Candelariella blastidiata sp. nov. (Ascomycota, Candelariaceae) from Eurasia and North America, and a key for grey thalli Candelariella

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Abstract: Candelariella blastidiata Yakovchenko sp. nov. is described. This corticolous species is characterized by biatorine yellow apothecia, a grey squamulose thallus with marginal and lower side blastidia, 8-spored asci, and a northern circumpolar distribution. Candelariella subdeflexa has previously been confused with C. blastidiata, but our analyses of phenotypic and DNA sequence data revealed C. blastidiata should be distinguished from C. subdeflexa. A worldwide key for Candelariella species with grey thalli is provided.

Key words: Candelariales, Candelariella subdeflexa, lichens, new species, species pair, taxonomy

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Introduction

Candelariaceae Hakul. is characterized within Lecanoromycetes primarily by the foliose or crustose thallus with pulvinic acid and related pigments which give the Candelariaceae its distinctive yellow to orange-yellow colour. The family has a worldwide distribution and consists of four genera with the largest, Candelariella Müll. Arg., comprising c. 50 accepted species. Among the species of Candelariella, a small group can be recognized that share the following features: a grey thallus, lacking lichen substances detected by

high performance liquid chromatography (HPLC) and thin-layer chromatography (TLC), 8-spored asci and mostly lecanorine apothecia (Westberg 2005). The representatives of this group are predominantly found in open woodlands, steppes and prairies growing on bark, wood, plant debris or calcareous rocks, more often in dry and sun-exposed habitats. Such Candelariella species with a grey thallus are now frequently reported by many researchers (e.g. Malíček et al. 2008; Vondrák et al. 2008; Aptroot 2011; Westberg & Clerc 2012; Westberg & Sohrabi 2012; Yakovchenko et al. 2012; Gasparyan et al. 2014; Urbanavichus & Urbanavichene 2014; Kocakaya & Halici 2015).

Among the species with a grey thallus, Candelariella subdeflexa (Nyl.) Lettau is distinct in its biatorine apothecia and well-developed thallus formed by squamules with occasional conidiophores producing globose conidia on the lower side of the thallus (Westberg 2007). During a revision of Candelariaceae in the herbaria H, LE and TNS, the first author found several specimens identified as Candelariella subdeflexa with blastidiate squamules. The aim of this study is to compare these individuals with

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other related taxa employing morphological, chemical and DNA sequence data. A world-wide key to grey thalline *Candelariella* is also presented.

Materials and Methods

Specimens and phenotype studies

The specimens examined are housed in ALTB, G, GZU, H, LE, PRA, SMR and TNS. Morphological observations were made using a dissecting microscope. Hand-cut sections of apothecia and thalli were observed after mounting in water. Measurements are presented as the mean (\bar{x}) and range including 85% of the variation, bounded by the smallest and largest observed values, followed by the sample size (n) following Westberg (2005).

Lichen substances were examined using thin-layer chromatography (TLC) with solvent B' (hexane: methyl tertbutyl ether: formic acid, 140: 72: 18) and solvent C (toluene: acetic acid, 170: 30) (Culberson & Kristinsson 1970; Culberson & Johnson 1982) for the specimen in TNS.

Sequences and phylogenetic reconstructions

To test the monophyly of Candelariella blastidiata and its relationship with other species, ITS nrDNA sequences of fresh material and those retrieved from the NCBI database (GenBank) were used for molecular phylogenetic analysis. ITS nrDNA sequence data were used because GenBank includes a number of ITS sequences of Candelariella, whereas other loci are poorly represented. Furthermore, ITS single-locus genealogies are usually consistent with phenotypic data (Westberg et al. 2007, 2009; Westberg & Arup 2011; Tripp & Lendemer 2015). One or more different sequences of appropriate length (and preferably lacking uncertain nucleotides) were selected from every species of Candelariella. Our sampling comprised 26 species representing morphological variability within Candelariella, 10 of which were represented by two or more accessions. Information on the samples with GenBank Accession numbers is shown in Table 1.

Two apothecia or fragments of thallus (for sterile specimens) were used for DNA extraction. Samples were frozen in liquid nitrogen and made into a powder in tubes using sterile pestles. The DNeasy Plant Mini Kit (Qiagen, Germany) was used for DNA extraction following the manufacturer's instructions. The primer set was ITS1F (Gardes & Bruns 1993) and LR1 (Vilgalys & Hester 1990). PCR settings were as follows: 95 °C for 2 min, 35 cycles of 94 °C for 20 s, 53 °C for 60 s, 72 °C for 2 min, followed by 72 °C for 15 min and then maintained at 14 °C. The sequences obtained in this study and the reference sequences from GenBank were aligned by Geneious 6.0 (Biomatters Ltd., New Zealand) and manually optimized. The sequence of Pycnora xanthococca (Sommerf.) Hafellner was used as outgroup since Pycnora has been identified as a possible sister clade

to Candelariaceae (Bendiksby & Timdal 2013). Total alignment of 461 bp was used for the maximum likelihood (ML) and Bayesian analyses. Optimal substitution models were inferred separately for ITS1, 5.8S and ITS2 using PartitionFinder version 1.1.1 (Lanfear et al. 2012). The symmetrical substitution model with gamma-distribution (SYM+G) was selected for the 5.8S partition and Kimura 2-parameter with proportion of invariable sites (K80+I) for ITS1+ITS2. Bayesian inference with the Markov chain Monte Carlo (BMCMC) method (Larget & Shimon 1999) was performed using MrBayes 3.2.3 (Ronquist et al. 2012). Three parallel Bayesian analyses were run for 20 000 000 generations in six chains and every 200th generation was sampled. The first 50% of trees were discarded as burn-in and a 50% majority-rule consensus tree calculated from the remaining trees of three runs with the sumt command implemented in MrBayes 3.2.3. Bootstrap support values and BMCMC posterior probability were noted on the best scoring tree (numbers are shown only when ≥50% in both). The ML tree and 1000 bootstrap replicates were calculated using RAxML 8.0.26 (Stamatakis 2014) by raxmlGUI software version 1.3.1 (Silvestro & Michalak 2012), applying the GTRGAMMA model of substitution because RAxML does not support SYM and K80 models.

Results

Six ITS sequences of blastidiate specimens of *Candelariella* were successfully obtained. They formed a monophyletic clade in the ITS phylogeny of *Candelariella* with support values of 100/1·00 (bootstrap/posterior probability) in the phylogenetic tree (Fig. 1). Variability in ITS sequences among four of the European and Russian specimens is only 1 residue, but the Chinese sequence differs from others by 3 residues. A supported sister clade (88/1·00) includes three sequences of *C. subdeflexa*. Four constant residues differentiate *C. subdeflexa* from blastidiate specimens. Both species form a group well-separated from all other included species.

The close relationship of blastidiate specimens with *C. subdeflexa* is well explained by the close morphology (i.e. grey thallus with biatorine apothecia) as well as growing in similar habitats. However, we consider the two clades as distinct and morphologically recognizable species and one of them is described here as *Candelariella blastidiata* Yakovchenko. Specimens of *C. blastidiata* studied include samples without conidiophores (European specimens) and

Candelariella blastidiata-

Species	Locality	Voucher (Herbarium)	GenBank Acc. no.	Reference
Candelariella aggregata	USA: Colorado	Westberg 1080 (LD)	EF535157	Westberg et al. (2007)
C. antennaria	USA: Colorado	Westberg 1155 (LD)	EF535159	Westberg et al. (2007)
C. aurella	USA: Colorado	Westberg 1053 (LD)	EF535162	Westberg et al. (2007)
C. aurella	USA: Arizona	Westberg 150 (LD)	EF535163	Westberg et al. (2007)
C. biatorina	USA: California	Tucker & Bratt 34049 (SBBG)	EF535165	Westberg et al. (2007)
C. blastidiata	Russia: Repub. of Altai	Davydov 7716 (ALTB)	KX853128	This study
C. blastidiata	Russia: Orenburg Region	Vondrák 7428 & Khodosovtsev (TNS)	KX853127	This study (isotype)
C. blastidiata	Russia: Orenburg Region	Vondrák 7428 & Khodosovtsev (LE-L11031)	KX853123	This study (holotype)
C. blastidiata	Slovakia: Filakovo	Vondrák 10207 (PRA)	KX853125	This study
C. blastidiata	Slovakia: Revúca	Vondrák 9195 (PRA)	KX853124	This study
C. blastidiata	China: Xinjiang	Davydov 12272 (ALTB)	KX853126	This study
C. borealis	Canada: Nunavut	Westberg 2381 (LD)	EF535167	Westberg et al. (2007)
C. borealis	USA: Colorado	Westberg 1079 (LD)	EF535168	Westberg et al. (2007)
C. californica	USA: California	Westberg 1244 (LD)	EF535169	Westberg et al. (2007)
C. citrina	Mexico: Baja California Sur	Westberg 398 (LD)	EF535170	Westberg et al. (2007)
C. clarkii	USA: Colorado	Tripp & D'az 4876 (COLO)	KR052104	Tripp & Lendemer (2015)
C. complanata	Mexico: Baja California Sur	Westberg 383 (LD)	EF535174	Westberg et al. (2007)
C. coralliza	Sweden: Öland	Westberg 51 (LD)	EF535175	Westberg et al. (2007)
C. coralliza	China: Yanbian Korean Autonomous Prefecture	YK140035	KP226207	Cao et al. (2015)
C. corallizoides	Mexico: Baja California Sur	Westberg 341 (LD)	EF535176	Westberg et al. (2007)
C. corviniscalensis	USA: Colorado	Morse 15881 & Ladd (S)	GU967377	Westberg et al. (2011)
C. deppeanae	USA: Arizona	Westberg 585 (holotype) (LD)	EF535178	Westberg et al. (2007)
C. deppeanae	Mexico: Chihuahua	Nash 36526 (ASU)	EF535179	Westberg et al. (2007)
C. efflorescens	Austria: Steiermark	Arup L97286 (hb. Ulf Arup)	EF535180	Westberg et al. (2007)
C. granuliformis	Canada: Nunavut	Mattsson 5209 (UPS)	GU967375	Westberg et al. (2011)
C. kansuensis	USA: Arizona	Wetmore 55470 (MIN)	EF535181	Westberg et al. (2007)
C. kansuensis	USA: Arizona	Westberg s. n. (LD)	FJ959349	Westberg et al. (2009)
C. lutella	Norway: Troms	Westberg 2808 (LD)	EF535182	Westberg et al. (2007)
C. lutella	USA: Arizona	Westberg 206 (LD)	EF535183	Westberg et al. (2007)
C. medians	Sweden: Skåne	Arup L03165 (LD)	EF535184	Westberg et al. (2007)
C. placodizans	Canada: Nunavut	Westberg 206 (LD)	EF535187	Westberg et al. (2007)
C. placodizans	USA: Colorado	Westberg 1083 (LD)	EF535188	Westberg et al. (2007)
C. reflexa	Norway: Vestfold	Lindblom & Blom L61 (BG)	EF535190	Westberg et al. (2007)
C. rosulans	USA: Colorado	Westberg 1146 (LD)	EF535191	Westberg et al. (2007)
C. rosulans	USA: Arizona	Westberg 812 (LD)	EF535192	Westberg et al. (2007)
C. rosulans	USA: Colorado	Westberg 199 (LD)	EF535193	Westberg et al. (2007)
C. spraguei	USA: Colorado	Westberg 1037 (LD)	EF535194	Westberg et al. (2007)

Table 1 (continued)

Species	Locality	Voucher (Herbarium)	GenBank Acc. no. Reference	Reference
C. subdeflexa	USA: South Dakota	$Lendemer\ 1164\ (PH)$	EF535196	Westberg et al. (2007)
C. subdeflexa	USA: Arizona	Nash 38631 (ASU)	EF535197	Westberg et al. (2007)
C. subdeflexa	USA: Arizona	Westberg 660 (LD)	EF535198	Westberg et al. (2007)
C. terrigena	Canada: Nunavut	Lutzoni & Miadlikowska 07.13.03-19 (DITKF)	HQ650602	Schmull et al. (2011)
C. vitellina	USA: Oregon	Westberg 875 (LD)	EF535199	Westberg et al. (2007)
C. vitellina	Sweden: Öland	Westberg 49 (LD)	EF535200	Westberg et al. (2007)
C. xanthostigma	USA: California	Nash 32596 (ASU)	EF535201	Westberg et al. (2007)
C. xanthostigma	USA: Colorado	Westberg 1122 (LD)	EF535202	Westberg et al. (2007)
Pycnora xanthococca	Norway	E. Timdal 11646 (O L-163707)	KF360412	Bendiksby & Timdal (2013)

samples with a poorly developed layer of conidiophores at the base of the squamules (Altaian specimens: Russia, ED 7716; China, ED 12272). The Chinese specimen also shows slight morphological differences, with larger squamules and more abundant blastidia (Fig. 2A).

Candelariella blastidiata Yakovchenko sp. nov.

MycoBank No.: MB 817756

Thallus corticolous, grey, squamulose with blastidia on margins and/or the lower side of squamules 25–34 (–60) µm wide, apothecia biatorine, convex, yellow, (0.3-)0.5(-0.9) mm in diam; apothecial margin indistinct. Asci 8-spored, ascospores narrowly-ellipsoid, $(7.5-)1.0.0-1.4.5(-15.0) \times (3.5-)4.0-5.0$ µm, conidiophores absent or partly-developed on the lower side of the squamules.

Type: Russia, Orenburg Region, Buzuluk District, National Park 'Buzuluksky bor', Partizanskiy Village, bank of the Borovka River, 52°59'54"N, 52°07'42"E, 80 m, on bark of Salix, 2009, J. Vondrák JV7428 & A. Khodosovtsev [Obermayer, Lichenotheca Graecensis no. 405] (LE-L11031—holotype; ALTB, ASU, B, C, CANB, CANL, E, G, GZU, H, HAL, HMAS, M, MAF, MIN, O, PRA, TNS, UPS—isotypes labelled "C. subdeflexa").

(Figs 2A & B, 3A-C)

Thallus squamulose. Squamules first prostrate to soon ascending and erect, scattered to crowded and somewhat imbricate, ±rounded, (0.2-)0.3-0.4-0.5(-0.7) mm long (n = 29)with irregularly incised margin; squamules $(75-)103-122-141(-163) \mu m$ thick (n = 19); epinecral layer absent; cortex alveolate, (0–) 4–8–13(–25) μ m thick (n = 35), consisting of gelatinized hyphae with ±isodiametric cells, $(2.5-)4.0-5.5-7.0(-10.0) \mu m$ wide (n = 30); algae chlorococcoid, (18-)19-20-21(-25) μm diam. (n = 20), lower surface covered by blastidia and sometimes also with a layer of conidiophores at the base of the squamules; conidiophores $(15-)23-32-41(-50) \mu m$ (n = 11). Surface pale grey to pale brownish grey, smooth and matt. Vegetative propagules: $(25-)27-34-41(-60) \mu m$ blastidia, (n = 36), same colour as thallus or with greenish tint, marginal to spreading on the lower side of the squamules.

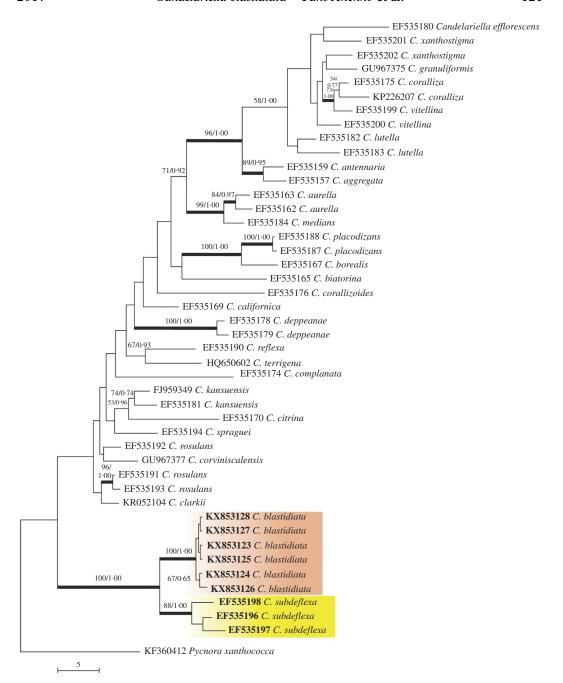


Fig. 1. Reconstruction of the Candelariella ITS phylogeny showing positions of C. blastidiata and C. subdeflexa. The tree was constructed by Maximum Liklihood (ML), and the reliability of each branch was tested by ML and Bayesian methods. Numbers at tree nodes indicate bootstrap values of ML (left) and BMCMC posterior probabilities (right) (only ML values ≥50% and PP values ≥0.50 are depicted). Thicker branches indicate where both the ML bootstrap value is ≥70% and the BMCMC posterior probability is ≥0.95. GenBank Accession numbers are given as OTU names (see Table 1). Pycnora xanthococca is the outgroup.

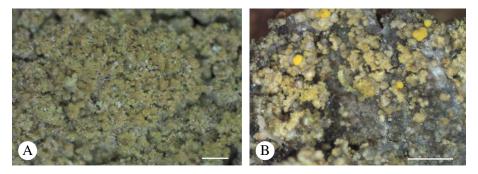


Fig. 2. Candelariella blastidiata. A, sterile specimen with abundant blastidia, China, Altai, ED 12272 (ALTB); B, isotype (ALTB). Scales: A = 200 µm; B = 500 µm. In colour online.

Apothecia biatorine, yellow, scattered to crowded and coalescing, (0.3-)0.4-0.5-0.5(-0.9) mm diam. (n = 32); disc first rounded and weakly convex, becoming irregular in outline and strongly convex with age; proper margin not visible, hidden below margin of the convex disc, of branched and anastomosing hyphae radiating towards the margin, consisting of ±rectangular cells c. $4.5-10.0 \times 1.5-2.5 \,\mu\text{m}$ (n = 10), ±similar epihymenium yellow-brown; throughout; hymenium colourless, c. 45·0-62·5 µm tall (n = 12); subhymenial layers colourless with oil droplets; paraphyses simple to branched, in midhymenium $1.5-2.5 \,\mu\text{m}$ wide (n = 15); tips cylindrical or weakly tapering, rarely narrowly clavate, up to 3 µm wide; asci clavate to widely-clavate, 8-spored, c. $30-50 \times 12.5-$ 15.0 μ m (n = 17); ascospores narrowly ellipsoid, with thin septa or rarely simple, straight slightly curved, (7.5-)10.0-12.0-14.5 $(-15.0) \times (3.5-)4.0-5.0-5.0 \,\mu\text{m} \ (n=42).$

Conidiogenous cells cylindrical to flask-shaped; conidia globose to widely teardrop-shaped, (2.5-)4.0-5.5-6.5(-7.5) µm diam. (n = 41). Pycnidia not seen.

Chemistry. Spot tests: apothecia K+ reddish, KC-, C-; thallus K-, KC-, C-. TLC: thallus, no substances; apothecia containing calycin, pulvinic acid, and pulvinic dilactone.

Etymology. The species is named with reference to the blastidia.

Ecology. Candelariella blastidiata was found on the bark of deciduous trees. It grows in open woodland or on solitary trees at elevations of 49–1050 m near rivers on trunks of Acer, Cercidium, Populus, Tilia and Salix. It often grows together with Candelariella antennaria Räsänen.

Distribution. Candelariella blastidiata is currently known from a small number of localities in Eurasia and North America: Central Europe, the European part of Russia (Orenburg, Samara Region), Siberia and the adjacent territory of China (Altai Mountains), and western North America (North Dakota and Arizona).

Additional specimens examined. Slovakia: Banská Bystrica Region: Revúca, Muráň, in village of Predná Hora, Muránská planina Mts, 48°46'21"N, 20°6'13"E, 760 m, on bark of Tilia cordata, 2011, J. Vondrák 9195 (PRA); Filakovo, Hajnáčka, Šurice, S-slope of hill Soví hrad, Cerová vrchovina upland, 48°13'34"N, 19°54'45"E, 250 m, on bark of solitary Acer campestre on sun-exposed rock, 2012, J. Vondrák 10207 (PRA).—Russia: Orenburg Region: Buzuluk District, National Park 'Buzuluksky bor', Koltu-Village, square 95, 52°53'03·13"N, 52°03'16·31"E, 64 m, on bark of Populus tremula and Salix alba, 2013, V. P. Travkin 167, 168 (SMR); same National Park, Borovo-opytniy Village, square 50, 52°59'52.87"N, 52°07'35.55"E, 81 m, on bark of Salix alba, 2015, V. P. Travkin 171 (SMR). Samara Region: Kinel District, Krasnosamarskiy Village, square 81, 52°59'45.61"N, 51°04'53.82"E, 49 m, on bark of Populus tremula, 2008, E. S. Korchikov 172, 173 (SMR). Republic of Altai: Ongudaisky District, Altai Mts, Terektinsky range, right bank of Yaloman River, 1 km upstream its mouth, on bark of Populus laurifolia, 50°31'27"N, 86°33'23"E, 770 m, 2009, E. A. Davydov 7716 (ALTB).— China: Xinjiang: Altai Mts, Kabinsky range, at 22 km

NNE of Keldynen-Bulak settlement, the valley of Terekty River, on bark of *Populus laurifolia*, 48°28'25"N, 86°41'29"E, 1050 m, 2005, *E. A. Davydov* 12272 (ALTB).—**USA:** *North Dakota:* Mercer County, Knife River Indian Villages National Historic Site, 0.5 mi NE of headquarters (50 mi NW of Bismarck), in woods along Knife River with dead green ash and box elder and small prairie knoll, sec. 33, T 145 N, R 84 W, on boulder, 47°20'23"N, 101°23'10"W, 1998, *C. M. Wetmore* 79964/51-2000 (GZU). *Arizona:* Pinal Co., between Boyce Thompson arboretum and Picket Post Mt., upper Sonoran Desert, 33°16'N, 111°10'W, 600–800 m, at the base of trunk of *Cercidium* sp., NE-exp. (very shady), 1988, *P. Clerc* 88/394 (G00295824).

Discussion

Candelariella blastidiata is easily recognized by the grey to brownish grey squamulose thallus, composed of prostrate to ascending and erect squamules with marginal blastidia spreading on the lower side of the squamules, the presence of biatorine apothecia with 8 spores per ascus and an absent or poorly developed layer of conidiophores at the base of the squamules (Fig. 3A & B).

The presence of biatorine apothecia and blastidia distinguishes *C. blastidiata* from other corticolous and lignicolous species with grey thalli: *Candelariella antennaria* Räsänen, *C. viae-lacteae* G. Thor & V. Wirth, *C. boikoi* Khodos. & S. Y. Kondr. and forms of *C. aurella* (Hoffm.) Zahlbr. with an eroded non-yellow thallus. It differs from *C. subdeflexa* by having blastidia and the absence of an epinecral layer. In addition, the layer of conidiophores on the lower side of the squamules is absent or poorly developed in *C. blastidiata*, whereas it is prominent in *C. subdeflexa* (Fig. 3D). In North America

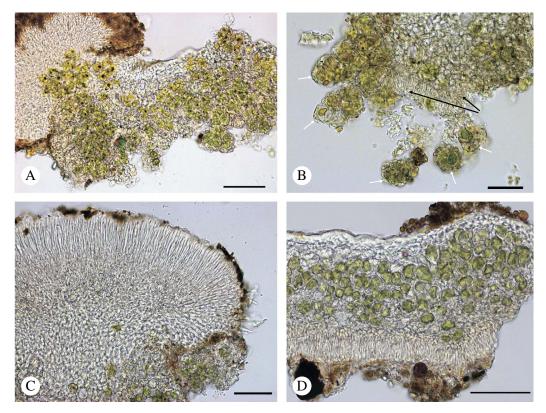


Fig. 3. A–C, Candelariella blastidiata, vertical sections through thallus; A, thallus with blastidia; B, lower part of thallus (white arrows, blastidia; black arrows, conidiophores); C, apothecium. D, Candelariella subdeflexa, vertical section through thallus with conidiophores on the lower side, lectotype (H–9506465). Scales: $A = 100 \,\mu m$; B–D = $50 \,\mu m$. In colour online.

both species occur together in North Dakota. They could be easily distinguished from each other morphologically: North American specimens of *C. subdeflexa* typically have a well-developed thallus composed of thick, shiny squamules forming a thick imbricate crust (Westberg & Clerc 2012) while *C. blastidiata* is distinguished by having flattened, matt squamules with marginal or lower-side blastidia.

Candelariella blastidiata can be found sterile and then its grey thallus is not likely to be identified as Candelariella. In a sterile state, C. blastidiata could be confused with C. viaelacteae G. Thor & V. Wirth. Candelariella antennaria with lecanorine apothecia might grow among squamules of sterile C. blastidiata and the combination of both species could be erroneously identified as C. viae-lacteae, which is characterized by lecanorine apothecia and a thallus composed of crowded granules resembling blastidia (Thor & Wirth 1990; Khodosovtsev et al. 2004; Khodosovtsev 2005). In such cases C. blastidiata is distinguished from C. viae-lacteae by permanent squamules while the thallus of C. viae-lacteae soon dissolves into granular blastidia. Some unrelated species (e.g. Caloplaca substerilis Vondrák et al.; Vondrák et al. 2013) might also be very similar to sterile crusts of Candelariella. blastidiata.

Morphological and molecular phylogenetic data support the close relationship of *C. subdeflexa* and *C. blastidiata* representing a so-called "species pair" (Poelt 1970) where *C. subdeflexa* reproduces by sexual propagules and *C. blastidiata* mostly by vegetative ones. Both species also possess asexual conidia as additional propagules; however, production of conidia appears to be less effective in *C. blastidiata*. Divergence by switching between reproductive modes has been shown for species in different families, such as *Parmeliaceae* (Poelt 1970; Miądlikowska *et al.* 2011), *Physciaceae* (Hafellner *et al.* 2012),

Teloschistaceae (Vondrák et al. 2016) and Umbilicariaceae (E. A. Davydov, D. Peršoh & G. Rambold, unpublished data), and often results in a reduction of sexual reproduction in species with vegetative propagules. As all our ITS sequences of C. blastidiata are from Eurasia and the two C. subdeflexa sequences are from North America, our results might be alternatively interpreted as an existence of two species that differ in geographical range and that have a blastidiate or non-blastidiate appearance. We cannot deny this possibility because of the limited availability of specimens.

Further specimens of *C. blastidiata* could probably be found amongst those labelled as "Candelariella subdeflexa" in herbaria. Specimens of *C. subdeflexa* from southern and central Europe (e.g. Austria, France, Germany, Italy, Spain and Switzerland) and from North Africa (Algeria) (Westberg & Clerc 2012) should be re-evaluated. The record from Russia (Obermayer 2012) is based on *C. blastidiata*, so *C. subdeflexa* should be excluded from the lichen flora of Russia.

Selected specimens of Candelariella subdeflexa examined. Algeria: Constantine: An Baumrinden am Wege gegen Balna bei Canstantine, 500 m, 1888, Flagey 1433 (H).—Afghanistan: Samangan: Kotal-i-Mirza Atbili Pass on the road from Pule-Khumri to Samangan (Aybak), N-facing slope, silt, at the base of a stump of *Pistacia vera*, 1250 m, 1972, P. Uotila 16702 (H).—USA: North Dakota: Ward County, 5.5 mi NNE of Sawyer (14 mi ESE of Minot), in grove of box elder (Acer negundo) and green ash (Fraxinus pennsylvanica) surrounded by cultivated fields, 48°06'N, 101°03'W, 470 m, 1979, T. D. Trana 9940 [distributed by C. M. Wetmore] 37-98 (GZU). Minnesota: Becker County, 1 mi N of Stinking Lake (18 mi NW of Detroit lakes), lone stream near farm with box elder and bur oak, sec. 18, T 140 N, R 43 W, 1978, C. M. Wetmore 35958 (H); Clay County, 8 mi NW of Hawley (18 mi NE of Moorhead), near gravel pit in grove of trees with box elder, cottonwood and willow, sec. 13, T 140 N, R 45 W, 1978, C. M. Wetmore 35984/233-81 (GZU).

Key to Candelariella with a grey thallus

1	On rocks, apothecia lecanorine	
	On bark, wood, plant debris, apothecia lecanorine or biatorine	

2(1)	Thallus indistinct to areolate with convex areoles, apothecia with yellow thalline margin, often partly excluded C. aurella (Hoffm.) Zahlbr. (grey thallus morphotype) Thallus distinct, areolate to squamulose, thalline margin persistent, greyish in the outer side
3(2)	Thallus areolate, thin; apothecia with thin, even thalline margin
	Thallus subsquamulose to squamulose, thick; apothecia with thick, crenulated and convoluted thalline margin
4(1)	On plant debris or shrubs in steppes and salt marshes
5(4)	Apothecia lecanorine
6(5)	Thallus uniformly composed of crowded spherical granules, unbranched to coralloid
7(6)	Thallus indistinct to amorphous crustose; thalline margin yellow, often greyish in the outer side, persistent; proper exciple does not form a stipe downward through the algal layer; on bark, rarely on wood
8(5)	Thallus without blastidia, often shiny, conidiophores on the lower side of the squamules present
	or poorly developed at the base of the squamules

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