

Neurons and Glia



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▼ INTRODUCTION

All tissues and organs in the body consist of cells. The specialized functions of cells and how they interact determine the functions of organs. The brain is an organ—to be sure, the most sophisticated and complex organ that nature has devised. But the basic strategy for unraveling its function is no different from that used to investigate the pancreas or the lung. We must begin by learning how brain cells work individually and then see how they are assembled to work together. In neuroscience, there is no need to separate *mind* from *brain*; once we fully understand the individual and concerted actions of brain cells, we will understand the origins of our mental abilities. The organization of this book reflects this “neurophilosophy.” We start with the cells of the nervous system—their structure, function, and means of communication. In later chapters, we will explore how these cells are assembled into circuits that mediate sensation, perception, movement, speech, and emotion.

In this chapter, we focus on the structure of the different types of cells in the nervous system: *neurons* and *glia*. These are broad categories, within which are many types of cells that differ based on their structure, chemistry, and function. Nonetheless, the distinction between neurons and glia is important. Although there are many neurons in the human brain (about 100 billion), glia outnumber neurons by tenfold. Based on these numbers, it might appear that we should focus our attention on glia for insights into the cellular functions of the nervous system. However, neurons are the most important cells for the unique functions of the brain. It is the **neurons** that sense changes in the environment, communicate these changes to other neurons, and command the body’s responses to these sensations. **Glia**, or **glial cells**, are thought to contribute to brain function mainly by insulating, supporting, and nourishing neighboring neurons. If the brain were a chocolate-chip cookie and the neurons were chocolate chips, the glia would be the cookie dough that fills all the other space and ensures that the chips are suspended in their appropriate locations. Indeed, the term *glia* is derived from the Greek word for “glue,” giving the impression that the main function of these cells is to keep the brain from running out of our ears! As we shall see later in the chapter, the simplicity of this view is probably a good indication of the depth of our ignorance about glial function. However, we still are confident that neurons perform the bulk of information processing in the brain. Therefore, we will focus 90% of our attention on 10% of brain cells: the neurons.

Neuroscience, like other fields, has a language all its own. To use this language, you must learn the vocabulary. After you have read this chapter, take a few minutes to review the key terms list and make sure you understand the meaning of each term. Your neuroscience vocabulary will grow as you work your way through the book.

▼ THE NEURON DOCTRINE

To study the structure of brain cells, scientists have had to overcome several obstacles. The first was the small size. Most cells are in the range of 0.01–0.05 mm in diameter. The tip of an unsharpened pencil lead is about 2 mm across; neurons are 40–200 times smaller. (For a review of the metric system, see Table 2.1.) This size is at or beyond the limit of what can be seen by the naked eye. Therefore, progress in cellular neuroscience was not possible before the development of the compound microscope in the late seventeenth century. Even then, obstacles remained. To observe brain tissue

Table 2.1 Units of Size in the Metric System

UNIT	ABBREVIATION	METER	
		EQUIVALENT	REAL-WORLD EQUIVALENT
Kilometer	km	10^3 m	About two-thirds of a mile
Meter	m	1 m	About 3 feet
Centimeter	cm	10^{-2} m	Thickness of your little finger
Millimeter	mm	10^{-3} m	Thickness of your toenail
Micrometer	μm	10^{-6} m	Near the limit of resolution for the light microscope
Nanometer	nm	10^{-9} m	Near the limit of resolution for the electron microscope

using a microscope, it was necessary to make very thin slices, ideally not much thicker than the diameter of the cells. However, brain tissue has a consistency like a bowl of Jello: not firm enough to make thin slices. Thus, the study of the anatomy of brain cells had to await the development of a method to harden the tissue without disturbing its structure and an instrument that could produce very thin slices. Early in the nineteenth century, scientists discovered how to harden, or “fix,” tissues by immersing them in formaldehyde, and they developed a special device called a microtome to make very thin slices.

These technical advances spawned the field of **histology**, the microscopic study of the structure of tissues. But scientists studying brain structure faced yet another obstacle. Freshly prepared brain has a uniform, cream-colored appearance under the microscope; the tissue has no differences in pigmentation to enable histologists to resolve individual cells. Thus, the final breakthrough in neurohistology was the introduction of stains that could selectively color some, but not all, parts of the cells in brain tissue.

One stain, still used today, was introduced by the German neurologist Franz Nissl in the late nineteenth century. Nissl showed that a class of basic dyes would stain the nuclei of all cells and also stain clumps of material surrounding the nuclei of neurons (Figure 2.1). These clumps are called *Nissl bodies*, and the stain is known as the **Nissl stain**. The Nissl stain is extremely useful for two reasons. First, it distinguishes neurons and glia from one another. Second, it enables histologists to study the arrangement, or **cytoarchitecture**, of neurons in different parts of the brain. (The prefix *cyto-* is from the Greek word for “cell.”) The study of cytoarchitecture led to the realization that the brain consists of many specialized regions. We now know that each region performs a different function.

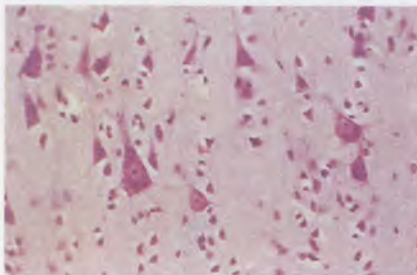


FIGURE 2.1

Nissl-stained neurons. A thin slice of brain tissue has been stained with cresyl violet, a Nissl stain. The clumps of deeply stained material around the cell nuclei are Nissl bodies. (Source: Hammersen, 1980, Fig. 493.)



FIGURE 2.2
Camillo Golgi (1843–1926).
 (Source: Finger, 1994, Fig. 3.22.)

The Golgi Stain

The Nissl stain, however, does not tell the whole story. A Nissl-stained neuron looks like little more than a lump of protoplasm containing a nucleus. Neurons are much more than that, but how much more was not recognized until the publication of the work of Italian histologist Camillo Golgi (Figure 2.2). In 1873, Golgi discovered that by soaking brain tissue in a silver chromate solution, now called the **Golgi stain**, a small percentage of neurons became darkly colored in their entirety (Figure 2.3). This revealed that the neuronal cell body, the region of the neuron around the nucleus that is shown with the Nissl stain, is actually only a small fraction of the total structure of the neuron. Notice in Figures 2.1 and 2.3 how different histological stains can provide strikingly different views of the same tissue. Today, neurohistology remains an active field in neuroscience, along with its credo: “The gain in brain is mainly in the stain.”

The Golgi stain shows that neurons have at least two distinguishable parts: a central region that contains the cell nucleus, and numerous thin tubes that radiate away from the central region. The swollen region containing the cell nucleus has several names that are used interchangeably: **cell body**, **soma** (plural: somata), and **perikaryon** (plural: perikarya). The thin tubes that radiate away from the soma are called **neurites** and are of two types: **axons** and **dendrites** (Figure 2.4).

The cell body usually gives rise to a single axon. The axon is of uniform diameter throughout its length, and if it branches, the branches generally extend at right angles. Because axons can travel over great distances in the body (a meter or more), it was immediately recognized by the histologists of the day that axons must act like “wires” that carry the output of the neurons. Dendrites, on the other hand, rarely extend more than 2 mm in

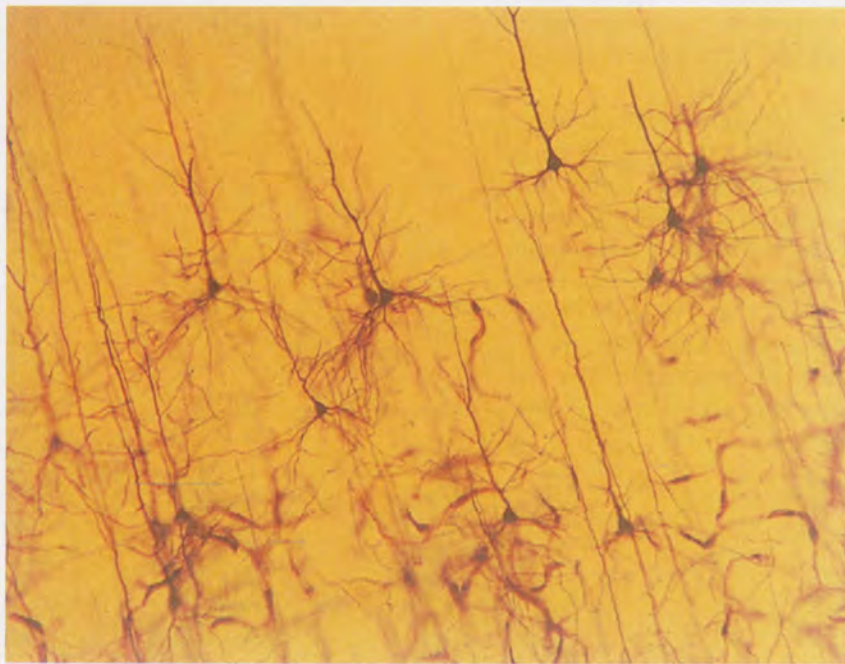


FIGURE 2.3
Golgi-stained neurons. (Source: Hubel, 1988, p. 126.)

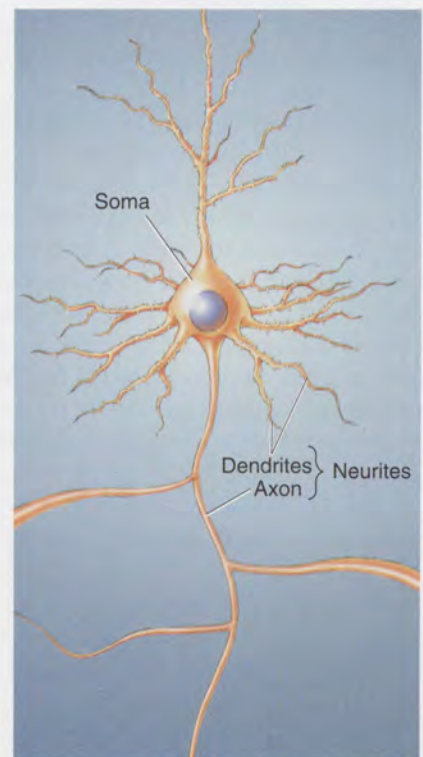


FIGURE 2.4
The basic parts of a neuron.

length. Many dendrites extend from the cell body and generally taper to a fine point. Early histologists recognized that because dendrites come in contact with many axons, they must act as the antennae of the neuron to receive incoming signals, or input.

Cajal's Contribution

Golgi invented the stain, but it was a Spanish contemporary of Golgi who used it to greatest effect. Santiago Ramón y Cajal was a skilled histologist and artist who learned about Golgi's method in 1888 (Figure 2.5). In a remarkable series of publications over the next 25 years, Cajal used the Golgi stain to work out the circuitry of many regions of the brain (Figure 2.6). Ironically, Golgi and Cajal drew completely opposite conclusions about neurons. Golgi championed the view that the neurites of different cells are fused together to form a continuous reticulum, or network, similar to the arteries and veins of the circulatory system. According to this reticular theory, the brain is an exception to the cell theory, which states that the individual cell is the elementary functional unit of all animal tissues. Cajal, on the other hand, argued forcefully that the neurites of different neurons are not continuous with one another and must *communicate by contact, not continuity*. This idea that the neuron adhered to the cell theory came to be known as the **neuron doctrine**. Although Golgi and Cajal shared the Nobel Prize in 1906, they remained rivals to the end.

The scientific evidence over the next 50 years weighed heavily in favor of the neuron doctrine, but final proof had to wait until the development of the electron microscope in the 1950s (Box 2.1). With the increased resolving power of the electron microscope, it was finally possible to show that the neurites of different neurons are not continuous with one another. Thus, our starting point in the exploration of the brain must be the individual neuron.



FIGURE 2.5
Santiago Ramón y Cajal (1852–1934).
(Source: Finger, 1994, Fig. 3.26.)

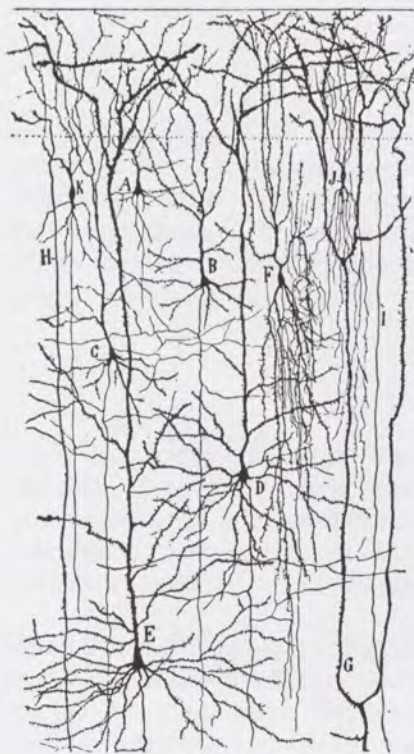


FIGURE 2.6
One of Cajal's many drawings of brain circuitry. The letters label the different elements Cajal identified in an area of the human cerebral cortex that controls voluntary movement. We will learn more about this part of the brain in Chapter 14. (Source: DeFelipe and Jones, 1998, Fig. 90.)

Box 2.1



OF SPECIAL INTEREST

Advances in Microscopy

The human eye can distinguish two points only if they are separated by more than about one-tenth of a millimeter ($100\ \mu\text{m}$). Thus, we can say that $100\ \mu\text{m}$ is near the *limit of resolution* for the unaided eye. Neurons have a diameter of about $20\ \mu\text{m}$, and neurites can measure as small as a fraction of a micrometer. The light microscope, therefore, was a necessary development before neuronal structure could be studied. But this type of microscopy has a

theoretical limit imposed by the properties of microscope lenses and visible light. With the standard light microscope, the limit of resolution is about $0.1\ \mu\text{m}$. However, the space between neurons measures only $0.02\ \mu\text{m}$ ($20\ \text{nm}$). No wonder two esteemed scientists, Golgi and Cajal, disagreed about whether neurites were continuous from one cell to the next. This question could not be answered until the electron microscope was developed and applied to biological specimens, which occurred only within the past 60 years or so.

The electron microscope uses an electron beam instead of light to form images, dramatically increasing the resolving power. The limit of resolution for an electron microscope is about $0.1\ \text{nm}$ —a million times better than the unaided eye. Our insights into the fine structure of the inside of neurons—the *ultrastructure*—have all come from electron microscopic examination of the brain.

Today, microscopes on the leading edge of technology use laser beams to illuminate the tissue and computers to create digital images (Figure A). Unlike the traditional methods of light and electron microscopy, which require tissue fixation, these new techniques give neuroscientists their first chance to peer into brain tissue that is still alive.



FIGURE A

A laser microscope and computer. (Source: Olympus.)

▼ THE PROTOTYPICAL NEURON

As we have seen, the neuron (also called a *nerve cell*) consists of several parts: the soma, the dendrites, and the axon. The inside of the neuron is separated from the outside by the limiting skin, the *neuronal membrane*, which lies like a circus tent on an intricate internal scaffolding, giving each part of the cell its special three-dimensional appearance. Let's explore the inside of the neuron and learn about the functions of the different parts (Figure 2.7).

The Soma

We begin our tour at the soma, the roughly spherical central part of the neuron. The cell body of the typical neuron is about $20\ \mu\text{m}$ in diameter. The watery fluid inside the cell, called the **cytosol**, is a salty, potassium-rich solution that is separated from the outside by the neuronal membrane. Within the soma are a number of membrane-enclosed structures called **organelles**.

The cell body of the neuron contains the same organelles that are found in all animal cells. The most important ones are the nucleus, the rough endoplasmic reticulum, the smooth endoplasmic reticulum, the Golgi apparatus, and the mitochondria. Everything contained within the confines

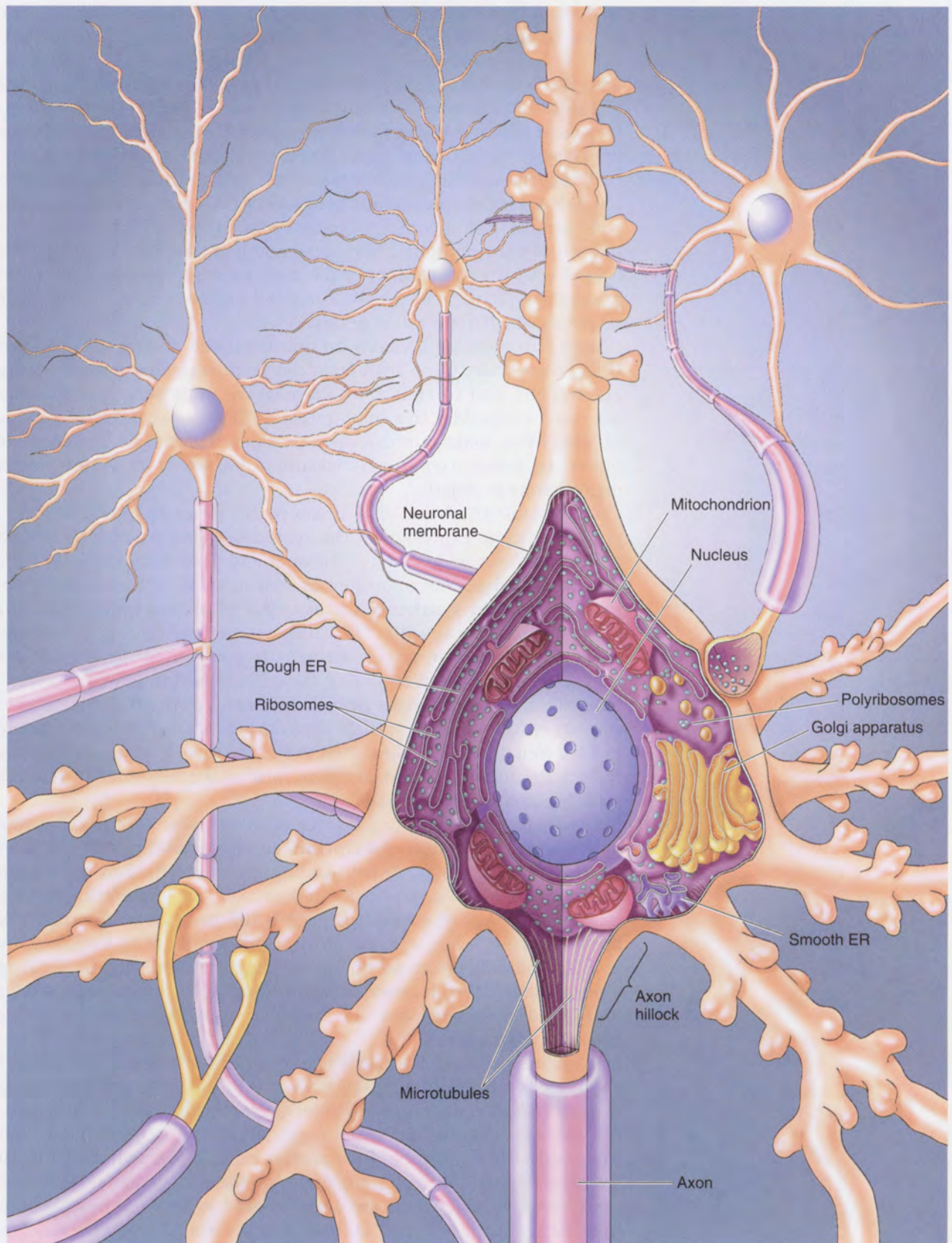


FIGURE 2.7
The internal structure of a typical neuron.

of the cell membrane, including the organelles but excluding the nucleus, is referred to collectively as the **cytoplasm**.

The Nucleus. Its name derived from the Latin word for “nut,” the **nucleus** of the cell is spherical, centrally located, and about 5–10 μm across. It is contained within a double membrane called the *nuclear envelope*. The nuclear envelope is perforated by pores that measure about 0.1 μm across.

Within the nucleus are **chromosomes**, which contain the genetic material, **DNA (deoxyribonucleic acid)**. Your DNA was passed on to you from your parents, and it contains the blueprint for your entire body. The DNA in each of your neurons is the same, and it is the same as the DNA in the cells of your liver and kidney. What distinguishes a neuron from a liver cell are the specific parts of the DNA that are used to assemble the cell. These segments of DNA are called **genes**.

Each chromosome contains an uninterrupted double-stranded braid of DNA, 2 nm wide. If the DNA from the 46 human chromosomes were laid out straight, end to end, it would measure more than 2 m in length. If we were to consider this total length of DNA as analogous to the string of letters that makes up this book, the genes would be analogous to the individual words. Genes can measure anywhere from 0.1 μm to several micrometers in length.

The “reading” of the DNA is known as **gene expression**. The final product of gene expression is the synthesis of molecules called **proteins**, which exist in a wide variety of shapes and sizes, perform many different functions, and bestow upon neurons virtually all of their unique characteristics. **Protein synthesis**, the assembly of protein molecules, occurs in the cytoplasm. Because the DNA never leaves the nucleus, there must be an intermediary that carries the genetic message to the sites of protein synthesis in the cytoplasm. This function is performed by another long molecule called **messenger ribonucleic acid**, or **mRNA**. Messenger RNA consists of four different nucleic acids strung together in various sequences to form a chain. The detailed sequence of the nucleic acids in the chain represents the information in the gene, just as the sequence of letters gives meaning to a written word.

The process of assembling a piece of mRNA that contains the information of a gene is called **transcription**, and the resulting mRNA is called the *transcript* (Figure 2.8a). Protein-coding genes are flanked by stretches of DNA that are not used to encode proteins but are important for regulating transcription. At one end of the gene is the **promoter**, the region where the RNA-synthesizing enzyme, *RNA polymerase*, binds to initiate transcription. The binding of the polymerase to the promoter is tightly regulated by other proteins called **transcription factors**. At the other end is a sequence of DNA called the *terminator* that the RNA polymerase recognizes as the end point for transcription.

In addition to the non-coding regions of DNA that flank the genes, there are often additional stretches of DNA within the gene itself that cannot be used to code for protein. These interspersed regions are called *introns*, and the coding sequences are called *exons*. Initial transcripts contain both introns and exons, but then, by a process called **RNA splicing**, the introns are removed and the remaining exons are fused together (Figure 2.8b). In some cases, specific exons are also removed with the introns, leaving an “alternatively spliced” mRNA that actually encodes a different protein. Thus, transcription of a single gene can ultimately give rise to several different mRNAs and protein products.

Messenger RNA transcripts emerge from the nucleus via pores in the nuclear envelope and travel to the sites of protein synthesis elsewhere in

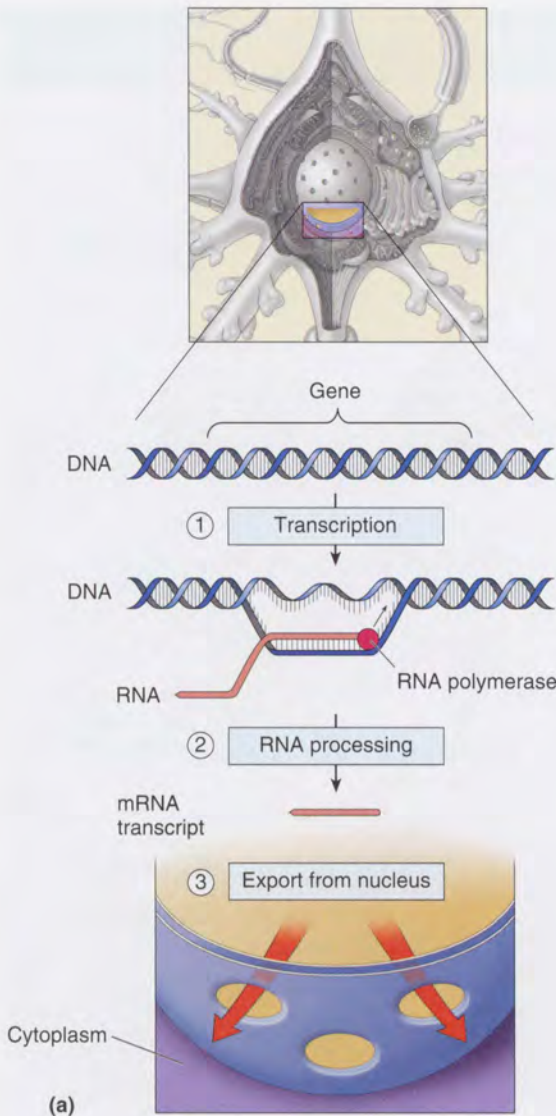
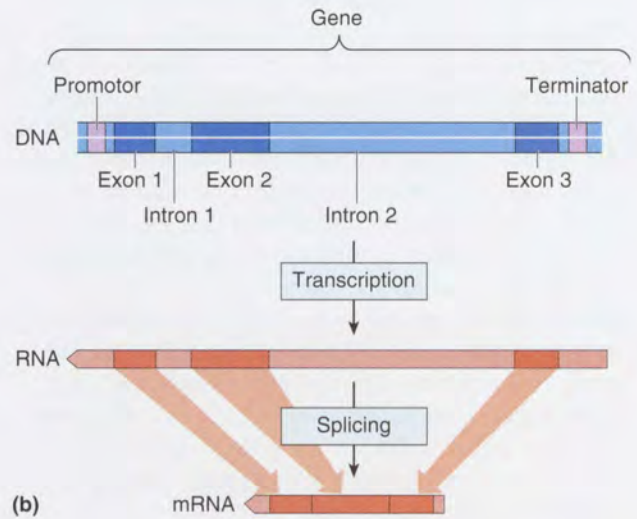


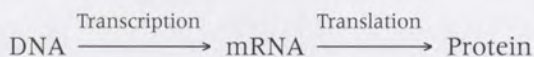
FIGURE 2.8

Gene transcription. (a) RNA molecules are synthesized by RNA polymerase and then processed into messenger RNA to carry the genetic instructions for protein assembly from the nucleus to the cytoplasm. **(b)** Transcription is initiated at the promoter region of the gene and stopped at the terminator region. The initial RNA must be spliced to remove the introns that do not code for protein.



the neuron. At these sites, a protein molecule is assembled much as the mRNA molecule was: by linking together many small molecules into a chain. In the case of protein, the building blocks are **amino acids**, of which there are 20 different kinds. This assembling of proteins from amino acids under the direction of the mRNA is called **translation**.

The scientific study of this process, which begins with the DNA of the nucleus and ends with the synthesis of protein molecules in the cell, is known as *molecular biology*. The “central dogma” of molecular biology is summarized as follows:



A new field within neuroscience is called *molecular neurobiology*. Molecular neurobiologists use the information contained in the genes to determine the structure and functions of neuronal proteins (Box 2.2).

Rough Endoplasmic Reticulum. Not far from the nucleus are enclosed stacks of membrane dotted with dense globular structures called **ribosomes**,

Box 2.2



BRAIN FOOD

Expressing One's Mind in the Post-Genomic Era

Sequencing the human genome—the entire length of DNA that comprises the genetic information in our chromosomes—was a truly monumental achievement, completed in 2003. The Human Genome Project identified all of the approximately 20,000 genes in human DNA. We now live in what has been called the “post-genomic era,” in which information about the genes expressed in our tissues can be used to diagnose and treat diseases. Neuroscientists are now using this information to tackle long-standing questions about the biological basis of neurological and psychiatric disorders, as well as to probe deeper into the origins of individuality. The logic goes as follows. The brain is the product of the genes expressed in it. Differences in gene expression between a normal brain and a diseased brain, or a brain of unusual ability, can be used to identify the molecular basis of the observed symptoms or traits.

The level of gene expression is usually defined as the number of mRNA transcripts synthesized by different cells and tissues to direct the synthesis of specific proteins. Thus, the analysis of gene expression requires a method to compare the relative abundance of various mRNAs in the brains of two groups of humans or animals. One way to perform such a comparison is to use DNA *microarrays*, which are created by robotic machines that arrange thousands of small spots of synthetic DNA on a microscope slide. Each spot contains a unique DNA sequence that will recognize and stick to a different specific mRNA sequence. To compare the gene expression in two brains, one begins by collecting a sample of mRNAs from each brain. The mRNA of one brain is labeled with a chemical tag that fluoresces green, and the mRNA of the other brain is labeled with a tag that fluoresces red. These samples are then applied to the microarray. Highly expressed genes will produce brightly fluorescent spots, and differences in the relative gene expression between the brains will be revealed by differences in the color of the fluorescence (Figure A).

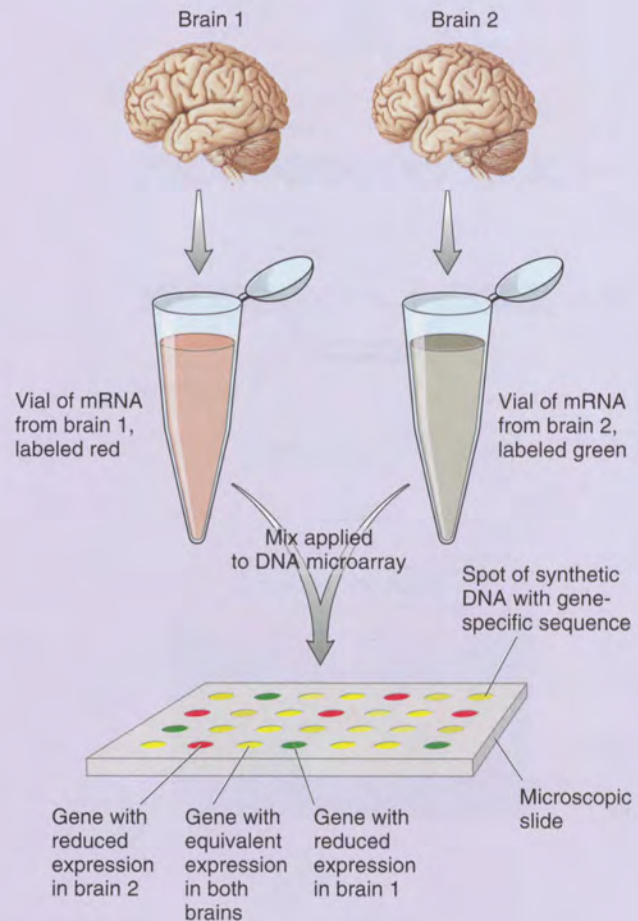


FIGURE A
Profiling differences in gene expression.

which measure about 25 nm in diameter. The stacks are called **rough endoplasmic reticulum**, or **rough ER** (Figure 2.9). Rough ER abounds in neurons, far more than in glia or most other non-neuronal cells. In fact, we have already been introduced to rough ER by another name: Nissl bodies. This organelle is stained with the dyes that Nissl introduced 100 years ago.

Rough ER is a major site of protein synthesis in neurons. RNA transcripts bind to the ribosomes, and the ribosomes translate the instructions contained in the mRNA to assemble a protein molecule. Thus, ribosomes take raw material in the form of amino acids and manufacture proteins using the blueprint provided by the mRNA (Figure 2.10a).

Not all ribosomes are attached to the rough ER. Many are freely floating and are called *free ribosomes*. Several free ribosomes may appear to be attached by a thread; these are called **polyribosomes**. The thread is a single strand of mRNA, and the associated ribosomes are working on it to make multiple copies of the same protein.

What is the difference between proteins synthesized on the rough ER and those synthesized on the free ribosomes? The answer appears to lie in the intended fate of the protein molecule. If it is destined to reside within the cytosol of the neuron, then the protein's mRNA transcript shuns the ribosomes of the rough ER and gravitates toward the free ribosomes.

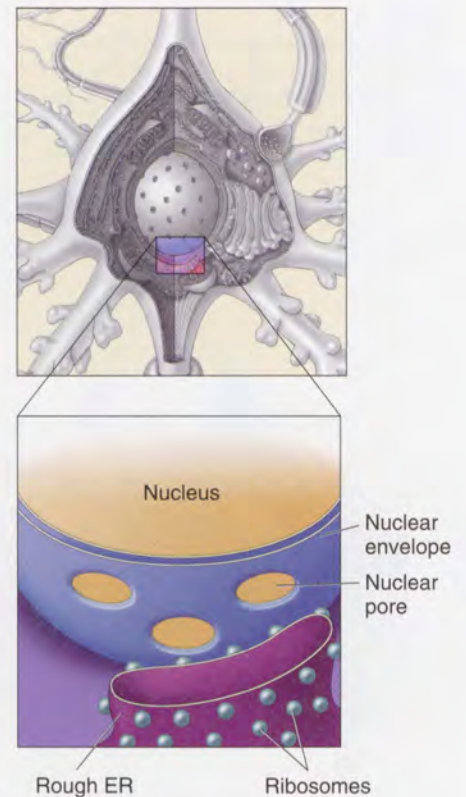


FIGURE 2.9
Rough endoplasmic reticulum, or rough ER.

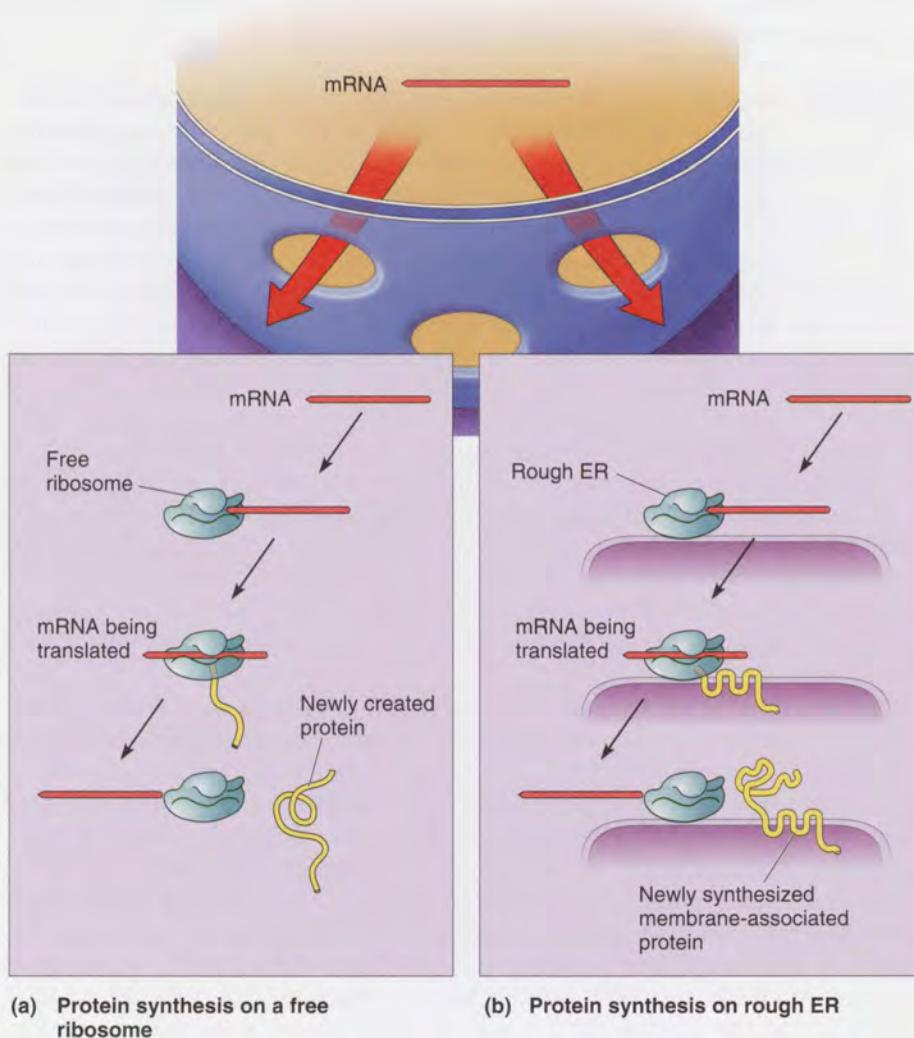


FIGURE 2.10
Protein synthesis on a free ribosome and on rough ER. Messenger RNA (mRNA) binds to a ribosome, initiating protein synthesis. **(a)** Proteins synthesized on free ribosomes are destined for the cytosol. **(b)** Proteins synthesized on the rough ER are destined to be enclosed by or inserted into the membrane. Membrane-associated proteins are inserted into the membrane as they are assembled.

FIGURE 2.11

The Golgi apparatus. This complex organelle sorts newly synthesized proteins for delivery to appropriate locations in the neuron.

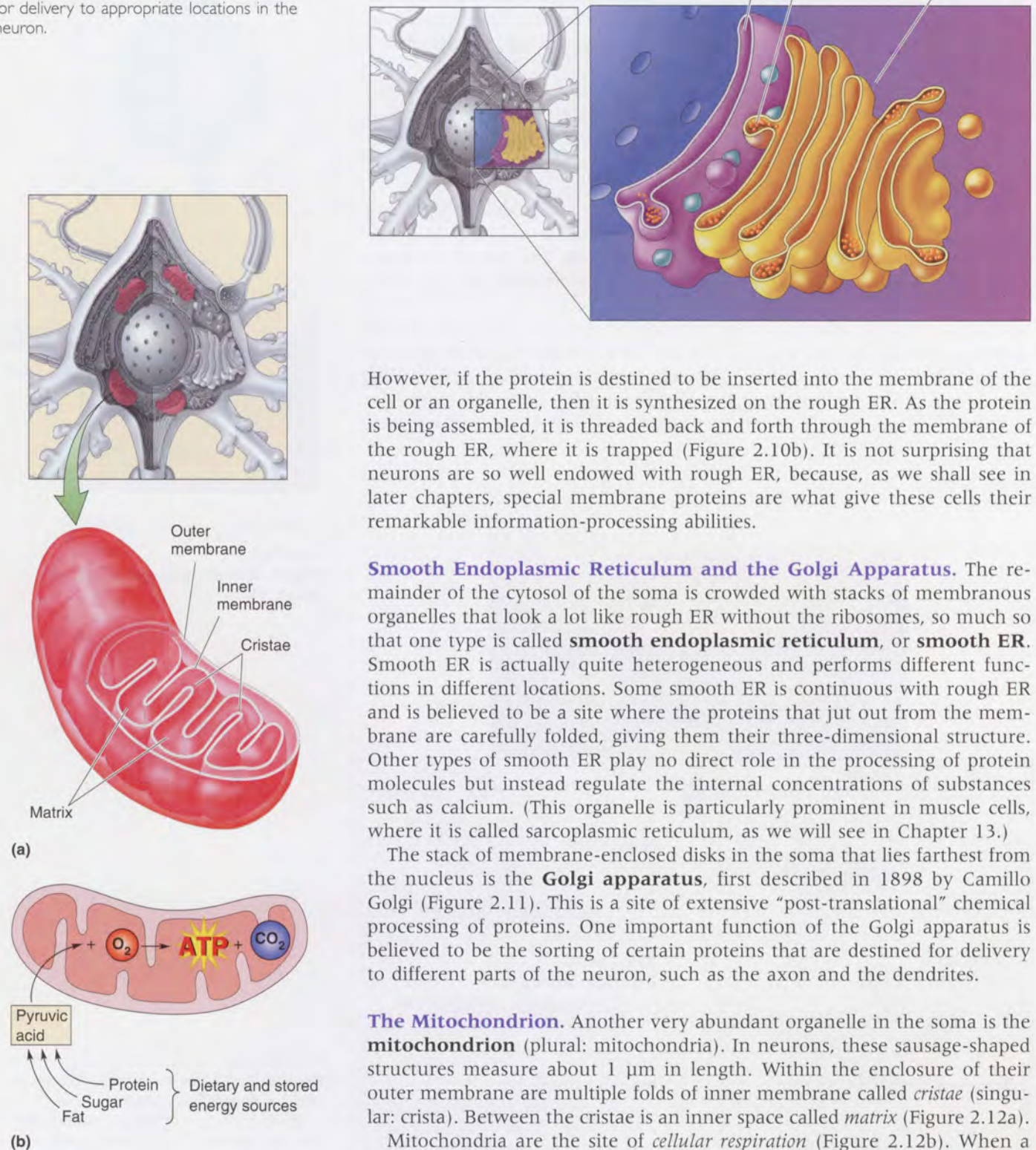


FIGURE 2.12

The role of mitochondria. (a) Components of a mitochondrion. (b) Cellular respiration. ATP is the energy currency that fuels biochemical reactions in neurons.

However, if the protein is destined to be inserted into the membrane of the cell or an organelle, then it is synthesized on the rough ER. As the protein is being assembled, it is threaded back and forth through the membrane of the rough ER, where it is trapped (Figure 2.10b). It is not surprising that neurons are so well endowed with rough ER, because, as we shall see in later chapters, special membrane proteins are what give these cells their remarkable information-processing abilities.

Smooth Endoplasmic Reticulum and the Golgi Apparatus. The remainder of the cytosol of the soma is crowded with stacks of membranous organelles that look a lot like rough ER without the ribosomes, so much so that one type is called **smooth endoplasmic reticulum**, or **smooth ER**. Smooth ER is actually quite heterogeneous and performs different functions in different locations. Some smooth ER is continuous with rough ER and is believed to be a site where the proteins that jut out from the membrane are carefully folded, giving them their three-dimensional structure. Other types of smooth ER play no direct role in the processing of protein molecules but instead regulate the internal concentrations of substances such as calcium. (This organelle is particularly prominent in muscle cells, where it is called sarcoplasmic reticulum, as we will see in Chapter 13.)

The stack of membrane-enclosed disks in the soma that lies farthest from the nucleus is the **Golgi apparatus**, first described in 1898 by Camillo Golgi (Figure 2.11). This is a site of extensive “post-translational” chemical processing of proteins. One important function of the Golgi apparatus is believed to be the sorting of certain proteins that are destined for delivery to different parts of the neuron, such as the axon and the dendrites.

The Mitochondrion. Another very abundant organelle in the soma is the **mitochondrion** (plural: mitochondria). In neurons, these sausage-shaped structures measure about 1 μm in length. Within the enclosure of their outer membrane are multiple folds of inner membrane called *cristae* (singular: *crista*). Between the cristae is an inner space called *matrix* (Figure 2.12a).

Mitochondria are the site of *cellular respiration* (Figure 2.12b). When a mitochondrion “inhales,” it pulls inside pyruvic acid (derived from sugars and digested proteins and fats) and oxygen, both of which are floating in the cytosol. Within the inner compartment of the mitochondrion, pyruvic acid enters into a complex series of biochemical reactions called the *Krebs cycle*, named after the German-British scientist Hans Krebs, who first proposed it

in 1937. The biochemical products of the Krebs cycle provide energy that, in another series of reactions within the cristae (called the electron-transport chain), results in the addition of phosphate to adenosine diphosphate (ADP), yielding **adenosine triphosphate (ATP)**, the cell's energy source. When the mitochondrion "exhales," 17 ATP molecules are released for every molecule of pyruvic acid that had been taken in.

ATP is the energy currency of the cell. The chemical energy stored in ATP is used to fuel most of the biochemical reactions of the neuron. For example, as we shall see in Chapter 3, special proteins in the neuronal membrane use the energy released by the breakdown of ATP into ADP to pump certain substances across the membrane to establish concentration differences between the inside and the outside of the neuron.

The Neuronal Membrane

The **neuronal membrane** serves as a barrier to enclose the cytoplasm inside the neuron and to exclude certain substances that float in the fluid that bathes the neuron. The membrane is about 5 nm thick and is studded with proteins. As mentioned earlier, some of the membrane-associated proteins pump substances from the inside to the outside. Others form pores that regulate which substances can gain access to the inside of the neuron. An important characteristic of neurons is that the protein composition of the membrane varies depending on whether it is in the soma, the dendrites, or the axon.

The function of neurons cannot be understood without understanding the structure and function of the membrane and its associated proteins. In fact, this topic is so important that we'll spend a good deal of the next four chapters looking at how the membrane endows neurons with the remarkable ability to transfer electrical signals throughout the brain and body.

The Cytoskeleton

Earlier, we compared the neuronal membrane to a circus tent that was draped on an internal scaffolding. This scaffolding is called the **cytoskeleton**, and it gives the neuron its characteristic shape. The "bones" of the cytoskeleton are the microtubules, microfilaments, and neurofilaments (Figure 2.13). By drawing an analogy with a scaffolding, we do not mean that the cytoskeleton is static. On the contrary, elements of the cytoskeleton are dynamically regulated and are very likely in continual motion. Your neurons are probably squirming around in your head even as you read this sentence.

Microtubules. Measuring 20 nm in diameter, **microtubules** are big and run longitudinally down neurites. A microtubule appears as a straight, thick-walled hollow pipe. The wall of the pipe is composed of smaller strands that are braided like rope around the hollow core. Each of the smaller strands consists of the protein *tubulin*. A single tubulin molecule is small and globular; the strand consists of tubulins stuck together like pearls on a string. The process of joining small proteins to form a long strand is called *polymerization*; the resulting strand is called a *polymer*. Polymerization and depolymerization of microtubules and, therefore, of neuronal shape can be regulated by various signals within the neuron.

One class of proteins that participate in the regulation of microtubule assembly and function are *microtubule-associated proteins*, or *MAPs*. Among other functions (many of which are unknown), MAPs anchor the microtubules to one another and to other parts of the neuron. Pathological changes in an axonal MAP, called tau, have been implicated in the dementia that accompanies Alzheimer's disease (Box 2.3).

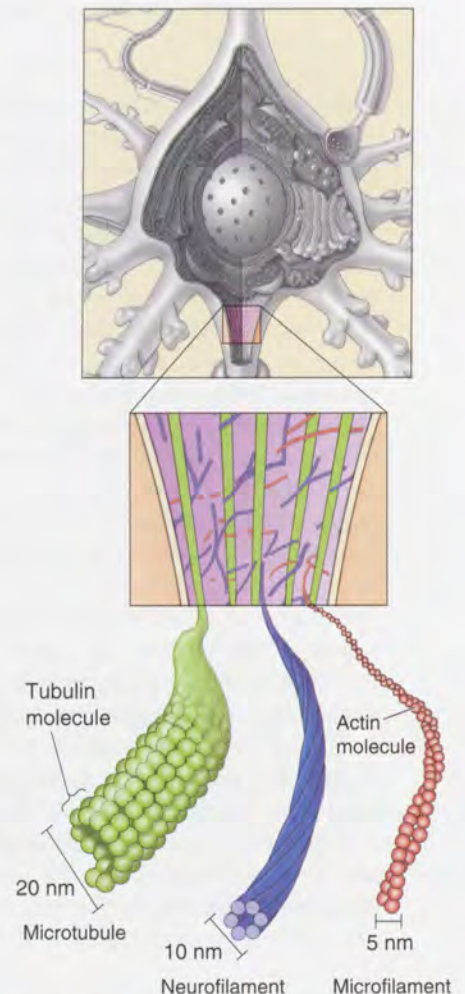


FIGURE 2.13

Components of the cytoskeleton.

The arrangement of microtubules, neurofilaments, and microfilaments gives the neuron its characteristic shape.

Box 2.3



OF SPECIAL INTEREST

Alzheimer's Disease and the Neuronal Cytoskeleton

Neurites are the most remarkable structural feature of a neuron. Their elaborate branching patterns, critical for information processing, reflect the organization of the underlying cytoskeleton. It is therefore no surprise that a devastating loss of brain function can result when the cytoskeleton of neurons is disrupted. An example is *Alzheimer's disease*, which is characterized by the disruption of the cytoskeleton of neurons in the cerebral cortex, a region of the brain crucial for cognitive function. This disorder and its underlying brain pathology were first described in 1907 by the German physician A. Alzheimer in a paper titled "A Characteristic Disease of the Cerebral Cortex." Below are excerpts from the English translation.

One of the first disease symptoms of a 51-year-old woman was a strong feeling of jealousy toward her husband. Very soon she showed rapidly increasing memory impairments; she could not find her way about her home, she dragged objects to and fro, hid herself, or sometimes thought that people were out to kill her, then she would start to scream loudly.

During institutionalization her gestures showed a complete helplessness. She was disoriented as to time and place. From time to time she would state that she did not understand anything, that she felt confused and totally lost. Sometimes she considered the coming of the doctor as an official visit and apologized for not having finished her work, but other times she would start to yell in the fear that the doctor wanted to operate on her; or there were times that she would send him away in complete indignation, uttering phrases that indicated her fear that the doctor wanted to damage her woman's honor. From time to time she was completely delirious, dragging her blankets and sheets to and fro, calling for her husband and daughter, and seeming to have auditory hallucinations. Often she would scream for hours and hours in a horrible voice.

Mental regression advanced quite steadily. After four and a half years of illness the patient died. She was completely apathetic in the end, and was confined to bed in a fetal position. (Bick et al., 1987, pp. 1–2.)

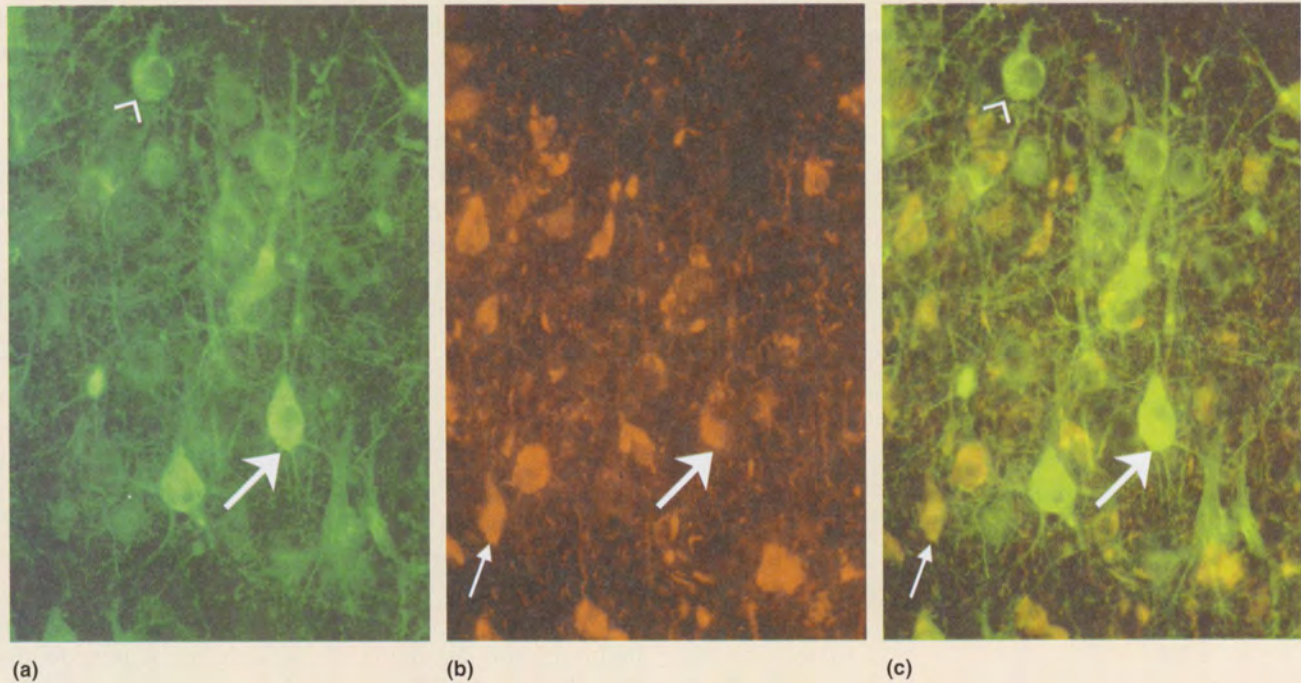
Following her death, Alzheimer examined the woman's brain under the microscope. He made particular note of changes in the "neurofibrils," elements of the cytoskeleton that are stained by a silver solution.

The Bielschowsky silver preparation showed very characteristic changes in the neurofibrils. However, inside an apparently normal-looking cell, one or more single fibers could be observed that became prominent through their striking thickness and specific impregnability. At a more advanced stage, many fibrils arranged parallel showed the same changes. Then they accumulated forming dense bundles and gradually advanced to the surface of the cell. Eventually, the nucleus and cytoplasm disappeared, and only a tangled bundle of fibrils indicated the site where once the neuron had been located.

As these fibrils can be stained with dyes different from the normal neurofibrils, a chemical transformation of the fibril substance must have taken place. This might be the reason why the fibrils survived the destruction of the cell. It seems that the transformation of the fibrils goes hand in hand with the storage of an as yet not closely examined pathological product of the metabolism in the neuron. About one-quarter to one-third of all the neurons of the cerebral cortex showed such alterations. Numerous neurons, especially in the upper cell layers, had totally disappeared. (Bick et al., 1987, pp. 2–3.)

The severity of the dementia in Alzheimer's disease is well correlated with the number and distribution of what are now commonly known as *neurofibrillary tangles*, the "tombstones" of dead and dying neurons (Figure A). Indeed, as Alzheimer speculated, tangle formation in the cerebral cortex very likely causes the symptoms of the disease. Electron microscopy reveals that the major components of the tangles are *paired helical filaments*, long fibrous proteins braided together like strands of a rope (Figure B). It is now understood that these filaments consist of the microtubule-associated protein *tau*.

Tau normally functions as a bridge between the microtubules in axons, ensuring that they run straight and

**FIGURE A**

Neurons in a human brain with Alzheimer's disease. Normal neurons contain neurofilaments but no neurofibrillary tangles. **(a)** Brain tissue stained by a method that makes neuronal neurofilaments fluoresce green, showing viable neurons. **(b)** The same region of the brain stained to show the presence of tau within neurofibrillary tangles, revealed by red fluorescence. **(c)** Superimposition of images in parts a and b. The neuron indicated by the arrowhead contains neurofilaments but no tangles and, therefore, is healthy. The neuron indicated by the large arrow has neurofilaments but also has started to show accumulation of tau and, therefore, is diseased. The neuron indicated by the small arrow in parts b and c is dead because it contains no neurofilaments. The remaining tangle is the tombstone of a neuron killed by Alzheimer's disease. (Source: Courtesy of Dr. John Morrison and modified from Vickers et al., 1994.)

parallel to one another. In Alzheimer's disease, the tau detaches from the microtubules and accumulates in the soma. This disruption of the cytoskeleton causes the axons to wither, thus impeding the normal flow of information in the affected neurons.

What causes the changes in tau? Attention is focused on another protein that accumulates in the brain of Alzheimer's patients, called *amyloid*. The field of Alzheimer's

disease research moves very fast, but the consensus today is that the abnormal secretion of amyloid by neurons is the first step in the process that leads to neurofibrillary tangle formation and dementia. Current hope for therapeutic intervention is focused on strategies to reduce the depositions of amyloid in the brain. The need for effective therapy is urgent: In the United States alone, more than 4 million people are afflicted with this tragic disease.

**FIGURE B**

Paired helical filaments of a tangle. (Source: Goedert, 1996, Fig. 2b.)

Microfilaments. Measuring only 5 nm in diameter, **microfilaments** are about the same thickness as the cell membrane. Found throughout the neuron, they are particularly numerous in the neurites. Microfilaments are braids of two thin strands, and the strands are polymers of the protein *actin*. Actin is one of the most abundant proteins in cells of all types, including neurons, and is believed to play a role in changing cell shape. Indeed, as we shall see in Chapter 13, actin filaments are critically involved in the mechanism of muscle contraction.

Like microtubules, actin microfilaments are constantly undergoing assembly and disassembly, and this process is regulated by signals in the neuron. Besides running longitudinally down the core of the neurites like microtubules, microfilaments are also closely associated with the membrane. They are anchored to the membrane by attachments with a meshwork of fibrous proteins that line the inside of the membrane like a spider web.

Neurofilaments. With a diameter of 10 nm, **neurofilaments** are intermediate in size between microtubules and microfilaments. They exist in all cells of the body as *intermediate filaments*; only in neurons are they called neurofilaments. The difference in name actually does reflect subtle differences in structure from one tissue to the next. An example of an intermediate filament from another tissue is keratin, which, when bundled together, makes up hair.

Of the types of fibrous structure we have discussed, neurofilaments most closely resemble the bones and ligaments of the skeleton. A neurofilament consists of multiple subunits (building blocks) that are organized like a chain of sausages. The internal structure of each subunit consists of three protein strands woven together. Unlike microfilaments and microtubules, these strands consist of individual long protein molecules, each of which is coiled in a tight, springlike configuration. This structure makes neurofilaments mechanically very strong.

The Axon

So far, we've explored the soma, organelles, membrane, and cytoskeleton. However, none of these structures is unique to neurons; they are found in all the cells in our body. Now we encounter the axon, a structure found only in neurons that is highly specialized for the transfer of information over distances in the nervous system.

The axon begins with a region called the **axon hillock**, which tapers to form the initial segment of the axon proper (Figure 2.14). Two noteworthy features distinguish the axon from the soma:

1. No rough ER extends into the axon, and there are few, if any, free ribosomes.
2. The protein composition of the axon membrane is fundamentally different from that of the soma membrane.

These structural differences translate into functional distinctions. Because there are no ribosomes, there is no protein synthesis in the axon. This means that all proteins in the axon must originate in the soma. And it is the different proteins in the axonal membrane that enable it to serve as the "telegraph wire" that sends information over great distances.

Axons may extend from less than a millimeter to over a meter long. Axons often branch, and these branches are called **axon collaterals**. Occasionally, an axon collateral will return to communicate with the same cell that gave rise to the axon or with the dendrites of neighboring cells. These axon branches are called *recurrent collaterals*.

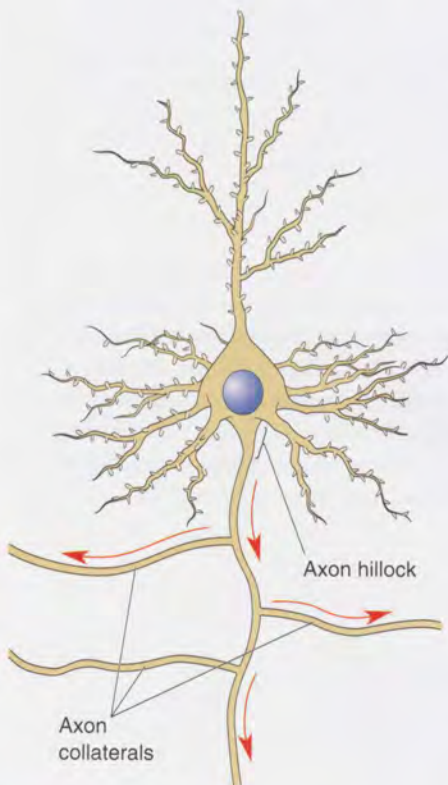


FIGURE 2.14
The axon and axon collaterals. The axon functions like a telegraph wire to send electrical impulses to distant sites in the nervous system. The arrows indicate the direction of information flow.

The diameter of an axon is variable, ranging from less than 1 mm to about 25 mm in humans and to as large as 1 mm in the squid. This variation in axon size is important. As will be explained in Chapter 4, the speed of the electrical signal that sweeps down the axon—the *nerve impulse*—varies depending on axonal diameter. The thicker the axon, the faster the impulse travels.

The Axon Terminal. All axons have a beginning (the axon hillock), a middle (the axon proper), and an end. The end is called the **axon terminal** or **terminal bouton** (French for “button”), reflecting the fact that it usually appears as a swollen disk (Figure 2.15). The terminal is a site where the axon comes in contact with other neurons (or other cells) and passes information on to them. This point of contact is called the **synapse**, a word derived from the Greek, meaning “to fasten together.” Sometimes axons have many branches at their ends, and each branch forms a synapse on dendrites or cell bodies in the same region. These branches are collectively called the **terminal arbor**. Sometimes axons form synapses at swollen regions along their length and then continue on to terminate elsewhere. Such swellings are called *boutons en passant* (“buttons in passing”). In either case, when a neuron makes synaptic contact with another cell, it is said to innervate that cell, or to provide **innervation**.

The cytoplasm of the axon terminal differs from that of the axon in several ways:

1. Microtubules do not extend into the terminal.
2. The terminal contains numerous small bubbles of membrane, called **synaptic vesicles**, that measure about 50 nm in diameter.
3. The inside surface of the membrane that faces the synapse has a particularly dense covering of proteins.
4. It has numerous mitochondria, indicating a high energy demand.

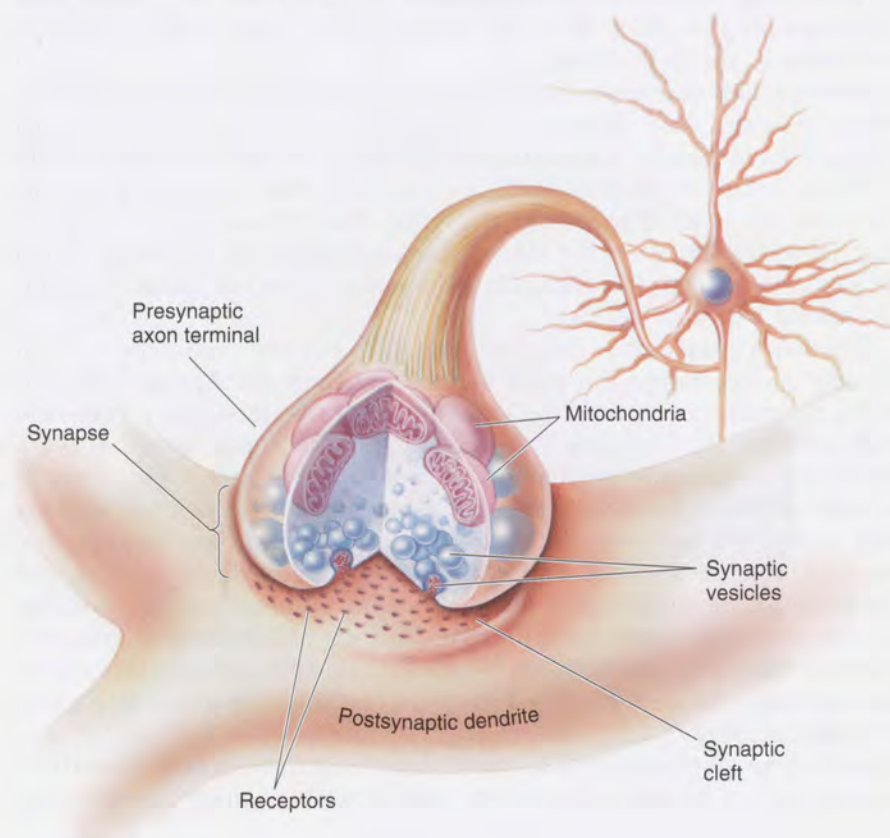


FIGURE 2.15

The axon terminal and the synapse. Axon terminals form synapses with the dendrites or somata of other neurons. When a nerve impulse arrives in the presynaptic axon terminal, neurotransmitter molecules are released from synaptic vesicles into the synaptic cleft. Neurotransmitter then binds to specific receptor proteins, causing the generation of electrical or chemical signals in the postsynaptic cell.

The Synapse. Although Chapters 5 and 6 are devoted entirely to how information is transferred from one neuron to another at the synapse, we provide a preview here. The synapse has two sides: *presynaptic* and *postsynaptic* (see Figure 2.15). These names indicate the usual direction of information flow, which is from “pre” to “post.” The presynaptic side generally consists of an axon terminal, while the postsynaptic side may be the dendrite or soma of another neuron. The space between the presynaptic and postsynaptic membranes is called the **synaptic cleft**. The transfer of information at the synapse from one neuron to another is called **synaptic transmission**.

At most synapses, information in the form of electrical impulses traveling down the axon is converted in the terminal into a chemical signal that crosses the synaptic cleft. On the postsynaptic membrane, this chemical signal is converted again into an electrical one. The chemical signal is called a **neurotransmitter**, and it is stored in and released from the synaptic vesicles within the terminal. As we will see, different neurotransmitters are used by different types of neurons.

This electrical-to-chemical-to-electrical transformation of information makes possible many of the brain’s computational abilities. Modification of this process is involved in memory and learning, and synaptic transmission dysfunction accounts for certain mental disorders. The synapse is also the site of action for many toxins and for most psychoactive drugs.

Axoplasmic Transport. As mentioned, one feature of the cytoplasm of axons, including the terminal, is the absence of ribosomes. Because ribosomes are the protein factories of the cell, their absence means that the proteins of the axon must be synthesized in the soma and then shipped down the axon. Indeed, in the mid-nineteenth century, English physiologist Augustus Waller showed that axons cannot be sustained when separated from their parent cell body. The degeneration of axons that occurs when they are cut is now called *Wallerian degeneration*. Because it can be detected with certain staining methods, Wallerian degeneration is one way to trace axonal connections in the brain.

Wallerian degeneration occurs because the normal flow of materials from the soma to the axon terminal is interrupted. This movement of material down the axon is called **axoplasmic transport**, first demonstrated directly by the experiments of American neurobiologist Paul Weiss and his colleagues in the 1940s. They found that if they tied a thread around an axon, material accumulated on the side of the axon closest to the soma. When the knot was untied, the accumulated material continued down the axon at a rate of 1–10 mm per day.

This was a remarkable discovery, but it is not the whole story. If all material moved down the axon by this transport mechanism alone, it would not reach the ends of the longest axons for at least half a year—too long a wait to feed hungry synapses. In the late 1960s, methods were developed to track the movements of protein molecules down the axon into the terminal. These methods entailed injecting the somata of neurons with radioactive amino acids. Recall that amino acids are the building blocks of proteins. The “hot” amino acids were assembled into proteins, and the arrival of radioactive proteins in the axon terminal was measured to calculate the rate of transport. Bernice Grafstein of Rockefeller University discovered that this *fast axoplasmic transport* (so named to distinguish it from *slow axoplasmic transport* described by Weiss) occurred at a rate as high as 1000 mm per day.

Much is now known about how axoplasmic transport works. Material is enclosed within vesicles, which then “walk down” the microtubules of the

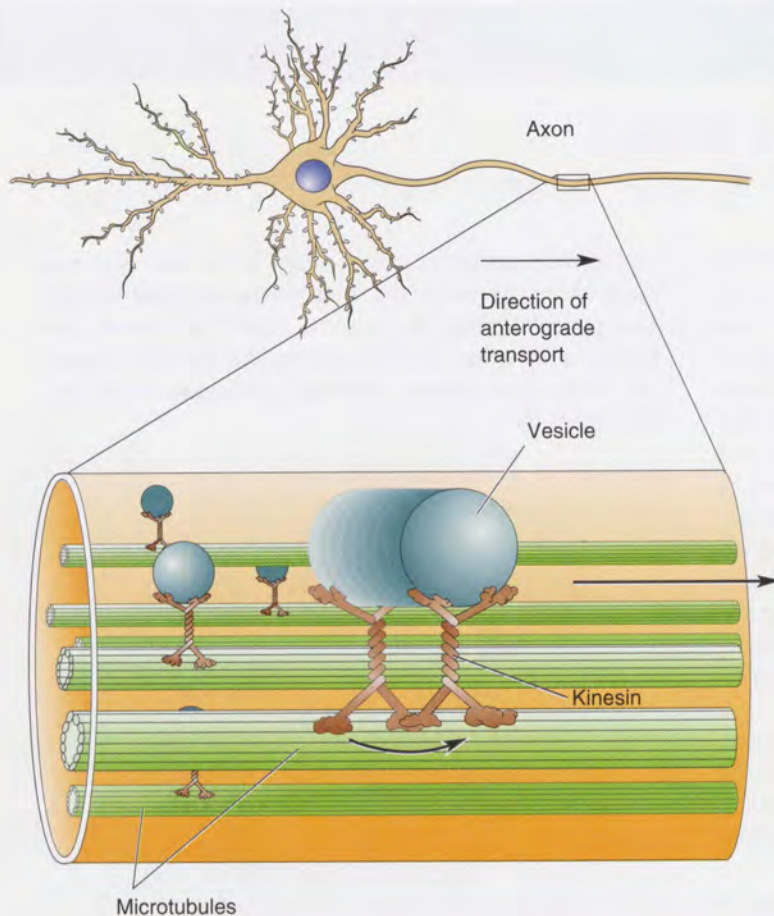


FIGURE 2.16

A mechanism for the movement of material on the microtubules of the axon. Trapped in membrane-enclosed vesicles, material is transported from the soma to the axon terminal by the action of the protein kinesin, which "walks" along microtubules at the expense of ATP.

axon. The "legs" are provided by a protein called *kinesin*, and the process is fueled by ATP (Figure 2.16). Kinesin moves material only from the soma to the terminal. All movement of material in this direction is called **anterograde transport**.

In addition to anterograde transport, there is a mechanism for the movement of material up the axon from the terminal to the soma. This process is believed to provide signals to the soma about changes in the metabolic needs of the axon terminal. Movement in this direction, from terminal to soma, is called **retrograde transport**. The molecular mechanism is similar to anterograde transport, except the "legs" for retrograde transport are provided by a different protein, *dynein*. Both anterograde and retrograde transport mechanisms have been exploited by neuroscientists to trace connections in the brain (Box 2.4).

Dendrites

The term *dendrite* is derived from the Greek for "tree," reflecting the fact that these neurites resemble the branches of a tree as they extend from the soma. The dendrites of a single neuron are collectively called a **dendritic tree**; each branch of the tree is called a *dendritic branch*. The wide variety of shapes and sizes of dendritic trees are used to classify different groups of neurons.

Because dendrites function as the antennae of the neuron, they are covered with thousands of synapses (Figure 2.17). The dendritic membrane under the synapse (the *postsynaptic* membrane) has many specialized protein molecules called **receptors** that detect the neurotransmitters in the synaptic cleft.

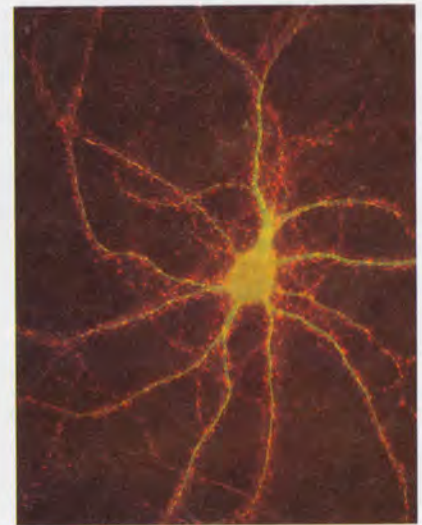


FIGURE 2.17

Dendrites receiving synaptic inputs from axon terminals. A neuron has been made to fluoresce green, using a method that reveals the distribution of a microtubule-associated protein. Axon terminals have been made to fluoresce orange-red, using a method to reveal the distribution of synaptic vesicles. The axons and cell bodies that contribute these axon terminals are not visible in this photomicrograph. (Source: *Neuron* 10 [Suppl.], 1993, cover image.)

Box 2.4



OF SPECIAL INTEREST

Hitching a Ride With Retrograde Transport

Fast anterograde transport of proteins in axons was shown by injecting the soma with radioactive amino acids. The success of this method immediately suggested a way to trace connections in the brain. For example, to determine where neurons in the eye send their axons, the eye was injected with radioactive proline, an amino acid. The proline was incorporated into proteins in the somata that were then transported to the axon terminals. By use of a technique called autoradiography, the location of radioactive axon terminals could be detected, thereby revealing the extent of the connection between the eye and the brain.

Researchers subsequently discovered that retrograde transport could also be exploited to work out connections in the brain. Strangely enough, the enzyme horseradish peroxidase (HRP) is selectively taken up by axon terminals and then transported retrogradely to the soma. A chemical reaction can then be initiated to visualize the location of the HRP in slices of brain tissue. This method is commonly used to trace connections in the brain (Figure A).

Some viruses also exploit retrograde transport to infect neurons. For example, the oral type of herpesvirus enters axon terminals in the lips and mouth and is then transported back to the parent cell bodies. Here the virus usually remains dormant until physical or emotional stress occurs (as on a first date), at which time it replicates and returns to the nerve ending, causing a painful cold sore. Similarly, the rabies virus enters the nervous system by

retrograde transport through axons in the skin. However, once inside the soma, the virus wastes no time in replicating madly, killing its neuronal host. The virus is then taken up by other neurons within the nervous system, and the process repeats itself again and again, usually until the victim dies.

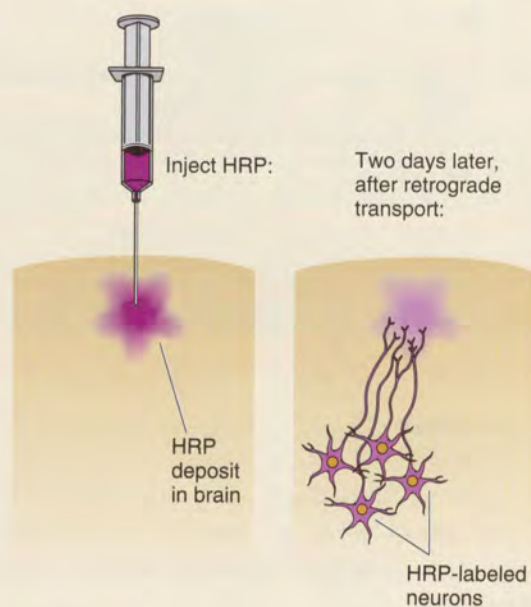


FIGURE A



The dendrites of some neurons are covered with specialized structures called **dendritic spines** that receive some types of synaptic input. Spines look like little punching bags that hang off the dendrite (Figure 2.18). The unusual morphology of spines has fascinated neuroscientists ever since their discovery by Cajal. They are believed to isolate various chemical reactions that are triggered by some types of synaptic activation. Spine structure is sensitive to the type and amount of synaptic activity. Unusual changes in spines have been shown to occur in the brains of individuals with cognitive impairments (Box 2.5). William Greenough of the University of Illinois at Urbana discovered that spine number is also very sensitive to the quality of the environment experienced during early development and in adulthood (Box 2.6).

FIGURE 2.18

Dendritic spines. This is a computer reconstruction of a segment of dendrite, showing the variable shapes and sizes of spines. Each spine is postsynaptic to one or two axon terminals. (Source: Harris and Stevens, 1989, cover image.)

Box 2.5



OF SPECIAL INTEREST

Mental Retardation and Dendritic Spines

The elaborate architecture of a neuron's dendritic tree is a good reflection of the complexity of its synaptic connections with other neurons. Brain function depends on these highly precise synaptic connections, which are formed during the fetal period and are refined during infancy and early childhood. Not surprisingly, this very complex developmental process is vulnerable to disruption. *Mental retardation* is said to have occurred if a disruption of brain development results in subaverage cognitive functioning that impairs adaptive behavior.

The use of standardized tests indicates that intelligence in the general population is distributed along a bell-shaped (Gaussian) curve. By convention, the mean intelligence quotient (IQ) is set to be 100. About two-thirds of the total population falls within 15 points (one standard deviation) of the mean, and 95% of the population falls within 30 points (two standard deviations). People with intelligence scores below 70 are considered to be mentally retarded if the cognitive impairment affects the person's ability to adapt his or her behavior to the setting in which he or she lives. Some 2–3% of humans fit this description.

Mental retardation has many causes. The most severe forms are associated with genetic disorders. An example is a condition called *phenylketonuria (PKU)*. The basic abnormality is a deficit in the liver enzyme that metabolizes the dietary amino acid phenylalanine. Infants born with PKU have an abnormally high level of the amino acid in the blood and brain. If the condition goes untreated, brain growth is stunted and severe mental retardation results. Another example is *Down syndrome*, which occurs when the fetus has an extra copy of chromosome 21, thus disrupting normal gene expression during brain development.

A second known cause of mental retardation is accidents during pregnancy and childbirth. Examples are maternal infections with German measles (rubella) and asphyxia during childbirth. A third cause of mental retardation is poor nutrition during pregnancy. An example is *fetal alcohol syndrome*, a constellation of developmental abnormalities that occur in children born to alcoholic mothers. A fourth cause, thought to account for the majority of cases, is environmental impoverishment—the lack of good nutrition, socialization, and sensory stimulation—during infancy.

While some forms of mental retardation have very clear physical correlates (e.g., stunted growth; abnormalities in the structure of the head, hands, and body), most cases have only behavioral manifestations. The brains of these individuals appear grossly normal. How, then, do

we account for the profound cognitive impairment? An important clue came in the 1970s from the research of Miguel Marin-Padilla, working at Dartmouth College, and Dominick Purpura, working at the Albert Einstein College of Medicine in New York City. Using the Golgi stain, they studied the brains of retarded children and discovered remarkable changes in dendritic structure. The dendrites of retarded children had many fewer dendritic spines, and the spines that they did have were unusually long and thin (Figure A). The extent of the spine changes was well correlated with the degree of mental retardation.

Dendritic spines are an important target of synaptic input. Purpura pointed out that the dendritic spines of mentally retarded children resemble those of the normal human fetus. He suggested that mental retardation reflects the failure of normal circuits to form in the brain. In the 3 decades since this seminal work was published, it has been established that normal synaptic development, including maturation of the dendritic spines, depends critically on the environment during infancy and early childhood. An impoverished environment during an early “critical period” of development can lead to profound changes in the circuits of the brain. However, there is some good news. Many of the deprivation-induced changes in the brain can be reversed if intervention occurs early enough. In Chapter 22, we will take a closer look at the role of experience in brain development.

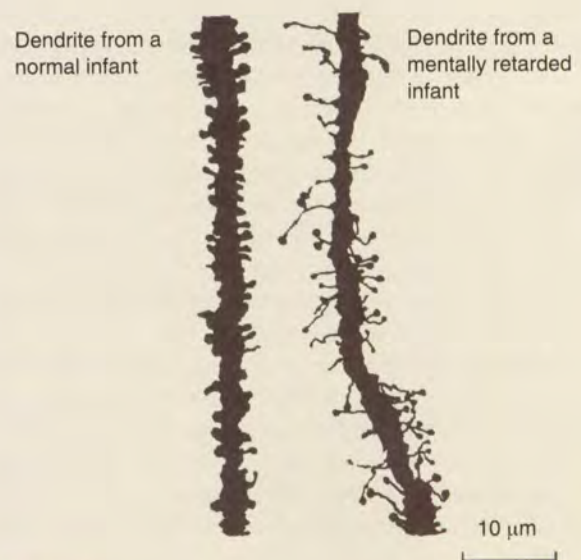


FIGURE A Normal and abnormal dendrites. (Source: Purpura, 1974, Fig. 2A.)

Box 2.6



PATH OF DISCOVERY

Spines and the Structural Basis of Memory

by William Greenough



It is a frequent misconception that scientific research results in simple, clear answers to questions. The truth is that almost every answer results in a whole battery of new questions. But the research serves to increase our understanding so that we know how to frame the new questions, and try to tackle them.

When we first saw polyribosomes below synapses in dendritic spines, we asked: "What are they doing away from the soma, where mRNA is usually translated?" An exciting possibility was that they might make special proteins that were involved in remodeling the synapse. I had seen that in animals exposed to enriched environments, new spines were made and other spines were remodeled from small to larger structures with—presumably—more efficient signaling. Could these synaptic polyribosomes be manufacturing crucial proteins that could account for the improved efficiency? Some kind of "memory proteins"?

I recalled a paper by Hollingsworth, in which he figured out a way to shear off synapses on spines, a preparation called synaptoneurosome (Figure A), and study them biochemically. My colleague Ivan Jeanne Weiler and I attempted to isolate the mRNA in synaptoneurosome to identify the crucial "memory proteins." Repeated failures, due apparently to mRNA degradation, both frustrated and slowed us. But eventually we found that some of the mRNA was being incorporated into polyribosomes, meaning that protein was being synthesized. We then discovered that this protein synthesis was strongly increased when we stimulated the synaptoneurosome with the neurotransmitter glutamate, and we were able to identify the glutamate receptor that was responsible. We now thought we were poised to isolate the "memory proteins" from the newly assembled polyribosomes.

We set up to harvest polyribosomes from stimulated and unstimulated synaptoneurosome to see which mRNAs were translated in response to glutamate. About this time, Jim Eberwine (University of Pennsylvania) told us about the mRNAs he had isolated from single dendrites. We looked for their translation in our new system, and the first one that was increased by stimulation turned out to be the fragile X mental retardation protein (FMRP). This has led

us to study brain abnormalities in fragile X syndrome, the largest cause of inherited mental retardation.

Do we now, at last, have a "memory protein"? Not yet, because FMRP is not a structural or receptor protein, but a protein that binds mRNA. It seems to orchestrate the translation of an impressive array of other mRNAs, few of which, at first glance, look like "memory proteins." But, if FMRP is a key to how the synapse changes itself, then we think we are still on the right track to understand synaptic remodeling. Indeed, people with fragile X syndrome have immature-appearing synapses.

So we have been wrong a lot of the time, and frustrated on a near-weekly basis. Nature is full of surprises, but we have increased our understanding of synaptic protein synthesis. Indeed, we are now asking new questions that may help us understand and possibly treat the leading inherited form of mental retardation. It was worth the effort.

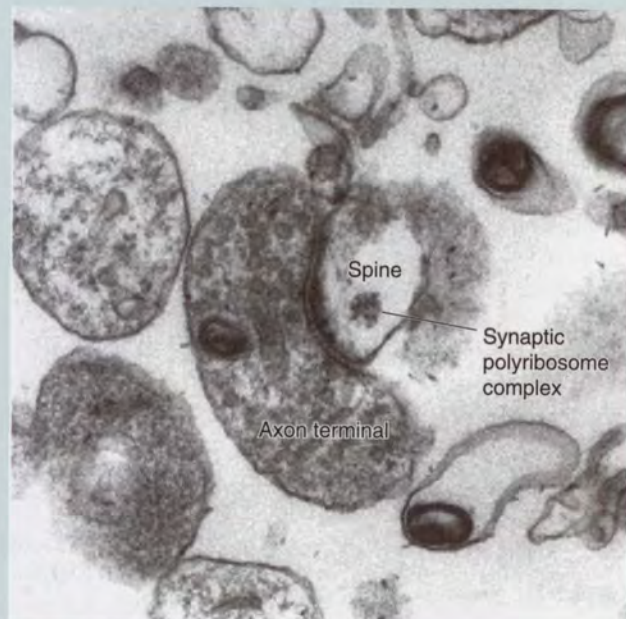


FIGURE A

Electron micrograph of a synaptoneurosome. The spine is about 1 micron in diameter. (Courtesy of William Greenough.)

For the most part, the cytoplasm of dendrites resembles that of axons. It is filled with cytoskeletal elements and mitochondria. One interesting difference was discovered by neuroscientist Oswald Steward. He found that polyribosomes can be observed in dendrites, often right under spines. Steward's research suggests that synaptic transmission can actually direct local protein synthesis in some neurons. In Chapter 25, we will see that synaptic regulation of protein synthesis is crucial for information storage by the brain.

▼ CLASSIFYING NEURONS

It is unlikely that we can ever hope to understand how each of the hundred billion neurons in the nervous system uniquely contributes to the function of the brain. But what if we could show that all the neurons in the brain can be divided into a small number of categories, and that within each category, all neurons function identically? The complexity of the problem might then be reduced to understanding the unique contribution of each category, rather than each cell. It is with this hope that neuroscientists have devised schemes for classifying neurons.

Classification Based on the Number of Neurites

Neurons can be classified according to the total number of neurites (axons and dendrites) that extend from the soma (Figure 2.19). A neuron that has a single neurite is said to be **unipolar**. If there are two neurites, the cell is **bipolar**, and if there are three or more, the cell is multipolar. Most neurons in the brain are **multipolar**.

Classification Based on Dendrites

Dendritic trees can vary widely from one type of neuron to another. Some have inspired elegant names like "double bouquet cells." Others have less interesting names, such as "alpha cells." Classification is often unique to a particular part of the brain. For example, in the cerebral cortex (the structure that lies just under the surface of the cerebrum), there are two broad classes: **stellate cells** (star-shaped) and **pyramidal cells** (pyramid-shaped) (Figure 2.20).

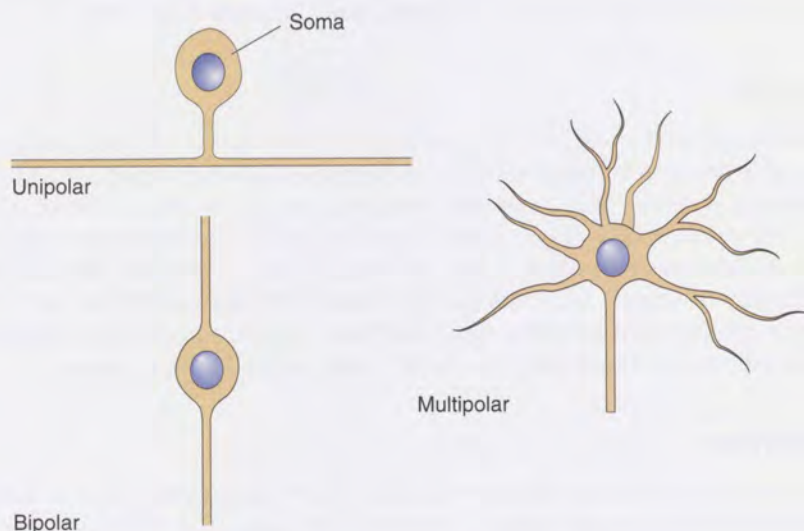
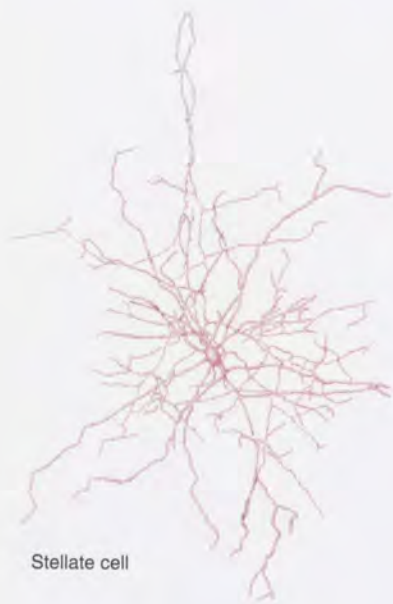


FIGURE 2.19

Classification of neurons based on the number of neurites.



Stellate cell



Pyramidal cell

Another simple way to classify neurons is according to whether their dendrites have spines. Those that do are called **spiny**, and those that do not are called **aspinous**. These dendritic classification schemes can overlap. For example, in the cerebral cortex, all pyramidal cells are spiny. Stellate cells, on the other hand, can be either spiny or aspiny.

Classification Based on Connections

Information is delivered to the nervous system by neurons that have neurites in the sensory surfaces of the body, such as the skin and the retina of the eye. Cells with these connections are called **primary sensory neurons**. Other neurons have axons that form synapses with the muscles and command movements; these are called **motor neurons**. But most neurons in the nervous system form connections only with other neurons. According to this classification scheme, these cells are all called **interneurons**.

Classification Based on Axon Length

Some neurons have long axons that extend from one part of the brain to the other; these are called *Golgi type I neurons*, or projection neurons. Other neurons have short axons that do not extend beyond the vicinity of the cell body; these are called *Golgi type II neurons*, or local circuit neurons. In the cerebral cortex, for example, pyramidal cells usually have long axons that extend to other parts of the brain and are therefore Golgi type I neurons. In contrast, stellate cells have axons that never extend beyond the cerebral cortex and are therefore Golgi type II neurons.

Classification Based on Neurotransmitter

The classification schemes presented so far are based on the morphology of neurons as revealed by a Golgi stain. Newer methods that enable neuroscientists to identify which neurons contain specific neurotransmitters have resulted in a scheme for classifying neurons based on their chemistry. For example, the motor neurons that command voluntary movements all release the neurotransmitter *acetylcholine* at their synapses. These cells are therefore also classified as *cholinergic*, meaning that they use this particular neurotransmitter. Collections of cells that use a common neurotransmitter make up the brain's neurotransmitter systems (see Chapters 6 and 15).

▼ GLIA

We have devoted most of our attention in this chapter to the neurons. While this decision is justified by the state of current knowledge, some neuroscientists consider glia to be the “sleeping giants” of neuroscience. One day, they suppose, it will be shown that glia contribute much more importantly to information processing in the brain than is currently appreciated. At present, however, the evidence indicates that glia contribute to brain function mainly by supporting neuronal functions. Although their role may be subordinate, without glia, the brain could not function properly.

Astrocytes

The most numerous glia in the brain are called **astrocytes** (Figure 2.21). These cells fill the spaces between neurons. The space that remains between the neurons and the astrocytes in the brain measures only about 20 nm wide. Consequently, astrocytes probably influence whether a neurite can grow or retract. And when we speak of fluid “bathing” neurons in the brain,

FIGURE 2.20
Classification of neurons based on dendritic tree structure. Stellate cells and pyramidal cells, distinguished by the arrangement of their dendrites, are two types of neurons found in the cerebral cortex.

it is more like a soaking from a hose than immersion in a swimming pool.

An essential role of astrocytes is regulating the chemical content of this *extracellular space*. For example, astrocytes envelop synaptic junctions in the brain, thereby restricting the spread of neurotransmitter molecules that have been released. Astrocytes also have special proteins in their membranes that actively remove many neurotransmitters from the synaptic cleft. A recent and unexpected discovery is that astrocytic membranes also possess neurotransmitter receptors that, like the receptors on neurons, can trigger electrical and biochemical events inside the glial cell. Besides regulating neurotransmitters, astrocytes also tightly control the extracellular concentration of several substances that have the potential to interfere with proper neuronal function. For example, astrocytes regulate the concentration of potassium ions in the extracellular fluid.

Myelinating Glia

Unlike astrocytes, the primary function of **oligodendroglial** and **Schwann cells** is clear. These glia provide layers of membrane that insulate axons. Boston University anatomist Alan Peters, a pioneer in the electron microscopic study of the nervous system, showed that this wrapping, called **myelin**, spirals around axons in the brain (Figure 2.22). Because the axon fits inside the spiral wrapping like a sword in its scabbard, the name *myelin sheath* describes the entire covering. The sheath is interrupted periodically, leaving a short length where the axonal membrane is exposed. This region is called a **node of Ranvier** (Figure 2.23).

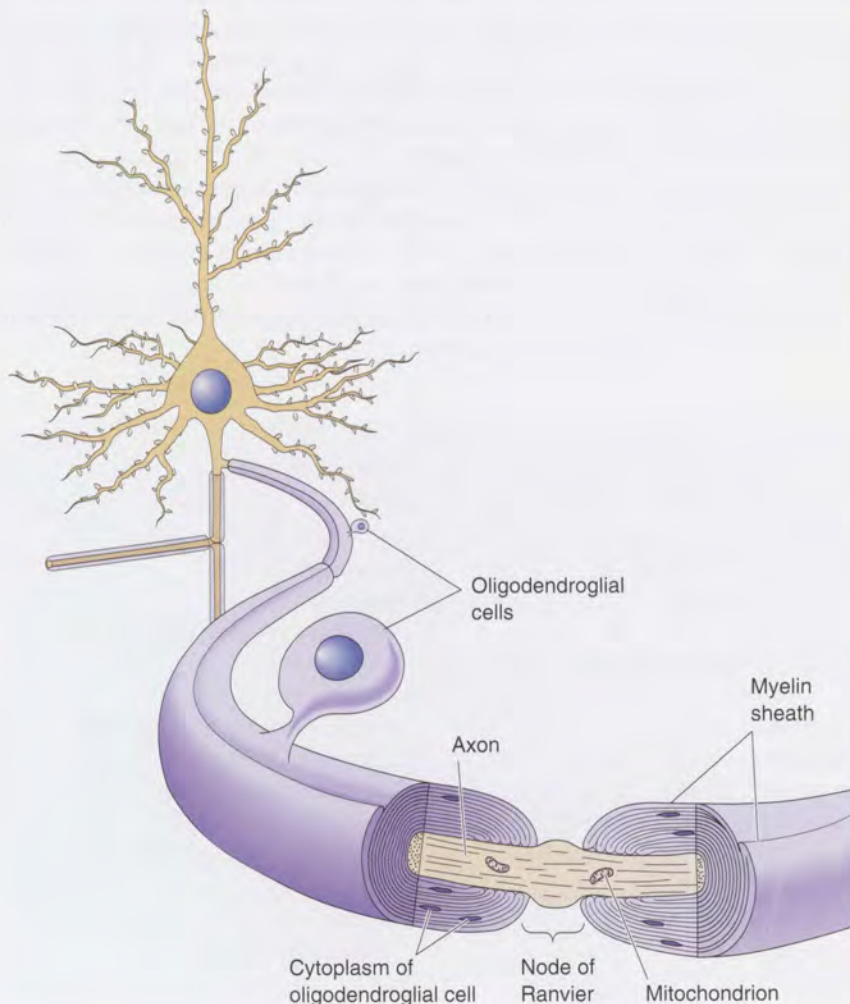


FIGURE 2.21

An astrocyte. Astrocytes fill most of the space in the brain that is not occupied by neurons and blood vessels.

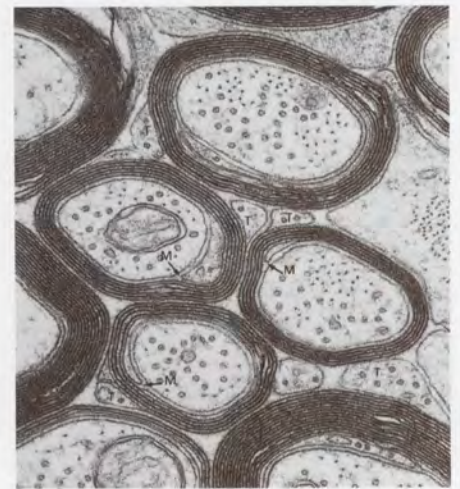


FIGURE 2.22

Myelinated optic nerve fibers cut in cross section. (Source: Courtesy of Dr. Alan Peters.)

FIGURE 2.23

An oligodendroglial cell. Like the Schwann cells found in the nerves of the body, oligodendroglia provide myelin sheaths around axons in the brain and spinal cord. The myelin sheath of an axon is interrupted periodically at the nodes of Ranvier.

We will see in Chapter 4 that myelin serves to speed the propagation of nerve impulses down the axon. Oligodendroglia and Schwann cells differ in their location and some other characteristics. For example, oligodendroglia are found only in the central nervous system (brain and spinal cord), while Schwann cells are found only in the peripheral nervous system (parts outside the skull and vertebral column). Another difference is that one oligodendroglial cell will contribute myelin to several axons, while each Schwann cell myelinates only a single axon.

Other Non-Neuronal Cells

Even if we eliminated every neuron, every astrocyte, and every oligodendroglial cell, other cells would still remain in the brain. For the sake of completeness, we must mention these other cells. First, special cells called **ependymal cells** provide the lining of fluid-filled ventricles within the brain, and they also play a role in directing cell migration during brain development. Second, a class of cells called **microglia** function as phagocytes to remove debris left by dead or degenerating neurons and glia. Finally, the vasculature of the brain—arteries, veins, and capillaries—would still be there.

▼ CONCLUDING REMARKS

Learning about the structural characteristics of the neuron provides insight into how neurons and their different parts work, because structure correlates with function. For example, the absence of ribosomes in the axon correctly predicts that proteins in the axon terminal must be provided by the soma via axoplasmic transport. A large number of mitochondria in the axon terminal correctly predicts a high energy demand. The elaborate structure of the dendritic tree appears ideally suited for the receipt of incoming information, and indeed, this is where most of the synapses are formed with the axons of other neurons.

From the time of Nissl, it has been recognized that an important feature of neurons is the rough ER. What does this tell us about neurons? We have said that rough ER is a site of the synthesis of proteins destined to be inserted into the membrane. We will now see how the various proteins in the neuronal membrane give rise to the unique capabilities of neurons to transmit, receive, and store information.



KEY TERMS

Introduction

neuron (p. 24)
glial cell (p. 24)

The Neuron Doctrine

histology (p. 25)
Nissl stain (p. 25)
cytoarchitecture (p. 25)
Golgi stain (p. 26)
cell body (p. 26)
soma (p. 26)
perikaryon (p. 26)
neurite (p. 26)
axon (p. 26)
dendrite (p. 26)
neuron doctrine (p. 27)

The Prototypical Neuron

cytosol (p. 28)
organelle (p. 28)
cytoplasm (p. 30)
nucleus (p. 30)
chromosome (p. 30)
DNA (deoxyribonucleic acid) (p. 30)
gene (p. 30)
gene expression (p. 30)
protein (p. 30)
protein synthesis (p. 30)
mRNA (messenger ribonucleic acid) (p. 30)
transcription (p. 30)
promoter (p. 30)

transcription factor (p. 30)
RNA splicing (p. 30)
amino acid (p. 31)
translation (p. 31)
ribosome (p. 31)
rough endoplasmic reticulum (rough ER) (p. 33)
polyribosome (p. 33)
smooth endoplasmic reticulum (smooth ER) (p. 34)
Golgi apparatus (p. 34)
mitochondrion (p. 34)
ATP (adenosine triphosphate) (p. 35)
neuronal membrane (p. 35)
cytoskeleton (p. 35)

microtubule (p. 35)
 microfilament (p. 38)
 neurofilament (p. 38)
 axon hillock (p. 38)
 axon collateral (p. 38)
 axon terminal (p. 39)
 terminal bouton (p. 39)
 synapse (p. 39)
 terminal arbor (p. 39)
 innervation (p. 39)
 synaptic vesicle (p. 39)
 synaptic cleft (p. 40)
 synaptic transmission (p. 40)
 neurotransmitter (p. 40)

axoplasmic transport (p. 40)
 anterograde transport (p. 41)
 retrograde transport (p. 41)
 dendritic tree (p. 41)
 receptor (p. 41)
 dendritic spine (p. 42)

Classifying Neurons

unipolar neuron (p. 45)
 bipolar neuron (p. 45)
 multipolar neuron (p. 45)
 stellate cell (p. 45)
 pyramidal cell (p. 45)
 spiny neuron (p. 46)

aspinous neuron (p. 46)
 primary sensory neuron (p. 46)
 motor neuron (p. 46)
 interneuron (p. 46)

Glia

astrocyte (p. 46)
 oligodendroglial cell (p. 47)
 Schwann cell (p. 47)
 myelin (p. 47)
 node of Ranvier (p. 47)
 ependymal cell (p. 48)
 microglial cell (p. 48)



REVIEW QUESTIONS

1. State the neuron doctrine in a single sentence. To whom is this insight credited?
2. Which parts of a neuron are shown by a Golgi stain that are not shown by a Nissl stain?
3. What are three physical characteristics that distinguish axons from dendrites?
4. Of the following structures, state which ones are unique to neurons and which are not: nucleus, mitochondria, rough ER, synaptic vesicle, Golgi apparatus.
5. What are the steps by which the information in the DNA of the nucleus directs the synthesis of a membrane-associated protein molecule?
6. Colchicine is a drug that causes microtubules to break apart (depolymerize). What effect would this drug have on anterograde transport? What would happen in the axon terminal?
7. Classify the cortical pyramidal cell based on (a) the number of neurites, (b) the presence or absence of dendritic spines, (c) connections, and (d) axon length.
8. What is myelin? What does it do? Which cells provide it in the central nervous system?



FURTHER READING

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