

## Anticancer Activity of Tirucallane Triterpenoids from *Amoora dasyclada*

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A new tetranortriterpene 3 $\alpha$ -acetoxy-24,25,26,27-tetranortirucalla-7-ene-23(21)-lactone (**3**), and eleven other compounds were isolated from the twigs of *Amoora dasyclada*. The structure of compound **3** was identified on the basis of spectroscopic data, and the bioactive experiments of **1** and **3–5** against AGZY 83-a (human lung cancer cells) and SMMC-7721 (human liver cancer cells) are documented. Among them, compound **5** exhibited a strong activity against SMMC-7721.

*Key words:* *Amoora dasyclada*, Tirucallane Triterpenoid, Anticancer Activity

### Introduction

In previous papers, we reported four new tirucallane-type triterpenoids (**1**, **2**, **4**, **5**) from the twigs of *Amoora dasyclada* (How et T. Chen) C. Y. Wu (Yang *et al.*, 2004a, b); here we present another tirucallane-type triterpenoid obtained during our continuing study on the same plant: 3 $\alpha$ -acetoxy-24,25,26,27-tetranortirucalla-7-ene-23(21)-lactone (**3**). Seven other compounds, taraxerone (**6**), taraxerol (**7**) and taraxerol acetate (**8**), scopoletin (**9**), stigmast-5-en-3 $\beta$ ,7 $\alpha$ -diol (**10**),  $\beta$ -sitosterol (**11**),  $\beta$ -sitosterol-D-glucoside (**12**), were isolated from the same source. The bioactive experiments of **1** and **3–5** against AGZY 83-a (human lung cancer cells) and SMMC-7721 (human liver cancer cells) were also assayed. Among them, compound **5** exhibited a strong activity against SMMC-7721 with the IC<sub>50</sub> value of  $8.41 \times 10^{-3}$   $\mu$ M/ml.

### Results and Discussion

Compound **3** (white needles) has a molecular formula of C<sub>28</sub>H<sub>42</sub>O<sub>4</sub> established by HR-ESI-MS (*m/z* 465.2989 [M + Na]<sup>+</sup>). The <sup>1</sup>H and <sup>13</sup>C NMR spectra of it were in good agreement with those of **2** indicating that **3** was also a tirucallane derivative (Yang *et al.*, 2004a). The IR spectrum showed the presence of a  $\gamma$ -lactone group (1784 cm<sup>-1</sup>); it was further confirmed by the long correlations between  $\delta_{\text{H}}$  4.37 (1H, t, *J* = 8.7 Hz, H-21 $\alpha$ ); 3.90 (1H,

t, *J* = 9.3 Hz, H-21 $\beta$ ) with  $\delta_{\text{C}}$  176.9 (s, C-23), 39.2 (d, C-20) and 34.2 (t, C-22);  $\delta_{\text{H}}$  2.52 (1H, dd, *J* = 19.0, 6.5 Hz, H-22 $\alpha$ ); 2.17 (1H, dd, *J* = 18.3, 13.8 Hz, H-22 $\beta$ ) with  $\delta_{\text{C}}$  72.4 (t, C-21), C-20 and C-23. The signals at  $\delta_{\text{H}}$  2.52, 2.17 showed a large coupling constant of 13.8 Hz and a small coupling constant of 6.5 Hz, respectively, which revealed axial orientations for H-20 and H-22 $\alpha$ . Comparing the <sup>1</sup>H and <sup>13</sup>C NMR spectra of **3** with those of **2**, except the  $\gamma$ -lactone group [ $\delta_{\text{C}}$  176.9 (s, C-23)], there was an acetoxy group in **3** [ $\delta_{\text{C}}$  170.7(s), 21.3 (q)], and the signals at  $\delta_{\text{H}}$  2.04 (3H, s) and 4.66 (bs) also indicated that an  $\alpha$ -acetoxy group was located at C-3 instead of a hydroxy group in **2**. The cross peaks between  $\delta_{\text{H}}$  2.04 (3H, s) and  $\delta_{\text{C}}$  170.7 (s, CH<sub>3</sub>COO),  $\delta_{\text{H}}$  4.66 (bs) and  $\delta_{\text{C}}$  170.7, 33.6 (s, C-4), 45.6 (d, C-5), 22.9 (t, C-2), 31.8 (t, C-1) in the HMBC spectrum supported this assumption.

The strongly negative optical rotation of **3** (–42.13°) suggested that it belongs to the tirucallanes (C-20 $\alpha$ ) (Sherman *et al.*, 1980; Jolad *et al.*, 1981). In the ROESY spectrum the strong cross peaks between  $\delta_{\text{H}}$  4.37 (H-21 $\alpha$ ) and 3.90 (H-21 $\beta$ ) with 1.35 (H-12a),  $\delta_{\text{H}}$  4.37 (H-21 $\alpha$ ) with 0.86 (Me-18),  $\delta_{\text{H}}$  3.90 (H-21 $\beta$ ) with 1.73 (1H, m, H-17) and 1.71 (H-12b) also indicated that **3** preferred H-20 $\alpha$  (C-20S) configuration to C-20R configuration proving the tirucallane-type triterpene (Mohamad *et al.*, 1999; Wang *et al.*, 2003). So compound **3** was a biodegraded product of tirucallane

with the loss of four carbon atoms at the side-chain and it was determined to be 3 $\alpha$ -acetoxy-24,25,26,27-tetranortirucalla-7-ene-23(21)-lactone (Fig. 1).

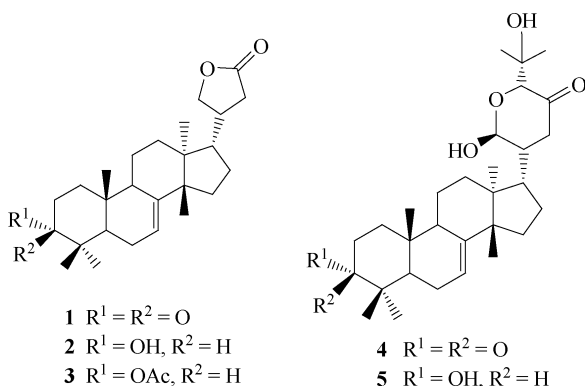


Fig. 1. The structures of compounds 1–5.

## Experimental

### General

Melting points: XRC-1 apparatus (Sichuan University, Sichuan, P.R. China), uncorrected. Optical rotations: Horiba SEAP-300 polarimeter (Kyoto, Japan). IR spectra: Bio-Rad (Richmond, CA, USA) FTS-135 infrared spectrophotometer. One- and two-dimensional NMR spectra: Bruker AM-400 or DRX-500 spectrometers (Karlsruhe, Germany). MS data: VG Autospec-3000 spectrometer (Manchester, England).

### Plant material

The twigs of *A. dasyclada* were collected in Xishuangbanna County of Yunnan Province, P.R. China, in January 2002. The plant was identified by Mr. Jingyun Cui, Xishuangbanna Tropical Botanical Garden, CAS, P.R. China.

### Extraction and isolation

The first step of the isolation was the same as previously described (Yang *et al.*, 2004b). Then fraction 2 was repeatedly chromatographed by CC over silica gel eluted with petroleum ether/EtOAc (from 1:0 to 8:2, v/v) to give compounds 6 (191 mg) and 8 (7 mg); fraction 4 was subject to repeated CC on silica gel eluted with petroleum ether/Me<sub>2</sub>CO (from 98:2 to 7:3) to obtain compounds 1 (440 mg), 3 (19 mg), 7 (140 mg) and 11 (2.1 g); fraction 5 was submitted to repeated CC on silica gel eluted with CHCl<sub>3</sub>/EtOAc (from 95:5 to 3:1) and then purified on a RP-18 column eluted with MeOH/H<sub>2</sub>O (from 1:1 to 1:0) to yield compounds 2 (20 mg) and 10 (17 mg); fraction 8 was repeatedly chromatographed by CC over silica gel eluted with CHCl<sub>3</sub>/Me<sub>2</sub>CO (from 95:5 to 3:1) and then purified on a RP-18 and Sephadex LH-20 column successively to afford compound 9 (21 mg); fraction 9 was repeatedly chromatographed over silica gel eluted with CHCl<sub>3</sub>/MeOH (from 95:5 to 1:1) to produce compound 12 (610 mg).

### Bioassays

An improved MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] colorimetric assay was performed in 96-well plates; the experimental details were like reported previously (Niu *et al.*, 2002).

The results of anticancer activity tests of the four tirucallane triterpenoids 1 and 3–5 against AGZY 83-a (human lung cancer cells) and SMMC-7721 (human liver cancer cells) are given in Table I. In the test, 1 and 3 showed inactivity to these two cell lines, 4 and 5 exerted weak activity against AGZY 83-a, 5 exhibited strong activity against SMMC-7721.

3 $\alpha$ -Acetoxy-24,25,26,27-tetranortirucalla-7-ene-23(21)-lactone (3): White needles. – M.p. 214–

Table I. Cytotoxicity<sup>a</sup> of compounds 1 and 3–5.

	<i>cis</i> -Platin <sup>c</sup>	IC <sub>50</sub> [ $\mu$ M/ml] <sup>b</sup>			
		1	3	4	5
AGZY 83-a	$5.673 \times 10^{-3}$	no activity	no activity	$0.065 \pm 0.013$	$0.050 \pm 0.005$
SMMC-7721	$3.947 \times 10^{-3}$	no activity	$0.171 \pm 0.044$	no activity	$0.018 \pm 0.005$

<sup>a</sup> AGZY 83-a, human lung cancer cells; SMMC-7721, human liver cancer cells.

<sup>b</sup> The IC<sub>50</sub> values are presented as means  $\pm$  SE.

<sup>c</sup> *cis*-Platin as positive control.

216 °C. –  $[\alpha]_D^{25}$  –42.13° (*c* 0.178, CHCl<sub>3</sub>). – IR (KBr):  $\nu$  = 2922, 1781, 1728, 1631, 1462, 1373, 1248, 1174, 1101, 1032, 1019 cm<sup>-1</sup>. – <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  = 5.26 (1H, d, *J* = 2.9 Hz, H-7), 4.37 (1H, t, *J* = 8.7 Hz, H-21 $\alpha$ ), 3.90 (1H, t, *J* = 9.3 Hz, H-21 $\beta$ ), 2.55 (1H, m, H-20), 2.52 (1H, dd, *J* = 19.0, 6.5 Hz, H-22 $\alpha$ ), 2.30 (1H, m, H-9), 2.17 (1H, dd, *J* = 18.3, 13.8 Hz, H-22 $\beta$ ), 2.05, 1.95 (each 1H, m, H-6), 2.04 (3H, s, CH<sub>3</sub>COO), 1.91, 1.32 (1H, m, H-16), 1.85, 1.67 (each 1H, m, H-2), 1.75 (1H, m, H-5), 1.73 (1H, m, H-17), 1.72, 1.38 (2H, m, H-1), 1.71, 1.35 (each 1H, m, H-12), 1.64–1.48 (4H, m, H-11 and H-15), 0.97 (3H, s, Me-30), 0.94 (3H, s, Me-29), 0.86 (3H, s, Me-18), 0.82 (3H, s, Me-28), 0.76 (3H, s, Me-19). – <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  = 31.8 (t, C-1), 22.9 (t, C-2), 78.2 (d, C-3), 36.6 (s, C-4), 45.6 (d, C-5), 23.8 (t, C-6), 118.6 (d, C-7), 144.9 (s, C-8), 48.4 (d, C-9), 34.9 (s, C-10), 17.3 (t, C-11), 31.9 (t, C-12), 43.7 (s, C-13), 50.6 (s, C-14), 34.1 (t, C-15), 27.3 (t,

C-16), 51.0 (d, C-17), 22.6 (q, C-18), 12.9 (q, C-19), 39.2 (d, C-20), 72.4 (t, C-21), 34.6 (t, C-22), 176.9 (s, C-23), 27.4 (q, C-28), 21.4 (q, C-29), 27.0 (q, C-30), 170.7 (s, CH<sub>3</sub>COO), 21.3 (q, CH<sub>3</sub>COO). – EI-MS: *m/z* = 442 (17, [M]<sup>+</sup>), 426 (13), 382 (9), 367 (100), 324 (3), 297 (4), 259 (13), 245 (6), 213 (5), 187 (13), 159 (11), 147 (9), 119 (12), 105 (9), 81 (5). – HR-ESI-MS: *m/z* = 465.2989 [M + Na]<sup>+</sup> (calcd. for C<sub>28</sub>H<sub>42</sub>O<sub>4</sub>Na, 465.2980).

Three known compounds **6–8** were identified by spectral analysis results and by comparison with the published data (Sakurai *et al.*, 1987). The *R<sub>f</sub>* values of **9–12** were coincident with the standard samples in different developing solvents.

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Jolad S. D., Hoffmann J. H., Schram K. H., and Cole J. K. (1981), Constituents of *Trichilia hispida* (Meliaceae). 4. Hispidals A and B, two new tirucallane triterpenoids. *J. Org. Chem.* **46**, 4085–4088.

Mohamad K., Martin M.-T., Litaudon M., Gaspard C., Sévenet T., and Païs M. (1999), Tirucallane triterpenes from *Dysoxylum macranthum*. *Phytochemistry* **52**, 1461–1468.

Niu X.-M., Li S.-H., Li M.-L., Zhao Q.-S., Mei S.-X., Wang S.-J., Lin Z.-W., and Sun H.-D. (2002), Cytotoxic ent-kaurane diterpenoids from *Isodon eriocalyx* var. *laxiflora*. *Planta Med.* **68**, 528–533.

Sakurai N., Yaguchi Y., and Inoue T. (1987), Triterpenoids from *Myrica rubra*. *Phytochemistry* **26**, 217–219.

Sherman M. M., Borris R. P., Ogura M., Cordell O. G., and Farnsworth N. R. (1980), 3*S*,24*S*,25-Trihydroxy-tirucall-7-ene from *Ailanthus excelsa*. *Phytochemistry* **19**, 1499–1501.

Wang L. Y., Wang N. L., Yao X. S., Miyata S., and Kitahara S. (2003), Euphane and tirucallane triterpene from the roots of *Euphane kansui* and their *in vitro* effects on the cell division of *Xenopus*. *J. Nat. Prod.* **66**, 630–633.

Yang S.-M., Ma Y.-B., Luo X.-D., Wu S.-H., and Wu D.-G. (2004a), Two new tetranortriterpenes from *Amoora dasyclada*. *Chin. Chem. Lett.* **15**, 1187–1190.

Yang S.-M., Ding L., Wu S.-H., Ma Y.-B., Luo X.-D., and Wu D.-G. (2004b), Two new tirucallane triterpenes with six-membered hemiacetal from *Amoora dasyclada*. *Z. Naturforsch.* **59b**, 1627–1629.