

# Study of a chemotaxonomic marker able to identify the genus *Aquilaria* (Thymelaeaceae)

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**Photo 1.**  
*Aquilaria rugosa*: flowers.

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## RÉSUMÉ

### Étude d'un marqueur chimiotaxonomique capable d'identifier le genre *Aquilaria* (Thymelaeaceae)

Le genre *Aquilaria* Lam. (Thymelaeaceae) comprend 21 espèces d'arbres (*The Plant List*, 2013) et se trouve principalement en Asie du Sud-Est. Lorsque l'arbre est infecté (champignons, bactéries), son bois devient brunâtre ou noirâtre (appelé bois d'agar) en raison de la sécrétion d'une oléorésine en réaction à l'infection. La résine est très parfumée et a été recherchée et utilisée pendant des siècles par les bouddhistes, les hindous et les musulmans pour faire de l'encens pour les cérémonies religieuses. Cette oléorésine se trouve principalement dans les espèces du genre *Aquilaria*, mais aussi dans quelques espèces des genres *Gyrinops* Gaertner et *Gonystylus* Teijsmann & Binnendijk. Il est difficile de faire la distinction entre ces espèces, et ce manque de connaissances taxonomiques a conduit à une surutilisation des arbres, mettant en danger ces espèces endémiques inscrites à l'Annexe II de la CITES. Nous avons utilisé la chimiotaxonomie comme outil de discrimination pour analyser les molécules polyphénoliques, métabolites secondaires, qui sont connus pour agir comme marqueurs taxonomiques dans d'autres plantes. En utilisant une technique d'analyse simple, efficace et peu coûteuse (chromatographie bidimensionnelle en couche mince), nous avons trouvé la même molécule polyphénolique dans les six espèces du genre *Aquilaria* étudiées, qui a été identifiée comme mangiférine par analyse colorimétrique et chromatographique en comparaison avec la mangiférine de contrôle. Au cours de ces analyses, nous avons trouvé une relation entre les genres *Aquilaria* et *Gyrinops* (botaniquement démontrée) par l'existence occasionnelle de mangiférine chez trois espèces du genre *Gyrinops*. Nous avons également constaté qu'un laps de temps de cent-quarante ans entre deux échantillons d'herbier de la même espèce n'avait pratiquement aucun effet sur la concentration de mangiférine dans la plante. Ces travaux ont montré les mérites de la chimiotaxonomie dans la recherche de marqueurs taxonomiques et l'originalité de l'analyse colorimétrique d'un métabolite de plante chromatographié en couche mince.

**Mots-clés :** *Aquilaria*, Thymelaeaceae, chimiotaxonomie, chromatographie bidimensionnelle sur couche mince, mangiférine, Asie du Sud-Est.

## ABSTRACT

### Study of a chemotaxonomic marker able to identify the genus *Aquilaria* (Thymelaeaceae)

The genus *Aquilaria* Lam. (Thymelaeaceae) comprises 21 tree species (*The Plant List*, 2013) and is mostly found in Southeast Asia. When the tree is infected (fungi, bacteria), its wood turns brownish or blackish (then called agarwood) due to the secretion of an oleoresin as a reaction to the infection. The resin is very fragrant and has been sought after and used for centuries by Buddhists, Hindus and Muslims to make incense for religious ceremonies. This oleoresin is mainly found in species of the genus *Aquilaria*, but also in a few species of the genera *Gyrinops* Gaertner and *Gonystylus* Teijsmann & Binnendijk. It is difficult to distinguish between these species, and this lack of taxonomic knowledge has led to over-use of the trees and endangering these endemic species listed in CITES Appendix II. We used chemotaxonomy as a discrimination tool to analyse polyphenolic molecules, which are secondary metabolites and known to act as taxonomic markers in other plants. Using a simple, effective and inexpensive analysis technique (two-dimensional thin-layer chromatography), we found the same polyphenolic molecule in the six species of the genus *Aquilaria* we studied, which was identified as mangiferin by chromatographic and colorimetric analysis in comparison with a mangiferin control molecule. During these analyses, we found a relationship between the genera *Aquilaria* and *Gyrinops* (*botanically demonstrated*) in the occasional existence of mangiferin in three species of the genus *Gyrinops*. We also found that a time laps of 140 years between two herbarium samples of the same species had virtually no effect on the mangiferin concentration in the plant. This study showed the merits of chemotaxonomy in seeking taxonomic markers, and the originality of colorimetric analysis of a thin-layer chromatographed plant metabolite.

**Keywords:** *Aquilaria*, Thymelaeaceae, chemotaxonomy, two-dimensional thin-layer chromatography, mangiferin, Southeast Asia.

## RESUMEN

### Estudio de un marcador quimiotaxonomico capaz de identificar al género *Aquilaria* (Thymelaeaceae)

El género *Aquilaria* Lam. (Thymelaeaceae) comprende 21 especies de árboles (*The Plant List*, 2013) y se encuentra principalmente en el Sudeste Asiático. Cuando el árbol está infectado (hongos, bacterias), su madera se convierte en oscura o negruzca (llamada madera de agar) a causa de la secreción de una oleorresina como reacción a la infección. La resina es muy perfumada y ha sido muy buscada y utilizada durante siglos por budistas, hindúes y musulmanes para producir incienso para las ceremonias religiosas. Esta oleorresina se encuentra principalmente en las especies del género *Aquilaria*, pero también en algunas especies de los géneros *Gyrinops* Gaertner y *Gonystylus* Teijsmann & Binnendijk. Es difícil distinguir estas especies, por lo que la falta de conocimientos taxonómicos ha llevado a una sobreutilización de los árboles, poniendo en peligro estas variedades endémicas inscrites en el anexo II de la CITES. Hemos utilizado la quimiotaxonomía como herramienta de discriminación para analizar las moléculas polifenólicas, metabolitos secundarios conocidos por actuar como marcadores taxonómicos en otras plantas. Utilizando una técnica de análisis simple, eficaz y barata (cromatografía bidimensional en capa fina), hemos encontrado la misma molécula polifenólica en las seis especies del género *Aquilaria* estudiadas, que ha sido identificada como mangiferina por análisis colorimétrico y cromatográfico en comparación con la mangiferina de control. Durante estos análisis, hemos encontrado una relación entre los géneros *Aquilaria* y *Gyrinops* (botánicamente demostrada) por la existencia ocasional de mangiferina en tres especies del género *Gyrinops*. También hemos constatado que un lapso de tiempo de ciento cuarenta años entre dos muestras de herbario de la misma especie no producía prácticamente ningún efecto en la concentración de mangiferina en la planta. Estos estudios han demostrado el valor de la quimiotaxonomía en la búsqueda de marcadores taxonómicos y la originalidad del análisis colorimétrico de un metabolito de planta cromatografiado en capa fina.

**Palabras clave:** *Aquilaria*, Thymelaeaceae, quimiotaxonomía, cromatografía bidimensional en capa fina, mangiferina, Sudeste Asiático.

## Introduction

The genus *Aquilaria* Lam. (Thymelaeaceae) is widespread from India to the island of New Guinea and more particularly in Southeast Asia as far as southern China. When the wood of certain species of *Aquilaria* is wounded and/or contaminated by a pathogen (e.g. a fungus), its defence strategy consists in secreting a very fragrant oleoresin, which plays a role in transforming the heartwood, which then becomes known as “Agarwood” (or Eagle Wood or Oud or Gaharu...).

The oleoresin secreted in the wood consists of a complex mixture of sesquiterpenes, phenylethylchromones and minor aromatic compounds, which give the warm, persistent and much appreciated aroma that particularly develops when the wood is heated. Agarwood has been used by Buddhists, Hindus and Muslims for centuries to make incense used in religious ceremonies (Qi *et al.*, 2005). With its medicinal properties, this oleoresin also plays an important role in traditional Chinese, Tibetan and Ayurvedic medicines, and is used as a sedative, analgesic, cardiostimulant, antirheumatic, etc. (Ueda *et al.*, 2006; Feng *et al.*, 2009; Qi *et al.*, 2009). The essential oils obtained by distilling agarwood are used in perfumery (such as “M7”, a perfume by Yves Saint Laurent).

It is worth noting that the economic value of the products obtained from *Aquilaria* is very high; 1 kg of essential oil obtained from agarwood costs at least 10,000 USD and 1 kg of incense obtained from the same wood costs around 1,000 to 1,300 USD.

The chemical composition of the oleoresin has already been analysed (see overview by Naef, 2011), but it cannot be used to sufficiently distinguish between the species. Oleoresin is mostly produced by species of the genus *Aquilaria* but also by some species of the genera *Gyrinops* Gaertner and *Gonystylus* Teijsmann & Binnendijk. The species of the last two genera are also difficult to distinguish from each other, but they might be a substitute for *Aquilaria* agarwood and form part of a preservation policy. This lack of taxonomic knowledge for these Asian species producing agarwood has led to over-use of the trees, endangering 17 endemic species of *Aquilaria* (Mabberley, 2008), which are all listed in CITES Appendix II.

As commercial use of these oleoresins is only possible with reliable knowledge of the species and with strict traceability, we used a chemotaxonomy analysis as a discrimination tool.

To that end we opted to study polyphenols, which are secondary metabolites found throughout the Plant Kingdom, but in highly varied forms. A composition difference in these metabolites is less often due to a fluctuation in the surrounding environment than to a genotypic difference, hence the taxonomic value of these substances (Bell, 1981). Polyphenol molecules are mainly synthesized in leaves and their role as chemotaxonomy markers has been demonstrated in numerous plants (Harborne *et al.*, 1971; Bate-Smith and Richens, 1973; Cooper-Driver and Swain, 1977; Andary *et al.*, 1988b; Williams *et al.*, 1988; Andary *et al.*, 1992).

Earlier research showed the existence of several types of polyphenols in *Aquilaria* spp. leaves: benzophenones (e.g. iriflophenone; Feng *et al.*, 2009), xanthone (e.g. mangiferin; Qi *et al.*, 2009), flavonoids (e.g. genkwanin, derivatives of luteolin and apigenin; Hara *et al.*, 2008; Qi *et al.*, 2009).

We conducted this study by comparing a batch of 40 specimens belonging to the genus *Aquilaria* (botanically analyzed) collected and dried in Laos and Cambodia, and some reference samples from the herbaria of Paris Museum (P) and Leiden (L). The latter samples corresponded to voucher species belonging to some close genera: *Aquilaria*, *Gyrinops*, *Gonystylus* and *Wikstroemia* Endlicher. *Aquilaria* and *Gyrinops* are classified in the sub-family of the *Aquilarioideae*, *Gonystylus* in that of the *Gonystiloideae* and *Wikstroemia* in that of the *Thymelaeoideae*. These three sub-families, with that of the *Gilgiodaphnoideae*, make up the family of the Thymelaeaceae (Hou, 1960). These sub-families, apart from the *Gilgiodaphnoideae*, are endemic to southern, tropical Asia.

We also took as an example of the use made of the genus *Aquilaria*, other than for its oleoresin, a quality study involving the analysis of a herbal tea comprising *Aquilaria* spp. leaf fragments, marketed as “Agarwood Tea”.

This chemotaxonomy analysis was carried out by two-dimensional thin-layer chromatography (quick and inexpensive method). The nature of the studied molecule was suggested by the position of each spot on the chromatogram, confirmed by comparison with the reference molecule and by a colorimetric analysis of the amount of colour in the Red, Green and Blue channels. This approach was totally original.



**Photos 2.**  
*Aquilaria rugosa*: flowers (a); fruits (b); tree (c);  
trees in plantation (d).

## Methodology

### Plant material

Forty specimens of *Aquilaria* spp. from Laos (24) and Cambodia (16) were collected and dried. We also studied a sample of *Aquilaria crassna* Pierre ex. Lecomte collected and dried in 2010 and kindly offered by Professor Kiet Le Cong in Vietnam.

Ten voucher specimens were provided by the National Natural History Museum (MNHN) in Paris (P):

- *Aquilaria agallocha* Roxb., Vidal 5800 (Thailand);
- *Aquilaria baillonii*, Chevalier 38441 (Indochina);
- *Aquilaria baillonii*, Poilane 30000 (Indochina);
- *Aquilaria crassna* Pierre ex Lecomte, Bejaud 364 (Cambodia);
- *Aquilaria crassna* Pierre ex Lecomte, Chevalier 1365 (Indochina);
- *Aquilaria crassna* Pierre ex Lecomte, Pierre 3619 Holotype (Cambodia);
- *Aquilaria malaccensis* Lam., Griffith 4382 (Burma/Malaysia);
- *Aquilaria sinensis* (Lour.) Spreng., Lei 864 (Hainan);
- *Aquilaria sinensis* (Lour.) Spreng., Bodinier 134 (Hong-Kong);
- *Aquilaria sinensis* (Lour.) Spreng., Chan 1047 (Hong-Kong).

And 12 specimens were provided by the National herbarium of Leiden (L), the Netherlands:

- *Aquilaria rugosa* L. C. Kiet & Kessler, Kiet 1940 Holotype (Vietnam);
- *Aquilaria rugosa* L. C. Kiet & Kessler, Kiet 1941 (Vietnam);
- *Gyrinops caudata* (Gilg) Domke, Hou 10629 (Indonesia);
- *Gyrinops ledermannii* Domke, Singadan 02 (New Guinea);
- *Gyrinops walla* Gaertn., Ridsdale 465 (South India);
- *Gonystylus bancanus* (Miq.), Kurz Awang Awang Irang SAN 97436 (Malaysia);
- *Gonystylus confusus* Airy Shaw, Niyomdham 960 (Thailand);
- *Gonystylus velutinus* Airy Shaw, Ambri W528 (Indonesia);
- *Wikstroemia indica* (L.) C.A. Mey., Hou 7612 (Thailand);
- *Wikstroemia meyeniana* Warb., Hou 6279 (Cambodia);
- *Wikstroemia polyantha* Merr., Maxwell 76-558 (Thailand);
- *Wikstroemia ridleyi* Gamble, Niyomdham & Sriboonma 1631 (Thailand).

Altogether, we analysed 6 *Aquilaria* species and 10 species belonging to three other genera: *Gyrinops*, *Gonystylus*, and *Wikstroemia*.

### Chromatographic analyses

For these chemotaxonomy studies, we chose to analyse polyphenol compounds from dried leaves. All the solvents were of “analysis” quality (VWR, France) and were as follows: acetic acid, hydrochloric acid, dichloromethane, ethanol and methanol.

### Preparation of plant extracts

The dried leaves were weighed out and ground in an electric grinder, then extracted at a rate of 0.2 g of plant in 10 ml of 80% methanol. Extraction was achieved by magnetic stirring at room temperature for 1 h. The resulting extract was filtered through filter paper (Labover, France) and the filtrate stored in brown glass bottles.

### Two-dimensional thin-layer chromatography (2D TLC)

Two-dimensional thin-layer chromatography involves using thin layers of cellulose industrially deposited on 20 x 20 cm aluminium plates (ref. 5552, Merck). Each plate was cut into four 10 x 10 cm squares. After the extract to be analysed had been deposited on one of the 10 x 10 cm squares, it underwent successive migrations perpendicular to each other (called first migration and second migration) in glass tanks (dimensions: 10 x 10 x 8 cm, ref. 022.510; CAMAG, France), using the following migration solvents:

- For type A polyphenols (flavonoids, coumarins, caffeic acid derivatives, xanthenes, etc.):
- solvent 1 (first migration): ethyl acetate – acetic acid – water (49:45:15; v/v);
- solvent 2 (second migration): ethyl acetate – acetic acid – water (10:30:70; v/v).

Solvent migration was halted 0.5 cm from the top of the plate for each solvent.

- For type B polyphenols (monomeric and oligomeric catechic tannins):
- a single migration with acetic acid – water (2:98; v/v).

Solvent migration was halted 0.5 cm from the top of the plate.

### Reference extract and molecule depositing

The reference molecules (supplied by Extra-Synthèse or Roth Sochiel, France) were as follows: acacetin, apigenin, apigenin-7-O-glucoside, caffeic acid, chorogenic acid, daphnetin, esculetin, fraxetin, genkwanin, isoorientine, kaempferol, luteolin, luteolin-7-O-glucoside, mangiferin, orientin, quercetin, scopoletin, rutin and vitexin.

Each reference molecule was solubilized to 0.5/1,000 in methanol then deposited on a chromatography plate at a rate of 1 µl of solution with a 1 µl micropipette (Microcaps Drumond, France).

Each plant extract to be analysed was deposited on a chromatography plate at a rate of 15 µl (using a 5 µl micropipette, Microcaps Drumond, France).

The chromatography of each extract was repeated three times.

### Mangiferin internal standard technique

We also applied the mangiferin internal standard technique for all the species: 1 µl of mangiferin standard solution was added to the 15 µl of plant extract deposited on the thin layer of cellulose. This result was compared to a chromatograph of the same plant extract without added mangiferin.

## Results

### Visualization and interpretation of the chromatograms

The plates were carefully dried with an electric hairdryer following each migration.

The chromatograms were observed either in natural light, or under UV light, before and after spraying (sprayer ref. 022.6100, CAMAG, France) and drying the reagent specific to each of the two chemical groups (type A and type B):

- For type A polyphenols (flavonoids, coumarins, caffeic acid derivatives, xanthenes):
- Neu's reagent (Neu, 1956): 1% amino-2-ethylidiphenylborinate in methanol (Sigma-Aldrich, France).

Type A polyphenols appear under UV light with specific fluorescences, ranging from blue to red, with intermediate colours depending on the chemical group. The colours are as follows (Dai *et al.*, 1995):

- flavonoids: yellow fluorescence in general but can range from yellow to red;
- coumarins: fluorescence that can range from blue to whitish blue;
- caffeic acid derivatives: more or less bright whitish blue fluorescence;
- hydroxyxanthenes: more or less pale yellow fluorescence.

However, the need to photograph the chromatograms to record the phenolic profiles of each extract sometimes raises a problem of colour rendition. For example, mangiferin, which, is seen with the naked eye as pale yellow fluorescence, appears more whitish on a photo.

- For type B polyphenols (flavan nuclei making up the more or less polymerized catechic tannins):
- DMCA reagent (dimethylaminocinnamaldehyde (Merck, France) at 0.1% in HCl-MeOH (10:90); Mc Murrouch and McDowell, 1978).

Catechic tannins appear with green colorations for polymers and blue for oligomers, under natural light, without noteworthy fluorescence under UV light.

### Chromatogram photography technique after visualization

Each chromatogram was placed in a photo chamber lit by a UV lamp (giving wavelengths of 254 or 366 nm, 2 x 8 W, Ref: 022.970, CAMAG, France). There was a hole at the top of the chamber to position a camera (Compatc, Panasonic – Lumix, DMC-FS10: 12 megapixels) to photograph the chromatograms (figure 1).

The distances and shots were identical for all the chromatograms.

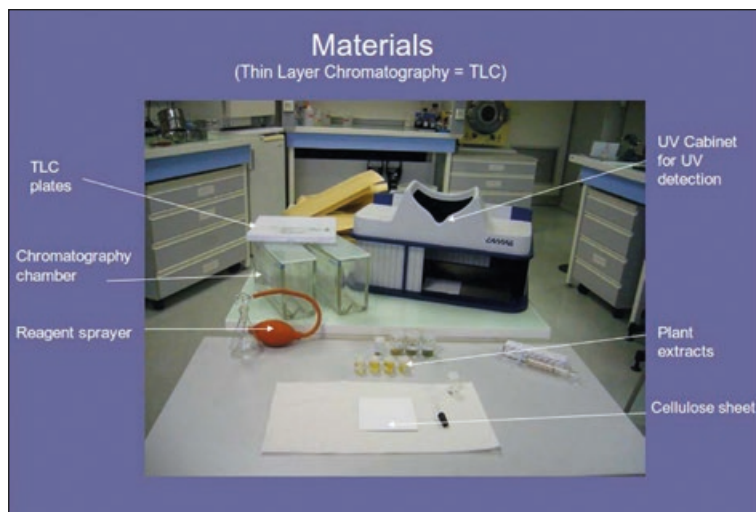
### Colour analysis method for fluorescence spots

After chromatography and photographing of the chromatogram, the fluorescent spots of the Type A molecules were measured by software (GIMP image editor, Google) that measured the colour of the molecule for the three channels, Red, Green and Blue (as a %), along with the Hue (in °) and Saturation (as a %).

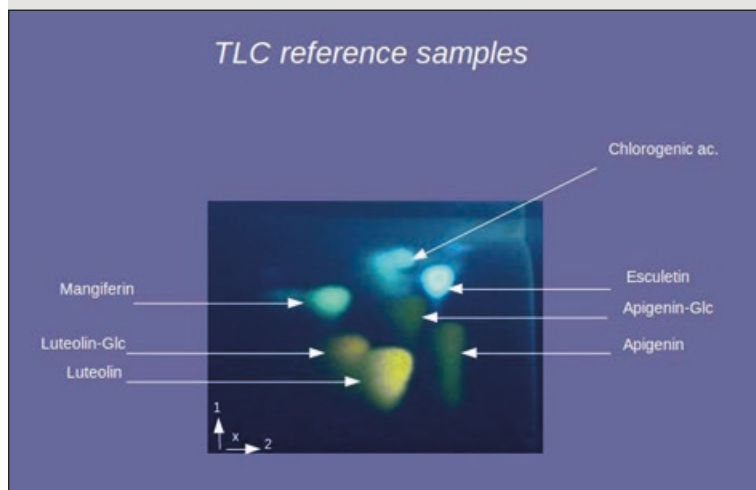
Our measurements were taken in the centre of the spot and validated by 3 replications.

Figure 2 shows the chromatographic image of a mixture of several reference molecules likely to be found in *Aquilaria* or other close genera (*Gyrinop*, *Gonystylus*, *Wikstroemia*) as mentioned in the introduction. In addition, colorimetric analyses of coloured spots corresponding to those molecules also showed us the possibility of distinguishing each molecule (table I).

An initial, unexpected result, was found when we compared holotype N° 3619 of *Aquilaria crassna* Pierre ex. Lecomte, collected in 1870 and conserved at the MNHN herbarium (P), with a sample of the same species collected and dried in 2010 by Professor Kiet Le Cong in Vietnam. When analysed under the same conditions, these two samples gave an analogous chromatographic image despite the 140-year difference in age between the two specimens, with only slightly toned-down fluorescence for the older one (figure 3).



**Figure 1.** Visualization of the thin-layer chromatography (TLC) material.



**Figure 2.** 2D TLC (two-dimensional thin-layer chromatography) of the different standard molecules after visualization with Neu's reagent. 1 and 2 are the order and two directions of migration of the two chromatographic solvents. X is the deposit of the solution to be chromatographed.

In the different species of the genus *Aquilaria*, we were able to identify mangiferin (C-glucoside of dihydroxyxanthone) (figure 4) by chromatographic and colorimetric comparison (measurement of colour parameters) with a reference mangiferin molecule (also used as an internal standard). In fact, the colorimetric analysis of all the spots corresponding to mangiferin found in the different taxa analysed gave results equivalent to the mangiferin standard (tables IIa and IIb). Alongside this molecule, we also found several flavonoids of the flavone group (particularly derivatives of luteolin and apigenin), along with some coumarins.

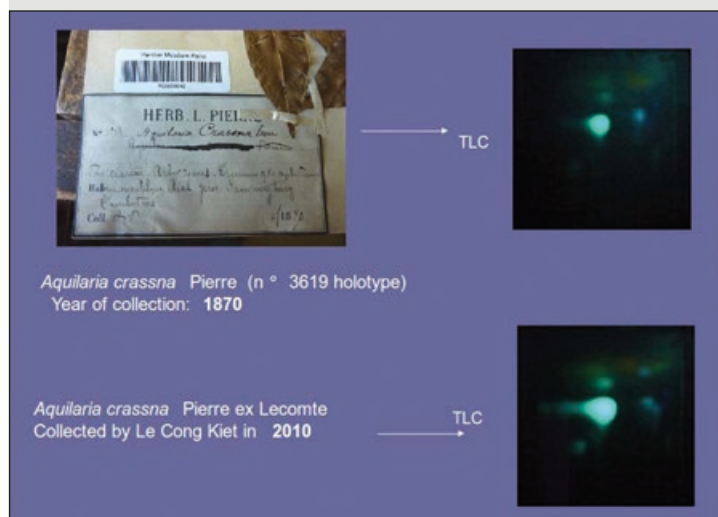
However, out of the six species of the *Aquilaria* genus studied, only one, *A.baillonii* (also noted as *Gyrinopsis baillonii* by Poilane) bearing the herbarium number 30000 (Poilane herbarium, MNHN), did not reveal the existence of mangiferin on the chromatographs (figure 4), whereas in the same MNHN herbarium, another sample bearing the same name, *A. baillonii*, number 38441 (Chevalier herbarium, MNHN) effectively corresponded to the expected chromatographic profile (with mangiferin) for species of the genus *Aquilaria*.

Within the genera *Gonystylus* and *Wikstroemia*, we were able to verify the absence of mangiferin in the different species studied. Indeed, we used the internal standard technique for mangiferin on all these species (see Methodology).

In addition, with the *W. indica* chromatogram (figure 5), we were able to see the presence of caffeic acid derivatives (dihydroxycinnamic acid derivatives) recognizable by their typical blue fluorescence, which we also found in the four species of *Wikstroemia* analysed.

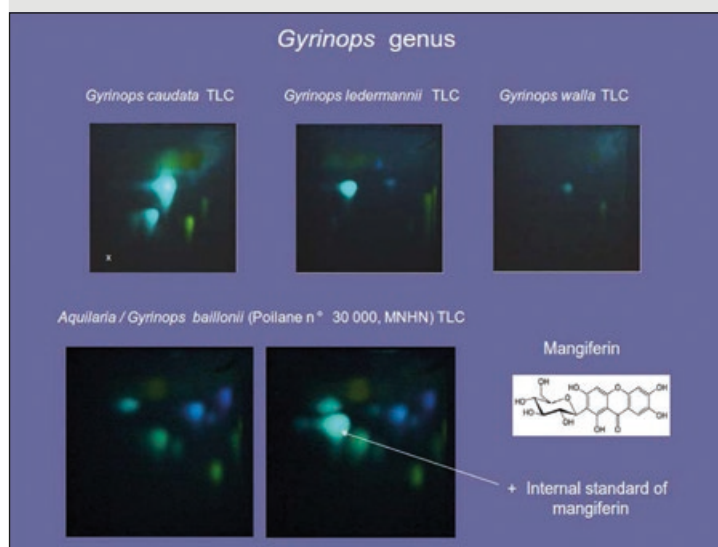
The analysis of the type B polyphenols (molecules with a flavan nuclei ex: tannins) showed a variable standard pattern that was difficult to interpret by our chromatographic tests, be it in *Aquilaria* or in the other genera. An example of the visualization of these tannins can be seen in figure 6 with sample N° 2, which shows an overall image of the tannins found in *Aquilaria*.

We then analysed the dried samples from Laos (24 samples) and Cambodia (16 samples) labelled "*Aquilaria*". The chromatographic profile of each sample was compared to that of the different voucher species (cf. Leiden (L) and Paris herbaria (P)). All these samples displayed a polyphenol profile very close to the standard schematic profile found in the genus *Aquilaria*.



**Figure 3.**

2D TLC comparing a sample from the holotype for the species *Aquilaria crassna* with the sample of the same species collected 140 years later.



**Figure 4.**

Chromatographic (2D TLC) comparison between three species of *Gyrinops*: *G. caudata*, *G. ledermannii* and *G. walla*. Use of a mangiferin internal standard on the 2D TLC of the species *Aquilaria/Gyrinops baillonii* to check for the absence of that molecule in the species.

**Table I.**

Assessment of the amount of Colour (as a %), Hue (in °) and Saturation (as a %) by standard-molecule for the three channels: red, green and blue (R,G, B); Glc: glucoside.

	Mangiferin	Luteolin	Luteolin-Glc	Apigenin	Apigenin-Glc	Chlorogenic acids	Esculetin
Red	15	71	89	31	32	72	79
Green	100	73	92	41	42	100	100
Blue	100	39	46	29	31	94	95
Hue	180	64	63	110	114	167	166
Saturation	65	60	62	55	59	75	60

**Table IIa.**

Comparative measurements of the different colorimetric parameters for mangiferin (Red, Green, Blue, Hue and Saturation) for each species of the genus *Aquilaria* analysed (collector name).

(Collector name)	<i>Aquilaria crassna</i> (Pierre)	<i>Aquilaria crassna</i> (Bejaud)	<i>Aquilaria baillonii</i> (Chevalier)	<i>Aquilaria crassna</i> (Le Cong)	<i>Aquilaria rugosa</i> (Le Cong)	<i>Aquilaria crassna</i> (Chevalier)
Red	8	24	42	45	54	58
Green	100	100	100	100	100	100
Blue	95	98	100	98	100	100
Hue	177	178	180	178	180	180
Saturation	92	76	58	55	46	40

**Table IIb.**

Comparative measurements of the different colorimetric parameters for mangiferin (Red, Green, Blue, Hue and Saturation) for each species of the genus *Aquilaria* analysed (collector name).

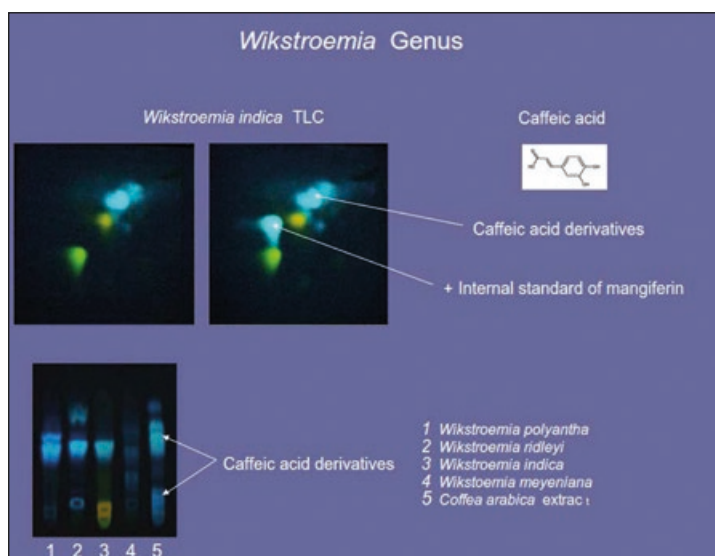
(Collector name)	<i>Aquilaria sinensis</i> (Lei)	<i>Aquilaria sinensis</i> (Bodinier)	<i>Aquilaria sinensis</i> (Chan)	<i>Aquilaria malacensis</i> (Griffith)
Red	16	27	29	48
Green	100	100	100	100
Blue	85	95	88	100
Hue	169	176	170	180
Saturation	84	73	71	52

As regards the quality analysis applied to the sample of *Aquilaria* spp. leaf fragments “Agarwood Tea”, we analysed sachets from two different places. Firstly some sachets from Laos (small-scale production, No. 1) and some sachets from Vietnam (industrial production, No. 2).

We analysed type A and type B polyphenols (see methodology) from the sachet contents. Interpretation of the chromatograms for the two sachets (Nos. 1 and 2) showed us that the type A polyphenol profiles found in *Aquilaria* were closely related (especially for mangiferin) (figure 6). However, the interpretation of the chromatographic profile of type B polyphenols (tannins) indicates that only Agarwood Tea N° 2 corresponds overall to the profile of tannins found in *Aquilaria*.

## Discussion

We chose a rapid, simple and inexpensive chromatographic analysis method to analyse our leaf extracts: two-dimensional cellulose thin-layer chromatography. This method offers greater accuracy than conventional one-dimensional chromatography. In fact, the positioning of the spots on the chromatogram is stricter and more reproducible, and the separation between spots is more efficient as each molecule migrates in two perpendicular directions, with a different



**Figure 5.**

Checking for the absence of mangiferin in *Wikstroemia indica* by the mangiferin internal standard technique (2D TLC) and for the presence of caffeic acid derivatives in the four species of *Wikstroemia* studied by one-dimensional TLC (with *Coffea arabica* extract as reference to caffeic acid derivatives).

elution solvent each time, hence with different separating power. Using thin layers of cellulose also prevents the oxidation of polyphenols, thus leading to high detection sensitivity, unlike the usual thin layer of silica gel.

Lastly, adding colorimetric analysis (measurement of the quantities of colour for the three channels: Red, Green and Blue) of the spots corresponding to the different molecules was very valuable, as it provided further confirmation of the identification of the analysed molecules. Among the values obtained, it was the “Hue” parameter that was most stable and which differentiated between mangiferin and the other molecules (table I), among other things.

In addition, we found that the colorimetric values obtained for mangiferin (which was found in each of the species of *Aquilaria* and in some of the species of *Gyrinops* analysed) were highly comparable for Blue and Green (tables IIa, IIb and III). However, the Red values varied (from 8 to 58, tables IIa and IIb) depending on some parameters that we were unable to control, such as concentrations, physiological stages, environmental conditions, etc. The Red values were also found to be inversely proportional to the Saturation values, which was normal according to the scale of RGB values obtained with the software used (GIMP colour editor, Google).

This type of identification by colorimetric analysis of molecules is very simple, inexpensive and, to our knowledge, has never been used before directly on thin-layer chromatograms. This method is perfect for analysing plant polyphenols, due to its sensitivity and specificity arising from the use of an identification reagent (Neu’s reagent), sensitive to within a nanogram of detected substance (Andary *et al.*, 1978), inducing different fluorescence colorations for most of the chemical groups studied.

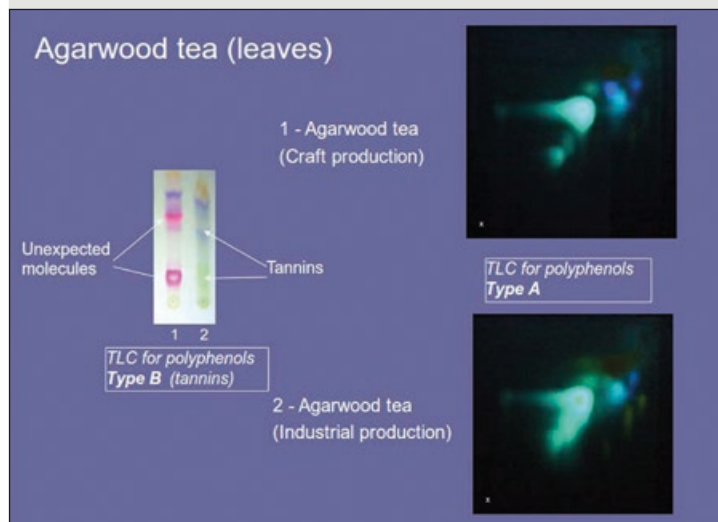
It should also be noted that application of this type of chromatography has proved very useful in the field, thanks to the development of a micro-chromatography kit using thin layers, enabling the simplified, but immediate analysis of leaves collected on site (Andary *et al.*, 1989).

Initially, we set out to check whether we could detect polyphenol molecules in both fresh and dried leaves, bearing in mind that herbarium samples may have been stored for numerous years and may also have undergone chemical decontamination. The result of this analysis showed that the polyphenol molecules of the leaves were well conserved and stood the test of time (140 years in our case), in their phytochemical environment *in planta* (figure 3).

**Table III.**

Comparative measurements of the different colorimetric parameters for mangiferin (R,G,B, Hue and Saturation) for each species of *Gyrinops* analysed.

Collector	<i>Gyrinops walla</i> Ridsdale	<i>Gyrinops ledermannii</i> Singadan	<i>Gyrinops caudata</i> Hou
Red	0	11	51
Green	27	100	100
Blue	31	100	100
Hue	186	180	180
Saturation	100	89	49



**Figure 6.**

Two-dimensional chromatography (type A polyphenols) and one-dimensional chromatography (type B polyphenols) of extracts 1 and 2 of *Aquilaria* Tea (leaves). For the type A polyphenols, the chromatograms are similar for the two extracts: small-scale production (1) and industrial production (2). For the type B polyphenols, the chromatogram shows a large difference between extracts 1 and 2. Only extract 2 displays the usual chromatographic pattern for tannins in *Aquilaria*. Extract 1 reveals unexpected molecules (shown in red) in this profile.

Chromatographic analysis of the different species showed us that the genus *Aquilaria* had a large number of type A polyphenols compared to the other three genera studied (*Gyrinops*, *Gonystylus* and *Wikstroemia*). Of these different polyphenols, mangiferin was the majority polyphenol in the six *Aquilaria* species analysed. Given the constant presence and high concentration of this molecule, we were able to consider it a chemotaxonomic marker of the genus *Aquilaria*.

It also turned out that this difference, the presence/absence of mangiferin, tallied with the doubt expressed by Poilane surrounding the identification of herbarium sample No. 30000. It should be noted that it is often very difficult to delimit species within the Thymelaeaceae. For instance, as indicated by Hou (1960), an important botanical feature for recognizing species belonging to a given genus may be missing in one or two species, thereby making identification of that genus less reliable. One example is the stamens that are generally free in the genus *Aquilaria*, and can be partially attached to the floral tube in some specimens, as in *A. cumingiana* (Decne.) Ridle.

Distinguishing between the different species of *Aquilaria* remains tricky, even in the presence of flowers and fruits, which are essential elements in recognizing plant species. In this case, the search for specific or generic chemical markers may be of great help in distinguishing between close genera, or better still, between species. Thus, *Gyrinopsis* Decne. has been considered as a section of the genus *Aquilaria* and even subsequently a synonymy of the genus *Aquilaria* (Hallier, 1922).



In the more recent classification of the Thymelaeaceae by Hou (1960), for the 12 species described for *Aquilaria* in Malaysia, seven species would appear to belong to the genus *Gyrinopsis*. The same author also demonstrated the close proximity that exists between the genus *Aquilaria* and the genus *Gyrinops*, though without merging these two genera. It turned out that the doubt expressed by Poilane regarding the identification of *Aquilaria baillonii* was backed up by our chemical analysis of the three species of the genus *Gyrinops* in which the mangiferin concentration was highly variable. In fact, in *G. caudata* and *G. ledermannii*, mangiferin was present (in different concentrations), whilst in the species *G. walla* there were only traces (figure 4). However, the role of mangiferin as a marker of the genus *Aquilaria* remains valid as this molecule not only marks species of this genus in a definite way, but also indicates the great phylogenetic proximity between *Aquilaria* and *Gyrinops* as well shown by Eurling and Gravendeel (2005) with phylogenetic analyses of DNA.

It is also interesting to note that for the four species of *Wikstroemia*, the constant existence of caffeic acid derivatives would be worth studying in a search for chemotaxonomic markers (figure 5). We have had the opportunity to demonstrate the value of these derivatives as genus or species markers in various plants (Andary *et al.*, 1988a, b; Andary *et al.*, 1992).

As regards the analysis of *Aquilaria* spp. leaf fragments marketed as a herbal tea (called Agarwood Tea), sample No. 1 (small-scale production) had probably been subjected to tannin degradation during the preparation process, which consisted (personal communication) of initial boiling for 10-15 min, followed by drying. The leaves were then finely broken up and boiled for another 5 min, then dried again. In addition, on the chromatogram (figure 6) for the one-dimensional thin-layer analysis of type B polyphenols (tannins), we found the appearance of unexpected molecules (reddish spots, sample No. 1) when compared to sample N° 2, which corresponded to a normal tannin profile found for the species of the genus *Aquilaria* studied. We were unable to interpret this difference any further as we did not know the preparation process for Agarwood Tea N° 2. However, it is sample No. 2 that should be used in the search for quality for commercial use.

## Conclusion

This work showed the possibility of using some very old herbarium samples, since the analysis of a herbarium sample from Paris (P) herbarium, which was 140 years old, revealed good conservation of its phenolic profile when compared to a sample of the same species collected and dried just a few months earlier, despite the leaves having undergone chemical decontamination. Comparative analysis of these phenolic molecules can therefore be very useful when seeking chemotaxonomic markers.

A chemical analysis of leaves from species belonging to four genera of the Thymelaeaceae revealed a large number of polyphenols (mangiferin, coumarins, derivatives of luteolin and apigenin and caffeic acid, etc.) making up a specific phenolic profile for each of these genera. It is interesting to

note that mangiferin, through its high concentration and constant presence in all the species of the genus *Aquilaria* analysed, constitutes a chemical marker for this genus, which is not the case for the genera *Gonystylus* and *Wikstroemia*, from which it was totally absent. However, the presence of mangiferin at very different concentrations in the three species of the genus *Gyrinops* studied confirmed the strong phylogenetic relation that is known to exist between *Aquilaria* and *Gyrinops*.

In addition, these results remove the uncertainty surrounding herbarium sample No. 30000 of the Poilane collection, deposited at MNHN herbarium (P) and labelled "*Aquilaria baillonii* or *Gyrinopsis baillonii*". In fact, as this sample does not contain any mangiferin, it cannot belong to the genus *Aquilaria*, but may belong to the genus *Gyrinops*.

The chromatography method we used in this analysis has the merits of being simple, practical and inexpensive. In addition, the colorimetric analysis of spots on the chromatograms belonging to various molecules gave an original result that improves the accuracy of the physicochemical authentication of chromatographed molecules contained in a plant extract.

This chromatographic analysis is not only an aid for the systematic of these species, but may also help in the *in situ* conservation of their genetic diversity. It may also be a traceability tool with a view to seeking species authenticity and quality in the various fields of use, as we were able to show by analysing dried leaf fragments sold in Asia as herbal tea (Agarwood Tea). This herbal tea is appreciated (cf. Aimi Zafirah *et al.*, 2017) for its sedating, anti-inflammatory, anti-microbial and anti-diabetic qualities.

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