

African Journal of Plant Science

Volume 8 Number 7, July 2014

ISSN 1996-0824



*Academic
Journals*

ABOUT AJPS

The **African Journal of Plant Science (AJPS)** (ISSN 1996-0824) is published Monthly (one volume per year) by Academic Journals.

African Journal of Plant Science (AJPS) provides rapid publication (monthly) of articles in all areas of Plant Science and Botany. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in AJPS are peer-reviewed.

Submission of Manuscript

Please read the **Instructions for Authors** before submitting your manuscript. The manuscript files should be given the last name of the first author

[Click here to Submit manuscripts online](#)

If you have any difficulty using the online submission system, kindly submit via this email ajps@academicjournals.org.

With questions or concerns, please contact the Editorial Office at ajps@academicjournals.org.

Editor

Prof. Amarendra Narayan Misra

*Center for Life Sciences, School of Natural Sciences,
Central University of Jharkhand,
Ratu-Lohardaga Road, P.O. Brambe-835205,
Ranchi, Jharkhand State,
India.*

Dr. Khaled Nabih Rashed

*Pharmacognosy Dept.,
National Research Centre,
Dokki, Giza, Egypt*

Associate Editors

Dr. Ömür Baysal

*Assoc. Prof.
Head of Molecular Biology and Genetic Department,
Faculty of Life Sciences,
Mugla Sıtkı Koçman University,
48000 -Mugla / TURKEY.*

Dr. Biswa Ranjan Acharya

*Pennsylvania State University
Department of Biology
208 Mueller Lab
University Park, PA 16802.
USA*

Dr. Pingli Lu

*Department of Biology
416 Life Sciences Building
Huck Institutes of the Life Sciences
The Pennsylvania State University
University Park, PA 16802
USA.*

Prof. H. Özkan Sivritepe

*Department of Horticulture Faculty of
Agriculture Uludag University Görükle
Campus Bursa 16059
Turkey.*

Dr. Nafees A. Khan

*Department of Botany
Aligarh Muslim University
ALIGARH-202002, INDIA.*

Prof. Ahmad Kamel Hegazy

*Department of Botany, Faculty of Science,
Cairo University, Giza 12613,
Egypt.*

Dr. Manomita Patra

*Department of Chemistry,
University of Nevada Las Vegas, Las Vegas,
NV 89154-4003.*

Dr. Annamalai Muthusamy

*Department of Biotechnology
Manipal Life Science Centre,
Manipal University,
Manipal – 576 104
Karnataka,
India.*

Dr. R. Siva

*School of Bio Sciences and Technology
VIT University
Vellore 632 014.*

Dr. Chandra Prakash Kala

*Indian Institute of Forest Management
Nehru Nagar, P.B.No. 357
Bhopal, Madhya Pradesh
India – 462 003.*

Instructions for Author

Electronic submission of manuscripts is strongly encouraged, provided that the text, tables, and figures are included in a single Microsoft Word file (preferably in Arial font).

The **cover letter** should include the corresponding author's full address and telephone/fax numbers and should be in an e-mail message sent to the Editor, with the file, whose name should begin with the first author's surname, as an attachment.

Article Types

Three types of manuscripts may be submitted:

Regular articles: These should describe new and carefully confirmed findings, and experimental procedures should be given in sufficient detail for others to verify the work. The length of a full paper should be the minimum required to describe and interpret the work clearly.

Short Communications: A Short Communication is suitable for recording the results of complete small investigations or giving details of new models or hypotheses, innovative methods, techniques or apparatus. The style of main sections need not conform to that of full-length papers. Short communications are 2 to 4 printed pages (about 6 to 12 manuscript pages) in length.

Reviews: Submissions of reviews and perspectives covering topics of current interest are welcome and encouraged. Reviews should be concise and no longer than 4-6 printed pages (about 12 to 18 manuscript pages). Reviews are also peer-reviewed.

Review Process

All manuscripts are reviewed by an editor and members of the Editorial Board or qualified outside reviewers. Authors cannot nominate reviewers. Only reviewers randomly selected from our database with specialization in the subject area will be contacted to evaluate the manuscripts. The process will be blind review.

Decisions will be made as rapidly as possible, and the journal strives to return reviewers' comments to authors as fast as possible. The editorial board will re-review manuscripts that are accepted pending revision. It is the goal of the AJFS to publish manuscripts within weeks after submission.

Regular articles

All portions of the manuscript must be typed double-spaced and all pages numbered starting from the title page.

The Title should be a brief phrase describing the contents of the paper. The Title Page should include the authors' full names and affiliations, the name of the corresponding author along with phone, fax and E-mail information. Present addresses of authors should appear as a footnote.

The Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The Abstract should be 100 to 200 words in length. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited.

Following the abstract, about 3 to 10 key words that will provide indexing references should be listed.

A list of non-standard **Abbreviations** should be added. In general, non-standard abbreviations should be used only when the full term is very long and used often. Each abbreviation should be spelled out and introduced in parentheses the first time it is used in the text. Only recommended SI units should be used. Authors should use the solidus presentation (mg/ml). Standard abbreviations (such as ATP and DNA) need not be defined.

The Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision. The results should be written in the past tense when describing findings in the authors' experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the Results but should be put into the Discussion section.

The Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

The Acknowledgments of people, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph form or repeated in the text.

Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or Powerpoint before pasting in the Microsoft Word manuscript file. Tables should be prepared in Microsoft Word. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

References: In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's name should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lower case letter like 'a' and 'b' after the date to distinguish the works.

Examples:

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998;

1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001) References should be listed at the end of the paper in alphabetical order. Articles in preparation or articles submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text (e.g., A. Kingori, University of Nairobi, Kenya, personal communication). Journal names are abbreviated according to Chemical Abstracts. Authors are fully responsible for the accuracy of the references.

Examples:

Chikere CB, Omoni VT and Chikere BO (2008). Distribution of potential nosocomial pathogens in a hospital environment. *Afr. J. Biotechnol.* 7: 3535-3539.

Moran GJ, Amii RN, Abrahamian FM, Talan DA (2005). Methicillinresistant *Staphylococcus aureus* in community-acquired skin infections. *Emerg. Infect. Dis.* 11: 928-930.

Pitout JDD, Church DL, Gregson DB, Chow BL, McCracken M, Mulvey M, Laupland KB (2007). Molecular epidemiology of CTXM-producing *Escherichia coli* in the Calgary Health Region: emergence of CTX-M-15-producing isolates. *Antimicrob. Agents Chemother.* 51: 1281-1286.

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications.* McGraw-Hill Inc., New York, pp. 591-603.

Short Communications

Short Communications are limited to a maximum of two figures and one table. They should present a complete study that is more limited in scope than is found in full-length papers. The items of manuscript preparation listed above apply to Short Communications with the following differences: (1) Abstracts are limited to 100 words; (2) instead of a separate Materials and Methods section, experimental procedures may be incorporated into Figure Legends and Table footnotes; (3) Results and Discussion should be combined into a single section.

Proofs and Reprints: Electronic proofs will be sent (e-mail attachment) to the corresponding author as a PDF file. Page proofs are considered to be the final version of the manuscript. With the exception of typographical or minor clerical errors, no changes will be made in the manuscript at the proof stage.

Fees and Charges: Authors are required to pay a \$550 handling fee. Publication of an article in the African Journal of Plant Science is not contingent upon the author's ability to pay the charges. Neither is acceptance to pay the handling fee a guarantee that the paper will be accepted for publication. Authors may still request (in advance) that the editorial office waive some of the handling fee under special circumstances

Copyright: © 2014, Academic Journals.

All rights Reserved. In accessing this journal, you agree that you will access the contents for your own personal use but not for any commercial use. Any use and or copies of this Journal in whole or in part must include the customary bibliographic citation, including author attribution, date and article title.

Submission of a manuscript implies: that the work described has not been published before (except in the form of an abstract or as part of a published lecture, or thesis) that it is not under consideration for publication elsewhere; that if and when the manuscript is accepted for publication, the authors agree to automatic transfer of the copyright to the publisher.

Disclaimer of Warranties

In no event shall Academic Journals be liable for any special, incidental, indirect, or consequential damages of any kind arising out of or in connection with the use of the articles or other material derived from the AJPS, whether or not advised of the possibility of damage, and on any theory of liability.

This publication is provided "as is" without warranty of any kind, either expressed or implied, including, but not limited to, the implied warranties of merchantability, fitness for a particular purpose, or non-infringement. Descriptions of, or references to, products or publications does not imply endorsement of that product or publication. While every effort is made by Academic Journals to see that no inaccurate or misleading data, opinion or statements appear in this publication, they wish to make it clear that the data and opinions appearing in the articles and advertisements herein are the responsibility of the contributor or advertiser concerned. Academic Journals makes no warranty of any kind, either express or implied, regarding the quality, accuracy, availability, or validity of the data or information in this publication or of any other publication to which it may be linked.

African Journal of Plant Science

Table of Content: Volume 8 Number 7, July 2014

ARTICLES

Drought stress and its intensity, the factor of strategies selection for drought tolerance in *Haloxylon aphyllum*

Naser Arabzadeh

Chemical profiles as chemotaxonomic tools for Loranthaceae in Nigeria

Jemilat Aliyu Ibrahim, Henry Omoreige Egharevba, Ibrahim Iliya, Florence Tarfa and Abiodun Emmanuel Ayodele

Efficacy of leaf extracts of some medicinal plants on growth of *Colletotrichum capsici* butler and bisby

Shinde J. U. and D. U. Gawai

Effects of crude extracts on some selected physiological parameters of French beans (*Phaseolus vulgaris*) infected with rust (*Uromyces appendiculatus*)

Menge D. M. S., Makobe M., Monda E. O. and Okemo P. O.

Fatty acid composition of the seed oil of *Chrysophyllum albidum* (G. Don)

Paul M. Osamudiamen and Lukman O. Afolabi

An ethnobotanical study of medicinal plants in Debre Libanos Wereda, Central Ethiopia

Seyoum Getaneh and Zerihun Girma

Community structure, regeneration potential and future dynamics of natural forest site in part of Nanda Devi Biosphere Reserve, Uttarakhand, India

Balwant Rawat, Sanjay Gairola, K. Chandra Sekar and R. S. Rawal

Chemical characterization and antimicrobial activity of essential oils and *Croton's* varieties modulator in the Brazilian's Northeast semiarid

Elissandra C. Angélico, Onaldo G. Rodrigues, José G. M. da Costa, Maria de Fátima A. Lucena, Vicente Queiroga Neto and Rosália Severo de Medeiros

Full Length Research Paper

Drought stress and its intensity, the factor of strategies selection for drought tolerance in *Haloxylon aphyllum*

Naser Arabzadeh

Agricultural and Natural Resources Researches Center, Sodoughi Avenue, P.O. Box 76175-538, Kerman, Iran.

Received 7 February, 2014; Accepted 16 July, 2014

Osmotic parameters of *Haloxylon aphyllum* were studied after inducing dryness. Water relations parameters with improving resistance to dryness of this species through inducing severe dryness and clarification of physiologic mechanisms of this plant in response to a low water and dryness were among the objectives of this study. For this purpose, the method of pressure chamber was employed. By this method, the pressure-volume curve was drawn and the parameters of water relations of the plant were obtained from analyzing them. A relatively mild dryness was induced to plants through a lack of irrigation. After two weeks, *Haloxylon* water potential reached -16.5 bars. A severe dryness was also induced to that but after four weeks of no irrigation, it was reduced to -27.2 bars. Relatively mild and severe dryness were repeated for six and 11 periods respectively. In both series of experiment, the control water potential that were being watered every two days once, remained fixed at about -12.7 bars. Based on the results, although the relatively mild dryness increased the elasticity of plant textures, but it had not a meaningful impact on its osmotic potential. Although the use of a relatively severe dryness decreased both osmotic potential and osmotic adjustment, but at the same time, it increased the elasticity too.

Key words: *Haloxylon*, water relations, water stress, drought resistance, osmotic potential, elasticity.

INTRODUCTION

Plants use two mechanisms which are: 1) tolerating dryness and 2) escaping from it in confronting with dryness (Turner, 1979; May and Milthorpe, 1962). *Haloxylon aphyllum* is among species which with the help of endurance mechanism is able to spend dry periods. These mechanisms have been studied in some of the species like various kinds of pine (Emadian, 1988; Grime, 1979; Bilan et al., 1979:1978; Youngman, 1965).

Haloxylon species like many other species, by osmotic

adjusting or increasing the elasticity of cellular wall in the condition of water tension maintain their turgescence better and consequently tolerate better drought periods (Emadian, 1988).

H. aphyllum like other multi-functional plants of *haloxylon* type is of great value and importance in protecting and supporting breakable ecosystems like deserts.

It is such that as a live windbreaker prevents soil

E-mail: arabzadeh_r_m@yahoo.com. Tel: +983412260531, +989131408872. Fax: +983412110395, +983412112990.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](#)

erosion (Tokasi et al., 2007; Safarnejad, 2005; Jafari et al., 2004), and as a correcting element increases the organic materials of soil and in a long term improves the structure of soil (Jafari et al., 2004), and increases the plant enrichment of the area under coverage (Bakhshi and Biroudian, 2008). In addition for those who live in deserts, it is an important source for the provision of fuel and fodder for cattle (Tokasi et al., 2007). As it has a broad emission spectrum in different soils from the viewpoint of texture (Javanshir et al., 1996), it can be noticed much more.

However, unfortunately, despite the high importance of haloxylon in biologic stabilization of sand lands, most of the planted plants in desert regions are facing the problem of dryness. Based on the confirmation of authorities and those in charge of desertification projects, after transferring the plants of *H. aphyllum* planted in vase to a natural plantation bed and despite observing principles related with post-plantation stages, after sometimes, a high percentage of transferred plants were afflicted with dryness and they had to be replanted. Taking action to replant instead of dried plants imposes relatively extravagant costs to the executive system of natural resources. According to the existing documents in the File Keeping Department of Agriculture Ministry of Iran, in average in each period, more than 10% of the vases with *H. aphyllum* were fading and became dried in the fields, and required replanting. In order to help reduce mortality percentage of planted plants in main beds of plantation, it seemed that placing one-year plants of *H. aphyllum* which were exposed to periodic dryness and possibility of their compatibility with unfavorable condition resulting reduction of rainfall and periodic droughts could be tested as an appropriate approach.

The main goal in conducting this research was firstly the study the changes of water relations parameters in order to improve resistance to dryness of *H. aphyllum* through induction of periodic dryness and also clarification of physiological mechanisms of this species in response to water shortage and dryness.

MATERIALS AND METHODS

The identified seeds of *H. aphyllum* were planted in plastic vases and were taken into care for one year. Plastic vases with an approximate capacity of three liters were selected in order to pave way for a better and a greater growth of the plants roots. The soil consisted of wind sand, soil and leaf-soil in proportions of 2, 1 and 1 respectively. Supply of necessary nutrition for plants during the experiment was done by including leaf-soil in the mentioned combination. At the same time, in order to prevent unwanted accumulation of water in vases, some fine holes were made in their bottom. After one year, the plants were transferred to a greenhouse.

Upon completion of one-month period of plants compatibility in greenhouse, the treatments of dryness induction was applied. For this purpose, a sufficient number of good and healthy plants were selected for the experiment. Half of them were considered for induction of dryness and the rest were selected as the controls that

were being watered every two days once. In this research, two series of experiments were conducted. The plants of the experiment of the first series received 6 periods of 7 to 14 days of dryness. During this period, the water potential (before sunrise) of their plant was measured every two days once by using the pressure chamber and according to Scholander method (Scholander et al., 1965). At this state, the water potential of haloxylon plants at the end of each period was reduced to -16.5 bars. The experiment second series plants, in addition to the mentioned dry periods, received five periods of dryness of 14 to 28 days too. In this series of tests, water potential of *H. aphyllum* plant was reduced at the end of each period to -27.2 bars. Induction of stress in mild (14-7 days) and severe (28-14 days) stresses were 6 and 6+5 (11) periods (replications) respectively. So every place (points) on the curve 3 is resultant 6 stress periods, and on the curve 4 is resultant 11 stress periods (repetitions).

It is worth mentioning that in order to measure the water potential of plants, separate plants were considered and in each measuring, 5 plants were cut and used. Plants were watered fully at the end of dryness period. Their before sunrise water potential in the day after irrigation was increased by -5.3 to -4.3 bars. Also, the control plants water potential was increased from -12.7 bars in both series of test to about -9.7 to -8.7 bars.

The impact of dryness induction on the elasticity and osmotic parameters of *H. aphyllum* plants became possible through an analysis of pressure-volume curves. These curves were prepared by using *Scholander method* (Figure 1).

The horizontal axis is the exit liquid volume (W_e) and its vertical axis is balancing pressure or $(\Psi_w)^{-1}$. Point E, is the place of intersection of osmotic line with W_e axis showing the rate of water which is exited under the infinite pressure from plant known as simplistic water (W_s). Point B is the place of conjunction of osmotic line with the axis of $(\Psi_w)^{-1}$ showing that Ψ_s of plant is in the condition of full turgescence.

As Figure 1 shows each pressure-volume curve has two outstanding parts: 1) curve parts which encompasses about 5 to 8 points; 2) direct part which includes 8 to 11 points. The direct part of curve was used to estimate Ψ_w , Ψ_p , & Ψ_s . According to the recommendation of Cutler et al. (1979), this action was performed by drawing a regression curve on at least 7 to 8 points of the last spots of pressure-volume curves. The overall equation of the curve is as follows:

$$(\Psi_{st})^{-1} = (\Psi_{s0})^{-1} - m \sum W_i$$

In the relations, $(\Psi_{s0})^{-1}$ is the inverse of primary osmotic potential of plant at full turgescence condition; m is the slope of regression which is under the influence of plant size and osmotic feature of texture and rate of exchange of plant water, $(\Psi_{st})^{-1}$ is the opposite of Ψ_s for t times of a pair from the data of pressure-volume and $\sum W_i$ is its corresponding accumulated quantity of exited liquid. With this assumption that the mentioned relation to be fully true in the considered range, by placing each pair of P_i and $\sum W_i$, it is possible to get the Ψ_s . The continuation of this line crosses the vertical axis in point B which specifies the inverse of osmosis potential in full turgescence $(\Psi_{s0})^{-1}$ (Tyree and Jarvis, 1982). Subsequently, the turgor potential (Ψ_p) of the plant in each point of pressure-volume curve was obtained through difference of Ψ_w and Ψ_s related to the same point. On the other hand, the correlation line cut off the horizontal axis in point E, which determines the volume of exited liquid from plant in an infinite pressure $[(\Psi_{s0})^{-1} m^{-1}]$ (Figure 1). Active osmotic water of plant ($W\Psi_s$) was calculated in form of $W_s(W_0 - W_d)^{-1}$ and its inactive osmotic water in form of $1 - W_s(W_0 - W_d)^{-1}$ (Cutler et al., 1979).

The existing data in the part of curvature of pressure-volume curve was used to estimate the average of the absolute value of elasticity ($\bar{\epsilon}$) of *H. aphyllum* (Tyree and Jarvis, 1982). To achieve

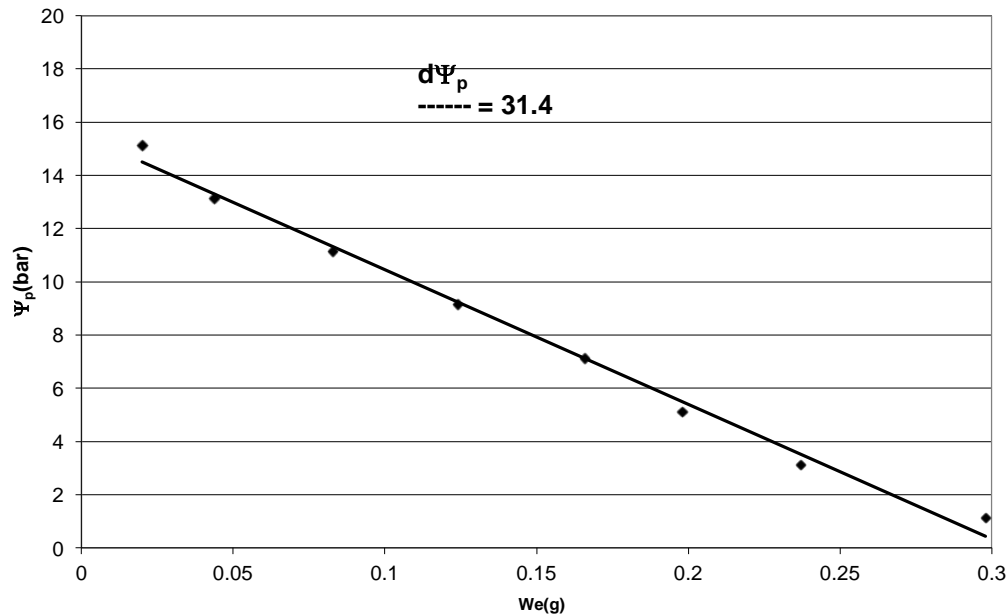


Figure 1. Pressure-volume curve of a plant of *Haloxylon aphyllum*.

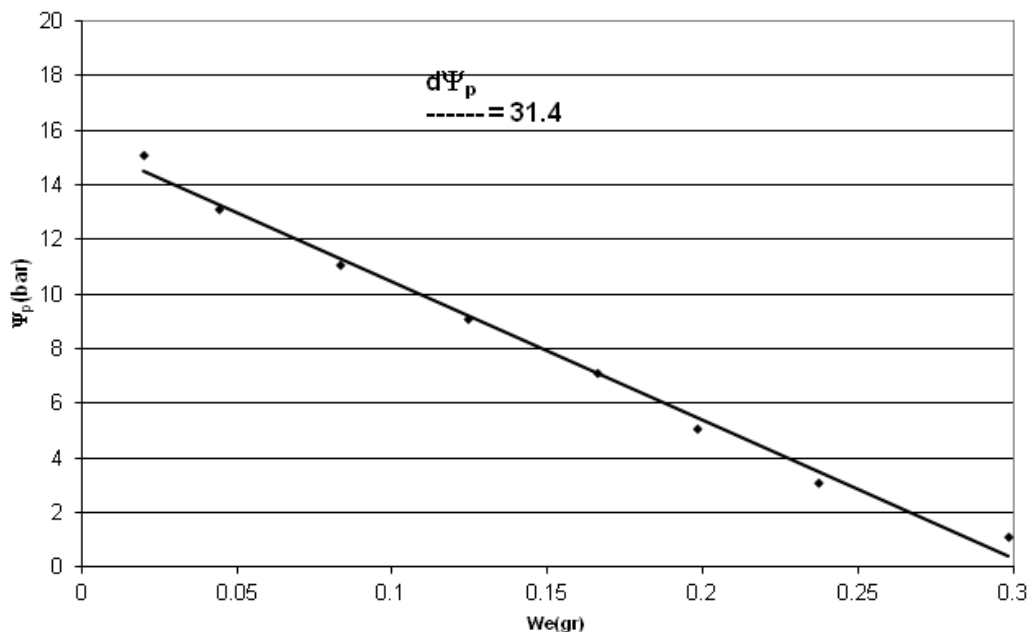


Figure 2. Calculating the rate of elasticity changes of cellular wall in *Haloxylon aphyllum* textures by using the curvature part of pressure-volume curve. Horizontal axis is exitted volume (W_e), and vertical axis is swollen potential. In order to calculate the average of absolute value of the elasticity ($\bar{\mathcal{E}}$), the curve slope of $d\Psi_p (dW_e)^{-1}$ of plant was used.

this parameter, it was necessary that firstly Ψ_p of the plant in the part of curvature of the pressure-volume curve be calculated. This action was performed by using the osmotic line and method of estimation of osmotic, turgor and water potentials. Then the obtained turgor potentials [(Ψ_p) s] in the mentioned limit with the

volume of corresponding condensed exitted liquid was drawn in coordinates sheet and their regression equation was calculated (Figure 2). The ($\bar{\mathcal{E}}$) was also calculated from the product of the slope of line [$(d\Psi_p) (dW_e)^{-1}$] in W_s .

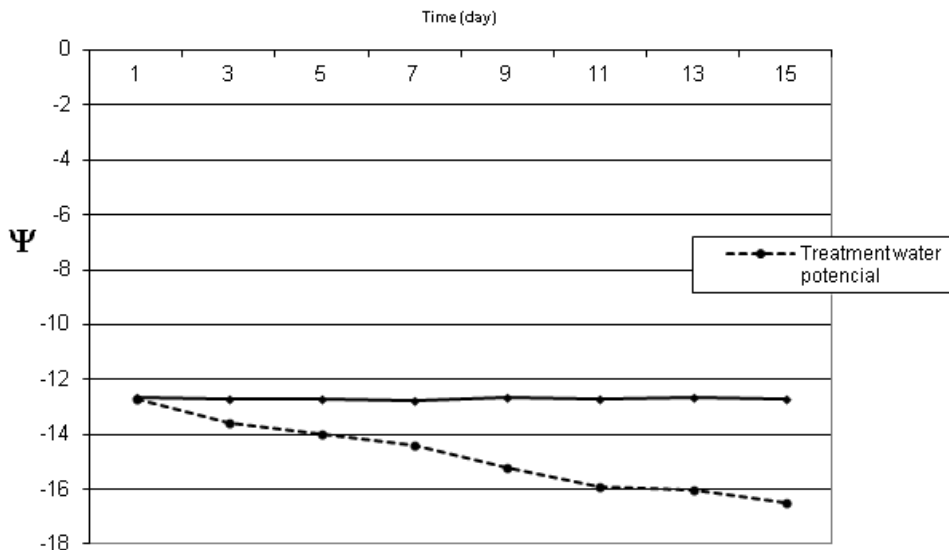


Figure 3. Relationship between plant water potential (Ψ_w) and time in *Haloxylon aphyllum* after watering. Pre-dawn plant water potential was measured every other day during two-week no-watering period.

Table 1. Comparing the result of water relations in *Haloxylon aphyllum* under mild dryness induction with control plants.

| Treatment | Ψ_{wo} (Bar) | Ψ_{po} (Bar) | Ψ_{so} (Bar) | Ψ_{wTLP} (Bar) | Ψ_{sTLP} (Bar) | $\bar{\epsilon}$ (Bar) | $W\Psi_s$ (%) |
|-----------------------|----------------------|----------------------|----------------------|------------------------|------------------------|---------------------------|------------------|
| STRESSED ¹ | -2.50** | 12.30** | -14.80 | -20.90 | -23.10 | 44.8** | 14.00** |
| CONTROL ² | -5.90 | 07.30 | -13.20 | -22.50 | -21.40 | 30.50 | 28.80 |

¹Seedlings, in addition to six early dry periods, five-day dry period 7 to 14 also received. The average water potential (before sunrise) at the end of their term limit had been reduced to -16.5 bars. Value of each parameter treated seedlings is related to seven seedlings. ²Both seedlings were irrigated once a day. Amount of each parameter of control seedlings is related to five seedlings.

$$\bar{\epsilon} = W_s [(d\Psi_p) (dW_e)^{-1}]$$

The used statistical method was t- student (Snedecor and Cochran, 1980).

RESULTS

Results of mild dryness induction

At the end of each period of mild dryness induction, water potential (Ψ_w) of *H. aphyllum* reduced about -16.5 bars; whereas Ψ_w of control remained at a higher range, that is in average -12.7 bars (Figure 3).

Parameters of active osmotic water ($W\Psi_s$) and pressure potential at the condition of moisture full saturation (Ψ_{po}) reduced and increased significantly and reached from 28.8 to 14% and was promoted from 7.3 to 12.3 bars, respectively. Their elasticity modulus ($\bar{\epsilon}$) and water potential (Ψ_{wo}) had a highly significant increase

(level 99%). Thus they accelerated from 30.5 to 44.8% and from -5.9 to -2.5 bars but the osmotic potential of tension treatments in full turgescence did not show a meaningful difference as compared with control (Table 1).

Results of severe dryness induction

After applying severe dryness induction, the water potential of *H. aphyllum* diminished in average up to about -27.2 bars, whereas the water potential of controls more or less remained fixed at the level of mild dryness induction (-12.7 bars) (Figure 4).

Water potential and pressure potential in fully saturation moisture condition increased significantly ($P < 1\%$), so that with the promotion to a higher level, it reached to the level of -2 and 18.8 bars respectively. At the same time, like the mild dryness tension, very meaningful increase of elasticity ($\bar{\epsilon}$) and very meaningful reduction of osmotic active water ($W\Psi_s$) were observed.

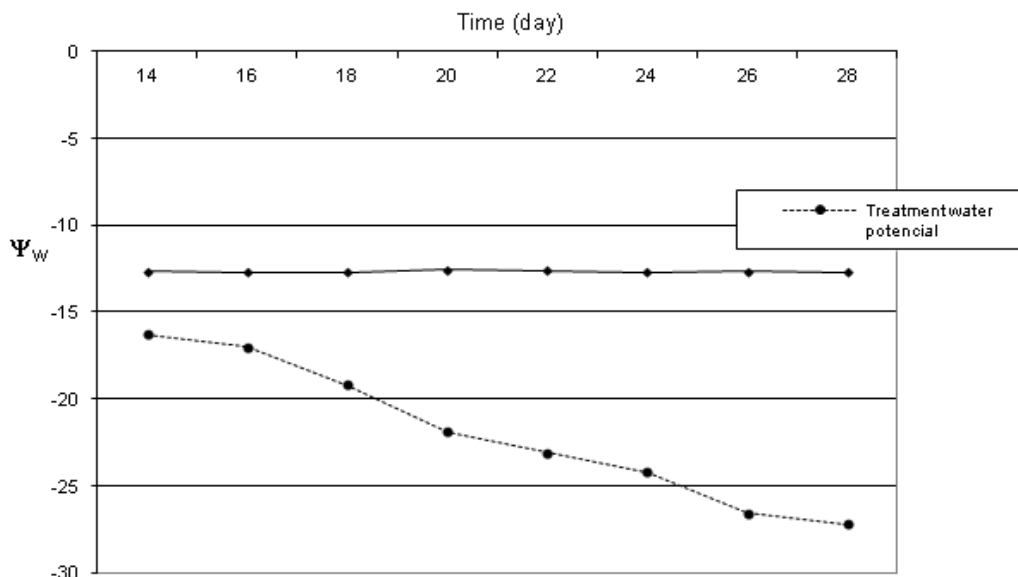


Figure 4. Relationship between plant water potential (Ψ_w) and time in *Haloxylon aphyllum* after watering. Pre-dawn plant water potential was measured every other day during a four-week period of no-watering.

However, concurrent with these changes, the osmotic potential in the condition of moisture full saturation (Ψ_{s0}) and dryness threshold (Ψ_{stlp}) reduced significantly ($P < 1\%$) and decreased from -13.5 to -20.8 bars and from -20.9 to -30.6 bars respectively.

DISCUSSION

Primarily it seemed that the reaction of parameters of water relations (like Ψ_{w0} , Ψ_{s0} , Ψ_{p0} , Ψ_{wtp} , $\bar{\varepsilon}$) of *Haloxylon aphyllum* in mild dryness condition was in conflict with their corresponding parameters in the control treatment (Table 1), because measured absolute value of elasticity ($\bar{\varepsilon}$) of under the treatment plants was 44.8 bars that was more elastic than $\bar{\varepsilon}$ of control plants (30.5 bars) significantly. On the other, the measured osmotic active water ($W\Psi_s$) in treated and controlled plants was 14 and 28.8% respectively, and this difference was meaningful in the level of 99%. Possibly, the concept of this with regard to the relation of ($\bar{\varepsilon}$) is that the more elastic plant needs a less water to maintain its turgescence. In addition, in the condition of full turgescence, the water potential of plants which were given dryness was higher than water potential of control plants.

Plant physiologists believe that many plants of dry regions are able to maintain their turgescence through mechanism of elasticity and consequently increase resistance to dryness. This feature is true for *Pinus taeda* (Emadian and Newton, 1989), *Pseudotsuga menziesii* (Joly and Zaerr, 1987), *Dubautia ciliolata* (Robichaux and

Canfield, 1985), and *Juglans nigra* (Parker and Pallardy, 1985). On the other, though the elasticity of treated plants increased, but their osmotic potential was without any meaningful change. In other words, in condition of mild dryness induction, no meaningful difference was observed at any statistical level between Ψ_{s0} and Ψ_{stlp} in control and dryness tension treatments (Table 1). With regard to mentioned condition, it seems that in the mild dryness condition, *H. aphyllum* through increase of elasticity is able to maintain its turgescence and continues its growth. Apparently, this will be possible only in lieu of an exchange with reduction of osmotic adjustment.

Water relations parameters reaction in severe stress (28-14 days without irrigation) took place in continuing mild dryness condition, and osmotic adjustment was activated (Table 2) so that in addition to increase cell wall elasticity of shoot tissues (such as effect of periodic mild stress) osmotic potential of the tissue significantly decreased compared to control tissue, and continued the life of stressed-plants. After induction, severe periodic dryness on plants (*Haloxylon aphyllum*), the increase of water and turgor potentials (300 and 250%, respectively) and reduction of osmotic potential (more than 50%) in the condition of moisture saturation and at the dryness threshold was considerable.

The results showed that the induction of periodic mild dryness was not able to have an impact on the increase of resistance to dryness in *Haloxylon aphyllum*. It only changed the strategy of plant in confronting with stresses resulting from drought. Instead of that, periodic induction of severe dryness could make ideal changes in osmotic parameters, and not only caused the viable and

Table 2. Comparing the result of water relations in *Haloxylon aphyllum* under severe dryness induction with control plants.

| Treatment | Ψ_{wo} (Bar) | Ψ_{po} (Bar) | Ψ_{so} (Bar) | Ψ_{wTLP} (Bar) | Ψ_{sTLP} (Bar) | ϵ (Bar) | $W\Psi_s$ (%) |
|-----------|----------------------|----------------------|----------------------|------------------------|------------------------|---------------------|------------------|
| MDI | -2.50 | 12.30 | -14.80 | -20.90 | -23.10 | 44.80 | 14.00 |
| SDI | -2.00 | 18.80 | -20.80 | -20.30 | -30.60 | 44.50 | 14.50 |

Table 3. Comparing the result of water relations in *Haloxylon aphyllum* under two water regimes: mild dryness induction and severe dryness induction.

| Treatment | Ψ_{wo} (Bar) | Ψ_{po} (Bar) | Ψ_{so} (Bar) | Ψ_{wTLP} (Bar) | Ψ_{sTLP} (Bar) | ϵ (Bar) | $W\Psi_s$ (%) |
|-----------|----------------------|----------------------|----------------------|------------------------|------------------------|---------------------|------------------|
| Stressed | -2.00** | 18.80** | -20.80** | -20.30** | -30.60** | 44.50** | 14.50** |
| Control | -5.80 | 07.50 | -13.50 | -29.70 | -20.90 | 31.80 | 26.50 |

¹Seedlings, in addition to six early dry periods, five-day dry period 14 to 28 also received. The average water potential (before sunrise) at the end of their term limit had been reduced to -27.2 bars. Value of each parameter treated seedlings is related to seven seedlings. ²Both seedlings were irrigated once a day. Amount of each parameter of control seedlings is related to five seedlings.

freshness of plant but also enabled plants to bear the severe condition of dryness and could maintain their water potential at a very high level. Comparing the parameters of water relations of *H. aphyllum* in mild dryness with severe dryness condition (Table 3) confirm the importance of the mentioned subject matter.

Mild dryness induction and severe dryness induction

Though under severe and longer dryness induction, cellular wall elasticity of *H. aphyllum* was maintained, however, osmotic adjustment became active for maintaining cellular turgescence. Therefore plants remained alive and continued their physiological and biochemical activities. Reduction of osmotic potential in the condition of saturation (by 6 bars) and reduce osmotic potential at threshold of wilt and dryness (by 7.5 bars) along with compatibility of plants to frequent dryness tension may enable them (compared to plants that were not exposed to any intensity of drought) tolerate drought periods resulting from factors affecting moisture shortage, and may be protected from the deleterious effects of drought periods in the same range of drought that seedlings similar to control wilt.

This idea can be put forth as the symbol of increase of resistance to dryness in *H. aphyllum* (which had received and accustomed to severe periodic dryness stresses) and to be used as a base for future research. Anyway, a decisive comment on this issue demands further studies and supplementary studies.

Conclusion

The results of this research showed that *H. aphyllum*

tolerates dryness well. If dryness be mild and its period be short, this species will be able to maintain its turgescence and continue its life by increasing elasticity mechanism. Apparently, this action is possible only in lieu of an exchange with reduction of osmotic adjustment. On the other hand, if dryness be severe and its period be long, for keeping its turgescence, in addition to increase elasticity, it uses the mechanism of osmotic adjustment too. In each of the two mentioned condition, maintaining turgescence in dryness condition cause the continuation of physiologic and biochemical activities of this species and make its growth and viability possible despite excessive and long dryness. Based on the results of this research, the executive officials of forestry and production of plants departments recommend that if they use *Haloxylon aphyllum* for biologic stability of sand lands in arid and semi-arid zones, prior to transfer of plants to main field, they should place them under five to eight periods of dryness induction for at least three to four weeks.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to Organization of Researches, Education and Expansion of Agriculture for supplying me with necessary credits and funds to implement this project. I also wish to thank Dr. Ramezanali Khavari-Nejad, Dr. Hossein Heidari Sharifabad and Dr. Fazlollah Emadian for their consultation in developing and implementing this research project.

REFERENCES

- Bilan MV, Hagan CT, Carter HB (1979). Stomata opening, transpiration, and needle moisture in loblolly pine seedlings from two Texas seed sources. *For. Sci.* 23:457-462.
- Bilan MV, Leach H, Davies G (1978). Root development in loblolly pine (*Pinus taeda*) from two Texas seed sources. In root from of planted trees (E. Van Eerden and J. M. Kinghorn, eds.) Joint Rep., No*, P. 17-22. Brit Colum Ministry of Forests/ Can. For Serv. Victoria, Brit Colum.
- Bakhshi J, Biroudian N (2008). A study on the effect of *Haloxylon aphyllum* plantation on richness and similarity understory species in Ardestan area. *Pajouhesh & Sazandegi*, 79:2-10. <http://www.magiran.com/viewpdf.asp?no=1>.
- Cutler JM, Shahan KW, Steponkus PL (1979). Characterization of internal water relations of rice by a pressure-volume method. *Crop Sci.* 19:681-984.
- Emadian SF (1988). Physiological responses of loblolly pine to silicon and water stress. Ph. D. Dissertation. Texas A&M University. Colleger Station, Texas, U.S. A.
- Emadian SF, Newton RG (1989). Growth enhancement of Loblolly Pine, *J. Plant Physiol.* 134:98-103.
- Grime JP (1979). *Plant strategies and vegetation processes*. Wiley, New York.
- Jafari M, Azarnivand H, Tavakoli H, Zehtabian G, Esmailzadeh H (2004). Investigation on different vegetation effects on sand dunes stabilization and improvement in Kashan. *Pajouhesh & Sazandegi*, 64:16-21. http://www.sid.ir/fa/VEWSSID/J_pdf/560138364m03.pdf
- Javanshir K, Dastmalchi H, Emarati A (1996). An ecologic survey of *Haloxylon*, *Populus* and *Tamarix* species in Iran deserts. The Second International Congress of Desertification, 1996, Iran. *Jehade Sazandegi*, 550pp, P. 1 <http://idochp2.irandoc.ac.ir/fulltextmanager/fulltext15/SE/38/38803.pdf>
- Joly RJ, Zaerr JB (1987). Alternation of cell wall elasticity in Douglas-fir during periods of water deficit. *J. Plant Physiol.* 83:418-422.
- May LH, Milthorpe FL (1962). Drought resistance of crop plants. *Field Crop Abst.*, 15:171-179.
- Parker WC, Pallardy SG (1985). Genotypic variation in tissue water relations of leaves and roots of black walnut (*Juglans nigra*) seedlings. *Physiologia Plantarum J.* 64:1:105-110. <http://62.60.154.17/ebook2/16352.pdf>
- Robichaux RH, Canfield JE (1985). Tissue elastic properties of eight Hawaiian *Dubautia* species that differ in habitat and diploid chromosome number. *Oecologia.* 66:77-80.
- Safarnejad A (2005). Comparison of saxaul species (*Haloxylon* spp) for its improvement and expansion in desert areas. *Pajouhesh & Sazandegi*, 67:51-57. http://www.sid.ir/fa/VEWSSID/J_pdf/5601384MT07.pdf
- Scholander PF, Hammel HT, Brad ED, Hemmingsen EA (1965). Sap pressure in the vascular plants. *Crop Sci.* 148:339-346.
- Snedecor GW, Cochran WG (1980). *Statistical Methods*, 7th ed., The Iowa State University Press, Ames, Iowa.
- Turner NC (1979). Drought resistance and adaptation to water deficit in crop plants. In *Stress Physiology in crop plants* (H. Mussell and R.C. Staples, eds.), pp. 87-103, John Wiley and Sons, N.Y.
- Tyree MT, Jarvis PG (1982). Water in tissues and cells. In *Physiology and Ecology II. Water relations and carbon assimilation* (O. L. Lange, P.S. Nobel, C.B. Osmond and H. Zigler, Eds). 12B:35-77. Springer-Verlag, Berlin.
- Youngman AL (1965). An ecotypic differentiation approach to the study of isolated populations of *Pinus taeda* in south central Texas, Ph. D. Thesis, Univ. Texas, Austin: Diss Abst. Int. B. 27, 3006.

Full Length Research Paper

Chemical profiles as chemotaxonomic tools for Loranthaceae in Nigeria

Jemilat Aliyu Ibrahim^{1*}, Henry Omoreige Egharevba¹, Ibrahim Iliya², Florence Tarfa³ and Abiodun Emmanuel Ayodele⁴

¹Department of Medicinal Plant Research and Traditional Medicine, National Institute for Pharmaceutical Research and Development, PMB 21, Garki, Abuja, Nigeria.

²Department of Pharmacy, University of Maiduguri, Maiduguri, Borno State, Nigeria.

³Department of Medicinal Chemistry and Quality Control, National Institute for Pharmaceutical Research and Development, PMB 21, Garki, Abuja, Nigeria.

⁴Department of Botany, University of Ibadan, Ibadan, Oyo State, Nigeria.

Received 10 February, 2014; Accepted 26 June, 2014

The Loranthaceae species are widespread throughout most regions of the world, and are used for various medicinal and ethnopharmacological purposes. However, the species vary in their pharmacological activity, sometimes in correlation with the species from same ecological region or host plant, due to variation in the chemical profiles. This has led to great emphasis on caution in identification and collection for use. The wide array of secondary metabolites in Loranthaceae species are believed to be of chemotaxonomic importance. In this study, the leaves of seven Nigeria species from different ecological locations were screened for the profiles of their secondary metabolites with a view towards establishing chemotaxonomic significance. The results show the complete absence of alkaloid from all the species. Over 80% of the species tested positive for balsam, flavonoids and phenols, more than 70% tested positive for tannins, 60% for saponins and about 50% tested positive for glycosides and volatile oils. Resins, phlobatannin, terpenes, sterols and anthraquinones were present in less than 50% of the species. Some metabolites were completely absent in one or more species. The patterns displayed could be of chemotaxonomic importance for Loranthaceae in Nigeria.

Key words: Loranthaceae, chemotaxonomy, secondary metabolites, Nigeria.

INTRODUCTION

Mistletoes are widespread throughout Africa, North America, Asia, Europe, Australia and Malaesia, with the American mistletoe (*Phoradendron serotinum*) and the

European mistletoe (*Viscum album*) particularly well known. Different species growing on different hosts may synthesize toxic compounds and protein such as lectins

*Corresponding author. E-mail: sadiqoyene@yahoo.com. Tel: +2348058293853.

and alkaloids with varying pharmacological activities (Preston et al., 2010). Thus, both the mistletoe and its host have shared responsibility in determining the pharmacological activity of the species. The distributions of these compounds or metabolites in different parts of a plant also vary (Preston et al., 2010). These pharmacological effects are due to variation in the chemical profiles especially the profiles of secondary metabolites. Secondary metabolites are proteins, glycosides, phenolics, steroids, saponins, terpenes, alkaloids and other chemical substances. Takhtajan (1973) suggested that secondary metabolites are compounds that may have taxonomic relevance.

Eighty percent or more of the world's population is estimated to depend primarily on traditional medicine for the treatment of ailments (Cunningham, 1993), and as a matter of fact, the use of medicinal plants is the main means of treatment by traditional healers. Also, many useful compounds, which are today used for treatment of life threatening diseases, were isolated from medicinal plants e.g. Artemisinin from *Artemisia annua* L. and Vincristine from *Catharantus roseus* (L.) G. Don (Dana, 2012; Aslam et al., 2010). The Loranthaceae, a parasitic family with mistletoes members are often considered useful as medicinal plants. It has been documented that mistletoes have immeasurable medicinal and traditional uses (Burkil, 1995; Erturk et al., 2003). The biological activities of immunomodulatory and antitumor effect of some mistletoe may be attributed to the presence of metabolites like lectins, viscotoxins and alkaloids found in the parasites (Stirpe et al., 1982; Bussing et al., 1996; Fernandez et al., 1998; Stein et al., 1999; Mengs et al., 2002).

The wide array of secondary metabolites in Loranthaceae sp. is believed to be of chemotaxonomic importance. Chemotaxonomic studies of 12 Loranthaceae and Viscaceae species namely, *Viscum rotundifolium* L.f., *Viscum capensis* L.f., *Viscum combreticola* Engl., *Viscum obovatum* Harv., *Viscum obscurum* Thunb., *Viscum verrucosum* Harv., *Loranthus dregei* Eckl. & Zehy., *Loranthus minor* Sprague, *Loranthus oleifolius* (Wendl.) Cham. & Schldl., *Loranthus rubromarginatus* Engl., *L. zeyheri* Harv. and *Loranthus* sp. were carried out in South Africa (Tilney and Lubke, 1974), and chlorogenic acid was found in all the species. Gedalovich-Shedletzky et al. (1989) analyzed and compared the chemical composition of viscin mucilage from three mistletoe species. Chemical analyses of different extracts from *Agelanthus dodoneifolius* yielded components such as triterpenes, sterols, carotenoides, saponosides, anthracenosides, anthocyanosides and tannins (Traoré, 2000). However, chemotaxonomic information on the West African or Nigerian species is unavailable. To clarify the status of Loranthaceae in the region, a revision of the Nigerian species was carried out recently and about 15 species were documented for the region (Ibrahim and Ayodele, 2011). This study aimed to

determine the profile of some basic secondary metabolites in the Nigerian species, which could be of chemotaxonomic significance.

MATERIALS AND METHODS

All reagents used were of analytical grade and were purchased from Zayo-Sigma Abuja, Nigeria. TLC plates used were also from the same source.

Plant collection and preparation

Twenty-seven specimens belonging to seven species were collected from the field through a field survey across host plant species and geographical location (Table 1). The specimens include *Agelanthus dodoneifolius* (4), *Globimetula braunii* (4), *Phragmanthera capitata* (2), *Phragmanthera nigritana* (1), *Tapinanthus bagwensis* (4), *Tapinanthus cordifolius* (4) and *Tapinanthus globiferus* (8). Vouchers specimens were deposited at the University of Ibadan Herbarium (UIH)

The leaves of each specimen were air-dried for one week at ambient temperature, and then pulverized using a mortar and pestle. The powdered leaf samples were used for the phytochemical screening and thin layer chromatographic (TLC) profiling.

Phytochemical screening

The presence of basic secondary metabolites including saponins, alkaloids, tannins, flavonoids, sterols, phenols, glycosides, resins, balsam, volatile oil, phlobatannin, terpenes and anthraquinones were determined using standard methods (Evans, 2002; Sofowora, 1993; Brain and Turner, 1975; Segelman et al., 1971).

TLC profiling

Twenty-four specimens representing seven species were examined. The specimens are: *T. globiferus* (9), *T. bangwensis* (2), *T. cordifolius* (2), *P. capitata* (2), *P. nigritana* (1), *G. braunii* (4) and *A. dodoneifolius* (4). A list of specimens and their corresponding numbers on the TLC plates are presented in Table 3.

Two grams of powdered leaf samples of each specimen were macerated in 20 ml of acetone for 24 h and filtered using filter papers. The extracts were spotted on three different pre-coated silica gel normal-phase TLC plates of dimension 12.5 by 8.5 cm. The dry spots were developed in a TLC tank of solvent system of ethylacetate : chloroform : methanol : water, in the ratio of 15:8:4:1. The developed spots were visualized by spraying the first plate with Vanillin in sulphuric acid reagent, the second plate with Gibbs reagent and the third plate with Dragendoff reagent for detection of terpenoids, phenolics and alkaloids, respectively. The retention factors (R_F values) were calculated for all the spots as distance moved by spot from the origin divided by distance moved by solvent front (Table 2).

RESULTS

Phytochemical screening

The result of the phytochemical screening for secondary

Table 1. Preliminary phytochemical screening of secondary metabolites from Loranthaceae species in Nigeria including taxa, hosts, localities, collection numbers and metabolites studied.

| Taxa | Host | Locality/No. | Gly | Rsn | Blm | Fla | Tnn | Akd | V.oil | Ptn | Spn | Tep | Str | Phn | Atq |
|---------------------------------|--------------------------------|--------------|-----|-----|-----|-----|-----|-----|-------|-----|-----|-----|-----|-----|-----|
| <i>Agelanthus dodoneifolius</i> | <i>Parkia biglobosa</i> | Jos 65 | | - | + | - | + | - | | - | + | - | - | + | ++ |
| <i>Agelanthus dodoneifolius</i> | <i>Parkia biglobosa</i> | Suleija 77 | + | - | - | + | + | - | + | - | - | - | + | + | |
| <i>Agelanthus dodoneifolius</i> | <i>Casuarina</i> sp. | Yola 119 | | + | + | + | + | - | | - | + | - | + | | - |
| <i>Agelanthus dodoneifolius</i> | <i>Vitellaria paradoxa</i> | Yola 118 | - | | + | + | + | - | - | - | + | + | - | + | |
| <i>Globimetula braunii</i> | <i>Persea americana</i> | Calabar 90 | | - | + | - | + | - | | - | + | - | - | + | - |
| <i>Globimetula braunii</i> | <i>Cola</i> sp. | Calabar 92 | | - | + | - | + | - | | - | ++ | - | - | + | + |
| <i>Globimetula braunii</i> | <i>Cola</i> sp. | Ibadan 97 | + | - | + | + | - | - | + | - | + | + | - | | - |
| <i>Globimetula braunii</i> | <i>Theobroma cacao</i> | Ibadan 102 | - | | + | + | - | - | - | - | + | + | - | - | |
| <i>Phragmanthera capitata</i> | <i>Persea americana</i> | Calabar 93 | | + | + | - | + | - | | - | + | - | - | + | + |
| <i>Phragmanthera capitata</i> | <i>Persea americana</i> | Calabar 89 | | - | + | + | + | - | - | - | - | + | - | | - |
| <i>Phragmanthera nigritana</i> | <i>Citrus</i> sp. | Chaza 78 | - | | + | + | + | - | - | - | + | + | - | + | |
| <i>Tapinanthus bangwensis</i> | <i>Newboldia leavis</i> | Ibadan 46 | | + | - | - | - | - | | - | - | - | - | | - |
| <i>Tapinanthus bangwensis</i> | <i>Citrus medica</i> | Ibadan 40 | + | - | - | + | - | - | - | - | - | - | + | - | - |
| <i>Tapinanthus bangwensis</i> | <i>Cola acuminata</i> | Ibadan | - | - | + | + | - | - | | - | + | + | + | + | - |
| <i>Tapinanthus bangwensis</i> | <i>Theobroma cacao</i> | Ibadan | - | - | + | + | - | - | | - | + | - | - | + | - |
| <i>Tapinanthus cordifolius</i> | <i>Citrus aurantifolia</i> | Jos 63 | + | + | + | + | + | - | + | - | - | - | - | + | |
| <i>Tapinanthus cordifolius</i> | <i>Cassia</i> sp. | Jos 86 | - | | + | + | + | - | - | - | + | + | - | + | |
| <i>Tapinanthus cordifolius</i> | <i>Syzygium eucalyptoide</i> | Jos | + | + | + | + | | - | + | - | + | + | + | | - |
| <i>Tapinanthus cordifolius</i> | <i>Ficus</i> sp. | Jos | + | + | | + | | - | + | + | + | + | + | | - |
| <i>Tapinanthus globiferus</i> | <i>Piliostigma thoninngii</i> | Kano 29 | | - | + | + | + | - | | - | - | - | - | + | + |
| <i>Tapinanthus globiferus</i> | <i>Azadirachta indica</i> | Yola116 | | - | + | + | + | - | | - | - | - | - | + | - |
| <i>Tapinanthus globiferus</i> | <i>Tectona grandis</i> | Kano 33 | | - | + | + | + | - | | - | + | - | - | + | - |
| <i>Tapinanthus globiferus</i> | <i>Parinari curatellifolia</i> | Kano34 | + | + | + | + | + | - | + | - | - | - | + | + | |
| <i>Tapinanthus globiferus</i> | <i>Zyzyphus</i> sp. | Yola 115 | | - | + | + | + | - | | - | - | - | + | | - |
| <i>Tapinanthus globiferus</i> | Unknown | Kano 27 | + | - | + | + | + | - | + | - | - | - | - | + | |
| <i>Tapinanthus globiferus</i> | <i>Vitex doniana</i> | Suleija 73 | - | | + | + | + | - | - | - | + | + | - | + | |
| <i>Tapinanthus globiferus</i> | <i>Gmelina arborea</i> | Suleija 71 | | + | + | + | + | - | | - | + | - | + | | - |

+ = Present; - = absent; ++ = abundant; Gly = glycoside; Rsn = resins; Blm = balsam; Fla = flavonoids; Tnn = tannins; Akd = alkaloids; V.oil = volatile oil; Ptn = phlobatannin; Spn = saponin; Tep = terpenes; Str = sterols; Phn = phenols; Atq = anthraquinone.

metabolites of species of Loranthaceae on different hosts from different localities is presented in Table 1. Alkaloids were absent in all the species of Loranthaceae screened. Flavonoids

were present in all except few specimens, *A. dodoneifolius* on *Parkia biglobosa* from Jos, the two specimens of *G. braunii* collected on an unidentified host from Calabar, *P. capitata* on

Persea americana from Calabar and *Tapinanthus bangwensis* on *Citrus medica* from Ibadan (Table 1). Balsams occurred in all the specimens of the Loranthaceae species except few species like *A.*

Table 2. R_F values of phenolic spots from TLC profile of Loranthaceae species in Nigeria.

| Taxa | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | |
|--------------------------------------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| | | | | | | | | | | | | | | 0.06 | 0.06 | | 0.04 | 0.04 | | | 0.06 | | | 0.04 | |
| | | | | | | 0.07 | 0.10 | | | | | | | 0.09 | 0.10 | 0.07 | 0.09 | | | | | | | | |
| | | | | 0.12 | | | | 0.14 | | | 0.14 | | | | | 0.14 | | 0.13 | 0.14 | | | 0.12 | 0.12 | 0.13 | |
| | | | | | | 0.16 | | | | | | 0.16 | 0.16 | | | | 0.17 | | | | | 0.16 | 0.16 | | |
| | | | | 0.20 | | | 0.19 | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | 0.23 | 0.25 | 0.25 | 0.25 | 0.25 | | | | 0.25 | 0.23 | 0.26 |
| | 0.29 | 0.30 | | 0.27 | 0.29 | 0.30 | 0.30 | 0.30 | 0.29 | 0.27 | 0.29 | 0.27 | 0.29 | 0.29 | 0.29 | 0.29 | 0.29 | 0.29 | 0.29 | | 0.29 | 0.30 | | 0.30 | |
| | | | | 0.32 | | | | | | | | 0.32 | | | | | | | | | | | 0.32 | | |
| | 0.36 | | | 0.37 | | | | | | | 0.35 | 0.36 | 0.36 | 0.36 | 0.35 | 0.35 | 0.35 | 0.35 | | | | | 0.37 | 0.37 | |
| | | | | | | | 0.39 | | | | | 0.40 | | | 0.40 | | | | | | 0.40 | | | | |
| | | | | | | | | | | | 0.46 | | | | | | | 0.43 | 0.43 | | | 0.39 | | | |
| R_F values of spots | 0.49 | | 0.48 | 0.50 | | 0.49 | 0.50 | 0.50 | 0.49 | | | 0.49 | 0.49 | | | 0.49 | | 0.49 | 0.49 | 0.48 | 0.49 | 0.50 | 0.50 | 0.49 | |
| | 0.53 | | 0.53 | | | 0.52 | 0.53 | | 0.52 | | 0.53 | 0.52 | | | | 0.53 | 0.52 | 0.53 | | 0.53 | | | | | |
| | 0.56 | | | 0.55 | | 0.56 | | | 0.55 | | | 0.56 | 0.56 | | | 0.56 | | | | 0.55 | | | | | |
| | 0.59 | | 0.59 | 0.58 | | | 0.59 | 0.59 | | | | 0.59 | 0.58 | | 0.58 | | 0.59 | | | | | | | | |
| | | | | | | | | | 0.63 | | | | | 0.63 | | | | 0.63 | 0.63 | 0.63 | | | | | |
| | 0.66 | 0.66 | 0.66 | 0.65 | 0.65 | 0.65 | 0.65 | 0.65 | | | | 0.66 | 0.66 | | | 0.65 | 0.65 | | | | 0.65 | | | | |
| | | | | | | | | | | | | 0.68 | | | | | | 0.68 | 0.68 | 0.68 | | 0.69 | 0.69 | 0.68 | |
| | 0.73 | 0.73 | 0.73 | 0.72 | 0.72 | 0.72 | 0.72 | 0.72 | 0.72 | 0.71 | 0.71 | | | 0.71 | 0.71 | 0.71 | 0.71 | 0.71 | 0.72 | 0.71 | 0.71 | 0.71 | 0.73 | 0.73 | |
| | | | | | | | | | | | | | 0.78 | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | 0.84 | 0.84 | 0.84 | 0.84 | | 0.85 | 0.84 | 0.84 |
| | | | | | | | | 0.91 | 0.91 | 0.91 | 0.92 | 0.91 | | | | | | 0.91 | 0.91 | 0.91 | | | | | |
| No. of spots | 8 | 3 | 5 | 10 | 3 | 8 | 9 | 8 | 7 | 3 | 7 | 13 | 8 | 6 | 8 | 11 | 12 | 13 | 11 | 7 | 5 | 10 | 9 | 9 | |

Key: See Table 3.

dodoneifolius on *P. biglobosa* from Suleija and *T. bangwensis* on *N. laevis* and *C. medica* (Table 1). Each of the four specimens of *T. bangwensis* on different host plants from Ibadan lacked tannins and also the two specimens of *G. braunii* from Ibadan lacked tannin (Table 1). Phenolics were also found to occur in most of the specimens screened except *G. braunii* on *Theobroma cacao* and *T. bangwensis* on *C. medica* (Table 1).

Figure 1 shows percentage response of the specimens to the metabolites screened. From this study, none of the metabolites occurred in all specimens or even all species (Figures 1 and 2).

Figure 2 present the percentage of species responding to metabolites in each location while Figure 3 shows the percentage response to metabolites by the species. Generally, about 90% of the species tested positive for balsam and phenols, while 76% tested positive for tannins, 63% for saponins and less than 5% for phlobatannin (Figure 1). All the samples of *G. braunii* tested positive to balsam and saponins, all *P. capitata* tested positive to balsams, flavonoids and tannins, *P. nigriflora* tested positive to balsams, flavonoids, tannins, saponins, terpenes and phenols, while samples of *T.*

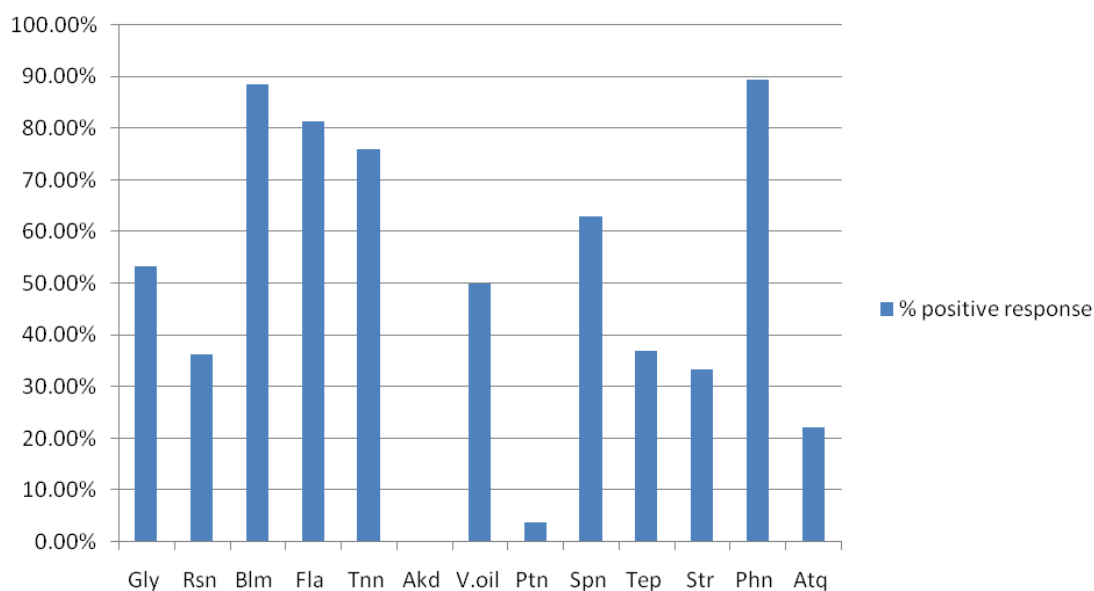
bangwensis varied in their chemical profiles with no consistent positive indication for a particular metabolite. However, over 75% tested positive to balsams, flavonoids, and phenols, while about 75% also tested positive to glycosides, resins, balsams, volatile oils, saponins and terpenes. All samples of *T. globiferus* tested positive to balsam, flavonoids and tannins and 75% was positive to phenols.

TLC profiling

The TLC profiling of the specimens of

Table 3. List of specimens, hosts and their corresponding extract spot number on the TLC plates {(Figures 4-6) and Table 2}.

| Specimen number on TLC plate | Name of Parasites | Name of Host | Host Family | Locality of collection/collection no. |
|------------------------------|---------------------------------|--------------------------------|---------------------------|---------------------------------------|
| 1 | <i>Tapinanthus globiferus</i> | <i>Piliostigma thoninngii</i> | Fabaceae-ceasalpinioideae | Kano 29 |
| 2 | <i>T. globiferus</i> | <i>Azadirachta indica</i> | Meliaceae | Yola116 |
| 3 | <i>T. globiferus</i> | <i>Tectona grandis</i> | Verbanaceae | Kano 33 |
| 4 | <i>T. globiferus</i> | <i>Parinari curattelifolia</i> | Chrysobalanaceae | Kano34 |
| 5 | <i>T. globiferus</i> | <i>Zyzyphus sp</i> | Rhamnaceae | Yola 115 |
| 6 | <i>T. globiferus</i> | <i>Terminalia avicenoides</i> | Combretaceae | Kano 35 |
| 7 | <i>T. globiferus</i> | - | - | Kano 27 |
| 8 | <i>T. globiferus</i> | <i>Vitex doniana</i> | Verbanaceae | Suleija 73 |
| 9 | <i>T. globiferus</i> | <i>Gmelina arborea</i> | „ | Suleija 71 |
| 10 | <i>Tapinanthus bangwensis</i> | <i>Newboldia leavis</i> | Bignoniaceae | Ibadan 46 |
| 11 | <i>T. bangwensis</i> | <i>Citrus medica</i> | Rutaceae | Ibadan 40 |
| 12 | <i>Tapinanthus cordifolius</i> | <i>Citrus auranthifolia</i> | „ | Jos 63 |
| 13 | <i>T. cordifolius</i> | <i>Cassia sp.</i> | Fabaceae-ceasalpinioideae | Jos 86 |
| 14 | <i>Phragmanthera capitata</i> | <i>Persea americana</i> | Lauraceae | Calabar 93 |
| 15 | <i>P. capitata</i> | <i>Persea americana</i> | „ | Calabar 89 |
| 16 | <i>Phragmanthera nigritana</i> | <i>Citrus sp.</i> | Rutaceae | Chaza, Suleija 78 |
| 17 | <i>Globimetula braunii</i> | <i>Persea americana</i> | Lauraceae | Calabar 90 |
| 18 | <i>G. braunni</i> | <i>Cola sp.</i> | Sterculiaceae | Calabar 92 |
| 19 | <i>Globimetula braunni</i> | <i>Cola sp.</i> | „ | Ibadan 97 |
| 20 | <i>G. braunni</i> | <i>Theobroma cacao</i> | „ | Ibadan 102 |
| 21 | <i>Agelanthus dodoneifolius</i> | <i>Parkia biglobosa</i> | Fabaceae-mimosoideae | Jos 65 |
| 22 | <i>A. dodoneifolius</i> | <i>Parkia biglobosa</i> | „ | Suleija 77 |
| 23 | <i>A. dodoneifolius</i> | <i>Casuarina sp.</i> | Casuarinaceae | Yola 119 |
| 24 | <i>A. dodoneifolius</i> | <i>Butryospermum parkii</i> | Sapotaceae | Yola 118 |

**Figure 1.** Percentage response of the total specimens of Loranthaceae species in Nigeria to presence of secondary metabolites. Gly = Glycoside; Rsn = Resins; Blm = Balsam; Fla = Flavonids; Tnn = Tannins; Akd = Alkaloids; V.oil = Volatile oil; Ptn = Phlobatannin; Spn = Saponin; Tep = Terpenes; Str = Sterols; Phn = Phenols; Atq = Anthraquinone

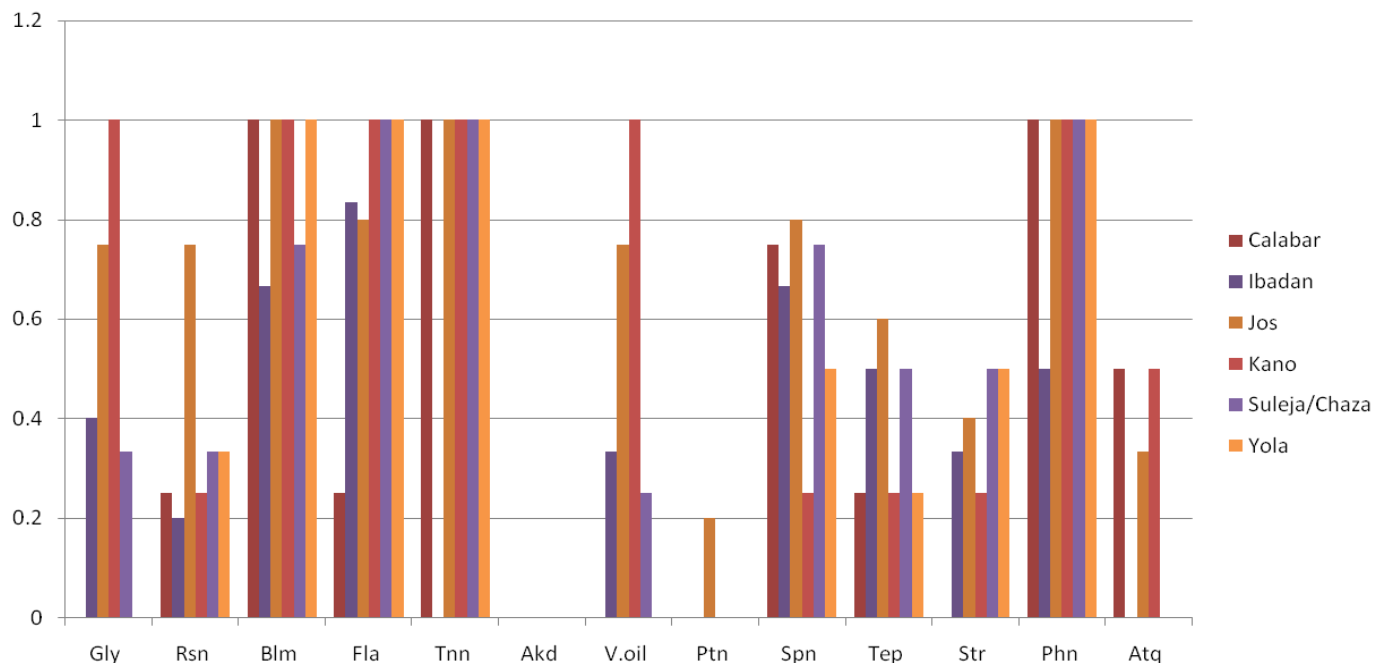


Figure 2. Percentage of species of Loranthaceae in Nigeria responding to secondary metabolites by location. Gly = Glycoside; Rsn = resins; Blm = balsam; Fla = flavonids; Tnn = tannins; Akd = alkaloids; V.oil = volatile oil; Ptn = phlobatannin; Spn = saponin; Tep = terpenes; Str = sterols; Phn = phenols; Atq = anthraquinone.

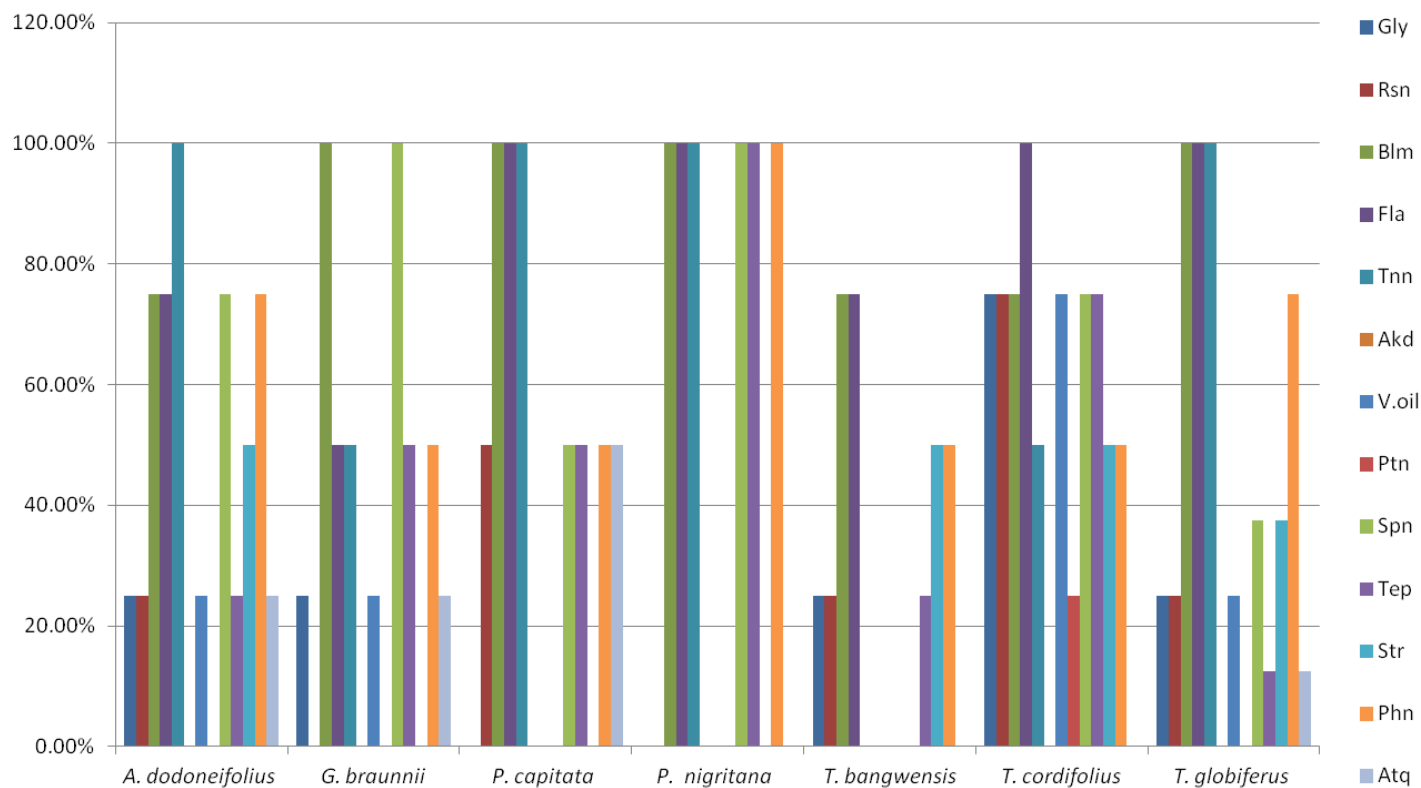


Figure 3. Percentage response of Loranthaceae species to secondary metabolites by species. Gly = Glycoside; Rsn = resins; Blm = balsam; Fla = flavonids; Tnn = tannins; Akd = alkaloids; V.oil = volatile oil; Ptn = phlobatannin; Spn = saponin; Tep = terpenes; Str = sterols; Phn = phenols; Atq = anthraquinone.

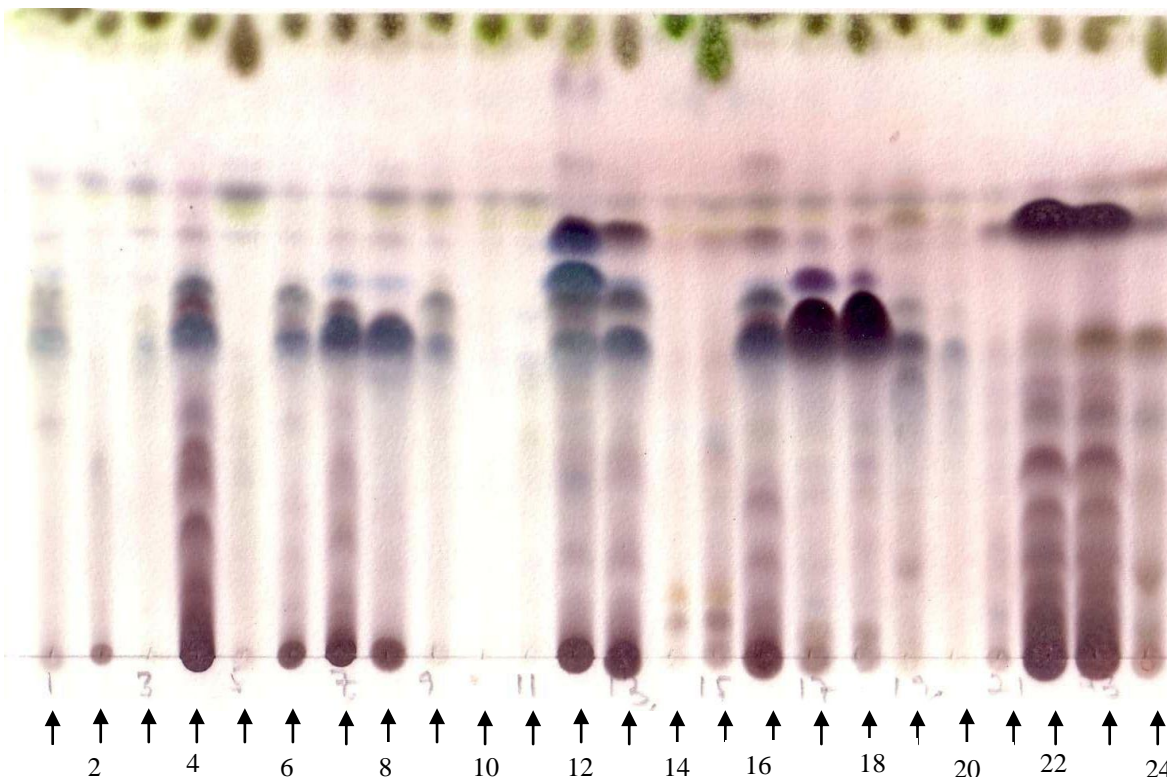


Figure 4. TLC profile of Loranthaceae specimens sprayed with Gibbs reagent.

Loranthaceae sp. using Gibbs, vanillin-sulphuric acid and Dragendoff spray reagents for TLC are shown in Figures 4, 5 and 6, respectively. In Figures 4, 5 and 6, Gibbs reagent were used for visualizing phenolics, vanillin in sulphuric acid for terpenoids, while Dragendoff reagent was used to see if alkaloids were present on the TLC plate, respectively. Table 2 shows the R_f values of spots found on the TLC plate in Figure 4, which reveals that all the specimens had phenolics in them although to varying degree judging from the numbers of spots. *T. cordifolius* on *Cassia* sp. from Jos (spot 12), *G. braunii* on *P. americana* (spot 17) and *G. braunii* (spot 18) from Calabar have the highest number of spots of 13, 13 and 12, respectively (Table 2). An intermediate number of spots were found in *T. globiferus* on *P. curatellifolia* from Kano (spot 4), *P. nigritana* on *Citrus* sp. from Suleija (spot 16) and *G. braunii* on *Cola* sp. from Ibadan (spot 19) with 10, 11 and 11 spots respectively (Table 2). The lowest spots are found in *T. globiferus* on *A. indica* (spot 2), *T. globiferus* on *Tectona grandis* (spot 3), *T. globiferus* on *Zyzyphus* sp. (spot 5), *T. bangwensis* on *Newboldia laevis* (spot 10) and *Agelanthus dodoneifolius* on *P. biglobosa* (spot 21) with 3, 5, 3, 3 and 5, respectively (Table 2). Spots with R_f values of 0.29 - 0.32 and 0.71 - 0.73 are found to be present in over 90% of the specimens. In Figure 5, terpenoids were only observed in some of the specimens. Alkaloids were absent from all the specimens studied (Figure 6).

DISCUSSION

The phytochemical analysis and the TLC profiling showed variation in the constituent secondary metabolites among various species irrespective of their host and ecological location (Table 1; Figures 4 and 5). Variation in secondary metabolites among the same mistletoe species occurring on different host plants have been observed in earlier studies (Deeni and Sadiq, 2002; Ibrahim et al., 2009). The only consistent pattern from this study was the lack of alkaloids from all the specimens analyzed (Table 1; Figures 1, 2 and 6). It is a known fact that quantitative and qualitative information on secondary metabolites is useful for taxonomic classification of plants (Harborne, 1968; Takhtajan, 1973). Hence absence of alkaloids, and the number of species testing positive for balsam and phenols appears to be of chemotaxonomic significance among the species in Nigeria. Alkaloids were not recorded for any of the Nigerian Loranthaceae specimens studied but Sanchez-Areola et al. (2004) recorded the presence of alkaloids in *Psittacanthus calyculatus*, a New World Loranthaceae endemic to Mexico (Kuijt, 2009).

The TLC R_f in Table 2 shows that there were similar phenolic compounds (R_f values of 0.29 - 0.32 and 0.71 - 0.73) present in most of the specimens, over 90% of the species and this further reinforced the fact that phenolics could be a source of analytical marker compound(s) for

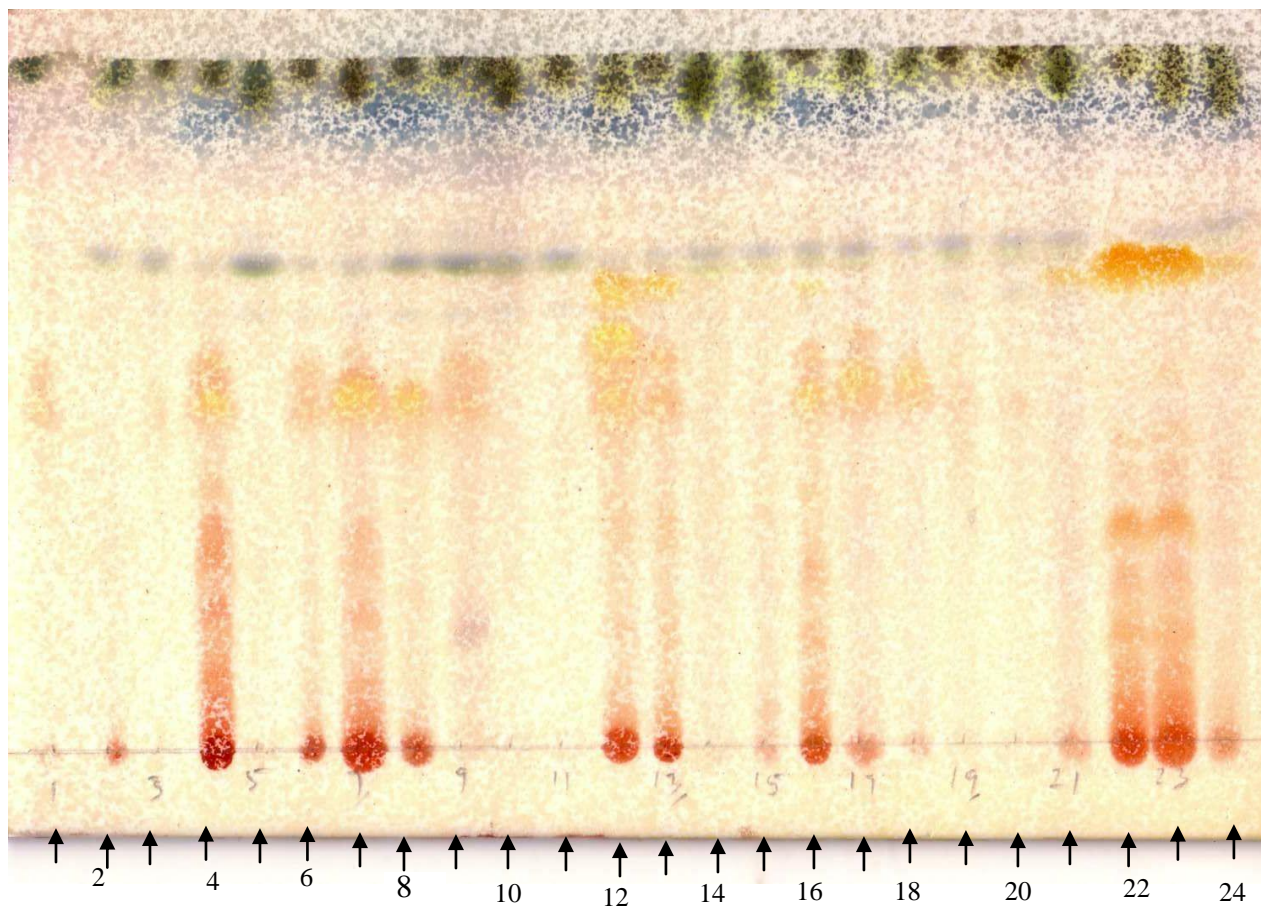


Figure 5. TLC profile of Loranthaceae specimens sprayed with Vanillin-sulphuric acid reagent.

standardization of herbal preparations from these species. High amounts of phenolics have long been known to be a phytochemical feature of parasitic flowering plants and they are said to occur at a level that is generally higher than the host plant (Khanna et al., 1968; Salatino et al., 1993). The study reveals that *G. braunii* specimens irrespective of their hosts or locations are rich in phenolic compounds as compared to other species while *T. globiferus* and *T. bangwensis* are depauperate in phenolics as compared to other species. Also of note is the absence of glycosides in the *Phragmanthera* species and total absence of tannins from all the specimens from Ibadan. These findings may be of chemotaxonomic importance. Thus, the presence of balsams and phenols could be used in specific combination with morphological characteristics and biogeographical distribution ranges for the delineation of genera and species in the family (Crockett and Robson, 2011).

Research on dwarf mistletoes (Viscaceae) in North America indicates that plant chemistry, particularly secondary metabolites, plays an important role in determining interactions between host and parasite (Snyder, 1996). This may not be applicable to Nigerian Loranthaceae

because of the variation noted in the metabolites present in the same species on different hosts. Differences in chemical profiles of the various species studied underscore why the specific choice of species for the treatment of a particular ailment is very important. This study has shown that some species may not possess a particular metabolite that is common in other species. For instance, the absence of glycosides in *Phragmanthera* species or tannins and saponins in *T. bangwensis* may result in major pharmacological differences. Although the correlation between host and chemical profile of the species was not clearly defined in this study, it is believed that the host could play a role in the observed chemical profile of the plant or species. The influence of host chemistry on the chemical constituents of the parasite on different hosts might justify why the host is as important as the parasite in pharmacognosy, ethnopharmacology and ethnomedicine, and why the use of these Loranthaceae in the treatment of an ailment is often dependent on a particular or specific host (Burkill, 1995; Snyder 1996; Adodo, 2002; Olapade, 2002; Preston et al., 2010), for instance, in Brazil, where there is preference for *Cladocolea micrantha* growing on cashew tree (*Anacardium occidentale*) for the treatment of tumors

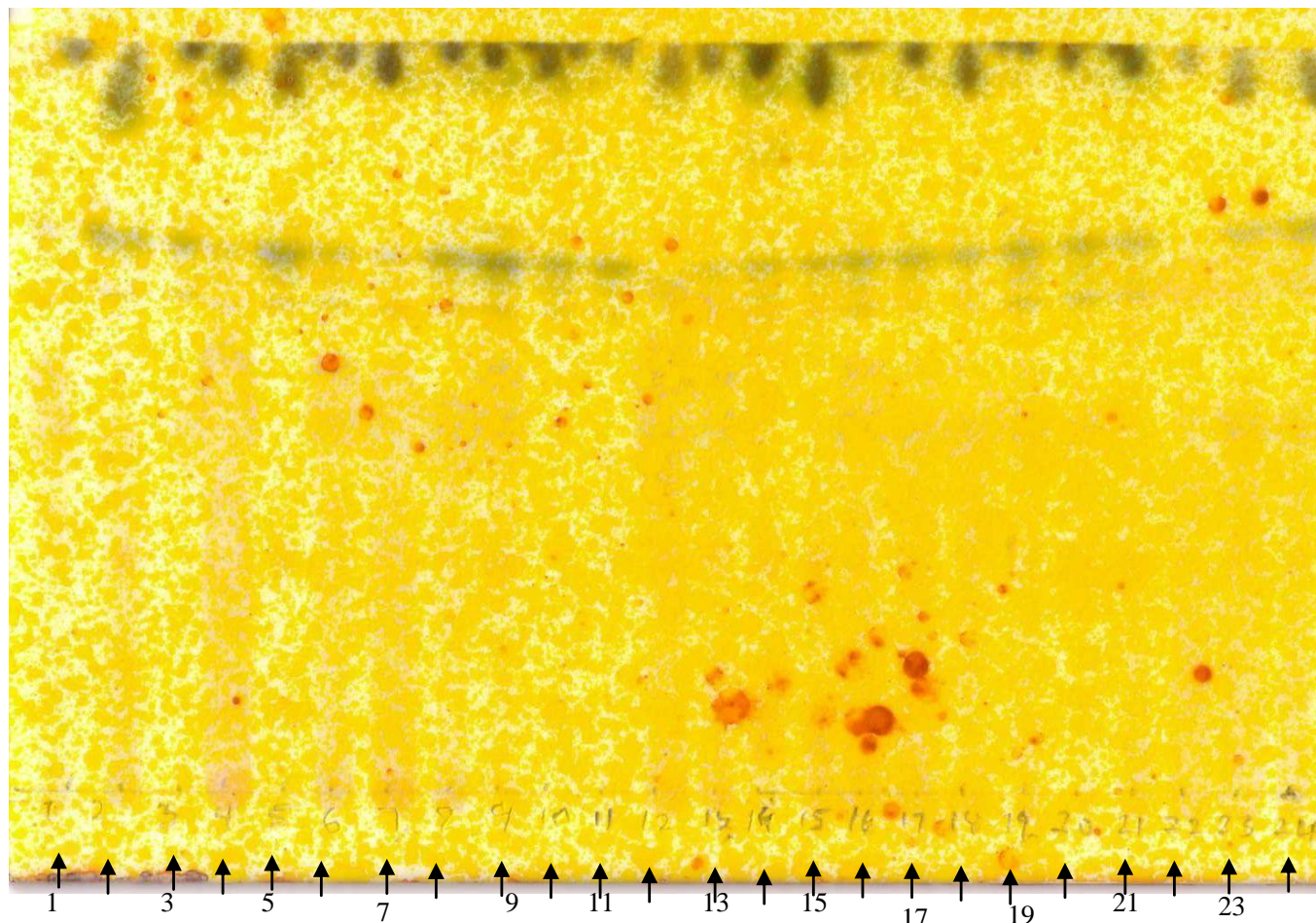


Figure 6. TLC profiling of Loranthaceae specimens sprayed with Dragendoff reagent.

and inflammatory diseases (Adodo, 2002; Olapade, 2002; Guimaraes et al., 2007).

Conclusion

From this investigation, species of Loranthaceae in Nigeria might not be delineated by scoring presence or absence of their secondary metabolites qualitatively or quantitatively due to variations which occur on same species from different hosts but the occurrence of similar metabolites like phenolics and balsam in most, if not all the species irrespective of the host and locality is useful taxonomically as a marker for the group. It is therefore our recommendation that caution should be exercised in the use of Loranthaceae as phytomedicine because of the chemical variations which exist in the same species found on different hosts. The same species collected from two different hosts might have different pharmacological effects in the body. The group is currently working on determining a phytochemical marker for the family Loranthaceae in Nigeria.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors are grateful to the following people who rendered assistance during field trips for specimen collection: Dr Theresa Omara-Achong, Oyepeju M.K.O, Baba Nafi of Keji village, Pastor Frank of University of Calabar, Dr. Colman Goji, Muazzam Ibrahim, Tanko Garba, Mrs. Sumbo Wahab and Mr. Owolabi; also Mr. John Apev, who assisted in the laboratory analysis.

REFERENCES

- Adodo A (2002). Nature Power: A Christian approach to Herbal Medicine. St Benedict Monastery, Ewu-Esan, Edo State, Nigeria, pp. 207.
- Aslam J, Khan SH, Siddiqui ZH, Fatima Z, Maqsood M, Bhat MA, Nasim SA, Ilah A, Ahmad IZ, Khan SA, Mujib A, Sharma MP (2010). *Catharanthus roseus* (L.) G.Don. An Important Drug: It's Applications

- and Production. Pharmacie Globale (IJCP). 1(4):1-16.
- Brain KR, Turner TD (1975). The practical evaluation of phytopharmaceuticals. Wright Scientechnica, Bristol, pp. 81-82.
- Burkill HM (1995). The useful plants of West Tropical Africa. 2nd ed. Vol. 3. Families J-L. Royal Botanic Gardens, Kew, pp. 857.
- Bussing A, Suzart K, Bergmann J, Pfeiller U, Schietzel M, Schweizer K (1996). Introduction of apoptosis in human lymphocytes treated with *Viscum album* L. is mediated by the mistletoe lectins. Cancer Lett. 99:59-72.
- Crockett SL, Robson NKB (2011). Taxonomy and Chemotaxonomy of the Genus *Hypericum* (Invited Review). Med. Aromat. Plant Sci. Biotechnol. 5(1):1-13.
- Cunningham AB (1993). Africa Medicinal Plants: Setting priorities at the interface between conservation and primary health care. People and plant working paper 1. Paris. UNESCO. <http://unesdoc.unesco.org/images/0009/000967/096707e.pdf>
- Dana GD (2012). *Artemisia annua*, Artemisinin, ACTs & Malarial Control in Africa. Tradition, Science and Public Policy. Washington D.C, pp. 17-21. www.mmv.org/artemisinin
- Deeni YY, Sadiq NM (2002). Antimicrobial properties and phytochemical constituents of the leaves of African mistletoe (*Tapinanthus dodoneifolius* (DC) Danser) (Loranthaceae): an ethnomedicinal plant of Hausaland, Northern Nigeria. J. Ethnopharmacol. 83:235-240
- Erturk O, Hati H, Yayli N, Demirbag Z (2003). Antimicrobial activity of *Viscum album* L. subsp. *Abietis* (Wiesb). Turk. J. Biol. 27:255-258.
- Evans WC (2002). *Trease and Evans Pharmacognosy*, 15th Ed., W.B. Sanders, London, pp.183-393.
- Fernandez T, Wagner ML, Varela BG, Ricco RA, Hajos SE, Gurni AA, Alvarez E (1998). Study of an Argentine mistletoe, the hemiparasitic *Ligaria cuneifolia* (R.et P.) Tiegh. (Loranthaceae). J. Ethnopharmacol. 62:25-34.
- Gedalovich-Shedletzky E, Delmer DP, Kuijt J (1989). Chemical composition of Viscin mucilage from three mistletoe species - a comparison. Ann. Bot. 64(3):249- 252.
- Guimaraes AC, Kuster RM, Amaral ACF, Ferreira JLP, Sinia AC (2007). Histological study of the leaf and stem of the Amazonian medicinal mistletoe *Cladocolea micrantha* (Loranthaceae). Int. J. Bot. 3(2):218-221.
- Harborne JB (1968). The use of secondary chemical characters on the systematics of higher plants. Chemotaxonomy and Serotaxonomy. J. G. Hawkes. Ed. Academic Press London
- Ibrahim JA, Ayodele AE (2011). Taxonomic Revision of the Nigerian Loranthaceae. Nigerian J. Bot. 24(1):153-188.
- Ibrahim JA, Ayodele AE, Okhale SE, Jegede AI, Kunle OF (2009). The taxonomic significance of *Agelanthus dodoneifolius* (DC.) Polh. & Wiens in relation to its hosts. Nigerian J. Bot. 22(1):89-101.
- Khanna SK, Viswanathan PN, Tewari CP, Krishnan PS, Sanwal GG (1968). Biochemical aspects of parasitism by angiosperm parasites: phenolics in parasites and hosts. Physiologia Plantarum. 21:949-959.
- Kuijt J (2009). Monograph of *Psittacanthus* (Loranthaceae). Systematic Botany Monographs. American Society of Plant Taxonomists, Ann Harbor. Vol. 86.
- Mengs U, Gothel D, Leng-Peschlow E (2002). Mistletoe extracts standardized to mistletoe lectins in oncology: review on current status of preclinical research. Anticancer Res. 22:1399-1407.
- Olapade EO (2002). The herbs for good health: the 50th Anniversary Lecture of the University of Ibadan. NARL Specialist Clinic, Ibadan, Nigeria, pp. 230.
- Preston AL, An M, Watson DM (2010). Chemical profile differences in endemic parasitic weeds: a study of host-parasite chemical profiles in select mistletoe and *Eucalyptus* species. Seventeenth Australasian Weeds Conference.
- Salatino A, Kraus JE, Salatino MLF (1993). Contents of tannins and their histological localization in young and adult parts of *Struthanthus vulgaris* Mart. (Loranthaceae). Ann. Bot. 72:409-414.
- Sanchez-Areola E, Maiti RK, Trujillo-Perez B (2004). Morpho-anatomical characters and secondary metabolites from *Psittacanthus calyculatus* (Loranthaceae). Int. J. Exp. Bot. 53:119-121.
- Segelman AB, Farnworth NR, Quimby MW (1971). Biological and physicochemical evaluation of plants III. False Negative saponin test results induced by the presence of tannins. Planta Medica. 19:304.
- Snyder M, Fineschi B, Linhart YB, Smith RH (1996). Multivariate discrimination of host use by dwarf mistletoe *Arceuthobium vaginatum* subsp. *cryptopodum*: inter- and intraspecific comparisons. J. Chem. Ecol. 22:295-305.
- Sofowora A (1993). Medicinal Plants and Traditional medicine in Africa. 2nd Ed., Spectrum Books Limited, Ibadan, Nigeria, pp. 145-148.
- Stein GM, Pfueller U, Schietzel M, Bussing A (1999). Thionins from *Viscum album* L. influence of the viscotoxins on the activation of granulocytes. Anticancer Res. 19:1037-1042.
- Stirpe F, Sandvig K, Olsnes S, Pihl A (1982). Action of viscum, a toxic lectin from mistletoe on cells in culture. J. Biol. Chem. 257:1371-1377.
- Takhtajan A (1973). The Chemical approach to plant classification with Special reference to the higher taxa of Magnoniales. In: Chemistry in Botanical classification, pp. 275-285.
- Tilney PM, Lubke RA (1974). Chemotaxonomic study of twelve species of the family Loranthaceae. J. South Afr. Bot. 40(4):315-332.
- Traoré R (2000). Etude pharmacologique chez l'animal de l'extrait aqueux de *Tapinanthus dodoneifolius* (DC). Danser (Loranthaceae) utilisée en thérapie anti-asthmatique au Burkina Faso. Thèse de pharmacie. FSS. Université d'Ouagadougou, Burkina Faso.

Short Communication

Efficacy of leaf extracts of some medicinal plants on growth of *Colletotrichum capsici* butler and bisby

Shinde J. U. and D. U. Gawai*

Botany Research Laboratory and Plant Disease Clinic, P. G. Department of Botany, N. E. S. Science College, Nanded. (M. S.), India.

Received July 26, 2011; Accepted 25 January, 2012

Attempts were made to determine the effect of leaf extracts of *Azadirachta indica*, *Ocimum sanctum*, *Tridax procumbens*, *Clerodendron innermis*, *Cathranthus roseus*, *Ricinus communis*, *Citrus limon* against *Colletotrichum capsici*. Out of these medicinal plants tested, 15% alcoholic extract of *Azadirachta indica* and *O. sanctum* was found inhibitory for the growth of *Colletotrichum capsici*. The results show that extracts from leaves of different plants vary in their effects on growth of *C. capsici*. It is evident from the results that aqueous and alcoholic leaf extracts of *A. indica* and *O. sanctum* exhibited strong fungitoxicity against *C. capsici*. Alcoholic extract of all the seven plants showed significant result as compared to aqueous extracts.

Key words: Chilli, anthracnose, leaf extract, *Colletotrichum capsici*.

INTRODUCTION

The total area under the cultivation of chilli crop in India is about 0.7 to 0.9 million hectares. It is grown extensively in Tamil nadu, Andhra pradesh, Karnataka and Maharashtra. It is grown as a rain fed crop in most parts of Andhra pradesh and as an irrigated crop in other areas. There are several varieties of chilli grown in India and some are non-pungent with large sized fruits that are used mainly as vegetables. Chilli is one of the important crops grown for its valuable fruit in making spices and condiments. It forms a part of the Indian diet. The fruits are used either dry or raw. It is used in green as well as dry powder form, rich source of vitamin A and vitamin C among the vegetable.

In chilli, there are various types of diseases but anthracnose is a serious disease of chilli found in India, caused by *Colletotrichum capsici* where it occurs in severe form in all the southern states. The disease is developed due to hot and humid conditions. The disease has been identified in all the chilli producing regions of the world and has become a serious constrain in chilli production whenever the crop is grown. Anthracnose causes extensive pre- and post-harvest damage to chilli fruits causing anthracnose lesions. These fungal infection are known to cause heavy damages and impair the quality of fruit seeds. Even small anthracnose lesions on chilli fruits reduce their marketable value (Manandhar

*Corresponding author. E-mail: dilip.gawai777@gmail.com

et al., 1995). Anthracnose disease can occur on leaves, stems and both pre- and post-harvest fruits (Isaac, 1992). Typical fruit symptoms are circular or angular sunken lesions, with concentric rings of acervuli that are often wet and produce pink to orange conidial masses under severe disease pressure, lesions which may coalesce. Conidial masses may occur in concentric rings on the lesions. Management and control of the anthracnose disease are still under extensive research (Yoon et al., 2004). Many studies have concluded that disease management practices are often inadequate to eliminate the diseases. Breeding to develop the long-lasting resistant varieties has also not been successful due to involvement of multiple *Colletotrichum* species in anthracnose infection.

MATERIALS AND METHODS

Collection of samples

Diseased chilli fruits were collected in polyethylene bags from fields and local market of Nanded city of Maharashtra State (India).

Identification of pathogen

The diseased chilli fruits were preliminary observed for sporulation characters like asexual or sexual spores or fruiting structures under compound microscope and their Identification was confirmed with the help of latest manuals (Subramanian, 1971; Jha 1993). Pure cultures of the identified fungus was prepared and maintained on Czapek dox agar slants for further experiments.

Preparation of plant extracts

Seven common and easily available plants like *Azadirachta indica*, *Ocimum sanctum*, *Tridax procumbens*, *Clerodendron innermis*, *Cathranthus roseus*, *Ricinus cummunis*, *Citrus limon* were selected. The leaves of the plant were collected separately, surface sterilized with 0.1% HgCl₂ and washed repeatedly with sterile distilled water for several times and kept for drying in hot air oven (Metalab) at 60°C temperature for 48 h.

Aqueous extract

The dried leaves of selected plants were crushed separately into fine powder with the help of blender 5, 10 and 15 g each of the plant powder was dissolved separately in 100 ml sterilized hot distilled water and filtered through Whatman No.1 filter paper. The filtrates were used as 5, 10 and 15% concentrations of aqueous plant extracts, respectively.

Alcoholic extracts

For alcoholic extract 5, 10, 15 g of each sun-dried medicinal plant material, were cut into small pieces and then macerated by blender 1 to 2 mm separately and the powder produced was blended in ethyl alcohol (1:10 w/v) and extracted under cold conditions for 24 h. The resultant extract was filtered through a glass wool filter and then rinsed with a small quantity (30 ml) of 96% alcohol. The extracts were evaporated under reduced pressure at 40°C.

Subsequently, the extracts were diluted by distilled water and stored in the deep freezer at -10°C (Fardos, 2009).

Evaluation of plant extracts against *C. capsici*

The effect of 5, 10, and 15% aqueous and alcoholic leaf extract was determined by measuring the mycelial dry weight. 50 ml of glucose nitrate medium was poured into each flask containing different concentrations (5, 10 and 15%) of the respective extracts (2 ml each). With a sterile cork borer (3 mm), mycelial disc of seven days old cultures of the isolates were inoculated in the flask and incubated at 28 ± 2°C. After seven days, the content of flasks were filtered through Whatman No. 1 filter paper The content were dried at 70°C for 24 h and percentage inhibition of mycelial growth was evaluated using the poisoned food techniques (PFT), and calculated using the formula given by Vincent (1927) and Ogbebor et al. (2007).

$$\% \text{ Inhibition} = \frac{100 (\text{Control} - \text{Treatment})}{\text{Control}}$$

RESULTS AND DISCUSSION

Biological control of fruit rot and dieback of chilli with plant products tested in many laboratories and field trials showed that the *O. sanctum* leaf extract and neem (*A. indica*) oil could restrict growth of the anthracnose fungus (Jeyalakshmi and Seetharaman, 1998). It is clearly evident from the results that the aqueous and alcoholic leaf extracts of all the plants tested against *C. capsici* significantly reduced mycelial dry weight, leaf extract of *A. indica* showed high percentage inhibition of mycelial dry weight (60.62, 71.05 and 81.57%) at 5, 10 and 15% aqueous extract while leaf extract of *Ricinus cummunis* showed very low percentage inhibition of mycelial dry weight (25, 37.36 and 35.52) (Tables 1 and 2). The alcoholic leaf extracts of all the plants tested were found to be more effective as compared to aqueous leaf extracts. The leaf extract of plants which vary in their effect on growth of *C. capsici* may be due to differential effect of active ingredient present in plants. Percentage inhibition of mycelial dry weight of *C. capsici* was highly inhibited in 15% alcoholic leaf extract of *A. indica* followed by leaf extract *O. sanctum*. Upadhyaya and Gupta (1990) reported the control of *Curvularia lunata* with extracts of *Ocimum sanctum*. Singh et al. (1993) reported the effectiveness of aqueous extracts of *O. sanctum* and *A. indica* in the control of disease development in banana. In this study, the differences in the inhibition of mycelial growth of *C. capsici* may be due to variations in fungitoxicity of leaf extract. Similarly, Kurucheve et al. (1997) observed that the variation in the inhibitory effect of plant extracts may be due to qualitative and qualitative differences in antifungal principles. The strong fungitoxicity exhibited by the leaf extract may be due to presence of chemical constituents including tannins, glycosides, alkaloids and flavonoids (Harborne, 1984).

It is clear from the results that all the leaf extracts of seven plants exhibited antifungal activity. Among these, many

Table 1. Effect of aqueous leaf extracts of some medicinal plants on *C. capsici*.

| Plant name | % inhibition of mycelial dry weight at different concentrations | | |
|---|---|-------|-------|
| | 5% | 10% | 15% |
| <i>Azadirachta indica</i> A. Juss. | 60.62 | 71.05 | 81.57 |
| <i>Ocimum sanctum</i> L. | 51.31 | 65.78 | 76.31 |
| <i>Clerodendrum inerme</i> (L.) Gaertn. | 35.52 | 50.00 | 53.94 |
| <i>Tridax procumbens</i> L. | 44.73 | 57.89 | 68.42 |
| <i>Catharanthus roseus</i> (L.) G. Don | 42.10 | 55.26 | 63.15 |
| <i>Ricinus communis</i> L. | 25.00 | 37.36 | 35.52 |
| <i>Citrus limon</i> L. | 31.57 | 42.89 | 46.05 |

Table 2. Effect of alcoholic leaf extracts of some medicinal plants on *C. capsici*.

| Plant name | % inhibition of mycelial dry weight at different concentrations | | |
|---|---|-------|-------|
| | 5% | 10% | 15% |
| <i>Azadirachta indica</i> A.Juss. | 68.42 | 76.31 | 89.47 |
| <i>Ocimum sanctum</i> L. | 63.15 | 75.00 | 84.21 |
| <i>Clerodendrum inerme</i> (L.) Gaertn. | 53.94 | 68.42 | 72.36 |
| <i>Tridax procumbens</i> L. | 57.89 | 72.36 | 76.31 |
| <i>Catharanthus roseus</i> (L.)_G.Don | 54.47 | 71.05 | 75.00 |
| <i>Ricinus communis</i> L. | 50.00 | 62.63 | 67.36 |
| <i>Citrus limon</i> L. | 52.63 | 65.26 | 71.05 |

workers have reported antifungal activities of different plant species and stressed the importance of plants as possible sources of natural fungicides (Tewari, 1995; Lakshmanan, 1990; Singh et al., 1993; Ogbemor et al., 2005).

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENT

Authors are thankful to the Principal, N.E.S. Science College, Nanded for providing facilities.

REFERENCES

- Fardos MB (2009). Antifungal activity of some medicinal plant used in Jeddash, Saudi Arabia Mycopathol. 7(1):51-57.
- Harborne JB (1984). Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. 2nd Ed. Chapman and Hall, New York, 288pp.
- Isaac S (1992). Fungal Plant Interaction. London: Chapman and Hall Press. p. 115.
- Jeyalakshmi C, Seetharaman K (1998). Biological control of fruit rot and die-back of chilli with plant products and antagonistic microorganisms. Plant Dis. Res. 13:46-48.
- Jha DK (1993). A Text book of Seed Pathology, Vikas Publishing house Pvt. Ltd. New Delhi; 132pp (Reprint 1995).
- Kuruchev V, Gerard EJ, Jayaraj J (1997). Screening of higher plants for fungitoxicity against *Rhizoctonia solani* in vitro. Indian Phytopathol. 50(2):235-241.
- Lakshmanan P (1990). Effect of certain plant extracts against *Corynespora cassicola*. Ind. J. Mycol. Plant Path. 20(3):267-269.
- Ogbemor ON, Adekunle AT, Enobakhare DA (2007). Inhibition of *Colletotrichum gloeosporioides* (Penz) Sae. Causal organism of rubber (*Hevea brasiliensis* Muell. Arg.) leaf spot using plant extracts. Afr. J. Biotech. 6(3):213-218.
- Manandhar JB, Hartman GL, Wang TC (1995). Anthracnose development on pepper fruits inoculated with *Colletotrichum gloeosporioides*. Plant Dis. 79:380-383.
- Singh HNP, Prasad MM, Sinha KK (1993). Efficacy of leaf extracts of some medicinal plants against diseases development in banana. Lett. Appl. Microbiol.. 17(6):269 – 271.
- Subramanian C.V. (1971). Hypomyces- An account of Indian species except *Cercospora*. ICAR, New Delhi.
- Tewari SN (1995). *Ocimum sanctum* La botanical fungicide for rice blast control. Trop. Sci. 35:263-273.
- Upadhyaya ML, Gupta RC (1990). Effect of extracts of some medicinal plants on the growth of *Curvularia lunata*. Indian J. Myco. Plant Path. 20(2):144-145.
- Vincent JM (1927). Distortion of fungal hyphae in the presence of certain inhibitors. Nature, 850pp.
- Yoon JB, Yang DC, Lee WP, Ahn SY, Park HG (2004). Genetic resources resistant to anthracnose in the genus *Capsicum*. J. Korean Soc. Hortic. Sci. 45:318-323.

Full Length Research Paper

Effects of crude extracts on some selected physiological parameters of French beans (*Phaseolus vulgaris*) infected with rust (*Uromyces appendiculatus*)

Menge D. M. S.¹, Makobe M.¹, Monda E. O.² and Okemo P. O.²

¹Department of Botany, Faculty of Science, Jomo Kenyatta University of Agriculture and Technology, P.O Box 62000-00200, Nairobi, Kenya.

²Department of Plant and Microbial Sciences, School of Pure and Applied Sciences, Kenyatta University, Nairobi, Kenya.

Received 7 February, 2012; Accepted 12 June, 2014

Rust (*Uromyces appendiculatus*) is a major foliar disease that reduces yield and pod quality in beans. The field trial of French beans was established at Jomo Kenyatta University of Agriculture and Technology (JKUAT). Single plant extracts and combinations of *Boscia angustifolia*, *Zanthoxylum chalybeum* and *Melea volkensii* were used to evaluate their effect on *U. appendiculatus* in the field. During the growing period, beans were infected with rust from natural inoculum at the field. Physiological responses such as carbon dioxide assimilation, transpiration (E), stomatal conductance (gs), and photosynthetic rate (Pn) of French beans treatments were examined after extracts of three antifungal plants were sprayed. *B. angustifolia* - *Z. chalybeum* combination and single plant treatment *M. volkensii* had positive effects on enhancing the rate of photosynthesis in bean plants. The high regressions between stomatal conductance and rate of transpiration in the treatments indicated that stomatal conductance and rate of transpiration were interdependent and it was interpreted to mean that stomatal conductance enhanced rate of transpiration at different times of the day. These plant extracts however caused an increase in the rate of transpiration of the bean plants, which resulted in loss of water. Results reveal bioactive potential of the flora from *M. volkensii* and a combination of *B. angustifolia* and *Z. chalybeum* to produce metabolites with potential applications as botanical pesticides.

Key words: Antifungal, beans, physiological responses, rust.

INTRODUCTION

The importance of the French beans is due to their high nutritive value in both energy and protein contents. Therefore, increasing the crop production is one of the most important targets of agricultural policy in several countries. The bean rust fungus (*Uromyces appendiculatus*) is of worldwide importance as a yield-

reducing disease of *Phaseolus vulgaris* L., potentially cause yield losses up to 50% (Venette and Jones, 1982; Berger et al., 1995; De Jesus Junior et al., 2001). Under severe disease, it completely defoliates the plant and can cause 100% crop failure (Steadman et al., 2002). Rust result in harmful effects on growth, most physiological

activities and the yield of beans. On global scale, studies have shown that some plant species have antifungal compounds (Fabry et al., 1996; Okemo et al., 2003). Within this context, natural products from plants seem to be a good alternative since numerous plants have the potential to control phytopathogenic fungi, and have much prospect to be used as a fungicide. Despite the many studies performed on biological control, relatively little is known about the role of the plant extracts (*Boscia angustifolia*, *Melea volkensii* and *Zanthoxylum chalybeum*) applied on the physiological parameters of the plants. In this study, we hypothesized that antifungal plant extracts might influence physiological activities of bean plants. Therefore, this study aimed at studying the role of selected plant extracts (added singly or in combination) in influencing photosynthetic activities of bean plants and finding an explanation for the above role based on test attributes.

MATERIALS AND METHODS

Study site

Field studies were carried out at JKUAT in Thika District. The university is located at latitude 1°05 S and longitude 37°00 E. It lies at an altitude of 1525 m above sea level and it receives an annual rainfall of 850 mm. Temperatures range from 13 and 26°C.

Collection and processing of plant materials

The samples of desired plants (*B. angustifolia*, *M. volkensii* and *Z. chalybeum*) from previous experiments (Omwenga, 2009; Kiswii, 2009) for antifungal activity were collected from different parts of the country (Samburu, Mombasa, Mwingi, Kakamega forest and Nakuru) in clean sacks (Table 1). The plants were identified and verified at Jomo Kenyatta University of Agriculture and Technology (Taxonomy unit, Department of Botany). Voucher specimens were deposited in the herbarium. The samples were labeled and deposited in the botany laboratory. The plant leaves and roots were dried separately at room temperature for a period of 1-2 weeks and then ground separately to powder using a grinding mill at 8000 rpm (Type 8 lab mill). The powder was stored in plastic bags at room temperature until the time required. Two kilograms of each plant sample was soaked and left overnight to allow extraction of the crude active compounds. The supernatant was filtered in several layers of muslin cloth and volumes adjusted to 20 L. (Stoll, 2000). A combination of *B. angustifolia*, *M. volkensii* and *Z. chalybeum* extracts was used because previous experiments (Menge, 2011) revealed a better synergism between the two in reducing the disease severity. A normal washing bar soap ground to powder and dried was used as a sticker at a rate of 1 g per litre of water extracts. Untreated control was used containing water and soap

only without the extracts. During the growing period, beans were infected with rust from natural inoculum at the field. Seeds were obtained from Regina Seed Company and planted at a spacing of 30 cm between rows and 10 cm between plants within the rows (Monda et al., 2003). French bean seeds commercially available coated with thiram were used to control root rots. French bean variety Amy seeds were planted in 4 × 3 m plots each separated by a 1 m path between the treatments and the replications. Amy is high yielding as compared to other varieties therefore it is grown by most farmers. Di-ammonium phosphate was used at planting, at a rate of 200 kg/ha mixed well before seed placement. Calcium ammonium nitrate was applied at a rate of 100 kg/ha at trifoliolate leaf stage.

Experimental design type

The experiment was carried out in a randomized complete design, and data analyzed using analysis of variance; and comparison of means was made by using Duncan's multiple range test. The treatments consisted of six plant extracts, copper hydroxide 61.4% (Kocide DF: metallic copper equivalent 40% formulated as a dry flowable) as a positive control and a negative water control. A spray regime of once a week using a knap sack was employed from the fifteen days after planting until flowering. The extracts were used as protectants. The fungicide was applied at a rate of 2.5 kg ha⁻¹ according to the manufacturers' recommendations. There were a total of seven hundred and sixty plants per replicate. Overhead irrigation twice a week and weeding were done as necessary.

CO₂ exchange measurements

Three different types of leaf gas exchange measurements were made on plants from the interior rows of the plots. First, at approximately weekly intervals, measurements of carbon dioxide assimilation rate were made at 0900, 1200, and 1500 h at the JKUAT farm. Mature, fully illuminated upper canopy leaves were measured at their nominal daytime growth. Daylight patterns of carbon dioxide assimilation rate were measured by the infrared gas analyzer (IRGA). IRGA was used as a null point instrument that allows the flow of carbon dioxide into the system at a rate equivalent to the rate of uptake of the leaf. The amount of carbon dioxide assimilated by the leaf was read directly from the IRGA. French bean leaf tissues from ten selected plants from each treatment were enclosed in the leaf chamber (leaf chamber = 2.5 cm²) one at a time. The air flow rate through the chamber remained fixed. The carbon dioxide assimilation was monitored for 1 min for each leaf by the IRGA connected in an open gas flow system. During measurement of CO₂ assimilation rate the following parameters were also recorded using IRGA; stomatal conductance, and transpiration. IRGA determines the rates of photosynthesis (AN) and transpiration (E), as:

$$AN = (\text{Air flux} \times \Delta\text{CO}_2) / \text{Leaf area}$$

$$E = (\text{Air flux} \times \Delta\text{H}_2\text{O}) / \text{Leaf area}$$

*Corresponding author. E-mail: dominicmenge@yahoo.co.uk.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](#)

Abbreviations: PAR, Photosynthetic active radiation; C₃, carbon parameters; JKUAT, Jomo Kenyatta University of Agriculture and Technology; P_n, photosynthetic rate; IRGA, infrared gas analyzer; E, transpiration; gs, stomatal conductance; Cu²⁺, copper; kg/ha, kilograms per hectare.

Table 1. Selected antifungal plant extracts for the study and parts of the plants used.

| Family | Scientific Name | Common/local name | Parts used |
|---------------|------------------------------|-------------------|--------------|
| Capparidaceae | <i>Boscia angustifolia</i> | Mulule (Kamba) | Leaves, stem |
| Rutaceae | <i>Zanthoxylum chalybeum</i> | Mjafari (Swahili) | Leaves, stem |
| Rutaceae | <i>Melea volkensii</i> | Mukau (Kamba) | Leaves, stem |

From Fick's first law of diffusion, other parameters (stomatal conductance and CO₂ assimilation rate) were then calculated.

RESULTS

Stomata conductance (gs) and transpiration rate

The diurnal changes in gs, rate of transpiration and photosynthetic rate under the antifungal treatments are as shown in Figures 1a and b. The stomatal conductance in Figure 1a followed the same pattern in all the treatments being highest at 9:00 am, dropped at midday and maintained low levels in the late afternoon. However, there were significant differences in stomatal conductance ($P=0.0173$) in the treatments at 9:00 am and was rated as *B. angustifolia* - *Z. chalybeum* ($77.7 \text{ mol/m}^2\text{sec}^{-1}$) combination having the highest stomatal conductance followed by, *M. volkensii* ($46.3 \text{ mol/m}^2\text{s}^{-1}$) and Kocide DF ($39.18 \text{ mol/m}^2\text{s}^{-1}$), respectively. *M. volkensii* plant extract showed lowest stomatal conductance as compared to the other plant extract combination. The stomatal conductance for commercial fungicide (Kocide DF) was significantly lower ($25.6 \text{ mol/m}^2\text{s}^{-1}$) than other treatments at 12:00 pm. *B. angustifolia* - *Z. chalybeum* ($41 \text{ mol/m}^2\text{s}^{-1}$), *M. volkensii* ($41.5 \text{ mol/m}^2\text{s}^{-1}$) and untreated control ($38.3 \text{ mol/m}^2\text{s}^{-1}$) were not significantly different from each other at 12:00 pm. There were no significant differences in stomatal conductance at 15:00 pm of all treatments ($P=0.1235$). This showed that apart from controlling fungal attack the treatments had some influence on stomatal conductance. This behavior was observed in all the four treatments in the three months growth period. Figure 1b shows the diurnal changes in the rate of transpiration in the four treatments. The rate of transpiration was highest at 9:00 am coinciding with highest stomatal conductance and dropped at noon when stomatal conductance also dropped and maintained low levels in the early afternoon and evening when stomatal conductance and PAR were low. There were significant differences ($P=0.003$) in transpiration rates of the treatments at 9:00 am. *B. angustifolia* - *Z. chalybeum* ($2.065 \text{ mol/m}^2/\text{s}$) and *M. volkensii* single plant extracts ($1.353 \text{ mol/m}^2/\text{s}$) had significantly the highest rate of transpiration as compared to other treatments. There were significant differences ($P=0.0015$) in the rates of transpiration among the treatments at 12:00 pm. Kocide DF ($0.76 \text{ mol/m}^2/\text{s}$) had significantly the lowest rate of transpiration while there

were no differences in *B. angustifolia* - *Z. chalybeum* ($1.12 \text{ mol/m}^2/\text{s}$), *M. volkensii* ($1.135 \text{ mol/m}^2/\text{s}$) and untreated control ($1.067 \text{ mol/m}^2/\text{s}$). There were significant differences ($P<0.05$) in transpiration rates of the treatments at 15:00 am. *B. angustifolia* - *Z. chalybeum* ($0.67 \text{ mol/m}^2/\text{s}$), *M. volkensii* ($0.78 \text{ mol/m}^2/\text{s}$) and Kocide DF ($0.77 \text{ mol/m}^2/\text{s}$) treated beans had lower rates of transpiration at 15:00 pm than the untreated control ($1.3 \text{ mol/m}^2/\text{s}$) at 15:00 pm. Generally both the single and combinations of plant extracts had a positive effect on the rate of transpiration. The high positive regressions ($r^2 >0.9$) and the regression equations are summarized in Table 2.

Effect of treatment on CO₂ assimilation and photosynthetic rate (Pn)

Figures 1 and 2 show diurnal changes in photosynthetic (Pn) and CO₂ assimilation rates in the four treatments, respectively. The more the negative CO₂ assimilation the more CO₂ is absorbed from the environment as shown in Figure 2. The CO₂ assimilation reached a peak at 9:00 am and decreased sharply at noon and eventually maintained low levels in the afternoon. CO₂ assimilation followed the same pattern as that of stomatal conductance. There were significant differences ($P<0.001$) in CO₂ assimilation rates among treatments at 9:00 am. *B. angustifolia* - *Z. chalybeum* (577.933 ppm) treated bean plants had significantly lowest CO₂ assimilation rate while there were no differences between *M. volkensii* (679.5 ppm), Kocide DF (641.364 ppm) and untreated control (651.154 ppm) in CO₂ assimilation rate at 9:00 am. There were no significant differences ($P>0.002$) in CO₂ assimilation rate of all treatments at 12:00 pm however they ranged from untreated control (362 ppm) being the highest then followed by *B. angustifolia* - *Z. chalybeum* (328.33 ppm), *M. volkensii* (320.33 ppm) and Kocide DF (304.18 ppm), respectively. Likewise, at 15:00 pm there were no differences ($P=0.1425$) in CO₂ assimilation rates of all treatments. The relationship between stomatal conductance and CO₂ assimilation was described by low insignificant positive regressions in each treatment as shown in Table 3. The low R² indicated the two parameters were very slightly interrelated. The diurnal pattern of rate of photosynthesis among the treatments was the same being highest at the morning, 9:00 am dropped at noon and remained low in the afternoon (15:00 pm). There were significant

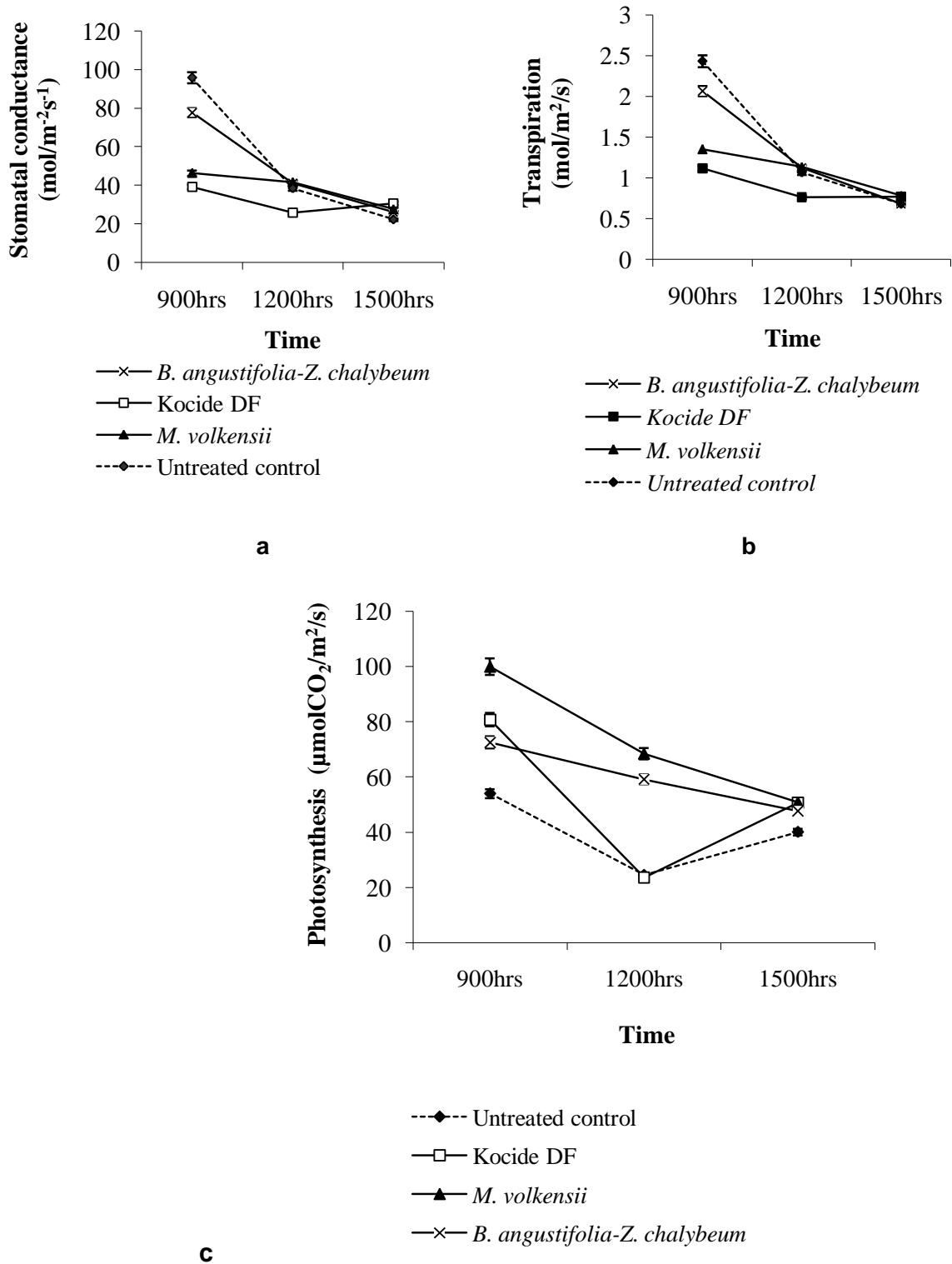


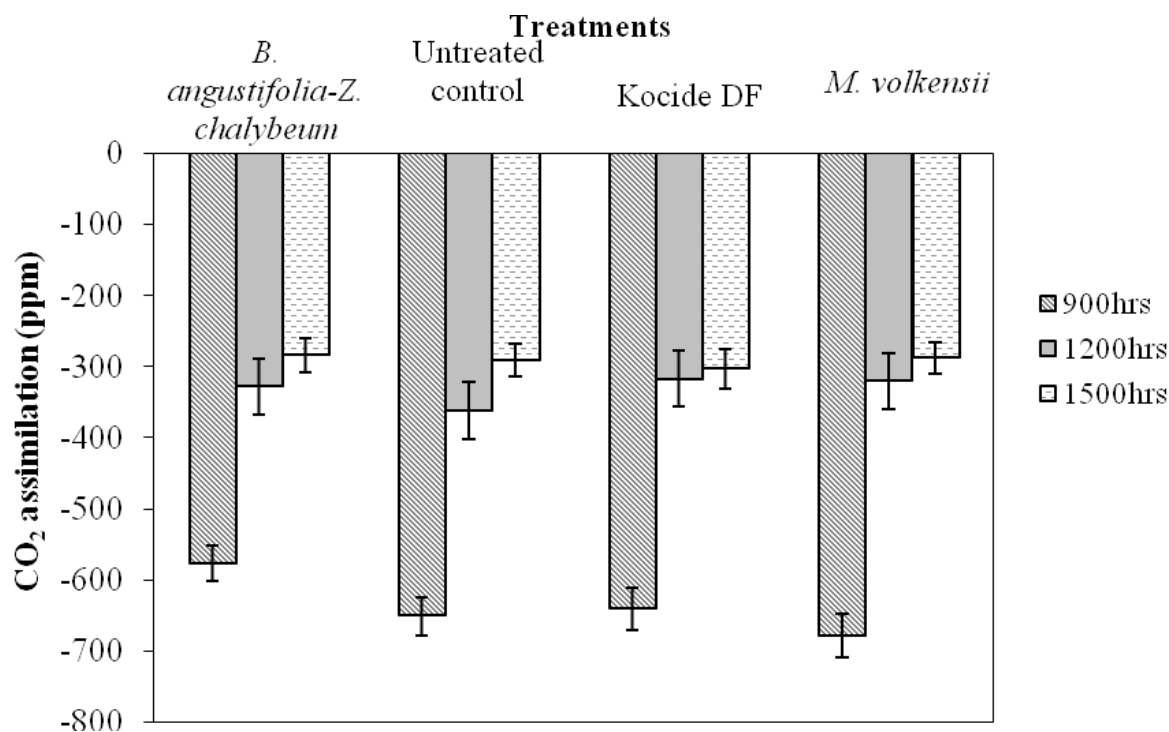
Figure 1. Daily diurnal courses of stomatal conductance (a), rate of transpiration (b) in French beans (Amy variety) exposed to various treatments. Each point represents the mean ± standard error of six replications.

differences (P=0.0021) in the rate of photosynthesis among the treatments at 9:00 am. The rate of photosynthesis was rated highest in *M. volkensis* (99.9

μmolCO₂/m²/s) as compared to the combination *B. angustifolia - Z. chalybeum* (72.5 μmolCO₂/m²/s) and untreated control (53.9 μmolCO₂/m²/s), respectively.

Table 2. The relationship between stomatal conductance and rate of transpiration among the four treatments.

| Treatment | Equation | R ² |
|--|------------------------|----------------|
| <i>B. angustifolia</i> - <i>Z. chalybeum</i> | $y = 44.851x - 10.454$ | 0.9829 |
| Untreated control | $y = 41.604x - 4.882$ | 0.9656 |
| Kocide DF | $y = 38.824x - 3.8036$ | 0.9449 |
| <i>M. volkensii</i> | $y = 37.407x - 0.7395$ | 0.9396 |

**Figure 2.** Daily courses of CO₂ assimilation in French beans exposed to various antifungal plant extracts and a commercial fungicide under natural conditions.**Table 3.** Linear relationships between CO₂ and the rate of photosynthesis.

| Treatment | Equation | R ² |
|--|------------------------|----------------|
| <i>B. angustifolia</i> - <i>Z. chalybeum</i> | $y = 4.0963x + 74.228$ | 0.5873 |
| Untreated control | $y = 0.0369x + 39.852$ | 0.0801 |
| Kocide DF | $y = 0.1134x + 7.1438$ | 0.4250 |
| <i>M. volkensii</i> | $y = 0.0994x + 27.677$ | 0.6596 |

There were significant differences ($P=0.0132$) in the rate of photosynthesis among the treatments at 12:00 pm. *M. volkensii* ($68.38 \mu\text{molCO}_2/\text{m}^2/\text{s}$) had significantly the highest photosynthetic rate at 12:00 pm followed by *B. angustifolia* - *Z. chalybeum* ($59.1 \mu\text{molCO}_2/\text{m}^2/\text{s}$). However, there were no differences between Kocide DF ($23.51 \mu\text{molCO}_2/\text{m}^2/\text{s}$) and untreated control ($24.4 \mu\text{molCO}_2/\text{m}^2/\text{s}$) at 12:00pm. *M. volkensii* (50.77

$\mu\text{molCO}_2/\text{m}^2/\text{s}$) and Kocide DF ($50.7 \mu\text{molCO}_2/\text{m}^2/\text{s}$) revealed significantly the highest rates of photosynthesis although they were not different from each other at 15:00pm. Untreated control ($39.98 \mu\text{molCO}_2/\text{m}^2/\text{s}$) had the lowest photosynthetic as compared to other treatments at 15:00 pm. Generally, *M. volkensii* had a positive effect on rate of photosynthesis as compared to other *B. angustifolia* - *Z. chalybeum*.

DISCUSSION

Effect of treatment on some selected physiological parameters

Generally, French bean leaves showed higher values of stomatal conductance with consequent higher transpiration. The high positive regressions ($r^2 > 0.9$) were obtained in the four treatments. This indicated that stomatal conductance and rate of transpiration were interdependent and it is interpreted to mean that stomatal conductance enhanced rate of transpiration at different times of the day.

The response of stomata to transpiration was used by Monteith (1995a), who re-analyzed 52 sets of published measurements at canopy scale of humidity responses on 16 species of monocots in terms of the relation between stomatal conductance and transpiration. *B. angustifolia* - *Z. chalybeum* combination and single plant treatment *M. volkensisii* had a positive effect on stomatal conductance of bean plants. This suggests that these antifungal plant extracts in general may have interfered with any one of the several biosynthetic pathways or energy production pathways. Commercial control (Kocide DF) had the lowest stomatal conductance of all treatments however; Kocide DF plots had the lowest water loss as compared to others, this indicates they were better at water conservation. Stomata showed a slight opening tendency until 1200 noon, when decreases in stomatal conductance were likely cut down in high transpiration (E) values. Since similar stomatal conductance values were observed during morning, changes in E values suggest that stomatal aperture was more than sufficient to support maximal E values since early hours of morning.

The high regressions between stomatal conductance and rate of transpiration in the four treatments indicated that stomatal conductance and rate of transpiration were interdependent and it was interpreted to mean that stomatal conductance enhanced rate of transpiration at different times of the day. The differences in R^2 values in four treatments were insignificant meaning that concerning these two parameters, the French beans responded to the treatments the same way. This pattern was maintained throughout the growing period. Therefore, the sources of variation in stomatal conductance and the rate of transpiration were treatment, time.

The daily diurnal courses conformed to Zeiger et al. (1981) study which showed that at dawn, stomatal conductance usually increases very rapidly because the entrained rhythm is in correct phase, and also there is a great sensitivity to low photon fluxes of blue light at this time. Stomatal conductance then increases gradually towards a maximum value in late morning or early afternoon before declining noticeably later in the day. This partial closure in the afternoon is thought to be driven by the entrained rhythm, and it is not unusual for

the stomata to be nearly closed before dusk. The responsiveness of stomata to light and CO_2 depends on leaf age and past treatment. As leaves become older, the stomata often become less responsive and may open partly, even at midday. It is difficult to generalize stomatal behavior because so many contradictory reports occur in the literature. Stomatal activity is affected by numerous internal and external factors which often interact in complex ways that sometimes are overlooked by investigators.

Daily course of CO_2 assimilation was similar for all evaluated treatments. In early morning, the sharp increase in photosynthetic photon flux density seems to be the main cause of CO_2 assimilation increase. Considering the highest CO_2 assimilation values, no significant difference was found between treatments under natural condition. Maximal CO_2 assimilation rates were reached around 9.00 am until 12:00 pm when reductions were recorded. Low stomatal conductance is known to cause decrease in CO_2 assimilation values by reducing the CO_2 available, which may be indicated by decreased intercellular CO_2 concentration values (Jones, 1998; Nobel, 1999). Commercial control (Kocide DF) treated plants had the lowest carbon dioxide released as compared to all other treatments because of its low stomata conductance.

Photosynthetic rates (P_n) among the four treatments followed a trend whereby they were at the peak at 9:00 am reducing gradually towards the afternoon and at 15:00 pm. The main sources of variation in the P_n might have been due to treatment and the time of the day. The explanation for the above stated interactions being significant could be that these factors were affecting the photosynthesis rates dependently. The antifungal plant extract had a positive effect on the rate of photosynthesis than other treatments. Therefore, the results suggest that the photosynthetic capacity of 'commercial control (Kocide DF) treated beans' were constrained at natural condition by low stomata conductance. Low stomata conductance in the commercial control (Kocide DF) treated plants might have affected the photosynthetic activity. The inactivation of Rubisco (ribulose-bisphosphate carboxylase/oxygenase) a key-enzyme of Calvin cycle and its two accompanying enzymes, that is, Rubisco activase and carbonic anhydrase under the stress conditions caused by copper and lead (not examined) may be regarded as another possible factor (Vojtechova and Leblova, 1991). This indicated that plant extract treatments were leaf physiology friendly as compared to the copper containing Kocide DF. The untreated control highest transpiration rates might have been caused by high disease severity. Rust caused increased transpiration (E) from infected tissues after sporulation in untreated control. Transpiration before sporulation, which potentially is by a mainly stomatal pathway, is inhibited, probably by stomatal closure; rust is known to inhibit stomatal opening in the light in other

diseases, e.g. bean (*P. vulgaris*) infected by either *U. phaseoli* (Duniway and Durbin, 1971b) or *Uromyces appendiculatus* and this effect has recently been confirmed for Faba bean rust (Tissera, unpublished results). In the present study, it was noted that at each sample time, more variability in transpiration rate occurred in rusted tissue than in healthy tissue. This variability probably occurred because the number of lesions per unit area of leaf was not controlled. Durbin (1978) stated that when sporulation occurred, transpiration from bean leaves infected with rust increased by as much as 50%. Where net photosynthesis was concerned, infection induced opposing changes in the four treatments; net photosynthesis in healthy leaves increased because gross photosynthesis was stimulated and photorespiration was inhibited. Net photosynthesis per plant and ultimately plant growth of the untreated control reduced because infection inhibits the growth of leaf area.

Photosynthesis is closely related to crop growth and yield, and higher photosynthetic rate of leaves is one of the important factors for high crop yield. The results showed that after flowering, the leaves gradually aged, the net Pn, E and gs of leaves gradually declined. Commercial control (Kocide DF) contains copper metal that might have caused low productivity. This could be attributed to its contents that can hamper the process of photosynthesis. It being a micronutrient, copper improves plant growth at natural concentrations. However, at higher concentrations, it also proves very toxic to plants. The phytotoxic effects related to higher concentrations of copper include inhibition of photosynthetic efficiency and as a result reduced crop productivity. The process of photosynthesis was adversely affected by Cu toxicity. Plants exposed to copper formulated fungicide (Kocide DF) showed a decline in photosynthetic rate, which might have resulted from distorted chloroplast structure, restrained photosynthesis of chlorophyll and carotenoids, inhibited activities of Calvin cycle enzymes, as well as deficiency of CO₂ as a result of stomatal closure (Vojtechova and Leblova, 1991).

A strong relationship exists between Kocide DF application and a decrease in photosynthesis and it is believed to result from stomatal closure. Increased rates of respiration and loss of chlorophyll from the leaf tissue apparently were the major factors responsible for the reduction of photosynthetic rates on diseased untreated control leaves.

The photoinhibition mechanism could have a character of photoprotection or represent damaging in PSII reaction centers (Osmond, 1994). The maximum CO₂ assimilation values observed are in agreement with the measurements performed by Souza et al. (2003) in common bean study. Transpiration exhibited similar trend to photosynthesis suggesting that an appreciable part of the inhibition of the two processes is related to increased stomatal resistance as a result of stomatal closure.

Conclusion

B. angustifolia - *Z. chalybeum* combination and single plant treatment of *M. volkensii* had positive effects on enhancing the rate of photosynthesis in bean plants. The high regressions between stomatal conductance and rate of transpiration in the treatments indicated that stomatal conductance and rate of transpiration were interdependent and it was interpreted to mean that stomatal conductance enhanced rate of transpiration at different times of the day. These plant extracts however caused an increase in the rate of transpiration of the bean plants, which resulted in loss of water.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors would like to appreciate all those who are directly or indirectly involved in accomplishing this work. Their special gratitude goes to the supervisors: Dr. Martha Makobe, Dr. Ethel O. Monda and Prof. Paul O. Okemo. Dr. Ethel O. Monda ensured the financial and administrative support. Special thanks go to the members of Botany Department and friends for their tremendous support during the study period.

REFERENCES

- Berger RD, Hau B, Weber GE, Bacchi LMA, Bergamin FA, Amorim L (1995). A simulation model to describe epidemics of rust of *Phaseolus* beans I. Development of the model and sensitivity analysis. *Phytopathol.* 85:715-721.
- De Jesus Junior WC, Do Vale FXR, Coelho RR, Hau B, Zambolim L, Costa LC, Bergamin FOA (2001). Effects of angular leaf spot and rust on yield loss of *Phaseolus vulgaris*. *Phytopathol.* 91:1045-1053.
- Duniway JM, Durbin RD (1971b). "Some effects of *Uromyces phaseoli* on the transpiration rate and stomatal response of bean leaves." *Phytopathol.* 61:114-119.
- Durbin RD (1978). The biochemistry of fungal and bacterial toxins and their modes of action, In Callow IA, ed, *Biochemicid Plant Pathology*. John Wiley & Sons Ltd., Toronto. pp. 137-162.
- Jones HG (1998). Drought tolerance and water use efficiency. In: Smith J. C.A. and Griffiths. H. (eds.), *Water Deficits, Plant Responses from Cell to Community*. BIOS Oxford, UK pp. 193-203.
- Kiswii MT (2009). *Aspergillus flavus* and aflatoxin levels in stored maize in eastern kenya and antifungal activity of some plant extracts. MSc Thesis, Kenyatta University, Kenya. pp. 144.
- Menge DMS, Makobe M, Monda EO, Okemo PO (2011). An *in vitro* and *in vivo* antifungal activity of selected crude extracts against bean rust (*Uromyces appendiculatus*). *Afr. Students J.* 1:76. <http://www.scribd.com/doc/49784524/THE-AFRICAN-STUDENTS-JOURNAL-NO-1>
- Monda EO, Ndegwa A, Munene S (2003). French beans production constraints in Nkuene and Abogeta division of Meru Central district in Kenya. Paper presented at 6th Biennial African Crop Science conference, 12-17 October 2003, Nairobi, Kenya.
- Monteith JL (1995a). Accommodation between transpiring vegetation and the convective boundary layer. *J. Hydrol.* 166:251-263.

- Nobel PS (1999). *Physiochemical and environmental plant physiology*. Academic Press, San Diego, 635p.
- Omwenga OE (2009). Ethnobotanical survey, antimicrobial efficacy and preliminary phytochemical screening of selected anti-diarrhoeal medicinal plants used by the Samburu Community, Wamba, Samburu District, Kenya. MSc Thesis, Kenyatta University, Kenya. pp. 125.
- Osmond CB (1994). What is photoinhibition? Some insights from comparisons of shade and sun plants. In: Baker NR and Bowyer JR (eds) *Photoinhibition of Photosynthesis: From Molecular Mechanisms to the Field*, pp. 1-24. BIOS Scientific Publishers, Oxford.
- Souza TPO, Alzate-Marin AL, Faleiro FG, De Barros EG (2003). Pathosystem common bean-*Uromyces appendiculatus*: host resistance, pathogen specialization, and breeding for rust resistance. *Pest Technol.* 2:56-69.
- Stoll FG (2000). *Management of plant pests and disease, by natural methods*. 2nd edition John Wiley and Sons Press NY.
- Venette JR, Jones DA (1982). Yield losses associated with severity of bean rust (*Uromyces phaseoli*) on pinto beans (*Phaseolus vulgaris* UI-114). *Phytopathol.* 72:794.
- Zeiger E, Farquhar GD, Cowan IR (1981). *Stomatal function*. Stanford University Press, California. 503p.

Short Communication

Fatty acid composition of the seed oil of *Chrysophyllum albidum* (G. Don)

Paul M. Osamudiamen* and Lukman O. Afolabi

Department of Chemical Sciences, Bells University of Technology, Ota, Ogun State, Nigeria.

Received 18 January, 2014; Accepted 26 June, 2014

The fatty acid composition of seed oil of *Chrysophyllum albidum* was determined by gas chromatography-mass spectroscopy (GC-MS). The seed oil contained much important fatty acids with linoleic (26.21%) and palmitic (14.41%) acids being the most abundant unsaturated and saturated fatty acids, respectively. The total unsaturation for the oil was 67.61%. These results confirm that the oil of *C. albidum* is of industrial importance.

Key words: *Chrysophyllum albidum*, fatty acid, seed oil, methyl esters

INTRODUCTION

African star apple (*Chrysophyllum albidum* G. Don) is a tropical edible fruit tree. It belongs to the family of Sapotaceae which has up to 800 species and make up almost half of the order (Ehiagbonare et al., 2008). It is primarily a forest tree species and its natural occurrences have been reported in diverse ecozones in Nigeria, Uganda, Niger Republic, Cameroon and Cote d'Ivoire (Bada, 1997). The plant often grows to a height of 36.5 m though it may be smaller (Bada, 1997). Preliminary studies indicated that the oil was non-drying, based on its iodine value (Table 1) (Osamudiamen and Lukman, 2012). This paper is a report on the fatty acid constituents of the oil.

MATERIALS AND METHODS

Sample preparation

The oil was extracted using soxhlet extractor with n-hexane for 8 h and the fatty acid profile of the seed oil was obtained as fatty acid methyl esters. This was prepared by using commercial aqueous

HCl as described by Ichihara et al. (2010). The reagent was made from 9.7 ml commercial concentrated HCl (35%w/w) diluted with 41.5 ml of methanol and 0.30 ml of the reagent solution were added in this order. The tube was vortexed and then heated at 100°C for 1 h. After cooling, 1 ml of hexane and 1 ml of water were added for extraction of methyl esters in the hexane phase. This was then analyzed with gas chromatography-mass spectrophotometer.

RESULTS AND DISCUSSION

The fatty acid composition of the seed oil investigated revealed that the fatty acid range was from 12:0 to 20:1 (Table 2). The saturated fatty acids comprised of lauric acid (12:0), palmitic acid (16:0) and stearic acid (18:0) while the unsaturated fatty acid comprised of two monoenes namely: vaccenic acid (18:1) and eicosenoic acid (20:1), one diene: linoleic acid (18:2) and one triene: α -linolenic acid (18:3). Linoleic (25.44%) and linolenic acid (26.12%) are the predominant unsaturated fatty acid. Linoleic acid undoubtedly is one of the most

*Corresponding author. E-mail: pmosamudiamen@gmail.com. Tel: +2348073809762.

Table 1. Physicochemical characteristics of *C. albidum* oil.

| Parameter | <i>C. albidum</i> oil |
|--|-----------------------|
| Colour | Dark yellow |
| State at room temperature | Liquid |
| Specific gravity (25°C) | 0.89±0.12 |
| Refractive index (25°C) | 1.66±0.04 |
| Mean molecular mass (g) | 1332.54±0.70 |
| Acid value (mgKOH/g) | 3.56±0.59 |
| Free fatty acid (%) | 1.80±0.21 |
| Saponification value (mgKOH/g) | 126.30±0.70 |
| Iodine value (mg iodine/g) | 31.06±0.70 |
| Peroxide value (MgO ₂ /g oil) | 1.76±0.50 |

Table 2. Fatty acid composition of *C. albidum*.

| Systematic name | Lipid number | Trivial name | Percentage |
|--------------------------------|--------------|------------------|------------|
| Dodecanoic acid | 12:0 | Lauric acid | 0.46 |
| n-hexadecanoic acid | 16:0 | Palmitic acid | 14.41 |
| Octadecanoic acid | 18:0 | Stearic acid | 4.38 |
| Trans-11-octadecanoic acid | 18:1 | Vaccenic acid | 14.73 |
| 9,12-octadecadienoic acid(z,z) | 18:2 | Linoleic acid | 25.44 |
| 9,12,15-octadecatrienoic acid | 18:3 | α-Linolenic acid | 26.12 |
| Cis-13-eicosenoic acid | 20:1 | Paullinic acid | 1.32 |
| Unsaturated acids | | | 67.61 |
| Saturated acids | | | 19.25 |
| Unidentifiables | | | 13.14 |
| Total acid | | | 100.00 |

important polyunsaturated fatty acids in human food because of its prevention of distinct heart vascular diseases (Omode et al., 1995). It is well known that dietary fat rich in linoleic acid, apart from preventing cardiovascular disorders such as coronary heart diseases and atherosclerosis, also prevents high blood pressure (Vles and Gottenbos, 1989). The presence of one of the three essential fatty acids in the seed oils make them nutritionally valuable. Palmitic (14.41%) and stearic acid (4.38%) are also present in high proportion in the oil, which are of nutritional significance

Conclusion

This study has shown that the fatty acid composition of *Chrysophyllum albidum* was predominantly linoleic, linolenic and palmitic acids. The fatty acid profile suggests the possible application of the seed and its oil as a potential industrial resource.

Conflict of Interests

The author(s) have not declared any conflict of interests.

REFERENCES

- Bada SO (1997). Preliminary Information on the Ecology of *Chrysophyllum albidum* (G.Don) in West and Central Africa: In Proceedings of a National Workshop on the Potentials of Star Apple in Nigeria. (eds Denton DA., Ladipo D.O, Adetoro M.A. and Serum MB) pp. 16-25.
- Ehiagbonare JE, Onyibe HI, Okoegwale EE (2008). Studies on the isolation of normal and abnormal seedlings of *Chrysophyllum albidum*: A step towards sustainable management of the taxon in the 21st century. *Sci. Res Essay.* 3(12):567-570.
- Ichihara K, Fukubayashi Y (2010). Preparation of fatty acid methyl esters for gas-liquid Chromatography. *J. Lipid Res.* 51:635-640.
- Omode AA, Fatoki OS, Olaogun KA (1995). Physicochemical Properties of Some Under-Exploited and Non-conventional Oil Seeds. *J. Agric. Food Chem.* 43:2850-2853.
- Osamudiamen PM, Afolabi LO (2012). Physicochemical Characteristics, Proximate and Mineral Compositions of the Underutilized Seed and Oil of *Chrysophyllum Albidum* from Ibadan, Nigeria. *Electron. J. Environ. Agric. Food Chem.* 11(4):351-357.
- Vles RO, Gottenbos JJ (1989). Nutritional Characteristics and Food Uses of Vegetable Oils. In G. Robblen, R. K. Downey, & A. Ashri (Eds.), *Oil crops of the world*, New York, USA: McGraw Hill. pp. 36-86.

Full Length Research Paper

An ethnobotanical study of medicinal plants in Debre Libanos Wereda, Central Ethiopia

Seyoum Getaneh¹ and Zerihun Girma^{2*}

¹Department of Biology, Arba Minch University, P.O. Box 21, Arba Minch, Ethiopia.

²Department of Wildlife and Eco-tourism, Hawassa University, P.O. Box 5, Hawassa, Ethiopia.

Received 18 May, 2013; Accepted 11 June, 2014

An ethnobotanical study of medicinal plants in Debre Libanos Wereda, in central Ethiopia, was carried out from October 2008 to June 2009. A total of 60 informants were interviewed that include knowledgeable farmers, monks, nuns, herbalist farmers and full time herbalists. A total of 83 medicinal plants classified under 77 genera and 46 families were collected. Asteraceae were the most prominent family (7) species and (6) genera, followed by Fabaceae and Lamiaceae that contain four species in three genera each. These plant species were found to be used in treating 50 different types of human and livestock diseases. The majority (77.1%) were wild species whereas 22.9% of the reported medicinal plant species were cultivated in home gardens. Higher numbers of species (46.6%) were harvested for their leaves, followed by roots, seeds and fruit (14.56, 13.59 and 6.80%, respectively). Vast knowledge on the traditional uses of these plants is conveyed from one generation to the next generation through words of mouth. As a result, there is a need for urgent biodiversity conservation of the area and the indigenous traditional ethnobotanical knowledge.

Key words: Ethnobotany, medicinal plant, herbalist, disease, mode of preparation.

INTRODUCTION

Ethnobotany is the study of the relationships between plants and people with a particular emphasis on traditional cultures. The traditional use of plants to fulfill daily needs dates back to the beginning of human civilization and continues to date. Still traditional medicinal plant knowledge is the integral part of culture of many Asian and African countries indigenous community (Subramanyam et al., 2008; Bekalo et al., 2009). In Ethiopia, utilization of medicinal plant remedies in preventing or curing various ailments still plays a

significant role in most parts of the country (Birhan et al., 2011; Giday and Teklehaymanot, 2013; Tolossa et al., 2013). Particularly, traditional herbal healing is widely practiced throughout the rural population as their primary healthcare system (Yineger and Yewhalaw, 2007; Seid and Tsegay, 2011).

There is a high expectation of enormous traditional knowledge and use of medicinal plant species in Ethiopia due to the existence of diverse languages, cultures, beliefs and significant geographical diversity which

*Corresponding author E-mail: zerihun.girma@yahoo.com.

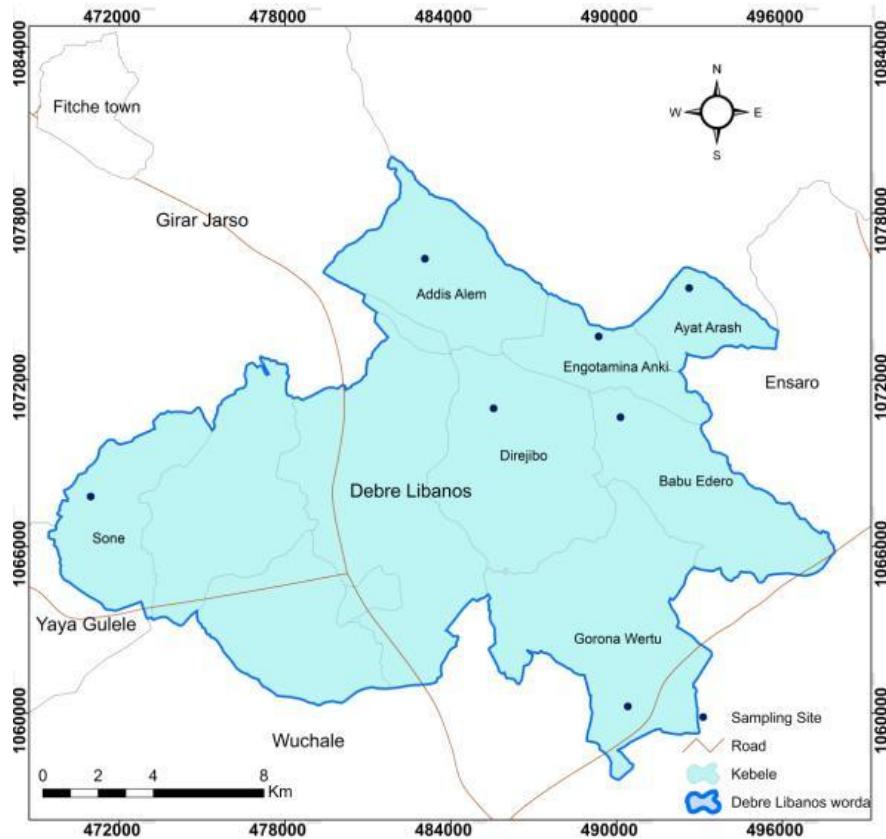


Figure 1. Location map of the study area. Data Source: Ethio-gis collected by ECSA (Ethiopian Central Statistics Authority, 2008). Software ArcGIS 9.2 was also used.

avored the formation of different habitat for medicinal plant (Cunningham et al., 2001). In Ethiopia, it has been estimated that traditional remedies are the most important and sometimes the only source of therapeutics for nearly 80% of the population of which 95% of traditional medicinal preparations are of plant origin (Hamilton, 2003). Much of the knowledge on traditional medicine is available in rural communities. Most of them are perpetuated by word of mouth within family or small community. However, since cultural systems are highly dynamic, these skills are likely to be lost when the communities emigrate to towns or regions, or if the local ecology is significantly changed (Suleman and Alemu, 2012). Furthermore, the high population pressure and its related consequences like increased need for agricultural land, settlement, fuel wood, house construction, and income generation have led to an extreme reduction of medicinal plant in all over their ranges (Bekalo et al., 2009; Belayneh et al., 2012). Teklehaymanot et al. (2006), attempted to study the medicinal plants of the present study area. But, the study covered small area as compared with the total coverage of Dere Libanos Wereda. In addition to this, the study was not able to determine the knowledge difference between the villagers and the Monastery dwellers. From this very fact, the

present study on the use and management of traditional medicinal plants by indigenous people in Debre Libanos Wereda has been initiated to complete the remaining task or to fill the research gap indicated by Teklehaymanot et al. (2006).

MATERIALS AND METHODS

The study area

The study area is located in central part of Ethiopia at about 104 km north of the capital city and situated between $38^{\circ} 05' 01''$ to $38^{\circ} 05' 51''$ E longitude and $9^{\circ} 40' 11''$ to $9^{\circ} 40' 51''$ N latitude (Figure 1). The total area coverage of the study area is 29,776 hectares. The study area encompasses ten peasant associations (rural administrative divisions) and one Administrative town. Interviews were carried out in seven peasant associations (sampling sites) (Figure 1). The total population of the study area is 62,830, of this about 90.4%, that is, 56,798 people live in rural area. The altitudinal range of the study area is between 1500 and 2700 m.a.s.l (Kemal et al., 1996).

The area is characterized by bimodal rainfall during the long rainy season (June to September) and shorter rainy season (March to April). The highest average monthly rainfall was recorded in July (353.99 mm) and the lowest in November (5.5 mm). The daily average maximum temperature was recorded in the month August (17.67°C) and daily average minimum temperature 6.14°C

(Kemal et al., 1996).

The most frequent soil type in the area is blackish and red soil. Black soil being dominant of all constitutes about 56% (16,675 hectare), while the red soil comprises 38% (11,341 ha) and the rest 6% (1,787 hectare) are mixture of different soil type. The soil texture is 63% clay, 27% silt and 10% sand (Kemal et al., 1996). The vegetation of the study area is characterized by, *Acacia-Commiphora* dominate at lower elevations, old remnant afro montane forest in the middle altitudes, and grasslands dominant at higher elevations in the highlands. The old afromontane forest in the middle altitude is owned by Debre Libanos Monastery of Ethiopian Oriental Orthodox Church. The common vegetations in the study area are mostly remnants of trees in agricultural fields, bushes, shrubs and secondary forests. The common plant species of the study area include *Allophylus abyssinicus* (Hochst.) Radlk, *Acacia abyssinica* Benth., *Prunus africana* (Hook.f.) Kalkman, *Juniperus procera* Hochst. ex Endl. and *Olea europaea* subsp. *cuspidata* (Wall. & G.Don) Cif.

Methods

Ethnobotanical data were collected from November 2008 to January 2009. Data were collected through observation, site and informant selection, semi structured interviews, guided field walks with informants, and group discussions to obtain indigenous knowledge of the local community on medicinal plants. Local people were involved during the whole period of investigation as a guide, informants, and as technical assistances. Seven informants were asked to rank five medicinal plants that are used to treat ascariasis. Seven informants were selected from local people for pair wise comparison of medicinal plants used to treat wound.

Two types of interviews were carried out; semi structured interviews and focused group discussion. A total of 60 (50 males and 10 females) informants that include knowledgeable farmers, monks, nuns, herbalist farmers and full time herbalists, were selected. The informants were grouped into three age groups, young (27-35), adult (36-50) and elderly (above 50) to see how the knowledge varies with age. Among them 15 key informants were selected based on the information obtained from knowledgeable elders and local authorities (Development Agents workers and Peasant associations administration leaders). The other 45 informants were selected randomly from the local people of the study area to record general knowledge about plant use. This was done by tossing a coin and using him/her as informant whenever head of the coin face up if he/she volunteered to participate.

Semi-structural interviews were conducted in a place where the informants were most comfortable and during the time they have chosen. Discussion was made with volunteer traditional healers and knowledgeable farmers on the knowledge and use of important medicinal plants. At the time of discussion all informants were allowed to talk freely without interruption.

Plant identification was carried out by voucher specimens collection. Preliminary identification of the collected specimens was made in the field, and then they were dried, deep-frozen and identified in the National Herbarium (ETH), housed in Addis Ababa University using the published volumes of the Flora of Ethiopia and Eritrea and by comparing with authentic herbarium specimens and finally confirmed by assistance of taxonomists.

Data analysis

Data was analyzed and summarized using descriptive statistics such as percentage and frequency. Filter facility was employed to identify the most common ailments in the study area. In addition to this preference ranking, paired comparisons were made for some selected medicinal plants based on methods given by Martin (1995)

and Cotton (1996). In the preference ranking exercise, an integer value (1, 2, 3, 4 and 5) was given, where the most important medicinal plant was given the highest value (5), while the least important is assigned a value of "1". These numbers were summed for all respondents, giving overall ranks to the medicinal plants. Informant consensus factor was also computed to evaluate the reliability of information recorded during the interview using the formula below (Martin, 1995):

$$ICF = \frac{n_{ur} - n_i}{n_{ur} - 1}$$

Where n_{ur} is citations in each category, n_i = number of species used.

RESULTS

A total of 83 medicinal plant species, distributed across 46 families and 77 genera, have been reported to be utilized by the local people in Debre Libanos Wereda as a remedies against various human and livestock ailments (Table 1). Asteraceae appeared as the most prominent family that contains 7 species, within 6 genera, followed by Lamiaceae and Fabaceae (4 species, 3 species). 83 medicinal plant species reported by local peoples in the study area as remedies for human and livestock diseases, 77 species (92.7%) are reported as a remedy for human ailments and 6 species (7.3%) for livestock.

Herbs were the dominant life forms among the reported medicinal plants that contained 32 species (38.6%) followed by shrubs with 30 species (36.1%). Trees (15 species, 18.1) and climbers (6 species, 7.2) were least abundant life forms of medicinal plants recorded from the area. Among the medicinal plant species recorded in the study area, the majority 64 species (77.1%) were wild, whereas 19 species (22.9 %) were cultivated in home gardens.

Plant parts widely used by local people in the study area to treat human and livestock diseases include leaves, root, seed, and fruits. The maximum percentage of the species (46.6%) was harvested for their leaves followed by roots, seed and fruit (14.56, 13.59 and 6.80%) respectively as a source of remedies. Other parts consist of only 18.45% (Table 1).

A significant percentage (54.3%) of the medicinal plant was used in fresh form for remedy preparations. Relatively few medicinal plants (34.9 %) were reported to be used in dried form and the remaining very few medicinal plants were reported to be used either fresh or dried (10.8%). Very common methods of remedy preparation in the study area were reported to be through crushing or pounding the usable part by using wooden or stone made material that cover 50.64% followed by squeezing (12.84%), chewing (11.0%), decoctions (7.34%), covering (6.42%), chopping (3.67%) and smoke (1.84%) (Figure 2). Local peoples also used additives such as butter, edible oil for wound and skin disease, and they used coffee, honey, and local beverages like

Table 1. Plant species used for treatment of human and livestock diseases and their mode of preparation and application in Debre Libanos Wereda, Ethiopia.

| S/N | Botanical name | Family name | Local name | Disease treated | Mode of preparation and application |
|-----|---|----------------|---------------|--|--|
| 1 | <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Girar | Goat intestinal parasite Nose bleeding | Crush the seed together with fruit and make a juice by mixing with cold water; give to the infected animal as a drink Squeeze flashy leave and drop to nostrils. |
| 2 | <i>Achyranthes aspera</i> L. | Amaranthaceae | Tilenj | Anthrax Eye | Crush the leaves and mix with fruit of <i>Cucumis ficifolium</i> and make a juice with cold water then give the cattle as a drink. Squeeze the leave and drop the fluid in to the eye. |
| 3 | <i>Allium cepa</i> L. | Alliaceae | Keye shinkurt | Asma malaria | Squeeze the bulb and take one tea spoon every morning Chew the bulb and swallow it |
| 4 | <i>Allium sativum</i> L. | Alliaceae | Nech shinkurt | Common cold Snake bit Abdominal pain | Crush the bulb and swallow it. Additionally, insert the bulb in the nostrils. Crush the bulb and put it on the site of bites and tie it Crush the bulb and mix with honey take a tea spoon each morning. |
| 5 | <i>Allophylus abyssinicus</i> (Hochst.) Radlkofer | Sapindaceae | Enbis | Skin itching | Squeeze the leave and rub on the skin |
| 6 | <i>Amaranthus caudatus</i> L. | Amaranthaceae | Bahr tef | Intestinal disorder Diarrhea | Crush the seed and mix with black teff flour then make a bread to eat. Squeeze the leaves and make a juice |
| 7 | <i>Artemisia rehan</i> L. | Asteraceae | Arity | Diarrhea | Boil leaves with water and dink half cup of coffee the hot decoction every day |
| 8 | <i>Asparagus africanus</i> Lam. | Asparagaceae | Serity | Abdominal pain. Tooth ache | Fresh root of this plant is chewed and swallowed Chew the root and hold it near the infected teeth. |
| 9 | <i>Bersama abyssinica</i> Fresen. | Melanthaceae | Azamir | Ascaris | Fresh leaves are boiled in water and drunk for three consecutive days. |
| 10 | <i>Brassica oleracea</i> L. | Brassicaceae | Yabesha gomen | (Stomack burn | Boil the leaves and mixed with oil of <i>Nigela sativa</i> L. and eat with injera. |
| 11 | <i>Brusea antidysenterica</i> J.F.Mill. | Simaroubaceae | Abalo | Wound and Skin itching | Crush leaves and mix with butter then cover the wound |
| 12 | <i>Buddleja polystachya</i> Fresen. | Loganiaceae | Anfar | Eye disease | Squeeze the leave and drop on the infected eye |
| 13 | <i>Calotropis procera</i> (Ait.) Ait.f. | Asclepiadaceae | Kobo | Wound | Squeeze fresh leave and pour the content on wound, or put the leave on fire then cover the wound with fired leave. |
| 14 | <i>Calpurnia aurea</i> (Alt.) Benth. | Fabaceae | Digita | Abdominal pain External parasite | Crush the leave and mix with coffee powder then make a juice with water. Chop the leave and put it in water for few days then wash the affected site. |

Table 1. Contd.

| | | | | | |
|----|---|---------------|------------------|---|--|
| 15 | <i>Capparis tomentosa</i> Lam. | Capparidaceae | Gumero | Spiritual disorder For any sudden ailment | Crush the root and put it on fire and smoke the bedroom of the patient Crush the root and mix with Tela then a glass of the mixture is given as a drink. |
| 16 | <i>Carica papaya</i> L. | Caricaceae | Papaya | Amoeba | Crush seeds and mix with honey and water then drink the juice each morning for a week. |
| 17 | <i>Carissa spinarum</i> L. | Apocynaceae | Agam | Spiritual disorder (unable to sleep at night) Evil eye | Make a powder from the root and mix with water, put a spoon full in to a cup of coffee and drink Smoke the root all over the body of the patient |
| 18 | <i>Catha edulis</i> (Vahl.) Forssk. ex Endl. | Celastraceae | Chat | Coughing | Boil the leave and stem with water then add honey and then set it aside to get cooled. At last drink a glass of cold mixture. |
| 19 | <i>Citrus limon</i> (L.) Burm.f. | Rutaceae | Lomi | Asma | Boil the leave of <i>Citrus limon</i> together with stem of sugarcane then drink hot decoction. |
| 20 | <i>Clematis simensis</i> Perr and Guill | Ranunculaceae | Yeazo hareg | Wound | Leaf of <i>Clematis simensis</i> are crushed, smashed and tied on wound. |
| 21 | <i>Clerodendrum myricoides</i> (Hochst.) R.Br.ex Vatke. | Verbenaceae | Misireg | Spider poison | Crush leaves and mix with butter then rub on the affected skin. |
| 22 | <i>Cordia africana</i> Lam. | Boraginaceae | Wanza | Gastric ulcer | The seed is Chewed and swallowed |
| 23 | <i>Croton marcostachyus</i> Del. | Euphorbiaceae | Bisana | Skin disease | The leaves are squeezed and the content is dropped on the infected site. |
| 24 | <i>Cucumis ficifolium</i> A.Rich. | Cucurbitaceae | Yemdir enboay | Wound Snake bite | Roots are Crushed and mixed with butter are put on wound. Chewing the root and swallowing the juice only |
| 25 | <i>Cucurbita pepo</i> L. | Cucurbitaceae | Duba | Tape worm | Roost the seed and eat |
| 26 | <i>Datura stramonium</i> L. | Solanaceae | Astenagir | Dandruff Wound | Crush leaves and seed of this plant mix with butter then apply the paste to affected area (head). Leaves are crushed and applied to affected area. |
| 27 | <i>Diplolophuium africanum</i> (Turez.) | Apiaceae | Feres zeng | Ascariasis | Squeeze leaves, mix with water and drink (for children) |
| 28 | <i>Dodonaea angustifolia</i> L.f. | Sapindaceae | Kitkita | Wound | Leaves are crushed and applied on wound |
| 29 | <i>Dorstenia barnimiana</i> Schweinf. | Moraceae | Work bameda | Leprosy | Crush the root, mix with seeds of <i>Lepidium sativum</i> L. and sorghum flour and then extract local alcohol (Areke) from it, then the patient drinks it until he/she recovers. |
| 30 | <i>Dovyalis abyssinica</i> (A.Rich.) Warb. | Flacortiaceae | Yabesha Qoshm | Hypertension Acariasis Bleeding gum | Root and stem tuber are smashed, mixed with 'Tela' and drink Boil seeds with water and drink. Eat fresh fruit |
| 31 | <i>Ekebergia capensis</i> Sparmm. | Meliaceae | Sembo | Skin rush | Crush leaves and mix with butter and apply on the site of infection. |

Table 1. Contd.

| | | | | | |
|----|---|---------------|---------------|---|---|
| 32 | <i>Eucalyptus globulus</i> Labil | Myrtaceae | Nech bahirzaf | Sudden physiological change | Boil leaves in water and inhale the vapor |
| 33 | <i>Euphorbia abyssinica</i> J.F.Gmel. | Euphorbiaceae | Qulqual | External parasite Wound | The latex (milky juice) is applied to affected area. Milky juice(latex) is applied on wound |
| 34 | <i>Ferula communis</i> L. | Apiaceae | Inslal | Hypertension Unable to urinate | Crush leaves add to boiled tea and drink. Crush leaves mix with water and drink. |
| 35 | <i>Ficus vasta</i> Forssk. | Moraceae | Warka | Constant lose of weight in cattle | Chop the bark boil it in water pour to the moth of the cattle |
| 36 | <i>Ficus palmata</i> Forssk. | Moraceae | Beles | Skin rush | Crushed leaves are mixed with butter then the paste is applied on the affected site. |
| 37 | <i>Glinus lotoides</i> L. | Aizoaceae | Metere | Tooth ache | Chew the root and hold tightly near the infected teeth. |
| 38 | <i>Glycine wightii</i> (Wight & Arn.) Verdc. | Fabaceae | Yelam chew | Tape worm | Crush seed and mix with Lin seed and then eat it. |
| 39 | <i>Grewia flavescens</i> Juss. | Tiliaceae | Lenquata | Constant lose of weight in cattle) | Crush fresh seed and leaves make a juice like fluid pour to the moth of the cattle. |
| 40 | <i>Helinus mystacinus</i> (Ait.) E. Mey.ex Steude. | Rhamnaceae | Shnbirit | Bloating | Crush leaves mix with water and drink. |
| 41 | <i>Jasminum floribundum</i> R.Br | Oleaceae | Tembele | 'Globa' (Cattle disease) | Leaves and stem are Crushed and boiled in water and put aside until it gets cooled. Then pour in to the mouth of the cattle. |
| 42 | <i>Laggera tomentosa</i> (Sch. Bip.ex A. Rich.) Oliv. & Hiern | Asteraceae | Nech kese | Eye treatment | Squeeze leave and put dropsthrough ear. |
| 43 | <i>Leonotis ocymifolia</i> (Burm.f.) Iwarsson | Lamiaceae | Eras kimir | Tape worm | Leaves are crushed, mixed with water and then the patient will take a glass of the mixture as a drink. |
| 44 | <i>Lepidium sativum</i> L. | Brassicaceae | Feto | Common cold | Hold fresh leaves tightly in to the nostrils |
| 45 | <i>Linum usitatissimum</i> L. | Linaceae | Telba | Abdominal pain | Squeeze leaves add in a cup of coffee and drink. |
| 46 | <i>Lycopersicon esculentum</i> Mill. | Solanaceae | Timatim | Abdominal pain | Crush seed and mix with other dishes ('Injera') |
| 47 | <i>Maesa lanceolata</i> Forssk. | Myrsinaceae | Kelawa | Intestinal wound | Crush the seeds in to powder and mix with water then drink a glass of juice before food each morning until recovery |
| 48 | <i>Momordica foetida</i> Schumach. & Thonn. | Cucurbitaceae | Ababure | Weak feeling | Squeeze the fruit and mix with <i>Allium cepa</i> L. Mix the juice in a cup of tea and drink it every morning |
| | | | | Skin disease | |
| | | | | Characterized by itching and black spots | Crush seed and mix with butter then apply on infected site. |
| | | | | Swelling wound | Roots are crushed mixed with butter and applied on the affected site. |

Table 1. Contd.

| | | | | | |
|----|--|----------------|--------------|--|---|
| 49 | <i>Myrsine africana</i> L. | Myrsinaceae | Kechemo | Endoparasites (Tape worm and Ascaris) | Crush fruits and make a juice then drink. |
| 50 | <i>Myrtus communis</i> L. | Myrtaceae | Ades | Headache | Crush leaves and boil with water then drink with cup. |
| 51 | <i>Nicotiana tabacum</i> L. | Solanaceae | Timbaho | To remove leeches from cattle mouth | Crush dry leaves mix with water and give it to cattle as a drink. |
| 52 | <i>Ocimum lamiifolium</i> Hochst. ex Benth. | Lamiaceae | Damakese | Headache and cough | Leaves are Squeezed and drunk alone or with coffee. |
| 53 | <i>Osyris quadripartita</i> . Decn. | Santalaceae | Keret | Abdominal pain Urine problem | Chewing the stem and swallowing the fluid only. Crush and boil with water and drink. |
| 54 | <i>Otostegia integrifolia</i> Benth. | Lamiaceae | Tungut | Abdominal pain Evil eye | Crush laves mix with water and drink. Burn stem and leaves and in hale the smoke |
| 55 | <i>Otostegia fruticosa</i> (Forssk.)Schweinf.ex Penzig | Lamiaceae | Geram tungut | Tonsillitis | Squeeze leaves while adding few drops of water and drink the juice. |
| 56 | <i>Pentas schimperiana</i> | Rubiaceae | Ese zeye | Snake bite | Chewing the root and swallowing only the liquid part of root. |
| 57 | <i>Phytolacca dodecandra</i> L.Herit. | Phytolaccaceae | Indod | Abortion Toothache | Crush the root and mix with water and drink. Chew the stem and hold it tightly to the infected teeth. |
| 58 | <i>Premna schimperi</i> Engl. | Verbenaceae | Chocho | Eye disease | Squeeze fresh leaves and drop a drop of the extract on the affected eye. |
| 59 | <i>Prunus persica</i> (L.) Batsch. | Rosaceae | Kok | Appetite | Eat fruits |
| 60 | <i>Pterolobium stellatum</i> (Frossk.) Brenan | Fabaceae | Kentafa | Goiter | Crush leaves mix with butter then apply the paste and tie it and cover it |
| 61 | <i>Rhus retinorrhoea</i> A.Rich. | Anacardiaceae | Tilem | Liver infection | Roots of <i>Rhus retinorrhoea</i> crushed together with flower of <i>Catha edulis</i> and (<i>Rumex nervosus</i>) and mixed with water and add small amount salt then drink |
| 62 | <i>Ricinus communis</i> L. | Euphorbiaceae | Gulo | Eye disease | Leaves are slightly heated and applied on the affected part of the eye |
| 63 | <i>Rosa abyssinica</i> Lindley. | Rosaceae | Kega | Gastric | Eat fruit and seeds |
| 64 | <i>Rumex nervosus</i> Vahl | Polygonaceae | Enboacho | Anti-bleeding | Crush dried leaves in to powder and apply on the cut. |
| 65 | <i>Rumex abyssinicus</i> Jacq | Polygonaceae | Mekemeko | Hypertension | Crush root in to powder mix with bubs of <i>Allium sativum</i> add the mixture in to boiled water and drink the hot decoction in a cup |
| 66 | <i>Rumex nepalensis</i> Spreng. | Polygonaceae | Tult | Diarrhea | Crush root mixed with water and drink |

Table 1. Contd.

| | | | | | |
|----|--|------------------|------------------|-------------------------------|--|
| 67 | <i>Ruta chalepensis</i> L. | Rutaceae | Tenadam | Abdominal pain | Stem and leaves are crushed and added in to a cup of tea or coffee and drunk or chewing fresh stem and leaves and swallowing |
| 68 | <i>Schinus molle</i> L. | Anacardiaceae | Kundo berbere | Tuberculosis | Crushed seeds are mixed with honey and then eaten. |
| 69 | <i>Sesamum angustifolium</i> (Oliv.) Engl. | Pedaliaceae | Selit | Ear defect | Extract oil from the seed and drop the extract it the ear |
| 70 | <i>Solanecio gigas</i> (Vatke) C.Jeffrey. | Asteraceae | Yeshe koko gomen | Skin disease | Leaves are crushed and pasted on affected body |
| 71 | <i>Solanum anguivi</i> Lam. | Solanaceae | Dekak enboay | Skin disease (itching) | Dry seeds are crushed, mixed with oil and applied on the site. For fresh seeds no need of using oil. |
| 72 | <i>Steganotaenia araliaceae</i> Hochst. | Apiaceae | yegibmrkuz | To remove leaches from cattle | Crush leaves boil with water then pour in to the mouth of the cattle |
| 73 | <i>Stephania abyssinica</i> (Dillon. et A.Rich.) Walp. | Menispermaceae | Yayit hareg | Wound | Roots are Crushed and mixed with milk and applied on wound. |
| 74 | <i>Trigonella foenum-graecum</i> L. | Fabaceae | Abish | Abdominal pain | Crush seeds and make a juice by mixing it with water then add hone to drink |
| 75 | <i>Verbascum sinaiticum</i> Benth | Scrophulariaceae | Yahiya joro | Skin disease | Leaves are crushed and mixed with water wash the affected part with the mixture and tie with the soaked leave. |
| 76 | <i>Verbena officinalis</i> L. | Verbenaceae | Atuch | Abdominal pain | Dry roots are crushed and mixed with water and drunk. For fresh root, chew and swallow only the liquid part of root. |
| 77 | <i>Vernonia amygdalina</i> Del. | Asteraceae | Girawa | Abdominal pain | Crush leaves in to powder and mix with water then drink. |
| 78 | <i>Vernonia bipontini</i> Vatke | Asteraceae | Gobez tekes | Toothache Eye disease | Chew leaves and hold it close to the infected teeth. Squeeze leaves and drop one or two drops of the extract on the eye |
| 79 | <i>Withania somnifera</i> (L.) Dunal | Solanaceae | Gizaw | Spiritual disease. | Smoke the entire body of the patient with dried leaves |
| 80 | <i>Xanthium strumarium</i> L. | Asteraceae | derkus | Skin rash | Leaves are crushed and mixed with butter and applied on the infected site. |
| 81 | <i>Ximenia caffra</i> Sond. | Olacaceae | Atat | Snake bite | Take seven leaves of this plant and chew it well then swallow the juice |
| 82 | <i>Zehneria scabra</i> (Linn.f.)Sond | Asteraceae | Hareg ressa | Wound | Leaves are crushed and mixed with oily substance and applied on the infected site. |
| 83 | <i>Zingiber officinale</i> Rosc. | Zingiberaceae | Zingible | Abdominal pain | Direct eating or crushed and mixed with tea and drunk. |

'Tela' and 'Areke' for those plants having bitter taste.

The most common mode of administration in the study area was oral that cover 49.4%. Most of the

remedies prescribed by traditional healers are applied in various ways such as drinking like a

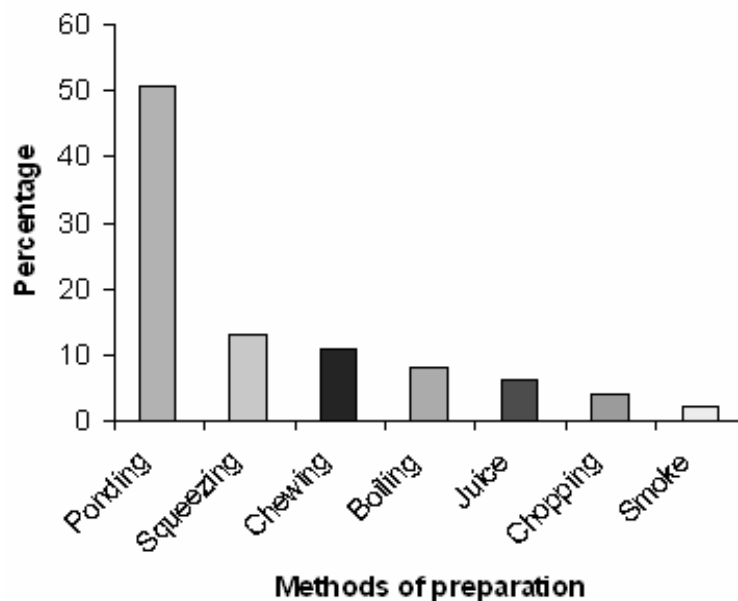


Figure 2. Mode of preparation of medicinal plants.

Table 2. Informant consensus factor of medicinal plants of Debre Libanos Wereda, Ethiopia by categories of diseases.

| Categories (diseases) | No of species | Percentage of total species | No. of informant | Percentage of no. of informant | ICF (%) |
|---|---------------|-----------------------------|------------------|--------------------------------|---------|
| Skin diseases and wounds | 21 | 25.3 | 45 | 19.7 | 54.5 |
| Respiratory infections and cough | 5 | 6 | 11 | 4.8 | 60 |
| Intestinal parasites, abdominal pain, diarrhea. | 14 | 16.9 | 52 | 22.8 | 74.5 |
| Malaria and hypertension | 4 | 4.8 | 8 | 3.5 | 57.1 |
| Snake bites and spider poisons | 5 | 6 | 9 | 4 | 50 |
| Common cold, headache and sudden physiological change | 7 | 8.4 | 55 | 24 | 88.9 |
| Spiritual disorder | 5 | 6 | 6 | 2.6 | 20 |
| Swelling and bleeding | 7 | 8.4 | 9 | 4 | 25 |
| Organs disease (eye, teeth, kidney, liver) | 10 | 12 | 18 | 7.9 | 47.1 |
| Cattle disease (bloating anthrax, leaches and external parasites) | 5 | 6 | 15 | 6.6 | 71.4 |

juice, take a drop of squeezed plant part or chewing and swallow the liquid part only, etc. Dermal is the second most important route of administration of traditional medicine which covers 33.7%. There are various ways of dermal application of traditional medicine. For example, they may apply as a form of paste, coated and tie or crushed the plant part and put the powder on the affected part and so on. Only few medicinal plants were reported to be administered through oral and nasal, eye, nose and ear (about 16.9%).

Almost all traditional healers in the study area do not have sufficient knowledge on dosages. An ethnobotanical data in this study showed that all administrations are not standardized. Healers determine the dosages based on age, physical appearance and

strength of the disease. Children are given small doses of medicine than considered in case of adult patient. Small amount such as drops, hand palms, coffee cups are applied as small dosage. For larger dosages, they use water glasses or other local materials that are used for drinking.

The highest percent informant consensus factor (% ICF) value was obtained with problems associated with common cold, headache and sudden physiological change (febrile illness characterized by fever, headache, skin rash and muscle spasm) (88.9%) followed by problems related to intestinal parasites, abdominal pain, diarrhea (74.5%). The lowest %ICF value was obtained in diseases associated with spiritual disorder (20%) (Table 2).

Table 3. Informant consensus.

| Botanical name of medicinal plant | No. of informants | Percentage |
|-----------------------------------|-------------------|------------|
| <i>Glinus lotoides</i> | 60 | 100 |
| <i>Ocimum lamiifolium</i> | 60 | 100 |
| <i>Ruta chalepensis</i> | 60 | 100 |
| <i>Zingiber officinale</i> | 60 | 100 |
| <i>Allium sativum</i> | 55 | 91.7 |
| <i>Croton macrostachyus</i> | 50 | 83 |
| <i>Brucea antidysenterica</i> | 49 | 81.7 |
| <i>Vernonia amygdalina</i> | 45 | 75 |
| <i>Lepidium sativum</i> | 38 | 63 |
| <i>Cucumis ficifolius</i> | 35 | 58 |

Table 4. Preference ranking of medicinal plants of Debre Libanos Wereda, Ethiopia used to treat Ascariasis.

| Lists of medicinal plants | Informant labeled from R1-R7 | | | | | | | Total | Rank |
|------------------------------------|------------------------------|----|----|----|----|----|----|-------|------|
| | R1 | R2 | R3 | R4 | R5 | R6 | R7 | | |
| <i>Ruta chalepensis</i> L. | 3 | 4 | 4 | 3 | 3 | 4 | 3 | 24 | 3 |
| <i>Malva verticillata</i> L. | 5 | 5 | 4 | 4 | 5 | 5 | 5 | 33 | 1 |
| <i>Diplophium africanum</i> Turcz. | 4 | 3 | 3 | 2 | 1 | 2 | 3 | 18 | 5 |
| <i>Vernonia amygdalina</i> Delile | 3 | 3 | 4 | 3 | 3 | 3 | 2 | 21 | 4 |
| <i>Lepidium sativum</i> L. | | 4 | 5 | 5 | 4 | 4 | 4 | 31 | 2 |

The study revealed that some medicinal plants are well known for their medicinal value among the herbalist and the local community at large than the other. As a result, local informants cited such plants repeatedly as a remedy of different diseases. For example, *Glinus lotoides* L., *Ocimum lamiifolium* Hochst. ex Benth and *Ruta chalepensis* L. were cited by 60 informants (100%) as a source of remedy for tapeworm, sudden physiological change, and abdominal pain respectively (Table 3). *Allium sativum* L. was cited by 55 informants (91.7%) as a remedy for malaria, common cold and other illness while *Croton macrostachyus* Hochst. ex Delile was cited by 50 informants (83%) for gastric ulcer. On the other hand, *Lepidium sativum* L. and *Cucumis ficifolius* A.Rich. were cited as the least commonly used medicinal plants by the informants (58%) (Table 3).

Preference ranking was made for five most important medicinal plants that are used to treat abdominal problems associated with ascariasis. Among the five selected medicinal plants *Malva verticillata* L. stands first followed by *L. sativum* L. (Table 4).

The result of pair wise comparison indicates that *Brucea antidysenterica* J.F.Mill. followed by *Calotropis procera* (Aiton) Dryand. and *Stephania abyssinica* (Quart.-Dill. & A.Rich.) Walp. ranked first, second and third, respectively for the treatment of wound. The other two less preferred medicinal plants for wound treatment were *Clematis simensis* Fresen. and *Verbascum sinaiticum* Benth. (Table 5).

DISCUSSION

The number of reported medicinal plants (83 species, 77 genera and 46 families) and their uses by the community witness show how rich the area is in terms of medicinal plants diversity and the depth of the local indigenous knowledge on medicinal plants and their applications. A number of studies elsewhere in Ethiopia have reported similar number of species of medicinal plants as traditional medicines against human and livestock alignments (Yinger and Yehwalahu, 2007; Bekalo, 2009; Tolosa et al., 2013). The relatively higher number of traditional medicinal plant species documented from the study area is mainly attributed to strict conservation of forest resources of the monastery. The monastery harbors various medicinal plants including those medicinal plants that have been widely used long time ago. The monastery has played a key role in conserving the remnant afro montane forest which is a source of most medicinal plant species, even though any plant in the monastery is strictly forbidden to be used as traditional medicine. Furthermore, the acceptance of folk medicine and the limited access to public healthcare services in the community may be factors contributing to the knowledge of medicinal species in local medical practices. However, the traditional systems and religious beliefs that generally restrict the way of transferring indigenous knowledge might have constrained, to some extent, the free flow of information on medicinal plants in

Table 5. Pair wise comparison of medicinal plant of Debre Libanos Wereda, Ethiopia used to treat wound.

| Medicinal plant | Informant labeled from R1-R7 | | | | | | | Total | Rank |
|--|------------------------------|----|----|----|----|----|----|-------|------|
| | R1 | R2 | R3 | R4 | R5 | R6 | R7 | | |
| <i>Brucea antidysenterica</i> J.F.Mill. | 4 | 5 | 4 | 5 | 5 | 4 | 5 | 32 | 1 |
| <i>Calotropis procera</i> (Aiton) Dryand. | 4 | 5 | 5 | 5 | 4 | 4 | 3 | 30 | 2 |
| <i>Clematis simensis</i> Fresen. | 3 | 4 | 2 | 3 | 5 | 3 | 4 | 24 | 4 |
| <i>Stephania abyssinica</i> (Quart.-Dill. & A.Rich.) | 4 | 3 | 5 | 4 | 3 | 4 | 5 | 28 | 3 |
| <i>Verbascum sinaiticum</i> Benth. | 4 | 3 | 4 | 2 | 3 | 4 | 2 | 22 | 5 |

this study. Secondly, it could be attributed to the scope of the study that attempted to study the medicinal plants in there agro ecological zones, lowland, middle elevations and highlands unlike Teklehaymanto et al. (2006) that confined his study only in the monastery and its immediate vicinity.

The trees and shrubs constitute more than 70% of the traditional medicinal plant in the study area. This could be attributed to various factors. First it can be related with the floristic composition of the vegetation of the area, which is dominated by herbs and exotic *Eucalyptus* sp. in the high lands and *Acacia-commiphora* woodland in the low lands. Secondly, a high usage of herbs could be an indication of their abundance, since the area receives relatively high amount of rain fall that fosters the flourishing of herbs. Thirdly, the affinity to use herbs as traditional medicinal plant could be attributed to strong bioactive compounds. Studies in various parts of the world have raveled that herbs contain phytochemicals like alkaloids and falvanoids that have strong anti bacterial and anti fungal effects (Legesse et al., 2011). A number of studies carried out elsewhere in Ethiopia have documented that herbs and shrubs are plant species mostly used by indigenous communities of Ethiopia as treatment against various human and livestock ailments (Teklehymanot et al., 2006; Bekalo et al., 2009; Mesfine et al., 2009). For example, Konta people use more of herbs (about 68 species) than trees (20 species) (Bekalo et al., 2009) in a similar pattern as reported from India, where about 19 out of 54 species were herbs and shrubs were about 12 species (Ayyanar and Ignacimuthu, 2005). More than half of the Zay plant remedies were also obtained from herbs (Giday et al., 2003)

Most of medicinal plants (75%) utilized by local people of the study area are collected from wild; only few (25%) are harvested from home garden. There is little tradition or practice by local people to cultivate medicinal plants. Plants were harvested and processed only when needs aroused. The use of uncultivated plants is a common practice in Ethiopia (Giday and Ameni, 2003) and this has been creating an additional pressure on the populations of wild plants besides enviromental degradation and deforestation. For example, similarly Gebre (2005) found that about 76.5% of the remedies were reported from wild. The same author also noted that most of the medicinal plants are under the threat as long as the

destruction and fragmentation of wild habitat continues. This is also true in the present study area. Furthermore, Asfaw and Woldu (1997) reported that only 6% of the plants maintained in home gardens in Ethiopia are primarily cultivated for their medicinal value even though many other plants grown for non-medicinal purposes are used for preparation of medicines.

In the present study, the leaf is one of the most extensively used plant parts in preparation of traditional herbal medicine followed by root and seed. The common use of leaf in the preparation of remedies could partly be due to the relative ease of finding this plant part. The practice of using leave part for remedies preparation helps to reduce the rate of threat on plant species or helps for sustainable harvesting of plants since removal of an appreciable amount of leaf is tolerated by the plant. Roots appeared also to be the second most plant part commonly used by the healers in the current study area. This could be associated with the fact that roots remain in the soil and are easily available, even during the long dry seasons in arid and semi-arid areas. But, harvesting roots for medicinal value could possibly put a strain on the survival of the plant since aerial parts of the plant are highly dependent on underground parts (root) for physical support and physiological process. In agreement with our study, similar studies in other parts of Ethiopia reported that roots and leaves are indeed the most commonly used medicinal plant parts (Bekalo et al., 2009; Bussmann et al., 2011). Inspection of the results on number of preparations and plant parts used may lead to the conclusion that harvesting medicinal plants for use in traditional medicine is not destructive to the natural vegetation of the study area since leaves are the most frequently sought parts of the plant. On the other hand, it may also lead to the conclusion that harvesting of medicinal plants is likely to be destructive because the second most frequently used part is the root.

According to the present survey, a significant number (56.88%) of the medicinal plants were to be used in fresh form in remedy preparations. This indicates that local people of the study area are highly dependent on fresh remedies that may put medicinal plants under serious threat, since there is no habit of preservation or storage plant parts for later use. Ethnobotanical studies of medicinal plants elsewhere in Ethiopia have documented the same mode of preparation (Seifu, 2004; Gebre, 2005;

Amenu, 2007; Beyene, 2007).

Very dominate methods of remedy preparation in the study area were reported to be through crushing (grinding) followed by squeezing, and chewing. Local people also used additives such as butter, edible oil for wound and skin diseases, and cup of coffee, honey, and local beverages like 'Tela' and 'Areke' for those plants having bitter taste. This finding is consistent with some reports elsewhere in Ethiopia (Tekelhaymanot and Giday, 2007; Bekalo et al., 2009; Flatie et al., 2009) but disagrees with some reports where other methods of remedy preparation are employed (Abebe and Ayeahu, 1993; Yirga 2010). It is likely that these differences are associated with the differences in culture and knowledge in different socio-cultural groups.

The choice of oral administration may be related to the use of some solvents or additives (butter, edible oil, coffee, honey and local beverages like 'Tela' and 'Areke') that are commonly believed to serve as a vehicle to transport the remedies. The additives are also important to minimize discomfort, improve the taste and reduce adverse effects such as vomiting and diarrhea, and enhance the efficacy and healing conditions (Etana, 2010). Similar findings were reported by many other researchers, indicating the oral route as the most preferred mode of administration (Filate et al., 2009; Mesfine et al., 2009; Addisie et al., 2012).

Dosage is not always well measured in most of the traditional medicine practitioners. The result of this study also showed that all administrations are not standardized. Other similar findings were reported by many other researchers (Yinger and Yehwalahu, 2007; Tolosa et al., 2013). Although, most of the remedies were reported to have no serious adverse effects except vomiting and temporary inflammations. This could be attributed to the low toxicity of the remedy preparations of the medicinal plant species used by the traditional healers in the study area. However, the toxicity of some medicinal plants and their potential to do harm is a common complaint among those who would like traditional medicine to be standardized. It is commonly believed that traditional practitioners either do not know the strength of their own medicines or do not bother to fit doses to the size or body weight of the patients (Hillenbrand, 2006). However, it is known that some traditional healers do give different dosages and frequency of application depending on age, sex and other condition or vary the medicine itself on such differences.

The highest numbers of plant species were reported to be used for treatment of intestinal parasites, abdominal pain and diarrhea indicating that there is relatively high consensus on the treatment of gastro intestinal problems with the medicinal plants of the area. This is mainly attributed to the highest prevalence of gastro intestinal problems ailments prevalent in the study area. A similar analysis found high value of informant consensus factor (ICF) for gastrointestinal illness in similar studies carried

out in different parts of Ethiopia (Tolosa et al., 2013). According to the information obtained from Debere Libanos local health center, intestinal parasites were the first among the top ten diseases treated in the health center in the year 2008. The frequent occurrence of these diseases might have given the chance for herbalists to develop diversified knowledge associated with this problem. The ICF results could be useful in prioritizing medicinal plants for further scientific validation of plants and plant products (Giday et al., 2006; Subramanyam et al., 2008) as pharmacologically effective remedies are expected from plants with higher ICF values (Trotter and Logan, 1986; Etuk and Mohammed, 2009). Indeed, documentation of inherently rich traditional ethno-medicinal knowledge based on ICF values have provided valuable information on new pharmacological dimensions for better health care of livestock and humans regarding many ailments (Etuk and Mohammed, 2009) and also assist conservation and management of rare, gradually vanishing important ethno-medicinal plant species

Traditional healers of the study area use different plants for the same ailment. But, when all plants are available at the same time they prefer one over the other. This is mostly done based on the effectiveness of the plants to cure the ailment. As a result, both preference ranking and paired comparison revealed *Malva verticillata* and *Brucea antidysenterica* as the most effective medicinal plants for the treatment of ascariasis and wound, respectively, at least in the context of local people. This indicates that indigenous people of the study area have sufficient knowledge on the healing power of medicinal plants.

It has been agreed that threats to biodiversity are increasing dramatically from time to time in such a way that the rate of biodiversity loss outweigh the rate of recruitments. Informants have pointed out that deforestation to expand agricultural land and for firewood collection are the major threats that threatened the survival of medicinal plants. Similar studies in different localities of Ethiopia have documented that medicinal plants are under severe anthropogenic threats. For example, wild habitats are subjected to the loss of a number of plant species due to different anthropogenic factors such as firewood collection (24.8%); frequent fire (22.3%) and harvesting medicinal plants for use in construction (19%) (Bekalo et al., 2009). A study conducted in Sekoru District (Yinger and Yehwalahu, 2007) has also showed that, there are different threats to medicinal plants such as deforestation (40%), drought (17.5%), agricultural expansion (12.5%) and fire (12.5%).

This study revealed that, most of the knowledge on herbal remedies is handed down to the younger members of the community by elders, who are 41-50 years old. This hints at the fact that ethno-medicinal knowledge is concentrated in the elderly members of the community and the relative difficulty in its transfer from the elders to the young generation. This might be related

to the waning of interest of the young generation on indigenous knowledge. Furthermore, since all most all part of the forest belongs to Debre Libanos Monastery of Ethiopian Oriental Orthodox Church, harvesting of any part of medicinal plant is strictly forbidden. A canon called "GIZIT" in the church language prohibits harvesting of any plant from the church compound. Move over, person who uses medicinal plants from the church compound or nearby area is believed not to get the expected cure from the health problem because of "GIZIT". As a result, many medicinal plants that are rare in other parts of the study area are found in the church compound or nearby area. These might appear to prevent the use of available medicinal plants by the herbalist and impart the expansion of the traditional medicine somehow. However, the strict conservation of the plant species by monastery is utmost important from the whole biodiversity conservation point of view. As a means for sustainable utilization of medicinal plants, plants in the church compounds and nearby area must be protected and used as a source of seeds for requirement and herbalist can now plant the seeds and grow the medicinal plants and use them as they wish. It might seem that herbalists are restricted to use. Different studies in different areas showed that medicinal plant knowledge and transfer of knowledge to the young generation have been affected by modernization (having access to modern education and health service) and environmental change (Bekalo, 2009; Tolosa et al., 2013). Furthermore, Western style health care services as provided by governments and NGOs, in particular in rural areas, seem to have contributed to a decline in traditional knowledge, in part because the local population simply regards Western medicine as more effective and safer.

Conclusion

The study area is home for several medicinal plants. These plants have a great value for the health problems of poor local people. Herbalists and knowledgeable farmers are using these plants to cure human and livestock diseases. The young generations have not shown much interest in this life long accumulated knowledge. This tendency of disinterestedness in traditional medicinal practices is likely to be one of the major causes for losing this wealth of knowledge in the near future. Therefore, it is important that the government create awareness among community members on the significance of preserving traditional knowledge and conserving medicinal plants before they disappear, and thereby ensure the rights of people to apply their traditional practices which are known for their proven safety and effectiveness.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENTS

Our special gratitude goes to the Department of Biology, Addis Ababa University for providing fund and facilities. We also duly acknowledge all herbalist and traditional healers who provided us information. Our appreciation goes to all those that helped us in data collection process. We duly appreciate the inputs of the reviewers.

REFERENCES

- Abebe D, Ayehu A (1993). Medicinal plants and enigmatic health practice of north Ethiopia. Berhanina Selam Printing Enterprise, Addis Ababa, Ethiopia.
- Addisie Y, Yared D, Kumar PA, Tomas Z, Awol A (2012). Traditional medicinal plants used by people in Libo-Kemkem district, south Gondar, Ethiopia. *Asian J. Agric. Sci.* 4:171-176.
- Amenu E (2007). Use And Management Of Medicinal Plants By Indigenous People of Ejaji area (Chelya Wereda) West Shoa, Ethiopia: An Ethnobotanical Approach. MSc Thesis, Addis Ababa University, Addis Ababa, Ethiopia.
- Asfaw Z, Woldu Z (1997). Crop Association of Home gardens in Wolayta and Gurage in Southern Ethiopia. *SINET: Ethiopian J. Sci.* 20:73-90.
- Ayyanar M, Ignacimuthu S (2005). Traditional knowledge of Kani tribals and Kouthalai of Tirunelveli hills, Tamil Nadu, India. *J. Ethnopharmacol.* 102:246-255.
- Bekalo TH, Woodmatas SD, Woldemariam ZA (2009). An ethnobotanical study of medicinal plants used by local people in the lowlands of Konta Special Woreda, southern nations, nationalities and peoples regional state, Ethiopia. *J. Ethnobiol. Ethnomed.* 5:26.
- Belayneh A, Asfaw Z, Demissew S, Bussa NF (2012). Medicinal plants potential and use by pastoral and agro-pastoral communities in Erer Valley of Babile Wereda, Eastern Ethiopia. *J. Ethnobiol. Ethnomed.* 8:42.
- Beyene T (2008). Ethnobotanical Study of Medicinal Plants In Adigrat Wereda and Adjacent Kebeles In Ganta-Afeshum Wereda, Eastern Tigray, Ethiopia: An Ethnobotanical Approach. MSc Thesis, Addis Ababa, Ethiopia.
- Birhan W, Giday M, Teklehaymanot T (2011). The contribution of traditional healers' clinics to public health care system in Addis Ababa, Ethiopia: A cross-sectional study. *J. Ethnobiol. Ethnomed.* 7:39.
- Bussmann RW, Swartzinsky P, Worede A, Evangelista P (2011). Plant use in Odo-Bulu and Demaro, Bale region, Ethiopia. *J. Ethnobiol. Ethnomed.* 7:28.
- Cotton CM (1996). *Ethnobotany: Principles and Applications*. John Wiley and Sons, New York, USA.
- Cunningham AB (2001). *Applied ethnobotany: People, wild plant use and conservation* London and Sterling, VA: Earthscan Publications Ltd.
- Etana B (2010). Ethnobotanical study of traditional medicinal plants of Goma Woreda, Jimma zone of Oromia region. Ethiopia: MSc Thesis, Addis Ababa University, Addis Ababa.
- Etuk EU, Mohammed BJ (2009). Informant consensus selection method: reliability assessment on medicinal plants used in north western Nigeria for the treatment of diabetes mellitus. *Afr. J. Pharm. Pharmacol.* 3:496-500.
- Flatie T, Gedif T, Asres K, Gebre-Mariam T (2009). Ethnomedical survey of Berta ethnic group Assosa Zone, Benishangul-Gumuz regional state, mid-west Ethiopia. *J. Ethnobiol. Ethnomed.* 5:14.
- Gebre T (2005). Ethnobotanical Study of Medicinal Plants in the Konso special Wereda (SNNPR), Ethiopia. MSc Thesis, Addis Ababa University, Addis Ababa, Ethiopia.
- Giday M, Ameni G (2003). An Ethnobotanical Survey on Plants of Veterinary Importance in two Woredas of Southern Tigray, Northern Ethiopia. *SINET: Ethiopian J. Sci.* 26:123-136.
- Giday M, Asfaw Z, Elmqvist T, Woldu Z (2003). An ethnobotanical study

- of medicinal plant used by The Zay People In Ethiopia. *J. Ethnopharmacol.* 85:43-52.
- Giday M, Teklehaymanot T (2013). Ethnobotanical study of plants used in management of livestock health problems by Afar people of Ada'ar District, Afar Regional State, Ethiopia. *J. Ethnobiol. Ethnomed.* 9:8.
- Hamilton A (2003). *Medicinal Plants and Conservation: issues and approaches.* Panda House, Catteshall Lane, London.
- Hillenbrand E (2006). Improving Traditional-Conventional Medicine Collaboration: Perspectives from Cameroonian Traditional Practitioners. *Nordic J. Afr. Stud.* 15:1-15.
- Kemal A, Mamo S, Megersa A, Mogos N (1996). Ecological study of Élan Mountain and Debre Libanos Monastery Natural Forest Areas. Environmental Protection And Land Use Planning Team. Oromia Agricultural Development Bureau, North Shewa Zone Agricultural Department, Fiche, Ethiopia.
- Legesse M, Ameni G, Mamo G, Medhin G, Bjune G, Abebe F (2011). Knowledge of cervical tuberculosis lymphadenitis and its treatment in pastoral communities of the Afar region, Ethiopia. *BMC Public Health* 11:157.
- Martin GJ (1995). *Ethnobotany: A method Manual.* Chapman and Hall, London, UK.
- Mesfin F, Demissew S, Teklehymanot T (2009). An ethnobotanical study of medicinal plants in Wonago Woreda, SNNPR, Ethiopia. *J Ethnobiol. Ethnomed.* 5:28.
- Seid MA, Tsegay BA (2011). Ethnobotanical survey of traditional medicinal plants in Tehuledere district, South Wollo, Ethiopia. *J. Med. Plant Res.* 5:6233-6242.
- Seifu T (2004). *Ethnobotanical And Ethnopharmaceutical Studies on Medicinal Plants Of Chifra District, Afar Region, North Eastern Ethiopia: An Ethnobotanical Approach.* MSc Thesis, Addis Ababa, Ethiopia.
- Subramanyam R, Steven NG, Murugesan M, Balasubramaniam V, Muneer M (2008). Consensus of the 'Malasars' traditional aboriginal knowledge of medicinal plants in the Velliangiri holy hills, India. *J. Ethnobiol. Ethnomed.* 4:8.
- Suleman S, Alemu T (2012). A survey on utilization of ethnomedicinal plants in Nekemte town, East Wellega (Oromia), Ethiopia. *J. Herbs Spices Med. Plants* 18:34-57.
- Teklehaymanot T, Gidey M (2007). Ethnobotanical Study of Medicinal Plants used by People in Zegie Peninsula, Northwestern Ethiopia. *J. Ethnobiol. Ethnomed.* 3:12.
- Teklehaymanot T, Gidey M, Medhin G, Mekonnen Y (2006). Knowledge and Use of Medicinal Plants by People around Debre Libanos Monastery in Ethiopia. *J. Ethnopharmacol.* 111:271-283.
- Tolossa K, Debela E, Athanasiadou S, Tolera A, Ganga G, Houdijk JG (2013). Ethno-medicinal study of plants used for treatment of human and livestock ailments by traditional healers in South Omo, Southern Ethiopia. *J. Ethnobiol. Ethnomed.* 9:32.
- Trotter RT, Logan MH (1986). Informants consensus: a new approach for identifying potentially effective medicinal plants. In *plants in Indigenous Medicine and Diet.* Edited by Etkin NL. Bedford Hill, NY, USA: Redgrave Publishing Company pp.91-112.
- Yineger H, Yewhalaw D (2007). Traditional medicinal plant knowledge and use by local healers in Sekoru District, Jimma Zone, South western Ethiopia. *J. Ethnobiol. Ethnomed.* 3:24.
- Yirga G (2010). Use of traditional medicinal plants by indigenous people in Mekele town, capital city of Tigray regional state of Ethiopia. *J. Med. Plants Res.* 4:1799-1804.

Full Length Research Paper

Community structure, regeneration potential and future dynamics of natural forest site in part of Nanda Devi Biosphere Reserve, Uttarakhand, India

Balwant Rawat^{1,2*}, Sanjay Gairola¹, K. Chandra Sekar¹ and R. S. Rawal¹

¹G.B. Pant Institute of Himalayan Environment and Development, Kosi-Katarmal, Almora 263 643, Uttarakhand, India.

²Society for the Conservation of Nature, Parnkuti Anantpur University Road, Rewa, Madhya Pradesh, India.

Received 20 April, 2014; Accepted 26 June, 2014

Realizing the overarching values of forests and considering their depletion at unprecedented rate, conservation of forests has emerged as the prime objective across the globe. Forest vegetation of Pindari-Sunderdhunga-Kafni (PSK), a protected area, part of Nanda Devi Biosphere Reserve in west Himalaya was analyzed for structure, composition and development of future compositional patterns. Forest vegetation surveys were carried out enumerating ten 10 x 10 m quadrat for tree species in each of 30 forest stand complemented by shrub (five 2 x 2 m quadrat) and herb (ten 1 x 1 m quadrat) surveys within each stand. Floristic richness reveals 332 plant species from 11 representative forest communities. Broadly, the demographic profiles exhibited progressive structures suggesting long term persistence of the communities/species. Differences in regeneration behavior of various species are indicative of future structure and dynamics of the communities. Data sets in the present study established target site in NDBR as potential sites for long-term ecological monitoring under various change scenarios.

Key words: Natural forests, regeneration pattern, population structure, compositional changes.

INTRODUCTION

Forest composition, community structure and diversity patterns are important ecological attributes significantly correlated with prevailing environmental as well as anthropogenic variables (Gairola et al., 2008; Ahmad et al., 2010). Forests are always characterized by their three main life stages called seedling (newly emerged plants), sapling (established plants stands between seedling and tree) and tree (tree undisturbed of micro environmental

conditions). The number (density) and type (richness) of trees define the structure and composition of forest (Shankar, 2001; Mishra et al., 2003). Species richness patterns in relation to the environment need to be understood before drawing conclusions on the effect of biodiversity on ecosystem processes. Numerous problems regarding the study of species richness need to be clarified, including the role of disturbance (Huston,

*Corresponding author. E-mail: balwantkam@gmail.com. Tel: 91-9675399782. Fax: 91-05962-241150.

1994), and the relative importance of biotic versus abiotic factors (Cornell and Lawton, 1992; Austin and Gaywood, 1994). The number of tree individuals at seedling, sapling and tree level reveals population structure and their establishment at seedling and sapling level represent regeneration status (Baduni and Sharma, 2001; Bhandari, 2003). The nature of forest communities depends on the ecological characteristics in sites, species diversity and regeneration status of species (Criddle et al., 2003; Todaria et al., 2010). The tree species strata, that is, seedling, sapling and tree layers of the plant communities that maintain the population structure of forest get affected by micro-environmental factors which vary with seasonal changes (Khumbongmayum et al., 2006; Kharkwal et al., 2009). Environmental variation within a small geographical area makes altitudinal gradients ideal for investigating several ecological and biogeographical hypotheses (Korner, 1998). Hence, it becomes necessary to understand the species richness, population structure, germination and establishment of seedlings and saplings across seasons and altitude for maintenance of forest (Khumbongmayum et al., 2006; Rao, 1988). Complete absence of seedlings and saplings of tree species in a forest indicates poor regeneration, while presence of sufficient number of young individuals in a given species population indicates successful regeneration (Saxena and Singh, 1984). Presence of sufficient number of seedlings, saplings and young trees is greatly influenced by interaction of biotic factors of the environment (Boring et al., 1981; Aksamit and Irving, 1984).

Realizing the overarching values of forests and considering their depletion at unprecedented rate, conservation of forests has emerged as the prime objective. As such, it is globally accepted that the depletion of forests has many ecological, social and economic consequences; one among these is loss of biodiversity (Jha et al., 2000). Forests form the renewable natural resource on earth and occupy very unique position among the various natural resources by supporting life on earth in several ways and providing services that cannot be substituted by any other means.

Since its inception (1988), the diverse ecosystems and their components in Nanda Devi Biosphere Reserve (NDBR) have remained attraction of researches. The representative ecosystems and their components in the reserve have shown evidences of change with time and space. In particular, the variations and changes in plant communities have been reported to be highly dependent on geographical, environmental and anthropogenic factors. Besides, differences in soil parameters, fire intensity, over harvesting and other kinds of disturbances contribute to the variation in vegetation from one stand to another or even within a community. Therefore, the reserve management, most often, looks for authentic and precise information on structure and composition of vegetation so as to address diverse issues of conserva-

tion and management at different levels ranging from species and community to landscape level. The forests in NDBR not only form diverse representative ecosystems but also are the home for many rare and endangered species. While the core zone of reserve consists of 10% forests, the buffer zone has nearly 27% area under forests. These forests help in maintaining rich floral (angiosperms- 699, gymnosperms- 11, pteridophytes- 137, bryophytes- 146, lichens- 77 and fungi- 128 spp.) and faunal (mammals- 29, birds- 243, insects- 229, molluscs- 14, amphibian- 8, annelids- 6, reptiles- 3 and pisces- 1) diversity in the reserve (Rawal and Rawat, 2012). In recognition of its uniqueness, NDBR has been included in World Network of Biosphere Reserves (WNBR) by UNESCO since 2004. Also, the Nanda Devi and the Valley of Flowers National Parks, forming core zone of the reserve, have been inscribed on the World Heritage List by UNESCO under Natural Criteria vii and x.

Though, studies on different aspects of biodiversity have been carried out in NDBR of Himalaya viz. natural resource utilization (Joshi, 2002; Joshi and Samant, 2004; Silori, 2001), ecosystem functions (Adhikari, 1992; Garkoti, 1992, 2008; Singh et al., 1994; Garkoti and Singh, 1995; Gairola, 2005), management and development (Rawal and Rawat, 2012), threat assessment (Kala et al., 1998; Kala, 2005; Joshi, 2002; Joshi and Samant, 2004), ethonobiological enumerations (Joshi et al., 2000; Kala, 2005; Rawat et al., 2013) and floristic analysis (Joshi, 2002; Gairola, 2005; Sekar and Rawat, 2011; Rawat, 2013) but a systematic approach on the population structure and seasonal regeneration pattern of forest communities in NDBR with respect to their long term existence, is still lacking. Under the provision of protected areas, the need for understanding the structure and regeneration pattern in forests have been already emphasized to mitigate the ongoing challenges like overexploitation, deforestation etc., that emerged along with the present changing climate and socio-economic scenario.

Therefore, an understanding of the processes that affect regeneration of forest species is of crucial importance to both ecologists and forest managers in protected areas. Keeping the above in mind, seasonal phytosociological investigations have been carried out in a part of Nanda Devi Biosphere Reserve of Uttarakhand.

MATERIALS AND METHODS

Study area and site selection

The Nanda Devi Biosphere Reserve (NDBR), which forms the extensive study area, was designated as Biosphere Reserve by Government of India on 18th January, 1988. The reserve has a unique combination of diverse ecosystems including traditional agro ecosystems, various types of temperate forests, alpine meadows, glaciers, etc. It represents the west Himalayan highland (2b) province of the biogeographic zone-Himalaya and lies between

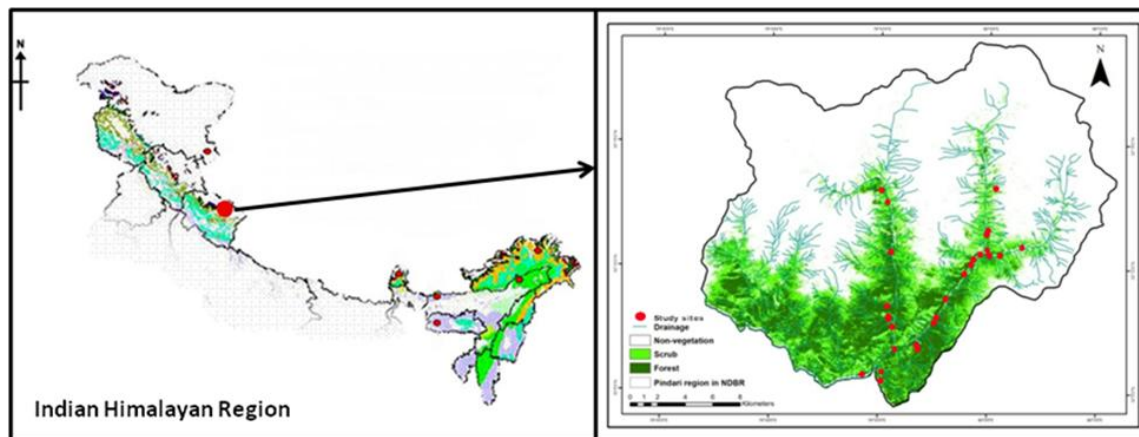


Figure 1. Location of study site of Pindari-Sunderdhunga-Kafni (right) in Nanda Devi Biosphere Reserve, western Himalaya (left).

30°06' and 31°04' north latitude and 79°13' and 80°17' east longitudes (Figure 1), and covers a total of 6,407.03 km² (Core zone 712.12 km²; Buffer zone 5,148.57 km², Transition zone 546.34 km²). One representative sites (Pindari-Sunderdhunga-Kafni: PSK in Kumaun region) in the buffer zone of NDBR formed the intensive study sites. Extensive surveys were conducted during 2008-2012 in these sites.

Sampling and data collection

Attempt was made to reach maximum possibly approachable stands. Standard phytosociological methods were followed to obtain quadrat data (Grieg-Smith, 1957; Misra, 1968; Kershaw, 1973; Muller-Dombois and Ellenberg, 1974; Dhar et al., 1997). In general, from each quadrat, circumference at breast height (CBH at 1.37 m from the ground) of all tree individual was recorded. Based on this information, individuals were considered as tree >31 cm; sapling 11-30 cm; seedling <11 cm CBH. Further, the individuals of tree species were grouped into six arbitrary CBH classes (A: <10; B: 11-30; C: 31-60; D: 61-120; E: >120 cm). The total number of individuals, belonging to each of the above classes, was calculated for each species in individual stand and stand information was pooled to represent community. Size class A and B represented seedlings and saplings, respectively. Other classes (C-E) represented tree classes. Relatively density of species in a particular size class was calculated as a percentage of total number of individuals in all size classes.

The quadrat information was pooled for calculating density, frequency, total basal area and their relative values (Misra, 1968; Muller-Dombois and Ellenberg, 1974). Following Whittaker (1975) and Pielou (1975), species richness was considered simply as the number of species per unit area. Species diversity index was computed using Shannon-Wiener information function (Shannon and Weiner, 1963). Statistical analysis (*t*-test and correlation coefficient (*r*) and coefficient determination (*r*²) and similarity indices were calculated using SPSS version 16 to determine the relationship between different phytosociological parameters.

Community structure and regeneration patterns

Seasonal investigation (Negi, 1995) on population structure and regeneration behavior of all tree species in PSK site was carried out during summer season: May-June, rainy season: mid July to August

and winter season: November-December in the years 2009 and 2010. Eleven representative forest communities were identified for studying detailed population structure to predict the future compositional changes in parent communities.

Regeneration status of species was determined based on population size of seedlings and saplings (Khan et al., 1987; Shankar, 2001; Bhuyan et al., 2003): good regeneration, if seedlings > saplings > trees; fair regeneration, if seedlings > or = saplings ≤ trees; fair regeneration, if the species survives only in sapling stage, but no seedlings (saplings may be <, > or = trees). If a species is present only in tree form, it is considered as not regenerating, while species having no trees but only seedlings is considered as 'new' species.

RESULTS

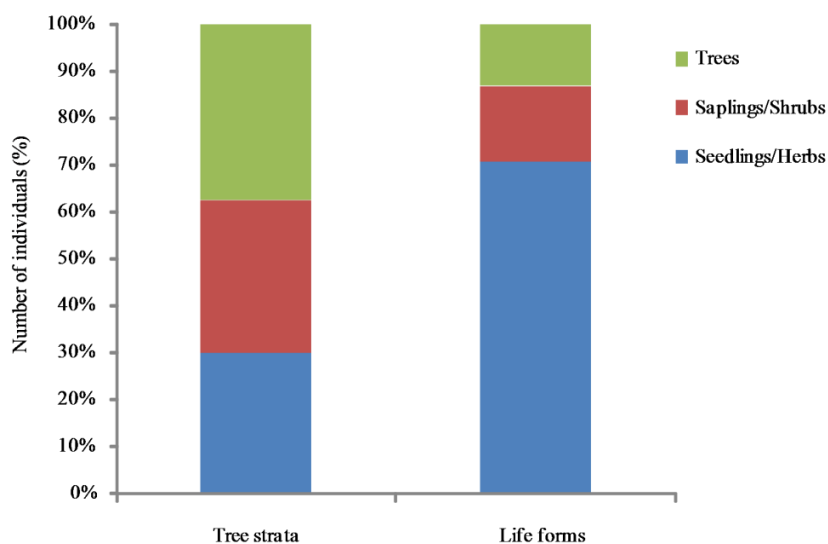
Site characteristics, floristic diversity and demographic patters

Eleven representative forest communities were distributed between 2025 to 3343 m asl. The community types, site representation and important species (IVI) are presented in Table 1. In general, 332 plant species were recorded from target site in NDBR. Of these, greater proportion (70.8%; 235 spp.) was of herbs. Shrubs constituted 16.3% (54 spp.), and trees 13.0% (43 spp.). Considering various taxonomic groups of the total 332 species in PSK site, 88.9% were angiosperms, 1.2% gymnosperms and 9.9% pteridophytes.

Proportional distribution of individuals in three broad tree strata (tree, sapling and seedling) and life forms (tree, shrubs, and herbs) in the study area and representative sites are depicted in Figure 2. In general, PSK site represented 43 tree species. Representation as compared to total tree species across different strata indicated maximum proportion of trees (93.2%) followed by saplings (81.4%) and seedlings (74.4%). The overall population structure for target sites and the entire reserve have been presented (Figure 3). As reflected, PSK site

Table 1. Site characteristics and dominant species across forest communities in PSK site.

| Community types (PSK site) | Altitude (masl) | Slope (°) | No. of stands | Important species (IVI value) |
|--|-----------------|-----------|---------------|--|
| <i>Alnus nepalensis</i> (Utis) | 2025 | 40-45 | 2 | <i>Alnus nepalensis</i> (128.2) <i>Ulmus wallichiana</i> (32.9) |
| Mixed Oak- Deciduous | 2217 | 40-55 | 3 | <i>Quercus floribunda</i> (79.8) <i>Aesculus indica</i> (34.6) |
| <i>Hippophae salicifolia</i> (Chuck) | 2452 | 5-15 | 3 | <i>Hippophae salicifolia</i> (232.0) <i>Alnus nepalensis</i> (32.6) |
| <i>Quercus floribunda</i> (Tilonj Oak) | 2504 | 35-50 | 4 | <i>Quercus floribunda</i> (127.1) <i>Rhododendron arboreum</i> (42.7) |
| <i>Quercus semecarpifolia</i> (Kharsu Oak) | 2669 | 35-50 | 4 | <i>Quercus semecarpifolia</i> (112.7) <i>Rhododendron arboreum</i> (37.9) <i>Quercus floribunda</i> (35.3) |
| Mixed-deciduous | 2773 | 40-55 | 3 | <i>Acer cappadocicum</i> (49.4) <i>Ulmus wallichiana</i> (28.1) <i>Rhododendron arboreum</i> (26.9) |
| Mixed Silver fir-Oak | 2855 | 30-45 | 2 | <i>Abies pindrow</i> (68.5) <i>Quercus semecarpifolia</i> (33.8) <i>Aesculus indica</i> (30.4) |
| Mixed Silver fir-Rhododendron-Maple | 2860 | 40-50 | 3 | <i>Rhododendron barbatum</i> (74.0) <i>Abies pindrow</i> (38.9) <i>Ilex dipyrena</i> (27.5) |
| <i>Abies pindrow</i> (Silver fir) | 2970 | 60-65 | 2 | <i>Abies pindrow</i> (99.9) <i>Rhododendron barbatum</i> (47.6) <i>Betula utilis</i> (44.0) |
| Mixed Birch-Silver fir | 3238 | 50-60 | 2 | <i>Betula utilis</i> (126.4) <i>Abies pindrow</i> (67.9) <i>Taxus wallichiana</i> (32.6) |
| <i>Betula utilis</i> (Birch) | 3343 | 50-65 | 2 | <i>Betula utilis</i> (183.4) <i>Euonymous fimbriatus</i> (29.1) <i>Rhododendron campanulatum</i> (20.8) |

**Figure 2.** Proportional distribution of species richness across different tree strata and life forms in PSK site in NDBR.

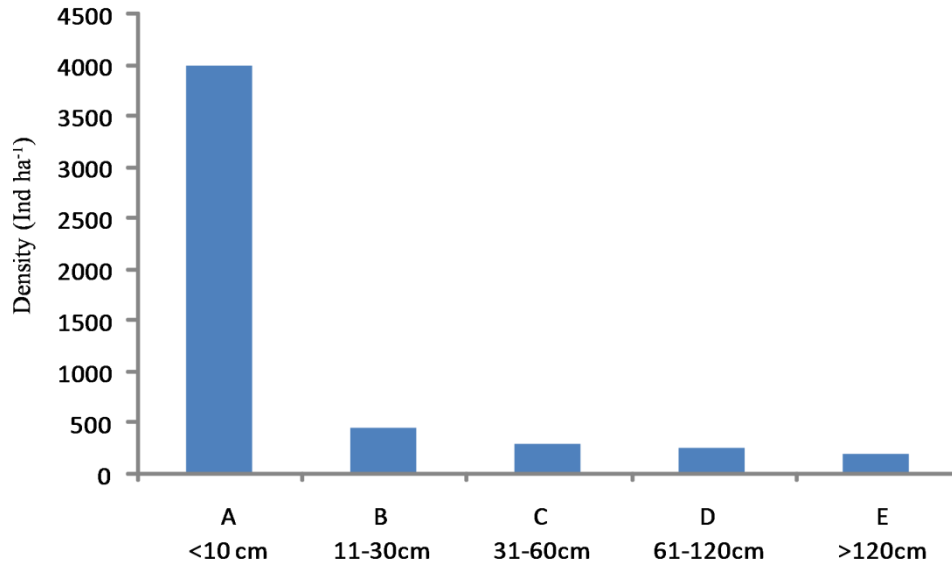


Figure 3. Mean density-diameter distribution of trees in PSK site.

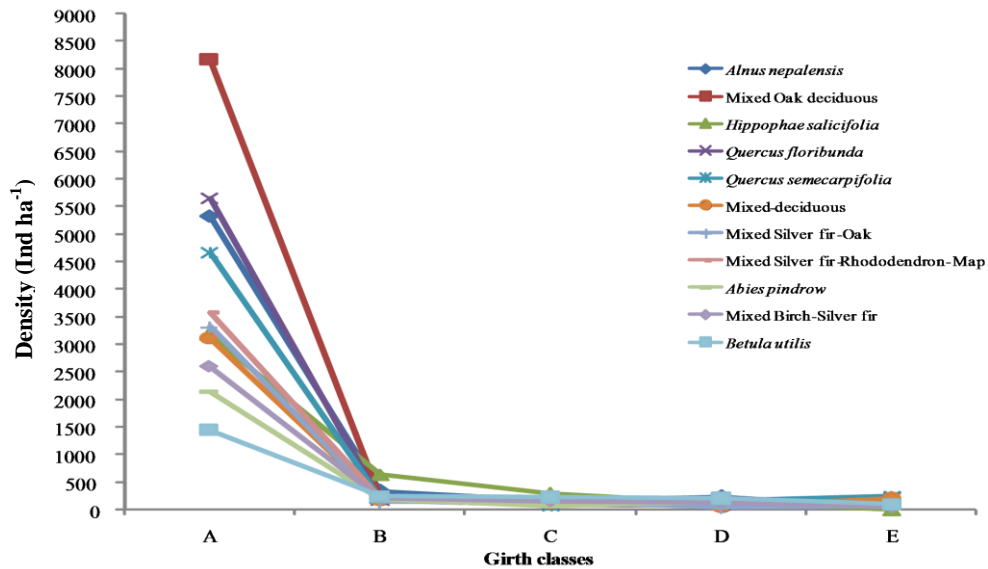


Figure 4. Density-diameter distribution of trees in different forest communities in PSK site.

has considerably larger number of individuals in seedling stage and more gradual decline of individuals towards higher tree size classes. The profile of demography for different forests in PSK site exhibited more or less similar patterns (Figure 4).

Community diversity and distribution pattern

Detailed quantitative ecological parameters in different forest communities are presented in Table 2. Considering the forest composition, tree species richness was highest

in *Quercus semecarpifolia* and Mixed Silver fir-Oak communities (23 spp. each) and minimum in *Hippophae salisifolia* community (4 spp.). Sapling species richness peaked in *Q. floribunda* (18 spp.) followed by Mixed deciduous and Mixed Silver fir-Oak communities (15 spp. each). Lowest richness was recorded in *H. salicifolia* community (3 spp.). Species richness at seedling stage was maximum in *Quercus floribunda* (19 spp.) followed by *Quercus semecarpifolia* and MIXED community (16 spp.) and lowest in *H. salicifolia* and *Betula utilis* communities (5 spp. each). The details of species richness in different tree layers and across

Table 2. Quantitative ecological parameters in different forest communities in PSK site.

| Community types (PSK site) | Species richness | | | Species density (Ind ha ⁻¹) | | | Species diversity | | |
|---|------------------|---------|------|---|-----------|-----------|-------------------|------------|------------|
| | Seedling | Sapling | Tree | Seedling | Sapling | Tree | Seedling | Sapling | Tree |
| <i>Alnus nepalensis</i> | 9 | 9 | 14 | 5325± 878 | 345 ± 53 | 460± 49 | 2.20 ± 0.3 | 2.38 ± 0.3 | 2.38 ± 0.2 |
| Mixed Oak deciduous | 13 | 12 | 21 | 8170± 3012 | 233 ± 33 | 480 ± 50 | 2.28 ± 0.2 | 2.66 ± 0.1 | 3.37 ± 0.1 |
| <i>Hippophae salicifolia</i> | 5 | 3 | 4 | 3167 ± 609 | 633 ± 179 | 423 ± 174 | 0.70 ± 0.2 | 0.64 ± 0.3 | 0.73 ± 0.1 |
| <i>Quercus floribunda</i> | 19 | 18 | 20 | 5650 ± 1613 | 170 ± 27 | 378 ± 40 | 2.46 ± 0.4 | 2.48 ± 0.1 | 2.84 ± 0.2 |
| <i>Quercus semecarpifolia</i> | 16 | 14 | 23 | 4663 ± 819 | 200 ± 37 | 473 ± 42 | 2.27 ± 0.3 | 2.10 ± 0.3 | 2.73 ± 0.4 |
| Mixed-deciduous | 16 | 15 | 21 | 3117 ± 348 | 197 ± 41 | 407 ± 73 | 2.34 ± 0.3 | 2.47 ± 0.2 | 3.31 ± 0.1 |
| Mixed Silver fir-Oak | 9 | 13 | 21 | 3325 ± 469 | 145 ± 12 | 260 ± 16 | 2.27 ± 0.1 | 2.39 ± 0.1 | 3.20 ± 0.1 |
| Mixed Silver fir- Rhododendron-Maple | 14 | 15 | 23 | 3583 ± 866 | 200 ± 26 | 367 ± 39 | 2.51 ± 0.1 | 2.67 ± 0.3 | 3.22 ± 0.3 |
| <i>Abies pindrow</i> | 9 | 10 | 17 | 2150 ± 653 | 190 ± 8 | 320 ± 49 | 2.08 ± 0.4 | 2.55 ± 0.1 | 2.71 ± 0.2 |
| Mixed Birch-Silver fir | 9 | 11 | 10 | 2600 ± 694 | 210 ± 0 | 355 ± 102 | 2.49 ± 0.3 | 2.64 ± 0.1 | 2.02 ± 0.1 |
| <i>Betula utilis</i> | 5 | 6 | 8 | 1450 ± 531 | 235 ± 20 | 535 ± 116 | 1.50 ± 0.1 | 1.81 ± 0.3 | 1.55 ± 0.2 |

communities are presented.

The tree density ranged from 260 (Mixed Silver fir-Oak community) to 535 ind ha⁻¹ in *B. utilis* community. In the case of saplings, maximum density was recorded in *H. salicifolia* community (633 ind ha⁻¹) and minimum in Mixed Silver fir-Oak community (145 ind ha⁻¹). Seedling density, however, peaked in Mixed Oak deciduous community (8170 ind ha⁻¹) followed by *Q. floribunda* (5650 ind ha⁻¹) and *Alnus nepalensis* community (5325 ind ha⁻¹). The minimum seedling density was recorded in *B. utilis* community (1450 ind ha⁻¹).

While considering the diversity index, highest value for tree layer was in the case of Mixed Oak deciduous community (3.37) followed by Mixed deciduous (3.31) and Mixed Silver fir-Rhododendron-Maple community (3.22). Whereas, Mixed Silver fir-Rhododendron-Maple community (2.67) followed by Mixed Oak deciduous (2.66) and Mixed Birch-Silver fir community (2.64) showed highest values in sapling layer. Mixed Silver fir-Rhododendron-

Maple community (2.51) also peaked for seedling diversity followed by Mixed Birch-Silver fir (2.49) and *Q. floribunda* community (2.46). *H. salicifolia* community invariably had lowest diversity values across three tree strata (tree - 0.73, sapling - 0.64, seedling - 0.70).

Regeneration status and seasonal behavior

In the target site, out of the 43 tree species, 16.3% showed good, 46.5% fair, 25.6% no, 7.0% poor regeneration and remaining 4.7% were represented only by seedlings and saplings (Table 3). ANOVA based analysis revealed uneven variation in density values across the seasons (Figure 5). Starting from summer (2009), a significant ($p>0.05$) increase in the number of seedlings was observed with onset of rainy season (2009).

Afterward, the seedling density gradually decreased in winter (2009) and summer (2010) and increased significantly in the next rainy

season (2010). In the year 2009, the average seedling density was measured about 2,867 ind ha⁻¹ in summer that reached 3,491 ind ha⁻¹ in the rainy season. In winter 2009, gradual decrease in seedling density (3,182 ind ha⁻¹) was recorded. Similar trends were observed in 2010. A linear regression line showed gradual but non-significant ($p>0.05$) increase in seedling density across the years.

The growth and establishment of seedling is irrespective of the altitude (Figure 6). *A. nepalensis* and mixed oak deciduous in lower altitudinal zone, *Abies-Rhododendron-Maple* in mid altitudinal zone and *B. utilis* in high altitude zone showed remarkable regeneration and seedling establishment.

DISCUSSION

Compositional diversity

Considering the floristic richness of representative

Table 3. Density and regeneration status in different forest communities in PSK site.

| Species | No. of individuals ha ⁻¹ | | | Status |
|--------------------------------|-------------------------------------|---------|-------|--------|
| | Seedling | Sapling | Tree | |
| <i>Abies pindrow</i> | 2937.5 | 109.2 | 297.5 | Fair |
| <i>A. spectabilis</i> | - | 5.0 | 10.0 | No |
| <i>Acer acuminatum</i> | 362.5 | 35.0 | 49.2 | Fair |
| <i>A. caesium</i> | 545.8 | 33.3 | 61.7 | Fair |
| <i>A. cappadocicum</i> | 1916.7 | 120.8 | 165.8 | Fair |
| <i>Aesculus indica</i> | - | 7.5 | 101.7 | No |
| <i>Alnus nepalensis</i> | 1970.8 | 220.0 | 274.2 | Fair |
| <i>Betula alnoides</i> | 1029.2 | 23.3 | 84.2 | Fair |
| <i>Betula utilis</i> | 1125.0 | 185.0 | 570.0 | Fair |
| <i>Buxus wallichiana</i> | 58.3 | 10.0 | - | New |
| <i>Carpinus viminea</i> | 154.2 | 14.2 | 66.7 | Fair |
| <i>Celtis australis</i> | - | - | 10.0 | No |
| <i>Cornus macrophylla</i> | - | 3.3 | 45.0 | No |
| <i>Corylus jacquemontii</i> | 50.0 | 10.0 | 33.3 | Poor |
| <i>Elaeagnus parvifolia</i> | - | - | 10.0 | No |
| <i>Euonymus fimbriatus</i> | 675.0 | 83.3 | 115.0 | Fair |
| <i>Eurya acuminata</i> | - | 5.0 | - | No |
| <i>Fraxinus micrantha</i> | - | - | 35.8 | No |
| <i>Hippophae salicifolia</i> | 3654.8 | 546.7 | 346.7 | Good |
| <i>Ilex dipyrrena</i> | 1941.7 | 118.3 | 102.5 | Good |
| <i>Juglans regia</i> | 16.7 | - | 42.5 | Poor |
| <i>Lyonia ovalifolia</i> | 1170.8 | 51.7 | 124.2 | Fair |
| <i>Mahonia borealis</i> | 33.3 | - | 5.0 | Poor |
| <i>Meliosma dilleniaefolia</i> | 287.5 | 34.2 | 31.7 | Good |
| <i>Neolitsea pallens</i> | 3929.2 | 126.7 | 123.3 | Good |
| <i>Prunus cornuta</i> | - | - | 35.0 | No |
| <i>Pyrus lanata</i> | 387.5 | 12.5 | 37.5 | Fair |
| <i>Pyrus pahsia</i> | 12.5 | 6.7 | - | New |
| <i>P. vestita</i> | 83.3 | 5.0 | 61.7 | Poor |
| <i>Quercus floribunda</i> | 6037.5 | 64.2 | 287.5 | Fair |
| <i>Q. incana</i> | - | - | 11.7 | No |
| <i>Q. semecarpifolia</i> | 3037.5 | 19.2 | 194.2 | Fair |
| <i>Rhododendron arboretum</i> | 5820.8 | 272.5 | 310.0 | Fair |
| <i>R. barbatum</i> | 4887.5 | 225.8 | 267.5 | Fair |
| <i>R. campanulatum</i> | 650.0 | 53.3 | 65.0 | Fair |
| <i>Rhus punjabensis</i> | - | - | 2.5 | No |
| <i>Salix daphnoides</i> | 628.6 | 70.0 | 96.7 | Fair |
| <i>Symplocos chinensis</i> | 258.3 | 49.2 | 70.8 | Fair |
| <i>S. ramosissima</i> | - | 3.3 | 33.3 | No |
| <i>Syringa emodi</i> | 183.3 | 32.5 | 8.3 | Good |
| <i>Taxus wallichiana</i> | 912.5 | 116.7 | 114.2 | Good |
| <i>Ulmus wallichiana</i> | 633.3 | 35.8 | 152.5 | Fair |
| <i>Viburnum nervosum</i> | 487.5 | 64.2 | 2.5 | Good |

site, the PSK site with over 332 plant species emerged as more species rich and representative as compared to other sites in NDBR (Rawat, 2013). This can be attributed to the existence of more diverse and broadleaf forest

communities (11 forest communities). In general, PSK site has 72% coverage of broadleaf dominated forests and nearly 28% conifer dominated ones, which is much higher than any other site in NDBR (Joshi, 2002; Joshi

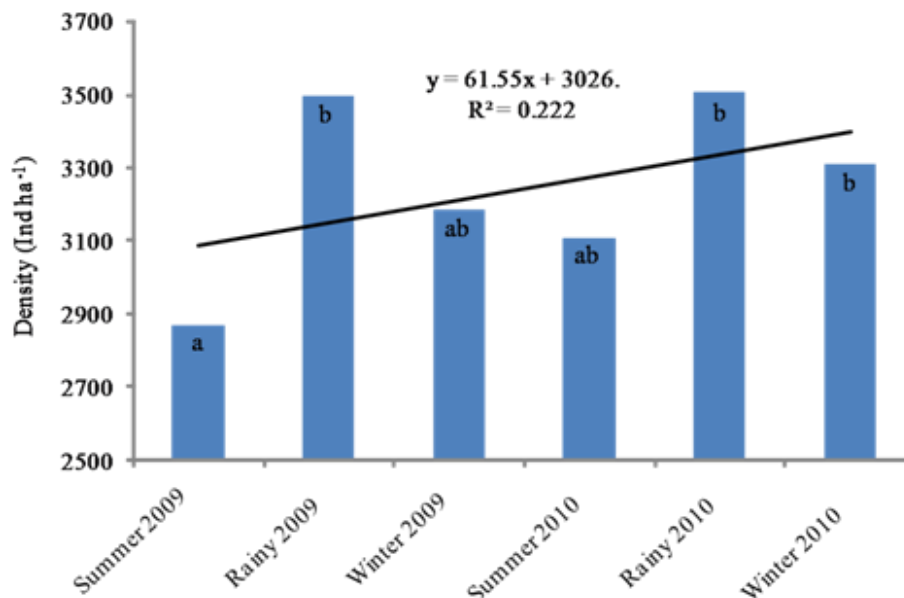


Figure 5. Average seasonal recruitment pattern in the entire forest communities. ANOVA was applied against the average density; same letters denote non-significant difference.

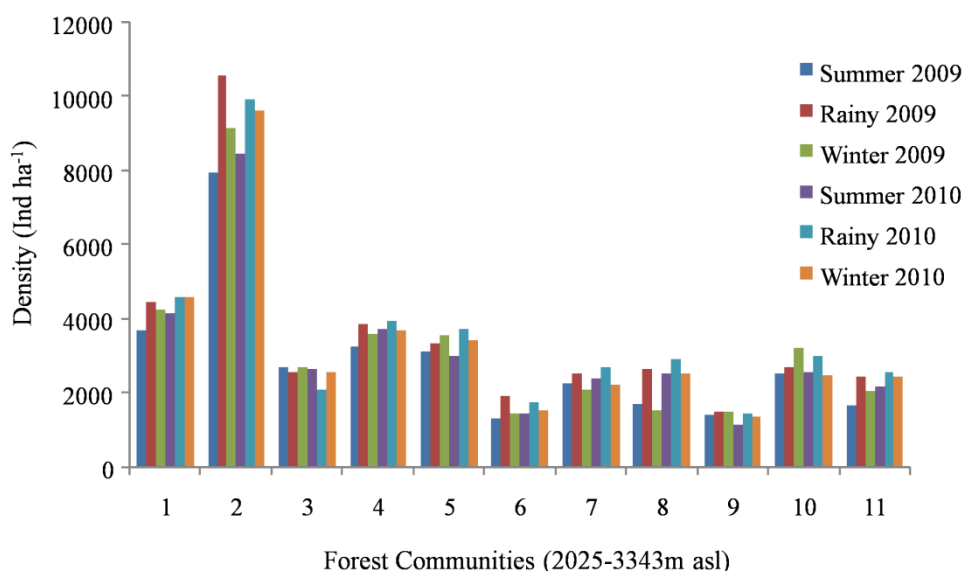


Figure 6. Seasonal recruitment patterns in different forest communities in PSK site in NDBR. [1. *Alnus nepalensis*; 2. Mixed Oak-deciduous; 3. *Hippophae salicifolia*; 4. *Quercus floribunda*; 5. *Quercus semecarpifolia*; 6. Mixed-deciduous; 7. Mixed Silver fir-Oak; 8. Mixed Silver fir-Rhododendron-Maple; 9. *Abies pindrow*; 10. Mixed Birch-Silver fir; 11. *Betula utilis*].

and Samant, 2004; Rawat, 2013). The explanations for this lies in the fact that PSK site supports more mesic (moist) conditions. In general, coniferous communities are broadly reported to be species poor as compared to broadleaf communities (Singh and Singh, 1992).

The mean tree density (260-535 ind ha⁻¹) was comparable to the values (320-1670 ind ha⁻¹) reported in

earlier studies pertaining to low and high altitude forests of west Himalaya (Bankoti, 1990; Joshi and Samant, 2004; Gairola et al., 2008; Garkoti, 2008). The density values closely corresponded with the values (270-610 ind ha⁻¹) recorded from the same region two decade ago (Bankoti, 1990), however, these values fall in lower range of values reported for the region (Zobel and Singh,

1997).

The mean values range for seedling density (1450-8170 ind ha⁻¹) was considerably higher as compared to earlier reports for high altitude forest in similar areas (Bankoti, 1990; Joshi, 2002; Joshi and Samant, 2004). The range of mean sapling density (145-633 ind ha⁻¹) falls within the lower range (40-6667 ind ha⁻¹) reported earlier (Bankoti, 1990; Joshi, 2002; Joshi and Samant, 2004; Gairola et al., 2008).

Current state of tree species richness (4-23 species) was comparable to earlier reports (24-42 species) from high altitude forests of the region. Similarly, the diversity values (0.73-3.37) are within the reported range (0.45-3.29) from the region (Bankoti, 1990; Adhikari et al., 1991; Joshi, 2002; Gairola, 2005; Gairola et al., 2008). The trends of species richness and diversity indicate forests in mid to higher altitudes are more diverse than the lower altitude forests in the west Himalaya.

Current demographic profiles

The size class distribution gives a demographic profile of the region which may indicate future prospects of target communities (Gairola, 2005). In general, the forest communities showed greater accumulation of individual in seedlings and a significant decline towards sapling and tree size classes. This structure reveals that the conversion from seedling to sapling is not proportional. This can be explained on account of greater mortality of seedlings due to severe winters. Similar conclusions have been drawn by other workers elsewhere (Khumbongmayum et al., 2006).

Further, large scale extraction of biomass, particularly of selected species, has also been reported to bring in structural changes in plant communities in the region and elsewhere (Thadani and Ashton, 1995; Singh et al., 1997; Spurr and Barnes, 1980; Cairns and Moen, 2004; Shrestha et al., 2007). As such, disturbances have been observed to exert profound effect on forest development, since they alter vegetation and release growing space, making it available for other species to occupy (Oliver and Larson, 1990; Mishra et al., 2003; Mishra et al., 2004; Gairola et al., 2008). The lower density of the higher girth classes of trees in the region, as compared to intermediate or lower girth classes, can be attributed to the relatively high mortality of large canopy trees (Goff et al., 1975; Lorimer et al., 2001).

Seasonal regeneration pattern

While considering the demographic profile, greater accumulation of seedlings can be attributed to occasional mast seedling and recruitment for some dominant species (*Q. floribunda*, *Q. semecarpifolia*). However, the long-term persistence of such recruits was later confirmed

by two years seasonal investigation on seedling survival patterns. Therefore, if this trend continues, the forest communities are likely to have increased dominance of such species.

Further, the seasonal recruitment patterns suggest that the regenerating species are now established and if this trend continues, the forest communities will sustain long in future. The overall population structure of tree species reveals that seedlings populations dominate tree populations and the fluctuation in population density in various seasons is related to the prevailing environmental factors. Germination of freshly dispersed seeds is high for most of the species during the rainy season, which is the wettest season. Lieberman and Li (1992) and Swaine et al. (1990) have observed similar patterns in tropical dry forest at Pinkwae, Ghana. Adverse effects of soil moisture stress and unfavorable temperature on survival of plant species may be responsible for reduction of seedling population during winter season (Perira and Kozlowski, 1977; Schulte and Marshall, 1983; Khumbongmayum et al., 2006). The gradual decrease in recruitments in summer season can be attributed to the anthropogenic pressure in form of lopping and grazing. Evolutionary history of grazing and environmental moisture or primary productivity interacts in determining species adaptations for tolerance or avoidance of herbivores and in community responses to grazing (Milchunas and Lauenroth, 1993). The average of fluctuations in recruitment density across altitude and seasons revealed established regeneration.

Expected changes in forests vis- a-vis representativeness

Broadly, the demographic profiles exhibited progressive structures suggesting long term persistence of the communities/species in these sites. However, diverse trends of density and richness of recruits helps us to depict the status of species in different forest communities (Khan et al., 1987; Shankar, 2001; Bhuyan et al., 2003). In this respect, following patterns across recruitment layers are noticeable for different communities:

Sapling layer

1. Communities having high representation of dominant species: *Alnus nepaensis*, *H. salicifolia*, Mixed deciduous, Mixed Silver fir-Rhododendron-Maple, *B. utilis* communities.
2. Communities having high representation of co-dominant species: Mixed Birch-Silver fir
3. Communities having poor representation of dominant species but highest of co-dominant species: *Q. floribunda*, *Q. semecarpifolia* communities.

4. Communities having poor representation of dominant and co-dominant species and high representation of other species: Mixed Oak deciduous, Mixed Silver fir-Oak communities.

Seedling layer

1. Communities having high seedling representation of dominant species: Mixed Oak deciduous, *H. salicifolia*, *Q. floribunda*, *Q. semecarpifolia*, Mixed Oak deciduous, Mixed Silver fir-Oak, Mixed Silver fir-Rhododendron-Maple, *Abies pindrow*, *B. utilis* communities.

2. Communities having sufficient representation of both dominant and co-dominant species, and accompanied by high representation of other species as well: Mixed Birch-Silver fir community.

Therefore, based on above trends of seedlings and saplings, various combinations and trends of communities can be drawn. For example, (i) the communities with greater representation of dominant species in both seedling and sapling stage would suggest further strengthening of dominant species; (ii) the communities with greater representation of both dominant and co-dominant in sapling and seedling layers would indicate the composition remains unchanged in future; (iii) the communities having greater proportion of seedling and saplings of co-dominant would indicate possible dominance of such species in future; (iv) the communities having greater representation of seedlings and saplings of the species other than the dominants and the co-dominants would indicate likely future changes in composition of target communities.

The demographic profiles of some of the dominant and some relatively less prominent tree species definitely require attention. For example, in the case of two dominant species, *Q. floribunda* and *Q. semecarpifolia*, in spite of their greater seedling numbers both were less prominently represented in sapling layer. Certain relatively less prominent species like *Celtis australis*, *Elaeagnus parvifolia*, *Fraxinus micrantha*, *Prunus cornuta*, *Quercus incana* and *Rhus punjabensis* were, however, represented only in tree layer suggesting that these species are not properly regenerating through near past and in the present. Therefore, long-term persistence of such species is in question. Besides, *Abies spectabilis*, *Aesculus indica*, *Symplocos ramosissima* with representation only in tree and sapling class and *Cornus macrophylla*, *Juglans regia*, *Mahonia borealis*, *Viburnum nervosum* in tree and seedling class only would require attention. On the contrary, species like *Buxus wallichiana*, *Pyrus pahasia*, *Eurya acuminata* having individuals in sapling and seedling class only indicated their recent introduction in respective communities.

Differences in regeneration behavior of various species are indicative of future structure and dynamics of the communities under natural circumstances. Present study

reveals good regeneration and exemplifies regeneration of tree species which is largely dependent on the prevailing environmental factors and anthropogenic threat, and if the existing ecological factors are not jeopardized, the future maintenance of the tree species in PSK site in NDBR will be sustained. However, presence of 'new' and 'not regenerating' species must be taken into consideration at the time of development of policies and plan for proper conservation and management of respective forest communities.

Conclusion

1. The data sets available, and generated through this study, provide enough bases for establishing PSK site in NDBR as potential sites for long-term ecological monitoring under various change scenarios.

2. Comparatively, the PSK site of the reserve supported greater diversity of plant communities and species as compared to other sites in the reserve.

3. Communities in target site broadly exhibited progressive demographic profiles which suggested long-term persistence of communities. However, unusually greater accumulation of seedling in PSK site with indications of successful establishment is indicative of possible changes in composition of communities in this site. Also, various community specific patterns of demography were revealed.

4. Assessment and analysis of changes in structure and composition of different forest types provides baseline data for developing priorities for conservation of other representative landscapes in the reserve as well as in the Himalaya.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors are grateful to the Director of the Institute for providing the necessary facilities to carry out this work. We are thankful to In-House Project-08 (2007-2012) for providing the financial facilities throughout the study period. Sincere thanks goes to DST (SERB/LS-933/2013) and CSIR (09/560 (0015)/2011-EMRI), Government of India for the financial support. We also thank the local inhabitants for their generous help during extensive field visits. Society for Conservation of Nature (SCON) is highly acknowledged for kind suggestions and support from time to time.

REFERENCES

Adhikari BS (1992). Biomass productivity and nutrient cycling of Kharsu

- oak and Silver fir forests in Central Himalaya. Ph.D. Thesis (Botany), Kumaun University, Nainital, India.
- Adhikari BS, Rikhari HC, Rawat YS, Singh SP (1991). High altitude forests: composition diversity and profile structures in a part of Kumaun Himalaya. *Trop. Ecol.* 32:86-97.
- Ahmad I, Ahmad MSA, Hussain M, Ashraf M, Ashraf MY, Hameed M (2010). Spatiotemporal aspects of plant community structure in open scrub rangelands of submountainous Himalayan plateaus. *Pak. J. Bot.* 42(5):3431-3440.
- Aksamit SE, Irving FD (1984). Prescribed burning for lowland black spruce regeneration in Northern Minnesota. *Can. J. For. Res.* 14:107-113.
- Austin MP, Gaywood M (1994). Current problems of environmental gradients and species response curves in relation to continuum theory. *J. Veg. Sci.* 5:473-482.
- Baduni NP, Sharma CM (2001). Population structure and community analysis on different aspects of Sal savanna forest type in outer Garhwal Himalaya. *Indian Forester* 127(9):1001-1011.
- Bankoti NS, Rawal RS, Samant SS, Pangtey YPS (1992). Forest vegetation of inner hill ranges in Kumaun, Central Himalaya. *Trop. Ecol.* 33:41-53.
- Bhandari BS (2003). Blue pine (*Pinus wallichiana*) forest stands of Garhwal Himalaya: Composition, population structure and diversity. *J. Trop. For. Sci.* 15(1): 26-36.
- Bhuyan P, Khan ML, Tripathi RS (2003). Tree diversity and population structure in undisturbed and human-impacted stands of tropical wet evergreen forest in Arunachal Pradesh, Eastern Himalayas, India. *Biodivers. Conserv.* 12(8):1753-1773.
- Boring LR, Monk CD, Swank WT (1981). Early regeneration of a clear cut southern Appalachian forest. *Ecol.* 62:1244-1253.
- Cairns DM, Moen J (2004). Herbivory influences tree lines. *J. Ecol.* 92:1019-1024.
- Cornell HV, Lawton JH (1992). Species interactions, local and regional processes, and limits to the richness of ecological communities: a theoretical perspective. *J. Anim. Ecol.* 61:1-12.
- Criddle RS, Church JN, Smith BN, Hansen LD (2003). Fundamental causes of the global patterns of species range and richness. *Russ. J. Plant Physiol.* 50:192-199.
- Dhar U, Rawal RS, Samant SS (1997). Structural diversity and representativeness of forest vegetation in a protected area of Kumaun Himalaya, India: implications for conservation. *Biodivers. Conserv.* 6:1045-1062.
- Gairola S (2005). Assessment of diversity pattern in sub-alpine forests of west Himalaya: recruitment strategy, litterfall and nutrient return. Ph.D. Thesis (Forestry), HNB University, Srinagar (Garhwal), India.
- Gairola S, Rawal RS, Todaria NP (2008). Forest vegetation patterns along an altitudinal gradient in sub-alpine zone of west Himalaya, India. *Afr. J. Plant Sci.* 2:42-48.
- Garkoti SC (1992). High altitude forests of central Himalaya: Productivity and nutrient cycling. Ph.D. Thesis (Botany), Kumaun University, Nainital, India.
- Garkoti SC (2008). Estimates of biomass and primary productivity in a high-altitude maple forest of the west central Himalaya. *Ecol. Res.* 23:41-49.
- Garkoti SC, Singh SP (1995). Variation in forest biomass and net primary productivity in the high mountains of central Himalaya. *J. Veg. Sci.* 6:23-28.
- Goff FG, West DC (1975). Canopy-under storey interaction effects on forest population structure. *For. Sci.* 21:98-108.
- Greig-Smith P (1957). *Quantitative Plant Ecology*. Academic Press, New York.
- Huston MA (1994). *Biological diversity: The coexistence of species on changing landscapes*. Cambridge University Press, Cambridge.
- Jha CS, Dutt CBS, Bawa KS (2000). Deforestation and land use changes in Western Ghats, India. *Curr. Sci.* 79:231-238.
- Joshi HC (2002). Assessment of habitat diversity, forest vegetation and human dependence in the buffer zone of Nanda Devi Biosphere Reserve of west Himalaya. Ph.D. thesis (Botany), Kumaun University, Nainital, India.
- Joshi HC, Arya SC, Samant SS (2000). Diversity, distribution and indigenous uses of plant species in Pindari area of Nanda Devi Biosphere Reserve- II. *Indian J. For.* 24:514-536.
- Joshi HC, Samant SS (2004). Assessment of forest vegetation and conservation priorities of communities in part of Nanda Devi Biosphere Reserve, West Himalaya. Part I. *Int. J. Sustain. Dev. World Ecol.* 11:326-336.
- Kala CP (2005). Indigenous uses, population density and conservation of threatened medicinal plants in Protected Areas of the Indian Himalayas. *Conserv. Biol.* 19(2):368-378.
- Kala CP, Rawat GS, Uniyal VK (1998). Ecology and conservation of the Valley of Flowers National Park, Garhwal Himalaya. Report, Wildlife Institute of India, Dehradun, India.
- Kershaw KA (1973). *Quantitative and dynamic plant ecology*. Second edition. Edward Arnold Limited, London.
- Khan ML, Rai JPN, Tripathi RS (1987). Population structure of some tree species in disturbed and protected sub-tropical forests of north-east India. *Acta Oecologica-Oecologia Applicata* 8(3):247-255.
- Kharkwal G (2009). Qualitative analysis of tree species in evergreen forests of Kumaun Himalaya, Uttarakhand, India. *Afr. J. Plant Sci.* 3(3):049-052.
- Khumbongmayum AD, Khan ML, Tripathi RS (2006). Biodiversity conservation in sacred groves of Manipur, northeast India: population structure and regeneration status of woody species. *Biodivers. Conserv.* 15:2439-2456.
- Korner C (1998). A re-assessment of high elevation treeline positions and their explanation. *Oecologia* 115:445-459.
- Lieberman D, Li M (1992). Seedling recruitment patterns in a tropical dry forest in Ghana. *J. Veg. Sci.* 3:375-382.
- Lorimer CG, Dahir SE, Nordhiem EV (2001). Tree mortality rates and longevity in mature and old growth hemlock-hardwood forests. *J. Ecol.* 89:960-971.
- Milchunas DG, Lauenroth WK (1993). Quantitative effects of grazing on vegetation and soils over a global range of environments. *Ecol. Monographs* 63:327-366.
- Mishra BP, Tripathi OP, Tripathi RS, Pandey HN (2004). Effect of anthropogenic disturbance on plant diversity and community structure of a sacred grove in Meghalaya, northeast India. *Biodivers. Conserv.* 13:421-436.
- Mishra BP, Tripathi RS, Tripathi OP, Pandey HN (2003). Effects of disturbance on the regeneration of four dominant and economically important woody species in a broad-leaved subtropical humid forest of Meghalaya, north east India. *Curr. Sci.* 84(11):1449-1453.
- Misra R (1968). *Ecological Work Book*. Oxford & IBH Publishing Company, Calcutta.
- Muller-Dombois D, Ellenberg H (1974). *Aims and methods of vegetation ecology*. John Wiley and Sons, New York.
- Negi SS (1995). *Uttarakhand: Land and People*. MD Publications PVT LTD, Delhi.
- Oliver CD, Larson BC (1990). *Forest Stand dynamics*. Government document, McGraw Hill, Inc. New York, U.S.
- Perira JS, Kozlowski TT (1977). Water relations and drought resistance of young *Pinus banksiana* and *Pinus resinosa* plantation trees. *Can. J. For. Res.* 7:132-137.
- Pielou EC (1975). *Ecological diversity*. Willey-Interscience, New York, pp. 165.
- Rao PB (1988). Effects of environmental factors on germination and seedling growth in *Quercus floribunda* and *Cupressus torulosa*, tree species of central Himalaya. *Ann. Bot.* 61:531-540.
- Rawal RS, Rawat B (2012). Nanda Devi Biosphere Reserve-west Himalaya, India. In: Palni LMS, Rawal RS, Rai RK, Reddy SV. *Compendium on Indian Biosphere Reserve. Progression during two decades of conservation*. Ministry of Environment and Forests, Government of India.
- Rawat B (2013). Changes in vegetation diversity and plant responses in Nanda Devi Biosphere Reserve over the last two decades. PhD thesis. Submitted to Kumaun University Nainital, Uttarakhand, India.
- Rawat B, Sekar KC, Gairola S (2013). Ethnomedicinal plants of Sunderdhunga valley, western Himalaya, India - traditional use, current status and future scenario. *Indian Forester* 139(1):61-68.
- Saxena AK, Singh JS (1984). Tree population structure of certain Himalayan forests and implications concerning the future composition. *Veg.* 58:61-69.
- Schulte PJ, Marshall PE (1983). Growth and water relations of black locust and pine seedlings exposed to controlled water-stress. *Can. J.*

- For. Res. 13: 334-338.
- Sekar KC, Rawat B (2011). Diversity, utilization and conservation of ethno-medicinal plants in Devikund - A high altitude, sacred wetland of Indian Himalaya. *Med. Plants* 3(2):105-112.
- Shankar U (2001). A case study of high tree diversity in a sa1 (*Shorea robusta*)- Dominated lowland forest of Eastern Himalaya: Floristic composition, regeneration and conservation. *Curr. Sci.* 81:776-786.
- Shannon CZ, Weiner W (1963). The mathematical theory of communications. Univ. Illinois Press, Urbane.
- Shrestha BB, Ghimire B, Lekhak HD, Jha PK (2007). Regeneration of treeline Birch (*Betula utilis* D. Don) forest in a trans-Himalayan dry valley in central Nepal. *Mountain Res. Dev.* 27:259-267.
- Silori CS (2001). Status and distribution of anthropogenic pressure in the buffer zone of Nanda Devi Biosphere in western Himalaya, India. *Biodivers. Conserv.* 10:1113-1130.
- Singh JS, Singh SP (1992). Forests of the Himalaya. Structure, Functioning and Impact of Man. Nainital, India: Gyanodaya Prakashan; and Delhi, India: Fine Art Press.
- Singh SP, Adhikari BS, Zobel DB (1994). Biomass productivity, leaf longevity and forest structure along an altitudinal gradient in central Himalaya. *Ecol. Monogr.* 64:401-421.
- Singh SP, Rawat YS, Garkoti SC (1997). Failure of brown oak (*Quercus semecarpifolia*) to regenerate in central Himalaya: a case of environmental semisurprise. *Curr. Sci.* 73:371-374.
- Spurr SH, Barnes BV (1980). *Forest Ecology*. John Wiley, New York, USA.
- Swaine MD, Lieberman D, Hall JB (1990). Structure and dynamics in a tropical dry forest in Ghana. *Veg.* 88:31-51.
- Todaria NP, Pokhriyal P, Uniyal P, Chauhan DS (2010). Regeneration status of tree species in forest of Phakot and Pathri Rao watersheds in Garhwal Himalaya. *Curr. Sci.* 98(2):171-175.
- Whittaker RH (1975). *Communities and ecosystems*, II edition. MacMillan Publishing Co. Inc., New York, pp. 385.
- Zobel DB, Singh SP (1997). Himalayan forests and ecological generalizations. *BioSci.* 47:735-745.

Full Length Research Paper

Chemical characterization and antimicrobial activity of essential oils and Croton's varieties modulator in the Brazilian's Northeast semiarid

Elissandra C. Angélico¹, Onaldo G. Rodrigues^{2*}, José G. M. da Costa³, Maria de Fátima A. Lucena², Vicente Queiroga Neto² and Rosália Severo de Medeiros²

¹Programa de Pós-Graduação em Ciência Animal, Universidade Federal de Campina Grande- UFCG, Av. Sta Cecília, Bairro Jatobá, s/n, 58700970, Patos-PB. Cx Postal 64, Tel. 83-3511-3000, Patos – PB, Brasil.

²Laboratório Multiusuário em Pesquisas Ambientais, Universidade Federal de Campina Grande- UFCG, Av. Sta Cecília, Bairro Jatobá, s/n, 58700970, Patos-PB. Cx Postal 64, Tel. 83-3511-3000, Patos – PB, Brasil.

³Laboratório de Pesquisa de produtos Naturais- LPPN- Universidade Regional do Cariri, URCA, Campus do Pimenta, 63105-000, Crato-Ceará, Brasil.

Received 10 June, 2014; Accepted 3 July, 2014

This study aimed to analyze the chemical composition and evaluate the antibacterial activity of essential oils and modulator of *Croton heliotropiifolius* Kunth and *Croton blanchetianus* Baill. The oils were obtained with distilled water and their components identified with GC/MS. Eucalyptol (16.9%), β -caryophyllene (15.9%) and germacrene-D (14.5%) for *C. heliotropiifolius* and cedrol (28.4%), eucalyptol (17, 4%) and α -pinene (10.5%) for *C. blanchetianus* were observed. The antimicrobial activity and minimum inhibitory concentration (MIC) was determined by the diffusion method and microdilution with standard bacteria Gram positive and negative. Preliminary results of the antibacterial activity showed that both oils were more effective against Gram-positive strain of *Bacillus cereus*. When verifying the minimum inhibitory concentration, it was observed that the essential oil of *C. heliotropiifolius* showed inhibitory activity only for the lineage multiresistance *Staphylococcus aureus* (MR 358), 512 μ g/mL with MIC. For the essential oil of *C. blanchetianus*, the result was significant for *S. aureus* with an MIC of 64 mg/mL. The oil of *C. blanchetianus* potentiated the antibiotic amikacin, kanamycin and gentamycin against *B. cereus* strain, showing a synergistic effect.

Key words: Phytochemistry, antimicrobial, bacterial resistance, secondary metabolites.

INTRODUCTION

A significant number of bacterial populations have acquired resistance to antimicrobial. Antibiotics are important therapeutic agents commonly used for the control of bacterial

infectious diseases. However, resistance to antibiotics has become a global public health problem. Bacterial infections caused by resistant strains are problems in

*Corresponding author. E-mail: onaldo@cstr.ufcg.edu.br.

numerous hospitals around the world, especially in patients compromised by age, illness and treatments with immunity-suppressant drugs (Lima et al., 2013). The knowledge on the therapeutic properties of medicinal plants obtained from the folk medicine, has been accumulated over centuries and the empirical knowledge often symbolizes the only therapy of various communities and ethnic groups. In Brazil, medicinal plants represent a rich source of which antimicrobial agents may be obtained. Plants are used medicinally in different infections and are a source of many potent and powerful drugs (Kubmarawa et al., 2007; Almeida et al., 2013).

Croton blanchetianus and *Croton heliotropiifolius*, popularly known as “velame-do-campo”, are shrub widely distributed in Brazilian north east occurring in vegetations (Ceará, paraíba and Pernambuco) (Gomes, 2006).

The interest on plants for investigations of new antimicrobials is due to the wide variety of chemical substances present in different parts of the plants with pharmacological action, as coumarin, flavonoids, terpenoids, alkaloids and tannins. In this sense, the most recent research involves screening of plant extracts and essential oils, to thus meet secondary metabolites with relevant biological activity. Presented for the first time in this work is the chemical composition of essential oil of *C. blanchetianus* and *C. heliotropiifolius* varieties obtained from the Caatinga's biome, and antimicrobial modulatory activities against bacterial strains.

MATERIALS AND METHODS

The leaves of *C. blanchetianus* and *C. heliotropiifolius* were collected in October and November 2009 at 8:00 p.m. at the site “São José do Bofim” municipality of Patos-PB, between geographic coordinates of latitude 07° 08' 20" S and longitude 037° 18' 06" W. Then a representative sample of each species was identified by Professor Maria de Fátima Souza and deposited at the Caatinga's Herbarium in the Federal University of Campina Grande (UFCG), under registration number # 496 and # 497, respectively.

The essential oils from fresh leaves of *C. blanchetianus* (820.0 g) and *C. heliotropiifolius* (652.95 g) were obtained by hydrodistillation using handset type Clevenger, for a period of two hours. Then the oil was dried with anhydrous sodium sulfate to remove excess water and kept in the refrigerator within 30 days until analysis. The identification of the essential oils chemical components of *C. blanchetianus* leaves and *C. heliotropiifolius* were obtained by gas chromatography-mass spectrometry (GC/EM), in a Hewlett-Packard model 5971 spectrometer operating at 70 eV ionization energy. Fused silica capillary column DB-5 (30m x 0.25 mm d.i., 0.25 µm film thickness) and helium carrier gas stream 1 mL/min split were used. The injector and detector temperatures were programmed from 250 to 200°C, respectively. The column temperature was programmed 35 and 180°C at 4°C/min and, then the 180°C at 280°C at 10°C/min. Mass spectra were obtained at 30 and 450 m/z. Individual components were identified by matching their mass spectra, 70 eV, with the database, using the library built and spectrometer with two other computers using retention indices as a pre-selection as well as by visual comparison of the fragmentation pattern with those reported in the literature.

Five standard bacterial strains used were ceded by the Oswaldo

Cruz Foundation for the preliminary assessment of the antibacterial activity of three Gram-negative: *Pseudomonas aeruginosa* (15442), *Klebsiella pneumoniae* (10031) and *Escherichia coli* (25922), two Gram-positive *Staphylococcus aureus* (ATCC 12692), *Bacillus cereus* (ATCC 33018) and a multiresistant strain isolated from clinical material *S. aureus* (MR 358), which have been revived amid middle Brain Heart Infusion (BHI) and incubated for 24 h at 37°C.

The antibacterial activity was determined by agar diffusion method by gel cavity. The bacteria were inoculated, using a swab sterile in Petri dishes previously prepared with Agar Muller-Hilton made cavities which (6 mm in diameter) is filled with 20 µL prepared with the essential oils of the plants solutions (diluted in DMSO) in the following concentrations: 10; 5; 2.5; 1.25; 0.6 and 0.3%. Then the plates were incubated at 37°C on the bacterial incubator for 24 h. Assays were performed in duplicate, together with the positive control antibiotic amikacin (30 µg) and chloramphenicol (30 µg) and as a negative control DMSO and distilled water. It was considered as the final result of each sample of the average of the measurements of the halos and susceptible to inhibition zone at or above 10 mm diameter corresponding to the diameter of the cavity in the culture medium.

The tested essential oils showed antimicrobial activity in the preliminary assessment submitted for the determination of minimum inhibitory concentration (MIC) by broth microdilution technique, based on the document CLSI/NCCLS M7-A6 to bacterium (NCCLS 2003).

The same pattern of the previous test strains, which were used, were inoculated in middle brain heart infusion broth (BHI) broth at 3.8% and incubated for 24 h at 35 ± 2°C. After this preculture, the standardization of the inoculum was done, which consisted of the preparation of a bacterial suspension in BHI at 3.8%, turbidity corresponding to 0.5 Scale McFarland (10⁸ UFC/mL). Then, this suspension was diluted to 10⁶ UFC/mL in broth BHI at 10%, and volumes 100 µL were then homogenized in 96 well microdilution plate supplemented with different concentrations of oil, resulting in a final inoculum of 5 x 10⁵ UFC/mL.

Solutions of essential oils were prepared using 10 mg of samples solubilized in 1 mL of DMSO (SIGMA, Brazil) obtaining an initial concentration of 10 mg/mL. From this concentration, dilutions were made in distilled water to obtain a stock solution of 1024 mg/mL. Final concentrations of the oils in the culture medium were 512, 256, 128, 64, 32, 16 and 8 µg/mL.

The tests were performed in duplicate and the plates were incubated at 35 ± 2°C for 24 h. A developer was added to each well, 25 µL of resazurin (sodium resazurin SIGMA, Brazil) prepared in sterile distilled water at a concentration of 0.01% (w/v) for a period of 30 min at room temperature. The negative control was gotten with 100 µL BHI broth plus the standardized bacterial inoculum. The minimum inhibitory concentration (MIC) was defined as the lowest concentration that is able to completely inhibit microbial growth in the microdilution wells as detected by the naked eye. The reading of the results for MIC determination was considered as positive for the wells that were blue and those who had negative staining, red coloring.

For the evaluation of the essential oil as a modulator of antibiotic activity, the MIC conventional aminoglycoside antibiotics (neomycin, kanamycin, amikacin and gentamicin), were determined in the absence and presence of the essential oil by the microdilution method, modified from the paper CLSI/NCCLS M7-A6 to bacterial (NCCLS 2003). Subinhibitory concentrations were used (MIC 1/8) in BHI at 10%. Two strains of bacteria transferred patterns were used in the test by Oswaldo Cruz Foundation, with two Gram-positive *B. cereus* (ATCC 33018) and *S. aureus* (ATCC 12692). The standard strains were selected because of the essential oil of *C. blanchetianus* providing the lowest MICs in antibacterial assays.

Solutions of the antibiotics were prepared with addition of distilled water to obtain a concentration corresponding to 1024 µg/mL. A

Table 1. Chemical constituents identified in the essential oils from the leaves of *C. heliotropiifolius* and *C. blanchetianus*.

| Constituent | <i>C. heliotropiifolius</i> | | <i>C. blanchetianus</i> | |
|----------------------|-----------------------------|-------|-------------------------|-------|
| | RI | % | RI | % |
| α -Pineno | 939 | 3.5 | 939 | 10.5 |
| β -Pineno | - | - | 980 | 3.0 |
| Sabineno | 976 | 2.4 | - | - |
| β -Mirceno | 991 | 5.5 | 991 | 1.5 |
| p -Cimeno | 1026 | 5.4 | 1026 | 4.2 |
| Eucaliptol | 1033 | 16.9 | 1033 | 17.4 |
| γ -Terpineno | 1062 | 4.5 | - | - |
| Linalool | 1098 | 1.1 | 1098 | 1.5 |
| Acetato de bornila | 1285 | 3.5 | - | 1.3 |
| Acetato de terpinila | 1346 | 1.7 | - | - |
| α -Copaeno | 1346 | 1.7 | - | - |
| β -Cariofileno | 1418 | 15.9 | 1418 | 3.8 |
| α -Humuleno | 1452 | 2.1 | 1452 | 1.3 |
| Germacreno D | 1480 | 14.5 | - | - |
| Biciclogermacreno | 1494 | 10.4 | - | - |
| δ -Cadineno | 1524 | 1.1 | - | - |
| Espatulenol | 1576 | 3.7 | 1576 | 2.8 |
| caryophyllene oxide | 1581 | 1.7 | 1581 | 1.2 |
| Viridiflorol | 1590 | 1.2 | - | - |
| Cedrol | - | - | 1589 | 28.4 |
| Alloaromadendreno | - | - | 1458 | 1.2 |
| p -Cimen-8-ol | - | - | 1189 | 1.3 |
| Criptona | - | - | 1186 | 1.3 |
| Monoterpenos | - | 13.2 | - | 39.2 |
| Sesquiterpenos | - | 80.7 | - | 10.3 |
| Total identified | | 98.8% | | 79.4% |

*Relative retention Indices (literature values).

volume of 100 mL of each solution of antibiotics was diluted seriously with wells containing BHI broth and 10% bacterial suspension diluted (1:10). Final concentrations of the antibiotics in the culture medium was 512, 256, 128, 64, 32, 16, 8, 4 and 2 μ g/mL. After incubation of the plates at 37°C for 24 h, the essential oils' action on the effect of antibiotics was demonstrated by the use of resazurin as specified above.

RESULTS AND DISCUSSION

The essential oils obtained with the leaves hydrodistillation showed yields of 0.075% and 0.72% for *C. heliotropiifolius* and *C. blanchetianus* relative to the weight of the fresh material used. By analyzing CG-EM it was possible to identify and quantify the constituents 23 (Table 1). The essential oil of *C. heliotropiifolius* constituents 18 account for 98.8%, with 13.2% monoterpenes and 80.7% identified sesquiterpenes. Among the compounds identified were eucalyptol (16.9%) as major compound followed by β -cariofileno (15.9%) and germacreno-D (14.5%). 15 components

were identified, *C. blanchetianus* (79.4%), being 39.2% monoterpenes and 10.3% sesquiterpenes constituents. The major compounds were cedrol (28.4%), eucaliptol (17.4%) and α -pineno (10.5%). The chemical profile of the oils was similar, but showed a different proportion species showing a high amount of oil of sesquiterpenes for *C. heliotropiifolius*, whereas for *C. blanchetianus* there is a predominance of monoterpenes. However, this chemical composition is consistent with literature data for *Croton* species whose essential oils are characterized by the predominance of monoterpenes and sesquiterpenes as major components. Silva (2008) with the essential oil extracted from the leaves of *C. heliotropiifolius* identified the presence of α -pinene, sabinene, linalool, bornila of acetate, beta-caryophyllene, germacrene D, δ -cadinene, α -humulene, bicyclogermacrene, spathulenol and eucalyptol as major compounds, corroborating the data of this study. According to Silva et al. (2010), the chemical composition of the essential oil of *C. sonderianus* presents the following constituents:

Table 2. Mean values of the halo of inhibition of bacterial growth in mm of the essential oil *C. blanchetianus*.

| Mean of inhibition zones (mm diameter) (Mean + standard deviation) | | | | | | | | | |
|--|----------------------|-----------------------|----------|--------|----------|-----|-----|----------|----------|
| Plants | Bacterial | Oil concentration (%) | | | | | | Control | |
| | | 10 | 5 | 2.5 | 1.25 | 0.6 | 0.3 | CLO | AMI |
| C. h* | <i>P. aeruginosa</i> | | | | | | | 23.5±2.1 | 23.5±0.7 |
| | <i>S. aureus</i> | 5.5±7.77 | | | | | | 0.0±0.0 | 19.5±0.7 |
| | <i>S. aureus</i> M.R | | | | | | | 21.0±1.4 | 21.5±3.5 |
| | <i>E. coli</i> | | | | | | | 0.0±0.0 | 19.0±4.2 |
| | <i>K. pneumoniae</i> | 5±7.07 | | | | | | 0.0±0.0 | 26.5±2.1 |
| | <i>B. cereus</i> | 11.5±2.12 | | | | | | 17.5±0.7 | 22.0±4.2 |
| C. b** | <i>P. aeruginosa</i> | - | 4.5±6.36 | - | - | - | - | 23.5±2.1 | 23.5±0.7 |
| | <i>S. aureus</i> | 4±5.65 | 3.5±4.94 | - | - | - | - | 0.0±0.0 | 19.5±0.7 |
| | <i>S. aureus</i> M.R | 6.5±9.19 | 5±7.07 | - | - | - | - | 21.0±1.4 | 21.5±3.5 |
| | <i>E. coli</i> | - | - | - | - | - | - | 0.0±0.0 | 19.0±4.2 |
| | <i>K. pneumoniae</i> | 4.5±6.36 | 5±7.07 | - | - | - | - | 0.0±0.0 | 26.5±2.1 |
| | <i>B. cereus</i> | 13.5±4.94 | 8.5±0.7 | 5±7.07 | 3.5±4.94 | - | - | 17.5±0.7 | 22.0±4.2 |

*Concentration used: (CLO) chloramphenicol 30 µg; (AMI) amikacin 30 µg.

Table 3. Values in mg/mL minimum inhibitory concentration (MIC) of essential oils from leaves of *C. heliotropiifolius* and *C. blanchetianus*.

| Bacterial | MIC (µg/mL) | |
|----------------------|-------------|--------|
| | EOCh* | EOCb* |
| <i>P. aeruginosa</i> | ≥ 1024 | ≥ 1024 |
| <i>S. aureus</i> | ≥ 1024 | 64 |
| <i>S. aureus</i> M.R | 512 | ≥ 1024 |
| <i>E. coli</i> | ≥ 1024 | 512 |
| <i>K. pneumoniae</i> | ≥ 1024 | ≥ 1024 |
| <i>B.cereus</i> | ≥ 1024 | 256 |

*EOCb: Essential oil of *C. blanchetianus* EOCh: Essential oil of *C. heliotropiifolius*.

spathulenol (18.32%), β-caryophyllene (14.58%) and caryophyllene oxide (8.54%), bicyclogermacrene (16.29%), β-phellandrene (15.42%) and β-caryophyllene (13.82%).

In this study, the essential oil extracted from the leaves of *C. heliotropiifolius* (Table 1) showed the presence of α-pinene, sabinene, linalool, bornil of acetate, beta-caryophyllene, germacrene D, δ-cadinene, α-humulene, bicyclogermacrene, spathulenol and eucalyptol as major compounds, corroborating the data of this study. The cedrol constituents, and alloaromadendreno criptona present in the essential oil of *C. blanchetianus* were only identified in this work, which may be related to the existence of chemotypes of this species found only in the Caatinga's biome.

As for the susceptibility testing (Table 2), for the preliminary determination of antibacterial activity, it was observed that both essential oils showed activity against

the Gram-positive strain *B. cereus* (ATCC 33018), with an average inhibition zone of 11.5 to 13.5 mm for *C. heliotropiifolius* and *C. blanchetianus* at a concentration of 10% as shown in Table 2. Gram-negative strains *Pseudomonas aeruginosa*, *Escherichia coli* and *Staphylococcus aureus* multiresistant, were resistant to the essential oil of *C. heliotropiifolius* as there was no formation on a zone of inhibition.

For oil *C. blanchetianus*, the *E. coli* strain was resistant at all concentrations, whereas in the other ones, the inhibition zone was less than 10 mm showing a high resistance of these microorganisms to the action of the components present in the oil at the concentrations tested. However, the demonstrated effect of essential oils on the same line can be attributed to the proportion of all components present in the oils. Regarding the assessment of the minimum inhibitory concentration (MIC) (Table 3), it was found that the essential oil of

Table 4. Values of MIC (mg/mL) of aminoglycosides in the absence and presence of the essential oil from the leaves of *C. blanchetianus*.

| Antibiotics | <i>S. aureus</i> (ATCC) | | <i>B. cereus</i> (ATCC) | |
|-------------|-------------------------|----------------|-------------------------|------------------|
| | MIC | EOCb (64µg/mL) | MIC | EOCb (256 µg/mL) |
| Amicacine | 8 | 8 | 8 | 4 |
| Neomicine | 32 | 32 | 8 | 8 |
| Kanamicine | 16 | 16 | 8 | 2 |
| Gentamicin | 2 | 2 | 4 | 2 |

*EOCb: Essential oil of *C. blanchetianus*.

C. heliotropiifolius showed inhibitory activity only for multidrug-resistant *S. aureus* strain (MR 358) with MIC 512 µg/mL. While the essential oil of *C. blanchetianus* showed inhibitory activity against *E. coli* strains (MIC 512 µg/mL), *B. cereus* (MIC 256 mg/mL) and *S. aureus* representing the most significant result with a lower MIC of 64 µg/mL. However, at the concentrations tested, the oils were not able to inhibit growth of the strains *K. pneumoniae* and *P. aeruginosa* MIC \geq considering 1024 µg/mL.

Given the results, it is observed that the Gram-negative strains were resistant to the action of oils, mainly the species *C. heliotropiifolius*, thus, absence of this inhibitory activity of essential oils on the Gram-negative bacteria. Silva et al. (2007) perhaps showed that the existing structural differences between bacteria, and Gram-positive are considered more sensitive to exposure to antibacterial products while, in general, negative bacteria were more resistant to the action with this essential oil because they had a rich outer membrane lipopolysaccharide, responsible for the hydrophilic surface, hindering the penetration of hydrophobic substances such as the constituents of many essential oils. Gonçalves (2007) reported that the extract of *C. blanchetianus* showed resistance against the strains *E. coli*, *S. epidermidis*, *S. aureus* and *S. typhimurium*. For *C. heliotropiifolius*, germacrene-D identified as the major constituent, proved to be inactive against *Bacillus subtilis*, *B. cereus*, *S. aureus*, *P. aeruginosa* and *E. coli*, as assessed by the methods of diffusion Agar and microdilution broth (Deuschle et al., 2007).

Studies on *C. blanchetianus* and *C. heliotropiifolius* species with antibacterial activity are still lacking, probably due to the fact that these species are native to the Caatinga with seasonal variations and mainly produce oils with low income. However, antimicrobial activity with essential oils to other species of general Croton, as *Croton nepetaefolius*, *Croton tiglium* and *Croton lechleri* L. is observed (Froldi et al., 2009).

Due to the complex nature of the essential oils, it would be difficult to give attribute to the activity observed at any single component present in it. Therefore, to know the mode of action of the oils it would be necessary to

examine separately each component of the essential oil and mixture to see if the chemical constituents have single or synergy inhibition.

The bacterial strains that showed the lowest inhibitory concentrations (MICs) were referred for modulating the activity of the essential oil against aminoglycosides.

In the result analysis (Table 4), it was observed that the essential oil of *C. blanchetianus* did not potentiate the activity of aminoglycoside antibiotics when tested against the *S. aureus* strain in 64 µg/mL concentration (MIC 1/8), the oil demonstrated a synergistic action on the activity of kanamycin, amikacin and gentamicin in the interaction with the strain *B. cereus*, with the most significant value of kanamycin with reduced MIC of 8 to 2µg/mL.

Oliveira et al. (2006) showed that the essential oil of some plants species in combination with the antibiotic gentamicin showed an antagonistic effect on *S. aureus* strain. Rodrigues et al. (2009) reported that the antibiotic activity gentamicin against *P. aeruginosa* was enhanced in the presence of the essential oil of *Croton zehntneri*, having a synergistic effect. However, the literature has not yet reported on the potentiating action of essential oils of the species studied with regards to the activity of aminoglycoside antibiotics.

Generally, the characteristics of the interference exerted by the essential oil on the antibiotic action varies according to the type of the antibiotic, the type of essential oil in combination tested, and the type of bacterial strain tested.

Conclusions

From the results obtained, it is concluded that the chemical constituents present in the essential oils of plants belonging to the class of monoterpenes and sesquiterpenes, have major compounds such as eucalyptus, β -caryophyllene and germacrene-D in the species *C. heliotropiifolius* and cedrol, eucalyptol and α -pinene in *C. blanchetianus*. Considering these results, we can concluded that the plants' essential oils when evaluated at a concentration of 10% have a higher antibacterial activity against the Gram-positive *B. cereus*

strain. Regarding the evaluation of the minimum inhibitory concentration, the essential oil of *C. heliotropifolius* is only active against the multidrug-resistant *S. aureus* strain with an MIC of 512 mg/mL. With the essential oil of *C. blanchetianus*, the antibacterial action with a MIC ranging 512-64 mg/mL proves to be more efficient for *S. aureus* (64 mg/mL). The essential oil of *C. blanchetianus* acts synergistically with antibiotics amikacin, kanamycin and gentamicin against the strain *B. cereus*. Although essential oils do not present inhibitory activity against all pathogenic strains tested, these results are promising and indicate that they are a source of natural products that possess antibacterial activity thus providing an important contribution to enlarge the biological knowledge of the species.

Conflict of Interests

The author(s) have not declared any conflict of interests.

ACKNOWLEDGEMENT

The National Council for Scientific and Technological Development (CNPq) is acknowledged for financial support.

REFERENCES

- Almeida TS, Rocha JBT, Rodrigues FFR, Campos AR, Costa JGM (2013). Chemical composition, antibacterial and antibiotic modulatory effect of *Croton campestris* essential oils. *Ind. Crops Prod.* 44:630-633.
- Deuschle R, Camargo T, Alves SH (2007). Fracionamento do extrato diclorometânico de *Senecio desiderabilis* Velloso e avaliação da atividade antimicrobiana. *Revista Brasileira de Farmacognosia.* 17: 71-75.
- Froldi AG, Zagottob G, Fillipini CR (2009). Activity of sap from *Croton lechleri* on rat vascular and gastric smooth muscles. *Phytomed.* 16:768-775.
- Gomes APS (2006). Revisão das espécies sulamericanas de *Croton L. subgen. Croton sect. Argyroglossum Baill.* (Crotonoideae - Euphorbiaceae). 124p. Tese de Doutorado. Universidade Federal Rural de Pernambuco, Recife-PE, Brasil.
- Gonçalves AL (2007). Estudo da atividade antimicrobiana de algumas árvores medicinais nativas com potencial de conservação/recuperação de florestas tropicais. 209p. Tese de Doutorado, Programa de Pós-graduação em Ciências Biológicas, Universidade Estadual Paulista.
- Kubmarawa D, Ajoku GA, Enwerem NM, Okorie DA (2007). Preliminary phytochemical and antimicrobial screening of 50 medicinal plants from Nigeria. *Afr. J. Biotechnol.* 14:1690-1696.
- Lima TB, Pinto MFS, Ribeiro SM, de Lima LA, Viana JC, Júnior NG, Cândido ES, Dias SC, Franco OL (2013). Bacterial resistance mechanism: what proteomics can elucidate: Review. *Faseb J.* 27:1291-1303.
- NCCLS- National Committee for Clinical Laboratory Standards (2003). *Methods for Dilution Antimicrobial Susceptibility Tests for bacteria that grow aerobically.* 6. ed. Wayne, PA: NCCLS Approved Standard M7-A6.
- Oliveira RAG, Lima EO, Vieira WL (2006). Estudo da interferência de óleos essenciais sobre a atividade de alguns antibióticos usados na clínica. *Revista Brasileira de Farmacognosia.* 16:77-82.
- Rodrigues FFG, Costa JGM, Coutinho HDM (2009). Synergy effects of the anybiotcs gentamicin and the essential oil of *Croton zehntneri*. *Phytomed.* 16:1052-1055.
- Silva JG, Souza IA, Higino JS (2007). Atividade antimicrobiana do extrato de *Anacardium occidentale* Linn em amostras multirresistentes de *Staphylococcus aureus*. *Revista Brasileira de Farmacognosia.* 17:572-577.
- Silva FKS (2008). Contribuição ao Estudo Fitoquímico de *Croton rhamnifolius* (Euphorbiaceae). 162f. Dissertação (Mestrado em Química Orgânica) - Universidade Federal do Ceará, Fortaleza.
- Silva LPM, Maia PVM, Nascimento TMG, Moraes JSC, Souza SMC, Lahlou SAN, Cardoso JHL (2010). *Croton sonderianus* essential oil samples distinctly affect airway smooth muscle (Report). *Int. J. Phytother. Phytopharmacol.* 17:721-725.



African Journal of Plant Science

Related Journals Published by Academic Journals

- *International Journal of Plant Physiology and Biochemistry*
- *Journal of Botany and Plant Pathology*
- *African Journal of Food Science*
- *International Journal of Biodiversity and Conservation*
- *Journal of Yeast and Fungal Research*

academicJournals