

1 **Structure and function of nematode communities across the Indian**  
2 **western continental margin and its oxygen minimum zone**

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11

12 **Abstract.** We studied patterns of nematode distribution along the western Indian continental  
13 margin to determine the influence of habitat heterogeneity and low oxygen levels on the  
14 community's taxonomic and functional structure. A single transect, perpendicular to the coast at  
15 14° N latitude was sampled from 34 to 2546 m depth for biological and environmental variables  
16 during August 2007. The oxygen minimum zone was extending from 102 m to 1001 m.  
17 Nematodes were identified to species (described and undescribed) and classified according to  
18 biological/functional traits. A total of 110 nematode species belonging to 24 families found along  
19 the transect. Three depth zones were identified, the shelf (depth range 34-102 m; highest  
20 nematode mean density:  $176.6 \pm 37$  ind/10 cm<sup>2</sup>), the slope (525-1524m;  $124.3 \pm 16$  ind/10 cm<sup>2</sup>),  
21 and the basin (2001-2546 m;  $62.9 \pm 2$  ind/10 cm<sup>2</sup>). Across the entire study area, the dominant  
22 species were *Terschellingia longicaudata*, *Desmodora* sp 1, *Sphaerolaimus gracilis*, and  
23 *Theristus ensifer*; their maximum density was at shelf stations. Nematode communities differed  
24 in species composition according to the zones, *Chromadorita* sp 2 (2.78%) *Sphaerolaimus*  
25 *gracilis* (2.21%) were dominant in shelf whereas *Terschellingia Longicaudata* (4.73%) and  
26 *Desmodora* sp 1 (4.42%) were dominant in the slope. However, in the deep basin *Halalaimus* sp  
27 1(1.11%) *Acantholaimus elegans* (1.11%) were seen dominant. The information in a particular  
28 functional group was not a simple reflection of the information in species abundance. Ecological  
29 information captured by adult length, adult shape, and life-history strategy was less site-specific  
30 and thus differed notably from information contained in other taxonomic groups. The functional  
31 composition of nematodes was strongly linked to the organic carbon and dissolved oxygen  
32 concentration. The OMZ harboured seven exclusively species: *Pselionema* sp 1, *Choanolaimus*  
33 sp 2, *Halichoanolaimus* sp 1, *Cobbia dentata*, *Daptonema* sp 1, *Trissonchulus* sp 1, and  
34 *Minolaimus* sp 1. Moreover, in our study, species diversity was higher on the shelf than on the  
35 slope or in the basin. Correlation with a number of environmental variables indicated that food  
36 quantity (measured as the organic carbon content and chlorophyll content) and oxygen level were  
37 the major factors that influenced the nematode community structure and function.

38  
39 Key words: oxygen minimum zone, habitat heterogeneity, Nematoda, functional traits, deep sea,  
40 Arabian Sea, Indian continental margin.

## 41 **1 Introduction**

42

43 The continental margin exhibits great topographical heterogeneity in terms of geomorphological  
44 features (e.g., Canyons and seamounts, shelf, slope, rise, marginal highs) and their related  
45 environmental conditions (e.g., depth, pressure, temperature, salinity, light, dissolved oxygen,  
46 sediment characteristics) (Levin et al., 2001). This heterogeneous environment is of great  
47 ecological interest because of its wide range gradients in living conditions and the high faunal  
48 diversity it supports on local, regional, and global scales (Vanreusel et al., 2010). The western  
49 Indian continental margin, located in the eastern Arabian Sea of the northern Indian Ocean,  
50 includes a series of complex environments including shelf, slope, and a permanent oxygen-  
51 depleted zone. In general, oxygen minimum zones (OMZs) are defined as layers of the water  
52 column where dissolved oxygen (DO) concentrations fall below  $0.5 \text{ ml l}^{-1}$ . Oxygen minima are  
53 common in the world oceans (Kamykowski and Zentara, 1990) and are found at intermediate  
54 depths in the Arabian Sea, off western Mexico, and off Peru and northern Chile. The Arabian Sea  
55 OMZ is one of the strongest and most intense in the world, with oxygen concentrations  $< 2 \mu\text{M}$  in  
56 its core (Helly and Levin, 2004; Paulmier and Ruiz-Pino, 2009; Zettler et al., 2009). The OMZ in  
57 the Arabian Sea is spread over  $285,000 \text{ km}^2$  of the benthic area. The entire OMZ occupies  
58 approximately the 150–1000 m depth range and is the thickest of the three major OMZs of the  
59 open ocean (Banse et al., 2014). Although the Arabian Sea covers only 2% of the surface area of  
60 the World Ocean, it is one of the most biologically productive regions (Ryther and Menzel,  
61 1965).

62 The entire western Indian continental margin (shelf to slope) supports a high benthic biodiversity  
63 (Ingole et al., 2010). Free-living nematodes are prominent members of the meiobenthos along the  
64 ocean continental margin, often constituting  $>90\%$  of all metazoa (Vincx et al., 1994). Moreover,  
65 they are more tolerant than macro- and other meiofauna to anoxic conditions (Giere, 1993;  
66 Moodley et al., 1997). Studies like Jensen (1987) and Murrell and Fleeger (1989) reported on  
67 effects of low-oxygen condition on nematofauna in sandy sediments at shallow depths and from  
68 the Gulf of Mexico, respectively found that nematode abundance was unaffected by oxygen  
69 concentration. Cook et al. (2000) studied nematodes from the Arabian Sea OMZ and concluded  
70 that nutrient input is more important than the oxygen concentration. However in another  
71 experimental study in the tidal flats, hypoxia caused a dramatic change in the nematode  
72 community composition with reduction in diversity and abundance of all dominant nematodes

73 except for genus *Odontophora* (Van Colen et al., 2009). Niera et al. (2013) found that nematode  
74 generic and trophic diversity was lowest, at the most oxygen-depleted site along a central Chile  
75 margin. De Torch et al. (2013) in a short incubation experiment found that nematodes were not  
76 affected by anoxic conditions. He further concluded that responses to stress depend on the species  
77 and the frequency of the hypoxic periods. In contrast to several studies on the effects of anoxia  
78 on Nematoda at higher taxonomic levels little is known about their response at species level.  
79 Several transitional settings in the western Indian continental margin, including the shelf, slope  
80 (long stretch of OMZ) and basin, provide multiple oxygen and other environmental gradients.  
81 These settings allowed us to investigate how oxygen and environmental factors affect and  
82 modulate the structure and function of nematodes community at species level. An exclusive study  
83 from the Indian western continental margin with reference to nematode tolerance in the OMZ is  
84 therefore important. Furthermore, nematode community structure from the western Indian  
85 continental margin (including the OMZ) of the Arabian Sea remains unstudied. In general  
86 information on nematode species data is missing from the Arabian Sea and the present study can  
87 provide important basic information on nematode community structure.

88 However, traditional taxonomic based methods of nematode community analyses may not fully  
89 account for their diverse roles in ecosystem function (Schratzberger et al., 2007) therefore the  
90 inclusion of functional analysis has been recommended (de Jonge et al., 2006).

91 Many previous studies (e.g. Bremner et al., 2003; Bostrom et al., 2006) on macrobenthic  
92 invertebrates have shown that linking the taxonomic and functional aspects can reveal different  
93 relationships between assemblages and in order to explore the relationship between biodiversity  
94 and ecosystem function nematode could be considered the ideal model organism (Danovaro et al.,  
95 2008).

96 Nematode species in functional groups share several morphological traits (Chalcraft and  
97 Resetarits, 2003), and use of these traits may provide additional information on changes in  
98 biodiversity and also facilitate better comparison with other geographical regions (Bremner,  
99 2008). Giere (1993) showed that a classification based on buccal structures of marine nematodes  
100 had applications in a variety of marine habitats. Thistle and Sherman (1985) found that nematode  
101 tail shape formed an important biological trait especially in locomotion and reproduction.  
102 Furthermore, body size is known to influence many traits of an animal, such as its life history,  
103 physiology, and energy requirements, whereas species morphotype represents adaptations to life  
104 in oxygen-poor and sulphide-rich sediments (Jensen, 1987a). Bongers et al., (1990) classified the

105 genera (along a scale of 1 to 5) from r-selected 'colonisers' to K-selected 'persisters', according  
106 to their life history, to assess the condition of terrestrial and freshwater habitats but also of marine  
107 environments from the shelf.

108  
109 In this study we examined the nematode community structure along a habitat gradient using  
110 traditional diversity measures, taxonomic properties, and the functional-group concept based on  
111 biological traits so as to answer the following questions.

112  
113 Does the heterogeneous gradient like shelf, slope and deep basin of western Indian  
114 continental margin affect marine nematode community structurally and functionally?

115  
116 What are the patterns and drivers of variation in nematode composition and diversity  
117 (structural and functional) along the western Indian margin; i.e., are oxygen levels the main  
118 driver, or are other factors (sediment, productivity etc.) more important?

119

## 120 **2 Materials and Methods**

121

### 122 **2.1 Study area**

123 A single transect was selected perpendicular to the coast of the Arabian Sea at 14° N latitude  
124 (Fig. 1). The bottom topography in the Arabian Sea has following characteristics i.e. the  
125 Carlsberg Ridge is in the southern part of Arabian Sea and Murray Ridge is in the north part,  
126 however in the mid-Arabian Sea (from 14° N to 21° N) there is a topographic slope. The  
127 continental shelf is 120 km wide off the southern, narrows to about 60 km off 11° N and widens  
128 to about 350 km off the Gulf of Cambay (Shetye and Shenoi, 1988).

129 Surface circulation in the Arabian Sea is controlled by the seasonal variation in various types of  
130 winds. During the SW monsoon, biological productivity in the Arabian Sea lies mainly around  
131 the centers of seasonal upwelling off Arabian Peninsula, Somalia and southwest India (Qasim,  
132 1977). The Arabian Sea, is characterized by a very pronounced midwater oxygen minimum zone  
133 (OMZ) between 150 to 1250 m and is over 1000m thick extends vertically from the bottom of  
134 the euphotic layer (~100 m) to ~1000 m (Wishner et al., 1990). The variations in the intensity of  
135 the OMZ, related to upwelling intensity and thermocline ventilation by Indian Ocean Water

136 (Reichart et al., 1998). It is located directly beneath the productive upwelling region (de Sousa et  
137 al., 1996; Morrison et al., 1998).

138

139

## 140 2.2 Sampling

141 A detailed benthic sampling was performed on board ORV *Sagar Kanya* during August 2007  
142 (cruise no. SK 237). Eight stations were sampled, at water depths of 34, 48, 102, 525, 1001,  
143 1524, 2001 and 2546 m, by means of a spade box corer (50 × 50 × 50 cm size). According to Rao  
144 and Veerayya (2000) the transect can be divided into five regions—shelf (34, 48, and 102 m),  
145 upper slope (525 m), mid slope (1001 m), lower slope (1524 m), and basin (2001–2546 m). For  
146 our purposes, however, all three slope depths were considered a single region; the study area was  
147 therefore divided into only three depth zones—shelf, slope, and basin (Ingole et al., 2010). On the  
148 basis of oxygen concentration, the OMZ was defined as extending from 102 m to 1001 m. For  
149 nematode assemblages, a PVC core (5.7 cm diameter) was used for sub-sampling. At each  
150 station, two sub samples were sliced per cm down to 5 cm sediment depth and fixed in buffered  
151 4% formalin. Separate one sub-core was collected for organic carbon ( $C_{org}$ ), sediment  
152 chlorophyll-*a* (Chl-*a*) and they were frozen at  $-20^{\circ}\text{C}$ . Bottom-water dissolved oxygen (DO)  
153 measurements were taken with a DO sensor attached to the CTD for depths down to 1524 m,  
154 below this depth; water collected in Niskin bottles was used for DO.

155

## 156 2.3 Laboratory analysis

157 Dissolved oxygen was analyzed by Winkler's method (Strickland and Parsons, 1968). Sample  
158 bottles are stored upright in ice chest, dark location and were analyzed after a period of 8-9 hours.  
159 Chl-*a* was estimated by means of an acetone extraction method using fluorometer (Holm-Hansen  
160 and Riemann, 1978). Total-carbon analyses were carried out in freeze-dried sediments with an  
161 NCS 2500 (Model-EA / NA1110) CNS analyzer. Inorganic carbon was analyzed with a  $\text{CO}_2$   
162 Culometer analyzer, and the percentage of  $\text{CaCO}_3$  was calculated. Percentage of  $C_{org}$  was  
163 calculated by subtraction of inorganic from total carbon. Sediment granulometry was determined  
164 by a Malvern Laser Analyzer (Model—Hydro 2000MU).

165 Meiofauna samples were washed over a 500- $\mu\text{m}$  mesh and then sieved on a 32- $\mu\text{m}$  mesh to retain  
166 the meiofauna fraction. The retained fraction on 32- $\mu\text{m}$  mesh was then elutriated by the  
167 centrifugation-flotation technique using LUDOX (Vincx 1996; Heip et al., 1985). Nematodes

168 were counted through a stereo-microscope, and were picked and mounted on permanent glass  
169 slides for taxonomic identification. In all, 1798 nematodes were examined from the whole area.  
170 They were identified up to described and morphotype species level from the pictorial keys of  
171 [Platt and Warwick \(1983\)](#) and the NeMys database ([Vanaverbeke et al., 2015](#)). For identification  
172 to species level, each morphotype was sketched and measured by means of a camera lucida so  
173 that all specimens of the same genus could be compared.

174

## 175 **2.4 Nematode functional trait analysis**

176 Nematode species were classified according to their buccal morphology, tail shape, adult length,  
177 adult shape, and life history.

178

179 Feeding types: Individual nematode species were assigned to four feeding categories according to  
180 [Wieser \(1953\)](#): selective deposit feeders (1A), non-selective deposit feeders (1B), epigrowth  
181 feeders (2A), and predators/omnivores (2B).

182

183 Tail shape: [Thistle and Sherman \(1985\)](#) developed a functional-trait scheme based on tail shape.  
184 Tail types are diverse and variable and suggested as an effective method of characterizing  
185 nematode communities. Nematodes were assigned to four tail-shape groups, which are common  
186 in free-living marine nematodes: short/round, elongated/filiform, conical, and clavate.

187

188 Total length and body shape: Total length and the maximum body width for adult nematodes  
189 were measured, and from the results we calculated the length-width ratio. [Soetaert et al. \(2002\)](#)  
190 suggested three body-shape morphologies: stout, slender, and long/thin. Length-width ratio is a  
191 measure of a nematode's body shape; long/thin animals have high ratios, and stout animals low  
192 ratios. From measured length, each species was therefore assigned to one of four length groups (<  
193 1 mm, 1–2 mm, 2–4 mm, >4 mm) and three shape categories (stout, with a length–width ratio <  
194 18; slender, with a length–width ratio of 18–72; and long/thin, with a length–width ratio > 72).

195

196 Life history (C-P score) : Nematodes were allocated to life-history groups according to [Bongers](#)  
197 [\(1990\)](#) and [Bongers et al. \(1991\)](#); genera are classified on a scale from 1 to 5 from colonisers (1,  
198 short life cycle, high reproduction rates, high colonisation ability, tolerance of various types of  
199 disturbance) to persisters (5, long life cycles, low colonisation ability, few offspring, sensitivity to

200 disturbance). The species belonging to Monhysterid family were assigned to the c-p 2 class  
201 (“general opportunists”) as advised by Bongers et al. (1995) and latter mentioned by Pape et al.  
202 (2013) and as such there were no nematodes belonging to c-p class 1 (“enrichment  
203 opportunists”).

204 A biological traits matrix was constructed by assigning to each nematode species its affinity to  
205 each trait category and the percent (%) contribution of each species.

206

## 207 **2.5 Statistical analysis**

208 The similarity analysis at the species level was based on two types of similarity measures: the  
209 Bray-Curtis dissimilarity (standardised, square-root transformed) (Bray and Curtis, 1957), based  
210 on the relative abundances of nematode genera, and ordination using the Jaccard similarity index  
211 based on presence-absence (Clarke, 1993). Differences between the samples were visualized by  
212 means of non-metric multidimensional scaling (nMDS) plots.

213 PERMANOVA designed with two factors: “station” (combined all the station in the zone) nested  
214 in “zones” (shelf, slope and deep basin) was applied to detect the statistical significance of  
215 differences in pair-wise comparison of nematode communities from different zones  
216 (PERMANOVA Anderson, 2005; Anderson et al., 2008).

217 Diversity was expressed in terms of the expected number of species in a sample, EG (51). The  
218 other diversity indices were calculated by means of Margalef’s index (Margalef, 1968) for  
219 species richness (d), Pielou’s index (Pielou, 1966) for species evenness (J’), and the Shannon–  
220 Wiener index (Shannon and Weaver, 1963) for species diversity (H’ by using loge).

221 Environmental variables were then subjected to principal-components analysis (PCA) to identify  
222 the spatial patterns based on environmental data. A lower triangular Euclidean distance matrix  
223 relating to the ordination was constructed (Clarke and Green, 1988). Prior to the calculation of  
224 the Euclidean distance resemblance matrix, the data were checked for uniform distribution  
225 followed by normalisation (subtracting the mean and dividing by the standard deviation, for each  
226 variable) before analysis.

227 Analysis of collinearity was tested using draftsman plot and the associated standard product  
228 moment correlation coefficient between all pairs of variables and those with correlations ( $r^2$ ) >  
229 0.9 were omitted from the model. If distribution of residuals was skewed, natural logarithm  
230 transformation was applied to the response variable until assumptions were met by the best  
231 model. The relationships of taxonomic and functional traits with environmental variables were



232 examined by mean of the BIOENV procedure (Clarke and Ainsworth, 1993), which calculates  
233 rank correlations between a similarity matrix derived from biological data and matrices derived  
234 from the environmental variables, thereby defining a set of variables that 'best explain' the biotic  
235 structure. RELATE and stepwise distance-based linear model permutation test (DistLM,  
236 McArdle and Anderson, 2001) was performed to identify which set of environmental variables  
237 predict the multivariate variation to nematode species assemblages. The adjusted  $R^2$  was used as  
238 a selection criterion to enable the fitting of the best explanatory environmental variables in the  
239 model. Euclidean distance was used as resemblance measure in all DISTLM procedures. Results  
240 were visualized with a distance-based redundancy analysis (dbRDA) (Anderson et al., 2008).

241 To identify the species that would characterize the three zones compared (shelf, slope, and basin)  
242 we performed the Indicator Species Analysis or IndVal (Dufrene and Legendre, 1997) using  
243 multi-level pattern analysis (De Caceres et al., 2010) in the R environment (R. Development Core  
244 Team, 2010) with the 'indicspecies' function. The statistical significance of the relationship  
245 between the species and site was tested by means of Monte Carlo randomizations with 1000  
246 permutations. The details of the method are described by Dufrene and Legendre (1997). All the  
247 analyses were performed with the procedures in the R software (R Development Core Team,  
248 2010; Dimitriadou et al., 2011) and the PERMANOVA+ module of the PRIMER v6 software  
249 (Anderson et al., 2008; Clarke and Gorley, 2006). A schematic diagram was prepared using the  
250 taxonomic and functional dataset in order to represent the pattern in the benthic zones of western  
251 Indian continental margin.

252

253

## 254 **3 Results**

255

### 256 **3.1 Environmental parameters**

257 The sediment texture was variable, as the shelf region and upper slopes (525 m) were  
258 characterised by silty and sandy facies, whereas the sediments at the mid slope (1001 m), lower  
259 slope (1524 m), and basin were characterised by clayish silts. The salinity of the bottom water  
260 varied little, but temperature was decreased with depth. The bottom-water DO ranged from 0.08  
261 to 2.3 ml l<sup>-1</sup>. The lowest near-bottom oxygen levels were recorded on the upper slope and mid  
262 slope (0.08 and 0.28 ml l<sup>-1</sup>, respectively), defined as the OMZ. Oxygen values began to increase  
263 at the lower slope. Higher values of DO were observed in the basin (Table 1). The OMZ extended

264 from 102 to 1001 m in the study area; its core was located at 525 m, where the lowest DO (0.08  
265 ml l<sup>-1</sup>) value was recorded.

266 The OMZ was also characterised by higher values of sediment Chl-*a* and organic carbon  
267 (Table 1) than were the shelf and basin. A PCA ordination constructed from the eight  
268 environmental factors (DO, sediment chlorophyll, C<sub>org</sub>, CaCO<sub>3</sub>, C:N ratio, clay, silt, and sand  
269 (Fig. 2) showed that the first two components accounted for about 81% of the variability of the  
270 data (PC1 explained 43.5 %, PC2 33.3 %). Two shelf stations were characterised by high coarse  
271 sand and CaCO<sub>3</sub> content, whereas one (at depth 102 m) clustered mainly with the slope stations,  
272 largely because of higher organic-matter content and was clearly separated from the others on the  
273 basis of its higher silt content. Basin samples were characterised by higher DO level (Fig. 2).

274

### 275 3.2 Nematode density and community structure

276 A total of 110 nematode species belonging to 24 families were found along the transect.  
277 Nematode density was higher at the shelf stations than on the slope or in the basin. The lowest  
278 average density was observed in the basin with 62.9±2 ind/10 cm<sup>2</sup>. The mean nematode density  
279 in the slope region was 124.3±16 ind/10 cm<sup>2</sup> and the lowest density was observed on the lower  
280 slope (98.1 ind/10 cm<sup>2</sup>) while upper slope station near to the shelf was represented with the  
281 density of 155.2 ind/10 cm<sup>2</sup> however the mid slope has density of 120.0 ind/10 cm<sup>2</sup> the highest  
282 average nematode density 176.6±37 ind/10 cm<sup>2</sup> was observed at shelf depth.

283 nMDS (non-metric multidimensional scaling) based on nematode species abundance (Bray-  
284 Curtis) and presence/absence data illustrates clearly the extent to which the three zones differ  
285 (Fig. 3 and 4). The nMDS plot based on the Bray-Curtis similarity measure indicated three  
286 groups of samples (Fig. 3). The larger group included the slope stations and the deepest shelf  
287 station—those stations that form the OMZ. The PERMANOVA community results indicated  
288 significant differences between the three zones (p=0.03) but the nMDS based on  
289 presence/absence separated only the slope and basin. The stations on the shelf were distributed  
290 randomly (Fig. 4), and the PERMANOVA community results confirmed insignificant differences  
291 between the different zone (p=0.12). Pair-wise PERMANOVA, comparison between shelf and  
292 deep basin nematode assemblages differ significantly as p=0.046 whereas the pairwise  
293 comparison between slope and shelf, slope and deep basin was not significant (Table 2).

294

295 Thirty-eight species were identified that were found only in the shelf region. Of these,  
296 *Chromadorita* sp 2 (2.78%) and *Sphaerolaimus gracilis* (2.21%) were dominant. *Terschellingia*  
297 *Longicaudata* (4.73%) and *Desmodora* sp 1 (4.42) (3.13%), dominated the slope, and  
298 *Halalaimus sp 1* (1.11%) and *Acantholaimus elegans* (1.11%), dominated the basin. Across the  
299 entire study area, however, the dominant species were *Terschellingia longicaudata*, (15.2 %),  
300 *Desmodora* sp 1 (14.5%), *Sphaerolaimus gracilis* (11.7%), and *Theristus ensifer* (9.1%); their  
301 maximum contributions came from shelf stations (Table 3).

302  
303 Only seven species were found exclusively in the OMZ: *Pselionema* sp 1, *Choanolaimus* sp 2,  
304 *Halichoanolaimus* sp 1, *Cobbia dentata*, *Daptonema* sp 1, *Trissonchulus* sp 1, and *Minolaimus* sp  
305 1. Only nine species were found exclusively in the basin: *Acantholaimus calathus*,  
306 *Acantholaimus mirabilis*, *Sphaerolaimus* sp 1, *Subsphaerolaimus* sp 1, *Pierrickia* sp 2, *Sabatieria*  
307 *praedatrix*, *Epacanthion* sp 1, *Longicyatholaimus* sp 1, and *Oncholaimus attenuatus*. The top 10  
308 dominant species from OMZ from the remaining areas are given in Table 4.

309 The IndVal index produced a list of indicator species for each group of sites: four genera  
310 were indicative of the shelf area and five genera of the slope (Table 5). *Viscosia viscosia* (P =  
311 0.005; stat value=0.751) was a good indicator of the shelf, whereas *Choanolaimus* sp 2 was  
312 strongly and significantly (P = 0.005; stat value = 0.950) associated with the slope.  
313 *Acantholaimus mirabilis* (P = 0.005; stat value=1.000) was significantly associated with the  
314 basin. In addition, a number of indicator species were highly significantly (P = 0.005) associated  
315 with the shelf, slope, or basin. Table 5 lists which species were associated with which zones. The  
316 conditional probability or positive predictive value of the species and the conditional probability  
317 of finding the species at sites belonging to any of the zones are shown in Table 5 for the relevant  
318 species, but those species with the highest IndVal value for the set of all the samples from the  
319 three zones (e.g., *Acantholaimus elegans*, *Acantholaimus filicaudatus*, *Anoplostoma blanchardi*,  
320 *Anoplostoma* sp 1, *Synonchiella* sp 1, *Desmoscolex* sp 1, *Tricoma* sp 1, and *Siphonolaimus*  
321 *ewensis*) were not amenable to statistical testing because of the lack of an external group for  
322 comparison.

323 The three benthic zones differed significantly in the diversity indices (Margalef's d;  
324 Pielou's J' evenness; estimated total number of species, ES (51); and log2 Shannon-Wiener Index  
325 for species diversity, H') of nematode assemblages (PERMANOVA: P < 0.05). The average  
326 values of diversity indices in each zone are given in Table 6.

### 327 3.3 Functional Traits

328 Functional attributes of nematode communities in the Arabian Sea are listed in **Tables 7** and **8**.  
329 The geographic separation was less pronounced in the ordinations derived from functional  
330 characteristics of nematode communities, although the basin and slope samples were clumped  
331 into groups but shelf stations were seen randomly distributed (**Fig.5**). As shown by the results of  
332 pair-wise PERMANOVA, three zones containing nematode assemblages significantly different  
333 (shelf vs deep basin; slope vs deep basin) differ significantly. Assemblages were dominated by  
334 deposit feeders (1B, 1A) followed by predators (2B), whereas proportions of epigrowth feeders  
335 (2A) were comparatively low (**Table 7**). The shelf area was dominated by non-selective deposit  
336 feeders (1B) and predators (2B), whereas the slope area stations showed the highest proportions  
337 (29–48%) of the predators/omnivores (group 2B). Similarly the dominance of predators (2B) was  
338 also seen in OMZ stations covered from the depth of 102 to 1001m. The shelf and basin areas  
339 were dominated by deposit feeders (mainly the non-selective 1B group).

340 Clavate and conical were the most prevalent tail shapes at the majority of stations (**Table**  
341 **7**); the average proportion of conical tails was higher on at the slope and OMZ stations (33–49%).  
342 Clavate tails dominated at all stations, but slightly less in the slope and so in the inside the OMZ  
343 (1001m) than on the shelf and in the basin area. Average adult lengths of nematodes ranged from  
344 1 to 4 mm, and the majority of all recorded individuals occurred in the two intermediate length  
345 classes (1–2 and 2–4 mm). The OMZ stations showed the highest proportion in the 1–2-mm  
346 category (36–62%); the size 2–4 mm was the second dominant (28–39%). Moreover, the basin  
347 was also dominated by these two length categories.

348 The majority of the nematodes were slender, and the proportion decreased with depth  
349 (from 82% in the shelf to 76% in the basin), whereas stout animals accounted for between 2 and  
350 13%. Long/thin animals continuously decreased with increasing depth until the lower slope (at  
351 the core of OMZ), but they were more abundant (17–24%) in the basin than were those of slender  
352 or stout shape. Over 76% of all identified individuals attained a coloniser-persister (C-P) score of  
353 2 to 3. The shelf, slope, basin regions harbored mostly colonisers in the range of 2–3. Extreme  
354 persisters (C-P score of 5) and colonisers were completely absent (**Table 8**).

355 The information in a particular functional group was not a simple reflection of the species  
356 abundance data. Ecological information captured by adult length, adult shape, and life-history  
357 strategy was less site-specific and thus differed notably from information contained in taxonomic  
358 groups.

359 Moreover the biological trait matrix didn't reveal any particular notable relationships  
360 between traits. However certain trends has been observed for example, 1-2 and 2-4mm length  
361 size was generally correlated with slender and colonisers (2-3 C-P score) whereas > 4mm length  
362 size nematodes had higher C-P scores with long/thin body shape. Some other combinations  
363 occurs such as, non-selective feeders with clavate tail shape, category while predators followed  
364 the similar trend as was conical tail shape category (Table 9).

365  
366 The schematic model represents the exact trend of each functional trait at each benthic  
367 zone. The relative abundance of each trait was plotted as area graphs and then a schematic figure  
368 is prepared in order to see the pattern according to the zones. For example, the shelf region favors  
369 the dominance of clavate tail shape, epigrowth feeding, slender body shape, and coloniser life  
370 style with length in the range of 1–2 mm. Similarly the OMZ shows an increasing abundance of  
371 some traits like round tail shape, stout body shape, 2-4mm and greater prevalence of omnivores  
372 and deposit feeders. Coloniser life style (2-3) were increased while persister (4) were dropped  
373 down to lower numbers in OMZ however, in the basin, the sediment characteristics changed from  
374 silty sand to clay. Feeding habit and tail shape also differed in the basin (Fig. 6).

### 378 **3.4 Influence of abiotic parameters on structure and function of nematode community**

379 The RELATE analyses indicated that the pattern based on the environmental variables is  
380 significantly related to the patterns inherent to the nematode community structure both taxonomic  
381 ( $r = 0.37$ ,  $p = 0.01$ ) and functionally ( $r = 0.41$ ,  $p = 0.01$ ). Results of BIOENV analyses revealed  
382 several notable relationships of species abundance data and biological traits with environmental  
383 parameters. For example feeding types have the best correlation with  $C_{org}$ , sediment chl *a*, and  
384 silt. Body size was well correlated with DO, clay, and  $C_{org}$ , whereas tail shape was correlated  
385 with clay, DO, silt, and body length with sand, clay, and C:N ratio. However, C-P score was  
386 significantly correlated with clay and DO (Table 10). Moreover in case of species abundance data  
387 bottom water DO, clay, and  $C_{org}$  were proved to be the best three parameters which correlated  
388 significantly ( $r = 0.712$ ,  $p = 0.02$ ). Of the 10 set of environmental parameters, 6 were retained for  
389 further analysis based on collinearity analysis (Draftsman plot). Only 4 variables salinity,  
390 temperature, water chl-*a* and C:N ratio with correlation  $r^2$  values .0.9 (considered redundant)

391 were omitted for the DISTLM procedures; the remaining variables and their pair-wise spearman  
392 correlations are shown in [Table 11](#).

393  
394 Analyses with distance-based linear models (DistLM) indicated that the 6 abiotic variables  
395 related to the variation in nematode community structure explained 97 % of its total variation,  
396 although not all variables were significant. Similarly for the functional traits of the 8 variables, 6  
397 were used by the DISTLM procedure to construct the best-fitting model, together explaining 96  
398 % of total variation. The DistLM analysis allowed the identification of those abiotic variables that  
399 were best correlated with the observed distribution patterns of taxonomic and functional traits of  
400 nematode species ([Table 11, 12](#)). Results based on the abundance and presence/absence data were  
401 virtually identical; therefore, only the results based on the abundance data and functional traits  
402 are reported. Variables such as the  $C_{org}$  and near-bottom oxygen content showed the highest  
403 correlations ( $P < 0.05$ ) with the nematode assemblages, whereas the functional traits were mainly  
404 explained by with total organic carbon input ( $P < 0.05$ ). [Figs 7, 8](#) shows the DISTLM results by  
405 means of a dbRDA plot, with the species abundance and biological traits superimposed. The  
406 vectors of the environmental variables retained by the DISTLM procedure as fitting the best  
407 explanatory model indicate the important role of OC in the slope area and chl a and sand in the  
408 shelf whereas DO and clay showed their importance to basin.

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## 423 4 Discussion

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### 425 4.1 Nematode density and community structure

426

427 Nematode density was higher in the shelf regions than on the slope or in the basin, but the lowest  
428 density was observed in the basin (2001–2546 m), where oxygen content was highest. Nematode  
429 densities and biomass along the regular slope generally decrease with water depth, surface  
430 primary productivity and distance offshore (Soltwedel, 2000).

431 The role of DO is crucial in structuring meiofaunal communities, and oxygen limitation might  
432 directly control meiofauna composition within the OMZ (Neira et al., 2001). Of the eight stations  
433 presented here, three are true OMZ stations, where DO contents are less than 0.5 ml l<sup>-1</sup>.  
434 Nematode abundance was highest in the center of OMZ (525 m) and decreased to its lowest value  
435 in the lower part of the OMZ (1001 m). Metazoan meiofauna in total and more specific nematode  
436 densities often reach maximum values at lowest oxygen concentrations within OMZs (reviewed  
437 in Levin, 2003).

438 In the present paper, nematode densities were lower to those reported by Cook et al. (2000) for  
439 the bathyal Oman margin (range: 494–2495 ind.10 cm<sup>2</sup>). Moreover they found the highest  
440 abundances inside the OMZ (700 m) not in the center of the OMZ (400 m), however, Levin et al.  
441 (1991) found the highest densities of nematodes (190 ind.10 cm<sup>2</sup>) in the center of the OMZ  
442 (at 745 m), where high concentrations of organic carbon and pigments were found. In our study  
443 the combined OMZ stations showed higher density than non-OMZ stations, perhaps as a result of  
444 the ability of some species to tolerate low-oxygen conditions to take advantage of abundant food  
445 (Levin et al., 1991, Cook et al., 2000). Although data do not exist to identify which, if any, of the  
446 factors shapes the nematode community assemblages, we note that C<sub>org</sub> and near-bottom oxygen  
447 content showed the highest correlations (P < 0.05). Similarly many previous OMZ investigations  
448 have noted a correlation between food quality and meiofaunal abundance and the absence of  
449 oxygen. Like Cook et al. (2000) reported a strong positive correlation between nematode density  
450 and food quality, measured as the hydrogen index, along the transect through the Oman margin  
451 OMZ. Meiofaunal densities correlated only with sediment chlorophyll a concentration, and not  
452 with oxygen or other sediment parameters along a similar OMZ transect on a seamount off  
453 Mexico (Levin et al., 1991).



454 Moreover, in general nematode abundance did not follow any particular gradient though it was  
455 lowest in the deep basin stations. Generally, depth can have a major influence on nematode  
456 density (Udalov et al., 2005; Soltwedel, 2000). The pattern of quantitative distribution of  
457 nematode abundance is described for intertidal and upper subtidal (<50 m) zones, the average  
458 world-wide nematode density is 1530 ind/10 cm<sup>2</sup>, for continental slope (1000–2500 m) it drops to  
459 430 ind/10 cm<sup>2</sup>, for lower slope (2500–3500 m) – 360 ind/10 cm<sup>2</sup>, for abyssal and hadal depths  
460 (>5000 m) this value is still lower (140 ind/10 cm<sup>2</sup>) (Mokievsky et al., 2004; Soltwedel, et al.,  
461 2003). Large-scale comparisons of nematode community structure in other ocean basins indicate  
462 that continental-shelf nematode communities differ significantly from slope communities  
463 (Vanreusel et al., 2010).

464 The nMDS based on nematode species abundance and presence-absence separated the  
465 three zones, and the difference was confirmed with PERMANOVA (p=0.03). Group 1 comprised  
466 the stations of the shelf region, with the exception of the deepest shelf station, which was  
467 clumped with the slope area to form the OMZ stations, where opportunistic species such as  
468 *Pselionema* sp 1, *Choanolaimus* sp 2, *Halichoanolaimus* sp 1, *Cobbia dentata*, *Daptonema* sp 1,  
469 *Trissonchulus* sp 1 and *Minolaimus* sp 1 were found. The exclusive presence of these species in  
470 OMZ may be the result of OMZ sediments that favor the success of nematode species that are  
471 tolerant (colonists) or adapted (endemics). In addition to the seven species found exclusively in  
472 the OMZ a number of species were found exclusively outside it. It is reasonable to speculate that  
473 OMZs have isolated hypoxia-tolerant nematode species due to their different life strategies, small  
474 size, less mobile with no pelagic larval stage. Therefore there could be some species which can  
475 tolerate the oxygen minima while others cannot. However the exact mechanisms of adaptation or  
476 preferences of species to the anoxic environment remains unclear. Several authors have pointed  
477 out that nematode species can successfully cope with anoxic conditions by developing  
478 appropriate physiological and behavioural mechanisms (e.g., symbiosis with bacteria, which we  
479 also observed, and migration to “oxygen islands,” sensu Reise and Ax (1979) and Wetzel et al.  
480 (1995). Mouthless and gutless nematodes carrying endosymbionts have been observed previously  
481 in deep-sea environments (Ingels et al., 2011).

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485 In present study nematode species have not developed any obvious adaptations, but they must  
486 have certain tolerance for anoxic conditions. The dominant species in OMZ like *Terschellingia*  
487 *longicaudata*, *Desmodora* sp 1, and *Sphaerolaimus gracilis*, however, have been recognised  
488 extensively as tolerant (Schratzberger et al., 2006). Adaptations in *Terschellingia* sp and  
489 *Sphaerolaimus* sp—the presence of dark, often multilayered intracellular globules in the  
490 intestinal cells—are often pointed out typical for sulphidic muds. However, their significance is  
491 ambiguous and their adaptive value for the thiobiotic life rather disputed. Moreover, the  
492 deposition of insoluble metal sulphides in intracellular inclusions in *Terschellingia longicaudata*,  
493 has been suggested to be a mechanism of detoxification of sulfide (Nicholas et al., 1987).  
494 Further, some specimens belonging to OMZ showed some morphological differences while some  
495 were observed with epibionts like the greatest numbers of specimens of *Desmodora* sp. Some  
496 species were observed to have unidentified blackish gut content. A small nematode was found in  
497 the gut of the *Metalinhomoeus* sp 1, which has a very small buccal cavity.

498  
499 Even though the majority of the species identified in our study were common and recorded from  
500 each of the three benthic zones, the IndVal index identified some species that can serve as  
501 indicators of the shelf, slope, and basin. The IndVal index is an important tool for ecosystem  
502 monitoring and assessments because it expresses a value for species or genera that can  
503 characterize the particular environment (Dufrene and Legendre, 1997). The genus *Acantholaimus*  
504 (*Acantholaimus mirabilis* significantly  $p = 0.005$  associated with basin) is quite typical of the  
505 deep sea and is rarely found in shallow water; only one species has been described from intertidal  
506 sediments (Platt and Zhang, 1982). The genus *Acantholaimus* increased in relative abundance  
507 with increasing depth in many deep-sea areas (Soetaert and Heip, 1995). Moreover, *Viscosia*  
508 *viscosia* was a good indicator of the shelf because it often inhabits the surface layers of sediment  
509 and is apparently capable of floating (Fonseca-Genevois et al., 2006).

510  
511 The IndVal index reaches its maximum (100%) when individuals of the target species are  
512 observed at all the sites of one group and at no sites of any other (Dufrière and Legendre, 1997).  
513 For those species that were found in equal proportions in each of the three zones, the association  
514 with the set of all the sites cannot be statistically tested, because no external group is available for  
515 comparison, so these species cannot be treated as indicators of a particular habitat but can be  
516 considered cosmopolitan.

517 Species diversity was higher on the shelf than on the slope or in the basin. This difference in  
518 diversity appears to be partly due to the bottom-water DO gradient, which includes values that  
519 are below the oxygen tolerance of many nematode species. Therefore very few species were  
520 present on the slope, especially inside the OMZ. Moreover, the results of a macrofauna study  
521 from this area have shown that different physiographic provinces and an oxygen gradient have a  
522 greater influence on the species composition and diversity than do other oceanographic  
523 conditions (Ingole et al., 2010). The species number and diversity were positively correlated to  
524 DO and negatively correlated with Chl-*a* and C<sub>org</sub>. Species diversity is not always correlated with  
525 the organic enrichment of the sediment; Schratzberger and Warwick (1998) observed a decrease  
526 in species richness possibly due to anoxia and the release of toxic substances under high input of  
527 organic loading. The lower species diversity of the nematode community under hypoxic  
528 conditions is accompanied by high dominance of a very few highly abundant species. Nematode  
529 species especially from OMZs seem able to develop some functional adaptations, as they must  
530 have a certain tolerance for sulphidic and/or anoxic conditions.

531

#### 532 **4.2 Functional Traits**

533 Marine organisms are often challenged by fluctuations in the aquatic environment, which they  
534 must survive, regulate, tolerate, or resist (Odiete, 1999). The different gradient of biological,  
535 chemical, and physical properties in the aquatic environment makes it more complicated. Among  
536 them one of the major challenges is the depletion of DO, but some organisms are able to cope by  
537 means of extreme changes in their body morphology and physiology (Schneider and Bush-  
538 Brown, 2003). In our study the distinct features of the three zones, were also reflected in the  
539 functional-trait approach.

540 Deposit feeders were more abundant in shelf and basin areas than at most of the slope-  
541 area (OMZ) stations, where predators/omnivores were most abundant. Epigrowth feeders were  
542 more abundant at lower shelf stations and at some stations of the OMZ region, probably because  
543 of the higher organic content of the sediment. OMZ often support mats of large sulfur-oxidising  
544 bacteria, including *Thioploca*, *Beggiatoa*, *Thiomargarita* which are often conspicuous features of  
545 the sediment surface there (Levin, 2003). These bacterial mats have been observed previously at  
546 the Oman margin (Levin et al., 1997).

547 Several previous workers have predicted that *Thioploca* mats represent a significant source of  
548 food for the OMZ faunas of the Peru–Chile margin (Gallardo, 1977, Arntz et al., 1991; Levin et

549 al., 2000). Epigrowth feeders have been found to feed on microbiota by scraping them off solid  
550 surfaces or mucus threads with their teeth. Higher amounts of organic matter enriched the growth  
551 of diatoms and ciliates in the OMZ, which can contribute significantly to food for epigrowth  
552 feeders, but in general the proportion of predators/omnivores and epigrowth feeders was low in  
553 the our study. Gambi et al. (2003) suggested that the low prevalence of predatory and omnivorous  
554 nematodes can be attributed to the absence of freshly dead organisms provided they acquire large  
555 teeth and capable of ingesting other animals.

556 The dominance of the deposit feeders in silty sediments of the basin is in agreement with  
557 most previous deep-sea studies (Tietjen, 1984; Jensen, 1988; Netto et al., 2005). Jensen (1988)  
558 found that the deep sea fauna in the Norwegian Sea was dominated by deposit feeders and had  
559 few predators and scavengers. Deposit feeders were also the dominant group in the Puerto Rico  
560 trench and the Hatteras abyssal plain. The selective deposit feeders (microvores) are the main  
561 consumers of deep sea bacteria (Ingels et al., 2011) because deposit-feeding nematodes have  
562 minute buccal cavities, restricting them to small particulate food or dissolved organic matter. The  
563 relative proportion of each of the four Wieser feeding types in a community depends on the  
564 nature of the available food, which may perhaps explain their prominence on exposed substrata  
565 (Platt and Warwick, 1980). BIOENV analysis showed that feeding types correlated well with  
566  $C_{org}$ , sediment chlorophyll, and silt, which represent the substratum condition and feeding  
567 content.

568 In nematodes, the tail plays an important role in locomotion, which in turn depends on  
569 sediment type (Riemann, 1974). Our BIOENV analysis revealed that tail type in nematodes was  
570 significantly correlated with sediment texture and  $C_{org}$ . Nematodes with clavate tail shape  
571 dominated at each depth, although to a greater degree in the shelf and basin regions (Table 3).  
572 Riemann (1974) considered this type of tail morphology to be typical of the inhabitants of the  
573 interstitial spaces in sand. The conical tail shape was observed more frequently in slope regions  
574 than on the shelf; again Riemann (1974) suggested conical tail could be a special adaptation to  
575 fine sand and muddy sediments, where only an incomplete interstitial system exists. The  
576 proportion of long-tail shape and round shape was very less as compared to the above mentioned  
577 categories but the scant availability of data on this aspect prevents comparison and any  
578 generalised conclusion.

579 Vanhove et al. (1995) and Soetaert et al. (2002) noted that length and width are important  
580 functional attributes for describing chemical stress, metabolic rate, the ability to move or migrate,

581 and vulnerability to predation. In fact, size of animals is an integrative feature strongly correlated  
582 with their morphology, locomotion, feeding mode, and other characteristics. Most authors have  
583 related nematode length and width to granulometry, in concordance with the BIOENV analysis  
584 which revealed that body shape was significantly correlated with sand and clay.

585 In our case, slender nematodes were dominant in the shelf region, and their lengths were  
586 mainly in the ranges of 2–4 mm and 1–2 mm. The slope area harbored more long/thin nematodes,  
587 as was the case in earlier studies (Jensen, 1987). Pronounced body elongation in nematodes and  
588 greater surface-volume ratio in thiobiotic species are adaptive characters related to low oxygen  
589 partial pressure and epidermal uptake of dissolved organic matter (Jensen, 1987). Further,  
590 increased length under suboxic or anoxic conditions reflects increased mobility. The increasing  
591 trends of long/thin nematodes from shelf to deeper stations possibly signify that large body size  
592 could facilitate easy burrowing through the sediment, but below the OMZ, the long/thin  
593 nematodes were even more abundant, suggesting the relationship with fine clay and silt  
594 sediments where only an incomplete interstitial system exists. In general comparison, however,  
595 the organisms inhabited in the oxygenated environments are found to be of size and shape a  
596 different from those of organisms from the OMZs (Schneider and Bush-Brown, 2003).

597  
598 Some biological traits, usually less accessible but with direct functional roles (Hodgson et  
599 al., 1999) like life history, are widely used in freshwater and terrestrial habitats (Bongers, 1990).  
600 Over 75% of all identified individuals attained a C-P score of 2 to 3. Animals from the OMZ  
601 represented a higher contribution to this range. Under eutrophic conditions, colonisers are more  
602 numerous, whereas the number of persisters remains constant, but persisters (scores 3–4) were  
603 more often seen in deeper waters, perhaps because most deep-water nematodes are deposit  
604 feeders, which often score 3–4 on the C-P scale (Bongers et al., 1991). Bongers and Ferris (1999)  
605 also distinguished two types of opportunists: enrichment and general opportunists. The genus  
606 *Monhystera* (family Monhysteridae), which was represented with only two species  
607 (*Diplolaimella* sp 1, *Diplolaimelloides* sp 1) in present study were classified as c-p class 2  
608 (general opportunist) (Bongers, 1990).

609 The trait matrix revealed that some combinations of functional groups were significantly  
610 correlated. Stout body shape was positively correlated with clavate, long tail and non-selective  
611 feeding type because these traits showed the same increasing trend from shelf to basin (Fig. 4). A  
612 similar trend was evident for long/thin tail shape with 4 mm and clavate shape with non-selective

613 feeding, which showed very high positive correlation (0.8). In general, large organisms were  
614 more prominent at eutrophic sites, whereas smaller organisms become more dominant in  
615 oligotrophic environments (Thiel, 1975). This pattern clearly explains the abundance of long/thin  
616 nematodes in the slope region of the Arabian Sea. The analysis of a combination of biological  
617 traits seems to be a more reliable approach for assessing the functional structure of nematode  
618 communities than was relying on single functional groups (Schratzberger et al., 2007). Our  
619 schematic model presented the trait responses in different zones (Fig. 4). The outcome confirmed  
620 that functional-trait analysis is of additional ecological importance, and the information captured  
621 by the biological-trait matrix was not a simple reflection of the information contained in  
622 taxonomy. Nematodes with different functional characteristics differ in their abilities to respond  
623 to environmental stresses and disturbance, thereby providing resilience to the community.  
624 Environmental conditions thus influence the importance of functional complementarity in  
625 structuring communities (Hooper et al., 2005).

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#### 628 **4.3 Environmental parameters and their impact on the nematode community**

629 The Arabian-Sea OMZ spreads over 285,000 km<sup>2</sup> and is the second most intense OMZ in the  
630 world tropical ocean (Kamykowski and Zentara, 1990). In this region oxygen concentrations and  
631 organic matter are inversely correlated (Levin and Gage, 1998), and this condition represents a  
632 major challenge for the adaptation of many species in OMZs.

633 Results of BIOENV analysis revealed the best correlation between the functional trait and  
634 individual parameter i.e. the importance of C<sub>org</sub>, sediment chlorophyll, and silt content for feeding  
635 strategy. Organic enrichment is an important ecological process in marine sediments (Kelly and  
636 Nixon, 1984), and it is related to the presence of fine sediment, a low hydrodynamic regime, and  
637 low DO (Snelgrove and Butman, 1994). Nematode assemblages were dominated by deposit-  
638 feeding organisms, which can make full use of both particulate organic matter and the associated  
639 bacterial biomass. Moreover the content of organic matter can partially explain the spatial  
640 patterns of distribution of free-living nematodes in some habitats (Ólafsson and Elmgren, 1997;  
641 Schratzberger et al., 2006). Body size was well correlated with DO, clay, and C<sub>org</sub>, whereas tail  
642 shape was correlated with clay, DO, and silt and body length with sand, clay, and C:N ratio. This  
643 pattern shows the properties of sediment-related factors, which are also important for  
644 morphological characteristics of species. The highest values of sediment C<sub>org</sub> on the slope are

645 probably due to the diverse topographic features there, and the associated hydrodynamic  
646 processes play an important role in the enrichment of  $C_{org}$  (Rao and Veerayya, 2000). The  
647 DistLM analysis indicated that DO concentration and organic content were the only parameters  
648 that were significantly correlated with nematode species and functional data respectively.

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## 652 **5 Conclusions**

653

654 Free-living nematodes are an abundant and diverse component of the meiofauna in the western  
655 continental margin of Arabian Sea. They follow the general trends of decreasing nematode  
656 abundance and diversity with increasing depth observed among other benthic taxa along the  
657 continental shelf and in the deep sea. We aimed to determine the nematode community structure  
658 and functional traits in the Indian western continental margin, including the OMZ. The effects of  
659 heterogeneous environments were more evident in taxonomic groups than in functional traits.  
660 Although numerous factors could be important for the formation and persistence of nematode  
661 communities, the presence of the OMZ seems to be paramount. Along with organic content, DO  
662 plays a vital role in structuring nematode communities and certain functional adaptations among  
663 different species may play an important role in determining the ecosystem function. From our  
664 study, no single parameter can be determined to cause the observed patterns in the three benthic  
665 zones. Our results support the previous finding that, nematodes are the most tolerant taxon of  
666 environmental variation but that within the nematode community only a few species can tolerate  
667 oxygen minima. Our study has provided the first insight into the ecosystem functional diversity  
668 from coastal to deep-sea nematode fauna from the Arabian Sea, but only further work will reveal  
669 whether this pattern is broadly applicable.

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Table 1. Location and depth of sampling stations and sediment texture.

| Depth                  | 34          | 48         | 102        | 525        | 1001           | 1524        | 2001        | 2546        |
|------------------------|-------------|------------|------------|------------|----------------|-------------|-------------|-------------|
| Lat(°N)                | 13°54.26'   | 13°59.88'  | 14°00.29'  | 14°00.24'  | 14°00.25'      | 14°00.30'   | 14°00.09'   | 13°59.55'   |
| Long(°E)               | 74°18.97'   | 74°00.03'  | 73°29.94'  | 73°13.97'  | 73°08.11'      | 72°57.22'   | 71°13.21'   | 70°48.40'   |
| Temperature,<br>°C     | 23.2        | 20.84      | 18         | 10.7       | 7.85           | 5           | –           | –           |
| Salinity<br>(psu)      | 35.8        | 35.5       | 35.1       | 35.3       | 35.2           | 35          | –           | –           |
| DO, ml l <sup>-1</sup> | 0.69        | 0.56       | 0.38       | 0.08       | 0.28           | 1.35        | 2.3         | 2.3         |
| Sediment<br>Chl— µg-l  | 1.4         | 0.2        | 0.6        | 0.7        | 2.1            | 0.6         | 0.6         | 0.2         |
| C <sub>org</sub> (%)   | 1.9         | 0.8        | 1.5        | 3.8        | 4.4            | 2.2         | 0.3         | 0.9         |
| C:N                    | 11.0        | -          | 10.3       | 10.8       | 8.9            | 8.2         | 4.6         | 4.9         |
| Clay (%)               | 8.3         | 9.8        | 3.0        | 12.9       | 13.2           | 8.7         | 15.1        | 15.6        |
| Silt (%)               | 75.6        | 39.7       | 38.5       | 53.6       | 84.5           | 89.4        | 79.2        | 71.7        |
| Sand (%)               | 16.0        | 50.5       | 58.5       | 33.5       | 2.3            | 1.9         | 5.7         | 12.7        |
| Texture                | Clayey silt | Silty sand | Silty sand | Sandy silt | Clayey<br>silt | Clayey silt | Clayey silt | Clayey silt |
| CaCO <sub>3</sub>      | 26.8        | 69.2       | 65.1       | 44.5       | 28.3           | 24.5        | 49.3        | 53.7        |

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1028 Table 2. Results of pair-wise comparison (samples from different zones) PERMANOVA  
 1029 analyses (based on Bray–Curtis similarity measure). Data was fourth-root transformed;  
 1030 resemblance was calculated using Bray-Curtis. Bold values indicate significant differences at  $p <$   
 1031 0.05

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| 1033 | Groups                   | t      | P(perm) | Unique perms | P(MC)        |
|------|--------------------------|--------|---------|--------------|--------------|
| 1034 |                          |        |         |              |              |
| 1035 | Taxonomic Shelf vs slope | 1.7529 | 0.317   | 3            | 0.264        |
| 1036 | Shelf vs Deep basin      | 3.3538 | 0.332   | 3            | <b>0.046</b> |
| 1037 | Slope vs Deep basin      | 6.0733 | 0.352   | 3            | 0.067        |
| 1038 |                          |        |         |              |              |
| 1039 | Functional traits        |        |         |              |              |
| 1040 | Shelf vs slope           | 2.2523 | 0.346   | 3            | 0.181        |
| 1041 | Shelf vs Deep basin      | 5.0419 | 0.354   | 3            | <b>0.013</b> |
| 1042 | Slope vs Deep basin      | 18.508 | 0.342   | 3            | <b>0.028</b> |
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Table 3. Average relative abundances (%) of the most abundant species belonging to each of three benthic zones. Only the top 5 dominant species from each zone are presented.

| Species                         | Shelf (%) | Species                            | Slope (%) | Species                      | Basin (%) |
|---------------------------------|-----------|------------------------------------|-----------|------------------------------|-----------|
| <i>Chromadorita</i> sp 2        | 2.78      | <i>Terschellingia Longicaudata</i> | 4.73      | <i>Halalaimus</i> sp 1       | 1.11      |
| <i>Sphaerolaimus gracilis</i>   | 2.21      | <i>Desmodora</i> sp 1              | 4.42      | <i>Acantholaimus elegans</i> | 1.11      |
| <i>Setosabatieria</i> sp 1      | 2.06      | <i>Theristus ensifer</i>           | 3.13      | <i>Setosabatieria</i> sp 1   | 0.92      |
| <i>Theristus heterospiculum</i> | 1.98      | <i>Sphaerolaimus gracilis</i>      | 2.36      | <i>Anoplostoma</i> sp 1      | 0.72      |
| <i>Synonchiella</i> sp 1        | 1.68      | <i>Halichoanolaimus</i> sp 1       | 2.10      | <i>Daptonema circulum</i>    | 0.53      |

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1077 Table 4. The 10 most abundant species recorded from the oxygen minimum zone (OMZ) and  
1078 from areas outside that zone.

| Species                            | OMZ (%) | Species                         | Outside OMZ (%) |
|------------------------------------|---------|---------------------------------|-----------------|
| <i>Terschellingia longicaudata</i> | 41.5    | <i>Setosabatieria</i> sp 1      | 14.1            |
| <i>Desmodora</i> sp 1              | 37.2    | <i>Chromadorita</i> sp 2        | 11.2            |
| <i>Sphaerolaimus gracilis</i>      | 27.6    | <i>Sphaerolaimus gracilis</i>   | 8.9             |
| <i>Theristus ensifer</i>           | 24.9    | <i>Theristus heterospiculum</i> | 8.3             |
| <i>Halichoanolaimus</i> sp 1       | 15.3    | <i>Synonchiella</i> sp 1        | 7.5             |
| <i>Pterygonema</i> sp 1            | 14.2    | <i>Metasphaerolaimus</i> sp 2   | 6.4             |
| <i>Daptonema</i> sp 1              | 13.1    | <i>Daptonema</i> sp 2           | 6.3             |
| <i>Parasphaerolaimus</i> sp 1      | 12.1    | <i>Siphnolaimus ewensis</i>     | 6.0             |
| <i>Halichoanolaimus robustus</i>   | 11.5    | <i>Axonolaimus</i> sp 3         | 5.8             |
| <i>Trissonchulus</i> sp 1          | 11.5    | <i>Parasphaerolaimus</i> sp 1   | 5.8             |

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1088 Table 5. List of indicator species calculated by IndVal index, number of genera associated with  
 1089 the shelf group =4, number of genera associated to slope groups = 5, number of genera associated  
 1090 with the basin = 7.

|                                 | Association statistic | P value |
|---------------------------------|-----------------------|---------|
| <b>Shelf</b>                    |                       |         |
| <i>Viscosia viscosia</i>        | 0.978                 | 0.005   |
| <i>Oxystomina affinis</i>       | 0.976                 | 0.005   |
| <i>Aegiololaimus</i> sp 1       | 0.973                 | 0.005   |
| <i>Axonolaimus</i> sp 1         | 0.950                 | 0.045   |
| <b>Slope</b>                    |                       |         |
| <i>Choanolaimus</i> sp 2        | 1.000                 | 0.005   |
| <i>Cobbia dentata</i>           | 0.973                 | 0.041   |
| <i>Daptonema</i> sp 1           | 0.951                 | 0.041   |
| <i>Halichoanolaimus</i> sp 1    | 0.942                 | 0.042   |
| <i>Pselionema</i> sp 1          | 0.941                 | 0.042   |
| <b>Basin</b>                    |                       |         |
| <i>Acantholaimus mirabilis</i>  | 1.000                 | 0.005   |
| <i>Acantholaimus calathus</i>   | 1.000                 | 0.005   |
| <i>Sabatieria Ornata</i>        | 1.000                 | 0.005   |
| <i>Theristus heterospiculum</i> | 1.000                 | 0.040   |
| <i>Setosabatieria</i> sp 1      | 0.994                 | 0.030   |
| <i>Metasphaerolaimus</i> sp 2   | 0.994                 | 0.030   |
| <i>Dagda</i> sp 1               | 0.983                 | 0.045   |

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Table 6. Mean and standard error for diversity indices of the nematode communities at species level. d, Margalef's index for species richness; J', Pielou's index for species evenness; ES (51), estimated total number of species; H'(loge), log2 Shannon–Wiener Index for species diversity.

| Zone  | d            | J'          | ES(51)       | H'(loge)    |
|-------|--------------|-------------|--------------|-------------|
| Shelf | 13.53 ± 1.53 | 0.98 ± 0.01 | 40.54 ± 2.57 | 4.04 ± 0.10 |
| Slope | 7.89 ± 0.51  | 0.98 ± 0.01 | 31.69 ± 1.24 | 3.56 ± 0.06 |
| Basin | 7.49 ± 0.11  | 0.97 ± 0.0  | 28.16 ± 0.22 | 3.48 ± 0.01 |



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1114 Table 7. Functional trait matrix showing the percentages of all individuals belonging to four  
1115 nematode feeding categories (1A, no buccal cavity or a fine tubular one, selective deposit  
1116 (bacterial) feeders; 1B, large but unarmed buccal cavity, non-selective deposit feeders; 2A,  
1117 buccal cavity with scraping tooth or teeth, epigrowth (diatom) feeders; 2B, buccal cavity with  
1118 large jaws, predators/omnivores) and four tail-shape categories.

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| Depth | Feeding habits |    |    |    | Tail shapes |         |      |       |
|-------|----------------|----|----|----|-------------|---------|------|-------|
|       | 1A             | 1B | 2A | 2B | Clavate     | Conical | Long | Round |
| 34    | 12             | 46 | 15 | 27 | 62          | 32      | 5    | 1     |
| 48    | 14             | 39 | 20 | 27 | 45          | 41      | 13   | 1     |
| 102   | 17             | 28 | 19 | 36 | 40          | 44      | 15   | 1     |
| 525   | 20             | 17 | 5  | 58 | 44          | 41      | 14   | 0     |
| 1001  | 40             | 28 | 8  | 24 | 28          | 51      | 20   | 0     |
| 1524  | 30             | 19 | 8  | 42 | 31          | 51      | 17   | 1     |
| 2001  | 18             | 34 | 13 | 35 | 49          | 30      | 21   | 0     |
| 2456  | 24             | 33 | 14 | 29 | 40          | 29      | 31   | 0     |

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1134 Table 8. Functional trait matrix showing the percentages of all individuals belonging to four  
1135 body-length categories, three body-shape categories, and five life-history strategies (colonizer-  
1136 persister score).

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| Depth | <1<br>mm | 1–2<br>mm | 2–4<br>mm | >4<br>mm | Slender | Stout | Long/thin | 1 | 2  | 3  | 4  | 5 |
|-------|----------|-----------|-----------|----------|---------|-------|-----------|---|----|----|----|---|
| 34    | 10       | 35        | 51        | 4        | 82      | 3     | 6         | 0 | 49 | 46 | 5  | 0 |
| 48    | 29       | 28        | 24        | 19       | 76      | 2     | 14        | 0 | 34 | 53 | 13 | 0 |
| 102   | 15       | 46        | 31        | 8        | 82      | 13    | 4         | 0 | 33 | 64 | 3  | 0 |
| 525   | 21       | 36        | 40        | 3        | 89      | 3     | 7         | 0 | 15 | 84 | 1  | 0 |
| 1001  | 25       | 37        | 35        | 3        | 83      | 4     | 12        | 0 | 24 | 71 | 5  | 0 |
| 1524  | 28       | 39        | 30        | 3        | 80      | 9     | 11        | 0 | 17 | 76 | 7  | 0 |
| 2001  | 25       | 35        | 27        | 13       | 83      | 0     | 17        | 0 | 27 | 52 | 21 | 0 |
| 2546  | 19       | 36        | 29        | 16       | 76      | 0     | 24        | 0 | 26 | 49 | 26 | 0 |

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Table 9. Biological trait matrix along with contribution (%) of species at each zone

| Species                           | Zones     |           |           | Buccal morphology |    |    |    | Tail shape |     |    |    | Adult Length (mm) |     |     |    | Adult shape |    |     | Life history (c-p score) |   |   |
|-----------------------------------|-----------|-----------|-----------|-------------------|----|----|----|------------|-----|----|----|-------------------|-----|-----|----|-------------|----|-----|--------------------------|---|---|
|                                   | Shelf (%) | Slope (%) | Basin (%) | 1A                | 1B | 2A | 2B | R          | E/F | CO | CA | >1                | 1-2 | 2-4 | >4 | ST          | SL | L/T | 2                        | 3 | 4 |
| <i>Acantholaimus calathus</i>     | 0.11      | 0.00      | 0.00      | 0                 | 0  | 1  | 0  | 0          | 1   | 0  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Acantholaimus elegans</i>      | 0.19      | 0.23      | 1.11      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Acantholaimus filicaudatus</i> | 0.15      | 0.15      | 0.19      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Acantholaimus mirabilis</i>    | 0.31      | 0.00      | 0.00      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 1                 | 0   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Actarjania</i> sp 1            | 0.15      | 0.00      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 0  | 1  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 1                        | 0 | 0 |
| <i>Actinonema</i> sp 1            | 0.31      | 0.00      | 0.00      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Actinonema</i> sp 2            | 0.15      | 0.00      | 0.00      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Actinonema</i> sp 3            | 0.31      | 0.00      | 0.00      | 0                 | 0  | 1  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Aegiololaimus</i> sp 1         | 0.38      | 0.00      | 0.00      | 1                 | 0  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 0 | 1 |
| <i>Anoplostoma blanchardi</i>     | 0.69      | 0.69      | 0.50      | 0                 | 1  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Anoplostoma</i> sp 1           | 0.50      | 0.19      | 0.72      | 0                 | 1  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 0   | 1   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Anoplostoma</i> sp 2           | 0.38      | 0.38      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Araeolaimus</i> sp 1           | 0.19      | 0.04      | 0.00      | 1                 | 0  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Araeolaimus</i> sp 2           | 0.23      | 0.00      | 0.27      | 1                 | 0  | 0  | 0  | 0          | 0   | 1  | 1  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Axonolaimus</i> sp 1           | 1.03      | 0.00      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 1                        | 0 | 0 |
| <i>Axonolaimus</i> sp 2           | 0.15      | 0.04      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 0  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 1                        | 0 | 0 |
| <i>Axonolaimus</i> sp 3           | 1.45      | 0.00      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 0  | 1  | 0                 | 0   | 1   | 0  | 0           | 1  | 0   | 1                        | 0 | 0 |
| <i>Bathyeurstomina</i> sp 1       | 0.11      | 0.00      | 0.38      | 0                 | 0  | 0  | 1  | 0          | 0   | 1  | 0  | 0                 | 0   | 0   | 1  | 0           | 1  | 0   | 0                        | 0 | 1 |
| <i>Campylaimus</i> sp 1           | 0.19      | 0.00      | 0.00      | 0                 | 1  | 0  | 0  | 0          | 0   | 0  | 1  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |
| <i>Ceramonema yunfengi</i>        | 0.46      | 0.00      | 0.00      | 1                 | 0  | 0  | 0  | 0          | 0   | 1  | 0  | 0                 | 1   | 0   | 0  | 0           | 1  | 0   | 0                        | 1 | 0 |

|                               |      |      |      |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |
|-------------------------------|------|------|------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| <i>Cermonema attenuatum</i>   | 0.23 | 0.04 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |   |
| <i>Cheironchus</i> sp 1       | 0.23 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |   |
| <i>Choanolaimus</i> sp 2      | 0.04 | 0.38 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |   |
| <i>Chromadora</i> sp 1        | 0.23 | 0.34 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |   |
| <i>Chromadorita</i> sp 1      | 0.19 | 0.65 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |   |
| <i>Chromadorita</i> sp 2      | 2.78 | 0.19 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Cobbia dentata</i>         | 0.72 | 0.38 | 0.00 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Cobbia</i> sp 1            | 0.34 | 0.00 | 0.00 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Cobbia</i> sp 3            | 0.72 | 0.04 | 0.04 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Cobbia</i> sp 2            | 0.23 | 0.11 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Dagda</i> sp 1             | 0.31 | 0.00 | 0.27 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Daptonema circumum</i>     | 0.84 | 0.31 | 0.53 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Daptonema</i> sp 1         | 0.72 | 1.14 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| <i>Daptonema</i> sp 2         | 1.56 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Desmodora</i> sp 1         | 0.88 | 4.42 | 0.38 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| <i>Desmoscolex</i> sp 1       | 1.30 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 |
| <i>Diplolaimella</i> sp 1     | 0.11 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Diplolaimelloides</i> sp 1 | 0.04 | 0.04 | 0.00 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| <i>Dolicholaimus</i> sp 1     | 0.50 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Doliolaimus</i> sp 1       | 0.00 | 0.00 | 0.11 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Dorylaimopsis</i> sp 1     | 0.15 | 0.00 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Dorylaimopsis</i> sp 2     | 0.15 | 0.46 | 0.00 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 |
| <i>Epacanthion</i> sp 1       | 0.00 | 0.00 | 0.31 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Gammanema</i> sp 1         | 0.08 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 |
| <i>Halalaimus gracilis</i>    | 0.38 | 0.00 | 0.08 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 |
| <i>Halalaimus</i> sp 1        | 0.00 | 0.50 | 1.11 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| <i>Halalaimus</i> sp 2        | 0.00 | 0.23 | 0.53 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| <i>Halalaimus</i> sp 2        | 0.00 | 0.19 | 0.42 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |

|                                    |      |      |      |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |
|------------------------------------|------|------|------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| <i>Halanonchus</i> sp 1            | 0.23 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 |   |
| <i>Halichoanolaimus chordiurus</i> | 0.00 | 0.00 | 0.11 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Halichoanolaimus robustus</i>   | 0.95 | 1.60 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Halichoanolaimus</i> sp 1       | 0.08 | 2.10 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Halichoanolaimus</i> sp 2       | 0.11 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Hopperia</i> sp 1               | 0.95 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Longicytholaimus</i> sp 1       | 0.00 | 0.00 | 0.50 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Marylynnia</i> sp 1             | 0.23 | 0.00 | 0.00 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Mesacanthion</i> sp 1           | 0.19 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 |   | 0 | 1 | 0 | 0 |   | 1 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Metalinhomoeus longiseta</i>    | 0.08 | 0.00 | 0.50 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Metalinhomoeus</i> sp 1         | 0.19 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Metasphaerolaimus</i> sp 1      | 0.00 | 0.00 | 0.38 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Metasphaerolaimus</i> sp 2      | 1.33 | 0.04 | 0.27 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Meyersia minor</i>              | 0.08 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Meyersia</i> sp 1               | 0.23 | 0.15 | 0.23 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Meyersia</i> sp 2               | 0.46 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Microlaimus</i> sp 1            | 0.15 | 0.00 | 0.08 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Minolaimus</i> sp 1             | 0.04 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Oncholaimus attenuatus</i>      | 0.00 | 0.00 | 0.34 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| <i>Oncholaimus mediterraneus</i>   | 0.11 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 |
| <i>Oxystomina affinis</i>          | 0.42 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 |
| <i>Oxystomina</i> sp 1             | 0.61 | 0.08 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| <i>Paralinhomoeus</i>              | 0.11 | 0.08 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |

|                                |      |      |      |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |
|--------------------------------|------|------|------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| <i>filiformis</i>              |      |      |      |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |
| <i>Paralinhomoeus lepturus</i> | 0.34 | 0.84 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 0 |
| <i>Paralinhomoeus</i> sp 1     | 0.57 | 0.76 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Paralinhomoeus</i> sp 2     | 0.42 | 0.00 | 0.08 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 0 |
| <i>Parasphaerolaimus</i> sp 1  | 1.45 | 0.92 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Pierickia</i> sp 1          | 0.23 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <i>Pierrickia</i> sp 2         | 0.00 | 0.00 | 0.11 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <i>Prooncholaimus</i> sp 1     | 0.69 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
| <i>Pselionema hexalatum</i>    | 0.46 | 1.26 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Pselionema</i> sp 1         | 0.11 | 1.11 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Pterygonema Platti</i>      | 0.19 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Pterygonema</i> sp 1        | 0.34 | 1.83 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Richtersia</i> sp 1         | 0.23 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 |   | 0 | 0 | 1 | 0 |
| <i>Sabatieria Ornata</i>       | 0.65 | 0.00 | 0.11 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |   | 1 | 0 | 1 | 0 | 0 |
| <i>Sabatieria praedatrix</i>   | 0.00 | 0.00 | 0.11 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Sabatieria Pulchra</i>      | 0.50 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Sabatieria Punctata</i>     | 0.42 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Sabatieria</i> sp 1         | 0.88 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <i>Sabatieria</i> sp 2         | 0.42 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Sabatieria</i> sp 3         | 1.41 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Setosabatieria</i> sp 1     | 2.06 | 0.00 | 0.92 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Siphnolaimus ewensis</i>    | 1.11 | 0.88 | 0.38 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 |
| <i>Southerniella</i> sp 1      | 1.14 | 0.38 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Sphaerolaimus balticus</i>  | 0.15 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |

|                                    |      |      |      |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |   |
|------------------------------------|------|------|------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| <i>Sphaerolaimus gracilis</i>      | 2.21 | 2.36 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Sphaerolaimus</i> sp 1          | 0.00 | 0.00 | 0.42 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Subsphaerolaimus</i> sp 1       | 0.00 | 0.00 | 0.08 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Synonchiella</i> sp 1           | 1.68 | 0.34 | 0.19 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Terschellingia Longicaudata</i> | 1.18 | 4.73 | 0.04 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Theristus ensifer</i>           | 0.42 | 3.13 | 0.04 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Theristus heterospiculum</i>    | 1.98 | 0.00 | 0.08 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Theristus interstitialis</i>    | 0.92 | 0.00 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Tricoma</i> sp 1                | 0.19 | 0.00 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 |
| <i>Trissonchulus</i> sp 1          | 0.46 | 1.18 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Viscosia</i> sp 1               | 0.04 | 0.00 | 0.34 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Viscosia</i> sp 2               | 0.42 | 0.53 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 |
| <i>Viscosia viscosia</i>           | 0.50 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Voscosia</i> sp 3               | 0.15 | 0.00 | 0.00 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
| <i>Vasostoma</i> sp 1              | 0.57 | 0.57 | 0.00 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| <i>Wieseria</i> sp 1               | 0.23 | 0.31 | 0.00 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |

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1154 1A= selective deposit feeders, 1B= non-selective deposit feeders, 2A=epigrowth feeders, 2B= predators (Wiser, 1953). R= round. E/F=  
1155 elongated/filiform, CO= conical, CA= clavate. ST= stout, L/T= long/thin, C-P= coloniser-persister score (Bongers et al., 1991, 1995.  
1156 Pape et al., 2013), 1= species belonging trait, 0= species not belonging trait.

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1160 Table 10. Results from BIOENV analyses: Spearman rank correlation ( $\rho$ ) and significance level  
1161 (P) between nematode biological traits and environmental variables. Values  $P < 0.05$  are  
1162 significant. DO, dissolved oxygen.

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| Variable      | Environmental parameters              | Global test (Rho) | Significance level of sample statistics (%) |
|---------------|---------------------------------------|-------------------|---|
| Species       | DO, sediment chlorophyll              | 0.785             |   |
| Feeding types | TOC, sediment chlorophyll<br>Clay, DO | 0.44              | 3   |
| Body size     | Sand, Silt, C:N ratio                 | 0.46              | 2   |
| Tail shape    | Clay, bottom DO                       | 0.52              | 1   |
| Body length   | Sand, clay, C:N ratio                 | 0.61              | 7   |
| C-P value     | Clay, bottom DO                       | 0.53              | 8   |

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1178 Table 11. Result of distance-based linear model (DistLM) analyses showing the influence of  
1179 environmental parameters on nematode species abundance data and on functional biological traits  
1180 (Bray-Curtis similarity of square-root-transformed abundance).

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SEQUENTIAL TESTS

| Variable  | R <sup>2</sup> | SS(trace) | Pseudo-F | P     | Prop.    | Cumul.  | res.df |
|-----------|----------------|-----------|----------|-------|----------|---------|--------|
| DO, mll-1 | 0.4647         | 6420.3    | 52.087   | 0.007 | 0.4647   | 0.4647  | 6      |
| Chl µg-1  | 0.50376        | 539.58    | 0.39351  | 0.81  | 3.91E-02 | 0.50376 | 5      |
| Corg (%)  | 0.73435        | 3185.9    | 34.722   | 0.048 | 0.23059  | 0.73435 | 4      |
| Clay (%)  | 0.827          | 1280      | 16.065   | 0.22  | 9.26E-02 | 0.827   | 3      |
| Silt (%)  | 0.89849        | 987.83    | 14.088   | 0.34  | 7.15E-02 | 0.89849 | 2      |
| Sand (%)  | 0.97481        | 1054.4    | 30.302   | 0.254 | 7.63E-02 | 0.97481 | 1      |

MARGINAL TESTS

| Variable  | SS(trace) | Pseudo-F | P     | Prop.   |
|-----------|-----------|----------|-------|---------|
| DO, mll-1 | 321.67    | 2.3209   | 0.068 | 0.27892 |
| Chl µg-1  | 179.35    | 1.1049   | 0.362 | 0.15552 |
| Corg (%)  | 399.65    | 3.1819   | 0.008 | 0.34654 |
| Clay (%)  | 168.03    | 1.0233   | 0.391 | 0.14571 |
| Silt (%)  | 121.46    | 0.70635  | 0.606 | 0.10532 |
| Sand (%)  | 164.27    | 0.99661  | 0.426 | 0.14244 |

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1192 Table 12. Result of distance-based linear model (DistLM) analyses showing the influence of  
 1193 environmental parameters on nematode species functional biological traits (Bray-Curtis similarity  
 1194 of square-root-transformed abundance.

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MARGINAL TESTS

| Variable  | SS(trace) | Pseudo-F | P     | Prop.   |
|-----------|-----------|----------|-------|---------|
| DO, mll-1 | 6420.3    | 5.2087   | 0.013 | 0.4647  |
| Chl aµg-1 | 1424      | 0.68948  | 0.668 | 0.10307 |
| Corg (%)  | 4771.6    | 3.1655   | 0.048 | 0.34537 |
| Clay (%)  | 3635.9    | 2.1429   | 0.13  | 0.26316 |
| Silt (%)  | 1741.9    | 0.86558  | 0.426 | 0.12608 |
| Sand (%)  | 2235.4    | 1.1582   | 0.313 | 0.1618  |

SEQUENTIAL TESTS

| Variable  | R <sup>2</sup> | SS(trace) | Pseudo-F | P     | Prop.    | Cumul.  | res.df |
|-----------|----------------|-----------|----------|-------|----------|---------|--------|
| DO, mll-1 | 0.4647         | 6420.3    | 5.2087   | 0.007 | 0.4647   | 0.4647  | 6      |
| Chl aµg-1 | 0.50376        | 539.58    | 0.39351  | 0.81  | 3.91E-02 | 0.50376 | 5      |
| Corg (%)  | 0.73435        | 3185.9    | 3.4722   | 0.048 | 0.23059  | 0.73435 | 4      |
| Clay (%)  | 0.827          | 1280      | 1.6065   | 0.22  | 9.26E-02 | 0.827   | 3      |
| Silt (%)  | 0.89849        | 987.83    | 1.4088   | 0.34  | 7.15E-02 | 0.89849 | 2      |
| Sand (%)  | 0.97481        | 1054.4    | 3.0302   | 0.254 | 7.63E-02 | 0.97481 | 1      |

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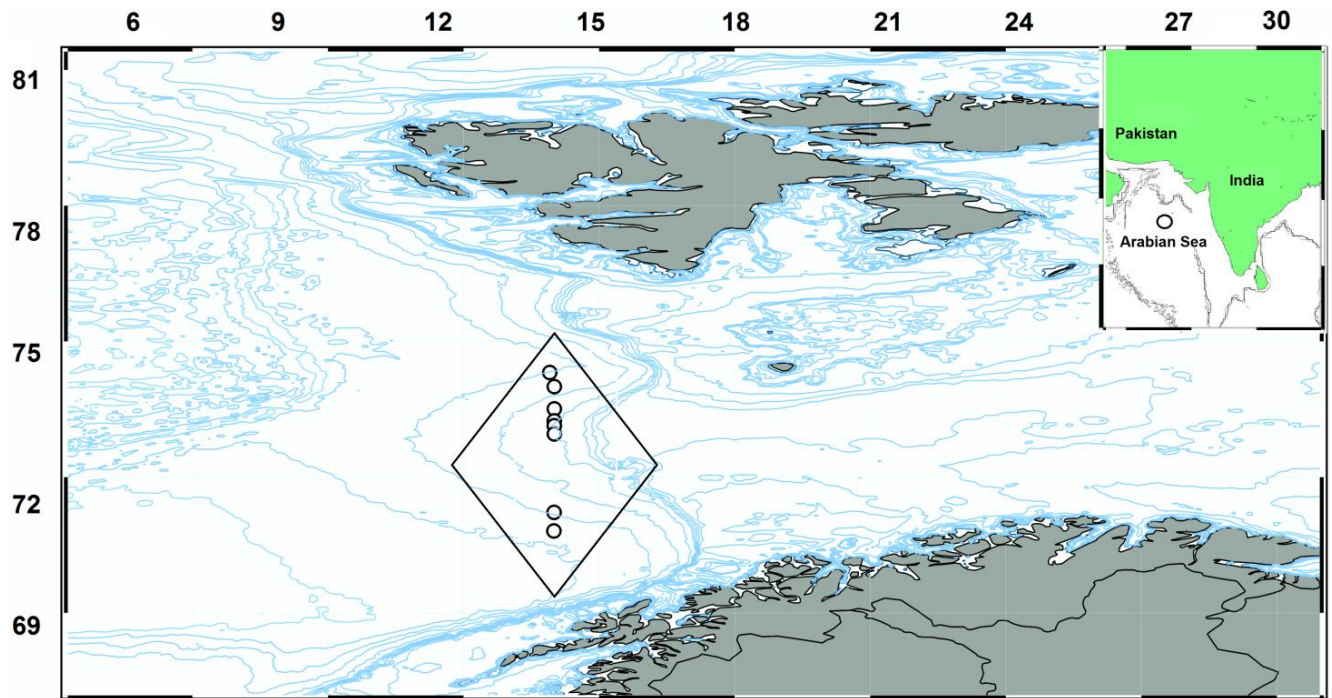
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1211 Figure 1. Map showing location of the 8 sampling stations (in circle) in the coast of Arabian  
1212 Sea along with depth contours and the positions of the sampling stations.

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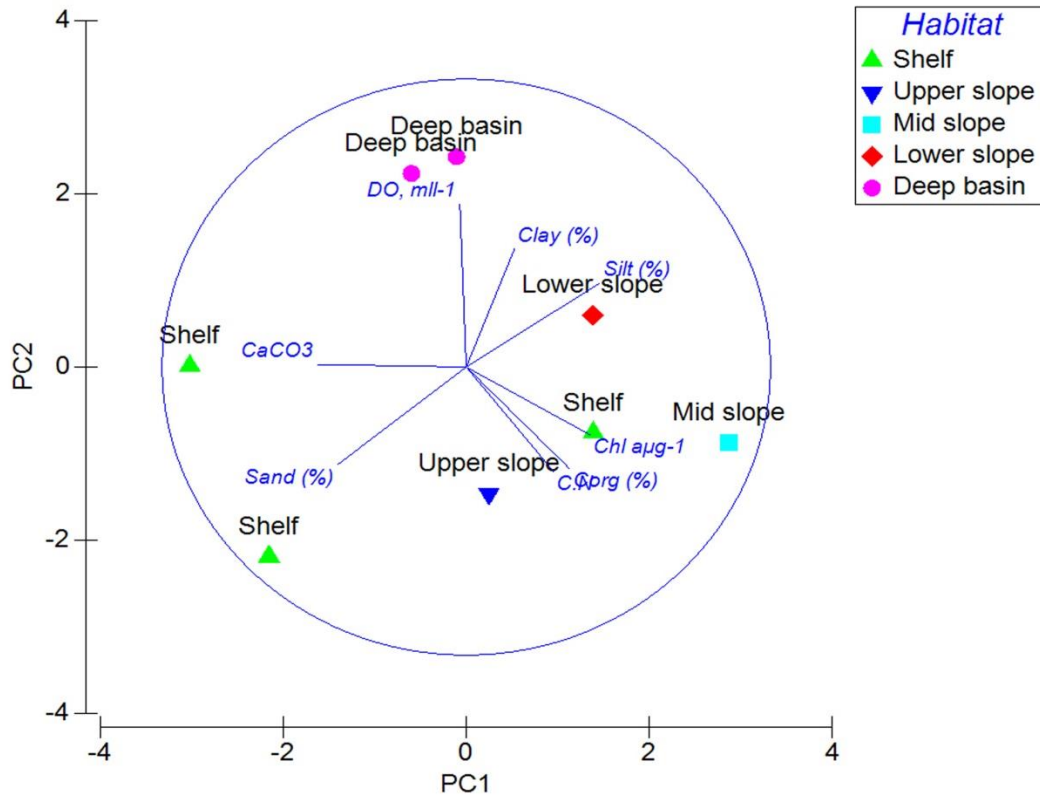
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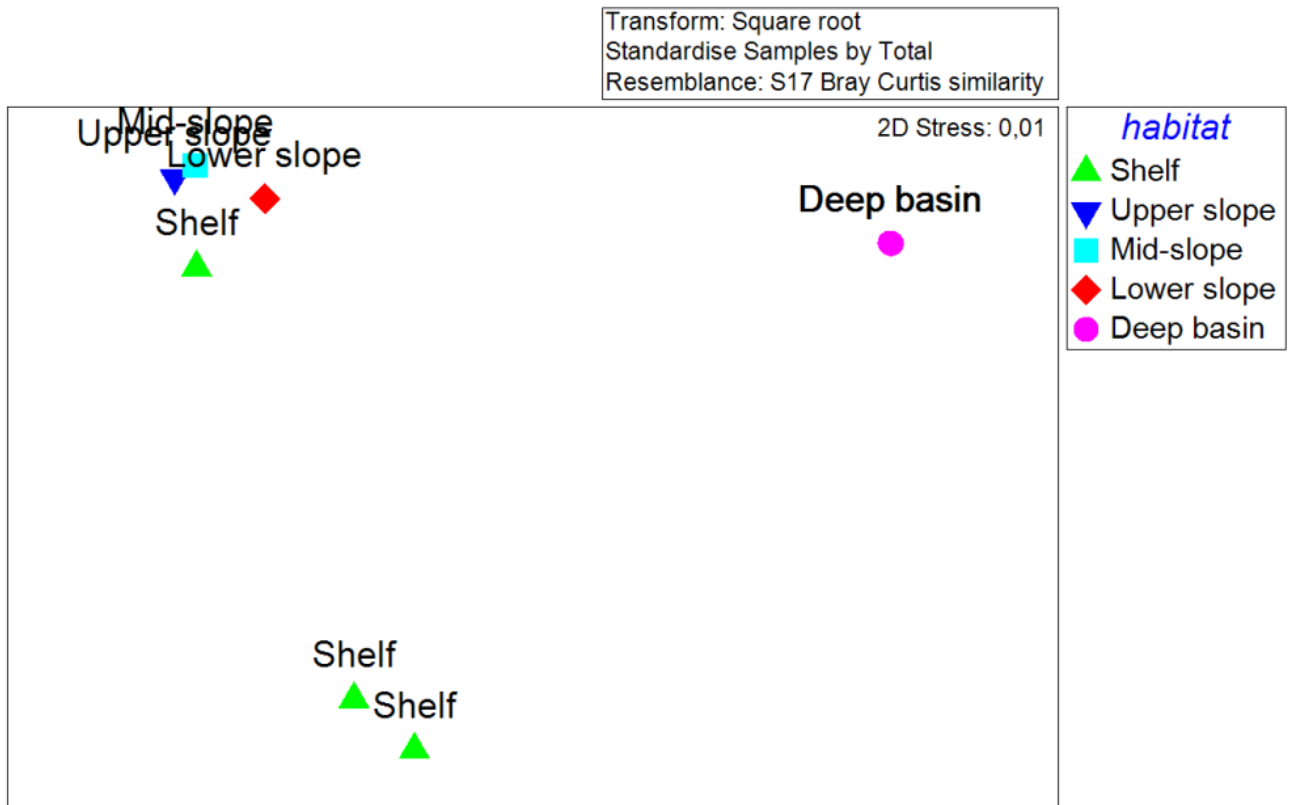
1223  
 1224 Figure 2. Principal component analysis derived from the contribution of parameters in each  
 1225 benthic zone. PC 1 and 2 accounted for 81 % of the total variation present.

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1242 Figure 3. nMDS ordination based on nematode species abundance using the Bray–Curtis  
1243 similarity index.

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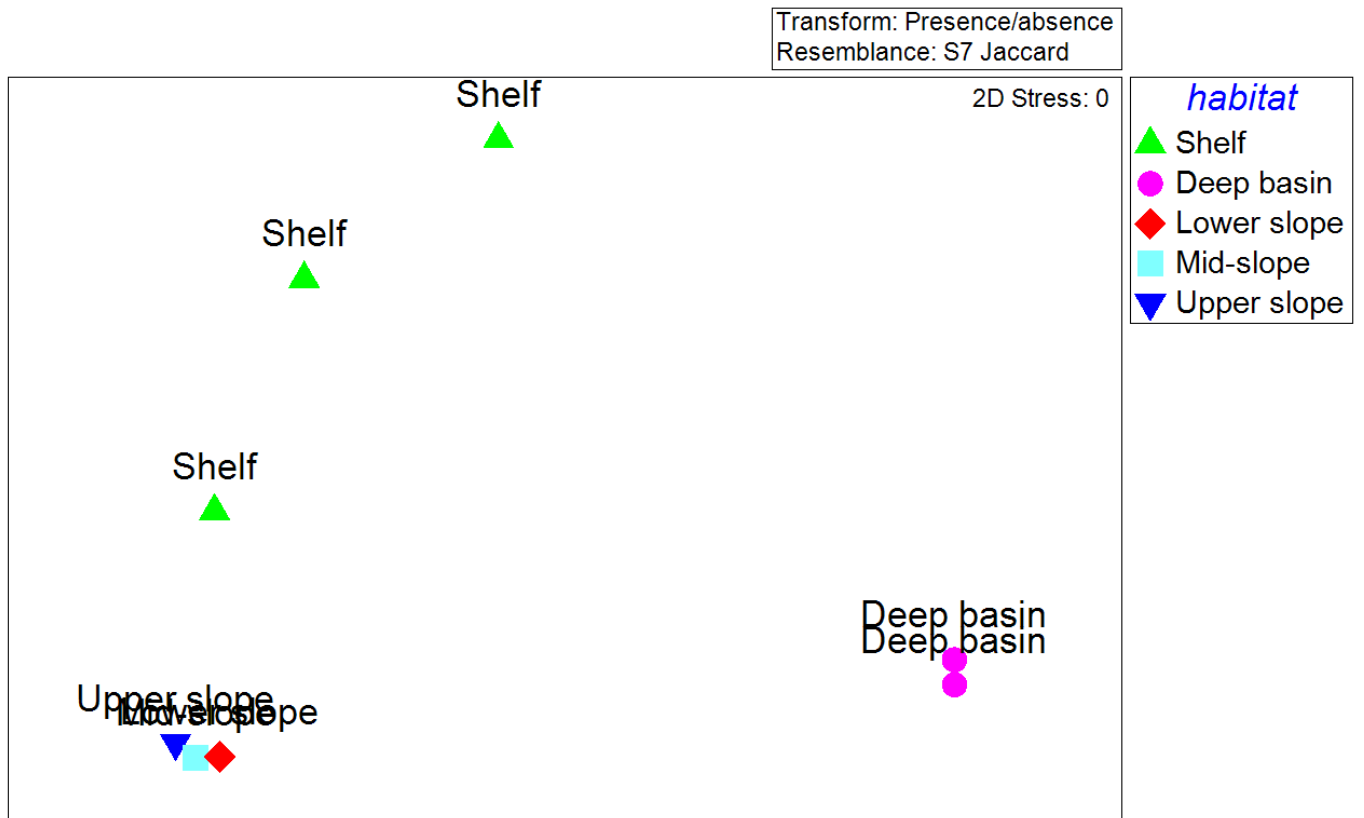
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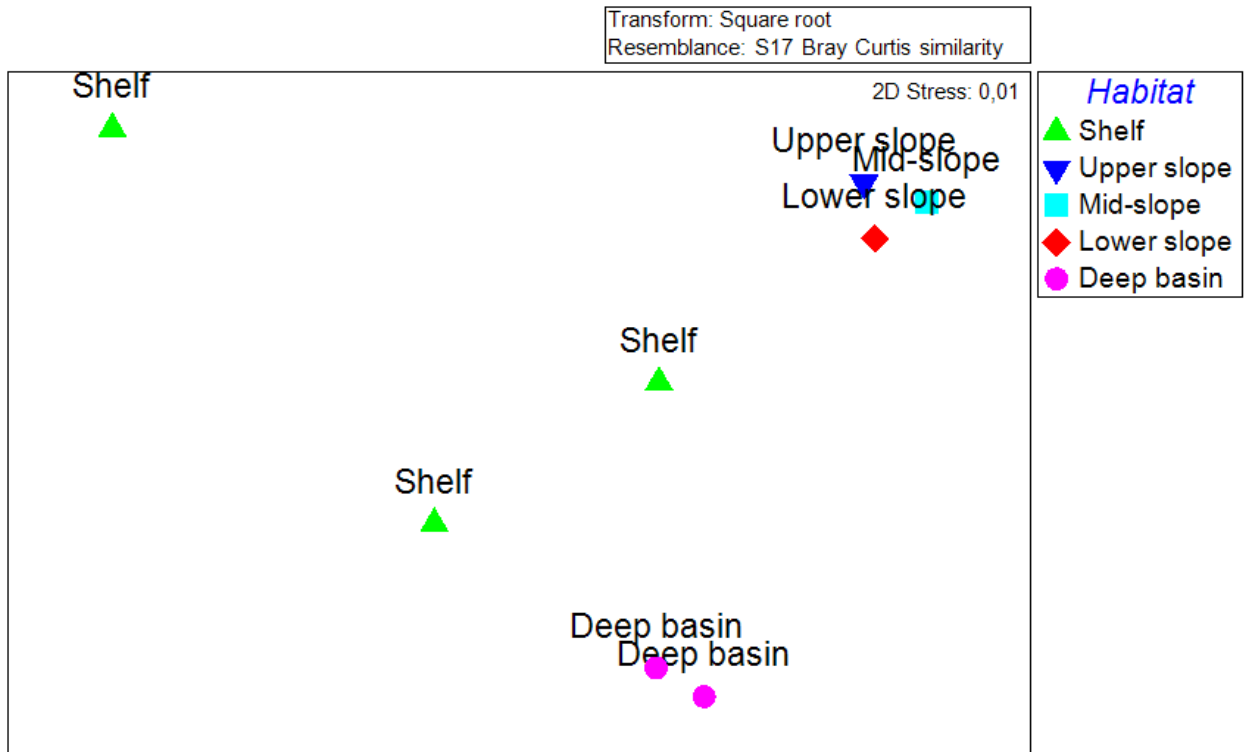
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Figure 4. nMDS ordination based on nematode species presence/absence.

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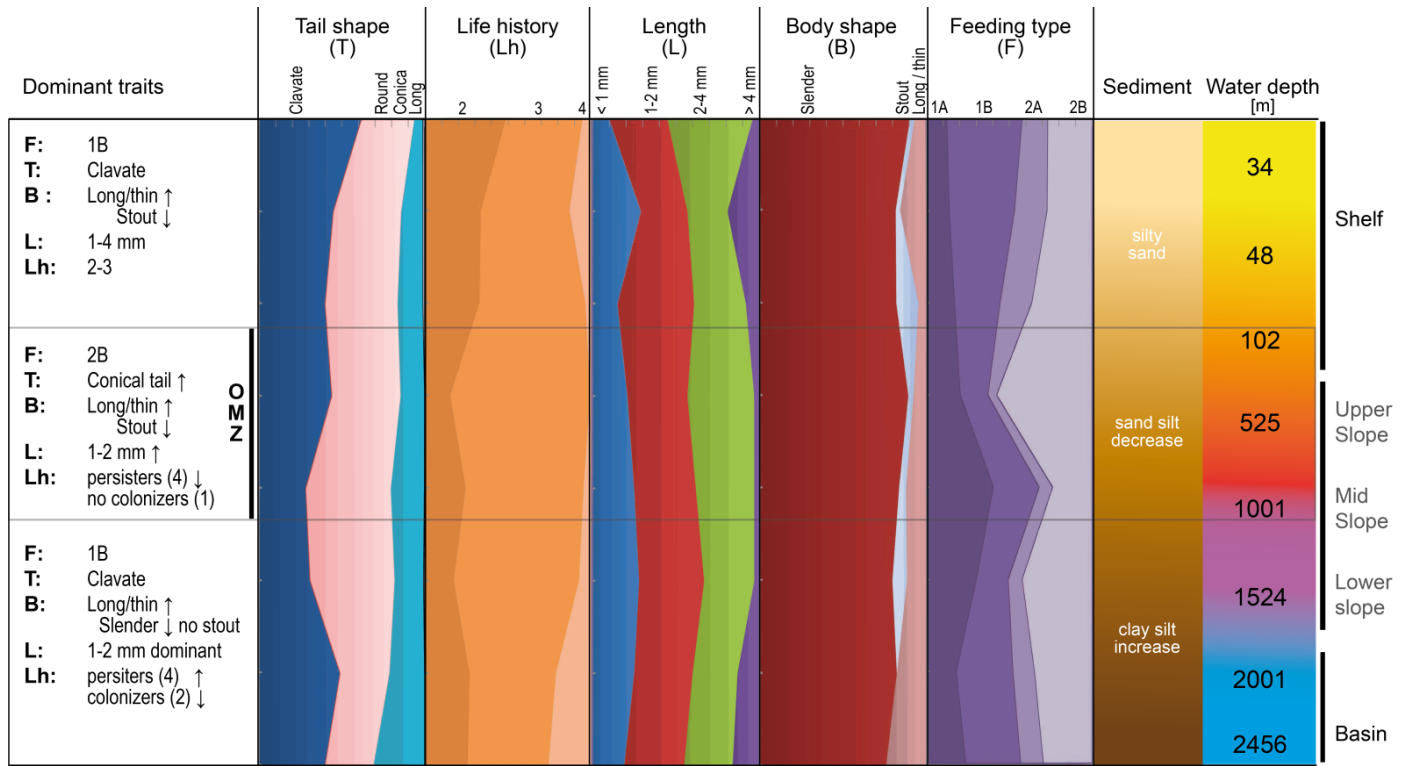


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Figure 5. nMDS ordination based on nematode species biological traits using the Bray–Curtis similarity index.

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Figure 6. Schematic model of the functional traits at each benthic zones

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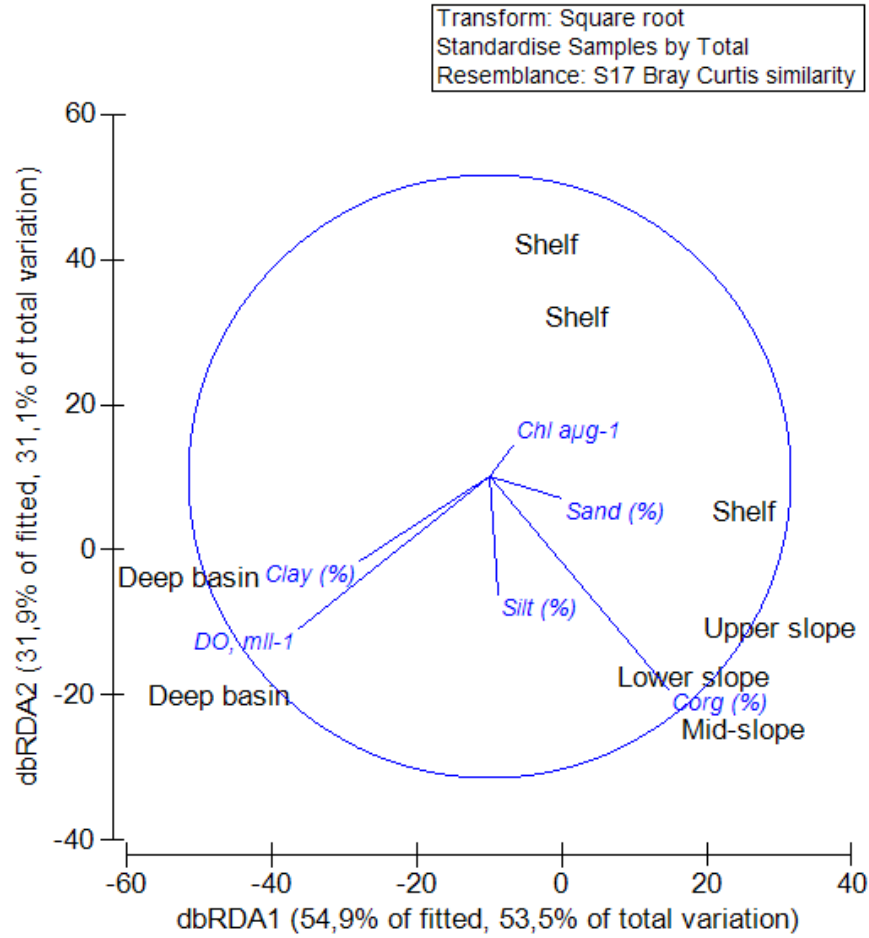
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1296 Figure 7. Distance-based redundancy (dbRDA) bubble plot illustrating the DISTLM model based  
 1297 on the species assemblage data and fitted environmental variables with their vector (strength and  
 1298 direction of effect of the variable on the ordination plot). Axis legends include % of variation  
 1299 explained by the fitted model and % of total variation explained by the axis.

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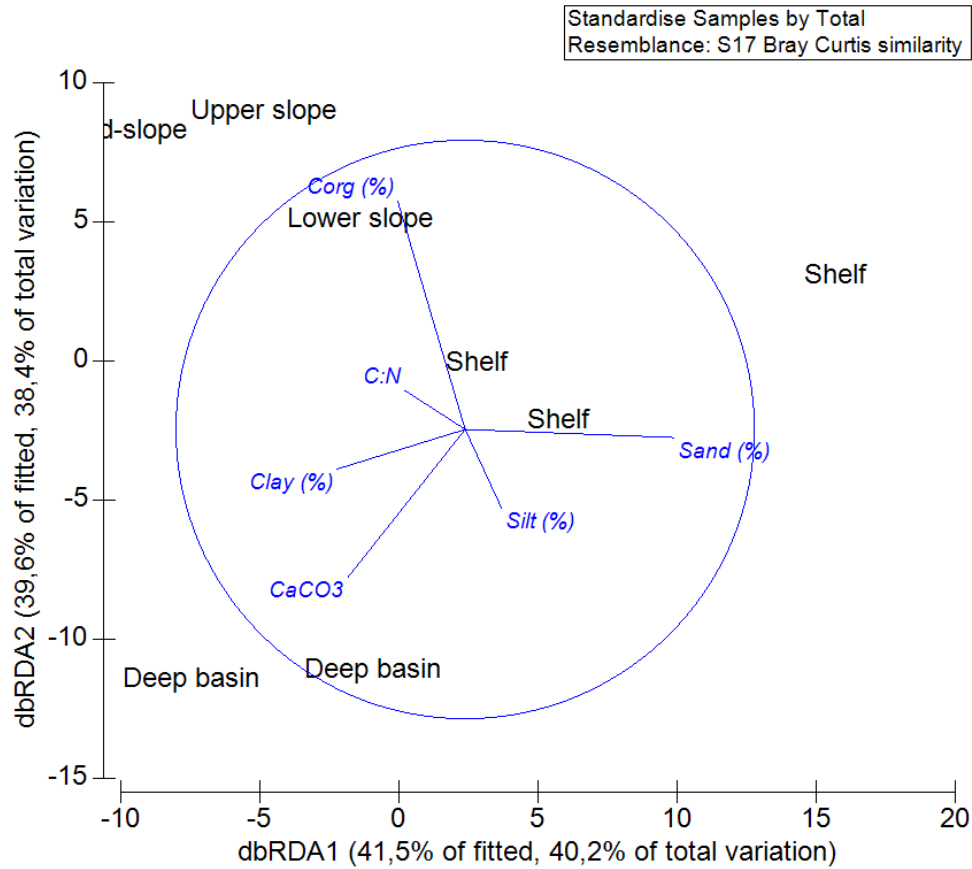
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1309 Figure 8. Distance-based redundancy (dbRDA) bubble plot illustrating the DISTLM model based  
 1310 on the species functional assemblage data and fitted environmental variables with their vector  
 1311 (strength and direction of effect of the variable on the ordination plot). Axis legends include % of  
 1312 variation explained by the fitted model and % of total variation explained by the axis.

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