# Molecular phylogeny of European Runcinida (Gastropoda, Heterobranchia): the discover of an unexpected pool of complex species, with special reference to the case of Runcina coronata 

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Received 31 March 2020; revised 11 April 2021; accepted for publication 22 May 2021


#### Abstract

Runcinida is a small heterobranch order of sea slugs with 61 known species distributed worldwide across temperate and tropical latitudes with two established families (Ilbiidae and Runcinidae). Little is known about the phylogenetic relationships within Runcinida. Here, we present the first molecular phylogeny of the order with an emphasis on European species and we discuss the taxonomic status of the type species Runcina coronata. Molecular phylogenetics based on the mitochondrial genes cytochrome $c$ oxidase subunit I and 16S rRNA and nuclear gene histone H3 is used to investigate relationships between species. Detailed morpho-anatomical worked was additionally employed to study Runcina coronata. Our results suggest the monophyly of Runcinida and showed that Runcina coronata is a complex of four species, namely: $R$. coronata proper, $R$. aurata and two new species here formally described ( $\boldsymbol{R}$. caletensis sp. nov. and $R$. tingensis sp. nov.).


ADDITIONAL KEYWORDS: biodiversity - cryptic species - Ilbiidae - Runcinidae - sea slugs - species delimitation.

## INTRODUCTION

The order Runcinida Burn, 1963 includes two recognized families, namely Ilbiidae Burn, 1963 and Runcinidae H.Adams \& A.Adams, 1854 (Bouchet et al., 2017), with 61 valid species distributed in the Atlantic and
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Mediterranean Sea (46 species) and the Indian and Pacific Ocean ( 15 species)(MolluscaBase,2021).Members of this taxon are mostly characterized by the presence of an undivided dorsal shield (notum), an undivided foot, small external gills around, or to the right side of, the anus (sometimes absent), lack of parapodial lobes and four gizzard plates (Burn, 1963; Miller \& Rudman, 1968; Thompson, 1976; Burn \& Thompson, 1998).

The family Ilbiidae is composed of three genera: Fofinha Moro \& Ortea, 2015, Ilbia Burn, 1963 and

Pseudoilbia Miller \& Rudman, 1968. Members of this family are characterized by the absence of shell, gills (Ilbia and Pseudoilbia) and/or gizzard plates (Pseudoilbia and Fofinha) and presence of a pedal furrow (Ilbia and Pseudoilbia).
The family Runcinidae includes eight genera: Edmundsina Ortea, 2013,Ildica Bergh, 1889,Karukerina Ortea, 2013, Lapinura Marcus \& Marcus, 1970, Metaruncina Baba, 1967, Runcina Forbes in Forbes \& Hanley, 1853, Runcinella Odhner, 1924 and Runcinida Burn, 1963. Members of this family are characterized by having a radula with a bilobed rachidian tooth, presence of gills (with up to four pinnulae) and four gizzard plates. The shell can be present or absent.
The systematics of Runcinida is confusing due to the lack of synapomorphic traits uniting all members in each family. Earlier referred by Odhner (1939) as Peltacea and subsequently named Runcinacea by Burn, 1963, the group was considered a suborder - the most 'derived' - within Cephalaspidea only sharing the presence of an external seminal groove (Odhner, 1939; Burn, 1963; Schmekel, 1985). Colosi (1915) first challenged the position of runcinids within Cephalaspidea and later Odhner (Odhner, 1968) suggested that Runcinacea should be ranked as an order. Recently, Bouchet et al. (2017: 330, paragraph e) in their revised Nomenclator and typification of gastropod and monoplacophoran families suggested the ending-ida for orders and proposed the name Runcinida to replace Runcinacea.
Over theyears,many authors have continued to consider runcinids as part of Cephalaspidea (Marcus \& Marcus, 1970; Kress, 1977; Schmekel, 1985; Thompson \& Brodie, 1988; Vaught, 1989; Gosliner, 1990, 1991; Cervera et al., 1991; Mikkelsen, 1993, 1996; Millard, 1997; Schmekel \& Cappellato, 2001, 2002), while several phylogenetic studies on Heterobranchia based on morphological (Dayrat \& Tillier, 2002; Wägele \& Klussmann-Kolb, 2005) and molecular data (Dayrat et al., 2001; Grande et al., 2004a, b; Vonnemann et al., 2005) questioned the inclusion of Runcinida in the Cephalaspidea. Malaquias et al. (2009), based on molecular phylogenetics, produced the first sound evidence for the inclusion of runcinids in their own order as early proposed by Odhner (in Franc, 1968). This was later supported by the molecular phylogenetic studies by Jörger et al. (2010), Wägele et al. (2014) and Oskars et al. (2015).

Most known runcinids species are concentrated in European waters (Atlantic Ocean, including the Mediterranean Sea) and belong to the genera Pseudoilbia, Runcina and Runcinella. Among these genera, Runcina is the most species-rich (31 species). However, the definition of its type species, Runcina coronata (Quatrefages, 1844), is problematic and
remains a matter of debate. The species $R$. coronata is reported from England southwards to the Mediterranean coast of France (Quatrefages, 1844; Alder \& Hancock, 1846; Vayssière, 1883; Schmekel \& Cappellato, 2002; Cervera et al., 2004; Ballesteros et al., 2016). It is defined by having an almost elliptical body with two small lobes on the front of the head and a rounded posterior end of the body. The notum is brown in colour with lighter edges and with characteristic small, dull, white spots forming a semicircle on the head zone behind the eyes and at the posterior end of the notum. Three small and slightly pinnate gills are situated on the right side of the anus (Quatrefages, 1844; Alder \& Hancock, 1846; Forbes \& Hanley, 1851).
Runcina coronata (as Pelta coronata) was described from Brehat, Bretagne (Atlantic coastline of France). Two years later, Alder \& Hancock (1846) found specimens at Torbay (southern England) similar to those described by Quatrefages and also to Limapontia nigra Johnston, 1835, but differing from the former by the presence of gills. Alder \& Hancock (1846) believed that the lack of gills in $R$. coronata was an erroneous observation by Quatrefages but, nevertheless, they did not designate any specific name for these specimens. Based on additional specimens also collected at Torbay, Forbes (1853) described the species Runcina hancocki, which mainly differed from $R$. coronata by the presence of external gills. Vayssière (1883), in his 'Monographie du Pelta', considered Runcina a synonym of Pelta, but in 1951, the genus name Pelta Quatrefages, 1844 was suppressed and only the name Runcina Forbes (in Forbes \& Hanley, 1853) was considered valid (see Lemche, 1967: opinion n. 811).

Runcina coronata was first reported from the Mediterranean Sea (Marseille, France) by Vayssière (1883). Pruvot-Fol (1954) suggested that the Mediterranean species R. calaritana Colosi, 1915 was conspecific with $R$. coronata and Burn (1963), after comparing the original description of $R$. coronata and the specimens studied by Vayssière, pointed out differences in the shape of the body, coloration and number of crests in the gizzard plates, suggesting these specimens to belong to R. calaritana. The species Runcina aurata García et al., 1986 (type locality: Club La Hacienda, Bay of Algeciras, Strait of Gibraltar) was synonymized with R. coronata by Cervera et al. (1991) after a detailed comparison of specimens of Runcina aurata with the original description of $R$. coronata.

This work provides the first approach to the study of the diversity and systematics of the European species of runcinids based on a molecular phylogenetic framework. The taxonomic status of the type species Runcina coronata (Quatrefages, 1844) is investigated combining molecular phylogenetics and morphoanatomical characters.

## MATERIAL AND METHODS

## TAXON SAMPLING

Samples were obtained from fieldwork conducted by the authors and colleagues, and through the study of museum collections. Voucher specimens are held in the collections of the Zoologische Staatssammlung München, ZSM (Munich, Germany), Museum Victoria, MV (Melbourne, Australia), Museu de Zoologia da Universidade de São Paulo, MZUSP (São Paulo, Brazil), University Museum of Bergen, ZMBN (Bergen, Norway) and Museo Nacional de Ciencias Naturales (Madrid, Spain). Seventy-seven specimens of Runcinida were included for molecular analyses, representing five genera: Ilbia (one specimen), Lapinura (two specimens), Metaruncina (one specimen), Runcina ( 71 specimens) and Runcinida (one specimen). The remaining two specimens were referred as 'runcinid' sp.. All the species used in this study are listed in Table 1 and were identified by comparison with primary literature. The aplysiid Aplysia dactylomela Rang, 1828 and the acteonoid Micromelo undatus (Bruguière, 1792) were used as outgroup taxa. In total, 189 novel sequences were generated during the present work and 91 were obtained from GenBank (Table 1).

For the study of the Runcina coronata speciescomplex, we included specimens from Swanage in southern England (c. 210 km from the type locality of $R$. coronata, which is Brehat in France), Cádiz in Andalusia, south-western Spain and Tangier in northwestern Morocco.

## DNA EXTRACTION, AMPLIFICATION AND SEQUENCING

Tissue samples were taken from the foot and DNA was extracted using the DNeasy Blood and Tissue Kit (Qiagen, Valencia, CA). Partial sequences of the mitochondrial cytochrome $c$ oxidase subunit I (COI) and 16 S rRNA (16S) and nuclear histone H3 (H3) genes were amplified by polymerase chain reaction (PCR) using the universal primers: LCO1490 and HCO2198 (Folmer et al., 1994 for COI); 16S ar-L and 16br-H (Palumbi et al., 1991 for 16S); and H3aF and H3aR (Colgan et al., 1998 for H3). Polymerase chain reactions were conducted in a $25 \mu \mathrm{~L}$ reaction volume containing $1 \mu \mathrm{~L}$ of both forward and reverse primers ( $10 \mu \mathrm{~mol} / \mathrm{L}$ ), $2.5 \mu \mathrm{~L}$ of dNTP ( $2 \mathrm{mmol} / \mathrm{L}$ ), a gene-dependent amount of magnesium chloride ( $25 \mathrm{mmol} / \mathrm{L}$ ), $0.25 \mu \mathrm{~L}$ of Qiagen DNA polymerase ( 5 units $/ \mu \mathrm{L}$ ), $5 \mu \mathrm{~L}$ of ' Q -solution' ( $5 \times$ ), $2.5 \mu \mathrm{~L}$ of Qiagen buffer (10×) (Qiagen Taq PCR Core Kit) and $2 \mu \mathrm{~L}$ of genomic DNA. Amplification of $C O I$ was performed with an initial denaturation for 5 min at $94{ }^{\circ} \mathrm{C}$, followed by $35-36$ cycles of 1 min at $94{ }^{\circ} \mathrm{C}$, 30 s at $45^{\circ} \mathrm{C}$ (annealing temperature) and 1 min at $72{ }^{\circ} \mathrm{C}$, with a final extension of 10 min at $72^{\circ} \mathrm{C}$. The

16S amplification began with an initial denaturation for 5 min at $94^{\circ} \mathrm{C}$, followed by $35-36$ cycles of 1 min at $94{ }^{\circ} \mathrm{C}, 30 \mathrm{~s}$ at 42 and $49^{\circ} \mathrm{C}$ (annealing temperatures) and 1 min at $72^{\circ} \mathrm{C}$, with a final extension of 10 min at $72{ }^{\circ} \mathrm{C}$. Amplification of H 3 was performed with an initial denaturation for 5 min at $94{ }^{\circ} \mathrm{C}$, followed by 35 cycles of 1 min at $94^{\circ} \mathrm{C}, 30 \mathrm{~s}$ at $52^{\circ} \mathrm{C}$ (annealing temperature) and 1 min at $72^{\circ} \mathrm{C}$, with a final extension of 10 min at $72^{\circ} \mathrm{C}$. Successful PCR products were sent to Macrogen, Inc for purification and sequencing on a 3730XL DNA sequencer (Applied Biosystems).

## PHYLOGENETIC ANALYSES

Sequences were edited in GENEIOUS v.10.2.3 (Kearse et al., 2012) and aligned using MAFFT (Katoh et al., 2009) implemented in GENEIOUS v.10.2.3 with default settings. Sequences from the protein-coding genes $C O I$ and $H 3$ were translated into amino acids to check for stop-codons. Hypervariable regions of the 16 S alignment where homology could not be confidently established were removed using GBlocks under relaxed settings (Talavera \& Castresana, 2007). Nevertheless, analyses including and excluding these regions provided similar results. Therefore, final analyses were performed including all nucleotides. Sequences of the COI, 16S and $H 3$ genes were trimmed to 658,489 and 328 nucleotides, respectively. Single gene and concatenated ( $\mathrm{H} 3+\mathrm{COI}+16 \mathrm{~S}$ ) analyses were performed. Saturation for the first-, second- and third-codon positions of the COI and $H 3$ genes was calculated in MEGA X (Kumar et al., 2018).

The best-fit evolutionary model for each gene was determined in jModelTest v.2.1.6 (Guindon \& Gascuel, 2003; Darriba et al., 2012), under the Akaike information criterion (AIC) (Akaike, 1974). The GTR+G+I model was selected for the COI and 16 S genes, and $\mathrm{K} 80+\mathrm{G}$ for the $H 3$ gene. Bayesian inference (BI) analyses were performed in MrBayes v.3.2.1 (Ronquist \& Huelsenbeck, 2003) with random starting trees and two parallel runs of $10^{7}$ generations. The models implemented were those estimated with jModelTest v.2.1.6. The combined dataset was partitioned among genes and the 'unlink' command was used to allow all parameters to vary independently within each partition. Convergence was checked in TRACER v.1.7.1 (Rambaut et al., 2018) with a burn-in of $25 \%$. Nodes with a posterior probability (PP) $\geq 0.95$ (Alfaro et al., 2003) were considered well supported and discussed. Maximum likelihood (ML) analysis was executed using RAxML v. 8 (Stamatakis, 2014) and node support was assessed with nonparametric bootstrapping (BS) with 5000 replicates. Nodes with bootstrap values (BS) $\geq 70$ (Hillis \& Bull, 1993) were considered significant and were discussed. Both BI
Table 1. List of specimens used for phylogenetic analysis. (*) New sequences generated for this study. Museum abbreviations: Museum Victoria collections NMVF), University Museum of Bergen (ZMBN), Zoologische Staatssammlung München, Germany (ZSM), Museo Nacional de Ciencias Naturales (MNCN), Museu de Zoologia da Universidade de São Paulo (MZUSP) and The Natural History Museum, London, United Kingdom (NHMUK)

| Species Preliminary Ids | Revised Ids | Locality | Voucher no. | 16S | COI | H3 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ilbia ilbi Burn, 1963 |  | Hamers Haven, Australia | NMVF234189 | MK323019 | - | MK322992 |
| Lapinura divae (Marcus \& Marcus, 1963) |  | Bermuda (GB) | ZMBN 82997 | KJ022825 | KF992195 | KJ022893 |
| Lapinura divae |  | Bermuda | ZMBN 82997 | MW144769* | - | MW182586* |
| Metaruncina nhatrangiensis Chernyshev, 2005 |  | Vietman (GB) | - | KP313664 | KP313668 | - |
| Metaruncina setoensis (Baba, 1954) |  | Okinawa, Japan | ZMBN 88231 | MW144770* | MW172338* | - |
| Pseudoilbia avellana (Schmekel \& Cappellato, 2001) | R. avellana | Palamos, Spain | MNCN 15.05/90415 | - | MW172346* | MW182594* |
| Runcina adriatica <br> T. Thompson, 1980 |  | Azores Islands, Portugal | MNCN 15.05/91518 | MW144774* | MW172339* | MW182587* |
| Runcina adriatica |  | Azores Islands, Portugal | ZMBN81756 | - | MW172341* | MW182589* |
| Runcina adriatica |  | Azores Islands, Portugal | ZMBN81756 | MW144773* | MW172344* | MW182591* |
| Runcina adriatica |  | Iz Island, Croatia | MNCN 15.05/88099 | MK323015 | MK322986 | MK323036 |
| Runcina adriatica |  | Ugljan Island, Croatia | MNCN 15.05/88100 | MK322996 | MK322967 | MK323022 |
| Runcina adriatica |  | Zut Island, Croatia | MNCN 15.05/88101 | MK322997 | MK322968 | - |
| Runcina adriatica |  | Otranto, Italy | MNCN 15.05/88102 | MK322993 | MK322966 |  |
| Runcina adriatica |  | Palamós, Spain | MNCN 15.05/90416 | - | MW172340* | MW182588* |
| Runcina adriatica |  | Cadaqués, Spain | MNCN 15.05/88103 | MK322994 | MK322987 | MK323020 |
| Runcina adriatica |  | Cadaqués, Spain | MNCN 15.05/90417 | MW144771* | MW172342* | - |
| Runcina adriatica |  | Cadaqués, Spain | MNCN 15.05/90418 | MW144772* | MW172343* | MW182590* |
| Runcina adriatica |  | Otranto, Italy | MNCN 15.05/90666 | MW144775* | MW172345* | MW182592* |
| Runcina cf. adriatica |  | Lecce, Italy | Pending number | MW144778* | MW172348* | MW182595* |
| Runcina cf. adriatica |  | Lecce, Italy | MNCN 15.05/91515 | MW144802* | MW172377* | MW182621* |
| Runcina cf. adriatica |  | La Strea, Italy | MNCN 15.05/91516 | MW144817* | MW172391* | MW182634* |
| Runcina cf. adriatica |  | Ugljan Island, Croatia | MNCN 15.05/91501 | MW144810* | MW172384* | MW182627* |
| Runcina cf. adriatica |  | Ugljan Island, Croatia | MNCN 15.05/91502 | MW144811* | MW172385* | MW182628* |
| Runcina cf. adriatica |  | Ugljan Island, Croatia | MNCN 15.05/91503 | MW144812* | MW172386* | MW182629* |
| Runcina africana Pruvot-Fol, 1953 |  | Tenerife, Canary Islands, Spain | NHMUK 20030791/1 | KJ022780 | DQ974680 | KJ022942 |
| Runcina africana |  | Port Lligant, Spain | MNCN 15.05/90651 | MW144776* | - | MW182593* |

Table 1. Continued

| Species Preliminary Ids | Revised Ids | Locality | Voucher no. | 16 S | COI | H3 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Runcina cf. bahiensis Cervera et al., 1991 | R. hornae | Cadaqués, Spain | MNCN 15.05/88104 | - | MK322988 | MK323005 |
| Runcina brenkoae <br> T. Thompson, 1980 |  | Split, Croatia | MNCN 15.05/88086 | MK322998 | MK322969 | MK323023 |
| Runcina brenkoae |  | Nin, Croatia | MNCN 15.05/88089 | MK323017 | MK322972 | MK323038 |
| Runcina brenkoae |  | Nin, Croatia | MNCN 15.05/88088 | MK323018 | MK322971 | MK323039 |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91509 | - | MW172399* | - |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91510 | - | MW172400* | MW182641* |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91511 |  | MW172401* | - |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91512 | - | MW172402* | - |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91513 | MW144787* | - | MW182644* |
| Runcina ferruginea |  | Iz Island, Croatia | MNCN 15.05/90667 | MW144788* | MW172361* | MW182607* |
| Runcina ferruginea |  | Ugljan Island, Croatia | MNCN 15.05/90668 | MW144786* | MW172359* | MW182605* |
| Runcina ferruginea |  | Split, Croatia | MNCN 15.05/90669 | - | MW172360* | MW182606* |
| Runcina ferruginea |  | Banyuls-sur-Mer, France | ZSMMOL 20142074 | - | MW172374* | MW182618* |
| Runcina hansbechi Schmekel \& Cappellato, 2001 | R. brenkoae | Banyuls-sur-Mer, France | ZSMMOL20142071 | MW144789* | MW172362* | MW182608* |
| Runcina cf. hansbechi | R. avellana | Mataró, Spain | MNCN 15.05/88108 | MK323028 | MK322973 | MK32300 |
| Runcina hornae Schmekel \& Cappellato, 2002 |  | Palamós, Spain | MNCN 15.05/88110 | MK323033 | MK322978 | - |
| Runcina cf. hornae |  | Iz Island, Croatia | MNCN 15.05/91504 | MW144802* | - | MW182622* |
| Runcina lusitanica Araujo et al., 2019 |  | Faro, Portugal | MNCN 15.05/88091 | MK323024 | MK322979 | MK322999 |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/200065 | - | MK322981 | MK323002 |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/88093 | - | MK322965 | MK323004 |
| Runcina brenkoae |  | Roses, Spain | MNCN 15.05/88087 | MK323001 | MK322970 | MK323026 |
| Runcina brenkoae |  | Roses, Spain | MNCN 15.05/88090 | MK323003 | MK322964 | - |
| Runcina brenkoae |  | Roses, Spain | MNCN 15.05/90422 | - | MW172349* | MW182596* |
| Runcina brenkoae |  | Banyuls-sur-Mer, France | ZSMMOL20142072 | MW144777* | MW172347* | - |
| Runcina coronata (Quatrefages, 1844) |  | Swanage, England | MNCN 15.05/88105 <br> MNCN/ADN 118951 | MK323031 | MK322976 | MK323010 |
| Runcina coronata |  | Swanage, England | MNCN 15.05/90423 | - | MW172356* | MW182602* |
| Runcina coronata | R. aurata | Cádiz, Spain | MNCN 15.05/88106 | MN057638 | - | MN057641 |
| Runcina coronata | R. aurata | Cádiz, Spain | MNCN 15.05/88107 | MN057639 | MN057640 | MN057642 |

Table 1. Continued

| Species Preliminary Ids | Revised Ids | Locality | Voucher no. | 16S | COI | H3 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Runcina coronata | R. aurata | Cádiz, Spain | MNCN 15.05/91500 | MW144785* | MW172358* | MW182604* |
| Runcina coronata | R. aurata | Cádiz, Spain | MNCN/ADN 118948 | MW144780* | MW172351* | MW182597* |
| Runcina coronata | R. aurata | Cádiz, Spain | MNCN/ADN 118950 | - | MW172353* | MW182599* |
| Runcina coronata | R. caletensis sp. nov | Cádiz, Spain | MNCN/ADN 118949 | MW144781* | MW172352* | MW182598* |
| Runcina coronata | $R$. caletensis sp. nov | Cádiz, Spain | MNCN 15.05/200113 | MW144783* | MW172355* | MW182601* |
| Runcina coronata | R. tingensis sp. nov | Tangier, Morocco | MNCN 15.05/91514 | MW144782* | MW172354* | MW182600* |
| Runcina coronata | $R$. tingensis sp. nov | Tangier, Morocco | MNCN 15.05/200114 | MW144784* | MW172357* | MW182603* |
| Runcina ferruginea Kress, 1977 |  | Newlyn, Cornwall, England | MNCN 15.05/88109 | MK323032 | MK322977 | MK323011 |
| Runcina ferruginea |  | Roses, Spain | MNCN 15.05/91517 | MW144779* | MW172350* | - |
| Runcina ferruginea |  | Lecce, Italy | MNCN 15.05/91508 | - | MW172397* | MW182639* |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/88092 | MK323035 | MK322980 | MK323014 |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/88094 | MK323027 | MK322989 | - |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/90424 | MW144790* | MW172363* | MW182609* |
| Runcina lusitanica |  | Faro, Portugal | MNCN 15.05/90425 | MW144791* | MW172364* | MW182610* |
| Runcina marcosi Araujo et al., 2019 |  | Roses, Spain | MNCN 15.05/88098 MNCN/ADN 118952 | MK323025 | MK322982 | MK323000 |
| Runcina marcosi |  | Mataro, Spain | MNCN 15.05/88095 | MK323037 | MK322983 | MK323016 |
| Runcina marcosi |  | Roses, Spain | MNCN 15.05/88097 | - | MK322990 | MK323012 |
| Runcina marcosi |  | Roses, Spain | MNCN 15.05/88096 | MK323034 | MK322984 | MK323013 |
| Runcina marcosi |  | Cadiz, Spain | MNCN 15.05/200066 | - | MK322985 | MK323006 |
| Runcina marcosi |  | Banyuls-sur-Mer, France | ZSM MOL 201442089 | MK323021 | MK322991 | MK322995 |
| Runcina marcosi |  | Roses, Spain | MNCN 15.05/90426 | - | MW172393* | - |
| Runcina marcosi |  | Roses, Spain | MNCN 15.05/90427 | MW144819* | MW172394* | MW182636* |
| Runcina ornata (Quatrefages, 1844) |  | Azores Islands, Portugal | ZMBN87929 | MW144796* | MW172368* | - |
| Runcina ornata |  | Azores Islands, Portugal | ZMBN87949 | MW144795* | MW172367* | - |
| Runcina sp. 3 | R. hornae | Mataro, Spain | MNCN 15.05/90654 | MW144818* | MW172392* | MW182635* |
| Runcina sp. 4 | R. hornae | Palamos, Spain | MNCN/ADN 118954 | - | MW172395* | MW182637* |
| Runcina sp. 8 | R. hornae | Roses, Spain | MNCN 15.05/90655 | MW144801* | MW172376* | MW182620* |
| Runcina sp. 10 | R. hornae | Mataro, Spain | MNCN 15.05/90656 | MW144822* | MW172404* | MW182643* |
| Runcina sp. 11 | R. hornae | Mataro, Spain | MNCN 15.05/90657 | MW144821* | MW172398* | MW182640* |

Table 1. Continued

| Species Preliminary Ids | Revised Ids | Locality | Voucher no. | 16S | COI | H3 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Runcina sp. 13 | R. hornae | Mataro, Spain | MNCN 15.05/90658 | - | MW172403* | MW182642* |
| Runcina sp. 15 | R. hornae | Cadaqués, Spain | MNCN 15.05/90659 | - | MW172369* | MW182614* |
| Runcina sp. 17 | R. adriatica | Cadaqués, Spain | MNCN 15.05/90419 | MW144798* | MW172371* | MW182616* |
| Runcina sp. 18 | R. hornae | Cadaqués, Spain | MNCN 15.05/90660 | MW144797* | MW172370* | MW182615* |
| Runcina sp. 19 | R. avellana | Roses, Spain | MNCN/ADN 118953 | MW144805* | MW172379* | - |
| Runcina sp. 19 | R. avellana | Roses, Spain | MNCN 15.05/90652 | MW144806* | MW172380* | MW182624* |
| Runcina sp. 20 | R. hornae | Roses, Spain | MNCN 15.05/90661 | MW144813* | MW172387* | MW182630* |
| Runcina sp. 20 | R. hornae | Roses, Spain | MNCN 15.05/90662 | MW144814* | MW172388* | MW182631* |
| Runcina sp. 20 | R. hornae | Roses, Spain | MNCN 15.05/90663 | MW144815* | MW172389* | MW182632* |
| Runcina sp. 21 |  | Zut Island, Croatia | MNCN 15.05/91505 | MW144816* | MW172390* | MW182633* |
| Runcina sp. 22 |  | Zut Island, Croatia | MNCN 15.05/91506 | MW144807* | MW172381* | - |
| Runcina sp. 22 |  | Zut Island, Croatia | MNCN 15.05/91507 | MW144809* | MW172383* | MW182626* |
| Runcina sp. 24 | R. hornae | Roses, Spain | MNCN 15.05/90664 | MW144820* | MW172396* | MW182638* |
| Runcina sp. 26 | R. avellana | Roses, Spain | MNCN 15.05/90653 | MW144808* | MW172382* | MW182625* |
| Runcina sp. 31 | R. adriatica | Es Caials (SP) | MNCN 15.05/90420 | MW144792* | MW172365* | MW182611* |
| Runcina sp. 33 | R. hornae | Port Lligant (SP) | MNCN 15.05/90665 | MW144793* |  | MW182612* |
| Runcina sp. 34 | R. adriatica | Port Lligant (SP) | MNCN 15.05/90421 | MW144794* | MW172366* | MW182613* |
| Runcinida marisae Chernyshev, 1998 |  | Vostok Bay, Russia | - | - | KP313667 | - |
| Runcinida valentinae Chernyshev, 2006 |  | Iturup Island, Russia | - | - | KP313666 | - |
| Runcinida sp. 1 |  | Cap Ferret, France | MNCN 15.05/90670 | MW144800* | MW172375* | MW182619* |
| Runcinid sp. 1 |  | Alagoas, Brazil | MZUSP103407 | MW144799* | MW172373* | - |
| Lapinura sp. 1 |  | Maui, Hawaii | MNCN 15.05/90671 | MW144804* | MW172378* | MW182623* |
| Runcinid sp. 2 |  | Quirimbas, Mozambique | ZMBN105119 | - | MW172372* | MW182617* |
| Aplysia dactylomela Rang, $1828$ |  | Cape Verde (GB) | NHMUK <br> 20030795/20030796 | KJ022921 | KF992168 | KJ022798 |
| Micromelo undatus <br> (Bruguière, 1792) |  | Tenerife, Canary Island (GB) | NHMUK 20030800 | KJ022944 | DQ974653 | KJ022778 |

and ML trees were visualized in FigTree v.1.4.3 (http://tree.bio.ed.ac.uk/software/figtree/). Minimum and maximum pairwise uncorrected $p$-distances of COI were calculated in MEGA X (Kumar et al., 2018).

## SPECIES DELIMITATION

For species delimitation analyses, the Automatic Barcode Gap Discovery (ABGD) (Puillandre et al., 2012) and Bayesian Poisson Tree Processes (bPTP) (Zhang et al., 2013) methods were used. For the ABGD analyses we used the fast-evolving COI gene using the ingroup sequences obtained in this study and those available in GenBank. We used the default setting ( $P_{\text {min }}=0.001, P_{\text {max }}=0.1$, Steps $=10$, $X=1.2, \mathrm{Nb}$ bins $=20$ ) under the three models of evolution, namely Jukes-Cantor (JC69), Kimura (K80) and Simple Distance. bPTP analyses were run with default parameters using the COI trees at the webserver (https://species.h-its.org/ptp/) (Zhang et al., 2013).

## MORPHOLOGY

The external morphology of the specimens used in our molecular phylogenies were, in most cases, studied based on photographs of live specimens ( 175 images) and from living animals collected during sampling activities. In the latter case, examination of the shape and coloration was carried out under a stereomicroscope. In order to identify all specimens, we compared these observations with original descriptions of species and specialized literature.
For the internal morphology, the animals were dissected either dorsally or ventrally, and the buccal bulbs and gizzards were extracted and dissolved in a solution of $10 \%$ sodium hydroxide to expose and clean them. The radulae and gizzard plates were then immersed in water, dried and mounted for scanning electron microscopy (SEM) with a Nova NanoSEM 450 available at the University of Cadiz (Cadiz, Spain). The reproductive systems were dissected out of the animals and examined and drawn using a stereomicroscope equipped with a camera lucida. All specimens previously identified as $R$. coronata used for morphological comparison are listed in the 'Examined material' in the Systematic description.

## RESULTS

## PhYLOGENETIC ANALYSES

The combined dataset yielded a sequence alignment of 1475 positions. We obtained 189 new sequences, 64 for $H 3,68$ for COI and 57 for 16S (Table 1). No saturation was observed in the $C O I$ and $H 3$ genes,
not even in the third-codon positions (not shown). The combined tree $(H 3+C O I+16 \mathrm{~S})$ provided better resolution than $H 3, C O I$ or 16 S separately (see Supporting Information, Figs S1-S3). The results of ML and BI trees rendered similar topologies, but bootstrap values were lower than posterior probabilities in some clades (Fig. 1).

Both ML and BI supported the monophyly of the order Runcinida ( $\mathrm{PP}=1 ; \mathrm{BS}=99$ ), with the species Ilbia ilbi Burn, 1963 (family Ilbiidae) sister to Clade A (family Runcinidae). Clade A ( $\mathrm{PP}=0.96 ; \mathrm{BS}=-$ ) is divided into two main sister subclades, here called Clade B (PP = 0.98, BS = - [no value was recovered]) and Clade C ( $\mathrm{PP}=1, \mathrm{BS}=78$ ).

Clade B was subdivided into three well-supported clades: the first, Clade D, including only the specimen labelled as runcinid sp. 2 from Mozambique; the second, Clade E ( $\mathrm{PP}=1$; $\mathrm{BS}=96$ ), containing Metaruncina setoensis (Baba, 1967) and Metaruncina nhatrangensis Chernyshev, 2005; and the third, Clade F ( $\mathrm{PP}=1 ; \mathrm{BS}=99$ ), with most of Runcina specimens and Pseudoilbia avellana (Schmekel \& Cappellato, 2001). Within Clade F, Runcina ornata appears as sister to the remaining Runcina specimens. Within the latter, three clades with maximum support correspond to well-established species, such as R. lusitanica Araujo et al., 2019, R. marcosi Araujo et al., 2019 and the type species, $R$. coronata. However, specimens attributed to Runcina adriatica Thompson, 1980, Runcina ferruginea Kress, 1977, Runcina hansbechi Schmekel \& Cappellato, 2001 and Runcina africana Pruvot-Fol, 1953 branched off in different clades. In addition, specimens early identified as Runcina cf. bahensis Cervera et al., 1991 and Runcina hornae Schmekel \& Cappellato, 2002 clustered together with several unidentified species from Spain $(\mathrm{PP}=1 ; \mathrm{BS}=100)$.

Clade C ( $\mathrm{PP}=1 ; \mathrm{BS}=78$ ) was subdivided into two clades: Clade G, containing specimens identified as Runcina ferruginea ( $\mathrm{PP}=1$; $\mathrm{BS}=100$ ); and Clade H ( $\mathrm{PP}=1 ; \mathrm{BS}=80$ ) with two subgroups, one including Lapinura divae (Marcus \& Marcus, 1970), Lapinura sp. 1, specimens identified as $R$. ferruginea and runcinid sp. $1(\mathrm{PP}=1 ; \mathrm{BS}=82)$, and the other with Runcinida marisae Chernyshev, 1998, Runcinida valentinae Chernyshev, 2006 and Runcinida sp. 1 ( $\mathrm{PP}=1 ; \mathrm{BS}=90$ ).

## Species delimitation

The ABGD analyses identified 31 groups with all three models of evolution implemented (Fig. 1A). Within these groups and for all models, the specimens identified as $R$. coronata (type species of the genus Runcina) split in four distinct groups: RC1, RC2, RC3


Figure 1. Phylogenetic hypothesis of the order Runcinida based on the combined genes $H 3, C O I$, and 16 S inferred by Bayesian analysis. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood analysis. A, ABGD results based on the COI dataset. B, bPTP results based on the COI dataset. Rectangles in Ilbia ilbi are missing since there is no COI sequence available. Abbreviations: ATL, Atlantic Ocean; MED, Mediterranean Sea. *, branches with maximum support. 1, refers to sequences from Genbank.
and RC4 (Fig. 2). The bPTP analysis showed the same result (Figs 1B, 2).

The minimum pairwise uncorrected $p$-distances for COI among groups of Runcinida recovered by ABGD are presented in Table 2, and ranged from $5.6 \%$ to $23.1 \%$. Within the genus Runcina, distances ranged from $6 \%$ to $17.7 \%$ between species (Table 2, groups VIXXIII; see Fig. 1). Regarding the specimens originally identified as $R$. coronata, the distances between the four groups (Fig. 2) were considerably higher compared with the distance between specimens within each group (Table 3). Groups RC1 (Runcina aurata) and RC4 (Runcina caletensis sp. nov.) contain specimens from the same locality (Cádiz, Spain) and the minimum distance between them was $7.8 \%$. Comparing both groups with group RC2 (Runcina coronata), which contains specimens from Swanage, England, the minimum distances were $6.3 \%$ for RC 1 and $8.8 \%$ for RC4. Finally, the minimum genetic distances when these three groups are compared with group RC3 (Runcina tingensis sp. nov.; specimens from Morocco) were $6.4 \%$ ( RC 1 ), $7.0 \%$ (RC2) and $9.9 \% ~(\mathrm{RC} 4)$ (Table 3).

Our morphological studies (see theme Systematic description below) and molecular results support four different species among the specimens originally identified as Runcina coronata. We present a redescription of $R$. coronata and $R$. aurata, and the description of the two new species.

## Systematic DESCRIPTION

Family Runcinidae H.Adams \& A.Adams, 1854

> (Figs 3A, B, 4A-C, 5A, 6A)

Runcina Forbes in Forbes \& Hanley, 1851.
Runcina coronata (Quatrefages, 1844).

## Synonymy

Pelta coronata Quatrefages, 1844: 151, pl.3, fig. IV.
Runcina hancocki Forbes (in Forbes \& Hanley), 1851: 611, pl. C.C.C, fig. 2.
R. calaritana Colosi, 1915: 1, figs 1-18.

Type locality: Bréhat, Bretagne, France ( $48^{\circ} 50^{\prime} 59^{\prime \prime} N$; $2^{\circ} 59^{\prime} 47{ }^{\prime \prime} \mathrm{W}$ ).

Examined material: MNCN 15.05/88105, Swanage ( $50^{\circ} 36^{\prime} 28^{\prime \prime} \mathrm{N} ; 1^{\circ} 56^{\prime} 45^{\prime}$ W), southern England, coll. Ian F. Smith, Apr 2016, 3 mm living animal, found by brushing the bases of stones with encrusting pink coralline algae (dissected and sequenced). MNCN $15.05 / 90423$, Swanage ( $50^{\circ} 36^{\prime} 28^{\prime \prime} \mathrm{N} ; 1^{\circ} 56^{\prime} 45^{\prime} \mathrm{W}$ ), southern England, coll. Ian F. Smith, Apr 2016, 4 mm living animal, found by brushing the bases of stones


Figure 2. Runcina coronata species-complex. Detail of Clade F extracted from the analyses illustrated in Figure 1. Continuous rectangles, ABGD analysis based on the COI dataset. Dotted rectangles, bPTP analysis based on the COI dataset. *, branches with maximum support. 1, refers to sequences from Genbank.
Table 2. Uncorrected $p$-distances based on the $C O I$ gene within the order Runcinida. Roman numbers are based on species delimitation groups represented in Figure 1

|  | III | IV | V | VI | VII | VIII | IX | X | XI | XII | XIII | XIV | XV | XVI |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| IV | 15.5 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| V | 18 | 15.8 |  |  |  |  |  |  |  |  |  |  |  |  |
| VI | 12.2-12.7 | 16.7-18.3 | 16.6-16.9 |  |  |  |  |  |  |  |  |  |  |  |
| VII | 14.9-15.4 | 17.4-18.3 | 17.6-18.5 | 8.2-9 |  |  |  |  |  |  |  |  |  |  |
| VIII | 14.5-15.2 | 18.1-19.1 | 16.8-17.6 | 9.8-10.9 | 9.9-12.2 |  |  |  |  |  |  |  |  |  |
| IX | 12.7-13.4 | 16.7-17.1 | 15.8-16.1 | 7.6-8.8 | 7.7-10.7 | 5.6-7 |  |  |  |  |  |  |  |  |
| X | 15.7-16.9 | 17.5-18.4 | 18-19.5 | 9.1-10.4 | 9.9-12.4 | 7.8-9.8 | 6.1-7.8 |  |  |  |  |  |  |  |
| XI | 15.5-15.9 | 17.9-18.7 | 16.7-17.4 | 9.8-11 | 9.8-13 | 11.1-12.7 | 9-10.5 | 11.1-13 |  |  |  |  |  |  |
| XII | 15.9-16.4 | 17.1-17.5 | 16.7-17.5 | 10.6-11.4 | 10.2-12.2 | 11.2-12.6 | $9.4-10.2$ | 10.2-12.5 | 10.3-11.3 |  |  |  |  |  |
| XIII | 14-15 | 16.2-17.1 | 17.8-18.5 | 7.2-9.3 | 7.2-11.1 | 9.6-11.7 | 9-10.5 | 8.8-11.5 | 9.6-11.3 | 8.4-10.4 |  |  |  |  |
| XIV | 15.4-15.5 | 18.3-18.7 | 16.9-17 | 10.5-10.9 | 8.9-10.6 | 11.3-13.1 | $9.4-10.5$ | 11.8-13.2 | 11.5-12.2 | 11.1-12 | 9.4-12 |  |  |  |
| XV | 15.7 | 18.9-19.7 | 16.7-17.4 | 10.5-11.6 | $9.3-11.1$ | 11.2-12.3 | $9.1-10.2$ | 9.9-11 | 10.9-11.8 | 10.9-12.4 | 10.4-12.6 | 6.3-7.3 |  |  |
| XVI | 16 | 19-19.8 | 18.4 | 10.8-12.2 | $9.2-11.4$ | 12.7-13.4 | 10.5-11.6 | 11.8-12.9 | 11-12.3 | 12.7-13.7 | 10.8-12.6 | 6.4-7.8 | 7-7.7 |  |
| XVII | 17 | 19.7 | 17.8-17.9 | 11.7-12.3 | 10-12.3 | 13.1-13.7 | 9.5-10 | 12.3-13.4 | 13.4-14.5 | 12.8-13.7 | 12.6-13.7 | 7.8-8 | 8.8-9.5 | 9.9-10.2 |
| XVIII | 15.6-17.6 | 17.8-19.4 | 17.2-19.7 | 9-11.3 | 10.2-13.1 | 10.5-13.6 | 9.5-11.8 | 11.6-13.9 | 11.4-13.8 | 10.9-14.4 | 9.4-11.8 | 11.6-13.5 | 12.3-14.6 | 12-13.7 |
| XIX | 15.1-15.7 | 18.2-19 | 16.7-17.8 | 10.4-11.2 | 10.1-12.3 | 10.2-11.7 | 8.4-9.9 | 9.9-12.4 | 10.7-12.4 | 11.1-13.2 | $9.6-12.3$ | 12.1-12.9 | 12.2-13.7 | 12.5-13.5 |
| XX | 16.9-18.7 | 17.7-19.2 | 19.4-20.3 | 11.5-13 | 11.6-13.6 | 13.3-15.3 | 11.3-13.1 | 11.3-14.4 | 12-14.9 | 11.5-13.7 | 11.4-13.9 | 12.8-14 | 12.8-14.5 | 12.3-13.8 |
| XXI | 17.9 | 19.4 | 18.4 | 12.8-13.6 | 14.4-14.8 | 14.4-14.8 | 13.6-13.9 | 14.6-15.3 | 14.1-14.9 | 12.9-13.3 | 11.8-13.7 | 13.9-14.1 | 14.1 | 13.4-13.6 |
| XXII | 15.4 | 16.8 | 16.3 | 9.6-10.4 | 10.1-12.1 | 10.7-11.1 | 9.6-10.4 | 12.2-13.9 | 11.6-12.5 | 12.4-13 | 9.8-11.1 | 12.6-13.3 | 12.3-13.1 | 12.7-13 |
| XXIII | 17.5-17.6 | 18.3-18.7 | 18.6-18.7 | 13.5-14.3 | 12.9-14.5 | 15.2-15.9 | 13.5-14.1 | 13.8-14.9 | 14.6-15.6 | 14.7-15.6 | 12.3-13.4 | 13.5-14.1 | 15.3-16.1 | 15.1-15.6 |
| XXIV | 18.3-18.5 | 18.8-19 | 20.8-21 | 14.4-15.2 | 16.7-18.2 | 18.7-19.3 | 16.6-17 | 18.2-19.5 | 16.6-18.1 | 15.5-16.3 | 16.4-17.2 | 16.1-16.6 | 17.2-18.1 | 17-18 |
| XXV | 16.9 | 20.5 | 19.5 | 16.7-17.3 | 16.7-18.8 | 18.3-18.8 | 15-15.9 | 18.1-18.8 | 15.6-16.7 | 17-17.5 | 16.2-17.7 | 16-16.6 | 17.2-17.8 | 16.9-17.8 |
| XXVI | 17.5 | 19.9 | 19.5 | 16.1-16.8 | 15.7-17.1 | 16.9-17.7 | 15.3-16.3 | 17.2-18.6 | 15.7-16.9 | 15.7-16.8 | 15-15.5 | 17.3-17.6 | 17.7-18.7 | 17.9-18 |
| XXVII | 16.9 | 18.3 | 20.1 | 16.6-16.9 | 17.9-18.8 | 18.5-19 | 16.7-17.9 | 17.4-18.6 | 17.5-18.1 | 17.2-17.6 | 16.7-17.9 | 17.6-17.8 | 18.5-19.2 | 18-18.2 |
| XXVIII | 21 | 21.4 | 20.5 | 17.8-19.1 | 20.1-21.1 | 20.7-21.5 | 19.6-19.9 | 20.1-21.1 | 20.1-21 | 21.3-21.5 | 19.6-20.5 | 19.2-19.3 | 20.6-21.3 | 20.8-21 |
| XXIX | 20.7 | 20.7 | 18.1 | 16.9-17.3 | 17.1-18.9 | 18.4-18.7 | 16.7-17 | 18.3-19.5 | 18.2-19.6 | 18.2-18.7 | 17.4-18.8 | 16.9-17.1 | 17.3-17.9 | 17.6-18.2 |
| XXX | 20.2 | 18.3 | 17.1 | 17.7-18.9 | 18.5-19 | 16.7-17.5 | 16.1-16.6 | 17.5-18.9 | 18-19 | 18.8-19.8 | 18-19.1 | 17.2-17.5 | 17.1-17.8 | 18.5-18.7 |
| XXXI | 19.3 | 20.5 | 19.3 | 17.4-17.7 | 17.5-19.8 | 18.5-19 | 16.7-17.4 | 18.5-20 | 17.9-19.6 | 18.2-18.6 | 17.8-19.2 | 18.2-18.5 | 18.4-19 | 19-19.7 |
|  | XVII | XVIII | XIX | XX | XXI | XXII | XXIII | XXIV | XXV | XXVI | XXVII | XXVIII | XXIX | XXX |
| III | 17 | 15.6-17.6 | 15-15.7 | 16.9-18.7 | 17.9 | 15.4 | 17.5-17.6 | 18.3-18.5 | 16.9 | 17.5 | 16.9 | 21 | 20.7 | 19.3 |
| IV | 19.7 | 17.8-19.4 | 18.2-19 | 17.7-19.2 | 19.4 | 16.8 | 18.3-18.7 | 18.8-19 | 20.5 | 19.9 | 18.3 | 21.4 | 20.7 | 18.3 |
| V | 17.8-17.9 | 17.2-19.7 | 16.7-17.8 | 19.4-20.3 | 18.4 | 16.3 | 18.6-18.7 | 20.8-21 | 19.5 | 19.5 | 20.1 | 20.5 | 18.1 | 17.1 |
| VI | 11.7-12.3 | 9-11.3 | 10.4-11.2 | 11.5-13 | 12.8-13.6 | 9.6-10.4 | 13.5-14.3 | 14.4-15.2 | 16.7-17.3 | 16.1-16.8 | 16.6-16.9 | 17.8-19.1 | 16.9-17.3 | 17.7-18.9 |
| VII | 10-12.3 | 10.2-13.1 | 10.1-12.3 | 11.6-13.6 | 14.4-14.8 | 10.1-12.1 | 12.9-14.5 | 16.7-18.2 | 16.7-18.8 | 15.7-17.1 | 17.9-18.8 | 20.1-21.1 | 17.1-18.9 | 18.5-19 |
| VIII | 13.1-13.7 | 10.5-13.6 | 10.2-11.7 | 13.3-15.3 | 14.4-14.8 | 10.7-11.1 | 15.2-15.9 | 18.7-19.3 | 18.3-18.8 | 16.9-17.7 | 18.5-19 | 20.7-21.5 | 18.4-18.7 | 16.1-16.6 |
| IX | 9.5-10 | 9.5-11.8 | 8.4-9.9 | 11.3-13.1 | 13.6-13.9 | $9.6-10.4$ | 13.5-14.1 | 16.6-17 | 15-15.9 | 15.3-16.3 | 16.7-17.9 | 19.6-19.9 | 16.7-17 | 16.7-17.5 |
| X | 12.3-13.4 | 11.6-13.9 | 9.9-12.4 | 11.3-14.4 | 14.6-15.3 | 12.2-13.9 | 13.8-14.9 | 18.2-19.5 | 18.1-18.8 | 17.2-18.6 | 17.4-18.6 | 20.1-21.1 | 18.3-19.5 | 17.5-18.7 |
| XI | 13.4-14.5 | 11.4-13.8 | 10.7-12.4 | 12-14.9 | 14.1-14.9 | 11.6-12.5 | 14.6-15.6 | 16.6-18.1 | 15.6-16.7 | 15.7-16.9 | 17.5-18.1 | 20.1-21 | 18.2-19.6 | 18-19 |
| XII | 12.8-13.7 | 10.9-14.4 | 11.1-13.2 | 11.5-13.7 | 12.9-13.3 | 12.4-13 | 14.7-15.6 | 15.5-16.3 | 17-17.5 | 15.7-16.8 | 17.2-17.6 | 21.3-21.5 | 18.2-18.7 | 18.8-19.8 |

Table 2. Continued

|  | XVII | XVIII | XIX | XX | XXI | XXII | XXIII | XXIV | XXV | XXVI | XXVII | XXVIII | XXIX | XXX |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| XIII | 12.6-13.7 | 9.4-11.8 | 9.6-12.3 | 11.4-13.9 | 11.8-13.7 | 9.8-11.1 | 12.3-13.4 | 16.4-17.2 | 16.2-17.7 | 15-15.5 | 16.7-17.9 | 19.6-20.5 | 17.4-18.8 | 18-18.8 |
| XIV | 7.8-8 | 11.6-13.5 | 12.1-12.9 | 12.8-14 | 13.9-14.1 | 12.6-13.3 | 13.5-14.1 | 16.1-16.6 | 16-16.6 | 17.3-17.6 | 17.6-17.8 | 19.2-19.3 | 16.9-17.1 | 17.2-17.5 |
| XV | 8.8-9.5 | 12.3-14.6 | 12.2-13.7 | 12.8-14.5 | 14.1 | 12.3-13.1 | 15.3-16.1 | 17.2-18.1 | 17.2-17.8 | 17.7-18.7 | 18.5-19.2 | 20.6-21.3 | 17.3-17.9 | 17.1-17.8 |
| XVI | 9.9-10.2 | 12-13.7 | 12.5-13.5 | 12.3-13.8 | 13.4-13.6 | 12.7-13 | 15.1-15.6 | 17-18 | 16.9-17.8 | 17.9-18 | 18-18.2 | 20.8-21 | 17.6-18.2 | 18.5-18.7 |
| XVII |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| XVIII | 12.7-14.9 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| XIX | 12.7-13.5 | 10.8-13.9 |  |  |  |  |  |  |  |  |  |  |  |  |
| XX | 13.6-14.5 | 11.5-14.8 | 9.9-11.6 |  |  |  |  |  |  |  |  |  |  |  |
| XXI | 16.5 | 12.4-13.9 | 13.2-14.1 | 9.1-9.8 |  |  |  |  |  |  |  |  |  |  |
| XXII | 12.2-12.4 | 12.9-14.9 | 12.4-13.3 | 13.9-15.5 | 16.5 |  |  |  |  |  |  |  |  |  |
| XXIII | 15-15.4 | 13.4-15.7 | 13.8-15.1 | 16.3-17.7 | 16.1-16.4 | 13.3-13.7 |  |  |  |  |  |  |  |  |
| XXIV | 16.6-16.9 | 17.8-19.2 | 17.8-18.7 | 16.3-18.2 | 17.9-18 | 18.3-18.5 | 19.1-19.9 |  |  |  |  |  |  |  |
| XXV | 17.8 | 19.4-21.1 | 17.6-18.3 | 19.3-20.7 | 20.3 | 17.3 | 18.9-19.1 | 17.7-17.8 |  |  |  |  |  |  |
| XXVI | 18.6-18.7 | 17.9-20.2 | 16.1-17.3 | 18.3-19 | 19.6 | 16.6 | 16.7-16.2 | 15.3-15.5 | 13.1 |  |  |  |  |  |
| XXVII | 18.8 | 18.6-20.9 | 17.2-17.7 | 18.6-19.5 | 18.9 | 17.9 | 20.1-20.4 | 16.3-16.4 | 13 | 15.7 |  |  |  |  |
| XXVIII | 20.5 | 20.2-22.1 | 19.8-20.5 | 22.3-23.1 | 22 | 21.1 | 20.1-20.9 | 20.7-20.9 | 18.8 | 18.1 | 19.6 |  |  |  |
| XXIX | 16 | 18-20.2 | 16.9-17.3 | 17.2-19.6 | 20.3 | 17.4 | 19.3-19.5 | 17.3-17.5 | 16.7 | 18.1 | 18.5 | 19.3 |  |  |
| XXX | 17.1 | 19.6-22 | 17.5-18 | 18.9-20.5 | 21.2 | 17.9 | 19.8-19.9 | 17.4-17.5 | 16.2 | 18.6 | 18.5 | 19.8 | 6.5 |  |
| XXXI | 17.4-17.7 | 18-20.2 | 16.7-17.8 | 18.5-19.8 | 20.1 | 18.1 | 19.9-20.1 | 16.9-17.1 | 17.4 | 19 | 19.3 | 19.1 | 16.6 | 16.4 |

Table 3. Uncorrected $p$-distances based on COI sequences for specimens previously identified as Runcina coronata

|  | Distance between groups (\%) |  |  | Distance within groups (\%) |
| :---: | :---: | :---: | :---: | :---: |
|  | RC1 | RC2 | RC3 |  |
| RC1 |  |  |  | 0.2-0.5 |
| RC2 | 6.3-7.3 |  |  | 1.0 |
| RC3 | 6.4-7.8 | 7.0-7.7 |  | 0.7 |
| RC4 | 7.8-8.0 | 8.8-9.5 | 9.9-10.2 | 0.0 |

with encrusting pink coralline algae (dissected and sequenced).

External morphology (Fig. 3A, B): Living specimens $3-4 \mathrm{~mm}$ in length. Body elongated and moderately broad. Lateral grooves on both sides between notum and foot. Notum smooth. Posterior part of the notum rounded. Foot slightly wider than the notum. Propodium rounded and wide, metapodium pointed. General ground colour of body dark brown. Front part of the notum dark brown on central zone between the eyes and pale fawn on the sides. White and yellowish spots all over the body, concentrated as semicircle bands behind the eyes and anterior to the notum end, and on the surface of the foot. Eyes small and visible. Three rounded gill laminae present on right of anus; the largest in the middle divided into two arcs. Gills yellowish with slightly brown margins. Anus located in median line of body, beneath the end of notum.

Internal anatomy (Figs 4A-C, 5A, 6A): Radular formula $19 \times 1.1 .1$ (MNCN 15.05/88105). Rachidian tooth bilobed with long and smooth lateral wings on each side. Central part of rachidian tooth contains a pair of pads, each possessing $9-11$ denticles. Size of denticles variable, with smaller and more developed denticles randomly distributed along pads. Small depression present between pads, with minute denticle present in some rows (Fig. 4A). Lateral teeth denticulate, elongate, hooked shape with 30-34 relatively long denticles (Fig. 4B). Triangular jaws present. Four gizzard plates with ten crests (Fig. 4C). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind the digestive gland divided into two lobes. Female gland opens to exterior through median size common genital duct (Fig. 5A). Male pore opens next to mouth, on the right side. Male copulatory organ elongated and cylindrical. Penial papilla absents. Prostate gland cylindrical, posterior to a narrow atrium. Slender seminal vesicle half size of prostate gland (Fig. 6A).

Distribution: Bréhat, Bretagne, France (Atlantic) (Quatrefages, 1844); Torbay, England. (Alder \&

Hancock, 1846, Forbes, 1853); Swanage, England (present study).

Remarks: A discussion of this species is included together with $R$. aurata in the 'Remarks' of the latter species.

## Runcina aurata García $\operatorname{ET}$ AL., 1986

(Figs 3C-F, 4D-F, 5B, 6B)
Type locality: Club La Hacienda, Cádiz, Spain ( $36^{\circ} 14^{\prime} 18^{\prime \prime} \mathrm{N} ; 5^{\circ} 18^{\prime} 36^{\prime \prime} \mathrm{W}$ )

Examined material: MNCN 15.05/91500, La Caleta (Cádiz) ( $36^{\circ} 31^{\prime} 59^{\prime \prime N} ; 6^{\circ} 18^{\prime} 31 "$ W), Andalusia, southwestern Spain, 8 April 2019, 3.5 mm living animal, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced). MNCN $15.05 / 88106$, La Caleta (Cádiz) $\left(36^{\circ} 31^{\prime} 59^{\prime} \mathrm{N} ; 6^{\circ} 18^{\prime} 31^{\prime} \mathrm{W}\right)$, Andalusia, south-western Spain, coll. Josep Romà, 18 April 2015, 2 mm in length preserved, depth 0.5-1.0 m . (dissected and sequenced). MNCN 15.05/88107, La Caleta (Cádiz) ( $36^{\circ} 31^{\prime} 59$ "N; $6^{\circ} 18^{\prime} 31^{\prime \prime} \mathrm{W}$ ), Andalusia south-western, Spain, coll. Josep Romà, 18 April 2015, 2 mm in length preserved, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced). MNCN:ADN 118948, La Caleta (Cádiz) ( $36^{\circ} 31^{\prime} 59^{\prime \prime} \mathrm{N} ; 6^{\circ} 18^{\prime} 31^{\prime \prime W}$ ), Andalusia southwestern, Spain, coll. Josep Romà, 17 May 2015, 1.5 mm in length preserved, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced). MNCN:ADN 118950, El Chato (Cádiz) ( $36^{\circ} 28^{\prime} 39^{\prime N}$ N; $6^{\circ} 15^{\prime} 49$ "W), Andalusia south-western, Spain, coll. Ana Bartual, 13 April 2015, 1 mm in length preserved, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced).

External morphology (Fig. 3C-F): Living specimen 3.5 mm length and preserved specimens $1-2 \mathrm{~mm}$ length. Body elongated and moderately broad. Lateral grooves on both sides between notum and foot. Anterior part of notum ('head') slightly bilobed. Posterior part of notum rounded. Propodium rounded, metapodium pointed. Foot as wide as notum. Foot extended beyond notum on rear part. Ground colour of body translucent pale fawn or yellowish. Digestive system visible as a broad brownish blotch. White spots on central zone of notum, behind eyes forming


Figure 3. Living animals of Runcina coronata (A, B) and Runcina aurata (C-F). A, MNCN $15.05 / 88105,3 \mathrm{~mm}$ in length, Swanage (southern England). B, MNCN $15.05 / 90423,4 \mathrm{~mm}$ in length, Swanage (southern England) (photos Ian F. Smith). C, MNCN/ADN 118948, 1.5mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). D, MNCN/ADN 118950, 1 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). E, MNCN 15.05/88106, 2 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). F, MNCN 15.05/88107, 2 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). Photos A and B, courtesy of Ian F. Smith; photos C-F by Ana Karla Araujo.
triangular patches and anterior to notum end. White spots maybe also absent. Black dots dispersed on notum and more concentrated on head zone. Eyes inconspicuous. Dark band on middle of dorsal surface of foot. Black dots may be present on ventral surface of foot. Four rounded and relatively large gills laminae to the right of anus. Gills yellowish
with slightly brown margins. Anus located in median line of body, beneath the end of notum.

Internal anatomy (Figs 4D-F, 5B, 6B): Radular formulae $12 \times 1.1 .1$ (MNCN 15.05/88106) and $13 \times 1.1 .1$ (MNCN 15.05/91500). Rachidian tooth bilobed with long and smooth lateral wings on each side. Central


Figure 4. Scanning electron micrographs of radula and gizzard plates. A-C, Runcina coronata. A, rachidian teeth (MNCN $15.05 / 88105$ ). B, lateral teeth (MNCN 15.05/88105). C, gizzard plate (MNCN 15.05/90423). D-F, Runcina aurata. D, rachidian teeth (MNCN 15.05/91500). E, lateral teeth (MNCN 15.05/88106). F, gizzard plate (MNCN 15.05/88106). G-I, Runcina caletensis (MNCN 15.05/200113) G, rachidian teeth. H, lateral teeth. I, gizzard plate. J-M, Runcina tingensis. J, rachidian teeth (MNCN 15.05/200114). L, lateral teeth (MNCN 15.05/200114). M, gizzard plate (MNCN 15.05/91514). Scale bars: A, B, E, J, L $=10 \mu \mathrm{~m} ; \mathrm{C}, \mathrm{F}, \mathrm{I}, \mathrm{M}=50 \mu \mathrm{~m} ; \mathrm{D}, \mathrm{G}=20 \mu \mathrm{~m} ; \mathrm{H}=5 \mu \mathrm{~m}$.
part contains pair of pads, each possessing 10-11 long, slender, pointed denticles. Size of denticles variable. Small denticles between large denticles. Small depression present between pads, with minute denticle present (Fig. 4D). Lateral teeth denticulate, elongate, hooked shape with $35-36$ long, pointed and samesize denticles (Fig. 4E). Triangular jaws present. Four gizzard plates with seven to nine crests (Fig. 4F). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind digestive gland,
opening to exterior through small size common genital duct (Fig. 5B). Male pore opens next to mouth, on the right side. Elongated and cylindrical male copulatory organ. Penial papilla not observed. Cylindrical and long prostate gland ends in slender and small seminal vesicle with black pigmentation (Fig. 6B).

Distribution: Cádiz, Strait of Gibraltar, Malaga and Murcia (southern Spain) (Templado, 1984; Garcia et al., 1986) and Azores Islands (Portugal) (Gosliner, 1990).


Figure 5. Female reproductive system. A. R. coronata, southern England (MNCN 15.05/90423). B. R. aurata, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/88106). C. R. caletensis, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/200113). D. R. tingensis, Tangier, north-western Morocco, Atlantic Ocean (MNCN 15.05/91514). Abbreviations: FM, female mass; CGD, common genital duct; GO, gonopore.

Remarks: Runcina coronata has been considered a taxonomically difficult species (see Introduction). Originally described from Brehat (Atlantic coast of France), this species was first reported in the Mediterranean Sea by Vayssière (1883) who identified specimens from Marseille (Mediterranean coast of France) as R. coronata. However, Burn (1963), based on morphological differences, especially the shape of the body and the colour pattern, suggested the
specimens identified by Vayssière (1883) could be R. calaritana. We cannot confidently attribute those specimens to a specific species, but we agree with Burn (1963) that they probably do not correspond to $R$. coronata. Pruvot-Fol (1954), and Cervera et al. (1991) regarded R. calaritana (Gulf of Cagliari, Sardinia, Italy) and R. aurata (from around the Strait of Gibraltar) conspecific with $R$. coronata. This problematic has ultimately created the perception


Figure 6. Male reproductive system. A, Runcina coronata, southern England (MNCN 15.05/90423). B, Runcina aurata, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/88106). C, Runcina caletensis, La Caleta, Cádiz, southwestern Spain, Atlantic Ocean (MNCN 15.05/200113). D, Runcina tingensis, Tangier, north-western Morocco, Atlantic Ocean (MNCN 15.05/91514). Shaded area indicates the presence of sperm. Abbreviations: MO, male opening; PP, penial papilla; PG, prostate gland; SV, seminal vesicle.
that $R$. coronata was present in the Mediterranean Sea (Schmekel \& Cappellato, 2002; Cervera et al., 2004; Ballesteros et al., 2016).

In general, the external and internal morphology of our specimens of $R$. coronata from Swanage
(England) are consistent with the original description of the species (Quatrefages, 1844), and with the description provided by Schmekel \& Cappellato (2002) based on specimens from Roscoff (Atlantic coast of France) and Plymouth (south of

England). However, compared with the description provided by García et al. (1986; specimens from the Strait of Gibraltar), our animals from England exhibit several differences, mainly in the shape of the body and colour pattern. The anterior and posterior ends of the notum are rounded, while in Spanish specimens it is pointed (García et al., 1986). The colour pattern of our specimens (Fig. 3A, B) differs drastically from those from the Strait of Gibraltar, which have a uniformly dark colour pattern, two whitish bands on both sides of the head and one white small band on the posterior right side of the notum (García et al., 1986). This suggests that likely specimens attributed to $R$. coronata by García et al. (1986) belong to a distinct species.
The original description of $R$. coronata describes briefly the male copulatory organ as 'a rather short testicular bag in the shape of a "club", with a seminal vesicle sometimes absent (Quatrefages, 1844).' Kress (1977) studied specimens from Plymouth (England) and provided additional anatomical data on the reproductive system. Comparatively, our specimens from Swanage (England) exhibit a similarly long and cylindrical prostate, but a slightly different seminal vesicle and common genital duct. Kress (1977) referred to a seminal vesicle 'considerably shorter than prostate' and a common genital duct forming a long loop, whereas in our material the seminal vesicle was approximately half the size of the prostate and the common genital duct was short (Fig. 6A).

The species $R$. aurata was described by García et al. (1986) from the southern coast of Spain (Cádiz, Strait of Gibraltar and Malaga). Gosliner (1990) reported the species from the Azores and suggested that a specimen illustrated and depicted by Thompson \& Brodie (1988: fig. 1E) from Plymouth as $R$. coronata was most likely $R$. aurata. Despite the fact that the description and illustration provided by Thompson \& Brodie (1988) are vague and lacking important information, the reference to the presence of a light area surrounding the eyes, suggests their identification as $R$. coronata to be correct.

The features of our specimens collected in Cádiz (Spain) are consistent with the original description of the species $R$. aurata (García et al., 1986). Externally, they differ from $R$. coronata by having a translucent yellow colour with black spots on the notum and on the ventral surface of the foot (Fig. 3; Table 4). Also, the number of gills is distinct: three gills in $R$. coronate, while our animals of $R$. aurata from Cádiz have four gills (Table 4). The original description of $R$. aurata refers, in fact, to three gills only, but the authors did not seem to have thoroughly looked at this character, which is difficult if not examined properly and across several specimens (García et al., 1986).

Concerning the radula, we observed some subtle differences between $R$. aurata and $R$. coronata, namely in the number of radular rows and shape of the denticles in the pads of the rachidian teeth (Table 4). However, in runcinids, these features can vary, even within species (Schmekel \& Cappellato, 2001; 2002; Araujo et al. 2019), and are, therefore, difficult to use

Table 4. Summary of diagnostic characters between Runcina coronata, Runcina aurata, Runcina caletensis and Runcina tingensis. Data based on Quatrefages (1844), Vayssière (1883), García et al. (1986), Cervera et al. (1991), Schmekel \& Cappellato (2002) and present study

|  | Runcina coronata | Runcina aurata | Runcina caletensis | Runcina tingensis |
| :---: | :---: | :---: | :---: | :---: |
| Colour pattern | Ground colour dark brown. White semicircles bands behind eyes and anterior to notum end. | Ground colour translucent pale fawn or yellowish. White spots behind eyes and anterior to notum end. | Ground colour translucent pale fawn or yellowish. Black dots on notum end. Two black longitudinal lines on the head. | Ground colour light brown. Triangular white patches behind eyes and anterior to notum end. |
| Gills | Three rounded | Four rounded and relatively large. | Three rounded | Two rounded |
| Shell | Absent | Absent | Absent | Absent |
| Radular formulae | $19 \times 1.1 .1$ | $12-13 \times 1.1 .1$ | $13 \times 1.1 .1$ | $12-14 \times 1.1 .1$ |
| Rachidian teeth | 9-11 short denticles on each pad | 10-11 long, slender, pointed denticles | 7-8 denticles stalactiteshaped. Depression between pads absent. | 7-10 short triangular denticles. |
| Lateral teeth | 30-34 denticles | 35-36 denticles | 33-36 denticles | 34-37 denticles |
| Gizzard plate | 10 crests | 7-9 crests | 7-10 crests | 8 crests |

in species identification. Regarding the gizzard plates, our specimens of $R$. coronata show ten crests in each plate, while in our animals of $R$. aurata it ranges from seven to nine, which is consistent with its original description (García, et al., 1986).
The male reproductive system in our specimens of $R$. aurata resembles the description of this organ by Gosliner (1990) for specimens from the Azores, but we could not observe a penial papilla. The seminal vesicle is shorter than in $R$. coronata and the atrium and male opening are broader (Fig. 6A, B). The female gland mass of $R$. aurata was never studied before and in our specimens of consists of one lobe, while in the studied specimens of $R$. coronata it is divided into two lobes (Fig. 5B).
The minimum uncorrected $p$-distance for the $C O I$ gene between $R$. coronata and $R$. aurata is $6.3 \%$ (Table 3) and, in addition to the phylogenetic tree, the species delimitation analyses suggested both species as valid (Fig. 2).

## Runcina caletensis Arauju, Pola, Malaquias \& Cervera, sp. nov.

(Figs 4G-I, 5C, 6C, 7A, B)
Zoobank registration: urn:lsid:zoobank. org:act:EC216698-6700-4607-9E35-515AB69BD17C

Examined material: Holotype: MNCN 15.05/200113, La Caleta (Cádiz) ( $36^{\circ} 31^{\prime} 59$ "N; $\left.6^{\circ} 18^{\prime} 31^{\prime \prime W}\right)$, Andalusia, south-western Spain, coll. Josep Romà, 17 May 2015, 3 mm living animal, depth $0.5-1 \mathrm{~m}$ (dissected and sequenced). Paratype: MNCN:ADN 118949, La Caleta (Cádiz) ( $36^{\circ} 31^{\prime} 59^{\prime \prime} \mathrm{N} ; 6^{\circ} 18^{\prime} 31^{\prime \prime} \mathrm{W}$ ), Andalusia, southwestern Spain, coll. Josep Romà, 16 June 2015, 1.5 mm in length preserved, depth $0.5-1.0 \mathrm{~m}$. (dissected and sequenced).

Etymology: Named after the type locality: the beach of La Caleta in Cádiz, Spain.

External morphology (Fig. 7A, B): Living and preserved specimens 3.0 mm and 1.5 mm in length, respectively. Body elongated and broad. Lateral grooves on both sides between notum and foot. Anterior part of notum ('head') slightly bilobed. Posterior part of notum rounded. Foot as wide as notum. Foot extends posteriorly beyond notum. Ground colour translucent pale fawn or yellowish. Digestive system visible as broad brownish blotch in juvenile specimens. Tiny white, black and yellow spots all over the body. White spots on the lobes of the head and on the anterior ventral surface of the foot. Triangular white patches behind the eyes. White semicircle anterior to the notum end. Very
few white spots in juvenile. Small black dots forming two longitudinal lines on head region. Black spots concentrated on the notum end posteriorly to white semicircle. In juveniles, only a few larger black spots are present along the margin of the notum. Eyes inconspicuous. Dark band on middle of posterior region of foot. Three rounded gill laminae located on right side of anus. Gills yellowish with slightly brown margins. Anus located in the midline of the body beneath the notal edge.

Internal anatomy (Figs 4G-I, 5C, 6C): Radular formula $13 \times 1.1 .1$ (15.05/200113). Rachidian tooth slightly bilobed with long and smooth lateral wings on each side. Central part of rachidian tooth contains a pair of pads, each possessing seven to eight developed denticles. Central depression between pads absent. Denticles long and pointed, decreasing in size towards middle of the tooth (Fig. 4G). Lateral teeth denticulate, elongate, hooked shaped with 33-36 long and pointed denticles (Fig. 4H). Triangular jaws present. Four gizzard plates with seven to ten crests (Fig. 4I). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind digestive gland. Female gland opens to exterior through common genital duct (Fig. 5C). Male pore opens next to mouth, on right side. Elongated and cylindrical male copulatory organ. Short, conical and unarmed penial papilla projects into large atrium. Cylindrical prostate gland strongly curved. Long and slender seminal vesicle with black pigmentation (Fig. 6C).

Distribution: Cádiz, southern Spain (present study).
Remarks: The species $R$. caletensis shares with $R$. coronata the presence of white bands and spots on the notum, and is overall externally similar to $R$. aurata. However, in $R$. caletensis the yellow ground colour is opaque, while in $R$. aurata the colour is translucent (Figs 3E, F, 7B). In addition, R. caletensis lacks black spots on the ventral surface of the foot, a feature present in $R$. aurata. Based on available data, $R$. caletensis is the only one among these species with rachidian teeth lacking the depression between the pads and with well-developed denticles of similar length along the masticatory edge of pads (Fig. 4G). The female gland mass of $R$. caletensis has a rounded shape and the common genital duct is larger than in the other two species (Fig. 5A-C). The seminal vesicle in $R$. caletensis is thinner than in $R$. coronata and more elongated than in $R$. aurata. Unlike for $R$. coronata and $R$. aurata, a penial papilla was observed in $R$. caletensis.

The minimum uncorrected $p$-distances for the COI gene is $7.8 \%$ between $R$. caletensis and $R$. coronata,


Figure 7. Living animals of Runcina caletensis (A, B) and Runcina tingensis (C, D). A, MNCN/AND 118949, 1.5 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). B, MNCN 15.05/200113, 3 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). C, MNCN $15.05 / 91514,1.5 \mathrm{~mm}$ in length, Tangier (north-western Morocco; Atlantic Ocean). D, MNCN $15.05 / 200114,2 \mathrm{~mm}$ in length, Tangier (north-western Morocco; Atlantic Ocean). A, B images by Ana Karla Araujo; C, D images courtesy of Naoufal Tamsouri.
and $8.8 \%$ between $R$. caletensis and $R$. aurata (Table 3). Species delimitation analyses recognized $R$. caletensis as a valid species (Fig. 2).

Runcina tingensis Araujo, Pola, Malaquias \& CERVERA, SP. NOV.
(Figs 4J-M, 5D, 6D, 7C, D)
Zoobank registration: urn:lsid:zoobank. org:act:84C29263-8B36-4FA7-A84A-94F4191CE491

Examined material: Holotype: MNCN 15.05/200114, Tangier ( $35^{\circ} 47^{\prime} 32 " \mathrm{~N} ; 5^{\circ} 49^{\prime} 04 " \mathrm{~W}$ ), Morocco, coll. Naoufal Tamsouri, 22 March 2018, 2 mm living animal, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced). Paratype: MNCN 15.05/91514, Tangier ( $35^{\circ} 47^{\prime} 32$ "N; $5^{\circ} 49^{\prime} 04^{\prime \prime}$ W), Morocco, coll. Naoufal Tamsouri, 22 March 2018, 1.5 mm fixed animal, depth $0.5-1.0 \mathrm{~m}$ (dissected and sequenced).

Etymology: The name tingensis refers to Tingi, the Greek name of Tangier (Morocco), the type locality of the species.

External morphology (Fig. 7C, D): Living and preserved specimens 2.0 mm and 1.5 mm in length, respectively. Body elongated. Lateral grooves on both sides between notum and foot present. Anterior part of notum ('head') straight, rounded on posterior end. Foot as wide as notum, with propodium and metapodium rounded; metapodium extends beyond notum. Ground colour of body light brown. Broad continuous light orange line on edge of notum. Digestive system visible as a broad brownish blotch. White spots concentrated behind eyes and on anterior end of notum forming a triangle. Yellowish dots dispersed on middle of notum. Dark small spots can be present behind anterior white spots on head region and rear part of notum. Eyes inconspicuous. Longitudinal mid-dorsal dark band on foot. Two rounded gills laminae on right side of anus. Gills light brown. Anus located on right lateral side beneath edge of notum, approximately in mid-region of body length.

Internal anatomy (Figs 4J-M, 5D, 6D): Radular formulae $14 \times 1.1 .1$ (MNCN 15.05/91514) and $12 \times 1.1 .1$ (MNCN 15.05/200114). Rachidian tooth bilobed with smooth lateral wings on each side. Central part of rachidian tooth contains pair of pads, each possessing seven to ten denticles. Denticles short, pointed. One inner denticle on each pad conspicuously more developed. Central small depression present between pads; small denticle in-between pads absent (Fig. 4J). Lateral teeth denticulate, elongate and hooked shaped with 34-37 long, thin, pointed denticles (Fig 4L). Triangular jaws present. Four gizzard plates with eight crests (Fig. 4M). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind
digestive gland. Opens to exterior through short and wide common genital duct (Fig. 5D). Male pore opens next to mouth, on right side. Male copulatory organ cylindrical. Short and unarmed penial papilla projects into round atrium. Prostate gland cylindrical and strongly curved with posterior part rounded. Seminal vesicle slender with middle part wider (Fig. 6D).

## Distribution: Tangier, Morocco (present study).

Remarks: Runcina tingensis resembles $R$. coronata by its dark colour and presence of small, white and yellow spots on the notum. However, $R$. tingensis has a broad, continuous, light orange line along the edge of the notum, while $R$. coronata has lighter regions only on the head and on the back of the notum. Compared with $R$. aurata and $R$. caletensis, the dark colour of $R$. tingensis contrasts with the translucent yellowish colour of these two species. The distribution of white spots is also distinctive among these species. In R. tingensis they form a triangular pattern on the notum (Fig. 7C, D), while in the other three species they form a semicircle (Figs 3, 7B). According to our observations, the radula of $R$. tingensis has, compared with $R$. aurata, $R$. coronata and $R$. caletensis, a prominently more developed denticle in each pad of the rachidian teeth (Fig. 4J). The female mass of $R$. tingensis differs from $R$. coronata and $R$. aurata by its elliptical shape and a larger common genital duct (Fig. 5), and its prostate is notably curved and the seminal vesicle enlarged in its central region (Fig. 6). A penial papilla was present as observed for $R$. caletensis.

The minimum uncorrected $p$-distances for the COI gene is $7 \%$ between $R$. tingensis and $R$. coronata, $6.4 \%$ between $R$. tingensis and $R$. aurata, and $9.9 \%$ between $R$. tingensis and $R$. caletensis (Table 3).

## DISCUSSION

Burn (1963) presented a systematic classification of the order Runcinida (as Runcinacea) based on morphoanatomical characters: in particular, shell, gills and radula. According to his classification, the rare Ildica nana Bergh, 1889 was the most primitive species followed by the genera Runcina, Runcinida and Runcinella, while the genus Ilbia was considered the most derived. Our results, suggest a sister-relationship between the genus Ilbia and the remaining members of Runcinida with genera Runcina, Runcinida, Metaruncina and Lapinura ( $\mathrm{PP}=1 ; \mathrm{BS}=99$ ). Unfortunately, samples of Ildica and Runcinella were not available to us and, therefore, we could not thoroughly test Burn's (1963) hypothesis, but despite
the limitations of our dataset, the results suggest the validity of the families Ilbiidae (with Ilbia) and Runcinidae (with Runcina, Runcinida, Metaruncina and Lapinura).

## RUNCINA DIVAE OR LAPINURA DIVAE?

The genus Lapinura (type species: Ildica divae Marcus \& Marcus, 1963), introduced by Marcus \& Marcus (1970) for specimens collected in Curaçao, Bonaire and Florida, is characterized by an external cupshaped larval shell. This genus was synonymized with Runcina by Clark (1984) after examining populations from Bermuda of supposed Lapinura divae in which some specimens lacked the external shell. Because of this 'variability', Clark disregarded the external shell and others characteristics that define the genus, emphasizing only the radular formula ( $N \times 1.1 .1$ ) and the presence of gizzard plates - common for most runcinids - to assign Lapinura divae to the genus Runcina.

Representatives of the Caribbean species Lapinura / Runcina divae from Bermuda branched off outside Clade F (the one with the type species of the genus Runcina) with other runcinids in Clade H. Therefore, we suggest that the genus Lapinura should be reinstated as valid and at least applied for the Caribbean species described by Marcus \& Marcus (1963). Moreover, we agree with Ortea et al. (2017) about a possible existence of more than one species being referred under the name Lapinura divae, due to the discrepancies (number of rows of the radula, presence/absence of the shell and number of crests of the gizzard plates) among specimens collected in the Caribbean Sea and Brazil (Marcus \& Marcus, 1963; Marcus \& Marcus, 1970; Thompson, 1977; Clark, 1984).

## What about European runcinids?

Three genera have been referred in European waters: Runcina (31 species), Runcinella (one species) and Pseudoilbia (one species). We have studied specimens of Runcina and Pseudoilbia but, unfortunately, specimens of Runcinella condio Moro \& Ortea, 2015, described from the Canary Islands, were not available for study.

## Runcinida

Here we add a fourth genus to the European fauna, namely Runcinida. To date, the genus Runcinida was restricted to the western Pacific Ocean and included three species, R. elioti (Baba, 1937), R. valentinae and $R$. marisae. However, according to our results, a specimen identified as Runcinida sp. 1, collected in

Cap Ferret, France (Atlantic coast), clustered together with $R$. marisae and $R$. valentinae ( $\mathrm{PP}=1, \mathrm{BS}=90$ ). Externally, all species of Runcinida can be distinguished by their unique colour pattern, with a dark-brown notum, yellowish or orange edge of notum and foot, and gills arranged in a semicircle above the anus. Our specimen (Runcinida sp. 1; Fig. 8) fits this colour pattern and arrangement of the gills, but differs from the other species in the genus by having small, black dots spread on the dorsal and ventral surfaces of the foot and several larger white spots on the notum (Fig. 8). Runcinida elioti (Baba, 1937) has fewer white spots too, but lacks small, black dots on the foot (Baba, 1937). Runcinida marisae has small, black dots on the ventral surface of the foot but white dots are absent (Chernyshev, 1998). Finally, Runcinida valentinae lacks black and white dots, but has a triangular orange patch on the anterior part of the notum (Chernyshev, 2006). The ABDG and bPTP recognized Runcinida sp. 1, R. marisae and $R$. valentinae as distinct species with COI uncorrected $p$-distances ranging between $6.5 \%$ to $16.6 \%$ (Table 2).

## Pseudoilbia avellana or Runcina avellana?

The species Runcina avellana was originally described by Schmekel \& Cappellato (2001) from Banyuls-surMer, French Mediterranean. These authors found an unusual radula ( $3 \times 1.0 .1$ ) and no gizzard plates in the single specimen examined, which they pointed out could be a juvenile. Ortea (2013), because of the absence of raquidian teeth and gizzard plates assigned the species to the genus Pseudoilbia, proposing the new combination name Pseudoilbia avellana.

Our specimens characterized by features consistent with the original description of $R$. avellana (sensu Schmekel \& Cappellato, 2001; Fig. 9), like the shape


Figure 8. Living animal of Runcinida sp. (MNCN 15.05/90670, 3 mm in length, Cap Ferret, north of France, Atlantic Ocean). Image courtesy of Marina Poddubetskaia.
of the body, presence of a round brownish mark on the centre of tail, general colour pattern, absence of gizzard plates and presence of gills, collected at Roses, Spain about 45 km from the type locality, clustered in the phylogenetic analyses among species of the genus Runcina (Clade F; Fig. 1), questioning the assignment by Ortea (2013) of this species to the genus Pseudoilbia. The latter genus is characterized by animals lacking gills, gizzard plates, shell and with radular formula 2.0.2 (Miller \& Rudman, 1968). Runcina avellana shares some of these features, but not all, and our phylogenetic results support its inclusion in Runcina. Moreover, the lack of rachidian teeth in R. avellana, reported by Schmekel \& Cappellato (2001), might be an artefact since only one apparently juvenile specimen was studied by these authors. Unfortunately, our effort in preparing the radula of this species was not successful and, thus, we could not study this structure. Additional specimens of $R$. avellana are necessary to permit a detailed study of its anatomy and comparison with other species of Runcina and Pseudoilbia.

## Runcina

The genus Runcina has been traditionally defined by the presence of up to four separated gills on the right side of the anus and a triseriate (1.1.1) radula with bilobed rachidian teeth and smooth or denticulated lateral teeth (Burn, 1963; Gosliner, 1991; Schmekel


Figure 9. Runcina avellana. A, schematic illustration taken from Schmekel \& Cappellato, 2001 (original description). B, living animal from Catalonia, north-eastern Spain (Mediterranean Sea) (MNCN 15.05/88108, 1,5 mm in length). Image B by Ana Karla Araujo.
\& Cappellato, 2001). Our analysis supports the monophyly of the genus Runcina ( $\mathrm{PP}=1 ; \mathrm{BS}=99$ ) but with $R$. avellana, which lacks rachidian tooth and, in addition, specimens resembling $R$. ferruginea from the Mediterranean coasts of Spain and France, and from Croatia clustered elsewhere in the tree together with other runcinids from Bermuda, Hawaii, Brazil, Japan, Russia and also from the Atlantic coast of France ( $\mathrm{PP}=1 ; \mathrm{BS}=78$ ).

We have included in our phylogenetic analysis about $39 \%$ of the nominal species of European Runcina (MolluscaBase, 2021) and, in addition, several unidentified specimens from Spain, Italy and Croatia (Fig. 1). Within Runcina, we retrieved a clade ( $\mathrm{PP}=1 ; \mathrm{BS}=100$ ) containing one specimen that we provisionally have identified as $R$. cf. bahiensis, one specimen provisionally identified as $R$. hornae and several unidentified specimens. Most specimens in this clade were collected in Catalonia (northeastern Mediterranean Spanish coast) and, despite the remarkable variation in colour pattern (Fig. 10), species delimitation analyses suggest they all belong to the same species (COI uncorrected $p$-distances varied between $0.0-2.0 \%$ ).

The species $R$. bahiensis was originally described from the Bay of Algeciras (Strait of Gibraltar, Spain) (Cervera et al., 1991) and R. hornae from Banyuls-sur-Mer (Mediterranean coast of France) (Schmekel \& Cappellato, 2002). Both species have been reported in several localities in Catalonia (Sánchez-Moyano et al., 2000; Ballesteros et al., 2016), and are regarded as differing in colour pattern, number of rows of radular teeth, shape of body and presence of small protuberances (Cervera et al., 1991; Schmekel \& Cappellato, 2002). Several of our specimens (Fig. 10BD) bear a colour pattern and body shape consistent with $R$. bahiensis, but none of them have the small protuberances characteristic of this species. The study of a radula of one specimen from Catalonia (Runcina sp. 3) matched the original description of the radula of R. hornae (Schmekel \& Cappellato, 2002; Fig. 11). Unfortunately, specimens of $R$. bahiensis from the type locality or nearby were not available for this study, but considering the radular similarities and the proximity to the type locality of $R$. hornae (c. 25.5 km ), we identify specimens is this clade as $R$. hornae, which is here demonstrated to be a species with remarkable colour variability (Fig. 10).

## Runcina coronata species complex

In the last few years, several complexes of species have been detected among heterobranch sea slugs (see, among others: Jörger \& Schrödl, 2013; Padula et al., 2014; Carmona et al., 2015; Krug et al., 2016; Korshunova et al., 2017; Austin et al., 2018). In

Runcinida, the first complex of species was unravelled by Araujo et al. (2019) for the species Runcina brenkoae with the description of two new species, namely R. marcosi and R. lusitanica.

In the current study, molecular and morphological data have showed that $R$. coronata hides a complex of at least four species, including two new to science and the previously described species $R$. aurata. Due to similarities with the original description by Quatrefages (1844), and the descriptions provided by Forbes (1851) and Schmekel \& Cappellato (2002), we regard our specimens from Swanage (England) conspecific with $R$. coronata.

The geographical distribution of $R$. coronata has been reported to extend from England to the French Mediterranean coast (Vayssière, 1883; Cervera et al., 2004). However, our results questioned the presence of $R$. coronata in the Iberian Peninsula, where most likely the records to this species belong to $R$. aurata (see in Results, 'Remarks' section of R. aurata). The morphological differences between specimens identified as $R$. coronata in the Mediterranean Sea and those from England and the Atlantic coast of France suggest, as previously stressed by Burn (1963), that animals studied by Vayssière (1883) from the Mediterranean are a distinct species and also that R. calaritana (Colosi, 1915) could be a valid name.

Therefore, we here restrict the distribution of $R$. coronata to southern England and the Atlantic coast of France, a limited geographical span supported by the direct development of the species (Schmekel \& Cappellato, 2001). The species $R$. aurata and $R$. caletensis, despite subtle differences, are externally difficult to distinguish and coexist in the same geographical area (Cádiz, south of Spain), whereas the species $R$. tingensis is so far only known from the north-western coast of Morocco.

This work has revealed several additional putative cases of hidden diversity among runcinids in Europe (e. g. R. adriatica and R. ferruginea), and our detailed study of the $R$. coronata species-complex has made it possible to redefine the type species of the genus, to clarify the taxonomic status of $R$. aurata and to describe two new species to science. In addition, we have provided the first modern approach to understanding relationships in the order Runcinida and a provisional framework to discuss the familial and generic classification of the group.

## ACKNOWLEDGEMENTS

We are grateful to all colleagues who provided specimens and photographs for this study, including Josep Romà, Naoufal Tamsouri, Ana Bartual, Ian F. Smith, Enric Madrenas, Miquel Pontes, Carles


Figure 10. Living animals of Runcina hornae. Specimens from Catalonia, north-eastern Spain (Mediterranean Sea). A, MNCN 15.05/88104, 1 mm in length. B, MNCN 15.05/90661, 1 mm in length. C, MNCN/AND 118954, 2 mm in length. D, MNCN 15.05/90656, 1.5 mm in length. E, MNCN 15.05/90660, 3 mm in length. F, MNCN $15.05 / 90655,3 \mathrm{~mm}$ in length. G, MNCN 15.05/90659, 2.5 mm in length. H, MNCN $15.05 / 90665,1.5 \mathrm{~mm}$ in length. I, MNCN $15.05 / 90658$, 1.5 mm in length. J, MNCN 15.05/90654, 2 mm in length. L, MNCN $15.05 / 90657$, 1 mm in length. M, MNCN $15.05 / 90662,2 \mathrm{~mm}$ in length. N, MNCN $15.05 / 90664,3 \mathrm{~mm}$ in length. O, MNCN $15.05 / 90663,1 \mathrm{~mm}$ in length. P, MNCN $15.05 / 88110,2 \mathrm{~mm}$ in length. Images A, E, G, H courtesy of Carlés Galià; images B, F, M, N, O courtesy of Marina Poddubetskaia; images C, D, I, J, L, P by Ana Karla Araujo.


Figure 11. Scanning electron micrographs of radula of Runcina hornae (MNCN 15.05/90654, 1 mm in length, Mataro, Spain). A, rachidian teeth. B, lateral teeth. Scale bars: $\mathrm{A}=5 \mu \mathrm{~m}$; $\mathrm{B}=10 \mu \mathrm{~m}$.

Galià, Marina Poddubetskaia, Alen Petani, Jakov Prkić, Cory Pittman and Jorge Antonio Domíguez Godino. We thank Bastian Brenzinger, Luis Ricardo Simone, Jon Anders Kongsrud and Melanie Mackenzie for lending specimens and processing loans of the ZSM, MZUSP, ZMBN and Museum Victoria collections, respectively. We also thank Juan González (SC-ICYT, UCA) for assistance with SEM. This study was supported by several research projects, 'Anfiatlantismo, alopatría y espécies crípticas en moluscos opistobranquios, con especial atención a los de la región Ibero-Balear-Macaronésica' funded by Ministry of Education and Science (CGL200605182/BOS) to J. L. Cervera, 'Diversidad de moluscos opistobranquios en Bermuda y biogeografía del Océano Atlántico' funded by Ministry of Science and Innovation (CTM2008-05228-E/MAR) to J. L. Cervera, 'Progess in Mediterranean Sea Slugs biodiversity' ASSEMBLE (7th FP) grant agreement no. 22779 to J. L. Cervera, 'La Caleta (Cádiz) e intermareales rocosos asociados: una ventana permanente al conocimiento de la biodiversidad marina' funded by Fundación Biodiversidad of the Ministry for
the Ecological Transition and 'Desentrañando la diversidad criptica en las regions Lusitánica y Mediterránea: Heterobranquios marinos (Mollusca), Sílidos (Annelida) y Caprélidos (Arthropoda, Pancrustacea) como casos de estudio' funded by the University of Cadiz (PR2018-039) to J. L. Cervera. A. K. Araujo holds a PhD grant from the Conselho Nacional de Desenvolvimento Científico e Tecnológico - CNPq/Brazil (Processo 205276/2014-8).

## REFERENCES

Akaike H. 1974. A new look at the statistical model identification. IEEE Transactions on Automatic Control 19: 716-723.
Alder J, Hancock A. 1846. Notices of some new and rare British species of naked Mollusca. Annals and Magazine of Natural History 18: 289-294.
Alfaro ME, Zoller S, Lutzoni F. 2003. Bayes or bootstraps? A simulation study comparing the performance of Bayesian Markov chain Monte Carlo sampling and bootstrapping in assessing phylogenetic confidence. Molecular Biology and Evolution 20: 255-266.
Araujo AK, Pola M, Malaquias MAE, Cervera JL. 2019. To be or not to be? What molecules say about Runcina brenkoae Thompson, 1980 (Gastropoda: Heretobranchia: Runcinida). Scientia Marina 83: 223-235.
Austin J, Gosliner T, Malaquias MAE. 2018. Systematic revision, diversity patterns, and trophic ecology of the tropical Indo-West Pacific sea slug genus Phanerophthalmus A.Adams, 1850 (Cephalaspidea, Haminoeidae). Invertebrate Systematics 32: 1336-1387.
Baba K. 1937. Opisthobranchia of Japan (I). Journal of the Department of Agriculture Kyûshû Imperial University 5: 195-236.
Baba K. 1967. Supplementary notes on the anatomy of Metaruncina setoensis (Baba, 1954), (N.G.) (Opisthobranchia, Cephalaspidea). Publications of the Seto Marine Biological Laboratory 15: 185-197.
Ballesteros M, Madrenas E, Pontes M. 2016. Actualización del catálogo de los moluscos opistobranquios (Gastropoda: Heterobranchia) de las costas catalanas. Spira 6: 1-28.
Bouchet P, Rocroi JP, Hausdorf B, Kaim A, Kano Y, Nützel A, Parkhaev P, Schrödl M, Strong EE. 2017. Revised classification, Nomenclator and Typification of Gastropod and Monoplacophoran Families. Malacologia 61: 1-256.
Burn R. 1963. Australian Runcinacea (Mollusca, Gastropoda). Australian Zoologist 13: 9-22.
Burn R, Thompson TE. 1998. Order Cephalaspidea. In: Beesley PL, Ross GJB, Wells A, eds. Mollusca: the southern synthesis. Fauna of Australia, Vol. 5. Melboure: CSIRO Publishing, Part B viii, pp. 565-1234.
Carmona L, Pola M, Gosliner TM, Cervera JL. 2015. Burnaia Miller, 2001 (Gastropoda, Heterobranchia, Nudibranchia): a facelinid genus with an Aeolidiidae's outward appearance. Helgoland Marine Research 69: 285-291.

Cervera JL, García-Gómez JL, García FJ. 1991. The genus Runcina Forbes \& Haley, 1851 (Opisthobranchia: Cephalaspidea) in the Strait of Gibraltar, with the description of a new species from the bay of Algeciras. Journal of Molluscan Studies 57: 199-208.
Cervera JL, Calado G, Gavaia C, Malaquias MAE, Templado J, Ballesteros M, García-Gómez JC, Megina C. 2004. An annotated and updated checklist of the opisthobranchs (Mollusca: Gastropoda) from Spain and Portugal (including islands and archipelagos). Boletín del Instituto Español de Oceanografía 20: 5-111.
Chernyshev AV. 1998. Runcinida marisae sp. nov. (Opisthobranchia, Runcinidae), a new runcinid mollusc from the Japan Sea. Ruthenica 8: 29-32.
Chernyshev AV. 2006. New data on molluesks of the Family Runcinidae (Gastropoda: Opisthobranchia) from Russian Far Eastern seas. Bulletin of the Russian Far East Malacological Society 10: 122-125.
Clark KB. 1984. New records and synonymies of Bermuda opisthobranchs (Gastropoda). The Nautilus 98: 85-97.
Colgan D, McLauchlan A, Wilson GDF, Livingston SP, Edgecombe GD. 1998. Histone H3 and U2 snRNA DNA sequences and arthropod molecular evolution. Australian Journal of Zoology 46: 419-437.
Colosi G. 1915. Osservazioni anatomo-istologiche sulla Runcina calaritana n. sp. Memorie della Reale Accademia delle Scienze di Torino 2, LXVI: 1-35.
Darriba D, Taboada GL, Doallo R, Posada D. 2012. jModelTest2: more models, new heuristics and parallel computing. Nature Methods 9: 696-704.
Dayrat B, Tillier S. 2002. Evolutionary relationships of euthyneuran gastropods (Mollusca): a cladistic re-evaluation of morphological characters. Zoological Journal of the Linnean Society 135: 403-470.
Dayrat B, Tillier A, Lecointre G, Tillie S. 2001. New clades of Euthyneura gastropods (Mollusca) from 28S rRNA sequences. Molecular Phylogenetics and Evolution 19: 225-235.
Folmer RO, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294-299.
Forbes E, Hanley S. 1851. A history of British Mollusca and their shells, 3. London: Van Voorst, 616.
García JC, López CM, Luque AA, Cervera JL. 1986. Descripción comparativa de Runcina aurata n. sp. Y R. coronata (Quatrefages, 1844) (Gastropoda: Opisthobranchia). Cahiers de Biologie Marine 27: 457-468.
Gosliner T. 1990. Opisthobranch Molluses from the Azores Islands. I. Runcinidae and Chromodorididae. Açoreana Supplement 135-166.
Gosliner TM. 1991. Four new species and a new genus of opisthobranch gastropods from the Pacific coast of North America. The Veliger 34: 272-290.
Grande C, Templado J, Cervera JL, Zardoya R. 2004a. Phylogenetic relationships among Opisthobranchia (Mollusca: Gastropoda) based on mitochondrial cox 1, trnV,
and rrnL genes. Molecular Phylogenetics and Evolution 33: 378-388.
Grande C, Templado J, Cervera JL, Zardoya R. 2004b. Molecular phylogeny of Euthyneura (Mollusca: Gastropoda). Molecular Biology and Evolution 21: 303-313.
Guindon S, Gascuel O. 2003. A simple, fast, and accurate method to estimate large phylogenies by maximumlikelihood. Systematic Biology 52: 696-704.
Hillis DM, Bull JJ. 1993. An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Systematic Biology 42: 182-192.
Jörger KM, Schrödl M. 2013. How to describe a cryptic species? Practical challenges of molecular taxonomy. Frontiers in Zoology 10: 1-27.
Jörger KM, Stöger I, Kano Y, Fukuda H, Knebelsberger T, Schrödl M. 2010. On the origin of Acochlidia and other enigmatic euthyneuran gastropods, with implications for the systematics of Heterobranchia. BMC Evolutionary Biology 10: 323.
Katoh K, Asimenos G, Toh H. 2009. Multiple alignment of DNA sequences with MAFFT. Methods in Molecular Biology 537: 39-64.
Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28: 1647-1649. doi:10.1093/ bioinformatics/bts199
Korshunova T, Martynov A, Bakken T, Picton B. 2017. External diversity is restrained by internal conservatism: new nudibranch mollusc contributes to the cryptic species problem. Zoologica Scripta 46: 683-692.
Kress A. 1977. Runcina ferruginea n. sp. (Opisthobranchia: Cephalaspidea), a new runcinid from Great Britain. Journal of the Marine Biological Association of the United Kingdom 57: 201-211.
Krug PJ, Vendetti JE, Valdés A. 2016. Molecular and morphological systematics of Elysia Risso, 1818 (Heterobranchia: Sacoglossa) from the Caribbean region. Zootaxa 4148: 001-137.
Kumar S, Stecher G, Knyaz C, Tamura K. 2018. MEGAX: Molecular Evolutionary Genetics Analysis across computing platforms. Molecular Biology and Evolution 35: 1547-1549.
Lemche H. 1967. Runcina Forbes, 1851 (Gastropoda): validated under the plenary powers. Opinion 811. Bulletin of Zoological Nomenclature 24: 89-90.
Malaquias MAE, Mackenzie-Dodds J, Bouchet P, Gosliner T, Reid DG. 2009. A molecular phylogeny of the Cephalaspidea sensu lato (Gastropoda: Euthyneura): Architectibranchia redefined and Runcinacea reinstated. Zoologica Scripta 38: 23-41.
Marcus Er, Marcus Ev. 1963. Opisthobranchs from Lesser Antilles. Studies on the Fauna of Curaçao and other Caribbean Islands 19: 1-76.
Marcus Er, Marcus Ev. 1970. Opisthobranchs from Curaçao and Faunistically related regions. Studies on the Fauna of Curaçao and other Caribbean Islands 122: 1-129.

Mikkelsen PM. 1993. Monophyly versus the Cephalaspidea (Gastropoda, Opisthobranchia) with an analysis of traditional cephalaspid characters. Bolletino Malacologico 29: 115-138.
Mikkelsen PM. 1996. The evolutionary relationships of Cephalaspidea s.1. (Gastropoda: Opisthobranchia): a phylogenetic analysis. Malacologia 37: 375-442.
Millard V. 1997. Classification of Mollusca. A classification of worldwide Mollusca. Pretoria: published by the author, 544.
Miller MC, Rudman WB. 1968. Two new genera and species of the superfamily Runcinoidea (Mollusca: Gastropoda: Opisthobranchia). Transactions of the Royal Society of New Zealand (Zoology) 10: 183-189.
Moro L, Ortea J. 2015. Nuevos taxones de babosas marinas de las Islas Canarias y de Cabo Verde (Mollusca: Heterobranchia). Vieraea 43: 21-86.
Odhner N. 1939. Opisthobranchia Mollusca from the western and northern coasts of Norway. Kongelige Norske Videnskabers Selskabs Skrifter 1: 1-93.
Odhner NH. 1968. Sous-classe des Opisthobranches. In: Franc A, Grassé PP, eds. Traité de Zoologie 5(3). Paris: Masson, 608-893.
Ortea J.2013. Descripción de un nuevo runcináceo (Gastropoda: Opisthobranchia) de las costas de Ghana, África Occidental. Revista de la Academia Canaria de Ciencias XXV: 9-14.
Ortea J, Moro L, Espinosa J. 2017. El género Lapinura (Marcus \& Marcus, 1963) (Mollusca: Runcinacea) en el Atlántico, con la descripción de nuevas especies de las islas de Cabo Verde y Costa Rica. Avicennia 21: 11-18.
Oskars TR, Bouchet P, Malaquias MAE. 2015. A new phylogeny of the Cephalaspidea (Gastropoda: Heterobranchia) based on expanded taxon sampling and gene markers. Molecular Phylogenetics and Evolution 89: 130-150
Padula V, Araújo AK, Matthews-Cascon H, Schrödl M. 2014. Is the Mediterranean nudibranch Cratena peregrina (Gmelin, 1791) present on the Brazilian coast? Integrative species delimination and description of Cratena minor n. sp. Journal of Molluscan Studies 80: 575-584.
Palumbi SR, Martin A, Roman S, Mcmillan W, Stice L, Grabowski G. 1991. The simple fools' guide to PCR. Department of Zoology and Kewalo Laboratory. Available from: palumbi.stanford.edu/SimpleFoolsMaster.pdf.
Pruvot-Fol A. 1954. Mollusques Opisthobranches. Faune de France 58: 460.
Puillandre N, Lambert A, Brouillet S, Achaz G. 2012. ABGD, Automatic Barcode Gap Discovery for primary species delimitation. Molecular Ecology 21: 1864-1877.
Quatrefages De J. 1844. Mémoire sur les Gastéropodes phlébentérés, ordre nouveau de la classe des gastéropodes. Annales des Sciences Naturelles (Zoologie), Serie 3, 1: 129-183.
Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. Systematic Biology 67: 901-904
Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572-1574.

Sánchez-Moyano JE, Estacio FJ, García-Adiego EM, García-Gómez JC. 2000. The molluscan epifauna of the alga Halopteris scorparia in southern Spain as a bioindicator of coastal environmental conditions. Journal of Molluscan Studies 66: 431-448.
Schmekel L. 1985. Aspects of evolution within the opisthobranchs. In: Trueman ER, Clarke MR, eds. The Mollusca. Evolution 10. Orlando: New York: Academic Press, 221-267.
Schmekel L, Cappellato D. 2001. Contributions to the Runcinidae. I. Six new species of thegenus Runcina (Opisthobranchia Cephalaspidea) in the Mediterranean. Vie et Milieu 51: 141-160.
Schmekel L, Cappellato D. 2002. Contributions to the Runcinidae. II. Three new species and comparative studies on five stablished species of Runcina (Opisthobranchia Cephalaspidea) in the Mediterranean. Vie et Milieu 52: 85-102.
Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post- analysis of large phylogenies. Bioinformatics 30: 1312-1313.
Talavera G, Castresana J. 2007. Improvement of phylogenies after removing divergent andambiguously aligned blocks from protein sequence alignments. Systematic Biology 56: 564-577.
Templado J. 1984. Moluscos de las praderas de Posidonia oceanica en las costas del Cabo de Palos (Murcia). Investigación Pesquera 48: 509-526.
Thompson TE. 1976. Biology of Opisthobranch Molluscs, Vol I. London: The Ray Society, 207.

Thompson TE. 1977. Jamaican opisthobranch molluses I. Journal of Molluscan Studies 43: 93-140.
Thompson TE, Brodie G. 1988. Eastern Mediterranean Opisthobranchia: Runcinidae (Runcinacea), with a review of Runcinid classification and a description of a new species from Fiji. Journal of Molluscan Studies 54: 339-346.
Vaught KC. 1989. A classification of the living Mollusca. Melbourne (FL): American Malacologists, 189.
Vayssière A. 1883. Les genres Pelta (Runcina) et Tylodina. Annales des Sciences Naturelles (Zoologie), Serie 6, 15: 1-46.
Vonnemann V, Schrödl M, Klussmann-Kolb A, Wägele H. 2005. Reconstruction of the phylogeny of the Opisthobranchia (Mollusca: Gastropoda) by means of 18 S and 28S rRNA gene sequences. Journal of Molluscan Studies 71: 113-125.
Wägele H, Klussmann-Kolb A. 2005. Opisthobranchia (Mollusca, Gastropoda) - more than just slimy slugs. Shell reduction and its implications on defence and foraging. Frontiers in Zoology 2: 1-18.
Wägele H, Klussmann-Kolb A, Verbeek E, Schröld M. 2014. Flashback and foreshadowing- a review of the taxon Opisthobranchia. Organisms Diversity \& Evolution 14: 133-149.
Zhang J, Kapli P, Pavlidis P, Stamatakis A. 2013. A general species delimitation method with applications to phylogenetic placements. Bioinformatics 29: 2869-287.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.
Figure S1. Phylogenetic hypothesis based on BI of the 16S gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.
Figure S2. Phylogenetic hypothesis based on BI of the COI gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.
Figure S3. Phylogenetic hypothesis based on BI of the $H 3$ gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.

