

Original Scientific Paper

***Aspicilia nigromaculata* sp. nov. (Megasporaceae, Pertusariales, Ascomycota) from Azad Jammu and Kashmir, Pakistan, evidence from morphology and DNA sequencing data**

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ABSTRACT:

A new species of Megasporaceae, *Aspicilia nigromaculata* sp. nov. is described and illustrated from Pakistan. A comparative morpho-anatomical study and ITS–LSU-based molecular analyses confirmed its position within the genus *Aspicilia*. It differs from the other species of the genus by the following set of features: a well-developed, grey to greenish grey frequently black spotted thick thallus, well-developed thicker peripheral and flat to slightly concave areoles, and small conidia 10–15 × 0.5–1 µm. Its positioning in a separate branch in the phylogenetic tree also makes it distinct from the other known species of the genus.

Keywords:

ITS nrDNA, nrLSU, phylogeny, saxicolous lichens

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INTRODUCTION

The currently accepted taxonomy of the lichenized family Megasporaceae (Pertusariales) includes eight genera i.e. *Aspicilia* A. Massal., *Circinaria* Link (SOHRABI *et al.* 2013a), *Lobothallia* (Clauzade & Cl. Roux) Hafellner (PAUKOV *et al.* 2019), *Megaspora* (Clauzade & Cl. Roux) Hafellner & V. Wirth, *Sagedia* Ach. (NORDIN *et al.* 2010), *Teuvoa* Sohrabi & S. Leavitt (SOHRABI *et al.* 2013b), *Aspiciliella* M. Choisy (ZAKERI *et al.* 2017) and *Oxneriarria* S.Y. Kondr. & Lökös (HAJI MONIRI *et al.* 2017). The lichen-forming ascomycete genus *Aspicilia* A. Massal. has a worldwide distribution and covers a miscellaneous collection of 200–250 species (SOHRABI *et al.* 2013c). It presents a considerable range of morphological variations (SOHRABI *et al.* 2011). Within the genus a number of species have radiating thalli and elongate, often ± diverging and ± branched marginal areoles, closely attached to the substratum (NORDIN *et al.* 2011). *Aspicilia nigromaculata* is described from Pakistan, based on a detailed morphological and anatomical characterization. The ITS and nrLSU based phylogenetic analyses also support the identity of this taxon as a novel species.

MATERIAL AND METHODS

Collection and preservation. The survey of the Jhelum Valley and the Peer Chinasi area, in the Muzaffarabad district of Azad Jammu and Kashmir, Pakistan, undertaken in 2020, focused on increasing the biotic knowledge of the lichens of Pakistan. Azad Jammu and Kashmir, which lies in the north-eastern part of Pakistan, comprises an area of 13297 km², with mountains ranging from 360 to 6325 m in altitude, and is characterised by rocky and undulating terrain (DAR *et al.* 2012). The collected samples were deposited in the herbarium of the Institute of Botany, University of the Punjab, Lahore (LAH).

Morphological characterisation. The specimens were examined macro and micromorphologically under a stereomicroscope (Meiji Techno, EMZ-5TR, Japan) and compound microscope (SWIFT M4000-D), with a 9MP camera system, respectively. For anatomical investigation, sections of the apothecia were cut manually and mounted in water, KOH. A minimum of twenty measurements in water were made for each diagnostic feature.

Chemical characterisation. The secondary chemistry was analysed using spot tests which were performed using KOH (10%; K), and a sodium hypochlorite solution (C). Thin layer chromatography was carried out using Solvent System G, following standard methods (ORANGE *et al.* 2010).

DNA extraction, PCR amplification and sequencing. Genomic DNA was extracted directly from a portion of the thallus with apothecia from each specimen using a modified 2% CTAB method (GARDES & BRUNS 1993). The ITS-nrDNA region (Internal Transcribed Spacer of the nrDNA) was amplified using the primer pair ITS1F (forward primer) (GARDES & BRUNS 1993) and ITS4 (reverse primer) (WHITE *et al.* 1990). The nrLSU region was also amplified by using the LROR as the forward and LR5 as the reverse primer for the nrLSU region (VILGALYS & HESTER 1990), following the amplification protocol of KHAN *et al.* (2018). The PCR products were visualized on a 1% agarose gel with ethidium bromide (SAMBROOK & RUSSEL 2001). The PCR products were then sent to Tsingke, China for sequencing. The sequences were aligned using BioEdit (HALL 1999).

BLAST analysis was used to retrieve highly similar sequences of both the ITS and nrLSU regions. Maximum query coverage and percent identity between related taxa were noted. The sequences retrieved from Genbank and proposed in the literature were used for initial alignment, which was trimmed and then realigned using web-PRANK with default settings (LÖYTYNOJA & GOLDMAN 2010). On the CIPRES Portal (MILLER *et al.* 2010) the HYK+G+I model was selected using jModelTest (POSADA 2008). Maximum likelihood analysis was also conducted using RAxML-HPC2 v. 8.1.11 on the CIPRES web server (STAMATAKIS 2014). 1000 bootstraps were used for rapid bootstrapping. FigTree v 1.4.3 (RAMBAUT *et al.* 2014) was used to display the phylogeny from the ML analysis.

RESULTS

An ITS sequence was successfully obtained from two specimens of the new species, described as *Aspicilia nigromaculata*. The sequences for the holotype of the new species, and a second collection (JV-07) were identical. An additional eighteen sequences of selected species of

Table 1. Specimens used in the ITS phylogenetic analyses of the *Aspicilia* species. New sequences are in bold

Name of species	Country/ Origin	Collection number	Genbank no. (ITS)	Reference
<i>Aspicilia abbasiana</i>	China	Ismayil et Abbas 20111154	KM609324	ISMAYIL <i>et al.</i> 2015; KONDRATYUK <i>et al.</i> 2016
<i>Aspicilia blastidiata</i>	Russia	Paukov AGP20111009-01	KX129963	PAUKOV <i>et al.</i> 2015, 2017
<i>Aspicilia blastidiata</i>	Russia	Paukov AGP20120801-01	KX159286	PAUKOV <i>et al.</i> 2015, 2017
<i>Aspicilia cinerea</i>	Sweden	Hafellner 37308	AF332111	IVANOVA & HAFELLNER 2001
<i>Aspicilia cinerea</i>	Austria	Kocourkova & Hafellner 46364	AF332112	IVANOVA & HAFELLNER 2001
<i>Aspicilia cuprea</i>	USA	Owe-Larsson 9112	EU057902	NORDIN <i>et al.</i> 2007
<i>Aspicilia dudinensis</i>	Sweden	Nordin 6036	EU057906	NORDIN <i>et al.</i> 2007
<i>Aspicilia epiglypta</i>	Sweden	Nordin 6303	EU057907	NORDIN <i>et al.</i> 2007
<i>Aspicilia epiglypta</i>	Sweden	Nordin 6305	HQ259261	NORDIN <i>et al.</i> 2011
<i>Aspicilia fluviatilis</i>	Sweden	Nordin 6188	HQ259264	NORDIN <i>et al.</i> 2011
<i>Aspicilia goettweicensis</i>	Austria	Vondrák 14026	KX159289	PAUKOV <i>et al.</i> 2017
<i>Aspicilia goettweicensis</i>	Russia	Paukov AGP20120513-03	KX159292	PAUKOV <i>et al.</i> 2017
<i>Aspicilia granulosa</i>	Sweden	Nordin 6174	HQ259265	NORDIN <i>et al.</i> 2011
<i>Aspicilia nigromaculata</i>	Pakistan	LAH36940	MW969628	This paper
<i>Aspicilia nigromaculata</i>	Pakistan	LAH36941	MW969629	This paper
<i>Aspicilia subdepressa</i>	France	Roux 24653	JF703123	ROUX <i>et al.</i> 2011
<i>Aspicilia subepiglypta</i>	Korea	100857 KoLRI	KY249607	KONDRATYUK <i>et al.</i> 2016
<i>Aspicilia subepiglypta</i>	Korea	110495 KoLRI	KY249608	KONDRATYUK <i>et al.</i> 2016
<i>Aspicilia subradians</i>	Sweden	Nordin 5984	HQ259267	NORDIN <i>et al.</i> 2011
<i>Aspicilia subradians</i>	Finland	Nordin 6370	HQ259268	NORDIN <i>et al.</i> 2011
<i>Circinaria esculenta</i>	Kazakhstan	Lvanov s. n. (UFU L-1743)	MK347507	PAUKOV <i>et al.</i> 2019
<i>Circinaria fruticulosa</i>	Russia	Paukov 3074 (UFU L-3256)	MK347508	PAUKOV <i>et al.</i> 2019
<i>Ochrolechia parella</i>	Antarctica	Park PCH080112-32	KJ607905	PARK <i>et al.</i> 2015
<i>Oxneriaria dendroplaca</i>	Finland	Nordin 6366	HQ259260	NORDIN <i>et al.</i> 2011
<i>Oxneriaria mashiginensis</i>	Sweden	Nordin 5790	EU057912	NORDIN <i>et al.</i> 2007
<i>Oxneriaria rivulicola</i>	Sweden	Nordin 5957	EU057922	NORDIN <i>et al.</i> 2007
<i>Oxneriaria verruculosa</i>	Norway	Owe-Larsson 9007	EU057940	NORDIN <i>et al.</i> 2007
<i>Oxneriaria verruculosa</i>	Sweden	Nordin 5942	EU057942	NORDIN <i>et al.</i> 2007

Table 2. Specimens used in the nrLSU phylogenetic analyses of the *Aspicilia* species. New sequences are in bold

Name of species	Country/Origin	Collection number	Genbank no. (ITS) Reference
<i>Aspicilia cinerea</i>	Sweden, Östergötland	Nordin 5542 (UPS)	HM060734 NORDIN et al. 2010
<i>Aspicilia cinerea</i>	Sweden, Dalarna	Hermansson 13275 (UPS)	HM060733 NORDIN et al. 2010
<i>Aspicilia dudinensis</i>	Sweden, Torne Lappmark	Nordin 6036 (UPS)	HM060748 NORDIN et al. 2010
<i>Aspicilia dudinensis</i>	Sweden, Torne Lappmark	Nordin 5971 (UPS)	HM060757 NORDIN et al. 2010
<i>Aspicilia epiglypta</i>	Sweden, Västergötland	Nordin 6303 (UPS)	HM060756 NORDIN et al. 2010
<i>Aspicilia epiglypta</i>	Sweden	Nordin 7037 (UPS)	MH255589 ZAKERI 2018
<i>Aspicilia laevata</i>	Sweden, Uppland	Tibell 23659 (UPS)	HM060730 NORDIN et al. 2010
<i>Aspicilia laevata</i>	Finland, Alandia	Nordin 5846 (UPS)	HM060735 NORDIN et al. 2010
<i>Aspicilia nigromaculata</i>	Pakistan	LAH36941	MW969630 This paper
<i>Aspicilia nikrapensis</i>	Svalbard	Ebbestad SVL2:1 (UPS)	HM060759 NORDIN et al. 2010
<i>Aspicilia nikrapensis</i>	Svalbard	Ebbestad SVL2:1 (UPS)	HM060759 NORDIN et al. 2010
<i>Aspicilia supertegens</i>	Sweden, Torne Lappmark	Nordin 6023 (UPS)	HM060751 NORDIN et al. 2010
<i>Aspicilia supertegens</i>	Norway, Troms	Owe-Larsson 9002 (UPS)	HM060742 NORDIN et al. 2010
<i>Circinaria caesiocinerea</i>	Sweden, Uppland	Tibell 22612 (UPS)	HM060731 NORDIN et al. 2010
<i>Circinaria emiliae</i>	Kazakhstan, Atyrau	Kulakov 3702 (UPS)	HM060728 NORDIN et al. 2010
<i>Circinaria emiliae</i>	Kazakhstan, Atyrau	Kulakov 3798 (UPS)	HM060729 NORDIN et al. 2010
<i>Circinaria gibbosa</i>	Sweden, Uppland	Nordin 5878 (UPS)	HM060740 NORDIN et al. 2010
<i>Circinaria hispida</i>	Turkey, Malatya	Candan 11 (ANES)	HM060760 NORDIN et al. 2010
<i>Circinaria leprosescens</i>	Sweden, Uppland	Nordin 5906 (UPS)	HM060749 NORDIN et al. 2010
<i>Oxneriaria dendroplaca</i>	Sweden, Torne Lappmark	Nordin 5952 (UPS)	HM060744 NORDIN et al. 2010
<i>Oxneriaria dendroplaca</i>	Finland, Enontekiö Lappi	Nordin 6366 (UPS)	HM060758 NORDIN et al. 2010
<i>Oxneriaria haeyrenii</i>	Sweden, Torne Lappmark	Nordin 5997 (UPS)	HM060755 NORDIN et al. 2010
<i>Oxneriaria mashiginensis</i>	Norway, Troms	Nordin 5708 (UPS)	HM060732 NORDIN et al. 2010
<i>Oxneriaria permutata</i>	Sweden, Torne Lappmark	Nordin 6027 (UPS)	HM060747 NORDIN et al. 2010
<i>Oxneriaria verruculosa</i>	Norway, Troms	Owe-Larsson 9007 (UPS)	HM060741 NORDIN et al. 2010

Aspicilia, five sequences of *Oxneriaria* S.Y. Kondr. & Lököös and two sequences of *Circinaria* Link were available from the literature (see Table 1 for voucher details). The aligned ITS1-5.8S-ITS2 region comprised 574 sites, of which 359 were conserved and 195 variable; 134 sites were parsimony informative. The tree resulting from the analyses of the ITS region is shown in Fig. 1. *Aspicilia nigromaculata* forms a well-supported clade with *A. subradians* (Nyl.) Hue (HQ259268, HQ259267), *A. fluviatilis* A. Nordin & Owe-Larss. (HQ259264), *A. granulosa* A. Nordin (HQ259265) and *A. epiglypta* (Norrl. ex Nyl.) Hue (HQ259261, EU057907). *Ochrolechia parella* (L.) A. Massal. (KJ607905) was selected for rooting the tree (PAUKOV & DAVYDOV 2020).

LSU sequences were newly prepared for one specimen (see Table 2 for voucher details). The tree resulting from the analyses of the LSU region is shown in Fig. 2. The Pakistan sequence is recovered as sister to *Aspicilia epiglypta* (GenBank no. MH255589, HM060756) (NORDIN et al. 2011).

Taxonomic treatment

Aspicilia nigromaculata Fayyaz, Afshan, Niazi & Khaliid sp. nov. (Figs. 1-3)

Diagnosis. A well-developed, grey to greenish grey frequently black spotted thick thallus, well-developed thicker peripheral and flat to slightly concave areoles, and small conidia 10–15 × 0.5–1 µm; differing from related species in the ITS and nrLSU region.

MycoBank No: MB839363

Holotype. PAKISTAN: Azad Jammu and Kashmir, Jhelum Valley (N 33°23', E 73°32'), 2500 m alt., on rock, 24 September 2020, Najam-ul-Sehar Afshan and Abdul Rehman Niazi (JV-07) (LAH36940—holotype), ITS GenBank accession number MW969628 LSU, Genbank accession number MW969630.

Thallus: crustose, areolate to rimose areolate, immarginate, frequently black spotted, thick, 8 to 10 cm wide,

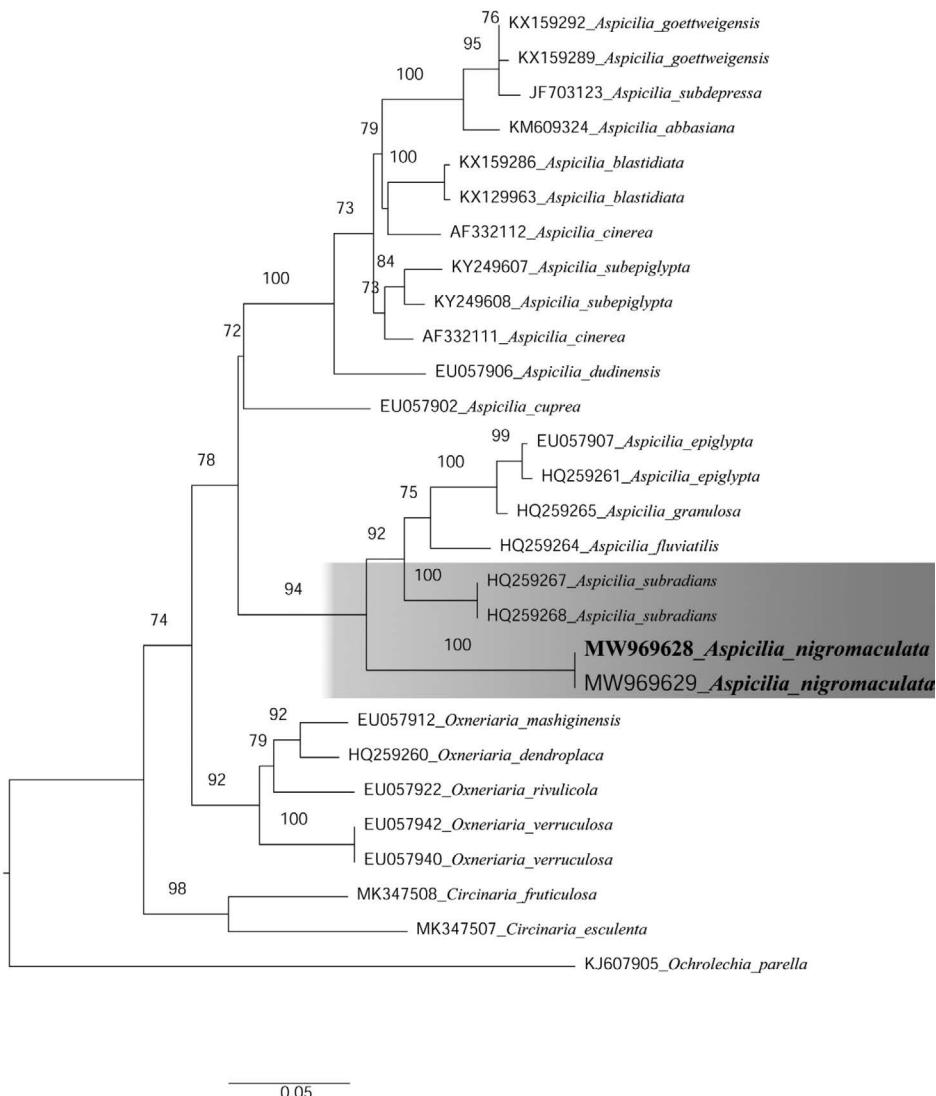


Fig. 1. Most likely phylogenetic relationship of *Aspicilia nigromaculata* with their associated taxa inferred from the nrITS region on rooting with *Ochrolechia parella* (L.) A. Massal. (KJ607905) as an outgroup.

epruinose. **Areoles:** 0.5 to 2 mm wide and 1 mm thick, thinner centrally but thicker towards the periphery, broadly attached to the stipitate, separated by distinct cracks, discrete to rarely contiguous, smooth to thinly wrinkled, angular to irregular, replicating by divisions, flat to slightly concave. **Upper surface:** grey to greenish grey. **Lower surface:** brown to black. **Prothallus:** not seen. **Upper cortex:** 20–30 µm thick, smooth, upper layer dark brown, hyaline adjacent to algal layer, paraplectenchymatous, *textura globularis*, cells 8–12 µm in diam. **Algal layer:** continuous, 40 to 60 µm thick. **Photobiont cells:** globose to sub-globose, 13–24 µm. **Medulla:** 60–110 µm thick, dark brown, hyphae prosoplectenchymatous 1–3 µm wide. **Pycnidia:** immersed, rare, indistinct, often black at the ostiole, 0.1–0.4 mm. **Conidiophores:** 13–15 × 2–2.5 µm. **Conidia:** filiform, acrogenous, straight, hyaline, 10–15 × 0.5–1 µm.

Chemistry—Cortex: K+ (yellowish-green), C-, KC-. **Medulla:** K-, C-, KC-, no substance detected by TLC.

Habitat and distribution. The known collections of the new species are from the Himalayan moist temperate forest of Azad Jammu and Kashmir, Pakistan. The specimens were found on siliceous rocks. The coniferous forest is dominated by species of *Pinus roxburghii* Sarg., *Quercus oblongata* D. Don, *Quercus glauca* Thunb and *Pyrus pashia* L. etc. The maximum daily temperature of the region varies from 20 to 30°C during the summer with an average temperature of 4°C during the winter, and there is moderate rainfall.

Etymology. The species epithet “*nigromaculata*” refers to the black-spotted thallus of the type.

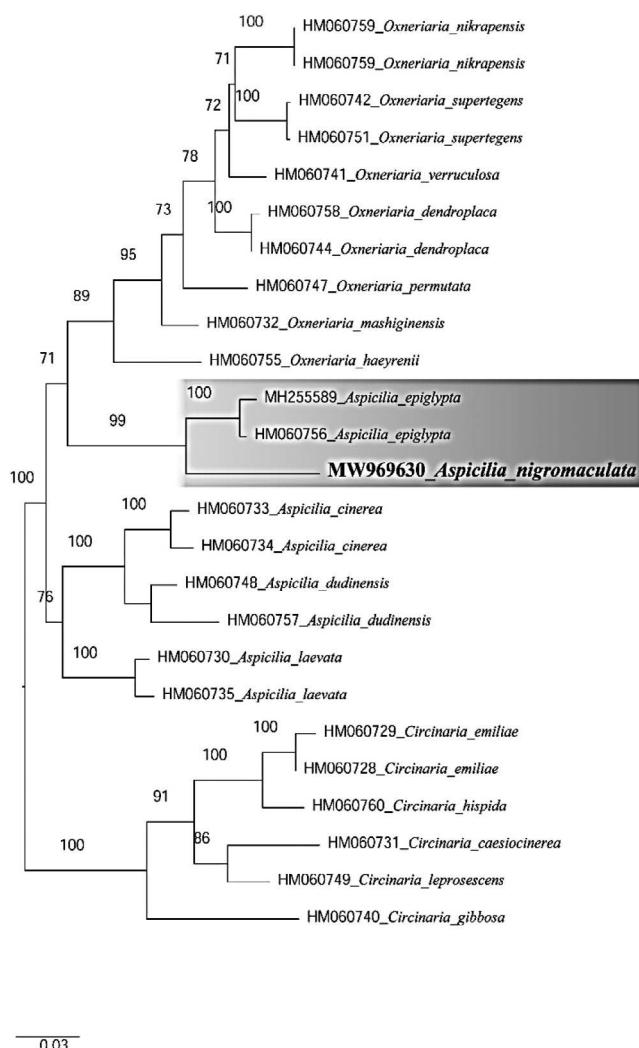


Fig. 2. Most likely phylogenetic relationship of *Aspicilia nigromaculata* with their associated taxa inferred from the nrLSU region.

Additional specimens examined. PAKISTAN. Azad Jammu and Kashmir, Muzaffarabad, Peer Chanasi (N 34°23', E 73°32', 2000 to 2924 m alt.), on rock, 26 September 2020, Najam-ul-Sehar Afshan and Abdul Rehman Niazi, (JV-07H) (LAH36941) (ITS GenBank accession number MW969629).

DISCUSSION

Two collections of this taxon were made from Peer Chanasi and Muzaffarabad (Azad Jammu & Kashmir). The two sites vary significantly in terms of elevation, vegetation and climatic conditions. However, the phylogenetic analyses confirmed that the two collections of *Aspicilia nigromaculata* form a monophyletic clade with good support (Figs. 1 & 2).

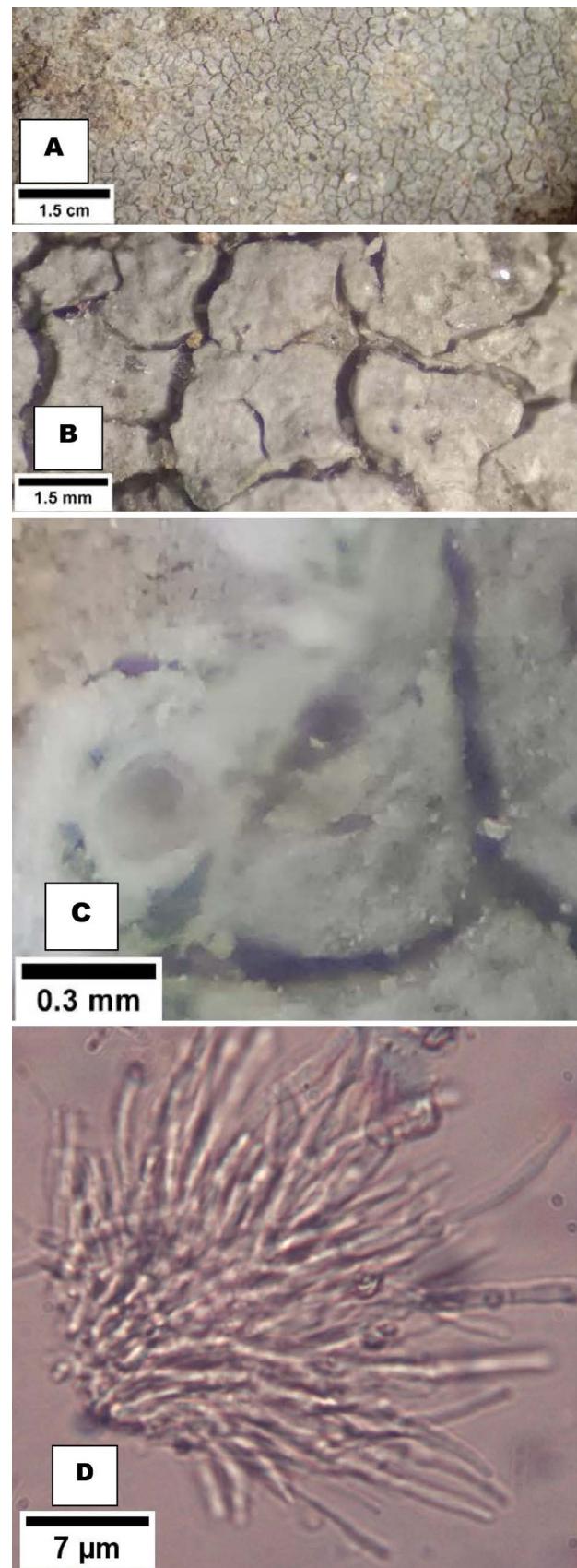


Fig. 3. *Aspicilia nigromaculata* Fayyaz, Afshan, Niazi & Khalid sp. nov. A. Thallus of the specimen, B. Areoles of the specimen, C. Pycnidia, D. Conidiomata

The areoles of *Aspicilia nigromaculata* are flat to slightly concave peripheral areoles and no apothecia were found in the present collections. The other species of *Aspicilia* have mostly convex areoles and apothecia are common. It shows certain morpho-anatomical differences from *A. subradians* i.e. in having flat to slightly convex well developed peripheral areoles, smaller conidia which are 10–15 µm and the prothallus is absent in our specimens (vs. marginal areoles poorly developed, central areoles sometimes convex and a distinct prothallus) (NORDIN *et al.* 2011). The new species differs from *Aspicilia fluviatilis* in having a thick, frequently black spotted, greenish grey thallus, a 20–34 µm thick cortex (vs. thin, often minutely white spotted, grey to ochraceous gray thallus, a 22–43 µm thick cortex). Similarly, the Pakistani collection also differs from *A. granulosa* in having a thick, greenish grey thallus (vs. a thin, green grey, brown grey to brownish thallus, and in lacking vegetative propagules (NORDIN *et al.* 2011). In the phylogenetic analyses the Pakistani collection also forms a separate subclade from the other *Aspicilia* species, suggesting it is not conspecific with any of the above-mentioned species.

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REFERENCES

- DAR M, COCHARD R, SHRESHTA R & AHMAD S. 2012. Plant resource utilization by local inhabitants around Machiara National Park, Azad Kashmir, Pakistan. *Journal of Food, Agriculture and Environment* **10**(3–4): 1139–1148.
- GARDES M & BRUNS TD. 1993. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. *Molecular Ecology* **2**(2): 113–118.
- HAJI MONIRI M, GROMAKOVA AB, LÓKÖS L & KONDRAKYUK SY. 2017. New members of the Megasporaceae (Pertusariales, lichen-forming Ascomycota): *Megaspore iranica* spec. nova and *Oxneriaria* gen. nova. *Acta Botanica Hungarica* **59**: 343–370.
- HALL TA. 1999. Bioedit: A user-friendly biological sequence alignment editor and analysis program, for windows 95/98/NT. *Nucleic Acid Symposium Series* **41**: 95–98.
- ISMAYIL G, ABBAS A & GUO S-Y. 2015. *Aspicilia volcanica*, a new saxicolous lichen from Northeast China. *Mycotaxon* **130**(2): 543–548.
- IVANOVA NV & HAFELLNER J. 2002. Searching for the correct placement of *Megaspore* by use of ITS1, 5.8 S and ITS2 rDNA sequence data. *Bibliotheca Lichenologica* **82**: 113–122.
- KHAN M, KHALID AN & LUMBSCH HT. 2018. A new species of *Lecidea* (Lecanorales, Ascomycota) from Pakistan. *MycoKeys* **38**: 25.
- KONDRAKYUK SY, LÓKÖS L, PARK J-S, JANG S-H & JEONG M-H. 2016. New *Aspicilia* species from South Korea proved by molecular phylogeny with a key to the Eastern Asian aspicilioid lichens. *Studia Botanica Hungarica* **47**(2): 227–249.
- LÖYTINOJA A & GOLDMAN N. 2010. webPRANK: a phylogeny-aware multiple sequence aligner with interactive alignment browser. *BMC Bioinformatics* **11**(1): 1–7.
- MILLER MA, PFEIFFER W & SCHWARTZ T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. *Proceedings of the Gateway Computing Environments Workshop (GCE)*, 14 November 2010, New Orleans, LA, pp. 1–8.
- NORDIN A, OWE-LARSSON B & TIBELL L. 2011. Two new *Aspicilia* species from Fennoscandia and Russia. *Lichenologist* **43**(1): 27–37.
- NORDIN A, SAVIĆ S & TIBELL L. 2010. Phylogeny and taxonomy of *Aspicilia* and Megasporaceae. *Mycologia* **102**: 1339–1349.
- NORDIN A, TIBELL L, OWE-LARSSON B. 2007. A preliminary phylogeny of *Aspicilia* in relation to morphological and secondary product variation. In: FRISCH A, LANGE U & STAIGER B (eds.), *Lichenologische Nebenstunden. Contributions to Lichen Taxonomy and Ecology in Honour of Klaus Kalb*. *Bibliotheca Lichenologica* **96**, pp. 247–266, J. Cramer in der Gebrüder Borntraeger Verlagsbuchhandlung, Berlin-Stuttgart.
- ORANGE A, JAMES PW & WHITE FJ. 2010. *Microchemical methods for the identification of lichens*. British Lichen Society.
- PARK CH, KIM KM, ELVEBAKK A, KIM OS, JEONG G & HONG SG. 2015. Algal and fungal diversity in Antarctic lichens. *Journal of Eukaryotic Microbiology* **62**(2): 196–205.
- PAUKOV A & DAVYDOV EA. 2020. *Aspicilia stalagmitica* (Megasporaceae) A new lichen species with isidia-like thalline outgrowths. *Turczaninowia* **23**(1): 5–12.
- PAUKOV A, DAVYDOV EA, NORDIN A, ROUX C, ŞENKARDEŞLER A, SOHRABI M, VONDRAK J, FROLOV IV, TEPTINA AY & SHIRYAEVA AS. 2019. Three new species, new combinations and a key to known species of *Lobothallia* (Megasporaceae). *The Lichenologist* **51**: 301–322.
- PAUKOV A, NORDIN A, TIBELL L, FROLOV I, VONDRAK J. 2017. *Aspicilia goettweigensis* (Megasporaceae, lichenized Ascomycetes) a poorly known and overlooked species in Europe and Russia. *Nordic Journal of Botany* **35**(5): 595–601.
- PAUKOV AG, YU TEPTINA A, PUSHKAREV EV. 2015. Heavy metal uptake by chemically distinct lichens from *Aspicilia* spp. growing on ultramafic rocks. *Australian Journal of Botany* **63**(2): 111–118.
- POSADA D. 2008. jModelTest: phylogenetic model averaging. *Molecular Biology and Evolution* **25**(7): 1253–1256.
- RAMBAUT A, SUCHARD MA, XIE D & DRUMMOND AJ. 2014. *FigTree 1.4. 2 Software*. Edinburgh: Institute of Evolutionary Biology. University of Edinburgh.
- ROUX C, NORDIN A, TIBELL L & SOHRABI M. 2011. Quelques espèces d'*Aspicilia* peu connues ou nouvelles des Pyrénées-Orientales (France). *Bulletin de la Société Linnéenne de Provence* **14**: 177–227.
- SAMBROOK J & RUSSEL DW. 2001. Rapid isolation of yeast DNA. In: SAMBROOK J & RUSSEL DW (eds.), *Molecular cloning: A laboratory manual*, pp. 631–632, Cold Spring Harbor Laboratory Press, New York.
- SOHRABI M, AHTI T & LITTERSKI B. 2011. *Aspicilia digita* sp. nov., a new vagrant lichen from Kyrgyzstan. *Lichenologist* **43**(1): 39–46.
- SOHRABI M, LEAVITT SD, HALICI MG, SHRESTHA G & STENROOS S. 2013b. *Teuvoa*, a new lichen genus in Megasporaceae (Ascomycota: Pertusariales), including *Teuvoa junipericola* sp. nov. *The Lichenologist* **45**: 347–360.

- SOHRABI M, STENROOS S, MYLLYS L, SØCHTING U, AHTI T & HYVÖNEN J. 2013c. Phylogeny and taxonomy of the ‘manna lichens’. *Mycological Progress* **12**(2): 231–269.
- SOHRABI M, STENROOS S, MYLLYS L, SØCHTING U, AHTI T & HYVÖNEN J. 2013a. Phylogeny and taxonomy of the “manna lichens.” *Mycological Progress* **12**(2): 231–269.
- STAMATAKIS A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**(9): 1312–1313.
- VILGALYS R & HESTER M. 1990. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* **172**(8): 4238–4246.
- WHITE TJ, BRUNS T, LEE SJWT & TAYLOR J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR Protocols: a Guide to Methods and Applications* **18**(1): 315–322.
- ZAKERI Z, DIVAKAR PK & OTTE V. 2017. Taxonomy and phylogeny of *Aspiciliella*, a resurrected genus of Megasporaceae, including the new species *A. portosantana*. *Herzogia* **30**: 166–176.

REZIME



***Aspicilia nigromaculata* sp. nov. (Megasporaceae, Pertusariales, Ascomycota) iz Azad Jammua i Kashmira, Pakistan, dokazi iz morfologije i podataka o sekvencama DNK**

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Nova vrsta u okviru Megasporaceae, *Aspicilia nigromaculata* sp. nov. je opisana iz Pakistana. Uporedne morfo-anatomske i molekularne analize zasnovane na ITS–LSU potvrđuju njegovu poziciju u okviru roda *Aspicilia*. Od drugih vrsta roda razlikuje se po sledećem skupu karakteristika: dobro razvijenim sivim do zelenkasto sivim, često crno pegavim debelim talusom, dobro razvijenim debljim perifernim i ravnim do blago konkavnim areolima i malim konidijama veličine 10–15 × 0.5–1 µm. Različitim od drugih poznatih vrsta roda ga čini i položaj u izdvojenoj grani na filogenetskom stablu.

Ključne reči: ITS nrDNA, nrLSU, filogenija, saksikolozni lišajevi

