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# Fascioliasis Due to *Fasciola hepatica* and *Fasciola gigantica* Infection: An Update on This 'Neglected' Neglected Tropical Disease

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#### Abstract

Fascioliasis, an infection due to the food- and water-borne trematodes *Fasciola hepatica* and *Fasciola gigantica*, is among the most neglected of the neglected tropical diseases. Among the estimated 91.1 million humans at risk for infection worldwide, as many as 17 million may be infected. Certain areas of the world bear the burden of the highest prevalence of

After reading this article, readers should be able to recognize the different morphologies of eggs from a variety of human parasites, understand the role of laboratory testing in the diagnosis of human fascioliasis, and appreciate the clinical and epidemiologic circumstances in which the diagnosis of infection with *Fasciola* is appropriate.

The neglected tropical diseases (NTD) comprise a group of (mostly infectious) conditions uncommonly recognized or diagnosed in the United States and the developed world; are less well understood than more common infections due to a of lack of research interest and/or insufficient funding; and, lastly, remain mysterious or unknown to health care providers because of minimal or no instruction regarding the diseases during our training.<sup>1</sup> However, the outlook for the NTD is

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#### Abbreviations

NTD, neglected tropical diseases; Fh, *Fasciola hepatica*; Fg, *Fasciola gigantica*; FBT, food-borne trematodiases; FAST-ELISA, Falcon assay screening test - enzyme-linked immunosorbent assay; EITB, enzyme-linked immunoelectrotransfer blot; ERCP, endoscopic retrograde cholangiopancreatography; MRCP, magnetic resonance cholangiopancreatography

infection. There, school-age children are the most likely to be infected. In the United States, rare cases have been reported among immigrants from endemic areas, returned travelers to endemic regions, and individuals residing in Hawaii, California, and Florida. Indigenous cases have almost always been associated with consumption of watercress. Diagnosis is made serologically most often, although stool examination for the eggs is fruitful if obtained when the adult worm is laying eggs. With an appropriate index of suspicion, laboratory and imaging studies often confirm the suspected diagnosis. Triclabendazole is the treatment of choice.

**Keywords:** Fascioliasis, *Fasciola hepatica*, *Fasciola gigantica*, trematodiasis, neglected tropical diseases, liver fluke

**Microbiology exam 71101** questions and corresponding answer form are located after this CE Update on page ##.

improving. For example, a new journal, PLoS Neglected Tropical Diseases, recently appeared with support of the Bill and Melinda Gates Foundation, which is also a pioneer in supporting the study of these diseases.<sup>2</sup> However, certain foodborne trematode infections in particular remain "neglected" NTD, according to the World Health Organization.<sup>1</sup> These include clonorchiasis (Chinese liver fluke disease), fascioliasis (sheep liver fluke disease), opisthorchiasis (fish liver fluke disease), and paragonimiasis (lung fluke disease). These diseases most often significantly affect large numbers of povertystricken individuals, generally in resource-limited regions, and receive very little interest from funding or government agencies.<sup>1</sup> As these diseases have complex life cycles and are rarely encountered in the United States, they receive little attention in the education of physicians, which furthers their enigmatic status.<sup>3</sup> This continuing education update will address the most commonly encountered (in the United States) of these "neglected" NTD, fascioliasis. The interested reader is referred to recent reviews of this topic for further information.<sup>3-11</sup>

#### Fasciola Epidemiology

Fascioliasis, a food- or water-borne trematodiasis due to infection by *Fasciola hepatica* (*Fh*) or *F. gigantica* (*Fg*), is currently

#### **CE Update**

#### Glossary

Autochthonous Coproantigens Coprological Originating where found; indigenous Antigens found in stool specimens Having to do with stool (for example, stool studies)

believed to affect as many as 17 million people worldwide,<sup>12</sup> with 91.1 million individuals at risk for infection.<sup>4</sup> Fasciola hepatica infects humans on all continents (except Antarctica), having the widest latitudinal, longitudinal, and altitudinal distribution of the food-borne trematodiases (FBT) and other parasitic and vector-borne diseases.<sup>13</sup> In contrast, Fg infection is more geographically constricted, occurring in the tropical regions of Africa, the Middle East, and Asia, where infection due to either species may occur.<sup>5</sup> Contrary to early thinking, prevalence of veterinary disease is not predictive of prevalence of human disease in endemic regions.<sup>13</sup> Prior to 1980, fascioliasis was thought to be a zoonosis of only mild, sporadic, or local importance. In the past 20 years however, a more complete understanding of disease epidemiology has emerged.<sup>14</sup> Mas-Coma and colleagues have proposed an epidemiologically useful classification, based on more recent studies, to better understand fascioliasis.14

Imported cases are those acquired in an endemic area but diagnosed in a region lacking indigenous fascioliasis, even in animals. Imported fascioliasis in the returned traveler exemplifies this situation.

Autochthonous, isolated, non-constant cases occur in areas where animal and human infections are recognized but are sporadic and not endemic. Indigenous cases resulting from ingestion of contaminated watercress in Hawaii are examples.

Endemic regions can be described as hypoendemic, mesoendemic, or hyperendemic, depending on whether coprological diagnosis on inhabitants reveals a prevalence of less than 1%, 1%-10%, or greater than 10%, respectively. Typically, in hypoendemic regions sanitation usually involves

#### Figure 1\_Key Points

- Most cases of fascioliasis seen in the United States will occur in immigrants and travelers returning from endemic regions
- Most cases worldwide occur in school-age children living in endemic areas (but those cases do not come to medical attention in the United States)
- Unlike other FBT, fascioliasis most often follows ingestion of contaminated aquatic vegetation or water—so vegetarians are at risk
- A high index of suspicion is necessary
- Acute disease is typified by fever, abdominal pain, and eosinophilia but no eggs in the stool
- Chronic disease is most often asymptomatic, but eggs may be shed and serology is positive
- Human Fasciola infection cannot be speciated on the basis of egg size
- Imaging studies often support the laboratory diagnosis

latrines or sewage disposal systems and outdoor defecation is unusual. In contrast, in hyperendemic areas, indiscriminate defecation is common, as latrines and waste management systems are absent. Human egg shedding contributes significantly to disease transmission in this circumstance.

Epidemic outbreaks can be characterized as occurring in areas where fascioliasis is endemic in animals but not humans and in areas where the disease is endemic in both animals and humans.

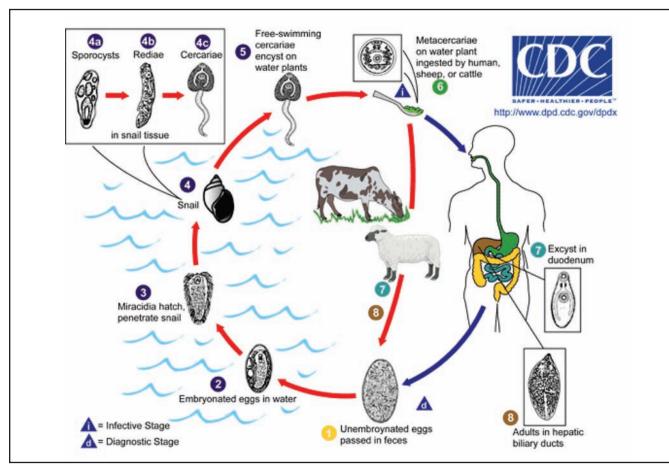
Areas of the world with the highest prevalence of fascioliasis include the highlands of Bolivia (the Bolivian Altiplano has the highest prevalence in the world<sup>14</sup>), Ecuador, and Peru; Cuba; Portugal and Spain; Turkey; the Nile Delta of Egypt and elsewhere in Africa; the northern regions of the Islamic Republic of Iran and elsewhere in Asia; and central Vietnam.<sup>7</sup>

Imported fascioliasis in the United States (and other developed countries) has been extensively reviewed.<sup>8,15-19</sup> Twenty-three cases of imported fascioliasis diagnosed in the United States have been summarized.<sup>19</sup> A handful of indigenous cases, primarily associated with consumption of contaminated watercress, have been identified in Hawaii, California, and Florida, as well.<sup>19</sup> Most cases seen in the United States will involve immigrants from endemic areas and returned travelers infected while visiting such regions (**Figure 1**).

The life cycle of *Fasciola* is shown in Figure 2.<sup>20</sup> Understanding the interaction between the parasite and its accidental human host at a molecular pathophysiological level is resulting from recent investigations.<sup>21</sup> Unlike the other FBT, in which consumption of raw or undercooked seafood is necessary for infection, humans most often become accidental hosts of Fasciola when they ingest aquatic vegetation on which the metacercariae have encysted. Implicated plants include (most commonly) watercress,<sup>6</sup> water morning glory (pak boong),<sup>8</sup> other aquatic plants, salads in endemic areas such as the highlands of South America,<sup>22</sup> and alfalfa juice (which is used as a medicinal tonic in Peru).<sup>23</sup> Another source of infection is drinking water contaminated with free-living (non-encysted) metacercariae. The least common mode of transmission is the consumption of raw or undercooked liver infected with immature or adult forms of the worm.

#### **Clinical Manifestations**

Given that more than half of cases are subclinical (asymptomatic),<sup>6</sup> human infection can be classified as acute or chronic based upon clinical manifestations and laboratory findings. These 2 classifications have been further subdivided into 6 phases (Table 1).<sup>7</sup> In a review of 173 cases,<sup>15</sup> clinical manifestations included abdominal pain in 125 (72%), fever in 99 (57%), constitutional symptoms in 76 (44%), urticaria in 22 (13%), itching in 17 (10%), respiratory symptoms in 14 (8%), headache in 8 (5%), and cardiac symptoms in 2 (1%). Laboratory and other evaluations revealed eosinophilia (>500/mm<sup>3</sup>) in 96%, leukocytosis (>10,000/mm<sup>3</sup>) in 64%, eggs in feces in 40%, and positive computed tomography in 70%. In chronic infection beyond the latent phase, clinical manifestations are those of the complications of fascioliasis, namely ascending cholangitis, cholelithiasis, cholecystitis, pancreatitis, biliary cirrhosis, and hepatic fibrosis. Unlike other FBT, fascioliasis has not yet been associated with the development of cholangiocarcinoma.<sup>24</sup> Typical fascioliasis results from the migration of immature worms through Glisson's



**Figure 2**\_Life cycle of *Fasciola*. Immature eggs are discharged in the biliary ducts and in the stool **①**. Eggs become embryonated in water **②**, eggs release miracidia **③**, which invade a suitable snail intermediate host **④**, including the genera *Galba, Fossaria*, and *Pseudosuccinea*. In the snail the parasites undergo several developmental stages (sporocysts **④**a, rediae **④**b, and cercariae **④**c). The cercariae are released from the snail **⑤** and encyst as metacercariae on aquatic vegetation or other surfaces. Mammals acquire the infection by eating vegetation containing metacercariae. Humans can become infected by ingesting metacercariae-containing freshwater plants, especially watercress **③**. After ingestion, the metacercariae excyst in the duodenum **⑦** and migrate through the intestinal wall, the peritoneal cavity, and the liver parenchyma into the biliary ducts, where they develop into adults **③**. In humans, maturation from metacercariae into adult flukes takes approximately 3 to 4 months. The adult flukes (*F. hepatica*: up to 30 mm by 13 mm; *F. gigantica*: up to 75 mm) reside in the large biliary ducts of the mammalian host. *F. hepatica* infect various animal species, mostly herbivores. From the Centers for Disease Control and Prevention Laboratory Identification of Parasites of Public Health Concern website.<sup>20</sup>

	Phase	Time Frame	Pathophysiology	Clinical Manifestations	Laboratory Findings
Acute	Incubation	Few days to few months	From ingestion of metacercariae to appearance of symptoms when they reach the liver parenchyma or other ectopic site	None	None
	Invasive	2-4 months	Worms migrate through liver tissue, causing necrosis, hemorrhage, and inflammation	Fever, abdominal pain, gastrointestinal disturbances, rashes, cough, hepatospleno- megaly, ascites, jaundice, anemia, right upper quadrant tenderness	High levels of eosinophils and immuno- globulin E often noted. Imaging and/or histopathologic studies often abnormal
	Latency	Months or years	Parasites mature and start to lay eggs when they reach the bile ducts	None or mild gastrointestinal disturbances with vague abdominal pain or tenderness	Intermittent eosinophilia may occur; eggs are shed in the stool intermittently
	Obstructive	Months or years	Parasites, fragments, and/or debris obstruct the biliary tree	Biliary colic, nausea, epigastric pain, jaundice, fever, right upper quadrant tenderness	Leukocytosis with cholangitis; anemia may ensue; imaging may be consistent with cholecystitis or reveal worms in the bile ducts
Chronic	Advanced chronic	Years	Consequences of long-term obstruction include calculi, bacterobilia, cholangitis, and cholecystitis	Chronic cholecystitis and cholangitis	Findings consistent with cholecystitis or cholangitis; egg laying ceases
	Post- infectious	Years	Most flukes have died, but calculi may persist with biliary cirrhosis and gallbladder atony	Chronicity of cholecystitis and/or cholangitis	Findings consistent with chronic cholecystitis or cholangitis may be seen

#### Figure 3\_Reference Laboratory for Fasciola Serology

Falcon assay screening test—enzyme-linked immunosorbent assay (FAST-ELISA) and enzyme-linked immunoelectrotransfer blot (EITB)^{28,29}

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Reference Laboratory for Fasciola Immunofluorescence on Worm Sections

For clinical consultation and immunofluorescence on worm sections:

Professor Jean Dupouty-Camet Parasitologie-Mycologie UFR Cochin 27 Fauberg St. Jacques, 75014 Paris, France

Telephone: 33-1-4234-1497 Fax: 33-1-4234-1496 Email: dupouyca@imaginet.fr and jean.dupouy-camet@cch.ap-hop-paris capsule and the liver parenchyma en route to the bile ducts, where they mature and take up permanent residence. Occasionally the metacercariae reach ectopic destinations, resulting in a cutaneous or visceral larva migrans picture reminiscent of strongyloidiasis, taeniasis, or gnathostomiasis.<sup>25-27</sup> The clinical findings with ectopic migration will depend upon the organs or tissues invaded.

#### **Diagnostic Approach to Fascioliasis**

Clearly, the appropriate diagnostic approach to the patient with possible fascioliasis depends first and foremost on a high index of suspicion, as well as the stage of the infection and the resources and expertise available.

#### Laboratory Evaluation

Eosinophilia, leukocytosis, and elevated inflammatory markers are common in acute infections (**Table 1**); anemia and/or elevated serum hepatic transaminases, bilirubin, or alkaline phosphatase are only occasionally transiently present during chronic infection. Within 2 to 4 weeks of becoming infected, a *Fasciola*-specific serologic response ensues, allowing confirmation of infection 5 to 7 weeks before eggs appear in

#### Table 2\_Differential Morphology of the Diagnostic Stages of Helminths Found in Humans: Eggs (Trematodes)

Species	Size	Shape	Color	Stage of Development When Passed
Schistosoma mansoni	140 µm × 66 µm Range, 114-180 µm × 45-73 µm	Elongated with prominent lateral spine near posterior end. Anterior end tapered and slightly curved.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Schistosoma japonicum	90 μm × 70 μm Range, 68-100 μm × 45-80 μm	Oval. Small lateral spine is often seen or may appear as a small hook or "knob" located in a depression in the shell.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Schistosoma haematobium	143 μm × 60 μm Range, 112-170 μm × 40-70 μm	Elongated with rounded anterior end and terminal spine at posterior end.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Schistosoma intercalatum	175 μm × 60 μm Range, 140-240 μm × 50-85 μm	Elongated with tapered anterior end and terminal spine. Sometimes spindle-shaped.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Schistosoma mekongi	69 μm × 56 μm* Range, 51-73 μm × 39-66 μm	Spherical. Small lateral spine, not always visible or may appear as a small "knob" in a depression in the shell.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Clonorchis sinensis	30 μm × I6 μm Range, 27-35 μm × 11-20 μm	Small, ovoidal, or elongated with broad rounded posterior end and a convex operculum resting on "shoulders." A small "knob" may be seen on the posterior end.	Yellow brown	Embryonated. Contains mature miracidium.
Opisthorchis felineus	30 μm × 12 μm Range, 26-30 μm × 11-15 μm	Elongated with operculum on anterior end and pointed terminal "knob" on posterior end.	Yellow brown	Embryonated. Contains mature miracidium.
Heterophyes heterophyes	28 μm × 15 μm Range, 28-30 μm × 15-17 μm	Small, elongated, or slightly ovoidal. Operculum. Slight "knob" at posterior end.	Yellow brown	Embryonated. Contains mature miracidium.
Metagonimus yokogawai	28 μm × 17 μm Range, 26-30 μm × 15-20 μm	Small, elongated, or ovoidal. Operculum. No "shoulders" at anterior end. Small "knob" often seen on posterior end.	Yellow or yellow brown	Embryonated. Contains mature miracidium.
Paragonimus westermani	85 μm × 53 μm Range, 68-118 μm × 39-67 μm	Ovoidal or elongated with thick shell. Operculum is slightly flattened and fits into shoulder area of shell. Posterior end is thickened. Egg often asymmetrical with 1 side slightly flattened.	Yellow brown to dark brown	Unembryonated. Filled with yolk material in which a germinal cell is imbedded. Cells are irregular in size.
Fasciola hepatica	145 μm × 80 μm Range, 120-150 μm × 63-90 μm	Ellipsoidal, thin shell. Small, indistinct operculum.	Yellow to light brown	Unembryonated. Filled with yolk cells in which an indistinct germinal cell is imbedded
Fasciolopsis buski	140 μm × 80 μm Range, 130-159 μm × 78-98 μm	Ellipsoidal, thin shell. Small, indistinct operculum.	Yellow brown.	Unembryonated. Filled with yolk cells in which an indistinct germinal cell is imbedded.

From the Centers for Disease Control and Prevention Parasites and Health Web site,<sup>41</sup> based on sizes of eggs in human fecal specimens.<sup>42,4</sup>

the feces. In the United States, serology is not commercially available and the Centers for Disease Control and Prevention refers Fasciola serology requests to a reference laboratory (Figure 3).<sup>28,29</sup> Despite some cross-reactivity of the Falcon assay screening test-enzyme-linked immunosorbent assay (FAST-ELISA) with serum from those with schistosomiasis (which generally can be clarified by the clinical scenario), the sensitivity of the test is reported to be 95%.<sup>30</sup> The sensitivity of the confirmatory enzyme-linked immunoelectrotransfer blot (EITB), using 12-, 17-, and 63-kiloDalton antigens, is reported to be 100%.<sup>30</sup> Positive serology can also be used to document infection in the chronic phase, when egg release may be intermittent or absent. Serology should revert to negative within a year of successful treatment. A variety of serodiagnostic methods have been described,<sup>31-39</sup> including 2 commercially available testing kits available in Europe and elsewhere.38,39

An ELISA assay to detect *Fh* coproantigens in preserved human stool samples has been described,<sup>40</sup> but the standard fecal study for the diagnosis of fascioliasis is the identification of eggs in the stool. In hyperendemic settings, children may shed thousands of eggs per gram of feces.<sup>3</sup> A tabular comparison (**Table 2**)<sup>41-43</sup> and **Figure 4**<sup>41</sup> illustrate the similarities and differences among the eggs of most human trematodes. **Figure 5** shows unstained wet mounts of *Fh* eggs.<sup>20</sup> Infection due to *Fh* and *Fg* classically has been differentiated by the

#### **Specific Features and Variations**

Lateral spine. Found in feces; in rare cases, in urine also. Eggs are discharged at irregular intervals and may not be found in every stool specimen. They are rare in chronic stages of infection.

Found in feces. Often coated with debris and may be overlooked.

- Terminal spine. Found in urine, occasionally in feces. Egg often covered with debris.
- Terminal spine long, slender with bent tip. Resembles *S. haematobium* egg except it is longer, is thinner, and has a longer spine. Found in feces. May have debris adhering to shell.
- Found in feces. Closely resembles *S. japonicum* egg except it is smaller. May be coated with debris.
- Small size, operculum and "knob" on posterior end. Shell often is covered by adhering debris.

Lacks prominent shoulders characteristic of Clonorchis and has more tapered end.

- Resembles *Clonorchis* egg but with less distinct "shoulders." Operculum is broader than in *Clonorchis*.
- Resembles *Clonorchis* and *Heterophyes* eggs. Shell is slightly thinner than *Heterophyes*. Operculum is broader than *Clonorchis*.
- Found in sputum, occasionally in feces. Resembles egg of *D. latum* but is larger, slightly asymmetrical, and the operculum is smaller and flatter. The widest part of the *Paragonimus* egg is usually anterior to the center; in a *D. latum*, the widest area is around the center.

Large size. Broadly oval eggs.

Large size. Resembles *F. hepatica* egg and cannot be easily distinguished from *Fasciola*.

geographic distribution of the organisms, when the 2 do not overlap. In regions endemic for both, however, it was thought that the size of the eggs shed in the feces could differentiate the 2 kinds. In livestock, the eggs of Fg are significantly larger than those shed by Fh, and this was thought to be true for human shedding as well. Recent work corrects this teaching, demonstrating that Fh eggs shed by humans are larger than those shed by livestock, while Fg eggs shed by humans are smaller than those shed by livestock, with considerable overlap. Thus, one cannot speciate human fascioliasis on the basis of the size of the eggs shed in the stool.<sup>44</sup>

Adult worms are found in the biliary tract, while migrating larvae can be found anywhere in the human host (most often in the skin or other sites of ectopic migration). Rarely, a worm will be retrieved from the patient (for example, by endoscopic retrograde cholangiopancreatography [ERCP]).<sup>45</sup> A comparison among the adult visceral trematodes of humans is demonstrated in **Figure 6**.<sup>41</sup> Unstained, formalin-fixed and carmine-stained adult *Fh* worms are shown in **Figure 7**.<sup>20</sup> An immunofluorescent antibody test with 90% sensitivity and specificity can be performed on worm sections, if the diagnosis is in doubt (**Figure 3**).

#### **Diagnostic Imaging**

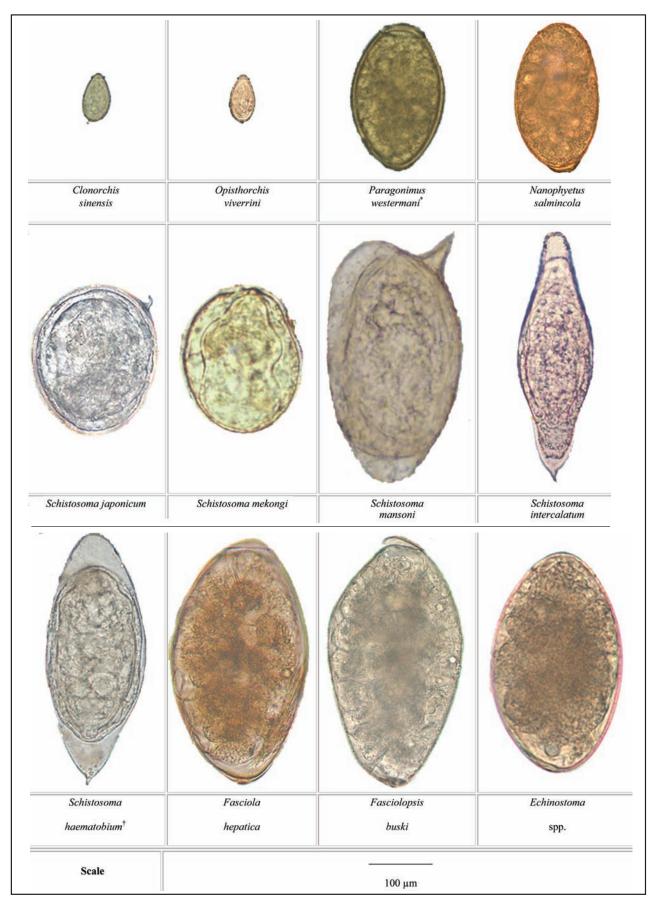
In the appropriate clinical context, imaging studies can be very useful to confirm the diagnosis of fascioliasis.<sup>46-48</sup> Typical findings include subcapsular hemorrhage; parenchymal nodular, ill-defined lesions, which may coalesce into tortuous or tubular tracks; filling defects in the biliary tract; and subcapsular, peribiliary, or periportal indistinct nodules and abscesses. Ultrasound, computed tomography, magnetic resonance imaging, and, to a lesser extent, nuclear medicine scanning all have a role. Endoscopic retrograde cholangiopancreatography and magnetic resonance cholangiopancreatography (MRCP) have been found to be useful in chronic disease when the adult worms occupy the biliary tree.

#### **Histopathology**

Rarely, a biopsy is performed, and a tissue diagnosis is made, most often when the diagnosis has not been considered. With ERCP removal of a worm, diagnosis can be confirmed as above. In ectopic fascioliasis, removal of the migrating metacercariae is diagnostic, as well. A liver biopsy typically reveals necrosis, acute and chronic inflammatory changes, debris, and occasionally fragments of migrating larvae.

#### **Treatment of Fascioliasis**

Unlike other helminth infections, fascioliasis responds very poorly to treatment with albendazole or praziquantel. The reason(s) for this lack of treatment effectiveness are not understood. Currently, the drug of choice is triclabendazole,<sup>49-52</sup> which is not commercially available in the United States. It may be ordered from Victoria Pharmacy, Zurich, Switzerland.<sup>53</sup> Outside of the United States, it is available from the manufacturer, Novartis (Basel, Switzerland), for both treatment of individual cases and for mass administration as part of



**Figure 4**\_Trematode eggs found in stool specimens of humans. From the Centers for Disease Control and Prevention Parasites and Health Web site.<sup>41</sup> <sup>1</sup>Usually found in respiratory specimens. <sup>†</sup>Usually passed in urine.

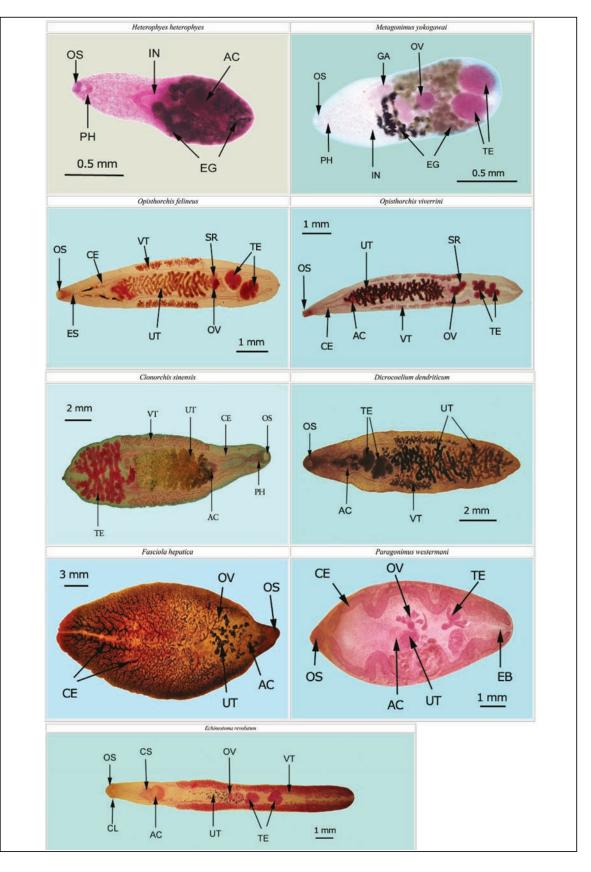


national control strategies.<sup>7</sup> The dose is 10 mg/kg by mouth once or twice, given with food. Alternatives include bithionol, available from the Centers for Disease Control and Prevention Drug Service, Atlanta, GA. The dose is 30-50 mg/kg by mouth on alternate days for 10-15 doses.<sup>53</sup> More recently, studies have suggested that nitazoxanide may be an effective alternative as well.<sup>53,54</sup> The dose is 500 mg by mouth twice daily for 7 days for adults. The dose is reduced to 100 mg twice daily for children aged 1-3 years and 200 mg twice daily for children aged 4-11 years. Those 12 years and above receive the adult dose.<sup>53</sup> There may be a role for treatment with artesunate in certain settings, based upon a pilot study.<sup>55</sup> As with all such diseases, prevention of infection by provision of clean water and safe food, control of sewage and waste, and healthy travel hygiene is preferable to treatment of established infection.

#### Conclusion

Fascioliasis is a disease rarely encountered in the United States. As a result, correct diagnosis is often delayed because the infection is not considered. As the laboratory plays an important role in confirming the diagnosis, it is important to consider the disease in a patient with risk factors and proceed with appropriate testing and treatment. *Acknowledgments:* The author would like to thank the medical library at Saint Peter's University Hospital for assistance with the literature review and Doctors Parvin Alizadeh, David Alcid, Kristina Feja, and Lawrence Frenkel for critical review of the manuscript.

- World Health Organization. The "Neglected" Neglected Worms. Action Against Worms. December 2007, issue 10. Available at: www.who.int/ neglected\_diseases/preventive\_chemotherapy/Newsletter10.pdf. Accessed July 6, 2010.
- 2. PLoS Neglected Tropical Diseases: A Peer-Reviewed Open Access Journal. Available at: www.plosntds.org/home.action. Accessed July 22, 2010.
- Mas-Coma S, Valero MA, Bargues MD. Fasciola, lymnaeids and human fascioliasis, with a global overview on disease transmission, epidemiology, evolutionary genetics, molecular epidemiology and control. *Adv Parasitol*. 2009;69:41-146.
- Keiser J, Utzinger J. Emerging foodborne trematodiasis. *Emerg Infect Dis.* 2005;11:1507-1514.
- 5. Keiser J, Utzinger J. Food-borne trematodiases. *Clin Microbiol Rev.* 2009;22:466-483.
- Tolan RW, Jr. 2001. Fascioliasis. eMedicine from WebMD. Updated June 15, 2010. Available at: emedicine.medscape.com/article/997890-overview. Accessed July 6, 2010.
- 7. World Health Organization. Report of the WHO Informal Meeting on Use of Triclabendazole in Fascioliasis Control. WHO Headquarters, Geneva, Switzerland: October 17-18, 2006. Available at: www.who.int/neglected\_ diseases/preventive\_chemotherapy/WHO\_CDS\_NTD\_PCT\_2007.1.pdf. Accessed July 6, 2010.



#### Figure 6

- AC = acetabulum (ventral sucker) IN = intestine
- CE = cecum •

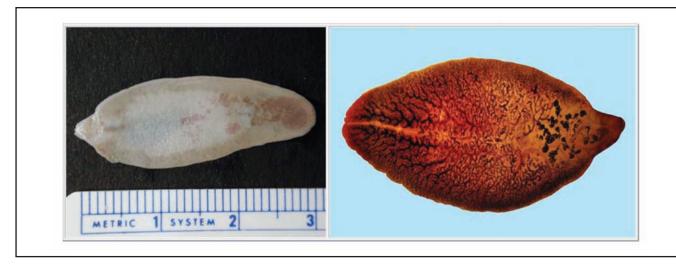
• CS = cirrus sac

CL = collar

- OS = oral sucker • • OV = ovary
  - PH = pharynx
- From the Centers for Disease Control and Prevention Parasites and Health Web site.<sup>41</sup>

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- EB = excretory bladder EG = eggs (within uterus)
- ES = esophagus •
- GA = genitoacetabulum
- SR = seminal receptacle
- TE = testes
- UT = uterus
- VT = vitellaria



**Figure 7**\_Adult *Fasciola hepatica*. Adults of *F. hepatica* are large and broadly flattened, measuring up to 30 mm long and 15 mm wide. The anterior end is cone shaped, unlike the rounded anterior end of *Fasciolopsis buski*. Adults reside in the bile ducts of the liver in the definitive host. Formalin-fixed, unstained (left) and carmine-stained adult *F. hepatica* (right). From the Centers for Disease Control and Prevention Laboratory Identification of Parasites of Public Health Concern Web site.<sup>20</sup>

- Price TA, Tuazon CU, Simon GL. Fascioliasis: Case reports and review. *Clin Infect Dis.* 1993;17:426-430.
- Garcia HH, Moro PL, Schantz PM. Zoonotic helminth infections of humans: Echinococcosis, cysticercosis and fascioliasis. *Curr Opin Infect Dis.* 2007;20:489-494.
- Marcos LA, Terashima A, Gotuzzo E. Update on hepatobiliary flukes: Fascioliasis, opisthorchiasis and clonorchiasis. *Curr Opin Infect Dis.* 2008;21:523-530.
- Alatoom A, Cavuoti D, Southern P, et al. *Fasciola hepatica* infection in the United States. *Lab Med.* 2008;39:425-428.
- 12. Hopkins DR. Homing in on helminths. Am J Trop Med Hyg. 1992;46:626-634.
- 13. Mas-Coma S. Epidemiology of fascioliasis in human endemic areas. *J Helminth*. 2005;79(3):207-216.
- 14. Mas-Coma MS, Esteban JG, Bargues MD. Epidemiology of human fascioliasis: A review and proposed new classification. *Bull WHO*. 1999;77:340-346.
- Arjona R, Riancho JA, Aguado JM, et al. Fascioliasis in developed countries: A review of classic and aberrant forms of the disease. *Medicine* (Baltimore). 1995;74;13-23.
- Graham CS, Brodie SB, Weller PF. Imported Fasciola hepatica infection in the United States and treatment with triclabendazole. Clin Infect Dis. 2001;33:1-5.
- 17. Jensenius M, Flægstad T, Stenstad T, et al. Fascioliasis imported to Norway. Scand J Infect Dis. 2005;37:534-537.
- Kang ML, Teo CHY, Wansaicheong GKL, et al. *Fasciola hepatica* in a New Zealander traveler. *J Travel Med.* 2008;15:196-199.
- Fried B, Abruzzi A. Food-borne trematode infections of humans in the United States of America. *Parasitol Res.* 2010;106:1263-1280.
- 20. Centers for Disease Control and Prevention. Laboratory Identification of Parasites of Public Health Concern. Available at: www.dpd.cdc.gov/dpdx/ HTML/ImageLibrary/Fascioliasis\_il.htm. Accessed July 23, 2010.
- Robinson MW, Menon R, Donnelly SM, et al. An integrated transcriptomics and proteomics analysis of the secretome of the helminth pathogen *Fasciola hepatica*: Proteins associated with invasion and infection of the mammalian host. *Mol Cell Proteomics*. 2009;8:1891-1907.
- Marcos L, Maco V, Terashima A, et al. Fascioliasis in relatives of patients with *Fasciola hepatica* infection in Peru. *Rev Inst Med Trop Sao Paulo*. 2005;47:219-222.
- Marcos L, Maco V, Samalvides F, et al. Risk factors for *Fasciola hepatica* infection in children: A case-control study. *Trans R Soc Trop Med Hyg.* 2006;100:158-166.
- Harinasuta T, Pungpak S, Keystone JS. Trematode infections. Opisthorchiasis, clonorchiasis, fascioliasis, and paragonimiasis. *Infect Dis Clin North Am.* 1993;7:699-716.
- Xuan LT, Hung NT, Waikagul J. Cutaneous fascioliasis: A case report in Vietnam. Am J Trop Med Hyg. 2005;72:508-509.

- Zhou L, Luo L, You C, et al. Multiple brain hemorrhages and hematomas associated with ectopic fascioliasis in brain and eye. *Surg Neurol.* 2008;69:516-521.
- Ying M, Xiaosu H, Wang B. A case of ectopic parasitism: *Fasciola bepatica* larvae burrow through a human brain and mimic cerebral aneurysm. *Trans R Soc Trop Med Hyg*, 2007;101:1051-1052.
- Espino AM, Duménigo BE, Fernández R, et al. Immunodiagnosis of human fascioliasis by enzyme-linked immunosorbent assay using excretory-secretory products. *Am J Trop Med Hyg.* 1987;37:605-608.
- 29. Hillyer GV, Soler de Galanes M, Rodriguez-Perez J, et al. Use of the Falcon assay screening test-enzyme-linked immunosorbent assay (FAST-ELISA) and the enzyme-linked immunoelectrotransfer blot (EITB) to determine the prevalence of human fascioliasis in the Bolivian Altiplano. *Am J Trop Med Hyg.* 1992;46:603-609.
- Hillyer GV. Serological diagnosis of *Fasciola hepatica*. Parasitol al Dia. 1993;17:130-136.
- Espino AM, Marcet R, Finlay CM. Detection of circulating excretory secretory antigens in human fascioliasis by sandwich enzyme-linked immunosorbent assay. *J Clin Microbiol.* 1990;28:2637-2640.
- 32. Carnevale S, Rodríguez MI, Santillán G, et al. Immunodiagnosis of human fascioliasis by an enzyme-linked immunosorbent assay (ELISA) and a micro-ELISA. *Clin Diagn Lab Immunol.* 2001;8:174-177.
- 33. Intapan PM, Maleewong W, Nateeworanart S, et al. Immunodiagnosis of human fascioliasis using an antigen of *Fasciola gigantica* adult worm with the molecular mass of 27 kDa by a dot-ELISA. *Southeast Asian J Trop Med Public Health*. 2003;34:713-717.
- 34. Wongkham C, Tantrawatpan C, Intapan PM, et al. Evaluation of immunoglobulin G subclass antibodies against recombinant *Fasciola gigantica* cathepsin L1 in an enzyme-linked immunosorbent assay for serodiagnosis of human fasciolosis. *Clin Diagn Lab Immunol.* 2005;12:1152-1156.
- Rokni MB, Samani A, Massoud J, et al. Evaluation of Dot-ELISA method using excretory-secretory antigens of *Fasciola hepatica* in laboratory diagnosis of human fasciolosis. *Iranian J Parasitol*. 2006;1:26-30.
- Espinoza JR, Maco V, Marcos L, et al. Evaluation of Fas2-ELISA for the serological detection of *Fasciola hepatica* infection in humans. *Am J Trop Med Hyg*, 2007;76:977-982.
- Tantrawatpan C, Maleewong W, Wongkham C, et al. Evaluation of immunoglobulin G4 subclass antibody in a peptide-based enzyme-linked immunosorbent assay for the serodiagnosis of human fascioliasis. *Parasitology*. 2007;134:2021-2026.
- DRG Diagnostics GmbH. DRG *Fasciola* IgG ELISA. Available at: www.drgdiagnostics.de/files/2010-03-fasciola.pdf. Accessed July 23, 2010.
- Gentaur. Fasciola hepatica IgG ELISA Kit. Available at: www.clonagen.com/ clonagen/6d7403a0-ee21-442d-884f-2ede890811e6/fasciola\_hepatica\_igg\_96\_ wells\_elisa\_product.aspx. Accessed July 23, 2010.

#### **CE Update**

- Ubeira FM, Muiño L, Adela Valero M, et al. MM3-ELISA detection of *Fasciola hepatica* coproantigens in preserved human stool samples. *Am J Trop Med Hyg.* 2009;81:156-162.
- Centers for Disease Control and Prevention. Parasites and Health: Intestinal Parasites: Comparative Morphology. Available at: www.dpd.cdc.gov/dpdx/html/ MorphologyTables.htm. Accessed July 23, 2010.
- 42. Harinasuta C, Kruatrachue M. The first recognized endemic area of bilharziasis in Thailand. *Ann Trop Med Parasitol.* 1962;56:314-322.
- Taylor RG, Moose JW. The egg from a human case of schistosomiasis in Laos. J Parasitol. 1971;57:78-80.
- Valero MA, Perez-Crespo I, Periago MV, et al. Fluke egg characteristics for the diagnosis of human and animal fascioliasis by *Fasciola hepatica* and *F. gigantica*. *Acta Trop.* 2009;111:150-159.
- Gulsen MT, Savas MC, Koruk M, et al. Fascioliasis: A report of five cases presenting with common bile duct obstruction. *Neth J Med.* 2006;64:17-19.
- Aksoy DY, Kerìmoğlu Ü, Oto A, et al. *Fasciola hepatica* infection: Clinical and computerized tomographic findings of ten patients. *Turk J Gastroenterol*. 2006;17:40-45.
- Koç Z, Ulusan Ş, Tokmak N. Hepatobiliary fascioliasis: Imaging characteristics with a new finding. *Diagn Interv Radiol.* 2009;15:247-251.
- Cantisani V, Cantisani C, Mortelé K, et al. Diagnostic imaging in the study of human hepatobiliary fascioliasis. *Radiol Med.* 2010;115:83-92.
- Aksoy DY, Kerimöglü U, Oto A, et al. Infection with Fasciola hepatica. Clin Microbiol Infect. 2005;11:859-861.

- Marcos LA, Tagle M, Terashima A, et al. Natural history, clinicoradiologic correlates, and response to triclabendazole in acute massive fascioliasis. *Am J Trop Med Hyg.* 2008;78:222-227.
- Fairweather I. Triclabendazole progress report, 2005-2009: An advancement of learning? J Helminthol. 2009;83:139-150.
- el-Morshedy H, Farghaly A, Sharaf S, et al. Triclabendazole in the treatment of human fascioliasis: A community-based study. *E Mediterr Health J.* 1999;5:888-894.
- 53. Centers for Disease Control and Prevention. Drugs for Parasitic Infections. Available at: www.dpd.cdc.gov/DPDx/html/PDF\_Files/MedLetter/ FlukeHermaphroditicInfection.pdf. Accessed July 24, 2010.
- 54. Favennec L, Jave Ortiz J, Gargala G, et al. Double-blind, randomized, placebocontrolled study of nitazoxanide in the treatment of fascioliasis in adults and children from northern Peru. *Aliment Pharmacol Ther.* 2003;17:265-270.
- Hien TT, Truong NT, Minh NH, et al. A randomized controlled pilot study of artesunate versus triclabendazole for human fascioliasis in central Vietnam. *Am J Trop Med Hyg*. 2008;78:388-392.

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