

EVALUATING THE STORABILITY OF CARYOPSES OF *CENCHRUS GLAUCUS* CV. CO1

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ABSTRACT

Germination of caryopses of *Cenchrus glaucus* was assessed under storage and germination improvement was noticed up to two months of storage due to the dormant nature of the species, which decreased gradually in the later periods. In storage, the caryopses stored in cloth bag absorbed more moisture which was parallel to the moisture fluctuations of the surrounding storage environment which also affects the storage behaviour of seeds. Halogen mixture treated seeds stored better in polythene bags preserved germination and vigour of seeds.

Key words: Caryopses, *Cenchrus*, Storage.

INTRODUCTION

Seed dormancy is the natural phenomenon for the survival of grasses in their undisturbed ecosystem. The seed dormancy in forage grasses prevents successful establishment of a new pastures. *Cenchrus* is an apomictic grass species, established from both seed and rooted slips.

In *Cenchrus glaucus* also seed germination itself is a problem and the fresh fluffs hinder the germination if sown as such. Lahiri and Kharabanda (1963) reported that fresh fluffs of both *Cenchrus* and *Lasirus* possessed inhibitors in the husk which prevents germination of fresh fluffs. More over handling with the fluffs during sowing or transport poses problem due its bulkiness and empty glumes. Deglumed seeds recorded ready germination than spikelets as such in *Cenchrus* sp. ((Parihar *et al.*, 1984b; Vijendradas, 1990; Venter and Rethman, 1992). But storability of the deglumed/ caryopses is not well documented. Hence an attempt was made to assess the storability of caryopses given with halogen treatment stored under ambient conditions.

MATERIALS AND METHODS

One month old fluffs of *Cenchrus glaucus* (Blou buffel) cv. CO1 were treated with sulphuric acid (purity 95%) @ 500 ml kg⁻¹ of seeds for 15min. and the caryopses were extracted after thorough washings for 4-5times. Then caryopses were dried back to 10 per cent moisture and cleaned by blowing

the seeds in the air blower at 0.55 mg pressure for 15 min and used for the study. Seeds were graded also with BSS 24 x 24 wire mesh sieve. Both treated and untreated seeds were packed in cloth or 700 gauge polythene bag (heat sealed) and stored under ambient conditions.

Treatments

T ₁	-	Seeds without any treatment
T ₂	-	Seeds treated with halogen mixture
C ₁	-	Stored in cloth bag
C ₂	-	Stored in polythene bag

The experiment was conducted with a CRD design in factorial concept with three factors *viz.*, period of storage, containers and seed treatment with four replications. The seeds were tested initially and subsequently at bimonthly intervals designated as P₀, P₂, P₄, P₆, P₈, P₁₀ and P₁₂ to assess the extent of loss in vigour and viability of seeds. The seeds were evaluated for moisture content (%), germination (%), root and shoot length (cm), vigour index values (Abdul-Baki and Anderson, 1973) and electrical conductivity of seed leachate (Presley, 1958). Moisture estimation was done by hot air oven method at 105 ± 2°C for 16h. Germination test was conducted on top of the paper media at 25°C and 90 ± 5% RH with the germination period of 14 days (Anonymous, 1990). Data were analysed following Snedecor and Cochran (1967).

RESULTS AND DISCUSSION

Germination and vigour of caryopses increased initially due to the dormancy nature of the species but it decreased gradually in further storage where the effect of treatments and containers has its own effect (Fig. 1 & 2).

Seed moisture has greater influence on germination of caryopses under storage. The moisture content of caryopses stored in cloth bag showed gradual increase during monsoon months (August-December), which again reduced during summer. Changes in seed moisture during storage were parallel to the changes in relative humidity of air in the ambient condition. The relative humidity during monsoon months was high compared to summer months. The effect was low in poly bags. Moisture content increased from 10.05 per cent (P_0) to 10.8 per cent at P_6 and P_8 which decreased to 10.24 per cent at P_{12} . (Fig.1.)

Germination of caryopses was increased by 25 per cent reaching 79 per cent during first two months of storage and then decreased gradually in the later periods to the minimum of ten per cent at the end of the storage period (Table 1). The initial increase in the germination per cent during first two months of storage was due to the release of dormancy which lies in the caryopses rather than the spiklet structures. Butler (1985) also opined that dormancy mechanisms of *C. ciliaris* lies within the caryopses rather than in the associated structures of fluff. Pandeya and Pathak (1978) recorded the presence of phenolics particularly anthocyanins in fresh glumes and other phenolics in fresh seeds of *Cenchrus ciliaris*. Das (1990) also recorded 23.7, 61.2 and 69.0 percentages of germination with deglumed seeds of *C. glaucus* after 1,2 and 10 months of storage respectively.

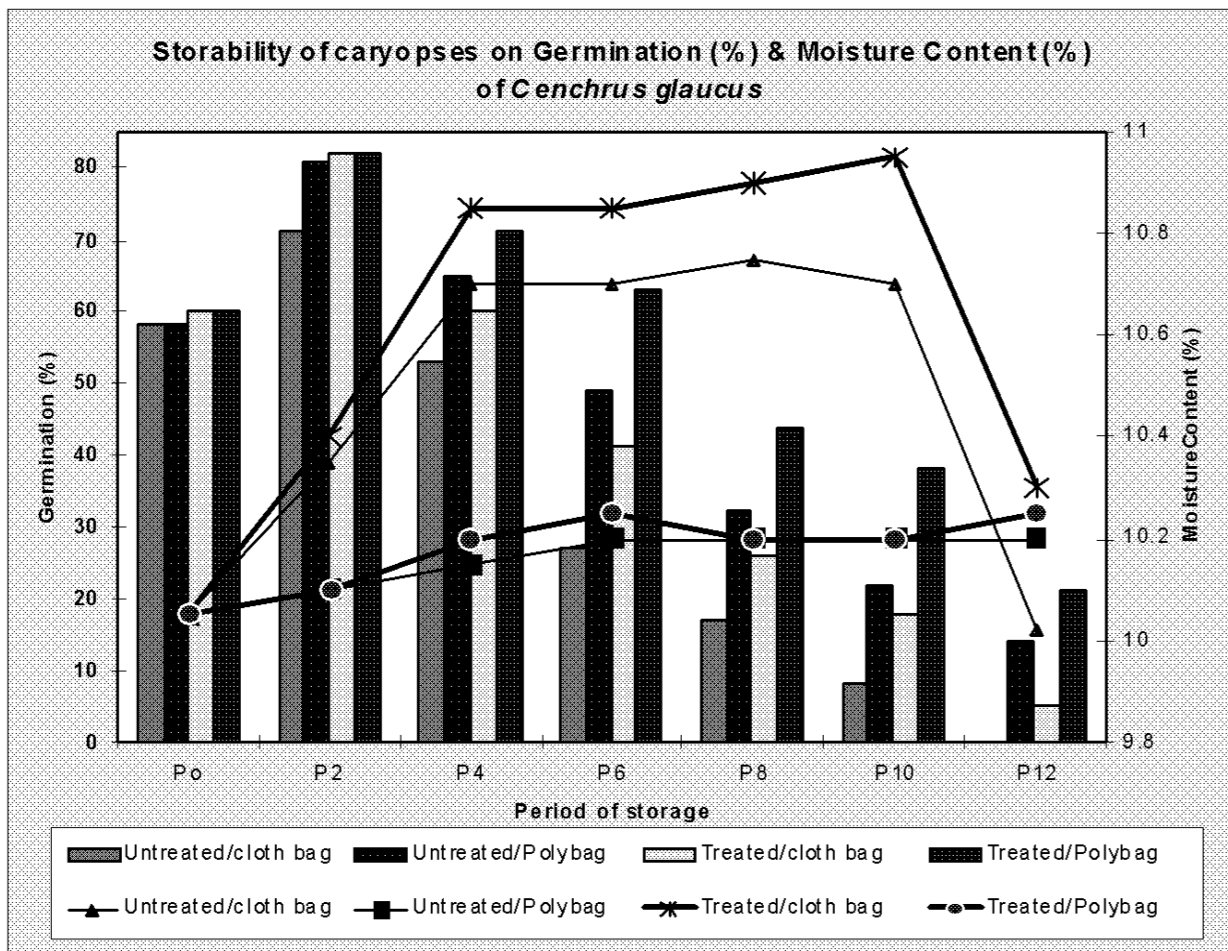


FIG.1 Effect of storage treatments on Germination and moisture content of caryopses of *Cenchrus glaucus*.

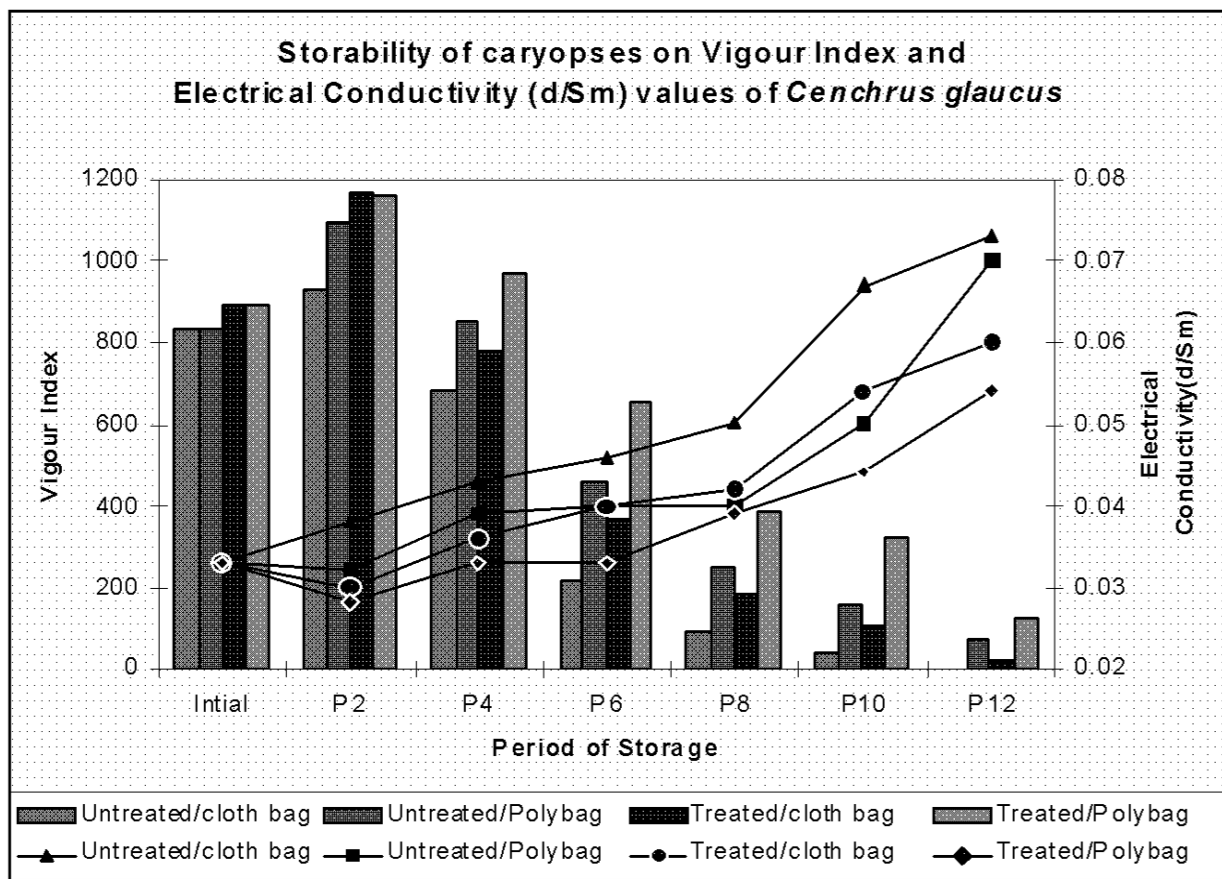


FIG 2. Effect of storage treatments on vigour status of seeds of *Cenchrus*.

The loss in germination was 87.3 per cent compared to the initial values. The tiny nature of true seeds associated with little protection and possibly because of the invisible injury caused during acid defluffing would have made them vulnerable for faster deterioration in storage. Similar results with the loss of viability of true seeds were noticed by several authors in grass seeds (Parihar *et al.* 1984a; Narayanasamy, 1994; White *et al.* 1999). The loss of viability was more (100 per cent) with untreated seeds stored in cloth bags than halogen treated seeds in polythene bag containers. (Fig.1) The possible effect of treatment was consistently associated with reduced production of volatile aldehydes, which are presumably the product of lipid peroxidation (Wilson and McDonald, 1986). The halogens act at the molecular level and stabilizes the C=C double bond in the unsaturated fatty acids of the lipo protein moiety and making less susceptible to lipid peroxidation and free radical reaction (Basu, 1994).

Seedling length both root and shoot length and vigour index of the stored seeds showed steady decline in vigour from the beginning of storage indicating the vigour loss of the true seeds which precedes the viability loss. The decrease was more in cloth bags compared to poly bags and likewise with untreated seeds.

The seeds stored in 700 gauge polythene bags sustained minimum deterioration in seedling length, vigour and membrane integrity as reflected through electrical conductivity (Fig.2) when compared to the seeds stored in the cloth bag. The 700 gauge polythene bag formed an effective barrier to moisture, vapour and gas and it would have also create oxygen tension and increase CO₂ accumulation inside the bag which would slowed down the respiration rate (Justice and Bass, 1978). Peel and Prondroff (1970) recommended sealed storage for *Panicum maximum* than open storage to maintain vigour and viability of seeds.

TABLE 1. Effect of treatments, containers and periods of storage on true seed germination (%) in *C. glaucus* cv. CO 1

Period of storage / treatments	T1			T2			Grand Mean
	C ₁	C ₂	M	C ₁	C ₂	M	
P ₀	58 (49.61)	58 (49.61)	58 (49.61)	60 (50.77)	60 (50.77)	60 (50.77)	59 (50.19)
P ₂	71 (57.45)	81 (64.18)	76 (60.80)	82 (64.93)	82 (64.93)	82 (64.93)	79 (62.87)
P ₄	53 (46.72)	65 (53.73)	59 (50.23)	60 (50.77)	71 (57.42)	66 (54.98v)	62 (52.16)
P ₆	27 (31.30)	49 (44.43)	38 (37.87)	41 (39.82)	63 (52.54)	52 (46.18)	45 (42.02)
P ₈	17 (24.27)	32 (34.44)	25 (29.36)	26 (3.64)	44 (41.56)	35 (36.10)	30 (32.73)
P ₁₀	8 (16.43)	22 (27.95)	15 (22.19)	18 (25.07)	38 (38.05)	28 (31.56)	22 (26.88)
P ₁₂	0 (0.383)	14 (21.92)	7 (11.15)	5 (12.86)	21 (27.27)	13 (20.66)	10 (15.61)
M	33 (32.31)	46 (42.32)	40 (37.32)	42 (39.27)	54 (47.51)	48 (43.39)	50 (44.91)
SE d	0.781	T 0.418	C 0.418	PT 1.105	TC 0.591	PC 1.051	PTC 1.563
CD (P ≤ 0.05)	1.601	0.856	0.856	2.264	1.210	2.264	3.201

Figures in parentheses are arc sine values. Analysis was done with the transformed values.

TABLE 2. Effect of treatments, containers and periods of storage on root & shoot length (cm) of seedlings of caryopses in *C. glaucus* cv. CO1

Period of storage / treatments	Root Length (Cm)												Grand Mean	
	T1			T2			M	C ₁	C ₂	M	C ₁	C ₂		
	C ₁	C ₂	M	C ₁	C ₂	C ₁								C ₂
P ₀	8.11	8.11	8.11	8.27	8.27	8.27	8.27	8.19	8.19	8.19	8.19	8.19	8.19	8.19
P ₂	7.62	7.68	7.65	8.18	8.07	8.13	8.07	7.90	7.88	7.89	7.90	7.88	7.89	7.89
P ₄	7.43	7.58	7.51	7.64	7.93	7.78	7.93	7.54	7.75	7.64	7.54	7.75	7.64	7.64
P ₆	3.45	4.65	4.05	4.26	5.22	4.74	5.22	3.86	4.94	4.40	3.86	4.94	4.40	4.40
P ₈	2.21	3.34	2.78	3.10	4.05	3.58	4.05	2.66	3.70	3.18	2.66	3.70	3.18	3.18
P ₁₀	1.93	3.16	2.54	2.88	3.83	3.36	3.83	2.40	3.50	2.95	2.40	3.50	2.95	2.95
P ₁₂	0.00	2.63	1.34	1.22	2.95	2.08	2.95	0.61	2.81	1.71	0.61	2.81	1.71	1.71
M	4.39	5.31	4.85	5.08	5.76	5.42	5.76	4.75	5.54	5.42	4.75	5.54	5.42	5.42
SE d	0.163	0.087	0.087	0.231	0.123	0.023	0.123	PTC	PTC	0.326	PTC	PTC	PTC	PTC
CD (P ≤ 0.05)	0.334	0.179	0.179	NS	NS	0.472	NS	NS	NS	NS	NS	NS	NS	NS
P ₀	6.26	6.26	6.26	6.53	6.53	6.53	6.53	6.40	6.40	6.40	6.40	6.40	6.40	6.40
P ₂	5.50	5.80	5.65	6.08	6.11	6.09	6.11	5.79	5.96	5.87	5.79	5.96	5.87	5.87
P ₄	5.39	5.57	5.48	5.33	5.76	5.54	5.76	5.36	5.66	5.51	5.36	5.66	5.51	5.51
P ₆	4.49	4.73	4.61	4.72	5.20	4.96	5.20	4.60	4.97	4.78	4.60	4.97	4.78	4.78
P ₈	3.18	4.44	3.81	3.80	4.78	4.29	4.78	3.49	4.61	4.05	3.49	4.61	4.05	4.05
P ₁₀	2.45	3.95	3.20	3.06	4.61	3.83	4.61	2.76	4.28	3.52	2.76	4.28	3.52	3.52
P ₁₂	0.00	2.65	1.32	2.34	3.05	2.70	3.05	1.17	2.85	2.01	1.17	2.85	2.01	2.01
M	3.89	4.77	4.33	4.55	5.15	4.85	5.15	4.22	4.96	4.85	4.22	4.96	4.85	4.85
SE d	0.154	0.082	0.082	0.217	0.217	0.116	0.217	PTC	PTC	0.307	PTC	PTC	PTC	PTC
CD (P ≤ 0.05)	0.315	0.168	0.168	0.445	0.445	NS	0.445	NS	NS	NS	NS	NS	NS	NS

Thus it was evident from the study that the polybags after treatment with halogen mixture @3g caryopses of *Cenchrus glaucus* could be stored in kg⁻¹ of seed for better storage.

REFERENCES

- Anonymous, (1990). Amendments to the International Rules for Seed Testing (1985) *ISTA*, Switzerland.
- Abdul-Baki, A.A. and Anderson, J.D. (1973) Vigour determination in soybean seed by multiple criteria. *Crop Sci.*, **13**: 630-632.
- Basu, R.N.(1994) An appraisal of research on wet and dry physiological seed treatments and their applicability with special reference to tropical and subtropical countries. *Seed Sci. & Technol.*, **22**: 107-126.
- Butler, J.E. (1985). Germination of Buffel grass. *Seed Sci. & Technol.*, **13** : 583-591.
- Das, L. D. V. (1990) Dormancy studies in *Cenchrus glaucus*. *Madras Agric. J.*, **77** : 254-256
- Justice, O.L. and Bass, L.N. (1978) Principles and Practices of Seed Storage. Agriculture Handbook No. 506. USDA, Washington. p.289.
- Lahiri, A.N. and Kharabanda, B.C. (1963) Germination studies on arid zone plants. II Germination inhibitors in the spikelet glumes of *Lasiurus indicus*, *Cenchrus ciliaris* and *Cenchrus setigerus*. *Ann. Arid Zone*, **1** : 114-126.
- Narayanasamy, S. (1994) Seed technological studies in deenanath grass (*Pennisetum pedicellatum* Trin.) M.Sc.(Ag.) Thesis, Tamil Nadu Agricultural University, Coimbatore.
- Pandeya, S.C. and Pathak, S.J. (1978). Germination behaviour of some ecotypes of anjan grass under dry storage and physical stress. *Proc. 1st Int. Rangeland Congress, USA*. pp.376-383.
- Parihar, S.S., Kanodia, K.C. and Rai, P.(1984a) Effect of age and removal of glumes on germination of *Cenchrus ciliaris*. *Indian J. Ecol.*, **11** : 313-316.
- Parihar, S.S., Kanodia, K.C. and Rai, P. (1984b) Seed germination studies with *Cenchrus setigerus*. 1. Effect of age and removal of glumes on germination. *Indian J. Range Mgmt.*, **5** : 5-9.
- Peel, A.C. and Prondroff, E.T. (1970) Storage of hamil grass (*Panicum maximum*) seeds. *Proc. Int. Seed Test. Assoc.*, **36** : 175-5.
- Presley, J.T. (1958). Relation of protoplast permeability to cotton seed viability and predisposition to seedling disease. *Pl. Dis. Reprt.*, **42** : 852.
- Snedecor, G.W. and Cochran, W.G. (1967) Statistical Methods. IOWA State University Press.
- Venter, P.S. and. Rethman, N.F.G. (1992) Germination of fresh seeds of thirty *Cenchrus ciliaris* ecotypes as influenced by seed treatment. *J. Grassland Soc. S. Africa*, **9**(4): 181-182
- Vijendradas, L.D. (1990). Dormancy studies in *Cenchrus glaucus*. *Madras agric. J.*, **77**(5-6): 254-256.
- White, N.D.G. Hulasave, R.B. and Jayas, D.S.(1999) Effect of storage conditions on quality loss of hullless and hulled oats and barley. *Can. J. Pl. Sci.*, **79**(4): 475-482.
- Wilson, D.O. and McDonald, M.B.(1986) A convenient volatile aldehyde assay for measuring seed vigour. *Seed Sci. & Technol.*, **14**: 259-268.