

Lower Enoplida of
the Seas of
the Soviet Union

List of Species of Lower Enoplida of the Soviet Union

Class NEMATODA

Subclass ADENOPHOREA

Order ENOPLIDA

1. Superfamily LEPTOSOMATOIDEA

1. Family LEPTOSOMATIDAE

1. Subfamily LEPTOSOMATINAE

1. Genus *Leptosomatum* Bastian, 1865

| | |
|---|----|
| 1. <i>L. arcticum</i> Filipjev, 1916..... | 76 |
| 2. <i>L. grebnickii</i> Filipjev, 1916..... | 77 |
| 3. <i>L. behringicum</i> Filipjev, 1916..... | 78 |
| 4. <i>L. bacillatum</i> (Eberth, 1863)..... | 79 |
| 5. <i>L. elongatum</i> Bastian, 1865..... | 81 |
| 6. <i>L. punctatum</i> (Eberth, 1863)..... | 82 |
| 7. <i>L. tetrophthalmum</i> Saveljev, 1912..... | 84 |
| 8. <i>L. breviceps</i> Platonova, 1967..... | 85 |
| 9. <i>L. gracile</i> Bastian, 1865..... | 86 |

2. Genus *Leptosomatides* Filipjev, 1918

| | |
|--|----|
| 10. <i>L. euxina</i> Filipjev, 1918..... | 89 |
| 11. <i>L. steineri</i> Filipjev, 1922..... | 90 |
| 12. <i>L. inocellatus</i> Platonova, 1967..... | 92 |
| 13. <i>L. crassus</i> Platonova, 1967..... | 93 |
| 14. <i>L. acutipapillosus</i> sp. nov..... | 95 |
| 15. <i>L. brevisetosus</i> sp. nov..... | 96 |
| 16. <i>L. marinae</i> sp. nov..... | 98 |

| | |
|--|-----|
| 3. Genus <i>Leptosomella</i> Filipjev, 1927 | |
| 17. <i>L. acrocerca</i> Filipjev, 1927..... | 100 |
| 2. Subfamily SYNONCHINAE | |
| 4. Genus <i>Synonchus</i> Cobb, 1893 | |
| 18. <i>S. murmanicus</i> Filipjev, 1927..... | 103 |
| 5. Genus <i>Anivanema</i> gen. nov. | |
| 19. <i>A. magna</i> sp. nov..... | 105 |
| 3. Subfamily THORACOSTOMATINAE | |
| 6. Genus <i>Deontostoma</i> Filipjev, 1916 | |
| 20. <i>D. papillatum</i> (Linstow, 1903)..... | 109 |
| 21. <i>D. arcticum</i> (Saveljev, 1912)..... | 110 |
| 22. <i>D. lobatum</i> (Steiner, 1916)..... | 112 |
| 7. Genus <i>Pseudocella</i> Filipjev, 1927 | |
| 23. <i>P. trichodes</i> (Leucart, 1849)..... | 116 |
| 24. <i>P. elegans</i> (Ditlevsen, 1926)..... | 118 |
| 25. <i>P. saveljevi</i> (Filipjev, 1927)..... | 120 |
| 26. <i>P. tenuis</i> Filipjev, 1946..... | 121 |
| 6 27. <i>P. coecum</i> (Saveljev, 1912)..... | 123 |
| 28. <i>P. gracilis</i> sp. nov..... | 124 |
| 29. <i>P. pseudocellum</i> (Filipjev, 1927)..... | 126 |
| 30. <i>P. bursata</i> Platonova, 1962..... | 126 |
| 31. <i>P. minor</i> Platonova, 1962..... | 128 |
| 32. <i>P. kurilensis</i> Platonova, 1962..... | 129 |
| 33. <i>P. mamillifera</i> Platonova, 1962..... | 132 |
| 34. <i>P. acuta</i> sp. nov..... | 134 |
| 35. <i>P. angusticeps</i> sp. nov..... | 135 |
| 36. <i>P. truncaticauda</i> sp. nov..... | 136 |
| 37. <i>P. rarisetosa</i> sp. nov..... | 138 |
| 38. <i>P. raddae</i> sp. nov..... | 139 |
| 2. Family ANTICOMIDAE | |
| 4. Subfamily ANTICOMINAE | |
| 8. Genus <i>Odontanticoma</i> gen. nov. | |
| 39. <i>O. dentifer</i> sp. nov..... | 143 |
| 40. <i>O. murmanica</i> (Filipjev, 1927)..... | 145 |

9. Genus *Anticoma* Bastian, 1865

| | |
|--|-----|
| 41. <i>A. eberthi</i> Bastian, 1865..... | 149 |
| 42. <i>A. insulaealbae</i> Filipjev, 1927..... | 151 |
| 43. <i>A. minor</i> Filipjev, 1927..... | 152 |
| 44. <i>A. arctica</i> Steiner, 1916..... | 153 |
| 45. <i>A. limalis</i> Bastian, 1865..... | 155 |
| 46. <i>A. pellucida</i> Bastian, 1865..... | 156 |
| 47. <i>A. novozemelica</i> (Filipjev, 1927)..... | 158 |
| 48. <i>A. brevisetosa</i> Platonova, 1967..... | 159 |
| 49. <i>A. filipjevi</i> Platonova, 1967..... | 160 |
| 50. <i>A. tenuicaudatooides</i> Gerlach and Riemann, 1974..... | 162 |
| 51. <i>A. grandis</i> Platonova, 1967..... | 162 |
| 52. <i>A. behringiana</i> sp. nov..... | 164 |
| 53. <i>A. curta</i> sp. nov..... | 166 |
| 54. <i>A. uschakovi</i> sp. nov..... | 168 |
| 55. <i>A. pacifica</i> sp. nov..... | 169 |
| 56. <i>A. pontica</i> Filipjev, 1918..... | 171 |
| 57. <i>A. platonovae</i> Sergeeva, 1972..... | 173 |

5. Subfamily PLATYCOMINAE

10. Genus *Platycomopsis* Ditlevsen, 1926

| | |
|--|-----|
| 58. <i>P. mesjatzevi</i> (Filipjev, 1927)..... | 175 |
|--|-----|

6. Subfamily BARBONEMATINAE

11. Genus *Barbonema* Filipjev, 1927

| | |
|--|-----|
| 59. <i>B. setifera</i> Filipjev, 1927..... | 179 |
|--|-----|

2. Superfamily ENOPLOIDEA

3. Family CRENOPHARYNGIDAE fam. nov.

12. Genus *Crenopharynx* Filipjev, 1934

| | |
|---|-----|
| 60. <i>C. marioni</i> (Southern, 1914)..... | 182 |
| 61. <i>C. gracilis</i> (Linstow, 1900)..... | 184 |
| 62. <i>C. armatus</i> sp. nov..... | 186 |

7

13. Genus *Nasinema* Filipjev, 1927

| | |
|---|-----|
| 63. <i>N. polare</i> (Steiner, 1916)..... | 187 |
| 64. <i>N. boreale</i> sp. nov..... | 188 |

4. Family RHABDODEMANIIDAE

14. Genus *Rhabdodemia* Baylis and Daubney, 1926

| | |
|---|-----|
| 65. <i>R. minor</i> (Southern, 1914)..... | 192 |
| 66. <i>R. gracilis</i> (Ditlevsen, 1919)..... | 193 |
| 67. <i>R. edentula</i> Platonova, 1974..... | 194 |
| 68. <i>R. latifaux</i> Platonova, 1974..... | 196 |
| 69. <i>R. hexonchia</i> Platonova, 1974..... | 197 |
| 70. <i>R. pontica</i> Platonova, 1965..... | 198 |
| 71. <i>R. marisalbi</i> Platonova, 1974..... | 199 |
| 72. <i>R. orientalis</i> Platonova, 1974 | 200 |
| 73. <i>R. angustissima</i> Platonova, 1974..... | 201 |

Introduction

ANATOMY AND MORPHOLOGY

8 **Size and shape of body.** As a group free-living marine Enoplida are large-sized nematodes. Of the families investigated, representatives of Leptosomatidae proved the longest with a length of 30 to 50 mm (10 to 20 mm on the average), while nematodes of Anticomidae and Rhabdodemaniidae are the shortest, not exceeding 7.0 to 8.0 mm (3.0 to 4.0 mm on the average). In family Crenopharyngidae forms somewhat longer than those in family Anticomidae predominate, ranging from 5.0 to 6.0 mm in length.

Nematodes are distinguished from many other organisms in that no clear-cut demarcation occurs between the head and the body. The following sections can be delineated in the body: 1) anterior trophic-sensory section, including oral organs, entire esophagus with chemo- and tangoreceptors, and the nerve ring, which is the main part of the nervous system; 2) middle trophic-genital section, with midgut and gonads; and 3) caudal section with caudal glands for attachment and such sensory organs as setae and papillae. A.A. Paramanov (1962) has observed that promorphologically two types of symmetries are evident in the trophic-sensory section of the body—bilateral symmetry as the base with radial symmetry superimposed. Organs situated in the trophic-genital section promorphologically correspond to bilateral symmetry in disposition. In the caudal section radial symmetry of the organs is observed, which should be considered a derivative of the bilateral symmetry. Thus radial structures are typical of the most mobile and active body parts, i.e., trophic-sensory and caudal sections, which serve as the central part of the sensory apparatus.

At first glance it would seem that the shape of the body in all nematodes is rather uniform. However, a closer scrutiny reveals that shape varies from filiform to pyriform. Variations encountered in free-living and plant nematodes are discussed here. It has been observed that in representatives of lower Enoplida a thin cylindrical and fusiform shape is much more common. A fusiform body is characteristic of nematodes of family Anticomidae; their body considerably tapers at both ends, narrowing in the region of the trophic-sensory and caudal sections. The tail in

anticomids is long and consists of two parts—base widely conical and rest extremely narrow and cylindrical.

In Crenopharyngidae the body shape is similar to that in Anticomidae but longer and tapering in the trophic-sensory section is not so abrupt. In representatives of family Leptosomatidae the body is slender, cylindrical, and tapers rather smoothly in the trophic-sensory part; the tail is usually short and bluntly rounded. In Rhabdodemaniidae the trophic-sensory end of the body narrows quite smoothly near the cephalic end; the tail is rather long, wide, and pointed at the tip.

As suggested by de Man the shape of the nematode body can be characterized by indices expressing ratios of its individual parts. In addition to these indices, measurements have also been given in the form of a formula as suggested by Cobb and modified by Filip'ev (1916).

Cuticle. Represented by a thick membrane covering the nematode body externally and also extending inside the oral aperture, esophagus, hind gut, vagina, and cervical pore. In all probability the cuticle of free-living nematodes plays a dual role—it protects their body from damage and serves as an external skeleton. It plays a particularly important role for nematodes inhabiting sea-breaker zones in which sharp particles of the bottom are moved about. The firmness of the cuticle in different groups of nematodes is ensured in various ways. In nematodes of the orders Chromadoridae and Desmoscolecidae the cuticle is annulated; it is also reinforced with different sclerotized structures such as spherical bodies, bands, denticles, etc., acquiring a great complexity. The cuticle of lower enoplids is relatively simple. In most cases it is absolutely smooth and very thick. In species of *Leptosomatum* and *Leptosomatides* extremely fine fascicles of criss-crossed fibers underlie the cuticle and play a supporting role, imparting great strength to the membrane. These fibers usually occur only in the anterior part of the nematode body.

On the surface of the cuticle longitudinal and lateral structures, termed "lateral fields," extend along both sides of the body. These lateral cuticular fields generally have their own pattern, which is distinct from the pattern of the cuticle. Along these fields small ridges occur, divided by longitudinal lines. The lateral fields lie against the longitudinal thickening of the hypoderm. Paramonov (1962) suggests that since the lateral field is situated between two masses of muscles (laterodorsal and lateroventral), it may be compared to a lateral cuticular crest, which is formed under the bilateral mechanical influence of these muscular masses. From this idea Paramonov concludes that the genesis of the lateral fields is associated with the locomotor function of the muscles. Thus the supporting role of the cuticle is revealed in the process of locomotion. This phenomenon (when external skeletal structures play a supporting role for musculature) is quite widespread in the animal world and hence nema-

todes are no exception in this respect.

Histologically the cuticle of enoplids is represented by a noncellular membrane secreted by the hypoderm. It is multilayered. As shown by Timm (1953), the cuticle in *Leptosomatum acephalatum* consists of six layers: an external homogeneous cortical layer, thin external layer with oblique fibers, thicker layer with fibers running at an angle to the previous layer, two layers similar to each other in structure which stain differently, and the innermost layer or basement membrane consisting of longitudinal fibers.

Studies of *Thoracostoma coronatum* and *Deontostoma magnificum* revealed that their cuticle is similar in structure to that of *Leptosomatum acephalatum*; hence Timm assumed that in all nematodes of family Leptosomatidae the cuticle is similar in structure. The cuticle lining the inner parts of the body consists of a different number of layers, that lining the rectum consists of three layers, while that lining the vulva consists of six and is sclerotized.

The relative thickness of the cuticle is directly related to its structure. The cuticle of enoplids, devoid of supportive structures, is significantly thicker than the annulated and sclerotized cuticle of chromadorids and
10 desmoscolecids, since its strength depends only on its thickness. For example, in *Cyatholaimus* and *Chromadora* (order Chromadorida) the thickness of the cuticle constitutes only 1/40 to 1/50 of the body width, while in enoplids the cuticle is much thicker.

Understandably in smaller forms, 2.0 to 5.0 mm long, the cuticle is thinner (2.0 to 5.0 μm) than in larger forms, up to 20 to 30 mm long, in which it may achieve a thickness of 17 to 20 μm . For this reason the absolute thickness of the cuticle should not be taken into consideration and only its relative thickness in comparison to size and thickness of the body examined.¹ Species of *Thoracostoma*, *Deontosoma*, and *Pseudocella* have a cuticle about 1/14 to 1/20 of the body width; *Leptosomatum* and *Leptosomatides*—1/15 to 1/18; *Synonchus* and *Cylicolaimus*—1/12 to 1/17; and *Anticoma*—1/14 to 1/20. The thickness of the cuticle reduces only rarely to 1/25 to 1/30 of the body width (as in the case of *Anticoma behringiana*, *Anticoma elegans* and *Anticoma filipjevi*). In Rhabdodemaeniidae the thickness of the cuticle constitutes about 1/12 to 1/16 of the body width.

Setae and papillae. These play the role of sensory organs. As outgrowths of the cuticle, they are thought to be derived from it. In lower enoplids a great variety in shape, size, and location is seen. Papillae are firmly attached to the body and immovable. Six in number, they are

¹Thickness of the cuticle and the corresponding width of the body were measured by me in the region of the nerve ring.

arranged in a circle around the oral aperture immediately behind the lips. In some free-living marine nematodes, labial papillae resemble setae (Filip'ev, 1921). In my material, *Leptosomatides acutipapillosus* possessed such setaceous papillae. In larger leptosomatids (*Thoracostoma*, *Deontostoma*, *Pseudocella*) papillae are well developed and distinctly visible. In Anticomidae labial papillae are arranged as in Leptosomatidae but due to their small size barely discernible. Contrary to females, which have only labial papillae, many males possess a system of preanal papillae (see p. 37).

Setae are the same in origin as the papillae but articulate with the body. All the nematodes of the families described by me have a crown of ten cephalic setae located immediately behind the cephalic capsule; eight are situated in pairs submedially and two single ones situated laterally. If interlobular grooves (p. 37) are present, the cephalic setae are located in them. The shape and size of the cephalic setae vary considerably within the limits of a genus.

In addition to cephalic setae, in many free-living nematodes setae also occur in the preneural region (anterior end of the nematode body from the head to the nerve ring). However, among leptosomatids these setae are absent in a number of species. In *Leptosomatum kerguelensis* they are rudimentary and transformed into papillae; in *L. clavatum* these setae are totally absent. It should be noted that in nematodes of *Leptosomatum* these setae are generally very small and, probably, tending toward reduction. In species of *Thoracostoma*, *Deontostoma*, and *Pseudocella* setae are invariably present but again generally small in size.

The setaceous armature in species of Synonchinae (family Leptosomatidae) varies notably. *Synonchus murmanicus* has a few small setae dispersed in the preneural part of the body. *Anivanema magna* is armed with long and powerful setae situated in regular longitudinal rows. In *Eusynonchus hirsutus* the entire preneural part is covered with longitudinal rows of fine and long setae.

The preneural setae in *Anticoma* (family Anticomidae) warrant special attention. Their arrangement in two longitudinal lateral rows, each 11 comprising three to eight setae, seems a characteristic feature of this genus. There are no setae outside these two rows in the preneural region of *Anticoma*. As shown by Micoletzky (1930) and Wieser (1953b), the number of setae changes with age. Adult *Anticoma campbelli* (Wieser, 1953c) have four to five setae in each row, while younger forms have only one to two. Hence until the last molt the number of cervical setae in *Anticoma* cannot be established.

The nature of the cervical setae in *Paranticoma*, and also in members of the family Anticomidae, is very interesting. These setae merge into a powerful spine inside which lies the duct of the cervical gland.

In addition to cephalic and cervical setae, males of many nematodes have a group of preanal setae, functionally entering into the composition of the copulatory organs (p. 37).

Lastly, small setae are present at the tip of the tail near the aperture of the caudal glands.

Head. A study of the head of nematodes of the order Enoplida is very important in the systematics of this group. Complex formations in the anterior end of the body, partly from the external cover (cuticle) and partly from the anterior end of the digestive system, together constitute a single structure in a morphological and functional sense. The degree of development and individual details of structure of these formations are considered the most important taxonomic features of the order Enoplida.

Works by Wieser (1953a) and Inglis (1964) in particular present a comparative anatomical analysis of the anterior body end of enoplids. Due to the fact that some of the propositions by Wieser were subjected to detailed criticism by Inglis, the scheme of the latter author appears better founded and I have utilized his terminology, translating some terms into Russian. The internal layer of the cuticle is significantly thickened at the place where the anterior end of the esophagus comes close to it; this thickening, resembling a cylinder or truncated cone, is called the cephalic capsule (Figure 1, 8). According to Inglis (1964) it consists of an endo- and mesocuticle. The outer covering of the anterior end of the esophagus also has a thickened cuticle. This thickening is labeled the esophageal capsule (Figure 1, 9) (according to Wieser, 1954—pharyngeal capsule). The junction of the esophageal and cephalic capsules is accentuated by a thickened wall encircling the body, which is called the cephalic ring (Figure 1, 5). Wieser calls this structure the “stomodaeal ring”. However, the former name appears more appropriate because Inglis showed that nerves innervating the labial sense organs always extend under this ring; for this reason the latter should be associated with the cephalic capsule and not with the esophagus. One more sclerotized structure—the cervical capsule—which is distinguished from the cephalic capsule by striations or punctation, is situated behind the cephalic capsule, often merging with it and extending into it (Figure 1, 10).

In Leptosomatidae and Anticomidae the cephalic capsule extends forward beyond the cephalic ring, a feature not seen in representatives of a number of other families of the order. However, its anterior part is always more weakly developed than the posterior part. Sometimes the anterior and posterior parts are so well demarcated that the cephalic capsule seems to consist of two separate elements. Kreis (1928) observed this phenomenon in species *Pseudocella saveljevi* (under the name *Thoracostoma conicaudatum*) and *Pseudocella filipjevi* and spoke of primary and secondary capsules (*primaer- und sekunderkapsel*).

In a number of cases, for example in *Thoracostoma*, *Deontostoma*, *Pseudocella*, and *Parabarbonema*, the anterior margin of the cephalic capsule forms six protuberances supplied with nerves; these extend toward
 12 the labial papillae and thus play the role of supporting structures (Figure 1, 11). The posterior part of the cephalic capsule may be divided into six lobes—four sublateral, one dorsal, and one ventral (Figure 1, 12). The lobes are separated from each other by interlobular grooves or furrows resembling arches or narrow fissures, each of which generally has a round anteriorly widened portion—the fenestra (Figure 1, 13). The lateral interlobular grooves, in which the amphids are situated, are much wider than the others. The margins of the lobes of the cephalic capsule may be smooth, or uneven, or wavy with more or less deep marginal grooves. The lobes may or may not exhibit openings or loculi of various shapes.

The oral cavity commences with the mouth and communicates with the esophageal cavity in the simplest cases (Figure 1, 5). With the development of sclerotized lips and deepening of the oral cavity, the oral opening varies in shape, becoming hexagonal or even circular for example. The oral cavity of Leptosomatidae in the original form is almost nil (for example in *Leptosomatum*). However, in other Leptosomatidae and also in other families of the order, it may change substantially. In species in which the esophagus reaches almost up to the anterior end of the body, as usually happens in Leptosomatidae and Anticomidae, the oral cavity remains undivided. However, in most families of the order the esophagus does not reach the anterior end; consequently two parts of the oral cavity communicate with each other—the onchial cavity, formed by the widening of the anterior esophagus (Figure 1, 7), and the buccal cavity situated in front of the onchial cavity (Figure 1, 3). Differences in the nature of the oral armature correspond to this division. The onchial cavity may exhibit falcate or lamellar processes—the onchia (Figure 1, 6). The onchia move by the action of the musculature of the esophagus, which undergoes specialization at this place and splits into separate muscular fascicles. All the structures of the buccal cavity are not directly connected with the musculature and are brought into motion only through the displacement of three thickened plates formed from the lining of the onchial cavity. The armature of the buccal cavity may be of two types—small denticles situated individually or in groups (odontia), or as in the case of Enoplida plates of thickened cuticle in the cavity (jaws) (Figure 1, 2). The jaws, as shown by Inglis (1964), form through the fusion of the cuticular lining of this cavity with the so-called buccal rods (longitudinal thickenings or ridges formed from the cuticle), and play the role of supporting structures in forms devoid of odontia. Due to the fact that in Leptosomatidae and Anticomidae a distinct division into buccal and onchial cavities does not appear, onchia in representatives of these families may be arranged near

the oral aperture and never form jaws; the only buccal armature in this case comprises odontia, or plates formed by their fusion. Moreover, odontia may be arranged at the level of the anteriormost onchia. Such an arrangement is in conformity with the circumstance that the esophageal musculature, in the form of anterior specialized muscles, extends far in front and is attached to the bases of the onchia.

In the anterior part of the body of representatives of order Enoplida a number of different cuticular elements are present, such as the cephalic capsule with the cuticle of the body wall covering both sides, cuticular lining of the oral cavity, and the esophageal capsule. These three structures, together with the tissues underlying them, confine externally both dorsally and ventrally a ring-shaped space filled with a liquid and termed the cephalic bladder (Inglis, 1964) (Figure 1, 4). Strictly speaking the cephalic bladder is precisely ring-shaped only in those forms in which the esophageal capsule is situated far away from the anterior end of the body and the oral cavity is narrower than the diameter of the integument around it. Even in such a case, however, it should be borne in mind that the oral cavity is trigonal and the entire ring appears to be represented by three separate parts corresponding to the sections of the esophagus, divided by narrow crosspieces in those places where the corners of the esophageal space come extremely close to the body wall (along the radius of the esophagus). In *Leptosomatidae* and *Anticomidae*, however, it often happens that the cephalic bladder is subdivided not into three, but into six parts due to specialized muscles extending toward the onchia and passing along the middle of the esophageal sections, crossing the parts of the cephalic bladder corresponding to each section, and thus dividing it into halves (three sections, each divided into two, for a total of six). The size of the bladder depends on the distance between the anterior body end and the esophagus. In *Leptosomatium* the esophagus extends almost to the anterior body end and hence there is no cephalic bladder. In *Anticoma* and genera close to it the cephalic bladder is very poorly developed. In those genera (*Parabarbonema*, *Synonchus*, *Thoracostoma*, and others) in which the esophageal capsule falls short of the anterior end by a significant distance, the cephalic bladder is well developed.

When the oral cavity is significantly developed, for example in *Cylico-laimus* and genera close to it, the size of the cephalic bladder shrinks due to an increase in the transverse diameter of the space of the oral cavity.

Thus a detailed study of the cephalic end makes it possible to draw some conclusions regarding the taxonomic structure of lower enoplids. In representatives of *Leptosomatidae* two lines of development can be traced: first, reinforcement of the cephalic and esophageal capsule and development of the cephalic bladder; and second, development of the oral cavity. *Anticomidae* is characterized by poor development of the cephalic capsule and the oral cavity.

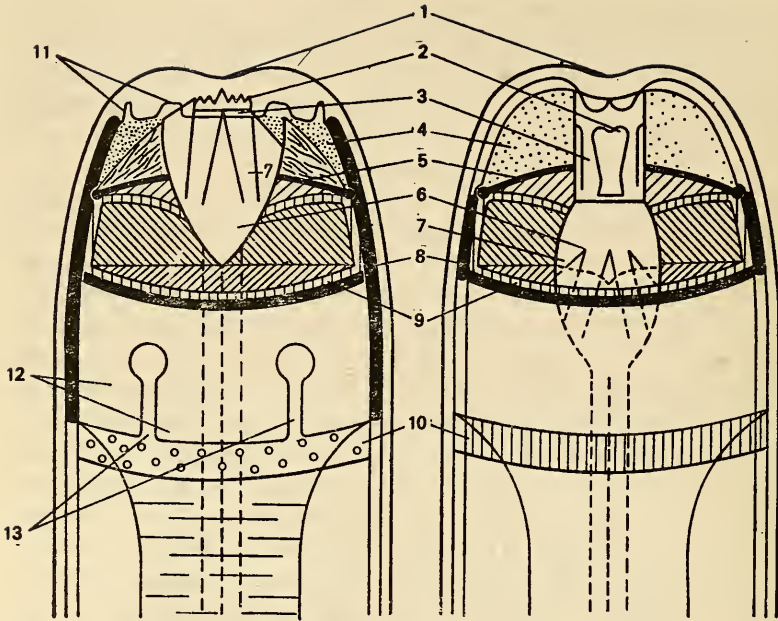


Figure 1. Scheme of head structure of nematodes of superfamilies Leptosomatoidea (left) and Enoploidea (right).

1—oral aperture; 2—odontia (mandibles); 3—buccal cavity; 4—cephalic bladder; 5—cephalic girdle or ring; 6—onchia; 7—onchial cavity; 8—cephalic capsule; 9—esophageal capsule; 10—cervical capsule; 11—anterior protuberances of cephalic capsule; 12—lobes of cephalic capsule; 13—interlobular grooves.

Hypoderm. The compact layer of the hypoderm, replacing the epithelium, underlies the cuticle which covers the body. The hypoderm has a distinctly cellular structure in free-living marine and a few fresh-water nematodes; it is syncytial in parasitic and most plant nematodes. The nuclei of the cells of the hypoderm disintegrate into a number of smaller nuclei (Martini, 1909). Localized thickenings of the hypoderm press into the pseudocoel and form four longitudinal cords—usually two lateral, one ventral, and one dorsal.

In nematodes of the order Enoplida there are eight hypodermal cords. In addition to the four mentioned above, four submedial cords are present. The cells of these cords are usually more stretched in width than in length. The hypoderm situated between the cords contains no nuclei. In transverse section the number of cells forming these cords differs in different leptosomatids. Thus three cells occur in *Synonchus strasseni* (Türk, 1903), five to six cells in *Thoracostoma setosum* (de Man, 1904), and four

to five cells in *Leptosomatium acephalatum* (Timm, 1953). According to the observations of Filip'ev (1916) the greater number of cells in the hypoderm is related to their increase in other organs also. According to Timm (1953) the dorsal and ventral cords in leptosomatids commence as a double row of large cells at the level of the amphids, while the lateral ones commence as a single row of large cells immediately behind the amphids, but beyond the eyes change into a layer consisting of a large number of cells (there are three rows of cells in the case of *Leptosomatium acephalatum*). Dorsal and ventral cords are wide anterior to the nerve ring but narrow behind it. Cells in the hypodermal cords are usually arranged in a single row between the eyes and the nerve ring. Beyond the nerve ring these cells reduce greatly in size (especially in the dorsal cord) and hence it is difficult to detect them. Lateral cords widen toward the base of the esophagus, become significantly thickened, and then extend along the entire body. These cords flatten, especially in sexually mature females, in the gonadal region. Throughout their length from the esophagus to the anus the lateral cords look like a thin, single, stretched cell, while the ventral cord alternately appears single-celled and double-celled. In the postanal region the ventral cord, wide in transverse section, consists of two cells. The dorsal cord disappears altogether but the lateral cords extend right up to the caudal glands and terminate at some distance from their openings.

The cells of different cords vary in form as well as in function (Filip'ev, 1921). Those of the dorsal and ventral cords accumulate fat and hence their shape is usually round, while those of the lateral cords may be rectangular or highly stretched.

Musculature. The nonspecialized musculature of the nematode is represented by a single layer of elongated, fusiform muscle cells underlying the hypoderm. It is divided into longitudinal muscular fields by the hypodermal cords. According to Schneider (1866) all nematodes can be divided into two groups on the basis of the number of cells in the muscular fields viz., "polymyarian" (large number of cellular rows in each muscular field with varying number of cells) and "meromyarian" (reduced and fixed small number of cells in each field). All representatives of order Enoplida belong to the first group. According to Timm (1953) in *Leptosomatium acephalatum* there are six to seven rows of cells in each muscular field in the anterior esophageal region, but commencing from the nerve ring and right up to the terminus the number of cell rows increases to ten to twelve.

15 As already mentioned, the somatic muscle cells of nematodes are fusiform. That part of the cell pressing (bulging) into the pseudocoel consists of only sarcoplasm with a nucleus and without myofibrillae. Myofibrillae are situated in the remaining part either in the shape of a

longitudinal groove (coelomyarian type) or in the shape of a flat field (platymyarian type). The specialized musculature of nematodes of the family Leptosomatidae was studied in detail by Timm (1953), but mainly in *Leptosomatium acephalatum*.

The valve of the caudal glands is stretched backward by a thin muscle which passes through the middle of these glands. In nematodes of *Leptosomatium* and *Thoracostoma* the retractor of the caudal glands has nuclei in its dorsal wall.

Esophageal and intestinal musculature are represented by short muscles extending from the body wall to the esophagus and intestine. These muscles are barely separated from each other and lie somewhat above or below the cardia. They support the mesenteric tissue, which at the level of the cardia is rather profuse, and exhibit no fixed pattern in disposition.

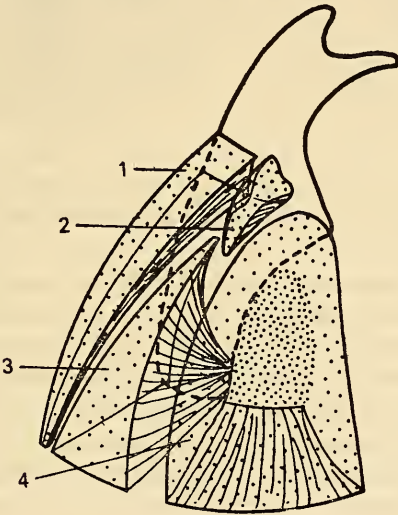
De Man (1886) described four median muscles in *Enoplus communis* of the family Enoplidae, proceeding anteriorly from the base of the esophagus, merging with four sublateral muscles, and binding them with the body near the submedian line. Each muscle consists of a single cell. Türk (1903) described similar musculature for *Synonchus* and Timm (1953) for *Leptosomatium*.

The structure of the specialized musculature of the head of enoplids was studied in detail by Inglis (1964); hence in analyzing these structures I have used his information extensively.

The musculature of the head is actually the end results of the transformation of the anterior end of the esophagus, which developed a series of separate muscles corresponding to the cephalic capsule. All these muscles are attached to the onchial region; the armature of the buccal cavity is completely devoid of musculature. These muscles originate from the external surface of the esophagus. The muscles of the head are represented, so to speak, by two rings situated transversely to the body axis, one of which lies inwardly and somewhat behind the other. Let us tentatively label the first ring posterior and the second anterior. Each ring consists of a few muscles. Moreover, their definite combination repeats itself in each esophageal section. In each section of the esophagus there are three muscles in the anterior ring and four in the posterior one, which are so arranged that the ends of the muscles of the anterior ring pass through the initial parts of the muscles of the posterior ring. In the anterior ring there is a medial unpaired muscle, narrow and thin at the 16 posterior margin but widening and thickening toward the anterior end (Figure 2, 1). It is bordered by a pair of muscles which are always significantly smaller and thinner (Figure 2, 2). In the posterior ring paired medial muscles are also found but they are separated from each other by the initial part of the medial muscle of the anterior ring (Figure 2, 3). Paired lateral muscles (Figure 2, 4) also border the medial muscles of the

posterior ring. The muscles of the anterior ring extend forward and inward up to their attachment with the onchial plate. The medial paired muscles of the posterior ring are also attached to the onchial plate but situated somewhat lower than the muscles of the anterior ring. The lateral muscles of the posterior ring are attached to the thin cuticle fringing the onchial plate.

The nonspecialized musculature of the esophagus in which the muscles are situated radially follows the specialized muscles of the head. This changeover from the two rings of specialized muscles to the radial musculature of the esophagus takes place abruptly with no distinct zone of transition. It should be noted that the opening through which the nerve passes to the labial sensory organs occurs between the medial and lateral muscles of the posterior ring.



15 Figure 2. Cephalic musculature of Enoplidae (only one section shown) (from Inglis, 1964).

1—medial unpaired muscle of anterior ring; 2—paired muscle of anterior ring; 3—medial paired muscle of posterior ring; 4—paired lateral muscle of posterior ring.

A similar arrangement of specialized muscles is mainly characteristic of representatives of family Enoplidae, which were studied by Inglis. I am inclined to assume that a similar scheme of muscular arrangement is also characteristic of lower representatives of the order. The beautiful diagrams of de Man (1904) substantiate this assumption, where in transverse sections taken in the cephalic region of *Deontostoma antarcticum* and *Thoracostoma setosum* all the muscles described by Inglis (Figure 3)

can be easily traced. It should be noted that in Anticomidae and the majority of Leptosomatidae the cephalic muscles pass forward through the cephalic bladder, dividing the latter into six small pockets or obliterating it altogether.

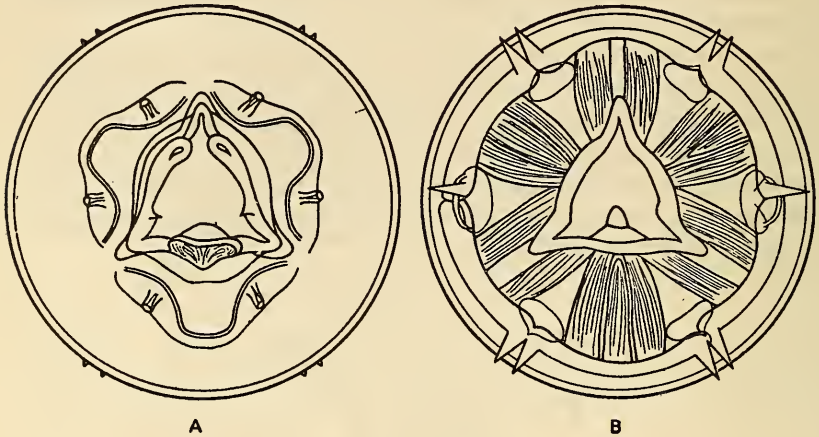


Figure 3. Structure of apical end of head of *Thoracostoma* (section at level of labial papillae—A; cephalic setae—B) (from de Man, 1904).

Specialized musculature of female. A V-shaped muscle extends from the subdorsal body wall to the dorsal part of the rectum, which originates from the posterior part of the rectum. This muscle bears two thin lateral connectives which approach the middle part of the caudal glands and are bound to the dorsal body wall by a third connective. Large nuclei lie in the central part of these connectives. Two small muscles, encircling the lower part of the caudal gland and extending toward the anterior part of the rectum, originate from the subdorsal body wall. These muscles raise the dorsal body wall during defecation.

Ten to twelve pairs of muscles, extending from the ventrolateral body wall toward the vagina and vulva, originate from the anterior and posterior parts of the vulva. The lateralmost pair bind the uterus and the vagina, while the centralmost pair bind the lips of the vulva. There is one nucleus in each of these muscles. All the nuclei are situated at one level, i.e., near the place of origin of the vulval muscles from the body wall. The number of pairs of these muscles probably varies in different species. Thus in *Leptosomatium acephalatum* there are 10 to 12 such muscles, while in *Thoracostoma coronatum* there are 6 to 8.

Five to six pairs of muscles, extending from the dorsolateral wall to the dorsal side of the vagina, are situated in the region of the vulva. They widen the vagina at the time eggs pass into it from the uterus. Two thick

muscles, each with one large nucleus, lie anterior and posterior to the vagina. These close the vulval opening. Two muscle cells lie dorsal to the vulva. Probably, they also assist in closing the vulval opening. In *Thoracostomum coronatum* there are two very large muscles on each side of the vagina which keep it closed.

The sphincter of the uterus is a highly reduced muscle situated near the uterine neck just ahead of the vagina. The muscular layer which on widening forms the sphincter extends further in the shape of a thin muscular investment around the vagina.

Specialized musculature of male. The V-shaped anal muscle of the male extends from the subdorsal wall of the body toward the posterior anal lip. The nucleus of this muscle lies near the body wall.

The copulatory musculature of the male is represented by numerous, thin, paired muscles (about 40 pairs), extending from the ventral side of the lateral walls to the subventral side of the body and terminating in the anterior part of the anus. Two to three pairs of short muscles extend from the ventrolateral wall of the body to the anterior lip of the anus; their function is to lift the anal lip. Türk (1903) described the copulatory muscles situated behind the anus in the case of *Synonchus*. Timm (1953) could not find similar muscles in *Leptosomatium acephalatum*.

The retractor musculature of the spicules is represented by paired muscles originating dorsal to the body wall, extending toward the lateral wall, and reaching the capituli of the spicules.

The protractor musculature of the spicules also consists of paired muscles. They form a longitudinal layer on the dorsal surface of the spicular pouch. Their nuclei are elongated.

The retractor musculature of the gubernaculum likewise consists of paired muscles which extend from the dorsal wall of the body and pass by the proximal end of the gubernaculum. In males the intestino-somatic musculature commences immediately after the esophagus, and consists of a thick muscular projection which surrounds the intestine but does not adhere to it. This projection is actually a series of irregular blocks of muscular tissues, which are attached to the somatic muscular layer or the body wall. Nuclei are usually found in the sarcoplasm of the muscular layer but may also be situated between the muscular projection (case) and the somatic layer. The testes and vas deferens may also be partially surrounded by the muscular case in the posterior part of the body. The muscular case then consolidates in large muscular blocks which are attached to the testes beyond their distal end.

Glands. These are always unicellular. In leptosomatids glands of the lateral fields look like large cells with a dark granular content. They are connected to the body wall by slender ducts. Near the outlet of these ducts a seta is often present. Bastian (1865) was the first to discover these

glands. Subsequently they were described by de Man (1893) in *Thoracostoma coronatum*; by Marion (1870) in *Thoracostoma zolae*; by Jägerskiöld (1901) in *Cylicolaimus magnus* and *Jaegerskioeldia acuticaudata*; by Türk (1903) in *Synonchus*, *Tuerkiana*, and *Paratuerkiana*; by de Man (1904) in *Thoracostoma setosum* and *Deontostoma antarcticum*; and by Steiner (1916) in *Leptosomatides steineri*.

Marion thought these glands served an adhesive function, secreting a substance whereby the nematode attaches itself to the substratum. Jägerskiöld assigned an excretory function to these glands. Zur Strassen (1904) considered them supporting cells of the nerve endings. Filip'ev (1921) deemed their nature glandular; surprisingly he reports these glands mostly for leptosomatids, whereas earlier authors mention them only for certain species in other groups of nematodes.

The cervical gland or renette is represented by a large unicellular structure in the anterior part of the nematode body (Figure 4). This gland is filled with a white secretion and stains darkly with hematoxylin. Two types of cervical glands are distinguishable: pyriform, with thick body, and short and slender duct; and tubular, with long stretched body, and long, thick duct. The renette body commences usually in the anterior part of the intestine or slightly above it, i.e., at the base of the esophagus. Usually this gland lies along the ventral line but sometimes it bends toward the dorsal side. In *Anticoma* (family Anticomidae) this gland is much larger and hence readily detected. It usually commences from the

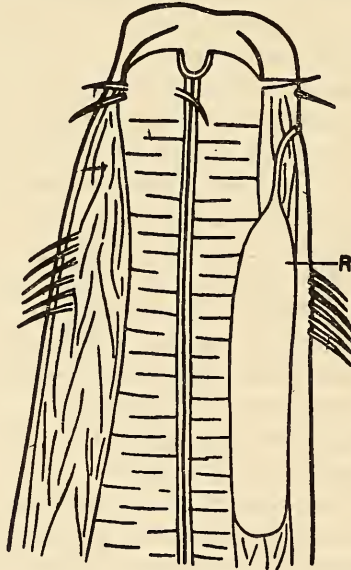


Figure 4. Anterior end of *Anticoma* (R—renette).

base of the esophagus and may open either in the pre- or postneural region. The position of the opening serves as a diagnostic character for species identification (pp. 148–174). Sometimes the glandular pore is highly displaced toward the anterior end and may open at the level of the amphids.

In species of Leptosomatidae the presence of the renette has proved controversial. According to Jägerskiöld (1901), Türk (1903), Steiner (1916), and Timm (1953) even the most thorough examination did not reveal the presence of this gland in *Synonchus*, *Tuerkiana*, *Paratuerkiana*, *Cylicolaimus*, *Thoracostoma*, nor in *Leptosomatium acephalatum* and *Leptosomatides steineri*. Filip'ev (1916) described the cervical pore in *Deontostoma papillatum* and de Man (1893) in *Thoracostoma coronatum*; the cervical gland per se they declared absent. Filip'ev (1921) expressed the possibility that the gland disappears during the cycle of development of the nematode.

The nature of the function of this gland was resolved after a long debate. Golovin (1902), Rauther (1907, 1909), and Stefansky (1922) investigated this problem experimentally by staining nematodes with vital stains. They arrived at the same conclusion—that the renette serves as an organ of excretion and osmoregulation. Stefansky (1922) and Paramonov (1962) positively attribute these functions to the renette but do not consider it the only organ of excretion. The excretory function is carried out by the renette, caudal glands, esophagus, and intestine.

In nematodes of the subclass Adenophorea the renette is compact and massive; a dendroid and very complex structure is characteristic of the renette in all members of the subclass Secernentea.

19 As indicated by Paramonov (1962), with a reduction in the protonephridial system in the ancestors of nematodes, a need for its replacement arose. In the earlier nematode stages of phylogenesis the excretory system was probably very bulky and consisted of a number of dermal glands. Afterwards, during the process of oligomerization three caudal and one cervical gland formed.

It seems to me that the absence of a renette in species of Leptosomatidae points to the fact that the process of its formation is still in the initial stages. In those forms in which this gland is absent its function is probably carried out by the glands of the lateral fields, caudal glands, the esophagus, and the intestine. The presence of glands in the lateral fields in leptosomatids (Filip'ev, 1921) also seems to support this idea. The fact that the renette also happens to be an organ of osmoregulation is substantiated by its absence in typical marine leptosomatids in which there is little need for such. Representatives of Anticomidae, largely found in pure fresh-water bodies, require a special organ of osmoregulation and the renette is quite well developed in anticomids. Its presence also bespeaks

the higher organization of Anticomidae.

There are three caudal glands, one on the dorsal side and two on the ventral.

Leidig (1854), followed by Eberth (1863), described these glands in the caudal region of nematodes. These scientists introduced the term "caudal gland". In structure these glands are reminiscent of the cervical gland and stain deeply with hematoxylin (Jägerskiöld, 1901). Like the cervical, two types of caudal glands are also distinguishable—tubular and pyriform. In lower enoplids both types are found (Figure 5), while various types may be seen in closely related forms. Hence this character lacks taxonomic significance. As can be seen from Figure 5, between the two aforementioned types of glands intermediate forms occur. All three caudal glands open at the extremity of the tail through a common caudal pore situated, most often, terminally. Occasionally this pore is somewhat displaced toward the ventral side. The caudal cone, sometimes longitu-

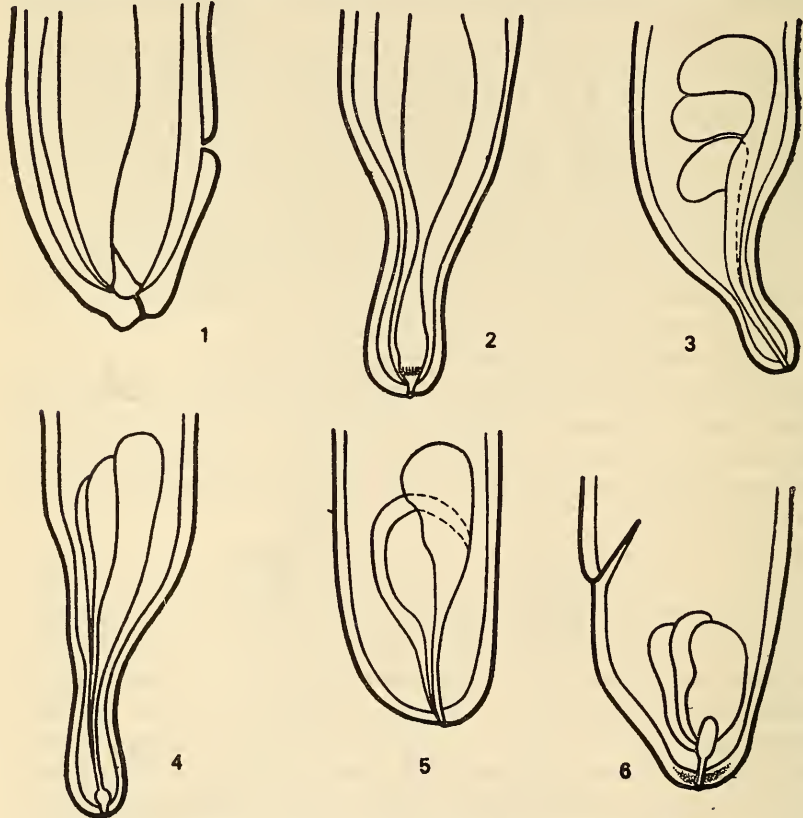


Figure 5. Shapes of caudal glands in lower Enoplida.

1—*Leptosomatum*; 2 to 4—*Synonchus*; 5—*Rhabdodemia*; 6—*Leptosomatides*.

nally striated (Figure 5, 1 and 2), usually lies in this pore. Cuticular folds usually develop around the caudal cone and provide support. The substance secreted by the caudal glands becomes firm on contact with water and hence these glands have been labeled "adhesive glands".

Body cavity. Primary form of body cavity—schizocoel devoid of epithelial lining and formed as a result of dispersion and disintegration of mesodermal cells. In more primitive nematodes it is usually difficult to observe the schizocoel as it is filled with nerve, muscle, and phagocytic cells. This led to the denial of its existence by Jägerskiöld (1901) in *Cyllocolaimus* and *Jaegerskiöldia*, by Türk (1903) in leptosomatids of the Mediterranean Sea, and by Timm (1953) in species of *Leptosomatium*, since none of these authors could detect a schizocoel in their material. De Man (1904) discovered only narrow slits in *Thoracostoma setosum*.

In intestinal and saprozoic nematodes living in an anisotonic medium the body cavity is represented by larger spaces containing liquid. In the changeover of nematodes to tissue parasitism in an isotonic medium the body cavity disappears and is filled with inflated cells which are parenchymatous in character (Filip'ev, 1937).

Phagocytic cells are located inside the body cavity. They are spherical and filled with a lustrous granular content. They have been studied in great detail by Golovin (1901). In nematodes placed in vital stains these cells stain quite rapidly. Other cells lose their stain on storage while phagocytic cells retain their stain for a long period; furthermore the stain sometimes crystallizes in their cytoplasm. These cells have an acidic reaction. Golovin directly proved their phagocytic nature; in transected worms they ingested India ink directly from sea water. Shimkevich (1899) was the first to record their phagocytic nature.

Phagocytic cells are scattered in large number throughout the length of the nematode body. According to Filip'ev (1921), the number ranges from 100 to 220 in nematodes of the order Enoplida. In some nematodes their disposition is irregular, while in others they are arranged mainly along the ventral line. Such a type of arrangement is characteristic of species of *Leptosomatium* and *Leptosomatides*.

These cells are more systematically disposed in more highly organized families of free-living nematodes such as Cyatholaimidae, Chromadoridae, and Axonolaimidae but their number markedly reduced. In *Axonolaimus setosus*, for example, there are only two large phagocytic cells (Filip'ev, 1918, 1921). They are comparable to the large phagocytic cells of parasitic nematodes.

Lipid cells are also present in the body cavity of nematodes. True, they have not been studied much and information about them is very scanty. Türk discovered such cells in *Synonchus strasseni*. They are dendritic in shape and situated around the intestine. Rauther (1907) observed

them in *Cylicolaimus* and *Thoracostoma*. Shimkevich (1899) and Golovin (1901) considered them homologues of phagocytic cells.

The observations of Timm (1953) in his more recent works should be noted. He has described similar cells in *Leptosomatum acephalatum*. In the female a paired row of such cells lies freely in the body cavity along the uterine region. A large number are disposed around the oviduct. The nucleus of these cells is located in the center of the cellular body. Some
21 of these cells are bifurcated (bifid) at one end. They are supported by the mesenteric connectives which link them with the body wall.

In *Enoplus* these cells branch and form a network on the intestine. They do not branch in *Deontostoma magnificum*.

Nervous system. The nervous system of nematodes parasitic in animals has been studied in detail, that of phytonematodes less so, and that of free-living nematodes almost not at all. Details of the nervous system of *Paroncholaimus zernovi* (family Oncholaimidae, order Enoplida) have been given by Filip'ev (1912, 1921), providing a basis for assuming that within the limits of the order Enoplida the general structural plan of the nervous system is similar. The scheme proposed by Filip'ev has been followed here. The nervous system of nematodes represents an extremely low level of organization. It is usually located in the hypoderm and only motor nerves and some part of the central nervous system distinguishable. The nerve ring or so-called circumesophageal ring constitutes its core and encircles the esophagus in its anterior half. In various species of Enoplida this ring occupies a different position in this part of the esophagus. Fibers and a small number of nerve cells enter into the composition of the nerve ring; Filip'ev (1912) has distinguished three types on the basis of their composition:

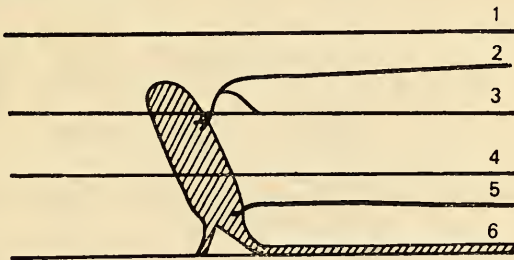
1. Total absence of any semblance of ganglia; nerve cells arranged in a single layer around the esophagus.

2. Barely discernible ganglia (which, as a matter of fact, are only groups of cells).

3. Readily distinguishable ganglia, densely fused with special "accompanying" and "supporting" cells.

Only the first type is seen in Enoplida—total absence of ganglia and presence of just a single layer of nerve cells around the esophagus. Nerve cords in the body of nematodes of the order Enoplida are situated as follows (Figure 6): The major and most powerful nerve—the ventral nerve, a pair of subventral nerves, a pair of subdorsal nerves, and a dorsal nerve located in the postneural part of the body. Two nerves occur on each side in the region of the lateral fields; Filip'ev labeled them the upper and lower lateral nerves. The ventral nerve splits into superficial inner fibers. The latter extend from the hypoderm and not reaching the level of the nerve ring join it in two nerve roots; the superficial nerve fibers

22 extend into the hypoderm right up to the level of the nerve ring and, here, like the inner fibers join the nerve ring in two nerve roots in the plane perpendicular to the body surface. Ultimately a small part of the superficial fibers extends further, into the preneural section. The subventral nerves merge with the roots of the inner fibers of the ventral nerve at the place of its junction with the nerve ring. Somewhat behind the ventral nerve the upper lateral nerves divide into two groups of nerve fibers, one extending into the preneural part and the other forming the nerve root joining the nerve ring. Subdorsal nerves take a sharp turn and from the hypoderm where they are located run deep and merge with the roots of the upper lateral nerves. The lower lateral and dorsal nerves do not communicate with the nerve ring and extend past it into the anterior part of the body. Thus six nerve cords of the postneural part extend into the preneural part—the dorsal, two dorsolateral, two ventrolateral, and the ventral. Two pairs of submedial and one pair of lateral nerves are situated deeper than these trunks, near the esophagus, and originate from the nerve cells situated anterior end posterior to the nerve ring. The submedial nerves exclusively innervate organs of the anterior body end. The lateral nerves branch into peripheral offshoots which form a compact system that innervates the amphids and various setae. Nerves running from the posterior body end and reaching the peripheral surface likewise innervate different setae.



21 Figure 6. Schematic diagram of the nervous system of nematodes of order Enoplida.

Nerves: 1—dorsal; 2—subdorsal; 3—dorsolateral; 4—ventrolateral; 5—subventral; 6—ventral.

Innervation of the organs of the posterior section of the body of free-living nematodes has not been well studied. One can only state that the copulatory organs, bursal musculature of the male, and the vulval region of the female are innervated by offshoots of the ventral and submedial nerves.

Organs of sense. Tangoreceptors. In all nematodes these organs are concentrated mainly in the cephalic region; however, in males they also

occur in the caudal section of the body. Cephalic setae, anal setae, and papillae supplied with nerves serve as tangoreceptors. As these have been discussed above, further comment is not necessary here.

Chemoreceptors: These are the lateral organs of amphids and constitute organs of chemical sensitivity. These are three main types of amphids in nematodes—cyathiform, circular, and spiral. Their structure is a most stable feature and usually characteristic of a large taxonomic group. Cyathiform amphids, causing a depression in the cuticle, are characteristic of the entire order Enoplida. A papilla with a nerve end intrudes into the closed posterior part. The anterior part of the amphid opens exteriorly. Amphid size in lower enoplids varies greatly; the width may range from 1/2 to 1/8 of the corresponding body diameter. Form is also variable, from circular to cyathiform to oval or elongated in both transverse and longitudinal directions (Figure 7). Location likewise is not constant; amphids may be situated immediately behind the cephalic capsule as in the case of leptosomatids (*Leptosomatium*, *Leptosomatides*), in the lateral interlobular grooves (*Synonchus*, *Thoracostoma*, *Deontostoma*, *Pseudocella*), or at some distance from the cephalic capsule; usually, however, this distance does not exceed two to three times the diameter of the head. In anticomids the amphids are arranged somewhat below the level of the cephalic setae and their size varies greatly in different species. It is interesting to note that in genus *Rhabdodemia* amphids are generally absent.

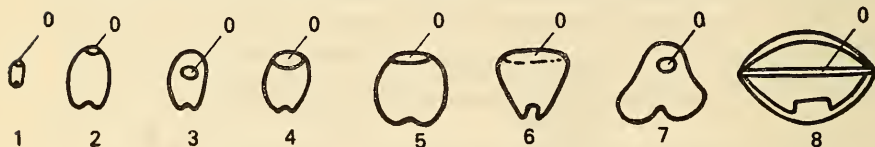


Figure 7. Shapes of amphids in nematodes of families Leptosomatidae and Anticomidae.

1—*Anticoma*; 2 to 4—*Pseudocella*; 5 to 7—*Leptosomatium*; 8—*Platicomopsis* (o—opening of amphid).

The functions of amphids have long been debated and range from glandular (de Man, 1886) to auricular (Marion, 1870; Bütschli, 1873). Zur Strassen (1904) put forward the proposition that amphids are organs of chemical sensitivity, i.e., chemoreceptors. This point of view is the most widely held today. However, the problem of the function of amphids as chemoreceptors cannot be considered conclusively resolved as experimental data supporting this supposition are still inadequate.

Inglis (1964), proceeding from the fact that the function of amphids as chemoreceptors has not been confirmed experimentally, is inclined to attribute to them an altogether different function, namely, receptors of

mechanical tension of the cuticle. Inglis' idea is extremely interesting, but unfortunately lacks experimental supportive confirmation.

Photoreceptors. Unlike the amphids, the shape of which is characteristic of a definite taxonomic group, photosensitive structures appear convergently in different groups of free-living nematodes. The presence or absence of these structures and their shape are features used in species diagnosis.

Photosensitive organs are always situated in the preneural region of nematodes. Of all the groups of nematodes studied in the present work, they were found only in leptosomatids (*Leptosomatium*, *Leptosomatides*, *Thoracostoma*, *Deontostoma*, and *Pseudocella*); their structure is quite complex (Figure 8) except in the last two genera. Each organ consists of dark red or brown pigment in a compact structure, which is circular or cyathiform with the lens situated anteriorly. Such "eyes" occur along the sides of the esophagus and their lenses project somewhat sideways. They are usually situated at the same level but in some cases (for example in *Leptosomatides euxina*) one may be situated much above the other. Some-

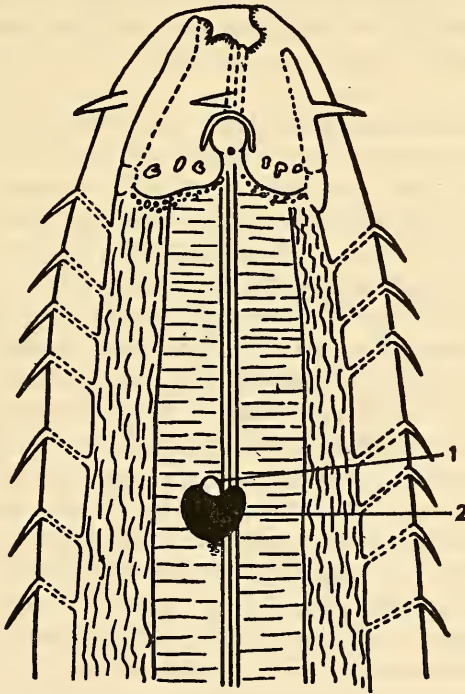


Figure 8. Structure of photosensitive organ in nematodes of genus *Thoracostoma*.

1—light-refractive lens; 2—pigment bowl.

times the pigment projects beyond the limits of the pigment bowl and forms a small dendroid overflow behind it.

In *Thoracostoma* and *Pseudocella* a tendency toward reduction of the photosensitive organs is observed. In *Thoracostoma* most species have "eyes" exhibiting the structure described above, but a number, for example *T. schizoepestilium*, *T. philippinensis*, *T. anocellatum*, *T. parasetosum*, and *T. brunni* are completely devoid of photosensitive organs. In *T. setosum* only the pigment bowl is present; the lens is absent.

In species of *Pseudocella* "eyes" with lenses are seldom found. Usually the photosensitive pigment occurs not in a compact mass, but in small spots of irregular shape and light red in color. Sometimes the pigment is dispersed in longitudinal strips with a series of lateral offshoots. Some representatives of this genus are absolutely devoid of pigment; in my material no pigment was discernible in *Pseudocella bursata*, *P. mamillifera*, *P. acuta*, and *P. angusticeps*.

24 It seems to me that in *Thoracostoma* and *Pseudocella* a process of gradual reduction of photosensitive structures is taking place but no loss in their occurrence. The reason for this is that these two genera are more highly organized in all the remaining details of their structure than in species of the genus *Deontostoma* and still more than in those of *Leptosomatium*.

Digestive system. The digestive system of nematodes commences with an oral opening which in some leads into an oral cavity. The latter is extremely varied in structure in lower enoplids as noted above. Here I shall only mention that there are species within the limits of the group of lower enoplids which are absolutely devoid of an oral cavity as well as species with a well-developed and armed oral cavity.

Wieser (1953b) divided all free-living marine nematodes into four groups on the basis of the structure of their oral cavity. Among lower enoplids, three of these four groups are found. Nematodes lacking a distinct buccal capsule belong to the first group. Capturing food in such a case is effected by sucking with the help of the esophageal muscles. *Anticoma*, *Paranticoma*, *Anticomopsis*, *Crenopharynx*, *Antopus*, *Barbonema*, *Leptosomatium*, *Leptosomatides*, *Leptosomella*, *Platycoma*, *Platicomopsis*, and *Triodontolaimus* should be included in this group.

Nematodes with a small oral cavity armed with small teeth or plates to scrape aquatic plants, algae, etc., or bacterial film on various underwater objects, belong to the second group. Representatives of Synonchynae and Thoracostominae (*Leptosomatidae*) belong to this group.

Finally, nematodes with an extremely large oral cavity with a thick sclerotized lining and powerful onchia which can pierce the membrane of aquatic plants and suck the contents of their cells belong to the third group. Furthermore, there are possibly predators which swallow small

organisms. Lower enoplids differ sharply with respect to type of food ingested and mode of feeding, which is reflected in the variation of structure of the oral apparatus.

The esophagus, which is comparatively uniform in shape in lower enoplids, follows immediately after the oral capsule. Usually the esophagus is straight and widens gradually toward the base. The esophagus of lower enoplids never exhibits true or pseudo bulbs. In transverse section the esophagus exhibits a triradial lumen lined internally with a cuticle with angles intruding inward and musculature arranged in the shape of three longitudinal bands, forming three sections in the esophagus (one dorsal and two subventral). Esophageal glands are situated (see below) in the middle of each section.

The musculature of the esophagus has been studied fairly well in parasitic nematodes (Looss, 1896; Schneider, 1902; Goldschmidt, 1904; Martini, 1916) and in free-living nematodes (Rauther, 1907; Filip'ev, 1921). It has been established that the esophagus in both groups is similar in structure. The tissue is composed of muscular, epithelial, nerve, and glandular cells. Muscular and epithelial cells are so closely arranged that their cellular boundaries are distinguished with great difficulty. The former contain radial muscular fibrillae, one end of which is attached to the internal angles of the esophageal tube and the other to the superficial layer of the esophagus. In lower enoplids muscular bands extend continuously throughout the esophagus. In those nematodes in which the esophagus has bulbs the epithelial tissue may be interrupted by the muscular tissue, forming a membrane in the region of the latter and thus imparting to the esophagus great firmness. Several epidermal cells, also containing fibrillae, extend from the anterior angles of the esophageal tube to its surface and are situated along the angles. As distinguished from those described above, these are not contractile but elastic fibers. Derivatives of the epidermal cells represent the external firm membrane
25 of the esophagus. Thus the muscular cells are surrounded, so to speak, by epithelial ones. It should be noted that the number of muscular and epithelial cells of the esophagus is greater in free-living nematodes than in parasitic ones. Thus in *Ascaris* there are 24 and in *Synonchus* 42. Moreover, the number in each species is constant (Filip'ev, 1921).

Rauther (1907) suggested that besides its basic function, the esophagus also fulfills an excretory one. In his experiments different vital stains accumulated and were then excreted in the angular bands of the esophagus. In his opinion the pigment strips sometimes observed in the esophagus are actually an accumulation of excrement in the esophageal lumen which is subsequently excreted. The lumen of the esophagus is lined with a cuticle that differs sharply from the external cuticle covering the body. Sometimes bulges are observed in the esophageal cuticle of

lower enoplids which resemble a stripe in longitudinal sections (Filip'ev, 1921). At the posterior end of the esophagus, where it joins the intestine, a special structure can be isolated which is known as the cardia. In the anterior part of the intestine the cardia projects in the shape of a triangle. Functionally, it serves as a valve, obstructing the movement of food from the intestine back to the esophagus. According to Filip'ev (1921) the cardia consists of clearly distinguishable cells devoid of musculature. Here Filip'ev differs in opinion from Türk (1903) who discovered a large number of nuclei and annulated muscular fibrillae in the cardia of *Synonchus*. Steiner (1915) noted the presence of sclerotized reinforcement in the cardia of *Leptosomatum sabangense*.

Timm (1953) in studying the cardia in sections from *Leptosomatum acephalatum* noted that it consists of esophageal tissue surrounded by a thin annular sphincter which, in turn, is surrounded by the nerve cells of the intestine.

The intestine of lower enoplids, like that of all free-living nematodes, is represented by a straight simple tube devoid of all outgrowths and blind pouches such as seen in parasitic nematodes. The intestinal wall consists of a single layer of cells. The number of intestinal cells varies considerably in different groups of nematodes. Thus in nematodes of Leptosomatidae and Oncholaimidae the number of cells is significantly more than in Monhysteridae and Chromadoridae (Bütschli, 1873; de Man, 1888; Cobb, 1893a).

According to Timm (1953) the number of intestinal cells in free-living nematodes varies from 123 to 5,000 and two types are distinguishable: unspecialized isocytes and specialized heterocytes. Isocytes are represented by rectangular cells of equal height. Heterocytes are oval cells, the size of which exceeds that of the isocytes by two to three times. As distinguished from the isocytes, these cells have a rough basophilic reticular net. Timm did not observe them in all leptosomatids. He found them in *Leptosomatum acephalatum* but not in *Deontostoma magnificum* and *Thoracostoma coronatum*. Türk (1903) observed them in *Synonchus strasseni*.

In addition to the foregoing cells, a rod-shaped layer ("rod border" according to Hyman, 1951; "bacillary layer" according to Timm, 1953) also occurs in the intestinal wall, in which rod-shaped basophilic elements can be distinguished. These structures have been described in detail by Timm (1953) for *Leptosomatum acephalatum*; Rauther (1907) did not find them in *Cylicolaimus*.

Inside the intestinal tube lies a smooth radially striated cuticle, and outside it a basal membrane formed partially from the intestinal cells and partially from the connective tissue (Martini, 1916).

In the middle part of the body in sexually mature individuals the intestine may be compressed by the gonads.

26 Timm (1953) noted that the intestine of males and females can be distinguished by cellular composition. For example, in *Leptosomatum acephalatum* he found that the intestine of males consisted not only of a smaller number of cells, but also fewer oval cells, and that the cells were flatter, almost cubical, and very much larger than in females.

The rectum (or hind gut) in females passes through the intestino-rectal valve surrounded by epithelial tissue. The entire rectum is lined with the outer cuticle, turned inside, which may extend even farther, i.e., into the intestine. Timm (1953) observed that in sections taken from the part somewhat anterior to the rectum in *Leptosomatum acephalatum* there were numerous digitiform structures directed into the lumen of the intestine. Probably numerous folds of the cuticle (digitiform in shape) occur here. The rectum is encircled by a thin muscular sphincter. In males the rectum and sperm duct fuse not far from the anus.

Glands participating in the process of digestion are closely associated with the digestive tract. According to Türk (1903) and Rauther (1907) the anterior part of these glands is represented by a simple tube, while the posterior part is dendroid. The dorsal gland differs somewhat from the subventral glands. Thus in *Cylicolaimus* and *Synonchus* the unbranched part of the dorsal gland runs farther backward than that in the subventral ones. Esophageal glands open either directly into the oral cavity or, in the case of its absence, into the esophagus somewhat posterior to the cephalic capsule. The position of the external openings of these glands varies in different species of nematodes. In nematodes of family Leptosomatidae the openings of the subventral glands are shifted more toward the anterior end and the dorsal gland is somewhat posterior in position. The duct of the subdorsal gland in *Leptosomatum acephalatum* opens immediately behind the oral cavity. In *Thoracostoma coronatum* this duct opens on a small denticle situated in the oral cavity. When two subdorsal glands are present, their ducts open at the same level but not on the same denticle. In *Deontostoma magnificum* the outlet of the dorsal gland opens on the dorsal denticle and the ducts of the subventral glands open somewhat anteriorly (Timm, 1953). Rauther (1907) discovered accessory "lateral" ducts of the glands at the level of the eyes in a *Thoracostoma* sp. Timm found such ducts in *Deontostoma magnificum*.

Reproductive system. The female reproductive system of nematodes is represented by a continuous tube, different sections of which serve respectively as the ovary, oviduct, uterus, and vagina (Figure 9). In lower enoplids these tubes comprise a pair, one anterior and the other posterior. The anterior and posterior position of the reproductive tubes is a secondary phenomenon. Originally these tubes were situated bilaterally. That they are situated on the right and left side of the intestines bespeaks this fact. Subsequently one tube reflexed and the other remained in its

original position. This occurred due to the intense stretching of the body in length and narrowing of the diameter (Filip'ev, 1921; Steiner, 1922b; Hyman, 1951; Paramonov, 1962).

In free-living nematodes the reproductive tubes are always positioned as described above. A bilateral arrangement of the reproductive tubes can be observed only in some parasitic (*Ascaris*) and phytonematodes (representatives of family Heteroderidae).

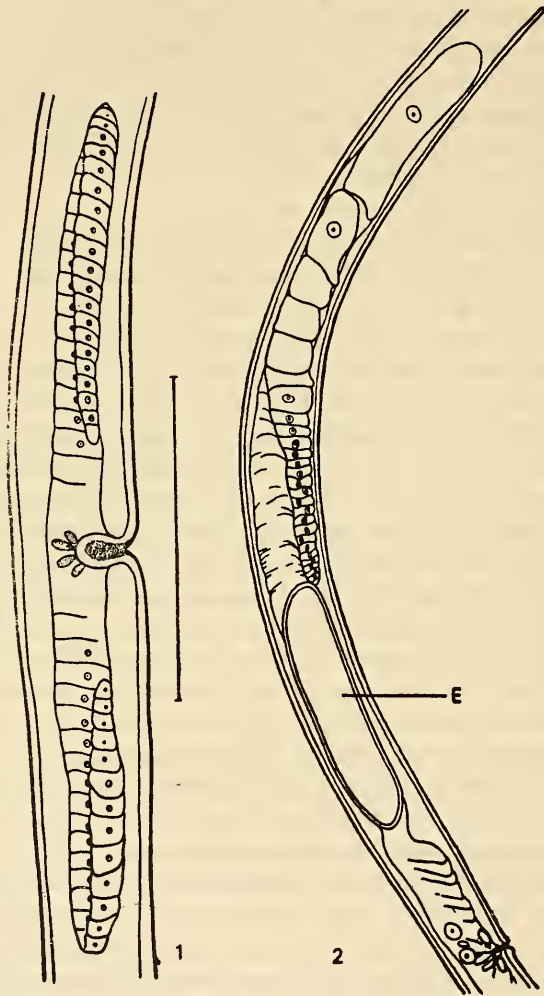


Figure 9. Structure of female reproductive system of nematodes.
 1—reproductive tube of *Thoracostoma*; 2—anterior reproductive tube of
Pseudocella (one large egg E visible in figure).

The terminal part of the ovaries of lower enoplids is always reflexed. The reflexed part is significant in length, usually not less than half the length of the straight part, but sometimes equal to it in length, i.e., the ends of the gonads reach the vagina (*Anticoma*).

The ends of the gonads from which the oogonia separate are syncytial in nature. The oogonia occur first in a cluster and then form a single row.

27 Externally the ovary is covered by a thin epithelium consisting of flat and stretched cells (Cobb, 1893; de Man, 1886).

The ovaries are somewhat displaced toward the dorsal side and attached to the muscular layer near the distal end of the uterus.

The cells of the ovarian epithelium may differ significantly in shape and size in different species of lower enoplids. Thus in *Thoracostoma coronatum* a large flat epithelial cell extends the length of the ovary. In *Leptosomatum acephalatum* extremely small cells are situated along the short germinal zone, but cells of pyramid shape are situated in the zone of growth; these are significantly larger than the cells of the germinal zone. Before the bend of the ovary the cells become very large and oval. As the genital tracts of nematodes are represented by a single continuous tube, demarcating the ovary and the oviduct becomes somewhat difficult. de Man (1886) distinguished these sections on the basis of their histology. In such a case the ovary is very long and the oviduct represented by a short area continuous with the uterus. Jägerskiöld (1901) and Filip'ev (1921) distinguished these two sections on the basis of their morphology, considering the ovary the entire part between the uterus and the depression in the contracted part of the blind sac. The oviduct is represented by that part of the reproductive tube which is separated due to the fact that the former does not join the end of the ovary itself, but a little posteriorly. Mature eggs are stored in this sac.

In sexually mature females the oviduct exhibits disintegrating spongy tissue throughout its length all along the surface of the ventral body wall between the intestine and the ovary. In such a condition the passage is almost invisible. In younger specimens the wall of the ovary has not disintegrated and the passage is distinctly visible. Timm (1953) observed such a phenomenon in *Leptosomatum acephalatum* but gave no explanation. In this same species he notes the absence of any trace of a spermatheca isolated from the reproductive canal. Spermatozoa were present in the uterus in the region of egg formation in the ovary. In many species spermatozoa often concentrate in the anterior part of the ovary, immediately behind the zone of egg formation. This region probably plays the role of the spermatheca.

28 Between the oviduct and uterus lies the zone of egg formation in which extremely large cells occur, filling the lumen of the reproductive canal. These cells surround the eggs within the limits of the egg-forming zone

or closely adhere to the eggs entering the uterus. The eggs either pass across the center of these cells or along one side. While entering this zone the eggs are devoid of a shell, but on exiting covered with a firm shell. Timm (1953) considers the function of these large active cells to be shell formation. They probably secrete an enzyme which activates the eggs to form a shell.

The uterus is represented by a relatively wider tube in which the walls consist of flat epithelial cells. Muscular fibers run only along its proximal end. Near the vagina the uterus has a reduced sphincter of thick muscular cells which is located on the ventral side. Unlike a large number of free-living nematodes (Chromadoridae and some Oncholaimidae) which have an unpaired uterus merging near the vagina, in lower enoplids the vagina is always paired and the passages of the two uteri separate.

The number and size of eggs vary considerably even within the limits of a single species. In the uterus of females of different species I have counted from one to nine eggs. The smallest eggs, 50 to 60 μm long, I found in *Anticoma* and the largest, 1,200 μm long, in *Pseudocella bursata*. Egg size, as mentioned above, varies within the limits of one and the same species. Thus in *Leptosomatium tetrophtalmum* eggs ranged from 154 to 206 μm in length, in *L. kerguelensis*—103 to 236 μm , in *Leptosomatides crassus*—154 to 360 μm , in *L. acutipapillosum*—154 to 257 μm , and in *Pseudocella trichodes*—206 to 412 μm .

The female genital pore (vulva) is situated on the ventral side, most often midbody. Sometimes it is shifted posteriorly and very rarely anteriorly. In females of some nematodes sclerotized granules encircle the vulval slit. Filip'ev (1922a) found this peculiarity in *Leptosomatides steineri*. I found it in other species—*Platycomopsis mesjatzevi* and *Deontostoma arcticum*. These granules are frequently found in species of genera *Leptosomatium* and *Leptosomatides*. In my material granules surrounding the vulval aperture occurred in four species of genus *Leptosomatium* and two species of *Leptosomatides*.

It is interesting to note that in females of *Paracycolaimus brevisetosus* I detected two vulval slits (Platonova, 1970). This phenomenon is an abnormality. A similar case has been described by Paramonov (1926).

The male genital tubes of lower enoplids are represented by paired testes which pass into two seminal vesicles and merge into a single vas deferens. The latter transforms into an unpaired muscular ejaculatory duct opening into the cloaca. The copulatory organ also opens into the cloaca.

As in the case of female gonads, in males also one testis is anterior and the other posterior (Figure 10). The beginning of the testis is represented by a syncytium, giving rise to spermatogonia. Mature spermatozoa collect in the posterior end of the testis or in the seminal vesicle.

The testis is covered externally by a thin epithelium composed of longitudinally stretched fusiform cells (de Man, 1886) or by an extremely thin and flat epithelium (Jägerskiöld, 1901).

The anterior part of the vas deferens consists of numerous low cells with indistinct cellular outlines. The number of cells decreases toward the posterior part of the vas deferens and their outlines become more distinct. The ejaculatory duct, situated next, consists of two rows of cells compressed longitudinally. There is almost no musculature in the anterior part of the duct but well-developed musculature present in the posterior part (Jägerskiöld, 1901; Türk 1903; Filip'ev, 1916).

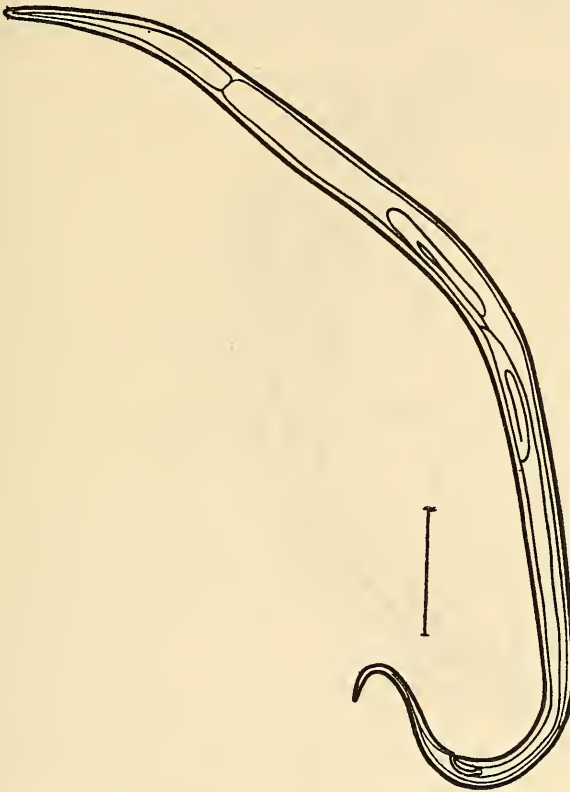


Figure 10. Diagrammatic sketch of reproductive tube in male.

The copulatory system of males is situated in a special sac of the cloaca and consists of a pair of spicules and a gubernaculum (Figure 11). The spicules are hollow sclerotized organs which curve variously. A plasmatic substance, part of those epidermal cells which form the spicules, develops inside them (Filip'ev, 1921). One can demarcate the capitulum of each

spicule at the proximal end from the constriction (cervix) following it. The body of the spicule has a lumen in the middle and a thin membrane in the ventral curvature, i.e., the velum. Sclerotized structures directing the movements of the spicules have been labeled "ruleks". This term was introduced in Soviet literature by Filip'ev to replace "gubernaculum", proposed by Looss. The gubernaculum may be paired or single. Mostly it is situated on the dorsal side of the spicules but sometimes ventrally (*Platycomopsis mesjatzevi*). Sometimes the gubernaculum is altogether absent (*Platycoma cephalata*, *Anticoma longisetosa*, *A. graciliceps*, *A. filipjevi*, *A. uschakovi*, *A. behringiana*, *A. brevisetosa*, and *Paranticoma antarctica*).

Filip'ev (1921, 1927) established some types of spicule structures

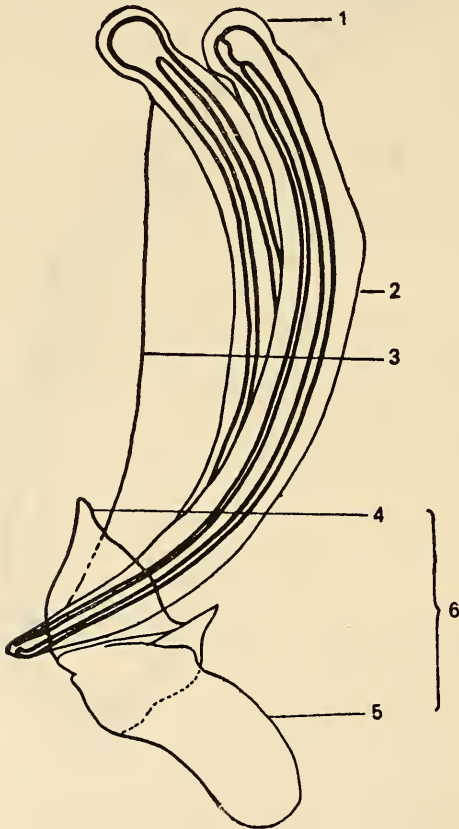


Figure 11. Diagrammatic structure of spicular apparatus of nematodes of genus *Pseudocella*.

1—capitulum of spicule; 2—body of spicule; 3—velum; 4—ventral process of gubernaculum; 5—dorsal process of gubernaculum; 6—gubernaculum.

(straight, arcuate, broad, lamelliform, flat, elongated, and complex), under which he formed subgroups. Of the seven main types, three are characteristic of lower enoplids: 1) arcuate (curved spicules which almost do not broaden in the middle; well-developed and complex gubernaculum always present); 2) broad (distinguished from the preceding type by flatter shape and presence of velum); and 3) lamelliform (with velum on ventral side). Broad spicules are characteristic of *Leptosomatum*, *Platycomopsis*, *Rhabdodemia*, and the majority of the species of Synonchinae, Barbonematinae, and Platycominae. Arcuate spicules are characteristic of *Cylicolaimus*, *Leptosomatides*, *Deontostoma*, *Thoracostoma*, and *Pseudocella*. Filip'ev placed representatives of Anticomidae under the third type, namely, lamelliform spicules.

It seems to me that this scheme of classification is much too broad. *Anticoma*, placed by Filip'ev under the lamelliform type, includes a significant diversity of spicule shapes. In my material for this genus I found arcuate, broad, and lamelliform spicules, with the gubernaculum either situated ventrally, dorsally, or absent (Figure 12).

In *Leptosomatum* the spicular shape is fairly uniform. These organs are slightly curved, sometimes with a narrow capitulum, and widen in the anterior third; the gubernaculum is simple in structure (without processes) and dorsally situated. All the species of this genus may be grouped under spicules of the broad type. *Platycomopsis* is characterized by weakly curved spicules, broad in the distal half, and a ventrally situated gubernaculum. Species of *Rhabdodemia* have spicules similar to those of *Leptosomatum*; *Barbonema* and *Platycoma* have broad, tubular spicules; *Cylicolaimus*, *Leptosomatides*, *Deontostoma*, *Thoracostoma*, and *Pseudocella* have arcuate spicules. However, some species of *Deontostoma*, *Thoracostoma*, and *Pseudocella* have lamelliform spicules, namely, *Deontostoma antarcticum*, *Thoracostoma setosum*, and *Pseudocella trichodes*. This group is characterized by a complex gubernaculum which always has a dorsal and rarely a ventral process. Often the gubernaculum is paired and forms a "funnel" in which the distal part of the spicules is located. In *Crenopharynx* the spicules are rather unique; they are very slender and long with a small round capitulum, and a simple slitlike gubernaculum. *Crenopharynx* is close to species of the family Phanodermatidae with respect to spicular structure.

Thus the structure of the copulatory apparatus within the limits of lower enoplids is highly diversified.

The sexual armature of the male, in addition to spicules, includes copulatory papillae, setae, and an accessory organ. Papillae and setae are situated in two submedial rows in the anal region. Slightly anterior to the anus an unpaired glandular structure occurs, constituting an accessory organ. It resembles a large papilla externally but inside one finds rather

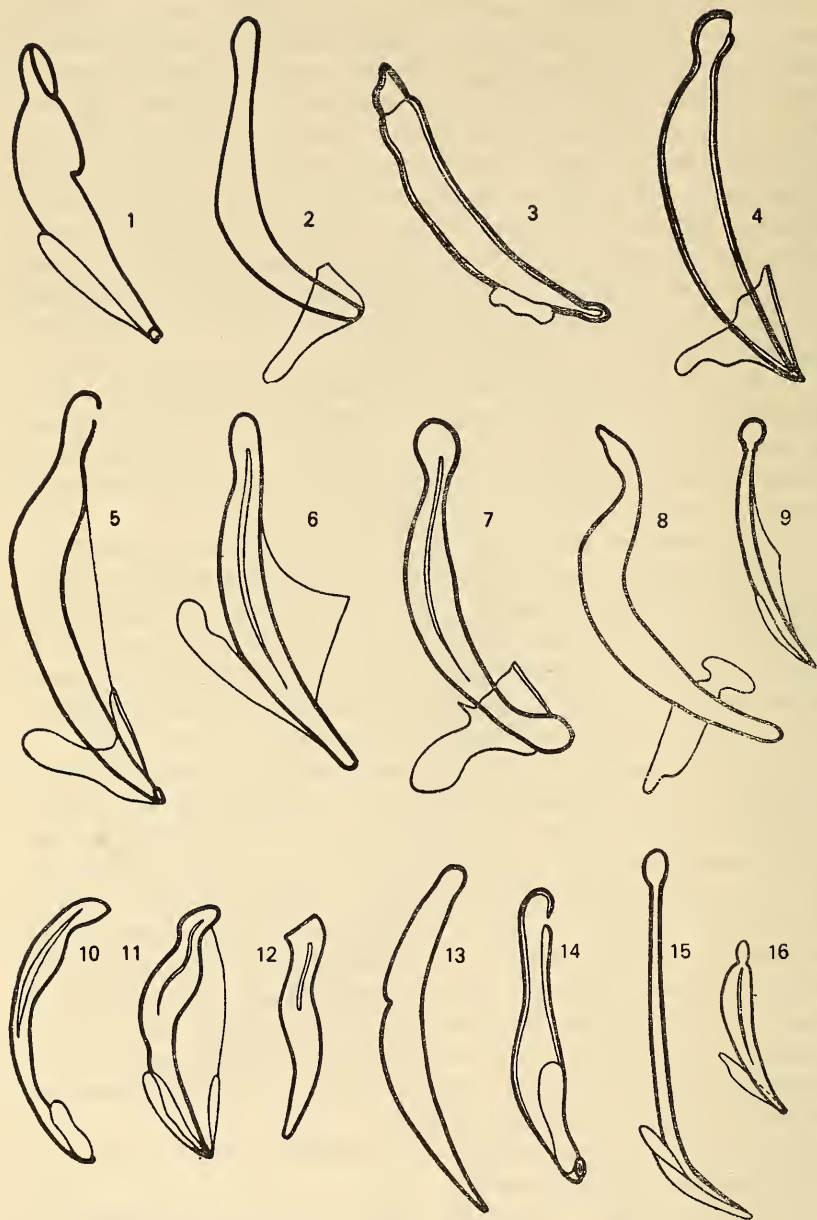


Figure 12. Types of spicular apparatus in lower Enoplida.

1 to 2—*Leptosomatium*; 3 to 4—*Synonchus*; 5—*Deontostoma*; 6—*Thoracosoma*; 7—*Pseudocella*; 8—*Cylicolaimus*; 9 to 11—*Anticoma*; 12—*Barbonema*; 13—*Platycoma*; 14—*Platycomopsis*; 15—*Crenopharynx*; 16—*Rhabdodemania*.

well-developed sclerotized tubes, annulations, or hemispheres. Sometimes sclerotized processes, directed downward or upward, are also present. The structure of the accessory organ and number and disposition of papillae and setae may serve as reliable species indexes. All these auxiliary copulatory armatures may be partially or fully absent. Representatives of *Rhabdodemanina* and *Leptosomatium* lack setae, papillae, and accessory organs. In species of *Anticomina* an accessory organ of the tubular type is always present but setae and papillae absent. In species of *Crenopharynx*, on the contrary, accessory organs are absent but preanal setae invariably present. In *Deontostoma*, *Thoracostoma*, and *Pseudocella* an accessory organ, papillae, and setae are usually present. Very rarely papillae or setae may be atrophied but an accessory organ invariably developed. When all three are present, the structure of the accessory organ as well as the disposition of the papillae and setae may vary significantly. In *Deontostoma arcticum* papillae with setae commence near the anus and extend a fairly significant distance. The accessory organ is situated in the middle of this row of papillae. In *Deontostoma papillatum* the papillae bear no setae and commence not from the anus but from the accessory organ. Males of *Deontostoma antarcticum* have only an accessory organ; papillae and setae are totally atrophied in them. In *Leptosomatium inocolatum* setae commence immediately behind the anus and papillae devoid of setae lie somewhat above the level of the accessory organ.

PHYLOGENETIC RELATIONS AND TAXONOMIC BASIS

Position of Order Enoplida in the Classification of Nematodes

The origin of nematodes and the position of their different groups in the taxonomic system of the class have yet to be conclusively resolved. Winslow (1960, p. 341) remarks that the position of nematodes in the classification of the animal kingdom and their phylogenetic relations have, as it were, slipped from the consciousness of researchers and hence their position is one of incertitude. In my opinion the latter contention is overly cautious. At present, most scientists engaged in the study of this group consider free-living marine nematodes extremely primitive and original forms (Filip'ev, 1921, 1934a; Schuurmans-Stekhoven, 1931; Hyman, 1951; Paramonov, 1962; and others). But Winslow is right that the question of the evolution of the class as a whole has yet to be fully resolved.

Researchers of this group have directed their attention mainly to parasitic nematodes and hence we now have considerable morphological, embryological, histological, and other data on this group. However, parasitic nematodes, being a specialized and highly organized group, can-

not alone illuminate the evolution of this class. The answer to this question must be sought among the most primitive and less specialized group of nematodes. For this reason, before searching for affinities between nematodes and other groups of animals, it is essential to note relations between orders and families within the class *per se*.

I do not claim to have conclusively established these affiliations as the amount of anatomical and histological information available in literature is, to my regret, inadequate. Hence only preliminary observations regarding this problem are given.

Filip'ev (1921) rightly suggested that highly simple forms, which exhibit no features of reduction or (atrophy), should be considered the most primitive nematodes, since the evolution of this group followed a line of specialization and simplification of some organic systems. Such nematodes belong to the order Enoplida.

33 Class Nematoda is divided into two subclasses—Secernentea comprising highly specialized and parasitic forms, and subclass Adenophorea in which mainly free-living forms, together with enoplids, a smaller number of parasitic nematodes, and phytonematodes are included. The families studied by me under the order Enoplida include exclusively marine forms. Of these families, Leptosomatidae comprises the least reduced and specialized group of nematodes with respect to structure and, for this reason, may probably be considered the most primitive family among free-living nematodes. The primitivity of this group is expressed in a number of anatomical and morphological peculiarities of structure.

The nervous system of nematodes of various groups is structured as follows. Around the nerve ring—the central part of the nervous system—nerve cells in species of Enoplida are situated in the most primitive manner; ganglia are totally absent and the nerve cells lie in a simple layer around the esophagus. In free-living nematodes of Monhysterida (*Siphonolaimus*, *Solenolaimus*) anatomically well defined, densely fused ganglia are present. In parasitic nematodes (*Ascaris*, *Mermis*, *Ancylostoma*) the situation of the cells of the central nervous system reflects an intermediate position between these two groups, i.e., less well-defined ganglia represented by only a group of cells. This is probably explained by a secondary reduction of the nervous system caused by a parasitic mode of life.

The peripheral nervous system has an extremely complex structure in species of Enoplida, while in free-living smaller forms, such as nematodes of Monhysterida, this nervous system is quite simple. A similar phenomenon is observed in parasitic nematodes (Filip'ev, 1921).

Probably the original form of the nematode nervous system should be considered such a structure as is found in enoplids, where the degree of integration of the central nervous system is very minimal and the peripheral nervous system exhibits no features of reduction. In more highly

organized free-living nematodes a distinct process of ganglionization, which leads to greater integration of the central nervous system, is observed. But reduction of the peripheral nervous system, brought about by the process of general reduction of cellular composition, accompanies this. Reduction of the central and peripheral nervous systems of parasitic nematodes is effected by a parasitic mode of life. In addition to a weakly integrated central nervous system and a highly developed peripheral system, nematodes of the order Enoplida have the most primitively constructed organs of chemical sensitivity—the amphids—which are cyathiform in shape. Other orders of free-living nematodes have more complexly constructed amphids—round, horseshoe-shaped, or spiral.

The body cavity in various groups of nematodes is developed to different degrees. In leptosomatids, where it has not been seen by a number of authors (Jägerskiöld, 1901; Türk, 1903; de Man, 1904; Timm, 1953), the body cavity is least developed. It is poorly developed in other families of the order Enoplida (Filip'ev, 1921). In other orders of free-living nematodes it is present as a small slit and achieves fuller development in saprozoic forms and intestinal parasites (Filip'ev, 1937). Thus the near absence of a body cavity in leptosomatids also points to their lower stage of organization.

Within the limits of the class the process of oligomerization of a number of organs has taken place (Dogel', 1954). This process can be observed by comparing the setaceous armature of free-living and parasitic nematodes.

34 Setae are extremely numerous and best developed in free-living nematodes, especially in marine forms (Filip'ev, 1918, 1921; Dogel', 1954). In parasitic forms they are considerably reduced. The original number of cephalic sensory organs of nematodes should be taken as six labial papillae and a crown of ten cephalic setae. In parasitic forms (Ascaridata, Spirurata, and others) the number of papillae and setae decreases either as a result of fusion or as a result of reduction. In free-living nematodes, in addition to cephalic setae, other setae are scattered all over the body or at least over the anterior end. In nematodes of the order Enoplida the crown of cephalic setae is usually situated in a single circle; in some species these setae are equal in length, while in others the four sublateral setae may be longer than the others. Only in Oxystomatidae is the crown of cephalic setae distinguished by two types—anterior, comprising six shorter setae, and posterior, comprising four relatively longer setae.

Beyond the limits of the order Enoplida the crown of cephalic setae has undergone more significant change. In most nematodes of the orders Chromadorida and Monhysterida the anterior crown bears structures resembling papillae and the posterior crown comprises four setae. In some nematodes of the order Desmoscolecida the four setae of the second

circle resemble cornuate processes. In some Chromadorida and Desmoscolecida a number of setae situated on the body merge with each other, forming powerful and strictly localized setae employed for locomotion. Preanal setae in some Desmoscolecida fuse and acquire the significance of copulatory setae. Thus, on the basis of the foregoing examples, one can distinctly trace the change of function and oligomerization of setae which originally had a sensory function.

In most species of Enoplida, especially in lower representatives, the cephalic crown of setae has a much simpler, nonproliferative structure, and the setae on the body are numerous and irregularly scattered. All these setae have only a sensory function. Such a type of arrangement of setae in nematodes is apparently more primitive.

Dermal glands are also subject to oligomerization. These glands are numerous within Leptosomatidae, while only a lone cervical gland is seen in Anticomidae.

Oligomerization can also be seen in the reproductive system. Paired reflexed gonads, paired genital ducts, and paired uteri fusing only near the genital pore should be considered the original form in nematodes. A reduction in the reproductive system takes place within the limits of the class. The commencement of this process is already evident in Enoplida, in some species of *Oncholaimus* (Filip'ev, 1921) where one female genital tube is significantly shorter than the other. In some cases even complete atrophy of one of the female genital tubes is seen. More often the anterior genital tube (Oncholaimidae) is reduced and less often the posterior genital tube (Oxystominidae).

No sign of reduction or fusion of organs is observed in the structure of the reproductive system of female leptosomatids and anticomids. Since no gonadal reduction or fusion of the reproductive tubes has occurred in them, the gonads are always reflexed. More often a single unpaired female genital tube is found in parasitic nematodes, but the phenomenon of polymerization may also be observed exceptionally. Thus there are four genital tubes in *Polydelphis*, six in *Hexametra*, and even nine to eleven in *Turgida*. This is considered quite natural for parasitic forms in connection with the need to produce a large number of eggs.

Paired gonads are extremely rare among males compared to females. Thus in Chromadorida males with a single testis are more common, in 35 Monhysterida the testis may be single or paired, in Enoplida the testes are generally paired, but in lower groups of this order they are always paired. The genital ducts (vas deferens and ejaculatory duct) are always unpaired in males. Sometimes oligomerization is evident in even the male copulatory apparatus; of the two spicules, one may be atrophied, which is mostly observed in parasitic nematodes.

Thus the female and male reproductive systems in leptosomatids are

least reduced and hence this group is considered much more primitive than the remaining groups of nematodes.

Phagocytic cells in nematodes of Enoplida are usually very numerous and scattered irregularly over the body. Their number in Leptosomatidae and Oncholaimidae ranges from 150 to 220 or more, and in Enchelidiidae never more than 70. Their structure and situation in enchelidiids are similar to those in oncholaimids. A more regular disposition of these cells is seen in Oxystominidae, where they concentrate near the lateral fields (Bütschli, 1874; de Man, 1907). In most leptosomatids the phagocytic cells are numerous and scattered irregularly over the body, but in *Leptosomatides euxina* they are situated mainly along the ventral line. In species of Chromadorida these cells are arranged in a few longitudinal rows (up to eight in number). In Araeolaimida the number of phagocytic cells is considerably reduced; in *Axonolaimus setosus* there are only two phagocytic cell (Filip'ev, 1918, 1921).

Extremely numerous and irregularly scattered phagocytic cells are thus found in enoplids.

It is interesting to review the changes in cellular composition of individual organs and tissues of nematodes. In nematodes of the order Enoplida eight cords formed by the hypoderm may be present; these cords invariably project. Submedian cords vary in number and may be absent in some species (Filip'ev, 1921). In such cases the total number of cords may decrease to four. In more highly organized free-living and parasitic nematodes the number of these cords is always constant and equal to four. Fluctuations in cellular composition of the lateral cords have been observed in Leptosomatidae. In *Synonchus strasseni* three to five rows of cells have been recorded (Türk, 1903), in *Thoracostoma setosum* five to six rows in the median cords (de Man, 1904), and in *Leptosomatum acephalatum* four to five rows in the lateral cords (Timm, 1953). Filip'ev (1916) relates the number of cellular rows in the hypodermal cords to the change in number of cells in other organs. The number of intestinal cells in different nematodes varies considerably (Bütschli, 1873; de Man, 1888; Maupas, 1900; Filip'ev, 1918, 1921; Timm, 1953). Their number is maximal in Leptosomatidae and Oncholaimidae of the order Enoplida and minimal in the orders Chromadorida and Monhysterida. In many monhysterids the intestine consists in all of one or two rows of cells. It should be noted that besides the large number of cells constituting the intestine in enoplids, Timm (1953) has observed considerable variation in the number of cells within the limits of species in some leptosomatids.

In the muscular fields situated between the hypodermal cords cellular composition also varies. As is well known, all members of the order Enoplida belong to the polymyarian group, i.e., a large number of cells occur in each muscular field. The majority of free-living nematodes also belong

to this group. Enoplids have a very large number of cells in the muscular fields (up to 12; Timm, 1953). A small number of free-living nematodes—Monhysteridae and Rhabditoides—and parasitic nematodes belong to the meromyarian group. Even a total degeneration of muscles (Filip'ev, 1934) has taken place in phytonematodes of the genera *Heterodera* and *Allantonema*. The same phenomenon has been observed in *Meloidogyne*.

36 During ontogenesis all nematodes pass through the stage of meromyarian musculature when the number of muscle cells is constant (Martini, 1903, 1906). By preserving this stage, adult meromyarian nematodes with a constant number of cells develop (Martini, 1908). With the multiplication of muscle cells the polymyarian type is obtained. It seems to me that Filip'ev is absolutely right in his assertion that it would be erroneous to consider young meromyarian individuals the ancestors of all meromyarian forms. Meromyarian forms should be considered regressive or neotenic but not primitive.

To summarize, of all these orders enoplids are the lowest organized group: cuticle smooth and totally devoid of protective armature of any kind; cephalic setae usually situated in one circle; amphids cyathiform; nervous system simple; and esophagus devoid of diverticula. Such features indicate the primitive structure of enoplids.

Taxonomy and Phylogenetic Relation of Families of Order Enoplida

To understand the problem of the taxonomic position of families of the order Enoplida attention should be given to the differences observed by Inglis in the cephalic structure and oral cavity of Leptosomatidae and Anticomidae on the one hand, and most representatives of the order on the other. In representatives of Leptosomatidae, especially higher ones such as *Thoracostoma*, the buccal cavity is poorly expressed and is not topographically demarcated from the onchial cavity; consequently the onchia are displaced far toward the anterior end and situated almost at the level of the odontia. Supporting structures, if developed, form as processes of the cephalic capsule. Because of the fusion of the buccal and onchial cavities and the anterior position of the onchia, the specialized musculature of the esophagus extends far forward toward the bases of the onchia, where it is attached after crossing the cephalic bladder. Non-specialized musculature in the posterior part of the esophagus remains undifferentiated, which is expressed exteriorly in the nonalveolar appearance of the esophagus. The same is observed in representatives of Anticomidae; the only differences are that the cephalic capsule and oral cavity with all its armature are weakly expressed.

An entirely different picture is seen in representatives of Phanoder-

matidae, Enoplidae, Oncholaimidae, and Enchelidiidae. Here the oral cavity is distinctly differentiated topographically into the anterior buccal cavity and the posterior onchial cavity; buccal rods may be situated in the former which play the role of supporting structures or develop later into mandibles, while in the posteriorly situated onchial cavity these rhabdoid structures develop into onchia. The sclerotized lining of both cavities may fuse imperceptibly, forming onchiobuccal plates, to the posterior (onchial) half of which the specialized musculature is attached, ensuring movement of the entire complex. Due to the posterior position of the onchia, the specialized musculature does not extend to the posterior margin of the buccal cavity and, consequently, does not cross the cephalic bladder. The nonspecialized musculature of the posterior part of the esophagus is differentiated into separate fascicles which externally impart an alveolar appearance to this part of the esophagus.

For a clear-cut picture of these undoubtedly independent lines of evolutionary development, one must imagine the structure of a hypothetical form which could have been their common ancestor. The cephalic capsule of such a form should resemble a cap at the anterior end of the body, i.e., of that part where the musculature of the esophagus joins the body wall. The cephalic and esophageal capsules in this form should not be differentiated and the cephalic bladder should be absent. The triradial internal lumen of the esophagus should open abruptly as a triangular oral opening so that there would be practically no buccal cavity. The lumen in the anterior part of the esophagus should remain constant in diameter throughout its entire length so that, strictly speaking, there would be no onchial cavity. Here, one could project that part of the esophagus would later develop into the onchial cavity. Because of the complete absence of oral armature, there would be no need for specialized musculature. The musculature of the esophagus would remain undifferentiated throughout its length.

Such a hypothetical form is very similar to members of the genus *Leptosomatum*. This gives one grounds for assuming the first evolutionary line (Leptosomatidae, Anticomidae, and Oxystominidae) or their primitive representatives as ancestral in relation to the second line, i.e., Enoplidae, Phanodermatidae, and others. Since it is indisputable that such forms as *Leptosomatum* are the most primitive within the limits of the first evolutionary line, it is clear that Leptosomatidae is the most primitive in this order.

In the order Enoplida, in addition to the families studied by Inglis (1964), there are other families related to these two phylogenetic branches; unfortunately information about them is absolutely inadequate to judge their taxonomic position. Of the two families grouped under the superfamily Tripiloidae and often considered the most primitive in the order

(de Coninck, 1965), Ironidae belongs to the leptosomatid-anticomid branch (Inglis, 1964). Tripilidae cannot be regarded as the most primitive in the order due to the presence of some specialized peculiarities of the esophagus, in particular the development of the esophago-intestinal valve. Lauratonematidae also cannot be regarded as primitive due to the reduction of the posterior gonad in males, and the annulated nature of its cuticle; the latter is uncharacteristic of representatives of the order as a whole.

If the evolution of the head in Leptosomatidae is examined, two parallel, but to some extent independent, processes can be traced. The first process is intensification of development of the cephalic and esophageal capsule, and the second process the formation and development of the oral cavity. The initial stages of the first process can be traced among genera close to *Leptosomatum*. In *Leptosomatides* and *Paraleptosomatides* the cephalic capsule is developed more powerfully than in *Leptosomatum* and there is every reason to assume that a developed cephalic bladder is also present. Moreover, in *Leptosomatides conisetosum* a depression occurs in the posterior margin in which the cephalic setae are situated. In *Leptosomatum reducta* there are outgrowths in the cephalic capsule resembling the supporting structures observed by Inglis in some of the forms studied by him. The commencement of the second process—development of the oral cavity—is also evident to some extent in nematodes of the group *Leptosomatum*. Mawson (1956, 1958b) observed a distinct oral cavity in *Leptosomatides conisetosum*. Both of these processes are developed in genera clustering around *Synonchus* and *Thoracostoma*. Genera of this group have a powerful cephalic capsule or often supporting structures in the anterior part. Sometimes they have a cephalic capsule continuing strongly backward or uniting in this part with the cervical capsule, which has deep clefts along its posterior margin. Representatives of these genera also have a fully developed esophageal capsule and a conspicuous cephalic bladder which is usually divided into six parts by the radii of the esophagus and fascicles of specialized musculature. At the same time they have an oral cavity in which onchia and odontia are present. True, the oral cavity is rather small in size.

In the other group of genera, those close to *Cycolaimus*, the cephalic capsule has not attained the significant development seen in the previous group, and at best resembles the cephalic capsule of some species of *Leptosomatides*. Contrarily, here the oral cavity is quite well developed (at least in males of those genera exhibiting sexual dimorphism). Moreover, widening of the oral cavity has taken place mainly at the expense of the onchial cavity, while the buccal cavity retains a relatively insignificant depth. Concomitantly, some peculiar types of cephalic bladder are seen; the bladder may extend significantly along the body but be reduced to a

narrow ring between the large oral cavity and the body wall. Structures such as odontia sometimes develop in the anterior part of the oral cavity; large onchia are situated somewhat posteriorly. The absence of onchia is probably explained by their wearing out since otherwise the reason for the development of an extensive onchial section in the oral cavity would not be understandable.

Thus three distinctly defined groups of genera, of which one is primary and the other two derived independently from it, are observed in Leptosomatidae.

Sometimes the genus *Triodontolaimus*, with only a single species—*Triodontolaimus acutus* (Villot)—is placed as a subfamily, in addition to the group of genera discussed above, in the family Leptosomatidae. The problem of its phylogenetic connection with the present Leptosomatidae is fairly complex. Judging from the description of de Man (1893, p. 116), the cephalic capsule in *T. acutus* is absent or at best very poorly developed. On the basis of his description, one can only gather that the head is posteriorly limited and demarcated from the rest of the body by a thin suture (p. 116); unfortunately this suture is not depicted in the illustrations. de Man makes no mention of the esophageal capsule, directing his attention to the presence of three symmetrical "teeth" with wide and concave bases. Subsequent information, namely, that musculature binds these bases, makes one assume that these "teeth" are onchia. The shifting forward of the onchia and the indivisibility of the small oral capsule indisputably indicate the relation of *Triodontolaimus* to the phylogenetic branch, Leptosomatidae—Oxystominidae. Simultaneously, the combination of peculiarities noted above (weak development of the cephalic capsule and powerfully developed onchia located in a comparatively small oral cavity) compel one to assume that *Triodontolaimus* is an extremely unique lateral offshoot of this phylogenetic branch. Hence it cannot be included in Leptosomatidae and merits isolation in an independent family. Because information on the morphology of *Triodontolaimus* is so meager, it seems more appropriate to assume that this group is derived from extremely primitive representatives of the family Leptosomatidae.

To solve the problem of the position of the family Anticomidae, it is necessary to establish whether the structural simplicity of the cephalic end is primary or simply the result of secondary simplification. An examination of the main group of genera of this family, clustering around *Anticoma*, revealed that some forms, such as *Cephalanticoma chitwoodi* (Inglis, 1964), possess an adequately developed cephalic capsule, a poorly developed cephalic bladder, and a small oral cavity with distinct onchia. In *Odontanticoma* the cephalic capsule is almost negligible but the oral cavity well developed (apparently at the cost of its onchial part).

These peculiarities, in association with small size, seem to indicate that the structural simplicity of the anticomid head is the result of secondary simplification due to general reduction in size. The presence of a renette also bespeaks the phylogenetic advancement of anticomids. The genus *Parabarbonema*, with only one species—*P. barba* Inglis, 1964—occupies an intermediate position between anticomids and leptosomatids. The structure of the cephalic capsule in this species is almost identical to that in members of *Leptosomatides*. The oral cavity is small and armed with three large onchia. Thus the anterior end of the body of anticomids could be regarded as the end result of simplification of the form type *Parabarbonema*. *Parabarbonema* is distinguished from *Anticoma* by the absence of a renette.

The taxonomic position of the other two groups of genera, clustered around *Platycoma* and *Barbonema*, is not really clear since information concerning these two genera is very scanty. Nevertheless both groups contain representatives with a highly reduced or even totally reduced cephalic capsule and an extremely small oral cavity; however, small but distinct onchia are present in *Proplatycoma sudafricana* (Inglis, 1964). Furthermore, one cannot refrain from mentioning the well-known resemblance in spicule structure in *Parabarbonema* and *Barbonema*.

Until fuller elucidation of the peculiarities of various anatomical characters is forthcoming, it is better to consider genera clustered around *Platycoma* and *Barbonema* independent of *Anticoma* and genera close to it, and also independent of ancestors analogous to *Parabarbonema*. The absence of a renette in *Parabarbonema* bespeaks the possibility of keeping *Anticoma* and genera close to it isolated in contrast to other genera in this family. If leptosomatids progressed along the path of complexity of the cephalic capsule and oral cavity, anticomids retrogressed along the path of simplification of these structures which, among other reasons, confirms the evolutionary independence of the family.

Two groups included by a number of authors in the composition of the family Leptosomatidae have been deliberately omitted by me in the foregoing discussion. *Rhabdodemia* has been variously included in the family Leptosomatidae (Filip'ev 1927; Schulz, 1932; Schuurmans-Stekhoven, 1946; Schuurmans-Stekhoven and Mawson, 1955), Oncholaimidae (Southern, 1914; Ditlevsen, 1926), Enoplidae (Wieser, 1959; Inglis, 1964; de Coninck, 1965), and sometimes established as an independent family (Filip'ev, 1934). As shown by Inglis (1964), *Rhabdodemia* belongs to the evolutionary branch Phanodermatidae–Enoplidae on the basis of structure of the anterior end of the body, and consequently must be excluded from the family Leptosomatidae. However, it is not easy to establish the position of this genus within the limits of the phylogenetic branch. Inglis (1964) in transferring this genus to Enoplidae, understood by him

in a very broad sense, notes features of its similarity with the family Enchelidiidae. Contrarily, de Coninck (1965) aligns *Rhabdodemanina* with the family Enoplidae in a special subfamily, Rhabdodemaniinae, including therein the genera *Chaetonema*, *Trichenoplus*, *Trileptium*, and *Donsinema*. In my opinion, the latter four genera have very little in common with *Rhabdodemanina* in the construction of the anterior end of the body. I think it is more appropriate to isolate *Rhabdodemanina* as a separate group (separate family), occupying a somewhat intermediate position among enoplids with unequal onchia (*Saveljevia*, *Parasaveljevia*, *Oxyonchus*), i.e., between Oncholaimidae and Enchelidiidae.

The taxonomic position of the genus *Crenopharynx* merits special discussion. Species now related to this genus were earlier included in *Stenolaimus* (Filip'ev 1927; Schuurmans-Stekhoven, 1950), and usually placed close to *Anticomina* or even listed as its synonym. Filip'ev (1934) suggested the name *Crenopharynx* for that group of species which has characters in common with *Stenolaimus* and placed the genus in Phanodermatidae (Filip'ev, 1927). Subsequently many authors related the genus *Crenopharynx* to the family Phanodermatidae, guided by the predominantly alveolar structure of the esophagus. Inglis (1964) showed that the structures of the buccal cavity of Phanodermatidae and Enoplidae were homologous through a detailed analysis of the cephalic structure of *Crenopharynx eina*. On these grounds he emphasized the need for including *Crenopharynx* in Phanodermatidae. However, this inclusion is improper. The homologue only proves the affinity of this genus to the phylogenetic line Phanodermatidae-Enoplidae and does not substantiate its inclusion
40 in either family. *Crenopharynx* can be sharply distinguished from present-day Phanodermatidae, type *Phanoderma*, as well as the aberrant genus *Dayellus* isolated by me in a separate family, by the presence of a triangular oral opening, primitive cephalic capsule with undeveloped cephalic bladder, and shallow buccal cavity devoid of distinctly formed buccal styli. Hence one is compelled to isolate *Crenopharynx* in an independent family to which it seems to me, the genus *Nasinema* should also be transferred. The genus *Nasinema* was formerly included by Filip'ev in Phanodermatidae.

The structure of the head, oral cavity, esophagus (alveolar in the posterior half), and general shape of the body bring *Nasinema* and *Crenopharynx* close together.

Representatives of the family Crenopharyngidae occupy, apparently, a much lower position in the phylogenetic branch to which they belong. Such an interpretation explains the similarity in structure of the cephalic end of Crenopharyngidae and Anticomidae. A poorly developed cephalic capsule and oral cavity are found so often among anticomids and lower leptosomatids that the similarity of these features cannot be overlooked.

Taxonomy of Family Leptosomatidae

The family Leptosomatidae was first established as a subfamily of Enoplidae by Filip'ev in his works of 1916 and 1918 to 1921 and only later raised to the rank of family (de Coninck and Schuurmans-Stekhoven, 1933).

Leptosomatidae comprises a primitive group of free-living marine nematodes with poorly expressed features of specialization. Filip'ev (1927) has stated that the specialization of this family is of such a low order that leptosomatids do not represent a single group but rather a series of isolated genera. However, descriptions of genera of Leptosomatidae since 1930 have directed my thoughts not to isolated genera, but to distinct groups of genera. It seems to me that these groups would be better classified as separate subfamilies.

Leptosomatinae, an extremely primitive subfamily, includes the genera *Leptosomatum*, *Leptosomatides*, *Leptosomella*, *Leptosomatina*, and *Paraleptosomatides* for which a short cephalic capsule with relatively fine walls and a narrow cephalic ring are characteristic. The oral cavity is absent. In most of these genera the tail is short and bluntly rounded. *Leptosomella* has a rather long and conical tail which narrows soon after the anus. Extremely short cephalic setae are also characteristic of this group. Usually the cervical setae, if present, are short. Only *Leptosomatina* Allgen, 1951 and *Leptosomella* Filip'ev 1927, which probably will be combined in the future due to many features of similarity, have long cephalic setae. It is regrettable that Filip'ev described his genus only on the basis of females and Allgen on the basis of males. As the material of these two genera was not available to me I cannot propose their combination at present and tentatively treat *Leptosomatina* and *Leptosomella* as independent genera.

In speaking about the merger of these two genera I have in view their type species—*Leptosomella acrocercera* and *Leptosomatina longisetum* respectively. They are very close in such characters as shape of head, presence of long cephalic setae, absence of eyes, thick cuticle, and absence of cervical setae. These very characters have been given by the authors as diagnostic features of the genera. Allgen later (1958a) described another species of *Leptosomatina*, namely, *L. appendixocaudatum*, in which the cephalic structure is entirely different from that in *L. longisetum*, and for which extremely short cephalic setae and an accessory organ
41 of unique structure are characteristic (latter absent in *L. longisetum*). The differences between *L. longisetum*, and *L. appendixocaudatum* appear to me so incomparably greater than the differences between *Leptosomatina longisetum* and *Leptosomella acrocercera* that a study of supplementary material will no doubt lead to their combination in one genus and the

isolation of *Leptosomatina appendixocaudatum* in an independent genus.

Of all the genera belonging to this group, *Leptosomatum* has the simplest spicular apparatus, i.e., weakly curved spicules and simple gubernaculum devoid of processes. All the remaining genera have a gubernaculum provided with a process, which may be only dorsal or dorsal and ventral.

Thoracostomatinae, isolated by de Coninck (1965), is characterized first and foremost by a highly developed cephalic capsule. It includes such genera as *Thoracostoma*, *Deontostoma*, *Pseudocella*, and *Synonchus* with genera close to it. The genus *Synonchus* originally embraced so many varied forms that many authors had a tendency, totally justified, to divide it into several subgenera. A contrary attempt to unite all forms with a well-developed cephalic capsule and clavate tail created much confusion in the taxonomy of this group. The fact that a series of species were transferred to *Thoracostoma* and some even to *Leptosomatum* (Villot, 1875) on the basis of the structure of the cephalic capsule, exacerbated this confusion. Cobb (1893a, p. 411) described the genus *Synonchus* as follows: "Worms of this genus are closely related to those of *Oncholaimus*. They have a pharynx armed with teeth in which the dorsal one is predominant and the remainder rudimentary. The pharynx is so small that the teeth occupy almost all the space when the mouth is closed. . . . A ventral accessory organ is present in males anterior to the anus." A detailed description of the two species of this genus—*S. hirsutus* and *S. fasciculatus*—came later.

Linstow (1900) described the Arctic species *Enoplus edentatus* in which the oral capsule lacks sclerotized plates and teeth, which characterize representatives of this genus. Due to the absence of the most distinctive character of the genus *Enoplus*, namely, sclerotized jaws, Jägerskiöld (1901) considered it improper to include this species in *Enoplus* and transferred it to *Thoracostoma*. He also gave a detailed description of a new species, *Thoracostoma acuticaudata*, but observed that he was relating it to *Thoracostoma* only tentatively, suggesting that future studies might isolate it in an independent genus. de Man (1904) held the same point of view. Türk (1903) gave a detailed description of two Neapolitan species which he included in *Thoracostoma*. Southern (1914) described two species from the southern coast of Ireland and assigned them to a new genus, *Fiacra*—*F. brevisetosa* and *F. longisetosa*. He considered the genus established by him close to *Thoracostoma*, *Enoplus*, and *Tridontolaimus*, and conceded that with future studies his species might have to be transferred to two different genera.

Filip'ev (1916) suggested the combination of all species with a conical tail, described as *Thoracostoma*, in a new genus—*Jaegerskioeldia*—with the type species *Thoracostoma acuticaudata* (Jägerskiöld). Later Filip'ev

(1927), acknowledging he had not known earlier about Cobb's description of the genus *Synonchus* (Cobb, 1893a), accepted the synonymy of these genera. Observing the great diversity of species in this genus, Filip'ev (1927) divided it into three subgenera: 1) *Fiacra* [type species, *S. (F.) longisetosum*] with long cephalic setae, large amphids, large arcuate spicules, and labial teeth; 2) *Synonchus* s. str. [type species, *S. (S.) fasciculatus*] with cephalic setae of average size, large amphids, large spicules, and without labial teeth; and 3) *Jaegerskioeldia* [type species, *S. (J.) acuticaudata*] with extremely short cephalic setae, small amphids, short spicules, and without labial teeth.

Quite recently it has been suggested (de Coninck, 1965) that these subgenera be elevated to independent genera. On studying those forms which constitute the genus *Synonchus* in its new dimensions, I have arrived at the conclusion that the latter is a composite group but undoubtedly liable to division into a series of genera. Thus, in addition to isolating the above-mentioned *Synonchus*, *Fiacra*, and *Jaegerskioeldia*, I also isolate *Eusynonchus* (type species, *Fiacra brevisetososa*), *Tuerkiana* (type species, *Thoracostoma strasseni*), and *Paratuerkiana* (type species, *Thoracostoma comes*), the descriptions of which are given in the taxonomic part of this work.

Thoracostoma (Synonchoides) galathea, described by Wieser (1956), combines characters of *Synonchus* (short cephalic capsule and buccal capsule with triangular plates) and *Thoracostoma* (shape of tail). As such, the form is isolated here in the subgenus *Synonchoides* of the genus *Thoracostoma*, stipulating that future studies will probably elevate it to the rank of genus. Another subgenus, *Corythostoma*, described by Wieser in the same work, also deserves in my opinion to be raised to the rank of genus as it differs notably from *Thoracostoma* in the structure of its cephalic capsule. The cephalic capsule of *Corythostoma* is wide and short with extremely wide, uniform interlobular grooves. The lateral grooves are not distinguishable in any way from the subventral and subdorsal grooves.

I propose that all the genera enumerated above be combined in the subfamily Synonchinae of the family Leptosomatidae. *Macronchus* Inglis, 1964 should also be transferred here; the author himself expressed doubts about its affinity with *Synonchus*. Finally, *Anivanema*, described by me in the present work, has likewise been placed in the family Leptosomatidae.

The history of the genus *Thoracostoma* warrants review. This genus was established by Marion in 1870 for four species from the Mediterranean Sea: *T. echinodon*, *T. dorylaimus*, *T. montredonense*, and *T. zolae*. To this genus Marion assigned worms with a long, almost nonattenuated body, with a sharply truncated head and a short tail. The presence of

permanent eyes and the absence of an accessory organ in the genital apparatus of the male (papillae present instead) are also characteristic of the genus, but a sclerotized cephalic capsule is the most important diagnostic feature. Species which had earlier been assigned to different genera—*Hemipsilus* (Leuckart, 1849), *Enoplus* (Eberth, 1863; Schneider, 1866), and *Leptosomatum* (Bastian, 1865; Villot, 1875)—were subsequently included in this genus. Later all nematodes with a sclerotized cephalic capsule were likewise included in the genus *Thoracostoma*. The significant difference in the structure of cephalic capsules compelled taxonomists to subsequently define a series of independent genera in Leptosomatidae, namely, *Tridontolaimus* de Man, 1893; *Rhabdodemia* Baylis and Daubney, 1926; *Synonchus* (*Jaegerskioeldia*) Filipjev, 1916; *Platycoma* Cobb, 1893; *Metacycolaimus* Stekhoven, 1946; *Cylicolaimus* de Man, 1890; *Synonchus* Cobb, 1893; *Deontostoma* Filipjev, 1916; *Leptosomatides* Filipjev, 1918; and also a genus already placed in another family—*Phanoderma* Bastian, 1865.

At the commencement of the twentieth century the limits of *Thoracostoma* were more or less firmly established. In 1916 Filip'ev isolated from *Thoracostoma* a new genus—*Deontostoma*. Filip'ev regarded the presence of a ventral process in the cephalic capsule of *Thoracostoma* and its absence in *Deontostoma* one of the basic differences between these two genera. He labeled this process rather inappropriately the "ventral tooth." This term misled foreign authors (Wieser, 1953c; Mawson, 1958a) and hence they refused to recognize the independence of genus *Deontostoma*.

On the basis of a thorough study of the work of Filip'ev and the material at my disposal, I consider it necessary to restore the independence of the genus isolated by Filip'ev (Platonova, 1962). The genus *Deontostoma* differs significantly from *Thoracostoma* in cephalic structure as well as in structure of the spicular apparatus. The cephalic capsule of representatives of *Deontostoma* is almost equal in width near the top and base and its entire wall is of the same thickness; as a result it has a radially symmetrical structure and appears symmetrical in a lateral view. In representatives of *Thoracostoma* the cephalic capsule narrows considerably from the base to the top; its ventral wall is significantly longer and thicker than the dorsal wall and due to this fact the capsule has a bilateral symmetrical structure and appears asymmetrical in a lateral view.

The spicules of males of *Deontostoma* are rather uniformly curved in their proximal part and devoid of sharp angles and grooves. The gubernaculum has distinct dorsal and ventral processes which are joined together by a curved plate. When they fuse, the halves of the gubernaculum form a sort of funnel in which the distal parts of the spicules lie. In *Thoracostoma* the spicules have a sharp break approximately midlength

and a velum often occurs on their internal margin. The terminal part of the gubernaculum encloses the distal end of the spicules. The processes of the gubernaculum are directed dorsally and although closely apposed to the spicule body are situated at a sharp angle to it.

These are the most distinctive characters differentiating *Deontostoma* and *Thoracostoma*. Several minor characters are detailed below.

Filip'ev (1927) divided *Thoracostoma* into two subgenera—*Thoracostoma* s. str. and *Pseudocella*. He related nematodes with distinct photosensitive eyes and the above-described spicular structure to subgenus *Thoracostoma*. To the subgenus *Pseudocella* he transferred species completely devoid of photosensitive pigment, or with pigment scattered throughout the anterior end of the body, or very rarely species with eyes always devoid of a lens. The spicules in this subgenus are smoothly arcuate and notably narrower than those of the subgenus *Thoracostoma* s. str. The gubernaculum has a caudal process directed at right angles to the longitudinal axis of the spicules. Filip'ev predicted that these two subgenera would be elevated to independent genera with the passage of time. Wieser (1956) notes that the structure of the spicular apparatus is a much more important character for the division of these two subgenera than the presence or absence of photosensitive organs since in all cases the latter may be totally absent even in *Thoracostoma* s. str.

A study of the species of *Pseudocella* permits me to supplement the diagnosis of this subgenus given by Filip'ev. All three genera are combined in a single subfamily, Thoracostominae, of the family Leptosomatidae.

If leptosomatids of the subfamily Thoracostominae reveal a tendency toward greater complexity of the cephalic capsule, then the genera *Cylicoliamus*, *Ritenbenkia*, and *Metacylicoliamus* exhibit complexity of the oral capsule. In this group together with greater complexity of the cephalic capsule, developed significantly less however than in Thoracostominae, one finds a powerful oral capsule with a thick sclerotized lining armed with onchia. Representatives of *Metacylicolaimus* have a relatively wide but not very deep oral cavity in which blunt onchia jut out. In *Ritenbenkia* the oral cavity is significantly longer than that in the previous genus, its wall much thicker, and the onchia and sclerotized plates arranged around the oral opening. Maximum development of the oral cavity occurs in *Cylicolaimus*, where it is very extensive, thick-walled, and usually armed with large teeth situated in its depth.

44 Such a characteristic feature as the presence of a highly developed oral cavity and associated changes in the structure of the head permit one to combine the above-mentioned genera with *Paracylicolaimus* in the subfamily Cylicolaiminae.

Taxonomy of Family Anticomidae

Like leptosomatids, anticomids were also isolated in a separate subfamily within Enoplidae by Filip'ev in his works of 1916 and 1918 to 1921. In much later works (1927, 1934) Filip'ev transferred the genus *Anticoma* to Leptosomatinae. de Coninck (1965) assigned Anticominae to the family Leptosomatidae but Hope and Murphy (1972) raised it to the rank of a family.

The family Anticomidae includes many diverse genera that can be grouped into different subfamilies. The genera *Anticoma*, *Paranticoma*, and *Anticomopsis*, as well as *Cephalanticoma* and *Odontanticoma*, newly described in the present work, *Antopus* (with a single species, *A. serialis*) and *Stenolaimus* (with a single species, *S. lepturus*) can all be included in Anticominae.

For a long time *Stenolaimus* occupied in extremely uncertain position in the classification of Anticomidae. Filip'ev (1934), conceding its heterogeneity, proposed *S. lepturus* as a synonym for *Anticoma* and proposed the genus *Crenopharynx* for the species *Stenolaimus marioni*, retaining it in the family Phanodermatidae. I agree with this concept only because this genus really is in need of a similar division into two independent genera. But *Stenolaimus lepturus* considerably differs from the genus *Anticoma* in structure of the spicules and situation and number of cervical setae, and must indisputably be regarded as an independent genus of the subfamily Anticominae. The position of *Crenopharynx* has already been discussed.

Barbonema and *Parabarbonema* differ so much from each other that each deserves isolation in a special subfamily—Barbonematinae and Parabarbonematinae.

Platycoma and *Platycomopsis* should also constitute a separate subfamily.

ECOLOGY

The ecology of free-living nematodes has been studied little and needs special attention and collection of material. As the basic purpose of this work is the taxonomic revision of the lower Enoplida, I had to utilize to some extent the casual and fragmentary information available while examining some problems regarding the ecology of this group.

The opinion that free-living nematodes were highly eurybiont prevailed among nematologists for many years. Kreis (1934) proposed that while inhabiting those parts of the sea which are advantageous to their existence, these nematodes do not exhibit a distinct affinity for definitive biotopes. This means that one and the same nematode species may be

found at different depths and in entirely different conditions, inhabiting littoral aquatic plants and sublittoral silts nonselectively.

Schuermans-Stekhoven (1931) studied the nematode fauna of the North Sea and Zuider Zee and discovered a large number of euryhaline forms among these organisms.

Gerlach (1958) notes that differences in biotopes with respect to fauna of free-living marine nematodes are not large; this fauna lives mainly in sandy beds and sublittoral facies. He thinks that the major percentage of these species are eurybionts. Gerlach, however, has noted a rather significant difference in nematodes in the littoral zone on the basis of biotopes.

It has also been established by Gerlach (1953, 1958) and Wieser (1951, 1953, 1960) that the distribution of nematodes reflects not so much a variety of biotopes, as a peculiarity in substratum, in particular the degree of its disintegration. In the distribution of nematodes on the basis of their association with various types of algae, species affinity of algae is not so significant as its external form, size, and degree of fragmentation of thallus. The same may be said about the substratum. Silt, fine sand, pebbles, and coarse sand should be examined first of all for the presence of interstices which may serve as shelters for the worms (p. 324).

Wieser (1953) has given an interesting ecological classification of free-living nematodes. It is based on the structure of the oral apparatus, which reflects the nature of feeding and consequently to some extent the type of life led by these organisms. All free-living nematodes are divided by Wieser into four groups (Figure 13).

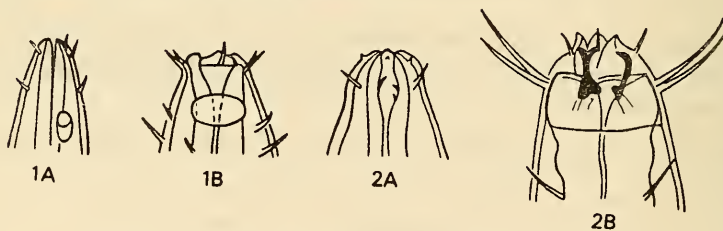


Figure 13. Types of structure of the oral cavity of nematodes (from Wieser, 1953b).

1A—*Anticoma*; 1B—*Paramonhystera*; 2A—*Microlaimus*; 2B—*Enoplodes*.

1. Group 1A. Forms lacking a distinct oral cavity and any kind of oral armature. Type of feeding, simply sucking with the help of the esophageal muscles. Food, detritus consisting of fine soft particles. Thrive among algae in the littoral zone and soft beds in the sublittoral.

2. Group 1B. Forms with well-developed cyathiform, conical, or cylindrical oral cavity which, like the previous group, are also devoid of an oral armature. Feeding as in group 1A. In addition, intake of detritus

effected by movement of lips and mouth. Food, detritus consisting of larger particle (including diatoms). Inhabit soft sand rich in detritus.

3. Group 2A. Oral cavity well-developed and armed with small sclerotized denticles or plates. Nematodes capable of scraping the surface of algae or piercing their membrane to suck cellular sap. Food, epiphytes and algae. Inhabit almost all biotopes.

4. Group 2B. Oral cavity large with a powerful armature of different types. Feeding as in the previous group but most of these nematodes are predators, swallowing small organisms, including other nematodes. Live in the littoral zone in coarse sand poor in detritus, and the sublittoral among shells and sand with larger particles.

Species of Leptosomatidae and Anticomidae play a significant role in the fauna of free-living marine nematodes. The number of their species ranges from 5 to 46 (or from 2 to 19% in relation to the total number of nematodes studied). My data and that collected from literature is presented in the Table which follows.

46 In some cases leptosomatids may be the predominant group. In my analysis of Lake Kergelen there were nematodes representing ten different families, of which 50.7% belong to Leptosomatidae (Platonova, 1968). In material also collected from Lake Kergelen, Schuurmans-Stekhoven

Proportion of lower Enoplida in the fauna of nematodes of some seas

| Water body | Number of individuals | | | Number of species | | | Author |
|--|-----------------------|----------|-----|-------------------|----------|-----|--------------------------------|
| | Total | Enoplids | | Total | Enoplids | | |
| | | Number | % | | Number | % | |
| Barents Sea | 1,040 | 203 | 19 | 121 | 26 | 21 | My data |
| North Atlantic, Norway-Green- land coast | — | — | — | 33 | 9 | 27 | Ditlevsen, 1926 |
| Norway, Trond- heims Fiord | 2,560 | 182 | 7 | 177 | 6 | 3.3 | Allgen, 1933 |
| Arctic Sea | — | — | — | 292 | 46 | 15 | Allgen, 1957 |
| North Sea | — | — | — | 206 | 11 | 5 | Schuurmans- Stekhoven, 1935 |
| Baltic Sea, Oresund | 3,000 | 248 | 8 | 222 | 5 | 2.2 | Allgen, 1935 |
| Kiel | 3,000 | 195 | 7 | 254 | 5 | 1.9 | Gerlach, 1958 |
| Mediterranean Sea | 614 | 54 | 8.7 | 338 | 24 | 7 | Schuurmans- Stekhoven, 1950 |
| Black Sea | 5,750 | 115 | 2 | 122 | 5 | 4 | Platonova, 1962a |
| Tropical part of Pacific Ocean | 970 | 142 | 14 | 109 | 15 | 13 | Allgen, 1961 |
| Antarctic and sub- antarctic seas | 5,582 | 546 | 9.8 | 343 | 21 | 6 | Allgen, 1959 |

and Mawson (1955) found that leptosomatids constituted 88.0% of the total population of nematodes.

It must be noted that when a ladle with a small catchment area, say 20 to 40 cm² (for example, the standard earthen bowls and other devices for making probes into the meiobenthos) was used for collecting nematodes, leptosomatids were rarely caught. They were most frequently found in bottom-combing probes where the sweep of the catching device was considerably larger. This fact, which I have noted a number of times, most probably indicates that the average distance between individuals of this group in nature is many times more than in other usually smaller marine nematodes. Possibly this is due to the larger size of leptosomatids (compared to other representatives of the meiobenthos in general and all other nematodes in particular) and also due to their greater mobility.

Hence leptosomatids should be included in the category of organisms of the macrobenthos in spite of their relatively smaller size. A predominant majority of other free-living nematodes are organisms of the meiobenthos, which is supported by their being found in large numbers during standard meiobenthic probes.

Biotopes. All the families examined in the present work are found by and large in all biotopes, from littoral water plants to deep water silt.

Leptosomatids live mainly among larger types of algae with a less fragmented thallus. These are usually brown algae (*Fucus*, *Ascophyllum*, *Laminaria*, *Alaria*) and rather rarely red algae (*Porphyra*, *Rhodimenia*, *Halosaccion*, *Lithothamnion*, and *Corallina*). There are numerous nematodes in these which belong to group 1A according to the classification of
47 Wieser (1953). Representatives of this group cannot utilize the algae they inhabit as food material due to the structure of their oral apparatus. Most probably the algae provide them with shelter and mechanical protection against the impact of sea breakers. As is well known, nematodes live in the surface layers of the seabed (Cobb, 1929; Filip'ev, 1934) and for this reason those which live in the zone of sea breakers and the littoral belt need that type of defense.

Wieser (1951) states that smaller forms of nematodes inhabit algae with a highly fragmented thallus (for example *Ceramium*) and large forms usually inhabit larger algae with a slightly truncated thallus. My observations confirm Wieser's. Among shrublike and highly dismembered algae I found small forms of Chromadoridae in abundance, but larger leptosomatids (average size 6 to 15 mm) in larger types of brown algae. Group 1B does not include lower Enoplida.

Representatives of group 2A are found in algae of the littoral zone in very significant numbers: some (for example *Pseudocella trichodes*) may be considered exclusive inhabitants of this biotope. They are capable of utilizing algae not only for shelter but also as food on the basis of their

oral structure. Group 2B was represented in my material by just six species of two genera—*Cylicolaimus* and *Rhabdodemania*. In the littoral zone these nematodes are almost nonexistent. Information in literature on the distribution of other species of these genera support my observations.

Fine sand and gravel are only slightly inhabited by nematodes as these organisms are easily swept away by waves from ground not protected by algae.

Silted sand in the sublittoral zone provides favorable conditions of habitation for nematodes. Here the impact of sea breakers is not felt, food reserves are sufficient, and far better aeration insured compared to silt. As shown by Wieser (1960) the degree of ground siltation exerts a great influence on the development of nematode fauna. Little silt preserves the interstices in which nematodes take refuge. When such spaces are filled by silt, a severe impoverishment of nematode fauna takes place. All three groups—1A, 2A, 2B—are widely represented in this biotope. As mentioned above, group 2B is found almost exclusively in silted sand beds of the sublittoral zone. Wieser (1951, 1953) believes that species of this group are capable of feeding on small organisms as well as algae, because of the presence of a large and well-armed oral cavity. Proceeding from the fact that species of this group are found very rarely in algae and live mainly in silted sand grounds, one may assume that these nematodes are mainly predators. As far as mollusks are concerned, Filip'ev (1927) and Schuurmans-Stekhoven (1950) think that this biotope always contains a rich fauna of nematodes. But leptosomatids are found among mollusks (lamellibranchs) rather rarely and usually singly. However, Filip'ev (1918) claims he often found species *Leptosomatum bacillatum* among bivalve mollusks.

Biotopes containing large numbers of invertebrates (independent of their taxonomic groups) are particularly rich in nematodes. Areas rich in polychaetes, mollusks, echinoderms, and sponges are equally rich in nematodes. Probably as a result of the vital functions of these organisms, the substratum is enriched with nutritive material utilized by nematodes.

Temperature. I have no experimental data on the influence of temperature on nematode fauna. Only this observation can be made: *Deontostoma arcticum*, *Pseudocella arcticum*, *P. coecum*, and *P. trichodes* are found predominantly in cold seas of the Arctic basin, while *Eusynonchus* 48 *hirsutus*, *Leptosomatum bacillatum*, and *Leptosomatides euxina* live in the warm waters of the Black Sea and the Mediterranean Sea. *Leptosomatum elongatum* may be considered eurythermal as it has been found in the cold waters of the subantarctic, Barents Sea, and Greenland Sea, as well as in the warm waters of tropical parts of the Indian and Pacific Oceans. Data on the remaining species discussed in this work are so meager that

at present it is difficult to comment on the influence of temperature on them.

The influence of temperature on nematode fauna is detailed by Gal'tsova in the present volume (p. 327).

Salinity. Filip'ev (1927), Schuurmans-Stekhoven (1931), and Allgen (1935) observed a reduction in number of free-living marine nematodes in relation to lower salinities. This is clearly evident for nematodes of family Leptosomatidae and Anticomidae. Most species of these families may be considered stenohaline. Fauna comprising leptosomatids and anticomids is much richer with respect to species in seas with a normal oceanic salinity (46 species, see Table). In such seas species of Leptosomatidae constitute 27% in relation to the total number of nematode species. In seas with lower salinities (Black Sea—17‰, Baltic Sea in region of Gulf of Kiel—13 to 19‰, and considerably diluted waters of Trondheims Fiord, Sea of Norway) the number of species of leptosomatids and anticomids falls to 4.0 to 6.0% or 1.9%. In the Sea of Azov (Filip'ev, 1922c; Mordukhai-Boltovskii, 1960) and in the eastern part of the Baltic Sea (Filip'ev, 1929; Schneider, 1906) leptosomatids are entirely absent.

Representatives of Thoracostomatinae (*Thoracostoma*, *Deontostoma*, *Pseudocella*) are probably rather sensitive to lower salinity. Thus in the western part of the Baltic Sea species of these genera are found very rarely and usually singly, while in the Black Sea they are totally absent. The subfamily Leptosomatinae (*Leptosomatum*, *Leptosomatides*, *Leptosomella*) is usually found in seas with normal salinity. However, two species of *Leptosomatum* (*L. punctatum* and *L. bacillatum*) and *Leptosomatides euxina* inhabit the Black Sea.

The family Anticomidae is, as a whole, more euryhaline. Moreover, species of *Anticoma*, which live in all seas except those with very low salinities, are the most euryhaline. Rhabdodemaniidae is adapted to great fluctuations of salinity. In my material there were specimens of *Rhabdodemia* from the Okhotsk, Barents, White, and Black Seas. According to Allgen (1929a) and Gerlack (1958) species of the genus are also found in the western part of the Baltic Sea.

The possibility is not ruled out that there may be other factors affecting the prevalence of leptosomatid fauna in the Black and Baltic Seas, but indubitably the considerable dilution of water is significant.

The problem of the ecological characteristics of individual species is incomparably more complex. Diversity of habitat and extremely wide distribution of species in various seas and under various conditions (depth, nature of the bottom, and salinity) indicate that many species of this group have adapted to a wide range of ecological factors.

It seems to me that the eurybiont nature of free-living marine nema-

todes has been rather exaggerated. In my material there were no species encountered in all biotopes in equal measure. Moreover, some species exhibit a distinct affinity for definite conditions. For example, *Pseudocella trichodes* is an inhabitant of large littoral algae, while *P. tenuis* and *P. elegans* live predominantly in depths of about 300 to 400 m in brown sublittoral silts.

It is difficult to express an opinion on the ecological affinity of species encountered in extremely diverse biotopes. At first sight they appear widely eurybiont and are so regarded by a number of authors (Schulz, 1932; Kreis, 1934; and others). However, it seems to me a careful analysis of the prevalence of these species makes it possible to pinpoint the biotope in which a definite species is encountered on a large scale or, at any rate, in a larger percentage than in other biotopes. Such a phenomenon bespeaks, first of all, the affinity of a species for certain conditions; the solitary occurrence of specimens of species in other biotopes can be attributed to wider limits of adaptation inherent in free-living nematodes, especially primitive marine forms. Sometimes cases of occurrence of nematodes of a single species in biotopes markedly different from each other can be explained by their drifting with fragments of algae or erosion of beds by sea breakers. Nematodes are also subject to wide-flung dispersal by currents due to their ability to cling to the surface film of the water. An example of a widely distributed species is *Pseudocella kurilensis*; definite conclusions about its ecological affinity can be derived from its prevalence. This species was found at five places in the littoral zones of Kuril Islands in pieces of algae; however, in all five places only solitary specimens were collected. Contrarily, as many as 49 specimens were found among a dense mass of sponges caught in a probe at 25 m depth on a slightly silted bed. In all probability the latter biotope is favorable for the prevalence of this species.

The species *Leptosomatides brevisetosum* seems to thrive in the littoral zone where it can find shelter in thickets of large algae, although solitary specimens have been recorded from a sandy bottom at a depth of 53 m.

It seems to me that grouping nematodes, as done by Wieser (1953), on the basis of their oral apparatus without taking into account their physiological indexes is incorrect. For example, in my material three species of *Pseudocella* would fall in group 2A but they apparently thrive under notably different conditions. *P. trichodes* is an inhabitant of littoral algae, while *P. tenuis* and *P. elegans* live in sublittoral silts. It is possible that *P. trichodes* is a more oxygen-loving form than *P. tenuis* and *P. elegans*.

The foregoing account shows that in spite of the few studies done to date the ecological characteristics of the family have been sufficiently defined to reveal its primitiveness. For a better understanding of the

ecology of this group, obviously more field and experimental studies are needed.

GEOGRAPHIC DISTRIBUTION

The geographic distribution of free-living marine nematodes of this group is steeped in confusion and even vacuity.

In the 1920's and 1930's many researchers considered free-living marine nematodes cosmopolitan. The hypothesis was put forward that zonal climatic factors play no role in the distribution of nematodes (Steiner, 1915; Kreis, 1934). Chitwood (1936a, 1936b, 1937, 1951, 1960) disputed this point of view on the grounds that species found by him on the coast of America differed significantly from those found on the European coast. Authors of a number of contemporary studies (Gerlach, Mawson, Meyl, Timm, and Wieser) subscribe to this point of view and believe that among free-living marine nematodes there are far fewer cosmopolitan forms than indicated earlier. Thus Wieser (1953c) found an extremely small number of species of nematodes, common in the fauna studied by him, on the coast of Chile and in coastal waters of Europe. Gerlach (1955) studied the fauna of marine nematodes of the Pacific coast
50 of Central America, and Meyl (1956, 1957) nematodes of the coast of Brazil; they found a large number of new forms not seen in areas studied earlier.

Schuermans-Stekhoven (1950) in his comprehensive work on nematodes of the Mediterranean Sea compared this fauna with that of the North, Baltic, Mramor, Black, and Azov Seas. He established that species of nematodes common to the North, Baltic, and Mediterranean Seas do not exceed 15% of all the nematode fauna of the Mediterranean Sea. Contrarily, the Mramor, Black, and Azov Seas have a much larger number of species common to the Mediterranean Sea (34.79%). For the Mramor this figure was 65.0%, the Black 29.5%, and the Azov 33.3%. Thus the fauna of the Mediterranean Sea reveals greater similarity to the fauna of the Mramor, Black, and Azov Seas than that of the more northern seas of Europe. Such an interrelation of the nematode fauna of these seas should be expected if the distribution of these organisms is subject to the same biogeographic phenomena as other groups of marine benthic organisms.

Data presented in a number of works by Allgen (1956a, 1956b, 1956c, 1957a, 1958b, 1958c) are none too clear in this respect. This author compares regions situated great distances apart. Comparing the Norwegian nematode fauna with that of other regions he states that at different points of the Norwegian coast significant percentages of species are found which are common to the Mediterranean Sea, tropical seas along

the coasts of Australia and South America, and also the Antarctic and subantarctic.

In other works (1934a, 1954b) Allgen has stated that several species are bipolar in distribution. All the bipolar species in the northern hemisphere live mainly in temperate waters (coast of Norway, England, and France) and in the southern hemisphere in the subantarctic region (islands of Terra del Fuego, Falklands, Campbell, Kergelen, MacCoury, Tasmania, New Zealand) or even in the Antarctic. Thus the distribution of these nematodes is similar to other groups of bipolar organisms. It is interesting that while discussing the reasons for the bipolar distribution of nematodes, Allgen refutes the present theory about the bipolarity of species; their settling in various oceanic depths (Chun, 1897), along the western coasts of continents, particularly in relation to Quaternary glaciation (Ortmann, 1896; Berg, 1920), and bipolarity as a result of a uniformly wide distribution during the Tertiary period (Deryugin, 1915). Allgen holds that bipolar species as well as species known to exist only in one hemisphere are, as a matter of fact, much more widely distributed and their known occurrences are casual and fail to reflect the size of the area of occurrence. Thus in his zoogeographic concepts Allgen reverts to the old opinion that free-living marine nematodes are cosmopolitan forms.

Not negating the possibility of a bipolar distribution of nematodes, I nonetheless hold that for an accurate analysis of this problem a thorough morpho-systematic study of bipolar species is essential. In the works mentioned above Allgen includes two species of Leptosomatidae among bipolar species. The first, *Pseudocella elegans*, was discovered by Ditlevsen (1922) in the subantarctic but as he had only one specimen, not sexually mature, he gave no description. Ditlevesen later discovered (1926) a large number of sexually mature nemotodes in Skagerrak, which he considered identical to the subantarctic specimen, and gave a description of the species. It seems to me that on the basis of the foregoing account one cannot conclude that this species is bipolar in distribution.

Allgen labeled the second species, *Thoracostoma coronatum*, bipolar 51 in distribution because he annexed it with *Triceratonema campbelli*. I cannot accept this contention because the identification of these two forms is erroneous; the former should be considered a Mediterranean form and the latter Antarctic.

Rhabdodemia minor, *Leptosomatium arcticum*, *Deontostoma arcticum*, and *Pseudocella trichodes* appear at first glance to be species with a bipolar distribution.

Rhabdodemia minor has been recorded from the coast of Ireland to that of Murmansk and once recorded by Allgen (1959) from the coast of Antarctica. The occurrence of this species in the Antarctic Ocean appears

dubious to me as only one female and two immature specimens were recorded, as a result of which identification could have been inaccurate. Specimens of *Leptosomatium arcticum* were recorded by Filip'ev from the Barents Sea near Murmansk (1916) and near the coast of Novaya Zemlya (1927). Mawson (1958b) found this species in the Antarctic and subantarctic. There are sufficient grounds to assume that here the authors were dealing with two different species. The species described by Filip'ev was based on a single female specimen and later a sexually immature specimen found by him near the coast of Novaya Zemlya. A large number of nematodes with mature males and females were at the disposal of Mawson. Having no opportunity to compare the males, on the basis of which a precise species identification is possible, Mawson made a mistake in identification and considered these two different species one.

The bipolar distribution of *Deontostoma arcticum* appears more valid. This species is often found in the lower Arctic and in boreal waters. It has occurred in large numbers along the southern coast of Chile (Wieser, 1953c) and in the Antarctic and subantarctic (Mawson, 1956, 1958b). *Pseudocella trichodes* is a species widely distributed in the littoral zone in Arctic and boreal waters, living mainly among larger algae. Allgen (1951, 1959) indicates the occurrence of this species on the south coast of Australia (one sexually immature specimen) and in the subantarctic in the sandy and silted-sand benthos of the sublittoral zone. *Leptosomatium gracile* and *Synonchus fasciculatus* have been recovered from the North Atlantic and Arctic Oceans. They were also detected by Allgen in the subantarctic region.

Thus the problem of the bipolarity of distribution of marine nematodes requires, in my opinion, a special exhaustive study.

If the distribution of marine nematodes is mainly subject to the same zoogeographic laws as the distribution of all other groups of marine benthic organisms, then one may essay, albeit in general outlines only, a zoogeographic classification of species and isolate groups of species characteristic for different seas. It should be noted beforehand, however, that specialists of other groups of marine benthos deal with organisms for which the zoogeographic characteristics are fairly clear. For marine nematodes this problem is still in the process of elucidation. To explain this phenomenon I want only to show the potentiality of a zoogeographic analysis of distribution of lower Enoplida. Marine nematodes in general have been studied too little to permit an exhaustive review. With the manifestation of affinity (adaptation) of an individual species to one or another zoogeographic entity, one can be guided by the scheme of zonation of the seas (to be exact, sublittoral) proposed by Ekman (1935, 1953), which is universally recognized.

Odontanticoma murmanica, *Anticoma insulaealbae*, *Leptosomatium*

tetrophtalmum, *Leptosomatides steineri*, *Pseudocella tenuis*, and *P. coecum* are known from numerous points only in the Kara and Barents Seas. On the basis of their type of distribution they can probably be considered Arctic species. Other Arctic species include *Deontostoma magnificentum* and *Ritenbenkia micropapillata*, and species described earlier in this work—*Anticoma filipjevi*, *A. grandis*, and *Pseudocella gracilis*. But since
52 these species were only found singly, it is difficult to make a statement about their distribution.

Anticoma arcticum *A. minor*, *Rhabdodemia minor*, *R. gracilis*, *R. scandinavia* and *Synonchus murmanicus* should probably be considered arctico-boreal species, because of their occurrence in the Kara and Barents Seas as well as in the boreal waters of Europe (North Sea and coast of England).

I cannot assign a number of species to a definite group because they constitute solitary specimens. However, as their habitats are widely dispersed in the Kara, Barents, and Norwegian Seas, the North Atlantic, and the North Sea, it may be that the following species will eventually be regarded as arctic, arctico-boreal, or less probably boreal species: *Anticoma strandi*, *A. brevisetosa*, *Odontanticoma vanoorti*, *Crenopharynx armatus*, *Barbonema setifera*, *Platycomopsis cobbi*, *P. mesjatzevi*, *Leptosomatides microlaimum*, *L. crassus*, *Leptosomatium arcticum*, *L. groenlandicum*, *L. breviceps*, *Leptosomella acrocerca*, *Pseudocella conicaudata*, *P. saveljevi*, *P. filipjevi*, and *Deontostoma lobatum*. There is an analogous collection of species known to occur only singly from the Bering Strait, Commander Islands, and the northern part of the Sea of Azov: *Anticoma behringiana*, *A. curta*, *A. uschakovi*, *Rhabdodemia ochotensis*, *R. breviceudata*, *Leptosomatium papillatum*, *L. grebnickii*, *L. behringicum*, *Pseudocella acuta*, *P. angusticeps*, and *Deontostoma papillatum*. Like the species in the previous list, they are also arctico-boreal or upper boreal. However, at present it is not possible to define their biogeographic characters.

Anticoma eberthi, *A. limalis*, *Crenopharynx marioni*, *Rhabdodemia laticauda*, *Leptosomatium gracile*, *Metacycolaimus flicaudatus*, *M. obtusidens*, *M. flagellicaudatus*, *M. effilatus*, and *Fiacra longisetosa* may be considered boreal-European forms distributed along the coasts of Norway, in the North and Baltic Seas, and along the coasts of England and France. From the records of solitary specimens it is possible that the following species may belong to this group: *Anticoma zosterae*, *A. microseta*, *Barbonema horridum*, *Rhabdodemia striata*, *R. major*, *Platycomopsis effilatus*, *Leptosomatium caecum*, *Jaegerskioeldia acuticaudata*, and *Eusynonchus brevisetosus*.

Species found by me near South Sakhalin and Kuril Islands may apparently be considered boreal or even lower boreal-Pacific forms:

Odontanticoma dentifer, *Leptosomatium acutipapillosum*, *L. brevisetosum*, *Anivanema magna*, *Pseudocella bursata*, *P. kurilensis*, *P. mamillifera*, and *P. truncaticauda*. Probably *Rhabdodemia illgi* from the Pacific coast of North America, known only from one habitat, could be transferred to this group.

A significant number of anticomids and leptocomids live in the Mediterranean Sea. Frequently found species include: *Anticoma pellicida*, *Platycoma cephalata*, *Crenopharynx paralepturus*, *Leptosomatium punctatum*, *L. bacillatum*, *Anivanema magna*, *Cylicolaimus* (?) *jaegerskioldi*, *Eusynonchus hirsutus*, *Tuerkiana strasseni*, *Paratuerkiana comes*, *Deontostoma montredonense*, *Thoracostoma coronatum*, and *T. zolae*.

Anticoma tyrrhenica, *Crenopharynx metagracilis*, *C. brevicaudatus*, *Pseudocella cavernicola*, and *P. citronicauda* have been recorded as isolated cases from the Mediterranean Sea. It is important to note that some of the species go beyond the limits of the Mediterranean Sea and are distributed in the north up to the coast of England, while others are not known beyond the limits of this sea.

From the Black Sea only five species have been recorded: *Anticoma pontica*, *Leptosomatides euxina*, *Rhabdodemia pontica*, *Leptosomatium punctatum* and *L. bacillatum*. Of these, the first three have been discovered only in the Black Sea to date.

- 53 In the Indo-West Pacific (Southeast Asia: Malayan Archipelago and along the Indian coast) single finds have been recorded for these species: *Anticoma ditlevseni*, *A. aberrans*, *A. procera*, *A. profunda*, *Anticomopsis filicauda*, *A. tenuicollis*, *Paranticoma elegans*, *P. bandaensis*, *P. profunda*, *Platycomopsis fliappendicatus*, *Leptosomatium ranjhai*, *L. keiense*, *Leptosomatides reducta*, *Leptosomatina longisetum*, *Thoracostoma philippinensis*, and *T. karachense*. To date these species have not been discovered outside the limits of this region.

Steiner and Albin (1933) and Allgen (1951) recorded the following species of anticomids and leptosomatids along the Pacific coast of Central America: *Anticomopsis tenuis*, *Leptosomatium pedroense*, *Deontostoma microlobatum*, *D. anchorilobatum*, *D. jollaensis*, *D. californicum*, and *Pseudocella panamaense*. Only isolated specimens were found, however.

Anticomids and leptosomatids are particularly numerous in the Antarctic and subantarctic. Species repeatedly encountered include: *Anticoma longissima*, *A. dahli*, *A. tenuis*, *A. filicauda*, *A. subsimilis*, *A. major*, *A. columbia*, *A. campbelli*, *A. australis*, *A. wieseri*, *Paranticoma antarctica*, *Crenopharynx antarcticus*, *Platycomopsis dimorphica*, *Deontostoma auclandiae*, *D. antarcticum*, *Thoracostoma anocellatum*, *T. vallini*, *T. angustifissulatum*, *T. campbelli*, *T. setosum*, *T. australe*, *Pseudocella brachychaites*, *P. tabarini*, and *P. polychaetes*. Species found only once: *Anticoma lata*, *A. kerguelensis*, *A. extensa*, *A. longisetosa*, *A. graciliceps*,

Paranticoma odhneri, *P. tubuliphora*, *Anticomopsis gibbonensis*, *Crenopharynx serialis*, *C. crassus*, *Rhabdodemia calicolaimus*, *Platycomopsis paracobbi*, *Leptosomatium australe*, *L. crassicutis*, *L. kerguelense*, *L. clavatum*, *Leptosomatides conisetosum*, *Deontostoma papillosum*, *D. demani*, *Thoracostoma chilensis*, *T. schizoepestylium*, *T. parasetosum*, and *T. brunni*. The study of marine nematodes is too inadequate to permit a statement as to which species are exclusively adapted to the subantarctic and which to the Antarctic. However, it may be noted that most of these species are known from both the subantarctic and the Antarctic.

The distribution of four species of *Anticomopsis* has given rise to doubts. *Anticomopsis pellucida* is widely distributed in the Mediterranean Sea. It has also been detected in the North Sea and the North Atlantic as far as the Barents Sea. Allgen (1927b) recorded this species for the subantarctic. *A. acuminata* has also been mainly found in the Mediterranean Sea and adjacent seas; it has also been recorded from Zond Islands (Micoletzky, 1930), the Pacific coast of North America (Wieser, 1959), and in Antarctic and subantarctic waters (Allgen, 1959). *A. limalis* is often found in the Arctic, North Atlantic, North Sea, and the western part of the Baltic Sea. Allgen recorded this species near the coast of California (1947b), for the Pacific and Atlantic coasts of Central America (1947a, 1951), the Caribbean Sea, the Philippines, the Hawaiian Islands, and the Antarctic and subantarctic (1959). *A. similis* has been recorded from the coast of Australia, Terra del Fuego, in the subantarctic, Antarctic, and Zond Islands. Probably these four species could have been classed as cosmopolitan had their taxonomic status not undergone so much confusion. As is mentioned in the taxonomic part of this work, some were combined repeatedly into one species and then separated subsequently. The limits of these species have not yet been distinctly defined. Hence the problem of their distribution must remain open.

Three species of *Leptosomatium* also appear at first glance to be cosmopolitan in distribution. *L. bacillatum* is widely distributed in the Mediterranean and adjacent seas and has also been recorded from the Norwegian and Caribbean Seas, on the coast of California and South Australia, and in the subantarctic and Antarctic. However, Allgen (1940a, 1943, 1947, 1951, 1959) had insufficient material at his disposal: most of it comprised females and sexually immature specimens. For this reason I am inclined to consider *L. bacillatum* a Mediterranean species. *L. sabangense* was first recorded from the islands of Zond by Steiner (1915). Later this author mentions the discovery of this species along the coast of Venezuela; as only a single sexually immature specimen was found I fear his identification may have been erroneous. Micoletzky recorded this species from the Red Sea (1922) and Allgen from the Mediterranean Sea, the coast of California, Panama (along the Pacific coast), Australia, and the Falkland

Islands. In every instance only one to three specimens were recovered and these too were sexually immature. For this reason I consider it an Indo-West-Pacific species. *L. elongatum* is widely distributed in the arctic and boreal regions. It has been found in the Mediterranean Sea, in the tropical part of the Indian Ocean, in the Pacific Ocean, on the coast of Panama and California, on the western coast of Australia, in the subantarctic and Antarctic.

It is extremely difficult to determine the geographic distribution of some species. *Anticoma ditlevseni* was first recorded by Micoletzky near the islands of Zond (233 specimens). Most of the specimens were sexually mature. Allgen has recorded the occurrence of one female of this species from the coast of Norway. *Anticomopsis typicus* was also recorded for the first time by Micoletzky near the islands of Zond. Luc and de Coninck (1959) discovered a male and female of this species in LaManshe. Wieser (1953c) recorded *Pseudocella kreisi* for the first time on the coast of Chile (one specimen which was sexually immature). Later (1956), he found one male off Sri Lanka (Ceylon) at a depth of 3,400 m. *Thoracostoma steineri* was described by Micoletzky (1922) on the basis of one female specimen found in the Red Sea. Later (1930) he found a significant number of sexually mature specimens of this species on the islands of Zond. Allgen found this species in the Mediterranean Sea and on the coast of California. Schuurmans-Stekhoven (1943, 1950) also discovered it in the Mediterranean Sea but his description differs significantly from that given by Micoletzky.

The material presented, it seems to me, bears testimony to the fact that free-living marine nematodes are not universally (ubiquitously) distributed organisms. Each zoogeographic region corresponds strictly to a well-defined group of species. Most of the data about the extremely wide distribution of species reflects an inadequate study of their systematics.

Studies of the zoogeographic characteristics of genera of lower enoplids are almost impossible today. All the genera, irrespective of species composition, are transoceanic and only some, mainly monotypic ones, have an affinity for definite regions.

In conclusion, it must be stated that the main problem in the analysis of the geographic distribution of nematodes is inadequate study of their fauna in most regions of the world. An accumulation of faunistic information in the future should significantly elucidate their zoogeographic analysis.

COLLECTION AND PROCESSING OF NEMATODES

Collecting nematodes is a very simple but laborious process. Information on this procedure is available in the works of Filip'ev (1926b) and

Goodey (1959). In the littoral zone samples are collected manually from the benthos and algae at rather shallow depths with the help of dredges operated from a boat. In the sublittoral zone benthos is scooped up by standard equipment used for such purposes, i.e., various types of trawls and dredges. Extracting nematodes from the seabed is not so easy. The method of "thorough-wash" is preferable for obtaining nematodes in a more concentrated mass. In this method the probe situated in Koch's cup is shaken vigorously and the contents poured into another container. Only larger, coarse particles of the benthos remain at the bottom of Koch's cup. This operation is repeated several times. The poured-off water contains nematodes and fine particles of benthos. It is filtered through a sieve with a fine mesh (No. 69) to remove the fine particles of silt. The benthos remaining on the sieve and containing a large number of nematodes is examined under a binocular microscope in Bogorov's chamber. Worms thus isolated are preserved in 70° alcohol or 4% formalin.

55 When nematodes are collected from algae the latter is shredded and separated in a large glass container. The material is then filtered through a sieve of large mesh to separate the algal fragments. The filtrate, containing small fragments of algae and nematodes, is subsequently passed through a sieve of fine mesh (No. 69). The remaining mass is either preserved as a whole or subjected to selection; nematodes freed from foreign particles and floating in clean water are preserved.

To make preparations of nematodes the worms are transferred from the preservation medium to a mixture of equal volumes of 96° alcohol, distilled water, and glycerine, and kept there for 3 to 12 days depending on the size and transparency of the nematodes. In this mixture the nematodes become transparent, after which they are transferred to pure glycerine for 12 to 20 hours. Nematodes processed in this manner are mounted in a glycerine-gelatin medium.

Systematics

Certain clarifications are necessary before proceeding with the taxonomic classification of Enoplida. First of all, the order Enoplida is divided into two superfamilies—Enoploidea and Leptosomatoidea. A characterization of each superfamily is presented, followed by a key to families which, in addition to serving the purpose of identification, gives some idea about the size of each superfamily. This information is essential to my later detailed discussion of the four families—two from each superfamily.

Species earlier assigned to just family Leptosomatidae and recorded from marine bodies of the Soviet Union are described herein and redistributed in four families—Leptosomatidae, Anticomidae, Rhabdodemaniidae, and Crenopharingidae. Species present in my material are not only described but depicted in original illustrations. All the measurements of body parts are based on my own material. For species not represented in my material but described by other authors from these seas, I have presented the published descriptions which, because they vary in style, could not be fitted into the scheme adopted for describing species at my disposal. Descriptions taken from literature are asterisked. Species encountered in the seas of the Soviet Union are numbered in the keys.

1. Order ENOPLIDA Schuurmans-Stekhoven and Coninck, 1933

Free-living nematodes of rather large size, complete in organization, and subject to less reduction (Filip'ev 1918, p. 37). Characters: cuticle almost always smooth, but nematodes of the family Lauratonematidae with annulations constitute an exception. Oral opening encircled by six papillae (occasionally setae); another circle, consisting of ten papillae, or two circles, one consisting of six and the other of four setae (occasionally papillae) situated on head. Occasionally setae of first circle totally reduced. Amphids cyathiform with transverse slit either elliptical or longitudinally elongated. Esophagus almost cylindrical, sometimes gradually broadening toward base. Esophageal glands often open sublaterally in lower part of oral cavity. Esophagi characteristic of the order either fuse with the cuticle anteriorly or show signs of an earlier attachment, disappearing subsequently due to powerful development of the oral cavity. Oral cavity later loses its capacity to extrude due to weak development and differentiation of muscles of anterior end of esophagus. Structure of oral cavity highly variable. Genital tubes mostly paired; family Lauratonematidae exceptional in which females have reproductive tubes with a reduced posterior part.

Key to Superfamilies of Order Enoplida

- 56 1 (2). Esophagus with wavy margin in posterior part, rarely cylindrical. Oral cavity sharply divided into two parts: buccal cavity with jaws or buccal stylets or simply lined with sclerotized cuticle, and onchial cavity often armed with onchia. Esophageal musculature extends anteriorly only up to anterior margin of onchial cavity.

-2. **Enoploidea** Baird, 1853.
 2 (1). Esophagus cylindrical and without wavy margin. Oral cavity either not expressed or without sharp division into buccal and onchial parts. Onchia, if present, displaced toward oral opening due to protrusibility of oral musculature almost up to oral opening.
1. **Leptosomatoidea** Filipjev, 1918.

1. Superfamily **LEPTOSOMATOIDEA** Filipjev, 1918

Sclerotized cephalic capsule well developed in many representatives. Oral cavity either completely reduced or with a simple structure and not divided into buccal and onchial parts. Additionally, its armature never represented by jaws or buccal stylets. Of the derivatives of the buccal cavity, only development of odontia (*Cylicolaimus* for example) possible. More often in forms with oral armature, odontia represented by onchia (derivatives of onchial cavity), which are notably shifted toward oral opening due to protrusibility of esophageal musculature. Esophagus widens gradually from anterior end toward base; alveolate structure never observed in posterior half. Cuticle covering the body thick, smooth, and often with fascicles of intersecting fibers in external layer. Amphids in most forms round or cyathiform. Some representatives of Oxystominidae have extremely narrow, and longitudinally elongated amphids.

Key to Families of Superfamily Leptosomatoidea

- 1 (8). Cephalic setae (10 in number) situated in single circle. Amphids usually cyathiform.
 2 (7). Cephalic capsule either poorly or well developed; oral cavity cyathiform, sometimes extremely short or not developed at all.
 3 (4). Cephalic capsule very poorly developed; oral cavity armed with three onchia which resemble mandibles.
 **Triodontolaimidae** de Coninck, 1965.
 [Only species: **Triodontolaimus acutus** (Villot, 1875)].
 4 (3). Oral cavity unarmed or its armature represented by typical onchia.
 5 (6). Cephalic capsule either absent or so poorly developed as to be undetectable. Body tapers distinctly toward both ends.
2. **Anticomidae** Filipjev, 1918.
 6 (5). Cephalic capsule always present and represented by powerful sclerotized structure in a number of forms. Body tapers slightly toward anterior end but negligibly toward posterior end.
1. **Leptosomatidae** Filipjev, 1916.
 7 (2). Cephalic capsule absent; oral cavity in the shape of a long, narrow cylinder. **Ironidae** de Man, 1876.

- 57 8 (1). Cephalic setae situated in two circles (six plus four); some setae reduced in some forms. Amphids with large openings, sometimes highly elongated in longitudinal direction.....
**Oxystominidae** Micoletzki, 1924.

1. Family LEPTOSOMATIDAE Filipjev, 1916

Large free-living marine nematodes, ranging from 10 to 50 mm in length. Cuticle thick, often multilayered. Ten cephalic setae always arranged in one circle. Mouth encircled by six papillae. Some genera with eyes or spots of photosensitive pigment. Esophagus straight, without bulbs. Cardia large, triangular. Most characteristic features of family: presence of sclerotized cephalic capsule with various degrees of complexity evident in endocuticle. More or less developed oral cavity armed with onchia or odontia in a number of forms. Gonads always paired and no sign of their reduction ever observed. Females with paired gonads, genital ducts, and uteri. Only testes paired in males, which later merge into an unpaired vas deferens and ejaculatory duct. Spicular apparatus of males highly diverse in structure but always consists of a pair of spicules and a gubernaculum. Gubernaculum with paired or unpaired processes. Most species of this family have an accessory organ, anal setae, and papillae. Caudal glands three, tubular or fusiform, and almost always open terminally at the tip of the tail.

Key to Subfamilies of Family Leptosomatidae

- 1 (4). Cephalic capsule highly developed, thick-walled.
 2 (3). Interlobular grooves of cephalic capsule differentiated into narrow furrows and wide fenestrae; lateral grooves differ in shape from the rest. Tail short and bluntly rounded.....
3. **Thoracostominae** de Coninck, 1965.
 3 (2). Interlobular grooves of cephalic capsule not differentiated and uniform in shape. Tail elongated and claviform.....
2. **Synonchinae** Platonova, 1970.
 4 (1). Cephalic capsule poorly developed, thin-walled.
 5 (6). Oral cavity highly developed, thick-walled, and armed with onchia.
**Cylicolaiminae** Platonova, 1970.
 6 (5). Oral cavity not developed.....1. **Leptosomatinae** Filipjev, 1916.

1. Subfamily LEPTOSOMATINAE Filipjev, 1916

Nematodes with poorly developed, short cephalic capsule. Cephalic suture straight or slightly wavy and without interlobular grooves. *Lepto-*

somatides conisetosum, with a cephalic suture with a deep groove, constitutes an exception.¹ Amphids always situated below line of cephalic suture. Cephalic ring narrow. Oral cavity poorly developed, almost absent. Esophagus encircled by nerve ring between anterior third and fourth part. Sclerotized granules sometimes present around the vulval slit on cuticular surface. Tail short and bluntly rounded in most representatives of family. Cephalic and preneural setae extremely short; latter sometimes absent. Posterior body end in males may or may not be armed with setae. Well-developed photosensitive eyes present in most representatives of subfamily.

Key to Genera of Subfamily Leptosomatinae

- 1 (2). Tail claviform.....3. **Leptosomella** Filipjev, 1927.
 2 (1). Tail conical, rounded, or acicular.
 3 (6). Eyes present.
 4 (5). Gubernaculum small, without processes.....
1. **Leptosomatum** Bastian, 1865.
 5 (4). Gubernaculum large, with large dorsal process.....
2. **Leptosomatides** Filipjev, 1918.
 6 (3). Eyes absent.
 7 (8). Spicules with velum. Gubernaculum with ventral and dorsal processes.....**Paraleptosomatides** Mawson, 1956.
 (Two species: **P. spiralis** Mawson, 1956
 and **P. elongatus** Mawson, 1956.)
 8 (7). Spicules without velum. Gubernaculum with dorsal process only.
**Leptosomatina** Allgen, 1951.
 (Two species: **L. longisetum** Allgen, 1951
 and **L. appendixocaudatum** Allgen, 1957.)

1. Genus *Leptosomatum* Bastian, 1865

Bastian, 1865: 144; Eberth, 1863: 18 (*Phanoglene* non Nordmann, 1841); de Man, 1893: 102; Filip'ev, 1961: 65; 1918: 42.

Type species. *L. elongatum* Bastian, 1865.

Nematodes of large size. Body tapers slightly toward anterior end and almost not at all posteriorly. Head slightly truncated. Cephalic capsule, though poorly developed, with walls which may vary significantly in width. Cephalic ring varies in position within limits of upper half of

¹It is possible that this species will be transferred in the future to another genus or even to another subfamily. Having no material at my disposal, I could not solve this problem just now.

cephalic capsule. Labial papillae distinct. Cephalic setae similar to preanal setae but occasionally rudimentary with appearance of small papillae. Anal setae absent. Amphids circular. Oral cavity poorly developed. Esophagus immediately apposed to oral opening. Lips and anterior part of esophageal wall often sclerotized and thickened. Vulva situated mid-body. Spicules simple in structure and slightly arcuate; in some species only proximal part curves sharply. Gubernaculum small and simple in structure. Most species with preanal glands but no distinct accessory organ on outlet on wall surface. Accessory organ well developed only in *L. keiense* and *L. ranjhai*; preanal papilla also present in latter species.

In the description of species of both *Leptosomatum* and *Leptosomatides* figures in Cobb's formula indicate the distances of the following parts of the body from the anterior end: 1. posterior end of cephalic capsule; 2. pigmented eyes; 3. nerve ring; 4. base of esophagus; 5. commencement of anterior genital tube; 6. vulva; 7. commencement of posterior genital tube; and 8. anus.

For the male the first four measurements are those given for the female. The dash which follows indicates the middle of the male body. The last figure indicates the distance from the anterior end to the anus. The corresponding width of the body is given below the line for those parts of the body described above.

Key to Species of Genus Leptosomatum

- 1 (38). Photosensitive eyes in preneural region.
- 2 (37). One pair of photosensitive eyes present, consisting of pigment bowl and light-refracting lens.
- 59 3 (4). Cephalic capsule very short; width exceeds length six times.8. *L. breviceps* Platonova, 1967.
- 4 (3). Cephalic capsule significantly longer; width exceeds length by not more than three times.
- 5 (12). Cephalic setae well developed.
- 6 (7). Head highly flattened.*L. australe* Linstow, 1906.
- 7 (6). Head anteriorly rounded.
- 8 (9). Amphids situated from anterior end at a distance equal to 3.5 times the cephalic diameter.*L. Micoletzky* Inglis, 1971.
- 9 (8). Amphids situated from anterior end at a distance equal to 1.0 to 1.5 times the cephalic diameter.
- 10 (11). Width of amphids equal to one-third corresponding body width.6. *L. punctatum* (Eberth, 1863).
- 11 (10). Width of amphids equal to one-fourth corresponding body width.*L. keiense* Micoletzky, 1930.

- 12 (5). Cephalic setae either rudimentary, resembling papillae, or more often absent.
- 13 (18). Males with accessory organ.
- 14 (15). Cuticular rays present on cephalic capsule.....
.....**L. ranjhai** Timm, 1960.
- 15 (14). Cuticular rays absent on cephalic capsule.
- 16 (17). Lens-shaped broadening occurs in anterior part of esophagus.
.....4. **L. bacillatum** (Eberth, 1863).
- 17 (16). Lens-shaped broadening does not occur in anterior part of esophagus.....9. **L. gracile** Bastian, 1865.
- 18 (13). Males without accessory organ.
- 19 (26). Cephalic setae rudimentary and do not project on surface of cuticle.
- 20 (21). Cephalic setae pierce cuticle and form a coronet on cephalic capsule.....**L. crassicutis** Platonova, 1958.
- 21 (20). Coronet on cephalic capsule absent.
- 22 (23). Head broadens and appears clavate in region of cephalic setae . .
.....**L. clavatum** Platonova, 1958.
- 23 (22). Broadening of head in cephalic region not evident.
- 24 (25). Amphids cordate.....**L. kerguelense** Platonova, 1958.
- 25 (24). Amphids transversely oval in shape.....
.....**L. sabangense** (Steiner, 1915).
- 26 (19). Cephalic setae short but nonetheless project on cuticular surface.
- 27 (28). Tail extremely short; length does not exceed three-fourths of anal diameter.....**L. pedroense** Allgen, 1947.
- 28 (27). Tail length exceeds anal diameter.
- 29 (30). Body diameter near eyes equal to 2.5 times cephalic diameter . .
.....3. **L. behringicum** Filipjev, 1916.
- 30 (29). Body diameter equal to twice cephalic diameter.
- 31 (34). Eyes situated at a distance twice cephalic diameter from anterior end.
- 32 (33). Lateral organs equal to one-sixth cephalic diameter from anterior end.....2. **L. grebnickii** Filipjev, 1916.
- 33 (32). Lateral organs equal to one-tenth cephalic diameter from anterior end.....1. **L. arcticum** Filipjev, 1916.
- 34 (31). Eyes situated at a distance thrice cephalic diameter from anterior end.
- 35 (36). Spicular capitulum demarcated from body of spicule by a distinct necklike constriction.....5. **L. elongatum** Bastian, 1865.
- 60 36 (35). Spicules with no necklike constriction.....
.....**L. acephalatum** Chitwood, 1936.
- 37 (2). Two pairs of photosensitive eyes present; on pair consists of pig-

- ment bowl and light-refracting lens and second pair simple collection of pigment 7. *L. tetrophtalmum* Saveljev, 1912.
- 38 (1). Photosensitive eyes absent.
- 39 (40). Cephalic setae extremely long; length equal to one-half cephalic diameter *L. caecum* Ditlevsen, 1923.
- 40 (39). Cephalic setae short.
- 41 (42). Esophagus granulated in anterior part *L. bathibium* Allgen, 1947.
- 42 (41). Esophagus not granulated.
- 43 (44). Width of amphid one-sixteenth corresponding cephalic diameter. *L. abissale* Allgen, 1915.
- 44 (43). Width of amphid one-third corresponding cephalic diameter. *L. groenlandicum* Allgen, 1954.

1. *Leptosomatium arcticum* Filipjev, 1916 (Figure 14)

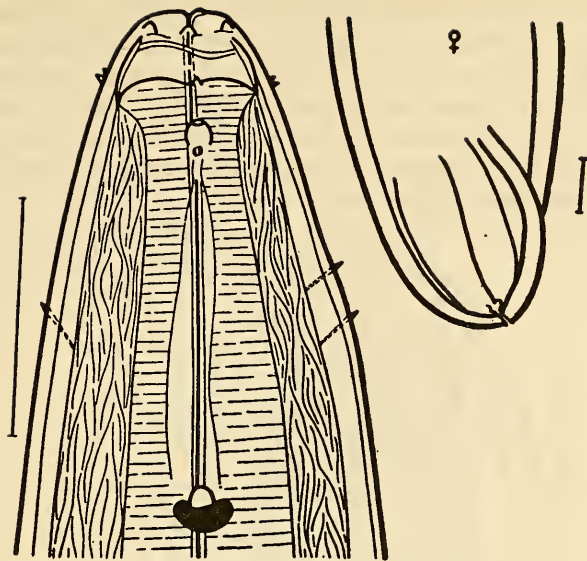
Filip'ev, 1916: 66-68, tab IV, fig. 1; Mawson 1958b: 315, fig. la-c.

$$1 \text{ ♀: } \frac{20}{52} \frac{120}{73} \frac{335}{140} \frac{1,275}{170} \frac{6,200}{-} \frac{7,550}{210} \frac{9,100}{-} \frac{10,725}{150} 10,900 \mu\text{m}; a = 52; \\ b = 9; c = 62; V = 69\%.$$

Body elongated, fairly thick, and tapers to almost 1/4 midbody diameter at anterior end and to 5/7 midbody diameter at posterior end. Width of cephalic capsule 2.6 its length. Cephalic ring situated midlength of cephalic capsule. Cephalic suture slightly sinuous. Labial papillae rather short. Cephalic setae, like cervical ones, extremely short; length does not exceed 4.0 μm (1/13 corresponding diameter). Amphids 7.0 μm in diameter and situated 30 μm from anterior end (1/40 esophageal length). Eyes situated 120 μm from anterior end (1/10 esophageal length).

- 61 Size of pigment bowl 5.0 $\mu\text{m} \times 9.0 \mu\text{m}$; diameter of lens 7.0 μm . Esophageal wall near mouth highly sclerotized. Esophagus thin and widens slightly in posterior part. Nerve ring situated 335 μm from anterior end (approximately 1/4 esophageal length). Vagina thick-walled. Sclerotized granules scattered on cuticle around vulva. Genital tubes not detected.

Filip'ev, who described the species, mentions the following characters which distinguish it from other species: 1) *L. elongatum*—small size, small amphids, and better developed cephalic capsule; 2) *L. bacillatum*—twice larger but has smaller amphids and eyes; 3) *L. tetrophtalmum*—absence of additional pair of eyes situated behind original eyes and more submedian position of vulva; 4) *L. grebnickii*—structure of cephalic capsule and much larger amphids; 5) *L. behringicum*—significantly better developed cephalic capsule, much larger amphids, and eyes and setal armature slightly reduced.



60

Figure 14. *Leptosomatum arcticum*.²

Geographic distribution. Barents Sea (littoral and sublittoral zones up to a depth of 5.0 m, among *Lithothamnion* sp. and *Laminaria* sp.). Occurrence in subantarctic (Mawson, 1958b) dubious.

2. *Leptosomatum grebnickii* Filipjev, 1916 (Figure 15)

Filip'ev, 1916: 68-70, tab. IV, fig. 2.

Holotype ♀: Zoological Institute, Academy of Sciences, USSR. Collection nos. 5778, 5779.

| | | | | | | | | |
|----|-----|-----|-------|-------|-------|--------|--------|---------------------------|
| 15 | 80 | 356 | 1,385 | 5,200 | 7,750 | 11,500 | 11,910 | 12,040 μm; a = 65; b = 9; |
| 60 | 100 | 107 | 124 | — | 185 | — | 120 | |

c = 92; V = 64%.

Body large and tapers to 1/5 midbody diameter at anterior end and to 5/7 midbody diameter at posterior end. Caudal length equal to width. Cuticle 7.0 μm thick, with fascicles of intersecting fibers. Cephalic capsule better developed than in *L. arcticum*; width 2.2 length. Cephalic ring situated in middle of cephalic capsule and narrower than in *L. arcticum*. Cephalic suture barely visible. Labial papillae larger and wider than in *L. arcticum*. Cephalic and cervical setae short, 5.0 μm (1/12 corresponding diameter). Amphids situated 30 μm from anterior end (1/4 esophageal

²Scale for Figures 14 to 86 corresponds to 50 μm.

length); diameter of amphids $9.0 \mu\text{m}$ ($1/6$ corresponding body diameter). Eyes $80 \mu\text{m}$ from anterior end ($1/17$ esophageal length). Pigment bowl $8.0 \mu\text{m} \times 12.0 \mu\text{m}$; diameter of lens $9.0 \mu\text{m}$. Esophageal wall near oral opening greatly thickened. Nerve ring at a distance of $365 \mu\text{m}$ from anterior end ($1/4$ length of esophagus). Esophagus long, narrow, and widens slightly toward posterior end. Sclerotized granules scattered around vulva on cuticle. Genital tubes not detected.

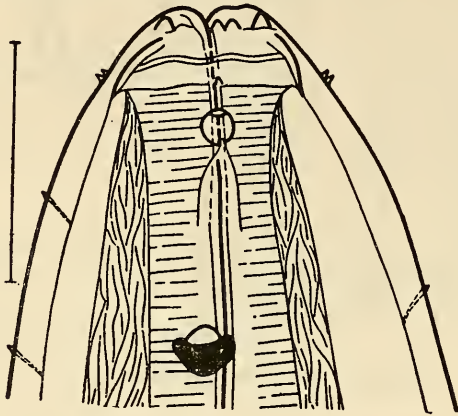


Figure 15. *Leptosomatum grebnickii*.

Very close to *L. arcticum* in structure, but differs from latter in wider, 62 shorter, and significantly less sclerotized cephalic capsule; significantly narrower cephalic ring; and much larger amphids.

Geographic distribution. Barents Sea.

3. *Leptosomatum behringicum* Filipjev, 1916 (Figure 16)

Filip'ev, 1916: 70-72.

Holotype ♀: Zoological Institute, Academy of Sciences, USSR. Collection nos. 5780, 5781.

| | | | | | | | | |
|----|-----|-----|-------|-------|-------|-------|-------|--------------------------------------|
| 80 | 82 | 295 | 1,100 | 2,650 | 5,300 | 8,700 | 9,673 | 9,750 μm ; a = 57; b = 9; |
| 50 | 100 | 115 | 145 | — | 170 | — | 80 | |

c = 126; V = 56%.

Body tapers to $5/28$ midbody diameter at anterior end and to $1/2$ midbody diameter at posterior end. Tail extremely short; length almost equal to width in anal region. Cuticle $4.0 \mu\text{m}$ thick, with fascicles of intersecting fibers. Cephalic capsule extremely short and wide; width exceeds length by 3.7 times. Cephalic ring very narrow and situated in upper part of cephalic capsule. Cephalic suture slightly sinuous. Labial papillae extremely minute. Cephalic setae also short, $2.0 \mu\text{m}$ long ($1/25$

corresponding diameter). Cervical setae not detected. Amphids situated $25\ \mu\text{m}$ from anterior end ($1/40$ esophageal length). Diameter of amphids $5.0\ \mu\text{m}$ ($1/9$ corresponding body diameter). Eyes situated $82\ \mu\text{m}$ from anterior end of body ($1/13$ esophageal length). Pigment bowl small, only $5.0\ \mu\text{m} \times 8.0\ \mu\text{m}$; diameter of lens $5.0\ \mu\text{m}$. Esophageal wall sclerotized in oral region. Esophagus slender and broadens slightly in posterior part. Nerve ring situated $295\ \mu\text{m}$ from anterior end ($1/4$ esophageal length). Vulva situated midbody. Eggs spherical and 150 to $160\ \mu\text{m}$ in diameter. Genital tubes not detected.

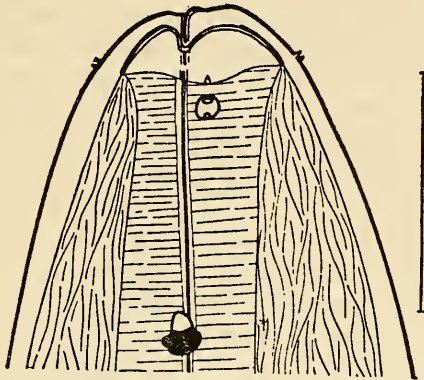


Figure 16. *Leptosomatum behringicum*.

Structurally close to *L. grebnickii* but distinguished from it in: 1) more weakly developed labial papillae; 2) more weakly developed setaceous armature; 3) position of cephalic ring situated in upper part of cephalic capsule in present species; 4) smaller size of amphids; and 5) smaller size of eyes.

Geographic distribution. Barents Sea.

4. *Leptosomatum bacillatum* (Eberth, 1863) (Figure 17)

Eberth, 1863: 19, 20, tab. II, figs. 1 to 4 (*Phanoglene*); Bastian, 1865: 146; Marion, 1870: 17, 18, pl. C., fig. 2 (*Stenolaimus macrosoma*); de Man, 1876: 103, tab. 8, fig. 9a, b; Filip'ev, 1918: 44-48, tab. 1, fig. 1; Allgen, 1940a: 488, Fig. 1a, b; Schuurmans-Stekhoven, 1950: 25-28, fig. 1a, b.

| | | | | | | | |
|-------|---|-------|---------|-------------|-------------|-------------|-------------|
| 2 ♂: | 11-13 | 73-75 | 280-285 | 1,090-1,110 | — | 9,207-9,260 | 9,300-9,355 |
| | 32 | 50-57 | 80-82 | 95-100 | 120 | 80-82 | |
| | μm ; a = 78-79; b = 8-9; c = 98-100. | | | | | | |
| 15 ♀: | 9-12 | 75-80 | 235-240 | 1,030-1,045 | 4,050-4,200 | 5,200-5,350 | × |
| | 27-30 | 55-60 | 70-80 | 95-100 | — | 120 | |

$$\times \frac{6,350-6,400}{-} \frac{8,522-8,620}{80-85} 8,600-8,700 \mu\text{m}; a=69-74; b=8-9; \\ c=110-145; V=60\%.$$

- 63 Body tapers to 1/4 midbody diameter at anterior end and 2/3 at posterior end. Head rounded. Tail length almost equal to width. Cuticle bilayered and 5.0 to 6.0 μm thick; thickness of exterior layer 1.0 μm . Fascicles of intersecting fibers extend into outer layer of cuticle. Cephalic capsule thick-walled; width three times length. Cephalic ring situated in uppermost part of cephalic capsule. Cephalic suture and labial papillae barely discernible. Cephalic and cervical setae extremely small, not exceeding 1.5 μm (1/20 corresponding diameter). Amphids situated 18 to 25 μm from anterior end (1/50 esophageal length) and 8.9 $\mu\text{m} \times 7.5 \mu\text{m}$ (1/4 to 1/5 corresponding body diameter). Diameter of amphid opening 2.5 μm . Eyes situated 73 to 80 μm from anterior end (1/13 to 1/14 esophageal length). Pigment bowl 12.0 $\mu\text{m} \times 10.0 \mu\text{m}$; diameter of lens 5.0 μm . Esophageal walls thick in this region. Two grooves with thick sclerotized walls form lens-shaped dilatations 15 to 18 μm from anterior end in wall of esophagus. Nerve ring situated 235 to 285 μm from anterior end (1/4 esophageal length). Esophagus and genital tubes usual in size for such structures. Uterus occupies 2/3 of entire genital tube. Eggs 200 to 250 $\mu\text{m} \times 70$ to 80 μm . Spicules straight, with rectangular head curved at an angle, and 90 μm long. Length of gubernaculum 25 μm . Accessory organ and anal papillae present but barely visible in my preparations.

The important characters distinguishing this species from others in the genus are: 1) lens-shaped expansions in anterior part of esophagus; and 2) straight spicules with almost rectangular head curved at an angle to the spicular body.

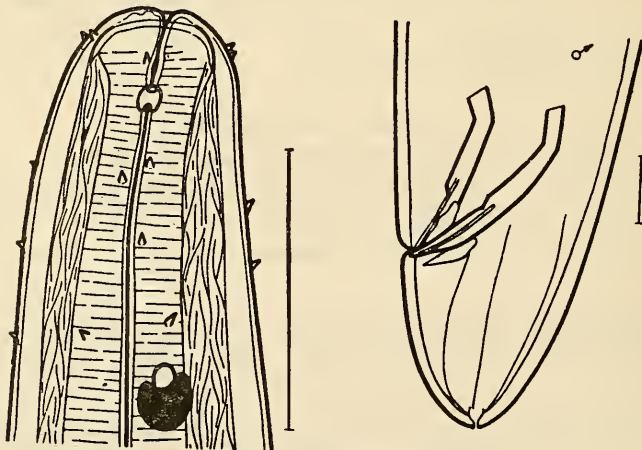


Figure 17. *Leptosomatum bacillatum*.

Geographic distribution. Black Sea (along the coastal strip among algae and on banks of mussels in silty and sandy benthos in a depth range of 10 to 96 m) and Mediterranean and Red Seas.

The distribution of this species given by Allgen is dubious. It is difficult to explain its presence on the coast of Norway (1940a), the Pacific coast of Panama and California (1947c), the Hawaiian Islands (1951), and all along the coast of Argentina right up to Terra del Fuego (1959). The occurrence of this species in the subantarctic and on the coast of Norway are particularly questionable, especially since the author had only one female specimen for Norway. It seems to me that another species of this genus was involved. Identification of females of species of this genus is a very difficult task.

64 5. *Leptosomatium elongatum* Bastian, 1865 (Figure 18)

Bastian, 1865: 145, pl. XII, figs. 156, 157; de Man, 1893: 103-107, pl. VI, fig. 9.

| | | | | | | |
|--------|---|---------|---------|-------------------|-------------|------------------|
| 4 ♀: | 21-22 | 104-109 | 545-566 | 1,957-2,111 | 5,819-6,386 | |
| | 23-25 | 46-48 | 75-78 | 116-127 | 137-170 | 162-182 |
| | 7,673-8,137 9,424-9,785 10,850-12,360 | | | | | |
| × | 170-200 | 167-182 | 135-137 | 11,000-12,535 μm; | | |
| | a=61-74; b=6; c=72-81; V=63-65%. | | | | | |
| 7 juv. | 11-18 | 18-31 | 86-104 | 375-463 | — | 4,752-8,291 |
| | 14-23 | 31-42 | 34-48 | 54-73 | 77-100 | 92-127 87-100 |
| | × 4,827-8,428 μm; a-53-66; b-4-5; c-63-65. | | | | | |

Body rather long and tapers to 10/37 to 5/21 midbody diameter at anterior end and 5/6 to 5/7 at posterior end. Length of tail equal to width. Cuticle 8.0 to 9.0 μm thick, with fascicles of intersecting fibers. Cephalic capsule with rather thick walls; width at base exceeds length by 2.2 times. Cephalic ring rather massive. Cephalic setae short, 2.32 μm long (1/20 corresponding diameter). Cervical setae even shorter, 1.5 μm long. Amphids 29 to 34 μm from anterior end (1/60 esophageal length) and 7.0 μm in diameter (1/8 corresponding body diameter). Eyes situated 104 to 109 μm from anterior end (1/20 esophageal length). Pigment bowl 9.28 μm × 11.6 μm; diameter of lens 5.8 μm. Lips somewhat thick. Nerve ring situated 545 to 566 μm from anterior end (1/4 esophageal length). Length of anterior female genital tube 1,751 to 1,854 μm and reflexed part 1,030 to 1,236 μm; of posterior tube 1,648 to 1,751 μm, and reflexed part 1,030 to 1,133 μm. In one female eggs 412 μm × 133 μm were detected in each uterine branch. Vulva surrounded by sclerotized granules and situated somewhat posterior to midbody.

Regrettably there was no male of this species in my material. Hence the distinctive features of this species are restricted to the cephalic struc-

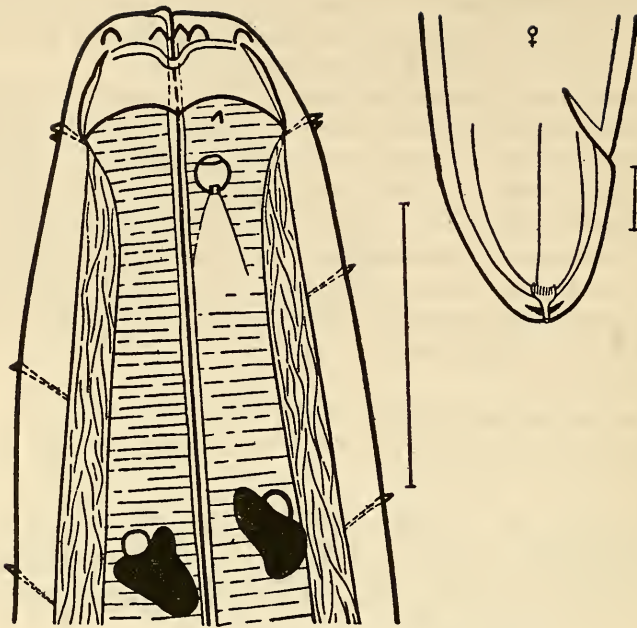


Figure 18. *Leptosomatum elongatum* Bastian.

65 ture, namely, its thick walls, cephalic ring situated in the upper part of capsule, and a distinct cephalic suture forming wide semicircular grooves separated by acicular projections.

Geographic distribution. This species in all probability is cosmopolitan. It has been found in the Barents Sea (predominantly in the littoral zone among shells or in sand), the Mediterranean Sea, coast of California, tropical part of the Indian Ocean, coast of Australia, and in the Antarctic and subantarctic.

6. *Leptosomatum punctatum* (Eberth, 1863) (Figure 19)

Eberth, 1863: 20, 21, tab. 2, figs. 5-7 (*Phanoglene*); Bastian, 1865: 145; Filip'ev, 1918: 48-50, tab. I, fig. 2.

| | | | | | | | | | |
|------|----|-----|-----|-------|-------|-------|-------|-------|--------------------------------------|
| 1 ♂: | 8 | 105 | 350 | 1,080 | — | 7,905 | | | 8,010 μm; a=80; b=7; c=77. |
| | 26 | 50 | 80 | 95 | 100 | 80 | | | |
| 1 ♀: | 10 | 100 | 360 | 1,100 | 4,006 | 4,158 | 5,437 | 8,359 | 8,470 μm; a=70; b=8; c=76; V=49%. |
| | 25 | 48 | 80 | 90 | — | 120 | — | 90 | |

Body tapers to 1/3 midbody diameter at anterior end and 10/13 at posterior end. Cuticle bilayered; outer and inner layers equal in thickness, 6.0 μm on the average. Cephalic capsule thick-walled; width exceeds

length by 3.2 times. Cephalic suture barely discernible. Cephalic ring not distinct in my preparations; it appeared narrow and was situated mid-length of the head. Labial papillae short. Cephalic and cervical setae rather long. Cephalic setae $6.0\ \mu\text{m}$ long ($1/4$ corresponding diameter); cervical setae slightly shorter, $5.0\ \mu\text{m}$ long. Amphids situated $30\ \mu\text{m}$ from anterior end ($1/30$ esophageal length); diameter of amphids $8.0\ \mu\text{m}$ ($1/3$ corresponding diameter). Eyes situated $105\ \mu\text{m}$ from anterior end ($1/10$ esophageal length). Pigment bowl $10\ \mu\text{m}$ wide; diameter of lens $4.0\ \mu\text{m}$. Anterior margin of cephalic capsule forms rectangular depression in oral region which Filip'ev (1918) called a "pocket". Esophageal wall highly sclerotized near oral opening; thickening extends up to level of amphids. Esophagus rather narrow, almost not expanding posteriorly. Male spicules approximately uniform in width almost throughout their length, narrowing only in the distal (sharply curved) part in the middle, and $65\ \mu\text{m}$ long. Gubernaculum $20\ \mu\text{m}$ long. Accessory organ situated $112\ \mu\text{m}$ anterior to anus. Anal papillae absent.

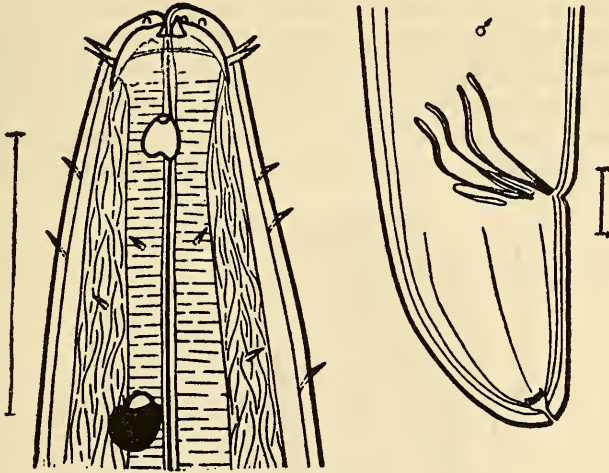


Figure 19. *Leptosomatum punctatum*.

Close to *L. bacillatum*, but differs in the following characters: 1) cephalic setae much longer, constituting $1/4$ corresponding cephalic diameter versus $1/20$ in *L. bacillatum*; 2) cuticular pockets present lateral to oral aperture in *L. punctatum*; and 3) spicules curved in middle portion (spicules straight in *L. bacillatum*).

Geographic distribution. Found mainly in the Black Sea in silt and among shells in a depth range of 40 to 100 m, and in algae of the coastal belt of the Mediterranean Sea.

7. *Leptosomatum tetrophtalmum* Saveljev, 1912 (Figure 20)
Savel'ev, 1912: 124.

| | | | | | |
|-------|--|---------------|---------------|-------------|-------------|
| 15 ♀: | 23-29 | 104-116 | 515 | 2,060-2,163 | 7,004-6,901 |
| | 23-25 | 52-55 | 81-92 | 125-137 | 175-207 |
| | 10,094-10,300 | 12,772-13,184 | 15,144-15,759 | | |
| × | 234-265 | 204-234 | 140-150 | | |
| | 15,306-15,934 μm; | | | | |
| | a = 58-73; b = 7-8; c = 91-94; V = 61-65%. | | | | |

Body very long and tapers to 1/4 midbody diameter at anterior end and 2/3 at posterior end. Caudal length slightly exceeds width. Cuticle thick, 9.3 to 10.4 μm, with fascicles of intersecting fibers. Cephalic capsule well developed but its wall not very thick. Cephalic suture distinctly visible and slightly sinuous. Width of cephalic capsule exceeds length by 1.8 times. Thin cephalic ring situated in anterior third of cephalic capsule. Labial papillae rather large, distinct. Cephalic setae short, 3.5 μm long (1/15 corresponding diameter). Cervical setae 2.0 μm long. Amphids with very thick wall, situated 23 to 34 μm from anterior end of body (equal to 1/60 to 1/90 esophageal length), spherical, and 7.0 μm in diameter (1/9 corresponding body diameter). Eyes situated 104 to 116 μm from anterior end (1/18 to 1/20 esophageal length). Pigment bowl 8.1 to 9.2 μm × 11.6 μm; diameter of lens 5.8 to 6.9 μm. Two more pigment spots situated posterior to original eyes. Oral cavity absent. Esophageal wall highly sclerotized in oral region. Nerve ring situated 515 μm from

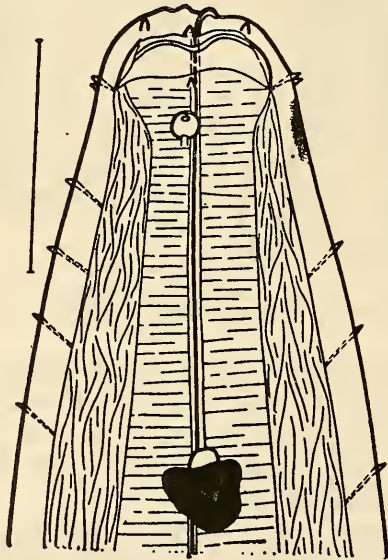


Figure 20. *Leptosomatum tetrophtalmum*.

anterior end (1/4 esophageal length). Length of anterior female genital tube 3,090 to 3,399 μm , and of posterior tube 2,678 to 2,884 μm ; respective lengths of reflexed parts 1,751 to 1,854 and 1,339 to 1,442 μm . Sclerotized granules occur around vulva on cuticle. Four to eight eggs 154 to 206 $\mu\text{m} \times 154$ to 185 μm found in each branch of uterus.

Distinguishing characters of this species: 1) large size; 2) presence of short setae; 3) thick-walled amphids, spherical in shape; and 4) presence of additional pigment spots without lens situated behind main eyes. The last feature is the most important from a diagnostic point of view.

Geographic distribution. Limited to the Barents Sea. Found in Lake Mogil'noe, Kola Bay, and on the shores of Novaya Zemlya.

8. *Leptosomatium breviceps* Platonova, 1967 (Figure 21)

Platonova, 1967: 829, fig. 1.

$$1 \text{ } \text{f} : \frac{6 \quad 87 \quad 382 \quad 1,184 \quad - \quad 4,377-7,779}{17-34 \quad 104 \quad 112 \quad 125 \quad - \quad 104-105} \quad 7,879 \text{ } \mu\text{m}; \quad a=56; \quad b=7; \\ c=79; \quad V=56\%.$$

Body tapers to 1/4 midbody diameter at anterior end and 10/13 at posterior end. Tail length equal to width. Cuticle 6.7 μm thick. Cephalic capsule extremely short; width exceeds length six times. Wall of cephalic capsule rather thin. Narrow cephalic ring passes medially through cephalic capsule. Cephalic suture straight. Labial papillae distinct; cephalic setae much longer than cervical ones. Length of cephalic setae 5.8 μm (1/6 corresponding diameter), and that of cervical setae 1.2 μm . Amphids situated 23.2 μm from anterior end (1/50 esophageal length).

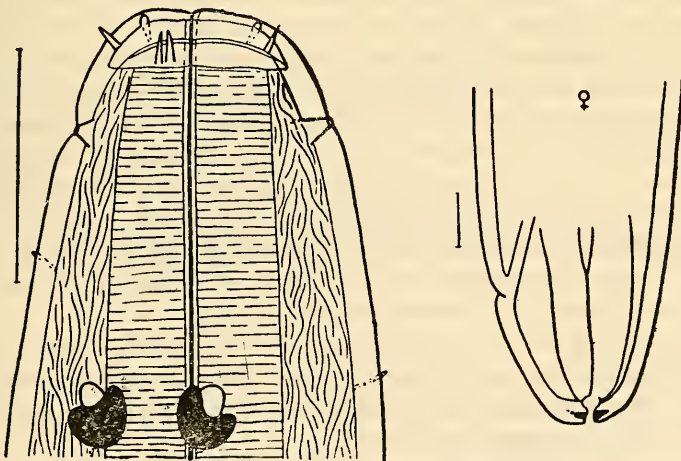


Figure 21. *Leptosomatium breviceps*.

Eyes situated $76 \mu\text{m}$ from anterior end ($1/13$ esophageal length). Pigment bowl $13.9 \mu\text{m} \times 12.7 \mu\text{m}$; diameter of lens $5.8 \mu\text{m}$. Nerve ring situated $382 \mu\text{m}$ from anterior end ($1/3$ esophageal length). Vulva situated at midbody. Female specimen examined was sexually immature and hence tubes not visible.

Distinguishing features of this species: 1) extremely short cephalic capsule (width six times greater than length; 2) medial situation of cephalic ring in cephalic capsule; 3) rather long cephalic setae; and 4) short cervical setae, $1/5$ length of cephalic setae.

Geographic distribution. This species has been found only in Kol'sk Gulf of the Barents Sea among *Lithothamnion* sp.

*9. *Leptosomatum gracile* Bastian, 1856 (Figure 22)

Bastian, 1865; 145; pl. XII, figs. 158–160; Steiner, 1916: 610–620, tab. 16, fig. 27c, d; tab. 29, fig. 27a, b, e–g; tab. 30, fig. 27h–o (description).

1 ♀: L = 13.3 mm; a = 68; b = 6; c = 74.

Cephalic capsule very short and thin-walled. Cephalic suture at level of cephalic papillae. Labial papillae distinct and usually situated in a group of six somewhat posterior to oral opening. Cephalic setae transform into papillae. Contrary to all other representatives of this family in which the cephalic setae or papillae substituted for them are always ten in number and situated in one circle, in *L. gracile* only six cephalic papillae were detected by Steiner. Neither papillae nor setae occur on the body. Amphids large, situated a short distance posterior to lateral cephalic setae, and with notably thickened anterior wall. In addition to well-developed eyes with lenses, pigment spots of irregular shape also occur near the pigment bowls. Of the three lips encircling oral opening, dorsal one highly thickened, sclerotized, and markedly projects above the other two. Esophageal wall also sclerotized. Nerve ring encircles esophagus at border of its anterior third. Vulval opening encircled by rather thick sclerotized lips. Cuticle around lips covered with minute sclerotized granules.

Despite the very brief description given by Bastian, Steiner noted the fallacy of making *L. gracile* synonymous with *L. elongatum* Bastian as done by de Man (1893). After studying specimens from the Barents Sea, Steiner arrived at the conclusion that species *L. elongatum* and *L. gracile*, although very similar, are nevertheless not identical.

Distinguishing features of this species: 1) cephalic capsule more weakly developed than in *L. elongatum*; 2) six cephalic papillae versus ten setae in *L. elongatum*; and 3) in addition to photosensitive eyes, irregular pigment spots also present in *L. gracile*.

Geographic distribution. Barents Sea and English Channel (shores of England).

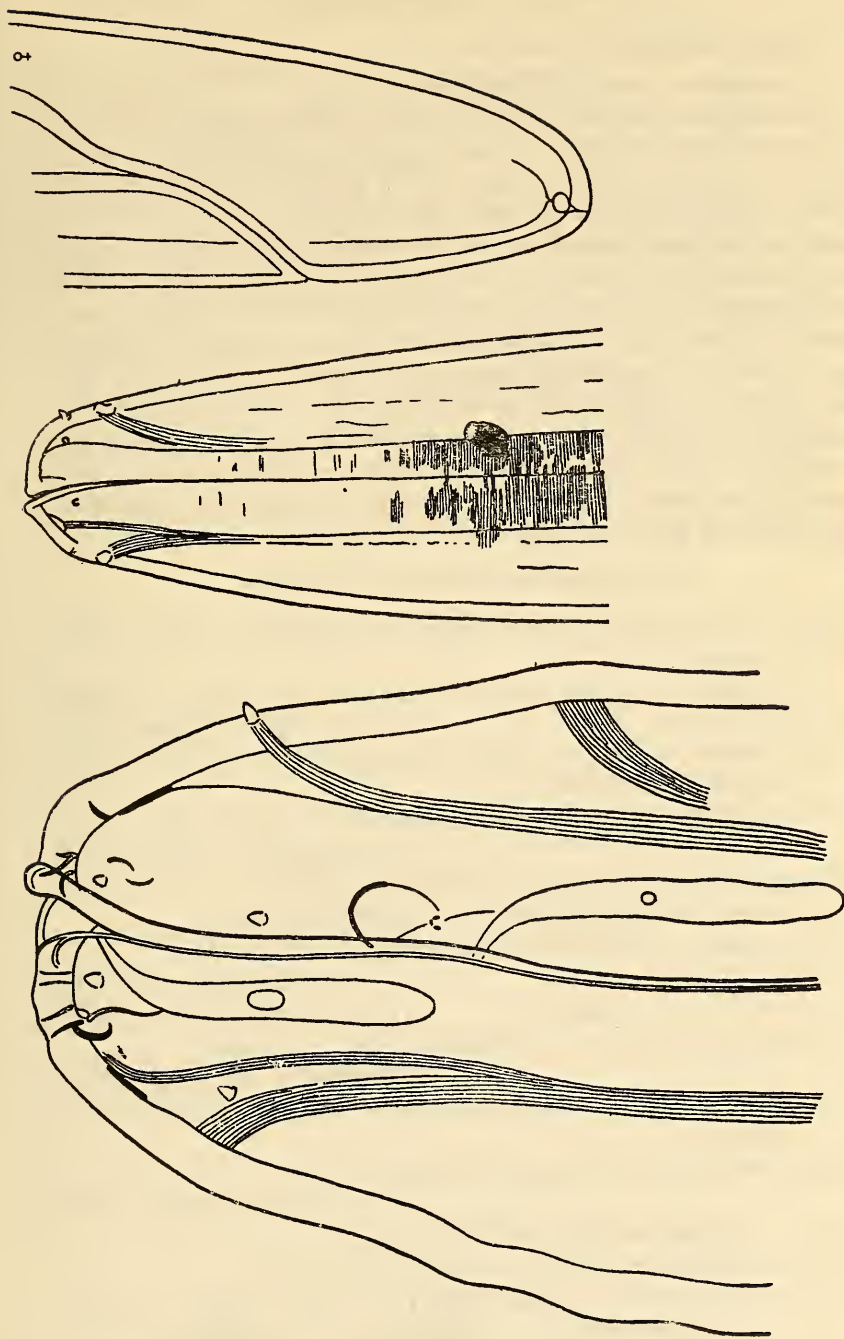


Figure 22. *Leptosomatium gracile* (from Steiner, 1916).

2. Genus *Leptosomatides* Filipjev, 1918

Filip'ev, 1918: 50, 51; 1922a: 98.

Type species: *L. euxina* Filipjev, 1918.

Nematodes of this genus highly resemble species of *Leptosomatum* in size, shape, and structure of cephalic capsule. Labial papillae distinctly visible, sometimes pointed, reminiscent of setae but usually short and blunt. Cephalic and cervical setae generally short. Unlike species of *Leptosomatum*, anal setae or papillae or sometimes both present in males of *Leptosomatides*. Amphids usually round and often taper toward posterior end. Oral cavity poorly developed. Vulva displaced posterior to midbody. Each spicule divided into two more or less equal parts—narrower proximal and wider distal. Occasionally velum situated on ventral aspect. Gubernaculum more complex than in species of *Leptosomatum*; always with paired capituli directed at right angles to spicular body. Dorsal process always present. Sometimes ventral process also present. Gubernaculum per se forms funnel around distal part of spicules. Well-developed accessory organ invariably present.

Key to Species of Genus Leptosomatides

- 1 (10). Amphids large; width exceeds one-eighth corresponding head diameter.
- 2 (3). Cephalic setae long; length about one-fifth corresponding head diameter.....**L. reducta** Timm, 1959.
- 3 (2). Cephalic setae short; length does not exceed one-tenth corresponding head diameter.
- 4 (5). Small denticles present in anterior part of esophagus.....
.....**L. antarcticum** Mawson, 1956.
- 5 (4). Denticles absent in anterior part of esophagus.
- 6 (9). Sclerotized granules present on cuticle around vulva.
- 70 7 (8). Cervical setae reduced. Body slender ($a = 74$ to 91).....
.....16. **L. marinae** sp. nov.
- 8 (7). Cervical setae present.....13. **L. crassus** Platonova, 1967.
- 9 (6). Sclerotized granules on cuticle around vulva absent.....
.....12. **L. inocellatus** Platonova, 1967.
- 10 (1). Amphids small; width less than one-eighth corresponding head diameter.
- 11 (12). Labial papillae acicular and resemble setae.....
.....14. **L. acutipapillosus** sp. nov.
- 12 (11). Labial papillae blunt, usual in shape.
- 13 (16). Gubernaculum with dorsal and ventral processes.
- 14 (15). Suture of cephalic capsule forms rather deep groove.....
.....**L. conisetosum** Schuurmans-Stekhoven and Mawson, 1955.

- 15 (14). Suture of cephalic capsule sinuous but does not form deep groove.....15. *L. brevisetosus* sp. nov.
 16 (13). Gubernaculum with only dorsal process.
 17 (18). Males devoid of preanal papillae.....
*L. microlaimum* Allgen, 1957.
 18 (17). Males with preanal papillae.
 19 (20). Cephalic setae long; length one-ninth corresponding head diameter.....10. *L. euxina* Filipjev, 1918.
 20 (19). Cephalic setae short; length does not exceed one-twentieth corresponding head diameter.....11. *L. steineri* Filipjev, 1922.

10. *Leptosomatides euxina* Filipjev, 1918 (Figure 23)

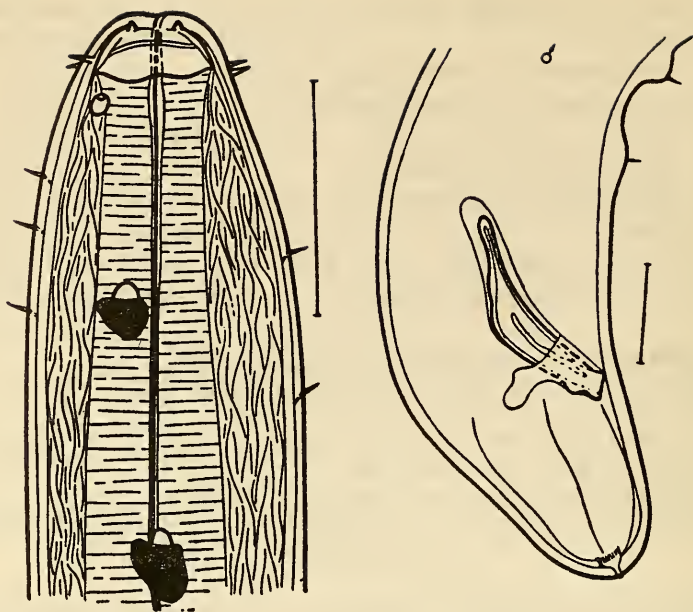
Filipjev, 1918: 51-54, tab. 1, fig. 3; 1922a; 98-100, tab. 1, fig. 1.

$$1 \text{ } \sigma: \frac{11 \ 100 \ 470 \ 1,800 \ - \ 8,877}{35 \ 60 \ 85 \ 95 \ 100 \ 100} \ 8,950 \ \mu\text{m}; \ a=89; \ b=5; \ c=108.$$

$$1 \text{ } \text{f}: \frac{14 \ 113 \ 555 \ 1,815 \ 4,567 \ 7,768 \ 8,897 \ 10,988}{39 \ 53 \ 86 \ 100 \ 115 \ 120 \ 112 \ 98} \ 11,078 \ \mu\text{m}; \ a=92; \\ b=6; \ c=123; \ V=70\%.$$

Body tapers to 2/5 midbody diameter at anterior end, but does not taper toward posterior end. Head narrow. Length of tail exceeds width by 1.2 times. Cuticle bilayered and 7.5 μm thick. Cephalic capsule with walls of average thickness. Cephalic suture distinct and only slightly sinuous. Capsule width exceeds length three times. Thin cephalic ring crosses cephalic capsule medially. Labial papillae small. Cephalic setae 4.0 μm long (1/9 corresponding diameter). Cervical setae slightly shorter, 3.0 μm . Amphids small, rounded, 5.0 μm in diameter (1/8 corresponding body diameter), and situated 16 μm from anterior end (1/100 esophageal length). Eyes situated asymmetrically: one lies 80 μm and the other 140 μm from anterior end (1/20 and 1/13 esophageal length respectively). Pigment bowl 13 $\mu\text{m} \times 13 \mu\text{m}$; diameter of lens 4.0 μm . Esophageal wall highly sclerotized in oral region and markedly thickened at level of amphids. Nerve ring situated 470 μm from anterior end (1/4 esophageal length). Two mature eggs, 430 $\mu\text{m} \times 90 \mu\text{m}$, found in each uterine branch. Spicules of males unequal in length; left spicule 110 μm long and right 90 μm . Neither has a distinct capitulum. Proximal part of spicules uniform in width, middle broadens considerably, almost doubles in width, and distal end blunt. Gubernaculum paired with fairly long processes, 65 μm . Ten pairs of protuberances with setae situated anterior to anus. Accessory organ present with lateral alate fortifying structures on cuticle.

Distinguishing features of this species: 1) location of amphids, which lie close to cephalic capsule, at a distance of 1/100 esophageal length from 71 anterior end; 2) thickening of esophageal wall at level of amphids;

Figure 23. *Leptosomatides euxina*.

3) unequal length of spicules; and 4) unequal distance of eyes from anterior end.

Geographic distribution. Limited to the Black Sea where it has been found in depths ranging from 25 to 200 m among shells and in phaseolin and terebellid silts.

11. *Leptosomatides steineri* Filipjev, 1922 (Figure 24)

Filip'ev, 1922a: 98; 1946: 159, fig. 2; Platonova, 1967: 829.

| | | | | | | |
|----------|--|---------|---------|-------------|-------------|---------------|
| 2 ♂: | 11 | 116 | 463-469 | 1,390-1,499 | — | 10,660-10,769 |
| | 34 | 45-46 | 81-83 | 110-112 | 112-120 | 120-125 |
| | | | | | | 115-125 |
| | × 10,780-10,894 μm; a=85-90; b=8; c=87. | | | | | |
| 13 ♀: | 17 | 104-116 | 433-510 | 1,566-1,823 | 5,686-6,973 | |
| | 29 | 42-52 | 81-94 | 122-132 | 137-162 | 150-187 |
| | | | | | | |
| | × $\frac{5,692-9,033}{155-287}$ $\frac{7,952-10,887}{145-200}$ $\frac{9,703-12,844}{112-137}$ 9,853-12,994 μm; | | | | | |
| | a=63; b=6-7; c=63-93; V=66-71%. | | | | | |
| 10 juv.: | 15-17 | 81-86 | 357-382 | 1,127-1,412 | — | |
| | 19-29 | 34-49 | 58-73 | 85-125 | 87-167 | 92-187 |
| | | | | | | |
| | × $\frac{5,247-9,527}{75-125}$ 5,359-9,652 μm; a=52-59; b=5-8; c=46-77. | | | | | |

Body long, tapers to 1/3 to 1/4 midbody diameter at anterior end and 2/3 at posterior end in female; no posterior attenuation seen in male. Length of tail almost equal to width. Cuticle thick, 10 to 14 μm ; fascicles of intersecting fibers present in outer layer. Cephalic capsule extremely short: width exceeds length by four times; wall very thick. Cephalic suture barely discernible and almost straight. Cephalic ring very narrow and lies in upper third of cephalic capsule. Labial papillae large. Cephalic setae extremely short, 2.3 μm long (1/20 corresponding diameter); cervical setae still shorter, 1.2 μm long. Amphids situated 17 to 29 μm from anterior end (1/60 to 1/80 esophageal length). Amphids 6.9 $\mu\text{m} \times$ 5.8 μm ; width 1/9 to 1/10 corresponding diameter. Eyes situated 116 μm from anterior end (1/12 to 1/15 esophageal length). Pigment bowl 9.3 $\mu\text{m} \times$ 12.7 μm ; diameter of lens 8.1 μm . Nerve ring situated 433 to 510 μm from anterior end (1/3 esophageal length). Length of anterior female genital tube 1,236 to 2,575 μm , posterior tube 1,030 to 2,163 μm , and their reflexed parts 927–2,060 μm and 721 to 1,442 μm respectively. One egg, 412 to 515 $\mu\text{m} \times$ 133 to 154 μm , seen in each uterine branch. Male spicules smoothly arcuate in midsection; length 137 μm and width in narrowest part 17.5 μm and in widest part 25 μm . Gubernaculum with long (30 μm) process. One postanal and 17 preanal pairs of papillae present on tail. Accessory organ with complex sclerotized skeleton situ-

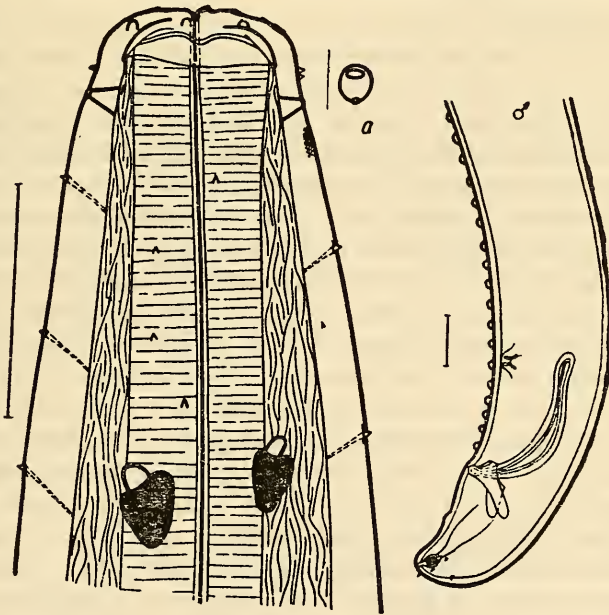


Figure 24. *Leptosomatides steineri* (a—amphid).

ated anterior to anus at a distance of 125 μm and provided with four processes, two directed dorsally and two ventrally.

Distinguishing features of this species: 1) extremely short cephalic capsule; 2) extremely short cephalic setae; 3) smoothly curved spicules of equal length; 4) one pair of postanal and 17 pairs of preanal papillae; and 5) accessory organ with complex structural processes (dorsal and ventral).

Geographic distribution. Arctic species. Found in Kara Sea in depths ranging from 140 to 360 m in sandy silt.

12. *Leptosomatides inocellatus* Platonova, 1967 (Figure 25)

Platonova, 1967: 829, fig. 1 (3, 4).

| | | | | | | | | | | |
|---|-------------------------------|--------------------------------|---------------------------------|-------------------------------|-------------------------------|--------------------------------|------------------------|-------|------|-------|
| 1 ♂: | $\frac{21}{40}$ | — | $\frac{510}{157}$ | $\frac{1,746}{225}$ | — | $\frac{9,831}{225}$ | 10,031 μm ; | a=37; | b=6; | c=50. |
| 2 ♀: | $\frac{15-23}{30-40}$ | $\frac{52-61}{52-61}$ | $\frac{515-618}{150-219}$ | $\frac{2,060-2,678}{177-290}$ | $\frac{5,768-7,539}{212-336}$ | | | | | |
| × | $\frac{8,652-9,599}{232-331}$ | $\frac{9,991-11,330}{225-336}$ | $\frac{13,390-15,264}{162-250}$ | 13,415-15,488 μm ; | | | | | | |
| a=45-58; b=6; c=70-108; V=52-64%. | | | | | | | | | | |
| 73 2 juv.: | $\frac{14-17}{17-19}$ | $\frac{34-40}{34-40}$ | $\frac{104-120}{104-120}$ | $\frac{1,390-1,644}{142-150}$ | — | $\frac{5,764-10,660}{112-125}$ | | | | |
| × 5,889-10,835 μm ; a=39-64; b=4-8; c=47-62. | | | | | | | | | | |

Body extremely long, tapers to 1/4 to 1/5 midbody diameter at anterior end and to 5/6 to 10/13 at posterior end. Length of tail almost equal to width. Cuticle very thick, 13 to 17 μm , bilayered, with intersecting fascicles of fibers in outer layer. Cephalic capsule short; width exceeds length three times. Cephalic wall rather thin. Cephalic suture faintly discernible and straight. Narrow cephalic ring situated medially in cephalic capsule. Labial papillae short. Cephalic setae short, but rather thick and massive, and 4.6 to 6.0 μm long (1/10 to 1/13 corresponding diameter). Cervical setae somewhat short, 3.5 to 4.0 μm long, situated in small pits, occasionally in groups of three to nine. Preanal setae of males significantly longer, up to 20 μm long. Amphids situated 29 to 35 μm (1/50 to 1/70 esophageal length) from anterior end, large, stretched longitudinally, 10.0 to 11.6 μm long, and width 1/6 corresponding diameter. Eyes absent. Nerve ring 510 to 618 μm from anterior end (1/4 esophageal length). Vulva median or posteromedian. Sclerotized granules around vulva absent. Length of anterior genital tube 1,545 to 2,884 μm , its bent part 1,133 to 2,678 μm , of posterior tube 1,200 to 2,020 μm , and of reflexed parts 1,133 to 1,648 μm . Sexually mature females with one egg 772 to 669 μm [sic] × 175 to 257 μm in each uterine branch. In females two to three pairs of papillae present on tail. Anterior male gonad 262 μm and posterior 1,442 μm long. Spicules equal in length but different in shape; right spicule smoothly arcuate,

with slightly thickened capitulum, followed by neck (cervix) widening into spicular body; left spicule significantly more irregular in shape. Spicular length 237 μm . Length of gubernaculum 50 μm . Accessory organ situated 150 μm anterior to anus. Setae in anal region number 20 pairs, of which 6 are postanal. Anterior to these setae lie 17 pairs of papillae.

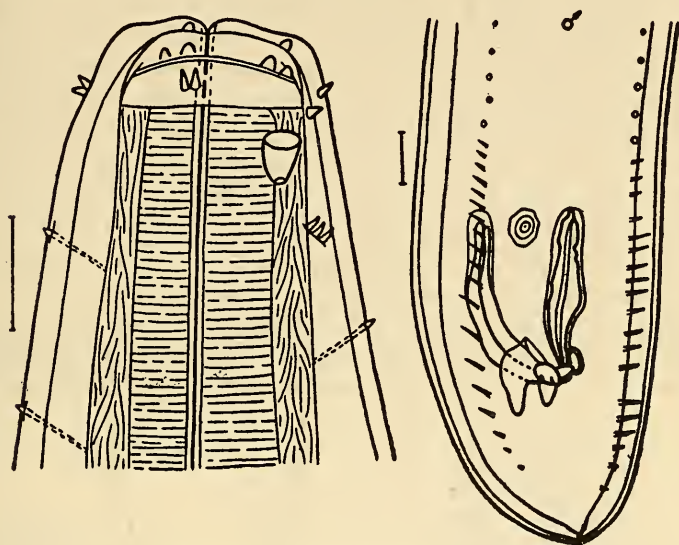


Figure 25. *Leptosomatides inocellatus*.

- 74 Distinguishing features of this species: 1) extremely thick cuticle; 2) cervical setae gathered in groups in small pits; 3) large amphids, stretched longitudinally; 4) complete absence of eyes and photosensitive pigment; 5) presence of two to three pairs of papillae on tail of female; 6) absence of sclerotized granules around vulva; 7) spicules significantly different in shape; and 8) presence of 20 pairs of setae and 17 pairs of papillae in anal region of male.

Geographic distribution. To date found only in the Kara Sea at depths ranging from 200 to 800 m.

13. *Leptosomatides crassus* Platonova, 1967 (Figure 26)

Platonova, 1967: 829-831, fig. 1 (5-7)

| | | | | | |
|-------------------------|--|-------------|--------------|-------------|-------------|
| $5 \text{ } \text{f}$: | 13-23 | 98-109 | 412-433 | 1,463-1,669 | 5,580-5,665 |
| | 21-26 | 43-48 | 81-89 | 125-150 | 167-200 |
| | 217-250 | | | | |
| \times | 6,407-7,828 | 9,191-9,888 | 9,497-12,154 | | |
| | 200-212 | 225-250 | 135-150 | | |
| | 9,622-12,329 μm ; | | | | |
| | a = 48-51; b = 7-8; c = 70-77; V = 64-67%. | | | | |

| | 10-11 | 50-63 | 220-270 | 980-991 | — |
|------------------------------------|--|-------|---------|---------|--------|
| 5 juv.: | 18-21 | 28-31 | 50-58 | 76-82 | 95-105 |
| | 115-120 | | | | |
| $\times \frac{4,032-4,287}{90-95}$ | 4,203-4,437 μm ; a=30-35; b=4; c=30-34. | | | | |

Body tapers to 1/5 midbody diameter at anterior end and 5/8 at posterior end. Length of tail almost equal to width. Cuticle thick, 11.6 to 14.0 μm , with fascicles of intersecting fibers in outer layer. Cephalic capsule with rather thick wall but poorly expressed cephalic suture; width exceeds length by 2.5 times. Extremely narrow cephalic ring situated in upper third of cephalic capsule. Labial papillae distinct. Cephalic setae extremely short, 2.32 μm in length (1/20 corresponding diameter). Cervical setae 1.6 μm long. Amphids 23 to 34 μm from anterior end (1/60 esophageal length) and 8.0 to 7.0 μm wide (1/6 to 1/8 corresponding diameter). Eyes 98 to 109 μm from anterior end (1/14 to 1/15 esophageal length). Pigment bowl 8.12 $\mu\text{m} \times 15.08 \mu\text{m}$; diameter of lens 7.0 to 8.0 μm . Esophageal wall sclerotized in oral region. Nerve ring 412 to 433 μm from anterior end (1/3 esophageal length). Length of anterior female genital tube 1,957 to 2,575 μm , of posterior tube 2,060 μm , and

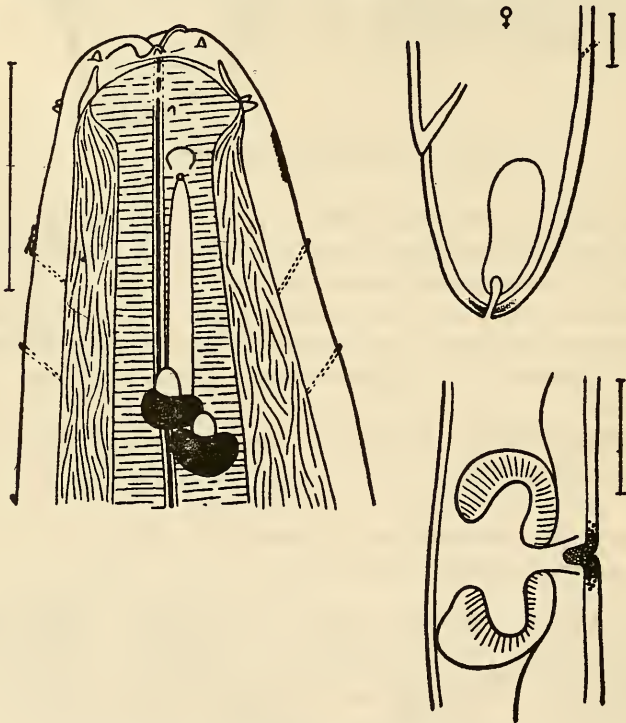


Figure 26. *Leptosomatides crassus* (below vulva).

of reflexed parts 515 to 1,030 μm and 824 to 927 μm respectively. Four to eight eggs, 154 to 360 $\mu\text{m} \times 824$ to 206 μm , found in uteri.

This species is distinguished from others in the genus (except *L. steineri*) by the following features: 1) long and broad body covered with thick cuticle; 2) relatively long cephalic capsule; and 3) extremely short cephalic and cervical setae. *L. crassus* differs from *L. steineri* in: 1) wider body; 2) cephalic capsule almost two times longer; 3) amphids situated closer to anterior end; 4) esophageal wall much thicker in oral region; 5) greater number of eggs in uteri of *L. crassus* (up to eight) than in *L. steineri* [in my material only one each, but according to Filip'ev (1946) up to four]. It should be noted that Filip'ev (1916) gives more taxonomic importance to such characters as size and number of eggs.

Geographic distribution. Found only in the Barents Sea in the littoral zone, predominantly under offshoots of brown algae.

14. *Leptosomatides acutipapillosus* sp. nov. (Figure 27)

Holotype ♀: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7034.

$$\begin{array}{cccccccc} 17 & 92 & 463 & 1,802 & 5,922 & 8,909 & 11,896 & 13,441 \\ \hline 25 & 46 & 81 & 125 & 162 & 187 & 192 & 200 & 132 \end{array} \quad 13,591 \mu\text{m};$$

$$a = 68; b = 7; c = 91; V = 66\%.$$

Paratypes.

$$\begin{array}{cccccc} 2 \text{ ♀:} & 17 & 92-109 & 459-463 & 1,695-1,802 & 5,929-6,330 \\ \hline & 25-29 & 46 & 81-100 & 125-137 & 162-177 & 187-242 \\ \times & 8,909-9,214 & 11,896-12,201 & 13,441-14,466 & & & \\ \hline & 192-225 & 200-212 & 132-150 & & & 13,591-14,641 \mu\text{m}; \\ & & & & & & a = 65-68; b = 7-9; c = 84-91; V = 62-66\%. \end{array}$$

$$\begin{array}{cccccc} 12 \text{ juv.:} & 10-17 & 69-104 & 300-408 & 969-1,438 & — \\ \hline & 14-25 & 31-46 & 48-98 & 72-150 & 97-175 & 120-187 \\ \times & 5,304-8,339 & & & & & \\ \hline & 87-125 & & & & & 5,429-8,469 \mu\text{m}; a = 44-48; b = 5-6; c = 44-70. \end{array}$$

Body tapers to 1/4 to 1/5 midbody diameter at anterior end and 5/7 to 2/3 at posterior end. Head round, tail small, slightly conically stretched and bluntly rounded at end. Length of tail exceeds width by 1.2 times. Cuticle bilayered, 9.0 to 10.0 μm thick, with fascicles of intersecting fibers situated in outer layer. Cephalic capsule with fairly thin wall and barely discernible suture. Narrow cephalic ring situated in upper third of cephalic capsule. Width of latter exceeds length by 2.7 times. Labial papillae with characteristic structure, pointed, and resemble setae. Cephalic setae short, 3.5 μm in length (1/13 corresponding body diameter). Cervical papillae 2.3 μm long. Amphids 29 μm from anterior end (1/60 eso-

phageal length) and $8.1 \mu\text{m}$ wide ($1/7$ to $1/8$ corresponding body diameter). Eyes 92 to $109 \mu\text{m}$ ($1/16$ to $1/18$ esophageal length). Pigment bowl 10.4 to $12.7 \mu\text{m}$; diameter of lens $7.0 \mu\text{m}$. Nerve ring situated about $1/4$ length of esophagus from anterior end. Length of anterior female genital tube $2,884$ to $2,987 \mu\text{m}$, of posterior tube $2,987$ to $3,000 \mu\text{m}$, and of reflexed parts $2,060$ to $1,751$ and $1,554$ to $1,957 \mu\text{m}$ respectively. Two to eight eggs, 154 to $257 \mu\text{m} \times 133$ to $154 \mu\text{m}$, found in each uterus.

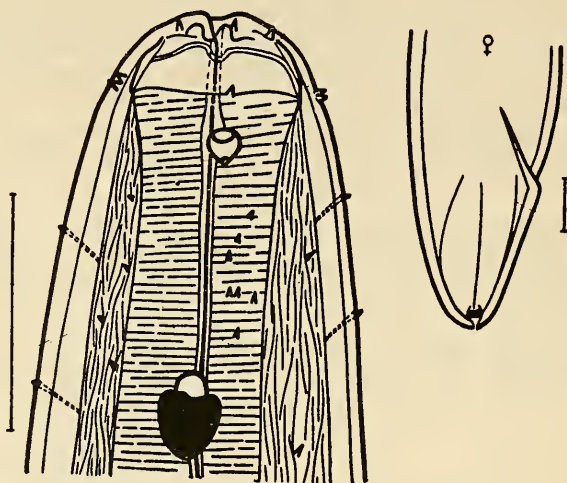


Figure 27. *Leptosomatides acutipapillosus* sp. nov.

- 76 Distinguishing features of this species: 1) conically stretched but obtusely rounded tail of female; 2) labial papillae pointed and resemble thick setae. Thick cuticle, relatively long cephalic capsule, and amphids with large openings are also characteristic of this species. Structurally close to *L. crassus* but further distinguished from it by shape of tail, pointed labial setae, somewhat wider cephalic ring and its lower position, and longer cephalic setae.

Geographic distribution. The only place of occurrence of this species has been the Sea of Okhotsk; found at a depth of 25 m in silted benthos.

15. *Leptosomatides brevisetosus* sp. nov. (Figure 28)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 6128.

| | | | | | | |
|----|----|-----|-------|-----|--------|---|
| 25 | 92 | 515 | 1,799 | — | 10,825 | 11,000 μm ; a=73; b=6; c=62. |
| 28 | 46 | 66 | 100 | 112 | 150 | |

Paratypes.

| | | | | | |
|-------|---|--------|---------|-------------|-------------|
| 8 ♂: | 10-11 | 81-92 | 450-515 | 1,799-1,854 | — |
| | 31-34 | 46-52 | 86-70 | 82-100 | 112-125 |
| × | $\frac{9,840-11,857}{137-160}$ 10,000-12,000 μm ; a = 73-92; b = 6-7; c = 62-84. | | | | |
| 23 ♀: | 11-23 | 92-125 | 470-618 | 1,892-2,472 | 5,022-8,170 |
| | 32-34 | 42-50 | 76-87 | 120-145 | 167-180 |
| × | $\frac{10,000-11,951}{180-250}$ $\frac{10,935-14,938}{260-200}$ [sic] $\frac{11,800-17,441}{140-150}$ 12,000-17,616 μm ; | | | | |
| | a = 56-80; b = 6-8; c = 70-100; V = 75-80%. | | | | |

Body tapers to 2/5 to 1/3 midbody diameter at anterior end and 10/13 to 5/8 at posterior end. In males body in region of spicular apparatus somewhat wider than midwidth. Cuticle thick, 7.5 μm , with fascicles of intersecting fibers in outer layer. Labial papillae short but very wide. Cephalic and cervical setae papillate, very short, with rounded ends. Length of cephalic setae 2.0 μm (1/15 to 1/16 corresponding diameter). Cephalic capsule short; length 2.8 times less than width. Cephalic suture barely discernible and sinuous. Cephalic ring shifted more anteriorly: situated in first quarter of capsule. Lips sclerotized. Fairly large amphids, width 8.12 μm (1/8 corresponding diameter), situated 25 to 32 μm from anterior end. Eyes 81 to 100 μm from anterior end. Pigment bowl 11.6 μm × 11.6 μm ; diameter of lens 5.8 μm . Nerve ring 450 to 618 μm from anterior end (1/4 esophageal length). Length of anterior female genital tube 3,000 to 3,200 μm , of posterior tube 2,884 to 2,900 μm , and of reflexed parts 1,700 to 1,820 μm and 2,000 to 2,060 μm respectively. Vulval lips thick and muscular with upper lip covering lower one. Genital armature of male represented by spicular apparatus, accessory organ, seven to nine pairs of pretubal, and one pair of postanal papillae. Length of spicules 140 to 170 μm . Each spicule divided into two parts: distal part significantly wider than proximal. Gubernaculum with dorsal and ventral processes. Accessory organ 150 to 160 μm from anus, with sclerotized cyathiform supports with two alate processes, one proceeding anteriorly and the other posteriorly.

Distinguishing features of this species: 1) amphids situated at a distance equal to two cephalic lengths from anterior end of body, i.e. situated farther away from anterior end compared to other species of the genus; 2) characteristic shape of amphids, anteriorly wide and tapering rather abruptly posteriorly; 3) absence of anal setae; 4) pretubal and postanal situation of papillae in males, but no papillae between anus and accessory organ; 5) cyathiform sclerotized structures with alate processes supporting accessory organ from inside; 6) distinct division of spicules into narrow proximal part and wide distal part; 7) presence of dorsal and

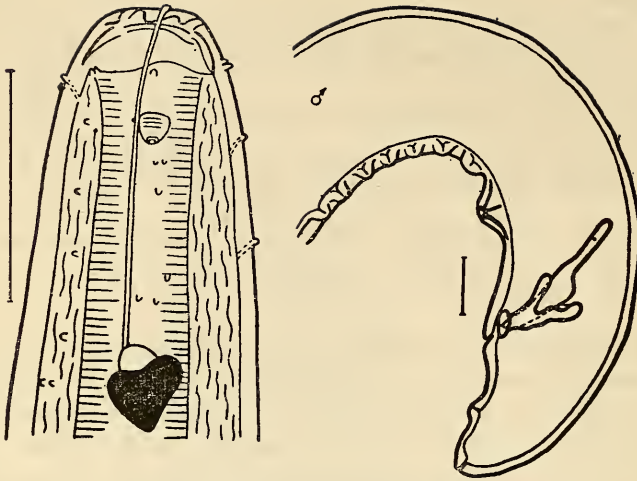


Figure 28. *Leptosomatides brevisetosus* sp. nov.

ventral processes on gubernaculum; and 8) extremely short, papillate cephalic and cervical setae.

Geographic distribution. Sea of Okhotsk (Kuril Islands). Found in the littoral zone under offshoots of algae and in sandy benthos.

16. *Leptosomatides marinae* sp. nov. (Figure 29)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7880.

$$\frac{18 \quad 84 \quad 465 \quad 1,834 \quad — \quad 12,474}{24 \quad 40 \quad 70 \quad 140 \quad 145 \quad 169 \quad 148} \quad 12,633 \text{ } \mu\text{m}; \text{ a} = 75; \text{ b} = 9, \text{ c} = 80.$$

Paratypes.

$$5 \text{ } \sigma: \frac{16-18 \quad 78-91 \quad 399-465 \quad 1,729-2,061 \quad —}{24-26 \quad 38-48 \quad 70-80 \quad 120-145 \quad 142-160 \quad 137-169}$$

$$\times \frac{10,573-14,323}{130-182} \quad 10,719-14,522 \text{ } \mu\text{m}; \text{ a} = 75-94; \text{ b} = 6-7; \text{ c} = 73-80.$$

$$78 \quad 8 \text{ } \sigma: \frac{15-23 \quad 78-96 \quad 399-465 \quad 1,662-2,221 \quad 6,184-7,620}{26-41 \quad 42-48 \quad 72-89 \quad 110-120 \quad 117-148 \quad 150-172}$$

$$\times \frac{8,578-10,844 \quad 10,706-14,900 \quad 12,701-15,964}{156-202 \quad 137-152 \quad 117-143} \quad 12,847-16,623 \text{ } \mu\text{m};$$

$$\text{ a} = 75-93; \text{ b} = 6-8; \text{ c} = 88-117; \text{ V} = 64-70\%.$$

Body tapers to 1/4 midbody diameter at anterior end and to not less than 2/3 at posterior end. Tail length somewhat exceeds width. Cuticle thick, 13.2 μm , and bilayered. Cephalic capsule thin-walled and width twice length. Narrow cephalic ring lies in upper part of cephalic capsule.

Cephalic setae very small and papillate. Cervical setae reduced. Amphids $8.4 \mu\text{m} \times 6.0 \mu\text{m}$ and situated immediately behind cephalic capsule. Eyes 80 to $90 \mu\text{m}$ from anterior end ($1/20$ esophageal length). Pigment bowl $12.0 \mu\text{m} \times 9.6 \mu\text{m}$; diameter of lens $6.0 \mu\text{m}$. Nerve ring situated in anterior third of esophagus. Length of anterior female genital tube $2,128$ to $3,724 \mu\text{m}$, of posterior tube $2,128$ to $4,056 \mu\text{m}$, and of reflexed parts $1,303$ to $2,128 \mu\text{m}$ and $1,130$ to $2,261 \mu\text{m}$ respectively. Ten eggs found in uterus. Vulval aperture encircled by sclerotized granules. Length of anterior male genital tube $2,257$ to $2,926 \mu\text{m}$ and posterior tube $1,130$ to $2,660 \mu\text{m}$. Spicules straight, wide, without capitulum, slightly acicular at proximal end, and with small protuberances on dorsal margin. Length of spicules 150 to $160 \mu\text{m}$. Gubernaculum large, 80 to $98 \mu\text{m}$ long, with an expansion at proximal end and pointed process directed ventrally. Accessory organ 150 to $180 \mu\text{m}$ anterior to anus.

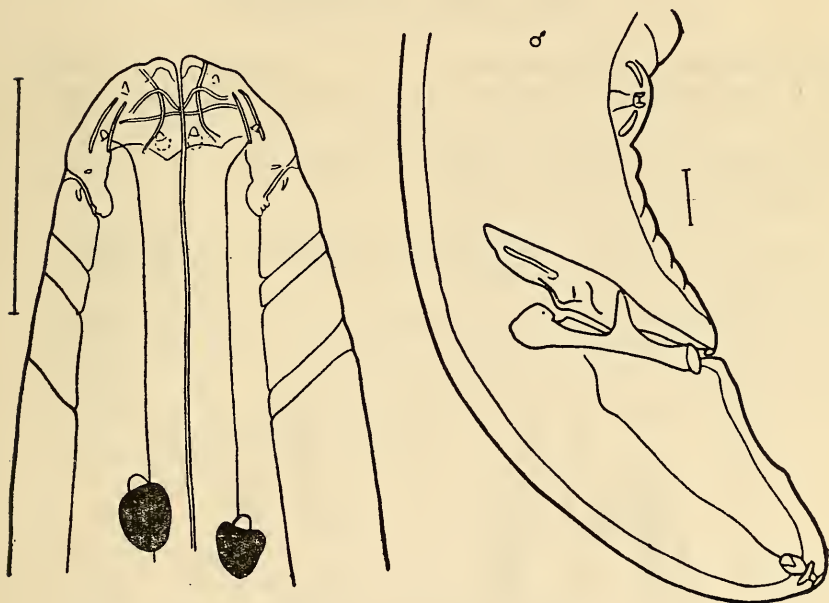


Figure 29. *Leptosomatides marinae* sp. nov.

This species is related to that group of forms in which females have sclerotized granules around the vulva. Structure of the spicular apparatus similar to that of *L. euxina*, but the gubernaculum differs significantly in structure.

- 79 **Geographic distribution.** Found around Kuril Islands—Kunashir (Gulf of Izmena) and Iturup (Pacific coast)—at a depth of 4.0 to 19.0 m in silted-sand among offshoots of algae, but more often in sponges.

3. Genus *Leptosomella* Filipjev, 1927

Filip'ev, 1927: 79.

Type species. *Leptosomella acrocerca* Filipjev, 1927.

Nematodes of this genus are significantly smaller in size than representatives of the two preceding genera. Cephalic capsule consists of three domes with an irregular suture. Oral cavity almost undeveloped. Cephalic setae long. In each pair of subventral and subdorsal setae one seta notably shorter than the other. Cervical setae absent. Distinct cervical gland present. Tail elongated and acicular.

Only one species has been discovered to date in the seas of the Soviet Union.

Key to Species of Genus Leptosomella

- 1 (2). Photosensitive eyes with lens *L. phaustra* Inglis, 1971.
- 2 (1). Photosensitive eyes without lens *L. acrocerca* Filipjev, 1927.

*17. *Leptosomella acrocerca* Filipjev, 1927 (Figure 30)
 Filip'ev, 1927: 80 (description).

| | | | | | |
|------|-------------|-------------|--|-------------|-------------|
| 3 ♀: | — | 195-300 | 560-980 | 1,530-2,570 | 1,830-2,900 |
| | 13-17 | 45-47 | 50-60 | — | 57-68 |
| × | 2,170-3,220 | 2,760-3,530 | 2,850-3,900 μm; a = 54-63; b = 4-7; c = 26-35. | | |
| | — | 40-52 | | | |

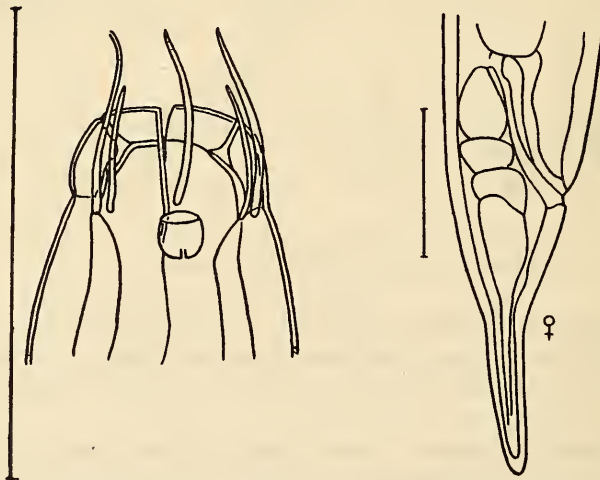


Figure 30. *Leptosomella acrocerca* (from Filip'ev 1927).

Body filiform and tapers posteriorly from preanal region. Tail conical and narrows notably toward posterior end. Cuticle smooth and comparatively thick ($4.0\ \mu\text{m}$). Head truncated; notably narrows at level of cephalic suture. Labial papillae distinct. Of the ten cephalic setae six are longer—13 to 15 μm in length (almost equal to cephalic diameter). Amphids situated immediately behind cephalic setae. Cephalic suture distinct. Cervical gland extremely large, $25\ \mu\text{m} \times 60\ \mu\text{m}$, and situated against posterior part of esophagus; its duct distinctly visible and its pore situated at a distance of 50 to 57 μm from anterior end. Nerve ring situated in anterior quarter of esophagus. Cardia $15.0\ \mu\text{m} \times 9.0\ \mu\text{m}$ in size. Rectum 40 to 45 μm long. Eggs $200\ \mu\text{m} \times 40\ \mu\text{m}$ in size. Vagina thick-walled.

Geographic distribution. Found only once in the Barents Sea at a depth of 36 m in gravel.

2. Subfamily SYNONCHINAE Platonova, 1970

Cephalic capsule short, rather thin-walled, but distinctly developed. Cephalic suture may be straight, but often bends and forms lobes and grooves. Interlobular grooves wide and not divided into furrows; formed by smoother curve of cephalic suture. All grooves identical in shape; lateral ones, in which amphids are located, do not differ in any way from the rest. Amphids may be situated inside lateral grooves as well as below level of cephalic suture. Cephalic ring narrow. Oral cavity small, often armed with onchia. Nerve ring encircles esophagus most often in its anterior third. Tail in majority of genera stretched but claviform at end. Posterior end in males covered with a large number of setae. Preneural part of body in both male and female usually covered with numerous long setae. Photosensitive eyes absent.

Key to Genera of Subfamily Synonchinae

- 1 (4). Tail short, wide, and blunt.
- 2 (3). Posterior margin of cephalic capsule serrated
 *Synonchoides* Wieser, 1956.
 (one species: *S. galatheae* Wieser, 1956).
- 3 (2). Posterior margin of cephalic capsule smooth, not serrated
 *Corythostoma* Wieser, 1956.
 (two species: *C. kreisi* Wieser, 1956.
 and *C. triaulolaimus* Hope, 1967).
- 4 (1). Tail elongated, claviform at end.
- 5 (8). Interlobular grooves narrow near posterior margin of capsule.

- 6 (7). Oral cavity armed with one dorsal tooth **Tuerkiana** Platonova, 1970.
- 7 (6). Oral cavity armed with three teeth 5. **Anivanema** Platonova, gen. nov.
- 8 (5). Interlobular grooves broad near posterior margin of capsule.
- 9 (14). Gubernaculum of spicular apparatus funnel-shaped.
- 10 (11). Amphids large, their width constituting one-third to one-half of corresponding body diameter **Fiacra** Southern, 1914.
- 11 (10). Amphids small, their width constituting one-eighth to one-seventh of corresponding body diameter.
- 12 (13). Oral cavity armed with three small teeth **Eusynonchus** Platonova, 1970.
- 13 (12). Oral cavity armed with one large tooth **Paratuerkiana** Platonova, 1970.
- 14 (9). Gubernaculum different shape (not funnel-shaped).
- 15 (16). Oral cavity not armed; totally devoid of teeth **Jaegerskioeidia** Filipjev, 1916.
- 81 16 (15). Oral cavity armed.
- 17 (18). Oral cavity armed with three teeth, of which dorsal tooth much larger than two subventral ones 4. **Synonchus** Cobb, 1893.
- 18 (17). Oral cavity armed with three teeth of equal size and three serrated plates **Macronchus** Inglis, 1964.
(one species: **M. shealsi** Inglis 1964).

4. Genus *Synonchus* Cobb, 1893

Cobb, 1893: 411, 412.

Type species: *Synonchus fasciculatus* Cobb, 1893.

Cephalic suture slightly wavy but does not form distinct interlobular grooves. Amphids situated below posterior margin of cephalic capsule. Oral cavity armed with one large dorsal and two small subventral onchia. Setae present in preneural region but sometimes small and few. Accessory organ present, but structure undescribed in type species. Spicules arcuate with distinct capituli. Gubernaculum simple in structure, without ventral process. Tail elongated and claviform at end.

In descriptions of species of genera *Synonchus* and *Anivanema* given below, figures in Cobb's formula denote distances from anterior end of body to: 1. posterior end of cephalic capsule; 2. nerve ring; 3. base of esophagus; 4. commencement of anterior genital tube; 5. vulva; 6. commencement of posterior genital tube; and 7. anus.

As stated earlier, measurements according to the formula are fewer for males.

Key to Species of Genus Synonchus

- 1 (2). Length of cephalic setae and width of amphids constitute one-third of corresponding diameters. . . . *S. fasciculatus* Cobb, 1893.
 2 (1). Length of cephalic setae constitutes one-tenth of corresponding diameter. . . . 18. *S. murmanicus* Filipjev, 1927.

18. *Synonchus murmanicus* Filipjev, 1927 (Figure 31)

Filip'ev, 1927: 88-90, pl. I, fig. 11, a-d.

| | | | | | | | |
|---------|--|---------|-------------|---------|---------------|---------|-----|
| | 29-31 | 618-648 | 2,163-2,234 | — | 11,433-12,740 | | |
| 2 ♂: | 34-40 | 63-71 | 205-212 | 288-309 | 309-360 | 206-225 | |
| | × 11,790-13,100 μm; a=32-42; b=5-6; c=32-42. | | | | | | |
| 2 ♀: | 29-34 | 618 | 2,163 | 4,223 | 6,798-7,828 | 9,785 | |
| | 40-46 | 63 | 167-200 | 216-309 | 360 | 257-329 | 339 |
| × | $\frac{11,948-13,184}{185-206}$ 12,308-13,472 μm; a=36-52; b=6; c=34-47; V=51-55%. | | | | | | |
| 8 juv.: | 23-29 | 515-648 | 1,545-2,038 | — | 7,931-10,896 | | |
| | 32-40 | 53-58 | 147-154 | 195-216 | 226-278 | 125-185 | |
| | × 8,181-11,233 μm; a=35-42; b=5; c=32-33. | | | | | | |

Large nematodes. Body tapers to 10/43 to 1/5 midbody diameter at anterior end and 2/3 to 10/17 at posterior end. Cephalic end rounded anteriorly. Tail of female conical and pointed toward end; that of male highly attenuate, elongated. Cuticle thick, 17 to 20 μm, and bilayered. Fascicles of intersecting fibers present in outer layer, due to which cuticle appears finely annulated. Width of cephalic capsule exceeds length by 2.0 to 2.3 times. Cephalic ring situated in middle of cephalic capsule. Lateral walls of latter with highly sclerotized thickenings. Labial papillae extremely small. Cephalic setae very short, 5.8 μm in length (1/10 corresponding diameter). Cervical papillae 2.2 μm and anal 17.5 to 20.0 μm long. Amphids situated immediately behind cephalic capsule and 9.0 to 82 11.0 μm wide (1/6 to 1/7 corresponding diameter). Nerve ring situated beyond anterior third of esophagus. Length of anterior female genital tube 2,575 μm, of posterior tube 2,987 μm, and of reflexed parts 1,751 μm and 1,854 μm respectively. One egg, 721 μm × 226 μm, present in each uterine branch. Vulva situated slightly posterior to midbody. Spicules slightly curved, almost uniform in width throughout their length; taper slightly toward distal end and sharply at proximal to form pointed capitulum. Length of spicules 175 μm and width in midsection 17 μm. Gubernaculum simple in structure, without processes, and projects slightly over ventral surface of spicules. Accessory organ 237 to 250 μm anterior to anus. Setae situated in anal region number 24 pairs; one pair almost at tip of tail. Caudal glands tubular.

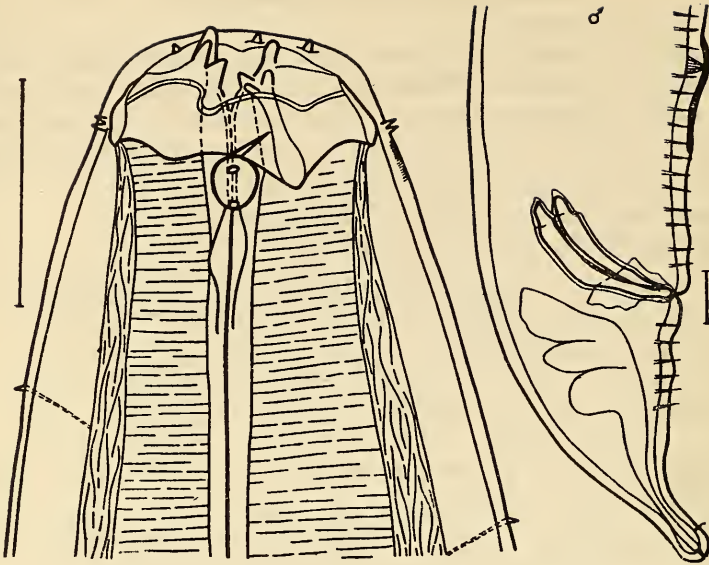


Figure 31. *Synonchus murmanicus*.

Distinguishing features of this species: 1) slightly wavy cephalic suture; 2) highly sclerotized thickenings of lateral walls of cephalic capsule; 3) short cephalic and cervical setae; 4) weakly arcuate spicules almost uniform in width throughout their length, with pointed capitulum significantly narrower than spicule body; 5) gubernaculum simple in structure, without processes; and 6) presence of about 20 pairs of setae in anal region of male.

Geographic distribution. Found only in the Barents Sea, predominantly in a depth range of 30 to 40 m, in silted sand.

5. Genus *Anivanema* gen. nov.

Type species. *A. magna* sp. nov.

Very large worms. Cephalic capsule well developed and reminiscent of *Thoracostoma* in structure. Interlobular grooves very deep, narrowing toward posterior margin of cephalic capsule. Margins of cephalic lobes serrated. Amphids large and situated in upper wider part of interlobular grooves. Three large onchia of equal length situated in upper part of oral cavity. Cephalic and preneural setae long and thick. Tail elongated and claviform at end.

83 19. *Anivanema magna* gen. nov., sp. nov (Figure 32)

Holotype ♀: Institute of Zoology, Academy of Sciences, USSR. Collection No. 6591.

| | | | | | | | |
|--------------------------|------|-------|-------|--------|--------|--------|------------------------|
| 34 | 1030 | 1,957 | 9,167 | 12,978 | 15,779 | 22,413 | 22,773 μm ; |
| 40 | 69 | 187 | 225 | 228 | 232 | 230 | |
| a=98; b=19; c=64; V=57%. | | | | | | | |

Body tapers to 1/6 midbody diameter at anterior end, but almost does not taper toward posterior end. Head anteriorly truncated. Tail tapers posteriorly from anus but terminates as a bulb. Cuticle thick, 10.0 μm . Cephalic capsule with deep interlobular grooves; depth 11.6 μm . Cephalic ring situated at end of first third of cephalic capsule; width at base of ring exceeds that at top by 1.7 times. Labial papillae well developed and wide. Cephalic and cervical setae long and thick.

Cephalic setae 17.4 μm long (1/3 corresponding body diameter) and cervical 15 μm long. Minute setae present on caudal bulb. Amphids large and situated in grooves of cephalic capsule. Oral cavity small, with thick sclerotized walls. Nerve ring encircles esophagus somewhat above its middle. Length of anterior female genital tube 3,810 μm , of posterior tube 2,800 μm , and of reflexed parts 3,000 μm and 2,200 μm respectively. One egg, 1,050 $\mu\text{m} \times 220 \mu\text{m}$, detected in genital tract. Vulva shifted slightly posterior to midbody.

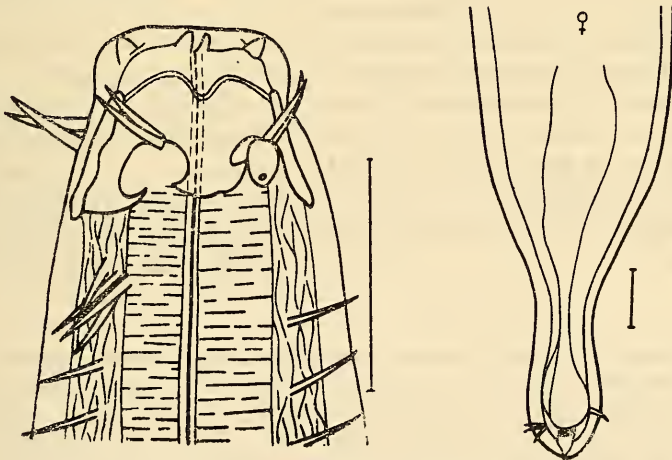


Figure 32. *Anivanema magna* gen. nov., sp. nov.

Distinguishing features of this species: 1) well-developed cephalic capsule with deep interlobular grooves; 2) long and strong setae, especially cephalic ones; 3) large and distinct labial papillae; and 4) extremely large size of body.

Geographic distribution. Found in the Gulf of Aniva in the Sea of Okhotsk in gravel at a depth of 23 m.

3. Subfamily THORACOSTOMATINAE de Coninck, 1965

Cephalic capsule powerfully developed, long, and thick-walled. Interlobular grooves very deep and divided into furrows and fenestrae which vary in shape. Lateral interlobular grooves, inside which amphids are situated, always significantly wider and differ in shape from subdorsal and subventral grooves. Cephalic lobes may be smooth or with perforations of different size and shape. Cephalic ring varies considerably in width and location. Oral cavity poorly developed and usually unarmed. Nerve ring encircles esophagus usually at border of its anterior third or at its mid-section. Tail wide, short, and obtusely rounded. Setae present on body but in very small numbers; shorter in length than those of representatives of subfamily Synonchinae. Photosensitive eyes or spots of photosensitive pigment always present in preneural part.

Key to Genera of Subfamily Thoracostomatinae

- 1 (2). Cephalic capsule with three distinct anteriorly directed processes.
.....**Triceratonema** gen. nov.
- 2 (1). Cephalic capsule of different shape.
- 3 (6). Cephalic capsule asymmetrical; ventral wall significantly thicker and longer than dorsal and forms a protuberance in front.
- 4 (5). Process of gubernaculum situated at a right angle to longitudinal axis of spicules.....7. **Pseudocella** Filipjev, 1946.
- 5 (4). Process of gubernaculum forms an acute angle with longitudinal axis of spicules.....**Thoracostoma** Marion, 1870.³
- 6 (3). Cephalic capsule symmetrical; entire wall even in length and thickness.....6. **Deontostoma** Filipjev, 1916.

³This genus includes the following species: *Thoracostoma echonodon*, Marion, 1870; *T. zolae* Marion, 1870; *T. coronatum* (Eberth, 1863); *T. setosum* (Linstow, 1896); *T. chilense* Steiner, 1922; *T. steineri* Micoletzky, 1922; *T. vallini* Allgen, 1927; *T. philippinensis* Allgen, 1951; *T. australe* Mawson, 1953; *T. anocellatum* Schuurmans-Stekhoven and Mawson, 1955; *T. angustifissulatum* Mawson, 1956; *T. bruuni* Wieser, 1956; *T. scizeopistylum* Mawson, 1958; *T. parasetosum* Mawson, 1958; *T. karachiense* Timm, 1959; and *T. trachygaster* Hope, 1967.

Descriptions of species by Allgen (*T. penduli* Allgen, 1954; *T. bracylobatum* Allgen, 1954; *T. multipapillosum* Allgen, 1957; *T. microchaetum* Allgen, 1957; *T. sivertseni* Allgen, 1958; *T. unifenestratum* Allgen, 1959; *T. falklandiae* Allgen, 1959; and *T. microfenestratum* Allgen, 1959) are so unsatisfactory and the illustrations so sketchy that I could not determine to which genus they should be assigned.

Genus *Triceratonoma* gen. nov.

Type species. *Thoracostoma campbelli* Ditlevsen, 1922.

Large nematodes. Powerful cephalic capsule notably differs in structure from cephalic capsule of other representatives of this subfamily. Instead of a single ventral outgrowth, there are three well-developed cornuate processes in *Triceratonema*, directed anteriorly and situated along the oral angles—one ventral and two sublateral. These outgrowths join into a narrow cephalic ring in the first third of the cephalic capsule. A strip of fine sclerotized granules occurs immediately behind the capsule. Large photosensitive eyes with lens also present. Spicules with small capituli; curve sharply in their anterior third. Velum situated on ventral margin of spicules. Gubernaculum well developed, especially in distal part; process directed dorsally at an acute angle to body of spicules.

Key to Species of Genus Triceratonema gen. nov.

- 1 (2). Lobes of cephalic capsule with isolated large perforations; margins smooth. Strip of sclerotized granules situated beyond cephalic capsule. Spicules curve sharply in proximal third.....
**T. campbelli** (Ditlevsen, 1922).
- 85 2 (1). Lobes of cephalic capsule with numerous minute perforations; margins highly sinuate. Sclerotized granules behind cephalic capsule absent. Spicules curve smoothly.....
**T. microlobatum** (Allgen, 1947).

6. Genus *Deontostoma* Filipjev, 1916

Marion, 1870: 27 (*Thoracostoma*); Linstow, 1892: 61; 1903: 276 (*Leptosomatium*); Savel'ev, 1912: 124 (*Thoracostoma*); Filip'ev 1916: 73; Platonova, 1962: 201; Hope, 1967: 323.

Type species: *Deontostoma arcticum* Filipjev, 1916.

Extremely large representatives of family Leptosomatidae, 10 to 37 mm long. Head usually truncated in front. Cephalic capsule cylindrical and does not narrow from base to top. All walls uniform in width and length and for this reason capsule appears symmetrical in profile. Cephalic suture may bend, forming marginal grooves. Cephalic lobes either entire or with small perforations of irregular shapes. Cephalic ring usually situated in anterior third of capsule. Nerve ring encircles esophagus in anterior third of cephalic capsule. Labial papillae massive and wide. Cephalic and cervical setae short; anal setae of male significantly longer. Photosensitive eyes present in all species of the genus. Vulva median or slightly posteromedian.

Labia of vulva may be unequal in width or protrude over body surface to various degrees. Spicules characteristic, rather uniformly curved in proximal part, and devoid of sharp grooves and angles. Gubernaculum with well-expressed dorsal and ventral processes joined by a curved plate. After fusing, halves of gubernaculum form a sort of funnel in which the distal part of the spicules lie. Twenty pairs of preanal papillae, some supplied with setae, situated anterior to anal opening in male. Accessory organ of male, or rather its sclerotized part, differs in structure in different species of the genus. In some cases this part is tubular or resembles a pivotal needle, while in others it is cyathiform.

In the description of species of genus *Deontostoma* the figures in Cobb's formula denote the distance of the following parts of the body from the anterior end: 1. posterior end of cephalic capsule; 2. eyes; 3. nerve ring; 4. base of esophagus; 5. commencement of anterior genital tube; 6. vulva; 7. commencement of posterior genital tube; and 8. anus.

Measurements are fewer for males than for females (see description of genus *Leptosomatium*).

Key to Species of Genus Deontostoma

- 1 (4). Interlobular grooves not divided into furrows and fenestrae; open widely posteriorly.
- 2 (3). Spicules with well-developed capituli and numerous protuberances.....**D. montredonense** (Marion, 1870).
- 3 (2). Spicules without distinctly defined capituli and protuberances.**D. zeae** (Inglis, 1964).
- 4 (1). Interlobular grooves distinctly divided into furrows and fenestrae.
- 5 (6). Four pairs of large rhomboid processes situated anterior to accessory organ. **D. californicum** (Steiner and Albin, 1933).
- 6 (5). Rhomboid processes anterior to accessory organ absent.
- 7 (8). Section of preanal region in male densely covered with papillate setae.....**D. papillosum** (Ditlevsen, 1922).
- 86 8 (7). Preanal region in male devoid of setae.
- 9 (26). Pigmented eyes present.
- 10 (13). Two pairs of pigmented eyes present: one pair distinct compact structures and second in shape of spots with vague contours.
- 11 (12). Esophageal lumen dilates in upper part of cephalic capsule. 22. **D. lobatum** (Steiner, 1916).
- 12 (11). Esophageal lumen without dilatation in upper part of cephalic capsule.....**D. pacifica** (Murphy, 1965).
- 13 (10). Only single pair of pigmented eyes present.
- 14 (21). Gubernaculum always with a ventrally directed process.

- 15 (18). Spicules with velum.
 16 (17). Accessory organ resembles a sucker.....
 **D. auclandiae** (Ditlevsen, 1922).
 17 (16). Accessory organ cyathiform with processes directed anteriorly
 and posteriorly..... **D. antarcticum** (Linstow, 1892).
 18 (15). Spicules without velum.
 19 (20). Cephalic capsule very wide and short; width twice length.....
 **D. washingtonensis** (Murphy, 1965).
 20 (19). Cephalic capsule much narrower; width exceeds length not more
 than 1.5 times..... **D. magnificum** (Timm, 1951).
 21 (14). Gubernaculum without ventrally directed process.
 22 (23). Small hollow spine situated in middle part of gubernaculum..
 21. **D. arcticum** (Saveljev, 1912).
 23 (22). Spine in gubernaculum absent.
 24 (25). Spicules with velum; preanal papillae in male thick and button-
 shaped..... 20. **D. papillatum** (Linstow, 1903).
 25 (24). Spicules without velum; preanal papillae in male normal in
 structure..... **D. jae** (Inglis, 1964).
 26 (9). Pigmented eyes absent..... **D. demani** (Mawson, 1956).

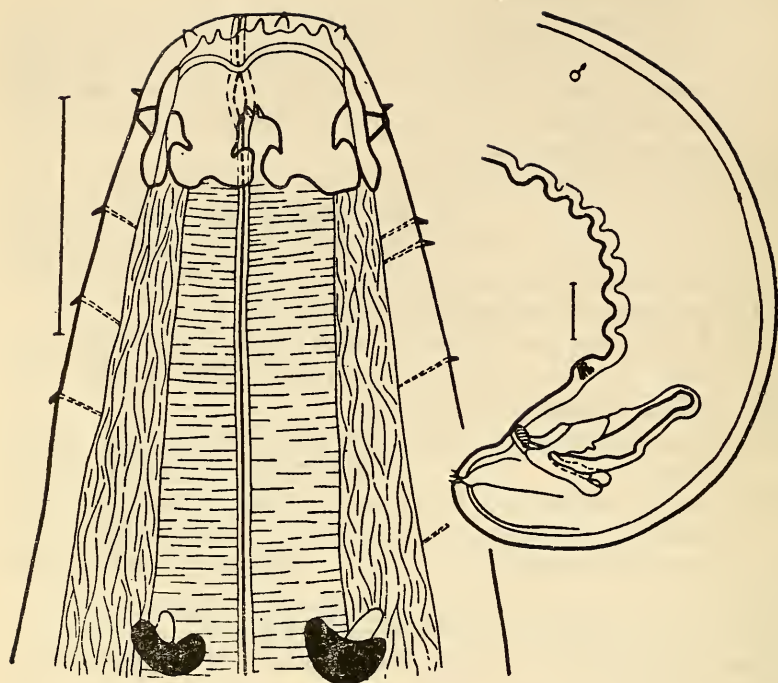
20. *Deontostoma papillatum* (Linstow, 1903) (Figure 33)

Linstow, 1903: 276, tab. 18, figs. 9, 10 (*Leptosomatum*); Filip'ev, 1916:
 79-82, tab. IV, fig. 5.

$$1 \text{ } \delta: \frac{48 \quad 153 \quad 675 \quad 2,050 \quad \text{---} \quad 17,900}{48 \quad 76 \quad 116 \quad 124 \quad 137 \quad 150 \quad 150} \quad 18,000 \text{ } \mu\text{m}; a=120;$$

$$b=8; c=180.$$

Body tapers to 1/3 midbody diameter at anterior end, but does not taper toward posterior end. Cuticle thick, 11.6 μm . Cephalic capsule tapers 1.5 times from base to top. Cephalic ring wide and situated at apical part of cephalic capsule. Cephalic lobes without perforations; margins sinuate and form shallow grooves 23.2 μm deep. Labial papillae wide, tapering slightly upwards. Cephalic and cervical setae extremely short and thin (length 1.32 and 3.45 μm respectively). Eyes situated three times length of cephalic capsule from anterior end. Pigment bowl 5.8 $\mu\text{m} \times 11.6 \mu\text{m}$ and lens 8.1 $\mu\text{m} \times 4.6 \mu\text{m}$. Spicular apparatus consists of two wide massive spicules of equal length (215 μm). Maximum width of spicules 75 μm and minimum 25 μm . Gubernaculum 115 μm long, with two processes—a wide and massive ventral one and a narrower dorsal one. Both processes closely apposed to body of spicules. Accessory organ with tubular sclerotized base situated at a distance of 100 μm anterior to anus. Seven pairs of preanal papillae situated anterior to accessory organ.



87

Figure 33. *Deontostoma papillatum*.

Distinguishing features of this species: 1) spicules extremely wide, massive, and encircled by two powerful processes of gubernaculum in distal part; 2) presence of seven pairs of very well-defined preanal papillae projecting on body surface in shape of buttons (species so-named because of this feature); 3) cephalic lobe of capsule without punctations but highly sinuate contours; and 4) cephalic ring wide and situated in apical part of cephalic capsule.

Geographic distribution. Known only from the Barents Sea.

21. *Deontostoma arcticum* (Saveljev, 1912) (Figure 34)

Filip'ev, 1916: 75-79, tab, IV, fig. 4; Schuurmans-Stekhoven, 1946: 15, fig. 1a-d. *Thoracostoma*—Savel'ev, 1912: 124; Allgen, 1933: 496; Wieser, 1953c: 28, fig. 11a-g; Mawson, 1956: 46, 47, fig. 6a-d; 1958b: 317, 318.

| | | | | | |
|-----|--|---------|---------|-------------|-----|
| 2♂: | 46-52 | 175-185 | 765-882 | 2,516-2,942 | — |
| | 52 71-75 | 104-112 | 187-192 | 262-362 | 370 |
| × | $\frac{24,832-27,825}{190-200}$ 25,000-28,000 μm ; a = 74-77; b = 9; c = 154-160. | | | | |

| | | | | | |
|---------|--|---------------|---------------|---------------|---------------|
| 2 ♀: | 41-46 | 162 | 721-787 | 2,575 | 11,845-12,360 |
| | 46 69-89 | 137 | 175-187 | 250 | — |
| | 16,274 | 19,982-20,394 | 26,883-27,017 | 27,033-27,192 | μm; |
| × | 300-375 | — | 165-200 | | |
| | a = 72-91; b = 10; c = 155-180; V = 59-60%. | | | | |
| 2 juv.: | 34 | 137-162 | 1,854-1,957 | — | 10,868-11,870 |
| | 34 52 | 75 | 185-187 | 200 | 112-137 |
| | × 11,000-12,000 μm; a = 57-60; b = 6; c = 83-92. | | | | |

Body tapers to 1/5 midbody diameter at anterior end and not less than 2/3 at posterior end. Cuticle thick, 10.4 to 14.0 μm. Cephalic capsule narrows to half from base upwards. Cephalic ring situated in first third of cephalic capsule. Cephalic lobes entire. Cephalic suture sinuate and forms small marginal grooves. Shape of interlobular grooves varies significantly (Figure 34). Sometimes they consist of wide fenestrae, the contours of which form offshoots below and long narrow furrows; in other cases contours of fenestrae more irregular and they themselves change into narrow but short furrows. Depth of interlobular grooves 23.2 to 26.6 μm. Labial papillae short and massive. Length of cephalic setae 4.6 to 5.8 μm (1/10 to 1/15 corresponding diameter) and of cervical papillae 2.3 to 5.8 μm. Anal setae of male also very short. Eyes situated at a distance of 3.5 to 4.0 times width of cephalic capsule. Size of pigment bowl 10.6 to 17.4 μm × 10.6 to 15.8 μm. Large eggs, 618 to 206 μm, seen in uterine tubes of female; three eggs found in anterior branch and one in posterior branch. Vulva slightly posterior to midbody and located in small depression; upper lip projects over lower one. Extremely small sclerotized granules situated around vulva on cuticular surface. Spicular apparatus consists of a pair of spicules with capituli differing in shape (Figure 34). Length of spicules 200 μm, maximum width 25 to 30 μm, and minimum 12.5 to 15.0 μm. Gubernaculum funnel-shaped and envelopes spicules at their distal end. Length of gubernaculum 100 to 125 μm and its dorsal process 30 μm. Hump-shaped accessory organ with sclerotized tube passing through it, situated 212 to 225 μm anterior to anus. Twenty-one to twenty-three pairs of preanal papillae bearing short and thin setae situated anterior to accessory organ.

This species is very close to *Deontostoma antarcticum* in structure but differs from the latter as follows: 1) lobes of cephalic capsule not perforated; 2) cephalic suture forms marginal grooves; 3) spicules without ventral velum; 4) shape of gubernaculum with ventral part much more massive and merging with dorsal part to form a more compact and narrower funnel; 5) accessory organ tubular in shape; and 6) vulva situated in depression, with prominent upper lip, and surrounded by minute sclerotized granules.

Nematodes classified under this species by Schuurmans-Stekhoven

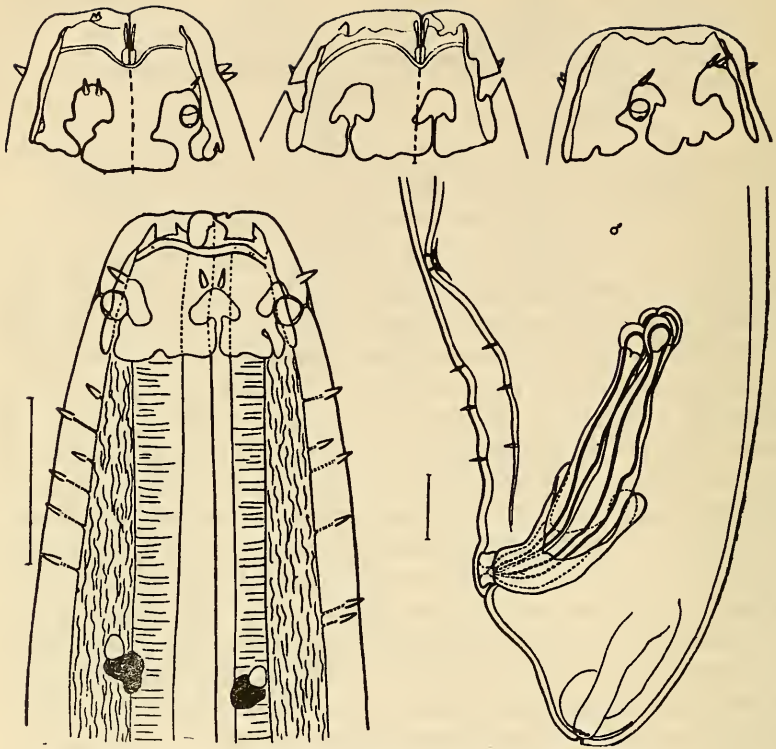


Figure 34. *Deontostoma arcticum*.
(Variability of interlobular grooves of cephalic capsule.)

(1946) differ significantly from *D. arcticum* in absence of eyes, structure of cephalic capsule, and shape of amphids. It is regretted that in the material of Schuurmans-Stekhoven there were no males, making identification very difficult. Nevertheless the form should probably be considered an independent species.

Geographic distribution. Bipolar. Found in the Barents Sea (in littoral zone among brown algae), Norwegian Sea, and North Sea, and also in Antarctic and subantarctic waters.

*22. *Deontostoma lobatum* (Steiner, 1916) (Figure 35)

Steiner, 1916: 620-623, tab. 16, fig. 29a, tab. 31, fig. 29b (*Thora-costoma*) (description).

L=6,196 μ m; a=45; b=5; c=57.

Body covered with thick cuticle. Cephalic capsule almost does not narrow from base to top. Cephalic lobes without punctations and their

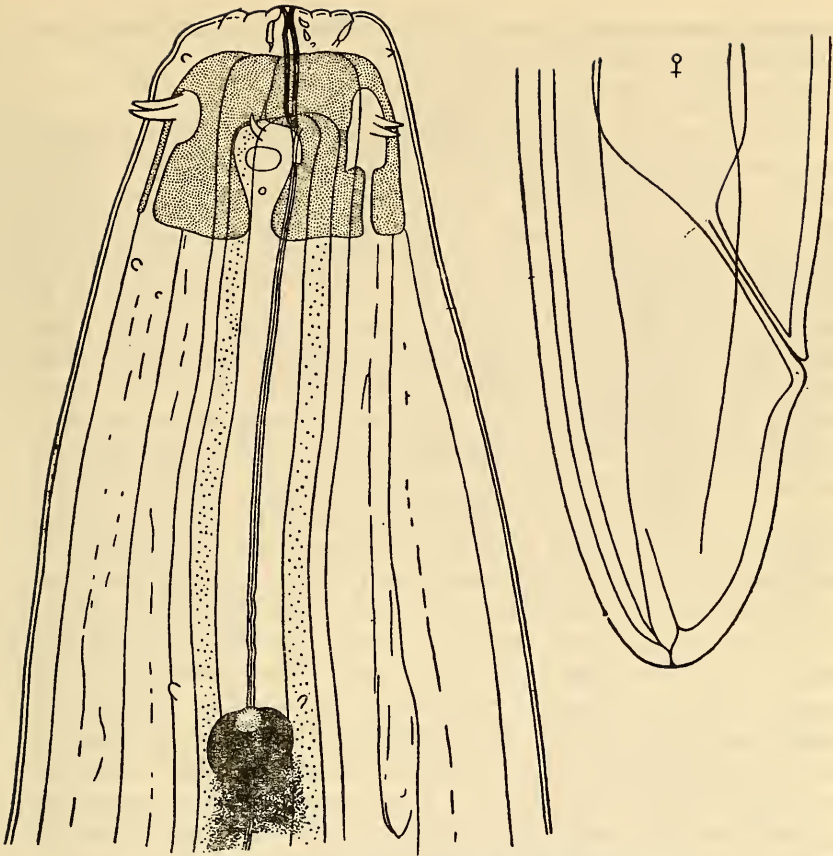


Figure 35. *Deontostoma lobatum* (from Steiner, 1916).

90 margins smooth (devoid of marginal grooves). Interlobular grooves with narrow furrows and rounded fenestrae. Labial papillae situated low, at level of upper margin of capsule. Cephalic setae short and wide. Cervical setae also short, occasionally dispersed throughout anterior part of body, mainly between anterior end and eyes. In addition to one pair of eyes, with pigment bowl and lens, another pair of irregular pigment spots are situated at a significant distance posterior to first pair. Esophageal lumen with dilatation in anterior part; walls of dilatation with a depression, opposite to which lies a dentoid process. Pores of esophageal glands open near dentoid process.

Geographic distribution. Barents Sea.

7. Genus *Pseudocella* Filipjev, 1927

Leuckart, 1849: 149 (*Hemipsilus*) (in part); Schneider, 1866: 58 (*Enoplus*) (in part); Bütschli, 1875: 42 (*Thoracostoma*) (in part); Filip'ev,

1916: 82 (*Thoracostoma*) (in part); 1927: 93; Wieser, 1954: 175; Platonova, 1962: 204; Hope, 1967: 309.

Type species. *Hemipsilus trochodes* Leucart, 1849.

Body 6.0 to 29.0 mm long. Cephalic capsule in the shape of a slightly truncated cone, distinctly narrowing from base to top. Ventral wall of cephalic capsule significantly well developed in length and thickness; hence in lateral view of capsule ventral wall looks like a stout, blunt protuberance directed anteriorly, and consequently capsule appears asymmetrical. Cephalic ring may differ in width but usually situated at posterior limit of first third of cephalic capsule. Band of sclerotized granules may lie posterior to capsule. Nerve ring encircles esophagus in its anterior third or fourth part. Labial papillae small but distinct. Cephalic and cervical setae longer than in representatives of genus *Deontostoma*. Characteristic peculiarity of the genus: absence of clearly expressed eyes provided with light-refracting lens. Instead of eyes, either a compact pigment spot present or, more often, photosensitive pigment dispersed in the form of large irregular spots all over anterior end of esophagus. In some species photosensitive pigment absent. Vulva usually situated mid-body; lips equal in width. Spicular apparatus differs considerably from that of genus *Deontostoma*. Spicules regularly arcuate and always with small but distinctly defined capituli and neck. Gubernaculum with two large processes of rectangular shape directed backward at right angles to longitudinal axis of spicules. In some species of this genus asymmetry in the development of the spicular apparatus is apparent, with one spicule significantly shorter than the other. Processes of the gubernaculum also asymmetrical. Anal setae and papillae of male vary considerably in number. Sometimes pretubal papillae absent but in some species even accessory organ absent.

In the description of species of genus *Pseudocella* given below, the figures in Cobb's formula denote the distance of the following parts from the anterior end: 1. posterior end of cephalic capsule; 2. nerve ring; 3. base of esophagus; 4. commencement of anterior genital tube; 5. vulva; 6. commencement of posterior genital tube; and 7. anus.

Figures are fewer for males than for females (see p. 74).

Key to Species of Genus Pseudocella

- 1 (2). Cervical setae absent.....**P. obliqua** (Ditlevsen, 1926).
- 2 (1). Cervical setae present.
- 91 3 (4). Interlobular grooves undeveloped....**P. filipjevi** (Kreis, 1928).
- 4 (3). Interlobular grooves well developed.
- 5 (12). Subdorsal and subventral interlobular grooves with distinct furrows.

- 6 (7). Subdorsal and subventral interlobular grooves narrow near posterior margin of cephalic capsule. . . . **P. tabarini** (Inglis, 1958).
- 7 (6). Subdorsal and subventral interlobular grooves do not narrow near posterior margin of cephalic capsule.
- 8 (9). Tail long, with pointed digitiform process at end. **P. citronicauda** (Wieser, 1954).
- 9 (8). Tail short and obtusely rounded.
- 10 (11). Cephalic capsule tapers distinctly from base to top; subdorsal and subventral interlobular grooves broaden posteriorly and form a triangle. **P. polychaites** Mawson, 1958.
- 11 (10). Cephalic capsule does not taper toward top; subdorsal and subventral interlobular grooves do not form an abrupt dilatation. **P. kreisi** (Wieser, 1953).
- 12 (5). Subdorsal and subventral interlobular grooves with well-defined furrows.
- 13 (24). Apophysis present on ventral aspect of tail of males.
- 14 (15). Process on tail of males resembles a bursa and provided with papillae. **P. bursata** Platonova, 1962.
- 15 (14). Apophysis on tail of males differs in structure; looks like a blunt protuberance.
- 16 (19). Cephalic ring narrow and situated in middle part of cephalic capsule.
- 17 (18). Large sclerotized granules present posterior to cephalic capsule. Photosensitive pigment merges into a single large spot. **P. truncaticauda** sp. nov.
- 18 (17). Sclerotized granules absent behind cephalic capsule. Photosensitive pigment dispersed in isolated small spots. **P. trichodes** (Leucart, 1849).
- 19 (16). Cephalic ring wide and situated in upper part of cephalic capsule.
- 20 (23). One unpaired accessory organ present.
- 21 (22). Lateral interlobular grooves with additional notches along margin which look like narrow slits. Amphids round. **P. mamillifera** Platonova, 1962.
- 22 (21). Lateral interlobular grooves with wide additional notches in upper part of margin. Amphids longitudinally elongated. **P. rarisetosa** sp. nov.
- 23 (20). In addition to unpaired accessory organ, pair of organs situated anterior to former. **P. angusticeps** sp. nov.
- 24 (13). Apophysis absent on ventral aspect of tail of males.
- 25 (26). Narrow longitudinal furrows extend along cephalic lobes. **P. panamaense** (Allgen, 1947).
- 26 (25). Longitudinal furrows absent on cephalic lobes.
- 27 (28). Cervical setae commence above level of cephalic suture, more or

- less from middle of amphids. **P. wieseri** Hope, 1967.
- 28 (27). Cervical setae commence below level of cephalic suture.
- 29 (38). Fenestrae in subdorsal and ventral interlobular grooves circular.
- 30 (31). Cephalic setae extremely short; length does not exceed one-tenth of corresponding diameter. . . . **P. brachychaites** Mawson, 1958.
- 92 31 (30). Cephalic setae significantly long; length not less than one-fourth of corresponding diameter.
- 32 (33). Small pointed onchium present on dorsal side of oral cavity. 29. **P. pseudocellum** (Filipjev, 1916).
- 33 (32). Onchia absent in oral cavity.
- 34 (35). Spicules unequal in length; right spicule longer by one-fourth than left. **P. cavernicola** (Wieser, 1954).
- 35 (34). Spicules equal in length.
- 36 (37). Photosensitive pigment totally absent. Spicules with small velum. 34. **P. acuta** sp. nov.
- 37 (36). Photosensitive pigment present. Spicules without velum. 38. **P. raddae** sp. nov.
- 38 (29). Fenestrae of subdorsal and subventral interlobular grooves anchor-shaped.
- 39 (46). Gubernaculum funnel-shaped, highly developed on all sides, and encloses distal part of spicules.
- 40 (41). Capituli of spicules rectangular. 28. **P. gracilis** sp. nov.
- 41 (40). Capituli of spicules round.
- 42 (43). Anchor-shaped additional cleft present above lateral interlobular grooves. 29. **P. tenuis** Filipjev, 1946.
- 43 (42). Additional cleft above lateral interlobular grooves absent.
- 44 (45). Body extremely slender; $a=95$ 25. **P. saveljevi** (Filipjev, 1927).
- 45 (44). Body significantly wide; $a=35$ to 56. 24. **P. elegans** (Ditlevsen, 1926).
- 46 (39). Gubernaculum not funnel-shaped, and does not enclose distal part of spicules.
- 47 (48). Gubernaculum small; dorsal process not developed. 31. **P. minor** Platonova, 1962.
- 48 (47). Gubernaculum large; dorsal process well developed.
- 49 (50). Cephalic ring narrow and situated in middle of cephalic capsule. 27. **P. coecum** (Saveljev, 1912).
- 50 (49). Cephalic ring wide and situated in upper part of cephalic capsule. 32. **P. kurilensis** Platonova, 1962.

23. *Pseudocella trichodes* (Leucart, 1849) (Figure 36)

Leucart, 1849: 149, tab. 3, figs. 1, 2, (*Hemipsilus*); Schneider, 1866; 58 (*Enoplus denticaudatus*); Bütschli, 1874: 42, tab. 8, fig. 33a-d; Savel'ev,

1912: 126; de Man, 1922: 251, Fig. 42a-b (*Thoracostoma schneideri*); de Man, 1888: 22, tabs. 2-3, figs. 12-12b; Linstow, 1900: 126, tab. 7, figs. 36-37; Southern, 1914: 39; Filip'ev, 1916: 88, tab. 4, fig. 7; Ditlevsen, 1919: 181 (*T. denticaudatum*); Filip'ev, 1927: 94-97, pl. I, fig. 12a-b; Kreis, 1928: 147; Allgen, 1935: 16; Schuurmans-Stekhoven, 1935a: 12 (*T. trichodes*), fig. 75; Allgen, 1957b: 8; 1959: 22, fig. 6a-b; Gerlach, 1958: 85; Steiner, 1916: 623, tab. 17, fig. 30a-b (*Thoracostoma* sp.); Platonova, 1967: 832.

| | | | | | |
|----------|--|---------|-------------|------------------------------------|-------------|
| 15 ♂: | 29-40 | 360-494 | 1,080-1,390 | — | 6,313-7,777 |
| | 20-23 | 46-52 | 112-125 | 155-167 | 205-207 |
| | × 6,393-7,948 μm; a=32-38; b=4-6; c=39-53. | | | | |
| 15 ♀: | 29-38 | 412-484 | 1,184-1,411 | 2,420-3,829 | |
| | 22-23 | 46-52 | 112-130 | 137-167 | 187-207 |
| | × 4,119-5,631 5,921-7,436 6,693-9,236 6,843-9,473 μm; | | | | |
| | 175-200 | 180-212 | 105-130 | a=33-57; b=6-7; c=60-90; V=56-61%. | |
| 15 juv.: | 24-39 | 309-463 | 844-1,338 | — | 3,522-6,797 |
| | 17 | 39-42 | 100-125 | 112-167 | 130-137 |
| | × 3,647-6,922 μm; a=28-46; b=4-5; c=36-69. | | | | |

- 93 Body rather elongated and tapers to 1/3 to 1/4 midbody diameter at anterior end, but almost does not taper posteriorly. Cuticle relatively thick, 8.1 to 10.0 μm. Tail rather short, rounded, and curves ventrally in males. Dentoid projection present on ventral aspect of tail. Cephalic capsule usual shape for this genus, tapering from base upward to half its length. Extremely narrow cephalic ring situated in anterior third of cephalic capsule. Lobes of cephalic capsule entire and without perforations. Small and narrow folds present on surface of lobes. Marginal grooves absent. Sublateral interlobular grooves resemble narrow furrows and dilate into wide round fenestrae. Amphids slightly elongated longitudinally; width 9.2 μm. Photosensitive pigment situated posterior to cephalic capsule at a distance somewhat less than its length. Longitudinal bands of pigment extend further. Labial papillae well defined. Length of cephalic setae 5.8 to 8.1 μm, and cervical 3.4 to 4.6 μm. Anal setae of male vary significantly in length: preanal setae 4.0 μm, and postanal 12.5 μm. Extremely small setae situated along margin of caudal pore. Nerve ring situated at end of first third of esophagus. Length of anterior female genital tube 1,493 to 2,060 μm, of posterior tube 1,648 to 2,369, and of reflexed parts 721 to 1,030 μm and 824 to 1,236 μm respectively. One to four eggs, 206 to 412 μm × 103 to 123 μm, seen in uteri. Spicules unequal in size and shape. Usually left spicule (187 to 200 μm) longer than right one (172 to 175 μm). Left spicule with larger capitulum, tapering into a significantly narrower body; blunt process occurs in upper part of spicu-

lar body. Right spicule more regular in form; narrow capitulum tapers into small cervix, which broadens into spicular body. Velum attached to spicules from ventral side. Gubernaculum paired; 32 to 40 μm long. Accessory organ 112 to 150 μm anterior to anus. Eight to ten pairs of preanal papillae situated anterior to accessory organ.

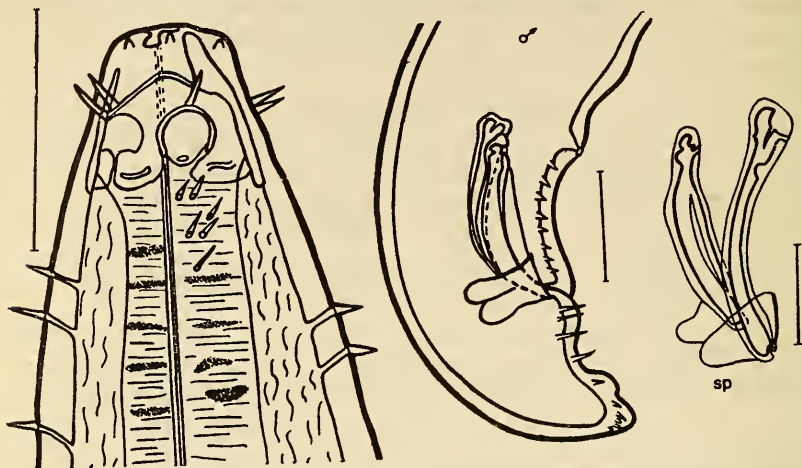


Figure 36. *Pseudocella trichodes*.

Distinguishing features of this species: 1) folds on lobes of cephalic capsule; 2) extremely narrow cephalic ring; 3) absence of marginal grooves in lobes of cephalic capsule; 4) characteristic form of interlobular grooves; 5) great differences in length of pre- and postanal setae in male; 94 6) characteristic structure of spicules, clearly differing from each other in shape and size; and 7) presence of dentoid process on ventral aspect of tail of males.

Geographic distribution. Found mainly in the seas of the Arctic basin, North Sea, and along western part of the Baltic Sea. Inhabits mainly littoral zone among brown algae. I do not accept Allgen's view that this species is ubiquitous (Allgen, 1957b). Even if the two findings of this species in the southern hemisphere on the southern shores of Australia and Argentina are taken into account (Allgen, 1951, 1959), this species can easily be considered bipolar in distribution.

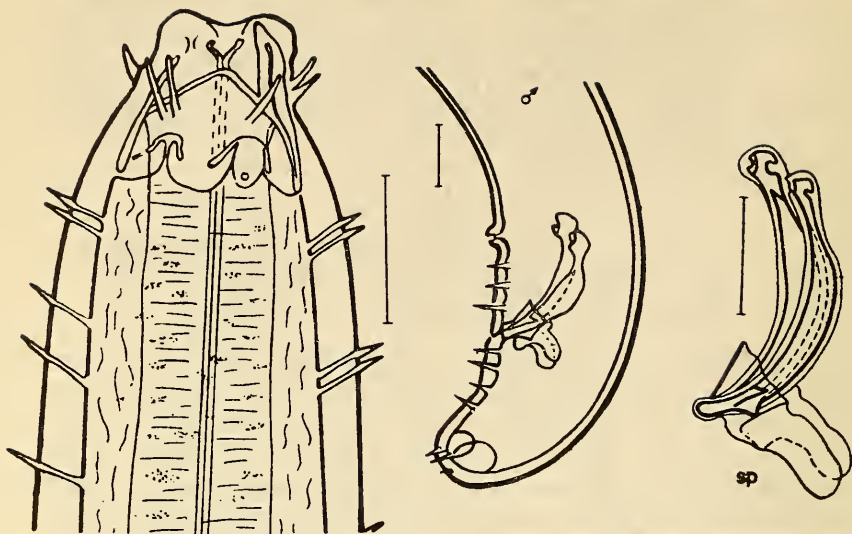
24. *Pseudocella elegans* (Ditlevsen, 1926) (Figure 37)

Ditlevsen, 1926: 25, tab. 5, figs. 3, 5, tab. 6, fig. 3, tab. 7, figs. 3-4 (*Thoracostoma*); Schuurmans-Stekhoven, 1935a: 14, fig. 76A-B; 1946: 17-19, fig. 2A-F; Filip'ev, 1946: 159; Allgen, 1957b: 8; Platonova, 1967: 832.

| | | | | | |
|----------|---|---------|-------------|-------------|-------------|
| 9 ♂: | 29-34 | 408-485 | 1,386-1,566 | — | 6,103-8,003 |
| | 20-26 | 42-46 | 95-132 | 112-175 | 130-200 |
| | 112-137 | | | | |
| | × 6,237-8,203 μm; a = 37-56; b = 4-5; c = 32-46. | | | | |
| 10 ♀: | 29-34 | 410-669 | 1,131-1,693 | 3,191-3,914 | 4,500-5,807 |
| | 20-25 | 42-51 | 112-142 | 150-188 | 167-225 |
| | 162-215 | | | | |
| | × $\frac{6,045-7,867}{125-137}$ $\frac{7,311-9,618}{125-140}$ 7,486-9,793 μm; a = 36-51; b = 5-6; | | | | |
| | c = 42-58; V = 50-63.5%. | | | | |
| 10 juv.: | 25-32 | 332-459 | 890-1,442 | — | 4,120-7,412 |
| | 19-23 | 34-46 | 72-112 | 100-162 | 112-188 |
| | 71-125 | | | | |
| | × 4,232-7,562 μm; a = 38-42; b = 5; c = 38-51. | | | | |

Body average in size, tapers to 1/7 to 1/10 midbody diameter at anterior end and almost does not taper toward posterior end. Cuticle 9.28 to 15.0 μm thick. Lobes of cephalic capsule rounded, sometimes with small number of fenestrae. Margin of cephalic capsule smooth, does not form peripheral (marginal) grooves. Depth of interlobular grooves 9.3 to 15.0 μm. Sublateral interlobular grooves narrow, anchor-shaped, while lateral interlobular grooves rounded and wide, with slit-shaped furrow on upper part. Amphid, 8.1 to 11.6 μm × 4.6 to 8.1 μm, situated in cavity of each interlobular groove. Ditlevsen (1926) in describing this species mistook the slit-shaped furrows in the upper part of the lateral grooves for horse-shoe amphids; hence he proposed the transference of this species to another genus. Sensory organs represented by cephalic setae 6.9 to 8.1 μm long, cervical setae 5.8 to 6.9 μm, and in males anal setae 12.5 to 15.0 μm. Photosensitive pigment dispersed throughout anterior body end; does not form small or large clusters. Length of anterior female genital tube 1,545 to 2,060 μm, of posterior tube 1,442 to 1,854, and of reflexed parts 824 to 1,030 μm and 824 to 1,080 μm respectively. One to three eggs, 204 to 357 μm × 102 to 153 μm, found in uterus. Length of anterior testis with seminal vesicles 515 to 721 μm and posterior testis 597 to 927 μm. Spicules unequal in length, 125 and 200 μm respectively. Either left or right spicule may be longer. Spicules equal in length exceptionally. Round capitulum tapers rather abruptly into narrow cervix on proximal end of spicule; midlength, spicules dilate again. Gubernaculum paired; structures fuse to form a funnel which encloses spicular bodies. Small velum present on ventral surface of spicules, and two processes, 30 to 40 μm long, present on dorsal surface. Accessory organ 87.5 to 125 μm anterior to anus; five to six pairs of papillae situated anterior to accessory organ.

Distinguishing features of this species: 1) rather small length in view of significant width; 2) sublateral grooves anchor-shaped; 3) absence of marginal grooves; 4) spicules unequal in length, with distinctly expressed round capitulum; and 5) gubernaculum paired; structures fuse and form a



94

Figure 37. *Pseudocella elegans*.

funnel enclosing spicular bodies.

Geographic distribution. Arctic-boreal species. Found in the Kara Sea (in depths ranging from 300 to 400 m in sandy silt), Greenland Sea, and Baltic Sea. One sexually immature specimen of this species Ditlevsen claims he collected in the subantarctic (Lake Campbell); this find appears dubious.

25. *Pseudocella saveljevi* (Filipjev, 1927) (Figure 38)

Filip'ev, 1927: 97, 98, pl. 1, fig. 13a-c (*Thoracostoma*); Kreis, 1928: 143-145, tab. 1, fig. 3 (*T. conicaudatum*); Platonova, 1967: 832.

$$1 \text{ ♂: } \frac{30}{27} \frac{250}{40} \frac{1,240}{68} \frac{—}{75} \frac{7,250}{77} \frac{7,360}{88} \mu\text{m}; a=95; b=6; c=67.$$

Body average in size, tapers to 1/3 midbody diameter at anterior end, and slightly broadens toward posterior end due to development of male copulatory apparatus. Cuticle 5.0 to 6.0 μm thick. Cephalic capsule rather short and broadens slightly toward base. Relatively wide cephalic ring situated in uppermost part of cephalic capsule. Lobes of cephalic capsule with narrow slit-shaped perforations. Median lobes with deep marginal grooves. Subventral and subdorsal interlobular grooves anchor-shaped. Narrow horseshoe-shaped cleft visible in upper part of wide lateral grooves. Band of extremely small sclerotized granules situated behind cephalic capsule. Labial papillae small. Length of cephalic and

cervical setae $7.0 \mu\text{m}$ and anal setae $12.5 \mu\text{m}$. Photosensitive pigment dispersed throughout anterior end of body. Nerve ring situated at border of first quarter of esophagus. Spicules unequal in length, left $105 \mu\text{m}$, and right $112 \mu\text{m}$, uniformly curved, with capituli of different shape. Gubernaculum paired; structures fuse and form funnel enveloping spicular bodies. Length of gubernacular process $65 \mu\text{m}$. Accessory organ $65 \mu\text{m}$ anterior to anus. Four pairs of preanal papillae situated anterior to accessory organ. Ten pairs of anal setae present in anal region.

This species is close to *Pseudocella coecum* in structure but differs from it in: 1) significantly small size; 2) structure of cephalic capsule: presence of slit-shaped punctations on lobes of cephalic capsule, presence of marginal grooves on median lobes, presence of small sclerotized granules behind cephalic capsule, and extremely wide cephalic ring; and 3) number of preanal papillae (four pairs versus eight to nine pairs in *P. coecum*).

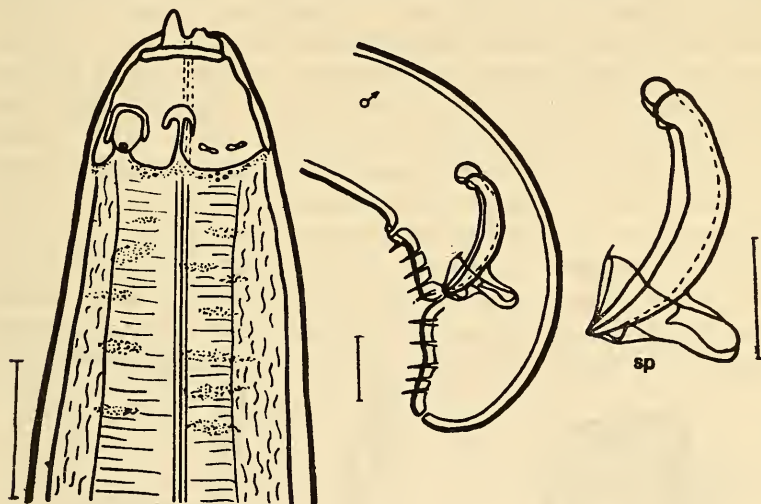


Figure 38. *Pseudocella saveljevi*.

Geographic distribution. Only isolated finds from the Barents Sea.

26. *Pseudocella tenuis* Filipjev, 1946 (Figure 39)

Filip'ev, 1946: 159-160, fig. 3a-c; Platonova, 1967: 832.

| | | | | | | |
|-------|--|---------|-------------|-------------|--------|-------------|
| 9 ♂: | 29-34 | 375-459 | 1,199-1,545 | — | — | 6,059-9,424 |
| | 20-23 | 38-48 | 70-105 | 87-125 | 85-175 | — |
| | × 6,171-9,574 μm ; a = 69-83; b = 5-6; c = 40-65. | | | | | |
| 10 ♀: | 23-32 | 357-669 | 1,236-1,550 | 3,275-4,377 | | × |
| | 19-23 | 37-74 | 73-92 | 80-110 | 87-125 | |

| | | | | | | |
|----------|--|-----------------------|-----------------------|-----------------------------|-------------|--------|
| × | 4,017-3,819 90-122 | 4,532-7,158 95-125 | 6,032-8,909 87-102 | 6,132-9,021 μm ; | | |
| | $a = 67-72$; $b = 5-6$; $c = 59-81$; $V = 58-64\%$. | | | | | |
| 10 juv.: | 23-30 | 341-408 | 1,030-1,438 | — | 3,914-8,240 | |
| | 18-23 | 34-45 | 74-100 | 90-112 | 100-130 | 82-100 |
| | $\times 4,014-8,340 \mu\text{m}$; $a = 39-64$; $b = 4-6$; $c = 40-83$. | | | | | |

Body average in size, tapers to 1/3 midbody diameter at anterior end and almost does not taper toward posterior end. Cuticle $8.2 \mu\text{m}$ (5.8 to $10.4 \mu\text{m}$) thick. Cephalic capsule highly similar to that of *Pseudocella elegans* but notably narrower and appears longitudinally stretched. Subdorsal and subventral interlobular grooves anchor-shaped. Amphids situated in furrows and not in fenestrae of lateral grooves; hence fenestrae with their outgrowths seem to be situated at a level higher than that of amphids. Interlobular grooves 11 to $15 \mu\text{m}$ deep. Lobes of cephalic capsule round, without marginal grooves. Extremely small number of slightly sclerotized granules located behind cephalic capsule. Cephalic ring narrow and situated in upper part of capsule. Amphids round, longitudinally stretched, and 8.1 to $9.2 \mu\text{m} \times 4.6$ to $5.8 \mu\text{m}$. Cephalic setae rather long, 6.9 to $8.1 \mu\text{m}$, cervical setae 4.6 to $12.5 \mu\text{m}$, and anal setae in male 10.0 to $12.5 \mu\text{m}$. Photosensitive pigment dispersed throughout anterior end of body. Length of anterior female genital tube 669 to $1,545 \mu\text{m}$, of ⁹⁷ posterior tube 824 to $1,339 \mu\text{m}$, and of reflexed parts 412 to $927 \mu\text{m}$ and 515 to $721 \mu\text{m}$ respectively. Two eggs, 255 to $357 \mu\text{m} \times 76$ to $91 \mu\text{m}$, found in each uterus. Length of spicules 100 to $145 \mu\text{m}$. Spicular body without distinct neck or cervix. Accessory organ situated 62 to $100 \mu\text{m}$ anterior to anus. Four to six pairs of papillae situated anterior to accessory organ.

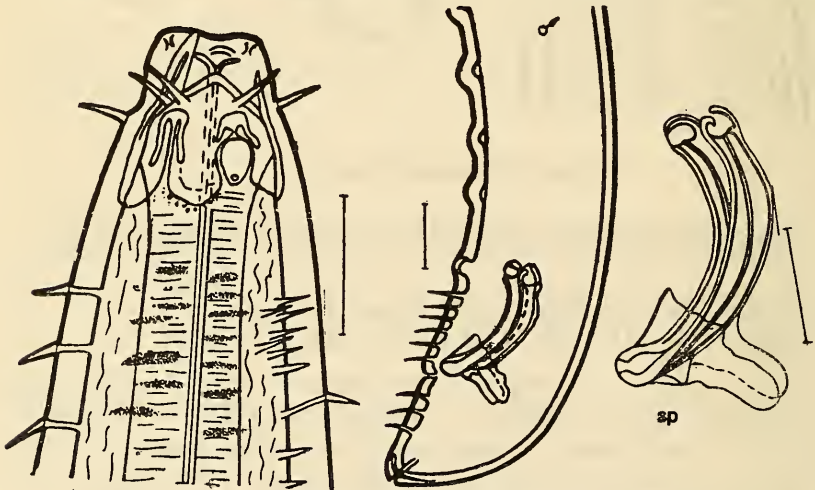


Figure 39. *Pseudocella tenuis*.

This species closely resembles *Pseudocella elegans* but differs from it in the following features: 1) significantly smaller width of body (in *P. elegans* $a=35-56$, in *P. tenuis* $a=68-72$); 2) shape of interlobular lateral grooves (in *P. elegans* wide and without distinctly expressed fenestrae; in *P. tenuis* narrow fenestrae with outgrowths situated above wide furrows); and 3) shape of spicules (curved in *P. elegans* with clearly distinguishable capitulum and neck; in *P. tenuis* long and massive with barely distinguishable capitulum, and neck totally absent).

Geographic distribution. Found only in the Kara Sea in depths ranging from 100 to 360 m, mainly in sandy silt.

27. *Pseudocella coecum* (Saveljev, 1912) (Figure 40)

Savel'ev, 1912: 124 (*Thoracostoma*); Filip'ev, 1927: 98-100, pl. 3, fig. 14a-g (*Thoracostoma*); Platonova, 1967: 832.

| | | | | | | | |
|---------|--------------------------------------|-----------------------|-------------------------------|-------------------------------|-------------------------------|---|----------------------------------|
| 1 ♂: | $\frac{34}{23}$ | $\frac{510}{58}$ | $\frac{1,530}{110}$ | — | $\frac{9,099}{200}$ | $\frac{9,261}{137}$ | μm ; a=48; b=6; c=57. |
| 2 ♀: | $\frac{34-38}{25-29}$ | $\frac{—}{58}$ | $\frac{463}{125}$ | $\frac{1,441-1,493}{172-175}$ | $\frac{4,325-4,480}{175-212}$ | $\frac{5,973-6,437}{187-200}$ | |
| | $\times \frac{7,621-8,394}{172-212}$ | | $\frac{8,859-9,630}{137-140}$ | | $\frac{9,009-9,805}{—}$ | μm ; a=46-48; | |
| | | | | | | b=6; c=56-60; V=66%. | |
| 2 juv.: | $\frac{29-40}{20-23}$ | $\frac{50-52}{50-52}$ | $\frac{112-125}{112-125}$ | $\frac{381-412}{137-162}$ | $\frac{1,153-1,215}{187-192}$ | $\frac{6,457-7,395}{105-125}$ | |
| | | | | | | $\times 6,607-7,515 \mu\text{m}$; a=38-39; b=6; c=44-62. | |

Body average in size, tapers to 1/3 to 1/4 midbody diameter at anterior end at base of cephalic capsule, and almost does not taper toward posterior end. Cuticle $8.1 \mu\text{m}$ thick. Cephalic capsule narrows to 2/5 from base toward middle [*sic*]. Narrow cephalic ring situated in middle of cephalic capsule. Lobes of cephalic capsule with large number of perforations, mostly minute, but in midlobular part merge to form rather large, round or slit-shaped loculi. In addition, lobular margins with thin transverse 97 folds and consequently appear rugulose. Occasionally, extremely fine sclerotized granules present under median lobe. Margins of cephalic lobes form small peripheral grooves. Subdorsal and subventral grooves anchor-shaped and $17.4 \mu\text{m}$ long. Lateral interlobular grooves wide, round, with canal-shaped depression in their upper part. Amphids located in lateral grooves large and only slightly elongated longitudinally. Amphids $9.2 \mu\text{m} \times 6.9 \mu\text{m}$. Labial papillae rather large. Cephalic setae $12 \mu\text{m}$ long, cervical setae $8.0 \mu\text{m}$, and anal setae in males $12.5 \mu\text{m}$. Photosensitive pigment dispersed in spots with irregular outlines. Length of female genital tubes 1,648 to 1,957 μm and reflexed parts 772 to 824 μm . Two to three eggs, 255 to 452 $\mu\text{m} \times 102$ to 125 μm , found in each branch of

uterus. Vulva shifted posterior to midbody (posteromedian in position). Spicules unequal in length; left $212 \mu\text{m}$, right $192 \mu\text{m}$. Gubernaculum with process $62.2 \mu\text{m}$ long and $15.0 \mu\text{m}$ wide at top. Accessory organ $87.5 \mu\text{m}$ anterior to anus. Preanal papillae barely visible in my preparations. Fourteen pairs of setae situated in anal region of male. Extremely small setae also present around outlet of caudal glands in both males and females. Extremely fine, isolated setae likewise present on dorsal surface of tail.

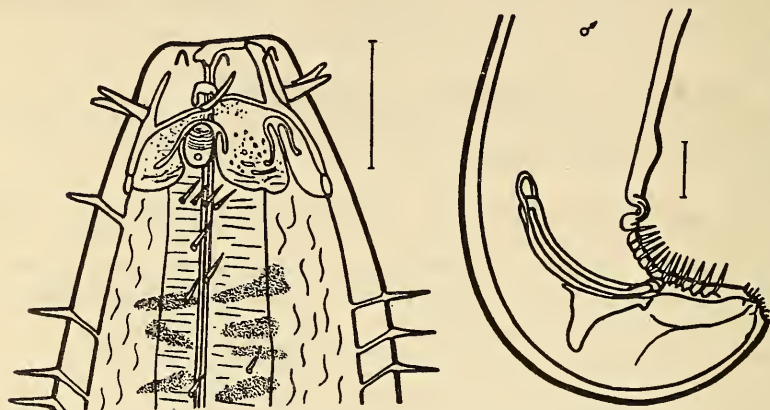


Figure 40. *Pseudocella coecum*.

Distinguishing features of this species: 1) structure of cephalic capsule: lobes with large number of punctations, lobular margins with fine transverse folds, subdorsal and subventral interlobular grooves anchor-shaped with extremely narrow furrows, small peripheral grooves present, and extremely minute sclerotized granules situated posterior to median lobe; 2) spicules unequal in length, rather narrow, smoothly arcuate, with round capitulum; and 3) extremely short setae, situated in groups in region of pore of caudal glands in both male and female. Short setae also present on dorsal aspect of tail.

Geographic distribution. Arctic species. Found in the Barents Sea (mainly in the littoral zone among brown algae), White Sea, and Kara Sea. Specimens primarily found at a depth of 30 m in sandy silt.

99 28. *Pseudocella gracilis* sp. nov. (Figure 41)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 5117.

| | | | | | |
|-------|-----|-------|-----|-------|--|
| 31 | 412 | 1,339 | — | 6,489 | 6,614 μm ; a = 65; b = 5; c = 53. |
| 19 40 | 75 | 87 | 102 | 125 | |

Body tapers to 2/5 midbody diameter at anterior end, and broadens slightly posteriorly in region of spicular apparatus. Cuticle 5.8 μm thick. Cephalic capsule narrows to 1/2 from base toward top. Cephalic ring narrow and situated in anterior third of cephalic capsule. Cephalic lobes with one to two large perforations. Subventral and subdorsal interlobular grooves anchor-shaped and 11.6 μm in depth. Lateral interlobular grooves wide, round, and with canalicular depression in upper part. Amphids located in grooves, elongated longitudinally, and 9.3 $\mu\text{m} \times 5.8 \mu\text{m}$. Labial papillae small. Cephalic setae 9.3 μm ; cervical setae 5.8 μm ; anal setae 12.5 μm . Photosensitive pigment dispersed in small spots throughout anterior part of esophagus. Spicules unequal in length; left 125 μm and right 150 μm long. Capitulum of spicules unique, not rounded as usually seen in *Pseudocella* but truncated at proximal end. Gubernaculum paired; structures fuse to form funnel. Length of gubernaculum processes 30 μm . Accessory organ 87.5 μm anterior to anus. Six pairs of papillae situated anterior to accessory organ, and thirteen pairs situated in anal region.

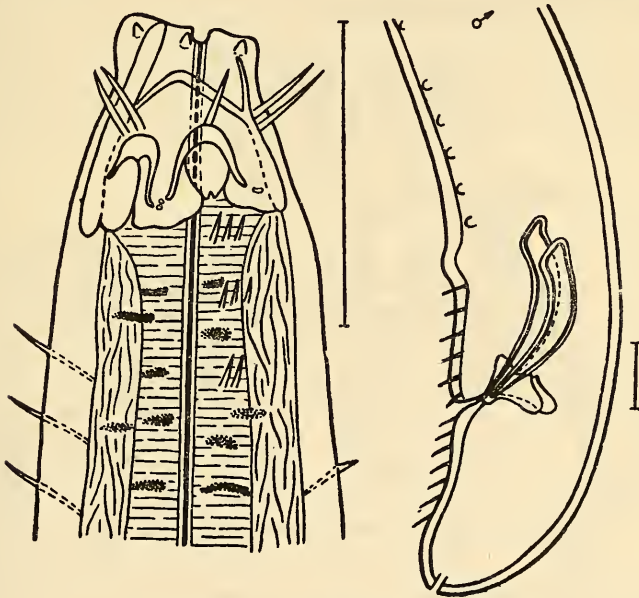


Figure 41. *Pseudocella gracilis* sp. nov.

This species is close to *P. saveljevi* but differs from it in these features: 1) greater body width; 2) larger number of anal papillae and setae; and 3) spicular structure—the primary distinguishing feature: spicules of *P. gracilis* with truncated almost rectangular capitulum, significantly longer, and much more smoothly arcuate than in *P. saveljevi*.

Geographic distribution. Found along the shores of Novaya Zemlya in the Strait of Matochkin Shar at a depth of 12 m in silted sand.

100 *29. *Pseudocella pseudocellum* Filipjev, 1927 (Figure 42)

Filip'ev 1916: 86-88, tab. IV, fig. 6 (*Thoracostoma coecum* nec Savel'ev, 1912) (description): 1927: 94.

1 ♀: L=13; a=57; b=7; c=71.

Body tapers to 1/4 midbody diameter at anterior end and to 1/2 posterior end. Cuticle 8.0 μm thick. Length of cephalic setae 12 μm . Amphids large, elongated-oval, and 13.0 $\mu\text{m} \times 8.0 \mu\text{m}$. Of all the interlobular grooves only lateral ones open widely posteriorly; remainder closely compressed in furrowed region. Cephalic lobes with perforations and transverse slits. Cephalic suture sinuous. Small sclerotized granules present behind median lobes. Small pointed onchia, curved upward, situated in oral cavity. Photosensitive pigment absent. Eggs, 150 $\mu\text{m} \times 260 \mu\text{m}$, found in uterine branches.

Geographic distribution. Barents Sea.

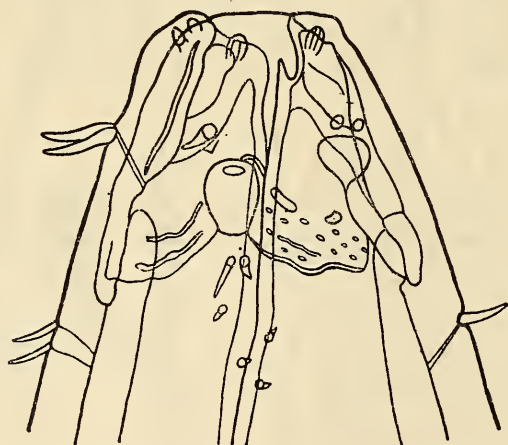


Figure 42. *Pseudocella pseudocellum* (from Filip'ev, 1916).

30. *Pseudocella bursata* Platonova, 1962 (Figure 43)

Platonova, 1962: 208, fig. 3A-G.

| | | | | | |
|-------|--|---------|-------------|---------------|---------------|
| | 46-55 | 790-800 | 2,800-3,060 | — | 24,650-27,608 |
| 10 ♂: | 48-50 | 70-75 | 170-175 | 180-185 | 180-187 |
| | × 25,000-28,000 μm ; a=148-151; b=8-9; c=69-71. | | | | |
| | 48-55 | 875-896 | 2,987-3,150 | 13,000-15,500 | |
| 10 ♀: | 40-50 | 75-82 | 162-182 | 175-197 | 180-200 × |

$$\times \frac{16,600-19,000}{220-224} \quad \frac{20,036-21,635}{198-210} \quad \frac{26,613-28,592}{178-187} \quad 27,000-29,000 \mu\text{m};$$

a = 135-140; b = 8-10; c = 61-70; V = 58-64%.

Body very long, tapers to $2/5$ midbody diameter at anterior end. Cuticle thickness 11 to 13 μm ; outer layer 7.5 to 8.0 μm . Cervical setae 17 μm long and situated in longitudinal rows. Interlobular grooves 16 to 20 μm deep and distinctly divided into fenestrae and furrows. Closed irregular groove present above fenestrae. Amphids 9.3 μm in diameter, located in each so-called lateral fenestra. Lobes of cephalic capsule with numerous minute and irregularly situated perforations. Margin of cephalic lobes sinuate, with marginal grooves. Band of large sclerotized granules situated posterior to cephalic capsule. Cephalic setae thick and long, 16 to 20 μm . Relatively large oral cavity with three onchia opens anterior to cephalic capsule. Six stout labial papillae situated around oral opening. Cephalic ring rather wide and irregular in shape. Neither photosensitive spots nor a trace of photosensitive pigment present. Length of anterior female genital tube 3,600 to 3,900 μm , of posterior tube 2,900 to 4,600 μm , and of reflexed parts 1,200 to 2,500 and 1,900 to 2,100 μm respectively. One very large egg (1,200 $\mu\text{m} \times 173 \mu\text{m}$) seen in uterus of some females. Spicules unequal in length; left 240 to 250 μm and right 190 to 200 μm . Gubernacular processes 67.5 μm long and differ in shape:

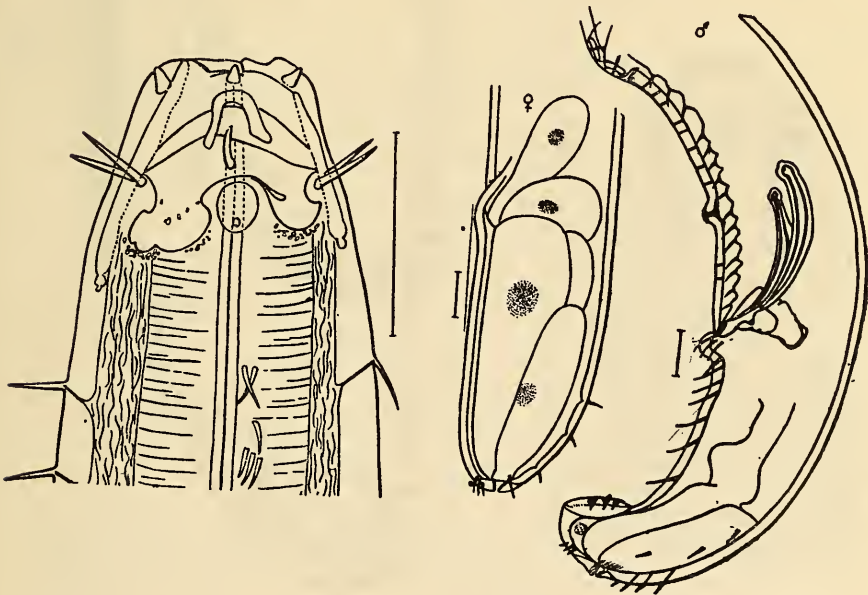


Figure 43. *Pseudocella bursata*.

one rounded at tip and the other rectangular. Accessory organ $75 \mu\text{m}$ anterior to anus. Pair of additional organs of different structure $100 \mu\text{m}$ anterior to accessory organ. Each consists of thin sclerotized duct running through a large papilla. Walls of papillae supported by annulated sclerotized thickenings. Eighteen to nineteen pairs of preanal papillae, each provided with a seta $27.5 \mu\text{m}$ long, and seven pairs of postanal setae $37.5 \mu\text{m}$ long, situated in anal region of male.

The characteristic cuticular structure on the ventral aspect of the tail of males resembles a bursa and is an interesting feature of this species; length of structure $76 \mu\text{m}$ and width at midlength $25 \mu\text{m}$.

This species significantly differs from other species of the genus in: 1) large size; 2) presence of oral cavity armed with teeth; 3) total lack of photosensitive pigment; and 4) presence of paired structures in addition to usual accessory organ.

Geographic distribution. Known only from the Bay of Aniva, Sea of Okhotsk, where it was found in gravel at a depth of 23 m.

102 31. *Pseudocella minor* Platonova, 1962 (Figure 44)

1 ♂: $\frac{29}{17} \frac{604}{42} \frac{1,020}{205} \frac{—}{132} \frac{4,850}{150} \frac{5,000}{140} \mu\text{m}; a=25; b=5; c=33.$

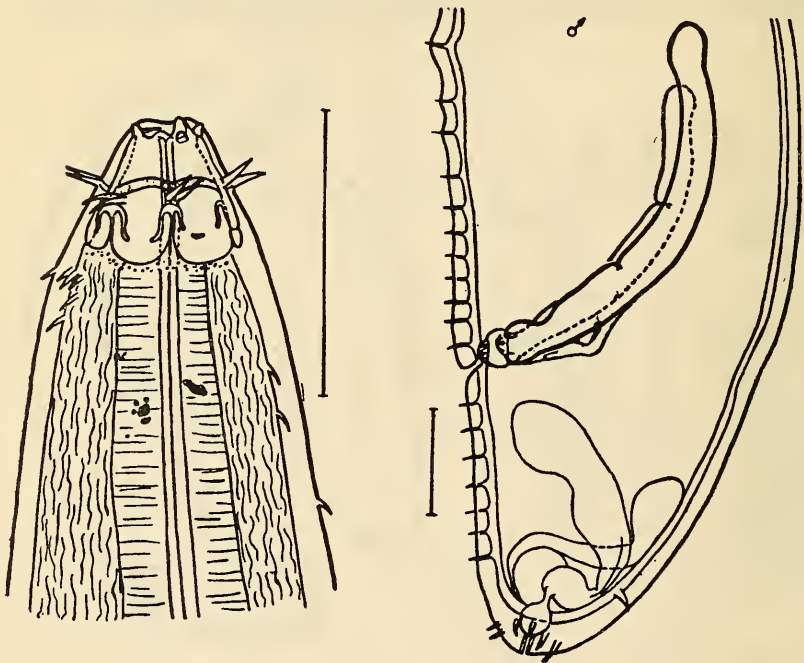


Figure 44. *Pseudocella minor*.

$$1 \text{ juv.: } \frac{17}{15} \frac{225}{22} \frac{575}{50} \frac{—}{67} \frac{1,985}{70} \frac{—}{70} 2,060 \text{ } \mu\text{m}; a=29; b=4; c=27.$$

Body tapers to 2/7 midbody diameter at anterior end. Cuticle 5.8 μm thick. Cephalic setae 4.8 μm and situated irregularly in groups or individually. Cephalic capsule opens in small oral cavity devoid of armature. Lobes of cephalic capsule separated by narrow interlobular anchor-shaped grooves 11.6 μm deep. Lobes with smooth margins devoid of peripheral grooves. Narrow band of extremely minute sclerotized granules lies posterior to cephalic capsule. Cephalic ring narrow and situated rather low in middle of cephalic capsule. Photosensitive pigment reduced to a few small granules. Genital armature of male represented by two thick, wide, arcuate spicules and gubernaculum with very short caudal processes. Spicules unequal in length, left 212 μm long and right 175 μm . Five pairs of very poorly developed preanal papillae 170 μm anterior to anus.

Distinguishing features of this species: 1) extremely small size; 2) cephalic ring situated in middle of cephalic capsule; 3) interlobular grooves anchor-shaped; 4) highly reduced photosensitive pigment; 5) gubernaculum with very short processes, not observed in other species of this genus; and 6) absence of accessory organ.

Geographic distribution. Found on the Pacific coast of Lake Shikotan (Kuril Islands) among littoral cospes of *Corallina* sp.

103 32. *Pseudocella kurilensis* Platonova, 1962 (Figure 45)
Platonova, 1962: 210–213, fig. 5A, B, and 6.

$$15 \text{ } \sigma: \frac{34-35}{22-25} \frac{437-459}{51-53} \frac{1,112-1,600}{105-112} \frac{—}{137-153} \frac{8,363-9,845}{162-187} \frac{—}{150-164}$$

× 8,500–10,000 μm ; a = 54–66; b = 5–9; c = 50–64.

$$15 \text{ } \text{♀}: \frac{34}{20-26} \frac{430-489}{52-53} \frac{1,400-1,600}{112-120} \frac{2,600-4,452}{156-160} \frac{—}{155-162}$$

× $\frac{4,800-6,452}{155-162} \frac{7,980-9,567}{152-160} \frac{8,884-10,825}{112-120} \frac{—}{—} \frac{9,000-11,000 \text{ } \mu\text{m};}{—}$

a = 60–70; b = 6–7; c = 56–62; V = 56–68%.

$$10 \text{ juv.: } \frac{30-40}{15-20} \frac{400-420}{49-50} \frac{980-1,020}{98-100} \frac{—}{110-120} \frac{5,842-6,000}{125-130} \frac{—}{118-120}$$

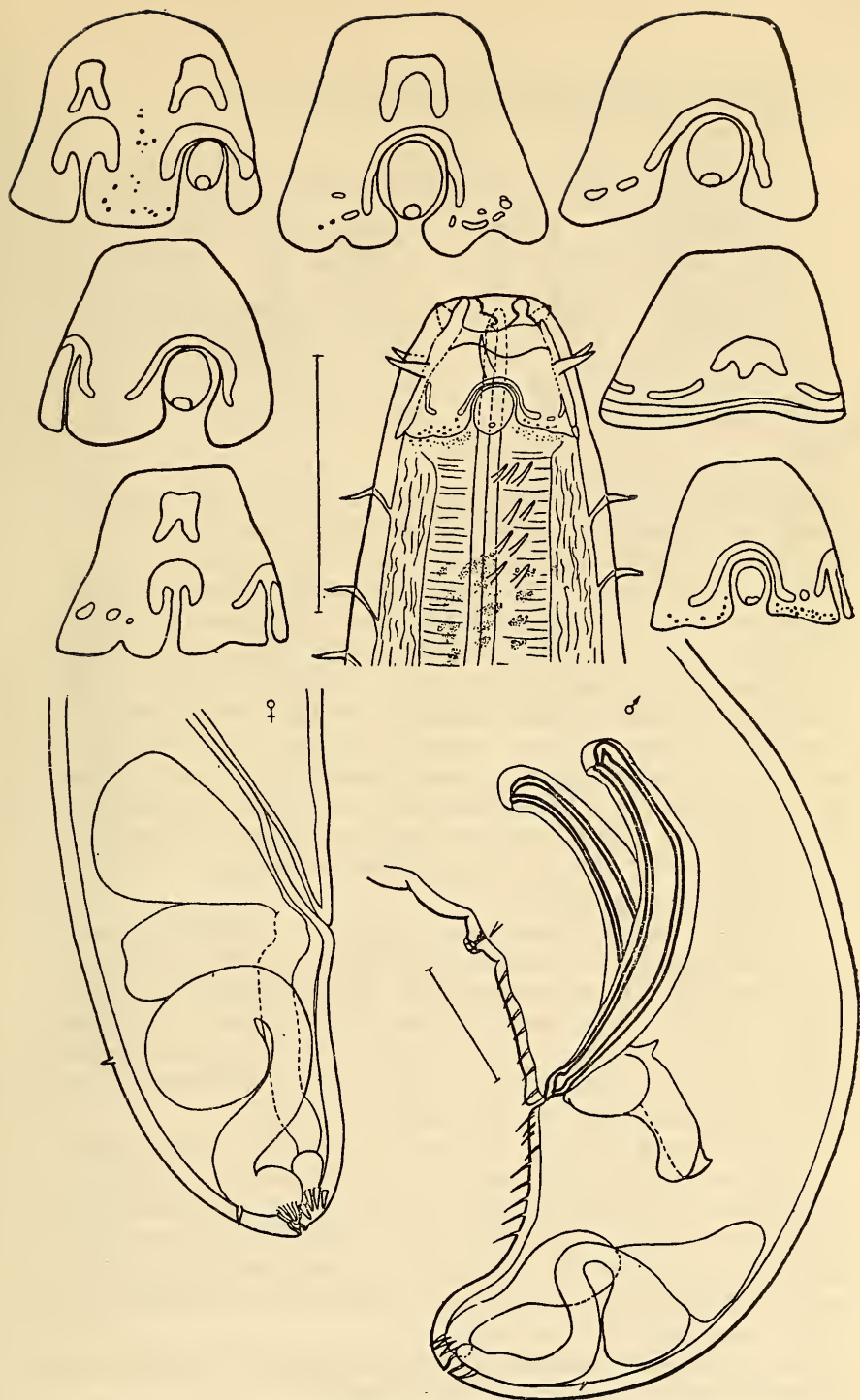
× 6,000–6,100 μm ; a = 46–50; b = 6; c = 60–61.

Body tapers to 2/7 midbody diameter at anterior end. Cuticle 4.6 to 6.9 μm thick. Length of cervical setae 6.0 to 8.0 μm in males and 5.0 to 5.8 μm in females. Number and arrangement of setae irregular; sometimes arranged in longitudinal rows or in groups of five to eight setae each. Amphids 6.9 μm in diameter. Subdorsal and subventral grooves

narrow with narrow furrows and small fenestrae; contours of latter form long posterior prominences. Hence interlobular grooves, like those in *P. minor*, anchor-shaped and 11.6 to 13.9 μm deep. Closed cleft present above grooves as in the case of *P. bursata*. Cephalic lobes with minute punctations which may merge into larger round or slit-shaped openings. Number and arrangement of punctations vary and probably not significant from a taxonomic point of view (Figure 45). Lobular margins occasionally with shallow peripheral grooves, sometimes sinuate, but otherwise smooth and rounded. Minute sclerotized granules, varying in size in some individuals, situated posterior to cephalic capsule. Some specimens had such granules only under some lobes. Length of cephalic setae 8.9 to 9.2 μm in females and 9.2 to 11.6 μm in males. Oral cavity devoid of oral armature and opens at top of cephalic capsule. Very wide cephalic ring situated slightly below oral cavity. Large irregular spots of red photosensitive pigment situated in anterior part of esophagus. Length of anterior female genital tube 2,000 to 2,678 μm , of posterior tube 2,500 to 2,700 μm , and of reflexed parts 700 to 721 and 720 to 824 μm respectively. Mature eggs 280 $\mu\text{m} \times 75 \mu\text{m}$. Spicular structure resembles that of *P. bursata*. Length of left spicule 175 to 200 μm and right spicule 212 to 250 μm . Gubernaculum with comparatively long processes. Tip of one process curves toward distal part of spicules and that of other process toward proximal part. Length of gubernacular processes 53.0 to 63.8 μm . Seven to eight pairs of postanal setae, 15 to 17 μm long, present in anal region. Accessory organ situated 87.5 μm anterior to anus. Five to nine pairs of preanal papillae situated anterior to accessory organ.

This species exhibits several characters which bring it close to *P. elegans*, *P. coecum*, and *P. saveljevi*. Nevertheless it differs substantially from *P. elegans* in: 1) structure of cephalic lobes (margin in *P. elegans* not rounded but sinuate with peripheral grooves); 2) presence of sclerotized granules behind cephalic capsule; 3) presence of punctations in cephalic lobes; 4) vulva situated midbody (in *P. elegans* vulva significantly displaced posterior to midbody— $V=70\%$); 5) much larger size and sharply differing indices. *P. kurilensis* differs from *P. coecum* in: 1) pigment dispersed throughout esophagus (not collected into small and compact clusters); 2) no exact localization of sclerotized granules (in *P. coecum* found only under median cephalic lobes); and 3) openings of amphids situated significantly low (in *P. coecum* situated in uppermost part). *P. kurilensis* differs from *P. saveljevi* in structure of male copulatory apparatus, especially in structure of gubernaculum.

Geographic distribution. Found on the coast of the Sea of Okhotsk and in the Pacific Ocean along Kuril Islands. Numerous specimens have been collected in the littoral zone under thalli of algae and in the sublittoral zone at a depth of 25 m among thickly massed sponges.



104 Figure 45. *Pseudocella kurilensis*. Various types of openings on cephalic capsule.

33. *Pseudocella mamillifera* Platonova, 1962 (Figure 46)
Platonova, 1962: 214–215, fig. 7A-3.

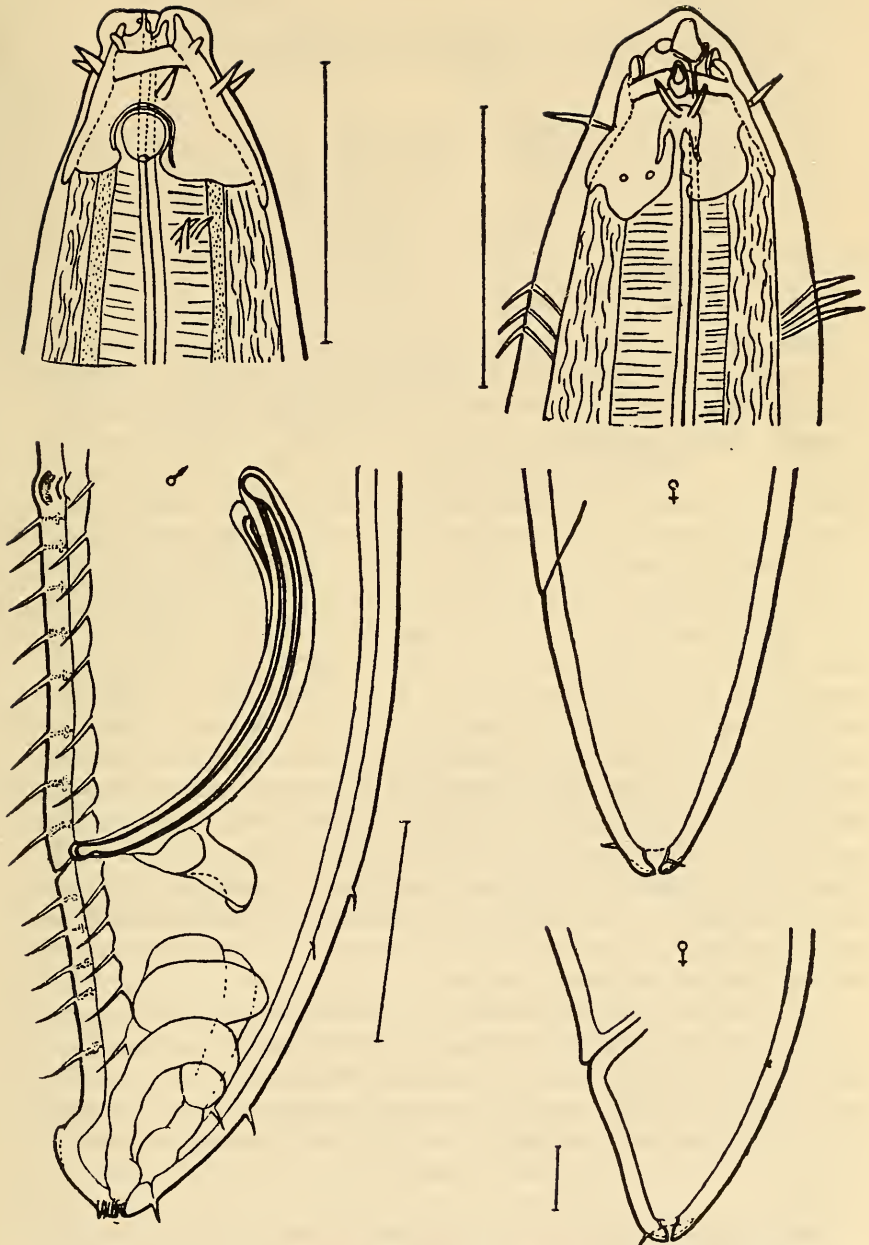
| | | | | | | | | | |
|-----|---------|--|------------------|-------------------------------|-----------------|-------------------------------|---------------------|-------------------------------|--|
| | 1 ♂: | $\frac{40}{17}$ | $\frac{484}{49}$ | $\frac{1,402}{137}$ | $\frac{—}{187}$ | $\frac{6,350}{187}$ | $\frac{6,500}{137}$ | | |
| | | $\frac{41-48}{23-29}$ | | $\frac{433-475}{54-55}$ | | $\frac{1,360-1,367}{150-153}$ | | $\frac{2,020-2,580}{187-193}$ | |
| 106 | 2 ♀: | | | | | | | $\frac{190-195}{87-112}$ | |
| | | $\frac{2,508-3,581}{200}$ | | $\frac{3,535-4,670}{195-197}$ | | $\frac{4,213-5,870}{87-112}$ | | 4,300–6,000 μm ; | |
| | | a = 21–30; b = 3–4; c = 46–49; V = 53–59%. | | | | | | | |
| | 1 juv.: | $\frac{40}{20}$ | $\frac{450}{50}$ | $\frac{1,200}{120}$ | $\frac{—}{150}$ | $\frac{5,200}{150}$ | $\frac{5,300}{145}$ | | |
| | | c = 56. | | | | | | | |

Body tapers to 10/37 midbody diameter at anterior end and 1/2 at posterior end. Cuticle 8.1 to 9.2 μm thick. Cervical setae very few and 7.0 to 11.6 μm . Cephalic capsule narrows gradually from base up to level of cephalic setae, then narrows abruptly, but broadens again toward top. Interlobular grooves, as in the preceding species, represented by narrow furrow and small fenestra 17.4 μm deep. Depth of lateral grooves 11.6 μm . Diameter of amphids 10.4 μm . Sublateral lobes of cephalic capsule with straight, slightly rounded margin; subventral lobes significantly longer and elongated in shape of tongue. Lobes lack peripheral grooves. Two round perforations present on each subventral lobe. Cephalic ring narrow and situated at level of cephalic setae. No trace of photosensitive pigment detected. Length of anterior female genital tube 927 to 947 μm , of posterior tube 927 to 1,030 μm , and of reflexed parts 515 to 721 μm and 618 to 825 μm respectively.

Copulatory system in male represented by two arcuate spicules unequal in length, left 200 μm and right 190 μm . Length of gubernacular process 20 μm . Thirteen setae, 20 μm long, arranged in anal region of male. Accessory organ situated 150 μm anterior to anus. Five preanal papillae situated anterior to accessory organ. Male has characteristic tail, bearing prominence on ventral aspect as in *P. trichodes*. Tail of female varies significantly in shape and size (Figure 46).

P. mamillifera is similar in structure to *P. trichodes* but differs from it in: 1) thinner and longer spicules; 2) presence of stout setae anterior to anus; 3) better developed accessory organ; 4) absence of photosensitive pigment; and 5) general shape of head. In *P. mamillifera* the ventral prominence is better developed, the cephalic capsule narrows abruptly in region of cephalic setae, and entire capsule narrows much more from the base to the top than in *P. trichodes*.

Geographic distribution. Found in the Sea of Okhotsk near the southern bank of Sakhalin at a depth of 53 m in sand.



34. *Pseudocella acuta* sp. nov. (Figure 47)

Holotype ♂. Institute of Zoology, Academy of Sciences, USSR. Collection No. 6011.

| | | | | | |
|----|-----|-------|-----|-------|----------------------------|
| 40 | 237 | 1,442 | — | 9,282 | 9,457 μm; a=45; b=7; c=54. |
| 23 | 58 | 117 | 175 | 210 | |

Paratypes.

| | | | | | |
|------|-------|---------|-------------|---------|--------------|
| 4 ♂: | 31 | 237-566 | 1,442-1,675 | — | 9,282-11,460 |
| | 20-25 | 52-58 | 112-137 | 137-175 | 167-210 |

× 9,457-11,635 μm; a=45-60; b=6-7; c=54-67.

| | | | | | |
|---------|-------|---------|-----------|---------|-------------|
| 5 juv.: | 29-34 | 160-463 | 856-1,493 | — | 7,345-8,085 |
| | 18-26 | 46-58 | 100-125 | 132-175 | 167-225 |

× 7,475-8,240 μm; a=37-45; b=55-87; c=53-57.

Body tapers considerably, to 1/8 to 1/10 midbody diameter at anterior end and 5/6 at posterior end. Cuticle 5.8 to 6.9 μm thick. Cephalic capsule narrows by 1/2 from base to top. Cephalic ring rather wide. Cephalic lobes with numerous minute perforations. Margin of each cephalic lobe with distinct peripheral groove. Sublateral interlobular grooves characteristic: extremely narrow furrows dilate abruptly into wide round fenestrae; offshoots from contours of latter extend downward. Depth of interlobular grooves 13.2 μm. Narrow strip of minute sclerotized granules located immediately behind cephalic capsule. Amphids round and diameter 6.9 to 8.1 μm. Oral opening surrounded by extremely short labial papillae. Cephalic setae 6.9 to 8.1 μm, cervical setae 5.8 to 6.9 μm and anal setae 17.4 μm. Photosensitive pigment totally absent. Nerve ring situated within limits of anterior third of esophagus. Length of anterior female genital tube 1,648 μm, of posterior tube 1,854, and of reflexed parts 824 μm and 927 μm respectively. Spicules unequal in length; right or left may be longer, and size 200 and 275 μm. Spicules with distinct but small capitulum, narrowing into neck. Small velum present on ventral aspect of spicules. Gubernaculum with long rectangular process, 45 to 67 μm. Accessory organ situated 87 to 105 μm anterior to anus. Nine to thirteen pairs of anal setae and six to nine pairs of preanal papillae present.

This species is closely related to *P. kurilensis* but differs in: 1) absence of photosensitive pigment; 2) furrows of interlobular grooves so narrow that lobes in region become contiguous (in *P. kurilensis* furrows usually much wider); 3) gubernacular process rectangular, slightly ventrally curved, and narrows toward tip up to 15 μm (in *P. kurilensis* narrows to 25 μm); and 4) tail somewhat pointed at end (in *P. kurilensis* more rounded).

Geographic distribution. Discovered in the northwestern part of the Sea of Okhotsk at a depth of 12 to 15 m in silted sand.

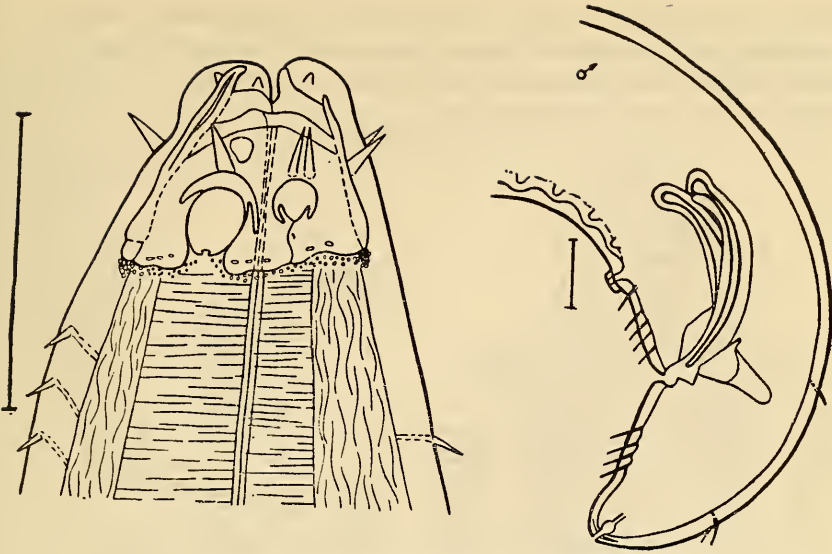


Figure 47. *Pseudocella acuta* sp. nov.

35. *Pseudocella angusticeps* sp. nov. (Figure 48)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 5002.

| | | | | | |
|----|-----|-------|-----|--------|---|
| 34 | 459 | 1,638 | — | 14,825 | 15,000 μm ; a=83; b=7; c=85. |
| 29 | 46 | 137 | 178 | 207 | |

Body tapers to 1/6 midbody diameter at anterior end and 2/3 at posterior end. Cuticle 9.3 μm thick. Cephalic capsule highly asymmetrical due to intense development of ventral outgrowth. Lateral interlobular grooves 11.6 μm deep, contain rounded amphids 12.7 μm in diameter. Lobes of cephalic capsule with narrow slit-shaped perforations. Periphery of lobes with very minute marginal grooves. Cephalic ring very wide. Labial papillae small. Length of cephalic setae 11.6 μm , and cervical setae 16.2 μm . Cervical setae cover anterior part of body rather densely. Twenty pairs of setae, 17.4 μm long, situated in anal region. Photosensitive pigment totally absent. Nerve ring encircles esophagus a little posterior to anterior third. Copulatory apparatus represented by two equal and arcuate spicules, 187.5 μm in length, with distinctly defined capitulum and neck broadening into spicular body. Gubernaculum with long rectangular process, 42.5 μm . Accessory organ situated 125 μm anterior to anus; another such organ 150 μm anterior to accessory organ. Cuticular thickening present on ventral side of tail.

This species is close to *P. kurilensis* but differs in: 1) significantly larger development of ventral outgrowth of cephalic capsule (which imparts a

different configuration to it); 2) absence of sclerotized granules posterior to cephalic capsule; 3) total absence of photosensitive pigment; 4) spicules equal in length and presence of two accessory organs; and 5) tail with cuticular thickening on ventral side.

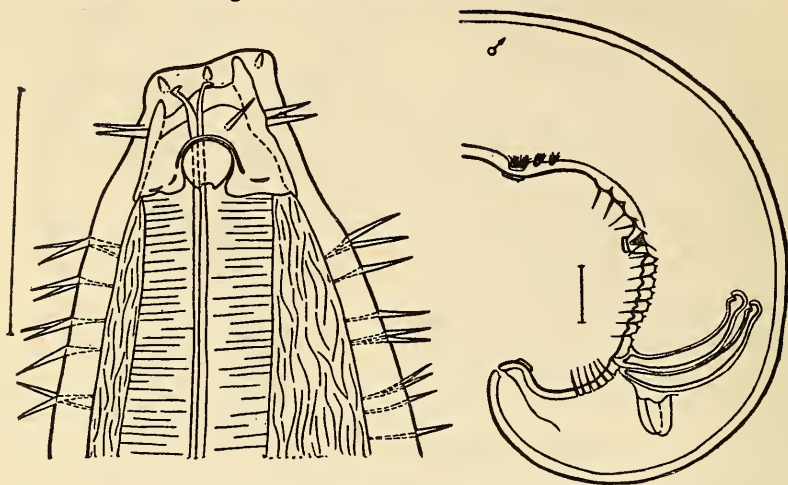


Figure 48. *Pseudocella angusticeps* sp. nov.

Geographic distribution. Found in Nagaev Bay (Sea of Okhotsk) at a depth of 28 m in silted sand.

36. *Pseudocella truncaticauda* sp. nov. (Figure 49)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 6594.

$$\frac{40 \quad 75 \quad 665 \quad - \quad 7,875}{23 \quad 63 \quad 75 \quad 150 \quad 165 \quad 162} \quad 8,075 \mu\text{m}; \quad a=49; \quad b=13; \quad c=41.$$

Body tapers to 1/7 midbody diameter at anterior end but almost does not narrow toward posterior end. Cuticle 11.6 μm thick. Cephalic capsule narrows rather abruptly; width at base three times greater than at top. Very narrow cephalic ring situated rather low, almost in middle of cephalic capsule. Cephalic lobes smooth, almost without marginal grooves, but with a small number of minute irregular perforations. Lateral interlobular grooves with relatively narrow furrows. Depth of grooves 15 μm . Band of large sclerotized granules situated posterior to cephalic capsule. Amphids round, slightly elongated in transverse direction, and 8.0 μm in diameter. Oral opening surrounded by short papillae. Length of cephalic setae 7.0 μm , cervical 10.4 μm , and anal 11.6 μm . Pigment spots irregular in shape and commence 34.8 μm posterior to cephalic capsule. Spicules unequal in length: left 237.5 μm and right 212.5 μm .

Both curve uniformly, are almost even in width throughout their length, and have a poorly delineated capitulum. First accessory organ situated 175 μm anterior to anus, and second 200 μm from first. Fourteen pairs of setae located in anal region. Body tapers rather abruptly posterior to anus in caudal region and widens slightly at tip of tail due to rather large protuberance on ventral side.

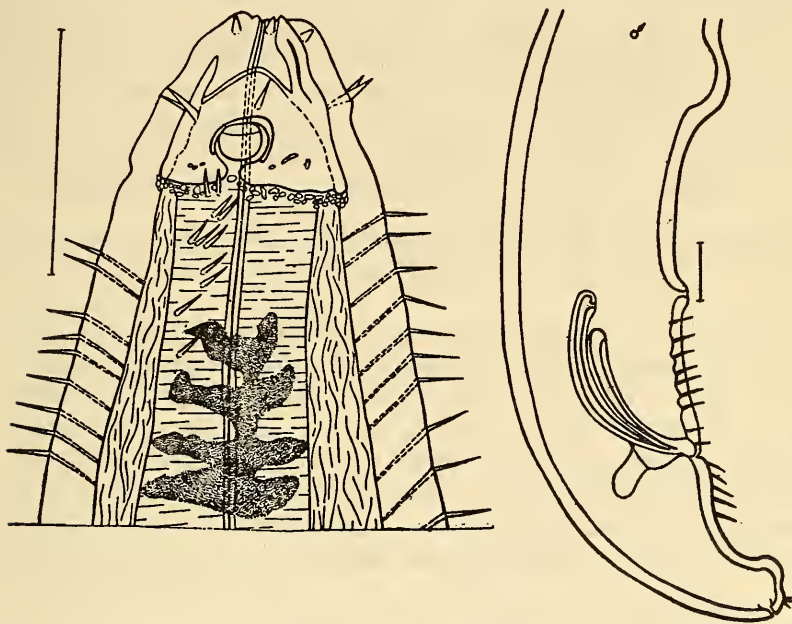


Figure 49. *Pseudocella truncaticauda* sp. nov.

This species is close to the male of *P. angusticeps* in structure of the posterior body part. Males of both species have two accessory organs and a protuberance on the ventral side of the tail. Males of *P. truncaticauda* differ however in: 1) spicules unequal in length, capituli poorly developed, and no cervix-like constriction (spicules of *Pseudocella angusticeps* equal in length with distinct capituli and long neck); 2) cephalic ring several times narrower and situated much lower than in the case of *P. angusticeps*; 3) lateral interlobular furrows much narrower than those in *P. angusticeps*; 4) band of large sclerotized granules situated posterior to cephalic capsule; and 5) photosensitive pigment present (in *P. angusticeps* there are neither granules nor pigment spots).

Geographic distribution. Found in the Sea of Okhotsk near Kuril Islands between Kunashir and Iturup, at a depth of 28 m among shingles (pebbles) with a mixture of silted sand.

110 37. *Pseudocella rarisetosa* sp. nov. (Figure 50)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7873.

| | | | | | |
|----|-----|-------|-----|-------|--|
| 45 | 391 | 1,390 | — | 7,312 | 7,476 μm ; a=31; b=5; c=46. |
| 25 | 40 | 150 | 242 | 127 | |

Paratypes.

| | | | | | |
|------|--|---------|-------------|---------|-------------|
| 3 ♂: | 48-50 | 360-412 | 1,133-1,339 | — | 6,592-7,107 |
| | 25-27 | 40-45 | 135-150 | 187-232 | 190-247 |
| | × 6,756-7,230 μm ; a=28-36; b=5-6; c=41-59. | | | | |

| | | | | | |
|------|--|---------|-------------|-------------|-------------|
| 2 ♀: | 45-51 | 370-432 | 1,236-1,339 | 4,326-4,480 | 6,695-7,621 |
| | 25 | 42 | 137 | 220 | 225 |
| | × 6,849-7,785 μm ; a=30-34; b=6; c=45-48; V=51-63%. | | | | |

| | | | | | | |
|---------|----|-----|-------|-----|-------|--|
| 1 juv.: | 40 | 391 | 1,287 | — | 7,158 | 7,302 μm ; a=33; b=6; c=51. |
| | 22 | 40 | 137 | 220 | 117 | |

Body tapers to 1/9 midbody diameter at anterior end and 1/2 at posterior end. Cuticle 7.0 μm thick. Width of cephalic capsule near basal part narrows to 2/7 at tip. Wide cephalic ring more or less situated at border of anterior third of cephalic capsule. Cephalic lobes smooth, without marginal grooves, and with a few minute perforations. Depth of interlobular grooves 15 μm . Rather broad accessory cleft extends in upper part of interlobular grooves. Amphids longitudinally elongated and 8.0 μm × 12.0 μm . Labial papillae fairly large. Cephalic setae 14 μm ,

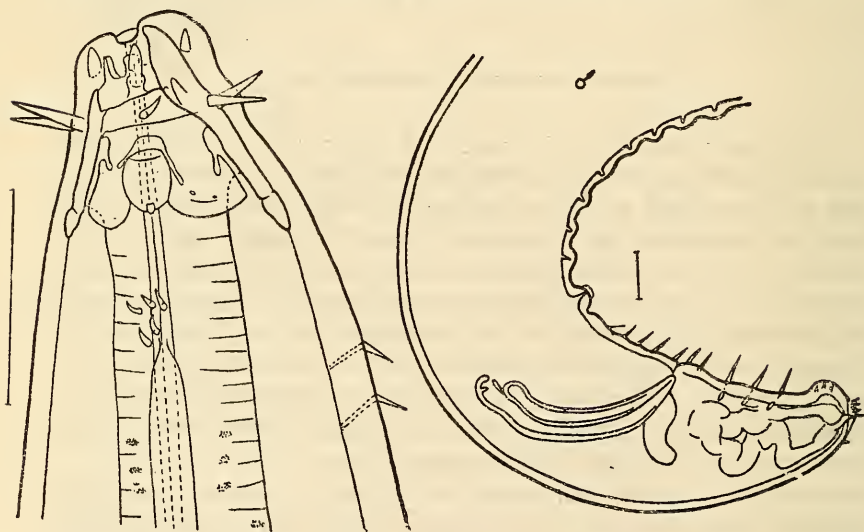


Figure 50. *Pseudocella rarisetosa* sp. nov.

cervical setae $6.0 \mu\text{m}$, and anal setae of male $25 \mu\text{m}$. Small pigment spots situated 80 to $100 \mu\text{m}$ from anterior end of body. Nerve ring encircles esophagus at border of anterior third. Length of female genital tubes 1,600 to $1,700 \mu\text{m}$. Spicules curve smoothly, unequal in length, with small capitulum; length 130 to $150 \mu\text{m}$. Gubernaculum with dorsal process and curves smoothly to ventral side. Accessory organ situated 80 to $100 \mu\text{m}$ anterior to anus; row of papillae situated anterior to accessory organ.

111 Protuberance on ventral surface of tail of male with three to four small setae. Similar small setae encircle opening of caudal glands.

P. rarisetosa resembles *P. angusticeps* in cephalic structure and caudal end but differs distinctly in: 1) small size; 2) shape of lateral interlobular grooves; and 3) absence of additional accessory organs.

Geographic distribution. Barents Sea.

38. *Pseudocella raddae* sp. nov. (Figure 51)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7916.

$$\frac{36 \quad 465 \quad 1,396 \quad - \quad 7,913}{24 \quad 48 \quad 120 \quad 148 \quad 195 \quad 169} \quad 8,072 \mu\text{m}; a=41; b=6; c=51.$$

Paratypes.

$$1 \text{ ♂: } \frac{32 \quad 372 \quad 1,103 \quad - \quad 7,088}{23-46 \quad 130 \quad 156 \quad 208 \quad 156} \quad 7,260 \mu\text{m}; a=35; b=7; c=42.$$

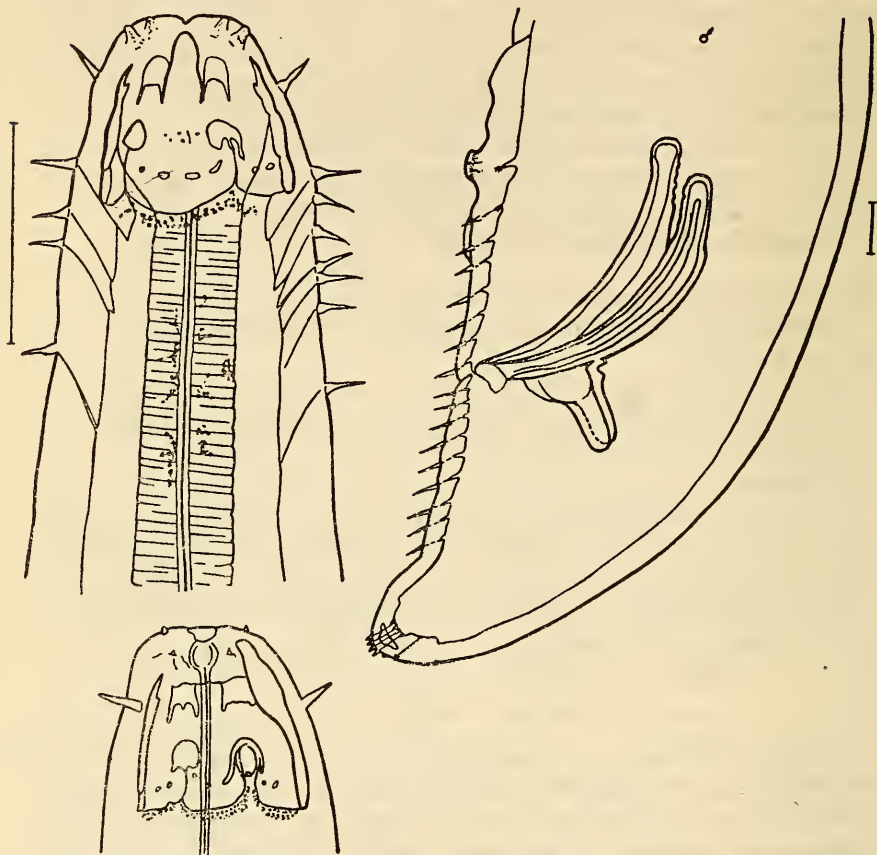
$$5 \text{ ♀: } \frac{32-38 \quad \quad \quad 399-465 \quad \quad \quad 1,263-1,529 \quad \quad \quad 2,952-4,423}{22-24 \quad 46-50 \quad \quad \quad 110-130 \quad \quad \quad 119-135 \quad \quad \quad 120-140}$$

$$112 \quad \times \frac{4,946-6,883 \quad 6,542-9,117 \quad 7,176-9,915}{135-161 \quad 122-143 \quad 101-130} \quad 7,300-10,087 \mu\text{m};$$

$$a=47-75; b=5-6; c=41-59; V=64-68\%.$$

Body tapers to 1/5 to 1/6 midbody diameter at anterior end and not less than 2/3 at posterior end. Cuticle $12 \mu\text{m}$ thick. Cephalic capsule narrows from base by slightly less than 1/2 toward top. Cephalic lobes with rather larger perforations. Margins of cephalic lobes without grooves. Sublateral interlobular grooves with extremely narrow furrows and wide rounded fenestrae; offshoots from contours of latter extend downward. Depth of interlobular grooves $16.8 \mu\text{m}$. Band of minute sclerotized granules situated posterior to cephalic capsule. Oral opening surrounded by large labial papillae. Cephalic setae $9.6 \mu\text{m}$, and cervical $7.2 \mu\text{m}$. Photosensitive pigment clusters in small irregularly shaped spots. Nerve ring encircles esophagus in anterior third. Length of anterior female genital tube 1,800 to $2,400 \mu\text{m}$, of posterior tube 1,500 to $2,200 \mu\text{m}$, and of reflexed parts 700 to $900 \mu\text{m}$. Vulva shifted slightly posterior to midbody (posteromedian position). Spicules slightly curved, with poorly developed capitulum, and distinct neck; length $182 \mu\text{m}$. Gubernaculum with large

process, 57 μm long. Cleft present on dorsal surface at base of process. Accessory organ situated 120 μm anterior to anus. Anal region of male armed with setae 12 μm in length. Small setae, 4.8 μm , situated at very tip of male tail.



111

Figure 51. *Pseudocella raddae* sp. nov.

This species is close to *P. acuta* but differs in: 1) shape of cephalic capsule; 2) presence of photosensitive pigment; 3) absence of spicular velum; and 4) presence of setae at tip of tail.

Geographic distribution. Discovered in littoral pools on the Pacific coast of Iturup (Kuril Islands).

2. Family ANTICOMIDAE Filipjev, 1918

Relatively minute forms; size more or less ranges from 2.0 to 10 mm. Body of majority of representatives fusiform, tapering toward both ends.

Cuticle thin and either single- or double-layered. Lips in most forms not developed and labial papillae, if present, extremely small. Nematodes of subfamily *Platycominae* with lips provided with large cuticular outgrowths constitute an exception. Oral cavity absent or poorly developed; occasionally armed with small onchia. Cephalic capsule poorly developed, and thin-walled; cephalic suture in most cases almost invisible. Cephalic setae arranged in one circle. Amphids cyathiform; only in representatives of subfamily *Platycominae* do amphids have unique contours, differing in shape in males and females. Photosensitive pigment absent. Esophagus gradually broadens toward base. Cardia longitudinally elongated. Gonads and genital tubes same structure as in representatives of *Leptosomatidae*. Spicular apparatus extremely diverse in different subfamilies. Accessory organ present in most cases. Caudal glands with highly stretched ducts; pores of latter open on tail terminally.

Key to Subfamilies of Family Anticomidae

- 1 (2). Cephalic setae long, many times exceeding cephalic diameter. Body tapers very slightly toward both ends.6. **Barbonematinae** subfam. nov.
- 113 2 (1). Cephalic setae short or insignificantly exceed cephalic diameter. Body distinctly tapers toward both ends.
- 3 (4). Tail long; posterior cylindrical part invariably exceeds conical part in length. Cervical setae often arranged in two longitudinal rows.4. **Anticominae** Filipjev, 1918.
- 4 (3). Tail comparatively short and often obtusely conical; cylindrical part, if present, significantly shorter than conical part. Cervical setae with different arrangement.
- 5 (6). Cephalic capsule developed. Individual cervical setae scattered throughout preneural part. . . . **Parabarbonematinae** subfam. nov.
- 6 (5). Cephalic capsule undeveloped. Cervical setae usually numerous and situated in clusters or transverse rows.5. **Platycominae** subfam. nov.

4. Subfamily ANTICOMINAE Filipjev, 1918

Small nematodes. Body fusiform and tapers notably toward both ends. Cephalic setae relatively short. Cervical setae situated laterally on both sides in longitudinal rows with two to six setae in each. In genus *Paranticoma* instead of setae there is a tube into which the pore of renette opens (renette always present in representatives of subfamily *Anticominae*). Renette elongated with long excretory ducts. Nerve ring encircles eso-

phagus in anterior third. Amphids round. Tail distinctly divided into two parts—wide conical part immediately behind anus followed by narrow cylindrical part, which may be very long and slender. Small protuberance usually present on terminal part of tail. Gubernaculum poorly developed or totally absent in most cases. Accessory organ tubular.

Key to Genera of Subfamily Anticominae

- 1 (2). Pore of renette situated on special setaceous outgrowth.
 **Paranticoma** Micoletzky, 1930.
- 2 (1). Setaceous outgrowth absent as renette pore lies directly on body.
- 3 (4). Two rows of "holes" in preneural region.
 **Antopus** Cobb, 1933.
- 4 (3). No "holes" in preneural region.
- 5 (6). Cervical setae situated in a few longitudinal rows; spicules long
 **Stenolaimus** Marion, 1870.
- 6 (5). Cervical setae situated in two longitudinal rows; spicules short.
- 7 (8). Accessory organ poorly developed and papillate in shape; gubernaculum absent. **Anticomopsis** Micoletzky, 1930.
- 8 (7). Accessory organ well developed and tubular in shape; gubernaculum usually present.
- 9 (12). Cephalic capsule poorly developed; cephalic suture not discernible.
- 10 (11). Oral cavity absent. 9. **Anticoma** Bastian, 1865.
- 11 (10). Oral cavity present. 8. **Odontanticoma** gen. nov.
- 12 (9). Cephalic capsule relatively well developed; cephalic suture distinctly visible. **Cephalanticoma** gen. nov.

114

Genus Cephalanticoma gen. nov.

Type species: *Anticoma chitwoodi* Inglis, 1964.

Cephalic capsule relatively well developed, with thick walls, and distinct and straight cephalic suture. Lips with well-developed papillae. Oral cavity small with three well-defined small onchia, one situated in each section of esophagus. Amphids situated on line of cephalic suture. Spicules thin, arcuate, and provided with velum. Gubernacular funnel encircles spicules at distal end; dorsal process of gubernaculum long, straight, and directed anteriorly along spicular axis. Tail very long and filiform.

8. *Genus Odontanticoma* gen. nov.

Type species: *Odontanticoma dentifer* sp. nov.

Cephalic capsule highly reduced. Oral cavity well developed and

armed with small distinct onchia. Oral opening encircled by well-developed lips. Labial papillae small. Amphids situated below line of cephalic suture. Spicules arcuate, wide in proximal part, with poorly developed capituli.

Figures given in Cobb's formula in the descriptions of genera *Odontanticoma* and *Anticoma* indicate distances from the anterior end up to the following parts: 1. posterior end of cephalic capsule or base of setae; 2. excretory pore of renette; 3. nerve ring; 4. basal part of esophagus; 5. commencement of anterior genital duct; 6. vulva; 7. commencement of posterior genital duct; and 8. anus.

For males these measurements are fewer (see p. 74).

Key to Species of Genus Odontanticoma

- 1 (2). Cephalic setae vary in length; in each pair of setae one notably shorter than the other. . . . **Odontanticoma vanoorti** Allgen, 1933.
- 2 (1). Cephalic setae equal in length.
- 3 (4). Onchia in oral cavity large. Cylindrical part of tail twice conical in length.39. **Odontanticoma dentifer** sp. nov.
- 4 (3). Onchia in oral cavity minute. Cylindrical part of tail equal to conical in length.40. **Odontanticoma murmanica** (Filipjev, 1927).

39. *Odontanticoma dentifer* sp. nov. (Figure 52)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7451.

| | | | | | | |
|------|----|-----|-----|----|-------|--------------------------------|
| 6 | 23 | 225 | 475 | — | 2,126 | 2,396 μ m; a=31; b=5; c=9. |
| 3-11 | 16 | 44 | 66 | 75 | 40 | |

Paratypes.

| | | | | | | |
|------|---|-------|---------|---------|-------|-------------|
| 3 ♂: | 6 | 22-24 | 206-230 | 452-487 | — | 2,100-2,369 |
| | 8-9 | 11-12 | 16-17 | 44-48 | 44-48 | 74-76 |
| | × 2,396-2,649 μ m; a=31-34; b=5; c=9. | | | | | |

| | | | | | | |
|------|-----------------------------------|-------------|-------------|----------------------|---------|--|
| 4 ♀: | 4-5 | 20-23 | 208-225 | 459-475 | 816-826 | |
| | 8-9 | 11-12 | 17-18 | 49-52 | 79-86 | |
| | 1,122-1,320 | 1,402-1,545 | 2,063-2,537 | 2,343-2,812 μ m; | | |
| | 86-92 | 91-93 | 48-51 | | | |
| | a=25-30; b=5-6; c=8-10; V=45-47%. | | | | | |

| | | | | | | |
|---------|---|-------|---------|---------|-------|-------------|
| 4 juv.: | 4 | 19-23 | 195-200 | 382-400 | — | 1,618-1,720 |
| | 7-8 | 10-11 | 15-16 | 48-50 | 71-73 | 76-78 |
| | × 1,847-1,950 μ m; a=24-25; b=5; c=8-9. | | | | | |

Body tapers to 1/8 to 1/10 midbody diameter at anterior end and 1/2 at posterior end. Head anteriorly truncated with distinct lips. Tail long; slender cylindrical part twice length of conical part. Cuticle thin, 2.3 μ m.

Cephalic capsule thin-walled. Cephalic width exceeds length by 1.5 times. Amphids situated one cephalic diameter from anterior end of body. Width of amphid $4.6 \mu\text{m}$ ($1/4$ corresponding diameter). Cephalic setae long, $8.0 \mu\text{m}$, longer than $1/2$ corresponding diameter. Cervical setae $4.6 \mu\text{m}$, situated in groups of five at a distance of three cephalic diameters from anterior end of body. Cervical pore located midway between amphids and first cervical seta. Oral cavity extensive, with three relatively large teeth. Nerve ring encircles esophagus slightly above its middle. Length of anterior female genital tube 306 to $496 \mu\text{m}$, of posterior tube 225 to $280 \mu\text{m}$, and of reflexed parts 285 to $350 \mu\text{m}$ and 200 to $250 \mu\text{m}$ respectively. One to two eggs, 91 to $102 \mu\text{m} \times 66 \mu\text{m}$, observed in genital tract of female. Spicular length $70 \mu\text{m}$. Velum thin and narrow, barely distinguishable. Gubernaculum small; encircles spicules at distal end and expands in ventral direction. Accessory organ situated $70 \mu\text{m}$ anterior to anus.

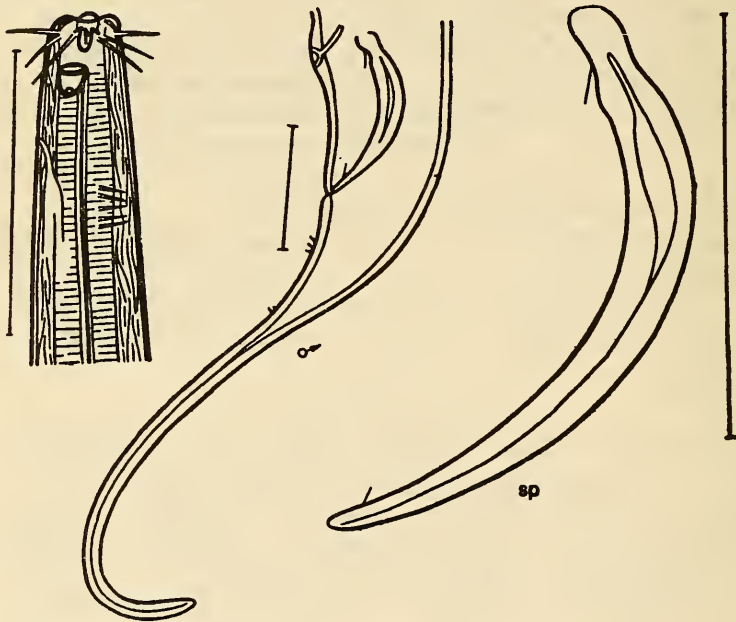


Figure 52. *Odontanticoma dentifer* gen. nov., sp. nov.

Distinguishing features of this species: 1) long cephalic setae; 2) well-developed oral cavity with onchia; 3) spicules arcuate, with slits, and expand in their widest part; and 4) gubernaculum expands toward anus.

Geographic distribution. Found in the Gulf of Terpeniya (Sakhalin, Sea of Okhotsk) at a depth of 36 to 39 m in a bed of fine sand.

*40. *Odontanticoma murmanica* Filipjev, 1927 (Figure 53)Filip'ev, 1927: 78-79, pl. 2, fig. 2a-e (*Anticoma*) (description).

| | | | | | | |
|------|----|-----|-----|-------|-------|--|
| 3 ♂: | — | 250 | 515 | — | 2,450 | 2,700-2,930 μm ; a=24; b=5; c=10. |
| | 16 | 64 | 95 | 116 | 60 | |
| 2 ♀: | — | 245 | 525 | 1,100 | 1,270 | 2,500-2,900 μm ; a=27; b=5; c=9. |
| | 16 | 62 | 90 | — | 105 | |
| | | | | | 52 | |

- 116 Conical and cylindrical parts of tail equal in length. Cuticle thick, 4.0 to 5.0 μm . Head slightly truncated. Labial papillae project slightly. Length of cephalic setae 8.0 μm (1/2 corresponding diameter). Amphids larger in males (4.0 $\mu\text{m} \times 5.0 \mu\text{m}$) than in females (2.0 $\mu\text{m} \times 3.0 \mu\text{m}$). Cervical setae 6.0 μm , situated two cephalic diameters from anterior end. Cuticle of head thicker than in rest of body. Cephalic suture barely visible. Cervical glands 60 $\mu\text{m} \times 30 \mu\text{m}$, situated against posterior part of esophagus. Pore of cervical gland situated at level of amphid or slightly above. Oral capsule thick-walled, 9.0 μm deep, and with short anterior vestibulum. Esophageal glands open at base of oral capsule. Cardia 35 to 45 μm . Testes separated from vas deferens by constriction. Length of spicules 75 to 80 μm (1/2 anal diameter). Gubernaculum narrow and

Figure 53. *Odontanticoma murmanica* (from Filip'ev, 1927).

long. Bursal musculature highly developed and situated four anal diameters anterior to anus. Accessory organ consists of sclerotized ring encircling opening with tube $23\ \mu\text{m}$ long; situated two anal diameters anterior to anus. Two short anal setae present. Length of vagina $35\ \mu\text{m}$.

Distinguishing features of this species: 1) situation of cervical pore at amphid level; 2) presence of highly elongated oral cavity, armed with small onchia; and 3) presence of well-developed spicular velum.

Geographic distribution. Found in the Barents and Kara Seas at depths ranging from 120 to 300 m in silty and muddy substrata.

9. Genus *Anticoma* Bastian, 1865

Eberth, 1863: 28 (*Odontobius*, part.); Bastian, 1865: 141; Bütschli, 1874: 35; de Man, 1878: 98; 1886: 51; Cobb, 1891: 765; Filip'ev, 1918: 63-65; 1927: 73; Micoletzky, 1930: 248; Wieser, 1953c: 14-16; Inglis, 1964: 325.

Type species: *Anticoma eberthi* Bastian, 1865.

Suture of cephalic capsule almost invisible. Lips not developed; labial papillae, if present, extremely small. Well-defined oral cavity absent. Amphids round or slightly elongated longitudinally. Nerve ring encircles esophagus slightly above its middle. Vulva anterior to midbody. Vulval glands well developed. Spicules differ in structure. Gubernaculum small.

As observed by Filip'ev (1927), this genus is extremely uniform and hence identification of species rendered difficult. Important taxonomic features of this genus are: structure of the spicular apparatus; location of excretory pore of cervical gland; position of cervical setae and amphids; and relative lengths of cephalic, cervical, and caudal setae. The number of cervical setae changes with the age of the nematode (Micoletzky, 1930; Wieser, 1953c) and for this reason cannot serve as a taxonomic character.

Key to Species of Genus Anticoma

- 1 (6). Cervical setae absent.
- 2 (3). Spicules with velum with processes directed ventrally.....
.....*A. curticauda* (Platonova, 1968).
- 3 (2). Spicules without such velum.
- 4 (5). Cylindrical part of tail exceeds conical part in length.....
.....*A. longissima* Allgen, 1958.
- 5 (4). Conical part of tail exceeds cylindrical part in length.....
.....*A. aberrans* Allgen, 1951.
- 6 (1). Cervical setae present.
- 7 (52). Excretory pore of renette anterior to cervical setae.

- 8 (11). Cuticle finely annulated.
- 9 (10). Amphids situated immediately behind anterior end of body . . .
 **A. wieseri** Mawson, 1958.
- 10 (9). Amphids situated one cephalic diameter away from anterior
 end of body **A. australis** Mawson, 1956.
- 11 (8). Cuticle smooth.
- 12 (17). Body extremely narrow ($a=60$ to 70).
- 13 (14). Cephalic setae extremely short, about one-sixth of correspond-
 ing diameter **A. tenuis** Allgen, 1930.
- 14 (13). Cephalic setae perceptibly longer, not less than one-fourth of
 corresponding diameter.
- 15 (16). Amphids very large, occupying one-third of corresponding dia-
 meter **A. procera** Micoletzky, 1930.
- 16 (15). Amphids very minute: width does not exceed one-sixth of corre-
 sponding diameter 44. **A. arctica** Steiner, 1916.
- 17 (12). Body relatively wide (a —does not exceed 30 to 40).
- 18 (19). Cephalic setae very short and appear papillate
 **A. microseta** Allgen, 1935.
- 19 (18). Cephalic setae significantly long and look like filaments.
- 20 (21). Gubernaculum with small caudally directed process
 45. **A. limalis** Bastian, 1865.
- 21 (20). Gubernaculum without caudal process.
- 22 (37). Cylindrical part of tail equal to conical in length.
- 23 (24). Cephalic setae short, constituting one-fifth of corresponding
 diameter 53. **A. curta** sp. nov.
- 24 (23). Cephalic setae significantly long, constituting one-third to one-
 half of corresponding diameter.
- 25 (26). Males with well-developed preanal setae
 55. **A. pacifica** sp. nov.
- 26 (25). Males without preanal setae.
- 27 (28). Body large; length about 4.0 mm 51. **A. grandis** sp. nov.
- 28 (27). Body significantly smaller; length about 2.0 mm.
- 29 (32). Spicules distinctly narrow in region of capituli.
- 30 (31). Cervical setae situated 1.5 cephalic diameters from anterior end
 **A. acuminata** (Eberth, 1863).
- 118 31 (30). Cervical setae situated three cephalic diameters from anterior
 end 56. **A. pontica** Filipjev, 1918.
- 32 (29). Spicules do not narrow in region of capituli but taper gradually
 from proximal end to distal end.
- 33 (36). Accessory organ situated at level of spicular capitulum.
- 34 (35). Amphids small; width less than one-fourth corresponding dia-
 meter **A. subsimilis** Cobb, 1914.
- 35 (34). Amphids large, width more than one-third corresponding dia-

- meter.....**A. zostera** Schulz, 1932.
- 36 (33). Accessory organ situated anterior to anus at a distance equal to twice length of spicules.....46. **A. pellucida** Bastian, 1865.
- 37 (22). Cylindrical part of tail twice or more length of conical part.
- 38 (39). Cephalic setae short; length not more than 1/4 corresponding head diameter.....43. **A. minor** Filipjev, 1927.
- 39 (38). Cephalic setae unusually long; length constitutes 1/3 to 1/2 corresponding head diameter.
- 40 (41). Accessory organ absent.....**A. allgeni** Platonova, 1968.
- 41 (40). Accessory organ present.
- 42 (45). Excretory pore distinctly shifted forward and situated one cephalic diameter from anterior end.
- 43 (44). Spicular length about 30 μm**A. profunda** Micoletzky, 1930.
- 44 (43). Spicular length about 60 μm48. **A. brevisetosa** Platonova 1967.
- 45 (42). Excretory pore situated more than two cephalic diameters from anterior end.
- 46 (47). Amphids large; width constitutes 1/2 corresponding head diameter.....54. **A. uschakovi** sp. nov.
- 47 (46). Amphids significantly small; width constitutes 1/3 to 1/4 corresponding head diameter.
- 48 (49). Cervical setae situated 4.5 cephalic diameters from anterior end47. **A. novozemelica** Filipjev, 1927.
- 49 (48). Cervical setae situated three cephalic diameters from anterior end.
- 50 (51). Preanal region and caudal end of male armed with setae.....**A. cobbi** Inglis, 1971.
- 51 (50). Preanal region and caudal end of male without setae.....52. **A. behringiana** sp. nov.
- 52 (7). Excretory pore posterior to cervical setae.
- 53 (56). Length of cephalic setae either equal to or exceeds cephalic diameter.
- 54 (55). Body tapers sharply toward cephalic end. Cephalic diameter 1/6 diameter at basal part of esophagus.....**A. graciliceps** Platonova, 1968.
- 55 (54). Body almost even in width throughout...**A. strandi** Allgen, 1940.
- 56 (53). Length of cephalic setae less than cephalic diameter.
- 57 (60). Excretory pore situated immediately behind cervical setae.
- 58 (59). Males with preanal and caudal setae...**A. campbelli** Allgen, 1932.
- 59 (58). Males without preanal and caudal setae.....**A. extensa** Wieser, 1953.
- 60 (57). Excretory pore situated at significant distance from cervical setae.
- 119 61 (62). Excretory pore situated near nerve ring: **A. tipica** Cobb, 1891.

- 62 (61). Excretory pore situated between cervical setae and nerve ring.
- 63 (64). Paired cephalic setae unequal in length.
 **A. litoris** Chitwood, 1936.
- 64 (63). All cephalic setae equal in length.
- 65 (72). Body large; length about 5.0 to 6.0 mm.
- 66 (67). Tail short (*c* more than 20); cylindrical part equal to conical part
 in length. 41. **A. eberthi** Bastian, 1865.
- 67 (66). Tail long (*c* not more than 10); cylindrical part twice length of
 conical part.
- 68 (69). Accessory organ situated 1.5 anal diameters from anus.
 **A. major** Mawson, 1956.
- 69 (68). Accessory organ more than 2.5 anal diameters from anus.
- 70 (71). Spicules with distinct narrow capitulum.
 49. **A. filipjevi** Platonova, 1967.
- 71 (70). Spicules without capitulum.
 50. **A. tenuicaudatoides** Gerlach and Riemann, 1974.
- 72 (65). Body small; length not more than 3.0 mm.
- 73 (74). Tail short (*c* about 30). 42. **A. insulaealbae** Filipjev, 1927.
- 74 (73). Tail long (*c* about 10).
- 75 (76). Spicular capitulum flat on ventral side.
 57. **A. platonovae** Sergeeva, 1972.
- 76 (75). Spicular capitulum without flat ventral side.
- 77 (78). Cervical setae situated immediately behind amphids.
 **A. dahli** Wieser, 1953.
- 78 (77). Cervical setae situated at a significant distance from amphids.
- 79 (80). Cylindrical part of tail four times longer than conical part.
 **A. trichura** Cobb, 1898.
- 80 (79). Cylindrical part of tail not more than twice length of conical
 part.
- 81 (82). Preanal setae entirely absent in males.
 **A. kerguelensis** Mawson, 1958.
- 82 (81). Preanal setae present in males.
- 83 (84). Length of cephalic setae 1/3 corresponding cephalic diameter.
 **A. lata** Cobb, 1898.
- 84 (83). Length of cephalic setae 1/2 corresponding cephalic diameter.
- 85 (86). Amphids less than one cephalic diameter from anterior end.
 **A. columba** Wieser, 1953.
- 86 (85). Amphids situated two cephalic diameters from anterior end.
 **A. ditlenseni** Micoletzky, 1930.

41. *Anticoma eberthi* Bastian, 1865 (Figure 54)

Bastian, 1865: 141, pl. XI, figs. 143–145; de Man, 1889: 186, tab. 5, fig. 3; Southern, 1914: 22; Filip'ev, 1918: 64; Platonova, 1967: 834.

$$1 \text{ ♂: } \frac{9}{16} \frac{29}{26} \frac{309}{34} \frac{700}{102} \frac{—}{150} \frac{4,408}{187} \frac{67}{67} 4,765 \text{ } \mu\text{m}; a=25; b=7; \\ c=12.$$

Body tapers to 1/10 midbody diameter at anterior end and 1/2 at posterior end. Head anteriorly truncated. Tail short; narrow cylindrical part equal in length to conical part. Cuticle 4.6 μm thick. Width of cephalic capsule almost three times its length. Amphids not detected. Cephalic setae 10 μm (1/2 corresponding diameter). Cervical setae situated 1.5 cephalic diameters from anterior end. Excretory pore of renette situated near nerve ring. Oral cavity funnel-shaped with thick sclerotized walls. Nerve ring encircles esophagus slightly above its middle. Length of spicules 100 μm ; spicules curved at end of first and second thirds of their length. Spicular capitulum merges gradually into spicular body. Concave oval area present on ventral aspect of spicular capitulum. Gubernaculum short, encircles distal end of spicules, narrows dorsally, and widens ventrally. Accessory organ situated 100 μm anterior to anus.

Distinguishing features of this species: 1) cephalic capsule broadens notably toward base (width at base exceeds that at top three times, whereas usually in species of genus *Anticoma* width exceeds length by two times); 2) extremely low position of excretory pore of cervical gland (situated near nerve ring); 3) characteristic concave oval area present on

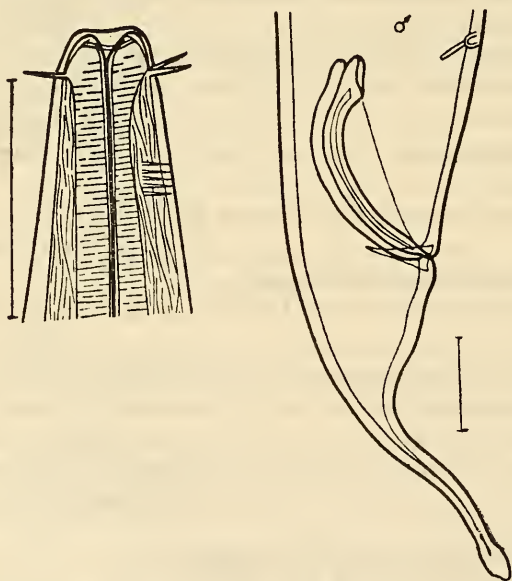


Figure 54. *Anticoma eberthi*.

ventral surface of spicular capitulum; and 4) tail narrows sharply in first half (from anus to midtail, tapers to 1/6).

Geographic distribution. Found in the Kara Sea (at a depth of 19 m on a sandy substratum), North Sea, and north Atlantic Ocean.

*42. *Anticoma insulaealbae* Filipjev, 1927 (Figure 55)

Filip'ev, 1927: 74, 75, pl. 1. fig. 1a-d (description).

1 ♀: $\frac{\text{---} \quad 325 \quad 720 \quad 1,115 \quad 2,625 \quad 3,630 \quad 4,890}{30 \quad 90 \quad 115 \quad \text{---} \quad 177 \quad \text{---} \quad 60}$ 5,080 μm ; a=29;
b=7; c=29.

1 ♂: $\frac{\text{---} \quad 270 \quad 580 \quad \text{---} \quad 4,410}{27 \quad 82 \quad 110 \quad 130 \quad 60}$ 4,560 μm ; a=35; b=8; c=30.

Tail conical, with dilatation at end, and in males curves toward ventral side. Cuticle smooth and 2.0 to 2.5 μm thick. Setae longer in lateral fields. Head round, slightly flattened anteriorly. Labial papillae very small. Length of cephalic setae 18 μm (2/3 corresponding diameter). Sublateral cephalic setae slightly shorter than remaining ones, 14 μm long. Amphids small, 7.0 $\mu\text{m} \times 6.0 \mu\text{m}$, and 6.0 μm posterior to lateral cephalic setae. Four cervical setae situated about two cephalic diameters from anterior end. Lateral fields narrow. Renette situated opposite posterior part of

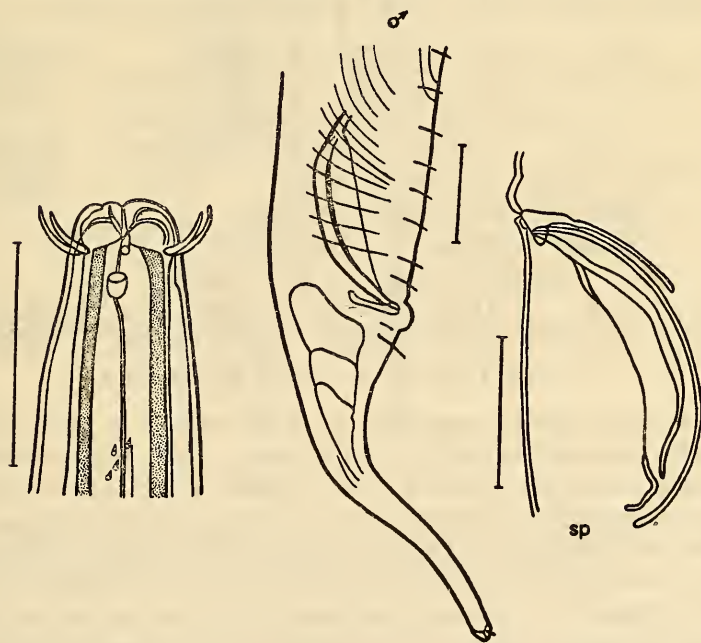


Figure 55. *Anticoma insulaealbae* (from Filip'ev, 1927).

esophagus. Cervical pore situated near limit of posterior third of pre-neural part. Dorsal glands open in anterior part; both subventral glands open somewhat posteriorly. Cardia round, 27 μm in diameter. Rectum long, 65 μm , and separated from intestine by distinct constriction. Anterior testis occupies 29%, posterior testis 31%, and vas deferens 71% of length of genital tube. Spicules curved, enlarged near capitulum, and with well-developed velum. Gubernaculum flat, closely apposed to spicules.

- 121 Length of spicules 100 μm (1.5 times longer than anal diameter). Length of gubernaculum 40 μm . Accessory organ situated 1.6 anal diameters anterior to anus. Nine pairs of subventral setae situated anterior to accessory organ, two pairs located postanally, and several pairs between anus and accessory organ. Slightly shorter setae situated on tail.

A. insulaealbae belongs to that group of species in which the cervical pore is shifted posteriorly, but differs from other representatives of this group in smaller size and longer filiform tail. It differs from *A. eberthi* in longer cephalic and cervical setae, situated farther from the anterior end, longer tail, and smaller measurements.

Geographic distribution. Found in the Kara Sea at a depth range of 15 to 20 m, predominantly in a sandy substratum.

43. *Anticoma minor* Filipjev, 1927 (Figure 56)

Filip'ev, 1927: 77, 78, pl. 2, fig. 5 a-d; Platonova, 1967: 834.

$$5 \text{ } \sigma: \begin{array}{cccccc} 4-5 & 12-15 & 192-207 & 401-422 & - & 1,648-1,751 \\ \hline 8-9 & 12-14 & 15-16 & 35-42 & 44-52 & 59-61 & 42-47 \end{array}$$

$$\times 1,887-2,006 \mu\text{m}; a=32-34; b=5; c=8.$$

$$12 \text{ } \text{f}: \begin{array}{cccccc} 4-5 & 15-17 & 157-197 & 316-448 & 597-782 \\ \hline 9-10 & 13-15 & 17-18 & 37-45 & 38-53 & 44-62 \end{array}$$

$$\times \begin{array}{ccc} 803-1,030 & 988-1,236 & 1,339-2,060 \\ \hline 46-64 & 43-64 & 30-37 \end{array} 1,579-2,310 \mu\text{m};$$

$$a=34-35; b=5; c=7-9, V=45-50\%.$$

$$8 \text{ } \text{juv}.: \begin{array}{cccccc} 4 & 11-12 & 162-197 & 275-290 & - & 1,133-1,442 \\ \hline 6-8 & 10-11 & 13-15 & 26-30 & 34-37 & 40-44 & 32-34 \end{array}$$

$$\times 1,333-1,652 \mu\text{m}; a=33-37; b=5; c=5-8.$$

- Small, rather slender nematodes. Body tapers to 1/5 to 1/6 midbody diameter at anterior end and 2/3 to 1/2 at posterior end. Head anteriorly truncated. Cuticle thin, 2.0 to 2.5 μm . Cephalic capsule extremely thin; width at base exceeds length by 2.0 to 2.5 times. Amphids 5.8 μm in diameter and situated 11 to 13 μm from anterior end. Length of cephalic setae 3.5 μm (1/3 to 1/4 corresponding diameter). Cervical setae (five in number) situated 37 to 41 μm from anterior end (distance equal to three cephalic diameters). Length of cervical setae 5.0 to 6.0 μm . Renette commences at posterior end of esophagus and excretory pore opens at level

of amphid. Wall of oral cavity slightly sclerotized. Esophagus long, dilating slightly toward base. Nerve ring encircles esophagus slightly below its middle. Length of anterior female genital tube 206 to 248 μm , of posterior tube 185 to 206 μm , and of reflexed parts 180 to 200 μm and 160 to 190 μm respectively. Vulva median or anteromedian. One mature egg, 100 to 120 $\mu\text{m} \times 40 \mu\text{m}$, seen in each uterus. Male genital tubes commence 570 to 600 μm from anterior end. Spicules arcuate, irregularly shaped, with large lamelliform velum; length 50 to 57 μm , i.e., almost equal to anal diameter. Gubernaculum simple; length 28 to 30 μm . Accessory organ 50 to 65 μm anterior to anus.

According to Filip'ev (1927) this species resembles *A. murmanica* and *A. limalis* in structure. It differs from the former in location of the cephalic setae, situated much closer to the anterior end, and from the latter in shape of spicules and rather long cephalic setae.

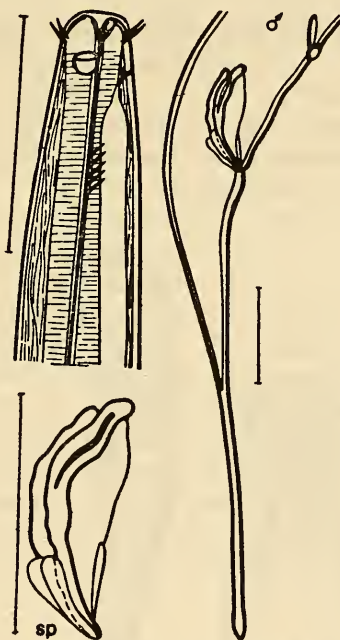


Figure 56. *Anticoma minor*.

Geographic distribution. Found in the Barents Sea from the coastal zone to a depth of 200 m on rocky and sandy substrata.

*44. *Anticoma arctica* Steiner, 1916 (Figure 57)

Steiner, 1916: 656–660, tab. 36, fig. 45a–f; Filip'ev, 1927: 75 (description); Allgen, 1928: 283; 1929a: 10.

| | | | | | |
|------|--------------------------------|-------------------------|-----------------------------|-------------------------|-----------------------------|
| 4 ♀: | $\frac{165-210}{10-12}$ | $\frac{495-590}{34-41}$ | — | $\frac{1,410-1,600}{—}$ | $\frac{1,650-1,800}{44-50}$ |
| | $\times \frac{2,260-2,480}{—}$ | | $\frac{2,720-3,135}{30-34}$ | | $2,850-3,300 \mu\text{m};$ |
| | $a=56-69; b=6-7; c=18-19.$ | | | | |

Characterized by unusually elongated and extremely slender body. Tail relatively short; width at narrowest part constitutes 1/10 of anal diameter. Cervical setae situated less regularly in nematodes described by Filip'ev compared to specimens described by Steiner. Cervical setae of this species situated two or three cephalic diameters from anterior end. Cephalic capsule very poorly developed and its cuticle appears to be a direct extension of thicker cuticle forming buccal capsule. Renette occupies 1/8 of esophagus, is slightly shifted toward right side, and opens at base of buccal capsule. Uteri in female short. Vaginal glands well developed.

Most important characteristics of this species: 1) highly elongated and slender body; 2) location of renette; 3) short tail; and 4) situation of cervical setae.

- 123 **Geographic distribution.** Found in the Barents Sea and western part of the Baltic Sea, predominantly in the coastal zone among algae and at a depth of about 30 m in a sandy bed.

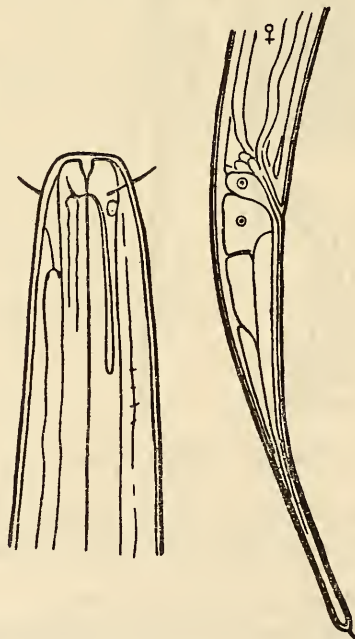


Figure 57. *Anticoma arctica* (from Steiner, 1916).

45. *Anticoma limalis* Bastian, 1865 (Figure 58)⁴

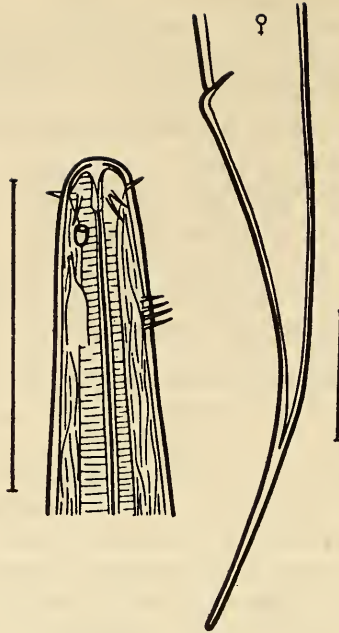
Bastian, 1865: 141, pl. XI, figs. 146–148; Bütschli, 1874: 271, 272, tab. IV, fig. 19a–c; Savel'ev, 1912: 108; Filip'ev, 1927: 76, pl. 12, fig. 3a–d; Allgen, 1931a: 97; 1931b: 211; 1932: 399; 1933: 11–12; 1935: 14–15; 1940b: 443; 1943: 267; 1954a: 5; 1957b: 6; Schuurmans-Stekhoven 1946: 35.

| | | | | | | | | | |
|---------|--|-------|---------|---------|-------|-------|-------|-------------|-----------------------|
| 1 ♀: | 5 | 22 | 214 | 430 | 935 | 1,185 | 1,500 | 2,569 | 2,927 μm ; |
| | 10 | 12 | 17 | 51 | 70 | 72 | 85 | 73 | |
| | a=34; b=7; c=8; V=40%. | | | | | | | | |
| 8 juv.: | 4–5 | 16–18 | 200–210 | 320–450 | — | — | — | 1,810–2,000 | |
| | 8 | 10 | 15–17 | 42–45 | 48–52 | 58–60 | — | 42–45 | |
| | × 2,110–2,305 μm ; a=36–38; b=5–7; c=7–8. | | | | | | | | |

Body tapers to 1/8 midbody diameter at anterior end and 5/12 at posterior end. Head slightly truncated anteriorly. Tail long; slender cylindrical part equal in length to conical part. Cuticle 2.0 to 3.0 μm thick. Cephalic capsule extremely thin; width twice length. Amphids situated almost immediately behind cephalic setae; extremely small, 3.0 μm × 2.0 μm , and somewhat longitudinally elongated. Length of cephalic setae 6.9 μm (slightly more than 1/2 corresponding diameter). Cervical setae commence three cephalic diameters from anterior end; length of setae 5.0 μm . Excretory pore of renette opens midway between cephalic and cervical setae. Oral cavity longitudinally elongated; walls sclerotized. Nerve ring encircles esophagus slightly anterior to its midlength. Length of anterior female genital tube 250 μm , of posterior tube 315 μm , and of reflexed parts 220 μm and 300 μm respectively. Eggs 100 μm × 70 μm . Vulva shifted anteriorly rather far away from midbody.

This species, as observed by Filip'ev (1927), resembles *A. pellucida* in size, various indexes (except c), and position of excretory pore of renette and vulva. The two species differ from each other in shape and size of tail (tail of *A. limalis* significantly longer and slender), location of amphids (in *A. limalis* situated somewhat nearer to anterior end), and length of cephalic setae (shorter in *A. pellucida*). They are more readily distinguished from each other by structure of spicular apparatus and gubernaculum.

⁴The taxonomic position of such species as *Anticoma acuminata*, *A. pellucida*, *A. limalis*, and *A. similis* has long been ambiguous. The first three species have been merged time and again, then separated on the basis of such characters as position of the excretory pore, length of tail, and position of the accessory organ in males (de Man, 1886; Filip'ev, 1918, 1927; Micoletzky, 1924, 1930; Schuurmans-Stekhoven, 1935a, 1950; and others). I have left the question of the identity of these species open because my material on *A. limalis* and *A. pellucida* is insufficient to determine their age and individual variability. Representatives of *A. acuminata* and *A. similis* were unfortunately absent in my material.



123

Figure 58. *Anticoma limalis*.

As there were no males in my material I cannot elaborate on a comparison of these two species.

Geographic distribution. Considering the information given in the literature, this species may be considered ubiquitous. However, it is found quite often in the Arctic, North Sea, and north Atlantic Ocean. Hence, I am inclined to consider *A. limalis* arctico-boreal. Information about its occurrence in other parts of the world oceans could possibly be based on the fact that some very similar species of *Anticoma* have been recorded under the name *A. limalis*.

46. *Anticoma pellucida* Bastian, 1865 (Figure 59)

Bastian, 1865: 142, pl. XI, figs. 149, 150; de Man, 1886: 53, figs. 9, 10; Ditlevsen, 1919: 159-161; 1923: 182; 1926: 20; Allgen, 1927a: 49; 1927b: 249; Schuurmans-Stekhoven, 1950: 29, 30, fig. 3A-B.

1 ♀: $\frac{5 \quad 11 \quad 193 \quad 408 \quad 714 \quad 969 \quad 1,224 \quad 2,210}{9 \quad 13 \quad 15 \quad 45 \quad 66 \quad 71 \quad 76 \quad 66 \quad 45}$ 2,440 μm ; a=32;
b=5; c=10; V=40%.

Body tapers to 1/8 midbody diameter at anterior end and 10/17 at posterior end. Head anteriorly round. Tail relatively small compared to

other species (significantly smaller than in *A. limalis*); conical part equal to cylindrical part in length; small bulge present at its terminus. Cuticle thin, only $2.3\ \mu\text{m}$. Walls of cephalic capsule distinctly visible in spite of insignificant size of head. Amphids located one cephalic diameter from anterior end, longitudinally elongated, and 2.3 to $3.4\ \mu\text{m}$ long. Length of cephalic setae $4.6\ \mu\text{m}$ ($1/3$ corresponding head diameter). Cervical setae $5.8\ \mu\text{m}$ long and commence 2.4 cephalic diameters from anterior end. Excretory pore of renette situated as in *A. limalis*, midway between cervical and cephalic setae. Oral cavity, as in *A. limalis*, also significantly elongated. Nerve ring encircles esophagus a little above its midlength. Length of anterior and posterior genital tubes of females $255\ \mu\text{m}$, and reflexed parts $178\ \mu\text{m}$. One egg, $102\ \mu\text{m} \times 51\ \mu\text{m}$, detected in genital tract. Vulva, as in *A. limalis*, significantly shifted anterior to midbody.



Figure 59. *Anticoma pellucida*.

Species highly similar to *A. limalis*; differences indicated under description of *A. limalis*.

Geographic distribution. Arctic and northern Atlantic, in the east up to the Kara Sea, and in the south up to the northern coast of France. 125 Extends even to western part of the Baltic Sea and the Mediterranean

Sea. Southernmost point of distribution is the western coast of Africa (Canary Islands). Allgen's (1927) report of this species in the subantarctic near Campbell Island appears dubious.

*47. *Anticoma novozemelica* (Filipjev, 1927) (Figure 60)

Filip'ev, 1927: 77, pl. 2, fig. 4a-c (*A. limalis* var. *novozemelica*) (description).

| | | | | | | | |
|------|---|-------|-------|-------|---------|-------------|-------|
| 2 ♂: | — | 20-23 | 320 | 730 | — | 3,065-3,260 | |
| | 13-14 | — | 35-82 | 75-82 | 100-102 | 52-55 | |
| | × 3,540-3,620 μm; a=35-36; b=5; c=8-10. | | | | | | |
| 1 ♀: | — | 330 | 715 | 1,300 | 1,800 | 2,160 | 3,230 |
| | 14 | 57 | 75 | — | 102 | — | 53 |
| | 3,760 μm; a=37; b=5; c=7. | | | | | | |

Filip'ev observed great similarity between this species and *A. limalis*. *A. novozemelica* differs from *A. limalis* in larger size, round and larger size of amphids (diameter 5.0 μm), and thinner walls of head. Oral cavity much more extensive than in *A. limalis*, and its thick sclerotized lining forms a small blunt outgrowth at the base. Spicules with velum and 80 μm long, or 1.7 to 1.8 times anal diameter. Gubernaculum relatively long, narrow, sharply expanded in distal part, and devoid of processes.

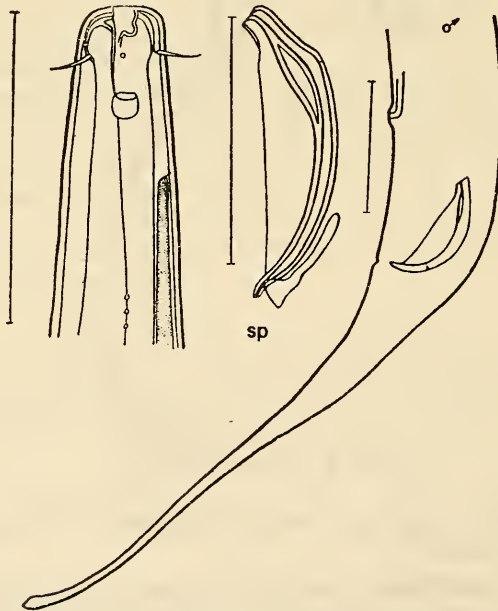


Figure 60. *Anticoma novozemelica* (from Filip'ev, 1927).

Geographic distribution. This species was recorded from the shores of Novaya Zemlya in the Strait of Matochkin Shar in the coastal zone in rhizoids of *Laminaria* sp.

48. *Anticoma brevisetosa* Platonova, 1967 (Figure 61)

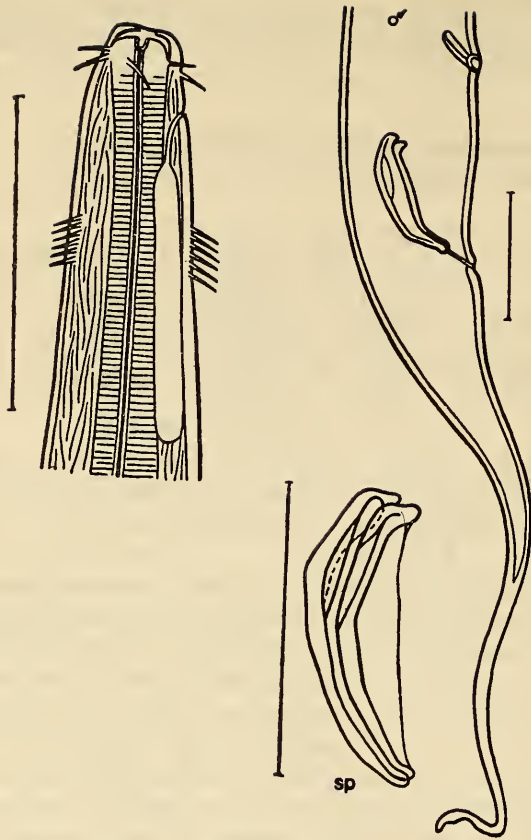
Platonov, 1967: 835, figs. 3-5.

| | | | | | | | | | | | |
|--|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-------------------------|-------------------------|-------------------------|-----------------------|---------|--------|--------|
| 1 ♂: | $\frac{5}{8}$ | $\frac{13}{11}$ | $\frac{160}{13}$ | $\frac{500}{40}$ | — | $\frac{2,100}{78}$ | $\frac{2,100}{48}$ | 2,355 μm ; | a = 39; | b = 5; | c = 9. |
| 7 ♀: | $\frac{4-5}{9-10}$ | $\frac{11-13}{11-12}$ | $\frac{13-15}{13-15}$ | $\frac{220-230}{48-51}$ | $\frac{425-440}{65-72}$ | $\frac{800-820}{62-78}$ | | | | | |
| × | $\frac{1,050-1,100}{70-80}$ | $\frac{1,300-1,396}{63-78}$ | $\frac{1,825-1,927}{40-51}$ | 2,080-2,222 μm ; | | | | | | | |
| a = 27-29; b = 5; c = 8; V = 49-50%. | | | | | | | | | | | |
| 10 juv.: | $\frac{3-4}{4-5}$ | $\frac{5}{5}$ | $\frac{15-17}{10-11}$ | $\frac{137-142}{23-25}$ | $\frac{240-255}{25-28}$ | — | $\frac{663-680}{30-32}$ | $\frac{20-22}{20-22}$ | | | |
| × 790-810 μm ; a = 25-26; b = 3; c = 6-7. | | | | | | | | | | | |

Body tapers to 1/8 to 1/10 midbody diameter at anterior end and not less than 1/2 at posterior end. Head anteriorly truncated. Tail long; fili-form cylindrical part twice length of conical part. Cuticle 3.5 to 4.6 μm thick. Cephalic capsule barely distinguishable because minute in size; width at base twice its length. Amphids not observed. Length of cephalic setae 3.5 to 4.6 μm (1/3 corresponding head diameter). Cervical setae situated 36 to 42 μm from anterior end in groups of six (distance slightly more than three cephalic diameters). They are equal in length to cephalic setae. Excretory pore situated one cephalic diameter from anterior end. Oral cavity very small and cyathiform. Nerve ring encircles esophagus medially. Length of anterior female genital tube 250 to 280 μm , of posterior tube 240 to 306 μm ; and of reflexed parts 180 to 193 μm and 180 to 193 μm respectively [*sic*]. Two mature eggs, 102 μm × 61 μm , observed in each uterus of female. Male genital tubes commence at a distance of 640 μm from anterior end. Spicules 67 μm long, smoothly arcuate, widen at proximal end, and gradually taper toward distal end. Spicules also taper slightly toward their capituli. Large elongated-oval cleft present in thickened part of spicules. Gubernaculum extremely small, almost invisible. Accessory organ situated 80 μm anterior to anus.

Structurally close to *A. limalis*, *A. brevisetosa* differs in: 1) shorter setae; 2) large number of cervical setae; 3) extremely small size of oral cavity; 4) position of excretory pore (much closer to anterior end); and 5) spicules with reduced gubernaculum.

127 **Geographic distribution.** Found in the Gulf of Kola Bay of the Barents Sea at a depth of 3.0 to 5.0 m among *Lithothamnion murmanicum*.

Figure 61. *Anticoma brevisetosa*.

49. *Anticoma filipjevi* Platonova, 1967 (Figure 62)
Platonova, 1967: 835, figs. 6-8.

$$1 \text{ } \sigma: \frac{8 \quad 180 \quad 305 \quad 670 \quad - \quad 5,300}{15 \quad 17 \quad 51 \quad 70 \quad 85 \quad 90 \quad 65} \quad 5,840 \text{ } \mu\text{m}; \quad a=65; \quad b=9;$$

$$c=10.$$

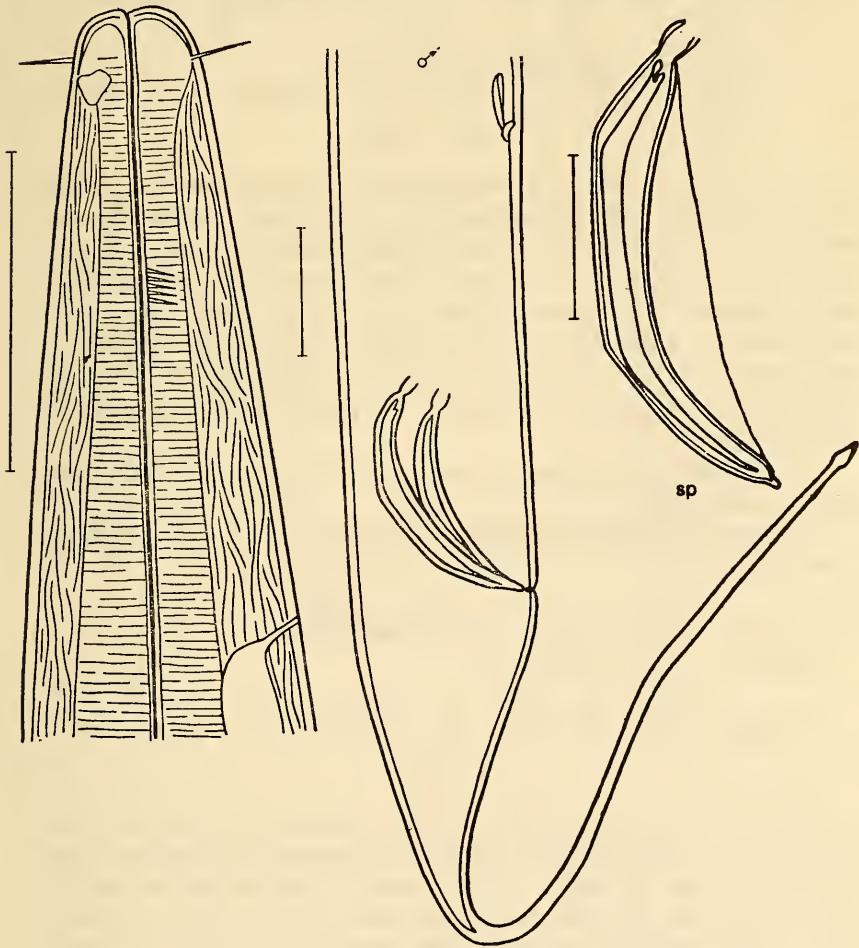
$$1 \text{ juv.}: \frac{6 \quad 215 \quad 280 \quad 610 \quad - \quad 3,610}{11 \quad 15 \quad 70 \quad 75 \quad 75 \quad 78 \quad 52} \quad 4,040 \text{ } \mu\text{m}; \quad a=51;$$

$$b=7; \quad c=8.$$

Body tapers to 1/5 midbody diameter at anterior end and 10/13 at posterior end. Head anteriorly rounded. Cylindrical part of tail twice longer than conical part. Cuticle 3.0 μm thick. Cephalic capsule very thin-walled; width three times length. Amphids situated immediately behind cephalic capsule. Cephalic setae rather long, 11 μm (about 1/2

corresponding head diameter). First of five cervical setae situated two cephalic diameters from anterior end. Excretory pore of renette situated significantly posterior to cervical setae, at 10 cephalic diameters from anterior end. Oral cavity almost undeveloped. Nerve ring encircles 128 esophagus somewhat anterior to its midlength. Spicules 95 μm long and curve at two points; capitulum small but distinctly developed. Spicular body dilates slightly and then tapers gradually toward distal end; wide slit extends along entire length of spicules. Velum extremely thin and narrow. Gubernaculum small and barely visible. Accessory organ 120 μm from anus.

Distinguishing features of this species: 1) large size; 2) location of



amphids immediately behind cephalic capsule; 3) relatively long cephalic setae; 4) position of excretory pore of cervical gland far away from cephalic end; and 5) shape of spicules—not smoothly arcuate but with a sharp bend at two points and presence of wide slit throughout their length. Structurally, *A. filipjevi* is close to *A. tenuicaudatoides*.

Geographic distribution. Found in the Kara Sea at a depth of 698 m in brown-gray silt.

*50. *Anticoma tenuicaudatoides* Gerlach and Riemann, 1974 (Figure 63) Filip'ev, 1946: 158, 159, fig. 1A-C (*tenuicaudata*) (description).

$$1 \text{ ♂: } \frac{35}{20} \frac{180}{35} \frac{305}{—} \frac{670}{70} \frac{—}{85} \frac{5,300}{90} \frac{64}{64} \quad 5,840 \text{ } \mu\text{m}; \quad a=65; \quad b=9; \\ c=18.$$

$$1 \text{ ♀: } \frac{35}{20} \frac{215}{35} \frac{280}{—} \frac{610}{75} \frac{2,630}{75} \frac{3,610}{52} \quad 4,100 \text{ } \mu\text{m}; \quad a=65; \quad b=9; \\ c=11; \quad V=64\%.$$

Body long. Tail extremely long (equal to 8.0 anal diameter in male and 9.5 in female), and very slender at tip, only 3.0 μm in width. Cuticle smooth and thin, only 3.0 to 4.0 μm . Cephalic setae about 10 to 11 μm , approximately equal to half cephalic width. Amphids small. Cervical setae commence 1.75 cephalic diameters from anterior end; they number four or five and differ slightly in size. Spicules usual in type and 95 μm long (equal to 1.4 anal diameter). Gubernaculum small, 30 μm long (equal to 0.4 anal diameter). Distance from anus to small anal tube 2.5 anal diameters.

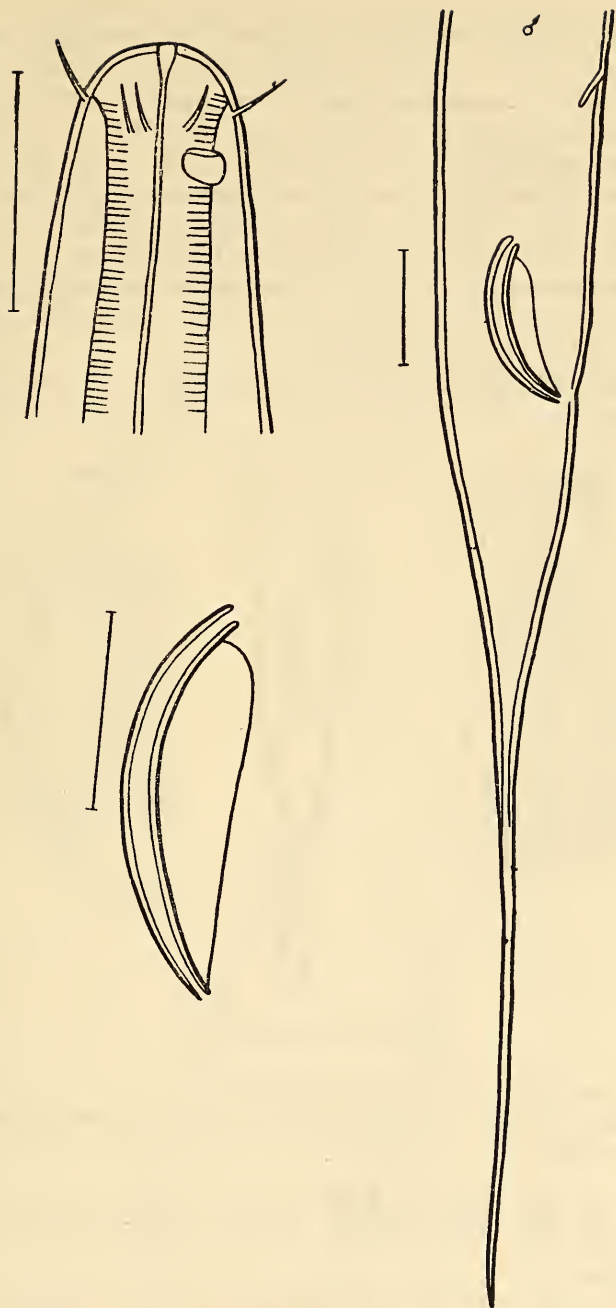
A. tenuicaudatoides belongs to that group of long-tailed species which possess a hind pore.

Geographic distribution. Found in the Kara Sea at a depth of 698 m in brown-gray silt.

51. *Anticoma grandis* Platonova, 1967 (Figure 64) Platonova, 1967: 835, figs. 9–11.

$$1 \text{ ♂: } \frac{8}{15} \frac{15}{17} \frac{270}{20} \frac{590}{60} \frac{—}{88} \frac{3,950}{105} \frac{65}{65} \quad 4,300 \text{ } \mu\text{m}; \quad a=41; \quad b=7; \\ c=12.$$

Body tapers to 1/7 midbody diameter at anterior end and 10/17 at posterior end. Head anteriorly flat. Tail rather long; conical part equal to cylindrical part in length. Cephalic capsule thin-walled and barely discernible; width twice length. Cuticle rather thick, 5.0 μm . Cephalic setae 6.0 μm (about 1/2 corresponding head diameter). Four cervical setae situated three cephalic diameters from anterior end; somewhat



smaller than cephalic setae, only $4.6 \mu\text{m}$ long. Amphids $5.0 \mu\text{m}$ and situated one cephalic diameter from anterior end. Excretory pore of renette opens almost at level of amphids. Oral cavity cyathiform and rather wide; wall highly sclerotized. Nerve ring encircles esophagus somewhat anterior to its midlength. Spicular length $105 \mu\text{m}$, capitulum undeveloped, and body with two curves. Wide slit extends along entire spicular length. Distinct wide velum with transverse striations present on ventral aspect of spicules. Gubernaculum small ($1/4$ spicular length) and simple in structure. Accessory organ situated $110 \mu\text{m}$ anterior to anus.

130 This species resembles *A. filipjevi* in size and external appearance. It differs in position of excretory pore of renette, well-developed and cyathiform oral cavity, and shape of spicules.

Geographic distribution. Found in the Kara Sea at a depth of 360 m in sandy silt.

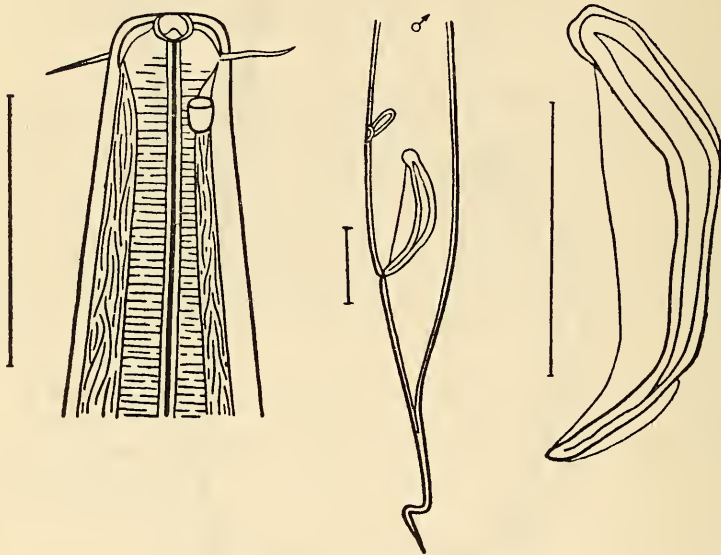


Figure 64. *Anticoma grandis*.

52. *Anticoma behringiana* sp. nov. (Figure 65)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7422.

| | | | | | | |
|---|----|-----|-----|----|-------|---------------------------------------|
| 5 | 19 | 229 | 499 | — | 2,250 | 2,540 μm ; a=26; b=6; c=9. |
| 9 | 12 | 15 | 68 | 85 | 97 | |

Paratypes.

| | | | | | | | |
|------|---|----|-----|-----|----|-------|---------------------------------------|
| 1 ♂: | 5 | 20 | 224 | 484 | — | 2,472 | 2,772 μm ; a=24; b=6; c=9. |
| | 8 | 12 | 16 | 45 | 68 | 75 | |

| | | | | | | |
|----------|--|-------|-----------------------------|---------|-----------------------------|-------------|
| 2 ♀: | 5-6 | 20-23 | 229-234 | 473-484 | 816-866 | 1,121-1,133 |
| | 8-9 | 12-14 | 16-17 | 45-47 | 63-65 | 72-74 |
| | × $\frac{1,401-1,442}{70-72}$ | | $\frac{2,064-2,163}{37-40}$ | | 2,400-2,771 μm ; | |
| | a=33-37; b=5-6; c=7-8; V=40-46%. | | | | | |
| 11 juv.: | 3-5 | 15-17 | 87-112 | 357-463 | — | 1,129-1,905 |
| | 6-8 | 8-12 | 13-15 | 19-25 | 47-62 | 50-95 |
| | 1,397-2,205 μm ; a=23-27; b=4-5; c=6-7. | | | | | |

Body slender and tapers to 1/5 to 1/7 midbody diameter at anterior end and 2/3 at posterior end. Head anteriorly truncated. Cylindrical part of tail 2.5 times longer than conical. Protuberance at caudal tip barely discernible. Cuticle thin, 2.3 μm . Cephalic capsule extremely thin; width at base twice length. Amphids, 5.8 μm \times 3.5 μm (1/4 corresponding diameter), situated 13 to 14 μm from anterior end (distance slightly more than one cephalic diameter). Length of cephalic setae 5.8 to 6.9

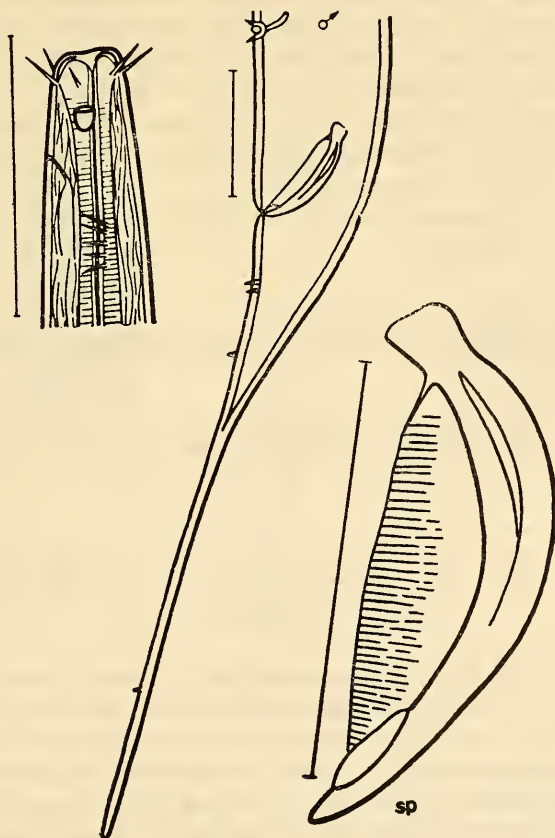


Figure 65. *Anticoma behringiana* sp. nov.

μm (1/2 corresponding head diameter). Cervical setae $5.0 \mu\text{m}$; anterior setae situated $34.8 \mu\text{m}$ from anterior end (distance equal to three cephalic diameters). Postanal part of tail covered with extremely short and sparse setae. Excretory pore of renette situated 19.7 to $23.2 \mu\text{m}$ from anterior end of body, i.e., about midway between amphids and first cervical seta. Oral cavity small and weakly sclerotized. Nerve ring encircles esophagus somewhat below its midlength. Length of anterior female genital tube 217 to $255 \mu\text{m}$, of posterior tube 280 to $309 \mu\text{m}$, and of reflexed parts 200 to $214 \mu\text{m}$ and 205 to $229 \mu\text{m}$ respectively. Vulva rather significantly shifted anterior to midbody. Females with mature eggs not detected. Spicules rather narrow, arcuate, slightly thickened in anterior third, and $60 \mu\text{m}$ long. Spicular capitulum cloven near top. Arcuate cleft evident in thickened part of spicules. Velum extremely thin. Gubernaculum minute, almost invisible. Accessory organ situated $70 \mu\text{m}$ anterior to anus.

A. behringiana is close to *A. brevisetosa* in structure, size, and indexes. However, body tapers much less from midregion to anterior end (five to seven times less). Cephalic setae longer and excretory pore of cervical gland situated farther from anterior end than in *A. brevisetosa*. Vulva significantly shifted anterior to midbody in *A. behringiana* but lies exactly midbody in *A. brevisetosa*. This species also differs in spicular shape and situation of accessory organ.

Geographic distribution. Found in the Bering Strait at a depth of 48 m in dark gray silted sand.

132 53. *Anticoma curta* sp. nov. (Figure 66)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7433.

$$\frac{4 \quad 9 \quad 175 \quad 372 \quad - \quad 1,442}{9 \quad 10 \quad 11 \quad 52 \quad 65 \quad 69 \quad 44} \quad 1,604 \mu\text{m}; a=23; b=4; c=8.$$

Paratype.

$$1 \text{ ♀: } \frac{5 \quad 13 \quad 175 \quad 382 \quad 626 \quad 806 \quad 1,112 \quad 1,622}{9 \quad 11 \quad 15 \quad 40 \quad 61 \quad 57 \quad 85 \quad 73 \quad 40} \quad 1,810 \mu\text{m}; \\ a=21; b=5; c=10; V=45\%.$$

Extremely small nematodes, comparable to minute *A. minor* in size. Body tapers to 1/8 to 1/9 midbody diameter at anterior end and 5/13 to 10/27 at posterior end. Head anteriorly truncated. Narrow cylindrical part of tail equal to conical in length; protuberance at terminal part barely distinguishable. Cuticle 2.3 to $3.4 \mu\text{m}$ thick. Cephalic capsule thin-walled; width at base twice length. Amphids in female (not detected in male) situated two cephalic diameters from anterior end, slightly

longitudinally elongated, and $3.5 \times 2.3 \mu\text{m}$. Cephalic setae 1.2 to $2.3 \mu\text{m}$ ($1/5$ to $1/6$ corresponding head diameter). Cervical setae fine and commence 29.0 to $34.8 \mu\text{m}$ from anterior end (distance equal to 3.0 to 3.7 cephalic diameters). Cervical setae equal to cephalic setae in length. Excretory pore of renette situated slightly more than one cephalic diameter from anterior end. Oral cavity relatively large and cyathiform. Nerve ring encircles esophagus somewhat anterior to its midlength. Length of anterior female genital tube $280 \mu\text{m}$, of posterior tube $306 \mu\text{m}$, and of reflexed parts $229 \mu\text{m}$ and $105 \mu\text{m}$ respectively. Only one egg, $102 \mu\text{m} \times 40 \mu\text{m}$, observed in genital tract of female. Spicules $42 \mu\text{m}$ long and relatively wide. Abrupt dilatation, occupying one-third spicular length, follows narrow capitulum; after dilatation spicules again narrow abruptly. Rather wide cleft along side of spicule. Velum not wide and may be thin. Gubernaculum reduced. Accessory organ situated $60 \mu\text{m}$ anterior to anus.

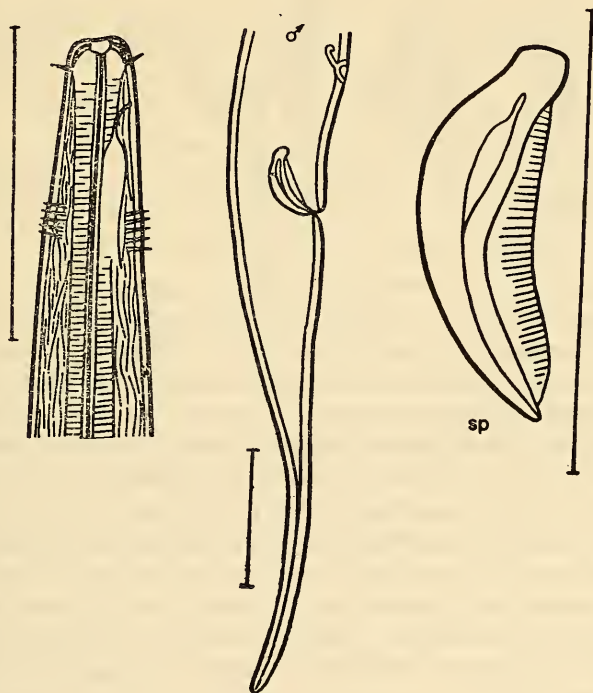


Figure 66. *Anticoma curta* sp. nov.

To some extent this species is close to *A. minor* but relatively thicker (in *A. minor*, $a = 32-35$; in *A. curta*, $a = 16-21$) and with shorter cephalic setae (equal to $1/3$ to $1/4$ corresponding diameter). These two species

differ in shape of oral cavity (funnel-shaped in *A. minor* and cyathiform in *A. curta*). Finally, it is very easy to distinguish these species on the basis of spicular structure.

Geographic distribution. Found in the Bering Strait at a depth of 48 m in dark gray sandy silt.

54. *Anticoma uschakovi* sp. nov. (Figure 67)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7467.

$$\frac{6 \quad 17 \quad 280 \quad 535 \quad - \quad 2,595}{11 \quad 15 \quad 19 \quad 85 \quad 120 \quad 130 \quad 70} \quad 2,857 \mu\text{m}; \quad a=22; \quad b=5; \quad c=10.$$

Paratypes.

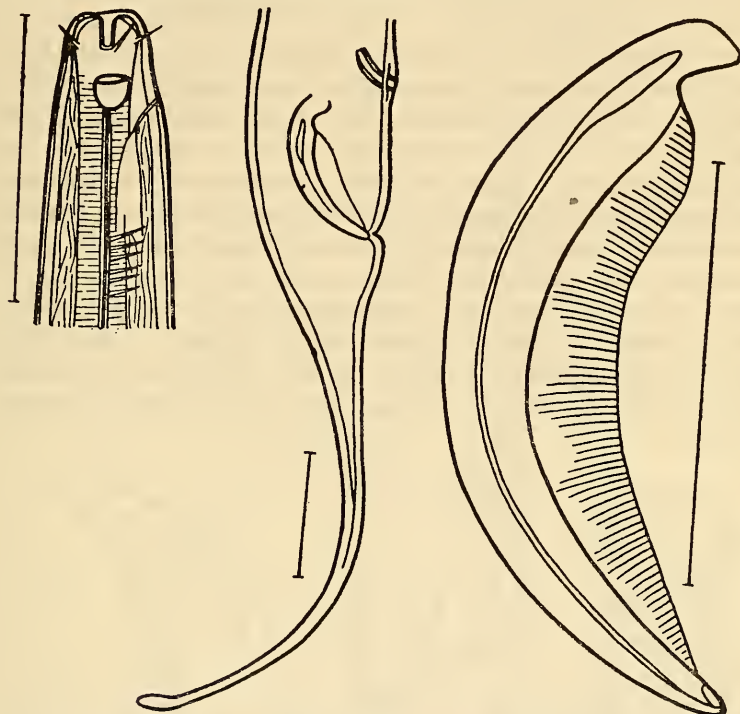
$$1 \text{ } \sigma: \frac{7 \quad 18 \quad 295 \quad 556 \quad - \quad 2,678}{10 \quad 14 \quad 20 \quad 71 \quad 96 \quad 112 \quad 51} \quad 2,978 \mu\text{m}; \quad a=26; \quad b=5; \\ c=10.$$

$$5 \text{ } \varnothing: \frac{5-7 \quad 16-18 \quad 280-300 \quad 570-618 \quad 1,040-1,071}{10-11 \quad 15-16 \quad 18-20 \quad 65-70 \quad 97-100 \quad 112-117} \\ \times \frac{1,460-1,500 \quad 1,933-1,989 \quad 2,882-2,987}{112-117 \quad 107-110 \quad 27-51} \quad 3,100-3,287 \mu\text{m}; \\ a=27-28; \quad b=5; \quad c=10-11; \quad V=46-47\%.$$

$$5 \text{ } \text{juv.}: \frac{4-5 \quad 12-15 \quad 153-158 \quad 387-392 \quad - \quad 2,026-2,266}{8 \quad 8-12 \quad 13-15 \quad 31-34 \quad 37-40 \quad 37-42 \quad 30-32} \\ \times 2,213-2,456 \mu\text{m}; \quad a=52-60; \quad b=6; \quad c=12-13.$$

- 134 Body tapers to 1/10 midbody diameter at anterior end and 1/2 at posterior end. Head anteriorly truncated. Filiform cylindrical part of tail twice length of conical part. Distinct thickening evident at caudal terminus. Cuticle 5.8 μm thick. Cephalic capsule with distinctly visible walls; width twice length. Amphids situated 11.6 μm (two cephalic diameters) from anterior end of body; width of amphids 8.2 μm (1/2 corresponding body diameter). Length of cephalic setae 5.8 μm (1/3 corresponding head diameter). First five cervical setae situated 34 to 40 μm from anterior end (three to four cephalic diameters). Excretory pore of renette situated near posterior margin of amphids. Oral cavity rectangular, with sclerotized walls, spacious, and reaches level of cephalic setae. Nerve ring encircles esophagus at midlength. Length of anterior female genital tube 413 to 429 μm , of posterior tube 480 to 500 μm , reflexed parts almost reach vulva. Not more than one egg, 137 $\mu\text{m} \times 66 \mu\text{m}$, seen in genital tract of female. Length of spicules 75 μm ; shape smoothly arcuate. Large ventral hump forms an abrupt dilatation in spicule in proximal part next to spicular capitulum. Narrowing of spicule more gradual and stops in proximal third. Remaining part of spicule narrow but more or less equal in width throughout length of spicular body. Velum narrow and thin. Accessory organ situated 75 μm anterior to anus.

Distinguishing features of this species: 1) well-developed oral cavity; 2) characteristic shape of tail (conical part extremely short and changes rather abruptly into cylindrical part, which is twice its length; and 3) spicules very large and different in shape.



133

Figure 67. *Anticoma uschakovi* sp. nov.

Geographic distribution. Found in the Bay of Nagaev (Sea of Okhotsk) at a depth of 27 m among pebbles and silt.

55. *Anticoma pacifica* sp. nov. (Figure 68)

Holotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7931.

| | | | | | | |
|----|----|-----|-----|-----|-------|----------------------------|
| 14 | 28 | 286 | 572 | — | 3,658 | 3,990 μm; a=24; b=7; c=12. |
| 28 | 32 | 44 | 100 | 130 | 166 | |

Paratypes.

| | | | | | | |
|------|--|-------|---------|---------|---------|---------|
| 4 ♂: | 14-16 | 20-28 | 200-300 | 598-638 | — | |
| | 25-30 | 32-26 | 42-48 | 98-110 | 104-109 | 132-143 |
| | × $\frac{3,125-3,524}{72-78}$ 4,055-4,414 μm; a=31-32; b=6-8; c=14-17. | | | | | |

$$4 \text{ ♀: } \frac{14-16}{24-30} \quad \frac{21-26}{31-38} \quad \frac{210-400}{40-46} \quad \frac{665-731}{90-100} \quad \frac{1,928-2,527}{111-117} \quad \frac{143-145}{143-145}$$

$$\times \frac{4,455-5,785}{57-72} \quad 4,774-6,104 \text{ } \mu\text{m};$$

$$a=32-42; b=7-9; c=15-19; V=40-42\%.$$

Body tapers to $2/9$ to $1/5$ midbody diameter at anterior end. Head anteriorly truncated. Length of cylindrical part of tail about equal to conical part. Distinct bulge evident at tip of tail. Cuticle $6.0 \text{ } \mu\text{m}$ thick. Cephalic capsule with rather thick walls; width 2.5 length. Amphids small, longitudinally elongated, and only $6.0 \text{ } \mu\text{m}$ wide ($1/3$ to $1/4$ corresponding neck diameter). Length of cephalic setae $12 \text{ } \mu\text{m}$ (slightly less than $1/2$ corresponding head diameter). Five cervical setae situated two cephalic diameters from anterior end. Renette short, pore opens at level of amphids. Oral cavity developed. Oral opening encircled by distinct lips. Nerve ring encircles esophagus more or less at midlength. Vulva slightly anterior to midbody length. Length of anterior female genital tube $480 \text{ } \mu\text{m}$, of posterior tube $500 \text{ } \mu\text{m}$. Two to seven eggs, varying in size (114 to

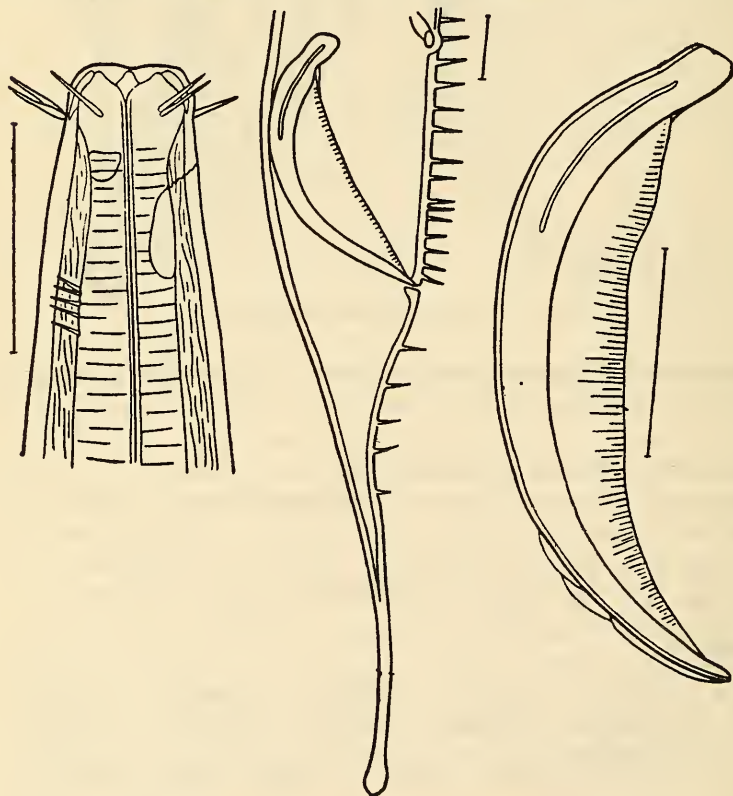


Figure 68. *Anticoma pacifica* sp. nov.

135 163 $\mu\text{m} \times 80 \mu\text{m}$ and 60 to 80 $\mu\text{m} \times 140 \mu\text{m}$), observed in genital tract of females. Spicules without capituli but with transversely striated velum; length of spicules 140 to 160 μm . Gubernaculum narrow and thin. Accessory organ 130 to 160 μm anterior to anus. Anal region of male armed with setae 13 μm long. Twenty pairs of preanal setae present, two pairs postanal, and single seta on tail.

Distinguishing features of this species: 1) presence of lips; 2) large size of body; 3) presence of anal setae in male; and 4) transverse striations on velum.

Geographic distribution. Found on the southern coast of Sakhalin in the Gulf of Aniva at a depth of 15 m among pebbles and fine sand.

56. *Anticoma pontica* Filipjev, 1918 (Figure 69)

Filip'ev, 1918: 66-69, tab. 2, fig. 6.

$$1 \text{ } \sigma: \frac{6 \quad 20 \quad 164 \quad 350 \quad - \quad 2,000}{9 \quad 13 \quad 18 \quad 42 \quad 45 \quad 72 \quad 45} \quad 2,210 \mu\text{m}; \quad a=30; \quad b=6; \quad c=10.$$

$$3 \text{ } \varnothing: \frac{5-7 \quad 19-22 \quad 168-183 \quad 360-381 \quad 935-950 \quad 1,200-1,212}{8-9 \quad 12-14 \quad 18-20 \quad 44-46 \quad 45-50 \quad 68-70 \quad 76-81}$$

$$\times \frac{1,525-1,562 \quad 2,256-2,298}{72-80 \quad 32-34} \quad 2,561-2,611 \mu\text{m};$$

$$a=32-33; \quad b=6-7; \quad c=8; \quad V=46-47\%.$$

$$4 \text{ } \text{juv.}: \frac{4-5 \quad 16-18 \quad 152-157 \quad 316-326 \quad - \quad 1,750-1,802}{6-8 \quad 10-11 \quad 16-17 \quad 32-34 \quad 38-40 \quad 60-62 \quad 30-32}$$

$$\times 1,950-2,002 \mu\text{m}; \quad a=32-34; \quad b=6; \quad c=10.$$

Body tapers to 1/9 to 1/10 midbody diameter at anterior end and 2/3 to 1/2 at posterior end. Head anteriorly round with slight flattening. Tail of female slightly longer than that of male. Narrow cylindrical part of tail equal in length to conical part. Cuticle 1.2 μm thick. Cephalic capsule poorly developed and barely distinguishable. Amphids round and 4.0 μm in diameter (1/4 corresponding neck diameter). Length of cephalic setae 7.0 μm (1/2 corresponding head diameter); five cervical setae commence three cephalic diameters from anterior end. Renette situated at posterior end of esophagus, with an extremely long duct opening 1.5 cephalic diameters from anterior end. Oral cavity small. Nerve ring encircles esophagus slightly anterior to its midlength. Length of anterior female genital tube 262 to 265 μm , of posterior tube 325 to 350 μm , and of reflexed parts 200 to 220 μm and 300 to 320 μm respectively. One egg, 75 $\mu\text{m} \times 42 \mu\text{m}$, seen in genital tract of one female. Spicules 80 μm long and curve mainly in anterior third. Capitulum of spicules elongated and rectangular. Velum located on ventral side of spicule. Gubernaculum small, 30 μm , envelops distal end of spicules, and broadens distally. Accessory organ situated 90 μm anterior to anus.

As noted by Filip'ev (1918), this species is very close to *A. pellucida* but differs in: 1) almost round amphids (longitudinally elongated in *A. pellucida*); 2) excretory pore of renette situated relatively close to anterior end; 3) spicules significantly narrower, not so sharply curved, and with a rectangular capitulum and velum. Gubernaculum without ventral process and accessory organ situated at level of spicular capitulum (in



Figure 69. *Anticoma pontica*.

A. pellucida spicules sharply curved in middle part, their capitulum rounded, and ventral velum absent). Gubernaculum with ventral process. Accessory organ situated two anal diameters from anus.

Geographic distribution. Possibly endemic to the Black Sea as has not been found beyond this limit.

57. *Anticoma platonovae* Sergeeva, 1972 (Figure 70)
Sergeeva, 1972: 1233, 1234, fig. 1 (description).

| | | | | | | |
|-----------------------|--|---------|---------|---------|-------|-------------|
| $3 \text{ } \sigma$: | — | 195-222 | 240-259 | 565-607 | — | 1,907-2,124 |
| | 5-6 | 32-34 | 34-46 | 45-48 | 49-52 | 34-38 |
| | $\times 2,008-2,254 \text{ } \mu\text{m}$; a=42-44; b=3-4; c=17-20. | | | | | |

Nematodes of average size. Body tapers to 1/8 to 1/9 midbody diameter at anterior end and 5/7 at posterior end. Cuticle smooth and $1.9 \text{ } \mu\text{m}$ thick. Head anteriorly rounded. Cephalic setae short. Cervical setae and amphids situated $14 \text{ } \mu\text{m}$ from anterior end (2.1 cephalic diameters). Size of amphids $5.0 \text{ } \mu\text{m}$. Excretory pore of cervical glands opens not less than 32.8 to 37.5 cephalic diameters from anterior end, and $45.5 \text{ } \mu\text{m}$ anterior to nerve ring. Nerve ring encircles esophagus 42.2 to $44.8 \text{ } \mu\text{m}$ anteriorly from its midlength. Oral cavity undeveloped. Esophagus widens toward base. Spicules curved in first and last quarters. Dorsal side of spicules smooth. On ventral side spicular capitulum forms a platform with a triangular process at its base. Velum extends from capitular tip. Guber-

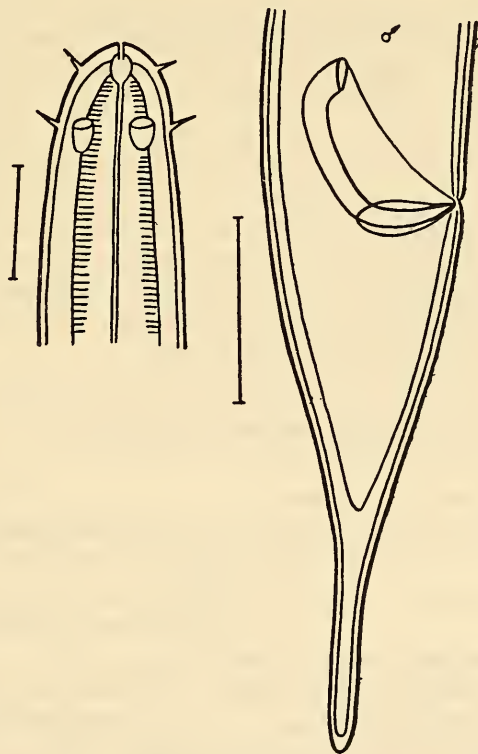


Figure 70. *Anticoma platonovae* (from Sergeeva, 1972).

naculum foliate and contiguous. Length of spicules 45.5 μm , of gubernaculum 17.0 μm . Two to three preanal setae present in male; accessory organ absent. Tail short; cylindrical part 5/7 of conical part.

Geographic distribution. Found in the Black Sea on the western coast (at a depth of 83 m in phaseolin silt), the southern coast (region of Yalta city in a depth range of 50 to 130 m in medium and phaseolin silts), and along the coast of Crimea.

137 *A. reflexa* Linstow, 1896, *A. filicauda* Mawson, 1956, and *A. calveti* Rouville, 1903 have not been included in the identification key due to their extremely brief and incomplete descriptions. *A. longicauda* Kreis, 1928 was described on the basis of females and the author assumed it might coincide with some known species in the future. *A. brevicauda* Kreis, 1963 was described on the basis of a sexually immature specimen, precluding its inclusion in the identification key.

5. Subfamily PLATYCOMINAE subfam. nov.

Nematodes of average size. Body does not taper toward both ends as much as in representatives of previous subfamily. Some forms with well-developed labial cuticular outgrowths. Cephalic setae long. Cervical setae numerous and situated in clusters or transverse rows. Amphids usually large; their structure tends to indicate sexual dimorphism: in males amphids significantly larger or with more complex configuration than in females. Renette not developed. Nerve ring situated at approximately midlength of esophagus. Tail rather short and wide, often pointed at end. Spicules usually small or reduced; gubernaculum highly developed, almost double spicular length. Additional copulatory armature always present and represented by one to two papillate accessory organs. Postanal papillae with setae may also be present.

138

Key to Genera of Subfamily Platycominae

- 1 (4). Lips with cuticular outgrowths.
- 2 (3). Two flat setae situated near each amphid...**Platycoma** Cobb, 1894.
(one species, **P. cephalata** Cobb, 1894).
- 3 (2). Four setae situated near each amphid.....
.....10. **Platycomopsis** Ditlevsen, 1926.
- 4 (1). Lips without cuticular outgrowths.
- 5 (6). Gubernaculum with dorsal process. Amphids in males differ in shape from those in females..... **Proplatycoma** gen. nov.
- 6 (5). Gubernaculum without process. Amphids in males same shape as in females, but larger..... **Pilosinema** gen. nov.

10. Genus *Platycomopsis* Ditlevsen, 1926

Ditlevsen, 1926: 30; Filip'ev, 1927: 81 (*Dactylonema*).

Type species: *P. cobbi* Ditlevsen, 1926.

Head demarcated from body by a distinct cervical constriction. Tail short, wide, and obtusely rounded or conical. Lips with cuticular outgrowths. Transverse row of four long setae situated posterior to each amphid. Slightly posterior to these, two subdorsal and subventral setae occur. Belt of granular cells situated in anterior part of body. Amphids very large, goblet-shaped. Spicules short. Gubernaculum curves slightly, without process. Accessory organ small, shifted rather far anterior to anus.

In the descriptions of species numbers in Cobb's formula denote distance from anterior end to: 1. posterior end of cephalic capsule; 2. amphids; 3. nerve ring; 4. base of esophagus; 5. commencement of anterior genital tube; 6. vulva; and 7. commencement of posterior genital tube. Number of figures in formula fewer for males (see earlier explanation).

Key to Species of Genus Platycomopsis

- 1 (2). Cephalic setae thin; length equal to cephalic diameter
 **P. cobbi** Ditlevsen, 1926.
 2 (1). Cephalic setae thick; length not more than two-thirds cephalic diameter.
 3 (4). Width of amphids exceeds one-half of corresponding diameter
 58 **P. mesjatzevi** Filipjev, 1927.
 4 (3). Width of amphids less than one-third of corresponding diameter
 **P. nuchapilosus** Vitiello, 1970.

58. *Platycomopsis mesjatzevi* (Filipjev, 1927) (Figure 71)

Filip'ev, 1927: 82-84, pl. I, fig. 10a-c (*Dactylonema*).

Lectotype ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7174.

| | | | | | | |
|-------|----|-----|-------|-----|-------|----------------------------|
| 17 | 23 | 463 | 1,235 | — | 6,436 | 6,648 μm; a=27; b=5; c=31. |
| 23-32 | 34 | 142 | 202 | 244 | 150 | |

Paralectotypes.

| | | | | | | | | |
|------|-------|-----|-------|-------|-------|-------|-------|-----------|
| 1 ♀: | 17 | 566 | 1,390 | 2,574 | 3,655 | 4,839 | 7,208 | 7,415 μm; |
| | 23-34 | 147 | 202 | 214 | 229 | 225 | 132 | |

a=32; b=5; c=35; V=49%.

| | | | | | | |
|------|-------|-------|---------|-------------|---------|---------|
| 2 ♂: | 17 | 23 | 465-515 | 1,235-1,339 | — | |
| | 19-23 | 30-34 | 34 | 142-162 | 200-212 | 225-244 |

$$\times \frac{6,436-7,261}{132-150} \quad 6,648-7,441 \mu\text{m}; \quad a=27-32; \quad b=5-6; \quad c=31-41.$$

$$10 \text{ juv.:} \quad \frac{13-15}{15-17} \quad \frac{29-32}{22-24} \quad \frac{412-463}{28-31} \quad \frac{1,235-1,390}{104-105} \quad \frac{\quad}{150-167} \quad \frac{\quad}{150-177}$$

$$\times \frac{4,892-5,870}{70-100} \quad 5,042-6,040 \mu\text{m}; \quad a=25-34; \quad b=5-6; \quad c=33-38.$$

- 139 Body tapers to 1/6 to 2/13 midbody diameter at anterior end and 5/8 to 10/17 at posterior end. Cephalic end rounded. Tail conical, rounded at tip. Cephalic capsule thin-walled, tapers slightly anteriorly, and width twice greater than length. Narrow cephalic ring situated in middle of cephalic capsule. Cephalic suture very narrow and barely distinguishable. Cuticle bilayered and 6.0 to 8.0 μm thick; thickness of outer layer 4.0 μm . Oral opening with three sclerotized lips. Cephalic setae thick and long, 19 to 25 μm (1.5 corresponding head diameter). Cervical setae slightly thinner and shorter, 15 to 17 μm . Anal setae absent. Extremely large amphids, transversely elongated, situated 23 μm from anterior end (1/6 esophageal length). Length of amphids 6.0 μm and width 17.4 μm (1/2 corresponding neck diameter). Three minute denticles situated in anterior part of small oral cavity. Esophagus dilates slightly anteroposteriorly and only at very base forms rather significant dilatation. Nerve ring situated slightly posterior to midesophagus. Vulva situated anterior to midbody; cuticle around it with sclerotized granules. Spicules extremely short (60 μm long), wide, uniform in width throughout their length, and bluntly rounded at proximal end. Gubernaculum paired, 137 to 160 μm

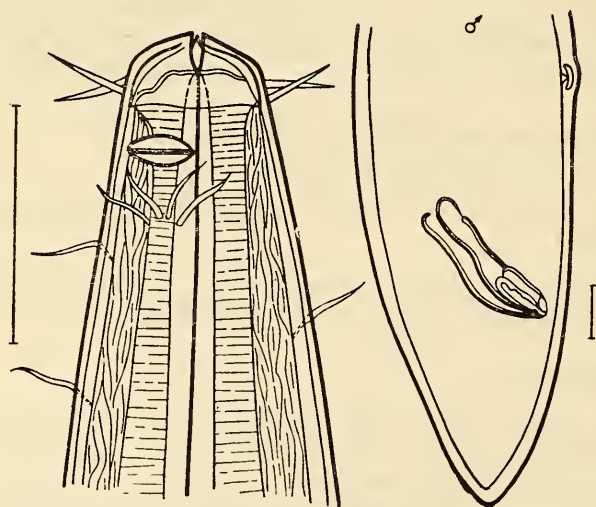


Figure 71. *Platycomopsis meszatevi*.

long, faintly curved, and narrows perceptibly in middle part. Accessory organ situated 229 to 280 μm anterior to anus.

Distinguishing features of this species: 1) extremely large amphids, elongated in transverse direction; 2) cephalic setae thick and long, exceeding corresponding diameter in length; 3) presence of group of four long setae situated behind each amphid; 4) narrow cephalic ring situated in middle of cephalic capsule; 5) presence of sclerotized granules in female on cuticle around vulva; and 6) conical tail rounded at tip.

Geographic distribution. Known only from the Barents Sea where it was found in a depth range of 30 to 200 m in silted sand.

Genus *Proplatycoma* gen. nov.

Type species: *Platycoma sudafricana* Inglis, 1966.

Nematodes characterized by sharp sexual dimorphism in structure of amphids. In female amphids typically cyathiform but slightly elongated in longitudinal direction. In male anterior end of amphids elongated anteriorly and sometimes with unique processes. Cuticular labial outgrowths absent. Cervical setae may be numerous and may be scattered over the preneural region in groups or singly.

Spicules small, blunt, and reduced. Gubernaculum significantly longer than spicules, curved in the middle, with large blunt dorsal process. Tail wide, terminally acicular.

Key to Species of Genus *Proplatycoma* gen. nov.

- 1 (2). Anterior part of amphids in male branched.....
..... *P. curiosa* (Gerlach, 1955).
- 2 (1). Anterior part of amphids in male unbranched.
- 3 (4). Anterior part of amphids in male conically pointed.....
..... *P. africana* (Gerlach, 1959).
- 4 (3). Anterior part of amphids in male arrow-shaped.....
..... *P. sudafricana* (Inglis, 1966).

Genus *Pilosinema* gen. nov.

Type species: *Platycomopsis filiappendicatus* Micoletzky, 1930.

Nematodes devoid of cuticular labial outgrowths. Characteristic sexual dimorphism. Amphids in male much larger than in female. Cervical setae situated in transverse rows behind amphids: three rows in male and two in female. In addition, small groups of setae scattered over preanal region. Spicules and gubernaculum straight and simple. Usually spicular apparatus difficult to detect. Accessory organ small and papil-

late. Tail distinctly divided into wide conical part and narrow cylindrical part which, however, may be extremely short.

Key to Species of Genus Pilosinema gen. nov.

- 1 (2). Tail conical, without cylindrical part **P. paracobbi** (Mawson, (1956).
 2 (1). Tail conical, with cylindrical terminal part.
 3 (4). Conical part of tail twice length of cylindrical part. Accessory organ of male two spicular lengths anterior to anus **P. filiappendicatus** (Micoletzky, 1930).
 4 (3). Conical part of tail thrice length of cylindrical part. Accessory organ of male one spicular length anterior to anus **P. dimorphica** (Mawson, 1956).

6. Subfamily BARBONEMATINAE subfam. nov.

Rather small nematodes. Body tapers markedly toward anterior end; tapering may be gradual or with abrupt constriction in preneural part. Tail tapers gradually and is rather slender. Oral cavity and lips not developed. Long cephalic setae arranged in two, three, or four circles. Cervical setae usually shorter than cephalic ones and quite sparse. Amphids observed in only one of the species described. Renette apparently absent. Position of nerve ring could not be determined. Spicules small and curved. Gubernaculum reduced. Accessory organ absent.

141 Probably the peculiarity of arrangement of cephalic setae made Gerlach (1956, 1957) transfer *Barbonema* to Oxystominidae. This appears questionable to me since in the type species (*B. setifera*, described by Filip'ev) the amphids have an entirely different shape than those in oxystominids. In the species described by Gerlach—*B. horridum* and *B. flagrum*—nothing is said about the structure of the amphids. These two species very much resemble each other in structure of the cephalic end and arrangement of setae, but differ greatly from *B. setifera*, so much so that they warrant placement in an independent genus. The structure of the spicular apparatus, however, brings *B. horridum* close to *B. setifera*.

Due to the fact that only the three species mentioned above are known at present, it seems advisable to me to postpone division of *Barbonema* until the discovery of a larger number of forms permits an improvement in the taxonomic system of barbonematines.

*11. Genus *Barbonema* Filipjev, 1927

Filip'ev, 1927: 81 (description).

Body tapers markedly toward anterior end. Head devoid of lips and oral capsule; armed with ten well-developed setae and one seta with a pyriform base situated posterior to the others. Lateral organ saccate. Spicules of male curved.

*59. *Barbonema setifera* Filipjev, 1927 (figure 72)

Filip'ev, 1927: 81, pl. 2, fig. 6a-c; (description).

$$1 \text{ ♂: } \frac{\text{---} \quad 90 \quad 480 \quad \text{---} \quad 2,180}{9 \quad 40 \quad 42 \quad 43 \quad 27} \quad 2,320 \text{ } \mu\text{m}; \quad a=53; \quad b=5; \quad c=16.$$

Body slender, filiform. Tail long, filiform, with a terminal bulge. Caudal length $52 \text{ } \mu\text{m}$; diameter at caudal end $0.23 \text{ } \mu\text{m}$. Cuticle smooth, thin, and 1.0 to $1.5 \text{ } \mu\text{m}$ thick. Head round; lips and papillae absent. Cephalic setae very long, $25 \text{ } \mu\text{m}$ (three cephalic diameters); four setae slightly more posterior than remaining six. Amphids situated immediately behind pyriform seta. Cardia very small. Intestine rudimentary. Spicules curved, S-shaped, flat in proximal part, and $35 \text{ } \mu\text{m}$ long. Gubernaculum and accessory organ absent.

Geographic distribution. Found only in the Barents Sea at a depth of 200 m in silted sand.

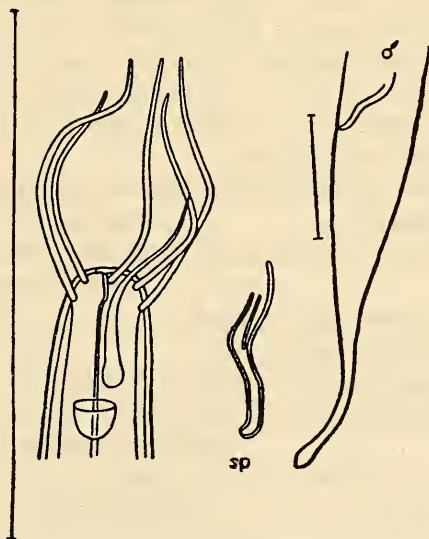


Figure 72. *Barbonema setifera*
(from Filip'ev, 1927).

2. Superfamily **ENOPLOIDEA** Baird, 1853

Sclerotized cephalic capsule not developed in most representatives. Oral capsule in most cases well developed and divided into buccal and onchial parts, as a result of which armature of oral cavity represented not only by onchia, but also by powerful jaws or buccal stylets—derivatives of buccal cavity. Entire esophageal musculature at anterior end of body, compared with that of *Leptosomatoidea*, appears markedly displaced posteriorly. Alveolate structure often observed in posterior half of esophagus. Cuticle similar in structure to that in representatives of superfamily *Leptosomatoidea* but significantly thinner. Amphids round or cyathiform.

Key to Families of Superfamily Enoploidea

- 1 (6). Oral cavity well developed, with powerful sclerotized lining, and often armed with well-developed onchia.
- 2 (3). Oral opening encircled by well-developed protruding lips; internal surface of latter often with distinct cuticular striations. 2. **Rhabdodemaniidae** de Coninck, 1955.
- 3 (2). Well-developed protruding lips absent.
- 4 (5). Sclerotized annular thickenings present on wall of oral cavity. Sexual dimorphism seen in structure of oral cavity. Latter absent in males of a number of forms. **Enchelidiidae** Filipjev 1918 (part.).
- 5 (4). Annular thickenings on walls of oral cavity absent. Sexual dimorphism not observed in structure of oral cavity. **Oncholaimidae** (Perrier, 1897).
- 6 (1). Oral cavity less distinct; wall poorly sclerotized.
- 7 (10). Oral cavity armed with jaws.
- 8 (9). Three distinct onchia present, of which one significantly larger than remaining two. **Oxyonchidae** de Coninck, 1965.
- 9 (8). Onchia absent; if present, small and uniform in length. **Enoplidae** Baird, 1853.
- 10 (7). Oral cavity unarmed; if armed, only buccal stylets present or structures similar to them.
- 11 (14). Cephalic capsule well developed and conical.
- 12 (13). Onchial plates twisted into tubes which look like spears in lateral view. **Thoracostomopsidae** Filipjev, 1927.
- 13 (12). Oral cavity with another type of armature, more often three small onchia. **Phanodermatidae** Filipjev, 1927.
- 14 (11). Cephalic capsule not developed or insignificantly developed and different in shape.
- 15 (16). Esophagus commences at a notable distance from anterior end

- of body and buccal stylets situated anterior to it.....
**Dayeliidae** fam. nov.
 16 (15). Esophagus commences at anterior end; buccal stylets absent.
 17 (18). Esophagus alveolate in posterior half. Spicules thin and long.
**3. Crenopharyngidae** fam. nov.
 18 (17). Esophagus not alveolate. Spicules short.....
**Enchelidiidae** Filipjev, 1918.
 (males of some genera).

3. Family CRENOPHARYNGIDAE fam. nov.

Body fusiform due to tapering toward anterior and posterior ends. Tail divided into wide conical and narrow cylindrical parts. Cephalic capsule almost undeveloped. Oral cavity small; onchial part absolutely undeveloped and buccal part poorly developed. Cephalic setae arranged in single circle. Renette present. Amphids always cyathiform.

On the basis of the foregoing characters this family resembles Anticomidae. Nevertheless a series of extremely substantial features prompted me not only to isolate Crenopharyngidae in an independent family, but even to transfer it to another superfamily.

Spicules in crenopharyngids extremely long and narrow. Accessory organ absent. Esophagus with alveolate structure in posterior half.

The family comprises two genera.

Key to Genera of Family Crenopharyngidae

- 1 (2). Two well-defined eyes present in preneural area.....
**13. Nasinema** Filipjev, 1927.
 2 (1). Eyes always absent.....**12. Crenopharynx** Filipjev, 1934.

12. Genus *Crenopharynx* Filipjev, 1934

Linstow, 1900: 128 (*Anoplostoma*); Southern, 1914: 23 (*Stenolaimus* non Marion, 1870); Filip'ev, 1934a: 9.

Type species: *Anoplostoma gracile* Linstow, 1900.

Cephalic capsule very poorly developed, with narrow cephalic ring. Lips encircling oral opening poorly developed. Labial papillae very small. Cervical setae either irregularly scattered over preneural part or totally absent. Amphids usually small, round, and situated in region of cephalic suture. Tail long; cylindrical part usually filiform. Spicules very long and narrow, with small and distinct capituli. Gubernaculum small, often with process directed dorsally. Eyes absent.

In descriptions of species of genus *Crenopharynx* figures in Cobb's

formula denote distance from anterior end of body to the following organs: 1. posterior end of cephalic capsule; 2. nerve ring; 3. base of esophagus; 4. commencement of anterior genital tube; 5. vulva; 6. commencement of posterior genital tube; and 7. anus.

Number of figures for males fewer than for females (see p. 74).

Key to Species of Genus Crenopharynx

- 1 (2). Well-defined dilatation at distal end of spicules.....61. **C. gracilis** Linstow, 1900.
 2 (1). Dilatation on distal end of spicules absent.
 3 (8). Cervical setae present.
 4 (5). Filiform cylindrical part of tail constitutes two-thirds of total caudal length. .**C. paralepturus** (Schuurmans-Stekhoven, 1950).
 5 (4). Filiform cylindrical part of tail constitutes not more than one-half of total caudal length.
 6 (7). Body narrow ($a=50-60$).....**C. metagracilis** (Schuurmans-Stekhoven, 1950).
 7 (6). Body wide ($a=25$).....**C. brevicaudatus** (Schuurmans-Stekhoven, 1950).
 8 (3). Cervical setae absent.
 9 (10). Oral cavity armed with onchia.....62. **C. armatus** sp. nov.
 10 (9). Oral cavity without onchia.
 11 (12). Cephalic setae not longer than one-third corresponding diameter.....**C. antarcticus** (Allgen, 1929).
 12 (11) Cephalic setae significantly long, constituting not less than one-half corresponding diameter.
 144 13 (14). Conical part of tail narrows abruptly into filiform cylindrical part.....**C. crassus** (Ditlevsen, 1930).
 14 (13). Conical part of tail reduces gradually into filiform cylindrical part.
 15 (18). Male with preanal setae.
 16 (17). Tail long (c =about 13).....60. **C. marioni** (Southern, 1914).
 17 (16). Tail significantly short (c =about 19).....**C. eina** Inglis, 1964.
 18 (15). Male without preanal setae.....**C. afra** Inglis, 1964.

60. *Crenopharynx marioni* (Southern, 1914) (Figure 73)

Southern, 1914: 23, tab. 4, fig. 12a-b; Ditlevsen, 1926: 10-11, pl. II, fig. 5, pl. III, figs. 3, 8, pl. IV, figs. 4, 5, pl. V, fig. 2, pl. VI, fig. 1; Schuurmans-Stekhoven, 1935a: 7, fig. 67, 1946: 32-33, fig. 7a-d; Allgen, 1954a: 12-13; 1957b: 6 (*Stenolaimus*).

| | | | | | |
|--|-------------|---------------|-------------|-----------------------------|-------------|
| 10 ♂: | 10-11 | 100-100 [sic] | 610-620 | — | 6,280-6,300 |
| | 13-15 19-20 | 42-48 | 90-96 | 200-210 | 100-105 |
| × 6,690-6,700 μm ; a = 30-34; b = 6; c = 13-16. | | | | | |
| 10 ♀: | 9-11 | 112-120 | 710-740 | 2,289-2,340 | |
| | 12-13 22-23 | 46-48 | 102-105 | 248-262 | |
| × | 3,099-3,145 | 4,250-4,380 | 5,942-6,112 | 6,442-6,617 μm ; | |
| | 230-238 | 220-226 | 103-110 | | |
| a = 26-29; b = 6; c = 13-15; V = 45-47%. | | | | | |
| 10 juv.: | 9-10 | 95-100 | 450-480 | — | 4,203-4,305 |
| | 12-13 13-17 | 38-40 | 60-70 | 170-192 | 80-100 |
| × 4,503-4,607 μm ; a = 24-26; b = 5; c = 12-15. | | | | | |

Body tapers to 1/14 to 1/17 midbody diameter at anterior end and 1/2 at posterior end. Tail long, with bulge at terminus; narrow cylindrical part somewhat longer than conical part. Cephalic capsule relatively well developed; width twice length. Cuticle thick, 5.8 μm . Cephalic setae long, 11.6 μm (1/2 corresponding head diameter). Setae absent in esophageal region. Setae 8.0 μm long, sparse in anal region. Amphids situated immediately behind lateral cephalic setae and slightly elongated longitudinally; diameter of amphids 4.6 μm (1/5 corresponding neck diameter). Cervical glands situated asymmetrically: one opens two cephalic diameters, and the other five cephalic diameters from anterior end. Oral cavity small, funnel-shaped, with thickly sclerotized walls. Nerve ring

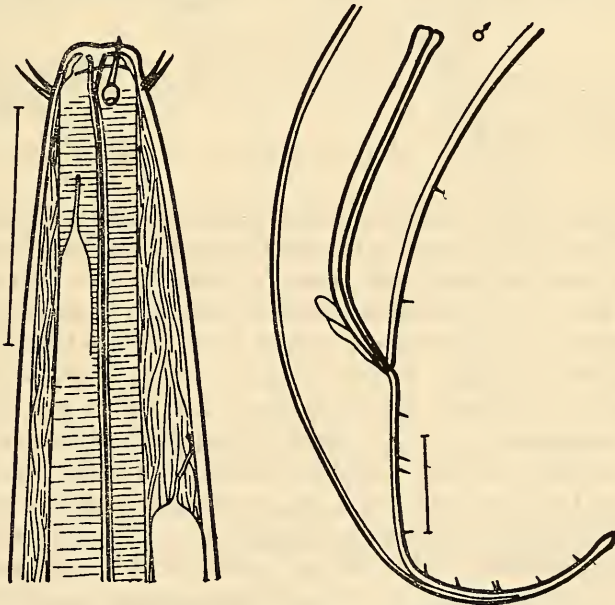


Figure 73. *Crenopharynx marioni*.

culum approximately equal to anal diameter and consists of two plates, which encircle terminal part of spicules from dorsal side; distal part of gubernaculum more complex and encircles spicules from lateral sides. Two to six eggs, 196 to 260 μm \times 10 to 170 μm , found in uteri.

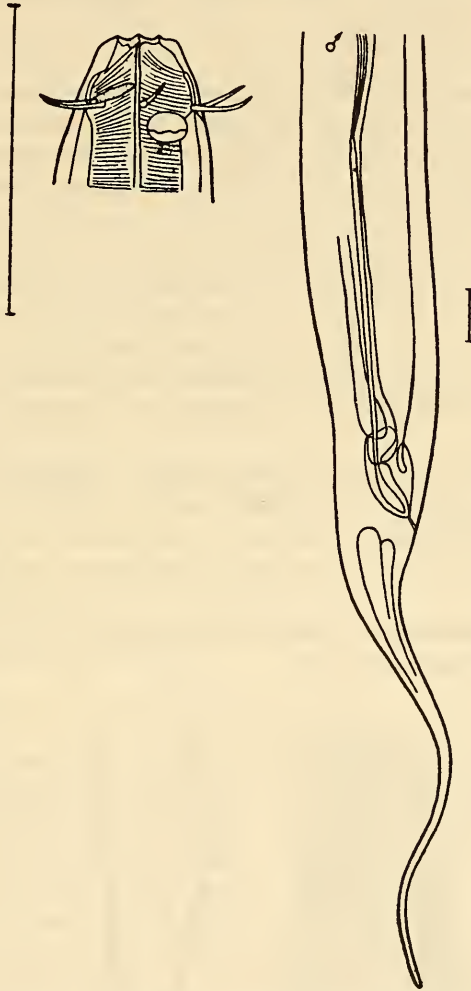


Figure 74. *Crenopharynx gracilis* (from Filip'ev, 1927).

This species is similar to *C. marioni* but differs in longer cephalic setae, more displaced cervical pore, slender tail, and absolutely different structure of spicules.

Geographic distribution. Found in the Barents and Kara Seas in silted beds in the sublittoral zone.

62. *Crenopharynx armatus* sp. nov (Figure 75)

Holotype ♀: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7546.

| | | | | | | | |
|-------------------|-----|-----|-------|-------|-------|-------|-----------------------------|
| 11 | 257 | 618 | 1,030 | 2,008 | 3,141 | 4,274 | 4,480 μm ; a=24; |
| 15 | 27 | 95 | 142 | 185 | 182 | 182 | |
| b=7; c=21; V=46%. | | | | | | | |

Body tapers to 1/12 midbody diameter at anterior end and 1/2 at posterior end. Head truncated. Tail short; narrow cylindrical part 1/4 length of conical part. Cephalic capsule well developed; width 2.4 length. Length of cephalic setae 11.6 μm (slightly less than 1/2 corresponding head diameter). Cuticle thick, 5.8 μm . Amphids round and small; diameter 4.0 μm (1/8 corresponding neck diameter). Pore of cervical gland situated 1.5 cephalic diameters from anterior end. Oral cavity rather spacious, with small onchia. Nerve ring encircles esophagus slightly anterior to its midlength. Length of anterior female genital tube 978 μm , of posterior tube 1,133 μm , and of reflexed parts 824 μm and 1,030 μm respectively. Vulva situated slightly anterior to midbody. No mature eggs observed in uterus.

In spite of only two females at my disposal, features differentiating this species from others of this genus could be delineated: 1) extremely short tail ($c=21$); 2) relatively well-developed cephalic capsule with distinct cephalic ring and thick walls; and 3) well-defined oral cavity, armed with onchia.

Geographic distribution. Found in the Barents Sea in the littoral zone at a depth of 18 m in a sandy bed.

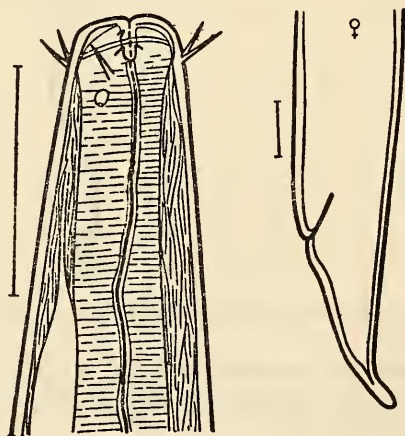


Figure 75. *Crenopharynx armatus* sp. nov.

13. Genus *Nasinema* Filipjev, 1927

Steiner, 1916: 596 (*Enchelidium*); Filip'ev, 1927: 118.

Type species: *Enchelidium polare* Steiner, 1916.

- 147 Cephalic capsule totally reduced. Lips and labial papillae not developed. Tail perceptibly shorter than in preceding genus, without filiform part, but with clavate bulge on which tube of excretory duct of cervical [sic] gland terminates. Cervical setae totally absent. Structure of amphids not known. Well-developed pigmented eyes present; pigment bowl cyathiform. Spicules long and narrow. Gubernaculum simple, without processes.

Key to Species of Genus Nasinema

- 1 (2). Small nematodes, body length about 2.0 mm.....
N. **stenolaima** Wieser, 1954.
 2 (1). Larger nematodes, body length 4.0 to 5.0 mm.
 3 (4). Body extremely slender (a =about 80)... 64. N. **boreale** sp. nov.
 4 (3). Body considerably wider (a =about 30).....
63. N. **polare** (Steiner, 1916).

*63. *Nasinema polare* (Steiner, 1916) (Figure 76)

Steiner, 1916: 596-600, tab. 16, fig. 28b, c, f, tab. 30, fig. 28a, d, e, g, h (*Enchelidium*) (description: 596).

1 ♀: L=5 mm; a = 32; b = 4; c = 18; V = 50%.

Body tapers toward both ends, but more significantly toward anterior end (about 1/9) than posterior end (about 1/2). Head anteriorly truncated. Tail tapers gradually from anus, without distinct filiform part, and terminates in rather wide tube. Buccal cavity small, funnel-shaped, with highly sclerotized walls. Esophageal lumen bounded by extremely thin walls up to its midlength, where walls become highly sclerotized; posterior third of esophagus differs significantly in alveolate structure. Nerve ring encircles esophagus about at its midlength. Excretory pore of renette situated 150 μ m from anterior end. Eyes situated 40 μ m from anterior end. Steiner assumed the presence of light-refractive lenses in these eyes, but in the specimen on which the description of the species is based these lenses were not discernible. Pigment bowls of eyes cyathiform. Vulva situated immediately posterior to midbody. Female genital tubes reflexed; reflexed part reaches uterus.

Geographic distribution. Discovered in the isthmus of the White Sea.

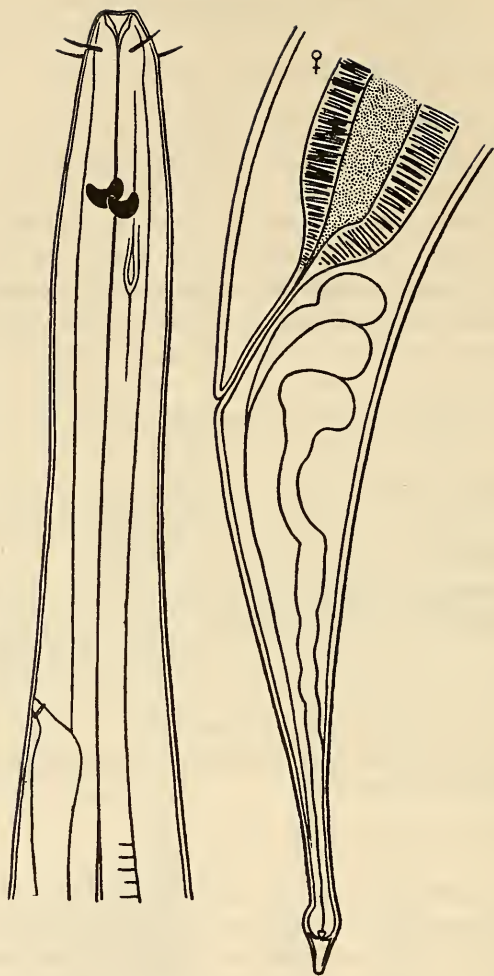


Figure 76. *Nasinema polare* (from Steiner, 1916).

148 64. *Nasinema boreale* sp. nov. (Figure 77)

Holotype. ♂: Institute of Zoology, Academy of Sciences, USSR. Collection No. 7927.

| | | | | | |
|----|-----|-----|----|-------|---|
| 12 | 221 | 690 | — | 5,120 | 5,932 μm ; a=81; b=10; c=30. |
| 12 | 15 | 49 | 62 | 72 | |

Paratype.

| | | | | | | |
|------|----|-----|-----|----|-------|--|
| 1 ♂: | 12 | 351 | 731 | — | 4,389 | 5,308 μm ; a=66; b=7; c=27. |
| | 12 | 15 | 49 | 54 | 80 | |

Body tapers to $1/6$ midbody diameter at anterior end and $2/3$ at posterior end. Head truncated. Tail rather long, without filiform part. Cuticle $2.0 \mu\text{m}$ thick. Buccal cavity poorly developed. Length of cephalic setae $5.0 \mu\text{m}$ and cervical $3.0 \mu\text{m}$. Nerve ring encircles esophagus in its anterior third. Posterior part of esophagus alveolate. Eyes considerably displaced toward anterior end and with distinct lens. In one specimen pigment a compact, oval, longitudinally elongated mass, and in another specimen scattered in small spots all over anterior part of body. Excretory pore of renette situated $50 \mu\text{m}$ from anterior end. Spicules very long (200 to $230 \mu\text{m}$), narrow, with small capituli. Gubernaculum simple and small, length $40 \mu\text{m}$. Sparsely scattered setae $5.0 \mu\text{m}$ long located in anal region of male.

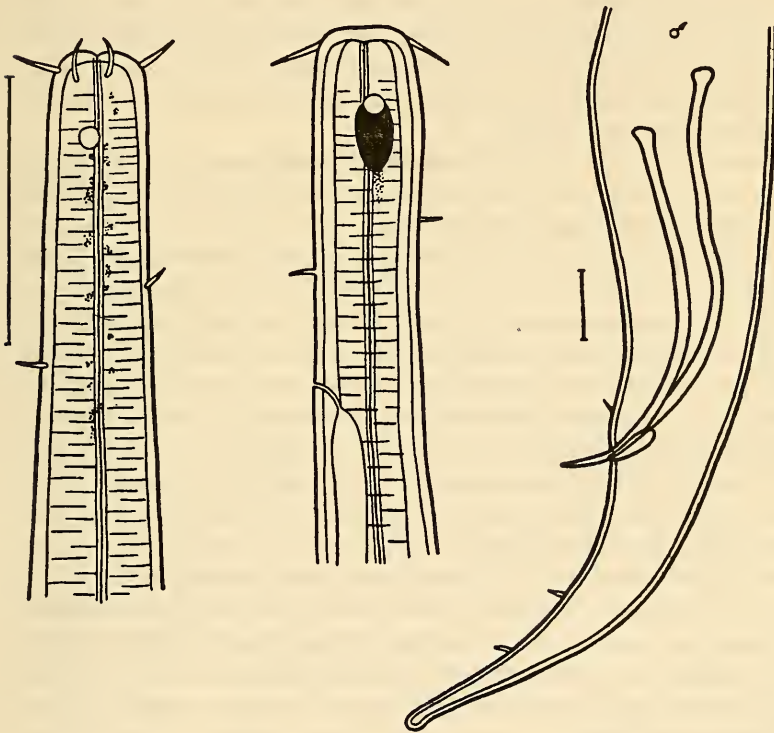


Figure 77. *Nasinema boreale* sp. nov.

N. boreale can be distinguished from the only other species known in the Soviet Union, namely *N. polare*, in position and shape of eyes, position of excretory pore of renette, more slender body, and shorter tail.

Geographic distribution. Found in the south terminus of Kunashir (Kuril Islands) at a depth of 5.0 m in sandy silt.

4. Family RHABDODEMANIIDAE Filipjev, 1934

This family includes only one genus—*Rhabdodemia*—and as such the basic characters coincide with those of the genus.

14. Genus *Rhabdodemia* Baylis and Daubney, 1926

Southern, 1914: 41 (*Demania* nec Lauvil, 1906); Ditlevsen, 1926: 15 (*Demania*); Baylis and Daubney, 1926: 112; Filip'ev, 1927: 90; Wieser, 1959: 9; Inglis, 1964: 322.

Type species: *Demania major* Southern, 1914.

Genus *Rhabdodemia* occupied an undefined taxonomic position for a long time. At present, there is a tendency to include it in family Enoplidae (de Coninck, 1965). I think this genus deserves isolation in an independent family which, indisputably, is phylogenetically close to family Enoplidae (Platonova, 1974).

Nematodes small in size, 2.8 mm long. Cuticle smooth. Body tapers slightly toward head but not toward anus; narrows slightly only in caudal region. Head distinctly demarcated from body. Oral cavity divided into buccal and onchial parts. Buccal cavity encircled by three well-developed lips with internal surface distinctly longitudinally striated. Striations vary in depth in different species. Sclerotized plate (mandible) enlarged and bifurcated in upper part, gradually narrows toward base, joins each lip from inner side. Anterior part of plate thickened and usually carries outgrowth directed outward (mandibular ring and mandibular outgrowth). Posterior part of mandible continues into onchial cavity where in most cases it is provided with an acicular and internally directed process, the onchium. Entire oral cavity with thick sclerotized lining, forming its wall. Cephalic end distinctly truncated. Tail relatively small, conical, and narrow slightly toward tip. Only cephalic setae present; cervical and preanal setae absent. Absence of amphids in type species is an interesting feature; no author has reported them to date. Esophagus and reproductive system as in leptosomatids, caudal glands tubular. Spicules weakly arcuate, wide in proximal part, and each terminates with distinct capitulum, which differs in structure in different species. Gubernaculum small, simple, and without process. Accessory organ absent.

In the descriptions of species of genus *Rhabdodemia*, figures in Cobb's formula denote distances from anterior end of body to the following parts: 1. posterior margin of oral cavity; 2. nerve ring; 3. esophageal base; 4. commencement of anterior genital tube; 5. vulva; 6. commencement of posterior genital tube; and 7. anus. As mentioned before, the number of measurements are fewer for males.

Key to Species of Genus *Rhabdodemanina*

- 1 (24). All cephalic setae equal in length.
- 2 (3). Tail short (c =about 40). Gubernaculum very long, almost equal to spicules in length.....
 **R. calycolaimus** Schuurmans-Stekhoven and Mawson, 1955.
- 3 (2). Tail much longer (c =about 20). Gubernaculum much shorter than spicules.
- 4 (5). Cephalic setae very long; length twice cephalic diameter.....
73. **R. angustissima** Platonova, 1974.
- 150 5 (4). Cephalic setae short; length significantly shorter than cephalic diameter.
- 6 (9). Oral cavity devoid of onchia.
- 7 (8). Lips poorly developed. Length of body 5.0 mm.....
 **R. striata** Schulz, 1932.
- 8 (7). Labia well-developed. Length of body 3.5 mm.....
67. **R. edentula** Platonova, 1974.
- 9 (6). Oral cavity armed with onchia.
- 10 (11). Spicular capitulum multiangular in shape.....
 **R. laticauda** Ditlevsen, 1926.
- 11 (10). Spicular capitulum undeveloped or different in shape (round and flattened ventrally or anteriorly).
- 12 (15). Small nematodes; length of body 2.0 to 3.0 mm.
- 13 (14). Six onchia situated in onchial cavity (two on each mandible)...
69. **R. hexonchia** Platonova, 1974.
- 14 (13). Three onchia situated in onchial cavity (one on each mandible).
70. **R. pontica** Platonova, 1965.
- 15 (12). Larger nematodes; length of body greater than 3.0 mm.
- 16 (17). Oral cavity very wide; length equal to width.....
68. **B. latifaux** Platonova, 1974.
- 17 (16). Oral cavity narrow; length exceeds width by 1.5 to 2.0 times.
- 18 (21). Spicules with well-developed capituli.
- 19 (20). Spicular capitulum round.....
 **R. scandinavica** Schuurmans-Stekhovveen, 1946.
- 20 (19). Spicular capitulum flattened on ventral side.....
71. **R. marisalbi** Platonova, 1974.
- 21 (18). Spicules without distinctly developed capituli.
- 22 (23). Cephalic setae thin, relatively long (length constitutes about one-half corresponding diameter). . 72. **R. orientalis** Platonova, 1974.
- 23 (22). Cephalic setae thick and short (length constitutes one-fifth corresponding diameter).....66. **R. gracilis** (Ditlevsen, 1919).
- 24 (1). Cephalic setae unequal in length; four submedian shorter than rest.

- 25 (34). Cephalic setae situated in one circle.
 26 (27). Extremely small nematodes; body length less than 2.0 mm. . . .
 **R. minima** Chitwood, 1936.
 27 (26). Nematodes significantly larger, body length greater than 3.0 mm.
 28 (29). Only one dorsal onchium present. **R. dura** Inglis, 1966.
 29 (28). Three onchia present.
 30 (31). All onchia equal in size. 65. **R. minor** (Southern, 1914).
 31 (30). Onchia differ in size; dorsal larger than ventrolateral.
 32 (33). Tail in male with large setae. Capitulum present.
 **R. mediterranea** Boucher, 1971.
 33 (32). Tail in male devoid of setae. Spicular capitulum absent.
 **R. nancyae** Inglis, 1964.
 34 (25). Cephalic setae situated in two circles.
 35 (36). Cephalic setae of anterior circle less than 1/3 length of setae in
 posterior circle. **R. illgi** Wieser, 1959.
 36 (35). Cephalic setae of anterior circle more than 1/2 length of setae in
 posterior circle or equal to them.
 37 (38). Spicules with distinctly developed capitulum.
 **R. major** (Southern, 1914).
 38 (37). Spicules without distinctly developed capitulum.
 **R. coronata** Gerlach, 1952.

151 65. *Rhabdodemanía minor* (Southern, 1914) (Figure 78)⁵

Southern, 1914: 43, tab. 7, fig. 21a-c (*Demanía*); Filip'ev, 1927: 92-93,
 pl. 3, fig. 9; Allgen, 1928: 283, 1929a: 10; 1935: 17; 1959: 29, fig. 11a-c.

| | | | | | |
|------|---|------------------|-------------------|------------------|----------------------|
| 2 ♂: | 18-19 19-20 | 102-240 55-62 | 440-660 75-80 | — 90-100 | 2,730-3,650 65-68 |
| | × 2,840-3,800 μm; a = 32-38; b = 6; c = 23-26. | | | | |
| 3 ♀: | 20-21 20 | 220-250 64-85 | 490-630 78-102 | 1,310-1,650 — | 1,620-2,010 — |
| | × 1,940-2,260 2,180-2,600 2,500-2,840 3,130-3,610 85-120 — — 60-92 × 3,270-3,760 μm; a = 31-39; b = 6-7; c = 23-27. | | | | |

Body rather short and thick. Cuticle thick, 5.0 μm. Cephalic setae differ in length; four submedian setae significantly shorter than remaining six. Onchia considerably displaced into depth of oral cavity. Cardia long and partially wedged into esophagus (up to 28 μm). Spicules straight, cuticular fold extends all along their length and over capitulum; spicular capitulum elongated. Eggs 200 μm × 70 μm.

⁵Description of species according to Southern (1914: 43) and Filip'ev (1927: 92, 93).

This species resembles *R. major* and *R. gracilis*. It differs from the former in: 1) shorter and thicker body; 2) length of esophagus and tail; and 3) length of cephalic setae. It differs from the latter in: 1) longer cephalic setae; 2) shape of tail; 3) thicker body; 4) straighter spicules; and 5) cuticular fold extending all along spicular body.

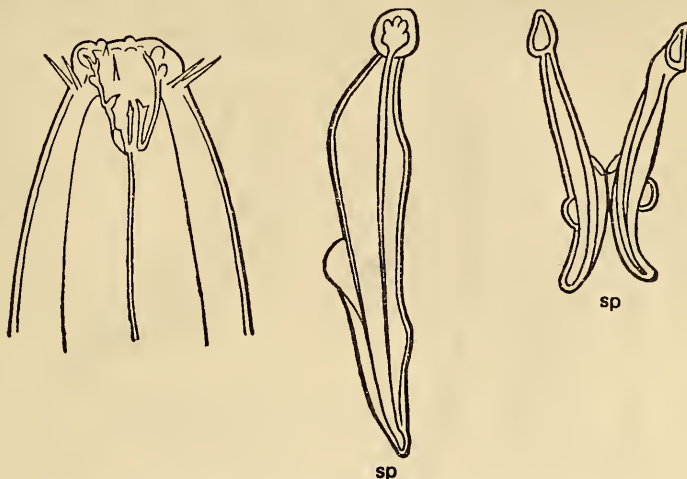


Figure 78. *Rhabdodemanian minor* (from Southern, 1914).

Geographic distribution. Probably an Arctico-boreal species as mainly found in the Barents, Norwegian, and Baltic Seas, and in the north Atlantic Ocean. The discovery of this species in the subantarctic by Allgen appears dubious.

66. *Rhabdodemanian gracilis* (Ditlevsen, 1919) (Figure 79)⁶

Ditlevsen, 1919: 187, tab. 10, fig. 6, tab. 11, figs. 1, 3, 7 (*Demanian*); Filip'ev, 1927: 91-92, pl. 3, fig. 8; Schuurmans-Stekhoven, 1935a; 12, fig. 73A-B; Allgen, 1935: 17.

| | | | | | | |
|-----|-----|--|---------|---------|-------------|-------------|
| 152 | 4 ♂ | 21-23 | 255-375 | 535-770 | — | 4,150-5,380 |
| | | 20-22 | 75-90 | 84-104 | 94-122 | 76-88 |
| | | × 4,300-5,550 μm; a = 34-55; b = 6; c = 23-33. | | | | |
| | 5 ♀ | 20-22 | 305-360 | 660-760 | 1,530-2,360 | 1,870-2,400 |
| | | 19-25 | 82-112 | 102-135 | — | — |
| | | × $\frac{2,620-3,630}{124-155}$ $\frac{3,090-4,540}{—}$ $\frac{3,480-5,150}{85-105}$ 5,000-6,220 μm; | | | | |
| | | a = 32-50; b = 7-9; c = 24-28. | | | | |

⁶Description of species according to Ditlevsen (1919: 187) and Filip'ev, (1927: 91-92).

Body short and thick; tapers considerably toward both ends but especially toward anterior end. Cephalic setae short and stout. Cuticle 5.0 to 7.0 μm thick. Weak pigmentation discernible in esophagus. Vulva situated somewhat posterior to midbody. Ovaries short; anterior one somewhat shorter than posterior one. Uteri large and highly elongated. Vulval glands and musculature well developed. Spicules curved, dilated in middle, and cuticular fold occupies two-thirds spicule length.

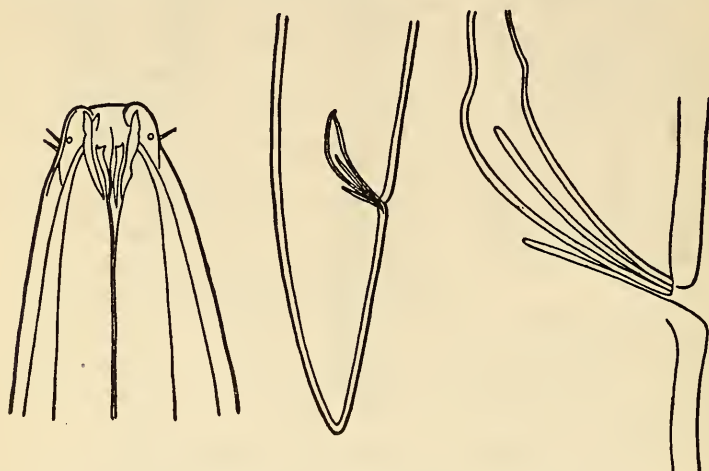


Figure 79. *Rhabdodemanía gracilis* (from Ditlevsen, 1919).

This species resembles *R. minor* in many respects; however distinguishing features between the two species have been given in their descriptions.

Geographic distribution. Found mainly in the western part of the Baltic Sea. Recorded by Filip'ev in the Kara and Barents Seas.

67. *Rhabdodemanía edentula* Platonova, 1974 (Figure 80)

Platonova, 1974: 1296, 1297, fig. 1.

| | | | | | |
|---------|---|---------|-------------|-------------|-------------|
| 5 ♂: | 15 | 187-190 | 480-595 | — | 2,300-4,253 |
| | — | 50-90 | 115-147 | 132-170 | 80-105 |
| | × 3,402-4,428 μm ; a = 26-28; b = 7-8; c = 23-25. | | | | |
| 4 ♀: | 15 | 515-550 | 1,170-1,246 | 1,840-2,018 | 2,540-3,254 |
| | 19 | 125-135 | 155-160 | 145 | 145 |
| × | $\frac{3,333-3,666}{90-100}$ 3,500-3,816 μm ; a = 24-26; b = 7; c = 20-25; V = 52-55%. | | | | |
| 4 juv.: | 13-14 | — | 370-412 | — | 1,918-1,957 |
| | 15-16 | — | 30-75 | 70-75 | 50 |
| | × 2,043-2,032 μm [sic]; a = 27-29; b = 5-6; c = 16. | | | | |

Body short, rather thick, and tapers considerably, to $1/6$ to $1/7$ mid-body diameter at anterior end and $5/8$ to $10/17$ at posterior end. Cuticle 153 7.0 to $8.0 \mu\text{m}$ thick. Tail posteriorly pointed. Cephalic setae 3.5 to $4.6 \mu\text{m}$ long ($1/4$ to $1/5$ corresponding head diameter). Lips well developed. Size of oral cavity $15.0 \mu\text{m} \times 8.5 \mu\text{m}$. Upper bifid ends of mandibles vary slightly in length. Mandibular ring well developed. Mandibular process large. Onchia absent. Pore of renette situated three cephalic diameters from anterior end. Length of anterior female genital tube 669 to $772 \mu\text{m}$, and of reflexed part 381 to $535 \mu\text{m}$.* In one female four eggs, 112 to $125 \mu\text{m} \times 62 \mu\text{m}$, observed in posterior branch of uterus. Vulva situated slightly posterior to midbody. Spicules 87 to $100 \mu\text{m}$ long, with capitulum flattened on ventral side, and an extremely short neck, which broadens into slightly curved spicular body. Length of gubernaculum $25 \mu\text{m}$.

The most characteristic feature of this species is the absence of onchia. Other distinguishing features: cephalic setae, mandibular ends unequal in size, shape of spicules.

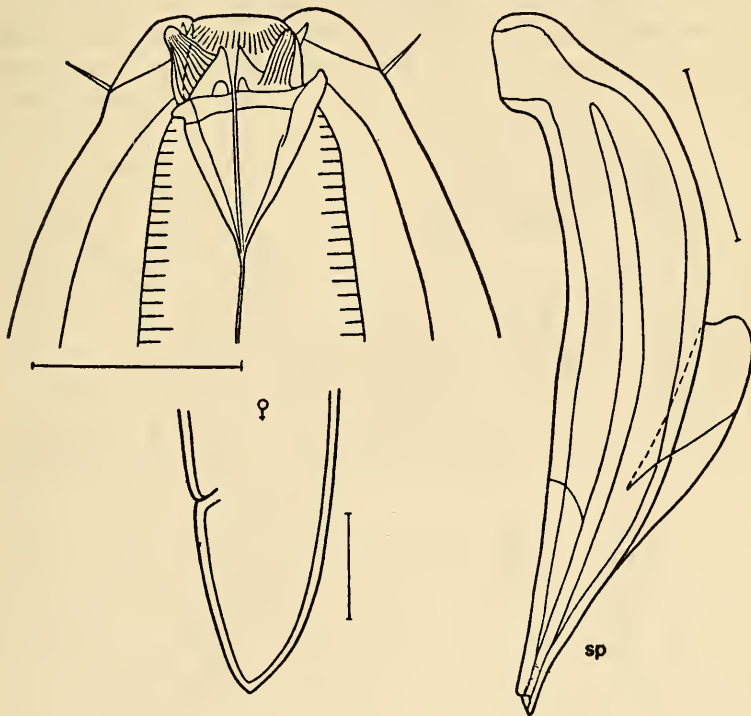


Figure 80. *Rhabdodemanía edentula*.

*Measurements of posterior tube not given in Russian original—General Editor.

Geographic distribution. Found in the Sea of Okhotsk (Bay of Nagaev) at a depth of 28 m in silted sand.

68. *Rhabdodemia latifaux* Platonova, 1974 (Figure 81)

Platonova, 1974: 1298, fig. 2.

$$1 \text{ ♂: } \frac{16 \text{ — } 566 \text{ — } 3,862}{25 \text{ — } 122 \text{ } 127 \text{ } 92} 4,048 \text{ } \mu\text{m}; a=32; b=7; c=21.$$

$$2 \text{ juv.: } \frac{15 \text{ — } 339\text{--}350 \text{ — } 1,569\text{--}1,935}{15 \text{ — } 70\text{--}72 \text{ } 75\text{--}80 \text{ } 50} 1,703\text{--}2,040 \text{ } \mu\text{m};$$

$$a=22\text{--}24; b=5\text{--}6; c=12\text{--}19.$$

Body tapers to 1/5 midbody diameter at anterior end and 10/13 at 154 posterior end. Tail tapers slightly and acicular at very tip. Cuticle 4.6 μm thick. Cephalic end blends with body. Length of cephalic setae 4.6 μm (1/3 corresponding head diameter). In addition to cephalic setae, a few short cervical setae present and a pair of extremely small setae located near excretory pore of caudal gland. Lips well developed. Size of oral cavity 16 μm \times 14 μm . Bifid ends of mandible equal. Mandibular ring poorly developed. Mandibular processes very small. Three small onchia situated in onchial part. Pore of renette situated two cephalic diameters from anterior end. There were no females of this species in my

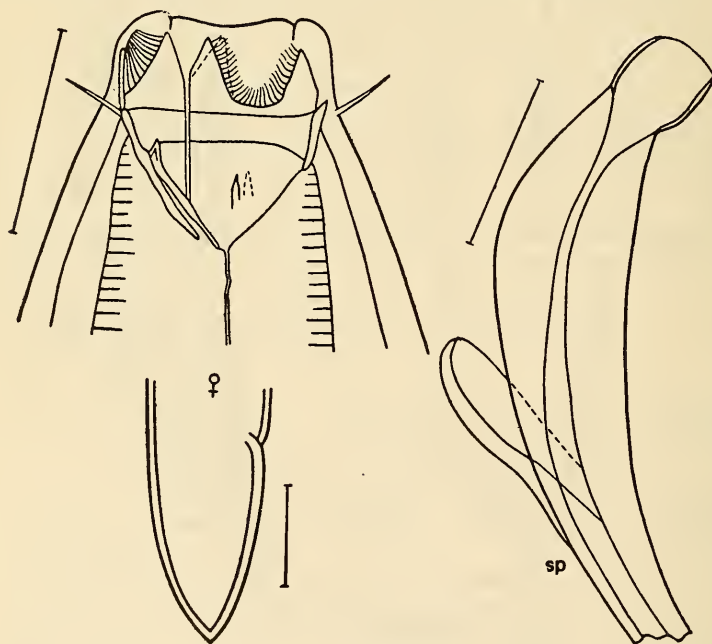


Figure 81. *Rhabdodemia latifaux*.

material. Spicules with narrow capitulum and 75 μm long. Gubernaculum trough-shaped and 17.5 μm long.

Geographic distribution. Found in the Barents Sea (Bay of Provideniya) at a depth of 4.0 m in rhizoids of *Laminaria* sp.

69. *Rhabdodemanina hexonchia* Platonova, 1974 (Figure 82)
Platonova, 1974: 1298, fig. 3.

$$4 \text{ } \sigma: \frac{8}{15} \quad \frac{160-162}{58} \quad \frac{350-412}{61-70} \quad \frac{\text{---}}{61-85} \quad \frac{2,300-2,781}{59-65} \quad 2,420-2,931 \text{ } \mu\text{m};$$

a = 34-40; b = 7; c = 16-19.

$$3 \text{ } \varnothing: \frac{8}{13-17} \quad \frac{170-174}{58} \quad \frac{380-391}{70} \quad \frac{900-1,050}{75-85} \quad \frac{1,670-1,822}{82-87}$$

× $\frac{2,400-2,697}{75-85} \quad \frac{2,862-2,906}{70-75} \quad 2,962-3,000 \text{ } \mu\text{m};$

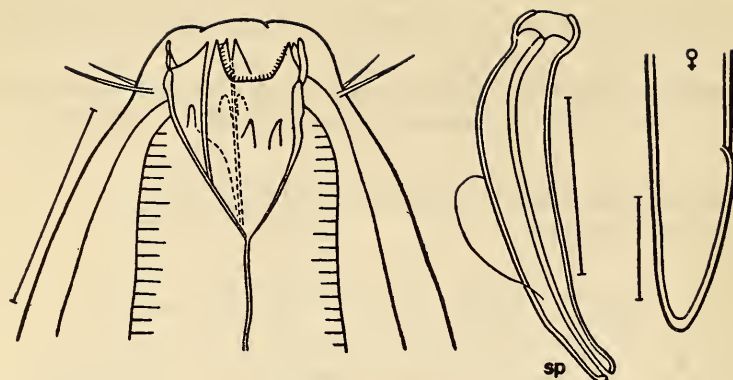
a = 34-36; b = 8; c = 21-29; V = 56%.

$$1 \text{ juv.}: \frac{7}{12} \quad \frac{174}{49} \quad \frac{360}{57} \quad \frac{\text{---}}{57} \quad \frac{1,802}{45} \quad 1,927 \text{ } \mu\text{m}; \text{ a} = 34; \text{ b} = 5; \text{ c} = 15.$$

Body tapers to 1/4 to 1/5 midbody diameter at anterior end and not more than 10/13 at posterior end. Tail long, obtusely rounded, and uni-
155 form in width throughout most of its length. Cuticle 3.5 μm thick. Cephalic setae long, 9.8 to 12.7 μm (slightly shorter than corresponding head diameter). Small setae present near excretory pore of caudal gland. Cervical constriction absent. Lips poorly developed, with faint striations. Oral cavity small, 8.0 μm \times 7.0 μm . Anterior end of mandible with shallow suture. Mandibular ring poorly developed and mandibular process absent. Onchia situated high, approximately on border between onchial and mandibular part. Two onchia present on each mandible, yielding a total of six. Excretory pore situated somewhat more than one cephalic diameter from anterior end. Length of anterior female genital tube 772 μm , of posterior tube 721 to 875 μm , and of reflexed parts 421 to 515 μm and 412 μm respectively. One egg, 200 μm \times 62 to 75 μm , present in each uterus. Vulva situated slightly posterior to midbody. Spicules weakly curved, with well-developed, round, and somewhat anteriorly flattened capitulum; spicular body narrows gradually toward distal end. Spicular length 50 μm . Gubernaculum lobate and 25 μm long.

Distinguishing features of these species: 1) long cephalic setae; 2) small head with wide oral cavity; 3) poorly developed mandibular ring and absence of mandibular process; 4) presence of six onchia instead of usual three; and 5) distinct shape of spicules.

Geographic distribution. Found in the Sea of Okhotsk near the western coast of Kamchatka at a depth of 22 m on a sandy bed.

Figure 82. *Rhabdodemanía hexonchia*.

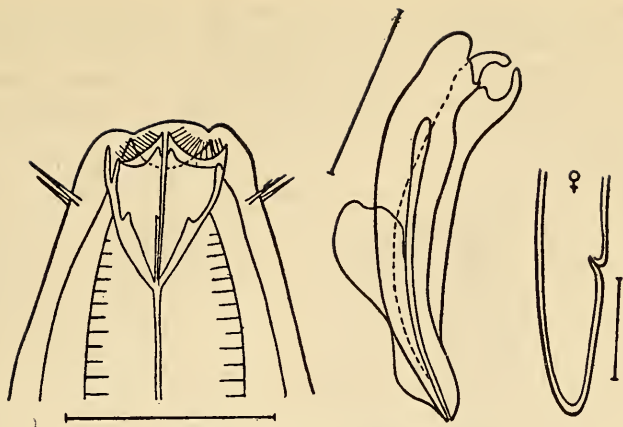
70. *Rhabdodemanía pontica* Platonova, 1965 (Figure 83)
Platonova, 1965; 100-102, fig. 1.

| | | | | | | |
|------|----------|--------------------------------|------------------|------------|----------------------|-----------------------------|
| 6 ♂: | 10 15 | 160-200 60-62 | 350-402 76-84 | — 78-85 | 1,900-2,200 50-60 | 2,000-2,352 μm ; |
| | | a = 24-26; b = 5-6; c = 14-16. | | | | |

| | | | | | | |
|------|----------|--|------------------|----------------------|-----------------------------|--|
| 4 ♀: | 10 16 | 200-209 56-58 | 460-470 76-86 | 927-990 90-94 | 1,339-1460 86-89 | |
| | | $\times \frac{1,808-1,920}{90-92}$ | | 2,220-2,500 55-62 | 2,382-2,656 μm ; | |
| | | a = 25-28; b = 5-6; c = 14-17 V = 56%. | | | | |

| | | | | | | |
|---------|---------|------------------|------------------|------------|----------------------|---|
| 8 juv.: | 8 11 | 150-174 42-48 | 309-360 45-57 | — 60-62 | 1,390-1,545 40-42 | $\times 1,527-1,657 \mu\text{m}$; a = 25-27; b = 4-5; c = 11-14. |
|---------|---------|------------------|------------------|------------|----------------------|---|

Body tapers to 1/5 to 5/27 midbody diameter at anterior end and 10/13 to 2/3 at posterior end. Tail long, bluntly rounded. Cuticle 2.3 to 156 3.5 μm thick. Length of cephalic setae 5.8 μm (1/3 corresponding head diameter). Except for cephalic, no other setae found on body. Head almost not demarcated from body. Lips poorly developed. Size of oral cavity 10.0 \times 5.0 μm . Mandibles with shallow suture anteriorly. Mandibular ring well developed but mandibular process absent. Onchia do not project much into oral cavity but rather long, occupying almost all of onchial cavity, and resemble highly sclerotized rays. Pore of renette situated two cephalic diameters from anterior end. Length of anterior female genital tube 412 to 469 μm , of posterior tube 463 to 469 μm , and of reflexed parts 257 to 309 μm and 310 μm respectively. Eggs, 206 $\mu\text{m} \times 72 \mu\text{m}$, observed in posterior uterus in just one female. Vulva situated somewhat posterior to midbody. Spicules 50 μm long, slightly curved, with rounded capitulum. Gubernaculum lobate and 17.5 μm long.

Figure 83. *Rhabdodemanía pontica*.

R. pontica differs from other species of the genus in: 1) small size; 2) absence of mandibular processes; 3) peculiarity of onchial structure; and 4) shape of spicules.

Geographic distribution. Found in the Black Sea in depths ranging from 3 to 120 m in silted sand and phaseolin silt.

71. *Rhabdodemanía marisalbi* Platonova, 1974 (Figure 84)
Platonova, 1974: 1299, fig. 5.

| | | | | | | | | |
|------|------------------------|---|---------|---------|-------------|-----------------|-------|-----------------|
| 2 ♂: | 20 | — | 619-700 | — | 4,368-4,775 | 4,550-4,997 μm; | | |
| | 29 | — | 105-121 | 121-129 | 81-97 | | | |
| | a=35-41; b=7; c=23-25. | | | | | | | |
| 1 ♀: | 20 | — | 503 | 2,007 | 2,559 | 4,602 | 5,705 | 5,907 μm; a=39; |
| | 30 | — | 130 | 140 | 150 | 130 | 101 | |
| | b=10; c=29; V=43%. | | | | | | | |

Large nematodes. Body tapers to 1/6 to 1/7 midbody diameter at anterior end and slightly more than 2/3 at posterior end. Cervical constriction poorly expressed. Tail acticular at tip. Cuticle 4.0 to 6.0 μm thick. Length of cephalic setae 6.0 μm (1/3 corresponding head diameter). Lips poorly developed. Size of oral cavity 20 μm × 9.0 μm. Anterior end of mandible with deep notch. Mandibular ring wide and highly sclerotized. Mandibular processes well developed. Onchia massive. Excretory pore situated two cephalic diameters from anterior end. Length of anterior female genital tube 552 μm, of posterior tube 1,043 μm.* Vulva displaced anterior to midbody. Spicular body narrows gradually from proximal

*Reflexed parts not mentioned in Russian original—General Editor.

157 to distal end; small velum on distal part extends to midlength of body; capitulum demarcated from body by distinct constriction and flattened on ventral side. Spicular length $81 \mu\text{m}$. Gubernaculum dilates gradually from distal to proximal end and terminates in shape of rounded hook; length $40 \mu\text{m}$.

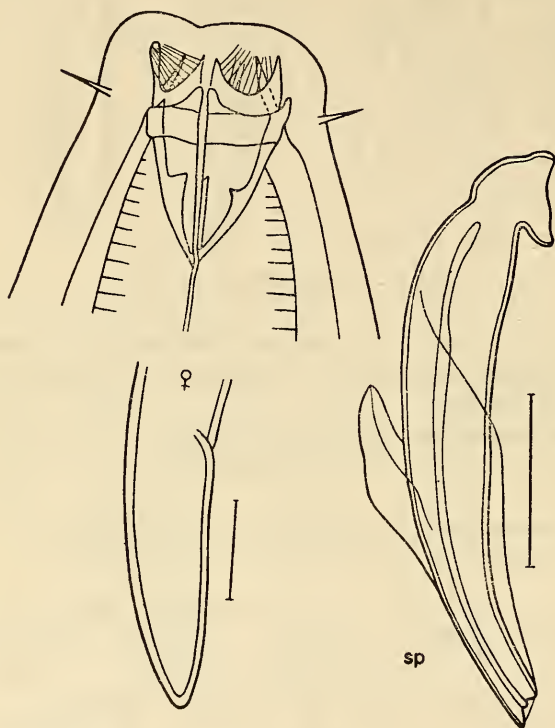


Figure 84. *Rhabdodemanía marisalbi*.

Distinguishing features of this species: 1) large size; 2) highly stretched oral cavity (length twice greater than width); 3) wide, well-developed mandibular ring and well-developed mandibular processes; and 4) shape of spicules.

Geographic distribution. Found in Chupin Inlet of the White Sea at a depth of 4.0 to 8.0 m in silted sand containing pebbles.

72. *Rhabdodemanía orientalis* Platonova, 1974 (Figure 85)
Platonova, 1974: 1300, fig. 6.

1 ♂: $\frac{16 \quad 263 \quad 627 \quad - \quad 3,887}{26 \quad 92 \quad 110 \quad 141 \quad 129} 4,089 \mu\text{m}; a=29; b=7; c=20.$

3 juv.: $\frac{9}{20} \quad \frac{109}{39} \quad \frac{278-407}{40-48} \quad \frac{—}{32-52} \quad \frac{1,134-2,363}{28-44} \quad 1,195-2,496 \mu\text{m};$
 $a=35-48; b=5-6; c=18-27.$

Rather large nematodes. Body tapers to 1/10 midbody diameter at anterior end and barely tapers toward posterior end. Tail blunt at tip. Cuticle $4.5 \mu\text{m}$ thick. Cephalic setae $9.0 \mu\text{m}$ long (1/2 corresponding head diameter). Lips well developed. Oral cavity $16.0 \mu\text{m} \times 9.0 \mu\text{m}$. Mandible with deep notch. Mandibular ring highly sclerotized, but rather narrow. Mandibular processes not large. Onchia well developed. Renette not readily discernible in my specimens; probably, its pore opens not far away from oral cavity. Females absent in my material. Spicules wide, narrow smoothly toward both ends, but without distinctly expressed capitulum. Spicular length $64.8 \mu\text{m}$. Gubernaculum small, funnel-shaped, and encloses distal end of spicules. Length of gubernaculum $25 \mu\text{m}$.

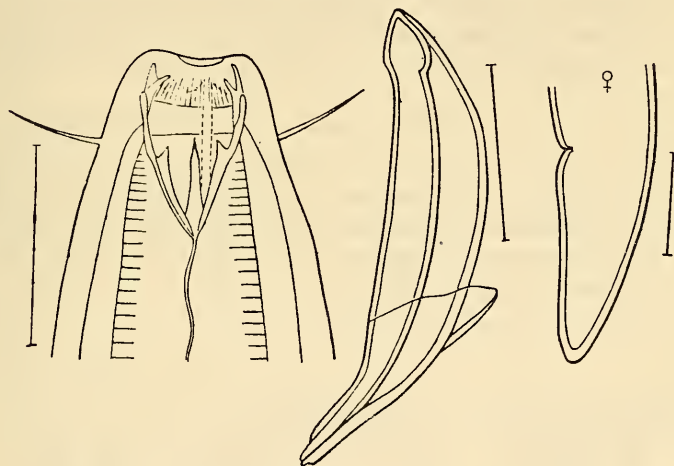


Figure 85. *Rhabdodemanía orientalis*.

R. orientalis resembles the White Sea species *R. marisalbi* in many ways but differs from it in: 1) longer cephalic setae; 2) much narrower mandibular ring; 3) wider body; and 4) shape of spicules.

Geographic distribution. Found in the Gulf of Pos'et (Sea of Japan) at a depth of 13 to 17 m in silted sand with shells.

73. *Rhabdodemanía angustissima* Platonova, 1974 (Figure 86)
 Platonova, 1974: 1300, fig. 7.

1 ♂: $\frac{11}{16} \quad \frac{182}{35} \quad \frac{405}{35} \quad \frac{—}{38} \quad \frac{3,257}{37} \quad 3,391 \mu\text{m}; a=89; b=8; c=25.$

| | | | | | | | | |
|---------|--------------|-----|-----|----|-------|--|-------|----------------------------------|
| 1 ♀: | 11 | 162 | 405 | — | 2,116 | — | 3,106 | 3,216 μm ; a=80; b=8; |
| | 24 | 20 | 24 | — | 40 | — | 32 | |
| | c=29; V=65%. | | | | | | | |
| 1 juv.: | 8 | 121 | 271 | — | 1,202 | 1,302 μm ; a=59; b=5; c=13. | | |
| | 12 | 19 | 21 | 22 | 20 | | | |

Body very narrow; tapers to 1/4 to 1/5 midbody diameter at anterior end and barely tapers toward anus. Tail wide, tip bluntly rounded. Head distinctly demarcated from body. Cuticle 5.0 μm thick. Lips well developed. Cephalic setae very long, 24 μm (length twice corresponding head diameter). Oral cavity 11.0 μm \times 7.0 μm . Anterior mandibular end with small notch. Mandibular ring poorly sclerotized. Mandibular processes not developed. Onchia narrow, long, and look like longitudinal rays. Pore of renette situated three cephalic diameters from anterior end of body. Genital tubes still undeveloped in specimen examined. Vulva shifted posteriorly to midbody. Spicules short and wide; proximal end wide and resembles capitulum. Spicular length 35.6 μm . Gubernaculum characteristic, extremely thin, narrow, and falcate at proximal end; length 10 μm .

Distinguishing features of this species: 1) extremely narrow body; 2) very long cephalic setae; 3) highly elongated oral cavity, with poorly developed mandibular ring, devoid of mandibular processes, and with long onchia that look like longitudinal rays; and 4) shape of spicules.

Geographic distribution. Found in the Sea of Okhotsk (Gulf of Aniva) in the littoral zone in a sandy bed.

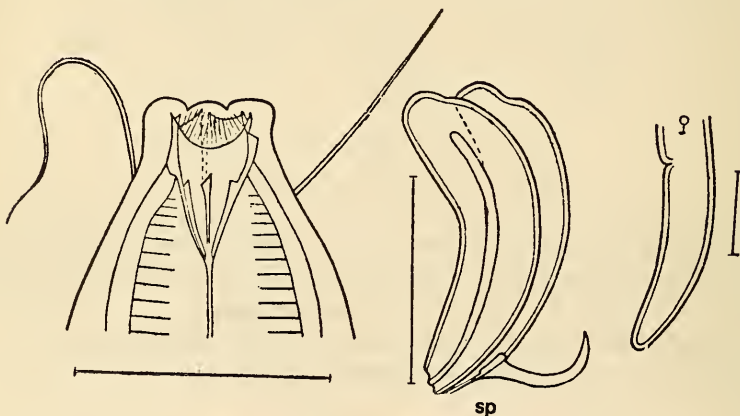


Figure 86. *Rhabdodemia angustissima*.

References

- 159– Allgen, C.A. 1927a. Über einige freilebende marine Nematoden von der
164 schwedischen Küste des Öresundes, *Zool. Anz.*, vol. 73, pp. 49–58.
- Allgen, C.A. 1927b. Freilebende marine Nematoden von den Campbell-
und Staten Inseln, *Nyt Mag. Naturvid.*, vol. 66, pp. 249–309.
- Allgen, C.A. 1928. Neue oder wenig bekannte freilebende marine
Nematoden von der schwedischen Westküste, *Zool. Anz.*, vol. 77,
pp. 281–307.
- Allgen, C.A. 1929a. Freilebende marine Nematoden aus der Umgebung
der staatlichen zoologischen Station Kristinenberg an der Westküste
Schwedens, *Capita Zool.*, 2, 8, 1–55.
- Allgen, C.A. 1929b. Über einige antarktische freilebende marine Nema-
toden, *Zool. Anz.*, vol. 84, pp. 126–140.
- Allgen, C.A. 1930. Freilebende marine Nematoden von der Stateninsel
(Feuerland-Archipel), I, *Zool. Anz.*, vol. 89, pp. 246–258.
- Allgen, C.A. 1931a. Weitere Beiträge zur Kenntnis des marinen Nema-
todenfauna der Campbell Insel, *Nyt. Mag., Naturvid*, vol. 70, pp.
97–198.
- Allgen, C.A. 1931b. Freilebende Nematoden aus dem Dröbakab-
schnitt des Oslo-fjords, *Zool. Jahrb. (Abt. Syst.)* vol. 61, pp. 211–262.
- Allgen, C.A. 1932. Über einige freilebende marine Nematoden aus der
Umgebung der biologischen Station auf der Insel Herdia (Nor-
wegen), *Arch. Naturgesch. (Neue Folge)*, vol. 26, pp. 399–432.
- Allgen, C.A. 1933. Freilebende Nematoden aus dem Trondheimsfjord,
Capita Zool., 4, 2, 1–162.
- Allgen, C.A. 1934a. Freilebende marine Nematoden aus Hallands
Värderö und der nahgelegenen Küste Schonens (Südschweden), *Folia
Zool. et Hydrobiol.*, 6, 3, 49–86. Riga.
- Allgen, C.A. 1934b. Bipolarität in der Verbreitung freilebender mariner
Nematoden, *Zool. Anz.*, vol. 105, pp. 331–334.
- Allgen, C.A. 1935. Die freilebende Nematoden des Öresunds, *Capita
Zool.*, 6, 3, 1–195.
- Allgen, C.A. 1936. Über einige freilebende marine Nematoden aus der
Strandfauna Norwegens, *Nyt Mag. Naturvid.*, vol. 76, pp. 245–272.
- Allgen, C.A. 1939. Über einige im reinen Schalensand der Westküste
Norwegens freilebende Nematoden, *Festschr. für E. Strand* vol. 5,
pp. 404–425.
- Allgen, C.A. 1940a. Über einige neue freilebende Nematoden von der
Nordwest- und Nordküste Norwegens, *Folia Zool. et Hydrobiol.*, 10,
1, 258–281. Riga.
- Allgen, C.A. 1940b. Weitere freilebende Nematoden insbesondere aus

- dem Schalensand- und Kiesboden der Strandzone Norwegens, *Folia Zool. et Hydrobiol.*, 10, 2, 487–508. Riga.
- Allgen, C.A. 1942. Die freilebenden Nematoden des Mittelmeers, *Zool. Jahrb. (Abt. Syst.)*, 76, 1–2, 1–102.
- Allgen, C.A. 1943. Norwegische marine Nematoden, *Zool. Jahrb. (Abt. Syst.)*, 76, 4, 267–322.
- Allgen, C.A. 1947a. Über einige Tiefsee—Nematoden aus dem Lopphavet im nördlichen Norwegen, *Det Kongelige Norske Videnskab. Selskab. Forhandl.*, vol. 19, pp. 7–11.
- Allgen, C.A. 1947b. On some free-living marine nematodes from Tobago, *Vid. Medd. Dansk. Naturh. Foren.*, vol. 110, pp. 45–65.
- Allgen, C.A. 1947c. West American marine nematodes, *Vid. Medd. Dansk. Naturh. Foren.*, vol. 110, pp. 65–219.
- Allgen, C.A. 1951. Pacific free-living marine nematodes, *Vid. Medd. Dansk. Naturh. Foren.*, vol. 113, pp. 263–411.
- Allgen, C.A. 1954a. Free-living marine nematodes from East Greenland and Jan Mayen, *Medd. om Groenland*, 107, 6, 5–44
- Allgen, C.A. 1954b. Das Bipolaritäts problem freilebender mariner Nematoden, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1953), 26, 8, 28–35.
- Allgen, C.A. 1956a. Vergleich zwischen den marinen Nematodenfaunen Norwegens und denen antarktischen Gebietes, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1955), 28, 15, 71–76.
- Allgen, C.A. 1956b. Vergleich zwischen den marinen Nematodenfaunen Norwegens und denen der Atlantikküste Europas, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1955), 28, 16, 77–83.
- Allgen, C.A. 1956c. Vergleich zwischen den marinen Nematodenfaunen Norwegens und denen des Mittelmeers sowie angrenzender nordöstlicher Meeresabschnitt, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1955), 28, 18, 84–95.
- Allgen, C.A. 1957a. Vergleich zwischen den marinen Nematodenfaunen Norwegens und der Tropen, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1956), 29, 9–10, 36–46.
- Allgen, C.A. 1957b. On a small collection of free-living marine nematodes from Greenland and some other arctic regions, *Medd. om Groenland*, 159, 3, 1–42.
- Allgen, C.A. 1958a. Zur Kenntnis norwegischen Nematoden, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1957), 30, 4, 22–27.
- Allgen, C.A. 1958b. Vergleich zwischen den marinen Nematoden Norwegens und denen der subtropischen Faunenengebiete in der südlichen Hemisphäre, *Det Kongelige Norske Videnskab. Selskab. Forhandl.* (1957), 30, 15, 93–98.
- Allgen, C.A. 1958c. Über einige freilebende marine Nematoden von

- der Ostküste Südamerikas, *Zool. Anz.*, vol. 160, pp. 206–217.
- Allgen, C.A. 1959. Free-living marine nematodes, IV, *Zool. Res. Swed. Antarct. Exp. 1901–1903*, 5, 2, 293 pp.
- Bastian, H.Ch. 1865. Monograph on Anguillulidae or free-living nematodes (marine, land, and fresh water), *Trans. Linn. Soc.*, vol. 25, pp. 73–184. London.
- Baylis, H.A. 1916. Some nemertines, free-living nematodes, and oligochaetes from the Falklands, *Ann. Mag. Nat. Hist.*, ser. 8, vol. 17, pp. 288–299.
- Baylis, H.A. and R. Daubney. 1926. *A Synopsis of Families and Genera of Nematodes*. Brit. Mus. (Nat. Hist.), London, 277 pp.
- Berg, L.S. 1920. Bipolyarnoe rasprostranenie organizmov i lednikovaya epokha (Bipolar distribution of organisms and the glacial epoch). *Izv. AN SSSR*, vol. 14, pp. 273–302.
- Bresslau, E. and J.H. Schuurmans-Stekhoven. 1940. Marine freilebende Nematoda aus der Nordsee, *Ouvrage édité par le Patrimoine du Mus. Roy. Hist. Nat. Belg.* Bruxelles, 74 pp.
- Bütschli, O. 1873. Beiträge zur Kenntnis der freilebenden Nematoden, *Nova Acta Leop.-Carol. Akad. Naturf.*, 36, 5, 144 pp.
- Bütschli, O. 1874. Zur Kenntnis der freilebenden Nematoden, insbesondere der des Kieler Hafens, *Abh. Senkenberg. Naturf. Ges. Frankf.-a.-Mein*, vol. 9, pp. 237–368.
- Chitwood, B.G. 1936a. Some marine nematodes of superfamily Enoploidea, *Trans. Amer. Micr. Soc.*, 55, 2, 208–213.
- Chitwood, B.G. 1936b. Some marine nematodes from North Carolina, *Proc. Helminth. Soc.* vol. 3, pp. 1–16. Washington.
- Chitwood, B.G. 1937. A new genus and ten new species of marine nematodes from North Carolina, *Proc. Helminth. Soc.*, vol. 4. pp. 54–59. Washington.
- Chitwood, B.G. 1951. North American marine nematodes, part 4, *Texas J. Sci.*, vol. 3, pp. 627–672.
- Chitwood, B.G. 1960. A Preliminary contribution to marine nemas (Adenophorea) of northern California, *Trans. Amer. Micr. Soc.*, 79, 4, 347–384.
- Chun, C. 1897. Die Beziehung zwischen dem arktischen und antarktischen Plankton. E. Nägele, Stuttgart, 64 pp.
- Cobb, N.A. 1891. *Anticoma*, a genus of free-living marine nematodes, *Proc. Linn. Soc.*, 5, 3, 765–774. N.S. Wales.
- Cobb, N.A. 1893a. *Tricoma* and other new Nematoda genera, *Proc. Linn. Soc.*, 8, 2, 389–420. N.S. Wales.
- Cobb, N.A. 1893b. Nematodes, mostly Australian and Fijian, *Macleay Mém. Vol., Linn. Soc. Misc. Publ.*, no. 13, pp. 3–59. N.S. Wales.

- Cobb, N.A. 1898. Australian free-living marine nematodes, *Proc. Linn. Soc.*, 23, 2, 383–407. N.S. Wales.
- Cobb, N.A. 1914. Antarctic marine free-living nematodes of the Shackleton Expedition, part 1, *Contrib. Sci. Nematology*, pp. 1–33
- Cobb, N.A. 1933. New nemic genera and species, with taxonomic notes, *J. Parasitol.*, 20, 2, 81–94.
- Deryugin, K.M. 1915. Fauna Kol'skogo zaliva i usloviya ee sushchestvovaniya (Fauna of Kola Bay and its living conditions). *Zap. Akad. Nauk, Fiz.-Mat. Otd.*, 34, 1, 928. St. Petersburg.
- Ditlevsen, H. 1919. Marine free-living nematodes from Danish waters, *Vid. Medd. Dansk. Naturh. Foren.* vol. 70, pp. 147–214.
- Ditlevsen, H. 1922. Marine free-living nematodes from Auckland and Campbell Islands, *Vid. Medd. Dansk. Naturh. Foren.*, vol. 73, pp. 1–32.
- Ditlevsen, H. 1923. Sur quelques Nématodes libres (Côtes de Brétagne et Rockal), *Bull. Soc. Zool.*, vol. 48, pp. 178–203. France.
- Ditlevsen, H. 1926. Free-living nematodes, part 6. In *The Ingolf Expeditions*, vol. 4, pp. 1–42.
- Ditlevsen, H. 1930. Marine free-living nematodes from New Zealand, *Vid. Medd. Dansk. Naturh. Foren.*, vol. 87, pp. 201–242.
- Dogel', V.A. 1954. Oligomerizatsiya gomologichnykh organov (Oligomerization of Homologous Organs). Izd. LGU, 367 pp.
- Eberth, C.J. 1863. *Untersuchungen über Nematoden*. W. Engelmann, Leipzig, 77 pp.
- Ekmann, S. 1935. *Tiergeographie des Meeres*. Akad. Verlagsgesellschaft, Leipzig, 542 pp.
- Ekmann, S. 1953. *Zoogeography of the Sea*. Sidwick and Jackson, London, 417 pp.
- Filip'ev, I.N. 1912. K voprosu o stroenii nervnoi sistemy u svobodnozhivushchikh nematod (Structure of the nervous system in free-living nematodes). *Tr. O-va Estestvoispyt*, vol. 43, pp. 205–215. St. Petersburg.
- Filip'ev, I.N. 1916. Svobodnozhivushchie nematody kolleksii Zoologicheskogo muzeya imperatorskoi Akademii Nauk (Free-living nematodes in the collection of the Zoological Museum, Imperial Academy of Sciences). *Ezheg. Zool. Muz. Imp. AN*, vol. 21, pp. 59–116. [English translation published by Amerind Pub. Co. Pvt. Ltd., New Delhi, India, 1973.]
- Filip'ev, I.N. 1918–1921. Svobodnozhivushchie morskije nematody okrestnostei Sevastopolya (Free-living marine nematodes of Sevastopol' environs). *Tr. Osoboi Zool. Lab. i Sevastopol'skoi Biol. St. Ross. AN*, ser. 2, no. 4, vol. 1/2, 614 pp.
- Filip'ev, I.N. 1922a. Novye dannye o svobodnykh nematodakh Chernogo

- morya (New data on free-living nematodes of the Black Sea). *Tr. Stavropol'skogo S.-Kh. Inst.*, 1, 16, 184. pp.
- Filip'ev, I.N. 1922b. Instruksiya dlya sobiraniya svobodnykh nematod (Instructions for the Collection of Free-living Nematodes). Izd. Petrogr. Agronom. Inst. Petrograd, 14 pp.
- Filip'ev, I.N. 1922c. O svobodykh nematodakh Azovskogo morya (Free-living nematodes of the Sea of Azov). *Tr. Stavropol'skogo S.-Kh.*, 1, 17, 185–208.
- Filip'ev, I.N. 1934b. Nematody vrednyye i poleznye v sel'skom khozyaystve (Useful and Harmful Nematodes in Agriculture). Sel'khozgiz, Moscow-Leningrad, 440 pp.
- Filip'ev, I.N. 1937. Klass kruglykh chervei (Nematoda). Rukovodstvo po zoologii. Bespozvonochnye [Class Nematoda (Roundworms). A Guide to Zoology. Invertebrates]. Biomedgiz, Moscow-Leningrad, vol. 1, pp. 556–628.
- Filip'ev, I.N. 1946. Svobodnozhivushchie nematody iz Severnogo Ledovitogo Okeana (Free-living nematodes from the Arctic Ocean). *Tr. Dreif. Eksp. Glavsevmorputi na Led. "G. Sedov" 1937–1940*, vol. 3, pp. 158–184.
- Filip'ev, I.N. [Filip'ev, I.N.]. 1927. Les nématodes libres des mers septentrionales appartenant à la famille des Enoplidae, *Arch. Naturgesch.*, 91, A, 4 (1925), 216.
- Filipjev, I.N. [Filip'ev, I.N.]. 1934a. Classification of free-living nematodes and their relation to parasitic nematodes, *Smiths. Misc. Coll.*, 89, 6, 1–63.
- Gerlach, S.A. 1953. Biozönotische Gliederung der Nematodenbesiedlung an den deutschen Küsten, *Ztschr. Morphol Ökol. Tiere*, 41, 5–6, 411–512.
- Gerlach, S.A. 1955. Zur Kenntnis der freilebenden marinen Nematoden von San Salvador, *Ztschr. Wiss. Zool.*, 158, 2–4, 249–303.
- Gerlach, S.A. 1956. Diagnosen neuer Nematoden aus der Kieler Bucht, *Kieler Meeresforsch.*, vol. 12, pp. 85–109.
- Gerlach, S.A. 1958. Die Nematodenfauna der sublitoralen Region in der Kieler Bucht, *Kieler Meeresforsch.*, vol. 14, pp. 64–92.
- Gerlach, S.A. 1959. Neue Meeres-Nematoden aus dem Supralitoral der deutschen Küsten, *Intern. Rev. Ges. Hydrobiol.*, 44, 3, 463–467.
- Gerlach, S.A. and F. Riemann. 1974. The Bremerhaven checklist of aquatic nematodes, *Veröff. Inst. Meeresforsch. Bremerh.*, Suppl. 4, pp. 405–734.
- Goldschmidt, R. 1904. Über die Cuticula von *Ascaris*. *Zool. Anz.*, vol. 28, pp. 259–266.
- Golovin, E.P. 1901, 1902. Nablyudeniya nad nematodami (Observations on Nematodes). *Tipolitografiya Imp. Univ., Kazan*, vol. 1, 149 pp.; vol. 2, 120 pp.

- Goodey, D.B. 1959. Laboratornye metody issledovaniya rastitel'nykh i pochvennykh nematod (Laboratory Methods of Studying Plant and Soil Nematodes). IL, Moscow, 85 pp.
- Hyman, L.H. 1951. *The Invertebrates*. Vol. 3: *Acantocephala, Aschelminthes and Entoprocta. The Pseudocoelomate Bilateria*. McGraw-Hill, New York, 572 pp.
- Inglis, W.G. 1958. A new species of the nematode genus *Thoracostoma* from the Antarctic, *Ann. Mag. Nat. Hist.*, ser. 13, 1, 1, 45-48.
- Inglis, W.G. 1964. The marine Enoplida (Nematoda): a comparative study of the head, *Bull. Brit. Mus. (Nat. Hist.)*, 11, 4, 265-376.
- Inglis, W.G. 1971. Marine Enoplida (Nematoda) from western Australia, *Trans. Roy. Soc.*, 95, 2, 65-78. S. Austr.
- Jägerskiöld, L.A. 1901. Weitere Beiträge zur Kenntnis der Nematoden, *Kongeliga Svenska Vet. Akad. Handl.*, 35, 2, 1-80.
- Kreis, H.A. 1928. Die freilebenden marinen Nematoden der Spitzbergen expedition von F. Römer und F. Schaudinn im Jahre 1898, *Mitt. Zool. Mus.*, vol. 14, pp. 131-197. Berlin.
- Kreis, H.A. 1934. Oncholaiminae Filipjev, 1916. Eine monographische Studie, *Capita Zool.*, 4, 5, 1-271.
- Leydig, F. 1854. Zoologisches, *Arch. Anat. Phys. Jahrg. 1854*, pp. 284-295.
- Leucart, R. 1849. Zur Kenntnis der Fauna von Island. I. Beitrag (Würmer), *Arch. Naturg.*, 15, 1, 149-208.
- Linstow, O. 1892. Helminthen aus Südgeorgien, *Jahrb. Hamburg. Wiss. Anstalten*, 9, 2, 59-77.
- Linstow, O. 1896. Nemathelminthen, *Ergebnisse Hamburg. Magalhaensische Sammelreise*, 3, 8, 3-21.
- Linstow, O. 1900. Die Nematoden, *Fauna Arctica*, vol. 1, pp. 117-132. Jena.
- Linstow, O. 1903. Entozoa des zoologischen Museums der keiserlichen Akademie der Wissenschaften zu St. Petersburg, *Ann. Mus. Zool.*, vol. 8, pp. 265-294. St. Petersburg.
- Linstow, O. 1906. Nematoda. National Antarctic Expedition 1901-1904, *Nat. Hist.*, vol. 3, pp. 1-4.
- Loos, C.A. 1896. Über den Bau der Ösophagus bei einiger Ascariden, *Zentralb. Bakt. Parasitenk.*, vol. 19, pp. 5-13.
- Luc, M. and L. de Coninck. 1959. Nématodes libres marins de la region Roscoff, *Arch. Zool. Expérim. Génér.*, vol. 98, fasc. 2, pp. 103-165.
- Man, J.G. de. 1876. Contribution à la connaissance des Nématodes marins du golf de Naples, *Tijdscher. Nederl. Dierk. Vereeniging*, vol. 2, pp. 78-196.
- Man, J.G. de. 1878. Contribution à la connaissance des Nématodes

- marins du golf de Naples, *Tijdschr. Nederl. Dierk. Vereeniging*, vol. 3, pp. 88–118.
- Man, J.G. de. 1886. *Anatomische Untersuchungen über freilebende Nordsee-Nematoden*. Paul Frohber, Leipzig, 82 pp.
- Man, J.G. de. 1888. Sur quelques Nématodes libres de la mer du Nord, nouveaux ou peu connus, *Mém. Soc. Zool.*, vol. 1, pp. 1–51. France.
- Man, J.G. de. 1889. Troisième note sur les Nématodes libres de la mer du Nord et de la Manche, *Mém. Soc. Zool.*, vol. 2, pp. 182–216. France.
- Man, J.G. de. 1890. Quatrième note sur les Nématodes libres de la mer du Nord et de la Manche, *Mém. Soc. Zool.*, vol. 2, pp. 169–194. France.
- Man, J.G. de. 1893. Cinquième note sur les Nématodes libres de la mer du Nord et de la Manche, *Mém. Soc. Zool.*, vol. 4, pp. 81–125. France.
- Man, J.G. de. 1904. *Nematodes libres—Res. Voy. S.Y. Belgica (1897–1898–1890)—Rap. Sci.* J.E. Buschmann, Anvers, 55 pp.
- Man, J.G. de. 1907. Sur quelques espèces nouvelles ou peu connus de Nématodes libres, habitant les côtes de la Zélande, *Mém. Soc. Zool.*, vol. 20, pp. 33–90. France.
- Man, J.G. de. 1922. Neue freilebende Nematoden aus der Zuiderzee, *Tijdschr. Nederl. Dierk. Vereeniging*, 18, 2, 124–134.
- Marion, A. 1870. Recherches zoologiques et anatomiques sur des Nématodes non parasites marins, *Ann. Sci. Nat.*, nos. 13–14, 102 pp.
- Martini, E. 1903. Über Furchung und Gastrulation des *Cuculanus elegans*, *Ztschr. Wiss. Zool.*, vol. 74, pp. 501–556.
- Martini, E. 1906. Über Subcuticula und Seitenfelder einiger Nematoden, *Ztschr. Wiss. Zool.*, vol. 81, pp. 699–766.
- Martini, E. 1908a. Die Konstanz histologischen Elementen bei Nematoden, *Verh. Anat. Gesellsch.*, vol. 22, pp. 132–134. Jena.
- Martini, E. 1908b–1909. Über Subcuticula und Seitenfelder einiger Nematoden, *Ztschr. Wiss. Zool.*, vol. 91, pp. 191–235; vol. 93, pp. 535–624.
- Martini, E. 1916. Die Anatomie der *Oxyuris curvula*, *Ztschr. Wiss. Zool.*, vol. 116, pp. 137–525.
- Maupas, E. 1900. Modes et formes de reproduction chez les Nématodes, *Arch. Zool. Exp.*, vol. 8, fasc. 3, pp. 463–624.
- Mawson, P.M. 1953. Some marine free-living nematodes from the Australian coast, *Trans. Roy. Soc.*, vol. 76, pp. 34–40. South Australia.
- Mawson, P.M. 1956–1958. Free-living nematodes. Sect. 1. Enoploidea from Antarctic stations. Sect. 2. Additional Enoploidea from Antarctic stations. Sect. 3. Enoploidea from subantarctic stations, *Brit. Aust. New Zeal. Antarct. Research Exped. 1929–1931. Reports—Series B.*

- (*Zool. and Bot.*): 1956, vol. 6, pt. 3, pp. 39–74; 1958a, vol. 4, pt. 13, pp. 293–305; 1958b, vol. 4, pt. 14, pp. 310–358.
- Meyl, A.H. 1956–1957. Beiträge zur freilebenden Nematodenfauna Brasiliens, I, *Nematologica*, 1, 4, 311–325; II, *Kieler Meeresforsch.*, vol. 13, pp. 125–133.
- Micoletzky, H. 1914. Freilebende Süßwasser-Nematoden der Ost-Alpen mit besonderer Berücksichtigung des Lunzer-Seengebietes, *Zool. Jahrb. (Abt. Syst.)*, 36, 4–5, 331–546.
- Micoletzky, H. 1922. Neue freilebende Nematoden aus Suez, *Sitz.-ber. Acad. Wiss. Wien Math.-Naturw. Klasse, Abt. 1*, vol. 131, pp. 77–103.
- Micoletzky, H. 1924. Weitere Beiträge zur Kenntnis freilebender Nematoden aus Suez, *Sitz.-ber. Acad. Wiss. Wien Math.-Naturw. Klasse, Abt. 1*, vol. 132, pp. 225–262.
- Micoletzky, H. and H. Kreis. 1930. Freilebende marine Nematoden von der Sunda-Inseln. I. Enoploidea, *Vid. Medd. Dansk Natur. Foren.*, vol. 87, pp. 243–399.
- Mordukhai-Boltovskoi, F.D. 1960. Katalog fauny svobodnozhivushchikh bespozvonochnykh Azovskogo morya (Catalogue of free-living invertebrates of the Sea of Azov). *Zool. Zhurn.*, 39, 10, 1454–1466.
- Ortmann, A. 1896. Über "Bipolarität" in der Verbreitung mariner Tiere, *Zool. Jahrb. (Abt. Syst.)*, 9, 4, 571–595.
- Paladian, G. 1962. Contribution à l'étude des Nématodes libres du littoral roumain de la mer Noire, *Trav. Mus. d'Hist. Nat. "Gr. Antipa"*, vol. 3, pp. 69–74.
- Paramonov, A.A. 1926. O sluchae bivul'varnosti u odnoi svobodnoi nematody (Case of bivulvarity in a free-living nematode). *Russk. Gidrobiol. Zhurn.*, 5, 10–12, 218–222.
- Paramonov, A.A. 1962. Osnovy fitogel'mintologii (Fundamentals of Phytohelminthology). Izd. AN SSSR, Moscow, 479 pp.
- Platonova, T.A. 1958. K faune nematod semeistva Leptosomatidae iz raiona ostrova Kergelen (Nematodes of family Leptosomatidae from Kerguelen Islands). *Byull. Sov. Antarkt. Eksp.*, no. 3, pp. 59–61.
- Platonova, T.A. 1962a. Nekotorye materialy po ekologii svobodnozhivushchikh nematod chernogo morya (Some data on the ecology of free-living nematodes of the Black Sea). In *Trudy 5-go Vsesoyuznogo Soveshchaniya Fitogel'mintologov. Samarkandskogo. Izd. Samarkandskogo Univ.*, pp. 200–227.
- Platonova, T.A. 1962b. Novye vidy nematod roda *Pseudocella* Filipjev i Kuril'skikh ostrovov i yuzhnogo Sakhalina (New species of nematodes of genus *Pseudocella* Filipjev from Kuril Islands and southern Sakhalin). *Issledovaniya Dal'nevostochnykh Morei SSSR*, vol. 8, pp. 200–218.
- Platonova, T.A. 1965. Novyi vid svobodrozhivushchei nematody iz

- Chernogo morya (New species of free-living nematodes of the Black Sea). In *Bentos*. Izd. Naukova Dumka, Kiev, pp. 100–102.
- Platonova, T.A. 1967. Svobodnozhivushchie morskije nematody semeistva Leptosomatidae Evropeiskoi Arktiki (Free-living marine nematodes of family Leptosomatidae of the European Arctic). *Zool. Zhurn.*, 46, 6, 828–839.
- Platonova, T.A. 1970. O sistematike Leptosomatidae (Nematoda) Sredizemnogo morya i prilizhashchikh vod Atlantiki [Systematics of Leptosomatidae (Nematoda) of the Mediterranean Sea and adjoining waters of the Atlantic Ocean]. *Zool. Zhurn.*, 49, 9, 1298–1305.
- Platonova, T.A. 1974. Ob'em i sistematicheskoe polozhenie roda *Rhabdodemia* (Nematoda, Enoplida) [Composition and systematic position of genus *Rhabdodemia* (Nematoda, Enoplida)]. *Zool. Zhurn.*, 53, 9, 1295–1303.
- Rauther, M. 1907. Über den Bau des Ösophagus und die Lokalisation der Nierenfunktion bei freilebenden Nematoden, *Zool. Jahrb. (Abt. Anat.)*, vol. 23, pp. 703–740.
- Rauther, M. 1909. Morphologie und Verwandtschaftsbeziehung der Nematoden, *Erg. Fortschr. Zool.*, vol. 1, pp. 491–596.
- Rouville, E. 1903. Révision des Nématodes libres, marins de la région de Cette, *Compt. Rend. Acad. Sci.*, vol. 137, pp. 1002–1003. Paris.
- Rouville, E. 1904. Révision des Nématodes libres, marins de la région de Cette, *Compt. Rend. Assoc. Franç. Avanc. Sci.*, vol. 33, pp. 788–797.
- Saveljev, S. [Savel'ev]. Zur Kenntnis der freilebenden Nematoden des Kolafjords und des Reliktensee Mogilnoje, *Tr. O-va Estestvoispyt*, vol. 43, pp. 108–126. St. Petersburg.
- Schimkewitsch, W.M. [Shimkevich, V.M.]. 1889. Über besondere Zellen in der Lebenshöhle der Nematoden, *Biol. Zentralbl.*, no. 19, p. 407.
- Schneider, A. 1866. *Monographie der Nematoden*. G. Reimer, Berlin, 375 pp.
- Schenider, G. 1906. Beiträge zur Kenntnis der im Uferschlamm des Finnischen Meerbusens freilebenden Nematoden, *Acta Soc. Fauna et Flora Fennica*, 27, 7, 1–42.
- Schneider, K. 1902. *Lehrbuch der vergleichenden Histologie der Tiere*. G. Fischer, Jena, 691 pp.
- Schneider, W. 1939. Fadenwürmer oder Nematoda, *Tierwelt Deutschlands*, vol. 36, 260 pp.
- Schulz, E. 1932. Zur Kenntnis mariner Nematoden aus der Kieler Bucht, *Zool. Jahrb. (Abt. Syst.)*, 62, 4, 329–430.
- Schulz, E. 1935. Marine Nematoden von Sicilien und Gran Canaria, *Zool. Anz.*, vol. 109, pp. 299–304.
- Schuermans-Stekhoven, J.H. 1931. Ökologische und morphologische

- Notizen über Zuiderseenematoden, *Ztschr. Morph. Ökol. Tiere*, 20, 4, 613–678.
- Schuurmans-Stekhoven, J.H. 1935a. Nematoda errantia. In *Tierwelt Nord- und Ostsee*, vol 5, pt. 28, 174 pp. Leipzig.
- Schuurmans-Stekhoven, J.H. 1935b. Nematodes libres capturés entre les hydraires dans le port de Plymouth, *Bull. Mus. Roy. Hist. Nat.*, 11, 33, 1–2. Belgium.
- Schuurmans-Stekhoven, J.H. 1943. Freilebende marine Nematoden des Mittelmeeres, *Zool. Jahrb. (Abt. Syst.)*, 76, 4, 323–380.
- Schuurmans-Stekhoven, J.H. 1946. Freilebende marine Nematoden des Skagerracks und der Umgebung von Stockholm, *Ark. Zool.*, vol. 37A, pp. 1–91.
- Schuurmans-Stekhoven, J.H. 1950. Free-living marine nemas of the Mediterranean. 1. The Bay of Villefranche, *Mém. Inst. Roy. Sci. Nat.*, ser. 2, fasc. 37, 220 pp. Belgium.
- Schuurmans-Stekhoven, J.H. and W. Adam. 1931. Free-living marine nemas of the Belgian coast, *Mém. Mus. Roy. Hist. Nat.*, no. 49, pp. 1–58. Belgium.
- Schuurmans-Stekhoven, J.H. and P.M. Mawson. 1955. On some free-living marine nematodes from Kerguelen Island, *Journ. Helminth.*, 29, 1–2, 87–104.
- Sergeeva, N.G. 1972. Novye vidy svobodnozhivushchikh nematod otriyada Enoplida Chernogo morya (New species of free-living nematodes of order Enoplida of the Black Sea). *Zool. Zhurn.*, 51, 8, 1233–1237.
- Southern, R. 1914. Nematelmia, Kinorhyncha and Chaetognatha, *Proc. Roy. Irish Acad.*, vol. 31, pt. 54, pp. 1–80.
- Steiner, G. 1915. Freilebende marine Nematoden von der Ostküste Sumatras, *Zool. Jahrb. (Abt. Syst.)*, vol. 38, pp. 233–244.
- Steiner, G. 1916. Freilebende Nematoden aus der Barentssee, *Zool. Jahrb. (Abt. Syst.)*, vol. 39, pp. 511–676.
- Steiner, G. 1922a. Beiträge zur Kenntnis mariner Nematoden, *Zool. Jahrb. (Abt. Syst.)*, vol. 44, pp. 1–68.
- Steiner, G. 1922b. Untersuchungen über den allgemeinen Bauplan des Nematoden-körpers, *Zool. Jahrb. (Abt. Anat.)*, vol. 43, pp. 1–96.
- Steiner, G. 1933. The nematode *Cylindrogaster longistoma* and its relationships, *J. Parasitol.*, 20, 1, 66–68.
- Steiner, G. and F. Albin. 1933. On the morphology of *Deontostoma californicum* n. sp., *J. Wash. Acad. Sci.*, vol. 23, pp. 25–30.
- Strassen, O.L. zur. 1904. *Antraconema*, eine neue Gattung freilebender Nematoden, *Zool. Jahrb., Suppl.* 7, pp. 301–346.
- Timm, R.W. 1951. A new species of marine nematode, *Thoracostoma magnificum*, with a note on possible “pigment cell” nuclei of the

- ocelli. *J. Wash. Acad. Sci.*, vol. 41, pt. 10, pp. 331–333.
- Timm, R.W. 1953. Observation on the monography and histological anatomy of marine nematode *Leptosomatum acephalatum* Chitwood, 1936 (Enopliidae, Leptosomatinae), *Amer. Midl. Nat.*, 49, 1, 229–248.
- Timm, R.W. 1959. New marine nematodes of superfamily Enoploidea from the Arabian Sea, *J. Bomb. Nat. Hist. Soc.*, vol. 56, pt. 2, pp. 204–210.
- Timm, R.W. 1960. A new species of *Leptosomatum* (Nematoda) from the Arabian Sea, *J. Helminth.*, 34, 3–4, 217–220.
- Türk, F. 1903. Über einige im Golf von Neapel freilebende Nematoden, *Mitt. Zool. Stat. Neapel*, vol. 16, pp. 281–347.
- Villot, A. 1875. Recherches sur les helminthes libres ou parasites des côtes de la Bretagne, *Arch. Zool. Exper.*, vol. 4, pp. 451–482.
- Vitiello, P. 1970. Nématodes libres marins des vases profondes du golf du Lion. 1. Enopliida, *Tethys*, 2, 1, 139–210.
- Wieser, W. 1951, 1954a. Untersuchungen über algenbewohnende Mikrofauna mariner Hartboden. I. Zur Ökologie und Systematik der Nematodenfauna von Plymouth, *Österr. Zool. Ztschr.*, 3, 3–4, 425–480; III. Zur Systematik der freilebenden Nematoden des Mittelmeeres, *Hydrobiologia*, 6, 12, (1954), 144–217.
- Wieser, W. 1952. Investigations of microfauna inhabiting seaweeds on a rocky coast. IV. Studies on the vertical distribution of fauna inhabiting seaweeds below the Plymouth laboratory, *J. Mar. Biol. Assoc.*, vol. 31, pt. 1, pp. 145–174. U.K.
- Wieser, W. 1953a. On the morphology of the head in family Leptosomatidae (marine free-living nematodes), with a key to all genera described, *Ark. Zool.*, ser. 2, 6, 1, 69–74.
- Wieser, W. 1953b. Die Beziehungen zwischen Mundhölengetalt, Ernährungsweise und Vorkommen bei freilebenden marinen Nematoden, *Ark. Zool.*, ser. 2, 4, 5, 439–484.
- Wieser, W. 1953c. Free-living marine nematodes. I. Enoploidea, *Lunds Univ. Arsskrift*, N. F. Avd. 2, 49, 6, 155.
- Wieser, W. 1954b. Beiträge zur Kenntnis der Nematoden submariner Höhlen. Ergebnisse der österreichischen Tyrrhenia Expedition, *Österr. Zool. Ztschr.*, 5, 12, 172–230.
- Wieser, W. 1956. Some free-living marine nematodes, *Galathea Repts.*, vol. 2 (Sci. Res. Dan. Deep-Sea Exp. 1950–1952), pp. 243–253.
- Wieser, W. 1959. Free-living nematodes and other small invertebrates of Puget Sound beaches, *Univ. Wash. Publ. Biol.*, vol. 19, pp. 1–170.
- Wieser, W. 1960. Benthic studies in Buzzard Bay. II. Meiofauna, *Limnol. and Oceanogr.*, 5, 2, 121–137.
- Wieser, W. and J. Kanwischer. 1961. Ecological and physiological studies

on marine nematodes from a small salt marsh near Wood Hole, Massachusetts, *Limnol. and Oceanogr.*, 6, 3, 262-270.

Winslow, W. 1960. Nematode ecology. In *Nematology* edited by J.N. Sasser and W.R. Jenkins. Univ. North Carolina Press, Chapel Hill, pp. 341-415.