



Australian lizards are outstanding models for reproductive biology research

Authors: Van Dyke, James U., Thompson, Michael B., Burridge, Christopher P., Castelli, Meghan A., Clulow, Simon, et al.

Source: Australian Journal of Zoology, 68(4) : 168-199

Published By: CSIRO Publishing

URL: <https://doi.org/10.1071/ZO21017>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Australian lizards are outstanding models for reproductive biology research

James U. Van Dyke^{A,S}, Michael B. Thompson^B, Christopher P. Burridge^C,
Meghan A. Castelli^{D,E}, Simon Clulow^F, Duminda S. B. Dissanayake^{D,E}, Caroline M. Dong^G,
J. Sean Doody^H, Danielle L. Edwards^I, Tariq Ezaz^D, Christopher R. Friesen^J,
Michael G. Gardner^{K,L}, Arthur Georges^D, Megan Higgie^M, Peta L. Hill^C,
Clare E. Holleley^E, Daniel Hoops^{N,O}, Conrad J. Hoskin^M, Deirdre L. Merry^C,
Julia L. Riley^P, Erik Wapstra^C, Geoffrey M. White^C, Sarah L. Whiteley^{D,E},
Martin J. Whiting^Q, Stephen M. Zozaya^R and Camilla M. Whittington^B

^ADepartment of Pharmacy and Biomedical Sciences, School of Molecular Sciences, La Trobe University, Wodonga Campus, Wodonga, Vic. 3690, Australia.

^BSchool of Life and Environmental Sciences, The University of Sydney, Sydney, NSW 2006, Australia.

^CSchool of Natural Sciences, University of Tasmania, Hobart, Tas. 7001, Australia.

^DInstitute for Applied Ecology, University of Canberra, Canberra, ACT 2617, Australia.

^EAustralian National Wildlife Collection, CSIRO, Canberra, ACT 2601, Australia.

^FCentre for Conservation Ecology and Genomics, Institute for Applied Ecology, University of Canberra, Canberra, ACT 2617, Australia.

^GSchool of Science and Engineering, Tulane University, New Orleans, LA 70118, USA.

^HDepartment of Integrative Biology, University of South Florida, St Petersburg Campus, St Petersburg, FL 33701, USA.

^ISchool of Natural Sciences, University of California Merced, Merced, CA 95344, USA.

^JSchool of Earth, Atmospheric and Life Sciences, University of Wollongong, Wollongong, NSW 2522, Australia.

^KCollege of Science and Engineering, Flinders University, Adelaide, SA 5042, Australia.

^LEvolutionary Biology Unit, South Australian Museum, Adelaide, SA 5001, Australia.

^MCollege of Science and Engineering, James Cook University, Townsville, Qld 4811, Australia.

^NMouse Imaging Centre, The Hospital for Sick Children, Toronto, ON M5T 3H7, Canada.

^ODepartment of Medical Biophysics, University of Toronto, Toronto, ON M5G 1L7, Canada.

^PDepartment of Biology, Dalhousie University, Halifax, NS B3H 4R2, Canada.

^QDepartment of Biological Sciences, Macquarie University, Sydney, NSW 2109, Australia.

^RResearch School of Biology, Australian National University, Canberra, ACT 2601, Australia.

^SCorresponding author. Email: j.vandyke@latrobe.edu.au

Abstract. Australian lizards are a diverse group distributed across the continent and inhabiting a wide range of environments. Together, they exhibit a remarkable diversity of reproductive morphologies, physiologies, and behaviours that is broadly representative of vertebrates in general. Many reproductive traits exhibited by Australian lizards have evolved independently in multiple lizard lineages, including sociality, complex signalling and mating systems, viviparity, and temperature-dependent sex determination. Australian lizards are thus outstanding model organisms for testing hypotheses about how reproductive traits function and evolve, and they provide an important basis of comparison with other animals that exhibit similar traits. We review how research on Australian lizard reproduction has contributed to answering broader evolutionary and ecological questions that apply to animals in general. We focus on reproductive traits, processes, and strategies that are important areas of current research, including behaviours and signalling involved in courtship; mechanisms involved in mating, egg production, and sperm competition; nesting and gestation; sex determination; and finally, birth in viviparous species. We use our review to identify important questions that emerge from an understanding of this body of research when considered holistically. Finally, we identify additional research questions within each topic that Australian lizards are well suited for reproductive biologists to address.

Keywords: social behaviour, mating behaviour, developmental biology, sexual conflict, chemical communication, squamate, reptile, ecology, evolution.

Received 14 May 2021, accepted 17 August 2021, published online 28 September 2021

Introduction

Australia is home to ~13% of the world's lizard biodiversity (~850 of ~7000 species: Uetz 2010), despite containing only 5% of the world's land area. Australian lizards are taxonomically distributed across seven families: Carphodactylidae, Diplodactylidae, Gekkonidae, Pygopodidae (all from the infraorder Gekkota), Scincidae, Agamidae, and Varanidae (Cogger 2018). Of these, the Scincidae are the most diverse Australian lineage, with ~460 species (Wilson and Swan 2021). The Carphodactylidae are endemic to Australia, and Australia is home to large proportions of the world's Diplodactylidae (~65%), Pygopodidae (~97%), Scincidae (~27%), and Varanidae (~50%). These species numbers and proportions are not exact, however, because the taxonomy of Australian lizards is still developing (Melville *et al.* 2021). Together, Australian lizards are distributed across the entire continent, and experience environments ranging from cool temperate forests and alpine meadows to hot deserts and tropical rainforests (Fig. 1) (Cogger 2018). Most of the lizard species in

Australia exist in only a few of these habitats, but some, like the pygopodid *Lialis burtonis* and the scincid *Tiliqua scincoides*, thrive across a range of habitat types. Australia's diverse environments, and the abilities of different lizard taxa to become specialists or generalists to survive within them, are likely major reasons for Australia's high lizard biodiversity (Pianka 1969; Powney *et al.* 2010; Skeels *et al.* 2020).

Australian lizard biodiversity, and its distribution across such variable environments, provides important opportunities for testing hypotheses about how traits evolve and function. In particular, Australian lizards exhibit multiple evolutionary origins of key innovations related to reproduction, including social behaviours (Gardner *et al.* 2016), signalling and reproductive tactics (Dong *et al.* 2021a; Stuart-Fox *et al.* 2021), viviparity (live birth: Blackburn 2015), and transitions in sex determination (Sarre *et al.* 2004). The multiple origins of these traits, processes, and strategies provide natural replication for robust tests of key evolutionary hypotheses (Garland and Adolph 1994): do the physiological and genetic

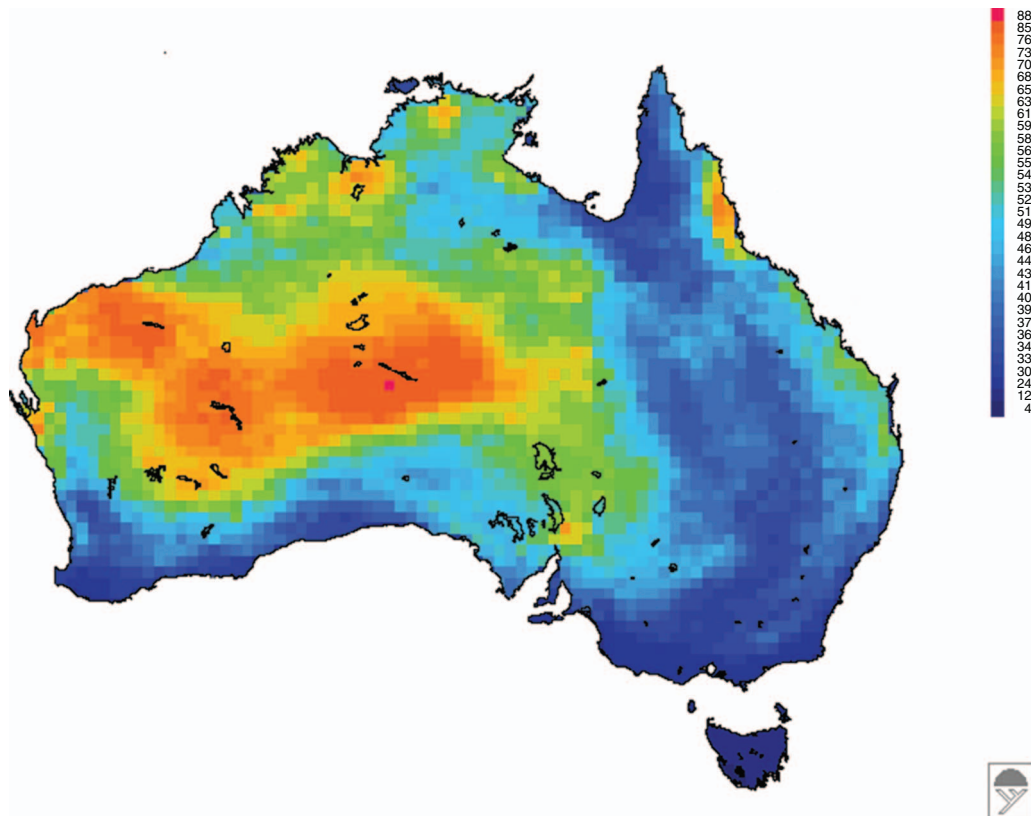


Fig. 1. Species richness map for Australian lizards. Colour coding represents the number of species in a 50 km × 50 km pixel, with cooler colours indicating fewer species (down to four) and warmer colours indicating more species (up to 88). (Reproduced from Cogger (2018), with permission from CSIRO Publishing.)



functions of these features evolve in the same ways? How do their functions change in species that are distributed across variable environments (Fig. 1)? The wide, overlapping distributions of some Australian lizard taxa (Cogger 2018) exhibiting variation in these traits provides a framework for answering these questions across species, and sometimes even within single species. Furthermore, many of these traits are shared with other animals, so the results gained from lizard research provide a basis for comparison with those in other taxa, which will help explain how these features evolve (Tinkle *et al.* 1970).

In addition to their utility for answering scientific questions, lizards are tractable research models. Many species are an ideal size to study using both captive and field-based approaches. They are big enough to carry dataloggers, yet small enough to house in captivity with relative ease and low costs (McDiarmid *et al.* 2012). Several Australian skinks and dragons exhibit high population densities, which enable the sample sizes required for robust ecological analyses. For these reasons, lizards around the world have been important model organisms for research in ecology, physiology, and life history evolution since the 1950s (Milstead 1967; Huey *et al.* 1983; Vitt and Pianka 1994). Early research on Australian lizards, in particular, contributed advances in understanding how placentas and viviparity evolve (Weekes 1935), and in community ecology (Pianka 1969, 1989).

Our objective with this review is to highlight the reproductive traits exhibited by Australian lizards that are of particular interest to biologists and students focussed on broader evolutionary and ecological questions that apply to animals more generally. Many Australian lizard experts contributed their expertise to highlight some important areas where Australian lizards have contributed, or could contribute, to answering broader questions in reproductive biology. We structured our review by covering topics in the sequence in which they occur during a reproductive cycle: behaviours and signalling involved in courtship; mechanisms involved in mating, egg production, and sperm competition; nesting and gestation; sex determination; and, finally, birth in viviparous species.

Pair bonds and sociality

Social behaviour can be as simple as an interaction between two individuals that briefly meet, or complex social interactions among multiple individuals that form large stable groups that interact over extended periods. Such social groups emerge as the result of either family members remaining together (e.g. fraternal societies) or through non-random associations of unrelated individuals (e.g. egalitarian societies). Both cases can result in individuals becoming mutually dependent on one another, sometimes forgoing their own reproduction and instead assisting with the reproduction of others. The emergence of complex social behaviour and societies has been highlighted as a major evolutionary transition (Maynard Smith and Szathmáry 1997). Thus, understanding the factors responsible for the evolutionary origins of such societies is a key challenge for biologists.

Traditionally, the flagship vertebrate taxa for understanding the evolution of complex sociality have been co-operatively breeding birds and mammals, organisms that live in complex societies (Rubenstein and Abbot 2017). Lizards, in contrast, have been largely ignored. However, a new social paradigm has emerged in lizards based on the recently recognised presence of stable, long-term, and sometimes complex social associations in multiple, independent taxa (Doody *et al.* 2013, 2021a; Gardner *et al.* 2016; Halliwell *et al.* 2017b). The best example of such complex social organisation comes from an Australian lineage of scincid lizards, the Egerniinae. The Egerniinae include 62 species in eight genera (*Egernia*, *Liopholis*, *Bellatorias*, *Lissolepis*, *Tiliqua*, *Cyclodomorphus*, *Corucia*, *Tribolonotus*; Gardner *et al.* 2008; Uetz 2010). Many species in the Egerniinae are characterised by the presence of long-term social associations both between adults, and between adults and their offspring, which result in the formation of stable family groups (Gardner *et al.* 2016; Whiting and While 2017; While *et al.* 2019). Crucially, the presence of these stable social associations and the diversity of social organisation (solitary, pair bonds, facultative family living, obligate family living) make the Egerniinae an outstanding model system for understanding the factors that have facilitated the origin and elaboration of complex social organisation.

The key foundation of social organisation in the Egerniinae is the presence of long-term pair bonds that form between males and females. Pair bonds were first identified in sleepy lizards (*Tiliqua rugosa*) (Bull 1988, 2000), where they are stable and may be life-long (Leu *et al.* 2015; Bull *et al.* 2017). Similar long-term pairings underpin social organisation across most social Egerniinae (Chapple 2003; Whiting and While 2017). The nature of these pair bonds can be variable across species. Although the predominant form of pair bonding within Egerniinae is monogamy, in some species, males form bonds with multiple females (Chapple and Keogh 2006; While *et al.* 2009b, 2011, 2019). In some species, such as *Egernia stokesii*, social bonding can extend to multiple adults of both sexes (Gardner *et al.* 2001, 2002; Duffield and Bull 2002), although often only a small number of individuals within these groups actually mate (Gardner *et al.* 2012).

Long-term data on pair bonding are relatively sparse for most Australian lizard species, except *T. rugosa*. However, pair stability over multiple breeding seasons has been identified for *Egernia cunninghami* (Stow and Sunnucks 2004), *E. saxatilis* (O'Connor and Shine 2003), *E. stokesii* (Duffield and Bull 2002; Gardner *et al.* 2002), and possibly *E. striolata* (Riley, unpubl. data) and *Liopholis whitii* (While, unpubl. data). While the occurrence of these long-term pair bonds is well established, we know relatively little about their functional significance. The reproductive benefits of maintaining a stable pair bond compared with switching partners, the short- and long-term trade-offs between maintaining single versus multiple social bonds, and the factors that initiate pair separation, remain largely unknown (but see Bull 2000; Leu *et al.* 2015). Addressing these questions has the potential to reveal the factors that promote and maintain the initial emergence of long-term pair bonds, which is relevant to other vertebrates in which family life is

based upon the maintenance of social monogamy (e.g. Young *et al.* 2019).

Stable social associations between males and females in the Egeriinae vary not only in their nature but in the extent to which they correspond to patterns of paternity acquisition. On average, members of the Egeriinae exhibit some of the lowest levels of female polyandry (i.e. mating with one or more males) in lizards (Uller and Olsson 2008), with genetic monogamy the most pervasive mating system. Despite the low levels of polyandry, most species exhibit at least some multiple mating and/or extra-pair mating. In social egeriine species, this ranges from 2.6% of litters including extra-pair paternity in *E. cunninghami* (Stow and Sunnucks 2004) to 10–30% for other species (e.g. Bull *et al.* 1998; Gardner *et al.* 2001; O'Connor and Shine 2003; Chapple and Keogh 2005; While *et al.* 2009b, 2014). In contrast, egeriine species that do not live in social groups exhibit higher levels of genetic polyandry. For example, 75% of *Tiliqua adelaidensis* litters show evidence of multiple mating (Schofield *et al.* 2014). Differences in the proportions of polyandry among species, coupled with variation in the extent of social complexity (see below), make the Egeriinae an excellent model for testing hypotheses about the role that genetic monogamy has played in the evolution of social organisation (e.g. Hughes *et al.* 2008; Cornwallis *et al.* 2010; Lukas and Clutton-Brock 2012).

Social aggregations in the Egeriinae are not just underpinned by prolonged associations between males and females, but also by prolonged associations between parents and their offspring. Indeed, offspring often delay dispersal and remain in their parents' refuge, sometimes for several years. The number and duration of these parent–offspring associations varies across the Egeriinae – from species in which these associations are absent (e.g. *T. rugosa*: Bull and Baghurst 1998), to species in which parents exhibit facultative associations with a single cohort of offspring (e.g. *L. whittii* (Chapple and Keogh 2006; While *et al.* 2009b), *Liopholis slateri* (Fenner *et al.* 2012), *Egernia saxatilis* (O'Connor and Shine 2003)), to species that live in large social groups with multiple cohorts of young. For example, *E. cunninghami* and *E. stokesii* can have up to five generations of offspring co-occurring in a single-family group (Gardner *et al.* 2001; Stow *et al.* 2001). Even in the mostly solitary *T. adelaidensis*, offspring can stay in natal burrows for several days before dispersing (Pearson *et al.* 2016). The associations between parents and offspring within the Egeriinae have several functional consequences that may constitute simple forms of parental care. For example, offspring gain increased access to basking sites, foraging opportunities, and retreat sites (Bull and Baghurst 1998; O'Connor and Shine 2004; Munch *et al.* 2018), which may result in an increase in early growth and survival (Botterill-James *et al.* 2016). Offspring also benefit from extended parent–offspring associations via a reduction in the risk of conspecific aggression and infanticide (O'Connor and Shine 2004; Sinn *et al.* 2008) and/or predation (Masters and Shine 2003; Watson *et al.* 2019). For example, the presence of a parent *E. saxatilis* eliminates aggression displayed towards offspring by unrelated adults (O'Connor and Shine 2004), and female *E. cunninghamii* actively chase off predatory snakes in

the presence of their offspring (Watson *et al.* 2019). Finally, offspring may inherit territories via prolonged parent–offspring associations, as suggested by the high levels of genetic relatedness within social groups of *E. stokesii* (Gardner *et al.* 2001) as well as the long-term residency of individuals in groups (Pearson *et al.* 2016).

Results from the Egeriinae clearly demonstrate the diversity in social traits that exist within the subfamily. Future research should experimentally test the factors that mediate the expression and nature of these simple forms of social behaviour, which have been predicted from observational studies (e.g. Halliwell *et al.* 2017a, 2017b) in a phylogenetic context. This combination will allow us to connect plasticity in social behaviour at the individual level with evolutionary divergence of social complexity at the population and species level. Such research has the potential to generate an understanding of how different social entities (males, females, offspring, siblings) initially come together, how such social associations are maintained in the face of inevitable conflicts that arise, and, ultimately, how the stabilisation of these associations provides a foundation for subsequent elaborations of more complex social behaviours. The growing appreciation that simple social behaviour, based around family life, has emerged independently in other lizard families (e.g. Cordylidae, Liolaemidae, Xantusiidae) (Fox and Shipman 2003; Davis *et al.* 2011; Gardner *et al.* 2016; Halliwell *et al.* 2017b) means that we can apply our understanding of social evolution in the Egeriinae to identify commonalities across transitions to social life in other social lizards. Their multiple origins of diverse social behaviours place the Egeriinae as an ideal model system for integrating a detailed understanding of social evolution with neurological, genomic, and developmental information. Combined, these origins will tell us whether social traits always evolve using the same mechanisms in different taxa. This approach has the potential to place the Egeriinae alongside other emerging socio-genomic/neuro-molecular vertebrate models, such as cichlids, frogs, butterfly fishes, birds, and mammals (e.g. O'Connell *et al.* 2012; Bukhari *et al.* 2019; Fischer *et al.* 2019; Nowicki *et al.* 2020) as a major future contributor to our understanding of what it takes to make an organism social.

Pheromones and reproduction in Australian squamates

Pheromones are chemicals that elicit a behavioural or physiological response in others of the same species (Karlson and Lüscher 1959; but see distinctions of Wyatt 2010), and are important to the social behaviour of squamates, including behaviour tied to reproduction. Pheromones influence behaviours such as locating, recognising, and choosing mates; antagonistic interactions that can determine the availability of mates; and parent–offspring recognition (reviewed by Houck 2009; Mason and Parker 2010; Martín and López 2011). Conceivably, pheromones could influence cryptic female choice after mating has already happened (e.g. choosing to use or reject sperm from certain males). Pheromones could also act as 'primers' that influence hormonal responses in the receiver (Bradbury and

Vehrencamp 2011), for example, by inducing or suppressing sexual receptivity in the receiver (Mendonça and Crews 2001). Many of the details of pheromones and reproductive behaviour remain to be tested in squamates.

In squamates, production of pheromones involved in reproductive signalling is controlled by reproductive hormones (Parker and Mason 2012; Parker and Mason 2014). Squamates produce pheromones from various regions of the body (Weldon *et al.* 2008). They are secreted from the skin (e.g. van Wyk and Mouton 1992; Mouton *et al.* 2010, 2014), including from specialised epidermal glands, such as generation glands and follicular pores (Cole 1966; Maderson 1972; Mayerl *et al.* 2015), or from specialised glands within the cloaca (Cooper and Grastka 1987; Cooper and Trauth 1992; Siegel *et al.* 2014). Pheromones identified from the scats of several Australian Egerniinae (Bull *et al.* 1999a, 1999b, 2000) are likely produced from glands in the cloaca and deposited onto the surface of the faeces, although this hypothesis remains untested. Variation in the presence of cloacal glands across squamates has not been widely assessed, but epidermal glands, which are visible to the naked eye and thus better known, vary widely in their presence among squamate lineages (García-Roa *et al.* 2017). Epidermal glands can vary between sexes, typically with glands being larger or present only in males (Mayerl *et al.* 2015). In some Australian lizards, such as *Amalosia* and *Nactus* geckos, the presence and absence of male epidermal pores vary even among closely related populations (Zozaya and Hoskin, pers. obs.). The *Ctenophorus maculatus* species complex exhibits variation in femoral pore size and number across environments (Edwards *et al.* 2015), suggesting that ecological factors may influence the evolution of epidermal glands in these species. Evolutionary trade-offs with other signalling modalities, such as visual displays, could also explain some variation in chemical signalling investment and the presence and extent of epidermal pores in other species (e.g. *Sceloporus*: Ossip-Klein *et al.* 2013; Campos *et al.* 2020) but has yet to be explored in Australian lizards. The drivers of variation in pheromone production, and its consequences for behaviour, are poorly understood areas ripe for further study, with *Amalosia* and *Nactus* geckos and agamids being potentially good models.

Squamates can detect chemicals in three ways: the nasal olfactory system, the vomeronasal (vomerofactory) system, and the gustatory (taste) system (reviewed in Schwenk 1995). Vomeroolfaction is generally regarded as the most important chemosensory system in squamates (Cooper 1994; Schwenk 1995), although nasal olfaction has been suggested as the primary chemosensory mode in geckos (Schwenk 1993). Beyond establishing the presence of taste buds (Schwenk 1985), gustation in squamates is essentially unstudied (Schwenk 1995; Mason and Parker 2010). Behaviours associated with chemosensory investigation are often used as proxies to measure and compare responses to chemical stimuli (Cooper 1998; Mason and Parker 2010). For example, the rate of tongue-flicking – an easily observed response tied to vomeroolfaction (Cooper 1994) – is the most commonly used proxy of chemosensory investigation in squamates (Cooper 1998; Mason and Parker 2010). Tongue-flicking and other proxies, however, vary in their suitability to address different

questions, and results are sometimes difficult to interpret (Cooper 1998). Future work should explore and test the link between proxies and behaviours of interest (e.g. tongue-flicking and mate preference), possibly including the development of new and better proxies. Identification of these links could be informed by more research on how chemicals are detected, how chemical signals are perceived (Romero-Diaz *et al.* 2021), and if and how detection and perception vary among groups.

Although most research on pheromones in squamates has focussed on European and New World species, several studies on Australasian taxa have focussed on pheromone-mediated discrimination (Table 1). Many of these studies relied on behavioural proxies and thus do not conclusively demonstrate a pheromonal role in reproduction, but they do suggest the potential for influencing mate choice and antagonistic behaviours that could lead to assortative mating. The link between pheromone variation and reproductive behaviours remains to be explicitly tested in most Australian taxa, except for the sea krait genus *Laticauda* (Shine *et al.* 2002b). Key to identifying, finding, and guarding potential or actual mates are behaviours such as assessing female receptivity (Head *et al.* 2005), and recognising and trailing mates (Bull *et al.* 1993a; Olsson and Shine 1998; Bull and Lindle 2002). Additionally, the capacity for reciprocal mother–offspring recognition (Head *et al.* 2008) is important to mediate postnatal conflict and care. Beyond these areas, more in-depth studies are needed to determine the precise ways that pheromones influence behaviour and reproduction.

Pheromonal research in Australia has focussed heavily on skinks, particularly those in the subfamily Egerniinae (Table 1), which exhibit complex sociality (Whiting and While 2017). However, pheromones are likely important to many squamate taxa. In fossorial taxa (e.g. *Anomalopus*, *Lerista*, *Aprasia*, blind snakes), pheromones are probably the most important, if not the only, signalling trait facilitating the finding and choosing of mates. Furthermore, recent work reveals divergence in the chemical blends of epidermal pore secretions among morphologically similar but genetically divergent gecko lineages, suggesting that pheromones may be important for mediating reproductive isolation in ‘cryptic species’ complexes (Zozaya *et al.* 2019). Continued research into the form and function of squamate pheromone systems is needed to better understand their influence on reproduction and their evolutionary consequences (e.g. speciation, evolution of sociality). Presently, the characterisation of chemical secretions has been done in two Australasian squamate genera: *Heteronotia* geckos (Zozaya *et al.* 2019) and *Laticauda* sea kraits (Shine *et al.* 2002b). Further work on chemical characterisation is needed in these and other groups to identify putative pheromone compounds for subsequent physiological and behavioural study. Exploring the potential links between pheromones and reproduction will require a better understanding of how compounds are detected and perceived, along with the design and execution of more biologically relevant behavioural and physiological assays. Australia’s extraordinary squamate diversity – with its corresponding diversity in morphology (for example, in epidermal pores), ecology, social systems, and behaviour, and the presence of

Table 1. Studies of Australasian squamates that tested for associations between pheromones and behaviours potentially linked to reproduction

Family	Species	Behaviour	References
Scincidae: Egerniinae	<i>Egernia stokesii</i>	Able to discriminate their own scent from the scent of other conspecifics, and group from non-group members, using both faeces and scent deposited on paper.	Bull <i>et al.</i> (2000)
	<i>Egernia striolata</i>	Able to discriminate their own scent from the scent of other conspecifics using both faeces and scent deposited on paper.	Bull <i>et al.</i> (1999a, 1999b)
	<i>Liopholis inornata</i> (as <i>Egernia inornata</i>)	Able to discriminate their own scent from the scent of other conspecifics using scent deposited on paper but not faeces.	Bull <i>et al.</i> (1999b)
	<i>Liopholis whitii</i>	Preference for scent of more closely related individuals.	Bordogna <i>et al.</i> (2016)
	<i>Tiliqua adelaidensis</i> <i>Tiliqua rugosa</i>	Males follow scent trails laid by females. Both sexes use pheromones to locate and trail pair-bonded mates, and mother–offspring recognition.	Ebrahimi <i>et al.</i> (2014) Bull <i>et al.</i> (1993a); Bull and Lindle (2002); Main and Bull (1996)
Scincidae: Eugongylinae	<i>Lampropholis similis</i> (as <i>L. coggeri</i>)	Males show greater interest in female versus male scents; females show greater interest in males or females from her own population.	Scott <i>et al.</i> (2015)
	<i>Carinascincus metallicus</i> (formerly <i>Niveoscincus metallicus</i>)	Males prefer to follow the scent of ‘partner’ female versus another female.	Olsson and Shine (1998)
Scincidae: Sphenomorphinae	<i>Eulamprus heatwolei</i>	Males use pheromones to assess female sexual receptivity.	Head <i>et al.</i> (2005)
		Pheromones facilitate reciprocal mother–offspring recognition.	Head <i>et al.</i> (2008)
Agamidae	<i>Ctenophorus pictus</i>	No female preference for epidermal pore secretions of more distantly related males.	Jansson <i>et al.</i> (2005)
Gekkonidae	<i>Heteronotia binoei</i>	Preliminary evidence that females discriminate among male epidermal pore secretions from different genetic lineages.	Zozaya, Hoskin, and Higgie, unpubl. data
Elapidae	<i>Cryptophis nigrescens</i>	Males prefer scent of larger females; females prefer scent of larger males.	Scott <i>et al.</i> (2013)
	<i>Emydocephalus annulatus</i>	No evidence that males use pheromones to locate females.	Shine (2005)
	<i>Laticauda colubrina</i> and <i>L. frontalis</i>	Pheromones (skin lipids) influence species discrimination and premating reproductive isolation between sympatric species.	Shine <i>et al.</i> (2002b)

several clades with worldwide distributions – present multiple potential model systems for pursuing lizard pheromone research.

Colour polymorphisms and alternative reproduction tactics in *Ctenophorus* spp

Intraspecific variation in male colour and reproductive traits

Colour polymorphism is the coexistence of two or more distinct, heritable morphs within an interbreeding population, with the rarest occurring too frequently to be solely the result of recurrent mutation (Huxley 1955). Colour polymorphic species are valuable for examining the relationships between traits under both natural and sexual selection because the colour morphs are a convenient visual indicator of specific reproductive behaviours and often carry physiological costs (Stuart-Fox *et al.* 2021). Trade-offs between body size, colouration, aggressiveness, and postcopulatory sexually

selected traits often occur in polymorphic species in which males compete for reproductive success using alternative mating tactics. For example, one morph can have higher fertilisation success via monopolising females through territoriality, another morph via relying on subterfuge mating tactics, and another by relying on highly competitive sperm (Sinervo and Lively 1996). Australia has two lizard species that have been used as model systems for such research: *Ctenophorus pictus* (painted dragon) and *C. modestus* (swift dragon). Male *C. pictus* are polymorphic in two aspects: head colour (red, orange, yellow, or blue: Olsson *et al.* 2007b), and the presence or absence of a yellow gular bib (Olsson *et al.* 2009a). Similarly, male *C. modestus* are polymorphic for throat colour (orange, yellow, yellow with an orange centre, and grey: Teasdale *et al.* 2013). Colour traits in both species are associated with different behavioural, physiological, and life-history traits. Both species have been used to study different aspects of alternative reproductive tactics and sexual

selection, with *C. pictus* as a model system for demonstrating the physiological costs associated with each morph, and *C. modestus* as a model for behaviour and aspects of coloration (e.g. visual conspicuousness, genetic and biochemical basis).

Colour traits in *C. pictus* are associated with trade-offs between investments in male aggressiveness, metabolism, condition loss, postcopulatory success, and telomere maintenance (a biomarker of stress and ageing) (Olsson *et al.* 2018a, 2018b), which are mediated by oxidative stress (Healey *et al.* 2007; Olsson *et al.* 2007a, 2009a; Healey and Olsson 2009; McDiarmid *et al.* 2017; Rollings *et al.* 2017; Friesen *et al.* 2017a, 2017b, 2020b, 2021). Bibbed and red-headed males invest more in traits that aid in precopulatory sexual selection, whereas yellow and non-bibbed males invest more in traits that aid in postcopulatory sexual selection, including sperm competition (Friesen *et al.* 2020a) (see the section *Multiple paternity, sperm competition, and postcopulatory sexual selection*). Red-headed and/or bibbed males tend to be more aggressive, and more likely to win male–male contests (Healey *et al.* 2007; McDiarmid *et al.* 2017). Bibbed males are also more attractive to females (McDiarmid *et al.* 2017). In contrast, yellow and non-bibbed males share similar reproductive tactics that include having larger testes (Olsson *et al.* 2009b) that produce more and faster sperm (Friesen *et al.* 2020c), but with significantly shorter copulation durations than either red or bibbed males (Olsson *et al.* 2009b; Friesen *et al.* 2020c). Although overall rates of multiple paternity are relatively low (~15% in the wild), yellow males have three times greater paternity success in head-to-head sperm competition trials over red males (Olsson *et al.* 2009b).

Bibbed male *C. pictus* do not lose paternity to neighbouring males in the field, probably because of effective territorial defence and mate guarding (Healey and Olsson 2009; Olsson *et al.* 2009a). Their investment in precopulatory traits carries a cost: both red-headed and bibbed males have significantly shorter blood cell telomeres (a potential marker of ageing and stress) than yellow-headed males and non-bibbed males (Olsson *et al.* 2007a; Friesen *et al.* 2017a, 2019). In contrast, their sperm telomeres are longer (Friesen *et al.* 2020c). Sperm telomere length is negatively related to sperm swimming velocity (Friesen *et al.* 2020c). The potential for sperm telomere length to influence sperm competitiveness or biases in female sperm storage are exciting ideas that remain to be tested (Olsson *et al.* 2018a, 2018b; Friesen *et al.* 2020c), as they may have implications for sperm competition across animals.

The male morphs of *C. modestus* exhibit different reproductive behavioural strategies, with orange as the most aggressive and grey as the least aggressive. The aggression level of yellow and orange-yellow morphs is conditional on the competitor's morph, where they are more aggressive to yellow and orange than to grey. The grey morph is the least bold, with similar boldness across the remaining morphs (Yewers *et al.* 2016). Differences in morph behavioural strategy correspond to differences in circulating baseline concentrations of androgen hormones. The aggressive orange morph has higher concentrations of androgens than either grey or yellow morphs (Yewers *et al.* 2017).

Although both *C. modestus* and *C. pictus* exhibit colour polymorphisms, the two species differ in their geographic distributions of morphs. All populations of *C. modestus* contain the same four morph types, but morph frequencies vary greatly among populations. However, variation in morph frequencies is not related to genetic differences or geographic distance among populations, but is strongly related to aridity index and vegetation cover (McLean *et al.* 2015). In contrast, not all populations of *C. pictus* have the same morph types. It is unclear if the apparent costs and benefits of alternative strategies influence fluctuations in morph frequency across years. From research on *Uta stansburiana* in the USA, fluctuations of morphs may be driven by female preference (Alonzo and Sinervo 2001) from year to year, but they also vary across geographic space from monomorphic to trimorphic (Corl *et al.* 2010b). Explanations for this variation centre on sexual selection as the cause of variance in morph number, given a positive relationship between sexual size dimorphism and morph number in *U. stansburiana* (Corl *et al.* 2010a). However, this relationship may not be indicative of variation in sexual selection (Chelini *et al.* 2021). Instead, both sexual size dimorphism and morph frequency may be influenced by common environmental factors in tandem, but independent of any relationship between them (Chelini *et al.* 2021). Environmental variables similarly influence the proportion of morphs in *C. modestus* (McLean *et al.* 2015), but this relationship has not yet been explored in *C. pictus*. By what mechanisms the environment regulates morph frequencies in polymorphic species is yet to be determined. It is possible that alternate numbers of morphs represent alternate resolutions of sexual conflict over mating dictated by environmental conditions (Svensson *et al.* 2020).

Intraspecific variation in female reproductive traits

In addition to variation in male colour morphs, lizard species also exhibit variation in reproductive output, including egg size, egg number, and reproductive frequency, which collectively determine total per-year fecundity. *Ctenophorus modestus* and *C. decresii* (tawny dragon) are sister species within the *C. decresii* species complex (McLean *et al.* 2014; Dong *et al.* 2021b) that differ in female reproductive strategy. *Ctenophorus modestus* females exhibit higher fecundity over a breeding season and produce larger clutches of smaller eggs, and more clutches per season (Dong *et al.* 2021b). In contrast, *C. decresii* females produce smaller clutches of larger eggs, and seldom more than one clutch per season. *Ctenophorus modestus* clutches also have longer incubation periods than do those of *C. decresii*.

The differing reproductive strategies of the two species may be attributable to environment-mediated selection. Both species are endemic to South Australia: *Ctenophorus modestus* occurs in the Flinders and Olary Ranges, whereas *C. decresii* occurs in the Mount Lofty Ranges, on the Fleurieu Peninsula, and on Kangaroo Island. The two species were long considered 'lineages' of *C. decresii sensu lato*, but were recently elevated to separate species on the basis of morphological and genetic differentiation (Dong *et al.* 2021b). Divergence between the two lineages may have resulted from isolation in separate

refugia during glacial–interglacial Pleistocene cycles (Byrne 2008; McLean *et al.* 2014). The evolution of differences in reproductive characteristics between the two species presents an opportunity to investigate the potential influence of environment-mediated selection on reproductive and other life history traits.

The differences in reproduction between *C. decrezii* and *C. modestus* may also be a result of climatic differences. The range of *C. modestus* extends across lower latitudes (-30.947° to -34.366°) with relatively warmer, semiarid conditions whereas climatic conditions of the higher latitudinal ranges of *C. decrezii* are relatively colder and temperate (-34.701° to -35.965°). Longer active seasons may allow for a higher clutch frequency in *C. modestus*, and, conversely, shorter active seasons constrain the breeding season of *C. decrezii*. A shorter active season also restricts the time for embryonic development and selects for a higher investment in the first clutch. This difference in reproductive output aligns with predictions that cold environments favour the production of fewer and larger eggs to increase offspring survival (Yampolsky and Scheiner 1996; Fischer *et al.* 2003; Morrison and Hero 2003). However, patterns of reproductive traits in lizards are not always globally congruent (Sun *et al.* 2013; Meiri *et al.* 2020). Counter-gradient variation in patterns of reproductive biology may be driven by many additional abiotic and biotic factors beyond latitude and climate (e.g. resource availability, predation risk, microhabitat preferences: Angilletta 2009; Mesquita *et al.* 2016). Furthermore, lizards may compensate for environmental factors in ways such as adjusting seasonal timing of reproduction (Warner and Shine 2007) or developmental mechanisms such as embryonic development rates (Oufiero and Angilletta 2006; Niewiarowski and Angilletta 2008), physiological pathways (Sun *et al.* 2013), and switching between semelparity and iteroparity (Eckhardt *et al.* 2017). Australia's diverse lizard fauna and geography present an ideal model system for testing hypotheses about how lizard reproductive traits evolve to cope with the constraints caused by different environments for several reasons. First, many of the ~850 lizard species overlap at least partially in their distributions (Wilson and Swan 2021), which presents an opportunity for replicated comparisons of environmental effects on reproduction. Second, some lizard species (and species complexes), such as *Lialis burtoni*, *Varanus gouldi*, and *Tiliqua scincoides*, have very large distributions across a range of different environments (Cogger 2018; Wilson and Swan 2021), which presents an excellent opportunity to study how reproductive variation is associated with specific environmental differences. The examples of geographic differences in reproductive output we present for *Ctenophorus* species are likely to also occur in other species with large distributions across varied environments.

Sexual selection on the neural control of reproduction

One of the few areas in neuroscience where lizards have made significant contributions is the neural control of reproductive behaviour (Lovern *et al.* 2004; Wade 2011). Across all vertebrates, two of the key brain regions controlling reproductive behaviour are the medial preoptic nucleus and the

ventromedial hypothalamic nucleus, both located in an area called the diencephalon. The medial preoptic nucleus drives male sexual behaviour and the ventromedial hypothalamic nucleus drives female behaviour (Numan 2014). Environmental influences, such as time of year and presence of conspecifics, affect reproductive behaviour in part by driving temporary changes to the size and the activity of these brain regions (Wade *et al.* 1993; Beck *et al.* 2008). In Australian agamid lizards, selection can drive more permanent changes in these brain regions over evolutionary time (Hoops *et al.* 2017).

Agamids are a particularly attractive lineage in which to study evolutionary changes in brain structure. They have the lowest known coefficient of brain allometry among lizards, and the highest known encephalisation quotient (Black 1983). Brain allometry is a measure of how closely changes in body size are paralleled by changes in brain size, and the encephalisation quotient estimates how 'enlarged' a brain is relative to a standard brain for a given body size (Platel 1979). The net result is that agamids have unusually large brains (for lizards) and relatively little variation in brain volume based on body size. These two factors are advantageous to researchers for practical reasons: larger brains are easier to extract, process, and measure accurately. Less variation with body size reduces overall variation in the data, which can make it easier to detect the influences of other variables on the brain.

Sexual selection drives both an increase in the volume of the medial preoptic nucleus and a decrease in the volume of the ventromedial hypothalamic nucleus in male *Ctenophorus* spp. dragons (Hoops *et al.* 2017). Thus, sexual selection possibly increases motivation and drive to perform male reproductive behaviours. However, brain region volumes do not differ across females in *Ctenophorus* species, suggesting that sexual selection does not drive changes in female reproductive behaviour. This suggests a passive role for females in reproduction, and is consistent with behavioural studies in some species of *Ctenophorus* (Lebas 2001; Olsson 2001; Jansson *et al.* 2005). In other lizard species, however, females can play very active roles in reproduction, including female-specific evolutionary changes in behaviour and motivation. For example, the female Lake Eyre dragon (*Ctenophorus maculosus*) displays unique reproductive colouration and behaviour that signals to males whether she is receptive to a male's advances (Olsson 1995). However, the brain structure of *C. maculosus* has not been compared with those of species with more passive reproductive behaviour. A behavioural innovation like that in female *C. maculosus* presents a unique opportunity to further understand how sexual selection can alter the brain.

Additional studies of brain anatomy across Australian lizards have great potential to develop our understanding of brain evolution. Approaches examining similar patterns of brain evolution with respect to sexual selection in other lizard species could reveal how generalisable the findings in the study of *Ctenophorus* spp. are to other taxa, and tease apart specific behaviours and ecological factors (such as territoriality or seasonality) that may influence how sexual selection shapes brain structure. There are additional topics, such as pair bonding (see *Pair bonds and sociality*) and parental care, where Australian lizards have the potential to

make important contributions to our understanding of the neural underpinnings of reproductive behaviour, and how these traits evolve. Australian lizards are potentially useful models for the study of brain anatomy and function because of their diversity of reproductive strategies. Our intimate understanding of this diversity can be leveraged towards understanding the underlying neural control, and what that means for the evolution of diverse reproductive strategies across vertebrates. The findings presented here represent the first step forward; the potential in this area is almost limitless.

Genital variation

Postcopulatory sexual selection is likely the main evolutionary force driving diversification in genital morphology in most animals (Simmons 2014; Brennan and Prum 2015). Understanding intrasexual and intersexual variation in genital morphology is critical for understanding mating-system dynamics, sexual conflict, and cryptic female choice. This knowledge is especially important given the diversity of mating systems within Australian lizards, each representing different opportunities for conflict and postcopulatory choice. Genital traits may also have an underappreciated role in understanding species diversification in lizards (Klaczko *et al.* 2015), as these traits are often involved in the development of reproductive isolation among putative species. Most of our understanding of genitalia in Australian lizards comes from work describing genital development in relation to sex determination (Whiteley *et al.* 2017). In squamates, males have hemipenes, whereas females may or may not have analogous structures called hemiclitoros (Böhme 1995; Martínez-Torres *et al.* 2015).

Research on hemipenal morphology is limited in Australian lizards. In a broadscale study across *Varanus*, including Australian species, hemipenal morphology is more phylogenetically informative than non-genital morphological characters; this pattern could reflect faster-evolving genitalia like that seen in *Anolis* species (Klaczko *et al.* 2015). Understanding such variation in Australian lizards may be important for understanding diversification dynamics in rapidly radiating Australian lizard lineages. Anecdotally, Australian agamids have larger hemipenes (relative to snout–vent length) compared with other ecologically similar lizard families on other continents (i.e. phrynosomatids: D. L. Edwards, pers. obs.). Greater hemipenal lengths are associated with greater copulation frequencies between species of *Anolis* lizards (Johnson *et al.* 2011). Differences in relative hemipenal size could therefore suggest fundamental differences in mating frequencies across lizard families. In *Liolaemus* species, hemipenal eversion is part of conspecific male–male aggressive displays (Ruiz-Monachesi *et al.* 2019). The potential for hemipenal morphology to play a similar role in Australian lizards remains to be investigated.

In contrast to males, very little is known about female genital evolution in squamates. Female genitals in squamates can be present as a rudimentary structure (Neaves *et al.* 2006), as hemiclitoros (Böhme 1995; Martínez-Torres *et al.* 2015), or as miniaturised hemipenes (Telemeco 2015). Even when present, they can also differ in colouration from conspecific

male hemipenes (Valdecantos and Lobo 2015). The functional role of female genitalia in squamates, and its variability, is unknown. *Varanus* spp., which are especially diverse across Australia, exhibit morphological diversity in hemiclitoros (Böhme 1995; Ziegler *et al.* 2005, 2007; Böhme and Ziegler 2009; Böhm *et al.* 2013), yet a comprehensive exploration of hemiclitoral morphology or even presence/absence in any Australian lizard genus, including *Varanus*, is currently lacking (but see King and Green 1999). Hemiclitoral morphology may relate to cryptic and postmating female control of reproduction. A prevailing paradigm is that precopulatory female choice is rare in lizards (Olsson and Madsen 1995; Olsson 2001; but see Sullivan and Kwiatkowski 2007). Variation in the presence/absence of cloacal glands, sensory innervation, epithelial wall thickness, and presence/absence of sperm crypts in lizards suggests that females have morphologically variable structures that enable postcopulatory choice (Sánchez-Martínez *et al.* 2007). Hemiclitori may have been retained for similar roles, such as expulsion of sperm. Studies should be undertaken using lizards with different mating systems and morphologies to determine the extent to which female lizards are able to control copulation duration and fertilisation in association with hemiclitoral and cloacal–vaginal morphology, which would determine mating biomechanics, sexual conflict over mating and the evolution of postcopulatory female choice (Friesen *et al.* 2014, 2016; Brennan 2016; Firman *et al.* 2017).

Work stemming from understanding genital development in relation to sex determination mode has provided some understanding of how lability in hemiclitoral structure develops. The central hypothesis is that genital variation occurs through differences in developmental programming among species. *Pogona vitticeps* exhibits temporary pseudohermaphroditism (TPH), whereby both ovaries and hemipenes are present at hatching (Whiteley *et al.* 2017, 2018). Females exhibit more variation in hemiclitoral phenotype than do males in hemipenal morphology (Whiteley *et al.* 2017). Thermostability of sex differentiation may play a role in an extended period of TPH, because other species with temperature-dependent or thermal influenced sex determination also show extended periods of TPH (Whiteley *et al.* 2018). While TPH complicates sex assignment based on hemipene eversion at birth in *P. vitticeps* (Whiteley *et al.* 2018), other Australian lizards (*Carinascincus* (formerly *Niveoscincus*) *ocellatus*) show clear genital differentiation at birth (Neaves *et al.* 2006). Resolving these differences among species is important for efforts to use genitalia to identify sex at birth. Given the diversity of sex determining mechanisms present within Australian lizards (section *Evolution of sex determination systems*), especially agamids, it is important to understand how genitalia develop in these different systems.

Multiple paternity, sperm competition, and postcopulatory sexual selection

Polyandrous mating systems are widespread throughout animal and plant taxa (Jennions and Petrie 2000; Pizzari and Wedell 2013; Taylor *et al.* 2014). Squamate reptiles are no exception (Uller and Olsson 2008; Friesen *et al.* 2020a), and

are excellent models to test critical questions about female promiscuity. Except for sperm, male lizards and snakes do not directly provide females with resources during courtship, mating, or after offspring are hatched/born, although females of territorial species may receive benefits from residing on a resource-rich site (Uller *et al.* 2010). Thus, the evolution of polyandry in lizards and snakes is simplified compared with vertebrates with extensive parental care (mammals and birds), so the fundamental costs and benefits of polyandry can be isolated (Uller and Olsson 2008; Kvarnemo and Simmons 2013). One benefit of polyandry is to ensure that a female has enough sperm to fertilise her ova. Sperm limitation may drive multiple mating in common lizards (*Zootoca vivipara*) (Uller and Olsson 2005). Multiple mating is associated with higher fecundity in lizards (Uller and Olsson 2005; LaDage *et al.* 2008; Noble *et al.* 2013; York and Baird 2019), but more studies specifically designed to tease apart the effects of sperm limitation and benefits of polyandry are needed. Social skinks and agamids with colour traits that may function in precopulatory sexual selection are ideal model systems, and both occur in Australia (section *Colour polymorphisms and alternative reproduction tactics in Ctenophorus spp.*).

A consequence of polyandry is that the battle for reproductive success does not always end after intrasexual competition for mates (usually males) or intersexual selection of mates (usually by females) – which together evolve as a result of precopulatory sexual selection (Darwin 1871; Andersson 1994). Instead, sexual selection continues within the reproductive tracts of promiscuous females, where the sperm of different males compete to fertilise eggs (Parker 1970; Parker and Pizzari 2010), and where female traits bias the contest for fertilisation success of different males (Thornhill 1983; Eberhard 1996; Arnqvist 2014). Together, these phenomena evolve as a result of postcopulatory sexual selection. Sperm competition is the postmating analogue of male–male competition for mates, which occurs when the ejaculates of more than one male overlap within the

reproductive tract of a polyandrous female (Parker 1998). Sperm competition between multiple males usually results in multiple paternity when there is no complete bias towards a single competitor. Multiple paternity is pervasive in squamates studied thus far, with more than 50% of clutches/litters exhibiting multiple paternity in the wild (Uller and Olsson 2008). Within lizards, rates of multiple paternity depend on how well males can monopolise access to females (Uller and Olsson 2008; Uller *et al.* 2010). Rates of multiple paternity are generally low (4–30%) in Australian territorial agamids (Table 2), but higher in non-social skinks (43–94%: Uller and Olsson 2008). The levels of polyandry in agamids (Table 2) contrast with those of ecologically similar families of lizards where multiple paternity is much higher (40–80%), for example, in the genera *Lacerta* in Europe (Fitze *et al.* 2005), and *Uta*, *Crotaphytus*, and *Sceloporus* in North America (Abell 1997; Zamudio and Sinervo 2000; Haenel *et al.* 2003; Peterson and Husak 2006).

Female sperm storage, where females store sperm for weeks to years after mating, increases the chance that sperm from different males will compete (Parker 1970). Sperm competition for fertilisation produces strong selection on sperm and ejaculate traits (Lüpold *et al.* 2020), which may impose energetic costs and concomitant trade-offs with precopulatory traits (body size, aggressiveness, colouration) as we find in some Australian lizards (see section *Colour polymorphisms and alternative reproduction tactics in Ctenophorus spp.*). Indeed, females of some Australian dragons store sperm across multiple clutches, creating the situation in which males may sire young posthumously (Olsson *et al.* 2009b). Female sperm storage could therefore shift male investment towards early-season mating success (Zamudio and Sinervo 2000). As a result, the evolution of female sperm storage, sperm longevity, and male lifespan are likely linked in some species. Females could select for sons that mature earlier, with longer-lived sperm, but die younger due to investments in early maturity. This pattern could be

Table 2. Multiple paternity in Australian lizards summed by species

Species	% Multiple paternity	Reference
Scincidae		
White's skink, <i>Liopholis whitii</i>	18.9% (23/122)	While <i>et al.</i> (2009b)
Cunningham's skink, <i>Egernia cunninghami</i>	2.6% (1/38)	Stow and Sunnucks (2004)
Spiny-tailed skink, <i>Egernia stokesii</i>	25.0% (4/16)	Gardner <i>et al.</i> (2000, 2002)
Tree Skink, <i>Egernia striolata</i>	13% (6/46)	Riley, unpubl. data
Southern water skink, <i>Eulamprus heatwolei</i>	56.4% (35/62)	Morrison <i>et al.</i> (2002); Stapley and Keogh (2005); Keogh <i>et al.</i> (2012)
Southern snow skink, <i>Carinascincus microlepidotus</i>	75.0% (6/8)	Olsson <i>et al.</i> (2005)
Spotted snow skink, <i>Carinascincus ocellatus</i>	93.8% (15/16)	Wapstra <i>et al.</i> , unpub. data (reported in Uller and Olsson 2008)
Southern grass skink, <i>Pseudemoia entrecasteauxii</i>	42.9% (12/28)	Stapley <i>et al.</i> (2003)
Pygmy blue-tongue, <i>Tiliqua adelaidensis</i>	75.0% (18/24)	Schofield <i>et al.</i> (2014)
Sleepy lizard, <i>Tiliqua rugosa</i>	19.0% (4/21)	Bull <i>et al.</i> (1998)
Agamidae		
Water dragon, <i>Intellagama lesueurii</i>	77.2% (17/22)	Frere <i>et al.</i> (2015)
Swift dragon, <i>Ctenophorus modestus</i>	2.7% (2/75)	Hacking <i>et al.</i> (2017); Rankin <i>et al.</i> (2016); Dong <i>et al.</i> , unpub. data
Tawny dragon, <i>Ctenophorus decresii</i>	0.0% (0/14)	Dong <i>et al.</i> , unpub. data
Jacky lizard, <i>Amphibolurus muricatus</i>	30.0% (20/67)	Warner <i>et al.</i> (2010)
Painted dragon, <i>Ctenophorus pictus</i>	15.0% (12/80)	Olsson <i>et al.</i> (2009a)
Ornate dragon, <i>Ctenophorus ornatus</i>	25.0% (5/20)	Lebas (2001)

common in short-lived annual lizards common in Australia, like *Ctenophorus pictus* and *C. fordi*, but has yet to be investigated.

Polyandry also has indirect genetic benefits in lizards and snakes (Wapstra and Olsson 2014), including positive fitness effects linked to offspring survival (Madsen *et al.* 1992; Eizaguirre *et al.* 2007), offspring dispersal (Laloi *et al.* 2009), improved heterozygosity for inbreeding avoidance, and ‘trading-up’ to a better mate (Olsson *et al.* 1996b; Laloi *et al.* 2011; While *et al.* 2014). Postcopulatory processes and conditions within the female reproductive tract may allow females to select sperm from males less-related to themselves (Olsson *et al.* 1996a, 1997, 2004), or, in species with XX/XY chromosomal sex determination, allow females to select sperm to determine the sexes of their offspring in a sex-biased environment (Olsson *et al.* 2007b; Cox and Calsbeek 2010). Nevertheless, the effects of multiple mating on offspring phenotypes are not always positive in lizards (Keogh *et al.* 2013; Noble *et al.* 2013), but these effects could also be cryptic in studies that do not analyse genetic interactions between the individuals participating in staged matings. In future work involving the assignment of parentage using molecular techniques, we encourage researchers to include enough markers to allow examination of genetic effects on the probability of paternity (e.g. relatedness) as well as male traits (genitalia, sperm, ejaculates) that may be used as cues by females to select paternity. Nearly nothing is known about female morphology, physiology, or behaviours that generate biases due to relatedness of their partners. Any polyandrous Australian species of lizard with a moderate length of sperm storage that demonstrates regular individual male–female interactions would be an ideal model for teasing apart the mechanisms of cryptic female choice. Dissociated mating and female sperm storage are relatively common in Australian skinks, including the genera *Carinascincus*, *Hemiergus*, and *Pseudemoia*, so these taxa might be useful models (Murphy *et al.* 2006).

Nesting ecology in Australian goannas

Most reptiles lack parental care and desert their eggs after depositing them in an excavated ground nest or burrow, or under objects such as rocks, logs, bark, or vegetation. Only a very small proportion of lizards attend their eggs (Shine 1988; Somma 2003), fuelling a perception that reproducing reptiles rarely engage in social behaviour after mating (but see *Pair bonds and sociality*). Moreover, excavating and backfilling shallow ground nests is conserved across many lizard families (Doody *et al.* 2009), with eggs predictably deposited in shallow nests in areas that are warm enough for successful development. In contrast, ‘deep-nesting’ Australian monitor lizards (‘goannas’) highlight that the ecology and evolution of nest site choice behaviour in lizards is anything but stereotypical or conserved.

Where the yellow-spotted monitor (*Varanus panoptes*) and its sister species, Gould’s monitor (*V. gouldii*), lay their eggs has only recently been identified. Aboriginal women suspected that eggs were laid in ‘warrens’, areas denuded of vegetation with multiple burrow entrances (Christian 2004), which was

confirmed by excavation beyond the terminus of open burrows. These warrens contain communal nests, solitary nests (with mothers returning to the same nest site year after year), nests with complex structures, and the deepest vertebrate nests in the world (up to 4 m deep: Doody *et al.* 2014, 2015, 2018a, 2018b). Moreover, the burrows provide refuges, foraging sites, aestivation sites and nesting sites for many other animals, which is the first demonstrated case of ecosystem engineering in lizards (Doody *et al.* 2021b).

Each warren contains multiple nests for the present (communal nesting) and previous years’ nesting (which may indicate traditional solitary nesting). For example, one portion of a *V. gouldii* warren contained 97 nests, including 21 nests with eggs (the rest were nests from previous years) (Doody *et al.* 2018a). Similarly, a *V. panoptes* warren contained 110 nests, including 11 with incubating eggs (Doody *et al.* 2018b). Extrapolation to the area of the entire *V. gouldii* warren predicted 53 nests with eggs, which implicates many mothers, despite the potential for multiple clutches per annum. When well fed in captivity, *V. panoptes* mothers can produce multiple clutches (D. Kirschner, unpubl. data), but this is less likely in nature, especially in a food-limited desert ecosystem (Doody *et al.* 2018b). Communal and traditional solitary nesting in these species may be related to the ease in constructing deep nesting burrows in soil that has been loosened by conspecifics within and among years, or some other benefit to mothers, eggs or hatchlings (Doody *et al.* 2009).

Both *V. panoptes* and *V. gouldii* nests are the deepest for any vertebrate: mean nest depths are 2.3–2.5 m (range = 1.0–3.6 m, $n = 162$) for *V. panoptes* and 3.0 m (range = 1.8–4.0 m, $n = 103$) for *V. gouldii* (Doody *et al.* 2014, 2015, 2018a, 2018b, 2020). Most ground-nesting reptiles deposit their eggs at depths of 20–250 mm, and even the gigantic leatherback sea turtle nests at depths averaging <1 m (Doody *et al.* 2014). These two goanna species probably nest deeply to maintain moist conditions during the long (~8 month) incubation period that spans the entire dry season, rather than depth being related to temperature (Doody *et al.* 2015). *Varanus panoptes* nests are deeper in a desert site than in a woodland site with higher rainfall (Doody *et al.* 2018b), and *V. panoptes* mothers nest deeper in drier years (Doody *et al.* 2021b). Decoupling of temperature from moisture in reptile nests (Doody *et al.* 2021b) demonstrates plasticity in depth due to a soil moisture gradient; in all other (shallower) reptile nests, shallower nesting means warmer but drier conditions, complicating the isolation of whether temperature or moisture is the cue for plasticity in nest depth.

Nesting is complex in both *V. panoptes* and *V. gouldii*, which construct elaborate helical burrows consisting of straight sections to a depth of ~1.5 m, followed by 2–7 tight spirals descending straight down and terminating in a slightly enlarged egg chamber. Fossil helical burrows date back to 255 Mya (*Diictodon*, a mammal-like therapsid: Smith 1987) and more recently from 22–34 Mya (*Palaeocastor*, a terrestrial beaver: Martin and Bennett 1977). Extant animal species with helical burrows include pocket gophers, prairie dogs, and scorpions (reviewed in Doody *et al.* 2015). The adaptive advantages of helical burrows are untested for any species, but

may include an antipredator function, improved drainage or hatchling escape, providing a more stable microclimate, an anticrowding mechanism, or mechanical advantage (reviewed in Doody *et al.* 2015). Interestingly, hatchling goannas appear not to use the spirals to emerge from the nest, but rather excavate a straight vertical tunnel, often through resistant soils (Doody *et al.* 2018a).

These varanids serve as ecosystem engineers for small animals, including 28 species of lizards (geckos, skinks, goannas), snakes, frogs, mammals, and numerous invertebrates. One warren contained 418 individual frogs (mostly *Uperoleia* spp.) that were aestivating there during the dry season (Doody *et al.* 2021a). The warrens also provided nesting habitat for another goanna species (*V. acanthurus*) and the gecko *Strophurus ciliaris* (Böhm *et al.* 2013; Doody *et al.* 2017, 2021b). These two goanna species thus join the burrowing tortoises of North America (*Gopherus*) and perhaps mound-nesting crocodilians as reptilian ecosystem engineers (Kinlaw and Grasmueck 2012; Somaweera *et al.* 2020).

Deep, complex communal nesting would not be expected to have evolved in desert or dry-adapted lizard species with shorter incubation periods, because the short incubation period would preclude the need for incubation over the entire dry season. The evolution of helical nesting remains enigmatic. Without knowing its value (adaptive or not), we cannot make predictions about its evolution, or lack thereof, in other lizard species. No other lizard species constructs such deep, helical burrows, but we predict that a third large, desert-inhabiting goanna with a long incubation period, the perentie (*V. giganteus*), may also nest at great depths due to its ecological similarity to its congeners. Deep helical nesting may also occur in populations of *V. griseus* near the Sahara Desert. As for why these goannas nest communally, the energy required to construct such deep complex burrows, and for the young to escape the burrows through resistant soils using their own tunnels, could select for mothers returning to communal nesting areas to take advantage of loosened soils. Returning to these sites would reduce the energetic costs of digging for both mothers and offspring, and thus allow both easier nesting, and easier escape from nests.

Viviparity and placentation

Australia is one of the best places in the world to study the evolution of viviparity and placentation, using reptiles as a model. Viviparity has evolved here repeatedly in both skinks and snakes (Blackburn 2015), and Australia has 20% of the reliably classified species that are bimodally reproductive: *Lerista bougainvillii* (Greer 1989; Qualls and Shine 1998), and *Saiphos equalis* (Smith and Shine 1997). In these species, individuals in some locations lay eggs, whereas those in other locations give birth to live young, providing a unique opportunity for comparative research on the evolution, costs, and mechanisms underpinning parity mode within a single species. Complex placentation and obligate placentotrophy have also evolved at least twice in Australian skinks, in the genera *Carinascincus* and *Pseudemoia* (Weekes 1935; Stewart and Thompson 2004). These represent 40% of the known origins of placentotrophy in lizards. A third skink species,

Eulamprus quoyii, also exhibits a modified chorioallantoic placenta (Murphy *et al.* 2011), though it is not highly placentotrophic (Thompson 1977; Thompson 1981). Together, viviparity and placentation represent some of the best opportunities for studying how novel complex traits evolve in vertebrates, because they have evolved repeatedly so many times (Griffith and Wagner 2017) and provide the phylogenetic replication necessary for comparisons to be statistically robust. In Australia, most research on viviparity and placentation has focussed on a relatively small group of skink species: Sphenomorphinae (*Eulamprus* spp., *Lerista bougainvillii*, *Saiphos equalis*), Eugongylineae (*Carinascincus* spp., *Pseudemoia* spp.), and Egerniinae (*Egernia* spp., *Liopholis* spp., *Tiliqua* spp.). The parity mode of additional species has been assigned based on observations of reproductive behaviour or dissection of gravid female museum specimens (Blackburn 2000), but many of these assignments require confirmation. It is unlikely that viviparity will be discovered in any lizard previously thought to be oviparous, but unexpected discoveries always have the potential to develop new avenues of research. For example, observation of both oviparity and viviparity *within an individual* (the first in a vertebrate) in *S. equalis* raises the possibility that reproductive mode may be plastic in some species (Laird *et al.* 2019).

All viviparous lizards studied so far have at least a simple placenta that provides respiratory gas exchange and a site of embryonic anchoring to the uterus (Blackburn 2015). Some species have more complex nutritive placentas, in which substantial quantities of organic nutrients are transported from the mother to embryos during pregnancy (obligate placentotrophy). Detecting this feature is more difficult than identifying viviparity alone. Placentotrophic species are identified by examination of the morphology of the placental tissues and comparisons of the dry masses of freshly ovulated eggs to those of newborn offspring (matrotrophy index: Thompson *et al.* 2000). Placentotrophic embryos increase in dry mass during development as mothers allocate nutrients to them, whereas non-placentotrophic embryos decrease in dry mass as they catabolise nutrients from their yolk (Stewart 1989; Swain and Jones 2000; Thompson *et al.* 2000). Placental transport of nutrients has also been demonstrated in Australian skinks using stable and radioisotopes (Thompson 1977; Swain and Jones 1997; Jones and Swain 2006; Itonaga *et al.* 2012b). Within Australian lizards, placental structure and function have been most well studied in the eugongyline skinks *Carinascincus* spp. and *Pseudemoia* spp. Even within these genera, the placental structures in *P. baudinii*, *P. rawlinsoni*, *C. palfreymani*, and *C. orocryptus* remain unstudied, and the degree of placental nutrient transport in *P. baudinii*, *P. rawlinsoni*, *C. greeni*, *C. palfreymani*, and *C. orocryptus* remains unknown. Furthermore, the amount of nutrients allocated across the placenta can vary within a species (Thompson and Speake 2006; Itonaga *et al.* 2012a; Van Dyke *et al.* 2014), so single estimates of matrotrophy index may not be representative of all individuals. The phylogenies for both *Carinascincus* and *Pseudemoia* remain poorly resolved (Hutchinson and Donnellan 1992; Brandley *et al.* 2015). Basic studies of placental structure and function in these taxa, and a

robust phylogeny, are needed to determine how the placenta has evolved in these species.

Research on Australian lizards underlies much of our understanding of how viviparity and placentation evolve and function in reptiles, and this work provides a basis for comparison with other viviparous taxa. Early in lizard pregnancies, a plasma membrane transformation (PMT) homologous to that of mammals occurs, which likely allows for embryos to anchor to the uterus (Murphy *et al.* 2000). This phenomenon was first recognised in Australian skinks. In *Pseudemoia* spp., the PMT is facilitated partially by changes in the distributions of desmosomes, tight junctions, occludins and cadherins in the uterine epithelium (Biazik *et al.* 2007, 2008, 2010; Wu *et al.* 2011). Once pregnancy begins, chorioallantoic and yolk sac placentas develop in all viviparous lizards, which facilitate contact between maternal and embryonic tissues (Thompson and Speake 2006). The functions of the two placentas likely vary across species, but the chorioallantoic placenta always has dense capillary beds that presumably facilitate placental exchange, especially for respiratory gases. In *Eulamprus tympanum*, angiogenesis in uterine and embryonic components of the chorioallantoic placenta increases the density and apposition of capillaries in both tissues (Parker *et al.* 2010a, 2010b). In *Saiphos equalis*, angiogenesis is driven in part by vascular endothelial growth factors (VEGFs; Murphy *et al.* 2010; Whittington *et al.* 2015). Carbonic anhydrases in both uterine and embryonic components of the chorioallantoic placenta likely facilitate transport of carbon dioxide in *Pseudemoia entrecasteauxii* (Van Dyke *et al.* 2015). Most viviparous lizards also transport small amounts of nutrients to their developing embryos (Thompson *et al.* 2000), especially inorganic ions like calcium (Linville *et al.* 2010), which may replace the role of the eggshell as an ion reserve in oviparous species (Herbert *et al.* 2006). In Australian lizards, evidence of obligate placentotrophy of organic nutrients is so far limited to skinks in the genera *Carinascincus* and *Pseudemoia*; candidate nutrient transporters in these taxa include lipoprotein lipase, and amino acid-transporting solute carriers (SLCs; Griffith *et al.* 2013, 2016). Gene expression analyses, particularly transcriptomic studies, have identified thousands of genes that are likely to be involved in pregnancy in Australian lizards (Griffith *et al.* 2016, 2017; Hendrawan *et al.* 2017; Foster *et al.* 2020). These genes likely contribute to key pregnancy-related functions including uterine remodelling, nutrient and respiratory gas transport, and immune regulation, and testing the functions of these genes is the next step in understanding their role in the evolution of viviparity.

Australian lizards also present an excellent opportunity to study the selective pressures that drive the evolution of viviparity and placentae, although these questions have received much less attention than the genetic, physiological, and morphological mechanisms that underpin both traits. Adaptive hypotheses for viviparity and placentation focus on the potential fitness benefits mothers gain by being able to continuously control the environment that developing embryos experience, and also on embryos' ability to impact maternal physiology to their own benefit (Tinkle and Gibbons 1977; Shine 1995; Crespi and Semeniuk 2004). Research integrating ecophysiological and molecular approaches is needed to test

how viviparity and placentotrophy impact fitness, and whether the fitness benefits of these traits overrides their concomitant fitness costs. For example, viviparity may allow mothers to improve the fitness of their individual offspring, but it also reduces the number of reproductive events a single female can have per year, and increases the physical burden on the mother (Tinkle and Gibbons 1977). Australian skinks are ideal models to test these adaptive hypotheses given their diversity of reproductive modes, especially in taxa that overlap in distribution and ecology, as do the genera *Pseudemoia*, *Carinascincus*, *Bassiana* (also known as *Acritoscincus*), and *Lampropholis*. The two reproductively bimodal species, *Lerista bougainvillii* and *Saiphos equalis*, also present an excellent opportunity to test how the fitness costs and benefits of reproductive mode vary with environment within single species. Simultaneously, they provide the opportunity to determine whether gene flow between viviparous and oviparous populations constrains the evolution of reproductive mode.

Sex reversal

Sex determination and differentiation are two developmental processes that govern the dichotomous phenotype, male or female. This variation is constrained to produce either male or female phenotypes. A sex-determining factor initiates the sexual differentiation cascade, and results in individuals that possess either ovaries (female) or testes (male), as well as sex-specific behaviours, morphologies, and physiologies. Lizard sex-determining factors could fall into diverse categories and we still have not identified a single sex-determining gene for lizards. In many lizard species, sex chromosomes determine female or male development, and these genetic sex determination (GSD) systems can be either female heterogametic (ZZ/ZW) or male heterogametic (XX/XY) (Bull 1980). Temperature-dependent sex determination (TSD) is also common among lizards (Harlow 2004), whereby sex is determined by the temperature at which the embryo develops during a sensitive period of incubation. Although some lizard lineages possess stable and conserved sex chromosomes, others exhibit rapid sex chromosome evolution, including transitions between male and female heterogamety and turnover in the chromosome pair recruited to determine sex (Sarre *et al.* 2011; Pennell *et al.* 2018). Sex chromosomes are predicted to play a central role in lizard evolution, adaptation, and speciation; even though the same reproductive phenotypes are achieved (male or female), episodic turnover in sex chromosomes and sex determining systems has broad evolutionary consequences (Ezaz *et al.* 2009; Sarre *et al.* 2011). Many transitions have occurred between TSD and GSD across the lizard phylogeny (Janzen and Krenz 2004) (see also section *Evolution of sex determination systems*).

Sex-determining systems exist as a continuum of genetic and environmental influence, with many species likely to occupy an intermediate position on the continuum from TSD to GSD (Sarre *et al.* 2004). Incubation temperatures are suspected of being capable of overriding genotypic sex-determining signals in ZZ/ZW and XX/XY systems in some

species (Shine *et al.* 2002a; Quinn *et al.* 2007; Holleley *et al.* 2015, 2016) in both oviparous and viviparous lizards (Shine *et al.* 2002a; Holleley *et al.* 2015, 2016; Cornejo-Páramo *et al.* 2020a; Wiggins *et al.* 2020; Dissanayake *et al.* 2021). Discordance between genotypic and phenotypic sex is known as sex reversal (Weber and Capel 2018). Naturally occurring

temperature-sex reversal has only been definitively identified in two vertebrates, both Australian lizards, which possess contrasting systems of GSD that simultaneously display temperature-dependent influences on sex. *Pogona vitticeps* is an agamid dragon lizard with a female heterogametic (ZZ/ZW) GSD system, but high incubation temperatures (>32°C) result

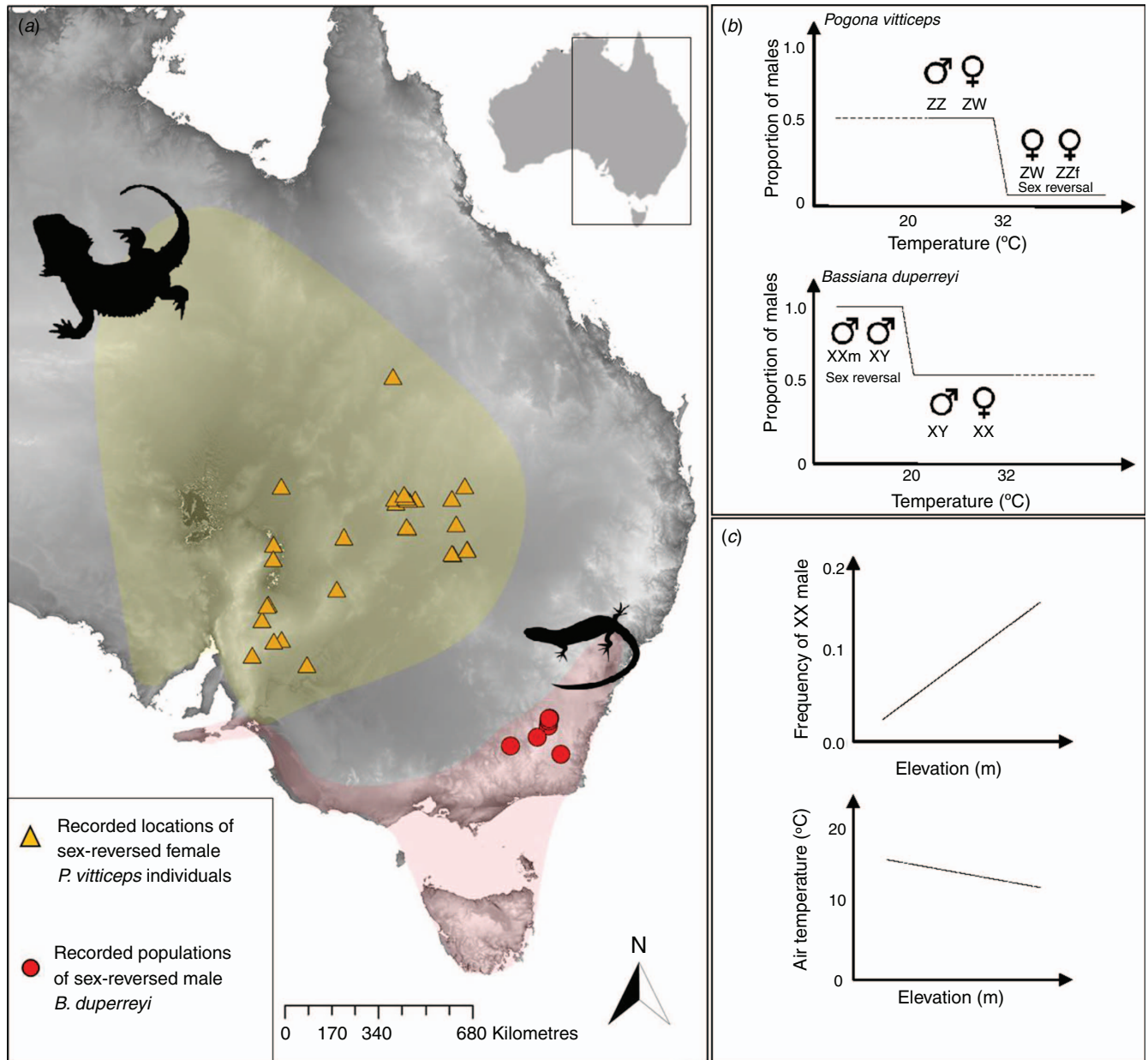


Fig. 2. Sex reversal in the Australian lizard species *Bassiana duperreyi* and *Pogona vitticeps*. (a) Distribution of sex reversal across the ranges of *P. vitticeps* (Castelli *et al.* 2021) and *B. duperreyi* (Dissanayake *et al.* 2021). The species range is indicated by the shaded areas. (b) Graphical representation of sex reversal characteristics (not to scale); top – *P. vitticeps*: sex reversal occurs when an individual with a male genotype (ZZ) is incubated at temperatures above 32°C, causing it to develop as a phenotypic female (ZZf) (Holleley *et al.* 2015); bottom – *B. duperreyi*: an individual with a female phenotype (XX) incubated at low temperatures will reverse its sex and develop as a phenotypic male (XXm) (Shine *et al.* 2002a; Radder *et al.* 2008). (c) Sex reversal frequency of *B. duperreyi* along an elevational gradient (Dissanayake *et al.* 2021). Underlying map generated using ArcGIS 10.5.1 (<http://www.esri.com>) and data from the Digital Elevation Model (Geoscience Australia) made available under Creative Commons Attribution 3.0 Australia (<https://creativecommons.org/licenses/by/3.0/au/legalcode>, last accessed 21 December 2020). The species range is indicated by the shaded areas (data from the Atlas of Living Australia website at <http://www.ala.org.au>; accessed 3 January 2021).

in reversal of the ZZ male genotype to a female phenotype (Quinn *et al.* 2007; Holleley *et al.* 2015). In contrast, *Bassiana duperreyi* has a male heterogametic (XX/XY) GSD system in which low incubation temperatures (<20°C) result in reversal of the XX genotype to a male phenotype (Shine *et al.* 2002a; Radder *et al.* 2008; Dissanayake *et al.* 2021).

The occurrence of sex reversal in wild populations of *P. vitticeps* and *B. duperreyi* provide unique opportunities to study the distribution and dynamics of sex reversal under different selective pressures in Australia (Fig. 2). Theoretical studies show that as the frequency of sex reversal increases in a population, and provided the reversed individuals are fertile, a likely response is the reduction and possible elimination of the W or Y chromosome under Fisher's frequency-dependent selection (Fisher 1930; Düsing 1884) and a transition to a pure TSD system (Bull 1981; Grossen *et al.* 2011; Holleley *et al.* 2015; Bókony *et al.* 2017; Geffroy and Wedekind 2020; Schwanz *et al.* 2020; Dissanayake *et al.* 2021). Alternatively, selection for the rarer sex may drive the evolution of higher temperature thresholds for sex reversal and the maintenance of GSD in this species (Schwanz *et al.* 2020). Across the natural range of *P. vitticeps*, sex reversal is spatially constrained, displaying no association with latitude or climatic variables, which is contrary to expectation based on laboratory observations (Castelli *et al.* 2021). In *B. duperreyi*, rates of sex reversal increase with elevation (from zero to 18.46% of XX individuals manifesting as phenotypic males: Dissanayake *et al.* 2021). This observation suggests that *B. duperreyi* has not been subject to selective pressures for rapid evolution in the threshold for sex reversal, and thus populations at high elevations may be susceptible to loss of the Y chromosome (Dissanayake *et al.* 2021). Evolution of the thermal threshold for sex reversal is possible in both species, and there exists some evolutionary capacity to buffer or moderate the effects of extreme climates (Castelli *et al.* 2021; Dissanayake *et al.* 2021).

Few species have been examined for instances of sex reversal in Australia or elsewhere (but see Wiggins *et al.* 2020; Whiteley *et al.* 2021a). The widespread occurrence of homomorphic sex chromosomes in lizards means that instances of sex reversal are challenging to detect. New bioinformatics tools and methods make it possible to identify and characterise sex chromosomes, leading to unanswered questions in sex determination mechanisms in lizards (Hill *et al.* 2018; Palmer *et al.* 2019; Cornejo-Páramo *et al.* 2020a; Dissanayake *et al.* 2020). Sex reversal, although not currently confirmed, may occur in several other Australian lizards: the southern water skink (*Eulamprus heatwolei*) (Cornejo-Páramo *et al.* 2020a), the spotted snow skink (*Carinascincus ocellatus*) (Cornejo-Páramo *et al.* 2020a) (see section *Evolution of sex determination systems*), the Jacky dragon (*Amphibolurus muricatus*) (Whiteley *et al.* 2021b), and several overseas species, including the common collared lizard (*Crotaphytus collaris*) (Wiggins *et al.* 2020), the multiocellated racerunner (*Eremias multiocellata*) (Wang *et al.* 2015), and the Japanese gecko (*Gekko japonicus*) (Tokunaga 1985).

So far, the molecular mechanisms of sex reversal in lizards are not fully understood (Georges and Holleley 2018; Castelli

et al. 2021; Whiteley *et al.* 2021c). The tantalising possibility that many traditionally regarded TSD species also have a cryptic underlying chromosomal component to sex determination is an important area of future research. Evolutionary modelling has shown that the Y or W chromosome can persist at low frequency despite high rates of sex reversal (Quinn *et al.* 2007; Schwanz *et al.* 2020). Beyond cryptic Y or W sex chromosomes, even more complex scenarios may exist that interact with thermal sex-determining cues, such as polygenic sex determination or *de novo* sex chromosomes (Mork *et al.* 2014). This is likely to be a very fruitful area of research using new genomic techniques. Finally, there are several evolutionary questions of interest concerning the transitions between GSD and TSD. For example, once the transition to TSD occurs, is there sufficient time for optimisation of the thresholds for sex determination (pivotal temperature) to avoid catastrophic sex ratio skew and demographic extinction under climate change? There will no doubt be many more surprises. The recent media report of sex reversal in an adult Boyd's forest dragon (*Lophosaurus boydii*) in response to loss of the sole male in its captive population (Mannix 2020) is one such example. If confirmed, this will provide exciting new avenues of research and the interaction of cytogenetic and genetic aspects of developmental programming in lizards.

Evolution of sex determination systems

Sex determination is so fundamental to reproduction that it is expected to be under strong purifying selection with highly conserved processes (Uller *et al.* 2007). Indeed, conserved GSD systems exist in therian mammals (male heterogamety; XX/XY) and birds (female heterogamety; ZZ/ZW) (Vicoso *et al.* 2013; Bachtrog *et al.* 2014). In contrast, reptilian sex determination mechanisms vary in a phylogenetically complex manner, which suggests multiple evolutionary transitions (Janzen and Phillips 2006; Pokorna and Kratochvil 2009). In lizards, there is GSD (XX/XY and ZZ/ZW are both common), TSD, and interactions between genes and temperature (GSD+EE or thermosensitive GSD) (Sarre *et al.* 2004). Sex chromosomes range from highly differentiated to morphologically indistinct pairs in lizards with GSD. Sex determination in lizards may be dosage dependent, where the homogametic sex (ZZ or XX) possesses two copies of a sex-determining gene and acquires a threshold for sexual phenotype, whereas the heterogametic sex (XY or ZW) possesses one copy and does not reach the threshold (Quinn *et al.* 2007). In addition, patterns of TSD are diverse: high temperatures can produce excess males ('FM' pattern), excess females ('MF' pattern) or an excess of females at both high and low temperatures with males produced at intermediate temperatures ('FMF' pattern) (Fig. 3). Finally, reaction norms to temperature can be steep, where the switch between male and female producing temperatures can occur over a small pivotal range (Fig. 3a–c), or shallow (Fig. 3d), where sex ratios vary gradually over a broad range of temperatures.

Work on Australian lizards has led to the important redefining of sex determination as existing as a continuum of states between pure GSD and pure TSD, rather than a

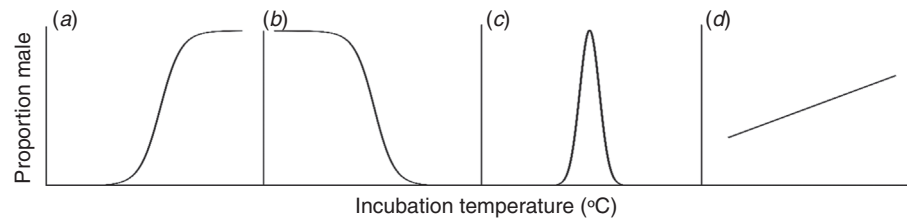


Fig. 3. Patterns of temperature-dependent sex determination (TSD). FM pattern (a, d) where males are produced at high temperatures; MF pattern (b) where females are produced at high temperatures; FMF pattern (c) where males are produced at an intermediate temperature and females are produced either side of this temperature. Reaction norms can be steep (a–c) or shallow (d).

dichotomous trait (Sarre *et al.* 2004). However, questions remain about how and why transitions between systems of sex determination occur. The drivers and mechanisms acting at the time of transitions are difficult to determine due to the divergence between lizards with different sex-determining systems. Australian lizards are central to understanding the evolutionary drivers of transitions, the mechanisms that underpin transitions and their consequences to species and populations. The adaptive benefit of GSD versus TSD has been the subject of extensive research in Australia (Shine *et al.* 1995; Shine 1999; Warner and Shine 2005, 2008; Pen *et al.* 2010; Wapstra and Warner 2010). TSD should be favoured when there are sex-specific advantages resulting from development at temperatures that produce an excess of the sex that benefits (Charnov and Bull 1977; but see Uller and Olsson 2006). GSD typically produces 50:50 sex ratios and is therefore favoured when seasonal temperature fluctuations are high and might otherwise produce sex ratio biases that are maladaptive for some individuals (Bulmer and Bull 1982; but see Dooren and Leimar 2003; Cornejo-Páramo *et al.* 2020b). Species with TSD may be particularly vulnerable to climate change because of the effect that highly-skewed sex ratios can have on population persistence if one sex becomes rare to the point of limiting mating frequency (Le Galliard *et al.* 2005; Boyle *et al.* 2014; Wedekind 2017; Valenzuela *et al.* 2019). While selection can favour biased sex ratios when sex-specific fitness benefits occur with incubation temperature (Charnov and Bull 1977), populations with sex ratios that are consistently biased towards one sex can experience deterioration of genetic diversity and therefore have reduced adaptive potential, with consequences for species distributions (Le Galliard *et al.* 2005; Mitchell *et al.* 2008; Mitchell and Janzen 2010). However, such population-wide skews are rare and often transient, because frequency-dependent selection favours females (or parents) producing the under-represented sex (Fisher 1958), which concomitantly can select for mechanisms to balance skews (Uller *et al.* 2007).

In reptiles, TSD was first discovered in *Agama agama* more than 50 years ago (Charnier 1966; Steele *et al.* 2018), followed by observations in turtles (Pieau 1972; Yntema 1976), the leopard gecko (Viets *et al.* 1993), and the tuatara (Cree *et al.* 1995), all of which are oviparous. Because viviparous females can behaviourally buffer the effects of temperature on offspring development, TSD was considered incompatible with viviparity (Bull 1980), but temperature effects on offspring sex have since been demonstrated in two viviparous

Australian lizards (Robert and Thompson 2001; Wapstra *et al.* 2004). These viviparous species differ from typical TSD species because they have distinctly shallow reaction norms best characterised by Fig. 3d. Australia has considerable lizard diversity, scientific expertise, and exceptional species amenable to advancing our understanding of evolutionary transitions in sex determination. One such species (*Carinascincus ocellatus*) is undergoing an incipient transition in sex determination, offering a rare opportunity to unravel the causes and consequences of such transitions.

An Australian model system for transitions in sex determination: Carinascincus ocellatus

The viviparous spotted snow skink, *C. ocellatus* (Fig. 4a), is a rare example of a viviparous species undergoing incipient divergence in sex determination (Wapstra *et al.* 2004; Pen *et al.* 2010; Cunningham *et al.* 2017). This small Tasmanian-endemic skink has a broad geographic and climatic distribution across Tasmania, with concomitant variation in life history (Wapstra *et al.* 1999, 2001; Wapstra and Swain 2001). In a warm, low elevation population, maternal basking opportunity (and therefore developmental temperature) influences the sex of her offspring; warm environmental conditions lead to an excess of female offspring in the population and cool conditions lead to an excess of males. In a high elevation population, sex ratios remain at parity regardless of environmental conditions and maternal basking opportunity (Pen *et al.* 2010; Cunningham *et al.* 2017) (Fig. 4b, c). The sex ratio response to temperature, initially observed in the laboratory in females from the low elevation population through altered basking opportunity (Wapstra *et al.* 2004), has since been demonstrated in the wild (Wapstra *et al.* 2009). There are potentially important consequences for this population-specific effect of environmental temperatures on offspring sex ratios (Fig. 5c) and for our understanding of the impact of climate change on species with TSD (especially where they follow a relatively rare shallow reaction norm) (Cunningham *et al.* 2020). Because warm temperatures produce a female-biased sex ratio in *C. ocellatus*, climate warming may result in population growth rather than decline as temperatures warm, as long as males do not become limiting (Rankin and Kokko 2007; Wapstra *et al.* 2009; Cunningham *et al.* 2017).

In *C. ocellatus*, divergence in sex determination could be driven by a combination of population-specific selection on

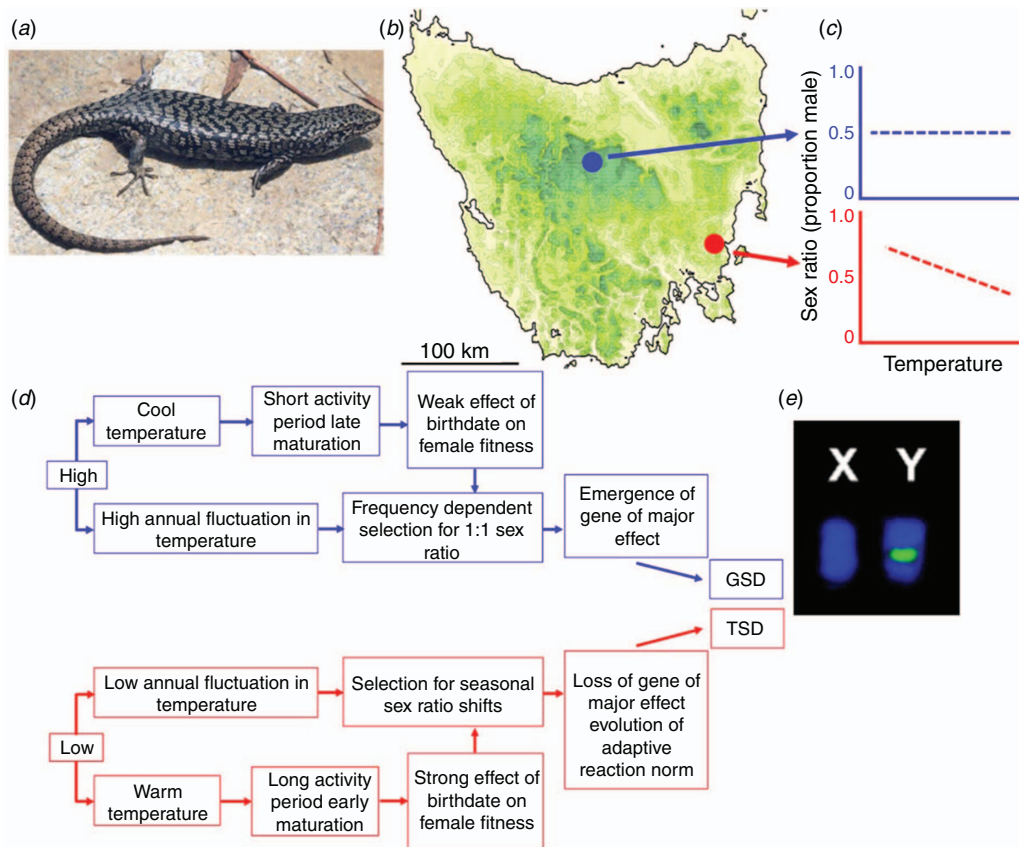


Fig. 4. *Carinascincus ocellatus* (a) high (1200 m above sea level (masl), GSD, blue) and low (50 masl, GSD+EE, red) elevation populations (b) with population specific sex-ratio responses to incubation temperature (c). A life history model parameterised with long-term field data predicts loss of sex determining genes at low elevation and emergence of sex determining genes at high elevation (adapted from Pen *et al.* 2010) (d). Sex chromosomes (e), labelled here with *C. ocellatus*-specific Y-linked probe (Hill *et al.* 2021a), are undifferentiated in both populations.

how mothers allocate the sexes of their offspring (resulting in thermosensitive sex determination at low elevation) along with selection against strongly biased sex ratios in variable, unpredictable environments (resulting in GSD at high elevation: Pen *et al.* 2010) (Fig. 4d). Specifically, in warm lowland populations, individual females that produce daughters under warmer conditions potentially have higher fitness because female offspring born early reach sexual maturity earlier and have a higher lifetime reproductive output than those born late. This situation selects for a link between gestational temperature, birth date, and offspring sex. Conversely, in the cold highlands selection does not occur because cooler conditions lead to slower growth with later sexual maturity, which creates a dissociation between temperature-dependent development, birth date, size at maturity, and sex-specific fitness benefits to mothers from sons or daughters. The high annual fluctuation in temperature in the cold highlands further selects against temperature effects in this model, because of the resultant frequency-dependent selection against the large skews in sex ratios (Fig. 4d).

The theoretical model (Fig. 4) established a potential adaptive explanation for the intraspecific variation in sex determination in *C. ocellatus*, but its mechanistic predictions

(emergence/loss of genes of major effect at high/low elevation) require testing. Sex chromosomes and sex-linked DNA in *C. ocellatus* differ only slightly between the populations (Hill *et al.* 2018, 2021a) and these populations diverged during the Pleistocene with negligible gene flow since divergence (Hill *et al.* 2021b). Specifically, and in contrast to some of the model predictions, both populations have XY heterogamety with morphologically undifferentiated X and Y sex chromosomes (Hill *et al.* 2021a) (Fig. 4e), and both populations largely share sex-linked genetic sequences (Hill *et al.* 2018). In addition, each population also possesses unique sex-linked genetic sequences. The *C. ocellatus* system highlights that only minor changes to the sex-linked genome are required for transitions in sex determination to occur: the high elevation population has more repeat and heterochromatin accumulation on the Y chromosome than the low elevation population (Hill *et al.* 2021a) and recombination among the sex-linked loci shared among populations is more suppressed in the high elevation population (Hill *et al.* 2018). This molecular work demonstrates that rather than loss/gain of genes of major effect as predicted, population-specific sex ratio responses to temperature are underpinned by sex-linked genes important for

sex determination in both populations. However, the theoretical model (Fig. 4) is useful because it establishes testable hypotheses for how sex determination evolves in viviparous lizards, and could easily be applied to other Australian viviparous species that exhibit temperature effects on offspring sex, like *Eulamprus tympanum* (Robert and Thompson 2001). Indeed, the model predictions are consistent for taxa starting with either XY or ZW chromosomal sex determination. The diversity of viviparous lizards in Australia may provide several useful models for further research if temperature is found to affect offspring sex in additional species.

Control over timing of birth

Most viviparous animals give birth to their offspring synchronously (within minutes or hours), although some lizard species complete the act of birth over several days. This phenomenon has been termed ‘birthing asynchrony’ and is defined as a spread in births separated by a minimum of 12 h (i.e. not within the same day; While *et al.* 2007). Birthing asynchrony is analogous to hatching asynchrony in birds, whereby the eggs of a clutch are laid and hatch over several days (Magrath 1990; Stoleson and Beissinger 1995; Amundsen and Slagsvold 1996; Stenning 1996). However, unlike hatching asynchrony, birthing asynchrony appears not to result from developmental asynchrony – birds are constrained to produce one embryo at a time and thus, to lay one egg at a time. In contrast, lizards that exhibit birthing asynchrony retain fully developed offspring and give birth to them one at a time (While *et al.* 2007), even when multiple offspring are present within the same uterus. This reproductive strategy occurs in multiple species within the Egerniinae, as well as four species within the African lizard family Cordylidae (Table 3).

The presence of birthing asynchrony raises several interesting questions relating to the proximate and ultimate mechanisms that facilitate this behaviour. Most work to date has focussed on the latter. Analogous hatching patterns in birds may be a mechanism that allows parents to mediate intrabrood

conflict during times of limited resource availability (Lack 1947; Stienen and Brenninkmeijer 2006). In Egerniinae, birthing asynchrony may provide a similar advantage by influencing the competitive environment of the brood and thus mediating offspring growth, survival, or dispersal (While and Wapstra 2009). Like hatching asynchrony in birds, birthing asynchrony results in a size hierarchy in a litter. While this size hierarchy does not impact the competitive ability of each offspring, it does alter the level of competition present within a single litter (While *et al.* 2009a; While and Wapstra 2009). Social Egerniinae often live in saturated habitats that include intense competition for resources, high levels of conspecific aggression and high juvenile mortality (Chapple 2003). Birthing asynchrony may operate as a trade-off between offspring mortality/dispersal and offspring mass/growth as a result of size hierarchy (While and Wapstra 2009). Indeed, when offspring are kept together in captivity, smaller individuals, which are frequently attacked by conspecifics, grow at a slower rate and behavioural development differs between dominant and submissive individuals (Riley *et al.* 2017). In the wild, such behaviour may promote the dispersal of the subdominant individual, allowing parents to modify the number of offspring they tolerate in their home range during periods of resource limitation. Interestingly, the only other family of lizards in which birthing asynchrony has been documented, the African Cordylidae, also includes group-living species (Mouton 2011). The co-occurrence of these traits across potentially two independent transitions to simple family life indicates the possibility of links between the evolution of complex sociality and that of birthing asynchrony. However, the functional consequences of birthing asynchrony and the size hierarchies it produces remain to be investigated.

While considerable work has attempted to understand the function of birthing asynchrony, we know almost nothing about its proximate mechanisms in either lizard clade. The answer is likely to lie in the co-option of the mechanisms that underpin the birthing process itself. These mechanisms span hormonal, neuronal, and embryonic effects, all of which interact to initiate and maintain the process of parturition. Lizards with asynchronous birth may have co-opted these

Table 3. Species within the squamate subfamily Egerniinae (Australian) and family Cordylidae (African) for which the presence of birthing asynchrony has been examined

Species	Birthing asynchrony	Mean spread (range)	Reference
(a) Subfamily Egerniinae			
<i>Egernia rugosa</i>	Yes	~2 days (0–10)	Peck <i>et al.</i> (2016)
<i>Egernia stokesii</i>	Yes	~3 days (1–12)	Duffield and Bull (1996)
<i>Egernia striolata</i>	Yes	~3 days (1–7)	Bonnett and Bull (2004); J. Riley, unpubl. data
<i>Liopholis whitii</i>	Yes	~3 days (0–10)	While <i>et al.</i> (2007); Chapple (2005)
<i>Lisssolepis coventryi</i>	Yes	2–3 days	Manning (2002)
<i>Tiliqua rugosa</i>	Limited	~0 days (0–8; most births synchronous)	Bull <i>et al.</i> (1993b)
<i>Tiliqua nigrolutea</i>	No	0	A. Edwards, pers. comm.
(b) Family Cordylidae			
<i>Cordylus macropholis</i>	Yes	~2 days (1–4)	Riley <i>et al.</i> , unpubl. data
<i>Karusasaurus polyzonus</i>	Yes	~2 days (1–3)	Riley <i>et al.</i> , unpubl. data
<i>Namazonurus peersi</i>	Yes	~2 days; most litters $n = 1$ offspring	Riley <i>et al.</i> , unpubl. data
<i>Ouroborus cataphractus</i>	Yes	~1 day; most litters $n = 1$ offspring	Riley <i>et al.</i> , unpubl. data

mechanisms to finely control when birth is initiated for each individual offspring while simultaneously preventing the birth of the remainder of the litter. For example, hormones actively influence the parturition process within the mother (Chaim and Mazor 1998) by way of their presence in the bloodstream and through changes in receptor expression in uterine tissue (Blanks and Thornton 2003). Additionally, dynamic changes in uterine innervation and neuronal receptor expression could facilitate the isolation of a single embryo within the uteri of a female. This type of regionally specific neuronal fluctuation is readily evident in the mammalian cervix at term pregnancy (Chávez-Genaro *et al.* 2006; Boyd *et al.* 2009) and exemplifies the capacity for the uterine environment to change rapidly and with precision. These mechanisms are further impacted by the presence of the developing embryos, and maternal–fetal hormone signalling provides an avenue for each embryo to influence the timing of its own birth (Challis *et al.* 2000; Liggins 2000). Although all the above mechanisms present intriguing targets for co-option during the evolutionary refinement of live birth, none have yet come under empirical scrutiny.

The integration of physiological, histological, and molecular techniques provides an opportunity to tease apart these competing mechanisms and generate a holistic understanding of the parturition process, and, more explicitly, how timing of birth is controlled by mothers and/or offspring. Specifically, the use of contraction bioassays to measure uterine contractile responses to key hormones (Paul *et al.* 2020), in combination with measuring changes in neuronal density across regions of the uterus to gauge the capacity for uterine relaxation, have the potential to demonstrate how this behaviour is achieved. Furthermore, combining contractile assays with transcriptomic data will allow identification of the regions of the genome that are associated with the contractile and relaxation responses. Such information should be compared across clades to examine whether similar mechanisms have been co-opted in independent lineages in a convergent pattern. Combined, this will enhance our understanding of how systemic reproductive innovations emerge, and will provide fundamental knowledge of how the process of live birth itself evolved. These results would have implications for a broad range of disciplines including evolutionary biology, conservation biology, and even human health.

Conclusions

We have identified several areas where research on Australian lizards has provided key advances in our understanding of animal reproduction, and areas where their potential as models for research has been underutilised. Australian lizards are particularly important models for studying the origins and mechanisms underlying sociality and mating systems. They exhibit a continuum of social behaviours, from largely solitary to highly social, which provides a framework for understanding how sociality evolves. Lizards communicate using chemical and/or visual signals, and these signals correlate with their mating behaviours. Thus, how their signals and mating behaviours coevolve is a potentially rich field for

future research, especially given the diversity that Australian lizards exhibit between communal sociality and solitary life. Furthermore, their mating behaviours feed into a complex arena of sexual conflict, where males and females both exhibit potential adaptations to increase their own fitness at the expense of their mate's. The fitness consequences of mating systems, as well as postmating mechanisms like cryptic female choice and sperm competition, thus provide a basis for understanding how mechanisms of communication might provide a starting point for more complex social behaviours to evolve.

Australian lizards are also important models for postfertilisation components of reproduction, especially the evolution of nesting decisions, viviparity, placentation, sex determination mechanisms, and asynchronous birth. Each of these traits has important consequences for how animal species survive in the face of environmental change. Our review shows how nesting decisions, TSD, and viviparity allow females to manipulate and potentially optimise the fitness of their offspring in response to environmental conditions like temperature, moisture, and population sex ratio. Placentation may provide a mechanism for females to maintain fetal survival in response to changes in food abundance during reproductive events. Birthing asynchrony allows the timing of parturition to be modified, possibly in response to social or environmental cues, so that all offspring are not born into the same environment.

Although we have covered many key topics, additional questions and areas of investigation where Australian lizards would provide excellent models for study remain. We have highlighted a list of outstanding key questions that were illuminated by our review in Table 4. It is not an exhaustive list, but we highlight questions that emerged after considering the review of all our topics. For example, communication, sociality, and environment-dependent variation (or fitness) are key topics that appear repeatedly in our review, and have major consequences for several areas of investigation. Our review demonstrates the utility of Australian lizards for addressing these questions.

In addition to the questions that emerged from our review, we also catalogued several topics that our review did not cover (Table 5). These topics were neglected by necessity rather than by choice, because no authors responded to our initial invitation to the Australian Society of Herpetologists listserv to cover these topics. There are certainly active research projects focusing on these topics that we did not review sufficiently as a result. However, the lack of response for these topics may indicate that these fields are areas for which lizards are relatively underutilised. We have suggested possible outstanding questions for which lizards would be ideal models to address.

Our review clearly highlights that the Australian dragons (Agamidae) and skinks (Scincidae) have contributed to understanding broad biological questions, in part because of their large number of species, and their special features such as sexual colour dimorphisms and reproductive modes. By contrast, we have highlighted only one study on goannas (Varanidae), and none on any of the gekkotans (Carphodactylidae, Diplodactylidae, Gekkonidae,

Table 4. Key questions that emerged from this synthesis of Australian lizard reproduction research

Topic of study	Synthesised key questions
Sociality	<p>What are the roles of polyandry and monogamy in suppressing or selecting for sociality?</p> <p>What are the molecular, neural, and hormonal mechanisms underpinning social traits and social evolution?</p> <p>How are social cues communicated and how do social cues and social complexity coevolve?</p>
Communication and signalling	<p>How do maternal and paternal behaviours become co-opted and refined during the early evolution of parental care?</p> <p>How do pheromonal and colour signalling combine to affect social and mating behaviours?</p> <p>How do colour and/or pheromonal signalling systems coevolve with mating strategy?</p> <p>Do specific colours communicate consistent signals across species, and how are those colours perceived and interpreted by the brain?</p> <p>How do pheromonal and colour signalling systems drive prezygotic isolation during speciation?</p>
Sexual selection, mating, and cryptic female choice	<p>What neural structures evolve to support complex communication and social behaviours associated with reproduction?</p> <p>How does sexual conflict evolve, and what are the interacting roles of male courtship strategy, female mating choice, sperm competition, and cryptic female choice?</p> <p>Can female copulatory and postcopulatory mechanisms override male courtship and precopulatory mechanisms to determine paternity, or <i>vice versa</i>?</p> <p>What is the role of large hemipenes in Australian agamids?</p> <p>If sociality reduces multiple paternity, how is the evolution of sperm competition affected?</p>
Female reproductive effort	<p>What environmental factors drive variation in the trade-offs between egg size, egg number, and reproductive frequency?</p> <p>What are the roles of proximate environmental constraints and ultimate selection pressures in driving variation in reproductive output? What is the function of hemiclitores in female lizards?</p>
Nesting ecology	<p>What environmental factors drive the evolution of communal and/or complex nesting behaviours?</p> <p>What environmental, ecological, and physiological features influence where, how, and when females construct their nests?</p> <p>How much flexibility is there in when a female oviparous lizard oviposits, does it involve embryonic diapause and what physiological and endocrinological mechanisms regulate it?</p>
Viviparity and placentation	<p>What are the genetic and physiological mechanisms required for viviparity and placentation to evolve?</p> <p>What are the selective pressures associated with the evolution of viviparity and placentation?</p>
Temperature-dependent sex determination	<p>Does TSD evolve as a result of sex-reversal, sex-specific fitness benefits of incubation temperature, or both?</p> <p>Alternatively, do sex-specific fitness benefits of incubation temperature drive the spread of TSD through a taxon after it evolves initially due to sex reversal?</p> <p>How common are TSD mechanisms (or sex-reversal) in viviparous taxa? Is their apparent scarcity in viviparity real, or a consequence of sampling?</p> <p>Is sex reversal unidirectional or bidirectional, does sex determination mechanism evolve because of biased sex ratios in any direction, or just one direction?</p>
Birth asynchrony	<p>How are sex-determining genes deleted, rearranged in chromosomes, or regulated when sex reversals occur?</p> <p>What is the function of asynchronous birth?</p> <p>What maternal and fetal physiological mechanisms control birth?</p> <p>How does asynchronous birth evolve?</p>

Table 5. Outstanding topics and questions neglected by our review

Topic	Outstanding questions
Evolutionary ecology and mechanisms of parthenogenesis	<p>Parthenogenesis occurs in Australian lizards (Scincidae: Menetia, Gekkonidae: Heteronotia: Kearney 2003), but there is far less literature on Australian lizards than on American teiid lizards. What are the mechanisms underlying parthenogenesis in Australian lizards, and how does it evolve?</p> <p>Is parthenogenesis in Australian lizards homologous or analogous with that in American lizards?</p>
Reproductive endocrinology	<p>How are reproductive behaviours and decisions hormonally controlled? This is an important area of research that Australian lizards are well suited to address (Jones 2017; Jones and Swain 2000). More specific questions include:</p> <p>How are spermatogenesis and spermiogenesis controlled in males?</p> <p>How do females 'decide' on number of ovarian follicles to recruit, number of eggs to ovulate, or egg size?</p> <p>How do maternal and fetal hormones interact to control or prolong pregnancy?</p> <p>How does the hormonal control of reproductive phenology change within a species across environments with different seasonal cycles?</p>
Reproductive responses to environmental novelty	<p>Australia has been especially impacted by environmental changes, including invasive species, habitat modification, and climate change. How has lizard reproduction changed as a result?</p> <p>Australia's smaller and shorter-lived species, such as annual agamids and some skinks, are ideal model organisms for testing how new selection pressures in novel environments drive changes in reproductive phenotype.</p> <p>Furthermore, the 'patchwork' of altered landscapes offers an opportunity for natural experiments on this topic. This work may also be critical to conserving species threatened by environmental change (Sinervo <i>et al.</i> 2010).</p>

Pygopodidae). Despite this fact, it seems unlikely that the latter families do not offer potential opportunities for significant research on broad reproductive questions. For example, the gekkonids are unusual for laying calcareous-shelled eggs, which provides a good comparison with the flexible-shelled eggs of species in the other gekkotan families, with implications for embryonic physiology and development (Andrews *et al.* 2013). Apart from the research on deep-nesting varanid lizards (see section *Nesting ecology in Australian goannas*), little is known about nests and nesting in most Australian lizards. Nest sites, nest structure, and environmental features, especially temperatures and water potentials, remain unknown for most species. Given the range of potential nest environments in Australia, from extreme aridity to moist temperate regions, investigations of nest and nesting behaviours, and their impact on embryonic physiology, are likely to be informative for the understanding of the evolution of reproductive strategies (viviparity, sex determination, incubation periods, rates of development and so on), and for predicting future effects of climate change. Gekkotans will be equally as important in these investigations as are dragons and skinks.

In summary, Australian lizards are diverse, providing opportunities for exciting research in reproductive biology. We have highlighted key areas of research that are ongoing, emerging, and relatively neglected. We hope that new generations of reproductive biologists and ecologists will be inspired by our review to consider these topics for their own research careers. To encourage those who are excited by the topics in our review, we have provided a list of the authors responsible for each section in our supplementary materials (Supplementary Table S1). We urge readers to contact the relevant authors for research opportunities in the topics that they find interesting. Australian lizards offer excellent opportunities to test important hypotheses in vertebrate reproductive biology. Their abundance and the diversity of environments they inhabit provide an important resource for ongoing ecological and evolutionary research.

Data availability statement

Data sharing is not applicable as no new data were generated or analysed during this study.

Conflicts of interest

The authors declare no conflicts of interest.

Declaration of funding

This research did not receive any specific funding.

Acknowledgements

We thank Professor Paul Cooper for the invitation to prepare this review *in lieu* of a plenary presentation by MBT at the 2020 meeting of the Australian and New Zealand Society of Comparative Physiology and Biochemistry, which was cancelled because of the COVID-19 pandemic. We thank two anonymous reviewers for their helpful comments. We acknowledge the use of the Atlas of Living Australia, grid.506668.b.

References

- Abell, A. J. (1997). Estimating paternity with spatial behaviour and DNA fingerprinting in the striped plateau lizard, *Sceloporus virgatus* (Phrynosomatidae). *Behavioral Ecology and Sociobiology* **41**, 217–226. doi:10.1007/s002650050382
- Alonzo, S. H., and Sinervo, B. (2001). Mate choice games, context-dependent good genes, and genetic cycles in the side-blotched lizard, *Uta stansburiana*. *Behavioral Ecology and Sociobiology* **49**, 176–186. doi:10.1007/s002650000265
- Amundsen, T., and Slagsvold, T. (1996). Lack's brood reduction hypothesis and avian hatching asynchrony: what's next? *Oikos* **76**, 613–620. doi:10.2307/3546359
- Andersson, M. B. (1994). 'Sexual Selection.' (Princeton University Press: Princeton, NJ.)
- Andrews, R. M., Thompson, M. B., and Green, V. W. (2013). Does low gas permeability of rigid-shelled gekkotan eggs affect embryonic development? *Journal of Experimental Zoology. Part A, Ecological and Integrative Physiology* **319**, 259–267.
- Angilletta, M. J. (2009). 'Thermal Adaptation: a Theoretical and Empirical Synthesis.' (Oxford University Press: Oxford, UK.)
- Arnqvist, G. (2014). Cryptic female choice. In 'The Evolution of Insect Mating Systems'. (Eds S. D. Simmons and L. W. Simmons.) pp. 204–220. (Oxford University Press: Oxford, UK.)
- Bachtrog, D., Mank, J.E., Peichel, C.L., Kirkpatrick, M., Otto, S.P., Ashman, T.L., Hahn, M.W., Kitano, J., Mayrose, I., Ming, R., Perrin, N., Ross, L., Valenzuela, N., and Vamosi, J.C. Tree of Sex Consortium(2014). Sex determination: why so many ways of doing it? *PLoS Biology* **12**, e1001899. doi:10.1371/journal.pbio.1001899
- Beck, L. A., O'Bryant, E. L., and Wade, J. S. (2008). Sex and seasonal differences in morphology of limbic forebrain nuclei in the green anole lizard. *Brain Research* **1227**, 68–75. doi:10.1016/j.brainres.2008.06.021
- Biazik, J. M., Thompson, M. B., and Murphy, C. R. (2007). The tight junctional protein occludin is found in the uterine epithelium of squamate reptiles. *Journal of Comparative Physiology. B, Biochemical, Systemic, and Environmental Physiology* **177**, 935–943. doi:10.1007/s00360-007-0192-1
- Biazik, J. M., Thompson, M. B., and Murphy, C. R. (2008). Claudin-5 is restricted to the tight junction region of uterine epithelial cells in the uterus of pregnant/gravid squamate reptiles. *The Anatomical Record* **291**, 547–556. doi:10.1002/ar.20677
- Biazik, J. M., Thompson, M. B., and Murphy, C. R. (2010). Desmosomes in the uterine epithelium of noninvasive skink placentae. *The Anatomical Record* **293**, 502–512. doi:10.1002/ar.21093
- Black, D. G. (1983). Encephalization of Australian lizards. Ph.D. Thesis, Monash University, Melbourne.
- Blackburn, D. G. (2000). Classification of the reproductive patterns of amniotes. *Herpetological Monograph* **14**, 371–377. doi:10.2307/1467051
- Blackburn, D. G. (2015). Evolution of vertebrate viviparity and specializations for fetal nutrition: a quantitative and qualitative analysis. *Journal of Morphology* **276**, 961–990. doi:10.1002/jmor.20272
- Blanks, A. M., and Thornton, S. (2003). The role of oxytocin in parturition. *BJOG* **110**, 46–51. doi:10.1016/S1470-0328(03)00024-7
- Böhm, M., Collen, B., Baillie, J. E. M., Bowles, P., Chanson, J., and Cox, N. *et al.* (2013). The conservation status of the world's reptiles. *Biological Conservation* **157**, 372–385. doi:10.1016/j.biocon.2012.07.015
- Böhme, W. (1995). Hemclitoris discovered: a fully differentiated erectile structure in female monitor lizards (*Varanus* spp.) (Reptilia: Varanidae). *Journal of Zoological Systematics and Evolutionary Research* **33**, 129–132.
- Böhme, W., and Ziegler, T. (2009). A review of iguanian and anguimorph lizard genitalia (Squamata: Chamaeleonidae; Varanoidea, Shinisauridae, Xenosauridae, Anguinae) and their phylogenetic significance: comparisons with molecular data sets. *Journal of Zoological Systematics and Evolutionary Research* **47**, 189–202. doi:10.1111/j.1439-0469.2008.00495.x

- Bókonyi, V., Kövér, S., Nemesházi, E., Liker, A., and Székely, T. (2017). Climate-driven shifts in adult sex ratios via sex reversals: the type of sex determination matters. *Philosophical Transactions of the Royal Society B: Biological Sciences* **372**, 20160325. doi:10.1098/rstb.2016.0325
- Bonnett, M., and Bull, C. M. (2004). *Egernia striolata* (tree skink) reproduction. *Herpetological Review* **25**, 389
- Bordogna, G., Cunningham, G., Fitzpatrick, L. J., Halliwell, B., MacGregor, H. E. A., Munch, K. L., Wapstra, E., and While, G. M. (2016). An experimental test of relatedness-based mate discrimination in a social lizard. *Behavioral Ecology and Sociobiology* **70**, 2139–2147. doi:10.1007/s00265-016-2217-9
- Botterill-James, T., Halliwell, B., Cooper-Scott, E., Uller, T., Wapstra, E., and While, G. M. (2016). Habitat structure influences the extent of parent-offspring association in a social lizard. *Frontiers in Social Evolution* **4**, 96.
- Boyd, J. W., Lechuga, T. J., Ebner, C. A., Kirby, M. A., and Yellon, S. M. (2009). Cervix remodeling and parturition in the rat: lack of a role for hypogastric innervation. *Reproduction* **137**, 739–748. doi:10.1530/REP-08-0507
- Boyle, M., Hone, J., Schwanz, L. E., and Georges, A. (2014). Under what conditions do climate-driven sex ratios enhance versus diminish population persistence? *Ecology and Evolution* **4**, 4522–4533. doi:10.1002/ece3.1316
- Bradbury, J. W., and Vehrencamp, S. L. (2011). 'Principles of Animal Communication.' 2nd edn. (Sinauer: Sunderland, MA.)
- Brandley, M. C., Bragg, J. G., Singhal, S., Chapple, D. G., Jennings, C. K., Lemmon, A. R., Lemmon, E. M., Thompson, M. B., and Moritz, C. (2015). Evaluating the performance of anchored hybrid enrichment at the tips of the tree of life: a phylogenetic analysis of Australian *Eugongylus* group scincid lizards. *BMC Evolutionary Biology* **15**, 62. doi:10.1186/s12862-015-0318-0
- Brennan, P. L. (2016). Studying genital coevolution to understand intermittent organ morphology. *Integrative and Comparative Biology* **56**, 669–681. doi:10.1093/icb/icw018
- Brennan, P. L., and Prum, R. O. (2015). Mechanisms and evidence of genital coevolution: the roles of natural selection, mate choice, and sexual conflict. In 'The Genetics and Biology of Sexual Conflict'. (Eds W. R. Rice and S. Gavrillets.) pp. 385–406. (Cold Spring Harbor Laboratory Press: Cold Spring Harbor, NY.)
- Bukhari, S., Saul, M., James, N., Bensky, M., Stein, L., Trapp, R., and Bell, A. (2019). Neurogenomic insights into paternal care and its relation to territorial aggression. *Nature Communications* **10**, 4437. doi:10.1038/s41467-019-12212-7
- Bull, J. J. (1980). Sex determination in reptiles. *The Quarterly Review of Biology* **55**, 3–21. doi:10.1086/411613
- Bull, J. J. (1981). Evolution of environmental sex determination from genotypic sex determination. *Heredity* **47**, 173–184. doi:10.1038/hdy.1981.74
- Bull, C. M. (1988). Mate fidelity in an Australian lizard *Trachydosaurus rugosus*. *Behavioral Ecology and Sociobiology* **23**, 45–49. doi:10.1007/BF00303057
- Bull, C. M. (2000). Monogamy in lizards. *Behavioural Processes* **51**, 7–20. doi:10.1016/S0376-6357(00)00115-7
- Bull, C. M., and Baghurst, B. (1998). Home range overlap of mothers and their offspring in the sleepy lizard, *Tiliqua rugosa*. *Behavioral Ecology and Sociobiology* **42**, 357–362. doi:10.1007/s002650050448
- Bull, M., and Lindle, C. (2002). Following trails of partners in the monogamous lizard, *Tiliqua rugosa*. *Acta Ethologica* **5**, 25–28. doi:10.1007/s10211-002-0063-4
- Bull, C. M., Bedford, G. S., and Schulz, B. A. (1993a). How do sleepy lizards find each other? *Herpetologica* **49**, 294–300.
- Bull, C. M., Pamula, Y., and Schulz, L. (1993b). Parturition in the sleepy lizard, *Tiliqua rugosa*. *Journal of Herpetology* **27**, 489–492. doi:10.2307/1564848
- Bull, C. M., Cooper, S. J. B., and Baghurst, B. C. (1998). Social monogamy and extra-pair fertilization in an Australian lizard, *Tiliqua rugosa*. *Behavioral Ecology and Sociobiology* **44**, 63–72. doi:10.1007/s002650050515
- Bull, C. M., Griffin, C. L., and Johnston, G. R. (1999a). Olfactory discrimination in scat-piling lizards. *Behavioral Ecology* **10**, 136–140. doi:10.1093/beheco/10.2.136
- Bull, C. M., Griffin, C. L., and Perkins, M. V. (1999b). Some properties of a pheromone allowing individual recognition, from the scats of an Australian lizard, *Egernia striolata*. *Acta Ethologica* **2**, 35–42. doi:10.1007/PL00012230
- Bull, C. M., Griffin, C. L., Lanham, E. J., and Johnston, G. R. (2000). Recognition of pheromones from group members in a gregarious lizard, *Egernia stokesii*. *Journal of Herpetology* **34**, 92–99. doi:10.2307/1565244
- Bull, C. M., Gardner, M. G., Sih, A., Spiegel, O., Godfrey, S. S., and Leu, S. (2017). Why is social behaviour rare in reptiles? Lessons from sleepy lizards. *Advances in the Study of Behavior* **49**, 1–26. doi:10.1016/bs.asb.2017.02.001
- Bulmer, M., and Bull, J. (1982). Models of polygenic sex determination and sex ratio control. *Evolution* **36**, 13–26. doi:10.1111/j.1558-5646.1982.tb05005.x
- Byrne, M. (2008). Evidence for multiple refugia at different time scales during Pleistocene climatic oscillations in southern Australia inferred from phylogeography. *Quaternary Science Reviews* **27**, 2576–2585. doi:10.1016/j.quascirev.2008.08.032
- Campos, S. M., Pruett, J. A., Soini, H. A., Zúñiga-Vega, J. J., Goldberg, J. K., Vital-García, C., Hews, D. K., Novotny, M. V., and Martins, E. P. (2020). Volatile fatty acid and aldehyde abundances evolve with behavior and habitat temperature in *Sceloporus* lizards. *Behavioral Ecology* **31**, 978–991. doi:10.1093/beheco/araa044
- Castelli, M. A., Georges, A., Cherryh, C., Rosauer, D. F., Sarre, S. D., Contador-Kelsall, I., and Holleley, C. E. (2021). Evolving thermal thresholds explain the distribution of temperature sex reversal in an Australian dragon lizard. *Diversity & Distributions* **27**, 427–438. doi:10.1111/ddi.13203
- Chaim, W., and Mazor, M. (1998). The relationship between hormones and human parturition. *Archives of Gynecology and Obstetrics* **262**, 43–51. doi:10.1007/s004040050226
- Challis, J., Sloboda, D., Matthews, S., Holloway, A., Alfaidy, N., Howe, D., Fraser, M., and Newnham, J. (2000). Fetal hypothalamic-pituitary adrenal (HPA) development and activation as a determinant of the timing of birth, and of postnatal disease. *Endocrine Research* **26**, 489–504. doi:10.3109/07435800009048560
- Chapple, D. G. (2003). Ecology, life-history, and behavior in the Australian scincid genus *Egernia*, with comments on the evolution of complex sociality in lizards. *Herpetological Monograph* **17**, 145–180. doi:10.1655/0733-1347(2003)017[0145:ELABIT]2.0.CO;2
- Chapple, D. G. (2005). Life history and reproductive ecology of White's skink, *Egernia whitii*. *Australian Journal of Zoology* **53**, 353–360. doi:10.1071/ZO05030
- Chapple, D., and Keogh, J. S. (2005). Complex mating system and dispersal patterns in a social lizard, *Egernia whitii*. *Molecular Ecology* **14**, 1215–1227. doi:10.1111/j.1365-294X.2005.02486.x
- Chapple, D., and Keogh, J. S. (2006). Group structure and stability in social aggregations of White's skink, *Egernia whitii*. *Ethology* **112**, 247–257. doi:10.1111/j.1439-0310.2006.01153.x
- Charnier, M. (1966). Action de la température sur le sex-ratio chez l'embryon d'*Agama agama* (Agamidae, Lacertilien). *Comptes Rendus des Seances de la Societe de Biologie et de Ses Filiales* **160**, 1470–1472.
- Charnov, E. L., and Bull, J. (1977). When is sex environmentally determined? *Nature* **266**, 828–830. doi:10.1038/266828a0

- Chávez-Genaro, R., Lombide, P., and Anesetti, G. (2006). A quantitative study of rat uterine sympathetic innervation during pregnancy and post partum. *Reproduction, Fertility and Development* **18**, 525–531. doi:10.1071/RD05053
- Chelini, M., Brock, K., Yeager, J., and Edwards, D. (2021). Environmental drivers of sexual dimorphism in a lizard with alternative mating strategies. *Journal of Evolutionary Biology* **34**, 1241–1255. doi:10.1111/jeb.13881
- Christian, K. (2004). *Varanus panoptes*. In 'Varanoid Lizards of the World'. (Eds E. R. Pianka and D. King.) pp. 423–429. (Indiana University Press: Bloomington.)
- Cogger, H. G. (2018). 'Reptiles & Amphibians of Australia.' 7th edn. (CSIRO Publishing: Melbourne.)
- Cole, C. J. (1966). Femoral glands in lizards: a review. *Herpetologica* **22**, 199–206.
- Cooper, W. E. (1994). Chemical discrimination by tongue-flicking in lizards: a review with hypotheses on its origin and its ecological and phylogenetic relationships. *Journal of Chemical Ecology* **20**, 439–487. doi:10.1007/BF02064449
- Cooper, W. E. (1998). Evaluation of swab and related tests as a bioassay for assessing responses by squamate reptiles to chemical stimuli. *Journal of Chemical Ecology* **24**, 841–866. doi:10.1023/A:1022373517653
- Cooper, W. E., and Grastka, W. R. (1987). Lingual responses to chemical fractions of urodaeal glandular pheromone to the skink *Eumeces laticeps*. *The Journal of Experimental Zoology* **242**, 249–253. doi:10.1002/jez.1402420302
- Cooper, W. E., and Trauth, S. E. (1992). Discrimination of conspecific male and female cloacal chemical stimuli by males and possession of a probable pheromone gland by females in a cordylid lizard, *Gerrhosaurus nigrolineatus*. *Herpetologica* **48**, 229–236.
- Corl, A., Davis, A. R., Kuchta, S. R., Comendant, T., and Sinervo, B. (2010a). Alternative mating strategies and the evolution of sexual size dimorphism in the side-blotched lizard, *Uta stansburiana*: a population-level comparative analysis. *Evolution* **64**, 79–96. doi:10.1111/j.1558-5646.2009.00791.x
- Corl, A., Davis, A. R., Kuchta, S. R., and Sinervo, B. (2010b). Selective loss of polymorphic mating types is associated with rapid phenotypic evolution during morphic speciation. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 4254–4259. doi:10.1073/pnas.0909480107
- Cornejo-Páramo, P., Dissanayake, D. S. B., Lira-Noriega, A., Martínez-Pacheco, M. L., Acosta, A., Ramírez-Suástegui, C., Méndez-de-la-Cruz, F. R., Székely, T., Urrutia, A. O., Georges, A., and Cortez, D. (2020a). Viviparous reptile regarded to have temperature-dependent sex determination has old XY chromosomes. *Genome Biology and Evolution* **12**, 924–930. doi:10.1093/gbe/evaa104
- Cornejo-Páramo, P., Lira-Noriega, A., Ramírez-Suástegui, C., Méndez-De-La-Cruz, F. R., Székely, T., Urrutia, A. O., and Cortez, D. (2020b). Sex determination systems in reptiles are related to ambient temperature but not to the level of climatic fluctuation. *BMC Evolutionary Biology* **20**, 103. doi:10.1186/s12862-020-01671-y
- Cornwallis, C., West, S., Davis, K., and Griffin, A. (2010). Promiscuity and the evolutionary transition to complex societies. *Nature* **466**, 969–972. doi:10.1038/nature09335
- Cox, R. M., and Calsbeek, R. (2010). Cryptic sex-ratio bias provides indirect genetic benefits despite sexual conflict. *Science* **328**, 92–94. doi:10.1126/science.1185550
- Cree, A., Thompson, M. B., and Daugherty, C. H. (1995). Tuatara sex determination. *Nature* **375**, 543. doi:10.1038/375543a0
- Crespi, B., and Semeniuk, C. (2004). Parent–offspring conflict in the evolution of vertebrate reproductive mode. *American Naturalist* **163**, 635–653. doi:10.1086/382734
- Cunningham, G. D., While, G. M., and Wapstra, E. (2017). Climate and sex ratio variation in a viviparous lizard. *Biology Letters* **13**, 20170218. doi:10.1098/rsbl.2017.0218
- Cunningham, G. D., While, G. M., Olsson, M., Ljungström, G., and Wapstra, E. (2020). Degrees of change: between and within population variation in thermal reaction norms of phenology in a viviparous lizard. *Ecology* **101**, e03136. doi:10.1002/ecy.3136
- Darwin, C. (1871). 'The Descent of Man, and Selection in Relation to Sex.' (John Murray: London.)
- Davis, A. R., Corl, A., Surget-Groba, Y., and Sinervo, B. (2011). Convergent evolution of kin-based sociality in a lizard. *Proceedings of the Royal Society B: Biological Sciences* **278**, 1507–1514. doi:10.1098/rspb.2010.1703
- Dissanayake, D. S. B., Holleley, C. E., Hill, L. K., O'Meally, D., Deakin, J. E., and Georges, A. (2020). Identification of Y chromosome markers in the eastern three-lined skink (*Bassiana duperreyi*) using *in silico* whole genome subtraction. *BMC Genomics* **21**, 667. doi:10.1186/s12864-020-07071-2
- Dissanayake, D. S. B., Holleley, C. E., Deakin, J. E., and Georges, A. (2021). High elevation increases the risk of Y chromosome loss in alpine skink populations with sex reversal. *Heredity* **126**, 805–816. doi:10.1038/s41437-021-00406-z
- Dong, C., Johnston, G., Stuart-Fox, D., Moussalli, A., Rankin, K., and McLean, C. (2021a). Elevation of divergent color polymorphic and monomorphic lizard lineages (Squamata: Agamidae) to species level. *Ichthyology & Herpetology* **109**, 43–54. doi:10.1643/h2020064
- Dong, C. M., Rankin, K. J., McLean, C. A., and Stuart-Fox, D. (2021b). Maternal reproductive output and F1 hybrid fitness may influence contact zone dynamics. *Journal of Evolutionary Biology* **34**, 680–694. doi:10.1111/jeb.13772
- Doody, J. S., Freedberg, S., and Keogh, J. S. (2009). Communal egg-laying in reptiles and amphibians: evolutionary patterns and hypotheses. *The Quarterly Review of Biology* **84**, 229–252. doi:10.1086/605078
- Doody, J. S., Burghardt, G. M., and Dinets, V. (2013). Breaking the social–asocial dichotomy: a role for reptiles in social behaviour research? *Ethology* **119**, 95–103. doi:10.1111/eth.12047
- Doody, J. S., James, H., Ellis, R., Gibson, N., Raven, M., Mahony, S., Hamilton, D. G., Rhind, D., Clulow, S., and McHenry, C. R. (2014). Cryptic and complex nesting in the yellow-spotted monitor, *Varanus panoptes*. *Journal of Herpetology* **48**, 363–370. doi:10.1670/13-006
- Doody, J. S., James, H., Colyvas, K., Mchenry, C. R., and Clulow, S. (2015). Deep nesting in a lizard, *déjà vu* devil's corkscrews: first helical reptile burrow and deepest vertebrate nest. *Biological Journal of the Linnean Society* **116**, 13–26. doi:10.1111/bij.12589
- Doody, J. S., Rhind, D., Green, B., Castellano, C., McHenry, C., and Clulow, S. (2017). Chronic effects of an invasive species on an animal community. *Ecology* **98**, 2093–2101. doi:10.1002/ecy.1889
- Doody, J. S., McHenry, C., Brown, M., Canning, G., Vas, G., and Clulow, S. (2018a). Deep, helical, communal nesting and emergence in the sand monitor: ecology informing paleoecology? *Journal of Zoology* **305**, 88–95. doi:10.1111/jzo.12543
- Doody, J. S., McHenry, C. R., Durkin, L., Brown, M., Simms, A., Coleman, L., Phizacklea, C., Jones, H., Phizacklea, O., and Clulow, S. (2018b). Deep communal nesting by yellow-spotted monitors in a desert ecosystem: indirect evidence for a response to extreme dry conditions. *Herpetologica* **74**, 306–310. doi:10.1655/Herpetologica-D-18-00004.1
- Doody, J. S., McGlashan, J., Fryer, H., Coleman, L., James, H., Soennichsen, K., Rhind, D., and Clulow, S. (2020). Plasticity in nest site choice behavior in response to hydric conditions in a reptile. *Scientific Reports* **10**, 16048. doi:10.1038/s41598-020-73080-6
- Doody, J. S., Burghardt, G. M., and Dinets, V. (2021a). 'The Secret Social Lives of Reptiles.' (Johns Hopkins University Press: Baltimore, MD.)

- Doody, J. S., Soennichsen, K. F., James, H., McHenry, C., and Clulow, S. (2021b). Ecosystem engineering by deep-nesting monitor lizards. *Ecology* **102**, e03271. doi:10.1002/ecy.3271
- Dooren, T. J. V., and Leimar, O. (2003). The evolution of environmental and genetic sex determination in fluctuating environments. *Evolution* **57**, 2667–2677. doi:10.1111/j.0014-3820.2003.tb01511.x
- Duffield, G. A., and Bull, C. M. (1996). Characteristics of the litter of the gidgee skink, *Egernia stokesii*. *Wildlife Research* **23**, 337–342. doi:10.1071/WR9960337
- Duffield, G. A., and Bull, C. M. (2002). Stable social aggregations in an Australian lizard, *Egernia stokesii*. *Naturwissenschaften* **89**, 424–427. doi:10.1007/s00114-002-0346-7
- Düsing, C. (1884). Die Regulierung des Geschlechtsverhältnisses bei der Vermehrung der Menschen, Tiere und Pflanzen. *Jenaische Zeitschrift Für Naturwissenschaft* **17**, 593–940. [translated by Edwards, A., 2000.
- Düsing, K. (1884). On the regulation of the sex ratio. *Theoretical Population Biology* **58**, 255–257]
- Eberhard, W. G. (1996). 'Female Control: Sexual Selection by Cryptic Female Choice.' (Princeton University Press: Princeton, NJ.)
- Ebrahimi, M., Godfrey, S. S., Fenner, A. L., and Bull, C. M. (2014). Mating behaviour in pygmy bluetongue lizards: do females 'attract' male lizards? *Australian Journal of Zoology* **62**, 491–497. doi:10.1071/ZO14055
- Eckhardt, F., Kappeler, P. M., and Kraus, C. (2017). Highly variable lifespan in an annual reptile, Labord's chameleon (*Furcifer labordi*). *Scientific Reports* **7**, 11397. doi:10.1038/s41598-017-11701-3
- Edwards, D. L., Melville, J., Joseph, L., and Keogh, J. S. (2015). Ecological divergence, adaptive diversification, and the evolution of social signaling traits: an empirical study in arid Australian lizards. *American Naturalist* **186**, E144–E161. doi:10.1086/683658
- Eizaguirre, C., Laloï, D., Massot, M., Richard, M., Federici, P., and Clobert, J. (2007). Condition dependence of reproductive strategy and the benefits of polyandry in a viviparous lizard. *Proceedings of the Royal Society B: Biological Sciences* **274**, 425–430. doi:10.1098/rspb.2006.3740
- Ezaz, T., Sarre, S. D., O'Meally, D., Graves, J. A. M., and Georges, A. (2009). Sex chromosome evolution in lizards: independent origins and rapid transitions. *Cytogenetic and Genome Research* **127**, 249–260. doi:10.1159/000300507
- Fenner, A. L., Pavey, C., and Bull, C. M. (2012). Behavioural observations and use of burrow systems by an endangered Australian arid-zone lizard, Slater's skink (*Liopholis slateri*). *Australian Journal of Zoology* **60**, 127–132. doi:10.1071/ZO12047
- Firman, R. C., Gasparini, C., Manier, M. K., and Pizzari, T. (2017). Postmating female control: 20 years of cryptic female choice. *Trends in Ecology & Evolution* **32**, 368–382. doi:10.1016/j.tree.2017.02.010
- Fischer, K., Brakefield, P. M., and Zwaan, B. J. (2003). Plasticity in butterfly egg size: why larger offspring at lower temperatures? *Ecology* **84**, 3138–3147. doi:10.1890/02-0733
- Fischer, E., Roland, A., Moskowitz, N., Tapia, E., Kyle, S., Coloma, L., and O'Connell, L. (2019). The neural basis of tadpole transport in poison frogs. *Proceedings of the Royal Society B: Biological Sciences* **286**, 20191084. doi:10.1098/rspb.2019.1084
- Fisher, R. A. (1930). 'The Genetical Theory of Natural Selection.' (Clarendon Press: Oxford, UK.)
- Fisher, R. A. (1958). 'The Genetical Theory of Natural Selection.' (Dover Publications, Inc.: New York.)
- Fitze, P. S., Galliard, J. F. L., Federici, P., Richard, M., and Clobert, J. (2005). Conflict over multiple-partner mating between males and females of the polygynandrous common lizards. *Evolution* **59**, 2451–2459. doi:10.1111/j.0014-3820.2005.tb00954.x
- Foster, C. S. P., Thompson, M. B., Van Dyke, J. U., Brandley, M. C., and Whittington, C. M. (2020). Emergence of an evolutionary innovation: gene expression differences associated with the transition between oviparity and viviparity. *Molecular Ecology* **29**, 1315–1327. doi:10.1111/mec.15409
- Fox, S. F., and Shipman, P. A. (2003). Social behavior at high and low elevations: environmental release and phylogenetic effects in *Liolaemus*. In 'Lizard Social Behavior'. (Eds S. F. Fox, J. K. McCoy, and T. A. Baird.) pp. 310–355. (Johns Hopkins University Press: Baltimore and London.)
- Frere, C. H., Chandrasoma, D., and Whiting, M. J. (2015). Polyandry in dragon lizards: inbred paternal genotypes sire fewer offspring. *Ecology and Evolution* **5**, 1686–1692. doi:10.1002/ece3.1447
- Friesen, C. R., Uhrig, E. J., Squire, M. K., Mason, R. T., and Brennan, P. L. R. (2014). Sexual conflict over mating in red-sided garter snakes (*Thamnophis sirtalis*) as indicated by experimental manipulation of genitalia. *Proceedings of the Royal Society B: Biological Sciences* **281**, 20132694. doi:10.1098/rspb.2013.2694
- Friesen, C. R., Uhrig, E. J., Mason, R. T., and Brennan, P. L. R. (2016). Female behaviour and the interaction of male and female genital traits mediate sperm transfer during mating. *Journal of Evolutionary Biology* **29**, 952–964. doi:10.1111/jeb.12836
- Friesen, C. R., Johansson, R., and Olsson, M. (2017a). Morph-specific metabolic rate and the timing of reproductive senescence in a color polymorphic dragon. *Journal of Experimental Zoology. Part A, Ecological and Integrative Physiology* **327**, 433–443. doi:10.1002/jez.2118
- Friesen, C. R., Wilson, M. R., Rollings, N., Sudyka, J., Whittington, C. M., Giraudeau, M., and Olsson, M. (2017b). Conditional handicaps in exuberant lizards: bright color in aggressive males is correlated with high levels of free radicals. *Frontiers in Ecology and Evolution* **5**, 1. doi:10.3389/fevo.2017.00001
- Friesen, C. R., de Graaf, S. P., and Olsson, M. (2019). The relationship of body condition, superoxide dismutase and superoxide with sperm performance. *Behavioral Ecology* **30**, 1351–1363. doi:10.1093/beheco/arz086
- Friesen, C. R., Kahrl, A. F., and Olsson, M. (2020a). Sperm competition in squamate reptiles. *Philosophical Transactions of the Royal Society B: Biological Sciences* **375**, 20200079. doi:10.1098/rstb.2020.0079
- Friesen, C. R., Noble, D. W. A., and Olsson, M. (2020b). The role of oxidative stress in postcopulatory selection. *Philosophical Transactions of the Royal Society B: Biological Sciences* **375**, 20200065. doi:10.1098/rstb.2020.0065
- Friesen, C. R., Rollings, N., Wilson, M., Whittington, C. M., Shine, R., and Olsson, M. (2020c). Covariation in superoxide, sperm telomere length and sperm velocity in a polymorphic reptile. *Behavioral Ecology and Sociobiology* **74**, 74. doi:10.1007/s00265-020-02855-8
- Friesen, C. R., Wilson, M., Rollings, N., Sudyka, J., Giraudeau, M., Whittington, C. M., and Olsson, M. (2021). Exercise training has morph-specific effects on telomere, body condition and growth dynamics in a color-polymorphic lizard. *The Journal of Experimental Biology* **224**, jeb242164. doi:10.1242/jeb.242164
- García-Roa, R., Jara, M., Baeckens, S., López, P., Damme, R. V., Martín, J., and Pincheira-Donoso, D. (2017). Macroevolutionary diversification of glands for chemical communication in squamate reptiles. *Scientific Reports* **7**, 9288. doi:10.1038/s41598-017-09083-7
- Gardner, M., Bull, C., Cooper, S., and Duffield, G. (2000). Microsatellite mutations in litters of the Australian lizard *Egernia stokesii*. *Journal of Evolutionary Biology* **13**, 551. doi:10.1046/j.1420-9101.2000.00189.x
- Gardner, M. G., Bull, C. M., Cooper, S. J. B., and Duffield, G. A. (2001). Genetic evidence for a family structure in stable social aggregations of the Australian lizard *Egernia stokesii*. *Molecular Ecology* **10**, 175–183. doi:10.1046/j.1365-294X.2001.01171.x

- Gardner, M. G., Bull, C. M., and Cooper, S. J. B. (2002). High levels of genetic monogamy in the group-living Australian lizard *Egernia stokesii*. *Molecular Ecology* **11**, 1787–1794. doi:10.1046/j.1365-294X.2002.01552.x
- Gardner, M. G., Hugall, A., Donnellan, S. C., Hutchinson, M., and Foster, R. (2008). Molecular systematics of social skinks: phylogeny and taxonomy of the *Egernia* group (Reptilia: Scincidae). *Zoological Journal of the Linnean Society* **154**, 781–794. doi:10.1111/j.1096-3642.2008.00422.x
- Gardner, M. G., Godfrey, S. S., Fenner, A. L., Donnellan, S. C., and Bull, C. M. (2012). Fine-scale spatial structuring as an inbreeding avoidance mechanism in the social skink *Egernia stokesii*. *Australian Journal of Zoology* **60**, 272–277. doi:10.1071/ZO12089
- Gardner, M. G., Pearson, S. K., Johnston, G. R., and Schwarz, M. P. (2016). Group living in squamate reptiles: a review of evidence for stable aggregations. *Biological Reviews of the Cambridge Philosophical Society* **91**, 925–936. doi:10.1111/brv.12201
- Garland, T., and Adolph, S. C. (1994). Why not to do two-species comparative studies: limitations on inferring adaptation. *Physiological Zoology* **67**, 797–828. doi:10.1086/physzool.67.4.30163866
- Geffroy, B., and Wedekind, C. (2020). Effects of global warming on sex ratios in fishes. *Journal of Fish Biology* **97**, 596–606. doi:10.1111/jfb.14429
- Georges, A., and Holleley, C. E. (2018). How does temperature determine sex? *Science* **360**, 601–602. doi:10.1126/science.aat5993
- Greer, A. E. (1989). 'The Biology and Evolution of Australian Lizards.' (Surrey Beatty and Sons: Chipping Norton, Australia.)
- Griffith, O. W., and Wagner, G. P. (2017). The placenta as a model for understanding the origin and evolution of vertebrate organs. *Nature Ecology and Evolution* **1**, 0072. doi:10.1038/s41559-017-0072
- Griffith, O. W., Ujvari, B., Belov, K., and Thompson, M. B. (2013). Placental lipoprotein lipase (LPL) gene expression in a placentotrophic lizard, *Pseudemoia entrecasteauxii*. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **320**, 465–470. doi:10.1002/jez.b.22526
- Griffith, O. W., Brandley, M. C., Belov, K., and Thompson, M. B. (2016). Allelic expression of mammalian imprinted genes in a matrotrophic lizard, *Pseudemoia entrecasteauxii*. *Development Genes and Evolution* **226**, 79–85. doi:10.1007/s00427-016-0531-x
- Griffith, O. W., Brandley, M. C., Whittington, C. M., Belov, K., and Thompson, M. B. (2017). Comparative genomics of hormonal signaling in the chorioallantoic membrane of oviparous and viviparous amniotes. *General and Comparative Endocrinology* **244**, 19–29. doi:10.1016/j.ygcen.2016.04.017
- Grossen, C., Neuenschwander, S., and Perrin, N. (2011). Temperature-dependent turnovers in sex-determination mechanisms: a quantitative model. *Evolution* **65**, 64–78. doi:10.1111/j.1558-5646.2010.01098.x
- Hacking, J., Stuart-Fox, D., and Gardner, M. (2017). Very low rate of multiple paternity detected in clutches of a wild agamid lizard. *Australian Journal of Zoology* **65**, 328–334. doi:10.1071/ZO18006
- Haenel, G. J., Smith, L. C., and John-Alder, H. B. (2003). Home-range analysis in *Sceloporus undulatus*. II. A test of spatial relationships and reproductive success. *Copeia* **2003**, 113–123. doi:10.1643/0045-8511(2003)003[0113:HRASUJ]2.0.CO;2
- Halliwell, B., Uller, T., Chapple, D., Gardner, M. G., Wapstra, E., and While, G. M. (2017a). Habitat saturation promotes delayed dispersal in a social reptile. *Behavioral Ecology* **28**, 515–522. doi:10.1093/beheco/arw181
- Halliwell, B., Uller, T., Holland, B., and While, G. M. (2017b). Live bearing promotes the evolution of sociality in reptiles. *Nature Communications* **8**, 2030. doi:10.1038/s41467-017-02220-w
- Harlow, P. (2004). Temperature-dependent sex determination in lizards. In 'Temperature-dependent Sex Determination in Vertebrates'. (Eds N. Valenzuela and V. A. Lance.) pp. 42–52. (Smithsonian Books: Washington, DC.)
- Head, M. L., Keogh, J. S., and Doughty, P. (2005). Male southern water skinks (*Eulamprus heatwolei*) use both visual and chemical cues to detect female sexual receptivity. *Acta Ethologica* **8**, 79–85. doi:10.1007/s10211-005-0003-1
- Head, M. L., Doughty, P., Blomberg, S. P., and Keogh, J. S. (2008). Chemical mediation of reciprocal mother–offspring recognition in the southern water skink (*Eulamprus heatwolei*): chemical mediation of kin recognition in a lizard. *Austral Ecology* **33**, 20–28. doi:10.1111/j.1442-9993.2007.01785.x
- Healey, M., and Olsson, M. (2009). Too big for his boots: are social costs keeping condition-dependent status signalling honest in an Australian lizard? *Austral Ecology* **34**, 636–640. doi:10.1111/j.1442-9993.2009.01968.x
- Healey, M., Uller, T., and Olsson, M. (2007). Seeing red: morph-specific contest success and survival rates in a colour-polymorphic agamid lizard. *Animal Behaviour* **74**, 337–341. doi:10.1016/j.anbehav.2006.09.017
- Hendrawan, K., Whittington, C. M., Brandley, M. C., Belov, K., and Thompson, M. B. (2017). The regulation of uterine proinflammatory gene expression during pregnancy in the live-bearing lizard, *Pseudemoia entrecasteauxii*. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **328**, 334–346. doi:10.1002/jez.b.22733
- Herbert, J. F., Lindsay, L. A., Murphy, C. R., and Thompson, M. B. (2006). Calcium transport across the uterine epithelium of pregnant lizards. *Herpetological Monographs* **20**, 205–211. doi:10.1655/0733-1347(2007)20[205:CTATUE]2.0.CO;2
- Hill, P. L., Burrridge, C. P., Ezaz, T., and Wapstra, E. (2018). Conservation of sex-linked markers among conspecific populations of a viviparous skink, *Niveoscincus ocellatus*, exhibiting genetic and temperature-dependent sex determination. *Genome Biology and Evolution* **10**, 1079–1087. doi:10.1093/gbe/evy042
- Hill, P., Shams, F., Burrridge, C. P., Wapstra, E., and Ezaz, T. (2021a). Differences in homomorphic sex chromosomes are associated with population divergence in sex determination in *Carinascincus ocellatus* (Scincidae: Lygosominae). *Cells* **10**, 291. doi:10.3390/cells10020291
- Hill, P. L., Wapstra, E., Ezaz, T., and Burrridge, C. P. (2021b). Pleistocene divergence in the absence of gene flow among populations of a viviparous reptile with intraspecific variation in sex determination. *Ecology and Evolution* **11**, 5575–5583. doi:10.1002/ece3.7458
- Holleley, C. E., O'Meally, D., Sarre, S. D., Marshall Graves, J. A., Ezaz, T., Matsubara, K., Azad, B., Zhang, X., and Georges, A. (2015). Sex reversal triggers the rapid transition from genetic to temperature-dependent sex. *Nature* **523**, 79–82. doi:10.1038/nature14574
- Holleley, C. E., Sarre, S. D., O'Meally, D., and Georges, A. (2016). Sex reversal in reptiles: reproductive oddity or powerful driver of evolutionary change? *Sexual Development* **10**, 279–287. doi:10.1159/000450972
- Hoops, D., Ullmann, J. F. P., Janke, A. L., Vidal-García, M., Gardner, T. S., Dwihapsari, Y., Merklings, T., Price, W. S., Endler, J. A., Whiting, M. J., and Keogh, J. S. (2017). Sexual selection predicts brain structure in dragon lizards. *Journal of Evolutionary Biology* **30**, 244–256. doi:10.1111/jeb.12984
- Houck, L. D. (2009). Pheromone communication in amphibians and reptiles. *Annual Review of Physiology* **71**, 161–176. doi:10.1146/annurev.physiol.010908.163134
- Huey, R. B., Pianka, E. R., and Schoener, T. W. (1983). 'Lizard Ecology: Studies of a Model Organism.' (Harvard University Press: Cambridge, MA.)

- Hughes, W., Oldroyd, B., Beekman, M., and Ratnieks, F. (2008). Ancestral monogamy shows kin selection is key to the evolution of eusociality. *Science* **320**, 1213–1216. doi:10.1126/science.1156108
- Hutchinson, M. N., and Donnellan, S. C. (1992). Taxonomy and genetic variation in the Australian lizards of the genus *Pseudemoia* (Scincidae: Lygosominae). *Journal of Natural History* **26**, 215–264. doi:10.1080/00222939200770091
- Huxley, J. (1955). Morphism and evolution. *Heredity* **9**, 1–52. doi:10.1038/hdy.1955.1
- Itonaga, K., Jones, S. M., and Wapstra, E. (2012a). Effects of maternal basking and food quantity during gestation provide evidence for the selective advantage of matrotrophy in a viviparous lizard. *PLoS One* **7**, e41835. doi:10.1371/journal.pone.0041835
- Itonaga, K., Wapstra, E., and Jones, S. M. (2012b). A novel pattern of placental leucine transfer during mid to late gestation in a highly placentalotrophic viviparous lizard. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **318**, 308–315. doi:10.1002/jez.b.22446
- Jansson, N., Uller, T., and Olsson, M. (2005). Female dragons, *Ctenophorus pictus*, do not prefer scent from unrelated males. *Australian Journal of Zoology* **53**, 279–282. doi:10.1071/ZO05024
- Janzen, F. J., and Krenz, J. G. (2004). Phylogenetics: which was first, TSD or GSD? In 'Temperature-Dependent Sex Determination in Vertebrates'. (Eds N. Valenzuela, and V. A. Lance) pp. 121–130. (Smithsonian Institution: Washington, DC.)
- Janzen, F. J., and Phillips, P. C. (2006). Exploring the evolution of environmental sex determination, especially in reptiles. *Journal of Evolutionary Biology* **19**, 1775–1784. doi:10.1111/j.1420-9101.2006.01138.x
- Jennions, M. D., and Petrie, M. (2000). Why do females mate multiply? A review of the genetic benefits. *Biological Reviews of the Cambridge Philosophical Society* **75**, 21–64. doi:10.1017/S0006323199005423
- Johnson, M. A., Cohen, R. E., Vandecar, J. R., and Wade, J. (2011). Relationships among reproductive morphology, behavior, and testosterone in a natural population of green anole lizards. *Physiology & Behavior* **104**, 437–445. doi:10.1016/j.physbeh.2011.05.004
- Jones, S. M. (2017). Variations upon a theme: Australian lizards provide insights into the endocrine control of vertebrate reproductive cycles. *General and Comparative Endocrinology* **244**, 60–69. doi:10.1016/j.ygcen.2015.09.004
- Jones, S. M., and Swain, R. (2000). Effects of exogenous FSH on follicular recruitment in a viviparous lizard *Niveoscincus metallicus* (Scincidae). *Comparative Biochemistry and Physiology. Part A, Molecular & Integrative Physiology* **127**, 487–493. doi:10.1016/S1095-6433(00)00279-8
- Jones, S. M., and Swain, R. (2006). Placental transfer of ³H-oleic acid in three species of viviparous lizards: a route for supplementation of embryonic fat bodies? *Herpetological Monographs* **20**, 186–193. doi:10.1655/0733-1347(2007)20[186:PTOHOA]2.0.CO;2
- Karlson, P., and Lüscher, M. (1959). 'Pheromones': a new term for a class of biologically active substances. *Nature* **183**, 55–56. doi:10.1038/183055a0
- Kearney, M. R. (2003). Why is sex so unpopular in the Australian desert? *Trends in Ecology & Evolution* **18**, 605–607. doi:10.1016/j.tree.2003.09.021
- Keogh, J. S., Noble, D. W., Wilson, E. E., and Whiting, M. J. (2012). Activity predicts male reproductive success in a polygynous lizard. *PLoS One* **7**, e38856. doi:10.1371/journal.pone.0038856
- Keogh, J. S., Umbers, K. D., Wilson, E., Stapley, J., and Whiting, M. J. (2013). Influence of alternate reproductive tactics and pre- and postcopulatory sexual selection on paternity and offspring performance in a lizard. *Behavioral Ecology and Sociobiology* **67**, 629–638. doi:10.1007/s00265-013-1482-0
- King, D., and Green, B. (1999). 'Goannas: The Biology of Varanid Lizards.' (UNSW Press: Kensington, NSW.)
- Kinlaw, A., and Grasmueck, M. (2012). Evidence for and geomorphologic consequences of a reptilian ecosystem engineer: the burrowing cascade initiated by the gopher tortoise. *Geomorphology* **157–158**, 108–121. doi:10.1016/j.geomorph.2011.06.030
- Klaczko, J., Ingram, T., and Losos, J. (2015). Genitals evolve faster than other traits in *Anolis* lizards. *Journal of Zoology* **295**, 44–48. doi:10.1111/jzo.12178
- Kvarnemo, C., and Simmons, L. W. (2013). Polyandry as a mediator of sexual selection before and after mating. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences* **368**, 20120042. doi:10.1098/rstb.2012.0042
- Lack, D. (1947). The significance of clutch-size. *The Ibis* **89**, 302–352. doi:10.1111/j.1474-919X.1947.tb04155.x
- LaDage, L. D., Gutzke, W. H., Simmons, R. A., and Ferkin, M. H. (2008). Multiple mating increases fecundity, fertility and relative clutch mass in the female leopard gecko (*Eublepharis macularius*). *Ethology* **114**, 512–520. doi:10.1111/j.1439-0310.2008.01495.x
- Laird, M. K., Thompson, M. B., and Whittington, C. M. (2019). Facultative oviparity in a viviparous skink (*Saiphos equalis*). *Biology Letters* **15**, 20180827. doi:10.1098/rsbl.2018.0827
- Laloi, D., Richard, M., Fédérici, P., Clobert, J., Teillac-Deschamps, P., and Massot, M. (2009). Relationship between female mating strategy, litter success and offspring dispersal. *Ecology Letters* **12**, 823–829. doi:10.1111/j.1461-0248.2009.01335.x
- Laloi, D., Eizaguirre, C., Fédérici, P., and Massot, M. (2011). Female choice for heterozygous mates changes along successive matings in a lizard. *Behavioural Processes* **88**, 149–154. doi:10.1016/j.beproc.2011.08.011
- Le Galliard, J. F., Fitze, P. S., Ferriere, R., and Clobert, J. (2005). Sex ratio bias, male aggression, and population collapse in lizards. *Proceedings of the National Academy of Sciences of the United States of America* **102**, 18231–18236. doi:10.1073/pnas.0505172102
- Lebas, N. (2001). Microsatellite determination of male reproductive success in a natural population of the territorial ornate dragon lizard, *Ctenophorus ornatus*. *Molecular Ecology* **10**, 193–203. doi:10.1046/j.1365-294X.2001.01174.x
- Leu, S., Burzacott, D., Whiting, M., and Bull, C. M. (2015). Mate familiarity affects pairing behaviour in a long-term monogamous lizard: evidence from detailed bio-logging and a 31-year field study. *Ethology* **121**, 760–768. doi:10.1111/eth.12390
- Liggins, G. C. (2000). The role of the hypothalamic–pituitary–adrenal axis in preparing the fetus for birth. *American Journal of Obstetrics and Gynecology* **182**, 475–477. doi:10.1016/S0002-9378(00)70241-9
- Linville, B. J., Stewart, J. R., Ecay, T. W., Herbert, J. F., Parker, S. L., and Thompson, M. B. (2010). Placental calcium provision in a lizard with prolonged oviductal egg retention. *Journal of Comparative Physiology. B, Biochemical, Systemic, and Environmental Physiology* **180**, 221–227. doi:10.1007/s00360-009-0400-2
- Lovern, M. B., Holmes, M. M., and Wade, J. (2004). The green anole (*Anolis carolinensis*): a reptilian model for laboratory studies of reproductive morphology and behavior. *ILAR Journal* **45**, 54–64. doi:10.1093/ilar.45.1.54
- Lukas, D., and Clutton-Brock, T. (2012). Cooperative breeding and monogamy in mammalian societies. *Proceedings. Biological Sciences* **279**, 2151–2156. doi:10.1098/rspb.2011.2468
- Lüpold, S., de Boer, R. A., Evans, J. P., Tomkins, J. L., and Fitzpatrick, J. L. (2020). How sperm competition shapes the evolution of testes and sperm: a meta-analysis. *Philosophical Transactions of the Royal Society B: Biological Sciences* **375**, 20200064. doi:10.1098/rstb.2020.0064

- Maderson, P. F. A. (1972). The structure and evolution of holocrine epidermal glands in sphaerodactylina and eublepharina gekkonid lizards. *Copeia* **1972**, 559–571. doi:10.2307/1442930
- Madsen, T., Shine, R., Loman, J., and Håkansson, T. (1992). Why do female adders copulate so frequently? *Nature* **355**, 440–441. doi:10.1038/355440a0
- Magrath, R. D. (1990). Hatching asynchrony in altricial birds. *Biological Reviews of the Cambridge Philosophical Society* **65**, 587–622. doi:10.1111/j.1469-185X.1990.tb01239.x
- Main, A. R., and Bull, C. M. (1996). Mother–offspring recognition in two Australian lizards, *Tiliqua rugosa* and *Egernia stokesii*. *Animal Behaviour* **52**, 193–200. doi:10.1006/anbe.1996.0164
- Manning, B. (2002). Notes on the captive breeding of the swamp skink (*Egernia coventryi*). *Herpetofauna* **32**, 35–38.
- Mannix, L. (2020). Doris the lady lizard now appears to be male, stunning scientists. *The Sydney Morning Herald*, 17 December 2020. Available at: <https://www.smh.com.au/national/doris-the-lady-lizard-now-appears-to-be-male-stunning-scientists-20201217-p56obk.html>
- Martin, L. D., and Bennett, D. K. (1977). The burrows of the Miocene beaver *Palaeocastor*, western Nebraska, USA. *Palaeogeography, Palaeoclimatology, Palaeoecology* **22**, 173–193. doi:10.1016/0031-0182(77)90027-X
- Martín, J., and López, P. (2011). Pheromones and reproduction in reptiles. In 'Hormones and Reproduction of Vertebrates'. (Eds D. O. Norris, and K. H. Lopez.) pp. 141–167. (Academic Press: San Diego, CA.)
- Martínez-Torres, M., Rubio-Morales, B., Piña-Amado, J. J., and Luis, J. (2015). Hemipenes in females of the Mexican viviparous lizard *Barisia imbricata* (Squamata: Anguillidae): an example of heterochrony in sexual development. *Evolution & Development* **17**, 270–277. doi:10.1111/ede.12134
- Mason, R. T., and Parker, M. R. (2010). Social behavior and pheromonal communication in reptiles. *Journal of Comparative Physiology. A, Neuroethology, Sensory, Neural, and Behavioral Physiology* **196**, 729–749. doi:10.1007/s00359-010-0551-3
- Masters, C., and Shine, R. (2003). Sociality in lizards: family structure in free-living King's skinks *Egernia kingii* from southwestern Australia. *Australian Zoologist* **32**, 377–380. doi:10.7882/AZ.2002.015
- Mayerl, C., Baeckens, S., and Damme, R. V. (2015). Evolution and role of the follicular epidermal gland system in non-ophidian squamates. *Amphibia-Reptilia* **36**, 185–206. doi:10.1163/15685381-00002995
- Maynard Smith, J., and Szathmáry, E. (1997). 'The Major Transitions in Evolution.' (Oxford University Press: Oxford, UK.)
- McDiarmid, R. W., Foster, M. S., Guyer, C., Gibbons, J. W., and Chernoff, N. (2012). 'Reptile Biodiversity: Standard Methods for Inventory and Monitoring.' (University of California Press: Berkeley, CA.)
- McDiarmid, C. S., Friesen, C. R., Ballen, C., and Olsson, M. (2017). Sexual coloration and sperm performance in the Australian painted dragon lizard, *Ctenophorus pictus*. *Journal of Evolutionary Biology* **30**, 1303–1312. doi:10.1111/jeb.13092
- McLean, C. A., Stuart-Fox, D., and Moussalli, A. (2014). Phylogeographic structure, demographic history and morph composition in a colour polymorphic lizard. *Journal of Evolutionary Biology* **27**, 2123–2137. doi:10.1111/jeb.12464
- McLean, C. A., Stuart-Fox, D., and Moussalli, A. (2015). Environment, but not genetic divergence, influences geographic variation in colour morph frequencies in a lizard. *BMC Evolutionary Biology* **15**, 156. doi:10.1186/s12862-015-0442-x
- Meiri, S., Avila, L., Bauer, A. M., Chapple, D. G., Das, I., Doan, T. M., Doughty, P., Ellis, R., Grismer, L., Kraus, F., Morando, M., Oliver, P., Pincheira-Donoso, D., Ribeiro-Junior, M. A., Shea, G., Torres-Carvajal, O., Slavenko, A., and Roll, U. (2020). The global diversity and distribution of lizard clutch sizes. *Global Ecology and Biogeography* **29**, 1515–1530. doi:10.1111/geb.13124
- Melville, J., Chapple, D. G., Keogh, J. S., Sumner, J., Amey, A., and Bowles, P. *et al.* (2021). A return-on-investment approach for prioritization of rigorous taxonomic research needed to inform responses to the biodiversity crisis. *PLoS Biology* **19**, e3001210. doi:10.1371/journal.pbio.3001210
- Mendonça, M., and Crews, D. (2001). Control of attractivity and receptivity in female red-sided garter snakes. *Hormones and Behavior* **40**, 43–50. doi:10.1006/hbeh.2001.1665
- Mesquita, D. O., Costa, G. C., Colli, G. R., Costa, T. B., Shepard, D. B., Vitt, L. J., and Pianka, E. R. (2016). Life-history patterns of lizards of the world. *American Naturalist* **187**, 689–705. doi:10.1086/686055
- Milstead, W. (1967). 'Lizard Ecology: A Symposium.' (University of Missouri Press: Columbia, MO.)
- Mitchell, N. J., and Janzen, F. J. (2010). Temperature-dependent sex determination and contemporary climate change. *Sexual Development* **4**, 129–140. doi:10.1159/000282494
- Mitchell, N. J., Kearney, M. R., Nelson, N. J., and Porter, W. P. (2008). Predicting the fate of a living fossil: how will global warming affect sex determination and hatching phenology in tuatara? *Proceedings of the Royal Society B: Biological Sciences* **275**, 2185–2193. doi:10.1098/rspb.2008.0438
- Mork, L., Czerwinski, M., and Capel, B. (2014). Predetermination of sexual fate in a turtle with temperature-dependent sex determination. *Developmental Biology* **386**, 264–271. doi:10.1016/j.ydbio.2013.11.026
- Morrison, C., and Hero, J. M. (2003). Geographic variation in life-history characteristics of amphibians: a review. *Journal of Animal Ecology* **72**, 270–279. doi:10.1046/j.1365-2656.2003.00696.x
- Morrison, S., Scott Keogh, J., and Scott, I. (2002). Molecular determination of paternity in a natural population of the multiply mating polygynous lizard *Eulamprus heatwolei*. *Molecular Ecology* **11**, 535–545. doi:10.1046/j.0962-1083.2002.01450.x
- Mouton, P. L. F. N. (2011). Aggregation behaviour of lizards in the arid western regions of South Africa. *African Journal of Herpetology* **60**, 155–170. doi:10.1080/21564574.2011.562926
- Mouton, P. L. F. N., Van Rensburg, D. J., and Van Wyk, J. H. (2010). Epidermal glands in cordylid lizards, with special reference to generation glands. *Zoological Journal of the Linnean Society* **158**, 312–324. doi:10.1111/j.1096-3642.2009.00547.x
- Mouton, P. L. F. N., Flemming, A. F., and Broeckhoven, C. (2014). Generation gland morphology in cordylid lizards: an evolutionary perspective. *Journal of Morphology* **275**, 456–464. doi:10.1002/jmor.20227
- Munch, K., Noble, D., Budd, L., Row, A., Wapstra, E., and While, G. M. (2018). Maternal presence facilitates plasticity in offspring behavior: insights into the evolution of parental care. *Behavioral Ecology* **29**, 1298–1306. doi:10.1093/beheco/ary122
- Murphy, C. R., Hosie, M. J., and Thompson, M. B. (2000). The plasma membrane transformation facilitates pregnancy in both reptiles and mammals. *Comparative Biochemistry and Physiology. Part A, Molecular & Integrative Physiology* **127**, 433–439. doi:10.1016/S1095-6433(00)00274-9
- Murphy, K., Hudson, S., and Shea, G. (2006). Reproductive seasonality of three cold-temperate viviparous skinks from southeastern Australia. *Journal of Herpetology* **40**, 454–464. doi:10.1670/0022-1511(2006)40[454:RSOTCV]2.0.CO;2
- Murphy, B. F., Parker, S. L., Murphy, C. R., and Thompson, M. B. (2010). Angiogenesis of the uterus and chorioallantois in the eastern water skink *Eulamprus quoyii*. *The Journal of Experimental Biology* **213**, 3340–3347. doi:10.1242/jeb.046862
- Murphy, B. F., Parker, S. L., Murphy, C. R., and Thompson, M. B. (2011). Placentation in the eastern water skink (*Eulamprus quoyii*): a placentome-like structure in a lecithotrophic lizard. *Journal of Anatomy* **218**, 678–689. doi:10.1111/j.1469-7580.2011.01368.x

- Neaves, L., Wapstra, E., Birch, D., Girling, J. E., and Joss, J. M. P. (2006). Embryonic gonadal and sexual organ development in a small viviparous skink, *Niveoscincus ocellatus*. *Journal of Experimental Zoology. Part A, Comparative Experimental Biology* **305A**, 74–82. doi:10.1002/jez.a.249
- Niewiarowski, P. H., and Angilletta, M. J. (2008). Countergradient variation in embryonic growth and development: do embryonic and juvenile performances trade off? *Functional Ecology* **22**, 895–901. doi:10.1111/j.1365-2435.2008.01441.x
- Noble, D. W., Keogh, J. S., and Whiting, M. J. (2013). Multiple mating in a lizard increases fecundity but provides no evidence for genetic benefits. *Behavioral Ecology* **24**, 1128–1137. doi:10.1093/beheco/art040
- Nowicki, J., Pratchett, M., Walker, S., Coker, D., and O'Connell, L. (2020). Gene expression correlates of social evolution in coral reef butterflyfishes. *Proceedings of the Royal Society B: Biological Sciences* **287**, 20200239. doi:10.1098/rspb.2020.0239
- Numan, M. (2014). 'Neurobiology of Social Behavior: Toward an Understanding of the Prosocial and Antisocial Brain.' (Academic Press: New York, NY.)
- O'Connell, L., Matthews, B., and Hofmann, H. (2012). Isotocin regulates paternal care in a monogamous cichlid fish. *Hormones and Behavior* **61**, 725–733. doi:10.1016/j.yhbeh.2012.03.009
- O'Connor, D., and Shine, R. (2003). Lizards in 'nuclear families': a novel reptilian social system in *Egernia saxatilis* (Scincidae). *Molecular Ecology* **12**, 743–752. doi:10.1046/j.1365-294X.2003.01777.x
- O'Connor, D., and Shine, R. (2004). Parental care protects against infanticide in the lizard *Egernia saxatilis* (Scincidae). *Animal Behaviour* **68**, 1361–1369. doi:10.1016/j.anbehav.2004.02.014
- Olsson, M. (1995). Forced copulation and costly female resistance behavior in the Lake Eyre dragon, *Ctenophorus maculosus*. *Herpetologica* **51**, 19–24.
- Olsson, M. (2001). No female mate choice in Mallee dragon lizards, *Ctenophorus fordi*. *Evolutionary Ecology* **15**, 129–141. doi:10.1023/A:1013865624146
- Olsson, M., and Madsen, T. (1995). Female choice on male quantitative traits in lizards – why is it so rare? *Behavioral Ecology and Sociobiology* **36**, 179–184. doi:10.1007/BF00177794
- Olsson, M., and Shine, R. (1998). Chemosensory mate recognition may facilitate prolonged mate guarding by male snow skinks, *Niveoscincus microlepidotus*. *Behavioral Ecology and Sociobiology* **43**, 359–363. doi:10.1007/s002650050502
- Olsson, M., Gullberg, A., Shine, R., Madsen, T., and Tegelström, H. (1996a). Paternal genotype influences incubation period, offspring size, and offspring shape in an oviparous reptile. *Evolution* **50**, 1328–1333. doi:10.1111/j.1558-5646.1996.tb02372.x
- Olsson, M., Shine, R., Madsen, T., Gullberg, A., and Tegelström, H. (1996b). Sperm selection by females. *Nature* **383**, 585. doi:10.1038/383585a0
- Olsson, M., Shine, R., Madsen, T., Gullberg, A., and Tegelström, H. (1997). Sperm choice by females. *Trends in Ecology & Evolution* **12**, 445–446. doi:10.1016/S0169-5347(97)85751-5
- Olsson, M., Madsen, T., Ujvari, B., and Wapstra, E. (2004). Fecundity and MHC affects ejaculation tactics and paternity bias in sand lizards. *Evolution* **58**, 906–909. doi:10.1111/j.0014-3820.2004.tb00423.x
- Olsson, M., Ujvari, B., Wapstra, E., Madsen, T., Shine, R., and Bensch, S. (2005). Does mate guarding prevent rival mating in snow skinks? A test using AFLP. *Herpetologica* **61**, 389–394. doi:10.1655/04-85.1
- Olsson, M., Healey, M., and Astheimer, L. (2007a). Afternoon T: testosterone level is higher in red than yellow male polychromatic lizards. *Physiology & Behavior* **91**, 531–534. doi:10.1016/j.physbeh.2007.04.025
- Olsson, M., Schwartz, T., Uller, T., and Healey, M. (2007b). Sons are made from old stores: sperm storage effects on sex ratio in a lizard. *Biology Letters* **3**, 491–493. doi:10.1098/rsbl.2007.0196
- Olsson, M., Healey, M., Wapstra, E., and Uller, T. (2009a). Testing the quality of a carrier: a field experiment on lizard signalers. *Evolution* **63**, 695. doi:10.1111/j.1558-5646.2008.00569.x
- Olsson, M., Schwartz, T., Uller, T., and Healey, M. (2009b). Effects of sperm storage and male colour on probability of paternity in a polychromatic lizard. *Animal Behaviour* **77**, 419–424. doi:10.1016/j.anbehav.2008.10.017
- Olsson, M., Wapstra, E., and Friesen, C. (2018a). Ectothermic telomeres: it's time they came in from the cold. *Philosophical Transactions of the Royal Society B: Biological Sciences* **373**, 20160449. doi:10.1098/rstb.2016.0449
- Olsson, M., Wapstra, E., and Friesen, C. R. (2018b). Evolutionary ecology of telomeres: a review. *Annals of the New York Academy of Sciences* **1422**, 5–28. doi:10.1111/nyas.13443
- Ossip-Klein, A. G., Fuentes, J. A., Hews, D. K., and Martins, E. P. (2013). Information content is more important than sensory system or physical distance in guiding the long-term evolutionary relationships between signaling modalities in *Sceloporus* lizards. *Behavioral Ecology and Sociobiology* **67**, 1513–1522. doi:10.1007/s00265-013-1535-4
- Oufiero, C. E., and Angilletta, M. J. Jr (2006). Convergent evolution of embryonic growth and development in the eastern fence lizard (*Sceloporus undulatus*). *Evolution* **60**, 1066–1075. doi:10.1111/j.0014-3820.2006.tb01183.x
- Palmer, D. H., Rogers, T. F., Dean, R., and Wright, A. E. (2019). How to identify sex chromosomes and their turnover. *Molecular Ecology* **28**, 4709–4724. doi:10.1111/mec.15245
- Parker, G. A. (1970). Sperm competition and its evolutionary consequences in insects. *Biological Reviews of the Cambridge Philosophical Society* **45**, 525–567. doi:10.1111/j.1469-185X.1970.tb01176.x
- Parker, G. A. (1998). Sperm competition and the evolution of ejaculates: towards a theory base. In 'Sperm Competition and Sexual Selection'. (Eds T. R. Birkhead, and A. P. Møller.) pp. 3–49. (Academic Press: San Deigo, CA.)
- Parker, M. R., and Mason, R. T. (2012). How to make a sexy snake: estrogen activation of female sex pheromone in male red-sided garter snakes. *The Journal of Experimental Biology* **215**, 723–730. doi:10.1242/jeb.064923
- Parker, M. R., and Mason, R. T. (2014). A novel mechanism regulating a sexual signal: the testosterone-based inhibition of female sex pheromone expression in garter snakes. *Hormones and Behavior* **66**, 509–516. doi:10.1016/j.yhbeh.2014.07.007
- Parker, G. A., and Pizzari, T. (2010). Sperm competition and ejaculate economics. *Biological Reviews of the Cambridge Philosophical Society* **85**, 897–934.
- Parker, S. L., Manconi, F., Murphy, C. R., and Thompson, M. B. (2010a). Uterine and placental angiogenesis in the Australian skinks, *Ctenotus taeniolatus*, and *Saiphos equalis*. *The Anatomical Record* **293**, 829–838. doi:10.1002/ar.21052
- Parker, S. L., Murphy, C. R., and Thompson, M. B. (2010b). Uterine angiogenesis in squamate reptiles: implications for the evolution of viviparity. *Herpetological Conservation and Biology* **5**, 330–334.
- Paul, J. W., Kemsley, J. O., Butler, T. A., Tolosa, J. M., Thompson, M. B., Smith, R., and Whittington, C. M. (2020). A comparison of uterine contractile responsiveness to arginine vasopressin in oviparous and viviparous lizards. *Journal of Comparative Physiology. B, Biochemical, Systemic, and Environmental Physiology* **190**, 49–62. doi:10.1007/s00360-019-01254-4
- Pearson, S. K., Godfrey, S. S., Bull, C. M., and Gardner, M. (2016). Larger lizards live longer in the group-living *Egernia stokesii*. *Australian Journal of Zoology* **64**, 182–191. doi:10.1071/ZO16024
- Peck, S., Gardner, M. G., Seddon, J. M., and Baxter, G. (2016). Life-history characteristics of the yakka skink, *Egernia rugosa*, indicate long-term social structure. *Australian Journal of Zoology* **64**, 335–343. doi:10.1071/ZO16061

- Pen, I., Uller, T., Feldmeyer, B., Harts, A., While, G. M., and Wapstra, E. (2010). Climate-driven population divergence in sex-determining systems. *Nature* **468**, 436–438. doi:10.1038/nature09512
- Pennell, M. W., Mank, J. E., and Peichel, C. L. (2018). Transitions in sex determination and sex chromosomes across vertebrate species. *Molecular Ecology* **27**, 3950–3963. doi:10.1111/mec.14540
- Peterson, C. C., and Husak, J. F. (2006). Locomotor performance and sexual selection: individual variation in sprint speed of collared lizards (*Crotaphytus collaris*). *Copeia* **2006**, 216–224. doi:10.1643/0045-8511(2006)6[216:LPASSI]2.0.CO;2
- Pianka, E. R. (1969). Habitat specificity, speciation, and species density in Australian desert lizards. *Ecology* **50**, 498–502. doi:10.2307/1933908
- Pianka, E. R. (1989). Desert lizard diversity: additional comments and some data. *American Naturalist* **134**, 344–364. doi:10.1086/284985
- Pieau, C. (1972). Effects de la temperature sur le developpement des glandes genitales chez les embryons de deux Chelonians, *Testudo graeca* L. et *Emys orbicularis* L. *Comptes rendus de l'Académie des Sciences (Paris)* **274**, 719–722.
- Pizzari, T., and Wedell, N. (2013). The polyandry revolution. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences* **368**, 20120041. doi:10.1098/rstb.2012.0041
- Platel, M. R. (1979). Brain weight–body weight relationships. In 'Biology of the Reptilia. Vol. 9'. (Eds C. Gans, R. G. Northcutt, and P. Ulinski.) pp. 147–171. (Academic Press: London.)
- Pokorna, M., and Kratochvil, L. (2009). Phylogeny of sex-determining mechanisms in squamate reptiles are sex chromosomes an evolutionary trap? *Zoological Journal of the Linnean Society* **156**, 168–183. doi:10.1111/j.1096-3642.2008.00481.x
- Powney, G. D., Grenyer, R., Orme, C. D. L., Owens, I. P. F., and Meiri, S. (2010). Hot, dry and different: Australian lizard richness is unlike that of mammals, amphibians and birds. *Global Ecology and Biogeography* **19**, 386–396. doi:10.1111/j.1466-8238.2009.00521.x
- Qualls, C. P., and Shine, R. (1998). *Lerista bougainvillii*, a case study for the evolution of viviparity in reptiles. *Journal of Evolutionary Biology* **11**, 63–78. doi:10.1007/s000360050066
- Quinn, A. E., Georges, A., Sarre, S. D., Guarino, F., Ezaz, T., and Graves, J. A. M. (2007). Temperature sex reversal implies sex gene dosage in a reptile. *Science* **316**, 411. doi:10.1126/science.1135925
- Radder, R. S., Quinn, A. E., Georges, A., Sarre, S. D., and Shine, R. (2008). Genetic evidence for co-occurrence of chromosomal and thermal sex-determining systems in a lizard. *Biology Letters* **4**, 176–178. doi:10.1098/rsbl.2007.0583
- Rankin, D., and Kokko, H. (2007). Do males matter? The role of males in population dynamics. *Oikos* **116**, 335–348. doi:10.1111/j.0030-1299.2007.15451.x
- Rankin, K. J., McLean, C. A., Kemp, D. J., and Stuart-Fox, D. (2016). The genetic basis of discrete and quantitative colour variation in the polymorphic lizard, *Ctenophorus decresii*. *BMC Evolutionary Biology* **16**, 179. doi:10.1186/s12862-016-0757-2
- Riley, J. L., Noble, D. W., Byrne, R. W., and Whiting, M. J. (2017). Early social environment influences the behaviour of a family-living lizard. *Royal Society Open Science* **4**, 161082. doi:10.1098/rsos.161082
- Robert, K. A., and Thompson, M. B. (2001). Sex determination. Viviparous lizard selects sex of embryos. *Nature* **412**, 698–699. doi:10.1038/35089135
- Rollings, N., Friesen, C. R., Sudyka, J., Whittington, C., Giraudeau, M., Wilson, M., and Olsson, M. (2017). Telomere dynamics in a lizard with morph-specific reproductive investment and self-maintenance. *Ecology and Evolution* **7**, 5163–5169. doi:10.1002/ece3.2712
- Romero-Diaz, C., Campos, S. M., Herrmann, M. A., Soini, H. A., Novotny, M. V., Hews, D. K., and Martins, E. P. (2021). Composition and compound proportions affect the response to complex chemical signals in a spiny lizard. *Behavioral Ecology and Sociobiology* **75**, 42. doi:10.1007/s00265-021-02987-5
- Rubenstein, D. R., and Abbot, P. (2017). 'Comparative Social Evolution.' (Cambridge University Press: Cambridge, UK.)
- Ruiz-Monachesi, M. R., Paz, A., and Quipildor, M. (2019). Hemipenes eversion behavior: a new form of communication in two *Liolaemus* lizards (Iguania: Liolaemidae). *Canadian Journal of Zoology* **97**, 187–194. doi:10.1139/cjz-2018-0195
- Sánchez-Martínez, P. M., Ramírez-Pinilla, M. P., and Miranda-Esquivel, D. R. (2007). Comparative histology of the vaginal–cloacal region in Squamata and its phylogenetic implications. *Acta Zoologica* **88**, 289–307. doi:10.1111/j.1463-6395.2007.00282.x
- Sarre, S. D., Georges, A., and Quinn, A. (2004). The ends of a continuum: genetic and temperature-dependent sex determination in reptiles. *BioEssays* **26**, 639–645. doi:10.1002/bies.20050
- Sarre, S. D., Ezaz, T., and Georges, A. (2011). Transitions between sex-determining systems in reptiles and amphibians. *Annual Review of Genomics and Human Genetics* **12**, 391–406. doi:10.1146/annurev-genom-082410-101518
- Schofield, J., Gardner, M. G., Fenner, A. L., and Bull, C. M. (2014). Promiscuous mating in the endangered Australian lizard *Tiliqua adelaidensis*: a potential windfall for its conservation. *Conservation Genetics* **15**, 177–185. doi:10.1007/s10592-013-0529-0
- Schwenk, L. E., Georges, A., Holleley, C. E., and Sarre, S. D. (2020). Climate change, sex reversal and lability of sex-determining systems. *Journal of Evolutionary Biology* **33**, 270–281. doi:10.1111/jeb.13587
- Schwenk, K. (1985). Occurrence, distribution and functional significance of taste buds in lizards. *Copeia* **1985**, 91–101. doi:10.2307/1444795
- Schwenk, K. (1993). Are geckos olfactory specialists? *Journal of Zoology* **229**, 289–302. doi:10.1111/j.1469-7998.1993.tb02637.x
- Schwenk, K. (1995). Of tongues and noses: chemoreception in lizards and snakes. *Trends in Ecology & Evolution* **10**, 7–12. doi:10.1016/S0169-5347(00)88953-3
- Scott, M. L., Whiting, M. J., Webb, J. K., and Shine, R. (2013). Chemosensory discrimination of social cues mediates space use in snakes, *Cryptophis nigriscens* (Elapidae). *Animal Behaviour* **85**, 1493–1500. doi:10.1016/j.anbehav.2013.04.003
- Scott, M. L., Llewelyn, J., Higgie, M., Hoskin, C. J., Pike, K., and Phillips, B. L. (2015). Chemoreception and mating behaviour of a tropical Australian skink. *Acta Ethologica* **18**, 283–293. doi:10.1007/s10211-015-0213-0
- Shine, R. (1988). Parental care in reptiles In 'Biology of the Reptilia. Vol. 16'. (Eds C. Gans, and R. Huey.) pp. 275–330. (Liss: New York.)
- Shine, R. (1995). A new hypothesis for the evolution of viviparity in reptiles. *American Naturalist* **145**, 809–823. doi:10.1086/285769
- Shine, R. (1999). Why is sex determined by nest temperature in many reptiles? *Trends in Ecology & Evolution* **14**, 186–189. doi:10.1016/S0169-5347(98)01575-4
- Shine, R. (2005). All at sea: aquatic life modifies mate-recognition modalities in sea snakes (*Emydocephalus annulatus*, Hydrophiidae). *Behavioral Ecology and Sociobiology* **57**, 591–598. doi:10.1007/s00265-004-0897-z
- Shine, R., Elphick, M. J., and Harlow, P. S. (1995). Sisters like it hot. *Nature* **378**, 451–452. doi:10.1038/378451a0
- Shine, R., Elphick, M. J., and Donnellan, S. (2002a). Co-occurrence of multiple, supposedly incompatible modes of sex determination in a lizard population. *Ecology Letters* **5**, 486–489. doi:10.1046/j.1461-0248.2002.00351.x
- Shine, R., Reed, R. N., Shetty, S., Lemaster, M., and Mason, R. T. (2002b). Reproductive isolating mechanisms between two sympatric sibling species of sea snakes. *Evolution* **56**, 1655–1662. doi:10.1111/j.0014-3820.2002.tb01477.x
- Siegel, D. S., Trauth, S. E., Rheubert, J. L., Rabe, B., Ruopp, B., Miralles, A., Murray, C. M., and Aldridge, R. D. (2014). Novel cloacal glands in snakes: the phylogenetic distribution of ventral urodaeal glands in Thamnophiini. *Herpetologica* **70**, 279–289. doi:10.1655/HERPETOLOGICA-D-13-00097

- Simmons, L. W. (2014). Sexual selection and genital evolution. *Austral Entomology* **53**, 1–17. doi:10.1111/aen.12053
- Sinervo, B., and Lively, C. M. (1996). The rock–paper–scissors game and the evolution of alternative male strategies. *Nature* **380**, 240–243. doi:10.1038/380240a0
- Sinervo, B., Méndez-de-la-Cruz, F., Miles, D. B., Heulin, B., Bastiaans, E., Villagrán-Santa Cruz, M., Lara-Resendiz, R., Martínez-Méndez, N., Calderón-Espinosa, M. L., Meza-Lázaro, R. N., Gadsden, H., Avila, L. J., Morando, M., De la Riva, I. J., Sepulveda, P. V., Rocha, C. F. D., Ibarjúengoytia, N., Puntriano, C. A., Massot, M., Lepetz, V., Oksanen, T. A., Chapple, D. G., Bauer, A. M., Branch, W. R., Clobert, J., and Sites, J. W. (2010). Erosion of lizard diversity by climate change and altered thermal niches. *Science* **328**, 894–899. doi:10.1126/science.1184695
- Sinn, D. L., While, G. M., and Wapstra, E. (2008). Maternal care in a social lizard: links between female aggression and offspring fitness. *Animal Behaviour* **76**, 1249–1257. doi:10.1016/j.anbehav.2008.06.009
- Skeels, A., Esquerré, D., and Cardillo, M. (2020). Alternative pathways to diversity across ecologically distinct lizard radiations. *Global Ecology and Biogeography* **29**, 454–469. doi:10.1111/geb.13044
- Smith, R. M. (1987). Helical burrow casts of therapsid origin from the Beaufort Group (Permian) of South Africa. *Palaeogeography, Palaeoclimatology, Palaeoecology* **60**, 155–169. doi:10.1016/0031-0182(87)90030-7
- Smith, S. A., and Shine, R. (1997). Intraspecific variation in reproductive mode within the scincid lizard *Saiphos equalis*. *Australian Journal of Zoology* **45**, 435–445. doi:10.1071/ZO97023
- Somaweera, R., Nifong, J., Rosenblatt, A., Brien, M. L., Combrink, X., Elsey, R. M., Grigg, G., Magnusson, W. E., Mazzotti, F. J., and Percy, A. (2020). The ecological importance of crocodylians: towards evidence-based justification for their conservation. *Biological Reviews of the Cambridge Philosophical Society* **95**, 936–959. doi:10.1111/brv.12594
- Somma, L. A. (2003). 'Parental Behavior in Lepidosaurian and Testudinian Reptiles: a Literature Survey.' (Krieger Publishing Company: Malabar.)
- Stapley, J., and Keogh, J. S. (2005). Behavioral syndromes influence mating systems: floater pairs of a lizard have heavier offspring. *Behavioral Ecology* **16**, 514–520. doi:10.1093/beheco/ari019
- Stapley, J., Hayes, C. M., and Scott Keogh, J. (2003). Population genetic differentiation and multiple paternity determined by novel microsatellite markers from the mountain log skink (*Pseudemoia entrecasteauxii*). *Molecular Ecology Notes* **3**, 291–293. doi:10.1046/j.1471-8286.2003.00429.x
- Steele, A. L., Wibbels, T., and Warner, D. A. (2018). Revisiting the first report of temperature-dependent sex determination in a vertebrate, the African redhead agama. *Journal of Zoology* **306**, 16–22. doi:10.1111/jzo.12560
- Stenning, M. J. (1996). Hatching asynchrony, brood reduction and other rapidly reproducing hypotheses. *Trends in Ecology & Evolution* **11**, 243–246. doi:10.1016/0169-5347(96)10030-6
- Stewart, J. R. (1989). Facultative placentotrophy and the evolution of squamate placentation: quality of eggs and neonates in *Virginia striatula*. *American Naturalist* **133**, 111–137. doi:10.1086/284904
- Stewart, J. R., and Thompson, M. B. (2004). Placental ontogeny of the Tasmanian scincid lizard, *Niveoscincus ocellatus* (Reptilia: Squamata). *Journal of Morphology* **259**, 214–237. doi:10.1002/jmor.10179
- Stienen, E. W., and Brenninkmeijer, A. (2006). Effect of brood size and hatching sequence on pre fledging mortality of sandwich terns: why lay two eggs? *Journal of Ornithology* **147**, 520–530. doi:10.1007/s10336-006-0075-3
- Stoleson, S. H., and Beissinger, S. R. (1995). Hatching asynchrony and the onset of incubation in birds, revisited. *Current Ornithology* **12**, 191–270. doi:10.1007/978-1-4615-1835-8_6
- Stow, A., and Sunnucks, P. (2004). High mate and site fidelity in Cunningham's skinks (*Egernia cunninghami*) in natural and fragmented habitat. *Molecular Ecology* **13**, 419–430. doi:10.1046/j.1365-294X.2003.02061.x
- Stow, A. J., Sunnucks, P., Briscoe, D. A., and Gardner, M. G. (2001). The impact of habitat fragmentation on dispersal of Cunningham's skink (*Egernia cunninghami*): evidence from allelic and genotypic analyses of microsatellites. *Molecular Ecology* **10**, 867–878. doi:10.1046/j.1365-294X.2001.01253.x
- Stuart-Fox, D., Aulsebrook, A., Rankin, K., Dong, C., and McLean, C. (2021). Convergence and divergence in lizard colour polymorphisms. *Biological Reviews of the Cambridge Philosophical Society* **96**, 289–309. doi:10.1111/brv.12656
- Sullivan, B. K., and Kwiatkowski, M. A. (2007). Courtship displays in anurans and lizards: theoretical and empirical contributions to our understanding of costs and selection on males due to female choice. *Functional Ecology* **21**, 666–675. doi:10.1111/j.1365-2435.2007.01244.x
- Sun, B. J., Li, S. R., Xu, X. F., Zhao, W. G., Luo, L. G., Ji, X., and Du, W. G. (2013). Different mechanisms lead to convergence of reproductive strategies in two lacertid lizards (*Takydromus wolteri* and *Eremias argus*). *Oecologia* **172**, 645–652. doi:10.1007/s00442-012-2524-4
- Svensson, E. I., Willink, B., Duryea, M. C., and Lancaster, L. T. (2020). Temperature drives pre-reproductive selection and shapes the biogeography of a female polymorphism. *Ecology Letters* **23**, 149–159. doi:10.1111/ele.13417
- Swain, R., and Jones, S. M. (1997). Maternal–fetal transfer of 3H-labelled leucine in the viviparous lizard *Niveoscincus metallicus* (Scincidae: Lygosominae). *The Journal of Experimental Zoology* **277**, 139–145. doi:10.1002/(SICI)1097-010X(19970201)277:2<139::AID-JEZ5>3.0.CO;2-Q
- Swain, R., and Jones, S. M. (2000). Facultative placentotrophy: half-way house or strategic solution? *Comparative Biochemistry and Physiology. Part A, Molecular & Integrative Physiology* **127**, 441–451. doi:10.1016/S1095-6433(00)00275-0
- Taylor, M. L., Price, T. A. R., and Wedell, N. (2014). Polyandry in nature: a global analysis. *Trends in Ecology & Evolution* **29**, 376–383. doi:10.1016/j.tree.2014.04.005
- Teasdale, L. C., Stevens, M., and Stuart-Fox, D. (2013). Discrete colour polymorphism in the tawny dragon lizard (*Ctenophorus decresii*) and differences in signal conspicuousness among morphs. *Journal of Evolutionary Biology* **26**, 1035–1046. doi:10.1111/jeb.12115
- Telemeco, R. S. (2015). Sex determination in southern alligator lizards (*Elgaria multicarinata*; Anguillidae). *Herpetologica* **71**, 8–11. doi:10.1655/Herpetologica-D-14-00033
- Thompson, J. (1977). The transfer of amino acids across the placenta of a viviparous lizard, *Sphenomorphus quoyii*. *Theriogenology* **8**, 158. doi:10.1016/0093-691X(77)90120-0
- Thompson, J. (1981). A study of the sources of nutrients for embryonic development in a viviparous lizard, *Sphenomorphus quoyii*. *Comparative Biochemistry and Physiology. Part A, Physiology* **70**, 509–518. doi:10.1016/0300-9629(81)92562-7
- Thompson, M. B., and Speake, B. K. (2006). A review of the evolution of viviparity in lizards: structure, function and physiology of the placenta. *Journal of Comparative Physiology. B, Biochemical, Systemic, and Environmental Physiology* **176**, 179–189. doi:10.1007/s00360-005-0048-5
- Thompson, M. B., Stewart, J. R., and Speake, B. K. (2000). Comparison of nutrient transport across the placenta of lizards differing in placental complexity. *Comparative Biochemistry and Physiology. Part A, Molecular & Integrative Physiology* **127**, 469–479. doi:10.1016/S1095-6433(00)00277-4

- Thornhill, R. (1983). Cryptic female choice and its implications in the scorpionfly *Harpobittacus nigriceps*. *American Naturalist* **122**, 765–788. doi:10.1086/284170
- Tinkle, D. W., and Gibbons, J. W. (1977). 'The Distribution and Evolution of Viviparity in Reptiles.' (Museum of Zoology, University of Michigan: Ann Arbor, MI.)
- Tinkle, D. W., Wilbur, H. M., and Tilley, S. G. (1970). Evolutionary strategies in lizard reproduction. *Evolution* **24**, 55–74. doi:10.1111/j.1558-5646.1970.tb01740.x
- Tokunaga, S. (1985). Temperature-dependent sex determination in *Gekko japonicus* (Gekkonidae, Reptilia). *Development, Growth & Differentiation* **27**, 117–120. doi:10.1111/j.1440-169X.1985.00117.x
- Uetz, P. (2010). The original descriptions of reptiles. *Zootaxa* **2334**, 59–68. doi:10.11646/zootaxa.2334.1.3
- Uller, T., and Olsson, M. (2005). Multiple copulations in natural populations of lizards: evidence for the fertility assurance hypothesis. *Behaviour* **142**, 45–56. doi:10.1163/1568539053627749
- Uller, T., and Olsson, M. (2006). No seasonal sex-ratio shift despite sex-specific fitness returns of hatching date in a lizard with genotypic sex determination. *Evolution* **60**, 2131–2136. doi:10.1111/j.0014-3820.2006.tb01850.x
- Uller, T., and Olsson, M. (2008). Multiple paternity in reptiles: patterns and processes. *Molecular Ecology* **17**, 2566–2580. doi:10.1111/j.1365-294X.2008.03772.x
- Uller, T., Pen, I., Wapstra, E., Beukeboom, L. W., and Komdeur, J. (2007). The evolution of sex ratios and sex-determining systems. *Trends in Ecology & Evolution* **22**, 292–297. doi:10.1016/j.tree.2007.03.008
- Uller, T., Stuart-Fox, D., and Olsson, M. (2010). Evolution of primary sexual characters in reptiles. In 'Evolution of Primary Sexual Characters in Animals'. (Eds J. L. Leonard, and A. Córdoba-Aguilar.) pp. 425–452. (Oxford University Press: Oxford.)
- Valdecantos, S., and Lobo, F. (2015). First report of hemiclitoris in females of South American liolaemid lizards. *Journal of Herpetology* **49**, 291–294. doi:10.1670/13-124
- Valenzuela, N., Litterman, R., Neuwald, J. L., Mizoguchi, B., Iverson, J. B., Riley, J. L., and Litzgus, J. D. (2019). Extreme thermal fluctuations from climate change unexpectedly accelerate demographic collapse of vertebrates with temperature-dependent sex determination. *Scientific Reports* **9**, 4254. doi:10.1038/s41598-019-40597-4
- Van Dyke, J. U., Griffith, O. W., and Thompson, M. B. (2014). High food abundance permits the evolution of placentotrophy: evidence from a placental lizard, *Pseudemoia entrecasteauxii*. *American Naturalist* **184**, 198–210. doi:10.1086/677138
- Van Dyke, J. U., Lindsay, L. A., Murphy, C. R., and Thompson, M. B. (2015). Carbonic anhydrase II is found in the placenta of a viviparous, matrotrophic lizard and likely facilitates embryo–maternal CO₂ transport. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **324**, 636–646. doi:10.1002/jez.b.22621
- van Wyk, J. H., and Mouton, P. F. N. (1992). Glandular epidermal structures in cordylid lizards. *Amphibia-Reptilia* **13**, 1–12. doi:10.1163/156853892X00175
- Vicoso, B., Kaiser, V. B., and Bachtrog, D. (2013). Sex-biased gene expression at homomorphic sex chromosomes in emus and its implication for sex chromosome evolution. *Proceedings of the National Academy of Sciences of the United States of America* **110**, 6453–6458. doi:10.1073/pnas.1217027110
- Viets, B. E., Tousignant, A., Ewert, M. A., Nelson, C. E., and Crews, D. (1993). Temperature-dependent sex determination in the leopard gecko, *Eublepharis macularius*. *The Journal of Experimental Zoology* **265**, 679–683. doi:10.1002/jez.1402650610
- Vitt, L. J., and Pianka, E. R. (1994). 'Lizard Ecology: Historical and Experimental Perspectives.' (Princeton University Press: Princeton, NJ.)
- Wade, J. S. (2011). Relationships among hormones, brain and motivated behaviors in lizards. *Hormones and Behavior* **59**, 637–644. doi:10.1016/j.yhbeh.2010.08.014
- Wade, J. S., Huang, J. M., and Crews, D. (1993). Hormonal control of sex differences in the brain, behavior and accessory sex structures of whiptail lizards (*Cnemidophorus* species). *Journal of Neuroendocrinology* **5**, 81–93. doi:10.1111/j.1365-2826.1993.tb00366.x
- Wang, C., Tang, X., Xin, Y., Yue, F., Yan, X., Liu, B., An, B., Wang, X., and Chen, Q. (2015). Identification of sex chromosomes by means of comparative genomic hybridization in a lizard, *Eremias multiocellata*. *Zoological Science* **32**, 151–156. doi:10.2108/zs130246
- Wapstra, E., and Olsson, M. (2014). The evolution of polyandry and patterns of multiple paternity in lizards. In 'Reproductive Biology and Phylogeny of Lizards and Tuatara'. (Eds J. L. Rheubert, D. S. Siegel, and S. E. Trauth.) pp. 576–601. (CRC Press: Boca Raton, FL, USA.)
- Wapstra, E., and Swain, R. (2001). Geographic and annual variation in life-history traits in a temperate zone Australian skink. *Journal of Herpetology* **35**, 194–203. doi:10.2307/1566108
- Wapstra, E., and Warner, D. A. (2010). Sex allocation and sex determination in squamate reptiles. *Sexual Development* **4**, 110–118. doi:10.1159/000272459
- Wapstra, E., Swain, R., Jones, S. M., and O'Reilly, J. (1999). Geographic and annual variation in reproductive cycles in the Tasmanian spotted snow skink, *Niveoscincus ocellatus* (Squamata: Scincidae). *Australian Journal of Zoology* **47**, 539–550. doi:10.1071/ZO99038
- Wapstra, E., Swain, R., and O'Reilly, J. M. (2001). Geographic variation in age and size at maturity in a small Australian viviparous skink. *Copeia* **2001**, 646–655. doi:10.1643/0045-8511(2001)001[0646:GVIAAS]2.0.CO;2
- Wapstra, E., Olsson, M., Shine, R., Edwards, A., Swain, R., and Joss, J. M. (2004). Maternal basking behaviour determines offspring sex in a viviparous reptile. *Biology Letters* **271**, S230–S232.
- Wapstra, E., Uller, T., Sinn, D. L., Olsson, M., Mazurek, K., Joss, J., and Shine, R. (2009). Climate effects on offspring sex ratio in a viviparous lizard. *Journal of Animal Ecology* **78**, 84–90. doi:10.1111/j.1365-2656.2008.01470.x
- Warner, D. A., and Shine, R. (2005). The adaptive significance of temperature-dependent sex determination: experimental tests with a short-lived lizard. *Evolution* **59**, 2209–2221. doi:10.1111/j.0014-3820.2005.tb00929.x
- Warner, D. A., and Shine, R. (2007). Fitness of juvenile lizards depends on seasonal timing of hatching, not offspring body size. *Oecologia* **154**, 65–73. doi:10.1007/s00442-007-0809-9
- Warner, D. A., and Shine, R. (2008). The adaptive significance of temperature-dependent sex determination in a reptile. *Nature* **451**, 566–568. doi:10.1038/nature06519
- Warner, D. A., Woo, K. L., Van Dyk, D. A., Evans, C. S., and Shine, R. (2010). Egg incubation temperature affects male reproductive success but not display behaviors in lizards. *Behavioral Ecology and Sociobiology* **64**, 803–813. doi:10.1007/s00265-009-0897-0
- Watson, G., Green, D., and Watson, J. (2019). Observations supporting parental care by a viviparous reptile: aggressive behaviour against predators demonstrated by Cunningham's skinks. *Australian Journal of Zoology* **67**, 180–183. doi:10.1071/ZO20024
- Weber, C., and Capel, B. (2018). Sex reversal. *Current Biology* **28**, R1234–R1236. doi:10.1016/j.cub.2018.09.043
- Wedekind, C. (2017). Demographic and genetic consequences of disturbed sex determination. *Philosophical Transactions of the Royal Society B: Biological Sciences* **372**, 20160326. doi:10.1098/rstb.2016.0326
- Weekes, H. C. (1935). A review of placentation among reptiles with particular regard to the function and evolution of the placenta. *Proceedings of the Zoological Society of London* **105**, 625–645. doi:10.1111/j.1096-3642.1935.tb01686.x

- Weldon, P. J., Flachsbarth, B., and Schulz, S. (2008). Natural products from the integument of non-avian reptiles. *Natural Product Reports* **25**, 738–756. doi:10.1039/b509854h
- While, G. M., and Wapstra, E. (2009). Effects of basking opportunity on birthing asynchrony in a viviparous lizard. *Animal Behaviour* **77**, 1465–1470. doi:10.1016/j.anbehav.2009.03.001
- While, G. M., Jones, S. M., and Wapstra, E. (2007). Birthing asynchrony is not a consequence of asynchronous offspring development in a non-avian vertebrate, the Australian skink *Egernia whitii*. *Functional Ecology* **21**, 513–519. doi:10.1111/j.1365-2435.2007.01272.x
- While, G. M., Uller, T., and Wapstra, E. (2009a). Offspring performance and the adaptive benefits of prolonged pregnancy: experimental tests in a viviparous lizard. *Functional Ecology* **23**, 818–825. doi:10.1111/j.1365-2435.2009.01544.x
- While, G. M., Uller, T., and Wapstra, E. (2009b). Within-population variation in social strategies characterize the social and mating system of an Australian lizard, *Egernia whitii*. *Austral Ecology* **34**, 938–949. doi:10.1111/j.1442-9993.2009.02002.x
- While, G. M., Uller, T., and Wapstra, E. (2011). Variation in social organization influences the opportunity for sexual selection in a social lizard. *Molecular Ecology* **20**, 844–852. doi:10.1111/j.1365-294X.2010.04976.x
- While, G. M., Uller, T., Bordogna, G., and Wapstra, E. (2014). Promiscuity resolves constraints on social mate choice imposed by population viscosity. *Molecular Ecology* **23**, 721–732. doi:10.1111/mec.12618
- While, G. M., Gardner, M., Chapple, D. G., and Whiting, M. J. (2019). Stable social grouping in lizards. In 'Evolutionary and Mechanistic Perspectives'. (Eds A. Russel, and V. Bells.) pp. 321–343. (CRC Press: New Hampshire, USA.)
- Whiteley, S. L., Holleley, C. E., Ruscoe, W. A., Castelli, M., Whitehead, D. L., Lei, J., Georges, A., and Weisbecker, V. (2017). Sex determination mode does not affect body or genital development of the central bearded dragon (*Pogona vitticeps*). *EvoDevo* **8**, 25. doi:10.1186/s13227-017-0087-5
- Whiteley, S. L., Weisbecker, V., Georges, A., Gauthier, A. R. G., Whitehead, D. L., and Holleley, C. E. (2018). Developmental asynchrony and antagonism of sex determination pathways in a lizard with temperature-induced sex reversal. *Scientific Reports* **8**, 14892. doi:10.1038/s41598-018-33170-y
- Whiteley, S. L., Castelli, M. A., Dissanayake, D. S. B., Holleley, C. E., and Georges, A. (2021a). Temperature induced sex reversal in reptiles: prevalence, discovery, and evolutionary implications. *Sexual Development*. doi:10.1159/000515687
- Whiteley, S. L., Georges, A., Weisbecker, V., Schwanz, L. E., and Holleley, C. E. (2021b). Ovaries suggest cryptic genetic influence in a reptile model for temperature-dependent sex determination. *Proceedings of the Royal Society B: Biological Sciences* **288**, 20202819. doi:10.1098/rspb.2020.2819
- Whiteley, S. L., Holleley, C. E., Wagner, S., Blackburn, J., Deveson, I. W., Graves, J. A. M., and Georges, A. (2021c). Two transcriptionally distinct pathways drive female development in a reptile with both genetic and temperature dependent sex determination. *PLOS Genetics* **17**, e1009465. doi:10.1371/journal.pgen.1009465
- Whiting, M., and While, G. M. (2017). Sociality in lizards. In 'Comparative Social Evolution'. (Eds D. R. Rubenstein, and P. Abbot.) pp. 390–426. (Cambridge University Press: Cambridge.)
- Whittington, C. M., Grau, G. E., Murphy, C. R., and Thompson, M. B. (2015). Unusual angiogenic factor plays a role in lizard pregnancy but is not unique to viviparity. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **324**, 152–158. doi:10.1002/jez.b.22615
- Wiggins, J. M., Santoyo-Brito, E., Scales, J. B., and Fox, S. F. (2020). Gene dose indicates presence of sex chromosomes in collared lizards (*Crotaphytus collaris*), a species with temperature-influenced sex determination. *Herpetologica* **76**, 27–30. doi:10.1655/Herpetologica-D-19-00036
- Wilson, S., and Swan, G. (2021). 'A Complete Guide to Reptiles of Australia.' 6th edn. (Reed New Holland: Wahroonga, NSW.)
- Wu, Q., Thompson, M. B., and Murphy, C. R. (2011). Changing distribution of cadherins during gestation in the uterine epithelium of lizards. *Journal of Experimental Zoology. Part B, Molecular and Developmental Evolution* **316B**, 440–450. doi:10.1002/jez.b.21419
- Wyatt, T. D. (2010). Pheromones and signature mixtures: defining species-wide signals and variable cues for identity in both invertebrates and vertebrates. *Journal of Comparative Physiology. A, Neuroethology, Sensory, Neural, and Behavioral Physiology* **196**, 685–700. doi:10.1007/s00359-010-0564-y
- Yampolsky, L. Y., and Scheiner, S. M. (1996). Why larger offspring at lower temperatures? A demographic approach. *American Naturalist* **147**, 86–100. doi:10.1086/285841
- Yewers, M. S. C., Pryke, S., and Stuart-Fox, D. (2016). Behavioural differences across contexts may indicate morph-specific strategies in the lizard *Ctenophorus decresii*. *Animal Behaviour* **111**, 329–339. doi:10.1016/j.anbehav.2015.10.029
- Yewers, M. S. C., Jessop, T. S., and Stuart-Fox, D. (2017). Endocrine differences among colour morphs in a lizard with alternative behavioural strategies. *Hormones and Behavior* **93**, 118–127. doi:10.1016/j.yhbeh.2017.05.001
- Yntema, C. L. (1976). Effects of incubation temperatures on sexual differentiation in the turtle, *Chelydra serpentina*. *Journal of Morphology* **150**, 453–461. doi:10.1002/jmor.1051500212
- York, J. R., and Baird, T. A. (2019). Sexual selection on female collared lizards favours offspring production with multiple males. *Animal Behaviour* **147**, 17–23. doi:10.1016/j.anbehav.2018.10.020
- Young, R., Ferkin, M., Ockendon-Powell, N., Orr, V., Phelps, S., Pogány, Á., Richards-Zawacki, C., Summers, K., Székely, T., Trainor, B., Zachar, G., O'Connell, L., and Hofmann, H. (2019). Conserved transcriptomic profiles underpin monogamy across vertebrates. *Proceedings of the National Academy of Sciences of the United States of America* **116**, 1331–1336. doi:10.1073/pnas.1813775116
- Zamudio, K. R., and Sinervo, B. (2000). Polygyny, mate-guarding, and posthumous fertilization as alternative male mating strategies. *Proceedings of the National Academy of Sciences of the United States of America* **97**, 14427–14432. doi:10.1073/pnas.011544998
- Ziegler, T., Gaulke, M., and Böhme, W. (2005). Genital morphology and systematics of *Varanus mabitang* Gaulke & Curio, 2001 (Squamata: Varanidae). *Current Herpetology* **24**, 13–17. doi:10.3105/1345-5834(2005)24[13:GMASOV]2.0.CO;2
- Ziegler, T., Schmitz, A., Koch, A., and Böhme, W. (2007). A review of the subgenus *Euprepisaurus* of *Varanus* (Squamata: Varanidae): morphological and molecular phylogeny, distribution and zoogeography, with an identification key for the members of the *V. indicus* and the *V. prasinus* species groups. *Zootaxa* **1472**, 1. doi:10.11646/zootaxa.1472.1.1
- Zozaya, S. M., Higgie, M., Moritz, C., and Hoskin, C. J. (2019). Are pheromones key to unlocking cryptic lizard diversity? *American Naturalist* **194**, 168–182. doi:10.1086/704059

Handling Editor: Paul Cooper