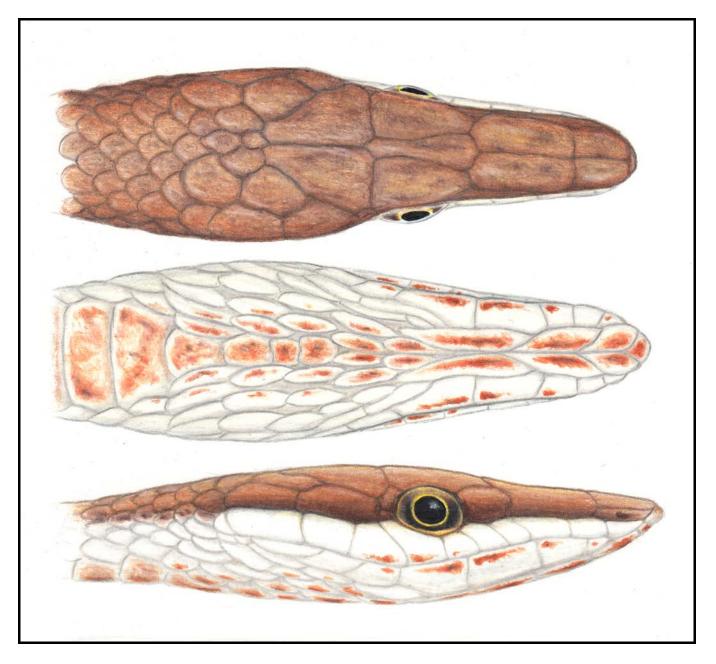
## BULLETIN of the Chicago Herpetological Society



Volume 55, Number 12 December 2020



## BULLETIN OF THE CHICAGO HERPETOLOGICAL SOCIETY Volume 55, Number 12 December 2020

Book Review: Indigo Snake: All You Need to Know about Indigo Snake Care, Housing and Feeding by Paul Jason Holly Carter	229
Notes on Reproduction of the Green Treefrog, Dryophytes cinereus (Anura: Hylidae), from Oklahoma Stephen R. Goldberg	229
Adventures with Dr. Rosen-Part 2	233
Index to Scientific Names of Amphibians and Reptiles for Volume 55 (2020)	239
Author – Title Index for Volume 55 (2020)	241
Herpetology 2020	243
New CHS Members This Month	244
Advertisements	244

Cover: A recently described Central American species: Köhler's vine snake, Oxybelis koehleri Jadin et al., 2020. Drawing by Justin T. Sipiorski.

#### STAFF

Editor: Michael A. Dloogatch—madadder0@aol.com Copy editor: Joan Moore

#### **2020 CHS Board of Directors**

President: John Gutierrez Vice-president: Jessica Wadleigh Treasurer: John Archer Recording Secretary: Gail Oomens Media Secretary: Annalisa Kolb Membership Secretary: Mike Dloogatch Sergeant-at-arms: Mike Scott Members-at-large: Rachel Bladow Jenny Hanson Tom Mikosz Immediate Past President: Rich Crowley

**The Chicago Herpetological Society** is a nonprofit organization incorporated under the laws of the state of Illinois. Its purposes are education, conservation and the advancement of herpetology. Meetings are announced in this publication, and are normally held at 7:30 P.M., the last Wednesday of each month. Membership in the CHS includes a subscription to the monthly *Bulletin*. Annual dues are: Individual Membership, \$25.00; Family Membership, \$28.00; Sustaining Membership, \$50.00; Contributing Membership, \$100.00; Institutional Membership, \$38.00. Remittance must be made in U.S. funds. Subscribers outside the U.S. must add \$12.00 for postage. Send membership dues or address changes to: Chicago Herpetological Society, Membership Secretary, 2430 N. Cannon Drive, Chicago, IL 60614.

**Manuscripts** published in the *Bulletin of the Chicago Herpetological Society* are not peer reviewed. Manuscripts and letters concerning editorial business should be e-mailed to the editor, <u>mdloogatch@chicagoherp.org</u>. Alternatively, they may be mailed to: Chicago Herpetological Society, Publications Secretary, 2430 N. Cannon Drive, Chicago, IL 60614. **Back issues** are limited but are available from the Publications Secretary for \$2.50 per issue postpaid.

Visit the CHS home page at <http://www.chicagoherp.org>.

**The** *Bulletin of the Chicago Herpetological Society* (ISSN 0009-3564) is published monthly by the Chicago Herpetological Society, 2430 N. Cannon Drive, Chicago IL 60614. Periodicals postage paid at Chicago IL. **Postmaster:** Send address changes to: Chicago Herpetological Society, Membership Secretary, 2430 N. Cannon Drive, Chicago IL 60614.

## Book Review: Indigo Snake: All You Need to Know about Indigo Snake Care, Housing and Feeding by Paul Jason 2020. Privately published. Softbound \$6.00. Kindle edition \$2.99.

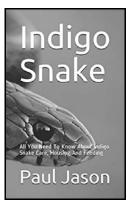
#### Reviewed By: Holly Carter drymarchonzz@hotmail.com

I often go in search of basic information books about reptiles and amphibians for our club, the Hoosier Herpetological Society, to resell. I recently became aware of a small book about care of indigo snakes in captivity that was listed on Amazon.com. I thought this might be a great addition to our collection, so I ordered one book to check it out.

After receiving this book, I was very unhappy with everything about it. It did have a great photo on the

cover, but I found out this had been copied from the internet. The book did not give credit for the cover photograph. Nor did it include any form of identification for the author.

The text covers 18 pages. An additional eight pages are blank, and the rest should have been for all the mistakes in it. The content is badly presented, makes no sense, and has no redeeming features. It looks to me like a bad translation from



some foreign language into English. For example, instead of "cold weather breeder" we see "iciness breeder." Where the word "female" should describe the snake, we see "woman" or "lady."

And then there is the matter of the word spacing in the book. The following two lines will give you the idea: After receiving this book, I was

very unhappy with everything about it.

This book was obviously produced by a company offering "print on demand," as a real publisher would certainly not have touched it. I am aware of information on the internet not always being truthful, but this book is beyond sad.

As if all this is not bad enough, a search within Amazon.com for the author's name reveals similar titles dealing with 34 different reptile species and five amphibians. To say nothing of a large number covering assorted bird and mammal species. I wrote to Amazon to let them know, as they could not have read or even opened these books before listing them, but no action has been taken and I am not optimistic.

Bulletin of the Chicago Herpetological Society 55(12):229-232, 2020

#### Notes on Reproduction of the Green Treefrog, Dryophytes cinereus (Anura: Hylidae), from Oklahoma

Stephen R. Goldberg Whittier College, Biology Department Whittier, CA 90608 sgoldberg@whittier.edu

#### Abstract

I conducted a histological examination of gonads from 43 *Dryophytes cinereus* adults from Oklahoma consisting of 27 males and 16 females. Males contained sperm from all months examined: March to July and September, October. The two smallest mature males (sperm in lumina of seminiferous tubules) measured 43 mm SVL and were from July and October. Females in spawning condition were found in February, April to July. The smallest mature female (spawning condition) measured 41 mm SVL and was from February. I found no evidence (gravid females containing postovulatory follicles from a recent spawning) to indicate *D. cinereus* spawns more than once during the same year in Oklahoma.

*Dryophytes cinereus* (Schneider, 1799) occurs in the southeastern United States from southern Texas to Florida and Delaware and western Tennessee to parts of Missouri, Arkansas, Illinois, Kentucky, Indiana, and has been introduced into Puerto Rico and Bahamas (Frost, 2020). In Oklahoma it occurs in the southeast corner (Bragg, 1943). *Dryophytes cinereus* breeding in Oklahoma occurs mainly from spring to early summer (Sievert and Sievert, 2011) in a variety of aquatic habitats (Redmer and Brandon, 2005). Eggs are deposited near the surface of water in small packs of jelly, often attached to floating vegetation (Tipton et al., 2012). In southern Illinois, sexual maturity is reached in 1 year (Garton and Brandon, 1975). Wright and Wright (1933) reported *D. cinereus* reproduction (no specific locality) occurred from April 15 to August 15. In the current paper I present data on the *D. cinereus* reproductive cycle from a histological examination of gonadal material from Oklahoma. The biology of *D. cinereus* is summarized in Redmer and Brandon (2003). Utilization of museum collections for obtaining reproductive data

Table 1. Two monthly stages in the spawning cycle of 16 adult female
D. cinereus from Oklahoma.

Month	n	Ready to spawn condition	Not in spawning condition
February	1	1	0
March	1	0	1
April	7	6	1
May	3	2	1
June	3	3	0
July	1	1	0

avoids removing additional animals from the wild.

A sample of 45 *D. cinereus* from Oklahoma collected 1956 to 1999 (Appendix) consisting of 27 adult males (mean SVL = 47.5 mm  $\pm$  3.1 SD, range = 43–55 mm), 16 adult females (mean SVL = 47.3 mm  $\pm$  4.3 SD, range = 41–55 mm) and two unsexed subadults (SVLs = 38 mm) was examined from the herpetology collection of the Sam Noble Museum of Natural History (OMNH), Norman, Oklahoma USA (Appendix). An unpaired *t*–test was used to test for differences between adult male and female SVLs (Instat, vers. 3.0b, Graphpad Software, San Diego, CA).

A small incision was made in the lower part of the abdomen of the 43 adults and the left testis was removed from males and a piece of the left ovary from females. Gonads were embedded in paraffin, sections were cut at 5  $\mu$ m and stained with Harris hematoxylin followed by eosin counterstain (Presnell and Schreibman, 1997). Histology slides were deposited at OMNH. No histology was done on the two subadults.

There was no significant difference between mean SVL of adult males versus adult females of D. cinereus (t = 0.15, df = 41, P = 0.88). The testicular morphology of D. cinereus is similar to that of other anurans as described in Ogielska and Bartmañska (2009a). Within the seminiferous tubules, spermatogenesis occurs in cysts which are closed until the late spermatid stage is reached; cysts then open and differentiating sperm reach the lumina of the seminiferous tubules (Ogielska and Bartmañska, 2009a). All 26 D. cinereus adult males were undergoing sperm formation (= spermiogenesis) in which clusters of sperm filled the seminiferous tubules. A ring of germinal cysts was located on the inner periphery of each seminiferous tubule. By month, numbers of D. cinereus males (N = 27) exhibiting spermiogenesis were: March (N = 2), April (N = 2), May (N = 7), June (N = 7), July (N = 7), September (N = 1), October (N = 1). The three smallest mature males in my study (spermiogenesis) measured 43 mm SVL and were from July (N = 2)(OMNH 39820, 41878) and October (N =1) (OMNH 47981). Wright and Wright (1933) reported adult D. cinereus males ranged from 37 to 59 mm in body size.

The ovaries of *D. cinereus* are typical of other anurans in consisting of paired organs located on the ventral sides of the kidneys; in adults they are filled with diplotene oocytes in various stages of development (Ogielska and Bartmañska, 2009b). Mature oocytes are filled with yolk droplets; the layer of surrounding follicular cells is thinly stretched. Two stages were present in the spawning cycle (Table 1): (1) "Ready to Spawn

Condition" in which mature oocytes predominated; (2) "Not in Spawning Condition" in which previtellogenic or atretic oocytes predominated. The smallest mature female *D. cinereus* (ready to spawn) measured 41 mm SVL (OMNH 41786) and was from February. Wright and Wright (1933) reported adult *D. cinereus* females ranged from 41 to 63 mm in body size. *Dryophytes cinereus* females of adult size, (OMNH 44423, SVL = 41 mm) from March and (OMNH 47183, SVL = 45 mm) from April (Table 1) were not in spawning condition and contained previtellogenic oocytes. It is conceivable these two females may have spawned later in the year. I cannot speculate as to when the two unsexed subadults (SVLs = 38 mm) (OMNH 41884, 47574) would have reached maturity.

Atretic follicles were noted in the ovaries of 3/13 (23%) of the D. cinereus females that were in spawning condition (Table 1). Atresia is a widespread process occurring in the ovaries of all vertebrates (Uribe Aranzábal, 2009). It is common in the amphibian ovary (Saidapur, 1978) and is the spontaneous digestion of a diplotene oocyte by its own hypertrophied and phagocytic granulosa cells which invade the follicle and eventually degenerate after accumulating dark pigment (Ogielska and Bartmañska, 2009b). See Saidapur and Nadkarni (1973) and Ogielska et al. (2010) for detailed descriptions of follicular atresia in the frog ovary. Abundant late atresia was noted in one non-spawning adult female from May (OMNH 38175, SVL = 46 mm). Oocytes had been replaced by vacuolated black-pigment-containing cells. It is likely atresia prevented this D. cinereus from spawning. Atresia plays an important role in fecundity by influencing numbers of ovulated oocytes (Uribe Aranzábal, 2011). Incidences of follicular atresia increase late in the reproductive period (Saidapur, 1978). Saved energy will be presumably utilized during a subsequent reproduction.

Times of breeding for *D. cinereus* throughout its range are shown in Table 2. My finding of one *D. cinereus* February female from Oklahoma (OMNH 41786) in spawning condition is somewhat early in the year although reproduction in Florida has also been reported to occur in February (Krysko et al., 2019). Because I lacked *D. cinereus* female samples from late summer and autumn, the duration of female reproduction in Oklahoma is not known, although my September and October males (OMNH 41883, 47981) were producing sperm, indicating breeding would have been possible.

There is a report that, in Georgia, *D. cinereus* (as *Hyla cinerea*) produces multiple egg clutches in the same reproductive season (Perrill and Daniel, 1983), however, I found no evidence that this occurs in Oklahoma. Multiple spawnings would have been suggested by the presence of mature follicles (upcoming spawning) and the concurrent presence of post-ovulatory follicles (recent spawning) (*sensu* Redshaw, 1972). Goldberg (2018) used the above criteria and reported multiple clutches were produced by the congener *D. wrightorum* in Arizona.

#### Acknowledgment

I thank Cameron D. Siler (OMNH) for permission to examine *D. cinereus* and Jessa L. Watters (OMNH) for facilitating the loan.

Locality	Breeding Period	Source
Alabama	April to August	Mount, 1975
Arkansas	Late April to August	Trauth et al., 2004
Florida	March to August	Carr, 1940
Florida	April to August	Gunzburger, 2006
Florida	February to September	Krysko et al., 2019
Georgia	April into July	Wright, 1932
Georgia	mid-April to mid-August	Jensen et al., 2008
Illinois	June and July	Garton and Brandon, 1975
Illinois	late May to August	Phillips et al., 1999
Kentucky	calls mid-May to August	Barbour, 1971
Louisiana	April to September	Dundee and Rossman, 1989
Louisiana	mid-March to late August	Boundy and Carr, 2017
Maryland	calls April to August	Cunningham and Nazdrowicz, 2018
Missouri	June to early July	Johnson, 2000
North Carolina	April to September	Dorcas et al., 2007
Tennessee	spring and summer	Niemiller and Reynolds, eds. 2021
Texas	March to October	Tipton et al., 2012
Carolinas to Virginia	May or June	Beane et al., 2010
Southeast	spring through summer	Dorcas and Gibbons, 2008

#### Literature Cited

Barbour, R. W. 1971. Amphibians and reptiles of Kentucky. Lexington: The University Press of Kentucky.

- Beane, J. C., A. L. Braswell, J. C. Mitchell, W. M. Palmer and J. R. Harrison III. 2010. Amphibians and reptiles of the Carolinas and Virginia. Second edition, revised and updated. Chapel Hill: The University of North Carolina Press.
- Boundy, J., and J. L. Carr. 2017. Amphibians and reptiles of Louisiana: An identification and reference guide. Baton Rouge: Louisiana State University Press.
- Bragg, A. N. 1943. Observations on the ecology and natural history of Anura XV. The hylids and microhylids in Oklahoma. Great Basin Naturalist 4(3-4):62-80.

Carr, A. F., Jr. 1940. A contribution to the herpetology of Florida. University of Florida Publication, Biological Science Series 3(1):1-118.

Cunningham, H. R., and N. H. Nazdrowicz, editors. 2018. The Maryland amphibian and reptile atlas. Baltimore: The Johns Hopkins University Press.

Dorcas, M., and W. Gibbons. 2008. Frogs and toads of the Southeast. Athens: The University of Georgia Press.

- Dorcas, M. E., S. J. Price, J. C. Beane and S. C. Owen. 2007. The frogs and toads of North Carolina, field guide and recorded calls. Raleigh: North Carolina Wildlife Resources Commission.
- Dundee, H. A., and D. A. Rossman. 1989. The amphibians and reptiles of Louisiana. Baton Rouge: Louisiana State University Press.
- Frost, D. R. 2020. Amphibian species of the world: an online reference. Version 6.0 (accessed 28 November 2020). Electronic Database accessible at <a href="https://amphibiansof.the.world.amnh.org/index.php">https://amphibiansof.the.world.amnh.org/index.php</a>. New York: American Museum of Natural History.
- Garton, J. S., and R. A. Brandon. 1975. Reproductive ecology of the green treefrog, *Hyla cinerea*, in southern Illinois (Anura: Hylidae). Herpetologica 31(2):150-161.
- Goldberg, S. R. 2018. Notes on reproduction of the Arizona treefrog, *Dryophytes wrightorum* (Anura: Hylidae) from Arizona. Sonoran Herpetologist 31(4):63-64.
- Gunzburger, M. S. 2006. Reproductive ecology of the green treefrog (*Hyla cinerea*) in northwestern Florida. American Midland Naturalist 155(2):321-328.
- Jensen, J. B., C. D. Camp, W. Gibbons and M. J. Elliott, editors. 2008. Amphibians and reptiles of Georgia. Athens: University of Georgia Press.

Johnson, T. R. 2000. The Amphibians and reptiles of Missouri. second edition. Jefferson City: Missouri Department of Conservation.

Krysko, K. L. K. M. Enge and P. E. Moler. 2019. Amphibians and reptiles of Florida. Gainesville: University of Florida Press.

Mount, R. H. 1975. The reptiles and amphibians of Alabama. Auburn, Alabama: Auburn University Agricultural Experiment Station.

Niemiller, M. L., and R. G. Reynolds, editors. 2011. The amphibians of Tennessee. Knoxville: The University of Tennessee Press.

- Ogielska, M., and J. Bartmañska. 2009a. Spermatogenesis and male reproductive system in Amphibia Anura. Pp. 34-99. *In*: M. Ogielska, editor, Reproduction of amphibians. Enfield, New Hampshire: Science Publishers.
- Ogielska, M., and J. Bartmañska. 2009b. Oogenesis and female reproductive system in Amphibia Anura. Pp. 153-272. *In*: M. Ogielska, editor, Reproduction of amphibians. Enfield, New Hampshire: Science Publishers.
- Ogielska, M., B. Rozenblut, R. Augustyñska and A. Kotusz. 2010. Degeneration of germ line cells in amphibian ovary. Acta Zoologica (Stockholm) 91(3):319-327.
- Perrill, S. A., and R. E. Daniel. 1983. Multiple egg clutches in Hyla regilla, H. cinerea and H. gratiosa. Copeia 1983(2):513-516.
- Phillips, C. A., R. A. Brandon and E. O. Moll. 1999. Field guide to amphibians and reptiles of Illinois. Champaign: Illinois Natural History Survey Manual 8.
- Presnell, J. K., and M. P. Schreibman. 1997. Humason's animal tissue techniques. Fifth edition. Baltimore: The Johns Hopkins University Press.
- Redmer, M., and R. A. Brandon. 2003. *Hyla cinerea* (Schneider) Green treefrog. Catalogue of American Amphibians and Reptiles 766.1-766.14.
- Redmer, M., and R. A. Brandon. 2005. Hyla cinerea (Schneider, 1799) Green Treefrog. Pp. 452-454. In: M. Lannoo, editor. Amphibian declines: The conservation status of United States species. Berkeley: University of California Press.
- Redshaw, M. R. 1972. The hormonal control of the amphibian ovary. American Zoologist 12(2):289-306.
- Saidapur, S. K. 1978. Follicular atresia in the ovaries of nonmammalian vertebrates. Pp. 225-244. In: G. H. Bourne, J. F. Danielli and K. W. Jeon, editors, International Review of Cytology, Vol. 54. New York: Academic Press.
- Saidapur, S. K., and V. B. Nadkarni. 1973. Follicular atresia in the ovary of the frog *Rana cyanophlyctis* (Schneider). Acta Anatomica 86(3-4):559-564.
- Sievert G., and L. Sievert. 2011. A field guide to Oklahoma's amphibians and reptiles. Oklahoma City: Oklahoma Department of Wildlife Conservation.
- Tipton, B. L., T. L. Hibbitts, T. D. Hibbitts, T. J.Hibbitts and T. J. Laduc. 2012. Texas amphibians: A field guide. Austin: University of Texas Press.
- Trauth, S. E., H. W. Robison and M. W. Plummer. 2004. The amphibians and reptiles of Arkansas. Fayetteville: The University of Arkansas Press.
- Uribe Aranzábal, M. C. 2009. Oogenesis and female reproductive system in Amphibia Urodela. Pp. 273-304. In: M. Ogielska, editor, Reproduction of amphibians. Enfield, New Hampshire: Science Publishers.

- Wright, A. H. 1932. Life-histories of the frogs of Okefinokee Swamp, Georgia. North American Salientia (Anura) No. 2. New York: The Macmillan Company.
- Wright, A. H., and A. A. Wright. 1933. Handbook of frogs and toads of the United States and Canada. Ithaca, New York: Comstock Publishing Associates.

#### Appendix

Forty-five *D. cinereus* Oklahoma examined by county from from the herpetology collection of the Sam Noble Museum., University of Oklahoma (OMNH), Norman, Oklahoma.

Johnston: OMNH 43582; Latimer: OMNH 41875, 41878; Le Flore: OMNH: 40007, 40008, 41883–41885, 42950–42952, 42954, 47193; Marshall: OMNH 46490, 47574, 48025; McCurtain: OMNH 38175, 38176, 39818–39821, 44410, 44421, 44423, 47182, 47183, 47185, 47186, 47188, 48117; Muskogee: OMNH 39581, 47981; Pushmataha: OMNH 41895–41897, 44429–44431; Rogers: OMNH: 41786, 41904–41906; Wagoner: OMNH 47853, 47854.

<sup>------. 2011.</sup> Hormones and the female reproductive system of amphibians. Pp. 55-81. *In*: D. O. Norris and K. H. Lopez, editors, Hormones and reproduction of vertebrates, Volume 2. Amphibians. Amsterdam: Elsevier.

#### Adventures with Dr. Rosen – Part 2

#### Roger A. Repp 9044 N. Valgrind Lane Tucson, AZ 85743 repproger22@gmail.com

As none of you may remember, last month's column centered on my friend Dr. Philip C. Rosen, who passed away on 18 September 2020. While his death impacted many of us greatly, the prevailing attitude here in Tucson has been one of celebrating his presence while he was among us, while feeling genuine gratitude for the privilege of having had him in our midst. Last month's column discussed two field trips that involved Phil and me leading a film crew to some rattlesnake dens, and ended with suggestions that Phil was a rare combination of herpetologist and conservation biologist. Should you not remember any of this, no worries! But I remind you that if you are to remember any of it, you might have to read it first. In any case, knowing that at least two of you will read everything that I write, we will proceed with the message of fun with Phil while he was among us, while touching on some of his great accomplishments in conservation.

Another aspect of Phil's final days that was discussed last month was his seeking data entry help in order to get some of his notes from ink and paper into a computerized format. He also needed some help with his life's work-a herpetological history of Avra Valley. I know that to be specifically true, as he asked me to help him with that. My reasons for turning him down were many-fold, the best being this damnable pandemic. The nature of the work would have required some face time. I "catch" things easily, and as a result, might have given Phil more than he wanted from me. Had the opportunity to work closely with Phil during his last days arisen at any other time, I would have jumped on it. But in saying that, I am also acutely aware that I am in the same position as Phil. I have over 30 years of my own ink and paper notes that need to be organized if they are ever to count for anything. In this day and age where any one of us can be dead in two weeks, that is a somewhat sobering notion. Moving on:

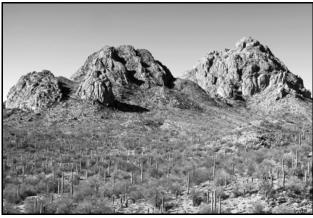


Figure 1. Ragged Top, the crown jewel of the Ironwood Forest National Monument. Image by the Author.

eastern boundary is the west side of the Tucson Mountains. Its western boundary is a series of loosely connected, low elevation mountain ranges. Of these ranges, Ragged Top is by far the most impressive (Figure 1). As the population of Tucson has grown, it has surged into Avra Valley. Agriculture, urban sprawl, and drought have created major changes in the landscape, as well as in the herpetofauna that inhabit it. One of the more interesting (if not depressing) developments in Avra Valley was the rapid extirpation of one species of fossorial snake that was quickly replaced by another. The Shovel-nosed Snake (*Chionactis annulata*) was once common here, but the last specimen documented was in 1979. The Variable Sandsnakes (*Chilomeniscus stramineus*) began to move in as the Shovel-nosed Snakes declined (Figure 2).

From 1988 to current, the author has been herping Avra Valley extensively. I can personally attest to the complete absence of the Shovel-nosed Snake, and the abundance of the Variable Sandsnake. Nobody would know the how and why of



Avra Valley is the next major valley west of Tucson. Its

Figure 2. (Left) Western Shovel-nosed Snake (*Chionactis annulata*). (Right) Variable Sandsnake (*Chilomeniscus stramineus*). The latter is an actual Avra Valley specimen. As briefly and inadequately explained in the text, the Shovel-nosed Snake was once common in Avra Valley, but according to Rosen, was replaced by the Variable Sandsnake. See text and Rosen(2003, 2008). Photo credits: (Left) Jim Rorabaugh; (Right) Fred Wilson.

this more than Phil, yet this aspect of his research here is but one grain of sand on his termite mound of historical knowledge of the place. I am still trying to gain access to a number of his papers, but for now, see Rosen (2003, 2008).

I don't know how much of Phil's life and knowledge was neatly wrapped up before he passed away. (I'm working on getting answers. It is like drilling through hardened tool steel!) What I do know is the thoroughness of his attention to detail when in the field. I am not alone in that knowledge. His reputation for the comprehensive documentation of any field outing was legendary among his peers. In last month's column, I described his turn-by-turn documenting of our drive to a rattlesnake den. At the time, I thought he might be doing this so he could later revisit "my" den on his own. Or perhaps "show off" to a fellow documenter? I did not realize at the time that Phil always did this sort of thing. His explosive revelations of longterm data, as well as his meticulous reports to various agencies, earned him many grants. Some of these grants were of the sixfigure kind. Yes, Dr. Rosen's reputation for careful, big picture documentation of field work everywhere he went led to the funding of many projects.

There were very few times in my "career" as an avocational herpetologist that I was paid to do herp-related field work. It's possible that I could have earned more money than I did with my herping hobby, but this thing called "a real job" (a damn good one at that) got in the way. Even with outstanding vacation benefits, my ability to do the real field work-every day for weeks and months at a pop-was limited. But there was the occasional academic windfall in herpetology for me, and the most money that I ever received was through Dr. Rosen. He hired me to provide historical data on the newly created Ironwood Forest National Monument (IFNM). For once, my ink and paper notes were actually going to pay off. (But the message "don't quit your day job, son" was always fiscally clear.) The creation of IFNM in the year 2000 was a parting gift to all from the Clinton administration. Large chunks of this monument line the western edge of Avra Valley. Hence, when a substantial grant was made available to survey IFNM, Phil was the obvious choice to land it. His pre-existing knowledge of the herpetological history of Avra Valley, coupled with his outstanding reputation, made him the best person for the job. He began the IFNM survey work in 2002, and hired me in the fall of that year. At that point in time, I had amassed 14 years of data on the herps of both Avra Valley and IFNM, twelve of which were before the monument was created. I had also published several articles in various gray-literature publications about Ragged Top. This rugged mountain is one of the crown jewels of IFNM. Phil hired me to produce a spreadsheet that listed all my encounters with Desert Iguanas (Dipsosaurus dorsalis), Chuckwallas (Sauromalus ater), and Sonoran Desert Tortoises (Gopherus morafkai) within Avra Valley and IFNM. These were the three species that he knew to be important indicator species of the region. He also always had a personal interest in all of the smaller fossorial snakes in any region. He wanted me to list any Groundsnakes (Sonora semiannulata), Shovel-nosed Snakes (Chionactis annulata), Blackheaded Snakes (Tantilla hobartsmithi), and Variable Sandsnakes (Chilomeniscus stramineus) found there. I had not found any of the first three species in Avra Valley, making my job easy

in this regard. He let me off the hook with the Variable Sandsnakes, of which I had found exactly 50 through the years. He already had scads of historical data on them, from many of the exact locations and time period as my data included. How we did not accidentally bump into each other while flipping nearly every board in Avra Valley will forever remain a mystery to me. In addition to the spreadsheet I was creating, Phil also wanted me to mark locations on topo maps that he provided. (I did not purchase a GPS unit until the year 2001. In this day and age of cell phones, pin drops and Google Earth, it is nearly impossible to even remember the days of paper maps and compasses. For the bulk of my early experiences herping IFNM, GPS units were of such size that they had to be carried in suitcases! And the cost of such units was prohibitive.) When I finished the spreadsheet and topo map-marking for Phil, there was still some money left in the pot for me. Enough money remained in my personal budget to use five billable days in the field. One of these "herpfor-a-paycheck days" was later spent in the company of Dennis Caldwell. That name, and the persona behind it, will be highlighted in a future column on the subject of Dr. Rosen. My day with Caldwell was but one of the five remaining funded days. Dr. Rosen wanted to be with me for the other four. Hot-DIGGITY-damn! I was going to get paid to herp with Phil!

Earlier in this article, I explained that the western boundary of Avra Valley is "a series of loosely connected, low-lying mountain ranges." I did not hang names on these ranges, as they are rather unremarkable and have no bearing on this article. But two of these ranges, Ragged Top and the Sawtooth Mountains, have everything to do with my words of today. We start by commenting that it is well that folk other than academic herpetologists named these ranges, or they would be known as "Ragged-topped" and "Saw-toothed" Mountains. Putting smartassed insinuations aside, both places are distinctive landmarks when viewed at a distance, and downright spectacular to behold when one is herping among them. The author must confess that it is taking every ounce of restraint that he possesses to not launch into thousands of words describing his experiences and subsequent devotion to Ragged Top. Suffice it to say that Ragged Top is the place that transformed me from a purposeless field herper into a serious student of the natural history of the herpetofauna in our region. It was here that I first witnessed the phenomenon of individual reptiles demonstrating fidelity to their shelters. The day that Dennis Caldwell (there's that name again) and I found our very first Chuckwalla was 9 March 1991. This was after we spent roughly a year trying to find a local population of them to visit. That day was also the same day that my first-ever repeating reptile was found. But I had to wait until December for that to be made known (Repp, 1996, 1998a,b). My first "repeater" was a tortoise. One day later, 10 March 1991, my very first repeating Chuckwalla was found. It was a young adult male Chuck (as they will be called at will throughout this column) viewed inside a classic boulder crevice (Figure 3). Probably the very last time that I will ever visit that crevice was 22 January 2018, nearly 27 years later. There was a huge adult Chuck within the crack on that day. While I can't say for certain that it was the same Chuck that was first viewed in 1991, neither can I say that it wasn't. As part of my efforts to address the possibility that it even could be the same Chuck, I emailed Craig



**Figure 3**. The author's longest-running repeating Chuckwalla (*Sauro-malus ater*) crevice. A "Chuck" has been observed inside with every visit since March of 1991. The author speculates that the same Chuck may have been seen over a 27-year time span here. Image by the author.

Ivanyi, codirector of the Arizona Sonora Desert Museum, with the question of how long captive specimens have been known to live. Craig put the right people on the mission. In jig time, records and quarantine technician Mary Powell-McConnell responded with the answer that our local species, Sauromalus ater, has survived in captivity for 29 years. It is certainly worth the words to add that an insular form of Chuck, the San Esteban Chuckwalla (Sauromalus varius), has been documented to live 42 years! Armed with this information, it is safe to say that it is possible that the Chuck under discussion could be the same one viewed over a 27-year time period. The young adult/huge adult situation with that particular Chuck lends more evidencehowever flimsy or speculative-to that possibility. Since this may be the only column where I ever write about Chuckwallas, I am including Figure 4. This is an example of a "huge," and perhaps long-lived adult male Chuckwalla from Ragged Top. I would like nothing more than to be able to go back to visit this particular crevice, with its now-huge repeating Chuckwalla within. But it is a 3-mile round trip over rugged terrain to get there and back. And I am now suddenly very old. This particular Chuck crevice needs a set of younger eyes and legs to monitor, but I have had zero luck with my efforts to recruit a younger version of me to do any of this sort of thing.

Getting back to *all* the Ragged Top crevice Chucks, the only thing that I can prove about these or any other wild herps viewed in shelters is the propensity of that particular shelter to consistently produce an occupant. I have two more Chuck cracks that are also in the running for being repeating champs. We go to my notes to see what I had to say about finding one of them:

1993 Fri Jan 15 Location: Rag Top E. and proper... Herps:... Also 5 Chuckwallas, 2 on R.T. East. These interest me. 1 (one) is in white rocks, below first outcrop...

#### Rosen and the White Rocks Chuckwalla

Upon receiving my spreadsheet, and studying my comments, Dr. Rosen wanted to see some of my long-term Chuck crevices, as well as anything else that might come our way. When I discovered that he was actually taking GPS readings off my topo



**Figure 4**. A whopper! They *do* get bigger, but the Chuckwalla in this image is the only large adult from Ragged Top that this author has ever captured and handled. Captive specimens have been documented to live as long as 29 years. Image by Daniel M. Bell.

map marking procedures, I made him aware that said marks might be wildly inaccurate. His solution to this problem was that we hike to these places, and nail them with accurate coordinates. Thus it came to pass that on 16 January of 2003, we headed for the eastern buttes of Ragged Top. During the ride out, Phil had many questions about my search methods. It was not until a few months later that I learned that he had never hunted for Chuckwallas by using a mirror or flashlight to shine rock crevices. He had made several trips to various places where Chucks could be seen basking on the top of boulders, such as Organ Pipe National Park (ORPI), but never had he tried crack-hunting for them. Since we have just mentioned it, his work at ORPI eventually became his dissertation. In addition to the mud turtles of Quitobaquito Springs, he also road cruised and worked the flats extensively for lizards and snakes. His list of papers regarding the herps of ORPI is staggering in its number and variety. In what was to become his trademark with later efforts, he used his extensive field notes and highly-honed analytical skills to document cyclical fluctuations of herps in the park. I was blessed to see his 21 November 2000 program for the Tucson Herpetological Society (see Appendix), where he did everything with his numbers that I hope to some day do with mine. This presentation was packed with information from his dissertation (Rosen, 2000).

Getting back to our January 2003 visit to Ragged Top, I appreciated his questions, as they gave me an opportunity to explain exactly how we would be working the slopes east of Ragged Top. Out of hundreds of visits to the place, and hundreds of observations of them, *never* have I seen a Chuck outside of a crevice there. This was despite the fact that my search methods always involve looking for herps on top of boulders as well as inside and beneath them. They obviously must bask and move about, but for whatever reason, they seem to be much more wary than Chucks from other locations. It is my impression that they see us coming from a long way off, and nosedive for cover.

We arrived at my favored parking spot at 0957 hrs. The ambient air temp was  $15^{\circ}$ C ( $59^{\circ}$ F); the cloud cover was 50% thin, wispy clouds; the humidity 20%; the wind speed was calm. It was shaping up to be the kind of day that I often designate as "Arizona perfect." By 1000 hrs, we began our first crunchy footsteps up the gentle incline of what I call the Front Ridge. We stayed in motion all day long, and did not return to the truck until 1752 hours, nearly eight hours later. For the entire effort, Phil performed his duties exactly as instructed. He traveled parallel with me, following the same contours, yet always maintaining a comfortable five- to ten-meter distance. This assured that we were always shining different locations, while being able to easily converge when a find was made, or when we arrived at one of my mapped locations for tortoises or Chuckwallas.

In all, I would guess that we traveled perhaps five miles during our eight-hour hike. We moved in somewhat linear fashion along a series of hills that have no name other than the names that I have given them. We generally moved from east-towest, first assailing the "Front Ridge," to "Holy Ground," to "Turtle Hill." We then crossed the undulating bajada to "Mount Badass," this being my moniker for Ragged Top in the days when I wrote about the place while trying to conceal its actual identity (see Repp, 1996, 1998a). From there, we hoofed back across the bajada, and battled the lengthy and featureless uphill stretch behind the Front Ridge to visit "Repp Hill," ending the journey with the steep, crumbly and slippery slopes of "Burnett Hill." From there, we slipped and slid our way back down to Repp Wash, where we had parked. There would have been a time when this sort of effort would have easily landed us five tortoises, five Chucks, three Western Lyresnakes (Trimorphodon biscutatus), three Gila Monsters (Heloderma suspectum), and five Western Diamond-backed Rattlesnakes (Crotalus atrox). Back in the day-before a relentless and heartless drought wiped out nearly every herp in the region – we used to jokingly say to each other: "Like my old grandpappy always said, 'You can't go wrong at Ragged Top, son!"" But that was then, between the years 1991 and 1996. This was now, where every visit after 1996 yielded only tortoise corpses and the occasional Chuck. On this day with Phil, instead of documenting living herps, we were taking GPS locations for the ghosts of the longdead ones. Perhaps the day will come when I am depressed enough to describe the things that passed before my eyes during that horrible year of 1996 in one of these columns.

Getting back to my hike with Dr. Rosen, while we were initially ascending Front Ridge, I found a western pipistrelle bat (*Pipistrellus hesperus*) in known lyresnake crevice number 2 (a crevice found to contain a lyresnake in 1992). As this species of bat is on the menu for lyresnakes, I have always noted when one is found in a known lyresnake crack. I have even noticed that if I pay a visit to a lyresnake crack soon *after* a bat is observed inside, a lyresnake will often appear. Very close to this crack, we paused to note the handful of fragments left over from the death of my most beloved tortoise of all, number 23. I had first found this tortoise — a four-inch-long juvenile — in January of 1994. In February of the same year, I was working with a grad student who processed and marked it as number 23 for me. I actually had several incidental encounters with #23 between then and 13 December 1996, when I found it fresh dead in its burrow. It had died so recently that the smell of death was still upon it, and maggots were seen moving in and out of it. When Phil and I visited it this day, just over seven years later, tortoise #23 was reduced to but a few bits and pieces of what had once been thicker carapace fragments.

Our one and only living find of the day was at the White Rocks. But the find would not have happened without the presence and diligence of Phil. Within an hour of me revealing my crack-hunting style for finding Chucks, the pupil became the master. Prior to this outing, the White Rocks Chuckwalla had never let me down. With every visit, anywhere from one to three Chucks had been observed here (Figure 5). But on this day, my heart sank when I looked into the known crack that had been so consistent for almost exactly ten years and found it to be empty! Had it been up to me, that would have been the end of the White Rocks part of the day. We would have just continued on for another seven hours and found nothing but a bat in a crack for an all-day effort. But Phil continued to search every crevice in the framework of that rock structure, working his way to the very top. And then it was "Oh my, look!" And I was "Yeah? Yeah?" And he was "Oh yeah!" Sure enough, he had found a huge adult Chuckwalla (see Figure 4 again for size reference) in a previously undiscovered crevice in the White Rocks formation. This find was basically a game-saving tackle on Phil's part. This particular Chuck was the only living herp of note to show up on this day. The find also kept the streak of the White Rocks alive. While I hope to get back to the White Rocks at least once this winter, until that happens, my most recent visit was 26 November 2019. With that visit, there was one Chuck visible in the usual crevice there, but it was not one of the two seen in Figure 5. This one was almost the exact size of the one seen in the righthand image of Figure 5, but it had a complete tail.



Figure 5. (Left) A large adult male Chuckwalla and (Right) a smaller female viewed and photographed in the same "White Rocks" crevice 15 years apart. Photo credits: (Left) Daniel M. Bell, 12 March 1994, (Right) Hans-Werner Herrmann, 24 January 2009.

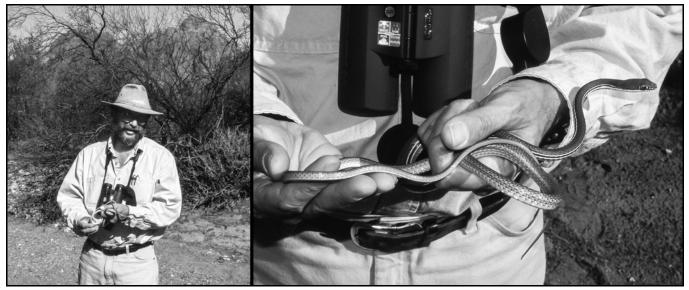


Figure 6. (Left) Dr. Philip C. Rosen with a Sonoran Whipsnake (*Masticophis bilineatus*) found and captured near Ragged Top. (Right) Close-up of the snake in the image to the left. Images by the author, 1 June 2003.

#### He likes it!

On 1 June 2003, Phil and I had one last survey day left for Ironwood Forest National Monument. There was as yet one portion of the monument to be surveyed, and that was the West Silver Bell Mountains. For whatever reason, despite the fact that we had technically already surveyed Ragged Top, he wanted to go back there again before assailing the West Silver Bell Mountains. The world can be grateful for many reasons that Phil and I did not always think alike. Had our roles been reversed, there is no way in hell that I would ever be inspired to go back to a place where two accomplished field herpers found only three dead tortoises and a Chuckwalla for eight solid hours of steep and difficult herping effort. Perhaps he merely wanted one last look at the rugged spectacle of Ragged Top itself before calling his survey finished (see Figure 1). In any case, he was the boss. Where I come from, bosses are not questioned, but obeyed. He was driving, and he made the slight detour to visit Ragged Top while we were, in theory, on our way to the West Silver Bells. And I am so very glad that he did.

One can visit a place hundreds of times, and think that one has seen it all. But that one surprise find – be it something extraordinary or not-can really be exciting. In this case, we speak of the Sonoran Whipsnake (Masticophis bilineatus) (MABI) that Phil and I cornered and captured at 0742 hours on this impulse visit that Dr. Rosen made possible. I can't remember all of the details of the find. I don't remember who saw it first. I don't remember which of us was the one to eventually grab it. What I do remember is one of us initially seeing it, and losing sight of it. I also remember the focused camaraderie between the two of us as we both surgically scrutinized the vicinity of that first visual. And then we had it on the run, losing it again when it slipped under a dense hackberry thicket. For ten minutes, we circled that hackberry, shining every opening in the helter-skelter snarl of woody stalks that comprise this *miserable* shrub. We eventually spotted a small bit of flank in the deepest recesses of the heart of the plant, and one of us gave it poke that sent it sprawling into the hands of the other. And when a herp of

any kind winds up in the hands of Dr. Rosen, one can bet that a thorough processing will follow. We GPSed the location of the find. We documented the ambient air temp as being 30.5°C (89.3 °F), with 0% clouds, 20% humidity, and just a puff of a breeze. Working together, we learned that our MABI was 798 mm (31.4 inches) long, with a tail length of 358 mm (14 inches). We also learned that this snake was a male by "cracking it." (Bending the tail at the cloaca and gently squeezing the hemipenes out.) And finally, we photographed it (Figure 6). While MABI can be common at higher elevations among oak trees and along riparian corridors, finding one in the harsh, hot and arid land of Chuckwallas and Desert Iguanas is quite another matter. Finding, capturing and processing this snake with Phil was a high point in my field outings with him. Had I encountered it without him, that same MABI would likely go something like: "MABI, ~1 m long, RTE, 0742 hours" in my notes. And there would be no capture or subsequent photo. Dr. Rosen had a way of nailing something nine ways to Sunday when in the field, and I'm glad that he led the charge to what will surely be the first and last Sonoran Whipsnake that I ever see at Ragged Top.

#### Epilogue (23 November 2020)

Just yesterday, when the words that fall above were deemed nearly complete, I came upon a resource previously unknown to me that led me directly to the heart of some of the publications of Dr. Philip Clark Rosen. This happened just in the nick of time to keep me from repeating the same mistake made in my first column about him. That mistake was suggesting that Dr. Rosen was primarily an aquatic herpetologist. That was an inane and inaccurate statement.

My big find was an academic website called ResearchGate that lists 87 publications written by Phil, dating from 1981 to early 2020. Sadly, this is not a complete listing, as I am finding more that were *not* included in ResearchGate on an almost daily basis as I delve into Phil's life. I have emailed some requests to those who knew Phil best to see if he left us a current *CV* before passing away. At this point in the day, it is not looking good, but time will tell. [But wait! There's more!] And time *did* tell! It spoke volumes. It said "Here Roger! Here is Phil's *CV*." Yes, by the end of the day today, my old friend Dennis Caldwell came through by emailing me a copy of a 2013 version of Dr. Rosen's *CV*. I have yet to thoroughly digest it, but for now, I know enough to be a *lot* more careful not to paint Dr. Rosen *or* his studies with too broad of a brush. Yes, there were many times that he kept his hiking boots dry during his illustrious herping career.

#### Acknowledgments

The author is deeply indebted to Dennis Caldwell and Jim Rorabaugh, who have not only contributed information for this column, but are *about* to contribute even more. The poor bastards have no idea how much they will be *hammered* in the days ahead. I also wish to extend my gratitude to Craig Ivanyi, Howard Byrne, Stephane Poulin and Mary Powell-McConnell of the Arizona Sonora Desert Museum for their assistance with the life expectancy of captive Chuckwallas. Special thanks are also always in order to Mike Dloogatch and Joan Moore for their editorial support and help, as well as photo editor Steve Barten who always provides skillful assistance (but with maximum complaints about my horrible photography).

This here is Roger Repp, signing off from Southern Arizona, where the turtles are strong, the snakes are handsome, and the lizards are above average.

#### Literature Cited

- Repp, R. A. 1996. Ides of March '92. Sonoran Herpetologist 9(2):9-12.
  - . 1998a. Ides of March revisited. Sonoran Herpetologist 11(8):86-89.

-----. 1998b. Wintertime observations on five species of reptiles in the Tucson Area: Sheltersite selections / Fidelity to sheltersites / Notes on behavior. Bulletin of the Chicago Herpetological Society 33(3):49-56.

Rosen, P. C. 2000. A monitoring study of vertebrate community ecology in the northern Sonoran Desert, Arizona. Ph.D. dissertation. University of Arizona, Tucson. 307 pp.

------. 2003. Distribution and ecology of amphibians and reptiles at Ironwood Forest National Monument. I. Desert Iguana, Chuckwalla, and Desert Tortoise. Final report section, Biological Survey of Ironwood Forest National Monument, Arizona-Sonora Desert Museum to Bureau of Land Management. 52 pp.

------. 2008. 2007 survey results for the Tucson Shovel-nosed Snake (*Chionactis occipitalis klauberi*), with evidence for ecological change in south-central Arizona. Final report to town of Marana and Arizona Game and Fish Department.

#### Appendix:

#### Additional Selections from the Work of Philip C. Rosen about Avra Valley and the Ironwood Forest National Monument

- Rosen, P. C. 2000. Lizard and snake population trends in context of the vertebrate ecosystem at Organ Pipe Cactus National Monument. Presentation to the Tucson Herpetological Society, November 21, 2000.
  - ------. 2003. Avra Valley snakes: Marana survey report for Tucson Shovel-nosed Snake (*Chionactis occipitalis klauberi*). Prepared for the Town of Marana, University of Arizona.
- ———. 2004. Avra Valley snakes: Marana survey report for the Ground Snake (Sonora semiannulata). Final Report to Town of Marana on Ground Snake. 17 pp.

-----. 2008. Ecological restoration in Tucson-owned Avra Valley lands in context of conservation of the Western Groundsnake and Tucson Shovel-nosed Snake. Unpublished report to City of Tucson, Office of Conservation and Sustainable Development.

April 73-92

## Index to Scientific Names of Amphibians and Reptiles for Volume 55 (2020)

January 1-24 February 25-48 March 49-72 Abronia taeniata 37 Acris crepitans 213 blanchardi 213 Acrochordus granulatus 52 Agalychnis dacnicolor 103 Agkistrodon contortrix 187, 205 Alligator mississippiensis 205 Ambystoma flavipiperatum 69 jeffersonianum 124 laterale 113 maculatum 5, 18, 124 ordinarium 69 tigrinum 113, 144 velasci 37 Amerotyphlops 77 Anaxyrus 213 americanus 3, 18, 124 charlesmithi 213 fowleri 3, 18, 213 microscaphus 213 punctatus 153 terrestris 213 Antillotyphlops 77 Apalone spinifera 18, 113 spinifera 7 Aquiloeurycea cephalica 37 galeanae 33, 34, 35, 36 Argyrophis 77 Asiatyphlops 77 Aspidoscelis 87 sonorae 154 tigris 149 Atelopus elegans 160 varius 157-162 zeteki 160 Atractaspis boulengeri 54, 58 Barisia imbricata 33, 35, 36, 37 Bitis gabonica 93, 97, 98 Boa constrictor 70 Bogertophis subocularis 210 Boiga irregularis 69 Bolitoglossa platydactyla 37 Bothrops atrox 91 Bufo alvarius 127 bufo 205 Bungarus caeruleus 205 Caiman crocodilus 10, 187 Calabaria reinhardtii 93, 94, 97 Calamaria gervaisi 26

May 93-112 June 113-132 gervaisii 25 Candoia paulsoni 52 Causus maculatus 93, 97, 98 Chacophrys pierottii 102 Chamaeleo chamaeleon 186 Chelydra serpentina 7, 18, 69, 73, 113, 125, 151 Chilomeniscus stramineus 233, 234 Chionactis annulata 233, 234 Chiropterotriton priscus 33, 34, 35, 36, 37 terrestris 37 Chrysemys picta 73, 74, 151 marginata 7, 18, 113 Clemmys guttata 7, 18 Coluber constrictor 125 hydrus 133 Conopsis lineata 37 Cophosaurus texanus 85 Craugaster augusti 35, 36 Crocodylus adelynhoserae 68 halli 68 jackyhoserae 68 Crotalus aquilus 37 atrox 35, 36, 37, 40-45, 83, 129, 166, 170, 178, 180, 182, 184 225.236 cerastes 83, 178, 180 cerberus 88, 89 horridus 70 lepidus 35, 36, 37, 83, 209 lutosus 129 molossus 34, 35, 36, 37, 42, 83, 85, 166, 167, 170, 178, 180, 182, 184 morulus 35, 36, 37 oreganus oreganus 151 pricei 35, 36, 37 ruber 225 scutulatus 35, 36, 37, 83, 178, 180 tigris 83, 147, 166-171, 178-185 triseriatus 171 willardi 41, 83 Crocodylus niloticus 216 Crotaphopeltis hotamboeia 54, 56, 58 Cryptobranchus alleganiensis 187 Cubatyphlops 77 Cyclocorus nuchalis nuchalis 25-26 Dasypeltis fasciata 93, 94, 97 Dendroaspis jamesoni 54, 56, 57, 58

July 133-152 August 153-172 September 173-188 jamesoni 93, 95, 97 polylepis 173 Diadophis punctatus 37, 125 Dipsadoboa underwoodi 54, 56, 58 viridis 54, 56, 58 Dipsosaurus dorsalis 234 Drymarchon couperi 71 Dryophis vittatus 243 Dryophytes cinereus 229-232 eximius 37 plicatus 37 Duellmanohyla uranochroa 160 Elaphe climacophora 174 Eleutherodactylus cystignathoides 34, 35, 36, 37 saxatilis 213 Elgaria 194 Emydoidea blandingii 9, 18, 73, 74, 113, 171 Emys orbicularis 10, 138 Enhydris caspia 133 Eretmochelys imbricata 69 Eryx 186 jaculus 186 miliaris 186 nogaiorum 186 tataricus 186 Eumeces brevirostris 37 japonicus 174 lynxe 37 Eurycea bislineata 124 cirrigera 214 multiplicata griseogaster 213 multiplicata 213 tynerensis 213, 214 Ficimia hardyi 37 Gastrophryne olivacea 213 Gekko japonicus 174 Geochelone platynota 71 Geophis latifrontalis 37 mutitorques 37 Gerrhonotus farri 192, 219 infernalis 33, 35, 36, 37, 192, 194, 220 lazcanoi 192, 219 liocephalus 192 lugoi 192, 219 ophiurus 192 parvus 34, 35, 36, 192-196,

October 189-208 November 209-228 December 229-244 219-222 Glyptemys insculpta 9, 18 Gopherus agassizii 71 morafkai 42, 83, 168, 234 Graptemys geographica 9, 18, 125 pseudogeographica 73 Gyrinophilus porphyriticus 18, 124 porphyriticus 7 Hapsidophrys smaragdinus 93, 94, 97 Heloderma suspectum 43, 44, 83, 167, 170, 236 Hemidactylus frenatus 154 mabouia 54, 56 Heterodon platirhinos 10, 18 Hyla avivoca 213 boans 100 chrysoscelis 100 eximia 37 geographica 100 miotympanum 37 plicata 37 pseudopuma 100 rosenbergi 100 versicolor 5, 18, 99-100, 124 Hynobius kimurae 174 lichenatus 174 nigrescens 174 tokyoensis 174 Iguana iguana 154 Incilius alvarius 127 periglenes 157-162 Indotyphlops braminus 77-81 pammeces 77 Isthmohyla rivularis 160 Kinixys erosa 54, 55, 93, 94 Kinosternon arizonense 19-22 flavescens 19 Lacerta bilineata 186 Lampropeltis annulata 37 greeri 37 mexicana 34, 35, 36, 37 triangulum 18, 37, 113-116, 125 syspila 126 triangulum 10

Lepidochelys olivacea 154

Lepidophyma gaigeae 37 Lepidothyris striatus 54, 56 Leptodactylus fuscus 171 labyrinthicus 102 Leptodeira septentrionalis 37 Leptophis ahaetulla 102 diplotropis 101-105 Limaformosa savorgnani 54, 58 Liophis poecilogyrus 102 Lithobates berlandieri 37, 103, 121-123, 213 blairi 163-165, 213 brownorum 213 catesbeianus 5, 18, 124, 213 clamitans 5, 18, 120, 125 forreri 213 neovolcanicus 153-157 palustris 5, 18, 125, 213, 214 pipiens 5, 18, 125 sphenocephalus 213 utricularius 214 sylvaticus 5, 18, 125 vaillanti 214 vibicarius 160 Madatyphlops 77 Malayopython reticulatus 50 Malayotyphlops 77 Masticophis 129, 131 bilineatus 64-65, 82-90, 237 flagellum 83, 179, 225, 226, 227 piceus 127-131 schotti 35, 36, 37 taeniatus 129 Mauremys leprosa 10 Mecistops cataphractus 97 Mehelya poensis 93, 96, 97 Melanosuchus niger 187 Micrurus tener 35, 36 Naia annulata annulata 54, 57, 58 melanoleuca 54, 57, 93, 95, 97 nigricollis 54, 57, 59 samarensis 25-26 Natriciteres fuliginoides 54, 58, 59, 93, 96, 97 Natrix natrix 134 scutata 134, 136, 138 tessellata 133-140 Necturus maculosus 18, 141-146, 175-177, 206 maculosus 7 Nerodia sipedon 18, 125 sipedon 11 Norops sagrei 154 Notophthalmus

viridescens 18, 99, 124 viridescens 7 Onychodactylus japonicus 174 Oopholis adelynhoserae 68 jackyhoserae 68 Orthosuchus stormbergi 216 Oxybelis aeneus 243 brevirostris 243 fulgidus 243 koehleri 243 microphthalmus 243 potosiensis 243 rutherfordi 243 wilsoni 243 Pantherophis bairdi 35, 36, 37 vulpinus 113, 115 Pelophylax ridibundus 133, 136, 138, 139 Pelusios gabonensis 54, 55 Philothamnus 94 carinatus 93, 95 heterodermus 93, 95, 97 nitidus nitidus 93, 95 Phrynohyas venulosa 102 Phrynosoma cornutum 199 orbiculare 33, 34, 35, 36, 37 solare 86, 199-204 Phyllodactylus tuberculosus 103 Pituophis catenifer 83, 148 deppei 33, 34, 35, 36, 37 Plestiodon brevirostris 37 dicei 33, 34, 35, 36, 37 lynxe 37 Plethodon angusticlavius 214 caddoensis 214 cinereus 124, 243 glutinosus 124, 214 hubrichti 243 metcalfi 214 montanus 71 yonahlossee 214 Podarcis muralis 186 Polemon collaris 93, 96, 97 fulvicollis 54, 58 Psammophis mossambicus 54, 58 Pseudacris clarkii 197-198 crucifer 5, 18, 118, 125 nigrita clarkii 197 streckeri 61-63 Pseudemys concinna 10 Pseudoeurycea cephalica 37

Pseudotriton montanus diastictus 214 Pternohlya fodiens 21 Python natalensis 10 sebae 93, 96 Ramphotyphlops braminus 77, 79 Rana berlandieri 37, 206 blairi 118, 163, 164 sphenocephala 118 sylvatica 117-120 vavapaiensis 206 Regina grahamii 189-191 septemvitatta 11, 18 Rhadinaea gaigeae 37 montana 37 Rheohyla miotympanum 37 Rhinella marina 26 Rhinocheilus lecontei 83 Salamandra salamandra 206 Salvadora grahamiae 35, 36, 37 hexalepis 83 Sauromalus ater 234, 235 varius 235 Scaphiopus couchii 213 Sceloporus aeneus 37 bicanthalis 37 chaneyi 34, 35, 36, 37 clarkii 86, 88, 89, 90 couchii 35, 36, 37 cyanogenys 35, 36, 37 goldmani 35, 36 grammicus 35, 36, 37 jarrovii 186 magister 150 melanorhinus 154 minor 35, 36 occidentalis 154 olivaceus 35, 36, 37 ornatus 34, 35, 36 parvus 35, 36, 37 poinsettii 35, 36, 37 samcolemani 37 scalaris 37, 41 torquatus 35, 36, 37 tristicus 186 virgatus 186 Scincella gemmingeri 37 Sclerophrys camerunensis 54, 56 Sistrurus catenatus 11, 66-67, 205 miliarius 70 Smilisca baudinii 103 fodiens 21, 103

Sonora semiannulata 234 Spea multiplicata 151 Sternotherus odoratus 9, 18, 74 Storeria dekayi 18, 37, 113 dekayi 11 Takydromus tachydromoides 174-175 Tantilla hobartsmithi 234 rubra 34, 35, 36, 37 Terrapene carolina 18 carolina 10, 206 coahuila 186 Tetrapodophis amplectus 50 Thamnophis brachystoma 11, 18 eques 37 exsul 34, 35, 36, 37 pulchrilatus 35, 36, 37 sauritus septentrionalis 12, 18 scalaris 37 sirtalis 18, 125 semifasciatus 113, 115 sirtalis 12, 187 sumichrasti 37 Thrasops flavigularis 93, 95, 97 Tlalocohyla smithii 103 Toxicodryas blandingii 93, 95, 97 pulverulenta 93, 95, 97 Trachemys scripta 73 elegans 3, 10, 18 Trachycephalus typhonius 101-105 Trachylepis affinis 54, 56 albilabris 93, 94 Trimorphodon biscutatus 236 lambda 42, 85 vilkinsonii 210 Tropidonotus gracilis 133 tantalus 133 Typhlina bramina 77 Typhlops biminiensis 79 braminus 77 Varanus jobiensis 47 Xantusia sanchezi 154 Xerotyphlops 77

## Author-Title Index for Volume 55 (2020)

January 1-24 February 25-48 March 49-72	April 73-92 May 93-112 June 113-132	July 133-152 August 153-172 September 173-188	October 189-208 November 209-228 December 229-244
Abernethy, K. A. See Pauwels, O. S. G.			
Araldi, A. See Pauwels, O. S. G.			
Archer, J. What You Missed at the Janua	ary Meeting: Mike Stefan	i	
Archer, J. What You Missed at the Febru	uary Meeting: Mike Redr	ner	
Banda-Leal, J. See Lazcano, D.			
Barker, D., and T. Barker Do Snakes H	ave Necks?		
Barker, T. See Barker, D.			
Barten, S. The Milksnake House: Eastern	n Milksnake, Lampropelt	is triangulum, Oviposition inside the T	op of a Chimney 113
Bauer, A. M. See Das, I.			
Boundenga, L. See Pauwels, O. S. G.			
Brecko, J. See Pauwels, O. S. G.			
Burger, R. M. On the Herpetological Co	llections of a Former Rou	ugh Rider, Colonel Martin Lalor Crimn	nins
Carlino, P. See Pauwels, O. S. G.			
Carter, H. Book Review: Indigo Snake:	All You Need to Know ab	oout Indigo Snake Care, Housing and F	<i>Feeding</i> by Paul Jason 229
Carter, R. Herpetological Art in the Indi	anapolis Zoo and the Adj	acent White River Gardens	
Carter, R. Herpetological Art at the Hou	ston Zoo, June 2018 .		
Cavataio, J. T. See Kutok, N. J.			
Chirio, L. See Pauwels, O. S. G.			
<b>Coleman, J. L.</b> Anthropogenic Drivers at Variable Harlequin Toad and Address		tangling the Disappearances of the Gol	
Crowley, R. A Letter from the Outgoing	CHS President		1
	Predation on Lithobates r	alas, A. Rodríguez-López and L. D. V neovolcanicus (Hillis & Frost, 1985) by xico	y Quiscalus mexicanus
Cruz-Sáenz, D. See also García Mata, E.	. S.		
Das, I., and A. M. Bauer Unissued Phila	telic Essays of Basutolan	d 1933, and the Lesotho Crocodile Stat	mp Issues
Dorn, A. D. See Kutok, N. J.			
Favela-Lara, S. See Lazcano, D.			
Ferrer, D. M. See Lazcano, D.			
Fonteyn, D. See Pauwels, O. S. G.			
-	of Western Mexico 23: P	<b>íos-Martínez, L. A. Hernández-Dávil</b> a redation by a Pacific Coast Parrot Snak pality Huejutla de Reyes, Hidalgo, Mex	e (Leptophis diplotropis)
García-Salas, J. A. See Lazcano, D.			
Goldberg, S. R. Notes on Reproduction	of Strecker's Chorus Frog	g, Pseudacris streckeri (Anura: Hylidae	e), from Oklahoma 61
Goldberg, S. R. Notes on Reproduction	of Rio Grande Leopard F	rogs, <i>Lithobates berlandieri</i> (Anura: Ra	anidae), from Texas 121
Goldberg, S. R. Notes on Reproduction	of Plains Leopard Frogs,	Lithobates blairi (Anura: Ranidae), fro	m Oklahoma
Goldberg, S. R. Notes on Reproduction	of the Spotted Chorus Fro	og, Pseudacris clarkii (Anura: Hylidae)	, from Oklahoma and Texas 197
Goldberg, S. R. Notes on Reproduction	of the Green Treefrog, Dr	ryophytes cinereus (Anura: Hylidae), fr	om Oklahoma
Gray, B. The Herpetofauna of Presque Is	sle State Park, Erie, Penns	sylvania: Annotated Checklist and Com	prehensive Bibliography 2
Hartzell, S. M. Forty-two New Township	p Records for Amphibian	s and Reptiles in Columbia County, Pe	nnsylvania, USA 124
Hernández-Bocardo, S. C. See Lazcano	, D.		

Hernández-D	Dávila, L. A. See García Mata, E. S.	
Jamen, C. P.	See Kutok, N. J.	
Kadeyeva, M	I. See Pauwels, O. S. G.	
Kaya, A. B.	See Pauwels, O. S. G.	
Kovshar, V.	See Pauwels, O. S. G.	
Krause, R.	Letter	111
	<b>J. T. Cavataio, A. D. Dorn, C. P. Jamen and T. D. Schramer</b> The End of a Quarter-century Hiatus: Reconfirming <i>grahamii</i> Baird and Girard, 1853 (Graham's Crayfish Snake) in DuPage County, Illinois, USA	189
	<b>J. A. García-Salas, J. Banda-Leal, L. D. Wilson, S. Favela-Lara, M. E. Solis-Baraja and S. Pacheco-Treviño</b> Mexican Herpetofauna 34: Herpetofauna Associated with Pine Forests in Nuevo León, Mexico	27
, ,	<b>S. C. Hernández-Bocardo and L. D. Wilson</b> Notes on Mexican Herpetofauna 35: Use of Artificial Shelters by Pairs of <i>otus parvus</i> Knight and Scudday, 1985 (Squamata: Anguidae), with Notes on Behavior and Interactions in Captivity	192
	J. Banda-Leal, D. M. Ferrer and L. D. Wilson Notes on Mexican Herpetofauna 36: A New Locality in Nuevo León, for the Pygmy Alligator Lizard, <i>Gerrhonotus parvus</i> Knight & Scudday, 1985 (Squamata: Anguidae)	219
Lazcano, D.	See also Cruz-Sáenz, D.	
Lazcano, D.	See also García Mata, E. S.	
Makemba, R	. N. See Pauwels, O. S. G.	
McDowell, W	V. T. A Brief History of the American Literature and a Collection Note on <i>Takydromus tachydromoides</i> (Family	
Lacertida	ae)	174
Moreels, M.	J. See Pauwels, O. S. G.	
Morelle, S.	See Pauwels, O. S. G.	
Ngoubangoy	e, B. See Pauwels, O. S. G.	
Osisly, R. S	See Pauwels, O. S. G.	
Pacheco-Trev	viño, S. See Lazcano, D.	
Palis, J. G.	Solving the Mystery of Wood Frogs at Snake Road	117
Pallemaerts,	L. See Pauwels, O. S. G.	
Pauly, A. S	ee Pauwels, O. S. G.	
· · ·	<b>S. G., and J. Brecko</b> A Case of Predation by <i>Naja samarensis</i> (Elapidae) on <i>Cyclocorus nuchalis nuchalis</i> phiidae) on Mindanao Island, Philippines	25
	S. G., A. B. Kaya, L. Boundenga, P. Carlino, L. Chirio, M. J. Moreels, S. Morelle, B. Ngoubangoye, L. Pallemaerts A. Abernethy Miscellanea Herpetologica Gabonica XV	54
-	S. G., A. Pauly, A. Araldi, R. N. Makemba, D. Fonteyn, R. Oslisly and A. M. Whittaker Miscellanea Herpetologica a XVI	93
Coloniza	S. G., M. Kadeyeva, V. Kovshar, A. Sakharbayev, F. A. Sarayev, S. Sarsengaliyev, S. Ukhov and S. Yerbulekov ation of Artificial Islands in the Kazakh Sector of the Caspian Sea by the Semiaquatic Snake <i>Natrix tessellata</i> (Squamata: ae)	133
Repp, R, A.	Turtle speak? An Unusual (Unique?) Display of Mass Social Interaction among Arizona Mud Turtles (Kinosternon	
arizonen	se) in Pima County, Arizona	19
Repp, R. A.	The Odyssey of 10,000 Beers: The First Six-pack of the Suizo Mountain Project	40
Repp, R. A.	MABI I'm Amazed!	64
Repp, R. A.	MABI I'm Amazed! (Part 2 of 2)	82
Repp, R. A.	The Great Action Jackson Coachwhip Capers	106
Repp, R. A.	Nastycophis—a Snake as Great as its Name (Part 3)	127
Repp, R. A.	Memo to the Neighborhood Roadrunners: Leave My Yard Lizards Alone, or Else	147
Repp, R. A.	Kim, the Serendipitous Tiger Rattlesnake	166
Repp, R. A.	Notes on the Tiger Rattlesnake (Crotalus tigris) in the Vicinity of Tucson, Arizona: Comparative Frequency of	1
		178
Repp, R. A.		199
Repp, R. A.	Adventures with Dr. Rosen—Part 1	
Repp, R. A.	Adventures with Dr. Rosen – Part 2	233

Bulletin of the Chicago Herpetological Society 55(12):243, 2020

## Herpetology 2020

In this column the editorial staff presents short abstracts of herpetological articles we have found of interest. This is not an attempt to summarize all of the research papers being published; it is an attempt to increase the reader's awareness of what herpetologists have been doing and publishing. The editor assumes full responsibility for any errors or misleading statements.

#### **COMPETITION BETWEEN SALAMANDER SPECIES**

T. R. Brophy and N. Reichenbach [2020, The Herpetological Bulletin 152:1-6] note that the Peaks of Otter salamander, Plethodon hubrichti, is a montane species found at altitudes above 442 m within a 117 km<sup>2</sup> area of the Blue Ridge Mountains in central Virginia, USA. In areas where this species is sympatric with the eastern red-backed salamander (Plethodon cinereus) it seemed likely that P. hubrichti populations were either depressed or eliminated. The habitability of areas beyond the current range boundaries for P. hubrichti is supported by several disjunct populations in areas sympatric with P. cinereus. From 2009 to 2012 the authors tested whether P. hubrichti was negatively impacted by competition with P. cinereus by removing P. cinereus from treatment plots at three sympatric field locations. The number of surface-active (SA) P. hubrichti increased significantly more on treatment plots than on corresponding reference plots, whereas the number of SA P. cinereus decreased significantly more on treatment plots than on reference plots. The removal of every one P. cinereus from the treatment plots led to an increase of 0.69 P. hubrichti. These results emphasize the importance of conserving mature hardwood forests along the perimeter of the P. hubrichti distribution, where it is sympatric with P. cinereus, so as to prevent future range contraction of this vulnerable species.

#### NOT WITHERING ON THE EVOLUTIONARY VINE

R. C. Jadin et al. [2020, Organisms Diversity & Evolution 20(4):723-746] note that the vine snake genus, Oxybelis, currently is composed of four taxa despite numerous studies suggesting and describing multiple taxa within the O. aeneus complex. The authors utilize a multilocus molecular dataset (i.e., cyt b, ND4, 12S, 16S, cmos, PRLR, 3663 bp) to conduct phylogenetic analyses to assess the evolutionary history of Oxybelis. The molecular analyses find three major lineages of Oxybelis (i.e., O. aeneus complex, O. brevirostris, O. fulgidus complex) with a sister relationship between O. brevirostris and the O. aeneus complex to the exclusion of the O. fulgidus complex. More specifically, O. aeneus appears to harbor at least five taxa currently unrecognized while O. fulgidus was found to be paraphyletic with respect to O. wilsoni, suggesting cryptic diversity and novel taxa in that clade as well. Additionally, the authors use morphological data in concert with the molecular analyses and the literature to support removing Oxybelis microphthalmus Barbour and Amaral, 1926; Oxybelis potosiensis Taylor, 1941; and Dryophis vittatus Girard, 1854 from the synonymy of O. aeneus. Finally, they describe two new species, O. koehleri and O. rutherfordi, from Central America and northern South America, respectively.

## Advertisements

For sale: **highest quality frozen rodents**. I have been raising rodents for over 30 years and can supply you with the highest quality mice available in the U.S. These are always exceptionally clean and healthy with no urine odor or mixed in bedding. I feed these to my own reptile collection exclusively and so make sure they are the best available. All rodents are produced from my personal breeding colony and are fed exceptional high protein, low fat rodent diets; no dog food is ever used. Additionally, all mice are flash frozen and are separate in the bag, not frozen together. I also have ultra low shipping prices to most areas of the U.S. and can beat others shipping prices considerably. I specialize in the smaller mice sizes and currently have the following four sizes available: Small pink mice (1 day old -1 gm), \$25/100; Large pink mice (4 to 5 days old -2 to 3 gm), \$27.50/100; Small fuzzy mice (7 to 8 days old -5 to 6 gm), \$30/100; Large fuzzy mice / hoppers (10 to 12 days old -8 to 10 gm), \$35/100 Contact Kelly Haller at 785-224-7291 or by e-mail at kelhal56@hotmail.com

Free to a good home. Back issues of the CHS *Bulletin* from October 2012 to the present. Boxed and ready to be picked up at my home in Oak Park. Rob Streit, 708-383-6830.

Line ads in this publication are run free for CHS members — \$2 per line for nonmembers. Any ad may be refused at the discretion of the Editor. Submit ads to <u>mdloogatch@chicagoherp.org</u>.

## NEW CHS MEMBERS THIS MONTH

Jessica Daly Michael Deutsch Andrew and Brittany Hedman Luke Larter Corrie Navis Hope Nye Eric Stitt Holly Zak Scott Zonis



## **UPCOMING MEETINGS**

Until in-person meetings again become possible the Chicago Herpetological Society will be holding monthly general meetings online via Zoom Webinar. The December Webinar will take place on Wednesday, December 30, at 7:30 P.M. Chicago time, and will feature two half-hour-long presentations. **Dr. Steve Barten**, a veterinarian and outstanding nature photographer, will split his segment between interesting veterinary cases and interesting episodes of field herping. **Cindy Rampacek**, a Milwaukee resident and former CHS board member, will speak about some of the many cases she has worked on involving animal control and rescues.

Viewers will be able to submit questions to be answered by the speaker following each segment.

Please check the CHS website or Facebook page each month for information on the program. Information about attending a Zoom Webinar can be found here:

<a href="https://support.zoom.us/hc/en-us/articles/115004954946-Joining-and-participating-in-a-webinar-attendee-">https://support.zoom.us/hc/en-us/articles/115004954946-Joining-and-participating-in-a-webinar-attendee-</a>

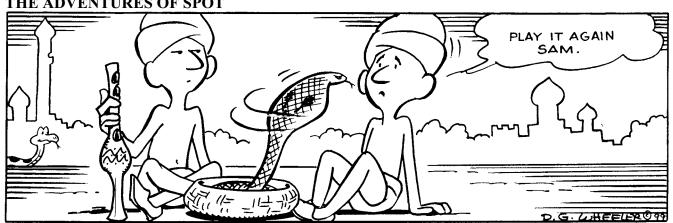
## **Board of Directors Meeting**

Are you interested in how the decisions are made that determine how the Chicago Herpetological Society runs? And would you like to have input into those decisions? The next board meeting will be held online. If you wish to take part, please email <u>akolb@chicagoherp.org</u>.

## **ELECTION RESULTS**

As a result of the elections held online and by mail, the following officers and members-at-large will serve on the CHS Board of Directors for the year 2021.

President: John Archer Vice-president: Rachel Bladow Treasurer: John Archer Recording Secretary: Rich Crowley Media Secretary: Stephanie Dochterman Membership Secretary: Mike Dloogatch Sergeant-at-arms: Tom Mikosz Members-at-large: Amelia Pollack-Cotter Kyle Houlihan Margaret Ann Paauw Immediate Past President: John Gutierrez



## THE ADVENTURES OF SPOT

Periodicals Postage Paid at Chicago IL

# CHICAGO HERPETOLOGICAL SOCIETY Affiliated with the Chicago Academy of Sciences

2430 North Cannon Drive • Chicago, Illinois 60614