

International bottom trawl survey in the Mediterranean (Medits)

Instruction manual

Version 5



April 2007

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PREAMBLE

The first version of this manual was prepared in 1994 within a co-ordination between several research Institutes from the four Mediterranean member States of the European Union. The target was to conduct a common bottom trawl survey in the Mediterranean in which all participants use the same gear and the same methodology. The manual was revised in 1995 after the 1994 survey to take methodological improvements acquired during the first survey into account. During the course of the project, several improvements were made. A new version of the manual has been established when necessary to allow for improvements follow-up. In any case, the Medits co-ordination committee has taken care that the amendments do not disrupt the consistency of the series. The third version of this manual was edited in 1999. The fourth version referred to the surveys carried out since 2000.

This fifth version is an updated of the fourth version, including the improvements adopted by the Medits group since 2005. They have been fully applied for the Medits surveys since 2005. Furthermore, the manual includes all the areas covered by the Medits surveys up to 2007.

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Introduction

This document is the fifth version of a manual elaborated in the frame of an international project to harmonise the samplings by bottom trawling of the demersal resources in the Mediterranean Sea (MEDITS program). It is the reference document for the surveys to be conducted from 2007 onwards by the research institutes contributing in the Medits surveys on the continental shelves and slopes in the Mediterranean (Fig. 1).

The manual describes the sampling gear characteristics, the sampling methodology and the treatment of the samples. Finally it gives the specifications of the data files for data storage and exchange.

[1] Specifications of the sampling gear

1.1 The trawl

The sampling gear is a bottom trawl made of four panels. The drawing of this trawl (IFREMER reference GOC 73) is given in figure 2. This gear was calculated to be operated by a vessel with a towing power of at least 368 kW (500 ch) and 4.5 tons of bollard pull.

On the plan in figure 2 the mesh sizes are indicated in bar length. The mesh numbers in height correspond to well finished and joined netting sections; the joining mesh should then be subtracted when cutting. The numbers of mesh in width do not include the side seams and those should then be added when cutting.

The floats of the headline, which are 40, should resist to an immersion of 1300 m. Their diameter should be around 20 cm, their individual buoyancy of 2.7 kgf ($\pm 5\%$), the total buoyancy of the 40 floats being around 108 kgf ($\pm 5\%$). The 40 floats should be distributed along the headline as follow (Fig. 3 and 7): from the end of each wing, one float every 1.50 m, 5 times; then one pair of floats every 1.50 m on the whole remaining length; in the headline bosom a small adjustment of the spacing is necessary. With this number of floats the vertical opening of the trawl should reach 2.4 to 2.6 m depending on the horizontal opening.

The weighting chain of 120 kg (3×40) should be secure to the foot rope every 17 cm (with a hanging height of at most 8 cm). A supplementary chain of 15 kg (around 6.50 m and a diameter of 10 mm) should in addition been secured symmetrically on both parts of the belly bosom in the same way as the first one (garland of 17 cm in length).

1.2 The rigging

The general drawing of the rigging is given in figure 3. Various details of mounting and connection are shown in figure 4. The upper legs length is 30 m; the lower legs length is 29 m, plus the adjustment chain of 1m.

To maintain the geometry of the trawl as constant as possible two bridles length are defined according to the depth. They are given in the following table:

Depth (in meters)	10 - 200	200 - 800
Bridles length (in meters)	100	150

Accordingly with some experiment made on board the RV/L'Europe in June 2000, it is recommended to increase the bridle length to 200 m in depths deeper than 500 m. This modification is not compulsory but it can favour a better and faster contact of the trawl with the ground.

1.3 The doors

The doors are also normalised. They are of type Morgere WH S (Fig.5). The adopted doors correspond to the size number 8. The warp is shackled in the fore hole of the bracket sheet (see arrow 1 in Fig. 5). The short parts of the external crowfoot are shackled in the most back part of the backside sheets, upper and lower (see arrow 2 in Fig. 5). The length of the back-strops (shackles not included) are as follow:

- long external back-strops: 1.60 m
- short upper and lower back-strops: 0.65 m ($\pm 10\%$).

1.4 Warp diameter and length

Taking the characteristics of the trawl and the rigging into account the warps should have a diameter of 16 mm, at least of 14 mm, at most of 20 mm. The length of warps to be shot is determined by the depth. The recommended relationship between depth and warp length is given in figure 6. Although in certain peculiar circumstances some adaptations can be made to this relationship, it is recommended to respect the depth/warp length ratio as far as possible.

For the vessels which are not equipped with a device to measure the length of shot warp, it is recommended to standardise the position of the last mark on the warp, for example at the most back warp block.

1.5 Complementary equipments

The systematic use of a device to control the trawl geometry (vertical and horizontal openings, contact with the bottom) is highly recommended. The sensors should be positioned as shown in figure 7. If it is not possible, measurements of the trawl geometry should be made at various depths on board each vessel at the beginning of the survey to establish a graph. For each haul it will be noted in the data files if the indications of trawl opening are estimated or measured *in situ*.

A security device allowing to get back the trawl by the codend can be installed. As far as possible, it is recommended to secure this lazy line as shown in figure 7 and to take care of its fixations.

[2] Sampling methodology

2.1 Vessel characteristics

The vessels used for the Medits surveys might have a horse power of at least 370 kW to be able to tow the standard sampling gear (traction at ground run: 4.5 tons). It is strongly recommended that the same vessel and crew be used as much as possible every year in each area to carry out the survey series in view of reducing variations between years due to vessel effect. The list of the vessels used since the beginning of the survey series is given in **annex I**.

2.2 Period of the survey

The period of the Medits survey is centred on June (from May to July).

2.3 Hauls localisation

The hauls are positioned following a depth stratified sampling scheme with random drawing of the positions within each stratum. The number of positions in each stratum is proportional to the surface of these strata. Except in the case of peculiar problems (damages noted in previous years, etc.), the hauls are made in the same position from year to year. The decision to make a haul in a given place should not be influenced by the presence of fish shoal eventually detected with the sounder or the sonar.

The following depths are fixed in all areas as strata limits:

- 10 - 50 m,
- 50 - 100 m,
- 100 - 200 m,
- 200 - 500 m,
- 500 - 800 m.

Furthermore the strata are limited by lines more or less perpendicular to the coast, depending on the geographical characteristics of each area. The adopted stratification schemes are shown in figure 1. It is strongly recommended to strictly maintain the same scheme between years. The strata are described in **annex II**. The target number of hauls by area is given in **annex III**.

The *Posidonia sp.* grasslands are excluded from the sampling scheme and should never be trawled.

2.4 Operating the gear

2.4.1 Sampling period in the day

The hauls must be made only during daylight. The daylight period is defined as the time between 30 minutes after sunrise and 30 minutes before sunset.

2.4.2 Haul speed and duration

The standard fishing speed is 3 knots on the ground. This recommended speed is a very important target to insure the best trawl geometry. The actual speed as well as the covered distance should be monitored and recorded.

It is highlighted that a speed lower than 2.8 knots can have a negative effect on the verticality and the stability of the doors which can lie down and get stuck in the mud. A speed greater than 3.2 knots can take the trawl off the ground at great depths.

The haul duration is fixed at 30 minutes on depths less than 200 m and at 60 minutes at depths more than 200 m.

2.4.3 Haul start and end definition

The start of the haul is defined as the moment from which the trawl geometry (vertical and horizontal) is stabilised (cf. § 3.4.5.). The end of the haul is defined as the moment of the beginning of warp hauling.

2.4.4 Haul orientation

In general the hauls are made at constant depth. The depth variations during the haul should not exceed $\pm 5\%$ relatively to the initial depth. The discrepancies to this target should be recorded. In case of important difference between the depth under the vessel and the depth at which the trawl is, the recorded depth is those at which the trawl is. As far as possible and in respect of the previous constraints the hauls should be rectilinear. If for some reasons that is not possible the turning will be as wide as possible for not to disrupt the trawl geometry. In all cases the fields "PARCOU" and "DISTANCE" of the "A" data file (see § 6.2.) should be precisely documented.

2.4.5 Managing the end of shooting operations and the start of the haul

After the complete shooting of the warps and the braking of the winches a relatively high speed (5-6 knots) will be maintained for around 1 minute to allow the trawl to well draw both in length and in width.

The speed will then be strongly reduced (even to 0) during the needed time (variable depending on each vessel and each depth: for example 2-3 minutes at 500 m) in such a way that the doors reach the ground.

Once the doors are on the ground a speed lower than the normal one (2.5-2.7 knots) should be maintained to allow the trawl to get down to the ground.

Once the net is well stabilised the speed will be increased towards the standard speed (3 knots); this moment is defined as the real start of the haul.

Except some peculiar situations in which some adaptations – as small as possible – to these management would be absolutely necessary, it is recommended to respect as precisely as possible the above described procedures.

For the vessels using a device such SCANMAR the trawl can be considered as well stabilised as soon as its vertical opening is between 2 and 3 m.

For the vessels without such a device, preliminary trials will be made before the survey. These trials will target to determine, ship by ship, the time needed to correctly operate the trawl, taking the depth and the working practice of each skipper into account.

2.4.6 Setting of the trawl on the ground

It is important that the gear stay in good contact with the ground during the whole haul. This point should be checked regularly either by acoustic device during the haul, by the observation of the chains wear or by the observation of benthic organisms in the catches after the haul.

2.4.7 Trawl geometry while fishing

The trawl is designed to have a vertical opening between 2 and 3 meters at the various depths if the above mentioned adjustments are respected.

When a device like SCANMAR is used the vertical and horizontal (between the wings) opening should be checked as often as possible, once the trawl is stabilised. The average values of these two parameters (disregarding the obviously aberrant values) will be reported in the data file for each haul.

2.4.8 Wear of the trawl

It has not been foreseen any system preventing the bosom of the trawl against rubbing on the ground. It is then recommended to change the rubbing pieces of the trawl as often as needed, particularly when they have lost their initial resistance characteristics.

[3] Treatment of the catches

3.1 Samplings

On board on the vessel the catches are split into five categories (**Annex IV**):

- Category A: Fish
- Category B: Crustaceans *Decapoda* and *Stomatopoda*
- Category C: Cephalopods
- Category D: Other commercial species
- Category E: Other non commercial animal species

For each species the total weight and number of individuals is recorded

When the catch of a given species or a fraction of a given species (e.g. juveniles) is too abundant to be measured *in extenso* it seems reasonable to make a sub-sample not exceeding 100 individuals

The common coding system adopted for the complete set of species (**Annex XV**) is a RUBIN like coding system as defined in the NCC standard¹ even if this international coding system has been no longer maintained for some years. This coding system appears to be a very practical one and it would be very easy in the future to build a correspondence table with any new coding system. In respect with the NCC recommendations and as the MEDITS coding is not strictly identical to the RUBIN one (different use, species not referenced to in the RUBBIN code) the "name" of this code has been changed and is for the purpose of the MEDITS "**FM list**".

The species identifications are made according to Fisher and *al*, 1987². For the fish species not included in this work the descriptions from Whitehead *et al*, 1984³ have been used. Furthermore, a correspondence with the most updated revisions by international bodies (e.g. Fishbase⁴ for fish) is given. This review is based on the one maintained by the Ifremer's Fishery information system⁵. Nevertheless, it is underlined that the species coding has to be kept strictly identical in the data base, even if the species name has been changed.

As no zero value is included in the tables, it is important to precise the extend of species potentially recorded from the catch. Coding for this information is given in **Annex V**.

A reference list of 38 species of fish, crustaceans and cephalopods is given in **Annex VI**. For all these species the length frequencies by sex must be reported as well as the maturity stage of the gonads (see below). Since 2006, it is recommended to include all the selacians in this group.

3.2 Biological parameters

3.2.1 Measurement units

For fish the length is ever the overall length, the tail being extended following its longer dimension. The measurement unit is the lower half centimetre.

For Crustaceans the length is measured in term of cephalo-thoracic length at the lower millimetre (**Annex VII**).

For Cephalopods the length is the mantle length at the lower half centimetre (**Annex VII**).

¹ NCC: Nordic code centre (Stockholm).

² Fisher W., M.L., Bauchaud et M. Shneider (rédict.), 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche (révision 1). Méditerranée et mer Noire (volumes I et II). Projet GCP/INT/422/EEC. FAO, Rome: 1530 p.

³ Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1984. Poissons de l'Atlantique du nord-est et de la Méditerranée (3 volumes). UNESCO, Paris.

⁴ Froese R. & D. Pauly eds, 2002. FishBase. World Wide Web electronic publication. www.fishbase.org.

⁵ Woehrling D., 2004. Le référentiel taxinomique du système d'information halieutique de l'Ifremer. Ifremer, DRV/RH/DT/04-04. 12 p.

If a given team wish, for its own works, to make complementary observations on other species or of an other nature, it is kindly invited to inform the MEDITS Group (Co-ordination and Steering Committees) to eventually allow to normalise the methodology with other research teams.

3.2.2 Sex and maturity

The sex is defined following four categories: male, female, undetermined (impossible to determine it by eye) and not determine (the individual has not been examined). The sexual maturity is defined with reference to the identification keys given in **Annex VIII** for the fish and **Annex IX** for the Crustaceans and Cephalopods. For Crustaceans the maturity stages is recorded only for the females.

The individuals of hermaphrodite species being currently changing sex are qualified by the sex of whom the gonad is the more developed

3.3 Other parameters

The bottom water temperature should be recorded at the start and the end of each haul. This information is stored in an exchange file with the format defined in **Annex XIII**. The recommended sensor is the Vemco minilog TDR -5 to 35°C which records both temperature and depth. It should be fixed on the bosom head line. It is important that the clock of the computer which receives the data from the sensor is exactly set accordingly with the UT time (GMT) to have the same times as in the "A" file. The data should be recorded in one file by haul and not in one file for several hauls. The temperatures from all the hauls (beginning and end) should be kept and reported in the file of type 4 (**D**) and they should correspond to the official time of beginning and end of the haul, assuming that the trawl begins and stops to work properly at these official times.

[4] Inter-calibration of the work at sea

To try an inter-calibration of the working methods between the various vessels two possibilities are recommended: an exchange of scientist on board the vessels and where and when it could be possible a co-ordination of trawling operation, together by the two vessels, at the border of the areas covered by these two vessels respectively.

To favour the exchange of scientists one place will be reserved on board of each vessel for the eventual boarding of a scientist from another team. In addition each co-ordination group will do their best to send scientist from their own team on board of other vessels participating in the project. It is expected that the reports of these boarding help to identify eventual differences in the working methodology.

Where and when different teams are in charge of adjacent working areas, they are invited, although it seems rather difficult and time consuming, to act in concert together to try to organise some common hauls in parallel to reach an inter-calibration between the two vessels.

[5] Data exchange formats

5.1 General information

Standard formats are defined for the storage and to facilitate exchange of the data produced by the Medits surveys. The exchange files are in an ASCII format.

5.2 Files type

Four files types are defined to store and exchange the data:

Type A: Characteristics of each haul (**Annex X**)

Type B: Catch of each haul in weight, number and number by sex (**Annex XI**)

Type C: Biological parameters for the species in the reference list (length, sex, maturity) (**Annex XII**)

Type D: Temperature data (**Annex XIII**)

Type T: List of hauls by stratum (**Annex XIV**)

The files names are defined as follow:

Position	Variable	Possible values
Character 1	Files type	A (haul characteristics) B (catch by haul) C (biological parameters) D (temperature) T (list of hauls)
Character 2-3	Country	MA, ES, FR, IT, SL, HR, AL, MO, ML, GR, CY
Character 4	Area	1 to 5 for Italy, 1 to 3 for Greece, 1 to 2 for Spain, _ (underscore) for the other countries
Character 5-8	Year	2000, 2001 , etc.
Character 9	Separator	. (point)
Character 10-12	Extension	TXT for A, B, C, D files, DAT for S and T

5.3 Files structure and information's coding

The exchange files format are described in **Annexes X to XIV**. Complementary coding tables used to fill in the data files are given in annexes referred above.

—

Figures

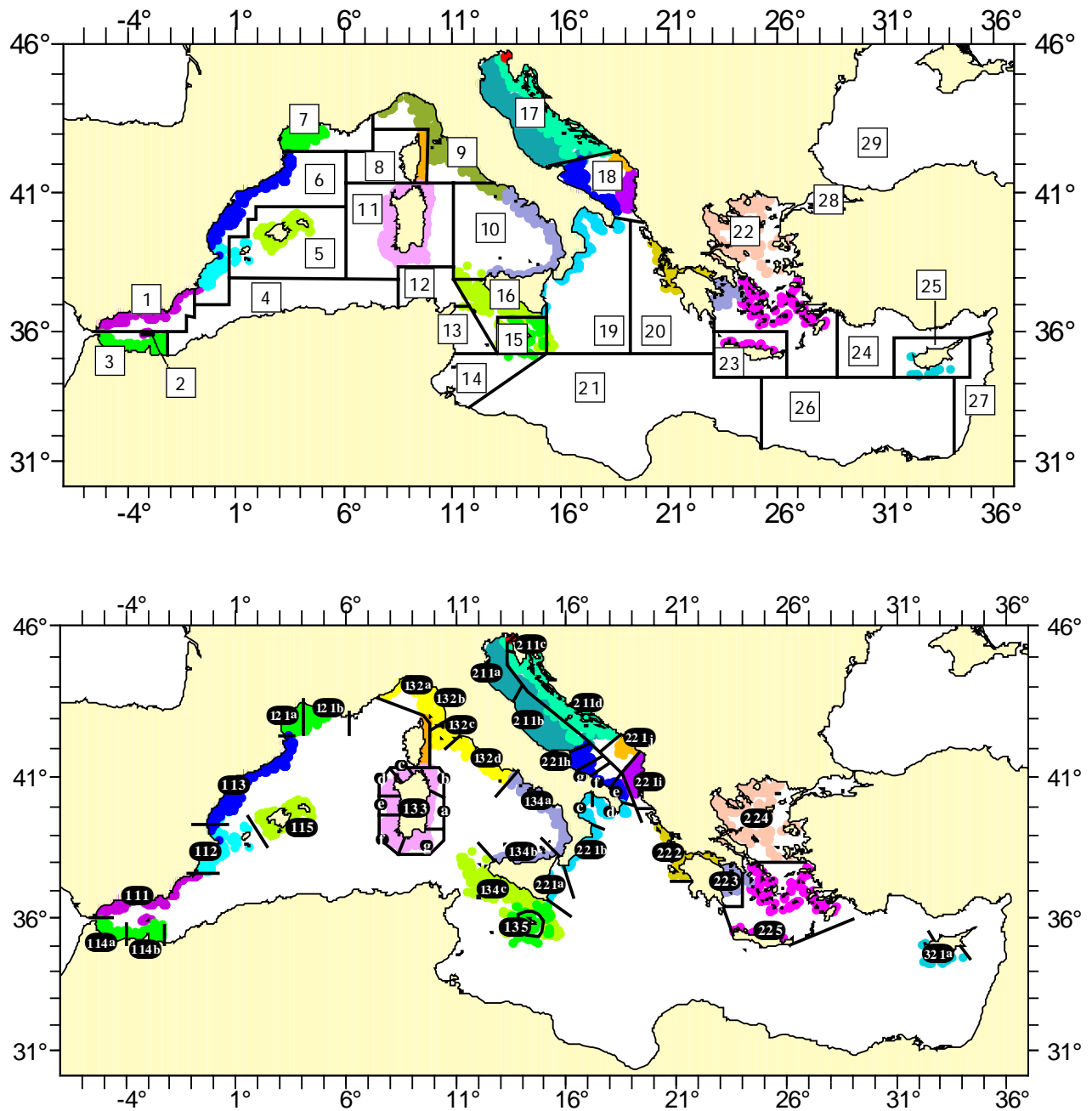


Fig. 1. General map of the area covered by the programme. Top: the GFCM GSAs, Down: the Medits strata. Coloured: areas covered by the Medits surveys.

The designations used and the presentation of cartographic data imply no line as for the juridical status of the various areas neither as for the border lines between countries.

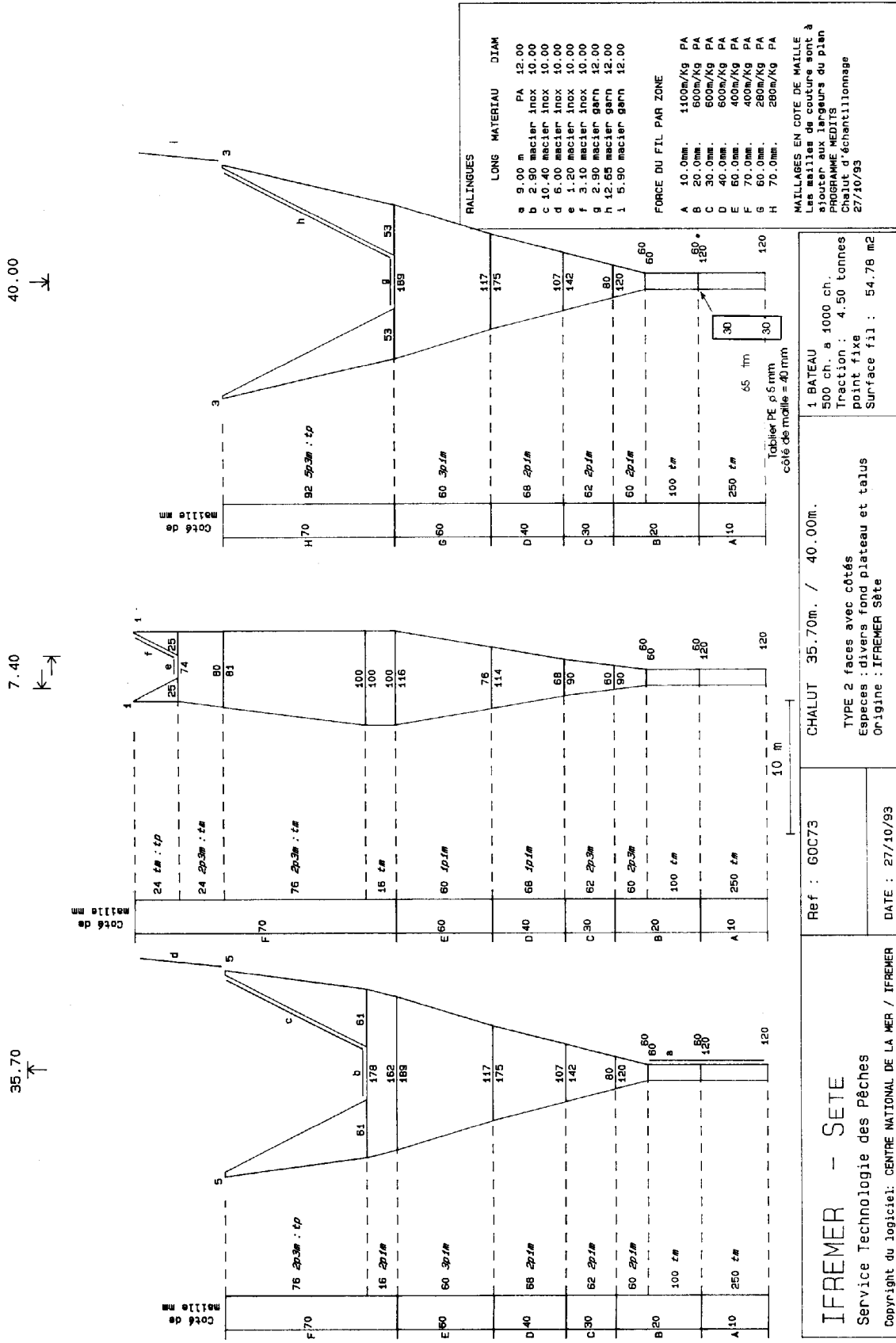


Fig. 2. Plan of the trawl GOC 73.

Schéma de gréement du chalut GOC 73

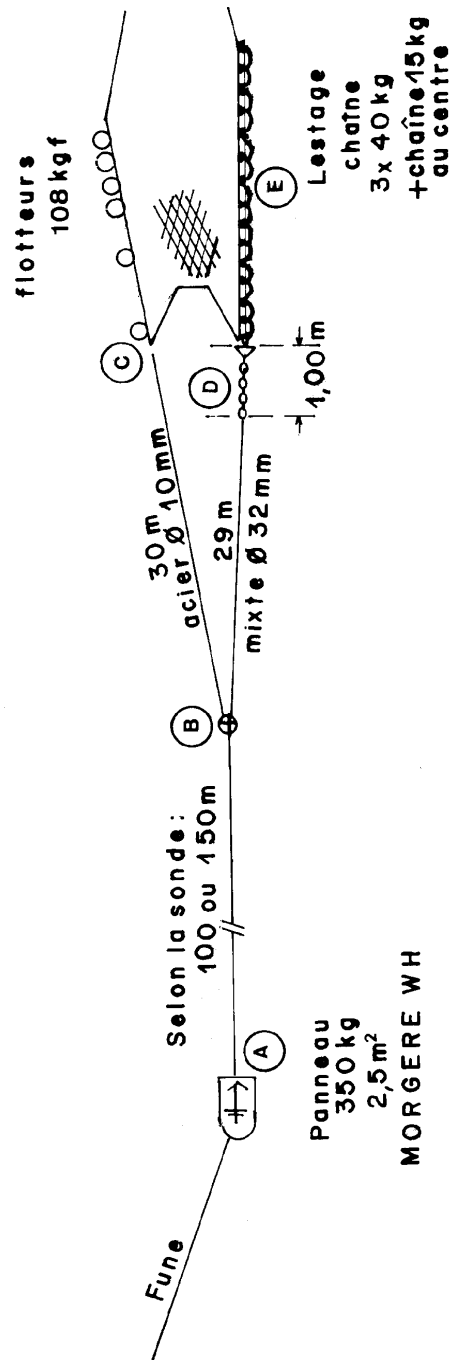


Fig. 3. Diagram of the rigging.

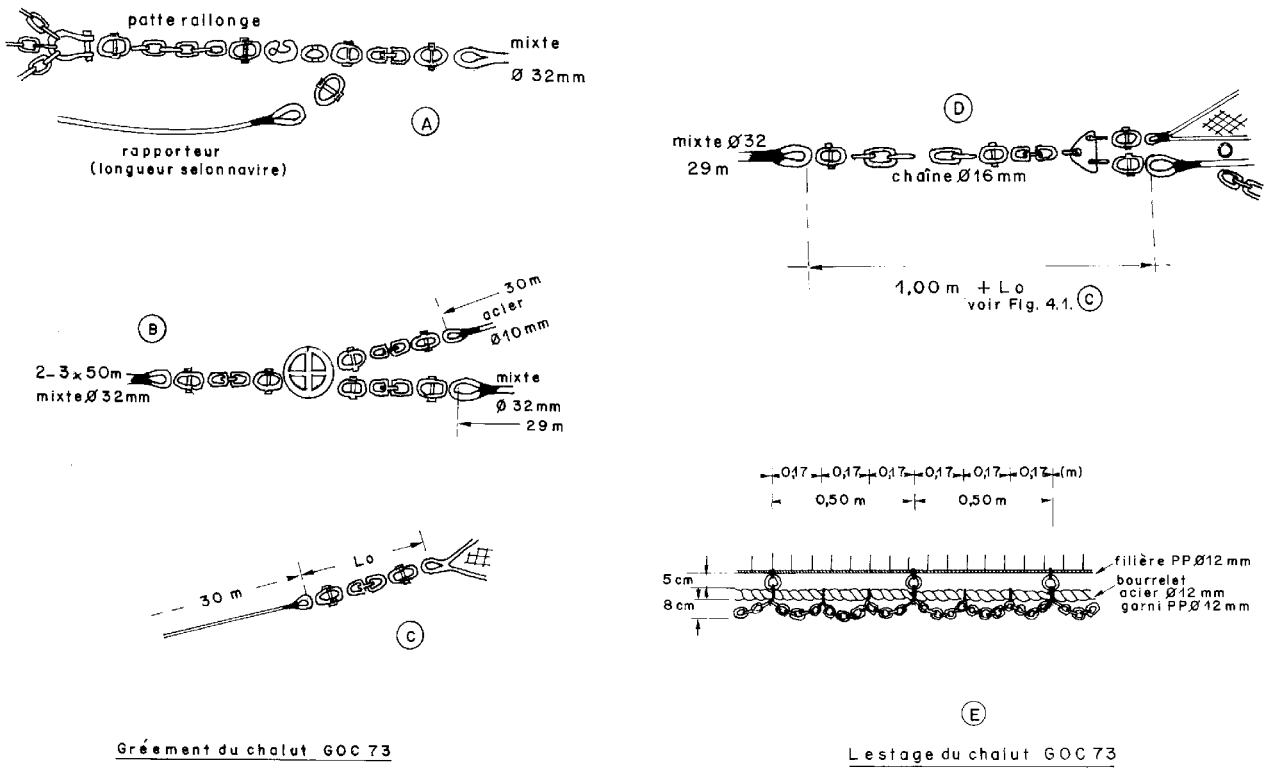
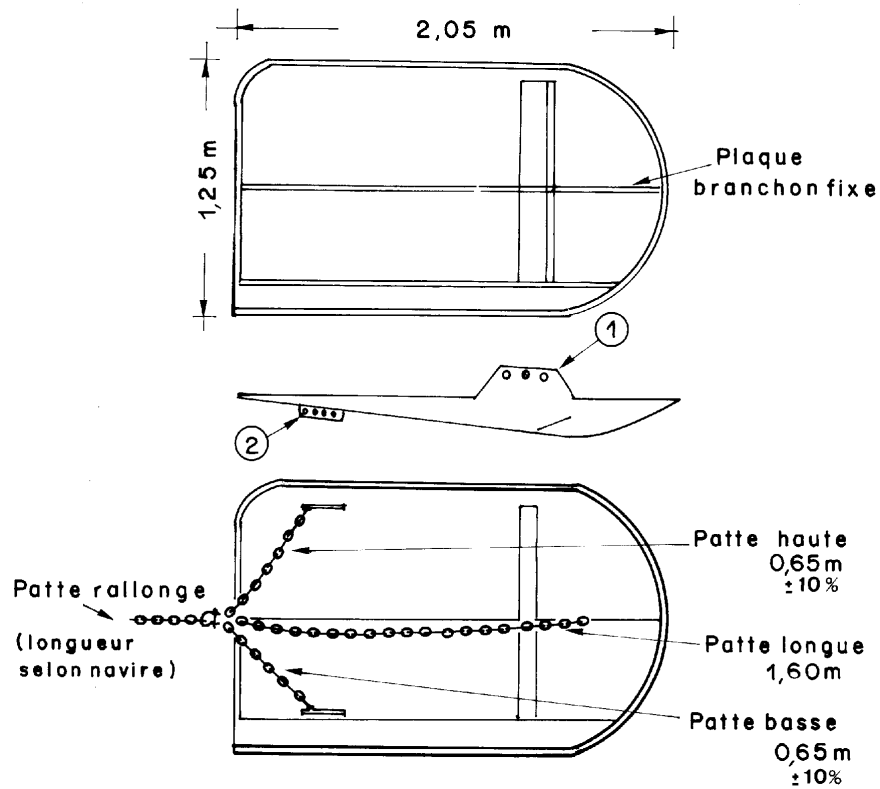


Fig. 4. Various details of the rigging.

(2,6 m² ; 350 kg)



Les longueurs des pattes sont indiquées
manilles non comprises

Fig. 5. Drawing of a door Morgère WHS (8).

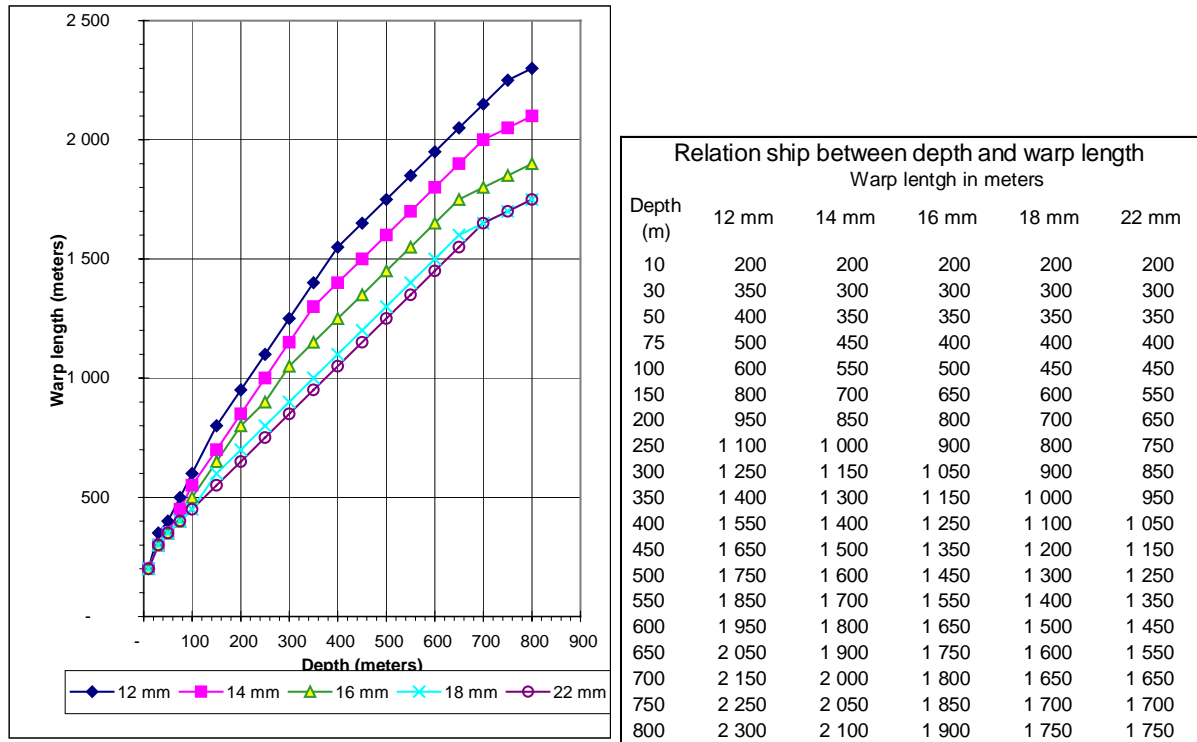
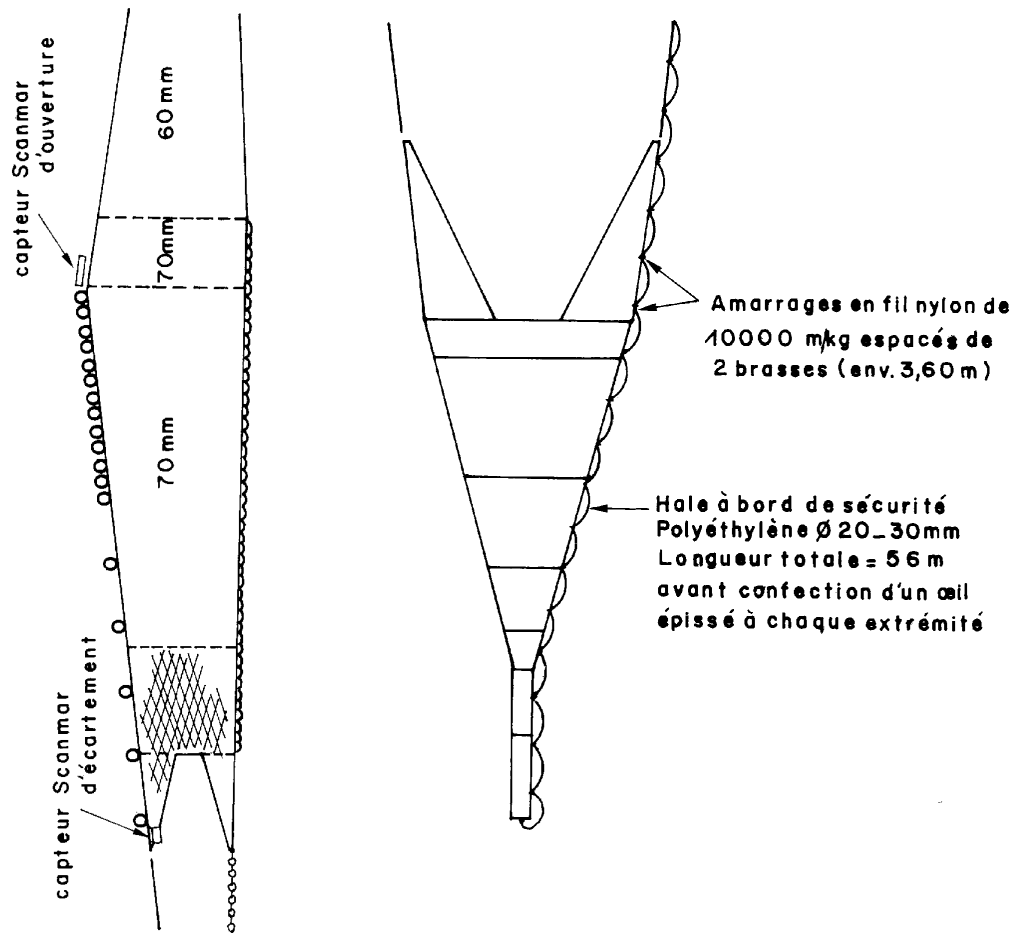


Fig. 6. Relationship between depth and warp length for the trawl GOC 73.



The security lazy line is secured first at the fore part of the upper starboard wing and second behind the codend. Between this two points, this port is secured around every two fathoms, alongside the seam between the upper face and the lateral starboard face of the trawl.

Fig. 7. Position of the geometry sensors and drawing of the lazy line.

[6] Annexes

- I. Code of countries, vessels and gear**
- II. Stratification scheme**
- III. Target number of hauls by area**
- IV. Code of recorded species, of general observations on hauls and of quadrants**
- V. Code of faunistic categories. Form to introduce new species**
- VI. List of reference species**
- VII. Standard lengths for Crustaceans and Cephalopods**
- VIII. Codes of sexual maturity for fish**
- IX. Codes of sexual maturity for Crustaceans and Cephalopods**
- X. Format of the type A files (Data on hauls)**
- XI. Format of the type B files (Catches by haul)**
- XII. Format of the type C files (Biological parameters)**
- XIII. Format of the type D files (Temperature data)**
- XIV. Format of the type T files (List of hauls by stratum)**
- XV. FM list of species codes**

I. Codes for countries, vessels and gear

Codes for countries

Code	Country
ALB	Albania
CYP	Cyprus
ESP	Spain
FRA	France
GRC	Greece
HRV	Croatia
ITA	Italy
MAL	Malta
MAR	Morocco
MON	Montenegro
SLO	Slovenia

Vessel codes and characteristics

Vessel code	Vessel Name	Type	Lenght (m)	Tonnage (TJB)	Year	Material	Power (kW)	Warp diam (mm)	Warp length (m)
AND	Andrea	R	29.5	211	1998	aluminium	1300	14	2250
BIM	Bianca Maria	P	26.81	116	1988	wood	485	12	3000
CHA	Charif Alidrissi	R	41	397	1986	steel	808	22	3000
COR	Cornide de Saavedra	R	66.7	1524	1970	steel	1651	29	2700
DAP			0	0	0		0	0	0
DEM	Demetrios	P	27.77	78.24	1991	steel	537	12	3000
EGU	Elisa Guidotti	P	29	69	1991	bois	330	14	2500
EVA	Evagelistria	P	29.1	59.45	2000	steel	497	12	1800
FRP	Francesco Padre	P	25	88	1984	steel	660	14	3000
FUL	Fulmine	P	29	147.2	0	wood	736	14	2500
GAB			0	0	0		0	0	0
GIS			0	0	0		0	0	0
IGO	Igor	P	22.5	102	1979	iron	345	14	2500
IRO	Ioannis Rossos	P	26.3	115.75	1986	iron	368	12	3000
LEU	L'Europe	R	29.6	259.69	1993	aluminium	690	16	2700
LIB			0	0	0		0	0	0
NAU	Nautilus	P	28.4	138	1991	iron	600	14	2500
NAV	Francisco Paula Navarro	R	30.5	178	1987	wood	750	18	2200
NUS	Nuovo Splendore	P	29.45	134.51	1967	wood	685	16	2450
PAR	Kapetan Paraschos	P	26.1	85.71	1989	wood	386	12	2000
PAS			0	0	0		0	0	0
PEC	Pasquale e Cristina	P	33.06	158.77	1996	wood	923	16	2500
PRI	Principessa I	P	32	165	1995	steel	403	14	2500
ROS	Roselys	R	0	0	0	wood	0	0	0
SAN	Sant'Anna	P	32.2	197.1	1981	steel	744	14	3100

Codes for the gear

Nature	Gear	MEDITS code	Comments
Trawl	Large opening and 4 faces	GOC73	Standard for all vessels
Rigging	With legs	GC73	Standard for all vessels
Doors	Morgère WH S8	WHS8	Standard for all vessels

II. Stratification scheme (by stratum number))

GSA	Country	Stratum	Depth (m)	Surface (km ²)	Area
1	Spain	11101 a	10-50	510	Alboran Sea
1	Spain	11102 a	50-100	1951	
1	Spain	11103 a	100-200	1086	
1	Spain	11104 a	200-500	3461	
1	Spain	11105 a	500-800	4912	
2	Spain	11106 b	10-50	0	Alboran Island
2	Spain	11107 b	50-100	130	Alboran Island
2	Spain	11108 b	100-200	132	
2	Spain	11109 b	200-500	221	
2	Spain	11110 a	500-800	350	
3	Morocco	11401 a	10-50	355	West Morocco
3	Morocco	11402 a	50-100	444	
3	Morocco	11403 a	100-200	487	
3	Morocco	11404 a	200-500	3580	
3	Morocco	11405 a	500-800	1108	
3	Morocco	11406 b	10-50	878	East Morocco
3	Morocco	11407 b	50-100	1098	
3	Morocco	11408 b	100-200	938	
3	Morocco	11409 b	200-500	3507	
3	Morocco	11410 b	500-800	1446	
5	Spain	11501 a	10-50	0	West Balears
5	Spain	11502 a	50-100	1170	West Balears
5	Spain	11503 a	100-200	1773	
5	Spain	11504 a	200-500	1123	
5	Spain	11505 a	500-800	2030	
5	Spain	11507 b	50-100	2255	East Balears
5	Spain	11508 b	100-200	1472	
5	Spain	11509 b	200-500	1518	
5	Spain	11510 b	500-800	1315	
6	Spain	11201 a	10-50	1130	Valenciana
6	Spain	11202 a	50-100	4095	
6	Spain	11203 a	100-200	3302	
6	Spain	11204 a	200-500	4242	
6	Spain	11205 a	500-800	3159	
6	Spain	11301 a	10-50	1896	Tramontana
6	Spain	11302 a	50-100	7219	
6	Spain	11303 a	100-200	3587	
6	Spain	11304 a	200-500	2477	
6	Spain	11305 a	500-800	1399	
7	France	12101 a	10-50	1482	West Gulf of Lions
7	France	12102 a	50-100	3911	
7	France	12103 a	100-200	819	
7	France	12104 a	200-500	709	
7	France	12105 a	500-800	660	
7	France	12106 b	10-50	696	East Gulf of Lions
7	France	12107 b	50-100	2610	
7	France	12108 b	100-200	1734	
7	France	12109 b	200-500	653	
7	France	12110 b	500-800	586	
8	France	13101 a	10-50	0	North East Corsica
8	France	13102 a	50-100	521	North East Corsica
8	France	13103 a	100-200	234	
8	France	13104 a	200-500	920	
8	France	13105 a	500-800	867	

GSA	Country	Stratum	Depth (m)	Surface (km ²)	Area
8	France	13106 b	10-50	0	South East Corsica
8	France	13107 b	50-100	524	South East Corsica
8	France	13108 b	100-200	153	
8	France	13109 b	200-500	383	
8	France	13110 b	500-800	960	
9	Italy	13201 a	10-50	657	North Ligurian Sea
9	Italy	13202 a	50-100	729	
9	Italy	13203 a	100-200	658	
9	Italy	13204 a	200-500	1737	
9	Italy	13205 a	500-800	2093	
9	Italy	13206 b	10-50	2053	East Ligurian Sea
9	Italy	13207 b	50-100	1598	
9	Italy	13208 b	100-200	3186	
9	Italy	13209 b	200-500	2449	
9	Italy	13210 b	500-800	879	
9	Italy	13211 c	10-50	945	North Tyrrhenian Sea
9	Italy	13212 c	50-100	1506	
9	Italy	13213 c	100-200	2732	
9	Italy	13214 c	200-500	2828	
9	Italy	13215 c	500-800	3071	
9	Italy	13216 d	10-50	2107	Central Tyrrhenian Sea
9	Italy	13217 d	50-100	2159	
9	Italy	13218 d	100-200	4302	
9	Italy	13219 d	200-500	3573	
9	Italy	13220 d	500-800	3148	
10	Italy	13401 a	10-50	1194	South East Tyrrhenian Sea
10	Italy	13402 a	50-100	1224	
10	Italy	13403 a	100-200	2095	
10	Italy	13404 a	200-500	3238	
10	Italy	13405 a	500-800	5248	
10	Italy	13406 b	10-50	622	South West Tyrrhenian Sea
10	Italy	13407 b	50-100	1003	
10	Italy	13408 b	100-200	1224	
10	Italy	13409 b	200-500	1966	
10	Italy	13410 b	500-800	2441	
11	Italy	13301 a	10-50	822	South East Sardinia
11	Italy	13302 a	50-100	382	
11	Italy	13303 a	100-200	351	
11	Italy	13304 a	200-500	589	
11	Italy	13305 a	500-800	502	
11	Italy	13306 b	10-50	910	North East Sardinia
11	Italy	13307 b	50-100	1592	
11	Italy	13308 b	100-200	839	
11	Italy	13309 b	200-500	765	
11	Italy	13310 b	500-800	855	
11	Italy	13311 c	10-50	627	North Sardinia
11	Italy	13312 c	50-100	796	
11	Italy	13313 c	100-200	512	
11	Italy	13314 c	200-500	500	
11	Italy	13315 c	500-800	242	
11	Italy	13316 d	10-50	431	North West Sardinia
11	Italy	13317 d	50-100	541	
11	Italy	13318 d	100-200	896	
11	Italy	13319 d	200-500	471	
11	Italy	13320 d	500-800	335	
11	Italy	13321 e	10-50	1096	West Sardinia
11	Italy	13322 e	50-100	446	

GSA	Country	Stratum	Depth (m)	Surface (km ²)	Area
11	Italy	13323 e	100-200	927	
11	Italy	13324 e	200-500	412	
11	Italy	13325 e	500-800	260	
11	Italy	13326 f	10-50	783	South West Sardinia
11	Italy	13327 f	50-100	987	
11	Italy	13328 f	100-200	2335	
11	Italy	13329 f	200-500	1620	
11	Italy	13330 f	500-800	1041	
11	Italy	13331 g	10-50	705	South Sardinia
11	Italy	13332 g	50-100	350	
11	Italy	13333 g	100-200	768	
11	Italy	13334 g	200-500	1060	
11	Italy	13335 g	500-800	1227	
15	Malta	13501 a	10-50	152	Malta
15	Malta	13502 a	50-100	1473	
15	Malta	13503 a	100-200	3076	
15	Malta	13504 a	200-500	3353	
15	Malta	13505 a	500-800	2526	
16	Italy	13411 c	10-50	2979	Strait of Sicily
16	Italy	13412 c	50-100	5943	
16	Italy	13413 c	100-200	5565	
16	Italy	13414 c	200-500	6972	
16	Italy	13415 c	500-800	9927	
17	Italy	21101 a	10-50	17300	North Adriatic Sea
17	Italy	21102 a	50-100	8200	
17	Italy	21103 a	100-200	0	
17	Italy	21104 a	200-500	0	
17	Italy	21105 a	500-800	0	
17	Italy	21106 b	10-50	4700	Central Adriatic Sea
17	Italy	21107 b	50-100	10350	
17	Italy	21108 b	100-200	14950	
17	Italy	21109 b	200-500	3900	
17	Italy	21110 b	500-800	950	
17	Slovenia	21111 c	10-50	184	North Adriatic-Slovenia
17	Slovenia	21112 c	50-100	0	
17	Slovenia	21113 c	100-200	0	
17	Slovenia	21114 c	200-500	0	
17	Slovenia	21115 c	500-800	0	
17	Croatia	21116 d	10-50	7308	North East Adriatic-Croatia
17	Croatia	21117 d	50-100	14785	
17	Croatia	21118 d	100-200	7225	
17	Croatia	21119 d	200-500	2409	
17	Croatia	21120 d	500-800	0	
18	Italy	22121 e	10-50	261	South West Adriatic Sea
18	Italy	22122 e	50-100	509	
18	Italy	22123 e	100-200	1348	
18	Italy	22124 e	200-500	332	
18	Italy	22125 e	500-800	860	
18	Italy	22126 f	10-50	329	South West Adriatic Sea
18	Italy	22127 f	50-100	599	
18	Italy	22128 f	100-200	1809	
18	Italy	22129 f	200-500	472	
18	Italy	22130 f	500-800	350	
18	Italy	22131 g	10-50	290	South West Adriatic Sea
18	Italy	22132 g	50-100	689	
18	Italy	22133 g	100-200	1214	
18	Italy	22134 g	200-500	260	

GSA	Country	Stratum	Depth (m)	Surface (km ²)	Area
18	Italy	22135 g	500-800	336	
18	Italy	22136 h	10-50	1702	South West Adriatic Sea
18	Italy	22137 h	50-100	1307	
18	Italy	22138 h	100-200	1407	
18	Italy	22139 h	200-500	707	
18	Italy	22140 h	500-800	492	
18	Albania	22141 i	10-50	568	South East Adriatic-Albania
18	Albania	22142 i	50-100	2231	
18	Albania	22143 i	100-200	2186	
18	Albania	22144 i	200-500	1840	
18	Albania	22145 i	500-800	1910	
18	Montenegro	22146 j	10-50	280	South Adriatic-Montenegro
18	Montenegro	22147 j	50-100	1100	
18	Montenegro	22148 j	100-200	1700	
18	Montenegro	22149 j	200-500	1150	
18	Montenegro	22150 j	500-800	770	
19	Italy	22101 a	10-50	412	North-Western Ionian Sea (East Sicily)
19	Italy	22102 a	50-100	377	
19	Italy	22103 a	100-200	334	
19	Italy	22104 a	200-500	650	
19	Italy	22105 a	500-800	641	
19	Italy	22106 b	10-50	326	North-Western Ionian Sea (South Calabria)
19	Italy	22107 b	50-100	225	
19	Italy	22108 b	100-200	257	
19	Italy	22109 b	200-500	939	
19	Italy	22110 b	500-800	1370	
19	Italy	22111 c	10-50	599	North-Western Ionian Sea (North Calabria)
19	Italy	22112 c	50-100	321	
19	Italy	22113 c	100-200	393	
19	Italy	22114 c	200-500	1327	
19	Italy	22115 c	500-800	1190	
19	Italy	22116 d	10-50	787	North-Western Ionian Sea (Apulia)
19	Italy	22117 d	50-100	778	
19	Italy	22118 d	100-200	1680	
19	Italy	22119 d	200-500	1439	
19	Italy	22120 d	500-800	2302	
20	Greece	22201 a	10-50	2916	East Ionian Sea
20	Greece	22202 a	50-100	4365	
20	Greece	22203 a	100-200	2536	
20	Greece	22204 a	200-500	3158	
20	Greece	22205 a	500-800	3848	
22	Greece	22301 a	10-50	2467	Argosaronikos
22	Greece	22302 a	50-100	587	
22	Greece	22303 a	100-200	7143	
22	Greece	22304 a	200-500	6074	
22	Greece	22305 a	500-800	8645	
22	Greece	22401 a	10-50	8645	North Aegean Sea
22	Greece	22402 a	50-100	8489	
22	Greece	22403 a	100-200	15823	
22	Greece	22404 a	200-500	19774	
22	Greece	22405 a	500-800	15426	
22	Greece	22501 a	10-50	4918	South Aegean Sea (encl. GSA 23: Crete)
22	Greece	22502 a	50-100	4090	
22	Greece	22503 a	100-200	13269	
22	Greece	22504 a	200-500	18100	
22	Greece	22505 a	500-800	22224	
25	Cyprus	32101 a	10-50	796	Cyprus

GSA	Country	Stratum	Depth (m)	Surface (km ²)	Area
25	Cyprus	32102 a	50-100	717	
25	Cyprus	32103 a	100-200	918	
25	Cyprus	32104 a	200-500	2245	
25	Cyprus	32105 a	500-800	6430	

III. Target number of hauls by area

Country	GSA	Strata	Surface (km ²)	No Hauls	Area
Spain	1, 2	111	12753	46	Northern Alboran Sea
Morocco	3	114	13841	63	Southern Alboran Sea
Spain	5	115	12656	60	Balearic Islands
Spain	6	112-113	32506	92	Northern Spain
France	7, 8	121, 131	18422	95	Gulf of Lions & Corsica
Italy	9	132	42410	120	Ligurian, North and Central Tyrrhenian Sea
Italy	10	134a-b	20255	70	Central and Southern Tyrrhenian Sea
Italy	11	133	26975	98	Sardinia
Malta	15	135	10580	45	Malta
Italy	16	134c	31386	120	Strait of Sicily
Italy	17	211a-b	60350	121	Northern Adriatic Sea
Slovenia	17	211c	184	2	Northern Adriatic Sea
Croatia	17	211d	31727	60	Northern Adriatic Sea
Italy	18	221e-h	15273	58	Southern Adriatic Sea
Albania	18	221i	8735	40	Southern Adriatic Sea
Montenegro	18	221j	5000	20	Southern Adriatic Sea
Italy	19	221a-d	16347	70	North-Western Ionian Sea
Greece	20	222	16823	32	Eastern Ionian Sea
Greece	22	223	24916	21	Aegean Sea (Argosaronikos)
Greece	22	224	68157	65	Aegean Sea (North)
Greece	22	225	62601	61	Aegean Sea (South)
Cyprus	25	321	11106	26	Cyprus

V. Codes for recorded species, of the observations on hauls and of quadrants

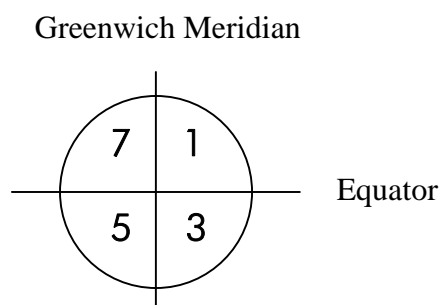
Codes of recorded species (Position 83 in the file 1)

MEDITS code	Nature	Comments
0	No standard species recorded	
1	Only the species of the reference list are recorded	See Annex VI
2	The species of the reference list plus some others are recorded	
3	All the caught species are recorded	See Annex XV
4	Species from a national list	

Coding of the observations (Position 109 in the file 1)

Meditis code	Nature	Comments
0	No problem	
1	Slight plugging of the net	
2	Heavy plugging of the net	
3	High abundance of jellyfish	
4	High abundance of plants in the net	
5	Tears of the net	
6	High abundance of benthos	
7		
8		
9	Other	

Coding of the quadrants (Positions 38 and 60 in the file 1)



VI. List of the reference species

Scientific name	Date ¹	CODE	Common name	
			Français	English
<i>Aspitrigla cuculus</i>	1998	ASPI CUC	Grondin rouge	Red gurnard
<i>Boops boops</i>	2006	BOOPBOO	Bogue	Bogue
<i>Citharus linguatula</i>	1994	CITH MAC	Feuille	Spotted flounder
<i>Eutrigla gurnardus</i>	1994	EUTR GUR	Grondin gris	Grey gurnard
<i>Galeus melastomus</i>	1998	GALU MEL	Chien espagnol	Blackmouth catshark
<i>Helicolenus dactylopterus</i>	1994	HELI DAC	Rascasse de fond	Rockfish
<i>Lepidorhombus boscii</i>	1994	LEPM BOS	Cardine à quatre taches	Four-spotted megrim
<i>Lophius budegassa</i>	1994	LOPH BUD	Baudroie rousse	Black-bellied angler
<i>Lophius piscatorius</i>	1994	LOPH PIS	Baudroie commune	Angler
<i>Merluccius merluccius</i>	1994	MERL MER	Merlu commun	European hake
<i>Micromesistius poutassou</i>	1994	MICM POU	Merlan bleu	Blue whiting
<i>Mullus barbatus</i>	1994	MULL BAR	Rouget-barbet de vase	Red mullet
<i>Mullus surmuletus</i>	1994	MULL SUR	Rouget-barbet de roche	Striped red mullet
<i>Pagellus acarne</i>	1994	PAGE ACA	Pageot acarné	Axillary seabream
<i>Pagellus bogaraveo</i>	1994	PAGE BOG	Dorade rose	Blackspot seabream
<i>Pagellus erythrinus</i>	1994	PAGE ERY	Pageot commun	Common pandora
<i>Sparus pagrus</i>	> 1996	SPAR PAG	Pagre commun	Common seabream
<i>Phycis blennoides</i>	1994	PHYI BLE	Phycis de fond	Greater forkbeard
<i>Raja clavata</i>	1994	RAJA CLA	Raie bouclée	Thornback ray
<i>Scyliorhinus canicula</i>	1998	SCYO CAN	Petite roussette	Smallspotted catshark
<i>Solea vulgaris</i>	1994	SOLE VUL	Sole commune	Common sole
<i>Spicara flexuosa</i>	1994	SPIC FLE	Gerle	Picarel
<i>Spicara smaris</i>	1998	SPIC SMA	Picarel	Picarel
<i>Trachurus mediterraneus</i>	1994	TRAC MED	Chinchard à queue jaune	Mediterranean horse mackerel
<i>Trachurus trachurus</i>	1994	TRAC TRA	Chinchard d'Europe	Atlantic horse mackerel
<i>Trigla lucerna</i>	2006	TRIGLUC	Grondin-perlon	Tub gurnard
<i>Trigloporus lastoviza</i>	1998	TRIP LAS	Grondin camard	Streaked gurnard
<i>Trisopterus minutus capelanus</i>	1994	TRIS CAP	Capelan	Poor-cod
<i>Zeus faber</i>	1994	ZEUS FAB	Saint-Pierre	John dory
Selacians ²	2006			
<i>Aristaeomorpha foliacea</i>	1994	ARIS FOL	Gambon rouge	Giant red shrimp
<i>Aristeus antennatus</i>	1994	ARIT ANT	Crevette rouge	Blue and red shrimp
<i>Nephrops norvegicus</i>	1994	NEPR NOR	Langoustine	Norway lobster
<i>Parapenaeus longirostris</i>	1994	PAPE LON	Crevette rose du large	Deep-water pink shrimp
<i>Eledone cirrhosa</i>	1994	ELED CIR	Poulpe blanc	Horned octopus
<i>Eledone moschata</i>	1997	ELED MOS	Elédone musquée	Musky octopus
<i>Illex coindetti</i>	1994	ILLE COI	Encornet rouge	Broadtail squid
<i>Loligo vulgaris</i>	1994	LOLI VUL	Encornet	European squid
<i>Octopus vulgaris</i>	1994	OCTO VUL	Pieuvre	Common octopus
<i>Sepia officinalis</i>	1994	SEPI OFF	Seiche commune	Common cuttlefish

¹ Year in which the species was introduced in the list (or removed if the year is preceded by >)

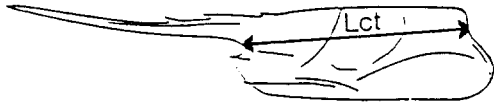
² It is recommended to carry out the observations referring to this list to all the selacian species in the GSAs where it is technically possible. To allow coherent analyses of the results, it is highlighted that the decision to enlarge or not biological observations on selacians must be applied consistently during all the surveys.

Ref. Common names: Fischer W., M.L. Bauchot, M. Schneider (rédacteurs), 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche. (Révision 1). Méditerranée et Mer Noire Zone de pêche 37. Rome, FAO, vol 1 et 2, 1530 p.

VII. Standard length for Crustaceans and Cephalopods

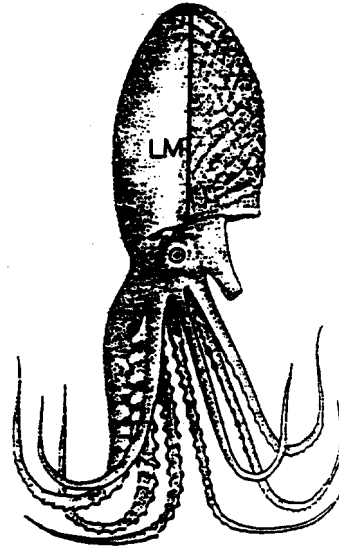
Crustaceans

Lct: cephalo-thoracic length



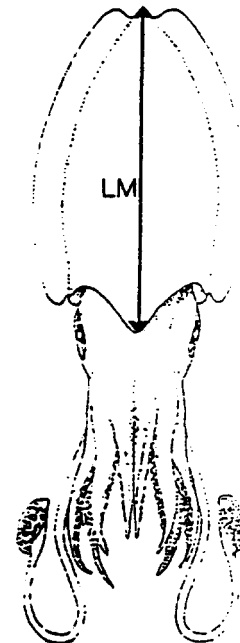
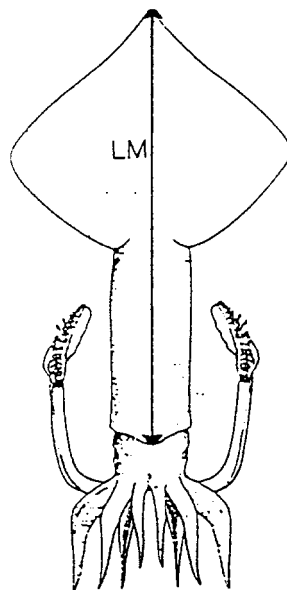
Cephalopods octopoda

LM: mantle length



Cephalopods decapoda

LM: mantle length



VIII. Code of sexual maturity for fish

bony fish

SEX	GONAD ASPECT	MATURATION STATE	STAGE	MEDITS
U	Sex not distinguished by naked eye. Gonads very small and translucent, almost transparent. Sex undetermined.	UNDETERMINED	0	0
F	Small pinkish and translucent ovary shorter than 1/3 of the body cavity. Eggs not visible by naked eye.	IMMATURE = VIRGIN	1	1
M	Thin and whitish testis shorter than 1/3 of the body cavity.			
F	Small pinkish/reddish ovary shorter than 1/2 of the body cavity. Eggs not visible by naked eye.	VIRGIN-DEVELOPING *	2a	2
M	Thin whitish testis shorter than 1/2 of the body cavity.			
F	Pinkish-reddish/reddish- orange and translucent ovary long about 1/2 of the body cavity. Blood vessels visible. Eggs not visible by naked eye.	RECOVERING *	2b	
M	Whitish/pinkish testis, more or less symmetrical, long about 1/2 of the body cavity.			
F	Ovary pinkish-yellow in colour with granular appearance, long about 2/3 of the body cavity. Eggs are visible by naked eye through the ovarian tunica, which is not yet translucent. Under light pressure, eggs are not expelled.	MATURING	2c	
M	Whitish to creamy testis long about 2/3 of the body cavity. Under light pressure, sperm is not expelled.			
F	Ovary orange-pink in colour, with conspicuous superficial blood vessels, long from 2/3 to full length of the body cavity. Large transparent, ripe eggs are clearly visible and could be expelled under light pressure. In more advanced conditions, eggs escape freely.	MATURE/SPAWNER	3	3
M	Whitish-creamy soft testis long from 2/3 to full length of the body cavity. Under light pressure, sperm could be expelled. In more advanced conditions, sperm escapes freely.			
F	Reddish ovary shrunk to about 1/2 length of the body cavity. Flaccid ovarian walls; ovary may contain remnants of disintegrating opaque and/or translucent eggs.	SPENT	4a	4
M	Bloodshot and flabby testis shrunken to about 1/2 length of the body cavity.			
F	Pinkish and translucent ovary long about 1/3 of the body cavity. Eggs not visible by naked eye.	RESTING *	4b	
M	Whitish/pinkish testis, more or less symmetrical, long about 1/3 of the body cavity.			

 Adult specimens

* : WARNING ! Be careful. These stages could be confused each other.

Elasmobranchs (oviparous)

SEX	GONAD ASPECT	MATURATION STATE	STAGE	MEDITS	
N	The specimens aren't sexed.	NOT DETERMINED	0	0	
F	Ovary is barely discernible with small isodiametric eggs. Distal part of oviducts is thick-walled and whitish. The nidamental glands are less evident.	IMMATURE / VIRGIN	1	1	
M	Claspers are small and flaccid and do not reach the posterior edge of the pelvic fins. Spermducts not differentiated. Testis small and narrow .				
F	Whitish and/or few yellow maturing eggs are visible in the ovary. The distal part of oviducts (uterus) is well developed but empty. The nidamental glands are small.	MATURING	2	2	
M	Claspers are larger, but skeleton still flexible. They extend to the posterior edge of the pelvic fins. Spermducts well developed eventually beginning to meander.				
F	Ovaries contain yellow eggs (large yolk eggs). The nidamental glands are enlarged and oviducts are distended.	MATURE	3a	3	
M	Claspers extends well beyond the posterior edge of the pelvic fin and their internal structure is generally hard and ossified. Testis greatly enlarged. Spermducts meandering over almost their entire length.				
F	Ovary walls transparent. Oocytes of different sizes, white or yellow. Nidamental glands large. Egg-cases more or less formed in the oviducts (Extruding Stage).	MATURE/EXTRUDING-ACTIVE	3b		
M	Clasper longer than tips of posterior pelvic fin lobes, skeleton hardened with axial cartilages hardened and pointed. Spermducts largely. Sperm flowing on pressure from cloaca (Active Stage).				
F	Ovary walls transparent. Oocytes of different sizes, white or yellow. Oviducts appear much enlarged, collapsed and empty. The nidamental glands diameter are reducing.	RESTING	4		4
M	Clasper longer than tips of posterior pelvic fin lobes, skeleton hardened with axial cartilages still hardened. Spermducts empty and flaccid.				

 Adult specimens

IX Codes of sexual maturity for Crustaceans and Cephalopods

Crustaceans


SEX	REPRODUCTIVE APPARATUS ASPECT	COLOURING OF FRESH OVARY	MATURATION STATE	STAGE	MEDITS
U	Sex not distinguished by naked eye. Sex undetermined	translucid	UNDETERMINED	0	0
F	Ovary hardly visible in transparence. After dissection of the tegument ovary is small and lobes are flaccid, stringy and poorly developed. <i>A. foliacea</i> and <i>A. antennatus</i> no spermatophores on thelycum.	Whitish or traslucid	IMMATURE = VIRGIN *	1	1 FEMALE
M	Petasma is not much visible, and there are not spermatic masses (emi-spermatophores) on the seminal ampullae, located on side of the V pair of pereopods. <i>A. foliacea</i> and <i>A. antennatus</i> : long rostrum.				
F	Ovary status to develop. Cephalic and lateral lobes are small but distinguishable by naked eye. Abdominal extension are thin and just visible.	<i>A. foliacea</i> : flesh coloured; <i>A. antennatus</i> : Ivory coloured with orange pink-violet dotting. <i>N. norvegicus</i> : cream. <i>P. longirostris</i> : cream orange.	VIRGIN DEVELOPING **	2a	2 FEMALE
M	Petasma appears visible and nearly or completely joined, but there are no spermatic masses in the seminar ampullae. <i>A. foliacea</i> & <i>A. antennatus</i> : long or intermediate rostrum.				
F	Ovary status to re-develop. Cephalic and lateral lobes are small but distinguishable by naked eye. Abdominal extension are thin and just visible. Occasionally presence of spermatophores in <i>A. foliacea</i> and <i>A. antennatus</i> .	<i>A. foliacea</i> : flesh coloured; <i>A. antennatus</i> : Ivory coloured with orange pink-violet dotting. <i>N. norvegicus</i> : cream. <i>P. longirostris</i> : cream orange.	RECOVERING**	2b	
M	Petasma appears completely joined, but there are no spermatic masses in the seminar ampullae. <i>A. foliacea</i> & <i>A. antennatus</i> : short rostrum.				
F	Ovary developed and occupies almost entirely the dorsal portion. The cephalic and lateral lobes are much developed and have a turgid consistence.	<i>A. foliacea</i> : light and dark grey; <i>A. antennatus</i> : lilla; <i>N. norvegicus</i> : light green; <i>P. longirostris</i> : light green or grey green.	MATURING OR ALMOST MATURE	2c	
M					
F	Turgid ovary extends to the whole dorsal portion, covery the organs below. Lobes and extensions well developed, in particular the abdominal extention are much evident. Oocytes well visible.	<i>A. foliacea</i> : black; <i>A. antennatus</i> : violet; <i>N. norvegicus</i> : dark grey; <i>P. longirostris</i> : brigh green or olive green.	MATURE	2d	
M	Petasma is perfectly visible and completely joined. Spermatic masses in seminar ampullae. <i>A. foliacea</i> & <i>A. antennatus</i> : small rostrum.				
F	Resting ovary. Presence of spermatophores in <i>A. foliacea</i> and <i>A. antennatus</i> .	Uncoloured.	RESTING ADULT*	2e	
F (<i>N. norvegicus</i>)	Eggs on pleiopods		BERRIED	3	

 Adult specimens

*, **: WARNING ! Be careful. These stages could be confused each other.

Cephalopods

SEX	REPRODUCTIVE APPARATUS ASPECT	EGGS SIZE (mm)	SPERMATOPHORES DEVELOPMENT	MATURATION STATE	STAGE	MEDITS
U	Sex not distinguished by naked eye. Sex undetermined.	Total absence of eggs.	Total absence of spermatophores.	UNDETERMINED	0	0
F	Small and translucent Nidamental Glands (NG) / Oviducal Glands (OG). Ovary is semi-transparent, stringy and lacking granular structure Small semi-transparent NG / OG. Oviduct meander not visible.	<i>L. vulgaris</i> & <i>I. coindetii</i> : no eggs <i>S. officinalis</i> : $\phi < 2\text{mm}$ <i>E. moschata</i> : $\phi < 4\text{mm}$ <i>E. cirrhosa</i> $\phi < 2\text{mm}$ <i>O. vulgaris</i> $\phi < 1\text{mm}$	Total absence of spermatophores	IMMATURE = VIRGIN	1	1
M	Testis small. Spermatophoric complex (SC) semi-transparent with not visible Vas deferens. Penis appears as a small prominence of SC.					
F	NG / OVG enlarged. NG covering some internal organs. Whitish ovary with granular structure clearly visible, not reaching the posterior half of the mantle cavity. Oviduct meander clearly visible.	Very small eggs	Absence of spermatophores	DEVELOPING	2a	
M	Enlarged testis with structure not clearly visible. The Vas deferens whitish or white and the spermatophoric organ with white streak.					
F	Large NG covering the viscera below. Ovary occupies the whole posterior half of mantle cavity, containing reticulated oocytes of all sizes tightly packed and probably a few ripe ova at its proximal part. Oviducts fully developed but empty.	<i>L. vulgaris</i> & <i>I. coindetii</i> : maturing eggs visible by naked eye. <i>S. officinalis</i> : 2,1mm < ϕ < 4mm <i>E. moschata</i> : 4mm < ϕ < 11mm <i>E. cirrhosa</i> : 2mm < ϕ < 5mm <i>O. vulgaris</i> : 1mm < ϕ < 2mm	<i>L. vulgaris</i> , <i>I. coindetii</i> and <i>S. officinalis</i> : few immature spermatophores in Needham's sac. <i>E. moschata</i> , <i>E. cirrhosa</i> , <i>O. vulgaris</i> : few spermatophores, barely developed and not functional	MATURING	2b	2
M	The Vas deferens white, meandering, enlarged. The Needham's sac (SS) with structureless whitish particles inside. Normally the Needham's sac is without functional spermatophores but sometimes some immature/abortive ones could occur. The testis tight, crispy, with visible structure.					
F	Large NG as previously. Ovary containing higher percentage of large reticulated eggs and some large ripe ova with smooth surface. In Teuthoidea ripe ova in oviducts.	<i>L. vulgaris</i> & <i>I. coindetii</i> : amber- colored and isodiametric eggs in oviducts and in part of the ovary ($\phi = 2\text{mm}$ in <i>Loligo</i> and $\phi = 1\text{mm}$ in <i>Illex</i>). <i>S. officinalis</i> : medium eggs (4,1mm < ϕ < 6,0mm) and big eggs (6,1mm < ϕ < 8mm) <i>E. moschata</i> : $\phi > 1\text{mm}$ (striped eggs). <i>E. cirrhosa</i> : $\phi > 5\text{mm}$ <i>O. vulgaris</i> : $\phi > 2\text{mm}$	Well developed spermatophores	MATURE	3a	3
M	Testis as before. Spermatophores packed in the Needham's sac.					
F	NG/OG large but soft and running. Ovary shrank and flaccid, with only immature oocytes attached to the central tissue and a few loose large ova in the coelom. In Teuthoidea oviduct may contain some mature ova but is no longer packed.	Few large ova	Disintegrating spermatophores	SPENT	3b	
M	Disintegrating spermatophores in the Needham's sac and the penis.					

 Adult specimens

X. Format of the type A files (Data on the haul)

Code)	Name	Position	Type*	Range	Comments
TYPENR	Type of file	1 - 2	2A	TA	Fixed value
PAYS	Country	3 - 5	3A	See Annexe I	ISO code
BATEAU	Vessel	6 - 8	3A	See Annexe I	MEDITS code
ENGIN	Gear	9 - 13	5A	See Annexe I	MEDITS code
GREEMENT	Rigging	14 - 17	4A	See Annexe I	MEDITS code
PANNEAUX	Doors	18 - 21	4A	See Annexe I	MEDITS code
AN	Year	22 - 25	4N		Ex: 2000, 2001
MOIS	Month	26 - 27	2N	1 to 12	
JOUR	Day	28 - 29	2N	1 to 28/29/30/31	
NOTRAI	Haul number	30 - 32	3N	1 to 999	One series by vessel/year
FERCHA	Codend closing	33 - 33	1A	S, C	S: without: C: controlled
HDEB	Shooting time	34 - 37	4N	0 à 2400	In UT Ex: 7 h 25 min > 725.
QUADEB	Shooting quadrant	38 - 38	1N	1, 3, 5, 7	See Annexe X
LATDEB	Shooting latitude	39 - 45	7N	3400 to 4600	Ex: 36° 40,22' > 3640,22.
LGNDEB	Shooting longitude	46 - 52	7N	0 to 2900	Ex: 4° 19,84' > 419,84.
PRODEB	Shooting depth	53 - 55	3N	0, 10 to 800	At the trawl position, in meters; unknown: 0
HFIN	Hauling time	56 - 59	4N	0 to 2400	In UT Ex: 7 h 25 min > 725.
QUAFIN	Hauling Quadrant	60 - 60	1N	1, 3, 5, 7	See Annexe X
LATFIN	Hauling latitude	61 - 67	7N	3400 to 4600	Ex: 36° 40,22' > 3640,22.
LGNFIN	Hauling longitude	68 - 74	7N	0 to 2900	Ex: 4° 19,84' > 419,84.
PROFIN	Hauling depth	75 - 77	3N	0, 10 à 800	At the trawl position, in meters; unknown: 0
DUREE	Haul duration	78 - 79	2N	5 to 90	In minutes
VALID	Validity	80 - 80	1A	V, I	V: valid; I: invalid. (1)
PARCOU	Course	81 - 81	1A	R, N	R: rectilinear N: not rectilinear
ESPENR	Recorded species	82 - 83	2N	See Annex IV	MEDITS code
DIST	Distance	84 - 87	4N	1000 to 9999	Distance over ground in meters
OUPER	Vertical opening of the trawl	88 - 90	3N	10 to 99	In decimetres
ECAIL	Wing opening	91 - 93	3N	50 to 250	In decimetres
PRGÉO	Geometrical precision	94 - 94	1A	M, E	M: measured; E: estimated.
LONBRA	Bridles length	95 - 97	3N	100 to 200	In meters
LONFUN	Warp length	98 - 101	4N	100 to 2200	In meters
DIAFUN	Warp diameter	102 - 103	2N	10 to 30	In millimetres
STAHYDRO	Hydrological station	104 - 108	5A		National coding
Observ	Observations	109 - 109	1N	1 to 9	MEDITS code (Annex IV)

* All numerical fields (N) are right justified; all alphanumeric fields (A)fields are left justified

Note:

(1) For the invalid hauls (I), no information on species is needed.

XI. Format of the type B files (Catches by haul)

Code	Name	Position	Type*	Range	Comments
TYPENR	Type of file	1 - 2	2A	TB	Fixed value
PAYS	Country	3 - 5	3A	See Annexe I	ISO code
BATEAU	Vessel	6 - 8	3A	See Annexe I	MEDITS code
AN	Year	9 - 12	4N		Ex: 2000, 2001
NOTRAI	Haul number	13 - 15	3N	1 to 999	One series by vessel/year.
FERCHA	Codend closing	16 - 16	1A	S, C	S: without; C: controlled
PARTIT	Part of the codend	17 - 17	1A	A, M, P, S	Mandatory if FERCHA = C. A: anterior; M: middle; P: posterior; S sum of the three parts
CATFAU	Faunistic category	18 - 18	1A	A to E See Annexe V	MEDITS code, filled in only if the 3 following fields are empty.
GENRE	Genus	19 - 22	4A	See Annex XV	Following the reference list
ESP	Species	23 - 25	3A	See Annex XV	Following the reference list
LIREF	Name of the reference list	26 - 27	2A	See Annex XV	NCC or MEDITS code
PTOT	Total weight in the haul	28 - 34	7N	0 to 9999999, space	For the given species, in grams
NBTOT	Total number in the haul	35 - 41	7N	0 à 9999999	For the given species. Should be equal to the sum of the 3 following fields
NBFEM	Nb of females in the haul	42 - 48	7N	0 to 9999999	
NBMAL	Nb of males	49 - 55	7N	0 to 9999999	
NbInd	Nb of undetermined	56 - 62	7N	0 to 9999999	Undetermined or not determined

* All numerical fields (N) are right justified; all alphanumeric fields (A) fields are left justified

XII. Format of type C files (biological parameters)

Code	Name	Position	Type*	Range	Comments
TYPENR	Type of file	1 - 2	2A	TC	Fixed value
PAYS	Country	3 - 5	3A	See Annex I	ISO code
BATEAU	Vessel	6 - 8	3A	See Annex I	MEDITS code
AN	Year	9 - 12	4N		Ex 2000, 2001
NOTRAI	Haul number	13 - 15	3N	1 to 999	One series by vessel/year
FERCHA	Codend closing	16 - 16	1A	S, C	S: without; C: controlled
PARTIT	Part of the codend	17 - 17	1A	A, M, P, S	Mandatory if FERCHA = C. A: anterior; M: middle; P: posterior; S sum of the three parts
GENRE	Genus	18 - 21	4A	See Annex XV	Following the reference list
ESP	Species	22 - 24	3A	See Annex XV	Following the reference list
CODLON	Length classes code	25 - 25	1A	m, 0, 1	Types of classes: m: 1 mm; 0: 0.5 cm; 1: 1cm
PFRAC	Weight of the fraction** in the whole haul	26 - 31	6N	0 to 999999	In grams
PECHAN	Weight of the sample really measured in this fraction	32 - 37	6N	0 à 999999	In grams
SEXE	Sex	38 - 38	1A	M, F, I, N	M: male; F: female; I: unde- termined; N: not determined.
NBSEX	Nb of individual of the above se measured in the sample	39 - 44	6N	1 to 999999, space	Unknown: space
CLALON	Length class	45 - 48	4N	1 to 9999	Identifier: lower limit of the class in millimetres; ex: 30.5- 31 cm = 305 (with CODLON = 0); 30-31 cm = 300 (with CODLON = 1) and 26-27 mm = 26 (with CODLON = m)
MATUR	Maturity	49 - 49	1N	0 to 4. See Annexes VIII & IX	0: not determined; 1: immature; 2: maturing; 3: mature or spawning; 4: post-spawning
MATSUB	Maturity sub-staging	50 - 50	1A	See Annexes VIII & IX	Sub-stages of maturity, from a to e (optional)
NBLON	No of individuals in the length class and the maturity stage	51 - 57	6N	1 to 999999	The length classes without any individual are excluded from the file. The sum of No of individuals par class and sex is the No of individuals in the sex and the sample.

* All numerical fields (N) are right justified; all alphanumeric fields (A) fields are left justified

** The word "Fraction" means any sub-group of individual from the total catch of a species (males, females, large sized individuals, small individuals, juveniles, etc.) on which it could be proceed to a sub-sample. For example: total weight = 1000 g which is divided into 100g of big individuals and 900 g of small. The big individuals will be entirely measured (PFRAC = 100; PECHAN = 100). The small ones will be sub-sampled with a ratio of 1/10 (PFRAC + 900; PECHAN = 90)

XIII. Format of type D files (Temperature data) and codes for the temperature measuring systems

Format of type 4 files (Temperature data)

Code	Name	Position	Type*	Range	Comments
TYPENR	Type of file	1 - 2	2A	TD	Fixed value
PAYS	Country	3 - 5	3A	See Annexe I	ISO code
BATEAU	Vessel	6 - 8	3A	See Annexe I	MEDITS code
AN	Year	9 - 12	4N		Ex: 2000, 2001
NOTRAI	Haul number	13 - 15	3N	1 to 999	One series by vessel/year
TODEB	Bottom temperature (beginning of the haul)	16 - 20	5N	0.00 to 30.00	in C° with two decimal positions
TOFIN	Bottom temperature (end of the haul)	21 - 25	5N	0.00 to 30.00	in C° with two decimal positions
METHOD	Measuring system	26 - 27	2A	See infra	MEDITS code

* All numerical fields (N) are right justified; all alphanumeric fields (A) fields are left justified

MEDITS Codes for the temperature measuring systems

Code	Systeme
VA	Vemco- Minilog TDR -5 to +35 C°
XA	XBT
SA	SCANMAR
CTD	CTD probe

XIV. Format of type T files (List of hauls by stratum)

Code)	Name	Position	Type*	Range	Comments
TYPENR	Type of file	1 - 2	2A	TR	Fixed value
AN	Year	3 - 6	4N		Ex: 2000, 2001
PAYS	Country	7 - 9	3A	See Annexe I	ISO code
BATEAU	Vessel	10 - 12	3A	See Annexe I	MEDITS code
NOTRAI	Haul number	13 - 15	3N	1 to 999	One series by vessel/year
N°STRATE	Number of the stratum	16 - 20	5N	See Annex II	

XV. FM list of species codes

**FAUNISTIC LIST OF THE MEDITERRANEAN
To be used in the trawl surveys**

Name of the list: FM

WARNING

The present list is destined to code the marine species encountered in the Mediterranean. It has been built following the principle used in the Nordic Code Centre (Stockholm). For most of the species the codes are identical to those proposed by the NCC. However some species can be coded differently. In addition numerous Mediterranean species are not included in the NCC code and have been added. So the present list is specific. It has to be referred as the FM list.

The initial list was made to be used during the surveys conducted by Ifremer in the western Mediterranean (French and Algerian coasts). Its use has been spread to the International survey MEDITS since 1994.

The first fish list has been established accordingly to the following work:

Hureau J.-C. et Th. Monod (réd.), 1973. Catalogue des poissons de l'Atlantique du nord-est et de la Méditerranée. Unesco, Paris, Vol I, xxii + 683 p.; vol II, 331 p. [réimpression comprenant le *Supplément 1978*, par E. Tortonese et J. -C. Hureau (réd), en 1979].

The reference of the species following this work is reported as "C" (for Clofnam) in the column "Source" with number which is attributed to this species in the Catalogue in the column "Reference".

This list has been increased with reference to the following works:

- Fisher W., M.L., Bauchot et M. Schneider (rédact.), 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche. (Révision 1). Méditerranée et mer Noire. Zone de pêche 37. Volume I. Végétaux et Invertébrés. Volume II. Vertébrés. Publication préparée par la FAO, résultat d'un accord entre la FAO et la Commission des Communautés Européennes (Projet GCP/INT/422/EEC) financée conjointement par ces deux organisations. Rome, FAO, 1530 p.

The reference of the species coming from this book are reported as "F" (for FAO) in the "Source" with the reference given to this species.

- Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1984. Poissons de l'Atlantique du nord-est et de la Méditerranée. Vol. I. UNESCO, Paris, 510 p.
- Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1986. Poissons de l'Atlantique du nord-est et de la Méditerranée. Vol. II et III. UNESCO, Paris, 511-1473.

For most of the Invertebrates, the species have been named accordingly to the following works:

- Zariquiey Alvarez R., 1968. Crustaceos decapodos ibéricos. Invest. Pesq. 32, 510 p.
- Riedl R., 1963. Fauna und flora der Adria. Paul Parey Ed. – 640pp.

The references to these works are mentioned as Z and R respectively in the column "Source".

The scientific names in the list are those of the last update of these various works. The words between parenthesis are the scientific names used in a former coding.

The source file of this list is located at the "Ecologie et modèles pour l'halieutique" department of Ifremer in Nantes. To allow to maintain the uniqueness of this file, the participating teams are invited to contact this department to include any new species (see the form below).

The species codes included in the data tables are based on the FM list. So, to maintain the consistency of the data series, they cannot be changed even if a species name is reviewed. In the last column of the following table, an updated valid name based on the Ifremer taxonomic reference table is given.

Medits Code	Scientific Name	Source	Reference	CATFAU	CODLON	Valid Name (Ifremer reference)
ABRAVER	<i>Abralia veranyi</i>	F	ENOP	C	0	<i>Abralia veranyi</i> (Rüppell, 1844)
ABRRALB	<i>Abra alba</i>	R	RIEDL	E	0	<i>Abra alba</i> (Wood W., 1802)
ACANEXI	<i>Acanthephyra eximia</i>	Z	Z	B	m	<i>Acanthephyra eximia</i> S.I. Smith, 1884
ACANPEL	<i>Acanthephyra pelagica</i>	Z	Z	B	m	<i>Acanthephyra pelagica</i> (Risso, 1816)
ACATPAL	<i>Acantholabrus palloni</i>	C	145.2.1	A	0	<i>Acantholabrus palloni</i> (Risso, 1810)
AEQUOPE	<i>Aequipecten opercularis</i>	F	PECT Aeq 1	D	0	<i>Aequipecten opercularis</i> (Linnaeus, 1758)
ALCYPAL	<i>Alcyonium palmatum</i>			D	0	<i>Alcyonium palmatum</i> Pallas, 1766
ALEPROS	<i>Alepocephalus rostratus</i>	C	30.1.1	A	0	<i>Alepocephalus rostratus</i> Risso, 1820
ALLOMED	<i>Alloteuthis media</i>	F	LOLIG Allot 3	C	0	<i>Alloteuthis media</i> (Linnaeus, 1758)
ALLOSPP	<i>Alloteuthis</i> spp	F	LOLIG Allot	C	0	<i>Alloteuthis</i> Wülker, 1920
ALLOSUB	<i>Alloteuthis subulata</i>	F	LOLIG Allot 2	C	0	<i>Alloteuthis subulata</i> (Lamarck, 1798)
ALOPVUL	<i>Alopias vulpinus</i>	C	9.1.1	A	0	<i>Alopias vulpinus</i> (Bonnaterre, 1788)
AOSFAL	<i>Alosa fallax</i>	C	33.6.3	A	0	<i>Alosa fallax</i> (Lacepède, 1803)
ALPHGLA	<i>Alpheus glaber</i>	F	ALPH Alph 5	B	m	<i>Alpheus glaber</i> (Olivi, 1792)
ALPHPLA	<i>Alpheus platydactylus</i>	Z	Z	B	m	<i>Alpheus platydactylus</i> Coutière, 1897
AMYGLUT	<i>Amygdalum luteum</i>		D'Onghia	E	0	<i>Amygdalum politum</i> (Verrill & Smith, 1880)
ANADDIL	<i>Anadara diluvii</i>	F	ARC Anad 3	D	0	<i>Anadara demiri</i> (Piani, 1981)
ANAMRIS	<i>Anamathia rissoana</i>	Z	Z	B	m	<i>Anamathia rissoana</i> (Roux, 1828)
ANAPBIC	<i>Anapagurus bicorniger</i>	Z	Z	B	m	<i>Anapagurus bicorniger</i> A. Milne-Edwards & Bouvier, 1892
ANAPLAE	<i>Anapagurus laevis</i>	Z	Z	B	m	<i>Anapagurus laevis</i> (Bell, 1845)
ANARGRA	<i>Anarchias euryurus (grassii)</i>	C	73.3.1	A	0	<i>Anarchias euryurus</i> (Lea, 1913)
ANCINIC	<i>Ancistroteuthis lichtensteini</i>	F	ONYCHO	C	0	<i>Ancistroteuthis lichtensteini</i> (De Férussac & D'Orbigny, 1839)
ANGUANG	<i>Anguilla anguilla</i>	C	71.1.1	A	0	<i>Anguilla anguilla</i> (Linnaeus, 1758)
ANTHANT	<i>Anthias anthias</i>	C	124.2.1	A	0	<i>Anthias anthias</i> (Linnaeus, 1758)
ANTOMEG	<i>Antonogadus megalokynodon</i>	C	101.19.2	A	0	<i>Gaidropsarus biscayensis</i> (Collett, 1890)
ANTOSPP	<i>Antonogadus</i> spp.	C	101.19	A	0	<i>Gaidropsarus</i> Rafinesque, 1810
APHIMIN	<i>Aphia minuta</i>	C	162.2.1	A	0	<i>Aphia minuta</i> (Risso, 1810)
APOGIMB	<i>Apogon imberbis</i>	C	127.1.1	A	0	<i>Apogon imberbis</i> (Linnaeus, 1758)
APORPES	<i>Aporrhais pespelecani</i>	F	APOR Apor 1	E	0	<i>Aporrhais pespelecani</i> (Linnaeus, 1758)
APORSER	<i>Aporrhais serresianus</i>	F	APOR Apor 2	D	0	<i>Aporrhais serresianus</i> (Michaud, 1828)
APTECAE	<i>Apterichthys caecus</i>	C	86.2.1	A	0	<i>Apterichtus caecus</i> (Linnaeus, 1758)
ARGESPY	<i>Argentina sphyraena</i>	C	46.1.1	A	0	<i>Argentina sphyraena</i> Linnaeus, 1758
ARGOOLE	<i>Argobuccinum olearium</i>	F	CYM Argo 1	D	0	<i>Ranella olearium</i> (Linnaeus, 1758)
ARGRACU	<i>Argyropelecus aculeatus</i>	C	38.2.2	A	0	<i>Argyropelecus aculeatus</i> Valenciennes, 1850
ARGRHEM	<i>Argyropelecus hemigymnus</i>	C	38.2.1	A	0	<i>Argyropelecus hemigymnus</i> Cocco, 1829
ARGYREG	<i>Argyrosomus regius</i>	C	137.2.1	A	0	<i>Argyrosomus regius</i> (Asso, 1801)
ARIOBAL	<i>Ariosoma balearicum</i>	C	82.2.1	A	0	<i>Ariosoma balearicum</i> (Delaroche, 1809)
ARISFOL	<i>Aristaeomorpha foliacea</i>	F	ARIST Aris 1	B	m	<i>Aristaeomorpha foliacea</i> (Risso, 1827)
ARITANT	<i>Aristeus antennatus</i>	F	ARIST Arist 1	B	m	<i>Aristeus antennatus</i> (Risso, 1816)
ARMIMAC	<i>Arminia maculata</i>	F	NAT Natic 1	D	0	<i>Armina maculata</i> Rafinesque, 1814
ARMITIG	<i>Armina tigrina</i>	R	RIEDL	E	0	<i>Armina tigrina</i> Rafinesque, 1814
ARNOIMP	<i>Arnoglossus imperialis</i>	C	196.2.2	A	0	<i>Arnoglossus imperialis</i> (Rafinesque, 1810)
ARNOLAT	<i>Arnoglossus laterna</i>	C	196.2.1	A	0	<i>Arnoglossus laterna</i> (Walbaum, 1792)
ARNORUP	<i>Arnoglossus rueppelli</i>	C	196.2.4	A	0	<i>Arnoglossus rueppelii</i> (Cocco, 1844)
ARNOTHO	<i>Arnoglossus thori</i>	C	196.2.5	A	0	<i>Arnoglossus thori</i> Kyle, 1913
ASPICUC	<i>Aspitrigla cuculus</i>	C	185.2.1	A	0	<i>Aspitrigla cuculus</i> (Linnaeus, 1758)
ASPIOBS	<i>Aspitrigla obscura</i>	C	185.2.2	A	0	<i>Chelidonichthys obscurus</i> (Bloch & Schneider, 1801)
ASTRSPP	<i>Astropecten</i> spp.			E	0	<i>Astropecten</i> Gray, 1840

MeditS Code	Scientific Name	Source	Reference	CATFAU CODLON	Valid Name (Ifremer reference)
ATELROT	<i>Atelecyclus rotundatus</i>	Z	Z	D 0	<i>Atelecyclus rotundatus</i> (Olivi, 1792)
ATRIFRA	<i>Atrina fragilis</i> (= <i>Pinna pectinata</i>)	F	PINN Attr 4	D 0	<i>Atrina pectinata</i> (Linnaeus, 1767)
AULOFIL	<i>Aulopus filamentosus</i>	C	50.1.1	A 0	<i>Aulopus filamentosus</i> (Bloch, 1792)
BALICAR	<i>Balistes carolinensis</i>	C	201.1.2	A 0	<i>Balistes capriscus</i> Gmelin, 1789
BASOPRO	<i>Bathysolea profundicola</i>	C	198.2.1	A 0	<i>Bathysolea profundicola</i> (Vaillant, 1888)
BATHDUB	<i>Bathypterois dubius</i>	F	CHLOR	A 0	<i>Bathypterois dubius</i> Vaillant, 1888
BATHMED	<i>Bathypterois mediterraneus</i>	C	53.1.2	A 0	<i>Bathypterois dubius</i> Vaillant, 1888
BATISPO	<i>Bathypolypus sponsalis</i>	F	OCT Bath 2	C 0	<i>Bathypolypus sponsalis</i> (Fischer & Fischer, 1892)
BATYMAR	<i>Bathynectes maravigna</i>	F	PORT	B m	<i>Bathynectes maravigna</i> (Prestandrea, 1839)
BATYSUP	<i>Bathynectes superbus</i>	Z	Z	B m	<i>Bathynectes maravigna</i> (Prestandrea, 1839)
BELLAPO	<i>Bellotia apoda</i>	C	172.3.1	A 0	<i>Bellotia apoda</i> Giglioli, 1883
BENGLA	<i>Benthoosema glaciale</i>	C	58.2.1	A 0	<i>Benthoosema glaciale</i> (Reinhardt, 1837)
BENTROB	<i>Benthocometes robustus</i>	C	172.4.1	A 0	<i>Benthocometes robustus</i> (Goode & Bean, 1886)
BERYDEC	<i>Beryx decadactylus</i>	C	112.1.1	A 0	<i>Beryx decadactylus</i> Cuvier, 1829
BERYSPL	<i>Beryx splendens</i>	C	112.1.2	A 0	<i>Beryx splendens</i> Lowe, 1834
BLENBAS	<i>Lipophrys (Blennius) basiliscus</i>	C	164.1.3	A 0	<i>Salaria basiliscus</i> (Valenciennes, 1836)
BLENCRI	<i>Scartella (Blennius) cristata</i> (<i>crinitus</i>)	C	164.1.6	A 0	<i>Scartella cristata</i> (Linnaeus, 1758)
BLENGAT	<i>Parablennius (Blennius)</i> <i>gattorugine</i>	C	164.1.8	A 0	<i>Parablennius gattorugine</i> (Linnaeus, 1758)
BLENOCE	<i>Blennius ocellaris</i>	C	164.1.1	A 0	<i>Blennius ocellaris</i> Linnaeus, 1758
BLENPAV	<i>Lipophrys (Blennius) pavo</i>	C	164.1.12	A 0	<i>Salaria pavo</i> (Risso, 1810)
BLENSPP	Blenniidae	C	164.	A 0	Blenniidae
BLENSPY	<i>Aidablennius (Blennius) sphynx</i>	C	164.1.17	A 0	<i>Aidablennius sphynx</i> (Valenciennes, 1836)
BLENTEN	<i>Parablennius (Blennius)</i> <i>tentaculari</i>	C	164.1.18	A 0	<i>Parablennius tentacularis</i> (Brünnich, 1768)
BOOPBOO	<i>Boops boops</i>	C	139.2.1	A 0	<i>Boops boops</i> (Linnaeus, 1758)
BOROANT	<i>Borostomias antarcticus</i>			C 0	<i>Borostomias antarcticus</i> (Lönnberg, 1905)
BOTHPOD	<i>Bothus podas</i>	C	196.1.1	A 0	<i>Bothus podas</i> (Delaroche, 1809)
BRACRII	<i>Brachioteuthis riisei</i>	F	BRACHIO Bra. 2	C 0	<i>Brachioteuthis riisei</i> (Steenstrup, 1882)
BUCCCOR	<i>Buccinum corneum</i>	F	BUCC Buc 1	D 0	<i>Buccinum corneum</i> (Linnaeus, 1758)
BUCCHUN	<i>Buccinum humphreysianum</i>	F	BUCC	D 0	<i>Buccinum humphreysianum</i> Bennet, 1824
BUCCSPP	<i>Buccinum</i> spp.	F	BUCC	E 0	<i>Buccinum</i> Linnaeus, 1758
BUGLLUT	<i>Buglossidium luteum</i>	C	198.3.1	A 0	<i>Buglossidium luteum</i> (Risso, 1810)
CALAGRA	<i>Calappa granulata</i>	F	CAL Cal 2	B m	<i>Calappa granulata</i> (Linnaeus, 1758)
CALCTUB	<i>Calcinus tubularis</i>	Z	Z	B m	<i>Calcinus tubularis</i> (Linnaeus, 1767)
CALICHI	<i>Calyptrea chinensis</i>		D'Angelo	E 0	<i>Calyptrea chinensis</i> (Linnaeus, 1758)
CALLRIS	<i>Callionymus risso</i>	C	163a.1.7.	A 0	<i>Callionymus risso</i> Lesueur, 1814
CALLRUB	<i>Callanthias ruber</i>	C	124.3.1	A 0	<i>Callanthias ruber</i> (Rafinesque, 1810)
CALMLYR	<i>Callionymus lyra</i>	C	163a.1.1	A 0	<i>Callionymus lyra</i> Linnaeus, 1758
CALMMAC	<i>Callionymus maculatus</i>	C	163a.1.3	A 0	<i>Callionymus maculatus</i> Rafinesque, 1810
CALMPHA	<i>Synchiropus (Callionymus)</i> <i>phaeton</i>	C	163a.1.4	A 0	<i>Synchiropus phaeton</i> (Günther, 1861)
CALMRIS	<i>Callionymus risso</i>	C	163a.1.7	A 0	<i>Callionymus risso</i> Lesueur, 1814
CALMSPP	<i>Callionymus</i>			A 0	<i>Callionymus</i> Linnaeus, 1758
CALOCOR	<i>Calocarides coronatus</i>			B m	<i>Calocarides coronatus</i> (Trybom, 1904)
CALOMAC	<i>Calocaris macandreae</i>	Z	Z	B m	<i>Calocaris macandreae</i> Bell, 1846
CANCCAN	<i>Cancellaria cancellata</i>	F	GASTEROPOD A F14	E 0	<i>Cancellaria cancellata</i> (Linnaeus, 1767)
CANIGRA	<i>Calliostoma granulatum</i>	F	TROCH	D 0	<i>Calliostoma granulatum</i> (Von Born, 1778)
CAPOAPE	<i>Capros aper</i>	C	123.1.1	A 0	<i>Capros aper</i> (Linnaeus, 1758)

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CARAHIP	<i>Caranx hippos</i>	C	131.1.1	A	0	<i>Caranx hippos</i> (Linnaeus, 1766)
CARARHO	<i>Caranx rhonchus</i>	C	131.1.5	A	0	<i>Caranx rhonchus</i> Geoffroy Saint-Hilaire, 1817
CARCPLU	<i>Carcharhinus plumbeus</i>	C	13.1.7	A	0	<i>Carcharhinus plumbeus</i> (Nardo, 1827)
CARCSP	<i>Carcharhinus</i> spp	C	13.1	A	0	<i>Carcharhinus</i> Blainville, 1816
CARDACU	<i>Acanthocardia aculeata</i>	F	CARD Acan 1	E	0	<i>Acanthocardia aculeata</i> (Linnaeus, 1758)
CARDECH	<i>Acanthocardia (Cardium) echinata</i>	F	CARD Acan 2	D	0	<i>Acanthocardia echinata</i> (Linnaeus, 1758)
CARDSPI	<i>Acanthocardia spinosa</i>		D'Angelo	E	0	<i>Acanthocardia spinosa</i> (Solander, 1786)
CARISPP	<i>Cardiomya</i> spp.	R	RIEDL	E	0	<i>Cardiomya</i> Adams A., 1864
CARISTE	<i>Caridion steveni</i>	F	HIPPOL	B	0	<i>Caridion steveni</i> Lebour, 1930
CARPACU	<i>Carapus acus</i>	C	175.1.1	A	0	<i>Carapus acus</i> (Brünnich, 1768)
CASSECH	<i>Cassidaria echinophora</i>	F	CASS Cass 1	D	0	<i>Galeodea echinophora</i> (Linnaeus, 1758)
CASSAB	<i>Phalium (Cassis) saburon</i>	F	CAS Phal 2	D	0	<i>Phalium saburon</i> (Bruguière, 1792)
CASSTYR	<i>Cassidaria tyrrhena</i>	F	CASS Cass 2	D	0	<i>Galeodea rugosa</i> (Linnaeus, 1771)
CATAALL	<i>Cataetix alleni</i>	C	172.6.1	A	0	<i>Cataetix alleni</i> (Byrne, 1906)
CECACIR	<i>Centracanthus cirrus</i>	C	141.1.1	A	0	<i>Centracanthus cirrus</i> Rafinesque, 1810
CENONIG	<i>Centrolophus niger</i>	C	176.1.1	A	0	<i>Centrolophus niger</i> (Gmelin, 1789)
CENTGRA	<i>Centrophorus granulosus</i>	C	16.1.2	A	0	<i>Centrophorus granulosus</i> (Bloch & Schneider, 1801)
CENTUYA	<i>Centrophorus uyato</i>	C	16.2.4	A	0	<i>Centrophorus uyato</i> (Rafinesque, 1810)
CEPHVOL	<i>Dactylopterus (Cephalacanthus) volitans</i>	C	193.1.1	A	0	<i>Dactylopterus volitans</i> (Linnaeus, 1758)
CEPOMAC	<i>Cepola rubescens (macrophthalma)</i>	C	128.1.1	A	0	<i>Cepola macrophthalma</i> (Linnaeus, 1758)
CERAMAD	<i>Ceratoscopelus maderensis</i>	C	58.4.1	A	0	<i>Ceratoscopelus maderensis</i> (Lowe, 1839)
CHAUSLO	<i>Chauliodus sloani</i>	C	40.1.1	A	0	<i>Chauliodus sloani</i> Bloch & Schneider, 1801
CHEOLAB	<i>Chelon labrosus</i>	C	181.2.1	A	0	<i>Chelon labrosus</i> (Risso, 1827)
CHIMMON	<i>Chimaera monstrosa</i>	C	26.1.1	A	0	<i>Chimaera monstrosa</i> Linnaeus, 1758
CHLAOPE	<i>Chlamys opercularis</i>	F	PECT Aeq 1	E	0	<i>Aequipecten opercularis</i> (Linnaeus, 1758)
CHLAVAR	<i>Chlamys varia</i>	F	PECT Chlam 1	E	0	<i>Chlamys varia</i> (Linnaeus, 1758)
CHLOGRA	<i>Chlorotocus crassicornis (gracilipes)</i>	Z	Z	B	m	<i>Chlorotocus crassicornis</i> (A. Costa, 1871)
CHROCHR	<i>Chromis chromis</i>	C	144.1.1	A	0	<i>Chromis chromis</i> (Linnaeus, 1758)
CIRCCAS	<i>Circomphalus casinus</i>	F	VEN	D	0	<i>Venus casina</i> Linnaeus, 1758
CIROBOR	<i>Cirolana borealis</i>			E	0	<i>Cirolana borealis</i> Lilljeborg, 1851
CITHMAC	<i>Citharus linguatula (macrolepidotus)</i>	C	194.1.1	A	0	<i>Citharus linguatula</i> (Linnaeus, 1758)
CLOPBIC	<i>Chlopsis bicolor</i>	C	77.1.1	A	0	<i>Chlopsis bicolor</i> Rafinesque, 1810
CLORAGA	<i>Chlorophthalmus agassizi</i>	C	55.1.1	A	0	<i>Chlorophthalmus agassizi</i> Bonaparte, 1840
COBLGAL	<i>Coryphoblennius galerita</i>	C	164.2.1	A	0	<i>Coryphoblennius galerita</i> (Linnaeus, 1758)
COELCOE	<i>Coelorhynchus coelorhynchus</i>	C	99.12.1	A	0	<i>Caelorinchus caelorhynchus caelorhynchus</i> (Risso, 1810)
COELOCC	<i>Coelorhynchus occa (C. labiatus)</i>	C	99.12.2	A	0	<i>Caelorinchus occa</i> (Goode & Bean, 1885)
CONGCON	<i>Conger conger</i>	C	82.1.1	A	0	<i>Conger conger</i> (Linnaeus, 1758)
CORIJUL	<i>Coris julis</i>	C	145.4.1	A	0	<i>Coris julis</i> (Linnaeus, 1758)
CORYGUN	<i>Coryphaenoides guentheri</i>	C	99.13.2	A	0	<i>Coryphaenoides guentheri</i> (Vaillant, 1888)
CRANSPP	<i>Crangon</i> sp	F	CRANG	B	m	<i>Crangon</i> J.C. Fabricius, 1798
CRASGIG	<i>Crassostrea gigas</i>	F	OSTR Crass 1	E	0	<i>Crassostrea gigas</i> (Thunberg, 1793)
CRASSPP	<i>Crassostrea</i> spp.	F	OSTR	E	0	<i>Crassostrea</i> Sacco, 1897
CUBIGRA	<i>Cubiceps gracilis</i>	C	177.2.1	A	0	<i>Cubiceps gracilis</i> (Lowe, 1843)
CUSPCUS	<i>Cuspidaria cuspidata</i>			E	0	<i>Cuspidaria cuspidata</i> (Olivi, 1792)
CYCLPIG	<i>Cyclothone pygmaea</i>	C	37.4.8	A	0	<i>Cyclothone pygmaea</i> Jespersen & Tåning, 1926
CYCLSPP	<i>Cyclothone</i> spp.	C	37.4	A	0	<i>Cyclothone</i> Goode & Bean, 1882

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CYMACOR	<i>Cymatium corrogatum</i>	F	CYM Cym 1	D 0	<i>Cymatium corrogatum</i> (Lamarck, 1816)
CYMBOLL	<i>Cymbium olla</i>			E 0	<i>Cymbium olla</i> (Linnaeus, 1758)
CYNPFER	<i>Cynoponticus ferox</i>	C	79.1.1	A 0	<i>Cynoponticus ferox</i> Costa, 1846
DALOIMB	<i>Dalophis imberbis</i>	C	86.3.1	A 0	<i>Dalophis imberbis</i> (Delaroche, 1809)
DARDARR	<i>Dardanus arrosor</i>	Z	Z	B m	<i>Dardanus arrosor</i> (Herbst, 1796)
DARDCAL	<i>Dardanus calidus</i>	Z	Z	B m	<i>Dardanus calidus</i> (Risso, 1827)
DASICEN	<i>Dasyatis centroura</i>	C	22.1.2	A 0	<i>Dasyatis centroura</i> (Mitchill, 1815)
DASIPAS	<i>Dasyatis pastinaca</i>	C	22.1.1	A 0	<i>Dasyatis pastinaca</i> (Linnaeus, 1758)
DASITOR	<i>Dasyatis tortonesi</i>	C	22.1.4	A 0	<i>Dasyatis pastinaca</i> (Linnaeus, 1758)
DASIVIO	<i>Dasyatis violacea</i>	C	22.1.3	A 0	<i>Pteroplatytrygon violacea</i> (Bonaparte, 1832)
DENTDEN	<i>Dentex dentex</i>	C	139.3.1	A 0	<i>Dentex dentex</i> (Linnaeus, 1758)
DENTGIB	<i>Dentex gibbosus</i>	C	139.3.3	A 0	<i>Dentex gibbosus</i> (Rafinesque, 1810)
DENTMAC	<i>Dentex macrophthalmus</i>	C	139.3.4	A 0	<i>Dentex macrophthalmus</i> (Bloch, 1791)
DENTMAR	<i>Dentex maroccanus</i>	C	139.3.5	A 0	<i>Dentex maroccanus</i> Valenciennes, 1830
DENTSPP	<i>Dentalium spp</i>			E 0	<i>Dentalium</i> Linnaeus, 1758
DIAPHOL	<i>Diaphus holti</i>	C	58.6.5.	A 0	<i>Diaphus holti</i> Tåning, 1918
DIAPMET	<i>Diaphus metopoclampus</i>	C	58.6.7	A 0	<i>Diaphus metopoclampus</i> (Cocco, 1829)
DIAPRAF	<i>Diaphus rafinesquei</i>	C	58.6.9	A 0	<i>Diaphus rafinesquii</i> (Cocco, 1838)
DIAPSP	<i>Diaphus spp.</i>	C	58.6.	A 0	<i>Diaphus</i> Eigenmann & Eigenmann, 1890
DICAMAY	<i>Dicranodromia mayheuxi</i>	Z	Z	B m	<i>Dicranodromia mayheuxii</i> A. Milne-Edwards, 1883
DICELAB	<i>Dicentrarchus labrax</i>	C	124.4.1	A 0	<i>Dicentrarchus labrax</i> (Linnaeus, 1758)
DICEPUN	<i>Dicentrarchus punctatus</i>	C	124.4.2	A 0	<i>Dicentrarchus punctatus</i> (Bloch, 1792)
DICOCUN	<i>Dicologlossa cuneata</i>	C	198.4.2	A 0	<i>Dicologlossa cuneata</i> (Moreau, 1881)
DIODITA	<i>Diodora italica</i>			E 0	<i>Diodora italica</i> (Defrance, 1820)
DIPGBIM	<i>Diplacogaster bimaculata</i>	C	208.2.1	A 0	<i>Diplecogaster bimaculata bimaculata</i> (Bonnaterre, 1788)
DIPLANN	<i>Diplodus annularis</i>	C	139.4.1	A 0	<i>Diplodus annularis</i> (Linnaeus, 1758)
DIPLCER	<i>Diplodus cervinus cervinus</i>	C	139.4.2.	A 0	<i>Diplodus cervinus cervinus</i> (Lowe, 1838)
DIPLPUN	<i>Diplodus puntazo</i>	C	139.8.1	A 0	<i>Diplodus puntazzo</i> (Cetti, 1777)
DIPLSAR	<i>Diplodus sargus</i>	C	139.4.3	A 0	<i>Diplodus sargus sargus</i> (Linnaeus, 1758)
DIPLVUL	<i>Diplodus vulgaris</i>	C	139.4.4	A 0	<i>Diplodus vulgaris</i> (Geoffroy Saint-Hilaire, 1817)
DORHTHO	<i>Dorhynchus thomsoni</i>	Z	Z	B m	<i>Dorhynchus thomsoni</i> Wyville & Thomson, 1873
DORILAN	<i>Dorippe lanata</i>	Z	Z	B m	<i>Medorippe lanata</i> (Linnaeus, 1767)
DORITHO	<i>Dorhynchus thomsoni</i>	Z	Z	C m	<i>Dorhynchus thomsoni</i> Wyville & Thomson, 1873
DORSVER	<i>Doris verrucosa</i>	R	RIEDL	E 0	<i>Doris verrucosa</i> Linnaeus, 1758
DOSISPP	<i>Dosinia spp</i>			D 0	<i>Dosinia Scopoli</i> , 1777
DROMPER	<i>Dromia personata</i>	F	DROM Drom 1	B m	<i>Dromia personata</i> (Linnaeus, 1758)
DUSSELO	<i>Dussumieria elopsoides</i>	X	X	A	<i>Dussumieria elopsoides</i> Bleeker, 1849
EBALCRA	<i>Ebalia cranchi</i>	Z	Z	B 0	<i>Ebalia cranchi</i> Leach, 1817
EBALNUX	<i>Ebalia nux</i>	Z	Z	B m	<i>Ebalia nux</i> A. Milne-Edwards, 1883
EHEMIR	<i>Echelus myrus</i>	C	84.1.1	A 0	<i>Echelus myrus</i> (Linnaeus, 1758)
ECHIDEN	<i>Echiodon dentatus</i>	C	175.2.2	A 0	<i>Echiodon dentatus</i> (Cuvier, 1829)
ELECRIS	<i>Electrona rissoi</i>	C	58.8.1	A 0	<i>Electrona risso</i> (Cocco, 1829)
ELEDCIR	<i>Eledone cirrhosa</i>	F	OCT Eled 1	C 0	<i>Eledone cirrhosa</i> (Lamarck, 1798)
ELEDMOS	<i>Eledone moschata</i>	F	OCT Eled 2	C 0	<i>Eledone moschata</i> (Lamarck, 1798)
ELEDSPP	<i>Eledone spp</i>	F	OCT	C 0	<i>Eledone</i> Leach, 1817
ENGRENC	<i>Engraulis encrasicolus</i>	C	35.1.1	A 0	<i>Engraulis encrasicolus</i> (Linnaeus, 1758)
EPHIGUT	<i>Ephippion guttiferum</i>	C	204.1.1	A 0	<i>Ephippion guttifer</i> (Bennett, 1831)
EPIGCON	<i>Epigonus constanciae</i>	C	127.2.3	A 0	<i>Epigonus constanciae</i> (Giglioli, 1880)
EPIGDEN	<i>Epigonus denticulatus</i>	C	127.2.2	A 0	<i>Epigonus denticulatus</i> Dieuzeide, 1950
EPIGTEL	<i>Epigonus telescopus</i>	C	127.2.1	A 0	<i>Epigonus telescopus</i> (Risso, 1810)

Medits Code	Scientific Name	Source	Reference	CATFAU	CODLON	Valid Name (Ifremer reference)
EPINAEN	<i>Epinephelus aeneus</i>	C	124.5.1	A	0	<i>Epinephelus aeneus</i> (Geoffroy Saint-Hilaire, 1817)
EPINALE	<i>Epinephelus alexandrinus</i>	C	124.5.2	A	0	<i>Epinephelus costae</i> (Steindachner, 1878)
EPINCAN	<i>Epinephelus caninus</i>	C	124.5.3	A	0	<i>Epinephelus caninus</i> (Valenciennes, 1843)
EPINGUA	<i>Epinephelus guaza</i>	C	124.5.4	A	0	<i>Epinephelus marginatus</i> (Lowe, 1834)
EPINSPP	<i>Epinephelus</i> spp	C	124.5	A	0	<i>Epinephelus</i> Bloch, 1793
ERETKLE	<i>Eretmophorus kleinenbergi</i>	C	103.1.1	A	0	<i>Eretmophorus kleinenbergi</i> Giglioli, 1889
ERGACLO	<i>Ergasticus clouei</i>	Z	Z	B	m	<i>Ergasticus clouei</i> A. Milne-Edwards, 1882
ETHUMAS	<i>Ethusa mascarone</i>	Z	Z	B	m	<i>Ethusa mascarone</i> (Herbst, 1785)
ETMOSPI	<i>Etmopterus spinax</i>	C	16.6.1	A	0	<i>Etmopterus spinax</i> (Linnaeus, 1758)
EUCHLIG	<i>Euchirograpsus liguricus</i>	Z	Z	B	m	<i>Euchirograpsus liguricus</i> H. Milne-Edwards, 1853
EUPHSPP	Euphausiidae			B	m	Euphausiidae
EURYASP	<i>Eurynome aspera</i>	Z	Z	B	m	<i>Eurynome aspera</i> (Pennant, 1777)
EUTRGUR	<i>Eutrigla gurnardus</i>	C	185.3.1	A	0	<i>Eutrigla gurnardus</i> (Linnaeus, 1758)
EVERBAL	<i>Evermannella balboi</i> (= <i>balbo</i>)	C	60.1.1	A	0	<i>Evermannella balbo</i> (Risso, 1820)
FLEXFLE	<i>Flexopecten flexuosus</i>	F	PETC Flex	E	0	<i>Chlamys flexuosa</i> (Poli, 1795)
FUNCWOO	<i>Funchalia woodwardi</i>	F	PEN	B	m	<i>Funchalia woodwardi</i> Johnson, 1867
FUSIROS	<i>Fusinus rostratus</i>	F	FASC Fus 1	E	0	<i>Fusinus rostratus</i> (Olivi, 1792)
FUSTUND	<i>Fusituris undatiruga</i>			E	0	<i>Fusituris undatiruga</i> (Bivona, 1832)
GADAMAR	<i>Gadella maraldi</i>	C	103.3.1	A	0	<i>Gadella maraldi</i> (Risso, 1810)
GADIARG	<i>Gadiculus argenteus</i>	C	101.5.1	A	0	<i>Gadiculus argenteus argenteus</i> Guichenot, 1850
GADUMER	<i>Merlangius merlangus</i>	C	101.7.1	A	0	<i>Merlangius merlangus</i> (Linnaeus, 1758)
GAIDMED	<i>Gaidropsarus mediterraneus</i>	C	101.20.1	A	0	<i>Gaidropsarus mediterraneus</i> (Linnaeus, 1758)
GAIDVUL	<i>Gaidropsarus vulgaris</i>	C	101.20.4	A	0	<i>Gaidropsarus vulgaris</i> (Cloquet, 1824)
GALADIS	<i>Galathea dispersa</i>	Z	Z	B	m	<i>Galathea dispersa</i> Bate, 1859
GALAINTE	<i>Galathea intermedia</i>	Z	Z	B	m	<i>Galathea intermedia</i> Liljeborg, 1851
GALANEX	<i>Galathea nexa</i>	Z	Z	B	m	<i>Galathea nexa</i> Embleton, 1834
GALEGAL	<i>Galeorhinus galeus</i>	C	13.3.1	A	0	<i>Galeorhinus galeus</i> (Linnaeus, 1758)
GALIDEC	<i>Galeoides decadactylus</i>	C	182.1.1	A	0	<i>Galeoides decadactylus</i> (Bloch, 1795)
GALUATL	<i>Galeus atlanticus</i>	F	SCYL Gal 11	A	0	<i>Galeus atlanticus</i> (Vaillant, 1888)
GALUMEL	<i>Galeus melastomus</i>	C	11.3.1	A	0	<i>Galeus melastomus</i> Rafinesque, 1810
GENNELE	<i>Gennadas elegans</i>	F	ARIST	B	m	<i>Gennadas elegans</i> S.I. Smith, 1882
GEPYDAR	<i>Gephyroberyx darwini</i>	C	115.1.1	A	0	<i>Gephyroberyx darwini</i> (Johnson, 1866)
GERYLON	<i>Geryon longipes</i>	F	GER Ger 2	B	m	<i>Geryon longipes</i> A. Milne-Edwards, 1882
GIBBSPP	<i>Gibbula</i> sp		D'Angelo		0	<i>Gibbula</i> Risso, 1826
GLOSLEI	<i>Glossanodon leioglossus</i>	C	46.2.1	A	0	<i>Glossanodon leioglossus</i> (Valenciennes, 1848)
GLOSVAL	<i>Glossodoris valenciennesi</i>	R	RIEDL	E	0	<i>Hypselodoris picta</i> (Schultz)
GLOUHUM	<i>Glossus humanus</i>	F	GLOSS Gloss 1	E	0	<i>Glossus humanus</i> (Linnaeus, 1758)
GNATMYS	<i>Gnathophis mystax</i>	C	82.3.1	A	0	<i>Gnathophis mystax</i> (Delaroche, 1809)
GOBICOL	<i>Deltentosteus</i> (<i>Gobius</i>) <i>colonialis</i>	C	162.10.2	A	0	<i>Deltentosteus collonianus</i> (Risso, 1820)
GOBIFRI	<i>Lesueurigobius</i> (<i>Gobius</i>) <i>friesii</i>	C	162.16.2	A	0	<i>Lesueurigobius friesii</i> (Malm, 1874)
GOBIGEN	<i>Gobius geniporus</i>	C	162.1.8	A	0	<i>Gobius geniporus</i> Valenciennes, 1837
GOBILIN	<i>Crystallogobius</i> (<i>Gobius</i>) <i>linearis</i>	C	162.9.1	A	0	<i>Crystallogobius linearis</i> (Düben, 1845)
GOBINIG	<i>Gobius niger</i>	C	162.1.1	A	0	<i>Gobius niger</i> Linnaeus, 1758
GOBIQUA	<i>Deltentosteus</i> (<i>Gobius</i>) <i>quadrifasciatus</i>	C	162.10.1	A	0	<i>Deltentosteus quadrifasciatus</i> (Valenciennes, 1837)
GOBISAN	<i>Lesueurigobius</i> (<i>Gobius</i>) <i>sanzoi</i>	C	162.16.4	A	0	<i>Lesueurigobius sanzoi</i> (De Buen, 1918)
GOBISPP	<i>Gobius</i> spp	C	162	A	0	<i>Gobius</i> Linnaeus, 1758
GOBISUE	<i>Lesueurigobius suerii</i>	C	162.16.1	A	0	<i>Lesueurigobius suerii</i> (Risso, 1810)
GONERHO	<i>Goneplax rhomboides</i> (= <i>angulata</i>)	Z	Z	B	m	<i>Goneplax rhomboides</i> (Linnaeus, 1758)

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GONICOC	<i>Gonichthys coccoi</i>	C	58.9.1	A 0	<i>Gonichthys cocco</i> (Cocco, 1829)
GONODEN	<i>Gonostoma denudatum</i>	C	37.1.1	A 0	<i>Gonostoma denudatum</i> Rafinesque, 1810
GYMACIC	<i>Gymnammodytes cicereus</i>	C	147.2.1	A 0	<i>Gymnammodytes cicereus</i> (Rafinesque, 1810)
GYMNALT	<i>Gymnura altavela</i>	C	22.2.1	A 0	<i>Gymnura altavela</i> (Linnaeus, 1758)
HADRCRA	<i>Hadriana craticuloides</i>	F	MUR	D 0	<i>Hadriana craticuloides</i> (Vokes, 1964)
HELIDAC	<i>Helicolenus dactylopterus</i>	C	184.2.1	A 0	<i>Helicolenus dactylopterus dactylopterus</i> (Delaroche, 1809)
HEPTPER	<i>Heptranchias perlo</i>	C	3.2.1	A 0	<i>Heptranchias perlo</i> (Bonnaterre, 1788)
HETEDIS	<i>Heteroteuthis dispar</i>	F	SEPIOL	C 0	<i>Heteroteuthis dispar</i> (Rüppell, 1845)
HEXAGRI	<i>Hexanchus griseus</i>	C	3.1.1	A 0	<i>Hexanchus griseus</i> (Bonnaterre, 1788)
HEXAVIT	<i>Hexanchus nakamurai</i> (vitulus)	C	3.1.2	A 0	<i>Hexanchus nakamurai</i> Teng, 1962
HINIINC	<i>Hinia incrassata</i>	F	NASS Hin	E 0	<i>Nassarius incrassatus</i> (Ström, 1768)
HINIRET	<i>Hinia reticulata</i>	F	NASS Hin 1	E 0	<i>Nassarius reticulatus</i> (Linnaeus, 1758)
HIPPHIC	<i>Hippocampus hippocampus</i>	C	97.4.1	A 0	<i>Hippocampus hippocampus</i> (Linnaeus, 1758)
HISTBON	<i>Histioteuthis bonnellii</i>	F	HISTIO	C 0	<i>Histioteuthis bonnellii</i> (De Férussac, 1835)
HISTREV	<i>Histioteuthis reversa</i>	F	HISTIO	C 0	<i>Histioteuthis reversus</i> (Verrill, 1880)
HISTSPP	<i>Histioteuthis</i> spp	F	HISTIO	C 0	<i>Histioteuthis</i> D'Orbigny, 1848
HOMAVUL	<i>Homarus vulgaris</i>	F	NEPH Hom 1	B m	<i>Homarus gammarus</i> (Linnaeus, 1758)
HOMOBAR	<i>Homola barbata</i>	Z	Z	B m	<i>Homola barbata</i> (J.C. Fabricius, 1793)
HOPLATL	<i>Hoplostethus atlanticus</i>	C	115.2.2	A 0	<i>Hoplostethus atlanticus</i> Collett, 1889
HOPLMED	<i>Hoplostethus mediterraneus</i>	C	115.2.1	A 0	<i>Hoplostethus mediterraneus mediterraneus</i> Cuvier, 1829
HYGOBEN	<i>Hygophum benoiti</i>	C	58.10.2	A 0	<i>Hygophum benoiti</i> (Cocco, 1838)
HYGOHIG	<i>Hygophum hygomii</i>	C	58.10.1	A 0	<i>Hygophum hygomii</i> (Lütken, 1892)
HYMEITA	<i>Hymenocephalus italicus</i>	C	99.5.1	A 0	<i>Hymenocephalus italicus</i> Giglioli, 1884
HYMPSPP	<i>Hymenopenaeus</i> sp	Z	Z	B m	<i>Hymenopenaeus</i> Smith, 1882
HYPESPP	Hyperiidæ			E 0	Hyperiidæ
HYPOPIC	<i>Hyporhamphus picarti</i>	C	93.2.1	A 0	<i>Hyporhamphus picarti</i> (Valenciennes, 1847)
ICHTOVA	<i>Ichthyococcus ovatus</i>	C	37.6.1	A 0	<i>Ichthyococcus ovatus</i> (Cocco, 1838)
ILLECOI	<i>Illex coindetii</i>	F	OMMAS III 1	C 0	<i>Illex coindetii</i> (Verany, 1839)
ILLESPP	<i>Illex</i>	F		C 0	<i>Illex</i> Steenstrup, 1880
INACCOM	<i>Inachus communissimus</i>	Z	Z	B m	<i>Inachus communissimus</i> Rizza, 1839
INACDOR	<i>Inachus dorsettensis</i>	Z	Z	B m	<i>Inachus dorsettensis</i> (Pennant, 1777)
INACTHO	<i>Inachus thoracicus</i>	Z	Z	B m	<i>Inachus thoracicus</i> P. Roux, 1830
JAXENOC	<i>Jaxea nocturna</i>			B m	<i>Jaxea nocturna</i> Nardo, 1847
LABRVIR	<i>Labrus viridis</i>	C	145.1.4	A 0	<i>Labrus viridis</i> Linnaeus, 1758
LABSBIM	<i>Labrus bimaculatus</i>	C	145.1.1	A 0	<i>Labrus mixtus</i> Linnaeus, 1758
LAEVCAR	<i>Laevicardium oblongum</i>	F	CARD Laev 1	E 0	<i>Laevicardium oblongum</i> (Gmelin, 1791)
LAGOLAG	<i>Lagocephalus lagocephalus</i>	C	204.2.1	A 0	<i>Lagocephalus lagocephalus lagocephalus</i> (Linnaeus, 1758)
LAMACRO	<i>Lampanyctus crocodilus</i>	C	58.12.1	A 0	<i>Lampanyctus crocodilus</i> (Risso, 1810)
LAMAPUS	<i>Lampanyctus pusillus</i>	C	58.12.10	A 0	<i>Lampanyctus pusillus</i> (Johnson, 1890)
LAMASPP	<i>Lampanyctus</i> spp	C	58.12	A 0	<i>Lampanyctus</i> Bonaparte, 1840
LAMPGUT	<i>Lampris guttatus</i>	C	105.1.1	A 0	<i>Lampris guttatus</i> (Brünnich, 1788)
LAPPFAS	<i>Lappanella fasciata</i>	C	145.7.1	A 0	<i>Lappanella fasciata</i> (Cocco, 1833)
LATRSP					<i>Latreillia</i> P. Roux, 1830
LEPALEP	<i>Lepadogaster lepadogaster</i>	C	208.4.1	A 0	<i>Lepadogaster lepadogaster lepadogaster</i> (Bonnaterre, 1788)
LEPICAU	<i>Lepidopus caudatus</i>	C	155.4.1	A 0	<i>Lepidopus caudatus</i> (Euphrasen, 1788)
LEPMBOS	<i>Lepidorhombus boscii</i>	C	195.2.2	A 0	<i>Lepidorhombus boscii</i> (Risso, 1810)
LEPMWHS	<i>Lepidorhombus whiffiagonis</i>	C	195.2.1	A 0	<i>Lepidorhombus whiffiagonis</i> (Walbaum, 1792)
LEPOLEP	<i>Lepidion lepidion</i>	C	103.6.1	A 0	<i>Lepidion lepidion</i> (Risso, 1810)

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LEPTCAV	<i>Lepidotrigla cavillone</i>	C	185.4.1	A	0	<i>Lepidotrigla cavillone</i> (Lacepède, 1801)
LEPTDIE	<i>Lepidotrigla dieuzeidei</i>	C	185.4.2	A	0	<i>Lepidotrigla dieuzeidei</i> Blanc & Hureau, 1973
LESTSPD	<i>Lestidiops sphyrenoides</i>	C	63.2.1	A	0	<i>Lestidiops sphyrenoides</i> (Risso, 1820)
LESTSPP	<i>Lestidiops</i> spp.	C	63.2	A	0	<i>Lestidiops</i> Hubbs, 1916
LICHAMI	<i>Lichia amia</i>	C	131.5.1	A	0	<i>Lichia amia</i> (Linnaeus, 1758)
LIGUENS	<i>Ligur ensiferus</i>	Z	Z	B	m	<i>Ligur ensiferus</i> (Risso, 1816)
LISSCHI	<i>Lissa chinagra</i>	Z	Z	B	m	<i>Lissa chinagra</i> (J.C. Fabricius, 1775)
LITHMOR	<i>Lithognathus mormyrus</i>	C	139.5.1	A	0	<i>Lithognathus mormyrus</i> (Linnaeus, 1758)
LIZAAUR	<i>Liza aurata</i>	C	181.3.2	A	0	<i>Liza aurata</i> (Risso, 1810)
LIZARAM	<i>Liza ramada</i>	C	181.3.1	A	0	<i>Liza ramado</i> (Risso, 1810)
LIZASAL	<i>Liza saliens</i>	C	181.3.4	A	0	<i>Liza saliens</i> (Risso, 1810)
LOBIDOF	<i>Lobianchia dofleini</i>	C	58.14.12	A	0	<i>Lobianchia dofleini</i> (Zugmayer, 1911)
LOBIGEM	<i>Lobianchia gemellarii</i>	C	58.14.1	A	0	<i>Lobianchia gemellarii</i> (Cocco, 1838)
LOLIFOR	<i>Loligo forbesi</i>	F	LOLIG Lolig 2	C	0	<i>Loligo forbesi</i> Steenstrup, 1856
LOLISPP	<i>Loligo</i>	F		C	0	<i>Loligo</i> Lamarck, 1798
LOLIVUL	<i>Loligo vulgaris</i>	F	LOLIG Lori 1	C	0	<i>Loligo vulgaris</i> Lamarck, 1798
LOPHBUD	<i>Lophius budegassa</i>	C	210.1.2	A	0	<i>Lophius budegassa</i> Spinola, 1807
LOPHPIS	<i>Lophius piscatorius</i>	C	210.1.1	A	0	<i>Lophius piscatorius</i> Linnaeus, 1758
LOPHSPP	<i>Lophius</i>	C		A	0	<i>Lophius</i> Linnaeus, 1758
LOPOTYP	<i>Lophogaster typicus</i>			E	0	<i>Lophogaster typicus</i>
LUNACAT	<i>Lunatia catena</i>	F	NAT	D	0	<i>Euspira catena</i> (da Costa, 1778)
LUNAFUS	<i>Lunatia fusca</i>		D'Onghia	E	0	<i>Euspira fusca</i> (De Blainville, 1825)
LUTRSPP	<i>Lutraria</i> spp.	R	RIEDL	E	0	<i>Lutraria</i> Lamarck, 1799
MACOSCO	<i>Macrorhamphosus scolopax</i>	C	96.1.1	A	0	<i>Macroramphosus scolopax</i> (Linnaeus, 1758)
MACRLIN	<i>Macropodia linaresi</i>	Z	Z	B	m	<i>Macropodia linaresi</i> Forest & Zariquiey-Alvarez, 1964
MACRLON	<i>Macropodia longipes</i>	Z	Z	B	m	<i>Macropodia tenuirostris</i> (Leach, 1814)
MACRROS	<i>Macropodia rostrata</i>	F	MAJI	B	m	<i>Macropodia rostrata</i> (Linnaeus, 1761)
MAJACRI	<i>Maja crispata</i>	F	MAJI Maja	B	m	<i>Maja crispata</i> Risso, 1827
MAJASQU	<i>Maja squinado</i>	F	MAJI Maja 1	B	m	<i>Maja squinado</i> (Herbst, 1788)
MAURMUE	<i>Maurolicus muelleri</i>	C	37.8.1	A	0	<i>Maurolicus muelleri</i> (Gmelin, 1789)
MCPIARC	<i>Liocarcinus arcuatus</i>	F	PORT Lioc 3	B	m	<i>Liocarcinus navigator</i> (Herbst, 1794)
MCPICOR	<i>Liocarcinus corrugatus</i>		Zariquiey	B	m	<i>Liocarcinus corrugatus</i> (Pennant, 1777)
MCPIDEP	<i>Liocarcinus (Macropipus) depurator</i>	F	PORT Lioc 4	B	m	<i>Liocarcinus depurator</i> (Linnaeus, 1758)
MCPIMAC	<i>Liocarcinus maculatus</i>	F	PORT Lioc	B	m	<i>Liocarcinus maculatus</i> (Risso, 1827)
MCPIPUB	<i>Necora (Macropipus) puber</i>	F	PORT Neco 1	B	m	<i>Necora puber</i> (Linnaeus, 1767)
MCPITUB	<i>Macropipus tuberculatus</i>	F	PORT Macro 1	B	m	<i>Macropipus tuberculatus</i> (P. Roux, 1830)
MEGANOR	<i>Meganyctiphanes norvegica</i>			B	m	<i>Meganyctiphanes norvegica</i>
MELAATL	<i>Melanostigma atlanticum</i>	C	170.6.1	A	0	<i>Melanostigma atlanticum</i> Koefoed, 1952
MERLMER	<i>Merluccius merluccius</i>	C	100.1.1	A	0	<i>Merluccius merluccius</i> (Linnaeus, 1758)
MICMPOU	<i>Micromesistius poutassou</i>	C	101.8.1	A	0	<i>Micromesistius poutassou</i> (Risso, 1827)
MICOSAB	<i>Microcosmus sabatieri</i>	F	PYUR Micr 2	D	0	<i>Microcosmus sabatieri</i> Roule, 1885
MICRMCS	<i>Microstoma microstoma</i>	C	46.1.3	A	0	<i>Microstoma microstoma</i> (Risso, 1810)
MICUAZE	<i>Microchirus azevia</i>	C	198.5.2	A	0	<i>Microchirus theophila</i> (Risso, 1810)
MICUBOS	<i>Microchirus boscanion</i>			A	0	<i>Microchirus boscanion</i> (Chabanaud, 1926)
MICUOCE	<i>Microchirus ocellatus</i>	C	198.5.3	A	0	<i>Microchirus ocellatus</i> (Linnaeus, 1758)
MICUVAR	<i>Microchirus variegatus</i>	C	198.5.1	A	0	<i>Microchirus variegatus</i> (Donovan, 1808)
MOLAMOL	<i>Mola mola</i>	C	207.1.1	A	0	<i>Mola mola</i> (Linnaeus, 1758)
MOLVDYP	<i>Molva dipterygia</i>	C	101.14.2	A	0	<i>Molva dipterygia</i> (Pennant, 1784)
MOLVMOL	<i>Molva molva</i>	C	101.14.1	A	0	<i>Molva molva</i> (Linnaeus, 1758)
MONOHIS	<i>Monochirus hispidus</i>	C	198.6.1	A	0	<i>Monochirus hispidus</i> Rafinesque, 1814

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MORAMOR	<i>Mora moro</i>	C	103.7.1	A 0	<i>Mora moro</i> (Risso, 1810)
MORIRUG	<i>Morio rugosa</i>			E 0	<i>Galeodea rugosa</i> (Linnaeus, 1771)
MUGICEP	<i>Mugil cephalus</i>	C	181.1.1	A 0	<i>Mugil cephalus</i> Linnaeus, 1758
MUGISPP	Mugilidae	C	181.	A 0	Mugilidae
MULLBAR	<i>Mullus barbatus</i>	C	138.1.1	A 0	<i>Mullus barbatus</i> Linnaeus, 1758
MULLSUR	<i>Mullus surmuletus</i>	C	138.1.2	A 0	<i>Mullus surmuletus</i> Linnaeus, 1758
MUNICUR	<i>Munida curvimana</i>	Z	Z	B m	<i>Munida curvimana</i> A. Milne-Edwards & Bouvier, 1894
MUNIINT	<i>Munida intermedia</i>	Z	Z	B m	<i>Munida intermedia</i> A. Milne-Edwards et Bouvier, 1899
MUNIIRI	<i>Munida iris</i>	Z	Z	B m	<i>Munida rutlanti</i> Zariquiey-Alvarez, 1952
MUNIPER	<i>Munida perarmata</i> (= <i>tenuimana</i>)	Z	Z	B m	<i>Munida perarmata</i> A. Milne-Edwards & Bouvier, 1894
MUNIRUG	<i>Munida rugosa</i>	Z	Z	B m	<i>Munida rugosa</i> (J.C. Fabricius, 1775)
MUNISPP	<i>Munida</i>	Z	Z	B m	<i>Munida</i> Leach, 1820
MUNITEN	<i>Munida tenuimana</i>	Z	Z	B m	<i>Munida perarmata</i> A. Milne-Edwards & Bouvier, 1894
MURAHEL	<i>Muraena helena</i>	C	73.1.1	A 0	<i>Muraena helena</i> Linnaeus, 1758
MUREBRA	<i>Bolinus (Murex) brandaris</i>	F	MUR Bol 1	D 0	<i>Bolinus brandaris</i> (Linnaeus, 1758)
MURETRU	<i>Murex trunculus</i>	R	RIEDL	D 0	<i>Hexaplex trunculus</i> (Linnaeus, 1758)
MUSTAST	<i>Mustelus asterias</i>	C	13c.5.2	A 0	<i>Mustelus asterias</i> Cloquet, 1821
MUSTMED	<i>Mustelus mediterraneus</i>	C	13c.5.3	A 0	<i>Mustelus punctulatus</i> Risso, 1827
MUSTMUS	<i>Mustelus mustelus</i>	C	13c.5.1	A 0	<i>Mustelus mustelus</i> (Linnaeus, 1758)
MYCOPUN	<i>Myctophum punctatum</i>	C	58.1.1	A 0	<i>Myctophum punctatum</i> Rafinesque, 1810
MYCOSPP	Myctophidae	C	58	A 0	Myctophidae
MYCTRUB	<i>Mycteroperca rubra</i>	C	124.6.1	A 0	<i>Mycteroperca rubra</i> (Bloch, 1793)
MYLIAQU	<i>Myliobatis aquila</i>	C	23.1.1	A 0	<i>Myliobatis aquila</i> (Linnaeus, 1758)
MYTIGAL	<i>Mytilus galloprovincialis</i>	F	MYTIL Mytil 1	D 0	<i>Mytilus galloprovincialis</i> Lamarck, 1819
MYTISPP	Mytilidae spp.	F	MYTIL	D 0	Mytilidae Rafinesque, 1815
NANSOBI	<i>Nansenia oblita</i>	C	46.4.2.	A 0	<i>Nansenia oblita</i> (Facciola, 1887)
NASSSPP	Nassariidae			D 0	Nassariidae Iredale, 1916
NATIMIL	<i>Naticarius millepunctatus</i>		D'Angelo	E 0	<i>Natica stercusmuscarum</i> (Gmelin, 1791)
NATISPP	Naticidae	F	NAT	D 0	Naticidae Guilding, 1834
NAUCDUC	<i>Naucrates ductor</i>	C	131.6.1	A 0	<i>Naucrates ductor</i> (Linnaeus, 1758)
NEMISCO	<i>Nemichthys scolopaceus</i>	C	76.1.1	A 0	<i>Nemichthys scolopaceus</i> Richardson, 1848
NEORCAR	<i>Neorossia caroli</i>	F	SEPIOL	C 0	<i>Neorossia caroli</i> (Joubin, 1902)
NEPRNOR	<i>Nephrops norvegicus</i>	F	NEPH Neph 1	B m	<i>Nephrops norvegicus</i> (Linnaeus, 1758)
NEROMAC	<i>Nerophis maculatus</i>	C	97.2.1	A 0	<i>Nerophis maculatus</i> Rafinesque, 1810
NEROOPH	<i>Nerophis ophidion</i>	C	97.2.2	A 0	<i>Nerophis ophidion</i> (Linnaeus, 1758)
NETOBRE	<i>Dysomma (Nettodarus) brevirostris</i>	C	81.1.1	A 0	<i>Dysomma brevirostre</i> (Facciola, 1887)
NETMEL	<i>Nettastoma melanurum</i>	C	80.1.1	A 0	<i>Nettastoma melanurum</i> Rafinesque, 1810
NEZUAEQ	<i>Nezumia aequalis</i>	C	99.9.1	A 0	<i>Nezumia aequalis</i> (Günther, 1878)
NEZUSCL	<i>Nezumia sclerorhynchus</i>	C	99.9.2	A 0	<i>Nezumia sclerorhynchus</i> (Valenciennes, 1838)
NOTABON	<i>Notacanthus bonapartei</i>	C	89.1.2	A 0	<i>Notacanthus bonaparte</i> Risso, 1840
NOTORIS	<i>Notolepis rissoi</i>	C	63.4.1	A 0	<i>Arctozenus risso</i> (Bonaparte, 1840)
NOTSBOL	<i>Notoscopelus bolini</i>	C	58.17.5	A 0	<i>Notoscopelus bolini</i> Nafpaktitis, 1975
NOTSELO	<i>Notoscopelus elongatus</i>	C	58.17.3	A 0	<i>Notoscopelus elongatus</i> (Costa, 1844)
NOTSKRO	<i>Notoscopelus kroyerii</i>	C	58.17.4	A 0	<i>Notoscopelus kroyeri</i> (Malm, 1861)
OBLAMEL	<i>Oblada melanura</i>	C	139.6.1	A 0	<i>Oblada melanura</i> (Linnaeus, 1758)
OCENERI	<i>Ocenebra erinacea</i>	R	RIEDL	E 0	<i>Ocenebra erinaceus</i> (Linnaeus, 1758)
OCTODEP	<i>Octopus defilippi</i>	F	OCT Oct 10	C 0	<i>Octopus defilippi</i> Verany, 1851
OCTOMAC	<i>Octopus macropus</i>	F	OCT Oct 2	C 0	<i>Octopus macropus</i> Risso, 1826

Medits Code	Scientific Name	Source	Reference	CATFAU	CODLON	Valid Name (Ifremer reference)
OCTOSAL	Octopus salutii	F	OCT Oct 23	C	0	Octopus salutii Verany, 1836
OCTOSPP	Octopus spp	F	OCT Oct	C	0	Octopus Cuvier, 1797
OCTOTET	Pteroctopus tetracirrhus	F	OCT Pter 1	C	0	Pteroctopus tetracirrhus (Delle Chiaje, 1830)
OCTOVUL	Octopus vulgaris	F	OCT Oct 1	C	0	Octopus vulgaris Cuvier, 1797
OCYTTUB	Ocythoe tuberculata	F	OCY ocy 1	C	0	Ocythoe tuberculata Rafinesque, 1814
ODONFER	Odontaspis ferox	C	5.1.1	A	0	Odontaspis ferox (Risso, 1810)
ODONTAU	Eugonphodus (Odontaspis) taurus	C	5.1.3	A	0	Carcharias taurus Rafinesque, 1810
OEDALAB	Oedalechilus labeo	C	181.4.1	A	0	Oedalechilus labeo (Cuvier, 1829)
OLIGATE	Oligopus ater	C	172.1.1	A	0	Grammonus ater (Risso, 1810)
ONYCBAN	Onychoteuthis banksi	F	ONYCHO	C	0	Onychoteuthis banksii (Leach, 1817)
ONYCSPP	Onychoteuthis spp	F	ONYCHO	C	0	Onychoteuthis Lichtenstein, 1818
OPDIBAR	Ophidion barbatum	C	173.1.1	A	0	Ophidion barbatum Linnaeus, 1758
OPDIROC	Ophidion rochei	C	173.1.2+3	A	0	Ophidion rochei Müller, 1845
OPHCRUF	Ophichthus rufus	C	86.1.2	A	0	Ophichthus rufus (Rafinesque, 1810)
OPHISER	Ophisurus serpens	C	86.4.1	A	0	Ophisurus serpens (Linnaeus, 1758)
OPHOFRA	Ophiothrix fragilis	R	RIEDL	E	0	Ophiothrix fragilis (Abildgaard)
OPISSPP	Opisthobranchia spp			E	0	Opisthobranchia Milne-Edwards, 1848
OPLOSPP	Oplophoridae	Z	Z	B	m	Oplophoridae Dana, 1852
OPTOAGA	Opisthoteuthis agassizii		FAUNA IBER	C	m	Opisthoteuthis agassizii Verrill, 1883
OSTREDU	Ostrea edulis	F	OSTR Ostr 1	D	0	Ostrea edulis Linnaeus, 1758
OSTRSPP	Ostrea spp.	R	RIEDL	E	0	Ostrea Linnaeus, 1758
OXYNCEN	Oxynotus centrina	C	15.1.1	A	0	Oxynotus centrina (Linnaeus, 1758)
PAGEACA	Pagellus acarne	C	139.7.2	A	0	Pagellus acarne (Risso, 1827)
PAGEBOG	Pagellus bogaraveo	C	139.7.3	A	0	Pagellus bogaraveo (Brünnich, 1768)
PAGEERY	Pagellus erythrinus	C	139.7.1	A	0	Pagellus erythrinus (Linnaeus, 1758)
PAGIERE	Paguristes eremita			B	m	Paguristes eremita (Linnaeus, 1767)
PAGUALA	Pagurus alatus	Z	Z	B	m	Pagurus alatus (J.C. Fabricius, 1775)
PAGUCUA	Pagurus cuanensis			B	m	Pagurus cuanensis Bell, 1845
PAGUEXC	Pagurus excavatus	Z	Z	B	m	Pagurus excavatus (Herbst, 1791)
PAGUFOR	Pagurus forbesii	Z	Z	B	m	Pagurus forbesii Bell, 1845
PAGUPRI	Pagurus prideauxi	Z	Z	B	m	Pagurus prideaux Leach, 1815
PALIELE	Palinurus elephas	F	PALIN Palin 1	B	m	Palinurus elephas (J.C. Fabricius, 1787)
PALIMAU	Palinurus mauritanicus	F	PALIN Palin 3	B	m	Palinurus mauritanicus Gruvel, 1911
PALISPP	Palinurus	F	PALIN	B	m	Palinurus Weber, 1795
PANDPRO	Pandalina profunda	F	PANDL	B	m	Pandalina profunda Holthuis, 1946
PAPANAR	Parapandalus narval	F	PANDL Parapnd	B	m	Plesionika narval (J.C. Fabricius, 1787)
PAPELON	Parapenaeus longirostris	F	PEN Parap 1	B	m	Parapenaeus longirostris (Lucas, 1846)
PAPOHUM	Parapristipoma humile	C	136.3.1	A	0	Parapristipoma humile (Bowdich, 1825)
PAPOOCT	Parapristipoma octolineatum	C	136.3.2	A	0	Parapristipoma octolineatum (Valenciennes, 1833)
PARALEP	Paraliparis leptochirus	C	192.3.3	A	0	Paraliparis leptochirus (Tortonese, 1959)
PARLCOR	Paralepis coregonoides	C	63.1	A	0	Paralepis coregonoides Risso, 1820
PARLSPE	Paralepis speciosa	F	PARALEP	A	0	Paralepis speciosa Bellotti, 1878
PAROCUV	Paromola cuvieri	F	HOM Par 1	B	m	Paromola cuvieri (Risso, 1816)
PARTANG	Parthenope angulifrons	Z	Z	B	m	Parthenope angulifrons Latreille, 1825
PARTMAC	Parthenope macrochelos	Z	Z	B	m	Parthenope macrochelos (Herbst, 1790)
PARTMAS	Parthenope massena	Z	Z	B	m	Parthenope massena (P. Roux, 1830)
PASIMUL	Pasiphaea multidentata	F	PASI Pasi 1	B	m	Pasiphaea multidentata Esmark, 1866
PASISIV	Pasiphaea sivado	F	PASI Pasi 2	B	m	Pasiphaea sivado (Risso, 1816)
PECTJAC	Pecten jacobaeus	F	PECT Pect 1	D	0	Pecten jacobaeus (Linnaeus, 1758)
PECTMAX	Pecten maximus	F	PECT	D	0	Pecten maximus (Linnaeus, 1758)

MeditS Code	Scientific Name	Source	Reference	CATFAU CODLON	Valid Name (Ifremer reference)
PECTSPP	Pecten	F	PECT	D 0	Pecten Müller O.F., 1776
PELTATR	Peltodoris atromaculata	R	RIEDL	E 0	Discodoris atromaculata (Bergh, 1880)
PENAKER	Penaeus kerathurus	F	PEN Pen 1	B m	Penaeus kerathurus (Forskål, 1775)
PENPHO	Pennatula phosphorea			D 0	Pennatula phosphorea Linnaeus, 1758
PERCGRA	Periclimenes granulatus	Z	Z	B m	Periclimenes granulatus Holthuis, 1950
PERICAT	Peristedion cataphractum	C	186.1.1	A 0	Peristedion cataphractum (Linnaeus, 1758)
PHALGRA	Phallium granulatum	F	CASS Phal 1	D 0	Phallium granulatum (Von Born, 1778)
PHILECH	Philocheras echinulatus	F	CRANG	B m	Philocheras echinulatus (M. Sars, 1861)
PHRYREG	Phrynorhombus regius	C	195.3.1	A 0	Phrynorhombus regius (Bonnaterre, 1788)
PHRYSPP	Phrynorhombus	C	195.3.1	A 0	Phrynorhombus Günther, 1862
PHYIBLE	Phycis blennoides	C	101.15.2	A 0	Phycis blennoides (Brünnich, 1768)
PHYIPHY	Phycis phycis	C	101.15.1	A 0	Phycis phycis (Linnaeus, 1766)
PHYLTRU	Phylonotus (Murex)(=Trunculariopsis)	F	MUR Phyl 1	D 0	Hexaplex trunculus (Linnaeus, 1758)
PHYSDAL	Physiculus dalwigki	C	103.8.1	A 0	Physiculus dalwigki Kaup, 1858
PILUSPI	Pilumnus spinifer	Z	Z	B m	Pilumnus hirtellus forma spinifer H. Milne-Edwards, 1834
PILUVIL	Pilumnus villosissimus	Z	Z	B m	Pilumnus villosissimus (Rafinesque, 1814)
PINNOB	Pinna nobilis	F	PINN Pinn 1	D 0	Pinna nobilis Linnaeus, 1758
PINNPEC	Pinna pectinata	R	RIEDL	E 0	Atrina pectinata (Linnaeus, 1767)
PINOPIN	Pinnotheres pinnotheres	Z	Z	B m	Nepinnotheres pinnotheres (Linnaeus, 1758)
PISAARN	Pisa armata	Z	Z	B m	Pisa armata (Latreille, 1803)
PISANOD	Pisa nodipes	Z	Z	B m	Pisa nodipes (Leach, 1815)
PISILON	Pisidia longicornis	Z	Z	B m	Pisidia longicornis (Linnaeus, 1767)
PLATFLE	Platichthys flesus	C	197.8.1	A 0	Platichthys flesus (Linnaeus, 1758)
PLEOMED	Plectorhinchus mediterraneus	C	136.4.1	A 0	Plectorhinchus mediterraneus (Guichenot, 1850)
PLERMEC	Pleurobranchaea meckely	R	RIEDL	E 0	Pleurobranchaea meckelii Meckel, 1813
PLESACA	Plesionika acanthonotus	Z	Z	B m	Plesionika acanthonotus (S.I. Smith, 1882)
PESANT	Plesionika antigai	Z	Z	B m	Plesionika antigai Zariquiey-Alvarez, 1955
PLESEDW	Plesionika edwardsii	F	PANDL Plesio 2	B m	Plesionika narval (J.C. Fabricius, 1787)
PLESGIG	Plesionika gigliolii	Z	Z	B m	Plesionika gigliolii (Senna, 1902)
PLESHET	Plesionika heterocarpus	F	PANDL Plesio 8	B m	Plesionika heterocarpus (A. Costa, 1871)
PLESMAR	Plesionika martia	F	PANDL Plesio 1	B m	Plesionika martia martia (A. Milne-Edwards, 1883)
PLEUPIL	Pleurobrachia pileus			E 0	Pleurobrachia pileus
POLARIS	Polyacanthonotus rissoanus	C	89.2.1	A 0	Polyacanthonotus rissoanus (De Filippi & Verany, 1857)
POLBHEN	Polybius henslowi	F	PORT	B m	Polybius henslowii Leach, 1820
POLCTYP	Polycheles typhlops	Z	Z	B m	Polycheles typhlops Typhlops Heller, 1862
POLYAME	Polyprion americanum	C	124.7.1	A 0	Polyprion americanus (Bloch & Schneider, 1801)
POMABEN	Pomadasys incisus (bennetti)	C	136.1.1	A 0	Pomadasys incisus (Bowdich, 1825)
POMSMAR	Pomatoschistus marmoratus	C	162.21.4	A 0	Pomatoschistus marmoratus (Risso, 1810)
POMSMIC	Pomatoschistus microps	C	162.21.5	A 0	Pomatoschistus microps (Krøyer, 1838)
POMSMIN	Pomatoschistus minutus	C	162.21.1	A 0	Pomatoschistus minutus (Pallas, 1770)
POMTSAL	Pomatomus saltator	C	129.1.1	A 0	Pomatomus saltatrix (Linnaeus, 1766)
PONIKUH	Pontinus kuhlii	C	184.3.1	A 0	Pontinus kuhlii (Bowdich, 1825)
PONPNOR	Pontophilus norvegicus	Z	Z	B m	Pontophilus norvegicus (M. Sars, 1861)
PONPSPI	Pontophilus spinosus	F	CRANG Pontop 1	B m	Pontophilus spinosus (Leach, 1815)
PONTCAT	Pontocaris cataphractus	Z	Z	B m	Aegaeon cataphractus (Olivi, 1792)
PONTLAC	Pontocaris lacazei	F	CRANG Pont 1	B m	Aegaeon lacazei (Gourret, 1887)
PRIOGLA	Prionace glauca	C	13.8.1	A 0	Prionace glauca (Linnaeus, 1758)
PROCEDU	Processa edulis	F	PROC Proc 2	B m	Processa edulis crassipes Nouvel & Holthuis,

MeditS Code	Scientific Name	Source	Reference	CATFAU	CODLON	Valid Name (Ifremer reference)
						1957
PROCMED	<i>Processa canaliculata</i> (mediterranea)	F	PROC Proc 1	B	m	<i>Processa canaliculata</i> Leach, 1815
PROCNOU	<i>Processa nouveli</i>	F	PROC	B	m	<i>Processa nouveli</i> holthuisi Al-Adhub & Williamson, 1975
PROSSPP	<i>Prosobranchia</i> spp			E	0	<i>Prosobranchia</i> Milne Edwards, 1848
PSAMMIC	<i>Psamechinus microtuberculatus</i>	R	RIEDL	E	0	<i>Psammechinus microtuberculatus</i>
PSENPEL	<i>Psenes pellucidus</i>	C	177.3.2	A	0	<i>Psenes pellucidus</i> Lütken, 1880
PSETMAX	<i>Psetta maxima</i>	C	195.4.1	A	0	<i>Psetta maxima</i> (Linnaeus, 1758)
PSEVCAR	<i>Pseudosimnia carnea</i>		D'Angelo	E	m	<i>Pseudosimnia carnea</i> (Poiret, 1789)
PTEAPEL	<i>Pteragogus pelycus</i>	X	X	A?		<i>Pteragogus pelycus</i> Randall, 1981
PTEOBOV	<i>Pteromylaeus bovinus</i>	C	23.2.1	A	0	<i>Pteromylaeus bovinus</i> (Geoffroy Saint-Hilaire, 1817)
PTERHIR	<i>Pteria hirundo</i>	F	PTER	D	0	<i>Pteria hirundo</i> (Linnaeus, 1758)
PUNTPUN	<i>Diplodus</i> (<i>Puntazzo</i>) <i>puntazzo</i>	C	137.8.1	A	0	<i>Diplodus puntazzo</i> (Cetti, 1777)
RAJAALB	<i>Raja alba</i>	C	21.1.18	A	0	<i>Rostroraja alba</i> (Lacepède, 1803)
RAJAAST	<i>Raja asterias</i>	C	21.1.2	A	0	<i>Raja asterias</i> Delaroche, 1809
RAJABAT	<i>Raja batis</i>	C	21.1.10	A	0	<i>Dipturus batis</i> (Linnaeus, 1758)
RAJABRA	<i>Raja brachyura</i>	C	21.1.3	A	0	<i>Raja brachyura</i> Lafont, 1873
RAJACIR	<i>Raja circularis</i>	C	21.1.14	A	0	<i>Leucoraja circularis</i> (Couch, 1838)
RAJACLA	<i>Raja clavata</i>	C	21.1.4	A	0	<i>Raja clavata</i> Linnaeus, 1758
RAJAFUL	<i>Raja fullonica</i>	C	21.1.13	A	0	<i>Leucoraja fullonica</i> (Linnaeus, 1758)
RAJAMEL	<i>Raja melitensis</i>	C	21.1.21	A	0	<i>Leucoraja melitensis</i> (Clark, 1926)
RAJAMIR	<i>Raja miraletus</i>	C	21.1.1	A	0	<i>Raja miraletus</i> Linnaeus, 1758
RAJAMON	<i>Raja montagui</i>	C	21.1.7	A	0	<i>Raja montagui</i> Fowler, 1910
RAJANAE	<i>Raja naevus</i>	C	21.1.15	A	0	<i>Leucoraja naevus</i> (Müller & Henle, 1841)
RAJAOXY	<i>Raja oxyrinchus</i>	C	21.1.12	A	0	<i>Dipturus oxyrinchus</i> (Linnaeus, 1758)
RAJAPOL	<i>Raja polystigma</i>	C	21.1.22	A	0	<i>Raja polystigma</i> Regan, 1923
RAJARDA	<i>Raja radula</i>	C	21.1.23	A	0	<i>Raja radula</i> Delaroche, 1809
RAJASPP	<i>Raja</i>	C	21.1.12	A	0	<i>Raja</i> Linnaeus, 1758
RAJAUND	<i>Raja undulata</i>	C	21.1.25	A	0	<i>Raja undulata</i> Lacepède, 1802
REGAGLE	<i>Regalecus glesne</i>	C	106.1.1.	A	0	<i>Regalecus glesne</i> Ascanius, 1772
RHINCEM	<i>Rhinobatos cemiculus</i>	C	19.1.2	A	0	<i>Rhinobatos cemiculus</i> Geoffroy Saint-Hilaire, 1817
RHINRHI	<i>Rhinobatos rhinobatos</i>	C	19.1.1	A	0	<i>Rhinobatos rhinobatos</i> (Linnaeus, 1758)
RHIPMAR	<i>Rhinoptera marginata</i>	C	24.1.1	A	0	<i>Rhinoptera marginata</i> (Geoffroy Saint-Hilaire, 1817)
RHYNHEP	<i>Rhynchogadus hepaticus</i>	C	103.9.1	A	0	<i>Rhynchogadus hepaticus</i> (Facciola, 1884)
RICHFRE	<i>Richardina fredericii</i>	Z	Z	A	0	<i>Richardina fredericii</i> Lo Bianco, 1903
RISSDES	<i>Rissoides desmaresti</i>	F	SQUIL	D	0	<i>Rissoides desmaresti</i> (Risso, 1816)
RISSPAL	<i>Rissoides pallidus</i>	F	SQUIL	B	m	<i>Rissoides pallidus</i> (Giesbrecht, 1910)
ROCHCAR	<i>Rochinia carpenteri</i>	Z	Z	B	m	<i>Rochinia carpenteri</i> (Wyville & Thomson, 1873)
RONDMIN	<i>Rondeletiola minor</i>	F	SEPIOL	C	0	<i>Rondeletiola minor</i> (Naef, 1912)
ROSSMAC	<i>Rossia macrosoma</i>	F	SEPIOL Ross 1	C	0	<i>Rossia macrosoma</i> (Delle Chiaje, 1828)
SADASAR	<i>Sarda sarda</i>	C	158.4.1	A	0	<i>Sarda sarda</i> (Bloch, 1793)
SALOTRU	<i>Salmo trutta trutta</i>	C	45.1.2	A	0	<i>Salmo trutta trutta</i> Linnaeus, 1758
SARDPIL	<i>Sardina pilchardus</i>	C	33.3.1	A	0	<i>Sardina pilchardus</i> (Walbaum, 1792)
SARIAUR	<i>Sardinella aurita</i>	C	33.4.1	A	0	<i>Sardinella aurita</i> Valenciennes, 1847
SARIMAD	<i>Sardinella maderensis</i>	C	33.4.2	A	0	<i>Sardinella maderensis</i> (Lowe, 1838)
SARPSAL	<i>Sarpa salpa</i>	C	139.9.1	A	0	<i>Sarpa salpa</i> (Linnaeus, 1758)
SCAEUNI	<i>Scaevurgus unicolor</i>	F	OCT Scae 1	C	0	<i>Scaevurgus unicolor</i> (Delle Chiaje, 1838)
SCALSCA	<i>Scalpelum scalpelum</i>	R	Riedl	B	m	<i>Scalpellum scalpellum</i> (Linnaeus, 1758)
SCAPNIG	<i>Scaphander lignarius</i>			D	0	<i>Scaphander lignarius</i> (Linnaeus, 1758)

MeditS Code	Scientific Name	Source	Reference	CATFAU CODLON	Valid Name (Ifremer reference)
SCHEOVA	Schedophilus ovalis	C	176.3.2	A 0	Schedophilus ovalis (Cuvier, 1833)
SCIAUMB	Sciaena umbra	C	137.1.1	A 0	Sciaena umbra Linnaeus, 1758
SCOBSAU	Scomberesox saurus	C	91.1.1	A 0	Scomberesox saurus saurus (Walbaum, 1792)
SCOHRHO	Scophthalmus rhombus	C	195.1.1	A 0	Scophthalmus rhombus (Linnaeus, 1758)
SCOMPNE	Scomber (Pneumatophorus) japonicus	C	156.1.2	A 0	Scomber japonicus Houttuyn, 1782
SCOMSCO	Scomber scombrus	C	156.1.1	A 0	Scomber scombrus Linnaeus, 1758
SCORELO	Scorpaena elongata	C	184.1.3	A 0	Scorpaena elongata Cadenat, 1943
SCORLOP	Scorpaena loppei	C	184.1.5	A 0	Scorpaena loppei Cadenat, 1943
SCORMAD	Scorpaena maderensis	C	184.1.6	A 0	Scorpaena madurensis Valenciennes, 1833
SCORNOT	Scorpaena notata	C	184.1.7	A 0	Scorpaena notata Rafinesque, 1810
SCORPOR	Scorpaena porcus	C	184.1.1	A 0	Scorpaena porcus Linnaeus, 1758
SCORSCO	Scorpaena scrofa	C	184.1.8	A 0	Scorpaena scrofa Linnaeus, 1758
SCYLARC	Scyllarus arctus	F	SCYL Scylr 1	B m	Scyllarus arctus (Linnaeus, 1758)
SCYLLAT	Scyllarides latus	F	SCYL Scyld 1	B m	Scyllarides latus (Latreille, 1803)
SCYLPYG	Scyllarus pygmaeus	F	SCYL Scylr 2	B m	Scyllarus pygmaeus (Bate, 1888)
SCYMLIC	Dalatias (Scymnorhinus) licha	C	16.4.3	A 0	Dalatias licha (Bonnaterre, 1788)
SCYOCAN	Scyliorhinus canicula	C	11.1.1	A 0	Scyliorhinus canicula (Linnaeus, 1758)
SCYOSTE	Scyliorhinus stellaris	C	11.1.2	A 0	Scyliorhinus stellaris (Linnaeus, 1758)
SEPENEG	Sepietta neglecta	F	SEPIOL	C 0	Sepietta neglecta Naef, 1916
SEPEOBS	Sepietta obscura	F	SEPIOL	C 0	Sepietta obscura Naef, 1916
SEPEOWE	Sepietta oweniana	F	SEPIOL	C 0	Sepietta oweniana (D'Orbigny, 1839)
SEPESPP	Sepietta spp.	F	SEPIOL	C 0	Sepietta Naef, 1912
SEPIELE	Sepia elegans	F	SEP Sep 3	C 0	Sepia elegans De Blainville, 1827
SEPIOFF	Sepia officinalis	F	SEP Sep 1	C 0	Sepia officinalis Linnaeus, 1758
SEPIORB	Sepia orbignyana	F	SEP Sep 4	C 0	Sepia orbignyana De Férussac, 1826
SEPIOSP	Sepia	F	SEP Sep 1	C 0	Sepia Linnaeus, 1758
SEPOAFF	Sepiola affinis	F	SEPIOL	C 0	Sepiola affinis Naef, 1912
SEPOINT	Sepiola intermedia	F	SEPIOL	C 0	Sepiola intermedia Naef, 1912
SEPOLIG	Sepiola ligulata	F	SEPIOL	C 0	Sepiola ligulata Naef, 1912
SEPOROB	Sepiola robusta	F	SEPIOL	C 0	Sepiola robusta Naef, 1912
SEPORON	Sepiola rondeleti	F	SEPIOL	C 0	Sepiola rondeleti Leach, 1817
SEPOSPP	Sepiola spp	F	SEP	C 0	Sepiola Leach, 1817
SERAATR	Serranus atricauda	C	124.1.2	A 0	Serranus atricauda Günther, 1874
SERACAB	Serranus cabrilla	C	124.1.1	A 0	Serranus cabrilla (Linnaeus, 1758)
SERAHEP	Serranus hepatus	C	124.1.3	A 0	Serranus hepatus (Linnaeus, 1758)
SERASCR	Serranus scriba	C	124.1.4	A 0	Serranus scriba (Linnaeus, 1758)
SERGARC	Sergestes arcticus	Z	Z	B m	Sergestes arcticus Krøyer, 1855
SERGROB	Sergestes robustus	Z	Z	B m	Sergestes robustus (S.J. Smith, 1882)
SERGSAR	Sergestes sargassi (= henseni)	Z	Z	B m	Sergestes sargassi Ortmann, 1893
SERIDUM	Seriola dumerili	C	131.9.1	A 0	Seriola dumerili (Risso, 1810)
SOLEIMP	Solea impar	C	198.1.2	A 0	Solea impar Bennett, 1831
SOLEKLE	Solea kleini	C	198.1.3	A 0	Synaptura kleinii (Risso, 1827)
SOLELAS	Solea lascaris	C	198.1.4	A 0	Solea lascaris (Risso, 1810)
SOLESEN	Solea senegalensis	C	198.1.6	A 0	Solea senegalensis Kaup, 1858
SOLEVUL	Solea vulgaris	C	198.1.1	A 0	Solea solea (Linnaeus, 1758)
SOLOMEM	Solenocera membranacea	F	SOLENO Soleno	B m	Solenocera membranacea (Risso, 1816)
SPARAUR	Sparus aurata	C	139.1.1	A 0	Sparus auratus Linnaeus, 1758
SPARCAE	Pagrus (Sparus) coeruleostictus	C	139.11.2	A 0	Pagrus caeruleostictus (Valenciennes, 1830)
SPARPAG	Pagrus (Sparus) pagrus	C	139.11.3	A 0	Pagrus pagrus (Linnaeus, 1758)
SPHOCUT	Sphoeroides cutaneus	C	204.3.2	A 0	Sphoeroides pachygaster (Müller & Troschel, 1848)

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SPHYSPY	<i>Sphyraena sphyraena</i>	C	180.1.1	A	0	<i>Sphyraena sphyraena</i> (Linnaeus, 1758)
SPICFLE	<i>Spicara flexuosa</i>	C	141.2.2	A	0	<i>Spicara maena</i> (Linnaeus, 1758)
SPICMAE	<i>Spicara maena</i>	C	141.2.1	A	0	<i>Spicara maena</i> (Linnaeus, 1758)
SPIC SMA	<i>Spicara smarís</i>	C	141.2.3	A	0	<i>Spicara smarís</i> (Linnaeus, 1758)
SPIC SPP	<i>Spicara</i>	C	141.2.3	A	0	<i>Spicara Rafinesque</i> , 1810
SPISSPP	<i>Spisula spp</i>	F	MACTR	E	0	<i>Spisula Gray</i> , 1837
SPISSUB	<i>Spisula subtrucata</i>	F	MACTR	E	0	<i>Spisula subtruncata</i> (Da Costa, 1778)
SPODCAN	<i>Spondyliosoma cantharus</i>	C	139.10.1	A	0	<i>Spondyliosoma cantharus</i> (Linnaeus, 1758)
SPRASPR	<i>Sprattus sprattus</i>	C	33.5.1	A	0	<i>Sprattus sprattus sprattus</i> (Linnaeus, 1758)
SQUAACA	<i>Squalus acanthias</i>	C	16.1.1	A	0	<i>Squalus acanthias</i> Linnaeus, 1758
SQUABLA	<i>Squalus blainvillei</i>	C	16.1.2	A	0	<i>Squalus blainville</i> (Risso, 1827)
SQUIMAN	<i>Squilla mantis</i>	F	SQUIL Squil 5	B	m	<i>Squilla mantis</i> (Linnaeus, 1758)
SQUTACU	<i>Squatina aculeata</i>	C	17.1.2	A	0	<i>Squatina aculeata</i> Cuvier, 1829
SQUTOCL	<i>Squatina oculata</i>	C	17.1.3	A	0	<i>Squatina oculata</i> Bonaparte, 1840
SQUTSPP	<i>Squatina spp</i>	C	17.1	A	0	<i>Squatina Duméril</i> , 1806
SQUTSQU	<i>Squatina squatina</i>	C	17.1.1	A	0	<i>Squatina squatina</i> (Linnaeus, 1758)
STEPDIA	<i>Stephanolepis diaspros</i>	C	202.1.2	A	0	<i>Stephanolepis diaspros</i> Fraser-Brunner, 1940
STICREG	<i>Stichopus regalis</i>	F	STICH Stich 1	D	0	<i>Eotichopus regalis</i>
STOLLEU	<i>Stoloteuthis leucoptera</i>	F	SEPIOL	C	0	<i>Stoloteuthis leucopterus</i> (Verrill, 1878)
STOMBOA	<i>Stomias boa</i>	C	41.1.1	A	0	<i>Stomias boa boa</i> (Risso, 1810)
STROFIA	<i>Stromateus fiatola</i>	C	179.1.1	A	0	<i>Stromateus fiatola</i> Linnaeus, 1758
SUBECAR	<i>Suberites carnosus</i>			D	0	<i>Suberites carnosus</i> (Johnston, 1842)
SUBEDOM	<i>Suberites domuncula</i>			D	0	<i>Suberites domuncula</i> (Olivi, 1792)
SUBESPP	<i>Suberites spp.</i>			D	0	<i>Suberites Nardo</i> , 1833
SYMBVER	<i>Symbolophorus veranyi</i>	C	58.19.1	A	0	<i>Symbolophorus veranyi</i> (Moreau, 1888)
SYMDCIN	<i>Symphodus cinereus</i>	C	145.9.3	A	0	<i>Symphodus cinereus</i> (Bonnaterre, 1788)
SYM DMED	<i>Symphodus mediterraneus</i>	C	145.9.6	A	0	<i>Symphodus mediterraneus</i> (Linnaeus, 1758)
SYMDOCE	<i>Symphodus ocellatus</i>	C	145.9.9	A	0	<i>Symphodus ocellatus</i> (Forsskål, 1775)
SYM DROS	<i>Symphodus rostratus</i>	C	145.9.1	A	0	<i>Symphodus rostratus</i> (Bloch, 1791)
SYM DTIN	<i>Symphodus tinca</i>	C	145.9.12	A	0	<i>Symphodus tinca</i> (Linnaeus, 1758)
SYMPLIG	<i>Symphurus ligulatus</i>	C	199.2.2	A	0	<i>Symphurus ligulatus</i> (Cocco, 1844)
SYM PNIG	<i>Symphurus nigrescens</i>	C	199.2.1	A	0	<i>Symphurus nigrescens</i> Rafinesque, 1810
SYNDSAU	<i>Synodus saurus</i>	C	51.1.2	A	0	<i>Synodus saurus</i> (Linnaeus, 1758)
SYNGACU	<i>Syngnathus acus</i>	C	97.1.1	A	0	<i>Syngnathus acus</i> Linnaeus, 1758
SYNGPHL	<i>Syngnathus phlegon</i>	C	97.1.3.	A	0	<i>Syngnathus phlegon</i> Risso, 1827
SYNGTAE	<i>Syngnathus taenionotus</i>	C	97.1.6	A	0	<i>Syngnathus taenionotus</i> Canestrini, 1871
SYNGTYP	<i>Syngnathus typhle</i>	C	97.1.8	A	0	<i>Syngnathus typhle</i> Linnaeus, 1758
TAENGRA	<i>Taeniura grabata</i>	C	22.4.1	A	0	<i>Taeniura grabata</i> (Geoffroy Saint-Hilaire, 1817)
TELLSPP	<i>Tellina spp</i>	F	TELL	E	0	<i>Tellina</i> Linnaeus, 1758
TETHFIM	<i>Tethys fimbria</i>			C	0	<i>Tethys fimbria</i> Linnaeus, 1767
THAMPOI	<i>Thalamita poissonii</i>	Y	Y	B		<i>Thalamita poissonii</i> (Audouin, 1826)
TODASAG	<i>Todarodes sagittatus</i>	F	OMMAS Todarod	C	0	<i>Todarodes sagittatus</i> (Lamarck, 1798)
TODIEBL	<i>Todaropsis eblanae</i>	F	OMMAS Todarod	C	0	<i>Todaropsis eblanae</i> (Ball, 1841)
TORPMAR	<i>Torpedo marmorata</i>	C	20.1.2	A	0	<i>Torpedo marmorata</i> Risso, 1810
TORPNOB	<i>Torpedo nobiliana</i>	C	20.1.3	A	0	<i>Torpedo nobiliana</i> Bonaparte, 1835
TORPSPP	<i>Torpedo</i>	C	20.1.1	A	0	<i>Torpedo Houttuyn</i> , 1764
TORPTOR	<i>Torpedo torpedo</i>	C	20.1.1	A	0	<i>Torpedo torpedo</i> (Linnaeus, 1758)
TRACMED	<i>Trachurus mediterraneus</i>	C	131.10.3	A	0	<i>Trachurus mediterraneus</i> (Steindachner, 1868)
TRACPIC	<i>Trachurus picturatus</i>	C	131.10.4	A	0	<i>Trachurus picturatus</i> (Bowdich, 1825)
TRACTRA	<i>Trachurus trachurus</i>	C	131.10.1	A	0	<i>Trachurus trachurus</i> (Linnaeus, 1758)

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TRAHARA	<i>Trachinus araneus</i>	C	148.1.2	A 0	<i>Trachinus araneus</i> Cuvier, 1829
TRAHDRA	<i>Trachinus draco</i>	C	148.1.1	A 0	<i>Trachinus draco</i> Linnaeus, 1758
TRAHRAD	<i>Trachinus radiatus</i>	C	148.1.3	A 0	<i>Trachinus radiatus</i> Cuvier, 1829
TRARTRA	<i>Trachyrhynchus trachyrhynchus</i>	C	99.1.1	A 0	<i>Trachyrincus scabrus</i> (Rafinesque, 1810)
TRAYCRI	<i>Trachyscorpia cristulata</i>	C	184.7.1	A 0	<i>Trachyscorpia cristulata echinata</i> (Koehler, 1896)
TRIGLUC	<i>Trigla lucerna</i>	C	185.1.2	A 0	<i>Chelidonichthys lucernus</i> (Linnaeus, 1758)
TRIGLYR	<i>Trigla lyra</i>	C	185.1.1	A 0	<i>Trigla lyra</i> Linnaeus, 1758
TRIILEP	<i>Trichiurus lepturus</i>	C	155.1.1	A 0	<i>Trichiurus lepturus</i> Linnaeus, 1758
TRIPLAS	<i>Trigloporus lastoviza</i>	C	185.5.1	A 0	<i>Chelidonichthys lastoviza</i> (Bonnaterre, 1788)
TRISCAP	<i>Trisopterus minutus capelanus</i>	C	101.11.1	A 0	<i>Trisopterus minutus</i> (Linnaeus, 1758)
TRISLUS	<i>Trisopterus luscus</i>	C	101.11.3	A 0	<i>Trisopterus luscus</i> (Linnaeus, 1758)
TRITNOD	<i>Charonia (Triton) rubicunda</i> (nodifer)	F	CYM Char 1	D 0	<i>Charonia lampas</i> (Linnaeus, 1758)
TURRCOM	<i>Turritella comunis</i>	F	D12	E 0	<i>Turritella communis</i> Risso, 1826
TURRSIM	<i>Turris similis</i>		D'Angelo	E 0	<i>Fusiturris similis</i> (Bivona And., 1838)
TURRSPP	<i>Turritella</i> spp	F	TURR	D 0	<i>Turritella</i> Lamarck, 1799
UMBAMED	<i>Umbraculum mediterraneum</i>			E 0	<i>Umbraculum umbraculum</i> (Röding, 1798)
UMBRCAN	<i>Umbrina canariensis</i>	C	137.4.2	A 0	<i>Umbrina canariensis</i> Valenciennes, 1843
UMBRCIR	<i>Umbrina cirrosa</i>	C	137.4.1	A 0	<i>Umbrina cirrosa</i> (Linnaeus, 1758)
UMBRRON	<i>Umbrina ronchus</i>	C	137.4.3	A 0	<i>Umbrina ronchus</i> Valenciennes, 1843
UPENMOL	<i>Upeneus moluccensis</i>	X	X	A	<i>Upeneus moluccensis</i> (Bleeker, 1855)
URANSCA	<i>Uranoscopus scaber</i>	C	149.1.1	A 0	<i>Uranoscopus scaber</i> Linnaeus, 1758
VENUSPP	<i>Venus</i> spp.	F	VEN	D 0	<i>Venus</i> Linnaeus, 1758
VENUVER	<i>Venus verrucosa</i>	F	VEN Ven 1	D 0	<i>Venus verrucosa</i> Linnaeus, 1758
VINCATT	<i>Vinciguerria attenuata</i>	C	37.12.1	A 0	<i>Vinciguerria attenuata</i> (Cocco, 1838)
VINCPOW	<i>Vinciguerria poweriae</i>	C	37.12.3	A 0	<i>Vinciguerria poweriae</i> (Cocco, 1838)
XANTCOU	<i>Medaeus (Xantho) couchi</i>	Z	Z	B m	<i>Monodaeus couchi</i> (Couch, 1851)
XENOCRI	<i>Xenophora crispa</i>	F	XENOPH	E m	<i>Xenophora crispa</i> (Koenig, 1825)
XENOSPP	<i>Xenophora</i> spp	F	XENOPH	E m	<i>Xenophora</i> Fischer Von Waldheim, 1807
XIPHGLA	<i>Xiphias gladius</i>	C	161.1.1	A 0	<i>Xiphias gladius</i> Linnaeus, 1758
ZEUSFAB	<i>Zeus faber</i>	C	120.1.1	A 0	<i>Zeus faber</i> Linnaeus, 1758
ZOSTOPH	<i>Zostoricessor ophiocephalus</i>	C	162.26.1	A O	<i>Zosterisessor ophiocephalus</i> (Pallas, 1814)

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