

Pollen morphology of tribes Aptosimeae and Myoporeae supports the phylogenetic pattern in early-branching Scrophulariaceae revealed by molecular studies

Authors: Mosyakin, Sergei L., and Tsymbalyuk, Zoya M.

Source: Willdenowia, 45(2): 209-222

Published By: Botanic Garden and Botanical Museum Berlin (BGBM)

URL: https://doi.org/10.3372/wi.45.45207

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

SERGEI L. MOSYAKIN^{1*} & ZOYA M. TSYMBALYUK¹

Pollen morphology of tribes *Aptosimeae* and *Myoporeae* supports the phylogenetic pattern in early-branching *Scrophulariaceae* revealed by molecular studies

Abstract

Mosyakin S. L. & Tsymbalyuk Z. M.: Pollen morphology of tribes *Aptosimeae* and *Myoporeae* supports the phylogenetic pattern in early-branching *Scrophulariaceae* revealed by molecular studies. – Willdenowia 45: 209–222. 2015. – Version of record first published online on 15 July 2015 ahead of inclusion in August 2015 issue; ISSN 1868-6397; © 2015 BGBM Berlin.

DOI: http://dx.doi.org/10.3372/wi.45.45207

Pollen morphology of six species representing three genera of tribe Aptosimeae and six species representing two genera of tribe Myoporeae (Scrophulariaceae) is described and illustrated using light microscopy (LM) and scanning electron microscopy (SEM). Pollen grains in Aptosimeae are 3-syncolporate, prolate, sometimes oblate-spheroidal in shape; mainly medium-sized, occasionally in some taxa small; exine sculpture striate, rarely with microperforations and small granules. Pollen grains in Myoporeae are 3-colpate-diorate, spheroidal, rarely oblate-spheroidal or prolate in shape; mainly medium-sized, occasionally in some taxa small; exine sculpture foveolate, microreticulate, rugulate-microperforate, rugulate, sometimes with small granules. Two major pollen types, 3-syncolporate (Aptosimeae) and 3-colpate-diorate (Myoporeae) are recognized by aperture types. Within these pollen types, eight subtypes are distinguished (three in Aptosimeae, five in Myoporeae) based on pollen size, exine sculpture and details of colpi and endoapertures. Our analysis of palynomorphological data (both published earlier and newly reported here) outlines the same main clades as those revealed by molecular phylogenetic studies, corresponding to Aptosimeae, Leucophylleae+Myoporeae and Androya, which differ by their pollen aperture types: 3-syncolporate, 3-colpate-diorate, and 3-colporate, respectively. There are also some differences in exine sculpture patterns. Thus, palynomorphological data are mainly consistent with recent results of molecular phylogenetic studies. Considerations on ancestral pollen character states in early-branching Scrophulariaceae are provided. It is assumed that the colporate type was probably ancestral in *Scrophulariaceae*; however, the ancestral status of the colpate type cannot be excluded as well.

Additional key words: exine sculpture, systematics, phylogeny, palynomorphology

Introduction

Current opinions on taxonomy and phylogeny of early-branching clades of *Scrophulariaceae*

Judging from the phylogenetic pattern revealed by recent molecular phylogenetic studies (Olmsted & al. 2001; Oxelman & al. 2005; Tank & al. 2006), the first early-branching lineage of the re-circumscribed *Scrophulariaceae* Juss. (s.str.) is represented by the tribe *Hemimerideae*, which is sister to the clade containing all other members of *Scrophulariaceae*. That large clade, in turn, comprises two subclades, one containing represent-

atives of three tribes, *Aptosimeae*, *Leucophylleae* and *Myoporeae* in their current updated circumscriptions, and another grouping the remaining representatives of *Scrophulariaceae*.

The tribe *Aptosimeae*, as outlined in recent studies (Olmsted & Reeves 1995; Olmsted & al. 2001; Fischer 2004; Oxelman & al. 2005; Tank & al. 2006; Takhtajan 2009), is a group represented by three genera (*Anticharis* Endl., *Aptosimum* Burch. ex Benth., and *Peliostomum* E. Mey. ex Benth.) and c. 40 species geographically restricted to tropical and southern Africa and southern Asia from the Arabian Peninsula to Malesia. Two genera, *Ap*-

¹ M. G. Kholodny Institute of Botany, National Academy of Sciences of Ukraine, 2 Tereshchenkivska Street, Kyiv (Kiev), 01601 Ukraine; *e-mail: s_mosyakin@hotmail.com (author for correspondence), palynology@ukr.net

tosimum with c. 20 species and Peliostomum with seven species, occur only in tropical and southern Africa, and representatives of Anticharis (with c. 14 species) also occur in southern Asia. South African taxa of Aptosimeae have a distinctive pollination syndrome. These species are reported to be pollinated by several taxa of a specialized group of pollen wasps (Hymenoptera, tribe Masarini), which are behaviourally distinct from all other aculeate wasps, approaching in that respect the true bees. Widespread association of Aptosimeae with these pollen wasps and floral specialization of species of wasp genera Celonites and Masarina have been established (Gess & Gess 2010). The tribe is also of special interest because one of its genera, Anticharis, contains the only known group of species of Scrophulariaceae having C4 photosynthesis (Sage & al. 2011; Khosravesh & al. 2012), which occurs very rarely in Lamiales (also registered in Blepharis Juss. in the Acanthaceae).

The Aptosimeae clade is sister to the clade Androya+Leucophylleae+Myoporeae (Oxelman & al. 2005; Tank & al. 2006). In the working version of the system of Lamiales (Olmstead 2012), Aptosimeae also contain Stemodiopsis Engl. (perhaps erroneously). Oxelman & al. (2005) placed Stemodiopsis in Lindernieae, "[a] novel strongly supported clade of taxa earlier assigned to Scrophulariaceae..." with "unclear relationships to the rest of Lamiales." Later, Rahmanzadeh & al. (2005) placed this genus in Gratiolaceae (tribe Stemodieae, validly published in Reveal 2012). In publications that followed (Tank & al. 2006; Schäferhoff & al. 2010) Stemodiopsis was placed in Linderniaceae. The newest molecular phylogenetic data by Fisher & al. (2013) confirmed the close relationships of Stemodiopsis with other genera of Linderniaceae.

The Myoporeae clade in a strict sense currently includes the genera Eremophila R. Br. (more than 200 species, Australia, mostly arid regions in the west), Myoporum Sol. ex G. Forst. (c. 30 species, mostly Australian, with some extending to southeastern Asia, islands of the Pacific and Indian oceans), Bontia L. (monotypic, the Caribbean area), Pentacoelium Zucc. (monotypic, from southern Japan through the Ryukyu Islands and Taiwan to southeastern mainland China), and three recently described genera endemic to a small southwestern region in Western Australia: Diocirea Chinnock (four species), and monotypic Calamphoreus Chinnock and Glycocystis Chinnock (Chinnock 2007; Olmstead 2012). In his detailed taxonomic revision of the group, Chinnock (2007) preferred to treat it at the family rank (as Myoporaceae) but admitted that combining Myoporaceae and Leucophylleae is a reasonable option and that the combined "Myoporaceous clade" is "...possibly best treated at subfamily level within the Scrophulariaceae s.str." (Chinnock 2007: 84). Considering generic relationships, Chinnock (l.c.) indicated that "Diocirea and Calamphoreus can be considered derivatives of Eremophila" and assumed that Glycosystis is closely related to one of the endemic Australian sections of *Myoporum*.

Androya H. Perrier, the monotypic genus represented by A. decaryi H. Perrier, is sometimes placed in Myoporeae or treated as a genus unassigned to any tribe (see Olmstead 2012). In earlier versions of his system, Takhtajan (1987, 1997) placed Androya in Buddlejaceae. Takhtajan (2009) later moved it to Myoporaceae, together with Bontia, Eremophila (incl. Pholidia R. Br. and Stenochilus R. Br.) and Myoporum.

The Leucophylleae clade contains Capraria L. (four species), Eremogeton Standl. & L. O. Williams (monotypic) and Leucophyllum Bonpl. (c. 16 species, including Faxonanthus Greenm.; see Henrickson & Flyr 1985). Geographically this group is restricted to Central America and the Caribbean, southernmost regions of North America, and some parts of South America (see the map in Gándara & Sosa 2013). In all recent versions of Takhtajan's system (Takhtajan 1987, 1997, 2009), Leucophylleae contained just two genera, Eremogeton and Leucophyllum, while Capraria was placed in the tribe Gratioleae of Scrophulariaceae s.l. Rahmanzadeh & al. (2005) included Capraria in Gratiolaceae tribe Gratioleae, which contradicts the placement suggested by Oxelman & al. (2005) and Gándara & Sosa (2013). The generic status of Faxonanthus and the phylogenetic relationships of Eremogeton remained unresolved until recently, due to the lack of molecular data (Oxelman & al. 2005). Gándara & Sosa (2013) filled that gap and included in their molecular study representatives of all three genera, paying special attention to Leucophyllum. They demonstrated that this genus in its traditional circumscription is not monophyletic, since both Capraria and Eremogeton are rooted within it. The alternative taxonomic options they suggested are (1) to submerge all three entities in one genus or (2) to split Leucophyllum into three genera, preserving the generic rank for Capraria and Eremogeton.

Molecular studies (Oxelman & al. 2005; Tank & al. 2006; Gándara & Sosa 2013) convincingly revealed that the sister clades corresponding to re-circumscribed tribes *Leucophylleae* and *Myoporeae* are together, in turn, sister to *Androya*. Reveal (2012) united tribes *Aptosimeae*, *Leucophylleae* and *Myoporeae* in *Scrophulariaceae* subfam. *Myoporoideae*.

Published data on pollen morphology

Scarce information is available on pollen grains of representatives of *Aptosimeae*. As far as we know, only pollen grains of *Aptosimum depressum* Burch. were studied by Erdtman (1952) using light microscopy. The situation with data on pollen morphology of *Myoporeae* and *Leucophylleae* is better. In particular, Erdtman (1952, light microscopy only) studied pollen of *Bontia daphnoides* L., *Myoporum laetum* G. Forst., *M. sandwicense* A. Gray, *Pholidia adenotricha* F. Muell. ex Benth. (now *Eremophila adenotricha* (F. Muell. ex Benth.) F. Muell.), *P. alternifolia* (R. Br.) Wettst. (now *E. alternifolia* R. Br.) and *Capraria biflora* L. and mentioned some affinity of pollen

grains of C. biflora to those in some Myoporaceae (recognized then as a separate family). Punt & Leenhouts (1967; light microscopy only) studied pollen grains of the monotypic genus Androya restricted to Madagascar. Minkin & Eshbaugh (1989) provided information on pollen grains of Leucophyllum texanum Benth. (light and scanning electron microscopy). Niezgoda & Tomb (1975) studied pollen grains of 14 species of Scrophulariaceae (tribe Leucophylleae and C. biflora) and 14 species of Myoporaceae using light, scanning and transmission electron microscopy. The most extensive palynomorphological study of Myoporeae (Australian taxa) was recently carried out by Chinnock (2007), who studied pollen grains of 28 species, including four species of *Diocirea*, two species of the monotypic Calamphoreus and Glycocystis, 19 species of Eremophila and three species of Myoporum. Pollen grains of Bontia daphnoides and Pentacoelium bontioides Siebold & Zucc. were also studied for comparison.

Considering new data on phylogenetic relationships of early-branching (basal) lineages of *Scrophulariaceae* and the virtual absence of information on pollen grains of *Aptosimeae*, we decided to partially fill this gap. In order to obtain additional data for comparison, we also studied some species of *Myoporeae*. The main objective of the study was to analyse and compare data on pollen morphology of early-branching lineages of *Scrophulariaceae* with molecular phylogenetic evidence for revealing possible ways of morphological evolution and hypothetical ancestral character states of pollen in this family.

Material and methods

Pollen from six species belonging to all three genera of *Aptosimeae* was sampled in the herbarium of the Missouri Botanical Garden, St Louis, Missouri, U.S.A. (MO). Pollen from six species representing two genera (*Eremophila* and *Myoporum*) of *Myoporeae* was sampled in the herbarium of the M.G. Kholodny Institute of Botany, the National Academy of Sciences of Ukraine, Kyiv (Kiev), Ukraine (KW); herbarium codes according to Thiers (2015+). Investigated specimens are cited according to the label information.

Pollen morphology was studied using light microscopy (LM) and scanning electron microscopy (SEM). For light microscopy studies (LM, Biolar, ×700), the pollen was acetolysed following Erdtman (1952). For size determinations, 20 measurements were taken along the polar and equatorial axes for each species.

For scanning electron microscopy (SEM, JSM-6060LA), pollen grains were treated with 96% ethanol, then the samples were sputter-coated with gold at the Center of Electron Microscopy of the M.G. Kholodny Institute of Botany (National Academy of Sciences of Ukraine, Kyiv). The micrographs were minimally edited with Adobe Photoshop 6.0 to enhance the images. Terminology used in descriptions of pollen grains mainly

follows the glossaries by Punt & al. (1994) and Tokarev (2002), with minor adjustments.

Results

General description of pollen grains

Aptosimeae

Pollen grains in monads, radially symmetrical, isopolar, 3-syncolporate, prolate, sometimes oblate-spheroidal; mainly medium-sized, occasionally in some taxa small: P[olar axis]=25.3–45.2 μ m, E[quatorial diameter]=15.9–33.2 μ m. Outline in polar view 3-lobate, in equatorial view elliptic, sometimes circular. Colpi 2.4–6.6 μ m wide, fused on apocolpia, with distinct, strict and slightly thickened margins; colpus membrane smooth.

Endoapertures mainly distinct, rarely indistinct, lalongate or sometimes lolongate, elliptic or circular, $4-7.9 \mu m \log, 2.7-9.3 \mu m$ wide, sometimes covered by margins of colpi.

Exine thin, 0.7–2 µm thick. Exine layers invisible, sometimes tectum two times thinner than lower layers. Exine sculpture striate, rarely with microperforations and small granules; apertures membrane psilate, granulate and granulate-verrucate.

Myoporeae

Pollen grains in monads, radially symmetrical, isopolar, 3-colpate-diorate, spheroidal, rarely oblate-spheroidal or prolate; mainly medium-sized, occasionally in some taxa small: $P=17.3-35.9~\mu m$, $E=14.6-33.2~\mu m$. Outline in polar view 3-lobate, in equatorial view elliptic or circular. Colpi $2.7-6.6~\mu m$ wide, with distinct \pm strict thickenings, or with slight to indistinct thickenings; margins uneven, slightly tapering to pointed or rounded ends; colpus membrane smooth.

Endoapertures distinct or indistinct, $4-13.3 \mu m$ long, $1.3-5.3 \mu m$ wide, lalongate, elliptic, margins uneven.

Exine 1.3–2.7 µm thick. Tectum two times thinner than infratectum. Columellae indistinct or distinct, short, arranged sparsely. Exine sculpture foveolate, microreticulate, rugulate-microperforate, rugulate, sometimes with small granules. Colpus membrane smooth, sometimes granulate

Pollen types and subtypes

Pollen grains in the studied taxa can be subdivided into two basic types, based on their aperture types, with three and five subtypes, respectively. The subtypes are separated mainly according to the pollen size, exine sculpture, and details of the colpi and endoapertures.

Type I: 3-syncolporate.

This type includes all three genera of Aptosimeae.

Subtype Ia: P=25.3–33.2 μm, E=15.9–22.6 μm. Sculpture striate, rarely with microperforations. Colpi 2.7–4 μm wide. *Anticharis*.

- 1. Endoapertures distinct, elliptic, 4–6.6 μm long, 2.7–5.3 μm wide. Colpus membrane smooth. *Anticharis imbricata*.
- 2. Endoapertures indistinct, mainly circular, rarely elliptic, $4-6.6~\mu m$ long, $4-6.6~\mu m$ wide. Colpus membrane smooth and granulate. *Anticharis linearis*.

Subtype Ib: P=25.3–34.6 μm, E=18.6–23.9 μm. Sculpture striate, rarely with microperforations and small granule. Colpi 2.4–4 μm wide. *Peliostomum*.

- 1. Endoapertures distinct, circular or elliptic, 4–5.3 μm long, 2.7–6.6 μm wide, covered by margins of colpi. Colpus membrane smooth, rarely granulate-verrucate. *Peliostomum leucorrhizum*.
- 2. Endoapertures distinct, circular or elliptic, 4–6.6 μm long, 2.7–4 μm wide, covered by margins of colpi. Colpus membrane smooth, rarely granulate. *Peliostomum virgatum*.

Subtype Ic: P=34.6–45.2 μ m, E=22.6–33.2 μ m. Sculpture striate. *Aptosimum*.

- 1. Colpi 2.4–2.7 μm wide. Endoapertures distinct, lolongate, elliptic, 5.3–7.9 μm long, 7.9–9.3 μm wide. Colpus membrane smooth or granulate-verrucate. *Aptosimum spinescens*.
- 2. Colpi 2.7–6.6 µm wide. Endoapertures distinct, circular, 6.6–7.9 µm long, 6.6–7.9 µm wide. Colpus membrane granulate and granulate-verrucate. *Aptosimum indivisum*.

Type II: 3-colpate-diorate.

This type includes all studied members of *Myoporeae*.

Subtype IIa: P=17.3–23.9 μm, E=18.6–23.9 μm. Sculpture foveolate. Colpi 2.7–5.3 μm wide, with pointed ends. Endoapertures indistinct, elliptic, 5.3–7.9 μm long, 1.3–2.4 μm wide. *Eremophila rotundifolia, E. sturtii*.

Subtype IIb: P=18.6–22.6 μm, E=18.6–21.3 μm. Sculpture microreticulate. Colpi 2.7–3.3 μm wide, with pointed ends. Endoapertures distinct or indistinct, elliptic, 4–6.6 μm long, 2.7–5.3 μm wide. *Myoporum oppositifolium*.

Subtype IIc: P=21.3–25.3 μm, E=22.6–23.9 μm. Sculpture microreticulate. Colpi 3.3–5.3 μm wide, with pointed ends. Endoapertures distinct, elliptic, 5.3–9.3 μm long, 2.7–3.3 μm wide. *Eremophila elderi*.

Subtype IId: $P=27.9-35.9~\mu m$, $E=26.6-31.9~\mu m$. Sculpture rugulate-microperforate. Colpi 4–5.3 μm wide, with pointed ends. Endoapertures distinct, elliptic, $7.9-10.6~\mu m$ long, $2-2.4~\mu m$ wide. *Eremophila glabra*.

Subtype IIe: P=27.9–34.6 μm, E=25.3–33.2 μm. Sculpture rugulate. Colpi 2.7–6.6 μm wide, with rounded ends. Endoapertures distinct or indistinct, elliptic, 4–13.3 μm long, 2–4 μm wide. *Eremophila debilis*.

Descriptions of pollen grains

Tribes, genera within tribes and species within genera are listed alphabetically. The summary of pollen meas-

urements for all studied taxa is provided in Table 1. Main pollen morphology characters are additionally summarized in Table 2.

Aptosimeae

Anticharis Endl.

Anticharis imbricata Schinz (Fig. 1A, G; 3A–D).

LM — Pollen grains 3-syncolporate, prolate, in polar view 3-lobed, in equatorial view elliptic. P=25.3–33.2 µm, E=15.9–22.6 µm. Colpi 2.7–4 µm wide, with distinct, strict margins, slightly thickened, fused on apocolpia; colpus membrane smooth, sometimes with granules over endoaperture. Endoapertures distinct, elliptic, 4–6.6 µm long, 2.7–5.3 µm wide. Exine 0.7–1.3 µm thick. Exine layers invisible. Exine sculpture invisible.

SEM — Sculpture striate, rarely with microperforations. Colpus membrane smooth, sometimes granular over endoaperture.

Specimen investigated — Namibia: grid ref. 2114 AA, flats SW of Brandberg West Mine, sandy stony open flats with scattered *Welwitschia* plants, 14 May 1976, *Oliver & Müller* 6666 (MO).

Anticharis linearis Hochst. ex Asch. (Fig. 1B, H; 3E–H).

LM — Pollen grains 3-syncolporate, prolate, in polar view 3-lobed, in equatorial view elliptic. P=26.6–30.6 μm, E=19.9–22.6 μm. Colpi 2.7–4 μm wide, with distinct, strict margins, slightly thickened, fused on apocolpia; colpus membrane smooth, sometimes with granules over endoaperture. Endoapertures indistinct, mainly circular, rarely elliptic, 4–6.6 μm long, 4–6.6 μm wide. Exine 0.7–1.3 μm thick. Exine layers invisible. Exine sculpture invisible.

SEM — Sculpture striate, rarely with microperforations. Colpus membrane smooth and granulate, sometimes granular over endoaperture.

Specimen investigated — Namibia: grid ref. 2618 AA, low bush plains c. 45 km N of Keetmanshoop, 13 Mar 1988, *P. Goldblatt & J. Manning 8748* (MO).

Aptosimum Burch. ex Benth.

Aptosimum indivisum Burch. ex Benth. (Fig. 1C, I; 3I_I)

LM — Pollen grains 3-syncolporate, prolate, in polar view 3-lobed, in equatorial view elliptic. P=37.2–45.2 μm, E=27.9–31.9 μm. Colpi 2.7–6.6 μm wide, with distinct, strict margins, slightly thickened, fused on apocolpia; colpus membrane smooth, sometimes with granules over endoaperture. Endoapertures distinct, circular, 6.6–7.9 μm long, 6.6–7.9 μm wide. Exine 1.1–2 μm thick. Exine layers invisible. Exine sculpture indistinct.

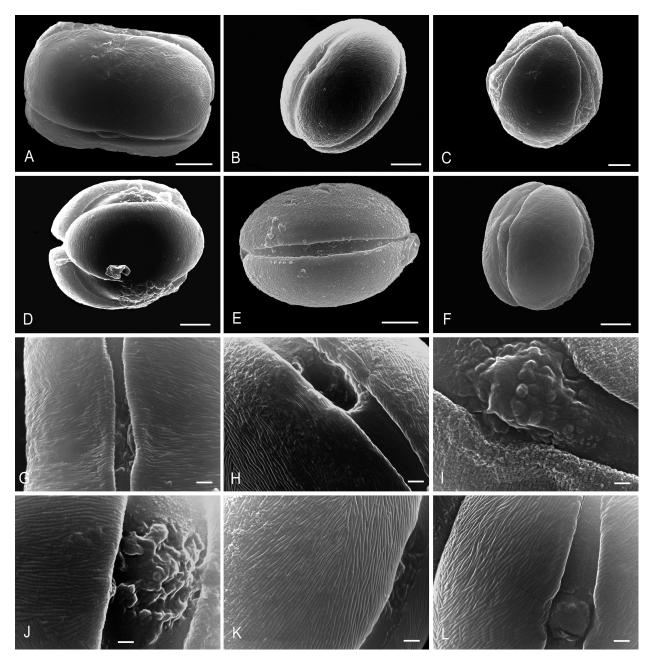


Fig. 1. Pollen grains of Aptosimeae (SEM) – A, G: Anticharis imbricata; B, H: A. linearis; C, I: Aptosimum indivisum; D, J: A. spinescens; E, K: Peliostomum leucorrhizum; F, L: P. virgatum; A–B, D–F: equatorial view; C: equatorial and polar view; exine sculpture: G, J, L: striate; H, K: striate with microperforations; I: granulate and granulate-verrucate aperture membranes. – Scale bars: $A-F=5 \mu m$; $G-L=1 \mu m$.

SEM — Sculpture striate. Colpus membrane granulate and granulate-verrucate, sometimes granular over endoaperture.

Specimen investigated — SOUTH AFRICA: NORTHERN CAPE: grid ref. 3119 (Calvinia) BD, along rail tracks near Downes Siding, 31°29'03"S, 19°57'04"E, 3606 ft, 12 Sep 2004, Peter Goldblatt & L. J. Porter 12415 (MO).

Aptosimum spinescens (Thunb.) Weber (Fig. 1D, J; 3M-P).

LM — Pollen grains 3-syncolporate, prolate, rarely oblate-spheroidal, in polar view 3-lobed, in equatorial view elliptic, sometimes circular. P=34.6–39.9 μm, E=22.6–33.2 μm. Colpi 2.4–2.7 μm wide, with distinct, strict margins, slightly thickened, fused on apocolpia; colpus membrane smooth, sometimes with granules over endoaperture. Endoapertures distinct, elliptic, 5.3–7.9 μm long, 7.9–9.3 μm wide. Exine 1.1–1.3 μm thick. Exine layers invisible. Exine sculpture indistinct.

SEM — Sculpture striate. Colpus membrane smooth or granulate-verrucate, sometimes granular over endoaperture.

Specimen investigated — Namibia: grid ref. 2416 DB, Maltehöhe district, N of Maltehöhe, about 45 km on road to Walvis Bay, 24°40′495″S, 16°49′513″E, 1341 m, dry grassland, sandy loam soil, dryland (veld), level flat slope, 11 Mar 1995, *P. M. Burgoyne 3472* (MO).

Peliostomum E. Mey. ex Benth.

Peliostomum leucorrhizum E. Mey. ex Benth. (Fig. 1E, K; 3Q-T).

LM — Pollen grains 3-syncolporate, prolate, in polar view 3-lobed, in equatorial view elliptic. P= 25.3–31.9 μm, E=18.6–22.6 μm. Colpi 2.7–4 μm wide, with distinct, strict margins, slightly thickened, fused on apocolpia; colpus membrane smooth, with granules over endoaperture. Endoapertures distinct, circular or elliptic, sometimes lolongate, 4–5.3 μm long, 2.7–6.6 μm wide, covered by margins of colpi. Exine 1.1–1.3 μm thick. Exine layers usually invisible; sometimes visible, with tectum two times thinner than lower layers. Exine sculpture invisible.

SEM — Sculpture striate, rarely with microperforations and small granules. Colpus membrane smooth, rarely granulate-verrucate.

Specimen investigated — SOUTH AFRICA: NORTHERN CAPE PROVINCE: 31 km N of Britstown (on N 13), karroid veld and sandy lax shrub steppe, 29 Sep 1974, *B. Nordenstam & J. Lundgren 2106* (MO).

Peliostomum virgatum E. Mey. ex Benth. (Fig. 1F, L; 3U-X).

LM — Pollen grains 3-syncolporate, prolate, in polar view 3-lobed, in equatorial view elliptic. P= 27.9–34.6 μ m, E=19.9–23.9 μ m. Colpi 2.4–4 μ m wide, with distinct, strict margins, slightly thickening, fused on apocolpia; colpus membrane smooth, with granules over endoaperture. Endoapertures distinct, circular or elliptic, 4–6.6 μ m long, 2.7–4 μ m wide, covered by margins of colpi. Exine 0.7–1.3 μ m thick. Exine layers usually invisible; sometimes visible, with tectum two times thinner than lower layers. Exine sculpture invisible.

SEM — Sculpture striate, rarely with microperforations and small granules. Colpus membrane smooth, rarely granulate.

Specimen investigated — SOUTH AFRICA: NORTHERN CAPE: grid ref. 2917 DC, Namaqua National Park, Keerom, Keurbos homestead, 29°59'39"S, 17°39'38.5"E, 464 m, level succulent karroo, well-drained sand soil, granite, full sun, 12 Aug 2009, *Bester S. P. 9438* (MO), det. J. Ready, Sep 2009.

Myoporeae Eremophila R. Br.

Eremophila debilis (Andrews) Chinnock (= *Myoporum debile* (Andrews) R. Br.) (Fig. 2E, K; 4Q–T).

LM — Pollen grains 3-colpate-diorate, prolate, oblate-spheroidal, rarely spheroidal, in polar view 3-lobed, in equatorial view elliptic or circular. P=27.9–34.6 μm, E=25.3–33.2 μm. Colpi 2.7–6.6 μm wide, with distinct, strict and thickened, or indistinct margins, slightly tapering to rounded ends; aperture membranes smooth. Endoapertures distinct or indistinct, elliptic, 4–13.3 μm long, 2–4 μm wide. Exine 2–2.7 μm thick. Tectum two times thinner than infratectum. Columellae distinct, short, sparsely arranged. Exine sculpture distinct, microreticulate.

SEM — Sculpture rugulate, sometimes with small granules. Colpus membrane smooth, sometimes also with small granules.

Specimen investigated — AUSTRALIA: QUEENSLAND: Darling Downs district, Yelarbon, around cemetery, c. 1 km from P.O., 28°34'S, 150°45'E, 240 m, topography flat, brown loamy soil, damaged *Eucalyptus* woodland, 10 Oct 1983, *E. M. Canning* 5820 & B. Rimes (KW).

Eremophila elderi F. Muell. (Fig. 2A, G; 4A–D).

LM — Pollen grains 3-colpate-diorate, spheroidal, rarely oblate-spheroidal and prolate, in polar view 3-lobed, in equatorial view circular. P=21.3–25.3 μm, E=22.6–23.9 μm. Colpi 3.3–5.3 μm wide, with distinct margins, slightly tapering to pointed ends; aperture membranes smooth. Endoapertures distinct, elliptic, 5.3–9.3 μm long, 2.7–3.3 μm wide. Exine 2–2.4 μm thick. Tectum two times thinner than infratectum. Columellae distinct or indistinct, short, arranged sparsely. Exine sculpture distinct, reticulate.

SEM — Sculpture microreticulate, rarely with small granules near colpi. Lumina of reticulum small, rounded, elongated or rounded-angular. Colpus membrane smooth, sometimes with small granules.

Specimen investigated — Australia: South Australia: NW region, N-facing slopes of N-most extremity of groups of prominent inselbergs between Deering Hills and Mann Ranges, c. 18 km NE of Mt Cooperinna, 26°15'00"S, 130°05'30"E), common between rocks on hillside, 8 Nov 1978, N. N. Donner 6619 (KW).

Eremophila glabra (R. Br.) Ostenf. (Fig. 2B, H; 4E–H).

LM — Pollen grains 3-colpate-diorate, prolate, rarely spheroidal and oblate-spheroidal, in polar view 3-lobed, in equatorial view elliptic, rarely circular. P=27.9–35.9 µm, E=26.6–31.9 µm. Colpi 4–5.3 µm wide, with distinct, \pm strict, thickened margins, slightly tapering to pointed ends; aperture membranes smooth. Endoapertures distinct, elliptic, with uneven margins, 7.9–10.6 µm long, 2–2.4 µm wide. Exine 2.4–2.7 µm thick. Tectum two times thinner than infratectum. Columellae mainly indistinct or distinct, short, sparsely arranged. Exine sculpture distinct, microreticulate.

SEM — Sculpture rugulate-microperforate, rare-

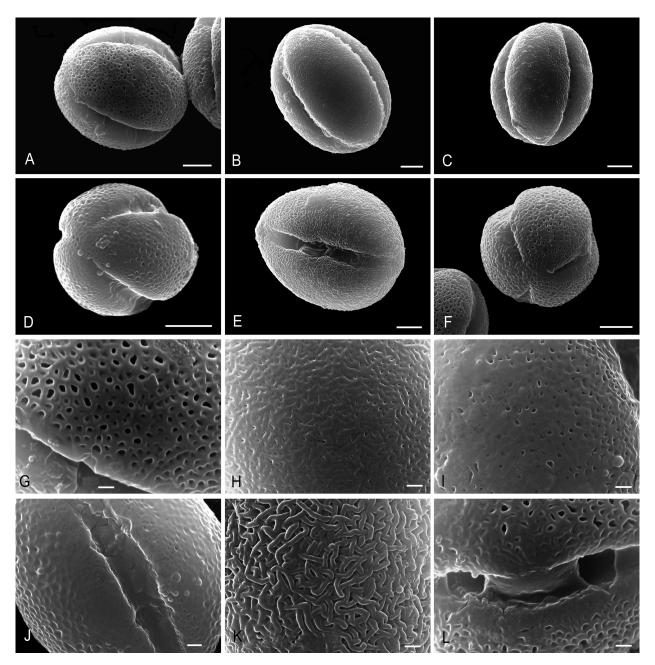


Fig. 2. Pollen grains of *Eremophila* and *Myoporum* (SEM) – A, G: *E. elderi*; B, H: *E. glabra*; C, I: *E. rotundifolia*; D, J: *E. sturtii*; E, K: *E. debilis*; F, L: *M. oppositifolium*; A–C, E: equatorial view; D, F: polar view; exine sculpture: G, L: microreticulate; H: rugulate-microperforate; I–J: foveolate; K: rugulate. – Scale bars: A–F = 5 µm; G–L = 1 µm.

ly with small granules near colpi. Colpus membrane smooth, sometimes with small granules.

Specimen investigated — Australia: South Australia: N Eyre Peninsula, foot of Iron Duke, at S end of Middleback Range, c. 50 km SW of Whyalla, growing in deep red sand, 29 Dec 1970, A. E. Orchard 2942 (KW).

Eremophila rotundifolia F. Muell. (Fig. 2C, I; Fig. 4 I–L). *LM* — Pollen grains 3-colpate-diorate, prolate and oblate-spheroidal, rarely spheroidal, in polar view 3-lobed, in equatorial view elliptic or circular. P=18.6–23.9 μm, E=18.6–23.9 μm. Colpi 4–5.3 μm wide, with indistinct,

 \pm strict margins, slightly tapering to pointed ends; aperture membranes smooth. Endoapertures indistinct, elliptic, with uneven margins, 5.3–6.6 μ m long, 1.3–2.4 μ m wide. Exine 1.1–1.6 μ m thick. Tectum two times thinner than infratectum. Columellae distinct, short, sparsely arranged. Exine sculpture distinct, microreticulate.

SEM — Sculpture foveolate. Colpus membrane smooth.

Specimen investigated — Australia: South Australia: Lake Eyre Basin, Beresford Hill, 29°16'S, 136°40'E, 80–100 m, extinct spring mound, limestone formation, 4 Oct 1978, *K. Chorney 1020* (KW).

Eremophila sturtii R. Br. (Fig. 2D, J; 4M-P).

LM — Pollen grains 3-colpate-diorate, prolate, rarely spheroidal, in polar view 3-lobed, in equatorial view elliptic, rarely circular. P=17.3–21.3 μ m, E=14.6–18.6 μ m. Colpi 2.7–5.3 μ m wide, with indistinct, uneven margins, slightly tapering to pointed ends; aperture membranes smooth. Endoapertures indistinct, elliptic, with uneven margins, 5.3–7.9 μ m long, 1.3–2.4 μ m wide. Exine 1.1–1.6 μ m thick. Tectum two times thinner than infratectum. Columellae indistinct. Exine sculpture mainly indistinct, microreticulate.

SEM — Sculpture foveolate, rarely with granules near colpi. Colpus membrane smooth, sometimes with small granules.

Specimen investigated — AUSTRALIA: QUEENSLAND: Warrego district, c. 25 km SW of Eulo on Hungerford road, flood plain of Paroo River, 28°20'S, 144°54'E, grey sandy clay soil, Eucalyptus largiflorens open woodland with well-developed shrubby layer of Eremophila mitchellii and E. sturtii, 13 Sep 1973, R. J. Henderson H2048 & D. E. Boyland (KW).

Myoporum Sol. ex G. Forst.

Myoporum oppositifolium R. Br. (Fig. 2F, L; 4U–X).

LM — Pollen grains 3-colpate-diorate, spheroidal, rarely oblate-spheroidal and prolate, in polar view 3-lobed, in equatorial view circular. P=18.6–22.6 μm, E=18.6–21.3 μm. Colpi 2.7–3.3 μm wide, with distinct, slightly thickening or indistinct margins, slightly tapering to ± pointed ends; aperture membranes smooth. Endoapertures distinct or indistinct, elliptic, 4–6.6 μm long, 2.7–5.3 μm wide. Exine 2–2.7 μm thick. Tectum two times thinner than infratectum. Columellae indistinct, short, sparsely arranged. Exine sculpture distinct, reticulate.

SEM — Sculpture microreticulate, sometimes with small granules. Lumina of reticulum small, rounded, elongated or rounded-angular. Colpus membrane smooth, sometimes with small granules.

Specimen investigated — Australia: Western Australia: SW division, Augusta c. 115 km SSW of Bunbury, near golf links, 12 Oct 1967, A. M. Ashby 2377 (KW).

Discussion

Comparative pollen morphology of Aptosimeae

The palynomorphological data showed that pollen grains of the studied taxa of *Aptosimeae* have similar aperture types and sculpture; however, they differ in their size and some features of apertures (Table 1, 2). Thus, species of *Aptosimum* have the largest pollen grains and endoapertures as compared to two other genera, *Anticharis* and

Peliostomum (Table 1). Pollen grains of the two studied species of Peliostomum have distinctive features in the structure of their apertures: endoaperture edges are covered with edges of colpi, a feature not observed in Anticharis and Aptosimum.

The two studied species of *Aptosimum* differ in their pollen size and structure of apertures. Pollen grains of *A. indivisum* are larger, with wider colpi and circular endoapertures, while pollen grains of *A. spinescens* are smaller, with narrower colpi and elliptic endoapertures.

The studied species of *Anticharis* differ in the shape of endoapertures (Table 2).

Species of *Peliostomum* have pollen grains similar in size, colpi structure and exine sculpture. In both *P. leu-corrhizum* and *P. virgatum* we observed both circular and elliptic endoapertures; however, in addition to that, lolongate endoapertures also sometimes occur in *P. leuco-rrhizum*.

Probably the specialized pollination by pollen wasps (Gess & Gess 2010) mentioned above is partly responsible for at least some peculiar pollen morphology features of *Aptosimeae* revealed in the present study.

Comparative pollen morphology of *Myoporeae*

Our data showed that pollen grains in the studied species of tribe *Myoporeae* are similar by their 3-colpate-diorate type of apertures but differ in their size, exine sculpture and structural features of apertures (Table 1, 2).

The studied species of *Eremophila* differ in their pollen size, exine sculpture and structural features of apertures. The smallest pollen grains are characteristic of *Eremophila elderi*, *E. rotundifolia* and *E. sturtii*. Pollen grains of *E. rotundifolia* and *E. sturtii* are similar in having narrow endoapertures and foveolate sculpture; in contrast, pollen grains of *E. elderi* have wider endoapertures and microreticulate exine sculpture. Pollen grains of *E. debilis* and *E. glabra* are larger than those in the other three species. These two species differ by their colpi structure: colpi in *E. debilis* have rounded ends, whereas in *E. glabra* the ends are pointed. Pollen grains of these two taxa also differ from those in all other species by their exine sculpture (Table 2).

Pollen of *Myoporum oppositifolium* is similar to pollen of *Eremophila rotundifolia* and *E. sturtii* in size and colpi structure; however, it differs by exine sculpture and endoaperture structure (Table 1, 2).

In general, our data are comparable to data provided by Niezgoda & Tomb (1975), with some additional features revealed. In particular, we found in members of *Eremophila* some exine sculpture types that were not reported earlier: foveolate, rugulate-microperforate and rugulate. Niezgoda & Tomb (1975) mentioned that rugulate exine sculpture occurs only in two studied species, *Capraria biflora* and *Leucophyllum pruinosum* I. M. Johnst. Our studies demonstrated that this sculpture pattern also occurs in some representatives of *Eremophila*.

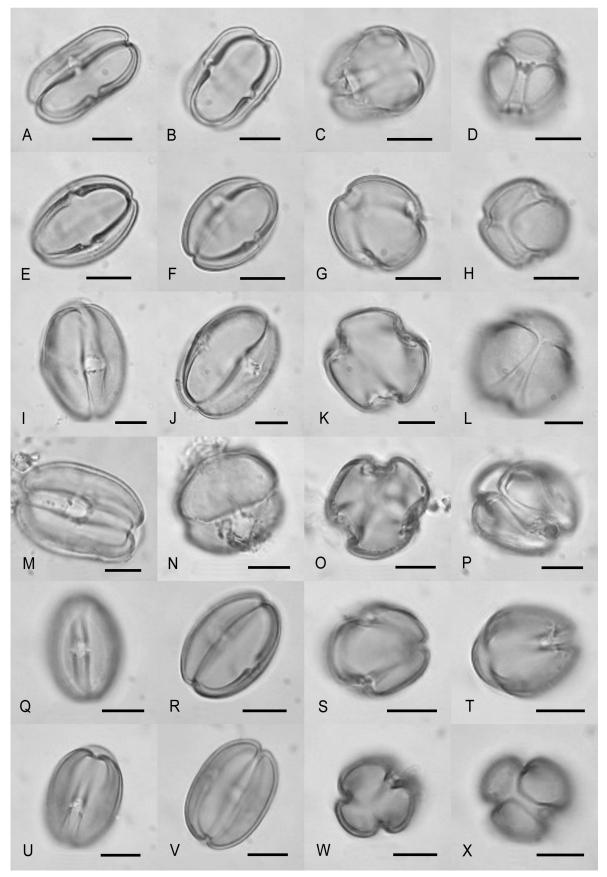


Fig. 3. Pollen grains of *Aptosimeae* (LM) – A–D: *Anticharis imbricata*; E–H: *A. linearis*; I–L: *Aptosimum indivisum*; M–P: *A. spinescens*; Q–T: *Peliostomum leucorrhizum*; U–X: *P. virgatum*; A, B, E, F, I, J, M, N, Q, R, U, V: equatorial view; C, D, G, H, K, L, O, P, S, T, W, X: polar view. – Scale bars: $A-X=10~\mu m$.

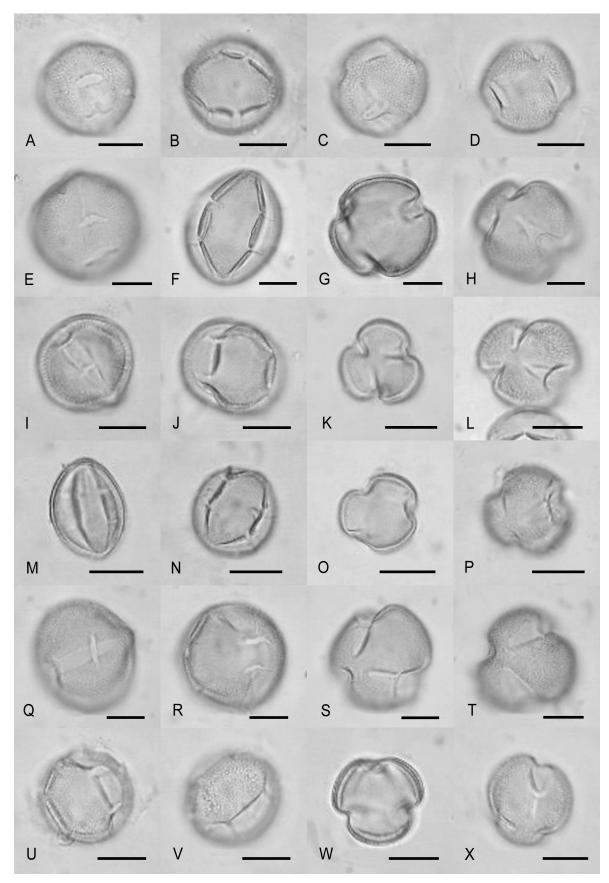


Fig. 4. Pollen grains of Eremophila and Myoporum (LM) – A–D: E. elderi; E–H: E. glabra; I–L: E. rotundifolia; M–P: E. sturtii; Q–T: E. debilis; U–X: M. oppositifolium; A, B, E, F, I, J, M, N, Q, R, U, V: equatorial view; C, D, G, H, K, L, O, P, S, T, W, X: polar view. – Scale bars: $A-X=10~\mu m$.

TT 11 1 C		1		4 11	1		
Table 1. Summar	vot no	len measi	irements /	AΠι	dimensi	ons are	ın ıım

Taxon	Polar axis	Equatorial axis	Colpi width	Endoaperture length	Endoaperture width	Exine thickness
Anticharis imbricata	25.3-33.2	15.9–22.6	2.7-4.0	4.0-6.6	2.7-5.3	0.7-1.3
Anticharis linearis	26.6-30.6	19.9-22.6	2.7-4.0	4.0-6.6	4.0-6.6	0.7-1.3
Aptosimum indivisum	37.2-45.2	27.9-31.9	2.7-6.6	6.6–7.9	6.6–7.9	1.1-2.0
Aptosimum spinescens	34.6-39.9	22.6-33.2	2.4-2.7	5.3-7.9	7.9–9.3	1.1-1.3
Peliostomum leucorrhizum	25.3-31.9	18.6-22.6	2.7-4.0	4.0-5.3	2.7-6.6	1.1-1.3
Peliostomum virgatum	27.9-34.6	19.9-23.9	2.4-4.0	4.0-6.6	2.7-4.0	0.7-1.3
Eremophila debilis	27.9-34.6	25.3-33.2	2.7-6.6	4.0-13.3	2.0-4.0	2.0-2.7
Eremophila elderi	21.3-25.3	22.6-23.9	3.3-5.3	5.3-9.3	2.7-3.3	2.0-2.4
Eremophila glabra	27.9-35.9	26.6-31.9	4.0-5.3	7.9–10.6	2.0-2.4	2.4-2.7
Eremophila rotundifolia	18.6-23.9	18.6-23.9	4.0-5.3	5.3-6.6	1.3-2.4	1.1-1.6
Eremophila sturtii	17.3-21.3	14.6-18.6	2.7-5.3	5.3-7.9	1.3-2.4	1.1-1.6
Myoporum oppositifolium	18.6–22.6	18.6–21.3	2.7–3.3	4.0-6.6	2.7-5.3	2.0-2.7

Chinnock (2007) reported that pollen grains of almost all genera (selected representatives of *Calamphoreus*, *Eremophila*, *Glycocystis* and *Myoporum*) have reticulate exine (coarsely and finely reticulate), and only all four species of *Diocirea* have punctitegillate (perforate) exine sculpture. However, our new data indicated that an exine sculpture pattern very similar to punctitegillate (perforate) also occurs in some species of *Eremophila*, in particular, *E. rotundifolia* and *E. sturtii*. Because of that, we believe that this peculiar exine pattern cannot be regarded as a reliable character distinguishing *Diocirea* and *Eremophila*.

Species of *Eremophila* studied by us belong to different sections (following the system proposed by Chinnock 2007) and differ by their pollen morphology. However, at present we have insufficient palynological data to analyse whether these characters support the sections, and pollen studies of additional taxa of that genus are needed.

Comparison of palynomorphological and molecular phylogenetic evidence

According to molecular phylogenetic data (Oxelman & al. 2005; Khosravesh & al. 2012), *Anticharis* and *Peliostomum* are sister groups within *Aptosimeae*. Palynomorphological data confirm the close relationship between these two genera, as shown above. It has been also shown that within *Anticharis* three clades can be outlined reflecting the gradual transition to C₄ photosynthesis (Khosravesh & al. 2012). *Aptosimum* is sister to the *Anticharis+Peliostomum* clade, and pollen morphology data also support that pattern, since the morphological differences between pollen grains of *Anticharis* and *Peliostomum* are exceeded by the differences between those two genera and *Aptosimum*.

According to molecular phylogenetic data (Oxelman & al. 2005), the genera *Bontia*, *Eremophila* and *Myopo*-

rum (Myoporeae in the strict sense) form a clade sister to the Capraria+Leucophyllum clade (Leucophylleae). Our palynomorphological data, as well as data of other authors (Niezgoda & Tomb 1975), are compatible with the molecular phylogenetic data. Pollen grains of all studied representatives of these genera are 3-colpate-diorate, with foveolate, microreticulate, reticulate, rugulate-perforate and/or rugulate exine sculpture.

The monotypic Madagascan genus *Androya* has been revealed by molecular phylogenetic analysis (Oxelman & al. 2005) as the most early-branching clade, which is sister to the clade containing all other genera of *Myoporeae* and *Leucophylleae*. According to Punt & Leenhouts (1967), pollen grains of *Androya* are 3-colporate, with smooth exine sculpture, thus differing considerably from pollen grains of other taxa of *Myoporeae* and *Leucophylleae*.

Oxelman & al. (2005) indicated that members of *Leucophylleae* (*Capraria* and *Leucophyllum*) are closely related to *Myoporeae* according to anatomical and palynomorphological evidence. Considering possible relationships of *Aptosimeae*, they mentioned that "[m]onophyly of *Aptosimeae* is strongly supported by our chloroplast DNA sequences, and a sister group relation with *Androya*, *Leucophylleae*, and *Myoporeae* is moderately supported. All have 3-colpate, diporate [sic!] pollen (Erdtman 1952; Niezgoda & Tomb 1975; Punt 1980)" (Oxelman & al. 2005: 419). The word "diporate" in their text is most probably an error for "diorate".

In fact, our data indicate that pollen grains in the clades *Aptosimeae*, *Leucophylleae+Myoporeae* and *Androya* evidently differ by their aperture types: 3-syncolporate, 3-colpate-diorate, and 3-colporate, respectively. There are also some differences in exine sculpture patterns: striate in *Aptosimeae*; rather diverse (but not striate nor smooth) in *Leucophylleae+Myoporeae*, a more diverse and speciesrich group; and smooth in *Androya* (Table 2).

Table 2. Summary of pollen morphological characters: original data compared with literature data. "—" = no data reported.

Taxon	Apertures	Colpus membrane	Endoaperture	Exine sculpture	Source of data
Aptosimeae					
Anticharis imbricata	3-syncolporate	smooth	elliptic	striate, rarely with microperforations	original data
Anticharis linearis	3-syncolporate	smooth and granulate	mainly circular, rarely elliptic	striate, rarely with microperforations	original data
Aptosimum indivisum	3-syncolporate	granulate and granu- late-verrucate	circular	striate	original data
Aptosimum spinescens	3-syncolporate	smooth or granulate- verrucate	elliptic	striate	original data
Peliostomum leucorrhizum	3-syncolporate	smooth, rarely granulate-verrucate	circular or elliptic	striate, rarely with microperforations and small granules	original data
Peliostomum virgatum	3-syncolporate	smooth, rarely granulate	circular or elliptic	striate, rarely with microperforations and small granules	original data
Myoporeae					
Eremophila debilis	3-colpate-diorate	smooth, some- times also with small granules	elliptic	rugulate, sometimes with small granules	original data
Eremophila elderi	3-colpate-diorate	smooth, some- times with small granules	elliptic	microreticulate, rarely with small granules near colpi (small lumina)	original data
Eremophila glabra	3-colpate-diorate	smooth, some- times also with small granules	elliptic	rugulate-microperforate, rarely with small granules near colpi	original data
Eremophila rotundifolia	3-colpate-diorate	smooth	elliptic	foveolate	original data
Eremophila sturtii	3-colpate-diorate	smooth, some- times with small granules	elliptic	foveolate, rarely with granules near colpi	original data
Eremophila (all studied spp.)	3-colpate-diorate	_	elliptic	finely reticulate (lumina of irregular size)	Niezgoda & Tomb (1975)
Eremophila (all studied spp.)	3-colpate-diorate	_	_	finely reticulate	Chinnock (2007)
Myoporum oppositifolium	3-colpate-diorate	smooth, some- times with small granules	elliptic	microreticulate, some- times with small gra- nules (small lumina)	original data
Myoporum acuminatum R. Br.	3-colpate-diorate	_	elliptic	finely reticulate (very large lumina)	Niezgoda & Tomb (1975)
Myoporum (all studied spp. except M. acuminatum)	3-colpate-diorate	_	elliptic	finely reticulate	Niezgoda & Tomb (1975)
Myoporum (all studied spp.)	3-colpate-diorate	_	_	finely or coarsely reticulate	Chinnock (2007)
Pentacoelium	3-colpate-diorate	_	_	finely reticulate	Chinnock (2007)
Diocirea (all studied spp.)	3-colpate-diorate	_		punctitegillate	Chinnock (2007)
Calamphoreus	3-colpate-diorate	_		finely reticulate	Chinnock (2007)
Glycocystis	3-colpate-diorate		_	finely reticulate	Chinnock (2007)
Bontia daphnoides	3-colpate-diorate	_	_	finely reticulate	Chinnock (2007)
Bontia daphnoides	3-colpate-diorate	_	elliptic	finely reticulate (lumina of irregular size)	Niezgoda & Tomb (1975)
Leucophylleae					
Capraria biflora	3-colpate-diorate	_	elliptic	rugulate	Niezgoda & Tomb (1975)
Leucophyllum pruinosum	3-colpate-diorate	_	elliptic	rugulate	Niezgoda & Tomb (1975)
Leucophyllum (all studied spp. except L. pruinosum)	3-colpate-diorate	_	elliptic	finely reticulate (lumina smaller near colpi)	Niezgoda & Tomb (1975)

Taxon	Apertures	Colpus membrane	Endoaperture	Exine sculpture	Source of data
Leucophylleae (continued)					
Leucophyllum texanum	3-colpate-diorate	psilate	_	reticulate	Minkin & Esh- baugh (1989)
Faxonanthus pringlei	3-colpate-diorate	_	elliptic	finely reticulate (small lumina)	Niezgoda & Tomb (1975)
Eremogeton grandiflorus	3-colpate-diorate	_	elliptic	finely reticulate (large irregular lumina)	Niezgoda & Tomb (1975)
Androya clade					
Androya decaryi	3-colporate	smooth	indistinct	smooth	Punt & Leenhouts (1967)

Our palynomorphological data, as well as other palynomorphological (Niezgoda & Tomb 1975) and molecular (Oxelman & al. 2005) evidence, indicate that the *Androya* clade probably merits recognition as a separate tribe, if *Leucophylleae* and *Myoporeae* are also recognized as separate tribes. An alternative logical solution would be an extended *Myoporeae*, re-circumscribed to include *Leucophylleae* and *Androya*.

Possible ancestral pollen character states in Scrophulariaceae

Pollen grains of representatives of all these tribes are characterized by the complex colporate type (combining ectoapertures and endoapertures). It may be assumed that this type is ancestral in the family *Scrophulariaceae*; however, the ancestral status of the colpate type cannot be excluded as well. If we postulate the colporate type as being ancestral in the family, then the 3-colporate type with smooth exine sculpture is probably the most ancient pattern, being also typical for many representatives of Buddlejeae and Teedieae (Tsymbalyuk & Mosyakin 2013; Mosyakin & Tsymbalyuk 2015), as well as for Androya. Further evolutionary changes of this type could have led to the formation of more advanced 3-colpatediorate pollen with rugulate and reticulate exine sculpture (peculiar to Leucophylleae+Myoporeae) and 3-syncolporate pollen with mainly striate sculpture (typical for Aptosimeae, as shown in this article), either in parallel or consecutively.

Our original data on pollen morphology of *Hemimerideae*, most probably the basalmost clade of *Scrophulariaceae*, will be published later. However, it is already evident from our preliminary data that two subclades of that tribe have colpate and colporate pollen, respectively, which is consistent with our hypothesis on pollen evolution patterns in early-branching *Scrophulariaceae*.

Conclusions

Our analysis of available palynomorphological data (both published earlier and newly reported here) clearly outlines the same main clades as those revealed by molecular phylogenetic studies (Oxelman & al. 2005; Gándara & Sosa 2013): Aptosimeae (3-syncolporate pollen grains, striate exine sculpture); Leucophylleae+Myoporeae (3-colpate-diorate pollen grains, foveolate, microreticulate, rugulate-microperforate, rugulate and/or reticulate exine sculpture); and Androya (3-colporate pollen grains, smooth exine sculpture). The 3-colporate pollen type with smooth exine sculpture, typical for Androya and occurring also in many representatives of Buddlejeae and Teedieae (Tsymbalyuk & Mosyakin 2013; Mosyakin & Tsymbalyuk 2015), is hypothesized to be ancestral in Scrophulariaceae. The clades Leucophylleae+Myoporeae and Aptosimeae have more advanced pollen types: 3-colpate-diorate with mainly rugulate and reticulate exine in the former, and 3-syncolporate with mainly striate sculpture in the latter.

Acknowledgements

The authors express their gratitude to James C. Solomon, Head Curator of the Missouri Botanical Garden herbarium (MO), Tatyana V. Shulkina, and staff members of MO, and to Nataliya M. Shyian, Head Curator of the National Herbarium of Ukraine (KW; herbarium of the M. G. Kholodny Institute of Botany, National Academy of Sciences of Ukraine), for their cooperation and assistance in extracting pollen samples from herbarium specimens. Special thanks are due to Peter H. Raven for his long-term support and encouragement of the senior author's research at the MO herbarium. Kind help and cooperation of Dmytro O. Klymchuk, Head of the Center of Electron Microscopy (M. G. Kholodny Institute of Botany), is greatly appreciated. The authors are grateful to two anonymous reviewers for their detailed review of the manuscript, valuable comments and suggestions, and to Nicholas Turland for his skilful editorial comments.

References

Chinnock R. J. 2007: *Eremophila* and allied genera: a monograph of the plant family *Myoporaceae*. – Dural: Rosenberg.

- Erdtman G. 1952: Pollen morphology and plant taxonomy: an introduction to palynology 1. Angiosperms.
 Waltham: Chronica Botanica Co.; Stockholm: Almquist & Wiksell.
- Fischer E. 2004: *Scrophulariaceae*. Pp. 333–432 in: Kubitzki K. (ed.), The families and genera of vascular plants **7.** Berlin: Springer.
- Fischer E., Schäferhoff B. & Müller K. 2013: The phylogeny of *Linderniaceae* the new genus *Linderniella*, and new combination within *Bonnaya*, *Craterostigma*, *Lindernia*, *Micranthemum*, *Torenia* and *Vandellia*. Willdenowia **43:** 209–238.
- Gándara E. & Sosa V. 2013: Testing the monophyly and position of the North American shrubby desert genus *Leucophyllum (Scrophulariaceae: Leucophylleae).* Bot. J. Linn. Soc. **171:** 508–518.
- Gess S. K. & Gess F. W. 2010: Pollen wasps and flowers in southern Africa. Pretoria: SANBI. SANBI Biodiversity Series 18.
- Henrickson J. & Flyr L. D. 1985: Systematics of *Leuco-phyllum* and *Eremogeton* (*Scrophulariaceae*). Sida **11:** 107–172.
- Khoshravesh R., Akhani H., Sage T. L., Nordenstam B. & Sage R. F. 2012: Phylogeny and photosynthetic pathway distribution in *Anticharis* Endl. (*Scrophulariaceae*). J. Exp. Bot. 63: 5645–5658.
- Minkin J. P. & Eshbaugh W. H. 1989: Pollen morphology of the *Orobanchaceae* and rhinanthoid *Scrophulariaceae*. Grana **28:** 1–18.
- Mosyakin S. L. & Tsymbalyuk Z. M. 2015: Pollen morphology of the southern African tribe *Teedieae*, an early-branching lineage of crown *Scrophulariaceae*. Willdenowia **45:** 65–75.
- Niezgoda Ch. J. & Tomb A. S. 1975: Systematic palynology of tribe *Leucophylleae* (*Scrophulariaceae*) and selected *Myoporaceae*. Pollen & Spores 17: 495–516.
- Olmstead R. [with the help of Albach D., Bremer B., Cantino P., dePamphilis C., Garnock-Jones P., Harley R., Ihlenfeldt H.-D., Lohmann L., Mathews S., McDade L., Norman E., Oxelman B., Reveal J., Scotland R., Smith J., Wagstaff S., Wallander E., Weber A., Wolfe A., Wortley A., Young N., Zjhra M. & others] 2012: A synoptical classification of the *Lamiales*. Version 2.4 [updated 26 Jul 2012]. Published at http://depts. washington.edu/phylo/Classification.pdf
- Olmstead R. G., dePamphilis C. W., Wolfe A. D. & Young A. D. 2001: Disintegration of the *Scrophulariaceae*. Amer. J. Bot. **88:** 348–361.
- Olmstead R. G. & Reeves P. A. 1995: Evidence for the polyphyly of the *Scrophulariaceae* based on chloroplast *rbcL* and *ndhF* sequences. Ann. Missouri Bot. Gard. **82:** 176–193.

- Oxelman B., Kornhall P., Olmstead R. G. & Bremer B. 2005: Further disintegration of the *Scrophulariaceae*. Taxon **54**: 411–425.
- Punt W., Blackmore S., Nilsson S. & Le Thomas A. 1994: Glossary of pollen and spore terminology. – Utrecht: LPP Foundation.
- Punt W. & Leenhonts P. W. 1967: Pollen morphology and taxonomy in the *Loganiaceae*. Grana Palynol. **7:** 469–516.
- Rahmanzadeh R., Müller K., Fischer E., Bartels D. & Borsch T. 2005: The *Linderniaceae* and *Gratiolaceae* are further lineages distinct from the *Scrophulariaceae* (*Lamiales*). Pl. Biol. (Stuttgart) **7:** 67–78.
- Reveal J. L. 2012: An outline of a classification scheme for extant flowering plants. Phytoneuron **2012–37:** 1–221.
- Sage R. F., Christin P. A. & Edwards E. J. 2011: The C₄ plant lineages of planet Earth. J. Exp. Bot. **62**: 3155–3169.
- Schäferhoff B., Fleischmann A., Fischer E., Albach D. C. Borsch T., Heubl G. & Müller K. F. 2010: Towards resolving *Lamiales* relationships: insights from rapidly evolving chloroplast sequences. – BMC Evol. Biol. 10: 352–374.
- Takhtajan A. L. 1987: Systema Magnoliophytorum. Leningrad: Nauka. [Тахтаджян А. Л. 1987: Система магнолиофитов. Ленинград: Наука].
- Takhtajan A. L. 1997: Diversity and classification of flowering plants. – New York: Columbia University Press.
- Takhtajan A. 2009: Flowering plants. Berlin: Springer.
 Tank D. C., Beardsley P. M., Kelchner S. A. & Olmstead R. G. 2006: Review of the systematics of *Scrophulariaceae* s.l. and their current disposition. Austral. Syst. Bot. 19: 289–307.
- Thiers B. 2015+ [continuously updated]: Index herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden: published at http://sweetgum.nybg.org/ih/ [accessed 3 Jul 2015].
- Tokarev P. I. 2002: Morphology and ultrastructure of the pollen grains. Moscow: KMK Scientific Press. [Токарев П. И. 2002: Морфология и ультраструктура пыльцевых зерен. Москва: Товарищество научных изданий КМК].
- Tsymbalyuk Z. M. & Mosyakin S. L. 2013: Atlas of pollen grains of representatives of *Plantaginaceae* and *Scrophulariaceae*. Kyiv: Nash Format. [Цимбалюк З. М., Мосякін С. Л. 2013: Атлас пилкових зерен представників родин *Plantaginaceae* та *Scrophulariaceae*. Київ: Наш Формат].