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Editorial Board Member of *World Journal of Gastroenterology*,
Dr. Mukaddes Esrefoglu, Professor, Department of Histology and Embryology,
Inonu University, 44280 Malatya, Turkey

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EDITORIAL OFFICE
Jian-Xia Cheng, Director
Jin-Lei Wang, Vice Director
World Journal of Gastroenterology
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-59080039
Fax: +86-10-85381893
E-mail: wjg@wjgnet.com
<http://www.wjgnet.com>

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History, ethics, advantages and limitations of experimental models for hepatic ablation

Seok Ling Ong, Gianpiero Gravante, Matthew S Metcalfe, Ashley R Dennison

Seok Ling Ong, Gianpiero Gravante, Matthew S Metcalfe, Ashley R Dennison, Department of Hepatobiliary and Pancreatic Surgery, Leicester General Hospital, Leicester LE5 4PW, United Kingdom

Author contributions: Ong SL, Gravante G, Metcalfe MS, Dennison AR were responsible for the acquisition of data, analysis and interpretation, drafting the article or revising it critically for important intellectual content and final approval of the version to be published; Ong SL, Dennison AR were responsible for substantial contributions to conception and design.

Correspondence to: Dr. Gianpiero Gravante, Department of Hepatobiliary and Pancreatic Surgery, Leicester General Hospital, Gwendolen Road, Leicester LE5 4PW, United Kingdom. ggravante@hotmail.com

Telephone: +44-116-2588244 Fax: +44-116-2584708

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combine advantages of both previous models.

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Abstract

Numerous techniques developed in medicine require careful evaluation to determine their indications, limitations and potential side effects prior to their clinical use. At present this generally involves the use of animal models which is undesirable from an ethical standpoint, requires complex and time-consuming authorization, and is very expensive. This process is exemplified in the development of hepatic ablation techniques, starting experiments on explanted livers and progressing to safety and efficacy studies in living animals prior to clinical studies. The two main approaches used are *ex vivo* isolated non-perfused liver models and *in vivo* animal models. *Ex vivo* non perfused models are less expensive, easier to obtain but not suitable to study the heat sink effect or experiments requiring several hours. *In vivo* animal models closely resemble clinical subjects but often are expensive and have small sample sizes due to ethical guidelines. Isolated perfused *ex vivo* liver models have been used to study drug toxicity, liver failure, organ transplantation and hepatic ablation and

INTRODUCTION

Progress in science and medicine depends on new ideas being developed, tested and introduced into mainstream practice. Animal experimentation is used not only in medicine and surgery but also in science, the pharmaceutical industry, the military and educational establishments. In medicine it is essential to establish beyond doubt the safety of a drug, instrument or procedure before it even begins phase 1 of clinical trials. To establish the safety of this heterogeneous range of treatments the medical profession has frequently had no option but to the use of live animals. Although this practice is highly undesirable and many people are morally opposed to these experiments, unless and until viable alternatives are available legislation prevents the introduction of these new drugs and procedures without this prior testing. Indeed virtually all drugs and surgical treatments that are in use today have been tested on animals.

Although the best treatment for liver tumour remains surgical resection^[1] the majority of patients are still affected by unresectable lesions. To these patients hepatic ablation provides an opportunity to increase survival^[2]. Experimental research over the last twenty years has seen the development of a number of ablative modalities and

several of these have been applied clinically^[3,4]. Ablation experiments are now focused to increase the diameter of the ablation zone to achieve better tumor margins^[5-12], or to conduct safe ablations on lesions positioned close to major vessels^[13-16]. Numerous experimental models are available, each with different advantages, disadvantages and ethical implications. It is therefore imperative that the researcher that wants to explore the field of experimental hepatic ablation is aware of their characteristics.

HISTORICAL BACKGROUND OF ANIMAL EXPERIMENTATION

The Greeks almost 2500 years ago described the earliest recorded animal experiments. Aristotle (384-322 BC), and Erasistratus (304-258 BC)^[17,18] both performed studies on live animals and in 2nd century Rome Galen who is known as the father of vivisection dissected live goats and pigs^[19]. These studies continued into the Roman era and were then passed on to medical schools in Arabia. The practice died out after this and was absent completely in the Dark Ages only being revived in Italy in the 16th century. Subsequently live animals have been used throughout history to study a wide range of problems and to assess new treatments particularly drugs and vaccines. In the 17th century, many pivotal discoveries came to light because of these studies including the understanding of lung function and the circulation of the blood. In the 1840's general anesthesia emerged (initially ether and chloroform) and it became possible to study unconscious animals. In 1881, 250 experiments were carried out in Britain, the first year that records were kept of the procedures carried out on animals. The following year (1822) the first animal protection law was passed in Britain and in 1876 the Cruelty to Animals Act came into being and was the first law specifically designed to regulate animal testing.

Many advances in our understanding did result from these animal studies including those in basic science and also medicine. These included Antoine Lavoisier demonstrating that respiration was a form of combustion using guinea pigs in calorimeters^[20], Stephen Hales measuring blood pressure in the horses, and in the 1880's Louis Pasteur demonstrating the concept of "germs" by giving anthrax to sheep^[21]. Also in the 1880's and 1890's Emil von Behring was able to isolate the diphtheria toxin and not only demonstrate its effects in guinea pigs but also, by 1898 produce immunity by the injection of a mixture of toxin and antitoxin, for which he was awarded the Nobel Prize in Physiology and Medicine in 1901. In 1921 Banting ligated the pancreatic duct of dogs and demonstrated that pancreatic isolates could be used to keep these diabetic animals alive, and by 1922 working with John Macloed he isolated insulin from bovine sources and famously treated Leonard Thompson, a 14-year-old diabetic boy with diabetes^[22].

OBJECTIONS TO LIVE ANIMAL EXPERIMENTS

Despite these advances and those that took place subsequently many people remained understandably opposed to any form of live animal experimentation. Claude Bernard was known as the "prince of vivisectors" and the father of physiology^[23] but ironically his wife Marie Francoise Martin founded the first anti-vivisection society in France in 1883 and famously wrote in 1865 that "the science of life is a superb and dazzlingly lighted hall which may be reached only by passing through a long and ghastly kitchen"^[23,24]. Opposition to animal experimentation continued and many eminent scientists voiced their disapproval. Charles Darwin in 1871 wrote to Ray Lancaster in reply and stated "You ask about my opinion on vivisection. I quite agree that it is justifiable for real investigations on physiology; but not for mere damnable and detestable curiosity. It is a subject which makes me sick with horror, so I will not say another word about it, else I shall not sleep tonight"^[25].

Objections to live animal experiments emerged in the United States in the 1860's and Henry Bergh founded the American Society for the Prevention of Cruelty to Animals. The first movement which specifically opposed vivisection was the American Antivivisection Society, which was founded in 1883. The continued opposition to the use of animals together with the dramatic increase in the number of procedure that were being performed (in 2002 in the United Kingdom 2.7 million live animal experiments were authorized and in the United States, it is estimated that between 19 and 29 million experiments were performed although exact estimates were difficult because 90% of the animals were rats, mice and other species recently exempt from legislation) encouraged many researchers in science and medicine to seek other methods to test drugs and procedures and much of this ethos has been incorporated into legislation or guidelines. Although British law requires that any new drug be tested on at least two different species of animals (one must be a large non-rodent), United Kingdom regulations in respect of animal experimentation are very strict and the Animals Act of 1986 requires that no animal experiments be conducted if there is a realistic alternative. It also exhorts investigators to examine their research carefully to determine the smallest number of animals that may be used to answer the questions posed by the research.

There remains however a dilemma with both ethical and legislative dimensions. All scientists agree that animal experimentation is unacceptable if there is a viable alternative but legislation will not allow the introduction of new products without rigorous testing. In 1988, the American Medical Association published a white paper defending biomedical research in animals^[26], and stated that "In fact, virtually every advance in medical science

in the 20th century, from antibiotics and vaccines to antidepressant drugs and organ transplants, has been achieved either directly or indirectly through the use of animals in laboratory experiments". In Europe European Centre for the Validation of Alternative Methods was established in October 1991 by a communication from the Commission to the Council and the Parliament. The aim was to try and organize research and exchange information to limit the need for animal experimentation. As defined in the communication it was felt that this could be achieved in four ways: (1) to coordinate the validation of alternative test methods at the European Union level; (2) to act as a focal point for the exchange of information on the development of alternative test methods; (3) to set up, maintain and manage a database on alternative procedures; and (4) to promote dialogue between legislators, industries, biomedical scientists, consumer organizations and animal welfare groups, with a view to the development, validation and international recognition of alternative test methods.

Although there has been considerable progress in these areas, notably with the use of fresh and cryo-preserved cells (particularly hepatocytes) problems remain when it is necessary to investigate a treatment prior to its use in patients. The problem is particularly acute in the field of biotechnology in which many of the devices that are proposed and developed are potentially harmful and effects in living tissue are essentially unknown. Today it is certainly possible to computer model some of the effects (widely used in the pharmaceutical industry to predict the usefulness and safety of new drugs) but many results remain unpredictable. Fresh tissue can be used and will provide answers if the procedure takes a short time and its effects and results do not rely on changes (particularly at the cellular level) that require interaction with living cells or evolve over a significant time period (several hours or longer). Unfortunately many devices do produce changes due to interaction with normally vascularized tissue and frequently take prolonged periods to produce effects. With these the only way to date that the results could be examined in detail was in living animals. In addition although prototypes could be studied in small animals instruments that have been developed for human use because of their size can often only be studied in large animals.

LIVER ABLATION EXPERIMENTAL MODELS

The number of patients with liver lesions, which are potentially suitable for treatment, continues to rise with advances in critical care, anesthesia, surgery and oncology. At present, the gold standard remains surgical resection that produces long-term survivors and cure in a significant number of patients. The advent of laparoscopic resection has further widened the indications but by far the most attractive concept remains ablation of

the liver (for primary or secondary tumors) containing the tumour. The ablation techniques currently in place for the treatment of liver tumors include radiofrequency ablation (RFA), microwave ablation (MWA), cryotherapy, percutaneous ethanol injection, high intensity focused ultrasound, interstitial laser photocoagulation, electrolysis and bimodal electric tissue ablation^[4]. The additional availability of methods for real-time monitoring to ensure that a sufficient volume is ablated (to engulf the tumor and reduce the likelihood of local recurrence) has seen an upsurge in the number of procedures performed and the consequent emergence of survival data that are very encouraging. In addition there is evidence that ablative techniques produce immunologically active components that may confer some extra survival advantage but also systemic effects with potentially harmful consequences.

The development of any new hepatic ablative therapy is a long process that requires a detailed understanding of the basic physical and chemical properties of the ablative modality and their application in clinical settings. It includes designing equipment suitable for clinical application, identifying methods of assessing tissue response to ablation, optimizing treatment monitoring, exploring local and systemic effect of the ablation, and finally, demonstrating the safety and efficacy in humans. Understanding of the tissue temperature changes and radiological evolution of lesion generated by each ablative modality is crucial before it can be used in a clinical setting. Further problems occur with some of the techniques that rely on temperature changes for their effectiveness. Particularly with RFA and MWA the effect can be significantly attenuated in the vicinity of large blood vessels due to the "heat sink effect" where the large flow in the vessels conducts heat away sufficiently rapidly to interfere with the treatment and potentially leave viable tumor cells around these vessels. Flows within the hepatic vessels have been shown to reduce the ablative energy via heat sink effects resulting in a greater energy requirement when ablating in close proximity to larger vessels with an associated higher risk of vascular and bile duct complications^[15,27]. For the same reasons, thermal ablation techniques particularly are influenced by the effects of the separate occlusion of the hepatic artery, portal vein or both (Pringle maneuver). This delicate balance between safety and efficacy of ablation has been the focus of many experimental studies^[15,28,29].

One further attraction of ablative techniques is the fact that they can often be performed percutaneously or laparoscopically and considerable effort has been expended in the development of suitable probes. These probes can be complex and are generally modeled initially on computers to predict the likely outcome after the delivery of a set amount of energy. Unfortunately the alterations in tissue resistance and electrical and thermal conductivity consequent upon the use of the probes, together with the complex designs (and frequent use of multiple probes or probes that assume unpredictable

shapes after deployment) means that to date it has been essential to test the devices in animal models. These will also be necessary to “calibrate” all new devices and produce standard dose-response curves, so that in addition to real-time monitoring a prediction can be made about the amount of energy that needs to be delivered to ablate a predetermined volume.

The increasing popularity and deployment of these techniques means that to accomplish these aims would require the use of a large number of experimental animals, and because of this it is valuable to consider whether there is an alternative approach that can produce reliable data of sufficient quality to facilitate the incorporation of these methods into clinical practice without the need for subsequent large animal studies. *Ex vivo* perfused liver models have been developed recently for the study of the hepatic toxicity of new pharmacological agents^[30-32], and in an attempt to improve the outcome from organ transplantation. Organ transplantation models, which use pulsatile perfusion after harvesting, would seem to offer an excellent alternative that could closely mimic the *in vivo* situation and allow data to be collected for up to 24 h. In a sufficiently well developed perfusion model in which parameters can be accurately controlled it seems likely that data generated would be genuinely useful and achieve the aim of avoiding the use of live animal models.

EX VIVO NON-PERFUSED MODELS

The most commonly used model for the study of hepatic ablation is liver explanted from animals. Bovine and porcine livers are most frequently used because of their similarity to human livers in terms of size and density, and because they are farm animals that are readily available and inexpensive. In addition long-term use has resulted in a familiarity with the animals' anatomy that is essential in studies on the liver where the lobulation and blood supply are an important determinants of outcome. Although the organ is not perfused the position of the ablation in respect of vessels and bile ducts remains important. Their size also allows a number of ablations to be carried out on each individual model, which minimizes the number of animals required.

These models can be used to examine the relationship between ablative energy and lesion size, to assess the effect of different antenna configurations on lesion size and to study the dielectric properties of liver tissues^[33-36]. Crocetti *et al*^[37] used *ex vivo* calf livers to assess the feasibility and validity of fusion imaging that combines ultrasound and computed tomography in the monitoring of RFA. Unfortunately the results of this study were not directly transferable to the clinical setting because of the absence of respiratory excursion and subject motions of this model. The absence of perfusion within *ex vivo* models also limits their ability to assess the effect of blood flow on the size of the lesion created by

ablation with a set amount of energy. Furthermore detailed studies of the cellular changes in the ablated area cannot be conducted due to the ischemia produced by explantation and the rapid cessation of metabolic and physiochemical processes.

IN VIVO MODELS

In view of the limitation of *ex vivo* models particularly with respect to the absence of blood flow, *in vivo* animal models have been widely used for the study of hepatic ablation. Data generated from these studies are very reliable and the close similarity to results in humans allows the experience to be directly transferred to clinical practice. Small laboratory animal models such as murine models are easier to handle but have significant differences in organ size, function and anatomy compared to the human liver. The relatively small organ size also limits the number of ablations that can be carried out per liver. In contrast, *in vivo* porcine liver closely resembles human livers in size, vascularity and metabolic function and although the shape is different the organ is also lobulated in a similar way to the human liver. This allows comparisons to be drawn with respect of the positioning of probes, real dosage levels and consequences of treatment. The result is that the porcine model is by far the most commonly used at present for the final evaluation of all ablative techniques.

The two major approaches to date for the study of hepatic ablation in animals, *ex vivo* non-perfused liver models and *in vivo* animal studies, and their respective advantages and limitations are compared in Table 1. A number of studies have been carried out simultaneously on *in vivo* and *ex vivo* non-perfused models to try and assess the magnitude of the changes produced and also to evaluate the specific differences. The volume of the ablated regions is always significantly smaller in the *in vivo* models when similar energy of ablation is applied^[5,38,39]. These findings can probably be attributed to the presence of vascular flow, which removes some of the energy (heat) during the treatment in the *in vivo* models and hence these models are used to examine the influence of vascular occlusion on the size of ablated area. *In vivo* models also allow histological study of different zones, and the evolution of these zones in lesions generated by thermal ablations.

Results generated from *in vivo* models are directly transferable to clinical practice and are at present the gold standard for pre-clinical experimental studies of hepatic ablation. However the use of living animals requires specific laboratory facilities and research teams with expertise in anesthetizing and handling different species. The size and temperament of larger animal models such as porcine and bovine models poses practical problems when animals need to be anesthetized, moved or examined post-procedure (particularly if blood sampling is required).

Table 1 Advantages and limitations of *ex vivo* and *in vivo* models for experimental study of hepatic ablation

Models	Applications	Advantages	Limitations
<i>Ex vivo</i> (non perfused) models	Compare the efficacy of different antenna configurations	Allow histological examination of whole lesion to study zones of ablation	Non-physiological
	Trial of different energy setting	Cheap Larger study sample size Easy to manipulate during experiment Does not require ethical approval/animal license	Homogenous parenchyma Absence of respiratory excursion and subject motions Lack of cooling effect secondary to tissue perfusion Unable to study heat sink effect
<i>In vivo</i> models	Study of lesion evolution over time	Small animals	Small animals
	Histological examination of lesion	Easier to handle	Small volume of liver
	Study of heat sink effect and the effect of bile duct cooling	Cheaper	
	Study of systemic responses to ablation	Ability to have larger sample size	Limit number of ablation on each liver
	Study of the effect of large volume ablation (in larger animals)		Not suitable for the study of large volume ablation
		Large animals Closer resemblance to human liver in terms of size and physiology More ablations can be carried out in each liver	Large animals Size and temperament poses challenges during anesthesia Difficult vascular access in porcine models Also limited by strict ethic regulation Small study sample size
			Common limitations
			Expensive
			Expertise in animal handling and anesthesia is required
			Duration of study is limited to the lifespan of the model
Isolated perfused <i>ex vivo</i> liver models	Study of lesion evolution over time	Cheaper than <i>in vivo</i> experiments	Absence of interacting organ systems which may have a role in generating systemic response
	Study of heat sink effect and the effect of bile duct cooling	Sophisticated and accurate manipulation of hepatic inflow (e.g., Pringle manoeuvre)	Unable to assess the impact of ablation on end organs
	Study of early inflammatory response	Does not require ethical approval Greater control of perfusion characteristics (e.g., portal vein and hepatic arterial flows and pressures)	Perfusion circuit itself may activate some degree of systemic response

EX VIVO PERFUSED MODELS

Ex vivo non-perfused and *in vivo* models have significant advantages with respect to specific scientific questions but an alternative model, the isolated perfused *ex vivo* model, theoretically would have most of the advantages of both. A normothermic liver perfusion system using autologous blood maintains physiological and metabolic functions and the hemodynamic changes resemble those of *in vivo* models with good preservation of liver architecture^[40-42]. The organ would be perfused over a number of hours, and within this period valuable data could be produced within a relatively inexpensive model with no ethical problems and no requirement for licensing (Table 1)^[43-45]. This should permit the histological study of lesions generated by hepatic thermal ablation, real-time monitoring of the evolution of the lesions and experiments designed to study the “heat sink” effect with different modalities. In addition, with a sufficiently sophisticated and stable model it should also be possible (by venous sampling) to study the early inflammatory responses following hepatic ablation^[44,46].

Liver perfusion studies date back as far as the 19th century when they were first performed for research on physiological function of the liver^[47]. A number of

studies have been conducted subsequently to establish valid models of liver perfusion using asanguineous perfusates or autologous blood^[41,48-50]. These models have been used for physiological research, treatment of liver failure, studies of metabolism and toxicity of new pharmacological compounds and in transplant units for the recovery and preservation of organs, which are to be implanted^[51-53]. Hildebrand *et al.*^[54] have advocated the use of a perfused *ex vivo* liver model for the training in laparoscopic RFA.

For the *ex vivo* perfused model the main limitation is the limited lifespan and the determination of intervals during this lifespan when valid data can be extracted. Nevertheless perfusion models, and particularly those using autologous blood and pulsatile perfusions, have become very sophisticated and physiological parameters can be very accurately maintained for many hours. The interaction of blood with non-biological surfaces of the perfusion circuit does activate a number of biological pathways producing some degree of systemic response and interpretation of responses to ablation will have to take these into account. However the response will be the same for each experiment and should not interfere with comparisons of different dose responses or different modalities^[55]. The absence of other interacting

organs may also affect the extent of systemic responses following hepatic ablation but this model still potentially represents an exciting new method of studying these new devices while avoiding the need to use live animals. The true potential and weakness of an isolated perfused liver model for the study of hepatic ablation remain to be seen (and will be determined by future research) but recent advances have certainly developed the model to the stage that merits this research^[56].

FUTURE DIRECTIONS AND ADVANCES

To overcome the limitations of disconnecting the liver from the remaining organs, Chung *et al.*^[56] recently created the first multiorgan *ex vivo* perfused model in which the liver was serially connected with the kidney. The viability of both organs and the inflammatory reaction elicited have been investigated in two separate studies^[56,57]. The addition of the kidney to the circuit improved the biochemical milieu of the circuit and consented a more physiological environment for those experiments requiring strict conditions^[57]. At the same time, the combination of two organs in the circuit did not increase the cytokine production compared to the classic “liver-alone” circuit^[56] and therefore could be used to test the inflammatory reaction produced by ablative techniques^[44] without significant interferences by the newly added organ.

CONCLUSION

The key advantages of isolated perfused liver models are the avoidance of live animal experiments, the ability to control accurately physiological parameters and to analyze morphological changes following hepatic ablation. In addition detailed study of the “heat sink” effect and changes consequent upon bile duct perfusion can be assessed. Other organs and tissues from the same animals from which the livers are procured can also be used in other research leading to an overall reduction in the number of animals required. Validation of the role of isolated perfused liver models for experimental studies of hepatic ablation is essential to establish their position in this field of research.

REFERENCES

- 1 **Wong SL**, Mangu PB, Choti MA, Crocenzi TS, Dodd GD, Dorfman GS, Eng C, Fong Y, Giusti AF, Lu D, Marsland TA, Michelson R, Poston GJ, Schrag D, Seidenfeld J, Benson AB. American Society of Clinical Oncology 2009 clinical evidence review on radiofrequency ablation of hepatic metastases from colorectal cancer. *J Clin Oncol* 2010; **28**: 493-508 [PMID: 19841322 DOI: 10.1200/JCO.2009.23.4450]
- 2 **Cho YK**, Kim JK, Kim MY, Rhim H, Han JK. Systematic review of randomized trials for hepatocellular carcinoma treated with percutaneous ablation therapies. *Hepatology* 2009; **49**: 453-459 [PMID: 19065676 DOI: 10.1002/hep.22648]
- 3 **Bhardwaj N**, Strickland AD, Ahmad F, Dennison AR, Lloyd DM. Liver ablation techniques: a review. *Surg Endosc* 2010; **24**: 254-265 [PMID: 19554370 DOI: 10.1007/s00464-009-0590-4]
- 4 **Gannon CJ**, Curley SA. The role of focal liver ablation in the treatment of unresectable primary and secondary malignant liver tumors. *Semin Radiat Oncol* 2005; **15**: 265-272 [PMID: 16183480 DOI: 10.1016/j.semradonc.2005.04.004]
- 5 **Hines-Peralta AU**, Pirani N, Clegg P, Cronin N, Ryan TP, Liu Z, Goldberg SN. Microwave ablation: results with a 2.45-GHz applicator in ex vivo bovine and in vivo porcine liver. *Radiology* 2006; **239**: 94-102 [PMID: 16484351 DOI: 10.1148/radiol.2383050262]
- 6 **Awad MM**, Devgan L, Kamel IR, Torbensen M, Choti MA. Microwave ablation in a hepatic porcine model: correlation of CT and histopathologic findings. *HPB (Oxford)* 2007; **9**: 357-362 [PMID: 18345319 DOI: 10.1080/13651820701646222]
- 7 **Simon CJ**, Dupuy DE, Iannitti DA, Lu DS, Yu NC, Aswad BI, Busuttil RW, Lassman C. Intraoperative triple antenna hepatic microwave ablation. *AJR Am J Roentgenol* 2006; **187**: W333-W340 [PMID: 16985103 DOI: 10.2214/AJR.05.0804]
- 8 **Goldberg SN**, Solbiati L, Hahn PF, Cosman E, Conrad JE, Fogle R, Gazelle GS. Large-volume tissue ablation with radio frequency by using a clustered, internally cooled electrode technique: laboratory and clinical experience in liver metastases. *Radiology* 1998; **209**: 371-379 [PMID: 9807561]
- 9 **Mulier S**, Miao Y, Mulier P, Dupas B, Pereira P, de Baere T, Lencioni R, Leveillee R, Marchal G, Michel L, Ni Y. Electrodes and multiple electrode systems for radiofrequency ablation: a proposal for updated terminology. *Eur Radiol* 2005; **15**: 798-808 [PMID: 15711846 DOI: 10.1007/s00330-004-2584-x]
- 10 **Mulier S**, Ni Y, Frich L, Burdior F, Denys AL, De Wispelaere JF, Dupas B, Habib N, Hoey M, Jansen MC, Lacrosse M, Leveillee R, Miao Y, Mulier P, Mutter D, Ng KK, Santambrogio R, Stippel D, Tamaki K, van Gulik TM, Marchal G, Michel L. Experimental and clinical radiofrequency ablation: proposal for standardized description of coagulation size and geometry. *Ann Surg Oncol* 2007; **14**: 1381-1396 [PMID: 17242989 DOI: 10.1245/s10434-006-9033-9]
- 11 **Yu NC**, Lu DS, Raman SS, Dupuy DE, Simon CJ, Lassman C, Aswad BI, Iannitti D, Busuttil RW. Hepatocellular carcinoma: microwave ablation with multiple straight and loop antenna clusters--pilot comparison with pathologic findings. *Radiology* 2006; **239**: 269-275 [PMID: 16493013 DOI: 10.1148/radiol.2383041592]
- 12 **Shen P**, Geisinger KR, Zagoria R, Levine EA. Pathologic correlation study of microwave coagulation therapy for hepatic malignancies using a three-ring probe. *J Gastrointest Surg* 2007; **11**: 603-611 [PMID: 17393259 DOI: 10.1007/s11605-006-0046-2]
- 13 **Wemyss-Holden SA**, Dennison AR, Finch GJ, Hall Pd Pde L, Maddern GJ. Electrolytic ablation as an adjunct to liver resection: experimental studies of predictability and safety. *Br J Surg* 2002; **89**: 579-585 [PMID: 11972547 DOI: 10.1046/j.1365-2168.2002.02064.x]
- 14 **Wemyss-Holden SA**, de la M Hall P, Robertson GS, Dennison AR, Vanderzon PS, Maddern GJ. The safety of electrolytically induced hepatic necrosis in a pig model. *Aust N Z J Surg* 2000; **70**: 607-612 [PMID: 10945557 DOI: 10.1046/j.1440-1622.2000.01907.x]
- 15 **Metcalfe MS**, Mullin EJ, Texler M, Berry DP, Dennison AR, Maddern GJ. The safety and efficacy of radiofrequency and electrolytic ablation created adjacent to large hepatic veins in a porcine model. *Eur J Surg Oncol* 2007; **33**: 662-667 [PMID: 17412548 DOI: 10.1016/j.ejso.2007.02.011]
- 16 **Wemyss-Holden SA**, Dennison AR, Berry DP, Maddern GJ. Local ablation for unresectable liver tumors: is thermal best? *J Hepatobiliary Pancreat Surg* 2004; **11**: 97-106 [PMID: 15127271 DOI: 10.1007/s00534-002-0715-9]
- 17 **Singer CJ**. Greek biology and Greek medicine. London: Oxford University Press, 1922: 6-14

- 18 **Wilson LG.** Erasistratus, Galen, and the pneuma. *Bull Hist Med* 1959; **33**: 293-314 [PMID: 13845095]
- 19 **Prendergast JS.** The Background of Galen's Life and Activities, and its Influence on his Achievements. *Proc R Soc Med* 1930; **23**: 1131-1148 [PMID: 19987627]
- 20 **Buchholz AC, Schoeller DA.** Is a calorie a calorie? *Am J Clin Nutr* 2004; **79**: 899S-906S [PMID: 15113737]
- 21 **Mock M, Fouet A.** Anthrax. *Annu Rev Microbiol* 2001; **55**: 647-671 [PMID: 11544370 DOI: 10.1146/annurev.micro.55.1.647]
- 22 **Gorden P.** Non-insulin dependent diabetes--the past, present and future. *Ann Acad Med Singapore* 1997; **26**: 326-330 [PMID: 9285027]
- 23 **Croce P.** Vivisection or science? An Investigation into Testing Drugs and Safeguarding Health. London and New York: Zed Books, 1999: 11-30
- 24 **Rudacille D.** The scalpel and the butterfly: the conflict. New York: Farrar, Straus and Giroux, 2000: 19-20
- 25 **Bowlby J.** Charles Darwin: a new life. London and New York: W.W.Norton, 1992: 420-421
- 26 **Hendee WR.** Use of animals in biomedical research: the challenge and response. AMA white paper. Chicago: American Medical Association, 1989
- 27 **Teratani T, Yoshida H, Shiina S, Obi S, Sato S, Tateishi R, Mine N, Kondo Y, Kawabe T, Omata M.** Radiofrequency ablation for hepatocellular carcinoma in so-called high-risk locations. *Hepatology* 2006; **43**: 1101-1108 [PMID: 16628706 DOI: 10.1002/hep.21164]
- 28 **Welp C, Siebers S, Ermert H, Werner J.** Investigation of the influence of blood flow rate on large vessel cooling in hepatic radiofrequency ablation. *Biomed Tech (Berl)* 2006; **51**: 337-346 [PMID: 17155870 DOI: 10.1515/BMT.2006.067]
- 29 **Thanos L, Mylona S, Galani P, Pomoni M, Pomoni A, Koskinas I.** Overcoming the heat-sink phenomenon: successful radiofrequency thermal ablation of liver tumors in contact with blood vessels. *Diagn Interv Radiol* 2008; **14**: 51-56 [PMID: 18306146]
- 30 **Vuppugalla R, Mehvar R.** Selective effects of nitric oxide on the disposition of chlorzoxazone and dextromethorphan in isolated perfused rat livers. *Drug Metab Dispos* 2006; **34**: 1160-1166 [PMID: 16621933 DOI: 10.1124/dmd.105.009050]
- 31 **Strubelt O, Kremer J, Tilse A, Keogh J, Pentz R, Younes M.** Comparative studies on the toxicity of mercury, cadmium, and copper toward the isolated perfused rat liver. *J Toxicol Environ Health* 1996; **47**: 267-283 [PMID: 8604150 DOI: 10.1080/009841096161780]
- 32 **Hung DY, Siebert GA, Chang P, Anissimov YG, Roberts MS.** Disposition kinetics of propranolol isomers in the perfused rat liver. *J Pharmacol Exp Ther* 2004; **311**: 822-829 [PMID: 15192084 DOI: 10.1124/jpet.104.070011]
- 33 **Brace CL, Laeseke PF, van der Weide DW, Lee FT.** Microwave Ablation With a Triaxial Antenna: Results in ex vivo Bovine Liver. *IEEE Trans Microw Theory Tech* 2005; **53**: 215-220 [PMID: 18079981 DOI: 10.1109/TMTT.2004.839308]
- 34 **Lee JM, Han JK, Kim HC, Choi YH, Kim SH, Choi JY, Choi BI.** Switching monopolar radiofrequency ablation technique using multiple, internally cooled electrodes and a multi-channel generator: ex vivo and in vivo pilot study. *Invest Radiol* 2007; **42**: 163-171 [PMID: 17287646]
- 35 **Navarro AC, Burdío F, Berjano EJ, Güemes A, Burdío JM, Sousa R, Lozano R, Tejero E, de Gregorio MA.** Small ablation zones created previous to saline infusion result in enlargement of the coagulated area during perfusion RF ablation: an ex vivo experimental study. *Physiol Meas* 2007; **28**: N29-N37 [PMID: 17664615 DOI: 10.1088/0967-3334/28/6/N02]
- 36 **O'Rourke AP, Lazebnik M, Bertram JM, Converse MC, Hagness SC, Webster JG, Mahvi DM.** Dielectric properties of human normal, malignant and cirrhotic liver tissue: in vivo and ex vivo measurements from 0.5 to 20 GHz using a precision open-ended coaxial probe. *Phys Med Biol* 2007; **52**: 4707-4719 [PMID: 17634659 DOI: 10.1088/0031-9155/52/15/022]
- 37 **Crocetti L, Lencioni R, Debeni S, See TC, Pina CD, Bartolozzi C.** Targeting liver lesions for radiofrequency ablation: an experimental feasibility study using a CT-US fusion imaging system. *Invest Radiol* 2008; **43**: 33-39 [PMID: 18097275]
- 38 **Solazzo SA, Ahmed M, Liu Z, Hines-Peralta AU, Goldberg SN.** High-power generator for radiofrequency ablation: larger electrodes and pulsing algorithms in bovine ex vivo and porcine in vivo settings. *Radiology* 2007; **242**: 743-750 [PMID: 17244719 DOI: 10.1148/radiol.2423052039]
- 39 **Wang Y, Sun Y, Feng L, Gao Y, Ni X, Liang P.** Internally cooled antenna for microwave ablation: results in ex vivo and in vivo porcine livers. *Eur J Radiol* 2008; **67**: 357-361 [PMID: 17768024 DOI: 10.1016/j.ejrad.2007.07.015]
- 40 **Imber CJ, St Peter SD, de Cnarruzabeitia IL, Lemonde H, Rees M, Butler A, Clayton PT, Friend PJ.** Optimisation of bile production during normothermic preservation of porcine livers. *Am J Transplant* 2002; **2**: 593-599 [PMID: 12201359 DOI: 10.1034/j.1600-6143.2002.20703.x]
- 41 **Imber CJ, St Peter SD, Lopez de Cnarruzabeitia I, Pigott D, James T, Taylor R, McGuire J, Hughes D, Butler A, Rees M, Friend PJ.** Advantages of normothermic perfusion over cold storage in liver preservation. *Transplantation* 2002; **73**: 701-709 [PMID: 11907414 DOI: 10.1097/00007890-200203150-00008]
- 42 **Butler AJ, Rees MA, Wight DG, Casey ND, Alexander G, White DJ, Friend PJ.** Successful extracorporeal porcine liver perfusion for 72 hr. *Transplantation* 2002; **73**: 1212-1218 [PMID: 11981411]
- 43 **Gravante G, Ong SL, Metcalfe MS, Sorge R, Fox AJ, Lloyd DM, Maddern GJ, Dennison AR.** Changes in acid-base balance during electrolytic ablation in an ex vivo perfused liver model. *Am J Surg* 2012; **204**: 666-670 [PMID: 20451173 DOI: 10.1016/j.amjsurg.2009.12.019]
- 44 **Gravante G, Ong SL, Metcalfe MS, Sorge R, Overton J, Lloyd DM, Maddern GJ, Dennison AR.** Cytokine response of electrolytic ablation in an ex vivo perfused liver model. *ANZ J Surg* 2010; **80**: 537-541 [PMID: 20795969 DOI: 10.1111/j.1445-2197.2010.05380.x]
- 45 **Gravante G, Ong SL, West K, McGregor A, Maddern GJ, Metcalfe MS, Lloyd DM, Dennison AR.** Patterns of histological changes following hepatic electrolytic ablation in an ex-vivo perfused model. *Pathol Oncol Res* 2012; **18**: 1085-1089 [PMID: 22706978]
- 46 **Gravante G, Ong SL, Metcalfe MS, Sorge R, Sconocchia G, Orlando G, Lloyd DM, Dennison AR.** Cytokine response to ischemia/reperfusion injury in an ex vivo perfused porcine liver model. *Transplant Proc* 2009; **41**: 1107-1112 [PMID: 19460492 DOI: 10.1016/j.transproceed.2009.02.054]
- 47 **Bernard C.** Sur le mécanisme de la formation du sucre dans le foie. *C R hebd Acad Sci* 1855; **41**: 461-469
- 48 **Hobbs KE, Hunt AC, Palmer DB, Badrick FE, Morris AM, Mitra SK, Peacock JH, Immelman EJ, Riddell AG.** Hypothermic perfusion as a method of short-term porcine liver storage. *Br J Surg* 1968; **55**: 862 [PMID: 5686999]
- 49 **Hobbs KE, Hunt AC, Palmer DB, Badrick FE, Morris AM, Mitra SK, Peacock JH, Immelman EJ, Riddell AG.** Hypothermic low flow liver perfusion as a means of porcine hepatic storage for six hours. *Br J Surg* 1968; **55**: 696-703 [PMID: 5676704 DOI: 10.1002/bjs.1800550913]
- 50 **Drapanas T, Zemel R, Vang JO.** Hemodynamics of the isolated perfused pig liver: metabolism according to routes of perfusion and rates of flow. *Ann Surg* 1966; **164**: 522-537 [PMID: 5927633]
- 51 **Abouna GM, Ashcroft T, Hull C, Hodson A, Kirkley J,**

- Walder DN. The assessment of function of the isolated perfused porcine liver. *Br J Surg* 1969; **56**: 289-295 [PMID: 4952478 DOI: 10.1002/bjs.1800560413]
- 52 **Abouna GM**, Fisher LM, Still WJ, Hume DM. Acute hepatic coma successfully treated by extracorporeal baboon liver perfusions. *Br Med J* 1972; **1**: 23-25 [PMID: 5061783 DOI: 10.1136/bmj.1.5791.23]
- 53 **Powell GM**, Hughes HM, Curtis CG. Isolated perfused liver technology for studying metabolic and toxicological problems. *Drug Metabol Drug Interact* 1989; **7**: 53-86 [PMID: 2699284 DOI: 10.1515/DMDI.1989.7.1.53]
- 54 **Hildebrand P**, Kleemann M, Roblick U, Mirow L, Bruch HP, Bürk C. Development of a perfused ex vivo tumor-mimic model for the training of laparoscopic radiofrequency ablation. *Surg Endosc* 2007; **21**: 1745-1749 [PMID: 17332954 DOI: 10.1007/s00464-007-9216-x]
- 55 **Lappegård KT**, Fung M, Bergseth G, Riesenfeld J, Mollnes TE. Artificial surface-induced cytokine synthesis: effect of heparin coating and complement inhibition. *Ann Thorac Surg* 2004; **78**: 38-44; discussion 44-45 [PMID: 15223398 DOI: 10.1016/j.athoracsur.2004.02.005]
- 56 **Chung WY**, Gravante G, Al-Leswas D, Alzarraa A, Sorge R, Ong SL, Pollard C, Lloyd DM, Metcalfe MS, Dennison AR. Addition of a kidney to the normothermic ex vivo perfused porcine liver model does not increase cytokine response. *J Artif Organs* 2012; **15**: 290-294 [PMID: 22476783]
- 57 **Chung WY**, Gravante G, Al-Leswas D, Alzarraa A, Sorge R, Ong SL, Pollard C, Lloyd DM, Metcalfe MS, Dennison AR. The autologous normothermic ex vivo perfused porcine liver-kidney model: improving the circuit's biochemical and acid-base environment. *Am J Surg* 2012; **204**: 518-526 [PMID: 23010618]

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Bowel preparation prior to colonoscopy: A continual search for excellence

Matthew L Bechtold, Abhishek Choudhary

Matthew L Bechtold, Abhishek Choudhary, Division of Gastroenterology, University of Missouri, Columbia, MO 65201, United States

Matthew L Bechtold, Division of Gastroenterology and Hepatology, University of Missouri Health Sciences Center, Five Hospital Drive, Columbia, MO 65212, United States

Author contributions: Bechtold ML wrote the manuscript; Choudhary A discussed the subject and provided critical revisions to the manuscript; both authors approved the final version of the manuscript.

Correspondence to: Matthew L Bechtold, MD, FACP, Division of Gastroenterology and Hepatology, University of Missouri Health Sciences Center, Five Hospital Drive, Columbia, MO 65212, United States. bechtoldm@health.missouri.edu

Telephone: +1-57-38821013 Fax: +1-57-38844595

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INVITED COMMENTARY ON HOT ARTICLES

As clinical gastroenterologists, we read the recent article by Tajika *et al*^[1] describing a new lower volume bowel preparation prior to colonoscopy with considerable interest. This article examined the use of mosapride and only 1.5 L of polyethylene glycol (PEG) for bowel preparation prior to colonoscopy. Given the continued need and search for more options for bowel preparation, we recommend reading and possibly implementing the information in this article.

Colorectal cancer is a common and devastating disease which affects many patients worldwide. Being in top three of cancers and cancer-related deaths in the United States, colorectal cancer has become a focus of gastroenterology, with much of our practice being significantly impacted by colorectal cancer screening and surveillance^[2].

Over the past 10 years, colonoscopy has become the screening test of choice for colorectal cancer. Unlike other non-invasive colorectal cancer screening tools, such as fecal occult blood testing and computed tomographic (CT) colonography, colonoscopy is the only modality which encompasses both diagnostic and therapeutic potential. However, colonoscopy is dependent on adequacy of bowel preparation for complete visualization of the

Abstract

Bowel preparation prior to colonoscopy is essential to maximize the benefits of colonoscopy. Numerous bowel preparations have been studied, ranging from 4 L polyethylene glycol (PEG) to split-dose regimens to 2 L PEG with an adjunct laxative (senna, bisacodyl, ascorbic acid). Due to the large volume of PEG required for adequate bowel preparation, many studies have focused on reducing this large volume to only 2 L PEG with the addition of an adjunct. Recently, a randomized controlled trial by Tajika *et al* showed that the addition of mosapride to only 1.5 L PEG was non-inferior to mosapride and 2 L PEG for bowel cleansing but did provide improvements in patient tolerance. This study offers yet another potential bowel preparation for patients undergoing colonoscopy and may trigger further studies with 1.5 L PEG with an adjunct. In this letter, we discuss the current state of bowel preparation prior to colonoscopy and offer information to guide clinicians on choosing the appropriate bowel preparation for their patients.

Table 1 Common bowel preparations utilized prior to colonoscopy

First tier			
Split-dose PEG			
2 L the night prior and 2 L morning of colonoscopy			
Second tier			
Full-dose PEG	2 L PEG + Adjunct	Sodium phosphate	Sulfate prep
4 L the night prior	Ascorbic acid Bisacodyl Senna Magnesium Mosapride	(Tablet)	(Na, K, Mg)
Third tier			
Miralax + Gatorade	Magnesium citrate	Enemas	1.5 L + Adjunct Mosapride

PEG: Polyethylene glycol.

colonic mucosa. If visualization is compromised, missed lesions, prolonged procedure time, and increased patient discomfort become increasingly possible, significantly impacting patients and healthcare costs^[3,4]. Therefore, bowel preparation prior to colonoscopy is extremely important for an adequate colonoscopic examination.

The optimal bowel preparation prior to colonoscopy must adequately clear the fecal material to visualize the underlying mucosa but also must be tolerable to the patient. Even if the bowel preparation is exceptional at clearing the colon of feces, if it is not palatable, the patients will likely not complete the full preparation. Over the past few years, many studies have been performed with varying uses of PEG, such as split-dose PEG and PEG with adjuncts, in an effort to improve patient tolerability without decreasing the efficacy of the bowel preparation. A meta-analysis in *Gastrointestinal Endoscopy* in 2011 demonstrated that split-dose PEG (consisting of 2 L PEG the night prior and 2 L PEG the morning of the procedure), improved both the cleansing of the colon and patient tolerability as compared to full-dose PEG (4 L the night prior to colonoscopy)^[5]. Subsequently, the split-dose PEG has become a very common bowel preparation and is the preferred bowel preparation of the American College of Gastroenterology. However, patients are still required to consume a large volume of PEG. Therefore, many studies have elected to reduce the volume of PEG to 2 L and add a laxative adjunct (ascorbic acid, bisacodyl, senna, magnesium, *etc.*) and monitor for bowel cleansing and patient tolerability^[6-12]. However, these studies resulted in conflicting results.

In 2011, two meta-analyses were published as abstracts in the *American Journal of Gastroenterology* showing that 2 L PEG with ascorbic acid cleansed the colon as well as 4L PEG but was equally tolerated^[13]; however, 2 L PEG with bisacodyl demonstrated equal bowel cleansing and improved patient tolerability^[14]. Despite the results, limitations have been placed on using bisacodyl with PEG due to risk of ischemic colitis. Therefore, the search continues for an adequate low-dose PEG bowel preparation.

In the randomized controlled trial by Tajika *et al.*^[11] mosapride (15 mg) was utilized as an adjunct laxative to low-dose PEG (1.5 L *vs* 2 L). The study revealed that mosapride with 1.5 L PEG resulted in equal bowel cleansing and increased tolerability as compared to mosapride with 2 L PEG. Given these results, mosapride with 1.5 L PEG appears promising as an alternative bowel preparation prior to colonoscopy. However, given a few limitations within this study, the results need to be interpreted cautiously.

First, this study utilized a non-inferiority model for analysis. Most bowel preparation studies are performed using a superiority model due to limitations of the non-inferiority model. The non-inferiority model is dependent upon the margin at which non-inferiority is defined. If the margin is not accurate, the results may be significantly affected. In this study, mosapride with 1.5 L PEG was non-inferior to mosapride with 2 L PEG; however, it appears that 2 L PEG with mosapride group has many more excellent and good preparations as compared to 1.5 L PEG with mosapride group (right colon: 88 *vs* 60; left colon: 97 *vs* 82). Given this difference, especially in the right colon, it is difficult for us to believe that equality of cleansing exists for these two regimens. Therefore, this leads us to believe that the sample size estimation and the non-inferiority margin may not be as accurate as hoped. Second, although the Aronchick scale has been shown to be a valuable tool in assessing bowel preparation, it is designed for the entire colon^[15]. This study used the Aronchick scale but divided it among right and left colon. Ideally, the Ottawa or Boston bowel preparation scales which utilize different segments of the colon with numerical scores may have been a better choice for bowel preparation assessment^[16,17].

At this point, based upon the available literature, we believe the ideal bowel preparation prior to colonoscopy is the split-dose PEG given 2 L the night prior and 2 L the day of the colonoscopy (Table 1). However, given its large volume, it may not be suited for all patients. Furthermore, given the number of other potential options including the preparation described in this study, alternative bowel preparations are readily available and may be utilized in certain cases without adversely affecting bowel cleansing.

The choice of bowel preparation is extremely important for patients and practices. Adequate bowel preparation which is tolerable to patients will likely enhance satisfaction among patients, who would be more likely to return for another colonoscopy. It will also likely reduce the need for early repeat procedures due to poor prep quality and overall costs. Given the number of preparations available, bowel preparation prior to colonoscopy in the future may be tailored to the needs and desires of the patients. Although split-dose PEG seems to be the gold-standard at this time, other preparations may be utilized to enhance patient satisfaction without adversely affecting overall preparation quality. The choice is up to you and your patient.

REFERENCES

- 1 **Tajika M**, Niwa Y, Bhatia V, Kawai H, Kondo S, Sawaki A, Mizuno N, Hara K, Hijioka S, Matsumoto K, Kobayashi Y, Saeki A, Akabane A, Komori K, Yamao K. Efficacy of mosapride citrate with polyethylene glycol solution for colonoscopy preparation. *World J Gastroenterol* 2012; **18**: 2517-2525 [PMID: 22654449 DOI: 10.3748/wjg.v18.i20.2517]
- 2 **Siegel R**, Naishadham D, Jemal A. Cancer statistics, 2012. *CA Cancer J Clin* 2012; **62**: 10-29 [PMID: 22237781 DOI: 10.3322/caac.21149]
- 3 **Kim WH**, Cho YJ, Park JY, Min PK, Kang JK, Park IS. Factors affecting insertion time and patient discomfort during colonoscopy. *Gastrointest Endosc* 2000; **52**: 600-605 [PMID: 11060182 DOI: 10.1067/mge.2000.109802]
- 4 **Rex DK**, Imperiale TF, Latinovich DR, Bratcher LL. Impact of bowel preparation on efficiency and cost of colonoscopy. *Am J Gastroenterol* 2002; **97**: 1696-1700 [PMID: 12135020 DOI: 10.1111/j.1572-0241.2002.05827.x]
- 5 **Kilgore TW**, Abdinoor AA, Szary NM, Schowengerdt SW, Yust JB, Choudhary A, Matteson ML, Puli SR, Marshall JB, Bechtold ML. Bowel preparation with split-dose polyethylene glycol before colonoscopy: a meta-analysis of randomized controlled trials. *Gastrointest Endosc* 2011; **73**: 1240-1245 [PMID: 21628016 DOI: 10.1016/j.gie.2011.02.007]
- 6 **Marmo R**, Rotondano G, Riccio G, Marone A, Bianco MA, Stroppa I, Caruso A, Pandolfo N, Sansone S, Gregorio E, D'Alvano G, Procaccio N, Capo P, Marmo C, Cipolletta L. Effective bowel cleansing before colonoscopy: a randomized study of split-dosage versus non-split dosage regimens of high-volume versus low-volume polyethylene glycol solutions. *Gastrointest Endosc* 2010; **72**: 313-320 [PMID: 20561621 DOI: 10.1016/j.gie.2010.02.048]
- 7 **Sharma VK**, Chockalingham SK, Ugheoke EA, Kapur A, Ling PH, Vasudeva R, Howden CW. Prospective, randomized, controlled comparison of the use of polyethylene glycol electrolyte lavage solution in four-liter versus two-liter volumes and pretreatment with either magnesium citrate or bisacodyl for colonoscopy preparation. *Gastrointest Endosc* 1998; **47**: 167-171 [PMID: 9512283 DOI: 10.1016/S0016-5107(98)70351-7]
- 8 **Hookey LC**, Depew WT, Vanner SJ. Combined low volume polyethylene glycol solution plus stimulant laxatives versus standard volume polyethylene glycol solution: a prospective, randomized study of colon cleansing before colonoscopy. *Can J Gastroenterol* 2006; **20**: 101-105 [PMID: 16482236]
- 9 **DiPalma JA**, Wolff BG, Meagher A, Cleveland Mv. Comparison of reduced volume versus four liters sulfate-free electrolyte lavage solutions for colonoscopy colon cleansing. *Am J Gastroenterol* 2003; **98**: 2187-2191 [PMID: 14572566 DOI: 10.1111/j.1572-0241.2003.07690.x]
- 10 **Eli C**, Fischbach W, Bronisch HJ, Dertinger S, Layer P, Rünzi M, Schneider T, Kachel G, Gröger J, Köllinger M, Nagell W, Goerg KJ, Wanitschke R, Gruss HJ. Randomized trial of low-volume PEG solution versus standard PEG + electrolytes for bowel cleansing before colonoscopy. *Am J Gastroenterol* 2008; **103**: 883-893 [PMID: 18190651 DOI: 10.1111/j.1572-0241.2007.01708.x]
- 11 **Adams WJ**, Meagher AP, Lubowski DZ, King DW. Bisacodyl reduces the volume of polyethylene glycol solution required for bowel preparation. *Dis Colon Rectum* 1994; **37**: 229-233; discussion 233-234 [PMID: 8137669 DOI: 10.1007/BF02048160]
- 12 **Ker TS**. Comparison of reduced volume versus four-liter electrolyte lavage solutions for colon cleansing. *Am Surg* 2006; **72**: 909-911 [PMID: 17058733]
- 13 **Godfrey JD**, Clark RE, Choudhary A, Ashraf I, Matteson ML, Bechtold ML. Low-volume polyethylene glycol and ascorbic acid for bowel preparation prior to colonoscopy: A meta-analysis [abstract]. *Am J Gastroenterol* 2011; **106**: S527 (1376)
- 14 **Clark RE**, Godfrey JD, Choudhary AMD, Ashraf I, Matteson ML, Bechtold ML. Low-volume polyethylene glycol and bisacodyl for bowel preparation prior to colonoscopy: A meta-analysis [abstract]. *Am J Gastroenterol* 2011; **106**: S528 (1377)
- 15 **Aronchick CA**, Lipshutz WH, Wright SH, DuFrayne F, Bergman G. Validation of an instrument to assess colon cleansing [abstract]. *Am J Gastroenterol* 1999; **94**: 2667
- 16 **Rostom A**, Jolicoeur E. Validation of a new scale for the assessment of bowel preparation quality. *Gastrointest Endosc* 2004; **59**: 482-486 [PMID: 15044882 DOI: 10.1016/S0016-5107(03)02875-X]
- 17 **Lai EJ**, Calderwood AH, Doros G, Fix OK, Jacobson BC. The Boston bowel preparation scale: a valid and reliable instrument for colonoscopy-oriented research. *Gastrointest Endosc* 2009; **69**: 620-625 [PMID: 19136102 DOI: 10.1016/j.gie.2008.05.057]

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Internal frontier: The pathophysiology of the small intestine

Haruhiko Sugimura, Satoshi Osawa

Haruhiko Sugimura, Department of Tumor Pathology, Hamamatsu University School of Medicine, 1-20-1 Handayama, Higashi-ku, Hamamatsu 431-3192, Japan

Satoshi Osawa, Endoscopic and Photodynamic Medicine, Hamamatsu University School of Medicine, 1-20-1 Handayama, Higashi-ku, Hamamatsu 431-3192, Japan

Author contributions: Sugimura H wrote the manuscript; Osawa S provided the materials and contributed to discussion about the manuscript.

Correspondence to: Haruhiko Sugimura, MD, PhD, Department of Tumor Pathology, Hamamatsu University School of Medicine, 1-20-1 Handayama, Higashi-ku, Hamamatsu 431-3192, Japan. hsugimur@hama-med.ac.jp

Telephone: +81-53-4352220 Fax: +81-53-4352225

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INVITED COMMENTARY ON HOT ARTICLES

Juan Rosai's Surgical Pathology (9th edition) contains 45 pages on the small intestine, compared with 80 pages on the large intestine^[1]. The actual length of the small intestine is much longer than that of the large intestine, and the vital importance of the small intestine is well known. Why then are fewer pages devoted to diseases of the small intestine? Obviously, the access of surgical pathologists and gastroenterologists to the small intestine is limited, compared with access to the large intestine and appendix. Actually, the old version of Morson and Dawson's textbook Gastrointestinal Pathology^[2] is fairer: the same number of pages is devoted to the small intestine and to the large intestine, probably because this book is a product of the century during which autopsies made major contributions to our knowledge base. This issue of the *World Journal of Gastroenterology* has a wide-ranging review article on the pathophysiology of the small intestine by Professor Basson and his colleagues^[3]. These authors are surgeons, and a tremendous amount of data based on their own clinical experiences and those of others, as well as on animal experiments, are comprehensively discussed in their review. This review article addresses conditions such as starvation and parenteral nutrition in patients with severe trauma or pancreatitis and in patients with postoperative short-gut syndrome for various reasons including bariatric surgery for obesity. Although these

Abstract

Even though the small intestine occupies a major portion of the abdominal space and is essential for life, in most pathology textbooks any chapter on small intestinal diseases, especially in human beings, is typically shorter than those for other gastrointestinal organs. Clinical and experimental investigations of the small intestine in various clinical situations, such as nutrition management, obesity interventions, and emergency care, have elucidated several important biological problems associated with the small intestine, the last frontier of gastroenterology. In this issue, a review by Professor Basson and his team at Michigan State University sheds light on the changes in the human small intestine under various conditions based on their clinical and surgical experience. With the advent of recent innovations in enteroscopy, a form of endoscopy used to examine deep within the small intestine, the issue that they highlighted, i.e., mucosal adaptation and atrophy of the human small intestine, has emerged as a major and manageable challenge for gastroenterologists in general, including the readers of the *World Journal of Gastroenterology*.

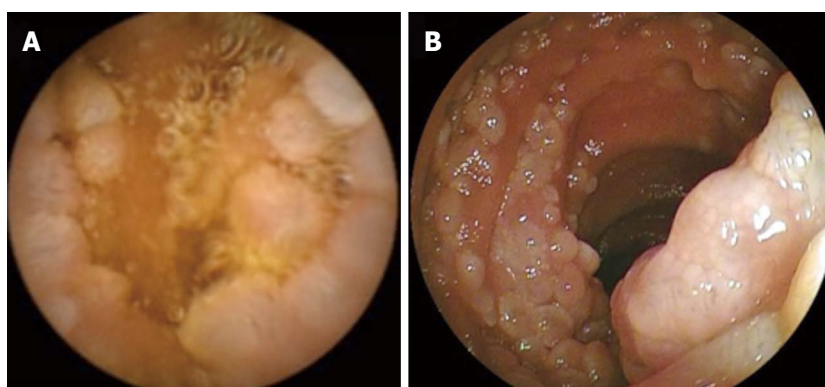


Figure 1 Endoscopy. Images obtained using capsule endoscopy (A) and double-balloon enteroscopy (B) in a 61-year-old man with follicular lymphoma who visited us because of gastrointestinal bleeding from an unknown cause and a hemoglobin concentration of 9.7 g/dL.

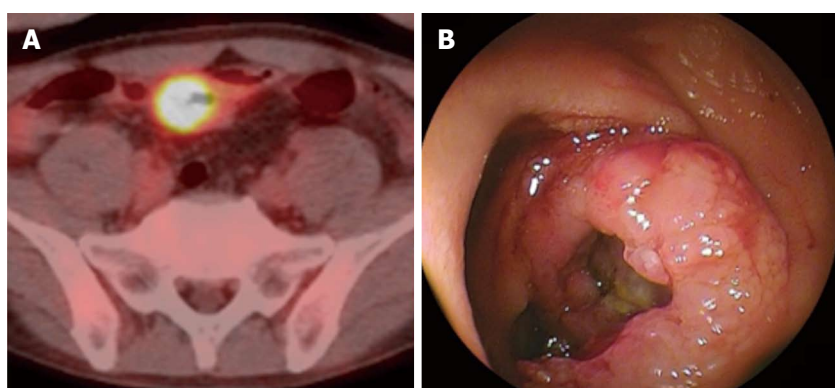


Figure 2 A mass positive by positron emission computed tomography actually was a tumor arising the mucosa of the small intestine of a 48-year-old woman who had suffered Crohn disease. A: Positron emission computed tomography/computed tomography showed an accumulation in site of wall thickening of ileum; B: Image obtained by double-balloon enteroscopy.

conditions are undoubtedly serious, recognition and explicit statements by medical professionals regarding the importance of the small intestine, and of its diseases as a “great burden of human distress”, have only recently appeared in medical literature^[4]. Most clinical conditions that highlight the importance of the small intestine are familiar only to experts in a particular area of medicine. These conditions are not typically a concern of non-surgical gastroenterologists, such as pathologists who are accustomed to diagnosing neoplasms of the large intestine, grading inflammatory bowel disease, and validating ischemic necrosis of the resected small intestine in cases with an acute abdomen. The small intestine is most often investigated because of its involvement in a disease originating from another organ, such as the mesothelium^[5]. Changes in the mucosa of the small intestine itself are an exceptional category among the slides that pile up next to the microscope.

The situation is now changing. As avid readers of *World Journal of Gastroenterology* may have noticed^[6-9], recent progress in enteroscopy has enabled areas deep within the small intestine to be reached, providing clinicians active in the wider branches of gastroenterology with the opportunity to evaluate changes in the small intestine, which have previously only been observed by surgeons. This

also means that many non-surgeons must commit to the management of the small intestine, based on findings and evaluations associated with specimens of small intestine obtained using these newly developed modalities. For ordinary pathologists, for example, the concepts mainly discussed in this review, i.e., adaptation and atrophy of the small intestine, may be new and unfamiliar. These lesions were encountered only after specific “congenital or acquired disease or medical and surgical intervention” had occurred in the patients. When specimens obtained by enteroscopy become routine in the future, however, the concepts and knowledge of adaptation and atrophy of the small intestine will become an essential “must” for all practitioners including general pathologists.

One of the important conditions that the authors of this review addressed is the change in the small intestine in subjects receiving total parental nutrition. This lifesaving modality has many variations in terms of its nutritional regimen and the effects of these variations on pathophysiology, that is, on the grade of adaptation of the small intestine. These effects on the small intestinal mucosa and the consequent outcomes of individual patients have been thoroughly investigated and published in many scientific articles, but the scientific evidence in human beings remains insufficient according to the

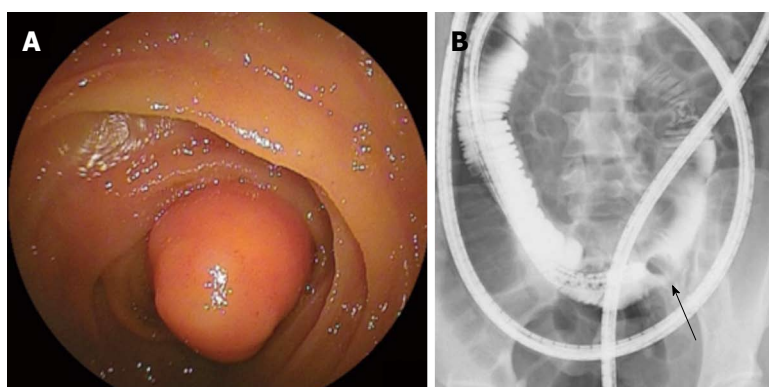


Figure 3 Identification of lipoma clarified the reason of intermittent abdominal discomfort in a 29-year-old female. Endoscopic finding (A) and selective small bowel series (B) obtained using double-balloon enteroscopy. This required surgical intervention. An arrow in the B indicates a defect by a lipoma.

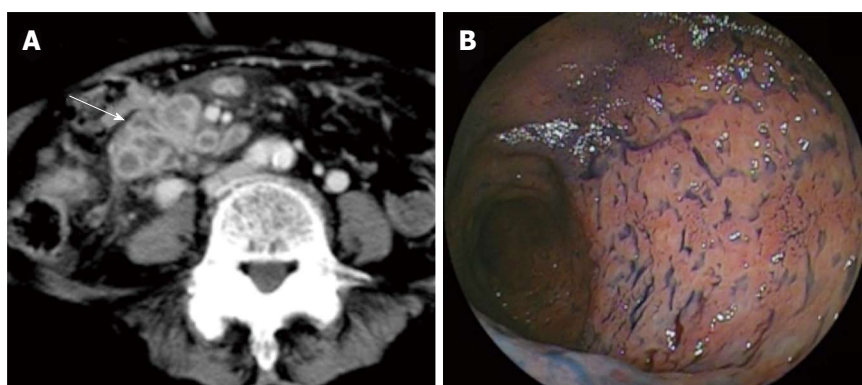


Figure 4 Intestinal tuberculosis is also revealed by double-balloon enteroscopy. A: Computed tomography showed wall thickening of the ileum with contrast enhancement (arrow); B: Double-balloon enteroscopy showed destruction of the small intestinal villi.

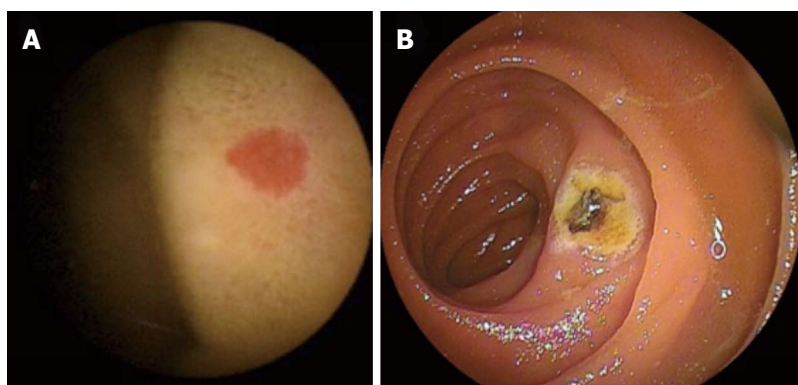


Figure 5 Angioectasia. A: Diagnosis of angioectasia was made by capsule endoscopy; B: Argon plasma coagulation successfully treated the lesion.

above-mentioned authors. The concept of mucosal adaptation^[10] includes proliferation, functional augmentation, and cellular differentiation. Biochemical changes include alterations in molecules related to apoptosis, proliferation, signal transduction, and fatty acid metabolism. These meticulous frameworks in intestinal cells and tissues have so far been revealed by studies using *in vivo* and *in vitro* manipulative systems. From now on, in the era of enteroscopy, lingering questions such as “what is the reality and examples of mucosal adaptation in human clinical settings?” and “how can we validate the accumulated ex-

perimental findings in human beings and exploit them in clinical practice?” will be answered.

Small intestinal enteroscopy is now available in ordinary hospitals, thus facilitating the detection of previously unobserved pathological conditions. Capsular endoscopy, the latest wireless version of enteroscopy, has become a popular practical procedure since the publication of a seminal report on this modality a decade ago^[11]. In the minds of laymen, this technique seems like a dream^[12]. The use of capsular endoscopy and refined enteroscopy using a double-balloon method^[13] in clinical practice have

revealed thousands of new anecdotal findings (Figure 1), and the rapid accumulation of this kind of basic knowledge of the small intestine will help to set up principles of surveillance for mucosal adaptation and atrophy of the small intestine (especially morphological changes) in various clinical conditions. For example, introduction of capsular endoscopy and a double-balloon method disclosed previously unrecognized lesions such as adenocarcinoma arising from Crohn disease in the small intestine (Figure 2)^[14], submucosal lipoma (Figure 3), tuberculosis at the terminal ileum (Figure 4), and angioectasia (Figure 5). None of these lesions were accessible until the recent development of capsule endoscopy and the double balloon method. The morphology is new to pathologists and endoscopists, and these developments will critically influence the managements of the patients. The concepts that Professor Basson and colleagues have illuminated in their review will soon become an important guidepost for evaluating the histopathology of the small intestine in daily practice and for patient care by a broader range of gastroenterologists.

REFERENCES

- 1 Rosai J. Rosai and Ackerman's Surgical Pathology. 9th ed. Edinburgh: Mosby, 2004: 615-872
- 2 Morson BC, Dawson IM, Day DW, Jass JR, Price AB, Williams GT. Morson & Dawson's gastrointestinal pathology. 3 ed. London: Blackwell Scientific Publications, 1989
- 3 Shaw D, Gohil K, Basson MD. Intestinal mucosal atrophy and adaptation. *World J Gastroenterol* 2012; **18**: 6357-6375 [PMID: 23197881 DOI: 10.3748/wjg.v18.i44.6357]
- 4 Tappenden KA. Emerging therapies for intestinal failure. *Arch Surg* 2010; **145**: 528-532 [PMID: 20566971 DOI: 10.1001/archsurg.2010.102]
- 5 Kim JH, Kwon KY, Jeon YK, Nam JH, Choi C, Hyeon CL, Choi YD. Mucin-positive epithelial mesothelioma of the peritoneum: small bowel involvement. *Pathol Int* 2011; **61**: 756-761 [PMID: 22126385 DOI: 10.1111/j.1440-1827.2011.02732.x]
- 6 Kav T, Sivri B. Is enteroscopy necessary for diagnosis of celiac disease? *World J Gastroenterol* 2012; **18**: 4095-4101 [PMID: 22919241 DOI: 10.3748/wjg.v18.i31.4095]
- 7 Oh TG, Chung JW, Kim HM, Han SJ, Lee JS, Park JY, Song SY. Primary intestinal lymphangiectasia diagnosed by capsule endoscopy and double balloon enteroscopy. *World J Gastrointest Endosc* 2011; **3**: 235-240 [PMID: 22110841 DOI: 10.4253/wjge.v3.i11.235]
- 8 Park SC, Chun HJ, Kang CD, Sul D. Prevention and management of non-steroidal anti-inflammatory drugs-induced small intestinal injury. *World J Gastroenterol* 2011; **17**: 4647-4653 [PMID: 22180706 DOI: 10.3748/wjg.v17.i42.4647]
- 9 Tee HP, How SH, Kaffes AJ. Learning curve for double-balloon enteroscopy: Findings from an analysis of 282 procedures. *World J Gastrointest Endosc* 2012; **4**: 368-372 [PMID: 22912911 DOI: 10.4253/wjge.v4.i8.368]
- 10 Drozdowski L, Thomson AB. Intestinal mucosal adaptation. *World J Gastroenterol* 2006; **12**: 4614-4627 [PMID: 16937429]
- 11 Iddan G, Meron G, Glukhovskiy A, Swain P. Wireless capsule endoscopy. *Nature* 2000; **405**: 417 [PMID: 10839527 DOI: 10.1038/35013140]
- 12 Muñoz-Navas M. Capsule endoscopy. *World J Gastroenterol* 2009; **15**: 1584-1586 [PMID: 19340899 DOI: 10.3748/wjg.15.1584]
- 13 Yamamoto H, Sekine Y, Sato Y, Higashizawa T, Miyata T, Iino S, Ido K, Sugano K. Total enteroscopy with a nonsurgical steerable double-balloon method. *Gastrointest Endosc* 2001; **53**: 216-220 [PMID: 11174299]
- 14 Kodaira C, Osawa S, Mochizuki C, Sato Y, Nishino M, Yamada T, Takayanagi Y, Takagaki K, Sugimoto K, Kanaoka S, Furuta T, Ikuma M. A case of small bowel adenocarcinoma in a patient with Crohn's disease detected by PET/CT and double-balloon enteroscopy. *World J Gastroenterol* 2009; **15**: 1774-1778 [PMID: 19360924 DOI: 10.3748/wjg.15.1774]

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Recurrent hepatitis C virus after transplant and the importance of plasma cells on biopsy

Eric R Kallwitz

Eric R Kallwitz, Section of Hepatology, Loyola University Medical Center, Maywood, IL 60153, United States

Author contributions: Kallwitz ER was responsible for the entire content, drafting and editing of the manuscript.

Correspondence to: Eric R Kallwitz, MD, Assistant Professor of Medicine, Section of Hepatology, Loyola University Medical Center, 2160 S First Avenue, Maywood, IL 60153, United States. ekallwitz@lumc.edu

Telephone: +1-708-2162538 Fax: +1-708-2166299

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INVITED COMMENTARY ON HOT ARTICLES

Hepatitis C virus (HCV) is the most common indication for liver transplantation in the United States. Recurrence of hepatitis C is nearly universal after transplantation and ensuing graft dysfunction occurs commonly. Ten percent of recipients progress to cirrhosis within 3 years of transplant^[1] demonstrating that some patients develop aggressive recurrent HCV. Plasma cell hepatitis, diagnosed histologically, is one of a number of conditions associated with adverse outcomes and graft failure in patients with posttransplant HCV. Plasma cell hepatitis can develop in the context of interferon based therapy or in the absence of treatment with interferon. Levitsky *et al*^[2] present a multicenter case-control study which included a large subset of patients found to have plasma cell hepatitis associated with interferon therapy for HCV. The manuscript should be read with interest by transplant hepatologists as it highlights important concepts regarding plasma cell hepatitis in patients with HCV after transplant. First, plasma cell hepatitis is under recognized. Second, the pathologic process resulting in plasma cell hepatitis is poorly understood. Finally, with a paucity of data, it is not possible to determine the best treatment for transplant recipients with this condition.

The case control series by Levitsky *et al*^[2] reported that the incidence of any immune mediated graft dysfunction on interferon based therapy varied by center, ranging between 3.2%-16.3%. Persons found to have immune mediated graft dysfunction on HCV therapy had significantly worse survival, and more graft failure leading to higher rates of retransplantation. Plasma cell hepatitis was the most common manifestation of interferon induced immune mediated graft dysfunction. It is important to note that plasma cell hepatitis was also commonly identified prior to the use of interferon. The incidence of plasma cell hepatitis

Abstract

Hepatitis C virus (HCV) is the leading indication for liver transplantation in the United States. It recurs universally after transplant but the rate of fibrosis and the development of graft failure is variable. Different donor and recipient features have been demonstrated to impact fibrosis. Plasma cell hepatitis, a histologic finding, is one feature associated with poor graft and patient outcomes. The pathogenic mechanism resulting in plasma cell hepatitis is poorly understood, with evidence suggesting a role for both the HCV and the immune system. A recent publication described plasma cell hepatitis in a larger context of immune mediated graft dysfunction in transplant recipients receiving interferon based therapy. This manuscript will highlight the topic of plasma cell hepatitis and provide commentary on the lack of recognition, the data regarding pathophysiologic mechanisms and the potential management options.

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Key words: Hepatitis C virus; Plasma cells; Biopsy; Sustained virological response

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Table 1 Generalized histologic and clinical features seen in post transplant patients with hepatitis C

Pathologic entity	Histologic features	Clinical and laboratory features
Plasma cell hepatitis	Plasma cells (often in sheets) Centrilobular necrosis	HCV PCR positive ANA or ASMA may be positive (often with low titers) Low level of immunosuppression Can be caused by interferon based therapy
<i>De novo</i> autoimmune hepatitis	Lymphoplasmacytic infiltrate Interface hepatitis	Positive ANA or ASMA Elevated immunoglobulins In persons on treatment HCV RNA often not detected (occurs in other settings in addition to HCV) Low level of immunosuppression Can be caused by interferon based therapy
Acute cellular rejection	Mixed inflammatory infiltrate Endotheliitis Nonsuppurative Cholangitis Centrilobular necrosis (variable)	Low level of immunosuppression Can be caused by interferon based therapy
Recurrent hepatitis C	Lymphocyte Aggregates Portal based fibrosis FCH is one variant (Cholestasis, Apoptosis, Fibrosis)	HCV PCR positive In FCH, markedly high viral load In FCH, high level of immunosuppression

HCV: Hepatitis C virus; FCH: Fibrosingcholestatic hepatitis; ANA: Antinuclear antibodies; ASMA: Anti-smooth muscle antibody; PCR: Polymerase chain reaction.

on pretreatment biopsies was much more common in persons who developed immune mediated graft dysfunction (36.5%) compared to the control group (7.7%, $P = 0.003$) on treatment^[2]. Fourteen of the cases labeled as plasma cell hepatitis by the central pathologist were not recognized by the local pathologist who initially interpreted the liver biopsy^[2]. The authors conclude that plasma cell hepatitis often predicts the development of immune mediated graft dysfunction occurring during interferon based treatment. In addition, the author's recommend that clinicians should not reduce immunosuppression doses and should not initiate interferon based therapy in those with immune features including plasma cell hepatitis on pretreatment biopsies.

The informative and interesting conclusions of this article deserve further comment. A main feature of the article from Levitsky and colleagues is that plasma cell hepatitis is under recognized and is often mistaken for recurrent hepatitis C or other forms of rejection^[2]. A histologic scoring system was developed in 2008^[3]. Diagnostic features of plasma cell hepatitis include numerous plasma cells, often in sheets or clusters, accompanied by centrilobular necrosis. Despite the existence of standardized criteria, it is not surprising that plasma cell hepatitis is under recognized. With few publications describing plasma cell hepatitis, it is not topical to hepatologists and pathologists. Additionally, in post-transplant patients, other processes such as recurrent HCV, *de novo* autoimmune hepatitis and acute cellular rejection present alternative diagnoses. Table 1 highlights clinical and histologic features which might help distinguish the different diagnoses. It is important to realize significant overlap does exist between the pathologic process making a definite diagnosis impossible in some cases. The recent publication in a high profile journal will hopefully lead to better recognition of this disorder.

The pathogenesis of plasma cell hepatitis has yet to be defined. It has been described both as a manifestation of hepatitis C^[4,5] and as a form of rejection^[3]. Evidence exists for both possibilities. Prior series have shown that HCV

plays a role. A two subject case series examined plasma cell hepatitis as a lymphoproliferative disorder^[5]. Both patients in this series had serum or urine protein electrophoresis demonstrating a monoclonal protein and RNA probes for hepatitis C were positive within the plasma cell infiltrate^[5]. An association between plasma cell hepatitis and mixed cryoglobulinemia has not been studied. In the current article, patient survival with plasma cell hepatitis was improved with a sustained virologic response to treatment for HCV^[2]. Although graft failure and retransplantation occurred in some cases, five year survival was above 80% and similar to control subjects^[2]. Additionally, there was a trend toward improved graft survival in cases of immune mediated graft dysfunction when hepatitis C was eradicated. In Kaplan-Meier analysis, the majority of graft loss occurred early after transplant with approximately 60% graft survival and no further graft loss occurring after two years in the group that achieved a sustained virological response (SVR)^[2]. In contrast, the group that did not achieve an SVR continued to develop graft loss during the entire period of follow up and graft survival was less than 40% at five years^[2]. It is additionally notable that SVR rates in series of patients with plasma cell hepatitis ranged between 40%-67%^[2,3]. Given that poor outcomes are observed with plasma cell hepatitis in persons who never received interferon^[3,6], prospective, randomized data are needed to compare outcome of interferon treatment versus no interferon treatment with respect to HCV eradication.

Other studies suggest the development of plasma cell hepatitis is an immune mediated event. Explanted livers of post transplant patients who later develop plasma cell hepatitis were more likely to have extensive plasma cell infiltrates^[6] suggesting that immunologic predisposition exists even prior to transplant. However, not all persons with plasma cell infiltrates on explant will develop plasma cell hepatitis, and additional factors after transplant appear to play a role. There are data describing the development of plasma cell hepatitis in the setting of lowered immunosup-

pression^[2,3]. In a series including 38 subjects with plasma cell hepatitis, 31 had either recently lowered immunosuppressant dosing or subtherapeutic drug levels^[3]. In the series by Levitsky *et al*^[2] significantly more patients with immune mediated graft dysfunction had a reduction in immunosuppression prior to interferon based therapy. Additionally, more subjects with immune mediated graft dysfunction had immunosuppression reduced during therapy^[2]. One would expect interferon would have a role in the development of an immune mediated event and the contribution of interferon to the development of plasma cell hepatitis is not quite clear. In a retrospective series, interferon was not associated with the development of plasma cell hepatitis and its use did not impact outcome once plasma cell hepatitis developed^[3]. In the series published by Levitsky *et al*^[2] persons with existing plasma cell hepatitis had worsened immune mediated graft dysfunction after their immunosuppression was reduced and interferon was started. Increasing baseline immunosuppression prior to initiating interferon in patients with plasma cell hepatitis should be considered for future study, especially given data showing improved outcomes with augmenting immunosuppression alone^[3].

With an immune predisposition, plasma cell hepatitis and *de novo* autoimmune hepatitis have overlapping features. They are nearly histologically indistinguishable, and some refer to them interchangeably^[7]. A few subtle clues suggest that these processes have a different underlying pathophysiology. Case series have described *de novo* autoimmune hepatitis developing in conjunction with interferon based therapy with elevated autoimmune titers, undetected HCV RNA levels and pretreatment biopsies showing no plasma cells^[8]. In addition, in transplant recipients for indications other than HCV, *de novo* autoimmune hepatitis has been shown to respond well to steroid therapy^[9], whereas plasma cell hepatitis in HCV infected recipients typically does not^[3].

Plasma cell hepatitis represents an important entity which is likely under reported as the result of poor recognition. Agreement on standardized nomenclature distinguishing plasma cell hepatitis from *de novo* autoimmune hepatitis in the post transplant setting may improve recognition. Plasma cell hepatitis best refers to plasma cell infiltration in the setting of post transplant hepatitis C. *De novo* autoimmune hepatitis best refers to plasma cell infiltration that occurs commonly with positive autoimmune titers, steroid responsiveness and, in the setting of interferon based therapy, may be best reserved when in a lymphoplasmacytic infiltrate develops without active viremia. As shown in Table 1, it must be recognized that overlap between the two conditions in both pathology and pathophysiologic mechanisms exist such that diagnostic certainty will not always occur.

Currently, the best management of plasma cell hepatitis that develops independent of HCV therapy is unclear. Limited data showed that augmentation of immunosuppression

without the addition of prednisone may be of benefit^[3]. Once on interferon based therapy, achieving an SVR was also shown to benefit patient survival^[2]. The recommendation by Levitsky *et al*^[2] that interferon should not be initiated in patients with plasma cell hepatitis may be overreaching based on the data presented. It also would suggest an alternate option with better outcomes existed. A practical approach may be augmenting baseline immunosuppression and a repeat liver biopsy. If the liver biopsy shows decreased immune features than interferon based therapy might be attempted. Ultimately, until there is better prospective data, responses to this entity will likely be reflective of single center experiences.

REFERENCES

- 1 **Berenguer M.** Natural history of recurrent hepatitis C. *Liver Transpl* 2002; **8**: S14-S18 [PMID: 12362293 DOI: 10.1053/jlts.2002.35781]
- 2 **Levitsky J, Fiel MI, Norvell JP, Wang E, Watt KD, Curry MP, Tewani S, McCashland TM, Hoteit MA, Shaked A, Saab S, Chi AC, Tien A, Schiano TD.** Risk for immune-mediated graft dysfunction in liver transplant recipients with recurrent HCV infection treated with pegylated interferon. *Gastroenterology* 2012; **142**: 1132-1139.e1 [PMID: 22285805 DOI: 10.1053/j.gastro.2012.01.030]
- 3 **Fiel MI, Agarwal K, Stanca C, Elhajj N, Kontorinis N, Thung SN, Schiano TD.** Posttransplant plasma cell hepatitis (de novo autoimmune hepatitis) is a variant of rejection and may lead to a negative outcome in patients with hepatitis C virus. *Liver Transpl* 2008; **14**: 861-871 [PMID: 18508382 DOI: 10.1002/lt.21447]
- 4 **Khettry U, Huang WY, Simpson MA, Pomfret EA, Pomposelli JJ, Lewis WD, Jenkins RL, Gordon FD.** Patterns of recurrent hepatitis C after liver transplantation in a recent cohort of patients. *Hum Pathol* 2007; **38**: 443-452 [PMID: 17188331 DOI: 10.1016/j.humpath.2006.08.028]
- 5 **Tun HW, Krishna M, Menke DM.** Hepatitis C-related post-transplant plasma cell proliferative disorder with hepatitis C virus in neoplastic plasma cells: a new posttransplant disease entity? *Transplant Proc* 2004; **36**: 2692-2696 [PMID: 15621126]
- 6 **Ward SC, Schiano TD, Thung SN, Fiel MI.** Plasma cell hepatitis in hepatitis C virus patients post-liver transplantation: case-control study showing poor outcome and predictive features in the liver explant. *Liver Transpl* 2009; **15**: 1826-1833 [PMID: 19938116 DOI: 10.1002/lt.21949]
- 7 **Fiel MI, Schiano TD.** Plasma cell hepatitis (de-novo autoimmune hepatitis) developing post liver transplantation. *Curr Opin Organ Transplant* 2012; **17**: 287-292 [PMID: 22498651 DOI: 10.1097/MOT.0b013e3283536622]
- 8 **Berardi S, Lodato F, Gramenzi A, D'Errico A, Lenzi M, Bonfadini A, Morelli MC, Tamè MR, Piscaglia F, Biselli M, Sama C, Mazzella G, Pinna AD, Grazi G, Bernardi M, Andreone P.** High incidence of allograft dysfunction in liver transplanted patients treated with pegylated-interferon alpha-2b and ribavirin for hepatitis C recurrence: possible de novo autoimmune hepatitis? *Gut* 2007; **56**: 237-242 [PMID: 16798778 DOI: 10.1136/gut.2006.092064]
- 9 **Gabler WL.** National Toxicology program report on fluoride and cancer. *J Oreg Dent Assoc* 1991; **60**: 32-33 [PMID: 1856781 DOI: 10.1016/S0140-6736(97)06478-7]

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Mansour A Parsi, MD, Series Editor

Endoscopic management of difficult common bile duct stones

Guru Trikudanathan, Udayakumar Navaneethan, Mansour A Parsi

Guru Trikudanathan, Department of Internal Medicine, University of Connecticut Medical Center, Farmington, CT 06269, United States

Udayakumar Navaneethan, Mansour A Parsi, Digestive Disease Institute, Cleveland Clinic, Cleveland, OH 44195, United States

Author contributions: Trikudanathan G, and Navaneethan U reviewed the literature and wrote the initial draft manuscript; Parsi MA supervised the process, revised and approved the manuscript for submission.

Correspondence to: Mansour A Parsi, MD, Digestive Disease Institute, Cleveland Clinic, 9500 Euclid Ave, Cleveland, OH 44195, United States. parsim@ccf.org

Telephone: +1-216-4454880 Fax: +1-216-4446305

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INTRODUCTION

Common bile duct (CBD) stones are seen in approximately 7%-12% of patients who undergo cholecystectomy for symptomatic cholelithiasis and are a common indication for referral to a biliary endoscopist^[1]. They vary in size ranging from rather small (approximately 1-2 mm) to very large (> 3 cm). Endoscopic retrograde cholangiopancreatography (ERCP) with endoscopic sphincterotomy (ES) and basket or balloon extraction are well established therapeutic procedures for the management of CBD stones. It is estimated that nearly 85%-95% of all CBD stones can be managed effectively by these conventional endoscopic methods^[2,3]. Failure to clear the bile duct renders the patient vulnerable to biliary obstruction, cholangitis and pancreatitis, thereby increasing the morbidity^[4,5]. The occurrence of acute cholangitis is associated with significant mortality, especially in the elderly, underscoring the need for early intervention to clear the bile duct of stones and to relieve the obstruction to achieve adequate biliary drainage. Extraction of CBD stones is one of the most commonly performed procedures by therapeutic endoscopists. With novel advances in extraction techniques and instruments emerging routinely, it is vital to keep abreast of the new developments in order to improve the outcome. This review focuses on the alter-

Abstract

Endoscopy is widely accepted as the first treatment option in the management of bile duct stones. In this review we focus on the alternative endoscopic modalities for the management of difficult common bile duct stones. Most biliary stones can be removed with an extraction balloon, extraction basket or mechanical lithotripsy after endoscopic sphincterotomy. Endoscopic papillary balloon dilation with or without endoscopic sphincterotomy or mechanical lithotripsy has been shown to be effective for management of difficult to remove bile duct stones in selected patients. Ductal clearance can be safely achieved with peroral cholangioscopy guided laser or electrohydraulic lithotripsy in most cases where other endoscopic treatment modalities have failed. Biliary stenting may be an alternative treatment option for frail and elderly patients or those with serious co morbidities.

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native endoscopic management options for the treatment of difficult to remove CBD stones.

REVIEW CRITERIA

In July 2011, we searched MEDLINE from 1982 to the present using the Medical Subject Headings terms common bile duct stone, endoscopic retrograde cholangiopancreatography, difficult stone, endoscopy, and the key word “common bile duct stone”. Full papers and abstracts in English language were considered. Important developments in research, reports from centers of excellence, and our own clinical experience in managing them, form the basis of this review article.

FACTORS ASSOCIATED WITH DIFFICULT TO TREAT BILE DUCT STONES

Multiple factors have been postulated to govern the success or failure of endoscopic extraction of CBD stones. In approximately 10%-15% of patients, managing biliary stones becomes formidable primarily due to difficulties in accessing the bile duct (periampullary diverticulum, sigmoid shaped CBD, post-gastrectomy Billroth type II anatomy, Roux-en-Y-gastrojejunostomy), large number of stones (greater than 10), large size of stones (stones with a diameter > 15 mm which cannot be grasped with a basket), unusually shaped stones (barrel-shaped) or location of the stones (intra hepatic, cystic duct, proximal to strictures)^[6]. In addition, endoscopic management becomes challenging in Mirizzi syndrome, in which stones in the cystic duct cause obstruction of the main bile duct^[7]. Kim *et al*^[8] prospectively evaluated the factors contributing to technical difficulty during endoscopic clearance of CBD stones. They reported that older age (> 65 years), previous gastrojejunostomy, larger stone size (≥ 15 mm), impaction of stones, shorter length of the distal CBD arm (≤ 36 mm), and more acute distal CBD angulation (≤ 135 degrees) are all contributors to technical difficulty for endoscopic removal of bile duct stones^[8].

MANAGEMENT OF DIFFICULT STONES

In high risk patients, the risks and benefits of alternative techniques for removal of bile duct stones not amenable to conventional endoscopic techniques must be carefully balanced against each other and with surgery. The individual decision concerning the appropriate therapy is also influenced by the local expertise and the availability of the technical equipment.

CBD stones up to 1.5 cm in diameter can be extracted intact after endoscopic sphincterotomy. The rate of successful retrieval progressively declines with increasing size of the stone^[9]. Larger stones especially those with a diameter ≥ 2 cm may need fragmentation before removal to reduce the risk of stone impaction.



Figure 1 Example of a three-layered mechanical lithotripsy device with the basket, inner plastic sheath and an outer metal sheath. The stone is captured by the basket and crushed against the outer metal sheath.

Mechanical lithotripsy

In 1982, Riemann *et al*^[10], first introduced mechanical lithotripsy (ML). ML is currently the most widely used technique for fragmentation of stones. Contemporary lithotripter baskets have a high breaking strengths and have improved the success rate of ML for extraction of large CBD stones (> 2 cm) to well over 90% without serious complications^[11]. Broadly speaking, there are two types of baskets for ML. The type of basket used depends on whether lithotripsy is done on an elective (“through the scope”) basis or on an emergent basis (salvage device) for basket impaction^[12]. The ‘through the scope’ model is typically a three-layer system with the basket, inner plastic sheath, and an outer metal sheath (Figure 1). The stone is captured with the basket and the outer metal sheath is advanced to the stone which will be crushed against it. Sometimes, unexpectedly, stone and basket impaction can occur even during routine extraction of smaller stones. Under such circumstances, stone fragmentation can be done after removing the handle from the basket and the duodenoscope from the patient. An endotripter (a spiral metal sheath) is introduced under fluoroscopic guidance, over the basket wires, and the stone is crushed after connecting the bare basket wires to the crank handle (Figure 2). The broken basket and the stone are then removed. The shaft of the endotripter is generally shorter and thicker than that employed in standard ML^[13]. Although basket impaction can occur with through the scope lithotripsy baskets; it is more commonly encountered with extraction baskets, which have thinner wires and weaker handles not suitable for fragmentation of stones.

In patients with multiple large stones, lithotripters with a sleeve system can be employed multiple times without withdrawal of the endoscope, facilitating stone fragmentation^[14,15].

ML has been widely used as it is a readily available, cost effective, and simple procedure. Unfortunately the failure rate is high especially in patients with stones greater than 2.8 cm in diameter^[15]. In a retrospective study the size of the stone was the only factor that significantly af-



Figure 2 Image of a mechanical lithotripter which can be used as a "salvage device" for removal of impacted baskets.

affected the success or failure of bile duct clearance. In this study of 162 patients, the cumulative probability of bile duct clearance ranged from > 90% for stones with a diameter less than 10 mm to 68% for those greater than 28 mm in diameter ($P < 0.02$)^[15]. A subsequent prospective study by Garg *et al*^[16] however reported that stone size alone may not be important unless considered together with the diameter of the bile duct. They concluded that the only important predictive factor that compromised the success of mechanical lithotripsy was stone impaction in the bile duct, with either an inability to pass the basket proximal to the stone or a failure of the basket to open fully around the stone to allow it to be grasped properly^[16]. Although stone composition was not included in the study by Leung *et al*^[12], some endoscopists believe that stones that are hard and densely calcified (visualized on a plain radiograph) resist mechanical fragmentation, resulting in an extraction failure with standard baskets. Although the stones which are molded to the shape of the bile duct may be softer, they are more difficult to crush because they may not be easily engaged by the lithotripter basket^[12].

A multi-center study reported the rate of complications associated with ML to be around 3.6%^[13]. Among the spectrum of complications, basket impaction or fracture of the basket wire are uniquely associated with ML. Non-surgical interventions that have been utilized in this setting include extension of sphincterotomy, awaiting spontaneous passage of the impacted basket and stone after successful stent placement, use of a second lithotripter, extracorporeal shock wave lithotripsy (ESWL), laser lithotripsy, electrohydraulic lithotripsy, and transhepatic lithotripsy and stone dislodgement^[13-18]. Other complications include broken handle and perforation or injury to the bile duct^[13-18].

In about 10% of the patients ML proves to be cumbersome, protracted and ineffective^[6] wherein one has to resort to other methods such as electrohydraulic, or laser lithotripsy for stone fragmentation and subsequent removal.

Electrohydraulic lithotripsy

Initially used as an industrial tool for fragmenting rocks in mines, its application was extrapolated to medical use

when Koch attempted fragmentation of biliary stones using this technology^[19]. Electrohydraulic Lithotripsy (EHL) consists of a bipolar lithotripsy probe which discharges sparks with the aid of a charge generator in an aqueous medium. The sparks generated under water generate high-frequency hydraulic pressure waves, the energy of which is absorbed by nearby stones and results in their fragmentation^[20]. The shock waves can cause inadvertent injury or perforation of the bile duct wall if the probe is not deployed close to the stone and away from the ductal wall. EHL can be performed under fluoroscopic guidance by using centering balloons or direct cholangioscopic vision^[20]. The disadvantage of using only fluoroscopic guidance is related to the two dimensional imaging and the inability to confirm correct positioning of the probe. Therefore direct visualization is frequently preferred to avoid damage to the ductal wall^[21]. A cholangioscope is inserted through the instrument channel of the mother scope. One or two dedicated biliary endoscopists are needed for this procedure. Continuous irrigation with water during the procedure generates a fluid medium for propagation of the shock waves and in addition offers a clear view of the stones by flushing away the debris^[20].

The overall complication rate ranges from 7% to 9%^[22,23], with most common complications being hemobilia, cholangitis, and less commonly, ductal perforation. Binmoeller *et al*^[6], in one of the earlier large studies reported that EHL was successful in 63 of the 64 patients who had failed previous attempts of ML. Smaller published studies report stone fragmentation rates between 77%-100% for peroral EHL with minimal complications^[22,24-28]. Arya *et al*^[23], reported a stone fragmentation rate of 96% and final stone clearance of 90%. In a retrospective study of 94 patients who had failed stone extraction by conventional techniques, Hui *et al*^[29], compared the outcomes of EHL with further endoscopy to stenting alone in a subset of elderly and infirm patients. They demonstrated that EHL and further ERCPs had a higher success rate (80%) with a low complication rate (7.7%) and recommended that elderly and frail patients should be referred to tertiary centers for EHL in order to prolong survival and decrease biliary complications^[29].

The EHL equipment is compact, requires no special electricity, and is relatively inexpensive^[30]. Other advantages are that the EHL procedure does not require special training or protective gear. In the United States, use of EHL for fragmentation and removal of biliary stones is quite common in centers with special interest in biliary disorders.

Laser lithotripsy

In laser lithotripsy (LL), laser light at a particular wavelength is focused on the surface of the stone to induce a wave-mediated fragmentation. The pulsed laser energy utilized in stone fragmentation is in contrast to the continuous laser energy used in tumor ablation^[7]. The first successful use of pulsed laser for shock-wave lithotripsy of bile duct stones was reported in 1986^[31]. Since then

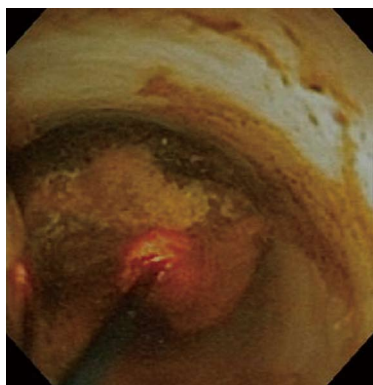


Figure 3 Laser lithotripsy of a bile duct stone under cholangioscopic guidance.

the technology has evolved and other laser types such as neodymium: yttrium-aluminum-garnet (Nd: YAG), flash lamp-pulsed dye (coumarin), the flash lamp-pulsed dye (rhodamine) with an automatic stone recognition system and the new Frequency Doubled Double Pulse Nd:YAG (FREDDY) system have been introduced^[32-35]. LL is typically performed perorally under cholangioscopic or fluoroscopic guidance or by the transhepatic approach. As with EHL, LL under direct visualization using a cholangioscope is often preferred to avoid damage to the ductal wall (Figure 3).

Based on some reports, ductal clearance can be accomplished in 64% to 97% of patients by using^[7,36] LL. In the majority of patients ductal clearance could be achieved in one session, although more sessions were required occasionally. LL has been demonstrated to be more effective than ESWL in terms of stone clearance rate and more rapid stone fragmentation with a shorter duration of treatment leading to a significant reduction in cost^[37,38]. In some centers where the laser equipment is available, laser lithotripsy has gained popularity and has managed to replace EHL as the primary modality for fragmentation of difficult to remove stones.

A recent innovation worth mentioning is the introduction of a double-lumen basket which allows passage of a laser probe for effective laser lithotripsy after the stone is captured by the basket^[39]. For a selected group of patients, this technique was shown to be feasible and effective, and the authors hope that continuous improvements in designs and construction materials would further enhance the success rate of this device.

Extracorporeal shockwave lithotripsy

In ESWL, high-pressure shock waves are generated outside the body (extracorporeal) by underwater spark gap (electrohydraulic) generated by piezoelectric crystals or electromagnetic membrane technology^[30]. The shock waves are focused by elliptical transducers to the designated target through a liquid or tissue medium which prevents the energy attenuation. ESWL is performed under ultrasound or fluoroscopic guidance. Since most

biliary stones are radiolucent and are not adequately visualized by fluoroscopy alone, placement of a nasobiliary tube for contrast instillation is required if ESWL is performed under fluoroscopy. In case of first generation lithotripters, the patients needed to be immersed in a water bath. Subsequent ESWL lithotripters employ water-filled compressible bags and a gel is applied to the skin surface for interface with the patient. Comparison between the lithotripters at a single center showed no significant difference in fragmentation of CBD stones^[40]. General anesthesia is usually needed as the discomfort produced may not be adequately controlled by conscious sedation. The critical determining factor for success of single ESWL session is stone size and microcrystalline structure and architecture of the stone^[41,42]. The presence or absence of bile duct stenosis can also influence the success of ESWL^[43]. Sauerbruch *et al*^[44], reported the efficacy of ESWL in achieving CBD stone fragmentation in over 90% of patients with minimal side effects.

In most institutions that already have access to ESWL for treatment of renal calculi, no other purchases of equipment needs to be made. For ESWL direct contact with the calculi is not needed, and multiple stones can be managed simultaneously^[41]. ESWL can be of particular help in patients with abnormal anatomy such as those who have undergone Billroth- II or Roux-en-Y surgeries in whom endoscopic access to the major papilla is difficult.

Although in general ESWL is tolerated well, it can be associated with adverse events such as transient biliary colic, subcutaneous ecchymosis, cardiac arrhythmia, self limited hemobilia, cholangitis, ileus and pancreatitis^[7,45]. Perinephric hematoma, biliary obstruction, bowel perforation, lung injury and splenic rupture are among the rarely reported complications^[42]. Multiple ESWL sessions may be required in a subset of patients to achieve ductal clearance, and endoscopic procedures between the ESWL sessions may become necessary to clear the bile duct of debris to assure drainage. The recurrence rate of CBD stones after ESWL clearance during a 1 to 2 year follow up was considerable and was around 14%^[45,46]. In a randomized trial comparing fluoroscopic guided ESWL and LL, LL was preferable not only for successful stone free rate (73% *vs* 97%), but also in terms of the number of sessions needed to clear the duct (3 in ESWL *vs* 1.2 in LL) and the duration of treatment^[30]. Another randomized trial comparing ultrasound guided ESWL and laser lithotripsy in the treatment of CBD stones refractory to conventional treatment clearly showed superior stone clearance rate and cost effectiveness in laser lithotripsy (52.4% *vs* 82%)^[40]. However, a prospective trial comparing EHL *vs* ESWL showed no difference in success rates for clearing the CBD, duration and cost of hospitalization between the two modalities^[24].

In the United States, ESWL is rarely performed for management of biliary stones and most centers prefer cholangioscopy-guided LL or EHL for this purpose.

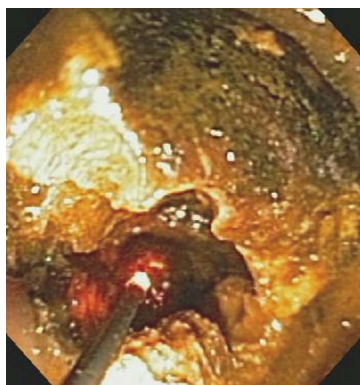


Figure 4 Direct peroral cholangioscopy guided laser lithotripsy of a bile duct stone. The red laser light makes targeting of the stone easier).

Cholangioscopy-guided lithotripsy

EHL or LL is ideally performed under direct visual control using a cholangioscope. The traditional cholangioscopy systems consist of a duodenoscope (also called “mother scope”) and a dedicated cholangioscope (also called “baby scope” or “daughter scope”) which is introduced into the bile duct through the accessory channel of the mother scope. The baby scope itself has an instrument channel through which the EHL or laser probe is introduced. Peroral cholangioscopy guided EHL or LL is cumbersome and can be labor intensive requiring an additional endoscopic unit and often participation of two skilled endoscopists, one to handle the duodenoscope and the other to maneuver the cholangioscope. The traditional cholangioscopes are capable of only 2-way steering, which may limit the field of view. These cholangioscopes are also extremely vulnerable to damage, requiring frequent expensive repairs. Furthermore, sharp angulations in the biliary tree may limit access into intrahepatic ducts or the cystic duct^[47].

To negate these limitations, the Spyglass Direct Visualization System (Boston Scientific Corp, Natick, Mass) was designed for single operator examination of the bile ducts, 4-way steering, and dedicated irrigation channels^[48,49]. In a large international multicenter study conducted at 10 centers in the United States and 5 centers in Europe, ductal clearance was successfully achieved in 71% of the patients using the Spyglass cholangioscopy system^[50]. Despite its effectiveness, the spyglass system has been underutilized mainly due to its fiber optic image quality which is inferior to the video image quality offered by the new videocholangioscopes^[51].

Direct peroral cholangioscopy

In direct peroral cholangioscopy (DPOC), an ultraslim upper endoscope is maneuvered across the biliary sphincter and into the bile duct for direct observation. With the introduction of high-definition ultraslim endoscopes with narrow band imaging capability, direct peroral cholangioscopy has gained popularity. This is mainly due to the many advantages of this technique. Compared to ductoscopy using a dedicated cholangioscope, direct cholangios-

copy has several advantages. It offers a single operator platform, digital image quality and simultaneous irrigation and therapeutic capabilities. The most profound disadvantage of DPOC, however, is the difficulty associated with traversing the biliary sphincter to gain access to the bile duct. This is mainly due to looping of the ultraslim upper endoscope in the stomach or in the duodenum. To enhance the success rate of DPOC, specialized accessories or techniques are needed to advance the ultra slim endoscope into the proximal biliary system.

Larghi and Waxman^[52] reported their experience in which the ultraslim upper endoscope was inserted with the aid of a guidewire placed during ERCP to maintain access. Additional use of manual pressure applied on the patient's abdomen has been shown to ease the passage of the ultraslim endoscope into the hilar area in some patients^[53]. The main drawback encountered with passage of an ultraslim upper endoscope over a guidewire for gaining access to the bile duct is the dislodgement of the guidewire from the bile duct and also large loop formation hindering the entrance of the endoscope into the biliary system. Moon *et al*^[54] demonstrated the ropeway technique using an intraductal balloon that can be anchored within an intrahepatic bile duct to advance an ultraslim upper endoscope into the biliary tree for performance of DPOC. However, withdrawal of the balloon may cause technical difficulties in maintaining access which underscores the need for other accessories to maintain the scope's position within the bile duct. The same group also reported use of an overtube balloon originally designed for double balloon enteroscopy to facilitate the introduction of an ultraslim upper endoscope into the biliary tree^[55]. However the large inner diameter of the overtube (10.8 mm) compared to the outer diameter of the ultraslim upper endoscopes (5.2-6 mm), makes it difficult to manipulate both instruments and results in discomfort to the patient, looping of the endoscope in the duodenum, and difficulty in reaching the proximal bile duct^[55,56].

In a recent study, we assessed utility of a novel anchoring balloon for performance of DPOC. Use of the anchoring balloon allowed consistent access to the biliary tree for performance of diagnostic and therapeutic DPOC distal to the confluence of the right and left hepatic ducts. More proximal access, however, was challenging owing to looping of the ultraslim endoscope after balloon removal.

Efforts are underway to develop the combined use of an intraductal anchoring balloon and an overtube especially designed for DPOC. Very recently, air embolism was reported following DPOC, resulting in a left sided hemiparesis^[57]. Endoscopists must be conscious of the fact that air embolism could remain asymptomatic, as in regional embolism (portal venous gas) or manifest as hypoxia, shock, cardiac arrest or cerebral ischemia as noted in this case^[57].

Once the hurdle of introducing an ultraslim upper endoscope into the bile duct has been overcome, a laser



Figure 5 Large diameter papillary balloon dilatation to remove bile duct stone.

or an EHL probe can easily be passed through the working channel of the ultraslim endoscope. Stones can be directly visualized before and during lithotripsy by DPOC (Figure 4). Lithotripsy is performed using this technique until the stones are satisfactorily fragmented to allow removal through the biliary sphincter. In a small study with 18 patients who had failed conventional endoscopic therapy including ML, DPOC guided EHL or LL was successful in approximately 90% of the patients with an average of 1.6 treatment sessions per patient^[58].

Specialized ultraslim upper endoscopes are being designed to facilitate access to the biliary tree for DPOC^[59]. Ultraslim upper endoscopes can be passed through the nasal cavity for performance of DPOC. Direct transnasal cholangioscopy has been reported for successful extraction of CBD stones^[60].

Endoscopic papillary balloon dilatation

Endoscopic papillary balloon dilation (EPBD) was introduced as an alternative to endoscopic sphincterotomy for removal of bile duct stones in 1980's^[61]. In an initial report involving 10 patients with CBD stones, biliary sphincteroplasty to 15 mm allowed removal of CBD stones in 6 patients^[61]. In the other 4 patients, ductal clearance required a combination of biliary sphincterotomy and mechanical lithotripsy. The use of large diameter papillary balloon dilatation (up to 20 mm in diameter) for management of difficult to remove biliary stones was reported by Ersoz *et al*^[62] in 2003. They reported a high success rate for stone removal. However, their complication rate was also high. Several reports have suggested that EPBD is associated with risk of severe pancreatitis which raises safety concern of this procedure^[61,63]. In our institution, we use EPBD selectively and try to avoid its use in those with high risk of post ERCP pancreatitis (Figure 5).

Since those initial reports, multiple studies have shown that EPBD alone or in combination with other techniques can be of use for management of difficult to remove biliary stones^[62-68]. EPBD is especially attractive in patients who are at risk for bleeding after endoscopic sphincterotomy or in those with altered anatomy

in whom a full sphincterotomy cannot be successfully achieved.

Some authors have suggested that the stone recurrence rate may also be higher with EPBD than with endoscopic sphincterotomy and mechanical lithotripsy^[69]. However the results of a Japanese multicentric trial with a mean follow up of 6.7 years demonstrated that there is lesser risk of stone recurrence following EPBD when compared with sphincterotomy^[70]. Further, a recent meta-analysis which included 15 randomized trials comparing EPBD and endoscopic sphincterotomy showed reduced risk of bleeding and infections and is especially indicated in older patients, those who are at risk for infection and coagulopathy^[71]. Despite its effectiveness, EPBD has been associated with serious complications^[63]. A higher risk of post ERCP pancreatitis has been observed which has been attributed to the inadequately loosened sphincter of Oddi and the intra mucosal hemorrhage and inflammation/edema around the papilla. This may cause compression of the pancreatic duct and may accentuate the risk of pancreatitis^[72]. In this regard, a randomized controlled trial demonstrated that a 5-min dilation time as opposed to the conventional 1-min time resulted in an adequately loosened sphincter of Oddi and consequently reduced the risk of post ERCP pancreatitis and improved its efficacy^[73]. The rate of these complications can be reduced by strict patient selection, avoidance of forced procedures, optimal dilation duration and immediate conversion to an alternative procedure if any difficulty is encountered during EPBD.

Endoscopic biliary stenting

In very old patients and those with serious co-morbidities where other endoscopic or surgical procedures may confer unacceptably high risks, endoscopic biliary stenting is a useful alternative^[69]. Biliary drainage by stenting is mandatory if ductal clearance cannot be achieved during ERCP or in between procedures in patients who require more than one session for ductal clearance. CBD stones have been reported to reduce in size in 60% of patients within one to two years after biliary stenting^[69].

Mechanical irritation of the stent on the stone is postulated to be one of the mechanisms.

In a study involving 28 geriatric patients with CBD stones refractory to conventional endoscopic removal, endoscopic biliary stent placement combined with oral ursodeoxycholic acid and terpene therapy for a mean of six months led to significant reduction in the size of CBD stone^[74]. Subsequently, endoscopic stone removal was successfully performed in 26 of 28 patients with a mean of 1.7 ERCP procedures. This combination therapy may be of use for treatment of difficult to remove CBD stones in a subset of patients with significant co-morbidities and intolerance to prolonged endoscopic treatment modalities.

In conclusion, the past several years have witnessed the emergence of new technologies and techniques for management of difficult to remove biliary stones. Treat-

ment of such stones is generally accomplished using a multimodal approach combining conventional techniques such as endoscopic sphincterotomy, use of extraction balloons and baskets and mechanical lithotripsy, with newer techniques such as cholangioscopy guided laser or electrohydraulic lithotripsy. Recent advances in the development of videocholangioscopes, single operator catheter-based cholangioscopes and specially-designed ultrathin upper endoscopes for DPOC have made lithotripsy under direct visual guidance safer, more reliable, and more routine. Future studies will certainly shed more light on the safety of different modalities for stone extraction and will help determine the best management approach for different subgroup of patients with difficult to remove bile duct stones.

REFERENCES

- Freitas ML, Bell RL, Duffy AJ. Choledocholithiasis: evolving standards for diagnosis and management. *World J Gastroenterol* 2006; **12**: 3162-3167 [PMID: 16718834]
- Samardzic J, Latic F, Kraljik D, Pitlovic V, Mrkovic H, Miskic D, Latic A, Delibegovic S. Treatment of common bile duct stones--is the role of ERCP changed in era of minimally invasive surgery? *Med Arh* 2010; **64**: 187-188 [PMID: 20645517]
- Strömberg C, Nilsson M. Nationwide study of the treatment of common bile duct stones in Sweden between 1965 and 2009. *Br J Surg* 2011; **98**: 1766-1774 [PMID: 21935910 DOI: 10.1002/bjs.7690]
- Cairns SR, Dias L, Cotton PB, Salmon PR, Russell RC. Additional endoscopic procedures instead of urgent surgery for retained common bile duct stones. *Gut* 1989; **30**: 535-540 [PMID: 2714686 DOI: 10.1136/gut.30.4.535]
- Parsi MA, Stevens T, Dumot JA, Zuccaro G. Endoscopic therapy of recurrent acute pancreatitis. *Cleve Clin J Med* 2009; **76**: 225-233 [PMID: 19339638 DOI: 10.3949/ccjm.76a.08017]
- Binmoeller KF, Brückner M, Thonke F, Soehendra N. Treatment of difficult bile duct stones using mechanical, electrohydraulic and extracorporeal shock wave lithotripsy. *Endoscopy* 1993; **25**: 201-206 [PMID: 8519238 DOI: 10.1055/s-2007-1010293]
- McHenry L, Lehman G. Difficult bile duct stones. *Curr Treat Options Gastroenterol* 2006; **9**: 123-132 [PMID: 16539873 DOI: 10.1007/s11938-006-0031-6]
- Kim HJ, Choi HS, Park JH, Park DI, Cho YK, Sohn CI, Jeon WK, Kim BI, Choi SH. Factors influencing the technical difficulty of endoscopic clearance of bile duct stones. *Gastrointest Endosc* 2007; **66**: 1154-1160 [PMID: 17945223 DOI: 10.1016/j.gie.2007.04.033]
- Lauri A, Horton RC, Davidson BR, Burroughs AK, Dooley JS. Endoscopic extraction of bile duct stones: management related to stone size. *Gut* 1993; **34**: 1718-1721 [PMID: 8282260 DOI: 10.1136/gut.34.12.1718]
- Riemann JF, Seuberth K, Demling L. Clinical application of a new mechanical lithotripter for smashing common bile duct stones. *Endoscopy* 1982; **14**: 226-230 [PMID: 7140657 DOI: 10.1055/s-2007-1021626]
- Schneider MU, Matek W, Bauer R, Domschke W. Mechanical lithotripsy of bile duct stones in 209 patients--effect of technical advances. *Endoscopy* 1988; **20**: 248-253 [PMID: 3168938 DOI: 10.1055/s-2007-1018186]
- Leung JW, Tu R. Mechanical lithotripsy for large bile duct stones. *Gastrointest Endosc* 2004; **59**: 688-690 [PMID: 15114312 DOI: 10.1016/S0016-5107(04)00174-9]
- Thomas M, Howell DA, Carr-Locke D, Mel Wilcox C, Chak A, Rajman I, Watkins JL, Schmalz MJ, Geenen JE, Catalano MF. Mechanical lithotripsy of pancreatic and biliary stones: complications and available treatment options collected from expert centers. *Am J Gastroenterol* 2007; **102**: 1896-1902 [PMID: 17573790 DOI: 10.1111/j.1572-0241.2007.01350.]
- Chung SC, Leung JW, Leong HT, Li AK. Mechanical lithotripsy of large common bile duct stones using a basket. *Br J Surg* 1991; **78**: 1448-1450 [PMID: 1773322 DOI: 10.1002/bjs.1800781214]
- Cipolletta L, Costamagna G, Bianco MA, Rotondano G, Piscopo R, Mutignani M, Marmo R. Endoscopic mechanical lithotripsy of difficult common bile duct stones. *Br J Surg* 1997; **84**: 1407-1409 [PMID: 9361600 DOI: 10.1002/bjs.1800841019]
- Garg PK, Tandon RK, Ahuja V, Makharia GK, Batra Y. Predictors of unsuccessful mechanical lithotripsy and endoscopic clearance of large bile duct stones. *Gastrointest Endosc* 2004; **59**: 601-605 [PMID: 15114300 DOI: 10.1016/S0016-5107(04)00295-0]
- Shim CS. How Should Biliary Stones be Managed? *Gut Liver* 2010; **4**: 161-172 [PMID: 20559517]
- Shim CS. How Should Biliary Stones be Managed? *Gut Liver* 2010; **4**: 161-172 [PMID: 20559517 DOI: 10.5009/gnl.2010.4.2.161]
- Koch H, Stolte M, Walz V. Endoscopic lithotripsy in the common bile duct. *Endoscopy* 1977; **9**: 95-98 [PMID: 891486 DOI: 10.1055/s-0028-1098497]
- Seitz U, Bapaye A, Bohnacker S, Navarrete C, Maydeo A, Soehendra N. Advances in therapeutic endoscopic treatment of common bile duct stones. *World J Surg* 1998; **22**: 1133-1144 [PMID: 9828721 DOI: 10.1007/s002689900532]
- Yoo KS, Lehman GA. Endoscopic management of biliary ductal stones. *Gastroenterol Clin North Am* 2010; **39**: 209-27, viii [PMID: 20478483 DOI: 10.1016/j.gtc.2010.02.008]
- Blind PJ, Lundmark M. Management of bile duct stones: lithotripsy by laser, electrohydraulic, and ultrasonic techniques. Report of a series and clinical review. *Eur J Surg* 1998; **164**: 403-409 [PMID: 9696440]
- Arya N, Nelles SE, Haber GB, Kim YI, Kortan PK. Electrohydraulic lithotripsy in 111 patients: a safe and effective therapy for difficult bile duct stones. *Am J Gastroenterol* 2004; **99**: 2330-2334 [PMID: 15571578 DOI: 10.1111/j.1572-0241.2004.40251.]
- Adamek HE, Buttmann A, Wessbecher R, Kohler B, Riemann JF. Clinical comparison of extracorporeal piezoelectric lithotripsy (EPL) and intracorporeal electrohydraulic lithotripsy (EHL) in difficult bile duct stones. A prospective randomized trial. *Dig Dis Sci* 1995; **40**: 1185-1192 [PMID: 7781432 DOI: 10.1007/BF02065522]
- Adamek HE, Maier M, Jakobs R, Wessbecher FR, Neuhauser T, Riemann JF. Management of retained bile duct stones: a prospective open trial comparing extracorporeal and intracorporeal lithotripsy. *Gastrointest Endosc* 1996; **44**: 40-47 [PMID: 8836715]
- Leung JW, Chung SS. Electrohydraulic lithotripsy with peroral choledochoscopy. *BMJ* 1989; **299**: 595-598 [PMID: 2508816 DOI: 10.1136/bmj.299.6699.595]
- Yasuda K, Nakajima M, Cho E, Mukai H, Kawai K. Comparison of peroral and percutaneous cholangioscopy. *Endoscopy* 1989; **21** Suppl 1: 347-350 [PMID: 2606084 DOI: 10.1055/s-2007-1012988]
- Moon JH, Cha SW, Ryu CB, Kim YS, Hong SJ, Cheon YK, Cho YD, Kim YS, Lee JS, Lee MS, Shim CS, Kim BS. Endoscopic treatment of retained bile-duct stones by using a balloon catheter for electrohydraulic lithotripsy without cholangioscopy. *Gastrointest Endosc* 2004; **60**: 562-566 [PMID: 15472679 DOI: 10.1016/S0016-5107(04)02012-7]
- Hui CK, Lai KC, Ng M, Wong WM, Yuen MF, Lam SK, Lai CL, Wong BC. Retained common bile duct stones: a comparison between biliary stenting and complete clearance of stones by electrohydraulic lithotripsy. *Aliment Pharmacol Ther* 2003; **17**: 289-296 [PMID: 12534415 DOI: 10.1046/

- j.1365-2036.2003.01415.]
- 30 **DiSario J**, Chuttani R, Croffie J, Liu J, Mishkin D, Shah R, Somogyi L, Tierney W, Song LM, Petersen BT. Biliary and pancreatic lithotripsy devices. *Gastrointest Endosc* 2007; **65**: 750-756 [PMID: 17383651 DOI: 10.1016/j.gie.2006.10.002]
- 31 **Lux G**, Ell C, Hochberger J, Müller D, Demling L. The first successful endoscopic retrograde laser lithotripsy of common bile duct stones in man using a pulsed neodymium-YAG laser. *Endoscopy* 1986; **18**: 144-145 [PMID: 2874019 DOI: 10.1055/s-2007-1018356]
- 32 **Ponchon T**, Gagnon P, Valette PJ, Henry L, Chavaillon A, Thieulin F. Pulsed dye laser lithotripsy of bile duct stones. *Gastroenterology* 1991; **100**: 1730-1736 [PMID: 1673442]
- 33 **Ell C**, Hochberger J, May A, Fleig WE, Bauer R, Mendez L, Hahn EG. Laser lithotripsy of difficult bile duct stones by means of a rhodamine-6G laser and an integrated automatic stone-tissue detection system. *Gastrointest Endosc* 1993; **39**: 755-762 [PMID: 8293896 DOI: 10.1016/S0016-5107(93)70259-X]
- 34 **Kim TH**, Oh HJ, Choi CS, Yeom DH, Choi SC. Clinical usefulness of transpapillary removal of common bile duct stones by frequency doubled double pulse Nd: YAG laser. *World J Gastroenterol* 2008; **14**: 2863-2866 [PMID: 18473411 DOI: 10.3748/wjg.14.2863]
- 35 **Hochberger J**, Bayer J, Maiss J, Tex S, Hahn EG. [Clinical results with a new frequency-doubled, double pulse Nd: YAG laser (FREDDY) for lithotripsy in complicated choledocholithiasis]. *Biomed Tech (Berl)* 1998; **43** Suppl: 172 [PMID: 9859311 DOI: 10.1515/bmte.1998.43.s1.172]
- 36 **Hochberger J**, Bayer J, May A, Mühldorfer S, Maiss J, Hahn EG, Ell C. Laser lithotripsy of difficult bile duct stones: results in 60 patients using a rhodamine 6G dye laser with optical stone tissue detection system. *Gut* 1998; **43**: 823-829 [PMID: 9824611 DOI: 10.1136/gut.43.6.823]
- 37 **Neuhaus H**, Zillinger C, Born P, Ott R, Allescher H, Röscher T, Classen M. Randomized study of intracorporeal laser lithotripsy versus extracorporeal shock-wave lithotripsy for difficult bile duct stones. *Gastrointest Endosc* 1998; **47**: 327-334 [PMID: 9609422 DOI: 10.1016/S0016-5107(98)70214-7]
- 38 **Jakobs R**, Adamek HE, Maier M, Krömer M, Benz C, Martin WR, Riemann JF. Fluoroscopically guided laser lithotripsy versus extracorporeal shock wave lithotripsy for retained bile duct stones: a prospective randomised study. *Gut* 1997; **40**: 678-682 [PMID: 9203950]
- 39 **Lee JE**, Moon JH, Choi HJ, Song AR, Jung EK, Cheon YK, Cho YD, Lee JS, Lee MS. Endoscopic treatment of difficult bile duct stones by using a double-lumen basket for laser lithotripsy--a case series. *Endoscopy* 2010; **42**: 169-172 [PMID: 19998219 DOI: 10.1055/s-0029-1215353]
- 40 **den Toom R**, Nijs HG, van Blankenstein M, Laméris JS, Schröder FH, Terpstra OT. Extracorporeal shock wave treatment of common bile duct stones: experience with two different lithotriptors at a single institution. *Br J Surg* 1991; **78**: 809-813 [PMID: 1873707 DOI: 10.1002/bjs.1800780714]
- 41 **White DM**, Correa RJ, Gibbons RP, Ball TJ, Kozarek RJ, Thirlby RC. Extracorporeal shock-wave lithotripsy for bile duct calculi. *Am J Surg* 1998; **175**: 10-13 [PMID: 9445230 DOI: 10.1016/S0002-9610(97)00234-1]
- 42 **Tandan M**, Reddy DN, Santosh D, Reddy V, Koppuju V, Lakhtakia S, Gupta R, Ramchandani M, Rao GV. Extracorporeal shock wave lithotripsy of large difficult common bile duct stones: efficacy and analysis of factors that favor stone fragmentation. *J Gastroenterol Hepatol* 2009; **24**: 1370-1374 [PMID: 19702905 DOI: 10.1111/j.1440-1746.2009.05919.x]
- 43 **Amplatz S**, Piazza L, Felder M, Comberlato M, Benvenuti S, Zancanella L, Di Fede F, de'Guelmi A, Bertozzo A, Farris P, Grasso T, Mega A, Chilovi F. Extracorporeal shock wave lithotripsy for clearance of refractory bile duct stones. *Dig Liver Dis* 2007; **39**: 267-272 [PMID: 17275426 DOI: 10.1016/j.dld.2006.11.003]
- 44 **Sauerbruch T**, Stern M. Fragmentation of bile duct stones by extracorporeal shock waves. A new approach to biliary calculi after failure of routine endoscopic measures. *Gastroenterology* 1989; **96**: 146-152 [PMID: 2642439 DOI: 10.1111/j.1572-0241.2004.30151.x]
- 45 **Parsi MA**, Stevens T, Lopez R, Vargo JJ. Extracorporeal shock wave lithotripsy for prevention of recurrent pancreatitis caused by obstructive pancreatic stones. *Pancreas* 2010; **39**: 153-155 [PMID: 19820418 DOI: 10.1097/MPA.0b013e3181bb1733]
- 46 **Conigliaro R**, Camellini L, Zuliani CG, Sassatelli R, Mortilla MG, Bertoni G, Formisano D, Bedogni G. Clearance of irretrievable bile duct and pancreatic duct stones by extracorporeal shockwave lithotripsy, using a transportable device: effectiveness and medium-term results. *J Clin Gastroenterol* 2006; **40**: 213-219 [PMID: 16633122 DOI: 10.1097/00004836-200603000-00008]
- 47 **Chathadi KV**, Chen YK. New kid on the block: development of a partially disposable system for cholangioscopy. *Gastrointest Endosc Clin N Am* 2009; **19**: 545-555 [PMID: 19917460 DOI: 10.1016/j.giec.2009.06.001]
- 48 **Chen YK**. Preclinical characterization of the Spyglass peroral cholangiopancreatography system for direct access, visualization, and biopsy. *Gastrointest Endosc* 2007; **65**: 303-311 [PMID: 17258991 DOI: 10.1016/j.gie.2006.07.048]
- 49 **Chen YK**, Pleskow DK. SpyGlass single-operator peroral cholangiopancreatography system for the diagnosis and therapy of bile-duct disorders: a clinical feasibility study (with video). *Gastrointest Endosc* 2007; **65**: 832-841 [PMID: 17466202 DOI: 10.1016/j.gie.2007.01.025]
- 50 **Chen YK**, Parsi MA, Binmoeller KF, Hawes RH, Pleskow DK, Slivka A, Haluszka O, Petersen BT, Sherman S, Deviere J, Meisner S, Stevens PD, Costamagna G, Ponchon T, Peetermans JA, Neuhaus H. Single-operator cholangioscopy in patients requiring evaluation of bile duct disease or therapy of biliary stones (with videos). *Gastrointest Endosc* 2011; **74**: 805-814 [PMID: 21762903 DOI: 10.1016/j.gie.2011.04.016]
- 51 **Parsi MA**. Peroral cholangioscopy in the new millennium. *World J Gastroenterol* 2011; **17**: 1-6 [PMID: 21218076 DOI: 10.3748/wjg.v17.i1.1]
- 52 **Larghi A**, Waxman I. Endoscopic direct cholangioscopy by using an ultra-slim upper endoscope: a feasibility study. *Gastrointest Endosc* 2006; **63**: 853-857 [PMID: 16650553 DOI: 10.1016/j.gie.2005.07.050]
- 53 **Park do H**, Park BW, Lee HS, Park SH, Park JH, Lee SH, Kim HS, Kim SJ. Peroral direct cholangioscopic argon plasma coagulation by using an ultraslim upper endoscope for recurrent hepatoma with intraductal nodular tumor growth (with videos). *Gastrointest Endosc* 2007; **66**: 201-203 [PMID: 17531236 DOI: 10.1016/j.gie.2006.11.037]
- 54 **Moon JH**, Ko BM, Choi HJ, Hong SJ, Cheon YK, Cho YD, Lee JS, Lee MS, Shim CS. Intraductal balloon-guided direct peroral cholangioscopy with an ultraslim upper endoscope (with videos). *Gastrointest Endosc* 2009; **70**: 297-302 [PMID: 19394010 DOI: 10.1016/j.gie.2008.11.019]
- 55 **Choi HJ**, Moon JH, Ko BM, Hong SJ, Koo HC, Cheon YK, Cho YD, Lee JS, Lee MS, Shim CS. Overtube-balloon-assisted direct peroral cholangioscopy by using an ultra-slim upper endoscope (with videos). *Gastrointest Endosc* 2009; **69**: 935-940 [PMID: 19327480 DOI: 10.1016/j.gie.2008.08.043]
- 56 **Tsou YK**, Lin CH, Tang JH, Liu NJ, Cheng CL. Direct peroral cholangioscopy using an ultraslim endoscope and overtube balloon-assisted technique: a case series. *Endoscopy* 2010; **42**: 681-684 [PMID: 20669079 DOI: 10.1055/s-0030-1255616]
- 57 **Efthymiou M**, Raftopoulos S, Antonio Chirinos J, May GR. Air embolism complicated by left hemiparesis after direct cholangioscopy with an intraductal balloon anchoring system. *Gastrointest Endosc* 2012; **75**: 221-223 [PMID: 21470606 DOI: 10.1016/j.gie.2011.01.038]

- 58 **Moon JH**, Ko BM, Choi HJ, Koo HC, Hong SJ, Cheon YK, Cho YD, Lee MS, Shim CS. Direct peroral cholangioscopy using an ultra-slim upper endoscope for the treatment of retained bile duct stones. *Am J Gastroenterol* 2009; **104**: 2729-2733 [PMID: 19623165 DOI: 10.1038/ajg.2009.435]
- 59 **Itoi T**, Moon JH, Waxman I. Current status of direct peroral cholangioscopy. *Dig Endosc* 2011; **23** Suppl 1: 154-157 [PMID: 21535223 DOI: 10.1111/j.1443-1661.2011.01114.x]
- 60 **Seo MS**, Moon JH, Choi HJ, Kim HK, Cheon YK, Cho YD, Lee MS. Bile-Duct Stone Removal under Direct Transnasal Cholangioscopy Using an Ultraslim Upper Endoscope. *Gut Liver* 2010; **4**: 428-429 [PMID: 20981228 DOI: 10.5009/gnl.2010.4.3.428]
- 61 **Staritz M**, Ewe K, Meyer zum Büschenfelde KH. Endoscopic papillary dilatation, a possible alternative to endoscopic papillotomy. *Lancet* 1982; **1**: 1306-1307 [PMID: 6123047 DOI: 10.1016/S0140-6736(82)92873-2]
- 62 **Ersoz G**, Tekesin O, Ozutemiz AO, Gunsar F. Biliary sphincterotomy plus dilation with a large balloon for bile duct stones that are difficult to extract. *Gastrointest Endosc* 2003; **57**: 156-159 [PMID: 12556775 DOI: 10.1067/mge.2003.52]
- 63 **Disario JA**, Freeman ML, Bjorkman DJ, Macmathuna P, Petersen BT, Jaffe PE, Morales TG, Hixson LJ, Sherman S, Lehman GA, Jamal MM, Al-Kawas FH, Khandelwal M, Moore JP, Derfus GA, Jamidar PA, Ramirez FC, Ryan ME, Woods KL, Carr-Locke DL, Alder SC. Endoscopic balloon dilation compared with sphincterotomy for extraction of bile duct stones. *Gastroenterology* 2004; **127**: 1291-1299 [PMID: 15520997 DOI: 10.1053/j.gastro.2004.07.017]
- 64 **Itoi T**, Itokawa F, Sofuni A, Kurihara T, Tsuchiya T, Ishii K, Tsuji S, Ikeuchi N, Moriyasu F. Endoscopic sphincterotomy combined with large balloon dilation can reduce the procedure time and fluoroscopy time for removal of large bile duct stones. *Am J Gastroenterol* 2009; **104**: 560-565 [PMID: 19174779 DOI: 10.1038/ajg.2008.67]
- 65 **Stefanidis G**, Viazis N, Pleskow D, Manolakopoulos S, Theocharis L, Christodoulou C, Kotsikoros N, Giannousis J, Sgouros S, Rodias M, Katsikani A, Chuttani R. Large balloon dilation vs. mechanical lithotripsy for the management of large bile duct stones: a prospective randomized study. *Am J Gastroenterol* 2011; **106**: 278-285 [PMID: 21045816 DOI: 10.1038/ajg.2010.421]
- 66 **Heo JH**, Kang DH, Jung HJ, Kwon DS, An JK, Kim BS, Suh KD, Lee SY, Lee JH, Kim GH, Kim TO, Heo J, Song GA, Cho M. Endoscopic sphincterotomy plus large-balloon dilation versus endoscopic sphincterotomy for removal of bile-duct stones. *Gastrointest Endosc* 2007; **66**: 720-76; quiz 768, 771 [PMID: 17905013 DOI: 10.1016/j.gie.2007.02.033]
- 67 **Attasaranya S**, Cheon YK, Vittal H, Howell DA, Wakelin DE, Cunningham JT, Ajmere N, Ste Marie RW, Bhattacharya K, Gupta K, Freeman ML, Sherman S, McHenry L, Watkins JL, Fogel EL, Schmidt S, Lehman GA. Large-diameter biliary orifice balloon dilation to aid in endoscopic bile duct stone removal: a multicenter series. *Gastrointest Endosc* 2008; **67**: 1046-1052 [PMID: 18178208 DOI: 10.1016/j.gie.2007.08.047]
- 68 **May GR**, Cotton PB, Edmunds SE, Chong W. Removal of stones from the bile duct at ERCP without sphincterotomy. *Gastrointest Endosc* 1993; **39**: 749-754 [PMID: 8293895 DOI: 10.1111/j.1572-0241.2004.30151.x]
- 69 **Lee DK**, Jahng JH. Alternative methods in the endoscopic management of difficult common bile duct stones. *Dig Endosc* 2010; **22** Suppl 1: S79-S84 [PMID: 20590778 DOI: 10.1111/j.1443-1661.2010.00960.x]
- 70 **Yasuda I**, Fujita N, Maguchi H, Hasebe O, Igarashi Y, Murakami A, Mukai H, Fujii T, Yamao K, Maeshiro K, Tada T, Tsujino T, Komatsu Y. Long-term outcomes after endoscopic sphincterotomy versus endoscopic papillary balloon dilation for bile duct stones. *Gastrointest Endosc* 2010; **72**: 1185-1191 [PMID: 20869711]
- 71 **Weinberg BM**, Shindy W, Lo S. Endoscopic balloon sphincter dilation (sphincteroplasty) versus sphincterotomy for common bile duct stones. *Cochrane Database Syst Rev* 2006; **(4)**: CD004890 [PMID: 17054222 DOI: 10.1002/14651858.CD004890.pub2]
- 72 **Baron TH**, Harewood GC. Endoscopic balloon dilation of the biliary sphincter compared to endoscopic biliary sphincterotomy for removal of common bile duct stones during ERCP: a metaanalysis of randomized, controlled trials. *Am J Gastroenterol* 2004; **99**: 1455-1460 [PMID: 15307859]
- 73 **Liao WC**, Lee CT, Chang CY, Leung JW, Chen JH, Tsai MC, Lin JT, Wu MS, Wang HP. Randomized trial of 1-minute versus 5-minute endoscopic balloon dilation for extraction of bile duct stones. *Gastrointest Endosc* 2010; **72**: 1154-1162 [PMID: 20869710 DOI: 10.1016/j.gie.2010.07.009]
- 74 **Han J**, Moon JH, Koo HC, Kang JH, Choi JH, Jeong S, Lee DH, Lee MS, Kim HG. Effect of biliary stenting combined with ursodeoxycholic acid and terpene treatment on retained common bile duct stones in elderly patients: a multicenter study. *Am J Gastroenterol* 2009; **104**: 2418-2421 [PMID: 19568225 DOI: 10.1038/ajg.2009.303]

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Prognostic value of innate and adaptive immunity in colorectal cancer

Fabio Grizzi, Paolo Bianchi, Alberto Malesci, Luigi Laghi

Fabio Grizzi, Paolo Bianchi, Luigi Laghi, Laboratory of Molecular Gastroenterology, Humanitas Clinical and Research Center, 20089 Milan, Italy

Alberto Malesci, Division of Gastroenterology, Humanitas Clinical and Research Center, 20089 Milan, Italy

Alberto Malesci, Department of Translational Medicine, University of Milan, 20089 Milan, Italy

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Correspondence to: Fabio Grizzi, PhD, Laboratory of Molecular Gastroenterology, Humanitas Clinical and Research Center, Via Manzoni 56, Rozzano, 20089 Milan, Italy. fabio.grizzi@humanitasresearch.it

Telephone: +39-2-82245161 Fax: +39-2-82244590

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nogenic and that host immune responses influence survival. Several lines of evidence support the concept that tumor stromal cells, are not merely a scaffold, but rather they influence growth, survival, and invasiveness of cancer cells, dynamically contributing to the tumor microenvironment, together with immune cells. Different types of immune cells infiltrate CRC, comprising cells of both the innate and adaptive immune system. A relevant issue is to unravel the discrepancy between the inhibitory effects on cancer growth exerted by the local immune response and the promoting effects on cancer proliferation, invasion, and dissemination induced by some types of inflammatory cells. Here, we sought to discuss the role played by innate and adaptive immune system in the local progression and metastasis of CRC, and the prognostic information that we can currently understand and exploit.

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Key words: Colorectal cancer; Immunity; Inflammation; Prognosis; Metastasis

Abstract

Colorectal cancer (CRC) remains one of the major public health problems throughout the world. Originally depicted as a multi-step dynamical disease, CRC develops slowly over several years and progresses through cytologically distinct benign and malignant states, from single crypt lesions through adenoma, to malignant carcinoma with the potential for invasion and metastasis. Moving from histological observations since a long time, it has been recognized that inflammation and immunity actively participate in the pathogenesis, surveillance and progression of CRC. The advent of immunohistochemical techniques and of animal models has improved our understanding of the immune dynamical system in CRC. It is well known that immune cells have variable behavior controlled by complex interactions in the tumor microenvironment. Advances in immunology and molecular biology have shown that CRC is immu-

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INTRODUCTION

Despite progresses in our biological and clinical knowledge, colorectal cancer (CRC) remains one of the major public health problems throughout the world^[1]. By its frequency, CRC ranks third in men and women worldwide^[1]. In addition, CRC continues to be one of the most common fatal types of cancer. Originally depicted as a multi-step dynamical disease, CRC develops slowly over several years and progresses through cytologically distinct benign and malignant states, from single crypt lesions through

adenoma, to malignant carcinoma with the potential for invasion and metastasis^[2,3]. According to the theory of multi-step carcinogenesis, colorectal epithelial cells accumulate a number of molecular changes to eventually become fully malignant^[4,5]. In spite of unifying theories, genetic and epigenetic events during the carcinogenesis process differ considerably from tumor to tumor. Thus, CRC is not a single disease; rather it encompasses different molecular and pathological entities with a wide range of clinical behaviors^[6]. At the molecular level, CRC is the tip of an iceberg the basis of which encloses a complex array of gene alterations, affecting supra-molecular processes. Essentially, like individual fingerprints, each tumor arises and behaves in a unique fashion that is unlikely to be exactly recapitulated by any other tumor. Nevertheless, molecular changes allows for a categorization of CRC, which is largely accepted, although likely to oversimplistic. It has been demonstrated that main genetic and epigenetic features, such as microsatellite instability (MSI), chromosomal instability, CpG island methylator phenotype (CIMP) or global DNA hypomethylation, lead to alterations of gene function on a genome-wide scale. It is known that activation of oncogenes, including *KRAS*, *BRAF*, *PIK3CA* and *TP53*, affects intracellular signaling pathways^[6,7]. The suppressor pathway is disrupted in CRC with chromosomal instability occurring in the majority of CRCs (approximately 85%), which have a molecular profile characterized by specific chromosomal amplifications and transformations, aneuploidy, and loss of heterozygosity^[6-8]. Differently, CRCs of the mutator pathway (approximately 15%) have a defective DNA mismatch repair system, which leads to accumulation of thousands of unrepaired mutations^[8]. This inability to repair DNA slippage errors and mismatches can easily be demonstrated because it results in variability in the length of DNA microsatellites, formed by repetitive sequences, that is MSI. It is accepted that MSI CRCs have a heterogeneous histological appearance, better prognosis due to a reduced metastatic potential, and a different response to chemotherapy^[9-14].

Histopathological examination reveals that likely other solid tumors, CRC are associated with diverse immune cell infiltrates^[15-20], and that in the cancer context, epithelial cells coexist with extracellular matrix components and non-neoplastic cell types, including fibroblasts, myofibroblasts, adipocytes, endothelial cells, pericytes, which collectively form the tumor stroma. Several lines of evidence support the concept that tumor stromal cells, are not merely a scaffold, but rather they influence growth, survival, and invasiveness of cancer cells, dynamically contributing to the tumor microenvironment, together with immune cells^[20-25]. The types of immune system cells that are found infiltrating CRC consist of cells of the innate immune system i.e., macrophages, neutrophils, mast cells and natural killer cells, as well as cells associated with an adaptive immune response i.e., T and B lymphocytes. Although it is commonly thought that an immune response localized to the tumor inhibit cancer growth, it is

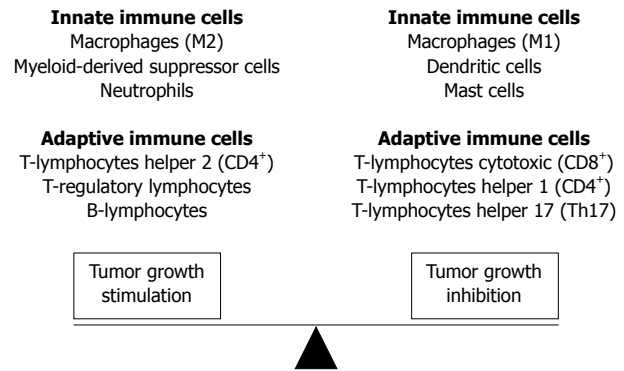


Figure 1 Immune cells have variable behavior controlled by complex interactions in the tumor microenvironment. Although it is commonly thought that an immune response localized to the tumor inhibit cancer growth, it is now clear that some types of tumor-associated inflammation can exert an opposite action.

clear that some types of tumor-associated inflammation may also exert an opposite action, at least at some point of CRC natural history^[26] (Figure 1). Here we sought to briefly review these two contradictory aspects of the immune response to CRC.

INNATE IMMUNITY AND COLORECTAL CANCER

It is well known that innate immunity represents the body's first defense or "gut reaction" to an abnormal situation, such as cancer, and does not involve specific recognition of immunogenic peptides, or antigens^[20]. Aside T and B lymphocytes, innate immune cells orchestrate an inflammatory environment that may function to either stimulate or inhibit cancer growth^[20]. Various innate immune cells have been implicated in CRC development and progression^[15,27,28]. Among these, macrophages, are a primary source of secreted pro-inflammatory cytokines, and are generally distinguished as type 1 (M1) or type 2 (M2)^[29-31]. M1s generally have an interleukin (IL) 12^{low}IL-10^{high} phenotype, show impaired expression of reactive nitrogen intermediates, poor antigen presentation and have tumoricidal capacity, while show high expression of angiogenic factors [including Vascular-endothelial growth factor (VEGF) epidermal-growth factor (EGF) and semaphorin 4D], metalloproteases (MMPs) and cathepsins as well as of the growth arrest-specific protein 6 (GAS6)^[29-32]. Additionally, M1s can support T-helper 1 (Th1) adaptive immunity^[33]. Conversely, M2s secrete immunosuppressive cytokines and promote tumor growth^[22,34]. It has been shown that cancer cells shape their interaction with macrophages by escaping phagocytosis and by promoting a M2-like polarization throughout chemokines and polarizing cytokines including chemokine (C-C motif) ligand 2 (CCL2), colony stimulating factor 1 (CSF1), macrophage slowing factor (MSF), tumor necrosis factor- α (TNF- α , IL-10 and transforming growth factor- β (TGF- β).

Among the cells with M2 phenotype, the tumor-associated macrophages (TAM) have been shown as capable of secreting proteases that enhance invasion and metastases, together with a range of cytokines inhibiting an adaptive tumor-specific immune response, and angiogenic factors that increase neovascularity. In patients with CRC, macrophages are usually found located around necrotic areas of tumor and the advancing tumor margin^[28]. Their role in CRC still remains controversial. While, it was originally thought that the main function of TAMs was a direct cytotoxic effects on tumor cells, phagocytosis apoptotic/necrotic cell debris, and present tumor-associated antigens to T lymphocytes, current evidences suggest that inflammation and TAMs can also promote tumor growth and metastasis. Kang *et al*^[35] have recently highlighted an association between intra-tumoral TAM densities with CRC malignant aggressiveness. Additionally, increased frequencies of intra-tumoral TAM have been associated with high levels of MMP type 2 and 9 expression in CRC cells^[36,37]. These findings are in accord with a previous cell-line study showing that co-culturing of tumor cells with macrophages enhances cancer cell migration, invasiveness, and MMP-2 and MMP-9 secretion^[35].

Kaler *et al*^[38] have recently established that macrophages promote Wnt signaling pathway in CRC cells and thus enhance their proliferation, and demonstrated that macrophages exert their pro-tumorigenic activity mainly through the release of IL-1 β . The same authors demonstrated that tumor necrosis factor related apoptosis inducing ligand (TRAIL) induced apoptosis of CRC cells is inhibited by macrophage derived IL-1 β , and showed that macrophages and recombinant IL-1 β counteract TRAIL-induced apoptosis through activation of Wnt signaling and stabilization of the nuclear transcription factor Snail in tumor cells.

A number of studies have also shown that macrophages can release a vast diversity of cytokines, proteolytic enzymes, growth factors, and inflammatory mediators that may directly influence and stimulate the growth and migration of tumor cells^[29-33]. Li *et al*^[39] first reported that IL-6 released by macrophage directly promotes CRC cell progression. They showed that monocyte/macrophage-derived IL-6 enhances migration of HT-29 CRC cells *in vitro*^[39]. Although, how IL-6 enhances HT-29 cell migration still remains unknown, it has been suggested that TGF- β may be involved in this process. Using monoclonal antibodies to neutralize IL-10 in macrophage supernatant, Li *et al*^[39] found that the IL-6-mediated effects on HT-29 CRC cells were all enhanced. Therefore, the interaction between IL-6 and IL-10 released from macrophage is indeed involved in CRC progression and prognosis. The above findings support the fact that TAMs play a regulatory role in the tumor microenvironment by modulating secretion of cytokines such as IL-6 and IL-10, thereby causing cancer cells to manipulate their microenvironment to facilitate cancer growth^[40,41].

TAM activation and maturation is under the influence

of the tumor microenvironment. TAMs retain a relatively immature phenotype, characterized by a low expression of differentiation-associated antigens in hypoxic microenvironments^[35,42,43]. Conversely to an M2 phenotype with pro-tumoral functions in CRC^[35], it has been shown that the macrophages, especially those secreting IL-12 and IL-23, infiltrating the tumor front are positively correlated with a favorable outcome, which implicates TAMs as possessing anti-tumor functions. Forssell *et al*^[44] stained with the pan-monocyte/macrophage marker CD68 a series of CRC specimens and showed that the higher macrophage infiltration along the tumor invasive front correlated with improved survival in colon cancer compared to rectal cancer. They concluded that a dense macrophage infiltration at the tumor front positively influences prognosis in colon cancer and that the degree of cell-to-cell contact may influence the balance between pro-tumorigenic and anti-tumorigenic properties of macrophages. High levels of tissue macrophages have been also associated with earlier disease stage, absence of nodal and lymphovascular metastases and an overall better prognosis. Zhou *et al*^[45] by analyzing the relationship between the density of TAMs and the potential of hepatic metastasis and survival have shown that a higher density of macrophages along the invasive front of CRC was associated with a higher 5-year survival rate. In addition, according to Forssell's scoring system that defines CD68 hot-spots as small areas among which the infiltration of macrophages was considerably above the average level of CD68-positive cells, the highest CD68 hot-spot was associated with both the incidence of hepatic metastasis and the interval between colon resection and the occurrence of hepatic metastasis^[45].

The mechanisms behind the anti-tumor effects of TAMs have still not been fully elucidated, and seem potentially be ascribed to the M1 phenotype, which is in part controlled by the CD4⁺T lymphocytes and the death of cancer cells^[45-47]. It has been ascertained that recruitment of TAMs contributes to the development of an adaptive immune response against cancer, and the balance between antigen availability and clearance through phagocytosis and subsequent degradation of senescent or apoptotic cells.

It is undeniable that the discrepancies in results between different studies may be due to a number of factors related to the location of the TAMs and the assessment methods employed. Recent studies have reported that different macrophage phenotypes localized to different regions of the carcinoma have variable effects on tumor cells^[18,19]. Furthermore, evidences have shown that the relationship between TAMs and tumor progression is tumor type-dependent.

Other innate immune system cells can be detected in CRC microenvironment, including mast cells (MCs), neutrophils, natural killer (NK) cells, and eosinophils.

Nagtegaal *et al*^[48] have shown that peritumoral MCs prevent local and distant recurrence, with improved survival as a consequence. The significant benefit of MCs on

tumor progression in CRC was also highlighted by Gounaris *et al.*^[49] who reported that depletion of MCs, either by drugs either in MC-deficient mice, led to remission of existing polyps. Similar to other immune cell types, high numbers of MCs are associated with earlier CRC disease stage and have been proposed as an independent prognostic marker for improved survival^[50]. It has also been reported that the number of MCs progressively decreases from normal mucosa through premalignant conditions and the lowest numbers are seen in cancers. Because of their location, MCs may prevent the metastasis of carcinomas that are restricted to the sub-mucosa. *In vivo* studies have observed that destruction of lymphatic vessels in the peritumoral infiltrate is always accompanied by MCs. This led to the hypothesis that MCs degranulate in the sub-mucosa when they come into contact with the inflammatory infiltrate or CRC cells, thus leading to the destruction of lymphatic vessels and thereby preventing further metastasis. Gulubova and Vlaykova^[51] proposed the MCs density along the invasive front of the primary CRC as a helpful tool for prognosis of patients after surgical therapy. In their study, it has been shown that patients with low MCs density had significantly better prognosis compared to those with high MCs density. In addition, Blatner *et al.*^[52] reported that in CRC, MCs contribute to systemic regulatory T-cell dysfunction. MCs have an intricate interaction with T-regulatory cells that controls the functions of both cell types in a reciprocal manner. MCs play also an important role in allograft acceptance, where they are required to sustain the peripheral tolerance mediated by T-regulatory cells. These latter can inhibit MCs differentiation and hinder degranulation by contact-dependent mechanisms and production of soluble factors, such as IL-10. Conversely, the activation and subsequent degranulation of MCs breaks peripheral tolerance. MCs degranulation or direct cell contact and secretion of IL-6 promote Th-17 conversion of T-regulatory cells with loss of both forkhead box P3 transcription factor (Foxp3) expression and T-cell-suppressive properties.

Neutrophils may form up to 15% of the inflammatory infiltrate associated with CRCs and this proportion increases within areas of tumor necrosis^[28]. In patients with rectal cancer, high concentrations of neutrophils have been shown as independent predictors of improved prognosis especially when microscopic abscesses form^[28]. However, an elevated neutrophil/lymphocyte ratio was, however, found by Halazun *et al.*^[53] led to a poorer survival time and higher rate of recurrence in CRC patients undergoing surgery for liver metastasis.

NK cells are granular lymphocytes that form part of the innate cellular immune response^[28]. In CRC, high numbers of NK cells in the inflammatory infiltrate has been associated with better prognosis^[28]. The number of NK cells decreases with increasing cancer stage. Additionally, it has been shown that the ratio of NK cells in the peripheral blood is an important prognostic indicator in CRC patients and it is of interest to note that 5-fluoro-

uracil-based chemotherapy increases the number of NK cells^[54,55].

Aside the above innate immune cell types, dendritic cells (DCs), antigen-presenting cells that are critical to the stimulation of effective anti-tumor adaptive immune responses, can become defective in the tumor microenvironment and aid in tumor immune evasion by failing to stimulate T lymphocytes. It has been suggested that the presence of DCs may be of significant benefit in patients with CRC. Xie *et al.*^[56] also demonstrated that the presence of DCs was found predominantly in early compared to later disease stages and mostly located in tumor surrounding tissue. Suzuki *et al.*^[57] showed the presence of mature CD83⁺ DCs at the cancer invasive front and by light and electron microscopy demonstrated their aggregation into clusters with lymphocytes, the majority of which were CD45RO⁺ T lymphocytes. They concluded that mature CD83⁺ DCs at the invasive margin promote T-cell activation for the generation of tumor specific immunity.

Although, there is growing evidence that the host innate immune system has a critical role in regulating carcinogenesis, the specific receptors involved and the importance of their interaction with commensal bacteria remain to be elucidated. Two major classes of innate immune receptors, the Toll-like receptors and Nod-like receptors, many of which are upstream of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), have been investigated^[58,59]. Particularly, the toll-like receptors have been implicated in promoting colon tumorigenesis. Fukata *et al.*^[60] have shown that toll-like receptor-4 (TLR4) is over-expressed in human colitis-associated cancer and, that mice deficient in TLR4 are markedly protected against colitis-associated neoplasia. A high TLR4 expression in the tumor microenvironment has recently been reported as a possible marker of disease progression in CRC^[60]. Conversely, the role of nod-like receptors in regulating colorectal tumorigenesis remains unclear. Chen *et al.*^[61] reported an increased intestinal permeability associated with enhanced inflammatory cytokine production and epithelial cell proliferation in Nod1-deficient mice. As the depletion of the gut microbiota suppressed tumor development in Nod1-deficient mice, a link should exist between commensal bacteria and host innate Nod1 signaling pathway involved in the development of inflammation-mediated CRC.

ADAPTIVE IMMUNITY AND COLORECTAL CANCER

It is today well recognized that cells of the adaptive immune system are recruited in CRC and colitis-associated tumours (CAC), where they have either pro- and anti-tumorigenic roles. T lymphocytes participate in inflammation, cancer development and progression, as well as in anticancer immunity^[20-22]. In CAC the adaptive immune system seems to have mainly a pro-tumorigenic role, while in CRC it may play a double-faced role, being the

balance between immune-surveillance (carried out by CD8⁺ and CD4⁺ T lymphocytes) and tumour-promoting inflammation (by various sub-types of T lymphocytes) to change over time, and eventually dictating disease progression. It has been suggested that immune-surveillance might mediate the early recognition and elimination of transformed cells and aberrant crypt foci, keeping small tumours in a dormant state^[62]. Immune-surveillance is retained important during the metastatic process, when small numbers or isolated tumoral cells travel and can be attacked by antitumor immune cells not inhibited by factors in the tumour microenvironment. Galon *et al*^[18] suggested that once human CRC become clinically detectable, the adaptive immune response plays a role in preventing tumour recurrence and metastasis. Independently, Chiba *et al*^[63] and Banerjee *et al*^[64] reported that intra-tumoral T cells modify the tumour stroma or CRC cells in a way that attenuate the metastatic potential.

Cytotoxic T lymphocytes (CD8⁺ T cells, or CTL) constitute one of the leading effector of antitumor immunity. In order for CD8⁺ T cells to recognize antigens, these need to be exposed on the tumour cells in association with the human leukocyte antigen (HLA) class I proteins^[65]. Upon encounter of a tumour cell antigen/HLA I complex for which their T cell receptor (TCR) is specific, CD8⁺ T lymphocytes clonally expand and differentiate^[65]. Once activated, cytotoxic T lymphocytes can mediate specific destruction of tumour cells through the release of lytic components *via* cell-cell interaction^[65,66]. Perforin, a cytolytic protein found in the granules of CD8 T-cells and NK cells, and enzymatic proteases, including granzyme B, are secreted determining cell death by disruption of the cell membrane and activation of the apoptotic pathway respectively. CD4⁺ T cells, which only respond to antigens presented by the HLA class II proteins expressed by DCs, are important for antitumor immunity. On the peculiar cytokine profile induced, CD4⁺ T lymphocytes are mainly subdivided in Th1 or Th2 lymphocytes^[67]. Th1 cells secrete cytokines such as interferon-gamma (IFN- γ) and TNF- α , and support cytotoxic T lymphocytes by producing IL-2, required for CD8⁺ T cells proliferation. Conversely, Th2 cells principally secrete IL-10, IL-4, and IL-5, and limit cytotoxic T lymphocytes proliferation.

Regulatory T cells (Treg cells) have been defined as a T-cell population that functionally suppresses an immune response by influencing the activity of another cell type^[68]. Treg cells have been categorized into two main classes based on their ontogeny: naturally occurring Treg (nTreg), which develop in the thymus and are present in mice and healthy humans from an early postnatal period, and Treg which can arise in the periphery (or *in vitro*)^[69]. nTreg are characterized by their high expression of CD25 (CD4⁺CD25⁺) and co-expression of the FOXP3^[69]. Needham *et al*^[70] reported that depletion of intra-tumoral Tregs enhances antitumor immunity and tumour rejection in mouse models. Clarke *et al*^[71] have shown that depletion of Tregs in the peripheral blood of patients

with CRC was recently shown to unmask CD4⁺ T-cell responses to tumour antigens. It is known that self-tumour antigens may also induce the preferential proliferation of CD4⁺CD25⁺FOXP3⁺ Treg cells. Tregs function to maintain immune homeostasis and limit acute inflammation. These cells interfere with T cell priming and can affect the antitumor function of effector cells *via* secretion of TGF- β and IL-10. A number of investigations suggest that Tregs infiltrating the tumour may adversely affect prognosis^[72-75]. Increased levels of Treg cells have been, however, associated with a favourable prognosis in CRC^[76-79]. The mechanisms of this dual effect, which seems dependent on the tissue type, are not well understood, although it is hypothesized that Tregs might suppress inflammation induced by growth promoting innate immune system cells.

Although the role of B lymphocytes in cancer has been overshadowed by the interest in developing T-cell-mediated cellular responses, it is now apparent that B lymphocytes can play a complementary role in the host response against tumour. B lymphocytes represent a cell population that express clonally diverse cell surface Ig receptors recognizing specific antigenic epitopes. In addition to the role of B lymphocytes in antibody production, these cells mediate/regulate several other functions fundamental for immune homeostasis. Of significant importance is the antigen-presenting role of B lymphocytes in the initiation of T-cell immune responses. Moreover, B lymphocytes can play a significant role in infection and autoimmunity as regulatory cells (indicated as Bregs) *via* the elaboration of suppressive cytokines, such as IL-10, TGF- β , or IL-4. The role played by B cells in cancer immunology remains still complex and somewhat controversial. Depending upon their state of activation, B lymphocytes have had divergent roles on T-cell differentiation and effector function. Oversimplifying, resting B lymphocytes have been reported to suppress T-cell-mediated antitumor immunity, by acting on both CD4⁺ and CD8⁺ T lymphocytes. In contrast, a number of reports suggest the efficacy of activated B lymphocytes in cellular immunotherapy of malignancies. In particular, activated B lymphocytes have been reported to enhance the ability to generate tumour-infiltrating lymphocytes *in vitro* involving anti-CD3 and IL-2.

The therapeutic targeting of tumours or components of the immune system with molecule-specific monoclonal antibodies (mAb) is now considered a viable treatment option for cancer patients. One of the currently applied antibodies in clinics is represented by rituximab (Rituxan) that targets B cells for elimination by binding the B cell-associated marker CD20. Interestingly, it has been recently developed a C57BL/6 TRAIL-sensitive tumour model with the aim of being able to use gene-targeted mice to better evaluate the innate and adaptive immune cells contributing to the tumoricidal activity of the MD5-1 mAb in more clinically relevant established tumours. C57BL/6 gene-targeted or immune cell-depleted mice were used to examine the antitumor activity of

MD5-1 against the TRAIL-sensitive mouse MC38 colon adenocarcinoma. It has been shown that an intact B cell compartment is critical for the therapeutic activity of MD5-1 against established tumours. B cells were confirmed to trigger tumour cell apoptosis by FcR-mediated cross-linking of the MD5-1 mAb *in vitro* and *in vivo* B lymphocytes were critical for directly triggering MD5-1-mediated tumour cell apoptosis.

Although the role of B-cells in human CRCs is still not completely characterized, B-cell-deficient mice exhibit spontaneous regression of MC38 colon carcinoma cells. Studies involving BCR transgenic mice indicated that B lymphocytes might inhibit antitumor T lymphocytes responses by antigen-nonspecific mechanisms. Shah *et al* investigated the role of B lymphocytes in tumour immunity by studying immune responses of mice genetically lacking B lymphocytes to primary tumours. They highlight that although the effects of B lymphocytes on anti-tumour response warrant further study, adoptive transfer of CD40(-/-) B lymphocytes into B lymphocytes-deficient mice resulted in restored growth of MC38 colon carcinoma cells suggesting additional factors other than CD40 are involved in dampening anti-tumour responses.

PROGNOSTIC INFORMATION OF IMMUNE CELL INFILTRATE

In contrast to infiltration of cells responsible for chronic inflammation, the presence of high numbers of T lymphocytes has been reported to be a positive prognostic factor in several cancers. The first reports on the beneficial effect of lymphocytic infiltration in CRC appeared already in the Eighties. They were later confirmed until recent most studies highlighting a prominent function for memory T lymphocytes and CD8⁺ T lymphocytes in predicting disease-free survival and overall survival. In general terms, it has been suggested that prognosis in patients with cancer is positively affected by (1) the presence of a tumour gene signature consistent with a type I adaptive immune response (i.e., increased antigen presentation, IFN- γ signalling, and TCR signalling); and (2) the presence of T cells that penetrate through tumour stroma and deeply infiltrate the parenchyma to become intra-tumoral T cells^[20]. Thus, besides a Th-1 response signature, the other key feature of an effective immune response is the ability of T cells to reach the site of the tumour and to infiltrate it. Because T-cell infiltration is not spatially homogeneous in CRC, attention has been focused on the predictive values of T lymphocytes located in the center of the tumor (CT), along the invasive margin (IM) and in lymphoid aggregate mainly detectable in proximity of the tumor (these aggregates are called tertiary lymphoid structures)^[80,81].

In a large series of CRCs, Pagès *et al*^[82] assessed the immune component of the tumoral microenvironment by a combination of high-throughput gene expression and immunophenotypic analyses, and evaluated its possible influence on tumour dissemination. They found

an association between evidence of an immune reaction within the tumour and the absence of tumour local invasion of vascular, lymphatic, and neural structures (collectively referred to as VELIPI). CRCs without VELIPI were associated with an enhanced immune cell infiltration and an increase of mRNA expression of adaptive Th1 effector T-cell markers [CD8, T-box transcription factor 21 (T-bet), IFN regulatory factor-1 (IRF-1), IFN- γ , granzyme B, and granzyme B]. The immunohistochemical analysis of adaptive immune markers i.e., CD3, CD8, granzyme B, and CD45RO, by tissue microarrays prepared from the CT and from its IM revealed a statistically significant correlation between the density of these immune cells and outcome for all patients but those with metastatic disease at diagnosis. Further, the combined analysis of both tumour regions improved the accuracy of survival prediction compared with single-region analysis^[18].

Independently, Deschoolmeester *et al*^[83] showed that the presence of a pronounced lymphocytic infiltration within the tumour is associated with improved survival. They found that CD3⁺ and CD8⁺ T lymphocytes within tumour nests and of CD3⁺ in the stroma had a major impact on the patients' overall survival. The improved survival associated with infiltration of T lymphocytes has been suggested to result from the effective suppression of micrometastases. Therefore, the densities of CD8⁺ T cells within the primary tumour might be a potential marker of the presence of a systemic immunosurveillance mechanism. In addition, tumour cells secrete substances in the stromal compartment, which might be recognized by the immune system that subsequently attack the tumour. A weak adaptive immune reaction correlated with a very poor prognosis even in patients with early tumour invasion. Conversely, a high density of adaptive immune cells correlated with a highly favourable prognosis whatever the local extent of the tumour and the regional lymph node invasion.

In mice, targeted disruptions of genes that encode critical components of the immune system (i.e., mice lacking: IFN- γ or its receptor, signal transducer and activator of transcription-1 mediating IFN- γ receptor signalling, perforin, recombination activating gene-2, or IL-12) induce an increased susceptibility of the host to tumours^[84]. These findings make it possible to hypothesize an immune-mediated control of tumour development by the adaptive compartment.

A number of studies have reported that MSI, CIMP, BRAF mutation, PIK3CA mutation, and tumour LINE-1 hypomethylation are associated with CRC prognosis and that lymphocytic infiltration is associated with many of these molecular variables. The associations of a prognostic biomarker with a given disease, strongly suggests its stage-dependency as outcome predictor. This is best exemplified by MSI CRC, whose overall prognostic advantage is associated with a low frequency of stage III and IV cases at diagnosis as compared to microsatellite stable counterpart. Most MSI CRCs show a pronounced

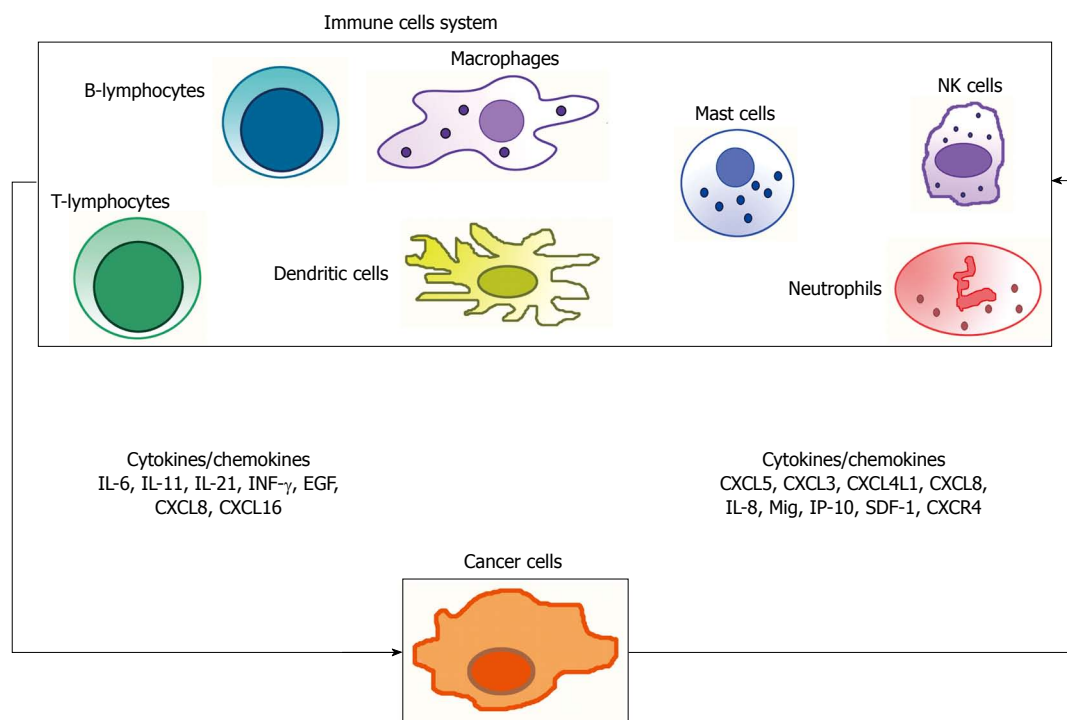


Figure 2 Anti- and pro-tumor immune response involves the interaction of several cell types of the adaptive as well as of the innate immune system, and an intricate network of products (i.e., cytokines and chemokines). IL: Interleukin; INF- γ : Intracellular interferon- γ ; EGF: Epidermal-growth factor; NK cells: Natural killer cells.

intra-tumoral inflammatory reaction (in fact a criterion for MSI testing), the mechanistic explanation of which, however, is still incompletely understood. Within these tumours, infiltrating lymphocytes have been identified as predominantly activated CD8⁺ T cells. The presence of these cytotoxic T lymphocytes has been attributed to the inherently greater production of abnormal peptides as a result of unreliable DNA repair in MSI-positive tumours. It is known that truncated peptides produced by frameshift mutations due to MSI may be immunogenic and contribute to the host immune response. However, little is still known about the interrelationship between tumour-infiltrating T lymphocytes, MSI status, and other tumour molecular features. It is indubitable that to define the prognostic effect of tumour-infiltrating T-cells independently of those potential confounders, large studies of CRC with extensive molecular characterization are needed. Additionally, caution is needed before incorporating tumour-infiltrating T cells into tumour staging. To minimise the risk of inappropriate tumour down-staging at diagnosis, survival data need to be confirmed in independent series of patients studied in the past decade. Moreover, the association has to be conclusively proven between low densities of tumour-infiltrating T cells and the clinical detection of metachronous metastases, which remains the most appropriate outcome measure for recognising a role of the local immune response in micrometastasis suppression. Recently, Laghi *et al*^[85] investigated the relationship between the density of CD3⁺ T infiltrating lymphocytes along the tumour invasive margin, and the occurrence of metachronous distant-organ

metastases after potentially curative resection, in a large, consecutive series of patients with deeply invading (pT3 or pT4) MSI-typed CRC, and no evidence of distant organ metastasis at diagnosis. They found that large areas of CD3⁺ cells at the invasive front of pT3 or pT4 CRCs are associated with a low risk of metachronous metastasis and consequently a survival advantage, only in patients with node-negative cancers, but not in patients whose cancers involved lymphnodes. Of interest, the prognostic advantage conferred by a high density of CD3⁺ cells was independent of tumour microsatellite status in patients with stage II CRC. CD3-immunostaining of CRC tissue might therefore be useful for selecting stage II patients who, because they are at very low risk for cancer progression, could be spared adjuvant treatments.

Nosho *et al*^[86] examined the prognostic role of tumour-infiltrating T-cell subsets in a database of 768 CRCs from two prospective cohort studies. They concurrently assessed the densities of CD3⁺, CD8⁺, CD45RO⁺, and FOXP3⁺ lymphocytes as well as other relevant molecular and pathological features, therefore making possible to evaluate the independent effect of each T-cell subset density on patient survival. They found that the density of CD45RO⁺ cells, but not that of CD3⁺, CD8⁺, or FOXP3⁺ cells, was an independent prognostic biomarker of longer survival in CRC patients. In contrast, Salama *et al*^[76] by analysing T-cell infiltrates in 967 CRCs including 593 stage II and 374 stage III cases, reported that FOXP3⁺ lymphocytes density had stronger prognostic significance than CD8⁺ and CD45RO⁺ cells, and predicted a better outcome. FOXP3⁺ lymphocytes were

found not associated with any histopathologic features. At multivariate analysis, stage, vascular invasion, and FOXP3⁺ cell density in tumoral tissue were independent prognostic indicators. These results led Salama *et al*^[76] to conclude that the inclusion of FOXP3⁺ cell density may help to improve the prognostication of early-stage CRC, although these authors do not explored this parameter with tumor stage.

CONCLUSION

It is accepted that human cancer is a complex dynamical disease^[87,88]. It is also now well recognized that cancers are not just composed of malignant cells, but that they are microenvironment consisting of many cell types, including a range of immune cells. It is indubitable that the antitumor immune response involves the interaction of several cell types and products (Figure 2), of the adaptive as well as of the innate immune system. It is also clear that CRC can escape immune surveillance using several strategies. The molecular profiles of the function and interaction of innate and adaptive immune cells, and the definition of tumor antigens have all led to build the basis of the knowledge of how the immune system modulates tumor growth and inhibition. The challenge remains to determine not only how the “rejection” pathway initiates in human malignancy, but also how that rejection is maintained. It is indubitable that the analysis of the type, quantity, location and the functions of the immune infiltrate becomes a primary step in understanding CRC natural history, and, in a clinical perspective, its prognostic determinants. A comprehensive analysis of all components of the lymphocytic infiltrates in the context of their localization, organization and impact at various steps of tumor progression remains largely, if not entirely, to be reported to prospective studies. In parallel, understanding the mechanisms of efficient immune reactions, the place where they are initiated, the cells and key cytokines and chemokines involved (Figure 2), and their impact at different stages of the disease should provide new tools and goals for more effective and less toxic targeted therapies.

REFERENCES

- Deschoolmeester V, Baay M, Specenier P, Lardon F, Vermorken JB. A review of the most promising biomarkers in colorectal cancer: one step closer to targeted therapy. *Oncologist* 2010; **15**: 699-731 [PMID: 20584808 DOI: 10.1634/theoncologist.2010-0025]
- Michor F, Iwasa Y, Lengauer C, Nowak MA. Dynamics of colorectal cancer. *Semin Cancer Biol* 2005; **15**: 484-493 [PMID: 16055342 DOI: 10.1016/j.semcancer.2005.06.005]
- Jass JR. Colorectal cancer: a multipathway disease. *Crit Rev Oncog* 2006; **12**: 273-287 [PMID: 17425506]
- Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; **61**: 759-767 [PMID: 2188735 DOI: 10.1016/0092-8674(90)90186-I]
- Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM, Bos JL. Genetic alterations during colorectal-tumor development. *N Engl J Med* 1988; **319**: 525-532 [PMID: 2841597 DOI: 10.1056/NEJM198809013190901]
- Ngino S, Chan AT, Fuchs CS, Giovannucci E. Molecular pathological epidemiology of colorectal neoplasia: an emerging transdisciplinary and interdisciplinary field. *Gut* 2011; **60**: 397-411 [PMID: 21036793 DOI: 10.1136/gut.2010.217182]
- Pino MS, Chung DC. The chromosomal instability pathway in colon cancer. *Gastroenterology* 2010; **138**: 2059-2072 [PMID: 20420946 DOI: 10.1053/j.gastro.2009.12.065]
- Boland CR, Goel A. Microsatellite instability in colorectal cancer. *Gastroenterology* 2010; **138**: 2073-2087.e3 [PMID: 20420947 DOI: 10.1053/j.gastro.2009.12.064]
- Imai K, Yamamoto H. Carcinogenesis and microsatellite instability: the interrelationship between genetics and epigenetics. *Carcinogenesis* 2008; **29**: 673-680 [PMID: 17942460 DOI: 10.1093/carcin/bgm228]
- Hyde A, Fontaine D, Stuckless S, Green R, Pollett A, Simms M, Sipahimalani P, Parfrey P, Younghusband B. A histology-based model for predicting microsatellite instability in colorectal cancers. *Am J Surg Pathol* 2010; **34**: 1820-1829 [PMID: 21107088 DOI: 10.1097/PAS.0b013e3181f6a912]
- Boland CR, Thibodeau SN, Hamilton SR, Sidransky D, Eshleman JR, Burt RW, Meltzer SJ, Rodriguez-Bigas MA, Fodde R, Ranzani GN, Srivastava S. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res* 1998; **58**: 5248-5257 [PMID: 9823339]
- Malesci A, Laghi L, Bianchi P, Delconte G, Randolph A, Torri V, Carnaghi C, Doci R, Rosati R, Montorsi M, Roncalli M, Gennari L, Santoro A. Reduced likelihood of metastases in patients with microsatellite-unstable colorectal cancer. *Clin Cancer Res* 2007; **13**: 3831-3839 [PMID: 17606714 DOI: 10.1158/1078-0432.CCR-07-0366]
- Gryfe R, Kim H, Hsieh ET, Aronson MD, Holowaty EJ, Bull SB, Redston M, Gallinger S. Tumor microsatellite instability and clinical outcome in young patients with colorectal cancer. *N Engl J Med* 2000; **342**: 69-77 [PMID: 10631274 DOI: 10.1056/NEJM200001133420201]
- Carethers JM, Smith EJ, Behling CA, Nguyen L, Tajima A, Doctolero RT, Cabrera BL, Goel A, Arnold CA, Miyai K, Boland CR. Use of 5-fluorouracil and survival in patients with microsatellite-unstable colorectal cancer. *Gastroenterology* 2004; **126**: 394-401 [PMID: 14762775 DOI: 10.1053/j.gastro.2003.12.023]
- McLean MH, Murray GI, Stewart KN, Norrie G, Mayer C, Hold GL, Thomson J, Fyfe N, Hope M, Mowat NA, Drew JE, El-Omar EM. The inflammatory microenvironment in colorectal neoplasia. *PLoS One* 2011; **6**: e15366 [PMID: 21249124 DOI: 10.1371/journal.pone.0015366]
- Peddareddigari VG, Wang D, Dubois RN. The tumor microenvironment in colorectal carcinogenesis. *Cancer Microenviron* 2010; **3**: 149-166 [PMID: 21209781 DOI: 10.1007/s12307-010-0038-3]
- Halama N, Michel S, Kloor M, Zoernig I, Pommerenke T, von Knebel Doeberitz M, Schirmacher P, Weitz J, Grabe N, Jäger D. The localization and density of immune cells in primary tumors of human metastatic colorectal cancer shows an association with response to chemotherapy. *Cancer Immun* 2009; **9**: 1 [PMID: 19226101]
- Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P, Cugnenc PH, Trajanoski Z, Fridman WH, Pagès F. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* 2006; **313**: 1960-1964 [PMID: 17008531 DOI: 10.1126/science.1129139]
- Galon J, Fridman WH, Pagès F. The adaptive immunologic microenvironment in colorectal cancer: a novel perspective. *Cancer Res* 2007; **67**: 1883-1886 [PMID: 17332313 DOI: 10.1158/0008-5472.CCR-07-0366]

- 10.1158/0008-5472.CAN-06-4806]
- 20 **Disis ML.** Immune regulation of cancer. *J Clin Oncol* 2010; **28**: 4531-4538 [PMID: 20516428 DOI: 10.1200/JCO.2009.27.2146]
- 21 **Mantovani A,** Romero P, Palucka AK, Marincola FM. Tumour immunity: effector response to tumour and role of the microenvironment. *Lancet* 2008; **371**: 771-783 [PMID: 18275997 DOI: 10.1016/S0140-6736(08)60241-X]
- 22 **Mantovani A,** Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* 2008; **454**: 436-444 [PMID: 18650914 DOI: 10.1038/nature07205]
- 23 **Bissell MJ,** Radisky D. Putting tumours in context. *Nat Rev Cancer* 2001; **1**: 46-54 [PMID: 11900251 DOI: 10.1038/35094059]
- 24 **Mueller MM,** Fusenig NE. Friends or foes - bipolar effects of the tumour stroma in cancer. *Nat Rev Cancer* 2004; **4**: 839-849 [PMID: 15516957 DOI: 10.1038/nrc1477]
- 25 **Bhowmick NA,** Neilson EG, Moses HL. Stromal fibroblasts in cancer initiation and progression. *Nature* 2004; **432**: 332-337 [PMID: 15549095 DOI: 10.1038/nature03096]
- 26 **Ferrone C,** Dranoff G. Dual roles for immunity in gastrointestinal cancers. *J Clin Oncol* 2010; **28**: 4045-4051 [PMID: 20644090 DOI: 10.1200/JCO.2010.27.9992]
- 27 **Saleh M,** Trinchieri G. Innate immune mechanisms of colitis and colitis-associated colorectal cancer. *Nat Rev Immunol* 2011; **11**: 9-20 [PMID: 21151034 DOI: 10.1038/nri2891]
- 28 **Salama P,** Platell C. Host response to colorectal cancer. *ANZ J Surg* 2008; **78**: 745-753 [PMID: 18844901 DOI: 10.1111/j.1445-2197.2008.04642.x]
- 29 **Biswas SK,** Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol* 2010; **11**: 889-896 [PMID: 20856220 DOI: 10.1038/ni.1937]
- 30 **Mantovani A,** Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol* 2010; **22**: 231-237 [PMID: 20144856 DOI: 10.1016/j.coi.2010.01.009]
- 31 **Sica A,** Larghi P, Mancino A, Rubino L, Porta C, Totaro MG, Rimoldi M, Biswas SK, Allavena P, Mantovani A. Macrophage polarization in tumour progression. *Semin Cancer Biol* 2008; **18**: 349-355 [PMID: 18467122 DOI: 10.1016/j.semcancer.2008.03.004]
- 32 **Martinez FO,** Gordon S, Locati M, Mantovani A. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *J Immunol* 2006; **177**: 7303-7311 [PMID: 17082649]
- 33 **Mantovani A,** Sica A, Locati M. New vistas on macrophage differentiation and activation. *Eur J Immunol* 2007; **37**: 14-16 [PMID: 17183610 DOI: 10.1002/eji.200636910]
- 34 **Rigo A,** Gottardi M, Zamò A, Mauri P, Bonifacio M, Krampfer M, Damiani E, Pizzolo G, Vinante F. Macrophages may promote cancer growth via a GM-CSF/HB-EGF paracrine loop that is enhanced by CXCL12. *Mol Cancer* 2010; **9**: 273 [PMID: 20946648 DOI: 10.1186/1476-4598-9-273].]
- 35 **Kang JC,** Chen JS, Lee CH, Chang JJ, Shieh YS. Intratumoral macrophage counts correlate with tumor progression in colorectal cancer. *J Surg Oncol* 2010; **102**: 242-248 [PMID: 20740582 DOI: 10.1002/jso.21617]
- 36 **Papadopoulos S,** Scorilas A, Arnogianaki N, Papapanayiotou B, Tzimogianni A, Agnantis N, Talieri M. Expression of gelatinase-A (MMP-2) in human colon cancer and normal colon mucosa. *Tumour Biol* 2001; **22**: 383-389 [PMID: 11786732 DOI: 10.1159/000050641]
- 37 **Herszényi L,** Sipos F, Galamb O, Solymosi N, Hritz I, Miheller P, Berczi L, Molnár B, Tulassay Z. Matrix metalloproteinase-9 expression in the normal mucosa-adenoma-dysplasia-adenocarcinoma sequence of the colon. *Pathol Oncol Res* 2008; **14**: 31-37 [PMID: 18347934 DOI: 10.1007/s12253-008-9004-5]
- 38 **Kaler P,** Galea V, Augenlicht L, Klampfer L. Tumor associated macrophages protect colon cancer cells from TRAIL-induced apoptosis through IL-1beta-dependent stabilization of Snail in tumor cells. *PLoS One* 2010; **5**: e11700 [PMID: 20661477 DOI: 10.1371/journal.pone.0011700]
- 39 **Li YY,** Hsieh LL, Tang RP, Liao SK, Yeh KY. Interleukin-6 (IL-6) released by macrophages induces IL-6 secretion in the human colon cancer HT-29 cell line. *Hum Immunol* 2009; **70**: 151-158 [PMID: 19272324 DOI: 10.1016/j.humimm.2009.01.004]
- 40 **Herbeuval JP,** Lelievre E, Lambert C, Dy M, Genin C. Recruitment of STAT3 for production of IL-10 by colon carcinoma cells induced by macrophage-derived IL-6. *J Immunol* 2004; **172**: 4630-4636 [PMID: 15034082]
- 41 **Li YY,** Chang JW, Hsieh LL, Yeh KY. Neutralization of interleukin (IL)-10 released by monocytes/macrophages enhances the up-regulatory effect of monocyte/macrophage-derived IL-6 on expressions of IL-6 and MUC1, and migration in HT-29 colon cancer cells. *Cell Immunol* 2010; **265**: 164-171 [PMID: 20851386 DOI: 10.1016/j.cellimm.2010.07.014]
- 42 **Lewis C,** Murdoch C. Macrophage responses to hypoxia: implications for tumor progression and anti-cancer therapies. *Am J Pathol* 2005; **167**: 627-635 [PMID: 16127144 DOI: 10.1016/S0002-9440(10)62038-X]
- 43 **Mantovani A,** Schioppa T, Porta C, Allavena P, Sica A. Role of tumor-associated macrophages in tumor progression and invasion. *Cancer Metastasis Rev* 2006; **25**: 315-322 [PMID: 16967326 DOI: 10.1007/s10555-006-9001-7]
- 44 **Forssell J,** Oberg A, Henriksson ML, Stenling R, Jung A, Palmqvist R. High macrophage infiltration along the tumor front correlates with improved survival in colon cancer. *Clin Cancer Res* 2007; **13**: 1472-1479 [PMID: 17332291 DOI: 10.1158/1078-0432.CCR-06-2073]
- 45 **Zhou Q,** Peng RQ, Wu XJ, Xia Q, Hou JH, Ding Y, Zhou QM, Zhang X, Pang ZZ, Wan DS, Zeng YX, Zhang XS. The density of macrophages in the invasive front is inversely correlated to liver metastasis in colon cancer. *J Transl Med* 2010; **8**: 13 [PMID: 20141634 DOI: 10.1186/1479-5876-8-13]
- 46 **Umehura N,** Saio M, Suwa T, Kitoh Y, Bai J, Nonaka K, Ouyang GF, Okada M, Balazs M, Adany R, Shibata T, Takami T. Tumor-infiltrating myeloid-derived suppressor cells are pleiotropic-inflamed monocytes/macrophages that bear M1- and M2-type characteristics. *J Leukoc Biol* 2008; **83**: 1136-1144 [PMID: 18285406 DOI: 10.1189/jlb.0907611]
- 47 **DeNardo DG,** Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, Coussens LM. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. *Cancer Cell* 2009; **16**: 91-102 [PMID: 19647220 DOI: 10.1016/j.ccr.2009.06.018]
- 48 **Nagtegaal ID,** Marijnen CA, Kranenbarg EK, Mulder-Stapel A, Hermans J, van de Velde CJ, van Krieken JH. Local and distant recurrences in rectal cancer patients are predicted by the nonspecific immune response; specific immune response has only a systemic effect--a histopathological and immunohistochemical study. *BMC Cancer* 2001; **1**: 7 [PMID: 11481031 DOI: 10.1186/1471-2407-1-7]
- 49 **Gounaris E,** Erdman SE, Restaino C, Gurish MF, Friend DS, Gounari F, Lee DM, Zhang G, Glickman JN, Shin K, Rao VP, Poutahidis T, Weissleder R, McNagny KM, Khazaie K. Mast cells are an essential hematopoietic component for polyp development. *Proc Natl Acad Sci USA* 2007; **104**: 19977-19982 [PMID: 18077429 DOI: 10.1073/pnas.0704620104]
- 50 **Nielsen HJ,** Hansen U, Christensen IJ, Reimert CM, Brünner N, Moesgaard F. Independent prognostic value of eosinophil and mast cell infiltration in colorectal cancer tissue. *J Pathol* 1999; **189**: 487-495 [PMID: 10629548 DOI: 10.1002/(SICI)1096-9896(199912)189:]
- 51 **Gulubova M,** Vlaykova T. Prognostic significance of mast cell number and microvascular density for the survival of patients with primary colorectal cancer. *J Gastroenterol Hepatol* 2009; **24**: 1265-1275 [PMID: 17645466 DOI: 10.1111/j.1440-1746.2007.05009.x]

- 52 **Blatner NR**, Bonert A, Beckhove P, Cheon EC, Krantz SB, Strouch M, Weitz J, Koch M, Halverson AL, Bentrem DJ, Khazaie K. In colorectal cancer mast cells contribute to systemic regulatory T-cell dysfunction. *Proc Natl Acad Sci USA* 2010; **107**: 6430-6435 [PMID: 20308560 DOI: 10.1073/pnas.0913683107]
- 53 **Halazun KJ**, Aldoori A, Malik HZ, Al-Mukhtar A, Prasad KR, Toogood GJ, Lodge JP. Elevated preoperative neutrophil to lymphocyte ratio predicts survival following hepatic resection for colorectal liver metastases. *Eur J Surg Oncol* 2008; **34**: 55-60 [PMID: 17448623 DOI: 10.1016/j.ejso.2007.02.014]
- 54 **Vesely P**, Tusková M, Melichar B. Phenotype of peripheral blood leukocytes and survival of patients with metastatic colorectal cancer. *Int J Biol Markers* 2005; **20**: 126-133 [PMID: 16011044]
- 55 **Holcombe RF**, Jacobson J, Dakhil SR, Stewart RM, Betting KS, Kannan K, Macdonald JS. Association of immune parameters with clinical outcome in stage III colon cancer: results of Southwest Oncology Group Protocol 9009. *Cancer Immunol Immunother* 1999; **48**: 533-539 [PMID: 10602891 DOI: 10.1007/s002620050602]
- 56 **Xie ZJ**, Jia LM, He YC, Gao JT. Morphological observation of tumor infiltrating immunocytes in human rectal cancer. *World J Gastroenterol* 2006; **12**: 1757-1760 [PMID: 16586547]
- 57 **Suzuki A**, Masuda A, Nagata H, Kameoka S, Kikawada Y, Yamakawa M, Kasajima T. Mature dendritic cells make clusters with T cells in the invasive margin of colorectal carcinoma. *J Pathol* 2002; **196**: 37-43 [PMID: 11748640 DOI: 10.1002/path.1018]
- 58 **Akira S**, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006; **124**: 783-801 [PMID: 16497588 DOI: 10.1016/j.cell.2006.02.015]
- 59 **Franchi L**, McDonald C, Kanneganti TD, Amer A, Núñez G. Nucleotide-binding oligomerization domain-like receptors: intracellular pattern recognition molecules for pathogen detection and host defense. *J Immunol* 2006; **177**: 3507-3513 [PMID: 16951308]
- 60 **Fukata M**, Hernandez Y, Conduah D, Cohen J, Chen A, Breglio K, Goo T, Hsu D, Xu R, Abreu MT. Innate immune signaling by Toll-like receptor-4 (TLR4) shapes the inflammatory microenvironment in colitis-associated tumors. *Inflamm Bowel Dis* 2009; **15**: 997-1006 [PMID: 19229991 DOI: 10.1002/ibd.20880]
- 61 **Chen GY**, Shaw MH, Redondo G, Núñez G. The innate immune receptor Nod1 protects the intestine from inflammation-induced tumorigenesis. *Cancer Res* 2008; **68**: 10060-10067 [PMID: 19074871 DOI: 0008-5472.CAN-08-2061]
- 62 **Terzić J**, Grivennikov S, Karin E, Karin M. Inflammation and colon cancer. *Gastroenterology* 2010; **138**: 2101-2114.e5 [PMID: 20420949 DOI: 10.1053/j.gastro.2010.01.058]
- 63 **Chiba T**, Ohtani H, Mizoi T, Naito Y, Sato E, Nagura H, Ohuchi A, Ohuchi K, Shiiba K, Kurokawa Y, Satomi S. Intraepithelial CD8+ T-cell-count becomes a prognostic factor after a longer follow-up period in human colorectal carcinoma: possible association with suppression of micrometastasis. *Br J Cancer* 2004; **91**: 1711-1717 [PMID: 15494715]
- 64 **Banerjee A**, Bustin SA, Dorudi S. The immunogenicity of colorectal cancers with high-degree microsatellite instability. *World J Surg Oncol* 2005; **3**: 26 [PMID: 15890075 DOI: 10.1186/1477-7819-3-26]
- 65 **Paschen A**, Eichmüller S, Schadendorf D. Identification of tumor antigens and T-cell epitopes, and its clinical application. *Cancer Immunol Immunother* 2004; **53**: 196-203 [PMID: 14689239 DOI: 10.1007/s00262-003-0479-3]
- 66 **Loose D**, Van de Wiele C. The immune system and cancer. *Cancer Biother Radiopharm* 2009; **24**: 369-376 [PMID: 19538060 DOI: 10.1089/cbr.2008.0593]
- 67 **Barnas JL**, Simpson-Abelson MR, Yokota SJ, Kelleher RJ, Bankert RB. T cells and stromal fibroblasts in human tumor microenvironments represent potential therapeutic targets. *Cancer Microenviron* 2010; **3**: 29-47 [PMID: 21209773 DOI: 10.1007/s12307-010-0044-5]
- 68 **Zou W**. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol* 2006; **6**: 295-307 [PMID: 16557261 DOI: 10.1038/nri1806]
- 69 **Saurer L**, Mueller C. T cell-mediated immunoregulation in the gastrointestinal tract. *Allergy* 2009; **64**: 505-519 [PMID: 19210347 DOI: 10.1111/j.1398-9995.2009.01965.x]
- 70 **Needham DJ**, Lee JX, Beilharz MW. Intra-tumoural regulatory T cells: a potential new target in cancer immunotherapy. *Biochem Biophys Res Commun* 2006; **343**: 684-691 [PMID: 16563349 DOI: 10.1016/j.bbrc.2006.03.018]
- 71 **Clarke SL**, Betts GJ, Plant A, Wright KL, El-Shanawany TM, Harrop R, Torkington J, Rees BI, Williams GT, Gallimore AM, Godkin AJ. CD4+CD25+FOXP3+ regulatory T cells suppress anti-tumor immune responses in patients with colorectal cancer. *PLoS One* 2006; **1**: e129 [PMID: 17205133 DOI: 10.1371/journal.pone.0000129]
- 72 **Khan AR**, Dovedi SJ, Wilkinson RW, Pritchard DI. Tumor infiltrating regulatory T cells: tractable targets for immunotherapy. *Int Rev Immunol* 2010; **29**: 461-484 [PMID: 20839911 DOI: 10.3109/08830185.2010.508854]
- 73 **Curiel TJ**, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, Evdemon-Hogan M, Conejo-Garcia JR, Zhang L, Burow M, Zhu Y, Wei S, Kryczek I, Daniel B, Gordon A, Myers L, Lackner A, Disis ML, Knutson KL, Chen L, Zou W. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004; **10**: 942-949 [PMID: 15322536 DOI: 10.1038/nm1093]
- 74 **Hiraoka N**, Onozato K, Kosuge T, Hirohashi S. Prevalence of FOXP3+ regulatory T cells increases during the progression of pancreatic ductal adenocarcinoma and its premalignant lesions. *Clin Cancer Res* 2006; **12**: 5423-5434 [PMID: 17000676 DOI: 10.1158/1078-0432.CCR-06-0369]
- 75 **Kobayashi N**, Hiraoka N, Yamagami W, Ojima H, Kanai Y, Kosuge T, Nakajima A, Hirohashi S. FOXP3+ regulatory T cells affect the development and progression of hepatocarcinogenesis. *Clin Cancer Res* 2007; **13**: 902-911 [PMID: 17289884 DOI: 10.1158/1078-0432.CCR-06-2363]
- 76 **Salama P**, Phillips M, Griew F, Morris M, Zeps N, Joseph D, Platell C, Iacopetta B. Tumor-infiltrating FOXP3+ T regulatory cells show strong prognostic significance in colorectal cancer. *J Clin Oncol* 2009; **27**: 186-192 [PMID: 19064967 DOI: 10.1200/JCO.2008.18.7229]
- 77 **Correale P**, Rotundo MS, Del Vecchio MT, Remondo C, Migali C, Ginanneschi C, Tsang KY, Licchetta A, Mannucci S, Loiacono L, Tassone P, Francini G, Tagliaferri P. Regulatory (FoxP3+) T-cell tumor infiltration is a favorable prognostic factor in advanced colon cancer patients undergoing chemo or chemioimmunotherapy. *J Immunother* 2010; **33**: 435-441 [PMID: 20386463 DOI: 10.1097/CJI.0b013e3181d32f01]
- 78 **Suzuki H**, Chikazawa N, Tasaka T, Wada J, Yamasaki A, Kitaura Y, Sozaki M, Tanaka M, Onishi H, Morisaki T, Katanano M. Intratumoral CD8(+) T/FOXP3 (+) cell ratio is a predictive marker for survival in patients with colorectal cancer. *Cancer Immunol Immunother* 2010; **59**: 653-661 [PMID: 19908042 DOI: 10.1007/s00262-009-0781-9]
- 79 **Frey DM**, Droezer RA, Viehl CT, Zlobec I, Lugli A, Zingg U, Oertli D, Kettelhack C, Terracciano L, Tornillo L. High frequency of tumor-infiltrating FOXP3(+) regulatory T cells predicts improved survival in mismatch repair-proficient colorectal cancer patients. *Int J Cancer* 2010; **126**: 2635-2643 [PMID: 19856313]
- 80 **Pagès F**, Galon J, Dieu-Nosjean MC, Tartour E, Sautès-Fridman C, Fridman WH. Immune infiltration in human tumors: a prognostic factor that should not be ignored. *Oncogene* 2010; **29**: 1093-1102 [PMID: 19946335 DOI: 10.1038/onc.2009.416]
- 81 **Zlobec I**, Lugli A. Invasive front of colorectal cancer: dynamic interface of pro-/anti-tumor factors. *World J Gastro-*

- enterol* 2009; **15**: 5898-5906 [PMID: 20014453 DOI: 10.3748/wjg.15.5898]
- 82 **Pages F**, Berger A, Camus M, Sanchez-Cabo F, Costes A, Molitor R, Mlecnik B, Kirilovsky A, Nilsson M, Damotte D, Meatchi T, Bruneval P, Cugnenc PH, Trajanoski Z, Fridman WH, Galon J. Effector memory T cells, early metastasis, and survival in colorectal cancer. *N Engl J Med* 2005; **353**: 2654-2666 [PMID: 16371631 DOI: 10.1056/NEJMoa051424]
 - 83 **Deschoolmeester V**, Baay M, Van Marck E, Weyler J, Vermeulen P, Lardon F, Vermorken JB. Tumor infiltrating lymphocytes: an intriguing player in the survival of colorectal cancer patients. *BMC Immunol* 2010; **11**: 19 [PMID: 20385003 DOI: 10.1186/1471-2172-11-19]
 - 84 **Dunn GP**, Old LJ, Schreiber RD. The three Es of cancer immunoediting. *Annu Rev Immunol* 2004; **22**: 329-360 [PMID: 15032581 DOI: 10.1146/annurev.immunol.22.012703.104803]
 - 85 **Laghi L**, Bianchi P, Miranda E, Balladore E, Pacetti V, Grizzi F, Allavena P, Torri V, Repici A, Santoro A, Mantovani A, Roncalli M, Malesci A. CD3+ cells at the invasive margin of deeply invading (pT3-T4) colorectal cancer and risk of post-surgical metastasis: a longitudinal study. *Lancet Oncol* 2009; **10**: 877-884 [PMID: 19656725 DOI: 10.1016/S1470-2045(09)70186-X]
 - 86 **Nosho K**, Baba Y, Tanaka N, Shima K, Hayashi M, Meyerhardt JA, Giovannucci E, Dranoff G, Fuchs CS, Ogino S. Tumour-infiltrating T-cell subsets, molecular changes in colorectal cancer, and prognosis: cohort study and literature review. *J Pathol* 2010; **222**: 350-366 [PMID: 20927778 DOI: 10.1002/path.2774]
 - 87 **Grizzi F**, Chiriva-Internati M. Cancer: looking for simplicity and finding complexity. *Cancer Cell Int* 2006; **6**: 4 [PMID: 16480511 DOI: 10.1186/1475-2867-6-4]
 - 88 **Grizzi F**, Di Ieva A, Russo C, Frezza EE, Cobos E, Muzzio PC, Chiriva-Internati M. Cancer initiation and progression: an unsimplifiable complexity. *Theor Biol Med Model* 2006; **3**: 37 [PMID: 17044918 DOI: 10.1186/1742-4682-3-37]

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Gastrointestinal radiation injury: Symptoms, risk factors and mechanisms

Abobakr K Shadad, Frank J Sullivan, Joseph D Martin, Laurence J Egan

Abobakr K Shadad, Laurence J Egan, Department of Gastroenterology, University Hospital Galway, University Hospital Galway, 34562 Galway, Ireland

Abobakr K Shadad, Laurence J Egan, Department of Pharmacology and Therapeutics, School of Medicine, University Hospital Galway, 34562 Galway, Ireland

Frank J Sullivan, Joseph D Martin, Department of Radiation Oncology, National University of Ireland Galway, University Hospital Galway, 34562 Galway, Ireland

Author contributions: Shadad AK wrote the review; Sullivan FJ, Martin JD and Egan LJ revised the contents.

Correspondence to: Laurence J Egan, Professor, Department of Pharmacology and Therapeutics, School of Medicine, University Hospital Galway, 34562 Galway, Ireland. laurence.egan@nuigalway.ie

Telephone: +353-91-495370 Fax: +353-91-495572

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various clinical manifestations of post-radiation gastrointestinal symptoms, to discuss possible patient and treatment factors implicated in normal gastrointestinal tissue radiosensitivity and to outline different mechanisms of intestinal tissue injury.

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Abstract

Ionising radiation therapy is a common treatment modality for different types of cancer and its use is expected to increase with advances in screening and early detection of cancer. Radiation injury to the gastrointestinal tract is important factor working against better utility of this important therapeutic modality. Cancer survivors can suffer a wide variety of acute and chronic symptoms following radiotherapy, which significantly reduces their quality of life as well as adding an extra burden to the cost of health care. The accurate diagnosis and treatment of intestinal radiation injury often represents a clinical challenge to practicing physicians in both gastroenterology and oncology. Despite the growing recognition of the problem and some advances in understanding the cellular and molecular mechanisms of radiation injury, relatively little is known about the pathophysiology of gastrointestinal radiation injury or any possible susceptibility factors that could aggravate its severity. The aims of this review are to examine the

INTRODUCTION

Radiation delivery methods

Radiation therapy can be delivered by three main methods. External beam radiotherapy is a method in which radiation beam is delivered from outside the body to the target tumour through two or three-dimensional beam arrays using linear accelerators. Advances in planning and delivery techniques such as 3-dimensional simulation and intensity modulated radiation therapy are associated with a reduced risk of normal tissue toxicity and allow a higher radiation dose to be used compared to conventional two dimensional methods^[1-4]. Enhanced target definition of both tumours and surrounding normal tissues and combining beams of varying intensity in intensity modulated radiation therapy allow for better dose delivery and improved tumour control with less toxicity^[5,6]. Quantitative dose tracking to both normal tissues and tumors in the form of dose-volume histograms^[7] provide a graphic display of a simulated radiation treatment plan and generate valuable information on the dose distribution within the volume of interest^[8]. These are now common planning

tools in modern external beam radiation delivery. Brachytherapy is an internal form of radiation therapy where the radiation sources are implanted within or in close proximity to the target tumour to deliver a high dose of localized radiation. This procedure is a highly effective dose delivery method for certain tumours such as prostate and gynaecological cancers. The third method of the radiation therapy is the systemic administration of radioactive particles which is termed radioisotope therapy. In this method radioactive particles (radionuclides) are injected into the blood stream to be adsorbed specifically by the targeted tissue such as thyroid gland^[9]. Gastrointestinal radiation injury most commonly occurs following external beam therapy.

A highly precise radiation delivery can be achieved through newer techniques such as image guided radiotherapy techniques which allow verification of the target position on a daily basis to account for internal target motion^[10]. Stereotactic radiation therapy^[11] can focus a narrow radiation beams on a small target such as early cancer or metastatic lesions^[12].

TREATMENT RELATED RISK FACTORS FOR GASTROINTESTINAL INJURY

Radiation dose, fractionation and field size

Radiation dose is a major determinant of the severity of acute and late normal tissue toxicity^[13-20], the desired optimal radiation dose is defined as the dose that maximizes the difference between “tumor” and “normal tissue” damage within the sigmoid shape dose-effect relationship curve^[11]. With respect to the gastrointestinal tract, the severity of toxicity is reported as Grades of severity to different symptoms or clinical manifestations ranging from minor symptomatic changes to severe life threatening complications. Multiple toxicity grading systems have been developed to assess adverse events of cancer treatment^[21]. Generally, Grade 1 and 2 radiation injury are frequent and they are often requiring no treatment although they can cause a considerable effect on patient quality of life. Examples of commonly used toxicity grading system to assess radiation injury severity are the Radiation Therapy Oncology Group^[22] and the Common Terminology Criteria for Adverse Events grading system (Table 1)^[21]. Radiation dose per fraction and altered fractionation schedules are important factors linked to increased risk of intestinal radiation toxicity^[23]. The radiosensitivity of the cell depends on two factors, the intrinsic radiosensitivity which is linearly related to the radiation dose and it represents the initial slope of the cell survival curve (alpha). The second factor is (beta) which represents the curvature of the cell survival curve and it is a factor of dose-per fraction and dose-rate variations in radiobiology^[24].

The alpha/beta ratio represents the dose at which the linear and quadratic components of the Linear-Quadratic model contribute equally to cell killing and has been shown to have a connection to early and late radiation

Table 1 Example of some gastrointestinal symptoms grades following radiation injury

Grade	Gastrointestinal symptoms
	Nausea
1	Loss of appetite without alteration in eating habits
2	Oral intake decreased without significant weight loss, dehydration or malnutrition; IV fluids indicated < 24 h
3	Inadequate oral caloric or fluid intake; IV fluids, tube feedings, or TPN indicated ≥ 24 h
4	Life-threatening consequences
5	Death
	Anorexia
1	Loss of appetite without alteration in eating habits
2	Oral intake altered without significant weight loss or malnutrition; oral nutritional supplements indicated
3	Associated with significant weight loss or malnutrition (e.g., inadequate oral caloric and/or fluid intake); IV fluids, tube feedings or TPN indicated
4	Life-threatening consequences
5	Death
	Haemorrhage-GI
1	Mild, intervention (other than iron supplements) not indicated
2	Symptomatic and medical intervention or minor cauterization indicated
3	Transfusion, interventional radiology, endoscopic, or operative intervention indicated; radiation therapy (i.e., hemostasis of bleeding site)
4	Life-threatening consequences; major urgent intervention indicated
5	Death
	Ulceration-GI
1	Asymptomatic, radiographic or endoscopic findings only
2	Symptomatic; altered GI function (e.g., altered dietary habits, oral supplements); IV fluids indicated < 24 h
3	Symptomatic and severely altered GI function (e.g., inadequate oral caloric or fluid intake); IV fluids, tube feedings, or TPN indicated ≥ 24 h
4	Life-threatening consequences
5	Death
	Incontinence anal
1	Occasional use of pads required
2	Daily use of pads required
3	Interfering with ADL; operative intervention indicated
4	Permanent bowel diversion indicated
5	Death

According to the Common Terminology Criteria for Adverse Events system v 3.0. IV: Intravenous; GI: Gastrointestinal; ADL: Activities of daily living; TPN: Total parenteral nutrition.

response^[25]. In radiotherapy of tumors with long turn-over time such as prostate cancer, the alpha/beta ratio is smaller than that of early reacting normal tissues. In this case, hypo-fractionation will be a better strategy for radiotherapy than the many small fractions used for other tumors^[24,26,27].

A data analysis of 918 head and neck cancer patients reported a variable prevalence of mucositis between patients treated with continuous hyperfractionated accelerated radiotherapy (CHART) and patients received conventional fractionation radiotherapy. The incidence of Grade 3 confluent mucositis reported after CHART was 75% compared to 44% following conventional fractionation radiotherapy^[28].

Modification of the radiation delivery regimes through

Table 2 Summary of risk factors for gastrointestinal radiation injury

Risk factors	
Radiation techniques	Treatment volume, total dose, fractionation dose and schedules
Combined modality therapies	Surgery
Medical co-morbidities	Chemotherapy: Particularly concurrent
Genetic susceptibility	Vascular disease, connective tissue disease, inflammatory bowel disease, HIV Single nucleotide polymorphism, ataxia telangiectasia

HIV: Human immunodeficiency virus.

hypofractionation was also suggested to be safer than conventionally fractionated conformal radiotherapy in a randomized study of prostate cancer radiotherapy^[29].

Treatment field size and intestinal volume irradiated are important factor and a key determinant of radiation toxicity. Bowel toxicity was found to be directly related to the volume of small intestine irradiated^[13]. Moreover, irradiation to a larger volume of small intestine was reported to increase the operative mortality in rectal cancer patients treated with anterior and posterior field irradiation technique^[30,31]. Furthermore, the impact of the field volume has been demonstrated in a randomized study of prostate cancer radiotherapy. Patients who were treated with conformal shielding with 48% less volume irradiation had less rectal toxicity at 5 years than patients treated with conventional radiotherapy, despite identical radiation dose^[32,33] (Table 2).

Combined modality approaches

Combined modality therapy increases the risk of radiation toxicity. Surgery or concurrent chemotherapy is linked to an increased incidence of radiation toxicity. Previous abdominal surgery increases the risk of radiation toxicity^[34]. Anatomical changes that increases intestinal exposure to radiation such as postoperative small intestine prolapse into the pelvic cavity^[13,35] or surgical adhesions that fix intestinal segments within the radiation field can all predispose part of the intestine to receive higher doses of radiation^[36]. Combining prostatectomy with radiotherapy can increase rectal toxicity during prostate cancer treatment^[37]. An analysis of acute toxicity was performed in 405 prostate patients in The European Organization for Research and Treatment of Cancer randomized trial 22863. In those patients it was reported that among other factors, previous genitourinary surgery was found to be predictive of lower gastrointestinal radiation toxicity^[38].

Combining chemotherapy with radiation has been reported to increase the rate of acute intestinal toxicity, while the long term effect of this combination is not clear^[20,39]. Oral mucositis was reported in more than 90% of patients treated with a combined chemo-radiotherapy regime for head and neck cancer^[40,41] in comparison to another study which reported an incidence of oral muco-

stitis in 62% of patients treated with radiotherapy alone^[42]. Concurrent chemotherapy with radiation has also been reported to increase the risk of oesophageal radiation injury by 12-fold^[43].

In a study of cervical cancer patients treated with chemoradiotherapy, the incidence of Grade 3 late intestinal toxicity increased from 10% to 26% in patients treated with both mitomycin and fluorouracil compared to a fluorouracil alone group, suggesting a possible role for the type of chemotherapeutic agent used as determinant of severity of gastrointestinal toxicity^[44].

Different mechanisms have been suggested to explain the sensitizing effect of adding chemotherapy in increasing the risk of intestinal radiation injury. Examples of possible mechanisms are alterations in cell cycle kinetics, or synchronization of replicating cell populations. Halopyrimidines such as fluorouracil, fluorodeoxyuridine and iododeoxyuridine may sensitize tumors both by inhibiting effective DNA repair and by increasing the amount of radiation induced DNA damage^[45-47].

SYMPTOMS RESULTING FROM RADIATION INJURY TO THE GASTROINTESTINAL TRACT

Acute and chronic gastrointestinal radiation injury

During external beam radiotherapy, ionising radiation enters and exits the body and therefore affects normal tissues surrounding the target tumour. The gastrointestinal tract which extends over a large surface area and any part of the gastrointestinal tract that falls within the radiation field can be affected, resulting in acute and chronic symptoms of gastrointestinal radiation injury (Table 3).

Clinical manifestations of gastrointestinal radiation injury can present during or soon following radiotherapy. These symptoms are related to acute mucosal injury and inflammation. Delayed symptoms occur a few months or years after radiotherapy and are attributed to a chronic process of transmural fibrosis and vascular sclerosis. Typically symptoms are considered "acute" if they occur within the course of treatment or up to 90 d following treatment. These are generally reversible. Chronic side effects are much less common and occur more than 90 d post radiation; they are less likely to reverse. The onset of delayed symptoms has been reported as much as after 30 years following radiotherapy^[48].

Mouth, pharynx, and oesophagus

During radiotherapy for head and neck or thoracic cancer, the upper gastrointestinal tract falls within the radiation field. Irradiation to this area results in acute mucosal injury causing mucositis and ulceration which manifests within the first two weeks in 30%-60% of patients, causing dysphagia and odynophagia^[49]. In a study of 254 non-small-cell lung cancer patients, acute toxicity of Grade 2 or worse has been reported in 78% of patients^[50]. Mucositis is debilitating, can be a dose limiting side effect and

Table 3 Acute and chronic manifestations of gastrointestinal radiation injury

Clinical manifestations	Radiation tolerance dose TD5/5, TD50/5 (Gy)	Gastrointestinal organ
Oral mucositis occurs in > 90% of patients with concurrent chemotherapy ^[40] Xerostomia and altered saliva composition	Parotid gland: TD5/5 (32) TD50/5 (46) ^[60,179,180]	Mouth, salivary glands, hypopharynx, parotid
Acute Grade 3-4 oesophageal injuries occur in 46% with concurrent chemotherapy ^[181] Dose > 58 Gy predicts Grade 3-5 acute oesophagitis ^[54] 60 Gy resulted in Grade 3 toxicity in 42% ^[182] Radiation can lead to late stricture and/or perforation of the oesophagus ^[53,60]	TD5/5 (55-60) TD50/5 (68-72) ^[60,179]	Oesophagus
40 Gy: Severe late toxicity in 7% including ulceration, gastritis and small-bowel obstruction/perforation ^[183] Elevated liver enzymes in 5% ^[184]	TD5/5; (50-60) TD50/5 (65-70) ^[60,179]	Stomach
(31.3-37 Gy resulted in RILD in 9.4% ^[66,185] 45 Gy cause 5% Grade 3-4 toxicity and 14% with con- current chemotherapy ^[71] Diarrhoea, abdominal pain in 20-70% ^[72] Transmural fibrosis lead- ing to obstruction in 5%-10% ^[68,77-78]	TD5/5; (30-50) TD50/5; (40-55) ^[60,179] TD5/5; (40-50)	Liver Small intestine
Intestinal fistulation occurs at a rate of 0.6% to 4.8% ^[68,79] 50 Gy 5 year estimate of small bowel obstruction is 11% ^[186] Colitis in 25%-50% of pa- tients ^[186] Grade 2-3 acute proctitis 40% ^[91] Chronic rectal symptoms in 6.7%-31% ^[91] Acute symptoms of anus and rectal injury occur in up to 75% of patients during radiotherapy ^[187]	TD50/5; (55-60) ^[60,179,180] Colon TD5/5; (45-55) TD50/5 (55-65) ^[60,179,180] Rectum TD5/5; (60-61.38) TD50/5 (80-81.38) ^[60,179,180]	Colon and rectum

TD5/5: Radiation dose associated with 5% of patients' risk of delayed toxicity in 5 years; TD50/5: The radiation dose associated with 50% of patients' risk of delayed toxicity in 5 years. RILD: Radiation-induced liver disease.

is difficult to treat. Severe symptoms may require therapy interruption, or the provision of an alternative nutritional route to avoid dehydration and malnutrition. Therefore, elective percutaneous endoscopic gastrostomy tube insertion before radiotherapy is a recognized practice and it is associated with improved quality of life and a lower rate of hospital admissions^[51,52]. In severe cases, acute radiation oesophagitis can lead to more serious complications such as significant bleeding or oesophageal perforation^[53].

Clinical predictors for acute oesophageal toxicity

include maximum radiation dose. A maximum dose of > 58 Gy was reported to predict the risk of Grade 3-5 oesophageal toxicity in 207 non-small-cell lung cancer patients treated with 3-dimensional conformal radiotherapy^[54] while a dose of 50 Gy was significantly associated with Grade 2 or worse oesophagitis in another cohort 36 non-small-cell lung cancer patients^[55]. In both studies patients received concurrent chemotherapy which is also a risk factor for oesophageal toxicity. The volume of the irradiated oesophagus has also been suggested to predict acute oesophageal toxicity^[55]. In another series of 208 non-small-cell lung cancer patients treated with three dimensional conformal radiotherapy, concurrent chemotherapy and maximal point dose to the oesophagus > 60 Gy were found to be significantly associated with the risk of Grade 3-5 oesophageal injury^[56]. The Quantitative Analyses of Normal Tissue Effects in the Clinic paper by Werner-Wasik *et al.*^[57] published in 2010 reviewed the published data on the dose-volume effect and concluded that it was not possible to identify the best threshold volumetric parameter for oesophageal irradiation given the variety of the volumetric metrics in the published data.

Delayed symptoms of oesophageal injury can manifest after several months following radiotherapy and include chronic ulceration, fistulisation or chronic dysphagia. Dysphagia can be secondary to tissue fibrosis and stricture formation or due to motility disorder induced by muscular or nerve injuries. Delayed oesophageal toxicity has been reported in 17% of non-small-cell lung cancer patients^[50] and the median and maximal time to the onset of late toxicity was 5 and 40 mo respectively after radiotherapy.

Stomach and duodenum

Gastric injury during radiotherapy occurs when the stomach falls within the radiation field of an adjacent tumour. Nausea, vomiting, dyspepsia and abdominal pain has been reported to occur early after radiotherapy to the upper abdomen in 50% of patients^[58]. These symptoms result from acute mucosal injury causing erosions and ulceration of gastric and duodenal mucosa.

Later symptoms include chronic dyspepsia and abdominal pain due to chronic ulceration secondary to mucosal injury^[59]. Rarely, gastric wall fibrosis can lead to gastric outlet obstruction. The radiation dose associated with 5% and 50% of patients risk of delayed gastric toxicity in 5 years (TD5/5 and TD50/5) have been estimated at 50 Gy and 65 Gy respectively for gastric ulceration or perforation. An accurate data on dose-volume constrain for partial gastric irradiation is not available. However, the threshold dose of 45 Gy to the whole stomach has been associated with ulceration in 5%-7% of patients^[60,61].

Liver injury

Following irradiation to the liver, radiation induced liver disease can occur in patients with normal pre-radiotherapy liver function, causing anicteric hepatomegaly and mild alkaline phosphatase serum level elevation. A more

severe derangement of liver function occurs in patients with pre-existing liver disease^[62]. Radiation induced liver disease can progress to fibrosis, cirrhosis, and liver failure^[63]. Abdominal imaging with computed tomography scan or magnetic resonance imaging can be helpful to show low-attenuation injury areas or areas of atrophy in the irradiated liver segment^[64].

Risk factors for radiation induced liver disease include baseline liver dysfunction, Hepatitis B virus carrier status^[65], mean dose > 30 Gy for partial liver radiotherapy, concurrent chemotherapy and volume of liver irradiated^[66,67].

Small intestine

The small intestine receives irradiation during radiotherapy of pelvic or abdominal malignancies. The degree of injury depends on the radiation dose and the volume of intestinal segment that falls within the radiation field^[68,69]. Significant correlation has been suggested between the volume of irradiated small bowel and the likelihood of acute toxicity, regardless of the radiation dose delivered^[70]. Other predictors of acute small intestine toxicity include the use of concurrent chemotherapy. This effect has been reported in 186 cervical cancer patients who received 45 Gy preoperative pelvic radiotherapy alone where 5% of patients experienced Grade 3-4 toxicity in comparison to 14% of 183 patients who received radiotherapy and weekly cisplatin^[71]. The relatively fixed portions of the small intestine such as the duodenum and the terminal ileum are at increased risk of radiation toxicity as they are more susceptible to receive higher doses of radiation than the mobile parts of small intestine.

The radiation dose associated with delayed small bowel toxicity have been estimated by Emami *et al*^[60]. The TD5/5 and TD50/5 doses for one third of small bowel irradiation were estimated at 50 Gy and 60 Gy respectively. The TD5/5 and TD50/5 for the whole-organ irradiation were 40 Gy and 55 Gy respectively. These doses estimates remained as a guide for two decades and more recent data were consistent with these ranges^[60,61].

Clinically, nausea, vomiting and abdominal pain are early symptoms that can occur during the first two weeks following abdominal radiotherapy, and may be mediated by the release of inflammatory cytokines following radiotherapy. Diarrhoea and abdominal pain occur during the first two weeks of radiotherapy to abdominal or pelvic malignancies in 20% to 70% of patients^[72]. This may be a result of direct radiation injury to the small intestinal mucosa causing epithelial atrophy, and reduced mucosal blood flow^[73].

The acute symptoms usually settle within three weeks after completion of radiotherapy and the intestinal epithelium regenerates from stem cells at the base of the crypts.

Delayed symptoms of radiation small intestinal injury manifest months to years after radiotherapy with symptoms of diarrhoea, recurrent abdominal pains and malabsorption.

Chronic diarrhoea following radiotherapy can result from different pathophysiological processes such as bile salt malabsorption, bacterial overgrowth, fat malabsorption, rapid intestinal transit or lactose intolerance^[74]. More chronic symptoms occur as a result of pathological abnormalities to the intestinal vascular compartment resulting in intestinal ischemia as well as progressive intestinal fibrosis leading to structural abnormalities such as strictures and fistulation. Bacterial overgrowth contributes to malabsorption and diarrhoea, particularly in patients with intestinal strictures^[73,75].

Intestinal obstruction can complicate 5% to 10% of severe small intestinal radiation injury^[76]. The rate of severe small intestinal complications following radiotherapy for rectal cancer can vary considerably depending on the tumor and treatment characteristics, Reports indicate rates of 0.8% to 13% for small intestinal obstruction^[68,77,78] and 0.6% to 4.8% for intestinal fistulation^[68,79]. Patients with severe small intestinal injury have a poor prognosis since surgery to manage strictures is complex and has been associated with poor outcomes^[73,80].

Colon and rectum

During abdominal and pelvic radiotherapy, the colon and rectum are commonly affected as their anatomical locations fall within the radiation field of a variety of cancers. The fixed portions of the colon, the caecum and the rectum are at greater risk of receiving higher doses of radiation than the remainder of the colon.

Acute radiation injury to the rectum and anal canal can result in a diversity of symptoms such as abdominal pain, diarrhoea, tenesmus, rectal pain, urgency, rectal discharge, incontinence, and fresh rectal bleeding. These symptoms occur primarily as a consequence of direct mucosal damage^[81-83]. Acute radiation injury to the colon can be severe and in 5%-15% can lead to therapy interruption or treatment plan alteration^[84].

A recent study showed that 47% of women who received radiotherapy for cervical or endometrial cancer reported symptoms of radiation intestinal injury affecting quality of life within 3 mo following therapy completion^[85]. These results are consistent with a previous structured questionnaire study^[86] which showed that 53% of patients had reported bowel symptoms significantly affecting their quality of life, whilst 81% of patients in the study described new-onset gastrointestinal problems after receiving radiotherapy.

The recent data on dose-volume effect in radiation induced rectal injury was reviewed by Michalski *et al*^[87]. The incidence of Grade > 2 injury from different studies was variable according to dose, treatment parameters and scale used in each study. Among the recent studies reported, an incidence of 13.5% and 16% of Grade > 2 rectal injury. Identified predictors for Grade > 2 rectal injury or rectal bleeding include the volume of the rectum irradiated and a total radiation dose > 60 Gy during 3-dimensional conformal radiotherapy. The effect of concurrent chemotherapy has been observed in the

European Organization for Research and Treatment of Cancer study where patients received 45 Gy preoperative radiotherapy or radiotherapy and 5-fluorouracil (5-FU). Grade > 2 diarrhoea occurred in 17% of radiotherapy alone group compared to 38% of radiotherapy and 5-FU group^[88].

Delayed symptoms of radiation colonic injury are insidious and usually follow a progressive course. They can manifest after a latent period of few months or years. One study has reported a 15% incidence of bowel toxicity 20 years after receiving radiotherapy in a cervical cancer cohort^[89]. Severe life threatening complications occur after radiotherapy such as fistulation, sepsis, perforation or bleeding at a rate between 4%-8% within 5-10 year time after radiotherapy^[90]. Patients suffer from a variety of symptoms, such as abdominal pain, changing bowel habits with intermittent diarrhoea. Constipation can result from altered colonic motility due to fibrosis and stricture formation. An abnormal bowel transit can manifest as recurrent pain and increased risk of obstruction or pseudo-obstruction secondary to fecal loading. Excessive fibrosis can cause loss of ano-rectal compliance and manifests as urgency and frequency^[73]. Faecal incontinence has been reported in up to 20% of patients and significantly reduces patients' quality of life^[86,91].

Unlike radiation injury to the small bowel, radiation injury to the colon does not compromise nutrient absorption and malabsorption is uncommon^[73].

Adverse effects of radiation to the pelvis primarily affect the colon and the rectum. However, other organs can be irradiated causing increased morbidity for example injury to the urinary tract or the genital system resulting in symptoms affecting quality of life were reported in 30% of patients^[92-95]. A rare but serious complication of prostate brachytherapy is recto-vesical fistula which occurs with a low frequency of 1 in 250 to 1 in 1000 patients implanted^[96-98].

Radiation exposure increases the risk of a secondary malignancy. Patients who received radiotherapy were shown to have significantly higher risk of developing second cancers both overall and in the areas that were exposed to the radiation field^[99]. In an analysis of testicular cancer survivors which included 28 843 men, the risk of a second cancer was estimated. The patterns of second cancer suggested that many factors may be involved, including previous treatment received, but the precise roles of different factors is still to be clarified. It has been reported that secondary leukaemia was associated with both radiotherapy and chemotherapy, whereas excess cancers of the stomach, bladder, and possibly pancreas were associated mainly with radiotherapy^[100]. This risk also includes colorectal cancer, which can occur more than 10 to 20 years after radiation exposure^[99,101].

PATIENT RELATED RISK FACTORS

Patient factors and individual variations

Individual patient phenotypic factors have been suggest-

ed to influence the susceptibility to intestinal radiation injury. It was reported that older patient age is associated with an increased risk of developing reduced organ function after radiotherapy^[102-104]. Body habitus has been reported as another susceptibility factor, where thin patients with narrow antero-posterior diameter can suffer an increased risk of intestinal radiation toxicity compared to normal individuals^[36]. Smoking status has been associated with risk of chronic intestinal toxicity^[20,105] as well as previous history of surgery^[13,35,36].

Medical co-morbidities

Vascular disease: Co-morbid vascular disease such as hypertension, diabetes mellitus and atherosclerosis were suggested to predispose patients to an increased vascular injury following radiation and subsequent intestinal wall ischemia and impaired tissue repair^[74]. The microocclusive vascular disease in addition to increased blood viscosity in diabetes mellitus were suggested to predispose to intestinal tissue ischemia^[106,107]. One study investigated the possible effect of diabetes mellitus during prostate cancer radiotherapy. The study reported higher rates of late genitourinary/gastrointestinal toxicities in diabetic patients than in non-diabetics (34% and 23% respectively). It was also noticed that diabetics developed complications earlier than the non-diabetics (10 mo and 24 mo respectively)^[108].

Inflammatory bowel disease: Co-morbid inflammatory bowel disease (IBD) is considered in some institutions as a relative contraindication to radiotherapy for concerns of greater risk of acute and late side effects^[109-111]. Intolerance to radiotherapy in IBD patients has been demonstrated in case reports^[112,113] and in a larger retrospective analysis where the incidence of severe acute and late events was 21% and 29% respectively^[114,115]. However, in a large retrospective analysis in patients with colorectal cancer, the data on treatment modality received for 170 colorectal cancer patients with history of IBD found no significant difference in cancer treatment modalities between patients with or without history of IBD. This observation points out that a history of IBD was not a barrier to receive radiotherapy treatment in this patient group^[116].

It has been postulated that co-morbid IBD and intestinal inflammation may alter the acute tissue response to radiotherapy through inflammatory mediators, growth factors and cytokine cascades produced at the site of intestinal injury. Some mediators and cytokines were suggested to decrease the sensitivity to radiation injury e.g., fibroblast growth factor 2, prostaglandin-E2, tumor necrosis factor- α and interleukin (IL)-1 and IL-11. However, others were suggested to have mixed effects e.g., IL-12 protecting bone marrow-derived cells but sensitizing intestinal epithelial cells to radiation injury^[117-123].

Collagen vascular diseases: Collagen vascular diseases (CVD) increases the risk of both acute and chronic radia-

tion toxicity, as has been reported by Chon *et al*^[105] in 4 different studies in patients with and without CVD. On the other hand, radiation may cause an acute exacerbation of systemic symptoms in patients with CVD^[124], possibility through release of fibroblast-triggering mediators by the inflammatory cells^[105].

Human immunodeficiency virus infection: It has been reported that human immunodeficiency virus (HIV) infection induces a state of radiosensitivity because severe mucositis was observed in HIV patients who received radiotherapy for the treatment of Kaposi sarcoma^[125,126]. Support for this hypothesis was found by an increased radiosensitivity of skin fibroblasts of HIV patients with Kaposi sarcoma compared to healthy control^[127]. It was also noted in a study involving 59 HIV positive patients that T-lymphocytes of HIV infected individuals were considerably more sensitive to X-rays compared to that of HIV negative donors^[128]. Housri *et al*^[129] reviewed the recent evidence and suggested recommendations for radiotherapy in HIV patients, based on the strength of the best available evidence, and classified according to Strength of Recommendation Taxonomy. There was no conclusive evidence to support the need for special precautions for HIV patients during radiotherapy^[130].

Genotypic variations

It has been suggested that patient's genotype may impact their individual susceptibility to radiation toxicity. This can occur through inherited germ-line mutations in genes involved in DNA damage detection, DNA repair or cell cycle regulation^[131-133]. Recently the term Radiogenomics has been introduced to refer to the science that aims to predict clinical radiosensitivity and to optimize radiotherapy treatment from individual genetic profiles^[134].

Genetic variations are thought to be a key determinant of normal tissue radiosensitivity and may account for up to 80% of the inter-individual variations in normal tissue reaction to radiotherapy^[135,136]. Support for this hypothesis was provided in a study of breast cancer radiotherapy, which reported the incidence and time to development of radiation-induced telangiectasia^[137]. The results of the study revealed a wide range of variation suggesting that patient-related factors can explain 81%-90% of the patient-to-patient variation in telangiectasia level seen after radiotherapy despite similar radiation treatment given. The results further supported reports of other studies^[138,139].

The state of extreme tissue radiosensitivity which has been identified in patients with germ-line mutations in genes involved in DNA damage detection or DNA repair e.g., Nijmegen breakage syndrome, Fanconi's anemia and Ataxia telangiectasia has supported the potential role of genetic variations as an important determinant of individual's radiation response. Nevertheless, this risk is probably confined to patients and carriers of those mutant genes and is not known to be relevant to other patients receiving radiotherapy^[31,45,140,141].

Candidate gene studies, [single nucleotide polymorphism (SNP) association studies] have investigated the role of many genes which have been linked to different elements of the mechanisms related to the pathogenesis of radiation toxicity. Genes investigated include those involved in DNA repair such as *ATM*, *BRCA1*, *BRCA2*^[142,143], apoptosis such as *TP53*, *BCL2*^[144,145], antioxidant enzymes such as *SOD1*^[146], and growth factors *FGF2*^[147,148] and *VEGF*^[147,148]. In this regard, an association has been suggested between candidate SNPs in the genes *TGFB1*, *SOD2*, *XRCC3*, *XRCC1* and late radiation toxicity in breast cancer patients^[133,149]. Similarly, SNP association studies in pelvic tissue have suggested correlations between some risk genes such as *XRCC1*, *XRCC3*, *TGFB1*, *OGG1* and an increased risk of developing late gastrointestinal and genitourinary radiation toxicity following radiotherapy^[150-152].

MECHANISMS OF RADIATION INJURY

Ionising radiation induces double strand breaks in DNA. This triggers activation of a signalling pathway that leads to activation of tumour suppressor p53. Depending on the extent of the DNA damage, which depends on the radiation dose, and on other factors in the cellular milieu, p53 activation leads to cell cycle arrest and DNA double strand break repair, or apoptosis. In cancer radiotherapy, apoptosis of tumour cells is the desired outcome. However, intestinal crypt epithelial cells are quite sensitive to radiation and the killing of these cells leads to mucosal injury^[153,154]. Specifically, when the dose of radiation is sufficient to kill all of the epithelial stem cells in a crypt, then as the epithelial cells migrate up the crypt and are eventually shed into the intestinal lumen; the crypt cannot be repopulated with epithelial cells, and consequently involutes. When this happens to a large proportion of crypts in a region of intestine, normal barrier function is lost which leads to the exposure of the normally sterile lamina propria to luminal microbes. This triggers an acute inflammatory response associated with immune cellular infiltrates; T lymphocytes, macrophages and neutrophils causing loss of epithelial cells as well as degradation of the extracellular matrix in the lamina propria due to enzymes and mediators released by the immune cells^[155]. A further damage to the mucosal and submucosal tissues are caused by reactive oxygen metabolites which are produced in large amounts by activated leukocytes in the inflamed mucosa and this can induce significant damage to various cellular components, including structural and regulatory proteins, carbohydrates, lipids, DNA, and RNA^[51].

During radiotherapy, ionising radiation kills crypt epithelial stem cells. As a result, crypts involute and epithelial barrier integrity is lost. This provides access of luminal microbes and their products to innate immune cells in the lamina propria, with activation of immune cells. An impaired recognition of bacterial translocation can further exacerbate the inflammatory process and promote

stricture formation by two possible mechanisms. The bacterial wall antigens could cause a secondary excessive up-regulation of pro-inflammatory transcription factors, such as nuclear factor κ B^[156]. This might be followed by prolonged macrophage activation and induction of NADPH oxidase expression^[157] leading to a further increase in oxygen radical secretion to eradicate bacteria leading to further tissue destruction^[155]. Meanwhile translocated bacteria could directly stimulate neighboring mesenchymal cells via pattern recognition receptors leading to increased activation of immune cells^[156,158].

The acute inflammatory process continues but eventually, after the cessation of radiation, through poorly understood mechanisms, crypts start to regenerate and this restores normal epithelial barrier function, which is followed by resolution of inflammation^[159]. In some patients this inflammatory process becomes exaggerated for unknown reasons with severe ulceration and inflammatory process runs a chronic course characterised by extensive fibrosis and intestinal ischemia^[160,161]. Recent observations in animal models of radiation injury indicate that repair after radiation may depend on the recruitment of mesenchymal stem cells from the bone marrow to the site of radiation injury. Mesenchymal stem cell mobilization and engraftment is thought to be induced by cytokines and potentially specific homing induced by chemokines, all of these are released by inflammation^[162,163].

The final pathological outcome of the radiation injury in the intestinal tissue will depend on a complex crosstalk between various cellular components of the tissue within the extracellular matrix which eventually determine the process of tissue recovery or long term complications^[73,164].

Radiation injury to the vascular compartment is thought to be a key feature in the pathological processes of intestinal radiation injury as well as an important determinant of both acute and chronic effects after radiotherapy^[165,166]. It has been regarded as a major component in the initiation, progression and maintenance of delayed intestinal tissue damage and enhanced fibrosis which lead to loss of mucosal function and stricture formation^[167,168].

Endothelial cell apoptosis has been implicated as the primary lesion leading to epithelial stem cell dysfunction and subsequent intestinal tissue damage following radiotherapy. Support for this hypothesis was found by identifying a state of radioresistance following inhibition of endothelial apoptosis in experimental mice. Radiation-induced crypt damage, organ failure, and death from radiation injury were all prevented when the endothelial apoptosis was inhibited pharmacologically, by the administration of fibroblast growth factor or genetically by deletion of the acid sphingomyelinase gene^[169]. However, subsequent studies challenged this concept by demonstrating an epithelial cell apoptosis at lower radiation doses which is insufficient to cause endothelial cell death. This result has been enforced further by experiments using high dose Boron therapy radiation, specifically targeted to the endothelium. The results showed no effect

on epithelial stem cell survival^[168,170].

Formation of new blood vessels (angiogenesis) is a crucial requirement for tumour growth and survival^[171]. The tumour vasculature is prone to hypoxia which results in further production of proangiogenic factors by tumour cells. Angiogenesis inhibitors target tumour endothelial cells and cause inhibition of new vessel formation and a transient tumour hypoxia^[172,173]. Although tumour hypoxia has been linked to increasing tumour radio-resistance, studies have shown that the administration of angiogenesis inhibitors improves tumour oxygenation and response to radiotherapy^[174-177]. Different mechanism has been suggested for the radio-sensitising effect of combining angiogenesis inhibitors with radiotherapy^[173]. Mazon *et al.*^[178] has recently reviewed the clinical trials on angiogenesis inhibitors, but despite the promising value of these new agents, the biological basis for their synergistic effect and the safety and efficacy of these agents are still to be determined.

CONCLUSION

Intestinal radiation injury is a significant clinical issue which is expected to increase in prevalence due to improved survival of cancer patients as well as to increased availability of radiotherapy as an affordable treatment option. Radiotherapy treatment can cause a wide variety of gastrointestinal side effects. Following radiation injury to the gastrointestinal tract, symptoms can present acutely or after a long period of time. Although severe intestinal injury is less common with the development of advanced radiotherapy planning and delivery techniques, a less severe degree of injury is common and continues to affect a considerable proportion of patients, significantly reduces their quality of life, and an extra burden to the cost of health care. The accurate diagnosis and management of intestinal radiation injury represents a clinical challenge to practicing physicians in both gastroenterology and oncology. Despite the growing recognition of the problem and some advances in understanding the cellular and molecular mechanisms of radiation injury, relatively little is known about the pathophysiology of intestinal radiation injury or the exact factors that aggravate it. Patient and treatment related risk factors have been suggested although the exact influence posed by these factors is still to be better characterized. Combined modality therapies for cancer are commonly used and they increase the risk of radiation toxicity and further add to the problem. Medical co-morbid diseases such as vascular disease, inflammatory bowel disease and collagen vascular disease can pose a significant risk that can affect patient suitability to receive radiotherapy treatment. Genotypic variations can influence the risk of gastrointestinal radiation injury but future research findings on this area are needed to assess their clinical importance. A better understanding of the pathophysiology of radiation injury may provide the opportunity to develop more effective preventive and therapeutic strategies.

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REFERENCES

- Meyer J, Czito B, Yin FF, Willett C. Advanced radiation therapy technologies in the treatment of rectal and anal cancer: intensity-modulated photon therapy and proton therapy. *Clin Colorectal Cancer* 2007; **6**: 348-356 [PMID: 17311699 DOI: 10.3816/CCC.2007.n.003]
- Meyer JJ, Willett CG, Czito BG. Emerging role of intensity-modulated radiation therapy in anorectal cancer. *Expert Rev Anticancer Ther* 2008; **8**: 585-593 [PMID: 18402525 DOI: 10.1586/14737140.8.4.585]
- Willett CG. Technical advances in the treatment of patients with rectal cancer. *Int J Radiat Oncol Biol Phys* 1999; **45**: 1107-1108 [PMID: 10613301]
- Meliani E, Mageras GS, Fuks Z, Leibel SA, Niehaus A, Lorant H, Zelefsky M, Baldwin B, Kutcher GJ. Variation in prostate position quantitation and implications for three-dimensional conformal treatment planning. *Int J Radiat Oncol Biol Phys* 1997; **38**: 73-81 [PMID: 9212007]
- Ling CC, Burman C, Chui CS, Kutcher GJ, Leibel SA, LoSasso T, Mohan R, Bortfeld T, Reinstein L, Spirou S, Wang XH, Wu Q, Zelefsky M, Fuks Z. Conformal radiation treatment of prostate cancer using inversely-planned intensity-modulated photon beams produced with dynamic multileaf collimation. *Int J Radiat Oncol Biol Phys* 1996; **35**: 721-730 [PMID: 8690638]
- Fraass BA, Kessler ML, McShan DL, Marsh LH, Watson BA, Duseau WJ, Eisbruch A, Sandler HM, Lichter AS. Optimization and clinical use of multisegment intensity-modulated radiation therapy for high-dose conformal therapy. *Semin Radiat Oncol* 1999; **9**: 60-77 [PMID: 10196399]
- Drzymala RE, Mohan R, Brewster L, Chu J, Goitein M, Harms W, Urie M. Dose-volume histograms. *Int J Radiat Oncol Biol Phys* 1991; **21**: 71-78 [PMID: 2032898]
- Niemierko A, Goitein M. Dose-volume distributions: a new approach to dose-volume histograms in three-dimensional treatment planning. *Med Phys* 1994; **21**: 3-11 [PMID: 8164585 DOI: 10.1118/1.597361]
- Clarke SE. Radionuclide therapy in oncology. *Cancer Treat Rev* 1994; **20**: 51-71 [PMID: 8293428 DOI: 10.1016/0305-7372(94)90010-8]
- Chung HT, Xia P, Chan LW, Park-Somers E, Roach M. Does image-guided radiotherapy improve toxicity profile in whole pelvic-treated high-risk prostate cancer? Comparison between IG-IMRT and IMRT. *Int J Radiat Oncol Biol Phys* 2009; **73**: 53-60 [PMID: 18501530 DOI: 10.1016/j.ijrobp.2008.03.015]
- Ikushima H. Radiation therapy: state of the art and the future. *J Med Invest* 2010; **57**: 1-11 [PMID: 20299738]
- Lo SS, Fakiris AJ, Chang EL, Mayr NA, Wang JZ, Papiez L, Teh BS, McGarry RC, Cardenas HR, Timmerman RD. Stereotactic body radiation therapy: a novel treatment modality. *Nat Rev Clin Oncol* 2010; **7**: 44-54 [PMID: 19997074 DOI: 10.1038/nrclinonc.2009.188]
- Letschert JG, Lebesque JV, de Boer RW, Hart AA, Bartelink H. Dose-volume correlation in radiation-related late small-bowel complications: a clinical study. *Radiother Oncol* 1990; **18**: 307-320 [PMID: 2244018 DOI: 10.1016/0167-8140(90)90111-9]
- Mak AC, Rich TA, Schultheiss TE, Kavanagh B, Ota DM, Romsdahl MM. Late complications of postoperative radiation therapy for cancer of the rectum and rectosigmoid. *Int J Radiat Oncol Biol Phys* 1994; **28**: 597-603 [PMID: 8113102]
- Baglan KL, Frazier RC, Yan D, Huang RR, Martinez AA, Robertson JM. The dose-volume relationship of acute small bowel toxicity from concurrent 5-FU-based chemotherapy and radiation therapy for rectal cancer. *Int J Radiat Oncol Biol Phys* 2002; **52**: 176-183 [PMID: 11777636]
- Gunnlaugsson A, Kjellén E, Nilsson P, Bendahl PO, Willner J, Johnsson A. Dose-volume relationships between enteritis and irradiated bowel volumes during 5-fluorouracil and oxaliplatin based chemoradiotherapy in locally advanced rectal cancer. *Acta Oncol* 2007; **46**: 937-944 [PMID: 17851844 DOI: 10.1080/02841860701317873]
- Huang EY, Sung CC, Ko SF, Wang CJ, Yang KD. The different volume effects of small-bowel toxicity during pelvic irradiation between gynecologic patients with and without abdominal surgery: a prospective study with computed tomography-based dosimetry. *Int J Radiat Oncol Biol Phys* 2007; **69**: 732-739 [PMID: 17531397 DOI: 10.1016/j.ijrobp.2007.03.060]
- Tho LM, Glegg M, Paterson J, Yap C, MacLeod A, McCabe M, McDonald AC. Acute small bowel toxicity and preoperative chemoradiotherapy for rectal cancer: investigating dose-volume relationships and role for inverse planning. *Int J Radiat Oncol Biol Phys* 2006; **66**: 505-513 [PMID: 16879928 DOI: 10.1016/j.ijrobp.2006.05.005]
- Minsky BD, Conti JA, Huang Y, Knopf K. Relationship of acute gastrointestinal toxicity and the volume of irradiated small bowel in patients receiving combined modality therapy for rectal cancer. *J Clin Oncol* 1995; **13**: 1409-1416 [PMID: 7751886]
- Theis VS, Sripadam R, Ramani V, Lal S. Chronic radiation enteritis. *Clin Oncol (R Coll Radiol)* 2010; **22**: 70-83 [PMID: 19897345 DOI: 10.1016/j.clon.2009.10.003]
- Trotti A, Colevas AD, Setser A, Rusch V, Jaques D, Budach V, Langer C, Murphy B, Cumberlin R, Coleman CN, Rubin P. CTCAE v3.0: development of a comprehensive grading system for the adverse effects of cancer treatment. *Semin Radiat Oncol* 2003; **13**: 176-181 [PMID: 12903007 DOI: 10.1016/S1053-4296(03)00031-6]
- Cox JD, Stetz J, Pajak TF. Toxicity criteria of the Radiation Therapy Oncology Group (RTOG) and the European Organization for Research and Treatment of Cancer (EORTC) *Int J Radiat Oncol Biol Phys* 1995; **31**: 1341-1346 [PMID: 7713792]
- Denham JW. Influence of dose-rate on inflammatory damage and adhesion molecule expression after abdominal radiation in the rat. *Int J Radiat Oncol Biol Phys* 2000; **47**: 1460-1461 [PMID: 10939886]
- Fowler JF. The radiobiology of prostate cancer including new aspects of fractionated radiotherapy. *Acta Oncol* 2005; **44**: 265-276 [PMID: 16076699 DOI: 10.1080/02841860410002824]
- Thames HD, Bentzen SM, Turesson I, Overgaard M, Van den Bogaert W. Time-dose factors in radiotherapy: a review of the human data. *Radiother Oncol* 1990; **19**: 219-235 [PMID: 2281152]
- Fowler JF, Chappell RJ, Ritter MA. The prospects for new treatments for prostate cancer. *Int J Radiat Oncol Biol Phys* 2002; **52**: 3-5 [PMID: 11777616]
- Brenner DJ. Hypofractionation for prostate cancer radiotherapy--what are the issues? *Int J Radiat Oncol Biol Phys* 2003; **57**: 912-914 [PMID: 14575821]
- Bentzen SM, Saunders MI, Dische S, Bond SJ. Radiotherapy-related early morbidity in head and neck cancer: quantitative clinical radiobiology as deduced from the CHART trial. *Radiother Oncol* 2001; **60**: 123-135 [PMID: 11439207]
- Arcangeli G, Fowler J, Gomellini S, Arcangeli S, Saracino B, Petrongari MG, Benassi M, Strigari L. Acute and late toxicity in a randomized trial of conventional versus hypofractionated three-dimensional conformal radiotherapy for prostate cancer. *Int J Radiat Oncol Biol Phys* 2011; **79**: 1013-1021 [PMID: 20447774 DOI: 10.1016/j.ijrobp.2009.12.045]
- Goldberg PA, Nicholls RJ, Porter NH, Love S, Grimsey JE. Long-term results of a randomised trial of short-course low-dose adjuvant pre-operative radiotherapy for rectal cancer: reduction in local treatment failure. *Eur J Cancer* 1994; **30A**:

- 1602-1606 [PMID: 7530469]
- 31 **Kountouras J**, Zavos C. Recent advances in the management of radiation colitis. *World J Gastroenterol* 2008; **14**: 7289-7301 [PMID: 19109862]
 - 32 **Dearnaley DP**, Khoo VS, Norman AR, Meyer L, Nahum A, Tait D, Yarnold J, Horwich A. Comparison of radiation side-effects of conformal and conventional radiotherapy in prostate cancer: a randomised trial. *Lancet* 1999; **353**: 267-272 [PMID: 9929018 DOI: 10.1016/S0140-6736(98)05180-0]
 - 33 **O'Brien PC**. Radiation injury of the rectum. *Radiother Oncol* 2001; **60**: 1-14 [PMID: 11410298]
 - 34 **Kasibhatla M**, Clough RW, Montana GS, Oleson JR, Light K, Steffey BA, Jones EL. Predictors of severe gastrointestinal toxicity after external beam radiotherapy and interstitial brachytherapy for advanced or recurrent gynecologic malignancies. *Int J Radiat Oncol Biol Phys* 2006; **65**: 398-403 [PMID: 16542793 DOI: 10.1016/j.ijrobp.2005.12.008]
 - 35 **Waddell BE**, Rodriguez-Bigas MA, Lee RJ, Weber TK, Petrelli NJ. Prevention of chronic radiation enteritis. *J Am Coll Surg* 1999; **189**: 611-624 [PMID: 10589598]
 - 36 **Hauer-Jensen M**. Late radiation injury of the small intestine. Clinical, pathophysiologic and radiobiologic aspects. A review. *Acta Oncol* 1990; **29**: 401-415 [PMID: 2202341]
 - 37 **Morgan SC**, Waldron TS, Eapen L, Mayhew LA, Winquist E, Lukka H. Adjuvant radiotherapy following radical prostatectomy for pathologic T3 or margin-positive prostate cancer: a systematic review and meta-analysis. *Radiother Oncol* 2008; **88**: 1-9 [PMID: 18501455 DOI: 10.1016/j.radonc.2008.04.013]
 - 38 **Zurlo A**, Collette L, van Tienhoven G, Blank L, Warde P, Dubois J, Jeanneret W, Storme G, Bernier J, Kuten A, Pierart M, Bolla M. Acute toxicity of conventional radiation therapy for high-risk prostate cancer in EORTC trial 22863. *Eur Urol* 2002; **42**: 125-132 [PMID: 12160582]
 - 39 **Gérard JP**, Conroy T, Bonnetain F, Bouché O, Chapet O, Clouston-Dejardin MT, Untereiner M, Leduc B, Francois E, Maurel J, Seitz JF, Buecher B, Mackiewicz R, Ducreux M, Bedenne L. Preoperative radiotherapy with or without concurrent fluorouracil and leucovorin in T3-4 rectal cancers: results of FFCD 9203. *J Clin Oncol* 2006; **24**: 4620-4625 [PMID: 17008704 DOI: 10.1200/JCO.2006.06.7629]
 - 40 **Wilkes JD**. Prevention and treatment of oral mucositis following cancer chemotherapy. *Semin Oncol* 1998; **25**: 538-551 [PMID: 9783593]
 - 41 **Parulekar W**, Mackenzie R, Bjarnason G, Jordan RC. Scoring oral mucositis. *Oral Oncol* 1998; **34**: 63-71 [PMID: 9659522]
 - 42 **Jham BC**, Reis PM, Miranda EL, Lopes RC, Carvalho AL, Scheper MA, Freire AR. Oral health status of 207 head and neck cancer patients before, during and after radiotherapy. *Clin Oral Invest* 2008; **12**: 19-24 [PMID: 17876612 DOI: 10.1007/s00784-007-0149-5]
 - 43 **Werner-Wasik M**, Scott C, Graham ML, Smith C, Byhardt RW, Roach M, Andras EJ. Interfraction interval does not affect survival of patients with non-small cell lung cancer treated with chemotherapy and/or hyperfractionated radiotherapy: a multivariate analysis of 1076 RTOG patients. *Int J Radiat Oncol Biol Phys* 1999; **44**: 327-331 [PMID: 10760427]
 - 44 **Rakovitch E**, Fyles AW, Pintilie M, Leung PM. Role of mitomycin C in the development of late bowel toxicity following chemoradiation for locally advanced carcinoma of the cervix. *Int J Radiat Oncol Biol Phys* 1997; **38**: 979-987 [PMID: 9276362]
 - 45 **Yamada T**. Textbook of Gastroenterology. In: Cohen SBS, editor. Radiation injury in the gastrointestinal tract. 4 ed. Lippincott: Williams & Wilkins, 2003: 2760-2771
 - 46 **Heimbürger DK**, Shewach DS, Lawrence TS. The effect of fluorodeoxyuridine on sublethal damage repair in human colon cancer cells. *Int J Radiat Oncol Biol Phys* 1991; **21**: 983-987 [PMID: 1833363]
 - 47 **Bruso CE**, Shewach DS, Lawrence TS. Fluorodeoxyuridine-induced radiosensitization and inhibition of DNA double strand break repair in human colon cancer cells. *Int J Radiat Oncol Biol Phys* 1990; **19**: 1411-1417 [PMID: 2148170]
 - 48 **Andreyev J**. Gastrointestinal complications of pelvic radiotherapy: are they of any importance? *Gut* 2005; **54**: 1051-1054 [PMID: 16009675 DOI: 10.1136/gut.2004.062596]
 - 49 **Pico JL**, Avila-Garavito A, Naccache P. Mucositis: Its Occurrence, Consequences, and Treatment in the Oncology Setting. *Oncologist* 1998; **3**: 446-451 [PMID: 10388137]
 - 50 **Ahn SJ**, Kahn D, Zhou S, Yu X, Hollis D, Shafman TD, Marks LB. Dosimetric and clinical predictors for radiation-induced esophageal injury. *Int J Radiat Oncol Biol Phys* 2005; **61**: 335-347 [PMID: 15667951 DOI: 10.1016/j.ijrobp.2004.06.014]
 - 51 **Piquet MA**, Ozsahin M, Larpin I, Zouhair A, Coti P, Monney M, Monnier P, Mirimanoff RO, Roulet M. Early nutritional intervention in oropharyngeal cancer patients undergoing radiotherapy. *Support Care Cancer* 2002; **10**: 502-504 [PMID: 12353130 DOI: 10.1007/s00520-002-0364-1]
 - 52 **Raykher A**, Russo L, Schattner M, Schwartz L, Scott B, Shike M. Enteral nutrition support of head and neck cancer patients. *Nutr Clin Pract* 2007; **22**: 68-73 [PMID: 17242458]
 - 53 **Chowhan NM**. Injurious effects of radiation on the esophagus. *Am J Gastroenterol* 1990; **85**: 115-120 [PMID: 2405641]
 - 54 **Singh AK**, Lockett MA, Bradley JD. Predictors of radiation-induced esophageal toxicity in patients with non-small-cell lung cancer treated with three-dimensional conformal radiotherapy. *Int J Radiat Oncol Biol Phys* 2003; **55**: 337-341 [PMID: 12527046]
 - 55 **Patel AB**, Edelman MJ, Kwok Y, Krasna MJ, Suntharalingam M. Predictors of acute esophagitis in patients with non-small-cell lung carcinoma treated with concurrent chemotherapy and hyperfractionated radiotherapy followed by surgery. *Int J Radiat Oncol Biol Phys* 2004; **60**: 1106-1112 [PMID: 15519781 DOI: 10.1016/j.ijrobp.2004.04.051]
 - 56 **Qiao WB**, Zhao YH, Zhao YB, Wang RZ. Clinical and dosimetric factors of radiation-induced esophageal injury: radiation-induced esophageal toxicity. *World J Gastroenterol* 2005; **11**: 2626-2629 [PMID: 15849822]
 - 57 **Werner-Wasik M**, Yorke E, Deasy J, Nam J, Marks LB. Radiation dose-volume effects in the esophagus. *Int J Radiat Oncol Biol Phys* 2010; **76**: S86-S93 [PMID: 20171523 DOI: 10.1016/j.ijrobp.2009.05.070]
 - 58 **Henriksson R**, Bergström P, Franzén L, Lewin F, Wagenius G. Aspects on reducing gastrointestinal adverse effects associated with radiotherapy. *Acta Oncol* 1999; **38**: 159-164 [PMID: 10227436]
 - 59 **Coia LR**, Myerson RJ, Tepper JE. Late effects of radiation therapy on the gastrointestinal tract. *Int J Radiat Oncol Biol Phys* 1995; **31**: 1213-1236 [PMID: 7713784]
 - 60 **Emami B**, Lyman J, Brown A, Coia L, Goitein M, Munzenrider JE, Shank B, Solin LJ, Wesson M. Tolerance of normal tissue to therapeutic irradiation. *Int J Radiat Oncol Biol Phys* 1991; **21**: 109-122 [PMID: 2032882]
 - 61 **Kavanagh BD**, Pan CC, Dawson LA, Das SK, Li XA, Ten Haken RK, Miften M. Radiation dose-volume effects in the stomach and small bowel. *Int J Radiat Oncol Biol Phys* 2010; **76**: S101-S107 [PMID: 20171503 DOI: 10.1016/j.ijrobp.2009.05.071]
 - 62 **Lawrence TS**, Robertson JM, Anscher MS, Jirtle RL, Ensinger WD, Fajardo LF. Hepatic toxicity resulting from cancer treatment. *Int J Radiat Oncol Biol Phys* 1995; **31**: 1237-1248 [PMID: 7713785]
 - 63 **Dawson LA**, Ten Haken RK. Partial volume tolerance of the liver to radiation. *Semin Radiat Oncol* 2005; **15**: 279-283 [PMID: 16183482 DOI: 10.1016/j.semradonc.2005.04.005]
 - 64 **Yamasaki SA**, Marn CS, Francis IR, Robertson JM, Lawrence TS. High-dose localized radiation therapy for treatment of hepatic malignant tumors: CT findings and their relation to radiation hepatitis. *AJR Am J Roentgenol* 1995; **165**: 79-84 [PMID: 7785638]
 - 65 **Cheng JC**, Liu HS, Wu JK, Chung HW, Jan GJ. Inclusion of biological factors in parallel-architecture normal-tissue

- complication probability model for radiation-induced liver disease. *Int J Radiat Oncol Biol Phys* 2005; **62**: 1150-1156 [PMID: 15990021 DOI: 10.1016/j.ijrobp.2004.12.031]
- 66 **Dawson LA**, Normolle D, Balter JM, McGinn CJ, Lawrence TS, Ten Haken RK. Analysis of radiation-induced liver disease using the Lyman NTCP model. *Int J Radiat Oncol Biol Phys* 2002; **53**: 810-821 [PMID: 12095546]
 - 67 **Pan CC**, Kavanagh BD, Dawson LA, Li XA, Das SK, Miften M, Ten Haken RK. Radiation-associated liver injury. *Int J Radiat Oncol Biol Phys* 2010; **76**: S94-100 [PMID: 20171524 DOI: 10.1016/j.ijrobp.2009.06.092]
 - 68 **Perez CA**, Grigsby PW, Lockett MA, Chao KS, Williamson J. Radiation therapy morbidity in carcinoma of the uterine cervix: dosimetric and clinical correlation. *Int J Radiat Oncol Biol Phys* 1999; **44**: 855-866 [PMID: 10386643]
 - 69 **Miller AR**, Martenson JA, Nelson H, Schleck CD, Ilstrup DM, Gunderson LL, Donohue JH. The incidence and clinical consequences of treatment-related bowel injury. *Int J Radiat Oncol Biol Phys* 1999; **43**: 817-825 [PMID: 10098437]
 - 70 **Martin E**, Pointreau Y, Roche-Forestier S, Barillot I. [Normal tissue tolerance to external beam radiation therapy: small bowel]. *Cancer Radiother* 2010; **14**: 350-353 [PMID: 20598616 DOI: 10.1016/j.canrad.2010.03.013]
 - 71 **Keys HM**, Bundy BN, Stehman FB, Muderspach LI, Chafe WE, Suggs CL, Walker JL, Gersell D. Cisplatin, radiation, and adjuvant hysterectomy compared with radiation and adjuvant hysterectomy for bulky stage IB cervical carcinoma. *N Engl J Med* 1999; **340**: 1154-1161 [PMID: 10202166 DOI: 10.1056/NEJM199904153401503]
 - 72 **Classen J**, Belka C, Paulsen F, Budach W, Hoffmann W, Bamberg M. Radiation-induced gastrointestinal toxicity. Pathophysiology, approaches to treatment and prophylaxis. *Strahlenther Onkol* 1998; **174** Suppl 3: 82-84 [PMID: 9830465]
 - 73 **Hauer-Jensen M**, Wang J, Boerma M, Fu Q, Denham JW. Radiation damage to the gastrointestinal tract: mechanisms, diagnosis, and management. *Curr Opin Support Palliat Care* 2007; **1**: 23-29 [PMID: 18660720 DOI: 10.1097/SPC.0b013e3281108014]
 - 74 **Yeoh E**, Horowitz M, Russo A, Muecke T, Robb T, Maddox A, Chatterton B. Effect of pelvic irradiation on gastrointestinal function: a prospective longitudinal study. *Am J Med* 1993; **95**: 397-406 [PMID: 8213872]
 - 75 **Sher ME**, Bauer J. Radiation-induced enteropathy. *Am J Gastroenterol* 1990; **85**: 121-128 [PMID: 2301333]
 - 76 **Hauer-Jensen M**, Wang J, Denham JW. Bowel injury: current and evolving management strategies. *Semin Radiat Oncol* 2003; **13**: 357-371 [PMID: 12903023]
 - 77 **Ooi BS**, Tjandra JJ, Green MD. Morbidities of adjuvant chemotherapy and radiotherapy for resectable rectal cancer: an overview. *Dis Colon Rectum* 1999; **42**: 403-418 [PMID: 10223765]
 - 78 **Birgisson H**, Pahlman L, Gunnarsson U, Glimelius B. Late adverse effects of radiation therapy for rectal cancer - a systematic overview. *Acta Oncol* 2007; **46**: 504-516 [PMID: 17497318 DOI: 10.1080/02841860701348670]
 - 79 **Holm T**, Singnomklao T, Rutqvist LE, Cedermark B. Adjuvant preoperative radiotherapy in patients with rectal carcinoma. Adverse effects during long term follow-up of two randomized trials. *Cancer* 1996; **78**: 968-976 [PMID: 8780533 DOI: 10.1002/(SICI)1097-0142(19960901)78:]
 - 80 **Galland RB**, Spencer J. The natural history of clinically established radiation enteritis. *Lancet* 1985; **1**: 1257-1258 [PMID: 2860452]
 - 81 **Nussbaum ML**, Campana TJ, Weese JL. Radiation-induced intestinal injury. *Clin Plast Surg* 1993; **20**: 573-580 [PMID: 8324995]
 - 82 **Schultheiss TE**, Lee WR, Hunt MA, Hanlon AL, Peter RS, Hanks GE. Late GI and GU complications in the treatment of prostate cancer. *Int J Radiat Oncol Biol Phys* 1997; **37**: 3-11 [PMID: 9054871]
 - 83 **Babb RR**. Radiation proctitis: a review. *Am J Gastroenterol* 1996; **91**: 1309-1311 [PMID: 8677984]
 - 84 **Hauer-Jensen M**, Fink LM, Wang J. Radiation injury and the protein C pathway. *Crit Care Med* 2004; **32**: S325-S330 [PMID: 15118539]
 - 85 **Abayomi J**, Kirwan J, Hackett A. The prevalence of chronic radiation enteritis following radiotherapy for cervical or endometrial cancer and its impact on quality of life. *Eur J Oncol Nurs* 2009; **13**: 262-267 [PMID: 19640788 DOI: 10.1016/j.ejon.2009.02.007]
 - 86 **Gami B**, Harrington K, Blake P, Dearnaley D, Tait D, Davies J, Norman AR, Andreyev HJ. How patients manage gastrointestinal symptoms after pelvic radiotherapy. *Aliment Pharmacol Ther* 2003; **18**: 987-994 [PMID: 14616164]
 - 87 **Michalski JM**, Gay H, Jackson A, Tucker SL, Deasy JO. Radiation dose-volume effects in radiation-induced rectal injury. *Int J Radiat Oncol Biol Phys* 2010; **76**: S123-S129 [PMID: 20171506 DOI: 10.1016/j.ijrobp.2009.03.078]
 - 88 **Bosset JF**, Collette L, Calais G, Mineur L, Maingon P, Radoscovic-Jelic L, Daban A, Bardet E, Beny A, Ollier JC. Chemotherapy with preoperative radiotherapy in rectal cancer. *N Engl J Med* 2006; **355**: 1114-1123 [PMID: 16971718 DOI: 10.1056/NEJMoa060829]
 - 89 **Eifel PJ**, Levenback C, Wharton JT, Oswald MJ. Time course and incidence of late complications in patients treated with radiation therapy for FIGO stage IB carcinoma of the uterine cervix. *Int J Radiat Oncol Biol Phys* 1995; **32**: 1289-1300 [PMID: 7635768]
 - 90 **Denton AS**, Bond SJ, Matthews S, Bentzen SM, Maher EJ. National audit of the management and outcome of carcinoma of the cervix treated with radiotherapy in 1993. *Clin Oncol (R Coll Radiol)* 2000; **12**: 347-353 [PMID: 11202086]
 - 91 **Denham JW**, O'Brien PC, Dunstan RH, Johansen J, See A, Hamilton CS, Bydder S, Wright S. Is there more than one late radiation proctitis syndrome? *Radiother Oncol* 1999; **51**: 43-53 [PMID: 10386716]
 - 92 **Turini M**, Redaelli A, Gramegna P, Radice D. Quality of life and economic considerations in the management of prostate cancer. *Pharmacoeconomics* 2003; **21**: 527-541 [PMID: 12751912]
 - 93 **Bloch S**, Love A, Macvean M, Duchesne G, Couper J, Kissane D. Psychological adjustment of men with prostate cancer: a review of the literature. *Biopsychosoc Med* 2007; **1**: 2 [PMID: 17371571 DOI: 10.1186/1751-0759-1-2]
 - 94 **Henderson A**, Andreyev HJ, Stephens R, Dearnaley D. Patient and physician reporting of symptoms and health-related quality of life in trials of treatment for early prostate cancer: considerations for future studies. *Clin Oncol (R Coll Radiol)* 2006; **18**: 735-743 [PMID: 17168208]
 - 95 **Lev EL**, Eller LS, Gejerman G, Lane P, Owen SV, White M, Nganga N. Quality of life of men treated with brachytherapies for prostate cancer. *Health Qual Life Outcomes* 2004; **2**: 28 [PMID: 15198803 DOI: 10.1186/1477-7525-2-28]
 - 96 **Gelblum DY**, Potters L. Rectal complications associated with transperineal interstitial brachytherapy for prostate cancer. *Int J Radiat Oncol Biol Phys* 2000; **48**: 119-124 [PMID: 10924980]
 - 97 **Theodorescu D**, Gillenwater JY, Koutrouvelis PG. Prostatourethral-rectal fistula after prostate brachytherapy. *Cancer* 2000; **89**: 2085-2091 [PMID: 11066049 DOI: 10.1002/1097-0142(20001115)89:]
 - 98 **Shakespeare D**, Mitchell DM, Carey BM, Finan P, Henry AM, Ash D, Bottomley DM, Al-Qaisieh B. Recto-urethral fistula following brachytherapy for localized prostate cancer. *Colorectal Dis* 2007; **9**: 328-331 [PMID: 17432984 DOI: 10.1111/j.1463-1318.2006.01119.x]
 - 99 **Moon K**, Stukenborg GJ, Keim J, Theodorescu D. Cancer incidence after localized therapy for prostate cancer. *Cancer* 2006; **107**: 991-998 [PMID: 16878323 DOI: 10.1002/cncr.22083]
 - 100 **Travis LB**, Curtis RE, Storm H, Hall P, Holowaty E, Van

- Leeuwen FE, Kohler BA, Pukkala E, Lynch CF, Andersson M, Bergfeldt K, Clarke EA, Wiklund T, Stoter G, Gospodarowicz M, Sturgeon J, Fraumeni JF, Boice JD. Risk of second malignant neoplasms among long-term survivors of testicular cancer. *J Natl Cancer Inst* 1997; **89**: 1429-1439 [PMID: 9326912]
- 101 **Liauw SL**, Sylvester JE, Morris CG, Blasko JC, Grimm PD. Second malignancies after prostate brachytherapy: incidence of bladder and colorectal cancers in patients with 15 years of potential follow-up. *Int J Radiat Oncol Biol Phys* 2006; **66**: 669-673 [PMID: 16887293 DOI: 10.1016/j.ijrobp.2006.05.016]
 - 102 **Bentzen SM**, Overgaard M, Thames HD. Fractionation sensitivity of a functional endpoint: impaired shoulder movement after post-mastectomy radiotherapy. *Int J Radiat Oncol Biol Phys* 1989; **17**: 531-537 [PMID: 2506157]
 - 103 **Merrick GS**, Butler WM, Galbreath RW, Stipetich RL, Abel LJ, Lief JH. Erectile function after permanent prostate brachytherapy. *Int J Radiat Oncol Biol Phys* 2002; **52**: 893-902 [PMID: 11958881]
 - 104 **Honoré HB**, Bentzen SM, Møller K, Grau C. Sensori-neural hearing loss after radiotherapy for nasopharyngeal carcinoma: individualized risk estimation. *Radiother Oncol* 2002; **65**: 9-16 [PMID: 12413669]
 - 105 **Chon BH**, Loeffler JS. The effect of nonmalignant systemic disease on tolerance to radiation therapy. *Oncologist* 2002; **7**: 136-143 [PMID: 11961197]
 - 106 **Isselbacher KBEH**. Principles of Internal Medicine. New York: McGraw-Hill Company, 1994: 1922-1974
 - 107 **Rubin EFJ**. Pathology. Philadelphia: J.B. Lippincott Company, 1994: 651-686
 - 108 **Herold DM**, Hanlon AL, Hanks GE. Diabetes mellitus: a predictor for late radiation morbidity. *Int J Radiat Oncol Biol Phys* 1999; **43**: 475-479 [PMID: 10078625]
 - 109 **Matthews RH**. Collagen vascular disease and irradiation. *Int J Radiat Oncol Biol Phys* 1989; **17**: 1123-1124 [PMID: 2808050]
 - 110 **Hareyama M**, Nagakura H, Tamakawa M, Hyodo K, Asakura K, Horikoshi T, Oouchi A, Shido M, Morita K. Severe reaction after chemoradiotherapy of nasopharyngeal carcinoma with collagen disease. *Int J Radiat Oncol Biol Phys* 1995; **33**: 971 [PMID: 7591915]
 - 111 **Abu-Shakra M**, Lee P. Exaggerated fibrosis in patients with systemic sclerosis (scleroderma) following radiation therapy. *J Rheumatol* 1993; **20**: 1601-1603 [PMID: 8164225]
 - 112 **Tiersten A**, Saltz LB. Influence of inflammatory bowel disease on the ability of patients to tolerate systemic fluorouracil-based chemotherapy. *J Clin Oncol* 1996; **14**: 2043-2046 [PMID: 8683234]
 - 113 **Grann A**, Wallner K. Prostate brachytherapy in patients with inflammatory bowel disease. *Int J Radiat Oncol Biol Phys* 1998; **40**: 135-138 [PMID: 9422569]
 - 114 **Willett CG**, Ooi CJ, Zietman AL, Menon V, Goldberg S, Sands BE, Podolsky DK. Acute and late toxicity of patients with inflammatory bowel disease undergoing irradiation for abdominal and pelvic neoplasms. *Int J Radiat Oncol Biol Phys* 2000; **46**: 995-998 [PMID: 10705022]
 - 115 **Green S**, Stock RG, Greenstein AJ. Rectal cancer and inflammatory bowel disease: natural history and implications for radiation therapy. *Int J Radiat Oncol Biol Phys* 1999; **44**: 835-840 [PMID: 10386640]
 - 116 **Ali RA**, Dooley C, Comber H, Newell J, Egan LJ. Clinical features, treatment, and survival of patients with colorectal cancer with or without inflammatory bowel disease. *Clin Gastroenterol Hepatol* 2011; **9**: 584-9.e1-584-9.e2 [PMID: 21565283 DOI: 10.1016/j.cgh.2011.04.016]
 - 117 **Cohn SM**, Schloemann S, Tessner T, Seibert K, Stenson WF. Crypt stem cell survival in the mouse intestinal epithelium is regulated by prostaglandins synthesized through cyclooxygenase-1. *J Clin Invest* 1997; **99**: 1367-1379 [PMID: 9077547 DOI: 10.1172/JCI119296]
 - 118 **Du XX**, Doerschuk CM, Orazi A, Williams DA. A bone marrow stromal-derived growth factor, interleukin-11, stimulates recovery of small intestinal mucosal cells after cytoablative therapy. *Blood* 1994; **83**: 33-37 [PMID: 8274749]
 - 119 **Houchen CW**, George RJ, Sturmoski MA, Cohn SM. FGF-2 enhances intestinal stem cell survival and its expression is induced after radiation injury. *Am J Physiol* 1999; **276**: G249-G258 [PMID: 9887002]
 - 120 **Houchen CW**, Stenson WF, Cohn SM. Disruption of cyclooxygenase-1 gene results in an impaired response to radiation injury. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G858-G865 [PMID: 11052981]
 - 121 **Neta R**. Modulation of radiation damage by cytokines. *Stem Cells* 1997; **15** Suppl 2: 87-94 [PMID: 9368290 DOI: 10.1002/stem.5530150713]
 - 122 **Neta R**, Stiefel SM, Ali N. In lethally irradiated mice interleukin-12 protects bone marrow but sensitizes intestinal tract to damage from ionizing radiation. *Ann N Y Acad Sci* 1995; **762**: 274-280; discussion 280-281 [PMID: 7545367]
 - 123 **Neta R**, Stiefel SM, Finkelman F, Herrmann S, Ali N. IL-12 protects bone marrow from and sensitizes intestinal tract to ionizing radiation. *J Immunol* 1994; **153**: 4230-4237 [PMID: 7930625]
 - 124 **Robertson JM**, Clarke DH, Pevzner MM, Matter RC. Breast conservation therapy. Severe breast fibrosis after radiation therapy in patients with collagen vascular disease. *Cancer* 1991; **68**: 502-508 [PMID: 1648431]
 - 125 **Nisce LZ**, Safai B. Radiation therapy of Kaposi's sarcoma in AIDS. Memorial Sloan-Kettering experience. *Front Radiat Ther Oncol* 1985; **19**: 133-137 [PMID: 3920122]
 - 126 **Cooper JS**, Fried PR. Defining the role of radiation therapy in the management of epidemic Kaposi's sarcoma. *Int J Radiat Oncol Biol Phys* 1987; **13**: 35-39 [PMID: 2433259]
 - 127 **Formenti SC**, Chak L, Gill P, Buess EM, Hill CK. Increased radiosensitivity of normal tissue fibroblasts in patients with acquired immunodeficiency syndrome (AIDS) and with Kaposi's sarcoma. *Int J Radiat Biol* 1995; **68**: 411-412 [PMID: 7594966]
 - 128 **Baeyens A**, Slabbert JP, Willem P, Jozela S, Van Der Merwe D, Vral A. Chromosomal radiosensitivity of HIV positive individuals. *Int J Radiat Biol* 2010; **86**: 584-592 [PMID: 20545573 DOI: 10.3109/09553001003734576]
 - 129 **Ebell MH**, Siwek J, Weiss BD, Woolf SH, Susman J, Ewigman B, Bowman M. Strength of recommendation taxonomy (SORT): a patient-centered approach to grading evidence in the medical literature. *J Am Board Fam Pract* 2004; **17**: 59-67 [PMID: 15014055]
 - 130 **Housri N**, Yarchoan R, Kaushal A. Radiotherapy for patients with the human immunodeficiency virus: are special precautions necessary? *Cancer* 2010; **116**: 273-283 [PMID: 20014399 DOI: 10.1002/cncr.24878]
 - 131 **Coleman CN**. Molecular biology in radiation oncology. Radiation oncology perspective of BRCA1 and BRCA2. *Acta Oncol* 1999; **38** Suppl 13: 55-59 [PMID: 10612497]
 - 132 **Trenz K**, Rothfuss A, Schütz P, Speit G. Mutagen sensitivity of peripheral blood from women carrying a BRCA1 or BRCA2 mutation. *Mutat Res* 2002; **500**: 89-96 [PMID: 11890937]
 - 133 **Andreassen CN**. Can risk of radiotherapy-induced normal tissue complications be predicted from genetic profiles? *Acta Oncol* 2005; **44**: 801-815 [PMID: 16332587 DOI: 10.1080/02841860500374513]
 - 134 **West C**, Rosenstein BS, Alsner J, Azria D, Barnett G, Begg A, Bentzen S, Burnet N, Chang-Claude J, Chuang E, Coles C, De Ruyck K, De Ruysscher D, Dunning A, Elliott R, Fachal L, Hall J, Haustermans K, Herskind C, Hoelscher T, Imai T, Iwakawa M, Jones D, Kulich C, Langendijk JH, O'Neils P, Ozsahin M, Parliament M, Polanski A, Rosenstein B, Seminars D, Symonds P, Talbot C, Thierens H, Vega A, West C, Yarnold J. Establishment of a Radiogenomics Consortium. *Int J Radiat Oncol Biol Phys* 2010; **76**: 1295-1296 [PMID: 20338472 DOI: 10.1016/j.ijrobp.2009.12.017]

- 135 **Turesson I**, Bernefors R, Book M, Flogegård M, Hermansson I, Johansson KA, Lindh A, Sigurdardottir S, Thunberg U, Nyman J. Normal tissue response to low doses of radiotherapy assessed by molecular markers—a study of skin in patients treated for prostate cancer. *Acta Oncol* 2001; **40**: 941-951 [PMID: 11845959]
- 136 **Ho AY**, Atencio DP, Peters S, Stock RG, Formenti SC, Cesaretti JA, Green S, Haffty B, Drumea K, Leitzin L, Kuten A, Azria D, Ozsahin M, Overgaard J, Andreassen CN, Trop CS, Park J, Rosenstein BS. Genetic predictors of adverse radiotherapy effects: the Gene-PARE project. *Int J Radiat Oncol Biol Phys* 2006; **65**: 646-655 [PMID: 16751059 DOI: 10.1016/j.ijrobp.2006.03.006]
- 137 **Safwat A**, Bentzen SM, Turesson I, Hendry JH. Deterministic rather than stochastic factors explain most of the variation in the expression of skin telangiectasia after radiotherapy. *Int J Radiat Oncol Biol Phys* 2002; **52**: 198-204 [PMID: 11777639]
- 138 **Tucker SL**, Turesson I, Thames HD. Evidence for individual differences in the radiosensitivity of human skin. *Eur J Cancer* 1992; **28A**: 1783-1791 [PMID: 1389511]
- 139 **Tucker SL**, Geara FB, Peters LJ, Brock WA. How much could the radiotherapy dose be altered for individual patients based on a predictive assay of normal-tissue radiosensitivity? *Radiother Oncol* 1996; **38**: 103-113 [PMID: 8966222]
- 140 **Taylor AM**, Harnden DG, Arlett CF, Harcourt SA, Lehmann AR, Stevens S, Bridges BA. Ataxia telangiectasia: a human mutation with abnormal radiation sensitivity. *Nature* 1975; **258**: 427-429 [PMID: 1196376]
- 141 **Cesaretti JA**, Stock RG, Atencio DP, Peters SA, Peters CA, Burri RJ, Stone NN, Rosenstein BS. A genetically determined dose-volume histogram predicts for rectal bleeding among patients treated with prostate brachytherapy. *Int J Radiat Oncol Biol Phys* 2007; **68**: 1410-1416 [PMID: 17490827 DOI: 10.1016/j.ijrobp.2007.02.052]
- 142 **O'Driscoll M**, Jeggo PA. The role of double-strand break repair - insights from human genetics. *Nat Rev Genet* 2006; **7**: 45-54 [PMID: 16369571 DOI: 10.1038/nrg1746]
- 143 **Petrini JH**, Stracker TH. The cellular response to DNA double-strand breaks: defining the sensors and mediators. *Trends Cell Biol* 2003; **13**: 458-462 [PMID: 12946624]
- 144 **Taylor RC**, Cullen SP, Martin SJ. Apoptosis: controlled demolition at the cellular level. *Nat Rev Mol Cell Biol* 2008; **9**: 231-241 [PMID: 18073771 DOI: 10.1038/nrm2312]
- 145 **Branzei D**, Foiani M. Regulation of DNA repair throughout the cell cycle. *Nat Rev Mol Cell Biol* 2008; **9**: 297-308 [PMID: 18285803 DOI: 10.1038/nrm2351]
- 146 **Mikkelsen RB**, Wardman P. Biological chemistry of reactive oxygen and nitrogen and radiation-induced signal transduction mechanisms. *Oncogene* 2003; **22**: 5734-5754 [PMID: 12947383 DOI: 10.1038/sj.onc.1206663]
- 147 **Olsson AK**, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling - in control of vascular function. *Nat Rev Mol Cell Biol* 2006; **7**: 359-371 [PMID: 16633338 DOI: 10.1038/nrm1911]
- 148 **Weis SM**. Vascular permeability in cardiovascular disease and cancer. *Curr Opin Hematol* 2008; **15**: 243-249 [PMID: 18391792 DOI: 10.1097/MOH.0b013e3282f97d86]
- 149 **Andreassen CN**, Alsner J, Overgaard M, Overgaard J. Prediction of normal tissue radiosensitivity from polymorphisms in candidate genes. *Radiother Oncol* 2003; **69**: 127-135 [PMID: 14643949]
- 150 **De Ruyck K**, Van Eijkeren M, Claes K, Morthier R, De Paepe A, Vral A, Thierens H. Radiation-induced damage to normal tissues after radiotherapy in patients treated for gynecologic tumors: association with single nucleotide polymorphisms in XRCC1, XRCC3, and OGG1 genes and in vitro chromosomal radiosensitivity in lymphocytes. *Int J Radiat Oncol Biol Phys* 2005; **62**: 1140-1149 [PMID: 15990020 DOI: 10.1016/j.ijrobp.2004.12.027]
- 151 **De Ruyck K**, Van Eijkeren M, Claes K, Bacher K, Vral A, De Neve W, Thierens H. TGFbeta1 polymorphisms and late clinical radiosensitivity in patients treated for gynecologic tumors. *Int J Radiat Oncol Biol Phys* 2006; **65**: 1240-1248 [PMID: 16798416 DOI: 10.1016/j.ijrobp.2006.03.047]
- 152 **Damaraju S**, Murray D, Dufour J, Carandang D, Myrehaug S, Fallone G, Field C, Greiner R, Hanson J, Cass CE, Parliament M. Association of DNA repair and steroid metabolism gene polymorphisms with clinical late toxicity in patients treated with conformal radiotherapy for prostate cancer. *Clin Cancer Res* 2006; **12**: 2545-2554 [PMID: 16638864 DOI: 10.1158/1078-0432.CCR-05-2703]
- 153 **Merritt AJ**, Potten CS, Kemp CJ, Hickman JA, Balmain A, Lane DP, Hall PA. The role of p53 in spontaneous and radiation-induced apoptosis in the gastrointestinal tract of normal and p53-deficient mice. *Cancer Res* 1994; **54**: 614-617 [PMID: 8306319]
- 154 **Potten CS**, Merritt A, Hickman J, Hall P, Faranda A. Characterization of radiation-induced apoptosis in the small intestine and its biological implications. *Int J Radiat Biol* 1994; **65**: 71-78 [PMID: 7905913]
- 155 **Rieder F**, Brenmoehl J, Leeb S, Schölmerich J, Rogler G. Wound healing and fibrosis in intestinal disease. *Gut* 2007; **56**: 130-139 [PMID: 17172588 DOI: 10.1136/gut.2006.090456]
- 156 **Rogler G**, Brand K, Vogl D, Page S, Hofmeister R, Andus T, Kneuchel R, Baeuerle PA, Schölmerich J, Gross V. Nuclear factor kappaB is activated in macrophages and epithelial cells of inflamed intestinal mucosa. *Gastroenterology* 1998; **115**: 357-369 [PMID: 9679041]
- 157 **Hausmann M**, Spöttl T, Andus T, Rothe G, Falk W, Schölmerich J, Herfarth H, Rogler G. Subtractive screening reveals up-regulation of NADPH oxidase expression in Crohn's disease intestinal macrophages. *Clin Exp Immunol* 2001; **125**: 48-55 [PMID: 11472425]
- 158 **Rogler G**, Gelbmann CM, Vogl D, Brunner M, Schölmerich J, Falk W, Andus T, Brand K. Differential activation of cytokine secretion in primary human colonic fibroblast/myofibroblast cultures. *Scand J Gastroenterol* 2001; **36**: 389-398 [PMID: 11336164]
- 159 **Hovdenak N**, Fajardo LF, Hauer-Jensen M. Acute radiation proctitis: a sequential clinicopathologic study during pelvic radiotherapy. *Int J Radiat Oncol Biol Phys* 2000; **48**: 1111-1117 [PMID: 11072170]
- 160 **Stone HB**, Coleman CN, Anscher MS, McBride WH. Effects of radiation on normal tissue: consequences and mechanisms. *Lancet Oncol* 2003; **4**: 529-536 [PMID: 12965273]
- 161 **Denham JW**, Hauer-Jensen M. The radiotherapeutic injury—a complex 'wound'. *Radiother Oncol* 2002; **63**: 129-145 [PMID: 12063002]
- 162 **François S**, Mouisseddine M, Mathieu N, Semont A, Monti P, Dudoignon N, Saché A, Boutarfa A, Thierry D, Gourmelon P, Chapel A. Human mesenchymal stem cells favour healing of the cutaneous radiation syndrome in a xenogenic transplant model. *Ann Hematol* 2007; **86**: 1-8 [PMID: 17043780 DOI: 10.1007/s00277-006-0166-5]
- 163 **François S**, Bensidhoum M, Mouisseddine M, Mazurier C, Allenet B, Semont A, Frick J, Saché A, Bouchet S, Thierry D, Gourmelon P, Gorin NC, Chapel A. Local irradiation not only induces homing of human mesenchymal stem cells at exposed sites but promotes their widespread engraftment to multiple organs: a study of their quantitative distribution after irradiation damage. *Stem Cells* 2006; **24**: 1020-1029 [PMID: 16339642 DOI: 10.1634/stemcells.2005-0260]
- 164 **Haydont V**, Vozenin-Brotons MC. Maintenance of radiation-induced intestinal fibrosis: cellular and molecular features. *World J Gastroenterol* 2007; **13**: 2675-2683 [PMID: 17569135]
- 165 **Molla M**, Panes J. Radiation-induced intestinal inflammation. *World J Gastroenterol* 2007; **13**: 3043-3046 [PMID: 17589918]

- 166 **Earnest DLTJ**. Radiation enteritis and colitis. Gastrointestinal disease. Philadelphia: WB Saunders, 1989: 1369-1382
- 167 **Bentzen SM**. Preventing or reducing late side effects of radiation therapy: radiobiology meets molecular pathology. *Nat Rev Cancer* 2006; **6**: 702-713 [PMID: 16929324 DOI: 10.1038/nrc1950]
- 168 **Milliat F**, François A, Tamarat R, Benderitter M. [Role of endothelium in radiation-induced normal tissue damages]. *Ann Cardiol Angeiol (Paris)* 2008; **57**: 139-148 [PMID: 18579118 DOI: 10.1016/j.ancard.2008.02.015]
- 169 **Paris F**, Fuks Z, Kang A, Capodiec P, Juan G, Ehleiter D, Haimovitz-Friedman A, Cordon-Cardo C, Kolesnick R. Endothelial apoptosis as the primary lesion initiating intestinal radiation damage in mice. *Science* 2001; **293**: 293-297 [PMID: 11452123 DOI: 10.1126/science.1060191]
- 170 **Schuller BW**, Binns PJ, Riley KJ, Ma L, Hawthorne MF, Coderre JA. Selective irradiation of the vascular endothelium has no effect on the survival of murine intestinal crypt stem cells. *Proc Natl Acad Sci USA* 2006; **103**: 3787-3792 [PMID: 16505359 DOI: 10.1073/pnas.0600133103]
- 171 **Hahnfeldt P**, Panigrahy D, Folkman J, Hlatky L. Tumor development under angiogenic signaling: a dynamical theory of tumor growth, treatment response, and postvascular dormancy. *Cancer Res* 1999; **59**: 4770-4775 [PMID: 10519381]
- 172 **Bussink J**, Kaanders JH, van der Kogel AJ. Tumor hypoxia at the micro-regional level: clinical relevance and predictive value of exogenous and endogenous hypoxic cell markers. *Radiother Oncol* 2003; **67**: 3-15 [PMID: 12758235]
- 173 **Citrin D**, Ménard C, Camphausen K. Combining radiotherapy and angiogenesis inhibitors: clinical trial design. *Int J Radiat Oncol Biol Phys* 2006; **64**: 15-25 [PMID: 16377411 DOI: 10.1016/j.ijrobp.2005.03.065]
- 174 **Teicher BA**, Holden SA, Ara G, Dupuis NP, Liu F, Yuan J, Ikebe M, Kakeji Y. Influence of an anti-angiogenic treatment on 9L gliosarcoma: oxygenation and response to cytotoxic therapy. *Int J Cancer* 1995; **61**: 732-737 [PMID: 7768649]
- 175 **Lee CG**, Heijn M, di Tomaso E, Griffon-Etienne G, Ancukiewicz M, Koike C, Park KR, Ferrara N, Jain RK, Suit HD, Boucher Y. Anti-Vascular endothelial growth factor treatment augments tumor radiation response under normoxic or hypoxic conditions. *Cancer Res* 2000; **60**: 5565-5570 [PMID: 11034104]
- 176 **Griffin RJ**, Williams BW, Wild R, Cherrington JM, Park H, Song CW. Simultaneous inhibition of the receptor kinase activity of vascular endothelial, fibroblast, and platelet-derived growth factors suppresses tumor growth and enhances tumor radiation response. *Cancer Res* 2002; **62**: 1702-1706 [PMID: 11912143]
- 177 **Teicher BA**, Holden SA, Ara G, Sotomayor EA, Huang ZD, Chen YN, Brem H. Potentiation of cytotoxic cancer therapies by TNP-470 alone and with other anti-angiogenic agents. *Int J Cancer* 1994; **57**: 920-925 [PMID: 7515861]
- 178 **Mazeron R**, Anderson B, Supiot S, Paris F, Deutsch E. Current state of knowledge regarding the use of antiangiogenic agents with radiation therapy. *Cancer Treat Rev* 2011; **37**: 476-486 [PMID: 21546163 DOI: 10.1016/j.ctrv.2011.03.004]
- 179 **Prabhakar R**, Rath GK. A simple plan evaluation index based on the dose to critical structures in radiotherapy. *J Med Phys* 2011; **36**: 192-197 [PMID: 22228927 DOI: 10.4103/0971-6203.89965]
- 180 **Kehwar TS**. Analytical approach to estimate normal tissue complication probability using best fit of normal tissue tolerance doses into the NTCP equation of the linear quadratic model. *J Cancer Res Ther* 2005; **1**: 168-179 [PMID: 17998649]
- 181 **Choy H**, Akerley W, Safran H, Graziano S, Chung C, Williams T, Cole B, Kennedy T. Multiinstitutional phase II trial of paclitaxel, carboplatin, and concurrent radiation therapy for locally advanced non-small-cell lung cancer. *J Clin Oncol* 1998; **16**: 3316-3322 [PMID: 9779707]
- 182 **Antonadou D**, Coliarakis N, Synodinou M, Athanassiou H, Kouveli A, Verigos C, Georgakopoulos G, Panoussaki K, Karageorgis P, Throuvalas N. Randomized phase III trial of radiation treatment +/- amifostine in patients with advanced-stage lung cancer. *Int J Radiat Oncol Biol Phys* 2001; **51**: 915-922 [PMID: 11704311]
- 183 **Cosset JM**, Henry-Amar M, Burgers JM, Noordijk EM, Van der Werf-Messing B, Meerwaldt JH, van der Schueren E. Late radiation injuries of the gastrointestinal tract in the H2 and H5 EORTC Hodgkin's disease trials: emphasis on the role of exploratory laparotomy and fractionation. *Radiother Oncol* 1988; **13**: 61-68 [PMID: 3141982]
- 184 **Ogata K**, Hizawa K, Yoshida M, Kitamuro T, Akagi G, Kagawa K, Fukuda F. Hepatic injury following irradiation—a morphologic study. *Tokushima J Exp Med* 1963; **43**: 240-251 [PMID: 14049847]
- 185 **Jackson A**, Ten Haken RK, Robertson JM, Kessler ML, Kutcher GJ, Lawrence TS. Analysis of clinical complication data for radiation hepatitis using a parallel architecture model. *Int J Radiat Oncol Biol Phys* 1995; **31**: 883-891 [PMID: 7860402]
- 186 **Letschert JG**, Lebesque JV, Aleman BM, Bosset JF, Horiot JC, Bartelink H, Cionini L, Hamers JP, Leer JW, van Glabbeke M. The volume effect in radiation-related late small bowel complications: results of a clinical study of the EORTC Radiotherapy Cooperative Group in patients treated for rectal carcinoma. *Radiother Oncol* 1994; **32**: 116-123 [PMID: 7972904]
- 187 **Yeoh EE**, Botten R, Russo A, McGowan R, Fraser R, Roos D, Penniment M, Borg M, Sun W. Chronic effects of therapeutic irradiation for localized prostatic carcinoma on anorectal function. *Int J Radiat Oncol Biol Phys* 2000; **47**: 915-924 [PMID: 10863060]

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Gastrointestinal radiation injury: Prevention and treatment

Abobakr K Shadad, Frank J Sullivan, Joseph D Martin, Laurence J Egan

Abobakr K Shadad, Laurence J Egan, Department of Gastroenterology, University Hospital Galway, University Hospital Galway, 34562 Galway, Ireland

Abobakr K Shadad, Laurence J Egan, Department of Pharmacology and Therapeutics, School of Medicine, University Hospital Galway, 34562 Galway, Ireland

Frank J Sullivan, Joseph D Martin, Department of Radiation Oncology, National University of Ireland Galway, University Hospital Galway, 34562 Galway, Ireland

Author contributions: Shadad AK wrote the review; Sullivan FJ, Martin JD and Egan LJ revised the contents.

Correspondence to: Laurence J Egan, Professor, Department of Pharmacology and Therapeutics, School of Medicine, University Hospital Galway, 34562 Galway, Ireland. laurence.egan@nuigalway.ie

Telephone: +353-91-495370 Fax: +353-91-495572

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ferent agents have been used to prevent or minimize the severity of gastrointestinal injury induced by ionising radiation exposure, including biological, chemical and pharmacological agents. In this review we aim to discuss various technical strategies to prevent gastrointestinal injury during cancer radiotherapy, examine the different therapeutic options for acute and chronic gastrointestinal radiation injury and outline some examples of research directions and considerations for prevention at a pre-clinical level.

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Key words: Radiation enteritis; Radiation proctitis; Prevention; Treatment; Gastrointestinal radiation injury

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Abstract

With the recent advances in detection and treatment of cancer, there is an increasing emphasis on the efficacy and safety aspects of cancer therapy. Radiation therapy is a common treatment for a wide variety of cancers, either alone or in combination with other treatments. Ionising radiation injury to the gastrointestinal tract is a frequent side effect of radiation therapy and a considerable proportion of patients suffer acute or chronic gastrointestinal symptoms as a result. These side effects often cause morbidity and may in some cases lower the efficacy of radiotherapy treatment. Radiation injury to the gastrointestinal tract can be minimised by either of two strategies: technical strategies which aim to physically shift radiation dose away from the normal intestinal tissues, and biological strategies which aim to modulate the normal tissue response to ionising radiation or to increase its resistance to it. Although considerable improvement in the safety of radiotherapy treatment has been achieved through the use of modern optimised planning and delivery techniques, biological techniques may offer additional further promise. Dif-

INTRODUCTION

External beam radiotherapy is a common treatment modality for wide varieties of cancers. Approximately 70% of all cancer patients receive radiotherapy during the course of their disease, while radiotherapy plays a central role in 25% of all cancer cures^[1-2]. Radiotherapy is a cost effective treatment, accounting for only 5% of the total cancer care expenditure^[3]. This estimate has increased over the last decade with the wide use of modern technological innovations in simulation, delineation, dose calculation and radiation treatment delivery^[4].

Gastrointestinal radiation injury is an important problem for two main reasons. First, it causes morbidity associated with a significant economic burden^[5] and a significant reduction in patient's quality of life^[6,7]. Second, it limits the dose of radiation that can be used to control cancer, as radiotherapy-related side effects often limit the tolerability for patients. Clinical manifestations of gastrointestinal

radiation injury can present acutely during or soon following radiotherapy due to acute mucosal injury and inflammation or they can present insidiously within few months or years after radiotherapy due to a chronic process of transmural fibrosis and vascular sclerosis.

Radiation toxicity to the gastrointestinal tract can be reduced by either of two strategies: technical strategies, which aim to physically shift the radiation dose away from the normal tissues or through biological strategies which aim to modulate the cellular and tissue response to ionising radiation.

TECHNICAL STRATEGIES FOR PREVENTION

Attention to detail regarding both radiotherapy planning and radiation delivery methods can minimise the risk of intestinal toxicity during pelvic radiotherapy. Multiple field planning and delivery can reduce the radiation dose to normal intestine as well as minimise the volume of small intestine exposed to radiation in the pelvis^[8].

Physical manoeuvres and tissue expander techniques

The impact of different physical measures such as patient position during pelvic radiotherapy have been evaluated in a randomized trial of supine and prone positioning in patients undergoing conformal radiotherapy for prostate cancer. There were significant improvements for small bowel, rectal wall and bladder wall doses in the supine position^[9].

Another study utilised combined manoeuvres to displace the small intestine from the pelvis during radiotherapy by bladder distension, lower abdominal wall compression with the patient in prone position using two-field and four-field planning radiographs of contrast-enhanced small bowel^[10]. The results demonstrated in 50% of patients the four-field volume of pelvic small bowel was significantly less in the prone position than in the supine position. Similarly, other techniques such as the use of a “belly board” while the patient is in the prone position where the opening in the table allows the abdomen to fall below the level of the table, displaces the small bowel by the effect of the gravity during radiotherapy. These technique were found to reduce the volume of the small intestine within the pelvis by a mean of 66%^[11].

Pre-treatment small bowel contrast studies can assess the location and mobility of the small bowel and hence, determine the optimum treatment position that could minimise the volume of small intestine within the pelvis^[12]. Tissue expander techniques have been used to minimise the volume of exposed small bowel within the irradiation field during abdominal and pelvic external beam radiotherapy. Studies have shown a reduction of 50% in the risk of chronic intestinal complications with the use of intra-pelvic tissue expander^[13]. Other techniques used to minimise the radiation dose to the rectum include transperineal injection of human collagen to increase the distance between the prostate gland and anterior rectal wall.

The mean reduction in dose to the anterior rectal wall was 50% with no rectal toxicity reported^[14].

Prophylactic surgical techniques

Various prophylactic surgical therapies were also employed to reduce the small intestinal exposure during pelvic radiotherapy such as insertion of biodegradable mesh slings, intra-pelvic breast prostheses or omentoplasty during operative resection whenever postoperative radiotherapy may be indicated^[15-20]. Mesh slings were found to reduce the volume of small intestine exposed by 50%^[21,22] while other techniques such as pelvic reconstruction, omentoplasty and transposition of the large bowel were found to reduce the volume of bowel at risk by 60%^[22,23]. Prostate gland immobilization and rectal wall sparing has been suggested by the use of endorectal balloons during prostate cancer radiotherapy. Endorectal balloons are reported to be well tolerated by patients and showed a significant rectal wall sparing effect during three-dimensional conformal radiotherapy and intensity-modulated radiotherapy for prostate cancer^[24,25].

Optimised planning and delivery techniques

A significant improvement in the safety of radiotherapy treatment has been achieved through the use of optimised planning and delivery techniques which aim to reduce the field size, focus the radiation beam into the lesion and minimise the volume of exposed surrounding tissues in the radiation field^[10]. Multiple field arrangements may also reduce normal tissue toxicity during pelvic radiotherapy.

The dose-volume histogram is a plot of a cumulative dose-volume frequency distribution, that graphically summarizes the simulated radiation distribution within a volume of interest of a patient which would result from a proposed radiation treatment plan^[26]. It provides a graphic display of a simulated radiation treatment plan and generates valuable information on the dose distribution within the volume of interest^[27]. The dose-volume histogram is used to predict the development of radiation toxicity and to identify low and high risk patient groups^[28]. Careful review of the dose-volume histogram of both the intended targets as well as the structures of avoidance constitutes a standard part of the assessment of the radiation treatment plans in modern radiation therapy.

Three-dimensional conformal techniques which comprise computerised tomography scanning simulation techniques and intensity modulated radiation therapy are associated with a reduced risk of normal tissue toxicity and allow a higher radiation doses to be used compared to conventional two dimensional methods^[29-32]. Computerised tomography scan simulation techniques were found to reduce the volume of the bowel unintentionally irradiated by 5% compared to conventional radiotherapy^[33].

Intensity modulated radiation therapy allows clear definition of both target lesions and surrounding normal tissues and hence the ability to apply two different inten-

sities of radiation (high and low) within the same treatment field^[34,35]. It represents the ultimate combination of treatment planning and delivery which provides the most flexible delivery of radiation to complex targets while minimising radiation doses to surrounding critical structures. It does so by utilizing sophisticated planning algorithms such as inverse planning and multiple small beams of varying intensity, to optimally deliver the treatment. Intensity modulated radiation therapy was associated with a significant reduction in toxicity following pelvic radiotherapy in prostate cancer patients^[36]. It was also associated with a reduction of 15%-18% in intestinal volume unintentionally irradiated compared to conventional two dimensional radiotherapy, and a 10%-13% reduction in intestinal volume unintentionally irradiated compared to three-dimensional computerised tomography simulation techniques^[33]. Furthermore, intensity modulated radiation therapy has been found to reduce acute and late toxicity and maintains good long term quality of life even when high radiation doses are administered to prostate cancer patients^[37-39]. In 208 head and neck cancer patients treated with intensity modulated radiation therapy it was found that intensity modulated radiation therapy was associated with reduced late toxicities without jeopardising local cancer control and overall survival^[40]. The long-term quality of life has been compared among survivors of head and neck cancer patients whom were treated with either intensity modulated radiation therapy or three-dimensional conformal radiotherapy. The study results have shown that the early improvement in quality of life associated with intensity modulated radiation therapy was maintained and become more magnified over time^[41].

Brachytherapy is another advanced technique in which the source of radiation is implanted within the malignant tissues (interstitial brachytherapy) or within a cavity in its immediate vicinity (intra-cavitary brachytherapy). The sources can be permanently inserted and emits their radiation over a prolonged period of time (low dose rate) or temporary and emit over a short period (high dose rate). In this fashion, a high radiation dose is limited to the target lesion sparing the surrounding normal tissues. Brachytherapy alone or in combination with external beam radiotherapy can reduce normal tissue toxicity without compromising treatment efficacy in prostate cancer^[42,43]. Brachytherapy can be an option for patients with history of inflammatory bowel disease and it is well tolerated in low risk prostate carcinoma^[44].

Combining target definition on a daily basis and adjusting the treatments to account for target motion is referred to as image guided radiotherapy techniques^[45]. Stereotactic radiation therapy can focus an extremely narrow ionizing radiation beam on a small target from different directions using an immobilisation system. The high accuracy of this technique has enabled targeting a small lesions such as brain metastatic lesions^[46]. Stereotactic radiation therapy uses a small number of high doses of radiation to target small lesions. It has been used in

the treatment of early-stage non-small-cell lung cancer, prostate cancer, renal-cell carcinoma, and liver cancer, and in the treatment of oligometastases in the lung, liver, and spine^[47]. Stereotactic radiation therapy is associated with high local control rates of cancer^[48] and it was evaluated in the treatment of prostate cancer using an accelerated form of hypofractionation through fewer but larger treatment fractions. The early results of this new technique suggest that it may induce an initial prostate specific antigen response similar to that seen with conventional fractionation radiotherapy but with fewer acute side effects. Disease control and chronic toxicity have not yet been fully evaluated^[49]. The use of higher (often > 10 Gy) daily radiation doses in stereotactic radiation therapy needs to be carefully considered in view of the potential for more severe side effects^[50]. The impact of the image guided radiotherapy techniques has been shown in a large study with 331 patients treated with high-dose intensity modulated radiation therapy with fiducial marker-based position verification for prostate cancer which allowed daily correction of the prostate position using the fiducial markers. The results showed that high dose was well tolerated with lower rate of acute toxicity, which provide possibilities for further dose escalation^[51].

High-energy proton beams stop abruptly in the tissue at the end of their range and deposit most of their energy there, in this fashion, they provide dose distributions that are superior to X-rays when used in comparable beam configurations^[51]. In addition, they are more densely ionising along their treatment path, and therefore a higher quantum of energy to kill the cancer cell and high linear energy transfer. With proton beam radiation therapy a smaller volume of normal tissues is irradiated at high dose levels for most anatomic sites than is feasible with any photon technique^[52]. Clinical studies have shown that proton beam therapy in patients with hepatocellular carcinoma has been shown to be effective, safe and well tolerable^[53]. Proton beam therapy allows large tumour volumes to be irradiated to high doses without significant dose exposure to surrounding normal tissue which make it a promising modality for the treatment of large-volume tumours^[54]. However, they have yet to show significant advantages in comparison with the conventional radiation therapy.

MEDICAL THERAPY FOR GASTROINTESTINAL RADIATION INJURY

Although technical strategies have achieved a significant degree of normal tissue protection during radiotherapy delivery, biological strategies may offer additional future promise. A wide variety of pharmacological agents, nutritional supplements, biological response modifiers, and dietary measures have been investigated for potential benefit to prevent or minimize the severity of intestinal tissue injury induced by ionising radiation exposure.

SUPPORTIVE TREATMENT

Patient selection for radiotherapy

Patient risk profile for developing radiation toxicity merits a careful consideration while assessing individual patient for radiotherapy treatment. Co-morbid diseases such as vascular disease, hypertension, diabetes mellitus and atherosclerosis^[55], inflammatory bowel disease^[56-58], collagen vascular disease^[59] and human immunodeficiency virus (HIV) infection^[60,61] have been linked to an increased risk of toxicity following external beam radiotherapy.

Nutritional status and effect of diet during radiotherapy

Patient general health and nutritional status during radiotherapy may affect outcomes. Measures to improve patient nutrition such as dietary counselling, oral supplements and intensive nutrition intervention have been associated with an improved outcome in terms of body weight, morbidity, and quality of life during and after radiotherapy^[62,63]. McGough *et al.*^[64] reviewed the efficacy of nutritional intervention on bowel symptoms during pelvic radiotherapy in data from 2646 patients. They found no evidence base for nutritional interventions identified to mitigate bowel symptoms following radiotherapy. However, the role of probiotic supplements, low fat diet and elemental diet was recommended for further evaluation. Subsequent studies on elemental diet identified the difficulty of patients tolerance to large volumes of elemental diet regimen during radiotherapy period^[65,66]. The effect of high-potency probiotic preparation VSL3 has been investigated in double-blind, placebo-controlled trial of 490 pelvic radiotherapy patients. The results showed more diarrhoea and daily bowel movements in the placebo group compared with VSL3 recipients^[67].

Biological, chemical and pharmacological therapy

Biological, chemical or pharmacological interventions to ameliorate the effect of radiation injury on gastrointestinal tract can be categorized according to the time of administration with respect to radiation exposure. Agents administered prior to radiotherapy for normal tissue prophylaxis are described as radioprotectors. Agents administered during the course of radiotherapy to minimize the injury are called mitigators, while agents administered to ameliorate an established injury are called treatment^[68].

The complexity of the pathophysiology of intestinal radiation injury along with the increasing prevalence of the problem has attracted significant research interest in exploring the effectiveness of many agents. Treatment of delayed radiation injury is challenging as it is often refractory to different therapeutic modalities^[69].

At the pre-clinical level, there has been a significant interest to test the efficacy of different agents to modulate the biological process implicated in radiation induced tissue injury and normal tissue damage (Table 1).

At the clinical level, there are no formal trials to reliably assess the effectiveness of most of the suggested therapies; hence most of the evidence is obtained from

small studies which are often from a single centre. In addition, the results reported in many studies are often directly related to the radiation toxicity scale in use and its sensitivity, specificity as well as to methods of interpretation and grading of symptoms reported by patients.

Topical therapy

Topical therapy for radiation proctitis has been tried with different agents. Local instillation of formalin has been reported to be effective in the treatment of severe haemorrhagic radiation proctitis^[70]. Sodium butyrate enemas were reported to improve the acute symptoms of radiation proctitis with no impact on the incidence and severity of late proctitis^[71]. Steroid enemas and other topical steroid preparations, commonly prescribed for other anorectal inflammatory conditions have also been in use for symptom relief in radiation proctitis, but without conclusive evidence of efficacy.

Hyperbaric oxygen was suggested to exert its therapeutic role in chronic radiation proctitis through induction of neovascularization that reverses tissue hypoxia^[72,73] with trophic effect on vasculogenic stem cells^[74]. Treatment of chronic radiation proctitis with hyperbaric oxygen has shown promising results in a large randomized double-blind trial with 120 patients with long term follow up. Hyperbaric oxygen therapy significantly improved the healing responses in patients with refractory radiation proctitis with enhanced bowel-specific quality of life^[75].

A recent systematic review demonstrated the therapeutic benefit of hyperbaric oxygen based on the evidence and expert consensus opinion. The review demonstrated its efficacy in patients with radiation damage to the anus and rectum and refractory chronic radiation injury^[73].

Anti-inflammatory agents

Anti-inflammatory agents were suggested to reduce the severity of acute intestinal inflammation following irradiation. 5 aminosalicylic acid (5-ASA) has been studied, but most have reported no benefit or even worse symptoms with the use of 5-ASA compared to placebo^[76-79]. Sulfasalazine showed clinical improvement in symptoms in a case series of four patients^[80]. Balsalazide has been suggested to improve proctitis and toxicity grade in patients undergoing prostate cancer radiotherapy, in a study of 27 patients^[81]. Taken together, the clinical evidence on therapeutic role of 5-ASA are not sufficient to recommend routine use in radiation injury.

Antioxidants

Antioxidants were suggested to have cytoprotective effects by reducing cellular oxidative stress following radiation injury to intestinal tissue. A sustained therapeutic benefit after 4 wk therapy with vitamin E and C has been reported in a study involving twenty patients with chronic radiation proctitis^[82]. More studies will be required to assess the therapeutic role of different antioxidant agents.

Table 1 Examples of putative intestinal radioprotectants tested in pre-clinical studies

Biological agent	Suggested effect/mechanism	Suggested radioprotection
Glutamine, arginine enriched diet ^[103]	Enhanced mucosal healing	Protective effect on rat intestine compared to control
Vitamin- E ^[104]	Reduction of oxidative stress	Protective effect on rat intestine compared to control
Captopril ^[105]	Inhibition of pro-inflammatory enzyme angiotensin-1-converting enzyme	Protective effect on mouse intestine compared to control
Rofecoxib ^[106]	Selective Inhibition of cyclooxygenases -2 enzyme	Protective effect on rat intestine compared to control
Clopidogrel ^[107]	Inhibition of platelets aggregation with reduced vascular sclerosis	Protective effect on rat intestine
Thalidomide ^[108]	Protection to microvascular bed	Attenuated injury to rat endothelial cells of micro vascular bed compared to control
Simvastatin ^[109]	Attenuate endothelial cell injury	Attenuated delayed intestinal injury in rats
Glucagon-like peptide-2 (GLP-2) ^[110]	Increased mucosal mass	Intestinal trophic and protective effect to rat intestine
Octreotide ^[111,112]	Modulation the inflammatory effects mediated by over expression of NFκB	Ameliorated inflammation and injury in rat intestine
Prostaglandin E-2 ^[113]	Pro-proliferative and anti-apoptotic effect on intestinal epithelium	Increased crypts survival
Anti-Transforming growth factors beta receptor ^[114]	Biological inhibition of extracellular remodeling	Reduced intestinal injury and fibrosis compared to IgG treated control mice
Toll like receptor 5 agonist derived from Salmonella flagellin ^[115]	Activation of intestinal immune response via NFκB signalling pathway	Protective effect to mice intestine
Intestinal sterilisation ^[116]	Reduced number of translocated bacteria, sepsis and inflammation	Reduced intestinal cell apoptosis
Germ-free raised mice		
Probiotic bacteria ^[117] (<i>Lactobacillus species</i>)	Restored intestinal microbiota imbalance	Reduces intestinal damage, sepsis and death in rodents

Amifostine

Amifostine is a scavenger of reactive oxygen species. Its protective effects have been related to its ability to minimise the injurious effects of free radicals on intestinal cells^[83]. Amifostine has been investigated for both systemic and topical routes of administration during radiotherapy. Intravenous amifostine administered daily before radiotherapy has been shown to reduce the incidence of radiation proctitis during pelvic radiotherapy^[84,85]. Similarly, rectal suspension of amifostine in two doses (1 g in 18 patients and 2 g in 12 patients) administered daily before radiation therapy for prostate cancer has resulted in significant improvement in acute and late bowel quality of life toxicity parameters, more noticeable with higher doses^[86]. Despite the results of these studies and that of many others, the updated clinical practice guidelines for the prevention and treatment of mucositis published in 2007^[87] concluded that data on amifostine are mostly obtained from small, single-centre studies with conflicting results. The group concluded that intravenous amifostine daily administration prior to radiotherapy may prevent radiation proctitis in patients who are receiving standard-dose radiotherapy for rectal cancer (Level III evidence, grade B recommendation)^[87].

In previous studies, amifostine has been shown to reduce the incidence of xerostomia in patients during head and neck radiotherapy^[88]. The American Society for Clinical Oncology recommended its use in 2002 published guidelines^[89]. However, the recommendation has been subsequently withdrawn in the guidelines update in 2008 due to an insufficient data^[90].

Sucralfate

Protection of mucosal cells has been suggested with sucralfate (aluminium sucrose octasulfate). A beneficial effect of sucralfate has been previously reported in two double-blind placebo-controlled studies in patients who received pelvic radiotherapy. It reported that the frequency of defecation and stool consistency were improved by sucralfate^[91,92]. Sucralfate enemas have been evaluated in 26 patients with radiation-induced proctitis with persistent rectal bleeding and the results showed a reduction in the severity of rectal bleeding in all patients with sucralfate enemas after four weeks treatment^[93]. However, in another study the therapeutic effect of sucralfate rectal enemas were evaluated compared to placebo, both administered during the course of prostate cancer radiotherapy. The results showed no difference in the rate of rectal bleeding between sucralfate and placebo treated group after a median follow-up of five years^[94]. The therapeutic benefits of oral sucralfate have also been assessed in a prospective randomised placebo-controlled study on fifty one patients receiving pelvic radiotherapy. Results from 44 study subjects showed significantly increased diarrhoea in the sucralfate group which led to cessation of the trial. The study concluded that sucralfate cannot be recommended for prophylaxis of acute radiation to the rectum and may even worsen the symptoms^[95]. Similarly, the lack of effect of micronized sucralfate mouthwash has been reported in a randomized, controlled trial in radiation-induced oral mucositis patients^[96]. Despite extensive investigation, clinical data on sucralfate have shown variable results, which do not support its protective role

for routine clinical use.

ENDOSCOPIC MANAGEMENT, BIOPSY AND LASER CAUTERY

A variety of endoscopic therapies have been tried for the treatment of chronic radiation injury such as the heater probe, bipolar electrocoagulation and argon plasma coagulation with newer methods of endoscopic ablation such as radiofrequency ablation and cryotherapy^[97]. Endoscopic treatment has been used primarily aimed to treat bleeding rectal telengectasia. Argon plasma coagulation therapy has been shown to be effective and safe treatment for treatment of rectal bleeding resulting from chronic radiation injury^[98,99]. Caution should be applied however when considering the use of these invasive techniques following high dose treatment delivery. For instance the use of rectal wall biopsy and laser cautery has been associated with the development of recto-urinary fistulae^[100-102] in men treated with brachytherapy for prostate cancer.

CONCLUSION

Prevention of gastrointestinal toxicity starts with a thorough assessment of patient risk profile before radiotherapy. There is no prophylactic or therapeutic agent available to radiotherapy patients proven to mitigate the acute and chronic symptoms of gastrointestinal radiation injury or to allow safe radiation dose escalation for better control of cancer. Many therapeutic agents are in use, but often with little evidence base. The recent advances in radiotherapy planning and delivery techniques provide a considerable degree of protection to the gastrointestinal during external beam radiotherapy. Newer techniques such as image guided radiotherapy techniques, stereotactic therapy and proton beam therapy may confer additional protection. On the other hand, a better understanding of the pathophysiology of intestinal radiation injury will allow the development of more effective biological strategies that could increase the end organ resistance to radiation toxicity. In this regard, modulation of various cellular and cytokine pathways that have been implicated in the development of the acute pathological process of radiation injury can give future promise. In addition to targeting different mechanisms mediating chronic inflammatory and fibrotic process that underlie the delayed pathological changes in the gastrointestinal tract, such approaches may provide new therapeutics insights to this problem.

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REFERENCES

- 1 DeVita VT, Hellman SRS. Cancer: Principles and Practice of Oncology. Philadelphia: Lippincott Williams and

- Wilkins, 2005
- 2 Wang J, Boerma M, Fu Q, Hauer-Jensen M. Significance of endothelial dysfunction in the pathogenesis of early and delayed radiation enteropathy. *World J Gastroenterol* 2007; **13**: 3047-3055 [PMID: 17589919]
- 3 Ringborg U, Bergqvist D, Brorsson B, Cavallin-Ståhl E, Ceberg J, Einhorn N, Frödin JE, Järhult J, Lamnevik G, Lindholm C, Littbrand B, Norlund A, Nylén U, Rosén M, Svensson H, Möller TR. The Swedish Council on Technology Assessment in Health Care (SBU) systematic overview of radiotherapy for cancer including a prospective survey of radiotherapy practice in Sweden 2001--summary and conclusions. *Acta Oncol* 2003; **42**: 357-365 [PMID: 14596499]
- 4 Van de Werf E, Verstraete J, Lievens Y. The cost of radiotherapy in a decade of technology evolution. *Radiother Oncol* 2012; **102**: 148-153 [PMID: 21872955]
- 5 Sher DJ. Cost-effectiveness studies in radiation therapy. *Expert Rev Pharmacoecon Outcomes Res* 2010; **10**: 567-582 [PMID: 20950072 DOI: 10.1586/erp.10.51]
- 6 Bacon CG, Giovannucci E, Testa M, Kawachi I. The impact of cancer treatment on quality of life outcomes for patients with localized prostate cancer. *J Urol* 2001; **166**: 1804-1810 [PMID: 11586228]
- 7 Bacon CG, Giovannucci E, Testa M, Glass TA, Kawachi I. The association of treatment-related symptoms with quality-of-life outcomes for localized prostate carcinoma patients. *Cancer* 2002; **94**: 862-871 [PMID: 11857323 DOI: 10.1002/cncr.10248]
- 8 Waddell BE, Rodriguez-Bigas MA, Lee RJ, Weber TK, Petrelli NJ. Prevention of chronic radiation enteritis. *J Am Coll Surg* 1999; **189**: 611-624 [PMID: 10589598 DOI: S1072-7515(99)00199-4]
- 9 Bayley AJ, Catton CN, Haycocks T, Kelly V, Alasti H, Bristow R, Catton P, Crook J, Gospodarowicz MK, McLean M, Milosevic M, Warde P. A randomized trial of supine vs. prone positioning in patients undergoing escalated dose conformal radiotherapy for prostate cancer. *Radiother Oncol* 2004; **70**: 37-44 [PMID: 15036850 DOI: 10.1016/j.radonc.2003.08.007]
- 10 Gallagher MJ, Brereton HD, Rostock RA, Zero JM, Zekoski DA, Poyss LF, Richter MP, Kligerman MM. A prospective study of treatment techniques to minimize the volume of pelvic small bowel with reduction of acute and late effects associated with pelvic irradiation. *Int J Radiat Oncol Biol Phys* 1986; **12**: 1565-1573 [PMID: 3759581]
- 11 Shanahan TG, Mehta MP, Bertelrud KL, Buchler DA, Frank LE, Gehring MA, Kubsad SS, Utrie PC, Kinsella TJ. Minimization of small bowel volume within treatment fields utilizing customized "belly boards". *Int J Radiat Oncol Biol Phys* 1990; **19**: 469-476 [PMID: 2394624]
- 12 Green N, Iba G, Smith WR. Measures to minimize small intestine injury in the irradiated pelvis. *Cancer* 1975; **35**: 1633-1640 [PMID: 1148997]
- 13 Herbert SH, Solin LJ, Hoffman JP, Schultz DJ, Curran WJ, Lanciano RM, Rosenblum N, Hogan M, Eisenberg B, Hanks GE. Volumetric analysis of small bowel displacement from radiation portals with the use of a pelvic tissue expander. *Int J Radiat Oncol Biol Phys* 1993; **25**: 885-893 [PMID: 8478241]
- 14 Noyes WR, Hosford CC, Schultz SE. Human collagen injections to reduce rectal dose during radiotherapy. *Int J Radiat Oncol Biol Phys* 2012; **82**: 1918-1922 [PMID: 21514738]
- 15 Meric F, Hirschl RB, Mahboubi S, Womer RB, Goldwein J, Ross AJ, Schnauffer L. Prevention of radiation enteritis in children, using a pelvic mesh sling. *J Pediatr Surg* 1994; **29**: 917-921 [PMID: 7931970]
- 16 Tuech JJ, Chaudron V, Thoma V, Ollier JC, Tasseti V, Duval D, Rodier JF. Prevention of radiation enteritis by intrapelvic breast prosthesis. *Eur J Surg Oncol* 2004; **30**: 900-904 [PMID: 15336738 DOI: 10.1016/j.ejso.2004.06.012]
- 17 Waddell BE, Lee RJ, Rodriguez-Bigas MA, Weber TK, Pe-

- trelli NJ. Absorbable mesh sling prevents radiation-induced bowel injury during "sandwich" chemoradiation for rectal cancer. *Arch Surg* 2000; **135**: 1212-1217 [PMID: 11030884]
- 18 **Haider N**, O'Sullivan C, Corbally MT, Fitzgerald RJ. Abdominopelvic mesh compartmentalization reduces the complications of radiotherapy in children: a preliminary report. *Eur J Pediatr Surg* 2006; **16**: 348-351 [PMID: 17160781 DOI: 10.1055/s-2006-924521]
 - 19 **Soper JT**, Clarke-Pearson DL, Creasman WT. Absorbable synthetic mesh (910-polyglactin) intestinal sling to reduce radiation-induced small bowel injury in patients with pelvic malignancies. *Gynecol Oncol* 1988; **29**: 283-289 [PMID: 3345950]
 - 20 **Sezeur A**, Martella L, Abbou C, Gallot D, Schlienger M, Vibert JF, Touboul E, Martel P, Malafosse M. Small intestine protection from radiation by means of a removable adapted prosthesis. *Am J Surg* 1999; **178**: 22-25; discussion 25-26 [PMID: 10456697]
 - 21 **Dasmahapatra KS**, Swaminathan AP. The use of a biodegradable mesh to prevent radiation-associated small-bowel injury. *Arch Surg* 1991; **126**: 366-369 [PMID: 1847798]
 - 22 **Logmans A**, van Lent M, van Geel AN, Olofsen-Van Acht M, Koper PC, Wiggers T, Trimbos JB. The pedicled omentoplasty, a simple and effective surgical technique to acquire a safe pelvic radiation field; theoretical and practical aspects. *Radiother Oncol* 1994; **33**: 269-271 [PMID: 7716269]
 - 23 **Chen JS**, ChangChien CR, Wang JY, Fan HA. Pelvic peritoneal reconstruction to prevent radiation enteritis in rectal carcinoma. *Dis Colon Rectum* 1992; **35**: 897-901 [PMID: 1387358]
 - 24 **van Lin EN**, Hoffmann AL, van Kollenburg P, Leer JW, Visser AG. Rectal wall sparing effect of three different endorectal balloons in 3D conformal and IMRT prostate radiotherapy. *Int J Radiat Oncol Biol Phys* 2005; **63**: 565-576 [PMID: 16168848]
 - 25 **Woel R**, Beard C, Chen MH, Hurwitz M, Loffredo M, McMahon E, Ching J, Lopes L, D'Amico AV. Acute gastrointestinal, genitourinary, and dermatological toxicity during dose-escalated 3D-conformal radiation therapy (3DCRT) using an intrarectal balloon for prostate gland localization and immobilization. *Int J Radiat Oncol Biol Phys* 2005; **62**: 392-396 [PMID: 15890580]
 - 26 **Drzymala RE**, Mohan R, Brewster L, Chu J, Goitein M, Harms W, Urie M. Dose-volume histograms. *Int J Radiat Oncol Biol Phys* 1991; **21**: 71-78 [PMID: 2032898 DOI: 0360-3016(91)90168-4]
 - 27 **Niemierko A**, Goitein M. Dose-volume distributions: a new approach to dose-volume histograms in three-dimensional treatment planning. *Med Phys* 1994; **21**: 3-11 [PMID: 8164585]
 - 28 **Greco C**, Mazzetta C, Cattani F, Tosi G, Castiglioni S, Fodor A, Orecchia R. Finding dose-volume constraints to reduce late rectal toxicity following 3D-conformal radiotherapy (3D-CRT) of prostate cancer. *Radiother Oncol* 2003; **69**: 215-222 [PMID: 14643961]
 - 29 **Meyer J**, Czito B, Yin FF, Willett C. Advanced radiation therapy technologies in the treatment of rectal and anal cancer: intensity-modulated photon therapy and proton therapy. *Clin Colorectal Cancer* 2007; **6**: 348-356 [PMID: 17311699]
 - 30 **Meyer JJ**, Willett CG, Czito BG. Emerging role of intensity-modulated radiation therapy in anorectal cancer. *Expert Rev Anticancer Ther* 2008; **8**: 585-593 [PMID: 18402525 DOI: 10.1586/14737140.8.4.585]
 - 31 **Willett CG**. Technical advances in the treatment of patients with rectal cancer. *Int J Radiat Oncol Biol Phys* 1999; **45**: 1107-1108 [PMID: 10613301]
 - 32 **Melian E**, Mageras GS, Fuks Z, Leibel SA, Niehaus A, Lorant H, Zelefsky M, Baldwin B, Kutcher GJ. Variation in prostate position quantitation and implications for three-dimensional conformal treatment planning. *Int J Radiat Oncol Biol Phys* 1997; **38**: 73-81 [PMID: 9212007]
 - 33 **Nutting CM**, Convery DJ, Cosgrove VP, Rowbottom C, Padhani AR, Webb S, Dearnaley DP. Reduction of small and large bowel irradiation using an optimized intensity-modulated pelvic radiotherapy technique in patients with prostate cancer. *Int J Radiat Oncol Biol Phys* 2000; **48**: 649-656 [PMID: 11020560]
 - 34 **Ling CC**, Burman C, Chui CS, Kutcher GJ, Leibel SA, LoSasso T, Mohan R, Bortfeld T, Reinstein L, Spirou S, Wang XH, Wu Q, Zelefsky M, Fuks Z. Conformal radiation treatment of prostate cancer using inversely-planned intensity-modulated photon beams produced with dynamic multileaf collimation. *Int J Radiat Oncol Biol Phys* 1996; **35**: 721-730 [PMID: 8690638]
 - 35 **Fraass BA**, Kessler ML, McShan DL, Marsh LH, Watson BA, Dusseau WJ, Eisbruch A, Sandler HM, Lichter AS. Optimization and clinical use of multisegment intensity-modulated radiation therapy for high-dose conformal therapy. *Semin Radiat Oncol* 1999; **9**: 60-77 [PMID: 10196399]
 - 36 **Zelefsky MJ**, Fuks Z, Hunt M, Lee HJ, Lombardi D, Ling CC, Reuter VE, Venkatraman ES, Leibel SA. High dose radiation delivered by intensity modulated conformal radiotherapy improves the outcome of localized prostate cancer. *J Urol* 2001; **166**: 876-881 [PMID: 11490237]
 - 37 **Lips IM**, Dehnad H, van Gils CH, Boeken Kruger AE, van der Heide UA, van Vulpen M. High-dose intensity-modulated radiotherapy for prostate cancer using daily fiducial marker-based position verification: acute and late toxicity in 331 patients. *Radiat Oncol* 2008; **3**: 15 [PMID: 18495016]
 - 38 **Lips IM**, van Gils CH, van der Heide UA, Kruger AE, van Vulpen M. Health-related quality of life 3 years after high-dose intensity-modulated radiotherapy with gold fiducial marker-based position verification. *BJU Int* 2009; **103**: 762-767 [PMID: 18990145]
 - 39 **Marchand V**, Bourdin S, Charbonnel C, Rio E, Munos C, Campion L, Bonnaud-Antignac A, Lisbona A, Mahé MA, Supiot S. No impairment of quality of life 18 months after high-dose intensity-modulated radiotherapy for localized prostate cancer: a prospective study. *Int J Radiat Oncol Biol Phys* 2010; **77**: 1053-1059 [PMID: 19880259]
 - 40 **Toledano I**, Graff P, Serre A, Boisselier P, Bensadoun RJ, Ortholan C, Pommier P, Racadot S, Calais G, Alfonsi M, Favrel V, Giraud P, Lapeyre M. Intensity-modulated radiotherapy in head and neck cancer: results of the prospective study GORTEC 2004-03. *Radiother Oncol* 2012; **103**: 57-62 [PMID: 22296746]
 - 41 **Chen AM**, Farwell DG, Luu Q, Vazquez EG, Lau DH, Purdy JA. Intensity-Modulated Radiotherapy is Associated with Improved Global Quality of Life Among Long-Term Survivors of Head-and-Neck Cancer. *Int J Radiat Oncol Biol Phys* 2012; Epub ahead of print [PMID: 22300572]
 - 42 **Chen AB**, D'Amico AV, Neville BA, Earle CC. Patient and treatment factors associated with complications after prostate brachytherapy. *J Clin Oncol* 2006; **24**: 5298-5304 [PMID: 17114664]
 - 43 **Lee WR**, Bae K, Lawton C, Gillin M, Morton G, Firat S, Baikadi M, Kuettel M, Greven K, Sandler H. Late toxicity and biochemical recurrence after external-beam radiotherapy combined with permanent-source prostate brachytherapy: analysis of Radiation Therapy Oncology Group study 0019. *Cancer* 2007; **109**: 1506-1512 [PMID: 17340591 DOI: 10.1002/cncr.22560]
 - 44 **Peters CA**, Cesaretti JA, Stone NN, Stock RG. Low-dose rate prostate brachytherapy is well tolerated in patients with a history of inflammatory bowel disease. *Int J Radiat Oncol Biol Phys* 2006; **66**: 424-429 [PMID: 16887295]
 - 45 **Chung HT**, Xia P, Chan LW, Park-Somers E, Roach M. Does image-guided radiotherapy improve toxicity profile in whole pelvic-treated high-risk prostate cancer? Comparison between IG-IMRT and IMRT. *Int J Radiat Oncol Biol Phys* 2009; **73**: 53-60 [PMID: 18501530]
 - 46 **Ikushima H**. Radiation therapy: state of the art and the fu-

- ture. *J Med Invest* 2010; **57**: 1-11 [PMID: 20299738]
- 47 **Lo SS**, Fakiris AJ, Chang EL, Mayr NA, Wang JZ, Papiez L, Teh BS, McGarry RC, Cardenes HR, Timmerman RD. Stereotactic body radiation therapy: a novel treatment modality. *Nat Rev Clin Oncol* 2010; **7**: 44-54 [PMID: 19997074]
 - 48 **Dilling TJ**, Hoffe SE. Stereotactic body radiation therapy: transcending the conventional to improve outcomes. *Cancer Control* 2008; **15**: 104-111 [PMID: 18376377]
 - 49 **Buyyounouski MK**, Price RA, Harris EE, Miller R, Tomé W, Schefter T, Parsai EL, Konski AA, Wallner PE. Stereotactic body radiotherapy for primary management of early-stage, low- to intermediate-risk prostate cancer: report of the American Society for Therapeutic Radiology and Oncology Emerging Technology Committee. *Int J Radiat Oncol Biol Phys* 2010; **76**: 1297-1304 [PMID: 20338473 DOI: S0360-3016(09)03543-3]
 - 50 **Kavanagh BD**, Pan CC, Dawson LA, Das SK, Li XA, Ten Haken RK, Miften M. Radiation dose-volume effects in the stomach and small bowel. *Int J Radiat Oncol Biol Phys* 2010; **76**: S101-S107 [PMID: 20171503]
 - 51 **Loeffler JS**, Smith AR, Suit HD. The potential role of proton beams in radiation oncology. *Semin Oncol* 1997; **24**: 686-695 [PMID: 9422264]
 - 52 **Suit H**, Goldberg S, Niemierko A, Trofimov A, Adams J, Paganetti H, Chen GT, Bortfeld T, Rosenthal S, Loeffler J, Delaney T. Proton beams to replace photon beams in radical dose treatments. *Acta Oncol* 2003; **42**: 800-808 [PMID: 14968940]
 - 53 **Chiba T**, Tokuyue K, Matsuzaki Y, Sugahara S, Chuganji Y, Kagei K, Shoda J, Hata M, Abei M, Igaki H, Tanaka N, Akine Y. Proton beam therapy for hepatocellular carcinoma: a retrospective review of 162 patients. *Clin Cancer Res* 2005; **11**: 3799-3805 [PMID: 15897579]
 - 54 **Sugahara S**, Oshiro Y, Nakayama H, Fukuda K, Mizumoto M, Abei M, Shoda J, Matsuzaki Y, Thono E, Tokita M, Tsuboi K, Tokuyue K. Proton beam therapy for large hepatocellular carcinoma. *Int J Radiat Oncol Biol Phys* 2010; **76**: 460-466 [PMID: 19427743]
 - 55 **Yeoh E**, Horowitz M, Russo A, Muecke T, Robb T, Maddox A, Chatterton B. Effect of pelvic irradiation on gastrointestinal function: a prospective longitudinal study. *Am J Med* 1993; **95**: 397-406 [PMID: 8213872]
 - 56 **Matthews RH**. Collagen vascular disease and irradiation. *Int J Radiat Oncol Biol Phys* 1989; **17**: 1123-1124 [PMID: 2808050]
 - 57 **Hareyama M**, Nagakura H, Tamakawa M, Hyodo K, Asakura K, Horikoshi T, Oouchi A, Shido M, Morita K. Severe reaction after chemoradiotherapy of nasopharyngeal carcinoma with collagen disease. *Int J Radiat Oncol Biol Phys* 1995; **33**: 971 [PMID: 7591915]
 - 58 **Abu-Shakra M**, Lee P. Exaggerated fibrosis in patients with systemic sclerosis (scleroderma) following radiation therapy. *J Rheumatol* 1993; **20**: 1601-1603 [PMID: 8164225]
 - 59 **Chon BH**, Loeffler JS. The effect of nonmalignant systemic disease on tolerance to radiation therapy. *Oncologist* 2002; **7**: 136-143 [PMID: 11961197]
 - 60 **Nisce LZ**, Safai B. Radiation therapy of Kaposi's sarcoma in AIDS. Memorial Sloan-Kettering experience. *Front Radiat Ther Oncol* 1985; **19**: 133-137 [PMID: 3920122]
 - 61 **Cooper JS**, Fried PR. Defining the role of radiation therapy in the management of epidemic Kaposi's sarcoma. *Int J Radiat Oncol Biol Phys* 1987; **13**: 35-39 [PMID: 2433259]
 - 62 **Ravasco P**, Monteiro-Grillo I, Marques Vidal P, Camilo ME. Impact of nutrition on outcome: a prospective randomized controlled trial in patients with head and neck cancer undergoing radiotherapy. *Head Neck* 2005; **27**: 659-668 [PMID: 15920748 DOI: 10.1002/hed.20221]
 - 63 **Isenring EA**, Bauer JD, Capra S. Nutrition support using the American Dietetic Association medical nutrition therapy protocol for radiation oncology patients improves dietary intake compared with standard practice. *J Am Diet Assoc* 2007; **107**: 404-412 [PMID: 17324657]
 - 64 **McGough C**, Baldwin C, Frost G, Andreyev HJ. Role of nutritional intervention in patients treated with radiotherapy for pelvic malignancy. *Br J Cancer* 2004; **90**: 2278-2287 [PMID: 15162154 DOI: 10.1038/sj.bjc.6601868]
 - 65 **McGough C**, Baldwin C, Norman A, Frost G, Blake P, Tait D, Khoo V, Harrington K, Andreyev HJ. Is supplementation with elemental diet feasible in patients undergoing pelvic radiotherapy? *Clin Nutr* 2006; **25**: 109-116 [PMID: 16289498]
 - 66 **McGough C**, Wedlake L, Baldwin C, Hackett C, Norman AR, Blake P, Harrington K, Tait D, Khoo V, Frost G, Andreyev HJ. Clinical trial: normal diet vs. partial replacement with oral E028 formula for the prevention of gastrointestinal toxicity in cancer patients undergoing pelvic radiotherapy. *Aliment Pharmacol Ther* 2008; **27**: 1132-1139 [PMID: 18315590]
 - 67 **Delia P**, Sansotta G, Donato V, Frosina P, Messina G, De Renzis C, Famularo G. Use of probiotics for prevention of radiation-induced diarrhea. *World J Gastroenterol* 2007; **13**: 912-915 [PMID: 17352022]
 - 68 **Citrin D**, Cotrim AP, Hyodo F, Baum BJ, Krishna MC, Mitchell JB. Radioprotectors and mitigators of radiation-induced normal tissue injury. *Oncologist* 2010; **15**: 360-371 [PMID: 20413641]
 - 69 **Denton A**, Forbes A, Andreyev J, Maher EJ. Non surgical interventions for late radiation proctitis in patients who have received radical radiotherapy to the pelvis. *Cochrane Database Syst Rev* 2002; CD003455 [PMID: 11869662]
 - 70 **Chattopadhyay G**, Ray D, Chakravarty S, Mandal S. Formalin instillation for uncontrolled radiation induced haemorrhagic proctitis. *Trop Gastroenterol* 2010; **31**: 291-294 [PMID: 21568145]
 - 71 **Hille A**, Herrmann MK, Kertesz T, Christiansen H, Hermann RM, Pradier O, Schmidberger H, Hess CF. Sodium butyrate enemas in the treatment of acute radiation-induced proctitis in patients with prostate cancer and the impact on late proctitis. A prospective evaluation. *Strahlenther Onkol* 2008; **184**: 686-692 [PMID: 19107351 DOI: 10.1007/s00066-008-1896-1]
 - 72 **Fok TC**, Jan A, Peel SA, Evans AW, Clokie CM, Sándor GK. Hyperbaric oxygen results in increased vascular endothelial growth factor (VEGF) protein expression in rabbit calvarial critical-sized defects. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2008; **105**: 417-422 [PMID: 18206401]
 - 73 **Craighead P**, Shea-Budgell MA, Nation J, Esmail R, Evans AW, Parliament M, Oliver TK, Hagen NA. Hyperbaric oxygen therapy for late radiation tissue injury in gynecologic malignancies. *Curr Oncol* 2011; **18**: 220-227 [PMID: 21980249]
 - 74 **Milovanova TN**, Bhopale VM, Sorokina EM, Moore JS, Hunt TK, Hauer-Jensen M, Velazquez OC, Thom SR. Hyperbaric oxygen stimulates vasculogenic stem cell growth and differentiation in vivo. *J Appl Physiol* 2009; **106**: 711-728 [PMID: 19023021]
 - 75 **Clarke RE**, Tenorio LM, Hussey JR, Toklu AS, Cone DL, Hinojosa JG, Desai SP, Dominguez Parra L, Rodrigues SD, Long RJ, Walker MB. Hyperbaric oxygen treatment of chronic refractory radiation proctitis: a randomized and controlled double-blind crossover trial with long-term follow-up. *Int J Radiat Oncol Biol Phys* 2008; **72**: 134-143 [PMID: 18342453]
 - 76 **Baughan CA**, Canney PA, Buchanan RB, Pickering RM. A randomized trial to assess the efficacy of 5-aminosalicylic acid for the prevention of radiation enteritis. *Clin Oncol (R Coll Radiol)* 1993; **5**: 19-24 [PMID: 8424910]
 - 77 **Resbeut M**, Marteau P, Cowen D, Richaud P, Bourdin S, Dubois JB, Mere P, N'Guyen TD. A randomized double blind placebo controlled multicenter study of mesalazine for the prevention of acute radiation enteritis. *Radiother Oncol* 1997; **44**: 59-63 [PMID: 9288859]
 - 78 **Martenson JA**, Hyland G, Moertel CG, Mailliard JA, O'Fallon JR, Collins RT, Morton RF, Tewfik HH, Moore RL, Frank AR, Urias RE, Deming RL. Olsalazine is contraindicated during pelvic radiation therapy: results of a double-blind,

- randomized clinical trial. *Int J Radiat Oncol Biol Phys* 1996; **35**: 299-303 [PMID: 8635937]
- 79 **Sanguineti G**, Franzone P, Marcenaro M, Foppiano F, Vitale V. Sucralfate versus mesalazine versus hydrocortisone in the prevention of acute radiation proctitis during conformal radiotherapy for prostate carcinoma. A randomized study. *Strahlenther Onkol* 2003; **179**: 464-470 [PMID: 12835883 DOI: 10.1007/s00066-003-1082-4]
 - 80 **Goldstein F**, Khoury J, Thornton JJ. Treatment of chronic radiation enteritis and colitis with salicylazosulfapyridine and systemic corticosteroids. A pilot study. *Am J Gastroenterol* 1976; **65**: 201-208 [PMID: 7136]
 - 81 **Jahraus CD**, Bettenhausen D, Malik U, Sellitti M, St Clair WH. Prevention of acute radiation-induced proctosigmoiditis by balsalazide: a randomized, double-blind, placebo controlled trial in prostate cancer patients. *Int J Radiat Oncol Biol Phys* 2005; **63**: 1483-1487 [PMID: 16099600]
 - 82 **Kennedy M**, Bruninga K, Mutlu EA, Losurdo J, Choudhary S, Keshavarzian A. Successful and sustained treatment of chronic radiation proctitis with antioxidant vitamins E and C. *Am J Gastroenterol* 2001; **96**: 1080-1084 [PMID: 11316150]
 - 83 **Francois A**, Milliat F, Vozenin-Brotons MC. Bowel injury associated with pelvic radiotherapy. *Radiat Phys Chem* 2005; **72**: 399-407 [DOI: 10.1016/j.radphyschem.2004.04.140]
 - 84 **Liu T**, Liu Y, He S, Zhang Z, Kligerman MM. Use of radiation with or without WR-2721 in advanced rectal cancer. *Cancer* 1992; **69**: 2820-2825 [PMID: 1315211]
 - 85 **Athanassiou H**, Antonadou D, Coliarakis N, Kouveli A, Synodinou M, Paraskevidis M, Sarris G, Georgakopoulos GR, Panousaki K, Karageorgis P, Throuvalas N. Protective effect of amifostine during fractionated radiotherapy in patients with pelvic carcinomas: results of a randomized trial. *Int J Radiat Oncol Biol Phys* 2003; **56**: 1154-1160 [PMID: 12829154]
 - 86 **Simone NL**, Ménard C, Soule BP, Albert PS, Guion P, Smith S, Godette D, Crouse NS, Sciuto LC, Cooley-Zgela T, Camphausen K, Coleman CN, Singh AK. Intrarectal amifostine during external beam radiation therapy for prostate cancer produces significant improvements in Quality of Life measured by EPIC score. *Int J Radiat Oncol Biol Phys* 2008; **70**: 90-95 [PMID: 17855015]
 - 87 **Keefe DM**, Schubert MM, Elting LS, Sonis ST, Epstein JB, Raber-Durlacher JE, Migliorati CA, McGuire DB, Hutchins RD, Peterson DE. Updated clinical practice guidelines for the prevention and treatment of mucositis. *Cancer* 2007; **109**: 820-831 [PMID: 17236223 DOI: 10.1002/cncr.22484]
 - 88 **Antonadou D**, Pepelassi M, Synodinou M, Puglisi M, Throuvalas N. Prophylactic use of amifostine to prevent radiochemotherapy-induced mucositis and xerostomia in head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 2002; **52**: 739-747 [PMID: 11849797]
 - 89 **Schuchter LM**, Hensley ML, Meropol NJ, Winer EP. 2002 update of recommendations for the use of chemotherapy and radiotherapy protectants: clinical practice guidelines of the American Society of Clinical Oncology. *J Clin Oncol* 2002; **20**: 2895-2903 [PMID: 12065567]
 - 90 **Hensley ML**, Hagerty KL, Kewalramani T, Green DM, Meropol NJ, Wasserman TH, Cohen GI, Emami B, Gradishar WJ, Mitchell RB, Thigpen JT, Trotti A, von Hoff D, Schuchter LM. American Society of Clinical Oncology 2008 clinical practice guideline update: use of chemotherapy and radiation therapy protectants. *J Clin Oncol* 2009; **27**: 127-145 [PMID: 19018081]
 - 91 **Henriksson R**, Franzén L, Littbrand B. Effects of sucralfate on acute and late bowel discomfort following radiotherapy of pelvic cancer. *J Clin Oncol* 1992; **10**: 969-975 [PMID: 1588377]
 - 92 **Valls A**, Pestchen I, Prats C, Pera J, Aragón G, Vidarte M, Algara M. [Multicenter double-blind clinical trial comparing sucralfate vs placebo in the prevention of diarrhea secondary to pelvic irradiation]. *Med Clin (Barc)* 1999; **113**: 681-684 [PMID: 10650568]
 - 93 **Kochhar R**, Sriram PV, Sharma SC, Goel RC, Patel F. Natural history of late radiation proctosigmoiditis treated with topical sucralfate suspension. *Dig Dis Sci* 1999; **44**: 973-978 [PMID: 10235606]
 - 94 **O'Brien PC**, Franklin CI, Poulsen MG, Joseph DJ, Spry NS, Denham JW. Acute symptoms, not rectally administered sucralfate, predict for late radiation proctitis: longer term follow-up of a phase III trial--Trans-Tasman Radiation Oncology Group. *Int J Radiat Oncol Biol Phys* 2002; **54**: 442-449 [PMID: 12243820]
 - 95 **Hovdenak N**, Sørbye H, Dahl O. Sucralfate does not ameliorate acute radiation proctitis: randomised study and meta-analysis. *Clin Oncol (R Coll Radiol)* 2005; **17**: 485-491 [PMID: 16149294]
 - 96 **Dodd MJ**, Miaskowski C, Greenspan D, MacPhail L, Shih AS, Shiba G, Facione N, Paul SM. Radiation-induced mucositis: a randomized clinical trial of micronized sucralfate versus salt & soda mouthwashes. *Cancer Invest* 2003; **21**: 21-33 [PMID: 12643006]
 - 97 **Rustagi T**, Mashimo H. Endoscopic management of chronic radiation proctitis. *World J Gastroenterol* 2011; **17**: 4554-4562 [PMID: 22147960 DOI: 10.3748/wjg.v17.i41.4554]
 - 98 **de la Serna Higuera C**, Martín Arribas M, Rodríguez Gómez S, Pérez Villoria A, Martínez Moreno J, Betancourt González A. Efficacy and safety of argon plasma coagulation for the treatment of hemorrhagic radiation proctitis. *Rev Esp Enferm Dig* 2004; **96**: 758-764 [PMID: 15584849]
 - 99 **Latorre Sánchez M**, Sempere García-Argüelles J, Barceló Cerdá S, Huguet Malaves JM, Canelles Gamir P, Quiles Teodoro F, Medina Chuliá E. [Evaluation of the endoscopic response to argon plasma coagulation in patients with chronic radiation proctopathy]. *Rev Esp Enferm Dig* 2008; **100**: 619-624 [PMID: 19119787]
 - 100 **Gelblum DY**, Potters L. Rectal complications associated with transperineal interstitial brachytherapy for prostate cancer. *Int J Radiat Oncol Biol Phys* 2000; **48**: 119-124 [PMID: 10924980]
 - 101 **Theodorescu D**, Gillenwater JY, Koutrouvelis PG. Prostatourethral-rectal fistula after prostate brachytherapy. *Cancer* 2000; **89**: 2085-2091 [PMID: 11066049 DOI: 10.1002/1097-0142(20001115)89:]
 - 102 **Shakespeare D**, Mitchell DM, Carey BM, Finan P, Henry AM, Ash D, Bottomley DM, Al-Qaisieh B. Recto-urethral fistula following brachytherapy for localized prostate cancer. *Colorectal Dis* 2007; **9**: 328-331 [PMID: 17432984]
 - 103 **Ersin S**, Tuncyurek P, Esassolak M, Alkanat M, Buke C, Yilmaz M, Telefoncu A, Kose T. The prophylactic and therapeutic effects of glutamine- and arginine-enriched diets on radiation-induced enteritis in rats. *J Surg Res* 2000; **89**: 121-125 [PMID: 10729239 DOI: 10.1006/jsre.1999.5808]
 - 104 **Empey LR**, Papp JD, Jewell LD, Fedorak RN. Mucosal protective effects of vitamin E and misoprostol during acute radiation-induced enteritis in rats. *Dig Dis Sci* 1992; **37**: 205-214 [PMID: 1735337]
 - 105 **Yoon SC**, Park JM, Jang HS, Shinn KS, Bahk YW. Radioprotective effect of captopril on the mouse jejunal mucosa. *Int J Radiat Oncol Biol Phys* 1994; **30**: 873-878 [PMID: 7960990]
 - 106 **Keskek M**, Gocmen E, Kilic M, Gencturk S, Can B, Cengiz M, Okten RM, Koc M. Increased expression of cyclooxygenase-2 (COX-2) in radiation-induced small bowel injury in rats. *J Surg Res* 2006; **135**: 76-84 [PMID: 16780881]
 - 107 **Wang J**, Albertson CM, Zheng H, Fink LM, Herbert JM, Hauer-Jensen M. Short-term inhibition of ADP-induced platelet aggregation by clopidogrel ameliorates radiation-induced toxicity in rat small intestine. *Thromb Haemost* 2002; **87**: 122-128 [PMID: 11848440]
 - 108 **Kim KT**, Chae HS, Kim JS, Kim HK, Cho YS, Choi W, Choi KY, Rho SY, Kang SJ. Thalidomide effect in endothelial cell

- of acute radiation proctitis. *World J Gastroenterol* 2008; **14**: 4779-4783 [PMID: 18720539]
- 109 **Wang J**, Boerma M, Fu Q, Kulkarni A, Fink LM, Hauer-Jensen M. Simvastatin ameliorates radiation enteropathy development after localized, fractionated irradiation by a protein C-independent mechanism. *Int J Radiat Oncol Biol Phys* 2007; **68**: 1483-1490 [PMID: 17674978]
 - 110 **Torres S**, Thim L, Milliat F, Vozenin-Brotans MC, Olsen UB, Ahnfelt-Rønne I, Bourhis J, Benderitter M, François A. Glucagon-like peptide-2 improves both acute and late experimental radiation enteritis in the rat. *Int J Radiat Oncol Biol Phys* 2007; **69**: 1563-1571 [PMID: 18035212]
 - 111 **Wang J**, Zheng H, Hauer-Jensen M. Influence of Short-Term Octreotide Administration on Chronic Tissue Injury, Transforming Growth Factor beta (TGF-beta) Overexpression, and Collagen Accumulation in Irradiated Rat Intestine. *J Pharmacol Exp Ther* 2001; **297**: 35-42 [PMID: 11259525]
 - 112 **Olgaç V**, Erbil Y, Barbaros U, Oztetcan S, Giriş M, Kaya H, Bilge H, Güler S, Toker G. The efficacy of octreotide in pancreatic and intestinal changes: radiation-induced enteritis in animals. *Dig Dis Sci* 2006; **51**: 227-232 [PMID: 16416241 DOI: 10.1007/s10620-006-3113-3]
 - 113 **Stenson WF**. Prostaglandins and epithelial response to injury. *Curr Opin Gastroenterol* 2007; **23**: 107-110 [PMID: 17268236 DOI: 10.1097/MOG.0b013e3280143cb6]
 - 114 **Zheng H**, Wang J, Kotliansky VE, Gotwals PJ, Hauer-Jensen M. Recombinant soluble transforming growth factor beta type II receptor ameliorates radiation enteropathy in mice. *Gastroenterology* 2000; **119**: 1286-1296 [PMID: 11054386]
 - 115 **Burdelya LG**, Krivokrysenko VI, Tallant TC, Strom E, Gleiberman AS, Gupta D, Kurnasov OV, Fort FL, Osterman AL, Didonato JA, Feinstein E, Gudkov AV. An agonist of toll-like receptor 5 has radioprotective activity in mouse and primate models. *Science* 2008; **320**: 226-230 [PMID: 18403709]
 - 116 **Crawford PA**, Gordon JI. Microbial regulation of intestinal radiosensitivity. *Proc Natl Acad Sci USA* 2005; **102**: 13254-13259 [PMID: 16129828]
 - 117 **Ciorba MA**, Stenson WF. Probiotic therapy in radiation-induced intestinal injury and repair. *Ann N Y Acad Sci* 2009; **1165**: 190-194 [PMID: 19538306]

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Intestinal injury can be reduced by intra-arterial postischemic perfusion with hypertonic saline

Oleg Korniyushin, Michael Galagudza, Anna Kotslova, Gelfia Nutfullina, Nina Shved, Alexey Nevorotin, Valeriy Sedov, Timur Vlasov

Oleg Korniyushin, Anna Kotslova, Valeriy Sedov, Department of Surgery, I.P. Pavlov Federal Medical University, 197022 St. Petersburg, Russia

Michael Galagudza, Timur Vlasov, Department of Pathophysiology, I.P. Pavlov Federal Medical University, 197022 St. Petersburg, Russia

Michael Galagudza, Timur Vlasov, Institute of Experimental Medicine, V.A. Almazov Federal Heart, Blood and Endocrinology Centre, 197341 St. Petersburg, Russia

Gelfia Nutfullina, Nina Shved, Department of Pathology, I.P. Pavlov Federal Medical University, 197022 St. Petersburg, Russia

Alexey Nevorotin, Laboratory of Electron Microscopy, I.P. Pavlov Federal Medical University, 197022 St. Petersburg, Russia

Author contributions: Korniyushin O, Galagudza M and Kotslova A performed the experiments; Nutfullina G and Shved N performed histological analyses; Nevorotin A was involved in data analysis and manuscript editing; Sedov V and Vlasov T designed the study and wrote the manuscript.

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Correspondence to: Dr. Michael Galagudza, MD, Department of Pathophysiology, I.P. Pavlov Federal Medical University, 197022 St. Petersburg, Russia. galagudza@mail.ru

Telephone: +7-812-4997035 Fax: +7-812-4997069

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Abstract

AIM: To investigate the effect of local intestinal perfusion with hypertonic saline (HTS) on intestinal ischemia-reperfusion injury (IRI) in both *ex vivo* and *in vivo* rat models.

METHODS: All experiments were performed on male Wistar rats anesthetized with pentobarbital sodium given intraperitoneally at a dose of 60 mg/kg. *Ex vivo* vascularly perfused rat intestine was subjected to 60-min ischemia and either 30-min reperfusion with

isotonic buffer (controls), or 5 min with HTS of 365 or 415 mOsm/L osmolarity (HTS^{365mOsm} or HTS^{415mOsm}, respectively) followed by 25-min reperfusion with isotonic buffer. The vascular intestinal perfusate flow (IPF) rate was determined by collection of the effluent from the portal vein in a calibrated tube. Spontaneous intestinal contraction rate was monitored throughout. Irreversible intestinal injury or area of necrosis (AN) was evaluated histochemically using 2,3,5-triphenyltetrazolium chloride staining. *In vivo*, 30-min ischemia was followed by either 30-min blood perfusion or 5-min reperfusion with HTS^{365mOsm} through the superior mesenteric artery (SMA) followed by 25-min blood perfusion. Arterial blood pressure (BP) was measured in the common carotid artery using a miniature pressure transducer. Histological injury was evaluated in both preparations using the Chui score.

RESULTS: *Ex vivo*, intestinal IRI resulted in a reduction in the IPF rate during reperfusion ($P < 0.05$ vs sham). The postischemic recovery of the IPF rate did not differ between the controls and the HTS^{365mOsm} group. In the HTS^{415mOsm} group, postischemic IPF rates were lower than in the controls and the HTS^{365mOsm} group ($P < 0.05$). The intestinal contraction rate was similar at baseline in all groups. An increase in this parameter was observed during the first 10 min of reperfusion in the control group as compared to the sham-treated group, but no such increase was seen in the HTS^{365mOsm} group. In controls, AN averaged $14.8\% \pm 5.07\%$ of the total tissue volume. Administration of HTS^{365mOsm} for 5 min after 60-min ischemia resulted in decrease in AN ($5.1\% \pm 1.20\%$ vs controls, $P < 0.01$). However, perfusion of the intestine with the HTS of greater osmolarity (HTS^{415mOsm}) failed to protect the intestine from irreversible injury. The Chui score was lower in the HTS^{365mOsm} group in comparison with controls (2.4 ± 0.54 vs 3.2 ± 0.44 , $P = 0.042$), while intestinal perfusion with HTS^{415mOsm} failed to improve the Chui score. Intestinal reperfusion with HTS^{365mOsm} in

the *in vivo* series secured rapid recovery of BP after its transient fall, whereas in the controls no recovery was seen. The Chiu score was lower in the HTS^{365mOsm} group *vs* controls (3.1 ± 0.26 and 3.8 ± 0.22 , $P = 0.0079$ respectively), although the magnitude of the effect was lower than in the *ex vivo* series.

CONCLUSION: Brief intestinal postischemic perfusion with HTS^{365mOsm} through the SMA followed by blood flow restoration is a protective procedure that could be used for the prevention of intestinal IRI.

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Key words: Intestinal ischemia-reperfusion injury; Superior mesenteric artery; Perfusion; Hypertonic saline; Acute mesenteric ischemia; Intestinal perfusate flow rate

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INTRODUCTION

Acute mesenteric ischemia (AMI) is a life-threatening vascular emergency with a mortality rate as high as 60%-80%^[1]. It can be caused by local arterial obstruction, impairment of venous outflow, or a systemic hemodynamic disorder resulting in severe intestinal hypoxic injury. The rather non-specific clinical presentation of AMI can be an obstacle to its early recognition, thus delaying the proper treatment, namely revascularization^[2]. However, if AMI can be diagnosed unambiguously prior to the development of intestinal infarction, either embolectomy or the intra-arterial infusion of thrombolytic agents can materially improve the outcome^[3]. Unfortunately, even after timely revascularization, the positive effect can be compromised, at least in part, by the development of reperfusion injury, including edema, the no-reflow phenomenon, and the progressive death of mucosal cells^[4]. In experimental models, intestinal ischemia-reperfusion injury (IRI) has been shown to be alleviated by a variety of pre- and postconditioning procedures^[5,6], while various antioxidants, anesthetics, vasodilators, anti-inflammatory agents, and cytokine receptor blockers can also be helpful^[7,8]. Furthermore, the systemic administration of hypertonic saline (HTS) has given clearly positive results^[9,10]. The protective effect of HTS in these experiments is believed to be based on the following mechanisms: arteriolar vasodilation, osmotic stress-induced inhibition of inflammatory cells, and prevention of intestinal edema^[10-12]. It is to be noted that the application of either HTS or some of the above-mentioned drugs may have certain limitations, such as an increased risk of electrolyte imbalance^[12]

and the development of adverse drug reactions, in either case as a result of systemic administration. It has been hypothesized, therefore, that perfusion of the intestine with HTS through the superior mesenteric artery (SMA) might prove to be a more advantageous strategy, given its greater efficacy in delivering the hypertonic solution to the target.

The present study was an experimental investigation of the effect of local intra-arterial HTS administration during the early reperfusion phase on the functional and structural manifestations of intestinal injury. To the best of our knowledge, this approach has not yet been attempted.

MATERIALS AND METHODS

Chemicals and materials

2,3,5-triphenyltetrazolium chloride (TTC) was obtained from Sigma-Aldrich (St. Louis, MO, United States). Pentobarbital sodium was purchased from Apoteket (Umea, Sweden). Bovine serum albumin was obtained from Biowest (Nuaille, France). Dextran 60 000 was obtained from Biochemist (Saransk, Russian Federation). Other chemicals used for the preparation of both Krebs-Henseleit buffer and HTS were purchased from Acros Organics (Geel, Belgium).

Animals

All experiments were performed on male Wistar rats weighting 250-300 g. The animals were fed regular chow, and water was available *ad libitum*. The procedures were performed in accordance with the Guide for the Care and Use of Laboratory Animals and were approved by the local Ethics Committee. The animals were anesthetized with pentobarbital sodium given intraperitoneally at a dose of 60 mg/kg. Core body temperature was maintained at 37.0 ± 0.5 °C by a feedback-controlled heating pad (TCAT-2LV controller, PHYSITEMP Instruments Inc., United States).

Ex vivo experiments

Isolated perfused small intestine preparation: The intestine was isolated and perfused according to a previously described technique^[13]. Briefly, the abdominal cavity was opened by a midline incision. To prevent blood coagulation, 50 units of heparin sodium were injected into the inferior vena cava. The abdominal aorta was exposed from the level of the celiac trunk (CT) to the aortic bifurcation. The intestinal segment between the pyloric sphincter and the distal portion of the ileum was cut using an electrocautery unit. The branches of the SMA that supply the cecum and proximal part of the colon were exposed, ligated, and cut, followed by the removal of the proximal colon. The intestinal lumen was rinsed with warm (37 °C) saline. The aorta was cannulated through its infrarenal segment with subsequent advancement of the cannula tip to the SMA level. The aorta was then ligated immediately above the CT, and perfusion of the small

intestine with oxygenated (95% O₂ and 5% CO₂) Krebs-Henseleit buffer (KHB) was initiated through the SMA and CT at a pressure of 80 mmHg. KHB solution was modified to contain high molecular weight compounds (1.0% bovine serum albumin and 3.0% dextran 60 000) plus (all in mmol/L) 118.5 NaCl, 25.0 NaHCO₃, 4.7 KCl, 1.2 KH₂PO₄, 1.2 MgSO₄, 1.5 CaCl₂, 5.0 glucose, 2.0 lactate, 0.2 pyruvate. The portal vein was cannulated to ensure outflow of the perfusion fluid. After ligation of the splenic and gastric vessels, the exposed part of the intestine was placed into the water-jacketed bath filled with normal saline to maintain the temperature at 37 °C during the experiment. All surgical procedures described were performed within 50–60 min and were followed by a 20-min stabilization period.

Experimental protocol: After stabilization, ischemia of the intestine was induced by cessation of the perfusate flow for 60 min, followed by reperfusion. During the initial 5-min reperfusion, the perfusion was resumed with the modified KHB of either normal (physiological) or increased osmolality (315 vs 365 or 415 mOsm/L, designated as HTS^{365mOsm} or HTS^{415mOsm}, respectively), in each case delivered under a pressure of 80 mmHg, followed by perfusion with normotonic KHB for 25 min. The experiments were randomized and allocated to the following 4 groups: (1) Sham (*n* = 6): the intestine was continuously perfused for 110 min; (2) Controls (*n* = 6): 60-min intestinal ischemia, followed by 30-min reperfusion with normotonic KHB (osmolality 315 mOsm/L); (3) HTS^{365mOsm} (*n* = 5): 60-min intestinal ischemia, followed by 5-min reperfusion with HTS having an increased osmolality of 365 mOsm/L (KHB supplemented with 50 mmol NaCl), followed by 25-min reperfusion with normotonic KHB; and (4) HTS^{415mOsm} (*n* = 6): the same sequence of procedures as in the previous group, but using HTS with osmolality increased to 415 mOsm/L.

Assessment of vascular intestinal perfusate flow rate: The vascular intestinal perfusate flow (IPF) rate was determined each 5 min during both the stabilization and reperfusion periods by collection of the effluent from the portal vein in a calibrated tube.

Determination of luminal intestinal flow: The outflow from the intestinal lumen was determined in all experiments during the stabilization period. Luminal flow exceeding 0.5 mL/min is indicative of pathological intestinal secretion, which is usually due to ischemic injury of the intestine during surgical manipulations. Thus, the experiments with luminal flow exceeding 0.5 mL/min were excluded from the final analysis.

Histochemical analysis: We used the technique of visualizing the area of necrosis (AN) with 2,3,5-triphenyltetrazolium chloride (TTC), a method generally considered to be a “gold standard” for AN delineation in the heart in other studies, including our own^[14,15]. Briefly, immediately

after the end of reperfusion into the SMA, 10 mL of 0.5% TTC solution were administered for 30 s. Fifteen minutes after that, 3 equally spaced segments of the intestine with a total length of 3 cm were excised and fixed in 4% buffered formaldehyde solution for 24 h at room temperature. The samples were cut into 7–8 transverse slices, each 1.5 mm thick, which were photographed for further analysis using Adobe Photoshop CS. TTC-positive tissue, brick red in color, was considered to be viable, in contrast to the necrotic tissue, which had a pale appearance. The AN volume in each sample was calculated by multiplying the TTC-negative area by the slice thickness. The AN volume of the whole intestinal segment was calculated by summing the AN volumes of the slices and was expressed as the percentage of AN in relation to the total volume of the intestinal tissue being analyzed.

Evaluation of spontaneous intestinal motility: According to the original technique described by Hoffman *et al.*^[16], this parameter is assessed by recording the movement of fecal pellets down the intestine. We used a different approach to register the spontaneous intestinal contraction rate. In brief, a pair of soft rubber rings, hand-crafted from a black tube with a diameter close to that of the intestine, were cut at one point and carefully placed around the intestine to signify equally spaced anatomical locations throughout the experiments. Sixty-second video recordings (5 frames per second) were taken every 10 min during the stabilization and reperfusion periods and were digitized with VirtualDub software (Avery Lee, United States). Based on the distances between the rings, the values were plotted against time, followed by a Fourier transform and probability plotting of the frequency distribution. The data were presented as the rates of low-frequency intestinal contractions (Hz).

In vivo experiments

For the investigation of the effect of HTS perfusion on the intestinal IRI, we developed a technique for perfusion through the SMA, which, in our opinion, offers some advantages over the systemic administration of HTS used before. The infrarenal aorta was occluded with a microclip and cannulated with an 18G polyethylene cannula (Figure 1A). The micro clip was then removed and placed on the aorta between the SMA and the CT, after which the cannula was advanced up to the SMA level, where its tip was fixed by ligation (Figure 1B). The next step included 2 manipulations: the occlusion of the distal part of the superior mesenteric vein (SMV) and the incision of the ileocecal SMV branch to secure the outflow of the perfusate from the intestinal circulation. The oxygenated HTS maintained at 37 °C was delivered through the SMA with an infusion pump (SPACE infusion system, B. Brown, Germany) at a constant rate of 15 mL/min. The flow rate was selected on the basis of our pilot experiments showing that the mean blood flow in the SMA of animals of comparable weight, having blood pressure (BP) in the range between 100 and 120 mmHg,

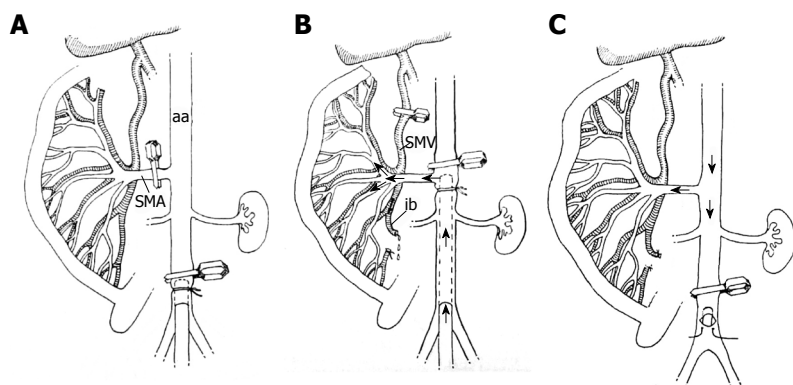


Figure 1 The sequential steps of the intestinal perfusion *in vivo*. A: The abdominal aorta (aa) is occluded by a clip, followed by cannulation of the vessel; the superior mesenteric artery (SMA) is clipped; B: The aortic clip is positioned above the SMA; the cannula is positioned at the level of SMA to allow its perfusion with hypertonic saline; the perfusate outflow is secured through the opened ileocecal branch (ib) of the superior mesenteric vein (SMV); the main trunk of SMV is clipped; C: The cannula is removed to allow blood flow through the SMA to resume simultaneously with the following procedures: the aortic clip is positioned as in Figure 1A; the clip occluding the SMV is removed, and bleeding from the ileocecal branch of the SMV is prevented by ligation.

was approximately 10% higher than 15 mL/min. Any appearance of blood in the perfusate was checked by visual inspection. At the end of the perfusion, the ileocecal SMV branch was ligated and the main trunk of the SMV was reopened. The aortic cannula was removed and the aortic incision closed with a suture (Figure 1C), so that blood flow was restored in both the aorta and the mesenteric vascular bed.

Experimental protocol: The animals were randomly allocated into 2 groups: (1) Controls ($n = 6$): intestinal ischemia by SMA occlusion for 30 min, followed by 30-min reperfusion with blood; and (2) HTS^{365mOsm} ($n = 5$): intestinal ischemia by SMA occlusion for 30 min, followed by 5-min reperfusion with HTS^{365mOsm}, followed by 25-min reperfusion with blood.

Blood pressure measurement: Arterial BP was measured in the common carotid artery using a miniature pressure transducer (Baxter, United States), to be monitored using PhysExp X4 software (Cardioprotect Ltd., St. Petersburg, Russian Federation). BP values were registered under the following conditions: at baseline, prior to SMA occlusion, 15 and 30 min after SMA occlusion, and at the 15th and 30th min of reperfusion.

Histological studies: At the end of reperfusion, intestinal samples were taken and routinely fixed in 10% buffered formaldehyde solution, embedded in paraffin, cut into 5 μ m sections, and stained with hematoxylin and eosin. The slides were analyzed under a light microscope by the pathologist blinded to the treatment mode used in each group. The results were scored from 0 to 5 according to the scale of Chiu *et al.*^[17]. The method is based on the well established relationship between the grade of intestinal injury, if any, and its structural manifestation (grade 0: normal villi; grade 1: development of subepithelial space; grade 2: moderate lifting of subepithelial layer; grade 3: massive epithelial lifting with a few tips denuded; grade 4: denuded villi with lamina propria and dilated capillaries

exposed; grade 5: digestion and disintegration of lamina propria with ulceration). In the present study, we rigorously followed this gradation as applied in similar trials by several groups^[10,18,19].

Statistical analysis

Data were analyzed with SPSS 11.0 (SPSS Inc. Software, Chicago, IL, United States). All values are expressed as mean \pm SD. Differences in the intestinal motility and IPF rate were tested by repeated-measures analysis of variance, followed by a Tukey post-hoc test. The non-parametric Mann-Whitney *U* test was used to determine the inter-group differences in AN value and histopathological score. $P \leq 0.05$ were considered significant.

RESULTS

Ex vivo experiments

Exclusions: Two animals were excluded from the final analysis (one in the control group and one in the HTS^{415mOsm} group) because the luminal flow exceeded 0.5 mL/min.

Intestinal perfusate flow rate: There were no inter-group differences in the IPF rate during the stabilization period (Figure 2). The IPF values remained unchanged over the entire 110-min perfusion period in the sham-treated group. In contrast, intestinal ischemia-reperfusion resulted in a reduction in the IPF rate during reperfusion ($P < 0.05$ vs sham). This reduction could have been due to IRI in the intestinal microcirculatory bed, resulting in no-re-flow. The postischemic recovery of the IPF rate did not differ between the controls and the HTS^{365mOsm} group. In the HTS^{415mOsm} group, postischemic IPF rates were lower than in the controls and the HTS^{365mOsm} group ($P < 0.05$).

Area of necrosis: TTC-negative intestinal tissue was not present in the samples obtained from the sham-treated group. In the controls, AN averaged $14.8\% \pm 5.07\%$ of the total tissue volume within the analyzed intestinal seg-

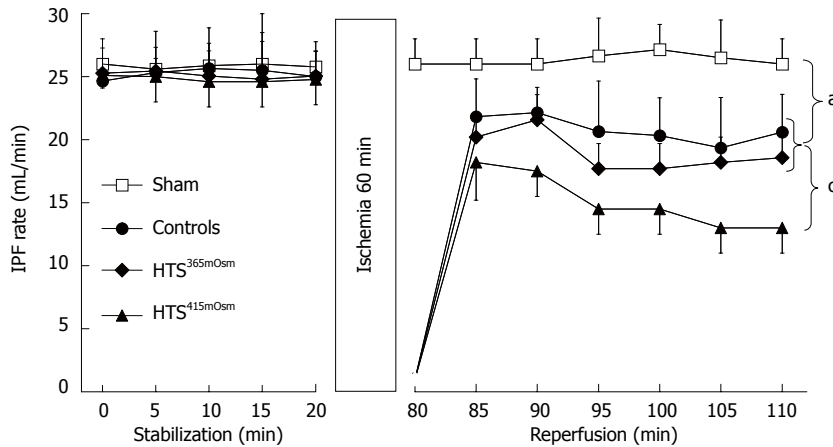


Figure 2 Intestinal perfusate flow rate in the *ex vivo* rat experiments with 60-min ischemia followed by varying conditions of reperfusion. The groups tested were: sham-operated (no ischemia at all, $n = 6$); controls (30 min reperfusion with normotonic buffer, $n = 5$); 2 groups with either HTS^{365mOsm} ($n = 5$) or HTS^{415mOsm} ($n = 5$), both subjected to 5-min reperfusion with the respective hypertonic solution followed by 25 min reperfusion with normotonic buffer. The pre-ischemic values of Intestinal perfusate flow (IPF) rate were similar in all groups. No changes were apparent in the shams over the whole period, whereas IPF values were lower in all the ischemia-reperfusion groups, although the groups showed different patterns. HTS^{415mOsm} resulted in the most prominent decrease in the IPF rate, with a further progressive fall to the end of the experiment. In contrast, IPF rate was less affected in the control and HTS^{365mOsm} groups, to a similar degree in both, while an appreciable degree of stabilization took place towards the end of the postischemic period. Postischemic IPF rate recovery did not differ between the controls and the HTS^{365mOsm} group, whereas postischemic IPF rate values were significantly lower in the HTS^{415mOsm} group, compared to both the controls and the HTS^{365mOsm} group. Data are mean \pm SD. ^a $P < 0.05$ vs sham; ^c $P < 0.05$ vs controls.

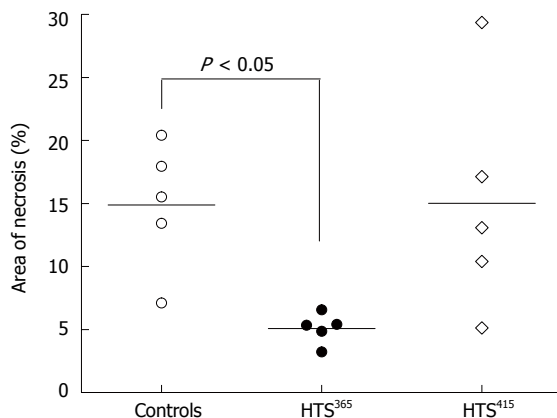


Figure 3 Area of necrosis within the isolated rat intestine after 60-min ischemia followed by the varying conditions of reperfusion. The area of necrosis (AN) is defined as the percentage of 2,3,5-triphenyltetrazolium chloride-negative in relation to the total intestinal tissue volume. Controls (reperfusion with normotonic saline) and the HTS^{415mOsm} group did not differ statistically regarding AN volume, whereas HTS^{365mOsm} administration resulted in a significantly lower value of AN. In both hypertonic saline (HTS) groups, the 5-min reperfusion with the appropriate hyperosmotic saline was followed by 25-min normotonic reperfusion. The data are presented as dot plots with median values.

ment (Figure 3). Administration of HTS^{365mOsm} for 5 min after 60-min ischemia resulted in decrease in AN ($5.1\% \pm 1.20\%$ vs controls, $P < 0.01$). However, perfusion of the intestine with the HTS of greater osmolarity (HTS^{415mOsm}) failed to protect the intestine from irreversible injury (AN = $15.0\% \pm 9.12\%$ vs controls, $P > 0.05$).

Intestinal motility: The rate of intestinal contraction-relaxation pulses, which was used as a measure of intestinal motility in the present study, was similar at baseline

in all groups (Figure 4). An increase in this parameter was observed during the first 10 min of reperfusion in the control group as compared to the sham-treated group, but no such increase was seen in the HTS^{365mOsm} group.

Histological data: Histological data from the intestinal samples obtained after 30-min reperfusion *ex vivo* are shown in Figure 5A. Sham-treated animals had normal or near-normal histology. The Chiu score was lower in the HTS^{365mOsm} group in comparison with controls (2.4 ± 0.54 vs 3.2 ± 0.44 , $P = 0.042$, Figure 5A), while intestinal perfusion with HTS^{415mOsm} failed to improve the Chiu score (3.6 ± 0.54 vs controls, $P > 0.05$).

In vivo experiments

Exclusions: Although the experimental model suggested is highly invasive and complex, the extensive training prior to the beginning of the main experimental series has minimized the number of animals excluded due to technical reasons. Only one rat was excluded in the control group because of the severe bleeding and low BP.

Blood pressure values during intestinal perfusion *in vivo*

Baseline BP values did not differ between the controls and the HTS^{365mOsm} group (Table 1, Figure 6). SMA occlusion did not cause appreciable changes in BP level in either group. Aortic occlusion during the 5-min SMA perfusion period in the HTS^{365mOsm} group resulted in a trend to BP elevation. Reinstitution of blood flow in the aorta caused a temporary decrease in BP that lasted for only 2-3 min before it returned to the baseline values (Table 1). In contrast, SMA reperfusion in controls caused a decrease in BP level that persisted during the entire reperfusion period (Table 1, Figure 6A).

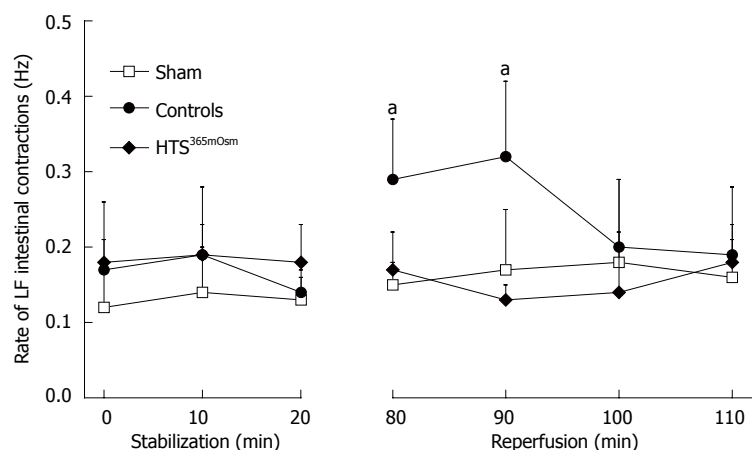


Figure 4 The rate of low frequency intestinal contractions in the *ex vivo* experiments. In shams, the intestine was continuously perfused for 110 min. In controls (60-min ischemia followed by 30-min reperfusion), intestinal contraction rate was significantly higher over the initial 10 min of reperfusion in comparison with the shams. Intestinal reperfusion with hyperosmotic saline in the HTS^{365mOsm} group prevented the increase in postischemic intestinal contraction rate. Data are mean \pm SD. ^a $P < 0.05$ vs sham. LF: Low frequency.

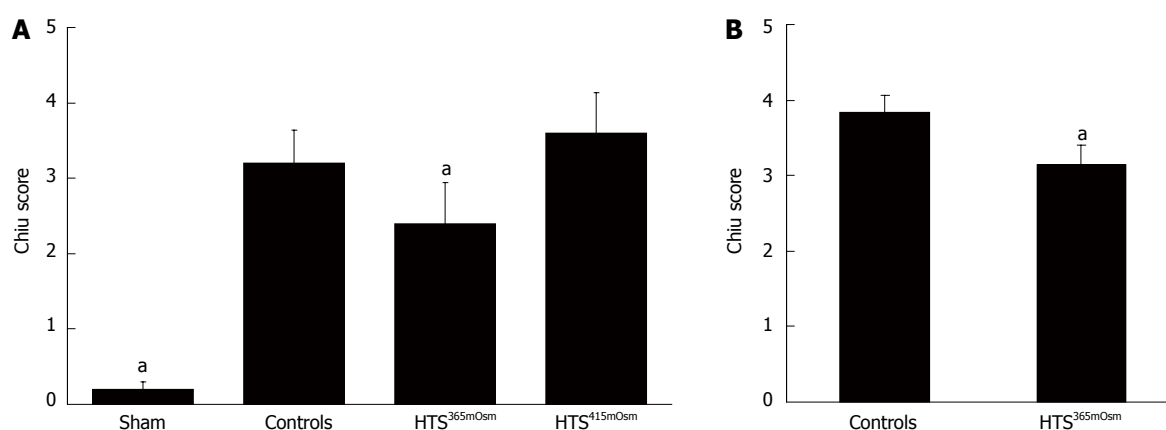


Figure 5 Histologically defined scores in the *ex vivo* (A) and *in vivo* (B) experiments. A, B: In the sham group (no ischemia at all, A) slight injury only is evident, probably due to the prolonged perfusion in the extracorporeal environment. Sixty-minute ischemia followed by 30-min normotonic reperfusion (controls) resulted in a significant injurious effect, which, however, was substantially less expressed in the HTS^{365mOsm} but not in the HTS^{415mOsm} group. The analogous difference between the controls and the HTS^{365mOsm} group is apparent in the *in vivo* experiments (B). The data are expressed as mean \pm SD; ^a $P < 0.05$ vs controls.

Table 1 Arterial blood pressure (mmHg) in the *in vivo* experiments (mean \pm SD)

	Controls ($n = 5$)	HTS ^{365mOsm} ($n = 5$)
Baseline	125 \pm 8	132 \pm 12
Before SMA occlusion	127 \pm 10	128 \pm 9
SMA occlusion		
15 min	118 \pm 11	125 \pm 7
30 min	121 \pm 9	123 \pm 6
SMA reperfusion		
15 min	68 \pm 12	118 \pm 13 ^a
30 min	74 \pm 8	126 \pm 11 ^a

SMA: Superior mesenteric artery. ^a $P < 0.05$ vs controls.

Histology: Histological data from the intestinal samples obtained after 30-min reperfusion *in vivo* are shown in Figure 5B. The Chiu score was lower in the HTS^{365mOsm} group *vs* controls (3.1 ± 0.26 and 3.8 ± 0.22 , $P = 0.0079$, respectively, Figure 5B), although the magnitude of the effect was lower than in the *ex vivo* series.

DISCUSSION

The results of this study indicate that the technique of

isolated *in vivo* vascular perfusion of the intestine is reproducible and reliable. Using this approach, intestinal perfusion with HTS^{365mOsm} over the initial 5 min of reperfusion resulted in significant protection of the intestine from IRI, as demonstrated using morphological data (per both TTC and Chiu *et al.*^[17]). In order to evaluate both the experimental and also the possible clinical implications of this strategy, it seems appropriate to review in detail the relevant findings from systemic HTS administration. A salvation effect on the intestinal IRI, as well as limitation of remote organ injury, were observed in rats that were subjected to 60-min SMA occlusion followed by 6-h reperfusion, when intravenous administration of HTS was performed during the last 5 min of ischemia^[10]. In parallel, an improvement in intestinal transit time, a reduction in mucosal injury, and limitation of intestinal myeloperoxidase activity were demonstrated using the same model; all effects were considered to be dependent on heme oxygenase-1 activity^[18]. In addition, HTS infusion performed within the first 5 min of reperfusion after 120 min of SMA occlusion in pigs resulted in a better recovery of both mean arterial blood pressure and intestinal microvascular perfusion in comparison with conventional isotonic resuscitation^[9]. In a somewhat different rat mod-

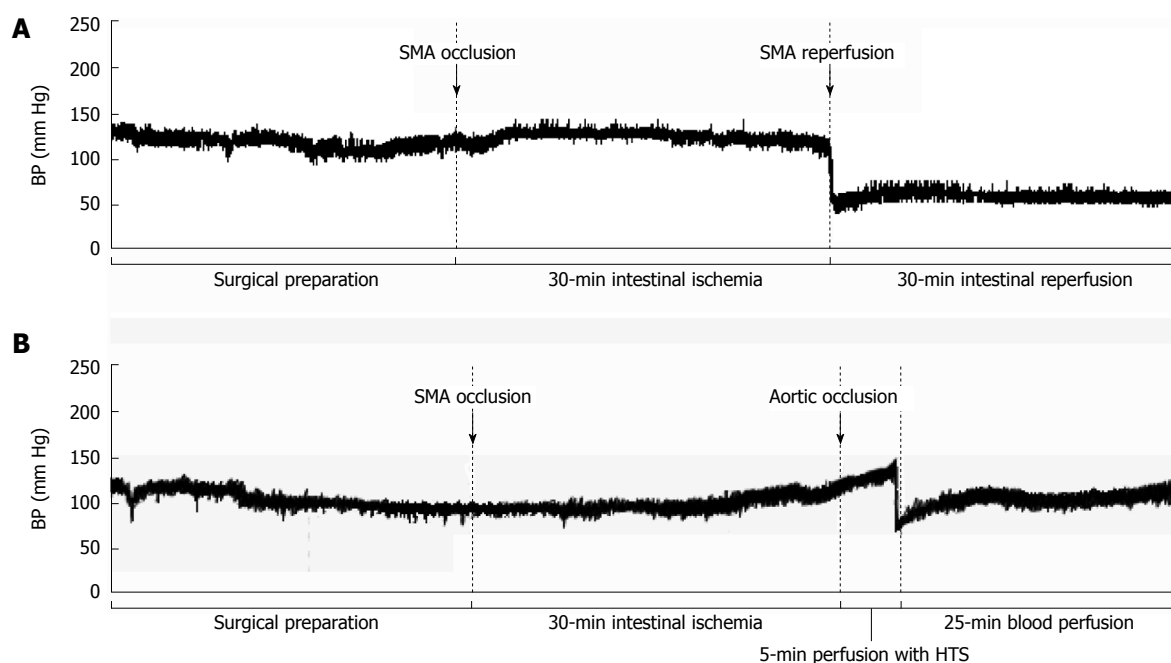


Figure 6 Representative samples of arterial blood pressure recordings in anesthetized rats subjected to intestinal ischemia-reperfusion *in vivo*. A: Thirty-minute superior mesenteric artery (SMA) occlusion followed by 30-min reperfusion (controls) resulted in a persistent blood pressure (BP) decrease over the entire reperfusion period; B: Thirty-minute SMA occlusion finalized by aortic clamping followed immediately by 5-min SMA perfusion with hypertonic saline (365 mOsm/L), with a subsequent 25-min reperfusion with blood, resulted in a transient drop with a rapid restoration of BP up to the initial values. HTS: Hypertonic saline.

el of intestinal IRI, this time induced by hemorrhagic shock followed by blood reinfusion, the administration of HTS for 25 min after the return of shed blood into circulation resulted in better post-resuscitation blood flow and improved endothelial function of the intestinal vasculature^[12]. It also suppressed the apoptotic death of intestinal epithelial cells^[20] and prevented intestinal mucosal injury^[21]. As to the remote effects of systemic HTS treatment, a more complete and stable restoration of BP^[12] as well as the prevention of systemic oxidative stress^[21] have been observed. However, in general, systemic HTS administration has several intrinsic drawbacks, including an increased risk of hypernatremia, plasma hyperosmolarity, cardiac arrhythmias, and heart failure^[12]. Apart from being free of the limitations inherent to systemic HTS administration, the technique of local HTS delivery seems to have 2 more additional advantages over a variety of protocols attempted previously^[22,23]. First, the delivery of perfusate directly into the SMA, with outflow from the opened vein, completely prevents the otherwise inevitable generalized spread throughout the systemic circulation of the metabolites and bacterial toxins accumulated during ischemia. Second, the pharmacological agents recommended for strengthening the protective effect of HTS^[24,25]—which could thus be potentially helpful to HTS administration *via* a local route—could be used at higher doses and therefore with higher efficiency, analogously with the strategy applied for regional perfusion in chemotherapy for certain neoplasms^[26].

The question might arise as to whether the HTS osmolarity value used in the present study was optimal. While the osmolarity of HTS for hypertonic resuscitation

averages approximately 7.5% NaCl, in our local delivery model we tested solutions with much lower osmolarities. Previous studies have tried values ranging from normal (300–310 mOsm/L) to nearly 400 mOsm/L for local organ anti-ischemic protection, particularly the heart; the optimal osmolarity was found to be 334 mOsm/L^[27]. Interestingly, in *ex vivo* experiments, osmolarity values above 400 mOsm/L apparently produced a negative effect; that is, HTS delivery resulted in a significant deterioration in postischemic IPF rate recovery as compared to controls (Figure 2). Although different mechanisms might be involved in the observed effect, the most likely are endothelial injury and impaired microvascular function; however, this matter requires further investigation. Based on these findings, in this study a solution with a lower osmolarity (HTS^{365mOsm}) was preferred and was found to be effective for the preservation of intestinal viability, not only *ex vivo*, but also in the *in vivo* model. A higher HTS osmolarity value than that optimal for the heart could be justified by the higher hydrophilicity of the intestinal tissue, which would thus also require more potent anti-edema protection. It should be noted, however, that further experimentation will be needed in order to determine more precisely the optimal HTS parameter(s) for the anti-IRI protection of intestinal tissue.

Although not fully understood, the mechanism of the intestine protection with HTS may include such factors as dilatation of precapillary arterioles^[12], inhibition of neutrophil-mediated reperfusion injury^[11], or prevention of intestinal edema due to reperfusion-induced water influx into the cells^[28]. At the molecular level, the major factor underlying the removal of excess water from the cells

has been shown to be the up-regulation of aquaporin 4 within the plasma membrane of the mucosal cells^[29].

As mentioned before, protective effects of local HTS delivery on reperfusion injury have also been found in the heart, suggesting that common mechanisms may be involved. The concept of controlled cardiac reperfusion was originally proposed in the late 1980s to protect the myocardium from IRI by modifying the composition and/or delivery rate of the perfusion fluid^[30]. One of the major modifications was the increased osmolality of the perfusate, which was shown to decrease the incidence of reperfusion-induced arrhythmias, to attenuate tissue injury, and to reduce edema in the isolated hearts of pig and rabbit^[27,31]. This promising approach is still awaiting implementation in clinical practice, initially in open heart surgery.

Local infusion of saline before reperfusion was also found to be protective in the brain. It was found that flushing the ischemic area of the brain with isotonic saline prior to reperfusion resulted in a reduced infarct volume and improved cerebral blood flow in the rat model^[32]. Since normotonic saline was used in this study, the authors attributed its protective effect to the local clearance of proinflammatory cytokines and metabolic end-products.

In the present study, the spontaneous intestinal motility was assessed with a technique that has not yet been employed in the experiments simulating IRI. The observation of significant augmentation of the intestinal contraction rate over the first 10 min of intra-arterial perfusate reflow after 60-min *in vitro* ischemia (Figure 4) is in general agreement with data showing a transient increase in both the contraction rate and baseline tension observed upon reoxygenation of the jejunal rings after they were subjected to 60-min *in vitro* hypoxia^[33]. In both situations, the increased motor activity of the intestine within the early postischemic period might be explained, in gross analogy with the postischemic heart response^[34], by sudden oxygen-evoked re-energizing of the cells immediately after severe hypoxia. The subsequent excessive activation of contractile machinery might, in turn, contribute to the progression of mucosal injury, most likely due to the adverse effect of hypermotility. Its complete prevention with HTS perfusion (Figure 4) may point to the solution of the relevant clinical problem.

One obvious limitation of our *in vivo* model is the need to advance the perfusion cannula through the infrarenal aorta, rather than directly into the SMA. As a result, the kidneys and hindlimbs were exposed to an additional 5 min of ischemia. However, this limitation is imposed by the small size of the animal and the need to restore blood flow in the mesenteric vascular bed. In order to make the results comparable, control animals also received 5-min occlusion of the infrarenal aorta. Future studies are needed to verify the protective effect of local HTS infusion in large animal models. For instance, the study might be repeated in pigs using an endovascular approach with percutaneous puncture in the femoral artery and balloon

occlusion of the SMA.

As to further extrapolations useful for clinical practice, the findings from the present study imply that intra-arterial HTS administration could contribute to the prevention, or at least amelioration, of 2 seemingly inevitable negative effects that would otherwise arise; that is, if “unprotected” reperfusion is applied. The first of these is reperfusion-induced intestinal damage, while the second includes whole-body disturbances, such as the systemic inflammatory response syndrome^[4], hemodynamic impairments^[35], and remote organ injuries^[36], all of which are associated with the massive washout of metabolites and bacterial toxins from the ischemia-affected intestine into the systemic circulation. Our *in vivo* model could lead to the development of a novel therapy that combines local perfusion of the ischemic region with mechanical restoration of the blood flow and/or the administration of pharmacological agents. As to the clinical translation of the model, it might be speculated that perfusion of the SMA in patients with AMI could be performed after laparotomy, either *via* a catheter directly placed in the SMA or during percutaneous intervention. The perfusate outflow could be ensured through the punctured SMV. It should be noted that the suggested technique delayed restoration of blood flow to the intestine by 5 min; however, it is unlikely that perfusion with an oxygen- and glucose-enriched solution could cause any ischemic/hypoxic injury to the intestine during this period. It is also of importance to determine the optimal osmolality value of the HTS, since the results of the present study have demonstrated the negative effect of HTS with excessive osmolality, i.e., 415 mOsm/L.

In conclusion, the results of our experiments show that isolated intestinal perfusion with HTS at the beginning of the reperfusion period can appreciably protect the intestine from IRI. Despite the positive effect of intra-arterial HTS delivery, aimed in the present study at establishing the validity of this approach in principle, its further improvement by optimization of the particular parameters during all steps of the procedure will be required before clinical trials can be contemplated.

COMMENTS

Background

Acute mesenteric ischemia (AMI) is a life-threatening vascular emergency with high mortality rate. The major treatment approach for AMI is rapid intestinal revascularization. Unfortunately, even after timely revascularization, its positive effect can be compromised, at least in part, by the development of intestinal reperfusion injury.

Research frontiers

Systemic administration of hypertonic saline (HTS) was shown to be protective in the experimental models of AMI. In the present study, the authors investigated the effect of local intra-arterial HTS administration during the early reperfusion phase on the functional and structural manifestations of intestinal injury.

Innovations and breakthroughs

The authors showed for the first time that isolated intestinal perfusion with HTS at the beginning of the reperfusion period can appreciably protect the intestine from ischemia-reperfusion injury. In addition, the technique of isolated *in vivo* vascular perfusion of the intestine is suggested and experimentally validated.

Applications

This *in vivo* model could lead to the development of a novel therapy that combines local perfusion of the ischemic intestine with mechanical restoration of the blood flow and/or the administration of pharmacological agents.

Terminology

Intestinal ischemia-reperfusion injury (IRI) develops because of the temporal complete or partial cessation of blood supply to the intestine. HTS is crystalloid or colloid solution with osmolarity value exceeding physiological. Potential benefits of HTS in the settings of AMI include prevention of edema, improvement in the microcirculation, and washout of potentially toxic metabolites.

Peer review

This paper investigated the effect of local intestinal perfusion with HTS on intestinal IRI in both *ex vivo* and *in vivo* rat models, to find that brief intestinal postischemic perfusion with HTS^{365mOsm} through the SMA followed by blood flow restoration was a protective procedure that could be used for the prevention of intestinal IRI. It is well written and organized.

REFERENCES

- Wyers MC. Acute mesenteric ischemia: diagnostic approach and surgical treatment. *Semin Vasc Surg* 2010; **23**: 9-20 [PMID: 20298945 DOI: 10.1053/j.semvasc Surg.2009.12.002]
- Berland T, Oldenburg WA. Acute mesenteric ischemia. *Curr Gastroenterol Rep* 2008; **10**: 341-346 [PMID: 18625147]
- Arthurs ZM, Titus J, Bannazadeh M, Eagleton MJ, Srivastava S, Sarac TP, Clair DG. A comparison of endovascular revascularization with traditional therapy for the treatment of acute mesenteric ischemia. *J Vasc Surg* 2011; **53**: 698-704; discussion 704-705 [PMID: 21236616 DOI: 10.1016/j.jvs.2010.09.049]
- Vollmar B, Menger MD. Intestinal ischemia/reperfusion: microcirculatory pathology and functional consequences. *Langenbecks Arch Surg* 2011; **396**: 13-29 [PMID: 21088974 DOI: 10.1007/s00423-010-0727-x]
- Takeshita M, Tani T, Harada S, Hayashi H, Itoh H, Tajima H, Ohnishi I, Takamura H, Fushida S, Kayahara M. Role of transcription factors in small intestinal ischemia-reperfusion injury and tolerance induced by ischemic preconditioning. *Transplant Proc* 2010; **42**: 3406-3413 [PMID: 21094787 DOI: 10.1016/j.transproceed.2010.06.038]
- Liu KX, Li YS, Huang WQ, Chen SQ, Wang ZX, Liu JX, Xia Z. Immediate postconditioning during reperfusion attenuates intestinal injury. *Intensive Care Med* 2009; **35**: 933-942 [PMID: 19190893 DOI: 10.1007/s00134-009-1428-1]
- Mallick IH, Yang W, Winslet MC, Seifalian AM. Ischemia-reperfusion injury of the intestine and protective strategies against injury. *Dig Dis Sci* 2004; **49**: 1359-1377 [PMID: 15481305 DOI: 10.1023/B:]
- Cámara CR, Guzmán FJ, Barrera EA, Cabello AJ, García A, Fernández NE, Caballero E, Ancer J. Ketamine anesthesia reduces intestinal ischemia/reperfusion injury in rats. *World J Gastroenterol* 2008; **14**: 5192-5196 [PMID: 18777596 DOI: 10.3748/wjg.14.5192]
- Jonas J, Heimann A, Strecker U, Kempinski O. Hypertonic/hyperoncotic resuscitation after intestinal superior mesenteric artery occlusion: early effects on circulation and intestinal reperfusion. *Shock* 2000; **14**: 24-29 [PMID: 10909889 DOI: 10.1097/00024382-200014010-00005]
- Gonzalez EA, Kozar RA, Suliburk JW, Weisbrodt NW, Mercer DW, Moore FA. Conventional dose hypertonic saline provides optimal gut protection and limits remote organ injury after gut ischemia reperfusion. *J Trauma* 2006; **61**: 66-73; discussion 73-74 [PMID: 16832251 DOI: 10.1097/01.ta.0000224190.65542.e2]
- Shukla A, Hashiguchi N, Chen Y, Coimbra R, Hoyt DB, Junger WG. Osmotic regulation of cell function and possible clinical applications. *Shock* 2004; **21**: 391-400 [PMID: 15087814 DOI: 10.1097/00024382-200405000-00001]
- Zakaria el R, Tsakadze NL, Garrison RN. Hypertonic saline resuscitation improves intestinal microcirculation in a rat model of hemorrhagic shock. *Surgery* 2006; **140**: 579-587; discussion 587-588 [PMID: 17011905 DOI: 10.1016/j.surg.2006.05.015]
- Gardemann A, Watanabe Y, Grosse V, Hesse S, Jungermann K. Increases in intestinal glucose absorption and hepatic glucose uptake elicited by luminal but not vascular glutamine in the jointly perfused small intestine and liver of the rat. *Biochem J* 1992; **283** (Pt 3): 759-765 [PMID: 1590766]
- Galagudza M, Vaage J, Valen G. Isoflurane and other commonly used anaesthetics do not protect the isolated buffer perfused mouse heart from ischemia-reperfusion injury. *Clin Exp Pharmacol Physiol* 2006; **33**: 315-319 [PMID: 16620294 DOI: 10.1111/j.1440-1681.2006.04368.x]
- Csonka C, Kupai K, Kocsis GF, Novák G, Fekete V, Bencsik P, Csont T, Ferdinandy P. Measurement of myocardial infarct size in preclinical studies. *J Pharmacol Toxicol Methods* 2010; **61**: 163-170 [PMID: 20188845 DOI: 10.1016/j.jvasc.2010.02.014]
- Hoffman JM, Brooks EM, Mawe GM. Gastrointestinal Motility Monitor (GIMM). *J Vis Exp* 2010; **(46)**: 2435 [PMID: 21189461 DOI: 10.3791/2435]
- Chiu CJ, McArdle AH, Brown R, Scott HJ, Gurd FN. Intestinal mucosal lesion in low-flow states. I. A morphological, hemodynamic, and metabolic reappraisal. *Arch Surg* 1970; **101**: 478-483 [PMID: 5457245 DOI: 10.1001/archsurg.1970.01340280030009]
- Attuwaybi B, Kozar RA, Gates KS, Moore-Olufemi S, Sato N, Weisbrodt NW, Moore FA. Hypertonic saline prevents inflammation, injury, and impaired intestinal transit after gut ischemia/reperfusion by inducing heme oxygenase 1 enzyme. *J Trauma* 2004; **56**: 749-758; discussion 758-759 [PMID: 15187737 DOI: 10.1097/01.TA.0000119686.33487.65]
- Guzmán-de la Garza FJ, Cámara-Lemarroy CR, Alarcón-Galván G, Cordero-Pérez P, Muñoz-Espinosa LE, Fernández-Garza NE. Different patterns of intestinal response to injury after arterial, venous or arteriovenous occlusion in rats. *World J Gastroenterol* 2009; **15**: 3901-3907 [PMID: 19701970 DOI: 10.3748/wjg.15.3901]
- Lu YQ, Gu LH, Huang WD, Mou HZ. Effect of hypertonic saline resuscitation on heme oxygenase-1 mRNA expression and apoptosis of the intestinal mucosa in a rat model of hemorrhagic shock. *Chin Med J (Engl)* 2010; **123**: 1453-1458 [PMID: 20819606]
- Powers KA, Zurawska J, Szasz K, Khadaroo RG, Kapus A, Rotstein OD. Hypertonic resuscitation of hemorrhagic shock prevents alveolar macrophage activation by preventing systemic oxidative stress due to gut ischemia/reperfusion. *Surgery* 2005; **137**: 66-74 [PMID: 15614283 DOI: 10.1016/j.surg.2004.05.051]
- Pang KS, Yuen V, Fayz S, te Koppele JM, Mulder GJ. Absorption and metabolism of acetaminophen by the in situ perfused rat small intestine preparation. *Drug Metab Dispos* 1986; **14**: 102-111 [PMID: 2868852]
- Scholtka B, Stümpel F, Jungermann K. Acute increase, stimulated by prostaglandin E₂, in glucose absorption via the sodium dependent glucose transporter-1 in rat intestine. *Gut* 1999; **44**: 490-496 [PMID: 10075955 DOI: 10.1136/gut.44.4.490]
- Shih HC, Huang MS, Lee CH. Estrogen augments the protection of hypertonic saline treatment from mesenteric ischemia-reperfusion injury. *Shock* 2011; **35**: 302-307 [PMID: 20926986 DOI: 10.1097/SHK.0b013e3181f8b420]
- Shih HC, Huang MS, Lee CH. Magnolol attenuates the lung injury in hypertonic saline treatment from mesenteric ischemia reperfusion through diminishing iNOS. *J Surg Res* 2012; **175**: 305-311 [PMID: 21704335 DOI: 10.1016/j.jss.2011.04.063]
- Sanki A, Kroon HM, Kam PC, Thompson JF. Isolated limb perfusion and isolated limb infusion for malignant lesions of the extremities. *Curr Probl Surg* 2011; **48**: 371-430 [PMID: 21704335 DOI: 10.1016/j.jss.2011.04.063]

- 21549235 DOI: 10.1067/j.cpsurg.2011.02.002]
- 27 **Weng ZC**, Nicolosi AC, Detwiler PW, Hsu DT, Schierman SW, Goldstein AH, Spotnitz HM. Effects of crystalloid, blood, and University of Wisconsin perfusates on weight, water content, and left ventricular compliance in an edema-prone, isolated porcine heart model. *J Thorac Cardiovasc Surg* 1992; **103**: 504-513 [PMID: 1545549]
- 28 **Radhakrishnan RS**, Radhakrishnan HR, Xue H, Moore-Olufemi SD, Mathur AB, Weisbrodt NW, Moore FA, Allen SJ, Laine GA, Cox CS. Hypertonic saline reverses stiffness in a Sprague-Dawley rat model of acute intestinal edema, leading to improved intestinal function. *Crit Care Med* 2007; **35**: 538-543 [PMID: 17205008 DOI: 10.1097/01.CCM.0000254330.39804.9C]
- 29 **Radhakrishnan RS**, Shah SK, Lance SH, Radhakrishnan HR, Xue H, Radhakrishnan GL, Ramaswamy US, Walker PA, Uray KS, Laine GA, Stewart RH, Cox CS. Hypertonic saline alters hydraulic conductivity and up-regulates mucosal/submucosal aquaporin 4 in resuscitation-induced intestinal edema. *Crit Care Med* 2009; **37**: 2946-2952 [PMID: 19770732 DOI: 10.1097/CCM.0b013e3181ab878b]
- 30 **Okamoto F**, Allen BS, Buckberg GD, Young H, Bugyi H, Leaf J. Perfusate composition: interaction of marked hyperglycemia and marked hyperosmolarity in allowing immediate contractile recovery after four hours of regional ischemia. *J Thorac Cardiovasc Surg* 1986; **92**: 583-593 [PMID: 3747586]
- 31 **Careaga G**, Argüero R, Chavez-Negrete A, Valero G, Portilla E, Garcia RM, Mendoza L, Angulo L, Miranda Y. Control of myocardial reperfusion injury with hypertonic-hyperosmotic solution in isolated rabbit heart. *Eur Surg Res* 1995; **27**: 269-276 [PMID: 7649214 DOI: 10.1159/000129409]
- 32 **Ding Y**, Li J, Rafols JA, Phillis JW, Diaz FG. Prereperfusion saline infusion into ischemic territory reduces inflammatory injury after transient middle cerebral artery occlusion in rats. *Stroke* 2002; **33**: 2492-2498 [PMID: 12364743 DOI: 10.1161/01.STR.0000028237.15541.CC]
- 33 **Bielefeldt K**, Conklin JL. Intestinal motility during hypoxia and reoxygenation in vitro. *Dig Dis Sci* 1997; **42**: 878-884 [PMID: 9149037]
- 34 **Abdallah Y**, Gkatzoflia A, Gligorievski D, Kasseckert S, Euler G, Schlüter KD, Schäfer M, Piper HM, Schäfer C. Insulin protects cardiomyocytes against reoxygenation-induced hypercontracture by a survival pathway targeting SR Ca²⁺ storage. *Cardiovasc Res* 2006; **70**: 346-353 [PMID: 16569400 DOI: 10.1016/j.cardiores.2006.02.020]
- 35 **Walensi M**, de Groot H, Schulz R, Hartmann M, Petrat F. Mesenteric ischemia-reperfusion injury: Clearly improved hemodynamics but only minor protection of the rat small intestine by (sub)therapeutic heparin sodium and enoxaparin doses. *J Surg Res* 2013; **179**: e57-e69 [PMID: 22494914 DOI: 10.1016/j.jss.2012.01.002]
- 36 **Varga J**, Tóth S, Staško P, Tóth S, Bilecová-Rabajdová M, Ostró A, Veselá J. Intestinal ischemia-reperfusion injury - the histopathological status of remote vital organs in acute and subacute phases. *Ann Transplant* 2012; **17**: 11-20 [PMID: 22466904]

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Characterization of focal liver masses using acoustic radiation force impulse elastography

Hana Park, Jun Yong Park, Do Young Kim, Sang Hoon Ahn, Chae Yoon Chon, Kwang-Hyub Han, Seung Up Kim

Hana Park, Division of Gastroenterology, Department of Internal Medicine, CHA Bundang Medical Center, CHA University, Gyeonggi-do 463-712, South Korea

Jun Yong Park, Do Young Kim, Chae Yoon Chon, Seung Up Kim, Department of Internal Medicine, Yonsei University College of Medicine, Seoul 120-752, South Korea

Sang Hoon Ahn, Kwang-Hyub Han, Department of Internal Medicine, Yonsei University College of Medicine, Seoul 120-752, South Korea

Author contributions: Park H designed the study, analyzed the data, performed the study, and wrote the paper; Park JY, Kim DY, Ahn SH, Chon CY collected the data; Han KH, Kim SU designed the study, collected the data, performed the study and wrote the paper.

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Correspondence to: Seung Up Kim, MD, Department of Internal Medicine, Yonsei University College of Medicine, 134 Shinchon-dong, Seodaemun-gu, Seoul 120-752, South Korea. ksukorea@yuhs.ac

Telephone: +82-2-3936884 Fax: +82-2-22281982

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Abstract

AIM: To investigate the diagnostic performance of acoustic radiation force impulse (ARFI) elastography for characterizing focal liver mass by quantifying their stiffness.

METHODS: This prospective study included 62 patients with a focal liver mass that was well visualized on conventional ultrasonography performed in our institution from February 2011 to November 2011. Among them, 12 patients were excluded for ARFI measurement failure due to a lesion that was smaller than the region of the interest and at an inaccessible loca-

tion (deeper than 8 cm) ($n = 7$) or poor compliance to hold their breath as required ($n = 5$). Finally, 50 patients with valid ARFI measurements were enrolled. If a patient had multiple liver masses, only one mass of interest was chosen. The masses were diagnosed by histological examination or clinical diagnostic criteria. During ultrasonographic evaluation, stiffness, expressed as velocity, was checked 10 times per focal liver mass and the surrounding liver parenchyma.

RESULTS: After further excluding three masses that were non-diagnostic on biopsy, a total of 47 focal mass lesions were tested, including 39 (83.0%) malignant masses [24 hepatocellular carcinomas (HCC), seven cholangiocellular carcinomas (CCC), and eight liver metastases] and eight (17.0%) benign masses (five hemangiomas and three focal nodular hyperplasias, FNH). Thirty-seven (74.0%) masses were confirmed by histological examination. The mean velocity was 2.48 m/s in HCCs, 1.65 m/s in CCCs, 2.35 m/s in metastases, 1.83 m/s in hemangiomas, and 0.97 m/s in FNHs. Although considerable overlap was still noted between malignant and benign masses, significant differences in ARFI values were observed between malignant and benign masses (mean 2.31 m/s vs 1.51 m/s, $P = 0.047$), as well as between HCCs and benign masses (mean 2.48 m/s vs 1.51 m/s, $P = 0.006$). The areas under the receiver operating characteristics curves (AUROC) for discriminating the malignant masses from benign masses was 0.724 (95%CI, 0.566-0.883, $P = 0.048$), and the AUROC for discriminating HCCs from benign masses was 0.813 (95%CI, 0.649-0.976, $P = 0.008$). To maximize the sum of sensitivity and specificity, an ARFI value of 1.82 m/s was selected as the cutoff value to differentiate malignant from benign liver masses. Furthermore, the cutoff value for distinguishing HCCs from benign masses was also determined to be 1.82 m/s. The diagnostic performance of the sum of the ARFI values for focal liver masses and the surrounding liver parenchyma to differentiate liver masses improved (AUROC = 0.853;

95%CI, 0.745-0.960; $P = 0.002$ in malignant liver masses *vs* benign ones and AUROC = 0.948; 95%CI, 0.896-0.992, $P < 0.001$ in HCCs *vs* benign masses).

CONCLUSION: ARFI elastography provides additional information for the differential diagnosis of liver masses. However, our results should be interpreted in clinical context, because considerable overlap in ARFI values existed among liver masses.

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Key words: Acoustic radiation force impulse; Focal liver mass; Hepatocellular carcinoma; Hemangioma; Focal nodular hyperplasia; Cholangiocellular carcinoma; Liver metastasis

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INTRODUCTION

Focal liver masses are being discovered at increasing rates due to the wide accessibility of modern high resolution imaging procedures. Conventional ultrasonography (US) is typically used as a first imaging modality to evaluate a focal liver mass. The differential diagnosis of a focal liver mass using imaging studies is based on the characteristics of the surrounding liver parenchyma and underlying clinical conditions such as cirrhosis and the characteristics of the mass itself. Although contrast-enhanced US, computed tomography (CT), and magnetic resonance imaging (MRI) can assess the morphology of a focal liver mass and its vasculature with a high level of diagnostic accuracy^[1-5], patients are exposed to potential risks, including contrast medium-induced side-effects and irradiation hazards^[6], particularly when repeated examinations are required.

Acoustic radiation force impulse (ARFI) elastography has been introduced as a new ultrasound imaging modality to evaluate tissue stiffness using the radiation forced-based imaging method. The tissue response to the radiation force is observed using conventional B-mode imaging pulses, and it is possible to display the quantitative shear wave velocity (m/s) of the ARFI image^[7,8]. Because velocity is directly related to tissue stiffness, ARFI imaging can be applied to evaluate tissue elasticity. Generally, the stiffer the tissue, the faster the shear wave propagates^[6,9-13]. Although several reports have indicated a good correlation of ARFI elastography with the other elastography systems such as transient elastography^[14,15] and with histological fibrosis grade^[14,16,17], ARFI elastography differs from other elastography systems which apply pressure manually to the surface of the organs or a

mechanical vibration to induce an elastic shear wave with only M-mode US imaging. Instead, ARFI uses short-duration acoustic pulses generated from a probe under real-time B-mode imaging to produce localized displacements in tissue^[18-20]. Furthermore, because ARFI elastography uses elastography with a flexible metering box of the region of the interest (ROI), it is the only elastography method suitable for quantifying focal liver mass stiffness.

Few reports have investigated the applicability of ARFI elastography to evaluate focal liver masses^[6,20-25]. Thus, we prospectively recruited patients with a focal liver mass and investigated the diagnostic performance of ARFI elastography to discriminate malignant liver masses and hepatocellular carcinoma (HCC) from benign masses by quantifying their stiffness.

MATERIALS AND METHODS

Subjects

From February 2011 to November 2011, a total of 62 patients with a focal liver mass that was well visualized on conventional US were prospectively recruited for this study. The subjects were referred to our institute for further evaluation of a focal liver mass from primary or secondary clinics, or had been diagnosed with a focal liver mass during a surveillance examination at our institute. Of these, 12 patients were excluded for the following reasons: (1) ARFI measurement failure due to a lesion that was smaller than the ROI and at an inaccessible location (deeper than 8 cm) ($n = 7$) or (2) poor compliance or inability to hold their breath as required ($n = 5$). Finally, 50 patients with valid ARFI measurements were enrolled. If a patient had multiple liver masses, only one mass of interest was chosen.

This study was approved by the institutional review board at Severance Hospital in Seoul, South Korea, and conformed to the ethical guidelines of the 1975 Declaration of Helsinki. Written informed consent was obtained from all participants.

Histological confirmation of the liver masses

We performed a targeted biopsy on masses when a confirmatory histological diagnosis was needed or the radiological diagnosis was not confirmative. Among the 50 masses in our study, 37 (74.0%) received targeted biopsy for histological confirmation. Masses that had any component of cholangiocellular carcinomas (CCC) or combined HCC (CHCC) were classified as CCC and the others as HCC.

Clinical diagnosis of the liver masses

A clinical diagnosis of HCC was made according to the American Association for the Study of Liver Disease recommendation^[26]. Briefly, patients were diagnosed with HCC if they had a tumor with a maximum diameter > 2 cm and exhibited typical features of HCC on dynamic CT, defined as enhancement in the arterial phase, early washout on the portal phase, and an α -fetoprotein level

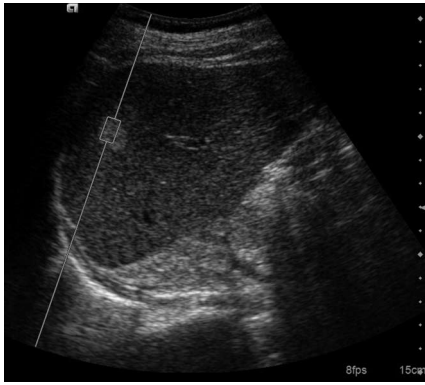


Figure 1 Measurement of acoustic radiation force impulse value within the region of interest of a focal liver mass. The shear wave velocity measured when the region of interest box was placed within the mass.

> 200 ng/mL. Overall, nine patients satisfied these criteria sufficiently to be diagnosed with HCC without a histological examination.

The presence of a hemangioma was diagnosed clinically in four patients, based on a combination of typical findings, determined using CT or MRI, and a lack of growth for at least 12 mo^[22]. A hemangioma appears as a mass with hypoattenuation on unenhanced CT, very high signal intensity on T2-weighted images, peripheral nodular enhancement in the arterial phase, and progressive filling-in of enhancement with no washout in later phases^[20].

ARFI elastography

ARFI elastography was performed with an Acuson S2000 ultrasound system (Siemens, Erlanger, Germany), using a 4-1 MHz curved array probe. ARFI elastography was performed by a single physician in all patients (2 years of experience with US and more than 100 examinations with ARFI elastography) who was blinded with regard to the clinical and biochemical data. For patients who underwent liver biopsy, ARFI elastography was performed just before the biopsy, on the same day. For patients with a solitary liver mass that was clinically diagnosed, ARFI elastography was performed at the time of enrollment in this study.

Details of the technical background and examination procedure have been described previously^[27]. Briefly, a B-mode US image of the lesion was identified utilizing a ROI, characterized by a box with a fixed dimension of 1 cm × 0.5 cm and a maximum depth of 8 cm. The ROI was entirely located in the lesion, and the ROI location was changed to cover large masses as much as possible without including any vascular or biliary structures (Figure 1). The potential presence of degeneration, such as necrotic, cystic, or calcified portions, was not included in the ROI. To evaluate background liver status of the focal mass, measurements were also taken in the surrounding liver parenchyma with the ROI within 2-3 cm from the target mass, taking care not to comprise any vascular or

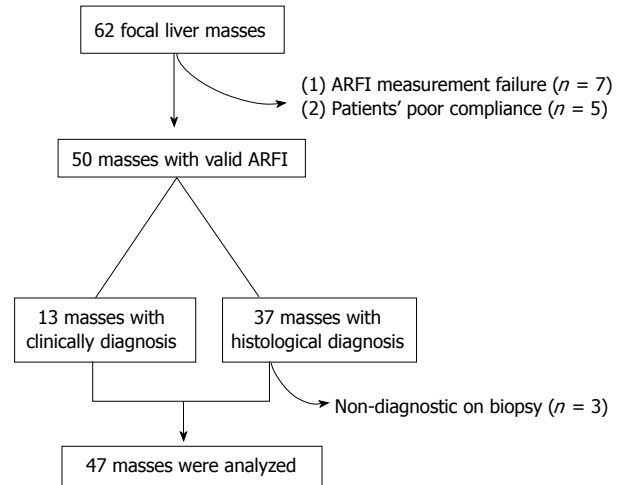


Figure 2 Recruitment algorithm.

biliary structure. To ensure quality of the ARFI measurement, 10 measurements were performed for each mass and surrounding liver parenchyma.

Statistical analysis

Continuous variables are expressed as medians and ranges. The χ^2 test or Fisher's exact test and the Mann-Whitney test were used to compare categorical and continuous variables, respectively. Receiver operating characteristics (ROC) curves and areas under the ROC curves (AU-ROC) were used to estimate diagnostic performance. The cutoff ARFI value for maximal diagnostic accuracy was selected by considering the highest sum of sensitivity and specificity. A P -value < 0.05 was considered statistically significant. The statistical analysis was performed using SPSS software ver. 18.0.0 (Chicago, IL, United States).

RESULTS

Final diagnosis of focal liver masses

We further excluded three masses that were non-diagnostic on biopsy due to a lack of sufficient tissue or ineffective targeting of the mass. Thus, 47 masses (34 histologically confirmed and 13 clinically diagnosed) were evaluated (Figure 2).

Overall, 15 HCCs, eight metastases (three from colorectal cancers, two from gallbladder cancer, one from pancreatic cancer, one from a gastrointestinal stromal tumor, and one from cervical cancer), seven CCCs, three focal nodular hyperplasias (FNH), and one hemangioma were diagnosed. Figure 3 describes pre-biopsy and post-biopsy diagnoses: 21 (61.8%) masses were consistent with the pre-biopsy diagnosis, and 10 masses diagnosed as HCC or CHCC at the time of pre-biopsy were finally confirmed as six HCCs and four CCCs through the biopsy. The histological diagnosis of the other three masses changed (two FNHs to one hemangioma and one HCC; one poorly-differentiated malignancy or metastatic mass to CCC).

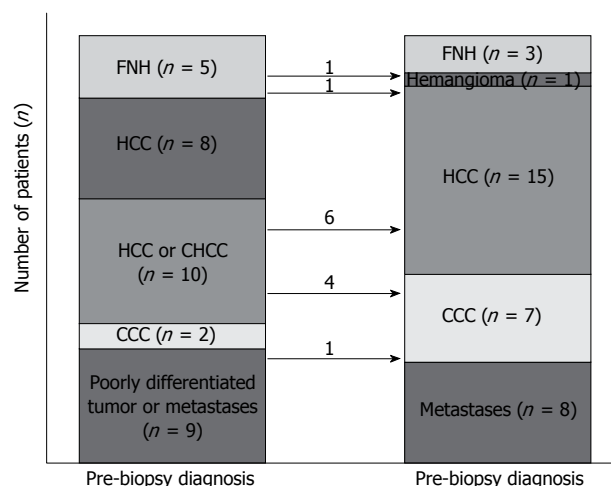


Figure 3 The flow of histological diagnosis. The values on arrows indicate the number of patients who obtained diagnoses differing from those pre-biopsy diagnoses after liver biopsy.

Characteristics and ARFI values of focal liver masses

The depth, location (right *vs* left lobe), and size of each focal liver mass are described in Table 1. ARFI values of HCCs and metastases were higher (2.48 and 2.35 m/s) than those of hemangiomas (1.83 m/s), CCCs (1.65 m/s), and FNH (0.97 m/s). ARFI values for CCCs, hemangiomas, and metastases were significantly higher than those for their surrounding liver parenchyma (mean 1.65 *vs* 1.07 m/s in CCCs; 1.83 m/s *vs* 1.10 m/s in hemangiomas; 2.35 m/s *vs* 1.45 m/s in metastases; all $P < 0.05$), whereas ARFI values for HCCs were similar to those of surrounding liver parenchyma (2.48 m/s *vs* 2.14 m/s, $P = 0.134$).

When HCCs, CCCs, and metastases were categorized into a malignant liver mass group ($n = 39$) and the others were stratified into a benign mass group ($n = 8$), a significant difference appeared in the between the malignant and benign liver masses (2.31 ± 1.05 m/s *vs* 1.51 ± 0.69 m/s, $P = 0.047$) (Figure 4A). However, considerable overlap in ARFI values was noted between malignant and benign masses. Additionally, a significant difference in the ARFI values was observed between the HCCs and the benign masses (2.48 ± 0.84 m/s *vs* 1.51 ± 0.69 m/s, $P = 0.006$) (Figure 4A). To address this overlap, we attempted further analysis taking into consideration the ARFI values of surrounding liver parenchyma as well as those of focal liver masses. When ARFI values of focal liver masses and their surrounding liver parenchyma were combined, the differences in ARFI values for each tumor type became more prominent (4.29 ± 1.22 m/s *vs* 2.92 ± 0.35 m/s for malignant masses *vs* benign masses, $P < 0.001$; 4.62 ± 0.96 m/s *vs* 2.92 ± 0.35 m/s for HCCs *vs* benign masses, $P < 0.001$) (Figure 4B).

Discrimination of malignant liver masses and HCCs from benign masses

The AUROC for discriminating the malignant masses from benign masses was 0.724 (95%CI, 0.566-0.883,

$P = 0.048$), and the AUROC for discriminating HCCs from benign masses was 0.813 (95%CI, 0.649-0.976, $P = 0.008$) (Table 2). To maximize the sum of sensitivity and specificity, an ARFI value of 1.82 m/s was selected as the cutoff value to differentiate malignant from benign liver masses. Furthermore, the cutoff value for distinguishing HCCs from benign masses was also determined to be 1.82 m/s (Table 2). Additionally, the diagnostic performance of the sum of the ARFI values for focal liver masses and the surrounding liver parenchyma to differentiate liver masses improved (AUROC = 0.853; 95%CI, 0.745-0.960; $P = 0.002$ in malignant liver masses *vs* benign ones and AUROC = 0.948; 95%CI, 0.896-0.992, $P < 0.001$ in HCCs *vs* benign masses) (Table 2).

DISCUSSION

ARFI elastography has been proposed as a new method for assessing liver stiffness^[7-9]. Although ARFI elastography uses shear wave velocity (m/s) to assess liver stiffness, which is similar to FibroScan[®], it exhibits several unique properties. First, ARFI elastography allows for the evaluation of deep tissue by generating a shear wave without the need for exertional compression^[7,28]. Second, ARFI elastography has the distinct advantage of being integrated into a conventional US system and can provide additional real-time information during a conventional US study^[14,15]. Third, ARFI elastography can be performed regardless of the presence of impediments such as ascites, although the reproducibility in this setting should be further investigated^[19,29]. Finally, ARFI elastography offers a flexible metering box at variable depths, allowing the examination of specific liver areas^[21,30]. Due to these characteristics, the clinical applicability of ARFI elastography has expanded to characterize and distinguish focal liver masses beyond a simple assessment of liver fibrosis^[6,20-25].

According to previous studies^[22,24,25], malignant liver masses are stiffer than benign masses, as reflected by higher ARFI values. Consistent with this finding, the ARFI values of malignant liver masses (2.31 m/s) or HCCs (2.48 m/s) in our cohort were significantly higher than those of benign masses (1.51 m/s) (all $P < 0.05$). However, when we consider that two types of malignant liver masses (HCCs and CCCs) with different ARFI values were stratified into one malignant mass group in our study, the accuracy of ARFI elastography for identifying malignant liver masses would change according to the proportion of HCCs and CCCs. Thus, an exact comparison between the ARFI values of HCCs and CCCs as well as those of benign masses should be performed in future, larger-scale studies. Additionally, the respective ARFI values of HCCs and metastatic masses in our study were 2.48 m/s and 2.35 m/s, which were similar to data from previous studies (2.45 to 2.63 m/s in HCC and 2.18 to 2.88 m/s in metastasis)^[22,23].

ARFI values for hemangiomas vary among reports^[6,20,22,23]. The ARFI values of hemangiomas reported

Table 1 Characteristics and acoustic radiation force impulse values of focal liver masses

Variables	Total masses (<i>n</i> = 47)	HCC (<i>n</i> = 24)	CCC (<i>n</i> = 7)	Metastases (<i>n</i> = 8)	Hemangioma (<i>n</i> = 5)	FNH (<i>n</i> = 3)
Histologic confirmation	34 (72.3)	15 (62.5)	7 (100)	8 (100)	1 (20.0)	3 (100)
Size, cm	5.0 (1.4-20.5)	4.9 (1.4-18.3)	9.4 (7.4-18.8)	8.0 (1.6-20.5)	1.9 (1.5-4.7)	2.7 (2.0-3.0)
Depth, cm	5.6 (2.7-7.8)	5.7 (2.7-7.8)	6.4 (4.8-7.2)	4.9 (3.5-7.6)	5.3 (4.5-6.5)	5.6 (2.8-7.8)
Right/left lobe	34 (72.3)/13 (27.7)	19 (79.2)/5 (20.8)	5 (71.4)/2 (28.6)	5 (62.5)/3 (37.5)	4 (80.0)/1 (20.0)	1 (33.3)/2 (66.7)
ARFI value, m/s						
Masses	2.23 ± 0.98	2.48 ± 0.84	1.65 ± 1.43	2.35 ± 1.18	1.83 ± 0.62	0.97 ± 0.48
Surrounding parenchyma	1.83 ± 0.73	2.14 ± 0.59	1.07 ± 0.49	1.45 ± 0.51	1.10 ± 0.14	1.63 ± 0.40
<i>P</i> value	0.029	0.134	0.015	0.043	0.013	0.581

Data was expressed as median (range) or *n* (%). HCC: Hepatocellular carcinoma; CCC: Cholangiocellular carcinoma; FNH: Focal nodular hyperplasia; ARFI: Acoustic radiation force impulse.

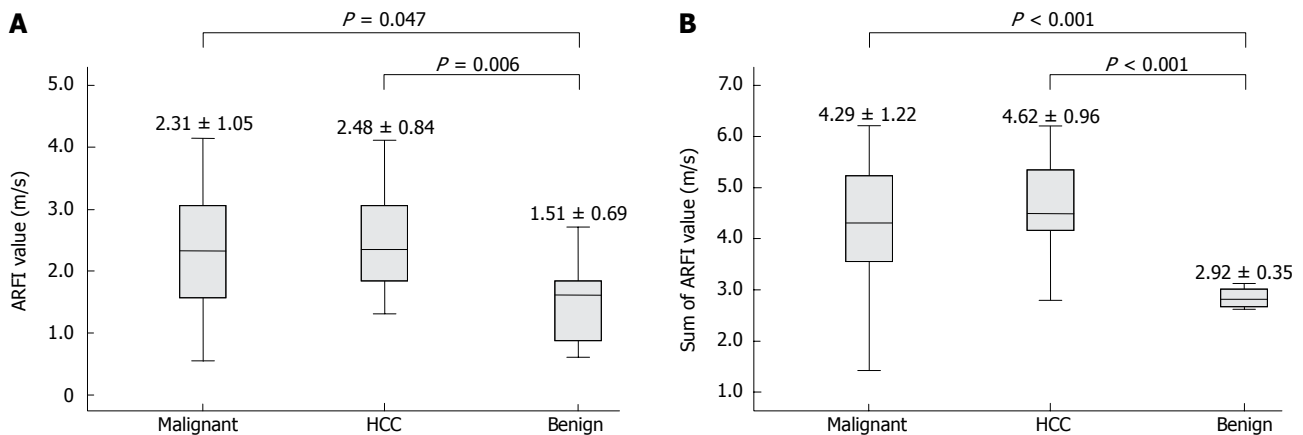


Figure 4 Acoustic radiation force impulse values of malignant masses, hepatocellular carcinoma, and benign masses. A: Acoustic radiation force impulse (ARFI) value of a focal liver mass; B: Sum of the ARFI values of liver mass and the surrounding liver parenchyma. HCC: Hepatocellular carcinoma.

Table 2 Optimal cutoff acoustic radiation force impulse values and corresponding diagnostic indices for discriminating malignant masses and hepatocellular carcinoma from benign liver masses

	Malignant <i>vs</i> benign		HCC <i>vs</i> benign	
	Masses	Masses with surrounding liver parenchyma	Masses	Masses with surrounding liver parenchyma
Cutoff ARFI value, m/s	1.82 ¹	3.72 ²	1.82 ¹	3.79 ²
AUROC (95% CI)	0.724 (0.566-0.883)	0.853 (0.745-0.960)	0.813 (0.649-0.976)	0.948 (0.896-0.992)
Sensitivity, %	71.8	71.8	79.2	87.5
Specificity, %	75.0	100	75.0	100
Positive predictive value, %	93.3	100	90.5	100
Negative predictive value, %	35.3	42.1	54.5	72.9

¹ARFI value of a focal liver mass; ²Sum of ARFI values of focal liver mass and surrounding liver parenchyma. ARFI: Acoustic radiation force impulse; HCC: Hepatocellular carcinoma; AUROC: Area under the receiver operating characteristics; CI: Confidence intervals.

in two previous studies (1.51 and 1.75 m/s, respectively) were comparable to our data (1.83 m/s)^[20,22,23], whereas the ARFI value for a hemangioma in another study was rather higher (2.36 m/s)^[22,23]. The variability in hemangioma ARFI values has been explained based on the amount of fibrotic septa that divide the dilated vascular spaces^[22]. That is, hemangiomas are composed of multiple vascular channels filled with blood so they would not be expected to be stiff, and therefore would have low ARFI values; in contrast, those including pathological patterns such as sclerosis, thrombosis of the vessels, or calcification, would be stiff and have high ARFI values^[20]. Because

relatively few hemangioma cases were analyzed in these studies, this discrepancy will need to be investigated *via* further study including a larger number of cases.

Some researchers have attempted to suggest a cutoff ARFI value for distinguishing malignant liver masses from benign masses. We obtained a high PPV (malignant masses *vs* benign masses, PPV 93.3%; HCC *vs* benign masses, 90.5%) using the cutoff ARFI value of 1.82 m/s, which maximized the sum of the sensitivity and specificity for distinguishing masses. Our cutoff value of 1.82 m/s for identifying malignant liver masses was slightly lower than those in previous studies (2.0 to 2.5 m/s)^[22,24,25],

which can be partially explained by the different methods used to categorize the masses. In one study, which reported a rather high cutoff value of 2.5 m/s, only metastases that had a high ARFI value of 4.18 m/s were categorized into a malignant group and compared with benign masses^[24], whereas another study that reported 2.22 m/s as a cutoff value for discriminating malignant masses from benign masses included FNHs, adenomas, and focal fatty change lesions in the benign mass group as well as hemangiomas^[25]. Thus, in this regard, further study is needed to prevent potential bias due to heterogeneity of the masses and to confirm the clinically applicable cutoff values for ARFI elastography. Although our cutoff value may be useful to physicians who encounter focal liver masses during routine US evaluation, it should be interpreted cautiously in the clinical context for several reasons. First, although our cutoff value would correctly characterize HCC and metastasis, it would mischaracterize CCC with a relatively low ARFI value and hemangioma with a relatively high ARFI value. Second, there was considerable overlap of ARFI values between malignant and benign masses.

Underlying fibrosis of the liver is a consideration for the differential diagnosis of a focal liver mass. Several previous studies have proposed that HCCs generally appear softer (lower ARFI values) than the surrounding liver^[21], whereas metastases and hemangiomas generally appear harder than that of the surrounding liver despite some controversies among studies^[21]. Similarly, we found that CCCs, hemangiomas, and metastases had higher ARFI values than those of surrounding liver parenchyma, whereas ARFI values of HCCs were statistically equivalent to those of the surrounding liver parenchyma. Because hemangiomas and metastases were evaluated in patients without chronic liver disease in most studies^[6,25], ARFI values seemed consistently higher than those of a background liver. In contrast, because HCCs were evaluated in patients with chronic liver disease and diverse degrees of background liver fibrosis, the comparative results between ARFI values of HCCs and those of background liver differed among studies based on the characteristics of each study cohort^[6,21,25]. That is, simultaneously measuring ARFI values of focal liver mass and the surrounding liver should focus on assessing the respective characteristics of the hepatic mass and surrounding fibrosis to prevent a misdiagnosis of the hepatic mass using the correlation of ARFI values between the liver mass and background liver parenchyma. However, when we used the sum of ARFI values of focal liver mass and the surrounding liver parenchyma, the diagnostic performance in terms of distinguishing liver mass improved. Thus, these controversial findings concerning the simultaneous measurement of liver masses and their surrounding liver parenchyma should be further investigated in future large-scale studies.

Although most cases (72.3%) were histologically confirmed in our study, the relative small sample size of our cohort and inclusion of patients with high ALT, which

has the potential of overestimating influences on ARFI values^[30,31], are potential limitations. Although ARFI elastography can freely locate the ROI box in a specific area within a mass and measure its stiffness, morphological characteristics of liver masses including heterogeneous components such as HCC with hemorrhage and fatty metamorphosis^[6,20,32] and lesion shapes were not considered in our study, which is a limitation of this study. Further investigation of how to evaluate such heterogeneous or morphologically varying liver masses using ARFI elastography and their influences on ARFI values of liver masses should be conducted.

In our study, we demonstrated the potential clinical utility of ARFI elastography for characterization of focal liver masses. Although this study had limitations and should be interpreted cautiously, our findings provide a useful reference for the differential diagnosis of a focal liver mass and will provide additional information to clinicians who are confronted with a need for an immediate diagnosis of a focal liver mass during a routine US examination before a further diagnostic imaging study such as contrast-enhanced US, CT or MRI. However, further studies with larger numbers of cases are warranted to assess the utility of ARFI elastography in the clinic.

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COMMENTS

Background

Acoustic radiation force impulse (ARFI) elastography can quantify tissue elasticity. Compared to previous elastography techniques, one of the distinguishing advantages of ARFI elastography is that it offers a flexible metering box at variable depths, which allows an examination of specific liver areas. With using this characteristic of ARFI elastography, authors investigated its applicability to evaluate focal liver masses.

Research frontiers

According to previous studies, malignant liver masses are stiffer than benign masses, as reflected by higher ARFI values. However, previous studies have showed somewhat controversial results with various median ARFI values for each focal liver mass and they have had limitations, such as most subjects not being supported by adequate histological confirmation.

Innovations and breakthroughs

Authors prospectively investigated 50 focal liver masses found during routine ultrasonography (US) study, and measured the stiffness of each focal liver mass using ARFI elastography. Focal liver masses included hepatocellular carcinomas (HCCs), cholangiocellular carcinomas, metastases, hemangiomas, and focal nodular hyperplasia. The advantage of our study over other previous studies is that majority of our subjects (72.3%) are confirmed with histological examinations.

Applications

Using ARFI elastography, physicians who are confronted with a need for an immediate diagnosis of a focal liver mass during a routine US examination can get additional information for the differential diagnosis of liver masses in clinical practice. Further future studies with larger numbers of cases are warranted to assess the utility of ARFI elastography.

Terminology

ARFI elastography is an emerging examination which can quantify tissue elasticity. ARFI elastography has the distinct advantage of being integrated into a conventional US system and can be checked simultaneously during a conventional US study. Furthermore, because ARFI elastography uses elastography with a flexible metering box of the region of the interest, it is suitable for quantifying focal liver mass stiffness.

Peer review

Although this study had limitations and should be interpreted cautiously, their results show the clinical applicability of ARFI elastography as a complementary diagnostic tool for the differential diagnosis of liver masses.

REFERENCES

- 1 Assy N, Nasser G, Djibre A, Beniashvili Z, Elias S, Zidan J. Characteristics of common solid liver lesions and recommendations for diagnostic workup. *World J Gastroenterol* 2009; **15**: 3217-3227 [PMID: 19598296]
- 2 Trillaud H, Bruel JM, Valette PJ, Vilgrain V, Schmutz G, Oyen R, Jakubowski W, Danes J, Valek V, Greis C. Characterization of focal liver lesions with SonoVue-enhanced sonography: international multicenter-study in comparison to CT and MRI. *World J Gastroenterol* 2009; **15**: 3748-3756 [PMID: 19673015]
- 3 Vilgrain V. Advancement in HCC imaging: diagnosis, staging and treatment efficacy assessments: hepatocellular carcinoma: imaging in assessing treatment efficacy. *J Hepatobiliary Pancreat Sci* 2010; **17**: 374-379 [PMID: 19924373 DOI: 10.1007/s00534-009-0230-3]
- 4 Hohmann J, Albrecht T, Hoffmann CW, Wolf KJ. Ultrasonographic detection of focal liver lesions: increased sensitivity and specificity with microbubble contrast agents. *Eur J Radiol* 2003; **46**: 147-159 [PMID: 12714231]
- 5 Numminen K, Isoniemi H, Halavaara J, Tervahartiala P, Makisalo H, Laasonen L, Hockerstedt K. Preoperative assessment of focal liver lesions: multidetector computed tomography challenges magnetic resonance imaging. *Acta Radiol* 2005; **46**: 9-15 [PMID: 15841734]
- 6 Gallotti A, D'Onofrio M, Romanini L, Cantisani V, Pozzi Mucelli R. Acoustic Radiation Force Impulse (ARFI) ultrasound imaging of solid focal liver lesions. *Eur J Radiol* 2012; **81**: 451-455 [PMID: 21330078 DOI: 10.1016/j.ejrad.2010.12.071]
- 7 Fahey BJ, Nightingale KR, Nelson RC, Palmeri ML, Trahey GE. Acoustic radiation force impulse imaging of the abdomen: demonstration of feasibility and utility. *Ultrasound Med Biol* 2005; **31**: 1185-1198 [PMID: 16176786 DOI: 10.1016/j.ultrasmedbio.2005.05.004]
- 8 Nightingale K, Palmeri M, Trahey G. Analysis of contrast in images generated with transient acoustic radiation force. *Ultrasound Med Biol* 2006; **32**: 61-72 [PMID: 16364798 DOI: 10.1016/j.ultrasmedbio.2005.08.008]
- 9 Piscaglia F, Salvatore V, Di Donato R, D'Onofrio M, Gualandi S, Gallotti A, Peri E, Borghi A, Conti F, Fattovich G, Sagrini E, Cucchetti A, Andreone P, Bolondi L. Accuracy of VirtualTouch Acoustic Radiation Force Impulse (ARFI) imaging for the diagnosis of cirrhosis during liver ultrasonography. *Ultraschall Med* 2011; **32**: 167-175 [PMID: 21321842 DOI: 10.1055/s-0029-1245948]
- 10 Palmeri ML, Wang MH, Rouze NC, Abdelmalek MF, Guy CD, Moser B, Diehl AM, Nightingale KR. Noninvasive evaluation of hepatic fibrosis using acoustic radiation force-based shear stiffness in patients with nonalcoholic fatty liver disease. *J Hepatol* 2011; **55**: 666-672 [PMID: 21256907 DOI: 10.1016/j.jhep.2010.12.019]
- 11 Kim SU, Jang HW, Cheong JY, Kim JK, Lee MH, Kim DJ, Yang JM, Cho SW, Lee KS, Choi EH, Park YN, Han KH. The usefulness of liver stiffness measurement using FibroScan in chronic hepatitis C in South Korea: a multicenter, prospective study. *J Gastroenterol Hepatol* 2011; **26**: 171-178 [PMID: 21175811 DOI: 10.1111/j.1440-1746.2010.06385.x]
- 12 Sporea I, Sirli R, Popescu A, Danilă M. Acoustic Radiation Force Impulse (ARFI)—a new modality for the evaluation of liver fibrosis. *Med Ultrason* 2010; **12**: 26-31 [PMID: 21165451]
- 13 Horster S, Mandel P, Zachoval R, Clevert DA. Comparing acoustic radiation force impulse imaging to transient elastography to assess liver stiffness in healthy volunteers with and without valsalva manoeuvre. *Clin Hemorheol Microcirc* 2010; **46**: 159-168 [PMID: 21135491 DOI: 10.3233/ch-2010-1342]
- 14 Friedrich-Rust M, Wunder K, Kriener S, Sotoudeh F, Richter S, Bojunga J, Herrmann E, Poynard T, Dietrich CF, Vermehren J, Zeuzem S, Sarrazin C. Liver fibrosis in viral hepatitis: noninvasive assessment with acoustic radiation force impulse imaging versus transient elastography. *Radiology* 2009; **252**: 595-604 [PMID: 19703889 DOI: 10.1148/radiol.2523081928]
- 15 Lupsor M, Badea R, Stefanescu H, Sparchez Z, Branda H, Serban A, Maniu A. Performance of a new elastographic method (ARFI technology) compared to unidimensional transient elastography in the noninvasive assessment of chronic hepatitis C. Preliminary results. *J Gastrointest Liver Dis* 2009; **18**: 303-310 [PMID: 19795024]
- 16 Boursier J, Isselin G, Fouchard-Hubert I, Oberti F, Dib N, Lebigot J, Bertrais S, Gallois Y, Calès P, Aubé C. Acoustic radiation force impulse: a new ultrasonographic technology for the widespread noninvasive diagnosis of liver fibrosis. *Eur J Gastroenterol Hepatol* 2010; **22**: 1074-1084 [PMID: 20440210 DOI: 10.1097/MEG.0b013e328339e0a1]
- 17 Goertz RS, Zopf Y, Jugl V, Heide R, Janson C, Strobel D, Bernatik T, Haendl T. Measurement of liver elasticity with acoustic radiation force impulse (ARFI) technology: an alternative noninvasive method for staging liver fibrosis in viral hepatitis. *Ultraschall Med* 2010; **31**: 151-155 [PMID: 20306380 DOI: 10.1055/s-0029-1245244]
- 18 Gallotti A, D'Onofrio M, Pozzi Mucelli R. Acoustic Radiation Force Impulse (ARFI) technique in ultrasound with Virtual Touch tissue quantification of the upper abdomen. *Radiol Med* 2010; **115**: 889-897 [PMID: 20082227 DOI: 10.1007/s11547-010-0504-5]
- 19 Takahashi H, Ono N, Eguchi Y, Eguchi T, Kitajima Y, Kawaguchi Y, Nakashita S, Ozaki I, Mizuta T, Toda S, Kudo S, Miyoshi A, Miyazaki K, Fujimoto K. Evaluation of acoustic radiation force impulse elastography for fibrosis staging of chronic liver disease: a pilot study. *Liver Int* 2010; **30**: 538-545 [PMID: 19874490 DOI: 10.1111/j.1478-3231.2009.02130.x]
- 20 Yu H, Wilson SR. Differentiation of benign from malignant liver masses with Acoustic Radiation Force Impulse technique. *Ultrasound Q* 2011; **27**: 217-223 [PMID: 22124386 DOI: 10.1097/RUQ.0b013e318239422e]
- 21 Fahey BJ, Nelson RC, Bradway DP, Hsu SJ, Dumont DM, Trahey GE. In vivo visualization of abdominal malignancies with acoustic radiation force elastography. *Phys Med Biol* 2008; **53**: 279-293 [PMID: 18182703 DOI: 10.1088/0031-9155/53/1/020]
- 22 Cho SH, Lee JY, Han JK, Choi BI. Acoustic radiation force impulse elastography for the evaluation of focal solid hepatic lesions: preliminary findings. *Ultrasound Med Biol* 2010; **36**: 202-208 [PMID: 20018432 DOI: 10.1016/j.ultrasmedbio.2009.10.009]
- 23 Heide R, Strobel D, Bernatik T, Goertz RS. Characterization of focal liver lesions (FLL) with acoustic radiation force impulse (ARFI) elastometry. *Ultraschall Med* 2010; **31**: 405-409 [PMID: 20652853 DOI: 10.1055/s-0029-1245565]
- 24 Davies G, Koenen M. Acoustic radiation force impulse elastography in distinguishing hepatic haemangiomas from metastases: preliminary observations. *Br J Radiol* 2011; **84**: 939-943 [PMID: 21385910 DOI: 10.1259/bjr/97637841]
- 25 Shuang-Ming T, Ping Z, Ying Q, Li-Rong C, Ping Z, Ruitive study.

- Zhen L. Usefulness of acoustic radiation force impulse imaging in the differential diagnosis of benign and malignant liver lesions. *Acad Radiol* 2011; **18**: 810-815 [PMID: 21419668 DOI: 10.1016/j.acra.2011.01.026]
- 26 **Bruix J**, Sherman M. Management of hepatocellular carcinoma. *Hepatology* 2005; **42**: 1208-1236 [PMID: 16250051 DOI: 10.1002/hep.20933]
- 27 **Son CY**, Kim SU, Han WK, Choi GH, Park H, Yang SC, Choi JS, Park JY, Kim do Y, Ahn SH, Chon CY, Han KH. Normal liver elasticity values using acoustic radiation force impulse imaging: a prospective study in healthy living liver and kidney donors. *J Gastroenterol Hepatol* 2012; **27**: 130-136 [PMID: 21679249 DOI: 10.1111/j.1440-1746.2011.06814.x]
- 28 **Quaia E**, Calliada F, Bertolotto M, Rossi S, Garioni L, Rosa L, Pozzi-Mucelli R. Characterization of focal liver lesions with contrast-specific US modes and a sulfur hexafluoride-filled microbubble contrast agent: diagnostic performance and confidence. *Radiology* 2004; **232**: 420-430 [PMID: 15286314 DOI: 10.1148/radiol.2322031401]
- 29 **Lim SM**, Kim SU. Acoustic radiation force impulse elastography is useful to exclude nonliver-related ascites. *Eur J Gastroenterol Hepatol* 2011; **23**: 1080-1081 [PMID: 21971343 DOI: 10.1097/MEG.0b013e32834be9e9]
- 30 **Yoon KT**, Lim SM, Park JY, Kim do Y, Ahn SH, Han KH, Chon CY, Cho M, Lee JW, Kim SU. Liver stiffness measurement using acoustic radiation force impulse (ARFI) elastography and effect of necroinflammation. *Dig Dis Sci* 2012; **57**: 1682-1691 [PMID: 22302243 DOI: 10.1007/s10620-012-2044-4]
- 31 **Lim SM**, Chung MJ, Han KH, Kim SU. Acoustic radiation force impulse elastography: a better option for patients with extrahepatic cholestasis. *Eur J Gastroenterol Hepatol* 2012; **24**: 215-216 [PMID: 22081004 DOI: 10.1097/MEG.0b013e32834e0789]
- 32 **Kwon HJ**, Kang MJ, Cho JH, Oh JY, Nam KJ, Han SY, Lee SW. Acoustic radiation force impulse elastography for hepatocellular carcinoma-associated radiofrequency ablation. *World J Gastroenterol* 2011; **17**: 1874-1878 [PMID: 21528062 DOI: 10.3748/wjg.v17.i14.1874]

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Inositol-requiring enzyme 1 α is required for gut development in *Xenopus laevis* embryos

Jing Guo, Xin-Xin Li, Jiao-Jiao Feng, Chen-Yang Yin, Xue-Jun Wang, Ning Wang, Li Yuan

Jing Guo, Xin-Xin Li, Jiao-Jiao Feng, Chen-Yang Yin, Xue-Jun Wang, Ning Wang, Li Yuan, Department of Biochemistry and Molecular Biology, Nanjing Medical University, Nanjing 210029, Jiangsu Province, China

Author contributions: Guo J, Li XX, Feng JJ and Yin CY performed the majority of the experiments; Wang XJ and Wang N provided vital reagents and analytical tools and revised the manuscript; Yuan L designed the study and wrote the manuscript; all authors read and approved the final manuscript.

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Correspondence to: Li Yuan, PhD, Associate Professor of Biochemistry and Molecular Biology, Department of Biochemistry and Molecular Biology, Nanjing Medical University, Nanjing 210029, Jiangsu Province, China. yuanli@njmu.edu.cn
Telephone: +86-25-86862895 Fax: +86-25-86862728

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Abstract

AIM: To investigate the role of inositol-requiring enzyme 1 α (IRE1 α) in gut development of *Xenopus laevis* embryos.

METHODS: *Xenopus* embryos were obtained with *in vitro* fertilization and cultured in 0.1 \times MBSH. One and half nanogram of IRE1 α , 1 ng of IRE1 α -GR mRNA, 1 ng of IRE1 α ΔC-GR mRNA, and 50 ng of IRE1 α morpholino oligonucleotide (MO) or XBP1(C)MO were injected into four blastomeres at 4-cell stage for scoring the phenotype and marker gene analysis. To rescue the effect of IRE1 α MO, 1 ng of IRE1 α -GR mRNA was co-injected with 50 ng of MO. For the activation of the GR-fusion proteins, dexamethasone was prepared as 5 mmol/L stock solutions in 100% ethanol and applied to the mRNA injected embryos at desired stages in a concentration of 10 μ mol/L in 0.1 \times MBSH. Embryos were

kept in dexamethasone up to stage 41. Whole-mount *in situ* hybridization was used to determine specific gene expression, such as IRE1 α , IRE1 β , Xbra and Xsox17 α . IRE1 α protein expression during *Xenopus* embryogenesis was detected by Western blotting.

RESULTS: In the whole-mount *in situ* hybridization analysis, xenopus IRE1 α and IRE1 β showed quite different expression pattern during tadpole stage. The relatively higher expression of IRE1 α was observed in the pancreas, and significant transcription of IRE1 β was found in the liver. IRE1 α protein could be detected at all developmental stages analyzed, from stage 1 to stage 42. Gain-of-function assay showed that IRE1 α mRNA injected embryos at tailbud stage were nearly normal and the expression of the pan-mesodermal marker gene Xbra and the endodermal gene Xsox17 α at stage 10.5 was not significantly changed in embryos injected with IRE1 α mRNA as compared to uninjected control embryos. And at tadpole stage, the embryos injected with IRE1 α -GR mRNA did not display overt phenotype, such as gut-coiling defect. Loss-of-function assay demonstrated that the IRE1 α MO injected embryos were morphologically normal before the tailbud stages. We did not observe a significant change of mesodermal and endodermal marker gene expression, while after stage 40, about 80% of the MO injected embryos exhibited dramatic gut defects in which the guts did not coil, but other structures outside the gastrointestinal tract were relatively normal. To test if the phenotypes were specifically caused by the knockdown of IRE1 α , a rescue experiment was performed by co-injection of IRE1 α -GR mRNA with IRE1 α MO. The data obtained demonstrated that the gut coiling defect was rescued. The deletion mutant of IRE1 α was constructed, consisting of the N-terminal part without the C-terminal kinase and RNase domains named IRE1 α ΔC, to investigate the functional domain of IRE1 α . Injection of IRE1 α ΔC-GR mRNA caused similar morphological alterations with gut malformation by interfering with the function of endogenous xIRE1 α . In order to investigate if IRE1 α /

XBP1 pathway was involved in gut development, 50 ng of XBP1 MO was injected and the results showed that knockdown of XBP1 resulted in similar morphological alterations with gut-coiling defect at tadpole stage.

CONCLUSION: IRE1 α is not required for germ layer formation but for gut development in *Xenopus laevis* and it may function *via* XBP1-dependent pathway.

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Key words: Inositol-requiring enzyme 1 α ; XBP1; *Xenopus laevis*; Gut; Development

Guo J, Li XX, Feng JJ, Yin CY, Wang XJ, Wang N, Yuan L. Inositol-requiring enzyme 1 α is required for gut development in *Xenopus laevis* embryos. *World J Gastroenterol* 2013; 19(2): 227-234 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v19/i2/227.htm> DOI: <http://dx.doi.org/10.3748/wjg.v19.i2.227>

INTRODUCTION

The endoplasmic reticulum (ER) plays a central role in the synthesis and modification of secretory and membrane proteins in all eukaryotic cells^[1-3]. Under normal conditions, these proteins are correctly folded and assembled in the ER. However, when cells are exposed to disturbed environment, such as overproduction of ER proteins, viral infection, and glucose deprivation, these proteins accumulate as unfolded or misfolded forms in the ER lumen and, consequently, cause ER stress. To maintain cellular homeostasis, cells induce some adaptive responses to ER stress. One of them is the unfolded protein response (UPR), which up-regulates the transcription of various genes to increase the protein-folding and protein-degradation activity in the ER^[4-7]. Inositol requiring enzyme-1 (IRE1) is an ER-located type I transmembrane protein with a kinase domain and RNase domain in the cytosolic region and has a unique function of relieving ER stress in cells. When the amino-terminal luminal region senses perturbations in the ER environment, *via* trans-autophosphorylation and activation of its RNase domain, IRE1 induces unconventional splicing of mRNA coding, a specific transcription factor for activating the UPR^[8-10]. IRE1 is highly conserved from yeast to humans, and two IRE1 paralogues have been reported in mammals: IRE1 α and IRE1 β ^[11-13]. Upon activation by ER stress, IRE1 performs an unconventional cytoplasmic splicing of XBP1 pre-mRNA, thus allowing the synthesis of active XBP1, which activates UPR target genes to restore the homeostasis of the ER^[10,14,15]. The spliced XBP1 mRNA is translated into a functional transcription factor to up-regulate gene expression for ER quality control. IRE1 is also reported to activate proapoptotic JNK signaling under ER stress conditions^[16].

IRE1 α is also known to be expressed ubiquitously in fetal and adult mice and to be essential for mammalian

developmental processes^[17]. Therefore, IRE1 α inactivation results in widespread developmental defects, leading to embryonic death after 12.5 d of gestation in mice^[18]. However, the cause of this embryonic lethality is not fully understood. These lines of evidence suggest that IRE1 α has a unique function in mammalian developmental processes, but it has been hitherto unclear in which tissues and how IRE1 α functions during embryogenesis.

Xenopus laevis is an excellent model system for studying organ development^[19]. However, little information is available about the function of the IRE1/XBP1 pathway during embryogenesis, although loss of function studies revealed that IRE1 or XBP1 is absolutely required for embryonic development of *C. elegans*^[20], *Drosophila*^[21] or mouse^[22]. In *Xenopus*, although the transcripts of two isoforms of IRE1, IRE1 α and IRE1 β show similar spatial expression in pre-neurula embryos, they are also differentially expressed following the onset of neurulation^[23]. IRE1 α is localized to the nervous system and mesoderm or endoderm-derived organs, such as pronephros and pancreas. However, the role of IRE1 α in organogenesis is still unclear. In the present study, both gain and loss of function analyses revealed that IRE1 α is required for gut development in *Xenopus*.

MATERIALS AND METHODS

Embryos

Wild type *Xenopus laevis* eggs were obtained by injecting 1000 IU of human chorionic gonadotrophin into the dorsal lymph sacs of adult females 6-8 h before egg collection. Eggs were fertilized *in vitro* with minced testes, dejellied with 2% cysteine hydrochloride (pH 7.8-8.0) 30 min after fertilization, and cultured in 0.1 \times MBSH (8.8 mmol/L NaCl, 0.24 mmol/L NaHCO₃, 0.1 mmol/L KCl, 0.082 mmol/L MgSO₄, 0.041 mmol/L CaCl₂, 0.033 mmol/L Ca(NO₃)₂, 1 mmol/L HEPES, pH 7.4). Staging of *Xenopus laevis* embryos was according to Nieuwkoop and Faber (1967).

Plasmids and constructs

IRE1 α ORF was amplified from a cDNA pool consisting of st.1, st.8, st.10, st.15, st.20 and st.28 cDNAs and subcloned to pCS2⁺ vector. The construct was named pCS2⁺-IRE1 α . To make expression constructs, complete coding region and the N-terminal region (aa 1-479) were amplified from the pCS2⁺-IRE1 α using PCR and subcloned to the BamHI-EcoRV sites on pCS2⁺-GR. The resulting constructs were designated as pCS2⁺IRE1 α -GR and pCS2⁺IRE1 α Δ C-GR.

In vitro RNA synthesis, antisense morpholino oligonucleotides and microinjection

Plasmids pCS2⁺IRE1 α , pCS2⁺IRE1 α -GR and pCS2⁺IRE1 α Δ C-GR were linearized with *NheI*. Capped mRNA for microinjection was synthesized with SP6 mMessage mMachineTM kit (Ambion) and cleaned up with RNeasy kit (Qiagen). The antisense morpholino

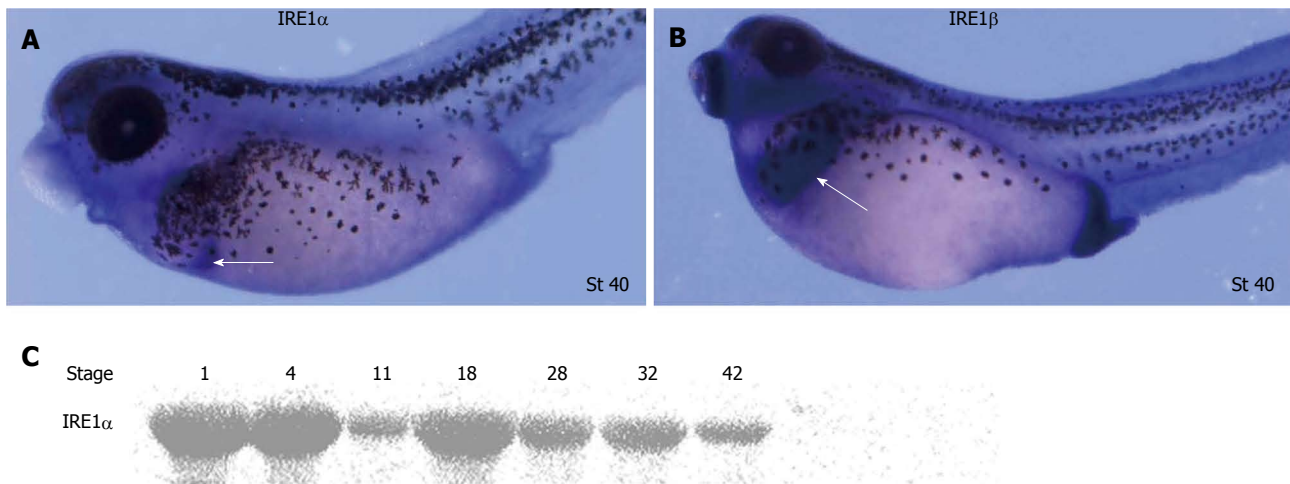


Figure 1 Expression pattern of xIRE1 in *Xenopus laevis* during development. A, B: Whole-mount *in situ* hybridizations revealed relatively high expression of inositol-requiring enzyme (IRE) 1 α in pancreas (white arrow in A) and relatively high expression of IRE1 β in liver (white arrow in B); C: Western blotting revealed temporal expression of IRE1 α during *Xenopus* embryogenesis.

oligonucleotide (Gene Tools) used for IRE1 α functional knockdown (IRE1 α MO) was: 5'-AAGAGAACCGC-CAGAGGC GCCATG T-3'; and an antisense morpholino oligonucleotide XBP1(C) MO designed to inhibit the cytoplasmic splicing of xXBP1 was: 5'-GACATCT-GGGCCTGCTC CTGC TGCA-3'. One and half nanogram of IRE1 α , 1 ng of IRE1 α -GR mRNA, 1 ng of IRE1 α Δ C-GR mRNA, and 50 ng of IRE1 α MO or XBP1(C)MO were injected into four blastomeres at 4-cell stage for scoring the phenotype, whole-mount *in situ* hybridization and marker gene analysis. For the activation of the GR-fusion proteins, dexamethasone (Sigma) was prepared as 5 mmol/L stock solutions in 100% ethanol and applied to the control and mRNA injected embryos at desired stages in a concentration of 10 μ mol/L in 0.1 \times MBS. Embryos were kept in dexamethasone up to stage 41. To rescue the effect of blocking xXBP1 splicing by IRE1 α MO, 1 ng of IRE1 α -GR mRNA was co-injected with 50 ng of MO.

***In vitro* translation**

In vitro protein translation was performed with TNT coupled Reticulocyte Lysate Systems (Promega) to test the efficiency of IRE1 α MO for blocking protein translation. One μ g of pCS2⁺IRE1 α plasmid was used either alone or together with 20 μ g IRE1 α MO for *in vitro* translation. Translation products were subjected to sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and chemiluminescent detection.

Whole-mount *in situ* hybridization

Whole-mount *in situ* hybridization was performed according to the protocol described elsewhere^[24]. The digoxigenin-labeled antisense probes were prepared as follows: pDrive-IRE1 α cut with *Hind*III, transcribed with T7 RNA polymerase; pCS2⁺XIRE1 β cut with *Xba*I, transcribed with T3 RNA polymerase; pBS-Xbra cut with *Sa*I, transcribed with T7 RNA polymerase; and pCS2⁺XSox17 α cut with *Cl*aI, transcribed with T7 RNA

polymerase.

Protein extraction and Western blotting

Embryos were homogenized in RIPA lysis buffer (20 mmol/L Tris, pH 8; 2 mmol/L EDTA, pH 8; 0.5% NP-40; 25 mmol/L β glycerophosphate; 100 mmol/L NaF; 100 mmol/L PMSF and phosphatase inhibitor cocktail). Lysates were centrifuged at 4 $^{\circ}$ C, 15 000 $\times g$, for 20 min, and the supernatants were added to 5 \times SDS loading buffer. Proteins were separated by 10% SDS-PAGE and transferred to nitrocellulose membrane (Millipore, Billerica, MA). Immunoblotting membranes were blocked with 5% milk in TBST. After several washes in TBST, membranes were incubated overnight with the first antibody [anti-IRE1 α antibody (Santa Cruz Biotechnology, Santa Cruz, CA), 1:500]. Loading controls of presumably constantly expressed proteins such as β -actin were used; however, their variability and increase in development precluded their use^[25]. Detection was then done with HRP-labeled secondary antibodies and enhanced chemiluminescence (ECL).

RESULTS

Expression of xIRE1 α in *Xenopus laevis* during development

Two forms of IRE1 genes, IRE1 α and IRE1 β , exist in *Xenopus laevis*. Whole-mount *in situ* hybridization revealed that IRE1 α and IRE1 β were expressed in a similar pattern from egg to gastrulation. In tailbud embryos, IRE1 α was detected in a domain that probably represented the dorsal pancreas anlage. IRE1 β was only observed in the hatching gland and cement gland until the hatched tadpole stage^[23]. To further explore the spatial expression patterns of IRE1 α and IRE1 β in *Xenopus* embryos at later stages, whole-mount *in situ* hybridizations were carried out. During tadpole stages, relatively high expression of IRE1 α was observed in the pancreas (Figure 1A), and significant transcription of IRE1 β was observed in the

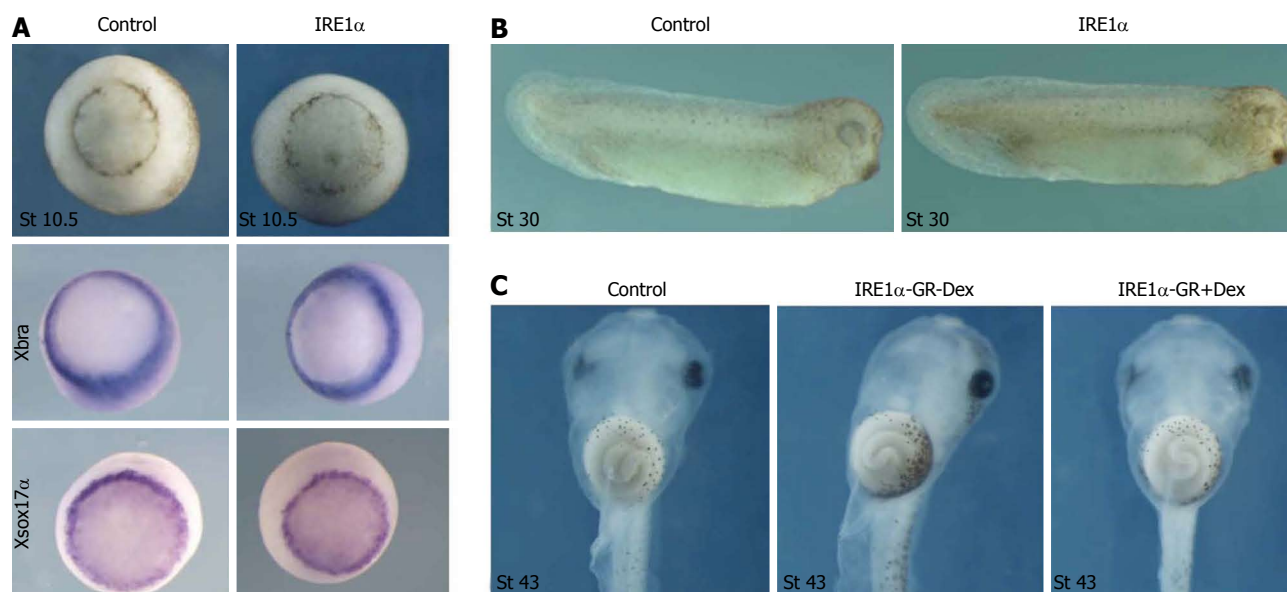


Figure 2 Overexpression of inositol-requiring enzyme 1 α affected neither mesoderm and endoderm formation nor gut development. A: Overexpression of inositol-requiring enzyme (IRE) 1 α did not change the phenotype and the expression of Xbra and Xsox17 α at stage 10.5; B: IRE1 α mRNA injected embryos at tailbud stage were nearly normal; C: Embryos injected with IRE1 α -GR mRNA did not display any defect until tadpole stage.



Figure 3 Inositol-requiring enzyme 1 α morpholino oligonucleotide blocks inositol-requiring enzyme 1 α translation in an *in vitro* transcription/translation assay. In this assay, xIRE1 α was effectively transcribed/translated from a pCS2+xIRE1 α construct. However, translation was dramatically decreased by addition of IRE1 α morpholino oligonucleotide (MO). IRE: Inositol-requiring enzyme.

liver (Figure 1B). IRE1 α protein could be detected at all developmental stages analyzed (Figure 1C).

IRE1 α gain of function

Different doses of IRE1 α mRNA were injected into all blastomeres at 4-cell stage. Even at a dose of 1.5 ng, injected embryos at tailbud stage were nearly normal (Figure 2B). Whole-mount *in situ* hybridization of embryos injected with IRE1 α mRNA revealed that the expression of the pan-mesodermal marker gene Xbra and the endodermal gene Xsox17 α at stage 10.5 was not significantly changed as compared to uninjected control embryos (Figure 2A). To test the function of IRE1 α in the later stage of development, the mRNA encoding a dexamethasone inducible variant of IRE1 α , referred to

as IRE1 α -GR, was injected into all four blastomeres at 4-cell stage *Xenopus* embryos. For the activation of the GR-fusion proteins, dexamethasone treatment was administered at embryonic stage 27. No apparent change of phenotype was observed even at tadpole stage (Figure 2C). Therefore, gain of function of IRE1 α did not lead to an apparent change of phenotype and we did not observe a significant change of mesodermal and endodermal marker gene expression.

IRE1 α MO blocking IRE1 α translation

To further explore the function of xIRE1 α during embryonic development, we performed a loss of function (LOF) analysis using an antisense morpholino oligonucleotide (IRE1 α MO) directed against xIRE1 α . *In vitro* protein translation was performed with TNT coupled Reticulocyte Lysate Systems to test the efficiency of IRE1 α MO for blocking protein translation. As shown in Figure 3, IRE1 α MO totally blocked IRE1 α translation.

IRE1 α loss of function

The IRE1 α MO-injected embryos were morphologically normal before the tailbud stages (Figure 4B) and we did not observe a significant change of mesodermal and endodermal marker gene expression (Figure 4A), but after stage 40, about 80% of the MO (50 ng)-injected embryos exhibited dramatic gut defects in which the guts did not coil, while other structures outside the gastrointestinal tract developed normally (Figure 4). To test if the phenotypes were specifically caused by the knockdown of IRE1 α , a rescue experiment was performed by co-injection of IRE1 α -GR mRNA with MO. The results indicated that the phenotype could be rescued (Figure 5). Taken together, these data indicate that IRE1 α is re-

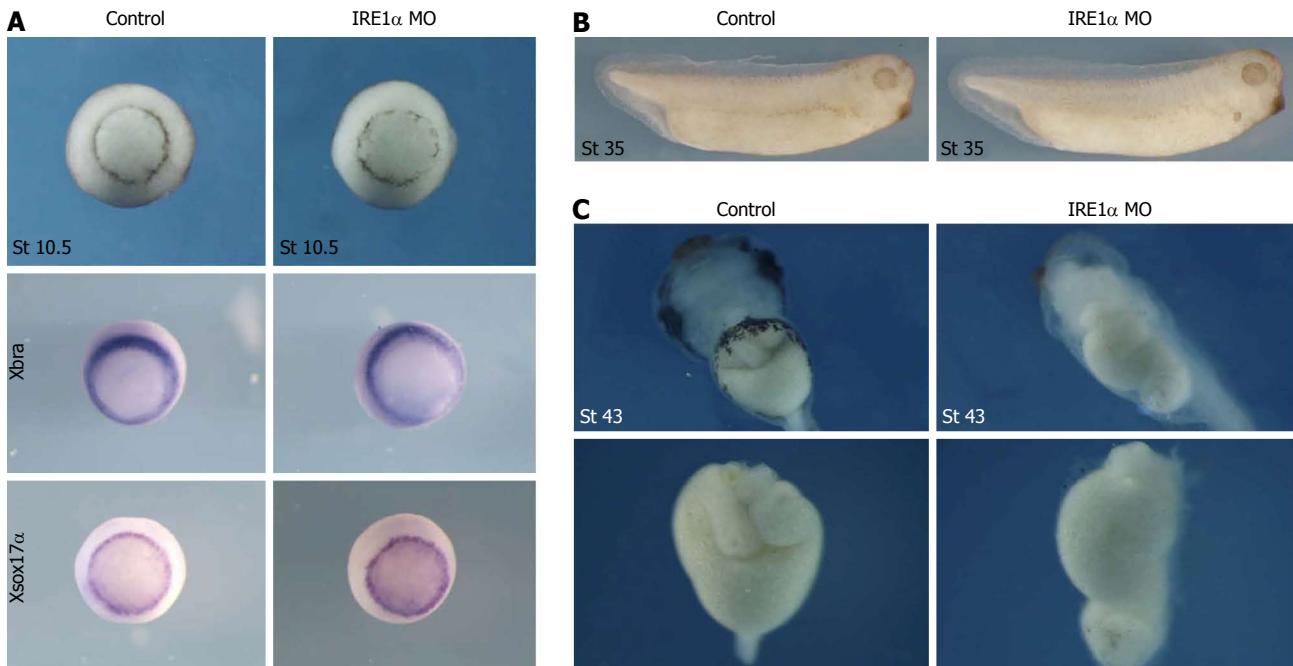


Figure 4 *Xenopus* inositol-requiring enzyme 1 α is required for gut development. A: Knockdown of x IRE1 α did not change the phenotype and the expression of Xbra and Xsox17 α at stage 10.5; B: The inositol-requiring enzyme (IRE) 1 α morpholino oligonucleotide (MO)-injected embryos were morphologically normal before the tailbud stages; C: IRE1 α knockdown upon injection of 50 ng of MO resulted in a gut defective phenotype. Surgically-resected guts from embryos were shown in C under panel. Coiled structure of gut was not detected in IRE1 α MO-injected embryos at stage 43. MO injection was repeated 5 times in a total of 278 embryos.



Figure 5 Rescue of xIRE1 α knockdown with IRE1 α -GR mRNA. Gut defective phenotype caused by 50 ng of morpholino oligonucleotide (MO) could be rescued by co-injection of 1 ng of inositol-requiring enzyme (IRE) 1 α -GR mRNA. Co-injection was done 3 times in a total of 258 embryos.

quired for proper gut development in *Xenopus*.

Deletion mutant of IRE1 α

IRE1 α is a Ser/Thr protein kinase and endoribonuclease that, upon activation, initiates the unconventional splicing of the mRNA of XBP1^[12]. The deletion mutant was constructed consisting of the N-terminal part without the C-terminal kinase and RNase domains (Figure 6A). Injection of IRE1 α Δ C-GR mRNA caused similar morphological alterations with gut malformation by interfering with the function of endogenous xIRE1 α (Figure 6B-D). These data further confirmed the notion that IRE1 α Δ C represents a dominant-negative form of IRE1 α .

XBP1 loss of function

IRE1 α is the most evolutionarily conserved branch of the UPR, and upon activation, initiates the unconventional splicing of the mRNA encoding the transcriptional

factor XBP1 to attenuate ER stress by mediating UPR. In other words, IRE1 α mediates XBP1 splicing. IRE1 α MO could knockdown IRE1 α expression, and then XBP1 splicing would be repressed. To test if knockdown of XBP1 might cause similar phenotype with IRE1 α knockdown, 50 ng of XBP1(C)MO was injected into four blastomeres at 4-cell stage for scoring the phenotype. As shown in Figure 7, injection of XBP1(C)MO caused similar phenotype with gut-coiling defect.

DISCUSSION

In eukaryotic cells, ER is responsible for the early steps in the maturation of most proteins in the secretory pathway, such as folding of the newly synthesized polypeptide chains and post-translational modifications that are essential for protein function^[26,27]. Nascent polypeptides are translocated to the ER lumen in an unfolded state, where

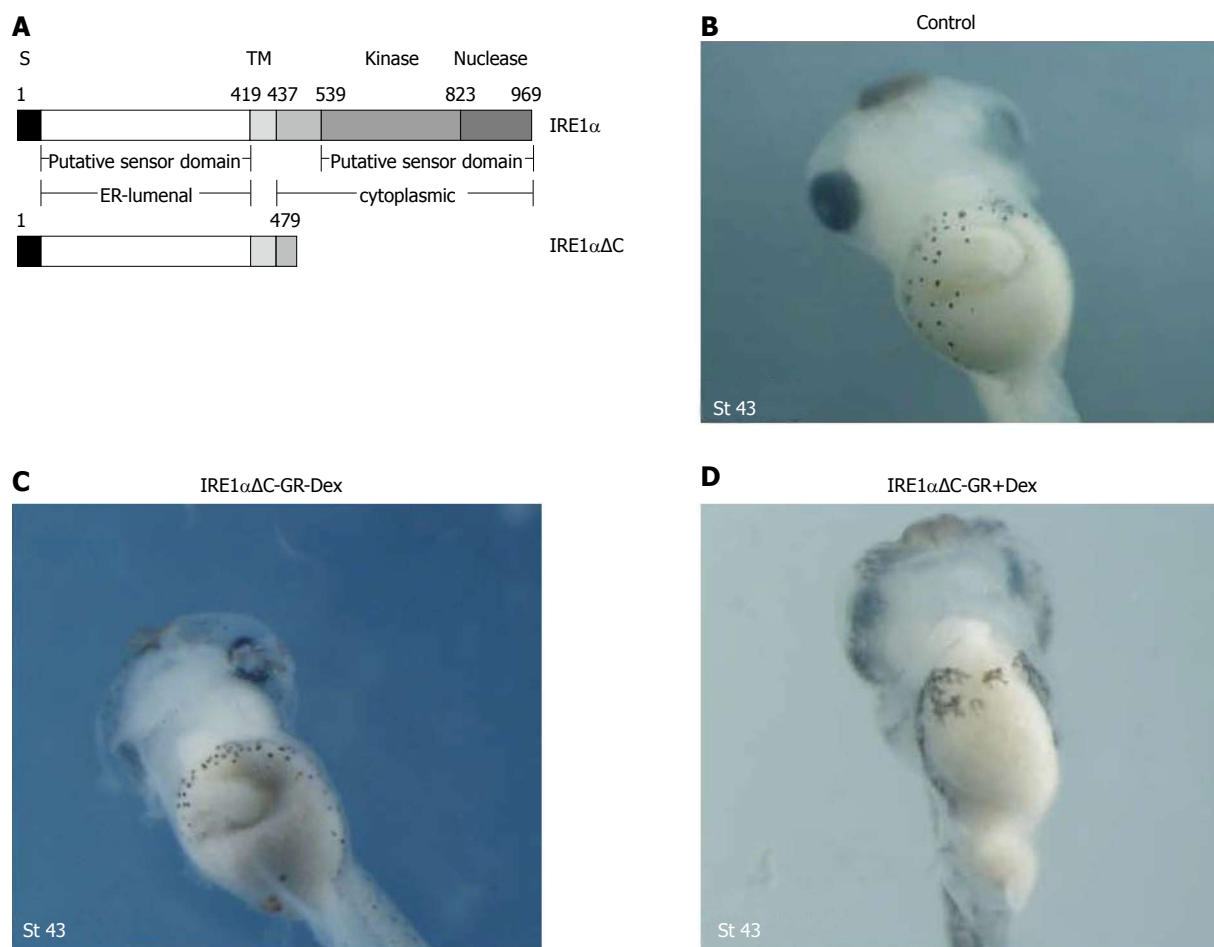


Figure 6 Injection of IRE1 α ΔC-GR mRNA caused morphological alterations with gut malformation. A: Diagram depicting the construction of deletion mutants. S and TM denote the signal peptide and the transmembrane domain; B-D: Gut defects were also observed with a dominant-negative mutant, IRE1 α ΔC, lacking the cytoplasmic kinase and RNase domains.

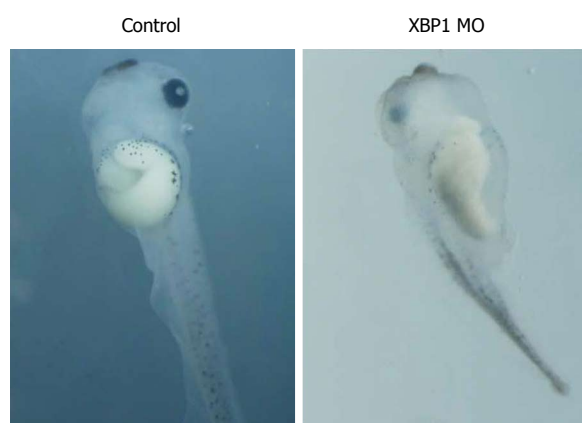


Figure 7 XBP1 knockdown affected gut development. Injection of XBP1(C) morpholino oligonucleotide (MO) caused similar phenotype with gut-coiling defect at stage 43.

they are processed for folding. However, the function of ER will be disrupted when the inflow of unfolded polypeptide chains exceeds the folding or processing capacity of the ER^[28,29]. This ER stress in turn leads to the activation of a series of adaptive pathways known as UPR to maintain ER homeostasis^[30]. The adaptive process, UPR,

has at least two distinct components^[4]. The first consists of the rapid and transient attenuation of new protein synthesis and can be considered to be an attempt on the part of the cell to limit the load on the folding apparatus in the ER. The second component consists of the upregulation of expression of genes whose products promote protein folding in the ER and degradation of malformed proteins. The latter UPR signaling pathways are transduced by three ER resident transmembrane proteins IRE1, PERK and ATF6 upon activation^[14], among which IRE1 functions as an endoribonuclease (RNase) to process XBP1 pre-mRNA to a mature form. Spliced XBP1 is a transcriptional activator that plays a fundamental role in the activation of a wide variety of UPR target genes^[31,32]. Two mammalian homologues of yeast IRE1 have been identified: IRE1 α and IRE1 β . IRE1 α is expressed in most cells and tissues, with highest levels of expression in the pancreas and placenta^[17]. IRE1 β expression is prominent only in intestinal epithelial cells^[13].

Accumulation of malformed proteins in the ER occurs under many pathophysiological conditions, in addition, it also takes place during embryonic development. During embryogenesis of vertebrates, such as *Xenopus*, germ layer induction, pattern formation, and morphoge-

netic movement are known to be mediated by secreted proteins, including fibroblast growth factors (FGFs)^[33], TGF- β /nodal/BMPs, and Wnts^[34,35]. Therefore, dysfunction of the ER should also interfere with the secretion of these proteins and consequently disrupt early embryonic development^[36].

Although the effect of ER stress on cellular physiology has been extensively investigated, little is known so far about how it affects early embryonic development. The previous study has verified the conservation of IRE1/XBP1 pathway in *Xenopus* embryos and the importance of IRE1 β for mesoderm formation in *Xenopus* embryos^[23]. In this study, we demonstrated that (1) IRE1 α mainly expressed in pancreas at tadpole stages and IRE1 α protein could be detected at all developmental stages analyzed (Figure 1); (2) although it did not cause overt phenotypes upon overexpression (Figure 2), specific knockdown of IRE1 α (Figure 4) or XBP1 (Figure 7) led to a gut-coiling defect, and injection of xIRE1 α Δ C-GR mRNA caused similar morphological alterations with gut malformation (Figure 6); and (3) IRE1 α -GR mRNA can rescue IRE1 knockdown phenotypes (Figure 5).

In conclusion, our loss and gain of function data support the notion that IRE1 α is required for gut development in *Xenopus* embryos. Our results demonstrate that homeostasis of ER and xIRE1 α functions are required for gut development in *Xenopus* embryos. We infer from our results that IRE1 regulates gut development through IRE1 α -XBP1 pathway.

COMMENTS

Background

Inositol-requiring enzyme 1 α (IRE1 α) is an endoplasmic reticulum (ER)-located type I transmembrane protein with a kinase domain and RNase domain in the cytosolic region. IRE1 α induces the unconventional splicing of XBP1 mRNA under ER stress condition. However, a XBP1-independent IRE1 α function also exists. IRE1 α is expressed ubiquitously in fetal and adult mice and is essential for mammalian developmental processes. However, the function of IRE1 α in specific organs and tissues remains incompletely understood.

Research frontiers

IRE1 α is confirmed to be essential during mammalian development. However, the IRE1 α conventional knockout mice showed embryonic lethality, and it has been reported that during development IRE1 α is required for B-cell differentiation, placental development and embryonic viability. And the function of IRE1 α in specific organs and tissues deserves to be illustrated.

Innovations and breakthroughs

Compared with previous studies, this study used the *Xenopus laevis* as animal model to study the function of IRE1 α during their development. After overexpressing and knockdown of the IRE1 α , the phenotype could be analyzed. And the results showed that at early stage, IRE1 α did not play a significant role in germ layer formation, however, at the stage of organogenesis, knockdown of IRE1 α or XBP1 caused gut-coiling defect. The results suggested that IRE1 α is not required for germ layer formation, but for gut development in *Xenopus laevis* and it may function via XBP1-dependent pathway.

Applications

The results in this study showed that IRE1 α does play a role in organogenesis, however, two questions remain to be answered: which organs or tissues in gut are affected and what is the underlying mechanism.

Peer review

This is an interesting article. The authors show in a systematic manner that the role of IRE1 α in the gut development of *Xenopus*. They also show that it acts through the XBP1 processing.

REFERENCES

- 1 Malhotra JD, Kaufman RJ. The endoplasmic reticulum and the unfolded protein response. *Semin Cell Dev Biol* 2007; **18**: 716-731 [PMID: 18023214 DOI: 10.1016/j.semcdb.2007.09.003]
- 2 Anelli T, Sitia R. Protein quality control in the early secretory pathway. *EMBO J* 2008; **27**: 315-327 [PMID: 18216874 DOI: 10.1038/sj.emboj.7601974]
- 3 Schröder M, Kaufman RJ. The mammalian unfolded protein response. *Annu Rev Biochem* 2005; **74**: 739-789 [PMID: 15952902 DOI: 10.1146/annurev.biochem.73.011303.074134]
- 4 Credle JJ, Finer-Moore JS, Papa FR, Stroud RM, Walter P. On the mechanism of sensing unfolded protein in the endoplasmic reticulum. *Proc Natl Acad Sci USA* 2005; **102**: 18773-18784 [PMID: 16365312 DOI: 10.1073/pnas.0509487102]
- 5 Xu C, Bailly-Maitre B, Reed JC. Endoplasmic reticulum stress: cell life and death decisions. *J Clin Invest* 2005; **115**: 2656-2664 [PMID: 16200199 DOI: 10.1172/JCI26373]
- 6 Wu J, Kaufman RJ. From acute ER stress to physiological roles of the Unfolded Protein Response. *Cell Death Differ* 2006; **13**: 374-384 [PMID: 16397578 DOI: 10.1038/sj.cdd.4401840]
- 7 Boyce M, Yuan J. Cellular response to endoplasmic reticulum stress: a matter of life or death. *Cell Death Differ* 2006; **13**: 363-373 [PMID: 16397583 DOI: 10.1038/sj.cdd.4401817]
- 8 Kimata Y, Ishiwata-Kimata Y, Ito T, Hirata A, Suzuki T, Oikawa D, Takeuchi M, Kohno K. Two regulatory steps of ER-stress sensor Ire1 involving its cluster formation and interaction with unfolded proteins. *J Cell Biol* 2007; **179**: 75-86 [PMID: 17923530 DOI: 10.1083/jcb.200704166]
- 9 Oikawa D, Kimata Y, Kohno K, Iwawaki T. Activation of mammalian IRE1 α upon ER stress depends on dissociation of BiP rather than on direct interaction with unfolded proteins. *Exp Cell Res* 2009; **315**: 2496-2504 [PMID: 19538957 DOI: 10.1016/j.yexcr.2009.06.009]
- 10 Calton M, Zeng H, Urano F, Till JH, Hubbard SR, Harding HP, Clark SG, Ron D. IRE1 couples endoplasmic reticulum load to secretory capacity by processing the XBP-1 mRNA. *Nature* 2002; **415**: 92-96 [PMID: 11780124 DOI: 10.1038/415092a]
- 11 Shen X, Zhang K, Kaufman RJ. The unfolded protein response--a stress signaling pathway of the endoplasmic reticulum. *J Chem Neuroanat* 2004; **28**: 79-92 [PMID: 15363493 DOI: 10.1016/j.jchemneu.2004.02.006]
- 12 Liu CY, Kaufman RJ. The unfolded protein response. *J Cell Sci* 2003; **116**: 1861-1862 [PMID: 12692187 DOI: 10.1242/jcs.00408]
- 13 Wang XZ, Harding HP, Zhang Y, Jolicoeur EM, Kuroda M, Ron D. Cloning of mammalian Ire1 reveals diversity in the ER stress responses. *EMBO J* 1998; **17**: 5708-5717 [PMID: 9755171 DOI: 10.1093/emboj/17.19.5708]
- 14 Yoshida H, Matsui T, Yamamoto A, Okada T, Mori K. XBP1 mRNA is induced by ATF6 and spliced by IRE1 in response to ER stress to produce a highly active transcription factor. *Cell* 2001; **107**: 881-891 [PMID: 11779464 DOI: 10.1016/S0092-8674(01)00611-0]
- 15 Oikawa D, Tokuda M, Hosoda A, Iwawaki T. Identification of a consensus element recognized and cleaved by IRE1 α . *Nucleic Acids Res* 2010; **38**: 6265-6273 [PMID: 20507909 DOI: 10.1093/nar/gkq452]
- 16 Urano F, Wang X, Bertolotti A, Zhang Y, Chung P, Harding HP, Ron D. Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1. *Science* 2000; **287**: 664-666 [PMID: 10650002 DOI: 10.1126/science.287.5453.664]
- 17 Tirasophon W, Welihinda AA, Kaufman RJ. A stress response pathway from the endoplasmic reticulum to the nucleus requires a novel bifunctional protein kinase/endonuclease (Ire1p) in mammalian cells. *Genes Dev* 1998; **12**: 1812-1824 [PMID: 9637683 DOI: 10.1101/gad.12.12.1812]

- 18 **Zhang K**, Wong HN, Song B, Miller CN, Scheuner D, Kaufman RJ. The unfolded protein response sensor IRE1 α is required at 2 distinct steps in B cell lymphopoiesis. *J Clin Invest* 2005; **115**: 268-281 [PMID: 15690081 DOI: 10.1172/JCI200521848]
- 19 **Kashiwagi K**, Kashiwagi A, Kurabayashi A, Hanada H, Nakajima K, Okada M, Takase M, Yaoita Y. *Xenopus tropicalis*: an ideal experimental animal in amphibia. *Exp Anim* 2010; **59**: 395-405 [PMID: 20660986 DOI: 10.1538/expanim.59.395]
- 20 **Shen X**, Ellis RE, Lee K, Liu CY, Yang K, Solomon A, Yoshida H, Morimoto R, Kurnit DM, Mori K, Kaufman RJ. Complementary signaling pathways regulate the unfolded protein response and are required for *C. elegans* development. *Cell* 2001; **107**: 893-903 [PMID: 11779465 DOI: 10.1016/S0092-8674(01)00612-2]
- 21 **Souid S**, Lepesant JA, Yanicostas C. The xbp-1 gene is essential for development in *Drosophila*. *Dev Genes Evol* 2007; **217**: 159-167 [PMID: 17206451 DOI: 10.1007/s00427-006-0124-1]
- 22 **Reimold AM**, Etkin A, Clauss I, Perkins A, Friend DS, Zhang J, Horton HF, Scott A, Orkin SH, Byrne MC, Grusby MJ, Glimcher LH. An essential role in liver development for transcription factor XBP-1. *Genes Dev* 2000; **14**: 152-157 [PMID: 10652269 DOI: 10.1101/gad.14.2.152]
- 23 **Yuan L**, Cao Y, Oswald F, Knöchel W. IRE1 β is required for mesoderm formation in *Xenopus* embryos. *Mech Dev* 2008; **125**: 207-222 [PMID: 18191552 DOI: 10.1016/j.mod.2007.11.010]
- 24 **Gawantka V**, Pollet N, Delius H, Vingron M, Pfister R, Nitsch R, Blumenstock C, Niehrs C. Gene expression screening in *Xenopus* identifies molecular pathways, predicts gene function and provides a global view of embryonic patterning. *Mech Dev* 1998; **77**: 95-141 [PMID: 9831640 DOI: 10.1016/S0925-4773(98)00115-4]
- 25 **Yashpal NK**, Li J, Wheeler MB, Wang R. Expression of β 1 integrin receptors during rat pancreas development-sites and dynamics. *Endocrinology* 2005; **146**: 1798-1807 [PMID: 15618357 DOI: 10.1210/en.2004-1292]
- 26 **Novosyadlyy R**, Kurshan N, Lann D, Vijayakumar A, Yakar S, LeRoith D. Insulin-like growth factor-I protects cells from ER stress-induced apoptosis via enhancement of the adaptive capacity of endoplasmic reticulum. *Cell Death Differ* 2008; **15**: 1304-1317 [PMID: 18437163 DOI: 10.1038/cdd.2008.52]
- 27 **Merquiol E**, Uzi D, Mueller T, Goldenberg D, Nahmias Y, Xavier RJ, Tirosh B, Shibolet O. HCV causes chronic endoplasmic reticulum stress leading to adaptation and interference with the unfolded protein response. *PLoS One* 2011; **6**: e24660 [PMID: 21949742 DOI: 10.1371/journal.pone.0024660]
- 28 **Back SH**, Lee K, Vink E, Kaufman RJ. Cytoplasmic IRE1 α -mediated XBP1 mRNA splicing in the absence of nuclear processing and endoplasmic reticulum stress. *J Biol Chem* 2006; **281**: 18691-18706 [PMID: 16644724 DOI: 10.1074/jbc.M602030200]
- 29 **Lin JH**, Li H, Yasumura D, Cohen HR, Zhang C, Panning B, Shokat KM, Lavail MM, Walter P. IRE1 signaling affects cell fate during the unfolded protein response. *Science* 2007; **318**: 944-949 [PMID: 17991856 DOI: 10.1126/science.1146361]
- 30 **Yuan L**, Cao Y, Knöchel W. Endoplasmic reticulum stress induced by tunicamycin disables germ layer formation in *Xenopus laevis* embryos. *Dev Dyn* 2007; **236**: 2844-2851 [PMID: 17849439 DOI: 10.1002/dvdy.21299]
- 31 **Iwawaki T**, Akai R, Yamanaka S, Kohno K. Function of IRE1 α in the placenta is essential for placental development and embryonic viability. *Proc Natl Acad Sci USA* 2009; **106**: 16657-16662 [PMID: 19805353 DOI: 10.1073/pnas.0903775106]
- 32 **Back SH**, Schröder M, Lee K, Zhang K, Kaufman RJ. ER stress signaling by regulated splicing: IRE1/HAC1/XBP1. *Methods* 2005; **35**: 395-416 [PMID: 15804613 DOI: 10.1016/j.jymeth.2005.03.001]
- 33 **Böttcher RT**, Niehrs C. Fibroblast growth factor signaling during early vertebrate development. *Endocr Rev* 2005; **26**: 63-77 [PMID: 15689573 DOI: 10.1210/er.2003-0040]
- 34 **Clevers H**. Wnt/ β -catenin signaling in development and disease. *Cell* 2006; **127**: 469-480 [PMID: 17081971 DOI: 10.1016/j.cell.2006.10.018]
- 35 **Kimelman D**. Mesoderm induction: from caps to chips. *Nat Rev Genet* 2006; **7**: 360-372 [PMID: 16619051 DOI: 10.1038/nrg1837]
- 36 **Kaufman RJ**. Orchestrating the unfolded protein response in health and disease. *J Clin Invest* 2002; **110**: 1389-1398 [PMID: 12438434 DOI: 10.1172/JCI0216886]

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Partially hydrolyzed guar gum in pediatric functional abdominal pain

Claudio Romano, Donatella Comito, Annalisa Famiani, Sabrina Calamarà, Italia Loddo

Claudio Romano, Donatella Comito, Pediatric Department, University of Messina, Messina 98100, Italy

Annalisa Famiani, Sabrina Calamarà, Italia Loddo, Pediatric Department, University of Messina, Messina 98100, Italy

Author contributions: Romano C and Comito D conducted the trial and recruited the cases; Famiani A, Calamarà S and Loddo I were involved in the elaboration of the data.

Correspondence to: Claudio Romano, Medical Doctor, PhD, Chief of Endoscopy and Gastroenterology Unit, Pediatric Department, University of Messina, Messina 98100, Italy. romanoc@unime.it

Telephone: +39-90-2212918 Fax: +39-90-2217005

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Abstract

AIM: To assess the effects of partially hydrolyzed guar gum (PHGG) diet supplement in pediatric chronic abdominal pain (CAP) and irritable bowel syndrome (IBS).

METHODS: A randomized, double-blind pilot study was performed in sixty children (8-16 years) with functional bowel disorders, such as CAP or IBS, diagnosed according to Rome III criteria. All patients underwent ultrasound, blood and stool examinations to rule out any organic disease. Patients were allocated to receive PHGG at dosage of 5 g/d ($n = 30$) or placebo (fruit-juice $n = 30$) for 4 wk. The evaluation of the efficacy of fiber supplement included IBS symptom severity score (Birmingham IBS Questionnaire), severity of abdominal pain (Wong-Baker Face Pain Rating Score) and bowel habit (Bristol Stool Scale). Symptom scores were completed at 2, 4, and 8 wk. The change from baseline in the symptom severity scale at the end of treatment and at 4 wk follow-up after treatment was the primary endpoint. The secondary endpoint was to evaluate compliance to supplementation with the PHGG in the

pediatric population. Differences within groups during the treatment period and follow-up were evaluated by the Wilcoxon signed-rank test.

RESULTS: The results of the study were assessed considering some variables, such as frequency and intensity of symptoms with modifications of the bowel habit. Both groups were balanced for baseline characteristics and all patients completed the study. Group A (PHGG group) presented a higher level of efficacy compared to group B (control group), (43% *vs* 5%, $P = 0.025$) in reducing clinical symptoms with modification of Birmingham IBS score (median 0 ± 1 *vs* 4 ± 1 , $P = 0.025$), in intensity of CAP assessed with the Wong-Baker Face Pain Rating Score and in normalization of bowel habit evaluated with the Bristol Stool Scale (40% *vs* 13.3%, $P = 0.025$). In IBS subgroups, statistical analysis shown a tendency toward normalization of bowel movements, but there was no difference in the prevalence of improvement in two bowel habit subsets. PHGG was therefore better tolerated without any adverse effects.

CONCLUSION: Although the cause of pediatric functional gastrointestinal disorders is not known, the results show that complementary therapy with PHGG may have beneficial effects on symptom control.

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Key words: Functional bowel disorders; Partially hydrolyzed guar gum; Pediatric chronic abdominal pain; Fiber diet

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 URL: <http://dx.doi.org/10.3748/wjg.v19.i2.235>

INTRODUCTION

Functional gastrointestinal disorders (FGIDs) are defined as a variable combination of chronic or recurrent gastrointestinal (GI) symptoms; they are age dependent and not explained by structural or biochemical abnormalities^[1]. Chronic abdominal pain (CAP) is the most common condition in FGIDs. It is usually functional without objective evidence of an underlying organic disorder. Apley *et al.*^[2] introduced the term recurrent abdominal pain (RAP) for the first time in pediatric literature, using it to describe a condition whereby children have experienced at least 3 bouts of pain, severe enough to affect activities, over a period of at least 3 mo. In 1999, the Pediatric Rome Working Group introduced standardized symptom-based criteria for pediatric FGIDs with the publication of the Rome II criteria^[3]. In 2006, the "Rome Committee"^[4] defined new diagnostic criteria (Rome III) for pediatric FGIDs, differentiating functional abdominal pain (FAP) from dyspepsia and irritable bowel syndrome (IBS) in that the pain is at a different site with normal bowel habits. The exact prevalence of CAP in children is not known. It seems to account for 2%-4% of all pediatric office visits^[5]. Several studies suggested that 13% of middle-school students and 17% of high school students have weekly experience of abdominal pain^[6,7]. According to these criteria, IBS is an FGID characterized by abdominal pain or discomfort, accompanied by altered bowel habits (constipation, IBS-c or diarrhea, IBS-d or alternating)^[8]. It has been shown that some factors, especially psychological factors, dietary habits, and frequency of exercise, are associated with onset and course of IBS^[9-11]. The term "functional" is not real but conceptual and the pathogenesis can be correlated with alterations of visceral sensitivity, increased intestinal permeability, chronic inflammation and presence of a genetic predisposition^[12]. Pharmacotherapy generally cannot be recommended for children with FAP, except in the context of clinical trials. Drugs should only be given in exceptional cases^[13,14]. Up to 40% of children undergo alternative or complementary therapy such as reassurance, phytotherapy, dietary restrictions or homeopathy^[15]. The putative benefit of such methods has not been documented by controlled clinical trials.

The beneficial effects of water-soluble dietary fibers have received attention as complementary therapy in FGIDs, especially in FAP and IBS, for their ability to modify bowel pattern, accelerate oral-to-anal transit and decrease intracolonic pressure and alleviate pain^[16,17]. Partially hydrolyzed guar gum (PHGG) is a vegetal, water-soluble, non-viscous, non-gelling dietary fiber that is derived from guar gum, a water-soluble, viscous, gelling polysaccharide found in the seeds of the guar plant. The saccharide component of guar gum is galactomannan^[18]. Parisi *et al.*^[19] showed, in an adult open trial, that PHGG supplementation is followed by a decrease of IBS symptoms, such as abdominal pain and bowel habit. Feldman *et al.*^[20], in a small, prospective, randomized,

double-blind, controlled trial, have revealed that fiber supplementation can improve symptoms in children with FAP. Despite this, there have been no recently available published randomized controlled trials (RCTs) to support the use of fiber in the treatment of CAP in a pediatric population.

The aim of this study was to assess the effect of PHGG diet supplement on CAP and IBS symptoms in paediatric population.

MATERIALS AND METHODS

Sixty patients were prospectively enrolled in the study and randomly assigned to one of 2 study arms (PHGG group or group A: 30 patients; placebo group or group B: 30 patients). Median age was 12.8 years (range 8-16 years) with a greater predominance of females (62% girls and 38% boys). CAP and IBS patients were defined according to the Rome III criteria.

All patients were identified into two subgroups: 21/30 (70%) and 19/30 (63%) with constipation-predominant IBS in group A and B respectively; 9/30 (30%) and 11/30 (37%) with diarrhoea-predominant IBS in group A and B respectively. At baseline, the two groups were not statistically different, with respect to age, sex, alterations in bowel movements, incidence and intensity of self-reported symptoms. Subjects' overall baseline demographic and clinical characteristics are summarized in Table 1. All patients underwent ultrasound, blood and stool examinations to exclude organic disease. Seven days before joining the study, patients were asked to not use any medication. The study was performed in accordance with the Declaration of Helsinki and was approved by the local ethics committee. Informed consent was obtained for each patient. All patients completed the trial without any dropouts. Figure 1 is a flow diagram showing the subjects' progression through the study. Patients were consecutively recruited from November 2010 to May 2011, in the Pediatric Gastroenterology Unit of the University of Messina, and randomly assigned to two groups (1:1, PHGG group or group A: 30 patients; placebo group or group B: 30 patients) to receive either a beverage of PHGG (Benefibra, Novartis Consumer Health) at a dosage of 5 g/d in 50 mL of fruit-juice ($n = 30$) or matching placebo (fruit-juice, $n = 30$) for 4 wk. For technical reasons of non-laboratory reproducibility of an inert and odorless powder, the placebo consisted of a fruit juice. As in other studies, PHGG was mixed with fruit juice during meals or between meals.

The manufacturer had no role in the conception, design or conduct of the study or in the analysis or interpretation of the data. Randomization was based on a computer-generated list. Supplementation was stopped after 4 wk and patients were followed up for a further 4 wk. GI symptoms were assessed with the "Birmingham IBS Symptom Questionnaire", "Wong Baker Faces Pain Rating Score" and "Bristol Stool Scale". The Birmingham IBS symptoms score consists of 11 questions based

Table 1 Baseline demographic and clinical data

	PHGG group A	Placebo group B	P value
n	30	30	
Age (yr)	12.3 ± 2.0	13.1 ± 1.5	0.16
Sex (male/female)	12/18	11/19	0.88
Self-reported pain	3.7 ± 1.2	3.5 ± 1.5	0.15
c-IBS	21/30 (70%)	19/30 (63%)	0.75
d-IBS	9/30 (30%)	11/30 (37%)	0.64

Values are mean ± SD or n (%). PHGG: Partially hydrolyzed guar gum; IBS: Irritable bowel syndrome.

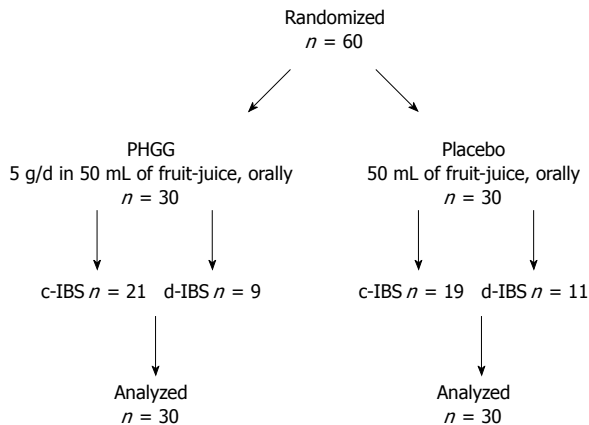


Figure 1 Flow diagram of the progress through the study. PHGG: Partially hydrolyzed guar gum; IBS: Irritable bowel syndrome.

on the frequency of IBS related symptoms. Each question had a standard response scale with symptoms all being measured on a 6-point Likert scale ranging from 0 = none of the time, to 5 = all of the time (Table 2). Wong-Baker Faces Pain Rating Score was used to evaluate CAP severity with a variable score from 0 = no hurt, to 5 = hurts worst. The Bristol Stool Scale classifies the form and consistency of stools into 7 categories (from separate hard lump to entirely liquid stools).

All patients were assessed clinically at 2, 4, and 8 wk (T1, T2, T3) by means of a physical examination and the scoring systems from baseline (Table 3). At T2, compliance with treatment (worse, unchanged, better) was also assessed. Adverse events or any use of other drugs were recorded. Primary outcome was reduction in the frequency and intensity of clinical symptoms and correlation with the improvement of character of stool. Secondary outcome was the evaluation of compliance and safety of PHGG in children.

Statistical analysis

Data are given as mean ± SD. Differences between groups were evaluated by the Kruskal-Wallis one-way analysis of variance (ANOVA). Nominal variables were analyzed with Pearson's chi-square test and Fisher's exact test when, in a 2 × 2 table, one cell had an expected frequency ≤ 5. Differences within groups during the treatment period and follow-up were evaluated by the Wilcoxon signed-rank test. The statistical level of sig-

Table 2 Modified by Birmingham Score Questionnaire^[36]

Constipation	Diarrhoea	Pain
Hard bowel motions	Loose, mushy or watery bowel motions	Discomfort or pain
Straining	Diarrhoea	Discomfort or pain after eating
Constipation	Leaked or soiled Urgency	Sleep problem
	Mucus or slime	

Each question had a standard response scale with symptoms all being measured on a 6-point Likert Scale ranging from 0 = none of the time to 5 = all of the time.

nificance was set at the 5% level ($P < 0.05$). The system utilized was IBM SPSS Statistics Processor.

RESULTS

The results of the study were assessed considering some main variables, such as improving frequency and intensity of the symptoms and modifications of the bowel habit.

Overall rating of frequency symptoms

At the enrolment visit (T0), mean score evaluation of principal IBS related symptoms (Birmingham Score) confirmed that symptoms were present almost every day, together with a strong functional disability in both groups. Overall, 15 of the 60 (24%) participants with IBS reported treatment success. Those in the PHGG group were more likely to have treatment success than those in the control group (43% *vs* 5%, $P = 0.025$). Responders with significant reduction of Birmingham IBS score (median 0 ± 1 *vs* 4 ± 1, $P = 0.025$) was shown in the PHGG group *vs* placebo at both 4 wk and 8 wk (Table 3). The total score and the three subscale scores for constipation, diarrhoea and pain symptoms of the Birmingham score were significantly improved at the 4 and 8 wk evaluations compared to the baseline in the PHGG group. The supplementation response was comparable both in IBS-d and IBS-c subgroups. In group B, no significant difference was found in comparisons at any evaluation time point for any subscale score.

Bowel habits

At baseline all patients were shown a wide range of alterations in bowel movements, evaluated with Bristol Stool Scale, without any difference in the two treatment groups. Effects of PHGG supplementation (5 g/d) for 4 wk on fecal output in IBS-d and IBS-c subsets *vs* placebo were also evaluated. In group A, there was a tendency toward normalization of bowel movements, which is highlighted by the progressive normalization of Bristol Stool Scale at type 3 or 4 (Table 3). In particular, 16 (26.6%) of 60 patients had normalized bowel habits: in the PHGG group the prevalence of improvement was 40% (12 patients), while it was 13.3% (4 patients) in the placebo group ($P = 0.025$). This result remained constant during the follow-up 4 wk. There was no difference in the prevalence of

Table 3 Outcome measures at baseline (T0), at 4 wk of supplementation (T2) and after 8 wk (mean \pm SD)

	Group A PHGG 5g/d (n = 30)			Group B Placebo (n = 30)		
	Baseline	4 wk (T2)	8 wk (T3)	Baseline	4 wk (T2)	8 wk (T3)
Birmingham Score ^a	28.5 \pm 7.16	24.3 \pm 6.02	23.0 \pm 6.15	29.5 \pm 6.94	28.4 \pm 8.39	28.7 \pm 7.54
Bristol Stool Score ^a						
IBS-c	1.00 \pm 1.02	2.02 \pm 1.50	2.32 \pm 1.50	1.16 \pm 0.89	1.76 \pm 1.04	1.65 \pm 1.08
IBS-d	5.02 \pm 0.63	4.01 \pm 0.16	4.07 \pm 0.12	5.54 \pm 0.32	4.86 \pm 0.96	4.89 \pm 0.73
Wong-Baker Score ^a						
	2.15 \pm 0.14	1.86 \pm 0.14	1.63 \pm 0.16	2.16 \pm 0.17	2.04 \pm 0.17	2.05 \pm 0.19

^aP < 0.05 vs placebo (Wilcoxon 2-sample test). PHGG: Partially hydrolyzed guar gum; IBS: Irritable bowel syndrome.

improvement in two bowel habit subsets ($P > 0.05$).

Intensity of the abdominal pain

There was no difference in pain intensity reported at baseline between the groups as Wong-Baker Face Pain Rating Score. During the course of study, there was a decrease in the intensity of pain in the group of children given PHGG, which was not seen in the placebo-supplemented group (Table 3). However, this result was not statistically significant ($P > 0.05$), compared with baseline at wk 4 and 8. Improvement of clinical symptoms in group A was correlated with a change of bowel habit and persisted 4 wk (T8) after cessation of PHGG supplementation. The clinical response was comparable both in IBS-d and IBS-c subgroups (Table 3). Analysis of the data confirmed optimal compliance and safety of PHGG dietary supplementation.

DISCUSSION

CAP is common in children and adolescents. In most children, CAP is functional without objective evidence of an underlying organic disorder. Children with CAP are more likely than children without CAP to have headache, joint pain, anorexia, vomiting, nausea and altered bowel habit assignable to IBS^[21]. Physicians must decide whether to order diagnostic tests or use conservative management. The presence of alarm symptoms or signs suggests higher pretest probability and prevalence of organic disease and may justify the performance of diagnostic tests. CAP can cause long absences from school and markedly worsens quality of life of the children and parents^[22,23].

In a recent American study, the diagnostic evaluation of CAP in a tertiary center in United States was found to cost approximately \$6000 per patient^[24]. The first treatment step is an age-appropriate assessment through the reassurance of the child and family on the absence of organic causes, but this does not mean that abdominal pain is not a real problem. Cognitive behavioral therapy, however, is an effective form of alternative treatment^[25,26].

A thorough review of literature, with a focus on RCTs, revealed a paucity of studies examining effectiveness of pharmacologic and dietary interventions. Definitive statements concerning therapeutic efficacy are quite

limited. Huertas-Ceballos *et al.*^[15], in a meta-analysis, failed to reveal any therapeutic benefit from a low-lactose or high-fiber diet for children with CAP. Therapeutic trials in adults with CAP associated with IBS symptoms have revealed a high rate of the placebo-response, confirming that non-pharmacological therapies alone are often adequate for many patients. PHGG is a soluble fiber with important properties, such as non-viscous texture, normal fermentation, non gelling, high hydrophilic potential and no interference with micronutrient absorption^[27]. There is clear evidence that fiber decreases whole gut transit time, accelerates oral-to-anal transit, and decreases intracolonic pressure reducing abdominal pain. Fiber may represent a mainstay in the FAP and IBS therapeutic algorithm. Results of fiber supplementation in the adult population in patients with FAP and IBS has produced contrasting results, and the main reason for the variation is correlated with different types of fiber used^[28,29]. The main distinction between soluble and insoluble fiber is essential as only soluble fiber such as PHGG dissolves in water and is widely metabolized in the large bowel, thus producing short-chain fatty acids, leading to selective stimulation of microbial growth^[30,31]. PHGG may also act as prebiotic, thus modulating intestinal microbiota. Weaver *et al.*^[32], in experimental studies in rats, demonstrated that PHGG administration was accompanied by a rise in butyrate concentrations of colonocytes. Tuohy *et al.*^[33] showed that PHGG supplementation in healthy volunteers caused selective increase in the percentage of *Bifidobacteria* and *Lactobacilli* with beneficial modulation of microbiota that has been reported to ameliorate IBS symptoms, with a decrease in pain and flatulence^[34].

Bijkerk *et al.*^[35] observed in an adult population that, although general fiber supplementation globally alleviates IBS symptoms, the beneficial effect is mainly associated with the use of soluble fiber rather than insoluble fiber. This study demonstrated that soluble fiber is effective in decreasing global IBS symptoms^[36] but was no better than a placebo. Some of these above mentioned studies on the use of fiber in adult populations were biased as they confirmed that the placebo response in IBS patients ranged from 20%-50%. In our study, the placebo response was much lower than expected.

In 2012, a systematic review identified 3 RCTs evalu-

ating fiber supplementation in children with FGIDs. Patients were supplemented with different dietary fiber types for 4-6 wk^[37]. Among these, the Feldman^[20] study, a randomized, double-blind, placebo-controlled trial, is the only study in children with CAP (26 for group) recruited from primary care practices and supplemented with soluble fibers. Improvement of symptoms in treated patients with fiber not was significant *vs* the placebo-group. In patients with IBS symptoms with modification of the bowel habit, water-soluble fibers, such as PHGG, decrease symptoms also with a prebiotic effect, beneficial modification of the intestinal microflora and selective increase of *Lactobacilli* and *Bifidobacteria*^[38]. PHGG was therefore tolerated and preferred by patients, indicating higher success of soluble fiber than bran or insoluble fiber. The present findings confirm the beneficial effects of PHGG at 5 g/d and in the short term (4 wk). Our study can be considered the first prospective, randomized, controlled, single-blind, clinical trial conducted with this particular fiber supplementation (PHGG) in pediatric CAP and IBS. Some limitations should caution against generalizing from the results of this study, such as the classification at baseline of CAP according to severity of symptoms (mild, moderate and severe) and lack of knowledge of dietary habits in patients enrolled. Given the good results obtained for the first time, it is important to confirm these preliminary data on a greater number of patients and also to consider the active role of liquid fiber in improvement of symptoms. The efficacy of this approach has proven how dietary management is more effective than pharmacological therapy in children with CAP and IBS.

In summary, fiber supplementation can be considered an important option in pediatric CAP and IBS. Water-soluble fiber, such as PHGG, is preferable to insoluble fiber. Moreover, initial studies have shown that fiber may act as a prebiotic, thus increasing the therapeutic benefits. Further placebo-controlled studies are needed to evaluate whether PHGG can also be seen as a maintenance therapy of CAP.

COMMENTS

Background

Functional bowel disorders, such as chronic abdominal pain (CAP), are frequent in children and similar to adult irritable bowel syndrome (IBS). Some children with CAP develop substantial disability and limitations in physical and psychosocial functions. There is little evidence of the efficacy of conventional medical treatment while there is a moderate evidence for the efficacy of complementary therapy (diet, fibers, low-lactose intake) in the adult population. Water-soluble fibers, such as oats, barley and gums in psyllium, can be safe in IBS symptoms.

Research frontiers

Water-soluble fibers are known for their ability to modify bowel patterns, accelerate oral-to-anal transit, decrease intracolonic pressure and alleviate pain. Functional abdominal disorders, such as abdominal pain and IBS, are frequent also in pediatric populations and should stimulate the trend to conservative therapy.

Innovations and breakthroughs

To date, there has been a limited number of studies regarding specific optional treatment in CAP and IBS. This is the first study in a pediatric population that showed a clinically significant improvement of the symptoms in pediatric functional gastrointestinal disease with dietary manipulation. The small sample

size and a low placebo effect may indicate a requirement for request powered and well designed randomized controlled trials on the clinical effectiveness and safety of dietary treatment.

Applications

The findings in this study indicate that fiber supplementation can be considered an important therapeutic option in pediatric IBS.

Peer review

It is very well written and the topic is very interesting for the readers. This is an important topic for gastroenterologists, clinicians, surgeons, Critical Care doctors and nutritionists.

REFERENCES

- 1 Clouse RE, Mayer EA, Aziz Q, Drossman DA, Dumitrascu DL, Mönnikes H, Naliboff BD. Functional abdominal pain syndrome. *Gastroenterology* 2006; **130**: 1492-1497 [PMID: 16678562 DOI: 10.1053/j.gastro.2005.11.062.]
- 2 Apley J, Naish N. Recurrent abdominal pains: a field survey of 1,000 school children. *Arch Dis Child* 1958; **33**: 165-170 [PMID: 13534750 DOI: 10.1136/ad.33.168.165]
- 3 Rasquin-Weber A, Hyman PE, Cucchiara S, Fleisher DR, Hyams JS, Milla PJ, Staiano A. Childhood functional gastrointestinal disorders. *Gut* 1999; **45** Suppl 2: II60-II68 [PMID: 10457047 DOI: 10.1136/gut.45.2008.ii60]
- 4 Rasquin A, Di Lorenzo C, Forbes D, Guiraldes E, Hyams JS, Staiano A, Walker LS. Childhood functional gastrointestinal disorders: child/adolescent. *Gastroenterology* 2006; **130**: 1527-1537 [PMID: 16678566]
- 5 Starfield B, Hoekelman RA, McCormick M, Benson P, Mendenhall RC, Moynihan C, Radecki S. Who provides health care to children and adolescents in the United States? *Pediatrics* 1984; **74**: 991-997 [PMID: 6504643]
- 6 Walker LS, Lipani TA, Greene JW, Caines K, Stutts J, Polk DB, Caplan A, Rasquin-Weber A. Recurrent abdominal pain: symptom subtypes based on the Rome II Criteria for pediatric functional gastrointestinal disorders. *J Pediatr Gastroenterol Nutr* 2004; **38**: 187-191 [PMID: 14734882 DOI: 10.1097/00005176-200402000-00016]
- 7 Son YJ, Jun EY, Park JH. Prevalence and risk factors of irritable bowel syndrome in Korean adolescent girls: a school-based study. *Int J Nurs Stud* 2009; **46**: 76-84 [PMID: 18722617 DOI: 10.1016/j.ijnurstu.2008.07.006]
- 8 Drossman DA, Corazzari E, Delvaux M, Spiller R, Talley NJ, Thompson WG. [Appendix B: Rome III diagnostic criteria for functional gastrointestinal disorders.] *Rev Gastroenterol Mex* 2010; **75**: 511-516 [PMID: 21169122]
- 9 Faresjö A, Grodzinsky E, Johansson S, Wallander MA, Timpka T, Akerlind I. Psychosocial factors at work and in every day life are associated with irritable bowel syndrome. *Eur J Epidemiol* 2007; **22**: 473-480 [PMID: 17484023 DOI: 10.1007/s10654-007-9133-2]
- 10 Saito YA, Locke GR, Weaver AL, Zinsmeister AR, Talley NJ. Diet and functional gastrointestinal disorders: a population-based case-control study. *Am J Gastroenterol* 2005; **100**: 2743-2748 [PMID: 16393229 DOI: 10.1111/j.1572-0241.2005.00288.x]
- 11 Kim YJ, Ban DJ. Prevalence of irritable bowel syndrome, influence of lifestyle factors and bowel habits in Korean college students. *Int J Nurs Stud* 2005; **42**: 247-254 [PMID: 15708012 DOI: 10.1016/j.ijnurstu.2004.06.015]
- 12 Faure C, Wieckowska A. Somatic referral of visceral sensations and rectal sensory threshold for pain in children with functional gastrointestinal disorders. *J Pediatr* 2007; **150**: 66-71 [PMID: 17188617 DOI: 10.1016/j.jpeds.2006.08.072]
- 13 Youssef NN, Di Lorenzo C. The role of motility in functional abdominal disorders in children. *Pediatr Ann* 2001; **30**: 24-30 [PMID: 11195731]
- 14 Ford AC, Talley NJ, Spiegel BM, Foxx-Orenstein AE, Schiller L, Quigley EM, Moayyedi P. Effect of fibre, antispasmodics, and peppermint oil in the treatment of irritable bowel syn-

- drome: systematic review and meta-analysis. *BMJ* 2008; **337**: a2313 [PMID: 19008265 DOI: 10.1136/bmj.a2313]
- 15 **Huertas-Ceballos A**, Logan S, Bennett C, Macarthur C. Pharmacological interventions for recurrent abdominal pain (RAP) and irritable bowel syndrome (IBS) in childhood. *Cochrane Database Syst Rev* 2008; (1): CD003017 [PMID: 18254013]
 - 16 **Connell AM**. The effects of dietary fiber on gastrointestinal motor function. *Am J Clin Nutr* 1978; **31**: S152-S156 [PMID: 101074]
 - 17 **Heizer WD**, Southern S, McGovern S. The role of diet in symptoms of irritable bowel syndrome in adults: a narrative review. *J Am Diet Assoc* 2009; **109**: 1204-1214 [PMID: 19559137 DOI: 10.1016/j.jada.2009.04.012]
 - 18 **Zuckerman MJ**. The role of fiber in the treatment of irritable bowel syndrome: therapeutic recommendations. *J Clin Gastroenterol* 2006; **40**: 104-108 [PMID: 16394869 DOI: 10.1097/01.mcg.0000196405.15110.bb]
 - 19 **Parisi G**, Bottona E, Carrara M, Cardin F, Faedo A, Goldin D, Marino M, Pantalena M, Tafner G, Verdianelli G, Zilli M, Leandro G. Treatment effects of partially hydrolyzed guar gum on symptoms and quality of life of patients with irritable bowel syndrome. A multicenter randomized open trial. *Dig Dis Sci* 2005; **50**: 1107-1112 [PMID: 15986863 DOI: 10.1007/s10620-005-2713-7]
 - 20 **Feldman W**, McGrath P, Hodgson C, Ritter H, Shipman RT. The use of dietary fiber in the management of simple, childhood, idiopathic, recurrent, abdominal pain. Results in a prospective, double-blind, randomized, controlled trial. *Am J Dis Child* 1985; **139**: 1216-1218 [PMID: 2998181]
 - 21 **Berger MY**, Gieteling MJ, Benninga MA. Chronic abdominal pain in children. *BMJ* 2007; **334**: 997-1002 [PMID: 17494020 DOI: 10.1136/bmj.39189.465718.BE]
 - 22 **Whitehead WE**, Burnett CK, Cook EW, Taub E. Impact of irritable bowel syndrome on quality of life. *Dig Dis Sci* 1996; **41**: 2248-2253 [PMID: 8943980 DOI: 10.1007/BF02071408]
 - 23 **Chassany O**, Geneve J, Abitbol JL. Specific quality of life questionnaire in irritable bowel syndrome. *Gastroenterology* 1995; **108**: A581 [DOI: 10.1016/0016-5085(95)26636-4]
 - 24 **Di Lorenzo C**, Colletti RB, Lehmann HP, Boyle JT, Gerson WT, Hyams JS, Squires RH, Walker LS, Kanda PT. Chronic Abdominal Pain In Children: a Technical Report of the American Academy of Pediatrics and the North American Society for Pediatric Gastroenterology, Hepatology and Nutrition. *J Pediatr Gastroenterol Nutr* 2005; **40**: 249-261 [PMID: 15735476 DOI: 10.1097/01.MPG.0000154661.39488.AC]
 - 25 **Chiou E**, Nurko S. Management of functional abdominal pain and irritable bowel syndrome in children and adolescents. *Expert Rev Gastroenterol Hepatol* 2010; **4**: 293-304 [PMID: 20528117 DOI: 10.1586/egh.10.28]
 - 26 **Lackner JM**, Mesmer C, Morley S, Dowzer C, Hamilton S. Psychological treatments for irritable bowel syndrome: a systematic review and meta-analysis. *J Consult Clin Psychol* 2004; **72**: 1100-1113 [PMID: 15612856 DOI: 10.1037/0022-006X.72.6.1100]
 - 27 **Bijkerk CJ**, Muris JW, Knottnerus JA, Hoes AW, de Wit NJ. Systematic review: the role of different types of fibre in the treatment of irritable bowel syndrome. *Aliment Pharmacol Ther* 2004; **19**: 245-251 [PMID: 14984370 DOI: 10.1111/j.0269-2813.2004.01862.x]
 - 28 **Giannini EG**, Mansi C, Dulbecco P, Savarino V. Role of partially hydrolyzed guar gum in the treatment of irritable bowel syndrome. *Nutrition* 2006; **22**: 334-342 [PMID: 16413751 DOI: 10.1016/j.nut.2005.10.003]
 - 29 **Giaccari S**, Grasso G, Tronci S, Allegretta L, Sponziello G, Montefusco A, Siciliano IG, Guarisco R, Candiani C, Chiri S. [Partially hydrolyzed guar gum: a fiber as coadjuvant in the irritable colon syndrome]. *Clin Ter* 2001; **152**: 21-25 [PMID: 11382164]
 - 30 **Marteau P**, Flourie B, Cherbut C, Corrèze JL, Pellier P, Seylaz J, Rambaud JC. Digestibility and bulking effect of ispaghula husks in healthy humans. *Gut* 1994; **35**: 1747-1752 [PMID: 7829013]
 - 31 **Parisi GC**, Zilli M, Miani MP, Carrara M, Bottona E, Verdianelli G, Battaglia G, Desideri S, Faedo A, Marzolino C, Tonon A, Ermani M, Leandro G. High-fiber diet supplementation in patients with irritable bowel syndrome (IBS): a multicenter, randomized, open trial comparison between wheat bran diet and partially hydrolyzed guar gum (PHGG). *Dig Dis Sci* 2002; **47**: 1697-1704 [PMID: 12184518]
 - 32 **Weaver GA**, Tangel C, Krause JA, Alpern HD, Jenkins PL, Parfitt MM, Stragand JJ. Dietary guar gum alters colonic microbial fermentation in azoxymethane-treated rats. *J Nutr* 1996; **126**: 1979-1991 [PMID: 8759370]
 - 33 **Tuohy KM**, Kolida S, Lustenberger AM, Gibson GR. The prebiotic effects of biscuits containing partially hydrolysed guar gum and fructo-oligosaccharides--a human volunteer study. *Br J Nutr* 2001; **86**: 341-348 [PMID: 11570986]
 - 34 **Slavin JL**, Greenberg NA. Partially hydrolyzed guar gum: clinical nutrition uses. *Nutrition* 2003; **19**: 549-552 [PMID: 12781858]
 - 35 **Bijkerk CJ**, de Wit NJ, Muris JW, Whorwell PJ, Knottnerus JA, Hoes AW. Soluble or insoluble fibre in irritable bowel syndrome in primary care? Randomised placebo controlled trial. *BMJ* 2009; **339**: b3154 [PMID: 19713235 DOI: 10.1136/bmj.b3154]
 - 36 **Roalfe AK**, Roberts LM, Wilson S. Evaluation of the Birmingham IBS symptom questionnaire. *BMC Gastroenterol* 2008; **8**: 30 [PMID: 18651941 DOI: 10.1186/1471-230X-8-30]
 - 37 **Horvath A**, Dziechciarz P, Szajewska H. Systematic Review of Randomized Controlled Trials: Fiber Supplements for Abdominal Pain-Related Functional Gastrointestinal Disorders in Childhood. *Ann Nutr Metab* 2012; **61**: 95-101 [PMID: 22889919]
 - 38 **Gibson GR**, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *J Nutr* 1995; **125**: 1401-1412 [PMID: 7782892]

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Atorvastatin and rosuvastatin do not prevent thioacetamide induced liver cirrhosis in rats

Haim Shirin, Efrat Sharvit, Hussein Aeed, Dov Gavish, Rafael Bruck

Haim Shirin, Gastroenterology Institute, Assaf Harofeh Medical Center, Zerifin 70300, Israel

Hussein Aeed, Department of Gastroenterology, The E. Wolfson Medical Center, Holon 58100, Israel

Dov Gavish, Department of Internal Medicine "A", The E. Wolfson Medical Center, Holon 58100, Israel

Rafael Bruck, Efrat Sharvit, Department of Gastroenterology, Tel Aviv Sourasky Medical Center, Tel Aviv 64239, Israel

Haim Shirin, Dov Gavish, Rafael Bruck, Sackler School of Medicine Tel-Aviv University, Tel-Aviv 69975, Israel

Author contributions: Aeed H performed the majority of the in vivo experiments; Sharvit E performed the majority of the in vitro experiments; Gavish D was involved in designing and editing the manuscript; Shirin H and Bruck R designed the study and wrote the manuscript.

Correspondence to: Haim Shirin, MD, Gastroenterology Institute, Assaf Harofeh Medical Center, Zerifin 70300, Israel. haimsh@asaf.health.gov.il

Telephone: +972-8-9779722 Fax: +972-8-9542047

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Abstract

AIM: To examine whether the administration of atorvastatin and rosuvastatin would prevent experimentally-induced hepatic cirrhosis in rats.

METHODS: Liver cirrhosis was induced by injections of thioacetamide (TAA). Rats were treated concurrently with TAA alone or TAA and either atorvastatin (1, 10 and 20 mg/kg) or rosuvastatin (1, 2.5, 5, 10 and 20 mg/kg) given daily by nasogastric gavage.

RESULTS: Liver fibrosis and hepatic hydroxyproline content, in the TAA-treated group was significantly higher than those of the controls [11.5 ± 3.2 vs 2.6 ± 0.6 mg/g protein ($P = 0.02$)]. There were no differences in serum aminotransferase levels in the TAA controls compared to all the groups treated concomitantly by

statins. Both statins used in our study did not prevent liver fibrosis or reduce portal hypertension, and had no effect on hepatic oxidative stress. Accordingly, the hepatic level of malondialdehyde was not lower in those groups treated by TAA + statins compared to TAA only. *In vitro* studies, using the BrdU method have shown that atorvastatin had no effect of hepatic stellate cells proliferation. Nevertheless, statin treatment was not associated with worsening of liver damage, portal hypertension or survival rate.

CONCLUSION: Atorvastatin or rosuvastatin did not inhibit TAA-induced liver cirrhosis or oxidative stress in rats. Whether statins may have therapeutic applications in hepatic fibrosis due to other etiologies deserve further investigation.

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Key words: Liver cirrhosis; Statins; Thioacetamide

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INTRODUCTION

Liver cirrhosis is one of the leading causes of morbidity and mortality worldwide. Hepatic stellate cells (HSC) play a major role in the pathogenesis of hepatic fibrosis^[1]. Injury to hepatocytes results in generation of lipid peroxides, which may have a direct stimulatory effect on matrix production by activated HSC^[2]. It has been suggested that aldehyde-protein adducts, including products of lipid peroxidation, modulate collagen gene expression in human fibroblasts^[3,4] and may be a link between tissue injury and hepatic fibrosis^[2,5]. Several

studies demonstrated that activation of HSC in culture is provoked by generation of free radicals and is blocked by anti-oxidants. This activation may involve the transcription factor c-myc and nuclear factor kappa B (NF- κ B)^[6,7]. Accordingly, antioxidants have been suggested as therapeutic modalities in experimental models^[8-10], and in patients with chronic liver injury^[11].

3-Hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors (statins) are used extensively to reduce serum cholesterol in an effort to reduce atherosclerotic cardiovascular morbidity and mortality. In addition to their cholesterol-lowering effect, statins demonstrate other biological effects (pleiotropic effects) that some of them may lead to clinical benefits. Those include anti-inflammatory^[12] and antioxidant effects^[13], inhibition of PDGF-stimulated proliferation and upregulation of tumor growth factor (TGF)- β signaling in murine mesangial cells and cultured heart cells^[14-16].

Over the past decade the potential effect of statins as anti-fibrotic agents has received increasing attention. The rationale for the anti fibrotic efficacy is based on the ability of statin compounds to: (1) decrease the growth of human Ito cells in vitro, independently of their effect on cholesterol synthesis^[17]; (2) inhibit the proliferation rate of HSC and reduction of the collagen protein steady state levels by both simvastatin and lovastatin^[18]; (3) inhibit steatosis, hepatic fibrosis and carcinogenesis in a rat model of non-alcoholic steatohepatitis (NASH)^[19]; and (4) enhance hepatic nitric oxide production and decrease hepatic vascular resistance in patients with cirrhosis and portal hypertension^[20]. In addition to inhibiting stellate cell activation, the anti-oxidative activity and the attenuation of inflammation by specific statin derivatives may also contribute to the inhibition of fibrosis. Despite the existing data suggesting that statin derivatives can inhibit fibrosis by various mechanisms, treatment with simvastatin or pravastatin did not decrease fibrosis-induced by bile duct ligation (BDL) or carbon tetrachloride (CCl₄) in rats^[21,22]. However, one recent study showed that very early atorvastatin treatment inhibits HSC activation and fibrosis in the BDL model *in vivo*^[23].

To further elucidate the anti-fibrotic activity of statins in the liver, we examined the effects of both atorvastatin and rosuvastatin in a well characterized model of chronic thioacetamide (TAA)-induced administration in which cirrhosis is mainly produced *via* the formation of reactive oxygen species. TAA undergoes an extensive metabolism to acetamide shortly after administration, and to the hepatotoxic reactive metabolite thioacetamide-S-oxide by the mixed function oxidase system^[24-26]. We hypothesized that inhibition of HSC activity in addition to the anti inflammatory and anti oxidative effects induced by statins may prevent the hepatic damage induced by TAA in rats.

Our results indicate that both atorvastatin and rosuvastatin did not diminish neither oxidative stress nor the development of TAA-induced cirrhosis in rats, and also had no effect on the proliferation of cultured HSC.

MATERIALS AND METHODS

Materials and animals

TAA, atorvastatin and rosuvastatin was purchased from Sigma (Sigma Chemical Co., St. Louis, MO). Male Wistar rats (250-300 g), obtained from Tel-Aviv University Animal Breeding Center, were kept in the animal breeding house of the E. Wolfson Medical Center and fed a Purina chow *ad libitum*. Animals were kept with a 12-h light-dark cycle at constant temperature and humidity and the rats had free access to tap water during the study period. Use of animals was in accordance with the National Institutes of Health Policy on the care and use of laboratory animals and was approved by the Animal Use and Care Committee of the E. Wolfson Medical Center.

Induction of liver cirrhosis

For induction of liver cirrhosis, rats were given intraperitoneal injections of thioacetamide, 200 mg/kg, twice a week for 12 wk, as previously described^[27]. Control rats were treated with intraperitoneal injections of NaCl 0.9%.

Analysis of liver histopathology

The rats were sacrificed at the completion of the treatment protocols, their livers were removed, and midsections of the left lobes of the livers were processed for light microscopy. This processing consisted of fixing the specimens in a 5% neutral formol solution, embedding the specimens in paraffin, making sections of 5 μ m thickness, and staining the sections with hematoxylin and eosin and Masson Trichrome. The tissue slices were scanned and scored blindly by two expert pathologists. The degree of fibrosis was expressed as the mean of 10 different fields in each slide, which had been classified on a scale of 0-4 according to Batts and Ludwig^[28].

Measurement of hepatic hydroxyproline levels

Quantitative determination of hepatic hydroxyproline content was performed as previously described^[29].

Measurement of hepatic malondialdehyde

For the determination of the hepatic content of malondialdehyde, liver tissue (5 g) was cut into small pieces using a razor blade, and homogenized after dilution in H₂O 1:10 w/v. Liver homogenate was centrifuged in 900 g for 5 min, and then the supernatant was collected and centrifuged in 20 000 rpm in Sorvall for 30 min using plastic tubes. The clear supernatant was obtained and malondialdehyde was measured and expressed as nmole/g wet tissue using the thiobarbituric acid method^[30]. Briefly, to 1 mL of 10% liver homogenate with 1.15% KCl were added 2 mL of fresh solution 15% w/v TCA, 0.375% w/v thiobarbituric acid, 0.25 mL/L HCl. The mixture was heated at 95 °C for 15 min. The solution was cooled to room temperature using tap water and centrifuged at 300 rpm for 10 min. Absorption of the pink supernatant was determined spectrophotometrically.

metrically at 532 nm.

Effect of atorvastatin on proliferation of primary hepatic stellate cells

Hepatic stellate cells were isolated from male rat using sequential pronase-collagenase digestion followed by Nycodenz (Sigma-Aldrich, Inc., St. Louis, MO, United States) density gradient centrifugation essentially as described previously with minor modifications^[1]. Briefly, the liver of male rat Wistar (300-400 g) was minced with scalpels and incubated with 100 mL of freshly prepared and filtered-GBSS with 65 mg pronase (Roche Molecular Biochemicals, Indianapolis, IN, United States), 50 mg collagenase (Worthington Biochemical Corporation, Lakewood, NJ, United States), and 0.5 mL 2.7% CaCl₂ for 15 min at 37 °C with 200 rpm shaking. Then 50 mL of freshly prepared and filtered-GBSS was added containing 12.5 mg pronase, 12.5 mg collagenase, 20 µg/mL DNase I (Sigma-Aldrich, Inc., St. Louis, MO, United States) and 0.25 mL 2.7% CaCl₂ for 30 min at 37 °C with 200 rpm shaking. The digested tissue was filtered through a sterile 150-µm metal mesh, and the cells were centrifuged at 2000 rpm for 7 min. The digested liver hepatic stellate cells were isolated on a 17.5% Nycodenz gradient centrifuged at 2700 rpm for 20 min. A stellate cell-enriched fraction was present in the upper layer. Cells were washed twice by centrifugation (1200 rpm, 4 °C, 5 min) in DMEM with 10% fetal calf serum (FCS), 100 µg/mL penicillin and 100 µg/mL of streptomycin.

Reagents

Atorvastatin (Sigma-Aldrich, Inc., St. Louis, MO, United States) stock solution was dissolved in DDW to a concentration of 5×10^{-4} mol/L. PDGF-BB (Peprotech Inc. NJ, United States) was dissolved in DDW to a concentration of 1 µg/mL. All the reagents were aliquot and stored in -20 °C until use.

Proliferation assays

The proliferation of HSC was examined by BrdU method (Exalpha biological, Inc. Watertown, MA, United States). Primary HSC were cultured for 14 d, after which they were trypsinized and plated at a density of 20 000 cell/well in 96 well plates in DMEM containing 10% FCS. The cells were incubated for 24 h, and then they were serum starved in DMEM + 0.5% FCS overnight. At the following day, the various stimuli were added in medium containing 0.5% FCS. HSC were exposed to either 30 ng/mL of PDGF (Peprotech, Inc. NJ, United States), and different concentration of Atorvastatin (5×10^{-8} mol/L, 10^{-7} mol/L, 5×10^{-7} mol/L) alone, or combination of the two. After 24 h the cells were tested for proliferation according to the manufacture instruction.

Western blotting

HSCs were plated on 100 mm plates at a density of 2×10^6 cells/plate. After 24 h, the medium was changed

to starvation medium (DMEM + 0.5% FCS) overnight. The following day, cells were incubated for 24 h with the different treatments according to which experiments were performed. Total proteins were extracted by incubating the cells for 30 min on ice in RIPA buffer containing a 1:100 dilution of a protease inhibitor cocktail (Sigma-Aldrich, Inc., St. Louis, MO, United States). After 20 min centrifugation at 14 000 rpm at 4 °C, extracts were normalized to total protein content, determined using the BCA Reagent (Sigma-Aldrich, Inc., St. Louis, MO, United States). Equal amounts of total protein were separated in 4%-12% bis-tris (BT) gels (NuPAGE, Gibco-BRL Life Technologies, Grand Island, NY), blotted onto Hybond C extra membranes, blocked overnight in 5% milk, and incubated with antibodies against α smooth muscle actin (α SMA), glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (Santa Cruz Biotechnology, Santa Cruz, CA, United States) and then incubated with horseradish peroxidase-conjugated secondary antibody. Afterwards, signals were detected by chemiluminescent. Expression of proteins was normalized to the expression of GAPDH.

Experimental design (in vivo experiments)

Control groups: (1) normal controls: injections of NaCl 0.9%; and (2) cirrhotic controls: TAA (200 mg/kg *ip*, twice weekly) injections for 12 wk.

Atorvastatin groups: (1) TAA + Atorvastatin 1 mg/kg daily; (2) TAA + Atorvastatin 10 mg/kg daily; and (3) TAA + Atorvastatin 20 mg/kg daily.

Rosuvastatin groups: (1) TAA + Rosuvastatin 2.5 mg/kg daily; (2) TAA + Rosuvastatin 5 mg/kg daily; (3) TAA + Rosuvastatin 10 mg/kg daily; and (4) TAA + Rosuvastatin 20 mg/kg daily.

Each subgroup included 6-7 rats. The statins were given *po*, started concurrently with TAA and continued during the study. When the treatments were completed after 12 wk the rats were sacrificed, their livers were removed and the weights of their spleens measured.

Statistical analysis

The data are presented as median (range). The significance of differences among different groups was determined by a student's *t*-test.

RESULTS

Induction of liver cirrhosis by thioacetamide

Intraperitoneal administration of TAA for 12 wk, resulted in a uniform coarse granulation of the surface of the rats' liver. Microscopic analysis revealed liver cirrhosis, characterized by mixed-sized fibrotic nodules in these TAA-treated rats. Neither atorvastatin nor rosuvastatin had any effect on liver enzymes when given alone or in addition to TAA.

Table 1 Effects of rosuvastatin and atorvastatin treatment on oxidative stress and liver fibrosis in thioacetamide-treated rats

	ALT	MDA nmole/g liver	Hydroxyproline (mg/g protein)	Fibrosis score (0-4)	Spleen weight (mg)
Rosuvastatin dose (mg/kg)					
Control	59 (44, 69)	30.6 (20.5, 31.1)	2.67 (1.9, 3.3)	0	1200 (1100, 1400)
Ros 2.5	52 (47, 58)	24.1 (19.8, 33.9)	2.7 (2.2, 3.5)	0	1020 (950, 1150)
Ros 5	50 (41, 57)	31.3 (26.9, 36.7)	2.8 (2.3, 3.1)	0	1100 (950, 1300)
R 10	56 (46, 64)	33.7 (27.7, 37.8)	2.3 (1.8, 3.7)	0	1000 (800, 1200)
R 20	44 (41, 47)	29.4 (23.9, 39.6)	3.9 (2.4, 4.2)	0	1200 (900, 1200)
TAA	63 (37, 79)	48.7 (38.4, 50.2)	11.2 (4.5, 15.4)	4 (1.5, 4)	1900 (1600, 2200)
TAA + R 2.5	53 (47, 63)	22 (19, 31.2)	7.5 (3.7, 13.1)	2.5 (1, 4)	1450 (1050, 1950)
TAA + R 5	54 (48, 67)	36.5 (24.8, 52.1)	11.0 (4.7, 16.2)	3.25 (1, 4)	1230 (1000, 1350)
TAA + R 10	51 (43, 60)	37.8 (27.5, 44.9)	11.8 (8.3, 16.2)	3 (2, 4)	1200 (1100, 1500)
TAA + R 20	51.5 (48, 55)	39.2 (29.6, 46.8)	12.3 (3.1, 14.3)	3.5 (1, 4)	1050 (900, 1300)
Atorvastatin dose (mg/kg)					
Control	59 (44, 69)	30.6 (20.5, 31.1)	2.67 (1.9, 3.3)	0	1200 (1100, 1400)
Ato 1	61 (55, 81)	32.8 (27.4, 33.6)	2.5 (2.1, 3.1)	0	1300 (1100, 1400)
Ato 10	56 (53, 59)	32.9 (29.1, 35.2)	3.4 (1.6, 3.7)	0	1200 (1100, 1300)
Ato 20	58 (48, 73)	30.1 (29.8, 35.3)	2.3 (1.95, 3)	0	1100 (1100, 1200)
TAA	63 (37, 79)	48.7 (38.4, 50.2)	11.2 (4.5, 15.4)	4 (1.5, 4)	1900 (1600, 2200)
TAA + Ato 1	49 (22, 70)	46.6 (35.8, 63.1)	9.1 (6.7, 16.1)	3 (2, 4)	1600 (900, 3000)
TAA + Ato 10	53 (47, 62)	43.4 (36.2, 53.1)	12.5 (6.9, 15.4)	4 (2, 4)	1500 (1400, 1700)
TAA + Ato 20	57 (43, 67)	48.2 (34.2, 58.5)	10.9 (5.3, 16.8)	4 (3, 4)	1500 (1300, 1700)

$n = 7$, in the groups that received thioacetamide (TAA), $n = 4$ in the control, rosuvastatin only treated groups and atorvastatin only treated groups. TAA, 200 mg/kg *ip* twice weekly for 12 wk. Rosuvastatin and atorvastatin had given daily by nasogastric gavage. Ros: Rosuvastatin; ALT: Alanine transaminase; MDA: Malondialdehyde.

Inhibition of thioacetamide-induced liver cirrhosis by statins

Compared to the rats which received TAA only, neither atorvastatin nor rosuvastatin had protective effect on the histopathologic score after 12 wk and TAA-induced liver cirrhosis was not inhibited by all the doses that were examined. Hepatic fibrosis was also quantitated by the measurement of hepatic hydroxyproline levels. The mean hydroxyproline levels of the TAA-treated group were similar to those of the TAA plus atorvastatin or rosuvastatin at all doses used.

Spleen weights

An indirect measure of portal hypertension was obtained by measuring the weights of the rats' spleens at the end of the experiment. Characteristic hemodynamic changes have previously been shown after 3 mo of TAA administration, i.e., upon TAA-induced liver cirrhosis^[27]. These changes include portal hypertension and hyperdynamic circulation which are accompanied with a significant increase in spleen weight. After 12 wk, the mean spleen weight of rats receiving TAA was about 30% higher than those receiving injections of 0.9% NaCl or statins only. The mean spleen weight of rats that received statins in addition to TAA was not lower than that of TAA alone (Table 1).

Hepatic levels of malondialdehyde and lipid peroxides

The hepatic levels of malondialdehyde measured after 12 wk, were not significantly different in the rats treated with TAA and statins compared to TAA only. Table 1 summarize the rosuvastatin and atorvastatin treatment effects on oxidative stress and liver fibrosis in TAA-

treated rats [median (range)], $n = 6-7$.

Effect of Atorvastatin on hepatic stellate cells proliferation and smooth muscle actin expression

Atorvastatin in different concentrations had no effect on HSC proliferation as examined by the BrdU method (Figure 1) and no effect on the expression of smooth muscle actin determined by western blot analysis (Figure 2).

DISCUSSION

Our major finding in the present study is that atorvastatin and rosuvastatin do not have a therapeutic value as a potential anti-oxidant or anti-fibrotic agents targeting increased oxidative stress or liver fibrosis induced by TAA in rats.

There are various experimental observations regarding the direct anti-fibrotic activity of the statins by the inhibition of stellate cell proliferation; lovastatin inhibits pancreatic stellate cell activation and alpha-smooth muscle actin expression^[31] and both simvastatin and lovastatin interferes with HSC activation *in vitro*^[20,21]. Fluvastatin reduces renal fibroblast proliferation and collagen type III production^[32] and suppresses oxidative stress and kidney fibrosis after ureteral obstruction^[33]. It is also possible that the anti-fibrotic effects of statins are mediated through mechanisms that stimulate fibroblast apoptosis *in vitro* and *in vivo* as it was shown in lung and renal fibroblasts^[32,34]. Alternatively, the anti-fibrotic effects of statins may be mediated through their newly recognized anti inflammatory^[12] and antioxidant mechanisms^[35-37]. These include the inhibition of myeloperoxidase derived and nitric oxide derived oxidants^[36],

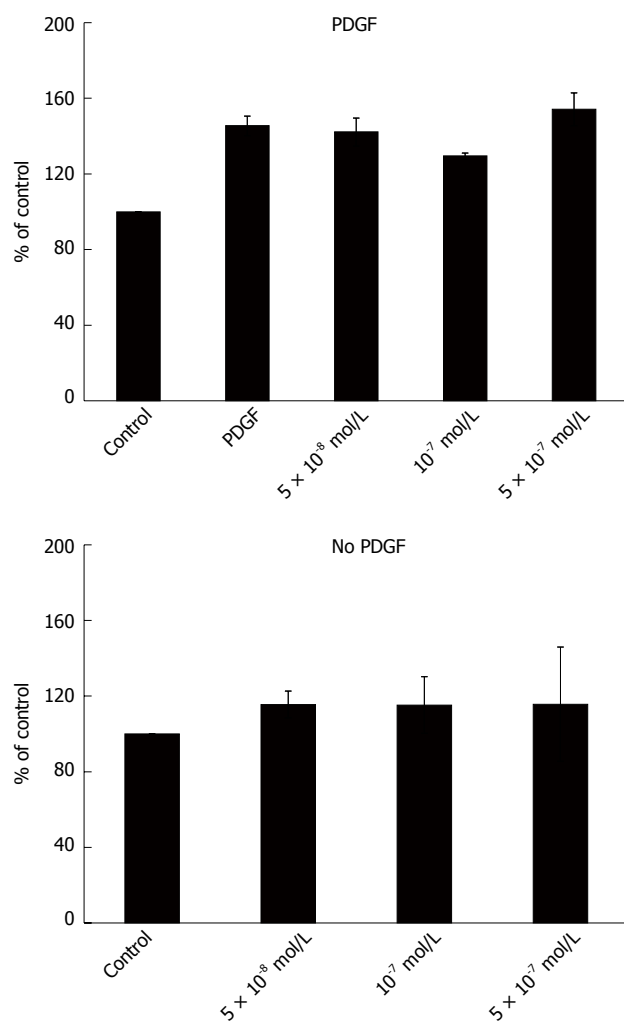


Figure 1 Effect of atorvastatin on hepatic stellate cells proliferation. PDGF: Platelet derived growth factor.

S-nitrosylation and activation of thioredoxin in endothelial cells^[37], decreased expression of essential NAD(P)H oxidase subunits and upregulation of catalase expression in vascular smooth muscle cells^[35]. Statins also induce the expression of a protein with antioxidant and anti-inflammatory functions, heme oxygenase-1 (HO-1), *in vitro* and *in vivo*^[33,38]. However, this effect is tissue specific demonstrating significant increase in liver HO-1 with simvastatin and lovastatin but not atorvastatin and rosuvastatin. The involvement of hydroxyl radicals and oxidative stress in TAA induced cirrhosis^[39], the antioxidative effects of different statins, including atorvastatin, and the antifibrotic effect of atorvastatin in a rat model of BDL as was reported recently by Trebicka *et al.*^[23], provided a good rationale for the assumption that statins may reduce or prevent fibrogenesis in this specific animal model.

The explanation for the unexpected failure of atorvastatin and rosuvastatin to inhibit fibrosis in this model of cirrhosis are not clear. Two studies demonstrated anti fibrogenic effects of atorvastatin in a CCl₄ induced fibrosis^[40,41]. Gardner *et al.*^[41] have shown that atorvas-

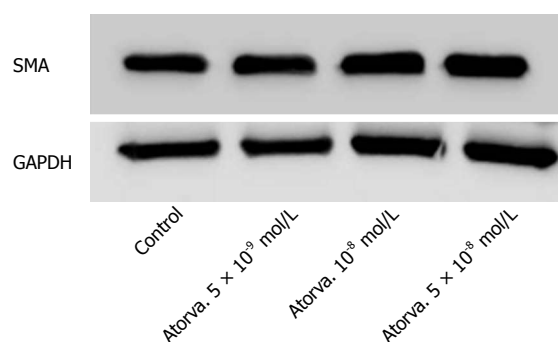


Figure 2 Atorvastatin effect on the expression of smooth muscle actin. GAPDH: Glyceraldehyde-3-phosphate dehydrogenase; SMA: Smooth muscle actin.

tatin exhibited statistically significant, although modest, suppression of CCl₄-induced fibrogenesis after 3 wk of treatment. This was shown only using a novel technique for measuring hepatic collagen synthesis *in vivo* through metabolic labeling with heavy water (²H₂O). Histopathology of the same tissues revealed no significant differences in fibrosis scores among groups that received co-treatment with atorvastatin, emphasizing the importance of the fibrosis measuring method.

Nevertheless the evidence that fibrosis was not inhibited by atorvastatin and with a more potent cholesterol metabolism pathway inhibitor, rosuvastatin, suggests that this effect is not selective and occurs independently of their HMG-CoA inhibition. Since we used much higher doses than those used clinically (up to 20 mg/kg) it does not seem that we are dealing with under dose treatment. Appropriate timing for the administration of the statins may also be critical. Later therapy in the BDL model, for example, lacked significant effects on fibrosis with no change in hepatic inflammation^[23]. The statin treatment in our study began in parallel to the administration of TAA. Although, it is possible that pretreatment by statins would be more effective, the fact that there was no improvement, neither in MDA levels nor in fibrosis after three months, do not support this hypothesis. Another possibility to explain these negative results might be the selection of the wrong statins. Indeed, the two statins that inhibited HSC proliferation *in vitro* were lovastatin and simvastatin^[18] and atorvastatin *in vivo*. In addition pitavastatin was the one that inhibited hepatic fibrosis in a choline-deficient L-amino acid defined diet liver fibrosis^[19]. However, despite these effects, even the latter mechanisms were not efficient to prevent or inhibit liver fibrosis, as it was demonstrated with simvastatin or pravastatin in two different animal models of cirrhosis (bile duct ligation and CCl₄), in rats^[21,22]. Moreover, antioxidant therapy in human clinical trials also lack or have minimal effect in chronic liver disease^[42,43]. Finally, statin-induced protein kinase C (PKC) activation in activated HSCs may interrupt statin-induced HSC apoptosis, thereby reducing its antifibrotic efficacy. Indeed Yang *et al.*^[44] recently demonstrated that simultaneous treatment with pravastatin and PKC inhibitor may

synergistically enhance antifibrotic efficacy in hepatic fibrosis induced by intraperitoneal injection of carbon tetrachloride or thioacetamide in mice. Finally, we also have to consider the possibility that the negative results may be due to the small numbers (type 2 error).

To further explore the effect of statins on liver fibrosis we examined the effect of several atorvastatin concentrations on the proliferation of primary HSC. We observed that atorvastatin had no inhibitory effect on HSC proliferation either in the presence or in the absence of PDGF (Figure 1). This finding is in discordance with several previous studies that demonstrated decreased HSC activation by statins^[17,18]. Moreover, using western blot analysis we found that atorvastatin (5×10^{-8} - 10^{-9} mol/L) had no effect on the expression of α smooth muscle actin further supporting the lack of effect shown in the proliferation assay.

It is of interest that treatment with atorvastatin or rosuvastatin in all doses did not reduce spleen weights, a parameter of portal hypertension. This is in contrast to the recently described therapeutic effects of simvastatin in patients with cirrhosis and portal hypertension. In patients with cirrhosis and portal hypertension simvastatin enhanced hepatic nitric oxide production and decreased hepatic resistance^[20]. Similarly, in cirrhotic rats, induced by BDL, atorvastatin has been shown to lower intrahepatic resistance and to decrease portal hypertension^[45].

Finally, oral atorvastatin and rosuvastatin were both well tolerated with no side effects, toxicity or mortality. Furthermore, transaminase levels, hepatic MDA and hydroxyproline and liver histopathology score did not aggravate. These data suggest that the use of atorvastatin and rosuvastatin is not associated with an increased risk of hepatotoxicity in damaged liver.

In summary, our results show that atorvastatin and rosuvastatin have no effect on TAA-induced liver cirrhosis. Obviously further studies are required to evaluate whether statins may have therapeutic applications, in the development of hepatic fibrosis induced by other etiologies.

COMMENTS

Background

Liver cirrhosis is one of the leading causes of morbidity and mortality worldwide. Hepatic stellate cells (HSC) play a major role in the pathogenesis of hepatic fibrosis. Several studies demonstrated that activation of HSC in culture is provoked by generation of free radicals. Accordingly, antioxidants have been suggested as therapeutic modalities in experimental models, and in patients with chronic liver injury.

Research frontiers

Previous studies mainly focused on the potential effect of statins as anti-fibrotic agents. However, whether the anti-oxidative activity and the attenuation of inflammation by specific statin derivatives may also contribute to the inhibition of stellate cell activation and fibrosis remain unclear. The authors hypothesized that inhibition of HSC activity in addition to the anti inflammatory and anti oxidative effects induced by statins may prevent the hepatic damage induced by TAA in rats.

Innovations and breakthroughs

To further elucidate the anti-fibrotic activity of statins in the liver, the authors examined the effects of both atorvastatin and rosuvastatin in thioacetamide (TAA)-

induced liver fibrosis in which cirrhosis is mainly produced via the formation of reactive oxygen species. The major finding was that both statins do not have a therapeutic value as a potential anti-oxidant or anti-fibrotic agents targeting increased oxidative stress or liver fibrosis induced by TAA.

Applications

The results show that atorvastatin and rosuvastatin have no effect on cirrhosis induced mainly by oxidative stress. Further studies are required to evaluate clarify the mechanism by which statins may have therapeutic applications, in hepatic fibrosis induced by other etiologies.

Terminology

3-Hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors (statins) are used extensively to reduce serum cholesterol in an effort to reduce atherosclerotic cardiovascular morbidity and mortality. In addition to their cholesterol-lowering effect, statins demonstrate also anti-inflammatory and antioxidant effects. TAA-induced cirrhosis is mainly produced via the formation of reactive oxygen species. TAA undergoes an extensive metabolism to acetamide shortly after administration, and to the hepatotoxic reactive metabolite thioacetamide-S-oxide by the mixed function oxidase system.

Peer review

Well written manuscript which presented that atorvastatin or rosuvastatin did not inhibit the formation of TAA-induced liver cirrhosis in rats. Further, no induction of oxidative stress or effect on HSC proliferation have been noted. The model is well presented, the experiments seems to be correct. Even if results are negative, and in partial or total contrast with previous studies, the study appears well conducted and the results discussed with honesty, and caution.

REFERENCES

- 1 **Tsukada S**, Parsons CJ, Rippe RA. Mechanisms of liver fibrosis. *Clin Chim Acta* 2006; **364**: 33-60 [PMID: 16139830 DOI: 10.1016/j.cca.2005.06.014]
- 2 **Bedossa P**, Houghlum K, Trautwein C, Holstege A, Chojkier M. Stimulation of collagen alpha 1(I) gene expression is associated with lipid peroxidation in hepatocellular injury: a link to tissue fibrosis? *Hepatology* 1994; **19**: 1262-1271 [PMID: 8175151 DOI: 10.1016/0270-9139(94)90876-1]
- 3 **Brenner DA**, Chojkier M. Acetaldehyde increases collagen gene transcription in cultured human fibroblasts. *J Biol Chem* 1987; **262**: 17690-17695 [PMID: 3693368]
- 4 **Houghlum K**, Brenner DA, Chojkier M. d-alpha-tocopherol inhibits collagen alpha 1(I) gene expression in cultured human fibroblasts. Modulation of constitutive collagen gene expression by lipid peroxidation. *J Clin Invest* 1991; **87**: 2230-2235 [PMID: 2040703 DOI: 10.1172/JCI115258]
- 5 **Houghlum K**, Bedossa P, Chojkier M. TGF-beta and collagen-alpha 1 (I) gene expression are increased in hepatic acinar zone 1 of rats with iron overload. *Am J Physiol* 1994; **267**: G908-G913 [PMID: 7977754]
- 6 **Meyer M**, Caselmann WH, Schlüter V, Schreck R, Hofschneider PH, Baeuerle PA. Hepatitis B virus transactivator MHBst: activation of NF-kappa B, selective inhibition by antioxidants and integral membrane localization. *EMBO J* 1992; **11**: 2991-3001 [PMID: 1639069]
- 7 **Lee KS**, Buck M, Houghlum K, Chojkier M. Activation of hepatic stellate cells by TGF alpha and collagen type I is mediated by oxidative stress through c-myc expression. *J Clin Invest* 1995; **96**: 2461-2468 [PMID: 7593635 DOI: http://]
- 8 **Liu SL**, Degli Esposti S, Yao T, Diehl AM, Zern MA. Vitamin E therapy of acute CCl4-induced hepatic injury in mice is associated with inhibition of nuclear factor kappa B binding. *Hepatology* 1995; **22**: 1474-1481 [PMID: 7590666 DOI: 10.1002/hep.1840220522]
- 9 **Brown KE**, Poulos JE, Li L, Soweid AM, Ramm GA, O'Neill R, Britton RS, Bacon BR. Effect of vitamin E supplementation on hepatic fibrogenesis in chronic dietary iron overload. *Am J Physiol* 1997; **272**: G116-G123 [PMID: 9038884]
- 10 **Boigk G**, Stroedter L, Herbst H, Waldschmidt J, Riecken EO, Schuppan D. Silymarin retards collagen accumulation

- in early and advanced biliary fibrosis secondary to complete bile duct obliteration in rats. *Hepatology* 1997; **26**: 643-649 [PMID: 9303494 DOI: 10.1002/hep.510260316]
- 11 **Houglum K**, Venkataramani A, Lyche K, Chojkier M. A pilot study of the effects of d-alpha-tocopherol on hepatic stellate cell activation in chronic hepatitis C. *Gastroenterology* 1997; **113**: 1069-1073 [PMID: 9322499 DOI: 10.1053/gast.1997.v113.pm9322499]
 - 12 **Crisby M**. Modulation of the inflammatory process by statins. *Timely Top Med Cardiovasc Dis* 2005; **9**: E3 [PMID: 15824762]
 - 13 **Takemoto M**, Liao JK. Pleiotropic effects of 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Arterioscler Thromb Vasc Biol* 2001; **21**: 1712-1719 [PMID: 11701455 DOI: 10.1161/hq1101.098486]
 - 14 **Yoshimura A**, Nemoto T, Sugeno Y, Inui K, Watanabe S, Inoue Y, Sharif S, Yokota N, Uda S, Morita H, Ideura T. Effect of simvastatin on proliferative nephritis and cell-cycle protein expression. *Kidney Int Suppl* 1999; **71**: S84-S87 [PMID: 10412745 DOI: 10.1046/j.1523-1755.1999.07121.x]
 - 15 **Park HJ**, Galper JB. 3-Hydroxy-3-methylglutaryl CoA reductase inhibitors up-regulate transforming growth factor-beta signaling in cultured heart cells via inhibition of geranylgeranylation of RhoA GTPase. *Proc Natl Acad Sci U S A* 1999; **96**: 11525-11530 [PMID: 10500210 DOI: 10.1073/pnas.96.20.11525]
 - 16 **Nogaki F**, Muso E, Yashiro M, Kasuno K, Kamata T, Ono T, Sasayama S. Direct inhibitory effects of simvastatin on matrix accumulation in cultured murine mesangial cells. *Kidney Int Suppl* 1999; **71**: S198-S201 [PMID: 10412775 DOI: 10.1046/j.1523-1755.1999.07151.x]
 - 17 **Mallat A**, Preaux AM, Blazejewski S, Dhumeaux D, Rosenbaum J, Mavrier P. Effect of simvastatin, an inhibitor of hydroxy-methylglutaryl coenzyme A reductase, on the growth of human Ito cells. *Hepatology* 1994; **20**: 1589-1594 [PMID: 7982659 DOI: 10.1002/hep.1840200631]
 - 18 **Rombouts K**, Kisanga E, Hellemans K, Wielant A, Schuppan D, Geerts A. Effect of HMG-CoA reductase inhibitors on proliferation and protein synthesis by rat hepatic stellate cells. *J Hepatol* 2003; **38**: 564-572 [PMID: 12713866 DOI: 10.1016/S0168-8278(03)00051-5]
 - 19 **Miyaki T**, Nojiri S, Shinkai N, Kusakabe A, Matsuura K, Iio E, Takahashi S, Yan G, Ikeda K, Joh T. Pitavastatin inhibits hepatic steatosis and fibrosis in non-alcoholic steatohepatitis model rats. *Hepatol Res* 2011; **41**: 375-385 [PMID: 21276150 DOI: 10.1111/j.1872-034X.2010.00769.x]
 - 20 **Zafra C**, Abalde JG, Turnes J, Berzigotti A, Fernández M, Garca-Pagán JC, Rodés J, Bosch J. Simvastatin enhances hepatic nitric oxide production and decreases the hepatic vascular tone in patients with cirrhosis. *Gastroenterology* 2004; **126**: 749-755 [PMID: 14988829 DOI: 10.1053/j.gastro.2003.12.007]
 - 21 **Oberti F**, Pilette C, Rifflet H, Maïga MY, Moreau A, Gallois Y, Girault A, le Bouil A, Le Jeune JJ, Saumet JL, Feldmann G, Calès P. Effects of simvastatin, pentoxifylline and spirinolactone on hepatic fibrosis and portal hypertension in rats with bile duct ligation. *J Hepatol* 1997; **26**: 1363-1371 [PMID: 9210625 DOI: 10.1016/S0168-8278(97)80473-4]
 - 22 **Huang HC**, Chang CC, Wang SS, Chan CY, Lee FY, Chuang CL, Hsin IF, Teng TH, Lin HC, Lee SD. Pravastatin for thioacetamide-induced hepatic failure and encephalopathy. *Eur J Clin Invest* 2012; **42**: 139-145 [PMID: 21749370 DOI: 10.1111/j.1365-2362.2011.02566.x]
 - 23 **Trebicka J**, Hennenberg M, Odenthal M, Shir K, Klein S, Granzow M, Vogt A, Dienes HP, Lammert F, Reichen J, Heller J, Sauerbruch T. Atorvastatin attenuates hepatic fibrosis in rats after bile duct ligation via decreased turnover of hepatic stellate cells. *J Hepatol* 2010; **53**: 702-712 [PMID: 20633948 DOI: 10.1016/j.jhep.2010.04.025]
 - 24 **Natarajan SK**, Thomas S, Ramamoorthy P, Basivireddy J, Pulimood AB, Ramachandran A, Balasubramanian KA. Oxidative stress in the development of liver cirrhosis: a comparison of two different experimental models. *J Gastroenterol Hepatol* 2006; **21**: 947-957 [PMID: 16724977 DOI: 10.1111/j.1440-1746.2006.04231.x]
 - 25 **Müller D**, Sommer M, Kretzschmar M, Zimmermann T, Buko VU, Lukivskaya O, Dargel R. Lipid peroxidation in thioacetamide-induced macronodular rat liver cirrhosis. *Arch Toxicol* 1991; **65**: 199-203 [PMID: 2053847 DOI: 10.1007/BF02307309]
 - 26 **Porter WR**, Neal RA. Metabolism of thioacetamide and thioacetamide S-oxide by rat liver microsomes. *Drug Metab Dispos* 1978; **6**: 379-388 [PMID: 28917]
 - 27 **Hori N**, Okanoue T, Sawa Y, Mori T, Kashima K. Hemodynamic characterization in experimental liver cirrhosis induced by thioacetamide administration. *Dig Dis Sci* 1993; **38**: 2195-2202 [PMID: 8261820 DOI: 10.1007/BF01299895]
 - 28 **Batts KP**, Ludwig J. Chronic hepatitis. An update on terminology and reporting. *Am J Surg Pathol* 1995; **19**: 1409-1417 [PMID: 7503362 DOI: 10.1097/0000478-199512000-00007]
 - 29 **Woessner JF**. The determination of hydroxyproline in tissue and protein samples containing small proportions of this imino acid. *Arch Biochem Biophys* 1961; **93**: 440-447 [PMID: 13786180 DOI: 10.1016/0003-9861(61)90291-0]
 - 30 **Wills ED**. Lipid peroxide formation in microsomes. General considerations. *Biochem J* 1969; **113**: 315-324 [PMID: 4390101]
 - 31 **Jaster R**, Brock P, Sparmann G, Emmrich J, Liebe S. Inhibition of pancreatic stellate cell activation by the hydroxy-methylglutaryl coenzyme A reductase inhibitor lovastatin. *Biochem Pharmacol* 2003; **65**: 1295-1303 [PMID: 12694870 DOI: 10.1016/S0006-2952(03)00075-3]
 - 32 **Ikeuchi H**, Kuroiwa T, Yamashita S, Hiramatsu N, Maeshima A, Kaneko Y, Hiromura K, Ueki K, Nojima Y. Fluvastatin reduces renal fibroblast proliferation and production of type III collagen: therapeutic implications for tubulointerstitial fibrosis. *Nephron Exp Nephrol* 2004; **97**: e115-e122 [PMID: 15331935 DOI: 10.1159/000079176]
 - 33 **Moriyama T**, Kawada N, Nagatoya K, Takeji M, Horio M, Ando A, Imai E, Hori M. Fluvastatin suppresses oxidative stress and fibrosis in the interstitium of mouse kidneys with unilateral ureteral obstruction. *Kidney Int* 2001; **59**: 2095-2103 [PMID: 11380811 DOI: 10.1046/j.1523-1755.2001.0590062095.x]
 - 34 **Tan A**, Levrey H, Dahm C, Polunovsky VA, Rubins J, Bitterman PB. Lovastatin induces fibroblast apoptosis in vitro and in vivo. A possible therapy for fibroproliferative disorders. *Am J Respir Crit Care Med* 1999; **159**: 220-227 [PMID: 9872842]
 - 35 **Wassmann S**, Laufs U, Müller K, Konkol C, Ahlborn K, Bäumer AT, Linz W, Böhm M, Nickenig G. Cellular antioxidant effects of atorvastatin in vitro and in vivo. *Arterioscler Thromb Vasc Biol* 2002; **22**: 300-305 [PMID: 11834532 DOI: 10.1161/hq0202.104081]
 - 36 **Shishehbor MH**, Brennan ML, Aviles RJ, Fu X, Penn MS, Sprecher DL, Hazen SL. Statins promote potent systemic antioxidant effects through specific inflammatory pathways. *Circulation* 2003; **108**: 426-431 [PMID: 12860913 DOI: 10.1161/01.CIR.0000080895.05158.8B]
 - 37 **Haendeler J**, Hoffmann J, Zeiher AM, Dimmeler S. Antioxidant effects of statins via S-nitrosylation and activation of thioredoxin in endothelial cells: a novel vasculoprotective function of statins. *Circulation* 2004; **110**: 856-861 [PMID: 15289372 DOI: 10.1161/01.CIR.0000138743.09012.93]
 - 38 **Hsu M**, Muchova L, Morioka I, Wong RJ, Schröder H, Stevenson DK. Tissue-specific effects of statins on the expression of heme oxygenase-1 in vivo. *Biochem Biophys Res Commun* 2006; **343**: 738-744 [PMID: 16563347 DOI: 10.1016/j.bbrc.2006.03.036]
 - 39 **Bruck R**, Shirin H, Aeed H, Matas Z, Hochman A, Pines M, Avni Y. Prevention of hepatic cirrhosis in rats by hydroxyl radical scavengers. *J Hepatol* 2001; **35**: 457-464 [PMID: 11682029 DOI: 10.1016/S0168-8278(01)00163-5]

- 40 **Schwartz YSh**, Dushkin MI, Komarova NI, Vorontsova EV, Kuznetsova IS. Cholesterol-induced stimulation of postinflammatory liver fibrosis. *Bull Exp Biol Med* 2008; **145**: 692-695 [PMID: 19110552 DOI: 10.1007/s10517-008-0175-6]
- 41 **Gardner JL**, Turner SM, Bautista A, Lindwall G, Awada M, Hellerstein MK. Measurement of liver collagen synthesis by heavy water labeling: effects of profibrotic toxicants and antifibrotic interventions. *Am J Physiol Gastrointest Liver Physiol* 2007; **292**: G1695-G1705 [PMID: 17347453 DOI: 10.1152/ajpgi.00209.2006]
- 42 **Medina J**, Moreno-Otero R. Pathophysiological basis for antioxidant therapy in chronic liver disease. *Drugs* 2005; **65**: 2445-2461 [PMID: 16296871 DOI: 10.2165/00003495-200565170-00003]
- 43 **Dey A**, Cederbaum AI. Alcohol and oxidative liver injury. *Hepatology* 2006; **43**: S63-S74 [PMID: 16447273 DOI: 10.1002/hep.20957]
- 44 **Yang JI**, Yoon JH, Bang YJ, Lee SH, Lee SM, Byun HJ, Myung SJ, Kim W, Lee HS. Synergistic antifibrotic efficacy of statin and protein kinase C inhibitor in hepatic fibrosis. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G126-G132 [PMID: 19910526 DOI: 10.1152/ajpgi.00299.2009]
- 45 **Trebicka J**, Hennenberg M, Laleman W, Shelest N, Biecker E, Schepke M, Nevens F, Sauerbruch T, Heller J. Atorvastatin lowers portal pressure in cirrhotic rats by inhibition of RhoA/Rho-kinase and activation of endothelial nitric oxide synthase. *Hepatology* 2007; **46**: 242-253 [PMID: 17596891 DOI: 10.1002/hep.21673]

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Hyperglycemia is a significant prognostic factor of hepatocellular carcinoma after curative therapy

Takanori Hosokawa, Masayuki Kurosaki, Kaoru Tsuchiya, Shuya Matsuda, Masaru Muraoka, Yuichiro Suzuki, Nobuharu Tamaki, Yutaka Yasui, Toru Nakata, Takashi Nishimura, Shoko Suzuki, Ken Ueda, Hiroyuki Nakanishi, Jun Itakura, Yuka Takahashi, Namiki Izumi

Takanori Hosokawa, Masayuki Kurosaki, Kaoru Tsuchiya, Shuya Matsuda, Masaru Muraoka, Yuichiro Suzuki, Nobuharu Tamaki, Yutaka Yasui, Toru Nakata, Takashi Nishimura, Shoko Suzuki, Ken Ueda, Hiroyuki Nakanishi, Jun Itakura, Yuka Takahashi, Namiki Izumi, Division of Gastroenterology and Hepatology, Musashino Red Cross Hospital, Tokyo 180-8610, Japan

Author contributions: Hosokawa T and Kurosaki M contributed equally to this work; Kurosaki M and Izumi N made substantial contributions to the conception and design of the study; Tsuchiya K, Matsuda S, Muraoka M, Suzuki Y, Tamaki N, Yasui Y, Nakata T, Nishimura T, Suzuki S, Ueda K, Nakanishi H, Itakura J, Takahashi Y and Izumi N collected the clinical data; Hosokawa T and Kurosaki M contributed to the analysis and interpretation of the data; Hosokawa T wrote the draft of the manuscript; Kurosaki M and Izumi N made critical revisions of the manuscript; and Izumi N obtained a research fund.

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Correspondence to: Namiki Izumi, MD, Division of Gastroenterology and Hepatology, Musashino Red Cross Hospital, 1-26-1 Kyonan-cho, Musashino-shi, Tokyo 180-8610, Japan. nizumi@musashino.jrc.or.jp

Telephone: +81-422-323111 Fax: +81-422-329551

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Abstract

AIM: To evaluate whether metabolic factors are related to distant recurrence of hepatocellular carcinoma (HCC) and survival after curative treatment.

METHODS: This retrospective study included 344 patients whose HCC was treated curatively by radio-frequency ablation (RFA) therapy. The mean age was 67.6 years and the mean observation period was 4.04 years. The etiological background of liver disease was

hepatitis B virus infection in 30, hepatitis C virus infection in 278, excessive alcohol drinking in 9, and other in 27 patients. The Child-Pugh classification grade was A ($n = 307$) or B ($n = 37$). The number of HCC nodules was one in 260, two in 61, and three in 23 patients. For surveillance of HCC recurrence after curative therapy with RFA, patients were radiologically evaluated every 3 mo. Factors associated with distant recurrence of HCC or survival were studied.

RESULTS: Inadequate maintenance of blood glucose in diabetic patients was associated with higher incidence of distant recurrence. The 1-, 2-, and 3-year recurrence rates were significantly higher in diabetic patients with inadequate maintenance of blood glucose compared with the others: 50.6% vs 26.8%, 83.5% vs 54.4%, and 93.8% vs 73.0%, respectively ($P = 0.0001$). Inadequate maintenance of blood glucose was an independent predictor of distant recurrence [adjusted relative risk 1.97 (95%CI, 1.33-2.91), ($P = 0.0007$)] after adjustment for other risk factors, such as number of HCC nodules [2.03 (95%CI, 1.51-2.73), $P < 0.0001$] and initial level of serum alpha fetoprotein (AFP) [1.43 (95%CI, 1.04-1.97), $P = 0.028$]. Obesity was not an independent predictor of recurrence. The incidence of distant recurrence did not differ between diabetic patients with adequate maintenance of blood glucose and non-diabetic patients. Among 232 patients who had HCC recurrence, 138 had a second recurrence. The 1-, 2-, and 3-year rates of second recurrence were significantly higher in diabetic patients with inadequate maintenance of blood glucose than in the others: 9.0% vs 5.9%, 53.1% vs 24.3%, and 69.6% vs 42.3%, respectively ($P = 0.0021$). Inadequate maintenance of blood glucose in diabetic patients [1.99 (95%CI, 1.23-3.22), $P = 0.0049$] and presence of multiple HCC nodules [1.53 (95%CI, 1.06-2.22), $P = 0.024$] were again significantly associated with second HCC recurrence. Inadequate maintenance of blood glucose in diabetic

patients was also a significant predictor of poor survival [2.77 (95%CI, 1.38-5.57), $P = 0.0046$] independent of excessive alcohol drinking [6.34 (95%CI, 1.35-29.7), $P = 0.019$], initial level of serum AFP [3.40 (95%CI, 1.88-6.18), $P < 0.0001$] and Child-Pugh classification grade B [2.24 (95%CI, 1.12-4.46), $P = 0.022$]. Comparing diabetic patients with inadequate maintenance of blood glucose *vs* the others, the 1-, 2-, and 3-year survival rates were significantly lower in diabetic patients with inadequate maintenance of blood glucose: 92% *vs* 99%, 85% *vs* 96%, and 70% *vs* 92%, respectively ($P = 0.0003$).

CONCLUSION: Inadequate maintenance of blood glucose in diabetic patients is a significant risk factor for recurrence of HCC and for poor survival after curative RFA therapy.

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Key words: Hyperglycemia; Hepatocellular carcinoma; Recurrence; Radio frequency ablation; Survival

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INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common cancers worldwide^[1] and its incidence has been increasing in many countries^[2]. Surgical resection, liver transplantation, and local ablation therapy, such as radiofrequency ablation (RFA) therapy, have been considered as efficient curative therapies for HCC. RFA therapy is now widely performed in patients with small HCC^[3] and a randomized controlled study demonstrated that the survival rates were similar in patients with small HCC receiving RFA or surgical resection^[4]. A characteristic of HCC is its high rate of recurrence after curative resection or local ablation therapy, reaching approximately 80% within 5 years^[5-7]. Identification of factors related to recurrence of HCC and therapeutic intervention targeting these factors may lead to prevention of frequent recurrence of HCC and improved survival.

Tumor factors, such as the number of HCC nodules and their size, are associated with the recurrence of HCC and survival prognosis^[8-10]. Another factor that is associated with the recurrence of HCC and survival is the hepatic reserve function at the time of HCC therapy^[8,10,11]. Hepatitis C virus (HCV) and hepatitis B virus (HBV) infection are the major causes responsible for 80% of HCC cases^[2] and antiviral therapy targeting HCV^[12,13] or

HBV^[14] has been shown to decrease HCC recurrence, and improve hepatic reserve function and survival. Non-alcoholic steatohepatitis (NASH) has also received attention as a cause of HCC^[15]. Metabolic factors, such as obesity and diabetes, are closely linked to the etiology of NASH. These metabolic factors have also been identified as risk factors for several other types of cancer. Obesity is associated with increased mortality rates of several cancers^[16,17] and diabetes is also reported as a risk factor for liver, pancreatic, renal, and colon cancers^[18,19]. If these metabolic factors are related to the recurrence of HCC, therapeutic intervention targeting these factors may lead to prevention of frequent recurrence of HCC and improved survival. The impact of diabetes on the recurrence of HCC after treatment has been discussed, but with conflicting results^[20-23].

In this study, factors contributing to the recurrence and prognosis of HCC after curative treatment were analyzed. We found that inadequate maintenance of blood glucose was related to the high rate of HCC recurrence and poor survival.

MATERIALS AND METHODS

Patients whose HCC was treated by RFA at the Musashino Red Cross Hospital were studied retrospectively for factors associated with recurrence of HCC and survival. The inclusion criteria were as follows: (1) HCC treated curatively with RFA at the Musashino Red Cross Hospital between 1999 and 2007; (2) maximum diameter of HCC nodule ≤ 3 cm; (3) number of HCC nodules ≤ 3 ; (4) no previous history of treatment for HCC; and (5) follow-up observation for at least 6 mo after RFA therapy. 344 patients met these criteria, including 140 women and 204 men, with a mean age of 67.6 years and mean observation time of 4.04 years. The clinical characteristics of the patients are summarized in Table 1. The etiological background of liver disease was HBV infection in 30, HCV infection in 278, excessive alcohol drinking (intake of ethanol ≥ 60 g/d for ≥ 5 years continuously) in 9, and non-B non-C non-alcoholic etiology in 27 patients. The Child-Pugh classification grade was either A ($n = 307$) or B ($n = 37$). The number of HCC nodules was one in 260, two in 61, and three in 23 patients. Thus, 260 patients had a single lesion, and 84 had multiple lesions. The maximum diameter of HCC nodules was 19.9 ± 0.3 mm.

Obesity was defined as a body mass index > 25 kg/m² according to the definition of the Japan Society for the Study of Obesity^[24]. Blood glucose was measured monthly for 6 mo after HCC treatment and the average value was determined. Inadequate maintenance of blood glucose was defined as an average value of blood glucose ≥ 200 mg/dL. The level of hemoglobin A1c (HbA1c) was not used in the present study because the lifespan of erythrocytes is shortened due to hypersplenism in patients with chronic hepatitis or cirrhosis, leading to lower HbA1c levels relative to the blood glucose level^[25]. Diagnosis of type 2 diabetes was made according to the

Table 1 Characteristics of patients undergoing curative radiofrequency ablation for hepatocellular carcinoma *n* (%)

Variable	Value
Sex (male/female)	204/140
Age(yr)	67.6 ± 8.4
Etiology of liver disease: HBV/HCV/NBNC	30/278/36
AST (IU/L)	84.0 ± 34.5
ALT (IU/L)	73.2 ± 36.5
GGT (IU/L)	82.9 ± 96.8
T-Chol (mg/dL)	157.8 ± 32.0
TG (mg/dL)	112.3 ± 55.7
Mean blood sugar (mg/dL)	139.3 ± 44.0
Diabetes mellitus	159 (48)
BMI > 25 kg/m ²	86 (25)
Maximum diameter of HCC nodule (mm)	19.9 ± 0.3
Number of HCC nodules: single/2 or 3	260/84
AFP (ng/mL)	214 ± 1025
Alcohol drinking > 60 g/d	9 (2.6)
Child-Pugh grade: A/B	307/37

HBV: Hepatitis B virus; HCV: Hepatitis C virus; NBNC: Neither HBV nor HCV; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; GGT: γ -glutamyltransferase; T-Chol: Total cholesterol; TG: Triglyceride; BMI: Body mass index; AFP: α -fetoprotein; HCC: Hepatocellular carcinoma.

American Diabetes Association criteria of a fasting blood glucose level ≥ 126 mg/dL (≥ 7.0 mmol/L) and/or HbA1c level ≥ 6.5 ^[26]. After initial treatment of HCC by RFA, the ablated area was confirmed by contrast-enhanced computed tomography (CT) within one week. If the ablated area was not sufficient, then RFA therapy was repeated until the HCC nodule was completely ablated.

HCC surveillance and diagnosis of recurrence

Diagnosis of HCC was based on abdominal ultrasonography, contrast-enhanced CT, magnetic resonance imaging (MRI), or angiography. Classical HCC was diagnosed for tumors showing vascular enhancement with washout on at least two types of diagnostic imaging. Tumor biopsy was used to diagnose tumors with non-classical imaging findings.

For surveillance of HCC recurrence after curative therapy with RFA, patients were evaluated by abdominal ultrasonography, contrast-enhanced CT, or contrast-enhanced MRI every three months. Recurrence of HCC was diagnosed based on a new lesion detected by ultrasonography showing vascular enhancement with washout on CT or MRI. If the tumor was not hypervascular, a tumor biopsy was performed to confirm the diagnosis.

Statistical analysis

For analysis of survival and recurrence, the time of initial RFA treatment was defined as day zero. Survival rate and recurrence rate were analyzed by the Kaplan-Meier method and log rank test. Multivariate analysis was performed using a Cox proportional hazard model. Data were analyzed using StatView Version 5.0 (SAS Institute Inc, Cary, North Carolina, United States) and IBM-SPSS statistics version 18 (IBM SPSS Inc, Chicago, IL, United States). Statistical significance was set at $P < 0.05$.

RESULTS

Factors associated with HCC recurrence

Of the 344 patients whose HCC was curatively treated by RFA, 232 had HCC recurrence. The 1-, 2-, and 3-year recurrence rates were 29.3%, 57.5%, and 75.2%, respectively. On univariate analysis, inadequate maintenance of blood glucose, higher initial level of serum AFP and multiple HCC nodules were significantly associated with HCC recurrence. Obesity ($P = 0.06$) and diabetes ($P = 0.65$) were not significantly associated with HCC recurrence.

Thirty-seven patients had diabetes with inadequate maintenance of blood glucose, 122 patients had diabetes with adequate maintenance of blood glucose, and 185 patients did not have diabetes. The HCC recurrence rate was significantly higher in diabetic patients with inadequate maintenance of blood glucose than in the others ($P = 0.0001$) (Figure 1A).

Comparing patients with diabetes ($n = 159$) and patients who did not have diabetes ($n = 185$), there was no significant difference in the recurrence rate ($P = 0.65$). Upon comparison of the three groups, i.e., the diabetes with inadequate maintenance of blood glucose group, the diabetes with adequate maintenance of blood glucose group, and the non-diabetes group, the recurrence rate was significantly higher in the diabetes with inadequate maintenance of blood glucose group than in the other two groups ($P = 0.0001$) (Figure 1B). On the other hand, there was no significant difference in the HCC recurrence rate between the diabetes patients with adequate maintenance of blood glucose group and the non-diabetes group.

In terms of the number of HCC nodules, namely, single ($n = 260$) *vs* multiple ($n = 84$), the recurrence rate was significantly higher in patients with multiple HCC nodules ($P = 0.0001$). Within each subgroup of patients with single and multiple HCC nodules, diabetes with inadequate maintenance of blood glucose was significantly associated with recurrence of HCC (single, $P = 0.006$; multiple, $P = 0.025$) (Figure 2A, B). In terms of the initial level of serum AFP ≥ 100 ng/mL ($n = 70$) *vs* < 100 ng/mL ($n = 274$), the recurrence rate was significantly higher in patients with AFP ≥ 100 ng/mL ($P = 0.018$). Within each subgroup of patients with AFP ≥ 100 ng/mL and < 100 ng/mL, diabetes with inadequate maintenance of blood glucose was associated with a higher rate of recurrence (AFP ≥ 100 ng/mL, $P = 0.005$; AFP < 100 ng/mL, $P = 0.017$) (Figure 2C, D).

Independent risk factors for distant recurrence of HCC on multivariate analysis were inadequate maintenance of blood glucose in diabetic patients [adjusted relative risk, 1.97 (95%CI, 1.33-2.91), $P = 0.0007$], multiple HCC nodules [2.03 (1.51-2.73), $P < 0.0001$], and AFP ≥ 100 ng/mL [1.43 (1.04-1.97), $P = 0.028$] (Table 2).

Factors associated with second recurrence

Among the 232 patients who had HCC recurrence, 138 had a second recurrence. Regarding second recurrence, inadequate maintenance of blood glucose in diabetic pa-

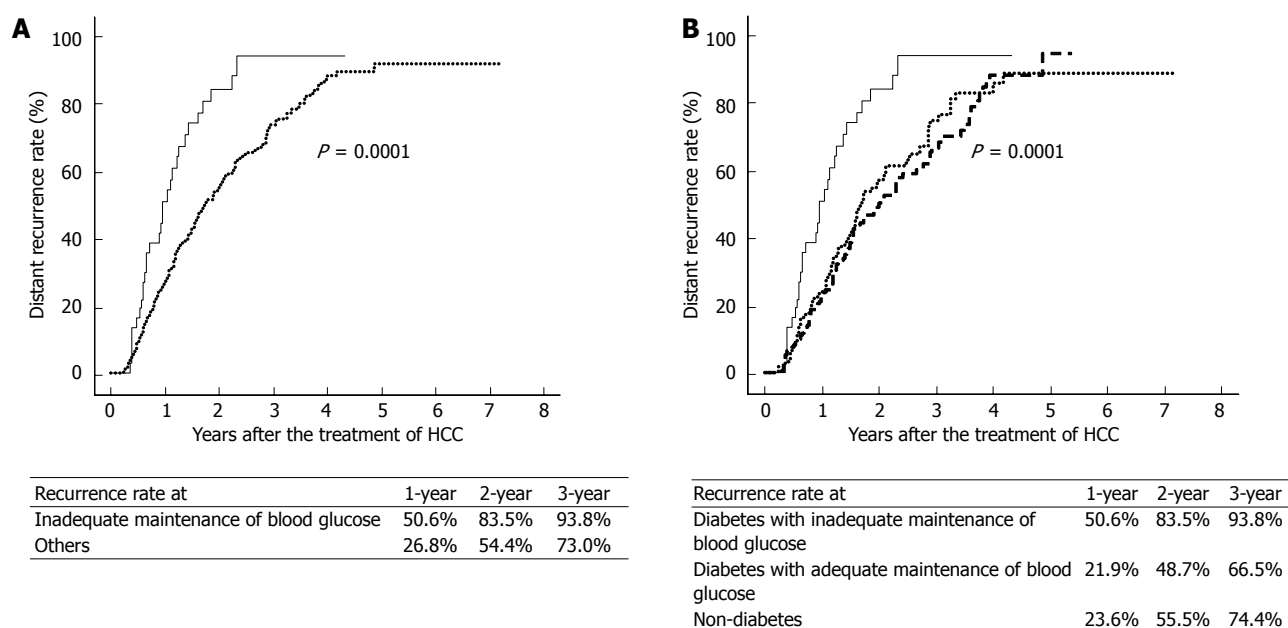


Figure 1 Kaplan-Meier curves showing a higher rate of hepatocellular carcinoma recurrence in diabetic patients with hyperglycemia. A: The cumulative incidence of the recurrence of hepatocellular carcinoma (HCC) was significantly higher in diabetic patients with inadequate maintenance of blood glucose (blood glucose ≥ 200 mg/dL solid line) than in the others (dotted line) ($P = 0.0001$); B: The HCC recurrence rate was significantly higher in diabetic patients with inadequate maintenance of blood glucose (solid line) than in diabetic patients with adequate maintenance of blood glucose (blood glucose < 200 mg/dL, broken line) or non-diabetic patients (dotted line) ($P = 0.0001$). There was no significant difference in HCC recurrence rate between diabetic patients with adequate maintenance of blood glucose and non-diabetic patients.

Table 2 Multivariate analysis of factors associated with recurrence of hepatocellular carcinoma

Factors	Odds ratio (95%CI)	P-value
First recurrence		
Inadequate maintenance of blood glucose	1.97 (1.33-2.91)	0.0007
Multiple HCC nodules	2.03 (1.51-2.73)	< 0.0001
AFP ≥ 100 ng/mL	1.43 (1.04-1.97)	0.028
Second recurrence		
Inadequate maintenance of blood glucose (mg/dL)	1.99 (1.23-3.22)	0.0049
Multiple HCC nodules	1.53 (1.06-2.22)	0.024

Inadequate maintenance of blood glucose was defined as an average of casual blood glucose of ≥ 200 mg/dL. HCC: Hepatocellular carcinoma; AFP: α -fetoprotein.

tients and multiple HCC nodules were again significantly associated with HCC recurrence. Obesity ($P = 0.18$), diabetes ($P = 0.31$) and initial level of serum AFP ($P = 0.08$) were not associated with second recurrence. In terms of the number of HCC nodules, namely, single *vs* multiple, the 1-, 2-, and 3-year recurrence rates were significantly higher in patients with multiple lesions (6.4% *vs* 6.1%, 39.3% *vs* 23.1%, and 52.5% *vs* 42.3%, respectively, $P = 0.013$). Upon comparing diabetic patients with inadequate maintenance of blood glucose *vs* the others, the rate of second recurrence was significantly higher in diabetic patients with inadequate maintenance of blood glucose ($P = 0.0021$) (Figure 3A). Upon comparing patients with diabetes *vs* patients who did not have diabetes, the rates of second recurrence were not significantly different (P

$= 0.31$). Upon comparison of the three groups, i.e., the diabetes with inadequate maintenance of blood glucose group, the diabetes with adequate maintenance of blood glucose group, and the non-diabetes group, the second recurrence rate was again significantly higher in the diabetes with inadequate maintenance of blood glucose group than in the other two groups ($P = 0.0035$) (Figure 3B). On the other hand, there was no significant difference in the second recurrence rate between the diabetes with adequate maintenance of blood glucose group and the non-diabetes group.

Independent risk factors for second recurrence of HCC on multivariate analysis were inadequate maintenance of blood glucose [1.99 (95%CI, 1.23-3.22), $P = 0.0049$] and multiple HCC nodules [1.53 (95%CI, 1.06-2.22), $P = 0.024$] (Table 2).

Factors associated with survival

There were 52 HCC-related or hepatic failure deaths. On univariate analysis, inadequate maintenance of blood glucose, excessive alcohol drinking, higher initial level of serum AFP and Child-Pugh classification grade B were significantly associated with survival. Obesity ($P = 0.81$) and diabetes ($P = 0.11$) were not significantly associated with survival.

Upon comparing diabetic patients with inadequate maintenance of blood glucose *vs* the others, the survival rate was significantly lower in patients with inadequate maintenance of blood glucose ($P = 0.0003$) (Figure 4A). Upon comparing diabetic patients *vs* non-diabetic patients, the survival rates were not significantly different

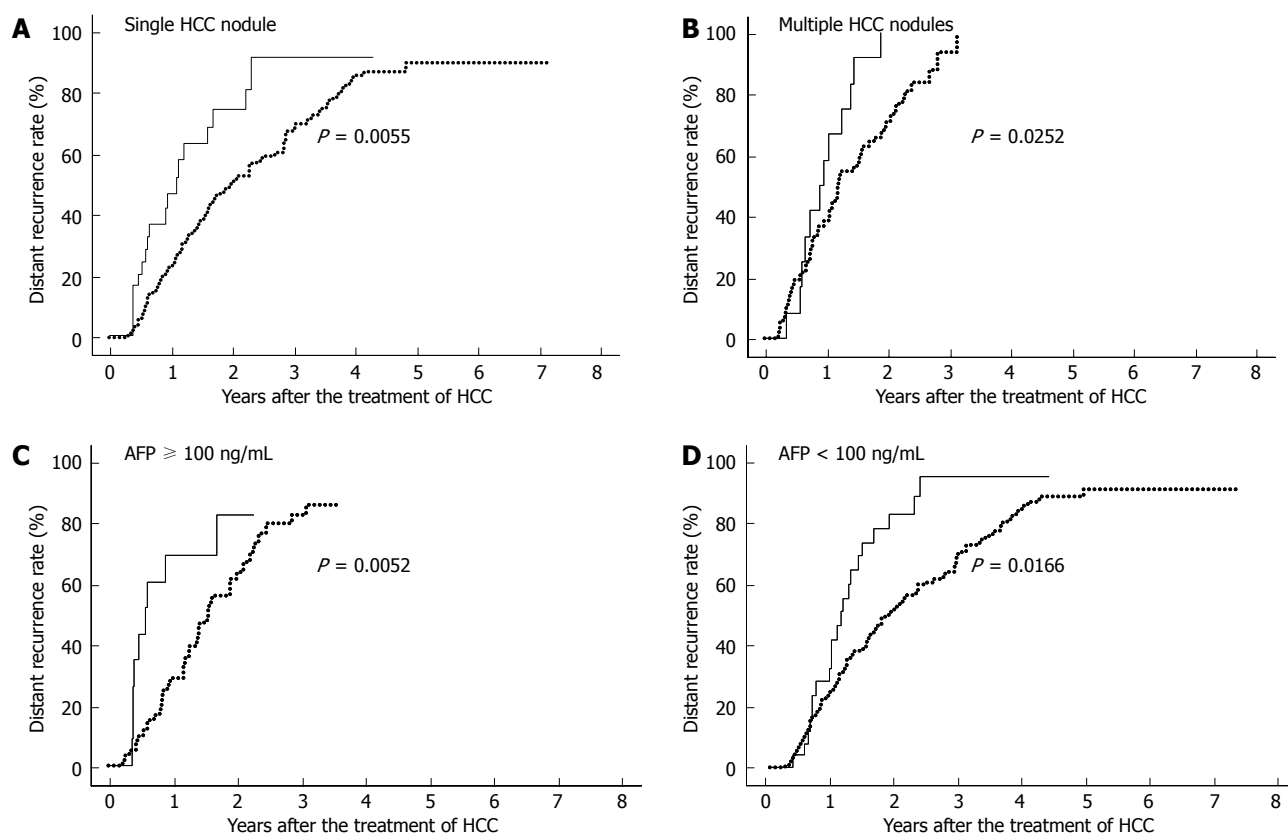


Figure 2 Diabetic patients with inadequate maintenance of blood glucose have higher rate of hepatocellular carcinoma recurrence after stratification by other risk factors. A: $P = 0.006$ for single hepatocellular carcinoma (HCC) nodule; B: $P = 0.025$ for multiple HCC nodules; C: $P = 0.005$ for AFP ≥ 100 ng/mL; D: $P = 0.017$ for α -fetoprotein (AFP) < 100 ng/mL. The cumulative incidence of the recurrence of HCC was significantly higher in diabetic patients with inadequate maintenance of blood glucose (solid line) than in the others (dotted line). after stratification by number of HCC nodules and by initial level of AFP.

Table 3 Multivariable analysis of factors associated with survival

Factors	Odds ratio (95%CI)	P-value
Inadequate maintenance of blood glucose	2.77 (1.38-5.57)	0.0046
Alcohol drinking ≥ 60 g/d	6.34 (1.35-29.7)	0.019
Child Pugh grade B	2.24 (1.12-4.46)	0.022
AFP ≥ 100 ng/mL	3.40 (1.88-6.18)	< 0.0001

Inadequate maintenance of blood glucose was defined as an average of casual blood glucose of ≥ 200 mg/dL. AFP: α -fetoprotein.

($P = 0.11$). of the survival rate was compared among the three groups, i.e., the diabetes with inadequate maintenance of blood glucose group, the diabetes with adequate maintenance of blood glucose group, and the non-diabetes group. The survival rate was significantly poorer in the diabetes with inadequate maintenance of blood glucose group than in the other two groups ($P = 0.0003$) (Figure 4B), while it did not differ between the diabetes with adequate maintenance of blood glucose group and the non-diabetes group.

The number of HCC nodules, which was a significant factor for HCC recurrence, was not related to survival ($P = 0.34$). Patients with excessive alcohol drinking had poor survival prognosis compared to those with non-excessive or no alcohol drinking ($P = 0.046$). Survival was

better in patients in Child-Pugh A class than in patients in Child-Pugh B class ($P = 0.0082$). AFP ≥ 100 ng/mL was associated with poor survival compared with AFP < 100 ng/mL ($P < 0.0001$).

On multivariate analysis, inadequate maintenance of blood glucose was a significant predictor of poor survival [2.77 (95%CI, 1.38-5.57), $P = 0.0046$] independent of excessive alcohol drinking [6.34 (95%CI, 1.35-29.7), $P = 0.019$], initial level of serum AFP ≥ 100 ng/mL [3.40 (95%CI, 1.88-6.18), $P < 0.0001$] and Child-Pugh classification grade B [2.24 (95%CI, 1.12-4.46), $P = 0.022$] (Table 3).

DISCUSSION

The impact of metabolic factors, such as hyperglycemia, diabetes and obesity, on distant recurrence and survival after curative RFA therapy for HCC was analyzed retrospectively. We identified that inadequate maintenance of blood glucose in diabetic patients was a significant and independent risk factor for early recurrence of HCC and a risk factor for poor survival, whereas obesity and diabetes were not. Diabetic patients with inadequate maintenance of blood glucose had a higher rate of HCC recurrence and poorer survival compared with diabetic patients with adequate maintenance of blood glucose and non-diabetic patients. In other words, even in patients with diabetes, if

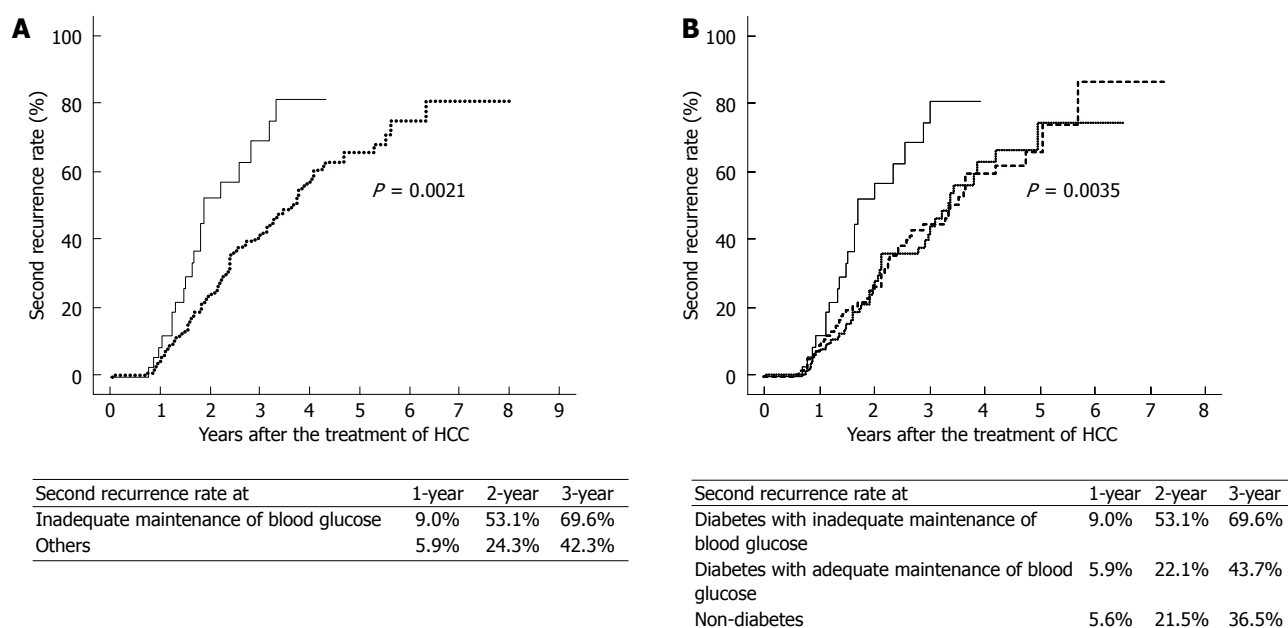


Figure 3 Kaplan-Meier curves showing a higher rate of second recurrence of hepatocellular carcinoma in diabetic patients with inadequate maintenance of blood glucose. **A:** The cumulative incidence of the second recurrence of hepatocellular carcinoma (HCC) was significantly higher in diabetic patients with inadequate maintenance of blood glucose (blood glucose ≥ 200 mg/dL solid line) than in the others (dotted line) ($P = 0.002$); **B:** The rate of second recurrence of HCC was significantly higher in diabetic patients with inadequate maintenance of blood glucose (solid line) than in diabetic patients with adequate maintenance of blood glucose (blood glucose < 200 mg/dL, broken line) or non-diabetic patients (dotted line) ($P = 0.004$). There was no significant difference in the rate of second recurrence of HCC between diabetic patients with adequate maintenance of blood glucose and non-diabetic patients.

the blood glucose was adequately maintained, the HCC recurrence rate and survival did not differ significantly compared with those in non-diabetic patients. These results indicate the possibility that adequate management of hyperglycemia may lead to reduction in the risk of HCC recurrence and improvement of overall survival.

The contribution of diabetes to the development of HCC has been confirmed in several reports^[27-30]. The impact of diabetes on the recurrence of HCC after treatment has also been discussed, but with conflicting results^[20-22]. A recent study from Taiwan demonstrated that diabetes may not affect the intra-hepatic HCC recurrence and survival after RFA^[23]. The results of the present study also indicated that diabetes itself is not a significant risk factor if the level of blood glucose is adequately managed. Rather, hyperglycemia was a significant risk factor for the recurrence of HCC. There may be several mechanisms involved in the relationship between hyperglycemia and HCC recurrence. Hyperglycemia promotes cancer cell proliferation in pancreatic cancer cells and breast cancer cells^[31-33] through accelerated cell cycle progression or through the production of reactive oxygen species, leading to activation of protein kinase C and increased DNA synthesis in cancer cells^[34]. A previous study in hepatitis C patients indicated that hyperglycemia after challenge with 75-g oral glucose tolerance test was associated with the risk for HCC while hyperglycemia at fasting was not^[35]. A possible reason for this result may be that patients with post-challenge hyperglycemia may have higher fluctuations in daily glucose levels that lead to oxidative stress^[35], because it was reported that acute fluctuations in blood glucose levels cause greater oxidative stress than

sustained chronic hyperglycemia^[36]. Taken together, a possible mechanism for the relationship between higher level of casual blood glucose and development of HCC in the present study may be that daily fluctuations in serum glucose levels caused greater oxidative stress. Alternatively, hyper-insulinemia or increased level of insulin-like growth factor, which are caused by hyperglycemia, may be related to carcinogenesis^[37-39]. Insulin levels were not measured in our study; therefore, the effects of insulin could not be identified.

Discussions are now taking place on methods of treating diabetes from the standpoint of cancer prevention. Control of hyperglycemia could reduce cancer incidence, which means that hyperglycemia could directly contribute to the development of cancer^[39]. The results of our study also showed that adequate management of hyperglycemia may lead to reduction in the risk of HCC recurrence and improvement of overall survival. Improvement in insulin resistance is undoubtedly the most important factor for the treatment of diabetes, but glycemic control is often difficult to achieve with dietary therapy, exercise, or insulin resistance-improving drugs alone. It was reported that metformin may be associated with a lower risk of cancer^[38] and there is a theoretical concern that exogenous insulin may be associated with an increased risk of cancer^[40]. In fact, a recent study reported that insulin therapy in patients with HCV infection is linked with the development of HCC^[41]. On the other hand, with insulin treatment, concomitant use of metformin has been reported to offset the carcinogenic risk of insulin^[42]. Whether glycemic control should be a priority, or whether avoiding hyper-insulinemia because

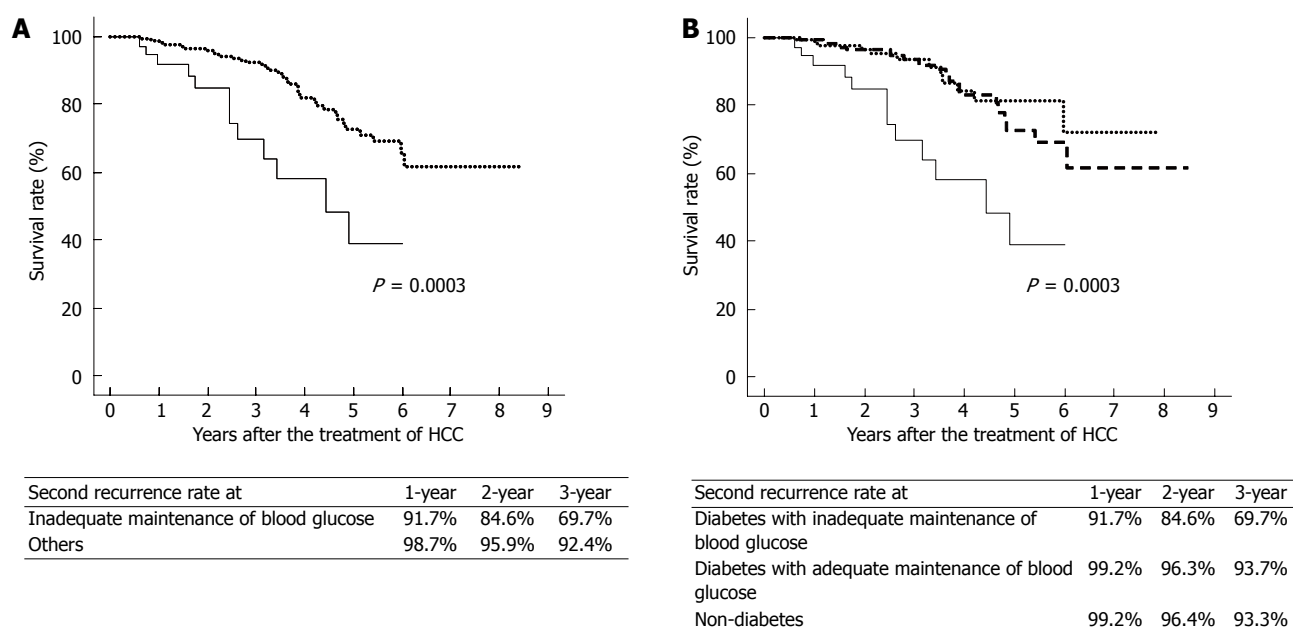


Figure 4 Patients with inadequate maintenance of blood glucose have a lower survival rate. A: The survival rate after curative local ablation therapy for hepatocellular carcinoma (HCC) was significantly lower in diabetic patients with inadequate maintenance of blood glucose (blood glucose ≥ 200 mg/dL solid line) than in the others (dotted line) ($P = 0.0003$); B: The survival rate was significantly lower in diabetic patients with inadequate maintenance of blood glucose (solid line) than in diabetic patients with adequate maintenance of blood glucose (blood glucose < 200 mg/dL, broken line) or non-diabetic patients (dotted line) ($P = 0.0003$). There was no significant difference in survival rate between diabetic patients with adequate maintenance of blood glucose and non-diabetic patients.

of therapy should be a priority, is an issue for future investigation.

In terms of survival of HCC patients, associations with liver function and tumor factors have been reported^[10], but conflicting results have been reported for the relationship with diabetes^[20,21]. These two studies involved heterogeneous groups of HCC patients treated with various therapies, including surgery, local ablation therapy and transcatheter arterial embolization. This heterogeneity may have led to the conflicting results, because the survival of HCC patients may be strongly affected by the initial treatment. Our study involved a homogeneous patient population, i.e., all patients were initially treated curatively by RFA. The results of our study suggest that glycemic control in diabetic patients, more so than diabetes itself, plays a role in survival. The mechanism by which glycemic control and survival are related is unknown, but frequent recurrence of HCC in hyperglycemic patients and the accumulation of damage in liver function because of repeated treatment intervention for HCC may lead to worsening survival.

In conclusion, inadequate maintenance of blood glucose in diabetic patients was a significant and independent risk factor for early recurrence of HCC and for poor survival. Adequate management of hyperglycemia in diabetic patients may lead to reduction in the risk of HCC recurrence and improvement in overall survival.

COMMENTS

Background

Hepatocellular carcinoma (HCC) is one of the most common cancers worldwide. Radiofrequency ablation (RFA) therapy is an efficient curative therapy

for HCC, but long-term survival is limited because of the high rate of distant recurrence of approximately 80% within 5 years. Identification of factors related to recurrence of HCC and therapeutic intervention targeting these factors may lead to prevention of frequent recurrence of HCC and improved survival.

Research frontiers

Metabolic factors, such as obesity and diabetes, have been identified as risk factors for several types of cancer, such as cancer of the liver, pancreas, kidney, and colon. These metabolic factors may be related to recurrence of HCC. The impact of diabetes on the recurrence of HCC after treatment has been discussed, but with conflicting results.

Innovations and breakthroughs

The authors identified that inadequate maintenance of blood glucose in diabetic patients was a significant and independent risk factor for early recurrence of HCC and a risk factor for poor survival, whereas diabetes was not. In other words, even in patients with diabetes, if the blood glucose was adequately maintained, then the HCC recurrence rate and survival did not differ significantly from those in non-diabetic patients.

Applications

The results of the present study indicate the possibility that adequate management of hyperglycemia in diabetic patients may lead to reduction in the risk of HCC recurrence and improvement of overall survival.

Peer review

This is an important study in which the effect of inadequate maintenance of blood glucose in diabetes has been shown as a significant risk factor for distant recurrence of hepatocellular carcinoma and poor survival after curative radiofrequency ablation therapy.

REFERENCES

- 1 Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin* 2005; **55**: 74-108 [PMID: 15761078 DOI: 10.3322/canjclin.55.2.74]
- 2 Bosch FX, Ribes J, Díaz M, Cléries R. Primary liver cancer: worldwide incidence and trends. *Gastroenterology* 2004; **127**: S5-S16 [PMID: 15508102 DOI: 10.1053/j.gastro.2004.09.011]
- 3 Arii S, Sata M, Sakamoto M, Shimada M, Kumada T, Shiina S, Yamashita T, Kokudo N, Tanaka M, Takayama T, Kudo M.

- Management of hepatocellular carcinoma: Report of Consensus Meeting in the 45th Annual Meeting of the Japan Society of Hepatology (2009). *Hepatol Res* 2010; **40**: 667-685 [PMID: 20633193 DOI: 10.1111/j.1872-034X.2010.00673.x]
- 4 **Chen MS**, Li JQ, Zheng Y, Guo RP, Liang HH, Zhang YQ, Lin XJ, Lau WY. A prospective randomized trial comparing percutaneous local ablative therapy and partial hepatectomy for small hepatocellular carcinoma. *Ann Surg* 2006; **243**: 321-328 [PMID: 16495695 DOI: 10.1097/01.sla.0000201480.65519.b8]
 - 5 **Tateishi R**, Shiina S, Teratani T, Obi S, Sato S, Koike Y, Fujishima T, Yoshida H, Kawabe T, Omata M. Percutaneous radiofrequency ablation for hepatocellular carcinoma. An analysis of 1000 cases. *Cancer* 2005; **103**: 1201-1209 [PMID: 15690326 DOI: 10.1002/cncr.20892]
 - 6 **Izumi N**, Asahina Y, Noguchi O, Uchihara M, Kanazawa N, Itakura J, Himeno Y, Miyake S, Sakai T, Enomoto N. Risk factors for distant recurrence of hepatocellular carcinoma in the liver after complete coagulation by microwave or radiofrequency ablation. *Cancer* 2001; **91**: 949-956 [PMID: 11251946]
 - 7 **Curley SA**, Izzo F, Ellis LM, Nicolas Vauthey J, Vallone P. Radiofrequency ablation of hepatocellular cancer in 110 patients with cirrhosis. *Ann Surg* 2000; **232**: 381-391 [PMID: 10973388 DOI: 10.1097/0000658-200009000-00010]
 - 8 **Poon RT**, Fan ST, Lo CM, Liu CL, Wong J. Intrahepatic recurrence after curative resection of hepatocellular carcinoma: long-term results of treatment and prognostic factors. *Ann Surg* 1999; **229**: 216-222 [PMID: 10024103 DOI: 10.1097/0000658-199902000-00009]
 - 9 **Tung-Ping Poon R**, Fan ST, Wong J. Risk factors, prevention, and management of postoperative recurrence after resection of hepatocellular carcinoma. *Ann Surg* 2000; **232**: 10-24 [PMID: 10862190 DOI: 10.1097/0000658-200007000-00003]
 - 10 **Kudo M**, Chung H, Haji S, Osaki Y, Oka H, Seki T, Kasugai H, Sasaki Y, Matsunaga T. Validation of a new prognostic staging system for hepatocellular carcinoma: the JIS score compared with the CLIP score. *Hepatology* 2004; **40**: 1396-1405 [PMID: 15565571 DOI: 10.1002/hep.20486]
 - 11 **Fuke H**, Sugimoto K, Shiraki K, Tanaka J, Beppu T, Yoneda K, Yamamoto N, Ito K, Takaki H, Nakatsuka A, Yamakado K, Takeda K, Takei Y. Predictive factors for distant recurrence of HCV-related hepatocellular carcinoma after radiofrequency ablation combined with chemoembolization. *Aliment Pharmacol Ther* 2008; **27**: 1253-1260 [PMID: 18221404 DOI: 10.1111/j.1365-2036.2008.03627.x]
 - 12 **Hagihara H**, Nouse K, Kobayashi Y, Iwasaki Y, Nakamura S, Kuwaki K, Toshimori J, Miyatake H, Ohnishi H, Shiraha H, Yamamoto K. Effect of pegylated interferon therapy on intrahepatic recurrence after curative treatment of hepatitis C virus-related hepatocellular carcinoma. *Int J Clin Oncol* 2011; **16**: 210-220 [PMID: 21152943 DOI: 10.1007/s10147-010-0150-x]
 - 13 **Miyake Y**, Takaki A, Iwasaki Y, Yamamoto K. Meta-analysis: interferon-alpha prevents the recurrence after curative treatment of hepatitis C virus-related hepatocellular carcinoma. *J Viral Hepat* 2010; **17**: 287-292 [PMID: 19732321 DOI: 10.1111/j.1365-2893.2009.01181.x]
 - 14 **Wong JS**, Wong GL, Tsoi KK, Wong VW, Cheung SY, Chong CN, Wong J, Lee KF, Lai PB, Chan HL. Meta-analysis: the efficacy of anti-viral therapy in prevention of recurrence after curative treatment of chronic hepatitis B-related hepatocellular carcinoma. *Aliment Pharmacol Ther* 2011; **33**: 1104-1112 [PMID: 21488914 DOI: 10.1111/j.1365-2036.2011.04634.x]
 - 15 **Bugianesi E**, Leone N, Vanni E, Marchesini G, Brunello F, Carucci P, Musso A, De Paolis P, Capussotti L, Salizzoni M, Rizzetto M. Expanding the natural history of nonalcoholic steatohepatitis: from cryptogenic cirrhosis to hepatocellular carcinoma. *Gastroenterology* 2002; **123**: 134-140 [PMID: 12105842 DOI: 10.1016/S1590-8658(02)90208-2]
 - 16 **Calle EE**, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N Engl J Med* 2003; **348**: 1625-1638 [PMID: 12711737 DOI: 10.1056/NEJMoa021423]
 - 17 **Renahan AG**, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet* 2008; **371**: 569-578 [PMID: 18280327 DOI: 10.1016/S0140-6736(08)60269-X]
 - 18 **Inoue M**, Iwasaki M, Otani T, Sasazuki S, Noda M, Tsugane S. Diabetes mellitus and the risk of cancer: results from a large-scale population-based cohort study in Japan. *Arch Intern Med* 2006; **166**: 1871-1877 [PMID: 17000944]
 - 19 **Hu FB**, Manson JE, Liu S, Hunter D, Colditz GA, Michels KB, Speizer FE, Giovannucci E. Prospective study of adult onset diabetes mellitus (type 2) and risk of colorectal cancer in women. *J Natl Cancer Inst* 1999; **91**: 542-547 [PMID: 10088625 DOI: 10.1093/jnci/91.6.542]
 - 20 **Toyoda H**, Kumada T, Nakano S, Takeda I, Sugiyama K, Kiriyaama S, Tanikawa M, Sone Y, Hisanaga Y. Impact of diabetes mellitus on the prognosis of patients with hepatocellular carcinoma. *Cancer* 2001; **91**: 957-963 [PMID: 11251947]
 - 21 **Huo TI**, Wu JC, Lui WY, Huang YH, Lee PC, Chiang JH, Chang FY, Lee SD. Differential mechanism and prognostic impact of diabetes mellitus on patients with hepatocellular carcinoma undergoing surgical and nonsurgical treatment. *Am J Gastroenterol* 2004; **99**: 1479-1487 [PMID: 15307864 DOI: 10.1111/j.1572-0241.2004.30024.x]
 - 22 **Komura T**, Mizukoshi E, Kita Y, Sakurai M, Takata Y, Arai K, Yamashita T, Ohta T, Shimizu K, Nakamoto Y, Honda M, Takamura T, Kaneko S. Impact of diabetes on recurrence of hepatocellular carcinoma after surgical treatment in patients with viral hepatitis. *Am J Gastroenterol* 2007; **102**: 1939-1946 [PMID: 17573788 DOI: 10.1111/j.1572-0241.2007.01354.x]
 - 23 **Chen WT**, Macatula TC, Lin CC, Lin CJ, Lin SM. Diabetes may not affect outcomes in hepatocellular carcinoma after radio-frequency ablation. *Hepatogastroenterology* 2011; **58**: 551-557 [PMID: 21661430]
 - 24 **Youkou A**, Hasegawa T, Suzuki K, Koya T, Sakagami T, Toyabe S, Arakawa M, Gejyo F, Narita I, Suzuki E. Influence of obesity on control in asthmatic Japanese patients defined by the Japanese definition of obesity. *Intern Med* 2011; **50**: 1911-1916 [PMID: 21921368 DOI: 10.2169/internalmedicine.50.5474]
 - 25 **Aizawa N**, Enomoto H, Imanishi H, Saito M, Iwata Y, Tanaka H, Ikeda N, Sakai Y, Takashima T, Iwai T, Moriwaki E, Shimomura S, Iijima H, Nakamura H, Nishiguchi S. Elevation of the glycated albumin to glycated hemoglobin ratio during the progression of hepatitis C virus related liver fibrosis. *World J Hepatol* 2012; **4**: 11-17 [PMID: 22312451 DOI: 10.5754/wjg.12064]
 - 26 **American Diabetes Association**. Diagnosis and classification of diabetes mellitus. *Diabetes Care* 2010; **33** Suppl 1: S62-S69 [PMID: 20042775]
 - 27 **Davila JA**, Morgan RO, Shaib Y, McGlynn KA, El-Serag HB. Diabetes increases the risk of hepatocellular carcinoma in the United States: a population based case control study. *Gut* 2005; **54**: 533-539 [PMID: 15753540 DOI: 10.1136/gut.2004.052167]
 - 28 **Lai MS**, Hsieh MS, Chiu YH, Chen TH. Type 2 diabetes and hepatocellular carcinoma: A cohort study in high prevalence area of hepatitis virus infection. *Hepatology* 2006; **43**: 1295-1302 [PMID: 16729295 DOI: 10.1002/hep.21208]
 - 29 **El-Serag HB**, Hampel H, Javadi F. The association between diabetes and hepatocellular carcinoma: a systematic review of epidemiologic evidence. *Clin Gastroenterol Hepatol* 2006; **4**: 369-380 [PMID: 16527702 DOI: 10.1016/j.cgh.2005.12.007]
 - 30 **Hassan MM**, Hwang LY, Hatten CJ, Swaim M, Li D, Abbruzzese JL, Beasley P, Patt YZ. Risk factors for hepatocellular carcinoma: synergism of alcohol with viral hepatitis

- and diabetes mellitus. *Hepatology* 2002; **36**: 1206-1213 [PMID: 12395331 DOI: 10.1053/jhep.2002.36780]
- 31 **Liu H**, Ma Q, Li J. High glucose promotes cell proliferation and enhances GDNF and RET expression in pancreatic cancer cells. *Mol Cell Biochem* 2011; **347**: 95-101 [PMID: 20960036 DOI: 10.1007/s11010-010-0617-0]
 - 32 **Okumura M**, Yamamoto M, Sakuma H, Kojima T, Maruyama T, Jamali M, Cooper DR, Yasuda K. Leptin and high glucose stimulate cell proliferation in MCF-7 human breast cancer cells: reciprocal involvement of PKC- α and PPAR expression. *Biochim Biophys Acta* 2002; **1592**: 107-116 [PMID: 12379472 DOI: 10.1016/S0167-4889(02)00276-8]
 - 33 **Yamamoto M**, Patel NA, Taggart J, Sridhar R, Cooper DR. A shift from normal to high glucose levels stimulates cell proliferation in drug sensitive MCF-7 human breast cancer cells but not in multidrug resistant MCF-7/ADR cells which overproduce PKC- β 1. *Int J Cancer* 1999; **83**: 98-106 [PMID: 10449615]
 - 34 **Nishikawa T**, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, Yorek MA, Beebe D, Oates PJ, Hammes HP, Giardino I, Brownlee M. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* 2000; **404**: 787-790 [PMID: 10783895 DOI: 10.1038/35008121]
 - 35 **Takahashi H**, Mizuta T, Eguchi Y, Kawaguchi Y, Kuwashiro T, Oeda S, Isoda H, Oza N, Iwane S, Izumi K, Anzai K, Ozaki I, Fujimoto K. Post-challenge hyperglycemia is a significant risk factor for the development of hepatocellular carcinoma in patients with chronic hepatitis C. *J Gastroenterol* 2011; **46**: 790-798 [PMID: 21331763 DOI: 10.1007/s00535-011-0381-2]
 - 36 **Monnier L**, Mas E, Ginot C, Michel F, Villon L, Cristol JP, Colette C. Activation of oxidative stress by acute glucose fluctuations compared with sustained chronic hyperglycemia in patients with type 2 diabetes. *JAMA* 2006; **295**: 1681-1687 [PMID: 16609090 DOI: 10.1001/jama.295.14.1681]
 - 37 **Fisher WE**, Boros LG, Schirmer WJ. Insulin promotes pancreatic cancer: evidence for endocrine influence on exocrine pancreatic tumors. *J Surg Res* 1996; **63**: 310-313 [PMID: 8661216 DOI: 10.1006/jsre.1996.0266]
 - 38 **Renahan A**, Smith U, Kirkman MS. Linking diabetes and cancer: a consensus on complexity. *Lancet* 2010; **375**: 2201-2202 [PMID: 20609959 DOI: 10.1016/S0140-6736(10)60706-4]
 - 39 **Giovannucci E**, Michaud D. The role of obesity and related metabolic disturbances in cancers of the colon, prostate, and pancreas. *Gastroenterology* 2007; **132**: 2208-2225 [PMID: 17498513 DOI: 10.1053/j.gastro.2007.03.050]
 - 40 **Giovannucci E**, Harlan DM, Archer MC, Bergenstal RM, Gapstur SM, Habel LA, Pollak M, Regensteiner JG, Yee D. Diabetes and cancer: a consensus report. *Diabetes Care* 2010; **33**: 1674-1685 [PMID: 20587728 DOI: 10.3322/caac.20078]
 - 41 **Kawaguchi T**, Taniguchi E, Morita Y, Shirachi M, Tateishi I, Nagata E, Sata M. Association of exogenous insulin or sulphonylurea treatment with an increased incidence of hepatoma in patients with hepatitis C virus infection. *Liver Int* 2010; **30**: 479-486 [PMID: 20040053 DOI: 10.1111/j.1478-3231.2009.02191.x]
 - 42 **Currie CJ**, Poole CD, Gale EA. The influence of glucose-lowering therapies on cancer risk in type 2 diabetes. *Diabetologia* 2009; **52**: 1766-1777 [PMID: 19572116 DOI: 10.1007/s00125-009-1440-6]

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Gastrointestinal tuberculosis is not associated with proton pump inhibitors: A retrospective cohort study

Kyoung Sup Hong, Seung Joo Kang, Jong Kyoung Choi, Ju Han Kim, Heewon Seo, Suehyun Lee, Jae-Woo Jung, Hye-Ryun Kang, Sang-Heon Cho, Joo Sung Kim

Kyoung Sup Hong, Jae-Woo Jung, Hye-Ryun Kang, Sang-Heon Cho, Department of Internal Medicine, Drug Safety Monitoring Center, Seoul National University Hospital, Seoul National University College of Medicine, Seoul 110799, South Korea

Seung Joo Kang, Jong Kyoung Choi, Joo Sung Kim, Department of Internal Medicine, Seoul National University Hospital, Seoul National University College of Medicine, Seoul 110799, South Korea

Ju Han Kim, Heewon Seo, Suehyun Lee, Division of Biomedical Informatics, Systems Biomedical Informatics Research Center, Seoul National University College of Medicine, Seoul 110799, South Korea

Author contributions: Hong KS and Kim JS designed the study and wrote the manuscript; Jung JW, Kang HR, and Cho SH contributed to the conception and design; Kang SJ contributed to the data analysis and interpretation; Kim JH contributed to the conception and data acquisition; and Choi JK, Seo H, and Lee S contributed to the data acquisition.

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Correspondence to: Dr. Joo Sung Kim, Department of Internal Medicine, Seoul National University Hospital, Seoul National University College of Medicine, Seoul 110799, South Korea. jooskim@snu.ac.kr

Telephone: +82-2-20722228 Fax: +82-2-7629662

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Abstract

AIM: To evaluate the effect of proton pump inhibitors (PPIs) on the development of gastrointestinal tuberculosis.

METHODS: All patients who were more than 20 years old and who had received a prescription for PPIs among those who visited Seoul National University Hospital from January 1, 2005 to December 31, 2009 were

identified. Due to the low sensitivity of the microbiologic test and the nonspecific pathologic findings, the diagnosis of gastrointestinal tuberculosis was confirmed through the presence of active ulcerations and the responses to anti-tuberculosis medications. The patients were divided into two groups according to treatment duration (group 1: ≤ 3 mo; group 2: > 3 mo) and were followed up from the time they took the first prescription of PPIs until their last visit. Logistic regression analysis was used to calculate the relative risks (RR) and 95%CI, adjusting for covariates.

RESULTS: Among the 61 834 patients exposed to PPIs (50 534 in group 1; 11 300 in group 2), 21 patients were diagnosed with PPI-associated gastrointestinal tuberculosis during 124 274 person-years of follow-up. Of 21 patients, the 12 who revealed only scar changes in the colonoscopy were excluded from the statistical analyses. Of those who remained, 2 were excluded because they underwent gastrointestinal endoscopy within 4 wk of the first prescription for PPIs. Longer exposure to PPI was associated with a higher mean age (55.0 ± 14.5 in group 1 vs 58.2 ± 13.3 in group 2, $P < 0.001$) and a higher Charlson co-morbidity index (0.50 ± 0.93 in group 1 vs 0.77 ± 1.14 in group 2, $P < 0.001$). The true incidence of active gastrointestinal tuberculosis was 0.65 per 1000 person-years in group 1 and 0.03 per 1 000 person-years in group 2. Like the less-than-three-month PPI treatment period in group 1, the over-three-month PPI therapy period in group 2 was not associated with increased risk of acquiring gastrointestinal tuberculosis, after adjusting for age and co-morbidities, whereas the Charlson co-morbidity index was associated with increased risk of acquiring gastrointestinal tuberculosis based on the score [RR: (reference 1) in group 1 vs 1.518 in group 2; 95% CI: 1.040-2.216, $P = 0.03$].

CONCLUSION: Long-term PPI therapy does not seem to be associated with increased risk of acquiring gas-

gastrointestinal tuberculosis, but a higher Charlson comorbidity index is associated with such.

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Key words: Proton pump inhibitor; Acid suppression; Tuberculosis; Gastrointestinal tuberculosis; Tuberculous colitis

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INTRODUCTION

Gut flora is closely related to human health and disease^[1]. The intestinal microflora is assumed to be affected by a series of factors that determine the intraluminal environment: the pH in the gastrointestinal tract, oxygen tension, nutrient availability, colonic physiology, bacterial interference, *etc.*^[2]. One of the major factors controlling the bacterial distribution in the gastrointestinal tract is the gastric acid barrier, which may be affected by the use of inhibitors of gastric acid secretion, gastrectomy, and dietary indiscretion and stress^[3]. The establishment of enteric infection has been considered directly related to gastric acidity reduction^[4,5]. Several recent studies associated proton pump inhibitors (PPIs) with a two- to three-fold increase in the risk of *Clostridium difficile* infection^[6-8].

PPIs are currently the most powerful gastric acid suppressants and the drug of choice for the treatment of gastroesophageal reflux disease and peptic ulcer. PPIs are very powerful acid suppressants whose effects mean the percentage time intragastric pH > 4 may increase from 20% at baseline to over 60% within a week^[9]. Today's life expectancy is longer than ever before, and the number of patients who take antiplatelet agents to prevent the onset of vascular diseases is increasing. Along with this, more and more patients are obliged to take prophylactic gastric acid suppressants, including PPIs, to prevent severe complications, such as peptic ulcer bleeding.

Tuberculosis is still an important health problem in many developing countries, including South Korea^[10]. Unlike in developing countries, the disease used to be uncommon in developed countries, but it has re-emerged in the Western countries as a result of the acquired immunodeficiency syndrome (AIDS) epidemic therein as well as the influx of immigrants from developing countries^[11-13]. Gastrectomy has been known to be a potential risk factor for tuberculosis for decades^[14-16]. To these authors' knowledge, however, there has been no report about a possible association between PPI use and gastrointestinal tuberculosis. This study was thus conducted to evaluate the effect of PPIs on the development of gastrointestinal tuberculosis.

MATERIALS AND METHODS

Setting and design

This study is part of a hospital-based longitudinal cohort study entitled "Seoul National University Hospital (SNUH) PPI Safety Study," in which these authors analyzed the data regarding patients who visited SNUH and who were treated with PPI between January 1, 2005, and December 31, 2009. SNUH is a large urban tertiary care center in Seoul, South Korea. The hospital's institutional review board approved the study with a waiver of informed consent.

Data sources

Data were obtained from a clinical data warehouse fully synchronized with the electronic medical recording (EMR) system created as part of the usual care. SNUH Clinical Data Warehouse (CDW) contains all the information from each visit, not only routine clinical data such as the demographics, diagnosis, medication profiles, laboratory results, and lengths of stay of inpatients since 2001 but also the electronic charts since 2004.

Patients and case definitions

All patients who were at least 20 years old at their first visit and who ingested PPI on prescription during the five-year screening period were included in the study. According to a large retrospective study including 225 Korean patients, histological examination of the colonoscopic biopsy specimens revealed caseous necrosis in only 11.1% of the patients, and acid-fast bacilli (AFB) in 17.3% of the patients. *Mycobacterium tuberculosis* was isolated from the culture of biopsy specimens in 29.3% of the patients^[17]. Even though granulomas were observed in 72.4% of the patients, granuloma is not a specific finding in intestinal tuberculosis. Due to the low sensitivity of the microbiologic test and the nonspecific pathologic findings, the diagnosis of active gastrointestinal tuberculosis was confirmed through both an endoscopic finding of active transverse ulcerations and a good response to anti-tuberculosis treatment, which was confirmed through a follow-up colonoscopy within three months from the initiation of the treatment. The definition of good therapeutic response was complete or near-complete healing of all the ulcerations in the follow-up colonoscopy.

The patients in whom colonoscopy revealed only scar changes were excluded from the statistical analysis for two reasons: (1) the exact temporal relationship between PPI ingestion and the development of gastrointestinal tuberculosis could not be verified; and (2) the diagnosis of tuberculosis could not be confirmed as there was no need for anti-tuberculosis treatment and due to the low sensitivity of the histological evaluation of the biopsy specimens. As tuberculosis is a chronic inflammatory disease, the patients who were diagnosed with gastrointestinal tuberculosis within four weeks of their first prescription for PPIs were also excluded from the statistical analyses.

The primary exposure of interest was receipt of PPIs. PPI

Table 1 Clinical information of the patients who were endoscopically diagnosed with suspicious tuberculosis

	Gender/ age	Group ¹	PPI duration before diagnosis (d)	Reason for endoscopy	Endoscopy findings	Pathology findings	Decision for analysis	Note
1	F/65	2	639	Diarrhea	Scar, cecum	No biopsy	Excluded	
2	F/73	1	844	Abdominal pain	Scar, cecum	No biopsy	Excluded	
3	F/65	2	1177	Routine check	Scar, cecum	No biopsy	Excluded	
4	F/60	1	415	Routine check	Scar, terminal ileum	No biopsy	Excluded	Small bowel resection due to tuberculosis 20 years ago
5	F/64	1	616	Anemia	Scar, descending colon	No biopsy	Excluded	
6	F/67	2	56	Abdominal pain	Scar, cecum	No biopsy	Excluded	
7	F/55	2	87	Routine check	Scar, cecum	No biopsy	Excluded	
8	F/82	2	523	Constipation	Scar, cecum	No biopsy	Excluded	
9	F/58	1	65	Lower abdominal pain	Scar, cecum	No biopsy	Excluded	
10	F/55	1	612	Routine check	Scar, cecum	No biopsy	Excluded	
11	M/52	1	894	Blood-tinged stool	Scar, cecum	No biopsy	Excluded	
12	F/61	2	29	Bloating	Scar, cecum	No biopsy	Excluded	
13	F/72	1	125	Hematochezia	Ulcers, cecum	Chronic active colitis	Included	
14	M/44	1	28	Anemia	Ulcers, cecum/ transverse colon	Non-caseating granuloma, positive PCR ²	Included	
15	F/63	2	55	Epigastric pain	Ulcer, gastric cardia	Non-caseating granuloma, positive PCR ²	Included	Palpable right supraclavicular lymph node
16	M/49	1	7	Melena	Ulcers, terminal ileum	Chronic active ileitis	Excluded	
17	M/41	1	574	Loose stool	Ulcers, cecum	Non-caseating granuloma, positive PCR ²	Included	
18	M/20	1	49	Lower abdominal pain	Ulcers, ileocecal valve	Chronic active colitis	Included	
19	F/66	1	1	Melena	Ulcers, terminal ileum	Chronic active ileitis	Excluded	
20	F/59	1	224	Lower abdominal pain	Ulcers, ileocecal valve	Chronic active colitis,	Included	
21	F/46	1	28	Epigastric pain	Ulcers, mid-esophagus	Non-caseating granu- loma, positive PCR ²	Included	

¹Patients with PPI treatment for three months or less as group 1, and patients with more-than-three-month PPI treatment as group 2; ²Polymerase chain reaction (PCR) for *Mycobacterium tuberculosis*. PPI: Proton pump inhibitor; M: Male; F: Female.

exposure was classified by overall dosing period before the first endoscopy showing suspicious gastrointestinal tuberculosis or the last prescription date in patients without gastrointestinal tuberculosis. The patients with less than three months of PPI treatment were defined as group 1, and those with three or more months of PPI treatment were defined as group 2. To normalize the different acid-suppressive capacities among the PPI regimens, omeprazole 20 mg, lansoprazole 30 mg, rabeprazole 20 mg, pantoprazole 40 mg, and esomeprazole 40 mg were defined as standard daily doses of PPI. To calculate the adjusted dosing period, the period of the half-dose regimen was multiplied by 0.5, and the period for the double-dose regimen was multiplied by 2.

Statistical analysis

The covariates that may influence the risk of acquiring gastrointestinal tuberculosis and those that may influence the exposure to PPIs were included in the analysis. These variables were age, sex, and co-morbidities. The co-morbidities were determined from the registered diagnosis by the attending physician using the International Classification of Diseases 10th Revision code. Quan's algorithm

was used to define the 17 Charlson co-morbidities, and the Charlson index was calculated^[18,19].

Unadjusted comparisons were performed using the *t*-test, ANOVA test, Mann-Whitney test, χ^2 test, or Fisher exact test, as appropriate. Logistic regression modeling was used to estimate the relative risk of acquiring gastrointestinal tuberculosis in multivariate analyses. Statistical analysis was performed using the SAS software, version 9.2 (SAS Institute Inc., Cary, North Carolina).

RESULTS

Among the patients who visited SNUH during the five-year study period, 61 834 patients received PPIs and 7 gastrointestinal tuberculosis cases were identified. After excluding the patients who were diagnosed with suspicious gastrointestinal tuberculosis through endoscopy prior to the first prescription of PPIs, a total of 21 patients were screened. Of these, 12 patients who revealed only scar changes in colonoscopy were excluded from the statistical analyses. Of those who remained, 2 were excluded because they underwent gastrointestinal endoscopy within 4 wk of the first prescription of PPIs (Table

Table 2 Summary of descriptive statistical analysis *n* (%)

	Group 1	Group 2
Patients (<i>n</i>)	50 534	11 300
Cases	6 (0.012)	1 (0.009)
Esophageal tuberculosis	1	0
Gastric tuberculosis	0	1
Colonic tuberculosis	5	0
Total follow-up (person-mo)	1 102 947	388 345
Average follow-up (mo)	21.8	34.4
Incidence per 1000 person/yr	0.65	0.03

1). Anti-tuberculosis medications (isoniazid, rifampicin, ethambutol, and pyrazinamide) were prescribed to the finally selected 7 patients. Follow-up endoscopy was performed 2 or 3 mo after the start of the anti-tuberculosis treatment, and revealed complete healing or almost-healed ulcerations in all 7 patients.

The true incidence of gastrointestinal tuberculosis was 0.65 per 1000 person-years in group 1 and 0.03 per 1000 person-years in group 2 (Table 2). The characteristics of each group are shown in Table 3. The mean age (\pm SD) was 55.0 ± 14.5 years in group 1 and 58.2 ± 13.3 years in group 2. Longer exposure to PPI was associated with a higher Charlson co-morbidity index and a higher age (Table 3).

Table 4 presents the demographic and clinical characteristics of the active-gastrointestinal-tuberculosis and non-tuberculosis groups. Due to the small number of patients in the active-tuberculosis group, there was no significant difference between the two groups.

Table 5 shows the results of the multivariable analysis using logistic regression. A longer PPI treatment period (over three months) was not associated with increased risk of acquiring active gastrointestinal tuberculosis. The Charlson index was associated with significantly increased risk of acquiring active gastrointestinal tuberculosis by over 50% per score 1.

DISCUSSION

This cohort study was conducted not only to evaluate the possible role of more-than-three-month PPI treatment but also to calculate the incidence rate of, and to find the risk factors for, acquiring active gastrointestinal tuberculosis in all the at-least-20-year-old patients who visited SNUH and who were treated with PPI between January 1, 2005 and December 31, 2009. As a result, more-than-three-month PPI treatment was found not to be associated with increased risk of acquiring active gastrointestinal tuberculosis. The annual incidence rate of tuberculosis was reported to be 97 per 100 000 in 2010 in the South Korean general population^[10]. The calculated incidence rate of active gastrointestinal tuberculosis seems to be much lower in PPI-treated patients in the present study (Table 2), even considering the reportedly small proportion of gastrointestinal tuberculosis in the whole tuberculosis population^[20].

A diagnosis of gastrointestinal tuberculosis can be

Table 3 Demographic and clinical characteristics of each group (mean \pm SD)

	Group 1	Group 2	<i>P</i> -value
No. of patients	50 534	11 300	
Age, yr	54.96 \pm 14.50	58.18 \pm 13.31	< 0.001
Gender, male (%)	52.5	46.7	< 0.001
PPI duration, d	25.17 \pm 21.51	285.78 \pm 437.02	< 0.001
Co-morbidities, <i>n</i> (%)			
AIDS	75 (0.15)	6 (0.05)	0.011
Cerebrovascular disease	3655 (7.23)	1201 (10.63)	< 0.001
Congestive heart failure	196 (0.39)	71 (0.63)	< 0.001
Chronic pulmonary disease	184 (0.36)	64 (0.57)	0.002
Dementia	332 (0.66)	159 (1.41)	< 0.001
DM	4438 (8.78)	1600 (14.16)	< 0.001
DM without chronic complication	4280 (8.47)	1532 (13.56)	< 0.001
DM with chronic complication	158 (0.31)	68 (0.60)	< 0.001
Liver disease, mild	3182 (6.30)	1475 (13.05)	< 0.001
Hemiplegia or paraplegia	57 (0.11)	8 (0.07)	0.213
Liver disease, moderate or severe	488 (0.97)	205 (1.81)	< 0.001
Any malignancy	62 (0.12)	13 (0.12)	0.833
Metastatic solid tumor	310 (0.61)	69 (0.61)	0.972
Myocardial infarction	598 (1.18)	121 (1.07)	0.313
Peripheral vascular disease	368 (0.73)	114 (1.01)	0.002
Peptic ulcer	7011 (13.88)	1569 (13.88)	0.977
Rheumatologic disease	284 (0.56)	421 (3.73)	< 0.001
Renal disease	498 (0.99)	342 (3.03)	< 0.001
Charlson index	0.50 \pm 0.93	0.77 \pm 1.14	< 0.001
Score = 0 <i>n</i> (%)	33297 (65.89)	6167 (54.58)	< 0.001
Score = 1 <i>n</i> (%)	15536 (30.75)	4363 (38.61)	
Score \geq 3 <i>n</i> (%)	1699 (3.36)	770 (6.81)	
History of admission (%)	29138 (57.66)	6692 (59.22)	0.002
History of ICU admission (%)	1668 (3.30)	352 (3.12)	0.315

PPI: Proton pump inhibitor; AIDS: Acquired immune deficiency syndrome; ICU: Intensive care unit; DM: Diabetes mellitus.

confirmed if a characteristic caseous granuloma, positive smear of AFB, or positive culture of mycobacterium is observed in the biopsy specimen^[12]. Compared to pulmonary tuberculosis, however, there is a relatively small absolute number of AFB in gastrointestinal tuberculosis^[21], and caseous granuloma is infrequently observed in patients with an early-stage disease or who have been treated with anti-tuberculosis medications^[22]. Many studies have been performed to assess the diagnostic accuracy of such histological markers, and there have been a number of reports showing granuloma in 41%-48%^[23-25], caseous granuloma in 8%-18%^[24-26], positive smear of AFB in 0%-100%^[24,25,27,28], and positive culture of AFB in 0%-69%^[24-26,28] of gastrointestinal-tuberculosis patients. As there have been many reports showing variable results, it seems impossible to set universal standards for diagnosing gastrointestinal tuberculosis. There are adjunct diagnostic modalities, such as polymerase chain reaction (PCR) for *Mycobacterium tuberculosis*, and endoscopic findings. Although PCR is a method that shows over 50% diagnostic sensitivity, its substantial false positivity limits its role to that of an adjunctive test for the diagnosis of gastrointestinal tuberculosis^[29]. In the present study, both non-caseating granuloma and positive PCR were observed in two patients with gastroesophageal

Table 4 Demographic and clinical characteristics of the active-tuberculosis and non-tuberculosis groups (mean \pm SD)

	Active tuberculosis	Non-tuberculosis ¹
No. of patients	7	61 790
Age, yr	50.6 \pm 17.7	55.6 \pm 14.3
Gender, male (%)	3 (43)	31 803 (51)
PPI duration	47.7 \pm 97.9	72.8 \pm 213.1
Charlson index	1.29 \pm 2.36	0.55 \pm 0.97
Score = 0 <i>n</i> (%)	5 (71)	39 459 (64)
Score = 1, 2 <i>n</i> (%)	0 (0)	19 899 (32)
Score \geq 3 <i>n</i> (%)	2 (29)	2467 (4)
History of admission	6 (86)	35 824 (58)

¹Patients without gastrointestinal tuberculosis irrespective of the temporal relationship with proton pump inhibitor treatment or disease activity. PPI: Proton pump inhibitor.

tuberculosis, but non-caseating granuloma was observed in only 2 of the 7 patients with intestinal tuberculosis, and PCR was positive in 1 of the 2 patients who showed granuloma.

The endoscopic findings of gastrointestinal tuberculosis are often nonspecific^[12,30]. Intestinal tuberculosis and Crohn's disease are chronic inflammatory bowel disorders that are difficult to differentiate from each other^[31,32]. A study on colonoscopic findings reported that four parameters (involvement of fewer than 4 segments, a patulous ileocecal valve, transverse ulcers, and scars or pseudopolyps) were more frequently observed in intestinal tuberculosis patients than in Crohn's disease patients. Four parameters (anorectal lesions, longitudinal ulcers, aphthous ulcers, and cobblestone appearance) were significantly more common in Crohn's disease patients than in intestinal tuberculosis patients. A systematic analysis of the 8 parameters of colonoscopy was very useful in the differential diagnosis as it could differentiate between intestinal tuberculosis and Crohn's disease with 87.5% accuracy^[33]. In the endemic areas of tuberculosis, it seems reasonable to prescribe anti-tuberculosis medications for patients with endoscopic findings favoring intestinal tuberculosis even if there is no specific histological finding from the biopsy specimens^[12]. In the present study, anti-tuberculosis medications were also prescribed for 5 patients with only nonspecific chronic inflammations found through biopsy. Intestinal tuberculosis was confirmed in all 5 patients through follow-up colonoscopy, which revealed good therapeutic responses as well as symptom relief.

This study has a number of strengths. First, to these authors' knowledge, this study is the first study that evaluated PPI's role in the development of gastrointestinal tuberculosis. Second, all the data in this study were extracted from the SNUH CDW system that is fully synchronized with the EMR system and is optimized for research. Using the SNUH CDW system, 61 834 patients among the over one million patients who visited SNUH during the 5-year study period were rapidly screened, and the patients with gastrointestinal tuberculosis were sensitively sought out *via* browsing endoscopy and

Table 5 Results of the multivariable analysis using logistic regression

Factor	Relative risk (95%CI)	P-value
PPI exposure		
Group 1	Reference 1	
Group 2	0.697 (0.083-5.891)	0.74
Age, increase by year	0.972 (0.923-1.023)	0.27
Charlson index, increase by score 1	1.518 (1.040-2.216)	0.03
Sex	1.697 (0.374-7.690)	0.49
History of admission	4.317 (0.501-37.220)	0.18

PPI: Proton pump inhibitor.

pathology reports. Most of all, compared to cohort studies using a public database, through which only the cumulative incidence rate could be estimated, it is notable that the true incidence rate of gastrointestinal tuberculosis in PPI-treated patients was calculated. The calculated incidence rate was as low as 0.65 per 1000 person-years in group 1 and 0.03 per 1000 person-years in group 2, and anti-tuberculosis medications were shown to be effective in all the patients with active gastrointestinal tuberculosis. Therefore, the risk of acquiring gastrointestinal tuberculosis does not seem to be clinically significant in PPI-treated patients.

This study, however, has several key limitations. First, the study was performed with patients from a single urbanized tertiary care hospital. In general, a hospital cohort is vulnerable to selection bias, and the results have poor generalizability. In South Korea, a single compulsory medical insurance takes effect. Due to the open medical delivery system, all the patients are practically free to visit tertiary care hospitals. In this study, 64% of the patients fall under Charlson index 0. Therefore, their possible difference from primary care patients in the aspect of clinical severity does not seem substantial. Another limitation of a hospital cohort is its dynamic nature, which enables the members to easily join or drop out. Fortunately, even if this study was performed in PPI-treated patients during a 5-year period, the patients could be followed up for 21.8 mo in group 1 and for 34.4 mo in group 2. Therefore, the adherence of the patients in this study seems to have been good.

The second limitation is the potentially different diagnostic sensitivity. This study is an observational study. Therefore, the patients did not necessarily undergo gastrointestinal endoscopy. The implementation of endoscopy was determined according to the patient's symptoms/signs, patient's will, and doctor's decision. There might have been differences in these factors between group 1 and group 2. In this study, most of the 12 patients with only ileo-colonic scars were females and underwent colonoscopy due to mild symptoms or as a routine check. Half of them belonged to group 2, showing a relatively higher incidence in group 2 compared to 1 in the patients with active gastrointestinal tuberculosis (Table 1). Presumably, this may be due to the more frequent requests for routine check-up from or the

stricter adherence to the hospital of female patients than male patients. In the 9 patients diagnosed with active gastrointestinal tuberculosis, however, there were nearly equal numbers of male and female patients. Endoscopy was performed to assess bloody diarrhea, anemia, and severe pain, which seemed to be symptoms suggesting active gastrointestinal tuberculosis (Table 1). In this study, statistical analysis was performed in the patients with active gastrointestinal tuberculosis, which could suggest that the results of this study were not highly biased by the patients' will or adherence.

In conclusion, long-term PPI therapy does not seem to be associated with increased risk of acquiring gastrointestinal tuberculosis whereas a higher Charlson comorbidity index is associated with such. These results, however, may not exempt further monitoring due to the small case number.

COMMENTS

Background

Proton pump inhibitors (PPIs) are currently the most powerful gastric acid suppressants. Several studies have associated acid suppression with increased risk of acquiring gastrointestinal infectious diseases, such as *Clostridium difficile* infection.

Research frontiers

Gastrectomy has been known to be a potential risk factor for tuberculosis, and gastric-acid suppression may play a role in its pathogenesis. There has been no report about a possible association between PPI use and gastrointestinal tuberculosis.

Innovations and breakthroughs

Seoul National University Hospital Clinical Data Warehouse is a database that is fully synchronized with an electronic medical recording system. Using the high-quality dataset, gastrointestinal-tuberculosis patients were efficiently screened, and the true incidence and relative risk of longer PPI treatment were calculated.

Applications

Long-term PPI therapy does not seem to be associated with increased risk of acquiring gastrointestinal tuberculosis.

Peer review

The response to therapy must be evaluated very strictly as this was adopted as a diagnostic criterion.

REFERENCES

- Guarner F, Malagelada JR. Gut flora in health and disease. *Lancet* 2003; **361**: 512-519 [PMID: 12583961 DOI: S0140-6736(03)12489-0]
- Hill MJ. Diet and the human intestinal bacterial flora. *Cancer Res* 1981; **41**: 3778-3780 [PMID: 7260945]
- Husebye E. The pathogenesis of gastrointestinal bacterial overgrowth. *Chemotherapy* 2005; **51** Suppl 1: 1-22 [PMID: 15855746 DOI: 81988]
- Howden CW, Hunt RH. Relationship between gastric secretion and infection. *Gut* 1987; **28**: 96-107 [PMID: 3546004]
- Simon GL, Gorbach SL. Intestinal flora in health and disease. *Gastroenterology* 1984; **86**: 174-193 [PMID: 6357937]
- Leonard J, Marshall JK, Moayyedi P. Systematic review of the risk of enteric infection in patients taking acid suppression. *Am J Gastroenterol* 2007; **102**: 2047-256; quiz 2057 [PMID: 17509031 DOI: AJG1275]
- Dial S, Delaney JA, Schneider V, Suissa S. Proton pump inhibitor use and risk of community-acquired *Clostridium difficile*-associated disease defined by prescription for oral vancomycin therapy. *CMAJ* 2006; **175**: 745-748 [PMID: 17001054 DOI: 175/7/745]
- Dial S, Delaney JA, Barkun AN, Suissa S. Use of gastric acid-suppressive agents and the risk of community-acquired *Clostridium difficile*-associated disease. *JAMA* 2005; **294**: 2989-2995 [PMID: 16414946 DOI: 294/23/2989]
- Miner PB, Allgood LD, Grender JM. Comparison of gastric pH with omeprazole magnesium 20.6 mg (Prilosec OTC) o.m. famotidine 10 mg (Pepcid AC) b.d. and famotidine 20 mg b.d. over 14 days of treatment. *Aliment Pharmacol Ther* 2007; **25**: 103-109 [PMID: 17229225 DOI: APT3129]
- WHO global tuberculosis control report 2010. Summary. *Cent Eur J Public Health* 2010; **18**: 237 [PMID: 21361110]
- Horvath KD, Whelan RL. Intestinal tuberculosis: return of an old disease. *Am J Gastroenterol* 1998; **93**: 692-696 [PMID: 9625110 DOI: S0002927098000872]
- Marshall JB. Tuberculosis of the gastrointestinal tract and peritoneum. *Am J Gastroenterol* 1993; **88**: 989-999 [PMID: 8317433]
- Snider DE, Roper WL. The new tuberculosis. *N Engl J Med* 1992; **326**: 703-705 [PMID: 1736110 DOI: 10.1056/NEJM199203053261011]
- Snider DE. Tuberculosis and gastrectomy. *Chest* 1985; **87**: 414-415 [PMID: 3979126]
- Steiger Z, Nickel WO, Shannon GJ, Nedwicki EG, Higgins RF. Pulmonary tuberculosis after gastric resection. *Am J Surg* 1976; **131**: 668-671 [PMID: 937642]
- Allison ST. Pulmonary tuberculosis after subtotal gastrectomy. *N Engl J Med* 1955; **252**: 862-863 [PMID: 14370443 DOI: 10.1056/NEJM195505192522006]
- Lee YJ, Yang SK, Myung SJ, Byeon JS, Park IG, Kim JS, Lee GH, Jung HY, Hong WS, Kim JH, Min YI. [The usefulness of colonoscopic biopsy in the diagnosis of intestinal tuberculosis and pattern of concomitant extra-intestinal tuberculosis]. *Korean J Gastroenterol* 2004; **44**: 153-159 [PMID: 15385724 DOI: 200409153]
- Sundararajan V, Quan H, Halfon P, Fushimi K, Luthi JC, Burnand B, Ghali WA. Cross-national comparative performance of three versions of the ICD-10 Charlson index. *Med Care* 2007; **45**: 1210-1215 [PMID: 18007172 DOI: 10.1097/MLR.0b013e3181484347]
- Charlson ME, Pompei P, Ales KL, MacKenzie CR. A new method of classifying prognostic comorbidity in longitudinal studies: development and validation. *J Chronic Dis* 1987; **40**: 373-383 [PMID: 3558716]
- Al Karawi MA, Mohamed AE, Yasawy MI, Graham DY, Shariq S, Ahmed AM, al Jumah A, Ghandour Z. Protean manifestation of gastrointestinal tuberculosis: report on 130 patients. *J Clin Gastroenterol* 1995; **20**: 225-232 [PMID: 7797832]
- Kalvaria I, Kottler RE, Marks IN. The role of colonoscopy in the diagnosis of tuberculosis. *J Clin Gastroenterol* 1988; **10**: 516-523 [PMID: 3053873]
- Tandon HD, Prakash A. Pathology of intestinal tuberculosis and its distinction from Crohn's disease. *Gut* 1972; **13**: 260-269 [PMID: 5033841]
- Bhargava DK, Kushwaha AK, Dasarathy S, Shriniwas P. Endoscopic diagnosis of segmental colonic tuberculosis. *Gastrointest Endosc* 1992; **38**: 571-574 [PMID: 1397913]
- Singh V, Kumar P, Kamal J, Prakash V, Vaiphei K, Singh K. Clinicocolonoscopy profile of colonic tuberculosis. *Am J Gastroenterol* 1996; **91**: 565-568 [PMID: 8633510]
- Shah S, Thomas V, Mathan M, Chacko A, Chandy G, Ramakrishna BS, Rolston DD. Colonoscopic study of 50 patients with colonic tuberculosis. *Gut* 1992; **33**: 347-351 [PMID: 1568653]
- Bhargava DK, Tandon HD, Chawla TC, Shriniwas BN, Kapur BM. Diagnosis of ileocecal and colonic tuberculosis by colonoscopy. *Gastrointest Endosc* 1985; **31**: 68-70 [PMID: 3922847]

- 27 **Pettengell KE**, Larsen C, Garb M, Mayet FG, Simjee AE, Pirie D. Gastrointestinal tuberculosis in patients with pulmonary tuberculosis. *Q J Med* 1990; **74**: 303-308 [PMID: 2385737]
- 28 **Sakai Y**. Colonoscopic diagnosis of the intestinal tuberculosis. *Mater Med Pol* 1979; **11**: 275-278 [PMID: 548670]
- 29 **Anand BS**, Schneider FE, El-Zaatari FA, Shawar RM, Claridge JE, Graham DY. Diagnosis of intestinal tuberculosis by polymerase chain reaction on endoscopic biopsy specimens. *Am J Gastroenterol* 1994; **89**: 2248-2249 [PMID: 7977255]
- 30 **Breiter JR**, Hajar JJ. Segmental tuberculosis of the colon diagnosed by colonoscopy. *Am J Gastroenterol* 1981; **76**: 369-373 [PMID: 7325151]
- 31 **Yang XS**, Zhang L, Shi XY, Zhang YL, Lv YM. [Current diagnostic status, clinical and pathologic manifestation for a Crohn's disease]. *Beijing Da Xue Xue Bao* 2006; **38**: 407-410 [PMID: 16892148]
- 32 **Liu TH**, Pan GZ, Chen MZ. Crohn's disease. Clinicopathologic manifestations and differential diagnosis from enterocolonic tuberculosis. *Chin Med J (Engl)* 1981; **94**: 431-440 [PMID: 6796347]
- 33 **Lee YJ**, Yang SK, Byeon JS, Myung SJ, Chang HS, Hong SS, Kim KJ, Lee GH, Jung HY, Hong WS, Kim JH, Min YI, Chang SJ, Yu CS. Analysis of colonoscopic findings in the differential diagnosis between intestinal tuberculosis and Crohn's disease. *Endoscopy* 2006; **38**: 592-597 [PMID: 16673312 DOI: 10.1055/s-2006-924996]

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Clinical outcomes and predictive factors in oral corticosteroid-refractory active ulcerative colitis

Han Ho Jeon, Hyun Jung Lee, Hui Won Jang, Jin Young Yoon, Yoon Suk Jung, Soo Jung Park, Sung Pil Hong, Tae Il Kim, Won Ho Kim, Jae Hee Cheon

Han Ho Jeon, Hyun Jung Lee, Hui Won Jang, Jin Young Yoon, Yoon Suk Jung, Soo Jung Park, Sung Pil Hong, Tae Il Kim, Won Ho Kim, Jae Hee Cheon, Department of Internal Medicine, Institute of Gastroenterology, Yonsei University College of Medicine, Seoul 120-752, South Korea

Won Ho Kim, Jae Hee Cheon, Brain Korea 21 Project for Medical Science, Yonsei University College of Medicine, Seoul 120-752, South Korea

Author contributions: Jeon HH performed the data analysis and wrote the manuscript; Cheon JH had original idea on this subject, performed colonoscopy, performed the data analysis, and wrote the manuscript; Lee HJ, Jang HW, Yoon JY, Jung YS, Park SJ, Hong SP, Kim TI, and Kim WH performed colonoscopy and reviewed the manuscript critically.

Correspondence to: Jae Hee Cheon, MD, PhD, Department of Internal Medicine, Yonsei University College of Medicine, 50 Yonsei-ro, Seodaemun-gu, Seoul 120-752, South Korea. geniushee@yuhs.ac

Telephone: +82-2-22281990 Fax: +82-2-3936884

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Abstract

AIM: To evaluate the clinical outcomes and prognostic factors after intravenous corticosteroids following oral corticosteroid failure in active ulcerative colitis patients.

METHODS: Consecutive patients with moderate to severe ulcerative colitis who had been treated with a course of intravenous corticosteroids after oral corticosteroid therapy failure between January 1996 and July 2010 were recruited at Severance Hospital, Seoul, South Korea. The disease activity was measured by the Mayo score, which consists of stool frequency, rectal bleeding, mucosal appearance at flexible sigmoidoscopy, and Physician Global Assessment. We retrospectively evaluated clinical outcomes at two weeks, one month, three months, and one year after the initiation of intravenous corticosteroid therapy. Two weeks out-

comes were classified as responders or non-responders. One month, three month and one year outcomes were classified into prolonged response, steroid dependency, and refractoriness.

RESULTS: Our study included a total of 67 eligible patients. At two weeks, 56 (83.6%) patients responded to intravenous corticosteroids. At one month, complete remission was documented in 18 (32.1%) patients and partial remission in 26 (46.4%). Eleven patients (19.7%) were refractory to the treatment. At three months and one year, we found 37 (67.3%) and 25 (46.3%) patients in prolonged response, ten (18.2%) and 23 (42.6%) patients in corticosteroid dependency, 8 (14.5%) and 6 (11.1%) patients with no response, respectively. Total 9 patients were underwent elective proctocolectomy within 1 year. The duration of oral corticosteroid therapy (> 14 d $vs \leq 14$ d, $P = 0.049$) and lower hemoglobin level (≤ 11.0 mg/dL $vs > 11.0$ mg/dL, $P = 0.02$) were found to be poor prognostic factors for response at two weeks. For one year outcome, univariate analysis revealed that only a partial Mayo score (≥ 6 $vs < 6$, $P = 0.057$) was found to be associated with a poor response.

CONCLUSION: The duration of oral corticosteroid therapy and lower hemoglobin level were strongly associated with poor outcome.

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Key words: Clinical outcome; Prognosis; Corticosteroid; Ulcerative colitis

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INTRODUCTION

Ulcerative colitis (UC) is a chronic inflammatory disease of the colon with unknown etiology and is characterized by a typical natural course with recurrent flares of mucosal inflammation. About 15% of patients with UC have been reported to have an overall chance of acute exacerbation and require admission to hospital and treatment with systemic corticosteroids^[1,2]. Systemic corticosteroids remain as the gold standard treatment of acute moderate to severe UC. However, about 15%-57% of UC patients remain steroid-dependent or refractory even after steroid treatment^[3]. Before the mid-1990s, proctocolectomy was the only treatment in steroid-unresponsive UC patients. Recently, intensive drug therapies have been shown to be effective in reducing and delaying the need for colectomy^[4,5].

For UC patients who fail to improve with a maximal dosage of 5-aminosalicylic acids, oral steroid therapy should be considered. Moreover, for those who also do not respond to orally administered steroids, hospital admission is usually required for intensive intravenous treatment^[6,7]. However, to the best of our knowledge, there has been no published study describing the clinical course of patients with acute attack of UC who were treated first with oral corticosteroids and subsequently with intravenous corticosteroids due to oral steroid failure. Before demonstrating the efficacy of second line drugs, the response and the clinical outcomes of intravenous corticosteroid therapy after oral administration need to be clarified. In addition, identification of early predictors of intravenous corticosteroid responsiveness could facilitate appropriate patient selection and well-timed administration of second line therapies in such UC patients. Clinically, earlier detection of response is important to help estimate risks, benefits, and duration of treatment, as this will enable the use of alternative drugs before complications of long term use of systemic corticosteroids can develop. However, data on predictive response factors of intravenous corticosteroids following oral corticosteroid therapy are lacking.

Here we sought to identify predictive clinical or biological factors associated with intravenous corticosteroid responsiveness in UC patients that initially did not respond to oral corticosteroids, as well as two week, one month, three month, and one year clinical outcomes of such treated patients with acute attack of UC.

MATERIALS AND METHODS

Patients

The clinical records of patients with active UC who were treated with intravenous corticosteroids immediately after treatment failure of oral corticosteroids at the Yonsei University College of Medicine, Seoul, South Korea between January 1996 and July 2010 were retrospectively evaluated. The diagnosis of ulcerative colitis was based on the accepted clinical, endoscopic, and his-

topathological criteria^[8]. The criteria for eligibility were male or female patients with a diagnosis of UC followed regularly for at least 1 year. The exclusion criteria were patients with a history of corticosteroid therapy at other hospitals, corticosteroid use for diseases other than UC and a follow-up duration of less than 1 year. All enrolled patients were initiated intravenous corticosteroid therapy after admission after failure of oral steroid therapy which was done at outpatient clinic.

Intravenous corticosteroid therapy was initiated with intravenous administration of 100 mg of hydrocortisone every eight hours. Intravenous corticosteroid therapy was continued for 1-2 wk, with the treatment duration depending on the individual patient conditions, followed by gradual tapering of corticosteroids. After clinical improvement of UC, the dose of intravenous hydrocortisone was reduced to 200 mg daily. If the patients had no clinical exacerbation of UC, they were administered 30 mg/d of oral corticosteroid therapy before discharge. Our oral corticosteroid tapering policy was to reduce prednisolone by 5 or 10 mg weekly for patients with improved clinical symptoms but to sustain the current dose of prednisolone for one week for patients with lasting clinical symptoms^[9,10]. All patients who were concomitantly taking sulfasalazine (2-4 g/d) or mesalamine (1.5-4 g/d) at the time of flare up continued therapy. Patients who took immunomodulators at the time of acute flare up also maintained their initial immunomodulator therapy while taking corticosteroids.

Our study is a retrospective study from a prospectively collected database. The data are stored in a form of Assess file as well as paper form. After then, the questionnaire including Mayo or partial Mayo scores including Physician Global Assessment (PGA) is updated every visit of the patient to outpatient clinics.

The disease activity was measured by the Mayo score, which consists of stool frequency, rectal bleeding, mucosal appearance at flexible sigmoidoscopy, and PGA^[11,12]. Each component was scored 0 to 3 points, and the total score ranged from 0 to 12 points. However, a measurement of this score necessarily requires invasive flexible sigmoidoscopy, which limits repeated measurement. Therefore, most actual disease activity was measured on a nine-point partial Mayo score, which excluded the mucosal appearance at endoscopy^[13]. At the initiation of oral corticosteroid therapy, demographic data including age, age at diagnosis, duration, gender, number of acute attacks, extent of disease, concomitant medications, duration of oral corticosteroid therapy, Mayo score, and partial Mayo score were collected. Partial Mayo score and laboratory parameters were also recorded at two weeks, one month, three months, and one year after the time of initiation of intravenous corticosteroid therapy. This study was approved by the institutional board of Severance Hospital.

Definitions

Clinical outcome was measured at two weeks, one month,

three months, and one year after the initiation of intravenous corticosteroids. The classification of response to intravenous corticosteroid therapy was adopted from previous studies with minor modification^[3,11,14,15]. Patients at two weeks were classified as responders or non-responders. Non-responder of intravenous therapy after oral corticosteroid therapy was defined as persistent active disease despite administration of intravenous corticosteroids over two weeks, death due to UC attack before day 14, proctocolectomy before day 14, or secondary alternative drug use such as cyclosporine and infliximab before day 14. Proctocolectomy was performed when the patient had intractable bloody diarrhea, a continued PGA score of 3, severe anemia, persistent abdominal pain, or severe malnutrition despite intensive medical treatment. One month outcomes were classified as complete remission, partial remission, or refractoriness. Complete remission was defined as a stool frequency $\leq 2/d$ with no rectal bleeding, stool urgency, fever, or any other systemic symptoms, and a PGA score of 0. Partial remission was defined as stool frequency $\leq 4/d$ or $\leq 50\%$ of initial stool frequency with regression of other clinical symptoms and a PGA score of 1 or 2. Refractoriness was defined as persistent active status despite administration of prednisolone up to 30 mg/d or the equivalent dose over the period of four weeks^[14]. Three month outcomes were classified into prolonged response, steroid dependency, and refractoriness. Prolonged response was defined as sustained complete or partial remission during the planned dose reduction or after the completion of corticosteroids^[3]. Steroid dependency was defined as need for the same corticosteroid dose for more than two weeks despite clinical improvement, requiring an increased dose, or restarting corticosteroid therapy within two weeks because of exacerbation of symptoms^[11,14]. Refractoriness was defined as no improvement of clinical symptoms despite continued corticosteroid use. Similarly, outcomes at one year were subdivided into three groups. Prolonged response was characterized by two conditions: maintaining complete or partial remission after discontinuation of corticosteroid therapy and requiring the same dose for more than two weeks or an increasing dose of corticosteroids only in the first three months. Steroid dependency was defined as restart of corticosteroid therapy due to recurrent flare-up of UC after the first three months or being unable to reduce prednisolone to 10 mg/day within three months. Non-response was defined in the same way as for the intermediate outcomes.

Statistical analysis

Continuous variables were presented as the mean \pm SD or median (range) and were compared using two-sample *t* tests. Categorical variables were compared by χ^2 tests or Fisher's exact test. Logistic regression analysis was performed to identify predictive variables of clinical outcomes. *P* values less than 0.05 were considered statistically significant. All the statistical analyses were performed using the statistical software package SPSS 12.0 for Win-

Table 1 Baseline characteristics of patients with active ulcerative colitis receiving intravenous corticosteroids after oral corticosteroid therapy failure

	Total patients (n = 67)
Gender (male/female)	39/28 (58.2/41.8)
Age (yr)	35 (13-78)
Age at diagnosis (yr)	31 (11-78)
Disease duration (mo)	24 (0-132)
Disease extent	
Proctitis	5 (7.5)
Left-sided colitis	21 (31.3)
Extensive colitis	41 (61.2)
First attack of UC	32 (47.8)
Number of previous flares	2 (1-4)
Initial disease activity	
Full Mayo score	9 (5-10)
Partial Mayo score	6 (3-7)
Initial prednisolone dose (mg)	
≥ 30 and < 40	35 (52.2)
≥ 40	32 (47.8)
Duration of oral corticosteroid use (d)	13 (3-50)
Maintenance before flare up	
None/salicylates/azathioprine	6/47/14
Concomitant medications	
Salicylates	53
Azathioprine	14
CRP (mg/dL)	10.9 (0.1-153.0)
ESR (mm/h)	40.0 (3.0-120.0)
Hemoglobin (mg/dL)	11.7 (6.4-16.5)
Albumin (mg/dL)	3.6 (2.3-5.0)

Data are expressed as absolute numbers (percentage) or median. UC: Ulcerative colitis; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate.

dows (SPSS Inc., Chicago, IL).

RESULTS

Patient characteristics

A total of 67 patients, 39 men and 28 women, with active UC were included in this study (Table 1). The median age at diagnosis was 31 years (range: 11-78 years), and median disease duration at the time of oral corticosteroid therapy was 24 mo (range: 0-132 mo). The median length of oral corticosteroid use before intravenous therapy was 13 d (range: 3-50 d). Forty-one (61.2%) patients had extensive disease, and 32 (47.8%) had their first attack of active UC.

Clinical outcomes after intravenous corticosteroid therapy

Clinical outcomes of patients were organized in a flow chart (Figure 1). At two weeks after treatment, 56 (83.6%) patients responded to intravenous systemic corticosteroids. Seven of the 11 non-responders underwent proctocolectomy before day 14. Three patients were treated with an intravenous tumor necrosis factor- α blocker, and one patient was treated with intravenous cyclosporine. At one month after treatment, 18 (32.1%) patients were in complete remission, 26 (46.4%) in partial remission, and 12 (21.5%) had no response. Collectively, 21.5% of

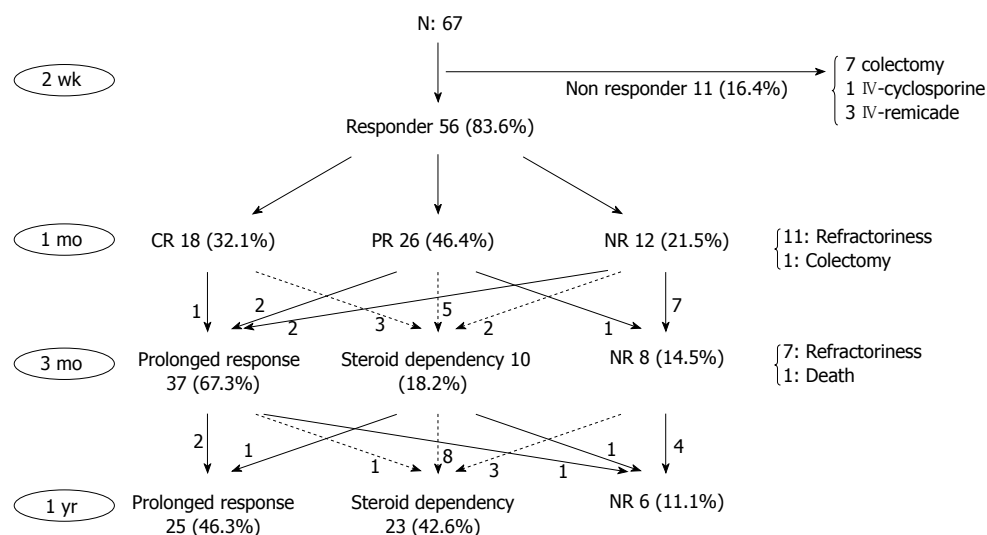


Figure 1 Clinical outcomes of patients with active ulcerative colitis treated with intravenous corticosteroids in oral corticosteroid refractory patients. CR: Complete remission; PR: Partial remission; NR: Non-response.

patients showed treatment failure to systemic corticosteroid therapy, and the other 78.5% of patients showed a partial or complete response to intravenous corticosteroid therapy. At three months, 37 patients had prolonged response, ten had steroid dependency, and seven showed no response. Of the 11 patients who were refractory to corticosteroid therapy at one month, two were in partial remission, two were steroid-dependent, seven had persistent refractoriness at three months, and one patient, a 59-year-old man who underwent proctocolectomy, died of pneumonia and sepsis. At one year, 25 of the 67 patients (37.3%) were categorized with prolonged response, 23 as steroid-dependent (34.3%), and six as non-responders (9.0%).

In our study, cytomegalovirus infection was detected in 5 patients. All were diagnosed by histologic examinations and were treated with ganciclovir. Of these, one patient underwent proctocolectomy within 14 d after the treatment. The rest of them responded to ganciclovir treatment. Finally, three patients were in partial remission and one patient was steroid dependent at one year.

Predictive factors for favorable outcomes

We performed univariate and multivariate analyses to detect clinical or laboratory factors capable of predicting poor outcomes of intravenous corticosteroid therapy after oral corticosteroid therapy failure. For the evaluation of intravenous corticosteroids at two weeks, we divided all patients into two groups: responders and non-responders who experienced death, proctocolectomy, or secondary medical treatment before two weeks. Patients were also divided into good responders and poor responders at one month, three months, and at one year after the corticosteroids therapy: good responders were those who showed a prolonged response, and poor responders were steroid-dependent or non-responsive. Multivariate analysis was carried out using the factors

that were found to be statistically significant by univariate analysis.

For two weeks outcomes, univariate analysis of predictors for non-responders showed that disease duration (> 24 mo), duration of oral corticosteroids use (> 14 days), and lower hemoglobin level (≤ 11 mg/dL) were associated with poor prognosis. According to multivariate analysis, the duration of oral corticosteroids use (> 14 d *vs* ≤ 14 d, $P = 0.049$) and lower hemoglobin level (≤ 11.0 mg/dL *vs* > 11.0 mg/dL, $P = 0.02$) remained predictive factors for non-responders (Table 2). No predictive factors for poor responders at one month or three months were identified (Tables 3 and 4). For one year outcome, univariate analysis revealed that only a partial Mayo score (≥ 6 *vs* < 6 , $P = 0.057$) was found to be associated with a poor response (Table 5).

DISCUSSION

We analyzed the clinical outcomes and identified predictive factors associated with corticosteroid responsiveness of patients who had an acute attack of UC and had been administered intravenous corticosteroids after a previous course of systemic oral corticosteroid therapy. There have been many previous studies that have reported the predictors of clinical response to systemic corticosteroid treatment in moderate to severe active UC patients^[16-19]. However, there are few reports of outcomes of a conservative approach of intravenous steroid treatment in moderate to severe active UC patients after oral corticosteroid therapy failure.

In our study, intravenous corticosteroid therapy following a failure after oral corticosteroid therapy was successful at inducing response at two weeks of treatment in 83.6% of patients. This result implies that intravenous corticosteroids could be administered when a lack of response to oral corticosteroid therapy is shown. This

Table 2 Comparison of clinical factors in immediate outcomes of the two groups of active ulcerative colitis patients receiving intravenous corticosteroids after oral corticosteroid therapy failure

	Responders (<i>n</i> = 56)	Non-responders ¹ (<i>n</i> = 11)	<i>P</i> -value		Odds ratio (95%CI)
			Univariate	Multivariate	
Gender (M/F)	32/24 (57.1/42.9)	7/4 (63.6/36.4)	0.690		
Age (yr)	37 (13-78)	39 (29-50)	0.458		
Disease duration (mo)					
≤ 24	34 (60.7)	3 (27.3)	0.041	0.123	3.38 (0.72-15.88)
> 24	22 (39.3)	8 (72.7)			
First attack of UC	26 (46.4)	6 (54.5)	0.273		
Disease extent			0.884		
Proctitis	4 (7.1)	1 (9.1)			
Left-sided colitis	17 (30.4)	4 (36.4)			
Extensive colitis	35 (62.5)	6 (54.5)			
Disease activity					
Full Mayo score (< 9/≥ 9)	27/24 (52.9/47.1)	3/8 (27.3/72.7)	0.122		
Partial Mayo score (< 6/≥ 6)	14/42 (25/75)	1/10 (9.1/90.9)	0.227		
Initial prednisolone dose (mg)			0.622		
≥ 30 and < 40	29 (51.8)	6 (54.5)			
≥ 40	27 (48.2)	5 (45.5)			
Duration of oral corticosteroid use (d)					
≤ 14	33 (58.9)	3 (27.3)	0.054	0.049	4.9 (1.01-23.81)
> 14	23 (41.1)	8 (72.7)			
Concomitant medications			0.809		
Salicylates	44 (78.6)	9 (81.8)			
Azathioprine	12 (21.4)	2 (18.2)			
CRP (mg/dL)					
≤ 8	37 (66.1)	11 (84.6)	0.303		
> 8	19 (33.9)	2 (15.4)			
ESR (mm/h)	40 (3-120)	40 (12-83)	0.999		
Hemoglobin (mg/dL)					
≤ 11	18 (32.1)	8 (72.7)	0.012	0.02	0.16 (0.03-0.75)
> 11	38 (67.9)	3 (27.3)			
Albumin (mg/dL)	3.63 (2.3-5.0)	3.56 (2.8-4.8)	0.331		

¹Non-responder group included patients that had secondary alternative drug use, proctocolectomy and death before day 14. M/F: Male/female; UC: Ulcerative colitis; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate.

result was partly explained by previous pharmacokinetic studies, which demonstrated delayed prednisolone absorption after an oral dose in patients with acute colitis as compared to healthy controls and higher plasma levels after intravenous steroid administration^[20,21]. In addition, intravenous steroid administration in the hospital ensures good treatment compliance.

The rate of failure of intravenous corticosteroid therapy in moderate to severe UC attacks was quite high, accounting for 30%-40% in recent studies investigating Western populations^[16-19,22,23]. Moreover, Meyers *et al*^[24] showed a better clinical improvement of intravenous corticotrophin therapy for acute UC in steroid-naïve patients than in patients who had already received steroid treatment. Considering that our enrolled patients were an oral corticosteroid failure group and approximately half of them were non steroid-naïve patients, our results at two weeks showed a much higher response rate to intravenous corticosteroid therapy compared with previous studies^[16-19,22,23]. Moreover, In our study, 13.4% of patients underwent proctocolectomy or experienced death within one year, while the incidence of cumulative proctocolectomy was reported to be about one-fifth that of exacerbation of UC^[25-27]. This difference in the results of steroid treat-

ment reaffirms the previous report that UC patients in the Korean population have a lower cumulative probability of proctocolectomy compared to those in Western countries^[28,29]. Taken together, these results suggest that Korean patients with UC might have a more favorable prognosis compared to Western counterparts. In order to investigate the clinical application of our results in the treatment of UC patients, further prospective studies are warranted.

One month after the initiation of intravenous corticosteroid therapy, 78.5% of patients (32.1% with complete response and 46.4% with partial response) showed clinical improvement, whereas 42.6% were dependent on steroids at one year. It has been previously reported that 22% of patients in a western study became steroid-dependent^[3]. Recently, Yoon *et al*^[10] reported that a steroid-naïve Korean UC patient showed a good response to steroid treatment, while 40% of UC patients eventually became steroid-dependent or refractory. Our study showed that the responses to intravenous corticosteroids following a failure after oral corticosteroid treatment were similarly favorable in the short term period. However, more than half of patients eventually became steroid-dependent or refractory in the long term.

Table 3 Comparison of clinical factors in one month outcomes of the two groups of active ulcerative colitis patients receiving intravenous corticosteroids after oral corticosteroid therapy failure

	Good responders ¹ (n = 44)	Poor responders ² (n = 12)	P-value univariate
Gender (M/F)	24/20 (54.5/45.5)	8/4 (66.7/33.3)	0.956
Age (yr)	37 (11-78)	38 (29-59)	0.342
Disease duration (mo)			
≤ 24	25 (56.8)	9 (75)	0.253
> 24	19 (43.2)	3 (3)	
First attack of UC	18 (40.9)	8 (66.7)	0.113
Disease extent			0.943
Proctitis	3 (6.8)	1 (8.3)	
Left-sided colitis	13 (29.5)	4 (33.3)	
Extensive colitis	28 (63.6)	7 (58.3)	
Disease activity			
Full Mayo score	23/19 (54.8/45.2)	4/5 (44.4/55.6)	0.574
($< 9/\geq 9$)			
Partial Mayo	12/32 (27.3/72.7)	2/10 (16.7/83.3)	0.452
score ($< 6/\geq 6$)			
Initial prednisolone dose (mg)			0.429
≥ 30 and < 40	24 (54.5)	5 (41.7)	
≥ 40	20 (45.5)	7 (58.3)	
Duration of oral corticosteroid use (d)			
≤ 14	26 (59.1)	7 (58.3)	0.962
> 14	18 (40.9)	5 (41.7)	
Concomitant medications			0.212
Salicylates	33 (75)	11 (91.7)	
Azathioprine	11 (25)	1 (8.3)	
CRP (mg/dL)			
≤ 8	28 (63.6)	9 (75)	0.461
> 8	16 (36.4)	3 (25)	
ESR (mm/h)	40.7 (3-120)	37.2 (12-83)	0.464
Hemoglobin (mg/dL)			
≤ 11	15 (34.1)	3 (25)	0.550
> 11	29 (65.9)	9 (75)	
Albumin (mg/dL)	3.6 (2.3-5.0)	3.45 (2.6-4.8)	0.295

¹Good responders included patients with prolonged response that maintained complete remission or partial remission after completion of corticosteroid therapy; ²Poor responders included patients with steroid dependence and non-response. M/F: Male/female; UC: Ulcerative colitis; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate.

Reports evaluating predictive factors for poor clinical outcomes have shown different and controversial results. Some found no predictive factors^[15,24,30] while others showed several factors related to clinical outcomes^[16-19,22,23]. In our study, the UC duration, the duration of oral corticosteroid use, and lower hemoglobin level were associated with failure of steroid treatment on univariate analysis. Of these variables, only duration of oral corticosteroid therapy and lower hemoglobin level were found to be independently associated with failure of corticosteroid therapy on multivariate analysis. Knowledge of the clinical factors associated with non-response to intravenous corticosteroid therapy after oral therapy may be useful in clinical decision-making. Our study was such an attempt to explain this situation. Our results showing the relationships between simple clinical factors, such as lower hemoglobin or duration of oral steroid use, and intravenous treatment outcome might help in predicting poor responders. Lower hemoglobin

Table 4 Comparison of clinical factors in three month outcomes of the two groups of active ulcerative colitis patients receiving intravenous corticosteroids after oral corticosteroid therapy failure

	Good responders ¹ (n = 37)	Poor responders ² (n = 18)	P-value univariate
Gender (M/F)	21/16 (56.8/43.2)	10/8 (55.6/44.4)	0.933
Age (yr)	35 (13-78)	41 (20-62)	0.146
Disease duration (mo)			
≤ 24	22 (59.5)	12 (66.7)	0.606
> 24	15 (40.5)	6 (33.3)	
First attack of UC	15 (40.5)	10 (55.6)	0.294
Disease extent			0.317
Proctitis	3 (8.1)	1 (5.6)	
Left-sided colitis	9 (24.3)	8 (44.4)	
Extensive colitis	25 (67.6)	9 (50)	
Disease activity			
Full Mayo score	20/14 (58.8/41.2)	7/9 (43.8/56.3)	0.318
($< 9/\geq 9$)			
Partial Mayo	11/26 (29.7/70.3)	3/15 (16.7/83.3)	0.297
score ($< 6/\geq 6$)			
Initial prednisolone dose (mg)			0.925
≥ 30 and < 40	19 (51.4)	9 (50)	
≥ 40	18 (48.6)	9 (50)	
Duration of oral corticosteroid use (d)			0.639
≤ 14	23 (62.2)	10 (55.6)	
> 14	14 (37.8)	8 (44.4)	
Concomitant medications			0.180
Salicylates	27 (73)	16 (88.9)	
Azathioprine	10 (27)	2 (11.1)	
CRP (mg/dL)			
≤ 8	24 (64.9)	12 (66.7)	0.895
> 8	13 (35.1)	6 (33.3)	
ESR (mm/h)	36.9 (3-120)	47.6 (6-120)	0.251
Hemoglobin (mg/dL)			
≤ 11	14 (37.8)	4 (22.2)	0.247
> 11	23 (62.2)	14 (77.8)	
Albumin (mg/dL)	3.73 (2.7-5.0)	3.44 (2.3-4.6)	0.082

¹Good responders included patients with prolonged response that maintained complete remission or partial remission after completion of corticosteroid therapy; ²Poor responders included patients with steroid dependence and non-response. M/F: Male/female; UC: Ulcerative colitis; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate.

level as a risk factor for poor response in our study is in accordance with an earlier reports, which reflects that initial severity of disease might be a significant predictor of poor clinical outcome after steroid treatment^[19,31,32]. Also, anemia is a common and important complication of IBD with a prevalence rate ranging from 8.8% to 66.6% in UC patients^[33,34]. The quality of life, an ability of work, and cognitive function can be impaired because of anemia in IBD patients^[35,36]. Impaired quality of life by anemia in UC patients could influence patient well-being sense and PGA. For this reason, lower hemoglobin level could be a risk factor for poor response in our study. However, none of the previous studies have evaluated duration of oral corticosteroid use before intravenous corticosteroids to predict outcome of corticosteroid therapy in patients with UC. We found that the duration of oral corticosteroid administration was an independent predictor of non-response to intravenous corticosteroid therapy on multivariate analysis in patients with acute exacerbation

Table 5 Comparison of clinical factors in one year outcomes of the two groups of active ulcerative colitis patients receiving intravenous corticosteroids after oral corticosteroid therapy failure

	Good responders ¹ (n = 25)	Poor responders ² (n = 29)	P-value univariate
Gender (M/F)	15/10 (60/40)	16/13 (55/45)	0.721
Age (yr)	35 (16-78)	39 (13-64)	0.445
Disease duration (mo)	30 (0-84)	29 (0-120)	0.325
≤ 24	14 (56)	20 (69)	
> 24	11 (44)	9 (31)	
First attack of UC	9 (36)	15 (51.7)	0.248
Disease extent			0.304
Proctitis	1 (4)	3 (10.3)	
Left-sided colitis	6 (24)	11 (37.9)	
Extensive colitis	18 (72)	15 (51.7)	
Disease activity			
Full Mayo score (< 9/≥ 9)	15/9 (62.5/37.5)	11/14 (44/56)	0.195
Partial Mayo score (< 6/≥ 6)	9/16 (36/64)	4/25 (13.8/86.2)	0.057
Initial prednisolone dose (mg)			0.785
≥ 30 and < 40	13 (52)	14 (48.3)	
≥ 40	12 (48)	15 (51.7)	
Duration of oral corticosteroid use (d)			0.337
≤ 14	17 (68)	16 (55.2)	
> 14	8 (32)	13 (44.8)	
Concomitant medications			0.771
Salicylates	19 (76)	23 (79.3)	
Azathioprine	6 (24)	6 (20.7)	
CRP (mg/dL)			
≤ 8	16 (64%)	19 (65.5)	0.907
> 8	9 (36%)	10 (34.5)	
ESR (mm/h)	34.1 (4-81)	46.3 (3-120)	0.171
Hemoglobin (mg/dL)			
≤ 11	9 (36)	8 (27.6)	0.507
> 11	16 (64)	21 (72.4)	
Albumin (mg/dL)	3.8 (2.7-4.7)	3.57 (2.3-5.0)	0.230

¹Good responders included patients with prolonged response that maintained complete remission or partial remission after corticosteroid therapy had finished; ²Poor responders included patients with steroid dependence and non-response. M/F: Male/female; UC: Ulcerative colitis; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate.

of UC. Patients with UC with more severe disease activity achieved remission less often than who tended to achieve remission earlier^[37]. The duration of oral corticosteroid therapy was likely to reflect the severity nature of UC in these non-responding patients. Additionally, prolonged oral corticosteroid therapy might be associated with mortality and morbidity such as infection and sepsis in UC patients undergoing surgery. With regard to the duration of intravenous corticosteroid treatment, the limit of 7-10 d for certifying the criteria of steroid resistance was based on historical series, which showed that the median time of remission of UC was 7.5 d and that prolonged treatment beyond 10 d did not increase the remission rate^[37]. In contrast with this point of view, a large retrospective study of single experienced hospital was in favor of more conservative approach, which entered into remission within the 21 d of treatment^[38]. Therefore it is difficult to define resistance to corticosteroid which

day after treatment is used as a limit marker. None of the previous studies have shown duration of oral corticosteroid administration was an independent predictor of non-response to intravenous corticosteroid therapy. This result demonstrates that optimal timing of intravenous corticosteroids after oral therapy was an important clinical factor of medical treatment response. In other words, use of oral corticosteroids for more than two weeks appears to be less effective in terms of clinical outcome. Additionally, long term oral corticosteroids use could lead to worse clinical outcomes and complications. An extended pre-operative use of steroids might increase the risk of surgical complications^[39,40]. UC patients undergoing an elective surgery have been shown to be at an increased risk of postoperative infectious complications in patients treated with corticosteroids^[40]. In our study, a total of 9 patients underwent elective proctocolectomy within 1 year. Among of them, one patient died of pneumonia and sepsis after proctocolectomy. Therefore, in the management of UC, the optimal timing of administration of intravenous corticosteroids after oral corticosteroid failure should be determined within two weeks after oral corticosteroid therapy. However, the optimal time limit with oral corticosteroid therapy in the face of response has not been clearly defined by randomized controlled trials. Therefore, large scaled, prospective studies are needed to confirm this result.

Yoon *et al*^[10] reported that partial Mayo score was a predictive factor of steroid dependency in steroid-naïve patients with UC. In our study, we also showed that initial higher partial Mayo score might be associated with long term poor prognosis of corticosteroid therapy. Considered overall, the partial Mayo scores could help to predict long term potential corticosteroid dependency or refractoriness in active UC patients who are treated with systemic corticosteroid therapy.

Our study on clinical outcomes and factors for response prediction after intravenous therapy after oral corticosteroids failure in active UC patient has clear clinical significance. There have been no previously published studies describing the clinical course of patients with acute attack of UC who were treated first with oral corticosteroids and subsequently intravenous corticosteroids. This study may provide valuable clinical data that can be used in the management of UC patients receiving oral corticosteroids therapy. Such studies could also be used to suggest optimal timing for administration of intravenous corticosteroids in UC patients who are treated with oral corticosteroids. Moreover, our enrolled UC patients possessed the quality of comparatively homogenous corticosteroid therapy indication and same dose of corticosteroids. These facts might be advantageous in terms of eliminating confounding factors.

There were several limitations in our study. Our study was a retrospective study in a single tertiary hospital. Mucosal healing has emerged as an important treatment goal in UC because evidence is accumulating that it can alter the clinical course of UC. However, evidence of cortico-

steroid's ability to promote mucosal healing is limited. A considerable portion of a period in this study was at moment before mucosal healing has emerged as an emerging parameter in UC. Then we could not evaluate the mucosal healing as a parameter of clinical outcomes in this study. Moreover, our study was not a placebo-controlled comparative study. Finally, the sample size was relatively small.

In conclusion, our study showed that most Korean patients with active UC responded well to intravenous corticosteroid therapy after oral corticosteroid therapy failure. However, a considerable number of patients turned out to be refractory to or dependent on this therapy. The duration of oral corticosteroid therapy and lower hemoglobin level were strongly associated with poor outcome. Further prospective studies are warranted to confirm these results and to determine the optimal timing and dose of corticosteroids.

COMMENTS

Background

Ulcerative colitis (UC) is a chronic inflammatory disease of the colon with unknown etiology and is characterized by a typical natural course with recurrent flares of mucosal inflammation. Systemic corticosteroids remain as the gold standard treatment of acute moderate to severe UC. For those who also do not respond to orally administered steroids, hospital admission is usually required for intensive intravenous treatment. However, there has been no published study describing the clinical course of patients with acute attack of UC who were treated first with oral corticosteroids and subsequently with intravenous corticosteroids due to oral steroid failure.

Research frontiers

This study showed that the duration of oral corticosteroids use (> 14 d vs ≤ 14 d, $P = 0.049$) and lower hemoglobin level (≤ 11.0 mg/dL vs > 11.0 mg/dL, $P = 0.02$) was predictive factors for non-responders. Lower hemoglobin level and duration of corticosteroids may be useful in clinical decision-making as the clinical factors associated with non-response to intravenous corticosteroid therapy after oral therapy.

Innovations and breakthroughs

These results showing the relationships between lower hemoglobin or duration of oral steroid use, and intravenous treatment outcome might help in predicting poor responders. Low hemoglobin level as a risk factor for poor response is in accordance with an earlier report which reflects that initial severity of disease might be a significant predictor of poor clinical outcome. However, none of the previous studies have evaluated duration of oral corticosteroid use before intravenous corticosteroids to predict outcome of corticosteroid therapy in patients with UC. This result demonstrates that optimal timing of intravenous corticosteroids after oral therapy was an important clinical factor of medical treatment response.

Peer review

This paper is well written and the authors highlight the limitation of the study appropriately in their discussion: Retrospective design, not strictly comparative in terms of therapies, and small numbers. Nonetheless, there is an important observation in terms of better management of patients with UC. Some of the findings have been previously reported (low hemoglobin as a risk factor for refractoriness) but it is valuable to see the principles applied to a different population. The comparative data is compelling and statistically significant using appropriate methods. Duration of oral steroid administration may be a very useful predictor of outcome in these cases. The authors are right to emphasize the need for further study in this area.

REFERENCES

- 1 Edwards FC, Truelove SC. The course and prognosis of ulcerative colitis. *Gut* 1963; **4**: 299-315 [PMID: 14084741 DOI: 10.1136/gut.4.4.299]

- 2 Abu-Suboh Abadía M, Casellas F, Vilaseca J, Malagelada JR. Response of first attack of inflammatory bowel disease requiring hospital admission to steroid therapy. *Rev Esp Enferm Dig* 2004; **96**: 539-44; 544-7 [PMID: 15449985 DOI: 10.4321/S1130-01082004000800003]
- 3 Faubion WA, Loftus EV, Harmsen WS, Zinsmeister AR, Sandborn WJ. The natural history of corticosteroid therapy for inflammatory bowel disease: a population-based study. *Gastroenterology* 2001; **121**: 255-260 [PMID: 11487534 DOI: 10.1053/gast.2001.26279]
- 4 Domènech E, Garcia-Planella E, Bernal I, Rosinach M, Cabré E, Fluvà L, Boix J, Gassull MA. Azathioprine without oral ciclosporin in the long-term maintenance of remission induced by intravenous ciclosporin in severe, steroid-refractory ulcerative colitis. *Aliment Pharmacol Ther* 2002; **16**: 2061-2065 [PMID: 12452938 DOI: 10.1046/j.1365-2036.2002.01385.x]
- 5 Kohn A, Daperno M, Armuzzi A, Cappello M, Biancone L, Orlando A, Viscido A, Annese V, Riegler G, Meucci G, Marrollo M, Sostegni R, Gasbarrini A, Peralta S, Prantera C. Infliximab in severe ulcerative colitis: short-term results of different infusion regimens and long-term follow-up. *Aliment Pharmacol Ther* 2007; **26**: 747-756 [PMID: 17697208 DOI: 10.1111/j.1365-2036.2007.03415.x]
- 6 Truelove SC, Witts LJ. Cortisone in ulcerative colitis; final report on a therapeutic trial. *Br Med J* 1955; **2**: 1041-1048 [PMID: 13260656 DOI: 10.1136/bmj.2.4947.1041]
- 7 Seo M, Okada M, Yao T, Ueki M, Arima S, Okumura M. An index of disease activity in patients with ulcerative colitis. *Am J Gastroenterol* 1992; **87**: 971-976 [PMID: 1642220]
- 8 Lennard-Jones JE. Classification of inflammatory bowel disease. *Scand J Gastroenterol Suppl* 1989; **170**: 2-6; discussion 16-19 [PMID: 2617184 DOI: 10.3109/00365528909091339]
- 9 Panaccione R, Rutgeerts P, Sandborn WJ, Feagan B, Schreiber S, Ghosh S. Review article: treatment algorithms to maximize remission and minimize corticosteroid dependence in patients with inflammatory bowel disease. *Aliment Pharmacol Ther* 2008; **28**: 674-688 [PMID: 18532990 DOI: 10.1111/j.1365-2036.2008.03753.x]
- 10 Yoon JY, Cheon JH, Park JJ, Hong SP, Kim TI, Kim WH. Clinical outcomes and factors for response prediction after the first course of corticosteroid therapy in patients with active ulcerative colitis. *J Gastroenterol Hepatol* 2011; **26**: 1114-1122 [PMID: 21299620 DOI: 10.1111/j.1440-1746.2011.06688.x]
- 11 D'Haens G, Sandborn WJ, Feagan BG, Geboes K, Hanauer SB, Irvine EJ, Lémann M, Marteau P, Rutgeerts P, Schölmerich J, Sutherland LR. A review of activity indices and efficacy end points for clinical trials of medical therapy in adults with ulcerative colitis. *Gastroenterology* 2007; **132**: 763-786 [PMID: 17258735 DOI: 10.1053/j.gastro.2006.12.038]
- 12 Schroeder KW, Tremaine WJ, Ilstrup DM. Coated oral 5-aminosalicylic acid therapy for mildly to moderately active ulcerative colitis. A randomized study. *N Engl J Med* 1987; **317**: 1625-1629 [PMID: 3317057 DOI: 10.1056/NEJM198712243172603]
- 13 Lewis JD, Chuai S, Nessel L, Lichtenstein GR, Aberra FN, Ellenberg JH. Use of the noninvasive components of the Mayo score to assess clinical response in ulcerative colitis. *Inflamm Bowel Dis* 2008; **14**: 1660-1666 [PMID: 18623174 DOI: 10.1002/ibd.20520]
- 14 Stange EF, Travis SP, Vermeire S, Reinisch W, Geboes K, Barakauskiene A, Feakins R, Fléjou JF, Herfarth H, Hommes DW, Kupcinskas L, Lakatos PL, Mantzaris GJ, Schreiber S, Villanacci V, Warren BF. European evidence-based Consensus on the diagnosis and management of ulcerative colitis: Definitions and diagnosis. *J Crohns Colitis* 2008; **2**: 1-23 [PMID: 21172194 DOI: 10.1016/j.crohns.2007.11.001]
- 15 Munkholm P, Langholz E, Davidsen M, Binder V. Frequency of glucocorticoid resistance and dependency in Crohn's disease. *Gut* 1994; **35**: 360-362 [PMID: 8150347 DOI: 10.1136/gut.35.3.360]
- 16 Ho GT, Mowat C, Goddard CJ, Fennell JM, Shah NB,

- Prescott RJ, Satsangi J. Predicting the outcome of severe ulcerative colitis: development of a novel risk score to aid early selection of patients for second-line medical therapy or surgery. *Aliment Pharmacol Ther* 2004; **19**: 1079-1087 [PMID: 15142197 DOI: 10.1111/j.1365-2036.2004.01945.x]
- 17 **Bernal I**, Mañosa M, Domènech E, Garcia-Planella E, Navarro M, Lorenzo-Zúñiga V, Cabré E, Gassull MA. Predictors of clinical response to systemic steroids in active ulcerative colitis. *Dig Dis Sci* 2006; **51**: 1434-1438 [PMID: 16868820 DOI: 10.1007/s10620-006-9103-7]
 - 18 **Kumar S**, Ghoshal UC, Aggarwal R, Saraswat VA, Choudhuri G. Severe ulcerative colitis: prospective study of parameters determining outcome. *J Gastroenterol Hepatol* 2004; **19**: 1247-1252 [PMID: 15482530 DOI: 10.1111/j.1440-1746.2004.03486.x]
 - 19 **Carbonnel F**, Gargouri D, Lémann M, Beaugerie L, Cattani S, Cosnes J, Gendre JP. Predictive factors of outcome of intensive intravenous treatment for attacks of ulcerative colitis. *Aliment Pharmacol Ther* 2000; **14**: 273-279 [PMID: 10735919 DOI: 10.1046/j.1365-2036.2000.00705.x]
 - 20 **Elliott PR**, Powell-Tuck J, Gillespie PE, Laidlaw JM, Lennard-Jones JE, English J, Chakraborty J, Marks V. Prednisolone absorption in acute colitis. *Gut* 1980; **21**: 49-51 [PMID: 7364320 DOI: 10.1136/gut.21.1.49]
 - 21 **Berghouse LM**, Elliott PR, Lennard-Jones JE, English J, Marks V. Plasma prednisolone levels during intravenous therapy in acute colitis. *Gut* 1982; **23**: 980-983 [PMID: 7129207 DOI: 10.1136/gut.23.11.980]
 - 22 **Travis SP**, Farrant JM, Ricketts C, Nolan DJ, Mortensen NM, Kettlewell MG, Jewell DP. Predicting outcome in severe ulcerative colitis. *Gut* 1996; **38**: 905-910 [PMID: 8984031 DOI: 10.1136/gut.38.6.905]
 - 23 **Benazzato L**, D'Incà R, Grigoletto F, Perissinotto E, Medici V, Angriman I, Sturniolo GC. Prognosis of severe attacks in ulcerative colitis: effect of intensive medical treatment. *Dig Liver Dis* 2004; **36**: 461-466 [PMID: 15285525 DOI: 10.1016/j.dld.2003.12.017]
 - 24 **Meyers S**, Sachar DB, Goldberg JD, Janowitz HD. Corticotropin versus hydrocortisone in the intravenous treatment of ulcerative colitis. A prospective, randomized, double-blind clinical trial. *Gastroenterology* 1983; **85**: 351-357 [PMID: 6305758]
 - 25 **Chakravarty BJ**. Predictors and the rate of medical treatment failure in ulcerative colitis. *Am J Gastroenterol* 1993; **88**: 852-855 [PMID: 8503379]
 - 26 **Mantzaris GJ**, Petraki K, Archavlis E, Amberiadis P, Kourteas D, Christidou A, Triantafyllou G. A prospective randomized controlled trial of intravenous ciprofloxacin as an adjunct to corticosteroids in acute, severe ulcerative colitis. *Scand J Gastroenterol* 2001; **36**: 971-974 [PMID: 11521989 DOI: 10.1080/003655201750305503]
 - 27 **Seo M**, Okada M, Yao T, Mataka H, Maeda K. Evaluation of the clinical course of acute attacks in patients with ulcerative colitis through the use of an activity index. *J Gastroenterol* 2002; **37**: 29-34 [PMID: 11824797 DOI: 10.1007/s535-002-8129-2]
 - 28 **Park SH**, Kim YM, Yang SK, Kim SH, Byeon JS, Myung SJ, Cho YK, Yu CS, Choi KS, Chung JW, Kim B, Choi KD, Kim JH. Clinical features and natural history of ulcerative colitis in Korea. *Inflamm Bowel Dis* 2007; **13**: 278-283 [PMID: 17206722 DOI: 10.1002/ibd.20015]
 - 29 **Kim ES**, Kim WH. Inflammatory bowel disease in Korea: epidemiological, genomic, clinical, and therapeutic characteristics. *Gut Liver* 2010; **4**: 1-14 [PMID: 20479907 DOI: 10.5009/gnl.2010.4.1.1]
 - 30 **Lee JH**, Cheon JH, Moon CM, Park JJ, Hong SP, Kim TI, Kim WH. Do patients with ulcerative colitis diagnosed at a young age have more severe disease activity than patients diagnosed when older? *Digestion* 2010; **81**: 237-243 [PMID: 20110709 DOI: 10.1159/000253850]
 - 31 **Park BJ**, Lee KJ, Hwang JC, Sin SJ, Chung JY, Cho SW. [Relapse rates of ulcerative colitis in remission and factors related to relapse]. *Korean J Gastroenterol* 2008; **52**: 21-26 [PMID: 19077487]
 - 32 **Park SM**, Han DS, Yang SK, Hong WS, Min YI. Clinical features of ulcerative colitis in Korea. *Korean J Intern Med* 1996; **11**: 9-17 [PMID: 8882472]
 - 33 **Wilson A**, Reyes E, Ofman J. Prevalence and outcomes of anemia in inflammatory bowel disease: a systematic review of the literature. *Am J Med* 2004; **116** Suppl 7A: 44S-49S [PMID: 15050885 DOI: 10.1016/j.amjmed.2003.12.011]
 - 34 **Gasche C**, Lomer MC, Cavill I, Weiss G. Iron, anaemia, and inflammatory bowel diseases. *Gut* 2004; **53**: 1190-1197 [PMID: 15247190 DOI: 10.1136/gut.2003.035758]
 - 35 **Wells CW**, Lewis S, Barton JR, Corbett S. Effects of changes in hemoglobin level on quality of life and cognitive function in inflammatory bowel disease patients. *Inflamm Bowel Dis* 2006; **12**: 123-130 [PMID: 16432377 DOI: 10.1097/01.MIB.0000196646.64615.db]
 - 36 **Gisbert JP**, Bermejo F, Pajares R, Pérez-Calle JL, Rodríguez M, Algaba A, Mancenido N, de la Morena F, Carneros JA, McNicholl AG, González-Lama Y, Maté J. Oral and intravenous iron treatment in inflammatory bowel disease: hematological response and quality of life improvement. *Inflamm Bowel Dis* 2009; **15**: 1485-1491 [PMID: 19408339 DOI: 10.1002/ibd.20925]
 - 37 **Meyers S**, Lerer PK, Feuer EJ, Johnson JW, Janowitz HD. Predicting the outcome of corticoid therapy for acute ulcerative colitis. Results of a prospective, randomized, double-blind clinical trial. *J Clin Gastroenterol* 1987; **9**: 50-54 [PMID: 3031150 DOI: 10.1097/00004836-198702000-00013]
 - 38 **Daperno M**, Sostegni R, Scaglione N, Ercole E, Rigazio C, Rocca R, Pera A. Outcome of a conservative approach in severe ulcerative colitis. *Dig Liver Dis* 2004; **36**: 21-28 [PMID: 14971812 DOI: 10.1016/j.dld.2003.04.001]
 - 39 **Faiz O**, Warusavitarne J, Bottle A, Tekkis PP, Clark SK, Darzi AW, Aylin P. Nonelective excisional colorectal surgery in English National Health Service Trusts: a study of outcomes from Hospital Episode Statistics Data between 1996 and 2007. *J Am Coll Surg* 2010; **210**: 390-401 [PMID: 20347730 DOI: 10.1016/j.jamcollsurg.2009.11.017]
 - 40 **Abera FN**, Lewis JD, Hass D, Rombeau JL, Osborne B, Lichtenstein GR. Corticosteroids and immunomodulators: post-operative infectious complication risk in inflammatory bowel disease patients. *Gastroenterology* 2003; **125**: 320-327 [PMID: 12891531 DOI: 10.1016/S0016-5085(03)00883-7]

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Dual probiotic strains suppress high fructose-induced metabolic syndrome

Do-Young Park, Young-Tae Ahn, Chul-Sung Huh, Robin A McGregor, Myung-Sook Choi

Do-Young Park, Young-Tae Ahn, Chul-Sung Huh, Korea Yakult Co., Ltd., Yongin, Gyeonggi 449-901, South Korea
Robin A McGregor, Myung-Sook Choi, Department of Food Science and Nutrition, Center for Food and Nutritional Genomics, Kyungpook National University, Daegu 702-701, South Korea
Author contributions: Park DY, Ahn YT, Huh CS and Choi MS contributed to the conception, design and acquisition of data; Park DY and McGregor RA contributed to the analysis and interpretation of data; Park DY, McGregor RA and Choi MS contributed to drafting the article or revising it critically for important intellectual content; all authors gave final approval of the version to be published.

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Correspondence to: Myung-Sook Choi, PhD, Professor, Department of Food Science and Nutrition, Center for Food and Nutritional Genomics, Kyungpook National University, 1370 Sankyuk Dong, Buk-gu, Daegu 702-701, South Korea. mschoi@knu.ac.kr

Telephone: +82-53-9506232 Fax: +82-53-9506229

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Abstract

AIM: To investigate the effect of novel probiotics on the clinical characteristics of high-fructose induced metabolic syndrome.

METHODS: Male Wistar rats aged 4 wk were fed a 70% w/w high-fructose diet ($n = 27$) or chow diet ($n = 9$) for 3 wk to induce metabolic syndrome, the rats were then randomized into groups and administered probiotic [*Lactobacillus curvatus* (*L. curvatus*) HY7601 and *Lactobacillus plantarum* (*L. plantarum*) KY1032] at 10^9 cfu/d or 10^{10} cfu/d or placebo by oral gavage for 3 wk. Food intake and body weight were measured once a week. After 6 wk, the rats were fasted for 12 h, then

anesthetized with diethyl ether and sacrificed. Blood samples were taken from the inferior vena cava for plasma analysis of glucose, insulin, C-peptide, total-cholesterol, triglycerides and thiobarbituric acid-reacting substances. Real-time polymerase chain reaction was performed using mouse-specific Taqman probe sets to assess genes related to fatty acid β -oxidation, lipogenesis and cholesterol metabolism in the liver. Target gene expression was normalized to the housekeeping gene, glyceraldehyde-3-phosphate dehydrogenase.

RESULTS: Rodents fed a high-fructose diet developed clinical characteristics of the metabolic syndrome including increased plasma glucose, insulin, triglycerides, total cholesterol and oxidative stress levels, as well as increased liver mass and liver lipids compared to chow fed controls. Probiotic treatment (*L. curvatus* HY7601 and *L. plantarum* KY1032) at high (10^{10} cfu/d) or low dosage (10^9 cfu/d) lowered plasma glucose, insulin, triglycerides and oxidative stress levels. Only high-dose probiotic treatment reduced liver mass and liver cholesterol. Probiotic treatment reduced lipogenesis *via* down-regulation of SREBP1, FAS and SCD1 mRNA levels and increased β -oxidation *via* up-regulation of PPAR α and CPT2 mRNA levels.

CONCLUSION: Probiotic *L. curvatus* HY7601 and *L. plantarum* KY1032 combined suppressed the clinical characteristics of high-fructose-induced metabolic syndrome, therefore, may provide a natural alternative for the treatment of diet-induced metabolic syndrome.

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Key words: Dyslipidemia; Fasting glucose; Gut microbiota; High-fructose diet; Inflammation; Insulin resistance; Lactobacillus; Metabolic syndrome; Oxidative stress; Probiotic

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INTRODUCTION

Metabolic syndrome is a rapidly growing worldwide pandemic, which is associated with a greater risk of multiple chronic pathologies including cardiovascular disease and Type 2 diabetes. Metabolic syndrome is characterised by a cluster of metabolic abnormalities including insulin resistance, elevated fasting glucose, elevated plasma triglycerides, elevated blood pressure, low-grade inflammation, abdominal obesity and reduced high-density lipoprotein (HDL)-cholesterol^[1,2]. There is no universal cause of metabolic syndrome, although major underlying risk factors include abdominal obesity, physical inactivity or diabetogenic diets^[1].

Diabetogenic diets with high fructose content have been strongly implicated in the development of metabolic syndrome, cardiovascular disease and Type 2 diabetes^[3]. Food manufacturers are increasingly using fructose corn syrup to increase sweetness, as well as the palatability of food and beverages. However, there is growing experimental evidence that excessive fructose intake can lead to metabolic abnormalities, including insulin resistance, dyslipidemia as well as abdominal adiposity in both animals and humans^[4]. Animals fed a high-fructose diet develop clinical characteristics of metabolic syndrome, therefore, high-fructose fed animals are particularly useful for assessing potential therapeutic interventions against metabolic syndrome^[5]. Excess dietary fructose can be converted to triglycerides through *de novo* lipogenesis, resulting in increased lipid accumulation in the liver and elevated blood lipid levels. Over a period of time prolonged high intracellular and systemic lipid levels can cause increased oxidative stress and inflammation, both of which can trigger insulin resistance, leading to increased blood glucose levels. Due to the lack of effective drugs to treat metabolic syndrome, there is growing interest in natural therapeutics to prevent or manage metabolic syndrome.

Over the past five years, probiotics have rapidly emerged as natural therapeutics with potential to target key risk factors associated with metabolic syndrome^[6,7]. Probiotics consist of single or multiple live bacterial species, which may directly or indirectly modulate gut microbial activity and improve host health. The human gut harbours between 10^{14} bacterial species collectively forming the gut microbiota^[8]. Gut microbial communities are proposed to provide the host with the ability to harvest otherwise inaccessible energy from the diet^[9-11] and modulate host genes associated with energy storage in adipose tissue^[12,12]. Probiotics have been widely assessed *in-vivo* in diet-induced obesity models^[13-17], however, different probiotic species even from the same family can exert variable effects on lipid accumulation and obesity^[18], therefore, it remains essential to assess the effectiveness

of probiotic strains in different animal disease models *in-vivo*. Probiotic yogurt containing multiple *Lactobacillus* strains has been reported to alleviate fasting blood glucose, plasma insulin and triglyceride in high-fructose fed rats^[19]. However, few other studies have considered the impact of probiotics on high-fructose diet-induced metabolic syndrome. To our knowledge, the dose-dependent and metabolic effects of probiotic treatment in high-fructose-induced metabolic syndrome remain to be established.

The aim of this study was to assess the dose-dependent effects of a probiotic consisting of *Lactobacillus curvatus* (*L. curvatus*) HY7601 and *Lactobacillus plantarum* (*L. plantarum*) KY1032 on hyperlipidemia, hyperglycemia, insulin resistance, oxidative stress and hepatic metabolism related gene expression in high-fructose fed rats with metabolic syndrome. We hypothesized that probiotic treatment may protect against dysregulated metabolism induced by a high-fructose diet in a dose-dependent manner.

MATERIALS AND METHODS

Animals, diets and experimental design

Male Wistar rats ($n = 36$) aged 4 wk were purchased from Jackson Laboratories (Bar Harbor, United States). All rats were individually housed under a constant temperature and humidity ($22 \pm 1^\circ\text{C}$, $55\% \pm 10\%$) with a 12 h light/12 h dark cycle. The experimental design consisted of a pretreatment phase (0-3 wk) and a treatment phase (3-6 wk). During the pretreatment phase, male Wistar rats were fed a 70% w/w high-fructose diet ($n = 27$) to induce metabolic abnormalities or a chow diet ($n = 9$) for 3 wk. The composition of the high-fructose diet was formulated according to Table 1. During the treatment phase, the placebo (HF) group ($n = 9$), low dose probiotic (LP) group ($n = 9$) and high dose probiotic (HP) group ($n = 9$) were fed the same high-fructose diet with placebo, 10^9 cfu probiotics or 10^{10} cfu probiotics, respectively, administered orally each day for a further 3 wk. The chow control group was fed the same chow diet with placebo administered orally each day for a further 3 wk. Freeze-dried *Lactobacillus* strains were produced by Culture Systems Inc. (United States), and packed with lactose according to Table 2. Each pack was resuspended in 500 μL distilled water prior to administration. Food intake and body weight were measured once a week. Before sacrifice, rats were fasted for 12 h and anesthetized with diethyl ether. Blood samples were taken from the inferior vena cava for plasma analysis. Epididymal adipose tissue and liver tissue were removed, rinsed with phosphate buffered saline, weighed and immediately frozen at -70°C . The experimental design was approved by the Ethics Committee of Korea Yakult Company Limited R and D Center.

Blood analysis

Plasma glucose, insulin and C-peptide concentrations

Table 1 Composition of high fructose diet

Ingredient	High fructose diet (g)
Casein	200.0
L-cystine	3.0
Fructose	700.0
Cellulose powder	50.0
Corn oil	25.0
Lard	20.0
Mineral Mix S10026	10.0
DiCalcium Phosphate	13.0
Calcium Carbonate	5.5
Potassium Citrate	16.5
Vitamin Mix V10001	10.0
Choline Bitartrate	2.0
Total	1000.0

Table 2 The composition of each supplement pack

	Placebo (mg)	10 ⁹ probiotics (mg)	10 ¹⁰ probiotics (mg)
<i>Lactobacillus curvatus</i> HY7601	-	5.0 (5 × 10 ⁸)	50.0 (5 × 10 ⁹)
<i>Lactobacillus plantarum</i> KY1032	-	2.5 (5 × 10 ⁸)	25.0 (5 × 10 ⁹)
Lactose	100.0	92.5	25.0
Total	100.0	100.0	100.0

Freeze-dried *Lactobacillus curvatus* HY7601 and *Lactobacillus plantarum* KY1032 concentration was 1 × 10⁸ and 2 × 10⁸ cfu/mg, respectively.

were determined using the glucose assay kit (Cayman, United States), insulin enzyme-linked immunosorbent assay (ELISA) kit (Millipore, United States) and rat C-peptide ELISA kit (EIAab, China), respectively, according to the manufacturer's instructions. Insulin resistance was assessed based on homeostasis model assessment of insulin resistance (HOMA-IR), calculated as the product of fasting plasma glucose (FPG) and insulin (FPI), divided by a constant^[20]. The equation was $[\text{FPG (mg/dL)} \times \text{FPI (}\mu\text{U/mL)}] / 2430$. Plasma total-cholesterol and triglyceride concentrations were determined using commercial kits (AsanPharm, South Korea). Plasma thiobarbituric acid-reacting substances (TBARS) were measured to assess oxidative stress as described previously^[21].

Hepatic lipid profile analysis

Hepatic lipids were extracted as previously reported^[22]. The dried lipid residues were dissolved in 1 mL of isopropanol for the triglyceride and cholesterol assays. Hepatic triglyceride and cholesterol concentrations were measured using the same commercial kits (AsanPharm, South Korea) used for the plasma analysis.

RT-qPCR

Total RNA was extracted from liver (15 mg) tissue using an RNAqueous kit (Ambion, United States). DNA was removed with a Turbo DNA-free kit (Ambion, United States). RNA integrity was verified and RNA quantified using a GeneQuant Pro spectrophotometer (GE Healthcare, United States). Total RNA (2 μg) was reverse-transcribed into cDNA with a high-capacity RNA-to-cDNA

Table 3 Catalog numbers of Taqman probes

Taqman probe	Catalog number
Peroxisome proliferative activated receptor α	Rn00566193_m1
Carnitine palmitoyl transferase 1	Rn00580702_m1
Carnitine palmitoyl transferase 2	Rn00563995_m1
Acyl-CoA oxidase 1	Rn01460628_m1
Sterol regulatory element binding protein-1	Rn01495769_m1
Fatty acid synthase	Rn00569117_m1
Stearoyl-CoA desaturase 1	Rn00594894_g1
Cholesterol 7 α -hydroxylase	Rn00564065_m1
Low density lipoprotein receptor	Rn00598442_m1

kit (Applied Biosystems Inc., United States). Then cDNA was amplified on a 7500 Real Time PCR System (Applied Biosystems Inc., United States) using mouse-specific Taqman probe sets (Table 3) under the following conditions: 95 °C for 10 min, followed by 40 cycles at 95 °C for 15 s, and 60 °C for 1 min. Target gene expression was normalized to the housekeeping gene, glyceraldehyde-3-phosphate dehydrogenase (catalog number Rn01775763_g1). Data was analyzed using the ABI 7500 System Sequence Detection software (Applied Biosystems Inc., United States) and presented as mean \pm SE.

Statistical analysis

All data were presented as mean \pm SE. Statistical analysis was performed using SPSS software (SPSS Inc., United States). Data were analyzed by one way analysis of variance, and the differences between experimental groups were evaluated using Duncan's multiple range test at the $P < 0.05$ level. Significant differences between chow control and HF control groups were determined using unpaired Student's t -test. Significant differences between week 3 and week 6 in each parameter were determined using the paired Student's t -test, and the values were considered statistically significant when $P < 0.05$.

RESULTS

Effects of probiotic treatment on food intake, body weight and tissue mass

Metabolic syndrome was induced in rodents by feeding them a high-fructose diet over 3 wk, while a control group was fed a chow diet. Rodents with metabolic syndrome were then randomized into three treatment groups and fed a high-fructose diet for a further 3 wk, with daily treatment of either probiotic (HP) at a dose of 1 × 10¹⁰ cfu/d, probiotic (LP) at a dose of 1 × 10⁹ cfu/d or placebo administered by oral gavage. The chow control group were fed the same diet for the same period and administered placebo by oral gavage. Average food intake was suppressed in the high-fructose diet fed rodents compared to the chow fed controls (Figure 1A). However, low or high dose probiotic treatment had no significant effect on food intake (Figure 1A). Body weight gain and epididymal fat mass were not significantly affected by high-fructose feeding or probiotic treatment regardless of dosage (Figure 1B and C). Importantly, average liver

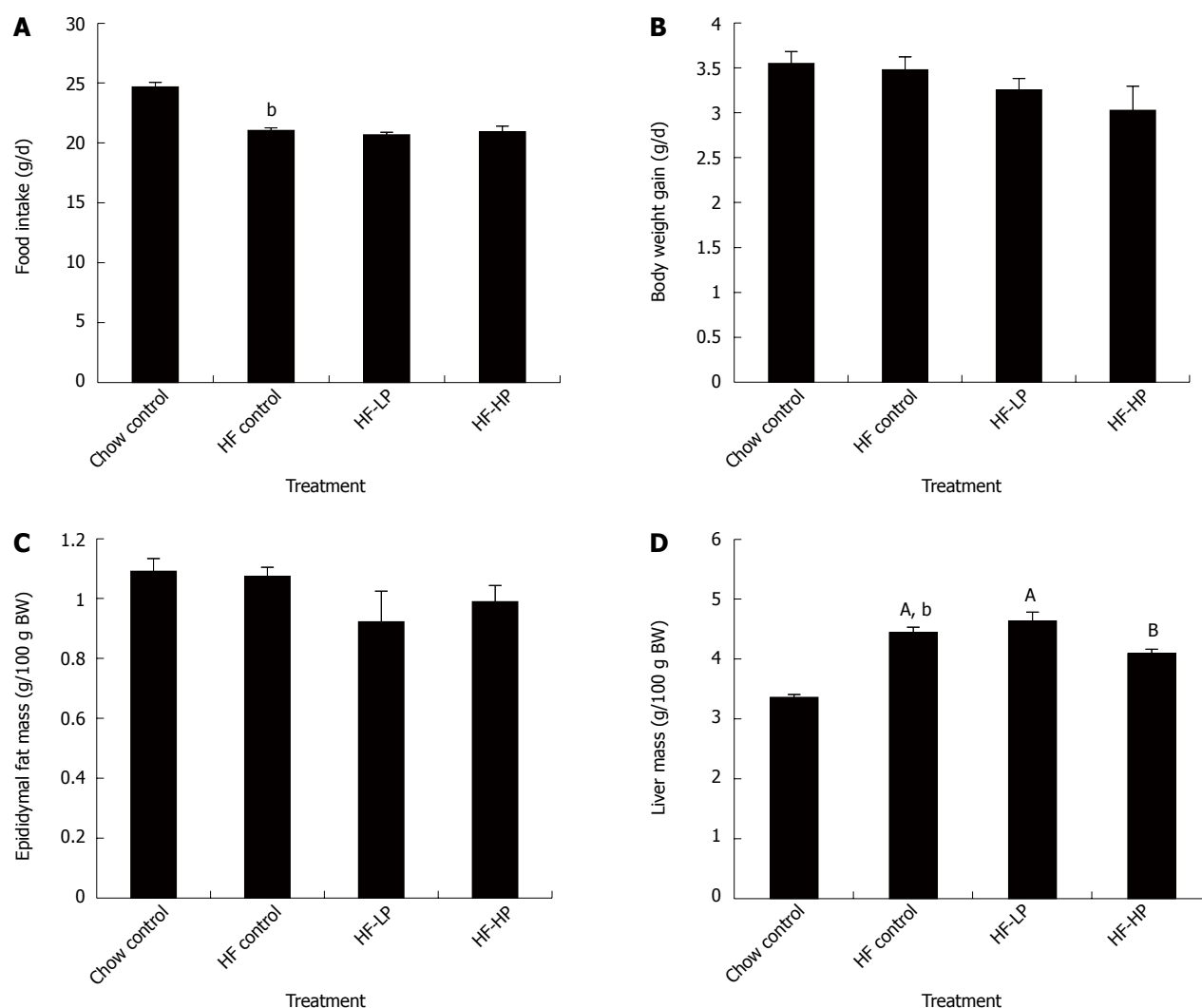


Figure 1 Effects of probiotic treatment on food intake (A), body weight gain (B), epididymal fat mass (C) and liver mass (D) in high-fructose diet-fed rats. Results are expressed as mean \pm SE. ^b $P < 0.01$ vs chow control by unpaired Student's *t*-test; ^{A,B} Bars with different capital letters are significantly different at $P < 0.05$ by Duncan's multiple range tests. HP: High dose probiotic; HF: High fructose diet; LP: Low dose probiotic; BW: Body weight.

mass, which was significantly increased by high-fructose diet, was significantly lower following HP treatment by 10% ($P < 0.01$) compared to high-fructose fed controls (Figure 1D).

Effects of probiotic treatment on dyslipidemia

Hypertriglyceridemia was effectively induced by a high-fructose diet within 3 wk (163.91 ± 6.50 mg/dL *vs* 49.26 ± 3.99 mg/dL, HF and chow group, respectively). However, subsequently 3-wk of probiotic treatment significantly reduced average plasma triglyceride levels by 46% compared to placebo treatment in the high-fructose fed rodents (Figure 2A). The probiotic treatment had no influence on either total cholesterol or HDL cholesterol levels, which were increased by high-fructose diet intake (Figure 2B and C).

Effects of probiotic treatment on hyperglycemia, insulin resistance and oxidative stress

The average plasma glucose levels of the high-fructose

fed rats were significantly higher than that of the chow fed controls at week 6, but were effectively reduced following LP and HP treatment by 24% ($P < 0.05$) and 14% ($P < 0.05$), respectively, (Figure 3A). The average plasma insulin levels of the high-fructose fed rats were significantly higher at both weeks 3 and 6 compared to the chow fed controls. However, after HP treatment plasma insulin levels were substantially lower (31%, $P < 0.05$) compared to the high-fructose fed rats. Moreover, the plasma insulin levels following LP and HP treatment were reduced by 33% ($P < 0.01$) and 29% ($P < 0.05$), respectively, compared to before treatment (Figure 3B). The HOMA-IR, a representative index of insulin resistance, was significantly higher in the high-fructose fed rats compared to the chow fed controls at week 6, but was significantly reduced by LP (35%) and HP (34%) treatment (Figure 3C). Furthermore, HOMA-IR following LP treatment was 25% ($P < 0.05$) lower compared to before treatment (Figure 3C). Plasma C-peptide is another biomarker associated with insulin resistance.

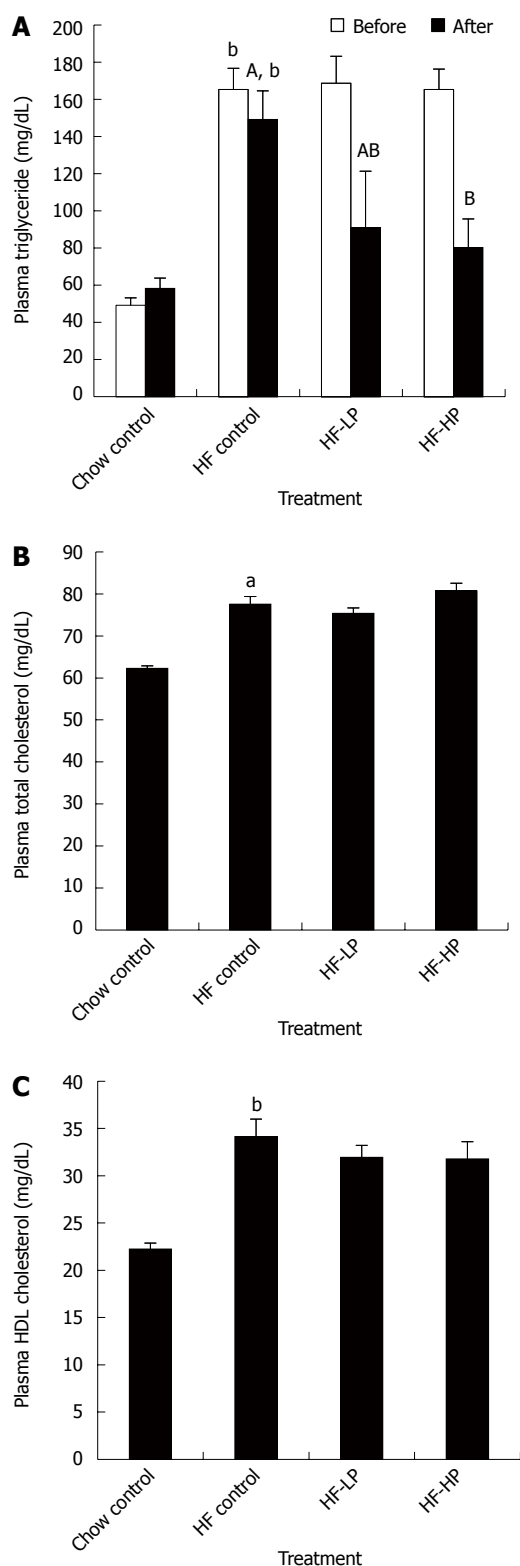


Figure 2 Effects of probiotic treatment on plasma triglyceride (A), plasma total cholesterol (B) and plasma high-density lipoprotein cholesterol (C) in high-fructose diet-fed rats. Results are expressed as mean \pm SE. ^a $P < 0.05$ and ^b $P < 0.01$ vs chow control by unpaired Student's *t*-test; ^{AB}Bars with different capital letters are significantly different at $P < 0.05$ by Duncan's multiple range tests. HF: High fructose diet; HP: High dose probiotic; LP: Low dose probiotic; HDL: High-density lipoprotein.

Plasma C-peptide level was 24% ($P < 0.05$) lower fol-

lowing HP treatment compared to placebo treatment in high-fructose fed rats (Figure 3D). Moreover, plasma C-peptide levels after HP treatment were 28% ($P < 0.05$) lower compared to before treatment (Figure 3D). In addition, probiotic treatment of either LP or HP decreased plasma TBARS levels by 37% ($P < 0.01$) and 50% ($P < 0.001$), respectively (Figure 3E).

Effects of probiotic treatment on hepatic lipid content and gene expression

Both hepatic triglyceride and cholesterol levels in the high-fructose fed rats were significantly higher than those in the chow fed controls. Probiotic treatment with either LP or HP tended to reduce hepatic triglyceride levels (Figure 4A), and HP significantly reduced (27%, $P < 0.05$) hepatic cholesterol levels compared to the high-fructose fed controls (Figure 4B).

In order to determine the mechanisms underlying the probiotic effect on hepatic lipid homeostasis, we examined the mRNA levels of hepatic genes associated with lipid metabolism (Figure 5A-C). High-fructose diet intake altered gene expression compared to chow diet intake. The changes in gene expression in the high-fructose group indicated decreases in fatty acid β -oxidation (PPAR α , CPT1, CPT2 and ACOX1), cholesterol uptake (LDLR) and bile acid synthesis (CYP7A1), and increases in fatty acid synthesis (SREBP1, FAS, SCD1). Importantly, HP treatment significantly reversed high-fructose-induced gene expression changes including up-regulation of PPAR α (+76%), CPT2 (+66%) and CYP7A1 (+71%), and down-regulation of SREBP1 (-30%), FAS (-54%) and SCD1 (-23%), although LP treatment did not cause any significant changes in hepatic lipid metabolism gene expression.

DISCUSSION

Recent studies indicate that the gut microbiota plays an important role in host lipid and glucose metabolism. Therefore, therapeutic probiotics which can manipulate the gut microbiota may also prevent some of the risk factors underlying the development of metabolic syndrome including dyslipidemia, elevated fasting glucose levels and insulin resistance^[23]. In the present study, we used a novel probiotic consisting of *L. curvatus* HY7601 and *L. plantarum* KY1302 isolated from Korean fermented cabbage. We showed that probiotic administered to high-fructose fed mice reversed the risk factors underlying the metabolic syndrome. Previous evidence from high-fructose diet-fed rat studies indicated that a probiotic-cultured yogurt called Dahi can also improve metabolic abnormalities^[19,24]. Importantly, in the present study, we established the dose-dependent effects of a novel combination of probiotic strains on metabolic syndrome.

High-fructose intake is reported to promote lipogenesis and suppress glucose intake^[25-27]. Consistent with other studies of high-fructose fed animals, hepatic gene expression analysis indicated that lipogenesis was increased *via*

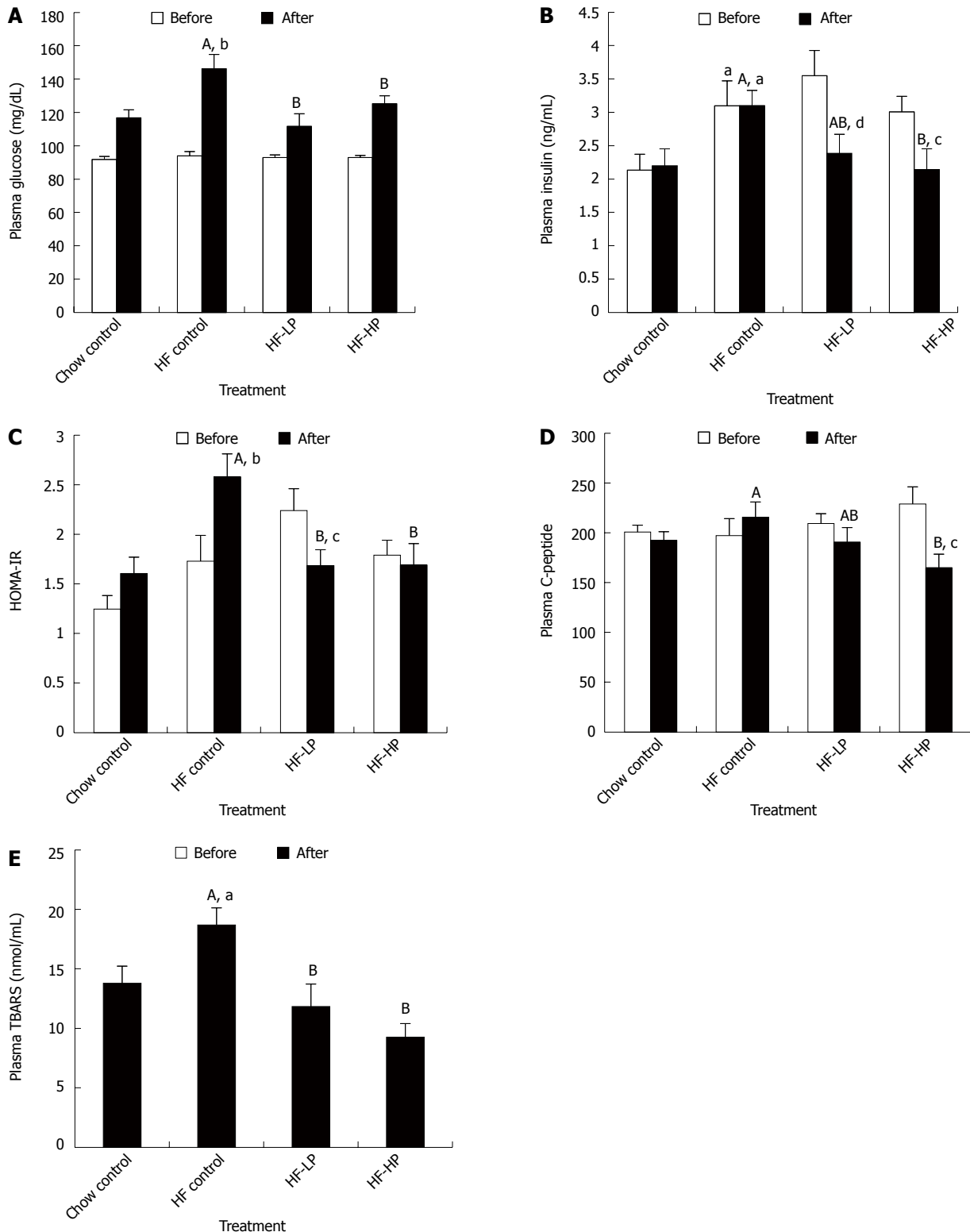


Figure 3 Effects of probiotic treatment on plasma glucose, plasma insulin, insulin resistance, plasma C-peptide and plasma thiobarbituric acid-reacting substances in high-fructose diet-fed rats. Results are expressed as mean \pm SE. ^a $P < 0.05$ and ^b $P < 0.01$ HF control vs chow control by unpaired Student's *t*-test; ^c $P < 0.05$ and ^d $P < 0.01$ week 6 vs week 3 by paired Student's *t*-test; ^{AB}Bars with different capital letters are significantly different at $P < 0.05$ by Duncan's multiple range tests. HF: High fructose diet; LP: Low dose probiotic; HP: High dose probiotic; TBARS: Thiobarbituric acid-reacting substances.

upregulation of SREBP1, FAS and SCD1, conversely β -oxidation was decreased *via* downregulation of PPAR α and PPAR α -regulated CPT1, CPT2 and ACOX1 expression^[25-27]. Increased lipogenesis and decreased β -oxidation

lead to excess accumulation of cellular lipids, as shown by the liver enlargement and hypertriglyceridemia in the high-fructose fed rats in the present study. Hypertriglyceridemia is known to be an important predictor of car-

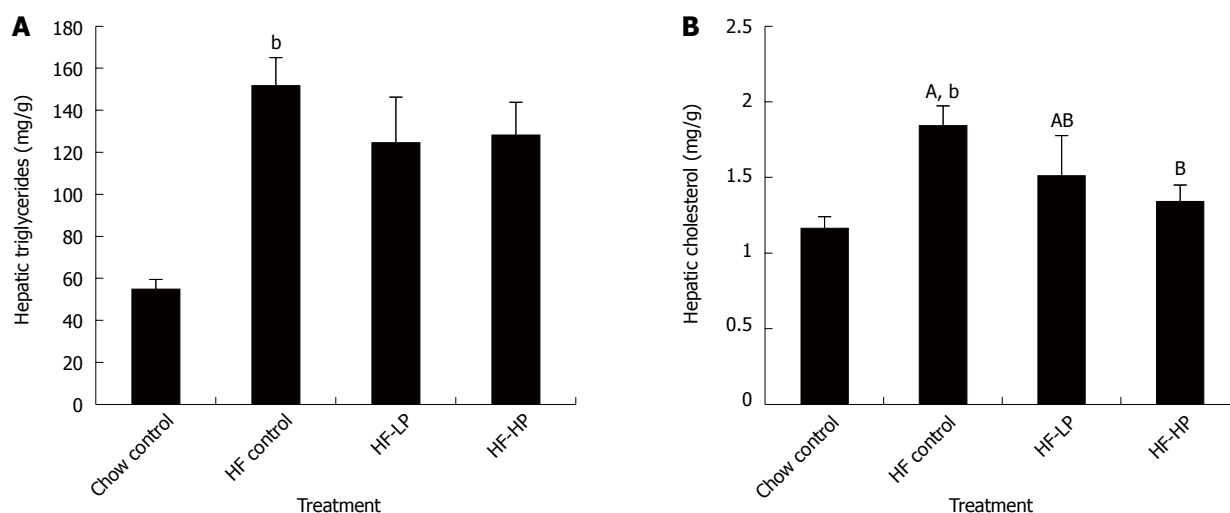


Figure 4 Effects of probiotic treatment on hepatic triglyceride and hepatic total cholesterol in high-fructose diet-fed rats. Results are expressed as mean \pm SE. ^b $P < 0.01$ HF control vs chow control by unpaired Student's *t*-test; ^{AB}Bars with different capital letters are significantly different at $P < 0.05$ by Duncan's multiple range tests. HF: High fructose diet; LP: Low dose probiotic; HP: High dose probiotic.

diovascular disease mortality in subjects with diabetes or impaired glucose tolerance^[28], therefore, reducing plasma triglyceride levels may improve long-term health.

Previous studies have shown that probiotics can alter the gut microbiota^[29], and direct gut microbiota manipulations in germ-free mice significantly affect host lipid metabolism^[2,12]. Here we showed that 10^{10} cfu/d probiotic treatment led to upregulated PPAR α and CPT2 expression reflecting activation of β -oxidation, and down-regulated SREBP1, FAS and SCD1 expression reflecting suppression of lipogenesis. Moreover, these probiotics induced transcriptional changes which resulted in a significant reduction in liver mass and plasma triglyceride levels. These findings are consistent with a previous report which showed that *L. plantarum* KY1032 inhibits lipid droplet accumulation during adipocyte differentiation. In contrast to plasma triglyceride levels, hepatic triglyceride levels were only slightly reduced by probiotic treatment. We hypothesized that the probiotic-induced increase in hepatic β -oxidation related gene expression was partly to clear excess hepatic triglycerides generated through high-fructose-induced *de novo* lipogenesis.

Contrary to hepatic triglyceride levels, cholesterol levels were significantly reduced by probiotic treatment, whereas plasma cholesterol levels were unchanged. We assessed whether the reduction in hepatic cholesterol following probiotic treatment was due to altered hepatic cholesterol metabolism related gene expression. High-fructose diet suppressed CYP7A1 expression, which encodes cholesterol 7 α -hydroxylase, the rate-limiting enzyme involved in the formation of bile from cholesterol. However, 10^{10} cfu/d probiotic treatment upregulated CYP7A1 expression indicating increased bile synthesis activity. Some probiotics are also reported to increase bile salt hydrolase activity^[30]. Hepatic LDLR expression remained unchanged, suggesting that probiotic treatment did not enhance uptake of plasma cholesterol into the liver, which is consistent with the absence of changes in

plasma cholesterol levels. However, the effect of probiotics on liver cholesterol metabolism may be partly dependent on the diet regime used, because in high-fat or high-cholesterol diet fed animals, probiotics are reported to decrease plasma and hepatic cholesterol levels together.

The probiotic effect on host lipid levels may also contribute to the observed improvement in glucose homeostasis. Excess intracellular lipids can inhibit insulin signalling, hence reduce insulin-stimulated glucose uptake leading to hyperglycemia and hyperinsulinemia as shown in this study. We observed that probiotic treatment at 10^9 or 10^{10} cfu/d significantly lowered plasma glucose, insulin and C-peptide concentrations, and reduced insulin resistance indicated by the reduced HOMA-IR index^[20,31]. Excess fructose intake is also reported to promote lipid peroxidation and oxidative stress is implicated in the pathogenesis of insulin resistance^[32,33]. In the present study, probiotic treatment at 10^9 or 10^{10} cfu/d significantly reversed the oxidative stress present in the high-fructose fed rats with metabolic syndrome, indicated by lower plasma TBARS levels. Some *Lactobacillus* strains are also reported to possess anti-oxidative activity. For example, probiotics have been reported to reduce exercise-induced oxidative stress, *via* increases in anti-oxidative activity, which helps neutralize reactive oxygen species^[34]. Furthermore, studies in human Type 2 diabetes show probiotic supplementation can increase superoxide dismutase and glutathione peroxidase activities^[35], which are anti-oxidants that help protect against oxidative stress.

While evidence is growing on the effects of probiotic treatment on many chronic pathologies, some issues remain to be addressed. Probiotics are widely assumed to modulate the gut microbiota, and exert health benefits *via* direct modification of gut microbial communities^[8]. Studies on diet-induced mice indicate that probiotics alter the gut microbiota^[29], similar to our recent experience with the same probiotic used in this study (unpublished observations). However, in another study using germ-

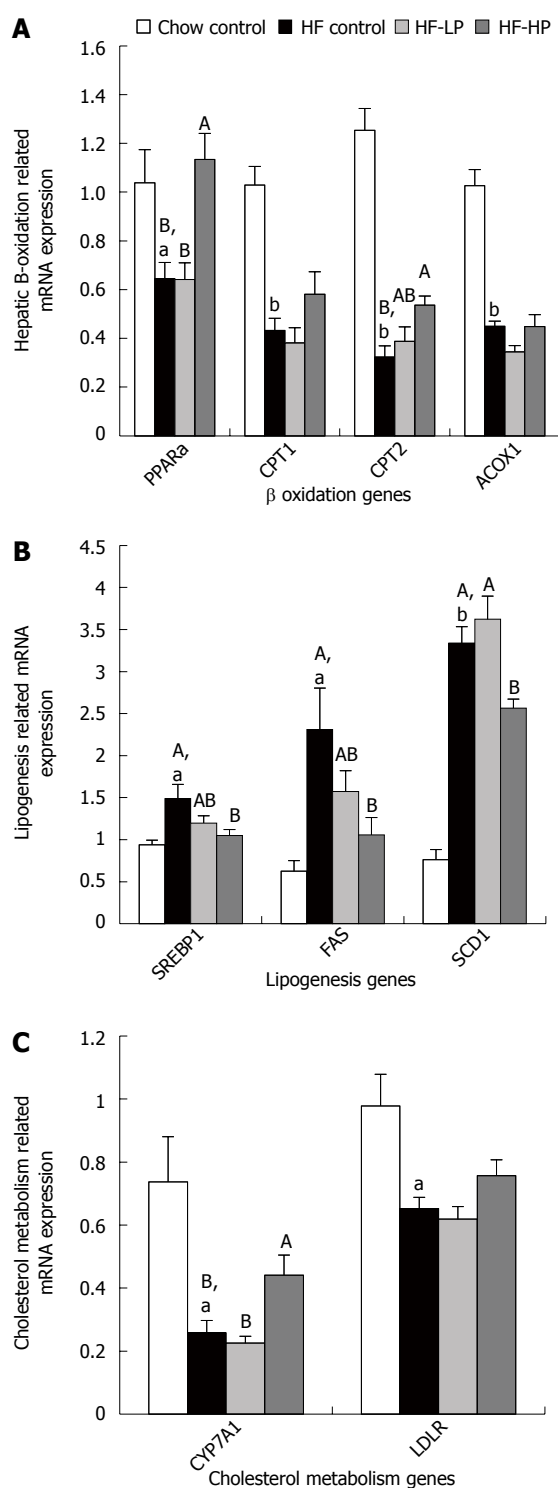


Figure 5 Effects of probiotic treatment on hepatic B-oxidation, lipogenesis and cholesterol metabolism related gene expression in high-fructose diet-fed rats. Results are expressed as mean \pm SE. ^a $P < 0.05$ and ^b $P < 0.01$ HF control vs chow control by unpaired Student's *t*-test; ^{AB}Bars with different capital letters are significantly different at $P < 0.05$ by Duncan's multiple range tests. HF: High fructose diet; LP: Low dose probiotic; HP: High dose probiotic; FAS: Fatty acid synthase; SREBP1: Sterol regulatory element-binding protein-1; PPARα: Peroxisome proliferator-activated receptor α; CPT: Carnitine palmitoyl-transferase; ACOX: Acyl-coenzyme A oxidase; SCD: Stearoyl-CoA desaturase; CYP7A1: Cholesterol 7α-hydroxylase gene; LDLR: Low-density lipoprotein receptor.

free mice transplanted with a small artificial gut microbial

community, a multi-species probiotic failed to change the gut microbiota^[36]. Whether multi-species probiotics are more effective than single-species probiotics is not clear, because few studies make this comparison. We used a two species probiotic in the present study, and based on our preliminary data the results suggest that combined species are more effective than either species alone. Finally, whether probiotics exert dose-dependent effects against metabolic syndrome has not been evaluated, nor has the minimum amount of live bacteria that is necessary for functional health benefits. Evidence from the present study indicates that probiotics containing 10^{10} cfu/d showed the greatest effectiveness against high-fructose-induced metabolic syndrome over a relatively short period, without any adverse effects. In contrast, probiotic treatment at 10^9 cfu/d exerted minimal effects on hypertriglyceridemia, but effectively improved hyperglycaemia and insulin resistance. In the case of hepatic gene expression, 10^9 cfu/d probiotic treatment had no modulatory effect on any of the genes tested, but probiotic treatment at 10^{10} cfu/d modulated lipogenesis and β-oxidation related genes.

In conclusion, probiotic *L. curvatus* HY7601 and *L. plantarum* KY1032 combined can suppress the clinical characteristics of high-fructose-induced metabolic syndrome, therefore, may provide a natural alternative for the treatment of diet-induced metabolic syndrome.

ACKNOWLEDGMENTS

Choi MS and McGregor RA declare no conflicts of interest. Park DY, Ahn YT and Huh CS are current or past employees of Korea Yakult Co., Ltd.

COMMENTS

Background

Metabolic syndrome is a growing health problem characterised by elevated blood sugar and blood lipid levels, insulin resistance, high blood pressure and abdominal obesity. Poor diet consisting of high sugar or high fat content is a major risk factor for metabolic syndrome. There is no universal treatment for metabolic syndrome. Probiotics found in fermented foods have emerged as a natural way of protecting against metabolic syndrome, but many probiotic strains have varying metabolic effects. Therefore, a major challenge for scientists is discovering probiotic strains, which may help protect against metabolic syndrome.

Research frontiers

Probiotic strains identified in kimchi, a traditional Korean fermented cabbage consumed regularly in South Asian countries, are reported to have various beneficial properties. Current research aims to determine whether these probiotic strains are effective in different animal models of disease including metabolic syndrome, Type 2 diabetes and obesity.

Innovations and breakthroughs

The authors showed that two probiotic strains, *Lactobacillus curvatus* (*L. curvatus*) HY7601 and *Lactobacillus plantarum* (*L. plantarum*) KY1032, can suppress metabolic abnormalities such as hypertriglyceridemia, hyperglycemia and insulin resistance in high-fructose-induced metabolic syndrome. These probiotic health benefits were associated with decreased lipogenesis and increased β-oxidation-related gene expression in the liver.

Applications

Probiotic with *L. curvatus* HY7601 and *L. plantarum* KY1032 may provide a natural supplement for the management of the underlying risk factors of metabolic syndrome.

Terminology

Probiotics consist of live micro-organisms which confer beneficial effects on host health. Hypertriglyceridemia is prolonged elevated triglyceride levels in blood. Hyperglycemia is prolonged elevated glucose levels in the blood. Insulin resistance is the inability of insulin to stimulate glucose uptake pathways in fat, skeletal muscle and the liver.

Peer review

This paper investigated the effect of novel probiotics on the clinical characteristics of high-fructose induced metabolic syndrome. The authors were suggesting a mechanism. One novel thing of this paper is that no one has looked at these specific microbes in this combination. It is well written.

REFERENCES

- 1 Grundy SM, Cleeman JI, Daniels SR, Donato KA, Eckel RH, Franklin BA, Gordon DJ, Krauss RM, Savage PJ, Smith SC, Spertus JA. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute scientific statement: Executive Summary. *Crit Pathw Cardiol* 2005; **4**: 198-203 [PMID: 18340209]
- 2 Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF, Gordon JI. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004; **101**: 15718-15723 [PMID: 15505215]
- 3 Malik VS, Popkin BM, Bray GA, Després JP, Hu FB. Sugar-sweetened beverages, obesity, type 2 diabetes mellitus, and cardiovascular disease risk. *Circulation* 2010; **121**: 1356-1364 [PMID: 20308626]
- 4 Stanhope KL, Schwarz JM, Keim NL, Griffen SC, Bremer AA, Graham JL, Hatcher B, Cox CL, Dyachenko A, Zhang W, McGahan JP, Seibert A, Krauss RM, Chiu S, Schaefer EJ, Ai M, Otokozawa S, Nakajima K, Nakano T, Beysen C, Hellerstein MK, Berglund L, Havel PJ. Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans. *J Clin Invest* 2009; **119**: 1322-1334 [PMID: 19381015]
- 5 de Moura RF, Ribeiro C, de Oliveira JA, Stevanato E, de Mello MA. Metabolic syndrome signs in Wistar rats submitted to different high-fructose ingestion protocols. *Br J Nutr* 2009; **101**: 1178-1184 [PMID: 19007450]
- 6 Everard A, Lazarevic V, Derrien M, Girard M, Muccioli GG, Neyrinck AM, Possemiers S, Van Holle A, François P, de Vos WM, Delzenne NM, Schrenzel J, Cani PD. Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *Diabetes* 2011; **60**: 2775-2786 [PMID: 21933985]
- 7 Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, Waget A, Delmée E, Cousin B, Sulpice T, Chamontin B, Ferrières J, Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* 2007; **56**: 1761-1772 [PMID: 17456850]
- 8 Sekirov I, Russell SL, Antunes LC, Finlay BB. Gut microbiota in health and disease. *Physiol Rev* 2010; **90**: 859-904 [PMID: 20664075]
- 9 Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, Egholm M, Henrissat B, Heath AC, Knight R, Gordon JI. A core gut microbiome in obese and lean twins. *Nature* 2009; **457**: 480-484 [PMID: 19043404]
- 10 Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; **444**: 1027-1031 [PMID: 17183312]
- 11 Bird AR, Brown IL, Topping DL. Starches, resistant starches, the gut microflora and human health. *Curr Issues Intest Microbiol* 2000; **1**: 25-37 [PMID: 11709851]
- 12 Bäckhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci USA* 2007; **104**: 979-984 [PMID: 17210919]
- 13 Lee K, Paek K, Lee HY, Park JH, Lee Y. Antiobesity effect of trans-10,cis-12-conjugated linoleic acid-producing *Lactobacillus plantarum* PL62 on diet-induced obese mice. *J Appl Microbiol* 2007; **103**: 1140-1146 [PMID: 17897219 DOI: 10.1111/j.1365-2672.2007.03336.x]
- 14 Lee HY, Park JH, Seok SH, Baek MW, Kim DJ, Lee KE, Paek KS, Lee Y, Park JH. Human originated bacteria, *Lactobacillus rhamnosus* PL60, produce conjugated linoleic acid and show anti-obesity effects in diet-induced obese mice. *Biochim Biophys Acta* 2006; **1761**: 736-744 [PMID: 16807088 DOI: 10.1016/j.bbali.2006.05.007]
- 15 Kondo S, Xiao JZ, Satoh T, Odamaki T, Takahashi S, Sugahara H, Yaeshima T, Iwatsuki K, Kamei A, Abe K. Antiobesity effects of *Bifidobacterium breve* strain B-3 supplementation in a mouse model with high-fat diet-induced obesity. *Biosci Biotechnol Biochem* 2010; **74**: 1656-1661 [PMID: 20699581]
- 16 Kang JH, Yun SI, Park HO. Effects of *Lactobacillus gasseri* BNR17 on body weight and adipose tissue mass in diet-induced overweight rats. *J Microbiol* 2010; **48**: 712-714 [PMID: 21046354 DOI: 10.1007/s12275-010-0363-8]
- 17 Chen JJ, Wang R, Li XF, Wang RL. *Bifidobacterium longum* supplementation improved high-fat-fed-induced metabolic syndrome and promoted intestinal Reg I gene expression. *Exp Biol Med (Maywood)* 2011; **236**: 823-831 [PMID: 21685239 DOI: 10.1258/ebm.2011.010399]
- 18 Yin YN, Yu QF, Fu N, Liu XW, Lu FG. Effects of four *Bifidobacteria* on obesity in high-fat diet induced rats. *World J Gastroenterol* 2010; **16**: 3394-3401 [PMID: 20632441]
- 19 Yadav H, Jain S, Sinha PR. Antidiabetic effect of probiotic dahi containing *Lactobacillus acidophilus* and *Lactobacillus casei* in high fructose fed rats. *Nutrition* 2007; **23**: 62-68 [PMID: 17084593]
- 20 Cacho J, Sevillano J, de Castro J, Herrera E, Ramos MP. Validation of simple indexes to assess insulin sensitivity during pregnancy in Wistar and Sprague-Dawley rats. *Am J Physiol Endocrinol Metab* 2008; **295**: E1269-E1276 [PMID: 18796548]
- 21 Do GM, Oh HY, Kwon EY, Cho YY, Shin SK, Park HJ, Jeon SM, Kim E, Hur CG, Park TS, Sung MK, McGregor RA, Choi MS. Long-term adaptation of global transcription and metabolism in the liver of high-fat diet-fed C57BL/6J mice. *Mol Nutr Food Res* 2011; **55** Suppl 2: S173-S185 [PMID: 21618427 DOI: 10.1002/mnfr.201100064]
- 22 Park DY, Ahn YT, Huh CS, Jeon SM, Choi MS. The inhibitory effect of *Lactobacillus plantarum* KY1032 cell extract on the adipogenesis of 3T3-L1 Cells. *J Med Food* 2011; **14**: 670-675 [PMID: 21554138 DOI: 10.1089/jmf.2010.1355]
- 23 Kootte RS, Vrieze A, Holleman F, Dallinga-Thie GM, Zoetendal EG, de Vos WM, Groen AK, Hoekstra JB, Stoes ES, Nieuwdorp M. The therapeutic potential of manipulating gut microbiota in obesity and type 2 diabetes mellitus. *Diabetes Obes Metab* 2012; **14**: 112-120 [PMID: 21812894]
- 24 Yadav H, Jain S, Sinha PR. Effect of skim milk and dahi (yogurt) on blood glucose, insulin, and lipid profile in rats fed with high fructose diet. *J Med Food* 2006; **9**: 328-335 [PMID: 17004894]
- 25 Basciano H, Federico L, Adeli K. Fructose, insulin resistance, and metabolic dyslipidemia. *Nutr Metab (Lond)* 2005; **2**: 5 [PMID: 15723702]
- 26 Miyazaki M, Dobrzyn A, Man WC, Chu K, Sampath H, Kim HJ, Ntambi JM. Stearoyl-CoA desaturase 1 gene expression is necessary for fructose-mediated induction of lipogenic gene expression by sterol regulatory element-binding protein-1c-dependent and -independent mechanisms. *J Biol Chem* 2004; **279**: 25164-25171 [PMID: 15066988]
- 27 Nagai Y, Nishio Y, Nakamura T, Maegawa H, Kikkawa R, Kashiwagi A. Amelioration of high fructose-induced meta-

- bolic derangements by activation of PPAR α . *Am J Physiol Endocrinol Metab* 2002; **282**: E1180-E1190 [PMID: 11934685]
- 28 **Fontbonne A**, Eschwège E, Cambien F, Richard JL, Ducimetière P, Thibault N, Warnet JM, Claude JR, Rosselin GE. Hypertriglyceridaemia as a risk factor of coronary heart disease mortality in subjects with impaired glucose tolerance or diabetes. Results from the 11-year follow-up of the Paris Prospective Study. *Diabetologia* 1989; **32**: 300-304 [PMID: 2666216]
- 29 **Murphy EF**, Cotter PD, Hogan A, O'Sullivan O, Joyce A, Fouhy F, Clarke SF, Marques TM, O'Toole PW, Stanton C, Quigley EM, Daly C, Ross PR, O'Doherty RM, Shanahan F. Divergent metabolic outcomes arising from targeted manipulation of the gut microbiota in diet-induced obesity. *Gut* 2012; Epub ahead of print [PMID: 22345653 DOI: 10.1136/gutjnl-2011-300705]
- 30 **Begley M**, Hill C, Gahan CG. Bile salt hydrolase activity in probiotics. *Appl Environ Microbiol* 2006; **72**: 1729-1738 [PMID: 16517616]
- 31 **Wu T**, Giovannucci E, Pischon T, Hankinson SE, Ma J, Rifai N, Rimm EB. Fructose, glycemic load, and quantity and quality of carbohydrate in relation to plasma C-peptide concentrations in US women. *Am J Clin Nutr* 2004; **80**: 1043-1049 [PMID: 15447918]
- 32 **Kelley GL**, Allan G, Azhar S. High dietary fructose induces a hepatic stress response resulting in cholesterol and lipid dysregulation. *Endocrinology* 2004; **145**: 548-555 [PMID: 14576175]
- 33 **Busserolles J**, Rock E, Gueux E, Mazur A, Grolier P, Rays-siguier Y. Short-term consumption of a high-sucrose diet has a pro-oxidant effect in rats. *Br J Nutr* 2002; **87**: 337-342 [PMID: 12064343]
- 34 **Martarelli D**, Verdenelli MC, Scuri S, Cocchioni M, Silvi S, Cecchini C, Pompei P. Effect of a probiotic intake on oxidant and antioxidant parameters in plasma of athletes during intense exercise training. *Curr Microbiol* 2011; **62**: 1689-1696 [PMID: 21400082]
- 35 **Ejtahed HS**, Mohtadi-Nia J, Homayouni-Rad A, Niafar M, Asghari-Jafarabadi M, Mofid V. Probiotic yogurt improves antioxidant status in type 2 diabetic patients. *Nutrition* 2012; **28**: 539-543 [PMID: 22129852]
- 36 **McNulty NP**, Yatsunenko T, Hsiao A, Faith JJ, Muegge BD, Goodman AL, Henrissat B, Oozeer R, Cools-Portier S, Gobert G, Chervaux C, Knights D, Lozupone CA, Knight R, Duncan AE, Bain JR, Muehlbauer MJ, Newgard CB, Heath AC, Gordon JI. The impact of a consortium of fermented milk strains on the gut microbiome of gnotobiotic mice and monozygotic twins. *Sci Transl Med* 2011; **3**: 106ra106 [PMID: 22030749]

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Clinical significance and risk factors of postembolization fever in patients with hepatocellular carcinoma

Chung Hwan Jun, Ho Seok Ki, Hoon Ki Lee, Kang Jin Park, Seon Young Park, Sung Bum Cho, Chang Hwan Park, Young Eun Joo, Hyun Soo Kim, Sung Kyu Choi, Jong Sun Rew

Chung Hwan Jun, Ho Seok Ki, Hoon Ki Lee, Kang Jin Park, Seon Young Park, Sung Bum Cho, Chang Hwan Park, Young Eun Joo, Hyun Soo Kim, Sung Kyu Choi, Jong Sun Rew, Division of Gastroenterology, Department of Internal Medicine, Chonnam National University Medical School, Gwangju 501-757, South Korea

Author contributions: Jun CH and Choi SK performed the majority of the study and wrote the manuscript; Cho SB, Park SY, and Park CH were involved in editing the manuscript; Ki HS, Lee GH, Park KJ were involved in data acquisition, analysis or interpretation; all authors were involved in revising and approving the final version for publication.

Correspondence to: Sung Kyu Choi, MD, Division of Gastroenterology, Department of Internal Medicine, Chonnam National University Medical School, 42 Jaebong-ro, Dong-Ku, Gwangju 501-757, South Korea. choisk@jnu.ac.kr

Telephone: +82-62-2206296 Fax: +82-62-2258578

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Abstract

AIM: To investigate tumor response and survival in patients with postembolization fever (PEF) and to determine the risk factors for PEF.

METHODS: Four hundred forty-three hepatocellular carcinoma (HCC) patients who underwent the first session of transcatheter arterial chemoembolization (TACE) between January 2005 and December 2009 were analyzed retrospectively. PEF was defined as a body temperature greater than 38.0 °C that developed within 3 d of TACE without evidence of infection. The tumor progression-free interval was defined as the interval from the first TACE to the second TACE based on mRECIST criteria. Clinical staging was based on the American Joint Committee on Cancer tumor, node, metastases (TNM) classification of malignant tumors. All patients were admitted before their 1st TACE treatment,

and blood samples were obtained from all patients before and after treatment. Clinicoradiological variables and host-related variables were compared between two groups: patients with PEF vs patients without PEF. Additionally, variables related to 20-mo mortality and tumor progression-free survival were analyzed.

RESULTS: The study population comprised 370 (85.4%) men and 73 (14.6%) women with a mean age of 62.29 ± 10.35 years. A total of 1836 TACE sessions were conducted in 443 patients, and each patient received between 1 and 27 (mean: 4.14 ± 3.57) TACE sessions. The mean follow-up duration was 22.23 ± 19.6 mo (range: 0-81 mo). PEF developed in 117 patients (26.41%) at the time of the first TACE session. PEF was not associated with 20-mo survival ($P = 0.524$) or computed tomography (CT) response ($P = 0.413$) in a univariate analysis. A univariate analysis further indicated that diffuse-type HCC ($P = 0.021$), large tumor size (≥ 5 cm) ($P = 0.046$), lipiodol dose (≥ 7 mL, $P = 0.001$), poor blood glucose control ($P = 0.034$), alanine aminotransferase (ALT) value after TACE ($P = 0.004$) and C-reactive protein (CRP) value after TACE ($P = 0.036$) served as possible risk factors correlated with PEF. The ALT value after TACE ($P = 0.021$) and lipiodol dose over 7 mL ($P = 0.011$) were independent risk factors for PEF in the multivariate analysis. For the 20-mo survival, poor blood sugar control ($P < 0.001$), portal vein thrombosis ($P = 0.001$), favorable CT response after TACE ($P < 0.001$), initial aspartate aminotransferase ($P = 0.02$), initial CRP ($P = 0.042$), tumor size ($P < 0.001$), TNM stage ($P < 0.001$) and lipiodol dose ($P < 0.001$) were possible risk factors in the univariate analysis. Tumor size ($P = 0.03$), poor blood sugar control ($P = 0.043$), and portal vein thrombosis ($P = 0.031$) were significant predictors of survival in the multivariate analysis. Furthermore, the tumor progression-free interval was closely associated with CRP > 1 mg/dL ($P = 0.003$), tumor size > 5 cm ($P < 0.001$), tumor type (poorly defined) ($P < 0.001$), and lipiodol dose (> 7 mL, $P < 0.001$).

CONCLUSION: PEF has no impact on survival at 20 mo or radiologic response. However, the ALT level after TACE and the lipiodol dose represent significant risk factors for PEF.

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Key words: Chemoembolization; Therapeutic; Fever; Carcinoma; Hepatocellular; Prognosis; Progression-free survival

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INTRODUCTION

Hepatocellular carcinoma (HCC) is the seventh most common carcinoma worldwide and the third most common cause of cancer-related mortality^[1]. In South Korea, the age-standardized incidence rate of HCC is 46.5 per 100 000 individuals^[2]. Recent advances in treatment, including liver transplantation, surgical resection, percutaneous ethanol injection therapy, radiofrequency ablation and transcatheter arterial chemoembolization (TACE) have improved the prognosis for patients with HCC^[3,4]. In addition, beneficial therapeutic options that may affect long-term cure include surgical resection, liver transplantation, and percutaneous ablation^[5]. However, these curative therapies are feasible only for a small subset of patients with HCC^[6]. Among the noncurative therapies, only chemoembolization, the most widely used treatment for unresectable HCC, has demonstrated a positive effect on survival^[7]. In contrast, TACE can be employed for any type of HCC irrespective of tumor size, location, or number^[8].

However, TACE inevitably results in a hypoxic insult to the HCC and the surrounding liver tissue^[9], and postembolization syndrome is common^[10]. Postembolization syndrome, which consists of temporary fever, ileus, and abdominal pain, is the most common side effect of chemoembolization, affecting 60% to 80% of patients with HCC^[5,11]. Postembolization fever after TACE is the most significant adverse effect, and it frequently affects the duration of hospitalization and causes the needless administration of antibiotics, although the fever is self-limited in most cases. However, few data about postembolization fever have been reported. Therefore, we evaluated the risk factors and prognostic significance of postembolization fever in patients with HCC.

MATERIALS AND METHODS

Patients and methods

Four hundred forty-three HCC patients who underwent

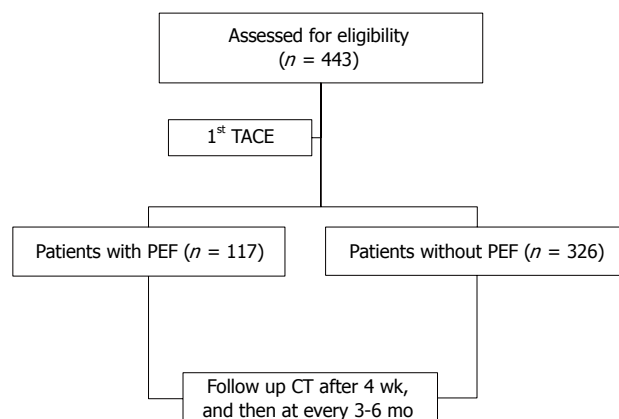


Figure 1 Flowchart of study patient enrollment. PEF: Postembolization fever; TACE: Transcatheter arterial chemoembolization; CT: Computed tomography.

the first session of TACE between January 2005 and December 2009 were analyzed retrospectively. The diagnosis of HCC was confirmed histologically or based on consistent findings obtained from at least two imaging techniques: ultrasonography, computed tomography (CT), magnetic resonance imaging, and/or selective hepatic arterial angiography^[12,13]. Clinical staging was determined based on the American Joint Committee on Cancer tumor, node, metastases (TNM) classification of malignant tumors^[14].

Clinicoradiological variables were compared between two groups (patients with PEF *vs* patients without PEF). The host-related variables included age, sex, viral status, cause of HCC, Child-Pugh score, Eastern Cooperative Oncology Group (ECOG) performance status, white blood cell counts, aspartate aminotransferase (AST), alanine aminotransferase (ALT), α -fetoprotein (AFP), and 20-mo mortality. The tumor-related variables included maximal tumor size, number of tumors, TNM stage, radiological findings (poorly defined or well defined), portal vein thrombosis, and CT response after 1st TACE.

All patients were admitted before their 1st TACE, and blood samples were obtained from all patients before and after treatment. Serum AFP, CRP, blood chemistry and ECOG score at admission were measured. After the 1st TACE, the patients were carefully followed. Dynamic CT was performed after 4 wk and then every 3 to 6 mo (Figure 1).

Our institutional review board did not require approval because the procedures were performed for clinical reasons. Informed consent was obtained from all patients after the nature and purpose of the TACE procedure had been fully explained.

Chemoembolization procedure

An arterial catheter was inserted into the femoral artery using the Seldinger method and placed in the hepatic artery. Tumor-feeding vessels were superselected whenever possible, and a solution containing 10 to 40 mg of doxorubicin hydrochloride (ADM; Dong-A Pharmacy, Seoul, Korea) and 0 to 40 mL of iodized oil (Lipiodol; Guer-

Table 1 Comparison of transcatheter arterial chemoembolization sessions with or without postembolization fever

Variables	Patients with PEF (<i>n</i> = 117)	Patients without PEF (<i>n</i> = 326)	<i>P</i> -value
Sex (M/F)	94/23	276/50	0.309
Age (yr)	62.26 ± 10.79	62.16 ± 9.98	0.971
Cause of HCC (HBV/ HCV/alcohol/other)	66/25/14/11	180/44/73/25	0.088
Tumor type (well defined/ poorly defined)	57/44	202/90	0.021
Portal vein thrombosis (yes/no)	9/93	39/253	0.292
Poor blood glucose control (yes/no)	29/88	116/210	0.039
Favorable tumor response	61 (52.5%)	155 (48.5%)	0.516
20-mo mortality	60 (51.7%)	172 (53.4%)	0.828
Initial WBC/mm ³	5969 ± 3497	5547 ± 2535	0.194
Initial AST U/L	68.87 ± 45.49	64.11 ± 44.79	0.358
Initial ALT U/L	50.84 ± 41.58	43.89 ± 33.16	0.129
Initial CRP mg/dL	1.23 ± 1.66	0.86 ± 0.93	0.116
Initial AFP IU/mL	2778 ± 8014	2600 ± 8656	0.857
Child-Pugh score	5.66 ± 0.72	5.77 ± 0.97	0.296
TNM stage	1.95 ± 1.05	1.83 ± 1.07	0.305
Interval from 1st TACE to 2 nd TACE (d)	157 ± 225	180 ± 270	0.412
Total TACE sessions	1.48 ± 3.45	4.02 ± 3.62	0.230

TACE: Transcatheter arterial chemoembolization; AFP: α -fetoprotein; PEF: Postembolization fever; TNM: Tumor, node, metastases; CRP: C-reactive protein; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; HCC: Hepatocellular carcinoma; M: Male; F: Female; HBV: Hepatitis B virus; HCV: Hepatitis C virus.

bet, Aulnay-sous-Bois, France) with absorbable gelatin particles (Gelfoam; Upjohn, Kalamazoo, Michigan) was injected through the catheter (5F) or microcatheter (2.8 or 3F). The doses of doxorubicin and iodized oil were individually determined according to tumor size, tumor extent, and the patient's underlying liver function.

Monitoring and management of postembolization fever

For the purpose of this study, we defined postembolization fever as a body temperature greater than 38.0 °C during the 3 d after TACE. Body temperature was measured *qid* by nurses using an axillary thermometer. Bacterial cultures from blood and urine and chest X-rays were performed for patients who had fevers after TACE to detect any potential infectious agents. Empirical broad-spectrum intravenous antibiotics were used to treat potential infections if there was fever but were discontinued when bacterial cultures did not reveal any causative agent and the fever had subsided. Ultrasonography or CT scans were performed if the fever persisted despite the use of antibiotics to detect the possible formation of an abscess. Acetaminophen or nonsteroidal anti-inflammatory drugs were used for symptom control, if necessary.

Response assessment

The efficacy of TACE was evaluated by comparing the CT scans obtained before and after chemoembolization in terms of iodized oil uptake patterns in the tumor that could be considered necrotic^[12] and tumor extent. The

iodized oil uptake was considered compact if the oily contrast medium was clearly dispersed through all viable target tumors but was noncompact in all other cases^[8]. Tumor response to TACE was defined as a compact uptake of iodized oil or at least a 30% decrease in the sum of the largest diameters of viable tumors, despite noncompact iodized oil uptake.

Definitions

PEF was defined as a body temperature greater than 38.0 °C that developed within 3 d of TACE without evidence of infection. Poor blood glucose control was defined as a mean blood glucose level > 200 mg/dL. Poorly defined tumor type was defined as diffuse-type HCC, whereas well-defined tumor type was defined as nodular HCC. Tumor progression-free survival was defined as the interval during and after treatment in which a patient remained alive and the disease did not worsen (in this case, the interval from the 1st TACE to the 2nd TACE).

Statistical analysis

Comparisons were performed using the student's *t* test for continuous variables and Pearson's χ^2 test. Factors that were significant in the univariate analysis were entered into a stepwise multivariate analysis to identify the most significant risk factors. The hazard function data were estimated using the Kaplan-Meier curve and compared using the log-rank test. Multivariate analyses were performed using the Cox proportional hazards model to identify prognostic factors. We performed statistical analyses using SPSS 17.0 (SPSS Inc., Chicago, United States). A *P*-value less than 0.05 was considered statistically significant.

RESULTS

Fever after TACE and clinical features

The study population consisted of 370 (85.4%) men and 73 (14.6%) women with a mean age of 62.29 ± 10.35 years. A total 1836 sessions of TACE were conducted in 443 patients between January 2005 and December 2009. Each patient received between 1 and 27 (mean, 4.14 ± 3.57) sessions of TACE. The mean follow-up duration was 22.23 ± 19.6 mo (range: 0–81 mo).

One hundred seventeen episodes of postembolization fever (26.41%) occurred in 443 HCC patients after the 1st TACE session. Most of the postembolization fever episodes peaked within the first two days after TACE. The post-TACE fever was usually self-limiting, with durations ranging from 1 to 10 d (mean: 1.72 ± 1.11 d). The infectious complication rate was 0.16% (3/1836 cases). Two cases of bacteremia and one case of liver abscess developed after 1836 TACE sessions. A comparison of the TACE sessions with and without fever is presented in Table 1.

Association of PEF with clinical variables

A univariate analysis indicated that diffuse-type HCC (*P* < 0.05), large tumor size (≥ 5 cm) (*P* < 0.05), lipiodol

Table 2 Multivariate and univariate analysis for postembolization fever, 20-mo mortality, and tumor progression-free survival

Variables	HR	95%CI	P-value
Multivariate analysis ¹			
ALT value after TACE	1.002	1.00-1.005	< 0.05
Lipiodol dose (< 7 mL)	0.539	0.329-0.881	< 0.05
Univariate analysis ²			
Poor BS control	2.673	1.77-4.034	< 0.01
Portal vein thrombosis	3.048	1.536-6.06	< 0.01
Poor CT response	2.638	1.785-3.891	< 0.01
Initial AST	1.006	1.001-1.011	< 0.05
Initial CRP	1.348	1.011-1.796	< 0.05
Tumor size	1.31	1.198-1.433	< 0.01
TNM	1.555	1.26-1.92	< 0.01
Lipiodol dose	1.12	1.058-1.185	< 0.01
Multivariate analysis ³			
Tumor size	1.252	1.108-1.414	< 0.01
Poor BS control	2.442	1.310-4.55	< 0.01
Portal vein thrombosis	3.344	1.021-10.98	< 0.05
Cox regression analysis ⁴			
Poor CT response after TACE	0.302	0.192-0.765	< 0.01
Lipiodol dose (\geq 7 mL)	0.494	0.279-0.874	< 0.05

¹Predictors for postembolization fever; ²Risk factors for 20-mo mortality;

³Predictors for 20-mo mortality; ⁴Predictors for tumor progression-free survival. HR: Hazard ratio; CRP: C-reactive protein; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; TNM: Tumor, node, metastases; TACE: Transcatheter arterial chemoembolization; BS: Base of support.

dose (\geq 7 mL) ($P < 0.01$), poor blood glucose control ($P < 0.05$), ALT value after TACE ($P < 0.01$) and CRP value after TACE ($P < 0.05$) were possible risk factors correlated with postembolization fever in patients with HCC.

A multivariate analysis using logistic regression showed that the ALT value after TACE ($P < 0.05$) and the lipiodol dose (\geq 7 mL) ($P < 0.05$) were independent predictive factors of postembolization fever (Table 2).

PEF was not associated with 20-mo survival ($P = 0.754$), 10-mo survival ($P = 0.524$) and CT response ($P = 0.461$) in the univariate analysis.

20-mo mortality

The univariate analysis revealed that poor blood sugar control ($P < 0.01$), portal vein thrombosis ($P < 0.01$), favorable CT response after TACE ($P < 0.01$), initial AST ($P < 0.05$), initial CRP ($P < 0.05$), tumor size ($P < 0.01$), TNM stage ($P < 0.01$) and lipiodol dose ($P < 0.01$) were possible risk factors correlated with 20-mo mortality (Table 2). A multivariate analysis using logistic regression showed that tumor size ($P < 0.01$), poor blood glucose control ($P < 0.01$) and portal vein thrombosis ($P < 0.05$) were independent risk factors for 20-mo mortality (Table 2).

Tumor progression-free survival

The progression-free survival in the poorly defined tumor (diffuse) type group was significantly shorter than in the well-defined (nodular) tumor type group ($P < 0.01$). Additionally, large size (size \geq 5 cm, $P < 0.01$), no antiviral treatment ($P < 0.05$), poor CT response ($P < 0.01$), lipiodol dose (dose \geq 7 mL, $P < 0.01$), antibiotic use

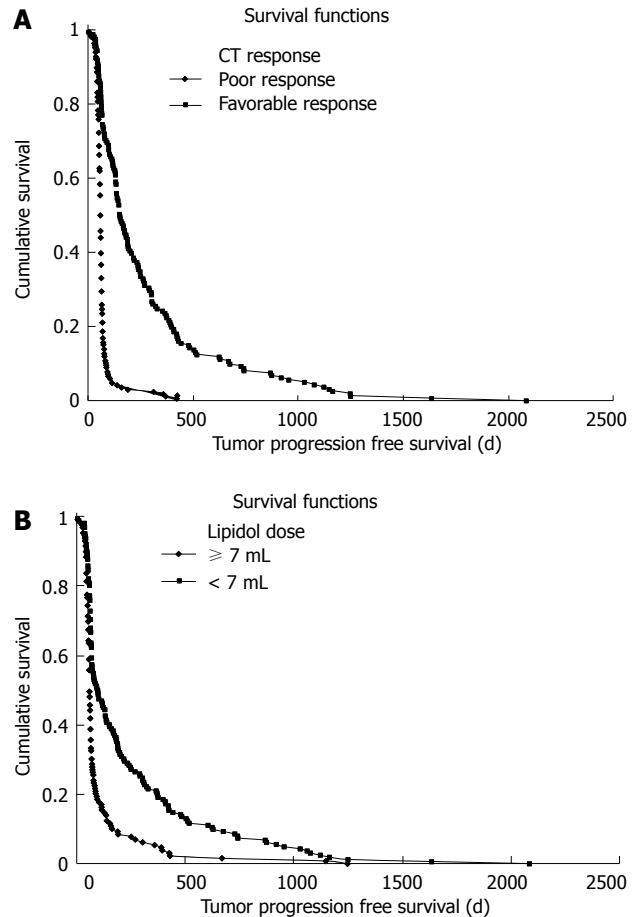


Figure 2 Kaplan-Meier curves of progression-free survival. A: According to computed tomography response (log-rank test, $P < 0.001$); B: According to lipiodol dose (log-rank test, $P < 0.001$).

($P < 0.05$) and CRP (CRP \geq 1 mg/dL, $P < 0.01$) were statistically significant factors in the univariate analysis (Figure 2A and B). In the multivariate Cox proportional hazard model for progression-free survival, CT response ($P < 0.01$) and lipiodol dose (\geq 7 mL) ($P < 0.05$) were identified as independent factors (Table 2).

DISCUSSION

TACE is one of the major treatment methods for unresectable HCC and has demonstrated survival benefits^[4,10-15]. Chemoembolization acts by obstructing the hepatic artery with embolization agents, usually gelatin, and introducing antitumor agents (e.g., cisplatin, doxorubicin, and mitomycin C) emulsified in iodized oil, thereby inducing extensive necrosis in large vascularized HCC tumors^[7]. TACE complications can be categorized as hepatic injuries, including deterioration of hepatic function, hepatic infarction, or intrahepatic biloma, and liver abscess; extrahepatic complications, including gastrointestinal bleeding, gallbladder or spleen infarction, and pulmonary embolism; and systemic complications, including postembolization syndrome and septicemia^[16]. The most frequent complication of chemoembolization is postembolization fever, which can typically be satisfactorily

alleviated with symptom treatment^[11,16]. Nevertheless, PEF frequently troubles patients, family members and physicians, and few data have been published concerning postembolization fever; therefore, we investigated the risk factors and clinical significance of PEF that developed after TACE in patients with HCC.

Infectious complications are very rare because of the standard antiseptic procedures associated with TACE. Another study reported that only 0.26% of HCC patients developed liver abscesses after TACE^[17]. Thus, antibiotic prophylaxis is usually not necessary in patients with HCC who are undergoing TACE^[10]. Although fevers were common (27%) in this study, they were generally not caused by an infectious process because very few patients had bacterial infections (0.16%). These fevers can often be adequately controlled with antipyretics, and in most cases, antibiotics are not necessary.

The pathogenesis of PEF remains unclear and complicated. The main aspects are as follows: (1) lipiodol-induced embolisms may result in ischemia, hypoxia, and necrosis in some normal hepatic cells; (2) chemotherapeutic drugs themselves have toxicities^[18]; (3) the procedure itself can lead to a considerable release of inflammatory factors^[19]; and (4) such stimuli as injury and drugs can contribute to stress responses in the human body. In the present study, the occurrence of PEF was closely associated with several clinical and laboratory variables, including poor blood glucose control, large tumor size (> 5 cm), poorly defined tumor type, post-TACE CRP level, lipiodol dose higher than 7 mL, and post-TACE ALT level in a univariate analysis. However, the multiple regression analysis showed that a lipiodol dose over 7 mL and the post-TACE ALT level were independent risk factors, which is similar to the result of another recent study^[20]. No difference was found between favorable CT responses and unfavorable CT responses regarding the presence of PEF, which is in concordance with another study showing that post-TACE fever was not associated with an enhanced tumor response in patients with HCC^[20-22].

PEF is thought to reflect extensive tumor necrosis, thereby representing the efficacy of chemoembolization^[22-24]. However, we observed no other robust association between PEF and survival in this study. Moreover, PEF did not independently affect progression-free survival, which may be an indirect indicator of treatment efficacy. These findings suggest that the previously described correlation between PEF and the extent of tissue necrosis cannot always be justified because the extent of tissue necrosis after chemoembolization is proportional to tumor mass, a factor that is independently associated with PEF.

Raoul *et al.*^[25] suggested that factors associated with poor TACE outcomes included Child-Pugh score, reduced liver function, AFP level, tumor size, tumor number, tumor type, portal vein thrombosis, multiple TACE sessions and lobar embolization. In our study, large tumor size, poor blood glucose control and portal vein thrombosis were independent risk factors for 20-mo mortality.

However, unlike in other studies, the Child-Pugh score was not a significant prognostic factor in our study. It was likely that most of the patients enrolled in our study had Child A (the Child score for patients with PEF was 5.66 *vs* 5.77 for those without PEF), and as a result, liver function did not affect the prognosis. According to Shim *et al.*^[22], hepatitis B virus infection, modified UICC stage (Stage 1), and response to chemoembolization are independent predictive factors for the tumor progression-free interval. In good agreement with other studies, we found that the TACE response and lipiodol dose (< 7 mL) were closely associated with tumor progression-free survival. Because the lipiodol dose was dependent on tumor size, progression-free survival was closely associated with tumor size and response to TACE.

This study had several limitations, including its retrospective design. Although we performed laboratory and culture studies of blood, urine, and ascites to detect hidden infections, it was impossible to rule out all infective complications in patients with PEF. In addition, the tumor response to chemoembolization may have been overestimated in patients with poorly defined HCC, particularly those with the diffuse infiltrative type of HCC, because of the difficulty of evaluating the degree of viable tumor. Finally, it may be difficult to determine whether the development of PEF after a single session of TACE was exclusively associated with overall survival because the mortality of TACE-treated HCC patients is subject to many other factors.

In conclusion, ALT levels after TACE and lipiodol dose were independent risk factors for postembolization fever in HCC patients. However, postembolization fever after TACE had no impact on survival at 20 mo or on the radiologic response.

COMMENTS

Background

Postembolization fever (PEF) is thought to reflect extensive tumor necrosis and thereby represent the efficacy of transcatheter arterial chemoembolization (TACE) in hepatocellular carcinoma (HCC). The aim of this study was to assess the tumor response and survival of patients with PEF after TACE and to determine the risk factors for PEF.

Research frontiers

TACE has improved the prognosis of patients with HCC. However, TACE inevitably results in a hypoxic insult to the HCC and the surrounding liver tissue, and postembolization fever is common. However, the clinical meaning of and the risks factor for PEF are not known.

Innovations and breakthroughs

PEF has been associated with overall survival in HCC patients. However, there is limited published data about PEF and the risk factors for PEF. Unlike other studies, their demonstrated that PEF was not associated with overall survival or radiological tumor response. Furthermore, this study demonstrated that alanine aminotransferase (ALT) levels after TACE and lipiodol doses were risk factors for PEF.

Applications

The results of this study imply that PEF is associated with lipiodol dose and ALT levels after TACE but does not affect prognosis; therefore, patients undergoing TACE who have risk factors for PEF should receive active or prophylactic antipyretics for PEF control.

Terminology

Postembolization fever was defined as a body temperature greater than 38.0 °C

that developed within 3 d of TACE without evidence of infection. Poorly defined tumor type was defined as diffuse-type HCC, whereas well-defined tumor type was defined as nodular HCC. Tumor progression-free survival was defined as the interval of time during and after treatment in which a patient remained alive and the disease did not worsen (in the case of this study, the interval from the 1st TACE to the 2nd TACE).

Peer review

Postembolization fever is a clinically relevant problem in the treatment of patients with HCC. The authors addressed potential causes and risk factors and found that the ALT level after TACE and the lipiodol dose served as independent risk factors for PEF after TACE and that PEF had no impact on 20-mo survival in HCC patients. These results were extracted and calculated from a large group of 443 patients who were treated and observed within a 5-year period.

REFERENCES

- 1 Yang JD, Roberts LR. Hepatocellular carcinoma: A global view. *Nat Rev Gastroenterol Hepatol* 2010; **7**: 448-458 [PMID: 20628345 DOI: 10.1038/nrgastro.2010.100]
- 2 Park JW. [Hepatocellular carcinoma in Korea: introduction and overview]. *Korean J Gastroenterol* 2005; **45**: 217-226 [PMID: 15843747 DOI: 200504302]
- 3 Mazzaferro V, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, Montalto F, Ammatuna M, Morabito A, Gen-nari L. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996; **334**: 693-699 [PMID: 8594428 DOI: 10.1056/NEJM199603143341104]
- 4 Llovet JM, Real MI, Montaña X, Planas R, Coll S, Aponte J, Ayuso C, Sala M, Muchart J, Solà R, Rodés J, Bruix J. Arterial embolisation or chemoembolisation versus symptomatic treatment in patients with unresectable hepatocellular carcinoma: a randomised controlled trial. *Lancet* 2002; **359**: 1734-1739 [PMID: 12049862 DOI: 10.1016/S0140-6736(02)08649-X]
- 5 Bruix J, Sala M, Llovet JM. Chemoembolization for hepatocellular carcinoma. *Gastroenterology* 2004; **127**: S179-S188 [PMID: 15508083 DOI: S0016508504016117]
- 6 Llovet JM, Bruix J, Gores GJ. Surgical resection versus transplantation for early hepatocellular carcinoma: clues for the best strategy. *Hepatology* 2000; **31**: 1019-1021 [PMID: 10733561 DOI: 10.1053/he.2000.6959]
- 7 Llovet JM, Bruix J. Systematic review of randomized trials for unresectable hepatocellular carcinoma: Chemoembolization improves survival. *Hepatology* 2003; **37**: 429-442 [PMID: 12540794 DOI: 10.1053/jhep.2003.50047]
- 8 Okada S. Transcatheter arterial embolization for advanced hepatocellular carcinoma: the controversy continues. *Hepatology* 1998; **27**: 1743-1744 [PMID: 9620352 DOI: 10.1002/hep.510270639]
- 9 Matsuda Y, Kawata S, Nagase T, Maeda Y, Yamasaki E, Kiso S, Ishiguro H, Matsuzawa Y. Interleukin-6 in transcatheter arterial embolization for patients with hepatocellular carcinoma. Effects of serine protease inhibitor. *Cancer* 1994; **73**: 53-57 [PMID: 8275438]
- 10 A comparison of lipiodol chemoembolization and conservative treatment for unresectable hepatocellular carcinoma. Groupe d'Etude et de Traitement du Carcinome Hépatocellulaire. *N Engl J Med* 1995; **332**: 1256-1261 [PMID: 7708069 DOI: 10.1056/NEJM199505113321903]
- 11 Chan AO, Yuen MF, Hui CK, Tso WK, Lai CL. A prospective study regarding the complications of transcatheter intraarterial lipiodol chemoembolization in patients with hepatocellular carcinoma. *Cancer* 2002; **94**: 1747-1752 [PMID: 11920537 DOI: 10.1002/cncr.10407]
- 12 Takayasu K, Arai S, Matsuo N, Yoshikawa M, Ryu M, Takasaki K, Sato M, Yamanaka N, Shimamura Y, Ohto M. Comparison of CT findings with resected specimens after chemoembolization with iodized oil for hepatocellular carcinoma. *AJR Am J Roentgenol* 2000; **175**: 699-704 [PMID: 10954453]
- 13 Boix J, Lorenzo-Zúñiga V, Moreno de Vega V, Domènech E, Gassull MA. Endoscopic resection of ampullary tumors: 12-year review of 21 cases. *Surg Endosc* 2009; **23**: 45-49 [PMID: 18398649 DOI: 10.1007/s00464-008-9866-3]
- 14 Kanematsu T, Furuta T, Takenaka K, Matsumata T, Yoshida Y, Nishizaki T, Hasuo K, Sugimachi K. A 5-year experience of lipiodolization: selective regional chemotherapy for 200 patients with hepatocellular carcinoma. *Hepatology* 1989; **10**: 98-102 [PMID: 2544499 DOI: S0270913989001254]
- 15 Lo CM, Ngan H, Tso WK, Liu CL, Lam CM, Poon RT, Fan ST, Wong J. Randomized controlled trial of transarterial lipiodol chemoembolization for unresectable hepatocellular carcinoma. *Hepatology* 2002; **35**: 1164-1171 [PMID: 11981766 DOI: 10.1053/jhep.2002.33156]
- 16 Chung JW, Park JH, Han JK, Choi BI, Han MC, Lee HS, Kim CY. Hepatic tumors: predisposing factors for complications of transcatheter oily chemoembolization. *Radiology* 1996; **198**: 33-40 [PMID: 8539401]
- 17 Ong GY, Changchien CS, Lee CM, Wang JH, Tung HD, Chuah SK, Chiu KW, Chiou SS, Cheng YF, Lu SN. Liver abscess complicating transcatheter arterial embolization: a rare but serious complication. A retrospective study after 3878 procedures. *Eur J Gastroenterol Hepatol* 2004; **16**: 737-742 [PMID: 15256974 DOI: 00042737-200408000-00003]
- 18 Nagano M, Nakamura T, Niimi S, Fujino T, Nishimura T, Murayama N, Ishida S, Ozawa S, Saito Y, Sawada J. Substitution of arginine for cysteine 643 of the glucocorticoid receptor reduces its steroid-binding affinity and transcriptional activity. *Cancer Lett* 2002; **181**: 109-114 [PMID: 12430185]
- 19 Distelhorst CW. Recent insights into the mechanism of glucocorticosteroid-induced apoptosis. *Cell Death Differ* 2002; **9**: 6-19 [PMID: 11803370 DOI: 10.1038/sj.cdd.4400969]
- 20 Li CP, Chao Y, Chen LT, Lee RC, Lee WP, Yuan JN, Yen SH, Lee SD. Fever after transcatheter arterial chemoembolization for hepatocellular carcinoma: incidence and risk factor analysis. *Scand J Gastroenterol* 2008; **43**: 992-999 [PMID: 19086281]
- 21 Wigmore SJ, Redhead DN, Thomson BN, Currie EJ, Parks RW, Madhavan KK, Garden OJ. Postchemoembolisation syndrome--tumour necrosis or hepatocyte injury? *Br J Cancer* 2003; **89**: 1423-1427 [PMID: 14562011 DOI: 10.1038/sj.bjc.6601329]
- 22 Shim JH, Park JW, Choi JI, Kim HB, Lee WJ, Kim CM. Does postembolization fever after chemoembolization have prognostic significance for survival in patients with unresectable hepatocellular carcinoma? *J Vasc Interv Radiol* 2009; **20**: 209-216 [PMID: 19084432 DOI: 10.1016/j.jvir.2008.10.021]
- 23 Hemingway AP, Allison DJ. Complications of embolization: analysis of 410 procedures. *Radiology* 1988; **166**: 669-672 [PMID: 3340761]
- 24 Yamada R, Sato M, Kawabata M, Nakatsuka H, Nakamura K, Takashima S. Hepatic artery embolization in 120 patients with unresectable hepatoma. *Radiology* 1983; **148**: 397-401 [PMID: 6306721]
- 25 Raoul JL, Sangro B, Forner A, Mazzaferro V, Piscaglia F, Bolondi L, Lencioni R. Evolving strategies for the management of intermediate-stage hepatocellular carcinoma: available evidence and expert opinion on the use of transarterial chemoembolization. *Cancer Treat Rev* 2011; **37**: 212-220 [PMID: 20724077 DOI: 10.1016/j.ctrv.2010.07.006]

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IL28B polymorphism and cytomegalovirus predict response to treatment in Egyptian HCV type 4 patients

Mostafa K El Awady, Noha G Bader El Din, Ashraf Tabll, Yaser El Hosary, Ashraf O Abdel Aziz, Hesham El Khayat, Mohsen Salama, Tawfeek H Abdelhafez

Mostafa K El Awady, Noha G Bader El Din, Ashraf Tabll, Tawfeek H Abdelhafez, Department of Microbial Biotechnology, National Research Center, Cairo, Giza 12622, Egypt
Yaser El Hosary, Department of Internal Medicine, National Research Center, Cairo, Giza 12622, Egypt

Ashraf O Abdel Aziz, Department of Tropical Medicine, Cairo University, Giza 12622, Egypt

Hesham El Khayat, Theodor Bilharz Research Institute, Giza 12622, Egypt

Mohsen Salama, National Liver Institute, Shebeen El Kom 32512, Egypt

Author contributions: El Awady MK designed the experiment, designed the new primers, and wrote the manuscript; Bader El Din NG, performed research work (IL28 PCR, CMV PCR, HCV PCR), edited the manuscript and made the final revision; Tabll A helped in the manuscript writing; El Hosary Y, Abdel Aziz AO, El Khayat H, Salama M provided blood samples; Abdelhafez TH participated in designing the new primers, and performed research work (IL28 PCR).

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Correspondence to: Dr. Mostafa El Awady, PhD, Department of Microbial Biotechnology, National Research Center, El-Beheouth Street 12622, Dokki, Cairo, Giza 12622, Egypt. mkawady@yahoo.com

Telephone: +20-12-3132640 Fax: +20-2-3370931

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Abstract

AIM: To test whether the status of positive cytomegalovirus (CMV) DNA detection adds to the predictive value of IL28B and to further categorize C/T allele carriers.

METHODS: This study included 166 chronic hepatitis C (CHC) patients who received combined interferon and ribavirin therapy for 48 wk, 84 spontaneous hepatitis C virus (HCV) resolvers who were positive for IgG anti-HCV antibody and negative for HCV RNA, and 100 healthy subjects who were negative for both HCV

antibodies and RNA as controls. Genomic DNA from peripheral blood was used for IL28B rs.12979860 single nucleotide polymorphism (SNP) and CMV DNA detection. A 139 bp fragment containing IL28B SNP was amplified in all subjects by polymerase chain reaction using a specifically designed primer. Then the IL28B rs.12979860 SNP was detected by restriction fragment length polymorphism (RFLP) genotyping. The presence of CMV DNA was tested by amplification of the *gB1* gene using nested polymerase chain reaction. The role of CMV and IL28B rs.12979860 SNP genotypes in determining the response rate to combined interferon therapy and clinical status of patients were statistically analyzed.

RESULTS: Current data showed that 67% of patients carrying the IL28B 12979860 C/C allele had a sustained viral response (SVR) while the genotypes C/T and TT were associated with lower SVR rates, 50% and 48%, respectively. SVR rates for the C/C allele were lower than other HCV genotypes and/or other populations. Genotype CC was associated with the response to interferon ($P = 0.025$). Genotype C/C was reduced from 48% in controls to 14% in CHC patients suggesting its protective role against progression to chronicity. The majority of spontaneously cleared subjects (86%) were C/C, confirming its protective role. The C/T allele was present in 71% of CHC patients compared with 38% of controls, so the use of IL28B SNP genotyping only in these patients may be of little value as a predictor of response. CMV reactivation occurred in 40% of CHC patients. Co-infection with CMV seriously diminished the response to interferon (IFN) therapy, with SVR rates in C/C genotypes 87.5% in CMV-negative patients and 12.5% in CMV-positive patients ($P < 0.0001$). SVR rates among C/T carriers were reduced to $< 50\%$ in patients with positive CMV DNA while the non-response rate doubled. These data indicate that a supplemental assay for CMV viremia adds to the prognostic value of IL28B genotyping.

CONCLUSION: The results suggest that both genetic (i.e., spontaneous) and therapeutic (IFN-based therapy) arms are complementary in the battle against HCV. CMV DNA testing may be of value to better predict the response to IFN, particularly in IL28B C/T carriers.

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Key words: Hepatitis C; Interleukin 28B; Genetic polymorphisms; Human cytomegalovirus; Spontaneous clearance

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INTRODUCTION

The current standard-of-care for chronic hepatitis C virus (HCV) infection includes weekly injections of pegylated interferon- α (peg-IFN) combined with daily oral ribavirin^[1,2] for 48 wk^[3]. During the first 3 mo of therapy, HCV viral loads usually fall to undetectable levels in response to interferon- α . However, in non-responder (NR) patients, HCV viral loads persist at or near pretreatment levels. Among those patients with an initial response to treatment, known as early virological responders, up to 50% will relapse after treatment is discontinued and are known as relapsed responders, whereas the remainder will have a sustained virological response (SVR) as determined by the absence of detectable viremia 6 mo after treatment has stopped^[4]. In Egypt, several reports stated that approximately 50% of patients infected with genotype 4, the most common among Egyptian HCV patients, achieve a SVR to this regimen. Genome-wide association studies have recently revealed that single nucleotide polymorphisms (SNPs) within or adjacent to IL28B (19q13) that codes for interferon- λ 3, predict spontaneous resolution of HCV^[5,6] and a likely SVR to IFN-based treatment in chronic hepatitis C (CHC) patients infected with genotypes other than type 4^[4,5,7-9]. Therefore, IL28B SNPs may have strong predictive value for the outcome of IFN-based therapy in the difficult to treat HCV genotype 4 patients, with the hope that IL28B SNPs will improve future decision-making in the management of CHC in Egypt. Nevertheless, it was estimated that IL28B variations account for about 15% of the inter-individual variability of SVR, thus supporting the necessity for additional predictor(s) of the response to treatment. Recent predictors include vitamin D deficiency, IFN-c-inducible protein-10 serum levels, steatosis/insulin resistance^[10-17], or IFN-stimulated genes (ISGs) such as OAS1^[18] and MxA^[19]. Therefore, biologi-

cal models for the prediction of SVR should include a variety of parameters besides IL28B genotyping, as this would offer better accuracy compared with IL28B typing alone^[13]. Recently two predictors for SVR in genotype 4 patients were reported; first, reactivation of CMV infection^[20] and second, SNP at exon 7 splice acceptor site of 2' 5'OAS gene^[18]. The role of several genetic factors that influence spontaneous or treatment-induced HCV clearance, such as ISGs and genes encoding the natural killer cell receptor KIR2DL3 and its ligand, and human leukocyte antigen C group 1^[21-23] should be re-analyzed parallel with the IL28 genotype. A major difficulty in decision-making for prediction of the response at the start of treatment is the high frequency of heterozygous carriers rs 12979860 C/T among chronic HCV patients, ranging from 45% to 70% of the CHC populations and provide vague predictive results. This means that the predictive value of rs 12979860 SNP will not be useful in almost half the patient population. The search for additional predictive factors for response is mandatory. Co-infection with other pathogens is, in some instances, an interfering factor against host genotype-based prediction. Several studies revealed comparable associations between IL28B variations and treatment-induced HCV clearance in patients co-infected with human immunodeficiency virus (HIV)^[5,24-26]. For example, one study in HIV co-infected patients reported 2-3-fold higher SVR rates for HCV genotypes 1 and 4 according to rs12979860 C/C *vs* C/T, T/T, but no difference for HCV genotype 3 according to rs12979860 C/C *vs* C/T, T/T^[26]. Recently, active CMV infection was shown by our laboratory^[20] to dramatically interfere with SVR rates in genotype 4 CHC patients, probably by inhibiting the JAK-STAT cascade. We, therefore, hypothesize that the role of CMV in the IL28B association with SVR should be a strong candidate for further categorization of C/T genotype carriers.

MATERIALS AND METHODS

Subjects

The present study comprised 350 subjects, including 166 CHC patients, 84 spontaneous HCV resolvers and 100 healthy control subjects.

Chronic HCV patients: All 166 patients had mild liver disease F0-F3 according to the Metavir classification^[27]. All subjects had undetectable HBV surface antigen (HBsAg), Anti-schistosoma antibodies (Abs), and autoimmune Abs. None of the patients had a history of long-term drug use, aflatoxins consumption or alcohol intake. All patients were infected with HCV genotype 4. Patients included 105 males and 61 females, with an age range of 19-57 years. None had uncontrolled type II diabetes mellitus, thyroid dysfunction or obesity. All patients received weekly injection of peg-IFN- α plus daily oral ribavirin treatment for 48 wk. Eighty six patients achieved a SVR, namely undetectable HCV RNA 24 wk after the end of

the treatment response that was achieved after 48 wk, i.e., a total of 72 wk. The remaining 80 patients were NR who failed to achieve a SVR, i.e., either did not respond to therapy after 12 wk or achieved undetectable viremia but failed to maintain the state of no viremia till 48 wk (breakthrough) or post 48 wk (viral relapse).

Spontaneous HCV resolvers: Eighty four subjects who tested positive for IgG anti-HCV Abs (3rd generation) and negative for HCV RNA were enrolled in the study. Patients included 53 males and 31 females, with an age range of 18-55 years.

Controls: One hundred healthy subjects who tested negative for both IgG anti HCV Abs and HCV RNA served as controls for the IL28B SNP and CMV reactivation studies. Also, they were negative for HBsAg, anti-schistosoma Abs, autoimmune Abs and CMV DNA. Subjects included 50 males and 50 females, with an age range of 18-64 years. None of subjects had a history of liver insult, either viral, metabolic or drug exposure.

Methods

Extraction of peripheral blood DNA: Peripheral blood was withdrawn from all subjects into EDTA solution and DNA was extracted using genomic DNA extraction kit (Qiagen, Milan Italy). Plasma samples were separated before DNA extraction and utilized for testing HBsAg, anti schistosomiasis Abs and autoimmune Abs (ANA, AMA, LKM1, ASMA). Purified genomic DNA was used for IL28B SNP analysis and for testing the presence of CMV DNA.

Detection of IL-28B rs12979860 C/T polymorphism: Genotyping for the IL-28B rs12979860 C/T polymorphism was performed according to Fabris *et al.*^[24] with minor modification by polymerase chain reaction based restriction fragment length polymorphism (PCR-RFLP). Using purified genomic DNA, a 139 base-pair (bp) product was obtained with the forward primer 5'-CCAGGGCCCCCTAACCTCTGCA-3' and the reverse primer 5'-GGGAGCGCGGAGTGCAATTCA-3', newly designed with the aid of NCBI Primer-Blast Tool (<http://www.ncbi.nlm.nih.gov/tools/primer-blast/>). Amplification was carried out in a total volume of 10 µL containing 10 mmol/L Tris-HCl (pH 8.3), 50 mmol/L KCl, Tween-20 0.01%, 0.2 mmol/L deoxyribonucleotides, 2-4 pmol of each primer, 2.0 mmol/L MgCl₂, 0.5 units hot-start Taq DNA polymerase (Righ Taq, Euroclone, Milan, Italy) and ~10 ng of genomic DNA. The thermal protocol for amplification included 35 cycles of denaturation (at 95 °C for 60 s), annealing (at 64 °C for 60 s), and polymerization (at 72 °C for 60 s). Ten µL of the amplicons were digested with 1 unit of the BstU-I restriction endonuclease (New England Biolabs, Hitchin, UK) in a total volume of 20 µL at 37 °C for 4-6 h or overnight. The fragments were resolved by electropho-

resis in 4% agarose gel after staining with ethidium bromide. A band of 139 bp indicated the TT genotype, 109 bp indicated the CC genotype; 139 + 109 bp indicated the CT genotype.

PCR amplification of the CMV gB gene: Primers for the first-round PCR were derived from the gB region of CMV, and PCR protocols were followed as described previously^[28,29]. The amplification mixture contained 10 pmol of each primer (CMV1 and CMV2), 0.2 mmol/L from each dNTP (Promega, Madison, WI, United States), 0.1 U Taq polymerase (Promega, Madison, WI, United States), 1 µL Taq buffer, 5 µL template, and distilled water to a final volume of 20 µL. A negative control tube (in which water replaced the DNA sample) was incorporated into each run. A sample from the first PCR product was used as a template in a second-round PCR (nested). Briefly, 1 µL of first round product was added to the reaction mixture, which is the same as the first-round reaction mix except for use of CMV3 and CMV4 as nested primers. The thermal cycling protocol was as follows: 1 min at 94 °C, 1 min at 55 °C and 1 min at 72 °C for 30 cycles. Then, the nested PCR products were analyzed with 3% agarose gel electrophoresis and stained with ethidium bromide. A result was considered positive when a clear 100 bp product was visible on the gel.

Primers used for the first and the second-round PCR were CMV1: 5'-GAGGACAACGAAATCCTGTTGGGCA-3'; CMV2: 5'-GTCGACGGTGGAGATACTGCTGAGG-3'; CMV3: 5'-ACCACCGCACTGAGGAATGTCAG-3'; and CMV4: 5'-TCAATCATGCGTTTGAAGAGGTA-3'.

Assessment of SVR status (real-time and RT-PCR of HCV RNA: As reported in our previous study^[20,29]. Disappearance of viremia was confirmed by a variety of tests including RT-PCR using nested primers derived from the highly conserved HCV 5'UTR and real-time PCR. These tests were done 3, 6, 12 and 18 mo after the start of IFN therapy. HCV RNA was extracted using the BIOZOL-total RNA extraction reagent kit (Hangzhou Bioer Technology, Hangzhou, China). Nested RT-PCR performed in a 25 µL reaction mixture containing 20 units of avian myeloblastosis virus reverse transcriptase (Clontech, Mountain View, CA, United States), 200-400 ng of total cellular RNA as template, 40 units of RNasin (Clontech), 0.2 µmol/L from each dNTP (QBIOSGENE, Carlsbad, CA, United States), and 50 pmol of the reverse primer P2. The reaction was incubated at 42 °C for 60 min and denatured at 98 °C for 10 min. Amplification of the highly conserved 5'-UTR sequences was done using two rounds of PCR with two pairs of nested primers. First-round amplification was done in 50 µL of reaction containing 50 pmol from each of the P1 forward primer and P2 reverse primer, 0.2 mmol/L from each dNTP, 10 µL from RT reaction mixture as template and two units of Taq DNA polymerase (Finnzyme, Woburn, MA, Unit-

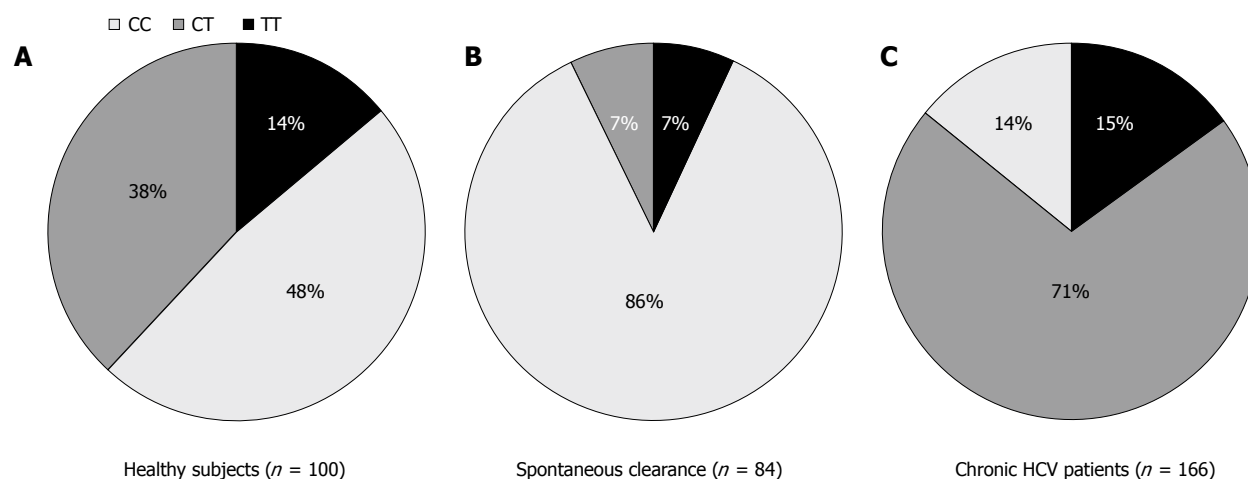


Figure 1 Distribution of IL28B rs12979860 single nucleotide polymorphism alleles among a healthy Egyptian population, spontaneous clearance and chronic hepatitis C patients. A: A hundred healthy subjects representing ethnic backgrounds of the Egyptian population were typed for IL28B single nucleotide polymorphism (SNP) rs 12979860. The distribution of different allele frequencies is depicted as a pie chart; B: Eighty four patients with spontaneous clearance from hepatitis C virus (HCV) infection were typed for IL28B SNP rs 12979860. The distribution of different allele frequencies is depicted as a pie chart; C: A hundred and sixty six chronic hepatitis C (CHC) patients with low F scores (F0-F2) were typed for IL28B SNP rs 12979860. The distribution of different allele frequencies is depicted as a pie chart.

ed States) in a 1 × buffer supplied with the enzyme. The thermal cycling protocol was as follows: 1 min at 94 °C, 1 min at 55 °C and 1 min at 72 °C for 30 cycles. The second round of amplification was done similar to the first round, except for use of the nested reverse primer p4 and forward primer p3 at 50 pmol each. A fragment of 171 bp length was identified in positive samples. Primer sequences were as follows: P1: 5'-AACTACTGTCTTCACGCAGAA-3'; P2: 5'-GGTGCACGGTCTACGAGACCTC-3'; P3: 5'-GTGCAGCCTCCAGGACCC-3'; and P4: 5'-ACTCGGCTAGCAGTCTCGCG-3'.

HCV quantitation was performed using the Fluorescence Quantitative Detection kit (Hangzhou Bioer Technology, Hangzhou, China). HCV RNA was reverse transcribed and a specific fragment was amplified with specific primers in a one-step RT-PCR reaction. The products were detected using specific Taqman-MGB Probes. The real-time RT-PCR reaction was done in 50 µL of reaction containing 25 µL RT-PCR mix, 2.5 µL Mn^{2+} , 1.5 µL HCV probe mix, and 1 µL internal control. Then 20 µL of the corresponding HCV RNA template were added. For the standard, 20 µL of the standards control 1-4 were added. All PCR tubes were placed in the real-time PCR Detection Instrument. The thermal cycling protocol was as follows: one cycle (30 s at 90 °C, 20 min at 61 °C and 1 min at 95 °C) followed by 30 cycles and 50 cycles (15 s at 95 °C, 1 min at 60 °C).

Statistical analysis

The significance of the response to IFN therapy was investigated by multivariate logistic regression analysis. CMV significantly affected the HCV response rate in C/T heterozygous carriers. The frequency of rs 12979860 SNP was compared in the control group *vs* the CHC patient group. All statistical analyses were performed using

SPSS 9.0 statistical software (SPSS Inc., Chicago, IL, United States). The odds ratio (OR) and 95% confidence interval (CI) were calculated to assess the relative risk confidence. A two tailed *P*-value 0.05 was considered statistically significant.

RESULTS

IL 28 C/C frequency in spontaneous resolvers of HCV infection

To test the hypothesis whether patients who spontaneously cleared HCV infection during or shortly following the acute phase bear the protective C/C allele more frequently than healthy subjects, we typed the IL28 B SNP (rs 12979860) in 84 spontaneous resolvers and 100 healthy subjects. The results of this study were previously described by our laboratory and are shown in Figure 1A and B, which clearly indicate the protective role of the C/C allele in patients who spontaneously cleared the virus, where C/C was present in 86% of spontaneous resolvers *vs* 48% of healthy subjects.

IL28B C/C frequency in chronic HCV patients

To examine the allele frequencies of the C/C genotype according to progression to chronic HCV infection, we compared the IL28B typing data derived from 166 chronic HCV patients with 84 subjects with spontaneous clearance of HCV. The results showed that the frequency of the protective C/C allele was markedly decreased from 86% in spontaneous resolvers to 48% in healthy controls to 14% in CHC patients regardless of their response to IFN therapy (Figure 1A-C). This decline was associated with a comparable increase in the heterozygous C/T from 7% in spontaneous resolvers to 38% in healthy subjects and to 71% in CHC patients.

Table 1 Statistical analyses of the different IL28B genotypes and alleles in non-responder and sustained viral response patients *n* (%)

	Non-responders (NR = 79)	Responders (SVR = 87)	Odds ratio	95%CI	<i>P</i> value
CC (<i>n</i> = 24)	8 (33.3)	16 (66.7)	4.575	1.376-7.846	0.025
CT (<i>n</i> = 117)	58 (49.6)	59 (50.4)	0.858	0.538-1.449	0.589
TT (<i>n</i> = 25)	13 (52)	12 (48)	0.978	0.758-1.759	0.464
C (<i>n</i> = 165)	74 (44.8)	91 (55.2)	1.636	0.773-3.465	0.198
T (<i>n</i> = 167)	84 (50.3)	83 (49.7)	0.798	0.598-1.349	0.604

Frequencies of genotypes and alleles at IL28B SNP were compared between SVR and NR patients. Genotype CC were associated with SVR ($P = 0.025$). SVR: Sustained viral response; NR: Non-responder.

Rate of SVR in patients with the rs12979860 C/C allele

Among 166 CHC patients treated with combined IFN and ribavirin therapy, 79 patients did not achieve a SVR while 87 patients achieved a SVR. The effect of different IL28B genotypes and alleles on the HCV response to IFN treatment are shown in Table 1. The genotype CC was associated with response to IFN ($P = 0.025$). Of the 24 CHC patients bearing the C/C allele, 16 (67%) achieved a SVR, while the other two genotypes C/T and TT were associated with lower SVR rates: 50% and 48%, respectively (Figure 2).

Association of CMV reactivation with SVR rates in C/C patients

The PCR results of the CMV *gB* gene showed that 66/166 (40%) of chronic HCV patients have detectable CMV DNA. Since we have previously shown that coinfection with CMV seriously diminished the response to IFN therapy, it was tempting to examine whether this co-infection is associated with lower SVR rates in C/C holders. The data shown in Figure 3 clearly demonstrate that positive CMV DNA dramatically reduced the SVR rates in C/C genotypes as represented by a 12.5% SVR rate in CMV-positive patients compared with 87.5% in CMV-negative patients ($P < 0.0001$).

Sorting of C/T genotype according to association with CMV infection

Since the C/T allele represents the majority of CHC patients (117/166) (71%), the use of IL28B SNP genotyping may be of little value as a predictor of response prior to starting the treatment regimen. Therefore, there is a great need to include another factor that was previously known to affect SVR rates. Table 2 shows the results of CMV testing in CT genotype patients. Specifically, CMV DNA was detected in 71% of NR patients and in 29% of SVR patients. In Figure 4, the SVR rates among C/T carriers was reduced to < 50% of its value in cases where patients had positive CMV DNA. On the other hand, the NR rate doubled in cases of positive CMV DNA. These data indicate that a supplemental assay for CMV viremia adds to the prognostic value of

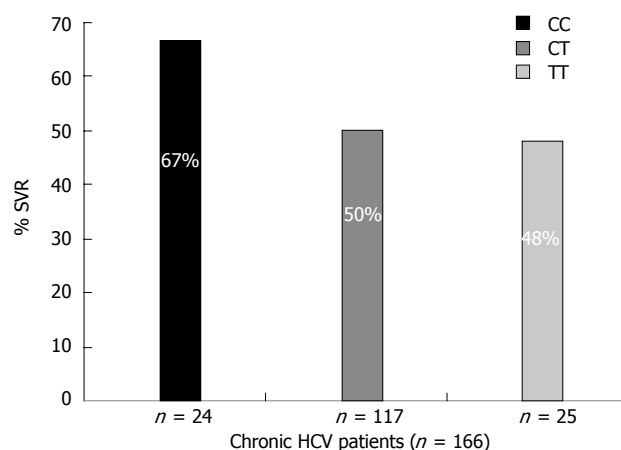


Figure 2 Rate of sustained viral response in IL28B variants. Among 166 chronic hepatitis C (CHC) patients treated with combined interferon (IFN) and ribavirin therapy, 80 patients did not achieve a sustained viral response (SVR) while 86 patients achieved a SVR. Among the 14% (24) of CHC patients bearing the C/C allele, 67% (16 patients) achieved a SVR, while the other two genotypes C/T and TT were associated with lower SVR rates; 50% and 48%, respectively. Genotype CC was associated with response to IFN ($P = 0.025$). HCV: Hepatitis C virus.

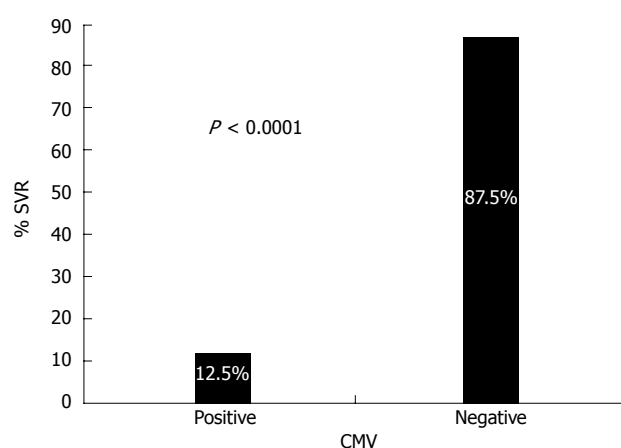


Figure 3 Association of cytomegalovirus reactivation with sustained viral response rates in C/C patients. The data shown clearly demonstrate that cytomegalovirus (CMV) reactivation has dramatically reduced the sustained viral response (SVR) rates in C/C genotypes as represented by a 12.5% SVR rate in CMV-positive patients compared with 87.5% in CMV-negative patients ($P < 0.0001$).

IL28B genotyping.

DISCUSSION

Several predictive factors have been reported recently for SVR rates for combination therapy in HCV infection. Viral factors, other than viral loads and genotypes were reported, including substitutions at core amino acids 70 and 91^[30,31], IFN sensitivity determining region variations^[32] and IFN and ribavirin response determining region variations^[33]. SNPs within a number of host genes were found to be associated with different treatment outcomes to therapy. These include IFN- α pathway genes^[34] and IFN-stimulated genes such as OAS^[18]

Table 2 Influence of cytomegalovirus infection on response rate to interferon combined therapy in chronic hepatitis C virus infected patients with IL28B C/T genotype *n* (%)

HCV response to treatment (<i>n</i> = 117)	IL 28B genotype (C/T) +ve CMV patients (<i>n</i> = 45)	IL 28B genotype (C/T) -ve CMV patients (<i>n</i> = 72)
NR (<i>n</i> = 59)	32 (71)	27 (38)
SVR (<i>n</i> = 58)	13 (29)	45 (62)

Reactivation of cytomegalovirus (CMV) was assessed by amplification of viral genome in all patients. Association of IL28B genotype C/T and CMV emia with response to interferon based therapy is illustrated in this table. SVR: Sustained viral response; NR: Non-responder.

and MxA^[19]. The widely studied IL28B SNP allele was reported to correlate with a favorable therapeutic response and was also associated with spontaneous clearance of HCV^[6]. Thomas *et al.*^[6] reported that the allele related to HCV clearance, namely rs 12979860 C/C, was the major allele in the majority of Asian and European countries. Conversely, patients of African ancestry had lower C/C allele frequency and SVR rates. The current data indicate that the Egyptian population infected mainly with genotype 4 has an intermediate response between that of the populations of African and European ancestries, i.e., the C/C allele frequency in normal subjects was 48% *vs* > 50% in Europeans and ~35% in Africans while SVR rates in the currently studied CHC patients were 67% *vs* ~80% in Europeans and ~50% in Africans. One of the most interesting aspects of the current study is the dramatic decline in C/C frequency in CHC patients (from 48% in controls to 14% in genotype 4 infections), thus allowing speculation that protective IL28B variations provide a more substantial advantage in acute HCV genotype 4. In a similar study on a German cohort, the rs12979860 C/C genotype was found in 49% of uninfected subjects and declined in chronic infection to 42.7% of HCV genotype 2 and 3 patients and 33.9% of HCV genotype 1 patients^[35]. Taking together the data from the German cohort and the data of the current study, we may speculate that HCV genotype 4 has the highest advantage of viral clearance during acute infection followed by genotype 1 and lastly genotypes 2 and 3.

Strong evidence from the current data on the significant protective effect of the IL28 B variant in acute HCV 4 is the high percentage of the C/C genotype among spontaneously cleared individuals, i.e., positive HCV Abs and negative HCV RNA compared with controls and CHC patients (86% *vs* 48 and 14%, respectively). The lack of a protective role of the C/C genotype in 14% of CHC patients suggests that, in CHC patients with the C/C genotype, other factors are involved in development of chronic infection. It is tempting to hypothesize that IL28B C/C in those 24 CHC patients was not sufficient to provide substantial tendency towards viral clearance and consequently explains the relatively lower figure for the C/C association with SVR rate in the present cohort compared with Asian and Euro-

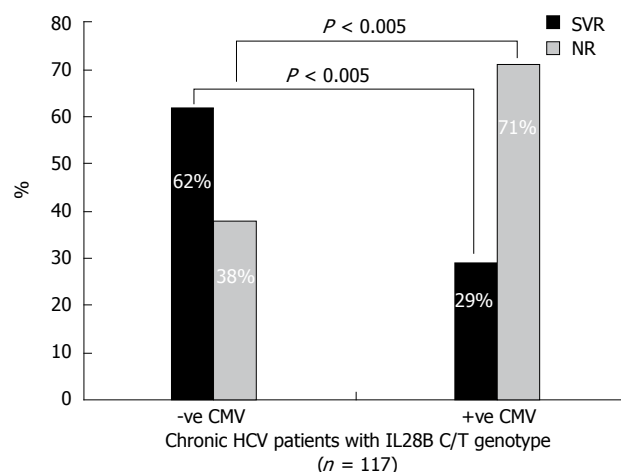


Figure 4 Sorting of C/T genotype according to association with cytomegalovirus infection. The sustained viral response (SVR) rates among C/T carriers, is reduced to < 50% of its value in cases where patients had cytomegalovirus (CMV) reactivation. On the other hand, the non-response rate is increased 2-fold in cases of CMV reactivation. HCV: Hepatitis C virus.

pean studies. In a recent study from our laboratory co-infection with active CMV viremia induced severe failure to achieve acceptable rates of SVR^[20]. Infection with CMV has evolved multiple mechanisms for disrupting the IFN-stimulated JAK/STAT signal transduction. It appears to inhibit IFN- α responsiveness by decreasing JAK1 protein, which is an essential component of IFN- α signaling^[36]. It was reported that CMV blocks IFN-stimulated gene factor 3 (ISGF3)-dependent (MHC class I, 2',5'-OAS, and MxA) and ISGF3-independent gene expression in infected cells. Moreover, the essential component of ISGF3, p48, is significantly decreased by CMV. Therefore decreased JAK1 and p48 protein would inhibit IFN- α stimulated signal transduction, transcription factor activation, and gene expression, thus it is likely to globally block IFN-stimulated responses in CMV-infected patients^[37]. The dual analysis of both roles of IL28B SNP and CMV co-infection clearly showed that C/C carriers not infected with CMV have a 7-fold higher rate of SVR than those C/C patients co-infected with CMV. One of the major problems in decision-making on an individual patient basis is the overwhelming preponderance of the IL28B C/T genotype among the current CHC cohort as well as other cohorts^[38,39]. The present results indicate that the IL28B SNP is not the only factor that influences the IFN-induced antiviral activities of host cells against viral infection. The present results showed a 2-fold increase in SVR rate in CMV-negative over CMV-positive C/T carriers and clearly highlight the significance of additional testing for CMV viremia besides IL28 genotyping to set a model for accurate prediction of response to combined therapy in C/T carriers.

In conclusion the current results allow us to speculate that both genetic (i.e., spontaneous) and therapeutic (IFN-based therapy) arms are complementary in the battle against HCV where poor spontaneous clearance is

associated with higher IFN-mediated response as in genotypes 2 and 3. On the other hand, the difficult to treat genotypes 1 and 4 have clearly better chances for spontaneous resolution. Furthermore, CMV DNA testing may be of value at the moment to make better prognostication on response to IFN particularly in heterozygous carriers. Needless to say the search for supplemental predictive host factors will never stop at least in the near future.

COMMENTS

Background

The current recommended treatment for chronic hepatitis C virus (HCV) infection is the combination of pegylated α -interferon (IFN) and ribavirin given for 48 wk. Both viral and host factors may contribute to the phenomenon that some patients with chronic hepatitis C respond well to IFN therapy but others do not. The host genetic factors mediate the vigor of the immune response of the host through the control of inter-individual differences in the production of intracellular antiviral proteins or some cytokines. Different studies revealed that single nucleotide polymorphisms (SNPs) within or adjacent to IL28B (19q13) that codes for interferon- λ 3, predict spontaneous resolution of HCV and likely SVR to IFN based treatment in chronic HCV patients infected with genotypes other than type 4. Since more than 93% of patients in Egypt are infected with HCV genotype 4, the authors investigated whether IL28B SNPs may be of predictive value for the outcome of IFN-based therapy in the difficult to treat HCV genotype 4 patients. Also the authors tested whether the status of positive cytomegalovirus (CMV) DNA detection added to the predictive value of IL28B and to further categorization of C/T allele carriers.

Research frontiers

HCV is a global health problem that infects more than 170 million people around the world and more than 15% of the Egyptian population. The response rate to combined IFN therapy does not exceed 50% of patients infected with HCV genotype 4, the most prevalent subtype in Egypt. Timing and rules for this therapy without major complications are not very well defined, so that infected patients not only suffer from severe adverse effects but there is also a contribution to evolution of more resistant strains of the virus. A research focus is how to determine and understand which HCV patients are likely to develop persistent infection, progressive liver disease or do not respond significantly to IFN therapy. These criteria are of utmost importance in disease control programs and treatment strategies. Therefore, there is an urgent need for biological models containing a variety of prognostic parameters to predict the response to IFN treatment in HCV-infected patients and to help make better decisions on treatment strategy.

Innovations and breakthroughs

The use of IL28B SNP analysis in predicting the response to combined IFN therapy is informative only in cases of CC or TT genotypes. Since the heterozygous genotype CT represents more than 70% of chronic HCV patients, there is a need for supplemental markers for making a decision on whether the HCV patient is a potential responder to combined IFN therapy or not. The authors' data clearly improve the decision-making by detecting CMV reactivation in the CT genotype group. CMV reactivation has been shown to dramatically interfere with the response to combined IFN treatment. The data presented in this manuscript are indeed important for hepatologists regarding the anticipated response to standard combined therapy for HCV genotype 4 patients.

Applications

The data of this manuscript help hepatologists make better decisions on response rate to combined interferon therapy for HCV genotype 4 patients.

Peer review

In this manuscript, the authors aimed to investigate whether the status of positive cytomegalovirus (CMV) DNA detection increases the predictive value of IL28B and to further categorize C/T allele carriers. The authors have found that CMV DNA testing might be valuable at the moment to make better decisions on response to IFN particularly in IL28B C/T carriers. Overall, this topic is very interesting.

REFERENCES

- 1 Ferenci P. Current Treatment for Chronic Hepatitis C. *Curr Treat Options Gastroenterol* 2004; **7**: 491-499 [PMID: 15527715 DOI: 10.1007/s11938-004-0008-2]
- 2 Hadziyannis SJ, Sette H, Morgan TR, Balan V, Diago M, Marcellin P, Ramadori G, Bodenheimer H, Bernstein D, Rizzetto M, Zeuzem S, Pockros PJ, Lin A, Ackrill AM. Peginterferon-alpha2a and ribavirin combination therapy in chronic hepatitis C: a randomized study of treatment duration and ribavirin dose. *Ann Intern Med* 2004; **140**: 346-355 [PMID: 14996676]
- 3 Lagging M, Wejstål R, Uhnöo I, Gerdén B, Fischler B, Fridman S, Josephson F, Karlström O, Sangfelt P, Schvarz R, Weiland O. Treatment of hepatitis C virus infection: updated Swedish Consensus recommendations. *Scand J Infect Dis* 2009; **41**: 389-402 [PMID: 20001276 DOI: 10.1080/00365540902998271]
- 4 Knapp S, Yee LJ, Frodsham AJ, Hennig BJ, Hellier S, Zhang L, Wright M, Chiaramonte M, Graves M, Thomas HC, Hill AV, Thursz MR. Polymorphisms in interferon-induced genes and the outcome of hepatitis C virus infection: roles of MxA, OAS-1 and PKR. *Genes Immun* 2003; **4**: 411-419 [PMID: 12944978 DOI: 10.1038/sj.gene.6363984]
- 5 Rauch A, Kutalik Z, Descombes P, Cai T, Di Iulio J, Mueller T, Bochud M, Battegay M, Bernasconi E, Borovicka J, Colombo S, Cerny A, Dufour JF, Furrer H, Günthard HF, Heim M, Hirschel B, Malinverni R, Moradpour D, Müllhaupt B, Witteck A, Beckmann JS, Berg T, Bergmann S, Negro F, Telenti A, Bochud PY. Genetic variation in IL28B is associated with chronic hepatitis C and treatment failure: a genome-wide association study. *Gastroenterology* 2010; **138**: 1338-145, 1338-145, [PMID: 20060832]
- 6 Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, Kidd J, Kidd K, Khakoo SI, Alexander G, Goedert JJ, Kirk GD, Donfield SM, Rosen HR, Tobler LH, Busch MP, McHutchison JG, Goldstein DB, Carrington M. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 2009; **461**: 798-801 [PMID: 19759533 DOI: 10.1038/nature08463]
- 7 Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, Heinzen EL, Qiu P, Bertelsen AH, Muir AJ, Sulkowski M, McHutchison JG, Goldstein DB. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 2009; **461**: 399-401 [PMID: 19684573 DOI: 10.1038/nature08309]
- 8 Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, Abate ML, Bassendine M, Spengler U, Dore GJ, Powell E, Riordan S, Sheridan D, Smedile A, Fragomeli V, Müller T, Bahlo M, Stewart GJ, Booth DR, George J. IL28B is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat Genet* 2009; **41**: 1100-1104 [PMID: 19749758 DOI: 10.1038/ng.447]
- 9 Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, Nakagawa M, Korenaga M, Hino K, Hige S, Ito Y, Mita E, Tanaka E, Mochida S, Murawaki Y, Honda M, Sakai A, Hiasa Y, Nishiguchi S, Koike A, Sakaida I, Imamura M, Ito K, Yano K, Masaki N, Sugauchi F, Izumi N, Tokunaga K, Mizokami M. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 2009; **41**: 1105-1109 [PMID: 19749757 DOI: 10.1038/ng.449]
- 10 Zeuzem S, Berg T, Moeller B, Hinrichsen H, Mauss S, Wedemeyer H, Sarrazin C, Hueppe D, Zehnter E, Manns MP. Expert opinion on the treatment of patients with chronic hepatitis C. *J Viral Hepat* 2009; **16**: 75-90 [PMID: 18761607]
- 11 Berg T, Sarrazin C, Herrmann E, Hinrichsen H, Gerlach T, Zachoval R, Wiedenmann B, Hopf U, Zeuzem S. Prediction

- of treatment outcome in patients with chronic hepatitis C: significance of baseline parameters and viral dynamics during therapy. *Hepatology* 2003; **37**: 600-609 [PMID: 12601358 DOI: 10.1053/jhep.2003.50106]
- 12 **Thompson AJ**, Muir AJ, Sulkowski MS, Ge D, Fellay J, Shianna KV, Urban T, Afdhal NH, Jacobson IM, Esteban R, Poordad F, Lawitz EJ, McCone J, Shiffman ML, Galler GW, Lee WM, Reindollar R, King JW, Kwo PY, Ghalib RH, Freilich B, Nyberg LM, Zeuzem S, Poynard T, Vock DM, Pieper KS, Patel K, Tillmann HL, Noviello S, Koury K, Pedicone LD, Brass CA, Albrecht JK, Goldstein DB, McHutchison JG. Interleukin-28B polymorphism improves viral kinetics and is the strongest pretreatment predictor of sustained virologic response in genotype 1 hepatitis C virus. *Gastroenterology* 2010; **139**: 120-9.e18 [PMID: 20399780 DOI: 10.1053/j.gastro.2010.04.013]
- 13 **Bitetto D**, Fattovich G, Fabris C, Ceriani E, Falletti E, Fornasiere E, Pasino M, Ieluzzi D, Cussigh A, Cmet S, Pirisi M, Toniutto P. Complementary role of vitamin D deficiency and the interleukin-28B rs12979860 C/T polymorphism in predicting antiviral response in chronic hepatitis C. *Hepatology* 2011; **53**: 1118-1126 [PMID: 21480318 DOI: 10.1002/hep.24201]
- 14 **Lange CM**, Bojunga J, Ramos-Lopez E, von Wagner M, Hassler A, Vermehren J, Herrmann E, Badenhop K, Zeuzem S, Sarrazin C. Vitamin D deficiency and a CYP27B1-1260 promoter polymorphism are associated with chronic hepatitis C and poor response to interferon- α 2a and ribavirin. *J Hepatol* 2011; **54**: 887-893 [PMID: 21145801 DOI: 10.1016/j.jhep.2010.08.036]
- 15 **Lange CM**, von Wagner M, Bojunga J, Berg T, Farnik H, Hassler A, Sarrazin C, Herrmann E, Zeuzem S. Serum lipids in European chronic HCV genotype 1 patients during and after treatment with pegylated interferon- α 2a and ribavirin. *Eur J Gastroenterol Hepatol* 2010; **22**: 1303-1307 [PMID: 20729742 DOI: 10.1097/MEG.0b013e32833de92c]
- 16 **Petta S**, Cammà C, Scazzone C, Tripodo C, Di Marco V, Bono A, Cabibi D, Licata G, Porcasi R, Marchesini G, Craxi A. Low vitamin D serum level is related to severe fibrosis and low responsiveness to interferon-based therapy in genotype 1 chronic hepatitis C. *Hepatology* 2010; **51**: 1158-1167 [PMID: 20162613 DOI: 10.1002/hep.23489]
- 17 **Lagging M**, Askarieh G, Negro F, Bibert S, Söderholm J, Westin J, Lindh M, Romero A, Missale G, Ferrari C, Neumann AU, Pawlotsky JM, Haagmans BL, Zeuzem S, Bochud PY, Hellstrand K. Response prediction in chronic hepatitis C by assessment of IP-10 and IL28B-related single nucleotide polymorphisms. *PLoS One* 2011; **6**: e17232 [PMID: 21390311 DOI: 10.1371/journal.pone.0017232]
- 18 **El Awady MK**, Anany MA, Esmat G, Zayed N, Tabll AA, Helmy A, El Zayady AR, Abdalla MS, Sharada HM, El Raziky M, El Akel W, Abdalla S, Bader El Din NG. Single nucleotide polymorphism at exon 7 splice acceptor site of OAS1 gene determines response of hepatitis C virus patients to interferon therapy. *J Gastroenterol Hepatol* 2011; **26**: 843-850 [PMID: 21182542 DOI: 10.1111/j.1440-1746.2010.06605.x]
- 19 **Hijikata M**, Ohta Y, Mishiro S. Identification of a single nucleotide polymorphism in the MxA gene promoter (G/T at nt -88) correlated with the response of hepatitis C patients to interferon. *Intervirology* 2000; **43**: 124-127 [PMID: 10971132 DOI: 10.1159/000025035]
- 20 **Bader el-Din NG**, Abd el-Meguid M, Tabll AA, Anany MA, Esmat G, Zayed N, Helmy A, el-Zayady AR, Barakat A, el-Awady MK. Human cytomegalovirus infection inhibits response of chronic hepatitis-C-virus-infected patients to interferon-based therapy. *J Gastroenterol Hepatol* 2011; **26**: 55-62 [PMID: 21175794 DOI: 10.1111/j.1440-1746.2010.06319.x]
- 21 **Askar M**, Avery R, Corey R, Lopez R, Thomas D, Pidwell D, Eghtesad B, Miller C, Fung J, Zein NN. Lack of killer immunoglobulin-like receptor 2DS2 (KIR2DS2) and KIR2DL2 is associated with poor responses to therapy of recurrent hepatitis C virus in liver transplant recipients. *Liver Transpl* 2009; **15**: 1557-1563 [PMID: 19877200 DOI: 10.1002/lt.21878]
- 22 **Carneiro VL**, Lemaire DC, Bendicho MT, Souza SL, Cavalcante LN, Angelo AL, Freire SM, Mendes CM, Santana N, Lyra LG, Lyra AC. Natural killer cell receptor and HLA-C gene polymorphisms among patients with hepatitis C: a comparison between sustained virological responders and non-responders. *Liver Int* 2010; **30**: 567-573 [PMID: 20456039 DOI: 10.1111/j.1478-3231.2010.02212.x]
- 23 **Khakoo SI**, Thio CL, Martin MP, Brooks CR, Gao X, Astemborski J, Cheng J, Goedert JJ, Vlahov D, Hilgartner M, Cox S, Little AM, Alexander GJ, Cramp ME, O'Brien SJ, Rosenberg WM, Thomas DL, Carrington M. HLA and NK cell inhibitory receptor genes in resolving hepatitis C virus infection. *Science* 2004; **305**: 872-874 [PMID: 15297676 DOI: 10.1126/science.1097670]
- 24 **Fabris C**, Falletti E, Cussigh A, Bitetto D, Fontanini E, Big-nulin S, Cmet S, Fornasiere E, Fumolo E, Fangazio S, Cerutti A, Minisini R, Pirisi M, Toniutto P. IL-28B rs12979860 C/T allele distribution in patients with liver cirrhosis: role in the course of chronic viral hepatitis and the development of HCC. *J Hepatol* 2011; **54**: 716-722 [PMID: 21146242 DOI: 10.1016/j.jhep.2010.07.019]
- 25 **Nattermann J**, Vogel M, Nischalke HD, Danta M, Mauss S, Stellbrink HJ, Baumgarten A, Mayr C, Bruno R, Tural C, Klausen G, Clotet B, Naumann U, Lutz T, Rausch M, Schewe K, Bienek B, Haerter G, Sauerbruch T, Rockstroh JK, Spengler U. Genetic variation in IL28B and treatment-induced clearance of hepatitis C virus in HIV-positive patients with acute and chronic hepatitis C. *J Infect Dis* 2011; **203**: 595-601 [PMID: 21257738 DOI: 10.1093/infdis/jiq098]
- 26 **Rallón NI**, Naggie S, Benito JM, Medrano J, Restrepo C, Goldstein D, Shianna KV, Vispo E, Thompson A, McHutchison J, Soriano V. Association of a single nucleotide polymorphism near the interleukin-28B gene with response to hepatitis C therapy in HIV/hepatitis C virus-coinfected patients. *AIDS* 2010; **24**: F23-F29 [PMID: 20389235 DOI: 10.1097/QAD.0b013e3283391d6d]
- 27 **Bedossa P**, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. The METAVIR Cooperative Study Group. *Hepatology* 1996; **24**: 289-293 [PMID: 8690394 DOI: 10.1002/hep.510240201]
- 28 **Jones RN**, Neale ML, Beattie B, Westmoreland D, Fox JD. Development and application of a PCR-based method including an internal control for diagnosis of congenital cytomegalovirus infection. *J Clin Microbiol* 2000; **38**: 1-6 [PMID: 10618053]
- 29 **Tabll A**, Shoman S, Ghanem H, Nabil M, El Din NG, El Awady MK. Assessment of human cytomegalovirus co-infection in Egyptian chronic HCV patients. *Virol J* 2011; **8**: 343 [PMID: 21740595 DOI: 10.1186/1743-422X-8-343]
- 30 **Akuta N**, Suzuki F, Kawamura Y, Yatsuji H, Sezaki H, Suzuki Y, Hosaka T, Kobayashi M, Kobayashi M, Arase Y, Ikeda K, Kumada H. Predictive factors of early and sustained responses to peginterferon plus ribavirin combination therapy in Japanese patients infected with hepatitis C virus genotype 1b: amino acid substitutions in the core region and low-density lipoprotein cholesterol levels. *J Hepatol* 2007; **46**: 403-410 [PMID: 17126448 DOI: 10.1016/j.jhep.2006.09.019]
- 31 **Akuta N**, Suzuki F, Sezaki H, Suzuki Y, Hosaka T, Someya T, Kobayashi M, Saitoh S, Watahiki S, Sato J, Kobayashi M, Arase Y, Ikeda K, Kumada H. Predictive factors of virological non-response to interferon-ribavirin combination therapy for patients infected with hepatitis C virus of genotype 1b and high viral load. *J Med Virol* 2006; **78**: 83-90 [PMID: 16299715 DOI: 10.1002/jmv.20507]

- 32 **Enomoto N**, Sakuma I, Asahina Y, Kurosaki M, Murakami T, Yamamoto C, Ogura Y, Izumi N, Marumo F, Sato C. Mutations in the nonstructural protein 5A gene and response to interferon in patients with chronic hepatitis C virus 1b infection. *N Engl J Med* 1996; **334**: 77-81 [PMID: 8531962 DOI: 10.1056/NEJM199601113340203]
- 33 **El-Shamy A**, Nagano-Fujii M, Sasase N, Imoto S, Kim SR, Hotta H. Sequence variation in hepatitis C virus nonstructural protein 5A predicts clinical outcome of pegylated interferon/ribavirin combination therapy. *Hepatology* 2008; **48**: 38-47 [PMID: 18537193 DOI: 10.1002/hep.22339]
- 34 **Welzel TM**, Morgan TR, Bonkovsky HL, Naishadham D, Pfeiffer RM, Wright EC, Hutchinson AA, Crenshaw AT, Bashirova A, Carrington M, Dotrang M, Sterling RK, Lindsay KL, Fontana RJ, Lee WM, Di Bisceglie AM, Ghany MG, Gretch DR, Chanock SJ, Chung RT, O'Brien TR. Variants in interferon-alpha pathway genes and response to pegylated interferon-Alpha2a plus ribavirin for treatment of chronic hepatitis C virus infection in the hepatitis C antiviral long-term treatment against cirrhosis trial. *Hepatology* 2009; **49**: 1847-1858 [PMID: 19434718 DOI: 10.1002/hep.22877]
- 35 **Sarrazin C**, Susser S, Doehring A, Lange CM, Müller T, Schlecker C, Herrmann E, Lötsch J, Berg T. Importance of IL28B gene polymorphisms in hepatitis C virus genotype 2 and 3 infected patients. *J Hepatol* 2011; **54**: 415-421 [PMID: 21112657 DOI: 10.1016/j.jhep.2010.07.041]
- 36 **Miller DM**, Cebulla CM, Sedmak DD. Human cytomegalovirus inhibition of major histocompatibility complex transcription and interferon signal transduction. *Curr Top Microbiol Immunol* 2002; **269**: 153-170 [PMID: 12224507 DOI: 10.1007/978-3-642-59421-2_10]
- 37 **Miller DM**, Zhang Y, Rahill BM, Waldman WJ, Sedmak DD. Human cytomegalovirus inhibits IFN-alpha-stimulated antiviral and immunoregulatory responses by blocking multiple levels of IFN-alpha signal transduction. *J Immunol* 1999; **162**: 6107-6113 [PMID: 10229853]
- 38 **Scott J**, Holte S, Urban T, Burgess C, Coppel E, Wang C, Corey L, McHutchison J, Goldstein D. IL28B genotype effects during early treatment with peginterferon and ribavirin in difficult-to-treat hepatitis C virus infection. *J Infect Dis* 2011; **204**: 419-425 [PMID: 21742841 DOI: 10.1093/infdis/jir264]
- 39 **Lutz P**, Wasmuth JC, Nischalke HD, Vidovic N, Grünhage F, Lammert F, Oldenburg J, Rockstroh JK, Sauerbruch T, Spengler U. Progression of liver fibrosis in HIV/HCV genotype 1 co-infected patients is related to the T allele of the rs12979860 polymorphism of the IL28B gene. *Eur J Med Res* 2011; **16**: 335-341 [PMID: 21813376 DOI: 10.1186/2047-783X-16-8-335]

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Lubiprostone induced ischemic colitis

Muhammed Sherid, Humberto Sifuentes, Salih Samo, Parakkal Deepak, Subbaramiah Sridhar

Muhammed Sherid, Salih Samo, Department of Internal Medicine, Division of Gastroenterology, Saint Francis Hospital, Evanston, IL 60202, United States

Humberto Sifuentes, Subbaramiah Sridhar, Section of Gastroenterology and Hepatology, Medical College of Georgia, Georgia Health Sciences University, Augusta, GA 30912, United States

Parakkal Deepak, Department of Internal Medicine, Division of Gastroenterology, NorthShore University HealthSystem, Highland Park, IL 60035, United States

Author contributions: Sherid M contributed to Study design, case presenter, literature review, data collection, data analysis, initial manuscript writing, manuscript review, approval of final version; Sifuentes H, Samo S, Deepak P, Sridhar S contributed to study design, literature review, data collection, data analysis, initial manuscript writing, manuscript review, approval of final version.

Correspondence to: Subbaramiah Sridhar, MBBS, MPH, FRCP (Edin), FRCP (Glasg), FRSS (Lond), FRCPC (Med and Gastro), FASGE, FACP, FACG, FASLM and S (United States), FAAG, MAAG, Professor of Medicine, Section of Gastroenterology and Hepatology, Medical College of Georgia, Georgia Health Sciences University, Augusta, GA 30912, United States. ssridhar@georgiahealth.edu

Telephone: +1-706-7212238 Fax: +1-706-7210331

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Abstract

Ischemic colitis accounts for 6%-18% of the causes of acute lower gastrointestinal bleeding. It is often multifactorial and more commonly encountered in the elderly. Several medications have been implicated in the development of colonic ischemia. We report a case of a 54-year old woman who presented with a two-hour history of nausea, vomiting, abdominal pain, and bloody stool. The patient had recently used lubiprostone with close temporal relationship between the increase in the dose and her symptoms of rectal bleeding. The radiologic, colonoscopic and histopathologic findings were all consistent with ischemic colitis. Her condition improved without any serious complications after the cessation of lubiprostone. This is the first reported case of ischemic

colitis with a clear relationship with lubiprostone (Naranjo score of 10). Clinical vigilance for ischemic colitis is recommended for patients receiving lubiprostone who are presenting with abdominal pain and rectal bleeding.

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Key words: Lubiprostone; Ischemic colitis; Gastrointestinal bleeding; Rectal bleeding; Colonoscopy

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INTRODUCTION

Ischemic colitis in patients with nonocclusive vascular disease is thought to result from a transient low flow state through an inadequate vascular system. Advanced age, aortic surgery, diabetes mellitus, hypertension, and peripheral vascular disease have been also suggested to be predisposing factors for ischemic colitis^[1]. It occurs more often in the splenic flexure and rectosigmoid junction which are known as the watershed areas. These areas are the distributions of the superior mesenteric artery and the inferior mesenteric artery for the splenic flexure and between the inferior mesenteric artery and the internal iliac artery for the rectosigmoid junction^[1].

The pathophysiology of developing ischemic colitis includes transient hypoperfusion that involve segments of the colon. Affected patients are often elderly or have a cardiovascular condition (congestive heart failure or an arrhythmia) which puts them at risk to develop a transient reduction in bowel perfusion. Less common causes of ischemic colitis include hypercoagulable states, vasculitis, embolus, colonic obstruction (strictures, colon cancer, diverticulitis)^[1,2]. Other causes include medications and substances such as amphetamines, alosetron, catechol-

amines (epinephrine, norepinephrine), cocaine, digitalis, nonsteroidal anti-inflammatory drugs, pseudoephedrine and triptans, *etc*^[3].

Lubiprostone was initially approved in 2006 by the US Food and Drug Administration for the treatment of idiopathic chronic constipation at a dose of 24 mcg twice daily^[4,6]. In 2008, it was approved for women with constipation predominant irritable bowel syndrome at dose of 8 mcg twice daily^[4,5,7]. Lubiprostone is a prostaglandin E1 derivative which acts by activating chloride channel type-2 (CLC-2) in the apical membrane of the intestinal epithelial cells. Consequently, it produces chloride rich fluid into the intestinal lumen with a passive sodium and water shift which increases the liquidity of the stool and promotes the gastrointestinal tract motility which in turn accelerates the bowel transit^[8]. Lubiprostone acts locally on the bowel epithelium and is minimally absorbed. It is well known to cause nausea, diarrhea, abdominal distention, and abdominal pain. Here we describe the first case report of ischemic colitis associated with lubiprostone usage.

CASE REPORT

A 54-year old female presented with a two-hour history of nausea, non-bloody, non-bilious vomiting associated with crampy generalized abdominal pain located to the epigastrium and left upper quadrant. She did not have any bowel movements 3 d prior to her presentation. In the emergency department, she had one bowel movement with hard stool followed by 6 watery, bloody stools. She did not have a fever or urinary symptoms. She did not have a history of recent antibiotic use, travel outside the United States, recent diet changes, or history of symptoms suggestive of inflammatory bowel disease. She suffers from diabetes mellitus, hypertension, hypercholesterolemia, hypothyroidism and chronic constipation. Her current medications included metformin (taking for 8 years), lisinopril (taking for 3 years), atorvastatin (taking for 4 years) and replacement thyroxine (taking for 4 years). She did not smoke or use illicit drugs.

On admission, her vital signs were essentially normal with a blood pressure of 134/70 mmHg without orthostatic changes, a heart rate of 61 bpm, temperature of 97.7 F°, respiratory rate of 16/min, oxygen saturation of 97% on room air, height of 162 cm, weight of 60.8 kg, and body mass index of 23.2. She did not have any episodes of hypotension either in the emergency department or during her hospitalization. Physical examination revealed a soft abdomen with mild epigastric and left upper quadrant tenderness. There was no distension, guarding, rebound tenderness, or organomegaly. The bowel sounds were normal. Rectal exam was unremarkable except for positive occult blood. The remainder of physical exam revealed no findings.

The laboratory studies were unremarkable including complete blood count, comprehensive metabolic panel, amylase, lipase, urinalysis, and urine toxicology. The stool

studies were negative for ova, parasites, clostridium difficile, and pathogenic bacteria. The hemoglobin A1c was 6.5%. A plain abdominal X-ray was unremarkable. A computed tomography (CT) scan of the abdomen and pelvis revealed thickening of the colonic wall involving the transverse and the proximal portions of the descending colon near the splenic flexure (Figure 1A). A colonoscopic examination showed mucosal changes consistent with ischemic colitis (mucosal edema and hyperemia with areas of hemorrhage, erosions, and frank ulcerations) affecting the descending colon, between 30 to 40 cm from the anal verge (Figure 1B-D). The histopathology examination showed ischemic loss of crypts with acute hemorrhage in lamina propria associated with acute fibrinopurulent exudate at colonic epithelial surface, consistent with ischemic colitis (Figure 1E, F).

A diagnosis of ischemic colitis was made based on the above findings. To clarify the etiology, a detailed history was taken including the use of over the counter and herbal medications. She stated that she had been taking lubiprostone for a period of 2 mo prior to her admission after she failed to respond to conventional laxatives for chronic constipation. The patient was initially started on 24 mcg of lubiprostone, but several hours after taking the drug, she developed nausea, vomiting and abdominal pain with bloody diarrhea which lasted for two days. She did not undergo any investigations, but her dose of lubiprostone was changed to 8 mcg daily as needed after she called her primary care physician. Twenty-four hours prior to her current admission she had taken a total of 24 mcg of lubiprostone. Her symptoms developed two hours after the last dose. This temporal relationship between the recurrence of her symptoms and the rechallenge of the drug confirms the diagnosis of ischemic colitis in the light of absence of other causes of ischemic colitis. This case scored a 10 on the Naranjo Nomogram for adverse drug reactions which places lubiprostone as a definitive cause of ischemic colitis in this case (definitive ≥ 9). The patient was treated with discontinuation of lubiprostone, intravenous fluids, bowel rest and intravenous antibiotics. Her hospitalization course was uneventful and she was discharged home without any complications 4 d later.

DISCUSSION

Ischemic colitis accounts for 6%-18% of the causes of acute lower gastrointestinal bleeding^[2]. The causes of ischemic colitis vary from systemic hypotension, aortoiliac surgery, atherosclerosis, thromboembolic events, vasculitis, and medications^[9].

The medications that have been implicated in the development of ischemic colitis have different mechanisms including decreasing blood flow *via* systemic hypotension with angiotensin-converting enzyme inhibitors, vasospasm with pseudoephedrine, thromboembolic events with oral contraceptives, vasculitis secondary to gold salts, and increasing intracolonic pressures with alos-

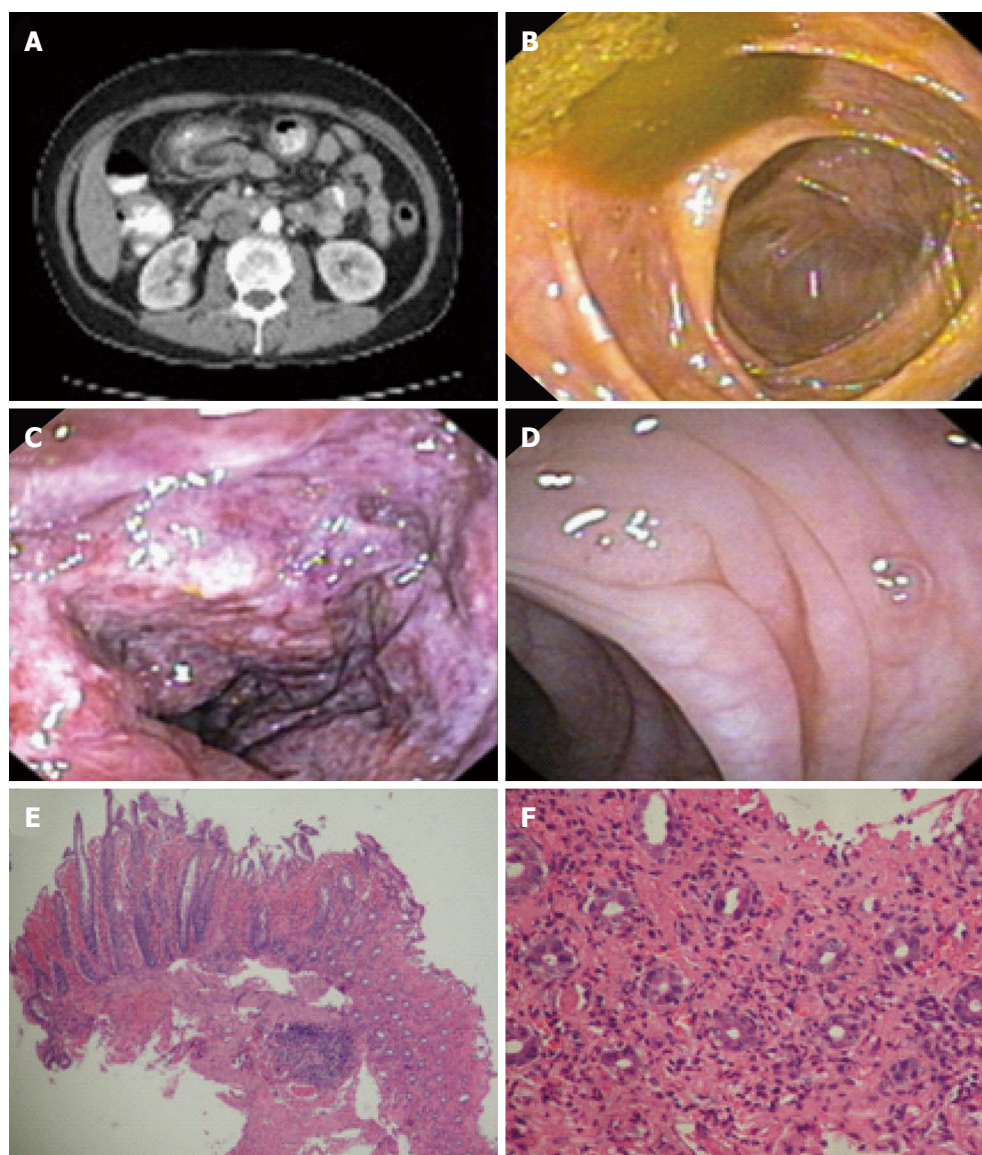


Figure 1 The laboratory studies results. A: Computed tomography scan shows thickening of the colonic wall involving the splenic flexure of the colon; B: Normal mucosa in the right colon (appendiceal orifice); C: Ischemic colitis features in the splenic flexure of the colon; D: Normal mucosa in sigmoid with a small polyp; E, F: Histopathology shows ischemic loss of crypts and acute hemorrhage in lamina propria (acute hemorrhagic necrosis replacing glands) associated with acute fibrinopurulent exudate at colonic epithelial surface, consistent with ischemic colitis.

etron^[3]. The mechanism of some drugs reported to cause ischemic colitis has not yet determined. Table 1 shows a list of medications and their mechanisms for causing ischemic colitis^[3].

Many case reports have linked ischemic colitis and other unusual associations such as herbal remedies (ma huang/ephedra), weight loss medications such as phentermine, following screening colonoscopies, certain chemotherapy agents such as bevacizumab and irinotecan, hepatitis C therapy with peginterferon and ribavirin, scuba diving, flying, snake bites, acute carbonic monoxide poisoning, using electrical muscle stimulation devices on the abdominal wall, and following long distance running^[3].

Lubiprostone has been shown to improve symptoms in patients with chronic idiopathic constipation. In a 48

wk prospective, multicenter, open-labeled trial performed by Lembo *et al*^[10] patients with chronic idiopathic constipation were taking lubiprostone 24 mcg twice daily as needed and the most common side effects were nausea (19.8%), diarrhea (9.7%), abdominal distension (6.9%), headache (6.9%), and abdominal pain (5.2%).

Lubiprostone has also been approved for the treatment of constipation predominant irritable bowel syndrome (IBS-C). In a randomized, placebo controlled trial, the overall rate of response based on a patient ranked assessment tool was significantly higher in the lubiprostone group using 8 mcg twice daily compared to placebo. The side effects were similar in both groups^[11].

In a phase 2, placebo controlled trial by Johanson *et al*^[12] lubiprostone was superior to placebo in IBS-C regarding the severity of constipation, stool consistency,

Table 1 Medications associated with ischemic colitis

Agent	Mechanism
Amphetamines	Vasoconstriction
Alosetron	
Catecholamines (epinephrine, norepinephrine)	
Cocaine	
Cyclosporine	
Digitalis	
Dopamine	
Ergot derivatives	
Nonsteroidal anti-inflammatory drugs	
Pseudoephedrine	
Triptans (Naratriptan, Rizatriptan, Sumatriptan)	
Vasopressin and vasopressin analogues	
Glycerin enema	Local vasospasm effect
Phosphosoda solution	
Angiotensin-converting enzyme inhibitors	Systemic hypotension
Antipsychotic (chlorpromazine)	
Beta blockers	
Barbiturates	
Diuretics	
Interleukin-2	
Tricyclic antidepressants	
Amphetamines	Vasculitis
Gold compounds	
Estrogens	
Progestational agents	Increased intracolonic pressure
Alosetron	
Danazol	Undetermined
Glycerin enema	
Carboplatin	
Flutamide	
Glutaraldehyde	
Hyperosmotic saline laxatives	
Interferon-alpha	
Mycophenolate mofetil	
Paclitaxol	
Simvastatin	
Tegaserod	

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frequency, straining, bloating, and abdominal pain in 1, 2, and 3 mo follow ups. Both diarrhea and nausea occurred more often in the lubiprostone group, especially with 24 mcg twice daily compared to 8 mcg twice daily.

Lubiprostone is a bicyclic fatty acid, derived from prostaglandin E1. It is a member of a class of drugs called prostones which, unlike prostaglandins, are not believed to cause direct smooth muscle contraction. It is metabolized in the upper part of the gastrointestinal tract by the microsomal carbonyl reductase system. Lubiprostone acts locally in gastrointestinal epithelium and its active metabolite, M3, absorbs systemically. The half-life of M3 is 0.9-1.4 h. M3 is used for the pharmacokinetic parameters of lubiprostone because the drug itself has limited systemic absorption and low bioavailability^[4,8].

Lubiprostone mainly acts on CLC-2 activators in the intestinal epithelium which is one of the nine types of chloride channels in the body which also includes the cystic fibrosis transmembrane conductance regulator (CFTR) (Figure 2)^[13].

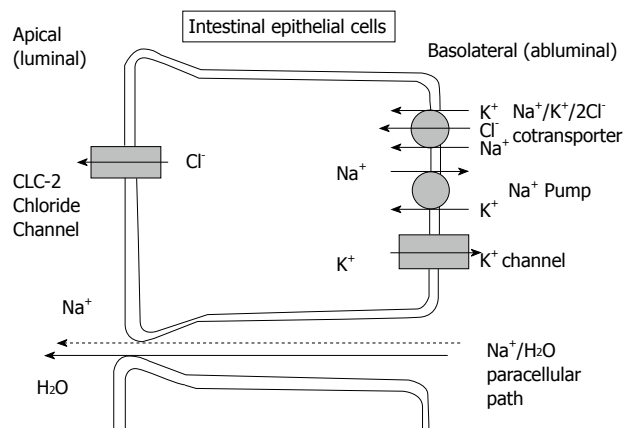


Figure 2 Mechanism of action of lubiprostone across epithelial intestinal cells^[8].

Activation of CLC-2 in the apical membrane of intestinal epithelial cells causes an active secretion of chloride ions from cells into the intestinal lumen followed by a passive secretion of sodium ions and water which results in increased isotonic fluid in the lumen which promotes small bowel and colonic transit by stimulating stretch receptors and smooth muscles in the intestine (Figure 2)^[8].

There is now controversy whether lubiprostone activates CLC-2 in the basolateral or apical aspect of intestinal epithelium, whether it activates mainly CFTR or CLC-2, or whether it directly stimulates smooth muscles *via* prostaglandin receptors^[8]. Bao *et al*^[14] showed in an experimental study that lubiprostone activates CLC-2 at low concentrations but activates CFTR at higher concentrations. It has also been shown that lubiprostone weakly activates prostaglandin receptors in different levels of the GI tract which could be responsible for some side effects such as nausea^[15].

In our patient, despite having other risk factors for ischemic colitis such as chronic constipation which has been shown to increase the risk of ischemic colitis by 2.7 folds in some studies^[16], the close temporal relationship between lubiprostone and occurrence of the symptoms of ischemic colitis makes the drug the most likely culprit of this event. When applying the Naranjo Nomogram in our patient, a score of 10 was obtained indicating a definite likelihood of lubiprostone as the cause of ischemic colitis in this case (Table 2).

To the best of our knowledge, this is the first report of lubiprostone induced ischemic colitis. We report that a direct causal relation by exclusion of other causes, the disappearance of symptoms after cessation of lubiprostone, recurrence of symptoms with self rechallenge, and a Naranjo Nomogram score of 10 all go in favor of the drug-induced ischemic colitis. The actual mechanism of lubiprostone causing ischemic colitis is not known, however, fast fluid shift into the intestinal lumen causing local hypoperfusion, rapid increase of the intracolonic pressure due to rapid fluid secretion into the bowel, or direct vasoconstriction caused by lubiprostone in higher doses

Table 2 Naranjo adverse drug reaction nomogram in our patient

	Yes	No	Our patient
Are there previous conclusive reports on this reaction?	1	0	0
Did the adverse event appear after the suspected drug was administered?	2	-1	2
Did the adverse reaction improve when the drug was discontinued or a specific antagonist was administered?	1	0	1
Did the adverse reaction reappear when the drug was readministered?	2	-1	2
Are there alternative causes (other than the drug) that could have, on their own, caused the reaction?	-1	2	2
Did the reaction appear when a placebo was given?	-1	1	1
Was the drug detected in the blood (or other fluids) in concentration known to be toxic?	1	0	0
Was the reaction more severe when the dose was increased or less severe when dose was decreased?	1	0	0
Did the patient have a similar reaction to the same or similar drugs in any previous exposure?	1	0	1
Was the adverse event confirmed by any objective evidence?	1	0	1
			10

Definite: score ≥ 9 ; Probable: 5-8; Possible: 1-4; Doubtful: ≤ 0 .

by stimulating prostaglandin receptors are the proposed pathophysiological mechanisms. In conclusion, lubiprostone may cause ischemic colitis, especially in some susceptible persons and greater awareness of this side effect is warranted with the increasing use of lubiprostone.

REFERENCES

- Theodoropoulou A**, Koutroubakis IE. Ischemic colitis: clinical practice in diagnosis and treatment. *World J Gastroenterol* 2008; **14**: 7302-7308 [PMID: 19109863 DOI: 10.3748/wjg.14.7302]
- Strate LL**. Lower GI bleeding: epidemiology and diagnosis. *Gastroenterol Clin North Am* 2005; **34**: 643-664 [PMID: 16303575 DOI: 10.1016/j.gtc.2005.08.007]
- Sherid M**, Ehrenpreis ED. Types of colitis based on histology. *Dis Mon* 2011; **57**: 457-489 [PMID: 21944389 DOI: 10.1016/j.disamonth.2011.05.004]
- AMITIZA®** (lubiprostone) [package insert]. Bethesda, MD: Sucampo Pharmaceuticals, 2009
- Schiller LR**, Camilleri M. Lubiprostone: viewpoints. *Drugs* 2006; **66**: 880-881 [PMID: 16706563 DOI: 10.2165/00003495-200666060-00016]
- Ford AC**, Suares NC. Effect of laxatives and pharmacological therapies in chronic idiopathic constipation: systematic review and meta-analysis. *Gut* 2011; **60**: 209-218 [PMID: 21205879 DOI: 10.1136/gut.2010.227132]
- Lang L**. The Food and Drug Administration approves lubiprostone for irritable bowel syndrome with constipation. *Gastroenterology* 2008; **135**: 7 [PMID: 18541153 DOI: 10.1053/j.gastro.2008.06.004]
- Lunsford TN**, Harris LA. Lubiprostone: evaluation of the newest medication for the treatment of adult women with constipation-predominant irritable bowel syndrome. *Int J Womens Health* 2010; **2**: 361-374 [PMID: 21151683 DOI: 10.2147/IJWH.S4537]
- Stamatakis M**, Douzinas E, Stefanaki C, Petropoulou C, Arampatzis H, Safioleas C, Giannopoulos G, Chatziconstantinou C, Xiromeritis C, Safioleas M. Ischemic colitis: surging waves of update. *Tohoku J Exp Med* 2009; **218**: 83-92 [PMID: 19478463 DOI: 10.1620/tjem.218.83]
- Lembo AJ**, Johanson JF, Parkman HP, Rao SS, Miner PB, Ueno R. Long-term safety and effectiveness of lubiprostone, a chloride channel (CIC-2) activator, in patients with chronic idiopathic constipation. *Dig Dis Sci* 2011; **56**: 2639-2645 [PMID: 21769655 DOI: 10.1007/s10620-011-1801-0]
- Drossman DA**, Chey WD, Johanson JF, Fass R, Scott C, Panas R, Ueno R. Clinical trial: lubiprostone in patients with constipation-associated irritable bowel syndrome--results of two randomized, placebo-controlled studies. *Aliment Pharmacol Ther* 2009; **29**: 329-341 [PMID: 19006537 DOI: 10.1111/j.1365-2036.2008.03881.x]
- Johanson JF**, Drossman DA, Panas R, Wahle A, Ueno R. Clinical trial: phase 2 study of lubiprostone for irritable bowel syndrome with constipation. *Aliment Pharmacol Ther* 2008; **27**: 685-696 [PMID: 18248656 DOI: 10.1111/j.1365-2036.2008.03629.x]
- Lipecta J**, Bali M, Thomas A, Fanen P, Edelman A, Fritsch J. Distribution of CIC-2 chloride channel in rat and human epithelial tissues. *Am J Physiol Cell Physiol* 2002; **282**: C805-C816 [PMID: 11880269 DOI: 10.1152/ajpcell.00291.2001]
- Bao HF**, Liu L, Self J, Duke BJ, Ueno R, Eaton DC. A synthetic prostone activates apical chloride channels in A6 epithelial cells. *Am J Physiol Gastrointest Liver Physiol* 2008; **295**: G234-G251 [PMID: 18511742 DOI: 10.1152/ajpgi.00366.2007]
- Bassil AK**, Borman RA, Jarvie EM, McArthur-Wilson RJ, Thangiah R, Sung EZ, Lee K, Sanger GJ. Activation of prostaglandin EP receptors by lubiprostone in rat and human stomach and colon. *Br J Pharmacol* 2008; **154**: 126-135 [PMID: 18332851 DOI: 10.1038/bjp.2008.84]
- Suh DC**, Kahler KH, Choi IS, Shin H, Kralstein J, Shetzline M. Patients with irritable bowel syndrome or constipation have an increased risk for ischaemic colitis. *Aliment Pharmacol Ther* 2007; **25**: 681-692 [PMID: 17311601 DOI: 10.1111/j.1365-2036.2007.03250.x]

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Lesson from an intriguing case of cryoglobulinemia

Michele Barone, Raffaele Licinio, Annabianca Amoruso, Maria Teresa Viggiani, Angela Maria Vittoria Larocca, Alfredo Di Leo

Michele Barone, Raffaele Licinio, Annabianca Amoruso, Maria Teresa Viggiani, Alfredo Di Leo, Department of Emergency and Organ Transplantation, University of Bari, 70100 Bari, Italy

Angela Maria Vittoria Larocca, Department of Biomedical Science and Human Oncology, University of Bari, 70100 Bari, Italy
Author contributions: Barone M, Licinio R, Amoruso A, Viggiani MT and Larocca AMV contributed equally to this work; Di Leo A approved the final version.

Correspondence to: Alfredo Di Leo, Professor, Department of Emergency and Organ Transplantation, University of Bari, Policlinico G. Cesare Sq 11, 70124 Bari, Italy. alfredo.dileo@uniba.it
Telephone: +39-80-5592577 Fax: +39-80-5593251

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Abstract

Cryoglobulinemia is a pathological condition usually associated with hepatitis C virus (HCV) chronic liver disease and less commonly with autoimmune or lymphoproliferative disorders. The possible association of cryoglobulinemia with hepatitis B virus (HBV) infection is not widely accepted. In our patient, serum negativity for HCV markers initially led us to consider two other causes of cryoglobulinemia. Myelodysplastic disorders were excluded on the basis of hematological studies, while serum markers for active HBV infection were positive. Surprisingly, the detection of HCV RNA in the cryocrit, even in the absence of anti-HCV antibodies, suggested a pathogenetic role of HCV in this case of cryoglobulinemia. Negative "first level" tests for HCV in the serum do not completely exclude HCV involvement in the pathogenesis of cryoglobulinemia. Analysis of the cryoprecipitate is always essential for diagnosis.

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Key words: Cryocrit; Hepatitis B virus; Hepatitis C virus; Myelodysplastic disorders; Cryoglobulinemia

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INTRODUCTION

Cryoglobulinemia is a pathological condition usually associated with hepatitis C virus (HCV) chronic liver disease^[1,2]. Cases of cryoglobulinemia associated with autoimmune or lymphoproliferative diseases are less common^[2], while the association with hepatitis B virus (HBV) infection is not widely accepted, the HBV virus being mostly associated with other immunocomplex-related disorders^[3-5].

Cryoglobulins are proteins that can precipitate at low temperatures ($< 4^{\circ}\text{C}$). The final product of this precipitate, termed the cryocrit, can be characterized on the basis of its composition: polyclonal immunoglobulins, monoclonal immunoglobulins or the presence of rheumatoid activity (RA). Three types of cryoglobulinemia have been identified, namely: type 1, including monoclonal cryoglobulins (IgM, IgG and IgA) without any RA; type 2, that includes a monoclonal component (usually IgM with RA) and a polyclonal component (usually IgM/IgG); type 3, that includes several polyclonal components and a component with RA (usually IgG/IgM)^[6].

Normally, cryoglobulinemia type 1 is associated with lymphoproliferative/myelodysplastic diseases such as multiple myeloma, Waldenstrom macroglobulinemia, chronic lymphatic leukemia, non-Hodgkin lymphoma, *etc.* Cryoglobulinemia type 2 is associated not only with lymphoproliferative diseases and plasma cellular dyscrasias but also with infectious and autoimmune diseases (rheumatoid arthritis, Sjogren's syndrome, *etc.*). Finally, cryoglobulinemia type 3 is frequently associated with autoimmune or infectious diseases. The mechanism responsible

for cryoglobulin formation during lymphoproliferative or autoimmune diseases is known, but the etiopathogenesis of the forms defined as “essential cryoglobulinemias”, which occur as isolated events, has still to be clarified. Among the essential forms, the most common variant is cryoglobulinemia type 2^[2]. The amount of cryoglobulins, expressed as a percentage of the serum volume, can vary significantly among patients and may change over time. It is therefore useful to evaluate the cryocrit, also during follow-up, in order to assess prognosis and treatment. Cryoglobulinemia characterization by protein electrophoresis and immunofixation is equally important to define the type^[2]. In 1966, Meltzer *et al*^[7] described a clinical triad characterized by palpable purpura, arthralgia and asthenia associated with nephropathy and neuropathy. It is widely accepted that presentation of the complete triad is rare in clinical practice. In fact, most patients are asymptomatic and purpura, often fleeting, is the only clinical manifestation. The frequent association of HCV infection with the great majority of cryoglobulins, initially defined as essential, suggested the involvement of this virus in the pathogenesis of mixed cryoglobulinemia. Actually, mixed cryoglobulinemia (type 2 or 3) is found in 50% of patients with chronic HCV infection^[1,4]. Therefore, in patients with mixed cryoglobulinemia, the possibility of HCV infection should always be considered, and serum tests should be performed for the detection of anti-HCV antibodies and HCV RNA.

CASE REPORT

Our patient was in apparent good health until the age of 73 when he underwent percutaneous angioplasty for acute non-Q myocardial infarction, after which he was prescribed anticoagulant and antihypertensive therapy. At the age of 77, he had an outpatient medical visit because of the first manifestation of purpura of the lower limbs, arthralgia and a sense of postural instability, raising the suspicion of a cryoglobulinemic syndrome. At this time, the cryocrit was positive (35%), and the patient underwent several serological and molecular investigations that excluded HCV (negative anti-HCV and HCV RNA) and HIV infection and demonstrated chronic HBV infection (surface antigen of the hepatitis B virus (HBsAg) 11 700 IU/mL, anti-HBsAg negative, hepatitis B core antibody positive, anti-hepatitis Be antibody positive, HBV-DNA 2 410 000 IU/mL, anti-HDV negative). Immediately after, he was admitted to our unit because of the onset of a hypertensive crisis that was not controlled by the administration of calcium antagonists, beta-blockers and loop diuretics. Renal function monitoring demonstrated the following values: creatinine 1.85 mg/dL, creatinine clearance 20 mL/min and 24-h proteinuria 1.3 g/24 h. Echotomographic examination showed ultrasonographic signs of cirrhosis, and hepatic elastometry yielded a stiffness value of 47.2 kPa. Other laboratory tests demonstrated aspartate aminotransferase \times 6.7 upper limit of normal (ULN), alanine aminotransferase \times 4.9 ULN and

gamma-glutamyl transpeptidase \times 5.2 ULN, and total bilirubin 1.8 mg/dL. On the basis of these data, the patient was started on antiviral treatment (0.5 mg entecavir every 72 h) and prednisone, 50 mg/d, to be tapered to 10 mg/d. Although the hypertensive crisis could presumably be ascribed to a cryoglobulin-induced nephropathy attributable to HBV, considering that the serum β 2-microglobulin levels were markedly increased (7.2 mg/L, < 2 ULN) and gamma-globulin showed a monoclonal peak, we also considered the possibility of cryoglobulinemia associated with a myeloproliferative disorder. The characteristics of the cryoprecipitate were therefore evaluated in order to define the type of cryoglobulinemia. We found rheumatoid factor positivity, lower C3 and C4 levels, 24% cryocrit, while serum immunofixation showed a K monoclonal IgM component, and cryocrit immunofixation a K monoclonal IgM component with polyclonal IgG. On the basis of these elements we made a diagnosis of mixed cryoglobulinemia type 2, that did not exclude a myeloproliferative disorder, so a peripheral blood smear and bone marrow biopsy were performed, and these excluded a myeloproliferative disease. During the period of hospitalization we observed an improvement in renal function (creatinine decreased to 1.4 mg/dL, 24-h creatinine clearance increased to 35 mL/min, and 24-h proteinuria decreased to 0.9 g) and a striking reduction of the cryocrit (6.1%). Surprisingly, cryoprecipitate characterization demonstrated the presence of HCV RNA (807 copies/mL, by m2000 Real Time System, Abbott, IL, United States, Figure 1) and for this reason, we rechecked the serum anti-HCV antibodies and HCV RNA, and the absence of both was confirmed (HCV RNA undetectable using 50 IU/mL as cut-off).

DISCUSSION

In the present case, the detection of HCV components in the cryocrit cast some doubt on the pathogenesis of the cryoglobulinemia that we initially attributed to HBV infection.

It has been reported that in chronic HCV patients with mixed cryoglobulinemia, HCV RNA may be undetectable in the plasma (due to the entrapment of HCV RNA in the cryoprecipitate) and the diagnosis of HCV infection is based on the presence of anti-HCV antibodies^[8]. However, in our patient anti-HCV antibodies were undetectable in the serum. This could be explained by interference between the two viruses that may even lead to inhibition of HCV replication in patients with chronic hepatitis C with a superimposed HBV infection^[9,10].

This experience suggests that negative tests for HCV detection in the serum (anti-HCV antibodies and HCV RNA) do not completely exclude the involvement of HCV in the pathogenesis of cryoglobulinemia, and show that analysis of the cryoprecipitate is always essential, since detection of HCV RNA in the cryoprecipitate is the most sensitive method. Once HCV infection has been excluded, investigations must take into account other causes

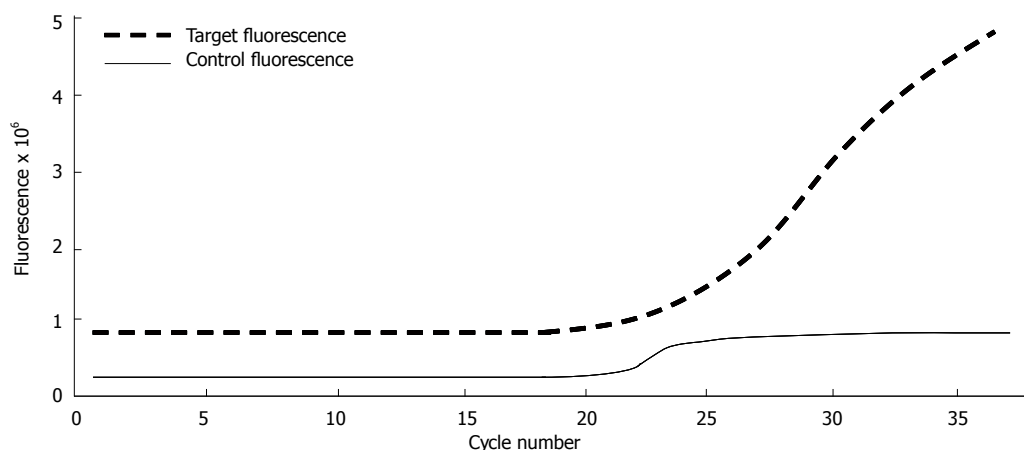


Figure 1 Real time polymerase chain reaction amplification plot for hepatitis C virus RNA. The presence of viral RNA started to become evident after 19.4 cycles and the amount of RNA was 807 copies/mL.

of cryoglobulinemia, such as autoimmune and myelo- or lymphoproliferative disorders and HBV infection.

REFERENCES

- 1 Ferri C, La Civita L, Longombardo G, Greco F, Bombardieri S. Hepatitis C virus and mixed cryoglobulinaemia. *Eur J Clin Invest* 1993; **23**: 399-405 [PMID: 8397090 DOI: 10.1111/j.1365-2362.1993.tb00782.x]
- 2 Trendelenburg M, Schifferli JA. Cryoglobulins are not essential. *Ann Rheum Dis* 1998; **57**: 3-5 [PMID: 9536813 DOI: 10.1136/ard.57.1.3]
- 3 Johnson RJ, Couser WG. Hepatitis B infection and renal disease: clinical, immunopathogenetic and therapeutic considerations. *Kidney Int* 1990; **37**: 663-676 [PMID: 1968522 DOI: 10.1038/ki.1990.32]
- 4 Liang TJ. Hepatitis B: the virus and disease. *Hepatology* 2009; **49**: S13-S21 [PMID: 19399811 DOI: 10.1002/hep.22881]
- 5 Cakir N, Pamuk ON, Umit H, Midilli K. Successful treatment with adefovir of one patient whose cryoglobulinemic vasculitis relapsed under lamivudine therapy and who was diagnosed to have HBV virologic breakthrough with YMDD mutations. *Intern Med* 2006; **45**: 1213-1215 [PMID: 17139120 DOI: 10.2169/internalmedicine.45.1816]
- 6 Brouet JC, Clauvel JP, Danon F, Klein M, Seligmann M. Biologic and clinical significance of cryoglobulins. A report of 86 cases. *Am J Med* 1974; **57**: 775-788 [PMID: 4216269 DOI: 10.1016/0002-9343(74)90852-3]
- 7 Meltzer M, Franklin EC, Elias K, McCluskey RT, Cooper N. Cryoglobulinemia--a clinical and laboratory study. II. Cryoglobulins with rheumatoid factor activity. *Am J Med* 1966; **40**: 837-856 [PMID: 4956871]
- 8 Bichard P, Ounanian A, Girard M, Baccard C, Rolachon A, Renversez JC, Cordonnier D, Seigneurin JM, Debru JL, Zarski JP. High prevalence of hepatitis C virus RNA in the supernatant and the cryoprecipitate of patients with essential and secondary type II mixed cryoglobulinemia. *J Hepatol* 1994; **21**: 58-63 [PMID: 7525697]
- 9 Alberti A, Pontisso P, Chemello L, Fattovich G, Benvegnù L, Belussi F, De Mitri MS. The interaction between hepatitis B virus and hepatitis C virus in acute and chronic liver disease. *J Hepatol* 1995; **22**: 38-41 [PMID: 7602074]
- 10 Sagnelli E, Coppola N, Messina V, Di Caprio D, Marrocco C, Marotta A, Onofrio M, Scolastico C, Filippini P. HBV superinfection in hepatitis C virus chronic carriers, viral interaction, and clinical course. *Hepatology* 2002; **36**: 1285-1291 [PMID: 12395342]

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Recurrent cervical esophageal stenosis after colon conduit failure: Use of myocutaneous flap

Young Jo Sa, Young Du Kim, Chi Kyung Kim, Jong Kyung Park, Seok Whan Moon

Young Jo Sa, Young Du Kim, Chi Kyung Kim, Seok Whan Moon, Department of Thoracic and Cardiovascular Surgery, St. Paul Hospital, The Catholic University of Korea, Seoul 130-709, South Korea

Jong Kyung Park, Department of General Surgery, St. Paul Hospital, The Catholic University of Korea, Seoul 130-709, South Korea

Author contributions: Sa YJ and Moon SW were responsible for research design and wrote the paper; Sa YJ, Kim YD, Park JK, and Moon SW performed surgical research; Sa YJ, Kim CK, and Moon SW analyzed and interpreted the data; Sa YJ and Moon SW designed the paper, drafted and revised the article, and obtained final approval.

Correspondence to: Seok Whan Moon, MD, PhD, Department of Thoracic and Cardiovascular Surgery, St. Paul Hospital, The Catholic University of Korea, Seoul 130-709, South Korea. swmoon@catholic.ac.kr

Telephone: +82-2-9582447 Fax: +82-2-9582447

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INTRODUCTION

A variety of total esophageal reconstruction procedures have been described; however, there is no universally-accepted method^[1]. Cervical reconstruction for esophageal stenosis of any etiology is complicated and can be more challenging in cases of recurrent cervical stenosis with previous colon conduit failure. Recently, we successfully treated upper cervical stenosis caused by corrosive strictures using colon conduits. However, these operations have been followed by recurrent stenosis. Many different approaches have been utilized in the treatment of cervical stenosis. These include local fasciocutaneous flaps, pedicled myocutaneous flaps, pedicled visceral flaps, free flaps, and free fasciocutaneous flaps^[2]. We present a successful patch-plasty with myocutaneous flap of sternocleidomastoid (SCM) muscle for treatment of recurrent cervical esophageal stenosis.

CASE REPORT

A 53-year-old male developed recurrent cervical esophageal stenosis 1.5 years after undergoing esophageal bypass surgery using a right colon conduit. He had received corrosive esophageal strictures three years earlier after a suicide attempt by potassium hydroxide ingestion. His presentation was complicated, with multi-level stenotic segments down the entire esophagus. He underwent

Abstract

A 53-year-old male developed cervical esophageal stenosis after esophageal bypass surgery using a right colon conduit. The esophageal bypass surgery was performed to treat multiple esophageal strictures resulting from corrosive ingestion three years prior to presentation. Although the patient underwent several endoscopic stricture dilatations after surgery, he continued to suffer from recurrent esophageal stenosis. We planned cervical patch esophagoplasty with a pedicled skin flap of sternocleidomastoid (SCM) muscle. Postoperative recovery was successful, and the patient could eat a solid meal without difficulty and has been well for 18 mo. SCM flap esophagoplasty is an easier and safer method of managing complicated and recurrent cervical esophageal strictures than other operations.

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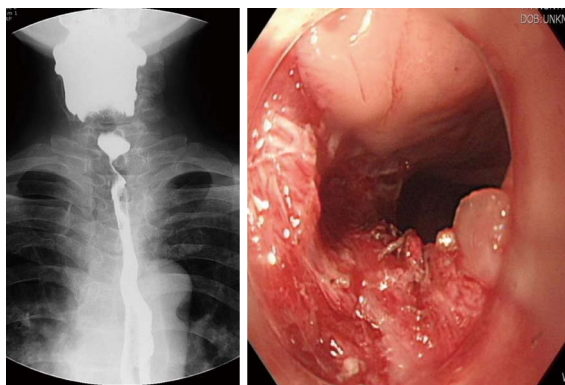


Figure 1 Esophagogram and endoscopic findings before patch esophagoplasty. The native esophagus was visualized with the esophagogram, but the colon conduit from bypass surgery was not visible. After insertion of an Endoscopic Varix Ligation tube, a stenotic lesion of the esophago-colonic anastomosis site was opened. Focal luminal stenosis by fibrosis was evident. Endoscopic dilatation was repeated using electrocauterization.

repeated balloon dilatation, but one cervical stricture progressed to near-complete obstruction just below the entrance of the esophagus as seen on endoscopy and barium esophagogram. The severity of stenosis below the proximal esophagus could not be evaluated. He had severe weight loss of 15 kg and required total parenteral nutrition, afterwards gaining back about 2.5 kg (43 kg and 173 cm, body mass index 14.37). Instead of feeding gastrostomy insertion, esophageal bypass surgery using a segment of the right colon was performed one year later. At surgery, the ileum was anastomosed with the cervical esophagus in a side-to-end manner, and its blood supply was preserved. Postoperative esophagogram showed that barium passed through the native esophagus and newly constructed colon conduit. The patient began to eat 7 d after surgery and returned home 14 d after surgery with a weight gain of 3 kg. On follow-up, he gained weight by additional 3 kg, but suffered from painful swallowing of solid food three mo after discharge. Follow-up endoscopy revealed cicatricial stenosis in the native esophagus proximal to the interposed colon, though the site of gastro-colon anastomosis was patent. Barium esophagogram showed mild stenosis at the level of the thoracic inlet and severe stenosis just above the anastomosis of the esophagus. The patient underwent repeated 7 times esophageal dilatation using Savary-Guillard or balloon dilators, and an episode of electrocauterization ablation to improve swallowing before the 2nd operation. Despite endoscopic dilatation using electrocauterization, he again developed pain on swallowing. Finally, he was admitted for surgical correction. We reviewed the esophagogram and endoscopic findings (Figure 1). The stenotic segment was 3 cm long and located in the proximal esophagus near the site of the previous esophago-colonic anastomosis. Fortunately, the lesion was easily opened with a dilator or balloon. This finding suggested that the stenotic segment was not caused by mechanical stenosis resulting from circumferential fibrosis, but was caused by twisted axis of esophageal anastomosis site possibly due to cicatri-

cial obstruction resulting from colon conduit failure. We planned to perform a patch enlargement operation with a myocutaneous flap of SCM muscle. Under endotracheal anesthesia, the patient was placed supine with neck extension and slight rotation for better exposure. A skin incision in the neck was made along the previous incision while preserving an elliptical skin island over the SCM muscle (Figure 2). Improved exposure of the stenotic cervical esophagus was obtained after SCM division with a skin island near the clavicle. There was a fibrotic band near the proximal esophagus instead of near the colon. The proximal esophagus and hypopharynx were exposed, and a small esophagotomy at the level of the cricoid cartilage was initially made and extended upward to the hypopharynx with a longitudinal incision of about 5 cm (Figure 2). The attached skin island of the SCM myocutaneous flap was rotated medially over the esophagopharyngeal opening and sutured in interrupted fashion with 3-0 black silk. The SCM muscle attached to the skin flap was then tacked into place around the esophagus and pharynx. A drain was placed, and the neck wound was closed after a skin tension-releasing procedure (Figure 2). Seven days later, an esophagogram showed good passage without tracheal aspiration (Figure 3). The patient could eat a solid meal without pain or resistance and returned home in ten days. He then returned to work and has been well with a gradual weight gain about 10 kg for 18 mo after the esophagoplasty.

DISCUSSION

Graft necrosis is the most dangerous complication of colon graft interposition with a reported mortality approaching 90% or more^[3]. Most instances of colonic graft ischemia are secondary to arterial insufficiency^[3]. Ischemic complications are expressed as frank colon necrosis, anastomotic leakage, and colonic stricture^[3]. Colonic segment necrosis and anastomotic leakage usually occur in the first postoperative week^[4], but in our case, localized strictures of the cervical region developed 1.5 years after the first bypass operation. The slow course is likely explained by slowly progressive ischemia from graft torsion, resulting in a non-necrotic colonic cicatricial stricture rather than colonic necrosis. This may be the cause of esophageal obstruction in our case. This cicatricial change in the esophageal anastomosis site might twist the esophageal axis, and cause painful swallowing. Also this lesion was easily dilated with esophageal ballooning.

Although dilatation is the primary method of treating esophageal strictures^[5], alimentary tract reconstruction is necessary for correction of severe and extensive strictures. Invasive surgeries like stomach pull-up, colon bypass, and free jejunal flap are not always possible when the prior operation was similarly invasive^[6].

In our case, the right colon had already been used during the previous bypass operation, and the graft stricture of the neo-esophagus resulted from ischemia. Thus, patch dilative strictureplasty with viable tissue was

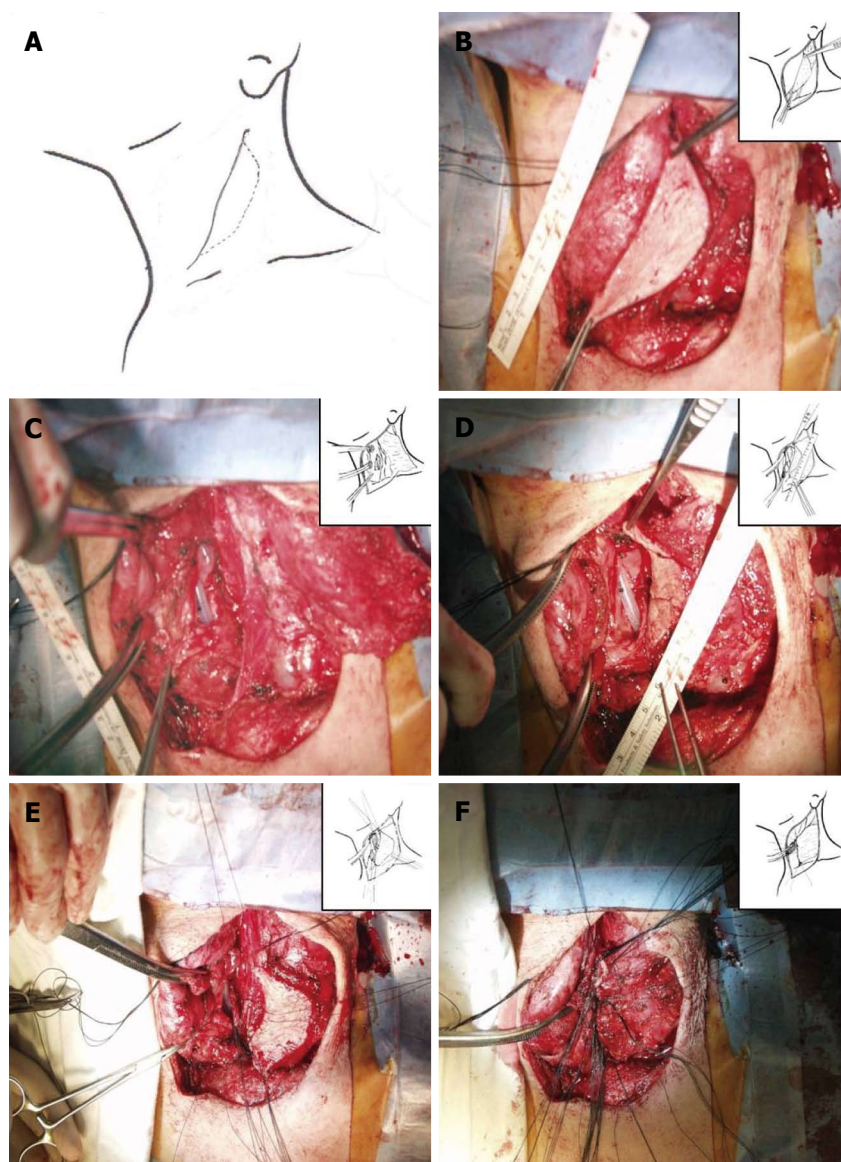


Figure 2 Photograph of the musculocutaneous sternocleidomastoid flap and esophagotomy during the operation. A: A skin incision was designed for an elliptical skin island based on the sternocleidomastoid muscle; B: A skin island (6 cm × 4 cm) on the sternocleidomastoid (SCM) muscle was prepared; C, D: Esophagotomy at the level of the cricoid cartilage was extended upward to pharynx and downward; E: The SCM muscle was severed near the clavicle and rotated. The skin flap was fixated to esophagus with interrupted silk sutures; F: The esophagoplasty site was covered with SCM muscle via multiple fixation sutures.

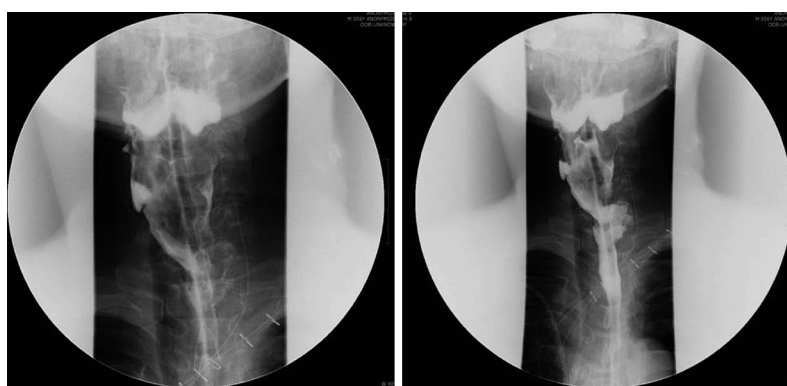


Figure 3 Esophagogram after sternocleidomastoid myocutaneous patch esophagoplasty. The stenotic lesion at the esophago-colonic anastomosis site was widened successfully and the dye passed through the colon conduit without resistance.

thought to be an ideal option for relieving the short, benign, recurrent cervical esophageal stricture.

The choice of tissue to be used as a patch to widen the esophagus varies. Among flaps, a myocutaneous flap has several advantages: its skin provides an adequate patch for the esophageal defect and the well-vascularized muscle forms a seal around the ischemic anastomosis

site, obliterates dead space, and conveys antibiotics^[7]. The SCM muscle is a logical choice for myocutaneous flap in this case because its anatomic proximity lessens trauma^[7]. Because SCM patch esophagoplasty avoids complete transection of the esophagus, coordinated peristalsis is also preserved and vagotomy is avoided^[7]. The SCM muscle has a segmental blood supply from branches of three

main vessels: the occipital artery superiorly, the superior thyroid artery in the middle segment, and a branch of the thyrocervical trunk inferiorly. The superior pedicle is more commonly used because it provides greater rotation and allows the SCM to reach the entire length of the ipsilateral cervical esophagus^[2]. The SCM myocutaneous flap does, however, have limitations. Safety is a major concern when the SCM flap is used in oncologic patients^[8]. In cases of radiation therapy, the SCM myocutaneous flap may be used only if the overlying skin is soft and supple^[2]. Furthermore, use of a SCM pedicled skin flap is limited to the cervical esophagus. In conclusion esophagoplasty with SCM myocutaneous flap is thought to be an efficient treatment of recurrent cervical esophageal strictures.

REFERENCES

- 1 **Park JK**, Sim SB, Lee SH, Jeon HM, Kwack MS. Pharyngo-enteral anastomosis for esophageal reconstruction in diffuse corrosive esophageal stricture. *Ann Thorac Surg* 2001; **72**: 1141-1143 [PMID: 11603426 DOI: 10.1016/S0003-4975(01)02961-7]
- 2 **Noland SS**, Ingraham JM, Lee GK. The sternocleidomastoid myocutaneous "patch esophagoplasty" for cervical esophageal stricture. *Microsurgery* 2011; **31**: 318-322 [PMID: 21500276 DOI: 10.1002/micr.20880]
- 3 **Wormuth JK**, Heitmiller RF. Esophageal conduit necrosis. *Thorac Surg Clin* 2006; **16**: 11-22 [PMID: 16696279 DOI: 10.1016/j.thorsurg.2006.01.003]
- 4 **Jain V**, Rodrigues GS, Gupta K. Ischaemic necrosis of subcutaneous colonic neoesophagus: an unusual complication of preternal hypertrophic scar. *Singapore Med J* 2006; **47**: 235-236 [PMID: 16518560]
- 5 **Ananthakrishnan N**, Parthasarathy G, Maraju NK, Kate V. Sternocleidomastoid muscle myocutaneous flap for corrosive pharyngoesophageal strictures. *World J Surg* 2007; **31**: 1592-1596 [PMID: 17551780 DOI: 10.1007/s00268-007-9120-5]
- 6 **Lin YD**, Jiang YG, Wang RW, Gong TQ, Zhou JH. Platysma myocutaneous flap for patch stricturoplasty in relieving short and benign cervical esophageal stricture. *Ann Thorac Surg* 2006; **81**: 1090-1094 [PMID: 16488729 DOI: 10.1016/j.athoracsur.2005.09.005]
- 7 **Cunningham DK**, Stern SJ, Burnett HF. Sternocleidomastoid myocutaneous esophagoplasty for benign cervical stricture. *Ann Thorac Surg* 2005; **79**: 1406-1407 [PMID: 15797094 DOI: 10.1016/j.athoracsur.2003.10.035]
- 8 **Lin CH**, Lin CH, Wu CW, Liao CT. Sternocleidomastoid muscle flap: an option to seal off the esophageal leakage after free jejunal flap transfer—a case report. *Chang Gung Med J* 2009; **32**: 224-229 [PMID: 19403014]

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A case of ascending colon variceal bleeding treated with venous coil embolization

Bong Suk Ko, Woo Tae Kim, Su Sun Chang, Eun Hye Kim, Seung Woo Lee, Won Seok Park, Yeon Soo Kim, Soon Woo Nam, Dong Soo Lee, Ji Chang Kim, Sang Bum Kang

Bong Suk Ko, Woo Tae Kim, Su Sun Chang, Eun Hye Kim, Seung Woo Lee, Won Seok Park, Yeon Soo Kim, Soon Woo Nam, Dong Soo Lee, Sang Bum Kang, Department of Internal Medicine, College of Medicine, The Catholic University of Korea, Daejeon St. Mary's Hospital, Daejeon 301-723, South Korea
Ji Chang Kim, Department of Radiology, College of Medicine, The Catholic University of Korea, Daejeon St. Mary's Hospital, Daejeon 301-723, South Korea

Author contributions: Ko BS wrote the paper; Kim WT, Chang SS, Kim EH, Lee SW, Park WS, Kim YS, Nam SW, and Lee DS collected the materials; Kim JC contributed radiologic interventions; and Kang SB designed the study.

Correspondence to: Sang Bum Kang, MD, Department of Internal Medicine, College of Medicine, The Catholic University of Korea, Daejeon St. Mary's Hospital, 64 Daeheungro, Jung-gu, Daejeon 301-723, South Korea. ksb1999@hanmail.net
Telephone: +82-42-2209823 Fax: +82-42-2219038

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INTRODUCTION

Colonic variceal bleeding is a very rare cause of lower gastrointestinal bleeding. Fewer than 100 cases of colonic variceal bleeding have been reported in English literature and few case reports exist in South Korea. However, most cases in South Korea were caused by different varix locations, etiologies and treatments. A single case in South Korea with ascending colonic variceal bleeding caused by alcoholic liver cirrhosis was treated with a hemicolectomy^[1]. We report the first case of an ascending colon variceal bleed caused by alcoholic liver cirrhosis that was treated with venous coil embolization in South Korea.

CASE REPORT

A 38-year-old female was admitted to our hospital with massive hematochezia and hemodynamic instability. She was diagnosed with alcoholic liver cirrhosis (Child-Pugh classification C) 6 years ago and had multiple hospitalizations with complications of esophageal variceal bleeding and hepatic encephalopathy. Upon physical examination, her initial blood pressure was 80/50 mmHg, her heart rate was 110 bpm, her respiration was 28 bpm and her

Abstract

A 38-year-old female with a history of alcoholic liver cirrhosis visited our hospital with a massive hematochezia. An esophagogastroduodenoscopy did not demonstrate any bleeding source, and a colonoscopy showed a massive hemorrhage in the ascending colon but without an obvious focus. The source of the bleeding could not be found with a mesenteric artery angiography. We performed an enhanced abdominal computed tomography, which revealed a distal ascending colonic varix, and assumed that the varix was the source of the bleeding. We performed a venous coil embolization and histoacryl injection to obliterate the colon varix. The intervention appeared to be successful because the vital signs and hemoglobin laboratory data remained stable and because the hematochezia was no longer observed. We report here on a rare case of colonic variceal bleeding that was treated with venous coil embolization.

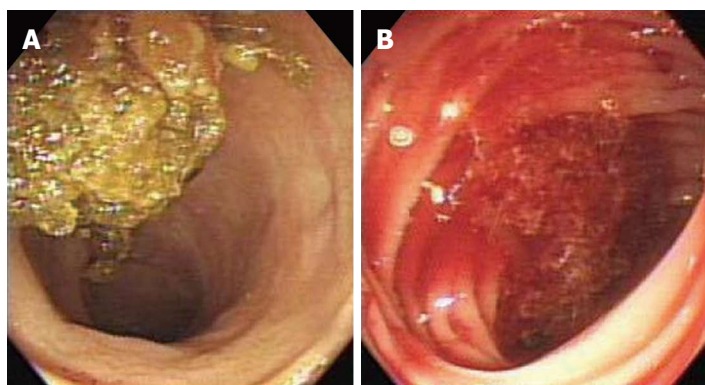


Figure 1 Initial colonoscopy. A: The terminal ileum showing yellowish-colored stool with no evidence of upper gastrointestinal bleeding; B: The ascending colon showing fresh blood without an observable source.

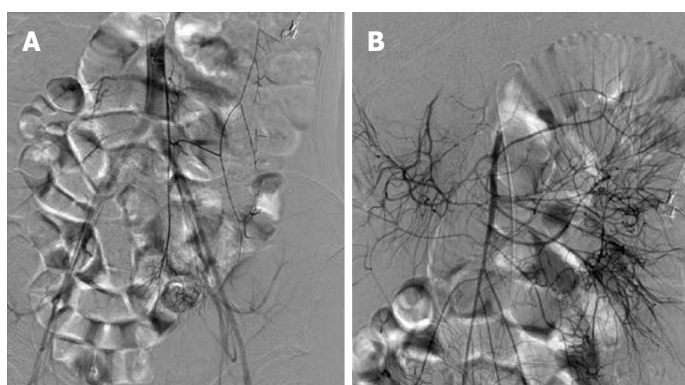


Figure 2 Mesenteric artery angiography. A: The inferior mesenteric artery angiography showing no source of bleeding; B: The superior mesenteric artery angiography showing no source of bleeding.

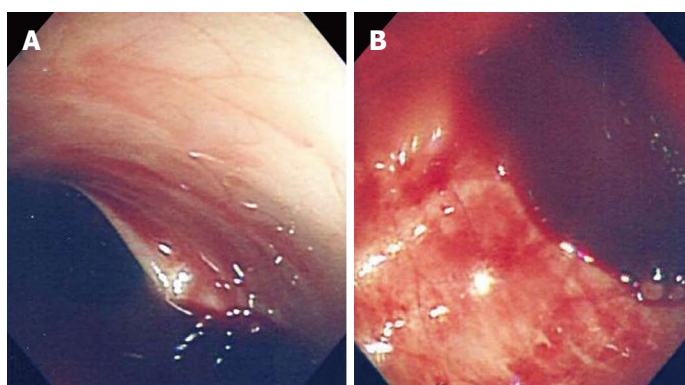


Figure 3 The second colonoscopy. A: Massive hemorrhage within the proximal ascending colon without a definite bleeding focus; B: Fresh blood within the distal ascending colon without a definite bleeding focus.

body temperature was 37 °C. No evidence of bleeding was detected with nasogastric tube irrigation. Abnormal laboratory data upon admission included the following: hemoglobin 7.0 g/dL, hematocrit 22.3%, platelets 59 000/mm³, serum creatinine 2.31 mg/dL, international normalized ratio 1.98, and albumin 2.1 g/dL.

An esophagogastroduodenoscopy demonstrated esophageal varices grade III but did not show signs of recent bleeding. The colonoscopy showed fresh blood from the ascending colon to the rectum, but there was no evidence of bleeding in the terminal ileum, which demonstrated that the origin of the bleeding could be assumed to be in the ascending colon (Figure 1). Diagnostic angiographies of the superior and inferior mesenteric arteries were performed, but the source of the bleeding could not be found (Figure 2). Even with 2 units of packed RBCs, the hemoglobin level dropped to

5.6 g/dL the next day, and the vital signs remained unstable.

A second colonoscopy was still unable to visualize the bleeding focus and only detected a massive hemorrhage in the ascending colon (Figure 3). We performed an enhanced abdominal computed tomography (CT), which revealed the formation of a varix surrounding and protruding inside the distal ascending colon by the portacaval shunt (Figure 4). The varix was thought to be the source of the gastrointestinal bleeding. We planned an operation, but a right hemicolectomy could not be performed due to poor vital signs and abnormal coagulopathy. Initially, we tried to perform a Balloon-Occluded Retrograde Transvenous Obliteration (BRTO) but failed due to the selection of the portacaval shunt. Because the patient had unstable vital signs that required immediate intervention, we performed a coil embolization and his-

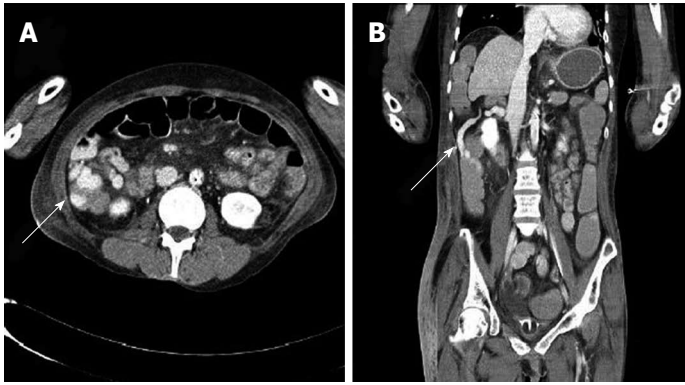


Figure 4 Abdominal computed tomography. A: Portal phase axial view with white arrow showing the varix surrounding and protruding inside the ascending colon formed by portacaval shunt; B: Coronal view with white arrow showing the colon varix.

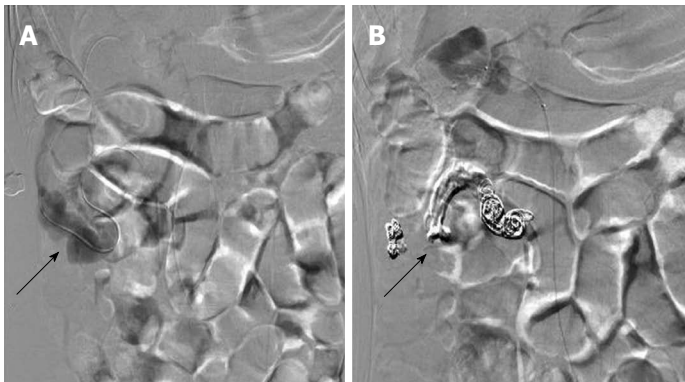


Figure 5 Venous coil embolization. A: Venography showing the varix within the ascending colon (black arrow); B: Coil embolization (black arrow) and histoacryl injection was performed to obliterate the varix.

toacryl injection to obliterate the varix (Figure 5). After the coil embolization and 4 units of packed red blood cells, the patient's hemoglobin level rose to 10.0 g/dL, where it stabilized, and the vital signs (blood pressure and heart rate) recovered. There were no further signs of bleeding, such as hematochezia. However, the patient developed complications of pneumonia and renal failure, which led to multiorgan failure. Even with ventilator care and continuous renal replacement therapy, the patient expired 2 wk after the intervention.

DISCUSSION

Colon varices are a very rare cause of lower gastrointestinal bleeding, with a reported incidence of 0.07%^[2]. Colon varices can be associated with several conditions, such as portal hypertension, portal venous obstruction, postsurgical changes and idiopathic factors, but the vast majority of colonic varices are related to portal hypertension caused by cirrhosis or portal vein obstruction. Embryologically derived anastomoses between the portal and systemic systems exist in the esophagus, terminal ileum, colon, retroperitoneum, and anterior abdominal wall^[3]. The most common sites of colonic varices are the rectum and cecum^[4]. Colon varices with portal hypertension are hypothesized to arise in patients in whom the colonic portosystemic venous collateral network is highly developed^[5]. The rate of colonic variceal bleeding that causes lower gastrointestinal bleeding with liver cirrhosis is approximately 1%-8%^[6,7]. Thus, if

a patient has a history of liver cirrhosis and has massive gastrointestinal bleeding, the possibility of colon variceal bleeding should also be considered. Other than portal hypertension, mesenteric venous or splenic vein obstruction, which may be due to thrombosis, tumor invasion, extrinsic compression, acute or chronic pancreatitis, mesenteric adhesions, or congenital anatomic variations, may cause a colon varix. With no etiology, colon varix is classified as idiopathic or primary^[5].

A colonoscopy is the principal method for the diagnosis of colon varices, but with massive bleeding, the diagnostic rate of a colonoscopy is 69% (range: 48%-90%) because the varices may be obscured by blood^[8]. Thus, in case of massive hematochezia, magnetic resonance imaging, mesenteric angiography and abdominal CT are alternative diagnostic tools^[9,10].

Management of colonic variceal bleeding has not been standardized. In a report describing a case of massive lower gastrointestinal bleeding with some hemodynamic instability caused by colonic varix, an operation was the optimal choice^[11]. However, most cases with colonic variceal bleeding are associated with cirrhotic liver, which makes many cases inoperable due to coagulopathy. Thus, interventional therapies are tested depending on the underlying etiology and the distribution of the varices. These therapies include transhepatic intravenous portosystemic shunt (TIPS), BRTO, endoscopic variceal ligation, somatostatin infusion, argon plasma coagulation, histoacryl injection, and coil embolization^[12-15]. Four other cases of colon variceal bleeding have been

described in South Korea. These cases had different varix locations, etiologies and treatments compared to our case. The first case was a 64-year-old female with liver cirrhosis who had descending colon variceal bleeding that was treated with a left hemicolectomy^[16]. The second case was a 24-year-old male who had idiopathic rectal variceal bleeding that was treated with low anterior resection^[2]. The third case was a 43-year-old male with ascending colon variceal bleeding due to alcoholic liver cirrhosis. This case was very similar to our case but was treated with a right hemicolectomy^[1]. The last case was a 33-year-old woman with a history of systemic lupus erythematosus who developed antiphospholipid syndrome and had ascending colon variceal bleeding that was treated with octreotide, a beta-blocker and warfarin^[17]. In our case, the patient was inoperable because of the poor vital signs and coagulopathy caused by the abnormal liver function. We initially tried to perform BRTO to obliterate the portacaval shunt, but the angiographic selection of the portacaval shunt failed. Thus, venous coil embolization along with a histoacryl injection was performed on the ascending colon varix. This technique seemed to be successful because the vital signs (blood pressure and heart rate) along with hemoglobin level remained stable, and no further evidence of gastrointestinal bleeding was observed after the intervention. This result is despite the fact that our patient expired due to complications of multiorgan failure because the patient already had terminal liver cirrhosis and had developed unstable vital signs before arriving in the emergency room. Together, these conditions made the patient susceptible to secondary infection and renal failure. Complications of the varix coil embolization procedure could also be considered after the intervention, such as necrosis, vessel perforation, non-targeting embolization and migration of the coil. However, although rate of complications has not been reported, this rate is thought to be very low because the procedure proceeds through a venous approach, and migration of the coil could be corrected during the procedure, and the patient did not complain of post-interventional symptoms such as abdominal pain, fever, or dyspnea. Thus, acute or delayed complications of the venous coil embolization were eliminated. A similar case in Switzerland included a 43-year-old male patient with ascending colon variceal bleeding due to alcoholic liver cirrhosis. The patient had TIPS initially to reduce the portosystemic pressure gradient, but the bleeding did not stop; the patient then underwent histoacryl injection and venous coil embolization. The patient recovered without further incidents^[18]. As noted above, there is no standardized management of colon variceal bleeding, but venous coil embolization could be an effective treatment option for patients with massive hematochezia by the colon varix. Even though our patient could not survive due to multiple organ failure and secondary infection, we believe that the successful venous coil embolization had

stopped the bleeding, which prevented a more aggressive progression. Our case is thought to be the first case in South Korea that used venous coil embolization to treat ascending colon variceal bleeding caused by alcoholic liver cirrhosis.

In conclusion, the treatment of colon variceal bleeding has not been well established. However, this case has shown that treatment with venous coil embolization on an ascending colon variceal bleeding caused by alcoholic liver cirrhosis is effective. We report here on the first case of ascending colon variceal bleeding caused by alcoholic liver cirrhosis that was treated with venous coil embolization in South Korea.

REFERENCES

- 1 Kim HU, Her KH, Kim SH, Kim BS, Kang YJ, Lee J, Kim KS. A case of variceal bleeding of the ascending colon associated with alcoholic liver cirrhosis. *Korean J Med* 2008; **75**: 215-220
- 2 Han JH, Jeon WJ, Chae HB, Park SM, Youn SJ, Kim SH, Bae IH, Lee SJ. A case of idiopathic colonic varices: a rare cause of hematochezia misconceived as tumor. *World J Gastroenterol* 2006; **12**: 2629-2632 [PMID: 16688816]
- 3 Edwards EA. Functional anatomy of the porta-systemic communications. *AMA Arch Intern Med* 1951; **88**: 137-154 [PMID: 14856444]
- 4 Sato T, Akaike J, Toyota J, Karino Y, Ohmura T. Clinicopathological features and treatment of ectopic varices with portal hypertension. *Int J Hepatol* 2011; **2011**: 960720 [PMID: 21994879 DOI: 10.4061/2011/960720]
- 5 Francois F, Tadros C, Diehl D. Pan-colonic varices and idiopathic portal hypertension. *J Gastrointest Liver Dis* 2007; **16**: 325-328 [PMID: 17925930]
- 6 Ganguly S, Sarin SK, Bhatia V, Lahoti D. The prevalence and spectrum of colonic lesions in patients with cirrhotic and noncirrhotic portal hypertension. *Hepatology* 1995; **21**: 1226-1231 [PMID: 7737627]
- 7 Hosking SW, Smart HL, Johnson AG, Triger DR. Anorectal varices, haemorrhoids, and portal hypertension. *Lancet* 1989; **1**: 349-352 [PMID: 2563507]
- 8 Abraham-Igwe C, Patel R. Idiopathic colonic varices: a case report. *Endoscopy* 2002; **34**: 680 [PMID: 12173098 DOI: 10.1055/s-2002-33235]
- 9 Chevallier P, Motamedi JP, Demuth N, Caroli-Bosc FX, Oddo F, Padovani B. Ascending colonic variceal bleeding: utility of phase-contrast MR portography in diagnosis and follow-up after treatment with TIPS and variceal embolization. *Eur Radiol* 2000; **10**: 1280-1283 [PMID: 10939490]
- 10 Choi JW, Lee CH, Kim KA, Park CM, Kim JY. Ectopic varices in colonic stoma: MDCT findings. *Korean J Radiol* 2006; **7**: 297-299 [PMID: 17143035]
- 11 Lopes LM, Ramada JM, Certo MG, Pereira PR, Soares JM, Ribeiro M, Areias J, Pinho C. Massive lower gastrointestinal bleeding from idiopathic ileocolonic varix: report of a case. *Dis Colon Rectum* 2006; **49**: 524-526 [PMID: 16395635 DOI: 10.1007/s10350-005-0279-2]
- 12 Allgaier HP, Ochs A, Haag K, Hauenstein KH, Tittor W, Rössle M, Blum HE. [Recurrent bleeding from colonic varices in portal hypertension. The successful prevention of recurrence by the implantation of a transjugular intrahepatic stent-shunt (TIPS)]. *Dtsch Med Wochenschr* 1995; **120**: 1773-1776 [PMID: 8549262 DOI: 10.1055/s-2008-1055541]
- 13 Anan A, Irie M, Watanabe H, Sohda T, Iwata K, Suzuki N, Yoshikane M, Nakane H, Hashiba T, Yokoyama M, Hi-

- gashihara H, Okazaki M, Sakisaka S. Colonic varices treated by balloon-occluded retrograde transvenous obliteration in a cirrhotic patient with encephalopathy: a case report. *Gastrointest Endosc* 2006; **63**: 880-884 [PMID: 16650568 DOI: 10.1016/j.gie.2005.11.038]
- 14 **Misra SP**, Dwivedi M. Ligation of a bleeding colonic varix using an upper gastrointestinal endoscope. *Endoscopy* 2006; **38**: 657 [PMID: 16673302 DOI: 10.1055/s-2006-925189]
 - 15 **Chen WC**, Hou MC, Lin HC, Chang FY, Lee SD. An endoscopic injection with N-butyl-2-cyanoacrylate used for colonic variceal bleeding: a case report and review of the literature. *Am J Gastroenterol* 2000; **95**: 540-542 [PMID: 10685765 DOI: 10.1111/j.1572-0241.2000.01782.x]
 - 16 **Song C**, Lee Y, Cho H, Kim D, Choi W, Park H. Massive gastrointestinal hemorrhage from the colonic varices. *J Korean Surg Soc* 1993; **44**: 923-928
 - 17 **Sohn W**, Lee HL, Lee KN. Variceal hemorrhage of ascending colon. *Clin Gastroenterol Hepatol* 2012; **10**: A24 [PMID: 21893130 DOI: 10.1016/j.cgh.2011.08.024]
 - 18 **Wiegand N**, Pfiffner R, Bauerfeind P. Ascending colonic variceal bleeding. *Gastrointest Endosc* 2006; **63**: 1073-104; discussion 1074 [PMID: 16733135 DOI: 10.1016/j.gie.2005.11.021]

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Atypical presentation of pseudomembranous colitis localized in adenomatous polyps

Cristian Hernández-Rocha, Jonathan Barra-Carrasco, Ana María Guzmán, Daniel Paredes-Sabja, Gabriel Lezcano, Pablo Zoroquiain, Manuel Álvarez-Lobos

Cristian Hernández-Rocha, Manuel Álvarez-Lobos, Department of Gastroenterology, Faculty of Medicine, Pontificia Universidad Católica de Chile, Marcoleta 367, Santiago 6510260, Chile
 Jonathan Barra-Carrasco, Daniel Paredes-Sabja, Laboratory of Mechanisms of Bacterial Pathogenesis, Department of Biological Sciences, Faculty of Biological Sciences, Universidad Andrés Bello, Santiago 6618000, Chile

Ana María Guzmán, Department of Clinical Laboratory, Faculty of Medicine, Pontificia Universidad Católica de Chile, Marcoleta 367, Santiago 6510260, Chile

Gabriel Lezcano, Pablo Zoroquiain, Department of Anatomopathology, Faculty of Medicine, Pontificia Universidad Católica de Chile, Marcoleta 367, Santiago 6510260, Chile

Daniel Paredes-Sabja, Department of Biomedical Sciences, College of Veterinary Medicine, Oregon State University, Corvallis, OR 97331, United States

Author contributions: Hernández-Rocha C, Barra-Carrasco J, Guzmán AM, Paredes-Sabja D and Álvarez-Lobos M wrote the paper; Lezcano G and Zoroquiain P reviewed the histopathological slides.

Correspondence to: Dr. Manuel Álvarez-Lobos, Department of Gastroenterology, Faculty of Medicine, Pontificia Universidad Católica de Chile, Marcoleta 367, Santiago, Chile. manalvarezl@gmail.com

Telephone: +56-2-3543820 Fax: +56-2-6397780

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Abstract

The most frequent cause of pseudomembranous colitis is *Clostridium difficile* (*C. difficile*) infection. This type of colitis is characterized by an endoscopic pattern of numerous small, yellowish or whitish plaques diffusely distributed, which typically compromises the rectum extending to proximal colon. Occasionally, the pseudomembranes compromise only the transverse or right colon, but their exclusive localization over polyps has not been reported. In this case report we have described a patient with symptoms compatible with *C. difficile* infection and positive for *C. difficile* toxigenic culture.

Colonoscopy examination showed two small polyps with a whitish surface, and histopathological analysis confirmed them to be pseudomembranes over tubular adenomas. The rest of the colonic mucosa was normal and no other cause was demonstrated. We suggest that this particular distribution might be due to a higher affinity for dysplastic cells such as adenomatous polyps of colon by *C. difficile* and/or its toxins.

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Key words: *Clostridium difficile*; Pseudomembranous colitis; Adenomatous polyps; Antibiotic-associated colitis; *Clostridium difficile* infections

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INTRODUCTION

The clinical spectrum of *Clostridium difficile* (*C. difficile*) infections may vary from mild diarrhea to life-threatening pseudomembranous colitis (PMC), which may localize from the proximal colon to the rectum^[1]. Localized forms of PMC have been reported to compromise the transverse and right colon^[2], but to the best of our knowledge, we have not found cases with exclusive localization of PMC over polyps. Below, we report a case with the pseudomembranes only covering the surface of adenomatous polyps.

CASE REPORT

An 86-year-old female receiving anticoagulant therapy due

to an aortic valve prosthesis with auricular fibrillation secondary to aortic stenosis was admitted with an incarcerated umbilical hernia. A hernioplasty was performed without complication, and she was treated with sodium piperacillin-tazobactam (4.5 g *tid*) intravenously for 7 d and discharged 10 d after surgery. Twenty-five days later, she presented hematochezia with hemodynamic compromise. She was admitted to the coronary unit, and an upper endoscopy showed a hiatal hernia with Cameron's ulcers; she was treated with omeprazole. The next day a colonoscopy showed a clean colonic mucosa, absence of blood, and two sessile 6-mm polyps in the transverse colon and in the right colon. The upper surface of both polyps was covered by an adherent and whitish layer and the surrounding mucosa was normal (Figure 1). The polyps were resected with cold biopsy forceps and sent for histopathological study. Strikingly, the day after colonoscopy, the patient developed fever with no apparent cause in the physical exam, while laboratory results showed a hematocrit of 29%, white blood cell count of 8700 cell/ μ L without band forms and C reactive protein was 10.2 mg/dL. The chest X-ray was normal and the blood and urine cultures were negative. She was empirically treated with sodium piperacillin-tazobactam, but three days later she presented frequent loose stools and diffuse abdominal pain. An enzyme immune assay (EIA) for *C. difficile* toxin was positive. The antibiotic was stopped and she was treated with oral metronidazole with an excellent response. Presence of *C. difficile* was further confirmed by real time-polymerase chain reaction (PCR) and toxigenic culture. PCR-ribotyping indicated that the clinical isolate that caused *C. difficile* infection was not an epidemic strain (non-ribotype 027 strain). A few days later, the histopathological study of the polyps showed two tubular adenomas with a low-grade epithelial dysplasia and pseudomembranes on their surface with a typical volcanic appearance (Figure 2).

DISCUSSION

It is our understanding that this is the first documented case in which the pseudomembranes are exclusively localized over adenomatous polyps with the rest of the colonic mucosa being intact. The term PMC is nonspecific and describes an acute mucosal injury characterized by an endoscopic pattern of numerous, discrete and small (2-5 mm), raised, round and yellowish plaques (pseudomembranes) distributed over an erythematous but un ulcerated mucosa^[1]. The rectum is frequently involved, but in a few cases the sigmoid and/or right colon is compromised^[2]. The histological pattern is characterized by a neutrophil-rich edema fluid in the lamina propria that bursts through tiny breaches to the surface epithelium, like a volcanic eruption, to form a characteristic punctate inflammatory pseudomembrane^[1,3]. The PMC is due, in nearly all cases, to *C. difficile*. Other infrequent causes are ischemic or infectious colitis^[4]. Piperacillin-tazobactam has been associated at lower rates with *C. difficile* infection than other antibiotics^[5]; however, in this case report, the

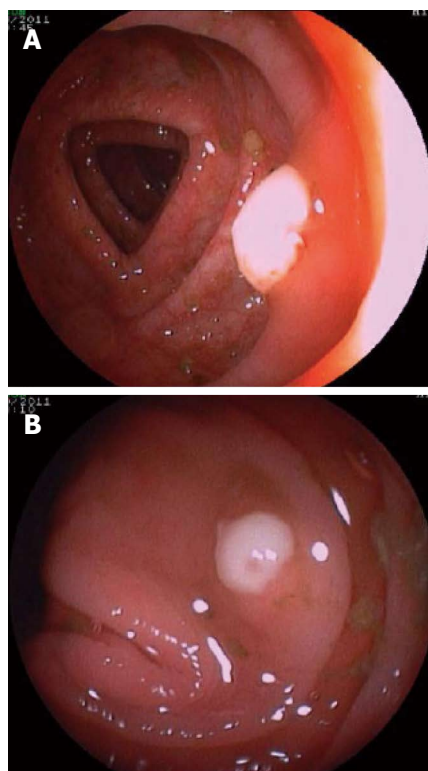


Figure 1 Colonoscopy images showing two polyps, 4 and 6 mm in size, in the right colon with adherent whitish surface. A: Polyp of 6 mm; B: Polyp of 4 mm. The mucosa round the polyps and in the rest of colon is normal.

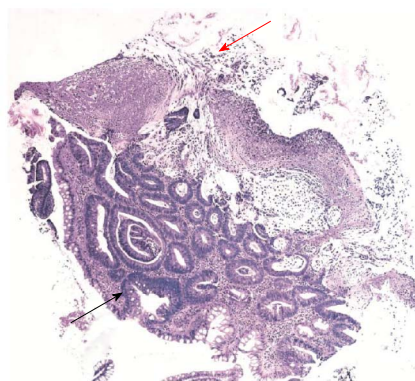


Figure 2 Low power view of a mucosal fragment. Histopathology shows: at the bottom, tubular structures lined by columnar epithelial cells with pseudostratified nuclei, consistent with low grade tubular adenoma (black arrow); at the top, ballooned crypts and intercrypt necrosis, an exudate featuring the classical "volcano" lesion (red arrow) surrounded by a laminated pseudomembrane composed of neutrophils, mucin and fibrin (hematoxylin and eosin, x 100).

clinical course and the study with EIA, PCR and toxigenic culture confirmed an association with *C. difficile* and the response to treatment with oral metronidazole was concordant.

The specific type of colonic cells where *C. difficile* infections starts is unclear, but it is known that at least enterotoxin A, one of its main virulence factors, has remarkable cytotoxicity for cells derived from human or animal cancer, more than for normal colonic epithelial

cells. One possible explanation, but by no means proven, is that the surface of neoplastic cells possesses a greater density of specific receptors for toxin A and/or toxin B as compared to less sensitive cell lines^[6]. Alternatively, it is well known that members of the *Clostridium* genus have a tropism for tumors mainly due to the hypoxic environment as a consequence of poor vascular irrigation^[7,8]. Thus, it seems plausible that colonic neoplastic cells might favor *C. difficile* colonization and PMC formation.

In conclusion, we propose that dysplastic changes of the colonic epithelium occurring as adenomatous polyps might represent a site of greater affinity for *C. difficile* toxin and/or tropism of *C. difficile* cells with the consequent formation of pseudomembranes, which were observed by an early colonoscopy in this patient.

REFERENCES

- 1 **Carpenter HA**, Talley NJ. The importance of clinicopathological correlation in the diagnosis of inflammatory conditions of the colon: histological patterns with clinical implications. *Am J Gastroenterol* 2000; **95**: 878-896 [PMID: 10763932 DOI: 10.1055/0000000000000000]
- 2 **Tedesco FJ**, Corless JK, Brownstein RE. Rectal sparing in antibiotic-associated pseudomembranous colitis: a prospective study. *Gastroenterology* 1982; **83**: 1259-1260 [PMID: 7129030 DOI: S0016508582002509]
- 3 **Price AB**, Davies DR. Pseudomembranous colitis. *J Clin Pathol* 1977; **30**: 1-12 [PMID: 838865 DOI: 10.1136/jcp.30.1.1]
- 4 **Bartlett JG**. Clinical practice. Antibiotic-associated diarrhea. *N Engl J Med* 2002; **346**: 334-339 [PMID: 11821511 DOI: 10.1056/NEJMcp011603]
- 5 **Mendez MN**, Gibbs L, Jacobs RA, McCulloch CE, Winston L, Guglielmo BJ. Impact of a piperacillin-tazobactam shortage on antimicrobial prescribing and the rate of vancomycin-resistant enterococci and *Clostridium difficile* infections. *Pharmacotherapy* 2006; **26**: 61-67 [PMID: 16509027]
- 6 **Kushnaryov VM**, Redlich PN, Sedmak JJ, Lyerly DM, Wilkins TD, Grossberg SE. Cytotoxicity of *Clostridium difficile* toxin A for human colonic and pancreatic carcinoma cell lines. *Cancer Res* 1992; **52**: 5096-5099 [PMID: 1516066]
- 7 **Wei MQ**, Ren R, Good D, Anné J. Clostridial spores as live 'Trojan horse' vectors for cancer gene therapy: comparison with viral delivery systems. *Genet Vaccines Ther* 2008; **6**: 8 [PMID: 18279524 DOI: 10.1186/1479-0556-6-8]
- 8 **Minton NP**. Clostridia in cancer therapy. *Nat Rev Microbiol* 2003; **1**: 237-242 [PMID: 15035028 DOI: 10.1038/nrmicro777]

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Anesthetic management of the SRS™ endoscopic stapling system for gastro-esophageal reflux disease

Ufuk Topuz, Tarik Umutoglu, Mefkur Bakan, Erdogan Ozturk

Ufuk Topuz, Tarik Umutoglu, Mefkur Bakan, Erdogan Ozturk, Department of Anesthesiology and Reanimation, Bezmialem Vakif University Faculty of Medicine, 34093 Istanbul, Turkey

Author contributions: Topuz U and Umutoglu T wrote this letter; Bakan M and Ozturk E revised the letter.

Correspondence to: Mefkur Bakan, MD, Assistant Professor, Bezmialem Vakif University, Vatan cad, Fatih, 34093 Istanbul, Turkey. mefkur@yahoo.com

Telephone: +90-536-2602699 Fax: +90-212-6217580

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Abstract

The SRS™ Endoscopic Stapling System (Medigus, Tel Aviv, Israel) is a new tool capable of creating a totally endoscopic fundoplication, combined with an endoscope, endoscopic ultrasound and a surgical stapler. SRS™ endoscopic stapling for gastro-esophageal reflux disease is a minimally invasive, outpatient procedure, which requires general anesthesia with positive-pressure ventilation. Keeping the patient on positive end-expiratory pressure (PEEP) may minimize the pressure gradient between the esophagus and the mediastinum, as well as help to prevent air from leaking around the screws and causing pneumomediastinum. In addition, in patients with hiatal hernia, higher PEEP levels may be required to increase intra-thoracic pressure and to force the stomach to slide into the abdomen for ease of endoscopy. We advise smoother emergence from anesthesia, taking precautions for retching, postoperative nausea and vomiting (PONV), while coughing and gagging during extubation and PONV may affect the success of the procedure. Total intravenous anesthesia with propofol and remifentanyl seems to be a good choice for these reasons.

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TO THE EDITOR

Endoscopic ultrasound was developed as a useful diagnostic tool and is being used in the treatment of various gastrointestinal diseases^[1,2]. The SRS™ Endoscopic Stapling System (Medigus, Tel Aviv, Israel) is a new tool capable of creating a totally endoscopic fundoplication, combined with an endoscope, endoscopic ultrasound, and a surgical stapler^[3]. This modified endoscope fires 3 rows of staples plicating the fundus onto the esophagus. While the anesthesia management of the SRS™ endoscopic fundoplication procedure has not been described, we would like to share our initial experiences with it on 12 patients.

After obtaining written informed consent, patients diagnosed with gastro-esophageal reflux disease, without or with ≤ 3 cm hiatal hernia, were scheduled for this procedure. Following at least 8 h fasting time, patients were premedicated with midazolam, metoclopramide, and a proton pump inhibitor. Total intravenous anesthesia (TIVA) with propofol and remifentanyl was used for both induction and maintenance of general anesthesia. After muscle relaxation was achieved with rocuronium, patients were endotracheally intubated. Patients were mechanically ventilated with oxygen/air mixture. During the procedure, the lumen of the stomach was inflated with air for better visualization. To prevent possible air leakage,

positive end-expiratory pressure (PEEP) 4-6 mmHg was applied. If a hiatal hernia was discovered during preliminary gastroscopy, the patient should be tilted 15 degrees head up and the PEEP level must be increased to 7-10 mmHg in order to depress the diaphragm and reduce the gastro-esophageal junction, so that it can be repositioned into the abdomen. Mean interventional time was 75 min; range was between 47-102 min according to the expertise of the operator. Paracetamol 1 g *iv* was administered for postoperative analgesia and ondansetron 4 mg *iv* for postoperative nausea and vomiting (PONV). Recovery was uneventful in all patients.

In conclusion, SRS™ endoscopic fundoplication for gastro-esophageal reflux disease is a minimally invasive, outpatient procedure, which requires general anesthesia with positive-pressure ventilation. Keeping the patient on PEEP may minimize the pressure gradient between the esophagus and the mediastinum and may help to prevent air from leaking around the screws and causing pneumo-mediastinum. In patients with hiatal hernia, higher PEEP levels may be required to increase intra-thoracic pressure

and force the stomach to slide into the abdomen for ease of endoscopy. It must be kept in mind that higher PEEP levels may be associated with arterial hypotension. We advise smoother emergence from anesthesia, taking precautions for retching, PONV, while coughing and gagging during extubation and PONV may affect the success of the procedure. TIVA with propofol and remifentanyl seems to be a good choice for these reasons.

REFERENCES

- 1 **Nishimura M**, Togawa O, Matsukawa M, Shono T, Ochiai Y, Nakao M, Ishikawa K, Arai S, Kita H. Possibilities of interventional endoscopic ultrasound. *World J Gastrointest Endosc* 2012; **4**: 301-305 [PMID: 22816010 DOI: 10.4253/wjge.v4.i7.301]
- 2 **Chavalitthamrong D**, Draganov PV. Endoscopic ultrasound-guided biliary drainage. *World J Gastroenterol* 2012; **18**: 491-497 [PMID: 22363114 DOI: 10.3748/wjg.v18.i6.491]
- 3 **Jafri SM**, Arora G, Triadafilopoulos G. What is left of the endoscopic antireflux devices? *Curr Opin Gastroenterol* 2009; **25**: 352-357 [PMID: 19342950 DOI: 10.1097/MOG.0b013e32832ad8b4]

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Editorial office

Jian-Xia Cheng, Director

Jin-Lei Wang, Vice Director

World Journal of Gastroenterology

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-59080039

Fax: +86-10-85381893

E-mail: wjg@wjgnet.com

<http://www.wjgnet.com>

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E-mail: bpgoffice@wjgnet.com

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Production center

Beijing Baishideng BioMed Scientific Co., Limited

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-85381892

Fax: +86-10-85381893

Representative office

USA Office

8226 Regency Drive,

Pleasanton, CA 94588-3144, United States

Telephone: +1-925-2238242

Fax: +1-925-2238243

Instructions to authors

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- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarrhoea. *Shijie Huaren Xiaohua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorseelaar RJ, Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

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DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRSA Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfeide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. Emerg Infect Dis serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

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