

ZITHOLELE CONSULTING ON BEHALF OF ESKOM HOLDINGS

**D615 - SOLAR INTEGRATION PROJECT
ENVIRONMENTAL IMPACT ASSESSMENT
DRAFT BIOPHYSICAL REPORT**

REFERENCE NUMBER: D615

Report No.: JW194/12/D615

5 October 2012



Jones & Wagener

Consulting Civil Engineers

59 Bevan Road PO Box 1434 Rivonia 2128 South Africa
Tel: 00 27 (0)11 519 0200 Fax: 00 27 (0)11 519 0201 email: post@jawsco.za

DOCUMENT APPROVAL RECORD

Report No.: JW194/12/D615

ACTION	FUNCTION	NAME	DATE	SIGNATURE
Prepared	Environmental Consultant	Konrad Kruger	5 October 2012	
Reviewed				
Approved				

RECORD OF REVISIONS AND ISSUES REGISTER

Date	Revision	Description	Issued to	Issue Format	No. Copies

EXECUTIVE SUMMARY

Zitholele Consulting On Behalf Of Eskom Holdings has appointed Jones and Wagener (Pty) Ltd (J&W), an independent company, to conduct an Environmental Impact Assessment (EIA) to evaluate the potential environmental and social impacts of the proposed project. The Environmental Assessment Practitioner (EAP) is Konrad Kruger.

This report aims to identify the surface water, terrestrial ecology and soils that could be impacted by the proposed Solar Integration Project. From the detailed assessments it became clear that the bulk of the sensitivities in the study area are located around the Orange River, where the sensitive habitats as well as the main farming activities occur. The Orange River is also the only perennial water body in the area and of utmost importance to the Province.

The routes to Ferrum provided a different environment with the occurrence of the red Kalahari sands and in some cases dunes. These red sands are susceptible to erosion and also “shifting”, and could be a tricky obstacle when constructing.

As a whole the study area is devoid of access routes and access to the alternatives that are far from existing provincial or national roads might be problematic.

The study identified preferred alternatives for each route, based on the potential impacts to sensitive features along the routes. In addition mitigation and management measures were proposed for each of the criteria assessed and with the successful implementation of these measures, it is the opinion of the consultant that the impacts from this proposed development are within the acceptable range.

ZITHOLELE CONSULTING ON BEHALF OF ESKOM HOLDINGS
D615 - SOLAR INTEGRATION PROJECT

DRAFT BIOPHYSICAL REPORT

REPORT NO: JW194/12/D615

<u>CONTENTS</u>	<u>PAGE</u>
1. INTRODUCTION.....	1
1.1 Background Information	1
1.2 Regional Setting	2
1.3 Project Team Details	2
1.4 Objectives of this Report	4
2. PROJECT DESCRIPTION.....	4
2.1 Project Location.....	4
2.1.1 Solar Substation to Ferrum Substation.....	4
2.1.2 Solar Substation to Aries and Nieuwehoop Substation	4
2.1.3 Solar Substation and 132 kV routes	5
2.2 Components of the Project.....	5
3. RECEIVING ENVIRONMENT	10
3.1 Introduction.....	10
3.2 Climate	10
3.2.1 Data Collection and Methodology	10
3.2.2 Regional Description	10
3.2.3 Sensitivities	11
3.3 Topography and Drainage.....	11
3.3.1 Data Collection and Methodology	11
3.3.2 Regional Description	14
3.3.3 Sensitivities	14
3.4 Soils.....	14
3.4.1 Data Collection and Methodology	14
3.4.2 Regional Description	14
3.4.3 Site Description	15
3.4.4 Sensitivities	20
3.5 Agricultural Potential (Land Capability)	25

3.5.1	<i>Data Collection and Methodology</i>	25
3.5.2	<i>Regional Description</i>	25
3.5.3	<i>Site Description</i>	25
3.5.4	<i>Sensitivities</i>	26
3.6	Terrestrial Biodiversity	29
3.6.1	<i>Data Collection and Methodology</i>	29
3.6.2	<i>Regional Description</i>	29
3.6.3	<i>Site description</i>	35
3.6.4	<i>Fauna</i> 41	
3.6.5	<i>Sensitivities</i>	41
3.7	Surface Water and Wetlands	48
3.7.1	<i>Data Collection and Methodology</i>	48
3.7.2	<i>Regional Description</i>	49
3.7.3	<i>Site description/delineation</i>	49
3.7.4	<i>Sensitivities</i>	53
3.8	Aquatic Ecology	57
4.	IMPACT ASSESSMENT METHODOLOGY	57
5.	IMPACT ASSESSMENT	58
5.1	Soils and Land Capability	58
5.1.1	<i>Existing impact</i>	58
5.1.2	<i>Additional impact</i>	59
5.1.3	<i>Mitigation/management measures</i>	62
5.1.4	<i>Cumulative impact</i>	62
5.2	Terrestrial Ecology	63
5.2.1	<i>Existing impact</i>	63
5.2.2	<i>Additional impact</i>	63
5.2.3	<i>Mitigation/management measures</i>	65
5.2.4	<i>Cumulative impact</i>	66
5.3	Surface Water and Wetlands	66
5.3.1	<i>Existing impact</i>	66
5.3.2	<i>Additional impact</i>	66
5.3.3	<i>Mitigation/management measures</i>	67
5.3.4	<i>Cumulative impact</i>	67
6.	PREFERRED ALTERNATIVES	67
7.	CONCLUSION AND WAY FORWARD	68

8. REFERENCES..... 68

LIST OF TABLES

Table 1: Agricultural Potential criteria.....	25
Table 2: Land Capability of the soils within the study site	26
Table 3: Soil and Land Capability Initial Impact Assessment.....	59
Table 4: Impacts to soils for each route alternative.....	59
Table 5: Additional impact by the proposed development to the soils and agriculture.....	60
Table 6: Soil and Land Capability Cumulative Impact Assessment	62
Table 7: Vegetation Initial Impact Assessment	63
Table 8: Vegetation Initial Impact Assessment – Lower Gariep Alluvial Vegetation	63
Table 9: Vegetation Additional Impact Assessment	63
Table 10: Vegetation Impact per route	64
Table 11: Vegetation Additional Impact Assessment – Lower Gariep Alluvial Vegetation.....	65

LIST OF FIGURES

Figure 1: Regional locality of the proposed study area.	3
Figure 2: Mean Annual Rainfall.....	10
Figure 3: Mean annual temperatures	11
Figure 4: Lightning Ground Flash Density.....	11
Figure 5: Landforms of the study area (Ferrum)	12
Figure 6: Landforms of the study area (Aries and Nieuwehoop).....	13
Figure 7: Dundee soil form.....	16
Figure 8: Rocky areas on site, just south of the Orange River.....	16
Figure 9: Mispah (left) and Glenrosa (right) soil forms	17
Figure 10: Namib soil form (left) red soils on site (right).....	19
Figure 11: Plooyburg (left) and Kimberley (right) soil forms	19
Figure 12: Mixed shallow soils on site.....	20
Figure 13: Coega soil form (left) and Brandvlei soil form (right).....	20
Figure 14: Soil map for the study area	21
Figure 15: Distribution line soils	22
Figure 16: Soil sensitivity map.....	23
Figure 17: Distribution Soil Sensitivity.....	24
Figure 18: Agricultural potential for the study area.....	27
Figure 19: Agricultural Potential for the Distribution Lines	28
Figure 20: Vegetation of the study area	31
Figure 21: Vegetation of the Ferrum Routes	32
Figure 22: Vegetation of the southern section of the study area.....	33
Figure 23: Vegetation for the Distribution Lines	34
Figure 24: Kathu Bushveld showing the vegetation as well as the impact of an existing power line	35
Figure 25: Olifantshoek Plains Thornveld from the air (left) and ground level (right).....	36
Figure 26: Koranna-Langeberg Mountain Bushveld.....	36
Figure 27: Gordonia Plains Shrubland	37
Figure 28: The salt pan found along the Ferrum-Solar 1 Route.....	37
Figure 29: Gordonia Duneveld showing the typical red dunes.....	38
Figure 30: Kalahari Karroid Shrubland	38
Figure 31: Bushmanland Arid Grassland	39
Figure 32: Lower Gariep Broken Veld showing protected “kokerboom” on the left.....	39
Figure 33: Lower Gariep Alluvial Vegetation, showing the encroachment from agriculture (left).....	40
Figure 34: Bushmanland Basin Shrubland surrounding Aries substation	41
Figure 35: Endangered habitat as well as Critical Biodiversity Areas (CBA's).....	43
Figure 36: Zoomed in view of endangered habitat crossing.....	44

Figure 37: CBA's in relation to the distribution lines..... 45

Figure 38: *Acacia erioloba*..... 46

Figure 39: *Acacia haematoxylon* (foreground) and *A. erioloba* (background)..... 47

Figure 40: Shepard's Tree..... 47

Figure 41: Surface Water features on site..... 51

Figure 42: National Wetland classification system (SANBI, 2009)..... 53

Figure 43: Surface Water Features 54

Figure 44: Crossing the Orange River..... 55

Figure 45: Distribution Routes Surface Water Map..... 56

Figure 46: Impact Assessment Methodology 57

APPENDICES

APPENDIX A – SPECIES LISTS

APPENDIX B – OTHER

TERMS AND ABBREVIATIONS

BA	Basic Assessment
CBA	Critical Biodiversity Area
DAFF	Department of Agriculture, Forestry and Fisheries
DEA	Department of Environmental Affairs
DWEA.....	Department of Water and Environmental Affairs
DWA	Department of Water Affairs
DWAF	Department of Water Affairs and Forestry
EA	Environmental Authorisation
EAP.....	Environmental Assessment Practitioner
EIA	Environmental Impact Assessment
EIR.....	Environmental Impact Report
EMPr.....	Environmental Management Program (NEMA)
GN	Government Notice
HIA.....	Heritage Impact Assessment
I&APs.....	Interested and Affected Parties
IEM	Integrated Environmental Management
IWULA	Integrated Water Use Licence Application
km	kilometres
kV.....	kilovolt
m.....	metres
m ³	cubic metres
mamsl	metres above mean sea level
NEMA	National Environmental Management Act
NEM:WA.....	National Environmental Management Waste Act
NFEPA.....	National Freshwater Ecosystem Priority Area
NWA	National Water Act
R	Regulation
S&EIR.....	Scoping and Environmental Impact Reporting Process
SAHRA	South African Heritage Resources Agency
SANBI.....	South African National Biodiversity Institute
SIA	Social Impact Assessment
SR.....	Scoping Report
ToR.....	Terms of Reference
WUL.....	Water Use Licence
WULA	Water Use Licence Application



Jones & Wagener

Consulting Civil Engineers

59 Bevan Road PO Box 1434 Rivonia 2128 South Africa
Tel: 00 27 (0)11 519 0200 Fax: 00 27 (0)11 519 0201 email: post@jawsco.za

ZITHOLELE CONSULTING ON BEHALF OF ESKOM HOLDINGS

D615 - SOLAR INTEGRATION PROJECT

DRAFT BIOPHYSICAL REPORT

REPORT NO: JW194/12/D615

1. INTRODUCTION

1.1 Background Information

Eskom Holdings SOC Limited (Eskom) is the main South African utility that generates, transmits and distributes electricity and whilst Eskom's reliance on coal fired power stations has allowed for the generation of some of the cheapest electricity in the world at ~R10/W, it has resulted in South Africa being the largest producer of greenhouse gases in Africa, and one of the Top 20 greenhouse gas producing countries in the world.

South Africa and Eskom have started to focus on more reliable energy generation and demonstration projects and research, undertaken by Eskom, have shown that both solar and wind energy show great potential in South Africa. As a result (and in view of reducing their carbon footprint) Eskom is looking to increase the renewable energy component of its supply mix to at least 1 600 MW by 2025.

The power supply crisis has also accelerated the need to diversify Eskom's energy mix and its move towards alternative energy sources such as nuclear power and natural gas, as well as various forms of renewable energy.

The Upington area has been identified as one of the highest solar radiation locations in the world, providing the best opportunities for using the sun to generate electricity. In an effort to utilise renewable energy resources to meet the growing demand for electricity, the South African Government proposes the establishment of a R 150 billion Solar Park at Klipkraal, ~15 km west of Upington in the Northern Cape. The Solar Park will use the sun's energy to eventually generate 1 500 MW of electricity.

Eskom is planning constructing a 100 MW Concentrating Solar Power (CSP) plant at the Solar Park. This employs an array of mirrors controlled by tracking systems to focus a large area of sunlight into a small beam. The resulting heat is used to generate electricity. CSP also has the backing of the World Bank, which views it as the only zero-emission technology that could potentially rival coal-fired power. Eskom received a positive Record of Decision (RoD), approving a 100 MW CSP facility for this project in August 2007.

The Department of Energy as well as several Independent Power Producers (IPPs) are busy with investigations to construct solar plants at the Solar Park which should source sufficient electricity to make up the 1 500 MW planned for the solar park.

The electricity generated at the Upington Solar Park (by IPP's and Eskom) will need to be integrated into the National Grid. The purpose of the Solar Park Integration Project

JONES & WAGENER (PTY) LTD REG NO. 1993/02655/07 VAT No. 4410136685

DIRECTORS: PW Day (Chairman) PrEng MS(Eng) FSAICE D Brink (CEO) PrEng BEng(Hons) FSAICE PG Gage PrEng CEng BS(Eng) GDE/MSAICE/AsStructE JP van der Berg PrEng PhD MEng MSAICE TT Goba PrEng MEng FSAICE GR Wardle (Alternate) PrEng MS(Eng) FSAICE
TECHNICAL DIRECTORS: JA Kempe PrEng BS(Eng) GDE/MSAICE/AsStructE JR Shamrock PrEng MS(Eng) MSAICE/MW/MJE Glendinning PrSciNat MS(Env Geochem) NJ Vermeulen PrEng PhD MEng MSAICE DC Rowe PrEng BS(Eng) MSAICE A Oosthuizen PrEng BEng(Hons) MSAICE
ASSOCIATES: BR Antrobus PrSciNat BS(Hons) MSAIEG MW Palmer MS(Eng) AMSAICE AJ Bain BEng AMSAICE HR Aschenborn PrEng BEng(Hons) MSAICE PJJ Smit BEng(Hons) AMSAICE R Puchner PrSciNat MS(Geol) MSAIEG MAEG TG le Roux PrEng MEng MSAICE M van Zyl PrSciNat BS(Hons) MIWM
CONSULTANTS: W Ellis PrEng CEng MStructE **FINANCIAL MANAGER:** HC Neveling BCom MBL

Member of Consulting Engineers South Africa

is to address the major infrastructural investments that Eskom will need to make in order to tie the Upington Solar Park into the National Grid. The proposed Solar Park Integration Project entails the construction of a substation at the Upington Solar Park, 400kV transmission lines to the east and south of Upington to feed the electricity into Eskom's National Grid as well as the construction of a number of 132kV power lines inter-linking the IPP solar plants with the Eskom Grid and distributing the power generated to Upington.

1.2 Regional Setting

This project is located in the Northern Cape Province between the towns of Kathu, Upington, Kakamas and Kenhardt as shown in Figure 1.

1.3 Project Team Details

The following project personnel were involved in the compilation of this report.

Konrad Kruger, BSc Hons (Geog)

Mr. Konrad Kruger graduated from the University of Pretoria with a BSc in Environmental Science in 2002 and BSc Honours in Geography in 2003. He has been involved in a variety of environmental projects in the last eight years and has undertaken a variety of specialist studies, mapping and environmental consulting. The specialist studies included vegetation assessments, soil mapping and agricultural assessments, wetland delineations, visual assessments and terrestrial ecological assessments. In terms of similar work he has undertaken the biophysical specialist studies for the following approved EIA's for transmission lines:

- Camden (Ermelo) to Mbewu (Empangeni) 765 kV power lines;
- 400 kV power lines from Kusile Power Station to Lulamisa (Diepsloot);
- 400 kV power lines from Kendal Power Station to Zeus (Secunda); and
- 400 kV Duvha – Minerva power line deviation.

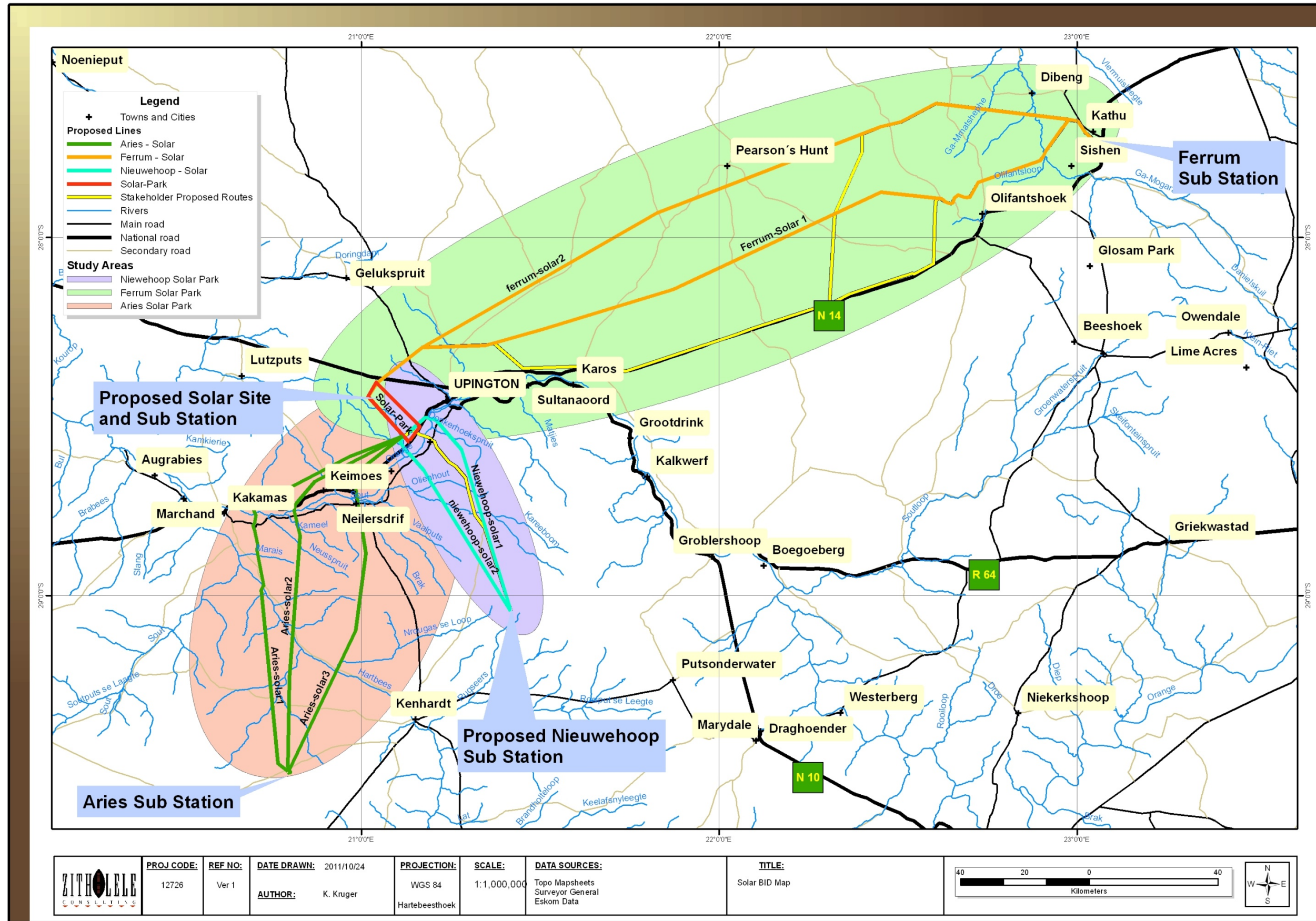


Figure 1: Regional locality of the proposed study area.

1.4 Objectives of this Report

This report aims to detail the specialist soil, agricultural potential, terrestrial ecology and surface water and wetland assessments that were undertaken for the Solar Integration Project. The information in this report will be included in the relevant EIA's for each of the Integration Project segments.

2. PROJECT DESCRIPTION

2.1 Project Location

As mentioned above, due to the size of the study area covered by this project, the description of the project is broken into three sections. The first section is the routes from the Solar Substation to the Ferrum Substation; the second is the two routes from the Solar Substation to the Aries and Nieuwehoop Substations. Lastly the third section of the route is made up of all the 132 kV lines that connect the Solar Substation with the surrounding solar projects in the park as well as with the Upington grid. Each of these is described in more detail below.

2.1.1 Solar Substation to Ferrum Substation

2.1.1.1. *Solar Ferrum Alternative 1*

Alternative 1 commences at the CSP outside of Upington traverses north-eastward approximately 15 km before turning eastwards for ~10 km. From here the route turns north-east again for ~ 120 km before meandering through the Langeberge for some 30 km. Lastly the route circles around Sishen Mine and into Kathu along other existing power lines to the existing Ferrum Substation.

2.1.1.2. *Solar Ferrum Alternative 2*

Alternative 2 commences at the CSP outside of Upington traverses north-eastward approximately 160 km before meandering through the Langeberge for some 10 km. Lastly the route joins Alternative 1 and circles around Sishen Mine and into Kathu along other existing power lines to the existing Ferrum Substation.

2.1.1.3. *Ferrum Stakeholder suggested Alternative:*

In addition to the alternatives mentioned above the stakeholders at the public meeting requested that an additional alternative be investigated during the EIA phase that is aligned with the N 14 highway rather than traversing through farming land

2.1.2 Solar Substation to Aries and Nieuwehoop Substation

2.1.2.1. *Solar Aries Alternative 1*

Alternative 1 commences at the CSP outside of Upington traverses south-westward along the Orange River and N14 Highway next to an existing 132 kV distribution line to just before Kakamas (about 60 km). There the line turns south, crosses over the Orange River and heads south for the 75 km to the Aries Substation, crossing over the Hartbees River.

2.1.2.2. *Solar Aries Alternative 2*

Alternative 2 commences at the CSP outside of Upington traverses south-westward along the Orange River and N14 Highway next to an existing 132 kV distribution line to 10 km before Kakamas (about 50 km). There the line turns south, crosses over the Orange River and heads south for the 75 km to the Aries Substation, crossing over the Hartbees River.

2.1.2.3. *Solar Aries Alternative 3*

Alternative 3 commences at the CSP outside of Upington traverses south-westward along the Orange River and N14 Highway next to an existing 132 kV distribution line up to 10 km after Loxtonvale (about 40 km). There the line turns south, crosses over the Orange River and heads south for the 75 km to the Aries Substation, crossing over the Hartbees River.

2.1.2.4. *Solar Nieuwehoop Alternative 1*

Alternative 1 commences at the CSP outside of Upington traverses north-eastward along the Orange River for 5 km. After Louisvale the line turn southeast, crosses over the Orange River and travels the approx. 60 km to the Nieuwehoop Substation, crossing over the Kareeboom River.

2.1.2.5. *Solar Nieuwehoop Alternative 2*

Alternative 2 commences at the CSP outside of Upington traverses south-westward for a very short distance (<2km) before turning southeast, crossing over the Orange River and travelling the approx. 60 km to the Nieuwehoop Substation, crossing over the Kareeboom River.

2.1.2.6. *Stakeholder suggested Alternative:*

In addition to the Nieuwehoop alternatives mentioned above the stakeholders at the public meeting requested that that an additional alternative be investigated during the EIA phase that is aligned along the local dirt road rather than traversing through farming land.

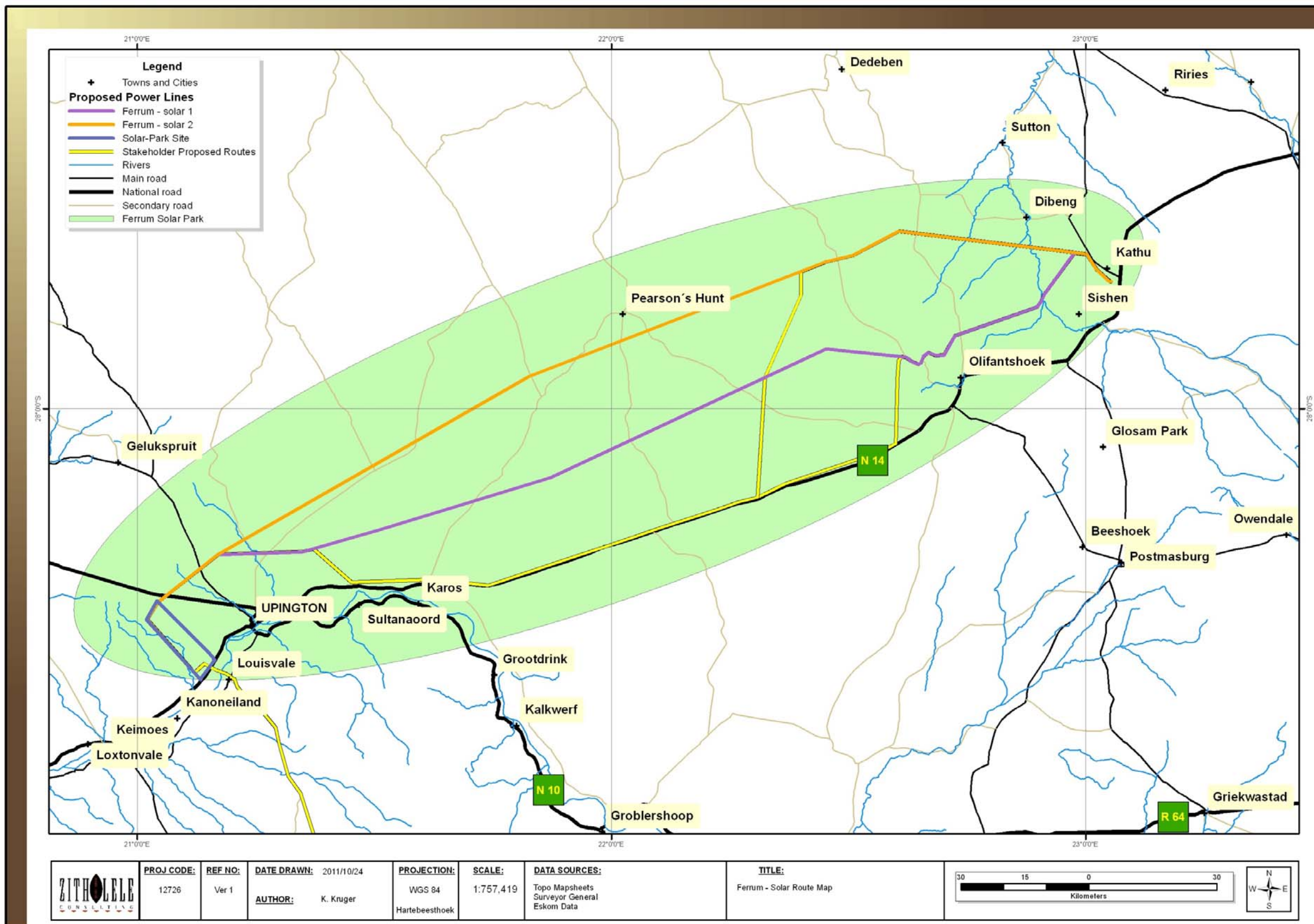
2.1.3 Solar Substation and 132 kV routes

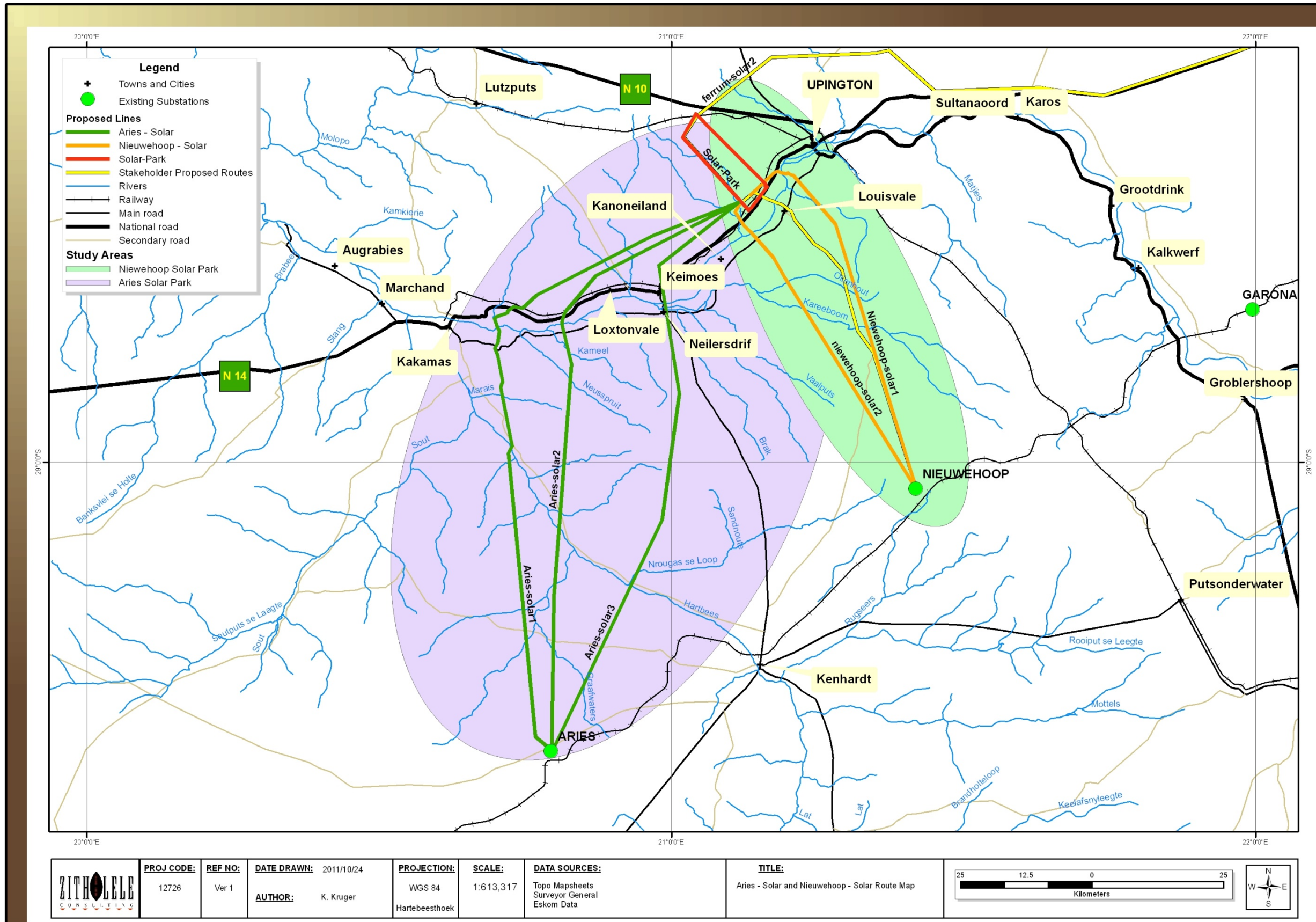
At the time of writing this report Eskom indicated that there are three potential locations for the proposed Solar Substation (refer to the Figure below) and provided the anticipated 132 kV power lines routes that will be utilised for each location. It should be noted that the Eskom CSP site has been approved in a previous EIA and it is assumed that all the line alignments inside this area have been included. The Eskom CSP will provide power to the DoE plant as well as the two IPP's on the adjacent properties as shown in the figure. In addition a 132 kV line will also provide power to the town of Upington via the Gordonia substation. As show on the map, no alternatives were given for these alignments and it is assumed that the 132 kV line to Upington will follow the existing line on site.

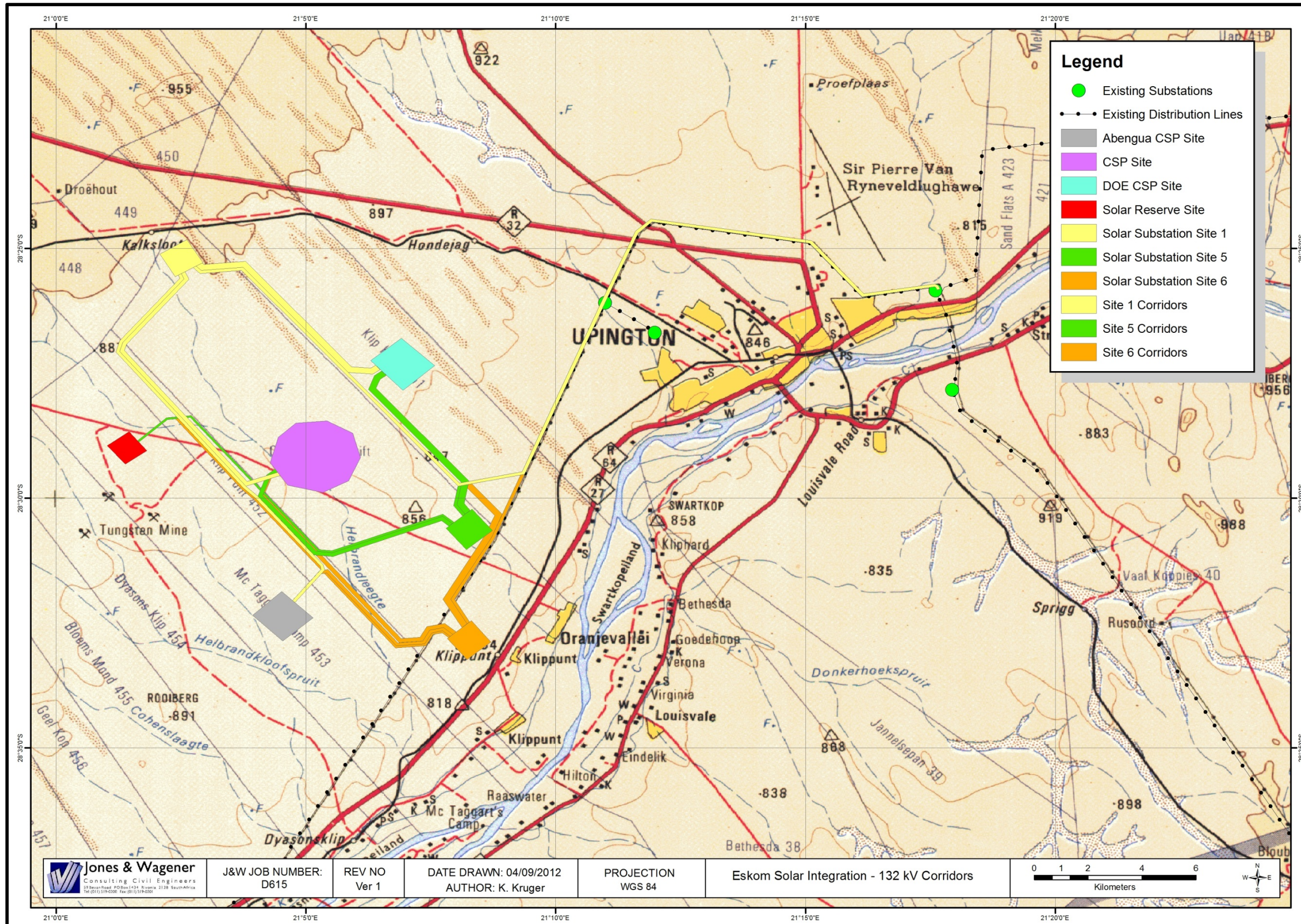
2.2 Components of the Project

The project under consideration includes the following main components once authorised:

- Power lines
 - 1 x 400kV to Ferrum;
 - 1 x 400 kV to Nieuwehoop;
 - 2 x 400 kV to Aries;
 - 2 x 132 kV to Gordonia; and
 - 11 x 132 kV to Eskom CSP, IPP's and DoE.
- Access routes;
- Construction camps;
- Construction, operation and maintenance of the above.







3. RECEIVING ENVIRONMENT

3.1 Introduction

This section provides a general description of the environment in which the proposed project will be located. The purpose of this section is to provide a perspective of the local environment within which the proposed infrastructure will exist and operate, with a view to identify sensitive issues/areas, such as wetlands or other ecological aspects, which need to be considered when conducting the impact assessment and designing the various components of the project.

3.2 Climate

3.2.1 Data Collection and Methodology

Broad scale meteoric data was obtained from the CSIR as well as information contained in the existing Eskom CSP site EIA. It should be noted that this was not a detailed study, but merely a desktop assessment as input into the other detailed studies.

3.2.2 Regional Description

Rainfall

The study area is located in the north western portion of South Africa. This area receives very variable late summer rainfall between February and April. The study area receives between 70 – 200mm of precipitation annually.

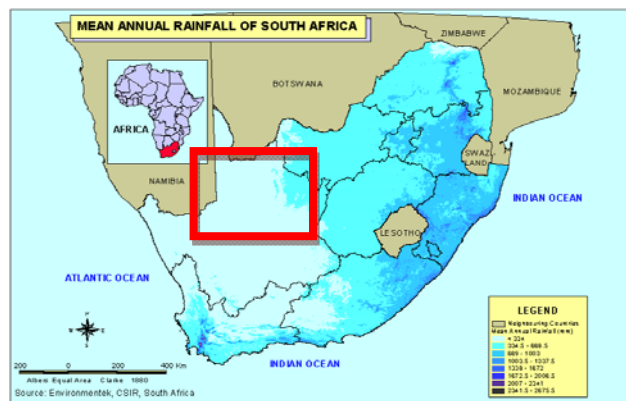


Figure 2: Mean Annual Rainfall

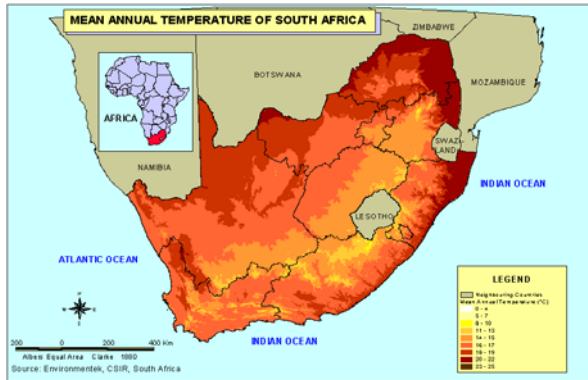


Figure 3: Mean annual temperatures

Temperature

The study area is located in one of the warmer parts of the country with mean max and min temperatures range from 40,6°C to -3,7°C with a mean annual temperature of 17,4°C.

3.2.3 Sensitivities

Wind

For the entire study area there is very low wind flow and no main wind direction. Whirl winds (dust devils) are common on hot summer days.

Lightning Strikes

The study area is located in an area with very low frequency of lightning strikes.

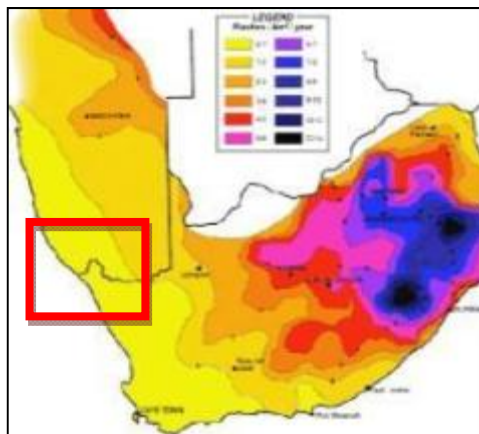


Figure 4: Lightning Ground Flash Density.

3.3 Topography and Drainage

3.3.1 Data Collection and Methodology

The topography data was obtained from the Surveyor General's 1:50 000 toposheet data for the region. Contours were combined from the topographical mapsheets to form a combined contours layer. Using the Arcview GIS software the landforms of the region was compiled and is shown in the Figure below.

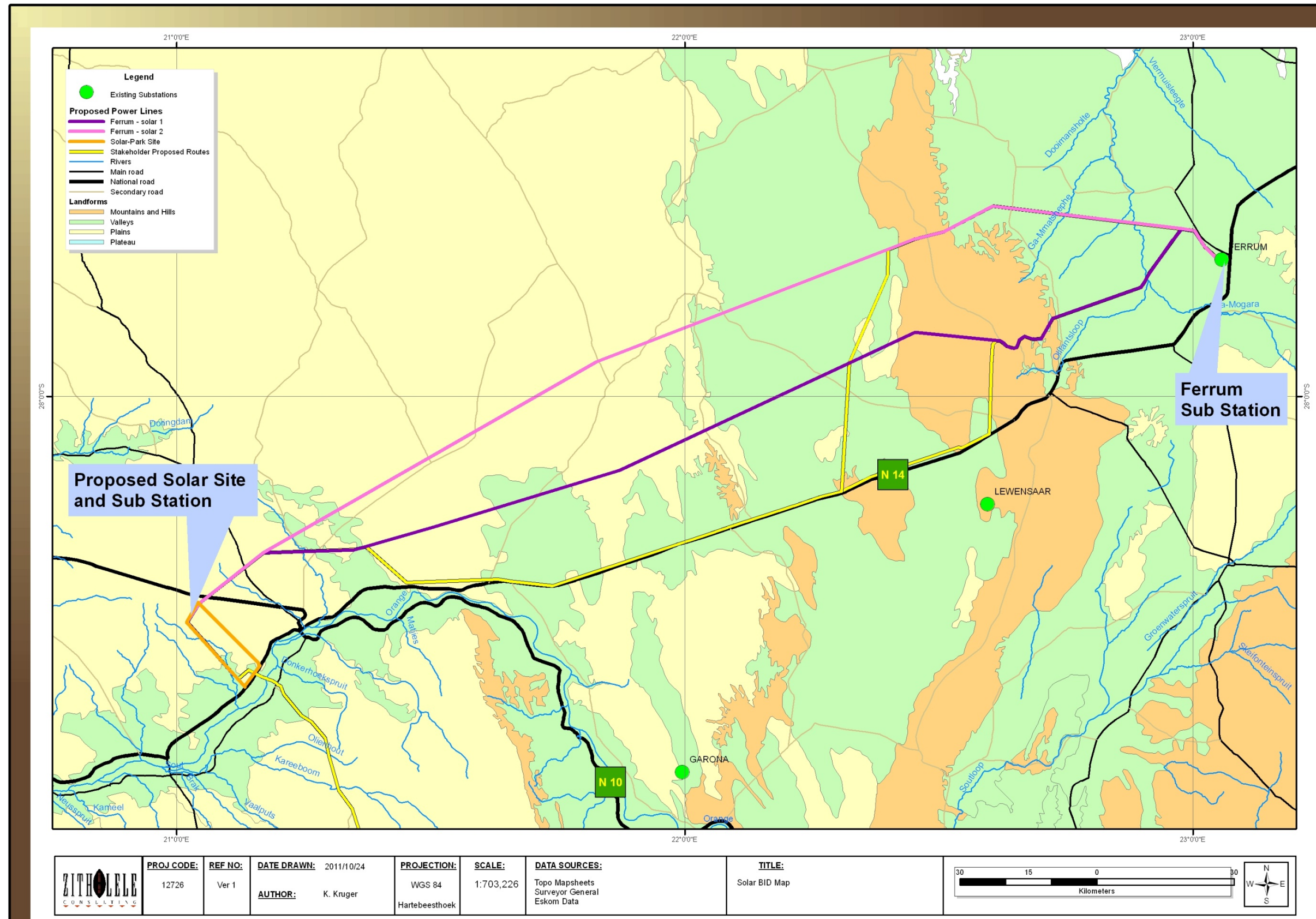


Figure 5: Landforms of the study area (Ferrum)

3.3.2 Regional Description

The altitude in the study area ranges from 600 mamsl (metres above mean sea level) to 1800 mamsl. The highest parts of the study area are in the eastern portions (Olifantshoek) and in the southern portions (Kenhardt) and the lowest portions are in the southern portions of the study area (Orange River).

The study area comprises of one major valley in the Orange River Basin and the Kalahari that all generally drain eastward. The area northeast of the Orange River is dominated by the Kalahari dunes and intermittent pans. On the eastern end of the study area the alternatives travel through the Langberge, a long linear mountain range that runs north-south through the study area. The area south of the Orange River is dominated by a flat plain with very few topographic features.

3.3.3 Sensitivities

In terms of topographical features no sensitivities exist although the dunefields in the Kalahari do afford some unique challenges discussed in more detail under the Soils Section below.

3.4 Soils

3.4.1 Data Collection and Methodology

The geological analysis was undertaken through desktop evaluation using a Geographic Information System (GIS) and relevant data sources. The geological data was taken from the Environmental Potential Atlas Data generated by the DEA. Soil data was obtained from the Department of Agriculture

The on-site soils assessment was conducted from August - October 2012. Soils were augered at 500-1 000m intervals along the proposed power line routes using a 150 mm bucket auger, up to refusal or 1.2 m. Soils were identified according to Soil Classification; a taxonomic system for South Africa (Memoirs on the Natural Resources of South Africa, no. 15, 1991). The following soil characteristics were documented:

- Soil horizons;
- Soil colour;
- Soil depth;
- Soil texture (Field determination);
- Wetness;
- Occurrence of concretions or rocks; and
- Underlying material (if possible).

3.4.2 Regional Description

The bulk (>30%) of the study area is covered by recent (Quaternary) alluvium and calcrete. Superficial deposits of the Kalahari Group are also present in the east. The extensive Palaeozoic diamictites of the Dwyka Group also outcrop in the area as do gneisses and metasediments of Mokolian age.

The soils derived from these geologies are mostly red-yellow apedal soils, freely drained with a high base status and < 300mm deep. Along the Orange River recent alluvial deposit from the River form the main soils forms.

3.4.3 Site Description

Following the site survey a number of soil forms were identified. The soils forms were grouped into management units and are described in detail in the sections below and Figure 14 illustrates the location of the soil types. The land capability (agricultural potential) of the abovementioned soil form is described in more detail in Section 3.5.

The management units are broken up into:

- Alluvial soils (Undifferentiated deep deposits);
- Rocky Areas;
- Sandy soils:
 - Red soils; and
 - Red and Yellow soils.

Each of these management units are described in more detail below.

3.4.3.1. Alluvial soils

These soils are mainly found along the Orange River floodplains and form the basis for most of the cultivation in the Northern Cape. The main soil form is the Dundee soil form which is shown below and typified by an Orthic A-horizon over a Stratified Alluvium. The stratification (layers) in the soil horizon is created by the deposition of material during flood events. The criteria for such a soil are as follows:

- is unconsolidated and contains stratifications caused by alluvial or colluvial deposition;
- directly underlies a diagnostic orthic or melanic A horizon, or occurs at the surface;
- does not qualify as diagnostic regic sand.

Unlike soil horizons that have developed by pedogenetic processes, stratified alluvium owes its distinguishing features to a depositional process and is thus not a sequence of so-called genetic horizons. Pedogenetic changes have been minimal and it is, properly, a C horizon or parent material. The rare occurrences of stratified colluvium are also accommodated by this concept.

Given time, homogenizing processes of soil formation will destroy the evidence of deposition: stratifications will disappear and be replaced by true genetic soil horizons, their kind depending upon the character of the particular material, the particular site and the particular external environment. However, alluvium is commonly utilized very intensively for crop production. For this practical reason, it has been regarded as desirable to recognise stratified alluvium as a diagnostic subsoil material. The classification reflects this importance of young alluvium by making provision, through a diagnostic horizon, for its easy inclusion. Other diagnostic subsoil horizons cater for the pedogenetic changes which affect alluvium with time.

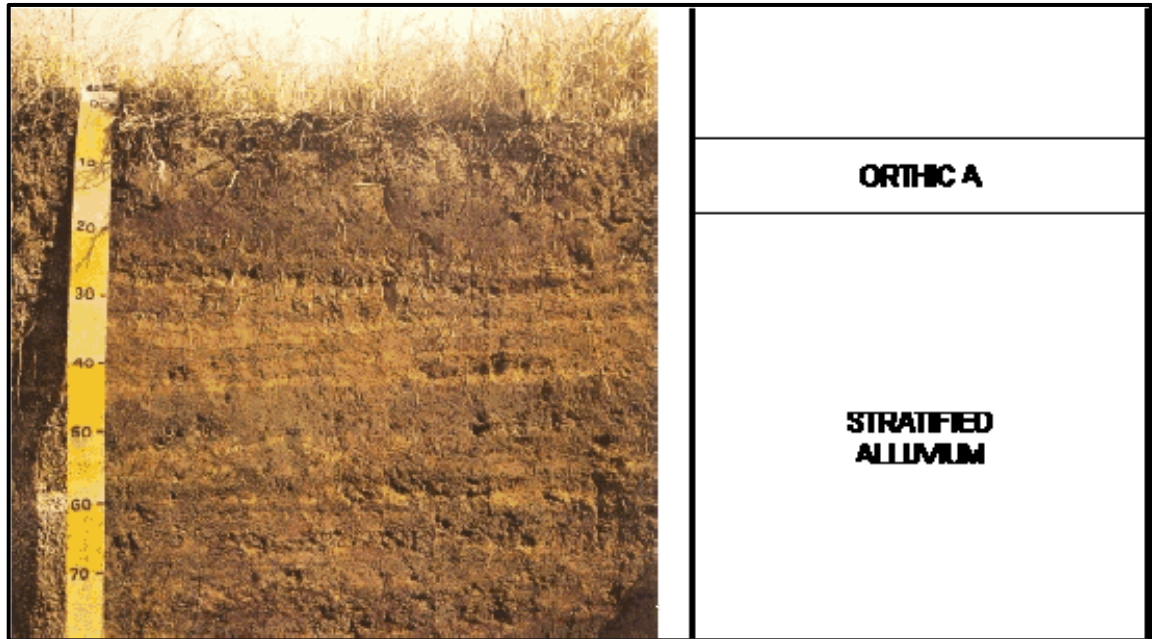


Figure 7: Dundee soil form

3.4.3.2. Rocky Areas

As shown on the Soils map for the study area there are two rocky soil types. The first is rocky areas with miscellaneous soils and the second is hard rock areas. In both cases the rock originates from shallow geology found throughout the study area. In the east of the study area the hard rock areas originate from the Langeberge and some isolated outcrops en route to Upington. The first unit of miscellaneous soils with rocky areas are found closer to the Orange River and is associated with the Inselbergs that can be found throughout the area. The soil forms that are found in these areas are illustrated below. These include the Mispah and Glenrosa soil forms and both are characterised by their shallow nature overlying a hard layer.



Figure 8: Rocky areas on site, just south of the Orange River

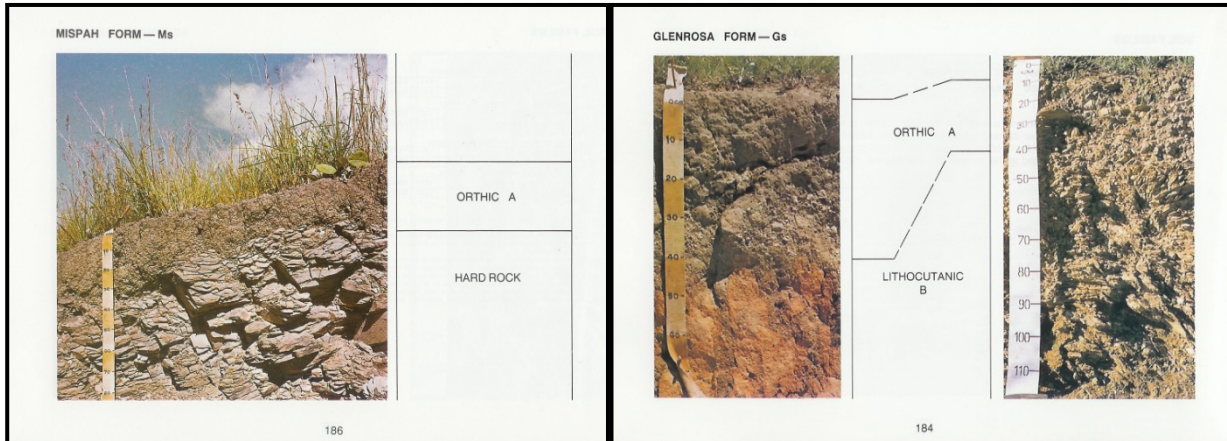


Figure 9: Mispah (left) and Glenrosa (right) soil forms

The lithocutanic B horizon found in the Glenrosa soil form has to comply to the following requirements:

- underlies a diagnostic topsoil horizon, either directly or via a stone-line, or an E horizon;
- merges into underlying weathering rock;
- has, at least in part, a general organization in respect of colour, structure or consistence which has distinct affinities with the underlying parent rock;
- has cutanic character expressed usually as tongues or prominent colour variegations caused by residual soil formation and illuviation resulting in the localization of one or more of clay, iron and manganese oxides, and organic matter in a non-homogenized matrix of geological material (saprolite) in a variable but generally youthful stage of weathering;
- lacks a laterally continuous horizon which would qualify as either a diagnostic pedocutanic B or prisma-cutanic B;
- does not qualify as a diagnostic podzol B, a neocarbonate B, a soft or hardpan carbonate horizon, or diagnostic dorbank;
- if the horizon shows signs of wetness, then more than 25% by volume has saprolite character.

The concept is one of minimal development of an illuvial B horizon in weathering rock. With the exception of its presence beneath an E horizon in Cartref form, the lithocutanic B occurs beneath a diagnostic topsoil horizon. In situ weathering of rock under a topsoil has produced a heterogeneous and, typically, highly variegated zone consisting of soil material (relatively well homogenized without traces of weathering rock) interspersed with saprolite or weathering rock in various stages of breakdown. The latter is recognised by its general organization with respect to structure, colour or consistence which still has distinct affinities with the parent rock. Furthermore, this zone grades into relatively unaffected and, eventually, fresh rock, sometimes at fairly shallow depth.

3.4.3.3. Sandy soils

This management unit described the bulk of the soils within the study area. Being an arid environment, very little pedogenesis has taken place and clay material is not common. Over the study area we have the red dunes of the Kalahari dominating the central region, and they are surrounded by deep red soil plains without dunes. To the

south and far east of the study area you find shallow red soils, mostly overlying calcrete and in the extreme south and eastern region you find mixed yellow and red soils with low clay percentages. Each of the soil forms found in these areas are illustrated and described below.

Deep red soils with and without dunes

The soil that dominates in these areas is the Namib soil form. This soil form is typified by a regic sand B-horizon that in the case of the study area is very red in colour. The illustration below shows a yellow version, however the photo on the right shows the colour of the soils within the study area. A regic sand has to meet the following criteria:

- is a recent deposit, usually aeolian, which, except for a possible darkening of the topsoil by organic matter, shows little or no further evidence of pedogenesis;
- is coarse textured and has little or no macroscopically visible structure; it may be massive or single grained; aeolian stratification (cross-bedding) may be present;
- may have any colour although "grey" as defined for the E horizon is common; aeolian stratification (cross-bedding), when present, prevents a material from qualifying as a diagnostic red or yellow-brown apedal B horizon or as an E-horizon;
- has mineralogical composition little, if any different from that of the parent material;
- has consistence that is loose, friable or soft;
- directly underlies an orthic A horizon or, if this is absent, occurs at the surface; and
- does not qualify as a neocutanic B, a neocarbonate B, an E horizon or as stratified alluvium.

The term regic (Gr.rhegos = blanket) is used here to convey the idea of cover sands in which, by virtue of their youth or environment, little or no profile development has taken place. The purpose of defining this class of materials as diagnostic is to provide a place in the classification for young sands of aeolian origin (red, yellow-brown or grey). Such materials often represent an important geographic entity in desert and littoral regions. Properties reflect minimal pedogenesis; essential is the fact that the mineralogical composition of the sand (e.g. quartz, feldspars, Ferro-magnesian minerals, shell fragments) is little if any different to that of the parent deposit and that there has been little if any clay formation.

Aeolian stratification (cross-bedding), if present, is diagnostic of regic sand; these should not, however, be confused with the more or less parallel, horizontally oriented lamellae which are common in certain E horizons. Because pedogenesis has been minimal in regic sand, changes within a sand body which are attributable to pedogenesis would tend to indicate the presence in the sand body of materials which do not qualify as regic sand. Regic sands are commonly but not necessarily deep.

When there is doubt as to whether a material qualifies as regic sand on the one hand or as a red or yellow-brown apedal B horizon on the other, regic sand is preferred when the sand body takes the form of a dune and, in the virgin state, vegetation is all but absent. The texture of regic sands is usually no finer than pure sand.

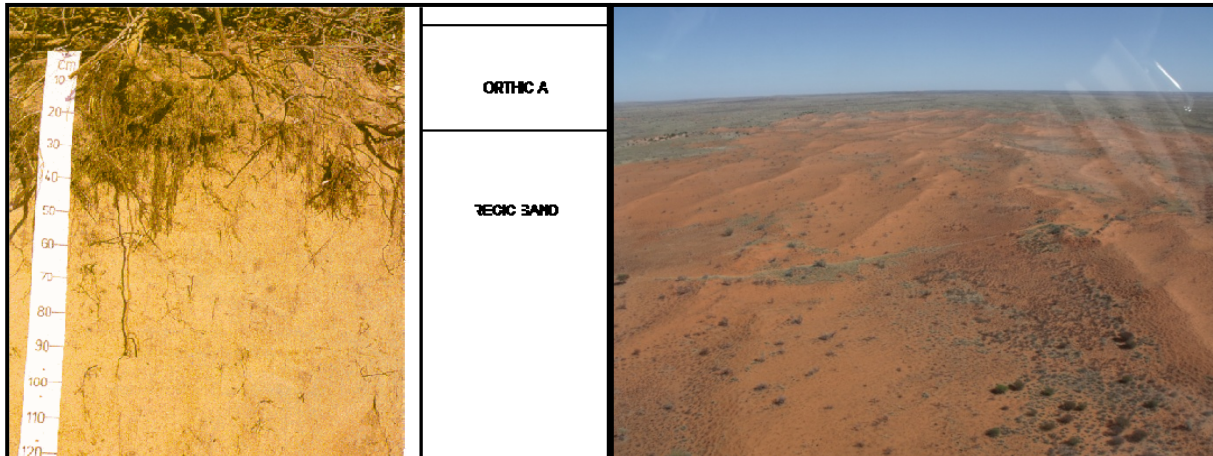


Figure 10: Namib soil form (left) red soils on site (right)

In cases where the regic sand horizon has undergone more pedogenesis this soil can be classified as a Hutton soil form, and in cases where the soil becomes shallow the Plooyburg soil form is found (as shown below).

Shallow red soils

The shallow red soils found throughout the study area most commonly overlie a calcrete layer, which in terms of the classification system is described as a soft Carbonate or a Hardpan Carbonate horizon. The dominant soils in this region are known as the Plooyburg and Kimberley soil forms as shown below. A Hardpan Carbonate layer is identified by the following criteria:

- is continuous throughout the pedon;
- is cemented by calcium and/or calcium-magnesium carbonates such as to be a barrier to roots and slowly permeable to water;
- is massive, vesicular or platy and extremely hard when dry and hard or very firm when moist;
- unless exposed by erosion, occurs beneath a melanic or orthic A, or yellow-brown apedal 8, red apedal B, neocutanic 8 or neocarbonate B horizon;
- does not qualify as diagnostic dorbank; and
- A laminar capping is common but not always present.

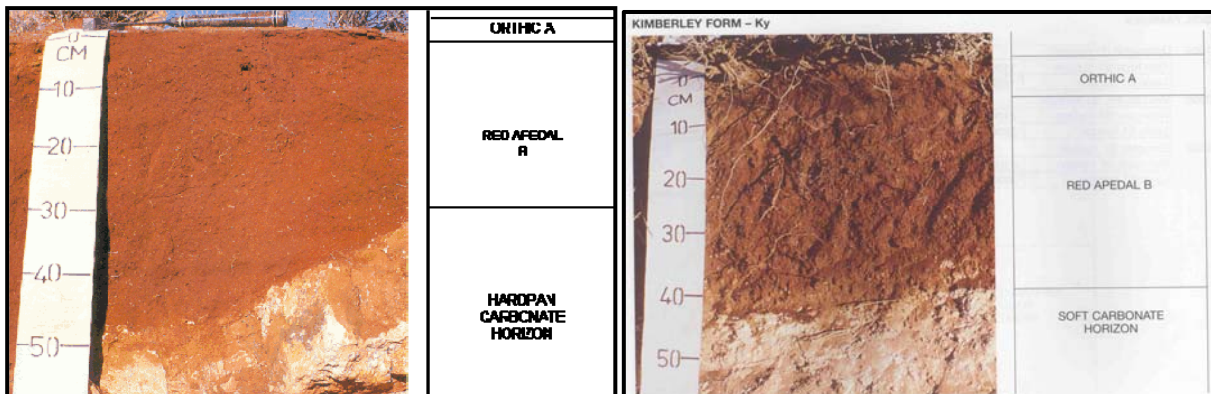


Figure 11: Plooyburg (left) and Kimberley (right) soil forms

Mixed red and yellow soils with very little clay

As the red sands of the Kalahari recede the soils start to become more diverse. These areas have a variety of soils including shallow calcrete, gravel plains and red or brown soils. Below are photos of the soils found in these areas.



Figure 12: Mixed shallow soils on site

The soil forms identified in this area include Coega, Brandvlei, Mispah, and Glenrosa. The latter two soils are described above, while the other two are shown below. Both the Coega and Brandvlei soils have carbonate horizons, in the Coega the concrete has hardened into an impenetrable layer.

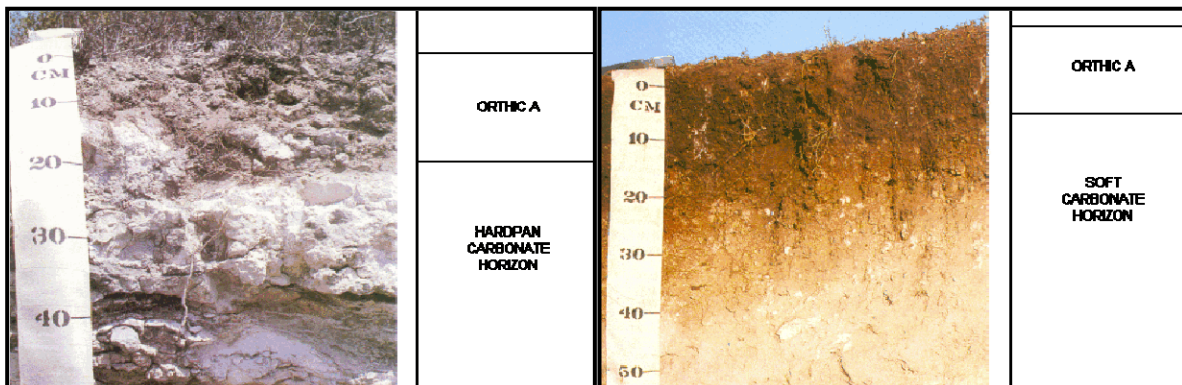


Figure 13: Coega soil form (left) and Brandvlei soil form (right)

3.4.4 Sensitivities

The potential sensitivities related to soils were screened using data from the Agricultural Research Council’s (ARC) website AGIS. The data from the ARC indicates that the area is prone to two potential sensitivities relating to soil – erosion and shifting sands. These are usually interrelated and in the dunefields of the Kalahari they are especially high as indicated in Figure 16 below. As shown in the map Ferrum – Solar 1 and 2 traverse large areas of potentially shifting sands.

The potential for shifting sands is caused by the high amounts of Sodium in the soil along with the sandy nature of the soil. These factors create an environment where soils easily disperse when water is introduced or erode when the vegetative cover is removed. These areas require special attention when constructing roads, erection and siting the pylon footings. These aspects are covered in the EMP and impact assessment.

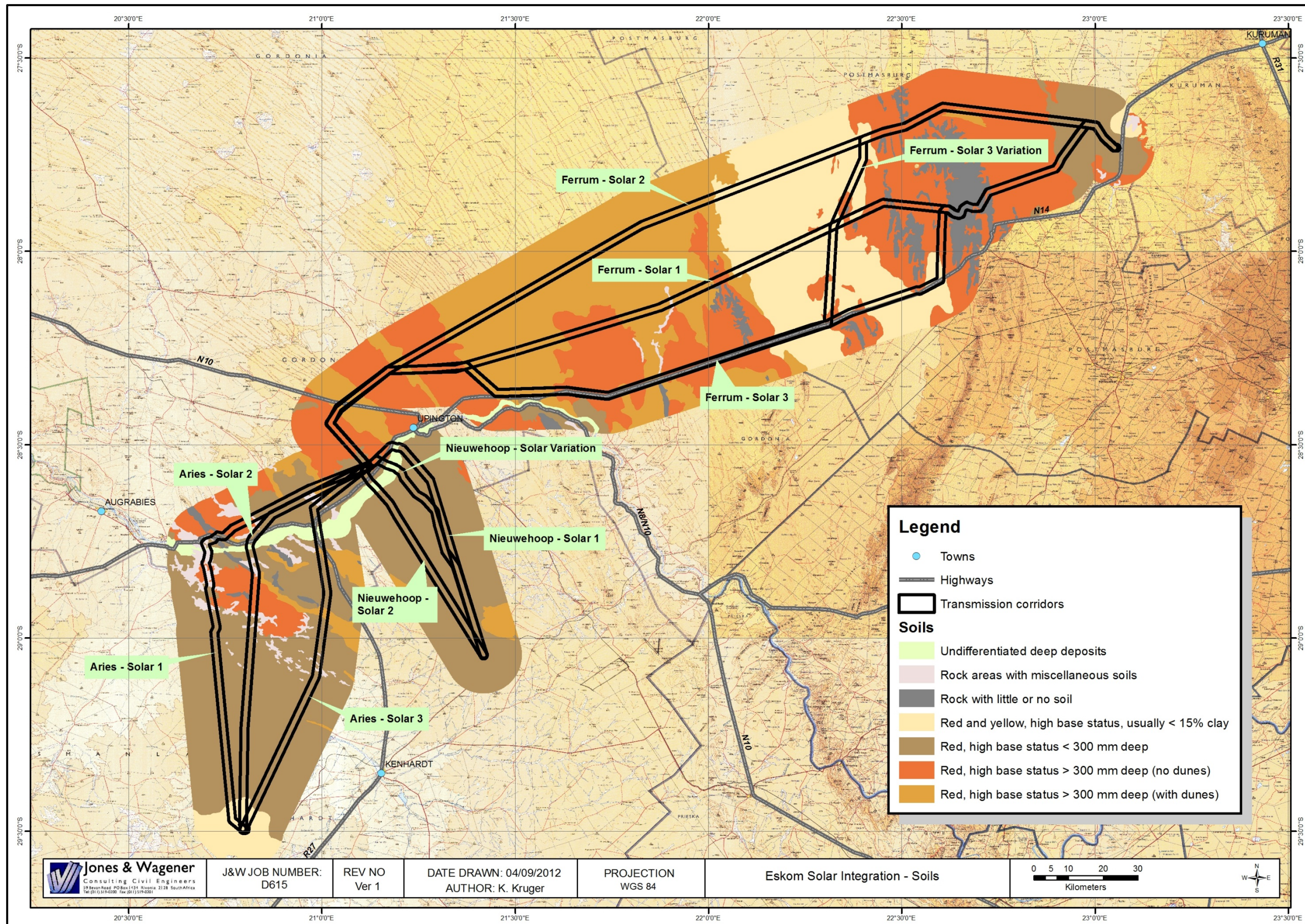


Figure 14: Soil map for the study area

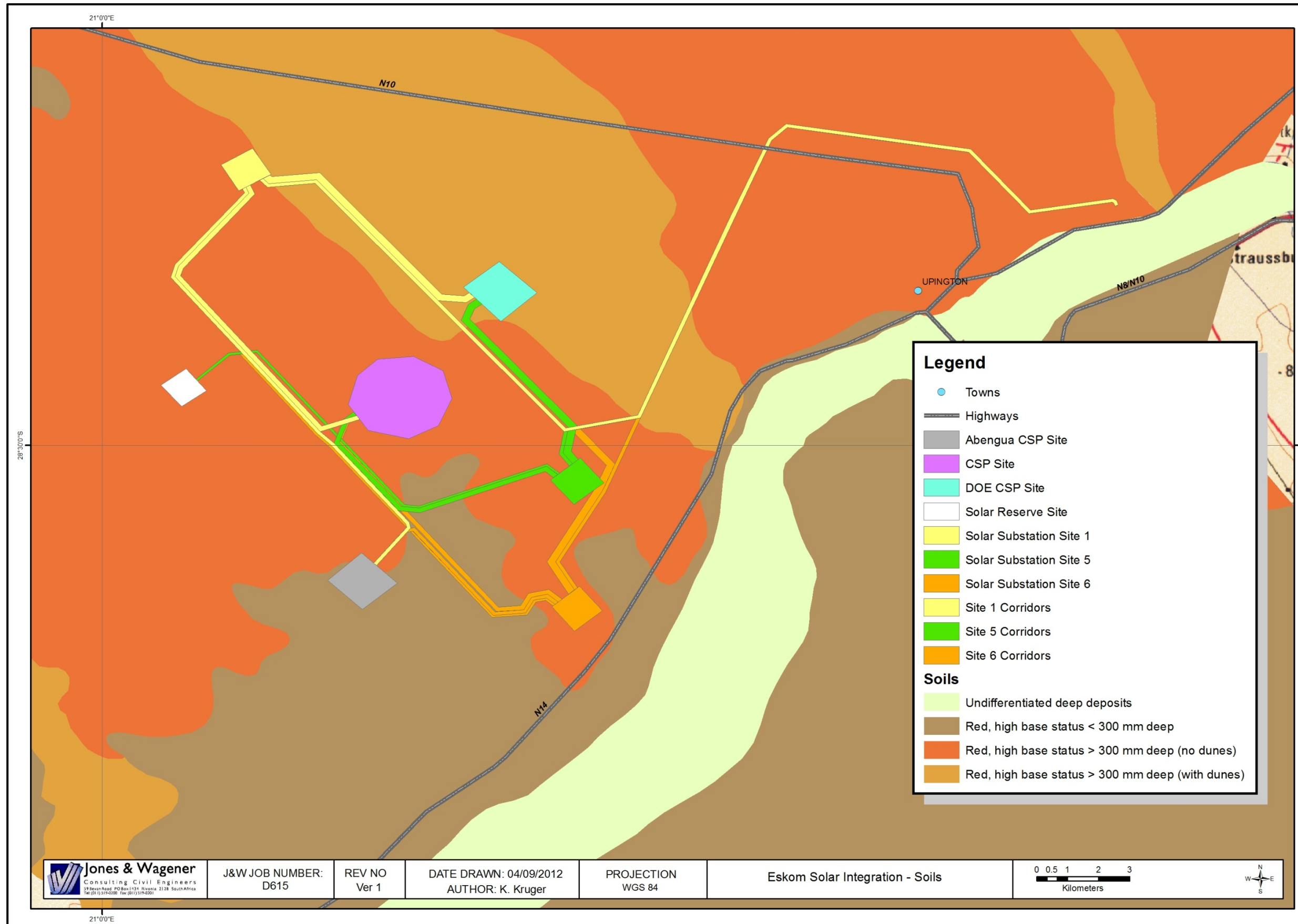


Figure 15: Distribution line soils

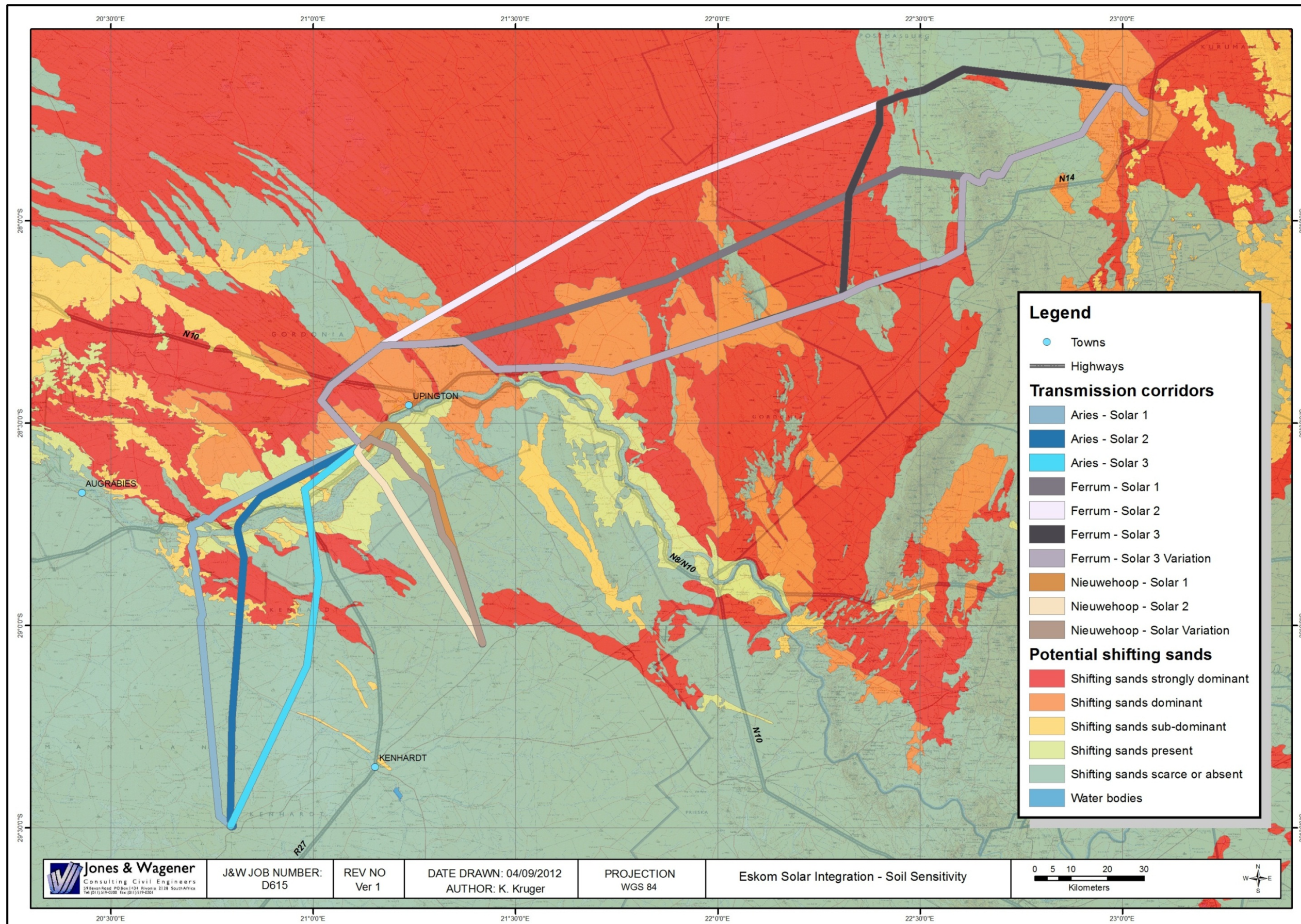


Figure 16: Soil sensitivity map

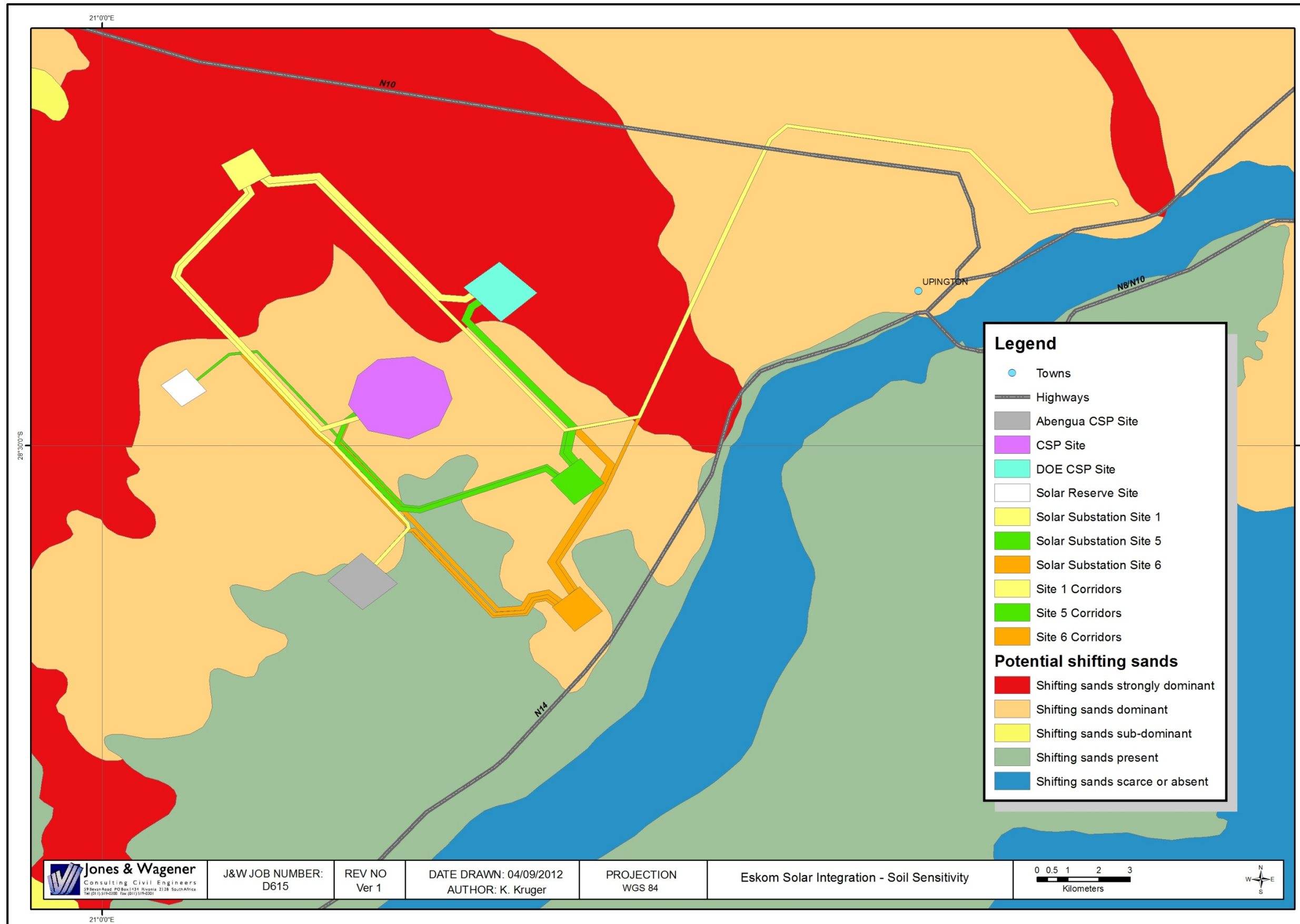


Figure 17: Distribution Soil Sensitivity

3.5 Agricultural Potential (Land Capability)

3.5.1 Data Collection and Methodology

Using the soil data collected during the site investigations and applying that to the land capability assessment methodology as outlined by the National Department of Agriculture¹, the agricultural potential/land capability of the site was determined.

3.5.2 Regional Description

Regionally the Northern Cape is not known for cultivation or high agricultural potential soils. The majority of the province is utilised for grazing of livestock due to the aridity and shallow soils that occur in the area.

3.5.3 Site Description

According to the land capability methodology, the potential for a soil to be utilised for agriculture is based on a wide number of factors. These are listed in the table below along with a short description of each factor.

Table 1: Agricultural Potential criteria

Criteria	Description
Rock Complex	If a soil type has prevalent rocks in the upper sections of the soil it is a limiting factor to the soil's agricultural potential
Flooding Risk	The risk of flooding is determined by the closeness of the soil to water sources.
Erosion Risk	The erosion risk of a soil is determined by combining the wind and water erosion potentials.
Slope	The slope of the site could potentially limit the agricultural use thereof.
Texture	The texture of the soil can limit its use by being too sandy or too clayey.
Depth	The effective depth of a soil is critical for the rooting zone for agricultural crops.
Drainage	The capability of a soil to drain water is important as most grain crops do not tolerate submergence in water.
Mechanical Limitations	Mechanical limitations are any factors that could prevent the soil from being tilled or ploughed.
pH	The pH of the soil is important when considering soil nutrients and hence fertility.
Soil Capability	This section highlights the soil type's capability to sustain agriculture.
Climate Class	The climate class highlights the prevalent climatic conditions that could influence the agricultural use of a site.

¹ Agricultural Research Council – Institute for Soil, Climate and Water (2002), *Development and Application of a Land Capability Classification System for South Africa*, Final Report to Directorate Agricultural Land Resource Management, National Department of Agriculture.

Criteria	Description
Land Capability / Agricultural Potential	The land capability or agricultural potential rating for a site combines the soil capability and the climate class to arrive at the sites potential to support agriculture.

The soils identified in Section 3.4 above were classified according to the methodology described above. The criteria mentioned above were evaluated in the table below.

Table 2: Land Capability of the soils within the study site

Soil	Agricultural	Sandy soils	Shallow Soil	Hard Rock
% on Site	1.7 %	48.3%	44.3%	5.7%
Rock Complex	None	None	Yes	Yes
Flooding Risk	High	None	None	None
Erosion Risk	Moderate	High	High	Very Low
Slope %	<4	<4	<4	>4
Texture	Loam	Sand	Sand	Rock/Sandy
Effective Depth	> 90 cm	> 30 cm	< 30 cm	< 10 cm
Drainage	Imperfect	Excellent	Poorly drained	Poorly drained
Mech Limitations	None	None	Rocks	Rocks
pH	> 5.5	> 5.5	> 5.5	> 5.5
Soil Capability	Class III	Class V	Class VI	Class VIII
Climate Class	Severe	Severe	Severe	Severe
Land Capability	Class III – Moderately Arable Land	Class VII – Grazing Land	Class VII – Grazing Land	Class VIII – Wildlife

No limitation	Low	Moderate	High	Very Limiting
---------------	-----	----------	------	---------------

The site is made up of three land capability classes, namely Class III, VII and VIII as shown in the Figure below. The Class III soils are suitable for cultivation but they have some restrictions – in this case flooding and climate. The Class VII soils have continuing limitations that cannot be corrected; in this case rock complexes, climate, stoniness, and a shallow rooting zone constitute these limitations. Class VIII soils are basically hard rock and have no agricultural use.

3.5.4 Sensitivities

Of the uses above, the agricultural soils located adjacent to the Orange River supports the agricultural cultivation core for the province. Impacts to these areas should be limited as the soils as well as the water sources are very limited.

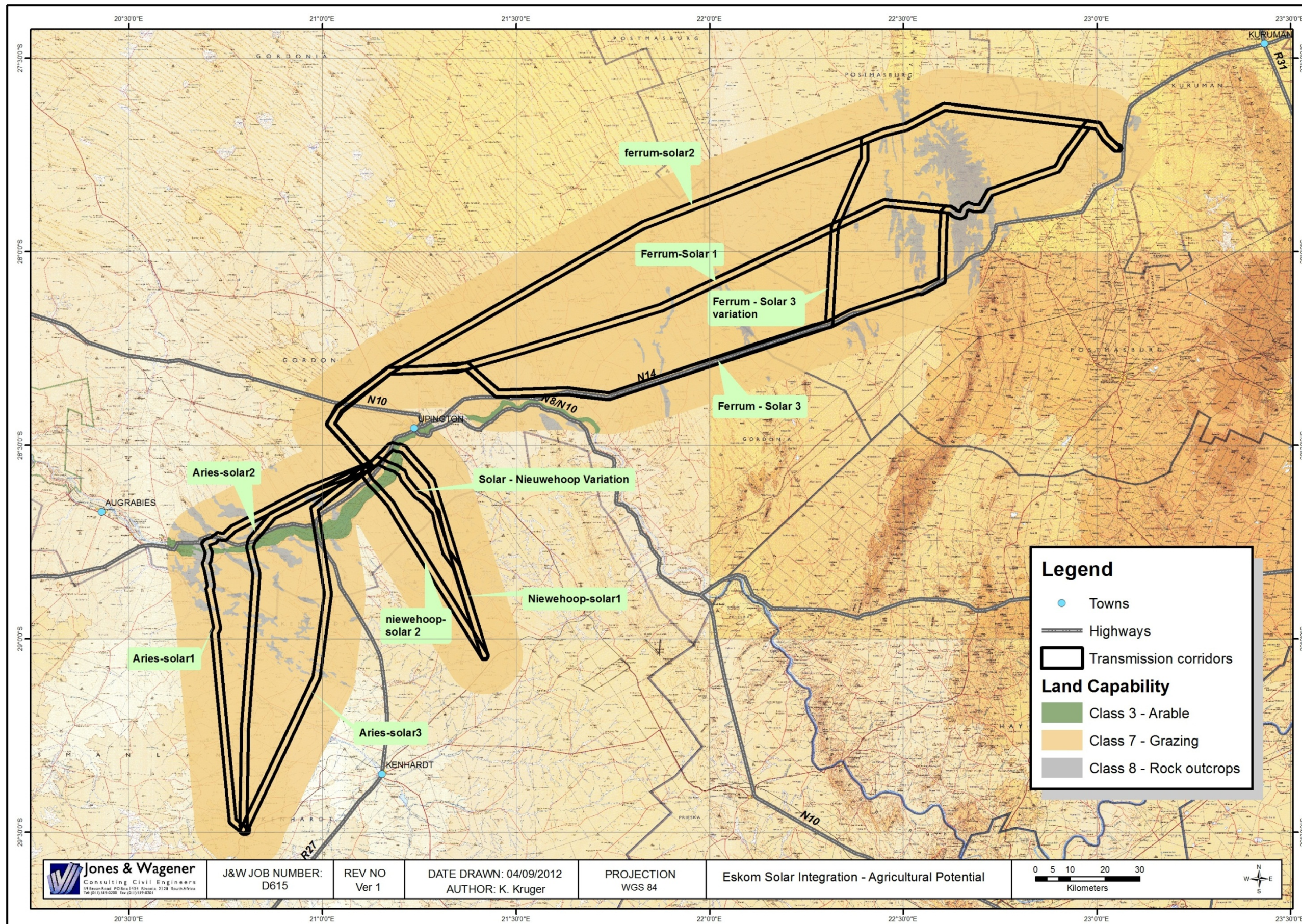


Figure 18: Agricultural potential for the study area

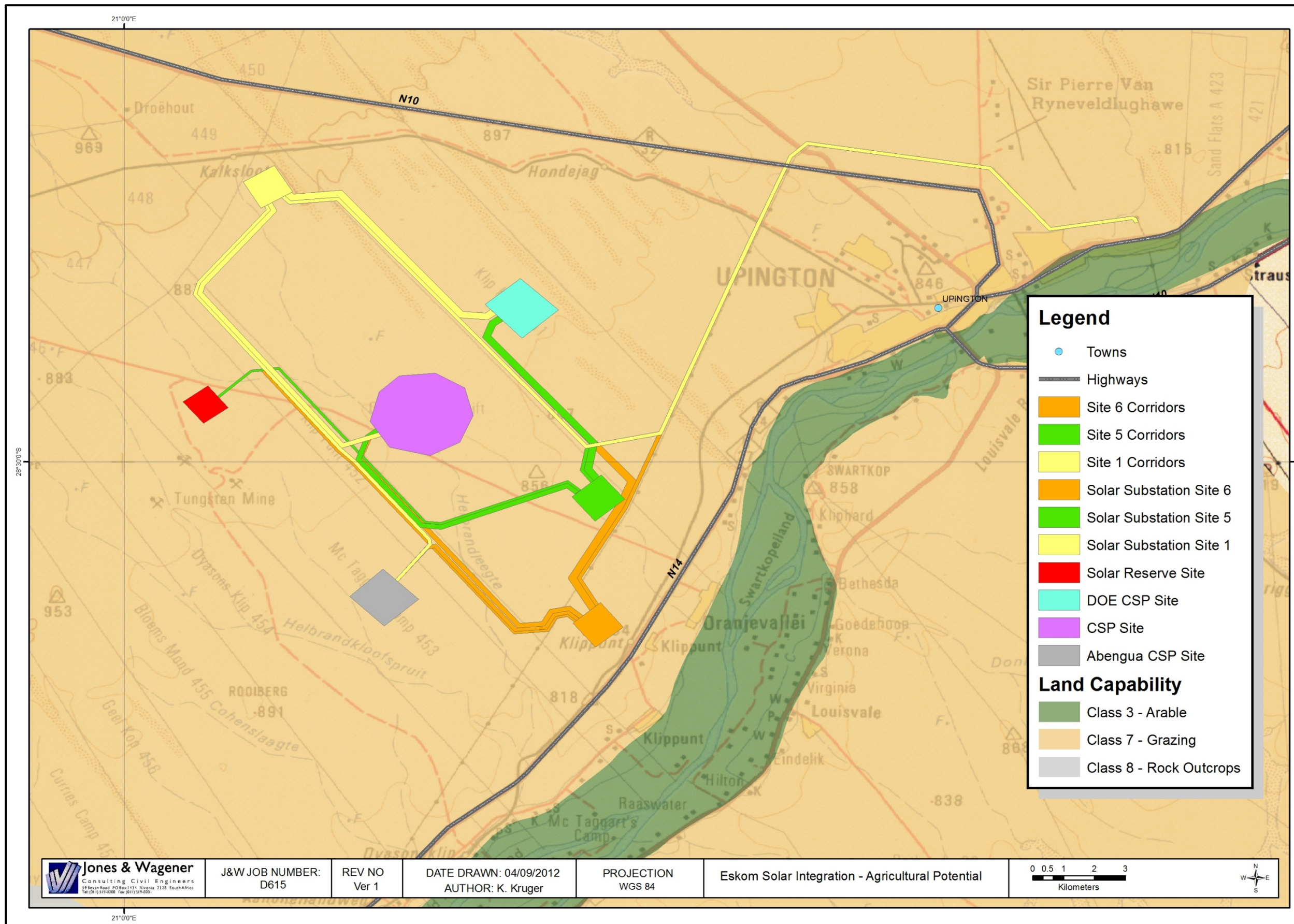


Figure 19: Agricultural Potential for the Distribution Lines

3.6 Terrestrial Biodiversity

3.6.1 Data Collection and Methodology

A literature review of the faunal and floral species that could occur in the area was conducted. The flora and fauna descriptions and data below are taken from *The Vegetation of South Africa, Lesotho and Swaziland* (Mucina and Rutherford 2006). Biodiversity data was obtained from the BGIS website for the Northern Cape provincial department and was used to conduct a desktop study of the area. This data consists of terrestrial components; ratings provide an indication as to the importance of the area with respect to biodiversity. Species information was obtained from the SIBIS website.

The detailed study involved extensive fieldwork, a literature review and a desktop study utilizing GIS. Site investigations were conducted from October 2011 to September 2012, from spring to summer. The area within the servitude was sampled using transects placed at 500 m intervals. At random points along these transect an area of 20 m x 20 m was surveyed. All species within the 20 m x 20 m quadrant were identified, photographed and their occurrence noted. Sensitive features such as ridges or wetlands were sampled by walking randomly through the area concerned and identifying all species within the area.

In addition to the references mentioned above, the following field guides were used:

- Guide to Grasses of Southern Africa (Frits van Oudtshoorn, 1999);
- Field Guide to Trees of Southern Africa (Braam van Wyk and Piet van Wyk, 1997);
- Field Guide to the Wild Flowers of the Highveld (Braam van Wyk and Sasa Malan, 1998);
- Problem Plants of South Africa (Clive Bromilow, 2001); and
- Medicinal Plants of South Africa (Ben-Erik van Wyk, Bosch van Oudtshoorn and Nigel Gericke, 2002)

Species lists were obtained from the SIBIS (*South African National Biodiversity Institute - Accessed through the SIBIS portal, sibus.sanbi.org, 2012-08-25*). In addition the following faunal guides were used on site and while compiling this report:

- Die Natuurlewe van Suider-Afrika, 'n veldgids tot diere en plante van die streek (Vincent Carruthers, 1997);
- Birds of Southern Africa (Ian Sinclair, 1994);
- Smithers' Mammals of Southern Africa, a field guide (Ed. Peter Apps, 2000);
- Sasol Owls and Owling in Southern Africa (Warwick Tarboton & Rudi Erasmus, 1998);
- Bats of Southern Africa (Peter John Taylor, 2000);

3.6.2 Regional Description

Nama-Karoo Biome

The Nama-Karoo Biome overlaps the main part of the study area and is a large landlocked biome in the central plateau of the western part of the country. The name is derived from the Khoi San word meaning "dry" and only the desert biome has higher variability in rainfall and the Kalahari greater extremes in temperature.

The flora in this biome is not particularly rich, and also has very low species endemism. *Asteraceae* (Asters), *Fabaceae* (Thorn Trees) and *Poaceae* (Grasses) are the dominant families found in the biome. The biome is a complex of extensive plains dominated by dwarf shrubs (< 1m tall) intermixed with grasses, succulents, geophytes and annual forbs. Small trees are limited to drainage lines or rocky outcrops. According to Mucina and Rutherford, the following vegetation types are found within the study area and this biome:

- Bushmanland Arid Grassland;
- Bushmanland Basin Shrubland;
- Kalahari Karroid Shrubland; and
- Lower Gariep Broken Veld.

Savanna Biome

Most Savanna has an herbaceous layer usually dominated by grass species and a discontinuous to sometimes very open tree layer. This is the most widespread biome in Africa. The savannah biome is found along the sandy dunefields to the north and east of Upington. Here the deeper soils allow for larger trees to establish themselves, especially Acacias with the intermittent shrubland in the areas between the dunes. Further to the east the topography and rainfall allows even bigger trees to establish themselves, especially around Kathu. Vegetation types found in this biome within the study area are:

- Gordonia Duneveld;
- Gordonia Plains Shrubland;
- Kathu Bushveld;
- Olifantshoek Plains Thornveld; and
- Koranna-Langeberge Mountain Bushveld.

Inland Azonal Vegetation

Also found in the study area is azonal vegetation, which is almost always associated with water bodies or wetlands. Within the study area the Orange River is the only perennial water source and the vegetation along its banks for a unique vegetation type i.e. the Lower Gariep Alluvial Vegetation. The flat alluvial terraces next to the river support a complex of riparian vegetation that will be assessed in detail during the EIA phase.

In addition the salt pans in the area is also recognised as a separate vegetation type known as the Southern Kalahari Salt Pans. These areas are generally devoid of vegetation but some specialist plants do survive here.

All the vegetation types mentioned above is illustrated in the maps below.

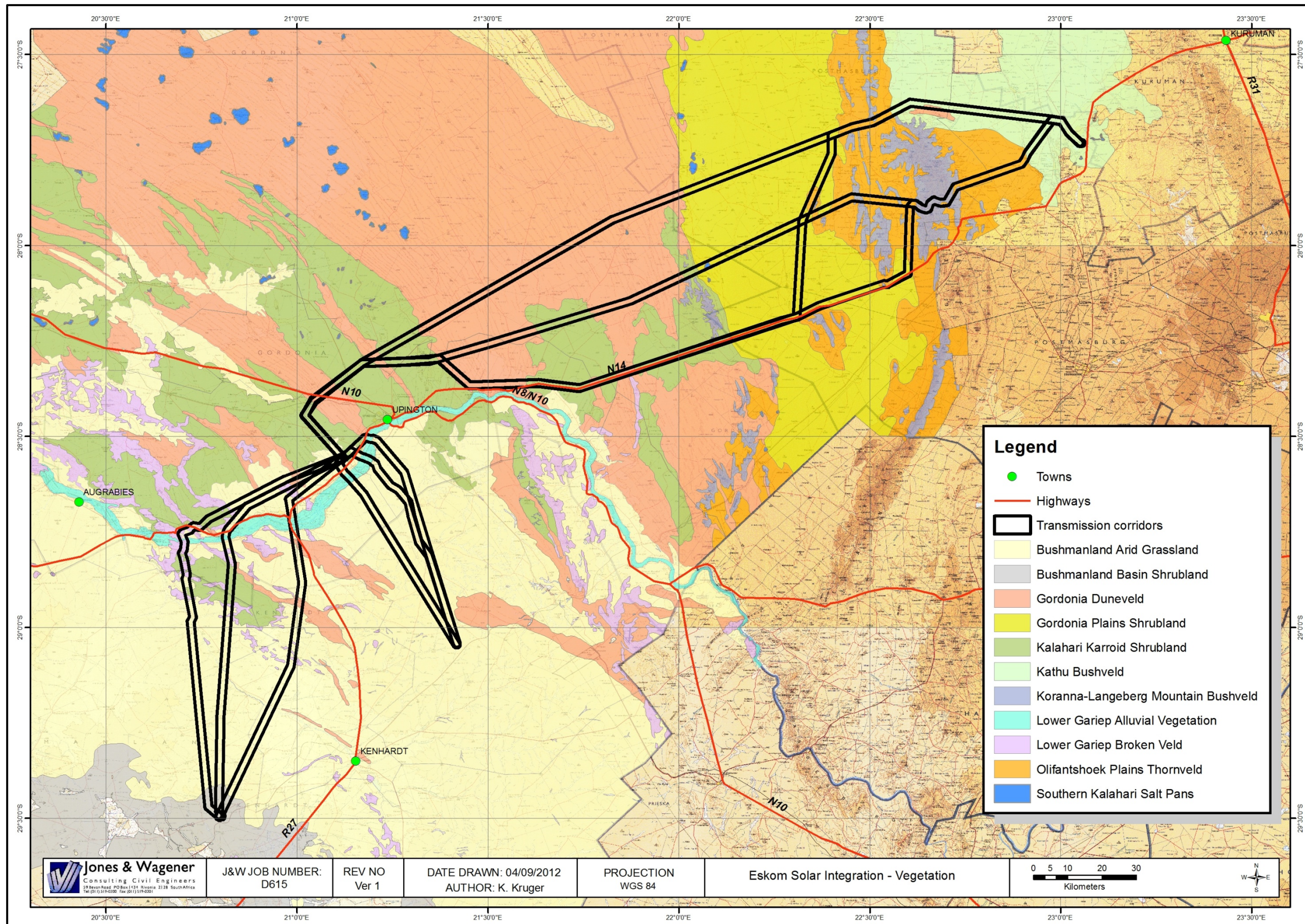


Figure 20: Vegetation of the study area

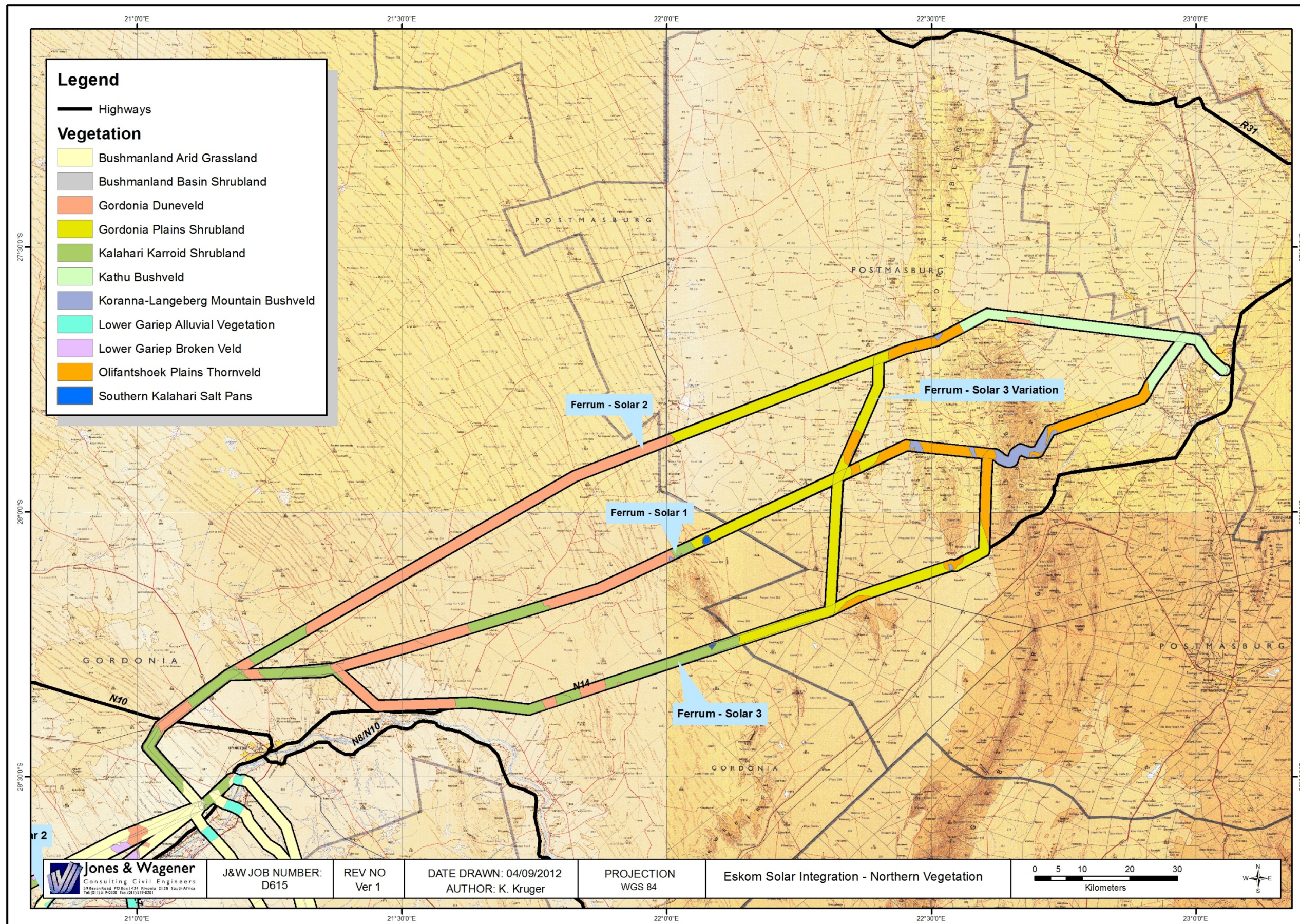


Figure 21: Vegetation of the Ferrum Routes

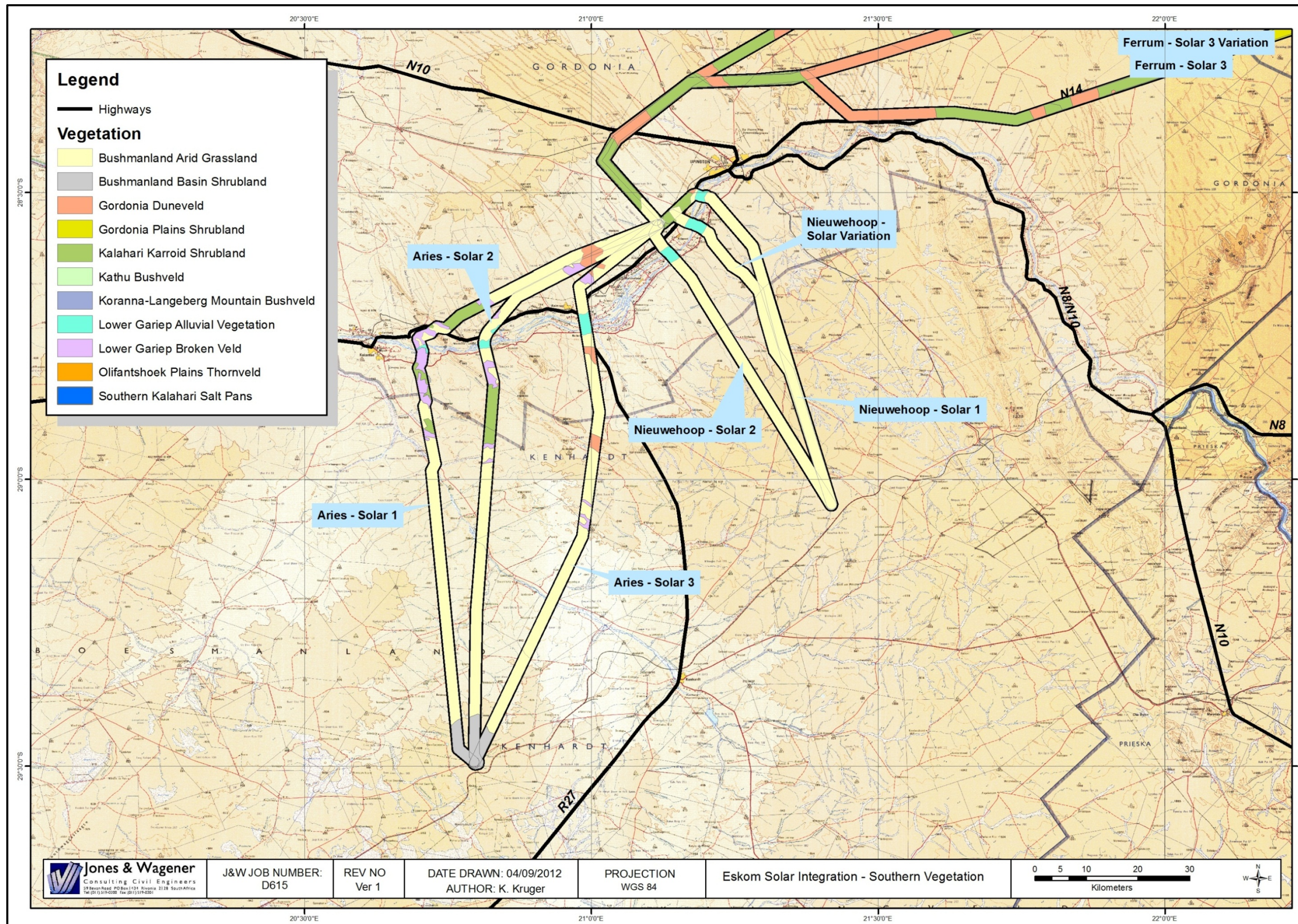


Figure 22: Vegetation of the southern section of the study area

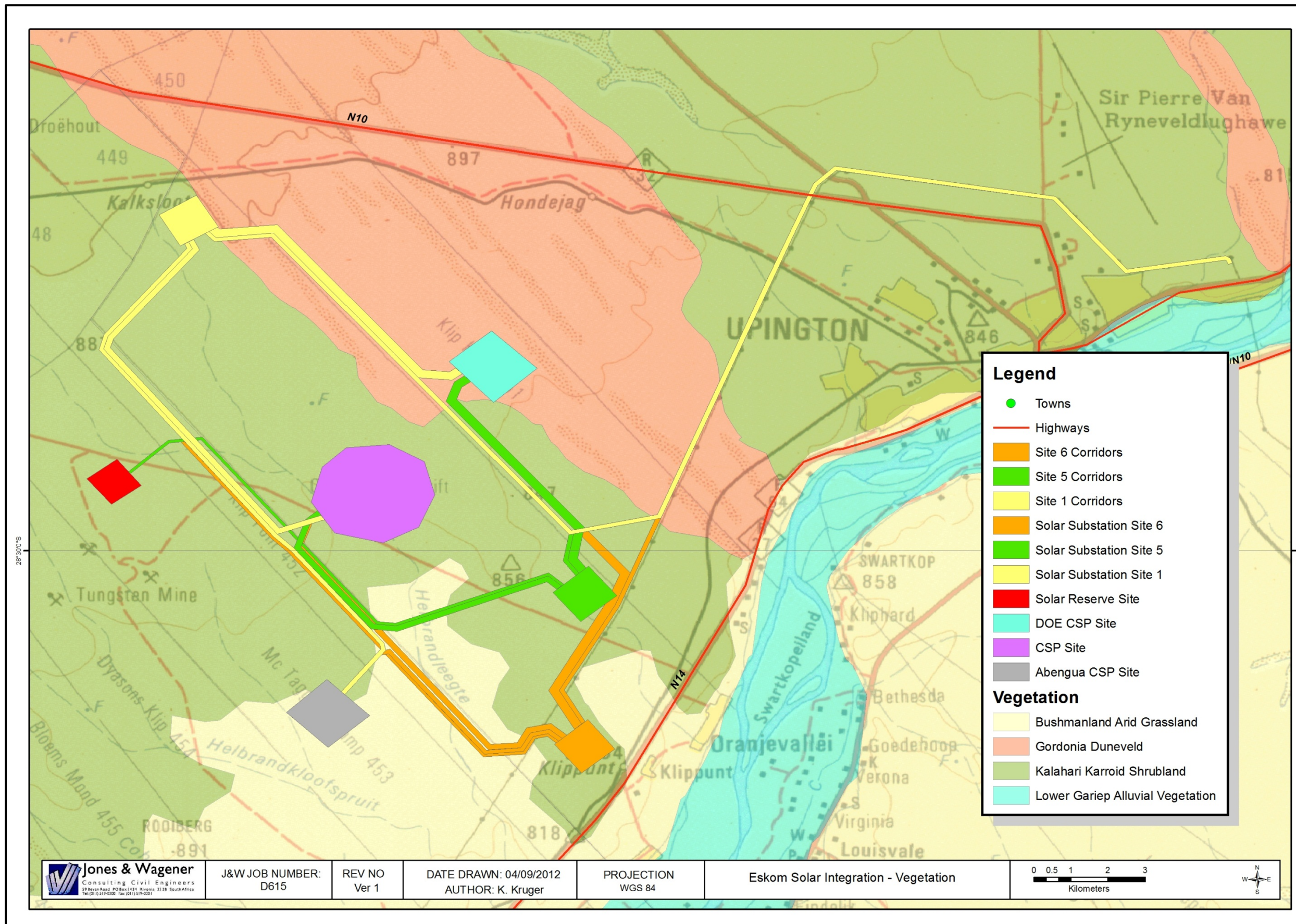


Figure 23: Vegetation for the Distribution Lines

3.6.3 Site description

In this section each of the vegetation/habitat types identified is described in more detail. This description starts at the easternmost section of the study area, at Kathu.

3.6.3.1. Ferrum to Solar Park Routes

Kathu Bushveld

This vegetation unit is found all around the Kathu area as the name suggests. The vegetation unit is typified by a medium-tall tree layer with *Acacia erioloba* in places, but the unit is mostly open with *Boscia albitrunca* as the other prominent tree. The shrub layer is the most important and dominant shrubs include *Acacia mellifera*, *Diospyros lycioides* and *Lycium hirsutum* with a variable grass layer. Dominant grasses include *Aristida meridionalis*, *Brachiaria nigropedata*, *Centropodia glauca*, *Eragrostis lehmanniana*, *Schmidtia pappophoroides* and *Stipagrostis ciliate*. Below are photographs taken from the helicopter flight over the study area. This unit is not threatened as only 1% of the vegetation unit has been transformed through mining in Kathu.



Figure 24: Kathu Bushveld showing the vegetation as well as the impact of an existing power line

Olifantshoek Plains Thornveld

The plains surrounding the town of Olifantshoek are dominated by thornveld and this vegetation type extends to all the plains downslope of the Korannaberg, Langeberg as well as the Asbestos Mountains. Here a wide variety of thorny trees and shrubs form an open mosaic with sparse grasses. The dominant trees are *Acacia luderitzii*, *Boschia albitrunca* and *Rhus tenuinervis*. The odd large *Acacia erioloba* can occur and the dominant grasses are *Schmidtia pappophoroides* and *Stipagrostis uniplumis*. This vegetation unit is not threatened as only 1% has been disturbed.



Figure 25: Olifantshoek Plains Thornveld from the air (left) and ground level (right)

Koranna-Langeberg Mountain Bushveld

This vegetation unit is found all along the Koranna and Langeberg Mountains. These rugged slopes support open shrubland with moderate grass cover. Dominant shrubs and small trees include *Acacia melifera* and *Croton gratissimus*. The grasses are dominated by *Aristida diffusa* and *Eragrostis curvula* with *Sarcostemma viminalis* a common succulent climber. Virtually no transformation has taken place so this vegetation unit is not threatened. The photographs below give an illustration of the typical vegetation found in this unit.

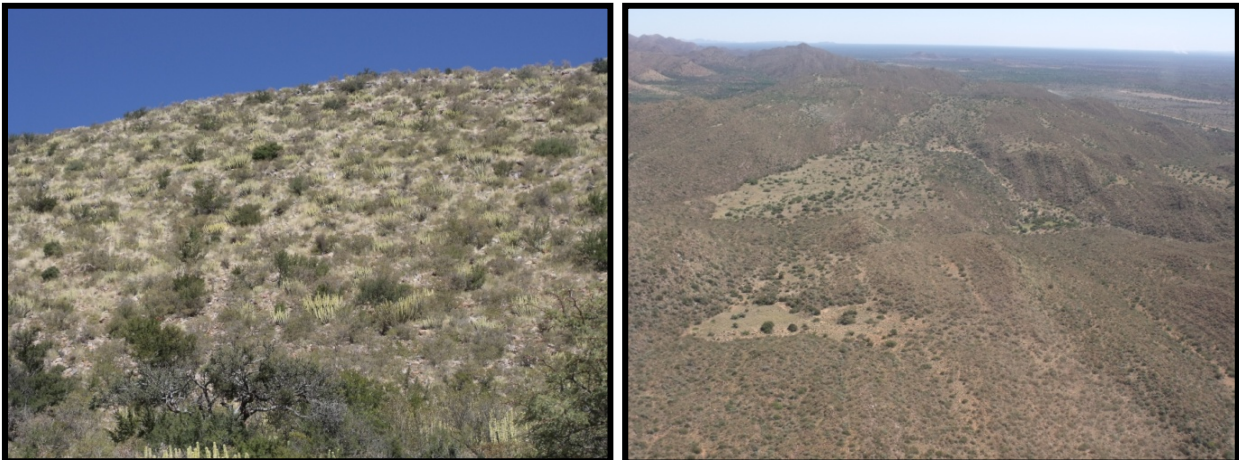


Figure 26: Koranna-Langeberg Mountain Bushveld

Gordonia Plains Shrubland

The Gordonia Plains Shrubland is found in a long band between the Kalahari dunes in the west and the Koranna and Langeberge in the east on the flat plains virtually devoid of dunes in between the two landscape features. These plains comprise of mainly open grassland with occasional shrubs *Rhigozum trichotomum*, *Grewia flava* and some scattered *Acacia haematoxylon* and *A. erioloba*. Dominant grasses include *Aristida meridionalis*, *Centropodia glauca*, *Eragrostis lehmanniana* and *Schmidtia kalahariensis*. Very little of this area has been disturbed and the vegetation type is not threatened. Please refer to some photographs of the typical vegetation within this unit below.



Figure 27: Gordonia Plains Shrubland

Southern Kalahari Salt Pans

The North West and Northern Cape Provinces house a number of intermittent endorheic, closed depressions (pans). These pans are vegetated by low grasslands although the centre of the pans is usually devoid of vegetation. The grasses are often dominated by *Sporobolus spp.* with a mixture of dwarf shrubs with an outer belt of *Lycium and/or Rhigozum*. Other species also include the succulent shrub *Zygophyllum tenue* and the grass *Enneapogon desvauxii*. This vegetation unit is subject to natural degradation – regeneration cycles controlled by the grazing of animals on the vegetation. In addition this vegetation unit is not threatened.



Figure 28: The salt pan found along the Ferrum-Solar 1 Route

Gordonia Duneveld

This vegetation unit covers a large expanse in the northern parts of the Northern Cape Province and is typified by the red Kalahari dunes. Several small pockets of dunes can be found scattered south of the Orange River. The dunes are parallel and about 3 – 8m above the plains. The vegetation comprises of open shrubland with *Stipagrostis amabilis* grasses dominating the dune crests, *Acacia haematoxylon* and *Acacia mellifera* trees on the slopes and *Rhigozum trichotomum* in the interdune “streets”. Other common species include *Grewia flava* shrubs, *Schmidtia kalahariensis* grasses and *Hermbsstaedtia fleckii* herbs. Area sensitive to overgrazing as removal of

vegetative cover can result in mobilisation of dune sands. This vegetation unit is well conserved and is not threatened.



Figure 29: Gordonia Duneveld showing the typical red dunes

Kalahari Karroid Shrubland

The Kalahari Karroid Shrubland forms alternating bands with the Gordonia Duneveld and usually occurs in the areas where the dunes do not occur. This vegetation type forms the transition between the Savanna biome and the Nama-Karoo biome as the tree elements reduce and shrubs and grasses start to dominate. Small trees and shrubs include *Acacia mellifera*, *Parkinsonia africana*, and *Rhigozum trichotomum*. Low shrubs dominate the area and include *Hermannia spinosa*, *Limeum aethiopicum* and *Phaeoptilum spinosum* while the common herbs include *Dicoma capensis*, *Chamaesyce inaequilatera*. Common grasses are *Aristida adscensionis*, *Enneapogon desvauxii*, *E. scaber* and *Stipagrostis obtusa*. This vegetation unit is not threatened although this area was the route of choice for early roads, which lead to the introduction of alien plants. The result is that some 25% of the unit has been colonised by scattered *Prosopis* species.



Figure 30: Kalahari Karroid Shrubland

3.6.3.2. Aries and Nieuwehoop to Solar Routes

This following section covers the vegetation found from Upington, to Kakamas and south to Kenhardt that is traversed by die routes from Aries to Solar Park as well as the routes from Nieuwehoop to the Solar Park.

Bushmanland Arid Grassland

This large vegetation unit comprises the grasslands between the shrublands to the north and east, the desert landscapes to the northwest and Namaqualand hills in the west. These extensive plains are dominated by white grasses mostly of the *Stipagrostis* genus giving the vegetation a semi-desert steppe character. In some low lying places the *Sasola* shrubs change the vegetation structure and in years of high rainfall a rich display of annual herbs and their flowers can be expected. Dominant grasses include *Aristida adscensionis*, *A. Congesta*, *Enneapogon desvauxii*, *Eragrostis nindensis*, *Schmidtia kalahariensis*, *Stipagrostis ciliate*, *S. obtusa* and *Cenchrus ciliaris*. Shrubs include *Lycium cinereum*, *Rhigozum trichotomum*, *Aptosimum spinescens*, *Hermannia spinosa* and *Pentzia spinescens*. Very little of this vegetation unit has been disturbed and hence the unit is not threatened.



Figure 31: Bushmanland Arid Grassland

Lower Gariep Broken Veld

This vegetation unit is found along the broken koppies and inselbergs around Keimoes and just before Kakamas as well as a few outcrops to the south. This rugged terrain is sparsely populated with vegetation that is dominated by shrubs with annuals present in spring in the form of perennial grasses and herbs. Dominant trees include *Aloe dichotoma* and *Acacia mellifera* with the dominant shrubs *Rhigozum trichotomum*, *Blepharis mitrata*. The dominant grasses include *Aristida adscensionis*, *Enneapogon desvauxii*, *E. scaber*, *Eragrostis nindensis*, *Stipagrostis obtusa*, *S. uniplumis*. The main dominant herb is *Forsskaolea candida*. This vegetation unit is also not threatened as there is less than 1% transformed. Below are photographs of the vegetation unit.



Figure 32: Lower Gariep Broken Veld showing protected “kokerboom” on the left

Lower Gariep Alluvial Vegetation

The Lower Gariep Alluvial Vegetation is found all along the alluvial floodplains and islands of the Orange River from Groblershoop to the Atlantic Ocean. These alluvial terraces support a variety of riparian thickets dominated by *Ziziphus mucronata*, *Euclea pseudebenus* and *Tamarix usneoides* along with reed beds with *Phragmites australis*. These are mixed with flooded grasslands and herblands on the terraces and banks of the river. Additional species in the riparian vegetation includes the trees and shrubs *Acacia karroo*, *Salix mucronata*, *Schotia afra* and *Gymnosporia linearis*. The grasslands and herblands include species such as *Tetragonia schenkii*, *Litogyne gariepina*, *Cynodon dactylon* and *Setaria verticillata*. This vegetation type has been extensively modified (>50% transformed) through agriculture (grapes and vegetables) as well as alluvial diamond mining. In addition this vegetation type is prone to invasion by *Nicotiana glauca* and *Argemone ochroleuca*. This vegetation type is therefore listed as endangered.



Figure 33: Lower Gariep Alluvial Vegetation, showing the encroachment from agriculture (left)

Bushmanland Basin Shrubland

The Bushmanland Basin Shrubland is found at the very southern extremities of the study area around the Aries substation. This is the northernmost part of a large basin centred around Brandvlei and Vanwyksvlei. These slightly irregular plains are dominated by a mixture of dwarf shrubs and “white” grasses and in years of high rainfall a number of annuals are also abundant. The dominant shrubs are *Lycium cinerum*, *Rhigozum trichotomum*, *Aptosimum spinescens*, *Hermannia spinosa*, *Pentzia spinescens*, *Zygopyllum micophyllum* and *Salsola tuberculata*. The dominant herbs and grasses are *Ganazia lichtensteinii*, *Leysera tenella*, *Aristida adscensionis*, *Enneapogon desvauxii*, *Stipagrostis obtuse* and *S. ciliate*. This vegetation unit is relatively undisturbed and not threatened.



Figure 34: Bushmanland Basin Shrubland surrounding Aries substation

3.6.3.3. *Distribution Routes*

The distribution lines cross over three main vegetation types including Bushmanland Arid Grassland, Kalahari Karroid Shrubland and Gordonia Duneveld as shown in Figure 23. These vegetation types have been described above.

3.6.4 Fauna

The habitats described above form the home for a variety of species and detailed lists of these are provided in Appendix A. In general the grasslands and shrub plains described above house species that can withstand the arid climate. Common species include the following:

- Mammals;
 - Bat-eared foxes;
 - Steenbok;
 - Scrub hare;
 - Springbok;
 - Aardvark;
 - Meerkat; and
 - Mongoose (variety).
- Reptiles
 - Puff adder; and
 - Leopard tortoise.

Avifauna has been specifically left out as that was a separate specialist study. In total an estimated 23 mammal, 17 reptile and 39 Arthropods as listed in the appendix.

3.6.5 Sensitivities

3.6.5.1. *Endangered Ecosystems*

Using data from SANBI on the protected and threatened ecosystems found in the study area Figure 35 was generated. The provincial data highlights Critical Biodiversity Areas (CBA's) as shown in yellow on the map. It also highlights biodiversity corridors

as shown in green on the map. Lastly the remaining pockets of endangered vegetation inside the CBA are shown in red.

From the map it can be seen that the Lower Gariep Alluvial vegetation adjacent to the Orange River is classified as a threatened ecosystem. Impacting this area requires approval as per the NEMA Listing 3 Regulations and NEM:BA. When evaluating the most preferred crossing of the Orange River and its environs, the threatened status of this vegetation unit should be considered as a critical factor in the evaluation.

3.6.5.2. Endangered Species

Further to the endangered ecosystem there is the consideration of protected and endangered species. In terms of the National Environmental Management: Biodiversity Act (NEM: BA, Act 10 of 2004) and the IUCN website the study area could contain the following endangered species:

- **Aloe pillansii** (Bastard Quiver Tree)
 - Status: Critically Endangered
- **Aloe ramosissima** (Maiden's Quiver Tree)
 - Status: Vulnerable
- **Mystromys albicaudatus** (White-tailed Mouse)
 - Status: Endangered
- **Pachypodium namaquanum** (Elephant's Trunk)
 - Status: Lower Risk/near threatened
- **Manis temminckii** (Pangolin)
 - Status: Vulnerable
- **Panthera pardus** (Leopard)
 - Status: Vulnerable

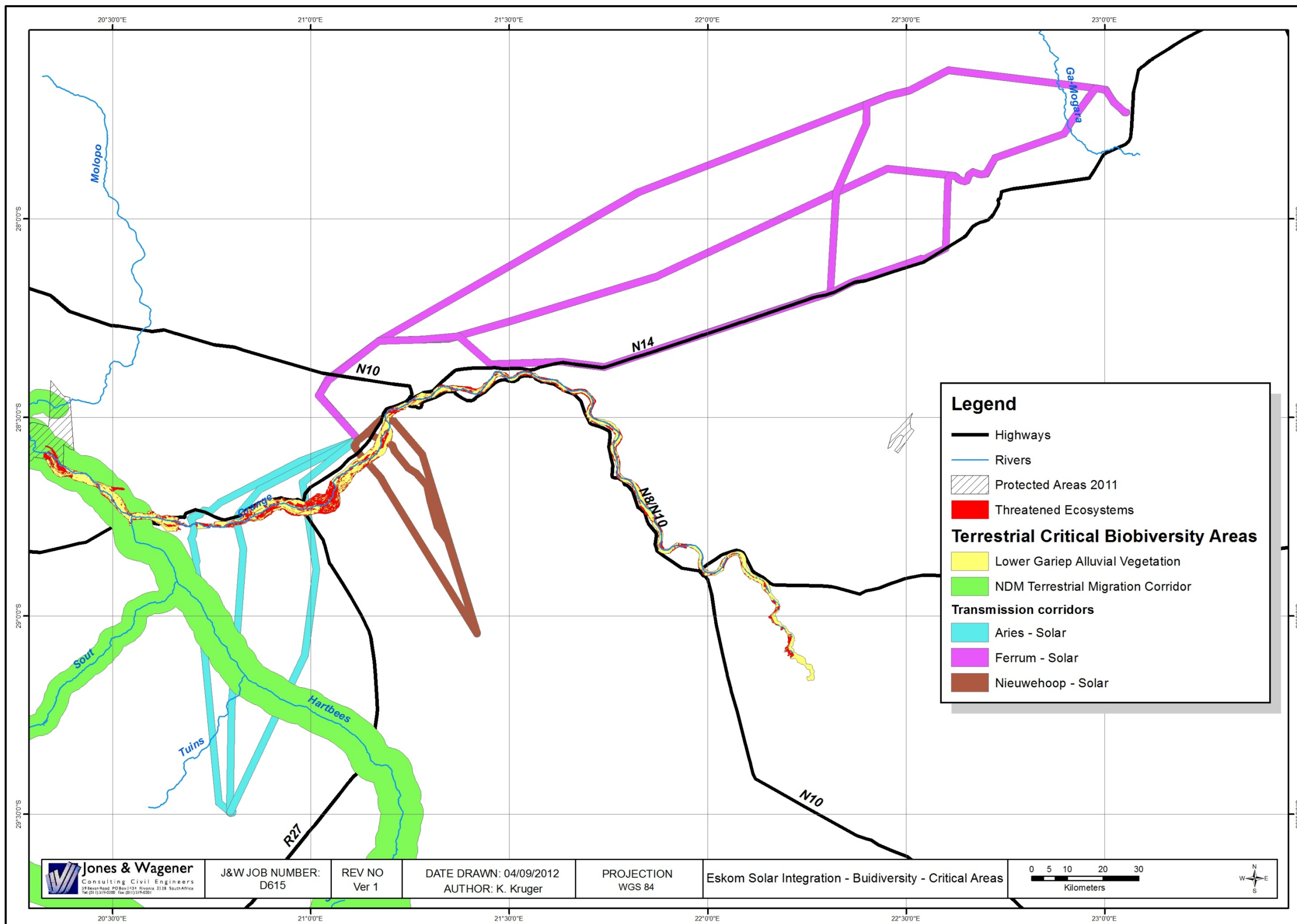


Figure 35: Endangered habitat as well as Critical Biodiversity Areas (CBA's)

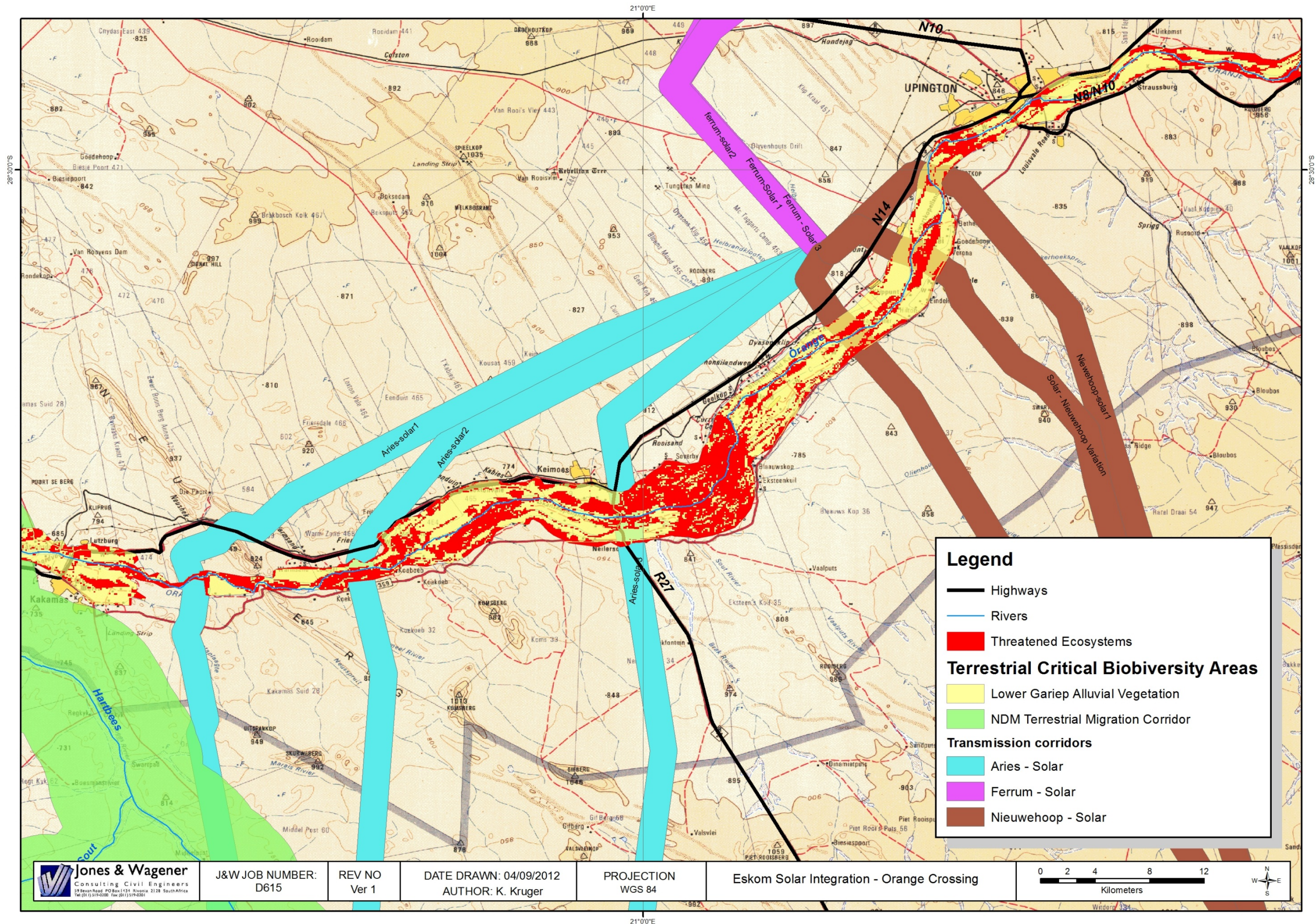


Figure 36: Zoomed in view of endangered habitat crossing

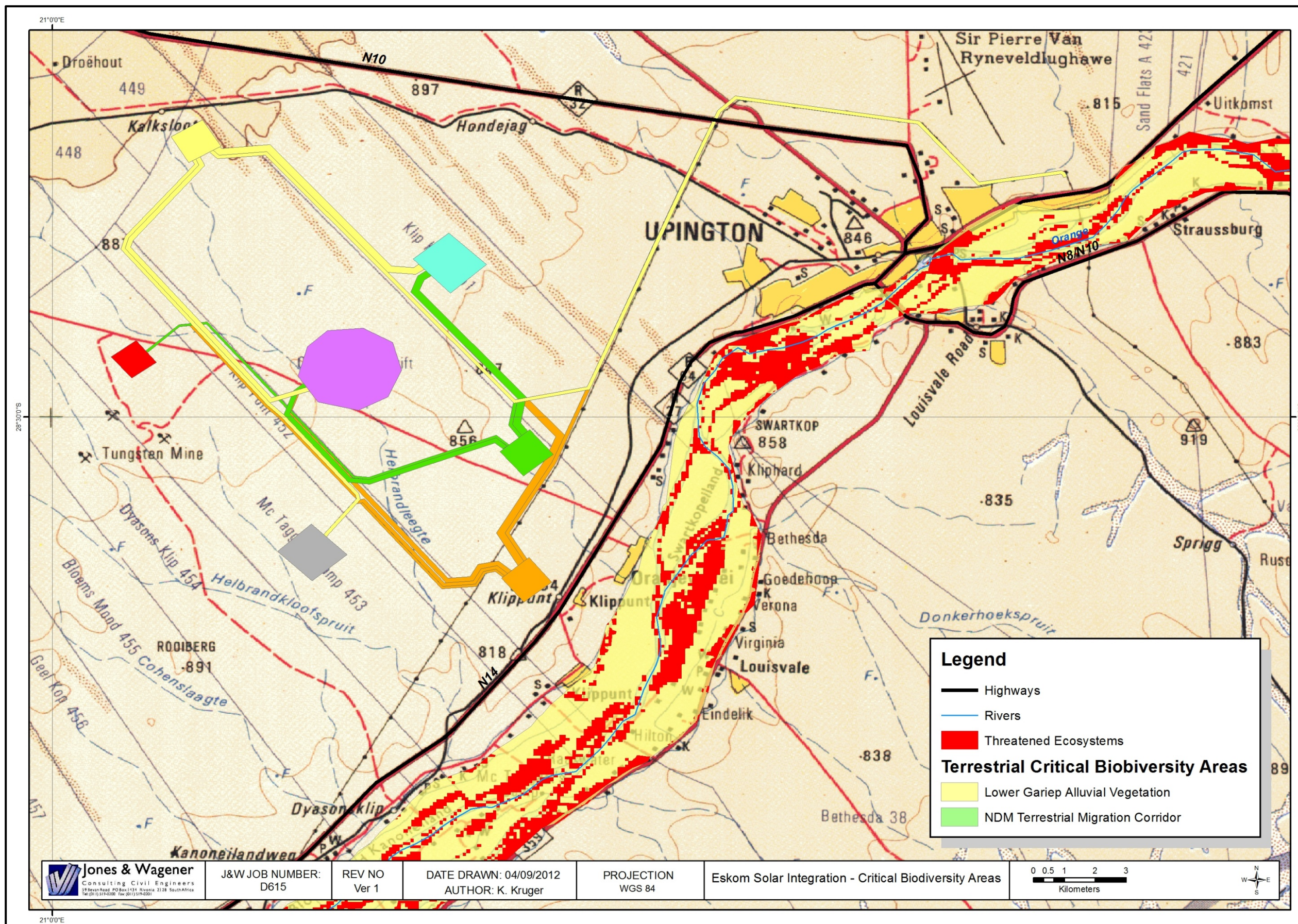


Figure 37: CBA's in relation to the distribution lines

3.6.5.3. Protected Species

In addition to the NEM: BA regulations, the Department of Agriculture, Forestry and Fisheries also have a list of protected trees that require a license to remove, crop or disturb prior to the activity. These trees are listed in terms of Section 15(1) of the National Forests Act, 1998, as amended. It should be noted that an EIA authorisation does not exempt the applicant from the NFA requirements.

The Environmental Management Plan (EMP) has a section that details how these requirements should be met. The species that could occur in the study area include (descriptions adapted from the South African National Biodiversity Institute's plant information website www.plantzafrica.com):

***Acacia erioloba* aka Camel Thorn, Kameeldoring / Mogohlo (NS) / Mogôthô (T)**

This large Acacia is found throughout the drier parts of southern Africa. It frequently occurs in areas of deeper sandy soils and groundwater, often found along dry river beds. The area around Kathu is especially rich in these trees and they occur sporadically throughout the study area. These trees can become quite large and range from a 2m spiny shrub to a 16m robust tree as shown below. Due to the potential height of these trees it is anticipated that they might require removal or pruning prior to construction of the power lines – applicant to ensure that the license is obtained from DAFF prior to the start of construction.

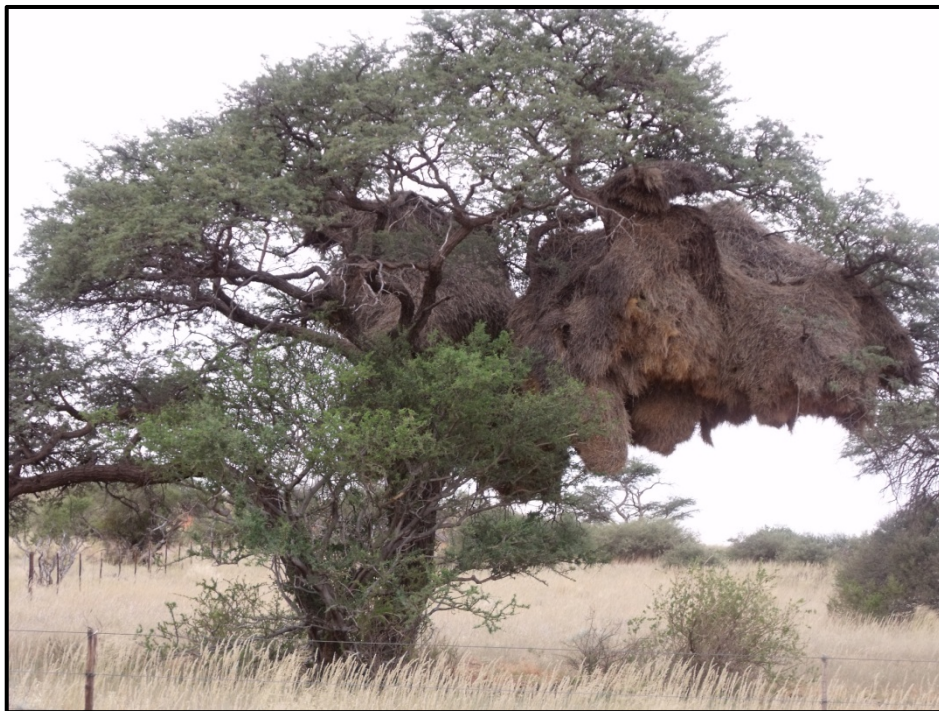


Figure 38: *Acacia erioloba*

***Acacia haematoxylon* aka Grey Camel Thorn, Vaalkameeldoring (A) / Mokholo (T)**

A shrub to medium-sized tree, 1.5 – 6m tall with an irregular crown. These trees are characteristic of the semi-desert and desert areas in South Africa. They occur on deep sandy soils and dunes as a shrub and larger specimens are found along drainage lines. These trees although similar in name to the larger Camel Thorn, are significantly smaller, with finer leaves of grey colour. The photo below was taken on site and shown

a Grey Camel Thorn in the foreground and a normal Camel Thorn in the back for comparison.



Figure 39: *Acacia haematoxylon* (foreground) and *A. erioloba* (background)

***Boscia albitrunca* aka Shepherd's tree, Witgat (A) / Mohlôpi (NS) / Motlhôpi (T) / Muvhombwe (V) / Umgqomogqomo (X) / Umvithi (Z)**

The Shepard's tree is the one tree in the Kalahari that does not shed its leaves, and hence provides a shady spot for animals and humans (hence the name). This small evergreen tree is characterised by an umbrella-shaped much branched crown and smooth white to grey bark. It is widespread throughout the study area covering almost all habitats. A photo of the tree is given below.



Figure 40: Shepard's Tree

3.7 Surface Water and Wetlands

3.7.1 Data Collection and Methodology

The surface water data was obtained from the National Freshwater Ecosystem Priority Area's (NFEPA) database from SANBI (2011). The data used included catchments, river alignments and river names. This information will be ground truthed during the specialist investigation.

3.7.1.1. Riparian Zones vs. Wetlands

Wetlands

The riparian zone and wetlands were delineated according to the Department of Water Affairs (DWA, previously known as the Department of Water Affairs and Forestry - DWAF) guideline, 2003: A practical guideline procedure for the identification and delineation of wetlands and riparian zones. According to the DWA guidelines a *wetland* is defined by the National Water Act as:

“land which is transitional between terrestrial and aquatic systems where the water table is usually at or near surface, or the land is periodically covered with shallow water, and which land in normal circumstances supports or would support vegetation typically adapted to life in saturated soil.”

In addition the guidelines indicate that wetlands must have one or more of the following attributes:

- Wetland (hydromorphic) soils that display characteristics resulting from prolonged saturation;
- The presence, at least occasionally, of water loving plants (hydrophytes); and
- A high water table that results in saturation at or near surface, leading to anaerobic conditions developing in the top 50 centimetres of the soil.

During the site investigation the following indicators of potential wetlands were identified:

- Terrain unit indicator;
- Soil form indicator;
- Soil wetness indicator; and
- Vegetation indicator.

Riparian Areas

According to the DWA guidelines a *riparian area* is defined by the National Water Act as:

“Riparian habitat includes the physical structure and associated vegetation of the areas associated with a watercourse which are commonly characterised by alluvial soils, and which are inundated or flooded to an extent and with a frequency sufficient to support vegetation of species with a composition and physical structure distinct from those of adjacent land areas”

The difference between Riparian Areas and Wetlands

According to the DWA guidelines the difference between a wetland and a riparian area is:

“Many riparian areas display wetland indicators and should be classified as wetlands. However, other riparian areas are not saturated long enough or often enough to develop wetland characteristics, but also perform a number of important functions, which need to be safeguarded... Riparian areas commonly reflect the high-energy conditions associated with the water flowing in a water channel, whereas wetlands display more diffuse flow and are lower energy environments.”

Delineation

The site was investigated for the occurrence / presence of wetlands and riparian areas, using the methodology described above and described in more detail in the DWA guidelines.

3.7.2 Regional Description

The surface water features in the study area is dominated by the Orange River, which is the largest river in South Africa and also the only perennial river in the study area. All the alternatives have to cross the Orange River and it is anticipated that the bulk of the alignments of the alternative routes will be determined by this river crossing.

Smaller rivers that also have to be crossed include the Ga-Mogara, Hartbees and Kareeboom rivers and some of their associated tributaries depending on the alternative route selected. These rivers are all non-perennial and only flow after storm events. The surface water features in the study area is dominated by the Orange River, which is the largest river in South Africa and also the only perennial river in the study area.

3.7.3 Site description/delineation

The site was investigated for the occurrence / presence of wetlands and riparian areas, using the methodology described above and described in more detail in the DWA guidelines.

3.7.3.1. *Terrain Unit Indicator*

The terrain on site varies from 600 metres above mean sea level (mamsl) to 1 800 mamsl. Terrain units on site include crest, slope, valley and plains. According to the DWA guidelines the valley bottom is the terrain unit where wetlands/drainage lines are most likely to occur, but the occurrence of wetlands is not excluded from any of the other terrain units.

3.7.3.2. *Soil Form Indicator*

Of the various soils identified in Section 3.4 above the alluvial soils are the main soil form that is can be an indicator of wetlands or drainage areas.

3.7.3.3. *Soil Wetness Indicator*

The soils on site were subjected to a soil wetness assessment. If soils showed signs of wetness within 50 cm of the soil surface, it was classified as a hydromorphic soil and divided into the following groups:

Temporary Zone

- Minimal grey matrix (<10%);
- Few high chroma mottles; and

- Short periods of saturation.

Seasonal Zone

- Grey matrix (>10%);
- Many low chroma mottles present; and
- Significant periods of wetness (>3 months / annum).

Permanent Zone

- Prominent grey matrix;
- Few to no high chroma mottles;
- Wetness all year round; and
- Sulphuric odour.

The Orange River and its surrounding areas were the only water body that had wetness within the top 50cm of the soil profile. Due to the aridity of the region, none of the other drainage lines or river beds shows signs of wetness, as they are just not saturated long enough to develop these signs.

3.7.3.4. Vegetation Indicator

From the vegetation assessment two vegetation units identified indicate the potential presence of water bodies, pans or wetlands. These include the Lower Gariep Alluvial vegetation and the Southern Kalahari Salt Pans. The Lower Gariep Alluvial vegetation is situated around the permanent water of the Orange River, while the pans are local depressions that collect water in periods of high rainfall, however these periods are very erratic and could be decades apart.

3.7.3.5. Delineated surface water features

According to the methodology that was followed for delineation of wetlands by DWA, there are three main surface water features present on site. These include:

- Rivers;
- Drainage Lines; and
- Pans.

Figure 10 illustrates the surface water bodies identified. It should be noted that although the area has a few rivers identified, the only perennial river is the Orange River. The rest of the study area is very arid, and the bulk of the drainage features are drainage lines with sandy beds that can be identified by the concentration of vegetation in these areas. These areas do however not classify as wetlands as they have no signs of wetness within the top 50cm of the soil profile. Please refer to the photographs below for a view of the Orange River as well as the dry drainage lines found on site.



Figure 41: Surface Water features on site

3.7.3.6. Classification of water bodies

The classification of the water bodies in the study area into different types was based on the method as defined in the National Wetland Classification System for South Africa (Figure 42), developed by the Freshwater Consulting Group for South African National Biodiversity Institute and the Working for Water Group.

This classification system has 6 levels of classification that in the end of level 5 described the functional wetland/water unit. This identification of the functional unit was the aim of this assessment. The classification of the wetlands on site proceeded as follows:

- Level 1 – System – Inland Ecosystem;
- Level 2 – Bioregion – Nama Karroo / Southern Kalahari
- Level 3 – Landscape Setting
 - a) Slope;
 - b) Plain; and
 - c) Valley floor.
- Level 4 – Hydrogeomorphic unit
 - a) Channels; and

Channel (river, including the banks): an open conduit with clearly defined margins that (i) continuously or periodically contains flowing water, or (ii) forms a connecting link between two water bodies. Dominant water sources include concentrated surface flow from upstream channels and tributaries, diffuse surface flow or interflow, and/or groundwater flow. Water moves through the system as concentrated flow and usually exits as such but can exit as diffuse surface flow because of a sudden change in gradient. Unidirectional channel-contained horizontal flow characterises the hydrodynamic nature of these units. Note that, for purposes of the classification system, channels generally refer to rivers or streams (including those that have been canalised) that are subject to concentrated flow on a continuous basis or periodically during flooding, as opposed to being characterised by diffuse flow (see unchannelled

valley-bottom wetland). As a result of the erosive forces associated with concentrated flow, channels characteristically have relatively obvious active channel banks.

b) Depressions.

Depression: a landform with closed elevation contours that increases in depth from the perimeter to a central area of greatest depth, and within which water typically accumulates. Dominant water sources are precipitation, ground water discharge, interflow and (diffuse or concentrated) overland flow. For ‘depressions with channeled inflow’, concentrated overland flow is typically a major source of water for the wetland, whereas this is not the case for ‘depressions without channeled inflow’. Dominant hydrodynamics are (primarily seasonal) vertical fluctuations. Depressions may be flatbottomed (in which case they are often referred to as ‘pans’) or round-bottomed (in which case they are often referred to as ‘basins’), and may have any combination of inlets and outlets or lack them completely. For ‘exorheic depressions’, water exits as concentrated surface flow while, for ‘endorheic depressions’, water exits by means of evaporation and infiltration.

- Level 5 – Level on inundation
 - a) Orange River – Perennial
 - b) Drainage Lines - Non-perennial – never inundated, saturation unknown; and
 - c) Pans - Non-perennial - never inundated, saturation unknown.

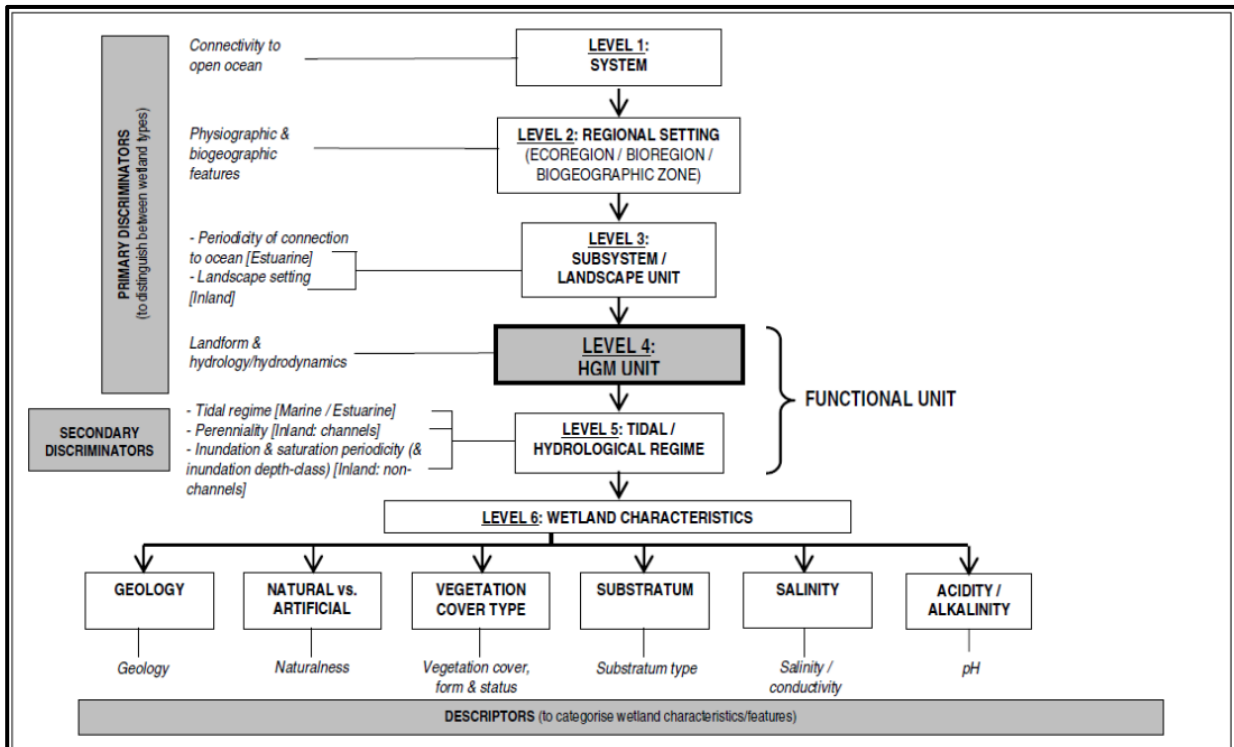


Figure 42: National Wetland classification system (SANBI, 2009)

Using the methodology above the following wetland types were identified on site as shown below in Figure 10:

- Orange River – Perennial River Channel
- Drainage Lines (channels) - Non-perennial – never inundated, saturation unknown; and
- Pans - Non-perennial - never inundated, saturation unknown.

3.7.4 Sensitivities

In the arid region of the Northern Cape, all water bodies are seen as highly sensitive and important features. The Orange River is the lifeline in this region and impacts to the river should be avoided as far as possible. Furthermore the drainage lines and pans are features that only hold or transport water in the unlikely event of a rainfall event. These features should also be avoided.

The maps below illustrate the water features identified and also provide a zoomed in view of the potential crossings over the Orange River.

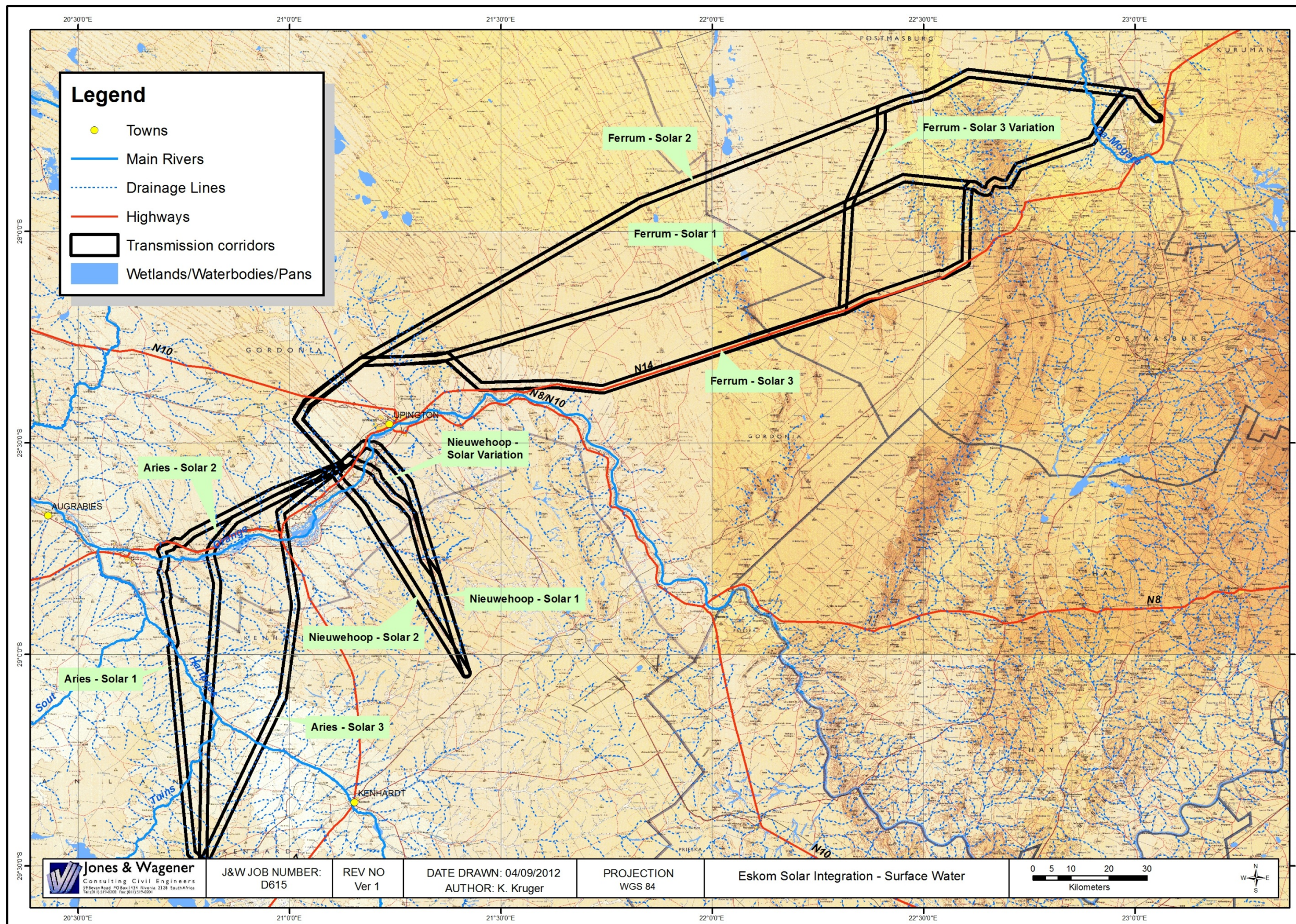


Figure 43: Surface Water Features

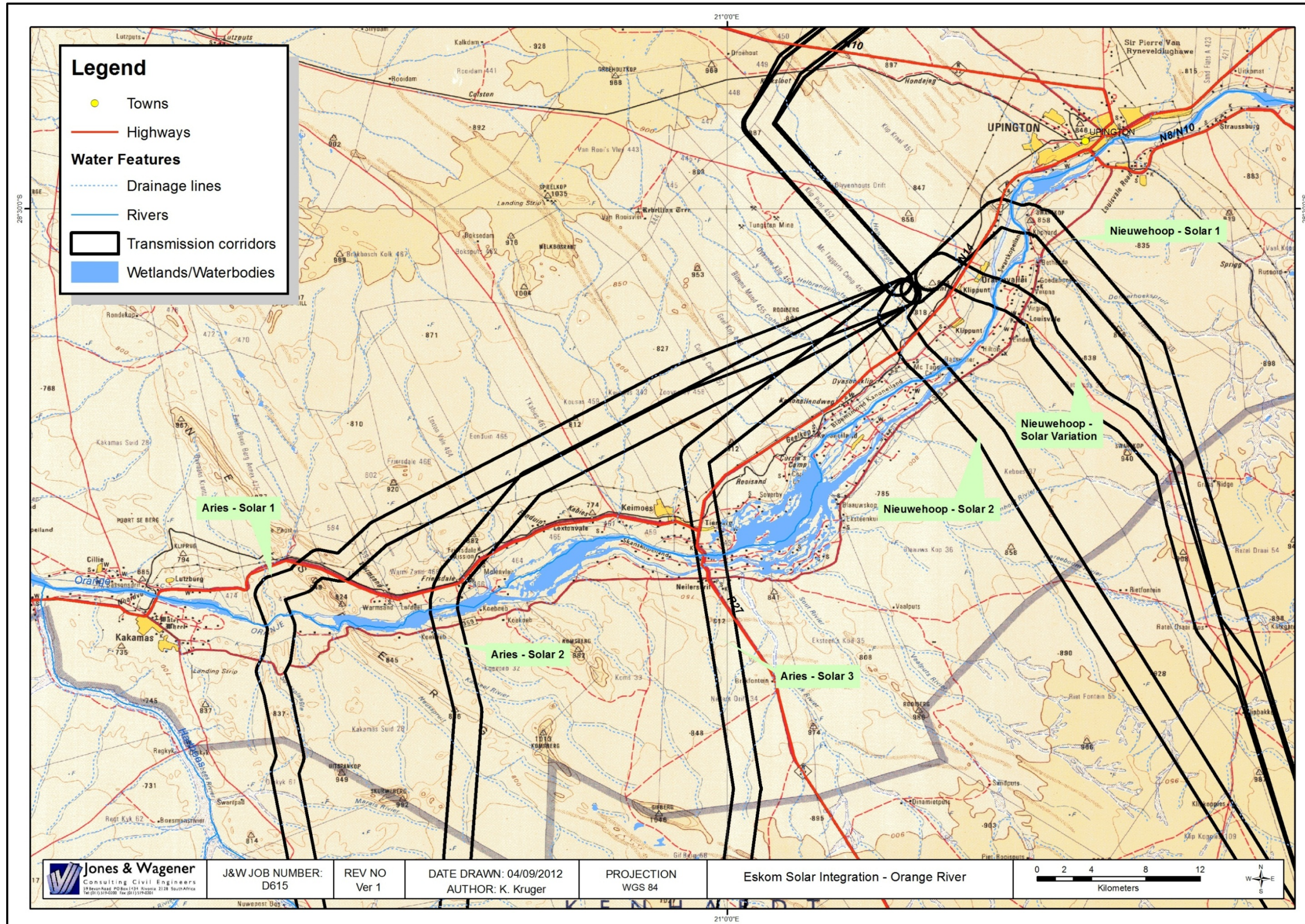


Figure 44: Crossing the Orange River

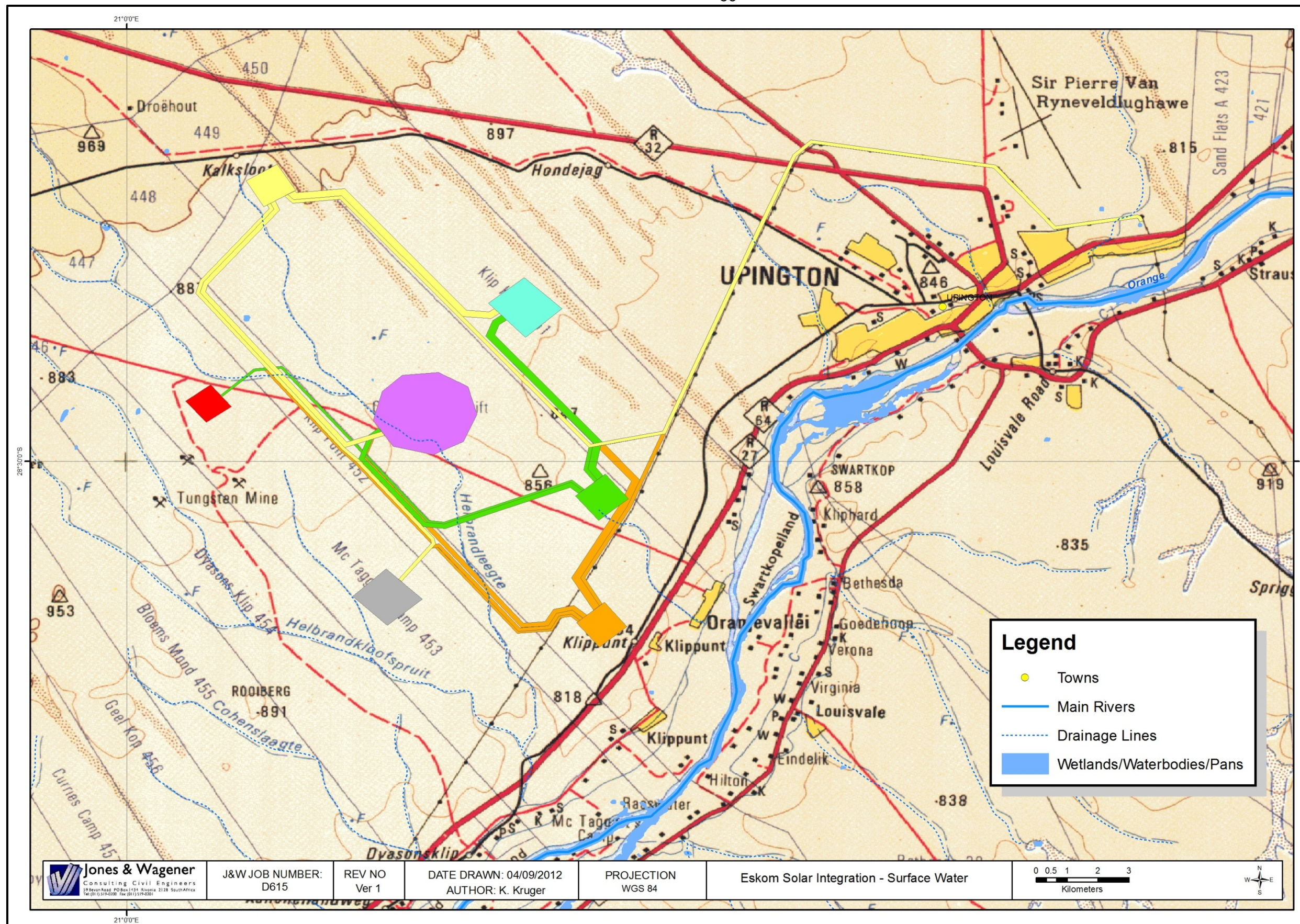


Figure 45: Distribution Routes Surface Water Map



3.8 Aquatic Ecology

To be completed with report from SAS.

4. IMPACT ASSESSMENT METHODOLOGY

In order to ensure uniformity, a standard impact assessment methodology has been utilised so that a wide range of impacts can be compared. The impact assessment methodology prescribed by Zitholele Consulting is given below.

To ensure uniformity, the assessment of impacts is addressed in a standard manner so that a wide range of impacts can be compared with each other. For this reason a clearly defined significance rating scale is provided to assess the significance (importance) of the associated impacts. The scale embraces the notion of extent and magnitude, but does not always clearly define these since their importance in the rating scale is very relative. For example, the magnitude (i.e. the size) of an area affected by atmospheric pollution may be extremely large (1000 km²) but the significance of this effect is dependent on the concentration or level of pollution. If the concentration were great, the significance of the impact would be HIGH or VERY HIGH, but if it were dilute it would be LOW or VERY LOW. Similarly, if 60 ha of a grassland type are destroyed the impact would be VERY HIGH if only 100 ha of that grassland type was known. The impact would be VERY LOW if the grassland type were common.

The potential significance of every environmental impact identified is determined by using a ranking scale, based on the following (the terminology is extracted from the DEAT guideline document on EIA Regulations, April 1998):

Occurrence

- Probability of occurrence (how likely is it that the impact may occur?), and
- Duration of occurrence (how long may it last?)

Severity

- Magnitude (severity) of impact (will the impact be of high, moderate or low severity?), and
- Scale/extent of impact (will the impact affect the national, regional or local environment, or only that of the site?)

In order to assess each of these factors for each impact, the following ranking scales were used:

Probability:

5 – Definite/don't know
4 – Highly probable
3 – Medium probability
2 – Low probability
1 – Improbable
0 – None

Duration:

5 – Permanent
4 – Long-term (ceases with the operational life)
3 – Medium-term (5-15 years)
2 – Short-term (0-5 years)
1 – Immediate

Scale:

5 – International
4 – National
3 – Regional (>5km)
2 – Local (<5km)
1 – Site only
0 – None

Magnitude:

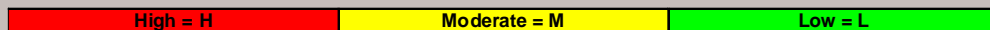
10 – Very high/don't know
8 – High
6 – Moderate
4 – Low
2 – Minor

Once the above factors had been ranked for each impact, the environmental significance of each was assessed using the following formula:

$$SP = (\text{magnitude} + \text{duration} + \text{scale}) \times \text{probability}$$

The maximum value is 100 significance points (SP). Environmental effects were rated as either of high, moderate or low significance on the following basis:

- More than 60 significance points indicated high environmental significance.
- Between 30 and 60 significance points indicated moderate environmental significance.
- Less than 30 significance points indicated low environmental significance.



Please note that **only negative impact will be ranked**

The degree of certainty of the assessment was judged on the following criteria:

Definite:	More than 90% sure of a particular fact.
Probable:	Between 70 and 90% sure of a particular fact, or of the likelihood of that impact occurring.
Possible:	Between 40 and 70% sure of a particular fact or of the likelihood of an impact occurring.
Unsure:	Less than 40% sure of a particular fact or the likelihood of an impact occurring.

Figure 46: Impact Assessment Methodology

5. **IMPACT ASSESSMENT**

The impact assessment is undertaken for each of the detailed biophysical fields assessed above i.e. soils and land capability, terrestrial ecology and surface water and wetlands. The assessment will aim to describe the impacts of each of the project areas and then also to differentiate between the available alternatives to identify the most suitable alternative for each of the main routes. Furthermore the assessment will also take cognisance of the expected different project phases i.e. construction and operations. At this stage it is not foreseen that the infrastructure will be decommissioned at any stage.

During construction the route will be surveyed, pegged and the soil nominations undertaken for each of the potential pylon foundations. The construction team will set up a construction camp in the study area and travel to site each day, transporting steel, workers and equipment to each of the tower sites. In some cases the power line servitude is cleared of vegetation to ease construction activities and to prevent possible electrical faults with nearby trees. The first step is the excavation of the pylon foundations, the reinforcing thereof and finally the concreting of the foundations. The equipment required to excavate the foundations can be manual labour, a TLB or in the case of hard rock – a drill rig will be required. The concrete will have to be transported via concrete trucks to the required locations.

After the foundations and footings have been installed the construction team will transport the various steel parts of the towers to the site and start erection of the pylons. This process again requires a lot of manual labour and often mobile cranes are used to assist with the erection of the towers. Once the tower are erected the stringing of the conductor cable commences, from tower to tower and the line is tensioned as per the requirements.

Once stringing and tensioning is complete the line is considered constructed, where after it will be tested prior to being commissioned. Once operational the line will conduct power along the approved route to the various substations. Operational and maintenance activities can include inspections via vehicle or helicopter and maintenance and repairs along the lines.

5.1 **Soils and Land Capability**

5.1.1 Existing impact

The soils and land capability as described in Section 3 highlights the study area as a sandy/calcrete area with very little cultivation. The area is arid and all the soils have a high base status as a result. The narrow area along the Orange River floodplain has been converted to high value agriculture with the cultivation of grapes, dates and vegetables. This industry is a major source of revenue for the province. The other areas are mainly utilised for grazing of cattle, sheep and goats with a couple of game farms operating in the area as well.

Along the Ferrum routes the existing impacts are found in the form of opencast iron ore mining at the mines around Kathu, linear structures such as the N14 highway to Upington and the compulsory farm roads. With the exception of the Kathu area, the soils along this route are relatively undisturbed. There are isolated cases where farm roads cross over dunes, removing vegetation and resulting in some minor erosion on the dune crests.

The Aries and Nieuwehoop area is similarly undisturbed, with urbanisation around Upington and the agricultural activities around the Orange River the only ones affecting the soils in this area. From an agricultural potential perspective this use of soil for agriculture is encouraged, hence the farming around the Orange River is not seen as a negative impact, as it might in the case of the terrestrial ecology assessment.

Around the proposed Eskom CSP site and the 132 kV routes to Gordonia substation the bulk of the area is also used for grazing land. There are a few activities along the route, such as the Duineveld landfill site, the Upington Airport as well as the town itself that have impacted upon the soils in this area.

The existing impact to soils for all four the study areas are rated as a Low Impact as shown in the table below.

Table 3: Soil and Land Capability Initial Impact Assessment

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Minor	<i>Site only</i>	<u>Medium Term</u>	<u>Definite</u>	Moderate
	2	1	3	5	30

5.1.2 Additional impact

5.1.2.1. Impact assessment

The additional impact of the proposed power lines will mainly be in the form of the clearing of the vegetation for the pylon sites, excavation of the foundations for the pylons, and the construction of access roads to the pylons (if required). In terms of impact to agriculture, grazing can continue under the power lines and in the servitudes as well as the planting of low growing crops. The activities that are limited is the use of large irrigation systems such as pivots, spraying of crops by planes and the planting of high growing crops such as fruit trees, windbreaks and palms.

The average area of a typical self-supporting tower footing is estimated at 14 m². Now there are various pylon design alternatives, but for this assessment worst case scenario is assumed which is the self-supporting towers along the entire route. With a tower placed every 350m you can calculate the potential impact to the soils. The potential impact for each route alternative is given below.

Table 4: Impacts to soils for each route alternative

Route Alternative	Length (km)	Foundation Impact to Soils (ha)	Agric Soils in corridor	Shifting Soils in corridor
Ferrum 1	212 km	0.85 ha	0 ha	17 683 ha
Ferrum 2	245 km	0.98 ha	0 ha	24 146 ha
Ferrum 3	279 km	1.12 ha	0 ha	18 032 ha
Ferrum 3 variation	267 km	1.07 ha	0 ha	15 051 ha
Aries 1	131 km	0.52 ha	74 ha	2 739 ha

Aries 2	121 km	0.48 ha	284 ha	2 450 ha
Aries 3	114 km	0.45 ha	651 ha	1 009 ha
Nieuwehoop 1	73 km	0.29 ha	327 ha	68 ha
Nieuwehoop 2	63 km	0.25 ha	497 ha	0 ha
Nieuwehoop variation	65 km	0.26 ha	630 ha	0 ha
Gordonia 1	29 km	0.12 ha	0 ha	36 ha

As shown in the table above when considering the potential impacts to soil and agriculture, the consideration is made for the impact to agricultural soils. But in this study area the soils also pose a risk to the potential development. The prevalence of shifting sands provide a potential risk to the stability of the pylons and the line overall.

In addition to the impact of the pylon foundations the potential impact of an access road must also be considered. It is assumed that the power lines will require an access road for the length of the route, hence the longer the route the larger the impact. However the Ferrum 3 and Ferrum 3 variation routes are aligned with a major road, the N14 in this case, and this road can be used for the transport of the bulk of the materials. Access roads will still be required from the highway to the specific pylons but this is a major advantage for these routes. The same applies to the Nieuwehoop 2 variation route, which is aligned along a provincial dirt road.

Once operational the impacts to the soil will remain, and if the construction activities have not been properly managed, wind erosion will start to occur in this phase. The utilisation and maintenance of roads will become important to limit the impacts.

Considering all the factors mentioned above, the potential impact to soils and agriculture and the potential risks for each of the alternatives are given in the table below.

Table 5: Additional impact by the proposed development to the soils and agriculture

Alternative	Significance	Spatial	Temporal	Probability	Rating
Ferrum 1	Low	Site	Long Term	Definite	45 - Moderate
Risk	Very High	Local	Long Term	High	64 - High
Ferrum 2	Low	Site	Long Term	Definite	45 - Moderate
Risk	Very High	Local	Long Term	High	64 - High
Ferrum 3	Minor	Site	Long Term	Definite	35 - Moderate
Risk	High	Local	Long Term	High	56 - Moderate
Ferrum 3 variation	Minor	Site	Long Term	Definite	35 - Moderate
Risk	High	Local	Long Term	High	56 - Moderate
Aries 1	Low	Site	Long Term	Definite	45 - Moderate
Risk	Moderate	Site	Long Term	High	44 - Moderate
Aries 2	Low	Site	Long Term	Definite	45 - Moderate
Risk	Moderate	Site	Long Term	High	44 - Moderate
Aries 3	Low	Site	Long Term	Definite	45 - Moderate
Risk	Moderate	Site	Long Term	High	44 - Moderate

Alternative	Significance	Spatial	Temporal	Probability	Rating
Nieuwehoop 1	Low	Site	Long Term	Definite	45 - Moderate
Risk	Minor	Site	Long Term	Medium	21 – Low
Nieuwehoop 2	Low	Site	Long Term	Definite	45 - Moderate
Risk	Minor	Site	Long Term	Medium	21 – Low
Nieuwehoop variation	Minor	Site	Long Term	Definite	35 - Moderate
Risk	Minor	Site	Long Term	Medium	21 – Low
Gordonia 1	Minor	Site	Long Term	Definite	35 - Moderate
Risk	Moderate	Site	Long Term	High	44 - Moderate

From the table above it can be seen that the impacts to soils and agriculture over the length of each of the alternative routes is regarded as a Moderate impact. The risk when considering the shifting sands and erosion is rated as a Low impact for the Nieuwehoop lines and a Moderate to High impact for the rest of the alternatives.

5.1.2.2. Preferred alternatives

It should be noted that the overall scale of the assessment makes it difficult to discern which of the routes are preferred, as the site conditions over the vast distances covered by these lines are very similar. Here we attempt to discuss the minor differences between the routes that the impact assessment table did not show.

Ferrum route

The routes to the Ferrum Substation in the east of the study area have all been rated as Moderate Impacts to soils, however upon closer inspection it can be seen that Ferrum 1 and 2 rated as 45, while Ferrum 3 and the variation to the route rated as a 35. This is due to the fact that the latter routes have main access roads in place that can be utilised without the need for major access road construction. In addition these two routes are also in an area with a slightly lower risk of shifting sands. Based on these criteria it is recommended that either Ferrum Route 3 or Ferrum Route 3 Variation be utilised.

Aries route

The Aries and Nieuwehoop lines cross over the Orange River and the Agricultural soils around the river. This is such a small area in comparison to the rest of the routes that the assessment cannot distinguish between the alternatives. However when evaluating only the crossings of the agricultural areas, a clearer assessment can be made. Hence this section will aim to address that, and such sections will also be included in the terrestrial ecology section and surface water section.

As indicated in Table 4 above the three Aries alternatives have varying levels of impact to the agricultural soils surrounding the river. Alternative 1 has a much smaller impact to agricultural soils than the other two alternatives. This is due to the environment down-stream of the Neus-weir. Here the Orange River flows through a number of sandstone outcrops and ridges and very little sediment has been deposited. Due to the smaller impact on the agriculture of the area – it is recommended that the Aries Alternative 1 route be utilised.

Nieuwehoop routes

As with the Aries routes above, the Nieuwehoop routes traverse over the Orange River and the surrounding farmland. From Table 4 it can be seen that in the case of the Nieuwehoop routes, that Alternative 1 has the smallest impact to agriculture and it is recommended to be utilised as the crossing point for the power line.

5.1.3 Mitigation/management measures

The following measures are proposed to manage and mitigate the potential impacts to soils and agricultural activities along the various routes.

- Utilise the alternative suggested above;
- Avoid unnecessary removal of vegetation cover;
- Use existing access roads as far as possible;
- If a new road is constructed, ensure that the Eskom erosion prevention guideline is followed and adhere to the Eskom tower construction specification TRMSCAAC1 Rev 3;
- Take land use into consideration when choosing pylon types, it is recommended that smaller footprint pylons be used in cultivated areas;
- Avoid placement of pylon footings in clay soils as well as on dunes, towers to be sited in between dunes in the so-called dune-streets;
- Avoid the construction of access roads through dunes;
- Spread absorbent sand on areas where oil spills are likely to occur, such as the refuelling area in the hard park;
- Oil-contaminated soils are to be removed to a contained storage area and bio-remediated or disposed of at a licensed facility;
- Use berms to minimise erosion where vegetation is disturbed, including hard parks, plant sites, borrow pit and office areas;
- If soils are excavated for the footing placement, ensure that the soil is utilised elsewhere for rehabilitation/road building purposes; and
- Ensure that soil is stockpiled in such a way as to prevent erosion from wind/storm water.

5.1.4 Cumulative impact

The cumulative impact of the power line construction and operations along with the impacts discussed in Section 5.1.1 slightly raise the impact score to 35, however the impact rating remains a Moderate impact as shown below.

Table 6: Soil and Land Capability Cumulative Impact Assessment

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Minor	<i>Site only</i>	<u>Long Term</u>	<u>Definite</u>	Moderate
	2	1	4	5	35

5.2 Terrestrial Ecology

5.2.1 Existing impact

In terms of the existing impact to the ecology of the study area, the vegetation has hardly been disturbed in most cases and the area is almost natural in appearance. All the vegetation units with the exception of the Lower Gariep Alluvial Vegetation show less than 5% transformation. Hence the impact on these areas is rated as a Low impact as shown below.

Table 7: Vegetation Initial Impact Assessment

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Minor	<i>Site only</i>	<u>Short term</u>	<u>Definite</u>	Low
	2	1	2	5	25

The impact to the Lower Gariep Alluvial Vegetation is another matter completely. Due to the high agricultural value of the soils and the proximity to the Orange River as a water source, this vegetation unit has been largely (50%) transformed by agriculture, to the point that it is endangered. This impact rates as a High impact as shown below.

Table 8: Vegetation Initial Impact Assessment – Lower Gariep Alluvial Vegetation

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Very High	<i>Regional</i>	<u>Long Term</u>	<u>Definite</u>	High
	10	3	4	5	85

5.2.2 Additional impact

The additional impact of the proposed power lines to the ecology will be the removal of vegetation within the servitude for the construction of the new power lines and the associated servitude roads. This is standard operating procedure for the construction of power lines. In addition to the impact to the vegetation, the noise and activity might scare local fauna away from the study area. The overall impact of each of the power line route alternatives on each vegetation unit is shown in the Table below. Please note that the areas indicated are for the entire corridor, not only the line.

The impact to vegetation if the standard operating procedure to clear the vegetation in the servitude is followed then the impact would be rated as a Moderate impact as shown below. Due to the bulk of the vegetation unit all being rated as not threatened, this rating applies to all the route alternatives.

Table 9: Vegetation Additional Impact Assessment

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Low	<i>Site only</i>	<u>Long-Term</u>	<u>High</u>	Moderate
	4	1	4	4	36

Table 10: Vegetation Impact per route

Route Alternative	Bushmanland Arid Grassland	Bushmanland Basin Shrubland	Gordonia Duneveld	Gordonia Plains Shrubland	Kalahari Karroid Shrubland	Kathu Bushveld	Koranna-Langeberg Mountain Bushveld	Olifantshoek Plains Thornveld	Southern Kalahari Salt Pans	Lower Gariep Alluvial Vegetation	Lower Gariep Broken Veld
Aries 1	15505	1616	550		2222					98	2404
Aries 2	14525	1767	637		1807					313	1339
Aries 3	15691	1238	969							656	623
Ferrum 1	837		11385	6382	10572	3999	2995	7943	177		
Ferrum 2	835		16308	7872	6690	9372	154	2511	3		
Ferrum 3	840		8208	11353	15820	9367	176	3584	77		
Ferrum 3 variation	832		7848	8476	15850	4059	2447	7789	59		
Niewehoop 1	11223		58		798					309	
Niewehoop 2	10245				15					491	
Nieuwehoop Variation	10518				362					603	
Gordonia	15		36		108						

However if considering the impact to the one endangered vegetation unit the impact is rated as a High impact as none of the potential Aries or Nieuwehoop routes can avoid impacting on the endangered vegetation.

Table 11: Vegetation Additional Impact Assessment – Lower Gariep Alluvial Vegetation

Impact	Significance	Spatial Scale	Temporal Scale	Probability	Rating
Impact to Soils	Very High	Site	Long Term	High	High
	10	1	4	4	60

5.2.2.1. Preferred alternatives

As mentioned in the soils section, the overall scale of the assessment makes it difficult to discern which of the routes are preferred, as the site conditions over the vast distances covered by these lines are very similar. Here we attempt to discuss the minor differences between the routes that the impact assessment table did not show.

Ferrum route

From a terrestrial ecology perspective the four alternatives for the proposed Ferrum route are very similar in nature. The area is largely natural with little impact to the environment. In terms of the potential impacts it is recommended that either Route 3 or the Route 3 variation be utilised as the use of existing access roads will significantly reduce the potential impact.

Aries route

The Aries and Nieuwehoop lines cross over the Orange River and the alluvial vegetation around the river. This is such a small area in comparison to the rest of the routes that the assessment cannot distinguish between the alternatives. However when evaluating only the crossings of the endangered habitats, a clearer assessment can be made.

As indicated in Table 9 above the three Aries alternatives have varying levels of impact to the endangered habitat (marked in green). Alternative 1 has a much smaller impact than the other two alternatives. This is due to the environment down-stream of the Neus-weir. Here the Orange River flows through a number of sandstone outcrops and ridges and very little riparian vegetation occurs. Due to the smaller impact on the endangered vegetation – it is recommended that the Aries Alternative 1 route be utilised.

Nieuwehoop routes

As with the Aries routes above, the Nieuwehoop routes traverse over the Orange River and the surrounding endangered habitat. From Table 9 it can be seen that in the case of the Nieuwehoop routes, that Alternative 1 has the smallest impact to the sensitive habitat and it is recommended to be utilised as the crossing point for the power line.

5.2.3 Mitigation/management measures

The following measures are proposed to manage and mitigate the potential impacts to terrestrial ecology along the various routes:

- General:

- No hunting or cooking to be permitted on site;
- All construction areas should be demarcated prior to construction to ensure that the footprint of the impacts are limited (including areas where vehicles may traverse);
- All alien invasive species on site should be removed and follow up monitoring and removal programmes should be initiated once construction is completed;
- Alternative 1 should be considered as the preferred alternative;
- Adhere to the ESKOM vegetation management guideline;
- The Environmental Control Officer should identify any sensitive along the servitude, particularly large terrestrial species and notify the fauna specialist of these so that advice can be given on how to best deal with the situation;
- The construction of new access roads in particular should be limited to a minimum; and
- All vehicle and pedestrian movement should be restricted to the actual construction site and, in the case of maintenance patrols, to the actual servitude.
- Sensitive habitat/species:
 - Removal of plants should be restricted to only trees that pose a risk to the power line. All other vegetation should not be cleared with the exception of the footprint excavations;
 - Once the route is pegged, identify all tree that require removal and identify if they require a permit from DAFF or NEM:BA;
 - The sensitive alluvial vegetation unit should be avoided and construction limited to 50 m from the edge of the endangered habitat if possible;
 - If construction has to take place inside the CBA, ensure that it takes place in areas that have already been disturbed;

5.2.4 Cumulative impact

If the abovementioned mitigation measures are implemented successfully, then the cumulative impact of the power lines and the existing impacts should not result in an impact larger than was assessed for the initial impacts.

5.3 Surface Water and Wetlands

5.3.1 Existing impact

5.3.2 Additional impact

5.3.3 Mitigation/management measures

- No construction camps or pylons should be placed within 50m from the edge of a surface water body.
- Demarcated areas where waste can be safely contained and stored on a temporary basis during the construction phase should be provided at the hard park;
- Waste is not to be buried on site;
- Hydro-carbons should be stored in a bunded storage area;
- All hazardous materials inter alia paints, turpentine and thinners must be stored appropriately to prevent these contaminants from entering the environment;
- Spill-sorb or similar type product must be used to absorb hydrocarbon spills in the event that such spills should occur;
- Care must be taken to ensure that in removing vegetation adequate erosion control measures are implemented;
- A storm-water management plan, including sufficient erosion-control measures, must be compiled in consultation with a suitably qualified environmental practitioner / control officer during the detailed design phase prior to the commencement of construction; and
- The propagation of low-growing dense vegetation suitable for the habitat such as grasses, sedges or reeds is the best natural method to reduce erosion potential in sensitive areas.

5.3.4 Cumulative impact

6. **PREFERRED ALTERNATIVES**

As part of the impact assessment undertaken in this report, it was aimed to determine the most suitable alternative to be utilised for each of the potential routes. Using the three detailed studies in the report, surface water, terrestrial ecology and soils it was determined that the following was the most preferred routes for each route:

Ferrum to Solar Park – Ferrum Option 3 or its variation;

Aries to Solar Park – Aries Option 1; and

Nieuwehoop to Solar Park – Nieuwehoop Option 1.

The Gordonia routes to the Solar Park all follow the same alignment along an existing 132 kV power line route and therefore no comment can be made on the preferred route.

7. CONCLUSION AND WAY FORWARD

In conclusion this report aimed to identify the surface water, terrestrial ecology and soils that could be impacted by the proposed Solar Integration Project. From the detailed assessments it became clear that the bulk of the sensitivities in the study area are located around the Orange River, where the sensitive habitats as well as the main farming activities occur. The Orange River is also the only perennial water body in the area and of utmost importance to the Province.

The routes to Ferrum provided a different environment with the occurrence of the red Kalahari sands and in some cases dunes. These red sands are susceptible to erosion and also “shifting”, and could be a tricky obstacle when constructing.

As a whole the study area is devoid of access routes and access to the alternatives that are far from existing provincial or national roads might be problematic.

The study identified preferred alternatives for each route, based on the potential impacts to sensitive features along the routes. In addition mitigation and management measures were proposed for each of the criteria assessed and with the successful implementation of these measures, it is the opinion of the consultant that the impacts from this proposed development are within the acceptable range.

8. REFERENCES

Department of Environmental Affairs, 2010a. *National Environmental Management Act, 1998 (Act 107 of 1998). Environmental Impact Assessment Regulations.* Government Gazette 33306, Government Notice R 543 of 18 June, 2010, Government Printer, Pretoria.

Department of Environmental Affairs, 2010b. *National Environmental Management Act, 1998 (Act 107 of 1998). Listing Notice 1: List of activities and competent authorities identified in terms of Section 24(2) and 24D.* Government Gazette 33306, Government Notice R 544 of 18 June, 2010, Government Printer, Pretoria.

Department of Environmental Affairs, 2010c. *National Environmental Management Act, 1998 (Act 107 of 1998). Listing Notice 2: List of activities and competent authorities identified in terms of Section 24(2) and 24D.* Government Gazette 33306, Government Notice R 545 of 18 June, 2010, Government Printer, Pretoria.

Department of Environmental Affairs, 2010d. *National Environmental Management Act, 1998 (Act 107 of 1998). Listing Notice 3: List of activities and competent authorities identified in terms of Section 24(2) and 24D.* Government Gazette 33306, Government Notice R 546 of 18 June, 2010, Government Printer, Pretoria.

- Department of Water Affairs and Forestry (DWAF), 2005. *A practical field procedure for identification and delineation of wetlands and riparian areas*. Department of Water Affairs and Forestry, Pretoria
- Eijkelkamp Agrisearch Equipment, 2004. *Revised Standard Soil Colour Charts*, Eijkelkamp, Giesbeek.
- Kotze et al, 2008. *Wetland Management Series, WET-EcoServices, A technique for rapidly assessing ecosystem services supplied by wetlands*, Water Research Commission, WRC Report TT 339/08,
- Nel, J.L., Murray, K.M., Maherry, A.M., Petersen, C.P., Roux, D.J., Driver, A., Hill, L., Van Deventer, H., Funke, N., Swartz, E.R., Smith-Adao, L.B., Mbona, N., Downsborough, L. and Nienaber, S. (2011). *Technical Report for the National Freshwater Ecosystem Priority Areas project*. WRC Report No. K5/1801. <http://bgis.sanbi.org/>
- SANBI, 2009. *Further Development of a Proposed National Wetland Classification System for South Africa. Primary Project Report*. Prepared by the Freshwater Consulting Group (FCG) for the South African National Biodiversity Institute (SANBI).
- Soil Classification Workgroup, 1991. *Soil Classification; a taxonomic system for South Africa*, Memoirs on the Natural Resources of South Africa, no. 15, Department of Agricultural Development, Pretoria.
- South Africa, Republic, 1998. *National Water Act, Act No. 36 of 1998*. Government Gazette 19182, Government Printer, Pretoria.
- South Africa, Republic, 1998. *National Environmental Management Act, Act No. 107 of 1998*. Government Gazette 19519, Government Printer, Pretoria.

...
Project Manager

...
An-other

...

Project Director
for Jones & Wagener

4 October 2012

*Document source: M:\D615 - Eskom Solar Integration Project
(ENV)\Report\D615_00_Rep_00_kk_Rev0_BiophysicalRep_Oct2012.docx
Document template: Scoping Report Template10.dotx*

ZITHOLELE CONSULTING ON BEHALF OF ESKOM HOLDINGS

D615 - SOLAR INTEGRATION PROJECT

DRAFT BIOPHYSICAL REPORT

Report: JW194/12/D615

APPENDIX A

SPECIES LISTS



ZITHOLELE CONSULTING ON BEHALF OF ESKOM HOLDINGS

D615 - SOLAR INTEGRATION PROJECT

DRAFT BIOPHYSICAL REPORT

Report: JW194/12/D615

APPENDIX B

OTHER



<u>FamilyName</u>	<u>SpeciesName</u>	<u>Category</u>
ACANTHACEAE	Acanthopsis disperma	Plants
ACANTHACEAE	Barleria greenii	Plants
ACANTHACEAE	Barleria lichtensteiniana	Plants
ACANTHACEAE	Barleria rigida	Plants
ACANTHACEAE	Blepharis mitrata	Plants
ACANTHACEAE	Monechma divaricatum	Plants
ACANTHACEAE	Monechma genistifolium subsp. australe	Plants
ACANTHACEAE	Monechma genistifolium subsp. genistifolium	Plants
ACANTHACEAE	Monechma incanum	Plants
ACANTHACEAE	Monechma spartioides	Plants
ACANTHACEAE	Acanthopsis hoffmannseggiana	Plants
ACANTHACEAE	Monechma desertorum	Plants
ACANTHACEAE	Monechma distichotrichum	Plants
ACANTHACEAE	Monechma sp.	Plants
AIZOACEAE	Aizoon asbestinum	Plants
AIZOACEAE	Aizoon schellenbergii	Plants
AIZOACEAE	Galenia africana	Plants
AIZOACEAE	Galenia crystallina	Plants
AIZOACEAE	Plinthus cryptocarpus	Plants
AIZOACEAE	Tetragonia arbuscula	Plants
AIZOACEAE	Tetragonia reduplicata	Plants
AIZOACEAE	Trianthema parvifolia var. parvifolia	Plants
AIZOACEAE	Mesembryanthemum noctiflorum subsp. stramineum	Plants
AIZOACEAE	Aizoon canariense	Plants
AIZOACEAE	Galenia herniariaefolia	Plants
AIZOACEAE	Galenia sarcophylla	Plants
AIZOACEAE	Plinthus karooicus	Plants
AIZOACEAE	Plinthus sericeus	Plants
AMARANTHACEAE	Amaranthus praetermissus	Plants
AMARANTHACEAE	Amaranthus thunbergii	Plants
AMARANTHACEAE	Hermbstaedia odorata var. odorata	Plants
AMARANTHACEAE	Leucosphaera bainesii	Plants
AMARANTHACEAE	Sericocoma avolans	Plants
AMARANTHACEAE	Amaranthus dinteri subsp. dinteri var. a	Plants
AMARANTHACEAE	Sericocoma pungens	Plants
AMARANTHACEAE	Sericorema remotiflora	Plants
AMARYLLIDACEAE	Haemanthus humilis subsp. humilis	Plants
AMARYLLIDACEAE	Nerine laticoma	Plants
AMARYLLIDACEAE	Crinum bulbispermum	Plants
AMARYLLIDACEAE	Crinum sp.	Plants
ANACARDIACEAE	Rhus lancea	Plants
ANACARDIACEAE	Searsia lancea	Plants
ANACARDIACEAE	Searsia pendulina	Plants
APOCYNACEAE	Adenium oleifolium	Plants
APOCYNACEAE	Cryptolepis decidua	Plants
APOCYNACEAE	Cynanchum orangeanum	Plants

APOCYNACEAE	<i>Gomphocarpus fruticosus</i> subsp. <i>fruticosus</i>	Plants
APOCYNACEAE	<i>Microloma incanum</i>	Plants
APOCYNACEAE	<i>Orbea lugardii</i>	Plants
APOCYNACEAE	<i>Orbea</i> sp.	Plants
APOCYNACEAE	<i>Pergularia daemia</i> var. <i>leiocarpa</i>	Plants
APOCYNACEAE	<i>Sarcostemma viminale</i> subsp. <i>viminale</i>	Plants
APOCYNACEAE	<i>Fockea sinuata</i>	Plants
APOCYNACEAE	<i>Gomphocarpus filiformis</i>	Plants
APOCYNACEAE	<i>Hoodia gordonii</i>	Plants
APOCYNACEAE	<i>Larryleachia dinteri</i>	Plants
APOCYNACEAE	<i>Larryleachia marlothii</i>	Plants
APOCYNACEAE	<i>Lavrania marlothii</i>	Plants
APOCYNACEAE	<i>Stapelia grandiflora</i> var. <i>grandiflora</i>	Plants
APOCYNACEAE	<i>Lavrania</i> sp.	Plants
APOCYNACEAE	<i>Orbea lutea</i> subsp. <i>lutea</i>	Plants
APOCYNACEAE	<i>Sarcostemma pearsonii</i>	Plants
APOCYNACEAE	<i>Tridentea marientalensis</i> subsp. <i>marientalensis</i>	Plants
ASPARAGACEAE	<i>Asparagus lignosus</i>	Plants
ASPARAGACEAE	<i>Asparagus pearsonii</i>	Plants
ASPARAGACEAE	<i>Asparagus stipulaceus</i>	Plants
ASPARAGACEAE	<i>Asparagus exuvialis</i> forma <i>exuvialis</i>	Plants
ASPARAGACEAE	<i>Asparagus suaveolens</i>	Plants
ASPARAGACEAE	<i>Asparagus denudatus</i>	Plants
ASPHODELACEAE	<i>Aloe claviflora</i>	Plants
ASPHODELACEAE	<i>Aloe dichotoma</i>	Plants
ASPHODELACEAE	<i>Aloe dichotoma</i> var. <i>dichotoma</i>	Plants
ASPHODELACEAE	<i>Aloe gariensis</i>	Plants
ASPHODELACEAE	<i>Aloe hereroensis</i> var. <i>hereroensis</i>	Plants
ASPHODELACEAE	<i>Trachyandra laxa</i> var. <i>laxa</i>	Plants
ASPHODELACEAE	<i>Aloe variegata</i>	Plants
ASPHODELACEAE	<i>Trachyandra jacquiniana</i>	Plants
ASPLENIACEAE	<i>Asplenium cordatum</i>	Plants
ASTERACEAE	<i>Arctotis leiocarpa</i>	Plants
ASTERACEAE	<i>Berkheya annectens</i>	Plants
ASTERACEAE	<i>Berkheya spinosissima</i> subsp. <i>spinosissima</i>	Plants
ASTERACEAE	<i>Dicoma capensis</i>	Plants
ASTERACEAE	<i>Dimorphotheca polyptera</i>	Plants
ASTERACEAE	<i>Eriocephalus microphyllus</i> var. <i>pubescens</i>	Plants
ASTERACEAE	<i>Felicia deserti</i>	Plants
ASTERACEAE	<i>Felicia muricata</i> subsp. <i>cinerascens</i>	Plants
ASTERACEAE	<i>Geigeria ornativa</i>	Plants
ASTERACEAE	<i>Geigeria pectidea</i>	Plants
ASTERACEAE	<i>Gorteria corymbosa</i>	Plants
ASTERACEAE	<i>Helichrysum gariense</i>	Plants
ASTERACEAE	<i>Hirpicium echinus</i>	Plants
ASTERACEAE	<i>Ifloga molluginoides</i>	Plants
ASTERACEAE	<i>Kleinia longiflora</i>	Plants

ASTERACEAE	<i>Laggera decurrens</i>	Plants
ASTERACEAE	<i>Leysera tenella</i>	Plants
ASTERACEAE	<i>Litogyne gariepina</i>	Plants
ASTERACEAE	<i>Nolletia arenosa</i>	Plants
ASTERACEAE	<i>Nolletia gariepina</i>	Plants
ASTERACEAE	<i>Osteospermum microcarpum</i> subsp. <i>microcarpum</i>	Plants
ASTERACEAE	<i>Pegolettia retrofracta</i>	Plants
ASTERACEAE	<i>Pentzia argentea</i>	Plants
ASTERACEAE	<i>Pentzia pinnatisecta</i>	Plants
ASTERACEAE	<i>Pentzia spinescens</i>	Plants
ASTERACEAE	<i>Pteronia mucronata</i>	Plants
ASTERACEAE	<i>Pteronia unguiculata</i>	Plants
ASTERACEAE	<i>Rosenia oppositifolia</i>	Plants
ASTERACEAE	<i>Senecio sisymbriifolius</i>	Plants
ASTERACEAE	<i>Tripteris microcarpa</i> subsp. <i>microcarpa</i>	Plants
ASTERACEAE	<i>Ursinia nana</i> subsp. <i>nana</i>	Plants
ASTERACEAE	<i>Amellus epaleaceus</i>	Plants
ASTERACEAE	<i>Amellus strigosus</i> subsp. <i>pseudoscabridus</i>	Plants
ASTERACEAE	<i>Amellus tridactylus</i> subsp. <i>arenarius</i>	Plants
ASTERACEAE	<i>Arctotis</i> sp.	Plants
ASTERACEAE	<i>Athanasia minuta</i> subsp. <i>minuta</i>	Plants
ASTERACEAE	<i>Berkheya spinosissima</i> subsp. <i>namaensis</i> var. <i>namaensis</i>	Plants
ASTERACEAE	<i>Chrysocoma ciliata</i>	Plants
ASTERACEAE	<i>Cineraria lobata</i> subsp. <i>lobata</i>	Plants
ASTERACEAE	<i>Dimorphotheca sinuata</i>	Plants
ASTERACEAE	<i>Eriocephalus ambiguus</i>	Plants
ASTERACEAE	<i>Eriocephalus ericoides</i> subsp. <i>ericoides</i>	Plants
ASTERACEAE	<i>Eriocephalus pauperrimus</i>	Plants
ASTERACEAE	<i>Eriocephalus spinescens</i>	Plants
ASTERACEAE	<i>Felicia clavipilosa</i> subsp. <i>clavipilosa</i>	Plants
ASTERACEAE	<i>Felicia hyssopifolia</i> subsp. <i>hyssopifolia</i>	Plants
ASTERACEAE	<i>Felicia muricata</i> subsp. <i>muricata</i>	Plants
ASTERACEAE	<i>Foveolina dichotoma</i>	Plants
ASTERACEAE	<i>Gazania lichtensteinii</i>	Plants
ASTERACEAE	<i>Geigeria filifolia</i>	Plants
ASTERACEAE	<i>Geigeria vigintiquamea</i>	Plants
ASTERACEAE	<i>Helichrysum herniarioides</i>	Plants
ASTERACEAE	<i>Helichrysum zeyheri</i>	Plants
ASTERACEAE	<i>Lasiopogon glomerulatus</i>	Plants
ASTERACEAE	<i>Nidorella microcephala</i>	Plants
ASTERACEAE	<i>Nidorella resedifolia</i> subsp. <i>resedifolia</i>	Plants
ASTERACEAE	<i>Osteospermum armatum</i>	Plants
ASTERACEAE	<i>Osteospermum muricatum</i> subsp. <i>longiradiatum</i>	Plants
ASTERACEAE	<i>Osteospermum pinnatum</i> var. <i>breve</i>	Plants
ASTERACEAE	<i>Osteospermum pinnatum</i> var. <i>pinnatum</i>	Plants
ASTERACEAE	<i>Osteospermum spinescens</i>	Plants
ASTERACEAE	<i>Othonna</i> sp.	Plants

ASTERACEAE	<i>Pentzia globosa</i>	Plants
ASTERACEAE	<i>Pteronia acuminata</i>	Plants
ASTERACEAE	<i>Pteronia glauca</i>	Plants
ASTERACEAE	<i>Pteronia leucoclada</i>	Plants
ASTERACEAE	<i>Pteronia sordida</i>	Plants
ASTERACEAE	<i>Pteronia</i> sp.	Plants
ASTERACEAE	<i>Rosenia glandulosa</i>	Plants
ASTERACEAE	<i>Rosenia humilis</i>	Plants
ASTERACEAE	<i>Senecio burchellii</i>	Plants
ASTERACEAE	<i>Senecio glutinarius</i>	Plants
ASTERACEAE	<i>Senecio niveus</i>	Plants
ASTERACEAE	<i>Sonchus oleraceus</i>	Plants
ASTERACEAE	<i>Trichogyne paronychioides</i>	Plants
ASTERACEAE	<i>Tripteris sinuata</i> var. <i>linearis</i>	Plants
ASTERACEAE	<i>Tripteris sinuata</i> var. <i>sinuata</i>	Plants
ASTERACEAE	<i>Geigeria ornativa</i> subsp. <i>ornativa</i>	Plants
ASTERACEAE	<i>Bidens bipinnata</i>	Plants
ASTERACEAE	<i>Helichrysum micropoides</i>	Plants
ASTERACEAE	<i>Pentzia</i> sp.	Plants
ASTERACEAE	<i>Senecio consanguineus</i>	Plants
ASTERACEAE	<i>Senecio trachylaenus</i>	Plants
ASTERACEAE	<i>Verbesina encelioides</i> var. <i>encelioides</i>	Plants
AYTONIACEAE	<i>Plagiochasma rupestre</i> var. <i>rupestre</i>	Plants
AZOLLACEAE	<i>Azolla filiculoides</i>	Plants
BIGNONIACEAE	<i>Rhigozum trichotomum</i>	Plants
BIGNONIACEAE	<i>Rhigozum obovatum</i>	Plants
BORAGINACEAE	<i>Codon royenii</i>	Plants
BORAGINACEAE	<i>Ehretia rigida</i> subsp. <i>rigida</i>	Plants
BORAGINACEAE	<i>Heliotropium ciliatum</i>	Plants
BORAGINACEAE	<i>Heliotropium supinum</i>	Plants
BORAGINACEAE	<i>Trichodesma africanum</i>	Plants
BRASSICACEAE	<i>Heliophila carnosa</i>	Plants
BRASSICACEAE	<i>Heliophila minima</i>	Plants
BRASSICACEAE	<i>Heliophila</i> sp.	Plants
BRASSICACEAE	<i>Heliophila trifurca</i>	Plants
BRASSICACEAE	<i>Coronopus integrifolius</i>	Plants
BRASSICACEAE	<i>Heliophila deserticola</i>	Plants
BRASSICACEAE	<i>Heliophila deserticola</i> var. <i>deserticola</i>	Plants
BRASSICACEAE	<i>Heliophila remotiflora</i>	Plants
BRASSICACEAE	<i>Lepidium africanum</i> subsp. <i>africanum</i>	Plants
BRASSICACEAE	<i>Lepidium desertorum</i>	Plants
BRASSICACEAE	<i>Lepidium schinzii</i>	Plants
BRASSICACEAE	<i>Sisymbrium burchellii</i> var. <i>burchellii</i>	Plants
BRYACEAE	<i>Bryum argenteum</i>	Plants
BURSERACEAE	<i>Commiphora gracilifrondosa</i>	Plants
CAMPANULACEAE	<i>Wahlenbergia denticulata</i> var. <i>denticulata</i>	Plants
CAPPARACEAE	<i>Boscia foetida</i> subsp. <i>foetida</i>	Plants

CAPPARACEAE	<i>Cadaba aphylla</i>	Plants
CAPPARACEAE	<i>Cleome angustifolia</i> subsp. <i>diandra</i>	Plants
CAPPARACEAE	<i>Cleome oxyphylla</i> var. <i>oxyphylla</i>	Plants
CAPPARACEAE	<i>Cleome paxii</i>	Plants
CAPPARACEAE	<i>Maerua gilgii</i>	Plants
CAPPARACEAE	<i>Boscia albitrunca</i>	Plants
CAPPARACEAE	<i>Cleome gynandra</i>	Plants
CARYOPHYLLACEAE	<i>Pollichia campestris</i>	Plants
CELASTRACEAE	<i>Gymnosporia linearis</i> subsp. <i>lanceolata</i>	Plants
CHENOPODIACEAE	<i>Atriplex semibaccata</i> var. <i>appendiculata</i>	Plants
CHENOPODIACEAE	<i>Salsola glabrescens</i>	Plants
CHENOPODIACEAE	<i>Salsola kali</i>	Plants
CHENOPODIACEAE	<i>Salsola namibica</i>	Plants
CHENOPODIACEAE	<i>Salsola tuberculata</i>	Plants
CHENOPODIACEAE	<i>Suaeda caespitosa</i>	Plants
CHENOPODIACEAE	<i>Suaeda merxmulleri</i>	Plants
CHENOPODIACEAE	<i>Bassia salsoloides</i>	Plants
CHENOPODIACEAE	<i>Chenopodium glaucum</i>	Plants
CHENOPODIACEAE	<i>Salsola aphylla</i>	Plants
CHENOPODIACEAE	<i>Salsola barbata</i>	Plants
CHENOPODIACEAE	<i>Salsola rabieana</i>	Plants
CHENOPODIACEAE	<i>Atriplex semibaccata</i> var. <i>typica</i>	Plants
COLCHICACEAE	<i>Colchicum melanthoides</i> subsp. <i>melanthoides</i>	Plants
COLCHICACEAE	<i>Ornithoglossum vulgare</i>	Plants
COLCHICACEAE	<i>Colchicum bellum</i>	Plants
COLCHICACEAE	<i>Ornithoglossum viride</i>	Plants
COLCHICACEAE	<i>Androcymbium melanthioides</i> subsp. <i>melanthioides</i>	Plants
CONVOLVULACEAE	<i>Convolvulus sagittatus</i>	Plants
CRASSULACEAE	<i>Adromischus</i> sp.	Plants
CRASSULACEAE	<i>Cotyledon orbiculata</i> var. <i>orbiculata</i>	Plants
CRASSULACEAE	<i>Crassula muscosa</i> var. <i>muscosa</i>	Plants
CRASSULACEAE	<i>Crassula sericea</i> var. <i>sericea</i>	Plants
CRASSULACEAE	<i>Tylecodon rubrovenosus</i>	Plants
CRASSULACEAE	<i>Crassula corallina</i> subsp. <i>corallina</i>	Plants
CRASSULACEAE	<i>Crassula corallina</i> subsp. <i>macrorrhiza</i>	Plants
CRASSULACEAE	<i>Cotyledon orbiculata</i> var. <i>dactyloopsis</i>	Plants
CUCURBITACEAE	<i>Coccinia rehmannii</i>	Plants
CUCURBITACEAE	<i>Cucumis africanus</i>	Plants
CUCURBITACEAE	<i>Citrullus lanatus</i>	Plants
CUCURBITACEAE	<i>Cucumis myriocarpus</i> subsp. <i>leptodermis</i>	Plants
CUCURBITACEAE	<i>Cucumis myriocarpus</i> subsp. <i>myriocarpus</i>	Plants
CUCURBITACEAE	<i>Cucumis sagittatus</i>	Plants
CUCURBITACEAE	<i>Corallocarpus schinzii</i>	Plants
CUCURBITACEAE	<i>Kedrostis capensis</i>	Plants
CYPERACEAE	<i>Cyperus capensis</i>	Plants
CYPERACEAE	<i>Cyperus longus</i> var. <i>tenuiflorus</i>	Plants
CYPERACEAE	<i>Cyperus marginatus</i>	Plants

CYPERACEAE	Scirpoides dioecus	Plants
CYPERACEAE	Cyperus bellus	Plants
CYPERACEAE	Bulbostylis hispidula	Plants
CYPERACEAE	Cyperus fulgens var. contractus	Plants
CYPERACEAE	Cyperus usitatus	Plants
EBENACEAE	Diospyros lycioides subsp. lycioides	Plants
EBENACEAE	Diospyros pallens	Plants
ERIOSPERMACEAE	Eriospermum bakerianum subsp. bakerianum	Plants
ERIOSPERMACEAE	Eriospermum flagelliforme	Plants
EUPHORBIACEAE	Euphorbia avasmontana var. avasmontana	Plants
EUPHORBIACEAE	Euphorbia gariepina subsp. balsamea	Plants
EUPHORBIACEAE	Euphorbia gariepina subsp. gariepina	Plants
EUPHORBIACEAE	Euphorbia glanduligera	Plants
EUPHORBIACEAE	Euphorbia inaequilatera var. inaequilatera	Plants
EUPHORBIACEAE	Euphorbia rudis	Plants
EUPHORBIACEAE	Euphorbia spinea	Plants
EUPHORBIACEAE	Euphorbia avasmontana var. sagittaria	Plants
EUPHORBIACEAE	Euphorbia mauritanica var. mauritanica	Plants
FABACEAE	Acacia erioloba	Plants
FABACEAE	Acacia karroo	Plants
FABACEAE	Acacia mellifera subsp. detinens	Plants
FABACEAE	Cullen tomentosum	Plants
FABACEAE	Cyamopsis serrata	Plants
FABACEAE	Hoffmannseggia lactea	Plants
FABACEAE	Indigastrum argyraeum	Plants
FABACEAE	Indigastrum argyroides	Plants
FABACEAE	Indigofera alternans var. alternans	Plants
FABACEAE	Indigofera heterotricha	Plants
FABACEAE	Indigofera holubii	Plants
FABACEAE	Indigofera pungens	Plants
FABACEAE	Indigofera rhytidocarpa subsp. rhytidocarpa	Plants
FABACEAE	Lebeckia spinescens	Plants
FABACEAE	Lotononis platycarpa	Plants
FABACEAE	Lotononis rabenaviana	Plants
FABACEAE	Melolobium candicans	Plants
FABACEAE	Parkinsonia africana	Plants
FABACEAE	Ptycholobium biflorum subsp. biflorum	Plants
FABACEAE	Tephrosia dregeana var. dregeana	Plants
FABACEAE	Acacia tortilis subsp. heteracantha	Plants
FABACEAE	Lessertia annularis	Plants
FABACEAE	Lessertia sp.	Plants
FABACEAE	Lotononis falcata	Plants
FABACEAE	Lotononis marlothii	Plants
FABACEAE	Melolobium exudans	Plants
FABACEAE	Prosopis glandulosa var. glandulosa	Plants
FABACEAE	Prosopis glandulosa var. torreyana	Plants
FABACEAE	Prosopis sp.	Plants

FABACEAE	<i>Prosopis velutina</i>	Plants
FABACEAE	<i>Requienia sphaerosperma</i>	Plants
FABACEAE	<i>Sutherlandia frutescens</i>	Plants
FABACEAE	<i>Acacia haematoxylon</i>	Plants
FABACEAE	<i>Acacia pendula</i>	Plants
FABACEAE	<i>Adenolobus garipensis</i>	Plants
FABACEAE	<i>Indigofera auricoma</i>	Plants
FABACEAE	<i>Lebeckia linearifolia</i>	Plants
FABACEAE	<i>Lessertia macrostachya</i> var. <i>macrostachya</i>	Plants
FABACEAE	<i>Melolobium macrocalyx</i>	Plants
FABACEAE	<i>Pomaria lactea</i>	Plants
FABACEAE	<i>Senna italica</i> subsp. <i>arachoides</i>	Plants
FABACEAE	<i>Tephrosia burchellii</i>	Plants
FABACEAE	<i>Calobota linearifolia</i>	Plants
FABACEAE	<i>Calobota spinescens</i>	Plants
GERANIACEAE	<i>Monsonia burkeana</i>	Plants
GERANIACEAE	<i>Monsonia luederitziana</i>	Plants
GERANIACEAE	<i>Sarcocaulon patersonii</i>	Plants
GERANIACEAE	<i>Monsonia umbellata</i>	Plants
GERANIACEAE	<i>Pelargonium minimum</i>	Plants
GERANIACEAE	<i>Sarcocaulon crassaule</i>	Plants
GERANIACEAE	<i>Monsonia glauca</i>	Plants
GIGASPERMACEAE	<i>Chamaebryum pottioides</i>	Plants
GISEKIACEAE	<i>Gisekia pharnacioides</i> var. <i>pharnacioides</i>	Plants
GISEKIACEAE	<i>Gisekia africana</i> var. <i>africana</i>	Plants
HYACINTHACEAE	<i>Dipcadi ciliare</i>	Plants
HYACINTHACEAE	<i>Dipcadi glaucum</i>	Plants
HYACINTHACEAE	<i>Drimia physodes</i>	Plants
HYACINTHACEAE	<i>Ledebouria undulata</i>	Plants
HYACINTHACEAE	<i>Ornithogalum suaveolens</i>	Plants
HYACINTHACEAE	<i>Ornithogalum tenuifolium</i> subsp. <i>tenuifolium</i>	Plants
HYACINTHACEAE	<i>Albuca setosa</i>	Plants
HYACINTHACEAE	<i>Dipcadi brevifolium</i>	Plants
HYACINTHACEAE	<i>Dipcadi gracillimum</i>	Plants
HYACINTHACEAE	<i>Dipcadi viride</i>	Plants
HYACINTHACEAE	<i>Drimia intricata</i>	Plants
HYACINTHACEAE	<i>Ornithogalum juncifolium</i> var. <i>juncifolium</i>	Plants
HYACINTHACEAE	<i>Ornithogalum unifolium</i>	Plants
HYACINTHACEAE	<i>Ornithogalum unifolium</i> var. <i>unifolium</i>	Plants
HYACINTHACEAE	<i>Dipcadi bakerianum</i>	Plants
HYACINTHACEAE	<i>Dipcadi papillatum</i>	Plants
HYACINTHACEAE	<i>Ledebouria</i> sp.	Plants
HYACINTHACEAE	<i>Ornithogalum tenuifolium</i> subsp. <i>aridum</i>	Plants
IRIDACEAE	<i>Gladiolus saccatus</i>	Plants
IRIDACEAE	<i>Ferraria divaricata</i> subsp. <i>divaricata</i>	Plants
IRIDACEAE	<i>Ferraria ferrariola</i>	Plants
IRIDACEAE	<i>Lapeirousia plicata</i> subsp. <i>plicata</i>	Plants

IRIDACEAE	<i>Moraea serpentina</i>	Plants
IRIDACEAE	<i>Moraea speciosa</i>	Plants
IRIDACEAE	<i>Moraea venenata</i>	Plants
IRIDACEAE	<i>Ferraria variabilis</i>	Plants
IRIDACEAE	<i>Moraea pallida</i>	Plants
IRIDACEAE	<i>Moraea polystachya</i>	Plants
LAMIACEAE	<i>Leucas capensis</i>	Plants
LAMIACEAE	<i>Salvia verbenaca</i>	Plants
LECANORACEAE	<i>Lecanora</i> sp.	Plants
LOASACEAE	<i>Kissenia capensis</i>	Plants
LOPHIOCARPACEAE	<i>Lophiocarpus polystachyus</i>	Plants
LORANTHACEAE	<i>Tapinanthus oleifolius</i>	Plants
LORANTHACEAE	<i>Septulina glauca</i>	Plants
MALVACEAE	<i>Hermannia abrotanoides</i>	Plants
MALVACEAE	<i>Hermannia coccocarpa</i>	Plants
MALVACEAE	<i>Hermannia minutiflora</i>	Plants
MALVACEAE	<i>Hermannia modesta</i>	Plants
MALVACEAE	<i>Hermannia spinosa</i>	Plants
MALVACEAE	<i>Hermannia stricta</i>	Plants
MALVACEAE	<i>Hermannia tomentosa</i>	Plants
MALVACEAE	<i>Hermannia vestita</i>	Plants
MALVACEAE	<i>Hibiscus elliotiae</i>	Plants
MALVACEAE	<i>Abutilon angulatum</i> var. <i>angulatum</i>	Plants
MALVACEAE	<i>Abutilon pycnodon</i>	Plants
MALVACEAE	<i>Hermannia bicolor</i>	Plants
MALVACEAE	<i>Hermannia gariepina</i>	Plants
MALVACEAE	<i>Radyera urens</i>	Plants
MALVACEAE	<i>Corchorus asplenifolius</i>	Plants
MALVACEAE	<i>Hermannia</i> sp.	Plants
MALVACEAE	<i>Melhania didyma</i>	Plants
MALVACEAE	<i>Sida rhombifolia</i> subsp. <i>rhombifolia</i>	Plants
MARSILEACEAE	<i>Marsilea macrocarpa</i>	Plants
MELIACEAE	<i>Nymania capensis</i>	Plants
MELIANTHACEAE	<i>Melianthus comosus</i>	Plants
MENISPERMACEAE	<i>Cissampelos capensis</i>	Plants
MESEMBRYANTHEMACEAE	<i>Dinteranthus wilmotianus</i>	Plants
MESEMBRYANTHEMACEAE	<i>Lithops bromfieldii</i>	Plants
MESEMBRYANTHEMACEAE	<i>Mesembryanthemum coriarium</i>	Plants
MESEMBRYANTHEMACEAE	<i>Mesembryanthemum crystallinum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Psilocaulon articulatum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Psilocaulon coriarium</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia barnardii</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia divaricata</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia kenhardtensis</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia</i> sp.	Plants
MESEMBRYANTHEMACEAE	<i>Aptenia geniculiflora</i>	Plants
MESEMBRYANTHEMACEAE	<i>Aridaria noctiflora</i> subsp. <i>straminea</i>	Plants

MESEMBRYANTHEMACEAE	<i>Brownanthus ciliatus</i> subsp. <i>ciliatus</i>	Plants
MESEMBRYANTHEMACEAE	<i>Dinteranthus pole-evansii</i>	Plants
MESEMBRYANTHEMACEAE	<i>Drosanthemum hispidum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Drosanthemum lique</i>	Plants
MESEMBRYANTHEMACEAE	<i>Lithops julii</i> subsp. <i>fulleri</i>	Plants
MESEMBRYANTHEMACEAE	<i>Mesembryanthemum guerichianum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Mesembryanthemum nodiflorum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Mesembryanthemum stenandrum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Psilocaulon granulicaule</i>	Plants
MESEMBRYANTHEMACEAE	<i>Psilocaulon subnodosum</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia ferox</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia vulvaria</i>	Plants
MESEMBRYANTHEMACEAE	<i>Lithops</i> sp.	Plants
MESEMBRYANTHEMACEAE	<i>Prenia tetragona</i>	Plants
MESEMBRYANTHEMACEAE	<i>Psilocaulon</i> sp.	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia canonotata</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia hamata</i>	Plants
MESEMBRYANTHEMACEAE	<i>Ruschia ruralis</i>	Plants
MOLLUGINACEAE	<i>Limeum aethiopicum</i> subsp. <i>aethiopicum</i> var. <i>aethiopicum</i>	Plants
MOLLUGINACEAE	<i>Limeum argute-carinatum</i> var. <i>argute-carinatum</i>	Plants
MOLLUGINACEAE	<i>Limeum myosotis</i> var. <i>confusum</i>	Plants
MOLLUGINACEAE	<i>Limeum sulcatum</i> var. <i>gracile</i>	Plants
MOLLUGINACEAE	<i>Hypertelis salsoloides</i> var. <i>salsoloides</i>	Plants
MOLLUGINACEAE	<i>Mollugo cerviana</i> var. <i>cerviana</i>	Plants
MOLLUGINACEAE	<i>Limeum fenestratum</i> var. <i>fenestratum</i>	Plants
MONTINIACEAE	<i>Montinia caryophyllacea</i>	Plants
MORACEAE	<i>Ficus cordata</i> subsp. <i>cordata</i>	Plants
NEURADACEAE	<i>Grielum humifusum</i> var. <i>humifusum</i>	Plants
NEURADACEAE	<i>Grielum humifusum</i> var. <i>parviflorum</i>	Plants
NYCTAGINACEAE	<i>Phaeoptilum spinosum</i>	Plants
OPHIOGLOSSACEAE	<i>Ophioglossum</i> sp.	Plants
OXALIDACEAE	<i>Oxalis haedulipes</i>	Plants
OXALIDACEAE	<i>Oxalis beneprotecta</i>	Plants
OXALIDACEAE	<i>Oxalis lawsonii</i>	Plants
PANNARIACEAE	<i>Psoroma hypnorum</i>	Plants
PAPAVERACEAE	<i>Argemone mexicana</i> forma <i>mexicana</i>	Plants
PARMELIACEAE	<i>Lichen</i> sp.	Plants
PASSIFLORACEAE	<i>Adenia repanda</i>	Plants
PEDALIACEAE	<i>Harpagophytum zeyheri</i> subsp. <i>sublobatum</i>	Plants
PEDALIACEAE	<i>Pterodiscus luridus</i>	Plants
PEDALIACEAE	<i>Rogeria longiflora</i>	Plants
PEDALIACEAE	<i>Sesamum capense</i>	Plants
PEGANACEAE	<i>Peganum harmala</i>	Plants
PHYLLANTHACEAE	<i>Phyllanthus humilis</i>	Plants
PHYLLANTHACEAE	<i>Phyllanthus maderaspatensis</i>	Plants
PLUMBAGINACEAE	<i>Dyerophytum africanum</i>	Plants
POACEAE	<i>Anthepphora pubescens</i>	Plants

POACEAE	<i>Aristida adscensionis</i>	Plants
POACEAE	<i>Aristida congesta</i> subsp. <i>barbicollis</i>	Plants
POACEAE	<i>Aristida engleri</i> var. <i>engleri</i>	Plants
POACEAE	<i>Aristida vestita</i> var. <i>vestita</i>	Plants
POACEAE	<i>Brachiaria glomerata</i>	Plants
POACEAE	<i>Cenchrus ciliaris</i>	Plants
POACEAE	<i>Centropodia glauca</i>	Plants
POACEAE	<i>Digitaria eriantha</i>	Plants
POACEAE	<i>Digitaria sanguinalis</i>	Plants
POACEAE	<i>Echinochloa holubii</i>	Plants
POACEAE	<i>Echinochloa stagnina</i>	Plants
POACEAE	<i>Enneapogon cenchroides</i>	Plants
POACEAE	<i>Enneapogon desvauxii</i>	Plants
POACEAE	<i>Enneapogon scaber</i>	Plants
POACEAE	<i>Eragrostis annulata</i>	Plants
POACEAE	<i>Eragrostis biflora</i>	Plants
POACEAE	<i>Eragrostis curvula</i>	Plants
POACEAE	<i>Eragrostis lehmanniana</i> var. <i>lehmanniana</i>	Plants
POACEAE	<i>Eragrostis nindensis</i>	Plants
POACEAE	<i>Eragrostis porosa</i>	Plants
POACEAE	<i>Eragrostis rotifer</i>	Plants
POACEAE	<i>Eriochloa fatmensis</i>	Plants
POACEAE	<i>Melinis repens</i> subsp. <i>grandiflora</i>	Plants
POACEAE	<i>Panicum arbusculum</i>	Plants
POACEAE	<i>Schmidtia kalahariensis</i>	Plants
POACEAE	<i>Setaria appendiculata</i>	Plants
POACEAE	<i>Setaria pumila</i>	Plants
POACEAE	<i>Setaria verticillata</i>	Plants
POACEAE	<i>Stipagrostis amabilis</i>	Plants
POACEAE	<i>Stipagrostis ciliata</i> var. <i>capensis</i>	Plants
POACEAE	<i>Stipagrostis namaquensis</i>	Plants
POACEAE	<i>Stipagrostis obtusa</i>	Plants
POACEAE	<i>Stipagrostis uniplumis</i> var. <i>neesii</i>	Plants
POACEAE	<i>Stipagrostis uniplumis</i> var. <i>uniplumis</i>	Plants
POACEAE	<i>Tragus berteronianus</i>	Plants
POACEAE	<i>Tragus racemosus</i>	Plants
POACEAE	<i>Triraphis ramosissima</i>	Plants
POACEAE	<i>Urochloa panicoides</i>	Plants
POACEAE	<i>Aristida congesta</i> subsp. <i>congesta</i>	Plants
POACEAE	<i>Aristida diffusa</i> subsp. <i>burkei</i>	Plants
POACEAE	<i>Chloris virgata</i>	Plants
POACEAE	<i>Dichanthium annulatum</i> var. <i>papillosum</i>	Plants
POACEAE	<i>Eragrostis brizantha</i>	Plants
POACEAE	<i>Eragrostis echinochloidea</i>	Plants
POACEAE	<i>Eragrostis homomalla</i>	Plants
POACEAE	<i>Eragrostis lehmanniana</i> var. <i>chaunantha</i>	Plants
POACEAE	<i>Eragrostis macrochlamys</i> var. <i>macrochlamys</i>	Plants

POACEAE	<i>Eragrostis macrochlamys</i> var. <i>wilmaniae</i>	Plants
POACEAE	<i>Eragrostis obtusa</i>	Plants
POACEAE	<i>Eragrostis procumbens</i>	Plants
POACEAE	<i>Eragrostis</i> x <i>pseud-obtusa</i>	Plants
POACEAE	<i>Fingerhuthia africana</i>	Plants
POACEAE	<i>Leptochloa fusca</i>	Plants
POACEAE	<i>Leucophrys mesocoma</i>	Plants
POACEAE	<i>Microchloa caffra</i>	Plants
POACEAE	<i>Oropetium capense</i>	Plants
POACEAE	<i>Panicum lanipes</i>	Plants
POACEAE	<i>Schismus barbatus</i>	Plants
POACEAE	<i>Schmidtia pappophoroides</i>	Plants
POACEAE	<i>Sporobolus ioclados</i>	Plants
POACEAE	<i>Sporobolus nervosus</i>	Plants
POACEAE	<i>Stipagrostis anomala</i>	Plants
POACEAE	<i>Stipagrostis hochstetteriana</i> var. <i>secalina</i>	Plants
POACEAE	<i>Tricholaena capensis</i> subsp. <i>capensis</i>	Plants
POACEAE	<i>Triraphis purpurea</i>	Plants
POACEAE	<i>Aristida vestita</i>	Plants
POACEAE	<i>Digitaria</i> sp.	Plants
POACEAE	<i>Dinebra retroflexa</i>	Plants
POACEAE	<i>Eragrostis aspera</i>	Plants
POACEAE	<i>Melinis repens</i> subsp. <i>repens</i>	Plants
POACEAE	<i>Melinis</i> sp.	Plants
POACEAE	<i>Phalaris canariensis</i>	Plants
POACEAE	<i>Setaria italica</i>	Plants
POACEAE	<i>Setaria</i> sp.	Plants
POACEAE	<i>Stipagrostis hochstetteriana</i> var. <i>hochstetteriana</i>	Plants
POLYGALACEAE	<i>Polygala seminuda</i>	Plants
POLYGALACEAE	<i>Polygala leptophylla</i> var. <i>armata</i>	Plants
POLYGONACEAE	<i>Oxygonum alatum</i> var. <i>alatum</i>	Plants
PORTULACACEAE	<i>Anacampseros baeseckeii</i>	Plants
PORTULACACEAE	<i>Anacampseros filamentosa</i> subsp. <i>filamentosa</i>	Plants
PORTULACACEAE	<i>Anacampseros filamentosa</i> subsp. <i>namaquensis</i>	Plants
PORTULACACEAE	<i>Avonia albissima</i>	Plants
PORTULACACEAE	<i>Portulaca hereroensis</i>	Plants
PORTULACACEAE	<i>Portulaca pilosa</i>	Plants
PORTULACACEAE	<i>Portulaca quadrifida</i>	Plants
PORTULACACEAE	<i>Talinum arnotii</i>	Plants
PORTULACACEAE	<i>Avonia ustulata</i>	Plants
PORTULACACEAE	<i>Portulaca oleracea</i>	Plants
PORTULACACEAE	<i>Talinum tenuissimum</i>	Plants
PORTULACACEAE	<i>Anacampseros filamentosa</i> subsp. <i>tomentosa</i>	Plants
PORTULACACEAE	<i>Portulaca kermesina</i>	Plants
RESEDACEAE	<i>Oligomeris dipetala</i> var. <i>dipetala</i>	Plants
RHAMNACEAE	<i>Ziziphus mucronata</i> subsp. <i>mucronata</i>	Plants
RICCIACEAE	<i>Riccia albornata</i>	Plants

RICCIACEAE	<i>Riccia okahandjana</i>	Plants
RUBIACEAE	<i>Kohautia cynanchica</i>	Plants
RUBIACEAE	<i>Kohautia ramosissima</i>	Plants
RUBIACEAE	<i>Kohautia caespitosa</i> subsp. <i>brachyloba</i>	Plants
SALICACEAE	<i>Salix mucronata</i> subsp. <i>mucronata</i>	Plants
SANTALACEAE	<i>Thesium lineatum</i>	Plants
SANTALACEAE	<i>Thesium hystricoides</i>	Plants
SCROPHULARIACEAE	<i>Anticharis senegalensis</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum albomarginatum</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum elongatum</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum lineare</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum lineare</i> var. <i>lineare</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum marlothii</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum procumbens</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum spinescens</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia argentea</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia aridicola</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia glutinosa</i>	Plants
SCROPHULARIACEAE	<i>Peliostomum leucorrhizum</i>	Plants
SCROPHULARIACEAE	<i>Antherothamnus pearsonii</i>	Plants
SCROPHULARIACEAE	<i>Cromidon minutum</i>	Plants
SCROPHULARIACEAE	<i>Diascia engleri</i>	Plants
SCROPHULARIACEAE	<i>Diascia</i> sp.	Plants
SCROPHULARIACEAE	<i>Hebenstretia integrifolia</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia</i> sp.	Plants
SCROPHULARIACEAE	<i>Lyperia tristis</i>	Plants
SCROPHULARIACEAE	<i>Manulea nervosa</i>	Plants
SCROPHULARIACEAE	<i>Manulea schaeferi</i>	Plants
SCROPHULARIACEAE	<i>Nemesia fleckii</i>	Plants
SCROPHULARIACEAE	<i>Selago albida</i>	Plants
SCROPHULARIACEAE	<i>Selago divaricata</i>	Plants
SCROPHULARIACEAE	<i>Veronica anagallis-aquatica</i>	Plants
SCROPHULARIACEAE	<i>Zaluzianskya diandra</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum junceum</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia atropurpurea</i> subsp. <i>pubescens</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia integerrima</i>	Plants
SCROPHULARIACEAE	<i>Selago paniculata</i>	Plants
SOLANACEAE	<i>Lycium bosciifolium</i>	Plants
SOLANACEAE	<i>Lycium cinereum</i>	Plants
SOLANACEAE	<i>Lycium oxycarpum</i>	Plants
SOLANACEAE	<i>Lycium pumilum</i>	Plants
SOLANACEAE	<i>Nicotiana glauca</i>	Plants
SOLANACEAE	<i>Solanum capense</i>	Plants
SOLANACEAE	<i>Lycium schizocalyx</i>	Plants
SOLANACEAE	<i>Solanum burchellii</i>	Plants
SOLANACEAE	<i>Solanum tomentosum</i> var. <i>tomentosum</i>	Plants
TAMARICACEAE	<i>Tamarix usneoides</i> E.Mey. ex Bunge x <i>T. ramosissima</i> Ledeb.	Plants

TECOPHILAEACEAE	<i>Cyanella lutea</i>	Plants
THYMELAEACEAE	<i>Gnidia polycephala</i>	Plants
UNKNOWN	Unknown sp.	Plants
URTICACEAE	<i>Forsskaolea candida</i>	Plants
VAHLIACEAE	<i>Vahlia capensis</i> subsp. <i>vulgaris</i> var. <i>longifolia</i>	Plants
VERBENACEAE	<i>Chascanum garipense</i>	Plants
VERBENACEAE	<i>Chascanum pumilum</i>	Plants
VERBENACEAE	<i>Chascanum incisum</i>	Plants
VERBENACEAE	<i>Chascanum pinnatifidum</i> var. <i>pinnatifidum</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus cristatus</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus pterophorus</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus terrestris</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus zeyheri</i> subsp. <i>zeyheri</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum dregeanum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum flexuosum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum gilfillanii</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum rigidum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum simplex</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum</i> sp.	Plants
ZYGOPHYLLACEAE	<i>Augea capensis</i>	Plants
ZYGOPHYLLACEAE	<i>Sisyndite spartea</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum lichtensteinianum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum retrofractum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum suffruticosum</i>	Plants
ZYGOPHYLLACEAE	<i>Fagonia sinaica</i> var. <i>minutistipula</i>	Plants

<u>FamilyName</u>	<u>SpeciesName</u>	<u>Category</u>
Agamidae	Agama anchietae	Animals
Anostomatidae	Henicus monstrosus	Animals
Apidae	Megachile sp.	Animals
Apidae	Meliturgula sp.	Animals
Apidae	Nomia sp	Animals
Apidae	Thyreus brachyaspis	Animals
Apidae	Thyreus calceatus	Animals
Araneidae	Gea infuscata	Animals
Baetidae	Unidentified Baetidae	Animals
Baetidae	Centroptilum sp.	Animals
Baetidae	Pseudocloeon magae	Animals
Braconidae	Iphiaulax dodsii	Animals
Buprestidae	Lampetis ocelligera	Animals
Buprestidae	Lampetis albomarginata chalcophoroides	Animals
Buprestidae	Lampetis amaurotica	Animals
Buprestidae	Lampetis amaurotica fuchsai	Animals
Buprestidae	Lampetis comorica	Animals
Buprestidae	Lampetis limbalis	Animals
Buprestidae	Ptosima sexmaculata	Animals
Buprestidae	Sphenoptera vinosa	Animals
Caenidae	Unidentified Caenidae	Animals
Cambalidae	Julomorpha fortis	Animals
Carabidae	Geobaenus ingenuus	Animals
Carabidae	Macrocheilus hybridus	Animals
Carabidae	Phloeozetus cordiger	Animals
Carabidae	Scarites sexualis	Animals
Carabidae	Trechodes babaulti	Animals
Chrysopidae	Italochrysa gigantia	Animals
Clariidae	Clarias gariepinus	Animals
Colubridae	Dasypeltis scabra	Animals
Colubridae	Dipsina multimaculata	Animals
Colubridae	Lycophidion capense	Animals
Cordylidae	Cordylus polyzonus	Animals
Coreidae	Homoeocerus trabeatus	Animals
Curculionidae	Platycopes gonopterus	Animals
Curculionidae	Porpacus cornirostris	Animals
Curculionidae	Hipporhinus subvittatus var.cinerascens	Animals
Cyprinidae	Labeobarbus kimberleyensis	Animals
Diapriidae	Ferrieropria	Animals
Eresidae	Gandanomeno depressus	Animals
Erirhididae	Hyposomus bevinsi	Animals
Eucoilidae	Gronotoma nitida	Animals
Euschmitiidae	genus ign. nr Penichrotes	Animals
Gekkonidae	Pachydactylus laevigatus	Animals
Gekkonidae	Pachydactylus montanus	Animals
Gekkonidae	Pachydactylus purcelli	Animals
Gekkonidae	Pachydactylus serval	Animals
Gekkonidae	Pachydactylus capensis	Animals
Gekkonidae	Pachydactylus latirostris	Animals

Gekkonidae	<i>Pachydactylus mariquensis</i>	Animals
Gekkonidae	<i>Chondrodactylus angulifer</i>	Animals
Gekkonidae	<i>Pachydactylus turneri</i>	Animals
Gekkonidae	<i>Ptenopus garrulus</i>	Animals
Gnaphosidae	<i>Zelotes cronwrighti</i>	Animals
Gnaphosidae	<i>Zelotes gooldi</i>	Animals
Gyrinidae	<i>Aulonogyrus alternatus</i>	Animals
Hesperiidae	<i>Alenia sandaster</i>	Animals
Hesperiidae	<i>Pyrgus fritillarius fritillum</i>	Animals
Hesperiidae	<i>Spialia mafa mafa</i>	Animals
Histeridae	<i>Exorhabdus marshalli</i>	Animals
Histeridae	<i>Hister stercorarius</i>	Animals
Histeridae	<i>Hister sulcimargo</i>	Animals
Histeridae	<i>Saprinus pseudocyanus</i>	Animals
Ichneumonidae	<i>Triclistus</i>	Animals
Ichneumonidae	<i>Paracollyria</i>	Animals
Keptageniidae	<i>Heptagenia sulphurea</i>	Animals
Lacertidae	<i>Pedioplanis lineoocellata</i>	Animals
Lacertidae	<i>Pedioplanis undata</i>	Animals
Leptoceridae	Unidentified Leptoceridae	Animals
Leptoceridae	<i>Leptecho</i> sp.	Animals
Leptoceridae	<i>Oecetis</i> sp.	Animals
Leptophlebiidae	<i>Adenophlebia peringueyella</i>	Animals
Libellulidae	<i>Trithemis arteriosa</i>	Animals
Liopteridae	<i>Paramblynotus hirsutebumus</i>	Animals
Lycaenidae	<i>Azanus ubaldus</i>	Animals
Lycaenidae	<i>Chilades trochylus</i>	Animals
Lycaenidae	<i>Aloeides arida</i>	Animals
Lycaenidae	<i>Aloeides barklyi</i>	Animals
Lycaenidae	<i>Aloeides molomo molomo</i>	Animals
Lycaenidae	<i>Brephidium metophis</i>	Animals
Lycaenidae	<i>Cacyreus lingeus</i>	Animals
Lycaenidae	<i>Iolus (Stugeta) bowkeri bowkeri</i>	Animals
Lycaenidae	<i>Trimenia macmasteri macmasteri</i>	Animals
Lycaenidae	<i>Tylopaedia sardonix peringueyi</i>	Animals
Lycaenidae	<i>Zizula hylax</i>	Animals
LYCAENIDAE	<i>Aloeides damarensis</i> subsp. <i>mashona</i>	Animals
LYCAENIDAE	<i>Chrysoritis pan</i> subsp. <i>lysander</i>	Animals
LYCAENIDAE	<i>Leptotes pirithous</i> subsp. <i>pirithous</i>	Animals
Lycaenidae	<i>Aloeides damarensis damarensis</i>	Animals
Lycaenidae	<i>Cigaritis namaqua</i>	Animals
Lycaenidae	<i>Cigaritis phanes</i>	Animals
Lycaenidae	<i>Iolus (Stugeta) subinfusata reynoldsi</i>	Animals
LYCAENIDAE	<i>Aloeides simplex</i>	Animals
LYCAENIDAE	<i>Stugeta bowkeri</i> subsp. <i>bowkeri</i>	Animals
LYCAENIDAE	<i>Stugeta subinfusata</i> subsp. <i>reynoldsi</i>	Animals
LYCAENIDAE	<i>Tarucus sybaris</i> subsp. <i>linearis</i>	Animals
Lycosidae	<i>Lycosa cretata</i>	Animals
Lygaeidae	<i>Spilostethus pandurus militaris</i>	Animals
Lygaeidae	<i>Spilostethus taeniatus</i>	Animals

Megachilidae	<i>Fidelia braunsiana</i>	Animals
Melittidae	<i>Meganomia binghami</i>	Animals
Membracidae	<i>Oxyrhachis subserrata</i>	Animals
Muscidae	Unidentified Muscidae	Animals
Naucoridae	<i>Laccocoris</i> sp.	Animals
Nemestrinidae	<i>Prosoeca robusta</i>	Animals
Nymphalidae	<i>Henotesia perspicua</i>	Animals
Nymphalidae	<i>Tarsocera namaquensis</i>	Animals
NYMPHALIDAE	<i>Acraea neobule</i> subsp. <i>neobule</i>	Animals
NYMPHALIDAE	<i>Acraea trimeni</i>	Animals
NYMPHALIDAE	<i>Danaus chrysippus</i> subsp. <i>orientis</i>	Animals
Nymphalidae	<i>Neptis jordani</i>	Animals
NYMPHALIDAE	<i>Acraea stenobea</i>	Animals
Pamphagidae	<i>Transvaalana draconis</i>	Animals
Perlidae	<i>Neoperla transvaalensis</i>	Animals
Pieridae	<i>Belenois aurota aurota</i>	Animals
Pieridae	<i>Catopsilia florella</i>	Animals
Pieridae	<i>Colotis eris eris</i>	Animals
Pieridae	<i>Pontia helice helice</i>	Animals
Pieridae	<i>Colotis agoye bowkeri</i>	Animals
Pieridae	<i>Colotis agoye agoye</i>	Animals
PIERIDAE	<i>Colotis euipe</i> subsp. <i>mediata</i>	Animals
PIERIDAE	<i>Pinacopteryx eriphia</i> subsp. <i>eriphia</i>	Animals
Plumariidae	<i>Myrmecopterina minor</i>	Animals
Pompilidae	<i>Priocnemis clypeatus</i>	Animals
Pompilidae	<i>Priocnemis fumipennis</i>	Animals
Pselaphidae	<i>Reichenbachia sulcicornis</i>	Animals
Pteromalidae	<i>Mesopolobus fasciiventris</i>	Animals
Ranidae	Unidentified Ranidae	Animals
Reduviidae	<i>Rhincoris rufigena</i>	Animals
Salticidae	<i>Festicula australis</i>	Animals
Salticidae	<i>Menemerus rubicundus</i>	Animals
Scarabaeidae	<i>Proagoderus gemmatus</i>	Animals
Scarabaeidae	<i>Microtochalus plagiger</i>	Animals
Scarabaeidae	<i>Liatongus quadripunctatus</i>	Animals
Scarabaeidae	<i>Onitis confusus</i>	Animals
Scarabaeidae	<i>Onthophagus orthocerus</i>	Animals
Scarabaeidae	<i>Oocamenta rufiventris</i>	Animals
Schendylidae	<i>Schendylurus caledonicus</i>	Animals
Scincidae	<i>Acontias lineatus</i>	Animals
Scincidae	<i>Mabuya</i> sp.	Animals
Scincidae	<i>Mabuya sulcata</i>	Animals
Scincidae	<i>Mabuya occidentalis</i>	Animals
Silphidae	<i>Silpha (Silpha) peringueyi</i>	Animals
Simuliidae	<i>Simulium bovis</i>	Animals
Sisyridae	Unidentified Sisyridae	Animals
Solpugidae	<i>Solpuguna collinita</i>	Animals
Sphecidae	<i>Tachytes labilis</i>	Animals
Sphecidae	<i>Laphyragogus pictus</i>	Animals
Staphylinidae	<i>Zyras (Camonia) conifera</i>	Animals

Stenopelmatidae	Sia (Maxentius) pallidus	Animals
Syrphidae	Paragus (Pandasyopthalmus) punctatus	Animals
Tabanidae	Mesomyia (Perisilvius) redunda	Animals
Tabanidae	Mesomyia(Mesomyia) aurantiaca	Animals
Tabanidae	Mesomyia(Mesomyia) costata	Animals
Tabanidae	Mesomyia(Mesomyia) namaquina	Animals
Tachinidae	Winthemia quadrata	Animals
Tenebrionidae	Strongylium lautum	Animals
Tenebrionidae	Strongylium muata	Animals
Tenebrionidae	Hypomelus vulpinus	Animals
Tenebrionidae	Eutrapela bicolor	Animals
Tenebrionidae	Strongylium perturbator	Animals
Testudinidae	Psammobates tentorius	Animals
Tetragnathidae	Nephila inaurata	Animals
Tettigoniidae	Hemihetrodes bachmanni	Animals
Thomisidae	Oxyptila	Animals
Tricorythidae	Tricorythus discolor	Animals
Tricorythidae	Unidentified Tricorythidae	Animals
Typhlopidae	Typhlops sp.	Animals
Unidentified Trichoptera	Unidentified Trichoptera	Animals
Viperidae	Bitis caudalis	Animals

FamilyName	SpeciesName	Category
ACANTHACEAE	Barleria macrostegia	Plants
ACANTHACEAE	Barleria rigida	Plants
ACANTHACEAE	Justicia puberula	Plants
ACANTHACEAE	Justicia thymifolia	Plants
ACANTHACEAE	Monechma divaricatum	Plants
ACANTHACEAE	Monechma incanum	Plants
ACANTHACEAE	Barleria irritans	Plants
ACANTHACEAE	Monechma genistifolium subsp. genistifolium	Plants
ACANTHACEAE	Blepharis mitrata	Plants
ACANTHACEAE	Barleria macrostegia	Plants
ACANTHACEAE	Blepharis integrifolia var. integrifolia	Plants
ACANTHACEAE	Blepharis marginata	Plants
ACANTHACEAE	Blepharis sp.	Plants
ACANTHACEAE	Glossochilus burchellii	Plants
ACANTHACEAE	Hypoestes forskalii	Plants
AIZOACEAE	Aizoon schellenbergii	Plants
AIZOACEAE	Plinthus karooicus	Plants
AIZOACEAE	Plinthus sericeus	Plants
AIZOACEAE	Tetragonia saligna	Plants
AIZOACEAE	Trianthema parvifolia var. parvifolia	Plants
AMARANTHACEAE	Aerva leucura	Plants
AMARANTHACEAE	Hermbstaedtia odorata var. odorata	Plants
AMARANTHACEAE	Sericorema remotiflora	Plants
AMARANTHACEAE	Alternanthera pungens	Plants
AMARANTHACEAE	Gomphrena celosioides	Plants
AMARANTHACEAE	Kyphocarpa angustifolia	Plants
AMARANTHACEAE	Pupalia lappacea var. lappacea	Plants
AMARANTHACEAE	Sericorema sericea	Plants
AMARYLLIDACEAE	Boophone disticha	Plants
AMARYLLIDACEAE	Haemanthus humilis subsp. humilis	Plants
ANACARDIACEAE	Searsia burchellii	Plants
ANACARDIACEAE	Searsia ciliata	Plants
ANACARDIACEAE	Searsia tridactyla	Plants
ANACARDIACEAE	Searsia tenuinervis	Plants
ANTHERICACEAE	Chlorophytum fasciculatum	Plants
APIACEAE	Deverra burchellii	Plants
APOCYNACEAE	Acokanthera oppositifolia	Plants
APOCYNACEAE	Fockea angustifolia	Plants
APOCYNACEAE	Piaranthus decipiens	Plants
APOCYNACEAE	Sarcostemma viminale subsp. viminale	Plants
APOCYNACEAE	Stapelia olivacea	Plants
APOCYNACEAE	Gomphocarpus tomentosus	Plants
APOCYNACEAE	Gomphocarpus tomentosus subsp. tomentosus	Plants
APOCYNACEAE	Microloma armatum var. burchellii	Plants
APOCYNACEAE	Pergularia daemia subsp. daemia	Plants
APOCYNACEAE	Raphionacme velutina	Plants

ASPARAGACEAE	<i>Asparagus suaveolens</i>	Plants
ASPARAGACEAE	<i>Asparagus bechuanicus</i>	Plants
ASPARAGACEAE	<i>Asparagus larycinus</i>	Plants
ASPARAGACEAE	<i>Asparagus exuvialis</i> forma <i>exuvialis</i>	Plants
ASPARAGACEAE	<i>Asparagus retrofractus</i>	Plants
ASPHODELACEAE	<i>Aloe hereroensis</i> var. <i>hereroensis</i>	Plants
ASPHODELACEAE	<i>Aloe</i> sp.	Plants
ASPHODELACEAE	<i>Bulbine narcissifolia</i>	Plants
ASTERACEAE	<i>Arctotheca calendula</i>	Plants
ASTERACEAE	<i>Arctotis leiocarpa</i>	Plants
ASTERACEAE	<i>Chrysocoma ciliata</i>	Plants
ASTERACEAE	<i>Chrysocoma obtusata</i>	Plants
ASTERACEAE	<i>Cineraria lyratiformis</i>	Plants
ASTERACEAE	<i>Cineraria vallis-pacis</i>	Plants
ASTERACEAE	<i>Dicoma anomala</i> subsp. <i>gerrardii</i>	Plants
ASTERACEAE	<i>Dicoma capensis</i>	Plants
ASTERACEAE	<i>Dicoma macrocephala</i>	Plants
ASTERACEAE	<i>Dimorphotheca sinuata</i>	Plants
ASTERACEAE	<i>Eriocephalus ericoides</i> subsp. <i>griquensis</i>	Plants
ASTERACEAE	<i>Euryops multifidus</i>	Plants
ASTERACEAE	<i>Felicia muricata</i> subsp. <i>cinerascens</i>	Plants
ASTERACEAE	<i>Felicia muricata</i> subsp. <i>muricata</i>	Plants
ASTERACEAE	<i>Gazania krebsiana</i> subsp. <i>serrulata</i>	Plants
ASTERACEAE	<i>Helichrysum argyrosphaerum</i>	Plants
ASTERACEAE	<i>Helichrysum cerastioides</i> var. <i>cerastioides</i>	Plants
ASTERACEAE	<i>Helichrysum pumilio</i> subsp. <i>pumilio</i>	Plants
ASTERACEAE	<i>Helichrysum zeyheri</i>	Plants
ASTERACEAE	<i>Hertia pallens</i>	Plants
ASTERACEAE	<i>Ifloga glomerata</i>	Plants
ASTERACEAE	<i>Leysera tenella</i>	Plants
ASTERACEAE	<i>Lopholaena cneorifolia</i>	Plants
ASTERACEAE	<i>Metalasia trivialis</i>	Plants
ASTERACEAE	<i>Nolletia ciliaris</i>	Plants
ASTERACEAE	<i>Osteospermum muricatum</i> subsp. <i>muricatum</i>	Plants
ASTERACEAE	<i>Pentzia incana</i>	Plants
ASTERACEAE	<i>Pentzia viridis</i>	Plants
ASTERACEAE	<i>Senecio consanguineus</i>	Plants
ASTERACEAE	<i>Tarchonanthus camphoratus</i>	Plants
ASTERACEAE	<i>Tarchonanthus obovatus</i>	Plants
ASTERACEAE	<i>Verbesina encelioides</i> var. <i>encelioides</i>	Plants
ASTERACEAE	<i>Geigeria ornativa</i> subsp. <i>ornativa</i>	Plants
ASTERACEAE	<i>Arctotis virgata</i>	Plants
ASTERACEAE	<i>Berkheya ferox</i> var. <i>tomentosa</i>	Plants
ASTERACEAE	<i>Dimorphotheca polyptera</i>	Plants
ASTERACEAE	<i>Eriocephalus ambiguus</i>	Plants
ASTERACEAE	<i>Eriocephalus microphyllus</i> var. <i>pubescens</i>	Plants
ASTERACEAE	<i>Felicia fascicularis</i>	Plants

ASTERACEAE	<i>Felicia hirta</i>	Plants
ASTERACEAE	<i>Foveolina dichotoma</i>	Plants
ASTERACEAE	<i>Helichrysum spiciforme</i>	Plants
ASTERACEAE	<i>Hirpicium echinus</i>	Plants
ASTERACEAE	<i>Othonna ramulosa</i>	Plants
ASTERACEAE	<i>Pentzia globosa</i>	Plants
ASTERACEAE	<i>Senecio radicans</i>	Plants
ASTERACEAE	<i>Geigeria pectidea</i>	Plants
ASTERACEAE	<i>Senecio sophioides</i>	Plants
ASTERACEAE	<i>Dicoma schinzii</i>	Plants
ASTERACEAE	<i>Gazania krebsiana</i> subsp. <i>arctotoides</i>	Plants
ASTERACEAE	<i>Geigeria brevifolia</i>	Plants
ASTERACEAE	<i>Kleinia longiflora</i>	Plants
ASTERACEAE	<i>Parthenium</i> sp.	Plants
ASTERACEAE	<i>Pegolettia retrofracta</i>	Plants
ASTERACEAE	<i>Rosenia humilis</i>	Plants
ASTERACEAE	<i>Senecio inaequidens</i>	Plants
AYTONIACEAE	<i>Plagiochasma rupestre</i> var. <i>rupestre</i>	Plants
BIGNONIACEAE	<i>Rhigozum brevispinosum</i>	Plants
BIGNONIACEAE	<i>Rhigozum obovatum</i>	Plants
BIGNONIACEAE	<i>Rhigozum trichotomum</i>	Plants
BORAGINACEAE	<i>Ehretia rigida</i> subsp. <i>rigida</i>	Plants
BORAGINACEAE	<i>Ehretia</i> sp.	Plants
BORAGINACEAE	<i>Heliotropium ciliatum</i>	Plants
BORAGINACEAE	<i>Trichodesma africanum</i>	Plants
BORAGINACEAE	<i>Heliotropium nelsonii</i>	Plants
BRYACEAE	<i>Bryum capillare</i>	Plants
BRYACEAE	<i>Bryum dichotomum</i>	Plants
BUDDLEJACEAE	<i>Buddleja saligna</i>	Plants
CAMPANULACEAE	<i>Wahlenbergia</i> sp.	Plants
CAPPARACEAE	<i>Boscia foetida</i> subsp. <i>foetida</i>	Plants
CAPPARACEAE	<i>Cleome kalachariensis</i>	Plants
CAPPARACEAE	<i>Cleome angustifolia</i> subsp. <i>diandra</i>	Plants
CAPPARACEAE	<i>Cleome monophylla</i>	Plants
CAPPARACEAE	<i>Cleome rubella</i>	Plants
CARYOPHYLLACEAE	<i>Corrigiola litoralis</i> subsp. <i>litoralis</i> var. <i>litoralis</i>	Plants
CARYOPHYLLACEAE	<i>Pollichia campestris</i>	Plants
CELASTRACEAE	<i>Putterlickia pyracantha</i>	Plants
CELASTRACEAE	<i>Putterlickia saxatilis</i>	Plants
CELASTRACEAE	<i>Gymnosporia buxifolia</i>	Plants
CHENOPODIACEAE	<i>Chenopodium carinatum</i>	Plants
CHENOPODIACEAE	<i>Chenopodium hederiforme</i> var. <i>undulatum</i>	Plants
CHENOPODIACEAE	<i>Exomis microphylla</i> var. <i>axyrioides</i>	Plants
CHENOPODIACEAE	<i>Salsola</i> sp.	Plants
CHENOPODIACEAE	<i>Atriplex suberecta</i>	Plants
COLCHICACEAE	<i>Colchicum melanthoides</i> subsp. <i>melanthoides</i>	Plants
COLCHICACEAE	<i>Ornithoglossum vulgare</i>	Plants

COMBRETACEAE	<i>Terminalia sericea</i>	Plants
CONVOLVULACEAE	<i>Evolvulus alsinoides</i>	Plants
CONVOLVULACEAE	<i>Ipomoea bolusiana</i>	Plants
CONVOLVULACEAE	<i>Ipomoea oenotheroides</i>	Plants
CONVOLVULACEAE	<i>Convolvulus multifidus</i>	Plants
CONVOLVULACEAE	<i>Convolvulus ocellatus</i> var. <i>ocellatus</i>	Plants
CONVOLVULACEAE	<i>Ipomoea hackeliana</i>	Plants
CONVOLVULACEAE	<i>Xenostegia tridentata</i> subsp. <i>angustifolia</i>	Plants
CONVOLVULACEAE	<i>Convolvulus boedeckerianus</i>	Plants
CONVOLVULACEAE	<i>Ipomoea obscura</i> var. <i>obscura</i>	Plants
CONVOLVULACEAE	<i>Seddera capensis</i>	Plants
CUCURBITACEAE	<i>Coccinia rehmannii</i>	Plants
CUCURBITACEAE	<i>Kedrostis crassirostrata</i>	Plants
CUCURBITACEAE	<i>Momordica balsamina</i>	Plants
CUCURBITACEAE	<i>Peponium caledonicum</i>	Plants
CUCURBITACEAE	<i>Acanthosicyos naudinianus</i>	Plants
CUCURBITACEAE	<i>Corallocarpus triangularis</i>	Plants
CUCURBITACEAE	<i>Cucumis africanus</i>	Plants
CUCURBITACEAE	<i>Cucumis myriocarpus</i> subsp. <i>myriocarpus</i>	Plants
CYPERACEAE	<i>Bulbostylis hispidula</i> subsp. <i>pyriformis</i>	Plants
CYPERACEAE	<i>Cyperus congestus</i>	Plants
CYPERACEAE	<i>Isolepis sepulcralis</i>	Plants
CYPERACEAE	<i>Isolepis setacea</i>	Plants
CYPERACEAE	<i>Schoenoplectus muricinux</i>	Plants
CYPERACEAE	<i>Schoenoplectus muriculatus</i>	Plants
CYPERACEAE	<i>Cyperus atriceps</i>	Plants
CYPERACEAE	<i>Bulbostylis burchellii</i>	Plants
CYPERACEAE	<i>Bulbostylis hispidula</i>	Plants
CYPERACEAE	<i>Cyperus difformis</i>	Plants
CYPERACEAE	<i>Cyperus margaritaceus</i> var. <i>margaritaceus</i>	Plants
CYPERACEAE	<i>Cyperus marlothii</i>	Plants
CYPERACEAE	<i>Cyperus squarrosus</i>	Plants
CYPERACEAE	<i>Fuirena pubescens</i> var. <i>pubescens</i>	Plants
CYPERACEAE	<i>Lipocarpa rehmannii</i>	Plants
CYPERACEAE	<i>Cyperus decurvatus</i>	Plants
DIPSACACEAE	<i>Scabiosa buekiana</i>	Plants
DRACAENACEAE	<i>Sansevieria aethiopica</i>	Plants
EBENACEAE	<i>Euclea undulata</i>	Plants
EBENACEAE	<i>Diospyros lycioides</i> subsp. <i>lycioides</i>	Plants
ERIOSPERMACEAE	<i>Eriospermum corymbosum</i>	Plants
EUPHORBIACEAE	<i>Clutia affinis</i>	Plants
EUPHORBIACEAE	<i>Croton gratissimus</i> var. <i>gratissimus</i>	Plants
EUPHORBIACEAE	<i>Euphorbia avasmontana</i> var. <i>avasmontana</i>	Plants
EUPHORBIACEAE	<i>Euphorbia inaequilatera</i> var. <i>inaequilatera</i>	Plants
EUPHORBIACEAE	<i>Euphorbia juttae</i>	Plants
EUPHORBIACEAE	<i>Euphorbia rectirama</i>	Plants
EUPHORBIACEAE	<i>Euphorbia mauritanica</i> var. <i>mauritanica</i>	Plants

EUPHORBIACEAE	<i>Tragia physocarpa</i>	Plants
FABACEAE	<i>Acacia erioloba</i>	Plants
FABACEAE	<i>Acacia erioloba</i> E.Mey. x <i>A. haematoxylon</i> Willd.	Plants
FABACEAE	<i>Acacia haematoxylon</i>	Plants
FABACEAE	<i>Acacia hebeclada</i> subsp. <i>hebeclada</i>	Plants
FABACEAE	<i>Acacia karroo</i>	Plants
FABACEAE	<i>Crotalaria damarensis</i>	Plants
FABACEAE	<i>Cullen tomentosum</i>	Plants
FABACEAE	<i>Cyamopsis serrata</i>	Plants
FABACEAE	<i>Indigofera alternans</i> var. <i>alternans</i>	Plants
FABACEAE	<i>Indigofera daleoides</i> var. <i>daleoides</i>	Plants
FABACEAE	<i>Indigofera damarana</i>	Plants
FABACEAE	<i>Indigofera heterotricha</i>	Plants
FABACEAE	<i>Indigofera rhytidocarpa</i> subsp. <i>rhytidocarpa</i>	Plants
FABACEAE	<i>Indigofera sessilifolia</i>	Plants
FABACEAE	<i>Leobordea platycarpa</i>	Plants
FABACEAE	<i>Listia heterophylla</i>	Plants
FABACEAE	<i>Lotononis crumanina</i>	Plants
FABACEAE	<i>Lotononis parviflora</i>	Plants
FABACEAE	<i>Melolobium calycinum</i>	Plants
FABACEAE	<i>Melolobium candicans</i>	Plants
FABACEAE	<i>Melolobium canescens</i>	Plants
FABACEAE	<i>Melolobium humile</i>	Plants
FABACEAE	<i>Pomaria burchellii</i> subsp. <i>burchellii</i>	Plants
FABACEAE	<i>Ptycholobium biflorum</i> subsp. <i>biflorum</i>	Plants
FABACEAE	<i>Senna italica</i> subsp. <i>arachoides</i>	Plants
FABACEAE	<i>Sutherlandia frutescens</i>	Plants
FABACEAE	<i>Sutherlandia humilis</i>	Plants
FABACEAE	<i>Sutherlandia microphylla</i>	Plants
FABACEAE	<i>Tephrosia dregeana</i> var. <i>dregeana</i>	Plants
FABACEAE	<i>Acacia mellifera</i> subsp. <i>detinens</i>	Plants
FABACEAE	<i>Crotalaria spartioides</i>	Plants
FABACEAE	<i>Indigofera vicioides</i> var. <i>vicioides</i>	Plants
FABACEAE	<i>Lessertia pauciflora</i> var. <i>pauciflora</i>	Plants
FABACEAE	<i>Parkinsonia africana</i>	Plants
FABACEAE	<i>Requienia sphaerosperma</i>	Plants
FABACEAE	<i>Tephrosia burchellii</i>	Plants
FABACEAE	<i>Calobota spinescens</i>	Plants
FABACEAE	<i>Indigofera alternans</i>	Plants
FABACEAE	<i>Calobota linearifolia</i>	Plants
FABACEAE	<i>Crotalaria orientalis</i> subsp. <i>orientalis</i>	Plants
FABACEAE	<i>Crotalaria podocarpa</i>	Plants
FABACEAE	<i>Elephantorrhiza elephantina</i>	Plants
FABACEAE	<i>Indigastrum argyraeum</i>	Plants
FABACEAE	<i>Indigofera</i> sp.	Plants
FABACEAE	<i>Melolobium exudans</i>	Plants
FABACEAE	<i>Melolobium macrocalyx</i> var. <i>macrocalyx</i>	Plants

FABACEAE	Melolobium villosum	Plants
FABACEAE	Rhynchosia confusa	Plants
FABACEAE	Rhynchosia totta var. totta	Plants
FABACEAE	Rhynchosia venulosa	Plants
FABACEAE	Tephrosia longipes subsp. longipes var. longipes	Plants
FABACEAE	Tephrosia purpurea subsp. leptostachya var. leptostachya	Plants
FABACEAE	Calobota cuspidosa	Plants
FISSIDENTACEAE	Fissidens sciophyllus	Plants
FISSIDENTACEAE	Fissidens submarginatus	Plants
GERANIACEAE	Monsonia luederitziana	Plants
GISEKIACEAE	Gisekia africana var. africana	Plants
GISEKIACEAE	Gisekia pharnacioides var. pharnacioides	Plants
GISEKIACEAE	Gisekia africana var. pedunculata	Plants
HYACINTHACEAE	Ledebouria undulata	Plants
HYACINTHACEAE	Dipcadi gracillimum	Plants
HYACINTHACEAE	Drimia physodes	Plants
HYACINTHACEAE	Dipcadi crispum	Plants
HYACINTHACEAE	Albuca sp.	Plants
IRIDACEAE	Moraea pallida	Plants
IRIDACEAE	Lapeirousia littoralis subsp. littoralis	Plants
IRIDACEAE	Ferraria glutinosa	Plants
IRIDACEAE	Lapeirousia littoralis subsp. caudata	Plants
JUNCACEAE	Juncus dregeanus subsp. dregeanus	Plants
LAMIACEAE	Acrotome inflata	Plants
LAMIACEAE	Ocimum americanum var. americanum	Plants
LAMIACEAE	Salvia namaensis	Plants
LAMIACEAE	Salvia verbenaca	Plants
LAMIACEAE	Stachys burchelliana	Plants
LAMIACEAE	Leucas capensis	Plants
LAMIACEAE	Stachys spathulata	Plants
LOBELIACEAE	Lobelia erinus	Plants
LOPHIOCARPACEAE	Corbichonia decumbens	Plants
LOPHIOCARPACEAE	Lophiocarpus polystachyus	Plants
LORANTHACEAE	Tapinanthus oleifolius	Plants
LORANTHACEAE	Tapinanthus forbesii	Plants
MALVACEAE	Abutilon austro-africanum	Plants
MALVACEAE	Grewia flava	Plants
MALVACEAE	Hermannia burkei	Plants
MALVACEAE	Hermannia comosa	Plants
MALVACEAE	Hermannia desertorum	Plants
MALVACEAE	Hermannia erodioides	Plants
MALVACEAE	Hermannia quartiniana	Plants
MALVACEAE	Hermannia sp.	Plants
MALVACEAE	Hermannia vestita	Plants
MALVACEAE	Malva pusilla	Plants
MALVACEAE	Melhania rehmannii	Plants
MALVACEAE	Sida cordifolia subsp. cordifolia	Plants

MALVACEAE	Waltheria indica	Plants
MALVACEAE	Hermannia abrotanoides	Plants
MALVACEAE	Hermannia burchellii	Plants
MALVACEAE	Hermannia linearifolia	Plants
MALVACEAE	Hermannia pulverata	Plants
MALVACEAE	Hermannia spinosa	Plants
MALVACEAE	Hermannia tomentosa	Plants
MALVACEAE	Radyera urens	Plants
MALVACEAE	Sida chrysantha	Plants
MALVACEAE	Hermannia jacobefolia	Plants
MALVACEAE	Hermannia linnaeoides	Plants
MALVACEAE	Hibiscus ludwigii	Plants
MALVACEAE	Hibiscus pusillus	Plants
MALVACEAE	Melhania burchellii	Plants
MALVACEAE	Melhania virescens	Plants
MALVACEAE	Pavonia burchellii	Plants
MALVACEAE	Sida ovata	Plants
MALVACEAE	Sida pseudocordifolia	Plants
MELIACEAE	Nymania capensis	Plants
MENISPERMACEAE	Cissampelos capensis	Plants
MESEMBRYANTHEMACEAE	Mestoklema arboriforme	Plants
MESEMBRYANTHEMACEAE	Ruschia sp.	Plants
MESEMBRYANTHEMACEAE	Trichodiadema pomeridianum	Plants
MOLLUGINACEAE	Limeum myosotis var. myosotis	Plants
MOLLUGINACEAE	Limeum viscosum subsp. transvaalense	Plants
MOLLUGINACEAE	Mollugo cerviana var. cerviana	Plants
MOLLUGINACEAE	Limeum aethiopicum subsp. aethiopicum var. aethiopicum	Plants
MOLLUGINACEAE	Limeum argute-carinatum var. argute-carinatum	Plants
MOLLUGINACEAE	Limeum aethiopicum var. intermedium	Plants
MORACEAE	Ficus cordata subsp. cordata	Plants
NEURADACEAE	Grielum humifusum var. parviflorum	Plants
NYCTAGINACEAE	Phaeoptilum spinosum	Plants
ORCHIDACEAE	Disperis macowanii	Plants
OXALIDACEAE	Oxalis lawsonii	Plants
OXALIDACEAE	Oxalis haedulipes	Plants
PEDALIACEAE	Sesamum capense	Plants
PEDALIACEAE	Harpagophytum procumbens subsp. procumbens	Plants
PEDALIACEAE	Sesamum triphyllum var. triphyllum	Plants
PHYLLANTHACEAE	Phyllanthus maderaspatensis	Plants
PHYLLANTHACEAE	Phyllanthus parvulus var. parvulus	Plants
PLUMBAGINACEAE	Dyerophytum africanum	Plants
POACEAE	Agrostis lachnantha var. lachnantha	Plants
POACEAE	Anthepphora pubescens	Plants
POACEAE	Aristida congesta subsp. barbicollis	Plants
POACEAE	Aristida congesta subsp. congesta	Plants
POACEAE	Aristida diffusa subsp. burkei	Plants
POACEAE	Aristida engleri var. engleri	Plants

POACEAE	<i>Aristida vestita</i>	Plants
POACEAE	<i>Brachiaria marlothii</i>	Plants
POACEAE	<i>Brachiaria nigropedata</i>	Plants
POACEAE	<i>Cenchrus ciliaris</i>	Plants
POACEAE	<i>Chloris virgata</i>	Plants
POACEAE	<i>Cynodon dactylon</i>	Plants
POACEAE	<i>Cynodon incompletus</i>	Plants
POACEAE	<i>Diandrochloa namaquensis</i>	Plants
POACEAE	<i>Diandrochloa pusilla</i>	Plants
POACEAE	<i>Digitaria eriantha</i>	Plants
POACEAE	<i>Digitaria glauca</i> var. <i>bechuanica</i>	Plants
POACEAE	<i>Digitaria seriata</i>	Plants
POACEAE	<i>Echinochloa</i> sp.	Plants
POACEAE	<i>Enneapogon scaber</i>	Plants
POACEAE	<i>Enneapogon scoparius</i>	Plants
POACEAE	<i>Eragrostis curvula</i>	Plants
POACEAE	<i>Eragrostis echinochloidea</i>	Plants
POACEAE	<i>Eragrostis gummiflua</i>	Plants
POACEAE	<i>Eragrostis lehmanniana</i> var. <i>lehmanniana</i>	Plants
POACEAE	<i>Eragrostis nindensis</i>	Plants
POACEAE	<i>Eragrostis obtusa</i>	Plants
POACEAE	<i>Eragrostis porosa</i>	Plants
POACEAE	<i>Eragrostis rigidior</i>	Plants
POACEAE	<i>Eragrostis rotifer</i>	Plants
POACEAE	<i>Eragrostis trichophora</i>	Plants
POACEAE	<i>Eragrostis</i> x <i>pseud-obtusa</i>	Plants
POACEAE	<i>Hyparrhenia hirta</i>	Plants
POACEAE	<i>Melinis nerviglumis</i>	Plants
POACEAE	<i>Melinis repens</i> subsp. <i>repens</i>	Plants
POACEAE	<i>Oropetium capense</i>	Plants
POACEAE	<i>Panicum gilvum</i>	Plants
POACEAE	<i>Panicum impeditum</i>	Plants
POACEAE	<i>Panicum kalaharensis</i>	Plants
POACEAE	<i>Panicum schinzii</i>	Plants
POACEAE	<i>Pogonarthria squarrosa</i>	Plants
POACEAE	<i>Schmidtia kalahariensis</i>	Plants
POACEAE	<i>Schmidtia pappophoroides</i>	Plants
POACEAE	<i>Stipagrostis ciliata</i> var. <i>capensis</i>	Plants
POACEAE	<i>Stipagrostis uniplumis</i> var. <i>uniplumis</i>	Plants
POACEAE	<i>Tragus berteronianus</i>	Plants
POACEAE	<i>Tragus koelerioides</i>	Plants
POACEAE	<i>Trichoneura grandiglumis</i>	Plants
POACEAE	<i>Urochloa panicoides</i>	Plants
POACEAE	<i>Anthephora argentea</i>	Plants
POACEAE	<i>Aristida meridionalis</i>	Plants
POACEAE	<i>Centropodia glauca</i>	Plants
POACEAE	<i>Enneapogon desvauxii</i>	Plants

POACEAE	<i>Eragrostis annulata</i>	Plants
POACEAE	<i>Eragrostis brizantha</i>	Plants
POACEAE	<i>Eragrostis cylindriflora</i>	Plants
POACEAE	<i>Eragrostis lehmanniana</i> var. <i>chaunantha</i>	Plants
POACEAE	<i>Eragrostis truncata</i>	Plants
POACEAE	<i>Merxmuellera</i> sp.	Plants
POACEAE	<i>Sporobolus</i> sp.	Plants
POACEAE	<i>Stipagrostis obtusa</i>	Plants
POACEAE	<i>Tragus racemosus</i>	Plants
POACEAE	<i>Triraphis purpurea</i>	Plants
POACEAE	<i>Sporobolus ioclados</i>	Plants
POACEAE	<i>Andropogon chinensis</i>	Plants
POACEAE	<i>Andropogon schirensis</i>	Plants
POACEAE	<i>Aristida adscensionis</i>	Plants
POACEAE	<i>Aristida engleri</i> var. <i>ramosissima</i>	Plants
POACEAE	<i>Aristida stipitata</i> subsp. <i>spicata</i>	Plants
POACEAE	<i>Brachiaria brizantha</i>	Plants
POACEAE	<i>Cymbopogon caesius</i>	Plants
POACEAE	<i>Cymbopogon pospischilii</i>	Plants
POACEAE	<i>Digitaria polyphylla</i>	Plants
POACEAE	<i>Digitaria sanguinalis</i>	Plants
POACEAE	<i>Eleusine coracana</i> subsp. <i>africana</i>	Plants
POACEAE	<i>Elionurus muticus</i>	Plants
POACEAE	<i>Enneapogon cenchroides</i>	Plants
POACEAE	<i>Eragrostis barrelieri</i>	Plants
POACEAE	<i>Eragrostis biflora</i>	Plants
POACEAE	<i>Eragrostis mexicana</i> subsp. <i>virescens</i>	Plants
POACEAE	<i>Eragrostis pallens</i>	Plants
POACEAE	<i>Eragrostis viscosa</i>	Plants
POACEAE	<i>Eustachys paspaloides</i>	Plants
POACEAE	<i>Fingerhuthia africana</i>	Plants
POACEAE	<i>Heteropogon contortus</i>	Plants
POACEAE	<i>Lamarckia aurea</i>	Plants
POACEAE	<i>Leptochloa fusca</i>	Plants
POACEAE	<i>Melinis repens</i> subsp. <i>grandiflora</i>	Plants
POACEAE	<i>Panicum coloratum</i> var. <i>coloratum</i>	Plants
POACEAE	<i>Panicum maximum</i>	Plants
POACEAE	<i>Setaria verticillata</i>	Plants
POACEAE	<i>Sporobolus fimbriatus</i>	Plants
POACEAE	<i>Stipagrostis uniplumis</i> var. <i>neesii</i>	Plants
POACEAE	<i>Urochloa stolonifera</i>	Plants
POLYGALACEAE	<i>Muraltia alopecuroides</i>	Plants
POLYGALACEAE	<i>Polygala leptophylla</i> var. <i>leptophylla</i>	Plants
POLYGALACEAE	<i>Polygala leptophylla</i> var. <i>armata</i>	Plants
POLYGALACEAE	<i>Polygala seminuda</i>	Plants
POLYGONACEAE	<i>Emex australis</i>	Plants
POLYGONACEAE	<i>Persicaria limbata</i>	Plants

PORTULACACEAE	<i>Talinum arnotii</i>	Plants
PORTULACACEAE	<i>Talinum caffrum</i>	Plants
PORTULACACEAE	<i>Talinum crispatum</i>	Plants
PORTULACACEAE	<i>Portulaca hereroensis</i>	Plants
PORTULACACEAE	<i>Portulaca kermesina</i>	Plants
POTTIACEAE	<i>Trichostomum brachydontium</i>	Plants
PROTEACEAE	<i>Leucadendron rubrum</i>	Plants
PTERIDACEAE	<i>Cheilanthes hirta</i> var. <i>brevipilosa</i>	Plants
PTERIDACEAE	<i>Cheilanthes multifida</i> var. <i>multifida</i>	Plants
PTERIDACEAE	<i>Pellaea calomelanos</i> var. <i>calomelanos</i>	Plants
RANUNCULACEAE	<i>Clematis brachiata</i>	Plants
RHAMNACEAE	<i>Helinus spartioides</i>	Plants
RHAMNACEAE	<i>Ziziphus mucronata</i> subsp. <i>mucronata</i>	Plants
RHAMNACEAE	<i>Phyllia</i> sp.	Plants
RICCIACEAE	<i>Riccia cavernosa</i>	Plants
RICCIACEAE	<i>Riccia okahandjana</i>	Plants
RICCIACEAE	<i>Riccia cupulifera</i>	Plants
RUBIACEAE	<i>Anthospermum rigidum</i> subsp. <i>rigidum</i>	Plants
RUBIACEAE	<i>Kohautia cynanchica</i>	Plants
RUBIACEAE	<i>Kohautia virgata</i>	Plants
SANTALACEAE	<i>Thesium hystricoides</i>	Plants
SANTALACEAE	<i>Thesium lineatum</i>	Plants
SANTALACEAE	<i>Thesium hystrix</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum junceum</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia integerrima</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia</i> sp.	Plants
SCROPHULARIACEAE	<i>Peliostomum leucorrhizum</i>	Plants
SCROPHULARIACEAE	<i>Selago mixta</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum albomarginatum</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum marlothii</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia atropurpurea</i> subsp. <i>pubescens</i>	Plants
SCROPHULARIACEAE	<i>Peliostomum organoides</i>	Plants
SCROPHULARIACEAE	<i>Selago divaricata</i>	Plants
SCROPHULARIACEAE	<i>Selago welwitschii</i> var. <i>australis</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum elongatum</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum spinescens</i>	Plants
SCROPHULARIACEAE	<i>Aptosimum lineare</i> var. <i>lineare</i>	Plants
SCROPHULARIACEAE	<i>Jamesbrittenia atropurpurea</i> subsp. <i>atropurpurea</i>	Plants
SCROPHULARIACEAE	<i>Sutera griquensis</i>	Plants
SOLANACEAE	<i>Lycium cinereum</i>	Plants
SOLANACEAE	<i>Lycium hirsutum</i>	Plants
SOLANACEAE	<i>Withania somnifera</i>	Plants
SOLANACEAE	<i>Lycium pumilum</i>	Plants
SOLANACEAE	<i>Solanum burchellii</i>	Plants
SOLANACEAE	<i>Solanum lichtensteinii</i>	Plants
SOLANACEAE	<i>Solanum panduriforme</i>	Plants
SOLANACEAE	<i>Solanum supinum</i> var. <i>supinum</i>	Plants

SOLANACEAE	<i>Solanum tomentosum</i> var. <i>tomentosum</i>	Plants
TAMARICACEAE	<i>Tamarix parviflora</i>	Plants
THYMELAEACEAE	<i>Gnidia kraussiana</i> var. <i>kraussiana</i>	Plants
THYMELAEACEAE	<i>Gnidia polycephala</i>	Plants
URTICACEAE	<i>Laportea peduncularis</i> subsp. <i>peduncularis</i>	Plants
VAHLIACEAE	<i>Vahlia capensis</i> subsp. <i>vulgaris</i> var. <i>linearis</i>	Plants
VAHLIACEAE	<i>Vahlia capensis</i> subsp. <i>vulgaris</i> var. <i>vulgaris</i>	Plants
VERBENACEAE	<i>Chascanum pinnatifidum</i> var. <i>pinnatifidum</i>	Plants
VERBENACEAE	<i>Lantana rugosa</i>	Plants
VERBENACEAE	<i>Chascanum adenostachyum</i>	Plants
VERBENACEAE	<i>Chascanum hederaceum</i> var. <i>hederaceum</i>	Plants
VERBENACEAE	<i>Chascanum schlechteri</i>	Plants
VISCACEAE	<i>Viscum rotundifolium</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus terrestris</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum flexuosum</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum incrustatum</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus zeyheri</i> subsp. <i>zeyheri</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum leptopetalum</i>	Plants
ZYGOPHYLLACEAE	<i>Tribulus excrucians</i>	Plants
ZYGOPHYLLACEAE	<i>Zygophyllum pubescens</i>	Plants

<u>FamilyName</u>	<u>SpeciesName</u>	<u>Category</u>
Agamidae	Agama atra	Lizards
Agamidae	Agama aculeata	Lizards
Apidae	Amegilla niveata	Animals
Apidae	Allodapula sp.	Animals
Apidae	Chalicodoma sp	Animals
Apidae	Heriades sp	Animals
Apidae	Meliturgula sp.	Animals
Apidae	Nomia sp	Animals
Apidae	Nomioides sp	Animals
Apidae	Pasites appletoni	Animals
Apidae	Unknown	Animals
Chrysopidae	Italochrysa gigantia	Animals
Colletidae	Colletes capensis	Animals
Colletidae	Colletes fascicularis	Animals
Hesperiidae	Kedestes lepenula	Animals
HESPERIIDAE	Spialia secessus	Animals
Lacertidae	Heliobolus lugubris	Animals
Lacertidae	Pedioplanis lineoocellata	Animals
Lycaenidae	Aloeides damarensis damarensis	Animals
Lycaenidae	Aloeides molomo molomo	Animals
Lycaenidae	Cigaritis ella	Animals
Lycaenidae	Crudaria leroma	Animals
Lycaenidae	Tylopaedia sardonix peringueyi	Animals
LYCAENIDAE	Aloeides damarensis subsp. mashona	Animals
LYCAENIDAE	Aloeides molomo subsp. krooni	Animals
LYCAENIDAE	Stugeta bowkeri subsp. bowkeri	Animals
Nemopteridae	Nemopterinae	Animals
NYMPHALIDAE	Acraea stenobea	Animals
NYMPHALIDAE	Vanessa cardui	Animals
Pieridae	Belenois aurota aurota	Animals
Pieridae	Colotis agoye bowkeri	Animals
Scarabaeidae	Pachynema crassipes	Animals
Scincidae	Mabuya spilogaster	Animals
Scincidae	Mabuya variegata	Animals
Testudinidae	Psammobates oculiferus	Animals