



# Integrative Organismal Biology

A Journal of the Society  
for Integrative and  
Comparative Biology

[academic.oup.com/icb](https://academic.oup.com/icb)



OXFORD  
UNIVERSITY PRESS



## REVIEW ARTICLE

### On the Success of the Hadal Snailfishes

M. E. Gerringer<sup>1</sup>

Friday Harbor Laboratories, University of Washington, Friday Harbor, WA 98250, USA

<sup>1</sup>E-mail: mgerringer@uw.edu

**Synopsis** Deep-sea trenches, depths 6000–11,000 m, are characterized by high pressures, low temperatures, and absence of sunlight. These features make up the majority of the deepest marine habitat—the hadal zone—home to distinct communities from those in the surrounding abyssal plains. The snailfishes, family Liparidae (Scorpaeniformes), have found notable success in the hadal zone from ~6000 to 8200 m, comprising the dominant ichthyofauna in at least six trenches worldwide. The hadal fish community is distinct from the abyssal community where elongate, scavenging fishes such as rattails (Macrouridae), cutthroat eels (Synphobranchidae), tripodfishes (Ipnopidae), eelpouts (Zoarcidae), and cusk eels (Ophidiidae) are most common. Until recently, little was known about the biology of these deepest-living fishes, or the factors that drive their success at hadal depths. Here, I review recent investigations spanning the abyssal–hadal boundary and discuss the factors structuring these communities, including the roles of pressure adaptation, feeding ecology, and life history. Hadal fishes show specialized adaptation to hydrostatic pressure both in accumulation of the pressure-counteractant trimethylamine *n*-oxide and in intrinsic changes to enzymes. Stomach content and amino acid isotope analyses, and jaw morphology suggest that suction-feeding predatory fishes like hadal liparids may find an advantage to descending into the trench where amphipods are increasingly abundant. Analysis of otolith growth zones suggest that snailfishes may be adapted to a seismically active, high-disturbance hadal environment by having relatively short life-spans. This review synthesizes the known literature on the planet’s deepest-living fishes and informs new understanding of adaptations to life in the trenches.

**Synopsis** Sobre el éxito de los Peces Caracol  
Las fosas marinas con profundidades de 6000 a 11.000 m se caracterizan por altas presiones, bajas temperaturas, y ausencia de luz solar. Estas características constituyen la mayoría de los hábitats marinos más profundos, la zona hadal, que albergan distintas comunidades que rodean a las planicies abisales. Los peces caracol de la familia Liparidae (Scorpaeniformes) conforman la ictiofauna dominante en zonas de 6000 a 8200 m y se encuentran en al menos 6 fosas a nivel mundial. La comunidad de peces caracol es distinta de la comunidad abisal en esta última los peces tienen cuerpos más elongados y suelen ser saprófitos como los peces cola de rata (Macrouridae), las anguilas de las familias Synphobranchidae, Zoarcidae, Ophidiidae. Hasta hace poco, la información acerca de la biología de estos peces de profundidad era casi nula así como los factores que impulsan su éxito en las profundidades. El objetivo de este artículo es hacer una revisión de las investigaciones recientes que abarcan el límite abisal-hadal y discutir los factores que estructuran estas comunidades, incluidos los roles de adaptación a la presión, ecología de la alimentación e historia de la vida. Los peces caracol muestran una adaptación especializada a la presión hidrostática, tanto en la acumulación del *n*-óxido de trimetilamina del agente de presión como en los cambios intrínsecos a las enzimas. El contenido del estómago y los análisis de isótopos de aminoácidos, y la morfología de la mandíbula sugieren que los peces depredadores que se alimentan por succión, como los liparidos, pueden encontrar una ventaja al descender a la zanja, donde los anfípodos son cada vez más abundantes. El análisis de las zonas de crecimiento de los otolitos sugiere que los peces caracol se pueden adaptar a un ambiente hadal sísmicamente activo y de alta perturbación al tener vidas relativamente cortas. Esta revisión sintetiza la literatura conocida sobre los peces de vida más profunda del planeta e informa una nueva comprensión de las adaptaciones a la vida en las trincheras.

Translated to Spanish by S. Hinojosa (hinojosa.silvia@gmail.com)

## Introduction

The hadal zone—predominately deep-sea trenches with depths ranging from 6000 to 11,000 m—represents 45% of the ocean's depth range but remains one of Earth's least explored habitats (e.g., Jamieson 2015). Taking their name from the Greek underworld, these parts of the ocean are characterized by conditions of high hydrostatic pressures, low temperatures, and complete absence of sunlight (Bruun 1957). The hadal zone is made up of 47 distinct habitats, including 27 subducting trenches, most of which are located around the Pacific Rim (Fig. 1; Stewart and Jamieson 2018). Trenches are long, narrow features, reaching lengths up to 4500 km and usually less than 100 km wide, made up of complex habitats including steep slopes and sedimentary basins (Stewart and Jamieson 2018).

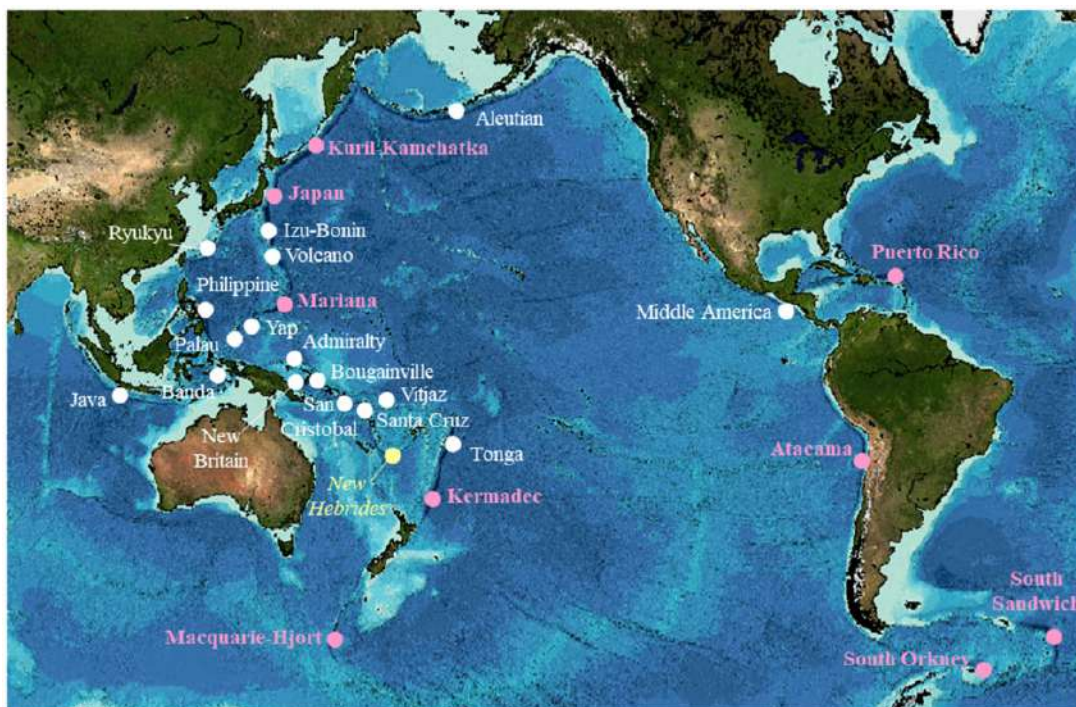
Many of the factors controlling the physiology and ecology of organisms found in hadal trenches are similar to those in the surrounding abyss and broader deep sea. However, a few exceptions make the hadal zone a distinct environment, most notably increased hydrostatic pressure, high levels of seismicity at plate boundaries, and a sloping topography distinct from the surrounding abyssal plains. With these environmental conditions, there are marked faunal transitions from the abyss to the hadal zone (Wolff 1970; Jamieson 2011). The hadal faunal community includes amphipods, tanaids, isopods, cumaceans, decapods, echinoderms, nematodes, polychaetes, copepods, molluscs, foraminifera, cnidarians, and fishes, at apparently high levels of endemism (Wolff 1958; Beliaev 1989; Jamieson et al. 2010; Jamieson 2015).

The first fish discovered from hadal depths was the cusk eel, *Bassogigas profundimus* (family Ophidiidae), caught in 1901 from 6035 m in the Moseley Trench by the Princess Alice expedition (Nielsen 1964). With additional exploration, however, a distinct shift in the fish community at the abyssal–hadal boundary was discovered (Figs. 2–5; Supplementary Note 1). Snailfishes (Liparidae, Scorpaeniformes, Fig. 6) now appear to be the most common vertebrates in the hadal environment. The shift from the abyssal to hadal fish community is an overlapping transition over about 500 m depth (Jamieson et al. 2011). For example, the macrourid *Coryphaenoides yaquinae* can, on occasion, be seen as deep as 7000 m, but they are most commonly found at depths less than 6000 m (Endo and Okamura 1992; Linley et al. 2017). In 1952, on the Danish *Galathea II* expedition—one of two prominent early voyages of hadal research—five individuals of the

hadal snailfish *Notoliparis kermadecensis* (Nielsen 1964) were caught between 6660 and 6770 m in the Kermadec Trench (Nielsen 1964). The other pioneering hadal expeditions, conducted aboard the Soviet vessel *Vityaz*, collected the snailfish, *Pseudoliparis amblystomopsis* (Andriashev 1955) in the Kuril–Kamchatka Trench at 7230 m in 1955 and in the Japan Trench to a maximum depth of 7579 m in 1957 (Nielsen 1964). Other hadal and deep-abyssal snailfishes were seen in the South Sandwich Trench, *Careproctus sandwichensis* at 5435–5453 m (Andriashev and Stein 1998), and in the Peru–Chile Trench (Fujii et al. 2010). Historical collections of hadal liparids have been few and far between (Table 1), limiting our understanding of the biology and ecology of these fishes.

As a family, snailfishes are found in a wide variety of habitats, from the intertidal to the hadal environment and from polar systems (Matallanas and Pequeno 2000) to the subtropics (Chernova et al. 2004). They are typically small, tadpole-shaped fishes which live in temperate to cold waters. Snailfishes display an array of interesting adaptations to distinct marine environments, including an antifreeze protein in the skin of Antarctic species (Evans and Fletcher 2001; Hobbs and Fletcher 2013), ossicles that may offer protection with minimum additional weight (Märss et al. 2010), and a subdermal extracellular matrix for increasing buoyancy and maintaining ideal body shape for drag reduction (Eastman et al. 1994; Gerringer et al. 2017a). The snailfishes are an understudied group, with many new species still being described (Chernova et al. 2004; Orr 2004; Chernova and Møller 2008; Stein and Drazen 2014) and with several species-level identifications and generic allocations still contentious (Kai et al. 2011; Orr et al., Accepted Manuscript.).

The abundance and significance of the family Liparidae at hadal depths (Table 2) has become increasingly clear through recent discoveries (e.g., Linley et al. 2016; Gerringer et al. 2017b). There are currently 12 species of hadal and near-hadal liparids (nine formally described, three observed) known from nine different trenches (Table 2). This marks a noticeable shift in the fish community at the abyssal–hadal boundary: from elongate, cosmopolitan species of the families Macrouridae (rattails), Ophidiidae (cusk eels), Ipnopidae (tripodfishes), Synphobranchidae (cutthroat eels), and Zoarcidae (eelpouts) on the abyssal plains (Wilson and Waples 1983; Nielsen and Merrett 2000; Milligan et al. 2016), to the hadal snailfishes (Liparidae) in the trenches. This review synthesizes the current



**Fig. 1** Hadal subduction trenches with depths exceeding 6000 m. Constructed using bathymetry data from the Global Multi-Resolution Topography (GMRT) synthesis data set (Ryan et al. 2009a) in GeoMapApp (www.geomapapp.org). List of trenches from Stewart and Jamieson (2018). Excludes 13 hadal trough and 7 trench fault habitats deeper than 6000 m (see map by Stewart and Jamieson [2018] for full hadal zone locations). Trenches with reports of snailfishes are highlighted in pink, trenches that have been sufficiently surveyed and shown to have no snailfishes shown in yellow.

scientific literature to address the following question: What evolutionary drivers have factored into the success of snailfishes in the trench environment relative to other potential colonizers?

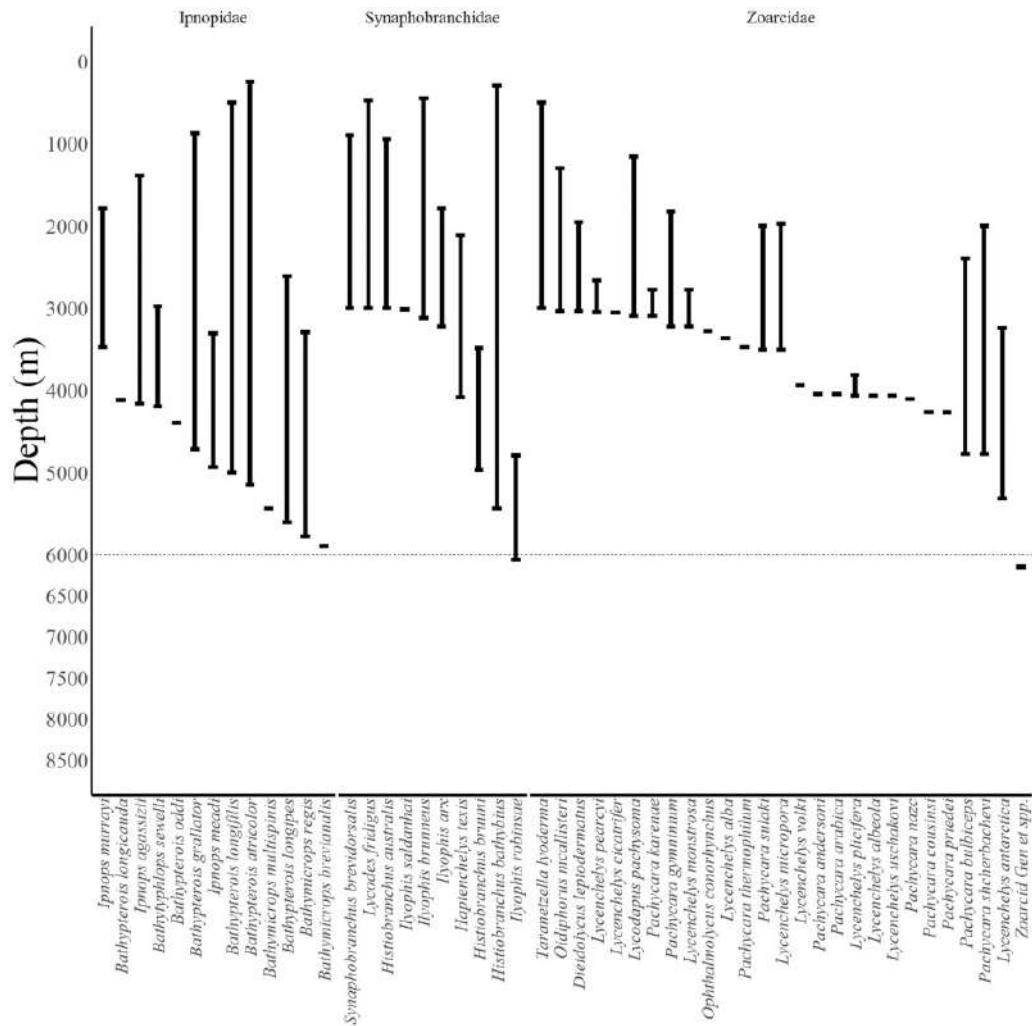
### Adaptations to high hydrostatic pressure

Perhaps the most intuitive factor that influences bathymetric ranges is hydrostatic pressure, long understood to limit radiation into the deep sea (Günther 1887). In the marine environment, pressures increase 1 atm with every 10 m depth, reaching as high as 1100 times atmospheric pressure (110 megapascals, MPa) at the ocean's deepest point. Naturally, only organisms that have adapted to high hydrostatic pressures are able to inhabit the hadal zone. Adaptations to high pressure occur on multiple scales, from the organismal to the molecular, from the limiting of gas-filled spaces such as swim bladders (Scholander 1954; Priede 2018) to homeoviscous adaptations to maintain cellular membrane fluidity under pressure through an increase in polyunsaturated fatty acid compositions (Sinensky 1974; Somero 1992).

Hydrostatic pressure is believed to set a depth limit for marine fishes nearly 2000 m shy of full-ocean depth. The depth limit hypothesis proposed

by Yancey et al. (2014) centers on the pressure-counteracting osmolyte trimethyl-amine oxide (TMAO). TMAO, a byproduct of lipid metabolism, increases concentration linearly with increasing habitat depth in fishes (Kelly and Yancey 1999). It appears that fishes living at greater pressures require TMAO to stabilize their proteins, a process thought to occur through the alteration of solvent interactions (e.g., Yancey and Siebenaller 1999). As the TMAO concentration increases, cellular osmolality also increases. At a certain depth—around 8200 m (Yancey et al. 2014)—fish cells contain such high TMAO concentrations that they reach the same osmolality as seawater. Adding additional TMAO to go deeper would pose an osmoregulatory problem, requiring a physiological shift to hyperosmotic life (Yancey et al. 2014). Hadal liparids have high TMAO concentrations and cellular osmolalities near isosmotic with seawater, suggesting that they are living at near the maximum depth for marine fishes (Yancey et al. 2014; Linley et al. 2016).

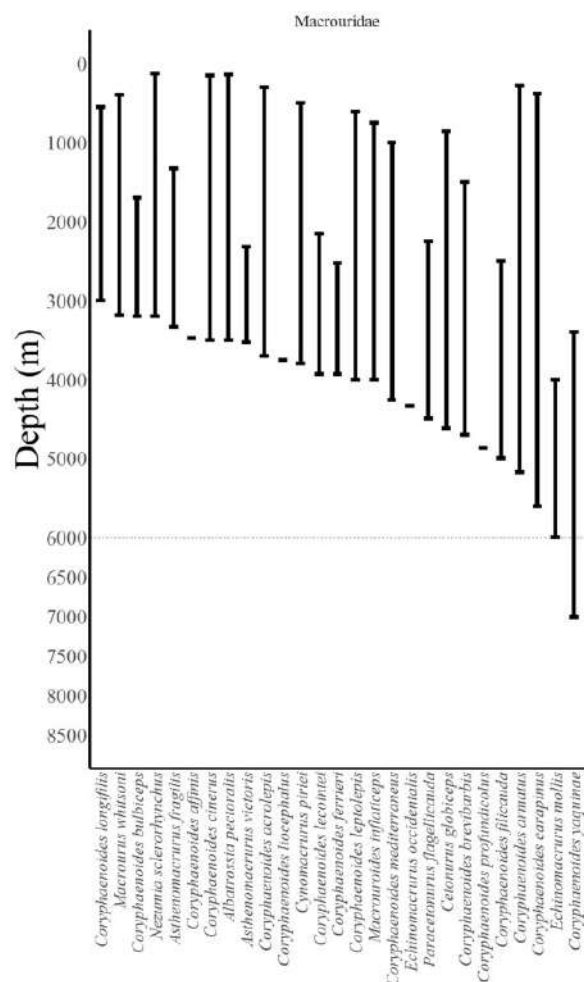
The accumulation of pressure-stabilizing molecules (piezolytes) such as TMAO represents an extrinsic adaptation, which alters the cellular milieu for protein function (reviewed by Yancey and Siebenaller 2015). However, organisms living under high



**Fig. 2** Reported depth distributions of deep-dwelling fishes (collections >3000 m) by family (Ipnopidae, Synphobranchidae, Zoarcidae). Abyssal–hadal boundary at 6000 m shown as dotted line. Data from FishBase (Froese and Pauly 2014) checked in the primary literature (Nielsen 1977; Andriashev and Stein 1998; Nielsen et al. 1999; Chernova et al. 2004; Chernova 2006; Duhamel and King 2007; Moller and King 2007; Jamieson et al. 2009; Linley et al. 2016, 2017; Geringer et al. 2017b). See Supplementary Note 1 for notes on record verifications.

hydrostatic pressures also exhibit intrinsic adaptations, where specific protein structures differ from those of their shallow-living counterparts. Several well-known pressure adaptations in other deep-sea taxa—such as high concentrations of polyunsaturated fatty acids to maintain membrane fluidity (Cossins and Macdonald 1986), reduced pressure sensitivities in  $\text{Na}^+ - \text{K}^+ - \text{ATPases}$  (Gibbs and Somero 1989), and adaptations of muscle actins (Swezey and Somero 1982)—likely allow hadal liparids to survive under pressure, although these adaptations have not yet been investigated to hadal depths. Geringer et al. (2017c) detailed a newly-discovered pressure adaptation in abyssal and hadal species—intrinsic changes in lactate dehydrogenases of hadal liparids and abyssal macrourids that allow

the enzymes to function better (increased maximum reaction rate,  $V_{\text{max}}$ ) at *in situ* pressures than they do at atmospheric pressure. Maximum reaction rates of lactate dehydrogenases, which catalyze the conversion of pyruvate to lactate in anaerobic glycolysis, in hadal liparids increased under habitat pressures of 60 MPa (water depth 6000 m). In shallow-living fishes, this enzyme was pressure inhibited. Adaptations to high pressure also occur at the organismal level in hadal liparids, which exhibit high volumes of gelatinous tissues and watery muscles to maintain buoyancy under high pressures without a gas bladder (Geringer et al. 2017a). The scope and diversity of pressure adaptations is great, and many more significant findings await in the study of hadal taxa.



**Fig. 3** Reported depth distributions of deep-dwelling fishes (collections >3000 m) by family (Macrouridae). Abyssal–hadal boundary at 6000 m shown as dotted line. Data from FishBase (Froese and Pauly 2014) checked in the primary literature (Nielsen 1977; Andriashev and Stein 1998; Nielsen et al. 1999; Chernova et al. 2004; Chernova 2006; Duhamel and King 2007; Moller and King 2007; Jamieson et al. 2009; Linley et al. 2016, 2017; Gerringer et al. 2017b). See Supplementary Note 1 for notes on record verifications.

### Feeding ecology

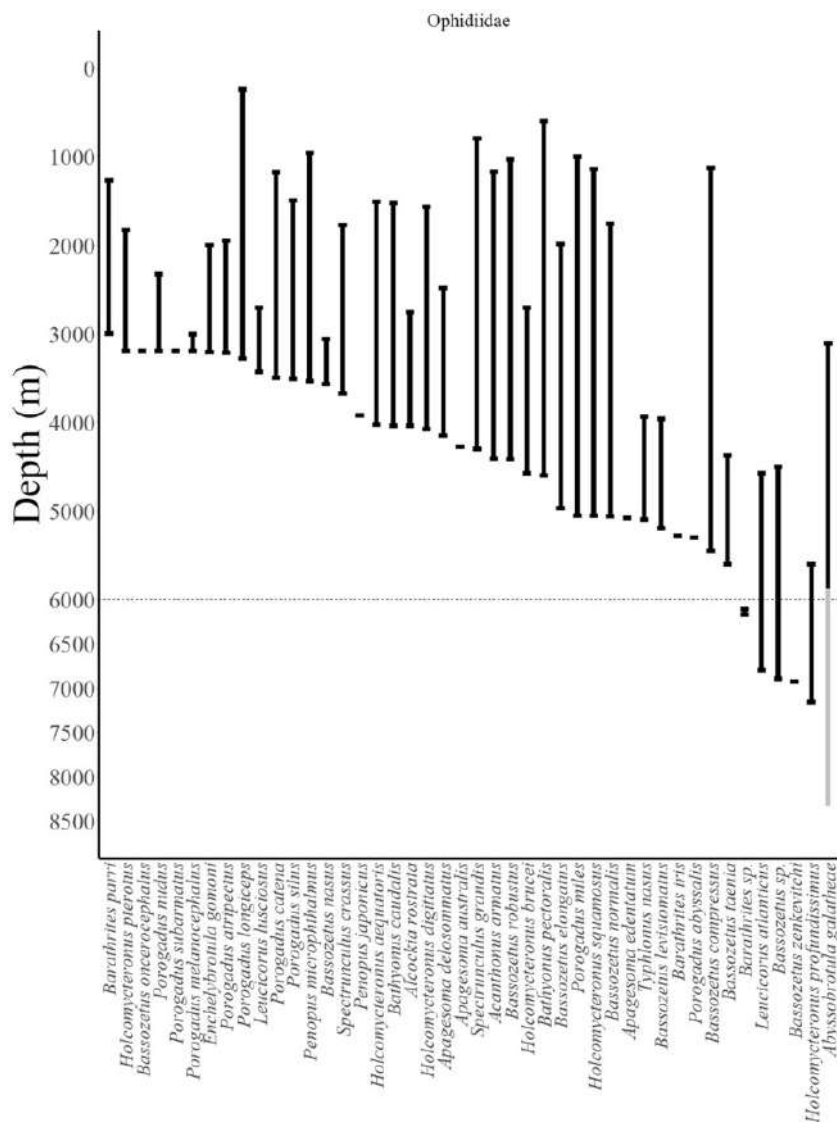
The v-shaped topography of hadal trenches is thought to funnel organic matter into the hadal zone, resulting in higher food availability for hadal organisms than for those in the food-limited abyss (George and Higgins 1979; Danovaro et al. 2003; Jamieson 2011; Ichino et al. 2015). This funneling is enhanced because hadal trenches are usually subduction zones—sites of high seismic activity that can trigger turbidity flows—bringing substantial quantities of sediment and organic matter into the trench (Itou et al. 2000; Oguri et al. 2013). Increased organic matter concentrations in the hadal zone seem to be accompanied by higher faunal abundances and

oxygen consumption that at abyssal depths (Beliaev 1989; Danovaro et al. 2002; Itoh et al. 2011; Glud et al. 2013; Wenzhöfer et al. 2016).

In hadal trenches, amphipod abundance increases substantially with increasing depth (Jamieson 2015), an effect that likely impacts the fish community (Gerringer et al. 2017d). Snailfishes may have a nutritional advantage in hadal trenches due to increased abundances of small crustacean prey such as amphipods (Gerringer et al. 2017d). Fishes that rely on piscivory or scavenging, such as macrourids or synphobranchids (reviewed by Drazen and Sutton 2017), might not have the same evolutionary pressures to invade hadal depths. Carrion is just as likely to fall in a trench as anywhere on the abyssal plains, indeed, far less likely considering the small global area of trenches compared with abyssal depths. Stomach contents and  $\delta^{15}\text{N}$  values of specific amino acids for two hadal snailfishes (*N. kermadecensis* and *P. swirei*) showed that amphipods are the most important prey item in both species (Gerringer et al. 2017d). This study revealed additional predator–prey relationships in the hadal zone—particularly that hadal liparids eat decapods, and, in some species, swimming polychaetes. Stomach content compositions and trophic positions were distinctly different for the abyssal and hadal species examined due differences in both feeding strategies and food availability.

A detailed functional morphology of abyssal and hadal fish feeding, similar to what has been conducted for Antarctic fishes (Bansode et al. 2014), may also further illuminate the role of trophic ecology in niche partitioning and depth zonation in the deep ocean. In addition to a shift in feeding strategy—from certain abyssal fishes that bite and tear their food to suction-feeding in hadal fishes—functional morphology and modeling of suction feeding parameters may reveal differences in optimum prey sizes for abyssal and hadal species. A difference in optimum prey size in relation to prey availability may partially account for the abundance of liparids in the hadal zone in comparison to the ophiidiids, which also suction feed but have only a shallow-hadal and abyssal range (Linley et al. 2017). Further, the strong pharyngeal jaw apparatus (Fig. 7) found in snailfishes from the Kermadec and Mariana trenches (Gerringer et al. 2017d) likely contributes to their ability to thrive on the large amphipod biomass at hadal depths.

Although few data exist on feeding ecology at hadal depths (Blankenship and Levin 2007; Gerringer et al. 2017d), recent studies have informed new understanding of trophic linkages in the hadal zone.



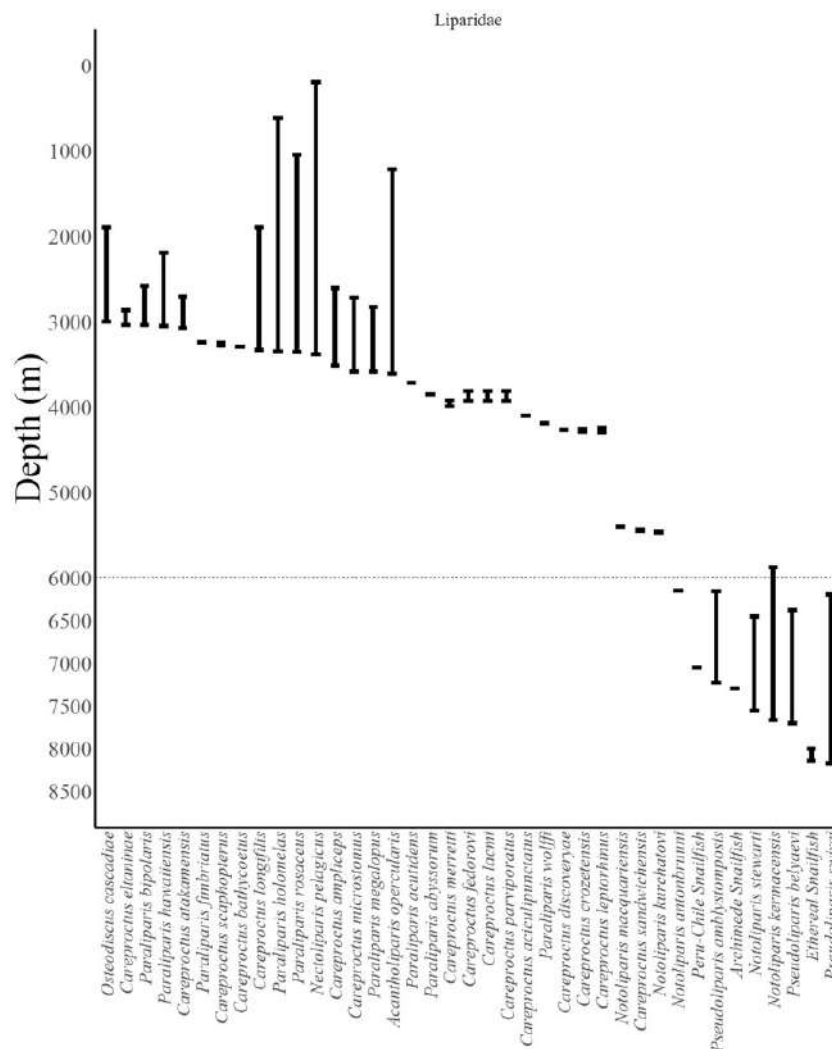
**Fig. 4** Reported depth distributions of deep-dwelling fishes (collections >3000 m) by family (Ophidiidae). Abyssal–hadal boundary at 6000 m shown as dotted line. Data from FishBase (Froese and Pauly 2014) checked in the primary literature (Nielsen 1977; Andriashev and Stein 1998; Nielsen et al. 1999; Chernova et al. 2004; Chernova 2006; Duhamel and King 2007; Moller and King 2007; Jamieson et al. 2009; Linley et al. 2016, 2017; Gerringer et al. 2017b). Uncertain record from open trawl shown in gray. See Supplementary Note 1 for notes on record verifications.

With additional analyses of stomach contents, stable isotopic compositions, and *in situ* feeding observations, the goal of assembling a hadal food web will be within reach. Further research is necessary to construct a complete food web; however, a preliminary outline is included in Fig. 8. Additional stomach contents and stable isotope analysis on other taxa—particularly amphipods, decapods, and holothurians—would be needed. Understanding the hadal food web is important not only as a matter of ecological interest, but also for tracking global patterns of carbon turnover. The hypothesis that trenches act as sinks of organic matter has been largely accepted (George and Higgins 1979;

Danovaro et al. 2002; Itoh et al. 2011; Glud et al. 2013; Ichino et al. 2015). Many authors note that accumulation of organic material would be significant for global carbon cycling, but the details of this process require further research (Oguri et al. 2013; Wenzhöfer et al. 2016).

### Life history

Hadal trenches are high-disturbance environments. Most trenches are located on subduction zones, sites of high seismic activity (Fig. 9). These seismic events can trigger rapid downslope movements of sediment known as turbidity flows, akin to undersea avalanches







**Fig. 6** Hadal liparids. *In situ* photographs of hadal snailfishes. a) c.f. *Notoliparis antonbruuni*, Atacama Trench, b) *Pseudoliparis swirei*, Mariana Trench, c) *Pseudoliparis belyaevi*, Japan Trench, d) *Notoliparis kermadecensis*, Kermadec Trench. Photos by Alan Jamieson, Newcastle University. Collections of hadal snailfish from the Mariana (e) and Kermadec (f) trenches, Hadal Ecosystem Studies cruises.

**Table 1** Global collections of hadal liparids by year

Trench	Species	Year	Depth	Ship	<i>n</i>
Kurile-Kamchatka	<i>Pseudoliparis amblystomopsis</i>	1953	7230	<i>Vitjaz</i>	1
Japan	<i>Pseudoliparis amblystomopsis</i>	1955	6156–7579	<i>Vitjaz</i>	5
Japan	<i>Pseudoliparis belyaevi</i>	1957	7579	<i>Vitjaz</i>	1
Kermadec	<i>Notoliparis kermadecensis</i>	1952	6660–6770	<i>Galathea</i>	5
Peru-Chile	<i>Notoliparis antonbruuni</i>	1966	6150	<i>Anton Bruun</i>	1
Japan	<i>Pseudoliparis amblystomopsis</i>	2009	7000	<i>Hakuho maru</i>	2
Kermadec	<i>Notoliparis kermadecensis</i>	2011	7002–7050	<i>Kaharoa</i>	6
Kermadec	<i>Notoliparis stewarti</i>	2011	7000	<i>Kaharoa</i>	1
Kermadec	<i>Notoliparis kermadecensis</i>	2014	6456–7554	<i>Thompson</i>	35
Kermadec	<i>Notoliparis stewarti</i>	2014	6456–7560	<i>Thompson</i>	6
Mariana	<i>Pseudoliparis swirei</i>	2014	6898–7966	<i>Falkor</i>	37
Mariana	<i>Pseudoliparis swirei</i>	2017	7581	<i>Shinyo-maru</i>	1

Collection year noted, depth presented in meters, *n* indicates the number of specimens collected. Current as of January 1, 2018. Ships operated out of the former Soviet Union, Denmark, USA, Japan, and New Zealand. Total: 101 specimens.

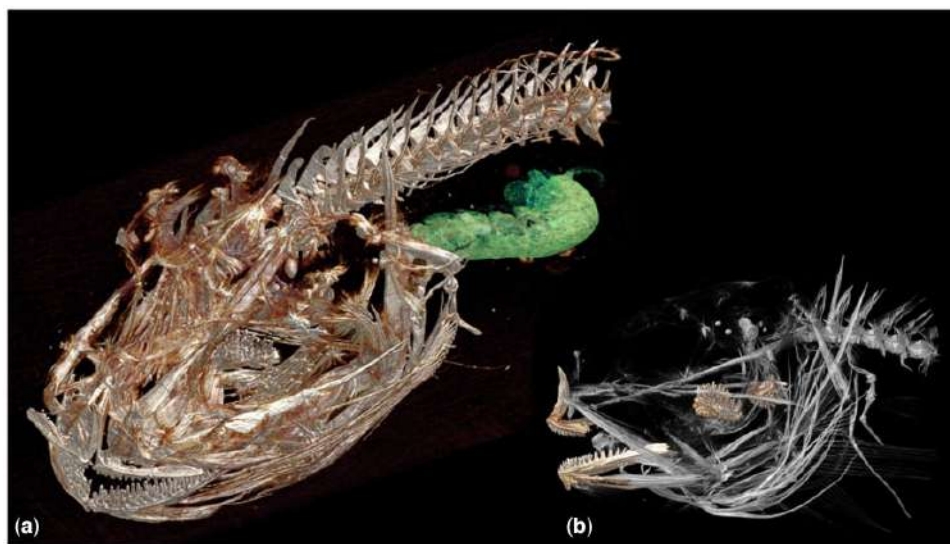
Future research should seek to validate these estimates and the age reading protocol. Although many methods for validation exist (reviewed by [Campana](#)

2001), most are not practical at either abyssal or hadal depths. Lead-radium dating may be used to validate the ages of abyssal rattails, *Coryphaenoides armatus*

**Table 2** Global geographic and bathymetric distribution of hadal liparids from observations and collections

Trench	Depth (m)	Species	Reference
Japan Trench	7420–7450	<i>Pseudoliparis amblystomopsis</i>	(Andriashev 1955)
	6380–7703	<i>Pseudoliparis belyaevi</i>	(Andriashev and Pitruk 1993; Fujii et al. 2010)
Kermadec Trench	5879–7669	<i>Notoliparis kermadecensis</i>	(Linley et al. 2016)
	6456–7560	<i>Notoliparis stewarti</i>	(Stein 2016)
Kuril-Kamchatka	6156–7587	<i>Pseudoliparis amblystomopsis</i>	(Andriashev 1955)
Macquarie-Hjort Trench	5400–5410	<i>Notoliparis macquariensis</i>	(Andriashev 1978)
Mariana Trench	6198–8178	<i>Pseudoliparis swirei</i>	(Gerringer et al. 2017b; Oguri and Noguchi 2017)
	8007–8145	Ethereal snailfish	(Linley et al. 2016)
Peru-Chile Trench	6150	<i>Notoliparis antonbruuni</i>	(Stein 2005)
	7049	Peru–Chile snailfish	(Linley et al. 2016)
Puerto Rico Trench	7300	Archimede snailfish	(Pérès 1965)
South Orkney Trench	5465–5474	<i>Notoliparis kurchatovi</i>	(Andriashev 1975)
South Sandwich Trench	5435–5453	<i>Careproctus sandwichensis</i>	(Andriashev and Stein 1998)

Includes records of abyssal species reported near trenches, current as of January 1, 2018. HOV *Archimede* observation in Puerto Rico Trench is probable, but anecdotal, see text for details. Undescribed species listed with common names.

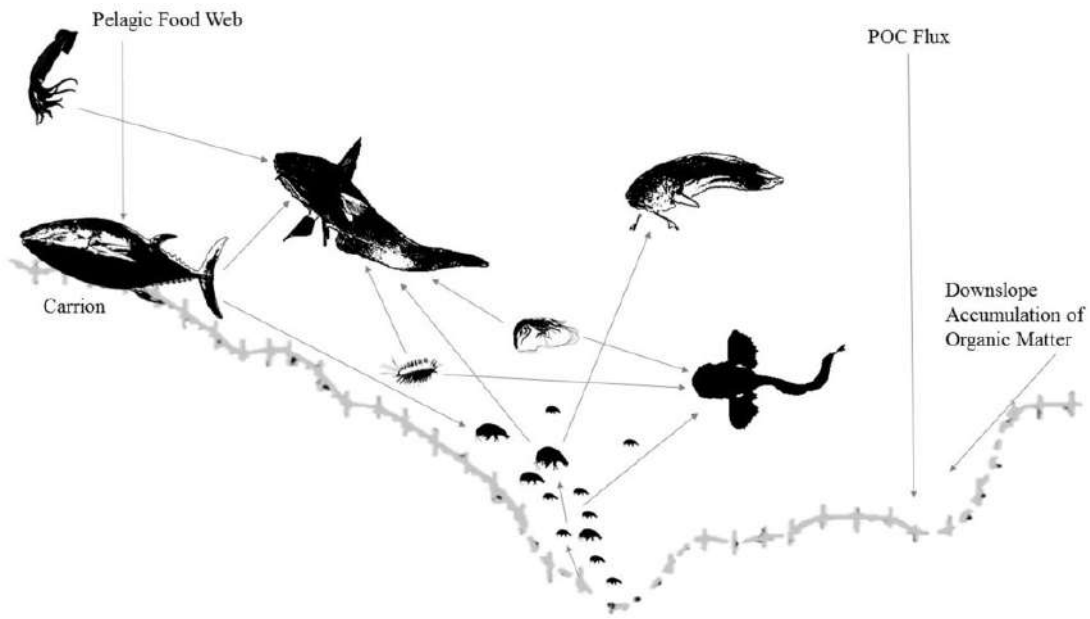


**Fig. 7** a) Skeletal morphology of holotype hadal snailfish, *P. swirei*, from micro-computed tomography. Amphipod prey visible in the stomach in green. Fish head length 18.9 mm. b) The strong pharyngeal jaw apparatus plays an important role in amphipod feeding.

and *C. yaquinae*, as has been applied to other species in the Macrourid family (Andrews et al. 1999). However, this method requires nearly a gram of early otolith growth (core extraction of first few years) material for instrumental analysis. Due to the small size of the hadal liparid otoliths, multiple individuals from discrete size or age classes would be needed to pool enough material for lead-radium dating. This type of analysis would require the combination of multiple collections to cover a thorough size range, but would be a valuable way to verify growth patterns in a common and important abyssal group. Validated life history data on these deep-living species would also

greatly inform discussions of the factors governing growth rate of deep-sea fishes (Cailliet et al. 2001; Drazen and Haedrich 2012), potentially clarifying the role of environmental variables such as temperature, pressure, and food supply. Other validation methods such as mark/recapture or outer ring analysis across multiple seasons are also impractical at this time (Gerringer et al. 2018).

It might be expected that hadal liparids would have benthic larvae—limiting dispersal and facilitating endemism. However, this was not supported by thermal history reconstructions based on oxygen isotopic composition of otoliths (Gerringer et al. 2018).

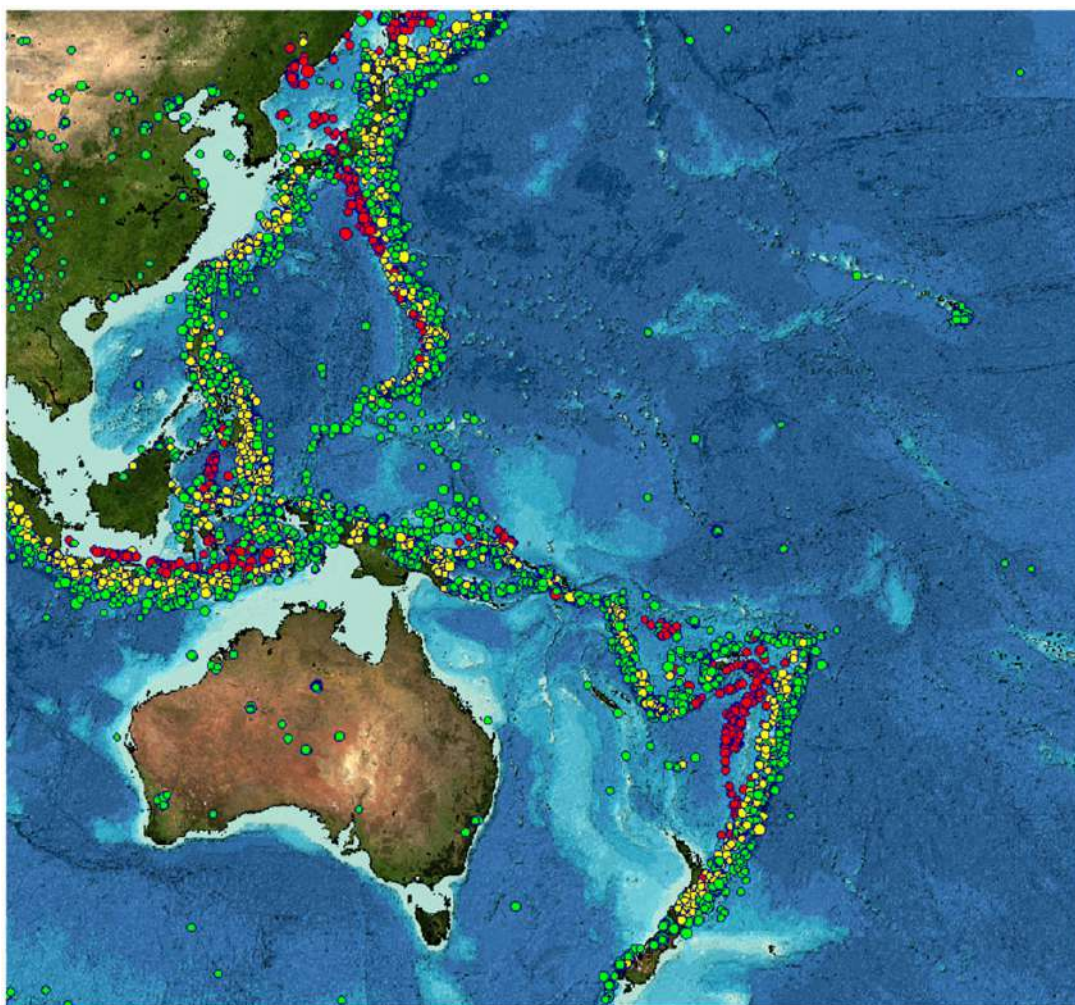


**Fig. 8** Generalized hadal food web derived from information presented here. Bathymetry based on Japan Trench (Fisher 1954). Arrows indicate known trophic linkages. Organisms shown include squid, macrourids, ophiidiids, liparids, decapods, polychaetes, and amphipods. Macrourids and ophiidiids are part of the abyssal/upper edges of the hadal food web. POC, particulate organic carbon. Drawings based on photos from MBARI, NOAA OER, Stuart Piertney, Alan Jamieson. Not to scale.

Oxygen isotope ratios in otoliths vary with temperature and seawater  $\delta^{18}\text{O}$ , allowing thermal histories of fishes to be calculated from measurements across otolith growth zones (Thorrold et al. 1997). Unexpectedly, large changes in oxygen isotopic compositions suggest temperature changes of greater than  $5^\circ\text{C}$  across ontogeny for hadal liparids from both the Mariana and Kermadec trenches (Gerringer et al. 2018). If temperature is the only factor influencing the oxygen composition of hadal liparid otoliths, these results would reflect an approximately 5000 m depth differential between larval and adult populations, according to the established conversions (Thorrold et al. 1997; Høie et al. 2004; Chang et al. 2015). According to the published literature and validation efforts, this change could not be explained by metabolically-mediated fractionation, pressure effects, excess protein accumulation, or instrumental drift. It is possible that an alternative unknown effect on carbonate chemistry is taking place.

If these snailfishes are indeed feeding in shallow waters before returning to the hadal zone, this is an incredible feat. Large vertical migrations from larval habitat to adult habitat have been documented in other species, for example in macrourids up to 1400 m (Lin et al. 2012) and up to 800 m in the jellynose fish *Ateleopus japonicus* (Shiao et al. 2017). Further, snailfishes in the closely-related genus *Liparis* often exhibit planktonic larval stages (Marliave and Peden 1989; Sokolovskii and

Sokolovskaya 2003; Walkusz et al. 2016) and thermal history reconstructions of the blacktail snailfish *C. melanurus* suggest similar life history traits (Gerringer et al. 2018). Although counterintuitive, a pelagic larval stage may also be supported by the interpretation of Munk (1964), who wrote “It seems fairly probable that all major constituents of the eyes of the deep-sea fishes examined have been fully differentiated before degeneration sets in, and it is suggested that the eyes may have been functionally normal in larvae and maybe also in young adults.” However, this would be one of the largest changes in hydrostatic pressure experienced by any metazoan and this finding certainly warrants further verification. Pressure effects, such as those explored in Gerringer et al. (2017c), would need to be investigated with this added complication of ontogenetic changes in habitat pressure. If hadal liparid larvae are indeed present at depths shallower than 1000 m (Gerringer et al. 2018), it should be possible to collect them by trawl, although similar surveys of deep-water snailfishes in Alaskan waters yielded few collections, despite the high number of known snailfish species in the area (Matarese et al. 2003). Although liparid larvae have few distinguishing characters (Kim et al. 1986a, 1986b; Marliave and Peden 1989) and confirming identifications morphologically would be difficult, genetic analysis could be used. This technique has been successfully applied in a number of systems for larval identification



**Fig. 9** Foci of large earthquakes (magnitude 5.0–9.0) in the Western Pacific over a 30-year period (1964–1995), showing hadal zones to be areas of high seismic activity. Earthquake data from the International Seismological Centre. Bathymetry from General Bathymetric Chart of the Oceans (GEBCO) and GMRT (Ryan et al. 2009b), map made in GeoMapApp (www.geomapp.org). Points are colored by earthquake depth: <50 km (green), 50–250 km (yellow), and >250 km (red).

(e.g., Hubert et al. 2010; Riemann et al. 2010). Data from the genetic markers (mitochondrial genes 16S, COI, and Cyt-b) analyzed in Geringer et al. (2017b) for *Notoliparis kermadecensis* and *N. stewarti* from the Kermadec Trench and *Pseudoliparis swirei* from the Mariana Trench would allow for positive identification of hadal liparid larvae collected from shallower depths over the trench.

### Evolution into the hadal environment

The current literature shows hadal liparids to be highly-specialized, endemic fishes that thrive in deep-sea trenches through a suite of adaptations. Hadal liparids have been captured and described from the Kuril–Kamchatka Trench (Andriashev 1955), Japan Trench (Andriashev 1955; Andriashev and Pitruk 1993), Kermadec Trench (Nielsen 1964), and Peru–Chile Trench (Stein 2005). In addition to

these known species, there are likely undiscovered species of hadal liparids in unexplored trenches. In 1964, for example, those diving the Puerto Rico Trench in the bathyscaphe *Archimede* reported that “at 7,300 m the community was characterized by the abundance of a liparidid fish (*Careproctus?*: about 200 individuals)” (Pérès 1965). Due to its geographic isolation from other hadal ecosystems, this finding would be of particular importance to the understanding of the adaptation and evolution of the planet’s deepest-living vertebrates. Megafauna collections in the Puerto Rico Trench would be a valuable target for future efforts, as would under-surveyed trenches such as the Aleutian, Philippine, and Java trenches.

The colonization of the deep sea by fishes, and by snailfishes in particular, is understood to be a radiation from shallow into the deep (Burke 1930; Priede and Froese 2013). Priede and Froese (2013) posit that

the evolution of liparids into the trenches are the result of independent speciation events. However, recent genetic analyses (mtCOI) show that hadal liparid species included in recent phylogenetic analyses (*N. stewarti*, *N. kermadecensis*, *P. swirei*, and *P. belyaevi*) form a strongly supported clade together when compared with extant species (Gerringer et al. 2017b; Orr et al., Accepted Manuscript.). Further, Orr et al. (Accepted Manuscript) show that this clade is basal to nearly all other liparids. Future phylogenetic analyses of multiple hadal liparids, building on the trees presented by Gerringer et al. (2017b) and Orr et al. (Accepted Manuscript), would be valuable in understanding connectivity between hadal fish populations and evolutionary patterns of dispersal. One interesting hypothesis for future exploration is a potential Antarctic origin of hadal fishes. Researchers have long noticed similarities between the Antarctic, Arctic, and deep-sea habitats (e.g., Thorson 1950). These common evolutionary drivers including cold temperatures and prolonged darkness, coupled by the deep-water currents radiating from Antarctica, may support an Antarctic origin for deep-sea taxa. Although evidence for Antarctic deep-sea origins has been found for octopods (Strugnell et al. 2008) and isopods (Hessler and Thistle 1975), no studies have yet addressed a potential Antarctic origin for hadal liparids.

Future phylogenetic work will also require careful consideration of taxonomic classifications, which are in need of revision across the snailfishes. With the collection of additional hadal liparid species, the two liparid genera *Notoliparis* and *Pseudoliparis*—which overlap in most taxonomic and ecological characteristics—should likely be synonymized (Gerringer et al. 2017b). Similar research has begun for hadal amphipods, which shows complex connectivity patterns and species overlap, particularly in the genus *Hirondellea* (Ritchie et al. 2015). It will be worthwhile to continue to explore how connected or distinct individual hadal trenches are from one another and the waters above, and whether these patterns vary across taxa. While it may be tempting to think of hadal trenches as remote, isolated habitats, evidence increasingly shows that trenches are closely tied to the surrounding ocean systems, even accumulating man-made pollutants at high concentrations (Jamieson et al. 2017).

Interestingly, there are now a few hadal trenches known to have more than one species of apparently endemic hadal snailfish. The Japan Trench has both *Pseudoliparis belyaevi* and *P. amblystomopsis* (Andriashev and Pitruk 1993). Recent work has revealed two genetically distinct populations in the

Kermadec Trench—*N. kermadecensis* and *Notoliparis stewarti* (Stein 2016; Gerringer et al. 2017b). There are also two snailfish populations in the Mariana Trench—*P. swirei* and the ethereal snailfish, which remains uncollected (Linley et al. 2016; Gerringer et al. 2017b). It is possible that the potential pelagic larval stage proposed from otolith microchemistry (Gerringer et al. 2018) provides a dispersal mechanism for these populations, for example for *P. amblystomopsis*, which is found in both the Japan and Kuril–Kamchatka trenches (Andriashev and Pitruk 1993). In the case of the ethereal snailfish and *P. swirei*, it is possible to envision distinct ecological niches. In form, these two fishes are very distinct, with the delicate ethereal snailfish almost certainly representing an undescribed genus. *Pseudoliparis swirei* is an active predator, feeding on amphipods and decapods in the water (Gerringer et al. 2017d). No ethereal snailfish specimens have been collected, so stomach content and stable isotope analyses have not been possible, but one fish was observed moving headfirst into the sediment in what appears to be a feeding behavior (Linley et al. 2016), suggesting that it may rely instead on benthic invertebrates. However, for *P. belyaevi* and *P. amblystomopsis*, and for *N. kermadecensis* and *N. stewarti*, there is no obvious driver of speciation. Indeed, they are indistinguishable in video. Collections of *N. kermadecensis* and *N. stewarti* have revealed no obvious distinctions in trophic ecology, pressure adaptation, or life history (Gerringer et al. 2017c, 2017d, 2018), despite the presence of two clades based on mitochondrial gene sequences (Gerringer et al. 2017b). Perhaps these populations were once geographically isolated and the genetic differences reflect some period of extended isolation in the past.

Evolution into the deep sea naturally requires adaptation to high hydrostatic pressure. However, reconstructing these evolutionary processes requires consideration of an important factor: pressure and temperature have interacting effects on biomolecules (e.g., Dahlhoff and Somero 1991; Somero 1992). An interesting result of pressure–temperature interaction is that a change in the temperature of the deep ocean would have profound and complex effects on organismal responses to pressure. As one example for consideration, the proposed depth limit of ~8200 m (Yancey et al. 2014) would likely be temperature dependent, as both pressure and temperature affect volume changes associated with physiological processes (reviewed by Somero 1992). Indeed, colonization of the deep sea is believed to have occurred during warmer temperatures (Priede and Froese

2013), which may have allowed easier adaptation to higher pressures. Eventually, it may be possible to develop a model that quantifies pressure and temperature interactions on a molecular level, allowing something like the agricultural growing degree-day, which relates growth rates to temperature (e.g., Neuheimer and Taggart 2007)—a growing-degree-day-MPa—to be applied in discussions of the rates of life and adaptation to different pressure–temperature regimes (Gerringer et al. 2017c).

### Future directions

Although recent decades have marked significant advancements in understanding of the hadal zone, there is still much that remains unknown about these habitats. Snailfishes have been found in at least six hadal trenches globally (Fig. 1 and Table 2). It is likely that additional hadal snailfish populations exist in trenches that have not yet been thoroughly explored. However, there is at least one trench, the New Hebrides, that does not appear to house an endemic hadal snailfish population (Linley et al. 2017). The reasons for this absence are not fully understood, but may relate to a distinct water mass input compared with trenches like the Kermadec, Mariana, and Japan, or to differences in surface productivity of overlying waters (Linley et al. 2017). Future exploration of trenches near the New Hebrides, such as the Vitjaz, Bougainville, San Cristobal, and New Britain, would help to clarify these drivers.

Hadal science in the last 50 years has been largely exploratory. The next phase of hadal research should strive for greater temporal and spatial understanding of deep-sea trenches, including of the fish community. From the videos, images, and samples of the hadal zone collected so far, it is already clear that there is great topographic and faunal heterogeneity within individual trenches and between trenches (Stewart and Jamieson 2018). The factors driving these changes warrant further exploration. Further, much of the work done at hadal depths has been on bait-attending fauna, due to technical constraints. Although useful, free-falling lander equipment provides a certain biased view of the community (e.g., Jamieson 2018). Alternative sampling and exploration methods, including remotely operated vehicles, will need to be employed to gain a more holistic view of hadal fauna and their physiology and ecology. In the future, *in situ* experiments would provide valuable additional information about life in the hadal zone. For example, existing methods of measuring metabolic rates in deep-living fishes (e.g., Drazen

et al. 2015) have been complicated by pressure effects on enzyme activities (Gerringer et al. 2017c). Studies using *in situ* oxygen consumption measurements on hadal fishes could be compared with shallower-living taxa to disentangle the roles of pressure, temperature, and food supply on metabolic rates that existing methods.

Additional exploration of abyssal ecosystems will also be needed to understand what makes the hadal zone distinct. Much of the work comparing the hadal liparids to abyssal fishes has been conducted using the prominent abyssal macrourids *C. armatus* and *C. yaquinae* as representative abyssal taxa. The macrourids are certainly an important abyssal group, with representatives in most ocean basins at a broad range of depths (Wilson and Waples 1983; Jamieson et al. 2009). Because there is a diversity of fishes living at abyssal depths, however, this is an oversimplification of the interactions at play in structuring the fish communities at abyssal and hadal depths. In particular, this view likely underrepresents the role of the cusk eels (Ophidiidae). Ophidiids are a largely understudied group, though they are wide-ranging and likely important members of the abyssal fish community (Nielsen and Merrett 2000; Uiblein et al. 2008; Nielsen and Møller 2011; Linley et al. 2017). To more thoroughly compare abyssal and hadal fishes, future research should focus on the biogeography, ecology, and physiology of the deep-dwelling cusk eels—to address their role in the abyssal ecosystem. Other abyssal fish families could also be compared in more detail in future work, including the eelpouts (Zoarcidae), cutthroat eels (Synphobranchidae), and tripodfishes (Ipnopidae), the last of which are very poorly studied as they are not attracted to baited cameras or traps. Furthering our understanding of deep bathyal and abyssal liparids would also provide insight into the evolution of fishes into the deep sea.

Hadal science is by necessity highly collaborative and interdisciplinary work. It will take continued cooperation between a host of scientists and engineers across borders to further our understanding of these communities in the ocean's greatest depths and their role in the global ocean ecosystem.

### Acknowledgments

Thanks to the captains and crews of the research vessels that supported the published work, especially the *RVs Thompson, Falkor, Kaharoa, Shinyo-Maru*, and *Centennial*. Thanks to Adam Summers for CT scanning and visualization. I am grateful to Jeffrey Drazen (University of Hawai'i at Mānoa), Paul

Yancey (Whitman College), Alan Jamieson, Thomas Linley (Newcastle University), James Orr (NOAA Fisheries, Alaska Fisheries Science Center), Allen Andrews (NOAA Fisheries, Pacific Islands Fisheries Science Center), Craig Smith, Anna Neuheimer, Brian Popp (University of Hawai'i at Mānoa), Logan Peoples (University of California San Diego), and Adam Summers (Friday Harbor Laboratories, University of Washington) for insightful comments, discussions, and draft reviews.

## Funding

This work was supported by the Seaver Institute, National Science Foundation (OCE #1130712 to J.C. Drazen), Schmidt Ocean Institute, the National Science Foundation Graduate Research Fellowship, and Friday Harbor Laboratories, University of Washington.

## Supplementary data

Supplementary data available at *IOB* online.

## References

- Andrews AH, Cailliet GM, Coale KH. 1999. Age and growth of the Pacific grenadier (*Coryphaenoides acrolepis*) with age estimate validation using an improved radiometric ageing technique. *Can J Fish Aquat Sci* 56:1339–50.
- Andriashev A. 1955. A new fish of the snailfish family (Pisces, Liparidae) found at a depth of more than 7 kilometers. *Trudy Inst Okeanol Akad Nauk SSSR* 12:340–4.
- Andriashev A. 1975. A new ultra-abyssal fish, *Notoliparis kurchatovi* gen. et sp. 7n. (Liparidae) from the South Orkney Trench (Antarctic). *Trudy Inst Okeanol Akad Nauk SSSR* 103:313–9.
- Andriashev AP. 1978. On the third species of the ultra-abyssal genus *Notoliparis* Andr. (Pisces, Liparidae), from the deep-waters of the Macquarie Trench, with notes about the zoogeographic and evolutionary interest of this discovery. *Trudy Inst Okeanol Akad Nauk SSSR* 112:152–61.
- Andriashev AP, Pitruk DL. 1993. Review of the ultra-abyssal (hadal) genus *Pseudoliparis* (Scorpaeniformes, Liparidae) with a description of a new species from the Japan Trench. *J Ichthyol* 33:325–30.
- Andriashev AP, Stein DL. 1998. Review of the snailfish genus *Careproctus* (Liparidae, Scorpaeniformes) in Antarctic and adjacent waters. Los Angeles (CA): Natural History Museum of Los Angeles County.
- Bansode MA, Eastman JT, Aronson RB. 2014. Feeding biomechanics of five demersal Antarctic fishes. *Polar Biol* 37:1835–48.
- Beliaev G. 1989. Deep-sea ocean trenches and their fauna (Ye Vinogradov M., editor). Moscow: Nauka Publishing House.
- Blankenship LE, Levin LA. 2007. Extreme food webs: foraging strategies and diets of scavenging amphipods from the ocean's deepest 5 kilometers. *Limnol Oceanogr* 52:1685–97.
- Bruun AF. 1957. General introduction to the reports and list of deep-sea stations. *Galathea Rep* 1:7–48.
- Burke V. 1930. Revision of fishes of family Liparidae. *Bull U S Natl Mus* i–xii:1–204.
- Cailliet GM, Andrews AH, Burton EJ, Watters DL, Kline DE, Ferry-Graham LA. 2001. Age determination and validation studies of marine fishes: do deep-dwellers live longer? *Exp Gerontol* 36:739–64.
- Campana S. 2001. Accuracy, precision and quality control in age determination, including a review of the use and abuse of age validation methods. *J Fish Biol* 59:197–242.
- Chang NN, Liu EY, Liao YC, Shiao JC. 2015. Vertical habitat shift of viviparous and oviparous deep-sea cusk eels revealed by otolith microstructure and stable-isotope composition. *J Fish Biol* 86:845–853.
- Chernova NV. 2006. New and rare snailfishes (Liparidae, Scorpaeniformes) with the description of four new species from the Southern Hemisphere and tropical east Pacific. *J Ichthyol* 46:S1–S14.
- Chernova NV, Møller PR. 2008. A new snailfish, *Paraliparis nigellus* sp. nov. (Scorpaeniformes, Liparidae), from the northern Mid-Atlantic Ridge—with notes on occurrence of *Pseudnos* in the area. *Mar Biol Res* 4:369–75.
- Chernova NV, Stein DL. 2002. Ten new species of *Pseudnos* (Pisces, Scorpaeniformes: Liparidae) from the Pacific and North Atlantic Ocean. *Copeia* 2002:755–78.
- Chernova NV, Stein DL, Andriashev AP. 2004. Family Liparidae Scopoli 1777. *Calif Acad Sci Annot Checklists Fishes* 31.
- Cossins AR, Macdonald AG. 1986. Homeoviscous adaptation under pressure. III. The fatty acid composition of liver mitochondrial phospholipids of deep-sea fish. *BBA* 860:325–35.
- Dahlhoff EP, Somero GN. 1991. Pressure and temperature adaptation of cytosolic malate dehydrogenases of shallow- and deep-living marine invertebrates: evidence for high body temperatures in hydrothermal vent animals. *J Exp Biol* 159:473–87.
- Danovaro R, Gambi C, Della Croce N. 2002. Meiofauna hotspot in the Atacama Trench, eastern South Pacific Ocean. *Deep Sea Res Part I Oceanogr Res Pap* 49:843–57.
- Danovaro R, Della Croce N, Dell'Anno A, Pusceddu A. 2003. A depocenter of organic matter at 7800 m depth in the SE Pacific Ocean. *Deep Sea Res Part I Oceanogr Res Pap* 50:1411–20.
- Drazen JC, Haedrich RL. 2012. A continuum of life histories in deep-sea demersal fishes. *Deep Sea Res Part I Oceanogr Res Pap* 61:34–42.
- Drazen JC, Sutton TT. 2017. Dining in the deep: the feeding ecology of deep-sea fishes. *Annu Rev Mar Sci* 9:337–66.
- Drazen JC, Friedman JR, Condon NE, Aus EJ, Gerringer ME, Keller AA, Clarke ME. 2015. Enzyme activities of demersal fishes from the shelf to the abyssal plain. *Deep Sea Res Part I Oceanogr Res Pap* 100:117–26.
- Duhamel G, King N. 2007. Deep-sea snailfish (Scorpaeniformes: Liparidae) of genera *Careproctus* and *Paraliparis* from the Crozet Basin (Southern Ocean). *Cybius* 31:379–87.
- Eastman JT, Hikida RS, Devries AL. 1994. Buoyancy studies and microscopy of skin and subdermal extracellular matrix of the Antarctic snailfish, *Paraliparis devriesi*. *J Morphol* 220:85–101.

- Endo H, Okamura O. 1992. New records of the abyssal grenadiers *Coryphaenoides armatus* and *C. yaquinae* from the western North Pacific. *Jpn J Ichthyol* 38:433–7.
- Evans RP, Fletcher GL. 2001. Isolation and characterization of type I antifreeze proteins from Atlantic snailfish (*Liparis atlanticus*) and dusky snailfish (*Liparis gibbus*). *Biochim Biophys Acta* 1547:235–44.
- Fisher RL. 1954. On the sounding of trenches. *Deep Res* 2:48–58.
- Froese R, Pauly D. 2014. FishBase. World Wide Web Electron Publ, November 2014.
- Fujii T, Jamieson AJ, Solan M, Bagley PM, Priede IG. 2010. A large aggregation of liparids at 7703 meters and a reappraisal of the abundance and diversity of hadal fish. *Bioscience* 60:506–15.
- George RY, Higgins RP. 1979. Eutrophic hadal benthic community in the Puerto Rico Trench. *Ambio Spec Rep* 6:51–8.
- Gerringer ME, Drazen JC, Linley TD, Summers AP, Jamieson AJ, Yancey PH. 2017a. Distribution, composition and functions of gelatinous tissues in deep-sea fishes. *R Soc Open Sci* 4:171063.
- Gerringer M, Linley T, Jamieson A, Goetze E, Drazen, J. 2017b. *Pseudoliparis swirei* sp. nov.: a newly-discovered hadal snailfish (Scorpaeniformes: Liparidae) from the Mariana Trench. *Zootaxa* 4358:161–77.
- Gerringer ME, Drazen JC, Yancey PH. 2017c. Metabolic enzyme activities of abyssal and hadal fishes: pressure effects and a re-evaluation of depth-related changes. *Deep Res Part I Oceanogr Res Pap* 125:135–46.
- Gerringer ME, Popp BN, Linley TD, Jamieson AJ, Drazen JC. 2017d. Comparative feeding ecology of abyssal and hadal fishes through stomach content and amino acid isotope analysis. *Deep Res Part I Oceanogr Res Pap* 121:110–20.
- Gerringer ME, Andrews AH, Huss GR, Nagashima K, Popp BN, Linley TD, Gallo ND, Clark MR, Jamieson AJ, Drazen JC. 2018. Life history of abyssal and hadal fishes from otolith growth zones and oxygen isotopic compositions. *Deep Res Part I Oceanogr Res Pap* 132:37–50.
- Gibbs A, Somero G. 1989. Pressure adaptation of Na<sup>+</sup>/K<sup>+</sup>-ATPase in gills of marine teleost fishes. *J Exp Biol* 143:475–92.
- Glud RN, Wenzhöfer F, Middelboe M, Oguri K, Turnewitsch R, Canfield DE, Kitazato H. 2013. High rates of microbial carbon turnover in sediments in the deepest oceanic trench on Earth. *Nat Geosci* 6:284–8.
- Günther A. 1887. Report on the deep-sea fishes collected by H.M.S. Challenger during the years 1873–76. *Voyag HMS Chall* 22:1–277.
- Hessler R, Thistle D. 1975. On the place of origin of deep-sea isopods. *Mar Biol* 32:155–65.
- Hobbs RS, Fletcher GL. 2013. Epithelial dominant expression of antifreeze proteins in cunner suggests recent entry into a high freeze-risk ecozone. *Comp Biochem Physiol Part A* 164:111–8.
- Høie H, Otterlei E, Folkvord A. 2004. Temperature-dependent fractionation of stable oxygen isotopes in otoliths of juvenile cod (*Gadus morhua* L.). *ICES J Mar Sci* 61:243–51.
- Hubert N, Delrieu-Trottin E, Irisson J-O, Meyer C, Planes S. 2010. Identifying coral reef fish larvae through DNA barcoding: A test case with the families Acanthuridae and Holocentridae. *Mol Phylogenet Evol* 55:1195–203.
- Ichino MC, Clark MR, Drazen JC, Jamieson A, Jones DOB, Martin AP, Rowden AA, Shank TM, Yancey PH, Ruhl HA. 2015. The distribution of benthic biomass in hadal trenches: a modelling approach to investigate the effect of vertical and lateral organic matter transport to the seafloor. *Deep Sea Res Part I Oceanogr Res Pap* 100:21–33.
- Itoh M, Kawamura K, Kitahashi T, Kojima S, Katagiri H, Shimanaga M. 2011. Bathymetric patterns of meiofaunal abundance and biomass associated with the Kuril and Ryukyu trenches, western North Pacific Ocean. *Deep Sea Res Part I Oceanogr Res Pap* 58:86–97.
- Itou M, Matsumura I, Noriki S. 2000. A large flux of particulate matter in the deep Japan Trench observed just after the 1994 Sanriku-Oki earthquake. *Deep Sea Res Part I Oceanogr Res Pap* 47:1987–98.
- Jamieson AJ. 2011. Ecology of deep oceans: hadal trenches. In: *Encyclopedia of life sciences(eLS)*. Chichester: John Wiley & Sons, Ltd. p. 1–8.
- Jamieson AJ. 2015. *The hadal zone: life in the deepest oceans*. Cambridge (UK): Cambridge University Press.
- Jamieson AJ. 2018. A contemporary perspective on hadal science. *Deep Res Part II Top Stud Oceanogr* 155:4–10.
- Jamieson AJ, Yancey PH. 2012. On the validity of the Trieste flatfish: dispelling the myth. *Biol Bull* 222:171–5.
- Jamieson AJ, Fujii T, Solan M, Matsumoto AK, Bagley PM, Priede IG. 2009. Liparid and macrourid fishes of the hadal zone: *in situ* observations of activity and feeding behaviour. *Proc R Soc B* 276:1037–45.
- Jamieson AJ, Fujii T, Mayor DJ, Solan M, Priede IG. 2010. Hadal trenches: the ecology of the deepest places on Earth. *Trends Ecol Evol* 25:190–7.
- Jamieson AJ, Kilgallen NM, Rowden AA, Fujii T, Horton T, Lörz A-N, Kitazawa K, Priede IG. 2011. Bait-attending fauna of the Kermadec Trench, SW Pacific Ocean: evidence for an ecotone across the abyssal–hadal transition zone. *Deep Sea Res Part I Oceanogr Res Pap* 58:49–62.
- Jamieson AJ, Malkocs T, Piertney SB, Fujii T, Zhang Z. 2017. Bioaccumulation of persistent organic pollutants in the deepest ocean fauna. *Nature* 1:24–27.
- Kai Y, Orr JW, Sakai K, Nakabo T. 2011. Genetic and morphological evidence for cryptic diversity in the *Careproctus rastrinus* species complex (Liparidae) of the North Pacific. *Ichthyol Res* 58:143–54.
- Kelly RH, Yancey PH. 1999. High contents of trimethylamine oxide correlating with depth in deep-sea teleost fishes, skates, and decapod crustaceans. *Biol Bull* 196:18–25.
- Kim YU, Park YS, Myoung JG. 1986a. Egg development and larvae of snailfish, *Liparis ingens* (Gibert et Burke). *J Korean Fish Soc* 19:368–74.
- Kim YU, Park YS, Myoung JG. 1986b. Egg development and larvae of the snailfish, *Liparis tanakai* (Gilbert et Burke). *J Korean Fish Soc* 19:380–6.
- Lin H-Y, Shiao J-C, Chen Y-G, Iizuka Y. 2012. Ontogenetic vertical migration of grenadiers revealed by otolith microstructures and stable isotopic composition. *Deep Sea Res Part I Oceanogr Res Pap* 61:123–30.
- Linley TD, Gerringer ME, Yancey PH, Drazen JC, Weinstock CL, Jamieson AJ. 2016. Fishes of the hadal zone including



- new species, in situ observations and depth records of Liparidae. *Deep Sea Res Part I Oceanogr Res Pap* 114:99–110.
- Linley TD, Stewart AL, McMillan PJ, Clark MR, Gerringer ME, Drazen JC, Fujii T, Jamieson AJ. 2017. Bait attending fishes of the abyssal zone and hadal boundary: community structure, functional groups and species distribution in the Kermadec, New Hebrides, and Mariana trenches. *Deep Sea Res Part I Oceanogr Res Pap* 121:38–53.
- Marliave JB, Peden AE. 1989. Larvae of *Liparis fucensis* and *Liparis callyodon*: is the “cottoid bubblemorph” phylogenetically significant? *Fish Bull* 87:735–43.
- Märss T, Lees J, Wilson MVH, Saat T, Špilev H. 2010. The morphology and sculpture of ossicles in the Cyclopteridae and Liparidae (Teleostei) of the Baltic Sea. *Est J Earth Sci* 59:263.
- Matallanas J, Pequeno G. 2000. A new snailfish species, *Paraliparis orcadensis* sp. nov. (Pisces: Scorpaeniformes) from the Scotia Sea (Southern Ocean). *Polar Biol* 23:298–300.
- Matarese AC, Blood DM, Picquelle SJ, Benson JL. 2003. Atlas of abundance and distribution patterns of ichthyoplankton from the Northeast Pacific Ocean and Bering Sea ecosystems based on research conducted by the Alaska Fisheries Science Center. NOAA Prof Pap NMFS 1:281.
- Milligan RJ, Morris KJ, Bett BJ, Durden JM, Jones DOB, Robert K, Ruhl HA, Bailey DM. 2016. High resolution study of the spatial distributions of abyssal fishes by autonomous underwater vehicle. *Sci Rep* 6:26095.
- Moller PR, King N. 2007. Two new species of *Pachycara* (Teleostei: Zoarcidae) from the sub-Antarctic southern Indian Ocean, and a range extension of *Lycenchelys antarctica*. *Copeia* 2007:586–93.
- Munk O. 1964. The eyes of three benthic deep-sea fishes caught at great depths. *Galathea Rep* 7:137–49.
- Neuheimer AB, Taggart CT. 2007. The growing degree-day and fish size-at-age: the overlooked metric. *Can J Fish Aquat Sci* 64:375–85.
- Nielsen JG. 1964. Fishes from depths exceeding 6000 meters. *Galathea Rep* 7:113–24.
- Nielsen JG. 1977. The deepest living fish, *Abyssobrotula galathea*, a new genus and species of oviparus ophidioids (Pisces, Brotulidae). *Galathea Rep* 14:41–8.
- Nielsen JG, Merrett NR. 2000. Revision of the cosmopolitan deep-sea genus *Bassozetus* (Pisces: Ophidiidae) with two new species. *Galathea Rep* 18:7–56.
- Nielsen JG, Møller PR. 2011. Revision of the bathyal cusk-eels of the genus *Bassogigas* (Ophidiidae) with description of a new species from off Guam, west Pacific Ocean. *J Fish Biol* 78:783–95.
- Nielsen J, Cohen DM, Markle DF, Robins CR. 1999. Ophidiiform fishes of the world (Order Ophidiiformes): an annotated and illustrated catalogue of pearlfishes, cusk-eels, brotulas, and other ophidiiform fishes known to date. *FAO Fish Synopsis* 18:178 p.
- Oguri K, Noguchi T. 2017. Deepest fish ever recorded, documented at depths of 8,178 m in Mariana Trench. JAMSTEC Press Release.
- Oguri K, Kawamura K, Sakaguchi A, Toyofuku T, Kasaya T, Murayama M, Fujikura K, Glud RN, Kitazato H. 2013. Hadal disturbance in the Japan Trench induced by the 2011 Tohoku–Oki earthquake. *Sci Rep* 3:1915.
- Orr JW. 2004. *Lopholiparis flerxi*: a new genus and species of snailfish (Scorpaeniformes: Liparidae) from the Aleutian Islands, Alaska. *Copeia* 2004:551–5.
- Orr J, Spies I, Stevenson D, Longo G, Kai Y, Ghods S, Hollowed M. Molecular phylogenetics of snailfishes (Liparidae: Cottodei) based on MtDNA and RADseq genomic analyses, with comments on selected morphological characters. *Zootaxa* 125 ms pp. Accepted Manuscript.
- Péres JM. 1965. Aperçu sur les résultats de deux plongées effectuées dans le ravin de Puerto-Rico par le bathyscaphe Archimède. *Deep Res Oceanogr Abstr* 12:883–91.
- Priede IG. 2018. Buoyancy of gas-filled bladders at great depth. *Deep Res Part I Oceanogr Res Pap* 132:1–5.
- Priede IG, Froese R. 2013. Colonization of the deep sea by fishes. *J Fish Biol* 83:1528–50.
- Riemann L, Alfredsson H, Hansen MM, Als TD, Nielsen TG, Munk P, Aarestrup K, Maes GE, Sparholt H, Petersen MI, et al. 2010. Qualitative assessment of the diet of European eel larvae in the Sargasso Sea resolved by DNA barcoding. *Biol Lett* 6:819–22.
- Ritchie H, Jamieson AJ, Piertney SB. 2015. Phylogenetic relationships among hadal amphipods of the Superfamily Lysianassoidea: implications for taxonomy and biogeography. *Deep Sea Res Part I Oceanogr Res Pap* 105:119–31.
- Ryan WBF, Carbotte SM, Coplan JO, O’Hara S, Melkonian A, Arko R, Weissel RA, Ferrini V, Goodwillie A, Nitsche F, et al. 2009a. Global multi-resolution topography synthesis. *Geochem Geophys Geosyst* 10:Q03014.
- Ryan W, Carbotte S, Coplan J, O’Hara S, Melkonian A, Arko R, Weissel R, Ferrini V, Goodwillie A, Nitsche F, et al. 2009b. Global multi-resolution topography (GMRT) synthesis data set. *Geochem Geophys Geosyst* 10:Q03014.
- Scholander PF. 1954. Secretion of gases against high pressures in the swimbladder of deep sea fishes. *Biol Bull* 107:260–77.
- Shiao J, Sui T, Chang N, Chang C. 2017. Remarkable vertical shift in residence depth links pelagic larval and demersal adult jellynose fish. *Deep Res* 121:160–8.
- Shinohara G, Yabe K, Nakaya M, Anma G, Yamaguchi S, Amaoka K. 1994. Deep-sea fishes collected from the North Pacific by the T/S Oshoro-maru. *Bull Fac Fish* 45:48–80.
- Sinenesky M. 1974. Homeoviscous adaptation: a homeostatic process that regulates the viscosity of membrane lipids in *Escherichia coli*. *Proc Natl Acad Sci U S A* 71:522–5.
- Sokolovskii AS, Sokolovskaya TG. 2003. Larvae and juveniles of the genus *Liparis* (Pisces: Liparidae) from the northwestern Sea of Japan. *Russ J Mar Biol* 29:305–15.
- Somero GN. 1992. Adaptations to high hydrostatic pressure. *Annu Rev Physiol* 54:557–77.
- Stein DL. 2005. Descriptions of four new species, redescription of *Paraliparis membranaceus*, and additional data on species of the fish family Liparidae (Pisces, Scorpaeniformes) from the west coast of South America and the Indian Ocean. *Zootaxa* 1019:1–25.
- Stein DL. 2016. Description of a new hadal *Notoliparis* from the Kermadec Trench, New Zealand, and redescription of *Notoliparis kermadecensis* (Nielsen) (Liparidae, Scorpaeniformes). *Copeia* 104:907–20.
- Stein D, Drazen J. 2014. *Paraliparis hawaiiensis*, a new species of snailfish (Scorpaeniformes: Liparidae) and the first

- described from the Hawaiian Archipelago. *J Fish Biol* 84:1519–26.
- Stewart HA, Jamieson AJ. 2018. Habitat heterogeneity of hadal trenches: considerations and implications for future studies. *Prog Oceanogr* 161:47–65.
- Strugnell JM, Rogers AD, Prodöhl PA, Collins MA, Allcock AL. 2008. The thermohaline expressway: the Southern Ocean as a centre of origin for deep-sea octopuses. *Cladistics* 24:853–60.
- Swezey R, Somero G. 1982. Polymerization thermodynamics and structural stabilities of skeletal muscle actins from vertebrates adapted to different temperatures and pressures. *Biochemistry* 21:4496–503.
- Thorrold SR, Campana SE, Jones CM, Swart PK. 1997. Factors determining  $^{13}\text{C}$  and  $^{18}\text{O}$  fractionation in aragonitic otoliths of marine fish. *Geochim Cosmochim Acta* 61:2909–19.
- Thorson G. 1950. Reproduction and larval ecology of marine bottom invertebrates. *Biol Rev* 25:1–45.
- Uiblein F, Nielsen JG, Møller PR. 2008. Systematics of the ophidiid genus *Spectrunculus* (Teleostei: Ophidiiformes) with resurrection of *S. crassus*. *Copeia* 2008:542–51.
- Walkusz W, Paulic JE, Wong S, Kwasniewski S, Papst MH, Reist JD, Sea B. 2016. Spatial distribution and diet of larval snailfishes (*Liparis fabricii*, *Liparis gibbus*, *Liparis tunicatus*) in the Canadian Beaufort Sea. *Oceanologia* 58:117–23.
- Wenzhöfer F, Oguri K, Middelboe M, Turnewitsch R, Toyofuku T, Kitazato H, Glud RN. 2016. Benthic carbon mineralization in hadal trenches: assessment by *in situ*  $\text{O}_2$  microprofile measurements. *Deep Res Part I Oceanogr Res Pap* 116:276–86.
- Wilson RR, Waples RS. 1983. Distribution, morphology, and biochemical genetics of *Coryphaenoides armatus* and *C. yaquinae* (Pisces: Macrouridae) in the central and eastern North Pacific. *Deep Sea Res Part A Oceanogr Res Pap* 30:1127–45.
- Wolff T. 1958. The hadal community, an introduction. *Deep Sea Res* 6:95–124.
- Wolff T. 1970. The concept of the hadal or ultra-abyssal fauna. *Deep Res Oceanogr Abstr* 17:983–1003.
- Yancey PH, Siebenaller JF. 1999. Trimethylamine oxide stabilizes teleost and mammalian lactate dehydrogenases against inactivation by hydrostatic pressure and trypsinolysis. *J Exp Biol* 202:3597–603.
- Yancey PH, Siebenaller JF. 2015. Co-evolution of proteins and solutions: protein adaptation versus cytoprotective micromolecules and their roles in marine organisms. *J Exp Biol* 218:1880–96.
- Yancey PH, Gerrerger ME, Drazen JC, Rowden AA, Jamieson AJ. 2014. Marine fish may be biochemically constrained from inhabiting the deepest ocean depths. *Proc Natl Acad Sci U S A* 111:4461–5.

### Synopsis Sobre o Sucesso dos Snailfishes Hadal

As fossas oceânicas profundadas (do inglês “deep-sea trenches”), cujas profundidades variam entre 6000 e 11.000 m, são caracterizadas por altas pressões, baixas temperaturas, e ausência de luz solar. Esses padrões caracterizam a maior parcela dos habitat marinho mais profundos—a zona hadal—região que abriga comunidades distintas se comparadas às planícies abissais adjacentes. Os peixe-caracóis, família Liparidae (Scorpaeniformes), obtiveram notável sucesso na zona hadal entre 6000–8200 m, faixa que abriga a ictiofauna dominante em pelo menos seis fossas distintas ao redor do mundo. A comunidade de peixes hadais é distinta da comunidade abissal, onde carneiros de corpo alongado, como os peixes granadeiros (Macrouridae), Sinafobranquídeos, peixes-tripé (Ipnopidae), Zoarcídeos, e cobrelos (Ophidiidae) são os mais comuns. Até recentemente, pouco se sabia sobre a biologia desses peixes que habitam regiões mais profundas, ou os fatores que impulsionam seu sucesso na em profundidades hadais. Aqui, eu reviso a literatura recente que compreende a fronteira abissal–hadal e discuto os fatores que estruturam essas comunidades, incluindo os papéis da adaptação à pressão, ecologia de alimentação e história de vida. Os peixes da regioao hadal apresentam uma adaptações específicas à pressão hidrostática, tanto no acúmulo do neutraliante de pressão *N*-óxido de trimetilamina quanto em alterações intrínsecas em enzimas. As análises de conteúdo estomacal e isótopos de aminoácidos e a morfologia mandibular sugerem que peixes predadores com alimentação por sucção, como os liparídeos da zona hadal, podem encontrar uma vantagem ao descender para as fossas, onde anfípodes são cada vez mais abundantes. A análise das zonas de crescimento dos otólitos sugerem que os peixes-caracóis podem estar adaptados a ambientes hadais altamente perturbados e sismicamente ativos por terem longevidade relativamente curta. Esta revisão sintetiza a literatura conhecida sobre os peixes dos ambientes mais profundos do planeta e apresenta uma nova interpretação das adaptações à vida em fossas oceânicas.

Translated to Portuguese by J.P. Fontenelle (jp.fontenelle@mail.utoronto.ca)

**Synopsis** Über den Erfolg der Scheibenbäuche des Hadals Tiefseeegräben mit einer Tiefe von 6000–11.000 m zeichnen sich durch hohe Drücke, niedrige Temperaturen, und der Abwesenheit von Sonnenlicht aus. Diese Eigenschaften machen den Großteil des tiefsten Lebensraums der Meere—der Hadal-Zone—aus, wo Gemeinschaften zu Hause sind, die sich distinkt von jenen in den umliegenden Abyssal-Ebenen unterscheiden. Die Scheibenbäuche, Familie Liparidae (Scorpaeniformes), haben in der Hadal-Zone von ~6000–8200 m einen beachtlichen Erfolg erzielt, sie stellen die dominierende Ichthyofauna in mindestens sechs Gräben weltweit. Die Gemeinschaft der Fische des Hadals unterscheidet sich von der abyssischen Gemeinschaft, in der vor allem längliche, aasfressende Fische wie Grenadierfische (Macrouridae), Grubenaale (Synphobranchidae), Netzaugenfische (Ipnopidae), Aalmuttern (Zoarcidae) und Bartmännchen (Ophidiidae) häufig sind. Bis vor kurzem war wenig über die Biologie dieser am tiefsten lebenden Fische bekannt oder über die Faktoren, die ihren Erfolg in den Tiefe des Hadals beeinflussen. Ich bespreche hier jüngste Untersuchungen, unter Einbezug der Grenze zwischen dem Abyssal und dem Hadal, und diskutiere dabei die Faktoren welche diese Gemeinschaften strukturieren, einschließlich der Rolle von Druckanpassungen, Nahrungsökologie und Life History. Fische des Hadals zeigen eine spezialisierte Anpassung an den hydrostatischen Druck sowohl durch die Ansammlung des druckgegenwirkenden Trimethylamin-*n*-oxids als auch durch intrinsische Veränderungen von Enzymen. Analysen von Mageninhalt und Aminosäure-Isotopen sowie die Morphologie des Kiefers legen nahe, dass Raubfische, welche ihre Nahrung durch Ansaugen aufnehmen, wie Hadal-Lipariden, einen Vorteil durch den Abstieg in den Graben haben könnten, wo Amphipoden zunehmend häufiger sind. Die Analyse der Otolith-Wachstumszonen lässt vermuten, dass Scheibenbäuche an eine seismisch aktive, viel gestörte Umwelt im Hadal durch eine relativ kurze Lebensdauer angepasst sein könnten. Dieser Aufsatz fasst die bekannte Literatur über die am tiefsten lebenden Fische des Planeten zusammen und vermittelt ein neues Verständnis von Anpassungen an das Leben in den Tiefseeegräben.

Translated to German by F. Klimm (frederike.klimm@neptun.uni-freiburg.de)