World Journal of *Rheumatology*

World J Rheumatol 2014 November 12; 4(3): 22-87





Published by Baishideng Publishing Group Inc

World Journal of Rheumatology

A peer-reviewed, online, open-access journal of rheumatology

Editorial Board

2011-2015

The *World Journal of Rheumatology* Editorial Board consists of 191 members, representing a team of worldwide experts in rheumatology. They are from 38 countries, including Argentina (2), Australia (4), Belgium (3), Brazil (3), Canada (2), Chile (1), China (16), Egypt (1), Finland (2), France (9), Germany (5), Greece (6), Hungary (2), India (3), Iran (2), Israel (6), Italy (11), Japan (2), Kuwait (1), Mexico (4), Morocco (2), Netherlands (3), Peru (1), Poland (1), Portugal (2), Qatar (1), Saudi Arabia (2), Slovakia (1), South Korea (4), Spain (7), Sweden (2), Switzerland (2), Thailand (1), Tunisia (1), Turkey (14), United Arab Emirates (1), United Kingdom (13), and United States (48).

EDITOR-IN-CHIEF

Jörg HW Distler, Erlangen

GUEST EDITORIAL BOARD MEMBERS

Yih-Hsin Chang, *Taichung* Jing-Long Huang, *Taoyuan* Pi-Chang Lee, *Taipei* Chin-San Liu, *Changhua* Ko-Hsiu Lu, *Taichung* Fuu-Jen Tsai, *Taichung* Chih-Shung Wong, *Taipei* Jeng-Hsien Yen, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Javier Alberto Cavallasca, Santa Fe Enrique Roberto Soriano, Buenos Aires



Australia Chang-Hai Ding, Melbourne

Davinder Singh-Grewal, Sydney Gethin Thomas, Brisbane Yin Xiao, Brisbane



Olivier Bruyère, *Liège* Nijs Jo, *Brussels* Jean-Yves Reginster, *Liège*



Simone Appenzeller, Cidade Universitaria Mittermayer Santiago, Nazaré Salvador Samuel K Shinjo, São paulo



Hong-Yu Luo, *Montreal* Guang-Ju Zhai, *St John's*







Jun-Min Chen, Fuzhou Sheng-Ming Dai, Shanghai Ai-Ping Lu, Beijing Chi Chiu Mok, Hong Kong Ling Qin, Hong Kong Han-Shi Xu, Guangzhou Qing-Yu Zeng, Shantou Peng Zhang, Shenzhen



Yasser Emad, Cairo



Rahman Shiri, Helsinki



Didier Attaix, *Theix* Francis Berenbaum, *Paris* Michel Jacques de Bandt, *Aulnay sous Bois* Pascal Laugier, *Paris* Pierre Miossec, *Lyon* M Djavad Mossalayi, *Bordeaux* Luc Mouthon, *Paris* Aleth Perdriger, *Rennes* Alain Saraux, *Brest*



Magali Cucchiarini, Homburg Thomas Jax, Neuss Friedrich Paul Paulsen, Erlangen Med H H Peter, Freiburg



Andrew P Andonopoulos, *Rion* Dimitrios Daoussis, *Patras* Kosmas I Paraskevas, *Athens* Grigorios Sakellariou, *Thessaloniki* Lazaros I Sakkas, *Larissa* Michael Voulgarelis, *Athens*



Laszlo Czirjak, Pecs András Komócsi, Pecs





Vikas Agarwal, Lucknow Srikantiah Chandrashekara, Bangalore Rajesh Vijayvergiya, Chandigarh



Nima Rezaei, Tehran Zahra Rezaieyazdi, Mashhad



Boaz Amichai, Ramat Gan George S Habib, Nazareth Illit Leonid Kalichman, Beer Sheva Igal Leibovitch, *Tel-Aviv* Ami Schattner, Rehovot Elias Toubi, Haifa



Silvano Adami, Verona Giuseppe Barbaro, Rome Mauro Cellini, Bologna Nicola Giordano, Siena Estrella Garcia Gonzalez, Siena Giovanni La Montagna, Napoli Claudio Lunardi, Verona Francesco Oliva, Rome Donato Rigante, Rome Dario Roccatello, Turin Maurizio Turiel, Milano



Yoshiya Tanaka, Kitakyushu Takashi Usui, Kyoto



Adel M A Alawadhi, Kuwait



Carlos Abud-Mendoza, San Luis Potosi Monica Vazquez-Del Mercado, Guadalajara José F Muñoz-Valle, Zapopan José Alvarez Nemegyei, Mérida



Zoubida Tazi Mezalek, Rabat Faissal Tarrass, Larache

Netherlands

Esmeralda Blaney Davidson, Nijmegen Timothy Ruben Radstake, Nijmegen

Nico M Wulffraat, Utrecht



Claudia Selene Mora-Trujillo, Lima







Elizabeth Benito-Garcia, Oeiras Alexandrina Ferreira Mendes, Coimbra



Mohammed Hammoudeh, Doha



Saudi Arabia

Almoallim Hani Mohammad, Jeddah Mohammed Tikly, Johannesburg



Slovakia

Ivica Lazúrová, Košice



Dae-Hyun Hahm, Seoul Young Mo Kang, Daegu Myeong Soo Lee, Daejeon Chang-Hee Suh, Suwon



Pedro Carpintero Benítez, Cordoba Francisco J Blanco, Coruña Vicente Giner Galvañ, Alcoy Segundo Gonzalez, Oviedo Narcis Gusi, Caceres Luis Martinez-Lostao, Zaragoza Gusi Narcis, Caceres

Spain





Aladdin Mohammad, Lund Ronald van Vollenhoven, Stockholm



Daniel Aeberli, Bern Hossein Hemmatazad, Zurich



Thailand

Prachya Kongtawelert, Chiang Mai





Aynur Akay, İzmir Deniz Evcik, Ankara Sibel Evigor, Izmir Ozgur Kasapcopur, Istanbul Suleyman Serdar Koca, Elazig Ugur Musabak, Ankara Demet Ofluoglu, Istanbul Salih Ozgocmen, Kayseri Cagatay Ozturk, Istanbul Mehmet Akif Ozturk, Ankara Ismail Sari, Izmir Mehmet Soy, Bolu Yavuz Yakut, Ankara Serap Yalın, Mersin



Ashok Kumar, Dubai



Ade O Adebajo, Sheffield Khalid Binymin, Mersyside Dimitrios P Bogdanos, London David D'Cruz, London Magdalena Dziadzio, London Edzard Ernst, Exeter Elena A Jones, Leeds Joseph G McVeigh, Belfast Sanjay Mehta, London Jonathan Rees, London Anita Williams, Salford Hazem M Youssef, Aberdeen Wei-Ya Zhang, Nottingham



United States

Cynthia Aranow, Manhasset Joseph R Berger, Lexington Vance Berger, Rockville Daniel Bikle, San Francisco Marc R Blackman, Washington Galina S Bogatkevich, Charleston Charles R Brown, Columbia Leigh F Callahan, Chapel Hill Hamid Chalian, Chicago Majid Chalian, Baltimore Sean Patrick Curtis, Rahway Barbara A Eberhard, New Hyde Park Luis R Espinoza, New Orleans Shu -Man Fu, Charlottesville Daniel E Furst, Los Angeles Reda Ebeid Girgis, Baltimore Alexei A Grom, Cincinnati Simon Helfgott, Boston Howard J Hillstrom, New York Gary S Hoffman, Cleveland Seung Jae Hong, Chicago





Meenakshi Jolly, *Chicago* M Firoze Khan, *Galveston* Irving Kushner, *Shaker Heights* Antonio La Cava, *Los Angeles* Yi Li, *Gainesville* Chuan-Ju Liu, *New York* Charles J Malemud, *Cleveland* Mahnaz Momeni, *Washington* Swapan K Nath, *Oklahoma* Ewa Olech, Oklahoma Alicia Rodríguez Pla, Dallas Chaim Putterman, Bronx Robert James Quinet, New Orleans Allison B Reiss, Mineola Lisa Georgianne Rider, Bethesda Bruce M Rothschild, Lawrence Hee-Jeong Im Sampen, Chicago Naomi Schlesinger, New Brunswick H Ralph Schumacher, Philadelphia Jasvinder A Singh, Birmingham Jianxun (Jim) Song, Hershey Yu-Bo Sun, Charlotte Thomas H Taylor, Norwich George C Tsokos, Boston Yu-Cheng Yao, Los Angeles Ping Zhang, Indianapolis Xiao-Dong Zhou, Houston



World Journal of Rheumatology

Contents		Four-monthly Volume 4 Number 3 November 12, 2014
REVIEW	22	Oral creatine supplementation: A potential adjunct therapy for rheumatoid arthritis patients Wilkinson TJ, O'Brien TD, Lemmey AB
	35	Gender differences in axial spondyloarthritis Neuenschwander R, Ciurea A
	44	Muscle wasting in rheumatoid arthritis: The role of oxidative stress Stavropoulos-Kalinoglou A, Deli C, Kitas GD, Jamurtas AZ
	54	Diagnosis and pharmacologic management of neuropathic pain among patients with chronic low back pain Yavuz F, Guzelkucuk U
MINIREVIEWS 62		Gout: A clinical overview and its association with cardiovascular diseases <i>Kienhorst LBE, Janssens HJEM, Janssen M</i>
	72	Quantifying synovial inflammation: Emerging imaging techniques Tripathi D, Agarwal V
	80	Does a biological link exist between periodontitis and rheumatoid arthritis? Joseph R, Jose Raj MG, Sundareswaran S, Kaushik PC, Nagrale AV, Jose S, Rajappan S



Contents	<i>World Journal of Rheumatology</i> Volume 4 Number 3 November 12, 2014			
APPENDIX I-V	Instructions to authors			
ABOUT COVER	Editorial Board Member of <i>World Journal of Rheumatology</i> , Grigorios Sakellariou, MD, Department of Rheumatology, 424 General MilitaryHospital Ring Road N.Efkarpias, Thessaloniki, 564 03, Greece			
AIM AND SCOPE	World Journal of Rheumatology (World J Rheumatol, WJR, online ISSN 2220-3214, DOI: 10.5499) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians. WJR covers topics concerning osteoarthritis, metabolic bone disease, connective tissue diseases, antiphospholipid antibody-associated diseases, spondyloarthropathies, acute inflammatory arthritis, fibromyalgia, polymyalgia rheumatica, vasculitis syndromes, periarticular rheumatic disease, pediatric rheumatic disease, miscellaneous rheumatic diseases, and rheumatology-related therapy, pain management, rehabilitation. We encourage authors to submit their manuscripts to WJR. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.			
INDEXING/ ABSTRACTING	World Journal of Rheumatology is now indexec	World Journal of Rheumatology is now indexed in Digital Object Identifier.		
FLYLEAF I-III	Editorial Board			
EDITORS FOR Resp	~ · ·	sible Science Editor: Yue-Li Tian g Editorial Office Director: Xiu-Xia Song		
NAME OF JOURNAL World Journal of Rheumatology	Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China Telephone: +86-10-59080039 Fax: +86-10-85381893 E-mail: editorialoffice@wignet.com	COPYRIGHT © 2014 Baishideng Publishing Group Inc. Articles pub- lished by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non- commercial License, which permits use, distribution, and		
ISSN ISSN 2220-3214 (online) LAUNCH DATE December 31, 2011 FREQUENCY Four-monthly	Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com PUBLISHER Baishideng Publishing Group Inc 8226 Regency Drive,	reproduction in any medium, provided the original work is properly cited, the use is non commercial and is other- wise in compliance with the license. SPECIAL STATEMENT All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the		



Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.22 World J Rheumatol 2014 November 12; 4(3): 22-34 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

REVIEW

Oral creatine supplementation: A potential adjunct therapy for rheumatoid arthritis patients

Thomas J Wilkinson, Thomas D O'Brien, Andrew B Lemmey

Thomas J Wilkinson, Thomas D O'Brien, Andrew B Lemmey, School of Sport, Health and Exercise Sciences, Bangor University, Wales LL57 2PZ, United Kingdom

Thomas D O'Brien, Research Institute for Sport and Exercise Sciences, Liverpool John Moores University, England L3 3AF, United Kingdom

Author contributions: Wilkinson TJ gathered literature information and drafted manuscript; O'Brien TD and Lemmey AB advised on search methodology and critically revised the manuscript.

Correspondence to: Andrew B Lemmey, Professor, School of Sport, Health and Exercise Sciences, Bangor University, George Building, Normal Site, Holyhead Road, Wales LL57 2PZ, United Kingdom. a.b.lemmey@bangor.ac.uk

Telephone: +44-1248-383932

Received: June 28, 2014 Revised: September 19, 2014 Accepted: October 1, 2014

Published online: November 12, 2014

Abstract

Creatine is one of the most popular forms of protein supplements and is known to improve performance in healthy athletic populations via enhanced muscle mass and adenosine triphosphate energy regeneration. Clinical use of creatine may similarly benefit patients with rheumatoid arthritis (RA), an inflammatory condition characterised by generalised muscle loss termed "rheumatoid cachexia". The adverse consequences of rheumatoid cachexia include reduced strength, physical function and, as a consequence, quality of life. Whilst regular high-intensity exercise training has been shown to increase muscle mass and restore function in RA patients, this form of therapy has very low uptake amongst RA patients. Thus, acceptable alternatives are required. The aim of this review is to consider the potential efficacy of creatine as an anabolic and ergonomic therapy for RA patients. To date, only one study has supplemented RA patients with creatine, and the findings from this investigation were inconclusive. However, trials in populations with similar losses of muscle mass and function as RA, including older adults and

those with other muscle wasting conditions, indicate that creatine is an efficacious way of improving muscle mass, strength and physical function, and may offer an easy, safe and cheap means of treating rheumatoid cachexia and its consequences.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Creatine supplementation; Nutritional supplement; Rheumatoid arthritis; Rheumatoid cachexia; Physical function

Core tip: Creatine supplementation primarily improves physical function by enhancing the re-synthesis of adenosine triphosphate *via* increased stores of phosphocreatine in the muscle. Through this pathway it provides greater levels of energy during physical activity and improves recovery. Creatine also augments muscle protein synthesis, thereby increasing muscle mass. These dual effects increase strength, reduce fatigue, and thereby improve function. In patients with conditions such as rheumatoid arthritis that are characterised by muscle loss and subsequent reductions in strength and physical function, creatine offers a potential therapeutic intervention for augmenting muscle mass and function that is safe, easy and inexpensive to administer.

Wilkinson TJ, O'Brien TD, Lemmey AB. Oral creatine supplementation: A potential adjunct therapy for rheumatoid arthritis patients. *World J Rheumatol* 2014; 4(3): 22-34 Available from: URL: http://www.wjgnet.com/2220-3214/full/v4/i3/22.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.22

INTRODUCTION

Patients with rheumatoid arthritis (RA) often experience a substantial loss of muscle mass (cachexia), which results in significant adverse consequences such as decreased strength, impaired physical function, and a reduction in



quality of life. Unfortunately, current drug treatments for RA do not attenuate this muscle loss, nor fully restore physical function^[1,2], and whilst exercise has been shown to be effective in restoring both muscle mass and function in RA patients (e.g.,^[3]) the lack of uptake and adherence to sufficiently intense training means this form of therapy is unlikely to be widely adopted. Nutritional supplements offer a potential alternate therapeutic intervention that would be more easily adopted. One such nutritional supplement is oral administration of creatine (Cr). Creatine is a popular form of protein supplementation that has been widely demonstrated to improve physical function via enhanced energy regeneration and increased muscle mass^[4]. Consequently, Cr supplementation potentially offers a low-cost and generally acceptable means for RA patients to restore muscle mass and functional capacity.

This article reviews the evidence regarding the potential of Cr as an adjunct treatment to improve muscle mass and function in RA patients. In the course of doing this, rheumatoid cachexia, its effect on patients, and the rationale for nutritional supplementation (such as Cr) to improve body composition and physical function will be discussed. Then the mechanisms and effectiveness of Cr in athletic populations will be described before we present a review of the existing evidence regarding the efficacy of Cr in RA-relevant clinical trials.

RHEUMATOID ARTHRITIS, CACHEXIA AND MUSCLE LOSS

Rheumatoid arthritis is an autoimmune disease predominantly affecting middle-aged and older females and is characterised by persistent synovitis, systemic inflammation, and the presence of specific autoantibodies^[5]. This inflammation is associated with damage to the articular cartilage and bone^[5], and a range of co-morbidities including cardiovascular disease^[6], obesity^[7], diabetes^[8], osteoporosis^[9], fatigue^[10] and depression^[11].

Additionally, RA is characterised by aberrant changes in body composition. The involuntary loss of muscle, often coupled with elevated adiposity, has been termed "rheumatoid cachexia"^[12], and occurs in approximately 67% of patients^[3,12-16] including those with controlled disease^[12]. Much like sarcopenia (muscle loss due to ageing^[17]), rheumatoid cachexia leads to a loss of strength^[18] and reduced physical functioning^[19,20] impairing performance of activities of daily living such as standing independently from a chair, walking, climbing stairs, and lifting and carrying^[21]. Additionally, muscle wasting impairs immune function^[22], and is a significant predictor of cardio-vascular disease and overall mortality^[23-28].

The aetiology of rheumatoid cachexia is multifactorial and may involve increased production of excess inflammatory cytokines such as tumour necrosis factor-alpha (TNF α) and interleukins -1 and -6 (IL-1, IL-6) which are also implicated in the pathophysiology of RA itself^(13,24-26). On a cellular level, several key signalling pathways, such as nuclear factor-kappa B (NF- κ B; catabolic) and insulingrowth factor- I (IGF- I; anabolic), regulate protein synthesis and degradation in the muscle^[27]. Changes to these pathways "tip" the metabolic activity from anabolic to catabolic, thereby inducing muscle wasting^[22].

TREATMENTS OF MUSCLE WASTING

Interventions that are effective in increasing muscle mass have been shown to improve physical function, reduce disability, and enhance quality of life in RA patients^[29]. However, efficacious and safe anabolic interventions which are widely acceptable to rheumatoid patients have yet to be identified.

Medication and drug treatments

Rheumatoid cachexia and relatively poor physical function remain prevalent even in RA patients with wellcontrolled disease activity (i.e., approximately 20%-30% below the level seen in age- and sex-matched sedentary healthy controls^[1,3]). Therefore, it is apparent that controlling disease activity alone is insufficient to restore body composition and function. Roubenoff et al^[13] hypothesised that TNF α was central in causing rheumatoid cachexia, so it might be expected that anti-TNF α biologics would be the pharmaceutical anti-rheumatic treatment most likely to reverse rheumatoid cachexia. However, even these agents have proved ineffective in this regard^[2,30,31]. In fact, in the trials conducted to date, anti-TNF α therapy have not only failed to increase lean mass in recent-onset^[2,30] and established^[31] RA patients, but appear to increase fat mass^[2] and more disturbingly, trunk fat mass^[31] relative to standard disease modifying anti-rheumatic drugs.

Similarly, yet to be published data from an on-going study by our group suggests that even in the current "Treat to Target" era, when disease activity is more tightly and successfully controlled, and clinical "remission" is regularly achieved, RA patients still experience significant loss of muscle (approximately 10%), increased adiposity (approximately 12%), and relatively poor physical function (approximately 20%-30% decreased), compared to age- and sex-matched healthy sedentary controls.

Progressive resistance training

High-intensity progressive resistance training (PRT) has been shown to substantially increase muscle mass, and as a consequence dramatically improve strength and restore normal levels of physical function in RA patients (*e.g.*,^[3,16,32]). However, patient uptake of exercise is poor^[33], and even patients who experience significant benefits of structured exercise cease training when supervision is withdrawn^[34]. Thus, sustained exercise training is unlikely to be widely adopted as a therapy for reversing cachexia and restoring function.

Nutritional supplementation

Anabolic nutritional supplementation offers a potential treatment option that is easily administered, inexpensive, and makes limited demands of the patient. It has been

Table 1 Summary of the results from the meta-analysis by Nissen et al Summary Summary							
Supplement $(n = \text{studies})$	Average dosage (maintenance dose)	Duration (wk)	Net lean mass change	Net strength change			
Cr (<i>n</i> = 18)	19.4 g/d for 5.3 d (6.7 g/d)	7.5	+0.36%/wk ^b	+1.09%/wk ^b			
HMB $(n = 9)$ Chromium (n = 12)	3 g/d 485 μg/d	8 11.2	,	+1.40%/wk ^b +0.25%/wk			
(n = 12) Androstenedione (n = 3)	200 mg/d	10.7	+0.05%/wk	-0.06%/wk			
Protein $(n = 4)$	1.15 g/kg per day	6.3	+0.12%/wk	-0.18%/wk			
DHEA $(n = 2)$	125 mg/d	10	+0.12%/wk	+0.06%/wk			

The net change is expressed as % change per week. Only Cr and HMB resulted in significant changes; ${}^{b}P < 0.005$. Cr: Creatine; HMB: β -hydroxy- β -methylbutyrate; DHEA: Dehydroepiandrosterone.

reported that up to 75% of RA patients believe that food and nutrition may play an important role in their symptom severity, with up to 50% of RA patients reportedly trying some form of dietary manipulation in an attempt to attenuate symptomology^[35]. Scientific evidence continues to suggest that diet should be part of routine care in those with wasting disorders (for review see Stamp^[35]).

Our group previously investigated the effects of 12 wk of a mixture of β -hydroxy- β -methylbutyrate, glutamine and arginine (HMB/GLN/ARG) protein supplementation in 40 RA patients^[15]. The results showed that both HMB/GLN/ARG and a control mixture of other, non-essential, amino acids (alanine, glutamic acid, glycine and serine) were effective in increasing muscle mass and improving physical function in RA patients. Thus it appears that protein per se is capable of significantly improving lean mass, total body protein and objective measures of physical function which reflect the ability to perform activities of daily living in RA patients.

Creatine, a combination of essential amino acids, has generally been shown to be more effective than other protein-based supplements in increasing lean mass. For example, Cribb et al^{36]} showed that Cr (1.5 g/kg per day for 11 wk) was able to significantly improve lean mass by +5.5%, compared to whey protein (+3.7%; P < 0.05) in 33 trained males. Further to this, in a meta-analysis^[37] of 48 studies, both lean mass and strength gain were unaffected by whey protein and other supplementation such as androstenedione when compared to a placebo treatment, and only supplementation with either Cr or HMB resulted in a significant gains (Table 1). The superior gains in lean mass and strength from Cr relative to HMB, combined with the additional benefits of Cr to energy production and recovery identifies Cr as a potentially highly effective adjunct treatment for improving rheumatoid cachexia and physical function.

WHAT IS CREATINE?

Creatine, or methylguanidine-acetic acid, is a naturally

occurring compound made from 3 amino acids; arginine, glycine, and methionine^[4], and is synthesized within the body, primarily in the liver, kidney and pancreas^[38].

Most (approximately 95%) of the total Cr pool is contained in skeletal muscle, with around 60% [75 mmol \cdot kg dry weight (dw)⁻¹] in the phosphorylated form, phosphocreatine (PCr)^[39,40], and the remaining 40% (50 mmol \cdot kg dw⁻¹) existing as free Cr^[41]. Muscle does not synthesize Cr itself but is dependent on Cr uptake through specific membrane sodium dependent transporters^[42].

WHAT DOES CREATINE DO?

Changes in ATP energy synthesis

Creatine performs many roles in the body, the most important of which is in generating energy for the muscles. Muscle relaxation and contraction, and therefore the movement of the body, is fuelled by energy liberated from the dephosphorylation of adenosine triphosphate (ATP).

ATP ↔ adenosine diphosphate (ADP) + phosphate (P) + energy (catalysed by the enzyme ATPase)

The ATP stores in the body are limited (concentration in skeletal muscle approximately 24 mmol \cdot kg/dw^[40]), and without a means of resynthesizing ATP at an equally rapid rate, maximal exercise exhausts these stores within 1-2 s^[43]. To overcome this storage limitation, the body is able to maintain a continuous ATP supply through different metabolic resynthesis pathways: either anaerobically in the cytosol, or aerobically in the mitochondrion.

As stated previously, Cr is primarily stored in the body in a phosphorylated form as PCr, with the muscle content of PCr 3-4 times higher than that of ATP^[41]. In a process called dephosphorylation, some energy for ATP resynthesis comes directly from the hydrolysis (splitting) of phosphate from PCr^[41].

PCr \leftrightarrow **Cr** + **P** + **Energy** [catalysed by the enzyme creatine kinase (CK)]

In this process, the liberated phosphate group can then combine with ADP in a reaction catalysed by CK to restore ATP levels^[44] and maintain high cellular ATP/ADP ratios^[45]:

ADP + **P** \leftrightarrow **ATP** + **Cr** (catalysed by CK)

As a consequence, it would be anticipated that increasing initial Cr stores and thereby delaying PCr depletion would enhance resynthesis of ATP and augment performance^[46,47]. Ingestion of Cr supplements (20 g a day for 5 d) has been shown to increase the total Cr and PCr concentration of human skeletal (Table 2), and indeed, reduced blood lactate concentrations have been observed after high-intensity^[48] and endurance exercise^[49]; although these findings are not universal^[50].

Changes in muscle mass and protein synthesis

Creatine is an osmotically active substance. Thus, as



Table 2 Changes in creatine and phosphocreatine levelsfollowing Cr supplementation						
	Mean baseline Cr levels ¹	Increase after 20 g/d for 5 d				
Creatine	Approximately 125 mmol · kg/dw ^[130] (90 to 160 mmol · kg/dw) ^[4]	+ 25 mmol · kg/dw (approximately 20%) ^[144]				
Phosphocreatine	Approximately 50 mmol · kg/dw ^[41]	+ 8 mmol · kg/dw (approximately 15%) ^[132]				

¹Typical values for an average 70 kg male. Cr: Creatine.

skeletal muscle cell Cr and PCr concentrations rise, the cell will rapidly draw in extracellular water *via* osmosis in order to maintain equilibrium^[51]. The uptake of Cr and water into the muscle accounts for the increases in body mass (approximately 1-2 kg) usually observed after a few days of supplementation (*e.g.*,^[52]). Total body water has been reported to increase up to 3 litres (+9%)^[45]; of which intra-cellular water has been shown to increase by between 0.77-3.0 litre (an increase of +3%-9% from baseline values) (*e.g.*,^[53-56]) in the absence of changes in extra-cellular water^[54].

The intramuscular uptake of Cr and the associated increase in intracellular water increases osmotic pressure, which in turn stimulates protein synthesis. Cellular hydration state is an important factor in controlling cellular protein turnover, *i.e.*, an increase in cellular hydration inhibits proteolysis and stimulates protein synthesis^[57], whereas cell shrinkage has opposite effects^[51,58-61]. However, it is unclear whether acute Cr supplementation augments muscle protein by this mechanism^[62,63].

Creatine has also been shown to stimulate muscle hypertrophy by inducing expression of muscle myogenic factors such as MRF4, MyoD and myogenin^[64].

Deldicque *et al*^(65,66) showed that the muscle gene ex-</sup>pression of IGF-I was raised following Cr supplementation. This finding was corroborated by Burke *et al*⁶⁷ who found increased muscle content of IGF-I as a result of Cr supplementation combined with 8 wk of PRT. These findings are highly relevant to Cr's anabolic potency as IGF-I produced locally in the muscle (mIGF-I) is thought to regulate adult skeletal muscle maintenance and hypertrophy^[68]. Conversely, Cr supplementation in conjunction with PRT has been shown to lower serum levels of myostatin^[69], a hormone that is highly expressed in RA synovial tissues and inhibits muscle growth by reducing myoblast (muscle) proliferation^[70-72] and thus is associated with muscle atrophy^[72] and joint destruction^[73]. The anabolic response to Cr supplementation is particularly evident in type II muscle fibres^[60,74], which is particularly interesting because RA patients present with preferential atrophy of type II fibres^[/5].

Reduction in inflammatory cytokines

Patients with RA exhibit high synovial levels and serum concentration of the cytokines $\text{TNF}\alpha$ and $\text{IL-1}\beta^{[22]}$. These cytokines, in addition to causing synovial inflam-

mation^[76], also modulate the expression of enzymes controlling muscle protein degradation^[27]. Bassit *et al*^[77] investigated the effects of Cr supplementation (20 g/d for 5 d prior to competition) on plasma levels of the proinflammatory cytokines: TNF α , IL-1 β , and prostaglandin E2 (PGE2) in triathletes after a half-ironman competition. These cytokines are typically raised following prolonged strenuous exercise^[78], but Cr supplementation attenuated the increases in TNF α by 42% and 64%, IL-1 β by 72% and 71%, and PGE2 by 85.5% and 91 %, 24 and 48 h post, respectively.

Creatine and bone degradation

RA patients are at 2-fold increased risk of having osteoporosis and approximately 28% of patients develop this condition^[9,14]. In wheelchair-independent patients experiencing Duchenne dystrophy, Cr supplementation was able to enhance bone mineral density (+3%) and reduce urinary cross-linked N telopeptides of type I collagen (NTx) excretion, a marker for bone resorption^[62]. In addition, Candow *et al*^[79] also reported a reduction in NTx (-27%) *vs* placebo (+13%; P < 0.005), and similar findings were reported by Chilibeck *et al*^[80] who showed that in elderly men, Cr was able to improve arm bone mineral density by +3.2% (P < 0.001) *vs* placebo (-1%) However, more research is needed in this area to understand the mechanisms behind this action.

Athletic performance

Creatine has repeatedly demonstrated efficacy in improving high-intensity short-term exercise performance and subsequent recovery. For example, in cycling, Cr supplementation has been shown to significantly enhance peak power output^[48,81,82] and maximal work^[39] during repetitive sprints. Similarly, runners who supplemented with Cr decreased their 100 m sprint time and total time for $6 \text{ m} \times 60 \text{ m}$ sprint intervals^[83], and highly trained football players improved their repeated sprint performance (6 m × 15 m sprints with 30 s recovery) and attenuated fatigue-induced decline in jumping ability following Cr supplementation^[84]. Creatine supplementation has also been found to be effective in improving performance of a variety of sustained high-intensity activities (e.g., kayaking for 5 min^[85]; 1000 m rowing^[86]; and running 300 and 1000 m intervals $(3-4 \text{ min rest})^{[87]}$). These functional benefits are attributed to increased ATP resynthesis, heightened availability of PCr in type II fibres, and increased total Cr stores^[41]. These effects may be particularly beneficial to older adults or clinical populations who experience difficulty performing short-term, high intensity activities such as hurrying for a bus, crossing roads, climbing stairs, or digging in the garden.

Creatine has also been shown to improve strength related measures. In an analysis of 22 studies, athletes supplementing with Cr had an average +8% greater increase in muscle strength than placebo (for a review see Rawson *et al*^[88]). Furthermore, Cr supplementation when combined with PRT has been shown to be more effective.

tive at increasing strength and weightlifting performance than PRT alone^[89,90]. Improvements in strength translate into increased work capacity, and thus improved ability to perform activities of daily living such as walking, carrying shopping, doing housework, *etc*^{16,19,21]}.

Although approximately 70%^[91] of short-term studies on Cr supplementation report some ergogenic benefit, the responses are often variable amongst individuals^[88], and supplementation generally does not result in improvements in endurance performance (e.g., repeated 6 km treadmill and terrain run performance^[48,58,92,93]).

CRITICAL REVIEW OF RELEVANT CLINICAL LITERATURE

Aim

The aim of this review is to examine existing evidence assessing the efficacy of Cr supplementation in improving muscle mass and physical function, with particular reference to its potential use in treating rheumatoid cachexia and its consequences. To achieve this we searched for, and extracted relevant data from published research papers in RA and other conditions for which findings are likely transferable to the RA population, e.g., ageing population and other musculoskeletal and wasting diseases.

Search methods

Peer-reviewed research articles were included in this review provided they: (1) investigated the effects of Cr supplementation in RA patients or other populations deemed relevant to RA (i.e., elderly populations (> 60 years) or musculoskeletal disorders featuring loss of muscle and physical function); (2) included body composition (muscle and/or fat mass) and/or physical function as outcome measures; and (3) conducted an intervention of any design in RA patients; or undertook a blinded placebocontrolled trial for non-RA populations, to ensure only evidence of higher certainty of evidence was included. As the purpose of this review is to investigate alternative treatments to high-intensity exercise for restoring muscle mass and physical function, data on the additive effects of Cr supplementation and PRT were excluded. Publications were also excluded if they were a literature review, thesis, abstract, or a letter or comment, and the search was limited to English language citations.

PubMed and Google Scholar were searched from April to May 2014 using the search term "creatine supplementation" combined with "cachexia"; "clinical"; "patient"; "older adults"; "elderly"; "sarcopenia" and "rheumatoid arthritis". In addition, the reference sections of the selected papers were hand-searched for relevant ancestral references. The title and abstract of each search result was first screened for relevance according to the inclusion criteria above, before full articles were obtained. Full-text articles were then screened before final inclusion in this review.

Search results

The initial search returned 758 articles, excluding dupli-

cates, of which 21 met the inclusion criteria and were selected for this review. One trail investigating Cr supplementation of RA patients was found^[94]. This study was not controlled in any way so is considered to provide evidence of low certainty. The body composition and physical function data extracted from trials in older adults are presented in Table 3, and data extracted from trials in other relevant clinical populations appear in Table 4.

Rheumatoid arthritis

Willer et al^[94] was the only study identified that completed a trial of Cr in an RA population. Twelve RA patients were un-blinded to the Cr supplementation and no placebo group or control arm existed. Participants were given oral Cr supplementation for 21 d using recommended doses (day 1-5: 20 g/d; day 6-21: 2 g/d) and the effects on muscle strength, subjectively assessed function during activities of daily living (Health Assessment Questionnaire), and disease activity were examined. It was found that Cr supplementation increased muscle strength in 8 out of the 12 patients by an average of +14% (P = 0.02), as determined by the muscle strength index (the mean of 8 strength measurements during flexion and extension of the knee and elbow/max sample strength $\times 100^{195}$). This increase in muscle strength was not associated with changes in skeletal muscle Cr or PCr levels. Routine clinical measures of disease activity and subjectively evaluated physical function showed no changes.

The authors attributed the "limited effectiveness of Cr" to "alterations in the kinetics of Cr in patients with RA (e.g., reduced transport into the muscle, increased metabolism and/or excretion)". However, this interpretation places emphasis on the subjectively assessed function, which was unchanged, rather than the objectively measured strength, which did improve significantly. It is known that the Health Assessment Questionnaire is weakly associated with objective measures of physical condition such as strength (r = -0.35) and joint mobility $(r = 0.27)^{[96]}$, and is often insensitive to changes in objective function (e.g., [3,96]). Additionally, only 12 patients were used in the study, and with Cr supplementation reported to be ineffective in approximately 30% of individuals^[46], it would be anticipated that only 8 of the RA patients in this investigation would see any benefit. Consistent with this prediction, strength increases were noted in 8 patients. Moreover, the study supplementation period only lasted 3 wk, much less than the 8-12 wk recommended by manufacturers and used by other studies. Thus, whilst the findings of Willer et al⁹⁴'s trial are inconclusive, they do provide some indications that Cr may be effective in the RA population. Clearly more research is needed in this area.

Ageing and sarcopenia Nine studies^[55,97-104] were identified that investigated the effects of Cr supplementation in older adults and met the inclusion criteria. Four of these studies, reported that Cr increased body mass by 0.49-1.86 $\rm kg^{[55,98,100,102]}$ and that



WJR www.wjgnet.com

Treatment arm (mean age <u>+</u> SD)	Supplementation period	Study design	Body composition changes	Physical function changes	Ref.
10 males	20 g/d for 10 d followed by 4	vs PL group (dextrose) (n	² Body density,	¹ Leg fatigue performance	[99]
(66.7 ± 1.9 yr)	g/d for 20 d	= 10)	² LM, ² %BF		
9 males	20 g/d for 5 d	vs PL group (sucrose)	¹ BM, ² LM	² Strength	[100]
(65.0 ± 2.1 yr)		(n = 8)			
10 females	0.3 g per kg/day for 7 d	vs PL group ($n = 6$)	No details	¹ Objective function tests, ² Endurance	[97]
$(67.0 \pm 6.0 \text{ yr})$				capacity	
10 males	0.3 g per kg/day for 7 d	vs PL group (powdered	¹ BM, ¹ LM	¹ Strength, ¹ Power, ¹ Objective function	[55]
(65.4 ± 1.5 yr)		cellulose) $(n = 8)$		tests	
15 females	0.3 g per kg/day for 7 d	vs PL group (powdered	¹ BM, ¹ LM, ² %BF	¹ Strength, ¹ Objective function tests	[98]
(63.3 ± 1.2 yr)		cellulose) ($n = 12$)			
7 males and 8 females	20 g/d for 7 d followed by	Cross-over design	² BM	¹ Strength, ¹ Endurance (cycling capacity	[101]
$(74.5 \pm 6.4 \text{ yr})$	10 g/d for 7 d			at fatigue threshold), ² Objective	
				function tests	
7 males	20 g/d for 5 d	vs PL group	¹ BM, ² LM	² MVC or contractile force	[102]
(72.5 ± 2.5 yr)		(maltodextrin) $(n = 5)$			
4 males and 4 females	20 g/d for 5 d followed by	vs PL (glucose) (n = 8)	² Lower limb	² Strength	[103]
(71.0 ± 1.9 yr)	3 g/d for 8 wk		volume, ² BM, ² %BF	² Endurance	
15 females	20 g/d for 5 d followed by	vs PL (dextrose) ($n = 15$)	¹ LM, ² FM	¹ Strength	[104]
(66.1 ± 4.8 yr)	5 g/d for 23 wk			¹ Objective function tests	

Table 3 Summary of studies investigating the effects of creatine supplementation on sarcopenia and function in older adults over 60 years

¹Significant increase/improvement; ²No significant change (^a*P* < 0.05 for interaction between placebo and Cr group). Cr: Creatine; PL: Placebo; BM: Body mass; %BF: Percent body fat; FM: Fat mass; LM: Lean mass; MVC: Maximal voluntary contraction; No details: No details are specified or this measure was not made.

this gain was predominantly lean mass (LM), with increases in muscle mass of up to +2.22 kg^[55]. In contrast, no significant changes in body mass or LM were found in the remaining five studies^[99-101,103,104], although a trend of increased LM (+0.3%) following Cr supplementation relative to placebo (P = 0.062) was found by one of these^[104]. As expected, no significant changes in % body fat subsequent to Cr supplementation in older subjects were reported^[55,99,100].

Three of the six studies that measured muscle strength changes reported improvements following Cr supplementation^[55,98,101]. Gotshalk *et al*^[55] reported strength increases of both maximal leg press (+7%-8%), knee extensor (+9%) and knee flexor muscles (+15%) in older males, whilst in females increases in leg press (+3.4% or 5.2 kg) and bench press (+4.4% or 1.7 kg) were found^[98]. In a cross-over design, Stout *et al*^[101] found that Cr significantly increased maximal isometric grip strength by +6.7%^[101]. Conversely, Jakobi *et al*^[102] found that 5 d of Cr supplementation was unable to increase elbow flexor maximal voluntary strength or any other muscle contractile properties (twitch and tetanic recordings from electrical stimulation of the muscles). Similar findings were reported by Rawson *et al*^[100], who found no significant effect on isometric elbow flexor strength after 5 d supplementation, and Bermon *et al*^[103] who found no increase in chest or strength compared to a placebo (P > 0.05).

All studies assessing short-term physical function reported significant improvements in lower-extremity functional tests such as the sit-to-stand in 30 s (SST-30) by up to $12^{0/55,97,98,105}$, and a tandem gait test by $6^{0/55}$ to $9^{0/98}$ following Cr supplementation. Lower body power (as assessed by a 10-s Wingate test) was shown to improve by $+11^{0/55}$ and Rawson *et al*⁹⁹ reported that leg fatigue (as

expressed as a % change in the total peak torque generate and assessed by 5×30 s knee extensions at 180° on an isokinetic dynamometer) was reduced by 9% (compared to a 5% increase in the placebo group, P < 0.05). Similar findings by Stout *et al*^{101]} showed lower body muscle endurance (cycling capacity at fatigue threshold) was improved by 15.6% compared to the placebo group. However, assessments of endurance capacity (*i.e.*, 1-mile walk test; and gross mechanical efficiency, ventilatory threshold, and peak oxygen intake determined during cycle ergometry) were not significantly improved following Cr supplementation^[97].

Trials in other clinical populations

One study^[106] trialled Cr supplementation in osteoarthritis (OA). OA is the most common form of arthritis, and as with RA, is characterised by joint damage, muscle weakness, poor physical function^[107], and predominantly affects females^[108]. In this investigation, Roy *et al*^[106] reported limited effects of Cr supplementation in OA patients recovering from total knee arthroplasty, despite a significant increase in serum Cr concentration, with no improvements in muscle strength (handgrip, dorsiflexion and quadriceps strength, 30-foot timed walk and 4-step climb) observed after 40 d (10 d pre surgery and 30 d post-surgery) of Cr supplementation relative to placebo.

One trial^[109] reported the use of Cr supplementation in fibromyalgia, another chronic syndrome of unknown etiology, characterized by some similarities in symptomology to RA, including pain, muscle dysfunction, disability and fatigue^[110]. Some of the fibromyalgia symptoms such as muscle dysfunction and fatigue could, in theory, be due to low muscle levels of ATP and PCr^[109]. A randomised controlled trial of Cr supplementation in fibromyalgia

Wilkinson TJ et al. Oral creatine supplementation

Condition	Treatment arm	Supplementation period	Control arm	Body composition changes	Physical function changes	Other effects	Ref.
Osteoarthritis	<i>n</i> = 18	10 g/d pre surgery; 5 g/ d for 30 d post-surgery	<i>vs</i> PL (<i>n</i> = 19) (dextrose)	² % BF, ² FM, ³ LM (CSA), ³ BW	³ Strength	³ PCr	[106]
Fibromyalgia	<i>n</i> = 16	20 g/d for 5 d followed by 5 g/d for 16 wk	vs PL (n = 16) (dextrose)	Not measured	¹ Strength	³ QoL scores, ³ Pain, ³ Cognition, ¹ PCr	[109]
Cancer (cachexia)	n = 16 (colorectal cancer)	20 g/d for 5 d followed by 5 g/d for 8 wk	vs PL (n = 15) (cellulose)	³ LM	¹ Strength	³ QoL scores	[118]
	n = 9 (adolescents with leukaemia (acute lymphoblastic)	2 sets of 8 wk (with a 6 wk wash out in- between)	vs control "natural history group" ($n = 50$)	³ LM, ² %BF	No details	³ Bone mineral content	[119]
Duchenne muscular	n = 18 (adolescents)	5 g/d for 8 wk	vs PL (n = 15) (vitamin C)	No details	¹ Strength	¹ PCr	[113]
dystrophy	n = 15 (adolescents)	5 g/d for 24 wk	vs PL (n = 16) (cocoa powder)	No details	³ Strength, ³ Objective function tests		[116]
	n = 30 (adolescents)	0.10 g per kg/d for 16 wk	Cross-over design (PL group dextrose)	¹ LM	¹ Strength	² Bone breakdown markers	[112]
	<i>n</i> = 15 (adolescents) (12 with DMD and 3 with Becker dystrophy)	3 g/d for 13 wk	Cross-over design (PL group maltodextrin)	No details	¹ Strength (MVC), ¹ Fatigue resistance		[62]
Mytonic muscular dystrophy 1 (DM1)	n = 34	5 g/d for 36 wk	Cross-over design (PL group dextrose)	³ LM	³ Strength, ³ Objective function tests		[115]
	<i>n</i> = 34	10.6 g/d for 10 d followed by 5.3 g/d for 45 d	Cross-over design (PL group cellulose)	³ LM	³ Strength	³ ADL, ³ QoL scores	[114]
Mytonic muscular dystrophy 2 (DM2)	<i>n</i> = 10	10 g/d for 13 wk	vs PL (n = 10)	No details	³ Strength	³ QoL scores	[117]

¹Significant increase/improvement; ²Significant decrease/reduction; ³No significant change (^a*P* < 0.05 for interaction between placebo and Cr group). Cr: Creatine; PL: Placebo; CSA: Cross sectional area; ADL: Activities of daily living; PCr: Phosphocreatine; QoL: Quality of life; MVC: Maximal voluntary contraction; BM: Body mass; FM: Fat mass; LM: Lean mass; PRT: Progressive resistance training; No details: No details are specified or this measure was not made.

patients^[109] found that muscle PCr content increased and muscle strength improved relative to the placebo group (leg-press by 9.8%, P = 0.02; chest-press by 1.2%, P = 0.02; and isometric hand-grip strength by 6.4%, P = 0.07) in the Cr group.

Myopathy is a muscle wasting disorder which primarily affects skeletal muscle. Much like rheumatoid cachexia seen in RA patients, this can cause a variety of complaints including progressive weakness and wasting of skeletal muscle, and fatigue (for a review see Kley et al^[111]). Seven trials of Cr supplementation in populations with myopathies were found, with these investigations finding mixed results on the efficacy of oral Cr. In a cross-over design trial in 30 Duchenne muscular dystrophy (DMD) adolescents^[112], the Cr supplementation phase increased lean mass by +0.7 kg and grip strength by approximately 20% compared to the placebo phase. In a similar design, Cr supplementation improved maximal strength and fatigue resistance in 15 other patients with DMD^[62]. Further to these trials, improvements in muscle PCr/P ratio and preservation of calf muscle strength were also reported by Banerjee *et al*^{113]} in 18 DMD patients.

In contrast, in cross-over design trials of patients with Mytonic muscular dystrophy 1 (DM1), Cr failed to induce any changes in muscle strength, lean mass or disease symptoms^[114,115], or improve function or strength in DMD patients^[116] or patients with myotonic dystrophy type 2 (DM2)^[117].

Two studies^[118,119] were found that reported trials of Cr supplementation in cancer patients. Up to 80% of cancer patients have associated muscle wasting which is termed "cancer cachexia"^[120]. Like other forms of cachexia, this is characterised by a preferential loss of skeletal muscle mass (with or without a loss of fat mass) which cannot be reversed through conventional methods of nutrition^[121]. In patients with cancer, Cr supplementation improved handgrip strength by 5.5% (P = 0.019)^[118] and reduced body fat accumulation (-3.5%; P < 0.05) relative to a placebo group^[119].

Review conclusions

Around 70% of RA patients are middle-aged or elderly females^[122], and the existing evidence indicates that Cr can be successful in countering the effects of sarcopenia



WJR www.wjgnet.com

in older populations independent of exercise training^[123], specifically in older females^[124,125]. Of nine included trials that have supplemented the elderly with Cr, only three^[100,102,103] found no beneficial effect on lean mass, strength, or physical function. However, the magnitude of effect appears to be reduced relative to that observed in young healthy individuals^[126], and the limited number of studies indicates that further work is needed to fully evaluate the role of Cr supplementation^[127].

Creatine has been shown to be effective in a range of clinical conditions^[128] including muscle wasting disorders^[62,112,113], and cancer cachexia^[118]. Despite the inconclusive findings of the solitary RA study^[94], of the twelve clinical trials identified, six showed positive effects of Cr on muscle mass and/or strength and function measures.

FACTORS AFFECTING CREATINE EFFECTIVENESS IN CERTAIN INDIVIDUALS OR POPULATIONS

Apart from inadequate supplement duration or dose, various other factors influence Cr effectiveness. It has been reported that 20%-30% of individuals do not respond to Cr supplementation; when "non-responsiveness" is defined as an increase in resting total muscle Cr of < 10mmol \cdot kg/dw following 5 d loading at 20 g per day^[46,88]. Syrotuik *et al*¹²⁹ found that based on pre-existing biological and physiological factors, "responders" (defined in that study as $\geq 20 \text{ mmol} \cdot \text{kg/dw}$ increase in intramuscular Cr) possessed a biological profile of (1) low initial levels of total Cr or PCr (approximately $< 110 \text{ mmol} \cdot \text{kg/dw}$); (2) higher percentage of type II fibres (> 63.1%); and (3) a higher preload muscle fibre cross-sectional area (CSA) (approximately > 1500 μ m²). For individuals whose initial muscle Cr concentrations approach 150 mmol · kg/dw, Cr supplementation does not appear to augment muscle Cr uptake, increase PCr resynthesis, or improve performance^[4,129,130]. Not surprisingly, optimal responses to Cr supplementation are generally observed in groups with reduced serum and muscle levels of Cr such as vegetarians and low meat eaters, which include many older individuals^[58,125,130,131].

Although the majority of the studies reviewed found benefits of Cr supplementation in the elderly, it has been suggested that uptake of Cr into muscle is reduced in older adults (> 60 years) relative to younger subjects^[99,132], and that subsequently older adults may require a longer Cr treatment period^[56].

SAFETY OF CREATINE

Concerns about possible side effects of Cr supplementation have been raised in lay publications, mailing lists and online forums. However, none of the studies included in this review reported any adverse incidents during the trials ranging from 5 d to 36 wk. This is consistent with other studies of long term (10 mo to 5 years) (*e.g.*,^[53,133,134]) or high dose Cr supplementation (10 g/d) (*e.g.*,^[135,136]) that have reported no adverse side effects. According to Walliman^[137], current evidence does "not hint towards any negative health effects of Cr". Therefore, the anecdotal reports remain unsubstantiated and may be unrelated to Cr supplementation^[44].

Concerns about the long-term safety of Cr have specifically been related to kidney function. Theoretically, the high nitrogen content (approximately 32%) of Cr could place additional strain on the kidney if taken in large excess for a long period of time^[133]. Glomerular filtration rate (GFR) is widely accepted as the best overall measure of kidney function, with serum and urine creatinine levels the most commonly used markers for estimating GFR^[136]. However, since Cr is converted to creatinine^[47], it is normal for individuals who take Cr supplements to have elevated creatinine levels^[138], thus falsely suggesting renal function impairment. Use of alternative GFR markers such as Cystatin C has shown that Cr supplementation does not promote renal dysfunction^[136].

There is currently limited research on the effects of Cr supplementation in patients with exiting low GFR. A prospective report^[139] suggests that short-term (35 d) Cr supplementation (5 d of 20 g/d followed by 5 g/d) does not affect kidney function in individuals with a single kidney and mildly decreased GFR. However, more research is needed in this area. Similarly, no evidence has emerged that Cr supplementation results in impaired liver function or liver damage^[41,44,140,141].

PRESCRIPTION OF CREATINE TO PATIENTS

Туре

Creatine supplements are usually taken as a tablet or powder (mixed with water), and exist in a variety of forms including Cr ethyl ester, Cr hydrochloride and the most commonly available Cr monohydrate (Cr complexed with a molecule of water). No differences in effectiveness have been found between these different Cr forms^[142].

"Loading" dosage

Cr should be "loaded" into the muscle (using a high dose) for the first few days followed by a lower maintenance dose^[41]. The most common "loading" dosage recommendation for Cr supplementation is 20 g/d (in four 5 g doses) for 5 d, as stores appear to be maximised within 5 to 6 d at this dose^[130]. Alternate loading phases exist including daily doses based on body mass such as 0.25 g/kg^[41] or 0.15 g/kg^[143]. However, a constant dose of 3 g/d, without an intensive loading phase, achieved an increase in total Cr levels equal to a standard 5 d loading protocol and subsequent maintenance phase after 28 d^[144].

"Maintenance" and frequency

Total muscle Cr can be maintained for at least 4-6 wk after the initial loading phase by the ingestion of small daily Cr doses of 2-5 g^[41,144]. This period of low dosage is called the "maintenance phase". Here, Cr is usually taken



in 8 to 12 wk cycles, with a 4 to 5 wk "washout" period in between to allow serum Cr to return to baseline levels.

CONCLUSION

RA is characterised by a loss of muscle which causes reduced strength and physical functioning. Current antirheumatic pharmaceutical treatments are unable to reverse the effects of cachexia, and although high intensity exercise is highly effective in rectifying body composition and restoring physical function, uptake of, and adherence to, exercise training by RA patients is poor. Thus, other treatment options need investigation. Oral Cr supplementation offers a potentially efficacious, cheap and widely acceptable therapy for achieving these outcomes in RA patients. Creatine works primarily by enhancing the re-synthesis of ATP *via* increased stores of PCr in the muscle, and thus improving recovery during and after physical activity. Creatine also augments muscle protein synthesis thereby increasing muscle mass.

This review found only one study in which RA patients were supplemented with $Cr^{[94]}$ and its findings, whilst promising, were inconclusive. However, trials in populations with similar presentation to RA (*i.e.*, reduced muscle mass and impaired physical function), including older females, indicate that Cr is an efficacious way to improve muscle mass, strength and physical function. Therefore, additional studies in RA populations are advocated, as confirmation of the efficacy of Cr supplementation would provide an easy, safe and effective means of reversing the effects of rheumatoid cachexia in the majority of the RA population.

REFERENCES

- 1 Lunt M, Watson KD, Dixon WG, Symmons DP, Hyrich KL. No evidence of association between anti-tumor necrosis factor treatment and mortality in patients with rheumatoid arthritis: results from the British Society for Rheumatology Biologics Register. *Arthritis Rheum* 2010; **62**: 3145-3153 [PMID: 20662063 DOI: 10.1002/art.27660]
- 2 Engvall IL, Tengstrand B, Brismar K, Hafström I. Infliximab therapy increases body fat mass in early rheumatoid arthritis independently of changes in disease activity and levels of leptin and adiponectin: a randomised study over 21 months. *Arthritis Res Ther* 2010; **12**: R197 [PMID: 20964833 DOI: 10.1186/ar3169]
- 3 Lemmey AB, Marcora SM, Chester K, Wilson S, Casanova F, Maddison PJ. Effects of high-intensity resistance training in patients with rheumatoid arthritis: a randomized controlled trial. Arthritis Rheum 2009; 61: 1726-1734 [PMID: 19950325 DOI: 10.1002/art.24891]
- 4 Casey A, Greenhaff PL. Does dietary creatine supplementation play a role in skeletal muscle metabolism and performance? *Am J Clin Nutr* 2000; **72**: 607S-617S [PMID: 10919967]
- 5 Scott DL, Wolfe F, Huizinga TW. Rheumatoid arthritis. Lancet 2010; 376: 1094-1108 [PMID: 20870100 DOI: 10.1016/ S0140-6736(10)60826-4]
- 6 Gremese E, Ferraccioli G. The metabolic syndrome: the crossroads between rheumatoid arthritis and cardiovascular risk. Autoimmun Rev 2011; 10: 582-589 [PMID: 21539940 DOI: 10.1016/j.autrev.2011.04.018]
- 7 García-Poma A, Segami MI, Mora CS, Ugarte MF, Terrazas

HN, Rhor EA, García E, Ramos MP, Alva M, Castañeda I, Chung CP. Obesity is independently associated with impaired quality of life in patients with rheumatoid arthritis. *Clin Rheumatol* 2007; **26**: 1831-1835 [PMID: 17340047]

- 8 Lee TJ, Park BH, Son HK, Song R, Shin KC, Lee EB, Song YW. Cost of illness and quality of life of patients with rheumatoid arthritis in South Korea. *Value Health* 2012; 15: S43-S49 [PMID: 22265066 DOI: 10.1016/j.jval.2011.11.020]
- 9 Haugeberg G, Uhlig T, Falch JA, Halse JI, Kvien TK. Bone mineral density and frequency of osteoporosis in female patients with rheumatoid arthritis: results from 394 patients in the Oslo County Rheumatoid Arthritis register. *Arthritis Rheum* 2000; 43: 522-530 [PMID: 10728744]
- 10 Helal AMH, Shahine EM, Hassan MM, Hashad DI, Moneim RA. Fatigue in rheumatoid arthritis and its relation to interleukin-6 serum level. *Egypt Rheum* 2012; 34: 153-157
- 11 Sheehy C, Murphy E, Barry M. Depression in rheumatoid arthritis--underscoring the problem. *Rheumatology* (Oxford) 2006; 45: 1325-1327 [PMID: 16908510]
- 12 Roubenoff R, Roubenoff RA, Ward LM, Holland SM, Hellmann DB. Rheumatoid cachexia: depletion of lean body mass in rheumatoid arthritis. Possible association with tumor necrosis factor. J Rheumatol 1992; 19: 1505-1510 [PMID: 1464859]
- 13 Roubenoff R, Roubenoff RA, Cannon JG, Kehayias JJ, Zhuang H, Dawson-Hughes B, Dinarello CA, Rosenberg IH. Rheumatoid cachexia: cytokine-driven hypermetabolism accompanying reduced body cell mass in chronic inflammation. J Clin Invest 1994; 93: 2379-2386 [PMID: 8200971]
- 14 Engvall IL, Svensson B, Tengstrand B, Brismar K, Hafström I. Impact of low-dose prednisolone on bone synthesis and resorption in early rheumatoid arthritis: experiences from a two-year randomized study. *Arthritis Res Ther* 2008; **10**: R128 [PMID: 18986531 DOI: 10.1186/ar2542]
- 15 Marcora S, Lemmey A, Maddison P. Dietary treatment of rheumatoid cachexia with beta-hydroxy-beta-methylbutyrate, glutamine and arginine: a randomised controlled trial. *Clin Nutr* 2005; 24: 442-454 [PMID: 15896432]
- 16 Marcora SM, Lemmey AB, Maddison PJ. Can progressive resistance training reverse cachexia in patients with rheumatoid arthritis? Results of a pilot study. J Rheumatol 2005; 32: 1031-1039 [PMID: 15940763]
- 17 Morley JE, Abbatecola AM, Argiles JM, Baracos V, Bauer J, Bhasin S, Cederholm T, Coats AJ, Cummings SR, Evans WJ, Fearon K, Ferrucci L, Fielding RA, Guralnik JM, Harris TB, Inui A, Kalantar-Zadeh K, Kirwan BA, Mantovani G, Muscaritoli M, Newman AB, Rossi-Fanelli F, Rosano GM, Roubenoff R, Schambelan M, Sokol GH, Storer TW, Vellas B, von Haehling S, Yeh SS, Anker SD. Sarcopenia with limited mobility: an international consensus. J Am Med Dir Assoc 2011; 12: 403-409 [PMID: 21640657 DOI: 10.1016/j.jamda.2011.04.014]
- 18 van Bokhorst-de van der Schueren MA, Konijn NP, Bultink IE, Lems WF, Earthman CP, van Tuyl LH. Relevance of the new pre-cachexia and cachexia definitions for patients with rheumatoid arthritis. *Clin Nutr* 2012; **31**: 1008-1010 [PMID: 22695407 DOI: 10.1016/j.clnu.2012.05.012]
- 19 Giles JT, Ling SM, Ferrucci L, Bartlett SJ, Andersen RE, Towns M, Muller D, Fontaine KR, Bathon JM. Abnormal body composition phenotypes in older rheumatoid arthritis patients: association with disease characteristics and pharmacotherapies. *Arthritis Rheum* 2008; 59: 807-815 [PMID: 18512711 DOI: 10.1002/art.23719]
- 20 Summers GD, Deighton CM, Rennie MJ, Booth AH. Rheumatoid cachexia: a clinical perspective. *Rheumatology* (Oxford) 2008; 47: 1124-1131 [PMID: 18448480 DOI: 10.1093/ rheumatology/ken146]
- 21 Baumgartner RN, Wayne SJ, Waters DL, Janssen I, Gallagher D, Morley JE. Sarcopenic obesity predicts instrumental activities of daily living disability in the elderly. *Obes Res* 2004;



Wilkinson TJ et al. Oral creatine supplementation

12: 1995-2004 [PMID: 15687401]

- 22 Walsmith J, Roubenoff R. Cachexia in rheumatoid arthritis. Int J Cardiol 2002; 85: 89-99 [PMID: 12163213]
- 23 Malmstrom TK, Miller DK, Herning MM, Morley JE. Low appendicular skeletal muscle mass (ASM) with limited mobility and poor health outcomes in middle-aged African Americans. J Cachexia Sarcopenia Muscle 2013; 4: 179-186 [PMID: 23532635 DOI: 10.1007/s13539-013-0106-x]
- 24 Rall LC, Rosen CJ, Dolnikowski G, Hartman WJ, Lundgren N, Abad LW, Dinarello CA, Roubenoff R. Protein metabolism in rheumatoid arthritis and aging. Effects of muscle strength training and tumor necrosis factor alpha. *Arthritis Rheum* 1996; **39**: 1115-1124 [PMID: 8670319]
- 25 Roubenoff R, Walsmith J, Lundgren N, Snydman L, Dolnikowski GJ, Roberts S. Low physical activity reduces total energy expenditure in women with rheumatoid arthritis: implications for dietary intake recommendations. *Am J Clin Nutr* 2002; **76**: 774-779 [PMID: 12324290]
- 26 Rall LC, Roubenoff R. Rheumatoid cachexia: metabolic abnormalities, mechanisms and interventions. *Rheumatology* (Oxford) 2004; 43: 1219-1223 [PMID: 15292530]
- 27 Fanzani A, Conraads VM, Penna F, Martinet W. Molecular and cellular mechanisms of skeletal muscle atrophy: an update. J Cachexia Sarcopenia Muscle 2012; 3: 163-179 [PMID: 22673968 DOI: 10.1007/s13539-012-0074-6]
- 28 Giles JT, Allison M, Blumenthal RS, Post W, Gelber AC, Petri M, Tracy R, Szklo M, Bathon JM. Abdominal adiposity in rheumatoid arthritis: association with cardiometabolic risk factors and disease characteristics. *Arthritis Rheum* 2010; 62: 3173-3182 [PMID: 20589684 DOI: 10.1002/art.27629]
- 29 **Lemmey AB**. Efficacy of progressive resistance training for patients with rheumatoid arthritis and recommendation regarding its prescription. *Int J Clin Rheum* 2011; **6**: 189-205
- 30 Marcora SM, Chester KR, Mittal G, Lemmey AB, Maddison PJ. Randomized phase 2 trial of anti-tumor necrosis factor therapy for cachexia in patients with early rheumatoid arthritis. *Am J Clin Nutr* 2006; 84: 1463-1472 [PMID: 17158431]
- 31 Metsios GS, Stavropoulos-Kalinoglou A, Douglas KM, Koutedakis Y, Nevill AM, Panoulas VF, Kita M, Kitas GD. Blockade of tumour necrosis factor-alpha in rheumatoid arthritis: effects on components of rheumatoid cachexia. *Rheumatology* (Oxford) 2007; 46: 1824-1827 [PMID: 18032540]
- 32 Sharif S, Thomas JM, Donley DA, Gilleland DL, Bonner DE, McCrory JL, Hornsby WG, Zhao H, Lively MW, Hornsby JA, Alway SE. Resistance exercise reduces skeletal muscle cachexia and improves muscle function in rheumatoid arthritis. *Case Rep Med* 2011; 2011: 205691 [PMID: 22203849 DOI: 10.1155/2011/205691]
- 33 Sokka T, Kautiainen H, Pincus T, Verstappen SM, Aggarwal A, Alten R, Andersone D, Badsha H, Baecklund E, Belmonte M, Craig-Müller J, da Mota LM, Dimic A, Fathi NA, Ferraccioli G, Fukuda W, Géher P, Gogus F, Hajjaj-Hassouni N, Hamoud H, Haugeberg G, Henrohn D, Horslev-Petersen K, Ionescu R, Karateew D, Kuuse R, Laurindo IM, Lazovskis J, Luukkainen R, Mofti A, Murphy E, Nakajima A, Oyoo O, Pandya SC, Pohl C, Predeteanu D, Rexhepi M, Rexhepi S, Sharma B, Shono E, Sibilia J, Sierakowski S, Skopouli FN, Stropuviene S, Toloza S, Valter I, Woolf A, Yamanaka H. Work disability remains a major problem in rheumatoid arthritis in the 2000s: data from 32 countries in the QUEST-RA study. *Arthritis Res Ther* 2010; 12: R42 [PMID: 20226018 DOI: 10.1186/ar2951]
- 34 Lemmey AB, Williams SL, Marcora SM, Jones J, Maddison PJ. Are the benefits of a high-intensity progressive resistance training program sustained in rheumatoid arthritis patients? A 3-year followup study. *Arthritis Care Res* (Hoboken) 2012; 64: 71-75 [PMID: 21671413 DOI: 10.1002/acr.20523]
- 35 **Stamp LK**, J MJ, Cleland LG. Diet and rheumatoid arthritis: a review of the literature. *Semin Arthritis Rheum* 2005; **35**: 77-94 [PMID: 16194694]

- 36 Cribb PJ, Williams AD, Stathis CG, Carey MF, Hayes A. Effects of whey isolate, creatine, and resistance training on muscle hypertrophy. *Med Sci Sports Exerc* 2007; **39**: 298-307 [PMID: 17277594]
- 37 Nissen SL, Sharp RL. Effect of dietary supplements on lean mass and strength gains with resistance exercise: a metaanalysis. J Appl Physiol (1985) 2003; 94: 651-659 [PMID: 12433852]
- 38 American College of Sports Medicine. The physiological and health effects of oral creatine supplementation. *Med Sci* Sports Exerc 2010; 32: 706-717
- 39 Casey A, Constantin-Teodosiu D, Howell S, Hultman E, Greenhaff PL. Creatine ingestion favorably affects performance and muscle metabolism during maximal exercise in humans. *Am J Physiol* 1996; 271: E31-E37 [PMID: 8760078]
- 40 Harris RC, Hultman E, Nordesjö LO. Glycogen, glycolytic intermediates and high-energy phosphates determined in biopsy samples of musculus quadriceps femoris of man at rest. Methods and variance of values. *Scand J Clin Lab Invest* 1974; 33: 109-120 [PMID: 4852173]
- 41 Bogdanis GC, Papaspyrou A, Maridaki M. Muscle Metabolism and Fatigue During Sprint Exercise: Effects Of Creatine Supplementation. Serb J Sport Sci 2007; 1: 37-57
- 42 **Longo N**, Ardon O, Vanzo R, Schwartz E, Pasquali M. Disorders of creatine transport and metabolism. *Am J Med Genet C Semin Med Genet* 2011; **157C**: 72-78 [PMID: 21308988 DOI: 10.1002/ajmg.c.30292]
- 43 **Burton DA**, Stokes K, Hall GM. Physiological effects of exercise. *Contin Educ Anaesth Crit Care Pain* 2004; **4**: 185-188
- 44 Kreider RB, Ferreira M, Wilson M, Grindstaff P, Plisk S, Reinardy J, Cantler E, Almada AL. Effects of creatine supplementation on body composition, strength, and sprint performance. *Med Sci Sports Exerc* 1998; 30: 73-82 [PMID: 9475647]
- 45 Bemben MG, Bemben DA, Loftiss DD, Knehans AW. Creatine supplementation during resistance training in college football athletes. *Med Sci Sports Exerc* 2001; 33: 1667-1673 [PMID: 11581550]
- 46 Greenhaff PL. Creatine and its application as an ergogenic aid. Int J Sport Nutr 1995; 5 Suppl: S100-S110 [PMID: 7550252]
- 47 Wyss M, Kaddurah-Daouk R. Creatine and creatinine metabolism. *Physiol Rev* 2000; 80: 1107-1213 [PMID: 10893433]
- 48 Balsom PD, Söderlund K, Sjödin B, Ekblom B. Skeletal muscle metabolism during short duration high-intensity exercise: influence of creatine supplementation. *Acta Physiol Scand* 1995; 154: 303-310 [PMID: 7572228]
- 49 Tang FC, Chan CC, Kuo PL. Contribution of creatine to protein homeostasis in athletes after endurance and sprint running. *Eur J Nutr* 2014; 53: 61-71 [PMID: 23392621 DOI: 10.1007/s00394-013-0498-6]
- 50 Engelhardt M, Neumann G, Berbalk A, Reuter I. Creatine supplementation in endurance sports. *Med Sci Sports Exerc* 1998; 30: 1123-1129 [PMID: 9662683]
- 51 Lang F, Busch GL, Ritter M, Völkl H, Waldegger S, Gulbins E, Häussinger D. Functional significance of cell volume regulatory mechanisms. *Physiol Rev* 1998; 78: 247-306 [PMID: 9457175]
- 52 Powers ME, Arnold BL, Weltman AL, Perrin DH, Mistry D, Kahler DM, Kraemer W, Volek J. Creatine Supplementation Increases Total Body Water Without Altering Fluid Distribution. J Athl Train 2003; 38: 44-50 [PMID: 12937471]
- 53 Poortmans JR, Francaux M. Long-term oral creatine supplementation does not impair renal function in healthy athletes. *Med Sci Sports Exerc* 1999; **31**: 1108-1110 [PMID: 10449011]
- 54 **Ziegenfuss TN**, Lowery LM, Lemon PW. Acute fluid volume changes in men during three days of creatine supplementation. *J Ex Physiol* 1998; **1**: 1-9
- 55 Gotshalk LA, Volek JS, Staron RS, Denegar CR, Hagerman FC, Kraemer WJ. Creatine supplementation improves muscular performance in older men. *Med Sci Sports Exerc* 2002; 34: 537-543 [PMID: 11880821]

Wilkinson TJ et al. Oral creatine supplementation

- 56 Chrusch MJ, Chilibeck PD, Chad KE, Davison KS, Burke DG. Creatine supplementation combined with resistance training in older men. *Med Sci Sports Exerc* 2001; 33: 2111-2117 [PMID: 11740307]
- 57 Ingwall JS, Weiner CD, Morales MF, Davis E, Stockdale FE. Specificity of creatine in the control of muscle protein synthesis. J Cell Biol 1974; 62: 145-151 [PMID: 4407046]
- 58 Balsom PD, Harridge SD, Söderlund K, Sjödin B, Ekblom B. Creatine supplementation per se does not enhance endurance exercise performance. *Acta Physiol Scand* 1993; 149: 521-523 [PMID: 8128901]
- 59 Bessman SP, Savabi F. The role of the phosphocreatine energy shuttle in exercise and muscle hypertrophy. Creatine and Creatine phosphate: Scientific and Clinical Perspectives. 1st ed. New York: Academic Press, 1988: 185-198
- 60 **Sipilä I**, Rapola J, Simell O, Vannas A. Supplementary creatine as a treatment for gyrate atrophy of the choroid and retina. *N Engl J Med* 1981; **304**: 867-870 [PMID: 7207523]
- 61 Häussinger D, Roth E, Lang F, Gerok W. Cellular hydration state: an important determinant of protein catabolism in health and disease. *Lancet* 1993; 341: 1330-1332 [PMID: 8098459]
- 62 Louis M, Lebacq J, Poortmans JR, Belpaire-Dethiou MC, Devogelaer JP, Van Hecke P, Goubel F, Francaux M. Beneficial effects of creatine supplementation in dystrophic patients. *Muscle Nerve* 2003; 27: 604-610 [PMID: 12707981]
- 63 Parise G, Mihic S, MacLennan D, Yarasheski KE, Tarnopolsky MA. Effects of acute creatine monohydrate supplementation on leucine kinetics and mixed-muscle protein synthesis. *J Appl Physiol* (1985) 2001; 91: 1041-1047 [PMID: 11509496]
- 64 Hespel P, Op't Eijnde B, Van Leemputte M, Ursø B, Greenhaff PL, Labarque V, Dymarkowski S, Van Hecke P, Richter EA. Oral creatine supplementation facilitates the rehabilitation of disuse atrophy and alters the expression of muscle myogenic factors in humans. *J Physiol* 2001; **536**: 625-633 [PMID: 11600695]
- 65 **Deldicque L**, Louis M, Theisen D, Nielens H, Dehoux M, Thissen JP, Rennie MJ, Francaux M. Increased IGF mRNA in human skeletal muscle after creatine supplementation. *Med Sci Sports Exerc* 2005; **37**: 731-736 [PMID: 15870625]
- 66 Deldicque L, Atherton P, Patel R, Theisen D, Nielens H, Rennie MJ, Francaux M. Effects of resistance exercise with and without creatine supplementation on gene expression and cell signaling in human skeletal muscle. *J Appl Physiol* (1985) 2008; **104**: 371-378 [PMID: 18048590]
- 67 Burke DG, Candow DG, Chilibeck PD, MacNeil LG, Roy BD, Tarnopolsky MA, Ziegenfuss T. Effect of creatine supplementation and resistance-exercise training on muscle insulin-like growth factor in young adults. *Int J Sport Nutr Exerc Metab* 2008; **18**: 389-398 [PMID: 18708688]
- 68 Adams GR. Invited Review: Autocrine/paracrine IGF-I and skeletal muscle adaptation. J Appl Physiol (1985) 2002; 93: 1159-1167 [PMID: 12183514]
- 69 Saremi A, Gharakhanloo R, Sharghi S, Gharaati MR, Larijani B, Omidfar K. Effects of oral creatine and resistance training on serum myostatin and GASP-1. *Mol Cell Endocrinol* 2010; 317: 25-30 [PMID: 20026378 DOI: 10.1016/j.mce.2009.12.019]
- 70 **Schiaffino S**, Dyar KA, Ciciliot S, Blaauw B, Sandri M. The role of myostatin in muscle wasting: an overview. *J Cachexia Sarcopenia Muscle* 2011; **2**: 143-151 [PMID: 21966641]
- 71 Schiaffino S, Dyar KA, Ciciliot S, Blaauw B, Sandri M. Mechanisms regulating skeletal muscle growth and atrophy. *FEBS J* 2013; 280: 4294-4314 [PMID: 23517348 DOI: 10.1111/ febs.12253]
- 72 Zimmers TA, Davies MV, Koniaris LG, Haynes P, Esquela AF, Tomkinson KN, McPherron AC, Wolfman NM, Lee SJ. Induction of cachexia in mice by systemically administered myostatin. *Science* 2002; 296: 1486-1488 [PMID: 12029139]
- 73 **Dankbar B**, Wunrau C, Wehmeyer C, Pap T. Myostatin-a new player in inflammatory bone loss. *Ann Rheum Diseases*

2011; 70: A75-A76

- 74 Söderlund K, Greenhaff PL, Hultman E. Energy metabolism in type I and type II human muscle fibres during short term electrical stimulation at different frequencies. *Acta Physiol Scand* 1992; 144: 15-22 [PMID: 1595349]
- 75 Wortmann RL. Inflammatory diseases of muscle. Textbook of Rheumatology. 4th ed. Philadelphia: WB Saunders Company, 1993: 1159–1188
- 76 Choy E. Understanding the dynamics: pathways involved in the pathogenesis of rheumatoid arthritis. *Rheumatology* (Oxford) 2012; **51** Suppl 5: v3-v11 [PMID: 22718924 DOI: 10.1093/rheumatology/kes113]
- 77 Bassit RA, Curi R, Costa Rosa LF. Creatine supplementation reduces plasma levels of pro-inflammatory cytokines and PGE2 after a half-ironman competition. *Amino Acids* 2008; 35: 425-431 [PMID: 17917696]
- 78 Morley JE, Baumgartner RN, Roubenoff R, Mayer J, Nair KS. Sarcopenia. J Lab Clin Med 2001; 137: 231-243 [PMID: 11283518]
- 79 Candow DG, Little JP, Chilibeck PD, Abeysekara S, Zello GA, Kazachkov M, Cornish SM, Yu PH. Low-dose creatine combined with protein during resistance training in older men. *Med Sci Sports Exerc* 2008; 40: 1645-1652 [PMID: 18685526 DOI: 10.1249/MSS.0b013e318176b310]
- 80 **Chilibeck PD**, Chrusch MJ, Chad KE, Shawn Davison K, Burke DG. Creatine monohydrate and resistance training increase bone mineral content and density in older men. *J Nutr Health Aging* 2005; **9**: 352-353 [PMID: 16222402]
- 81 **Tarnopolsky MA**. Potential benefits of creatine monohydrate supplementation in the elderly. *Curr Opin Clin Nutr Metab Care* 2000; **3**: 497-502 [PMID: 11085837]
- 82 Wiroth JB, Bermon S, Andreï S, Dalloz E, Hébuterne X, Dolisi C. Effects of oral creatine supplementation on maximal pedalling performance in older adults. *Eur J Appl Physiol* 2001; 84: 533-539 [PMID: 11482548]
- 83 Skare OC, Skadberg AR. Creatine supplementation improves sprint performance in male sprinters. *Scand J Med Sci Sports* 2001; 11: 96-102 [PMID: 11252467]
- 84 Mujika I, Padilla S, Ibañez J, Izquierdo M, Gorostiaga E. Creatine supplementation and sprint performance in soccer players. *Med Sci Sports Exerc* 2000; 32: 518-525 [PMID: 10694141]
- 85 McNaughton LR, Dalton B, Tarr J. The effects of creatine supplementation on high-intensity exercise performance in elite performers. *Eur J Appl Physiol Occup Physiol* 1998; 78: 236-240 [PMID: 9721002]
- 86 Rossiter HB, Cannell ER, Jakeman PM. The effect of oral creatine supplementation on the 1000-m performance of competitive rowers. J Sports Sci 1996; 14: 175-179 [PMID: 8737325]
- 87 Harris RC, Viru M, Greenhaff PL, Hultman E. The effect of oral creatine supplementation on running performance during maximal short-term exercise in man. J Physiol 1993; 467: 74-74
- 88 Rawson ES, Volek JS. Effects of creatine supplementation and resistance training on muscle strength and weightlifting performance. J Strength Cond Res 2003; 17: 822-831 [PMID: 14636102]
- 89 **Volek JS**, Kraemer WJ, Bush JA, Boetes M, Incledon T, Clark KL, Lynch JM. Creatine supplementation enhances muscular performance during high-intensity resistance exercise. *J Am Diet Assoc* 1997; **97**: 765-770 [PMID: 9216554]
- 90 Larson-Meyer DE, Hunter GR, Trowbridge CA, Turk JC, Ernest JM, Torman SL, Harbin PA. The effect of creatine supplementation on muscle strength and body composition during off-season training in female soccer players. J Strength Con Res 2000; 14: 434-442
- 91 Kreider RB. Effects of creatine supplementation on performance and training adaptations. *Mol Cell Biochem* 2003; 244: 89-94 [PMID: 12701815]
- 92 Stroud MA, Holliman D, Bell D, Green AL, Macdonald IA,

Greenhaff PL. Effect of oral creatine supplementation on respiratory gas exchange and blood lactate accumulation during steady-state incremental treadmill exercise and recovery in man. *Clin Sci* (Lond) 1994; **87**: 707-710 [PMID: 7874863]

- 93 Chilibeck PD, Magnus C, Anderson M. Effect of in-season creatine supplementation on body composition and performance in rugby union football players. *Appl Physiol Nutr Metab* 2007; 32: 1052-1057 [PMID: 18059577]
- 94 Willer B, Stucki G, Hoppeler H, Brühlmann P, Krähenbühl S. Effects of creatine supplementation on muscle weakness in patients with rheumatoid arthritis. *Rheumatology* (Oxford) 2000; **39**: 293-298 [PMID: 10788538]
- 95 Stucki G, Schönbächler J, Brühlmann P, Mariacher S, Stoll T, Michel BA. Does a muscle strength index provide complementary information to traditional disease activity variables in patients with rheumatoid arthritis? *J Rheumatol* 1994; 21: 2200-2205 [PMID: 7699619]
- 96 van den Ende CH, Breedveld FC, Dijkmans BA, Hazes JM. The limited value of the Health Assessment Questionnaire as an outcome measure in short term exercise trials. *J Rheumatol* 1997; 24: 1972-1977 [PMID: 9330941]
- 97 Cañete S, San Juan AF, Pérez M, Gómez-Gallego F, López-Mojares LM, Earnest CP, Fleck SJ, Lucia A. Does creatine supplementation improve functional capacity in elderly women? J Strength Cond Res 2006; 20: 22-28 [PMID: 16503684]
- 98 Gotshalk LA, Kraemer WJ, Mendonca MA, Vingren JL, Kenny AM, Spiering BA, Hatfield DL, Fragala MS, Volek JS. Creatine supplementation improves muscular performance in older women. *Eur J Appl Physiol* 2008; **102**: 223-231 [PMID: 17943308]
- 99 Rawson ES, Wehnert ML, Clarkson PM. Effects of 30 days of creatine ingestion in older men. Eur J Appl Physiol Occup Physiol 1999; 80: 139-144 [PMID: 10408325]
- 100 Rawson ES, Clarkson PM. Acute creatine supplementation in older men. Int J Sports Med 2000; 21: 71-75 [PMID: 10683103]
- 101 Stout JR, Sue Graves B, Cramer JT, Goldstein ER, Costa PB, Smith AE, Walter AA. Effects of creatine supplementation on the onset of neuromuscular fatigue threshold and muscle strength in elderly men and women (64 - 86 years). J Nutr Health Aging 2007; 11: 459-464 [PMID: 17985060]
- 102 Jakobi JM, Rice CL, Curtin SV, Marsh GD. Neuromuscular properties and fatigue in older men following acute creatine supplementation. *Eur J Appl Physiol* 2001; 84: 321-328 [PMID: 11374116]
- 103 Bermon S, Venembre P, Sachet C, Valour S, Dolisi C. Effects of creatine monohydrate ingestion in sedentary and weighttrained older adults. *Acta Physiol Scand* 1998; 164: 147-155 [PMID: 9805101]
- 104 **Gualano B**, Macedo AR, Alves CR, Roschel H, Benatti FB, Takayama L, de Sá Pinto AL, Lima FR, Pereira RM. Creatine supplementation and resistance training in vulnerable older women: A randomized double-blind placebo-controlled clinical trial. *Exp Gerontol* 2014; **53**: 7-15
- 105 Neves M, Gualano B, Roschel H, Lima FR, Lúcia de Sá-Pinto A, Seguro AC, Shimizu MH, Sapienza MT, Fuller R, Lancha AH, Bonfá E. Effect of creatine supplementation on measured glomerular filtration rate in postmenopausal women. *Appl Physiol Nutr Metab* 2011; 36: 419-422 [PMID: 21574777]
- 106 Roy BD, de Beer J, Harvey D, Tarnopolsky MA. Creatine monohydrate supplementation does not improve functional recovery after total knee arthroplasty. *Arch Phys Med Rehabil* 2005; 86: 1293-1298 [PMID: 16003653]
- 107 Slemenda C, Brandt KD, Heilman DK, Mazzuca S, Braunstein EM, Katz BP, Wolinsky FD. Quadriceps weakness and osteoarthritis of the knee. *Ann Intern Med* 1997; 127: 97-104 [PMID: 9230035]
- 108 Lawrence RC, Felson DT, Helmick CG, Arnold LM, Choi H, Deyo RA, Gabriel S, Hirsch R, Hochberg MC, Hunder GG, Jordan JM, Katz JN, Kremers HM, Wolfe F. Estimates of the

prevalence of arthritis and other rheumatic conditions in the United States. Part II. *Arthritis Rheum* 2008; **58**: 26-35 [PMID: 18163497 DOI: 10.1002/art.23176]

- 109 Alves CR, Santiago BM, Lima FR, Otaduy MC, Calich AL, Tritto AC, de Sá Pinto AL, Roschel H, Leite CC, Benatti FB, Bonfá E, Gualano B. Creatine supplementation in fibromyalgia: a randomized, double-blind, placebo-controlled trial. *Arthritis Care Res* (Hoboken) 2013; 65: 1449-1459 [PMID: 23554283 DOI: 10.1002/acr.22020]
- 110 Leader A, Amital D, Rubinow A, Amital H. An open-label study adding creatine monohydrate to ongoing medical regimens in patients with the fibromyalgia syndrome. *Ann N Y Acad Sci* 2009; **1173**: 829-836 [PMID: 19758235 DOI: 10.1111/ j.1749-6632.2009.04811.x]
- 111 Kley RA, Tarnopolsky MA, Vorgerd M. Creatine for treating muscle disorders. *Cochrane Database Syst Rev* 2011;
 (2): CD004760 [PMID: 21328269 DOI: 10.1002/14651858. CD004760.pub3]
- 112 Tarnopolsky MA, Mahoney DJ, Vajsar J, Rodriguez C, Doherty TJ, Roy BD, Biggar D. Creatine monohydrate enhances strength and body composition in Duchenne muscular dystrophy. *Neurology* 2004; 62: 1771-1777 [PMID: 15159476]
- 113 Banerjee B, Sharma U, Balasubramanian K, Kalaivani M, Kalra V, Jagannathan NR. Effect of creatine monohydrate in improving cellular energetics and muscle strength in ambulatory Duchenne muscular dystrophy patients: a randomized, placebo-controlled 31P MRS study. *Magn Reson Imaging* 2010; 28: 698-707 [PMID: 20395096 DOI: 10.1016/j.mri.2010.03.008]
- 114 Walter MC, Reilich P, Lochmüller H, Kohnen R, Schlotter B, Hautmann H, Dunkl E, Pongratz D, Müller-Felber W. Creatine monohydrate in myotonic dystrophy: a doubleblind, placebo-controlled clinical study. J Neurol 2002; 249: 1717-1722 [PMID: 12529796]
- 115 **Tarnopolsky M**, Mahoney D, Thompson T, Naylor H, Doherty TJ. Creatine monohydrate supplementation does not increase muscle strength, lean body mass, or muscle phosphocreatine in patients with myotonic dystrophy type 1. *Muscle Nerve* 2004; **29**: 51-58 [PMID: 14694498]
- 116 Escolar DM, Buyse G, Henricson E, Leshner R, Florence J, Mayhew J, Tesi-Rocha C, Gorni K, Pasquali L, Patel KM, Mc-Carter R, Huang J, Mayhew T, Bertorini T, Carlo J, Connolly AM, Clemens PR, Goemans N, Iannaccone ST, Igarashi M, Nevo Y, Pestronk A, Subramony SH, Vedanarayanan VV, Wessel H. CINRG randomized controlled trial of creatine and glutamine in Duchenne muscular dystrophy. *Ann Neurol* 2005; 58: 151-155 [PMID: 15984021]
- 117 Schneider-Gold C, Beck M, Wessig C, George A, Kele H, Reiners K, Toyka KV. Creatine monohydrate in DM2/ PROMM: a double-blind placebo-controlled clinical study. Proximal myotonic myopathy. *Neurology* 2003; 60: 500-502 [PMID: 12578937]
- 118 Norman K, Stübler D, Baier P, Schütz T, Ocran K, Holm E, Lochs H, Pirlich M. Effects of creatine supplementation on nutritional status, muscle function and quality of life in patients with colorectal cancer--a double blind randomised controlled trial. *Clin Nutr* 2006; 25: 596-605 [PMID: 16701923]
- 119 Bourgeois JM, Nagel K, Pearce E, Wright M, Barr RD, Tarnopolsky MA. Creatine monohydrate attenuates body fat accumulation in children with acute lymphoblastic leukemia during maintenance chemotherapy. *Pediatr Blood Cancer* 2008; 51: 183-187 [PMID: 18421708 DOI: 10.1002/pbc.21571]
- 120 Tan BH, Fearon KC. Cachexia: prevalence and impact in medicine. Curr Opin Clin Nutr Metab Care 2008; 11: 400-407 [PMID: 18541999 DOI: 10.1097/MCO.0b013e328300ecc1]
- 121 **Fearon K**, Strasser F, Anker SD, Bosaeus I, Bruera E, Fainsinger RL, Jatoi A, Loprinzi C, MacDonald N, Mantovani G, Davis M, Muscaritoli M, Ottery F, Radbruch L, Ravasco P, Walsh D, Wilcock A, Kaasa S, Baracos VE. Definition and

Wilkinson TJ et al. Oral creatine supplementation

classification of cancer cachexia: an international consensus. *Lancet Oncol* 2011; **12**: 489-495 [PMID: 21296615 DOI: 10.1016/S1470-2045(10)70218-7]

- 122 Symmons D, Turner G, Webb R, Asten P, Barrett E, Lunt M, Scott D, Silman A. The prevalence of rheumatoid arthritis in the United Kingdom: new estimates for a new century. *Rheumatology* (Oxford) 2002; **41**: 793-800 [PMID: 12096230]
- 123 Rawson ES, Venezia AC. Use of creatine in the elderly and evidence for effects on cognitive function in young and old. *Amino Acids* 2011; 40: 1349-1362 [PMID: 21394604 DOI: 10.1007/s00726-011-0855-9]
- 124 Aguiar AF, Januário RS, Junior RP, Gerage AM, Pina FL, do Nascimento MA, Padovani CR, Cyrino ES. Long-term creatine supplementation improves muscular performance during resistance training in older women. *Eur J Appl Physiol* 2013; **113**: 987-996 [PMID: 23053133 DOI: 10.1007/s00421-012 -2514-623053133]
- 125 Brose A, Parise G, Tarnopolsky MA. Creatine supplementation enhances isometric strength and body composition improvements following strength exercise training in older adults. J Gerontol A Biol Sci Med Sci 2003; 58: 11-19 [PMID: 12560406]
- 126 Moon A, Heywood L, Rutherford S, Cobbold C. Creatine supplementation: can it improve quality of life in the elderly without associated resistance training? *Curr Aging Sci* 2013; 6: 251-257 [PMID: 24304199]
- 127 Devries MC, Phillips SM. Creatine supplementation during resistance training in older adults-a meta-analysis. *Med Sci Sports Exerc* 2014; 46: 1194-1203 [PMID: 24576864]
- 128 Chung YL, Alexanderson H, Pipitone N, Morrison C, Dastmalchi M, Ståhl-Hallengren C, Richards S, Thomas EL, Hamilton G, Bell JD, Lundberg IE, Scott DL. Creatine supplements in patients with idiopathic inflammatory myopathies who are clinically weak after conventional pharmacologic treatment: Six-month, double-blind, randomized, placebocontrolled trial. *Arthritis Rheum* 2007; 57: 694-702 [PMID: 17471547]
- 129 Syrotuik DG, Bell GJ, Burnham R, Sim IL, Calvert RA, Maclean IM. Absolute and relative strength performance following creatine monohydrate supplementation combined with periodized resistance training. J Strength Con Res 2000; 14: 182-190
- 130 Harris RC, Söderlund K, Hultman E. Elevation of creatine in resting and exercised muscle of normal subjects by creatine supplementation. *Clin Sci* (Lond) 1992; 83: 367-374 [PMID: 1327657]
- 131 Delanghe J, De Slypere JP, De Buyzere M, Robbrecht J, Wieme R, Vermeulen A. Normal reference values for creatine, creatinine, and carnitine are lower in vegetarians. *Clin Chem* 1989; 35: 1802-1803 [PMID: 2758659]

- 132 **Stec ES**, Rawson MJ. Benefits of creatine supplementation for older adults. *Braz J Biomotric* 2010; **4**: 215-226
- 133 Poortmans JR, Auquier H, Renaut V, Durussel A, Saugy M, Brisson GR. Effect of short-term creatine supplementation on renal responses in men. *Eur J Appl Physiol Occup Physiol* 1997; 76: 566-567 [PMID: 9404870]
- 134 Poortmans JR, Francaux M. Renal dysfunction accompanying oral creatine supplements. *Lancet* 1998; 352: 234 [PMID: 9683236]
- 135 Earnest CP, Almada AL, Mitchell TL. High-performance capillary electrophoresis-pure creatine monohydrate reduces blood lipids in men and women. *Clin Sci* (Lond) 1996; 91: 113-118 [PMID: 8774269]
- 136 **Lemon PW**. Dietary creatine supplementation and exercise performance: why inconsistent results? *Can J Appl Physiol* 2002; **27**: 663-681 [PMID: 12501003]
- 137 Walliman T. Comment on "Creatine is an approved, effective and safe dietary supplement that is NOT causing toxic hepatitis!". *Food Chem Toxicol* 2013; 51: 453–454
- 138 Shao A, Hathcock JN. Risk assessment for creatine monohydrate. *Regul Toxicol Pharmacol* 2006; 45: 242-251 [PMID: 16814437]
- 139 Gualano B, Ugrinowitsch C, Novaes RB, Artioli GG, Shimizu MH, Seguro AC, Harris RC, Lancha AH. Effects of creatine supplementation on renal function: a randomized, double-blind, placebo-controlled clinical trial. *Eur J Appl Physiol* 2008; **103**: 33-40 [PMID: 18188581 DOI: 10.1007/ s00421-007-0669-3]
- 140 Mayhew DL, Mayhew JL, Ware JS. Effects of long-term creatine supplementation on liver and kidney functions in American college football players. *Int J Sport Nutr Exerc Metab* 2002; 12: 453-460 [PMID: 12500988]
- 141 Schröder H, Terrados N, Tramullas A. Risk assessment of the potential side effects of long-term creatine supplementation in team sport athletes. *Eur J Nutr* 2005; 44: 255-261 [PMID: 15309421]
- 142 Spillane M, Schoch R, Cooke M, Harvey T, Greenwood M, Kreider R, Willoughby DS. The effects of creatine ethyl ester supplementation combined with heavy resistance training on body composition, muscle performance, and serum and muscle creatine levels. *J Int Soc Sports Nutr* 2009; 6: 6 [PMID: 19228401 DOI: 10.1186/1550-2783-6-6]
- 143 Vorgerd M, Grehl T, Jager M, Muller K, Freitag G, Patzold T, Bruns N, Fabian K, Tegenthoff M, Mortier W, Luttmann A, Zange J, Malin JP. Creatine therapy in myophosphorylase deficiency (McArdle disease): a placebo-controlled crossover trial. *Arch Neurol* 2000; 57: 956-963 [PMID: 10891977]
- 144 Hultman E, Söderlund K, Timmons JA, Cederblad G, Greenhaff PL. Muscle creatine loading in men. J Appl Physiol (1985) 1996; 81: 232-237 [PMID: 8828669]

P-Reviewer: Kemal NAS, Turiel M S-Editor: Ji FF L-Editor: A E-Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.35 World J Rheumatol 2014 November 12; 4(3): 35-43 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

REVIEW

Gender differences in axial spondyloarthritis

Regula Neuenschwander, Adrian Ciurea

Regula Neuenschwander, Adrian Ciurea, Department of Rheumatology, University Hospital Zurich, 8091 Zurich, Switzerland Author contributions: Neuenschwander R and Ciurea A were involved in drafting the article and revising it critically for important intellectual content and English language; Both authors approved the final version to be published.

Correspondence to: Adrian Ciurea, MD, Department of Rheumatology, University Hospital Zurich, Gloriastrasse 25, 8081 Zurich, Switzerland. adrian.ciurea@usz.ch

Telephone: +41-44-2552932 Fax: +41-44-2554415 Received: July 11, 2014 Revised: August 18, 2014 Accepted: September 18, 2014 Published online: November 12, 2014

Abstract

Within the concept of axial spondyloarthritis (axSpA), relevant differences between men and women have been described for patients with the radiographic disease form [ankylosing spondylitis (AS)]. The subjective perception of disease activity (spinal and peripheral pain, fatigue, morning stiffness) has been shown to be higher in female than in male patients. Moreover, women experience more functional limitations and a lower quality of life, despite lower degrees of radiographic spinal damage. Peripheral clinical involvement (arthritis and enthesitis) is, additionally, more predominant in women. On the other hand, a higher level of objective signs of inflammation (C-reactive protein, erythrocyte sedimentation rate, magnetic resonance imaging of sacroiliac joints and spine) has been reported in men. Whether these differences might explain the better response to treatment with anti-tumor necrosis factor agents observed in male patients remains unclear. The underlying causes of the discrepancies are still unknown and genetic, environmental, cultural and/or societal factors may be involved. While AS is still more prevalent in men in a ratio of 2-3:1, the prevalence of males and females in patients with axSpA without radiographic sacroiliac damage is similar. Gender differences in this subgroup of patients have not been adequately addressed, and are particularly needed to further validate the Assessment of SpondyloArthritis international Society classification criteria.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Axial spondyloarthritis; Ankylosing spondylitis; Classification; Gender; Outcome

Core tip: In comparison to men, women with ankylosing spondylitis (AS) experience a higher subjective burden of disease despite lower objective signs of systemic inflammation and less spinal radiographic damage. A better response to treatment with tumor necrosis factor inhibitors has been demonstrated in male AS patients.

Neuenschwander R, Ciurea A. Gender differences in axial spondyloarthritis. *World J Rheumatol* 2014; 4(3): 35-43 Available from: URL: http://www.wjgnet.com/2220-3214/full/v4/i3/35.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.35

INTRODUCTION

Ankylosing spondylitis (AS) is still considered the prototype disease of a group of inflammatory rheumatic conditions, referred to as spondyloarthritides (SpA), and characterized by inflammation of the sacroiliac joints (SIJ), the spine, as well as peripheral joints and entheses^[1,2]. It was traditionally associated with a long diagnostic delay^[3], as the defining radiographic changes of the SIJ, described by the modified New York criteria^[4], usually develop gradually over several years. It is now regarded as part of a disease continuum, referred to as axial SpA (axSpA), defined by the 2009 Assessment of Spondylo-Arthritis international Society (ASAS) classification criteria^[5]. Within this concept, AS, also called radiographic axSpA, is opposed to nonradiographic (nr-) axSpA^[6,7]. In the absence of definite radiographic SIJ damage, patients can be classified as having nr-axSpA either in the presence of sacroiliitis on magnetic resonance imaging (MRI) plus at least one SpA feature or in the presence of human Leukocyte Antigen (HLA)-B27 plus at least two SpA



features^[5]. Relevant differences between men and women have been delineated for the AS subgroup and this review will particularly focus on new data published after the extensive 2008 survey by Lee *et al*^[8]. Recent studies have also highlighted differences in sex distribution between AS and nr-axSpA^[9-12]. Data on gender differences in the nr-axSpA subgroup are, however, only beginning to emerge^[13].

GENDER DIFFERENCES IN ANKYLOSING SPONDYLITIS

Distribution

AS was believed to affect predominantly men, with a ratio of 10 males for every female patient^[14]. It remained a relevant example of sex biased research for many decades as it was often carried out in military or veteran's hospitals. Underestimation of disease prevalence in the female population might have additionally been due to differences in disease phenotype, reluctance to perform X-rays of the pelvis and lumbar spine in women of child-bearing age, gender differences in the act of seeking a doctor's advice or a faster investigative approach in men with back pain and stiffness in physically demanding jobs. Studies conducted in the last decade reported a much smaller sex distribution difference, still favoring males, in the order of 2-3:1, also reflecting progress in imaging technologies and changing gender roles^[9,11,15]. A recent systematic analysis of 13 cross-sectional population studies revealed a mean gender ratio of 3.4:1 and some differences between geographic regions (3.8:1 in Europe and 2.3:1 in Asia)^[16].

Pathogenesis

The strong genetic association of AS with HLA-B27, discovered by Brewerton and Schlosstein in 1973^[17,18], was subsequently shown to presumably not be implicated in the unequal sex distribution, as the prevalence of HLA-B27 was similar in women and men^[19]. These findings should be confirmed in larger population studies, as the proportion of HLA-B27 positivity was significantly lower in women than in men in recent treatment studies^[20,21] and in the Swiss Clinical Quality Management (SCQM) axSpA cohort^[22]. While only a quarter of the genetic susceptibility to AS is currently explained by HLA-B27, sex differences have not been addressed in other discovered major histocompatibility complex (MHC)- or non-MHC genetic associations involved in the innate immune stimulation, the interleukin-23 pathway or peptide presentation^[23]. Potential sex differences have been described for the ANKH gene, coding for a protein regulating the cellular export of inorganic pyrophosphate^[24]. The association of ANKH with disease susceptibility remains, however, controversial^[25,26]. By contrast, no evidence for an involvement of the X-chromosome in the development of AS could be detected^[2/].</sup>

Genetic factors may also have an indirect impact on disease susceptibility by interacting with environmental factors influenced by gender. HLA-B27 is particularly interesting in this regard, as it might interact with bacterial antigens^[28]. The triggering role of Chlamydia trachomatis is clearly established for the development of reactive arthritis after urogenital infection with this bacterium and the disease has an important male predominance (up to 20:1)^[29]. A significant proportion of HLA-B27-positive patients with Chlamydia-induced SpA will eventually develop AS and Chlamydia trachomatis has also been detected by polymerase chain reaction in synovial tissues from patients with other SpA forms^[30].

Smoking represents another environmental and societal factor potentially influencing gender differences in rad-axSpA. Although studies linking smoking with incident AS are lacking, smoking was demonstrated to be associated with increased disease activity, impaired function and poorer quality of life in cross-sectional analyses^[31-38]. Furthermore, it was associated with future radiographic spinal progression in longitudinal studies^[39,40], probably through an amplifying effect of disease activity on radiographic damage^[41]. While the prevalence of smoking is higher in men than in women with AS^[38,42], a causal effect between smoking and gender differences in disease outcome, as depicted below, remains to be established.

No differences in gonadal and adrenal sex hormones have been identified in male and female patients with AS in comparison to healthy controls^[42]. A recent analysis of 448 women using the oral contraceptive pill in comparison to 123 female non-users failed to demonstrate any effect of the use of exogenous estrogens on the initiation or on the severity of AS^[43].

Disease onset and diagnostic delay

The age at symptom onset was similar in women and men with AS in most of the studies available to date, while a longer duration from disease onset to diagnosis has been detected in women^[3,44]. However, HLA-B27 has been shown to have a strong effect not only on the age at symptom onset^[9,21], but also on diagnostic delay^[15], and was not available in all patients. Additional investigations are therefore needed. We found an earlier disease onset in male patients in a large cohort of 1199 patients with AS (mean age at onset 26.3 years in men vs 29.3 years in women, P < 0.001)^[22]. The documented differences in HLA-B27 prevalence in men and women in this study (see above) may not fully account for the gender difference in disease onset, as women were on average 1.8 years older than men at the beginning of back pain in the subgroup of HLA-B27-positive patients^[22].

Signs and symptoms

A multitude of comparisons of male and female patients with AS have provided data on women having more pain at the level of the cervical spine and in peripheral joints, with the hip joints being more often involved in men^[45-56]. The intensity of symptoms (spinal pain, peripheral joint pain and swelling, areas of localized tenderness, fatigue, morning stiffness) was substantiated with the patientreported Bath Ankylosing Spondylitis Disease Activity Index (BASDAI)^[57] in the more recent literature. Women

36

presented with higher BASDAI, global pain, nocturnal pain, joint pain and fatigue scores in comparison to men^[58-66]. As these common symptoms overlap with the clinical features of fibromyalgia, which is more prevalent in women with AS than in men^[67-69], additional objective parameters to assess disease activity seem advisable. Whether the use of the recently defined Ankylosing Spondylitis Disease Activity Score (ASDAS) will prove helpful in this regard, as its calculation includes acute phase reactants [erythrocyte sedimentation rate (ESR) or C-reactive protein (CRP)] in addition to patient-reported parameters^[70], remains to be established.

Clinical findings

The report of women being more often treated with methotrexate, sulfasalazine and prednisone in an North American cohort suggested a higher prevalence of peripheral arthritis in women with AS^[56]. When analyzing the presence of current or previous peripheral synovitis, these gender differences with regard to the presence of peripheral arthritis in AS have been substantiated^[62,71]. In the Swiss SCQM axSpA Cohort (826 men and 373 women with AS), a significantly higher percentage of women had peripheral synovitis as well as a higher number of swollen joints at inclusion, while no differences have been observed with regard to the percentage of men and women with previous peripheral arthritis (unpublished results). The discrepancies might be explained by longer disease duration in men in this cohort, allowing for more clinical manifestations to accumulate. As demonstrated in another registry, the presence of peripheral arthritis seems to delay spinal radiographic progression (see below), independently of other confounding factors, including gender^[72].

Peripheral enthesitis evaluated by the Maastricht Ankylosing Spondylitis Enthesitis Score was also more frequently found in women in comparison to men with AS^[62,73]. The finding might be confounded by a potential overlap between enthesitic and fibromyalgia tender points.

Spinal mobility, as assessed by different clinical parameters, such as the tragus-to-wall distance, the Schober's s test or the Bath Ankylosing Spondylitis Metrology Index in more recent studies, was consistently more greatly impaired in women in comparison to men. Spinal mobility cannot be used as a proxy for radiographic damage in an individual patient^[74], as both inflammation and structural damage have been shown to contribute to its impairment^[75].

With regard to extra-articular manifestations - acute anterior uveitis, psoriasis and inflammatory bowel disease-only bowel inflammation was positively associated with the percentage of women in the evaluated AS studies of a systematic review and meta-analysis of 156 selected articles^[76].

Imaging studies

Radiographic differences between genders in AS have been analyzed in numerous studies^[34,45,46,48,49,51-56,62,77]. After

adjustment for age and disease duration and using either the Bath Ankylosing Spondylitis Radiography Index or the modified Stoke Ankylosing Spondylitis Spinal Score (mSASSS) in the more recent literature, men consistently presented with more important spinal changes on X-rays in comparison to women. This was confirmed in a recent 12-year prospective follow-up analysis, especially in the presence of HLA-B27 positivity^[78].

Male sex was also associated with MRI inflammation of the SIJ in several studies of patients with axSpA, including patients with AS, while HLA-B27 was an independent predictor of future MRI positivity^[21,79,80].

Laboratory findings

While ESR and CRP levels have been demonstrated to be higher in women in comparison to men in the general population^[81-83], acute phase reactants were either similar, or slightly more elevated in male patients with AS in comparison to females, suggesting a higher level of systemic inflammation in men^[20,59-61,84,85]. Elevated CRP was shown to be associated with radiographic progression in AS^[39]. As smoking is associated with pro-inflammatory effects and may raise CRP levels in a non-specific manner^[86], it remains unclear, whether higher acute phase reactant levels in AS might be explained in part by the higher prevalence of smokers in the male AS population.

Disease activity and radiographic progression

Higher disease activity, as measured by the ASDAS, which includes both patient-reported outcomes and acute-phase reactants, led to more spinal structural damage, especially syndesmophyte formation, in a recent 12-year longitudinal study^[78]. The authors highlighted the fact that the ASDAS outperformed all other disease activity measures (BASDAI and CRP, patient-reported global activity and CRP, back pain and CRP) in this analysis. A significant interaction was found between ASDAS and gender: the effect of ASDAS on the change in mSASSS was higher in males versus females (0.98 vs -0.06 units per 2 years and per additional unit of ASDAS). Whether an association exists between important mechanical forces acting on the spine (observed in male-dominated more intensive occupational activities) and formation of syndesmophytes, should be confirmed in future studies.

Functional outcomes

Self-reported functional ability in performing daily activities is usually assessed with the Bath Ankylosing Spondylitis Functional Index. A similar level of functional impairment has been demonstrated in women and men with AS^[20,56,61,64]. After adjustment for the level of radiographic spinal damage, however, the disability observed was more pronounced in female in comparison to male patients^[56]. The documented higher level of peripheral symptoms (arthritis and enthesitis) in women might be regarded as a confounder in this analysis^[87].

Quality of life

Health-related quality of life encompasses the individual



well-being considering social, emotional and physical aspects, as well as the effect of disease on a patient's wellbeing, mainly measured by the Short-Form Health Survey (SF-36) and the Ankylosing Spondylitis Quality of Life Questionnaire (ASQoL). It is significantly impaired in AS^[88]. In unadjusted analyses, women reported a significantly worse quality of life and a greater reduction in vitality than men^[88-90]. However, after adjustment for the normal differences in men and women's self-assessment of their health in the general population, the crude effect of AS on the quality of life was greater in men^[88].

Work disability

Analyses of gender differences with regard to work disability in AS led to conflicting results due to differences in economic environment, social security system, disability compensation system in different countries as well as comorbidities, disease duration and proportions of patients with manual jobs in the respective studies^[91-100]. The contributing factors to absenteeism (impossibility to attend work, either due to temporary sick leave or permanent worker disability) and presenteeism (reduced performance or productivity at work due to health reasons) in AS patients have been analyzed recently^[101]. Female sex, an impaired health-related quality of life and helplessness (a personal factor) were associated with presenteeism, while at-work limitations and lower quality of life contributed to sick leave.

Family life

In a US analysis, patients with AS were more likely to have never been married and men were more likely to divorce, especially in longstanding disease^[102]. In the same study, women with AS were less likely to have children than women in the general population, in contrast to men. The authors postulate that women might be more sensitive to the concern about having children with AS. The quality of marital life was shown to be characterized by a higher global distress and a higher probability of aggression from their partner in female SpA patients from Korea^[103]. Furthermore, in comparison to healthy individuals in Europe, a higher proportion of women with SpA was shown to be single or divorced^[104]. Disease activity was, moreover, higher in divorced than married female SpA patients. With regard to inheritance, a higher prevalence of AS among children and siblings of female index cases has been demonstrated, suggesting that women may require a higher genetic load to develop the disease^[44, 105]. The higher prevalence of a family history of SpA found in female patients with axSpA^[13,56,62,64], would be compatible with this hypothesis.

Response to treatment

Tumor necrosis factor (TNF) inhibitor treatment is recommended in patients with highly active AS and insufficient response to non-steroidal anti-inflammatory drugs^[106]. Elevated acute phase reactants have been shown to represent the most important predictor of treatment response^[84,107-113]. Gender differences in response to TNF inhibition or treatment survival have also been identified in most studies, which were, however, not adjusted for all known confounding factors. In the Groningen Leeuwarden AS observational cohort, male gender was a predictor of treatment response, while female gender predicted treatment discontinuation independently of other parameters^[111]. Female gender was also a predictor of anti-TNF-agent discontinuation in the South Swedish Arthritis Treatment Group Register and in the Danish nationwide DANBIO register^[84,113]. In a study of pooled data from four clinical control trials of etanercept, sulfasalazine or placebo in AS, female patients had less improvement in ASDAS than male patients^[20]. In contrast, female gender was an independent predictor of improvement in BASDAI and the Bath Ankylosing Spondylitis Functional Index in the British Society of Rheumatology Biologics Register^[110], while gender did not influence treatment responses in other studies^[107,112,113].

Mortality

An increased mortality was found in men but not women with AS (standard mortality rate 1.63 *vs* 1.38, P < 0.001) in a cohort of 677 patients followed since 1977, with circulatory disease being the most frequent cause of death^[114]. A trend towards increased mortality in women was only found after longer disease duration (35-40 years). The authors postulate that a larger study population with a longer time span of observation might be necessary to demonstrate excess mortality in women with AS. The finding that the increased mortality in AS was related to the degree of disease activity, however, points to a really existing gender difference in this regard.

GENDER DIFFERENCES IN NONRADIOGRAPHIC AXIAL SPONDYLOARTHRITIS

The most striking differences between patients with axSpA fulfilling or not fulfilling the modified New York classification criteria (AS vs nr-axSpA) are the unequal gender distribution (1:1 in nr-axSpA and favoring men in a ratio of 2-3:1 in AS) and the higher level of CRP in AS com-pared to nr-axSpA^[9-12]. Women as well as patients with low systemic inflammation might have a lower risk to develop radiographic spinal damage and remain longer in the nonradiographic disease stage. The risk of misdiagnosis in patients with other reasons for back pain in context of HLA-B27 positivity^[115] would represent an alternative, mutually not exclusive explanation for the higher prevalence of women in the nr-axSpA group. Comparisons between women and men with nr-axSpA, as well as between women with AS and women with nr-axSpA, along with prospective longitudinal data are therefore needed. Tournadre *et al*^[13] have analyzed the differences between female and male patients in the French cohort Devenir des Spondylarthropathies Indifférenciées Récentes of patients classified as having axSpA. Data are also presented in the subgroup of patients classified by the imaging and

WJR www.wjgnet.com

the clinical arms of the ASAS classification, respectively. Only patients in the clinical arm are available for analysis of gender differences in nr-axSpA in this study, as both patients with AS and patients with nr-axSpA are present in the imaging arm by definition. Higher self-reported disease activity parameters and functional impairment were found in women. Multivariate regression models confirmed the relationship between higher levels of BASDAI, ASDAS-CRP, fatigue and ASQoL and female gender^[13].

CONCLUSION

Female AS patients experience higher levels of pain and other self-reported disease activity parameters, a greater impairment of health-related quality of life and a reduced treatment response upon TNF inhibition. On the other hand, male AS patients present with higher objective measures of disease activity (acute phase reactants, inflammation of SIJ on MRI) and a more important radiographic spinal damage. The causes underlying these relevant differences remain largely unknown.

REFERENCES

- 1 **Dougados M**, Baeten D. Spondyloarthritis. *Lancet* 2011; **377**: 2127-2137 [PMID: 21684383]
- 2 van der Heijde D, Maksymowych WP. Spondyloarthritis: state of the art and future perspectives. *Ann Rheum Dis* 2010; 69: 949-954 [PMID: 20444753]
- 3 **Feldtkeller E**, Bruckel J, Khan MA. Scientific contributions of ankylosing spondylitis patient advocacy groups. *Curr Opin Rheumatol* 2000; **12**: 239-247 [PMID: 10910174]
- van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis Rheum* 1984; 27: 361-368 [PMID: 6231933]
- 5 Rudwaleit M, van der Heijde D, Landewé R, Listing J, Akkoc N, Brandt J, Braun J, Chou CT, Collantes-Estevez E, Dougados M, Huang F, Gu J, Khan MA, Kirazli Y, Maksymowych WP, Mielants H, Sørensen IJ, Ozgocmen S, Roussou E, Valle-Oñate R, Weber U, Wei J, Sieper J. The development of Assessment of SpondyloArthritis international Society classification criteria for axial spondyloarthritis (part II): validation and final selection. *Ann Rheum Dis* 2009; 68: 777-783 [PMID: 19297344]
- 6 Sieper J, van der Heijde D. Review: Nonradiographic axial spondyloarthritis: new definition of an old disease? *Arthritis Rheum* 2013; 65: 543-551 [PMID: 23233285 DOI: 10.1002/ art.37803]
- 7 Poddubnyy D, Sieper J. Similarities and differences between nonradiographic and radiographic axial spondyloarthritis: a clinical, epidemiological and therapeutic assessment. *Curr Opin Rheumatol* 2014; 26: 377-383 [PMID: 24807404 DOI: 10.1097/BOR.00000000000071]
- 8 Lee W, Reveille JD, Weisman MH. Women with ankylosing spondylitis: a review. *Arthritis Rheum* 2008; 59: 449-454 [PMID: 18311755 DOI: 10.1002/art.23321]
- 9 Rudwaleit M, Haibel H, Baraliakos X, Listing J, Märker-Hermann E, Zeidler H, Braun J, Sieper J. The early disease stage in axial spondylarthritis: results from the German Spondyloarthritis Inception Cohort. *Arthritis Rheum* 2009; 60: 717-727 [PMID: 19248087 DOI: 10.1002/art.24483]
- 10 **Kiltz U**, Baraliakos X, Karakostas P, Igelmann M, Kalthoff L, Klink C, Krause D, Schmitz-Bortz E, Flörecke M, Bollow

M, Braun J. Do patients with non-radiographic axial spondylarthritis differ from patients with ankylosing spondylitis? *Arthritis Care Res* (Hoboken) 2012; **64**: 1415-1422 [PMID: 22505331 DOI: 10.1002/acr.21688]

- 11 Ciurea A, Scherer A, Exer P, Bernhard J, Dudler J, Beyeler B, Kissling R, Stekhoven D, Rufibach K, Tamborrini G, Weiss B, Müller R, Nissen MJ, Michel BA, van der Heijde D, Dougados M, Boonen A, Weber U. Tumor necrosis factor α inhibition in radiographic and nonradiographic axial spondyloarthritis: results from a large observational cohort. *Arthritis Rheum* 2013; **65**: 3096-3106 [PMID: 23983141 DOI: 10.1002/art.38140]
- 12 Wallis D, Haroon N, Ayearst R, Carty A, Inman RD. Ankylosing spondylitis and nonradiographic axial spondyloarthritis: part of a common spectrum or distinct diseases? *J Rheumatol* 2013; **40**: 2038-2041 [PMID: 24187102]
- 13 Tournadre A, Pereira B, Lhoste A, Dubost JJ, Ristori JM, Claudepierre P, Dougados M, Soubrier M. Differences between women and men with recent-onset axial spondyloarthritis: results from a prospective multicenter French cohort. *Arthritis Care Res* (Hoboken) 2013; 65: 1482-1489 [PMID: 23463610 DOI: 10.1002/acr.22001]
- 14 West HF. Aetiology of Ankylosing Spondylitis. Ann Rheum Dis 1949; 8: 143-148 [PMID: 18623806]
- 15 Feldtkeller E, Khan MA, van der Heijde D, van der Linden S, Braun J. Age at disease onset and diagnosis delay in HLA-B27 negative vs. positive patients with ankylosing spondylitis. *Rheumatol Int* 2003; 23: 61-66 [PMID: 12634937 DOI: 10.1007/s00296-002-0237-4]
- 16 Dean LE, Jones GT, MacDonald AG, Downham C, Sturrock RD, Macfarlane GJ. Global prevalence of ankylosing spondylitis. *Rheumatology* (Oxford) 2014; 53: 650-657 [PMID: 24324212 DOI: 10.1093/rheumatology/ket387]
- 17 Brewerton DA, Hart FD, Nicholls A, Caffrey M, James DC, Sturrock RD. Ankylosing spondylitis and HL-A 27. Lancet 1973; 1: 904-907 [PMID: 4123836]
- 18 Schlosstein L, Terasaki PI, Bluestone R, Pearson CM. High association of an HL-A antigen, W27, with ankylosing spondylitis. N Engl J Med 1973; 288: 704-706 [PMID: 4688372 DOI: 10.1056/NEJM197304052881403]
- 19 Hill HF, Hill AG, Bodmer JG. Clinical diagnosis of ankylosing spondylitis in women and relation to presence of HLA-B27. Ann Rheum Dis 1976; 35: 267-270 [PMID: 984907]
- 20 van der Horst-Bruinsma IE, Zack DJ, Szumski A, Koenig AS. Female patients with ankylosing spondylitis: analysis of the impact of gender across treatment studies. *Ann Rheum Dis* 2013; 72: 1221-1224 [PMID: 23264358 DOI: 10.1136/annrheumdis-2012-202431]
- 21 Chung HY, Machado P, van der Heijde D, D'Agostino MA, Dougados M. HLA-B27 positive patients differ from HLA-B27 negative patients in clinical presentation and imaging: results from the DESIR cohort of patients with recent onset axial spondyloarthritis. *Ann Rheum Dis* 2011; **70**: 1930-1936 [PMID: 21803752 DOI: 10.1136/ard.2011.152975]
- 22 Ciurea A, Scherer A, Weber U, Neuenschwander R, Tamborrini G, Exer P, Bernhard J, Villiger PM, Kissling R, Michel BA, Stekhoven D. Age at symptom onset in ankylosing spondylitis: is there a gender difference? *Ann Rheum Dis* 2014; 73: 1908-1910 [PMID: 25104774 DOI: 10.1136/annrheumdis-2014-205613]
- 23 Robinson PC, Brown MA. The genetics of ankylosing spondylitis and axial spondyloarthritis. *Rheum Dis Clin North Am* 2012; 38: 539-553 [PMID: 23083754 DOI: 10.1016/ j.rdc.2012.08.018]
- 24 Tsui HW, Inman RD, Paterson AD, Reveille JD, Tsui FW. ANKH variants associated with ankylosing spondylitis: gender differences. *Arthritis Res Ther* 2005; 7: R513-R525 [PMID: 15899038 DOI: 10.1186/ar1701]
- 25 **Timms AE**, Zhang Y, Bradbury L, Wordsworth BP, Brown MA. Investigation of the role of ANKH in ankylosing spon-

dylitis. Arthritis Rheum 2003; **48**: 2898-2902 [PMID: 14558096 DOI: 10.1002/art.11258]

- 26 Pimentel-Santos FM, Ligeiro D, Matos M, Mourão AF, Vieira de Sousa E, Pinto P, Ribeiro A, Santos H, Barcelos A, Godinho F, Cruz M, Fonseca JE, Guedes-Pinto H, Trindade H, Brown MA, Branco JC. ANKH and susceptibility to and severity of ankylosing spondylitis. J Rheumatol 2012; 39: 131-134 [PMID: 22089454 DOI: 10.3899/jrheum.110681]
- 27 Hersh AH, Stecher RM, Solomon WM, Wolpaw R, Hauser H. Heredity in ankylosing spondylitis; a study of fifty families. *Am J Hum Genet* 1950; 2: 391-408 [PMID: 14837909]
- 28 Yu D, Kuipers JG. Role of bacteria and HLA-B27 in the pathogenesis of reactive arthritis. *Rheum Dis Clin North Am* 2003; 29: 21-36, v-vi [PMID: 12635498]
- 29 Bas S, Scieux C, Vischer TL. Male sex predominance in Chlamydia trachomatis sexually acquired reactive arthritis: are women more protected by anti-chlamydia antibodies? *Ann Rheum Dis* 2001; 60: 605-611 [PMID: 11350850]
- 30 Carter JD, Gérard HC, Espinoza LR, Ricca LR, Valeriano J, Snelgrove J, Oszust C, Vasey FB, Hudson AP. Chlamydiae as etiologic agents in chronic undifferentiated spondylarthritis. *Arthritis Rheum* 2009; 60: 1311-1316 [PMID: 19404948 DOI: 10.1002/art.24431]
- 31 Averns HL, Oxtoby J, Taylor HG, Jones PW, Dziedzic K, Dawes PT. Smoking and outcome in ankylosing spondylitis. *Scand J Rheumatol* 1996; 25: 138-142 [PMID: 8668955]
- 32 Ward MM. Predictors of the progression of functional disability in patients with ankylosing spondylitis. *J Rheumatol* 2002; **29**: 1420-1425 [PMID: 12136900]
- 33 Ward MM, Weisman MH, Davis JC, Reveille JD. Risk factors for functional limitations in patients with long-standing ankylosing spondylitis. *Arthritis Rheum* 2005; 53: 710-717 [PMID: 16208654 DOI: 10.1002/art.21444]
- 34 Doran MF, Brophy S, MacKay K, Taylor G, Calin A. Predictors of longterm outcome in ankylosing spondylitis. J Rheumatol 2003; 30: 316-320 [PMID: 12563688]
- 35 Kaan U, Ferda O. Evaluation of clinical activity and functional impairment in smokers with ankylosing spondylitis. *Rheumatol Int* 2005; 25: 357-360 [PMID: 14991231 DOI: 10.1007/s00296-004-0451-3]
- 36 Ward MM, Hendrey MR, Malley JD, Learch TJ, Davis JC, Reveille JD, Weisman MH. Clinical and immunogenetic prognostic factors for radiographic severity in ankylosing spondylitis. *Arthritis Rheum* 2009; 61: 859-866 [PMID: 19565552 DOI: 10.1002/art.24585]
- 37 Bodur H, Ataman S, Rezvani A, Buğdaycı DS, Cevik R, Birtane M, Akıncı A, Altay Z, Günaydın R, Yener M, Koçyiğit H, Duruöz T, Yazgan P, Cakar E, Aydın G, Hepgüler S, Altan L, Kırnap M, Olmez N, Soydemir R, Kozanoğlu E, Bal A, Sivrioğlu K, Karkucak M, Günendi Z. Quality of life and related variables in patients with ankylosing spondylitis. *Qual Life Res* 2011; 20: 543-549 [PMID: 20978859 DOI: 10.1007/s11136-010-9771-9]
- 38 Mattey DL, Dawson SR, Healey EL, Packham JC. Relationship between smoking and patient-reported measures of disease outcome in ankylosing spondylitis. *J Rheumatol* 2011; 38: 2608-2615 [PMID: 21965641]
- 39 Poddubnyy D, Haibel H, Listing J, Märker-Hermann E, Zeidler H, Braun J, Sieper J, Rudwaleit M. Baseline radiographic damage, elevated acute-phase reactant levels, and cigarette smoking status predict spinal radiographic progression in early axial spondylarthritis. *Arthritis Rheum* 2012; 64: 1388-1398 [PMID: 22127957 DOI: 10.1002/art.33465]
- 40 **Poddubnyy D**, Haibel H, Listing J, Märker-Hermann E, Zeidler H, Braun J, Sieper J, Rudwaleit M. Cigarette smoking has a dose-dependent impact on progression of structural damage in the spine in patients with axial spondyloarthritis: results from the GErman SPondyloarthritis Inception Cohort (GESPIC). *Ann Rheum Dis* 2013; **72**: 1430-1432 [PMID: 23625981 DOI: 10.1136/annrheumdis-2012-203148]

- 41 **Ramiro S**, van Tubergen AM, Landewé R, Stolwijk C, Dougados M, Van den Bosch F, van der Heijde D. Disease activity in male smokers has a >10-fold amplified effect on radiographic damage in comparison with female non-smokers in ankylosing spondylitis. *Arthritis Rheum* 2013; **65**: S640
- 42 Gooren LJ, Giltay EJ, van Schaardenburg D, Dijkmans BA. Gonadal and adrenal sex steroids in ankylosing spondylitis. *Rheum Dis Clin North Am* 2000; 26: 969-987 [PMID: 11084954]
- 43 Mahendira D, Thavaneswaran A, Carty A, Haroon N, Anton A, Passalent L, Alnaqbi KA, Savage L, Aslanyan E, Inman RD. Analysis of the effect of the oral contraceptive pill on clinical outcomes in women with ankylosing spondylitis. *J Rheumatol* 2014; **41**: 1344-1348 [PMID: 24931958 DOI: 10.3899/jrheum.130996]
- 44 Calin A, Brophy S, Blake D. Impact of sex on inheritance of ankylosing spondylitis: a cohort study. *Lancet* 1999; 354: 1687-1690 [PMID: 10568571]
- 45 TYSON TL, THOMPSON WA, RAGAN C. Marie-strümpell spondylitis in women. Ann Rheum Dis 1953; 12: 40-42 [PMID: 13031450]
- 46 HART FD, ROBINSON KC. Ankylosing spondylitis in women. Ann Rheum Dis 1959; 18: 15-23 [PMID: 13650453]
- 47 **McBryde AM**, McCollum DE. Ankylosing spondylitis in women. The disease and its prognosis. *N C Med J* 1973; **34**: 34-37 [PMID: 4509203]
- 48 Resnick D, Dwosh IL, Goergen TG, Shapiro RF, Utsinger PD, Wiesner KB, Bryan BL. Clinical and radiographic abnormalities in ankylosing spondylitis: a comparison of men and women. *Radiology* 1976; **119**: 293-297 [PMID: 1265258 DOI: 10.1148/119.2.293]
- 49 Braunstein EM, Martel W, Moidel R. Ankylosing spondylitis in men and women: a clinical and radiographic comparison. *Radiology* 1982; 144: 91-94 [PMID: 7089271 DOI: 10.1148/radiology.144.1.7089271]
- 50 Marks SH, Barnett M, Calin A. Ankylosing spondylitis in women and men: a case-control study. J Rheumatol 1983; 10: 624-628 [PMID: 6620264]
- 51 **Gran JT**, Ostensen M, Husby G. A clinical comparison between males and females with ankylosing spondylitis. *J Rheumatol* 1985; **12**: 126-129 [PMID: 3872366]
- 52 Will R, Edmunds L, Elswood J, Calin A. Is there sexual inequality in ankylosing spondylitis? A study of 498 women and 1202 men. *J Rheumatol* 1990; **17**: 1649-1652 [PMID: 2084239]
- 53 Kidd B, Mullee M, Frank A, Cawley M. Disease expression of ankylosing spondylitis in males and females. *J Rheumatol* 1988; 15: 1407-1409 [PMID: 3199401]
- 54 Jiménez-Balderas FJ, Mintz G. Ankylosing spondylitis: clinical course in women and men. J Rheumatol 1993; 20: 2069-2072 [PMID: 7516975]
- 55 Eustace S, Coughlan RJ, McCarthy C. Ankylosing spondylitis. A comparison of clinical and radiographic features in men and women. *Ir Med J* 1993; 86: 120-122 [PMID: 8360039]
- 56 Lee W, Reveille JD, Davis JC, Learch TJ, Ward MM, Weisman MH. Are there gender differences in severity of ankylosing spondylitis? Results from the PSOAS cohort. Ann Rheum Dis 2007; 66: 633-638 [PMID: 17127685]
- 57 Garrett S, Jenkinson T, Kennedy LG, Whitelock H, Gaisford P, Calin A. A new approach to defining disease status in ankylosing spondylitis: the Bath Ankylosing Spondylitis Disease Activity Index. J Rheumatol 1994; 21: 2286-2291 [PMID: 7699630]
- 58 Brophy S, Calin A. Ankylosing spondylitis: interaction between genes, joints, age at onset, and disease expression. J *Rheumatol* 2001; 28: 2283-2288 [PMID: 11669170]
- 59 Roussou E, Sultana S. Spondyloarthritis in women: differences in disease onset, clinical presentation, and Bath Ankylosing Spondylitis Disease Activity and Functional indices (BASDAI and BASFI) between men and women with spondyloarthritides. *Clin Rheumatol* 2011; **30**: 121-127 [PMID: 20882310 DOI: 10.1007/s10067-010-1581-5]



- 60 Slobodin G, Reyhan I, Avshovich N, Balbir-Gurman A, Boulman N, Elias M, Feld J, Mader R, Markovitz D, Rimar D, Rosner I, Rozenbaum M, Zisman D, Odeh M. Recently diagnosed axial spondyloarthritis: gender differences and factors related to delay in diagnosis. *Clin Rheumatol* 2011; 30: 1075-1080 [PMID: 21360100 DOI: 10.1007/s10067-011-1719-0]
- 61 Ibn Yacoub Y, Amine B, Laatiris A, Hajjaj-Hassouni N. Gender and disease features in Moroccan patients with ankylosing spondylitis. *Clin Rheumatol* 2012; **31**: 293-297 [PMID: 21796348 DOI: 10.1007/s10067-011-1819-x]
- 62 de Carvalho HM, Bortoluzzo AB, Gonçalves CR, da Silva JA, Ximenes AC, Bértolo MB, Ribeiro SL, Keiserman M, Menin R, Skare TL, Carneiro S, Azevedo VF, Vieira WP, Albuquerque EN, Bianchi WA, Bonfiglioli R, Campanholo C, Costa IP, Duarte AP, Gavi MB, Kohem CL, Leite NH, Lima SA, Meirelles ES, Pereira IA, Pinheiro MM, Polito E, Resende GG, Rocha FA, Santiago MB, Sauma Mde F, Sampaio-Barros PD. Gender characterization in a large series of Brazilian patients with spondyloarthritis. *Clin Rheumatol* 2012; **31**: 687-695 [PMID: 22203094 DOI: 10.1007/s10067-011-1890-3]
- 63 Roussou E, Sultana S. Early spondyloarthritis in multiracial society: differences between gender, race, and disease subgroups with regard to first symptom at presentation, main problem that the disease is causing to patients, and employment status. *Rheumatol Int* 2012; **32**: 1597-1604 [PMID: 21328058 DOI: 10.1007/s00296-010-1680-2]
- 64 Ortega Castro R, Font Ugalde P, Castro Villegas MC, Calvo Gutiérrez J, Muñoz Gomariz E, Zarco Montejo P, Almodóvar R, Mulero Mendoza J, Torre-Alonso JC, Gratacós Masmitjá J, Juanola Roura X, Ariza Ariza R, Fernández Dapica P, Linares Ferrando LF, Brito Brito ME, Cuende Quintana E, Vázquez Galeano C, Moreno Ramos MJ, Giménez Úbeda E, Rodríguez Lozano JC, Fernández Prada M, Queiro Silva R, Moreno Ruzafa E, Júdez Navarro E, Más AJ, Medrano Le Quement C, Ornilla E, Montilla Morales C, Pujol Busquets M, Clavaguera Poch T, Fernández-Espartero MC, Carmona Ortell L, Collantes Estévez E. Different clinical expression of patients with ankylosing spondylitis according to gender in relation to time since onset of disease. Data from REG-ISPONSER. *Reumatol Clin* 2013; **9**: 221-225 [PMID: 23474378 DOI: 10.1016/j.reuma.2012.09.008]
- 65 Dagfinrud H, Vollestad NK, Loge JH, Kvien TK, Mengshoel AM. Fatigue in patients with ankylosing spondylitis: A comparison with the general population and associations with clinical and self-reported measures. *Arthritis Rheum* 2005; 53: 5-11 [PMID: 15696569 DOI: 10.1002/art.20910]
- 66 Bianchi WA, Elias FR, Carneiro S, Bortoluzzo AB, Goncalves CR, da Silva JA, Ximenes AC, Bértolo MB, Ribeiro SL, Keiserman M, Skare TL, Menin R, Azevedo VF, Vieira WP, Albuquerque EN, Bonfiglioli R, Campanholo C, Carvalho HM, Costa IP, Duarte AP, Kohem CL, Leite NH, Lima SA, Meirelles ES, Pereira IA,Pinheiro MM, Polito E, Resende GG, Rocha FA, Santiago MB, Sauma MD, Valim V, Sampaio-Barros PD. Assessment of fatigue in a large series of 1492 Brazilian patients with Spondyloarthritis. *Mod Rheumatol* 2014 Jun 2; Epub ahead of print [PMID: 24884480 DOI: 10.31 09/14397595.2014.906049]
- 67 Almodóvar R, Carmona L, Zarco P, Collantes E, González C, Mulero J, Sueiro JL, Gratacós J, Torre-Alonso JC, Juanola X, Batlle E, Ariza R, Font P. Fibromyalgia in patients with ankylosing spondylitis: prevalence and utility of the measures of activity, function and radiological damage. *Clin Exp Rheumatol* 2010; 28: S33-S39 [PMID: 21176420]
- 68 Salaffi F, De Angelis R, Carotti M, Gutierrez M, Sarzi-Puttini P, Atzeni F. Fibromyalgia in patients with axial spondyloarthritis: epidemiological profile and effect on measures of disease activity. *Rheumatol Int* 2014; 34: 1103-1110 [PMID: 24509896 DOI: 10.1007/s00296-014-2955-9]
- 69 Haliloglu S, Carlioglu A, Akdeniz D, Karaaslan Y, Kosar A. Fibromyalgia in patients with other rheumatic diseases:

prevalence and relationship with disease activity. *Rheuma-tol Int* 2014; **34**: 1275-1280 [PMID: 24589726 DOI: 10.1007/ s00296-014-2972-8]

- 70 van der Heijde D, Lie E, Kvien TK, Sieper J, Van den Bosch F, Listing J, Braun J, Landewé R. ASDAS, a highly discriminatory ASAS-endorsed disease activity score in patients with ankylosing spondylitis. *Ann Rheum Dis* 2009; 68: 1811-1818 [PMID: 19060001 DOI: 10.1136/ard.2008.100826]
- 71 Geirsson AJ, Eyjolfsdottir H, Bjornsdottir G, Kristjansson K, Gudbjornsson B. Prevalence and clinical characteristics of ankylosing spondylitis in Iceland - a nationwide study. *Clin Exp Rheumatol* 2010; 28: 333-340 [PMID: 20406616]
- 72 Kim TJ, Lee S, Joo KB, Park DJ, Park YW, Lee SS, Kim TH. The presence of peripheral arthritis delays spinal radiographic progression in ankylosing spondylitis: Observation Study of the Korean Spondyloarthropathy Registry. *Rheumatology* (Oxford) 2014; **53**: 1404-1408 [PMID: 24609061 DOI: 10.1093/rheumatology/keu014]
- 73 Carneiro S, Bortoluzzo A, Gonçalves C, Silva JA, Ximenes AC, Bértolo M, Ribeiro SL, Keiserman M, Skare T, Menin R, Azevedo V, Vieira W, Albuquerque E, Bianchi W, Bonfiglioli R, Campanholo C, Carvalho HM, Costa Id, Duarte A, Kohem C, Leite N, Lima SA, Meirelles ES, Pereira IA, Pinheiro MM, Polito E, Resende GG, Rocha FA, Santiago MB, Sauma Mde F, Valim V, Sampaio-Barros PD. Effect of enthesitis on 1505 Brazilian patients with spondyloarthritis. *J Rheumatol* 2013; 40: 1719-1725 [PMID: 23858049 DOI: 10.3899/jrheum.121145]
- 74 Wanders A, Landewé R, Dougados M, Mielants H, van der Linden S, van der Heijde D. Association between radiographic damage of the spine and spinal mobility for individual patients with ankylosing spondylitis: can assessment of spinal mobility be a proxy for radiographic evaluation? *Ann Rheum Dis* 2005; 64: 988-994 [PMID: 15958757 DOI: 10.1136/ ard.2004.029728]
- 75 Machado P, Landewé R, Braun J, Hermann KG, Baker D, van der Heijde D. Both structural damage and inflammation of the spine contribute to impairment of spinal mobility in patients with ankylosing spondylitis. *Ann Rheum Dis* 2010; 69: 1465-1470 [PMID: 20498215]
- 76 Stolwijk C, van Tubergen A, Castillo-Ortiz JD, Boonen A. Prevalence of extra-articular manifestations in patients with ankylosing spondylitis: a systematic review and meta-analysis. *Ann Rheum Dis* 2013 Sep 2; Epub ahead of print [PMID: 23999006 DOI: 10.1136/annrheumdis-2013-203582]
- 77 Spencer DG, Park WM, Dick HM, Papazoglou SN, Buchanan WW. Radiological manifestations in 200 patients with ankylosing spondylitis: correlation with clinical features and HLA B27. J Rheumatol 1979; 6: 305-315 [PMID: 490525]
- 78 Ramiro S, van der Heijde D, van Tubergen A, Stolwijk C, Dougados M, van den Bosch F, Landewé R. Higher disease activity leads to more structural damage in the spine in ankylosing spondylitis: 12-year longitudinal data from the OASIS cohort. *Ann Rheum Dis* 2014; **73**: 1455-1461 [PMID: 24812292 DOI: 10.1136/annrheumdis-2014-205178]
- 79 van Onna M, Jurik AG, van der Heijde D, van Tubergen A, Heuft-Dorenbosch L, Landewé R. HLA-B27 and gender independently determine the likelihood of a positive MRI of the sacroiliac joints in patients with early inflammatory back pain: a 2-year MRI follow-up study. *Ann Rheum Dis* 2011; 70: 1981-1985 [PMID: 21859694 DOI: 10.1136/annrheum-dis-2011-200025]
- 80 Van Praet L, Jans L, Carron P, Jacques P, Glorieus E, Colman R, Cypers H, Mielants H, De Vos M, Cuvelier C, Van den Bosch F, Elewaut D. Degree of bone marrow oedema in sacroiliac joints of patients with axial spondyloarthritis is linked to gut inflammation and male sex: results from the GIANT cohort. *Ann Rheum Dis* 2014; **73**: 1186-1189 [PMID: 24276368 DOI: 10.1136/annrheumdis-2013-203854]
- 81 **Wong ND**, Pio J, Valencia R, Thakal G. Distribution of C-reactive protein and its relation to risk factors and coronary



heart disease risk estimation in the National Health and Nutrition Examination Survey (NHANES) III. *Prev Cardiol* 2001; 4: 109-114 [PMID: 11828186]

- 82 Khera A, McGuire DK, Murphy SA, Stanek HG, Das SR, Vongpatanasin W, Wians FH, Grundy SM, de Lemos JA. Race and gender differences in C-reactive protein levels. J Am Coll Cardiol 2005; 46: 464-469 [PMID: 16053959 DOI: 10.1016/j.jacc.2005.04.051]
- 83 Doran B, Zhu W, Muennig P. Gender differences in cardiovascular mortality by C-reactive protein level in the United States: evidence from the National Health and Nutrition Examination Survey III. Am Heart J 2013; 166: 45-51 [PMID: 23816020 DOI: 10.1016/j.ahj.2013.03.017]
- 84 Kristensen LE, Karlsson JA, Englund M, Petersson IF, Saxne T, Geborek P. Presence of peripheral arthritis and male sex predicting continuation of anti-tumor necrosis factor therapy in ankylosing spondylitis: an observational prospective cohort study from the South Swedish Arthritis Treatment Group Register. *Arthritis Care Res* (Hoboken) 2010; 62: 1362-1369 [PMID: 20506310 DOI: 10.1002/acr.20258]
- 85 Dagfinrud H, Kjeken I, Mowinckel P, Hagen KB, Kvien TK. Impact of functional impairment in ankylosing spondylitis: impairment, activity limitation, and participation restrictions. J Rheumatol 2005; 32: 516-523 [PMID: 15742446]
- 86 Yanbaeva DG, Dentener MA, Creutzberg EC, Wesseling G, Wouters EF. Systemic effects of smoking. *Chest* 2007; 131: 1557-1566 [PMID: 17494805]
- 87 Bethi S, Dasgupta A, Weisman MH, Learch TJ, Gensler LS, Davis JC, Reveille JD, Ward MM. Functional limitations due to axial and peripheral joint impairments in patients with ankylosing spondylitis: are focused measures more informative? *Arthritis Care Res* (Hoboken) 2013; 65: 607-614 [PMID: 23097327 DOI: 10.1002/acr.21878]
- 88 Dagfinrud H, Mengshoel AM, Hagen KB, Loge JH, Kvien TK. Health status of patients with ankylosing spondylitis: a comparison with the general population. *Ann Rheum Dis* 2004; 63: 1605-1610 [PMID: 15547084 DOI: 10.1136/ard.2003.019224]
- 89 Ward MM. Health-related quality of life in ankylosing spondylitis: a survey of 175 patients. *Arthritis Care Res* 1999; 12: 247-255 [PMID: 10689989]
- 90 Yılmaz O, Tutoğlu A, Garip Y, Ozcan E, Bodur H. Healthrelated quality of life in Turkish patients with ankylosing spondylitis: impact of peripheral involvement on quality of life in terms of disease activity, functional status, severity of pain, and social and emotional functioning. *Rheumatol Int* 2013; 33: 1159-1163 [PMID: 22955799 DOI: 10.1007/ s00296-012-2510-5]
- 91 Boonen A, de Vet H, van der Heijde D, van der Linden S. Work status and its determinants among patients with ankylosing spondylitis. A systematic literature review. J Rheumatol 2001; 28: 1056-1062 [PMID: 11361189]
- 92 **Boonen A**, Chorus A, Miedema H, van der Heijde D, van der Tempel H, van der Linden S. Employment, work disability, and work days lost in patients with ankylosing spondylitis: a cross sectional study of Dutch patients. *Ann Rheum Dis* 2001; **60**: 353-358 [PMID: 11247865]
- 93 Boonen A, Chorus A, Miedema H, van der Heijde D, Landewé R, Schouten H, van der Tempel H, van der Linden S. Withdrawal from labour force due to work disability in patients with ankylosing spondylitis. *Ann Rheum Dis* 2001; 60: 1033-1039 [PMID: 11602474]
- 94 Ward MM, Kuzis S. Risk factors for work disability in patients with ankylosing spondylitis. *J Rheumatol* 2001; 28: 315-321 [PMID: 11246669]
- 95 **Barlow JH**, Wright CC, Williams B, Keat A. Work disability among people with ankylosing spondylitis. *Arthritis Rheum* 2001; **45**: 424-429 [PMID: 11642641]
- 96 **Boonen A**, van der Heijde D, Landewé R, Spoorenberg A, Schouten H, Rutten-van Mölken M, Guillemin F, Dougados

M, Mielants H, de Vlam K, van der Tempel H, van der Linden S. Work status and productivity costs due to ankylosing spondylitis: comparison of three European countries. *Ann Rheum Dis* 2002; **61**: 429-437 [PMID: 11959767]

- 97 Boonen A, Chorus A, Landewé R, van der Heijde D, Miedema H, van der Tempel H, van der Linden S. Manual jobs increase the risk of patients with ankylosing spondylitis withdrawing from the labour force, also when adjusted for job related withdrawal in the general population. *Ann Rheum Dis* 2002; **61**: 658 [PMID: 12079917]
- 98 Mau W, Listing J, Huscher D, Zeidler H, Zink A. Employment across chronic inflammatory rheumatic diseases and comparison with the general population. *J Rheumatol* 2005; 32: 721-728 [PMID: 15801031]
- 99 Ariza-Ariza R, Hernández-Cruz B, Collantes E, Batlle E, Fernández-Sueiro JL, Gratacós J, Juanola X, Linares LF, Mulero J, Zarco P. Work disability in patients with ankylosing spondylitis. *J Rheumatol* 2009; **36**: 2512-2516 [PMID: 19833749 DOI: 10.3899/jrheum.090481]
- 100 Bakland G, Gran JT, Becker-Merok A, Nordvåg BY, Nossent JC. Work disability in patients with ankylosing spondylitis in Norway. J Rheumatol 2011; 38: 479-484 [PMID: 21285159 DOI: 10.3899/jrheum.100686]
- 101 Gordeev VS, Maksymowych WP, Schachna L, Boonen A. Understanding presenteeism in patients with ankylosing spondylitis: contributing factors and association with sick leave. *Arthritis Care Res* (Hoboken) 2014; 66: 916-924 [PMID: 24339444 DOI: 10.1002/acr.22253]
- 102 Ward MM, Reveille JD, Learch TJ, Davis JC, Weisman MH. Impact of ankylosing spondylitis on work and family life: comparisons with the US population. *Arthritis Rheum* 2008; 59: 497-503 [PMID: 18383414 DOI: 10.1002/art.23523]
- 103 Yim SY, Lee IY, Lee JH, Jun JB, Kim TH, Bae SC, Yoo DH. Quality of marital life in Korean patients with spondyloarthropathy. *Clin Rheumatol* 2003; 22: 208-212 [PMID: 14505212 DOI: 10.1007/s10067-003-0700-y]
- 104 Feldtkeller E, Lemmel EM. Quality of marital life in patients with spondyloarthropathy. *Clin Rheumatol* 2004; 23: 277-278 [PMID: 15168165 DOI: 10.1007/s10067-004-0872-0]
- 105 Miceli-Richard C, Said-Nahal R, Breban M. Impact of sex on inheritance of ankylosing spondylitis. *Lancet* 2000; 355: 1097-1098; author reply 1098 [PMID: 10744112 DOI: 10.1016/ S0140-6736(05)72217-0]
- 106 van der Heijde D, Sieper J, Maksymowych WP, Dougados M, Burgos-Vargas R, Landewé R, Rudwaleit M, Braun J. 2010 Update of the international ASAS recommendations for the use of anti-TNF agents in patients with axial spondyloarthritis. *Ann Rheum Dis* 2011; **70**: 905-908 [PMID: 21540200]
- 107 Rudwaleit M, Listing J, Brandt J, Braun J, Sieper J. Prediction of a major clinical response (BASDAI 50) to tumour necrosis factor alpha blockers in ankylosing spondylitis. *Ann Rheum Dis* 2004; 63: 665-670 [PMID: 15037444 DOI: 10.1136/ ard.2003.016386]
- 108 Davis JC, Van der Heijde DM, Dougados M, Braun J, Cush JJ, Clegg DO, Inman RD, de Vries T, Tsuji WH. Baseline factors that influence ASAS 20 response in patients with ankylosing spondylitis treated with etanercept. J Rheumatol 2005; 32: 1751-1754 [PMID: 16142873]
- 109 Rudwaleit M, Claudepierre P, Wordsworth P, Cortina EL, Sieper J, Kron M, Carcereri-De-Prati R, Kupper H, Kary S. Effectiveness, safety, and predictors of good clinical response in 1250 patients treated with adalimumab for active ankylosing spondylitis. *J Rheumatol* 2009; **36**: 801-808 [PMID: 19273449 DOI: 10.3899/jrheum.081048]
- 110 Lord PA, Farragher TM, Lunt M, Watson KD, Symmons DP, Hyrich KL. Predictors of response to anti-TNF therapy in ankylosing spondylitis: results from the British Society for Rheumatology Biologics Register. *Rheumatology* (Oxford) 2010; 49: 563-570 [PMID: 20032223 DOI: 10.1093/rheumatology/kep422]



- 111 Arends S, Brouwer E, van der Veer E, Groen H, Leijsma MK, Houtman PM, Th A Jansen TL, Kallenberg CG, Spoorenberg A. Baseline predictors of response and discontinuation of tumor necrosis factor-alpha blocking therapy in ankylosing spondylitis: a prospective longitudinal observational cohort study. *Arthritis Res Ther* 2011; 13: R94 [PMID: 21689401]
- 112 Vastesaeger N, van der Heijde D, Inman RD, Wang Y, Deodhar A, Hsu B, Rahman MU, Dijkmans B, Geusens P, Vander Cruyssen B, Collantes E, Sieper J, Braun J. Predicting the outcome of ankylosing spondylitis therapy. *Ann Rheum Dis* 2011; 70: 973-981 [PMID: 21402563 DOI: 10.1136/ard.2010.147744]
- 113 **Glintborg B**, Ostergaard M, Krogh NS, Dreyer L, Kristensen HL, Hetland ML. Predictors of treatment response and drug

continuation in 842 patients with ankylosing spondylitis treated with anti-tumour necrosis factor: results from 8 years' surveillance in the Danish nationwide DANBIO registry. *Ann Rheum Dis* 2010; **69**: 2002-2008 [PMID: 20511613 DOI: 10.1136/ard.2009.124446]

- 114 Bakland G, Gran JT, Nossent JC. Increased mortality in ankylosing spondylitis is related to disease activity. Ann Rheum Dis 2011; 70: 1921-1925 [PMID: 21784726]
- 115 Bakland G, Alsing R, Singh K, Nossent JC. Assessment of SpondyloArthritis International Society criteria for axial spondyloarthritis in chronic back pain patients with a high prevalence of HLA-B27. Arthritis Care Res (Hoboken) 2013; 65: 448-453 [PMID: 22833469 DOI: 10.1002/acr.21804]
- P- Reviewer: Feldtkeller E, Sakellariou GT S- Editor: Gong XM L- Editor: A E- Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.44 World J Rheumatol 2014 November 12; 4(3): 44-53 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

REVIEW

Muscle wasting in rheumatoid arthritis: The role of oxidative stress

Antonios Stavropoulos-Kalinoglou, Charikleia Deli, George D Kitas, Athanasios Z Jamurtas

Antonios Stavropoulos-Kalinoglou, Charikleia Deli, Athanasios Z Jamurtas, Department of Sport and Exercise Science, University of Thessaly, 42100 Trikala, Greece

George D Kitas, Department of Rheumatology, Dudley Group NHS Foundation Trust, Russell's Hall Hospital, Dudley, West Midlands DY1 2HQ, United Kingdom

George D Kitas, Arthritis Research UK Epidemiology Unit, University of Manchester, Manchester M13 9PT, United Kingdom

Athanasios Z Jamurtas, Institute of Human Performance and Rehabilitation, Centre for Research and Technology Thessaly, 42100 Trikala, Greece

Author contributions: Stavropoulos-Kalinoglou A, Deli C, Kitas GD and Jamurtas AZ contributed to this paper.

Supported by The research project is implemented within the framework of the Action «Supporting Postdoctoral Researchers » of the Operational Program "Education and Lifelong Learning" (Action's Beneficiary: General Secretariat for Research and Technology); and is co-financed by the European Social Fund (ESF) and the Greek State

Correspondence to: Antonios Stavropoulos-Kalinoglou, PhD, Department of Sport and Exercise Science, University of Thessaly, Trikala-Karyes Road, 42100 Trikala,

Greece. antonios.stav@gmail.com

Telephone: +30-24-31047038 Fax: +30-24-31047038

Received: April 17, 2014 Revised: September 1, 2014

Accepted: September 23, 2014

Published online: November 12, 2014

Abstract

Rheumatoid arthritis (RA), the commonest inflammatory arthritis, is a debilitating disease leading to functional and social disability. In addition to the joints, RA affects several other tissues of the body including the muscle. RA patients have significantly less muscle mass compared to the general population. Several theories have been proposed to explain this. High grade inflammation, a central component in the pathophysiology of the disease, has long been proposed as the key driver of muscle wasting. More recent findings however, indicate that inflammation on its own cannot fully explain the high prevalence of muscle wasting in RA. Thus, the

contribution of other potential confounders, such as nutrition and physical activity, has also been studied. Results indicate that they play a significant role in muscle wasting in RA, but again neither of these factors seems to be able to fully explain the condition. Oxidative stress is one of the major mechanisms thought to contribute to the development and progression of RA but its potential contribution to muscle wasting in these patients has received limited attention. Oxidative stress has been shown to promote muscle wasting in healthy populations and people with several chronic conditions. Moreover, all of the aforementioned potential contributors to muscle wasting in RA (i.e., inflammation, nutrition, and physical activity) may promote pro- or antioxidative mechanisms. This review aims to highlight the importance of oxidative stress as a driving mechanism for muscle wasting in RA and discusses potential interventions that may promote muscle regeneration via reduction in oxidative stress.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Rheumatoid arthritis; Oxidative stress; Muscle wasting; Inflammation; Cytokines; Exercise

Core tip: Muscle wasting is common in rheumatoid arthritis (RA) and associates with significant health burden. To date several theories have been proposed to explain why RA patients loose muscle mass but the exact underlying mechanisms are not clear. Oxidative stress is a key driver of muscle wasting in the general population; however, its potential role in muscle wasting in RA has not been studied. As it arises from this review, oxidative stress seems to contribute to muscle wasting in RA. Further research on the subject is warranted, especially focusing on the underlying mechanisms and potential interventions.

Stavropoulos-Kalinoglou A, Deli C, Kitas GD, Jamurtas AZ. Muscle wasting in rheumatoid arthritis: The role of oxidative stress. *World J Rheumatol* 2014; 4(3): 44-53 Available from:



URL: http://www.wjgnet.com/2220-3214/full/v4/i3/44.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.44

INTRODUCTION

Rheumatoid arthritis (RA) is the most common inflammatory arthritis, with a prevalence of approximately 1% in Europe and North America^[1]. It is an autoimmune disease affecting mainly synovial joints^[2] and associates with high-grade inflammation characterised by high levels of circulating pro-inflammatory cytokines, including tumour necrosis factor alpha (TNF- α), and the interleukins (IL) 1 and 6. These cytokines are produced in the inflamed synovium and are implicated in joint swelling, pain, and eventually destruction^[3]. As the condition progresses, patients very frequently lose their jobs^[4], functional ability (movement)^[1] and eventually their independence^[5]. Apart from this physical and psychological personal burden, RA has significant costs for the health and social care system^[6]. Thus, the efforts of the scientific community have focused on the identification and elimination of the potential causes of RA as well as effective treatments. Despite significant therapeutic progress in recent years a cure remains elusive^[1].

Apart from the joints, RA affects several other tissues of the body leading to systemic involvement and significant extra-articular manifestations^[7]. It is these extra-articular manifestations, and not RA itself, that significantly shorten the life of RA patients and add extra layers of morbidity. Cardiac and vascular conditions are especially common among these patients. Heart disease in RA is both more prevalent and more likely to lead to death than in the general population^[8]. The exact cause for this remains unknown, however genetic predisposition^[9-12], classical cardiovascular disease risk factors^[13,14] and the effects of systemic inflammation on the vasculature^[15,16] are all thought to contribute. Other extra-articular manifestations are observed in the skin, eyes, lungs, renal, nervous and gastrointestinal systems^[17].

The reasons for the development of such manifestations are not fully understood. It is believed that the endocrine functions of the pro-inflammatory cytokines (mainly TNF- α , IL-1 and IL-6) initiate a cascade of destructive processes in distant tissues, with reactive oxygen species playing a central role in cell membrane destruction and cellular death^[18].

OXIDATIVE STRESS IN RA

Free radicals and inflammation

Free radicals, such as reactive oxygen species (ROS), have been proposed to play a significant role both in the development and progression of inflammation, as well as its deleterious effects on cell structure and function at the site of the inflammation and in distant tissues^[19,20]. Free radicals, formed as by-products of normal biological processes - such as cellular metabolism in the mitochondrial

electron transport chain and reperfusion injury - are highly reactive agents that can cause physiological damage^[21]. Free radicals can damage all cellular components such as lipids, proteins and DNA. In the general population, they are counterbalanced by effective antioxidant defence mechanisms. However, in inflammatory conditions these defence mechanisms seem to be weakened^[22]. It is not clear which is the sequence of events but it seems likely that inflammation reduces the anti-oxidant response, thereby increasing the accumulation of free radicals^[19]. These further activate pro-inflammatory nuclear pathways (specifically Activator Protein one - AP-1 and nuclear factor kappa β - NF κ B) that transcribe cytokines and adhesion molecules involved in the modulation of inflammation^[23] resulting in further production of free radicals. Nitric oxide (NO) has a role in the regulation of vascular tone, superoxide free radical (O2 -) in fibroblast proliferation and hydrogen peroxide (H2O2) in the activation of pro-inflammatory transcription factors. Other control mechanisms which may be perturbed in inflammation include: the oxidative modification of low density lipoprotein, the oxidative inactivation of alpha-1-protease inhibitor, DNA damage, lipid peroxidation and heat shock protein formed with the activation of neutrophil, nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and endothelial cell xanthine dehydrogenase, which associate with oxidative stress and contribute significantly to the inflammatory process^[19,22,23].

Oxidative stress and RA activity

Oxidative stress is frequently reported in patients with RA. Cells present in inflamed joints (*e.g.*, macrophages, neutrophils and lymphocytes), have the ability to produce free radicals^[24,25]. These liberate superoxide radical, hydrogen peroxide, elastase, hypochlorus acid and eicosanoids^[26]. Another source of free radicals in RA is hypoxic reperfusion injury from elevated synovial cavity pressure during joint movement. A fivefold increase in mitochondrial ROS production in whole blood and monocytes of patients with RA compared with healthy subjects suggests that oxidative stress is a significant factor in RA^[26].

Free radicals are implicated in joint damage both indirectly (via increasing inflammation as described above) and directly. They can degrade joint cartilage, by attacking proteoglycans (integral components of structural tissues) and inhibiting proteoglycan synthesis^[27]. Indeed in patients with RA, serum and synovial fluid contain end products of lipid peroxidation which correlate with disease severity and activity^[28]. In parallel, anti-oxidant capacity in patients with RA seems to be significantly reduced^[29]. This low antioxidant status has been associated with low levels of tocopherols, beta-carotene and retinols and low activities of glutathione reductase and superoxide dismutase^[26]. RA has also been linked to low levels of reduced glutathione- an intracellular antioxidant- in synovial fluid T cells^[30]. Reduced glutathione, is among the most prominent defences against reactive oxygen species. It is a substrate for glutathione peroxidases, several trans-



ferases and several other enzymes and acts as a general radical quencher in cells by removing superoxide anions and hydrogen peroxide^[26]. Serum concentrations of antioxidative vitamins A, C and E are also significantly reduced in RA^[19,23,31].

MUSCLE WASTING IN RA

A significant but little studied extra-articular manifestation of RA is rheumatoid cachexia (RC). RC is characterised by a high rate of muscle mass and strength loss, typically with preservation or slight increase in fat mass^[32]. RC differs from other forms of cachexia such as those observed in cancer, chronic heart failure, kidney disease or chronic infection as it is rarely accompanied by a net weight loss^[33]. RC also differs from sarcopenia (age-related reduction in muscle mass observed in the elderly) as it occurs at a younger age and muscle mass loss progresses at a substantially higher rate^[34]. The prevalence of RA in the United Kingdom is 0.8%^[5]. The exact prevalence of RC is not known today as there is no consensus on its definition and assessment. However, at least 10%-20% of patients with controlled $RA^{[35,36]}$ and > 40% of patients with active RA^[37] suffer from muscle wasting. This makes it one of the most common complications of RA.

RC has been shown to associate with poorer disease outcomes including reduced quality of life, more fatigue and increased overall morbidity and mortality^[34,38,39], although the independent nature and directionality of many of these associations remain uncertain and require further study. Low muscle mass also associates with dysmetabolic states such as insulin resistance and type II diabetes^[40,41] and thus may be partly responsible for the increased cardiovascular risk observed in RA^[33,35,42]. Inflammation associated with the disease is consistently identified as the potential cause of these manifestations. Indeed, inflammatory cytokines produced at the site of the disease (*i.e.*, the joints) have endocrine functions and act on distant tissues such as the muscle^[43].

Mechanisms of muscle wasting in RA

Inflammation: High plasma concentrations of the inflammatory cytokines implicated in RA pathophysiology (TNF- α , IL-1 and IL-6) are thought to trigger muscle wasting^[34,44]. TNF- α -induced activation of the classical $NF_{\kappa}B$ pathway is now generally accepted to lead to inhibition of skeletal muscle differentiation and regeneration in a variety of muscle diseases, although this has not been confirmed yet as a mechanism of muscle wasting in RA patients. IL-1 among other cytokines has been shown to prevent the anabolic effect of insulin growth factor 1 (IGF-1) on myoblast differentiation, muscle protein synthesis, and myogenin expression^[44,45], while intravenous infusion of IL-6 in healthy volunteers led to net muscle protein degradation in healthy individuals^[46]. In RA patients, short term (3-6 mo) anti-TNF-a medication led to significant reduction of disease activity but did not improve body composition and had no effect of muscle mass^[47,48] suggesting that cytokines might not be the most significant contributors to muscle wasting in RA.

Physical inactivity: Physical inactivity is the strongest predictor of fat mass in RA^[49], while resistance exercise interventions may result in increased muscle mass and strength and partial reversal of muscle wasting in patients with RA^[50,51]. Therefore muscle wasting in RA seems to be a consequence of a negative spiral between the metabolic and functional consequences of inflammation which enhance muscle catabolism and the premature adoption of an increasingly sedentary lifestyle in which the anabolic stimulus of regular exercise is missing^[49] with consequent increase in fat mass. In line with this hypothesis is the observation that obesity is a common feature of RA and adds to the high risk for the metabolic syndrome and cardiovascular disease^[34,52,53].

Adiposity: It is reasonable to assume that there are parallels between the mechanisms that lead to sarcopenia in healthy sedentary elderly individuals and the mechanisms that lead to muscle mass loss in RA patients. It is also reasonable to assume that there are parallels between the impact of obesity on the rate of sarcopenia and the potential role that obesity plays in the mechanisms that lead to muscle wasting in RA. An inherent consequence of the adoption of a sedentary life-style, without a reduction in energy intake, is a gradual increase of the subcutaneous and visceral adipose tissue mass^[54]. Adipose tissue (especially visceral) is a well-known source of inflammation. In addition to adipocytes, pre-adipocytes and fibroblasts, up to 50% of the cell mass in adipose tissue of obese individuals are inflammatory cells such as monocytes and macrophages^[55,56]. Adipocytes and macrophages both are a source of inflammatory cytokines^[55,56]. In addition, the large adipose tissue stores in obese individuals are a constant source of lipolysis and lead to high circulatory concentrations of fatty acids and triglycerides. High plasma concentrations of inflammatory cytokines, FA and triglycerides contribute to the insulin resistance of skeletal muscle and its microvasculature^[54-56] via mechanisms outlined below.

Insulin resistance: The most striking change in skeletal muscle through a sedentary lifestyle is a reduction of the mitochondrial density^[54] and, therefore, of oxidative capacity of blood-borne fatty acids (NEFA's). Sedentary muscles also have a reduced capacity to oxidize the lipid droplets that are stored in the muscle in the vicinity of the mitochondria^[54,57,58]. This, combined with an increased supply of plasma fatty acids and triglycerides, leads to the accumulation of fatty acid metabolites (long-chain fatty acyl-coenzyme A, diacylglycerols and ceramides). Both these fatty acid metabolites and the exposure of the muscle to inflammatory cytokines activate serine kinases that lead to serine phosphorylation of insulin receptor substrate 1 (IRS-1) and prevent downstream activation of the insulin signalling cascade and, therefore, impair

WJR www.wjgnet.com

glucose transporter type 4 translocation and glucose uptake^[58]. Insulin resistance (IR) also leads to an imbalance between protein synthesis and degradation^[59] and is a major cause of the muscle mass loss in sedentary obese elderly individuals.

Endothelial dysfunction: The overload of the muscle with fatty acids, triglycerides and inflammatory cytokines also leads to major impairments in its associated vasculature. The endothelial cells that cover the luminal wall of feeding arteries, resistance arteries, and terminal arterioles (which control blood supply to the capillaries) normally dilate if they are exposed to meal-induced increases in insulin concentration^[54,60]. Insulin in endothelial cells activates the enzyme eNOS [endothelial nitric oxide (NO) synthase] and the resultant NO leads to dilation of the smooth muscle layer in arteries and arterioles. This mechanism ensures that in the period after meal ingestion maximal amounts of glucose, amino acids and insulin are channelled to the muscle to maximize glucose uptake, increase protein synthesis and reduce protein degradation^[54,60]. Vascular overload with lipids and inflammatory cytokines also leads to endothelial IR and reduces the supply of blood and nutrients to muscles of obese individuals^[54,60]

POTENTIAL ROLE OF OXIDATIVE STRESS IN MUSCLE WASTING IN RA

To our knowledge, there is no study today investigating the associations of oxidative stress with muscle wasting in RA. However, there are numerous reports in the general population and several other conditions showing that oxidative stress may be a very important underlying mechanism that drives muscle wasting.

Endothelial function and oxidative stress

Experiments in obese Zucker rats and incubated endothelial cells have shown that high concentrations of longchain fatty acylCoA and diacylglycerol activate protein kinase C (PKC)- β in aortic endothelium^[61] and prevent insulin-induced activation of IRS-1, Akt, eNOS phosphorylation and increases in NO production. A high lipid and cytokine load (*via* PKC-activation) also leads to induction of NADPH oxidase in the vasculature of patients with the metabolic syndrome, hypertension or cardiovascular disease^[54]. High NADPH oxidase activity will lead to excess production of superoxide anions (O₂-) which will scavenge NO thereby reducing basal and insulin-induced NO-production. Superoxide anions react with NO resulting in the formation of peroxynitrite and reducing the amount of NO available for vasodilation^[62].

Muscle disuse and oxidative stress

Physical inactivity, in a population with constantly high grade inflammation, such as those with RA, may lead to significant intramuscular accumulation of ROS, as is the case for muscle disuse (*e.g.*, due to limb immobilization

or bed rest) in the general population^[63]. Muscle atrophy from disuse is mainly attributed to oxidative stress, *i.e.*, reduces anti-oxidant capacity and increased ROS production^[64,65]. Mitochondria are the site for excessive ROS production^[65,66]; and ROS production, such as H₂O₂, is increased by up to 100% following 14 d of limb immobilization^[67]. Moreover, xanthine oxidase and NADPH oxidase contribute but to a lesser degree to ROS production in muscle disuse^[68,69]. Similarly, lipid peroxidation has been shown to associate with muscle atrophy^[70].

These affect the balance between protein synthesis and degradation^[71,72]. Specifically, disturbed redox balance due to intramuscular ROS accumulation, such as that of $H_2O_2^{[73,74]}$, may activate transcriptional factors that increase expression of apoptotic pathways, such as the NF- $\kappa\beta$ pathway and Foxo leading to severe protein degradation^[68,75]. Moreover, oxidative stress may activate calpain and caspase-3, further increasing proteolytic processes^[68,76]. Oxidation of muscle proteins themselves makes them susceptible to proteolytic damage^[77].

ROS accumulation may also inhibit signalling pathways controlling protein synthesis^[78,79]. It seems that ROS inhibit mRNA translation at an early stage; this reduces the ability of senescent satellite cells to become active and infiltrate the muscle cell^[79,80]. However, these studies where performed in cell cultures, and it is not clear if these processes also occur *in vivo*.

Aging and oxidative stress

Muscle wasting is commonly observed in the elderly, affecting their quality of life and independence^[81]. Oxidative stress has long been associated with aging related processes^[82]. The elderly exhibit increased concentrations of oxidative by-products compared to younger individuals^[83,84]. In normal aging, ROS participate in a number of processes aiding the transmission of signals within the muscle and affect gene expression^[85,86]. As is the case with disuse atrophy, in aging mitochondria are also the main site for ROS production. Aging mitochondria seem to produce larger amounts of ROS, and especially H2O2, compared to younger ones^[87]. The wasting effect of H₂O₂ seems to be mediated by the presence of the Copper and Zinc containing superoxide dismutase (Cu,ZnSOD)^[88,89]. Cu,ZnSOD has been shown to decrease with aging^[90]. In animal studies, SOD1 (gene encoding Cu,ZnSOD) knockout mice exhibited a form of rapid muscle wasting with similar characteristics to that of aging, including oxidative stress and weakness^[91].

PREVENTION OF OXIDATIVE STRESS IN RA

Nutritional interventions

The vast majority of studies in this field have focused on the use of exogenous anti-oxidative agents, such as the administration of vitamins (A, C, E) and omega-3 fatty acids. Vitamin E seems to uncouple joint inflammation and joint destruction in the transgenic KRN/NOD

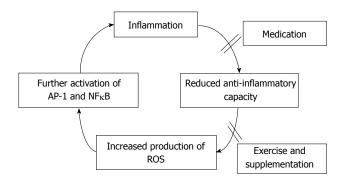


Figure 1 Hypothesis: Inflammation reduces anti-oxidant capacity which increases reactive oxygen species concentration, further activating proinflammatory pathways, entering the body into a vicious circle. Control of inflammation *via* medication might increase anti-inflammatory capacity of the body while exercise and supplementation may lead to increased anti-oxidant capacity both resulting in reduced oxidative stress and preserve muscle mass. ROS: Reactive oxygen species. AP: Activator protein; NF κ B: Nuclear factor kappa β .

mouse model of RA, with a beneficial effect on joint destruction^[92]. Some studies have even attributed therapeutic value to antioxidant supplementation as they reported better control^[93] and improvement of RA-related symptoms^[94]. Dietary interventions have been suggested to improve plasma levels of vitamin C, retinol and uric acid, which inversely correlate with variables related to disease activity^[95]. Moreover, proper dietary antioxidant nutrient intake may reduce generation of free radicals and improve antioxidant status in RA patients^[96]. Finally, intake of certain antioxidant micronutrients particularly beta-cryptoxanthine, supplemental zinc, and possibly diet in fruits and cruciferous vegetables have been suggested to protect against the development of RA^[97].

Increasing anti-oxidant capacity in RA is a very attractive and potentially effective intervention. In current clinical practice, vitamin and micronutrient supplementation is frequently prescribed. However, we now know, that polypharmacy (prescription of a large number of medications) is one of the most significant reasons why patients forget to take their pills^[98]. However, the most important question concerning use of antioxidants, is that of suicidal oxidative stress^[19]. In certain conditions, such as presence of transition metals, antioxidants can act as pro-oxidants^[99]. Similarly, high concentrations of antioxidants can cause the cell to undergo severe oxidative stress ultimately resulting in suicidal cell death^[100].

Medication

In very recent years, the anti-oxidant potential of antiTNF therapy has also been investigated. Infliximab plays an essential role as an anti-oxidative agent against advanced glycation end-product formation, oxidative DNA damage and lipid peroxidation^[101], whereas etanercept acts as a regulator against pentosidine formation, oxidative DNA damage, and lipid peroxidation in RA patients^[102].

Exercise and oxidative stress

In the general population, exercise has been shown to in-

crease anti-oxidant capacity. Working *via* the physiological concept of hormesis (an ancient practice where the induction of a sub-lethal dose of toxin was used to increase tolerance of the organism to withstand higher doses of toxins) acute exercise increases free radical production^[103], in a dose-response fashion (*i.e.*, increasing intensity, increases free radical production). This exercised-induced increase in free radicals is due to the increased electron leak from the mitochondria as well as the alterations in blood flow and oxygen supply that occur in response to exercise

However, it has been consistently observed that trained individuals have high levels of antioxidant enzymes and certain nonenzymatic antioxidants in muscle^[106] and demonstrate greater resistance to exercise-induced or -imposed oxidative stress^[107,108]. Most likely, these adaptations result from cumulative effects of repeated exercise bouts on the gene expression of anti-oxidant enzymes. However, the attenuation of oxidative stress by exercise is reduced in the aging muscle, warranting concomitant nutritional supplementation with anti-oxidants to elicit the greatest potential benefits^[109].

EXERCISE IN RHEUMATOID ARTHRITIS

Exercise is a useful tool, with constantly increasing clinical relevance to several conditions. In recent years, a large number of studies have investigated the safety of different exercise modalities in RA. Despite the common misconception that it may increase joint pain and damage, all of the studies indicate that properly designed exercise interventions are safe and beneficial for RA patients^[110]. de Jong *et al*^[111-113] have investigated the safety of intensive aerobic exercise (in the form of cycling) in > 200RA patients; they concluded that all patients were able to achieve the pre-determined intensity targets. However, they pointed out that patients with severe joint damage may need attention^[114]. Along similar lines, resistance training improved body composition and muscle mass without any adverse effects on disease activity^[50]. Finally, we have recently completed a randomised trial looking at the effects of intensive aerobic exercise on cardiovascular risk factors in RA patients^[115]. From these and other studies^[116-120], it is clear that exercise is a safe intervention for RA patients and its use in the clinical setting is gaining significant support. Moreover, it is evident that exercise is able to reverse muscle wasting and increase muscle mass in RA patients. Indeed, the regenerative capacity of the RA muscle seems to be unaffected as the number of satellite cells (muscular stem cells that are utilised for muscular regeneration) present in it are preserved^[121] but the stimulus for their activation (i.e., exercise) is absent.

CONCLUSION

The role of oxidative stress in muscle wasting has been clearly demonstrated in several studies. However, to date there is no study looking at this in RA patients. We suggest that there is significant scope for such research in RA as the potential mechanisms by which oxidative stress

WJR | www.wjgnet.com

drives muscle wasting have been already described in other populations. Identification of specific mechanisms induced by RA-associated inflammation could significantly aid towards improvement of pharmacological and non-pharmacological interventions aiming to counteract oxidative stress in RA. In addition to effective control of inflammation *via* medication, exercise and nutrition may prove significant aids towards the reduction of oxidative stress (Figure 1).

REFERENCES

- 1 Alamanos Y, Drosos AA. Epidemiology of adult rheumatoid arthritis. *Autoimmun Rev* 2005; **4**: 130-136 [PMID: 15823498 DOI: 10.1016/j.autrev.2004.09.002]
- 2 Symmons D, Turner G, Webb R, Asten P, Barrett E, Lunt M, Scott D, Silman A. The prevalence of rheumatoid arthritis in the United Kingdom: new estimates for a new century. *Rheumatology* (Oxford) 2002; **41**: 793-800 [PMID: 12096230 DOI: 10.1093/rheumatology/41.7.793]
- 3 **Buch M**, Emery P. The aetiology and pathogenesis of rheumatiod arthritis. *Hospital Pharmacist* 2002; **9**: 5-10
- 4 Callahan LF, Yelin EH. The social and economic consequences of rheumatic disease. In: Klippel JH CL, Stone JH, Weyand, CM, editors. Primer on the rheumatic diseases. 12th ed. Atlanta: Arthritis Foundation, 2001: 1-4
- 5 Westhoff G, Listing J, Zink A. Loss of physical independence in rheumatoid arthritis: interview data from a representative sample of patients in rheumatologic care. *Arthritis Care Res* 2000; **13**: 11-22 [PMID: 11094922 DOI: 10.1002/1529-0131(200002)13: 1<11::AID-ART4>3.0.CO;2-5]
- 6 Pugner KM, Scott DI, Holmes JW, Hieke K. The costs of rheumatoid arthritis: an international long-term view. *Semin Arthritis Rheum* 2000; 29: 305-320 [PMID: 10805355 DOI: 10.1016/s0049-0172(00)80017-7]
- 7 Turesson C, Jacobsson LT. Epidemiology of extra-articular manifestations in rheumatoid arthritis. *Scand J Rheumatol* 2004; 33: 65-72 [PMID: 15163106 DOI: 10.1080/030097403100 04621]
- 8 Maradit-Kremers H, Crowson CS, Nicola PJ, Ballman KV, Roger VL, Jacobsen SJ, Gabriel SE. Increased unrecognized coronary heart disease and sudden deaths in rheumatoid arthritis: a population-based cohort study. *Arthritis Rheum* 2005; 52: 402-411 [PMID: 15693010 DOI: 10.1002/art.20853]
- 9 Gonzalez-Gay MA, Gonzalez-Juanatey C, Lopez-Diaz MJ, Piñeiro A, Garcia-Porrua C, Miranda-Filloy JA, Ollier WE, Martin J, Llorca J. HLA-DRB1 and persistent chronic inflammation contribute to cardiovascular events and cardiovascular mortality in patients with rheumatoid arthritis. *Arthritis Rheum* 2007; 57: 125-132 [PMID: 17266100 DOI: 10.1002/ art.22482]
- 10 Panoulas VF, Nikas SN, Smith JP, Douglas KM, Nightingale P, Milionis HJ, Treharne GJ, Toms TE, Kita MD, Kitas GD. Lymphotoxin 252A& gt; G polymorphism is common and associates with myocardial infarction in patients with rheumatoid arthritis. *Ann Rheum Dis* 2008; 67: 1550-1556 [PMID: 18230628 DOI: 10.1136/ard.2007.082594]
- 11 Panoulas VF, Stavropoulos-Kalinoglou A, Metsios GS, Smith JP, Milionis HJ, Douglas KM, Nightingale P, Kitas GD. Association of interleukin-6 (IL-6)-174G/C gene polymorphism with cardiovascular disease in patients with rheumatoid arthritis: the role of obesity and smoking. *Atherosclerosis* 2009; 204: 178-183 [PMID: 18848327 DOI: 10.1016/j.atherosclerosis. 2008.08.036]
- 12 **Mattey DL**, Dawes PT, Nixon NB, Goh L, Banks MJ, Kitas GD. Increased levels of antibodies to cytokeratin 18 in patients with rheumatoid arthritis and ischaemic heart disease.

Ann Rheum Dis 2004; **63**: 420-425 [PMID: 15020337 DOI: 10.1136/ard.2003.008011]

- 13 Panoulas VF, Douglas KM, Milionis HJ, Stavropoulos-Kalinglou A, Nightingale P, Kita MD, Tselios AL, Metsios GS, Elisaf MS, Kitas GD. Prevalence and associations of hypertension and its control in patients with rheumatoid arthritis. *Rheumatology* (Oxford) 2007; 46: 1477-1482 [PMID: 17704521 DOI: 10.1093/rheumatology/kem169]
- 14 Toms TE, Panoulas VF, Douglas KM, Griffiths HR, Kitas GD. Lack of association between glucocorticoid use and presence of the metabolic syndrome in patients with rheumatoid arthritis: a cross-sectional study. *Arthritis Res Ther* 2008; 10: R145 [PMID: 19091101 DOI: 10.1186/ar2578]
- 15 Stevens RJ, Douglas KM, Saratzis AN, Kitas GD. Inflammation and atherosclerosis in rheumatoid arthritis. *Expert Rev Mol Med* 2005; 7: 1-24 [PMID: 15876361 DOI: 10.1017/ S1462399405009154]
- 16 Gonzalez A, Maradit Kremers H, Crowson CS, Ballman KV, Roger VL, Jacobsen SJ, O'Fallon WM, Gabriel SE. Do cardiovascular risk factors confer the same risk for cardiovascular outcomes in rheumatoid arthritis patients as in non-rheumatoid arthritis patients? *Ann Rheum Dis* 2008; 67: 64-69 [PMID: 17517756 DOI: 10.1136/ard.2006.059980]
- 17 Cojocaru M, Cojocaru IM, Silosi I, Vrabie CD, Tanasescu R. Extra-articular Manifestations in Rheumatoid Arthritis. *Maedica* (Buchar) 2010; 5: 286-291 [PMID: 21977172]
- 18 Prete M, Racanelli V, Digiglio L, Vacca A, Dammacco F, Perosa F. Extra-articular manifestations of rheumatoid arthritis: An update. *Autoimmun Rev* 2011; 11: 123-131 [PMID: 21939785 DOI: 10.1016/j.autrev.2011.09.001]
- 19 Mahajan A, Tandon VR. Antioxidants and Rheumatoid Arthritis. J Indian Rheumatol Assoc 2004; 12: 139-142
- 20 Winrow VR, Winyard PG, Morris CJ, Blake DR. Free radicals in inflammation: second messengers and mediators of tissue destruction. *Br Med Bull* 1993; **49**: 506-522 [PMID: 8221019]
- 21 Speakman JR. Body size, energy metabolism and lifespan. J Exp Biol 2005; 208: 1717-1730 [PMID: 15855403 DOI: 10.1242/ jeb.01556]
- 22 Vasanthi P, Nalini G, Rajasekhar G. Status of oxidative stress in rheumatoid arthritis. *Int J Rheum Dis* 2009; **12**: 29-33 [PMID: 20374313 DOI: 10.1111/j.1756-185X.2009.01375.x]
- 23 Sukkar SG, Rossi E. Oxidative stress and nutritional prevention in autoimmune rheumatic diseases. *Autoimmun Rev* 2004; **3**: 199-206 [PMID: 15110232 DOI: 10.1016/ j.autrev.2003.09.002]
- 24 Maly FE, Cross AR, Jones OT, Wolf-Vorbeck G, Walker C, Dahinden CA, De Weck AL. The superoxide generating system of B cell lines. Structural homology with the phagocytic oxidase and triggering via surface Ig. J Immunol 1988; 140: 2334-2339 [PMID: 2832475]
- 25 Baskol G, Demir H, Baskol M, Kilic E, Ates F, Kocer D, Muhtaroglu S. Assessment of paraoxonase 1 activity and malondialdehyde levels in patients with rheumatoid arthritis. *Clin Biochem* 2005; 38: 951-955 [PMID: 16055108 DOI: 10.1016/j.clinbiochem.2005.06.010]
- 26 Hassan MQ, Hadi RA, Al-Rawi ZS, Padron VA, Stohs SJ. The glutathione defense system in the pathogenesis of rheumatoid arthritis. *J Appl Toxicol* 2001; 21: 69-73 [PMID: 11180282 DOI: 10.1002/jat.736]
- 27 Hadjigogos K. The role of free radicals in the pathogenesis of rheumatoid arthritis. *Panminerva Med* 2003; 45: 7-13 [PMID: 12682616]
- 28 Rowley D, Gutteridge JM, Blake D, Farr M, Halliwell B. Lipid peroxidation in rheumatoid arthritis: thiobarbituric acidreactive material and catalytic iron salts in synovial fluid from rheumatoid patients. *Clin Sci* (Lond) 1984; 66: 691-695 [PMID: 6723205]
- 29 **Kamanli A**, Naziroğlu M, Aydilek N, Hacievliyagil C. Plasma lipid peroxidation and antioxidant levels in patients

with rheumatoid arthritis. *Cell Biochem Funct* 2004; **22**: 53-57 [PMID: 14695655 DOI: 10.1002/cbf.1055]

- 30 Kalpakcioglu B, Senel K. The interrelation of glutathione reductase, catalase, glutathione peroxidase, superoxide dismutase, and glucose-6-phosphate in the pathogenesis of rheumatoid arthritis. *Clin Rheumatol* 2008; 27: 141-145 [PMID: 17912575 DOI: 10.1007/s10067-007-0746-3]
- 31 Henderson CJ, Panush RS. Diets, dietary supplements, and nutritional therapies in rheumatic diseases. *Rheum Dis Clin North Am* 1999; 25: 937-968, ix [PMID: 10573768 DOI: 10.1016/s0889-857x(05)70112-5]
- 32 Roubenoff R, Roubenoff RA, Cannon JG, Kehayias JJ, Zhuang H, Dawson-Hughes B, Dinarello CA, Rosenberg IH. Rheumatoid cachexia: cytokine-driven hypermetabolism accompanying reduced body cell mass in chronic inflammation. J Clin Invest 1994; 93: 2379-2386 [PMID: 8200971 DOI: 10.1172/JCI117244]
- 33 Summers GD, Metsios GS, Stavropoulos-Kalinoglou A, Kitas GD. Rheumatoid cachexia and cardiovascular disease. Nat Rev Rheumatol 2010; 6: 445-451 [PMID: 20647995 DOI: 10.1038/nrrheum.2010.105]
- 34 **Roubenoff R**. Rheumatoid cachexia: a complication of rheumatoid arthritis moves into the 21st century. *Arthritis Res Ther* 2009; **11**: 108 [PMID: 19439037 DOI: 10.1186/ar2658]
- 35 Metsios GS, Stavropoulos-Kalinoglou A, Panoulas VF, Sandoo A, Toms TE, Nevill AM, Koutedakis Y, Kitas GD. Rheumatoid cachexia and cardiovascular disease. *Clin Exp Rheumatol* 2009; 27: 985-988 [PMID: 20149317]
- 36 Elkan AC, Håkansson N, Frostegård J, Cederholm T, Hafström I. Rheumatoid cachexia is associated with dyslipidemia and low levels of atheroprotective natural antibodies against phosphorylcholine but not with dietary fat in patients with rheumatoid arthritis: a cross-sectional study. *Arthritis Res Ther* 2009; **11**: R37 [PMID: 19284557 DOI: 10.1186/ ar2643]
- 37 Engvall IL, Elkan AC, Tengstrand B, Cederholm T, Brismar K, Hafstrom I. Cachexia in rheumatoid arthritis is associated with inflammatory activity, physical disability, and low bio-available insulin-like growth factor. *Scand J Rheumatol* 2008; **37**: 321-328 [PMID: 18666027 DOI: 10.1080/03009740802055984]
- 38 Kremers HM, Nicola PJ, Crowson CS, Ballman KV, Gabriel SE. Prognostic importance of low body mass index in relation to cardiovascular mortality in rheumatoid arthritis. *Arthritis Rheum* 2004; 50: 3450-3457 [PMID: 15529378 DOI: 10.1002/art.20612]
- 39 Giles JT, Bartlett SJ, Andersen RE, Fontaine KR, Bathon JM. Association of body composition with disability in rheumatoid arthritis: impact of appendicular fat and lean tissue mass. *Arthritis Rheum* 2008; 59: 1407-1415 [PMID: 18821641 DOI: 10.1002/art.24109]
- 40 Evans W. Functional and metabolic consequences of sarcopenia. J Nutr 1997; 127: 998S-1003S [PMID: 9164283]
- 41 Nair KS. Aging muscle. Am J Clin Nutr 2005; 81: 953-963 [PMID: 15883415]
- 42 **Kitas GD**, Erb N. Tackling ischaemic heart disease in rheumatoid arthritis. *Rheumatology* (Oxford) 2003; **42**: 607-613 [PMID: 12709534 DOI: 10.1093/rheumatology/keg175]
- 43 **Gabay C**, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 1999; **340**: 448-454 [PMID: 9971870 DOI: 10.1056/NEJM199902113400607]
- Bakkar N, Guttridge DC. NF-kappaB signaling: a tale of two pathways in skeletal myogenesis. *Physiol Rev* 2010; 90: 495-511 [PMID: 20393192 DOI: 10.1152/physrev.00040.2009]
- 45 Broussard SR, McCusker RH, Novakofski JE, Strle K, Shen WH, Johnson RW, Dantzer R, Kelley KW. IL-1beta impairs insulin-like growth factor i-induced differentiation and downstream activation signals of the insulin-like growth factor i receptor in myoblasts. *J Immunol* 2004; **172**: 7713-7720 [PMID: 15187154 DOI: 10.4049/jimmunol.172.12.7713]

- 46 van Hall G, Steensberg A, Fischer C, Keller C, Møller K, Moseley P, Pedersen BK. Interleukin-6 markedly decreases skeletal muscle protein turnover and increases nonmuscle amino acid utilization in healthy individuals. J Clin Endocrinol Metab 2008; 93: 2851-2858 [PMID: 18430776 DOI: 10.1210/ jc.2007-2223]
- 47 Metsios GS, Stavropoulos-Kalinoglou A, Douglas KM, Koutedakis Y, Nevill AM, Panoulas VF, Kita M, Kitas GD. Blockade of tumour necrosis factor-alpha in rheumatoid arthritis: effects on components of rheumatoid cachexia. *Rheumatology* (Oxford) 2007; 46: 1824-1827 [PMID: 18032540 DOI: 10.1093/rheumatology/kem291]
- 48 Marcora SM, Chester KR, Mittal G, Lemmey AB, Maddison PJ. Randomized phase 2 trial of anti-tumor necrosis factor therapy for cachexia in patients with early rheumatoid arthritis. *Am J Clin Nutr* 2006; 84: 1463-1472 [PMID: 17158431]
- 49 Stavropoulos-Kalinoglou A, Metsios GS, Smith JP, Panoulas VF, Douglas KM, Jamurtas AZ, Koutedakis Y, Kitas GD. What predicts obesity in patients with rheumatoid arthritis? An investigation of the interactions between lifestyle and inflammation. *Int J Obes* (Lond) 2010; **34**: 295-301 [PMID: 19859075 DOI: 10.1038/ijo.2009.220]
- 50 Marcora SM, Lemmey AB, Maddison PJ. Can progressive resistance training reverse cachexia in patients with rheumatoid arthritis? Results of a pilot study. *J Rheumatol* 2005; 32: 1031-1039 [PMID: 15940763 DOI: 10.2147/cia.s42136]
- 51 Lemmey AB, Marcora SM, Chester K, Wilson S, Casanova F, Maddison PJ. Effects of high-intensity resistance training in patients with rheumatoid arthritis: a randomized controlled trial. *Arthritis Rheum* 2009; 61: 1726-1734 [PMID: 19950325 DOI: 10.1002/art.24891]
- 52 Metsios GS, Stavropoulos-Kalinoglou A, Panoulas VF, Wilson M, Nevill AM, Koutedakis Y, Kitas GD. Association of physical inactivity with increased cardiovascular risk in patients with rheumatoid arthritis. *Eur J Cardiovasc Prev Rehabil* 2009; 16: 188-194 [PMID: 19238083 DOI: 10.1097/ HJR.0b013e3283271ceb]
- 53 Stavropoulos-Kalinoglou A, Metsios GS, Koutedakis Y, Nevill AM, Douglas KM, Jamurtas A, van Zanten JJ, Labib M, Kitas GD. Redefining overweight and obesity in rheumatoid arthritis patients. *Ann Rheum Dis* 2007; 66: 1316-1321 [PMID: 17289757 DOI: 10.1136/ard.2006.060319]
- 54 Wagenmakers AJ, van Riel NA, Frenneaux MP, Stewart PM. Integration of the metabolic and cardiovascular effects of exercise. *Essays Biochem* 2006; 42: 193-210 [PMID: 17144889 DOI: 10.1042/bse0420193]
- 55 **Berg AH**, Scherer PE. Adipose tissue, inflammation, and cardiovascular disease. *Circ Res* 2005; **96**: 939-949 [PMID: 15890981 DOI: 10.1161/01.RES.0000163635.62927.34]
- 56 Andersson CX, Gustafson B, Hammarstedt A, Hedjazifar S, Smith U. Inflamed adipose tissue, insulin resistance and vascular injury. *Diabetes Metab Res Rev* 2008; 24: 595-603 [PMID: 18756581 DOI: 10.1002/dmrr.889]
- 57 Shaw CS, Jones DA, Wagenmakers AJ. Network distribution of mitochondria and lipid droplets in human muscle fibres. *Histochem Cell Biol* 2008; **129**: 65-72 [PMID: 17938948 DOI: 10.1007/s00418-007-0349-8]
- 58 Shaw CS, Clark J, Wagenmakers AJ. The effect of exercise and nutrition on intramuscular fat metabolism and insulin sensitivity. *Annu Rev Nutr* 2010; 30: 13-34 [PMID: 20373917 DOI: 10.1146/annurev.nutr.012809.104817]
- 59 Wagenmakers AJ. Tracers to investigate protein and amino acid metabolism in human subjects. *Proc Nutr Soc* 1999; 58: 987-1000 [PMID: 10817167 DOI: 10.1017/s0029665199001305]
- 60 Barrett EJ, Eggleston EM, Inyard AC, Wang H, Li G, Chai W, Liu Z. The vascular actions of insulin control its delivery to muscle and regulate the rate-limiting step in skeletal muscle insulin action. *Diabetologia* 2009; 52: 752-764 [PMID: 19283361 DOI: 10.1007/s00125-009-1313-z]

- 61 Naruse K, Rask-Madsen C, Takahara N, Ha SW, Suzuma K, Way KJ, Jacobs JR, Clermont AC, Ueki K, Ohshiro Y, Zhang J, Goldfine AB, King GL. Activation of vascular protein kinase C-beta inhibits Akt-dependent endothelial nitric oxide synthase function in obesity-associated insulin resistance. *Diabetes* 2006; 55: 691-698 [PMID: 16505232 DOI: 10.2337/ diabetes.55.03.06.db05-0771]
- 62 Silver AE, Beske SD, Christou DD, Donato AJ, Moreau KL, Eskurza I, Gates PE, Seals DR. Overweight and obese humans demonstrate increased vascular endothelial NAD(P)H oxidase-p47(phox) expression and evidence of endothelial oxidative stress. *Circulation* 2007; 115: 627-637 [PMID: 17242275 DOI: 10.1161/CIRCULATIONAHA.106.657486]
- 63 **Powers SK**, Smuder AJ, Judge AR. Oxidative stress and disuse muscle atrophy: cause or consequence? *Curr Opin Clin Nutr Metab Care* 2012; **15**: 240-245 [PMID: 22466926 DOI: 10.1097/MCO.0b013e328352b4c2]
- 64 Falk DJ, Deruisseau KC, Van Gammeren DL, Deering MA, Kavazis AN, Powers SK. Mechanical ventilation promotes redox status alterations in the diaphragm. *J Appl Physiol* (1985) 2006; 101: 1017-1024 [PMID: 16675618 DOI: 10.1152/ japplphysiol.00104.2006]
- 65 Kavazis AN, Talbert EE, Smuder AJ, Hudson MB, Nelson WB, Powers SK. Mechanical ventilation induces diaphragmatic mitochondrial dysfunction and increased oxidant production. *Free Radic Biol Med* 2009; 46: 842-850 [PMID: 19185055 DOI: 10.1016/j.freeradbiomed.2009.01.002]
- 66 Powers SK, Hudson MB, Nelson WB, Talbert EE, Min K, Szeto HH, Kavazis AN, Smuder AJ. Mitochondria-targeted antioxidants protect against mechanical ventilation-induced diaphragm weakness. *Crit Care Med* 2011; **39**: 1749-1759 [PMID: 21460706 DOI: 10.1097/CCM.0b013e3182190b62]
- 67 Min K, Smuder AJ, Kwon OS, Kavazis AN, Szeto HH, Powers SK. Mitochondrial-targeted antioxidants protect skeletal muscle against immobilization-induced muscle atrophy. J Appl Physiol (1985) 2011; 111: 1459-1466 [PMID: 21817113 DOI: 10.1152/japplphysiol.00591.2011]
- 68 McClung JM, Judge AR, Talbert EE, Powers SK. Calpain-1 is required for hydrogen peroxide-induced myotube atrophy. *Am J Physiol Cell Physiol* 2009; **296**: C363-C371 [PMID: 19109522 DOI: 10.1152/ajpcell.00497.2008]
- 69 Whidden MA, McClung JM, Falk DJ, Hudson MB, Smuder AJ, Nelson WB, Powers SK. Xanthine oxidase contributes to mechanical ventilation-induced diaphragmatic oxidative stress and contractile dysfunction. *J Appl Physiol* (1985) 2009; 106: 385-394 [PMID: 18974366 DOI: 10.1152/japplphysiol.91106.2008]
- Kondo H, Miura M, Itokawa Y. Oxidative stress in skeletal muscle atrophied by immobilization. *Acta Physiol Scand* 1991; 142: 527-528 [PMID: 1950601 DOI: 10.1111/j.1748-1716.1991. tb09191.x]
- 71 Powers SK, Kavazis AN, DeRuisseau KC. Mechanisms of disuse muscle atrophy: role of oxidative stress. *Am J Physiol Regul Integr Comp Physiol* 2005; 288: R337-R344 [PMID: 15637170 DOI: 10.1152/ajpregu.00469.2004]
- 72 Powers SK, Kavazis AN, McClung JM. Oxidative stress and disuse muscle atrophy. *J Appl Physiol* (1985) 2007; 102: 2389-2397 [PMID: 17289908 DOI: 10.1152/japplphysiol.01202.2006]
- 73 Li YP, Chen Y, Li AS, Reid MB. Hydrogen peroxide stimulates ubiquitin-conjugating activity and expression of genes for specific E2 and E3 proteins in skeletal muscle myotubes. *Am J Physiol Cell Physiol* 2003; 285: C806-C812 [PMID: 12773310 DOI: 10.1152/ajpcell.00129.2003]
- 74 McClung JM, Van Gammeren D, Whidden MA, Falk DJ, Kavazis AN, Hudson MB, Gayan-Ramirez G, Decramer M, De-Ruisseau KC, Powers SK. Apocynin attenuates diaphragm oxidative stress and protease activation during prolonged mechanical ventilation. *Crit Care Med* 2009; **37**: 1373-1379

[PMID: 19242334 DOI: 10.1097/CCM.0b013e31819cef63]

- 75 Dodd SL, Gagnon BJ, Senf SM, Hain BA, Judge AR. Rosmediated activation of NF-kappaB and Foxo during muscle disuse. *Muscle Nerve* 2010; 41: 110-113 [PMID: 19813194 DOI: 10.1002/mus.21526]
- 76 McClung JM, Judge AR, Powers SK, Yan Z. p38 MAPK links oxidative stress to autophagy-related gene expression in cachectic muscle wasting. *Am J Physiol Cell Physiol* 2010; 298: C542-C549 [PMID: 19955483 DOI: 10.1152/ajpcell.00192.2009]
- 77 Smuder AJ, Kavazis AN, Hudson MB, Nelson WB, Powers SK. Oxidation enhances myofibrillar protein degradation via calpain and caspase-3. *Free Radic Biol Med* 2010; 49: 1152-1160 [PMID: 20600829 DOI: 10.1016/j.freeradbiomed.2010.06.025]
- 78 O'Loghlen A, Pérez-Morgado MI, Salinas M, Martín ME. N-acetyl-cysteine abolishes hydrogen peroxide-induced modification of eukaryotic initiation factor 4F activity via distinct signalling pathways. *Cell Signal* 2006; 18: 21-31 [PMID: 15907373 DOI: 10.1016/j.cellsig.2005.03.013]
- 79 Zhang L, Kimball SR, Jefferson LS, Shenberger JS. Hydrogen peroxide impairs insulin-stimulated assembly of mTORC1. *Free Radic Biol Med* 2009; 46: 1500-1509 [PMID: 19281842 DOI: 10.1016/j.freeradbiomed.2009.03.001]
- 80 **Kimball SR**, Jefferson LS. New functions for amino acids: effects on gene transcription and translation. *Am J Clin Nutr* 2006; **83**: 500S-507S [PMID: 16470021]
- 81 Marcell TJ. Sarcopenia: causes, consequences, and preventions. J Gerontol A Biol Sci Med Sci 2003; 58: M911-M916 [PMID: 14570858 DOI: 10.1093/gerona/58.10.m911]
- 82 HARMAN D. Aging: a theory based on free radical and radiation chemistry. *J Gerontol* 1956; **11**: 298-300 [PMID: 13332224 DOI: 10.1093/geronj/11.3.298]
- 83 Lass A, Sohal BH, Weindruch R, Forster MJ, Sohal RS. Caloric restriction prevents age-associated accrual of oxidative damage to mouse skeletal muscle mitochondria. *Free Radic Biol Med* 1998; 25: 1089-1097 [PMID: 9870563 DOI: 10.1016/s0891-5849(98)00144-0]
- 84 Broome CS, Kayani AC, Palomero J, Dillmann WH, Mestril R, Jackson MJ, McArdle A. Effect of lifelong overexpression of HSP70 in skeletal muscle on age-related oxidative stress and adaptation after nondamaging contractile activity. *FASEB J* 2006; 20: 1549-1551 [PMID: 16723383 DOI: 10.1096/fj.05-4935fie]
- 85 Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002; 82: 47-95 [PMID: 11773609 DOI: 10.1152/physrev.00018.2001]
- 86 Jackson MJ, Papa S, Bolaños J, Bruckdorfer R, Carlsen H, Elliott RM, Flier J, Griffiths HR, Heales S, Holst B, Lorusso M, Lund E, Øivind Moskaug J, Moser U, Di Paola M, Polidori MC, Signorile A, Stahl W, Viña-Ribes J, Astley SB. Antioxidants, reactive oxygen and nitrogen species, gene induction and mitochondrial function. *Mol Aspects Med* 2002; 23: 209-285 [PMID: 12079772 DOI: 10.1016/s0098-2997(02)00018-3]
- 87 **Sanz A**, Pamplona R, Barja G. Is the mitochondrial free radical theory of aging intact? *Antioxid Redox Signal* 2006; **8**: 582-599 [PMID: 16677102 DOI: 10.1089/ars.2006.8.582]
- 88 Muller FL, Liu Y, Van Remmen H. Complex III releases superoxide to both sides of the inner mitochondrial membrane. *J Biol Chem* 2004; 279: 49064-49073 [PMID: 15317809 DOI: 10.1074/jbc.M407715200]
- 89 Iñarrea P. Purification and determination of activity of mitochondrial cyanide-sensitive superoxide dismutase in rat tissue extract. *Methods Enzymol* 2002; 349: 106-114 [PMID: 11912900 DOI: 10.1016/s0076-6879(02)49326-3]
- 90 Jackson MJ. Skeletal muscle aging: role of reactive oxygen species. *Crit Care Med* 2009; **37**: S368-S371 [PMID: 20046122 DOI: 10.1097/CCM.0b013e3181b6f97f]
- 91 Muller FL, Song W, Liu Y, Chaudhuri A, Pieke-Dahl S,

Stavropoulos-Kalinoglou A et al. Muscle wasting and oxidative stress in RA

Strong R, Huang TT, Epstein CJ, Roberts LJ, Csete M, Faulkner JA, Van Remmen H. Absence of CuZn superoxide dismutase leads to elevated oxidative stress and acceleration of age-dependent skeletal muscle atrophy. *Free Radic Biol Med* 2006; **40**: 1993-2004 [PMID: 16716900 DOI: 10.1016/j.free radbiomed.2006.01.036]

- 92 De Bandt M, Grossin M, Driss F, Pincemail J, Babin-Chevaye C, Pasquier C. Vitamin E uncouples joint destruction and clinical inflammation in a transgenic mouse model of rheumatoid arthritis. *Arthritis Rheum* 2002; 46: 522-532 [PMID: 11840456]
- 93 Helmy M, Shohayeb M, Helmy MH, el-Bassiouni EA. Antioxidants as adjuvant therapy in rheumatoid disease. A preliminary study. *Arzneimittelforschung* 2001; **51**: 293-298 [PMID: 11367869 DOI: 10.1055/s-0031-1300040]
- 94 Jaswal S, Mehta HC, Sood AK, Kaur J. Antioxidant status in rheumatoid arthritis and role of antioxidant therapy. *Clin Chim Acta* 2003; 338: 123-129 [PMID: 14637276 DOI: 10.1016/ j.cccn.2003.08.011]
- 95 Hagfors L, Leanderson P, Sköldstam L, Andersson J, Johansson G. Antioxidant intake, plasma antioxidants and oxidative stress in a randomized, controlled, parallel, Mediterranean dietary intervention study on patients with rheumatoid arthritis. *Nutr J* 2003; **2**: 5 [PMID: 12952549 DOI: 10.1186/1475-2891-2-5]
- 96 Bae SC, Kim SJ, Sung MK. Inadequate antioxidant nutrient intake and altered plasma antioxidant status of rheumatoid arthritis patients. *J Am Coll Nutr* 2003; 22: 311-315 [PMID: 12897046 DOI: 10.1080/07315724.2003.10719309]
- 97 Cerhan JR, Saag KG, Merlino LA, Mikuls TR, Criswell LA. Antioxidant micronutrients and risk of rheumatoid arthritis in a cohort of older women. *Am J Epidemiol* 2003; **157**: 345-354 [PMID: 12578805 DOI: 10.1093/aje/kwf205]
- 98 Treharne GJ, Douglas KM, Iwaszko J, Panoulas VF, Hale ED, Mitton DL, Piper H, Erb N, Kitas GD. Polypharmacy among people with rheumatoid arthritis: the role of age, disease duration and comorbidity. *Musculoskeletal Care* 2007; 5: 175-190 [PMID: 17623274 DOI: 10.1002/msc.112]
- 99 Cao G, Sofic E, Prior RL. Antioxidant and prooxidant behavior of flavonoids: structure-activity relationships. *Free Radic Biol Med* 1997; 22: 749-760 [PMID: 9119242 DOI: 10.1016/ s0891-5849(96)00351-6]
- 100 Offer T, Russo A, Samuni A. The pro-oxidative activity of SOD and nitroxide SOD mimics. *FASEB J* 2000; 14: 1215-1223 [PMID: 10834943 DOI: 10.1016/s0891-5849(99)90541-5]
- 101 Kageyama Y, Takahashi M, Ichikawa T, Torikai E, Nagano A. Reduction of oxidative stress marker levels by anti-TNF-alpha antibody, infliximab, in patients with rheumatoid arthritis. *Clin Exp Rheumatol* 2008; 26: 73-80 [PMID: 18328150]
- 102 Kageyama Y, Takahashi M, Nagafusa T, Torikai E, Nagano A. Etanercept reduces the oxidative stress marker levels in patients with rheumatoid arthritis. *Rheumatol Int* 2008; 28: 245-251 [PMID: 17661050 DOI: 10.1007/s00296-007-0419-1]
- 103 Davison GW, Morgan RM, Hiscock N, Garcia JM, Grace F, Boisseau N, Davies B, Castell L, McEneny J, Young IS, Hullin D, Ashton T, Bailey DM. Manipulation of systemic oxygen flux by acute exercise and normobaric hypoxia: implications for reactive oxygen species generation. *Clin Sci* (Lond) 2006; 110: 133-141 [PMID: 16197367 DOI: 10.1042/CS20050135]
- 104 Urso ML, Clarkson PM. Oxidative stress, exercise, and antioxidant supplementation. *Toxicology* 2003; 189: 41-54 [PMID: 12821281 DOI: 10.1016/S0300-483X(03)00151-3]
- 105 Nikolaidis MG, Jamurtas AZ. Blood as a reactive species generator and redox status regulator during exercise. Arch Biochem Biophys 2009; 490: 77-84 [PMID: 19712664 DOI: 10.1016/j.abb.2009.08.015]
- 106 **Kostaropoulos IA**, Nikolaidis MG, Jamurtas AZ, Ikonomou GV, Makrygiannis V, Papadopoulos G, Kouretas D. Comparison of the blood redox status between long-distance and

short-distance runners. *Physiol Res* 2006; **55**: 611-616 [PMID: 16497108]

- 107 Ji LL, Leeuwenburgh C, Leichtweis S, Gore M, Fiebig R, Hollander J, Bejma J. Oxidative stress and aging. Role of exercise and its influences on antioxidant systems. *Ann N Y Acad Sci* 1998; **854**: 102-117 [PMID: 9928424 DOI: 10.1111/ j.1749-6632.1998.tb09896.x]
- 108 Nikolaidis MG, Jamurtas AZ, Paschalis V, Fatouros IG, Koutedakis Y, Kouretas D. The effect of muscle-damaging exercise on blood and skeletal muscle oxidative stress: magnitude and time-course considerations. *Sports Med* 2008; 38: 579-606 [PMID: 18557660 DOI: 10.2165/00007256-200838070-00005]
- 109 Fulle S, Protasi F, Di Tano G, Pietrangelo T, Beltramin A, Boncompagni S, Vecchiet L, Fanò G. The contribution of reactive oxygen species to sarcopenia and muscle ageing. *Exp Gerontol* 2004; **39**: 17-24 [PMID: 14724060 DOI: 10.1016/ j.exger.2003.09.012]
- 110 Metsios GS, Stavropoulos-Kalinoglou A, Veldhuijzen van Zanten JJ, Treharne GJ, Panoulas VF, Douglas KM, Koutedakis Y, Kitas GD. Rheumatoid arthritis, cardiovascular disease and physical exercise: a systematic review. *Rheumatology* (Oxford) 2008; **47**: 239-248 [PMID: 18045810 DOI: 10.1093/ rheumatology/kem260]
- 111 de Jong Z, Munneke M, Zwinderman AH, Kroon HM, Jansen A, Ronday KH, van Schaardenburg D, Dijkmans BA, Van den Ende CH, Breedveld FC, Vliet Vlieland TP, Hazes JM. Is a long-term high-intensity exercise program effective and safe in patients with rheumatoid arthritis? Results of a randomized controlled trial. *Arthritis Rheum* 2003; 48: 2415-2424 [PMID: 13130460 DOI: 10.1002/art.11216]
- 112 de Jong Z, Munneke M, Kroon HM, van Schaardenburg D, Dijkmans BA, Hazes JM, Vliet Vlieland TP. Long-term follow-up of a high-intensity exercise program in patients with rheumatoid arthritis. *Clin Rheumatol* 2009; 28: 663-671 [PMID: 19247575 DOI: 10.1007/s10067-009-1125-z]
- 113 de Jong Z, Vliet Vlieland TP. Safety of exercise in patients with rheumatoid arthritis. *Curr Opin Rheumatol* 2005; **17**: 177-182 [PMID: 15711232 DOI: 10.1097/01.bor.0000151400.33899.88]
- 114 de Jong Z, Munneke M, Zwinderman AH, Kroon HM, Ronday KH, Lems WF, Dijkmans BA, Breedveld FC, Vliet Vlieland TP, Hazes JM, Huizinga TW. Long term high intensity exercise and damage of small joints in rheumatoid arthritis. *Ann Rheum Dis* 2004; 63: 1399-1405 [PMID: 15479889 DOI: 10.1136/ard.2003.015826]
- 115 Stavropoulos-Kalinoglou A, Metsios GS, Veldhuijzen van Zanten JJ, Nightingale P, Kitas GD, Koutedakis Y. Individualised aerobic and resistance exercise training improves cardiorespiratory fitness and reduces cardiovascular risk in patients with rheumatoid arthritis. *Ann Rheum Dis* 2013; 72: 1819-1825 [PMID: 23155222 DOI: 10.1136/annrheumdis-2012-202075]
- 116 Allen SH, Minor MA, Hillman LS, Kay DR. Effect of exercise on the bone mineral density and bone remodelling indices in women with rheumatoid arthritis: 2 case studies. *J Rheumatol* 1993; 20: 1247-1249 [PMID: 8371231]
- 117 Anandarajah AP, Schwarz EM. Dynamic exercises in patients with rheumatoid arthritis. *Ann Rheum Dis* 2004; 63: 1359-1361 [PMID: 15479884 DOI: 10.1136/ard.2004.020693]
- 118 **Baslund B**, Lyngberg K, Andersen V, Halkjaer Kristensen J, Hansen M, Klokker M, Pedersen BK. Effect of 8 wk of bicycle training on the immune system of patients with rheumatoid arthritis. *J Appl Physiol* (1985) 1993; **75**: 1691-1695 [PMID: 8282621]
- 119 Bilberg A, Ahlmén M, Mannerkorpi K. Moderately intensive exercise in a temperate pool for patients with rheumatoid arthritis: a randomized controlled study. *Rheumatology* (Oxford) 2005; 44: 502-508 [PMID: 15728422 DOI: 10.1093/rheumatology/keh528]

120 de Jong Z, Munneke M, Lems WF, Zwinderman AH, Kroon HM, Pauwels EK, Jansen A, Ronday KH, Dijkmans BA, Breedveld FC, Vliet Vlieland TP, Hazes JM. Slowing of bone loss in patients with rheumatoid arthritis by long-term highintensity exercise: results of a randomized, controlled trial. *Arthritis Rheum* 2004; 50: 1066-1076 [PMID: 15077288 DOI: 10.1002/art.20117]

121 Beenakker KG, Duijnisveld BJ, Van Der Linden HM, Visser CP, Westendorp RG, Butler-Brown G, Nelissen RG, Maier AB. Muscle characteristics in patients with chronic systemic inflammation. *Muscle Nerve* 2012; 46: 204-209 [PMID: 22806369 DOI: 10.1002/mus.23291]

> P- Reviewer: La Montagna G, Sakkas L S- Editor: Ji FF L- Editor: A E- Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.54 World J Rheumatol 2014 November 12; 4(3): 54-61 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

REVIEW

Diagnosis and pharmacologic management of neuropathic pain among patients with chronic low back pain

Ferdi Yavuz, Umut Guzelkucuk

Ferdi Yavuz, The Clinic of Physical Medicine and Rehabilitation, Military Hospital of Etimesgut, 06790 Ankara, Turkey Umut Guzelkucuk, Department of Physical Medicine and Rehabilitation, TAF Rehabilitation Centre, Gulhane Military Medical Academy, 06790 Ankara, Turkey

Author contributions: Guzelkucuk U performed the review of literature; Yavuz F written and revised the article.

Correspondence to: Ferdi Yavuz, MD, The Clinic of Physical Medicine and Rehabilitation, Military Hospital of Etimesgut, The Street of Erler, 06790 Ankara, Turkey. ferdiyavuz@yahoo.com Telephone: +90-312-3461311 Fax: +90-312-2911009 Received: June 1, 2014 Revised: September 28, 2014

Accepted: October 14, 2014

Published online: November 12, 2014

Abstract

Chronic low back pain consists of both nociceptive and neuropathic mechanisms and can be classified as a mixed pain syndrome. Neuropathic component of chronic low back pain has often been under-recognized and under-treated by the physicians. Recent studies have demonstrated that approximately 20%-55% of chronic low back pain patients have neuropathic pain symptoms. An altered peripheral, spinal, and supraspinal processing of pain arising as a result of a lesion affecting the nerves system are the major contributor to neuropathic low back pain. The clinical evaluation is still the gold standard for assessment and diagnosis of neuropathic low back pain. Although diagnosis can be difficult due to the lack of reliable gold standard diagnostic test for neuropathic low back pain, screening tools may help non-specialists, in particular, to identify potential patients with neuropathic low back pain who require further diagnostic evaluation and pain management. Several screening tools for neuropathic pain have been developed and tested with different patient populations. Among the screening tools, the painDETECT questionnaire and the Standardized Evaluation of Pain are validated in patients with low back pain. The Standardized Evaluation of Pain may lead to more effective

in discriminating between neuropathic and nociceptive pain in patients with low back pain according to the higher rate of sensitivity and its validity in patients with low back pain. However, the most appropriate approach is still to combine findings on physical and neurologic examinations and patient's report in distinguishing neuropathic pain from nociceptive pain. The clinical examination including bedside sensory tests is still the best available tool for assessment and diagnosis neuropathic pain among patients with chronic low back pain. Due to the fact that chronic low back pain consists of both nociceptive and neuropathic mechanisms, a multimodal treatment approach is more rational in the management of patients with chronic low back pain. Therefore, combination therapy including drugs with different mechanisms of action should be given to the patients with chronic low back pain.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Low back pain; Neuropathic; Pharmacotherapy; Screening; Questionnaire

Core tip: Neuropathic component of chronic low back pain has often been under-recognized and undertreated by the physicians. Recent studies have demonstrated that approximately 20%-55% of chronic low back pain patients have neuropathic pain symptoms. The clinical examination including bedside sensory tests is still the best available tool for assessment and diagnosis neuropathic pain among patients with chronic low back pain. Combination therapy including drugs with different mechanisms of action should be given to the patients with chronic low back pain.

Yavuz F, Guzelkucuk U. Diagnosis and pharmacologic management of neuropathic pain among patients with chronic low back pain. *World J Rheumatol* 2014; 4(3): 54-61 Available from: URL: http://www.wjgnet.com/2220-3214/full/v4/i3/54.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.54



INTRODUCTION

Chronic low back pain (LBP) is defined as pain and disability lasting more than 3 mo. In adults, the incidence of chronic LBP is estimated about 6%-15%^[1]. Although there are multiple causes of LBP, about 85% of LBP patients have non-specific LBP. If the cause of LBP is not due to a specific pathology such as infection, tumor, osteoporosis, inflammatory disorders, disc pathologies, then it can be called non-specific LBP. About 10%-15% of the patients with non-specific LBP will go on to develop chronic, disabling LBP^[2,3]. The most common pain generator in chronic LBP is the facet joint (40%), the intervertebral disc (26%) and the sacroiliac joint (2%), respectively^[4].

Chronic LBP consists of both nociceptive and neuropathic mechanisms and can be classified as a mixed pain syndrome^[5,6]. Non-specific nociceptive pain is caused by an inflammatory response to tissue injury and usually described as a sharp or aching pain, while neuropathic pain is caused by damage to nerve tissues and usually described as a burning or heavy sensation, or numbness along the dermatom of the affected nerve^[7,8]. Neuropathic component of chronic LBP has often been underrecognized and under-treated by the physicians. Therefore, recent studies have demonstrated that approximately 20%-55% of chronic LBP patients have neuropathic pain symptoms^[6,9-11]. The presence of a neuropathic pain component is associated with more severe pain^[6], a greater number of comorbidities^[5], reduced quality of life^[12] and higher healthcare utilization costs^[13].

Mechanical and chemical pathophysiological mechanisms are thought to be responsible for neuropathic LBP. Mechanical pathomechanism consist of nerve root compression due to spinal stenosis or intervertebral disc herniation and lesions of nociceptive sprouts within the degenerated intervertebral disc. In chemical pathomechanism, chemokines and cytokines originating from the degenerative disc have been elucidated^[5,14-16]. In addition, the theoretical consideration of nerve roots as the only cause of neuropathic pain in chronic LBP is incorrect. Regarding the pathogenesis of degenerative and painful discs, it was reported that intervertebral discs have nerve ingrowth into the inner layers of the annulus fibrosis; as such, the intervertebral disc itself can be a source of neuropathic pain in patients with chronic LBP^[14]. Some various nerve-damaging mechanisms were shown in generating a neuropathic pain component in patients with chronic LBP^[5]. An altered peripheral, spinal, and supraspinal processing of pain arising as a result of a lesion affecting the nerves system are the major contributor to neuropathic LBP^[17-20].

SCREENING TOOLS FOR NEUROPATHIC PAIN AMONG PATIENTS WITH CHRONIC LBP

Since neuropathic LBP requires specific treatment, fa-

vouring the use of drugs with proven efficacy in the treatment of neuropathic pain such as opioids, tricyclic antidepressants and anticonvulsants^[21], identifying neuropathic pain from nociceptive pain is important. It is assumed that the treatment directed against the specific cause or particular pain mechanisms will induce better treatment response in the patients. Therefore, physicians should consider chronic LBP not only with nociceptive component but also with neuropathic component. The clinical evaluation is still the gold standard for assessment and diagnosis of neuropathic LBP. The diagnostic workup should include neurological and psychosocial evaluation. Although diagnosis can be difficult due to the lack of reliable gold standard diagnostic test for neuropathic LBP, screening tools may help to identify neuropathic pain component in patients with chronic LBP^[5]. An ideal screening tools should be brief, simple, valid, and sensitive. Several screening tools for neuropathic pain such as the Leeds Assessment of Neuropathic Symptoms and Signs $(LANSS)^{[10]}$, the Douleur Neuropathic Symptoms questions $(DN4)^{[22]}$, the ID-Pain^[23], the Neuropathic Pain Questionnaire $(NPQ)^{[24]}$, the Standardized Evaluation of Pain $(StEP)^{[25]}$, and the painDETECT questionnaire (PD-Q)^[11] have been developed and tested with different patient populations. These tools contain some weak and strong features, and they are insufficient to diagnose neuropathic pain in 10%-20% of the patients. However, providing immediate information and their ease of use for both clinicians and patients makes these screening tools attractive^[26].

Among the screening tools, the PD-Q and the StEP are validated in patients with LBP^[11,25]. The PD-Q consists of graduation of pain, pain course pattern and radiating pain. The graduation of pain subscale consists of seven items, and patients are asked to answer each item using a 6-point scale. The PD-Q score is calculated by addition of the each score in the questionnaire, with a maximum score of 38. Scores 19 or greater indicate that neuropathic mechanisms are likely to be involved in the pain; scores between 13 and 18 are uncertain but a neuropathic pain component may be present; scores of ≤ 12 are suggestive of nociceptive pain. Approximately 80% sensitivity and specificity have been found for the PD-Q. The StEP consists of six interview questions and ten physical tests^[25]. The StEP achieves higher sensitivity (92%) and specificity (97%) than the PD-Q which consists only interview questions in distinguishing neuropathic pain from nociceptive pain in patients with LBP. Straight-leg-raising test (Lasegue's sign), a reduced response to cold sensation and a reduced pinprick sensation are the most discriminatory StEP indicators for neuropathic pain^[27].

Although the other screening tools except for the PD-Q and the StEP are also used in some clinical studies for distinguishing neuropathic pain from nociceptive pain in patients with LBP^[10,28-31], none of them has been validated in patients with LBP. The LANSS scale^[10] and the DN4 questionnaire^[22] are another screening tools consist of physical tests such as sensation examination and interview questions. The LANSS comprises a seven-item



pain scale, including the sensory descriptors and items for sensory examination, with a maximum score of 24. Scores less than 12 indicate that neuropathic mechanisms are unlikely to be involved in the pain and scores 12 or greater indicate the opposite. The LANSS demonstrated sensitivity of 83% and specificity of 87% in distinguishing neuropathic pain from nociceptive pain. In the DN4 screening tool, three physical tests were using for determining light touch sensation, pinprick sensation and painful response^[25]. A score of at least 4/10 in this screening tool is indicative of neuropathic pain, with high sensitivity and specificity (82.9% and 89.9%, respectively).

The ID-Pain^[23] and the NPQ^[24] rely only on interview questions, and they don't include physical examinations. The ID-Pain is a six-item screening tool, scores ranged from 1 to 5, with a higher score indicative of pain that contains a neuropathic component. The NPQ consists a 12-item questionnaire form, and this scale demonstrated sensitivity of 66.6% and specificity of 74.4% in distinguishing neuropathic pain from nociceptive pain.

Based on the above mentioned clinical studies, it seems plausible that the StEP may lead to more effective in discriminating between neuropathic and nociceptive pain in patients with LBP according to the higher rate of sensitivity and its validity in patients with LBP. However, the most appropriate approach is still to combine findings on physical and neurologic examinations and patient's report in distinguishing neuropathic pain from nociceptive pain.

BEDSIDE SENSORY TESTS

The Neuropathic Pain Special Interest Group of the International Association for the Study of Pain recommend the sensory bedside examination which consists of touch, pinprick, pressure, cold, heat, and vibration sensations for patients presenting with possible neuropathic pain^[32]. In order to demonstrate sensory abnormalities in patients with chronic LBP, sensory symptoms and signs should be investigated carefully in the affected dermatome. Thus, these tests will help to physicians confirming or denying the presence of neuropathic pain. A piece of cotton wool can be used in touch sensation examination. Thermal sensation can be assessed by warm and cold objects. Vibration sense can be assessed by a 128-Hz tuning fork^[5,33]. The findings in the painful area should be compared with the findings in the non-painful area in contralateral side. The reported responses of the patient are recorded as the same, increased, or decreased, as compared with the normal area. Temporal summation, hypoalgesia to pinprick, allodynia to brush and cold, and hypoesthesia to light touch are discriminant findings for the neuropathic pain. The bedside sensory tests were also found more sensitive than quantitative sensory testing^[34,35]. In order to show a lesion of the somatosensory system suggesting possible neuropathic pain, careful clinical examination should be made. However, there is no gold standard finding to label a specific pain within an area of sensory abnormalities as neuropathic pain.

PHARMACOLOGIC MANAGEMENT OF NEUROPATHIC PAIN AMONG PATIENTS WITH CHRONIC LBP

In the treatment of neuropathic pain among patients with chronic LBP, there are many treatments consist of non-pharmacological and pharmacological. Thus, it is difficult for physicians to decide on convenient treatment method. In recently, some treatment guidelines which suggest a multimodal approach for the treatment of neuropathic pain have been developed^[36-40]. In the present review, we will only focus on pharmacological treatment of neuropathic pain among patients with chronic LBP. First-line medications recommended for neuropathic pain include tricyclic antidepressants, anticonvulsants, and opioid analgesics.

Antidepressants

Tricyclic antidepressants (TCAs) show their analgesic effect via some mechanisms in the central and peripheral nervous systems, including: reuptake inhibition of noradrenaline and serotonin neurotransmission; actions on opioid, adrenergic, serotonin, gamma-aminobutyric acid and N-methyl-D-aspartate receptors; and activation some ion channels^[41]. Their analgesic effects are independent of their antidepressant effect. TCAs are recommended in the NICE guidelines for patients with chronic LBP who showed inadequate treatment response to other drugs^[42]. TCAs have several side-effects such as sedation, dry mouth, blurred vision, and urinary retention. All of these side-effects are often due to their anticholinergic properties. Especially, elderly patients may be more susceptible to some of these effects. Therefore, TCAs should be used carefully in elderly patients^[43]. Data on the efficacy of antidepressants other than TCAs such as serotonin noradrenaline reuptake inhibitors-duloxetine and venlafaxine- and selective serotonin reuptake inhibitors (SSRIs)in chronic LBP are conflicting^[44-48].

In a systematic review by Staiger et al^[49], TCAs were found to produce moderate pain reductions for patients with chronic LBP while SSRIs were not found effective in pain reducing in patients with chronic LBP. In addition, conflicting results were found about the antidepressants whether they improve functional status of patients with chronic LBP. In the Cochrane review of 10 placebo-controlled clinical trials including antidepressants, the authors conclude that "there is no clear evidence that antidepressants are more effective than placebo in the management of patients with chronic LBP"^[50]. In randomized placebo controlled trial involving duloxetine, the reduction in weekly mean pain observed with duloxetine was significantly greater at higher doses (120 mg) than with placebo. However, there were no differences between duloxetine 20 or 60 mg and placebo. Although duloxetine 120 mg showed significant effect on reduc-

WJR www.wjgnet.com

ing pain level, adverse reactions were found significantly higher than placebo^[44]. In contrast to this study, further clinical studies showed that duloxetine 60 mg provided significantly greater pain reductionsthan placebo in patients with chronic LBP^[45,46].

Anticonvulsants

Anticonvulsant agents such as pregabalin and gabapentin show their analgesic effects by binding to the $\alpha 2$ - γ subunit of N-type voltage-gated calcium channels which leads to decreased release of neurotransmitters^[51]. Gabapentin is initiated at 300 mg/d and up titrated in 300-mg increments every 3-7 d according to tolerability, to a target dose of 1800-3600 mg/d in three divided doses. Pregabalin is initiated at 150 mg/d in two divided doses. After 7 d, the dose was elevated to target dose of 300-600 mg/d. The main side effects of these drugs include somnolence, dizziness and peripheral oedema, and caution is advised in patients with renal insufficiency^[37].

To date, there is no systematic reviews of anticonvulsants for chronic LBP. However, there are two clinical trials of gabapentin for chronic LBP^[52,53]. As the result of these studies, gabapentin showed small improvements in pain scores compared with placebo. There was no difference between gabapentin and placebo in according to the rates of adverse reactions. In the treatment of chronic LBP, there is no evidence to support the use of pregabalin. In two randomized trials, there were no significant difference between pregabalin and placebo groups in according to the reduction in weekly mean pain score. In addition, when the patients with treatment-refractory neuropathic pain, including those with chronic LBP had taken pregabalin as a monotherapy, pain relief and improvement in quality of life were found significantly lower than those patients with either oxycodone controlled release (CR) alone or the combination of oxycodone CR and pregabalin^[54,55].

Opioid analgesics

These drugs show their analgesic effect with binding to opioid receptors in the central nervous system, thus they regulate the pathways involved in the generation, transmission, and modulation of pain impulses^[56]. In clinical practice, the most important factors for the restriction of an opioid using are drug tolerability issues and adverse reactions. The most common adverse reactions are dry mouth, nausea, constipation, dizziness, drowsiness, pruritis and vomiting^[57]. In addition, the other concerns about opioids using in chronic non-malignant pain management are development of analgesic tolerance and dependence in susceptible patients. However, the short-term using of opioids is recommended in those patients with nociceptive and neuropathic pain who have unresponsive to firstline treatment and in those patients who have moderate to severe pain^[58]. Due to the absence of high-quality published trials, there are few meta-analyses and systematic reviews investigating opioids in the chronic pain setting.

In higher-quality trial, sustained-release oxymorphone or sustained-release oxycodone was found to be superior than placebo in the treatment of chronic LBP^[59]. In another study conducted by Schnitzer et al^{60]} tramadol was found more effective than placebo for short-term pain relief in the patients with chronic LBP. Two other trials of tramadol found that there were no significant differencesin benefits or harms between sustained-release and immediate-release tramadol for chronic LBP^[61,62]. There is no trial comparing tramadol with acetaminophen or opioid monotherapy, or with other NSAIDs. Another open-label, randomized multicenter study showed that transdermal fentanyl and sustained-release oral morphine provided similar pain relief in patients with chronic LBP^[63]. The meta-analysis investigating the use of opioids in chronic non-cancer pain, including chronic LBP reported that opioids were more effective than placebo for both nociceptive and neuropathic pain^[64]. Controversy exists as to whether opioids are effective for neuropathic component of chronic LBP.

Other drugs

Tapentadol is a centrally acting analgesic used to treat moderate to severe acute pain. This drug show its analgesic effect via acting as µ-opioid receptor agonist and providing noradrenaline re-uptake inhibition^[65]. In phase 3 studies, patients with chronic LBP showed good clinical results to tapentadol prolonged release (PR)^[66,67]. In these studies, tapentadol PR demonstrated similar analgesic efficacy compared with oxycodone CR. Gastrointestinal tolerability and the incidence of drug discontinuations were lower in patients using tapentadol PR than those patients using oxycodone CR^[68,69]. In another phase 3b study, the effectiveness and safety of tapentadol PR vs a combination of tapentadol PR and pregabalin were compared for the management of severe, chronic LBP with a neuropathic component. The authors found that tapentadol PR showed comparable improvements in pain intensity and quality-of-life measures to combination of tapentadol PR and pregabalin, with improved drug tolerability^[69]. Ascorbic acid (Vitamin C) is an anti-oxidant. This means it lowers the amount of free radicals produced from oxidation, like the reactive oxygen species (ROS). ROS are critically involved in the development and maintenance of neuropathic pain. So, free radical scavengers like ascorbic acid could be useful for treatment of neuropathic pain^[70]. However, there is no clinical study investigating tilidine and ascorbic acid in the management of neuropathic pain among patients with chronic LBP.

COMBINATION THERAPY

Since chronic LBP consists of both nociceptive and neuropathic mechanisms, combination therapy such as antidepressants and/or anticonvulsants plus opioids or NSAIIs might be rational in the treatment of chronic LBP^[71]. Treatment guidelines also recommend combination therapy in the treatment of neuropathic pain due to different causes as an option for patients who are unresponsive to the monotherapy^[7,36]. However, combination therapy is associated with some limitations consisting of adverse reactions and drug interactions^[39,72].

In the literature, the number of clinical studies investigating the effect of combination therapy for neuropathic pain component in patients with chronic LBP is very few. Although most of the available clinical studies have investigated combinations of an opioid plus another drugs, there is only one study investigating the efficacy of celecoxib plus pregabalin combination drug therapy in a mixed population of patients including chronic LBP^[/1]. In this study, the authors showed that combination therapy showed significantly greater reductions in LBP, and a similar frequency of adverse reactions, compared with either celecoxib or pregabalin alone. In the literature, there were two studies investigating the benefit of an opioid plus pregabalin. In the first study, the combination of oxycodone CR plus pregabalin was compared with either oxycodone CR or pregabalin alone in 409 patients with treatment-refractory neuropathic pain (most commonly due to radiculopathy). The authors found that LBP relief was faster and more substantial in the patients with combination therapy than in those patients with pregabalin monotherapy. The patients with combination therapy showed significantly greater improvements in quality of life than patients with either oxycodone CR or pregabalin using. Combination therapy also showed a superior safety profile to both monotherapies^[55]. In the second study, the authors investigated the benefit of combination of buprenorphine plus pregabalin in patients with chronic LBP. Pain reduction was found significantly greater in patients with combination therapy than in patients with buprenorphine monotherapy^[73]. There were also 2 studies examining the benefit of tramadol plus paracetamol in a combination therapy for the patients with chronic LBP. In these studies, significantly greater improvements in LBP severity were determined in patients with combination therapy than in patients with placebo. Adverse reactions were found more common with the combination therapy than with placebo^[74,75].

To sum up, combination therapy of pregabalin plus other analgesic drugs such as celecoxib, oxycodone CR and buprenorphine appears to be more effective in reducing neuropathic pain component whereas pregabalin monotherapy seems to be ineffective. Tramadol alone and in combination with paracetamol also appeared to be effective.

CONCLUSION

Presently, there is no available gold standard test for determining a neuropathic pain component in chronic LBP. Neurophysiological testing and screening tools have some limitations in the differentiation of a neuropathic component in chronic LBP patients. So that, bedside sensory tests is the still best available tool for assessment and diagnosis neuropathic pain among patients with chronic LBP. Due to the fact that chronic LBP consists of both nociceptive and neuropathic mechanisms, a multimodal approach to medication probably is more rational in the management of patients with chronic LBP. Therefore, combination therapy including drugs with different mechanisms of action should be given to the patients with chronic LBP. In the literature there is no clear evidence that antidepressants and opioids are effective in the management of neuropathic pain among patients with chronic LBP. In addition, there is no evidence to support the use of anti-convulsant drugs. In order to improve level of evidence in diagnosing and treating neuropathic LBP, further well-designed clinical studies investigating pharmacologic management in neuropathic pain among patients with chronic LBP are needed.

REFERENCES

- 1 Manchikanti L. Epidemiology of low back pain. *Pain Physician* 2000; **3**: 167-192 [PMID: 16906196]
- 2 Deyo RA, Phillips WR. Low back pain. A primary care challenge. *Spine* (Phila Pa 1976) 1996; 21: 2826-2832 [PMID: 9112706]
- 3 Carey TS, Garrett JM, Jackman AM. Beyond the good prognosis. Examination of an inception cohort of patients with chronic low back pain. *Spine* (Phila Pa 1976) 2000; 25: 115-120 [PMID: 10647169]
- 4 Manchikanti L, Singh V, Pampati V, Damron KS, Barnhill RC, Beyer C, Cash KA. Evaluation of the relative contributions of various structures in chronic low back pain. *Pain Physician* 2001; 4: 308-316 [PMID: 16902676]
- 5 Freynhagen R, Baron R. The evaluation of neuropathic components in low back pain. *Curr Pain Headache Rep* 2009; 13: 185-190 [PMID: 19457278 DOI: 10.1007/s11916-009-0032-y]
- 6 Romanò CL, Romanò D, Lacerenza M. Antineuropathic and antinociceptive drugs combination in patients with chronic low back pain: a systematic review. *Pain Res Treat* 2012; 2012: 154781 [PMID: 22619711 DOI: 10.1155/2012/154781]
- 7 Forde G. Adjuvant analgesics for the treatment of neuropathic pain: evaluating efficacy and safety profiles. *J Fam Pract* 2007; 56: 3-12 [PMID: 17270113]
- 8 Woolf CJ. Pain: moving from symptom control toward mechanism-specific pharmacologic management. Ann Intern Med 2004; 140: 441-451 [PMID: 15023710]
- 9 Beith ID, Kemp A, Kenyon J, Prout M, Chestnut TJ. Identifying neuropathic back and leg pain: a cross-sectional study. *Pain* 2011; **152**: 1511-1516 [PMID: 21396774 DOI: 10.1016/ j.pain.2011.02.033]
- 10 Kaki AM, El-Yaski AZ, Youseif E. Identifying neuropathic pain among patients with chronic low-back pain: use of the Leeds Assessment of Neuropathic Symptoms and Signs pain scale. *Reg Anesth Pain Med* 2005; **30**: 422-428 [PMID: 16135345]
- 11 Freynhagen R, Baron R, Gockel U, Tölle TR. painDETECT: a new screening questionnaire to identify neuropathic components in patients with back pain. *Curr Med Res Opin* 2006; 22: 1911-1920 [PMID: 17022849 DOI: 10.1185/030079906X132488]
- 12 O'Connor AB. Neuropathic pain: quality-of-life impact, costs and cost effectiveness of therapy. *Pharmacoeconomics* 2009; 27: 95-112 [PMID: 19254044 DOI: 10.2165/00019053-200 927020-00002]
- 13 Schmidt CO, Schweikert B, Wenig CM, Schmidt U, Gockel U, Freynhagen R, Tölle TR, Baron R, Kohlmann T. Modelling the prevalence and cost of back pain with neuropathic components in the general population. *Eur J Pain* 2009; 13: 1030-1035 [PMID: 19201230 DOI: 10.1016/j.ejpain.2008.12.003]
- 14 Coppes MH, Marani E, Thomeer RT, Groen GJ. Innervation of "painful" lumbar discs. Spine (Phila Pa 1976) 1997; 22:



2342-2349; discussion 2349-2350 [PMID: 9355214]

- 15 Peng B, Hou S, Wu W, Zhang C, Yang Y. The pathogenesis and clinical significance of a high-intensity zone (HIZ) of lumbar intervertebral disc on MR imaging in the patient with discogenic low back pain. *Eur Spine J* 2006; **15**: 583-587 [PMID: 16047210 DOI: 10.1007/s00586-005-0892-8]
- 16 Peng B, Wu W, Hou S, Li P, Zhang C, Yang Y. The pathogenesis of discogenic low back pain. J Bone Joint Surg Br 2005; 87: 62-67 [PMID: 15686239]
- 17 Wu G, Ringkamp M, Murinson BB, Pogatzki EM, Hartke TV, Weerahandi HM, Campbell JN, Griffin JW, Meyer RA. Degeneration of myelinated efferent fibers induces spontaneous activity in uninjured C-fiber afferents. *J Neurosci* 2002; 22: 7746-7753 [PMID: 12196598]
- 18 Amir R, Kocsis JD, Devor M. Multiple interacting sites of ectopic spike electrogenesis in primary sensory neurons. J Neurosci 2005; 25: 2576-2585 [PMID: 15758167 DOI: 10.1523/ JNEUROSCI.4118-04.2005]
- 19 Finnerup NB, Jensen TS. Mechanisms of disease: mechanism-based classification of neuropathic pain-a critical analysis. Nat Clin Pract Neurol 2006; 2: 107-115 [PMID: 16932532]
- 20 Baron R. Mechanisms of disease: neuropathic pain--a clinical perspective. Nat Clin Pract Neurol 2006; 2: 95-106 [PMID: 16932531]
- 21 **Finnerup NB**, Otto M, McQuay HJ, Jensen TS, Sindrup SH. Algorithm for neuropathic pain treatment: an evidence based proposal. *Pain* 2005; **118**: 289-305 [PMID: 16213659]
- 22 Bouhassira D, Attal N, Alchaar H, Boureau F, Brochet B, Bruxelle J, Cunin G, Fermanian J, Ginies P, Grun-Overdyking A, Jafari-Schluep H, Lantéri-Minet M, Laurent B, Mick G, Serrie A, Valade D, Vicaut E. Comparison of pain syndromes associated with nervous or somatic lesions and development of a new neuropathic pain diagnostic questionnaire (DN4). *Pain* 2005; **114**: 29-36 [PMID: 15733628 DOI: 10.1016/ j.pain.2004.12.010]
- 23 Portenoy R. Development and testing of a neuropathic pain screening questionnaire: ID Pain. *Curr Med Res Opin* 2006; 22: 1555-1565 [PMID: 16870080 DOI: 10.1185/030079906X115702]
- 24 **Krause SJ**, Backonja MM. Development of a neuropathic pain questionnaire. *Clin J Pain* 2003; **19**: 306-314 [PMID: 12966256 DOI: 10.1097/00002508-200309000-00004]
- 25 Scholz J, Mannion RJ, Hord DE, Griffin RS, Rawal B, Zheng H, Scoffings D, Phillips A, Guo J, Laing RJ, Abdi S, Decosterd I, Woolf CJ. A novel tool for the assessment of pain: validation in low back pain. *PLoS Med* 2009; 6: e1000047 [PMID: 19360087 DOI: 10.1371/journal.pmed.1000047]
- 26 Baron R, Binder A, Wasner G. Neuropathic pain: diagnosis, pathophysiological mechanisms, and treatment. *Lancet Neurol* 2010; 9: 807-819 [PMID: 20650402 DOI: 10.1016/ S1474-4422(10)70143-5]
- 27 Cruccu G, Truini A. Tools for assessing neuropathic pain. PLoS Med 2009; 6: e1000045 [PMID: 19360134 DOI: 10.1371/ journal.pmed.1000045]
- 28 Enthoven WT, Scheele J, Bierma-Zeinstra SM, Bueving HJ, Bohnen AM, Peul WC, van Tulder MW, Berger MY, Koes BW, Luijsterburg PA. Back complaints in older adults: prevalence of neuropathic pain and its characteristics. *Pain Med* 2013; 14: 1664-1672 [PMID: 24118796 DOI: 10.1111/ pme.12232]
- 29 El Sissi W, Arnaout A, Chaarani MW, Fouad M, El Assuity W, Zalzala M, Dershaby YE, Youseif E. Prevalence of neuropathic pain among patients with chronic low-back pain in the Arabian Gulf Region assessed using the leeds assessment of neuropathic symptoms and signs pain scale. *J Int Med Res* 2010; **38**: 2135-2145 [PMID: 21227019 DOI: 10.1177/14732300 1003800629]
- 30 Attal N, Perrot S, Fermanian J, Bouhassira D. The neuropathic components of chronic low back pain: a prospective multicenter study using the DN4 Questionnaire. *J Pain* 2011; **12**:

1080-1087 [PMID: 21783428 DOI: 10.1016/j.jpain.2011.05.006. Epub]

- 31 Walsh J, Rabey MI, Hall TM. Agreement and correlation between the self-report leeds assessment of neuropathic symptoms and signs and Douleur Neuropathique 4 Questions neuropathic pain screening tools in subjects with low backrelated leg pain. J Manipulative Physiol Ther 2012; 35: 196-202 [PMID: 22397741 DOI: 10.1016/j.jmpt.2012.02.001]
- 32 O'Connor AB, Dworkin RH. Treatment of neuropathic pain: an overview of recent guidelines. *Am J Med* 2009; **122**: S22-S32 [PMID: 19801049 DOI: 10.1016/j.amjmed.2009.04.007]
- 33 Cruccu G, Anand P, Attal N, Garcia-Larrea L, Haanpää M, Jørum E, Serra J, Jensen TS. EFNS guidelines on neuropathic pain assessment. *Eur J Neurol* 2004; **11**: 153-162 [PMID: 15009162 DOI: 10.1111/j.1468-1331.2004.00791.x]
- 34 Haanpää M, Attal N, Backonja M, Baron R, Bennett M, Bouhassira D, Cruccu G, Hansson P, Haythornthwaite JA, Iannetti GD, Jensen TS, Kauppila T, Nurmikko TJ, Rice AS, Rowbotham M, Serra J, Sommer C, Smith BH, Treede RD. NeuPSIG guidelines on neuropathic pain assessment. *Pain* 2011; **152**: 14-27 [PMID: 20851519 DOI: 10.1016/ j.pain.2010.07.031]
- 35 Baron R. Neuropathic pain: clinical, vol 5. In: Basbaum AI, Kaneko A, Shepherd GM, et al (eds). The Senses: a Comprehensive Reference. Amsterdam: Elsevier, 2008: 865-900
- 36 Moulin DE, Clark AJ, Gilron I, Ware MA, Watson CP, Sessle BJ, Coderre T, Morley-Forster PK, Stinson J, Boulanger A, Peng P, Finley GA, Taenzer P, Squire P, Dion D, Cholkan A, Gilani A, Gordon A, Henry J, Jovey R, Lynch M, Mailis-Gagnon A, Panju A, Rollman GB, Velly A. Pharmacological management of chronic neuropathic pain - consensus statement and guidelines from the Canadian Pain Society. *Pain Res Manag* 2007; **12**: 13-21 [PMID: 17372630]
- 37 Dworkin RH, O'Connor AB, Backonja M, Farrar JT, Finnerup NB, Jensen TS, Kalso EA, Loeser JD, Miaskowski C, Nurmikko TJ, Portenoy RK, Rice AS, Stacey BR, Treede RD, Turk DC, Wallace MS. Pharmacologic management of neuropathic pain: evidence-based recommendations. *Pain* 2007; 132: 237-251 [PMID: 17920770]
- 38 Jensen TS, Madsen CS, Finnerup NB. Pharmacology and treatment of neuropathic pains. *Curr Opin Neurol* 2009; 22: 467-474 [PMID: 19741531 DOI: 10.1097/WCO.0b013e3283311e13]
- 39 Dworkin RH, O'Connor AB, Audette J, Baron R, Gourlay GK, Haanpää ML, Kent JL, Krane EJ, Lebel AA, Levy RM, Mackey SC, Mayer J, Miaskowski C, Raja SN, Rice AS, Schmader KE, Stacey B, Stanos S, Treede RD, Turk DC, Walco GA, Wells CD. Recommendations for the pharmacological management of neuropathic pain: an overview and literature update. *Mayo Clin Proc* 2010; 85: S3-S14 [PMID: 20194146 DOI: 10.4065/mcp.2009.0649]
- 40 Attal N, Cruccu G, Baron R, Haanpää M, Hansson P, Jensen TS, Nurmikko T. EFNS guidelines on the pharmacological treatment of neuropathic pain: 2010 revision. *Eur J Neurol* 2010; **17**: 1113-1e88 [PMID: 20402746 DOI: 10.1111/j.1468-1331.2010.02999.x]
- 41 **Verdu B**, Decosterd I, Buclin T, Stiefel F, Berney A. Antidepressants for the treatment of chronic pain. *Drugs* 2008; **68**: 2611-2632 [PMID: 19093703 DOI: 10.2165/0003495-200868180 -00007]
- 42 National Collaborating Centre for Primary Care (UK). Low Back Pain: Early Management of Persistent Non-specific Low Back Pain [Internet]. London: Royal College of General Practitioners (UK), 2009: May. Available from: URL: http:// www.ncbi.nlm.nih.gov/books/NBK11702/
- 43 Amann U, Schmedt N, Garbe E. Prescribing of potentially inappropriate medications for the elderly: an analysis based on the PRISCUS list. *Dtsch Arztebl Int* 2012; **109**: 69-75 [PMID: 22368709 DOI: 10.3238/arztebl.2012.0069]
- 44 Skljarevski V, Ossanna M, Liu-Seifert H, Zhang Q, Chappell

A, Iyengar S, Detke M, Backonja M. A double-blind, randomized trial of duloxetine versus placebo in the management of chronic low back pain. *Eur J Neurol* 2009; **16**: 1041-1048 [PMID: 19469829 DOI: 10.1111/j.1468-1331.2009.02648.x]

- 45 Skljarevski V, Zhang S, Desaiah D, Alaka KJ, Palacios S, Miazgowski T, Patrick K. Duloxetine versus placebo in patients with chronic low back pain: a 12-week, fixed-dose, randomized, double-blind trial. *J Pain* 2010; 11: 1282-1290 [PMID: 20472510 DOI: 10.1016/j.jpain.2010.03.002]
- 46 Skljarevski V, Desaiah D, Liu-Seifert H, Zhang Q, Chappell AS, Detke MJ, Iyengar S, Atkinson JH, Backonja M. Efficacy and safety of duloxetine in patients with chronic low back pain. *Spine* (Phila Pa 1976) 2010; 35: E578-E585 [PMID: 20461028 DOI: 10.1097/BRS.0b013e3181d3cef6]
- 47 Skljarevski V, Zhang S, Chappell AS, Walker DJ, Murray I, Backonja M. Maintenance of effect of duloxetine in patients with chronic low back pain: a 41-week uncontrolled, doseblinded study. *Pain Med* 2010; 11: 648-657 [PMID: 20546509 DOI: 10.1111/j.1526-4637.2010.00836.x]
- 48 Dickens C, Jayson M, Sutton C, Creed F. The relationship between pain and depression in a trial using paroxetine in sufferers of chronic low back pain. *Psychosomatics* 2000; 41: 490-499 [PMID: 11110112 DOI: 10.1176/appi.psy.41.6.490]
- 49 Staiger TO, Gaster B, Sullivan MD, Deyo RA. Systematic review of antidepressants in the treatment of chronic low back pain. *Spine* (Phila Pa 1976) 2003; 28: 2540-2545 [PMID: 14624092]
- 50 Urquhart DM, Hoving JL, Assendelft WW, Roland M, van Tulder MW. Antidepressants for non-specific low back pain. *Cochrane Database Syst Rev* 2008; (1): CD001703 [PMID: 18253994 DOI: 10.1002/14651858.CD001703.pub3]
- 51 Sills GJ. The mechanisms of action of gabapentin and pregabalin. Curr Opin Pharmacol 2006; 6: 108-113 [PMID: 16376147]
- 52 McCleane GJ. Does gabapentin have an analgesic effect on background, movement and referred pain? A randomised, double-blind, placebo controlled study. *Pain Clinic* 2001; 13: 103-107 [DOI: 10.1163/156856901753420945]
- 53 Yildirim K, Sisecioglu M, Karatay S, Erdal A, Levent A, Ugur M. The effectiveness of gabapentin in patients with chronic radiculopathy. *The Pain Clinic* 2003; 15: 213-218 [DOI: 10.1163/156856903767650718]
- 54 Remmers AE, Sharma U, Lamoreaux L. Pregabalin treatment of patients with chronic low back pain. Abstract 660. Proceedings of the 19th Annual Meeting of the American Pain Society 2000. Atlanta, Georgia, 2000
- 55 Gatti A, Sabato AF, Occhioni R, Colini Baldeschi G, Reale C. Controlled-release oxycodone and pregabalin in the treatment of neuropathic pain: results of a multicenter Italian study. *Eur Neurol* 2009; 61: 129-137 [PMID: 19092248 DOI: 10.1159/000186502]
- 56 Grabois M. Management of chronic low back pain. Am J Phys Med Rehabil 2005; 84: S29-S41 [PMID: 15722781]
- 57 Moore RA, McQuay HJ. Prevalence of opioid adverse events in chronic non-malignant pain: systematic review of randomised trials of oral opioids. *Arthritis Res Ther* 2005; 7: R1046-R1051 [PMID: 16207320]
- 58 Deshpande A, Furlan A, Mailis-Gagnon A, Atlas S, Turk D. Opioids for chronic low-back pain. *Cochrane Database Syst Rev* 2007; (3): CD004959 [PMID: 17636781]
- 59 Hale ME, Dvergsten C, Gimbel J. Efficacy and safety of oxymorphone extended release in chronic low back pain: results of a randomized, double-blind, placebo- and active-controlled phase III study. J Pain 2005; 6: 21-28 [PMID: 15629415 DOI: 10.1016/j.jpain.2004.09.005]
- 60 Schnitzer TJ, Gray WL, Paster RZ, Kamin M. Efficacy of tramadol in treatment of chronic low back pain. J Rheumatol 2000; 27: 772-778 [PMID: 10743823]
- 61 Raber M, Hofmann S, Junge K, Momberger H, Kuhn D. Analgesic efficacy and tolerability of tramadol 100mg sus-

tained-release capsules in patients with moderate to severe low back pain. *Clin Drug Investig* 1999; **17**: 415-423 [DOI: 10.2165/00044011-199917060-00001]

- 62 **Sorge J**, Stadler T. Comparison of the analgesic efficacy and tolerability of tramadol 100mg sustained-release tablets and tramadol 50mg capsules for the treatment of chronic low back pain. *Clin Drug Investig* 1997; **14**: 157-164 [DOI: 10.2165 /00044011-199714030-00001]
- 63 **Allan L**, Richarz U, Simpson K, Slappendel R. Transdermal fentanyl versus sustained release oral morphine in strongopioid naïve patients with chronic low back pain. *Spine* (Phila Pa 1976) 2005; **30**: 2484-2490 [PMID: 16284584]
- 64 Furlan AD, Sandoval JA, Mailis-Gagnon A, Tunks E. Opioids for chronic noncancer pain: a meta-analysis of effectiveness and side effects. *CMAJ* 2006; **174**: 1589-1594 [PMID: 16717269 DOI: 10.1503/cmaj.051528]
- 65 Tzschentke TM, Jahnel U, Kogel B, Christoph T, Englberger W, De Vry J, Schiene K, Okamoto A, Upmalis D, Weber H, Lange C, Stegmann JU, Kleinert R. Tapentadol hydrochloride: a next-generation, centrally acting analgesic with two mechanisms of action in a single molecule. *Drugs Today* (Barc) 2009; **45**: 483-496 [PMID: 19834626 DOI: 10.1358/dot.2009.45.7.1395291]
- 66 Buynak R, Shapiro DY, Okamoto A, Van Hove I, Rauschkolb C, Steup A, Lange B, Lange C, Etropolski M. Efficacy and safety of tapentadol extended release for the management of chronic low back pain: results of a prospective, randomized, double-blind, placebo- and active-controlled Phase III study. *Expert Opin Pharmacother* 2010; **11**: 1787-1804 [PMID: 20578811 DOI: 10.1517/14656566.2010.497720]
- 67 Steigerwald I, Müller M, Davies A, Samper D, Sabatowski R, Baron R, Rozenberg S, Szczepanska-Szerej A, Gatti A, Kress HG. Effectiveness and safety of tapentadol prolonged release for severe, chronic low back pain with or without a neuropathic pain component: results of an open-label, phase 3b study. *Curr Med Res Opin* 2012; 28: 911-936 [PMID: 22443293 DOI: 10.1185/03007995.2012.679254]
- 68 Lange B, Kuperwasser B, Okamoto A, Steup A, Häufel T, Ashworth J, Etropolski M. Efficacy and safety of tapentadol prolonged release for chronic osteoarthritis pain and low back pain. *Adv Ther* 2010; 27: 381-399 [PMID: 20556560 DOI: 10.1007/s12325-010-0036-3]
- 69 Baron R, Martin-Mola E, Müller M, Dubois C, Falke D, Steigerwald I. Effectiveness and Safety of Tapentadol Prolonged Release (PR) Versus a Combination of Tapentadol PR and Pregabalin for the Management of Severe, Chronic Low Back Pain With a Neuropathic Component: A Randomized, Double-blind, Phase 3b Study. *Pain Pract* 2014 Apr 17; Epub ahead of print] [PMID: 24738609 DOI: 10.1111/papr.12200]
- 70 Kim HK, Park SK, Zhou JL, Taglialatela G, Chung K, Coggeshall RE, Chung JM. Reactive oxygen species (ROS) play an important role in a rat model of neuropathic pain. *Pain* 2004; **111**: 116-124 [PMID: 15327815 DOI: 10.1016/ j.pain.2004.06.008]
- 71 Romanò CL, Romanò D, Bonora C, Mineo G. Pregabalin, celecoxib, and their combination for treatment of chronic low-back pain. J Orthop Traumatol 2009; 10: 185-191 [PMID: 19921480 DOI: 10.1007/s10195-009-0077-z]
- 72 Chaparro LE, Wiffen PJ, Moore RA, Gilron I. Combination pharmacotherapy for the treatment of neuropathic pain in adults. *Cochrane Database Syst Rev* 2012; 7: CD008943 [PMID: 22786518 DOI: 10.1002/14651858.CD008943.pub2]
- 73 Pota V, Maisto M, Pace MC. Association of buprenorphine TDS and pregabalin in the treatment of low back pain. *Eur J Pain* 2007; 11: S83 [DOI: 10.1016/j.ejpain.2007.03.206]
- 74 Ruoff GE, Rosenthal N, Jordan D, Karim R, Kamin M. Tramadol/acetaminophen combination tablets for the treatment of chronic lower back pain: a multicenter, randomized, double-blind, placebo-controlled outpatient study. *Clin*

Ther 2003; **25**: 1123-1141 [PMID: 12809961 DOI: 10.1016/ S0149-2918(03)80071-1]

75 **Peloso PM**, Fortin L, Beaulieu A, Kamin M, Rosenthal N. Analgesic efficacy and safety of tramadol/ acetaminophen combination tablets (Ultracet) in treatment of chronic low back pain: a multicenter, outpatient, randomized, double blind, placebo controlled trial. *J Rheumatol* 2004; **31**: 2454-2463 [PMID: 15570651]

P- Reviewer: Beales DJ, Schencking M, Tangtrakulwanich B S- Editor: Ji FF L- Editor: A E- Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.62 World J Rheumatol 2014 November 12; 4(3): 62-71 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

MINIREVIEWS

Gout: A clinical overview and its association with cardiovascular diseases

Laura BE Kienhorst, Hein JEM Janssens, Matthijs Janssen

Laura BE Kienhorst, Matthijs Janssen, Department of Rheumatology, Rijnstate Hospital, 6880 TA Arnhem, The Netherlands Hein JEM Janssens, Department of Primary and Community Care, Radboud University Medical Centre, 6525 GA Nijmegen, The Netherlands

Hein JEM Janssens, Department of Clinical Research, Rijnstate Hospital, 6880 TA Arnhem, The Netherlands

Author contributions: All authors contributed to this paper. Correspondence to: Laura BE Kienhorst, MD, LLM, Department of Rheumatology, Rijnstate Hospital, PO Box 9555, 6880

TA Arnhem, The Netherlands. lkienhorst@rijnstate.nl

Telephone: +31-88-0055400 Fax: +31-88-0056612

Received: June 29, 2014 Revised: August 8, 2014 Accepted: September 4, 2014

Published online: November 12, 2014

Abstract

Gout is a common disease caused by the deposition of monosodium urate (MSU) crystals in patients with hyperuricemia, and characterized by very painful recurrent acute attacks of arthritis. The gold standard for diagnosing gout is the identification of MSU crystals in synovial fluid by polarization light microscopy. Arthritis attacks can be treated with anti-inflammatory medications, such as non-steroidal anti-inflammatory drugs, colchicine, oral prednisone, or intra-articular or intramuscular glucocorticoids. To prevent gout uric acid lowering therapy with for example allopurinol can be prescribed. When gout is adequately treated, the prognosis is good. Unfortunately, the management of gout patients is often insufficient. Gout is associated with dietary factors, the use of diuretics, and several genetic factors. Comorbidities as hypertension, chronic kidney disease, cardiovascular diseases, the metabolic syndrome, diabetes, obesity, hyperlipidemia, and early menopause are associated with a higher prevalence of gout. Xanthine oxidase and chronic systemic inflammation seem to play an important role in the pathophysiology of the association between gout and cardiovascular diseases. To prevent cardiovascular diseases gout patients must be early screened for cardiovascular risk factors.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Gout; Review; Clinical; Cardiovascular diseases

Core tip: Gout is a common disease caused by the deposition of monosodium urate (MSU) crystals in patients with hyperuricemia, and characterized by very painful recurrent acute attacks of arthritis. The gold standard for diagnosing gout is the identification of MSU crystals in synovial fluid. Arthritis attacks are treated with antiinflammatory medications, to prevent gout uric acid lowering therapy can be prescribed. When gout is adequately treated, the prognosis is good. Comorbidities as chronic kidney disease, cardiovascular diseases, and the metabolic syndrome are associated with gout. Gout patients must be early screened for cardiovascular risk factors.

Kienhorst LBE, Janssens HJEM, Janssen M. Gout: A clinical overview and its association with cardiovascular diseases. *World J Rheumatol* 2014; 4(3): 62-71 Available from: URL: http://www.wjgnet.com/2220-3214/full/v4/i3/62.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.62

INTRODUCTION

Gout is a common disease caused by the deposition of monosodium urate (MSU) crystals in patients with hyperuricemia, and characterized by very painful recurrent acute attacks of arthritis. Gout has been recognized as a clinical entity for a long period of time. Acute gout occurring in the first metatarsophalangeal (MTP-1) joint, first identified by the Egyptians in 2640 BC, was later recognized by Hippocrates in the 5th century BC, who



referred to it as "the unwalkable disease"^[1]. The first person to use the word gout (*gutta quam podagram vel artiticam vocant* - the gout that is called podagra or arthritis) was the Dominican monk Randolphus of Bocking, domestic chaplain to the Bishop of Chichester (1197-1258)^[2]. Through the ages gout was known as "the king of diseases and the disease of kings", because of its association with alcohol consumption, purine-rich diet and obesity^[3].

This review starts with a clinical overview on the epidemiology, pathogenesis, clinical presentation, risk factors, diagnosis, treatment, and prognosis of gout. Hereafter, the review discusses the association between gout and cardiovascular diseases.

CLINICAL OVERVIEW OF GOUT

Epidemiology

Gout is one of the most common rheumatic diseases with a prevalence of 1%-2% in the adult population in developed countries^[4]. The prevalence of gout is higher in men, and rises with age^[5]. Gout occurs four to ten times more often in men than in women among patients under the age of 65^[5]. In the elderly, gout has a somewhat more equal sex distribution^[5], possibly due to the fall of uricosuric estrogen in women after the menopause^[6,7]. Accumulating evidence suggests an increase in the prevalence of gout in the last decades, which might be caused by an increased longevity and an increased prevalence of factors that promote hyperuricemia such as obesity, the metabolic syndrome, chronic kidney disease, and dietary changes^[5,8-11].

Pathogenesis

Gout is caused by a disorder of the purine metabolism and results from MSU crystal deposition in and around the joints which is associated with hyperuricemia. The serum uric acid concentration is determined by the endogenous production of uric acid by synthesis and cell turnover, the exogenous supply via dietary intake, and renal (two-third) and intestinal (one-third) excretion^[12]. Hyperuricemia is the result of uric acid overproduction, uric acid underexcretion, or a combination of the two^[12]. Hyperuricemia is defined as a serum uric acid concentration that exceeds the solubility at physiologic temperature and pH $(0.38-0.40 \text{ mmol/L})^{[3,13]}$. Although hyperuricemia is necessary to develop gout, it is not sufficient to cause gout. Only one cohort study from 1987 investigated the association between the level of serum uric acid and the cumulative incidence of gout. Gout occurred in just 22% of the patients with a baseline serum uric acid of more than 0.54 mmol/L over a 5-year period^[14].

Necessary for the occurrence of gout arthritis is the formation of MSU crystals when hyperuricemia is present. The formation of MSU crystals depends on the solubility of uric acid in joint fluid. The solubility is influenced by factors such as temperature, pH, level of articular dehydration, concentration of cations, and the presence of nucleating agents (collagen, chondroitin sulfate, and nonaggregating proteoglycans)^[12]. Variation in these factors might explain partly the preference of gout attacks in the MTP-1 joint (the relatively low temperature of this peripheral joint)^[15] and in osteoarthritic joints (degeneration with decreased collagen and proteoglycans)^[16], and the nocturnal onset of the attack (articular dehydration)^[12,13]. However, these factors do not explain for example why gout does rarely occur in the MTP-5 joint which has probably a lower temperature than the MTP-1 joint, and gout is also rarely seen in often osteoarthritic hip joints.

MSU crystal formation leads to MSU crystal deposition in synovial fluids. MSU crystals are pro-inflammatory stimuli. MSU crystals are phagocytosed as particles by monocytes and cause an inflammatory response with the release of pro-inflammatory mediators as tumor necrosis factor (TNF)- α , interleukin (IL)-1b, and IL-6^[12,13]. Mechanisms by which MSU crystals activate cells in the joint and the role of these pro-inflammatory mediators are not yet fully explained. The generally accepted hypothesis is that MSU crystals activate monocytes *via* the inflammasome leading to IL-1b production^[17-21]. IL-1b can induce recruitment of other inflammatory cells within the joint to produce cytokines and chemotactic factors. This results in neutrophil influx to the joint, which is the hallmark of gouty arthritis.

Clinical presentation

Typically, a patient with a gout attack has an acute painful and swollen joint, which is often red and warm. The onset of the arthritis is abrupt. A gout attack usually affects one joint in the lower limbs. Most often, in 57% of the primary care patients^[22], the MTP-1 joint was involved^[23]. In 86% of primary care patients with gouty arthritis the lower leg was affected^[22]. Next most frequent locations are the mid-foot, the ankle and the knee^[16]. Gout attacks are self-limiting and resolve within 7-10 d. However, the arthritis attacks are often recurrent.

Recurrent gout attacks can lead to permanent joint damage and tophi depositions. Tophi can be found in or close to joints, in bursas, tendon sheaths, and in articular cartilage^[24]. Clinical experience shows that in some patients later in the course of the disease the gout attacks can occur more often, and it takes more days before the attack is resolved. Then the arthritis is more frequently polyarticular and spreads to the upper limbs^[16,23].

Risk factors

Many factors have been described as risk factors for the development of gout. However, the associations between these "risk" factors and gout are almost exclusively based on epidemiological studies, which of course cannot proof causal relations between these factors and gout. Epidemiological studies show that several dietary factors might increase the risk of gout, such as alcohol consumption^[25,26], purine-rich meat and seafood intake^[26,29]. The consumption of fructose-sweetened soft drinks^[26,29]. The consumption of dairy products^[28], skim milk powder^[30], folate, vegetables, and coffee are associated with a decreased prevalence of gout^[26]. According to epidemio-

logical studies the use of thiazide and loop diuretics, but not aldosteron antagonists, are associated with the risk of gout^[26,31,32]. However, these results might be confounded by cardiovascular indications^[33].

Other factors can cause the development of gout. Genetics (sex^[5], some genes such as *SLC2A9*, *ABCG2*, *SL-C17A3*, and *SLC22A12*^[3,13] and Asian descent^[34,35]), age^[5], and constitutional influences (body composition) are risk factors, and these cannot be influenced. Comorbidities as hypertension^[36], chronic kidney disease, cardiovascular diseases^[37-41], the metabolic syndrome^[36,42], diabetes^[42,43], obesity^[36], hyperlipidemia^[36], and early menopause^[6] are associated with a higher prevalence of gout^[26]. Nowadays, especially the association between gout and cardiovascular diseases is a large research field^[37-41,44,45], but the exact mechanism of why these diseases are associated is not fully understood.

Diagnosis

The gold standard for diagnosing gout is the identification of MSU crystals in synovial fluid or in a tophus by polarization light microscopy^[46]. The accuracy of the gold standard has been tested in only a few studies. The sensitivity of detection of MSU crystals was shown to be 69% with a specificity of 97%^[47]. After training the sensitivity of the detection of MSU crystals can become 95% with a specificity of 97%^[48]. In clinical practice most synovial fluid is aspirated from the affected joint during a gout attack. A longstanding opinion is that synovial fluid should be analyzed with a polarization microscope rapidly after aspiration, because the formation and solubility of MSU crystals might be affected by pH and temperature^[49]. A recent systematic review has shown that MSU crystals can also be detected in synovial fluid which has been stored for a maximum of 8 wk^[49].

Although there is a gold standard, in primary care synovial fluid analysis is often not possible or not available. Polarization light microscopes are expensive and almost only available at rheumatology departments. But approximately 90% of the gout patients are diagnosed and treated by primary care physicians^[50]. Primary care physicians diagnose gout without the gold standard, based on clinical signs and symptoms, which has demonstrated to have a limited predictive value^[22]. In patients with MTP-1 arthritis the diagnosis gout was right in only 77%, while primary care physicians supposed gout to be the diagnosis in 98% of the patients^[51]. Even in rheumatology departments the gold standard is not always used for diagnosing gout^[52,53].

Several criteria sets were developed to improve the validity of the clinical diagnosis, such as the American College of Rheumatology criteria^[54], which showed a limited sensitivity (90%; 79%) and specificity (64%; 70%) in primary^[55] and secondary care^[56] in MSU crystal-proven gout patients, respectively. A diagnostic rule to diagnose gout without joint fluid analysis developed in a primary care population of MSU crystal-proven gout patients had better results^[22], but the validity of this rule is unknown in secondary care.

Nowadays imaging techniques are increasingly used for diagnosing gout patients. Ultrasonography and dualenergy computed tomography (DECT) are promising but expensive methods for diagnosis and monitoring gout, but with yet an unknown validity in medical practice^[57-62]. Compared to the gold standard of synovial fluid aspiration the main advantage of these techniques is that they are non-invasive. A disadvantage of DECT is its high exposure to radiation.

Treatment

Standard treatment consists of anti-inflammatory drugs for gout attacks, sometimes followed by long-term preventive urate lowering therapy. Acute gout attacks are treated with non-steroidal anti-inflammatory drugs (NSAIDs), colchicine, oral prednisone, or intra-articular or intramuscular glucocorticoids. Unfortunately, there are only a few trials which compare the efficacy and safety of these therapeutics in gout patients. NSAIDs and prednisone have a comparable therapeutic effect during a gout attack^[63-65]. Low-dose colchicine has the same therapeutic effect after 24 h as high-dose colchicine, but with less adverse effects^[66]. Because of the lack of trials, the choice which anti-inflammatory therapy is prescribed is mainly based on comorbidity and comedications. In case of no restrictions by comorbidity and comedications, the cost of the therapy can be taken into account. In the United States, the cost of colchicine have risen after the rebranding of this therapeutic. IL-1b blockers such as canakinumab $^{\rm [67]}$, anakinra $^{\rm [68-70]}$, and rilonacept $^{\rm [71-75]}$ are new therapeutic opportunities in patients with gouty arthritis, but their efficacy and safety should be further tested. However, these new therapeutics are expensive, and should only be prescribed in patients with frequent gout attacks who failed or have contra-indications for the traditional anti-inflammatory drugs. Trials concerning the efficacy of non-pharmacological interventions for gout attacks are even more rare, probably due to ethical and practical difficulties to set up these type of trials $^{[6]}$. Only one trial was performed which shows that local ice therapy can be useful during gout attacks^[77].

Preventive uric acid lowering therapy is indicated in patients with two or more gout attacks per year, tophaceous gout, or a history of uric acid urolithiasis^[78,79]. The decision whether to start preventive uric acid lowering therapy or to accept frequent gout attacks and/or tophi should always be made in accordance with the patient. The aim of the uric acid lowering therapy is to decrease the frequency of gout attacks^[80,81] and/or to reduce tophi^[82] by sufficiently reducing the serum uric acid level. The target serum uric acid level should be at least below 0.36 mmol/L^[79]. A lower target serum uric acid of 0.30 mmol/L can be aimed for in case of severe gout (for instance tophaceous gout)^[78,83]. Based on evidence and experience the first choice uric acid lowering agent is the xanthine oxidase inhibitor allopurinol^[78,79]. The uricosuricum benzbromarone 100-200 mg per day and allopurinol 300-600 mg per day have comparable efficacy and safety profiles^[82,84]. Probenecid, an old uricosuricum, has moderate efficacy

as uric acid lowering therapeutic in patients with lack of effectiveness of or intolerance to allopurinol^[85]. A trial has shown similar effects of allopurinol 200-300 mg per day and febuxostat, a new xanthine oxidase inhibitor, 80-120 mg per day^[86], but in clinical practice the dose of allopurinol can be further enhanced until 600 mg. At this moment, because of high costs and little clinical experience, febuxostat should only be used when the target serum uric acid level cannot be reached by an appropriate dose of allopurinol, or when the patient is intolerant to allopurinol. In both cases benzbromarone is also a good and less expensive alternative. The uricase derivative rasburicase is now only registered for tumor lysis syndrome, but might be beneficial as uric acid lowering therapy in gout patients^[87-90]. A new uricase derivative pegloticase is proven to be useful in patients who are refractory to or intolerant for conventional therapy^[91-95]. Uricases should be administered intravenously with a risk of infusion reactions, and there always remains a risk for antibody formation due to the conjugation to proteins. The latter might impede the efficacy of uricases. The selective uric acid reabsorption inhibitor lesinurad might be another future treatment option^[96].

Only a few studies have compared the efficacy and safety of uric acid lowering monotherapy, and solely one study looked at combination therapy of two uric acid lowering therapeutics. The combination of lesinurad and febuxostat was well tolerated, and the target serum uric acid level was achieved in all patients^[97]. Based on clinical experience benzbromarone can be added to allopurinol when the target serum uric acid cannot be reached by allopurinol monotherapy. The dose of the uric acid lowering medications should be carefully increased to reduce adverse effects, and should be titrated based on serum uric acid levels^[78,79]. It is generally accepted that uric acid lowering therapy should be started under several months of prophylactic anti-inflammatory medications (colchicine or NSAIDs) to prevent paradoxal gout attacks at the start, although there are no studies to prove this^[78,79]. The uric acid lowering therapy should be continued lifelong.

In addition to the uric acid lowering therapy some other pharmacological measures can be helpful to reduce serum uric acid. When a gout patient is also diagnosed with hypertension, losartan could be considered as an-tihypertensive treatment, because of its small uric acid lowering effect^[98]. Vitamin C, a safe supplement, might have a very small uric acid lowering effect^[99,100], although a small randomized controlled trial in gout patients could not confirm this^[101].

Additional non-pharmacological measures, like dietary advices, to reduce serum uric acid may be useful, but their uric acid lowering effects are small (10%-18%) and therapeutically insufficient (*i.e.*, no reduce of the frequency of gout flares) in most patients^[26]. Observational studies showed that the intake of purine-rich meat and seafood, fructose-rich soft drinks, and alcohol should be reduced, and dairy intake and the consumption of vegetables should be encouraged^[26,78,79,102]. Trials concerning the efficacy of non-pharmacological interventions to lower serum uric acid are also lacking. The only trial of dietary intervention in gout patients suggested that skim milk powder enriched with glycomacropeptide and G600 milk fat extract might reduce the frequency of gout flares^[30].

Prognosis

Gout is a potentially curable disease. Unfortunately, the management of gout patients is often insufficient^[5,103-107]. An important reason is the limited use of uric acid lowering therapy. Only 30%-60% of the patients are still prescribed allopurinol one year after the start of the therapy^[4], and only 17% of the gout patients might be fully adherent to allopurinol therapy^[108]. The poor adherence is often, unfairly, blamed on gout patients unwilling to take uric acid lowering therapy. Lack of appropriate information from their doctor is an important factor which plays a role in the poor adherence. An observational study showed that patient education, individual lifestyle advice and slow upward titration of uric acid lowering therapy according to serum uric acid levels can improve the adherence to uric acid lowering therapy^[109].

Acute gout attacks and the presence of tophi account for a major component of the reported decreased healthrelated quality of life in gout patients, and are associated with decreased work productivity which leads to an economic burden for the society^[110-112]. This emphasizes the importance of the effective management of gout. Urate lowering therapy is cost-effective when patients have two or more recurrent attacks per year^[113].

THE ASSOCIATION BETWEEN GOUT AND CARDIOVASCULAR DISEASES

Nowadays, an important study field within gout research is the association between gout and cardiovascular diseases. The increasing interest in this association is probably due to its great clinical importance, because of the high prevalence of gout and cardiovascular diseases. This part of the review elaborates more on the association between gout and cardiovascular diseases.

The association of gout with cardiovascular diseases

Most studies looked at the association between hyperuricemia and cardiovascular diseases. Two systematic reviews of prospective cohort studies show that, after correction for traditional risk factors for cardiovascular diseases, patients with hyperuricemia have a significant higher risk for cardiac diseases^[45], cardiac mortality^[38], stroke^[37], and stroke-related mortality^[37]. The mean association of the risk for cardiac mortality was 12% per increase of the serum uric acid of 0.059 mmol/L^[38]. In women there was a stronger association between hyperuricemia and cardiovascular diseases and mortality than in men^[37,38]. Higher levels of hyperuricemia are stronger risk factors for cardiovascular diseases and mortality than lower levels of hyperuricemia^[114]. Interestingly, several studies observed a J-curve relationship between serum uric acid level and cardiovascular disease or all-cause mortality^[115,116]. A low serum uric acid level might be associated with a higher mortality, because uric acid can play a protective antioxidant role^[117]. It should be noticed that the definition of hyperuricemia differed between several studies and it was not always corrected for sex. Also, patients with hyperuricemia could be symptomatic (*i.e.*, gout) or asymptomatic. However, it is likely that the conclusions from studies about patients with hyperuricemia are also valid in patients with gout.

Some studies investigated the association between gout and cardiovascular diseases. Gout was shown to be associated with an increased risk for heart failure^[39] and myocardial infarction^[45]. Several prospective cohort studies showed that gout was also associated with cardiovascular mortality^[40,41,44,118] and with overall mortality^[39-41,44,118]. Gout is a stronger risk factor for cardiovascular diseases and mortality than hyperuricemia^[40,41]. Tophaceous gout was a very strong risk factor for cardiovascular mortality^[114]. Unfortunately, the diagnosis of gout was often not based on identification of MSU crystals, but on selfreport. In MSU crystal-proven gout the association might be stronger than in gout otherwise diagnosed, and therefore the association of gout and cardiovascular diseases can be underestimated.

The pathophysiology of the association of gout with cardiovascular diseases

The pathophysiological pathways that link gout with cardiovascular diseases are not fully clear. Gout might lead to cardiovascular diseases through endothelial dysfunction caused by oxidative stress through xanthine oxidase activation. Another pathway is based on chronic systemic inflammation in patients with gout, also in asymptomatic periods, which might lead to cardiovascular diseases. Both pathways are now discussed in more detail.

Accumulating evidence shows that xanthine oxidase plays a central role in the association of hyperuricemia and gout with cardiovascular diseases. Upregulation of xanthine oxidase activity rather than decreased renal excretion of uric acid is an important factor underlying the increased serum uric acid levels in heart failure patients^[119]. Endothelial dysfunction might be caused by accelerated inactivation of nitric oxide by reactive oxygen species, and xanthine oxidase is a source of reactive oxygen species production^[120].

Several studies suggest that allopurinol, a xanthine oxidase inhibitor, has cardioprotective effects. Most studies looked at indicators for higher cardiovascular risk. Allopurinol improved the endothelial function^[121] and resulted in an improved vasodilated capacity and peripheral blood flow in patients with heart failure^[122]. Allopurinol gave a significant blood pressure reduction in patients with hyperuricemia^[123-126]. In patients with chronic stable angina allopurinol increased the time to chest pain and the total exercise time^[127,128]. Allopurinol inhibits the oxidation of low-density lipoprotein, which plays an important role in the development of atherosclerosis^[129]. These mechanisms might contribute to a favorable effect

of allopurinol on the cardiovascular risk in gout patients. The effect of allopurinol on mortality was the topic of several studies. These studies showed that allopurinol reduced the mortality in heart failure patients^[130-133]. One recent study investigated the effect of allopurinol on cardiovascular outcome. Allopurinol was associated with a reduced risk of myocardial infarction^[134]. On contrary, benzbromarone, an uricosuricum, did not have beneficial cardioprotective effects^[129,135].

A different pathway which might link gout to cardiovascular diseases is based on chronic systemic inflammation. Low-grade chronic systemic inflammation can contribute to the development of cardiovascular diseases. Some evidence is found that in patients with hyperuricemia or gout low-grade chronic systemic inflammation is present. Serum uric acid levels were associated with C-reactive protein levels, TNF- α levels and IL-6 levels^[136]. Typical gout signs seen with ultrasonography are present in asymptomatic joints of patients with hyperuricemia or gout^[137]. This might imply that also in between gout attacks low-grade inflammation is present. Also in tophaceous gout, a severe form of gout with widespread urate deposition, more low-grade inflammation might be present compared to non-tophaceous gout. Tophaceous gout was shown to be stronger risk factor for cardiovascular diseases and mortality than non-tophaceous gout^[114].

CONCLUSION

Gout is no longer 'the king of diseases and the disease of kings', but a very common disease which is associated with cardiovascular diseases. Not only the gout attacks should be treated, but gout patients should also be screened and treated for cardiovascular risk factors.

REFERENCES

- Nuki G, Simkin PA. A concise history of gout and hyperuricemia and their treatment. *Arthritis Res Ther* 2006; 8 Suppl 1: S1 [PMID: 16820040]
- 2 Copeman WSC. A short history of gout and the rheumatic disease. Med Hist 1964; 8: 394–395
- 3 **Richette P**, Bardin T. Gout. *Lancet* 2010; **375**: 318-328 [PMID: 19692116 DOI: 10.1016/S0140-6736(09)60883-7]
- 4 Annemans L, Spaepen E, Gaskin M, Bonnemaire M, Malier V, Gilbert T, Nuki G. Gout in the UK and Germany: prevalence, comorbidities and management in general practice 2000-2005. *Ann Rheum Dis* 2008; 67: 960-966 [PMID: 17981913 DOI: 10.1136/ard.2007.076232]
- 5 Kuo CF, Grainge MJ, Mallen C, Zhang W, Doherty M. Rising burden of gout in the UK but continuing suboptimal management: a nationwide population study. *Ann Rheum Dis* 2014 Jan 15; Epub ahead of print [PMID: 24431399 DOI: 10.1136/annrheumdis-2013-204463]
- 6 Hak AE, Curhan GC, Grodstein F, Choi HK. Menopause, postmenopausal hormone use and risk of incident gout. *Ann Rheum Dis* 2010; 69: 1305-1309 [PMID: 19592386 DOI: 10.1136/ard.2009.109884]
- 7 Puig JG, Michán AD, Jiménez ML, Pérez de Ayala C, Mateos FA, Capitán CF, de Miguel E, Gijón JB. Female gout. Clinical spectrum and uric acid metabolism. *Arch Intern Med* 1991; 151: 726-732 [PMID: 2012455 DOI: 10.1001/archinte.1991.004 00040074016]

- 8 Roddy E, Doherty M. Epidemiology of gout. Arthritis Res Ther 2010; 12: 223 [PMID: 21205285 DOI: 10.1186/ar3199]
- 9 Saag KG, Choi H. Epidemiology, risk factors, and lifestyle modifications for gout. *Arthritis Res Ther* 2006; 8 Suppl 1: S2 [PMID: 16820041 DOI: 10.1186/ar1907]
- 10 Wallace KL, Riedel AA, Joseph-Ridge N, Wortmann R. Increasing prevalence of gout and hyperuricemia over 10 years among older adults in a managed care population. *J Rheumatol* 2004; **31**: 1582-1587 [PMID: 15290739]
- 11 Bhole V, de Vera M, Rahman MM, Krishnan E, Choi H. Epidemiology of gout in women: Fifty-two-year followup of a prospective cohort. *Arthritis Rheum* 2010; 62: 1069-1076 [PMID: 20131266 DOI: 10.1002/art.27338]
- 12 Choi HK, Mount DB, Reginato AM. Pathogenesis of gout. Ann Intern Med 2005; 143: 499-516 [PMID: 16204163 DOI: 10.7326/0003-4819-143-7-200510040-00009]
- 13 **Neogi T**. Clinical practice. Gout. *N Engl J Med* 2011; **364**: 443-452 [PMID: 21288096 DOI: 10.1056/NEJMcp1001124]
- 14 Campion EW, Glynn RJ, DeLabry LO. Asymptomatic hyperuricemia. Risks and consequences in the Normative Aging Study. *Am J Med* 1987; 82: 421-426 [PMID: 3826098 DOI: 10.1016/0002-9343(87)90441-4]
- 15 **Roddy E**. Revisiting the pathogenesis of podagra: why does gout target the foot? *J Foot Ankle Res* 2011; **4**: 13 [PMID: 21569453 DOI: 10.1186/1757-1146-4-13]
- 16 Roddy E, Zhang W, Doherty M. Are joints affected by gout also affected by osteoarthritis? Ann Rheum Dis 2007; 66: 1374-1377 [PMID: 17284542 DOI: 10.1136/ard.2006.063768]
- 17 Chen CJ, Shi Y, Hearn A, Fitzgerald K, Golenbock D, Reed G, Akira S, Rock KL. MyD88-dependent IL-1 receptor signaling is essential for gouty inflammation stimulated by monosodium urate crystals. J Clin Invest 2006; 116: 2262-2271 [PMID: 16886064 DOI: 10.1172/JCI28075]
- 18 Martinon F, Pétrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals activate the NALP3 inflammasome. *Nature* 2006; 440: 237-241 [PMID: 16407889 DOI: 10.1038/nature04516]
- 19 Joosten LA, Netea MG, Mylona E, Koenders MI, Malireddi RK, Oosting M, Stienstra R, van de Veerdonk FL, Stalenhoef AF, Giamarellos-Bourboulis EJ, Kanneganti TD, van der Meer JW. Engagement of fatty acids with Toll-like receptor 2 drives interleukin-1β production via the ASC/caspase 1 pathway in monosodium urate monohydrate crystalinduced gouty arthritis. *Arthritis Rheum* 2010; **62**: 3237-3248 [PMID: 20662061 DOI: 10.1002/art.27667]
- 20 Cronstein BN, Terkeltaub R. The inflammatory process of gout and its treatment. *Arthritis Res Ther* 2006; 8 Suppl 1: S3 [PMID: 16820042]
- 21 **Pope RM**, Tschopp J. The role of interleukin-1 and the inflammasome in gout: implications for therapy. *Arthritis Rheum* 2007; **56**: 3183-3188 [PMID: 17907163 DOI: 10.1002/art.22938]
- 22 Janssens HJ, Fransen J, van de Lisdonk EH, van Riel PL, van Weel C, Janssen M. A diagnostic rule for acute gouty arthritis in primary care without joint fluid analysis. *Arch Intern Med* 2010; **170**: 1120-1126 [PMID: 20625017 DOI: 10.1001/ archinternmed.2010.196]
- 23 Grahame R, Scott JT. Clinical survey of 354 patients with gout. Ann Rheum Dis 1970; 29: 461-468 [PMID: 5476673]
- 24 Forbess LJ, Fields TR. The broad spectrum of urate crystal deposition: unusual presentations of gouty tophi. *Semin Arthritis Rheum* 2012; **42**: 146-154 [PMID: 22522111 DOI: 10.1016/j.semarthrit.2012.03.007]
- 25 Choi HK, Atkinson K, Karlson EW, Willett W, Curhan G. Alcohol intake and risk of incident gout in men: a prospective study. *Lancet* 2004; 363: 1277-1281 [PMID: 15094272 DOI: 10.1016/S0140-6736(04)16000-5]
- 26 Singh JA, Reddy SG, Kundukulam J. Risk factors for gout and prevention: a systematic review of the literature. *Curr Opin Rheumatol* 2011; 23: 192-202 [PMID: 21285714 DOI:

10.1097/BOR.0b013e3283438e13]

- 27 Zhang Y, Chen C, Choi H, Chaisson C, Hunter D, Niu J, Neogi T. Purine-rich foods intake and recurrent gout attacks. *Ann Rheum Dis* 2012; 71: 1448-1453 [PMID: 22648933 DOI: 10.1136/annrheumdis-2011-201215]
- 28 Choi HK, Atkinson K, Karlson EW, Willett W, Curhan G. Purine-rich foods, dairy and protein intake, and the risk of gout in men. *N Engl J Med* 2004; **350**: 1093-1103 [PMID: 15014182 DOI: 10.1056/NEJMoa035700]
- 29 Choi HK, Curhan G. Soft drinks, fructose consumption, and the risk of gout in men: prospective cohort study. *BMJ* 2008; **336**: 309-312 [PMID: 18244959 DOI: 10.1136/ bmj.39449.819271.BE]
- 30 Dalbeth N, Ames R, Gamble GD, Horne A, Wong S, Kuhn-Sherlock B, MacGibbon A, McQueen FM, Reid IR, Palmano K. Effects of skim milk powder enriched with glycomacrope-ptide and G600 milk fat extract on frequency of gout flares: a proof-of-concept randomised controlled trial. *Ann Rheum Dis* 2012; **71**: 929-934 [PMID: 22275296 DOI: 10.1136/annrheum-dis-2011-200156]
- 31 Bruderer S, Bodmer M, Jick SS, Meier CR. Use of diuretics and risk of incident gout: a population-based case-control study. *Arthritis Rheumatol* 2014; 66: 185-196 [PMID: 24449584 DOI: 10.1002/art.38203]
- 32 Hueskes BA, Roovers EA, Mantel-Teeuwisse AK, Janssens HJ, van de Lisdonk EH, Janssen M. Use of diuretics and the risk of gouty arthritis: a systematic review. *Semin Arthritis Rheum* 2012; **41**: 879-889 [PMID: 22221907 DOI: 10.1016/j.se marthrit.2011.11.008]
- 33 Janssens HJ, van de Lisdonk EH, Janssen M, van den Hoogen HJ, Verbeek AL. Gout, not induced by diuretics? A casecontrol study from primary care. *Ann Rheum Dis* 2006; 65: 1080-1083 [PMID: 16291814 DOI: 10.1136/ard.2005.040360]
- 34 Kuo CF, Grainge MJ, See LC, Yu KH, Luo SF, Valdes AM, Zhang W, Doherty M. Familial aggregation of gout and relative genetic and environmental contributions: a nationwide population study in Taiwan. *Ann Rheum Dis* 2013; Epub ahead of print [PMID: 24265412 DOI: 10.1136/annrheumdis-2013-204067]
- 35 Lin KC, Lin HY, Chou P. Community based epidemiological study on hyperuricemia and gout in Kin-Hu, Kinmen. J *Rheumatol* 2000; 27: 1045-1050 [PMID: 10782835]
- 36 Choi HK, Ford ES, Li C, Curhan G. Prevalence of the metabolic syndrome in patients with gout: the Third National Health and Nutrition Examination Survey. *Arthritis Rheum* 2007; 57: 109-115 [PMID: 17266099 DOI: 10.1002/art.22466]
- 37 Kim SY, Guevara JP, Kim KM, Choi HK, Heitjan DF, Albert DA. Hyperuricemia and risk of stroke: a systematic review and meta-analysis. *Arthritis Rheum* 2009; 61: 885-892 [PMID: 19565556 DOI: 10.1002/art.24612]
- 38 Kim SY, Guevara JP, Kim KM, Choi HK, Heitjan DF, Albert DA. Hyperuricemia and coronary heart disease: a systematic review and meta-analysis. *Arthritis Care Res* (Hoboken) 2010; 62: 170-180 [PMID: 20191515 DOI: 10.1111/j.1365-2842.2009.02037.x]
- 39 **Krishnan E**. Gout and the risk for incident heart failure and systolic dysfunction. *BMJ Open* 2012; **2**: e000282 [PMID: 22337813 DOI: 10.1136/bmjopen-2011-000282]
- 40 Krishnan E, Svendsen K, Neaton JD, Grandits G, Kuller LH. Long-term cardiovascular mortality among middle-aged men with gout. Arch Intern Med 2008; 168: 1104-1110 [PMID: 18504339 DOI: 10.1001/archinte.168.10.1104]
- 41 Kuo CF, See LC, Luo SF, Ko YS, Lin YS, Hwang JS, Lin CM, Chen HW, Yu KH. Gout: an independent risk factor for allcause and cardiovascular mortality. *Rheumatology* (Oxford) 2010; 49: 141-146 [PMID: 19933595 DOI: 10.1093/rheumatology/kep364]
- 42 Li C, Hsieh MC, Chang SJ. Metabolic syndrome, diabetes, and hyperuricemia. *Curr Opin Rheumatol* 2013; **25**: 210-216 [PMID: 23370374 DOI: 10.1097/BOR.0b013e32835d951e]

- 43 Lai HM, Chen CJ, Su BY, Chen YC, Yu SF, Yen JH, Hsieh MC, Cheng TT, Chang SJ. Gout and type 2 diabetes have a mutual inter-dependent effect on genetic risk factors and higher incidences. *Rheumatology* (Oxford) 2012; **51**: 715-720 [PMID: 22179738 DOI: 10.1093/rheumatology/ker373]
- Choi HK, Curhan G. Independent impact of gout on mortality and risk for coronary heart disease. *Circulation* 2007; 116: 894-900 [PMID: 17698728 DOI: 10.1161/CIRCULA-TIONAHA.107.703389]
- 45 Kuo CF, Yu KH, See LC, Chou IJ, Ko YS, Chang HC, Chiou MJ, Luo SF. Risk of myocardial infarction among patients with gout: a nationwide population-based study. *Rheumatology* (Oxford) 2013; **52**: 111-117 [PMID: 22787006 DOI: 10.1093/rheumatology/kes169]
- 46 Zhang W, Doherty M, Pascual E, Bardin T, Barskova V, Conaghan P, Gerster J, Jacobs J, Leeb B, Lioté F, McCarthy G, Netter P, Nuki G, Perez-Ruiz F, Pignone A, Pimentão J, Punzi L, Roddy E, Uhlig T, Zimmermann-Gòrska I. EULAR evidence based recommendations for gout. Part I: Diagnosis. Report of a task force of the Standing Committee for International Clinical Studies Including Therapeutics (ESCISIT). Ann Rheum Dis 2006; 65: 1301-1311 [PMID: 16707533 DOI: 10.1136/ard.2006.055251]
- 47 **Gordon C**, Swan A, Dieppe P. Detection of crystals in synovial fluids by light microscopy: sensitivity and reliability. *Ann Rheum Dis* 1989; **48**: 737-742 [PMID: 2478085]
- 48 Lumbreras B, Pascual E, Frasquet J, González-Salinas J, Rodríguez E, Hernández-Aguado I. Analysis for crystals in synovial fluid: training of the analysts results in high consistency. *Ann Rheum Dis* 2005; 64: 612-615 [PMID: 15769916 DOI: 10.1136/ard.2004.027268]
- 49 Graf SW, Buchbinder R, Zochling J, Whittle SL. The accuracy of methods for urate crystal detection in synovial fluid and the effect of sample handling: a systematic review. *Clin Rheumatol* 2013; 32: 225-232 [PMID: 23138881 DOI: 10.1007/s10067-012-2107-0]
- 50 Owens D, Whelan B, McCarthy G. A survey of the management of gout in primary care. *Ir Med J* 2008; 101: 147-149 [PMID: 18624262]
- 51 Kienhorst LB, Janssens HJ, Fransen J, van de Lisdonk EH, Janssen M. Arthritis of the first metatarsophalangeal joint is not always gout: a prospective cohort study in primary care patients. *Joint Bone Spine* 2014; 81: 342-346 [PMID: 24468668 DOI: 10.1016/j.jbspin.2013.12.001]
- 52 **Pascual E**, Sivera F. Why is gout so poorly managed? Ann Rheum Dis 2007; **66**: 1269-1270 [PMID: 17881662 DOI: 10.1136/ard.2007.078469]
- 53 Pascual E, Sivera F, Andrés M. Synovial fluid analysis for crystals. *Curr Opin Rheumatol* 2011; 23: 161-169 [PMID: 21285711 DOI: 10.1097/BOR.0b013e328343e458]
- 54 Wallace SL, Robinson H, Masi AT, Decker JL, McCarty DJ, Yü TF. Preliminary criteria for the classification of the acute arthritis of primary gout. *Arthritis Rheum* 1977; 20: 895-900 [PMID: 856219]
- 55 Janssens HJ, Janssen M, van de Lisdonk EH, Fransen J, van Riel PL, van Weel C. Limited validity of the American College of Rheumatology criteria for classifying patients with gout in primary care. *Ann Rheum Dis* 2010; 69: 1255-1256 [PMID: 19910298 DOI: 10.1136/ard.2009.123687]
- 56 Malik A, Schumacher HR, Dinnella JE, Clayburne GM. Clinical diagnostic criteria for gout: comparison with the gold standard of synovial fluid crystal analysis. *J Clin Rheumatol* 2009; 15: 22-24 [PMID: 19125136 DOI: 10.1097/ RHU.0b013e3181945b79]
- 57 Rettenbacher T, Ennemoser S, Weirich H, Ulmer H, Hartig F, Klotz W, Herold M. Diagnostic imaging of gout: comparison of high-resolution US versus conventional X-ray. *Eur Radiol* 2008; 18: 621-630 [PMID: 17994238 DOI: 10.1007/s00330-007-0802-z]
- 58 Thiele RG, Schlesinger N. Diagnosis of gout by ultrasound.

Rheumatology (Oxford) 2007; **46**: 1116-1121 [PMID: 17468505 DOI: 10.1093/rheumatology/kem058]

- 59 Dalbeth N, Doyle AJ. Imaging of gout: an overview. Best Pract Res Clin Rheumatol 2012; 26: 823-838 [PMID: 23273794 DOI: 10.1016/j.berh.2012.09.003]
- 60 Glazebrook KN, Guimarães LS, Murthy NS, Black DF, Bongartz T, Manek NJ, Leng S, Fletcher JG, McCollough CH. Identification of intraarticular and periarticular uric acid crystals with dual-energy CT: initial evaluation. *Radiology* 2011; 261: 516-524 [PMID: 21926378 DOI: 10.1148/radiol.11102485]
- 61 **Manger B**, Lell M, Wacker J, Schett G, Rech J. Detection of periarticular urate deposits with dual energy CT in patients with acute gouty arthritis. *Ann Rheum Dis* 2012; **71**: 470-472 [PMID: 21859688]
- 62 Huppertz A, Hermann KG, Diekhoff T, Wagner M, Hamm B, Schmidt WA. Systemic staging for urate crystal deposits with dual-energy CT and ultrasound in patients with suspected gout. *Rheumatol Int* 2014; **34**: 763-771 [PMID: 24619560]
- 63 Janssens HJ, Janssen M, van de Lisdonk EH, van Riel PL, van Weel C. Use of oral prednisolone or naproxen for the treatment of gout arthritis: a double-blind, randomised equivalence trial. *Lancet* 2008; **371**: 1854-1860 [PMID: 18514729 DOI: 10.1016/S0140-6736(08)60799-0]
- 64 **Man CY**, Cheung IT, Cameron PA, Rainer TH. Comparison of oral prednisolone/paracetamol and oral indomethacin/ paracetamol combination therapy in the treatment of acute goutlike arthritis: a double-blind, randomized, controlled trial. *Ann Emerg Med* 2007; **49**: 670-677 [PMID: 17276548 DOI: 10.1016/j.annemergmed.2006.11.014]
- 65 **Underwood M**. Gout. *Clin Evid* (Online) 2011; **2011:** 1120 [PMID: 21575286]
- 66 Terkeltaub RA, Furst DE, Bennett K, Kook KA, Crockett RS, Davis MW. High versus low dosing of oral colchicine for early acute gout flare: Twenty-four-hour outcome of the first multicenter, randomized, double-blind, placebo-controlled, parallel-group, dose-comparison colchicine study. *Arthritis Rheum* 2010; 62: 1060-1068 [PMID: 20131255 DOI: 10.1002/ art.27327]
- 67 Schlesinger N, Alten RE, Bardin T, Schumacher HR, Bloch M, Gimona A, Krammer G, Murphy V, Richard D, So AK. Canakinumab for acute gouty arthritis in patients with limited treatment options: results from two randomised, multicentre, active-controlled, double-blind trials and their initial extensions. *Ann Rheum Dis* 2012; **71**: 1839-1848 [PMID: 22586173 DOI: 10.1136/annrheumdis-2011-200908]
- 68 So A, De Smedt T, Revaz S, Tschopp J. A pilot study of IL-1 inhibition by anakinra in acute gout. *Arthritis Res Ther* 2007; 9: R28 [PMID: 17352828 DOI: 10.1186/ar2143]
- 69 Chen K, Fields T, Mancuso CA, Bass AR, Vasanth L. Anakinra's efficacy is variable in refractory gout: report of ten cases. *Semin Arthritis Rheum* 2010; 40: 210-214 [PMID: 20494407 DOI: 10.1016/j.semarthrit.2010.03.001]
- 70 Ottaviani S, Moltó A, Ea HK, Neveu S, Gill G, Brunier L, Palazzo E, Meyer O, Richette P, Bardin T, Allanore Y, Lioté F, Dougados M, Dieudé P. Efficacy of anakinra in gouty arthritis: a retrospective study of 40 cases. *Arthritis Res Ther* 2013; 15: R123 [PMID: 24432362 DOI: 10.1186/ar4303]
- 71 Terkeltaub R, Sundy JS, Schumacher HR, Murphy F, Bookbinder S, Biedermann S, Wu R, Mellis S, Radin A. The interleukin 1 inhibitor rilonacept in treatment of chronic gouty arthritis: results of a placebo-controlled, monosequence crossover, non-randomised, single-blind pilot study. *Ann Rheum Dis* 2009; 68: 1613-1617 [PMID: 19635719 DOI: 10.1136/ard.2009.108936]
- 72 Schumacher HR, Sundy JS, Terkeltaub R, Knapp HR, Mellis SJ, Stahl N, Yancopoulos GD, Soo Y, King-Davis S, Weinstein SP, Radin AR. Rilonacept (interleukin-1 trap) in the prevention of acute gout flares during initiation of urate-lowering therapy: results of a phase II randomized, double-blind, pla-

cebo-controlled trial. *Arthritis Rheum* 2012; **64**: 876-884 [PMID: 22223180 DOI: 10.1002/art.33412]

- 73 Schumacher HR, Evans RR, Saag KG, Clower J, Jennings W, Weinstein SP, Yancopoulos GD, Wang J, Terkeltaub R. Rilonacept (interleukin-1 trap) for prevention of gout flares during initiation of uric acid-lowering therapy: results from a phase III randomized, double-blind, placebo-controlled, confirmatory efficacy study. *Arthritis Care Res* (Hoboken) 2012; 64: 1462-1470 [PMID: 22549879 DOI: 10.1002/acr.21690]
- 74 Terkeltaub RA, Schumacher HR, Carter JD, Baraf HS, Evans RR, Wang J, King-Davis S, Weinstein SP Rilonacept in the treatment of acute gouty arthritis: a randomized, controlled clinical trial using indomethacin as the active comparator. *Arthritis Res Ther* 2013; 15: R25 [DOI: 10.1186/ar4159]
- 75 Mitha E, Schumacher HR, Fouche L, Luo SF, Weinstein SP, Yancopoulos GD, Wang J, King-Davis S, Evans RR. Rilonacept for gout flare prevention during initiation of uric acid-lowering therapy: results from the PRESURGE-2 international, phase 3, randomized, placebo-controlled trial. *Rheumatology* (Oxford) 2013; **52**: 1285-1292 [PMID: 23485476 DOI: 10.1093/rheumatology/ket114]
- 76 Moi JH, Sriranganathan MK, Edwards CJ, Buchbinder R. Lifestyle interventions for acute gout. *Cochrane Database Syst Rev* 2013; 11: CD010519 [PMID: 24186771]
- 77 Schlesinger N, Detry MA, Holland BK, Baker DG, Beutler AM, Rull M, Hoffman BI, Schumacher HR. Local ice therapy during bouts of acute gouty arthritis. *J Rheumatol* 2002; 29: 331-334 [PMID: 11838852]
- 78 Khanna D, Fitzgerald JD, Khanna PP, Bae S, Singh MK, Neogi T, Pillinger MH, Merill J, Lee S, Prakash S, Kaldas M, Gogia M, Perez-Ruiz F, Taylor W, Lioté F, Choi H, Singh JA, Dalbeth N, Kaplan S, Niyyar V, Jones D, Yarows SA, Roessler B, Kerr G, King C, Levy G, Furst DE, Edwards NL, Mandell B, Schumacher HR, Robbins M, Wenger N, Terkeltaub R. 2012 American College of Rheumatology guidelines for management of gout. Part 1: systematic nonpharmacologic and pharmacologic therapeutic approaches to hyperuricemia. *Arthritis Care Res* (Hoboken) 2012; 64: 1431-1446 [PMID: 23024028]
- 79 Zhang W, Doherty M, Bardin T, Pascual E, Barskova V, Conaghan P, Gerster J, Jacobs J, Leeb B, Lioté F, McCarthy G, Netter P, Nuki G, Perez-Ruiz F, Pignone A, Pimentão J, Punzi L, Roddy E, Uhlig T, Zimmermann-Gòrska I. EULAR evidence based recommendations for gout. Part II: Management. Report of a task force of the EULAR Standing Committee for International Clinical Studies Including Therapeutics (ESCISIT). Ann Rheum Dis 2006; 65: 1312-1324 [PMID: 16707532 DOI: 10.1136/ard.2006.055269]
- 80 Perez-Ruiz F. Treating to target: a strategy to cure gout. Rheumatology (Oxford) 2009; 48 Suppl 2: ii9-ii14 [PMID: 19447780 DOI: 10.1093/rheumatology/kep087]
- 81 Shoji A, Yamanaka H, Kamatani N. A retrospective study of the relationship between serum urate level and recurrent attacks of gouty arthritis: evidence for reduction of recurrent gouty arthritis with antihyperuricemic therapy. *Arthritis Rheum* 2004; **51**: 321-325 [PMID: 15188314 DOI: 10.1002/ art.20405]
- 82 Perez-Ruiz F, Calabozo M, Pijoan JI, Herrero-Beites AM, Ruibal A. Effect of urate-lowering therapy on the velocity of size reduction of tophi in chronic gout. *Arthritis Rheum* 2002; 47: 356-360 [PMID: 12209479 DOI: 10.1002/art.10511]
- 83 **Perez-Ruiz F**, Herrero-Beites AM, Carmona L. A two-stage approach to the treatment of hyperuricemia in gout: the "dirty dish" hypothesis. *Arthritis Rheum* 2011; **63**: 4002-4006 [PMID: 21898351 DOI: 10.1002/art.30649]
- 84 Reinders MK, van Roon EN, Jansen TL, Delsing J, Griep EN, Hoekstra M, van de Laar MA, Brouwers JR. Efficacy and tolerability of urate-lowering drugs in gout: a randomised controlled trial of benzbromarone versus probenecid after failure of allopurinol. *Ann Rheum Dis* 2009; 68: 51-56 [PMID:

18250112 DOI: 10.1136/ard.2007.083071]

- 85 Pui K, Gow PJ, Dalbeth N. Efficacy and tolerability of probenecid as urate-lowering therapy in gout; clinical experience in high-prevalence population. *J Rheumatol* 2013; 40: 872-876 [PMID: 23457380 DOI: 10.3899/jrheum.121301]
- Tayar JH, Lopez-Olivo MA, Suarez-Almazor ME. Febuxostat for treating chronic gout. *Cochrane Database Syst Rev* 2012; 11: CD008653 [PMID: 23152264 DOI: 10.1002/14651858. CD008653.pub2]
- 87 Moolenburgh JD, Reinders MK, Jansen TL. Rasburicase treatment in severe tophaceous gout: a novel therapeutic option. *Clin Rheumatol* 2006; 25: 749-752 [PMID: 16247589 DOI: 10.1007/s10067-005-0043-y]
- 88 Richette P, Bardin T. Successful treatment with rasburicase of a tophaceous gout in a patient allergic to allopurinol. *Nat Clin Pract Rheumatol* 2006; 2: 338-342; quiz 343 [PMID: 16932713 DOI: 10.1038/ncprheum0214]
- 89 Richette P, Viguier M, Bachelez H, Bardin T. Psoriasis induced by anti-tumor necrosis factor therapy: a class effect? J Rheumatol 2007; 34: 438-439 [PMID: 17304662]
- 90 De Angelis S, Noce A, Di Renzo L, Cianci R, Naticchia A, Giarrizzo GF, Giordano F, Tozzo C, Splendiani G, De Lorenzo A. Is rasburicase an effective alternative to allopurinol for management of hyperuricemia in renal failure patients? A double blind-randomized study. *Eur Rev Med Pharmacol Sci* 2007; **11**: 179-184 [PMID: 17970234]
- 91 Sundy JS, Becker MA, Baraf HS, Barkhuizen A, Moreland LW, Huang W, Waltrip RW, Maroli AN, Horowitz Z. Reduction of plasma urate levels following treatment with multiple doses of pegloticase (polyethylene glycol-conjugated uricase) in patients with treatment-failure gout: results of a phase II randomized study. *Arthritis Rheum* 2008; **58**: 2882-2891 [PMID: 18759308 DOI: 10.1002/art.23810]
- 92 Anderson A, Singh JA. Pegloticase for chronic gout. Cochrane Database Syst Rev 2010; (3): CD008335 [PMID: 20238366 DOI: 10.1002/14651858.CD008335.pub2]
- 93 Sundy JS, Baraf HS, Yood RA, Edwards NL, Gutierrez-Urena SR, Treadwell EL, Vázquez-Mellado J, White WB, Lipsky PE, Horowitz Z, Huang W, Maroli AN, Waltrip RW, Hamburger SA, Becker MA. Efficacy and tolerability of pegloticase for the treatment of chronic gout in patients refractory to conventional treatment: two randomized controlled trials. *JAMA* 2011; 306: 711-720 [PMID: 21846852 DOI: 10.1001/jama.2011.1169]
- 94 Becker MA, Baraf HS, Yood RA, Dillon A, Vázquez-Mellado J, Ottery FD, Khanna D, Sundy JS. Long-term safety of pegloticase in chronic gout refractory to conventional treatment. *Ann Rheum Dis* 2013; 72: 1469-1474 [PMID: 23144450 DOI: 10.1136/annrheumdis-2012-201795]
- 95 Baraf HS, Becker MA, Gutierrez-Urena SR, Treadwell EL, Vazquez-Mellado J, Rehrig CD, Ottery FD, Sundy JS, Yood RA. Tophus burden reduction with pegloticase: results from phase 3 randomized trials and open-label extension in patients with chronic gout refractory to conventional therapy. *Arthritis Res Ther* 2013; **15**: R137 [PMID: 24286509]
- 96 Crittenden DB, Pillinger MH. New therapies for gout. Annu Rev Med 2013; 64: 325-337 [PMID: 23327525 DOI: 10.1146/ annurev-med-080911-105830]
- 97 Fleischmann R, Kerr B, Yeh LT, Suster M, Shen Z, Polvent E, Hingorani V, Quart B, Manhard K, Miner JN, Baumgartner S; on behalf of the RDEA594-111 Study Group. Pharmacodynamic, pharmacokinetic and tolerability evaluation of concomitant administration of lesinurad and febuxostat in gout patients with hyperuricaemia. *Rheumatology* (Oxford) 2014 Feb 8; Epub ahead of print [PMID: 24509406 DOI: 10.1093/ rheumatology/ket487]
- 98 Würzner G, Gerster JC, Chiolero A, Maillard M, Fallab-Stubi CL, Brunner HR, Burnier M. Comparative effects of losartan and irbesartan on serum uric acid in hypertensive patients with hyperuricaemia and gout. J Hypertens 2001; 19:

1855-1860 [PMID: 11593107]

- 99 Huang HY, Appel LJ, Choi MJ, Gelber AC, Charleston J, Norkus EP, Miller ER. The effects of vitamin C supplementation on serum concentrations of uric acid: results of a randomized controlled trial. *Arthritis Rheum* 2005; **52**: 1843-1847 [PMID: 15934094 DOI: 10.1002/art.21105]
- Juraschek SP, Miller ER, Gelber AC. Effect of oral vitamin C supplementation on serum uric acid: a meta-analysis of randomized controlled trials. *Arthritis Care Res* (Hoboken) 2011;
 63: 1295-1306 [PMID: 21671418 DOI: 10.1002/acr.20519]
- 101 Stamp LK, O'Donnell JL, Frampton C, Drake JM, Zhang M, Chapman PT. Clinically insignificant effect of supplemental vitamin C on serum urate in patients with gout: a pilot randomized controlled trial. *Arthritis Rheum* 2013; 65: 1636-1642 [PMID: 23681955 DOI: 10.1002/art.37925]
- 102 Nederlandse Vereniging voor Reumatologie (Richtlijn Jicht. 2013). Available from: URL: http://www.nvr.nl/richtlijnen/ richtlijnen2
- 103 Neogi T, Hunter DJ, Chaisson CE, Allensworth-Davies D, Zhang Y. Frequency and predictors of inappropriate management of recurrent gout attacks in a longitudinal study. J *Rheumatol* 2006; 33: 104-109 [PMID: 16267879]
- 104 Roddy E, Zhang W, Doherty M. Concordance of the management of chronic gout in a UK primary-care population with the EULAR gout recommendations. *Ann Rheum Dis* 2007; 66: 1311-1315 [PMID: 17504843 DOI: 10.1136/ard.2007.070755]
- 105 Mikuls TR, Farrar JT, Bilker WB, Fernandes S, Saag KG. Suboptimal physician adherence to quality indicators for the management of gout and asymptomatic hyperuricaemia: results from the UK General Practice Research Database (GPRD). *Rheumatology* (Oxford) 2005; 44: 1038-1042 [PMID: 15870145 DOI: 10.1093/rheumatology/keh679]
- 106 Doherty M, Jansen TL, Nuki G, Pascual E, Perez-Ruiz F, Punzi L, So AK, Bardin T. Gout: why is this curable disease so seldom cured? *Ann Rheum Dis* 2012; **71**: 1765-1770 [PMID: 22863577 DOI: 10.1136/annrheumdis-2012-201687]
- 107 De Vera MA, Marcotte G, Rai S, Galo JS, Bhole V. Medication adherence in gout: a systematic review. *Arthritis Care Res* (Hoboken) 2014; 66: 1551-1559 [PMID: 24692321 DOI: 10.1002/acr.22336]
- 108 Zandman-Goddard G, Amital H, Shamrayevsky N, Raz R, Shalev V, Chodick G. Rates of adherence and persistence with allopurinol therapy among gout patients in Israel. *Rheumatology* (Oxford) 2013; **52**: 1126-1131 [PMID: 23392592 DOI: 10.1093/rheumatology/kes431]
- 109 Rees F, Jenkins W, Doherty M. Patients with gout adhere to curative treatment if informed appropriately: proof-of-concept observational study. *Ann Rheum Dis* 2013; **72**: 826-830 [PMID: 22679303 DOI: 10.1136/annrheumdis-2012-201676]
- 110 Brook RA, Forsythe A, Smeeding JE, Lawrence Edwards N. Chronic gout: epidemiology, disease progression, treatment and disease burden. *Curr Med Res Opin* 2010; 26: 2813-2821 [PMID: 21050059 DOI: 10.1185/03007995.2010.533647]
- 111 Edwards NL, Sundy JS, Forsythe A, Blume S, Pan F, Becker MA. Work productivity loss due to flares in patients with chronic gout refractory to conventional therapy. J Med Econ 2011; 14: 10-15 [PMID: 21138339 DOI: 10.3111/13696998.2010 .540874]
- 112 Khanna PP, Nuki G, Bardin T, Tausche AK, Forsythe A, Goren A, Vietri J, Khanna D. Tophi and frequent gout flares are associated with impairments to quality of life, productivity, and increased healthcare resource use: Results from a crosssectional survey. *Health Qual Life Outcomes* 2012; **10**: 117 [PMID: 22999027 DOI: 10.1186/1477-7525-10-117]
- 113 Ferraz MB, O'Brien B. A cost effectiveness analysis of urate lowering drugs in nontophaceous recurrent gouty arthritis. J Rheumatol 1995; 22: 908-914 [PMID: 8587081]
- 114 **Perez-Ruiz F**, Martínez-Indart L, Carmona L, Herrero-Beites AM, Pijoan JI, Krishnan E. Tophaceous gout and high level of hyperuricaemia are both associated with increased risk

of mortality in patients with gout. *Ann Rheum Dis* 2014; **73**: 177-182 [PMID: 23313809 DOI: 10.1136/annrheumdis-2012-202421]

- 115 Culleton BF, Larson MG, Kannel WB, Levy D. Serum uric acid and risk for cardiovascular disease and death: the Framingham Heart Study. Ann Intern Med 1999; 131: 7-13 [PMID: 10391820 DOI: 10.7326/0003-4819-131-1-199907060-00003]
- 116 Fang J, Alderman MH. Serum uric acid and cardiovascular mortality the NHANES I epidemiologic follow-up study, 1971-1992. National Health and Nutrition Examination Survey. JAMA 2000; 283: 2404-2410 [PMID: 10815083 DOI: 10.1001/jama.283.18.2404]
- 117 Johnson RJ, Kang DH, Feig D, Kivlighn S, Kanellis J, Watanabe S, Tuttle KR, Rodriguez-Iturbe B, Herrera-Acosta J, Mazzali M. Is there a pathogenetic role for uric acid in hypertension and cardiovascular and renal disease? *Hypertension* 2003; **41**: 1183-1190 [PMID: 12707287 DOI: 10.1161/01. HYP.0000069700.62727.C5]
- 118 Teng GG, Ang LW, Saag KG, Yu MC, Yuan JM, Koh WP. Mortality due to coronary heart disease and kidney disease among middle-aged and elderly men and women with gout in the Singapore Chinese Health Study. *Ann Rheum Dis* 2012; 71: 924-928 [PMID: 22172492 DOI: 10.1136/ard.2011.200523]
- 119 Landmesser U, Spiekermann S, Dikalov S, Tatge H, Wilke R, Kohler C, Harrison DG, Hornig B, Drexler H. Vascular oxidative stress and endothelial dysfunction in patients with chronic heart failure: role of xanthine-oxidase and extracellular superoxide dismutase. *Circulation* 2002; **106**: 3073-3078 [PMID: 12473554 DOI: 10.1161/01.CIR.0000041431.57222.AF]
- 120 Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 2000; 87: 840-844 [PMID: 11073878 DOI: 10.1161/01.RES.87.10.840]
- 121 Dawson J, Quinn T, Walters M. Uric acid reduction: a new paradigm in the management of cardiovascular risk? *Curr Med Chem* 2007; 14: 1879-1886 [PMID: 17627523]
- 122 Doehner W, Schoene N, Rauchhaus M, Leyva-Leon F, Pavitt DV, Reaveley DA, Schuler G, Coats AJ, Anker SD, Hambrecht R. Effects of xanthine oxidase inhibition with allopurinol on endothelial function and peripheral blood flow in hyperuricemic patients with chronic heart failure: results from 2 placebo-controlled studies. *Circulation* 2002; **105**: 2619-2624 [PMID: 12045167 DOI: 10.1161/01.CIR.0000017502.58595.ED]
- 123 Feig DI, Soletsky B, Johnson RJ. Effect of allopurinol on blood pressure of adolescents with newly diagnosed essential hypertension: a randomized trial. *JAMA* 2008; 300: 924-932 [PMID: 18728266 DOI: 10.1001/jama.300.8.924]
- 124 Kanbay M, Ozkara A, Selcoki Y, Isik B, Turgut F, Bavbek N, Uz E, Akcay A, Yigitoglu R, Covic A. Effect of treatment of hyperuricemia with allopurinol on blood pressure, creatinine clearence, and proteinuria in patients with normal renal functions. *Int Urol Nephrol* 2007; **39**: 1227-1233 [PMID: 17701281]
- 125 Gois PH, Souza ER. Pharmacotherapy for hyperuricemia in hypertensive patients. *Cochrane Database Syst Rev* 2013;
 1: CD008652 [PMID: 23440832 DOI: 10.1002/14651858. CD008652.pub2]
- 126 Agarwal V, Messerli F. Effect of allopurinol on blood pressure (author response to Allopurinol on hypertension: insufficient evidence to recommend). *J Clin Hypertens* (Greenwich) 2013; **15**: 701 [PMID: 24034667 DOI: 10.1111/jch.12152]
- 127 Noman A, Ang DS, Ogston S, Lang CC, Struthers AD. Effect of high-dose allopurinol on exercise in patients with chronic stable angina: a randomised, placebo controlled crossover trial. *Lancet* 2010; **375**: 2161-2167 [PMID: 20542554 DOI: 10.1016/S0140-6736(10)60391-1]
- 128 Struthers A, Shearer F. Allopurinol: novel indications in cardiovascular disease. *Heart* 2012; 98: 1543-1545 [PMID: 22801998 DOI: 10.1136/heartjnl-2012-302249]
- 129 **Tsutsumi Z**, Moriwaki Y, Takahashi S, Ka T, Yamamoto T. Oxidized low-density lipoprotein autoantibodies in patients with primary gout: effect of urate-lowering therapy. *Clin*

Chim Acta 2004; 339: 117-122 [PMID: 14687901]

- 130 Luk AJ, Levin GP, Moore EE, Zhou XH, Kestenbaum BR, Choi HK. Allopurinol and mortality in hyperuricaemic patients. *Rheumatology* (Oxford) 2009; 48: 804-806 [PMID: 19447769 DOI: 10.1093/rheumatology/kep069]
- 131 Wei L, Mackenzie IS, Chen Y, Struthers AD, MacDonald TM. Impact of allopurinol use on urate concentration and cardiovascular outcome. *Br J Clin Pharmacol* 2011; **71**: 600-607 [PMID: 21395653 DOI: 10.1111/j.1365-2125.2010.03887.x]
- 132 Struthers AD, Donnan PT, Lindsay P, McNaughton D, Broomhall J, MacDonald TM. Effect of allopurinol on mortality and hospitalisations in chronic heart failure: a retrospective cohort study. *Heart* 2002; 87: 229-234 [PMID: 11847159 DOI: 10.1136/heart.87.3.229]
- 133 Wei L, Fahey T, Struthers AD, MacDonald TM. Association between allopurinol and mortality in heart failure patients: a long-term follow-up study. *Int J Clin Pract* 2009; 63: 1327-1333 [PMID: 19691616 DOI: 10.1111/j.1742-1241.2009.02118.x]
- 134 Grimaldi-Bensouda L, Alpérovitch A, Aubrun E, Danchin N,

Rossignol M, Abenhaim L, Richette P; the PGRx MI Group. Impact of allopurinol on risk of myocardial infarction. *Ann Rheum Dis* 2014 Jan 6; Epub ahead of print [PMID: 24395556 DOI: 10.1136/annrheumdis-2012-202972]

- 135 Ogino K, Kato M, Furuse Y, Kinugasa Y, Ishida K, Osaki S, Kinugawa T, Igawa O, Hisatome I, Shigemasa C, Anker SD, Doehner W. Uric acid-lowering treatment with benzbromarone in patients with heart failure: a double-blind placebo-controlled crossover preliminary study. *Circ Heart Fail* 2010; **3**: 73-81 [PMID: 19933411 DOI: 10.1161/CIRCHEARTFAILURE.109.868604]
- 136 Lyngdoh T, Marques-Vidal P, Paccaud F, Preisig M, Waeber G, Bochud M, Vollenweider P. Elevated serum uric acid is associated with high circulating inflammatory cytokines in the population-based Colaus study. *PLoS One* 2011; 6: e19901 [PMID: 21625475 DOI: 10.1371/journal.pone.0019901]
- 137 Chowalloor PV, Keen HI. A systematic review of ultrasonography in gout and asymptomatic hyperuricaemia. Ann Rheum Dis 2013; 72: 638-645 [PMID: 23291387 DOI: 10.1136/ annrheumdis-2012-202301]

P-Reviewer: Baran DA, Beltowski J S- Editor: Ji FF L- Editor: A E- Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.72 World J Rheumatol 2014 November 12; 4(3): 72-79 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

MINIREVIEWS

Quantifying synovial inflammation: Emerging imaging techniques

Deepak Tripathi, Vikas Agarwal

Deepak Tripathi, Vikas Agarwal, Department of Clinical Immunology, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow UP 226014, India

Author contributions: Tripathi D and Agarwal V contributed solely to this paper.

Supported by ICMR, No. 5/4-5/4/ortho/08/NCD-1

Correspondence to: Vikas Agarwal, MD, DM, Additional Professor, Department of Clinical Immunology, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Raebareli Road, Lucknow UP 226014, India. vikasagr@yahoo.com

Telephone: +91-52-22494318 Fax: +91-52-22668812

Received: April 12, 2014 Revised: July 23, 2014

Accepted: September 6, 2014

Published online: November 12, 2014

Abstract

Imaging techniques to assess synovial inflammation includes radiography, ultrasound, computed tomography, magnetic resonance imaging (MRI) and recently positron emission tomography. The ideal objective of imaging approaches are to quantify synovial inflammation by capturing features such as synovial hyperplasia, neo-angiogenesis and infiltration of immune cells in the synovium. This may enable clinicians to estimate response to therapy by measuring the improvement in the inflammatory signals at the level of synovium. Ultrasound can provide information regarding thickening of the synovial membrane and can reveal increased synovial blood flow using power Doppler technique. Bone marrow edema and synovial membrane thickness on MRI scan may serve as indicators for arthritis progression. Enhancement of the synovium on dynamic contrast MRI may closely mirror the inflammatory activity in the synovium. Diffusion tensor imaging is an advance MRI approach that evaluates the inflammation related to cell infiltration or aggregation in an inflamed synovium. In this review, we summarize the newer imaging techniques and their developments to evaluate synovial inflammation.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Synovitis; Imaging; Diffusion tensor imaging; Positron emission tomography; Computed tomography; Ultrasound

Core tip: Nowadays, more and more emphasis is being put on capturing the microscopic features of inflammation on non-invasive techniques of imaging. Emerging magnetic resonance imaging techniques seem to have potential to capture these molecular events and replace synovial histology in future. In this paper we have reviewed exciting recent advances in the field of imaging that pick up inflammatory signals from inflamed synovium and are likely to be available for routine clinical practice in near future.

Tripathi D, Agarwal V. Quantifying synovial inflammation: Emerging imaging techniques. *World J Rheumatol* 2014; 4(3): 72-79 Available from: URL: http://www.wjgnet.com/2220-3214/ full/v4/i3/72.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.72

INTRODUCTION

In chronic inflammatory joint diseases, synovium is the major site of inflammation. During inflammation, the phenotype of synovium is modified and it changes into thickened invasive tissue that erodes into surrounding soft tissues (cartilage, ligaments and tendons) and bone tissue. It is difficult to define the stages during the transformation of non-specific synovitis into aggressive invasive destructive synovitis^[1]. Infiltration of macrophages in the synovial membrane is of pathogenic importance because macrophages generate several pro-inflammatory cytokines for example interleukin-1 β (IL-1 β), interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α) activate synovial neovascularisation and cause damage to joint by various mechanisms^[2]. Greater understanding about the pathogenesis of



synovial inflammation has led to development of focused biologic treatments that minimize disease progression and tissue destruction. Development of biologics such as anti-TNF- α monoclonal antibodies, antibodies against IL-6, receptor activator for nuclear factor- κ B ligand, IL-17 and CD20 (B cell) improve arthritis symptoms by reducing the synovial inflammation^[3]. The full-benefits of newer treatments can only be noticed if tools are available to perfectly identify the site and severity of synovial inflammation before irreversible damage occurs. In this review, we focus on the current improvements in imaging modalities as well as how these newer imaging modalities can be applied in the monitoring of synovitis and application of these newer imaging modalities in effective control of synovial inflammation in various arthritic disorders.

RADIOGRAPHY

Virtually all patients undergo radiographs or X-rays of the joint at initial presentation to know the extent of the disease and damage to the joints. Sequential image analysis is commonly executed during the treatment to monitor synovitis progression or regression (erosion scores or joint space narrowing). It has following advantages; easy availability, extensive image acquisition of almost all the joints areas, latest digitised formats grant simple restoration and evaluation of images in future and comparatively very economical. Drawbacks include contact with ionising radiation, which is however comparatively low for one time of X-rays but can become high over a period of time during sequential X-rays^[4] and, most significantly, a lack of sensitivity in detecting early synovial inflammation and early joint damage^[5,6].

X-ray in synovitis progression

Plain X-rays are not beneficial in monitoring of progression of synovitis in early arthritis patients. X-rays of wrist and knee joint are abnormal in 15%-30% of patients at initial presentation who finally fulfils the diagnostic criteria of rheumatoid arthritis (RA)^[7]. In early RA, radiographs reveal non-specific synovial thickening and periarticular osteopenia, neither of which is diagnostic. However, in late RA characteristic erosions and their pattern of joint involvement may suggest diagnosis of RA but by then disease is far advanced. Despite this fact, radiographs have been extensively validated in a number of clinical trials. In ATTRACT (Anti TNF Therapy in RA with Concomitant Therapy) study, examining the effectiveness of infliximab in preventing progression of joint erosions^[8], the two radiologists had a high reliability with correlations varying between 0.84 at baseline to 0.92 at weeks 102 of total radiographic scores. However, radiographs provide a very little information regarding severity of inflammation and lacks quantifiable variables of inflammation in early arthritis. They are reliable indicators of damage.

ULTRASONOGRAPHY

Latest reports recommend that ultrasonography (US)

may be an important imaging technique to determine the degree of synovitis in active RA joints. Furthermore, it may be more sensitive in identifying active synovial inflammation than clinical joint examination^[9]. Recently, Le Boedec et al^{10]} reported that US may provide additional useful information beyond clinical joint examination in the shoulders and metatarsophalangeal joints. They further concluded that the usefulness of power Doppler and B mode ultrasound were reduced with low DAS28 score and shorter disease duration respectively^[10]. US can provide information about disease activity (synovial inflammation and tenosynovitis) and joint damage (bone erosions)^[11]. Hence, US not only may help in examination but may also help in monitoring outcome of treatment. Ultrasonic waves when hit the tissue interfaces, they are reflected back and these reflected waves (echo waves) are recorded to generate US images. Thickness of synovium in areas of joint capsule and tenosynovium can be detected by B mode grey scale US (GSUS)^[12]. Synovial inflammation in the knee of RA patients was examined through GSUS at 5.0 MHz in a study evaluating Yttrium-90 radiation synovectomy^[13]. In this study GSUS findings (suprapatellar effusion and synovial thickening) correlated well with the clinical and arthrographic findings.

US imaging approach is even more enhanced with the application of Doppler techniques^[14]. Flow of red blood cells, either towards the US probe or away from it, demonstrated by Colour Doppler US is suitable for analyzing excessive flow rate in blood vessels^[15]. Hypertrophy of the synovium is determined as non- displaceable, intraarticular, poorly compressible, which can be recognized by Doppler signal^[16]. Thickened synovium is much less compressible therefore on probe compress fluid movement is very less while in normal synovium fluid is displaced easily on probe compression. Sometimes, normal anatomical tissues may mimic synovial inflammation due to low reflectivity of US waves, particularly with US equipment of lower resolution^[17].

Power Doppler US (PDUS) is the most beneficial US technique in rheumatology. It analyses Doppler changes of the moving red blood cell, regardless of its route and rate, and is consequently well developed for the quantification of blood flow rate within the low flow synovium^[18,19]. PDUS can quantify inflammatory activity in the erosions of RA by revealing increased blood flow in the synovium of these patients. Histological analysis have confirmed that changes in power Doppler signals are associated with inflammatory changes in the synovial membrane^[20], however one must be cautious about artefact signals at bone synovium interface especially if gain setting of the US machine is high. Increased Doppler signal matches specifically with neutrophils recruitment and area fibrin accumulation^[21], however, no exact correlation between systemic vascular endothelial growth factor expression and neo-vascularisation was demonstrated. PDUS is sensitive but also more vulnerable to false signals. The software that measures PDUS is proprietary; therefore, results from one manufacturer may not be comparable with another.

Zaishideng®

POSITRON EMISSION TOMOGRAPHY

Positron emission tomography (PET) is a sensitive image acquisition technique that depends on ion emitting isotopes of radioactive element. Positron loses energy after it collides with atoms and is exhausted after colliding with an electron, leading to the emission of two gamma rays. After collision two gamma rays travel in opposite direction. The PET sensor captures signal of these two energy packets (photons) along with their relative position. PET sensors at the same time capture signal of these energy packet from various directions. Associated computed tomography (CT) scanning offers superior structural information of the tissues and spatial localization of the PET signals. A tracer molecule fluorodeoxyglucose [(18F) FDG] has been widely used in PET research. It has homology with the sugar molecule but it contains radioactive isotope, upon accumulation in the metabolic active cells it gives the signal which is captured by PET scanning and provide information about the location of inflamed tissue. Increased uptake of this sugar analogue is directed by the various sugar transporter receptors such as GLUT1 and GLUT3 on the cell surface, both are over expressed on hyper metabolic cells. This sugar analogue is phosphorylated by the hexokinase enzyme but it is not metabolised further in the glycolytic pathway and is trapped in the cells^[22].

Several attempts with the FDG PET in the estimation of synovial inflammation in arthritis demonstrated that there is enhanced uptake of 18F-FDG in the inflamed synovium^[23,24]. The semi-quantitative scoring of joint involvement and quantitative uptake of 18F-FDG has been shown to correlate with markers of inflammation^[25]. Another tracer molecule 11C R PK11195 has also been used in PET scanning which specifically binds to the receptors present on phagocytic cells. This group analysed joint of 11 inflammatory arthritis individuals and examine the joints with the invasive methods such as arthroscopic surgery and histology^[26]. It was observed that tracer molecule accumulation was higher in inflamed joints as compared to non- inflamed joints of the same individual. The uptake of tracer molecule correlated well with expression of benzodiazepine receptor and CD68 expression on macrophages in the synovium. Being highly sensitive and ability to capture multiple joints simultaneously, PET scan may be used to pick up sub clinical synovitis. Recently, PET analysis with 18F FDG tracer molecule was conducted in 18 inflammatory arthritis patients; four of them were in disease remission status^[27]. 18F FDG uptake was significantly different between patients with active RA vs patients in remission. Moreover, it has been reported that 18F FDG uptake varied with alterations in CRP and matrix metalloproteinase 3 levels in patients with RA receiving anti-TNF therapy^[28,29]. However, as radiation amount of a PET-CT is high, it cannot be recommended for routine screening of RA patients in clinical practice. To overcome this problem, a new technique, PET MRI is being developed^[30]. Currently, there is no data available regarding its role in evaluation of synovial inflammation. Synovial phagocytes cell were targeted with tracer molecule PK11195 in PET scanning to identify early synovial inflammation in 24 anticyclic citrullinated peptide antibody positive patients presenting with arthralgias only. PET scan was focused on hand joints only. Four patients were detected to have PET positive joints at baseline and all four developed RA within next 2 years. Amongst rest, five more developed RA but had negative PET scan at baseline. Among these five two developed arthritis of hand joints and rest three developed arthritis outside the field of view of PET scanner^[31].

CT SCANNING

Compared to other imaging approaches very few CT scan based investigations are available in the context of synovial inflammation. It may assist in diagnosis of several different kinds of inflammatory arthritides. Similar to conventional X-ray, it very well demonstrates the cortical bone architecture and is considered as the gold standard for the imaging of erosions in the joints against which other imaging techniques are evaluated^[32-34]. Multidetector CT generates quite excellent images which can be saved in a digital format and can be utilized to compare progression of the joint erosions during follow up. CT scan out-performed MRI in evaluating joint erosions at wrists in RA^[34]. Similar data was reported by Døhn *et al*^[35,36] for erosions at the metacarpophalangeal (MCP) joints during serial monitoring of patients of RA being treated with anti TNF- α therapy. CT scanning exposes to ionising rays but the influence of this is comparatively minimal as only the extremities are analysed. The major drawback of CT scanning is limited coverage of joint areas as compared to conventional X-ray imaging^[37].

Micro focal CT is a better image quality strategy which enables assessment of bone mineral density. This technique has been used for the analysis of erosions in RA. In a study it was observed that small erosions in joints were seen in both controls and RA subjects, but erosions > 1.9 mm diameter were specific for RA. Using this very technique it was reported that anti interleukin 6 receptor monoclonal antibody (tocilizumab) could repair bone erosions and has favourable effect on bone remodelling in RA^[38,39].

CT osteoabsorptiometry is another imaging strategy which has been utilized to evaluate periarticular osteopaenia in early inflammatory arthritis. It was observed that RA patients had significantly less mineralization at MCP joints compared to control subjects^[40]. Volumetric bone mineral density evaluated with the use of a quantitative CT (high resolution-peripheral quantitative CT, HR pQCT) system confirmed the involvement of trabecular bone compartment in the peri-articular osteopenia^[41]. Presently, all these newer CT scan techniques are only at research stages and are less likely to be available for routine clinical practice in near future.

Baishideng®

MRI IN SYNOVITIS

MRI is one of the most sensitive methods available to evaluate cartilage, synovium and bone tissue changes in the joint. MRI-based quantification of synovial thickening and synovial fluid volume indicate disease activity. The signal strength related with synovial thickening is intermediate to minimal on T1 weighted image, but higher on T2 weighted image due to excessive water of synovial fluid within the synovium and reflects the degree of inflammation^[42]. Synovial inflammation is further enhanced on T2 weighted image on MRI scanning. Differentiation of synovial inflammation from synovial fluid without using contrast agent is challenging; however, heavily T2 weighted images can differentiate between the two. Compared to joint effusion inflamed synovium has lower signal intensity on T2-weighted image^[43]. Contrast based T1 enhancement on MR imaging with kinetic study is helpful in differentiation of effusion from inflamed synovium^[44,45]. Gadolinium based contrast medium enhances inflamed synovium soon after administration however, it rapidly diffuses into synovial fluid compartment, resulting in equal signal strengths between synovium and the synovial fluid. This rate of equilibration of signals between the synovium membrane and the synovial fluid compartment may indicate the degree of leakiness of inflamed vessels in the synovium and thus intensity of inflammation^[45].

Dynamic contrast enhanced MRI in synovitis progression

Dynamic contrast enhanced MRI (DCE MRI) is an imaging technique which is used for evaluation of the pharmacokinetic factors relevant to the exchange of contrast material between intravascular and extra vascular spaces and indicate the presence of new blood vessels (neo angiogenesis) in inflamed joint. T1 weighted MRI images are obtained prior to and after administration of a T1-shortening diffusible contrast agent. Post contrast time intensity curve delineating the concentration of the contrast agent in the areas of synovium reflects the intensity of inflammation in the synovium. Information obtained from the time intensity curve can be examined semi quantitatively or quantitatively with the Toft's model based software. In the semi quantitative analysis, factors that define the form of the time intensity curve, for example uptake of contrast agent, maximum enhancement and wash out ratio are calculated^[46-49]. The degree of synovial inflammation is quantified by pharmacokinetic model by plotting time intensity curve which analyses diffusion of contrast agent from the vascular compartment to the extra vascular extracellular compartment^[46]. The rate of exchange of contrast agent between these two compartments and amount of contrast agent in the extracellular spaces depend upon the perfusion and leakiness of the blood vessels in the synovium. DCE MRI is being progressively utilized for the recognition of synovial inflammation in early inflammatory arthritis. DCE MRI results have been correlated strongly with histological severity of inflamed knee synovium. Not only this, it

was further utilized to monitor reduction in inflammation following intra articular steroid injection^[50-52]. Another study reported that many RA patients display enhanced capillary leak and vascularisation of the synovium^[53]. Efficacy of antiTNFa treatment in RA patients was observed by decrease in various DCE MRI parameters such as enhanced T1 relaxation time, volume transfer constant and fractional blood volume. Furthermore, volume transfer constant (kp) has been reported to be a good marker of vascularity in the synovium^[49]. In other studies in RA patients, color coded DCE MRI parameters such as; micro vessels density and permeability were overlapped over anatomical MRI images, it was observed that the distribution of information produced by kps and fractional blood volume computations matches the qualitative evaluation of signal strength on post contrast T1 images^[46,49]. Analyses of qualitative readings of colour coded parametric images were reliable and consistent on several image acquisitions of the same individual at different time intervals. Hence, DCE MRI represents a highly efficient technique which has potential to be noninvasive imaging biomarker for evaluation of alterations in vascularity of inflamed synovium. It can be utilized for monitoring efficacy of disease-modifying anti-rheumatic drugs (DMARD) and biologic therapies in RA and other inflammatory arthritis.

Advantages and disadvantages of various imaging techniques have been listed in Table 1.

Quantify synovial inflammation with diffusion tensor imaging

Diffusion tensor image (DTI) is a non contrast based image strategy that quantifies diffusion of fluid in vivo as well as provides details of the tissues at microscopic level^[54,55]. This imaging strategy has been used in the evaluation of structure of organised tissues such as neural tissues of the brain, heart muscle tissues and intervertebral disc^[56]. Because of presence of the cell membranes and other elements in in vivo system, diffusion of water molecules is restricted and these arrange themselves along a particular direction, along the length of tissue components rather than perpendicularly, on application of strong magnetic field. Restricted movement of water molecules in a particular direction is called as anisotropic diffusion. Anisotropy of water molecule can be utilized to obtain details about the cells organization at microscopic level^[57]. Diffusion of normal fluid is isotropic and it can be analyzed with only one diffusion parameter but in biological tissue anisotropic diffusion can be described by a 3×3 symmetric matrix. In biological tissues, complete diffusion matrix can be computed by calculating 6 independent matrix elements. The most widely used scalar indices that are based on DTI are the fractional anisotropy (FA) and mean diffusivity (MD)^[58-60]. FA is a measure of diffusion anisotropy and its minimum value "0" represents isotropic diffusion, *i.e.*, equal probability of diffusion in all directions. Whereas a maximum value of "1" represents highly restricted diffusion such as very thin fibres. MD on the other hand represents mean of



Imaging technique for quantification of synovial inflammation	Advantages	Limitations
Radiography	Cost effective,	Exposure to ionising radiation, which although relatively
	Ease of access,	low for one set of X-rays can cumulate over time with a
	Wide coverage of important joint regions, newer digitised formats	potential impact on patient longevity
	that allow easy retrieval and comparison of images longitudinally	Lack of sensitivity for detecting early joint damage and
	and relative low cost	inability to image the inflammatory processes within the joint that precede damage
		Very little information regarding severity of inflammation Lacks quantifiable variables of inflammation in early arthritis
Ultrasonography	It is sensitive, cost effective, can be performed by the treating physician in out-patient-department basis, and can be repeated as	Normal anatomical structures may have low reflectivity and mimic synovitis if careful attention is not paid to
	desired for serial monitoring of inflammation	technique, particularly with lower resolution equipment
	desired for serial monitoring of inflammation	
Positron	Sensitive, able to assess inflammation at molecular level	Operator dependent
emission tomography	Used for imaging sub-clinical synovitis because of their sensitivity	Costly, Still experimental No evidences available in early synovitis. Not available
emission tomography	and ability to capture many joints	at many centres and not being used in day to day clinical
	and ability to capture many joints	practices
		High radiation exposure
Computed tomography	Cost effective	Computed tomography scan upon whole body scanning
scanning	Multidetector helical CT produces very high-quality images which	leads to high dose radiation exposure
0	can be stored in a digitised format and compared with later images	It does provide less coverage than plain radiography as
	to determine erosion progression Gold standard for imaging of bone erosions	usually only one joint area is scanned
Magnetic	Sensitive techniques to assess soft tissue and bone changes in the joints	Time consuming,
resonance imaging	Very sensitive for the detection of early synovial inflammation	Relatively costly
	Diffusion Tensor based imaging can evaluate molecular event during synovial inflammation without contrast medium	Contrast based enhancement required for dynamic study

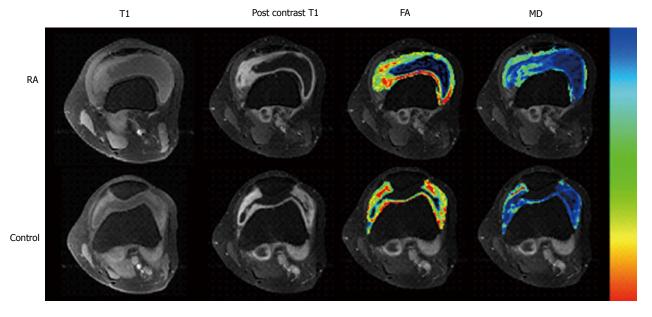


Figure 1 A 45-year-old male rheumatoid arthritis synovium shows strong enhancement on fat suppressed post-contrast T1-weighted axial image. Color coded fractional anisotropy (FA = 0.26) map from segmented region of enhanced synovial membrane overlaid on post-contrast fat-suppressed T1-weighted image show increased FA as compared to control (FA = 0.19). Mean diffusivity being inversely related to FA showed low values (1.01 × 10⁻³ mm²/s). Color red denotes increased whereas blue denotes decreased values. RA: Rheumatoid arthritis; MD: Mean diffusivity; FA: Fractional anisotropy.

molecular motion independent of direction of the tissue. It depends upon the size and integrity of the cells.

We have earlier evaluated DTI parameters to assess severity of synovial inflammation in eighteen RA patients and six healthy individuals. Considerably significant high

FA and reduced mean diffusivity were seen in RA individuals in comparison to controls (Figure 1). In this study we found a strong association between FA and synovial liquid IL-1 β and TNF- α levels^[61]. We also observed a positive correlation between cylindrical isotropy and sICAM which suggested that the adhered inflammatory molecules on synovium represent the planar model of diffusion tensor. This led us to speculate that limited movement of water molecule in the synovium of inflammatory arthritis patients was a consequence of inflammatory cell infiltration and aggregation^[62]. It has been suggested that this technique may replace synovial histology to evaluate the severity of inflammation and assess efficacy of disease modifying drug therapy^[63].

CONCLUSION

Various newer imaging techniques are being utilized to explore the pathogenesis of synovial inflammation and soft tissue disruptions that occur in various inflammatory and damaging arthritides. The new emerging techniques have capability to quantify synovial inflammation and vascularity and modifications in cartilage biochemistry as well. A combination of DTI with DCE MRI may capture microscopic features of inflammation such as cellular infiltration and increased vascularity and may emerge as powerful tools to evaluate severity of inflammation at the level of synovium. These imaging strategies are important for analyzing the clinical effectiveness of DMARDs and biologics. Developments in imaging techniques, such as the miniaturization of extremity magnet for DCE and DTI MRI, and automated software programs may make these techniques available for routine clinical usage.

REFERENCES

- 1 **Haywood L**, Walsh DA. Vasculature of the normal and arthritic synovial joint. *Histol Histopathol* 2001; **16**: 277-284 [PMID: 11193203]
- 2 Hoeben A, Landuyt B, Highley MS, Wildiers H, Van Oosterom AT, De Bruijn EA. Vascular endothelial growth factor and angiogenesis. *Pharmacol Rev* 2004; 56: 549-580 [PMID: 15602010 DOI: 10.1124/pr.56.4.3]
- 3 Quan LD, Thiele GM, Tian J, Wang D. The Development of Novel Therapies for Rheumatoid Arthritis. *Expert Opin Ther Pat* 2008; 18: 723-738 [PMID: 19578469 DOI: 10.1517/1354377 6.18.7.723]
- 4 Little MP, Tawn EJ, Tzoulaki I, Wakeford R, Hildebrandt G, Paris F, Tapio S, Elliott P. Review and meta-analysis of epidemiological associations between low/moderate doses of ionizing radiation and circulatory disease risks, and their possible mechanisms. *Radiat Environ Biophys* 2010; **49**: 139-153 [PMID: 19862545 DOI: 10.1007/s00411-009-0250-z]
- 5 McQueen FM. Magnetic resonance imaging in early inflammatory arthritis: what is its role? *Rheumatology* (Oxford) 2000; **39**: 700-706 [PMID: 10908686 DOI: 10.1093/rheumatology/39.7.700]
- 6 Niewold TB, Harrison MJ, Paget SA. Anti-CCP antibody testing as a diagnostic and prognostic tool in rheumatoid arthritis. *QJM* 2007; 100: 193-201 [PMID: 17434910 DOI: 10.1093/qjmed/hcm015]
- 7 Forslind K, Ahlmén M, Eberhardt K, Hafström I, Svensson B. Prediction of radiological outcome in early rheumatoid arthritis in clinical practice: role of antibodies to citrullinated peptides (anti-CCP). Ann Rheum Dis 2004; 63: 1090-1095 [PMID: 15308518 DOI: 10.1136/ard.2003.014233]
- 8 Maini RN, Breedveld FC, Kalden JR, Smolen JS, Furst D, Weisman MH, St Clair EW, Keenan GF, van der Heijde D, Marsters PA, Lipsky PE. Sustained improvement over two

years in physical function, structural damage, and signs and symptoms among patients with rheumatoid arthritis treated with infliximab and methotrexate. *Arthritis Rheum* 2004; **50**: 1051-1065 [PMID: 15077287 DOI: 10.1002/art.20159]

- 9 McAlindon T, Kissin E, Nazarian L, Ranganath V, Prakash S, Taylor M, Bannuru RR, Srinivasan S, Gogia M, McMahon MA, Grossman J, Kafaja S, FitzGerald J. American College of Rheumatology report on reasonable use of musculoskeletal ultrasonography in rheumatology clinical practice. *Arthritis Care Res* (Hoboken) 2012; 64: 1625-1640 [PMID: 23111854 DOI: 10.1002/acr.21836]
- 10 Le Boedec M, Jousse-Joulin S, Ferlet JF, Marhadour T, Chales G, Grange L, Hacquard-Bouder C, Loeuille D, Sellam J, Albert JD, Bentin J, Chary-Valckenaere I, D'Agostino MA, Etchepare F, Gaudin P, Hudry C, Dougados M, Saraux A. Factors influencing concordance between clinical and ultrasound findings in rheumatoid arthritis. *J Rheumatol* 2013; **40**: 244-252 [PMID: 23322464 DOI: 10.3899/jrheum.120843]
- 11 Brown AK. Using ultrasonography to facilitate best practice in diagnosis and management of RA. *Nat Rev Rheumatol* 2009; 5: 698-706 [PMID: 19901917 DOI: 10.1038/nrrheum.2009.227]
- 12 Alcalde M, D'Agostino MA, Bruyn GA, Möller I, Iagnocco A, Wakefield RJ, Naredo E. A systematic literature review of US definitions, scoring systems and validity according to the OMERACT filter for tendon lesion in RA and other inflammatory joint diseases. *Rheumatology* (Oxford) 2012; 51: 1246-1260 [PMID: 22378717 DOI: 10.1093/rheumatology/ kes018]
- 13 Cooperberg PL, Tsang I, Truelove L, Knickerbocker WJ. Gray scale ultrasound in the evaluation of rheumatoid arthritis of the knee. *Radiology* 1978; 126: 759-763 [PMID: 628753 DOI: 10.1148/126.3.759]
- 14 Halpern EJ. Contrast-enhanced ultrasound imaging of prostate cancer. *Rev Urol* 2006; 8 Suppl 1: S29-S37 [PMID: 17021624]
- 15 Cohen GS, Braunstein L, Ball DS, Roberto PJ, Reich J, Hanno P. Selective arterial embolization of idiopathic priapism. *Cardiovasc Intervent Radiol* 1996; **19**: 47-49 [PMID: 8653747]
- 16 Wakefield RJ, Balint PV, Szkudlarek M, Filippucci E, Backhaus M, D'Agostino MA, Sanchez EN, Iagnocco A, Schmidt WA, Bruyn GA, Kane D, O'Connor PJ, Manger B, Joshua F, Koski J, Grassi W, Lassere MN, Swen N, Kainberger F, Klauser A, Ostergaard M, Brown AK, Machold KP, Conaghan PG. Musculoskeletal ultrasound including definitions for ultrasonographic pathology. J Rheumatol 2005; 32: 2485-2487 [PMID: 16331793]
- 17 McNally EG. Ultrasound of the small joints of the hands and feet: current status. *Skeletal Radiol* 2008; **37**: 99-113 [PMID: 17712556 DOI: 10.1007/s00256-007-0356-9]
- 18 Newman JS, Adler RS, Bude RO, Rubin JM. Detection of soft-tissue hyperemia: value of power Doppler sonography. *AJR Am J Roentgenol* 1994; 163: 385-389 [PMID: 8037037 DOI: 10.2214/ajr.163.2.8037037]
- 19 Porta F, Radunovic G, Vlad V, Micu MC, Nestorova R, Petranova T, Iagnocco A. The role of Doppler ultrasound in rheumatic diseases. *Rheumatology* (Oxford) 2012; 51: 976-982 [PMID: 22253027 DOI: 10.1093/rheumatology/ker433]
- 20 Andersen M, Ellegaard K, Hebsgaard JB, Christensen R, Torp-Pedersen S, Kvist PH, Søe N, Rømer J, Vendel N, Bartels EM, Danneskiold-Samsøe B, Bliddal H. Ultrasound colour Doppler is associated with synovial pathology in biopsies from hand joints in rheumatoid arthritis patients: a cross-sectional study. *Ann Rheum Dis* 2014; **73**: 678-683 [PMID: 23475981 DOI: 10.1136/annrheumdis-2012-202669]
- 21 Wakefield TW, Strieter RM, Schaub R, Myers DD, Prince MR, Wrobleski SK, Londy FJ, Kadell AM, Brown SL, Henke PK, Greenfield LJ. Venous thrombosis prophylaxis by inflammatory inhibition without anticoagulation therapy. J Vasc Surg 2000; **31**: 309-324 [PMID: 10664500]
- 22 Zeman MN, Scott PJ. Current imaging strategies in rheu-

matoid arthritis. *Am J Nucl Med Mol Imaging* 2012; **2**: 174-220 [PMID: 23133812]

- 23 Palmer WE, Rosenthal DI, Schoenberg OI, Fischman AJ, Simon LS, Rubin RH, Polisson RP. Quantification of inflammation in the wrist with gadolinium-enhanced MR imaging and PET with 2-[F-18]-fluoro-2-deoxy-D-glucose. *Radiology* 1995; **196**: 647-655 [PMID: 7644624 DOI: 10.1148/radiology.196.3.7644624]
- 24 Elzinga EH, van der Laken CJ, Comans EF, Lammertsma AA, Dijkmans BA, Voskuyl AE. 2-Deoxy-2-[F-18]fluoro-D-glucose joint uptake on positron emission tomography images: rheumatoid arthritis versus osteoarthritis. *Mol Imaging Biol* 2007; **9**: 357-360 [PMID: 17902022 DOI: 10.1007/ s11307-007-0113-4]
- 25 Goerres GW, Forster A, Uebelhart D, Seifert B, Treyer V, Michel B, von Schulthess GK, Kaim AH. F-18 FDG whole-body PET for the assessment of disease activity in patients with rheumatoid arthritis. *Clin Nucl Med* 2006; **31**: 386-390 [PMID: 16785804 DOI: 10.1097/01.rlu.0000222678.95218.42]
- 26 van der Laken CJ, Elzinga EH, Kropholler MA, Molthoff CF, van der Heijden JW, Maruyama K, Boellaard R, Dijkmans BA, Lammertsma AA, Voskuyl AE. Noninvasive imaging of macrophages in rheumatoid synovitis using 11C-(R)-PK11195 and positron emission tomography. *Arthritis Rheum* 2008; 58: 3350-3355 [PMID: 18975347 DOI: 10.1002/art.23955]
- 27 Kubota K, Ito K, Morooka M, Mitsumoto T, Kurihara K, Yamashita H, Takahashi Y, Mimori A. Whole-body FDG-PET/CT on rheumatoid arthritis of large joints. *Ann Nucl Med* 2009; 23: 783-791 [PMID: 19834653 DOI: 10.1007/s12149-009-0305-x]
- 28 Beckers C, Jeukens X, Ribbens C, André B, Marcelis S, Leclercq P, Kaiser MJ, Foidart J, Hustinx R, Malaise MG. (18)F-FDG PET imaging of rheumatoid knee synovitis correlates with dynamic magnetic resonance and sonographic assessments as well as with the serum level of metalloproteinase-3. *Eur J Nucl Med Mol Imaging* 2006; **33**: 275-280 [PMID: 16247604 DOI: 10.1007/s00259-005-1952-3]
- 29 van der Laken CJ, Huisman MH, Voskuyl AE. Nuclear imaging of rheumatic diseases. *Best Pract Res Clin Rheumatol* 2012; 26: 787-804 [PMID: 23273792 DOI: 10.1016/j.berh.2012.10.006]
- 30 Miese F, Scherer A, Ostendorf B, Heinzel A, Lanzman RS, Kröpil P, Blondin D, Hautzel H, Wittsack HJ, Schneider M, Antoch G, Herzog H, Shah NJ. Hybrid 18F-FDG PET-MRI of the hand in rheumatoid arthritis: initial results. *Clin Rheumatol* 2011; **30**: 1247-1250 [PMID: 21590292 DOI: 10.1007/ s10067-011-1777-3]
- 31 Gent YY, Voskuyl AE, Kloet RW, van Schaardenburg D, Hoekstra OS, Dijkmans BA, Lammertsma AA, van der Laken CJ. Macrophage positron emission tomography imaging as a biomarker for preclinical rheumatoid arthritis: findings of a prospective pilot study. *Arthritis Rheum* 2012; **64**: 62-66 [PMID: 21898356 DOI: 10.1002/art.30655]
- 32 Canella C, Philippe P, Pansini V, Salleron J, Flipo RM, Cotten A. Use of tomosynthesis for erosion evaluation in rheumatoid arthritic hands and wrists. *Radiology* 2011; 258: 199-205 [PMID: 21045184 DOI: 10.1148/radiol.10100791]
- 33 Døhn UM, Ejbjerg BJ, Hasselquist M, Narvestad E, Court-Payen M, Szkudlarek M, Møller J, Thomsen HS, Ostergaard M. Rheumatoid arthritis bone erosion volumes on CT and MRI: reliability and correlations with erosion scores on CT, MRI and radiography. *Ann Rheum Dis* 2007; 66: 1388-1392 [PMID: 17606464 DOI: 10.1136/ard.2007.072520]
- 34 Perry D, Stewart N, Benton N, Robinson E, Yeoman S, Crabbe J, McQueen F. Detection of erosions in the rheumatoid hand; a comparative study of multidetector computerized tomography versus magnetic resonance scanning. J Rheumatol 2005; 32: 256-267 [PMID: 15693085]
- 35 **Døhn UM**, Ejbjerg BJ, Court-Payen M, Hasselquist M, Narvestad E, Szkudlarek M, Møller JM, Thomsen HS, Østergaard M. Are bone erosions detected by magnetic resonance

imaging and ultrasonography true erosions? A comparison with computed tomography in rheumatoid arthritis metacarpophalangeal joints. *Arthritis Res Ther* 2006; **8**: R110 [PMID: 16848914 DOI: 10.1186/ar1995]

- 36 Døhn UM, Ejbjerg B, Boonen A, Hetland ML, Hansen MS, Knudsen LS, Hansen A, Madsen OR, Hasselquist M, Møller JM, Ostergaard M. No overall progression and occasional repair of erosions despite persistent inflammation in adalimumab-treated rheumatoid arthritis patients: results from a longitudinal comparative MRI, ultrasonography, CT and radiography study. *Ann Rheum Dis* 2011; **70**: 252-258 [PMID: 20980282 DOI: 10.1136/ard.2009.123729]
- 37 Knevel R, Kwok KY, de Rooy DP, Posthumus MD, Huizinga TW, Brouwer E, van der Helm-van Mil AH. Evaluating joint destruction in rheumatoid arthritis: is it necessary to radiograph both hands and feet? *Ann Rheum Dis* 2013; **72**: 345-349 [PMID: 22580587 DOI: 10.1136/annrheumdis-2012-201391]
- 38 Stach CM, Bäuerle M, Englbrecht M, Kronke G, Engelke K, Manger B, Schett G. Periarticular bone structure in rheumatoid arthritis patients and healthy individuals assessed by high-resolution computed tomography. *Arthritis Rheum* 2010; 62: 330-339 [PMID: 20112404]
- 39 Finzel S, Rech J, Schmidt S, Engelke K, Englbrecht M, Schett G. Interleukin-6 receptor blockade induces limited repair of bone erosions in rheumatoid arthritis: a micro CT study. Ann Rheum Dis 2013; 72: 396-400 [PMID: 22586162 DOI: 10.1136/ annrheumdis-2011-201075]
- 40 Meirer R, Müller-Gerbl M, Huemer GM, Schirmer M, Herold M, Kersting S, Freund MC, Rainer C, Gardetto A, Wanner S, Piza-Katzer H. Quantitative assessment of periarticular osteopenia in patients with early rheumatoid arthritis: a preliminary report. *Scand J Rheumatol* 2004; **33**: 307-311 [PMID: 15513678 DOI: 10.1080/03009740410005890]
- 41 **Fouque-Aubert A**, Boutroy S, Marotte H, Vilayphiou N, Bacchetta J, Miossec P, Delmas PD, Chapurlat RD. Assessment of hand bone loss in rheumatoid arthritis by high-resolution peripheral quantitative CT. *Ann Rheum Dis* 2010; **69**: 1671-1676 [PMID: 20525847 DOI: 10.1136/ard.2009.114512]
- 42 Narváez JA, Narváez J, De Lama E, De Albert M. MR imaging of early rheumatoid arthritis. *Radiographics* 2010; 30: 143-163; discussion 163-165 [PMID: 20083591 DOI: 10.1148/ rg.301095089]
- 43 Narváez JA, Narváez J, Roca Y, Aguilera C. MR imaging assessment of clinical problems in rheumatoid arthritis. *Eur Radiol* 2002; 12: 1819-1828 [PMID: 12111074 DOI: 10.1007/ s00330-001-1207-z]
- 44 Ostergaard M, Ejbjerg B. Magnetic resonance imaging of the synovium in rheumatoid arthritis. *Semin Musculoskelet Radiol* 2004; 8: 287-299 [PMID: 15643570 DOI: 10.1055/ s-2004-861576]
- 45 Rand T, Imhof H, Czerny C, Breitenseher M, Machold K, Turetschek K, Trattnig S. Discrimination between fluid, synovium, and cartilage in patients with rheumatoid arthritis: contrast enhanced Spin Echo versus non-contrast-enhanced fat-suppressed Gradient Echo MR imaging. *Clin Radiol* 1999; 54: 107-110 [PMID: 10050739 DOI: 10.1016/S0009-9260(99)91070-X]
- 46 van der Leij C, van de Sande MG, Lavini C, Tak PP, Maas M. Rheumatoid synovial inflammation: pixel-by-pixel dynamic contrast-enhanced MR imaging time-intensity curve shape analysis--a feasibility study. *Radiology* 2009; 253: 234-240 [PMID: 19703863 DOI: 10.1148/radiol.2531081722]
- 47 Hodgson R, Grainger A, O'Connor P, Barnes T, Connolly S, Moots R. Dynamic contrast enhanced MRI of bone marrow oedema in rheumatoid arthritis. *Ann Rheum Dis* 2008; **67**: 270-272 [PMID: 17965120 DOI: 10.1136/ard.2007.077271]
- 48 Hodgson RJ, Barnes T, Connolly S, Eyes B, Campbell RS, Moots R. Changes underlying the dynamic contrastenhanced MRI response to treatment in rheumatoid arthritis. *Skeletal Radiol* 2008; 37: 201-207 [PMID: 18058095 DOI:

10.1007/s00256-007-0408-1]

- 49 Zierhut ML, Gardner JC, Spilker ME, Sharp JT, Vicini P. Kinetic modeling of contrast-enhanced MRI: an automated technique for assessing inflammation in the rheumatoid arthritis wrist. *Ann Biomed Eng* 2007; 35: 781-795 [PMID: 17340197 DOI: 10.1007/s10439-006-9249-7]
- 50 Boesen M, Østergaard M, Cimmino MA, Kubassova O, Jensen KE, Bliddal H. MRI quantification of rheumatoid arthritis: current knowledge and future perspectives. *Eur J Radiol* 2009; **71**: 189-196 [PMID: 19477615 DOI: 10.1016/ j.ejrad.2009.04.048]
- 51 Kubassova O, Boesen M, Cimmino MA, Bliddal H. A computer-aided detection system for rheumatoid arthritis MRI data interpretation and quantification of synovial activity. *Eur J Radiol* 2010; 74: e67-e72 [PMID: 19411154 DOI: 10.1016/j.ejrad.2009.04.010]
- 52 Peloschek P, Boesen M, Donner R, Kubassova O, Birngruber E, Patsch J, Mayerhöfer M, Langs G. Assessement of rheumatic diseases with computational radiology: current status and future potential. *Eur J Radiol* 2009; **71**: 211-216 [PMID: 19457632 DOI: 10.1016/j.ejrad.2009.04.046]
- 53 Ostergaard M, Stoltenberg M, Løvgreen-Nielsen P, Volck B, Sonne-Holm S, Lorenzen I. Quantification of synovistis by MRI: correlation between dynamic and static gadoliniumenhanced magnetic resonance imaging and microscopic and macroscopic signs of synovial inflammation. *Magn Reson Imaging* 1998; 16: 743-754 [PMID: 9811140 DOI: 10.1016/S0730-725X(98)00008-3]
- 54 Pierpaoli C, Jezzard P, Basser PJ, Barnett A, Di Chiro G. Diffusion tensor MR imaging of the human brain. *Radiology* 1996; 201: 637-648 [PMID: 8939209 DOI: 10.1148/radiology.201.3.8939209]
- 55 **Chen J**, Liu W, Zhang H, Lacy L, Yang X, Song SK, Wickline SA, Yu X. Regional ventricular wall thickening reflects changes in cardiac fiber and sheet structure during contraction: quantification with diffusion tensor MRI. *Am J Physiol*

Heart Circ Physiol 2005; **289**: H1898-H1907 [PMID: 16219812 DOI: 10.1152/ajpheart.00041.2005]

- 56 Hsu EW, Setton LA. Diffusion tensor microscopy of the intervertebral disc anulus fibrosus. *Magn Reson Med* 1999; 41: 992-999 [PMID: 10332883]
- 57 Le Bihan D, Mangin JF, Poupon C, Clark CA, Pappata S, Molko N, Chabriat H. Diffusion tensor imaging: concepts and applications. *J Magn Reson Imaging* 2001; 13: 534-546 [PMID: 11276097 DOI: 10.1002/jmri.1076]
- 58 Hasan KM, Parker DL, Alexander AL. Comparison of gradient encoding schemes for diffusion-tensor MRI. J Magn Reson Imaging 2001; 13: 769-780 [PMID: 11329200 DOI: 10.1002/ jmri.1107]
- 59 Hasan KM, Alexander AL, Narayana PA. Does fractional anisotropy have better noise immunity characteristics than relative anisotropy in diffusion tensor MRI? An analytical approach. *Magn Reson Med* 2004; **51**: 413-417 [PMID: 14755670 DOI: 10.1002/mrm.10682]
- 60 Pfefferbaum A, Sullivan EV. Disruption of brain white matter microstructure by excessive intracellular and extracellular fluid in alcoholism: evidence from diffusion tensor imaging. *Neuropsychopharmacology* 2005; 30: 423-432 [PMID: 15562292 DOI: 10.1038/sj.npp.1300623]
- 61 Agarwal V, Kumar M, Singh JK, Rathore RK, Misra R, Gupta RK. Diffusion tensor anisotropy magnetic resonance imaging: a new tool to assess synovial inflammation. *Rheumatology* (Oxford) 2009; 48: 378-382 [PMID: 19174567 DOI: 10.1093/rheumatology/ken499]
- 62 Gupta RK, Nath K, Prasad A, Prasad KN, Husain M, Rathore RK, Husain N, Srivastava C, Khetan P, Trivedi R, Narayana PA. In vivo demonstration of neuroinflammatory molecule expression in brain abscess with diffusion tensor imaging. *AJNR Am J Neuroradiol* 2008; 29: 326-332 [PMID: 17989372 DOI: 10.3174/ajnr.A0826]
- 63 **Buckland J**. Bye-bye biopsy? *Nat Rev Rheumatol* 2009; **5**: 236 [DOI: 10.1038/nrrheum.2009.36]

P-Reviewer: Bourgoin SG, Sakkas L S- Editor: Ji FF L- Editor: A E- Editor: Wu HL







Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.5499/wjr.v4.i3.80 World J Rheumatol 2014 November 12; 4(3): 80-87 ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

MINIREVIEWS

Does a biological link exist between periodontitis and rheumatoid arthritis?

Rosamma Joseph, MG Jose Raj, Shobha Sundareswaran, Priyanka Chand Kaushik, Amol Vijay Nagrale, Susan Jose, Sreeraj Rajappan

Rosamma Joseph, Priyanka Chand Kaushik, Amol Vijay Nagrale, Sreeraj Rajappan, Department of Periodontics, Government Dental College, Kottayam 686008, Kerala, India

MG Jose Raj, Department of Biochemistry, Government Medical College, Calicut 673008, Kerala, India

Shobha Sundareswaran, Department of Orthodontics, Government Dental College, Calicut 673008, Kerala, India

Susan Jose, Medical College Campus, Calicut 673008, Kerala, India

Author contributions: All authors have contributed to literature search, intellectual content, drafting and editing of the manuscript and language revision; all authors approved the final version to be published.

Correspondence to: Dr. Rosamma Joseph, Professor and Head, Department of Periodontics, Government Dental College, Medical College PO, Calicut 673008, Kerala,

India. drrosammajoseph@gmail.com

 Telephone: +91-944-6070599
 Fax: +91-495-2356781

 Received: June 28, 2014
 Revised: September 25, 2014

 Accepted: October 14, 2014
 Published online: November 12, 2014

Abstract

Periodontitis or Periodontal disease (PD) and Rheumatoid arthritis (RA) are two the most common chronic inflammatory diseases. Periodontitis is a biofilm associated destructive inflammatory disease of the periodontium caused by specific microorganisms. Rheumatoid arthritis is an autoimmune condition and is identified by elevated serum autoantibody titre directed against citrullinated peptides or rheumatoid factor. Periodontitis may involve some elements of autoimmunity. Recent studies have established that PD and RA show a common pathway and could be closely associated through a common dysregulation and dysfunction in inflammatory mechanism. The enzyme peptidyl arginine deiminase (PAD), expressed by Porphyromonas gingivalis (P. gingivalis) is responsible for the enzymatic deimination of arginine residuals to citrulline resulting in protein citrullination and its increased accumulation in RA.

Citrullination by PAD may act as a putative biologic link between PD and RA. Association of Human leukocytic antigen-DR4 antigen has been established both with RA and PD. Several interleukins and inflammatory mediators (ILs) and Nuclear factor kappa beta ligand are linked to these common chronic inflammatory diseases. Antibodies directed against heat shock protein (hsp 70 ab) of P. gingivalis, P. melanogenicus and P. intermedia are raised in PD as well as RA. Both the conditions share many pathological and immunological similarities. Bacterial infection, genetic susceptibility, altered immune reaction and inflammatory mediators considered responsible for RA are also associated with PD. So it is plausible that a biological link may exist between PD and RA. Therapies aimed at modifying the expression and effect of inflammatory mediators and effector molecules such as matrix metalloproteinases, proinflammatory cytokines and autoantibodies of structural proteins may probably reduce the severity of both RA and PD.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Periodontal disease; Rheumatoid arthritis; Citrullinated peptidase; *Porphyromonas gingivalis*; Inflammatory marker; Inflammation and autoantibody

Core tip: Periodontal disease (PD) and Rheumatoid arthritis (RA) share many pathological and immunological similarities. Recent studies have established significant association between the two. Bacterial infection, genetic susceptibility, altered immune reaction and inflammatory mediators considered responsible for RA are also associated with PD. So it is plausible that a biological link may exist between PD and RA. Therapies aimed at reduction of inflammatory mediators and effector molecules can probably reduce the severity of both RA and PD.

Joseph R, Jose Raj MG, Sundareswaran S, Kaushik PC, Nagrale AV, Jose S, Rajappan S. Does a biological link exist between periodontitis and rheumatoid arthritis? *World J Rheumatol* 2014; 4(3):



80-87 Available from: URL: http://www.wjgnet.com/2220-3214/ full/v4/i3/80.htm DOI: http://dx.doi.org/10.5499/wjr.v4.i3.80

INTRODUCTION

Periodontal disease (PD) is an immuno inflammatory disease of the periodontium which comprises of both hard and soft tissues like gingiva, periodontal ligament, cementum and alveolar bone. It results from a complex interaction between gram negative organisms, their byproducts and the response of the host^[1-4]. The resulting gingival inflammation leads to destruction of both the soft and hard tissues supporting the tooth^[5]. The prevalence is said to be as high as 80% to 90%^[6].

For periodontal tissues in a healthy state, a steady equilibrium exists between tissue destruction and repair. Periodontal destruction is initiated and progressed by specific periodontal microorganisms that colonize in plaque biofilm. Host microbial interaction determines the extent and severity of periodontal disease^[7-9]. A large number of different species of bacteria can be identified in the dental plaque^[10] but only a few of them are implicated in chronic periodontitis^[11,12]. To prevent exacerbated reactions and destruction of host tissues, an appropriate tolerance mechanism is required by the host to recognize and identify nonpathogenic and pathogenic bacteria^[13]. Pattern recognition receptors and microbe associated molecular patterns have a very significant role in periodontal inflammation and adaptive immune response^[13,14].

The equilibrium established between anti-inflammatory and proinflammatory cytokines (IL-1 α , IL-1 β , TNF- α , IL-6^[15], IL-7, IL-11, IL-17A, IL-17F, IFN-γ, IL-4, IL-10, IL-13, IL-16, IFN- α , TGF- $\beta^{[16,17]}$) is responsible for the net inflammatory response. Increased levels of IL-18, IL-12, IL-6, IL-17, TNF- α , and IFN- γ are reported in gingival tissues of chronic destructive periodontitis^[18,19]. In periodontitis both Th1 (IFN- γ , IL-2, TNF- α) and Th2 (IL-4, IL-5, IL-6, IL-13) type cytokines are observed^[20]. There is supporting evidence for the role of IL-17 and Th17 cells in periodontal disease^[21]. IL-17 induces IL-6 and IL-8 secretion by gingival fibroblasts and also upregulates MMP - Matrix Metalloproteinases (MMP-1) and MMP-3 in these cells^[22,23]. IL-17 also induces IL-1 β and TNF- α secretion from macrophages and gingival epithelial cells^[22,23]. Inflammatory cytokines are produced as a result of activation of toll like receptors of oral epithelial cells by the lipopolysaccharide of the gram negative periodontal pathogens^[24]. Recently it has been reported that pathogenesis of many systemic diseases are associated with these inflammatory mediators. The pathways bridging periodontal infection and systemic health include transient bacteremia via metastatic infection, injury and inflammation resulting from immunological response induced by periodontal pathogens^[25].

Recent studies have demonstrated that chronic periodontitis acts as a risk factor for systemic diseases like diabetes mellitus, cardiovascular disease, adverse pregnancy

Joseph R et al. Rheumatoid arthritis and periodontitis

outcomes, rheumatoid arthritis etc^[26-28].

Rheumatoid arthritis (RA) is a chronic inflammatory disease of articular joint with unknown etiology marked by a symmetric, peripheral polyarthritis and often results in joint damage and physical disability. The pathogenic hallmark of RA is synovial inflammation and proliferation, focal bone erosion and thinning of articular cartilage. Articular cartilage is an avascular tissue composed of a specialized matrix of collagen, proteoglycans and other proteins. Chondrocytes contribute to the unique cellular component. Cartilage is a highly responsive tissue that reacts to inflammatory mediators and mechanical factors, which in turn alters the balance between cartilage anabolism and catabolism. The structural damage to the mineralized cartilage and subchondral bone is mediated by osteoclasts^[29].

Worldwide prevalence of RA, an autoimmune condition is approximately 1%^[30]. It is diagnosed as chronic inflammatory polyarthritis when five or more joints are affected^[29]. On close observation, a number of similarities seem to exist between the supporting periodontal structures and articular joint (Table 1).

SIMILARITIES BETWEEN RA AND PD

Periodontitis is a destructive chronic inflammatory disease of the periodontium caused by biofilm associated specific microorganisms^[31-33]. Rheumatoid arthritis is an autoimmune condition and is characterized by elevated serum autoantibody titre directed against citrullinated peptides or rheumatoid factor (RF)^[34,35]. Autoantibodies such as RF and anti-citrullinated protein/peptide antibody (ACPA) may be found in the sera of RA patients long before clinical onset of disease^[27]. Periodontitis may also involve some elements of autoimmunity^[36]. Autoantibodies and specific T cells against host molecules, such as type 1 collagen, have been detected in periodontal disease^[23]. Recent studies have established statistically significant association between PD and RA^[37-41]. The likelihood of PD among patients with RA is high. Also a higher prevalence of RA has been reported among patients with moderate to severe PD^[41]. Joseph R. reported more periodontal destruction in RA group, pointing to a positive association between these diseases^[42]. When comparing patients with RA and those with PD, many similarities have been reported in terms of serum cytokine and gene expression profiles, increased levels of serum matrix metalloproteinases, reactive oxygen species, lipid mediators, and neutrophil associated enzymes^[5,43-46]. It has further been proposed that polymorphisms relating to genes encoding inflammatory cytokines might confer susceptibility to RA and PD^[47-49]. Table 2 depicts some similarities observed in the pathogenesis of RA and PD.

Role of Porphyromonas gingivalis and immune response

Periodontal pathogens like *Porphyromonas gingivalis* (*P. gingivalis*) can invade the blood vessels and endothelial cells and lead to persistent bacteremia. It has the ability to in-



81

Table 1 Similarities between periodontal structures and articular joint

Supporting periodontal structures	Articular joint
Periodontal structures comprise of cementum, alveolar bone,	Articular joints comprise of articular cartilage, bone, ligaments, synovial cavity, synovial
periodontal ligament, gingival crevicular fluid and gingiva	fluid, and synovial capsule
Cementum is an avascular tissue	Articular cartilage is an avascular tissue
Periodontal ligament is a thin connective tissue that surrounds	Synovial tissue is a thin layer of connective tissue. It consists primarily of two cell types-
the root connecting it to the alveolar bone	type A synoviocytes (macrophage derived) and type B synoviocyte (fibroblast derived)
Periodontal ligament is collagenous and consists of epithelial	Synovial fibroblasts are the most abundant and produce the structural components of the
rests of malassez, fibroblasts, osteoblasts and ground substances	joints including collagen, fibronectin and laminin
(hyaluronic acid and proteoglycans-fibronectin and laminin)	
Gingival crevicular fluid is an infiltrate of blood	Synovial fluid is an infiltrate of blood

Table 2 Similarities in pathogenesis periodontal disease and rheumatoid arthritis

PD	RA
Chronic immunoinflammatory disease	Chronic immunoinflammatory disease
Periodontal pathogen is the main etiological agent with some element of	Bacteria/peptide as an adjunct antigen in autoantibody production
autoimmunity	
HLA-DR antigen association	HLA-DR antigen association
Inflammatory infiltrate mainly consists of B cells, plasma cells, PMN, T cell,	Inflammatory infiltrate consists of T cell, B cell, plasma cell, dendritic cell,
dendritic cell, and macrophages	mast cell, macrophages, and few granulocytes
Increases level of IL-1, TNF-α, PGE2, MMPs, NF-κB, RANK/RANKL/OPG,	Increases level of IL-1, TNF- α , PGE2, MMPs
osteoclast activation	NF-κβ, RANK/RANKL/OPG, osteoclast activation
Th1, ↑ ed Th2 and Th 17	Th1 = Th2 and $Th 17$
Role of nitric oxide	Role of nitric oxide
Genetic and environmental influences	Genetic and environmental influences
Bacterial DNA of anaerobes and high antibody titres against heat shock	Bacterial DNA of anaerobes and high antibody titres against heat shock
protein of P.gingivalis, P. Melanogenicus and P. Intermedia ^[95]	protein of P. gingivalis, P. Melanogenicus and P. Intermedia ^[95]

PD: Periodontal disease; RA: Rheumatoid arthritis; IL: Inflammatory mediator; MMPs: Matrix metalloproteinases; HLA: Human leukocytic antigen; HLA-DR: Human leucocyte antigen- D related; RANK: Receptor activator of nuclear factor kappa-β; RANKL: Receptor activator of nuclear factor kappa-β ligand; OPG: Osteoprotegerin; *P. gingioalis: Porphyromonas Gingivalis; P. intermedia: Prevotella intermedia; P. melaninogenicus: Prevotella melaninogenicus;* TNF: Tumor necrosis factor; NF-κB: Nuclear factor - kappa β; IL: Interleukin; PGE: Prostaglandin E2.

vade primary chondrocytes of knee joints. As a result, cell cycle progression gets delayed, ultimately leading to accelerated apoptosis of these chondrocytes^[50]. The virulence of P. gingivalis is mainly associated with its trypsin like proteolytic activity and ability to produce arginine and lysine -specific cysteine endopeptidase like gingipain R and gingipain K respectively^[51]. Gingipain aids in evasion of host defense, tissue destruction and infection^[52,53]. It leads to activation of MMPs (1, 3 and 9) and degradation of host proteins (laminin, fibronectin and collagen)^[54]. Being the only identified bacterium with expression of peptidyl arginine deiminase (PAD), P. gingivalis and PAD represent a notable pathogenic element of RA^[55-58]. PAD catalyses the deimination of arginine residuals to citrulline, a form of post-translational protein modification^[59] which leads to an irreversible translation of arginine to citrulline^[56,59]. However one important difference is that PAD expressed by P. gingivalis and human PAD are not exactly homologous $^{[56,59]}$. It has been reported that in RA there is an increased citrullination of structure proteins^[60]. This probably accounts for the fact that P. gingivalis titre significantly correlates with ACPA titre in RA patients^[56,61-65]

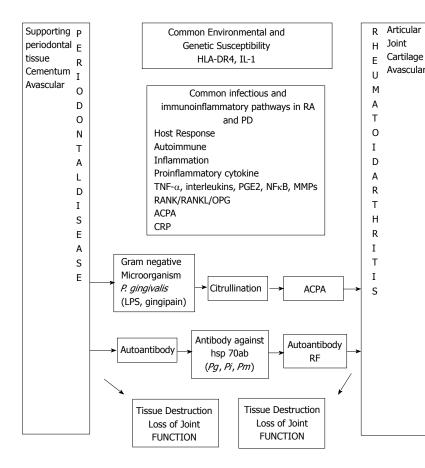
GENETIC PROFILE IN PD AND RA

The most potent disease risk gene in RA and PD is the genome on the human leukocytic antigen (HLA) re-

gion^[66]. HLA-DR4 antigen is associated with both RA and PD. This genetic association between these chronic inflammatory conditions also points to the biologic link between them^[65,67-69]. Hitchon and colleagues have reported an association between *P. gingivalis* and the presence of ACPA in a population with predominant RApredisposing HLA-DRB1 alleles. This gene-environment interaction may contribute to the breaking up of selftolerance to citrullinated proteins. It could also amplify autoimmune reactions which could predispose to RA^[70].

MARKERS OF INFLAMMATION IN PD AND RA

The synovial fluid of RA patients is rich in proinflammatory cytokines and many interleukins, (IL-1, IL-6, IL-8, IL-15, and IL-17) as well as NF- κ B ligand (RANKL) which can be linked with RA^[71,72]. Similar profile of inflammatory mediators has been identified in chronic periodontitis^[35,73,74]. Elevated serum levels of TNF- α is associated with both these chronic inflammatory diseases^[74,75]. Lipopolysaccharides and other bacterial byproducts stimulate the release of TNF- α and it upregulates the release of prostaglandin E2 (PGE2) and MMPs that stimulate osteoclast activation. These inflammatory processes ultimately lead to bone resorption in both RA and PD^[76].



Joseph R et al. Rheumatoid arthritis and periodontitis

Figure 1 Hypothetical model of biological link between rheumatoid arthritis and periodontal disease. ACPA: Anti-citrullinated protein/peptide antibody; CRP: C reative protein; HLA: Human leukocyte antigen; hsp: Heat shock protein; LPS: Lipopolysaccharide, MMPs: Matrix metalloprotienases; NF_KB: Nuclear factor kappa β ; OPG: Osteoprotegerin; PD: Periodontal disease; PGE2: Prostaglandin E2; RANK: Receptor activator of nuclear factor kappa β ; RANKL: Receptor activator of nuclear factor kappa β ; Igand; RA: Rheumatoid arthritis; RF: Rheumatoid factor; TNF- α : Tumor necrosis factor alpha; ACPA: Anti-citrullinated protein; RF: Rheumatoid factor; *P. gingivalis: Porphyromonas gingivalis*; IL: Interleukin.

EFFECTS OF THERAPY FOR RHEUMATOID ARTHRITIS ON PERIODONTAL DISEASE AND THERAPY FOR PERIODONTAL DISEASE ON RHEUMATOID ARTHRITIS

It has been suggested that treatment of periodontitis in patient with RA improved their response to RA therapy^[77-80]. Treatment of RA with disease modifying antirheumatic drugs (DMARDS) improves their periodontal condition due to its host modulatory effect, thus masking the gingival inflammation and actual periodontal destruction^[81-83]. Similarly, reduction in the systemic inflammation by the additional effect of periodontal therapy may also have been masked by DMARDs^[35]. Al-Katma et al^[84] assessed the role of scaling and root planning (SRP) on RA and demonstrated that there was an improvement in RA scores in the test group as compared to the control group. Advances in treatment of RA have identified novel therapeutic targets such as anticytokine therapy. Anti-TNF- α therapy used to control RA may also be beneficial in the management of periodontitis^[85-88]. Ortiz et al^[77] assessed the additional effect of non-surgical periodontal therapy (NSPT) in RA patients under anti-TNF- α therapy and reported that regardless of the medications, supportive periodontal therapy had a positive result on the clinical features of RA. In the absence of periodontal treatment anti-TNF- α therapy alone had no relevant outcome on the periodontal condition^[77].

DOES A BIOLOGIC LINK EXIST BETWEEN PERIODONTAL DISEASE AND RHEUMATOID ARTHRITIS?

First of all, PD and RA share many pathological and immunological similarities. A cyclic nature of disease activity is seen in both RA and PD. There is evidence to suggest that PD could act as a potential risk factor for RA^[24,89,90]. Similarly, RA subjects have significantly increased clinical attachment loss (CAL)^[19-21]. Increased levels of antibodies to periodontopathic bacteria are reported to have been identified in sera and synovial tissues of patients with RA^[63,70,81,91]. Correlation of serum level IgG antibodies to P. gingivalis with anticyclic citrullinated peptide indicates that serum protein citrullination via peptidyl arginine deiminase of P. gingivalis drives RA responses^[63,70]. Citrullination by PAD may act as a biologically plausible mechanistic link between PD and RA. Furthermore the presence of RA might predispose individuals to PD^[92,93]. Clinical trials suggest that treatment of PD has a significant effect on RA severity and vice versa^[84,94]

Second, it is suspected that *IL-1* gene polymorphism affects the cytokine protein in RA and PD. HLA DR4 antigen is associated with both the conditions which points to the biological link between the two^[67].

Third, it is reported that antibodies against heat shock protein (hsp 70 ab) of *P. gingivalis, P. melanogenicus and P. intermedia* are elevated not only in supporting periodontal tissues but also in synovial tissue of articular joints of RA patients^[91,95].

Joseph R et al. Rheumatoid arthritis and periodontitis

Fourth point, Both RA and PD have shown raised titres of IL-10, IL-1 α , IL-1 β , MMPs, TNF- α , LT- α and low titres of IL-1 α and IL-6^[45]. A common inflammatory marker dysfunction seems to be associated with both the articular joint and supporting periodontal tissue.

Bacterial infection, genetic susceptibility, altered immune reaction and inflammatory mediators considered responsible for RA are also associated with PD. So it is plausible that a biological link may exist between PD and RA (Figure 1).

EVIDENCES LINKING PD AND RA

Several studies have revealed that the prevalence of PD was high in patients with $RA^{[40,42]}$ and that PD severity was greater in RA patients^[45]. Mercado *et al*^[39] (2001) demonstrated a relation between RA and severity of periodontitis in a case control clinical study. Pischon *et al*^[40] (2009) in a cross sectional clinical study showed significantly more CAL in RA subjects as compared to non-RA subjects. They have concluded that oral hygiene may account for this association to some extent. Kobayashi *et al*^[49] (2007) reported that IL-1 and FCyR gene polymorphism have potential risk for RA and periodontitis.

Martinez-Martinez *et al*^{p61} (2009) in a case series clinical study on subjects with refractory RA and periodontitis found that *P. intermedia*, *P. gingivalis*, and *T. denticola* were the most predominant gram negative bacteria identified in synovial fluid, which substantiates the concept of anti-CCP and citrullinated structure protein. Dissick *et al*⁴¹¹ (2010) in a case control study demonstrated that RA+ patients have more moderate to severe periodontitis. They have reported that females and smokers are at more risk in the RA+/periodontitis complex

Okada et al^[81] (2011) demonstrated that corticosteroids, anti-rheumatic drugs, NSAIDs and TNF-α antagonists therapy improved the clinical features of periodontitis in RA patients. Presence of anti-PG IgG antibodies in RA+ patients may influence the serum RF level and periodontal health status^[81]. Mayer et al^[83] (2009) in a case control study concluded that TNF- α levels correlated with overall CAL and that inhibition of proinflammatory cytokines may account for the reduction of periodontal parameters. In another case control study, Ribeiro et al⁹⁴ (2005) evaluated the role of NSPT on RA status and found that RF decreased after periodontal intervention. The effects of NSPT in subjects with and without RA was studied by Pinho Mde et al^[97] (2009) and they stated that that the relation between RA and periodontal disease activity is unclear. The effect of NSPT on RA patients under anti-TNF- α was studied by Ortiz *et al*^[77] (2009) who inferred that NSPT had a positive effect on the clinical parameters of RA. Okada et al^[98] in (2013) suggested that periodontal treatment decreases the levels of antibodies to P. gingivalis and citrulline in patients with RA and Periodontitis. They concluded that these observations may reflect the role of P. gingivalis in the protein citrullination which is related to the pathogenesis of RA^[98]. Kaur et al^{78]} (2014) in a systematic review and meta analysis reported that non surgical periodontal therapy could lead to improvement in clinical and biochemical disease activity in RA.

Quirke *et al*^[99] in 2013 reported that *P. gingivalis* is seemingly distinctive among periodontal pathogens in having PPAD (*P. gingivalis* peptidylarginine deiminase) with potential to evoke autoimmune response. They opined that the peptidyl citrulline specific immune response to PPAD might break tolerance in RA and could be a target for therapy^[99]. Agnihotri *et al*^[100] in (2014) reviewed the link between RA and PD in the elderly and inferred that thorough understanding of the link between the two chronic inflammatory diseases might be beneficial in rendering better health care protection and betterment of the life style of aged individuals.

CONCLUSION

The relationship between RA and PD can be attributed to common dysfunction and dysregulation in inflammatory mechanisms. Apparently, the common factors are bacterial lipopolysaccharides and inflammatory mediators. Development of specific autoantibodies by citrullination of protein by *P. gingivalis* may be the connecting link between RA and PD. Therapies aimed at suppression of inflammatory mediators and effector molecules such as MMP, proinflammatory cytokines and autoantibodies of structural proteins may probably reduce the severity of both RA and PD.

REFERENCES

- Cochran DL. Inflammation and bone loss in periodontal disease. J Periodontol 2008; 79: 1569-1576 [PMID: 18673012 DOI: 10.1902/jop.2008.080233]
- 2 Page RC, Kornman KS. The pathogenesis of human periodontitis: an introduction. *Periodontol 2000* 1997; 14: 9-11 [PMID: 9567963]
- 3 **Offenbacher S**. Periodontal diseases: pathogenesis. *Ann Periodontol* 1996; **1**: 821-878 [PMID: 9118282]
- 4 Cantore S, Mirgaldi R, Ballini A, Coscia MF, Scacco S, Papa F, Inchingolo F, Dipalma G, De Vito D. Cytokine gene polymorphisms associate with microbiogical agents in periodontal disease: our experience. *Int J Med Sci* 2014; **11**: 674-679 [PMID: 24843315 DOI: 10.7150/ijms.6962]
- 5 Bartold PM, Marshall RI, Haynes DR. Periodontitis and rheumatoid arthritis: a review. J Periodontol 2005; 76: 2066-2074 [PMID: 16277578]
- 6 Saini R, Marawar PP, Shete S, Saini S. Periodontitis, a true infection. J Glob Infect Dis 2009; 1: 149-150 [PMID: 20300407 DOI: 10.4103/0974-777X.56251]
- 7 Kornman KS. Mapping the pathogenesis of periodontitis: a new look. *J Periodontol* 2008; **79**: 1560-1568 [PMID: 18673011 DOI: 10.1902/jop.2008.080213]
- 8 Van Dyke TE. Role of the neutrophil in oral disease: receptor deficiency in leukocytes from patients with juvenile periodontitis. *Rev Infect Dis* 1985; 7: 419-425 [PMID: 3895356]
- 9 Genco RJ, Van Dyke TE, Levine MJ, Nelson RD, Wilson ME. 1985 Kreshover lecture. Molecular factors influencing neutrophil defects in periodontal disease. J Dent Res 1986; 65: 1379-1391 [PMID: 3023465]
- 10 Paster BJ, Boches SK, Galvin JL, Ericson RE, Lau CN, Levanos VA, Sahasrabudhe A, Dewhirst FE. Bacterial diversity in human subgingival plaque. J Bacteriol 2001; 183: 3770-3783



WJR www.wjgnet.com

Joseph R et al. Rheumatoid arthritis and periodontitis

[PMID: 11371542]

- 11 Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL. Microbial complexes in subgingival plaque. J Clin Periodontol 1998; 25: 134-144 [PMID: 9495612]
- 12 Socransky SS, Haffajee AD, Ximenez-Fyvie LA, Feres M, Mager D. Ecological considerations in the treatment of Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis periodontal infections. *Periodontol 2000* 1999; 20: 341-362 [PMID: 10522230]
- 13 **Beutler B**. Toll-like receptors: how they work and what they do. *Curr Opin Hematol* 2002; **9**: 2-10 [PMID: 11753071]
- 14 **Beutler B**, Hoebe K, Du X, Ulevitch RJ. How we detect microbes and respond to them: the Toll-like receptors and their transducers. *J Leukoc Biol* 2003; **74**: 479-485 [PMID: 12960260]
- 15 Lamster IB, Novak MJ. Host mediators in gingival crevicular fluid: implications for the pathogenesis of periodontal disease. *Crit Rev Oral Biol Med* 1992; **3**: 31-60 [PMID: 1730070]
- 16 Graves DT, Delima AJ, Assuma R, Amar S, Oates T, Cochran D. Interleukin-1 and tumor necrosis factor antagonists inhibit the progression of inflammatory cell infiltration toward alveolar bone in experimental periodontitis. *J Periodontol* 1998; 69: 1419-1425 [PMID: 9926773]
- 17 Lappin DF, MacLeod CP, Kerr A, Mitchell T, Kinane DF. Anti-inflammatory cytokine IL-10 and T cell cytokine profile in periodontitis granulation tissue. *Clin Exp Immunol* 2001; 123: 294-300 [PMID: 11207661]
- 18 Górska R, Gregorek H, Kowalski J, Laskus-Perendyk A, Syczewska M, Madaliński K. Relationship between clinical parameters and cytokine profiles in inflamed gingival tissue and serum samples from patients with chronic periodontitis. *J Clin Periodontol* 2003; 30: 1046-1052 [PMID: 15002890]
- 19 Lester SR, Bain JL, Johnson RB, Serio FG. Gingival concentrations of interleukin-23 and -17 at healthy sites and at sites of clinical attachment loss. *J Periodontol* 2007; 78: 1545-1550 [PMID: 17668974]
- 20 Matsuki Y, Yamamoto T, Hara K. Detection of inflammatory cytokine messenger RNA (mRNA)-expressing cells in human inflamed gingiva by combined in situ hybridization and immunohistochemistry. *Immunology* 1992; 76: 42-47 [PMID: 1628899]
- 21 Gaffen SL, Hajishengallis G. A new inflammatory cytokine on the block: re-thinking periodontal disease and the Th1/ Th2 paradigm in the context of Th17 cells and IL-17. *J Dent Res* 2008; 87: 817-828 [PMID: 18719207]
- 22 Newman MG, Takei HH, Klokkevold PR, Carranza FA. Clinical Periodontology 10th ed. New Delhi: Elsevier Science Health Science Division, 2006: 239
- 23 Newman MG, Takei H, Klokkevold PR, Carranza FA. Carranza's Clinical Periodontology 11th edition (Elsevier Health Sciences, 2011: 281-282). Available from: URL: http://www.clinicalperiodontology.com/
- 24 **Tietze K**, Dalpke A, Morath S, Mutters R, Heeg K, Nonnenmacher C. Differences in innate immune responses upon stimulation with gram-positive and gram-negative bacteria. *J Periodontal Res* 2006; **41**: 447-454 [PMID: 16953821]
- 25 Akar H, Akar GC, Carrero JJ, Stenvinkel P, Lindholm B. Systemic consequences of poor oral health in chronic kidney disease patients. *Clin J Am Soc Nephrol* 2011; 6: 218-226 [PMID: 21115624 DOI: 10.2215/CJN.05470610]
- 26 Kuo LC, Polson AM, Kang T. Associations between periodontal diseases and systemic diseases: a review of the interrelationships and interactions with diabetes, respiratory diseases, cardiovascular diseases and osteoporosis. *Public Health* 2008; **122**: 417-433 [PMID: 18028967]
- 27 Löe H. Periodontal disease. The sixth complication of diabetes mellitus. *Diabetes Care* 1993; 16: 329-334 [PMID: 8422804]
- 28 Mercado F, Marshall RI, Klestov AC, Bartold PM. Is there a relationship between rheumatoid arthritis and periodontal disease? J Clin Periodontol 2000; 27: 267-272 [PMID: 10783841]
- 29 Longo D, Fauci A, Kasper D, Hauser S, Jameson J. Harrisons Manual of Medicine, 18th ed. New York: McGraw-Hill Education, 2012: 2738

- 30 **Sacks JJ**, Luo YH, Helmick CG. Prevalence of specific types of arthritis and other rheumatic conditions in the ambulatory health care system in the United States, 2001-2005. *Arthritis Care Res* (Hoboken) 2010; **62**: 460-464 [PMID: 20391499 DOI: 10.1002/acr.20041]
- 31 Slots J, Rams TE. New views on periodontal microbiota in special patient categories. J Clin Periodontol 1991; 18: 411-420 [PMID: 1890221]
- 32 **Socransky SS**, Haffajee AD. The bacterial etiology of destructive periodontal disease: current concepts. *J Periodontol* 1992; **63**: 322-331 [PMID: 1573546]
- 33 Wolff L, Dahlén G, Aeppli D. Bacteria as risk markers for periodontitis. J Periodontol 1994; 65: 498-510 [PMID: 8046566]
- 34 McInnes IB, Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. Nat Rev Immunol 2007; 7: 429-442 [PMID: 17525752]
- 35 Rutger Persson G. Rheumatoid arthritis and periodontitis - inflammatory and infectious connections. Review of the literature. J Oral Microbiol 2012; 4 [PMID: 22347541 DOI: 10.3402/jom.v4i0.11829]
- 36 Gemmell E, Yamazaki K, Seymour GJ. The role of T cells in periodontal disease: homeostasis and autoimmunity. *Periodontol 2000 2007*; 43: 14-40 [PMID: 17214833]
- 37 Greenwald RA, Kirkwood K. Adult periodontitis as a model for rheumatoid arthritis (with emphasis on treatment strategies) J Rheumatol 1999; 26: 1650-1653 [PMID: 10451056]
- 38 Kässer UR, Gleissner C, Dehne F, Michel A, Willershausen-Zönnchen B, Bolten WW. Risk for periodontal disease in patients with longstanding rheumatoid arthritis. *Arthritis Rheum* 1997; 40: 2248-2251 [PMID: 9416864]
- 39 Mercado FB, Marshall RI, Klestov AC, Bartold PM. Relationship between rheumatoid arthritis and periodontitis. J Periodontol 2001; 72: 779-787 [PMID: 11453241]
- 40 Pischon N, Pischon T, Kröger J, Gülmez E, Kleber BM, Bernimoulin JP, Landau H, Brinkmann PG, Schlattmann P, Zernicke J, Buttgereit F, Detert J. Association among rheumatoid arthritis, oral hygiene, and periodontitis. *J Periodontol* 2008; 79: 979-986 [PMID: 18533773 DOI: 10.1902/jop.2008.070501]
- 41 Dissick A, Redman RS, Jones M, Rangan BV, Reimold A, Griffiths GR, Mikuls TR, Amdur RL, Richards JS, Kerr GS. Association of periodontitis with rheumatoid arthritis: a pilot study. J Periodontol 2010; 81: 223-230 [PMID: 20151800 DOI: 10.1902/jop.2009.090309]
- 42 Joseph R, Rajappan S, Nath SG, Paul BJ. Association between chronic periodontitis and rheumatoid arthritis: a hospital-based case-control study. *Rheumatol Int* 2013; **33**: 103-109 [PMID: 22228465 DOI: 10.1007/s00296-011-2284-1]
- 43 Mercado FB, Marshall RI, Bartold PM. Inter-relationships between rheumatoid arthritis and periodontal disease. A review. J Clin Periodontol 2003; 30: 761-772 [PMID: 12956651]
- 44 de Pablo P, Chapple IL, Buckley CD, Dietrich T. Periodontitis in systemic rheumatic diseases. *Nat Rev Rheumatol* 2009; 5: 218-224 [PMID: 19337286 DOI: 10.1038/nrrheum.2009.28]
- 45 Havemose-Poulsen A, Sørensen LK, Stoltze K, Bendtzen K, Holmstrup P. Cytokine profiles in peripheral blood and whole blood cell cultures associated with aggressive periodontitis, juvenile idiopathic arthritis, and rheumatoid arthritis. J Periodontol 2005; 76: 2276-2285 [PMID: 16332240]
- 46 Kobayashi T, Murasawa A, Komatsu Y, Yokoyama T, Ishida K, Abe A, Yamamoto K, Yoshie H. Serum cytokine and periodontal profiles in relation to disease activity of rheumatoid arthritis in Japanese adults. *J Periodontol* 2010; **81**: 650-657 [PMID: 20429644 DOI: 10.1902/jop.2010.090688]
- 47 Havemose-Poulsen A, Sørensen LK, Bendtzen K, Holmstrup P. Polymorphisms within the IL-1 gene cluster: effects on cytokine profiles in peripheral blood and whole blood cell cultures of patients with aggressive periodontitis, juvenile idiopathic arthritis, and rheumatoid arthritis. *J Periodontol* 2007; **78**: 475-492 [PMID: 17335371]
- 48 **Kobayashi T**, Ito S, Kuroda T, Yamamoto K, Sugita N, Narita I, Sumida T, Gejyo F, Yoshie H. The interleukin-1 and

Fcgamma receptor gene polymorphisms in Japanese patients with rheumatoid arthritis and periodontitis. *J Periodontol* 2007; **78**: 2311-2318 [PMID: 18052703]

- 49 Kobayashi T, Murasawa A, Ito S, Yamamoto K, Komatsu Y, Abe A, Sumida T, Yoshie H. Cytokine gene polymorphisms associated with rheumatoid arthritis and periodontitis in Japanese adults. J Periodontol 2009; 80: 792-799 [PMID: 19405833 DOI: 10.1902/jop.2009.080573]
- 50 Pischon N, Röhner E, Hocke A, N'Guessan P, Müller HC, Matziolis G, Kanitz V, Purucker P, Kleber BM, Bernimoulin JP, Burmester G, Buttgereit F, Detert J. Effects of Porphyromonas gingivalis on cell cycle progression and apoptosis of primary human chondrocytes. *Ann Rheum Dis* 2009; 68: 1902-1907 [PMID: 19054824 DOI: 10.1136/ard.2008.102392]
- 51 **Ogrendik M**. Rheumatoid arthritis is linked to oral bacteria: etiological association. *Mod Rheumatol* 2009; **19**: 453-456 [PMID: 19554393 DOI: 10.1007/s10165-009-0194-9]
- 52 **Imamura T**. The role of gingipains in the pathogenesis of periodontal disease. *J Periodontol* 2003; **74**: 111-118 [PMID: 12593605]
- 53 Stathopoulou PG, Galicia JC, Benakanakere MR, Garcia CA, Potempa J, Kinane DF. Porphyromonas gingivalis induce apoptosis in human gingival epithelial cells through a gingipain-dependent mechanism. *BMC Microbiol* 2009; 9: 107 [PMID: 19473524 DOI: 10.1186/1471-2180-9-107]
- 54 Jie Bao G, Kari K, Tervahartiala T, Sorsa T, Meurman JH. Proteolytic Activities of Oral Bacteria on ProMMP-9 and the Effect of Synthetic Proteinase Inhibitors. *Open Dent J* 2008; 2: 96-102 [PMID: 19088890 DOI: 10.2174/1874210600802010096]
- 55 McGraw WT, Potempa J, Farley D, Travis J. Purification, characterization, and sequence analysis of a potential virulence factor from Porphyromonas gingivalis, peptidylarginine deiminase. *Infect Immun* 1999; 67: 3248-3256 [PMID: 10377098]
- 56 Rosenstein ED, Greenwald RA, Kushner LJ, Weissmann G. Hypothesis: the humoral immune response to oral bacteria provides a stimulus for the development of rheumatoid arthritis. *Inflammation* 2004; 28: 311-318 [PMID: 16245073]
- 57 Maresz KJ, Hellvard A, Sroka A, Adamowicz K, Bielecka E, Koziel J, Gawron K, Mizgalska D, Marcinska KA, Benedyk M, Pyrc K, Quirke AM, Jonsson R, Alzabin S, Venables PJ, Nguyen KA, Mydel P, Potempa J. Porphyromonas gingivalis facilitates the development and progression of destructive arthritis through its unique bacterial peptidylarginine deiminase (PAD). *PLoS Pathog* 2013; 9: e1003627 [PMID: 24068934 DOI: 10.1371/journal.ppat.1003627]
- 58 Abdullah SN, Farmer EA, Spargo L, Logan R, Gully N. Porphyromonas gingivalis peptidylarginine deiminase substrate specificity. *Anaerobe* 2013; 23: 102-108 [PMID: 23856045 DOI: 10.1016/j.anaerobe.2013.07.001]
- 59 Schellekens GA, Visser H, de Jong BA, van den Hoogen FH, Hazes JM, Breedveld FC, van Venrooij WJ. The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. *Arthritis Rheum* 2000; 43: 155-163 [PMID: 10643712]
- 60 Wegner N, Lundberg K, Kinloch A, Fisher B, Malmström V, Feldmann M, Venables PJ. Autoimmunity to specific citrullinated proteins gives the first clues to the etiology of rheumatoid arthritis. *Immunol Rev* 2010; 233: 34-54 [PMID: 20192991 DOI: 10.1111/j.0105-2896.2009.00850.x]
- 61 Haffajee AD, Socransky SS. Microbial etiological agents of destructive periodontal diseases. *Periodontol 2000* 1994; 5: 78-111 [PMID: 9673164]
- 62 Mikuls TR, Payne JB, Yu F, Thiele GM, Reynolds RJ, Cannon GW, Markt J, McGowan D, Kerr GS, Redman RS, Reimold A, Griffiths G, Beatty M, Gonzalez SM, Bergman DA, Hamilton BC, Erickson AR, Sokolove J, Robinson WH, Walker C, Chandad F, O'Dell JR. Periodontitis and Porphyromonas gingivalis in patients with rheumatoid arthritis. *Arthritis Rheumatol* 2014; 66: 1090-1100 [PMID: 24782175 DOI: 10.1002/art.38348]

- 63 Mikuls TR, Payne JB, Reinhardt RA, Thiele GM, Maziarz E, Cannella AC, Holers VM, Kuhn KA, O'Dell JR. Antibody responses to Porphyromonas gingivalis (P. gingivalis) in subjects with rheumatoid arthritis and periodontitis. *Int Immunopharmacol* 2009; **9**: 38-42 [PMID: 18848647 DOI: 10.1016/j.intimp.2008.09.008]
- 64 Liao F, Li Z, Wang Y, Shi B, Gong Z, Cheng X. Porphyromonas gingivalis may play an important role in the pathogenesis of periodontitis-associated rheumatoid arthritis. *Med Hypotheses* 2009; **72**: 732-735 [PMID: 19246161 DOI: 10.1016/ j.mehy.2008.12.040]
- 65 Mikuls TR, Thiele GM, Deane KD, Payne JB, O'Dell JR, Yu F, Sayles H, Weisman MH, Gregersen PK, Buckner JH, Keating RM, Derber LA, Robinson WH, Holers VM, Norris JM. Porphyromonas gingivalis and disease-related autoantibodies in individuals at increased risk of rheumatoid arthritis. *Arthritis Rheum* 2012; 64: 3522-3530 [PMID: 22736291 DOI: 10.1002/art.34595]
- 66 Kinloch AJ, Alzabin S, Brintnell W, Wilson E, Barra L, Wegner N, Bell DA, Cairns E, Venables PJ. Immunization with Porphyromonas gingivalis enolase induces autoimmunity to mammalian α-enolase and arthritis in DR4-IE-transgenic mice. *Arthritis Rheum* 2011; 63: 3818-3823 [PMID: 21953289 DOI: 10.1002/art.30639]
- 67 Marotte H, Farge P, Gaudin P, Alexandre C, Mougin B, Miossec P. The association between periodontal disease and joint destruction in rheumatoid arthritis extends the link between the HLA-DR shared epitope and severity of bone destruction. *Ann Rheum Dis* 2006; 65: 905-909 [PMID: 16284099]
- 68 Bonfil JJ, Dillier FL, Mercier P, Reviron D, Foti B, Sambuc R, Brodeur JM, Sedarat C. A "case control" study on the rôle of HLA DR4 in severe periodontitis and rapidly progressive periodontitis. Identification of types and subtypes using molecular biology (PCR.SSO). J Clin Periodontol 1999; 26: 77-84 [PMID: 10048640]
- 69 Kapitány A, Zilahi E, Szántó S, Szücs G, Szabó Z, Végvári A, Rass P, Sipka S, Szegedi G, Szekanecz Z. Association of rheumatoid arthritis with HLA-DR1 and HLA-DR4 in Hungary. *Ann N Y Acad Sci* 2005; **1051**: 263-270 [PMID: 16126967]
- 70 Hitchon CA, Chandad F, Ferucci ED, Willemze A, Ioan-Facsinay A, van der Woude D, Markland J, Robinson D, Elias B, Newkirk M, Toes RM, Huizinga TW, El-Gabalawy HS. Antibodies to porphyromonas gingivalis are associated with anticitrullinated protein antibodies in patients with rheumatoid arthritis and their relatives. *J Rheumatol* 2010; **37**: 1105-1112 [PMID: 20436074 DOI: 10.3899/jrheum.091323]
- 71 **Astry B**, Harberts E, Moudgil KD. A cytokine-centric view of the pathogenesis and treatment of autoimmune arthritis. *J Interferon Cytokine Res* 2011; **31**: 927-940 [PMID: 22149412 DOI: 10.1089/jir.2011.0094]
- Fardellone P, Séjourné A, Paccou J, Goëb V. Bone remodelling markers in rheumatoid arthritis. *Mediators Inflamm* 2014; 2014: 484280 [PMID: 24839355 DOI: 10.1155/2014/484280]
- 73 Bozkurt FY, Berker E, Akkuş S, Bulut S. Relationship between interleukin-6 levels in gingival crevicular fluid and periodontal status in patients with rheumatoid arthritis and adult periodontitis. *J Periodontol* 2000; 71: 1756-1760 [PMID: 11128925]
- 74 Cetinkaya B, Guzeldemir E, Ogus E, Bulut S. Proinflammatory and anti-inflammatory cytokines in gingival crevicular fluid and serum of patients with rheumatoid arthritis and patients with chronic periodontitis. *J Periodontol* 2013; 84: 84-93 [PMID: 22414257 DOI: 10.1902/jop.2012.110467]
- 75 Orita S, Koshi T, Mitsuka T, Miyagi M, Inoue G, Arai G, Ishikawa T, Hanaoka E, Yamashita K, Yamashita M, Eguchi Y, Toyone T, Takahashi K, Ohtori S. Associations between proinflammatory cytokines in the synovial fluid and radiographic grading and pain-related scores in 47 consecutive patients with osteoarthritis of the knee. *BMC Musculoskelet Disord* 2011; **12**: 144 [PMID: 21714933 DOI: 10.1186/1471-247 4-12-144]



- 76 Xue M, McKelvey K, Shen K, Minhas N, March L, Park SY, Jackson CJ. Endogenous MMP-9 and not MMP-2 promotes rheumatoid synovial fibroblast survival, inflammation and cartilage degradation. *Rheumatology* (Oxford) 2014 Jun 29; Epub ahead of print [PMID: 24982240]
- 77 Ortiz P, Bissada NF, Palomo L, Han YW, Al-Zahrani MS, Panneerselvam A, Askari A. Periodontal therapy reduces the severity of active rheumatoid arthritis in patients treated with or without tumor necrosis factor inhibitors. J Periodontol 2009; 80: 535-540 [PMID: 19335072 DOI: 10.1902/jop.2009.080447]
- 78 Kaur S, Bright R, Proudman SM, Bartold PM. Does periodontal treatment influence clinical and biochemical measures for rheumatoid arthritis? A systematic review and meta-analysis. *Semin Arthritis Rheum* 2014 Apr 28; Epub ahead of print [PMID: 24880982 DOI: 10.1016/j.semarthrit.2014.04.009]
- 79 Monsarrat P, Vergnes JN, Cantagrel A, Algans N, Cousty S, Kémoun P, Bertrand C, Arrivé E, Bou C, Sédarat C, Schaeverbeke T, Nabet C, Sixou M. Effect of periodontal treatment on the clinical parameters of patients with rheumatoid arthritis: study protocol of the randomized, controlled ESPERA trial. *Trials* 2013; **14**: 253 [PMID: 23945051 DOI: 10.1186/1745-6215-14-253]
- 80 Erciyas K, Sezer U, Ustün K, Pehlivan Y, Kisacik B, Senyurt SZ, Tarakçioğlu M, Onat AM. Effects of periodontal therapy on disease activity and systemic inflammation in rheumatoid arthritis patients. *Oral Dis* 2013; 19: 394-400 [PMID: 22998534 DOI: 10.1111/odi.12017]
- 81 Okada M, Kobayashi T, Ito S, Yokoyama T, Komatsu Y, Abe A, Murasawa A, Yoshie H. Antibody responses to periodontopathic bacteria in relation to rheumatoid arthritis in Japanese adults. *J Periodontol* 2011; 82: 1433-1441 [PMID: 21342003 DOI: 10.1902/jop.2011.110020]
- 82 Han JY, Reynolds MA. Effect of anti-rheumatic agents on periodontal parameters and biomarkers of inflammation: a systematic review and meta-analysis. J Periodontal Implant Sci 2012; 42: 3-12 [PMID: 22413068 DOI: 10.5051/jpis.2012.42.1.3]
- 83 Mayer Y, Balbir-Gurman A, Machtei EE. Anti-tumor necrosis factor-alpha therapy and periodontal parameters in patients with rheumatoid arthritis. *J Periodontol* 2009; 80: 1414-1420 [PMID: 19722791 DOI: 10.1902/jop.2009.090015]
- 84 Al-Katma MK, Bissada NF, Bordeaux JM, Sue J, Askari AD. Control of periodontal infection reduces the severity of active rheumatoid arthritis. J Clin Rheumatol 2007; 13: 134-137 [PMID: 17551378]
- 85 Kobayashi T, Yokoyama T, Ito S, Kobayashi D, Yamagata A, Okada M, Oofusa K, Narita I, Murasawa A, Nakazono K, Yoshie H. Periodontal and Serum Protein Profiles in Patients With Rheumatoid Arthritis Treated With Tumor Necrosis Factor Inhibitor Adalimumab. J Periodontol 2014 May 26; Epub ahead of print [PMID: 24857321]
- 86 Üstün K, Erciyas K, Kısacık B, Sezer U, Pehlivan Y, Öztuzcu S, Gündoğar H, Onat AM. Host modulation in rheumatoid arthritis patients with TNF blockers significantly decreases biochemical parameters in periodontitis. *Inflammation* 2013; **36**: 1171-1177 [PMID: 23649513 DOI: 10.1007/ s10753-013-9652-9]
- 87 Queiroz-Junior CM, Bessoni RL, Costa VV, Souza DG, Teixeira MM, Silva TA. Preventive and therapeutic anti-TNF-α therapy with pentoxifylline decreases arthritis and the associated periodontal co-morbidity in mice. *Life Sci* 2013; 93: 423-428 [PMID: 23911669 DOI: 10.1016/j.lfs.2013.07.022]
- 88 Savioli C, Ribeiro AC, Fabri GM, Calich AL, Carvalho J, Silva CA, Viana VS, Bonfá E, Siqueira JT. Persistent

periodontal disease hampers anti-tumor necrosis factor treatment response in rheumatoid arthritis. *J Clin Rheumatol* 2012; **18**: 180-184 [PMID: 22647860 DOI: 10.1097/ RHU.0b013e31825828be]

- 89 Scher JU, Bretz WA, Abramson SB. Periodontal disease and subgingival microbiota as contributors for rheumatoid arthritis pathogenesis: modifiable risk factors? *Curr Opin Rheumatol* 2014; 26: 424-429 [PMID: 24807405 DOI: 10.1097/ BOR.0000000000000076]
- 90 Scher JU, Abramson SB. Periodontal disease, Porphyromonas gingivalis, and rheumatoid arthritis: what triggers autoimmunity and clinical disease? *Arthritis Res Ther* 2013; 15: 122 [PMID: 24229458]
- 91 Moen K, Brun JG, Valen M, Skartveit L, Eribe EK, Olsen I, Jonsson R. Synovial inflammation in active rheumatoid arthritis and psoriatic arthritis facilitates trapping of a variety of oral bacterial DNAs. *Clin Exp Rheumatol* 2006; 24: 656-663 [PMID: 17207381]
- 92 Ramamurthy NS, Greenwald RA, Celiker MY, Shi EY. Experimental arthritis in rats induces biomarkers of periodontitis which are ameliorated by gene therapy with tissue inhibitor of matrix metalloproteinases. J Periodontol 2005; 76: 229-233 [PMID: 15974846]
- 93 Queiroz-Junior CM, Madeira MF, Coelho FM, Costa VV, Bessoni RL, Sousa LF, Garlet GP, Souza Dda G, Teixeira MM, Silva TA. Experimental arthritis triggers periodontal disease in mice: involvement of TNF-α and the oral Microbiota. *J Immunol* 2011; **187**: 3821-3830 [PMID: 21890656 DOI: 10.4049/jimmunol.1101195]
- 94 Ribeiro J, Leão A, Novaes AB. Periodontal infection as a possible severity factor for rheumatoid arthritis. J Clin Periodontol 2005; 32: 412-416 [PMID: 15811060]
- 95 Modi DK, Chopra VS, Bhau U. Rheumatoid arthritis and periodontitis: biological links and the emergence of dual purpose therapies. *Indian J Dent Res* 2009; 20: 86-90 [PMID: 19336867]
- 96 Martinez-Martinez RE, Abud-Mendoza C, Patiño-Marin N, Rizo-Rodríguez JC, Little JW, Loyola-Rodríguez JP. Detection of periodontal bacterial DNA in serum and synovial fluid in refractory rheumatoid arthritis patients. J Clin Periodontol 2009; 36: 1004-1010 [PMID: 19929953 DOI: 10.1111/ j.1600-051X.2009.01496.x]
- 97 Pinho Mde N, Oliveira RD, Novaes AB, Voltarelli JC. Relationship between periodontitis and rheumatoid arthritis and the effect of non-surgical periodontal treatment. *Braz Dent J* 2009; 20: 355-364 [PMID: 20126902]
- 98 Okada M, Kobayashi T, Ito S, Yokoyama T, Abe A, Murasawa A, Yoshie H. Periodontal treatment decreases levels of antibodies to Porphyromonas gingivalis and citrulline in patients with rheumatoid arthritis and periodontitis. *J Periodontol* 2013; 84: e74-e84 [PMID: 23701010 DOI: 10.1902/jop.2013.130079]
- 99 Quirke AM, Lugli EB, Wegner N, Hamilton BC, Charles P, Chowdhury M, Ytterberg AJ, Zubarev RA, Potempa J, Culshaw S, Guo Y, Fisher BA, Thiele G, Mikuls TR, Venables PJ. Heightened immune response to autocitrullinated Porphyromonas gingivalis peptidylarginine deiminase: a potential mechanism for breaching immunologic tolerance in rheumatoid arthritis. *Ann Rheum Dis* 2014; 73: 263-269 [PMID: 23463691 DOI: 10.1136/annrheumdis-2012-202726]
- 100 Agnihotri R, Gaur S. Rheumatoid arthritis in the elderly and its relationship with periodontitis: a review. *Geriatr Gerontol* Int 2014; 14: 8-22 [PMID: 23530652 DOI: 10.1111/ggi.12062]

P- Reviewer: Ardila CM, Haraszthy V, Kanzaki H S- Editor: Ji FF L- Editor: A E- Editor: Wu HL



WJR | www.wjgnet.com



Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx www.wjgnet.com World J Rheumatol 2014 November 12; 4(3): I-V ISSN 2220-3214 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

INSTRUCTIONS TO AUTHORS

GENERAL INFORMATION

World Journal of Rheumatology (World J Rheumatol, WJR, online ISSN 2220-3214, DOI: 10.5499) is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

Aim and scope

WJR covers topics concerning osteoarthritis, metabolic bone disease, connective tissue diseases, antiphospholipid antibody-associated diseases, spondyloarthropathies, acute inflammatory arthritis, fibromyalgia, polymyalgia rheumatica, vasculitis syndromes, periarticular rheumatic disease, pediatric rheumatic disease, miscellaneous rheumatic diseases, and rheumatology-related therapy, pain management, rehabilitation.

We encourage authors to submit their manuscripts to *WJR*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

WJR is edited and published by Baishideng Publishing Group (BPG). BPG has a strong professional editorial team composed of science editors, language editors and electronic editors. BPG currently publishes 43 OA clinical medical journals, including 42 in English, has a total of 15471 editorial board members or peer reviewers, and is a world first-class publisher.

Columns

The columns in the issues of WJR will include: (1) Editorial: The editorial board members are invited to make comments on an important topic in their field in terms of its current research status and future directions to lead the development of this discipline; (2) Frontier: The editorial board members are invited to select a highly cited cuttingedge original paper of his/her own to summarize major findings, the problems that have been resolved and remain to be resolved, and future research directions to help readers understand his/her important academic point of view and future research directions in the field; (3) Diagnostic Advances: The editorial board members are invited to write high-quality diagnostic advances in their field to improve the diagnostic skills of readers. The topic covers general clinical diagnosis, differential diagnosis, pathological diagnosis, laboratory diagnosis, imaging diagnosis, endoscopic diagnosis, biotechnological diagnosis, functional diagnosis, and physical diagnosis; (4) Therapeutics Advances: The editorial board members are invited to write high-quality therapeutic advances in their field to help improve the therapeutic skills of readers. The topic covers medication therapy, psychotherapy, physical therapy, replacement therapy, interventional therapy, minimally invasive therapy, endoscopic therapy, transplantation therapy, and surgical therapy; (5) Field of Vision: The editorial board members are invited to write commentaries on classic articles, hot topic articles, or latest articles to keep readers at the forefront of research and increase their levels of clinical research. Classic articles refer to papers that are included in Web of Knowledge and have received a large number of citations (ranking in the top 1%) after being published for more than years, reflecting the quality and impact of papers. Hot topic articles refer to papers that are included in Web of Knowledge and have received a large number of citations after being published for no more than 2 years, reflecting cuttingedge trends in scientific research. Latest articles refer to the latest published high-quality papers that are

included in PubMed, reflecting the latest research trends. These commentary articles should focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions. Basic information about the article to be commented (including authors, article title, journal name, year, volume, and inclusive page numbers; (6) Minireviews: The editorial board members are invited to write short reviews on recent advances and trends in research of molecular biology, genomics, and related cutting-edge technologies to provide readers with the latest knowledge and help improve their diagnostic and therapeutic skills; (7) Review: To make a systematic review to focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions; (8) Topic Highlight: The editorial board members are invited to write a series of articles (7-10 articles) to comment and discuss a hot topic to help improve the diagnostic and therapeutic skills of readers; (9) Medical Ethics: The editorial board members are invited to write articles about medical ethics to increase readers' knowledge of medical ethics. The topic covers international ethics guidelines, animal studies, clinical trials, organ transplantation, etc.; (10) Clinical Case Conference or Clinicopathological Conference: The editorial board members are invited to contribute high-quality clinical case conference; (11) Original Articles: To report innovative and original findings in rheumatology; (12) Research Report: To briefly report the novel and innovative findings in rheumatology; (13) Meta-Analysis: To evaluate the clinical effectiveness in rheumatology by using data from two or more randomised control trials; (14) Case Report: To report a rare or typical case; (15) Letters to the Editor: To discuss and make reply to the contributions published in W/R, or to introduce and comment on a controversial issue of general interest; (16) Book Reviews: To introduce and comment on quality monographs of rheumatology; and (17) Autobiography: The editorial board members are invited to write their autobiography to provide readers with stories of success or failure in their scientific research career. The topic covers their basic personal information and information about when they started doing research work, where and how they did research work, what they have achieved, and their lessons from success or failure.

Name of journal

World Journal of Rheumatology

ISSN

ISSN 2220-3214 (online)

Frequency

Four-monthly

Editor-in-chief

Jörg HW Distler, MD, Department of Internal Medicine 3, University of Erlangen-Nuremberg, Universitätsstr, 29, 91054 Erlangen, Germany

Editorial office

Jin-Lei Wang, Director Xiu-Xia Song, Vice Director World Journal of Rheumatology

Instructions to authors

Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China Telephone: +86-10-59080039 Fax: +86-10-85381893 E-mail: editorialoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

Publisher

Baishideng Publishing Group Inc 8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

Instructions to authors

Full instructions are available online at http://www.wjgnet.com/2220-3214/g_info_20100722180909.htm.

Indexed and abstracted in

Digital Object Identifier.

SPECIAL STATEMENT

All articles published in journals owned by the BPG represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including t-test (group or paired comparisons), chisquared test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, etc. The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (n). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the P value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJR* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje. org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human

studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of BPG, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is http://www.clinicaltrials.gov sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at: http://www.wjgnet.com/esps/. Authors are highly recommended to consult the ONLINE INSTRUC-TIONS TO AUTHORS (http://www.wjgnet.com/2220-3214/ g_info_20100722180909.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to



WJR www.wjgnet.com

wjrheumato@wjgnet.com, or by telephone: +86-10-85381891. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g. Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g. Telephone: +86-10-85381892 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision for acceptance is made only when at least two experts recommend an article for publication. Reviewers for accepted manuscripts are acknowledged in each manuscript, and reviewers of articles which were not accepted will be acknowledged at the end of each issue. To ensure the quality of the articles published in *WJR*, reviewers of accepted manuscripts will be announced by publishing the name, title/position and institution of the reviewer in the footnote accompanying the printed article. For example, reviewers: Professor Jing-Yuan Fang, Shanghai Institute of Digestive Disease, Shanghai, Affiliated Renji Hospital, Medical Faculty, Shanghai Jiaotong University, Shanghai, China; Professor Xin-Wei Han, Department of Radiology, The First Affiliated Hospital, Zhengzhou University, Zhengzhou, Henan Province, China; and Professor Anren Kuang, Department of Nuclear Medicine, Huaxi Hospital, Sichuan University, Chengdu, Sichuan Province, China.

Abstract

There are unstructured abstracts (no less than 200 words) and structured abstracts. The specific requirements for structured abstracts are as follows:

An informative, structured abstract should accompany each manuscript. Abstracts of original contributions should be structured into the following sections: AIM (no more than 20 words; Only the purpose of the study should be included. Please write the Aim in the form of "To investigate/study/..."), METHODS (no less than 140 words for Original Articles; and no less than 80 words for Brief Articles), RESULTS (no less than 150 words for Original Articles and no less than 120 words for Brief Articles; You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, *e.g.*, 6.92 ± 3.86 *vs* 3.61 ± 1.67 , P < 0.001), and CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Core tip

Please write a summary of less than 100 words to outline the most innovative and important arguments and core contents in your paper to attract readers.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRO-DUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both.

Illustrations

Figures should be numbered as 1, 2, 3, etc., and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Figures should be either Photoshop or Illustrator files (in tiff, eps, jpeg formats) at high-resolution. Examples can be found at: http://www.wignet.com/1007-9327/13/4520. pdf; http://www.wjgnet.com/1007-9327/13/4554.pdf; http:// www.wjgnet.com/1007-9327/13/4891.pdf; http://www. wjgnet.com/1007-9327/13/4986.pdf; http://www.wjgnet. com/1007-9327/13/4498.pdf. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ...etc. It is our principle to publish high resolution-figures for the printed and E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a



Instructions to authors

second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ${}^{a}P < 0.05$, ${}^{b}P < 0.01$ should be noted (P > 0.05 should not be noted). If there are other series of P values, ${}^{c}P < 0.05$ and ${}^{d}P < 0.01$ are used. A third series of P values can be expressed as ${}^{c}P < 0.05$ and ${}^{f}P < 0.01$. Other notes in tables or under illustrations should be expressed as ${}^{1}F_{}^{2}F_{}^{3}F_{}^{2}$, or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with \bullet , \circ , \blacksquare , \square , \land , *dc*, *etc.*, in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Pleased provide PubMed citation numbers to the reference list, e.g. PMID and DOI, which can be found at http://www.ncbi.nlm.nih. gov/sites/entrez?db=pubmed and http://www.crossref.org/Sim-pleTextQuery/, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wig.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

 Jung EM, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13. 6356]

Chinese journal article (list all authors and include the PMID where applicable)

2 Lin GZ, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarrhoea. Shijie Huaren Xiaohua Zazhi 1999; 7: 285-287

In press

3 Tian D, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. Proc Natl Acad Sci USA 2006; In press

Organization as author

4 Diabetes Prevention Program Research Group. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; 40: 679-686 [PMID: 12411462 PMCID:2516377 DOI:10.1161/01.HYP.0000035706.28494. 09]

Both personal authors and an organization as author

5 Vallancien G, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; 169: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju. 0000067940.76090.73]

No author given

6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325. 7357.184]

Volume with supplement

- 7 Geraud G, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; 42 Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/ j.1526-4610.42.s2.7.x]
- Issue with no volume
- 8 Banit DM, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (401): 230-238 [PMID: 12151900 DOI:10.10 97/00003086-200208000-00026]

No volume or issue

9 Outreach: Bringing HIV-positive individuals into care. HRSA Careaction 2002; 1-6 [PMID: 12154804]

Books

- Personal author(s)
- Sherlock S, Dooley J. Diseases of the liver and billiary system.
 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296
- Chapter in a book (list all authors)
- 11 Lam SK. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450
- Author(s) and editor(s)
- 12 Breedlove GK, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wieczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 Harnden P, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56 Conference tester
- Conference paper
- 14 Christensen S, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191
- Electronic journal (list all authors)
- 15 Morse SS. Factors in the emergence of infectious diseases. Emerg Infect Dis serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: http://www.cdc.gov/ ncidod/eid/index.htm

Patent (list all authors)

16 Pagedas AC, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1



Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express t test as t (in italics), F test as F (in italics), chi square test as χ^2 (in Greek), related coefficient as r (in italics), degree of freedom as v (in Greek), sample number as n (in italics), and probability as P (in italics).

Units

Use SI units. For example: body mass, m (B) = 78 kg; blood pressure, p (B) = 16.2/12.3 kPa; incubation time, t (incubation) = 96 h, blood glucose concentration, c (glucose) $6.4 \pm 2.1 \text{ mmol/L}$; blood CEA mass concentration, p (CEA) = 8.6 24.5 µg/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formal-dehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23243641.

The format for how to accurately write common units and quantums can be found at: http://www.wjgnet.com/2220-3214/g_info_20100725073806.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *t* concentration, A area, *l* length, *m* mass, *V* volume.

Genotypes: gyr.A, arg 1, c myc, c fos, etc. Restriction enzymes: EcoRI, HindI, BamHI, Kbo I, Kpn I, etc. Biology: H. pylori, E coli, etc.

Examples for paper writing

All types of articles' writing style and requirement will be found in the link: http://www.wignet.com/esps/NavigationInfo.aspx?id=15

RESUBMISSION OF THE REVISED MANUSCRIPTS

Authors must revise their manuscript carefully according to the

Instructions to authors

revision policies of BPG. The revised version, along with the signed copyright transfer agreement, responses to the reviewers, and English language Grade A certificate (for non-native speakers of English), should be submitted to the online system *via* the link contained in the e-mail sent by the editor. If you have any questions about the revision, please send e-mail to esps@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2220-3214/g_info_20100725073726.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2220-3214/g_info_20100725073445.htm.

Proof of financial support

For paper supported by a foundation, authors should provide a copy of the document and serial number of the foundation.

STATEMENT ABOUT ANONYMOUS PUBLICA-TION OF THE PEER REVIEWERS' COMMENTS

In order to increase the quality of peer review, push authors to carefully revise their manuscripts based on the peer reviewers' comments, and promote academic interactions among peer reviewers, authors and readers, we decide to anonymously publish the reviewers' comments and author's responses at the same time the manuscript is published online.

PUBLICATION FEE

WJR is an international, peer-reviewed, OA online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium and format, provided the original work is properly cited. The use is non-commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 698 USD per article. All invited articles are published free of charge.





Published by Baishideng Publishing Group Inc

8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

