MEG3/GTL2
MEG8/snoRNAs
PEG11

Chromosome 18

ELONGIN A3
IMPACT

Chromosome 19

PEG3/ZIM2
USP29
ZIM1
ZIM3
ZNF264

Chromosome 20

GNAS1
GNAS1-AS
L3MBTL
NNAT

Chromosome 15

ATP10C
GABRB3
HBII-13
HBII-52
IPW
MAGEL2
MKRN3
NDN
PAR-SN
PAR1
PAR5
PEG12
PWCR1
RASGRF1
SNRPN
UBE3A
UBE3A-AS
ZNF127-AS

Chromosome X

XIST

Am. J. hum. Genet. 42: 215-216, 1988

**Editorial : Uniparental Disomy: A Rare Consequence of the High Rate of
Aneuploidy in Human Gametes**

Dorothy Warburton

Department of Genetics and development of Pediatrics, Columbia University. New York

Am. J. hum. Genet. 42: 217-225, 1988

Uniparental Disomy as Mechanism for Human Genetic Disease

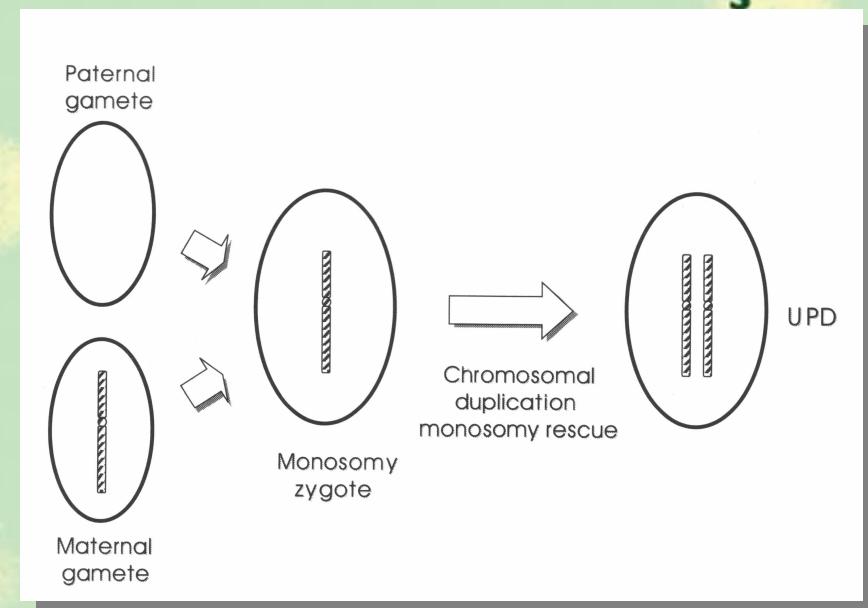
**J. Edward Spence, Ronald G. Percicante, Guillian M. Greig. Huntington F.
Willard. David H. Ledbetter. J. Fielding Heitmancik, Marilyn S. Pollack, William
E. O'Brien and Arthur L. Beaudet**

Howard Hughes Medical Institute, Institute of Molecular Genetics and Department of Microbiology and Immunology,
Baylor College of Medicine, Houston: Mercy Hospital, Watertown, NY: and Department of Medical Genetics,
University of Toronto, Toronto

The first thoroughly analyzed and described case of UPD. It was one involving maternal chromosome 7, responsible for cystic fibrosis in an unusually short girl who carried Gly542Ter mutation in her CFTR gene.

This article, of Beaudet's lab, with Ledbetter among the Authors and Spence as the Senior Author, was not only featuring the first case ever sighted of non-traditional recessive inheritance through reduction to homozygosity of the recessive mutant only carried by one of the two parents. It also offered a most comprehensive review of the possible mechanisms leading to the occurrence of UDP.

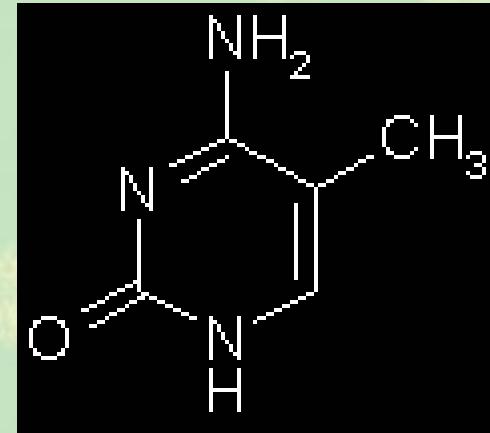
MONOSOMY RESCUE (fusione di un gamete nullisomico e uno normale monosomico seguita da duplicazione del cromosoma monosomico)



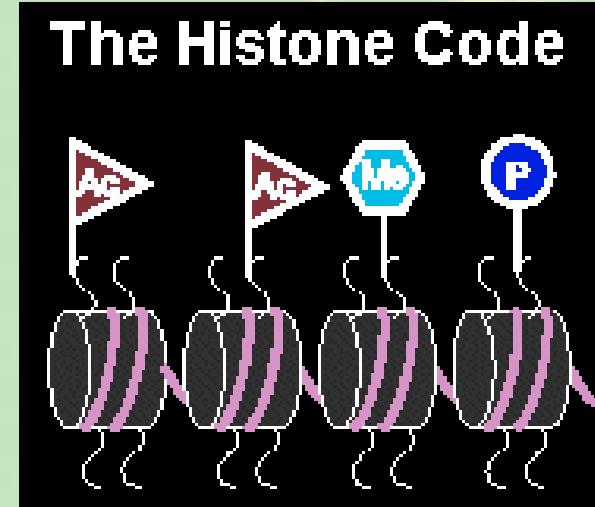
MODIFICAZIONI CHIMICHE DELLA CROMATINA



- Metilazione della citosina



- Metilazione, acetilazione, fosforilazione proteine istoniche





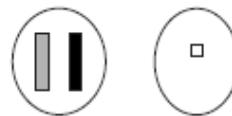
UPD and supernumerary marker chromosomes

Parents



MEIOTIC ORIGIN

Gametes



Zygote



Somatic cells



MITOTIC ORIGIN

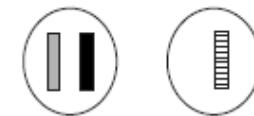


Figure 5. Formation of UPD associated with an additional marker chromosome. (A) The marker chromosome was formed in meiosis and a gamete with the marker chromosome but no normal homologue was fertilised by a normal gamete. (B) A disomic gamete is fertilised by a gamete with a marker chromosome formed in meiosis. (C) A disomic gamete is fertilised by a normal gamete and subsequent mitotic formation of the marker chromosome. In (B) and (C) not only heterodisomy as shown but also isodisomy is possible.

IUGR, diagnosi prenatale (età materna avanzata): 46,XX/45,X



RISULTATO

Cariotipo fetale: mos45,X/46,XX

OSSERVAZIONI

L'analisi è stata eseguita su 34 piastre metafasiche derivate da 18 colonie di 3 colture. E' presente una condizione di mosaismo con due linee cellulari: una con monosomia X (8 metafasi da 4 colonie di 3 colture) e l'altra con corredo cromosomico 46,XX (26 metafasi da 14 colonie di 3 colture). Si consiglia il colloquio con il genetista.

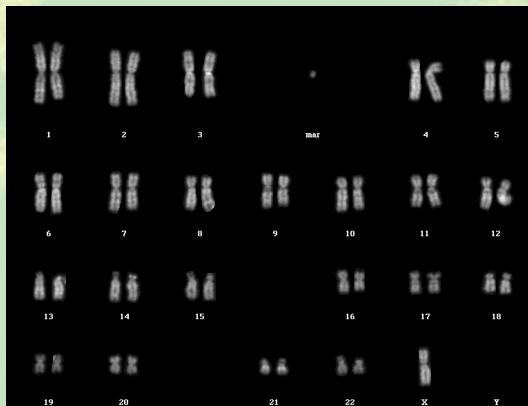
PNGR severo, fronte ampia e bombata (sospetta SRS)

RISULTATO Cariotipo: mos46,X,+mar[25]/45,X[7]/46,XX[68]

OSSERVAZIONI

L'analisi è stata eseguita su 100 metafasi con risoluzione di 450 bande per set apoide. E' presente una condizione di mosaismo con tre linee cellulari: una a 46 cromosomi con monosomia X e con presenza di un piccolo marcatore puntiforme (25%), una a 45 cromosomi con monosomia X (7%) e una con corredo cromosomico 46,XX (68%). L'applicazione della tecnica FISH (vedi referto allegato) ha mostrato segnale positivo del marcitore all'ibridazione con sonda centromerica specifica per il cromosoma 7. L'analisi molecolare (vedi referto allegato) ha permesso di identificare la presenza della disomia uniparentale materna del cromosoma 7. Si consiglia il colloquio con il genetista.

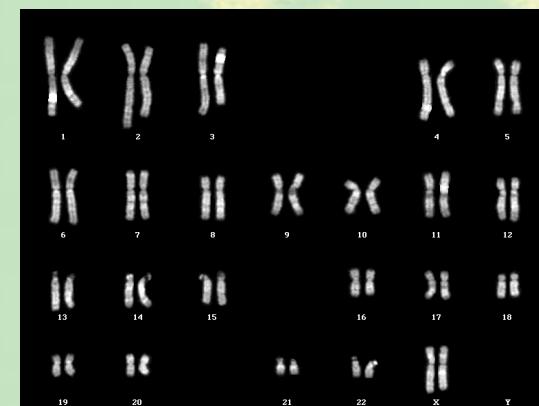
46,X,+mar (25%)

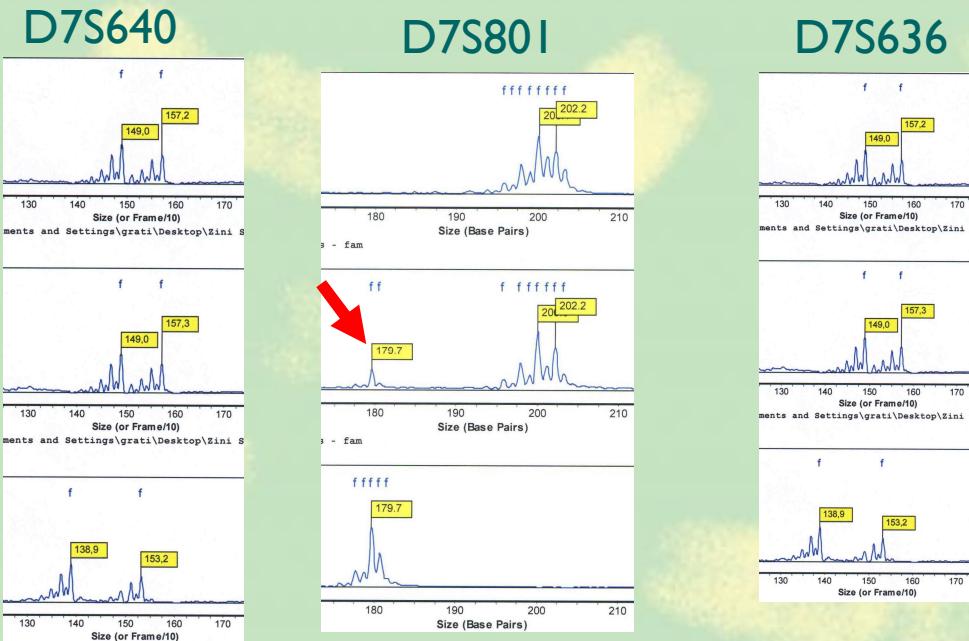


45,X (7%)



46,XX (68%)





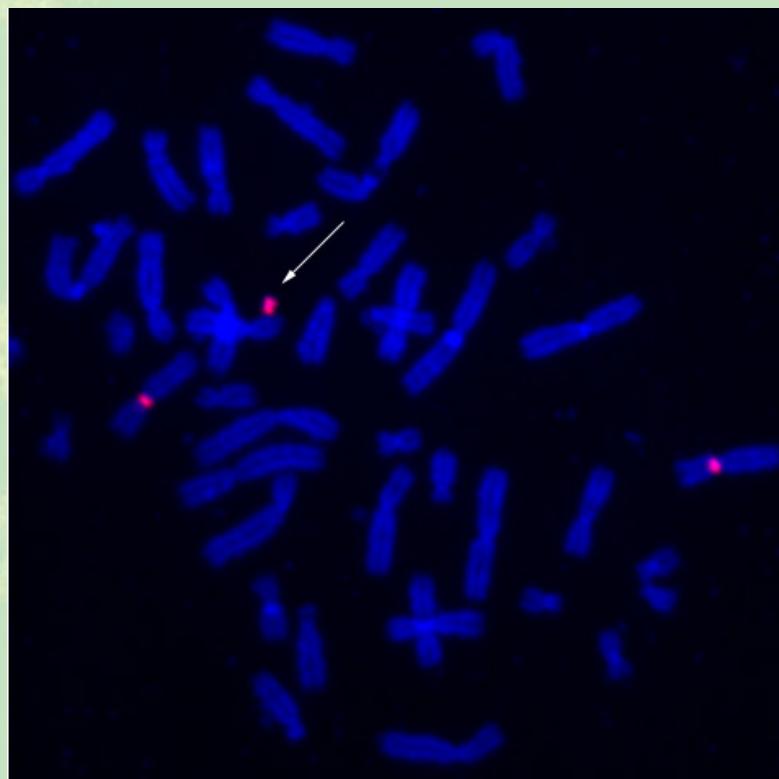
LOCI POLIMORFICI ANALIZZATI	ALLELI			RISULTATO
	MADRE	FIGLIA	PADRE	
D7S3069	1.1	1.1	2.3	UPD MAT
D7S801	2.3	(1).2.3	1.1	ETERO UPD MAT+ LINEA TRISOMICA CON CR7 PAT
D7S663	1.2	1.2	1.3	ESCLUSIONE UPD PAT
D7S480	1.3	1.3	2.4	ETERO UPD MAT
D7S640	1.4	1.4	2.3	ETERO UPD MAT
D7S636	2.4	2.4	1.3	ETERO UPD MAT
D7S483	1.2	1.2	2.2	ESCLUSIONE UPD PAT
NOS3	1.2	1.2	1.3	ESCLUSIONE UPD PAT

Risultato: L'analisi di segregazione allelica di 8 marcatori polimorfici localizzati lungo il cromosoma 7 ha evidenziato nella probanda una eterodisomia pura di entrambi i bracci; a livello del centromero (D7S801) è presente una condizione di mosaicismo tra la linea cellulare con eterodisomia materna e quella con la trisomia.

Il risultato ottenuto indica che è avvenuto un errore di non-disgiunzione alla meiosi I materna con la formazione di uno zigote trisomico per il cromosoma 7. Successivamente è avvenuta una ricombinazione mitotica con la perdita di entrambi i bracci dell'omologo di origine paterna; tale evento ha generato una condizione di UPD materna a livello di queste sequenze e mosaicismo per le regioni centromeriche.

MATERIALE:	sangue periferico
METODO:	IBRIDAZIONE IN SITU A FLUORESCENZA
SONDA UTILIZZATA:	Sonda alfa satellite specifica per i centromeri dei cromosomi X/Y. Sonda alfa satellite specifica per il centromero del cromosoma 7.
COMMENTO:	L'applicazione della tecnica FISH con l'utilizzo delle sonde sopra indicate, ha permesso di caratterizzare il marcitore come un derivativo del cromosoma 7 (vedere foto allegata: la freccia indica il cromosoma marcatore). Si consiglia il colloquio con il genetista.

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Prot.: BM 14515

Analisi: FISH eseguita con sonda alfa satellite specifica per il centromero del cromosoma 7.