



The island rule explains consistent patterns of body size evolution in terrestrial vertebrates

Ana Benítez-López^{1,2}✉, Luca Santini^{1,3,4}, Juan Gallego-Zamorano¹, Borja Milá⁵,
Patrick Walkden⁶, Mark A. J. Huijbregts^{1,7} and Joseph A. Tobias^{6,7}

Island faunas can be characterized by gigantism in small animals and dwarfism in large animals, but the extent to which this so-called ‘island rule’ provides a general explanation for evolutionary trajectories on islands remains contentious. Here we use a phylogenetic meta-analysis to assess patterns and drivers of body size evolution across a global sample of paired island-mainland populations of terrestrial vertebrates. We show that ‘island rule’ effects are widespread in mammals, birds and reptiles, but less evident in amphibians, which mostly tend towards gigantism. We also found that the magnitude of insular dwarfism and gigantism is mediated by climate as well as island size and isolation, with more pronounced effects in smaller, more remote islands for mammals and reptiles. We conclude that the island rule is pervasive across vertebrates, but that the implications for body size evolution are nuanced and depend on an array of context-dependent ecological pressures and environmental conditions.

From giant pigeons to dwarf elephants, islands have long been known to generate evolutionary oddities¹. Understanding the processes by which island lineages evolve remains a prominent theme in evolutionary biology, not least because they include many of the world’s most bizarre and highly threatened organisms². The classic insular pattern of both small-animal gigantism and large-animal dwarfism in relation to mainland relatives has been described as a macroevolutionary or biogeographical rule—the ‘island rule’^{3–5} (Fig. 1). However, previous research has cast doubt on the generality of this pattern⁶, suggesting that body size shifts are asymmetrical, with reduced size in some clades (for example, carnivores, heteromyid rodents and artiodactyls) or increased size in others (for example, murid rodents)^{7,8}. Even in these cases, the underlying mechanisms driving patterns of insular gigantism and dwarfism remain unclear.

Several mechanisms have been proposed to explain the island rule, including reduced predation, relaxed competition and resource limitation in island environments⁹. In theory, each of these factors may be accentuated in smaller, more isolated islands, where lower levels of interspecific competition and predation could lead to ‘ecological release’, allowing small-bodied species to increase in body size^{5,9}. Conversely, among large-bodied species, limited resource availability could select for smaller body sizes with reduced energy requirements, leading to insular dwarfism. Climatic conditions may also influence body size evolution on islands because primary productivity and associated resource availability are strongly influenced by climate^{9,10}. Although previous studies of body size evolution on islands have tested the effects of these different mechanisms, many have focused on relatively restricted geographic and taxonomic scales and did not directly address the island rule in its broad sense across multiple species within a taxon^{10–13}, with notable exceptions^{9,14–16}.

Most work on the island rule has been restricted to mammals (for example, refs. 4,7,14,17), although the hypothesis has also been tested in amphibians¹⁸, reptiles^{19–21}, birds^{15,22}, fish²³, insects²⁴, molluscs²⁵ and plants²⁶. The highly inconsistent results of these studies (for example, refs. 5,6,27) are perhaps unsurprising because they typically deal with single species or pool together data on different traits from numerous sources without controlling for variation in study design or accounting for sampling variance. Accordingly, a recent systematic review based on a simplified scoring system²⁷ concluded that empirical support for the island rule is not only potentially biased but also generally low, particularly for non-mammalian taxa. However, scoring approaches provide only limited information, as they do not account for heterogeneity between studies, taxonomic representativeness, sample size or precision in the estimates.

These limitations are best addressed with formal meta-analyses^{28,29}, hence we tested the island rule hypothesis by applying phylogenetic meta-regressions to a global dataset of 2,479 island-mainland comparisons for 1,166 insular and 886 mainland species of terrestrial vertebrates (Fig. 2 and Supplementary Dataset 1—GitHub link provided in Supplementary Information). Our analytical framework allows us to control for multiple types of variation, including data source, sample size imbalance, intraspecific and intrapopulation variability, and phylogenetic relatedness (see Methods). For each island-mainland comparison, we calculated the log response ratio (lnRR) as the natural logarithm of the ratio between the mean body size of individuals from an insular population M_i and that of mainland relatives M_m ($\ln\text{RR} = \log(M_i/M_m)$)³⁰. Then, we regressed lnRR against the body mass of the mainland population (M_m ; Fig. 1).

This framework provides a clear set of predictions in the context of evolutionary trajectories on islands^{4,6,14}. Specifically, as negative values of lnRR indicate dwarfism and positive values indicate

¹Department of Environmental Science, Institute for Wetland and Water Research, Radboud University, Nijmegen, The Netherlands. ²Integrative Ecology Group, Estación Biológica de Doñana, Spanish National Research Council (CSIC), Sevilla, Spain. ³Department of Biology and Biotechnologies “Charles Darwin”, Sapienza University of Rome, Rome, Italy. ⁴Institute of Research on Terrestrial Ecosystems (CNR-IRET), National Research Council, Monterotondo (Rome), Italy. ⁵Department of Biodiversity and Evolutionary Biology, National Museum of Natural Sciences, Spanish National Research Council (CSIC), Madrid, Spain. ⁶Department of Life Sciences, Imperial College London, Ascot, UK. ⁷These authors contributed equally: Mark A. J. Huijbregts, Joseph A. Tobias. ✉e-mail: ana.benitez@ebd.csic.es

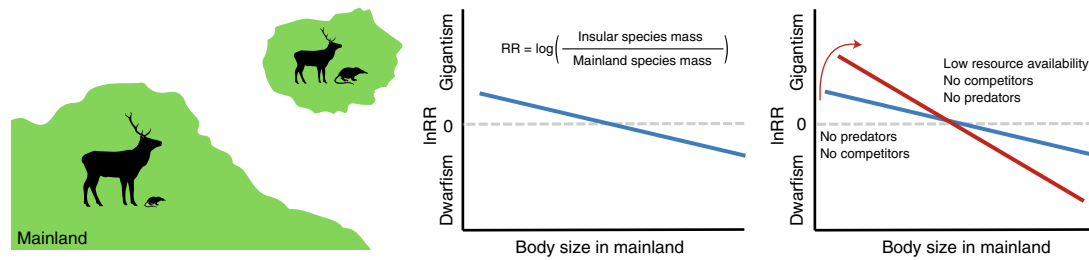


Fig. 1 | Conceptual figure showing body size evolution in island populations. According to the island rule, changes in body size of island populations are dependent on the body mass of mainland relatives, with small species tending to increase in size on islands (gigantism) and large species tending to decrease in size (dwarfism). By plotting the log response ratio (lnRR) between insular mass and mainland mass, against mainland mass, we can test if insular populations adhere to the rule (intercept >0 and slope <0; blue line). Mechanisms proposed to drive 'island rule' effects are mainly based on reduced predation, inter- and intraspecific competition, and food availability, suggesting that the relationship will steepen in small, remote islands (red line). Silhouettes extracted from 'phylopic' (www.phylopic.org).

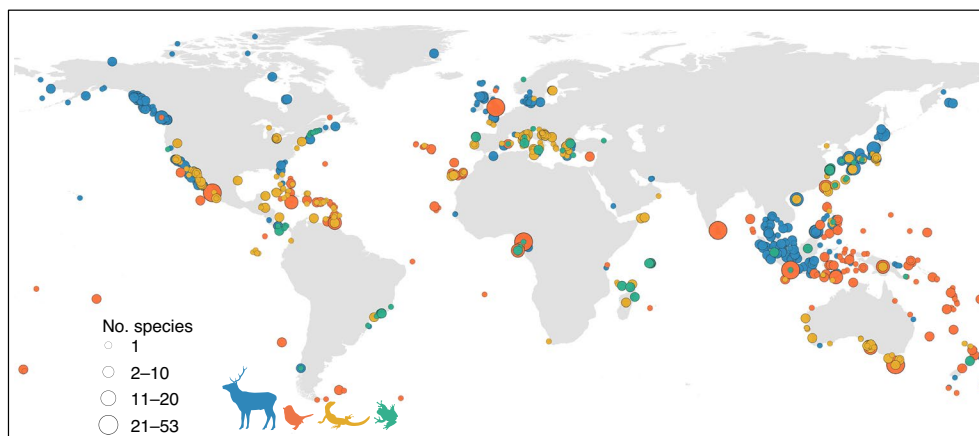


Fig. 2 | Location of island populations included in our analyses for mammals, birds, reptiles and amphibians. Mammals ($N = 1,058$) are shown in blue, birds ($N = 695$) in orange, reptiles ($N = 547$) in yellow and amphibians ($N = 179$) in green. The size of each point indicates the number of species sampled on each island; some points overlap. See Supplementary Fig. 1 for a four-panel figure with the location of insular populations separated for each taxonomic group. ArcMap 10.5. was used to create the map. Silhouettes extracted from 'phylopic' (www.phylopic.org).

gigantism, a positive intercept and negative slope of the lnRR–mainland mass relationship supports the island rule (Fig. 1). Given the contentiousness of the generality in the island rule, we assessed the robustness of our results against potential biases derived of regressing ratios^{31,32}, using small samples, imputing missing data or the influence of using data from the island rule literature or derived from other studies focused on unrelated questions (that is, publication bias; see Methods). Finally, we use our framework to assess how body size shifts are related to island size, island isolation, island productivity and climate, as well as species diet. The extent to which these different factors explain insular body size shifts allows us to re-evaluate a range of hypotheses for the mechanisms underlying 'island rule' effects on body size, including ecological release⁹, immigrant selection⁹, resource limitation^{9,33,34}, thermoregulation^{9,15,35}, water availability^{36,37} and starvation resistance^{9,33} (Supplementary Table 1 and Extended Data Fig. 1).

Results

The generality of the island rule. We found that lnRR and mainland body mass were negatively related for mammals, birds and reptiles, with small species tending to gigantism and large species to dwarfism (Fig. 3). The relationship was weakly negative but statistically non-significant for amphibians, with a tendency towards gigantism across all body sizes (Fig. 3 and Table 1). We obtained similar results using size ratios corrected for small sample size

(lnRR^A), or by regressing island mass against mainland mass, with support for the island rule across all groups except for amphibians (Supplementary Tables 3 and 4). This indicates that our analyses are robust to small sample size bias³⁸ or any potential spurious correlation associated to ratio regression models^{31,32} (Extended Data Fig. 2). Further, neither imputation nor publication bias influenced our results (Supplementary Tables 5 and 6), with no apparent differences between island–mainland comparisons sampled from studies formally testing the island rule or compiled from unrelated datasets.

Mainland body mass explained 11.4, 7.0 and 17.6% of the variance in mammals, birds and reptiles, respectively. The amount of further variance accounted for by phylogeny (0.0–29.8%), data source (1.8–25.1%), and species (25.9–53.2%) fluctuated widely among taxa (Extended Data Fig. 3). Phylogeny accounted for a relatively large amount of variance in mammals (20.1%) and reptiles (29.8%), but even in these cases the overall patterns were not driven by large effects in particular clades. Some groups tended towards gigantism and others towards dwarfism, while others contained both dwarfs or giants depending on body size (for example, Primata, Rodentia and Carnivora in mammals, and Viperidae, Scincidae and Iguanidae in reptiles; Extended Data Fig. 4).

Ecological mechanisms underlying body size evolution on islands. The pattern of body size evolution in our island–mainland comparisons provides some insight into the likely mechanisms

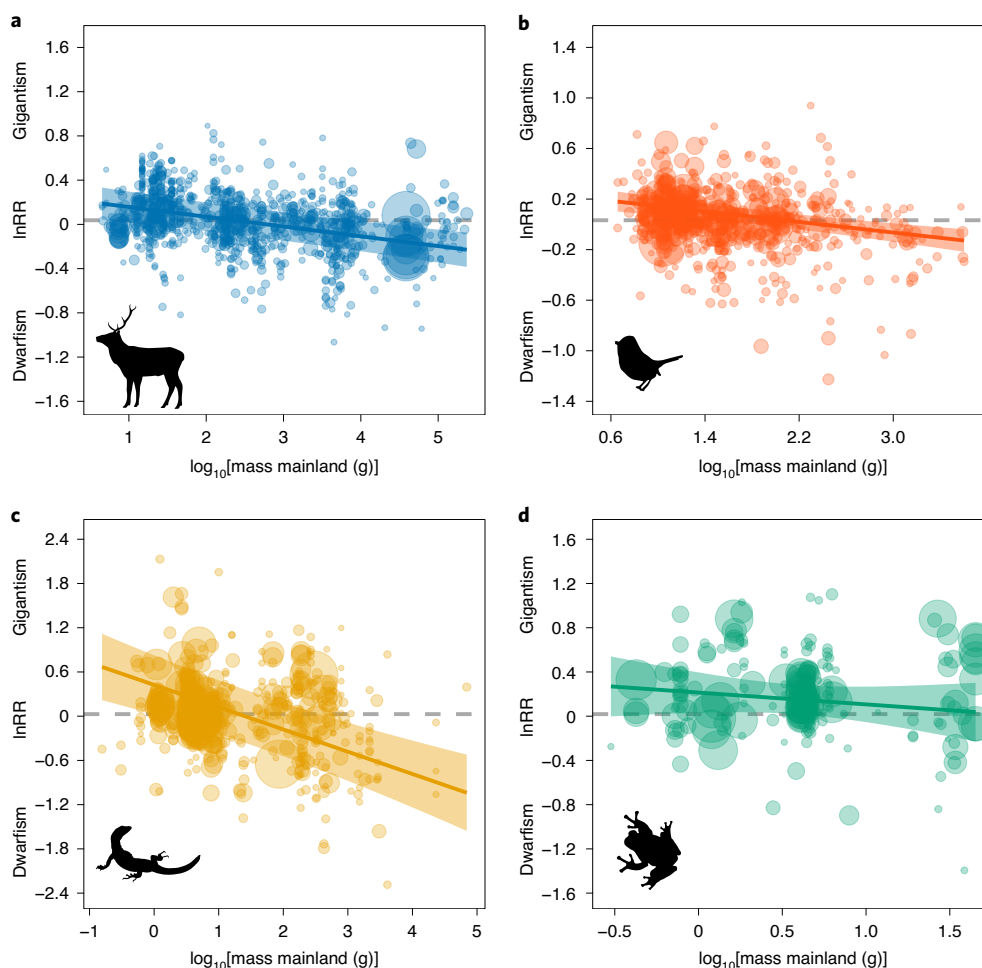


Fig. 3 | 'Island rule' effects in terrestrial vertebrates. a–d. Relationship between lnRR (log response ratio between island mass and mainland body mass) and body mass in the mainland for mammals ($N = 1,058$; **a**), birds ($N = 695$; **b**), reptiles ($N = 547$; **c**) and amphibians ($N = 179$; **d**). Models were fitted using phylogenetic multi-level meta-regression models with mainland body mass as moderator, and observation-level ID, source ID, species ID and phylogeny as random effects. lnRR > 0 indicates gigantism, lnRR < 0 indicates dwarfism and lnRR = 0 indicates stasis (no shift in body size from mainland to island populations). The size of points represents the inverse of the sampling variance for each paired island-mainland response ratio in the model. Shaded areas represent 95% confidence intervals. Note that y axes have different scales. Silhouettes extracted from 'phylopic' (www.phylopic.org).

driving 'island rule' effects (Extended Data Figs. 5–8, Supplementary Table 7 and Supplementary Dataset 2—GitHub link provided in Supplementary Information). Overall, insular size shifts arise through some combination of ecological release from predation and competition, resource limitation, biased colonization (that is, immigrant selection) and starvation resistance. The fact that no single factor explained island effects on body size is not surprising because some hypotheses shared overlapping predictions, making them difficult to disentangle.

Shifts in body mass of mammals were mostly explained by island size and spatial isolation (Omnibus test for moderators (QM) = 12.20, $P = 0.002$; Fig. 4a), resulting in more pronounced gigantism or dwarfism in small and remote islands. Birds showed similar size shifts in relation to spatial isolation and island area, but these were not statistically significant (Supplementary Table 7). In both mammals and birds, temperature had similar effects across the size range, with body size consistently larger in cool islands and smaller in warm islands (Extended Data Figs. 5e and 6e). Hence, in these groups, even large species that had undergone dwarfism were significantly larger in cool insular environments than in warm ones. Contrary to the starvation resistance hypothesis, small-sized birds did not become larger in highly seasonal islands, but large-sized

birds had reduced dwarfism on islands with high seasonality in temperatures (QM = 12.33, $P < 0.001$; Extended Data Fig. 6).

In reptiles, the combination of island area and spatial isolation were the most important factors explaining variation in body size (Fig. 4c), with productivity and seasonality also supported but with weaker effects (Extended Data Fig. 7). Similar to mammals, the tendency towards dwarfism or gigantism in large-bodied or small-bodied reptiles was more apparent in isolated small-sized islands, with stronger effects of area than isolation (Supplementary Table 7). The effects of productivity and seasonality were only partially in line with predictions, as small-sized species were larger on islands with high seasonality, but smaller on islands with high productivity (Extended Data Fig. 7). In turn, large-bodied reptiles were smaller on islands with low productivity and high seasonality.

Finally, the relationship between size ratio and mainland mass in amphibians was slightly steeper in small and remote islands (Fig. 4d), with island area being marginally more important than spatial isolation (Extended Data Fig. 8). The effect of seasonality was clearer, with amphibian species inhabiting islands with high seasonality (unpredictable environments) tending towards gigantism, whereas those from islands with low seasonality (predictable environments) being similar in size to mainland counterparts

(Extended Data Fig. 8). We found no effects of diet for any of the four taxa, or precipitation for amphibians, contrary to the water availability hypothesis.

Discussion

Based on comprehensive morphometric data from a worldwide sample of island fauna, we show consistent patterns of body size evolution across terrestrial vertebrates in accordance with predictions of the island rule. This finding was robust to alternative modelling approaches (island mass versus mainland mass regressions), small sample bias, data imputation and publication bias. Moreover, we have demonstrated that insular size shifts are contextual and depend not only on the body size of mainland relatives (island rule *sensu stricto*) but also on the physiographic and climatic characteristics of particular island environments⁹.

Repeated evolutionary trajectories on islands. We found a clear negative relationship between insular body size variation and the body mass of mainland individuals in mammals, birds and reptiles. Mainland body mass explains between 7.0 and 17.6% of the variation in insular size divergence in these three taxonomic groups, which is similar to that reported in smaller-scale studies of bats (15%), birds (13%), snakes (42%), non-volant and terrestrial mammals (11–21%) and turtles (8%)^{5,14,15,39,40}. Contrary to these earlier studies, our analyses are corrected not only for phylogenetic relatedness, but also for variability between species and intrapopulation variability, thereby strengthening the evidence for predictable evolutionary trajectories on islands. Nevertheless, the island rule provides only a partial explanation for these trajectories because substantial variation around the trend line remains unexplained. We also conducted the first multispecies test of island rule effects in amphibians, showing that the relationship goes in the expected direction but with a weak effect (1.4%), possibly because the body mass range in amphibians is narrower and limited to small sizes (~0.5–50 g) and thus most amphibians tend to gigantism on islands with reduced predation risk.

Our findings are in contrast with a number of studies rejecting the island rule, including a recent review of evidence from across mammals, birds and reptiles²⁷, as well as other taxon-specific studies focused on lizards^{20,41} and turtles²¹. On the other hand, the patterns we detect are consistent with analyses supporting the island rule in snakes¹⁹, mammals^{4,9} and birds^{5,15}. We conclude that the contradictory results of previous studies may have been related to sampling bias, heterogeneity between sources and species, and phylogenetic relatedness (that is, statistical non-independence). By accounting for these effects in our global models we are able to demonstrate that vertebrate animals evolve in largely consistent ways on islands. Further, we have shown that the island rule is not clade-specific and instead applies to numerous clades within major taxonomic groups, particularly in mammals and birds.

A corollary that emerges from the island rule is that body size converges on islands. Specifically, if insular environments select for intermediate body sizes, closer to the optimal size of the focal clade, then the size spectrum of organisms found on islands should be narrower compared with the mainland^{42,43}. Theoretically, the optimal body size towards which small and large species may converge in low-diversity systems such as islands should correspond to the point where the trend intersects the horizontal dashed line in the relationship between size ratio and mainland mass, at which point fitness is maximized⁴² (but see ref. ⁴⁴). Interestingly, the shift between dwarfism and gigantism in our models occurred at approximately 100–250 g in endotherms, slightly larger than the 100 g adult body mass proposed for mammals⁴² (but see ref. ⁴³), and the mode of the global body size distribution of birds that separate between small- and large-bodied species (60 g)^{22,45,46}. Additionally, our analyses suggest that the optimal body size for island reptiles should be ~20 g, which is marginally

Table 1 | Parameter estimates for the phylogenetic meta-regression models testing the generality of the island rule in terrestrial vertebrates

Class	<i>k</i>	Intercept (CI)	Slope (CI)	QM (P value)	<i>R</i> ² _m	<i>R</i> ² _c
Mammals	1,058	0.208 (0.052, 0.365)	−0.088 (−0.122, −0.055)	27.30 (<i>P</i> < 0.001)	11.4	67.8
Birds	695	0.216 (0.117, 0.315)	−0.104 (−0.145, −0.064)	25.40 (<i>P</i> < 0.001)	7.0	50.4
Reptiles	547	0.410 (0.006, 0.814)	−0.305 (−0.419, −0.190)	27.21 (<i>P</i> < 0.001)	17.6	84.2
Amphibians	179	0.195 (0.012, 0.377)	−0.107 (−0.320, 0.107)	0.96 (<i>P</i> = 0.328)	1.4	68.5

k, number of island-mainland comparisons (lnRR); CI, confidence interval; QM, test of moderators (\log_{10} (mainland mass)); *R*²_m, marginal *R*², estimated percentage of heterogeneity explained by the moderator (fixed effects); *R*²_c, conditional *R*², percentage of heterogeneity attributable to fixed and random effects.

higher than the modal body size of lepidosaurs (14.1 g)⁴⁷. Whether there is an optimal body size in island biotas has been the subject of much debate⁴⁴, but overall we expect that phenotypic variability in morphometric traits will be substantially narrowed if directional selection is operating in island assemblages, a feature that warrants further investigation. Additionally, optimal phenotypes should vary with the environmental characteristics of islands, in particular their area and isolation, climate, productivity and seasonality. For example, in mammals, our results suggest that the optimal body size would be ~100 g and ~900 g in warm and cold islands, respectively.

Ecological mechanisms influencing body size variation. Because body size is intimately linked to many physiological and ecological characteristics of vertebrates, it may be associated with a variety of environmental factors. As a consequence, the body size of colonizing species may predictably evolve as the result of selective pressures associated with insular environments (for example, low food resources, few competitors, no predators) and others that act across larger geographic scales (for example, climate). For mammals and reptiles, our results suggest that insular body size shifts are indeed governed by spatial isolation and island size, with individuals becoming dwarfs or giants in remote islands of limited size. Furthermore, the slope of the relationship between size ratio and mainland mass was slightly steeper for birds and amphibians in small remote islands than in large islands near continental land masses (Fig. 4). This points to a combination of resource limitation (with small islands having fewer resources to maintain large-sized organisms^{48,49}) along with release from interspecific competition and predation pressure in small, species-poor islands. The pattern is also consistent with biased colonization favouring larger individuals with higher dispersal abilities (immigration selection⁵⁰). Conversely, our results showed that body size divergence on islands close to the mainland was minimal, reflecting two non-mutually exclusive processes. First, many of these islands have been connected to the continent by land bridges so recently that phenotypic differences have not had time to accumulate. Second, regular dispersal between mainland and island populations promotes gene flow, with introgression counteracting divergent selection^{51,52}.

Besides island physiography (area and isolation), other relevant factors were temperature conditions in endotherms and resource availability and seasonality in ectothermic organisms. Mammals

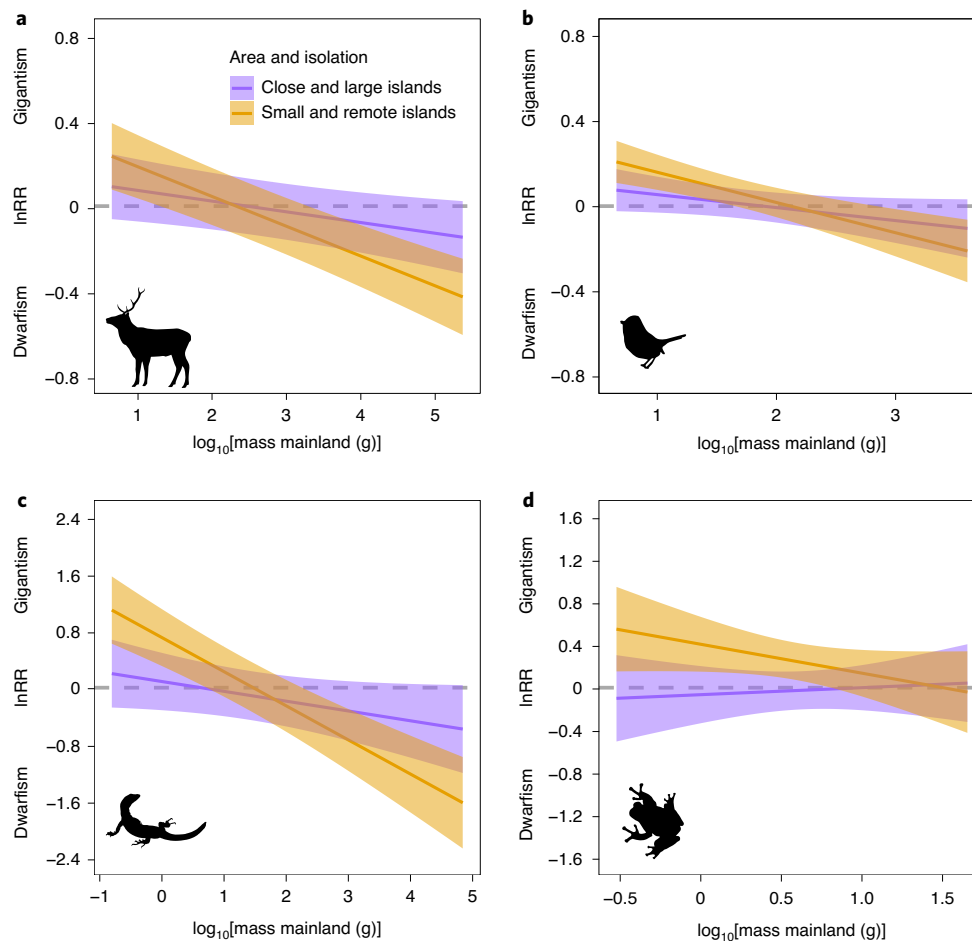


Fig. 4 | The effect of island area and spatial isolation on insular size shifts in terrestrial vertebrates. a–d. The InRR–mainland mass relationships for small and remote islands, and for close and large islands are shown for mammals ($N = 1,058$; **a**), birds ($N = 695$; **b**), reptiles ($N = 547$; **c**) and amphibians ($N = 179$; **d**). Continuous variables are represented at the 10% and 90% quantile for each extreme (close versus remote islands; small versus large islands). InRR > 0 indicates gigantism, InRR < 0 indicates dwarfism and InRR = 0 indicates stasis (no shift in body size from mainland to island populations). Shaded areas represent 95% confidence intervals. Silhouettes extracted from ‘phylopic’ (www.phylopic.org).

and birds both responded to island temperature in line with the heat conservation hypothesis, with small- and large-sized species exhibiting exacerbated gigantism and diminished dwarfism, presumably to conserve heat in colder, harsher insular environments. Additionally, temperature seasonality was an important determinant of the size of large-bodied birds, with populations on highly seasonal islands being similar in size to mainland populations. One possibility is that larger size in these cases may help maintain energy reserves during periods of low food availability, allowing them to thrive in otherwise hostile environments. Another possibility is that bird populations on highly seasonal islands—which tend to be situated at relatively high latitudes—are more often seasonally mobile or even migratory, potentially increasing gene flow with mainland populations or weakening adaptation to the local environment⁵³. These findings add new insights to previous results regarding the role of thermal and feeding ecology on morphological divergence in island birds^{54,55}. Traditionally, changes in feeding ecology were thought to be the prime force in driving morphological divergence in island birds^{54,55}. Yet, our results imply that physiological mechanisms related to heat conservation (‘thermoregulation hypothesis’) and energy constraints (‘starvation resistance hypothesis’) may also shape body size evolution in island birds.

In reptiles, we find some evidence that resource availability and seasonality are important factors explaining body size evolution,

with some deviations from the patterns predicted. As hypothesized, large species are much smaller on islands with low resource availability, and small species are larger on islands with high seasonality. Yet, unexpectedly, small species are larger on islands with low productivity, perhaps because increased intraspecific competition favours large individuals under the high population densities that reptiles often attain on islands^{56,57}.

Overall, most amphibians tended to gigantism, presumably as a result of increased growth rate or lower mortality due to reduced predation pressure on islands⁵⁸. Additionally, we found that body size of amphibians consistently increased on islands where resources were highly seasonal and unpredictable, perhaps to maximize energy reserves and withstand long periods without food, for example during aestivation or hibernation⁵⁹. We did not find a clear relationship between precipitation and body size, suggesting that water availability is not a key factor. It seems that gigantism in island amphibians is mostly driven by physiological mechanisms that maximize growth rate, particularly in smaller, more isolated islands. These findings should be further explored when more data on island–mainland pairwise populations of amphibians become available.

Body size evolution in extinct species. Our analyses focused on extant species for which we could gather information on the variation around the morphometric estimates, along with sample size

(essential for meta-analyses). The widespread extinction of large species on islands, including dwarf morphotypes of large species such as insular elephants in Sicily and the Aegean islands^{60,61}, may have masked the historical pattern of phenotypic variation on islands⁶². Giant insular birds^{54,63}, primates^{64,65} and lizards⁶⁶, along with large insular turtle species, went extinct during the Holocene and late Pleistocene⁶⁷, most likely because of overhunting and the introduction of invasive species^{68,69}. Overall, it is estimated that human colonization of oceanic islands was followed by the extinction of 27% of insular endemic mammals⁷⁰, as well as over 2,000 bird species in the Pacific region alone⁷¹, with these losses biased towards large-bodied, flightless, ground-nesting species⁶⁸. Extinct species may shed new light on size evolution in insular vertebrates because species extinctions have substantially altered the biogeography of body size in island faunas, potentially leading to downsized insular communities^{72,73}. For example, the predominance in our dataset of smaller-bodied organisms could reflect the extinction of large species on islands⁶⁸, or simply the fact that few islands support large species. Either way, further studies should include data from extinct species as this may alter or strengthen the signal that we report for extant species³⁹.

We foresee that, under global change, the extinction of insular species and the introduction of novel (invasive) species may trigger new equilibria, with concomitant shifts in the composition of insular communities and the opening of new niches to which species may respond via genetic adaptations and phenotypic plasticity. Recent evidence indicates that even introduced species on islands, which were not included in our analysis, predictably evolve towards dwarfism or gigantism^{74–76}. In theory, as the Anthropocene gathers pace, further extinctions will drive a decline in mean body size of the overall island community, pushing optimal body sizes towards the lower end of body size ranges in the different vertebrate groups.

Conclusions

Of the many evolutionary implications of living on islands—together known as the ‘island syndrome’²—the effects on body size are the most widely known and controversial. We have shown that these ‘island rule’ effects are widespread in vertebrate animals, although the evidence for amphibians is inconclusive. Morphological changes were directional for species at the extremes of the body size range in mammals, birds and reptiles, following the predicted pattern of convergence towards intermediate ‘optimum’ body sizes, in line with optimal body size theory^{42,43,45}. Although this convergence towards morphological optima may result from natural selection or phenotypic plasticity, the exact mechanism producing these changes on islands is still not well understood. Nonetheless, we found that consistent transitions towards intermediate body sizes were associated with a combination of factors, indicating a range of different ecological mechanisms. Our results highlight the contextual nature of insular size shifts, where island physiographic, climatic and ecological characteristics play a fundamental role in shaping body size evolution, reinforcing the idea that large-scale macroevolutionary patterns do not arise from single mechanisms but are often the result of multiple processes acting together^{77,78}.

Methods

Data collection. We collected baseline morphometric data from articles included in a recent assessment of the island rule²⁷, as well as other compilations assembled to test the hypothesis in reptiles²⁰, mammals⁶ and birds¹⁵. To expand this sample, we then performed a literature search (February 2020) in the Web of Science Core Collection using the following search string: ‘(island rule’ OR ‘island effect’ OR ‘island syndrome’ OR island*) AND (gigantism OR dwarfism OR ‘body size’ OR weight OR SVL OR snout-vent length OR length OR size) AND (mammal* OR bird* OR avian OR amphibia* OR reptile*)’ (Appendix 1). Because this search was complementary to the data we had gathered from previous compilations^{6,15,20,27}, we downloaded only the first 500 hits out of a total of 33,431 hits ordered by relevance, and removed duplicates already included in our dataset. We reviewed every island-mainland comparison reported in published studies and traced primary source

data when possible to extract original measurements. We also extracted data from all studies containing morphometric measurements for insular populations when these could be matched with equivalent data published elsewhere for relevant mainland taxa. We excluded problematic data, such as comparisons that were not supported by taxonomic or phylogenetic evidence, or that reported morphometric data restricted to single specimens or without sample size. In addition, we excluded comparisons based on extinct taxa as they are often known from very few or incomplete specimens (Supplementary Dataset 3—GitHub link provided in Supplementary Information).

It has been argued that research on the island rule might be prone to ascertainment bias, where researchers are more likely to notice and measure animals of extreme body size when conducting research on islands⁴¹. To help overcome this problem, we collected body size data not only from studies testing the island rule, or reporting dwarfism and gigantism in island fauna, but also from studies that did not specifically test hypotheses related to the island rule. We matched unpaired insular populations with independent data from mainland populations by performing species-specific searches in the Web of Science Core Collection and Google Scholar. We also compiled morphometric data for 442 insular and 407 mainland bird species from an independent global dataset of avian functional traits⁷⁹.

Large islands may be more ‘mainland like’ in relation to factors that are thought to affect body size (that is, competition, resource availability and predation⁵). Thus, when major islands were at least ten times larger than a nearby island, we treated the large island as the mainland comparison, following previous studies testing the island rule^{4,5,20}. Consequently, a single mid-sized island can simultaneously be treated as the continent in comparisons with smaller islands, and the island in comparisons with larger continents. When authors reported data referring to an entire archipelago instead of a specific island (3.2% of cases), we used the size of the largest island as island area. Removing these cases from our analyses did not qualitatively affect our results (Supplementary Table 8).

Our final dataset contained 529 data sources and 2,479 island-mainland comparisons^{7,10,17,36,58,79–601}. In total, we collated morphometric measurements for 63,561 insular and 154,875 mainland specimens representing mammals (1,058 island-mainland comparisons), birds (695 comparisons), reptiles (547 comparisons) and amphibians (179 comparisons) from across the globe (Fig. 2). A total of 2,068 island-mainland comparisons (83.4%) were within-species (for example, subspecies) comparisons and 411 (16.6%) were between-species comparisons. Insular populations were sampled from an array of islands varying widely in size (0.0009–785,753 km²), climate and level of spatial isolation (0.03–3,835 km from mainland). To explore the drivers of body size shifts in insular populations, we also sampled species with a wide range of average body masses (0.18–234,335 g). We collated data on body size indices (body mass, body length, cranial and dental measurements) of different taxa in island and mainland populations following strict morphological, phylogenetic and biogeographic criteria. Specifically, we always compared the same body size index for island and mainland populations. For within-species comparisons, we compared island and mainland populations based on the information given by the authors of the relevant study (for example, taking note of which mainland source populations are likely to inhabit a particular island because of colonization history or isolation via rising sea levels^{89,101,240,385,547}). When we matched comparisons independently, we used information published in the study reporting the insular form, selecting the geographically closest mainland population whenever possible. In addition, we prioritized latitudinal alignment of mainland and island populations to avoid confounding effects of latitudinal variation in body size. In the case of island endemics, we compared island populations with their closest mainland relative whenever these were identifiable by phylogenetic data or other information reported in each particular study. This usually meant selecting their sister species or the geographically closest representative of a sister clade or polytomy (Supplementary Dataset 1—GitHub link provided in Supplementary Information). If we could not reliably establish the closest mainland relative, we discarded the data (Supplementary Dataset 3—GitHub link provided in Supplementary Information).

When more than one body size index was reported in published studies, we prioritized those indices related most closely to body mass (Supplementary Table 2). For mammals, we selected indices in the following order of preference: body mass, body length, cranial length (greatest skull length or condylobasal length) and dentition (for example, canine length)⁵. For birds, preferred indices were body mass, wing length, tarsus length and bill length. Finally, for amphibians and reptiles, size was reported as body mass, snout-vent length, carapace length (for turtles) and total length (including snout-vent length and tail length). In all cases, we included measurements only for adults. To avoid size biases attributable to sexual size dimorphism, we calculated the pooled mean for both sexes and the combined standard deviation using standard formulae for combining groups⁶⁰². When information was available for only one sex (male or female), we restricted our size comparisons to the sex for which we had morphometric data in both mainland and island populations. Data from zoos or studies that could not be georeferenced were discarded.

To overcome the problem that different authors report size using different indices, we used allometric relationships to convert island and mainland size to

body mass equivalents, thereby enabling cross-taxa and cross-study comparisons. Although this conversion is imprecise, morphological indices and body mass are nonetheless highly correlated across the global scale and wide range of body sizes within our samples (providing more accurate predictions than simply assuming an exponent -3 , as in previous studies testing the island rule⁵⁰). We used published allometric relationships where available (Supplementary Table 2), or derived them based on published datasets^{47,179,603–608} and other data sources (Supplementary Dataset 4—GitHub link provided in Supplementary Information). To calculate allometric relationships, we used ordinary least square models of the \log_{10} -transformed body mass against the \log_{10} -transformed body size index (for example, condylobasal length; Supplementary Table 2 and Dataset 4).

For birds, we complemented published data with standardized morphometric measurements from 3,618 museum specimens and live individuals of 436 insular and 404 mainland bird species (see ref. ⁷⁹). We used wing length in the main analyses instead of tarsus length because the former is a better predictor of body mass in our dataset ($R^2_{\text{wing}} = 0.89$ versus $R^2_{\text{tarsus}} = 0.69$; Supplementary Table 2; see also ref. ⁶⁰⁹). Although wing length may change during moult or thereafter because of wear, these effects are negligible in relation to interspecific differences⁷⁹, and minimized by calculating averages across multiple individuals. Further, interobserver differences between measurements may explain some variation in wing length estimates, but again this bias was shown to have negligible effects in our dataset by comparing repeated measures from different observers (see ref. ⁷⁹). To assess the consistency in our results, we repeated analyses using tarsus length, another popular proxy of overall body size in birds⁶¹⁰. Our results were unchanged (Supplementary Fig. 2).

To select suitable comparisons for museum specimens, we first classified species as either insular or continental by overlapping International Union for Conservation of Nature range polygons with a geographic information system land layer including continental land masses. For each insular species we then identified continental sister species from avian phylogenies⁶¹¹, using the method described above. We excluded bird species that are highly pelagic or aerial (for example, swifts) and fully migratory species, because in these groups it is unclear whether insular and mainland forms experience different environments¹⁵. Further, we also excluded flightless bird species, because morphological changes may be owing to flightlessness rather than island dwelling per se¹⁵.

We calculated the response ratio (lnRR; equation (1)) as effect size in our meta-regressions, where we divided the mean body mass of individuals from an insular population, \bar{M}_i , by that of the nearest mainland relative, \bar{M}_m , and then applied the natural logarithm. Unlike unlogged ratios, the sampling distribution of lnRR is normal, particularly for small samples³⁰, and thus less prone to statistical artefacts associated with ratio-based regressions.

$$\ln\text{RR} = \ln \left(\frac{\bar{M}_i}{\bar{M}_m} \right) \quad (1)$$

Response ratios >0 indicate a shift towards larger sizes (gigantism) whereas ratios <0 indicate shifts towards smaller sizes (dwarfism). Besides mean measurements, we recorded measures of variation, that is, standard deviation, standard error or coefficient of variation, and sample sizes of the body size indices in island and mainland organisms. Standard deviation and sample sizes were used to calculate sampling variances (equation (2)), which were then used to weight each response ratio (coupled with the amount of heterogeneity, that is, the variance in the underlying effects)³⁰.

$$\hat{\sigma}^2(\ln\text{RR}) = \frac{\text{SD}_i^2}{N_i \bar{M}_i^2} + \frac{\text{SD}_m^2}{N_m \bar{M}_m^2}; \quad (2)$$

where SD_i and SD_m are the standard deviation of the mean body size in the insular and the mainland population, respectively, and N_i and N_m the sample sizes. Standard deviations (SD) were extracted from raw data when possible. If ranges were provided instead of standard deviation (or standard error or coefficient of variation), we calculated standard deviation following ref. ⁶¹². If neither ranges nor measures of variation were reported, but the reported sample size was >1 , we imputed standard deviation based on the coefficient of variation from all complete cases ('Bracken approach'⁶¹³). Imputation was done for 22% of all cases in mammals, 1.1% in birds, 11% in reptiles and 7.3% in amphibians, all within the upper limit of imputations ($<30\%$ of all cases per group) advised in previous studies³⁰.

For each study and island–mainland comparison, we compiled the mainland and island names, the study reference, the body size index used, the geographic coordinates, the distance to the closest mainland (spatial isolation, in kilometres) and the island area (in square kilometres). We completed missing data on island characteristics using the United Nations Environment Programme island database (<http://islands.unep.ch/>) and the Threatened Island Biodiversity Database (<http://tib.islandconservation.org/>). Missing information was calculated using Google Earth. Additionally, we extracted the normalized difference vegetation index (NDVI) as a proxy for resource availability on islands⁶¹⁴. We also calculated the standard deviation of the NDVI to assess seasonality in leaf or vegetation cover, as an index of seasonality in available resources. The NDVI was downloaded from the NASA Ames Ecological Forecasting Lab (<https://lpdaacsvc.cr.usgs.gov/appears/task/area>).

As climate influences both resource requirements and primary productivity, body size evolution should also be influenced by climatic conditions on islands. We thus extracted island climatic conditions from WorldClim version 2.0 (<http://worldclim.org>⁶¹⁵). Specifically, we used variables that are more closely associated with the proposed underlying mechanisms of Bergmann's rule (that is, thermoregulation and starvation resistance): mean annual temperature, annual precipitation, and seasonality of temperature and precipitation⁶¹⁶. We assumed that the time period for these bioclimatic variables (1970–2000), although not necessarily matching the actual time period of body size evolution in the insular populations, roughly represents the climatic conditions in the Holocene, a period relatively climatically stable where most of our populations became isolated (that is, after the Last Glacial Maximum; see also ref. ⁷⁹). Because climatic variability across cells substantially exceeds variation within cells in the Holocene, current layers are considered adequate for geographic comparisons. All spatial variables were downloaded at 0.1 degree resolution, and we averaged all cells per island to obtain a mean value of each environmental variable (for example, temperature, NDVI, precipitation and so on). Finally, for each species included in our dataset, we collated diet information from EltonTraits for birds and mammals⁶¹⁷, and from other sources for reptiles^{607,618}, and classified species as carnivores ($>50\%$ diet consisting of vertebrates) or non-carnivores ($<50\%$ diet consisting of vertebrates), following previous studies^{79,619}. As all amphibians in our dataset are carnivores⁶²⁰, we did not record their diet.

Data analyses. To test the island rule hypothesis, we used phylogenetic meta-regressions between lnRR and body mass of mainland relatives, following most previous studies of the island rule (for example, refs. ^{4,5,76,621,622}). A negative slope for this relationship would support the island rule (Fig. 1).

The use of multiple populations of the same species can overestimate the actual number of degrees of freedom, generating type 1 errors. We controlled for this by adding 'Species' as a random effect intercept in our analyses. Additionally, body size evolution in insular vertebrates is heavily influenced by phylogenetic effects, with species within entire clades seemingly showing either dwarfism or gigantism⁶. Thus, we accounted for phylogeny by including the phylogenetic relatedness correlation matrix as a random effect. The species term captures the similarities of effect sizes within the same species, while the phylogenetic term represents the similarity due to relatedness⁶²³. We also added 'Source' as a random effect intercept to account for between-source variability and the fact that we had multiple response ratios per study. In some cases, 'Source' represented the combination of two sources of data, one for the island size and one for the mainland size. Finally, we included an observation-level random effect, which represents the residual variance that needs to be explicitly modelled in a meta-analysis²⁹. Total heterogeneity and heterogeneity due to phylogeny, source and species identity were computed following ref. ²⁹.

We tested the robustness of our results against several potential limitations. As multiple island populations were often compared with a single mainland population, we accounted for these repeated measures in a variance–covariance matrix where the diagonal includes the sampling variances and the off-diagonals of the matrix represent the shared variance (covariance) among the response ratios due to the common mainland population⁶²⁴. Further, we compared our main results with models fitted with lnRR and sampling variances corrected for small sample size³⁸. Another potential problem is that regressions using ratios may lead to spurious correlations^{31,32}. Thus, we conducted an additional analysis testing the statistical significance of body size trends by regressing island mass against mainland mass, following previous studies^{4,5,20,41}. Phylogenetic meta-regressions were run using island mass as the response variable and mainland mass as the predictor (both transformed with natural log), with random effects as specified above, and sampling variance $\text{SD}_i^2/\bar{M}_i^2 \times N_i$. This approach has some limitations in being harder to visualize and less effective in considering the sampling variance of measurements (representing intrapopulation variability), yet nonetheless provides an alternative approach for assessing the robustness of our results, in line with previous studies^{4,5,20,41}. Finally, we assessed publication bias by testing the influence of data source on the relationship between size ratio and mainland mass. This involved comparing whether patterns differed in island–mainland pairs extracted from studies testing the island rule (38.6% of cases) versus pairs extracted from studies not testing the island rule (61.4% of cases).

Testing ecological hypotheses explaining insular size shifts. To evaluate the relative roles of key mechanisms proposed to influence body size evolution in island fauna, we compiled a further range of variables (Supplementary Table 1 and Extended Data Fig. 1). These included island area (linked to both resource limitation and to ecological release from both predation and competition) and spatial isolation (linked to reduced colonization from mainland populations for smaller taxa, that is, immigration selection⁵⁰). In addition, we included climatic and resource seasonality, which are linked to the starvation resistance hypothesis, and productivity and species diet, each of which are linked to resource limitation. Because body size evolution may be influenced by climate (for example, Bergmann's rule)^{9,621}, we also included mean temperature, which is linked to body size adaptations for enhancing heat conservation or dissipation (thermoregulation hypothesis). For amphibians, we included precipitation as a proxy for water supply linked to aquatic habitats, moisture and humidity (water availability hypothesis).

We modelled interactions between body size and each of the explanatory variables because we expected these factors to differentially affect species of different sizes, thus producing different effects in small, medium-sized and large species. In line with the ecological release and resource limitation hypotheses, we expected the slope of the $\ln R$ –mainland mass relationship to be steeper in smaller islands, isolated from the mainland and with fewer or no predators (Fig. 1). Further, if resource availability is a key factor, we also expected large species to undergo dwarfism on islands with low productivity^{48,49}, and for dwarfism to be accentuated in dietary niches with high energy requirements, including carnivory⁹. In addition, high seasonality in resources and in temperature was expected to result in increased gigantism in small-sized species, because energy reserves increase faster than energy depletion as body size increases (starvation resistance hypothesis)^{9,625}. We hypothesized that smaller species would benefit comparatively more by increasing in size than larger species. As amphibians are generally small sized, we also fitted a model for this group with only additive terms (mainland mass + $s\text{dNDVI}$) where seasonality in resources ($s\text{dNDVI}$) would result in larger body sizes for all species. Finally, mechanisms driven by thermoregulation and water availability predict that body size shifts are associated with temperature and rainfall, respectively. Mean temperature was expected to predominantly affect endotherms and small ectotherms with good thermoregulating abilities (reptiles and anurans) living on cold islands that, compared with similar-sized species on islands with a mild climate, would exhibit more pronounced gigantism to enhance heat conservation. We fitted the effect of mean temperature (T_{mean}) as an interactive (mainland mass $\times T_{\text{mean}}$) or additive term (mainland mass + T_{mean}) to assess whether only small species or all species would increase in size in low temperature islands (Supplementary Tables 1 and 7, and Extended Data Fig. 1).

Prior to modelling, all the moderators (explanatory variables) were inspected and \log_{10} transformed if necessary to meet normality assumptions in model errors. We considered a result to be significant when the 95% confidence interval did not cross zero. We assessed the explained heterogeneity using QM and the percentage of variance explained by the moderators using R^2 marginal⁶²⁶. All figures show the relationship between size response ratio and body mass, and how this might be altered by the mechanisms explained above.

All analyses were performed in R version 3.5.3⁶²⁷ using the packages metafor v2.0⁶²⁸ and metagear v0.4⁶²⁹ for the meta-regression models and data imputation, metaDigitise v1.0⁶³⁰ for data extraction from plots, ape v5.2⁶³¹ for estimating branch lengths and resolving polytomies, rotl v3.0.4⁶³² for building the phylogenies for our species by searching the Open Tree Taxonomy⁶³³ and retrieving the phylogenetic relationships from the Open Tree of Life⁶³⁴, sf v0.7-3⁶³⁵ and raster v2.7-15⁶³⁶ for spatial analyses, dplyr v0.8.0.1⁶³⁷ and reshape2 v1.4.3⁶³⁸ for data manipulation and ggplot2 v3.3.0.9000⁶³⁹ and ggpubr v0.1.8⁶⁴⁰ for data visualization. ArcMap 10.5 was used for Fig. 2. Silhouettes in figures were extracted from 'phylopic' (www.phylopic.org). The PRISMA checklist for systematic reviews is available in Appendix 3.

Reporting Summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

All data are available at https://github.com/anabenlop/Island_Rule and https://figshare.com/projects/Body_size_evolution_in_insular_vertbrates/89102.

Code availability

The code to conduct the analyses is available at https://github.com/anabenlop/Island_Rule.

Received: 1 May 2020; Accepted: 22 February 2021;

Published online: 15 April 2021

References

- Foster, J. B. Evolution of mammals on islands. *Nature* **202**, 234–235 (1964).
- Baeckens, S. & Van Damme, R. The island syndrome. *Curr. Biol.* **30**, R338–R339 (2020).
- Whittaker, R. J. & Fernández-Palacios, J. M. *Island Biogeography: Ecology, Evolution, and Conservation* (Oxford Univ. Press, 2007).
- Lomolino, M. V. Body size of mammals on islands: the island rule reexamined. *Am. Nat.* **125**, 310–316 (1985).
- Lomolino, M. V. Body size evolution in insular vertebrates: generality of the island rule. *J. Biogeogr.* **32**, 1683–1699 (2005).
- Meiri, S., Cooper, N. & Purvis, A. The island rule: made to be broken? *Proc. R. Soc. B* **275**, 141–148 (2008).
- Meiri, S., Dayan, T. & Simberloff, D. The generality of the island rule reexamined. *J. Biogeogr.* **33**, 1571–1577 (2006).
- Meiri, S., Dayan, T. & Simberloff, D. Body size of insular carnivores: little support for the island rule. *Am. Nat.* **163**, 469–479 (2004).
- Lomolino, M. V., Sax, D. F., Palombo, M. R. & van der Geer, A. A. Of mice and mammoths: evaluations of causal explanations for body size evolution in insular mammals. *J. Biogeogr.* **39**, 842–854 (2012).
- Millien, V. Relative effects of climate change, isolation and competition on body-size evolution in the Japanese field mouse, *Apodemus argenteus*. *J. Biogeogr.* **31**, 1267–1276 (2004).
- Angerbjörn, A. Gigantism in island populations of wood mice (*Apodemus*) in Europe. *Oikos* **47**, 47–56 (1986).
- Schillaci, M. A., Meijaard, E. & Clark, T. The effect of island area on body size in a primate species from the Sunda Shelf islands. *J. Biogeogr.* **36**, 362–371 (2009).
- Radtkey, R. R., Fallon, S. M. & Case, T. J. Character displacement in some *Cnemidophorus* lizards revisited: a phylogenetic analysis. *Proc. Natl Acad. Sci. USA* **94**, 9740–9745 (1997).
- McClain, C. R., Durst, P. A., Boyer, A. G. & Francis, C. D. Unravelling the determinants of insular body size shifts. *Biol. Lett.* **9**, 20120989 (2013).
- Clegg, S. M. & Owens, P. The 'island rule' in birds: medium body size and its ecological explanation. *Proc. R. Soc. B* **269**, 1359–1365 (2002).
- Raia, P. & Meiri, S. The island rule in large mammals: paleontology meets ecology. *Evolution* **60**, 1731–1742 (2006).
- Bromham, L. & Cardillo, M. Primates follow the 'island rule': implications for interpreting *Homo floresiensis*. *Biol. Lett.* **3**, 398–400 (2007).
- Montesinos, R., da Silva, H. R. & de Carvalho, A. L. G. The 'island rule' acting on anuran populations (Bufonidae: *Rhinella ornata*) of the Southern Hemisphere. *Biotropica* **44**, 506–511 (2012).
- Boback, S. M. Body size evolution in snakes: evidence from island populations. *Copeia* **2003**, 81–94 (2003).
- Meiri, S. Size evolution in island lizards. *Glob. Ecol. Biogeogr.* **16**, 702–708 (2007).
- Itescu, Y., Karraker, N. E., Raia, P., Pritchard, P. C. & Meiri, S. Is the island rule general? Turtles disagree. *Glob. Ecol. Biogeogr.* **23**, 689–700 (2014).
- Boyer, A. G. & Jetz, W. Biogeography of body size in Pacific island birds. *Ecography* **33**, 369–379 (2010).
- Herczeg, G., Gonda, A. & Merilä, J. Evolution of gigantism in nine-spined sticklebacks. *Evolution* **63**, 3190–3200 (2009).
- Palmer, M. Testing the 'island rule' for a tenebrionid beetle (Coleoptera, Tenebrionidae). *Acta Oecol.* **23**, 103–107 (2002).
- McClain, C. R., Boyer, A. G. & Rosenberg, G. The island rule and the evolution of body size in the deep sea. *J. Biogeogr.* **33**, 1578–1584 (2006).
- Biddick, M., Hendriks, A. & Burns, K. Plants obey (and disobey) the island rule. *Proc. Natl Acad. Sci. USA* **116**, 17632–17634 (2019).
- Lokatis, S. & Jeschke, J. M. The island rule: an assessment of biases and research trends. *J. Biogeogr.* **45**, 289–303 (2018).
- Gurevitch, J., Koricheva, J., Nakagawa, S. & Stewart, G. Meta-analysis and the science of research synthesis. *Nature* **555**, 175–182 (2018).
- Nakagawa, S. & Santos, E. S. Methodological issues and advances in biological meta-analysis. *Evol. Ecol.* **26**, 1253–1274 (2012).
- Hedges, L. V., Gurevitch, J. & Curtis, P. S. The meta-analysis of response ratios in experimental ecology. *Ecology* **80**, 1150–1156 (1999).
- Atchley, W. R., Gaskins, C. T. & Anderson, D. Statistical properties of ratios. I. Empirical results. *Syst. Zool.* **25**, 137–148 (1976).
- Prairie, Y. T. & Bird, D. F. Some misconceptions about the spurious correlation problem in the ecological literature. *Oecologia* **81**, 285–288 (1989).
- Phillips, L. B., Hansen, A. J. & Flather, C. H. Evaluating the species energy relationship with the newest measures of ecosystem energy: NDVI versus MODIS primary production. *Remote Sens. Environ.* **112**, 3538–3549 (2008).
- Olesen, J. M. & Valido, A. Lizards as pollinators and seed dispersers: an island phenomenon. *Trends Ecol. Evol.* **18**, 177–181 (2003).
- Zamora-Camacho, F., Reguera, S. & Moreno-Rueda, G. Bergmann's rule rules body size in an ectotherm: heat conservation in a lizard along a 2200-metre elevational gradient. *J. Evol. Biol.* **27**, 2820–2828 (2014).
- Valenzuela-Sánchez, A., Cunningham, A. A. & Soto-Azat, C. Geographic body size variation in ectotherms: effects of seasonality on an anuran from the southern temperate forest. *Front. Zool.* **12**, 37 (2015).
- Ashton, K. G. Do amphibians follow Bergmann's rule? *Can. J. Zool.* **80**, 708–716 (2002).
- Lajeunesse, M. J. Bias and correction for the log response ratio in ecological meta-analysis. *Ecology* **96**, 2056–2063 (2015).
- Lomolino, M. V. et al. Of mice and mammoths: generality and antiquity of the island rule. *J. Biogeogr.* **40**, 1427–1439 (2013).
- Boback, S. M. & Guyer, C. Empirical evidence for an optimal body size in snakes. *Evolution* **57**, 345–451 (2003).
- Meiri, S., Raia, P. & Phillimore, A. B. Slaying dragons: limited evidence for unusual body size evolution on islands. *J. Biogeogr.* **38**, 89–100 (2011).
- Brown, J. H., Marquet, P. A. & Taper, M. L. Evolution of body size: consequences of an energetic definition of fitness. *Am. Nat.* **142**, 573–584 (1993).
- Damuth, J. Cope's rule, the island rule and the scaling of mammalian population density. *Nature* **365**, 748–750 (1993).
- Raia, P., Carotenuto, F. & Meiri, S. One size does not fit all: no evidence for an optimal body size on islands. *Glob. Ecol. Biogeogr.* **19**, 475–484 (2010).

45. Maurer, B. The evolution of body size in birds. II. The role of reproductive power. *Evol. Ecol.* **12**, 935–944 (1998).
46. Blackburn, T. M. & Gaston, K. J. The distribution of body sizes of the world's bird species. *Oikos* **70**, 127–130 (1994).
47. Feldman, A., Sabath, N., Pyron, R. A., Mayrose, I. & Meiri, S. Body sizes and diversification rates of lizards, snakes, amphisbaenians and the tuatara. *Glob. Ecol. Biogeogr.* **25**, 187–197 (2016).
48. McNab, B. K. Minimizing energy expenditure facilitates vertebrate persistence on oceanic islands. *Ecol. Lett.* **5**, 693–704 (2002).
49. McNab, B. K. Geographic and temporal correlations of mammalian size reconsidered: a resource rule. *Oecologia* **164**, 13–23 (2010).
50. Durst, P. A. & Roth, V. L. Mainland size variation informs predictive models of exceptional insular body size change in rodents. *Proc. R. Soc. B* **282**, 20150239 (2015).
51. Tobias, J. A., Ottenburghs, J. & Pigot, A. L. Avian diversity: speciation, macroevolution, and ecological function. *Annu. Rev. Ecol. Evol. Syst.* **51**, 533–560 (2020).
52. Li, J.-W. et al. Rejecting strictly allopatric speciation on a continental island: prolonged postdivergence gene flow between Taiwan (*Leucodioptron taewanus*, Passeriformes Timaliidae) and Chinese (*L. canorum canorum*) hwameis. *Mol. Ecol.* **19**, 494–507 (2010).
53. Somveille, M., Rodrigues, A. S. & Manica, A. Why do birds migrate? A macroecological perspective. *Glob. Ecol. Biogeogr.* **24**, 664–674 (2015).
54. Blondel, J. Evolution and ecology of birds on islands: trends and prospects. *Vie Milieu* **50**, 205–220 (2000).
55. Grant, P. R. in *Evolution on Islands* (1998).
56. Novosolov, M. et al. Power in numbers. Drivers of high population density in insular lizards. *Glob. Ecol. Biogeogr.* **25**, 87–95 (2016).
57. Santini, L. et al. Global drivers of population density in terrestrial vertebrates. *Glob. Ecol. Biogeogr.* **27**, 968–979 (2018).
58. Castellano, S. & Giacoma, C. Morphological variation of the green toad, *Bufo viridis*, in Italy: a test of causation. *J. Herpetol.* **32**, 540–550 (1998).
59. Ashton, K. G. Body size variation among mainland populations of the western rattlesnake (*Crotalus viridis*). *Evolution* **55**, 2523–2533 (2001).
60. Athanassiou, A., van der Geer, A. A. & Lyras, G. A. Pleistocene insular Proboscidea of the eastern Mediterranean: a review and update. *Quat. Sci. Rev.* **218**, 306–321 (2019).
61. Herridge, V. L. & Lister, A. M. Extreme insular dwarfism evolved in a mammoth. *Proc. R. Soc. B* **279**, 3193–3200 (2012).
62. Faurby, S. & Svenning, J.-C. Resurrection of the island rule: human-driven extinctions have obscured a basic evolutionary pattern. *Am. Nat.* **187**, 812–820 (2016).
63. Steadman, D. W. *Extinction and Biogeography of Tropical Pacific Birds* (Univ. Chicago Press, 2006).
64. MacPhee, R. D. & Horowitz, I. New craniodental remains of the Quaternary Jamaican monkey *Xenothrix mcgregori* (Xenotrichini, Callicebinae, Pitheciidae), with a reconsideration of the Aotus hypothesis. *Am. Mus. Novit.* **2004**, 3434 (2004).
65. MacPhee, R. & Fleagle, J. Postcranial remains of *Xenothrix mcgregori* (Primates, Xenotrichidae) and other Late Quaternary mammals from Long Mile Cave, Jamaica. *Bull. Am. Mus. Nat. Hist.* **206**, 287–321 (1991).
66. Pregill, G. Body size of insular lizards: a pattern of Holocene dwarfism. *Evolution* **40**, 997–1008 (1986).
67. Hansen, D. M., Donlan, C. J., Griffiths, C. J. & Campbell, K. J. Ecological history and latent conservation potential: large and giant tortoises as a model for taxon substitutions. *Ecography* **33**, 272–284 (2010).
68. Boyer, A. G. Extinction patterns in the avifauna of the Hawaiian islands. *Divers. Distrib.* **14**, 509–517 (2008).
69. White, A. W., Worthy, T. H., Hawkins, S., Bedford, S. & Spriggs, M. Megafaunal meiolaniid horned turtles survived until early human settlement in Vanuatu, southwest Pacific. *Proc. Natl Acad. Sci. USA* **107**, 15512–15516 (2010).
70. Alcover, J. A., Sans, A. & Palmer, M. The extent of extinctions of mammals on islands. *J. Biogeogr.* **25**, 913–918 (1998).
71. Steadman, D. W. Prehistoric extinctions of Pacific island birds: biodiversity meets zooarchaeology. *Science* **267**, 1123–1131 (1995).
72. Pérez-Méndez, N., Jordano, P., García, C. & Valido, A. The signatures of Anthropocene defaunation: cascading effects of the seed dispersal collapse. *Sci. Rep.* **6**, 24820 (2016).
73. Heinen, J. H., van Loon, E. E., Hansen, D. M. & Kissling, W. D. Extinction-driven changes in frugivore communities on oceanic islands. *Ecography* **41**, 1245–1255 (2018).
74. van der Geer, A. A. E. Changing invaders: trends of gigantism in insular introduced rats. *Environ. Conserv.* **45**, 203–211 (2018).
75. van der Geer, A. A. E., Lomolino, M. V. & Lyras, G. 'On being the right size' - do aliens follow the rules? *J. Biogeogr.* **45**, 515–529 (2018).
76. Mathys, B. A. & Lockwood, J. L. Rapid evolution of great kiskadees on Bermuda: an assessment of the ability of the island rule to predict the direction of contemporary evolution in exotic vertebrates. *J. Biogeogr.* **36**, 2204–2211 (2009).
77. Quinn, J. F. & Dunham, A. E. On hypothesis testing in ecology and evolution. *Am. Nat.* **122**, 602–617 (1983).
78. McGill, B. J. & Nekola, J. C. Mechanisms in macroecology: AWOL or purloined letter? Towards a pragmatic view of mechanism. *Oikos* **119**, 591–603 (2010).
79. Pigot, A. L. et al. Macroevolutionary convergence connects morphological form to ecological function in birds. *Nat. Ecol. Evol.* **4**, 230–239 (2020).
80. Krause, M. A., Burghardt, G. M. & Gillingham, J. C. Body size plasticity and local variation of relative head and body size sexual dimorphism in garter snakes (*Thamnophis sirtalis*). *J. Zool.* **261**, 399–407 (2003).
81. Krystufek, B., Tvrtkovic, N., Paunovic, M. & Ozkan, B. Size variation in the northern white-breasted hedgehog *Erinaceus roumanicus*: latitudinal cline and the island rule. *Mammalia* **73**, 299–306 (2009).
82. Kubo, M. O. & Takatsuki, S. Geographical body size clines in sika deer: path analysis to discern amongst environmental influences. *Evol. Biol.* **42**, 115–127 (2015).
83. Kuchling, G., Rhodin, A. G., Ibarrondo, B. R. & Trainor, C. R. A new subspecies of the snakeneck turtle *Chelodina mccordi* from Timor-Leste (East Timor) (Testudines: Chelidae). *Chelonian Conserv. Biol.* **6**, 213–222 (2007).
84. Kuo, C.-Y., Lin, Y.-T. & Lin, Y.-S. Sexual size and shape dimorphism in an agamid lizard, *Japalura swinhonis* (Squamata: Lacertilia: Agamidae). *Zool. Stud.* **48**, 351–361 (2009).
85. Kurta, A. & Ferkin, M. The correlation between demography and metabolic rate: a test using the beach vole (*Microtus breweri*) and the meadow vole (*Microtus pennsylvanicus*). *Oecologia* **87**, 102–105 (1991).
86. Kutrup, B., Cakir, E., Colak, Z., Bulbul, U. & Karaoglu, H. Age and growth of the green toad, *Bufo viridis* (Laurenti, 1768) from an island and a mainland population in Giresun, Turkey. *J. Anim. Vet. Adv.* **10**, 1469–1472 (2011).
87. Kwet, A., Steiner, J. & Zillikens, A. A new species of *Adenomera* (Amphibia: Anura: Leptodactylidae) from the Atlantic rain forest in Santa Catarina, southern Brazil. *Stud. Neotrop. Fauna Environ.* **44**, 93–107 (2009).
88. Lampert, K. P., Bernal, X. E., Rand, A. S., Mueller, U. G. & Ryan, M. J. Island populations of *Physalaemus pustulosus*: history influences genetic diversity and morphology. *Herpetologica* **63**, 311–319 (2007).
89. Lawlor, T. E. The evolution of body size in mammals: evidence from insular populations in Mexico. *Am. Nat.* **119**, 54–72 (1982).
90. Lee, D. E. et al. Growth, age at maturity, and age-specific survival of the arboreal salamander (*Aneides lugubris*) on Southeast Farallon Island, California. *J. Herpetol.* **46**, 64–71 (2012).
91. Li, Y. et al. Reduced predator species richness drives the body gigantism of a frog species on the Zhoushan Archipelago in China. *J. Anim. Ecol.* **80**, 171–182 (2011).
92. Libois, R. M. & Fons, R. Le mulot des Iles d'Hyères: un cas de gigantisme insulaire. *Vie Milieu* **40**, 217–222 (1990).
93. Lichtenbelt, W. D. V. & Albers, K. B. Reproductive adaptations of the green iguana on a semiarid island. *Copeia* **3**, 790–798 (1993).
94. Lim, B. The distribution, food habits and parasite patterns of the leopard cat (*Prionailurus bengalensis*) in Peninsular Malaysia. *J. Wildl. Parks* **17**, 17–27 (1999).
95. Lin, L.-H., Mao, F., Chen, C. & Ji, X. Reproductive traits of the gray ratsnake *Ptyas korros* from three geographically distinct populations. *Curr. Zool.* **58**, 820–827 (2012).
96. Lindell, L. E., Forsman, A. & Merila, J. Variation in number of ventral scales in snakes: effects on body size, growth rate and survival in the adder, *Vipera berus*. *J. Zool.* **230**, 101–115 (1993).
97. Ljubicavljević, K., Džukić, G., Vukov, T. & Kalezić, M. Morphological variability of the Hermann's tortoise (*Testudo hermanni*) in the central Balkans. *Acta Herpetol.* **7**, 253–262 (2012).
98. Lo Cascio, P. & Corti, C. Indagini sull'ecologia dei rettili sauri della RNO e del SIC 'Isola di Lampedusa'. *Naturalista Sicil.* **32**, 319–354 (2008).
99. Lo Cascio, P. & Pasta, S. Preliminary data on the biometry and the diet of a micro-insular population of *Podarcis wagleriana* (Reptilia: Lacertidae). *Acta Herpetol.* **1**, 147–152 (2006).
100. Lo Valvo, M. & Giacalone, G. Biometrical analyses of a Sicilian green toad, *Bufo siculus* (Stöck et al. 2008), population living in Sicily. *Int. J. Morphol.* **31**, 681–686 (2013).
101. Long, E. S., Courtney, K. L., Lippert, J. C. & Wall-Scheffler, C. M. Reduced body size of insular black-tailed deer is caused by slowed development. *Oecologia* **189**, 675–685 (2019).
102. López-Martin, J., Ruiz-Olmo, J. & Padró, I. Comparison of skull measurements and sexual dimorphism between the Minorcan pine marten (*Martes martes minoricensis*) and the Iberian pine marten (*M. m. martes*): a case of insularity. *Mamm. Biol.* **71**, 13–24 (2006).
103. Lötters, S. et al. Bio-sketches and partitioning of sympatric reed frogs, genus *Hyperolius* (Amphibia: Hyperoliidae), in two humid tropical African forest regions. *J. Nat. Hist.* **38**, 1969–1997 (2004).

104. Luiselli, L., Filippi, E. & Capula, M. Geographic variation in diet composition of the grass snake (*Natrix natrix*) along the mainland and an island of Italy: the effects of habitat type and interference with potential competitors. *Herpetol. J.* **15**, 221–230 (2005).
105. Luiselli, L., Petrozzi, F., Mebert, K., Zuffi, M. A. L. & Amori, G. Resource partitioning and dwarfism patterns between sympatric snakes in a micro-insular Mediterranean environment. *Ecol. Res.* **30**, 527–535 (2015).
106. Luz, A. C., Vicente, L. & Monasterio, C. in *Mainland and Insular Lizards: A Mediterranean Perspective* (eds Corti, C. et al.) 111–123 (Firenze Univ. Press, 2006).
107. Lymberakis, P., Poulakakis, N., Kaliontzopoulou, A., Valakos, E. & Mylonas, M. Two new species of *Podarcis* (Squamata; Lacertidae) from Greece. *Syst. Biodivers.* **6**, 307–318 (2008).
108. Lynch, J., Conroy, J., Kitchener, A., Jefferies, D. & Hayden, T. Variation in cranial form and sexual dimorphism among five European populations of the otter *Lutra lutra*. *J. Zool.* **238**, 81–96 (1996).
109. Lyon, M. W. Jr. Mammals collected in eastern Sumatra by Dr. W.L. Abbott during 1903, 1906, and 1907, with descriptions of new species and subspecies. *Proc. US Natl Mus.* **34**, 619–679 (1908).
110. Lyon, M. W. Mammals of Banka, Mendanau, and Billiton islands, between Sumatra and Borneo. *Proc. US Natl Mus.* **31**, 575–612 (1906).
111. Lyon, M. W. Mammals collected by Dr. W.L. Abbott on Borneo and some of the small adjacent islands. *Proc. US Natl Mus.* **40**, 53–146 (1911).
112. Lyon, M. W. Tree shrews: an account of the mammalian family Tupaiidae. *Proc. US Natl Mus.* **45**, 1–188 (1913).
113. Lyon, M. W. Mammals collected by Dr. W.L. Abbott on the chain of islands lying off the western coast of Sumatra, with descriptions of twenty-eight new species and subspecies. *Proc. US Natl Mus.* **52**, 437–462 (1916).
114. Maddock, S. *Systematics and Phylogeography of Seychelles Amphibians*. PhD thesis, Univ. College London (2016).
115. Madeira, B. D. C. M. A. *Sexual Dimorphism and Reproductive Phenology of Common Birds in São Tomé Island: Conservation Implications*. PhD thesis, Univ. de Lisboa (2018).
116. Madsen, T. & Shine, R. Phenotypic plasticity in body sizes and sexual size dimorphism in European grass snakes. *Evolution* **47**, 321–325 (1993).
117. Mageski, M. et al. The island rule in the Brazilian frog *Phyllodytes luteolus* (Anura: Hylidae): incipient gigantism? *Zoologia* **32**, 329–333 (2015).
118. Magnanou, E., Fons, R., Blondel, J. & Morand, S. Energy expenditure in Crocidurinae shrews (Insectivora): is metabolism a key component of the insular syndrome? *Comp. Biochem. Physiol. A* **142**, 276–285 (2005).
119. Maharadatunkamsi, H. S., Kitchener, D. & Schmitt, L. Relationships between morphology, genetics and geography in the cave fruit bat *Eonycteris spelaea* (Dobson, 1871) from Indonesia. *Biol. J. Linn. Soc.* **79**, 511–522 (2003).
120. Major, R. E. Latitudinal and insular variation in morphology of a small Australian passerine: consequences for dispersal distance and conservation. *Aust. J. Zool.* **60**, 210–218 (2012).
121. Mallick, S., Driessen, M. & Hocking, G. Biology of the southern brown bandicoot (*Isodon obesulus*) in south-eastern Tasmania. II. Demography. *Aust. Mammal.* **20**, 339–347 (1998).
122. Malmquist, M. G. Character displacement and biogeography of the pygmy shrew in northern Europe. *Ecology* **66**, 372–377 (1985).
123. Manier, M. K. Geographic variation in the long-nosed snake, *Rhinocheilus lecontei* (Colubridae): beyond the subspecies debate. *Biol. J. Linn. Soc.* **83**, 65–85 (2004).
124. Manríquez-Morán, N. L., Cruz, M. V.-S. & Mendez-De La Cruz, F. R. Reproductive biology of the parthenogenetic lizard, *Aspidoscelis cozumela*. *Herpetologica* **61**, 435–439 (2005).
125. Marinelli, L. & Millar, J. S. The ecology of beach-dwelling *Peromyscus maniculatus* on the Pacific coast. *Can. J. Zool.* **67**, 412–417 (1989).
126. Martins, M., Araujo, M. S., Sawaya, R. J. & Nunes, R. Diversity and evolution of macrohabitat use, body size and morphology in a monophyletic group of neotropical pitvipers (*Bothrops*). *J. Zool.* **254**, 529–538 (2001).
127. Martins, M., Arnaud, G. & Avila-Villegas, H. Juvenile recruitment, early growth, and morphological variation in the endangered Santa Catalina Island rattlesnake, *Crotalus catalinensis*. *Herpetol. Conserv. Biol.* **7**, 376–382 (2012).
128. Marunouchi, J., Ueda, H. & Ochi, O. Variation in age and size among breeding populations at different altitudes in the Japanese newts, *Cynops pyrrhogaster*. *Amphib.-Reptil.* **21**, 381–396 (2000).
129. Matsui, M. & Ota, H. Parameters of fecundity in *Microhyla ornata* from the Yaeyama group of the Ryukyu Archipelago. *Jpn. J. Herpetol.* **10**, 73–79 (1984).
130. Matsui, M., Shimada, T. & Sudin, A. First record of the tree frog genus *Chiromantis* from Borneo with the description of a new species (Amphibia: Rhacophoridae). *Zool. Sci.* **31**, 45–51 (2014).
131. Matsui, M., Toda, M. & Ota, H. A new species of frog allied to *Fejervarya limnocharis* from the southern Ryukyus, Japan (Amphibia: Ranidae). *Curr. Herpetol.* **26**, 65–79 (2007).
132. Mazák, J. H. & Groves, C. P. A taxonomic revision of the tigers (*Panthera tigris*) of Southeast Asia. *Mamm. Biol.* **71**, 268–287 (2006).
133. McCord, W. P. & Iverson, J. B. A new species of *Ocadia* (Testudines: Batagurinae) from southwestern China. *Proc. Biol. Soc. Wash.* **107**, 52–59 (1994).
134. McCord, W. P. & Thomson, S. A. A new species of *Chelodina* (Testudines: Pleurodira: Chelidae) from northern Australia. *J. Herpetol.* **36**, 255–267 (2002).
135. McFadden, K. W. & Meiri, S. Dwarfism in insular carnivores: a case study of the pygmy raccoon. *J. Zool.* **289**, 213–221 (2013).
136. McLaughlin, J. F. & Roughgarden, J. Avian predation on *Anolis* lizards in the northeastern caribbean: inter-island contrast. *Ecology* **70**, 617–628 (1989).
137. Medway, L. Observations on the fauna of Pulau Tioman and Pulau Tulai. 2. The mammals. *Natl Mus. Singapore Bull.* **34**, 9–32 (1966).
138. Mees, G. F. A. *Systematic Review of the Indo-Australian Zosteropidae (Part III)* (EJ Brill, 1969).
139. Meijaard, E. & Groves, C. A taxonomic revision of the *Tragulus* mouse-deer (Artiodactyla). *Zool. J. Linn. Soc.* **140**, 63–102 (2004).
140. Meijaard, E. & Groves, C. P. Morphometrical relationships between South-east Asian deer (Cervidae, tribe Cervini): evolutionary and biogeographic implications. *J. Zool.* **263**, 179–196 (2004).
141. Meik, J. M., Lawing, A. M. & Pires-daSilva, A. Body size evolution in insular speckled rattlesnakes (Viperidae: *Crotalus mitchellii*). *PLoS ONE* **5**, e9524 (2010).
142. Melo, M. et al. Multiple lines of evidence support the recognition of a very rare bird species: the Príncipe thrush. *J. Zool.* **282**, 120–129 (2010).
143. Melo, M. & O’Ryan, C. Genetic differentiation between Príncipe Island and mainland populations of the grey parrot (*Psittacus erithacus*), and implications for conservation. *Mol. Ecol.* **16**, 1673–1685 (2007).
144. Melton, R. Body size and island *Peromyscus*: a pattern and a hypothesis. *Evol. Theory* **6**, 113–126 (1982).
145. Menzies, J. & Tyler, M. The systematics and adaptations of some Papuan microhylid frogs which live underground. *J. Zool.* **183**, 431–464 (1977).
146. Merriam, C. H. Revision of the shrews of the American genera *Blarina* and *Notiosorex*. *N. Am. Fauna* **10**, 5–34 (1895).
147. Merriam, C. H. Synopsis of the American shrews of the genus *Sorex*. *N. Am. Fauna* **10**, 57–125 (1895).
148. Merriam, C. H. Six new mammals from Cozumel Island, Yucatan. *Proc. Biol. Soc. Wash.* **14**, 99–104 (1901).
149. Merriam, C. H. Descriptions of ten new kangaroo rats. *Proc. Biol. Soc. Wash.* **20**, 75–80 (1907).
150. Miller, C. & Miller, T. Population dynamics and diet of rodents on Rangitoto Island, New Zealand, including the effect of a 1080 poison operation. *New Zeal. J. Ecol.* **19**, 19–27 (1995).
151. Miller, G. S. Mammals collected by Dr. W.L. Abbott in the region of the Indragiri River, Sumatra. *Proc. Acad. Nat. Sci. Phila.* **54**, 143–159 (1902).
152. Miller, G. S. *Catalogue of the Mammals of Western Europe (Europe Exclusive of Russia) in the Collection of the British Museum* (Order of the Trustees of the British Museum, 1912).
153. Millien, V. & Damuth, J. Climate change and size evolution in an island rodent species: new perspectives on the island rule. *Evolution* **58**, 1353–1360 (2004).
154. Molina-Borja, M. & Rodriguez-Dominguez, M. A. Evolution of biometric and life-history traits in lizards (*Gallotia*) from the Canary Islands. *J. Zool. Syst. Evol. Res.* **42**, 44–53 (2004).
155. Monadjem, A., McCleery, R. A. & Collier, B. A. Activity and movement patterns of the tortoise *Stigmochelys pardalis* in a subtropical savanna. *J. Herpetol.* **47**, 237–242 (2013).
156. Montesinos, R., da Silva, H. R. & Gomes de Carvalho, A. L. The ‘island rule’ acting on anuran populations (Bufonidae: *Rhinella ornata*) of the Southern Hemisphere. *Biotropica* **44**, 506–511 (2012).
157. Montgomery, C. E., Boback, S. M., Green, S. E., Paulissen, M. A. & Walker, J. M. *Cnemidophorus lemniscatus* (Squamata: Teiidae) on Cayo Cochino Pequeño, Honduras: extent of island occupancy, natural history, and conservation status. *Herpetol. Conserv. Biol.* **6**, 10–24 (2011).
158. Moratelli, R., Wilson, D. E., Novaes, R. L., Helgen, K. M. & Gutiérrez, E. E. Caribbean *Myotis* (Chiroptera, Vespertilionidae), with description of a new species from Trinidad and Tobago. *J. Mammal.* **98**, 994–1008 (2017).
159. Mori, A. & Hasegawa, M. Early growth of *Elaphe quadrivirgata* from an insular gigantic population. *Curr. Herpetol.* **21**, 43–50 (2002).
160. Mori, A., Ikeuchi, I. & Hasegawa, M. Calling activity of an anuran assemblage in a temporary pond in a dry forest of Madagascar. *Curr. Herpetol.* **34**, 140–148 (2015).
161. Mori, E. et al. Skull shape and Bergmann’s rule in mammals: hints from Old World porcupines. *J. Zool.* **308**, 47–55 (2019).
162. Motokawa, M. Geographic variation in the Japanese white-toothed shrew *Crociodura dsinezumi*. *Acta Theriol.* **48**, 145–156 (2003).
163. Motokawa, M. & Lin, L.-K. Geographic variation in the mole-shrew *Anourosorex squamipes*. *Mammal. Study* **27**, 113–120 (2002).

164. Motokawa, M., Lin, L.-K., Harada, M. & Hattori, S. Morphometric geographic variation in the Asian lesser white-toothed shrew *Crocidura shantungensis* (Mammalia, Insectivora) in East Asia. *Zool. Sci.* **20**, 789–795 (2003).
165. Munks, S. The breeding biology of *Pseudocheirus peregrinus viverrinus* on Flinders Island, Bass Strait. *Wildl. Res.* **22**, 521–533 (1995).
166. MVZ. *Museum of Vertebrate Zoology - Herpetological Collection* (University of California, Berkeley, accessed 20 August 2020); <https://mvz.berkeley.edu/mvzherp/>
167. Myers, S., Brown, G. & Kleindorfer, S. Divergence in New Holland honeyeaters (*Phylidonyris novaehollandiae*): evidence from morphology and feeding behavior. *J. Ornithol.* **151**, 287–296 (2010).
168. Nagorsen, D. W. in *Martens, Sables, and Fishers: Biology and Conservation* (eds Harestad, A. S. et al.), 85–97 (Cornell Univ. Press, 1994).
169. Nagy, Z. T., Glaw, F. & Vences, M. Systematics of the snake genera *Stenophis* and *Lycodyras* from Madagascar and the Comoros. *Zool. Scr.* **39**, 426–435 (2010).
170. Nanova, O. & Prôa, M. Cranial features of mainland and Commander Islands (Russia) Arctic foxes (*Vulpes lagopus*) reflect their diverging foraging strategies. *Polar Res.* **36**, 7 (2017).
171. Napier, P. H. *Catalogue of Primates in the British Museum (Natural History) and Elsewhere in the British Isles. Part III: Family Cercopithecidae, Subfamily Colobinae* Vol. 3 (British Museum (Natural History), 1985).
172. Nelson, E. Descriptions of new birds from the Tres Marias Islands, western Mexico. *Proc. Biol. Soc. Wash.* **12**, 5–11 (1898).
173. Nelson, E. W. The rabbits of North America. *N. Am. Fauna* **29**, 1–314 (1909).
174. Newman, D. G. Activity, dispersion, and population densities of Hamilton's frog (*Leiopelma hamiltoni*) on Maud and Stephens islands, New Zealand. *Herpetologica* **46**, 319–330 (1990).
175. Nguyen, S. T., Oshida, T., Dang, P. H., Bui, H. T. & Motokawa, M. A new species of squirrel (Sciuridae: Callosciurus) from an isolated island off the Indochina Peninsula in southern Vietnam. *J. Mammal.* **99**, 813–825 (2018).
176. Nijman, V. Group composition and monandry in grizzled langurs, *Presbytis comata*, on Java. *Folia Primatol.* **88**, 237–254 (2017).
177. Nor, S. M. *The Mammalian Fauna on the Islands at the Northern Tip of Sabah, Borneo* (Field Museum of Natural History, 1996).
178. Norman, F. & Hurley, V. Gonad measurements and other parameters from Chestnut Teal *Anas castanea* collected in the Gippsland Lakes region, Victoria. *Emu* **84**, 52–55 (1984).
179. Nowak, R. M. & Walker, E. P. *Walker's Mammals of the World* Vol. 1 (JHU, 1999).
180. O'Connell, D. P. et al. A sympatric pair of undescribed white-eye species (Aves: Zosteropidae: Zosterops) with different origins. *Zool. J. Linn. Soc.* **186**, 701–724 (2019).
181. Oates, J. F., Davies, A. G. & Delson, E. in *Colobine Monkeys: Their Ecology, Behaviour and Evolution* (eds Davies, A. G. & Oates, J. F.) 45–73 (Cambridge Univ. Press, 1994).
182. Oh, H.-S., Yoshinaga, Y., Kaneko, T., Iida, H. & Mori, T. Taxomic re-examination of the *Apodemus agrarius chejuensis*, comparing external and cranial morphological characters among four Asian *Apodemus* species. *J. Fac. Agric.* **47**, 373–386 (2003).
183. Ohdachi, S., Abe, H., Oh, H. & Han, S. Morphological relationships among populations in the *Sorex caecutiens/shinto* group (Eulipotyphla, Soricidae) in East Asia, with a description of a new subspecies from Cheju Island, Korea. *Mamm. Biol.* **70**, 345–358 (2005).
184. Okada, S., Izawa, M. & Ota, H. Growth and reproduction of *Gekko hokouensis* (Reptilia: Squamata) on Okinawajima Island of the Ryukyu Archipelago, Japan. *J. Herpetol.* **36**, 473–479 (2002).
185. O'Keeffe, J., O'Boyle, I. & Fogarty, U. *Observations on the Pathology of Tuberculosis in Badgers* Report No. 1898473838 (University College Dublin Centre for Veterinary Epidemiology and Risk Analysis, 1997).
186. Olsen, P., Debus, S., Czechura, G. & Mooney, N. Comparative feeding ecology of the grey goshawk *Accipiter novaehollandiae* and brown goshawk *Accipiter fasciatus*. *Aust. Field Ornithol.* **13**, 178–192 (1990).
187. Oneto, F., Ottonello, D. & Salvidio, S. Primi dati sulla biometria di *Euleptes euopaea* (Genè, 1839) dell'isola del Tino (La Spezia, Liguria). *Doriana* **8**, 1–8 (2008).
188. Oromi, N. et al. Geographical variations in adult body size and reproductive life history traits in an invasive anuran, *Discoglossus pictus*. *Zoology* **119**, 216–223 (2016).
189. Osgood, W. H. Natural history of the Queen Charlotte islands. *N. Am. Fauna* **21**, 1–87 (1901).
190. Osgood, W. H. Revision of the mice of the American genus *Peromyscus*. *N. Am. Fauna* **28**, 1–285 (1909).
191. Ota, H. Taxonomic redefinition of *Japalura swinhonis* Günther (Agamidae: Squamata), with a description of a new subspecies of *J. polygonata* from Taiwan. *Herpetologica* **47**, 280–294 (1991).
192. Ota, H., Lau, M. W., Weidenhofer, T., Yasukawa, Y. & Bogadek, A. Taxonomic review of the geckos allied to *Gekko chinensis* Gray 1842 (Gekkonidae Reptilia) from China and Vietnam. *Trop. Zool.* **8**, 181–196 (1995).
193. Owens, J. R. *Ecology and Behavior of the Bioko Island Drill (Mandrillus leucophaeus poensis)*. PhD thesis, Drexel Univ. (2013).
194. Pabijan, M., Gehring, P.-S., Koehler, J., Glaw, F. & Vences, M. A new microendemic frog species of the genus *Blommersia* (Anura: Mantellidae) from the east coast of Madagascar. *Zootaxa* **2978**, 34–50 (2011).
195. Pafilis, P. et al. Reproductive biology of insular reptiles: marine subsidies modulate expression of the 'island syndrome'. *Copeia* **2011**, 545–552 (2011).
196. Pafilis, P., Kapsalas, G., Lymberakis, P., Protopoulos, D. & Sotiropoulos, K. Diet composition of the Karpathos marsh frog (*Pelophylax cerigensis*): what does the most endangered frog in Europe eat? *Anim. Biodivers. Conserv.* **42**, 1–8 (2019).
197. Pafilis, P., Meiri, S., Foufopoulos, J. & Valakos, E. Intraspecific competition and high food availability are associated with insular gigantism in a lizard. *Naturwissenschaften* **96**, 1107–1113 (2009).
198. Pafilis, P. et al. Body size affects digestive performance in a Mediterranean lizard. *Herpetol. J.* **26**, 199–205 (2016).
199. Pagh, S. et al. Methods for the identification of farm escapees in feral mink (*Neovison vison*) populations. *PLoS ONE* **14**, e0224559 (2019).
200. Pahl, L. & Lee, A. Reproductive traits of 2 populations of the common ringtail possum, *Pseudocheirus peregrinus*, in Victoria. *Aust. J. Zool.* **36**, 83–97 (1988).
201. Palacios, F. & Fernández, J. A new subspecies of hare from Majorca (Balearic Islands). *Mammalia* **56**, 71–86 (1992).
202. Papakosta, M. A. Biometric variation in *Martes foina* from mainland Greece and the Aegean Islands. *Turk. J. Zool.* **41**, 654–663 (2017).
203. Parker, W. S. & Pianka, E. R. Comparative ecology of populations of the lizard *Uta stansburiana*. *Copeia* **1975**, 615–632 (1975).
204. Parrish, G. & Gill, B. Natural history of the lizards of the Three Kings Islands, New Zealand. *New Zeal. J. Zool.* **30**, 205–220 (2003).
205. Pasachnik, S. A. et al. Body size, demography, and body condition in *Utila* spiny-tailed iguanas, *Ctenosaura bakeri*. *Herpetol. Conserv. Biol.* **7**, 391–398 (2012).
206. Pearson, D. et al. Ecological notes on crowned snakes *Elapognathus coronatus* from the Archipelago of the Recherche in southwestern Australia. *Aust. Zool.* **31**, 610–617 (2001).
207. Pedrono, M. & Markwell, T. Maximum size and mass of the ploughshare tortoise, *Gechelone yniphora*. *Chelonian Conserv. Biol.* **4**, 190 (2001).
208. Perez-Ramos, E. et al. *Aspidoscelis sexlineata* (Sauria: Teiidae) in Mexico: distribution, habitat, morphology, and taxonomy. *Southwest. Nat.* **55**, 419–425 (2010).
209. Persson, S. et al. Influence of age, season, body condition and geographical area on concentrations of chlorinated and brominated contaminants in wild mink (*Neovison vison*) in Sweden. *Chemosphere* **90**, 1664–1671 (2013).
210. Peters, J. A. The snakes of the subfamily Dipsadinae. *Misc. Publ. Mus. Zool. Univ. Mich.* **114**, 1–224 (1960).
211. Pocock, R. The civet cats of Asia. Part II. *J. Bombay Nat. Hist. Soc.* **36**, 629–656 (1933).
212. Pocock, R. The geographical races of *Paradoxurus* and *Paguma* found to the east of the Bay of Bengal. *Proc. Zool. Soc. Lond.* **104**, 613–684 (1934).
213. Pocock, R. I. *The Fauna Of British India Including Ceylon And Burma* (Taylor & Francis, 1939).
214. Pons, J. M., Kirwan, G. M., Porter, R. F. & Fuchs, J. A reappraisal of the systematic affinities of Socotran, Arabian and East African scops owls (*Otus*, Strigidae) using a combination of molecular, biometric and acoustic data. *Ibis* **155**, 518–533 (2013).
215. Abe, H. Habitat factors affecting the geographic size variation of Japanese moles. *Mammal. Study* **21**, 71–87 (1996).
216. Abramov, A. V., Jenkins, P. D., Rozhnov, V. V. & Kalinin, A. A. Description of a new species of *Crocidura* (Soricomorpha: Soricidae) from the island of Phu Quoc, Vietnam. *Mammalia* **72**, 269–272 (2008).
217. Abramov, A. V. & Puzachenko, A. Y. Sexual dimorphism of craniological characters in Eurasian badgers, *Meles* spp. (Carnivora, Mustelidae). *Zool. Anz.* **244**, 11–29 (2005).
218. Adams, N. E., Dean, M. D. & Pauly, G. B. Morphological divergence among populations of *Xantusia riversiana*, a night lizard endemic to the Channel Islands of California. *Copeia* **106**, 550–562 (2018).
219. Aguilar-Moreno, M. et al. Dimorfismo sexual de *Aspidoscelis costata costata* (Squamata: Teiidae) en el sur del Estado de México, México. *Rev. Chil. Hist. Nat.* **83**, 585–592 (2010).
220. Ajtici, R. Morphological, biogeographical and ecological characteristics of Kotschy's gecko (*Cyrtodactylus kotschyi* Steindachner, 1870 Gekkonidae) from the mainland portion of its distribution range. *Fauna Balk.* **3**, 1–70 (2014).
221. Albrecht, G. H., Jenkins, P. D. & Godfrey, L. R. Ecogeographic size variation among the living and subfossil prosimians of Madagascar. *Am. J. Primatol.* **22**, 1–50 (1990).

222. Allen, G. M. *The Mammals of China and Mongolia* (American Museum of Natural History, 1938).
223. Allen, J. A. A preliminary study of the South American opossums of the genus *Didelphis*. *Bull. Am. Mus. Nat. Hist.* **16**, 249–279 (1902).
224. Allen, J. A. Mammals collected in Alaska and northern British Columbia by the Andrew J. Stone expedition of 1902. *Bull. Am. Mus. Nat. Hist.* **19**, 521–567 (1903).
225. Allen, J. A. Notes on American deer of the genus *Mazama*. *Bull. Am. Mus. Nat. Hist.* **34**, 521–553 (1915).
226. Allen, J. A. Review of the South American Scuriidae. *Bull. Am. Mus. Nat. Hist.* **34**, 147–309 (1915).
227. Allen, J. A. & Chapman, F. M. On a collection of mammals from the island of Trinidad, with descriptions of new species. *Bull. Am. Mus. Nat. Hist.* **5**, 203–234 (1898).
228. Allen, J. A., Kermode, F. & Andrews, R. C. The white bear of southwestern British Columbia. *Bull. Am. Mus. Nat. Hist.* **26**, 233–238 (1909).
229. Altunisik, A., Kalayci, T. E., Gul, C., Ozdemir, N. & Tosunoglu, M. A skeletochronological study of the smooth newt *Lissotriton vulgaris* (Amphibia: Urodela) from an island and a mainland population in Turkey. *Ital. J. Zool.* **81**, 381–388 (2014).
230. Amor, N. & Farjallah, S. Morphological variation of the African green toad, *Bufo boulengeri* (Amphibia: Anura) in Tunisia. *Pak. J. Zool.* **43**, 921–926 (2011).
231. Anderson, R. P. & Handley, C. O. Dwarfism in insular sloths: biogeography, selection, and evolutionary rate. *Evolution* **56**, 1045–1058 (2002).
232. Andrade, P. et al. Ecomorphological patterns in the Blackcap *Sylvia atricapilla*: insular versus mainland populations. *Bird Study* **62**, 498–507 (2015).
233. Andrews, R. M. Evolution of life histories: a comparison of *Anolis* lizards from matched island and mainland habitats. *Breviora* **454**, 1–51 (1979).
234. Angarita-Sierra, T. & Lynch, J. D. A new species of *Ninia* (Serpentes: Dipsadidae) from Chocó-Magdalena biogeographical province, western Colombia. *Zootaxa* **4244**, 478–492 (2017).
235. Angelici, F., Capizzi, D., Amori, G. & Luiselli, L. Morphometric variation in the skulls of the crested porcupine *Hystrix cristata* from mainland Italy, Sicily, and northern Africa. *Mamm. Biol.* **68**, 165–173 (2003).
236. Aowphol, A., Rujirawan, A., Taksintum, W., Arsirapot, S. & Mcleod, D. S. Re-evaluating the taxonomic status of *Chiromantis* in Thailand using multiple lines of evidence (Amphibia: Anura: Rhacophoridae). *Zootaxa* **3702**, 101–123 (2013).
237. Araújo, M. & Martins, M. The defensive strike of five species of lanceheads of the genus *Bothrops* (Viperidae). *Braz. J. Biol.* **67**, 327–332 (2007).
238. Arendt, W. J. & Arendt, A. L. Bill deformity in a pearly-eyed thrasher from Montserrat, West Indies. *Auk* **54**, 324–332 (1986).
239. Arntzen, J. & García-Paris, M. Morphological and allozyme studies of midwife toads (genus *Alytes*), including the description of two new taxa from Spain. *Contrib. Zool.* **65**, 5–34 (1995).
240. Aubret, F. Island colonisation and the evolutionary rates of body size in insular neonate snakes. *Heredity* **115**, 349–356 (2015).
241. Auer, M. & Taskavak, E. Population structure of syntopic *Emys orbicularis* and *Mauremys rivulata* in western Turkey. *Biologia* **59**, 81–84 (2004).
242. Aumann, T. *Aspects of the Biology of the Brown Goshawk Accipiter Fasciatus in South-Eastern Australia*. MSc thesis, Monash Univ. (1986).
243. Baier, F. & Hoekstra, H. E. The genetics of morphological and behavioural island traits in deer mice. *Proc. R. Soc. B* **286**, 20191697 (2019).
244. Baker-Gabb, D. J. *Comparative Ecology and Behaviour of Swamp Harriers Circus approximans, Spotted Harriers C. assimilis and Other Raptors in Australia and New Zealand* (Royal Australasian Ornithological Union, 1982).
245. Bakhuis, W. L. Size and sexual differentiation in the lizard *Iguana iguana* on a semi-arid island. *J. Herpetol.* **16**, 322–325 (1982).
246. Balham, R. W. Grey and mallard ducks in the Manawatu district, New Zealand. *Emu* **52**, 163–191 (1952).
247. Bangma, J. T. et al. Perfluorinated alkyl acids in plasma of American alligators (*Alligator mississippiensis*) from Florida and South Carolina. *Environ. Toxicol. Chem.* **36**, 917–925 (2017).
248. Bangma, J. T. et al. Variation in perfluoroalkyl acids in the American alligator (*Alligator mississippiensis*) at Merritt Island National Wildlife Refuge. *Chemosphere* **166**, 72–79 (2017).
249. Banks, R. A new insular subspecies of spiny pocket mouse (Mammalia: Rodentia). *Proc. Biol. Soc. Wash.* **80**, 101–104 (1967).
250. Barbo, F. E. et al. Another new and threatened species of lancehead genus *Bothrops* (Serpentes, Viperidae) from Ilha dos Franceses, southeastern Brazil. *Zootaxa* **4097**, 511–529 (2016).
251. Barbour, T. & Allen, G. M. The white-tailed deer of eastern United States. *J. Mammal.* **3**, 65–78 (1922).
252. Bartle, J. & Sagar, P. Intraspecific variation in the New Zealand bellbird *Anthornis melanura*. *Notornis* **34**, 253–306 (1987).
253. Barwick, R. E. *Studies on the Scincid Lizard Egernia cunninghami* (Gray, 1832). PhD thesis, Australian National Univ. (1965).
254. Başkale, E., Ulubeli, S. A. & Kaska, Y. Age structures and growth parameters of the Levantine frog, *Pelophylax bedriagae*, at different localities in Denizli, Turkey. *Acta Herpetol.* **13**, 147–154 (2018).
255. Bee, J. W. & Hall, E. R. *Mammals of Northern Alaska on the Arctic Slope* (Museum of Natural History, University of Kansas, 1956).
256. Bejakovic, D., Kalezic, M. L., Aleksic, I., Dzucic, G. & Crnobrnjašailovic, J. Female reproductive cycle and clutch traits in the Dalmatian wall lizard (*Podarcis melisellensis*). *Folia Zool.* **44**, 371–380 (1995).
257. Bell, B. D. Observations on the ecology and reproduction of the New Zealand leiopelmid frogs. *Herpetologica* **34**, 340–354 (1978).
258. Bell, R. C. & Irian, C. G. Phenotypic and genetic divergence in reed frogs across a mosaic hybrid zone on São Tomé Island. *Biol. J. Linn. Soc.* **128**, 672–680 (2019).
259. Bennett, D. & Hampson, K. Further Observations of *Varanus olivaceus* on the Polillo Islands. In *Wildlife and Conservation in the Polillo Islands* Polillo Project Final Report, Multimedia CD (eds Hampson, K. et al.) (Viper Press, 2003).
260. Benson, S. B. Two new pocket mice: genus *Perognathus*, from the Californias. *Univ. Calif. Publ. Zool.* **32**, 449–454 (1930).
261. Bentz, E. J., Rodríguez, M. R., John, R. R., Henderson, R. W. & Powell, R. Population densities, activity, microhabitats, and thermal biology of a unique crevice- and litter-dwelling assemblage of reptiles on Union Island, St. Vincent and the Grenadines. *Herpetol. Conserv. Biol.* **6**, 40–50 (2011).
262. Beovides-Casas, K. & Mancina, C. A. Natural history and morphometry of the Cuban iguana (*Cyclura nublila* Gray, 1831) in Cayo Siju, Cuba. *Anim. Biodivers. Conserv.* **29**, 1–8 (2006).
263. Berardo, F. *Habitat Preferenziali e Dinamica di Popolazione di Testudo hermanni Nelle Aree Costiere del Molise*. PhD thesis, Univ. Molise (2015).
264. Berry, R. The evolution of an island population of the house mouse. *Evolution* **18**, 468–483 (1964).
265. Berry, R., Jakobson, M. & Peters, J. The house mice of the Faroe Islands: a study in microdifferentiation. *J. Zool.* **185**, 73–92 (1978).
266. Bertolero, A. in *Enciclopedia Virtual de los Vertebrados Españoles* (eds Salvador, A. & Marco, A.) (Museo Nacional de Ciencias Naturales, 2015); <http://www.vertebradosibericos.org/>
267. Bertolero, A., Cheylan, M. & Nougarede, J.-P. Accroissement de la fécondité chez la tortue d'Hermann *Testudo hermanni hermanni* en condition insulaire: un contre-exemple du syndrome insulaire? *Rev. Ecol.* **62**, 93–98 (2007).
268. Bischoff, W. Bemerkungen zur innerartlichen Variabilität von *Gallotia atlantica* (Peters & Doria, 1882) (Lacertidae). *Bonn. Zool. Beitr.* **36**, 489–506 (1985).
269. Bishop, C. A. & Rouse, J. D. Polychlorinated biphenyls and organochlorine pesticides in plasma and the embryonic development in Lake Erie water snakes (*Nerodia sipedon insularum*) from Pelee Island, Ontario, Canada (1999). *Arch. Environ. Contam. Toxicol.* **51**, 452–457 (2006).
270. Blackburn, D. C. A new squeaker frog (Arthroleptidae: Arthroleptis) from Bioko island, Equatorial Guinea. *Herpetologica* **66**, 320–334 (2010).
271. Blondel, J., Perret, P., Anstett, M. C. & Thebaud, C. Evolution of sexual size dimorphism in birds: test of hypotheses using blue tits in contrasted Mediterranean habitats. *J. Evol. Biol.* **15**, 440–450 (2002).
272. Boback, S. M. A morphometric comparison of island and mainland boas (*Boa constrictor*) in Belize. *Copeia* **2006**, 261–267 (2006).
273. Bosc, V. *Inventaire du Phyllocladyle d'Europe Euleptes europaea sur l'îlot de Roscana (Corse du Sud)* (Conservatoire des Espaces Naturels de Corse, 2009).
274. Boye, P. Notes on the morphology, ecology and geographic origin of the Cyprus long-eared hedgehog (*Hemiechinus auritus dorotheae*). *Bonn. Zool. Beitr.* **42**, 115–123 (1991).
275. Braithwaite, L. & Miller, B. The mallard, *Anas platyrhynchos*, and mallard-black duck, *Anas superciliosa rogersi*, hybridization. *Wildl. Res.* **2**, 47–61 (1975).
276. Brandon-Jones, D. The taxonomic affinities of the Mentawai Islands Sureli, *Presbytis potenziani* (Bonaparte, 1856) (Mammalia: Primata: Cercopithecidae). *Raffles Bull. Zool.* **41**, 331–357 (1993).
277. Brasileiro, C. A., Haddad, C. E., Sawaya, R. J. & Sazima, I. A new and threatened island-dwelling species of *Cycloramphus* (Anura: Cycloramphidae) from southeastern Brazil. *Herpetologica* **63**, 501–510 (2007).
278. Brecko, J. et al. Functional and ecological relevance of intraspecific variation in body size and shape in the lizard *Podarcis melisellensis* (Lacertidae). *Biol. J. Linn. Soc.* **94**, 251–264 (2008).
279. Brisbin, I. L. Jr & Lenarz, M. S. Morphological comparisons of insular and mainland populations of southeastern white-tailed deer. *J. Mammal.* **65**, 44–50 (1984).
280. Brown, J. L., Maan, M. E., Cummings, M. E. & Summers, K. Evidence for selection on coloration in a Panamanian poison frog: a coalescent-based approach. *J. Biogeogr.* **37**, 891–901 (2010).
281. Bruschi, S. et al. Comments on the status of the Sardinian–Corsican lacertid lizard *Podarcis tiliguerta*. *Proc. Calif. Acad. Sci.* **57**, 225–245 (2006).

282. Buden, D. W. Morphological variation and distributional ecology of the giant Micronesian gecko (*Perochirus scutellatus*) of Kapingamarangi Atoll. *Pacif. Sci.* **52**, 250–258 (1998).
283. Burt, W. H. Descriptions of heretofore unknown mammals from islands in the Gulf of California, Mexico. *Trans. San Diego Soc. Nat. Hist.* **16**, 161–185 (1932).
284. Butynski, T. M., de Jong, Y. A. & Hearn, G. W. Body measurements for the monkeys of Bioko Island, Equatorial Guinea. *Primate Conserv.* **24**, 99–105 (2009).
285. Cabot, J. & Urdiales, C. The subspecific status of Sardinian warblers *Sylvia melanocephala* in the Canary Islands with the description of a new subspecies from Western Sahara. *Bull. Br. Ornithol. Club* **125**, 230–240 (2005).
286. Camerano, L. Ricerche intorno alla variazione del *Phyllodactylus europaeus* Gené. *Bull. Mus. Zool. Anat. Comp. R. Univ. Torino* **19**, 1–28 (1904).
287. Camps, D. in *Enciclopedia Virtual de los Vertebrados Españoles* (eds Salvador, A. & Barja, I.) (Museo Nacional de Ciencias Naturales, 2017); <http://www.vertebradosibericos.org/>
288. Capula, M. et al. in *Scripta Herpetologica: Studies on Amphibians and Reptiles in Honour of Benedetto Lanza* (eds Capula, M. & Corti, C.) 39–47 (Edizioni Belvedere, 2014).
289. Carbone, M. *Caratteristiche della Popolazione di Testudo hermanni Gmelin del Parco Naturale della Maremma*. MSc thesis, Univ. Genova (1988).
290. Carrascal, L. M., Moreno, E. & Valido, A. Morphological evolution and changes in foraging behaviour of island and mainland populations of blue tit (*Parus caeruleus*)—a test of convergence and ecomorphological hypotheses. *Evol. Ecol.* **8**, 25–35 (1994).
291. Carretero, M. & Llorente, G. Morphometry in a community of Mediterranean lacertid lizards, and its ecological relationships. *Hist. Anim.* **2**, 77–99 (1993).
292. Case, T. J., Cody, M. L. & Ezcurra, E. *A New Island Biogeography of the Sea of Cortés* (Oxford Univ. Press, 2002).
293. Case, T. J. & Schwaner, T. D. Island/mainland body size differences in Australian varanid lizards. *Oecologia* **94**, 102–109 (1993).
294. Castellano, S., Rosso, A., Doglio, S. & Giacoma, C. Body size and calling variation in the green toad (*Bufo viridis*). *J. Zool.* **248**, 83–90 (1999).
295. Castilla, A. M. & Bauwens, D. Reproductive characteristics of the lacertid lizard *Podarcis atrata*. *Copeia* **2000**, 748–756 (2000).
296. Catzeflis, F., Maddalena, T., Hellwing, S. & Vogel, P. Unexpected findings on the taxonomic status of east Mediterranean *Crocodyrus russula* auct. (Mammalia, Insectivora). *Z. Säugetierkd.* **50**, 185–201 (1985).
297. Celis-Diez, J. L. et al. Population abundance, natural history, and habitat use by the arboreal marsupial *Dromiciops gliroides* in rural Chiloé Island, Chile. *J. Mammal.* **93**, 134–148 (2012).
298. Ceriaco, L. M. et al. Description of a new endemic species of shrew (Mammalia, Soricomorpha) from Príncipe Island (Gulf of Guinea). *Mammalia* **79**, 325–341 (2015).
299. Chamberlain, J. The Block Island meadow mouse, *Microtus proventus*. *J. Mammal.* **35**, 587–589 (1954).
300. Charles-Dominique, P. Urine marking and territoriality in *Galago alleni* (Waterhouse, 1837—Lorisioidea, Primates)—a field study by radio-telemetry. *Z. Tierpsychol.* **43**, 113–138 (1977).
301. Chasen, F. & Kloss, C. B. On a collection of mammals from the lowlands and islands of North Borneo. *Bull. Raffles Mus.* **6**, 1–82 (1931).
302. Chen, S.-L. et al. Taxonomic status of the Korean populations of the genus *Scincella* (Squamata: Scincidae). *J. Herpetol.* **35**, 122–129 (2001).
303. Chermel, J. J., Olimpio, J. & Ximenez, A. Descrição de uma nova espécie do gênero *Cavia* Pallas, 1766 (Mammalia-Caviidae) das Ilhas dos Moleques do Sul, Santa Catarina, Sul do Brasil. *Biotemas* **12**, 95–117 (1999).
304. Chondropoulos, B. P. & Lykakis, J. J. Ecology of the Balkan wall lizard, *Podarcis taurica ionica* (Sauria: Lacertidae) from Greece. *Copeia* **1983**, 991–1001 (1983).
305. Clark, R. J. Herpetofauna of the islands of the Argo-Saronic Gulf, Greece. *Proc. Calif. Acad. Sci.* **35**, 23–36 (1967).
306. Clegg, S. M. et al. Microevolution in island forms: the roles of drift and directional selection in morphological divergence of a passerine bird. *Evolution* **56**, 2090–2099 (2002).
307. Clover, R. C. Phenetic relationships among populations of *Podarcis sicula* and *P. melisellensis* (Sauria: Lacertidae) from islands in the Adriatic Sea. *Syst. Biol.* **28**, 284–298 (1979).
308. Cole, C. J., Dessauer, H. C., Townsend, C. R. & Arnold, M. G. Unisexual lizards of the genus *Gymnophthalmus* (Reptilia, Teiidae) in the Neotropics: genetics, origin, and systematics. *Am. Mus. Novit.* **2994**, 1–29 (1990).
309. Colyn, M. Données pondérales sur les primates Cercopitheciidae d'Afrique Centrale (Bassin du Zaïre/Congo). *Mammalia* **58**, 483–488 (1994).
310. Copley, P., Read, V., Robinson, A. & Watts, C. in *Bandicoots and Bilbies* (eds Seebeck, J. H. et al.) 345–356 (Surrey Beatty & Sons Pty Ltd, 1990).
311. Cordero Rivera, A., Velo-Antón, G. & Galán, P. Ecology of amphibians in small coastal Holocene islands: local adaptations and the effect of exotic tree plantations. *Munibe* **25**, 94–103 (2007).
312. Corti, C. & Zuffi, M. Aspects of population ecology of *Testudo hermanni hermanni* from Asinara Island, NW Sardinia (Italy, western Mediterranean Sea): preliminary data. *Amphib.-Reptil.* **24**, 441–447 (2003).
313. Cowan, I. M. Insularity in the genus *Sorex* on the north coast of British Columbia. *Proc. Biol. Soc. Wash.* **54**, 95–107 (1941).
314. Cox, D. T. et al. Patterns of seasonal and yearly mass variation in West African tropical savannah birds. *Ibis* **153**, 672–683 (2011).
315. Creer, S., Chou, W. H., Malhotra, A. & Thorpe, R. S. Offshore insular variation in the diet of the Taiwanese bamboo viper *Trimeresurus stejnegeri* (Schmidt). *Zool. Sci.* **19**, 907–913 (2002).
316. Cruz-Elizalde, R. et al. Sexual dimorphism and natural history of the western Mexico whiptail, *Aspidoscelis costata* (Squamata: Teiidae), from Isla Isabel, Nayarit, Mexico. *North-West J. Zool.* **10**, 374–381 (2014).
317. Cumbo, V., Licata, F., Mercurio, E., Anz, S. & Lo Valvo, M. in *VIII Congresso Nazionale Societas Herpetologica Italica* (eds Di Cerbo, A. R. et al.) 401–404 (Ianieri Edizioni, 2010).
318. D'Angelo, S., Galia, F. & Lo Valvo, M. Biometric characterization of two Sicilian pond turtle (*Emys trinacris*) populations of south-western Sicily. *Rev. Esp. Herpetol.* **22**, 15–22 (2008).
319. Dagosto, M., Gebo, D. L. & Dolino, C. Positional behavior and social organization of the Philippine tarsier (*Tarsius syrichta*). *Primates* **42**, 233–243 (2001).
320. Darevsky, I. Two new species of the worm-like lizard *Dibamus* (Sauria, Dibamidae), with remarks on the distribution and ecology of *Dibamus* in Vietnam. *Asiatic Herpetol. Res.* **4**, 1–12 (1992).
321. David, P., Vidal, N. & Pauwels, O. S. A morphological study of Stejneger's pitviper *Trimeresurus stejnegeri* (Serpentes, Viperidae, Crotalinae), with the description of a new species from Thailand. *Russian J. Herpetol.* **8**, 205–222 (2001).
322. De la Cruz, J. O. & Casas, E. V. Jr Captive observations and comparative morphology of Philippine tarsier (*Carlito syrichta*) in Brgy. Hugpa, Biliran, Biliran: a preliminary study. *Philipp. J. Nat. Sci.* **20**, 46–54 (2015).
323. Decker, D. Systematics of the coatis, genus *Nasua* (Mammalia: Procyonidae). *Proc. Biol. Soc. Wash.* **104**, 370–386 (1991).
324. Delany, M. Variation in the long-tailed field-mouse (*Apodemus sylvaticus* (L.)) in north-west Scotland I. Comparisons of individual characters. *Proc. R. Soc. B* **161**, 191–199 (1964).
325. Delany, M. & Healy, M. Variation in the white-toothed shrews (*Crocodyrus* spp.) in the British Isles. *Proc. R. Soc. B* **164**, 63–74 (1966).
326. Delany, M. & Healy, M. Variation in the long-tailed field-mouse (*Apodemus sylvaticus* (L.)) in the Channel Islands. *Proc. R. Soc. B* **166**, 408–421 (1967).
327. Delaugerre, M. La variation géographique chez *Phyllodactylus europaeus* Gené (Reptilia, Sauria, Gekkonidae). Etude de la population de l'îlot Sperduto Grande (Sud de la Corse, réserve naturelle des Iles Lavezzi). *Publ. Soc. Linn. Lyon* **54**, 262–269 (1985).
328. Delaugerre, M. *Le Phyllodactyle d'Europe sur l'île de Port-Cros: mise en place d'un suivi géographique; la population de la Gabinière* (Parc national de Port-Cros, 2003).
329. Delaugerre, M. & Ouni, R. *Archipel de la Galite «Notes herpétologiques 2008»* (PIM Initiative, 2009).
330. Delaugerre, M. & Dubois, A. La variation géographique et la variabilité intrapopulationnelle chez *Phyllodactylus europaeus* (Reptilia, Sauria, Gekkonidae). *Bull. Mus. Natl. Hist. Nat.* **7**, 709–736 (1985).
331. Delibes de Castro, M. Sobre las ginetas de la Isla de Ibiza (*Genetta genetta isabellae* n. ssp.). *Doñana Acta Vertebr.* **4**, 139–160 (1977).
332. Delibes, M. & Amores, F. The stone marten *Martes foina* (Erxleben, 1777) (Mammalia, Carnivora) from Ibiza (Pitiusic, Balearic Islands). *Misc. Zool.* **10**, 335–345 (1986).
333. Delson, E., Terranova, C. J., Jungers, W. L., Sargis, E. J. & Jablonski, N. G. Body mass in Cercopitheciidae (Primates, Mammalia): estimation and scaling in extinct and extant taxa. *Anthropol. Pap. Am. Mus. Nat. Hist.* **83**, 1–159 (2000).
334. Dietzen, C., Garcia-del-Rey, E., Castro, G. D. & Wink, M. Phylogeography of the blue tit (*Parus teneriffae*-group) on the Canary Islands based on mitochondrial DNA sequence data and morphometrics. *J. Ornithol.* **149**, 1–12 (2008).
335. Djong, H. T., Matsui, M., Kuramoto, M., Nishioka, M. & Sumida, M. A new species of the *Fejervarya limnocharis* complex from Japan (Anura, Dicroglossidae). *Zool. Sci.* **28**, 922–929 (2011).
336. Dodd, C. K. Jr Population structure and the evolution of sexual size dimorphism and sex ratios in an insular population of Florida box turtles (*Terrapene carolina bauri*). *Can. J. Zool.* **75**, 1495–1507 (1997).
337. Donihue, C. M., Brock, K. M., Fougopoulos, J. & Herrel, A. Feed or fight: testing the impact of food availability and intraspecific aggression on the functional ecology of an island lizard. *Funct. Ecol.* **30**, 566–575 (2016).
338. Driessen, M. M. & Rose, R. K. *Isoodon obesulus* (Peramelemorphia: Peramelidae). *Mamm. Species* **47**, 112–123 (2015).

339. Du, W.-G., Ji, X. & Zhang, Y.-P. Inter-population variation in life-history traits of a Chinese lizard (*Takydromus septentrionalis*, Lacertidae). *Herpetol. J.* **16**, 233–237 (2006).
340. Du, W.-G., Ji, X., Zhang, Y.-P., Xu, X.-F. & Shine, R. Identifying sources of variation in reproductive and life-history traits among five populations of a Chinese lizard (*Takydromus septentrionalis*, Lacertidae). *Biol. J. Linn. Soc.* **85**, 443–453 (2005).
341. Dumbell, G. S. *The Ecology, Behaviour and Management of New Zealand Brown Teal, or Pateke (Anas aucklandica chlorotis)*. PhD thesis, Univ. Auckland (1987).
342. Dunham, A. E., Tinkle, D. W. & Gibbons, J. W. Body size in island lizards: a cautionary tale. *Ecology* **59**, 1230–1238 (1978).
343. Dunn, E. R. & Saxe, L. Jr. Results of the Catherwood-Chaplin West Indies expedition, 1948. Part V. Amphibians and reptiles of San Andrés and Providencia. *Proc. Acad. Nat. Sci. Phila.* **102**, 141–165 (1950).
344. Dunstone, N. et al. Uso del hábitat, actividad y dieta de la guíña (*Oncifelis guigna*) en el Parque Nacional Laguna San Rafael, XI Región, Chile. *B. Mus. Nac. Hist. Nat.* **51**, 147–158 (2002).
345. Elton, S. & Morgan, B. J. Muzzle size, paranasal swelling size and body mass in *Mandrillus leucophaeus*. *Primates* **47**, 151–157 (2006).
346. Esteves, R. G. *Comportamento Alimentar e Aspectos Reprodutivos de Bothrops insularis (Amaral, 1921) (Serpentes: Viperidae), na Criação Ex-Situ no Instituto Vital Brazil*. MSc thesis, Univ. Federal do Estado do Rio de Janeiro (2016).
347. Faaborg, J. & Winters, J. E. Winter resident returns and longevity and weights of Puerto Rican birds. *Bird-Band* **50**, 216–223 (1979).
348. Fang, Y.-P. & Lee, L.-L. Re-evaluation of the Taiwanese white-toothed shrew, *Crociodura tadae* Tokuda and Kano, 1936 (Insectivora: Soricidae) from Taiwan and two offshore islands. *J. Zool.* **257**, 145–154 (2002).
349. Faraone, F. P. *Indagini Sulla Variazione Fenotipica in Ambienti Insulari e Microinsulari delle Specie di Podarcis Wagler, 1820 (Reptilia, Lacertidae) Presenti in Sicilia e in Alcune Isole Circumsciliane*. PhD thesis, Univ. Palermo (2011).
350. Faraone, F. P., Giacalone, G. & Lo Valvo, M. in *Atti VIII Congresso Nazionale Societas Herpetologica Italica* (eds Di Cerbo, A. R. et al.) 247–252 (Ianeri Edizioni, 2010).
351. Fitch, H. S. Variation in clutch and litter size in New World reptiles. *Univ. Kans. Mus. Nat. Hist. Misc. Publ.* **76**, 1–76 (1985).
352. Fooden, J. Systematic review of the rhesus macaques, *Macaca mulatta* (Zimmermann, 1780). *Fieldiana Zool.* **96**, 1–180 (2000).
353. Fooden, J. & Albrecht, G. H. Latitudinal and insular variation of skull size in crab-eating macaques (Primates, Cercopithecidae: *Macaca fascicularis*). *Am. J. Phys. Anthropol.* **92**, 521–538 (1993).
354. Ford, H. The honeyeaters of Kangaroo Island. *S. Aust. Ornithol.* **27**, 134–138 (1976).
355. Fornasiero, S., Corti, C., Luiselli, L. & Zuffi, M. A. Sexual size dimorphism, morphometry and phenotypic variation in the whip snake *Hierophis viridiflavus* from a central Mediterranean area. *Rev. Ecol.* **62**, 73–85 (2007).
356. Forsman, A. Variation in sexual size dimorphism and maximum body size among adder populations: effects of prey size. *J. Anim. Ecol.* **60**, 253–267 (1991).
357. Forsman, A., Merila, J. & Ebenhard, T. Phenotypic evolution of dispersal-enhancing traits in insular voles. *Proc. R. Soc. B* **278**, 225–232 (2011).
358. Foster, J. *The Evolution of the Native Land Mammals of the Queen Charlotte Islands and the Problem of Insularity*. PhD thesis, Univ. British Columbia (1963).
359. Fox, N. C. Some morphological data on the Australasian harrier (*Circus approximans gouldi*) in New Zealand. *Notornis* **24**, 9–19 (1977).
360. Franco, M., Quijano, A. & Soto-Gamboa, M. Communal nesting, activity patterns, and population characteristics in the near-threatened monito del monte, *Dromiciops gliroides*. *J. Mammal.* **92**, 994–1004 (2011).
361. Fukada, H. Growth and maturity of the Japanese rat snake, *Elaphe climacophora* (Reptilia, Serpentes, Colubridae). *J. Herpetol.* **12**, 269–274 (1978).
362. Galán, P. Reproductive characteristics of an insular population of the lizard *Podarcis hispanica* from northwest Spain (Cies Islands, Galicia). *Copeia* **2003**, 657–665 (2003).
363. Ganem, G., Granjon, L., Ba, K. & Duplantier, J. M. Body size variability and water balance - a comparison between mainland and island populations of *Mastomys huberti* (Rodentia, Muridae) in Senegal. *Experientia* **51**, 402–410 (1995).
364. Garcés-Restrepo, M. F., Giraldo, A. & Carr, J. L. Population ecology and morphometric variation of the Chococo river turtle (*Rhinoclemmys nasuta*) from two localities on the Colombian Pacific coast. *Bol. Cient. Mus. Hist. Nat. Univ. Caldas* **17**, 160–171 (2013).
365. Garcia-Porta, J., Smid, J., Sol, D., Fasola, M. & Carranza, S. Testing the island effect on phenotypic diversification: insights from the *Hemidactylus* geckos of the Socotra Archipelago. *Sci. Rep.* **6**, 23729 (2016).
366. Gaulke, M. Overview on the present knowledge on *Varanus mabitang* Gaulke and Curio, 2001, including new morphological and meristic data. *Biawak* **4**, 50–58 (2010).
367. Geissmann, T. *Evolution of Communication in Gibbons (Hylobatidae)*. PhD thesis, Univ. Zürich (1993).
368. Giacalone, G., Abbate, M., Fritz, U., & Lo Valvo, M. Preliminary data on distribution, morphometric and genetic characterization of Hermann's tortoise in Sicily. In *Herpetologia Sardiniae 282–286* (LATINA: Edizioni Belvedere, 2008).
369. Giagia, E. V. *Cytotaxonomical Study of Eastern European Hedgehog *Erinaceus concolor* M. in Greece*. PhD thesis, Univ. Patras (1977).
370. Gil Escobedo, L. J. *Ámbitos de Hogar de la Iguana de Órgano *Ctenosaura palearis* (Sauria: Iguanidae) en el Bosque Tropical Estacionalmente Seco de Cabañas, Zacapa, Guatemala*. BSc thesis, Univ. San Carlos de Guatemala (2016).
371. Gill, B. & McLean, I. G. Morphometrics of the whitehead *Mohoua albicilla* on Little Barrier Island, New Zealand. *New Zeal. J. Zool.* **13**, 267–271 (1986).
372. Glaw, F., Hawlitschek, O., Glaw, K. & Vences, M. Integrative evidence confirms new endemic island frogs and transmarine dispersal of amphibians between Madagascar and Mayotte (Comoros Archipelago). *Sci. Nat.* **106**, 19 (2019).
373. Glaw, F., Koepler, J., Townsend, T. M. & Vences, M. Rivaling the world's smallest reptiles: discovery of miniaturized and microendemic new species of leaf chameleons (*Brookesia*) from northern Madagascar. *PLoS ONE* **7**, e31314 (2012).
374. Glaw, F., Nagy, Z. T., Franzen, M. & Vences, M. Molecular phylogeny and systematics of the pseudoxyrhopiine snake genus *Liopholidophis* (Reptilia, Colubridae): evolution of its exceptional sexual dimorphism and descriptions of new taxa. *Zool. Scr.* **36**, 291–300 (2007).
375. Glaw, F., Vences, M., Andreone, F. & Vallan, D. Revision of the *Boophis majori* group (Amphibia: Mantellidae) from Madagascar, with descriptions of five new species. *Zool. J. Linn. Soc.* **133**, 495–529 (2001).
376. Goldberg, S. R., Bursey, C. R. & Arreola, J. Gastrointestinal helminths from eight species of *Aspidoscelis* (Squamata: Teiidae) from Mexico. *West. N. Am. Nat.* **74**, 223–227 (2014).
377. Goldman, E. A. & Jackson, H. H. Raccoons of North and Middle America. *N. Am. Fauna* **60**, 1–153 (1950).
378. Goltsman, M., Kruchenkova, E. P., Sergeev, S., Volodin, I. & Macdonald, D. W. 'Island syndrome' in a population of Arctic foxes (*Alopex lagopus*) from Mednyi Island. *J. Zool.* **267**, 405–418 (2005).
379. González Quintero, E. P. *Análisis Taxonómico del Coyote (Canis latrans) de la Península de Baja California, México*. MSc thesis, Centro de Investigaciones Biológicas del Noroeste S.C (2004).
380. González Rossell, A. *Ecología y Conservación de la Iguana (Cyclura nubila nubila) en Cuba*. PhD thesis, Univ. Alicante (2018).
381. Granjon, L. & Cheylan, G. Biometric differentiation of black rat (*Rattus rattus*) populations in the west Mediterranean islands. *Mammalia* **54**, 213–231 (1990).
382. Grant, P. *A Systematic Study of the Terrestrial Birds of the Tres Marias Islands, Mexico* (Yale Peabody Museum of Natural History, 1965).
383. Grant, P. The coexistence of two wren species of the genus *Thryothorus*. *Wilson Bull.* **78**, 266–278 (1966).
384. Grant, P. *Further Information on the Relative Length of the Tarsus in Land Birds* (Yale Peabody Museum of Natural History, 1966).
385. Grant, P. Ecological and morphological variation of Canary Island blue tits, *Parus caeruleus* (Aves: Paridae). *Biol. J. Linn. Soc.* **11**, 103–129 (1979).
386. Grinnell, J. The species of the mammalian genus *Sorex* of west-central California. *Univ. Calif. Publ. Zool.* **10**, 179–195 (1913).
387. Grinnell, J., Dixon, J. S. & Linsdale, J. M. *The Fur Bearing Mammals of California* (Univ. California Press, 1937).
388. Guarino, F. M., Garcia, G. & Andreone, F. Huge but moderately long-lived: age structure in the mountain chicken, *Leptodactylus fallax*, from Montserrat, West Indies. *Herpetol. J.* **24**, 167–173 (2014).
389. Gursky, S. Effects of radio transmitter weight on a small nocturnal primate. *Am. J. Primatol.* **46**, 145–155 (1998).
390. Gutsche, A. & Streich, W. J. Demography and endangerment of the Utila Island spiny-tailed iguana, *Ctenosaura bakeri*. *J. Herpetol.* **43**, 105–113 (2009).
391. Hadi, S., Ziegler, T. & Hodges, J. K. Group structure and physical characteristics of simakobu monkeys (*Simias concolor*) on the Mentawai Island of Siberut, Indonesia. *Folia Primatol.* **80**, 74–82 (2009).
392. Hagen, B. Zur Kleinsäugerfauna Siziliens. *Bonn. Zool. Beitr.* **5**, 1–15 (1954).
393. Hai, B. T., Tu, L. N., Duong, V. T. & Son, N. T. Geographic variation in skull size and shape of *Crociodura dracula* (Mammalia: Soricidae) in Vietnam. In *Proc. 7th National Scientific Conference on Ecology and Biological Resources 670–677* (2017).
394. Harding, L. E. *Trachypithecus cristatus* (Primates: Cercopithecidae). *Mamm. Species* **42**, 149–165 (2010).

395. Hasegawa, M. Insular radiation in life-history of the lizard *Eumeces okadae* in the Izu Islands, Japan. *Copeia* **1994**, 732–747 (1994).
396. Hasegawa, M. & Moriguchi, H. in *Current Herpetology in East Asia* (eds Matui, M. et al.) 414–432 (Herpetological Society of Japan, 1989).
397. Hawlitschek, O., Nagy, Z. T. & Glaw, F. Island evolution and systematic revision of Comoran snakes: why and when subspecies still make sense. *PLoS ONE* **7**, e42970 (2012).
398. Heaney, L. R. Island area and body size of insular mammals: evidence from the tri-colored squirrel (*Callosciurus prevosti*) of Southeast Asia. *Evolution* **32**, 29–44 (1978).
399. Heaney, L. R. & Timm, R. M. Systematics and distribution of shrews of the genus *Crocidura* (Mammalia: Insectivora) in Vietnam. *Proc. Biol. Soc. Wash.* **96**, 115–120 (1983).
400. Hedges, S. B. & Thomas, R. At the lower size limit in amniote vertebrates: a new diminutive lizard from the West Indies. *Caribb. J. Sci.* **37**, 168–173 (2001).
401. Heinsohn, G. E. *Ecology and Reproduction of the Tasmanian Bandicoots (Perameles gunni and Isoodon obesulus)* (Univ. California Press, 1966).
402. Hemelaar, A. Age, growth and other population characteristics of *Bufo bufo* from different latitudes and altitudes. *J. Herpetol.* **22**, 369–388 (1988).
403. Heo, J.-H. et al. Can an invasive prey species induce morphological and behavioral changes in an endemic predator? Evidence from a South Korean snake (*Oocatochus rufodoratus*). *Asian Herpetol. Res.* **5**, 245–254 (2014).
404. Hernández-Gallegos, O., López Moreno, A. E., Méndez de la Cruz, F. R. & Walker, J. Home range of the parthenogenetic lizard *Aspidoscelis maslini* (Fritts, 1969), on a beach strand. *Herpetozoa* **31**, 83–86 (2018).
405. Hernández-Gallegos, O., López-Moreno, A. E., Méndez-Sánchez, J. F., Lloyd Rheubert, J. & Méndez-de la Cruz, F. R. Home range of *Aspidoscelis cozumela* (Squamata: Teiidae): a parthenogenetic lizard microendemic to Cozumel Island, Mexico. *Rev. Biol. Trop.* **63**, 771–781 (2015).
406. Hernandez-Salinas, U., Ramirez-Bautista, A., Pavon, N. P. & Rosas Pacheco, L. F. Morphometric variation in island and mainland populations of two lizard species from the Pacific coast of Mexico. *Rev. Chil. Hist. Nat.* **87**, 1–9 (2014).
407. Herrel, A., Cottam, M. D., Godbeer, K., Sanger, T. & Losos, J. B. An ecomorphological analysis of native and introduced populations of the endemic lizard *Anolis maynardi* of the Cayman Islands. *Breviora* **522**, 1–10 (2011).
408. Herrel, A., Spithoven, L., Van Damme, R. & De Vree, F. Sexual dimorphism of head size in *Gallotia galloti*: testing the niche divergence hypothesis by functional analyses. *Funct. Ecol.* **13**, 289–297 (1999).
409. Hershkovitz, P. Mammals of northern Colombia, preliminary report no. 6: rabbits (Leporidae), with notes on the classification and distribution of the South American forms. *Proc. US Natl Mus.* **100**, 327–375 (1950).
410. Hervias-Parejo, S. et al. Small size does not restrain frugivory and seed dispersal across the evolutionary radiation of Galapagos lava lizards. *Curr. Zool.* **65**, 353–361 (2019).
411. Heyer, M. M., Heyer, W. R. & de Sá, R. O. *Leptodactylus pentadactylus*. Catalogue of American Amphibians and Reptiles (CAAR) (ed. Price, A. H.) 1–48 (Society for the Study of Amphibians and Reptiles, 2012).
412. Heyer, W., Rand, A., Cruz, C., Peixoto, O. L. & Nelson, C. E. Frogs of Boracéia. *Arq. Zool.* **31**, 231–410 (1990).
413. Higgins, P. J. *Handbook of Australian, New Zealand and Antarctic birds: Parrots to Dollarbird* Vol. 4 (Oxford Univ. Press, 1999).
414. Higgins, P. J. & Davies, S. J. J. F. *Handbook of Australian, New Zealand and Antarctic Birds: Snipe to Pigeons* Vol. 3 (Oxford Univ. Press, 1996).
415. Higgins, P. J. & Peter, J. M. *Handbook of Australian, New Zealand and Antarctic Birds: Pardalotes to Shrike-Thrushes* Vol. 6 (Oxford Univ. Press, 2002).
416. Higgins, P. J., Peter, J. M. & Cowling, S. J. *Handbook of Australian, New Zealand and Antarctic Birds: Boatbill to Starlings* Vol. 7 (Oxford Univ. Press, 2006).
417. Higgins, P. J., Peter, J. M. & Steele, W. K. *Handbook of Australian, New Zealand and Antarctic Birds: Tyrant-Flycatchers to Chats* Vol. 5 (Oxford Univ. Press, 2001).
418. Hill, J. The Robinson collection of Malaysian mammals. *Bull. Raffles Mus.* **29**, 6–22 (1960).
419. Hitchmough, R. A. & McCallum, J. The mammals, birds, reptiles and freshwater fish of the eastern island group of the Bay of Islands. *Tane* **26**, 127–134 (1980).
420. Hofmeyr, M. D. Egg production in *Chersina angulata*: an unusual pattern in a Mediterranean climate. *J. Herpetol.* **38**, 172–179 (2004).
421. Hollingsworth, B. D. The systematics of chuckwallas (*Sauromalus*) with a phylogenetic analysis of other iguanid lizards. *Herpetol. Monogr.* **12**, 38–191 (1998).
422. How, R. A., Cowan, M. A., Teale, R. J. & Schmitt, L. H. Environmental correlates of reptile variation on the Houtman Abrolhos Archipelago, eastern Indian Ocean. *J. Biogeogr.* **47**, 2017–2028 (2020).
423. Huang, W.-S. Ecology and reproductive patterns of the agamid lizard *Japalura swinhonis* on an east Asian island, with comments on the small clutch sizes of island lizards. *Zool. Sci.* **24**, 181–188 (2007).
424. Hummelinck, P. W. *Studies on the Fauna of Curaçao, Aruba, Bonaire, and the Venezuelan Islands: No. 6* (Nijhoff, 1940).
425. Hutterer, R. Variation and evolution of the Sicilian shrew: taxonomic conclusions and description of a possibly related species from the Pleistocene of Morocco (Mammalia: Soricidae). *Bonn. Zool. Beitr.* **42**, 241–251 (1991).
426. Huyghe, K., Vanhooydonck, B., Herrel, A., Tadić, Z. & Van Damme, R. Morphology, performance, behavior and ecology of three color morphs in males of the lizard *Podarcis melisellensis*. *Integr. Comp. Biol.* **47**, 211–220 (2007).
427. Iliopoulou-Georgudaki, J. The relationship between climatic factors and forearm length of bats: evidence from the chiropterofauna of Lesbos island (Greece-east Aegean). *Mammalia* **50**, 475–482 (1986).
428. Inger, R. F., Stuart, B. L. & Iskandar, D. T. Systematics of a widespread Southeast Asian frog, *Rana chalconota* (Amphibia: Anura: Ranidae). *Zool. J. Linn. Soc.* **155**, 123–147 (2009).
429. Iverson, J. B., Hines, K. N. & Valiulis, J. M. The nesting ecology of the Allen Cays rock iguana, *Cyclura cyclura inornata* in the Bahamas. *Herpetol. Monogr.* **18**, 1–36 (2004).
430. Jacobs, D. Morphological divergence in an insular bat, *Lasiurus cinereus semotus*. *Funct. Ecol.* **10**, 622–630 (1996).
431. Jenkins, P. & Veitch, C. Sexual dimorphism and age determination in the North Island saddleback (*Philesturnus carunculatus rufaster*). *New Zeal. J. Zool.* **18**, 445–450 (1991).
432. Jenkins, P. D., Abramov, A. V., Bannikova, A. A. & Rozhnov, V. V. Bones and genes: resolution problems in three Vietnamese species of *Crocidura* (Mammalia, Soricomorpha, Soricidae) and the description of an additional new species. *ZooKeys* **2013**, 61–79 (2013).
433. Janssen, T. A. et al. Morphological characteristics of the lizard *Anolis carolinensis* from South Carolina. *Herpetologica* **51**, 401–411 (1995).
434. Jessop, R. Biometrics and moult of red-capped plovers in Victoria, Tasmania, South Australia and north west Australia. *Stilt* **17**, 29–35 (1990).
435. Jessop, T. S. et al. Maximum body size among insular Komodo dragon populations covaries with large prey density. *Oikos* **112**, 422–429 (2006).
436. Ji, X. & Wang, Z. W. Geographic variation in reproductive traits and trade-offs between size and number of eggs of the Chinese cobra (*Naja atra*). *Biol. J. Linn. Soc.* **85**, 27–40 (2005).
437. Ji, X., Xie, Y. Y., Sun, P. Y. & Zheng, X. Z. Sexual dimorphism and female reproduction in a viviparous snake, *Elaphe rufodorata*. *J. Herpetol.* **31**, 420–422 (1997).
438. Jimenez, J. E. & McMahon, E. in *Canids: Foxes, Wolves, Jackals, and Dogs: Status Survey and Conservation Action Plan* (eds Sillero-Zubiri, C. et al.) 50–55 (IUCN, 2004).
439. Joubert, L. & Cheylan, M. La tortue d'Hermann de Corse: résultat des recherches menées en 1985 et 1986. *Trav. Sci. Parc Nat. Rég. Réserves Nat. Corse* **22**, 1–54 (1989).
440. Judd, F. W. & Ross, R. K. Year-to-year variation in clutch size of island and mainland populations of *Holbrookia propinqua* (Reptilia, Lacertilia, Iguanidae). *J. Herpetol.* **12**, 203–207 (1978).
441. Kang, C. et al. Differential predation drives the geographical divergence in multiple traits in aposematic frogs. *Behav. Ecol.* **28**, 1122–1130 (2017).
442. Keast, A. Competitive interactions and the evolution of ecological niches as illustrated by the Australian honeyeater genus *Meliphreptus* (Meliphagidae). *Evolution* **22**, 762–784 (1968).
443. Keehn, J. E., Nieto, N. C., Tracy, C. R., Gienger, C. M. & Feldman, C. R. Evolution on a desert island: body size divergence between the reptiles of Nevada's Anaho Island and the mainland around Pyramid Lake. *J. Zool.* **291**, 269–278 (2013).
444. Kim, D.-I. et al. Patterns of morphological variation in the Schlegel's Japanese gecko (*Gekko japonicus*) across populations in China, Japan, and Korea. *J. Ecol. Environ.* **43**, 34 (2019).
445. Kim, S.-I., Oshida, T., Lee, H., Min, M.-S. & Kimura, J. Evolutionary and biogeographical implications of variation in skull morphology of raccoon dogs (*Nyctereutes procyonoides*, Mammalia: Carnivora). *Biol. J. Linn. Soc.* **116**, 856–872 (2015).
446. King, R. B. Body size variation among island and mainland snake populations. *Herpetologica* **45**, 84–88 (1989).
447. King, R. B. Variation in brown snake (*Storeria dekayi*) morphology and scalation: sex, family, and microgeographic differences. *J. Herpetol.* **31**, 335–346 (1997).
448. King, T. The birds of the Lesio-Louma and Lefini reserves, Batéké Plateau, Republic of Congo. *Malimbus* **33**, 1–41 (2011).
449. Kitchener, D., Hishah, S., Schmitt, L. & Maryanto, I. Morphological and genetic variation in *Aethalops alecto* (Chiroptera, Pteropodidae) from Java, Bali and Lombok Is, Indonesia. *Mammalia* **57**, 255–272 (1993).

450. Kitchener, D. & Schmitt, L. Morphological and genetic variation in *Suncus murinus* (Soricidae: Crocidurinae) from Java, Lesser Sunda Islands, Maluku and Sulawesi, Indonesia. *Mammalia* **58**, 433–452 (1994).
451. Kitchener, D., Schmitt, L., Hishig, S., How, R. & Cooper, N. Morphological and genetic variation in the bearded tomb bats (Taphozous: Emballonuridae) of Nusa Tenggara, Indonesia. *Mammalia* **57**, 63–84 (1993).
452. Klauber, L. M. The gopher snakes of Baja California: with descriptions of a new subspecies of *Pituophis catenifer*. *Trans. San Diego Soc. Nat. Hist.* **11**, 1–40 (1946).
453. Klauber, L. M. A new gopher snake (*Pituophis*) from Santa Cruz Island, California. *Trans. San Diego Soc. Nat. Hist.* **11**, 41–48 (1946).
454. Klauber, L. M. Some new and revived subspecies of rattlesnakes. *Trans. San Diego Soc. Nat. Hist.* **11**, 61–116 (1949).
455. Klauber, L. M. *Rattlesnakes: Their Habits, Life Histories, and Influence on Mankind* (Univ. California Press, 1997).
456. Kloss, C. B. On mammals collected in Siam. *J. Nat. Hist. Soc. Siam* **4**, 333–407 (1919).
457. Klütsch, C., Misof, B., Grosse, W. R. & Moritz, R. Genetic and morphometric differentiation among island populations of two *Norops* lizards (Reptilia: Sauria: Polychrotidae) on independently colonized islands of the Islas de Bahía (Honduras). *J. Biogeogr.* **34**, 1124–1135 (2007).
458. Knapp, C. R., Iverson, J. B. & Owens, A. K. Geographic variation in nesting behavior and reproductive biology of an insular iguana (*Cyclura cyclura*). *Can. J. Zool.* **84**, 1566–1575 (2006).
459. Kohno, H. Reptiles in a seabird colony: herpetofauna of Nakanokamishima Island of the Yaeyama group, Ryukyu Archipelago. *Isl. Stud. Okinawa* **9**, 73–89 (1991).
460. Koopman, K. F. & Diamond, J. Zoogeography of mammals from islands off the northeastern coast of New Guinea. *Am. Mus. Novit.* **2690**, 1–17 (1979).
461. Kraus, C., Trillmich, F. & Künkele, J. Reproduction and growth in a precocial small mammal, *Cavia magna*. *J. Mammal.* **86**, 763–772 (2005).
462. Proetzel, D., Ruthensteiner, B., Scherz, M. D. & Glaw, F. Systematic revision of the Malagasy chameleons *Calumma boettgeri* and *C. linotum* (Squamata: Chamaeleonidae). *Zootaxa* **4048**, 211–231 (2015).
463. Qualls, C. P., Shine, R., Donnellan, S. & Hutchinson, M. The evolution of viviparity within the Australian scincid lizard *Lerista bougainvillii*. *J. Zool.* **237**, 13–26 (1995).
464. Quin, D. G., Smith, A. P. & Norton, T. W. Eco-geographic variation in size and sexual dimorphism in sugar gliders and squirrel gliders (Marsupialia: Petauridae). *Aust. J. Zool.* **44**, 19–45 (1996).
465. Rader, J. A., Dillon, M. E., Chesser, R. T., Sabat, P. & del Rio, C. M. Morphological divergence in a continental adaptive radiation: South American ovenbirds of the genus *Cinclodes*. *Auk* **132**, 180–190 (2015).
466. Radočaj, M., Jelić, D., Karaica, D. & Kapelj, S. Morphological and reproductive traits of the insular population of *Podarcis siculus* (Reptilia: Lacertidae) from Krk Island (Croatia). *Hyla* **2**, 5–22 (2011).
467. Raia, P. et al. The blue lizard spandrel and the island syndrome. *BMC Evol. Biol.* **10**, 289 (2010).
468. Rajaratnam, R. *Ecology of Leopard Cat (Prionailurus bengalensis) in Tabin Wildlife Reserve, Sabah, Malaysia*. PhD thesis, Univ. Kebangsaan Malaysia (2000).
469. Rausch, R. L. Geographic variation in size in North American brown bears, *Ursus arctos* L., as indicated by condylobasal length. *Can. J. Zool.* **41**, 33–45 (1963).
470. Reboucas, R., da Silva, H. R., Sanuy, D. & Sole, M. Sexual maturity and growth of male toads (*Rhinella ornata*): a comparison between insular and mainland populations. *Zool. Anz.* **283**, 12–19 (2019).
471. Reboucas, R., da Silva, H. R. & Sole, M. Frog size on continental islands of the coast of Rio de Janeiro and the generality of the island rule. *PLoS ONE* **13**, e0190153 (2018).
472. Redfield, J. Distribution, abundance, size, and genetic variation of *Peromyscus maniculatus* on the Gulf Islands of British Columbia. *Can. J. Zool.* **54**, 463–474 (1976).
473. Renaud, S. et al. Morphometrics and genetics highlight the complex history of eastern Mediterranean spiny mice. *Biol. J. Linn. Soc.* **130**, 599–614 (2020).
474. Reynolds, R. G., Niemiller, M. L. & Fitzpatrick, B. M. Genetic analysis of an endemic archipelagic lizard reveals sympatric cryptic lineages and taxonomic discordance. *Conserv. Genet.* **13**, 953–963 (2012).
475. Rhodin, A. G. Chelid turtles of the Australasian Archipelago: a new species of *Chelodina* from Roti Island, Indonesia. *Breviora* **498**, 1–31 (1994).
476. Rickart, E. A., Heaney, L. R., Goodman, S. M. & Jansa, S. Review of the Philippine genera *Chrotomys* and *Celaenomys* (Murinae) and description of a new species. *J. Mammal.* **86**, 415–428 (2005).
477. Ridgway, R. *The Birds of North and Middle America: A Descriptive Catalogue of the Higher Groups, Genera, Species, and Subspecies of Birds Known to Occur in North America, from the Arctic Lands to the Isthmus of Panama, the West Indies and Other Islands of the Caribbean Sea, and the Galapagos Archipelago* (US Government Printing Office, 1904).
478. Rivas, G. A. et al. A distinctive new species of *Gonatodes* (Squamata: Sphaerodactylidae) from Isla La Blanquilla, Venezuela, with remarks on the distribution of some other Caribbean sphaerodactylid lizards. *S. Am. J. Herpetol.* **8**, 5–18 (2013).
479. Robertson, H., Whitaker, A. & Fitzgerald, B. Morphometrics of forest birds in the Orongorongo Valley, Wellington, New Zealand. *New Zeal. J. Zool.* **10**, 87–97 (1983).
480. Rocha, C. F. & Vrcibradic, D. Reproductive traits of two sympatric viviparous skinks (*Mabuya macrorhyncha* and *Mabuya agilis*) in a Brazilian restinga habitat. *Herpetol. J.* **9**, 43–53 (1999).
481. Rodríguez-Cabal, M. A., Amico, G. C., Novaro, A. J. & Aizen, M. A. Population characteristics of *Dromiciops gliroides* (Philippi, 1893), an endemic marsupial of the temperate forest of Patagonia. *Mamm. Biol.* **73**, 74–76 (2008).
482. Rog, S., Ryan, M. J., Mueller, U. & Lampert, K. P. Evidence for morphological and genetic diversification of túngara frog populations on islands. *Herpetol. Conserv. Biol.* **8**, 228–239 (2013).
483. Rogers, K., Rogers, A. & Rogers, D. *Bander's Aid: Supplement Number One* RAOU Report No. 67 (Royal Australasian Ornithologists Union, 1990).
484. Ross, R. K. & Judd, F. W. Comparison of lipid cycles of *Holbrookia propinqua* from Padre Island and mainland Texas. *J. Herpetol.* **16**, 53–60 (1982).
485. Rowe-Rowe, D. & Crafford, J. Density, body size, and reproduction of feral house mice on Gough Island. *S. Afr. J. Zool.* **27**, 1–5 (1992).
486. Runemark, A., Sagonas, K. & Svensson, E. I. Ecological explanations to island gigantism: dietary niche divergence, predation, and size in an endemic lizard. *Ecology* **96**, 2077–2092 (2015).
487. Sabater Pi, J. Contribution to the ecology of *Colobus polykomos satanas* (Waterhouse, 1838) of Rio Muni, Republic of Equatorial Guinea. *Folia Primatol.* **19**, 193–207 (1973).
488. Sagonas, K., Pafilis, P. & Valakos, E. D. Effects of insularity on digestion: living on islands induces shifts in physiological and morphological traits in island reptiles. *Sci. Nat.* **102**, 55 (2015).
489. Sahimi, H. N. M., Chubo, J. K., Mohd, M., Saripuddin, N. B. & Ab Rahim, S. S. The distribution and population density of Bornean tarsier, *Tarsius bancanus borneanus* (Elliot) in secondary and rehabilitated forests of Universiti Putra Malaysia, Bintulu Sarawak Campus, Sarawak, Malaysia. *Trop. Life Sci. Res.* **29**, 139–154 (2018).
490. Salvidio, S., Cresta, P. & Dolmen, D. The common toad *Bufo bufo* population of Hitra Island, central Norway. *Fauna Norv.* **14**, 51–55 (1993).
491. Sanches, V. Q. A. & Grings, D. R. Daily movement and habitat use of *Iguana iguana* (Linnaeus, 1758) in an urban second growth Amazonian forest fragment in Brazil. *Herpetol. Notes* **11**, 93–96 (2018).
492. Sanderson, J., Sunquist, M. E. & Iriarte, A. W. Natural history and landscape-use of guignas (*Oncifelis guigna*) on Isla Grande de Chiloé, Chile. *J. Mammal.* **83**, 608–613 (2002).
493. Sargis, E. J., Millien, V., Woodman, N. & Olson, L. E. Rule reversal: ecogeographical patterns of body size variation in the common treeshrew (Mammalia, Scandentia). *Ecol. Evol.* **8**, 1634–1645 (2018).
494. Sá-Sousa, P., Almeida, A., Rosa, H., Vicente, L. & Crespo, E. Genetic and morphological relationships of the Berenga wall lizard (*Podarcis bocagei berlengensis*: Lacertidae). *J. Zool. Syst. Evol. Res.* **38**, 95–102 (2000).
495. Sá-Sousa, P. & Harris, D. J. *Podarcis carbonelli* Pérez-Mellado, 1981 is a distinct species. *Amphib.-Reptil.* **23**, 459–468 (2002).
496. Sá-Sousa, P., Vicente, L. & Crespo, E. Morphological variability of *Podarcis hispanica* (Sauria: lacertidae) in Portugal. *Amphib.-Reptil.* **23**, 55–69 (2002).
497. Scali, S. et al. Continental versus Mediterranean European whip snake (*Hierophis viridiflavus*): a morphometric approach. *Biota* **3**, 161–166 (2003).
498. Scheffer, V. B. & Dalquest, W. W. A new shrew from Destruction Island, Washington. *J. Mammal.* **23**, 333–335 (1942).
499. Schifter, H., Cunningham-van Someren, G. & van Someren, G. C. The avifauna of the North Nandi Forest, Kenya. *Ann. Naturhist. Mus. Wien. B* **100**, 425–479 (1998).
500. Schlotfeldt, B. E. & Kleindorfer, S. Adaptive divergence in the superb fairy-wren (*Malurus cyaneus*): a mainland versus island comparison of morphology and foraging behaviour. *Emu* **106**, 309–319 (2006).
501. Schultz, A. H. Observations on the growth, classification and evolutionary specialization of gibbons and siamangs. *Hum. Biol.* **5**, 212–255 (1933).
502. Schultz, A. H. The relative weight of the testes in primates. *Anat. Rec.* **72**, 387–394 (1938).
503. Schwaner, T. D. & Sarre, S. D. Body size of tiger snakes in southern Australia, with particular reference to *Notechis ater serventyi* (Elapidae) on Chappell Island. *J. Herpetol.* **22**, 24–33 (1988).
504. Schwaner, T. D. & Sarre, S. D. Body size and sexual dimorphism in mainland and island tiger snakes. *J. Herpetol.* **24**, 320–322 (1990).
505. Schwartz, A. Three new mammals from southern Florida. *J. Mammal.* **33**, 381–385 (1952).
506. Schwartz, A. Snakes of the genus *Alsophis* in Puerto Rico and the Virgin Islands. *Stud. Fauna Curaçao Caribb. Isl.* **23**, 177–227 (1966).

507. Scott, D. Notes on the eastern hogsnoke, *Heterodon platyrhinus* Latreille (Squamata, Colubridae), on a Virginia barrier island. *Brimleyana* **12**, 51–55 (1986).
508. Selcer, K. W. & Judd, F. W. Variation in the reproductive ecology of *Holbrookia propinqua* (Sauria, Iguanidae). *Tex. J. Sci.* **34**, 125–135 (1982).
509. Senczuk, G., García, A., Colangelo, P., Annesi, F. & Castiglia, R. Morphometric and genetic divergence in island and mainland populations of *Anolis nebulosus* (Squamata: Polychrotidae) from Jalisco (Mexico): an instance of insular gigantism. *Ital. J. Zool.* **81**, 204–214 (2014).
510. Seock, M., Nam, K.-B. & Yoo, J.-C. Distribution and movement tendencies of short-tailed viper snakes (*Gloydius saxatilis*) by altitude. *Asian Herpetol. Res.* **8**, 39–47 (2017).
511. Serrano-Cardozo, V. H., Ramírez-Pinilla, M. P., Ortega, J. E. & Cortes, L. A. Annual reproductive activity of *Gonatodes albogularis* (Squamata: Gekkonidae) living in an anthropic area in Santander, Colombia. *S. Am. J. Herpetol.* **2**, 31–38 (2007).
512. Shaidani, N.-I. *The Biogeographic Origins and Trophic Ecology of Maine's Island Red-Backed Salamanders (Plethodon cinereus)*. MSc thesis, Univ. Maine (2017).
513. Sharples, C. M., Fa, J. E. & BELL, D. J. Geographical variation in size in the European rabbit *Oryctolagus cuniculus* (Lagomorpha: Leporidae) in western Europe and North Africa. *Zool. J. Linn. Soc.* **117**, 141–158 (1996).
514. Shekelle, M. Observations of wild Sangihe Island tarsiers *Tarsius sangirensis*. *Asian Primates J.* **3**, 18–23 (2013).
515. Shimada, T., Maeda, S. & Sakakibara, M. A morphological study of *Cynops pyrrhogaster* from the Chita Peninsula: rediscovery of the 'extinct' Atsumi race endemic to peninsular regions of Aichi Prefecture, central Japan. *Curr. Herpetol.* **35**, 38–52 (2016).
516. Shine, R. Venomous snakes in cold climates: ecology of the Australian genus *Drysdalia* (Serpentes: Elapidae). *Copeia* **1981**, 14–25 (1981).
517. Shine, R., Sun, L.-X., Zhao, E. & Bonnet, X. A review of 30 years of ecological research on the Shedao pitviper, *Gloydius shedaoensis*. *Herpetol. Nat. Hist.* **9**, 1–14 (2003).
518. Sicuro, F. L. & Oliveira, L. F. B. Variations in leopard cat (*Prionailurus bengalensis*) skull morphology and body size: sexual and geographic influences. *PeerJ* **3**, e1309 (2015).
519. Siler, C. D., McVay, J. D., Diesmos, A. C. & Brown, R. M. A new species of fanged frog, genus *Limnonectes* (Amphibia: Anura: Dicroglossidae) from southeast Mindanao Island, Philippines. *Herpetologica* **65**, 105–114 (2009).
520. Siliceo-Cantero, H. H., García, A., Reynolds, R. G., Pacheco, G. & Lister, B. C. Dimorphism and divergence in island and mainland *Anoles*. *Biol. J. Linn. Soc.* **118**, 852–872 (2016).
521. Simberloff, D., Dayan, T., Jones, C. & Ogura, G. Character displacement and release in the small Indian mongoose, *Herpestes javanicus*. *Ecology* **81**, 2086–2099 (2000).
522. Sinclair, E. Morphological variation among populations of the quokka, *Setonix brachyurus* (Macropodidae: Marsupialia), in Western Australia. *Aust. J. Zool.* **46**, 439–449 (1998).
523. Siqueira, C. C., Van Sluys, M., Ariani, C. V. & Rocha, C. F. Feeding ecology of *Thoropa miliaris* (Anura, Cycloramphidae) in four areas of Atlantic rain forest, southeastern Brazil. *J. Herpetol.* **40**, 520–525 (2006).
524. Slábová, M. & Frynta, D. Morphometric variation in nearly unstudied populations of the most studied mammal: the non-commensal house mouse (*Mus musculus domesticus*) in the Near East and northern Africa. *Zool. Anz.* **246**, 91–101 (2007).
525. Sleeman, D., Cussen, R., Southey, A. & O'Leary, D. The badgers *Meles meles* (L.) of Coney Island, Co. Sligo. *Ir. Nat. J.* **27**, 10–18 (2002).
526. Sleeman, D. P., Davenport, J., Cussen, R. E. & Hammond, R. F. The small-bodied badgers (*Meles meles* (L.)) of Rutland Island, Co. Donegal. *Ir. Nat. J.* **30**, 1–6 (2009).
527. Sleeman, P., Cussen, R., O'Donoghue, T. & Costello, E. Badgers (*Meles meles*) on Fenit Island, and their presence or absence on other Islands in Co. Kerry, Ireland. *Small Carniv. Conserv.* **24**, 10–12 (2001).
528. Smith, F. A. Evolution of body size among woodrats from Baja California, Mexico. *Funct. Ecol.* **6**, 265–273 (1992).
529. Smith, R. J. & Jungers, W. L. Body mass in comparative primatology. *J. Hum. Evol.* **32**, 523–559 (1997).
530. Smith, T. B. & Temple, S. A. Feeding habits and bill polymorphism in hook-billed kites. *Auk* **99**, 197–207 (1982).
531. Mitchell, C. & Mitchell, P. *Translocation of shore skink (Oligosoma smithi) from Mimiwhangata to Matakahe-Limestone Island (November/December 2007)* (Friends of Matakahe-Limestone Island Society, 2008).
532. Sody, H. Notes on some primates, carnivora, and the babirusa from the Indo-Malayan and Indo-Australian regions. *Treubia* **20**, 121–190 (1949).
533. Song, L., Fa-Hong, Y. & Xue-Fei, L. Cranial morphometric study of four giant flying squirrels (*Petaurista*) (Rodentia: Sciuridae) from China. *Zool. Res.* **33**, 119–126 (2012).
534. Sparkman, A. M. et al. Convergence in reduced body size, head size, and blood glucose in three island reptiles. *Ecol. Evol.* **8**, 6169–6182 (2018).
535. Stager, K. E. The avifauna of the Tres Marias Islands, Mexico. *Auk* **74**, 413–432 (1957).
536. Stamenković, S. & Matić, R. Morphological correlates of prey consumed by *Podarcis melisellensis* (Braun, 1877) and *P. siculus* (Rafinesque, 1810) (Sauria, Lacertidae) from two mainland regions in the eastern Adriatic area. *Arch. Biol. Sci.* **65**, 1015–1025 (2013).
537. Steven, D. M. Recent evolution in the genus *Clethrionomys*. *Symp. Soc. Exp. Biol.* **7**, 310–319 (1953).
538. Stoddart, D. M. & Braithwaite, R. W. A strategy for utilization of regenerating heathland habitat by the brown bandicoot (*Isodon obesulus*; Marsupialia, Peramelidae). *J. Anim. Ecol.* **48**, 165–179 (1979).
539. Storer, R. W. Variation in the red-tailed hawks of southern Mexico and Central America. *Condor* **64**, 77–78 (1962).
540. Strickland, D. & Norris, D. R. An example of phenotypic adherence to the island rule? Anticosti gray jays are heavier but not structurally larger than mainland conspecifics. *Ecol. Evol.* **5**, 3687–3694 (2015).
541. Struhsaker, T. T. *The Red Colobus Monkeys: Variation in Demography, Behavior, and Ecology of Endangered Species* (Oxford Univ. Press, 2010).
542. Stuart-Smith, J. F., Stuart-Smith, R. D., Swain, R. & Wapstra, E. Size dimorphism in *Rankinia tympanocryptis diemensis* (Family Agamidae): sex-specific patterns and geographic variation. *Biol. J. Linn. Soc.* **94**, 699–709 (2008).
543. Stubbs, D. & Shingland, I. R. The ecology of a Mediterranean tortoise (*Testudo hermanni*): a declining population. *Can. J. Zool.* **63**, 169–180 (1985).
544. Swarth, H. The lemming of Nunivak Island, Alaska. *Proc. Biol. Soc. Wash.* **44**, 101–104 (1931).
545. Swarth, H. S. in *University of California Publications in Zoology* Vol. 7 (eds Ritter, W. E. & Kofoed, C. A.) 9–172 (Univ. California Press, 1911).
546. Takada, Y., Sakai, E., Uematsu, Y. & Tateishi, T. Morphometric variation of house mice (*Mus musculus*) on the Izu Islands. *Mammal. Study* **24**, 51–65 (1999).
547. Takada, Y., Sakai, E., Uematsu, Y. & Tateishi, T. Morphological variation of large Japanese field mice, *Apodemus speciosus* on the Izu and Oki islands. *Mammal. Study* **31**, 29–40 (2006).
548. Takada, Y., Yamada, H. & Tateishi, T. Morphometric variation of Japanese wild mice on islands. *J. Mamm. Soc. Jpn* **19**, 113–128 (1994).
549. Takenaka, T. & Hasegawa, M. Female-biased mortality and its consequence on adult sex ratio in the freshwater turtle *Chinemys reevesii* on an island. *Curr. Herpetol.* **20**, 11–17 (2001).
550. Tamarin, R. H. Dispersal, population regulation, and K-selection in field mice. *Am. Nat.* **112**, 545–555 (1978).
551. Tanaka, K. Phenotypic plasticity of body size in an insular population of a snake. *Herpetologica* **67**, 46–57 (2011).
552. Tanaka, K. & Ota, H. Natural history of two colubrid snakes, *Elaphe quadrivirgata* and *Rhabdophis tigrinus*, on Yakushima Island, southwestern Japan. *Amphib.-Reptil.* **23**, 323–331 (2002).
553. Taverne, M. et al. Diet variability among insular populations of *Podarcis* lizards reveals diverse strategies to face resource-limited environments. *Ecol. Evol.* **9**, 12408–12420 (2019).
554. Taylor, E. H. The lizards of Thailand. *Univ. Kans. Sci. Bull.* **44**, 687–1077 (1963).
555. Taylor, H. L. & Cooley, C. R. A multivariate analysis of morphological variation among parthenogenetic teiid lizards of the *Cnemidophorus cozumela* complex. *Herpetologica* **51**, 67–76 (1995).
556. Terada, C., Tatsuzawa, S. & Saitoh, T. Ecological correlates and determinants in the geographical variation of deer morphology. *Oecologia* **169**, 981–994 (2012).
557. Terborgh, J., Faaborg, J. & Brockmann, H. J. Island colonization by Lesser Antillean birds. *Auk* **95**, 59–72 (1978).
558. Tessa, G., Crottini, A., Giacoma, C., Guarino, F. M. & Randrianirina, J. E. Comparative longevity and age at sexual maturity in twelve rainforest frogs of the genera *Boophis*, *Gephyromantis*, and *Mantidactylus* (Anura: Mantellidae) from Madagascar. *Phyllomedusa* **16**, 13–21 (2017).
559. Thomas, D. & Broughton, E. *Status of Three Canadian Caribou Populations North of 70 in Winter 1977* (Canadian Wildlife Service, 1978).
560. Thouless, C. & Bassri, K. A. Taxonomic status of the Farasan Island gazelle. *J. Zool.* **223**, 151–159 (1991).
561. Tsai, T. Sexual dimorphism of Chinese green tree viper. *Trimeresurus stejnegeri stejnegeri*. *Biol. Bull. Natl. Taiwan Normal Univ.* **33**, 13–22 (1998).
562. Uller, T. et al. Genetic differentiation predicts body size divergence between island and mainland populations of common wall lizards (*Podarcis muralis*). *Biol. J. Linn. Soc.* **127**, 771–786 (2019).
563. Ursin, E. *Geographical Variation in Apodemus sylvaticus and A. flavicollis (Rodentia, Muridae) in Europe, with Special Reference to Danish and Latvian Populations* (Munksgaard, 1956).
564. Valakos, E. D. & Polymeni, R. M. The food of *Cyrtodactylus kotschy* (Steindachner, 1870) (Sauria - Gekkonidae) during the wet season in the Mediterranean insular ecosystems of the Aegean. *Herpetol. J.* **1**, 474–477 (1990).

565. Van Heezik, Y. M., Cooper, J. & Seddon, P. J. Population characteristics and morphometrics of angulate tortoises on Dassen Island, South Africa. *J. Herpetol.* **28**, 447–453 (1994).
566. Van Weenen, J. *Aspects of the Ecology of Cunningham's Skink, Egernia cunninghami, on West Island, South Australia*. BSc thesis, Univ. Adelaide (1995).
567. Vanek, J. P. & Burke, R. L. Insular dwarfism in female eastern hog-nosed snakes (*Heterodon platirhinos*; Dipsadidae) on a barrier island. *Can. J. Zool.* **98**, 157–164 (2020).
568. Vanzolini, P. E. & Reboucas-Spieker, R. Distribution and differentiation of animals along the coast and in continental islands of the state of São Paulo, Brazil. 3. Reproductive differences between *Mabuya caissara* and *Mabuya macrorhyncha* (Sauria, Scincidae). *Pap. Avulsos Zool.* **29**, 95–109 (1976).
569. Vasconcelos, R. & Carranza, S. Systematics and biogeography of *Hemidactylus homoeolepis* Blanford, 1881 (Squamata: Gekkonidae), with the description of a new species from Arabia. *Zootaxa* **3835**, 501–527 (2014).
570. Velo-Anton, G. & Cordero-Rivera, A. Ethological and phenotypic divergence in insular fire salamanders: diurnal activity mediated by predation? *Acta Ethol.* **20**, 243–253 (2017).
571. Vences, M. Erste untersuchungen über die fortpflanzungsbiologie des Iberischen scheibenzünglers *Discoglossus galganoi* (Amphibia: Anura: Discoglossidae). *Acta Biol. Benrodis* **6**, 89–98 (1994).
572. Vences, M., Glaw, F., Mercurio, V. & Andreone, F. Review of the Malagasy tree snakes of the genus *Stenophis* (Colubridae). *Salamandra* **40**, 161–179 (2004).
573. Vences, M., Köhler, J., Pabijan, M. & Glaw, F. Two syntopic and microendemic new frogs of the genus *Blommersia* from the east coast of Madagascar. *Afr. J. Herpetol.* **59**, 133–156 (2010).
574. Ventura, J. & Fuster, M. J. L. Morphometric analysis of the black rat, *Rattus rattus*, from Congreso Island (Chafarinas Archipelago, Spain). *Orsis* **15**, 91–102 (2000).
575. Verdon, E. & Donnelly, M. A. Population structure of Florida box turtles (*Terrapene carolina bauri*) at the southernmost limit of their range. *J. Herpetol.* **39**, 572–577 (2005).
576. Vershinin, A. The biology and trapping of the ermine in Kamchatka. *Byull. Mosk. Ova. Ispyt. Prir. Otd. Biol.* **77**, 16–26 (1972).
577. Vervust, B., Grbac, I. & Van Damme, R. Differences in morphology, performance and behaviour between recently diverged populations of *Podarcis sicula* mirror differences in predation pressure. *Oikos* **116**, 1343–1352 (2007).
578. Villeneuve, A. R. Habitat selection and population density of the world's smallest chameleon, *Brookesia micra*, on Nosy Hara, Madagascar. *Herpetol. Conserv. Biol.* **12**, 334–341 (2017).
579. Vogel, P. & Sofianidou, T. The shrews of the genus *Crociodura* on Lesbos, an eastern Mediterranean island. *Bonn. Zool. Beitr.* **46**, 339–347 (1996).
580. Vogrin, M. Sexual dimorphism in *Podarcis sicula campestris*. *Turk. J. Zool.* **29**, 189–191 (2005).
581. Vrcibradic, D. & Rocha, C. F. Observations on the natural history of the lizard *Mabuya macrorhyncha* Hoge (Scincidae) in Queimada Grande Island, Sao Paulo, Brazil. *Rev. Bras. Zool.* **22**, 1185–1190 (2005).
582. Walker, R., Woods-Ballard, A. J. & Rix, C. E. Population density and seasonal activity of the threatened Madagascar spider tortoise (*Ptyxis arachnoides arachnoides*) of the southern dry forests; south west Madagascar. *Afr. J. Ecol.* **46**, 67–73 (2008).
583. Wang, Y., Li, Y., Wu, Z. & Murray, B. Insular shifts and trade-offs in life-history traits in pond frogs in the Zhoushan Archipelago, China. *J. Zool.* **278**, 65–73 (2009).
584. Watkins, G. G. Proximate causes of sexual size dimorphism in the iguanian lizard *Microlophus occipitalis*. *Ecology* **77**, 1473–1482 (1996).
585. Wayne, R. K. et al. A morphologic and genetic study of the island fox, *Urocyon littoralis*. *Evolution* **45**, 1849–1868 (1991).
586. West, A. G. *Variation in Mandible Shape and Body Size of House Mice Mus musculus Across the New Zealand Archipelago: A Trans-Tasman Comparison Using Geometric Morphometrics*. MSc thesis, Univ. Waikato (2017).
587. Whitehead, V. *Population Dynamics of the Eastern Box Turtle (Terrapene carolina carolina) in the Maryville College Woods, Maryville, Tennessee: A Report of a Senior Study*. BSc thesis, Maryville College (2017).
588. Woodman, N. et al. A new southern distributional limit for the Central American rodent *Peromyscus stirtoni*. *Caribb. J. Sci.* **38**, 281–284 (2002).
589. Wu, Z., Li, Y. & Murray, B. R. Insular shifts in body size of rice frogs in the Zhoushan Archipelago, China. *J. Anim. Ecol.* **75**, 1071–1080 (2006).
590. Xu, F., Adler, G. H. & Li, Y. Covariation in insular life-history traits of the rice frog (*Fejervarya limnocharis*) in eastern China. *Asian Herpetol. Res.* **4**, 28–35 (2013).
591. Yabe, T. Population structure and male melanism in the Reeves' turtle, *Chinemys reevesii*. *Jpn. J. Herpetol.* **15**, 131–137 (1994).
592. Yasukawa, Y., Ota, H. & Iverson, J. B. Geographic variation and sexual size dimorphism in *Mauremys mutica* (Cantor, 1842) (Reptilia: Bataguridae), with description of a new subspecies from the southern Ryukyus, Japan. *Zool. Sci.* **13**, 303–317 (1996).
593. Yurgenson, P. B. Ermines of the Far East territory. *Byull. Mosk. Ova. Ispyt. Prir. Otd. Biol.* **45**, 239–243 (1936).
594. Yusefi, G. H., Kiabi, B. H., Khalatbari, L., Faizolah, K. & Monteiro, N. M. Morphological analysis of Brandt's hedgehog (*Paraechinus hypomelas*) reflects the isolation history of Persian Gulf islands and has implications for taxonomy. *Biol. J. Linn. Soc.* **119**, 497–510 (2016).
595. Yustian, I. *Ecology and Conservation Status of Tarsius bancanus saltator on Belitung Island, Indonesia* (Cuvillier, 2007).
596. Ziegler, T., Vences, M., Glaw, F. & Böhme, W. Remarks on the genital morphology of the Malagasy snake genus *Liophidium* (Reptilia, Serpentes, Colubridae). *Acta Biol. Benrodis* **8**, 157–159 (1996).
597. Zihlman, A. L., Mootnick, A. R. & Underwood, C. E. Anatomical contributions to hybrid taxon and adaptation. *Int. J. Primatol.* **32**, 865–877 (2011).
598. Zuffi, M. & Gariboldi, A. Sexual dimorphism of the European pond terrapin, *Emys orbicularis* (L., 1758) from Italy. *Sci. Herpetol.* **1995**, 124–129 (1995).
599. Zuffi, M. A., Sacchi, R., Pupin, F. & Cencetti, T. Sexual size and shape dimorphism in the Moorish gecko (*Tarentola mauritanica*, Gekkota, Phyllodactylidae). *North-West. J. Zool.* **7**, 189–197 (2011).
600. Zuffi, M. A. L., Odetti, F., Batistoni, R. & Mancino, G. Geographic variation of sexual size dimorphism and genetics in the European pond turtle, *Emys orbicularis* and *Emys trinacris*, of Italy. *Ital. J. Zool.* **73**, 363–372 (2006).
601. Zug, G. R., Hedges, S. B. & Sunkel, S. Variation in reproductive parameters of three neotropical snakes, *Coniophanes fissidens*, *Dipsas catesbyi*, and *Imantodes cenchoa*. *Smithson. Contrib. Zool.* **300**, 1–20 (1979).
602. Higgins, J. P. et al. *Cochrane Handbook for Systematic Reviews of Interventions* (John Wiley & Sons, 2019).
603. Santini, L., Benítez-López, A., Ficetola, G. F. & Huijbregts, M. A. Length-mass allometries in amphibians. *Integr. Zool.* **13**, 36–45 (2018).
604. Lislevand, T., Figuerola, J. & Székely, T. Avian body sizes in relation to fecundity, mating system, display behavior, and resource sharing. *Ecology* **88**, 1605–1605 (2007).
605. Van Valkenburgh, B. in *Body Size in Mammalian Paleobiology: Estimation and Biological Implications* (eds Damuth, J. & MacFadden, B. J.) 181–206 (Cambridge Univ. Press, 1990).
606. Jones, K. E. et al. PanTHERIA: a species-level database of life history, ecology, and geography of extant and recently extinct mammals. *Ecology* **90**, 2648–2648 (2009).
607. Scharf, I. et al. Late bloomers and baby boomers: ecological drivers of longevity in squamates and the tuatara. *Glob. Ecol. Biogeogr.* **24**, 396–405 (2015).
608. Regis, K. W. & Meik, J. M. Allometry of sexual size dimorphism in turtles: a comparison of mass and length data. *PeerJ* **5**, e2914 (2017).
609. Gosler, A., Greenwood, J., Baker, J. & Davidson, N. The field determination of body size and condition in passerines: a report to the British Ringing Committee. *Bird Study* **45**, 92–103 (1998).
610. Rising, J. D. & Somers, K. M. The measurement of overall body size in birds. *Auk* **106**, 666–674 (1989).
611. Jetz, W., Thomas, G., Joy, J., Hartmann, K. & Mooers, A. The global diversity of birds in space and time. *Nature* **491**, 444–448 (2012).
612. Wan, X., Wang, W., Liu, J. & Tong, T. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. *BMC Med. Res. Methodol.* **14**, 135 (2014).
613. Bracken, M. in *Effective Care of the Newborn Infant* (eds Sinclair, J. C. & Bracken, M. B.) 13–20 (Oxford Univ. Press, 1992).
614. Pettorelli, N. et al. The normalized difference vegetation index (NDVI): unforeseen successes in animal ecology. *Clim. Res.* **46**, 15–27 (2011).
615. Fick, S. E. & Hijmans, R. J. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *Int. J. Climatol.* **37**, 4302–4315 (2017).
616. Blackburn, T. M., Gaston, K. J. & Loder, N. Geographic gradients in body size: a clarification of Bergmann's rule. *Divers. Distrib.* **5**, 165–174 (1999).
617. Wilman, H. et al. EltonTraits 1.0: species-level foraging attributes of the world's birds and mammals. *Ecology* **95**, 2027–2027 (2014).
618. Meiri, S. Traits of lizards of the world: variation around a successful evolutionary design. *Glob. Ecol. Biogeogr.* **27**, 1168–1172 (2018).
619. Benítez-López, A. et al. The impact of hunting on tropical mammal and bird populations. *Science* **356**, 180–183 (2017).
620. Solé, M. & Rödder, D. in *Amphibian Ecology and Conservation: A Handbook of Techniques* (ed. Dodd, C. K. Jr) 167–184 (Oxford Univ. Press, 2010).
621. McClain, C. R., Durst, P. A. P., Boyer, A. G. & Francis, C. D. Unravelling the determinants of insular body size shifts. *Biol. Lett.* **9**, 20120989 (2013).
622. Rozzi, R. Space-time patterns of body size variation in island bovids: the key role of predatory release. *J. Biogeogr.* **45**, 1196–1207 (2018).
623. Hadfield, J. & Nakagawa, S. General quantitative genetic methods for comparative biology: phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *J. Evol. Biol.* **23**, 494–508 (2010).

624. Lajeunesse, M. J. On the meta-analysis of response ratios for studies with correlated and multi-group designs. *Ecology* **92**, 2049–2055 (2011).
625. Lindstedt, S. L. & Boyce, M. S. Seasonality, fasting endurance, and body size in mammals. *Am. Nat.* **125**, 873–878 (1985).
626. Nakagawa, S. & Schielzeth, H. A general and simple method for obtaining R^2 from generalized linear mixed-effects models. *Methods Ecol. Evol.* **4**, 133–142 (2013).
627. R Core Team R: *A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2018).
628. Viechtbauer, W. Conducting meta-analyses in R with the metafor package. *J. Stat. Softw.* **36**, 1–48 (2010).
629. Lajeunesse, M. J. Facilitating systematic reviews, data extraction and meta-analysis with the metagear package for R. *Methods Ecol. Evol.* **7**, 323–330 (2016).
630. Pick, J. L., Nakagawa, S. & Noble, D. W. Reproducible, flexible and high-throughput data extraction from primary literature: the metaDigitise R package. *Methods Ecol. Evol.* **10**, 426–431 (2019).
631. Paradis, E., Claude, J. & Strimmer, K. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290 (2004).
632. Michonneau, F., Brown, J. W. & Winter, D. J. rotl: an R package to interact with the Open Tree of Life data. *Methods Ecol. Evol.* **7**, 1476–1481 (2016).
633. Rees, J. A. & Cranston, K. Automated assembly of a reference taxonomy for phylogenetic data synthesis. *Biodivers. Data J.* **5**, e12581 (2017).
634. Hinchliff, C. E. et al. Synthesis of phylogeny and taxonomy into a comprehensive tree of life. *Proc. Natl Acad. Sci. USA* **112**, 12764–12769 (2015).
635. Pebesma, E. Simple features for R: standardized support for spatial vector data. *R J.* **10**, 439–446 (2018).
636. Hijmans, R. J. et al. raster: Geographic Data Analysis and Modeling (2015); <https://rspatial.org/raster>
637. Wickham, H., Francois, R., Henry, L. & Müller, K. dplyr: A Grammar of Data Manipulation. R package version 0.4.3. <https://CRAN.R-project.org/package=dplyr> (2015).
638. Wickham, H. reshape2: Flexibly Reshape Data: A Reboot of the reshape Package. R package version 1 (2012); <https://github.com/hadley/reshape>
639. Wickham, H. *ggplot2: Elegant Graphics for Data Analysis* (Springer, 2016).
640. Kassambara, A. ggpubr: 'ggplot2' Based Publication Ready Plots. R package version 0.1. 6. (2017); <https://rpkgs.datanovia.com/ggpubr/>

Acknowledgements

We are grateful to K. B. Aubry, J. E. Keehn, S. Michaelides and D. Strickland for sharing their data with us, and to P. Peres-Neto and S. Nakagawa for useful discussion on the analytical framework. A.B.-L. was supported by a Juan de la Cierva-Incorporación grant (IJCI-2017-31419) from the Spanish Ministry of Science, Innovation and Universities. L.S. and M.A.J.H. were supported by the ERC project (62002139 ERC—CoG SIZE 647224). We thank numerous biological collections, in particular the Natural History Museum, Tring, for providing access to specimens. Bird trait data collection was supported by Natural Environment Research Council grant nos. NE/I028068/1 and NE/P004512/1 (to J.A.T.).

Author contributions

A.B.-L. conceived and coordinated the research, led the analyses and wrote the first draft; A.B.-L., L.S., J.G.-Z., M.A.J.H. and J.A.T. helped to develop the conceptual framework; L.S. compiled the environmental rasters; J.A.T., P.W. and B.M. provided morphometric data. All authors contributed to the data collection from the literature and to the writing of the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Extended data is available for this paper at <https://doi.org/10.1038/s41559-021-01426-y>.

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41559-021-01426-y>.

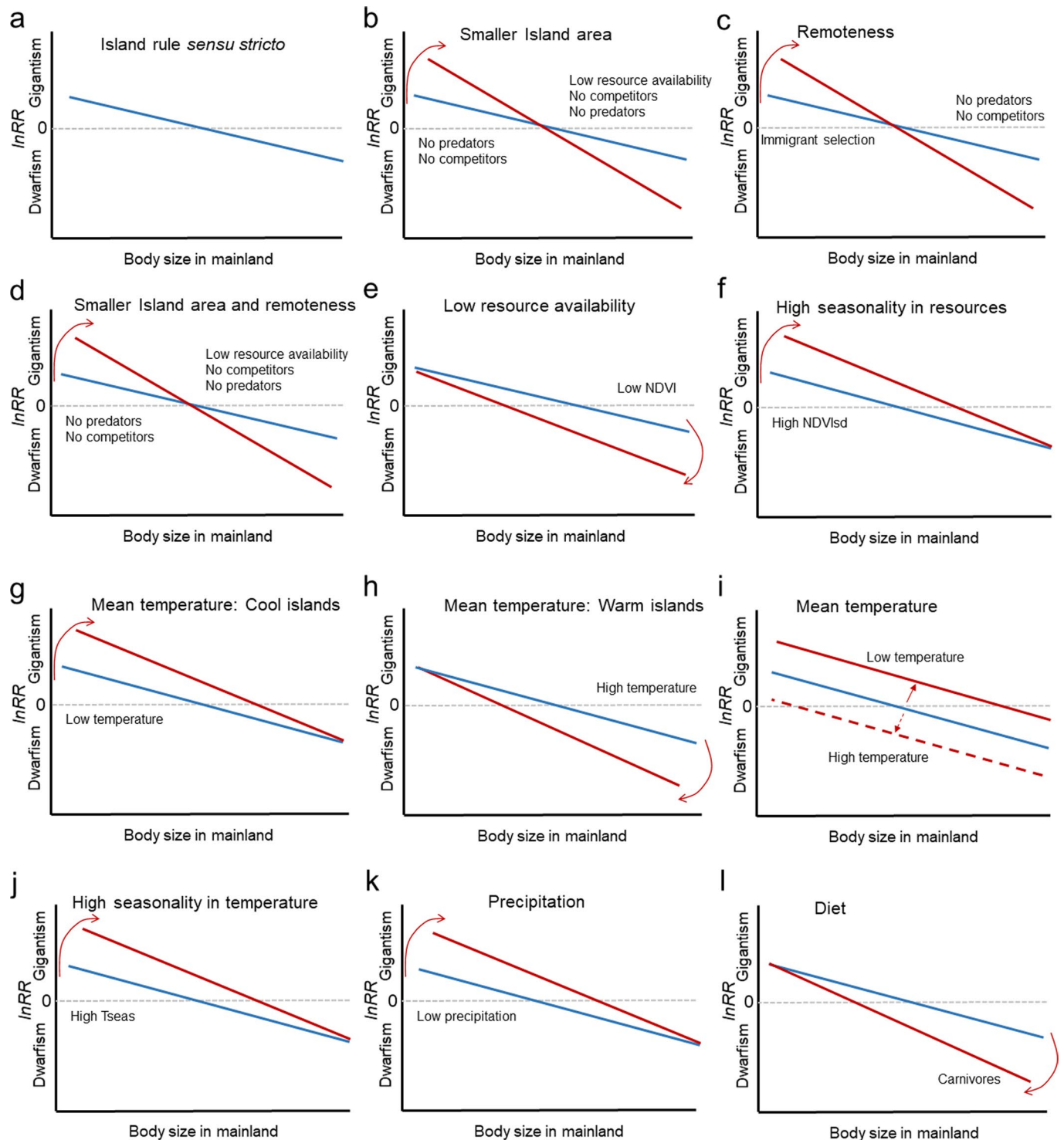
Correspondence and requests for materials should be addressed to A.B.-L.

Peer review information *Nature Ecology & Evolution* thanks Shai Meiri, Alfredo Sánchez-Tójar and Luis Valente for their contribution to the peer review of this work. Peer reviewer reports are available.

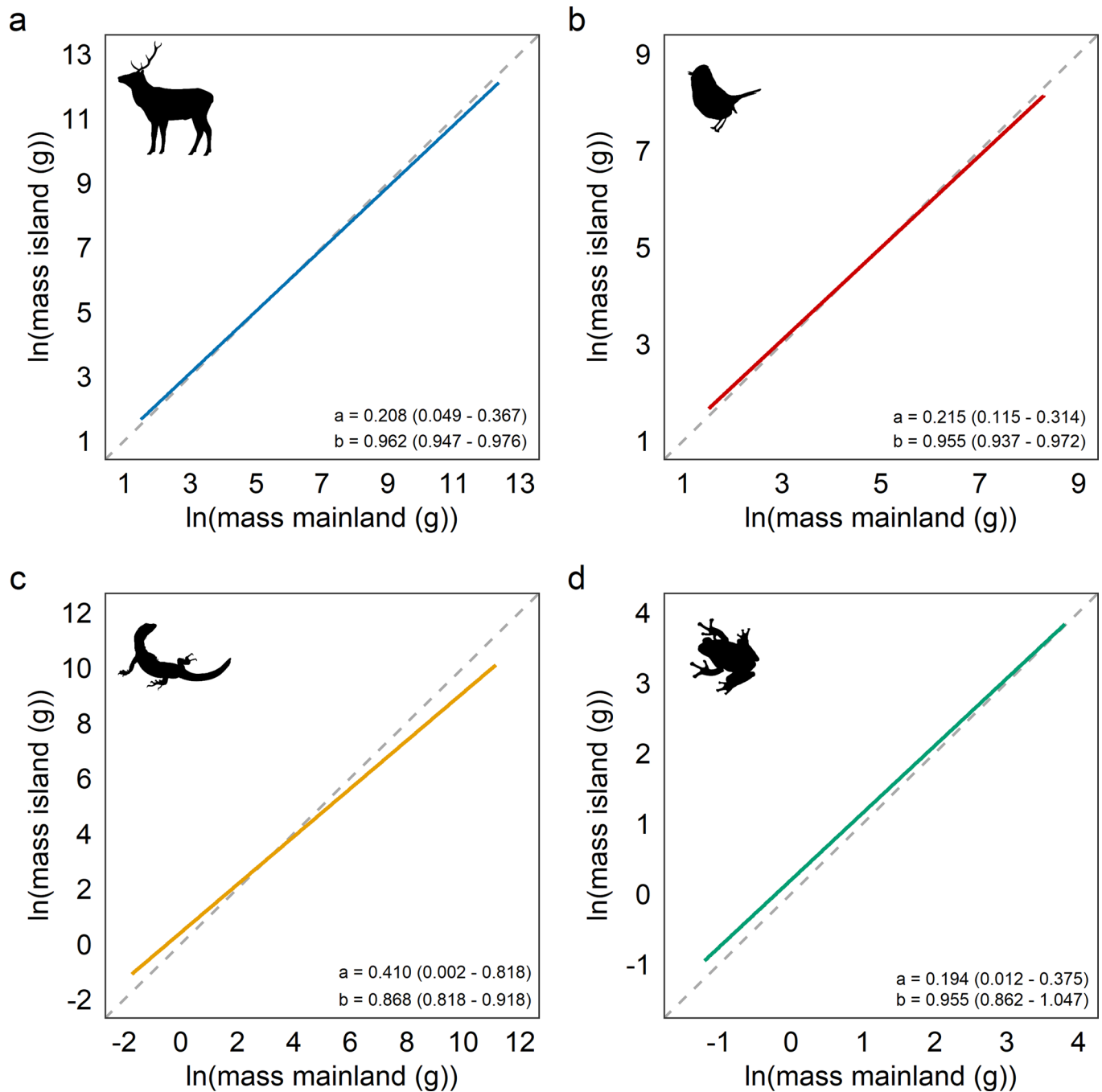
Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

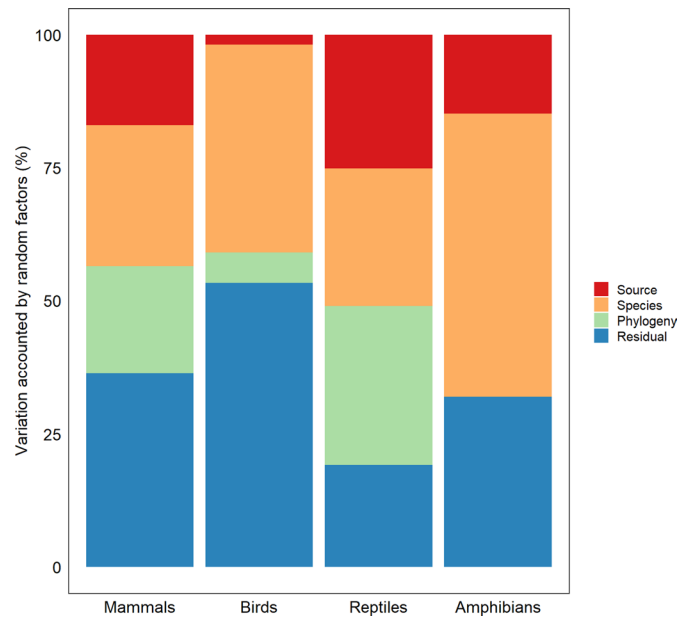
© The Author(s), under exclusive licence to Springer Nature Limited 2021



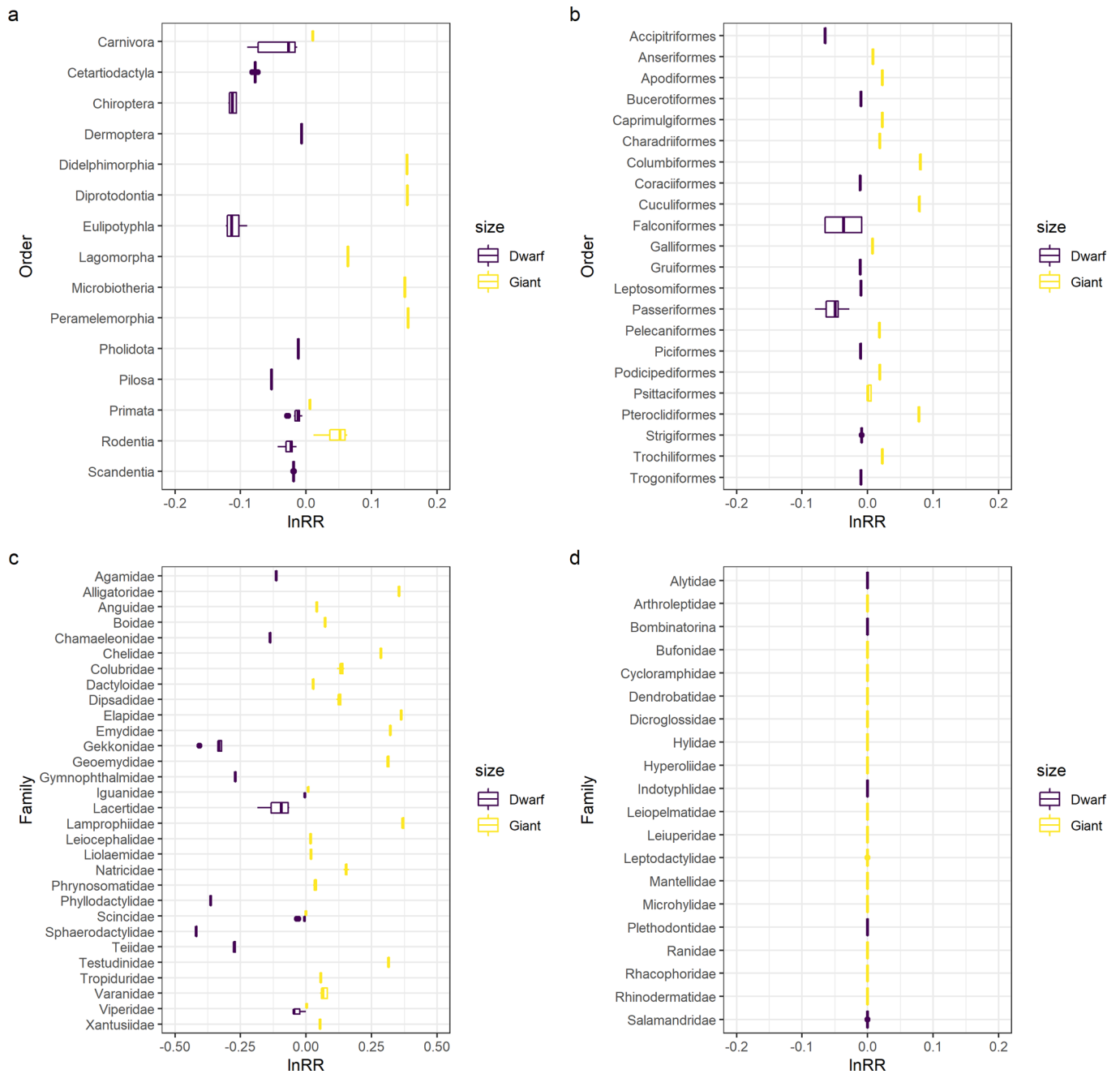
Extended Data Fig. 1 | Conceptual models depicting the different hypotheses tested to explain insular size shifts in vertebrates. Expected insular size shifts are depicted as a result of (a) Island rule effects; (b) island area; (c) spatial isolation (distance to mainland); (d) a combination of island area and distance to mainland; (e) primary productivity, (f) seasonality in resources; (g) mean temperature affecting mostly small species in cool islands; (h) mean temperature affecting mostly small species in warm islands; (i) mean temperature affecting all species; (j) seasonality in temperature; (k) precipitation; (l) species dietary preferences (carnivores vs non carnivores). Detailed information on the different hypotheses can be found in Supplementary Table 1.



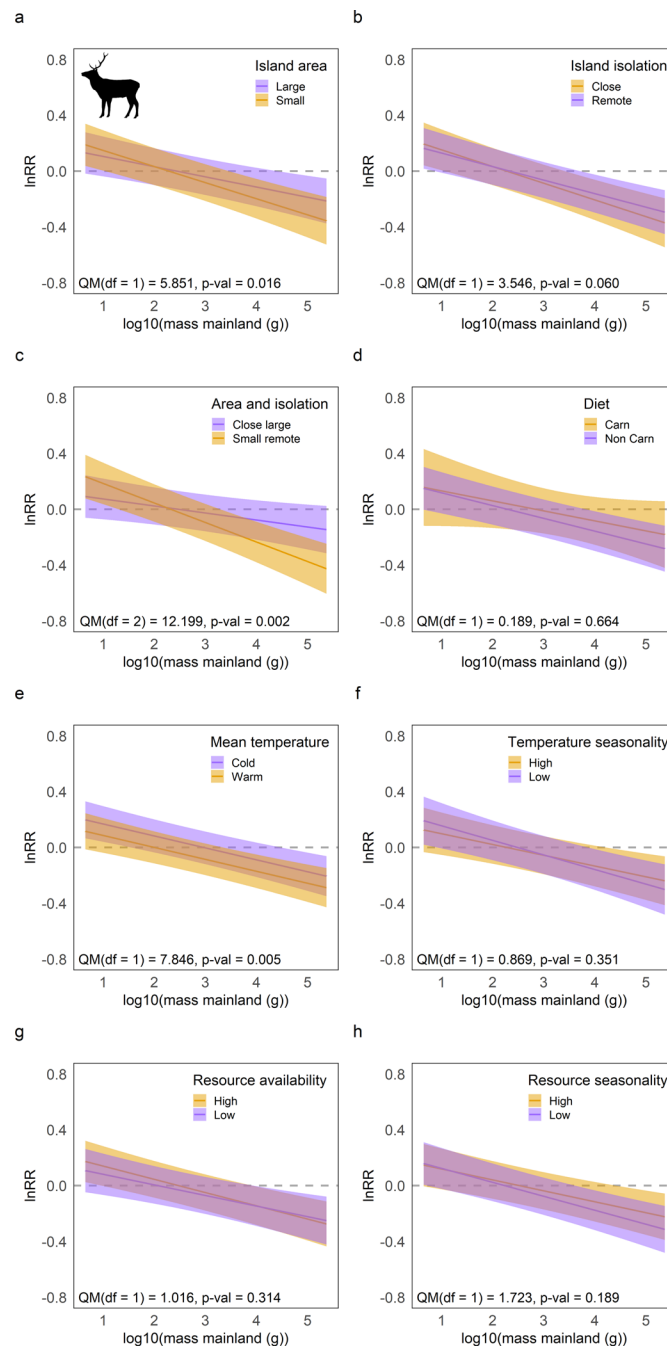
Extended Data Fig. 2 | Phylogenetic meta-regression models of island body size versus mainland body size. Mean body size (\ln -transformed mass, in g) on islands versus mainland is shown for (a) mammals, (b) birds, (c) reptiles and (d) amphibians. The dashed line has a slope of 1 and an intercept of 0. The solid lines represent the phylogenetic meta-regression slope estimate. The intercept a and slope b are presented in each plot along with CI. The island rule holds if the intercept is > 0 and the slope < 1 .



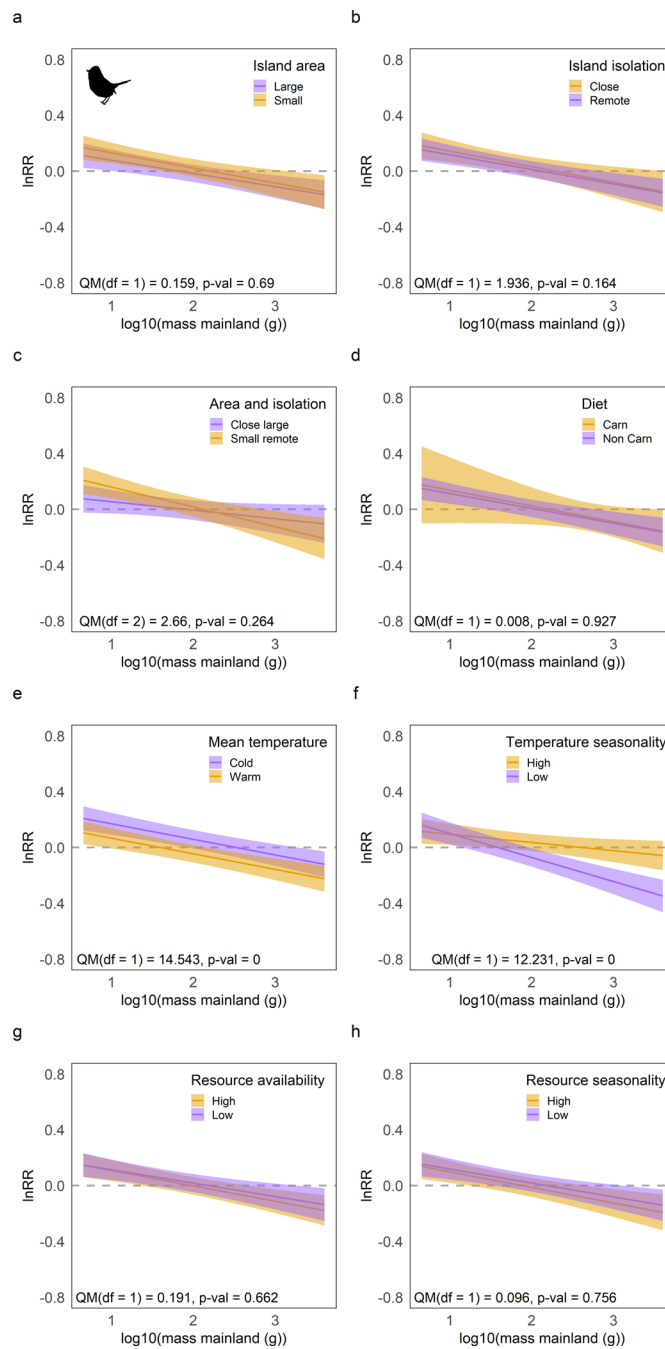
Extended Data Fig. 3 | Variation accounted for by random factors (Source, Species and Phylogeny) and residual variation. The amount of variance accounted for by phylogeny was the largest for reptiles and mammals, and low for birds. The extent of variance explained by data sources was larger for mammals and reptiles, and low for amphibians and birds. The residual variance was highest for birds, followed by mammals, amphibians and reptiles, indicating that other factors besides mainland body size may help to explain insular size shifts (see Extended Data Fig. 5–8).



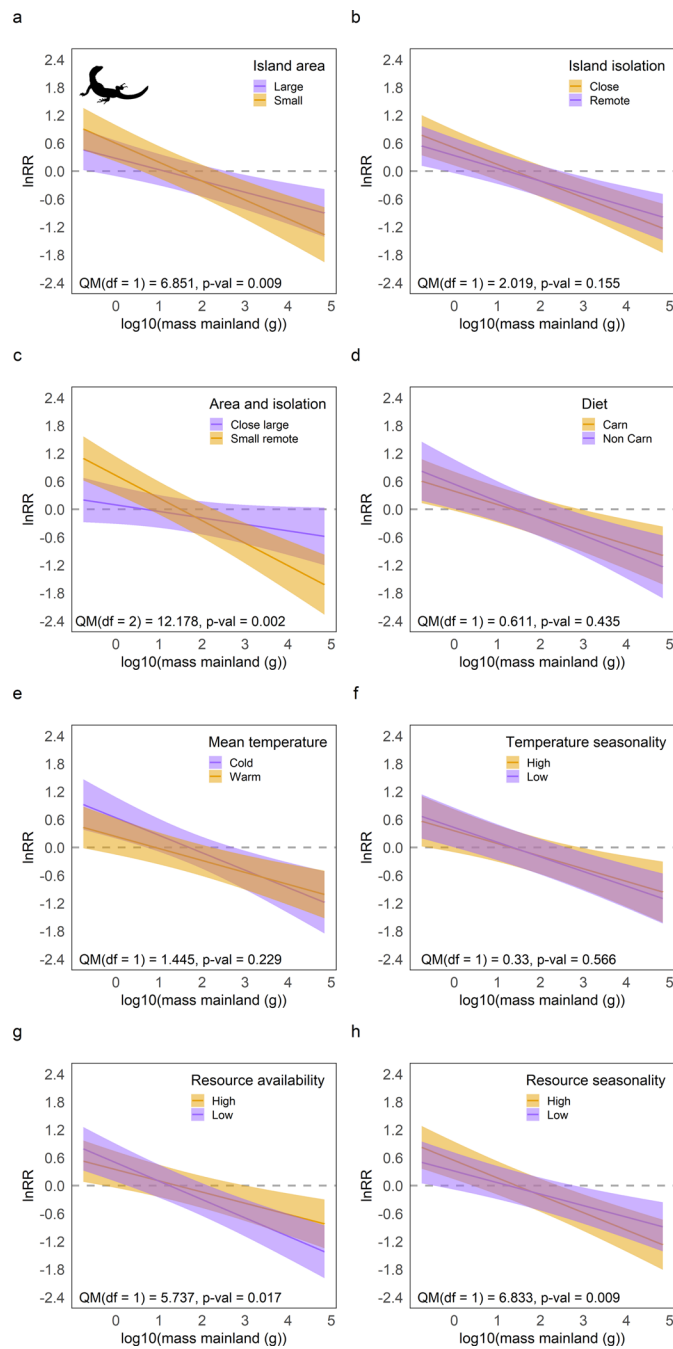
Extended Data Fig. 4 | Phylogenetic random effects across taxonomic orders. Boxplots of the variation in phylogenetic random effects across orders are shown for **(a)** mammals and **(b)** birds; and across families for **(c)** reptiles and **(d)** amphibians. Positive values indicate a tendency towards gigantism, whereas negative values indicate dwarfism.



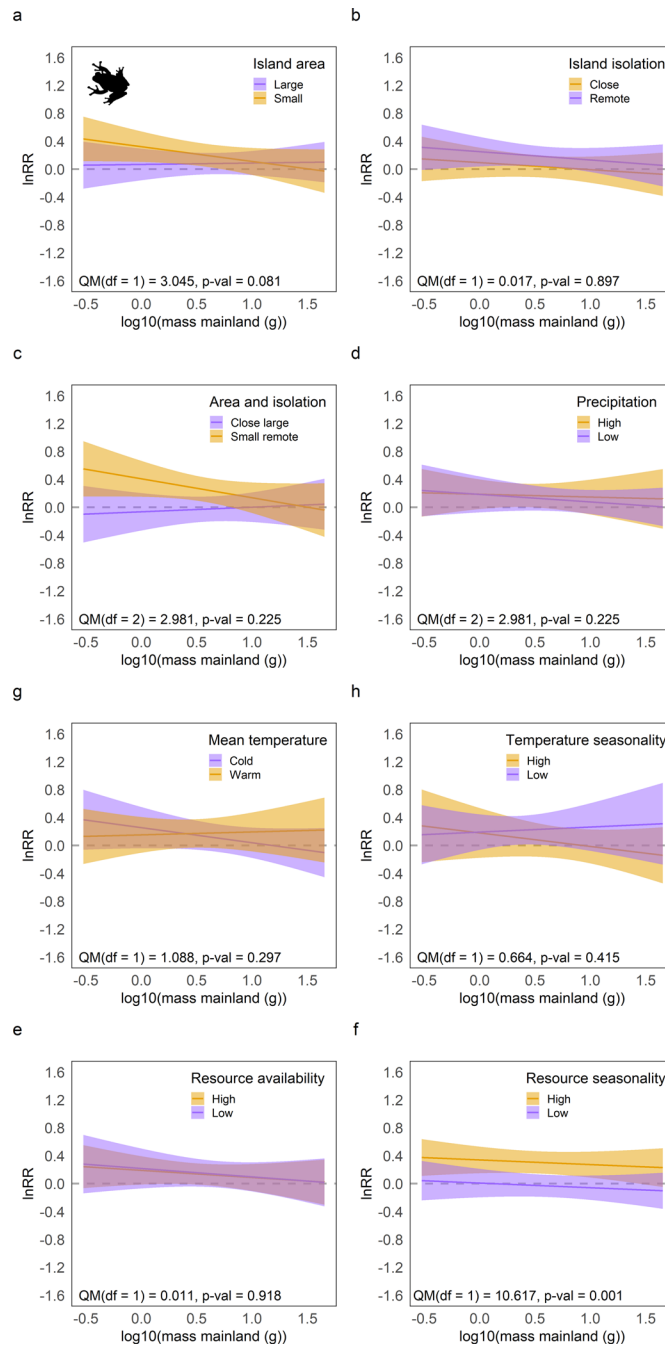
Extended Data Fig. 5 | Ecological factors explaining insular size shifts in mammals. Only 1-2 variables are displayed per plot, while keeping the other predictors at median values. Continuous variables are represented at the 10% and 90% quantile for each extreme (close vs remote or small vs large, and low vs high). $\ln RR > 0$ indicates gigantism, $\ln RR < 0$ indicates dwarfism, and $\ln RR = 0$ indicates no shift in body size in islands compared to mainland populations. Shaded areas represent 95% CI. QM indicates the explained heterogeneity (variance) by the interaction between each explanatory factor and body mass (for example mass:area in panel a), or the explanatory factor only in case of intercept-only models (for example temperature in this case). See Supplementary Table 7 for details.



Extended Data Fig. 6 | Ecological factors explaining insular size shifts in birds. Only 1-2 variables are displayed per plot, while keeping the other predictors at median values. Continuous variables are represented at the 10% and 90% quantile for each extreme (close vs remote or small vs large, and low vs high). InRR > 0 indicates gigantism, InRR < 0 indicates dwarfism, and InRR = 0 indicates no shift in body size in islands compared to mainland populations. Shaded areas represent 95% CI. QM indicates the explained heterogeneity (variance) by the interaction between each explanatory factor and body mass (for example mass:area in panel a), or the explanatory factor only in case of intercept-only models (for example temperature in this case). See Supplementary Table 7 for details.



Extended Data Fig. 7 | Ecological factors explaining insular size shifts in reptiles. Only 1-2 variables are displayed per plot, while keeping the other predictors at median values. Continuous variables are represented at the 10% and 90% quantile for each extreme (close vs remote or small vs large, and low vs high). $\text{InRR} > 0$ indicates gigantism, $\text{InRR} < 0$ indicates dwarfism, and $\text{InRR} = 0$ indicates no shift in body size in islands compared to mainland populations. Shaded areas represent 95% CI. QM indicates the explained heterogeneity (variance) by the interaction between each explanatory factor and body mass (for example mass:area in panel a), or the explanatory factor only in case of intercept-only models (for example temperature in this case). See Supplementary Table 7 for details.



Extended Data Fig. 8 | Ecological factors explaining insular size shifts in amphibians. Only 1-2 variables are displayed per plot, while keeping the other predictors at median values. Continuous variables are represented at the 10% and 90% quantile for each extreme (close vs remote or small vs large, and low vs high). InRR > 0 indicates gigantism, InRR < 0 indicates dwarfism, and InRR = 0 indicates no shift in body size in islands compared to mainland populations. Shaded areas represent 95% CI. QM indicates the explained heterogeneity (variance) by the interaction between each explanatory factor and body mass (for example mass:area in panel a), or the explanatory factor only in case of intercept-only models (for example seasonality in resources - sdNDVI in this case). See Supplementary Table 7 for details.

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection Data from figures was extracted with the package metaDigitise v1.0 in R 3.5.3

Data analysis Data was analysed using: 'R' v 3.5.3 and 'ArcMap' v 10.5

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

All data and code are available at https://github.com/anabenlop/Island_Rule and https://figshare.com/projects/Body_size_evolution_in_insular_vertbrates/89102

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	Phylogenetic meta-analysis of the generality of the island rule in vertebrates and potential mechanisms underlying insular size shifts. and
Research sample	We assembled a global dataset of 2,479 island-mainland comparisons for 1,166 insular and 886 mainland species of terrestrial vertebrates, including mammals (1,058 island-mainland comparisons), birds (695 comparisons) reptiles (547 comparisons) and amphibians (179 comparisons) spread over the globe. In total we included morphometric measurements of 154,875 mainland and 63,561 insular specimens from species covering a wide range of average body masses (0.18–234,335 g)
Sampling strategy	We collected data from articles included in a recent assessment of the island rule and previous compilations included in previous studies of reptiles, mammals, and birds, tracing original data sources when possible to extract original measurement data. We additionally sampled body size measurements from published studies that did not assess the island rule per se, or – in the case of birds – also from original morphometric data collected from museum and live specimens. Overall, we included information retrieved from peer-reviewed articles, live and museum specimens, and expedition reports.
Data collection	Morphological data obtained from the literature and from measurements of live caught individuals and preserved museum skins. Ecological data obtained from the literature. Environmental data was obtained from publically available spatially-explicit rasters.
Timing and spatial scale	Data collection started in February 2018 and continued up to November 2020, with major gaps in between due to focus on other projects. Note that all data come from literature except for data on museum specimens, which was collected for another study (Pigot et al. 2020. NEE 4, 230-239.
Data exclusions	We excluded problematic data, such as comparisons that were not supported by taxonomic or phylogenetic evidence, or which reported morphometric data restricted to single specimens or without sample size. In addition, we excluded comparisons based on extinct taxa since they are often known from very few or incomplete specimens. Excluded studies and reasons for exclusion are specified in Supplementary Dataset 3.
Reproducibility	No experiments were undertaken. All data and codes necessary to reproduce the findings of this study are available at https://github.com/anabenlop/Island_Rule
Randomization	Our sampled belonged to either island populations, or mainland populations of the same or closely related species. No randomization was required or applied.
Blinding	NA
Did the study involve field work?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals

No laboratory animals were used.

Wild animals

A significant proportion of the avian data are from museum specimens. Some data are included from wild-caught birds that were not harmed during data collection and subsequently released into the wild. In all cases, birds were caught by mist-netting, a passive, non-invasive technique which does not harm the individual birds.

Field-collected samples

No samples were taken from the field

Ethics oversight

Natural Environment Research Council

Note that full information on the approval of the study protocol must also be provided in the manuscript.