# **Journal of Vegetation Science**

# Dispersal syndrome influences the match between seed rain and soil seed bank of woody species in a Neotropical dry forest.

Journal:	Journal of Vegetation Science
Manuscript ID	JVS-S-05080
Manuscript Type:	Special feature article
Date Submitted by the Author:	09-Sep-2019
Complete List of Authors:	Jara-Guerrero, Andrea; Universidad Técnica Particular de Loja, Departamento de Ciencias Biológicas Espinosa, Carlos Iván; Universidad Técnica Particular de Loja, Ciencias Biológicas Méndez, Marcos; Universidad Rey Juan Carlos, Área de Biodiversidad y Conservación de la Cruz, Marcelino; Universidad Rey Juan Carlos Escudero, Adrián; Universidad Rey Juan Carlos, Área de Biodiversidad y Conservación
Keywords:	Regeneration strategies, seed dispersal, seed dispersal phenology, seed rain density, seed bank density, seasonally dry tropical forest, similarity, species composition



# 1 Dispersal syndrome influences the match between seed rain and soil seed bank of woody

- 2 species in a Neotropical dry forest
- 3 Seed compartments and vegetation similarity
- 4 Andrea Jara-Guerrero<sup>1</sup>, Carlos I. Espinosa<sup>1</sup>, Marcos Méndez<sup>2</sup>, Marcelino De la Cruz<sup>2</sup>, Adrian
- 5 Escudero<sup>2</sup>.
- 6
- 7 <sup>1</sup>Departamento de Ciencias Biológicas, Universidad Técnica Particular de Loja, CP.: 11-01-
- 8 608, Loja, Ecuador.
- 9 <sup>2</sup>Área de Biodiversidad y Conservación, Universidad Rey Juan Carlos, E-28933 Móstoles,
- 10 Spain.
- 11 Correspondence
- 12 Carlos I. Espinosa, Departamento de Ciencias Biológicas, Universidad Técnica Particular de
- 13 Loja, CP.: 11-01-608, Loja, Ecuador.
- 14 E-mail: <u>ciespinosa@utpl.edu.ec</u>
- 15
- 16 Funding information
- 17 Agencia Española de Cooperación Internacional y para el Desarrollo (AECID) (A/024796/09
- and A/030244/10); Spanish Ministerio de Ciencia, Islas-Espacio (CGL2009-13190-C03-02)
- 19 and ROOTS (CGL2015-66809-P); REMEDINAL 3-CM (CAM S2013/MAE-2719, 2014-
- 20 2018); Universidad Técnica Particular de Loja (PROY\_CCNN\_1054).
- 21
- 22
- 23

# 24 ABSTRACT

25	Aim
23	7 11111

26	Long-distance seed dispersal and the ability to form soil seed banks are alternative strategies
27	to plant regeneration. These strategies are known to be mediated by adaptations of seed
28	dispersal and reproductive phenology, which if analyzed together can provide valuable
29	information about the regeneration strategies of species. We explored whether dispersal
30	syndrome and/or seed dispersal phenology control the similarity between standing vegetation,
31	seed rain and soil seed bank in a seasonally dry tropical forest (SDTF) in Southern Ecuador.
32	Location
33	Arenillas Ecological Reserve, Southwestern Ecuador, the biogeographic region of Pacific
34	Coastal Ecuador.
35	Methods
36	We recorded standing vegetation of woody species in a 9 ha permanent plot and sampled soil
37	seed bank from 265 cores of 0.06 m2. Additionally, we sampled seed rain along 1 yr from 265
38	seed traps of 0.64 m2. We evaluated similarity in species composition, species richness, and
39	abundance between the three compartments for all species and for each dispersal syndrome.
40	Results
41	Our studied community showed a high similarity in species composition between
42	compartments. The similarity was influenced by dispersal syndrome and the seed dispersal

43 phenology. Seed rain – standing vegetation similarity was high in species that dispersed in the

- 44 rainy season, such as zoochorous species, while seed bank standing vegetation was higher
- 45 for authochorous species compared with the other dispersal syndromes.
- 46 Conclusions
- 47 In our SDTF, dispersal phenology seemed to influence whether seeds had direct germination
- 48 or entered the soil seed bank and influenced the match between seed rain and soil seed bank,
- 49 suggesting different regeneration strategies related to dispersal ability.
- 50 Key words
- 51 Regeneration strategies, seed dispersal, seed dispersal phenology, seed rain density, seed bank
- 52 density, seasonally dry tropical forest, similarity, species composition.

imilan, j

#### 54 Introduction

Seed dispersal is critical for the recruitment of new individuals (Schupp & Fuentes, 1995; 55 56 Nathan & Muller-Landau, 2000) and a key process for the regeneration of plant populations and communities (Howe & Smallwood, 1982). Recruitment may occur immediately upon the 57 arrival of seeds to the ground after seed release from the mother plant (*i.e.* seed rain), or be 58 delayed through the formation of transient or permanent soil seed banks (Uhl et al., 1981; 59 Álvarez-Buylla & Martínez-Ramos, 1990; Moles & Drake, 1999; Silva et al., 2016; Vandvik 60 et al., 2016). The relative importance of these two primary regeneration sources conform a 61 remarkable evolutionary dilemma with variations among habitats (Thompson, 2000; Vandvik 62 et al., 2016), and its clarification is a crucial step to understand the community structure and 63 composition and to apply appropriate management measures. 64 Several studies have focused on the understanding of the structure and composition of plant 65 communities based only in the standing vegetation (Bakker et al., 1996; Dungan et al., 2001; 66 Savadogo et al., 2017). However, as seed availability is a first ecological filter for plant 67 68 regeneration, the understanding the local structure and composition of plant communities requires a joint consideration of the standing vegetation, the spatial extent achieved by 69 dispersal (seed rain) and the temporal exploration obtained through the existence of soil seed 70 banks (Bakker et al., 1996; Nathan and Muller-Landau, 2000; Thompson, 2000; 71 Hopfensperger, 2007; Pärtel, 2014; Vandvik et al., 2016; Savadogo et al., 2017). 72 The match among these three community compartments -standing vegetation, seed rain and 73 soil seed bank- will depend on the filters exerted by the local environment on the regional 74 floristic pool along the history of the plant community (Caballero et al., 2008). Tight matches 75 among standing vegetation and soil seed banks have been documented in some pioneer 76

communities such as those of ephemerals in some drylands (Caballero et al., 2008). 77 Nevertheless, in mature communities, the match can be low due to inputs of seeds coming 78 from surrounding sites and communities, or due to the long-standing permanence in the soil 79 seed bank of species from previous successional stages (Beatty, 1991; Thompson, 2000; 80 81 Laskurain et al., 2004; Martins & Engel, 2007, Gomes et al., 2019), even if they are not currently present in the standing vegetation (Pärtel, 2014). Mismatches can also emerge from 82 the differential ability of plant species to establish soil seed banks. This implies that if most of 83 the species in the community rely on dispersal for their regeneration, the similarity between 84 standing vegetation and seed rain will be high, but the similarity between standing vegetation 85 and seed soil bank will be low. 86

The ability to establish soil banks has been studied in relation to a few plant functional traits. 87 e.g., regeneration guild (Vázquez-Yanes & Orozco Segovia, 1984, 1993; Garwood, 1989; 88 Chandrashekara & Ramakrishnan, 1993; Rees, 1997), life form (Garwood, 1989; Rees, 1997; 89 Thompson, 2000; Silva et al., 2016), seed chemistry (Thompson, 2000), seed size and 90 dispersal capacity (Bakker et al., 1996; Thompson, 2000; Valenta et al., 2015; Douh et al., 91 2018; Ssali et al., 2018), and phenology (Heydel & Tackenberg, 2017). For instance, 92 93 observational studies suggested that large seed size and long dispersal distance should be negatively related to persistence in soil (Klinkhamer et al., 1987; Bakker et al., 1996; 94 Thompson, 2000). In this sense, species from mature stages typically have bigger and heavier 95 seeds than pioneer species, which germinate immediately after dispersal without entering the 96 soil bank, thereby avoiding seed eaters (Swaine & Hall, 1983; Teketay, 2005). However, 97 98 information is scarce for those more complex interactions between plant functional traits and the establishment of soil seed banks. In particular, a relationship between dispersal syndrome 99 100 and persistence in the soil can be mediated by the tight evolutionary connection of dispersal

syndrome with dispersal phenology (Heydel & Tackenberg, 2017; Douh et al., 2018).
Namely, by confining dispersal to periods with relatively low air humidity and windy
weather, anemochorous species seem to benefit by increased flying distances. In autochorous
species, the formation of soil seed banks would be unnecessary when optimal conditions for
dispersal match those required for germination (Bakker et al., 1996).

It has been hypothesized that in seasonally dry tropical forest (SDTF) soil seed banks are 106 more important than in tropical rain forests due to the need of released seeds to survive the 107 dry and unfavourable season (Vázquez-Yanes & Orozco Segovia, 1984). However, the few 108 109 studies available have shown that soil seed banks of woody species are also variable among SDTFs. While some studies have revealed a high presence of standing woody species in the 110 soil bank (Garwood, 1989), other studies found a low similarity between the seed bank and 111 standing vegetation (Rico-Gray & García-Franco, 1992; Uasuf et al., 2009; Savadogo et al., 112 2017; Gomes et al., 2019). Among other factors, these discrepancies could be related to 113 114 differences in the seed banking ability of the different dispersal syndromes (Valenta et al., 2015). Unfortunately, the few works exploring the relationship between dispersal syndrome 115 and soil permanence of seeds in SDTF have not provided a clear explanation for these 116 discrepancies (Antunes et al., 1998; Barbosa, 2003; Rodrigues da Luz & Ferreira-Nunes, 117 2014; Valenta et al., 2015). In our opinion, such contradictory results are the consequence of 118 ignoring the connection between dispersal syndrome and dispersal phenology. If the need to 119 establish a soil seed bank depends on the lag between seed dispersal and the optimal 120 conditions for germination, species dispersing during the dry months should dominate seed 121 122 banks. In this sense, anemochorous trees are good candidates to be part of soil banks since they tend to disperse into periods with relatively low humidity and windy weather (Heydel & 123 Tackenberg, 2017), which in SDTFs coincides with the dry season. In SDTFs the dry season 124

125 has optimal conditions for dispersal of anemochorous species but poor germination conditions that induce the entrance of these seeds in the soil seed bank. Contrarily, the formation of soil 126 seed banks would be unnecessary for zoochorous species, in which seed dispersal occurs in 127 the rainy season, precisely during the activity peak for most animal populations. Thus, a 128 complex interaction between dispersal syndrome, dispersal phenology, and soil seed bank 129 formation could be operating in tropical dry forests. Such information is critically needed in 130 SDTF of the Neotropic, where forest remnant reach only a 10% of original distribution 131 (Banda et al., 2016), and where the common practice of direct seedling sowing is ineffective 132 due to high desiccation (Vieira et al., 2008). The knowledge of seed bank and seed rain 133 structure and its role in natural regeneration can be useful to implementation of alternative 134 restoration strategies (Vieira et al., 2008). 135

In this study, we quantified standing vegetation, seed rain, and soil seed bank in a 9 ha forest plot of SDTF with the aim to examine to what extent these two primary seed sources –seed rain and seed bank– are contributing to the community assemblage, and how the dispersal phenology and dispersal syndrome can explain those contributions. We developed this study in a small-protected area of SDTF in Southwestern Ecuador. This area represents one of the last SDTF remnants below 100 m a.s.l. along the South America pacific coast. We considered this area might be useful as reference site of "pristine forest" to guide restoration actions.

143 Materials and Methods

## 144 Study site

145 The study was conducted in the Arenillas Ecological Reserve (REA for its acronym in

146 Spanish), which is located in the southwestern most tip of Ecuador (03° 34' 15.44''S; 80° 08'

147 46.15"E, 30 m a.s.l.) (Fig. 1). Mean annual rainfall is 667 mm, with a dry season extending

148 from June to December (with less than 40 mm during this 7-month period). Annual mean temperature is  $25^{\circ}$ C (Huaquillas weather station, n = 45 years). REA holds one of the last 149 remnants of SDTF in the Ecuadorian coast, as well as lowland dry scrub communities and 150 mangroves in the lowest areas (Sierra, 1999). It was included within the Ecuadorian 151 Patrimony of the State Natural Areas in 2001 (BirdLife International, 2013) and has been 152 protected from extractive activities and man-driven perturbations for about 60 years. In the 153 center of REA, in one of the best-conserved sectors, we established a 9 ha permanent plot 154 (300 x 300 m) consisting of transitional vegetation between dry deciduous forest and lowland 155 156 scrub (Fig. 1).



157

Fig. 1 Left map: location of the Arenillas Ecological Reserve (square) in Ecuador. Central map: location of the permanent plot in "Pintag Nuevo" (black dot). The draft on the right shows the permanent plot with the spatial location of each individual (light gray dots) in the 300 x 300 m plot and shrubs (dark gray dots) in the central 100 x 100 m plot. The black dots show the location of sampling points for seed rain and soil bank.

### 163 Sampling of the standing vegetation

In the 9 ha of the permanent plot, all tree individuals with a diameter at breast height (DBH)  $\geq$ 5 cm were recorded in 2009 (Fig. 1). Additionally, in the central 1 ha plot, we recorded all shrub and subshrub individuals with DBH > 1 cm.

#### 167 Sampling of the seed rain

168 Seed traps were used to sample the seed rain. Traps were made of a mesh fabric with holes <

169 0.5 mm to allow water runoff while preventing seed loss. The fabric was placed on a square

metal frame 80 cm long  $(0.64 \text{ m}^2)$  located 80 cm above the ground (Stevenson & Vargas

171 2008). A heavy object was placed in the center of each trap to prevent seed loss by wind.

We placed 265 traps in a regular grid in an area of 220 x 220 m of forest (4.8 ha), in the central part of the 9 ha plot (Fig. 1). Traps were located in each corner and the center of each 20 x 20 m cells of the grid. The total sampled area for seed rain was 169.6 m<sup>2</sup>. Seed rain was collected monthly between August 2011 and July 2012 for 12 surveying times. All seeds falling into the traps were identified (see below) and counted. When the dispersal units were

177 whole fruits or their parts, the number of seeds contained in them was counted.

## 178 Sampling of the soil seed bank

Soil seed bank was sampled once between November and December 2011, at the end of the dry season, when most seeds had been dispersed and before germination started in the rainy season. This sampling date corresponds with the moment in which the soil seed samples reach their maximum diversity and density, i.e., it includes both the transient and the persistent soil seed bank (Caballero et al., 2005; Espinosa et al., 2013). As in the case of the seed rain, 265

soil samples were collected in 0.25 x 0.25 x 0.03 m squares located adjacent to the seed traps (Fig. 1), adding up to a total sampled area of 16.56 m<sup>2</sup>.

Seed bank composition was assessed using the germination method, which allows estimating 186 viable seeds in the soil (Bigwood & Inouve, 1988; Holzapfel et al., 1993; Caballero et al., 187 2008). Soil samples were placed in trays with a basal layer of pumice to control the moisture. 188 Travs were kept under controlled watering for six months, and emerging seedlings were 189 identified, counted and removed at 3-4 day intervals (Caballero et al., 2008). After six 190 months, the remaining material of each sample was watered with a gibberellic acid (GA3) 191 solution (1000 ppm) to stimulate germination in seeds with secondary dormancy (Caballero et 192 al., 2008). From here on, soil samples were kept under watering for two additional months. 193 Unidentified seedlings were transplanted for subsequent identification. 194

#### 195 Statistical analyses

From the mapping of the standing vegetation, we obtained the number of species present in the permanent plot, and the density (individuals per hectare) and basal area by species. Both density and basal area were estimated from the 9 ha plot for tree species and from the central 1 ha plot for shrub species. We also assembled two data sets for the seed density, one in the seed rain and another one in the soil bank. The density of each species in the three compartments –standing vegetation, seed rain and soil bank– was transformed by dividing each density value by that of the most abundant species in each compartment.

203 According to fruit and seed features, and direct observation in the field, species were

- 204 classified into three dispersal syndromes: zoochory, anemochory and autochory (Jara-
- Guerrero et al., 2015). To assess the contribution of seed rain and soil bank to woody
- 206 community assemblage we considered the similarity in species composition between the three

207	compartments. We used the Sørensen similarity index (Oksanen et al., 2012) with presence-
208	absence. To evaluated similarities based in abundance, we calculated Bray-Curtis similarity
209	using the transformed density of the three compartments, and also, comparing the basal area
210	of standing vegetation and transformed density of seed rain and seed bank. Both similarity
211	indices range from 0 to 1, where 1 indicates identical composition. The similarity was
212	analyzed both for all species together and for each dispersal syndrome separately.
213	We used an extension of the Fisher exact test for large contingency tables to test differences
214	in the frequency of species among dispersal syndromes in the three compartments.
215	Significance was obtained from 2000 Monte Carlo simulations (R Core Team, 2016).
216	To analyze the temporal pattern in species composition of the seed rain we used time-
217	constrained agglomerative clustering analysis (Legendre & Legendre, 1998), following
218	Ward's (1963) method. First, we computed a Euclidean distance matrix by using a species-
219	by-month matrix, with the total number of seeds for each species recorded in each matrix cell.
220	Second, to find the best cluster solution for the annual seed rain data, we tested clustering
221	results with 2 to 6 groups using cross-validation with 100 iterations for each set of groups.
222	The clustering with the lowest cross-validation residual error was retained. This analysis was
223	performed with the 'const.clust' package (Legendre, 2011). Third, we assessed whether any
224	species was significantly associated with the temporal clusters revealed previously, using the
225	"signassoc" function, from the 'indicspecies' package (De Cáceres and Legendre, 2009). The
226	null hypothesis tested was that the average abundance of a species was not higher in the target
227	month group than in other groups. All statistical analyses were carried out with the R
228	environment, version 3.2.4 (R Core Team, 2016).

229

#### 230 **Results**

# 231 Species richness and abundance in the standing vegetation, seed rain and seed bank

A total of 47 species were recorded pooling the three compartments studied, from which 23

were trees and 24 were shrubs (Appendix S1). All of them were present in the standing

vegetation, with 8211 individuals. *Handroanthus billbergii* and *Croton rivinifolius* were the

most abundant tree and shrub with 57 and 721 individuals per hectare, respectively (Appendix

S1). In the seed rain, we collected 15083 seeds (88.9 seeds  $m^{-2}$ ) from 33 woody species, with

a mean density ( $\pm$  standard deviation) per species of 2.69 ( $\pm$  4.95) seeds m<sup>-2</sup> (Appendix S1).

238 The main contribution to the seed rain came from *Alternanthera* sp., *Croton* spp. and

*Erythroxylum glaucum* (Appendix S1). In the seed bank, we recorded 613 emerged seeds

240 (37.01 seeds m<sup>-2</sup>) from 15 woody species, with a mean density ( $\pm$  standard deviation) of 2.47

 $\pm$  3.9 seeds m<sup>-2</sup> (Appendix S1). The main contribution to the seed bank came from

242 Alternanthera sp. and Croton spp. (Appendix S1). The only taxon that was abundant in the

three compartments was *Croton* spp.

## 244 Similarity among standing vegetation, seed rain and soil seed bank

All species recorded both in seed rain and in soil bank occurred in the standing vegetation; 33
were present in the seed rain and 15 in the soil bank. Seeds of 19 species from the standing
vegetation were recorded only in the seed rain, and seeds of one species (*Piptadenia flava*)
were recorded only in the soil bank. Fourteen species of the standing vegetation were absent
in both the seed rain and the seed bank, including three dominant trees (the two *Handroanthus*species and *Cynophalla mollis*).

In general, Sørensen similarity index based on presence-absence data showed a relatively high
 similarity among compartments and decreased from standing vegetation–seed rain through

256

seed rain-seed bank to standing vegetation-seed bank (Fig. 2A). For transformed density and

basal area data, the seed rain and soil seed bank were the components with the highest

similarity (Fig. 2B, C).



Fig. 2 Similarity in species composition among standing vegetation (SV), seed rain (SR) and
seed bank (SB) considering all species or each dispersal syndrome separately. Results are
based on presence-absence (A), density of the three compartments (B) and basal area of
species in standing vegetation (C). Width arrows is proportional to similarity.

# 261 Dispersal syndromes and similarity among compartments

262 The number of species of each dispersal syndrome did not vary significantly among

- 263 compartments (Table 1, Fisher's exact test, p = 0.55). Zoochory was the dispersal syndrome
- with most seeds in the seed rain, while the anemochory showed a highest density in the soil

- bank (Fig. 3A). Autochory was the dispersal syndrome with the highest density in the
- standing vegetation, while anemochory was the syndrome with the highest accumulated basal
- area (Fig. 3B, Appendix S1).
- **Table 1.** Number of species (percentage) of each dispersal syndrome in each compartment
- studied: standing vegetation, seed rain and seed bank.



270

Fig. 3 Abundance of the three dispersal syndromes in (A) the seed rain (black bars) and the
soil seed bank (grey bars) and (B) in the standing vegetation estimated as basal area (black
bars) or density (grey bars).

274 For anemochory and zoochory, Sørensen similarity index based on presence-absence data showed a relatively high similarity among compartments and decreased from standing 275 vegetation-seed rain through seed rain-soil bank to standing vegetation-soil bank (Fig. 2A). 276 From the three dispersal syndromes, autochorous species showed the highest similarities, with 277 standing vegetation and seed rain relationship showing the highest similarity. When we 278 considered the transformed density and basal area data, similarity among compartments was 279 dependent on dispersal syndrome (Fig. 2B, C). Autochorous species showed the highest 280 similarity between compartments, followed by anemochorous species (Fig. 2bB, C). 281 Zoochorous species showed the lowest similarities among compartments (Fig. 2B, C). The 282 match between standing vegetation and seed compartments was stronger with density than 283 with basal area. 284

# 285 Temporal variation in the seed rain

Seed rain fluctuated monthly in species richness (4–17 species: Fig. 4A) and seed abundance
(1.3–27.9 seeds m<sup>-2</sup>: Fig. 4B). The highest species richness was recorded at the transition from
the rainy to the dry season (between May to June), and the lowest in December, just before
the beginning of the rains (Fig. 4A). Seed abundance collected in traps showed a bimodal
pattern, reaching a maximum in February (rainy season) and another in July (dry season),
while the minimum was observed in December (transition between dry and rainy season)
(Fig. 4B).



293

Fig. 4 Temporal variation in woody species number (A) and abundance (B-E) in the seed rain.
Circles represent total richness, triangles represent anemochorous, quadrates represent
autochorous and diamonds represent zoochorous species. The gray area indicates the rainy
season. Wiskers indicate ± 1 standard error.

298 Chronological constrained clustering analysis separated three well-defined groups of months 299 regarding seed rain composition (cross-validation residual error = 0.773). The first group 300 corresponded to the months of the dry season, from August to December. The second group 301 corresponded to the beginning of rainy season, from January to March; while months of the 302 third group, April to July, corresponded to the transition from the rainy to dry season (Fig. 5). 303 Thirteen species showed a significantly positive association to one of the three compositional 304 groups (Fig. 5, Appendix S2). Among them, the anemochorous species were associated to the 305 first group (dry months), while zoochorous and autochorous species were associated to the second (rainy months) or third group (transition from the rainy to dry season). Along the first 306 months of the dry season we recorded a very low seed density of zoochorous species, most of 307 them corresponded to species that started the dispersal process during the rainy months. We 308 did not record seeds of zoochorous species at the end of the dry season (November and 309 December) (Fig. 4). By contrast, anemochorous species were recorded during the dry months, 310 and only two species continued their dispersal once the rains started. Only autochorous 311 species were present in the seed rain throughout the year, with a peak in richness at the 312 313 transition from the rainy to dry season (Fig. 4).



Fig. 5 Dendrogram presenting the results of the seed rain partitioning between months.

Species significantly associated are shown for each group. \*: p < 0.05, \*\*: p < 0.01; \*\*\*: p <</li>
0.001.

#### 318 DISCUSSION

- 319 As expected, the seed rain of woody species in this STDF showed a seasonal variation in
- species number and seed abundance. As observed in other studies (Swaine & Hall, 1983;
- 321 Douh et al., 2018; Escobar et al., 2018), in our SDTF the timing of seed dispersal seems to be

an important factor defining the input of seeds to the soil bank. Seed release was staggered along the year, with a dominance of anemochorous species in the dry season and zoochorous species in the wet season. In relation to this staggering, abundance in the soil seed bank was maximal for anemochorous woody species and minimal for zoochorous ones. Overall, these findings suggest that dispersal syndromes are a major factor explaining mismatches between abundance in the seed rain and in the soil bank in tropical dry forests.

The timing of seed release differed among dispersal syndromes, with most of zoochorous 328 329 species dispersing during the rainy season while anemochorous species dispersed in the dry season. This temporal segregation of dispersal syndromes has been found in temperate 330 ecosystems (Heydel & Tackenberg, 2017) and, to different extents, in other SDTFs (Frankie 331 et al., 1974; Gottsberger & Silberbauer-Gottsberger, 1983; Griz & Machado, 2001; Carvalho 332 & Sartori, 2015; see, however, Escobar et al., 2018 and Cortes-Flores et al., 2019 for an 333 availability of fleshy fruits throughout the year in other seasonally dry ecosystems of 334 Neotropic). This phenological separation of dispersal syndromes fits long-hold expectations 335 for reproductive events in SDTFs, that have traditionally been considered to occur at 336 337 opportune times from a performance perspective (Janzen, 1967). The rainy season coincides with the maximal activity of seed dispersers for zoochorous species, while the dry season 338 provides leafless canopies and windy conditions suitable for long-distance dispersal in 339 340 anemochorous species (Frankie et al., 1974; Gottsberger & Silberbauer-Gottsberger, 1983; Du et al., 2009). 341

Nevertheless, it is difficult to adjust all phases of a reproductive event, from flowering to seed
germination, to match the most suitable conditions for each phase. For example, flowering
phenology can constrain fruiting phenology (Primack, 1985; Bolmgren & Cowan, 2008;
Bolmgren & Eriksson, 2015). In our SDTF, dispersal timing seemed to influence subsequent

346 events, namely direct germination vs. entrance in the soil seed bank, and influenced the match between seed rain and soil seed bank. In SDTFs, a distinction has been suggested between 347 rapid-rainy and intermediate-dry syndromes as a function of the delay between seed release 348 and germination (Garwood, 1983). By dispersing and germinating during the rainy season, 349 zoochorous species were apparently following a rapid-rainy syndrome, adjusted to the 350 maximum size of animal populations. In addition, synchrony between dispersal and rain 351 maximizes the chances of seedling establishment and recruitment (Howe & Smallwood, 1982; 352 Garwood, 1983; Bullock, 1995; Fornara & Dalling, 2005; Jara-Guerrero et al., 2011; Espinosa 353 et al., 2012). This result concurs with the extended idea that zoochorous species depend 354 mainly on seed rain for regeneration (Bakker et al., 1996; Thompson, 2000). In our study, the 355 only three zoochorous species found in the soil bank were *Lantana* sp., which released seeds 356 during the dry season and Bursera graveolens and Cordia macrocephalla, which dispersed at 357 the end of the rainy season. Nevertheless, the abundance of the latter two species in the soil 358 bank was very low. All of them have a hard seed coat, a morphological trait that is considered 359 as a typical strategy to survive the dry season (Garwood, 1983; Baskin & Baskin, 1998), and 360 thus to facilitate the incorporation of seeds into soil banks. 361

Anemochorous species showed two strategies of dispersal timing. The most common strategy 362 was to disperse during the dry season when conditions are very poor to germination but very 363 favorable for efficient dispersal as the canopy is mostly open. These dry conditions lead to an 364 entrance of anemochorous seeds in the soil seed bank, at least in a transient way until the next 365 rainy season, i.e., an intermediate-dry syndrome (Garwood, 1983). The other strategy of this 366 guild was to disperse just immediately before or at the beginning of the rainy season. Species 367 with the latter strategy were characterized by large seeds (> 0.5 cm), i.e., *Handroanthus* 368 chrysantha, H. billbergii, Ceiba trischistandra and Eriotheca ruizii (Romero-Saritama, 2016). 369

Page 20 of 39

Jara-Guerrero et al. - 20

370 In this case, seeds find conditions for rapid germination and a relatively large period for guaranteeing seedlings with large sizes enough for surviving the first dry season. Survival to 371 this critical period is usually size dependent since large seedlings normally have a higher 372 chance of survival (Escudero et al., 1999, 2000). 373 In the case of autochorous species, which were dominant in the three studied compartments, 374 no clear seasonal pattern was found in the seed rain, suggesting a high variation in 375 germination requirements and regeneration strategies (Escobar et al., 2018). It is widely 376 377 accepted that interspecific competition promotes strong selection for differentiation in resource use (Grant, 1972; Pfennig & Pfennig, 2009). This character displacement (Brown & 378 Wilson, 1956) can promote evolutionary divergence in phenology of seed dispersal between 379 species with the same dispersal syndrome (see Snow, 1965, for similar reasoning concerning 380 zoochorous species). Thus, the formation of soil seed banks is advantageous in autochorous 381 382 species because it decreases competition caused by their limited dispersal capacity and keeps the seed viability during unfavorable periods for germination and seedling development 383 (Venable & Brown, 1988; Bakker et al., 1996). This strategy is particularly important in 384 highly stochastic environments such as SDTF, where the bet-hedging strategy typical of 385 species with soil banks dictates a sacrifice in the mean fitness to reduce fitness variance 386 (Evans et al., 2007). 387

The similarity between seed rain, soil seed bank and standing vegetation in our SDTF was higher than values reported for forest ecosystems (45% vs. an average of  $31\% \pm 3.7$ :

Hopfensperger, 2007), and among the highest for tropical forests (Hardesty & Parker, 2002;
Hopfensperger, 2007; Gomes et al., 2019). This remarkably high similarity can be related to
two not mutually exclusive factors. First, the input of seeds came mainly from local standing
vegetation, and there are no hidden species in the soil seed bank (Pärtel, 2014). In this SDTF,

394 dispersal was not very efficient in any species independently of the dispersal syndrome (Jara-Guerrero et al., 2015). Second, this high similarity could be at least partially associated with a 395 small degree of disturbance (Hopfensperger, 2007) in this well-preserved forest. In this sense, 396 high similarity could prevent the presence of pioneer species in the soil seed bank. Although 397 scarce, available information about the similarity between standing vegetation and soil seed 398 banks in SDTF is contradictory, with some studies showing high similarity (Garwood 1989) 399 and others finding a low similarity (Rico-Gray & García-Franco, 1992; Uasuf et al., 2009; 400 Savadogo et al., 2017, Gomes et al., 2019). This apparent discrepancy could be explained, at 401 least partially, if the effects of dispersal syndrome and dispersal phenology are explicitly 402 considered. 403

Although comparisons between these community compartments sometimes are based on 404 presence-absence data sets (e.g. Hall & Swaine, 1980; Dungan et al., 2001; Grombone-405 Guaratini et al., 2004; Reubens et al., 2007, Uasuf et al., 2009, Greulich et al. 2019), the 406 explicit consideration of the area basal and density as complementary estimates of 407 interspecies differences in performance provided new insights. First, these estimates 408 409 introduced important differences with the presence-absence comparisons. Second, these functional estimates weakened the strong match found in all the cases between the standing 410 vegetation and seed rain, which suggests that variations in the reproductive performance 411 412 between species reduce the similarity. In general, the density of standing vegetation was better matched with the abundance in the seed bank than the total basal area. This suggests 413 that the number of individuals per species, rather than the basal area, is a better descriptor of 414 the total number of seeds released. 415

In conclusion, the dispersal syndrome was a critical determinant of the timing of eventsrelated to seed dispersal, seed banking and germination in the studied SDTF. The

418 phenological staggering of seed release by different dispersal syndromes led to different window opportunities for direct germination or delayed emergence and, thus, for the presence 419 of species in the soil seed bank. Apparently, conditions for effective seed dispersal set the 420 stage in which subsequent phases of the reproductive event develop in these highly seasonal 421 environments. Earlier events can indirectly shape seed traits related to persistence in soil. 422 More studies about seed bank persistence are needed to determine the adaptive potential in 423 population terms of this separation between seed maturation, release, and emergence. In any 424 case, it seems evident that the correct interpretation of soil seed banks in SDTFs on the longer 425 demographic terms need to be evaluated considering the differences in dispersal ability of co-426 occurring species. 427

# 428 ACKNOWLEDGMENTS

We would like to thank to Jorge Armijos, Dalton Quizhpe, Santiago Erazo for practical
assistance in the field. We gratefully acknowledge to Ministerio del Ambiente of Ecuador for
access facilities to the REA.

## 432 AUTHOR CONTRIBUTIONS

433 AJG, MC, MM, and CE conceived and designed research; AJG, and CE performed research;

434 AJG, MC, MM, CE, and AE analyzed data; AJG, MC, MM, CE, and AE wrote the paper.

# 435 SUPPORTING INFORMATION

436 Please see Appendix S1 for data from woody species present in the standing vegetation, seed

- 437 rain and soil seed bank, and Appendix S2 for a summary of the association analysis of species
- 438 in the compositional groups of seed rain.

#### 440 **REFERENCES**

- Álvarez-Buylla, E.R. & Martínez-Ramos. M. (1990). Seed bank vs seed rain in the
  regeneration of a tropical pioneer tree. *Oecologia*, 84, 314-325.
- Antunes, N.B., Ribeiro, J.F. & Salomäo, A.N. (1998). Caracterização de frutos e sementes
  deseis especes vegetais em matas de galeria do distrito federal. *Revista Brasileira de Sementes*, 20, 112–119.
- Bakker, J.P., Poschlod, P., Strykstra, R.J., Bekker, R. & Thompson, K. (1996). Seed banks
  and seed dispersal: important topics in restoration ecology. *Acta Botanica Neerlandica*, 45, 461–490.
- Banda, K., Delgado-Salinas, A., Dexter, K.G., Linares-Palomino, R., Oliveira-Filho, A.,
  Prado, D., et al. (2016). Plant diversity patterns in neotropical dry forests and their
  conservation implications. *Science*, 353, 1383–1387.
- Barbosa, D.C.A. (2003). Estratégias de germinação e crescimento de espécies lenhosas da
  caatinga com germinação rápida. In: I.R. Leal, M. Tabarelli, J.M.C. Silva (Eds). *Ecologia e conservação da caatinga* (pp. 625–656). Editora Universitaria da UFPE,
- 455 Recife.
- Baskin, C.C., & Baskin, J.M. (1998). Seeds: Ecology, biogeography and evolution of *dormancy and germination*. Academic Press, San Diego, California.
- Beatty, S.W. (1991). Colonization dynamics in a mosaic landscape: the buried seed pool. *Journal of Biogeography*, 18, 553–563.
- Bigwood, D.W. & Inouye, D.W. (1988). Spatial pattern analysis of seed banks: an improved
  method and optimized sampling. *Ecology*, 69, 497–507.
- 462 Birdlife International. (2013). Important Bird Areas factsheet: Reserva Ecológica Arenillas.
- 463 Downloaded from http://www.birdlife.org on 12/03/2013.

- Bolmgren, K., & Cowan, P.D. (2008). Time-size tradeoffs: a phylogenetic comparative study
  of flowering time, plant height and seed mass in a north-temperate flora. *Oikos*, 117,
  466 424–429.
- 467 Bolmgren, K. & Eriksson, O. (2015). Are mismatches the norm? Timing of flowering,
- 468 fruiting, dispersal and germination and their fitness effects in *Frangula alnus*469 (Rhamnaceae). *Oikos*, 124, 639–648.
- Brown, W.L. & Wilson, E.O. (1956). Character displacement. *Systematic Zoology*, 5, 49–64.
- 471 Bullock, S.H. (1995). Plant reproduction in Neotropical dry forests. In S. Bullock, H.
- 472 Mooney, E. Medina (Eds.), *Seasonally dry tropical forests* (pp. 277–297). Cambridge
- 473 University Press, Cambridge, UK.
- Caballero, I., Olano, J.M., Luzuriaga, A.L. & Escudero, A. (2005). Spatial coherence between
  seasonal seed banks in a semi-arid gypsum community: density changes but structure
  does not. *Seed Science Research*, 15, 153–160.
- 477 Caballero, I., Olano, J.M., Loidi, J. & Escudero, A. (2008). A model for small-scale seed bank
  478 and standing vegetation connection along time. *Oikos*, 117, 1788-1795.
- 479 Carvalho, F.S. & Sartori, A.L.B. (2015). Reproductive phenology and seed dispersal
  480 syndromes of woody species in the Brazilian Chaco. *Journal of Vegetation Science*,
  481 26, 302–311.
- Chandrashekara, U.M., & Ramakrishnan, P.S. (1993). Germinable soil seed bank dynamics
  during the gap phase of a humid tropical forest in the Western Ghats of Kerala, India. *Journal of Tropical Ecology*, 9, 455-467.
- 485 Cortés-Flores, J., Cornejo-Tenorio, G., Urrea-Galeano, L., Andresen, E., González-
- 486 Rodríguez, A., & Ibarra-Manríquez, G. (2019). Phylogeny, fruit traits, and ecological
- 487 correlates of fruiting phenology in a Neotropical dry forest. *Oecologia*, 189, 159–169.

Journal of Vegetation Science

488	De Caceres, M., & Legendre, P. (2009). Associations between species and groups of sites:
489	indices and statistical inference. Ecology, 90, 3566-3574.
490	Douh, C., Daïnou, K., Loumento, J.J., Moutsambote, J.M., Fayolle, A., Tosso, F., Forni, E.,
491	Gourlet-Fleury, S., & Doucet, J.L. (2018). Soil seed bank characteristics in two central
492	African forest types and implications for forest restoration. Forest Ecology and
493	Management, 409, 766–776.
494	Du, Y., Mi, X., Liu, X., Chen, L., Ma, K. (2009). Seed dispersal phenology and dispersal
495	syndromes in a subtropical broad-leaved forest of China. Forest Ecology and
496	Management, 258, 1147–1152.
497	Dungan, R.J., Norton, D.A., & Duncan, R.P. (2001). Seed rain in successional vegetation,
498	Port Hills Ecological District, New Zealand. New Zealand Journal of Botany, 39, 115-
499	124.
500	Escobar, D.F.E., Silveira, F.A.O., & Morellato, L.P.C. (2018). Timing of seed dispersal and
501	seed dormancy in Brazilian savanna: two solutions to face seasonality. Annals of
502	Botany, 121, 1197–1209.
503	Escudero, A. Somolinos, R., Olano, J.M., & Rubio, A. (1999). Factors controlling the
504	establishment of Helianthemum squamatum an endemic gypsophile of semi-arid
505	Spain. Journal of Ecology, 87, 290–302.
506	Escudero, A., Albert, M.J., Pita, J.M., & Pérez-García, F. (2000). Inhibitory effects of
507	Artemisia herba-alba on the germination of the gypsophyte Helianthemum
508	squamatum. Plant Ecology, 148, 71–80.
509	Espinosa, C.I., De La Cruz, M., Luzuriaga, A.L., & Escudero, A. (2012). Bosques tropicales
510	secos de la región Pacífico Ecuatorial: diversidad, estructura, funcionamiento e
511	implicaciones para la conservación. Ecosistemas, 21, 167-179.

512	Espinosa, C.I., Luzuriaga, A.L., De La Cruz, M., Montero, M., Escudero, A. (2013). Co-
513	occurring grazing and climate stressors have different effects on the total seed bank
514	when compared to the persistent seed bank. Journal of Vegetation Science, 24, 1098-
515	1107.
516	Evans, M.K., Régis, F., Kane, M.J. & Venable, D.L. (2007). Bet Hedging via Seed Banking
517	in Desert Evening Primroses (Oenothera, Onagraceae): Demographic Evidence from
518	Natural Populations. The American Naturalist, 169, 184–194.
519	Fornara, D.A. & Dalling, J.W. (2005). Seed bank dynamics in five Panamanian forests.
520	Journal of Tropical Ecology, 21, 223–226.
521	Frankie, G.W., Baker. H.G., & Opler, P.A. (1974). Comparative phenological studies of trees
522	in tropical wet and dry forests in the lowlands of Costa Rica. Journal of Ecology, 62,
523	881–919.
524	Garwood, N. (1983). Seed germination in a seasonal tropical forest in Panama: a community
525	study. Ecological Monographs, 53, 159–181.
526	Garwood, N.C. (1989). Tropical soil seed banks: a review. In: M.A. Lek, V.T. Parker, R.L.
527	Simpson (Eds). Ecology of soil seed banks (pp. 149–209). Academic Press, San Diego,

528 CA.

Gomes, F.M., Oliveira, C.C., Rocha Miranda, R., Costa, R.C., & Loiola, M.I.B. (2019).
Relationships between soil seed bank composition and standing vegetation along
chronosequences in a tropical dry forest in north-eastern Brazil. *Journal of Tropical Ecology*, 35, 173–184.

533 Gottsberger, G., & Silberbauer-Gottsberger, I. (1983). Dispersal and distribution in the

534 cerrado vegetation of Brazil. Sonderbände des Naturwissenshaftlichen Vereins in

535 *Hamburg*, 7, 315–352.

536	Grant, P.R. (1972). Convergent and divergent character displacement. Biological Journal of
537	the Linnean Society, 4, 39–68.

- Greulich, S., Chevalier, R., & Villar, M. (2019). Soil seed banks in the floodplain of a large
  river: a test of hypotheses on seed bank composition in relation to flooding and
  established vegetation. *Journal of Vegetation Science*, 30, 732–745.
- Griz, L.M.S., & Machado, I.C.S. (2001). Fruiting phenology and seed dispersal syndromes in
  caatinga, a tropical dry forest in the northeast of Brazil. *Journal of Tropical Ecology*,
- 543 17, 303–321.
- 544 Grombone-Guaratini, M.T., De Freitas Leitão Filho, H., & Kageyama, P. (2004). The seed
- bank of a gallery forest in southeastern Brazil. *Brazilian Archives of Biology and Technology*, 47, 793–797.
- Hall ,J.B., & Swaine, M.D. (1980). Seed stocks in Ghanaian forest soils. *Biotropica*, 12, 256–
  263.
- Hardesty, B., & Parker, V. (2002). Community seed rain patterns and a comparison to adult
  community structure in a West African tropical forest. *Plant Ecology*, 164, 49–64.
- Heydel, F., & Tackenberg, O. (2017). How are the phenologies of ripening and seed release
  affected by species' ecology and evolution? *Oikos*, 126, 738–747.
- Holzapfel, C., Schmidt, W., & Shmida, A. (1993). The role of seed bank and seed rain in the
  recolonization of disturbed sites along an aridity gradient. *Phytocoenologia*, 23: 561–
  580.
- Hopfensperger, K.N. (2007). A review of similarity between seed bank and standing
  vegetation across ecosystems. *Oikos*, 116, 1438–1448.
- Howe, H.F., & Smallwood, J. (1982). Ecology of seed dispersal. *Annual Review of Ecology and Systematics*, 13, 201-228.

560	Janzen D. H. (1967). Synchronization of sexual reproduction of trees within dry season in
561	Central America. Evolution, 21, 620-637.
562	Jara-Guerrero, A., De La Cruz, M., & Méndez, M. (2011). Seed dispersal spectrum of woody
563	species in South Ecuadorian dry forests: environmental correlates and the effect of
564	considering species abundance. Biotropica, 43, 722-730.
565	Jara-Guerrero, A., De La Cruz, M., Espinosa, C.I., Méndez, M. & Escudero, A. (2015). Does
566	spatial heterogeneity blur the signature of dispersal syndromes on spatial patterns of
567	woody species? A test in a tropical dry forest. Oikos, 124, 1360-1366.
568	Klinkhamer, P.G.L., De Jong, T. J., Metz, J.A.J., & Val, J. (1987). Life history tactics of
569	annual organisms: the joint effects of dispersal and delayed germination. Theoretical
570	Population Biology, 32, 127–156.
571	Laskurain, N.A., Escudero, A., Olano, J.M., & Loidi, J. (2004). Seedling dynamics of shrubs
572	in a fully closed temperate forest: greater than expected. <i>Ecography</i> , 27,650–658.
573	Legendre, P., & Legendre, L. (1998). Numerical ecology (2nd ed.). Elsevier.
574	Legendre, P. (2011). const.clust: space- and time-constrained clustering package. Available at
575	http://adn.biol.umontreal.ca/~numericalecology/Rcode/
576	Martins, A.M., & Engel, V.L. (2007). Soil seed banks in tropical forest fragments with
577	different disturbance histories in southeastern Brazil. Ecological engineering, 31, 165-
578	174.
579	Moles, A.T., & Drake, D.R. (1999). Potential contributions of the seed rain and seed bank to
580	regeneration of native forest under plantation pine in New Zealand. New Zealand
581	Journal of Botany, 37, 83–93.
582	Nathan, R., & Muller-Landau, H. (2000). Spatial patterns of seed dispersal, their determinants
583	and consequences for recruitment. Trends in Ecology and Evolution, 15, 278–285.

584	Oksanen, J., F. Guillaume Blanchet, Kindt, R., Legendre, P., Minchin, P., O'hara, R. B.,
585	Simpson, G., Solymos, P., Stevens, M., & Wagner, H. (2012). Package "vegan".
586	Community Ecology Package. URL <u>http://vegan.r-forge.r-project.org/</u>
587	Pärtel, M. (2014). Community ecology of absent species: hidden and dark diversity. Journal
588	of Vegetation Science, 25, 1154–1159.
589	Pfennig, K.S., & Pfennig, D.W. (2009). Character displacement: ecological and reproductive
590	responses to a common evolutionary problem. Quarterly Review of Biology, 84, 253-
591	276.
592	Primack, R.B. (1985). Relationships among flowers, fruits and seeds. Annual Review of
593	Ecology, Evolution, and Systematics, 18, 409–430.
594	R Core Team. (2016). R: A language and environment for statistical computing. R
595	Foundation for Statistical Computing, Vienna, Austria.
596	Rees, M. (1997). Seed dormancy. In: Crawley, M.J. (Ed.). Plant Ecology (pp. 214–238).
597	Blackwell Science.
598	Reubens, B., Heyn, M., Gebrehiwot, K., Hermy, M., & Muys, B. (2007). Persistent soil seed
599	banks for natural rehabilitation of dry tropical forests in northern Ethiopia.
600	<i>Tropicultura</i> , 25, 204–214.
601	Rico-Gray, V., & García-Franco, J.G. (1992). Vegetation and soil seed bank of successional
602	stages in tropical lowland deciduous forest. Journal of Vegetation Science, 3, 617-
603	624.
604	Rodrigues Da Luz, G., & Ferreira-Nunes, Y. R. (2014). Seed germination of arboreal-shrub
605	species with different dispersal mechanisms in a Brazilian tropical dry forest. In: A.
606	Sánchez-Azofeifa, J. S. Powers, G. W. Fernandes, & M. Quesada (Eds.). Tropical dry

607	forests in the Americas: ecology, conservation, and management (pp. 281–299). CRC
608	Press, Boca Ratón, FL.
609	Romero-Saritama, J.M. (2016). Caracterización morfofisiológica de semillas de especies
610	leñosas distribuidas en dos zonas secas presentes en el Sur del Ecuador. Ecosistemas,
611	25, 93–100.
612	Savadogo, P., Sanou, L., Dayamba, D., Bognounou, F., & Thiombiano, A. (2017).
613	Relationships between soil seed banks and above-ground vegetation along a
614	disturbance gradient in the W National Park trans-boundary biosphere reserve, West
615	Africa. Journal of Plant Ecology, 10, 349–363.
616	Schupp, E., & Fuentes, M. (1995). Spatial patterns of seed dispersal and the unification of
617	plant population ecology. Écoscience, 2, 267–275.
618	Sierra, R. (Ed.). (1999). Propuesta preliminar de un sistema de clasificación de vegetación
619	para el Ecuador Continental. Proyecto INEFAN/GEF-BIRF y EcoCiencia. Quito,
620	Ecuador.
621	Silva, E.P., Figueiredo, F.G., Séfara, S., & Fernandes, L. (2016). Evaluation of the potential
622	of seed rain as an alternative for forest restoration in Permanent Preservation Areas.
623	<i>Revista Árvore</i> , 40, 21–28.
624	Snow, D.W. (1965). A possible selective factor in the evolution of fruiting seasons in tropical
625	forest. Oikos, 15, 274–281.
626	Ssali, F., Stein, R.M., & Sheil, D. (2018). Tree seed rain and seed removal, but not the seed
627	bank, impede forest recovery in bracken (Pteridium aquilinum (L.) Kuhn)-dominated
628	clearings in the African highlands. Ecology and Evolution, 8, 4224–4236.
629	Stevenson, P., & Vargas, I. (2008). Sample size and appropriate design of fruit and seed traps
630	in tropical forests. Journal of Tropical Ecology, 24, 95-105.

631	Swaine, M.D., & Hall, J.B. (1983). Early succession of cleared forest land in Ghana. Journal
632	<i>of Ecology</i> , 71, 601–627.
633	Teketay, D. (2005). Seed and regeneration ecology in dry Afromontane forests of Ethiopia: I.
634	Seed production - population structures. Tropical Ecology, 46, 29-44.
635	Thompson, K. (2000). The functional ecology of soil seed banks. In: M. Fenner (Ed.). Seeds:
636	the ecology of regeneration in plant communities (2nd ed., pp. 215–235). CAB
637	International. UK.
638	Uasuf, A., Tigabu, M. & Odén. P.C. (2009). Soil seed banks and regeneration of Neotropical
639	dry deciduous and gallery forests in Nicaragua. Bois et Forêts des Tropiques, 299, 49–
640	62.
641	Uhl, C., Clark, K., Clark, H., & Murphy, P. (1981). Early plant succession after cutting and
642	burning in the upper Rio Negro region in the Amazon Basin. Journal of Ecology, 69,
643	631-649.
644	Valenta, K., Steffens, T. S., Rafaliarison, R.R., Chapman, C.A., & Lehman, S.M. (2015).
645	Seed banks in savanna, forest fragments, and continuous forest edges differ in a
646	tropical dry forest in Madagascar. Biotropica, 47, 435-440.
647	Vandvik, V., Vandvik, V., Klanderud, K., Meineri, E., Måren, I.E., & Töpper, J. (2015). Seed
648	banks are biodiversity reservoirs, species-area relationships above versus below
649	ground. Oikos, 125, 218–228.
650	Vázquez-Yanes, C., & Orozco Segovia, A. (1984). Ecophysiology of seed germination in the
651	tropical humid forests of the world: a review. Tasks for Vegetation Science, 12, 37-50.
652	Vázquez-Yanes, C., & Orozco-Segovia, A. (1993). Patterns of seed longevity and
653	germination in the tropical rainforest. Annual Review of Ecology and Systematics, 24,
654	69–87.

655	Venable, D. L., & Brown, J.S. (1988). The selective interactions of dispersal, dormancy, and
656	seed size as adaptations for reducing risk in variable environments. American
657	Naturalist, 131, 360–384.
658	Vieira, D.L.M., de Lima, V.V., Sevilha, A.C., & Scariot, A. (2008). Consequences of dry-
659	season seed dispersal on seedling establishment of dry forest trees: Should we store

- seeds until the rains? Forest Ecology and Management, 256, 471-481. 660
- Ward, J. H., Jr. (1963). Hierarchical grouping to optimize an objective function. Journal of 661
- the American Statistical Association, 58, 236–244. 662

 I group.

to Revense only

Supporting information to the paper

Jara-Guerrero et al. Dispersal syndrome influences the match between seed rain and soil seed bank of woody species in a Neotropical dry forest. *Journal of Vegetation Science.* 

Appendix S1. Woody species present in the 9-ha plot of standing vegetation, seed rain and/or soil seed bank. Number of individuals per

hectare (No. Ind./ha), relative abundance as percentage of the total of individuals (RA), mean ± standard error DBH (cm), total number of

seeds, seed density seed /m<sup>2</sup> (s/m<sup>2</sup>), dispersal season (Season), growth form (GF.: T, tree; S, shrub; C, climber) and dispersal syndrome (Zo,

zoochory; An, anemochory; Au, autochory). NP: not present.

		Standing vegetation			Seed rain		Seed bank				
Family	Species	No. Ind./ha	RA	DBH	No. seeds	s/m²	No. seeds	s/m <sup>2</sup>	GF	Dispersal syndrome	
Achatocarpaceae	Achatocarpus pubescens	38	0.573	3.11 <u>+</u> 1.89	639	3.8	NP	NP	S	Zo	
Amaranthaceae	Alternanthera sp.	94	1.417	1.48 <u>+</u> 0.46	2979	17.6	210	12.7	S	An	
	Iresine diffusa	1	0.015	2.48 <u>+</u> NA	3	0.02	NP	NP	S	An	
	Verbesina sp.	25	0.377	1.78 <u>+</u> 1.06	133	0.8	NP	NP	S	An	
	<i>Viguiera</i> sp.	53	0.799	1.15 <u>+</u> 0.19	NP	NP	NP	NP	S	An	
Asteraceae	Asteraceae unknown	4	0.060	1.67 <u>+</u> 0.84	10	0.1	NP	NP	S	An	
Bignoniaceae	Handroanthus billbergii	515	7.765	28.01 <u>+</u> 12.62	NP	NP	NP	NP	Т	An	
	Handroanthus chrysantha	340	5.127	16.37 <u>+</u> 11.03	NP	NP	NP	NP	Т	An	
Bixaceae	Cochlospermum vitifolium	220	3.317	23.72 <u>+</u> 12.11	345	2.0	2	0.1	Т	An	

Boraginaceae	Cordia lutea	13	0.196	$10.62 \pm 5.02$	NP	NP	NP	NP	Т	Zo
-	Cordia macrocephala	148	2.232	$1.69 \pm 0.81$	50	0.3	15	0.9	S	Zo
	Cordia rosei	137	2.066	 1.84 + 1.05	885	5.2	NP	NP	S	Zo
Burseraceae	Bursera graveolens	110	1.659	- 21.21 + 7.7	131	0.8	8	0.5	Т	Zo
Cactaceae	Armatocereus sp.	391	5.896	$-6.42 \pm 3.08$	49	0.3	NP	NP	Т	Zo
Capparaceae	Colicodendron scabridum	194	2.925		NP	NP	NP	NP	Т	Zo
	Cynophalla mollis	476	7.177	$12.13 \pm 5.28$	NP	NP	NP	NP	Т	Zo
Caricaceae	Vasconcellea parviflora	19	0.286	2.12 <u>+</u> 1.36	NP	NP	NP	NP	S	Zo
Convolvulaceae	Ipomoea carnea	255	3.845	1.59 <u>+</u> 0.55	19	0.1	63	3.8	S	An
Erythroxylaceae	Erythroxylum glaucum	232	3.498	11.43 <u>+</u> 5.93	2851	16.8	NP	NP	Т	Zo
Euphorbiaceae	Croton rivinifolius	721	10.872	$1.45 \pm 0.45$	2919	17.2	170	10.3	S	Au
	Croton micans	217	3.272	3.67 <u>+</u> 2.12	27	0.2	10	0.6	S	Au
Fabaceae	Aeschynomene scoparia	43	0.648	$1.4 \pm 0.62$	41	0.2	9	0.5	S	An
	Chloroleucon mangense	254	3.830	13.93 ± 5.9	259	1.5	8	0.5	Т	Au
	Erythrina velutina	40	0.603	30.3 <u>+</u> 16.14	19	0.1	3	0.2	Т	Au
	Geoffroea spinosa	226	3.408	13.66 <u>+</u> 6.78	NP	NP	NP	NP	Т	Zo
	Leucaena trichodes	311	4.689	7.2 <u>+</u> 2.32	56	0.3	52	3.1	Т	Au
	Mimosa acantholoba	3	0.045	1.86 <u>+</u> 1.15	1	0.0	NP	NP	S	Au
	Piptadenia flava	55	0.829	8.06 <u>+</u> 2.39	NP	NP	1	0.1	Т	Au

\_\_\_\_\_

	Pithecellobium excelsum	21	0.317	3.43 + 2.17	24	0.1	NP	NP	S	Zo	
	Caesalpinia glabrata	188	2.835	- 15.09 + 7.64	10	0.1	NP	NP	Т	Au	
	Senna hicapsularis	31	0.467	142 + 047	NP	NP	NP	NP	S	Au	
	Senna mollissima	14	0.211	8 68 + 1 82	116	0.7	2	0.1	л Т	Au	
Lamiaceae	Hyptis sn	47	0.709	1.00 - 1.02	852	5.0	2 NP	NP	S	An	
Malnighiaceae	Malnighia emarginata	158	2.382	$1.97 \pm 0.05$ $4.12 \pm 2.17$	140	0.8	NP	NP	S	70	
Malyaceae	Byttharia flaruosa	130	6.665	$\frac{1}{1}$ 1 58 + 0 66	150	0.0	3	0.2	S	20 Au	
Walvaccac	Coiha trichistandra	442	0.060	$1.58 \pm 0.00$	ND	ND	J	ND	т	An	
	Eriothoog muizii	4	0.136	$0.99 \pm 0.00$	INF 165	1 O	ND	ND	I T	An	
	Malvaceae unknown spl	30	0.452	$114 \pm 0.21$	255	1.0	ND	ND	S	Au	
	Malvaceae unknown sp1	265	3.996	$1.14 \pm 0.21$	233	0.0	ND	ND	S	Au	
	Urana sp	1	0.015	$20.03 \pm 9.03$	420	0.0	ND	ND	S	Au	
Moragana	Eigus sp.	6	0.090	1.03 - 10A	1200	7.1	ND	ND	ъ	Au Zo	
Nyotaginagaaa	Ficus sp.	17	0.256	$21.00 \pm 0.04$	ND	/.I	ND	ND	I T	Zo	
Dolygonooooo	Coccoloba miziana	17	0.633	$10.91 \pm 0.95$	202	1 C	ND	ND	т	Z0	
Primulaceae	Ronellia sprucei	42	0.543	$10.20 \pm 2.00$	202 ND	1.2	ND	ND	I T	Zo	
Phampacasa	Zizinhus thyrsiflorg	1	0.015	$7.51 \pm NA$	ND	ND	ND	ND	Т	Z0	
Rubiaceae	Randia aurantiaca	55	0.829	7.31 - 10A 2.14 + 1.47	118	1NF	NP	NP	r S	20 70	
Verbenacca	Lantana sp	120	1.960	$2.14 \pm 1.47$	0 0	0.7	1NF	1NF 2 /	S	20	
verbenaceae	Битипи эр.	150		1.20 - 0.41	0	0.0	51	J. <del>1</del>	5	20	_

Supporting information to the paper

Jara-Guerrero et al. Dispersal syndrome influences the match between seed rain and soil seed bank of woody species in a Neotropical dry forest. *Journal of Vegetation Science.* 

Appendix S2. Summary of the association analysis of species in the compositional groups of seed rain. P-value of association is shown for

each species. The best group corresponds to the group with the lower p-value after Sidak's correction for multiple testing with 33 species.

Dispersal	$\wedge$	<b>D</b> (	<i>P-value</i>						
syndrome	Species	Best group	Best						
~j		8- ° - P	group	Group 1	Group 2	Group 3			
Anemochory	Aeschynomene scoparia	1	0.006	0.002	1	1			
Anemochory	Alternanthera sp.	3	0.140	0.79	0.77	0.049			
Anemochory	Asteraceae unknown	1	0.006	0.002	1	1			
Anemochory	Cochlospermum vitifolium	1	0.115	0.04	1	1			
Anemochory	Eriotheca ruizii	1	0.167	0.059	0.659	1			
Anemochory	Hyptis sp.	1	0.006	0.002	0.844	1			
Anemochory	Ipomoea carnea	1	0.276	0.102	1	0.606			
Anemochory	Iresine sp.	1	0.790	0.406	1	1			
Anemochory	Verbesina sp.	1	0.542	0.229	1	0.256			
Autochory	Byttneria flexuosa	3	0.059	1	1	0.02			

Autochory	Caesalpinia glabrata	1	0.281	0.104	1	0.644
Autochory	Chloroleucon mangense	3	0.469	0.437	1	0.19
Autochory	Croton micans	2	0.096	1	0.033	1
Autochory	Croton rivinifolius	2	0.416	1	0.164	0.482
Autochory	Erythrina velutina	3	0.115	0.7	0.915	0.04
Autochory	Leucaena trichodes	3	0.377	0.478	1	0.146
Autochory	Malvaceae sp. 1	3	0.687	1	1	0.321
Autochory	Malvaceae sp. 4	2	0.573	1	0.247	0.556
Autochory	Mimosa acantholoba	1	0.801	0.416	1	1
Autochory	Senna mollissima	1	0.386	0.15	1	1
Autochory	Urena sp.	3	0.059	1	1	0.02
Zoochory	Achatocarpus pubescens	2	0.021	1	0.007	0.793
Zoochory	Armatocereus sp.	3	0.686	1	0.471	0.32
Zoochory	Bursera graveolens	3	0.501	0.969	0.308	0.207
Zoochory	Coccoloba ruiziana	3	0.059	1	1	0.02
Zoochory	Cordia macrocephala	3	0.259	1	1	0.095
Zoochory	Cordia rosei	2	0.554	1	0.236	0.463

Zoochory	Erythroxylum glaucum	2	0.660	1	0.302	0.326
Zoochory	Ficus sp.	3	0.686	1	1	0.32
Zoochory	Lantana sp.	1	0.758	0.377	0.479	0.589
Zoochory	Malpighia emarginata	2	0.559	1	0.239	0.578
Zoochory	Pithecellobium excelsum	3	0.261	1	1	0.096
Zoochory	Randia aurantiaca	1	0.188	0.067	1	0.569