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An extraordinary new species of
Melanophryniscus (Anura, Bufonidae)
from southeastern Brazil

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ABSTRACT

We describe a new species of bufonid from a lowland, sandy soil, *restinga* habitat in the state of Espírito Santo, southeastern Brazil. Based on the shared occurrence of putative morphological synapomorphies of *Melanophryniscus* and the results of a phylogenetic analysis of DNA sequences of a broad sample of bufonids, and other anurans, we assign the new species to *Melanophryniscus*. The new species possesses several peculiar character states that distinguish it from all other *Melanophryniscus* including, but not limited to: fingers II, III, and V much reduced; nuptial pad with few enlarged, brown-colored spines on medial margin of finger II; seven presacral vertebrae, the last fused with the sacrum; and ventral humeral crest prominent, forming a spinelike projection.

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INTRODUCTION

Toads of the family Bufonidae are distributed in temperate and tropical areas worldwide, except for the Australo-Papuan region, Madagascar, Seychelles, and New Zealand, but they have been widely introduced in some of those areas where they did not naturally occur (Frost et al., 2006; Frost, 2011). It is a diverse family with nearly 50 genera and over 550 species. Although Bufonidae is monophyletic (Haas, 2003; Frost et al., 2006; Van Bocxlaer et al., 2010), the relationships among extant taxa are still not fully understood. *Melanophryniscus* is consistently recovered as the sister taxon of all other bufonids (Haas, 2003; Darst and Cannatella, 2004; Frost et al., 2006; Pramuk et al., 2007; Van Bocxlaer et al., 2010; Pyron and Wiens, 2011). Several morphological synapomorphies of Bufonidae exist (Ford and Cannatella, 1993; Frost et al., 2006), while few others support the less-inclusive clade of all Bufonidae excluding *Melanophryniscus* (and presumably *Truebella*, fide Frost et al., 2006; Pramuk, 2006).

We collected an unnamed and very peculiar bufonid toadlet during a herpetological survey in a *restinga* (sand-dune habitats covered mainly with herbaceous and shrubby xerophilous vegetation) fragment in the state of Espírito Santo, southeastern Brazil; *restingas* belong to the Atlantic rainforest biome and are common to the Brazilian coast (Suguio and Tessler, 1984; Eiten, 1992). Based on the presence of putative morphological synapomorphies of *Melanophryniscus* and results of a phylogenetic analysis of DNA sequences we assign the new species to *Melanophryniscus*.

MATERIAL AND METHODS

Fieldwork was carried out at Parque Estadual Paulo Cesar Vinha (PEPCV), a protected area in the *restinga* of Setiba, municipality of Guarapari, state of Espírito Santo, Brazil. Frogs were collected in pitfall traps with drift fences or by hand during active searches. Pitfall sampling occurred from December 2005 to November 2006 for 2–4 consecutive days, once a month, during that period. Traps were installed in sets of four buckets in a radial disposition (see Cechin and Martins, 2000, for details on pitfall traps). Three satellite buckets were placed 10 m from a central bucket and connected to it by 0.5 m tall drift fences. Five trap sets were installed in the area. Two sets in the open *Clusia* formation (for detailed description of vegetation formations at PEPCV see Pereira, 1990), two inside the forested area (*mata seca* of Pereira, 1990) and one in an area of herbaceous vegetation near the shore. Pitfall traps were checked once a day, in the morning. Traps were opened at 18:00 h (GMT -03:00 hrs) in the first day of sampling and closed at 18:00 h of the last day. Active searches for specimens were conducted from December 2005 through February 2007 for 2–4 consecutive days once a month. Searches were random in duration and habitat sampled. From November 2006–February 2007 a directed effort was made in search of evidence for reproductive activity of the new species. During that period searches focused on areas where individuals of the new species were previously found. Specifically, we actively searched the leaf litter, small crevices, burrows, small ponds, and phytotelmata (mostly bromeliads) looking for eggs, tadpoles, and breeding adults (i.e., calling males and amplexant pairs). Searches were conducted early in the morning and late in the afternoon, commonly

extending over the first few hours of night. Although several adults (of both sexes) and a few smaller specimens (apparently juveniles) were found during this directed effort, much to our frustration, no data on the reproductive biology of the species could be collected.

Live individuals were anesthetized and killed with a lethal dose of lidocaine, fixed in 10% formalin and preserved in 70% ethanol. Specimens used in the description and analyzed for comparisons are deposited in the following collections: American Museum of Natural History, New York (AMNH); Coleção de Anfíbios Célio F.B. Haddad, Universidade Estadual Paulista, Rio Claro, São Paulo, Brazil (CFBH); Museo Argentino de Ciencias Naturales “Bernardino Rivadavia”–CONICET, Buenos Aires, Argentina (MACN); Museu Nacional, Rio de Janeiro, Brazil (MNRJ); Coleção Herpetologica Oswaldo Rodrigues da Cunha, Museu Paraense Emílio Goeldi, Pará, Brazil (MPEG).

MORPHOLOGY: The following nine measurements were taken: SVL (snout-vent length), HL (head length; measured from tip of snout to posterior corner of the left eye), HW (head width; greatest width of head located between angles of the jaw), ED (eye diameter), IOD (interorbital distance; between anterior corners of the eyes), END (eye-nostril distance; from posterior margin of the nostril to anterior corner of the eye), THL (thigh length), TBL (tibia length), and FL (foot length; distal margin of tarsus to tip of fourth toe). All measurements are in millimeters, taken to the nearest 0.1 mm, and all were taken with an ocular micrometer attached to a Leica stereomicroscope, except for SVL, which was measured with digital calipers under the stereomicroscope. Descriptions of color in life were based on one paratype (CFBH 15745). Sex was determined by the presence of nuptial pads, vocal slits, and hypertrophied arms in males. This method of sexual identification proved successful by the dissection of one putative male and one putative female and direct observation of gonads and developing eggs.

Two paratypes, one male (CFBH 15745) and one female (CFBH 15735), were cleared and double-stained for osteological study, following the protocol by Taylor and Van Dyke (1985). Terminology follows Trueb (1973, 1993) for cranial and postcranial osteology and Fabrezi (1992, 1993) for carpal and tarsal osteology. Following Fabrezi and Alberch (1996), fingers are numbered II–V.

PHYLOGENETIC ANALYSES: To hypothesize the phylogenetic placement of the new species, we analyzed DNA sequences from a broad sample of bufonid and several nonbufonid clades. The analysis included sequences of the mitochondrial genes 12S, the intervening tRNA^{Val}, and a fragment of 16S, and portions of the nuclear genes exon 1 rhodopsin, exon 2 of chemokine receptor 4 (Cxcr4), and recombination activating gene 1 (RAG-1). Sequences for most terminals were obtained from Genbank (appendix 2). We generated sequences from the new species and from *Frostius erythrophthalmus*. Through the courtesy of Diego Baldo we included sequences of *Melanophryniscus devincenzii*. Laboratory protocols and primers employed followed Faivovich et al. (2010).

Phylogenetic analyses were performed under direct optimization in POY 5.0 (Varón et al., 2010, 2012) using equal weights for all transformations (substitutions and insertion/deletion events) and the parsimony optimality criterion, following the justification of Kluge and Grant (2006; see also Grant and Kluge, 2009; Wheeler, 2012). Contiguous sequences were preliminar-

ily delimited in fragments of putative homology to allow incorporation of partial sequences and accelerate dynamic homology cost calculations (Wheeler et al., 2006). Analyses were performed using the command “search,” which implements a driven search composed of random addition sequence Wagner builds (RAS), subtree pruning and regrafting (SPR) and tree bisection and reconnection (TBR) branch swapping, parsimony ratcheting (Nixon, 1999), and tree fusing (Goloboff, 1999), storing the shortest trees of each independent run and performing a final round of tree fusing on the pooled trees. Five independent runs, each consisting of four 12-hour driven searches, were implemented in parallel on a dual hexacore server at the Museu de Zoologia da Universidade de São Paulo. A final 24-hour run composed of three 8-hour driven searches of the implied alignment (Wheeler, 2003) derived from the optimal tree found under direct optimization was performed to search for additional most-parsimonious trees. Goodman-Bremer support values (Goodman et al., 1982; Bremer, 1988; see Grant and Kluge, 2008) were calculated using inverse constraints to search for next-most-optimal trees with 5 RAS + TBR analyses of the implied alignment.

RESULTS

At least two character states shared by the new species and all the remaining bufonids justify its inclusion within the family: the origin of the m. depressor mandibulae solely from the squamosal and the absence of teeth (Ford and Cannatella, 1993; Frost et al., 2006). The absence of the zygomatic rami, exostosed frontoparietals that diverge anteriorly, and fusion of the parasphenoid with the sphenethmoid and prootic are putative synapomorphies of *Melanophryniscus* (McDiarmid, 1971; Graybeal and Cannatella, 1995) that also occur in the new species. Additionally, our phylogenetic analysis of DNA sequence data resulted in six most-parsimonious trees of 33,389 steps that recovered the new species as the well-supported (Goodman-Bremer value of 26) sister of all included species of *Melanophryniscus*, which collectively are recovered as the sister of all remaining bufonids (fig. 1; see appendix 3 for entire tree). Although our analysis aimed only to study the position of the new species and should not be construed as a test of the most current hypotheses of bufonid relationships (Van Bocxlaer et al., 2010; Pylon

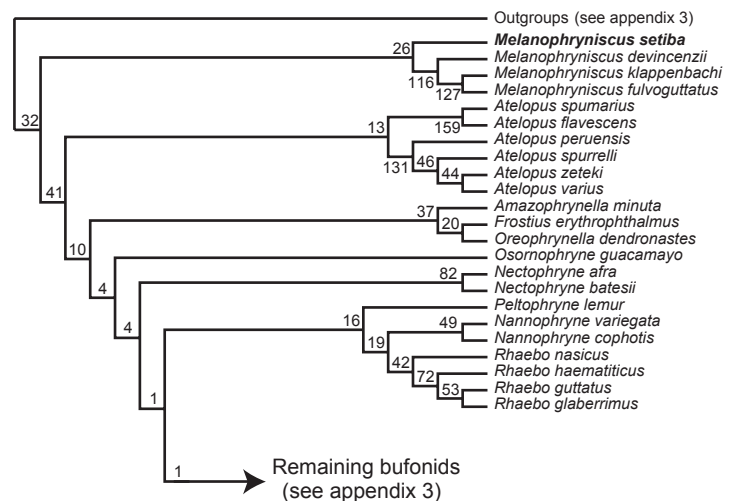


FIG. 1. Part of the strict consensus of 6 optimal trees of 33,389 equally weighted steps with Goodman-Bremer support values on each node. See appendix 3 for the full tree. The new species is in **boldface**.

and Wiens, 2011), we also note that *Frostius*, which has never been included in molecular phylogenetic analyses, was recovered as the sister taxon of our only exemplar of *Oreophrynella* (*O. dendronastes*) in a well-supported clade also containing *Amazophrynella minuta*.

***Melanophryniscus setiba*, sp. nov.**

Figures 2–12, table 1

HOLOTYPE: CFBH 17036, male (figs. 3–4); from Parque Estadual Paulo César Vinha (20°36'25"S; 40°25'01"W, near sea level), approximately 1.5 km from the beach, restinga of Setiba, municipality of Guarapari, state of Espírito Santo, Brazil; collected on 10 December 2005 by P.L.V. Peloso, S.E. Pavan, N.B. Thomazini, and R. Montesinos.

PARATOPOTYPES: CFBH 15727–33, three adult females, one adult male, one adult female, and two adult males, respectively, collected on 26–29 December 2005 by P.L.V. Peloso, R. Montesinos, and R.V. Peloso; CFBH 15734–41, one adult male, one adult female (15735, adult female cleared and double-stained), three adult males, one adult female, one adult male, and one subadult, respectively, collected on 08–24 January 2006 collected by P.L.V. Peloso, S.E. Pavan, R. Montesinos, B. Becacici, and J. Albino; CFBH 15742, MPEG 21940–41 three adult females, collected on 17–18 March 2006 by P.L.V. Peloso and R. Montesinos; MPEG 21944, adult male, collected in May 2006 by R. Kawada; CFBH 15743–45, three adult males (15745 cleared and double-stained), collected on 17 December 2006 by P.L.V. Peloso and S.E. Pavan; MPEG 21942–43, one adult male, and one adult female, respectively, collected by P.L.V. Peloso, D. Barbosa, P. Rubens, R. Kawada, and S.E. Pavan on 15 December 2007.

DIAGNOSIS: A small *Melanophryniscus* (SVL 13.8–16.1; table 1) with a moderately robust body that can be diagnosed by the combination of the following character states: (1) head wider than long; (2) snout round in dorsal view and slightly protruding in lateral view (profiles follow Heyer et al., 1990); (3) frontal macrogland (see Naya et al., 2004) absent; (4) fingers II, III, and V much reduced; (5) subarticular tubercles distinguishable on finger IV; (6) nuptial pad, with few enlarged, brown-colored keratinized spines at medial margin of finger II present in males; (7) finger and toe tips rounded, unexpanded; (8) toes I–IV connected by barely noticeable webbing; (9) frontoparietals heavily exostosed, fused, not diverging anteriorly; (10) pectoral girdle completely ossified, triangular; (11) sternum heavily mineralized and greatly reduced; (12) seven presacral vertebrae, the last fused with the sacrum; (13) ventral humeral crest prominent, forming a spinelike projection, more evident in males than females; (14) surfaces of head and dorsum slightly granular without keratinous spines; throat and chest smooth; and (15) color pattern: dorsum reddish brown to dark brown in life; two dorsal marks present on dorsum—one in form of an “X” anteriorly, and one in form of a “Λ” posteriorly; venter light colored with variable amounts of brown markings and spots; a ventral dark brown stain usually present at midbody.

COMPARISONS WITH OTHER SPECIES: *Melanophryniscus setiba* differs from all remaining species of *Melanophryniscus* by the strong coossification of the dorsal skull elements. Males of the new species are readily distinguished from males of all remaining species of *Melanophryniscus* by the presence of a bifurcated humeral spine and by nuptial pads with a few enlarged,

TABLE 1 Measurements of the type series of *Melanophryniscus setiba*. The paratype CFBH 15741 is likely a juvenile (very small SVL) and was excluded from this analysis. SD = standard deviation.

	Males (<i>n</i> = 14)			Females (<i>n</i> = 10)		
	Mean	SD	Range	Mean	SD	Range
SVL	15.2	0.6	14.4–16.1	15.2	0.8	13.8–16.0
HL	3.8	0.2	3.2–4.1	3.6	0.2	3.4–4.0
HW	5.7	0.2	5.3–6.1	5.7	0.2	5.3–6.1
ED	1.5	0.1	1.2–1.6	1.45	0.1	1.4–1.6
IOD	2.9	0.5	1.6–3.3	2.68	0.6	1.6–3.3
END	1.2	0.1	1.1–1.5	1.18	0.1	1.0–1.3
THL	6.7	0.1	6.4–7.0	6.26	0.4	5.6–6.7
TBL	6.2	0.2	5.8–6.7	5.73	0.5	5.2–6.9
FL	4.3	0.3	3.6–4.8	4.08	0.3	3.5–4.6

brown-colored spines on the base of finger II (taxonomic distribution of nuptial pads poorly known in *Melanophryniscus*, but, when present, composed of multiple minute, spines colored on its tips, on fingers II, III, and IV: see Discussion); presence of prominent ventral humeral crest, and reduced phalangeal formula of both hands and feet. The absence of a frontal macrogland readily separates the new species from all species in the *M. tumifrons* group (the macrogland is a putative synapomorphy of the *M. tumifrons* group; see Caramaschi and Cruz, 2002; Baldo and Basso, 2004; Naya et al., 2004). The new species further differs from all species in the *M. moreirae* and *M. stelzneri* groups by its almost smooth skin on dorsum and flanks (developed warts with an apical corneous spine in the *M. moreirae* group; and keratinized spines on skin in the *M. stelzneri* group). The diminutive size of *M. setiba* is rivaled only by *M. vilavelhensis* (12.8–17.2 mm; unassigned to any group; Steinbach-Padilha, 2008), with all other species >20 mm (summary in Steinbach-Padilha, 2008: 104).

DESCRIPTION OF THE HOLOTYPE (figs. 3–4): Body robust. Head large, wider than long; snout short, rounded in dorsal view and slightly protruding in lateral view (fig. 4A–B); nostrils small, not protuberant, directed anterolaterally, almost at tip of snout; frontal macrogland absent; canthus rostralis distinct, concave; loreal region nearly flat, vertical; eye diameter about half the interorbital distance, not protruding dorsally; postorbital crests absent; tympanum absent; supratympanic fold absent; vocal sac medial, not forming externally visible folds; vocal slits present, tongue long and narrow; premaxillary, maxillary, and vomerine teeth absent; choanae small, rounded. Strong coossification of the dorsal skull elements. Ventral humeral crest present, well developed, not projecting through the skin but externally visible as a protuberance. Arm and forearm hypertrophied (likely due to extensive development of musculature). Finger IV longest, followed by finger III; finger V reduced but distinguishable (the peculiar morphology of finger II and a much reduced finger V make judgments of relative finger lengths imprecise and unreliable); finger IV moderately robust; finger tips rounded; fingers connected by a thick web; nuptial pad with few enlarged, well-separated, brown-colored

keratinized spines along the proximal medial margin of finger II (fig. 4C); subarticular tubercles present and conspicuous on fingers III and IV; inner thenar tubercle elliptical, larger than outer metacarpal tubercle; palmar tubercle present, large, rounded; supernumerary tubercles on palm of hand present but barely distinguishable (fig. 4C). Legs short, slender; toes robust; toes I, II, and V very reduced; toe IV the longest, followed by toe III (others too small to make a confident statement of relative length; fig. 4D); toe tips rounded; barely noticeable webbing connecting toes II–V; subarticular tubercles present, large; inner metatarsal tubercles large, elliptical, larger than inner metatarsal tubercle (fig. 4D). Skin on head, dorsum, dorsal surfaces of arms, and legs very lightly granular. Gular region, chest, and ventral surfaces of arms and legs smooth.

MEASUREMENTS OF HOLOTYPE: SVL 15.1, HL 3.9, HW 5.5, ED 1.5, IOD 3.2, END 1.1, THL 6.4, TBL 5.9, FL 4.0.

COLOR OF THE HOLOTYPE IN PRESERVATIVE: Dorsum brown; two dorsal marks present, one in the form of an “X” anteriorly and one in the form of a “Λ” posteriorly; marks on the dorsum dark brown; dorsal arms and legs grayish brown. Throat brown (usually more pigmented in males than females); chest dark brown; venter light brown with a large dark brown stain at midbody and many dark spots and minor stains all over the ventral surface; ventral arms and legs cream color; tips of fingers and toes beige.

COLOR IN LIFE (based on paratype CFBH 15475, male, fig. 2): Iris black; pupil ring golden. Dorsum reddish brown to dark brown; marks on the dorsum dark brown; a few white spots present dorsolaterally; arms and legs brown dorsally. Throat brown; chest dark brown; venter light orange with large dark brown blotch at midbody, ventral arms and legs light orange. Palm and sole brownish red; tips of fingers and toes reddish orange.

VARIATION AND SEXUAL DIMORPHISM: Measurements of the type series are given in table 1. Males have robust arms and forearms (likely due to muscular hypertrophy) while they are slender in females (fig. 5). The humeral processes and bifurcated spine are variably visible externally among male specimens (fig. 5B–D), never visible externally in females (fig. 5A). Females lack nuptial spines and have slender arms and forearms. One cleared and double-stained female (CFBH 15735) possesses a distinct humeral spine, although it is not bifurcated and is much less developed than that of the male specimen (CFBH 15745). No sexual dimorphism in SVL was detected among 14 males and 10 females (Student’s *t*-test; $t = 0.139$, $df = 22$, $p = 0.891$), although females have significantly shorter legs than males ($t = -3.856$, $p < 0.05$ for THL; $t = -2.796$, $p < 0.05$ for TBL; $t = -2.021$, $p = 0.056$ for FL). Bidder’s organ is apparently absent (not detected from direct observation under a dissecting microscope or from histological sections).

The X- and Λ-shaped marks on the dorsum vary in intensity, but this variation does not seem to be ontogenetic or sexually dimorphic; some specimens have a light-brown dorsum while in others it is dark brown (colors in preservative, but such variations do occur in live specimens; reddish brown/dark brown). A few individuals have white spots on the dorsolateral surface near the thigh in life (fig. 2); in preservative these white spots tend to disappear. Ventral pattern of brown markings variable among individuals (fig. 5); throat uniformly dark colored

(brownish) or light (creamish) with brown spots and blotches; ventral midbody usually with a brown blotch, highly variable in shape and size, light or dark brown.

OSTEOLOGY: Description based on male CFBH 15745 and female CFBH 15735. Skull slightly longer than wide in dorsal view. Frontoparietals, prootics, exoccipitals, sphenethmoid complex, nasals, palatines, and, if actually present, vomers, forming a continuous, single, fused structure with unclear limits between individual endochondral bones (fig. 6A–D); dorsal surface of nasals and frontoparietals heavily exostosed with irregularly shaped outgrowths, allowing limits to be inferred (fig. 6B). Frontoparietals fused to each other and nasals, sphenethmoid, prootic, and ossified tectum synoticum. Frontoparietals with nearly triangular posterolateral processes covering posterior corner of orbit and anteromedial portion of otic capsule, extending laterally to anterolateral tip of otic capsule. Occipital groove covered along most of its length, exposed in some areas. Nasals fused medially, longer than wide, reaching anteriorly the tip of the snout; maxillary process short, not reaching pars facialis of the maxilla laterally. Premaxillae edentulous and in close contact with each other and with the maxillae (fig. 6C–D). Alary process of premaxilla laminar, expanded distally; directed slightly anteriorly in profile. Pars palatina well developed; pars dentalis nearly laminar. Palatine process pointed in ventral view, conspicuously thicker than pars palatina; maximum length about twice the width of the pars palatina. Maxilla edentulous; pars facialis of the maxilla with irregular free margin. Quadratojugal small and restricted to anteroventral part of quadrate (fig. 6E), contiguous with distal portion of ventral ramus of the squamosal; anterior process short and widely separated from the maxilla; quadrate unossified. Parasphenoid indistinguishably fused with sphenethmoid and prootic (fig. 6C). Vomers absent as discrete elements (i.e., either fused to underlying sphenethmoid complex or absent); dentigerous process absent; prechoanal process well developed; postchoanal process reduced. Palatines absent as discrete elements, likely fused to the sphenethmoid complex, as suggested by a distinct thickening of the bone in the corresponding area. Pterygoid triradiate; anterior ramus in contact with maxilla, reaching base of antorbital process, almost touching lateral tip of sphenethmoid complex; medial ramus in bony contact with otic capsule. Squamosal lacking zygomatic ramus; otic ramus with broad otic plate abutting crista parotica; ventral arm laterally flattened in its anterior section, resulting in bony sheath with pointed ventral process. Sphenethmoid extensively ossified, forming sphenethmoid complex due to fusion with several ossified nasal structures, including complete septum nasi. Orbitosphenoid cartilage completely ossified. Prootic fused with parasphenoid, exoccipital, and with the sphenethmoid complex anteriorly. Epiotic eminences low. Crista parotica broad, completely ossified. Occipital condyles broadly separated. Columellae and tympanic ring absent. Operculum cartilaginous, about two thirds of its volume ossified, with prominent point of attachment for the *m. opercularis* (fig. 6F). Fenestra ovalis with bony margins, except for the lateral wall in the male, which is cartilaginous.

Angulosplenic extended from the jaw articulation almost to mentomeckelian bones. Dentary widest at anterior termination of angulosplenic, overlapping mentomeckelian. Mentomeckelian elements small and syndesmotically united. Distal portion of each half of the jaw, including mentomeckelian, and distal portions of dentary and angulosplenic, curved ventrally, forming a wide V-shape in the area around the symphysis in frontal view.



FIG. 2. *Melanophryniscus setiba*, in life. Adult male, SVL 16.0 mm (CFBH 15745, paratype).

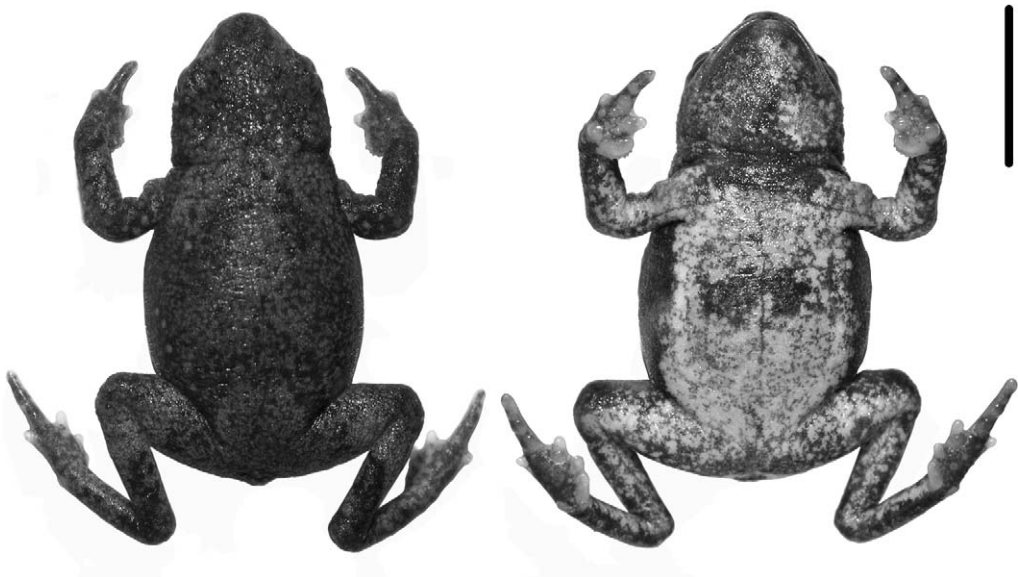


FIG. 3. *Melanophryniscus setiba*. Dorsal and ventral views of the holotype (CFBH 17036, male). Scale bar = 5.0 mm.

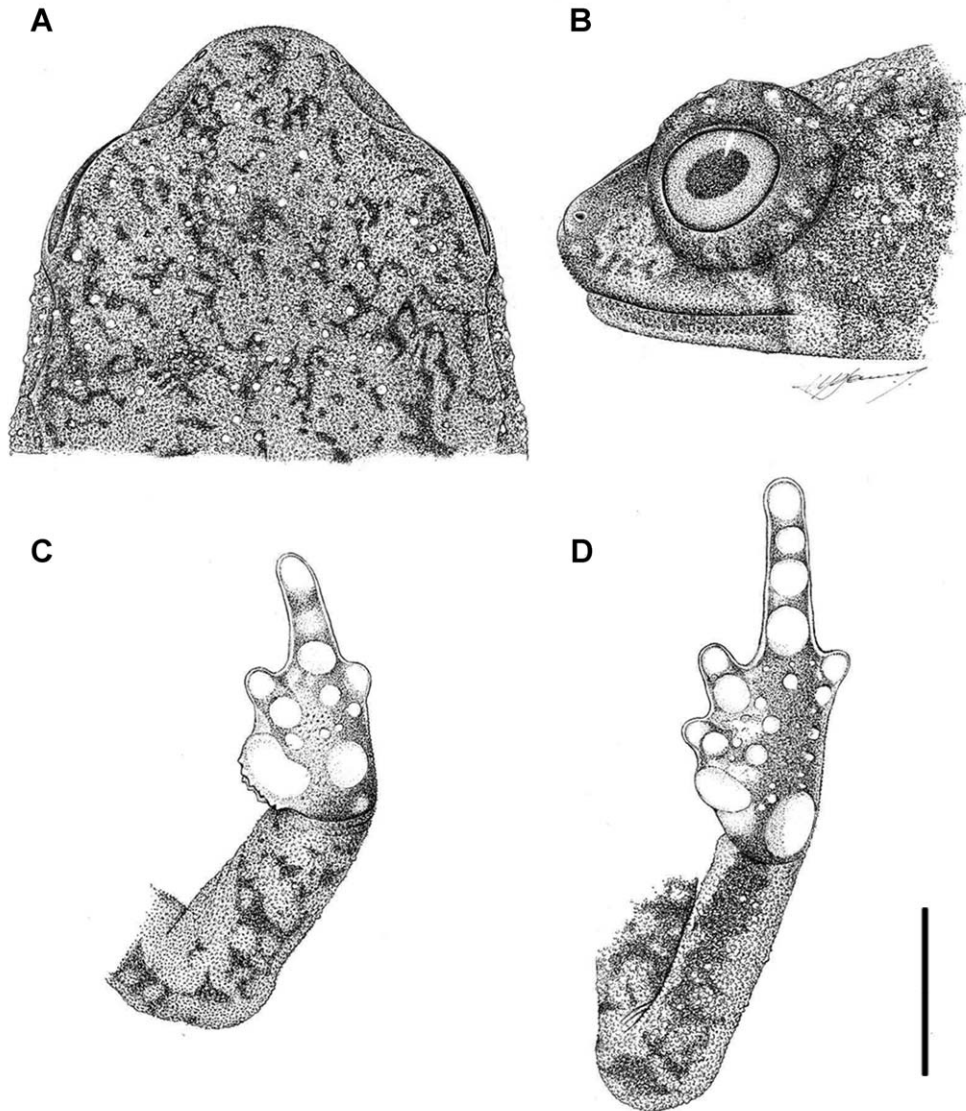


FIG. 4. *Melanophryniscus setiba*. Holotype (CFBH 17036, male). **A.** dorsal and **B.** lateral views of head; ventral views of **C.** hand and **D.** foot. Note the nuptial pad with keratinized spines at the medial margin of finger II. Scale bar = 2.0 mm.

Hyoid plate (fig. 7) about five times longer than wide (measured at narrowest point); not mineralized in female, slightly mineralized in male; hyalia moderately long, anterior processes oriented medially; hyalia inserted at limit between prootic and basal process. Anterolateral process present and broadly expanded, about 55% as long as hyoid plate; posterolateral processes absent. Posteromedial processes long, well ossified, noticeably curved dorsally. Larynx of male and female similar sized; female with complete cricoid; esophageal process small, triangular; bronchial processes long and slender (cricoid and bronchial processes of male damaged during preparation).

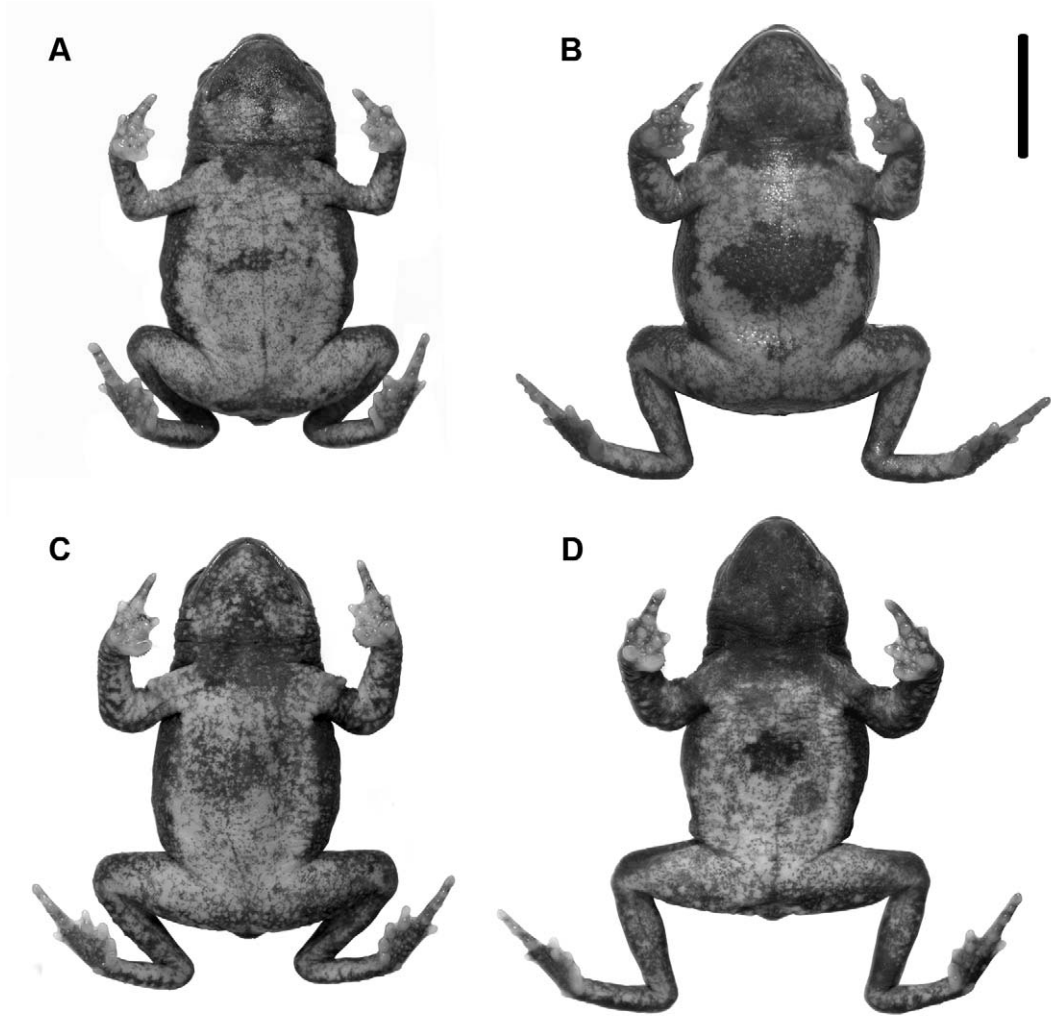


FIG. 5. Ventral pattern variation in *Melanophryniscus setiba*, **A**. CFBH 15739 female; **B**. CFBH 15745 male; **C**. CFBH 15733 male; and **D**. CFBH 15734 male. All paratypes. Scale bar = 5.0 mm.

Seven procoelous presacral vertebrae (fig. 8); first six vertebrae free, seventh fused with sacral vertebra, as evidenced by the intervertebral foramina between fused centra. Vertebrae II–VI with imbricated neural arches. Cotylar facets of atlas broadly separated. Transverse processes of presacral II directed anteriorly; transverse process of presacral III broader than those of presacrals II, IV–VI; processes with irregular flanges along its free margins. Neural arches of presacrals II–VI with irregular posterior margins; neural arches of presacrals III–V ornamented with elevated, anteriorly directed triangular shape with irregular base, those of presacrals VI and VII + sacrum ornamented with an irregular sagittal flange. Sacral diapophyses flattened, broadly expanded; irregular flanges along anterior and posterior margins. Iliosacral articulation with elongate sesamoid. Sacro-urostilar articulation bicondilar. Anterior half of

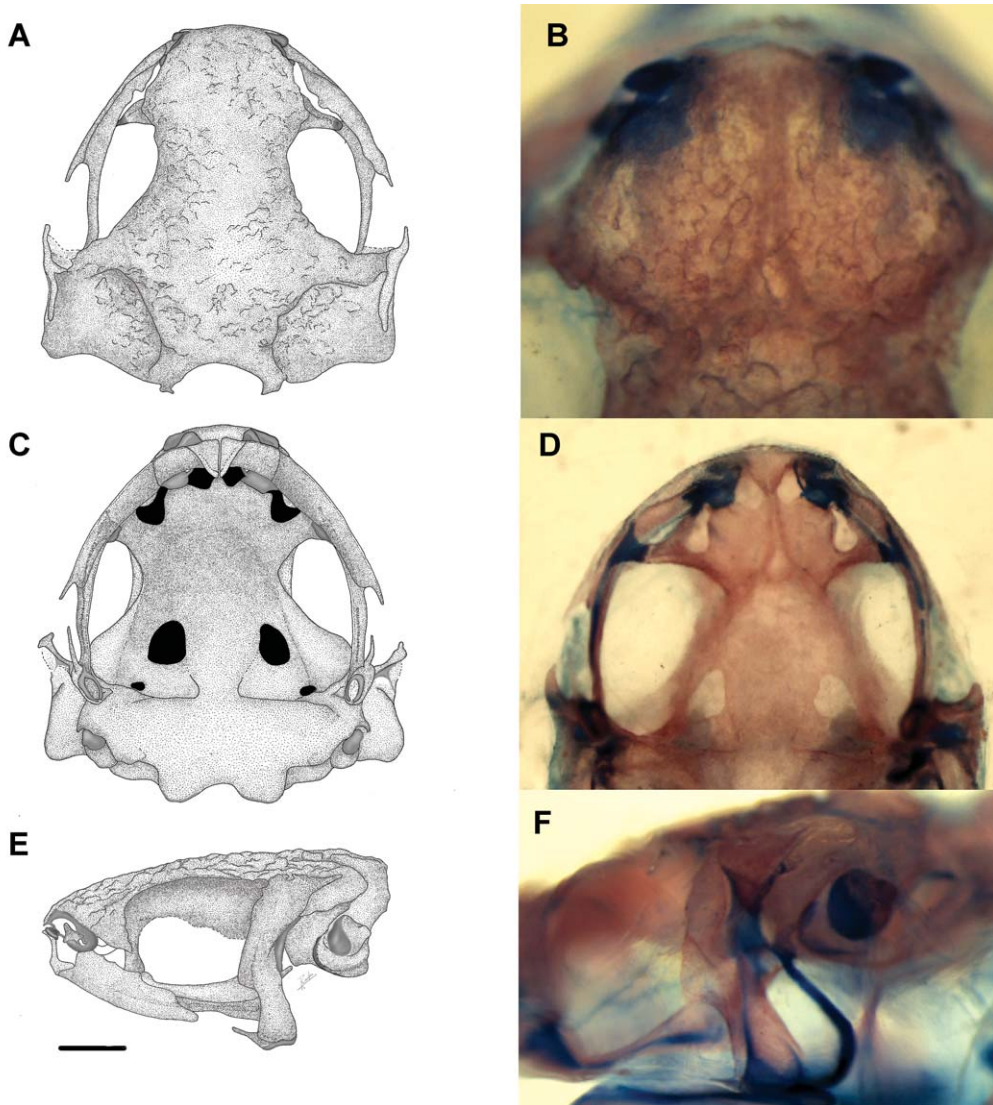


FIG. 6. Skull of *Melanophryniscus setiba*. Drawings (CFBH 15735, female) in A. dorsal, B. ventral, and C. lateral views. Photographs (CFBH 15745, male) showing details of D. nasal/frontoparietal region, E. ventral view of skull, and F. otic region.

urostyle bearing low, posteriorly tapered dorsal crest (fig. 8A); anterior portion of the crest with two parasagittal flanges; middle portion of urostyle with two lateral flanges, tapering and disappearing by the distal fifth of the bone.

Pectoral girdle (fig. 9) lacking omosternum. Clavicles straight and robust, fused to scapulae and coracoid. Partes acromialis larger than partes glenoidiales. Coracoids broadly dilated proximally. Epicoracoid cartilages heavily mineralized and reduced to a single, thin sliver of cartilage joining clavicles and coracoids. In the female the median section of the medial margin of the

coracoids is concave, and the epicoracoid discontinuous, establishing a fenestra between both coracoids. Sternum cartilaginous and heavily mineralized, round, greatly reduced. Cleithrum and ossified suprascapula indistinguishably fused.

Pelvic girdle (fig. 10) V-shaped. Ischia, pubis, and ilia fused. Iliac shaft cylindrical, elongate; dorsal crest developed along proximal 60% of ilium; dorsal prominence directed dorsolaterally. Preacetabular angle obtuse.

Humerus slender. In the male, it bears prominent ventral, medial, and lateral crests; ventral crest nearly 40% as long as humerus, bearing distinct, bicapitate, spinelike process near distal end of free margin (fig. 11); lateral and medial crests about 50% as long as humerus; medial crest higher than lateral crest. In the female, medial and lateral crests inconspicuous, forming low ridge; ventral crest present but less developed than in the male, lacking distinct spinelike process.

Carpus of right hand of female composed of radiale, ulnare, distal carpal 3–4–5, distal carpal 2, element Y, proximal element of prepollex, one sesamoid on the dorsal surface between radioulna and radiale, and one sesamoid on the ventral surface of distal carpal 3–4–5. Carpus of left hand differing in that proximal element of prepollex is absent as a discrete element, with equivalent space occupied by element Y; we consider it likely that in this hand the prepollex is fused with element Y. Terminal phalanges knoblike; reduced in

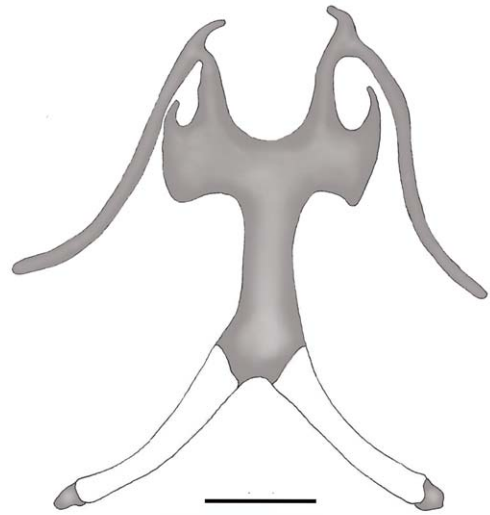


FIG. 7. Hyoid plate of *Melanophryniscus setiba* in ventral view (CFBH 15735, female). Scale bar = 1.0 mm.

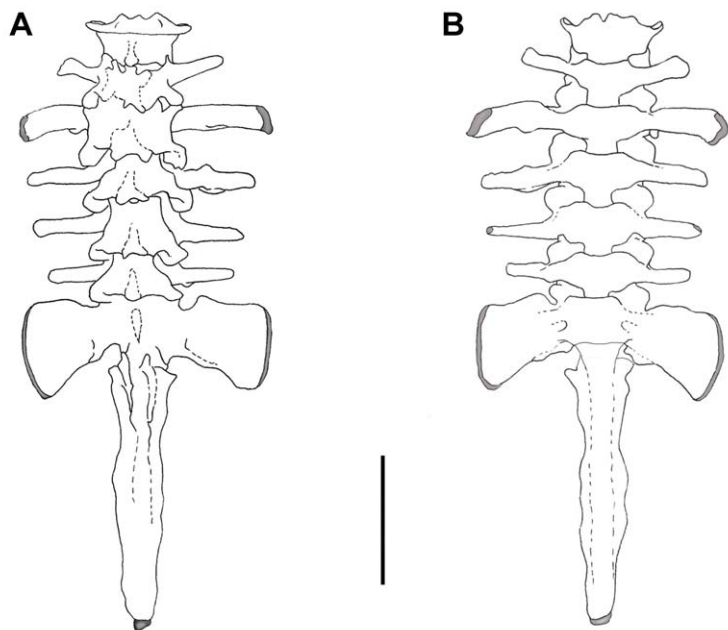


FIG. 8. Vertebral column of *Melanophryniscus setiba* (CFBH 15745, male) in A. dorsal, and B. ventral views. Scale bar = 2.0 mm.

digits II, III, V; elongate and slightly expanded in digit IV; penultimate phalanx of digit V reduced. Metacarpal II enlarged, bearing a peculiar, longitudinal bony outgrowth on medial margin, not ossified distally. Carpus of male (fig. 12A) with same morphology as female, with the addition that metacarpal II is even more enlarged medially due to hypertrophy of the bony outgrowth, which is fully ossified and has a distal process that covers the mediolateral section of metacarpal II. The relatively depressed longitudinal space delimited by this process and the element Y–prepollex coincident with space occupied by nuptial spines. Phalangeal formula of hand 1–2–3–2.

Tarsus (fig. 12B) composed of tibiale, fibulare, and three individual elements, including distal tarsal 2–3, an element that we interpret as distal tarsal 1 + element Y, and prehallux. Terminal phalanges knoblike, reduced on digits I–III and V, elongate on digit IV; penultimate phalanx on digit III reduced in the female. One sesamoid occurring in tarso-metatarsal joint at level of metatarsal IV, other between the proximal heads of tibiale and fibulare. Phalangeal formula of foot 1–2–2–4–2 (male), 1–2–3–4–2 (female).

DISTRIBUTION AND NATURAL HISTORY: *Melanophryniscus setiba* is known only from the type locality, where it was found in leaf litter of forested areas. Specimens were usually found in *mata seca* (“dry forest”). Several individuals were observed at the forest edge, less than 1 m away from an adjacent formation known as “open *Clusia* formation.” The open *Clusia* formation occurs in sandy soil and is characterized by discontinuities in the vegetation with small and large shrubs forming a mosaic structure (fig. 13).

Melanophryniscus setiba, like most other *Melanophryniscus* (see Santos and Grant, 2011), is apparently a diurnal species with a peak of activity recorded in late afternoon. One specimen was collected at night while walking on the leaf litter, while 10 others were collected when daylight was available (all others collected in pit-fall traps). Additional specimens were seen, but not collected, during the day (P.L.V.P., personal obs.). The species is a walking toad, with locomotion similar to that previously observed in other *Melanophryniscus* spp. and similar bufonids (McDiarmid, 1971; Baldo and Basso, 2004).

Despite a major, concentrated effort to obtain data on the reproductive biology of this species (see Material and Methods), no reproductive activity (e.g., calling males or amplexant pairs) or clues that could point to

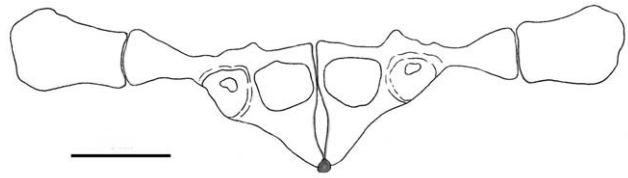


FIG. 9. Pectoral girdle of *Melanophryniscus setiba* (CFBH 15745, male) in ventral view. Scale bar = 2.0 mm.

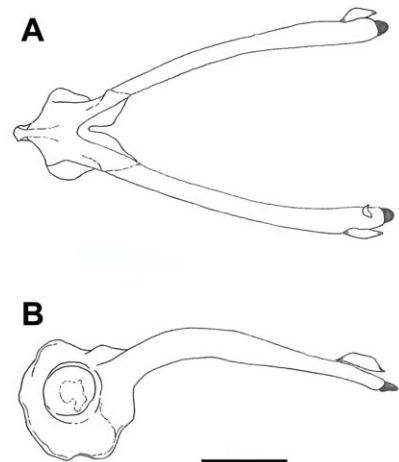


FIG. 10. Pelvic girdle of *Melanophryniscus setiba* (CFBH 15735, female) in **A.** dorsal, and **B.** lateral views. Scale bar = 1.0 mm.

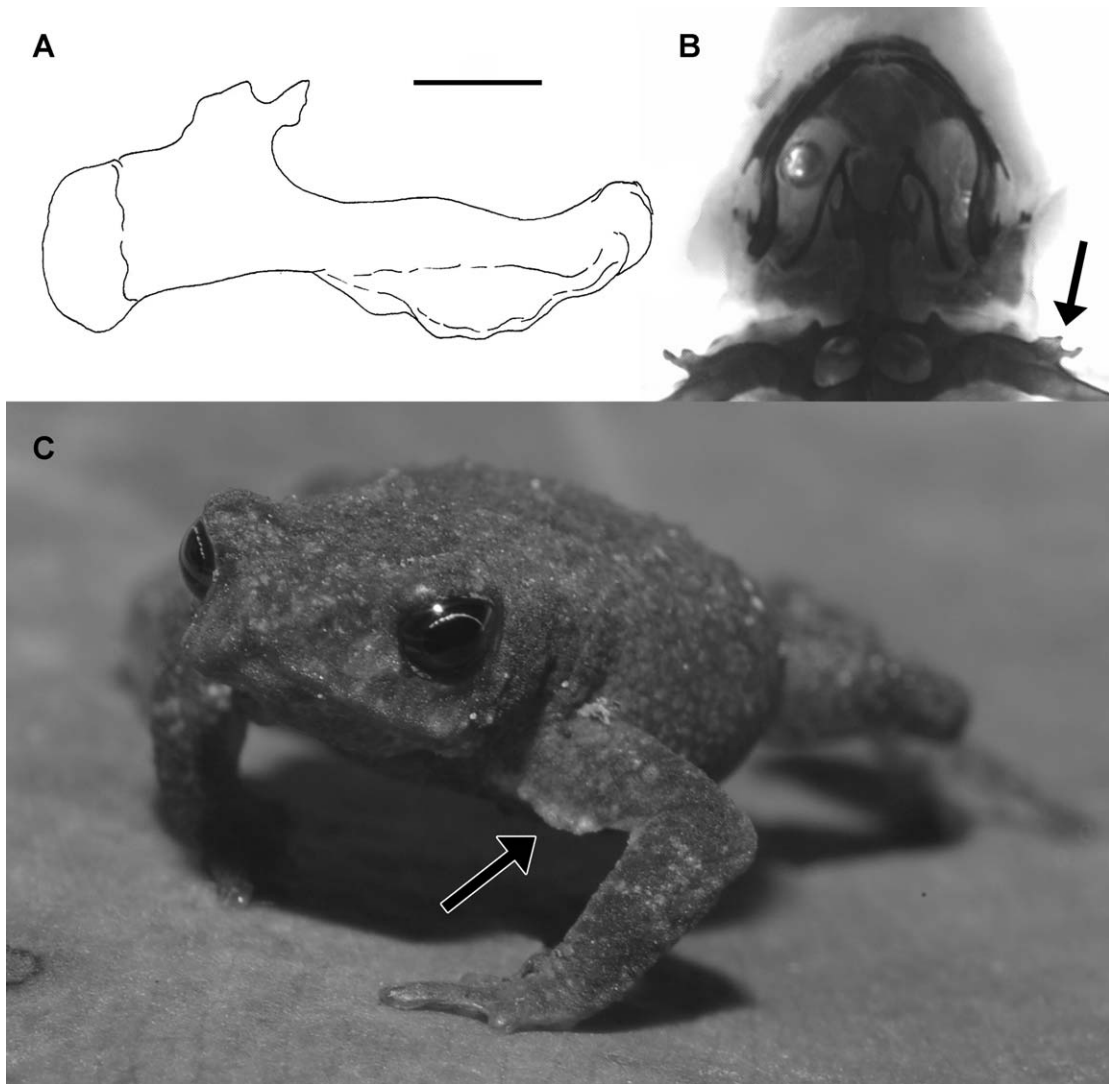


FIG. 11. Humerus and humeral crest/spine of a male *Melanophryniscus setiba* (CFBH 15745, paratype). **A.** Drawing of the humerus in dorsal view; scale bar = 2.0 mm. **B.** Photograph in ventral view; note slightly distinct morphology of spines in the right and left humerus. **C.** Same specimen, in life; note that humeral spine is externally visible in this sex but not protruding out of skin.

breeding site (e.g., eggs, tadpoles) were observed in the field. A dissected female (CFBH 15735) contained eight ova (ca. 2.0 mm each) with a pigmented animal pole and several other small immature eggs. The small ovarian complement and large size of ovarian eggs are suggestive of increased incubation time, and/or hatching in advanced developmental stages (Bradford, 1990; Summers et al., 2006, 2007). The type locality has a large quantity of bromeliad plants that could serve as a potential breeding site for *M. setiba* - phytotelm breeding is present at least in two *Melanophryniscus* species (*M. alipioi* Langone et al., 2008,

and *M. vilavelhensis* Steinbach-Padilha, 2008). Nevertheless, we note that the diameter of oviposited ova in water-breeding *Melanophryniscus* are 1.7–2.0 mm in *M. krauczuki* (Baldo and Basso, 2004), 2.0 in *M. montevidensis* (Langone et al., 2008), 2.0–2.5 in *M. moreirae* (Bokermann, 1967; Starret, 1967), 1.4–2.0 in *M. stelzneri* (Echeverria, 1998; Bustos Singer and Gutiérrez, 1997), and egg diameter in the phytotelm-breeding *M. alipioi* is 2.5–3.0 mm (Langone et al., 2008). The presence of vocal slits and a vocal sac in males suggests that this species vocalizes. However, no calling males were detected, despite several targeted searches at the type locality by the senior author.

Stomachs of two specimens (CFBH 15735, female, and CFBH 15745, male) contained several ants and one mite. The diets of previously studied species of *Melanophryniscus* consist predominantly of ants and mites as well (reviewed by Daly et al., 2008; see also Bonansea and Vaira, 2007; Quiroga et al., 2011). Mites and ants are probably the dietary source of lipophilic alkaloids found in defensive skin secretions of *Melanophryniscus* (Saporito et al., 2011), which suggests that *M. setiba* might also secrete these alkaloids.

Melanophryniscus setiba seems to be locally abundant, although it is difficult to observe due to its small size and cryptic coloration. Most activity was recorded from December through March, which coincides with the rainy season. Other leaf-litter frogs captured in the same forest patch with the new species were *Chiasmocleis carvalhoi*, *Leptodactylus natalensis*, and *Phyllodytes luteolus* cf. *crombiei*. Of these, *C. carvalhoi* and *P. cf. crombiei* were captured simultaneously with the new species in pitfall traps. Some hylids were also detected in the area where we found the new species, especially associated with bromeliads: *Aparasphenodon brunoi*, *Phyllodytes luteolus*, *Scinax agilis*, *S. alter* and *S. argyreornatus*. Among these, *S. argyreornatus* was the only one ever found in the leaf litter.

ETYMOLOGY: The specific name, *setiba*, is derived from the Tupi Guarani language and means “seashells in abundance.” Setiba is the popular name of the region where the type locality is located, likely due to the high number of mollusk shells found in the beaches of the region.

SUGGESTED COMMON NAMES: Restinga toadlet (English); sapinho-da-restinga (Portuguese).

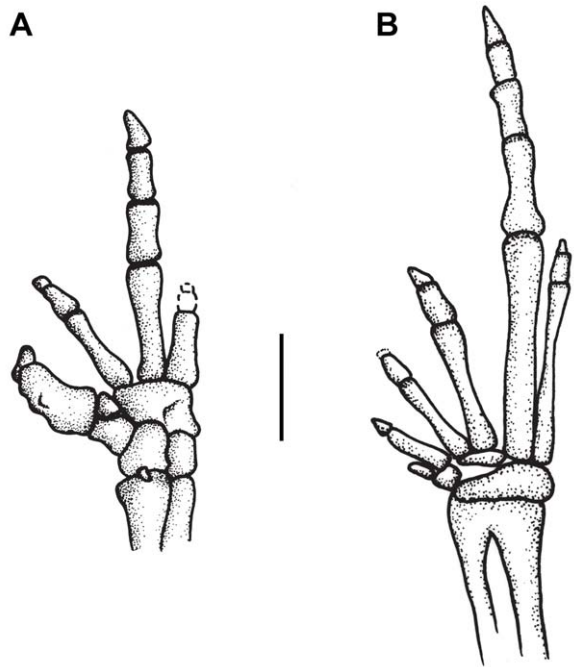


FIG. 12. Dorsal views of **A.** left hand and **B.** right foot of *Melanophryniscus setiba* (CFBH 15745, male). Scale bar = 2.0 mm.

DISCUSSION

Consistent with previous findings (e.g., Graybeal, 1997; Darst and Cannatella, 2004; Frost et al., 2006; Pramuk, 2006; Van Bocxlaer et al., 2010; Pyron and Wiens, 2011), our phylogenetic analysis recovered *Melanophryniscus* as the sister taxon of all other bufonids. *Melanophryniscus setiba*, in turn, was placed as the sister taxon of the remaining exemplars of *Melanophryniscus*. Although our taxon sampling does provide a rigorous test of the placement of *M. setiba* among bufonids and other anurans,⁷ it leaves the question of its placement among the species of *Melanophryniscus* largely unanswered. Of the 26 currently recognized species, only three were included in the present analysis. Following Cruz and Caramaschi (2003), *Melanophryniscus klappenbachii* and *M. fulvoguttatus* are exemplars of the *M. stelzneri* group, and *M. devincenzii* is an exemplar of the *M. tumifrons* groups. Unfortunately, we lack exemplars of the *M. moreirae* group and were also unable to include the recently named phytotelm breeders *M. alipioi* (Langone et al., 2008) and the diminutive *M. vilavelhensis* (unassigned to any group, Steinbach-Padilha, 2008). Importantly, we also failed to include *Truebella* (presumably related to *Melanophryniscus*; Pramuk, 2006). Sampling within *Melanophryniscus* must be greatly increased to test the placement of *M. setiba* relative to other species of this clade.

⁷ The number of recognized bufonid genera is a matter of ongoing discussion (see Frost et al., 2006, 2008, 2009; Pauly et al., 2009; Pyron and Wiens, 2011). We follow Frost (2011), who lists 48 genera, with the addition of *Amazophrynella* (Fouquet et al., 2012a, 2012b). We included 30 out of the 49 genera in our analysis. Most of the exclusions are unlikely to influence the position of the new taxa (e.g., the African genera *Altiphrynoides*, *Churamiti*), but some exclusions are, nonetheless, regrettable (see text). Our analysis was performed with the sole purpose of positioning the new species and we reserve ourselves to not comment any further on outgroup relationships and on the relationships within Bufonidae.



FIG. 13. Habitat of *Melanophryniscus setiba* at the type locality. **A.** General view of an open *Clusia* formation with a *mata seca* type of forest at background, and **B.** microhabitat of the species, sandy soil covered with leaf litter inside the *mata seca*.

McDiarmid (1971) proposed several diagnostic features of *Melanophryniscus*, and Graybeal and Cannatella (1995) further listed as putative synapomorphies: the “absence of the zygomatic ramus of the squamosal, exostosed frontoparietals that diverge anteriorly, ossified orbitosphenoid cartilage, frontoparietals always fused posteriorly, and parasphenoid fused to the underlying chondrocranium...” Most of those synapomorphies were found in the specimens examined by us, including *M. setiba*. The only inconsistency we observed regards the “frontoparietals that diverge anteriorly.” McDiarmid (1971) had already shown that the frontoparietals may not diverge anteriorly in some *Melanophryniscus* and this was confirmed by us when analyzing *M. moreirae*. The frontoparietals also do not diverge anteriorly in *M. setiba*. The apparent absence of a Bidder’s organ in *M. setiba* is consistent with Echeverria’s (1998) observations of *M. stelzneri*. The food items found on stomachs of two specimens of *M. setiba* (ants and mites) are the same food-item classes found on other studied species of *Melanophryniscus* (Filipello and Crespo, 1994; Daly et al., 2008, Quiroga et al., 2011).

In addition to putative synapomorphies *M. setiba* shares with other species of *Melanophryniscus*, it has a number of unusual character states:

(1) Nuptial pad structure. In addition to our report of nuptial pads in *M. setiba*, the taxonomic distribution of nuptial pads in *Melanophryniscus* is poorly known. Several papers dealing with the taxonomy of the genus have omitted any reference to nuptial pads (e.g., Klappenbach, 1968; Caramaschi and Cruz, 2002; Kwet et al., 2005; Di-Bernardo et al., 2006; Steinbach-Padilla, 2008), with the notable exceptions of Ceï (1980), Baldo and Basso (2004), Langone et al. (2008), and Baldo et al. (2012), who mention its presence but do not discuss taxonomic distribution. Nuptial pads are known to be present in *M. alipioi* (Langone et al., 2008), *M. cambaraensis* (Santos et al., 2010), *M. krauczuki* (Baldo and Basso, 2004), *M. pachyrhynchus* (Baldo et al., 2012), *M. rubriventris* (Vaira, 2005), and *M. stelzneri* (Ceï, 1980), and we have observed them in adult males of *M. devincenzii*, *M. moreirae*, *M. simplex*, and *M. tumifrons*. In all these species, the pad occurs on finger II and most frequently on III or even IV (the case of *M. rubriventris*). Our observations indicate that the pad is composed of many minute, keratinized spines. In contrast, the nuptial pad of *M. setiba* has only a few, enlarged keratinized spines at medial margin of finger II (fig. 4C). The enlarged spines present in the pad of *M. setiba* are extremely infrequent in bufonids and are most reminiscent to those described and illustrated by Duellman and Ochoa (1991) in *Nannophryne corynetes* (as *Bufo corynetes*).

(2) Presence of anterior process of hyoid. The anterior process is absent in *M. moreirae*, *M. rubriventris*, *M. krauczuki*, and *M. pachyrhynchus* (McDiarmid, 1971, 1972; Baldo and Basso, 2004; Baldo et al., 2012) but present and well developed in *M. setiba*. An anterior process on the hyale is also present in some species of *Atelopus*, (McDiarmid, 1971) and *Frostius* (Cannatella, 1986).

(3) Small ovarian complement. McDiarmid (1971) mentioned clutches with 78–237 eggs with black animal pole and light vegetal pole for *Melanophryniscus*; Baldo and Basso (2004) reported more than 109 eggs per clutch and up to 401 eggs per breeding event in *M. krauczuki*. We observed an ovarian complement of eight ova in *Melanophryniscus setiba*, the same number of eggs reported for a clutch in *M. vilavelhensis* (Steinbach-Padilha, 2008).

(4) Low phalangeal formula. Previously studied species of *Melanophryniscus* present phalangeal formulae of 2-2-3-3 and 2-2-3-4-3 for hands and feet, respectively (McDiarmid, 1971; Baldo and Basso, 2004). In contrast, the phalangeal formulae are 1-2-3-2 for hands and 1-2-2-4-2 or 1-2-3-4-2 in feet for *M. setiba*. Phalangeal loss, involving fingers II and V, and toes I, III, and V follows the general pattern described by Alberch and Gale (1985) in anurans (see also Yeh, 2002). Phalangeal losses occur also occur in other bufonids, such as some species of *Ateolopus*, *Dendrophryniscus*, *Didynamipus*, *Incilius*, *Mertensophryne*, and *Osornophryne* (Savage and Kluge, 1961; McDiarmid, 1971; Ruiz-Carranza and Hernández-Camacho, 1976; Grandison, 1981).

(5) Spinelike humeral process. The highly developed ventral crest of the humerus is apparently a novelty among bufonids. While a notable ventral crest was reported by Perret (1972) and Boistel and Amiet (2001) for *Wolterstorffina mirei* and *W. chirioi* respectively, it is not elaborated into a spine. Similar processes, in shapes that vary from an elevated ridge to a sharply pointed, curved spine, are present at least in Centrolenidae (Cisneros-Heredia and McDiarmid, 2007), Hylidae (Bokermann, 1965), Microhylidae (Matsui, 2009), Telmatobiidae (Lavilla and Sandoval, 1991), and Rhacophoridae (Kuramoto and Joshy, 2003).

CONSERVATION REMARKS

The coastal region of Brazil is one of the country's most disturbed and exploited areas (Morelato and Haddad, 2000). The demographic density of the coastal zone is much higher than the country's average (Ministério do Meio Ambiente, 2002) and, as a result, the remaining vegetated coastal strips have been intensely affected by human activity and a consequent degradation of the habitats (Morelato and Haddad, 2000).

Restingas are relatively fragile coastal environments and the clearing of vegetation results in further difficulties for habitat regeneration. Surprisingly, according to an extensive survey on the restingas of southeastern Brazil (Rocha et al., 2003), these coastal habitats, despite being in the highest human density zone, remain one of the least-known environments, not only in biodiversity, but also regarding conservation status of its remnants. The discovery of this species in Guarapari reinforces the notion that the Atlantic rainforest and its related habitats, such as restingas, are home to a great and still poorly known biodiversity. Several other vertebrate species have been recently described from restinga habitats across coastal Brazil (see examples in Rocha et al., 1997, 2000; Dias et al., 2002, Peixoto et al., 2003, Verrastro et al., 2003; Izecksohn et al., 2009). The high level of degradation of restinga habitats is a threat to survival of many species, especially to those endemic or rare.

Recognition of *Melanophryniscus setiba* as another case of endemism of restinga habitat seems tempting; however, it is possible that this species occurs in nearby forested areas outside the restinga domain. Some cases of amphibian species endemic to restinga habitats in southeastern Brazil were reported by Rocha et al. (2005)—*Scinax agilis*, *S. littoreus* and *Xenohyla truncata* (Hylidae), *Rhinella pygmaea* (Bufonidae), and *Leptodactylus marambaiae* (Leptodactylidae)—but at least *R. pygmaea* was shown not to be endemic to restingas (Silva et al., 2007).

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REFERENCES

- Alberch, P., and E.A. Gale. 1985. A developmental analysis of an evolutionary trend: digital reduction in amphibians. *Evolution* 39: 8–23.
- Baldo, D., and N.G. Basso. 2004. A new species of *Melanophryniscus* Gallardo, 1961 (Anura: Bufonidae), with comments on the species of the genus reported for Misiones, northeastern Argentina. *Journal of Herpetology* 38: 393–403.
- Baldo, D., et al. 2012. The taxonomic status of *Melanophryniscus orejasmirandai* Prigioni & Langone, 1987 “1986” (Anura: Bufonidae). *Zootaxa* 3235: 45–61.
- Boistel, R., and J.L. Amiet. 2001. Une nouvelle espèce de *Wolterstorffina* (Amphibia, Anura, Bufonidae) de l'étage afro-subalpin du Mont Okou (Cameroun). *Alytes* 18: 127–140.
- Bokermann, W.C.A. 1965 *Hyla langei*, a new frog from Paraná, southern Brazil. *Journal of the Ohio Herpetological Society* 5: 49–51.
- Bokermann, W.C.A. 1967. Observações sobre *Melanophryniscus moreirae* (Mir. Rib.) (Amphibia-Brachycephalidae). *Anais da Academia Brasileira de Ciências* 39: 301–306.
- Bonansea, M.I., and M. Vaira. 2007. Geographic variation of the diet of *Melanophryniscus rubriventris* (Anura: Bufonidae) in northwestern Argentina. *Journal of Herpetology* 41: 231–236.
- Bradford, D.F. 1990. Time and rate of embryonic incubation in amphibians: the influence of ovum size, temperature, and reproductive mode. *Physiological Zoology* 63: 1157–1180.

- Bremer, K. 1988. The limits of amino acid sequence data in angiosperm phylogenetic reconstruction. *Evolution* 42: 795–803.
- Bustos Singer, R., and M. Gutiérrez. 1997. Reproducción y desarrollo larval del sapo enano *Melanophryniscus stelzneri stelzneri* (Weyemberg, 1875) (Anura: Bufonidae). *Cuadernos de Herpetología* 11: 21–30.
- Cannatella, D.C. 1986. A new genus of bufonid (Anura) from South America, and phylogenetic relationships of the Neotropical genera. *Herpetologica* 42: 197–205.
- Caramaschi, U., and C.A.G. Cruz. 2002. Taxonomic status of *Atelopus pachyrhynchus* Miranda–Ribeiro, 1920, redescription of *Melanophryniscus tumifrons* (Boulenger, 1905), and descriptions of two new species of *Melanophryniscus* from the state of Santa Catarina, Brazil (Amphibia, Anura, Bufonidae). *Arquivos do Museu Nacional, Rio de Janeiro* 60: 303–314.
- Cechin, S.Z., and M. Martins. 2000. Eficiência de armadilhas de queda (*pitfall traps*) em amostragens de anfíbios e répteis no Brasil. *Revista Brasileira de Zoologia* 17: 729–740.
- Cei, J.M. 1980. Amphibians of Argentina. *Monitore Zoologico Italiano Monografia N.S.* 2: 1–609.
- Cruz, C.A.G., and U. Caramaschi. 2003. Taxonomic status of *Melanophryniscus stelzneri dorsalis* (Mertens, 1933) and *Melanophryniscus stelzneri fulvoguttatus* (Mertens, 1937) (Amphibia, Anura, Bufonidae). *Boletim do Museu Nacional, Nova Série, Zoologia* 500: 1–11.
- Cisneros-Heredia, D.F., and R.W. McDiarmid. 2007. Revision of the characters of Centrolenidae (Amphibia: Anura: Athesphatanura), with comments on its taxonomy and the description of new taxa of glassfrogs. *Zootaxa* 1572: 3–82.
- Daly, J.W., et al. 2008. Indolizidine 239Q and quinolizidine 275I. Major alkaloids in two Argentinian bufonid toads (*Melanophryniscus*). *Toxicon* 52: 858–870.
- Darst, C.R., and D.C. Cannatella. 2004. Novel relationships among hyloid frogs inferred from 12S and 16S mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 31: 462–475.
- Di-Bernardo, M., R. Maneyro, and H. Grillo. 2006. New species of *Melanophryniscus* (Anura: Bufonidae) from Rio Grande do Sul, southern Brazil. *Journal of Herpetology* 40: 261–266.
- Dias, E.J.R., C.F.D. Rocha, and D. Vrcibradic. 2002. A new *Cnemidophorus* (Squamata: Teiidae) from Bahia State, northeastern Brazil. *Copeia* 2002: 1070–1077.
- Duellman, W.E., and O.M. Ochoa. 1991. A new species of *Bufo* (Anura: Bufonidae) from the Andes of southern Peru. *Copeia* 1991: 137–141.
- Echeverria, D.D. 1998. Aspectos de la reproducción in vitro de y del desarrollo larval *Melanophryniscus stelzneri* (Weyembergh, 1875) (Amphibia, Bufonidae) con comentarios acerca del órgano de Bidder. *Alytes* 15: 158–170.
- Eiten, G. 1992. Natural Brazilian vegetation types and their causes. *Anais da Academia Brasileira de Ciências* 64: 35–65.
- Fabrezi, M. 1992. El carpo de los anuros. *Alytes* 10: 1–29.
- Fabrezi, M. 1993. The anuran tarsus. *Alytes* 11: 47–63.
- Fabrezi, M., and P. Alberch. 1996. The carpal elements of anurans. *Herpetologica* 52: 188–204.
- Faivovich, J., et al. 2005. Systematic review of the frog family Hylidae, with special reference to Hylinae: phylogenetic analysis and taxonomic revision. *Bulletin of the American Museum of Natural History* 294: 1–240.
- Faivovich, J., et al. 2010. The phylogenetic relationships of the charismatic poster frogs, Phyllomedusinae (Anura, Hylidae). *Cladistics* 26: 227–261.
- Filipello, A.M., and F.A. Crespo. 1994. Alimentación en *Melanophryniscus stelzneri* (Anura: Bufonidae). *Cuadernos de Herpetología* 8: 18–24.

- Ford, L.S., and D.C. Cannatella. 1993. The major clades of frogs. *Herpetological Monographs* 7: 94–117.
- Fouquet, A., et al. 2012. Molecular phylogeny and morphometric analyses reveal deep divergence between Amazonia and Atlantic Forest species of *Dendrophryniscus*. *Molecular Phylogenetics and Evolution* 62: 826–383.
- Fouquet, A., et al. 2012. *Amazonella* Fouquet *et al.*, 2012 (Anura: Bufonidae) junior homonym of *Amazonella* Lundblad, 1931 (Acari: Unionicolidae): proposed replacement by *Amazophrynella* nom. nov. *Zootaxa* 3244: 68.
- Frost, D.R. 2011. Amphibian species of the world: an online reference, version 5.5 (January, 2011). New York: American Museum of Natural History. Internet resource (<http://research.amnh.org/herpetology/amphibia/index.html>), accessed March 4, 2012.
- Frost, D.R., et al. 2006. The amphibian tree of life. *Bulletin of the American Museum of Natural History* 297: 1–370.
- Frost, D.R., et al. 2008. Is *The Amphibian Tree of Life* really fatally flawed? *Cladistics* 24: 385–395.
- Frost, D.R., R.W. McDiarmid, and J.R. Mendelson III. 2009. Response to the point of view of Gregory B. Pauly, David M. Hillis, and David C. Cannatella, by the Anuran Subcommittee of the SSAR/HL/ASIH Scientific and Standard English Names List. *Herpetologica* 65: 136–153.
- Goloboff, P.A. 1999. Analyzing large data sets in reasonable times: solutions for composite optima. *Cladistics* 15: 415–428.
- Goodman, M., C.B. Olson, J.E. Beeber, and J. Czelusniak. 1982. New perspectives in the molecular biological analysis of mammalian phylogeny. *Acta Zoologica Fennica* 169: 19–35.
- Grandison, A. 1981. Morphology and phylogenetic position of the West African *Didynamipus sjoestedti* Andersson, 1903 (Anura Bufonidae). *Monitore Zoologico Italiano Supplemento N.S.* 15: 187–215.
- Grant, T., et al. 2006. Phylogenetic systematics of dart-poison frogs and their relatives (Amphibia: Athesphatanura: Dendrobatidae). *Bulletin of the American Museum of Natural History* 299: 1–262.
- Grant, T., and A.G. Kluge. 2008. Credit where credit is due: the Goodman-Bremer support metric. *Molecular Phylogenetics and Evolution* 49: 405–406.
- Grant T., and A.G. Kluge. 2009. Parsimony, explanatory power, and dynamic homology testing. *Systematics and Biodiversity* 7: 357–363.
- Graybeal, A. 1997. Phylogenetic relationships of bufonid frogs and tests of alternate macroevolutionary hypotheses characterizing their radiation. *Zoological Journal of the Linnean Society* 119: 297–338.
- Graybeal, A., and D.C. Cannatella. 1995. A new taxon of Bufonidae from Peru, with descriptions of two new species and a review of the phylogenetic status of supraspecific bufonid taxa. *Herpetologica* 51: 105–131.
- Haas, A. 2003. Phylogeny of frogs as inferred from primarily larval characters (Amphibia: Anura). *Cladistics* 19: 23–90.
- Heyer, W.R., S.A. Rand, C.A.G. Cruz, O.L. Peixoto, and C.E. Nelson. 1990. Frogs of Boracéia. *Arquivos de Zoologia* 31: 231–410.
- Izecksohn, E., S.P. Carvalho-e-Silva, and O.L. Peixoto. 2009. Sobre *Gastrotheca fissipes* (Boulenger, 1888), com descrição de uma nova espécie (Amphibia, Anura, Amphignathodontidae). *Arquivos do Museu Nacional (Rio de Janeiro)* 67: 81–91.
- Klappenbach, M.A. 1968. Notas herpetológicas, IV. El género *Melanophryniscus* (Amphibia, Salientia) en el Uruguay, con descripción de dos nuevas especies. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo* 9: 1–12.
- Kluge, A.G., and T. Grant. 2006. From conviction to anti-superfluity: old and new justifications for parsimony in phylogenetic inference. *Cladistics* 22: 276–288.

- Kuramoto M., and S.H. Joshy. 2003. Two new species of the genus *Philautus* (Anura: Rhacophoridae) from the Western Ghats, southwestern India. *Current Herpetology* 22: 51–60.
- Kwet, A., R. Maneyro, A., Zillikens, and D. Mebs. 2005. Advertisement calls of *Melanophryniscus dorsalis* (Mertens, 1933) and *M. montevidensis* (Philippi, 1902), two parapatric species from southern Brazil and Uruguay, with comments on morphological variation in the *Melanophryniscus stelzneri* group (Anura: Bufonidae). *Salamandra* 41: 3–20.
- Langone, J.A., M.V. Segalla, M. Bornschein, and R.O. de Sá. 2008. A new reproductive mode in the genus *Melanophryniscus* Gallardo, 1961 (Anura: Bufonidae) with description of a new species from the state of Paraná, Brazil. *South American Journal of Herpetology* 3: 1–9.
- Lavilla, E.O., and P.E. Sandoval. 1991. A new Bolivian species of the genus *Telmatobius* (Anura: Leptodactylidae) with a humeral spine. *Amphibia-Reptilia* 20: 55–64.
- Matsui, M. 2009. A new species of *Kalophrynus* with a unique male humeral spine from Peninsular Malaysia (Amphibia, Anura, Microhylidae). *Zoological Science* 26: 579–585.
- McDiarmid, R.W. 1971. Comparative morphology and evolution of frogs of the Neotropical genera *Atelopus*, *Dendrophryniscus*, *Melanophryniscus* and *Oreophrynella*. *Bulletin of the Los Angeles County Museum of Natural History* 12: 1–66.
- McDiarmid, R.W. 1972. La posición sistemática de la rana *Atelopus rubriventris* Vellard. *Physis* 31: 15–21.
- Ministério do Meio Ambiente. 2002. Avaliação e identificação de áreas prioritárias para a conservação, utilização sustentável e repartição dos benefícios da biodiversidade nos biomas brasileiros. Brasília, Brazil: MMA/SBF.
- Morelato, L.P.C., and C.F.B. Haddad. 2000. Introduction: the Brazilian Atlantic Forest. *Biotropica* 32: 786–792.
- Naya, D.E., J.A. Langone, and R.O. de Sá. 2004. Características histológicas de la tumefacción frontal de *Melanophryniscus* (Amphibia: Anura: Bufonidae). *Revista Chilena de Historia Natural* 77: 593–598.
- Nixon, K.C. 1999. The parsimony ratchet, a new method for rapid parsimony analysis. *Cladistics* 15: 407–414.
- Pauly, G.B., D.M. Hillis, and D.C. Cannatella. 2009. Taxonomic freedom and the role of official lists of species names. *Herpetologica* 65: 115–128.
- Peixoto, O.L., U. Caramaschi, and E.M.X. Freire. 2003. Two new species of *Phyllodytes* (Anura, Hylidae) from the state of Alagoas, northeastern Brazil. *Herpetologica* 59: 235–246.
- Pereira, O.J. 1990. Caracterização fitofisionômica da restinga de Setiba – Guarapari/ES. In S. Watanabe (organizer), *Anais do II Simpósio de Ecossistemas da Costa Sul e Sudeste Brasileira*, 3: 207–219. São Paulo: ACIESP.
- Perret, J.L. 1972. Les espèces des genres *Wolterstorffina* et *Nectophrynoides* d'Afrique (Amphibia, Bufonidae) *Annales de la Faculté des Sciences du Cameroun* 11: 93–119.
- Pramuk, J.B. 2006. Phylogeny of south American *Bufo* (Anura: Bufonidae) inferred from combined evidence. *Zoological Journal of the Linnean Society* 146: 407–452.
- Pramuk, J.B., T. Robertson, J.W. Sites, Jr., and B.P. Noonan. 2007. Around the world in 10 million years: biogeography of the nearly cosmopolitan true toads (Anura: Bufonidae). *Global Ecology and Biogeography* 17: 72–83.
- Pyron, R.A., and J.J. Wiens. 2011. A large-scale phylogeny of Amphibia including over 2,800 species, and a revised classification of extant frogs, salamanders, and caecilians. *Molecular Phylogenetics and Evolution* 61: 543–583.
- Quiroga M.F., M.I. Bonansea, and M. Vaira. 2011. Population diet variation and individual specialization in the poison toad, *Melanophryniscus rubriventris* (Vellard, 1947). *Amphibia-Reptilia* 32: 261–265

- Rocha, C.F.D., H.G. Bergallo, and D. Peccinini-Seale. 1997. Evidence of an unisexual population of the Brazilian whiptail lizard genus *Cnemidophorus* (Teiidae) with description of a new species. *Herpetologica* 53: 374–382.
- Rocha, C.F.D., A.F.F. Araújo, D. Vrcibradic, and E.M. Costa. 2000. New *Cnemidophorus* (Squamata: Teiidae) from coastal Rio de Janeiro State, southeastern Brazil. *Copeia* 2000: 501–509.
- Rocha, C.F.D., H.G. Bergallo, M.A.S. Alves, and M.A. Van Sluys. 2003. A biodiversidade nos grandes remanescentes florestais do Estado do Rio de Janeiro e nas restingas da Mata Atlântica. Rio de Janeiro: Rima.
- Rocha, C.F.D., M.A. Van Sluys, H.G. Bergallo, and M.A.S. Alves. 2005. Endemic and threatened tetrapods in the restingas of the biodiversity corridors of Serra do Mar and of the Central da Mata Atlântica in eastern Brazil. *Brazilian Journal of Biology* 65: 159–168.
- Roelants, K., et al. 2007. Global patterns of diversification in the history of modern amphibians. *Proceedings of the National Academy of Sciences of the United States of America* 104: 887–892.
- Ruiz-Carranza, P.M., and J.I. Hernández-Camacho. 1976. *Osornophryne*, género nuevo de anfibios bufónidos de Colombia y Ecuador. *Caldasia* 11: 93–148.
- Santos, R.R., and T. Grant. 2011. Diel pattern of migration in a poisonous toad from Brazil and the evolution of chemical defenses in diurnal amphibians. *Evolutionary Ecology* 25: 249–258.
- Santos, R.R., S.B. Leonardi, V.Z.C. Caorsi, and T. Grant. 2010. Directional orientation of migration in an aseasonal explosive-breeding toad from Brazil. *Journal of Tropical Ecology* 26: 415–421.
- Saporito, R.A., M.A. Donnelly, T.F. Spande, and H.M. Garraffo. 2011. A review of chemical ecology in poison frogs. *Chemoecology*: 1–10.
- Savage, J.M., and A.G. Kluge. 1961. Rediscovery of the strange Costa Rica toad, *Crepidius epioticus* Cope. *Revista de Biología Tropical* 9: 39–51.
- Silva, G.R., S.P. Carvalho-e-Silva, and A.M.P.T. Carvalho-e-Silva. 2007. *Chaunus pygmaeus*. geographical distribution. *Herpetological Review* 38: 97.
- Starrett, P.H. 1967. Observations on the life history of frogs of the family Atelopodidae. *Herpetologica* 23: 195–204.
- Steinbach-Padilha, G.C. 2008. A new species of *Melanophryniscus* (Anura, Bufonidae) from the Campos Gerais region of southern Brazil. *Phyllomedusa* 7: 98–108.
- Suguio, K., and M.G. Tessler. 1984. Planícies costeiras de cordões litorâneos Quaternários do Brasil: origem e nomenclatura. In L.D. Lacerda, D.S.D. Araújo, R. Cerqueira, and B. Turcq (editors), *Restingas: origem, estrutura, processos*: 12–25. Niterói, Brazil: Centro Editorial da Universidade Federal Fluminense.
- Summers, K., C.S. McKeon, and H.E. Heying. 2006. The evolution of parental care and egg size: a comparative analysis in frogs. *Proceedings of the Royal Society Biological Sciences Series B* 273: 687–692.
- Summers, K., C.S. McKeon, H.E.J. Heying, and W. Patrick. 2007. Social and environmental influences on egg size evolution in frogs. *Journal of Zoology* 271: 225–232.
- Taylor, W., and G.C. Van Dyke. 1985. Revised procedures for staining and cleaning small fishes and other vertebrates for bone and cartilage study. *Cybium* 9: 107–119.
- Trueb, L. 1973. Bones, frogs and evolution. In J.L. Vial (editor), *Evolutionary biology of the anurans: contemporary research on major problems*: 65–132. Columbia: University of Missouri Press.
- Trueb, L. 1993. Patterns of cranial diversity among the Lissamphibia. In J. Hanken, and B.K. Hall (editors), *Patterns of structural and systematic diversity*: 65–132. Chicago: University of Chicago Press.

- Vaira, M. 2005. Annual variation of breeding patterns of the toad, *Melanophryniscus rubriventris* (Vellard, 1947). *Amphibia-Reptilia* 26: 193–199.
- Van Bocxlaer, I., et al. 2010. Gradual adaptation toward a range-expansion phenotype initiated the global radiation of toads. *Science* 327: 679–682.
- Varón, A., L.S. Vinh, W.C. Wheeler. 2010. POY version 4: phylogenetic analysis using dynamic homologies. *Cladistics* 26: 72–85.
- Varón, A., N. Lucaroni, L. Hong, and W.C. Wheeler. 2012. POY 5.0. Black Sabbath Development build b7e482d8509e R.C. New York: American Museum of Natural History. Program and documentation available online (<http://research.amnh.org/scicomp/projects/poy.php>).
- Verrastro, L., L. Veronese, C. Bujes, and M.M. Dias-Filho. 2003. A new species of *Liolaemus* from southern Brazil (Iguania: Tropiduridae). *Herpetologica* 59: 105–118.
- Wheeler, W.C. 2003. Implied alignment: a synapomorphy-based multiple sequence alignment method. *Cladistics* 19: 261–268.
- Wheeler, W.C. 2012. Trivial minimization of extra-steps under dynamic homology. *Cladistics* 28: 188–189.
- Wheeler, W.C., et al. 2006. Dynamic homology and phylogenetic systematics: a unified approach using POY. New York: American Museum of Natural History.
- Yeh, J. 2002. The effect of miniaturized body size on skeletal morphology in frogs. *Evolution* 56: 628–641.

APPENDIX 1

ADDITIONAL SPECIMENS EXAMINED

Atelopus spumarius: MPEG 2875 (Serra Norte, Carajás, Pará, Brasil). *Amazophrynella minuta*: MPEG 4260, 4560–61, 4553 (Faro, Pará, Brasil). *Frostius pernambucensis*: MNRJ 33431 *Melanophryniscus* sp.: MPEG 6076 (Itatiaia, São Paulo, Brasil). *Melanophryniscus moreirae*: AMNH 16997, 16999, CFBH 296–97; *Melanophryniscus krauczuki*: MACN 39957–58; *Melanophryniscus devincenzii*: MACN 36856; *Melanophryniscus montevidensis*: AMNH 71171–74; *Melanophryniscus rubriventris*: MACN 33218–19; *Melanophryniscus stelzneri*: AMNH 77710; *Melanophryniscus* sp. (*gr. tumifrons*) MACN 39959–61.

APPENDIX 2

SPECIES INCLUDED IN THE PHYLOGENETIC ANALYSIS

Genbank accession numbers for the sequences employed in the phylogenetic analysis. Sequences were produced by Darst and Cannatella (2004), Faivovich et al. (2005), Frost et al. (2006), Grant et al. (2006), Pramuk (2006), Pramuk et al. (2007), and Roelants et al. (2007). Species in bold are those for which we are providing original sequences.

	12S + tRNAVal + 16S	RAG-1	CXCR4	Rhodopsin Exon 1
<i>Acris crepitans</i>	AY843559	—	EF107468	AY844533
<i>Adenomus kelaartii</i>	EF107161	—	EF107447	—
<i>Allobates trilineatus</i>	DQ501998	—	—	DQ503184
<i>Allophryne ruthveni</i>	AY843564	—	—	AY844538
<i>Alsodes gargola</i>	AY843565	—	—	AY844539
<i>Amazophrynella minuta</i>	AY843582	DQ158346	DQ306496	AY844555
<i>Ameerga trivittata</i>	DQ502201	—	—	DQ503190
<i>Amietophrynus kisolensis</i>	DQ158464	—	DQ306560	—
<i>Amietophrynus maculatus</i>	DQ158469	DQ158389	DQ306533	DQ284005
<i>Amietophrynus poweri</i>	DQ158482	DQ158401	DQ306559	—
<i>Amietophrynus regularis</i>	DQ158485	DQ158404	DQ306523	DQ283858
<i>Amietophrynus steindachneri</i>	DQ158488	DQ158406	DQ306546	—
<i>Amietophrynus xeros</i>	DQ158499	DQ158414	DQ306561	—
<i>Amietophrynus brauni</i>	DQ158437	DQ158361	DQ306514	DQ284401
<i>Amietophrynus camerunensis</i>	DQ158439	DQ158363	DQ306555	DQ283979
<i>Amietophrynus garmani</i>	DQ158453	DQ158375	DQ306547	—
<i>Amietophrynus gracilipes</i>	DQ158456	DQ158378	DQ306522	—
<i>Amietophrynus gutturalis</i>	DQ158460	DQ158382	DQ306556	DQ284035
<i>Amietophrynus latifrons</i>	DQ283343	—	—	DQ283970
<i>Amietophrynus tuberosus</i>	DQ283362	—	—	DQ283984
<i>Anaxyrus americanus</i>	DQ158426	DQ158352	DQ306520	—
<i>Anaxyrus boreas</i>	DQ158498	—	DQ306499	DQ283871
<i>Anaxyrus cognatus</i>	DQ158444	DQ158367	DQ306502	—
<i>Anaxyrus debilis</i>	DQ158449	DQ158371	DQ306507	—
<i>Anaxyrus exsul</i>	DQ158450	DQ158372	DQ306550	—
<i>Anaxyrus fowleri</i>	DQ158451	DQ158373	DQ306505	—
<i>Anaxyrus microscaphus</i>	DQ158476	DQ158395	DQ306563	—
<i>Anaxyrus punctatus</i>	DQ283160	—	—	DQ283855
<i>Anaxyrus quercicus</i>	DQ158484	DQ158403	DQ306562	—
<i>Anaxyrus terrestris</i>	DQ158489	—	DQ306537	DQ283854
<i>Anaxyrus woodhousii</i>	DQ158498	DQ158413	DQ306551	DQ283875
<i>Ansonia longidigitata</i>	DQ283341	—	—	DQ283968
<i>Atelognathus patagonicus</i>	AY843571	—	—	AY844545
<i>Atelopus flavescens</i>	DQ283259	—	—	DQ283928
<i>Atelopus peruensis</i>	DQ158419	DQ158345	DQ306495	—
<i>Atelopus spumarius</i>	DQ283260	—	—	DQ283929
<i>Atelopus spurrelli</i>	DQ502200	—	—	—
<i>Atelopus varius</i>	AY325996	—	—	—

	12S + tRNAVal + 16S	RAG-1	CXCR4	Rhodopsin Exon 1
<i>Atelopus zeteki</i>	DQ283252	—	—	—
<i>Batrachyla leptopus</i>	AY843572	—	—	AY844546
<i>Brachycephalus ephippium</i>	DQ283091	—	—	—
<i>Bufo bufo</i>	DQ158438	DQ158362	DQ306504	—
<i>Bufo gargarizans</i>	DQ158428	DQ158353	DQ306531	DQ283905
<i>Centrolene prosoblepon</i>	AY843574	AY364223	AY364193	AY844548
<i>Ceratophrys cornuta</i>	AY326014	AY364218	DQ306491	—
<i>Ceratophrys cranwelli</i>	AY843575	—	—	—
<i>Chacophrys pierottii</i>	DQ283328	—	—	—
<i>Cochranella bejaranoi</i>	AY843576	—	—	AY844372
<i>Crossodactylus schmidtii</i>	AY843579	—	—	AY844552
<i>Cruziophyla calcarifer</i>	AY843562	—	—	AY844536
<i>Cryptobatrachus sp.</i>	AY326050	—	—	—
<i>Cycloramphus boraceiensis</i>	DQ283097	—	—	DQ283813
<i>Dendrobates auratus</i>	AY843581	—	AY364184	AY844554
<i>Didynamipus sjostedti</i>	AY325991	—	—	—
<i>Duttaphrynus melanostictus</i>	DQ158475	DQ158394	DQ306508	DQ283967
<i>Edalorhina perezi</i>	AY843585	—	—	AY844558
<i>Epipedobates boulengeri</i>	DQ283037	—	—	—
<i>Eupsophus calcaratus</i>	AY843587	—	—	AY844560
<i>Fejervarya limnocharis</i>	AY843588	—	—	AY844561
<i>Frostius erythrophthalmus</i>^a	JX961680	—	—	—
<i>Gastrotheca cornuta</i>	AY843591	—	—	—
<i>Gastrotheca fissipes</i>	AY843592	—	—	AY844564
<i>Gastrotheca pseustes</i>	AY326051	—	—	—
<i>Heleophryne purcelli</i>	AY843593	AY364221	AY364191	AY844565
<i>Hemiphractus helioi</i>	AY843594	AY844382	—	AY844566
<i>Hemisus marmoratus</i>	AY326070	AY364216	AY364186	DQ284029
<i>Hyalinobatrachium fleischmanni</i>	DQ283453	—	—	DQ284043
<i>Hyla cinerea</i>	AY549327	AY844408	DQ306493	AY844597
<i>Hylodes phyllodes</i>	DQ283096	—	—	DQ283812
<i>Hypodactylus sp.</i> ^b	AY326010	—	—	—
<i>Incilius alvarius</i>	DQ158425	DQ158351	DQ306516	DQ283933
<i>Incilius coccifer</i>	DQ158443	DQ158366	DQ306526	—
<i>Incilius valliceps</i>	DQ158493	DQ158409	DQ306545	—
<i>Incilus coniferus</i>	DQ158445	—	DQ306534	DQ283860
<i>Incilus luetkenii</i>	DQ158467	DQ158387	DQ306565	—
<i>Ingerophrynus divergens</i>	DQ283149	—	—	DQ283849
<i>Ingerophrynus galeatus</i>	DQ158452	DQ158374	DQ306506	DQ283995
<i>Ingerophrynus macrotis</i>	DQ158468	DQ158388	DQ306525	—
<i>Kaloula conjuncta</i>	AY326064	—	—	—
<i>Lepidobatrachus laevis</i>	DQ283152	—	EF107461	DQ283851
<i>Leptodactylus latrans</i>	DQ158417	DQ158343	DQ306492	AY844681
<i>Leptodactylus petersi</i>	DQ283063	—	—	DQ283790
<i>Leptophryne borbonica</i>	EF107164	—	EF107450	—

	12S + tRNAVal + 16S	RAG-1	CXCR4	Rhodopsin Exon 1
<i>Limnodynastes salmini</i>	AY326071	AY364219	AY364189	—
<i>Limnomedusa macroglossa</i>	AY843689	—	—	AY844682
<i>Lithodytes lineatus</i>	AY843690	—	—	AY844683
<i>Litoria aurea</i>	AY843691	—	—	AY844684
<i>Litoria freycineti</i>	AY843693	—	—	—
<i>Litoria infrafronata</i>	AY843694	—	—	AY844687
<i>Litoria meiriana</i>	AY843695	—	—	AY844688
<i>Lysapsus limellum</i>	AY843697	—	—	AY844690
<i>Mantidactylus femoralis</i>	AY843698	—	—	—
<i>Megaelosia goeldii</i>	DQ283072	—	—	DQ283797
<i>Melanophryniscus devincenzii</i>^c	JX961678	—	—	JX961676
<i>Melanophryniscus fulvoguttatus</i> ^d	DQ158421	DQ158347	DQ306494	—
<i>Melanophryniscus klappenbachi</i>	AY843699	—	—	DQ283765
<i>Melanophryniscus setiba</i>^e	JX961679	—	JX961675	JX961677
<i>Mertensophryne anotis</i>	AF220910	—	—	—
<i>Mertensophryne micranotis</i>	EF107207	—	EF107491	—
<i>Nannophryne cophotis</i>	DQ158446	DQ158369	DQ306540	—
<i>Nannophryne variegatus</i>	DQ158494	DQ158410	DQ306515	—
<i>Nectophryne afra</i>	DQ283360	—	—	DQ283981
<i>Nectophryne batesii</i>	DQ283169	—	—	—
<i>Nectophrynoides tornieri</i>	DQ283413	—	EF107490	DQ284018
<i>Neobatrachus sudelli</i>	AY843700	—	—	AY844691
<i>Odontophrynus americanus</i>	AY843704	—	—	AY844695
<i>Oreophrynella dendronastes</i>	DQ158422	—	—	—
<i>Osornophryne guacamayo</i>	AY326036	—	—	—
<i>Paratelmatobius sp.</i>	DQ283098	—	—	DQ283814
<i>Pedostibes hosii</i>	DQ283164	—	EF107449	DQ283859
<i>Peltophryne lemur</i>	DQ158465	DQ158386	DQ306513	—
<i>Phrynoidis asper</i>	DQ158431	DQ158356	—	DQ283848
<i>Phrynoidis juxtasper</i>	DQ158463	DQ158385	DQ306542	—
<i>Phyllobates bicolor</i>	DQ502181	—	—	—
<i>Phyllobates terribilis</i>	DQ502157	—	—	DQ503244
<i>Phyllodytes luteolus</i>	AY843721	—	—	AY844708
<i>Physalaemus cuvieri</i>	AY843729	—	—	AY844717
<i>Platymantis sp.</i>	AY326061	—	—	—
<i>Pleurodema brachyops</i>	AY843733	—	—	AY844721
<i>Pristimantis rhabdolaemus</i> ⁶	AY843586	—	—	AY844559
<i>Pristimantis thymelensis</i>	AY326009	—	—	—
<i>Pristimantis w-nigrum</i>	AY326004	DQ158344	—	—
<i>Pseudepidalea viridis</i>	DQ283279	—	—	DQ283940
<i>Pseudopaludicola falcipes</i>	AY843741	—	—	AY844728
<i>Pseudophryne bibroni</i>	AY843988	—	—	AY844729
<i>Rana temporaria</i>	AY326063	—	—	—
<i>Rhacophorus bipunctatus</i>	AY843750	—	—	—
<i>Rhaebo glaberrimus</i>	DQ158454	DQ158376	DQ306510	—
<i>Rhaebo guttatus</i>	DQ158459	DQ158381	DQ306497	DQ283994

	12S + tRNAVal + 16S	RAG-1	CXCR4	Rhodopsin Exon 1
<i>Rhaebo haematiticus</i>	DQ158461	DQ158383	DQ306501	DQ283861
<i>Rhaebo nasicus</i>	DQ158477	DQ158396	DQ306512	—
<i>Rhinella amboensis</i>	DQ283386	—	—	DQ284003
<i>Rhinella arenarum</i>	AY843573	AY844370	—	AY844547
<i>Rhinella arenarum</i>	DQ158429	DQ158354	DQ306529	AY844547
<i>Rhinella arequipensis</i>	DQ158430	DQ158355	DQ306564	—
<i>Rhinella arunco</i>	DQ158442	DQ158365	DQ306552	DQ283857
<i>Rhinella atacamensis</i>	DQ158433	DQ158357	DQ306541	—
<i>Rhinella castaneoticus</i>	DQ158440	DQ158364	DQ306539	—
<i>Rhinella cf. dapsilis</i>	DQ158448	DQ158370	DQ306532	—
<i>Rhinella cf. margaritifer</i>	DQ158471	DQ158390	DQ306554	—
<i>Rhinella cf. margaritifer</i>	DQ158491	—	DQ306511	—
<i>Rhinella festae</i>	DQ158423	DQ158349	DQ306521	—
<i>Rhinella granulosa</i>	DQ158457	DQ158379	DQ306557	DQ283966
<i>Rhinella humboldti</i>	DQ158434	DQ158358	—	—
<i>Rhinella limensis</i>	DQ158466	—	DQ306509	—
<i>Rhinella marina</i>	DQ158473	DQ158392	DQ306530	DQ283789
<i>Rhinella nesiotis</i>	DQ158478	DQ158397	DQ306500	—
<i>Rhinella ocellatus</i>	DQ158479	DQ158398	DQ306538	—
<i>Rhinella poeppigii</i>	DQ158481	DQ158400	DQ306517	—
<i>Rhinella schneideri</i>	DQ158480	DQ158399	DQ306528	DQ283791
<i>Rhinella spinulosa</i>	DQ158487	DQ158405	DQ306566	DQ283775
<i>Rhinella vellardi</i>	DQ158495	DQ158411	DQ306527	—
<i>Rhinella veraguensis</i>	DQ158496	DQ158412	DQ306524	—
<i>Rhinoderma darwini</i>	DQ283324	AY364222	AY364192	DQ283963
<i>Scaphiophryne marmorata</i>	AY843751	AY364205	AY364175	—
<i>Schismaderma carens</i>	DQ158424	DQ158350	DQ306519	DQ284027
<i>Scythrophrys sawayae</i>	DQ283099	—	—	DQ283815
<i>Sphaenorhynchus lacteus</i>	AY549367	—	—	AY844754
<i>Telmatobius bolivianus</i>	AY843769	—	—	AY844757
<i>Telmatobius verrucosus</i>	DQ283040	—	—	DQ283770
<i>Thoropa miliaris</i>	DQ283331	—	—	—
<i>Trachycephalus venulosus</i>	AY549362	AY844493	—	AY844707
<i>Trichobatrachus robustus</i>	AY843773	AY364212	AY364182	AY844760
<i>Vitreorana eurygnatha</i>	AY843595	—	—	AY844567
<i>Werneria mertensi</i>	DQ283348	—	—	DQ283974
<i>Wolterstorffina parvipalmata</i>	DQ283346	—	—	DQ283972

^a *Frostius erythrophthalmus* MNRJ 32399, Brazil: Bahia: Uruçuca: Parque Estadual Serra do Canduru.

^b These sequences were reported as *Phrynopus* sp. (KU 202652) by Darst and Cannatella (2004). However, they blast as *Hypodactylus brunneus* (KU 178258) with 99% of coverage and 99% of similarity; for that reason we label them as *Hypodactylus* sp.

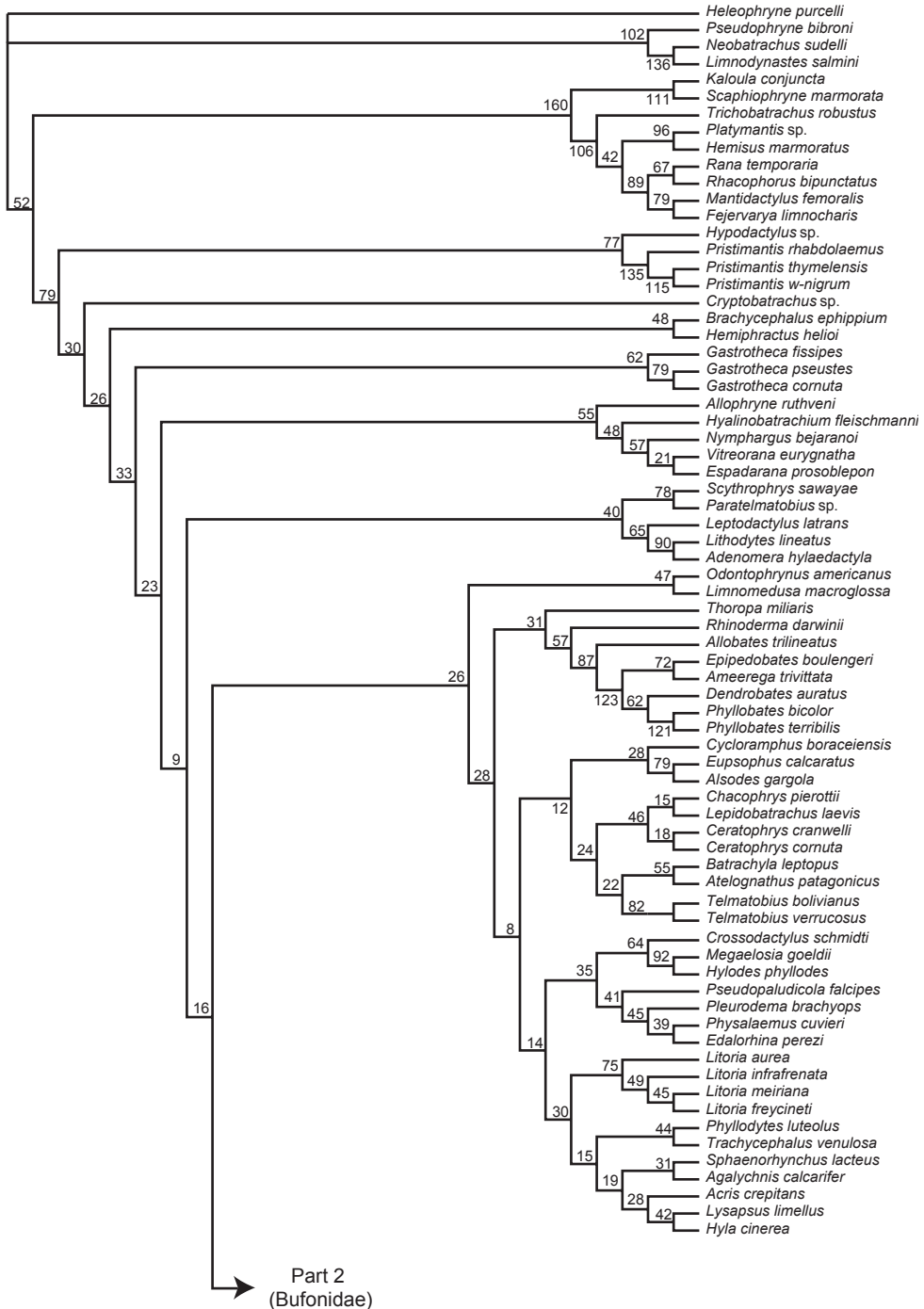
^c *Melanophryniscus devincenzii* BKT 0074, Uruguay: Tacuarembó: Punta del Laureles.

^d These sequences had been reported as *Melanophryniscus stelzneri* (KU 289071) by Pramuk (2006). The specimen is actually *M. fulvoguttatus* (Diego Baldo, personal commun.).

^e *Melanophryniscus setiba* CFBH 15748 (tissue collection number, CFBH-T 5088), Brazil: Espírito Santo: Guarapari: Parque Estadual Paulo César Vinha (20°36'25"S; 40°25'01"W).

APPENDIX 3: PHYLOGENETIC TREE

Strict consensus of six optimal trees of 33,389 equally weighted steps with Goodman-Bremer support values on each node. Part of the tree is shown in figure 1. The new species, *Melanophryniscus setiba*, is shown in **boldface**.





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