Review

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Traditional Uses, Chemical Profile and Biological Activities of *Piper hispidum* Sw.: a Review

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Abstract: Piper hispidum Sw. (Piperaceae) (syn. Piper hispidinervum) is a medicinal shrub distributed in Central and South America, widely used as an astringent, diuretic, stimulant for unblocking the liver and stopping hemorrhages. The plant has great interest among researchers due to the production of essential oil and important raw material for the chemical industry, which has a high demand for cosmetic, insecticide, and pesticide industries. In this review, traditional uses, phytochemicals, and biological activities of P. hispidum are comprehensively and systematically summarized through searching scientific databases, including Science Direct, PubMed, Google Scholar, Scopus, and Web of Science. Phytochemical studies revealed the presence of amides, benzoic acids, flavonoids, phenylpropanoids, butenolides, phenol, and essential oils; hence it has several activities, such as antioxidant, antibacterial, α -amylase, insecticidal, schistosomicidal, leishmanial, larvicidal, antiplasmodial, cytotoxicity, estrogenic and serotonergic properties. This review is expected to draw the attention of medical professionals and the general public towards P. hispidum as well as to open the door for detailed research in the future.

Keywords: Piper hispidum; Piper hispidinervum; amide; benzoic acid; essential oil; leishmanial.

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1. Introduction

The Piperaceae family belongs to the Nymphaeiflorae superorder, of the Piperales order, and includes approximately 2500 species distributed in five genera (*Piper*, *Peperomia*, *Zippelia*, *Manekia*, and *Verhuellia*). Piperaceae can be found in herbs, vines, shrubs, and trees more rarely [1-3]. *Piper* is the most economically and ecologically important genus of the Piperaceae family. It is represented by herbs, shrubs, and trees and is widely distributed in the world's tropical and subtropical regions. It consists of a wide variety of species of high economic value, as they are used as food aromas, perfumes, fish venom, insecticides, as well as in the treatment of gynecological and gastrointestinal disturbances, depression, anxiety, pain, and inflammations, as well as bacterial and fungal infections [4-6]. Moreover, *Piper* species are used to treat diseases, including fever, jaundice, rheumatism, and neuralgia, in various countries' folk medicine [7]. Chemical studies have shown that *Piper* has many classes of compounds, such as amides, alkaloids, flavonoids, lignans, neolignans, aristolactams, terpenes, steroids, and phenylpropanoids [8].

Piper hispidum Sw. is a herbaceous plant that grows in tropical and subtropical regions of the world, is widely distributed over a large geographic region, and can be found in Central America, the Andes, and South America. It is one of the plants that is abundantly used because of its medicinal properties [9,10]. Piper hispidinervum C.DC. is a synonym of Piper hispidum Sw., known as "pimenta-longa" in Brazil. This species resembles Piper aduncum L. to some extent but differs in its scarcely scabrous leaves, glabrous stem, and short peduncle. It is distributed throughout South America and is especially prominent in Acre's state in Brazil, and may extend into Amazonas [11].

The current review aims to provide a concise summary of the available information of *P. hispidum* (including *P. hispidinervum*), particularly concerning its traditional uses, phytochemistry, and biological activities. The pharmacological activities and chemical constituents of P. hispidum are also discussed by highlighting its pharmacological potential and identifying this plant as a natural source for potential drug compounds. This review was conducted through searches using Science Direct, PubMed, Google Scholar, Scopus, and Web of Science. The keywords used were "*Piper hispidum*", "*Piper hispidinervum*", "*Piper*", "phytochemistry", "biological activity", and "essential oils" articles over the period from the beginning of the database until May 2020. As a second search strategy, we included studies obtained by a manual search of the included studies' reference lists.

2. Plant Profile

The Plant List includes 88 synonym plant names of *P. hispidum* [12]. It is commonly known as *platanillo-de-cuba* and *bayuyo* (Cuba), *cordoncillo* (Mexico), *jaborandi*, *matico* or *aperta-joão*, and *falso-jaborandi* (Brazil) [13]. *P. hispidum* is a shrub with cylindrical and green stem, which has alternate leaves. The main anatomical characteristics that can be used in its identification are root with sclereids on cortical parenchyma, stem cortex with discontinuous strands of angular collenchyma, and vascular tissue constituted by two discontinuous circles of collateral vascular bundles. The leaf is dorsiventral and hypostomatic with tetracytic stomata. The hypodermis is discontinuous in the adaxial face, loose in the abaxial one, and presents a variable number of layers. Uniseriate epidermis and oil idioblasts occur in all organs [14].

3. Traditional Uses

Previous research has reported the use of *P. hispidum* to ease the pain of childbirth, anemia, and rheumatism in Nicaragua [15]. In addition, the inflorescence is applied topically for muscle aches [16]. The Q'eqchi Maya tribe in Guatemala used this plant to treat female reproductive disorders, including amenorrhea, dysmenorrhea, and menopause [17]. The plant's leaves are also traditionally used by the Chayahuitas, an Amazonian Peruvian ethnic group in Peru, to heal wounds and treat symptoms of cutaneous leishmaniasis [18]. It has also been used as an insecticide, astringent, diuretic, stimulant, liver treatment, and for stopping hemorrhages [19]. In Colombia, the tea of the decoction of *P. hispidum* leaves is useful for the treatment of malaria [20], while in Jamaica, the infusion of leaves in combination with *P. aduncum* is used to treat stomach pains and colds [21].

A leaf infusion is applied to kill head lice in Ecuador, while in Panama, it is used to treat conjunctivitis and diarrhea [22]. Meanwhile, the Totonacs ethnic group from Mexico used to treat mumps and tonsillitis and prevent tooth decay [23]. In South and Central America (particularly Brazil, Colombia, Ecuador, Guatemala, Honduras, Mexico, Panama, and Peru), it

is popularly used for snakebites, insect bites, skin cleansing, head lice, amygdalitis, mouth sores, and teeth whitening agent [24]. Meanwhile, *P. hispidinervum* oil is an important raw source of safrole, a chemical used to synthesize piperonyl butoxide (PBO), a vital ingredient of pyrethroid insecticides [25].

4. Phytochemistry

Up to now, 43 compounds have been reported from *P. hispidum* including eleven amides (1-11) [26-30], seven benzoic acids (12-18) [29,31], sixteen flavonoids (19-34) [30-35], five phenylpropanoids (35-39) [29,31,34], butenolides (40-42) [36], and one phenol (43) [33]. Amides are the predominant secondary metabolite constituents in *P. hispidum*. Their structures, names, plant part, and references are collected in Table 1 and Figure 1.

4.1. Amides.

Eleven amides (1-11) were successfully identified from the roots of P. hispidum. Recently, Lima et al. [26] reported the isolation of four amides (1-4), obtained by supercritical carbon dioxide extraction procedure. Amides (1) and (2) were found as a mixture, as determined by GC-MS. Another study, Alecio et al. [27] was successfully identified a new pyrrolidine amide, N-[7-(3',4'-methylenedioxyphenyl)-2(Z),4(Z)-heptadienoyl]pyrrolidine (5), along with two known amides, identified as N-[5-(3',4'-methylenedioxyphenyl)-2(E)pentadienoyl]-pyrrolidine **(6)** and N-[2-(3',4'-methylenedioxy-6'-methoxyphenyl)-2(Z)propenoyl]-pyrrolidine (7) from the dichloromethane extract of the dried leaves part. Besides, Friedrich et al. [29] and Ruiz et al. [30] managed to isolate N-trans-feruloyltyramine (8) and N-2-(3',4',5'-trimethoxyphenyl)ethyl-2-hydroxybenzamide (11) from the stems and leaves parts of P. hispidum, respectively. Meanwhile, Navickiene et al. [28] were successfully characterized two amides, known as (3Z,5Z)-N-isobutyl-8-(3',4'-methylenedioxy-phenyl)-*N*-[3-(6'-methoxy-3',4'-methylenedioxyphenyl)-2(*Z*)heptadienamide **(9)** and propenoyl]pyrrolidine (10).

4.2. Benzoic acids.

Seven benzoic acids (**12-18**) have been reported phytochemically from *P. hispidum* [29,31]. Friedrich *et al.* [29] were successfully isolated and characterized three new 4-hydroxybenzoic acid derivatives, 4-methoxy-3,5-bis-(3-hydroxy-3-methyl-1-butenyl)benzoate (**12**), 3-hydroxy-2-(1-hydroxy-1-methylethyl)-2,3-dihydrobenzofuran-5-carboxylic acid methyl ester (**13**), and 3-hydroxy-2-(1-hydroxy-1-methylethyl)-2,3-dihydrobenzofuran-5-carboxylic acid (**14**), along with nervogenic acid (**15**), nervogenic acid methyl ether (**16**), 2,2-dimethyl-8-(3-methyl-2-butenyl)-2*H*-chromene-6-carboxylic acid (**17**), and 4-hydroxy-3-(3-methyl-2-butenyl)benzoate (**18**) from the methanolic extract of the stems of *P. hispidum*.

4.3. Flavonoids.

Fourteen flavonoids (**19-34**) have been isolated phytochemically from *P. hispidum* [30-34]. It comprises six flavanones (**19-24**) and nine chalcones (**25-34**). Erika *et al.* [32] were successfully characterized two known flavonoids, identified as 5-hydroxy-7-methoxyflavanone (**20**) and 5-hydroxy-4',7-dimethoxyflavanone (**21**) from the ethanolic extract of the inflorescences of *P. hispidum*. In another study, Vieira *et al.* [33] were reported 6-hydroxy-5,7-dimethoxyflavanones (**23**), and

5,7,8-trimethoxyflavanones (**24**) from the Brazillian leaves extract. In the case of chalcones, Costa *et al.* [35] managed to isolate three compounds, known as 2'-hydroxy-4,4',6'-trimethoxychalcone (**31**), 2'-hydroxy-3,4,4',6'-tetramethoxychalcone (**32**), and 2',3-dihydroxy-4,4',6'-trimethoxychalcone (**33**) from the same part. In the meantime, 2',4',6-trimethoxychalcone (**25**), 2',6'-dihydroxy-4'-methoxychalcone (**27**), and 2'-hydroxy-3',4',6'-trimethoxychalcone (**28**) have been isolated from the inflorescences, fruits and leaves part of *P. hispidum*, respectively [30-33].

Table 1. Phytochemicals identified from *P. hispidum* and bioactivities.

No	Compounds	Part	Bioactivities	Ref
	AMIDES		Diometricular	
1	(Z)-3-(6-methoxybenzo[d][1,3]dioxol-5-yl)-1- (pyrrolidin-1-yl)prop-2-en-1-one	Leaves		[24]
2	(E)-3-(6-methoxybenzo[d][1,3]dioxol-5-yl)-1-(pyrrolidin-1-yl)prop-2-en-1-one	Leaves	Cytotoxicity: Inhibition of 9.19% (HepG2) and 62.11% (HL-60)	[24]
3	(2Z,4E)-7-(benzo[d][1,3]dioxol-5-yl)-1- (pyrrolidin-1-yl)hepta-2,4-dien-1-one	Leaves		[24]
4	(2E,4Z)-7-(benzo[d][1,3]dioxol-5-yl)-1- (pyrrolidin-1-yl)hepta-2,4-dien-1-one	Leaves		[24]
5	<i>N</i> -[7-(3',4'-methylenedioxyphenyl)-2(<i>Z</i>),4(<i>Z</i>)-heptadienoyl]pyrrolidine	Leaves	Antifungal: MIC value 8.0 µg against Cladosporium sphaerospermum	[25]
6	<i>N</i> -[5-(3',4'-methylenedioxyphenyl)-2(<i>E</i>)-pentadienoyl] pyrrolidine	Stems	Antifungal: MIC value 5.0 µg/mL against Cladosporium sphaerospermum	[26]
	1 7317	Leaves		[25]
7	<i>N</i> -[2-(3',4'-methylenedioxy-6'-methoxyphenyl)-2(Z)-propenoyl]-pyrrolidine	Leaves		[25]
8	N-trans-feruloyltyramine	Stems		[27]
9	(3Z,5Z)-N-isobutyl-8-(3',4'-methylenedioxy-phenyl)-heptadienamide	Stems	Antifungal: MIC value 5.0 µg/mL against Cladosporium sphaerospermum	[26]
10	N-[3-(6'-methoxy-3',4'-methylenedioxyphenyl)-2(Z)-propenoyl]pyrrolidine	Stems	Antifungal: MIC value 5.0 μg/mL against <i>Cladosporium sphaerospermum</i>	[26]
11	<i>N</i> -2-(3',4',5'-trimethoxyphenyl)ethyl-2-hydroxybenzamide	Leaves		[28]
	BENZOIC ACIDS			
12	4-Methoxy-3,5-bis-(3-hydroxy-3-methyl-1-butenyl)benzoate	Stems		[27]
13	3-Hydroxy-2-(1-hydroxy-1-methylethyl)-2,3- dihydrobenzofuran-5-carboxylic acid methyl ester	Stems		[27]
14	3-Hydroxy-2-(1-hydroxy-1-methylethyl)-2,3- dihydrobenzofuran-5-carboxylic acid	Stems		[27]
15	Nervogenic acid	Stems		[27]
16	Nervogenic acid methyl ether	Stems		[27]
	•	Fruits		[29]
17	2,2-Dimethyl-8-(3-methyl-2-butenyl)-2H-chromene-6-carboxylic acid	Stems		[27]
18	4-Hydroxy-3-(3-methyl-2-butenyl)benzoate FLAVONOIDS	Stems		[27]
19	5,7-Dihydroxyflavanone	Leaves		[28]
20	5-Hydroxy-7-methoxyflavanone	Fruits		[29]
		Inflores cences		[30]
21	5-Hydroxy-4',7-dimethoxyflavanone	Inflores cences		[30]
22	6-Hydroxy-5,7-dimethoxyflavanones	Leaves		[31]
23	8-Hydroxy-5,7-dimethoxyflavanones	Leaves		[31]
24	5,7,8-Trimethoxyflavanones	Leaves		[31]
25	2',4',6-Trimethoxychalcone	Inflores cences		[30]
26	2',4',6-Trihydroxychalcone	Leaves		[32]
27	2',6'-Dihydroxy-4'-methoxychalcone	Fruits		[29]
28	2'-Hydroxy-3',4',6'-trimethoxychalcone	Leaves	Antileishmanial: IC ₅₀ values of 0.8 μM against <i>Leishmania amazonensis</i> Cytotoxicity: IC ₅₀ values of 1.6 μM against peritoneal macrophages	[28]

No	Compounds	Part	Bioactivities	Ref
		Branch		[31]
29	2',4'-Dihydroxy-6'-methoxychalcone	Leaves	Antileishmanial: IC ₅₀ values of 8.0 μM	[28]
			against Leishmania amazonensis	
			Cytotoxicity: IC ₅₀ values of 18.2 μM against	
			peritoneal macrophages	
30	2',3'-Dihydroxy-4',6'-dimethoxychalcones	Branch		[31]
31	2'-Hydroxy-4,4',6'-trimethoxychalcone	Leaves	Antimicrobial: MIC value of 125 and 250	[33]
			μg/mL against S. aureus and C. albicans	
32	2'-Hydroxy-3,4,4',6'-tetramethoxychalcone	Leaves	Antimicrobial: MIC value of 250 and 500	[33]
			μg/mL against S. aureus and C. albicans	
33	2',3-Dihydroxy-4,4',6'-trimethoxychalcone	Leaves	Antimicrobial: MIC value of 125 and 250	[33]
			μg/mL against S. aureus and C. albicans	
34	2',4,6'-Trihydroxy-4'-methoxychalcone	Leaves	Antiplasmodial: IC ₅₀ values of 16.9 μg/mL	[32]
			(poW) and 10.4 μg/mL (Dd2) active against	
			both a chloroquine-sensitive and a resistant	
			strain of Plasmodium falciparum	
	PHENYLPROPANOIDS			
35	ω-Hydroxyisodillapiole	Stems	Cytotoxicity: IC ₅₀ values of 31.8 µg/mL	[27]
			against human bladder carcinoma cell line	
			ECV-304	
36	Dillapional	Stems	Cytotoxicity: IC ₅₀ values of 31.7 μg/mL	[27]
			against human bladder carcinoma cell line	
			ECV-304	
37	Dillapiole aldehyde	Stems	Cytotoxicity: IC ₅₀ values of 31.1 μg/mL	[27]
			against human bladder carcinoma cell line	
			ECV-304	
38	Dillapiole	Leaves		[32]
39	1-Allyl-2,3-(methylenedioxy)-4,5-dimethoxy-	Fruits		[29]
	benzene			
	BUTENOLIDES			
40	9,10-Methylenedioxy-5,6-Z-fadyenolide	Leaves	Estrogenic and serotonergic: Bound to the	[34]
			serotonin receptor 5-HT7 with IC50 values	
			of 16.1 and 8.3 μM, respectively, and using	
			GTP shift assays, it was found to be a partial	
44	5 (G P)		agonist of the 5-HT7 receptor	50.43
41	5,6-Z-Fadyenolide	Leaves		[34]
42	Piperolide	Leaves		[34]
	PHENOL			
43	4-(5' <i>E-n</i> -hexadecenyl)-phenol	Leaves		[31]

4.4. Phenylpropanoids.

Friedrich *et al.* [29] managed to isolate three phenylpropanoids, identified as ω -hydroxyisodillapiole (**34**), dillapional (**35**), and dillapiole aldehyde (**36**), from the part of the stem. Meanwhile, a long time ago, Burke and Nair, [31] reported the isolation of 1-allyl-2,3-(methylenedioxy)-4,5-dimethoxy-benzene (**39**) from the fruits of *P. hispidum*.

4.5. Butenolides.

Three butenolides, including one new compound, 9,10-methylenedioxy-5,6-Z-fadyenolide (**40**) were isolated from the leaves of *P. hispidum* collected from Guatemala [36]. Other butenolides were 5,6-Z-fadyenolide (**41**) and piperolide (**42**).

4.6. Phenol.

4-(5'E-n-hexadecenyl)-phenol (43) was the only phenol found in the leaves of P. hispidum, as reported by Vieira et al. [33].

Figure 1. Chemical structures of isolated compounds from *P. hispidum*.

4.7. Essential oils.

Essential oils are complex mixtures of volatile compounds, mainly terpenes and oxygenated aromatic and aliphatic compounds, such as phenols, alcohols, aldehydes, ketones, esters, lactones, coumarins, ethers, and oxides, biosynthesized and accumulated in many plants. These naturally occurring mixtures of volatile compounds have been gaining increasing interest because of their wide range of applications in pharmaceutical, sanitary, cosmetics, perfume, food, and agricultural industries. Several significant biological activities are attributed to essential oils, such as allelopathic, antibacterial, antifungal, antioxidant, anti-inflammatory, and anticancer activities [37-48].

The chemical compositions of *P. hispidum* essential oils have been reported from various origins including Brazil [49-56], Colombia [57-59], Cuba [60], France [61], Guatemala

[62], Panama [63], and Venezuela [64]. Most of the essential oils successfully identified monoterpenes as the major components, which are γ -terpinene, β -pinene, α -pinene, limonene, δ-3-carene, and α -copaene. In addition, sesquiterpenes were identified as the most dominant component, which are β -caryophyllene, γ -cadinene, curzerene, and germacrene D. Besides, oxygenated sesquiterpenes characterized as *trans*-nerolidol, β -eudesmol, and spathulenol have been reported from *P. hispidum* essential oils collected from Colombia, Cuba, and Guatemala, respectively. *P. hispidum* essential oils also presented a phenylpropanoid, dillapiole. Its richness in the leaves and roots part was collected from Brazil and Panama. Table 2 summarizes the data on the major components and biological activities of *P. hispidum* (and *P. hispidinervum*) [65-68] essential oils.

Table 2. Components identified from *P. hispidum* and *P. hispidinervum* essential oils and bioactivities.

Country	Parts	Total		Major components (%)	Bioactivities	Ref
_		No	%	-		
Brazil	Leaves	32	99.7	γ-Terpinene (30.9%), α-terpinene (14.4%), <i>p</i> -cymene (12.1%), α-selinene (9.0%), β-selinene (8.1%)	Antifugal: MIC and MFC values of 312.5 and >1250 μg/mL against <i>Rhizopus</i> oryzae	[15]
Brazil	Leaves	33	92.8	Khusimene (12.1%), β-pinene (12.0%), γ-cadinene (13.2%), ledol (8.8%)	Toxicity: LC ₅₀ value of 404.8 μg/mL against <i>Artemsia salina</i>	[49]
Brazil	Aerial parts	61	88.4	β-Caryophyllene (10.5%), α-humulene (9.5%), α-copaene (7.3%), limonene (6.9%), caryophyllene oxide (5.9%)	Cytotoxicity: IC ₅₀ value of >25 μg/mL against HCT-116, SKMEL19, and ACP03 cell lines Antifungal: Detection limit (DL) of 0.1 μg/mL (Cladosporium cladosporioides) and 1.0 μg/mL (Cladosporium sphareospermum) Antioxidant: Inhibition of 26.4% (DPPH); TEAC (303.1 mg ET/mL) AChE: Detection limit (DL) of 0.01 μg/mL	[50]
Brazil	Roots	8	99.9	Dilapiole (57.5%), elemicin (24.5%), apiol (10.2%)	(BE) 01 0:01 µg/mE	[20]
Brazil	Leaves	11	97.2	γ-Cadinene (25.13%), camphene (15.61%), α-guaiene (11.47%), γ-elemene (10.88%)		[51]
Brazil	Leaves	38	99.0	γ-Terpinene (27.3%), p-cymene (14.0%), α-terpinene (12.0%), α-selinene (8.4%), β-selinene (7.5%), terpinolene (6.5%)	Antimicrobial: MIC value of 937.5 µg/mL against Aeromonas hydrophila	[52]
Brazil	Leaves	36	75.2	β-pinene (9.76%), α-pinene (6.90%), (<i>E</i>)-nerolidol (6.30%), γ-cadinene (5.3%), δ-cadinene (5.0%)	,	[53]
Brazil	Fruits	99	70.0	limonene (16.3%), β-pinene (14.5%), α-pinene (13.5%), linalool (9.6%), α-terpineol (8.5%), 1,8-cineol (5.1%)		[54]
Brazil	Ripe fruits	47	98.1	α-Copaene (36.2%), β-pinene (7.5%), α-pinene (7.1%), (<i>E</i>)-nerolidol (7.0%)		[55]
	Unripe fruits	44	97.8	α-copaene (28.7%), α-pinene (13.9%), β-pinene (13.3%)		
Brazil	Fresh leaves	17	95.1	Germacrene D (33.9%), δ-3-carene (17.4%), (<i>E</i>)-caryophyllene (13.8%), bicyclogermacrene (7.1%)		[56] _
	dried leaves	21	94.1	Germacrene D (31.0%), δ -3-carene (19.1%), (E)-caryophyllene (14.9%), bicyclogermacrene (6.2%)	Cytotoxicity: Inhibition of 62.8% (HepG2) and 70.2% (HL-60) Larvicidal: LC ₅₀ value of 141.9 µg/mL against Aedes aegypti	

Country	Parts			Major components (%)	Bioactivities	Ref
•		No % 17 85.4				_
	Stems			Germacrene D (18.8%), (<i>E</i>)-caryophyllene (14.2%), δ-cadinene (10.6%), bicyclogermacrene (9.3%), α-ylangeno (6.5%), α-muurolol (6.2%)	Cytotoxicity: Inhibition of 47.7% (HepG2) and 16.3% (HL-60)	
Colombia	Aerial parts	44	99.4	trans-Nerolidol (23.6%), caryophyllene oxide (5.4%), β-elemene (5.1%), trans-β-caryophyllene (5.1%)	Antifungal: MIC values of 99 μg/mL (<i>T. rubrum</i>) and 125 μg/mL (<i>T. mentagrophytes</i>) Cytotoxicity: IC ₅₀ value of 51.7 μg/mL against Vero cell line	[57] [58]
Colombia	Leaves	17	87.5	δ-3-Carene (9.6%), p-cymene (10.9%), limonene (17.2%), elemol (14.1), γ-elemene (7.3%), β-eudesmol (5.7%)	Antifeedant: EC ₅₀ value of 48.0 µg/mL against Spodoptera littoralis Phytotoxic: Percentage control of 94.5% against Lolium perenne	[59]
Cuba	Leaves	25	93.5	β-eudesmol (17.5%), trans-6-vinyl- 4,5,6,7-tetrahydro-3,6-dimethyl-5- isopropenylbenzofuran (12.9%), γ- eudesmol (9.3%), α-eudesmol (8.1%), elemol (7.6%)	•	[60]
France	Leaves	64	90.5	Curzerene (15.7%), germacrene B (10.9%), α-selinene (10.5%), β-selinene (7.6%)	Antidermatophytic: MIC value of 62 μg/mL against Trichophyton mentagrophytes Antileishmanial: IC50 value of 3.4 μg/mL against Leishmania amazonensis Cytotoxicity: TD50 value of 35.5 μg/mL against BALB/c mice peritoneal macrophages	[61]
Guatemala	Leaves	23	74.9	Spathulenol (8.7%), β-caryophyllene (6.9%), (<i>E</i>)-nerolidol (6.8%), germacrene D (6.0%), caryophyllene oxide (5.9%)		[62]
Panama	Leaves	43	95.1	Dillapiole (57.7%), piperitone (10.0%)	Larvicidal: LC ₁₀₀ value of 250 μg/mL against <i>Aedes aegypti</i>	[63]
Venezuela	Leaves	34	95.2	α-pinene (15.3 %), β-pinene (14.8 %), β-elemene (8.1 %), caryophyllene oxide (7.8 %), δ-3-carene (6.9 %)	Antimicrobial: MBC values of 12.5 μg/mL against S. aureus, S. epidermides, S. saprophyticus, B. cereus, B. subtilis Cytotoxicity: IC50 values of 18.6 (HeLa), 27.7 (A549), 32.9 (MCF-7), 37.5 (Vero) μg/mL	[64]
Brazil	Leaves	26	98.6	Safrole (85.0%), terpinolene (5.4%)	Amoebicidal: At conc. of 0.5 mg/mL, the oil was lethal to 100% of the Acanthamoeba polyphaga trophozoites	[65]
Brazil	Leaves/ twigs	24	98.0	Safrole (77.9%), terpinolene (8.8%), bicyclogermacrene (3.7%)	Antifeedant: EC ₅₀ values of 0.4 and 17.7 μg/cm ² against <i>L. decemlineata</i> and <i>S. littoralis</i> , respectively	[66]
Brazil	Leaves	7	96.1	Safrole (89.9%), terpinolene (3.10%), β-	F	[67]
Brazil	Leaves/	22	99.4	bisabolene (1.70%) Safrole (64.3%), terpinolene (10.2%),		[68]

5. Biological activities

5.1. Antioxidant activity.

Research by Caceres *et al.* [69] reported the antioxidant activity by total phenolics, DPPH, and ABTS assays. In DPPH, the methanol extract of *P. hispidum* showed IC₅₀ values of 0.404 mg/mL (leaves) and 0.317 mg/mL (roots), whereas the dichloromethane extract gave IC₅₀ values of 0.391 mg/mL (leaves) and 0.263 mg/mL (roots), respectively. As for ABTS, the methanol extract demonstrated IC₅₀ values of 0.498 mg/mL (leaves) and 0.131 mg/mL (roots), whereas the dichloromethane extract gave IC₅₀ values of 0.158 mg/mL (leaves) and 0.164 mg/mL (roots). It is also reported that dichloromethane leaves extract revealed the highest phenolic content with 14.28 μg of gallic acid/mg extract.

5.2. Antibacterial activity.

P. hispidum has several medicinal properties and harbors a diversity of endophytes. Previously, four endophytic fungi from *P. hispidum* were used for obtaining crude ethyl acetate extracts that were tested against *Enterococcus hirae*, *Escherichia coli*, *Micrococcus luteus*, *Salmonella typhi*, and *Staphylococcus aureus*, using the cup plate technique. All bacteria were inhibited by the four extracts tested, except for *Enterococcus hirae* that was inhibited by only two extracts. The extract produced by *L. theobromae* was statistically the most effective against all bacteria except for *S. typhi*, being the extract of the Diaporthales endophyte more effective against it [70].

5.3. α -Amylase activity.

Orlandelli *et al.* [71] had evaluated the suitability of four agro-industrial wastes (corncob, pineapple peel, sugarcane bagasse, and wheat bran) as low-cost substrates for the α-amylase production by nine *P. hispidum* endophytes belonging to the genera *Bipolaris*, *Colletotrichum*, *Diaporthe*, *Phoma*, *Phyllosticta*, *Marasmius*, *Phlebia*, and *Schizophyllum* by using the starch-iodine method. Their study revealed that starchy substrates were less efficient than cellulose-rich substrates. Meanwhile, remarkable results were obtained for *Bipolaris* sp. JF767001 (4.14 U/mL) cultivated with pineapple peel, and for *Phlebia* sp. JF766997 (4.09 U/mL) and *S. commune* JF766994 (4.07 U/mL) with sugarcane bagasse.

5.4. Insecticidal activity.

Dos Santos *et al.* [72] evaluated the insecticidal and repellent potential of the acetonic leaf extract of *P. hispidum* on *Hypothenemus hampei* insects by topical application, contaminated surface, and repellent effect. They found that in the exposition in the contaminated surface, 100% of mortality was observed in the dilution of 25.0 mg/mL and 50 to 80% in dilutions of 5.0 to 0.004 mg/mL, while 0.0008 mg/mL and the control resulted in only 5% of mortality. Besides, in the topic application, 60 to 65% of mortality was observed with dilutions of 25.0 to 0.1 mg/mL. However, the repellence index was lower than the minimum value praised in the literature to consider a substance as a repellent.

5.5. Schistosomicidal effects.

The dichloromethane fraction of *P. hispidum* leaves (at 100 mg/mL) extract was found inactive in terms of mortality, number of separated worms, and number of worms with reduced motor activity [73].

5.6. Leishmanicidal activity.

The ethanolic extracts of P. hispidum showed leishmanicidal activity against promastigotes and axenic amastigotes of Leishmania amazonensis with IC₅₀ values of 5.0 μ g/mL and 69.0 μ g/mL, respectively [19].

5.7. Larvicidal activity.

Recently, Falkowski *et al.* [74] reported the leaves extract's larvicidal activity against *Aedes aegypti* laboratory strain susceptible to all insecticides. The extract exhibiting more than 50% larval mortality after 48 h of exposition at 100 μg/mL against the natural population were considered active. The leaves ethyl acetate extract of *P. hispidum* was found active with LC₅₀ values of 70.5 (95% CI 60.4–81.6) and 54.7 (95% CI 45.9–64.0) μg/mL at 24 and 48 h against the laboratory strain Paea. In another study, the ethanol leaves extract of *P. hispidum* showed larvicidal potential against *A. aegypti* larvae with an LC₅₀ value of 0.169 mg/mL [75].

5.8. Antiplasmodial activity.

The lipophilic extracts of P. hispidum proved to be active against both a chloroquine-sensitive and a resistant strain of *Plasmodium falciparum* with IC₅₀ values of 7.6 and 13.0 μ g/mL, respectively [34].

5.9. Cytotoxicity activity.

The lipophilic extracts of *P. hispidum* reported moderately active against human tumor cell lines SK-MEL30 (melanoma) and MCF-7 (mamma cell carcinoma) with IC₅₀ values of 24.1 and 25.1 μg/mL, respectively. Meanwhile, the extract was found inactive against KB (squamous carcinoma) and A549 (lung carcinoma), which gave IC₅₀ value >30 μg/mL [34].

5.10. Estrogenic and serotonergic activities.

Michel *et al.* [36] reported the *P. hispidum* leaf extract enhanced the expression of the estrogen-responsive reporter and endogenous genes in MCF-7 cells, demonstrating estrogen agonist effects.

6. Conclusions

In conclusion, the knowledge of traditional usages, origin, phytochemicals, and biological activities on *P. hispidum* has been comprehensively studied and have been well supported and clarified by modern pharmacological studies. Thus, it is suggested the potential application of *P. hispidum* on health and pharmaceutical industry. However, the present findings are still insufficient as the study of action mechanisms is still not clearly identified. Therefore, further efforts are needed, and more well-designed studies both in vitro and in vivo are required to establish the possibilities of *P. hispidum* to be used as a food supplement or for pharmaceutical purposes.

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Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Salleh, W.M.N.H.W.; Ahmad, F.; Yen, K.H.; Sirat, H.M. Chemical compositions, antioxidant and antimicrobial activities of essential oils of *Piper caninum* Blume. *Int. J. Mol. Sci.* **2011**, *12*, 7720-7731, https://doi.org/10.3390/ijms12117720.
- 2. Salleh, W.M.N.H.W.; Ahmad, F.; Yen, K.H.; Sirat, H.M. Chemical compositions, antioxidant and antimicrobial activity of the essential oils of *Piper officinarum* (Piperaceae). *Nat. Prod. Commun.* **2012**, 7, 1934578X1200701229, https://doi.org/10.1177/1934578X1200701229.
- 3. Salleh, W.M.N.H.W.; Ahmad, F.; Sirat, H.M.; Yen, K.H. Chemical compositions and antibacterial activity of the leaf and stem oils of *Piper porphyrophyllum* (Lindl.) NE Br. *EXCLI journal* **2012**, *11*, 399, https://doi.org/10.17877/DE290R-4900.
- 4. Salleh, W.M.N.H.W.; Ahmad, F.; Khong, H.Y. Chemical composition of *Piper stylosum* Miq. and *Piper ribesioides* Wall. essential oils, and their antioxidant, antimicrobial and tyrosinase inhibition activities. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas* **2014**, *13*, 488-497.
- 5. Salleh, W.M.N.H.W.; Ahmad, F.; Yen, K.H. Chemical compositions and antimicrobial activity of the essential oils of *Piper abbreviatum*, *P. erecticaule* and *P. lanatum* (Piperaceae). *Nat. Prod. Commun.* **2014**, 9, 1934578X1400901235, https://doi.org/10.1177/1934578X1400901235.
- 6. Salleh, W.M.N.H.W.; Hashim, N.A.; Ahmad, F.; Heng Yen, K. Anticholinesterase and antityrosinase activities of ten *Piper* species from Malaysia. **2014**, *4*, 527-531, https://doi.org/10.5681/apb.2014.078.
- 7. Salleh, W.M.N.H.W.; Kamil, F.; Ahmad, F.; Sirat, H.M. Antioxidant and anti-inflammatory activities of essential oil and extracts of *Piper miniatum*. *Nat. Prod. Commun.* **2015**, *10*, 2005-2008, https://doi.org/10.1177/1934578X1501001151.
- 8. Salleh, W.; Ahmad, F.; Yen, K.H. Chemical constituents from *Piper caninum* and antibacterial activity. *Journal of Applied Pharmaceutical Science* **2015**, *5*, 020-025, https://doi.org/10.7324/JAPS.2015.50604.
- 9. Mooney, H.A.; Field, C.; Yanes, C.V.; Chu, C. Environmental controls on stomatal conductance in a shrub of the humid tropics. *Proceedings of the National Academy of Sciences* **1983**, *80*, 1295, https://doi.org/10.1073/pnas.80.5.1295.
- 10. Wadt, L.H.d.O.; Ehringhaus, C.; Kageyama, P.Y. Genetic diversity of "*Pimenta Longa*" genotypes (*Piper* spp., Piperaceae) of the Embrapa Acre germplasm collection. *Genet. Mol. Biol.* **2004**, 27, 74-82, https://doi.org/10.1590/S1415-47572004000100013.
- 11. Yunker, T.G. Separata de hoenea: The Piperaceae of Brazil. *Instituto de Botânica Press, São Paulo, SP* **1972**, 2, 137-139.
- 12. The Plant List. Version 1 2001. Available online: http://www.theplantlist.org.
- 13. Michel, J.L.; Duarte, R.E.; Caceres, A.; Yao, P.; Bolton, J.L.; Huang, Y.; Caceres A.; Veliz M.; Soejarto, D.D.; Mahady, G.B. Medical potential of plants used by the Q'eqchi Maya of Livingston, Guatemala for the treatment of women's health complaints. *J. Ethnopharmacol.* **2007**, *114*, 92-101.
- 14. Albiero, A.L.M.; Paoli, A.A.S.; Souza, L.A.; Mourao, K.S.M. Morphology and anatomy of the vegetative organs of *Piper hispidum. Rev. Bras. Farmacogn.* **2006**, *16*(*3*), 379-391.
- 15. Lans, C.; Harper, T.; Georges, K.; Bridgewater, E. Medicinal and ethnoveterinary remedies of hunters in Trinidad. *BMC Complement. Altern. Med.* **2001**, *1*, 10, https://doi.org/10.1186/1472-6882-1-10.

- 16. Almeida, C.A.; Azevedo, M.; Chaves, F.; Roseo de Oliveira, M.; Rodrigues, I.A.; Bizzo, H.R.; Gama, P.E.; Alviano, D.S.; Alviano, C.S. *Piper* essential oils inhibit *Rhizopus oryzae* growth, biofilm formation, and rhizopuspepsin activity. *Can. J. Infect. Dis. Med. Microbiol.* **2018**, 5295619, https://doi.org/10.1155/2018/5295619.
- 17. Coe, F.G.; Anderson, G.J. Screening of medicinal plants used by the Garífuna of Eastern Nicaragua for bioactive compounds. *J. Ethnopharmacol.* **1996**, *53*, 29-50, https://doi.org/10.1016/0378-8741(96)01424-9.
- 18. Duke, J.A.; Vasquez, R. Amazonian Ethnobotanical Dictionary; CRC Press: 1994.
- Estevez, Y.; Castillo, D.; Pisango, M.T.; Arevalo, J.; Rojas, R.; Alban, J.; Deharo, E.; Bourdy, G.; Sauvain, M. Evaluation of the leishmanicidal activity of plants used by Peruvian Chayahuita ethnic group. *J. Ethnopharmacol.* 2007, 114, 254-259, https://doi.org/10.1016/j.jep.2007.08.007.
- 20. Morton, J.F. Atlas of medicinal plants of Middle America: Bahamas to Yucatan; Charles C. Thomas: 1981.
- 21. Facundo, V.A.; Pollli, A.R.; Rodrigues, R.V.; Militão, J.; Stabelli, R.G.; Cardoso, C.T. Fixed and volatile chemical constituents from stems and fruits of *Piper tuberculatum* Jacq. and from roots of *P*. hispidum. *Acta Amazon.* **2008**, *38*, 743-748, https://doi.org/10.1590/S0044-59672008000400018.
- 22. Otero, R.; Fonnegra, R.; Jiménez, S.L.; Núñez, V.; Evans, N.; Alzate, S.P.; García, M.E.; Saldarriaga, M.; Del Valle, G.; Osorio, R.G.; Díaz, A.; Valderrama, R.; Duque, A.; Vélez, H.N. Snakebites and ethnobotany in the northwest region of Colombia: Part I: Traditional use of plants. *J. Ethnopharmacol.* **2000**, *71*, 493-504, https://doi.org/10.1016/s0378-8741(00)00243-9.
- 23. Otero, R.; Núñez, V.; Barona, J.; Fonnegra, R.; Jiménez, S.L.; Osorio, R.G.; Saldarriaga, M.; Díaz, A. Snakebites and ethnobotany in the northwest region of Colombia: Part III: Neutralization of the haemorrhagic effect of *Bothrops atrox* venom. *J. Ethnopharmacol.* **2000**, *73*, 233-241, https://doi.org/10.1016/s0378-8741(00)00321-4.
- 24. Durant-Archibold, A.A.; Santana, A.I.; Gupta, M.P. Ethnomedical uses and pharmacological activities of most prevalent species of genus *Piper* in Panama: A review. *J. Ethnopharmacol.* **2018**, 217, 63-82, https://doi.org/10.1016/j.jep.2018.02.008.
- 25. Pimentel, F.A.; Cardoso, M.G.; Salgado, A.; Silva, V.; Zarcaroni, L.M.; Morais, A.R.; Nelson, D.L. Phytochemistry of *Piper hispidinervum* cultivated under the edafoclimatic conditions of Lavras, MG, Brazil. *Natural Products: An Indian Journal* **2006**, *2*, 89-94.
- 26. Lima, R.N.; Santos, A.D.C.; Ribeiro, A.S.; Cardozo-Filho, L.; Freitas, L.S.; Barison, A.; Costa, E.V.; Alves, P.B. Selective amides extraction and biological activity from *Piper hispidum* leaves using the supercritical extraction. *J. Supercrit. Fluids* **2020**, *157*, 104712, https://doi.org/10.1016/j.supflu.2019.104712.
- 27. Alécio, A.C.; Bolzani, V.d.S.; Young, M.C.M.; Kato, M.J.; Furlan, M. Antifungal amide from leaves of *Piper hispidum. J. Nat. Prod.* **1998**, *61*, 637-639, https://doi.org/10.1021/np9703656.
- Navickiene, H.M.D.; Alécio, A.C.; Kato, M.J.; Bolzani, V.d.S.; Young, M.C.M.; Cavalheiro, A.J.; Furlan, M. Antifungal amides from *Piper hispidum* and *Piper tuberculatum*. *Phytochemistry* 2000, 55, 621-626, https://doi.org/10.1016/S0031-9422(00)00226-0.
- 29. Friedrich, U.; Siems, K.; Solis, P.N.; Gupta, M.P.; Jenett-Siems, K. New prenylated benzoic acid derivatives of *Piper hispidum. ChemInform* **2005**, *36*, https://doi.org/10.1002/chin.200541203.
- 30. Ruiz, C.; Haddad, M.; Alban, J.; Bourdy, G.; Reategui, R.; Castillo, D.; Sauvain, M.; Deharo, E.; Estevez, Y.; Arevalo, J.; Rojas, R. Activity-guided isolation of antileishmanial compounds from *Piper hispidum*. *Phytochem. Lett.* **2011**, *4*, 363-366, https://doi.org/10.1016/j.phytol.2011.08.001.
- 31. Burke, B.; Nair, M. Phenylpropene, benzoic acid and flavonoid derivatives from fruits of jamaican *Piper* species. *Phytochemistry* **1986**, 25, 1427-1430, https://doi.org/10.1016/S0031-9422(00)81303-5.
- 32. Plazas, G.; Cuca, L.E.; Delgado, W.A. Flavonoids from inflorescences of *Piper hispidum* Kunth (Piperaceae) and acetylated derivatives. *Rev. Colomb. Quím.* **2008**, *37*, 135-144.
- 33. Vieira, P.C.; De Alvarenga, M.A.; Gottlieb, O.R.; Gottlieb, H.E. 4-Hexadecenylphenol and flavonoids from *Piper hispidum. Planta Med.* **1980**, *39*, 153-156, https://doi.org/10.1055/s-2008-1074918.
- 34. Jenett-Siems, K.; Mockenhaupt, F.P.; Bienzle, U.; Gupta, M.P.; Eich, E. *In vitro* antiplasmodial activity of Central American medicinal plants. *Trop. Med. Int. Health* **1999**, *4*, 611-615, https://doi.org/10.1046/j.1365-3156.1999.00456.x.
- 35. Costa, G.M.; Endo, E.H.; Cortez, D.A.G.; Nakamura, T.U.; Nakamura, C.V.; Dias Filho, B.P. Antimicrobial effects of *Piper hispidum* extract, fractions and chalcones against *Candida albicans* and *Staphylococcus aureus*. *J. Mycol. Med.* **2016**, *26*, 217-226, https://doi.org/10.1016/j.mycmed.2016.03.002.
- 36. Michel, J.L.; Chen, Y.; Zhang, H.; Huang, Y.; Krunic, A.; Orjala, J.; Veliz, M.; Soni, K.K.; Soejarto, D.D.; Caceres, A.; Perez, A.; Mahady, G.B. Estrogenic and serotonergic butenolides from the leaves of *Piper*

- hispidum Swingle (Piperaceae). *J. Ethnopharmacol.* **2010**, 129, 220-226, https://doi.org/10.1016/j.jep.2010.03.008.
- 37. Salleh, W.M.N.H.W.; Khamis, S.; Nadri, M.H.; Kassim, H.; Tawang, A. Essential oil composition and antioxidant activity of *Reinwardtiodendron cinereum* Mabb. (Meliaceae). *Nat. Volatiles Essent. Oils* **2020**, 7(4), 1-7, https://doi.org/10.37929/nveo.770245.
- 38. Salleh, W.M.N.H.W.; Khamis, S.; Nadri, M.H.; Kassim, H.; Tawang, A. Chemical composition and acetylcholinesterase inhibition of the essential oil of *Cyathocalyx pruniferus* (Maingay ex Hook.f. & Thomson) J.Sinclair. *Nat. Volatiles Essent. Oils* **2020**, *7*(4), 8-13, https://doi.org/10.37929/nveo.770303.
- 39. Salleh, W.M.N.H.W.; Khamis, S. Chemical composition of *Sarcotheca laxa* (Ridl.) Knuth essential oil and their bioactivities. *Riv. Ital. Sostanze Gr.* **2020**, *97*(*3*), 11-16.
- 40. Shakri, N.M.; Salleh, W.M.N.H.W.; Khamis, S.; Ali, N.A.M. Chemical characterization of *Goniothalamus macrophyllus* and *Goniothalamus malayanus* leaves' essential oils. *Z. Naturforsch. C J Biosci.* **2020**, *75*(11-12), 485-488, https://doi.org/10.1515/znc-2020-0090.
- 41. Shakri, N.M.; Salleh, W.M.N.H.W.; Khamis, S.; Ali, N.A.M. Composition of the essential oils of three Malaysian Xylopia species (Annonaceae). *Z. Naturforsch. C J Biosci.* **2020**, *75(11-12)*, 479-484, https://doi.org/10.1515/znc-2020-0096.
- 42. Shakri, N.M.; Salleh, W.M.N.H.W.; Khamis, S.; Ali, N.A.M.; Shaharudin, S.M. Chemical composition of the essential oils of four Polyalthia species from Malaysia. *Z. Naturforsch. C J Biosci.* **2020**, *75*(*11-12*), 473-478, https://doi.org/10.1515/znc-2020-0097.
- 43. Salleh, W.M.N.H.W.; Khamis, S. Chemical composition of the essential oil of *Diospyros wallichii* King & Gamble (Ebenaceae). *Nat. Volatiles Essent. Oils* **2020**, *7*(3), 12-17.
- 44. Salleh, W.M.N.H.W.; Khamis, S. Chemical composition and anticholinesterase inhibitory activity of *Pavetta graciliflora* Wall. ex Ridl. essential oil. *Z. Naturforsch. C J Biosci.* **2020**, *75*(11-12), 467-471, https://doi.org/10.1515/znc-2020-0075.
- 45. Azhar, M.A.M.; Salleh, W.M.N.H.W.; Khamis, S. Essential oil composition of three *Cryptocarya* species from Malaysia. *Z. Naturforsch. C J Biosci.* **2020**, 75(7-8), 297-301, https://doi.org/10.1515/znc-2020-0079.
- 46. Salleh, W.M.N.H.W.; Khamis, S.; Nafiah, M.A. Chemical composition and anticholinesterase inhibitory activity of *Dipterocarpus cornutus* Dyer essential oil. *Z. Naturforsch. C J Biosci.* **2020**, 75(5-6), 171-175, https://doi.org/10.1515/znc-2020-0028.
- 47. Salleh, W.M.N.H.W.; Khamis, S. Chemical composition and lipoxygenase activity of the leaves essential oil of *Rothmannia macrophylla* (Hook.f.) Bremek. *J. Essent. Oil Bear. Pl.* **2020**, 23(2), 331-336, https://doi.org/10.1080/0972060X.2020.1752817.
- 48. Anuar, M.Z.A.; Salleh, W.M.N.H.W.; Khamis, S.; Nafiah, M.A.; Said, Z.M. Essential oil composition of *Alseodaphne perakensis* (Gamble) Kosterm from Malaysia. *Nat. Prod. Res.* **2021**, *35*(*3*), 508-511, https://doi.org/10.1080/14786419.2019.1636245.
- 49. Assis, A.; Brito, V.; Bittencourt, M.; Silva, L.; Oliveira, F.; Oliveira, R. Essential oils composition of four *Piper* species from Brazil. *J. Essent. Oil Res.* **2013**, 25, 203-209, https://doi.org/10.1080/10412905.2013.767755.
- 50. da Silva, J.K.R.; Pinto, L.C.; Burbano, R.M.R.; Montenegro, R.C.; Guimarães, E.F.; Andrade, E.H.A.; Maia, J.G.S. Essential oils of Amazon *Piper* species and their cytotoxic, antifungal, antioxidant and anticholinesterase activities. *Ind. Crops Prod.* **2014**, *58*, 55-60, https://doi.org/10.1016/j.indcrop.2014.04.006
- Machado, S.M.F.; Militão, J.S.L.T.; Facundo, V.A.; Ribeiro, A.; Morais, S.M.; Machado, M.I.L. Leaf oils of two brazilian *Piper* species: *Piper arboreum* Aublet var. *latifolium* (C.DC) Yuncker and *Piper hispidum* Sw. *J. Essent. Oil Res.* 1994, 6, 643-644, https://doi.org/10.1080/10412905.1994.9699360.
- 52. Majolo, C.; Monteiro, P.C.; Nascimento, A.V.P.d.; Chaves, F.C.M.; Gama, P.E.; Bizzo, H.R.; Chagas, E.C. Essential oils from five Brazilian *Piper* species as antimicrobials against strains of *Aeromonas hydrophila*. *J. Essent. Oil Bear. Pl.* **2019**, 22, 746-761, https://doi.org/10.1080/0972060X.2019.1645047.
- 53. Ramos, C.S.; Soares, M.G.; da Silva, A.M.; Batista-Pereira, L.G.; Corrêa, A.G.; Kato, M.J. Electrophysiological responses of the Naupactus bipes beetle to essential oils from Piperaceae species. *Nat. Prod. Commun.* **2012**, *7*, 1934578X1200700835, https://doi.org/10.1177/1934578X1200700835.
- 54. Delgado, A.W.; Cuca, S.L.E. Composición química del aceite esencial de *Piper hispidum. Rev. Prod. Nat.* **2007**, *1*, 5-8.
- 55. Simeone, M.L.F.; Mikich, S.B.; Côcco, L.C.; Hansel, F.A.; Bianconi, G.V. Chemical composition of essential oils from ripe and unripe fruits of *Piper amalago* L. var. *medium* (Jacq.) Yunck and *Piper hispidum* Sw. *J. Essent. Oil Res.* **2011**, *23*, 54-58, https://doi.org/10.1080/10412905.2011.9700483.

- Lima, R.N.; Ribeiro, A.S.; Santiago, G.M.P.; Costa d'S.C.O.; Soares, M.B.; Bezerra, D.P.; Shanmugam, S.; dos S. Freitas, L.; Alves, P.B. Antitumor and *Aedes aegypti* larvicidal activities of essential oils from *Piper klotzschianum*, *P. hispidum*, and *P. arboretum. Nat. Prod. Commun.* 2019, 14, 1-6, https://doi.org/10.1177/1934578X19863932.
- 57. Benitez, N.P.; Meléndez León, E.M.; Stashenko, E.E. Essential oil composition from two species of Piperaceae family grown in Colombia. *J. Chromatogr. Sci.* **2009**, *47*, 804-807, https://doi.org/10.1093/chromsci/47.9.804.
- 58. Tangarife-Castaño, V.; Correa-Royero, J.B.; Roa-Linares, V.C.; Pino-Benitez, N.; Betancur-Galvis, L.A.; Durán, D.C.; Stashenko, E.E.; Mesa-Arango, A.C. Anti-dermatophyte, anti-Fusarium and cytotoxic activity of essential oils and plant extracts of *Piper* genus. *J. Essent. Oil Res.* **2014**, *26*, 221-227, https://doi.org/10.1080/10412905.2014.882279.
- 59. Jaramillo-Colorado, B.E.; Pino-Benitez, N.; González-Coloma, A. Volatile composition and biocidal (antifeedant and phytotoxic) activity of the essential oils of four Piperaceae species from Choco-Colombia. *Ind. Crops Prod.* **2019**, *138*, 111463, https://doi.org/10.1016/j.indcrop.2019.06.026.
- 60. Pino, J.A.; Marbot, R.; Bello, A.; Urquiola, A. Composition of the essential oil of *Piper hispidum* Sw. from Cuba. *J. Essent. Oil Res.* **2004**, *16*, 459-460, https://doi.org/10.1080/10412905.2004.9698771.
- 61. Houël, E.; Gonzalez, G.; Bessière, J.-M.; Odonne, G.; Eparvier, V.; Deharo, E.; Stien, D. Therapeutic switching: from antidermatophytic essential oils to new leishmanicidal products. *Mem. Inst. Oswaldo Cruz* **2015**, *110*, 106-113, https://doi.org/10.1590/0074-02760140332.
- 62. Cruz, S.M.; Cáceres, A.; Álvarez, L.E.; Apel, M.A.; Henriques, A.T. Chemical diversity of essential oils of 15 *Piper* species from Guatemala. **2011**, 39-46, https://doi.org/10.17660/ActaHortic.2012.964.4.
- 63. Santana, A.I.; Vila, R.; Cañigueral, S.; Gupta, M.P. Chemical composition and biological activity of essential oils from different species of *Piper* from Panama. *Planta Med.* **2016**, 82, 986-991, https://doi.org/10.1055/s-0042-108060.
- 64. Morales, A.; Rojas, J.; Moujir, L.M.; Araujo, L.; Rondón, M. Chemical composition, antimicrobial and cytotoxic activities of *Piper hispidum* Sw. essential oil collected in Venezuela. *J. Appl. Pharm. Sci.* **2013**, *3*, 16, https://doi.org/10.7324/JAPS.2013.3603.
- 65. Sauter, I.P.; Rossa, G.E.; Lucas, A.M.; Cibulski, S.P.; Roehe, P.M.; da Silva, L.A.A.; Rott, M.B.; Vargas, R.M.F.; Cassel, E.; von Poser, G.L. Chemical composition and amoebicidal activity of *Piper hispidinervum* (Piperaceae) essential oil. *Ind. Crops Prod.* 2012, 40, 292-295, https://doi.org/10.1016/j.indcrop.2012.03.025.
- 66. Andrés, M.F.; Rossa, G.E.; Cassel, E.; Vargas, R.M.F.; Santana, O.; Díaz, C.E.; González-Coloma, A. Biocidal effects of *Piper hispidinervum* (Piperaceae) essential oil and synergism among its main components. *Food Chem. Toxicol.* **2017**, *109*, 1086-1092, https://doi.org/10.1016/j.fct.2017.04.017.
- 67. Riva, D.; Simionatto, E.L.; Wisniewski Jr, A.; Salerno, A.R.; Schallenberger, T.H. Adaptation studies of *Piper hispidinervum* C.DC.(long pepper) species in Itajaí Valley-SC, by the chemical composition of essential oil obtained by microwave and traditional hydrodistillation. *Acta Amazon.* **2011**, *41*, 297-302, https://doi.org/10.1590/S0044-59672011000200016.
- 68. Filho, A.P.d.S.S.; Cunha, R.L.; Vasconcelos, M.A.M.d.; Zoghbi, M.d.G.B. Essential oil components of *Pogostemon heyneanus* Benth, *Piper hispidinervum* C. DC. and *Ocimum americanum* L. obtained in the Amazon. *J. Essent. Oil Bear. Pl.* **2010**, *13*, 347-352, https://doi.org/10.1080/0972060X.2010.10643833.
- 69. Cáceres, A.; Cruz, S.M.; Gaitán, I.; Guerrero, K.; Álvarez, L.E.; Marroquín, M.N. Antioxidant activity and quantitative composition of extracts of *Piper* species from Guatemala with potential use in natural product industry. *Acta Hortic*. **2011**, 77-84, https://doi.org/10.17660/ActaHortic.2012.964.9.
- 70. Orlandelli, R.C.; Alberto, R.N.; Almeida, T.T.; Azevedo, J.L.; Pamphile, J.A. In vitro antibacterial activity of crude extracts produced by endophytic fungi isolated from *Piper hispidum* Sw. *J. Appl. Pharm. Sci.* **2012**, 2, 137-141, https://doi.org/10.7324/JAPS.2012.21027.
- 71. Orlandelli, R.C.; Alberto, R.N.; Almeida, T.T.; Azevedo, J.L.; Pamphile, J.A. In vitro antibacterial activity of crude extracts produced by endophytic fungi isolated from *Piper hispidum* Sw. *J. Appl. Pharm. Sci.* **2012**, 2, 137-141, https://doi.org/10.4025/actascitechnol.v39i3.30067.
- 72. dos Santos, M.R.A.; Silva, A.G.; Lima, R.A.; Lima, D.K.S.; Sallet, L.A.P.; Teixeira, C.A.D.; Polli, A.R.; Facundo, V.A. Insecticidal activity of *Piper hispidum* (Piperaceae) leaves extract on (*Hypothenemus hampei*). *Rev. Bras. Botân.* **2010**, *33*, 319-324, https://doi.org/10.1590/S0100-84042010000200012.
- 73. Carrara, V.S.; Vieira, S.C.H.; De Paula, R.G.; Rodrigues, V.; Magalhães, L.G.; Cortez, D.A.G.; Da Silva Filho, A.A. *In vitro* schistosomicidal effects of aqueous and dichloromethane fractions from leaves and stems

- of Piper species and the isolation of an active amide from *P. amalago* L.(Piperaceae). *J. Helminthol.* **2014**, 88, 321, https://doi.org/10.1017/S0022149X13000205.
- 74. Falkowski, M.; Jahn-Oyac, A.; Odonne, G.; Flora, C.; Estevez, Y.; Touré, S.; Boulogne, I.; Robinson, J.-C.; Béreau, D.; Petit, P.; Azam, D.; Coke, M.; Issaly, J.; Gaborit, P.; Stien, D.; Eparvier, V.; Dusfour, I.; Houël, E. Towards the optimization of botanical insecticides research: Aedes aegypti larvicidal natural products in French Guiana. *Acta Trop.* **2020**, *201*, 105179, https://doi.org/10.1016/j.actatropica.2019.105179.
- 75. Porto, K.R.; Motti, P.R.; Yano, M.; Roel, A.R.; Cardoso, C.A.L.; Matias, R. Screening of plant extracts and fractions on *Aedes aegypti* larvae found in the state of Mato Grosso do Sul (linnaeus, 1762)(culicidae). *An. Acad. Bras. Cienc.* **2017**, 89, 895-906, https://doi.org/10.1590/0001-3765201720150017.