# A phylogenetic analysis of the genus Eunice (Eunicidae, polychaete, Annelida) 

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#### Abstract

Species of Eunice are distributed worldwide, inhabiting soft and hard marine bottoms. Some of these species play significant roles in coral reef communities and others are commercially important. Eunice is the largest and most poorly defined genus in Eunicidae. It has traditionally been subdivided in taxonomically informal groups based on the colour and dentition of subacicular hooks, and branchial distribution. The monophyly of Eunice and of its informal subgroups is tested here using cladistic analyses of 24 ingroup species based on morphological data. In the phylogenetic hypothesis resulting from the present analyses Eunice and its subgroups are paraphyletic; the genus may be divided in at least two monophyletic groups, Eunice s.s. and Leodice, but several species do not fall inside these two groups. Most of the traditional characters used in the taxonomy of Eunice are homoplasies; however, characters used for the first time in this study, such as certain jaw characters and characters derived from a close examination of chaetal variation along the body, are promising sources of phylogenetic signal. © 2007 The Linnean Society of London, Zoological Journal of the Linnean Society, 2007, 150, 413-434.


ADDITIONAL KEYWORDS: branchial distribution - Eunicida - jaw - Leodice - mandible - maxillae - phylogeny - subacicular hooks.

Eunice Cuvier, 1817 is the most species rich of the ten genera of the family Eunicidae, comprising around 220 species. It is distributed worldwide, but it is most common in shallow tropical waters (Fauchald, 1992). Species of Eunice inhabit soft and hard marine bottoms, many burrow into hard corals and calcareous algae or live in their crevices, and play significant roles in coral reef communities as bioeroders (Hutchings, 1986), but possibly also in assembling those communities (Roberts, 2005). Eunice includes some commercially important species used as bait in com-

[^0]mercial and leisure fishing (Gambi et al., 1994). Sexually mature Eunice specimens have a large size range, varying from a few mm to 6000 mm in length (Fauchald, 1992), and both very large and very small species have been described without adequate knowledge of ontogenetic variation.

The taxonomy within Eunice is difficult because of the poor knowledge of ontogenetic and intraspecific variation of characters (Steiner, Nogueira \& Amaral, 2002). In addition, many species have been described based on incomplete specimens (Miura, 1986), and several species are known just from type specimens (Fauchald, 1992). Furthermore, definition of the genus is problematic as it is based on the following plesiomorphies: presence of three antennae, a pair of palps, a pair of peristomial cirri, and a complete set of chaetal types (limbate, pectinate, aciculae, compound, and subacicular hooks) (Orensanz, 1990); features
that are also present in the well-defined eunicid genus Euniphysa (Lu \& Fauchald, 2000) and in another eunicean family, Onuphidae.

The poor definition of Eunice and the insufficient understanding of character variation led to the description of genera such as Leodice Lamarck, 1818, and Nicidion Kinberg, 1865, both of which were later considered junior synonyms of Eunice. Leodice, named for Leodice antennata Lamarck, 1818, was not clearly differentiated from Eunice, and both names were used interchangeably through the first third of the 20th century (Hartman, 1944). Nicidion included Eunice species lacking branchiae, a character not considered acceptable at either the generic (Hartman, 1944) or subgeneric levels (Fauchald, 1970) because it is highly variable, and individuals within the same 'Nicidion' species may or may not lack branchiae.

Currently Eunice is subdivided into taxonomically informal groups based on colour and dentition of subacicular hooks (Ehlers, 1868; Hartman, 1944) and branchial distribution patterns (Fauchald, 1970). Miura (1986) suggested that the Eunice informal groups based on the characteristics of subacicular hooks could correspond to genera or subgenera; he also suggested that the presence of few branchial filaments present along the whole body was the ancestral branchial distribution pattern in Eunice.

The monophyly of Eunice is currently disputed. Studies on the phylogeny of the order Eunicida using molecular data resulted in paraphyletic Eunice and Eunicidae (Struck, Westheide \& Purschke, 2002; Struck, Purschke \& Halanych, 2006). Fauchald (1992) attempted a phylogenetic analysis of Eunice based on morphological features of the type specimens to test the relationships within the genus. His results were generally inconclusive and the only consistent clade in all trees was a group that included all species with yellow hooks.

The purposes of the present study are to test the monophyly of the genus Eunice, as well as of its informal groups, based on colour and dentition of the subacicular hooks and branchial distribution patterns, and to provide a hypothesis of phylogeny for Eunice that can be used as a framework for future studies on Eunice and Eunicidae in general. For this we used an expanded morphological character set, compared with the one used by Fauchald (1992), and better preserved material than represented by the types.

## MATERIAL AND METHODS

## TAXONOMIC SAMPLING

We examined 24 Eunice species (Table 1) representing all taxonomically informal groups (Table 2) and the morphological diversity within these. Outgroup taxa
to test the monophyly of Eunice and/or root the phylogenetic analyses were species of other eunicid genera, Marphysa, Palola, and Lysidice, members of the other four best known families of the order Eunicida, Onuphidae, Dorvilleidae, Lumbrineridae, and Oenonidae, and members of Amphinomida, a probable sister group to Eunicida (Rouse \& Fauchald, 1997; but see Struck et al., 2006 for a discussion) (Table 3).

Each specimen was examined under stereo and compound microscopes. Specimens were dissected as described by Day (1967) in order to code the characters of the buccal apparatus. To determine chaetal variation along the body, six parapodia of each specimen were examined, one from the first and last $2.5 \%$ (determined by number of chaetigers) of the body, respectively, and four from the median chaetiger of each quarter of the body. We quantified branchial distribution as the percentage of continuous chaetigers in which branchiae are present, as used by Fauchald (1992).

## Specimens

The material examined in this study came from the following institutions: ECOSUR, Colecion de Referencia ECOSUR, Chetumal, Mexico; IBUFRJ, Departamento de Zoologia, Instituto de Biologia, UFRJ, Rio de Janeiro, Brazil; IRSNB, Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium; SMNH, Swedish Museum of Natural History, Stockholm, Sweden; USNM, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA.

## PhYLOGENETIC ANALYSES

We coded two different character matrices, one with composite characters (composite analyses) and another with reductive characters (reductive analyses) (sensu Strong \& Lipscomb, 1999), to check the sensitivity of our data set to the coding methods, which differ in the number of inapplicable characters and in the underlying assumptions about primary hypotheses of homology.

A matrix of 59 composite ( 74 reductive; Appendix 1) morphological characters coded for 37 taxa (24 Eunice ingroup taxa plus 13 outgroup taxa) (Table 4; Appendix 2) was analysed under maximum parsimony optimization criteria. Eleven characters (20 in the reductive matrix) are uninformative autapomorphies or polymorphisms; however, these were included in the analyses because they have been observed in other species of the appropriate genus or family (J. Zanol \& K. Fauchald, pers. observ.) not included here because of the limited taxonomic sampling. Alternatively, some of these characters represent polymorphic variation that is uninformative because of the way current analytic

Table 1. Ingroup taxa examined in the present study (see Table 2 for a description of the groups)


Table 2. Taxonomically informal groups employed in the subdivision of the genus Eunice (adapted from Hartman, 1944; and Fauchald, 1970), used in the present study in the choice of the species to be examined. Branchial distribution was quantified as the percentage of continuous chaetigers in which branchiae is present, as used in Fauchald (1992)
$\left.\begin{array}{llll}\hline & & \begin{array}{l}\text { Number } \\ \text { of species }\end{array} \\ \text { Groups } & \text { Characteristics } & \begin{array}{l}\text { Total of } \\ \text { in this }\end{array} \\ \text { species* } \\ \text { study }\end{array}\right]$
*Approximate total.

Table 3. Outgroup taxa examined in the present study

| Family | Species | Number, collection, and locality of specimens examined |
| :---: | :---: | :---: |
| Amphinomidae | Amphinome rostrata (Pallas, 1766) | 1 (USNM 51511) from Trinidad and Tobago, Matura Beach |
| Oenonidae | Oenone fulgida (Savigny, 1818) | 1 (USNM 53751) from, USA, Florida, Conch Key |
| Lumbrineridae | Lumbrineris fragilis (Müller, 1776) <br> Lumbrineris latreille Audouin \& Milne Edwards, 1834 | 1 (USNM 30097) from USA, Massachusetts 1 (USNM 53018) from USA, North Carolina |
| Dorvilleidae | Dorvillea sociabilis (Webster, 1879) | 1 (USNM 33830) from USA, Virginia |
| Onuphidae | Diopatra tridentata Hartman, 1944 Kinbergonuphis tenuis (Hansen, 1882) | 1 (IBUFRJ 375) from Brazil, Bahia 1 (IBUFRJ 376) from Brazil, Bahia |
| Eunicidae |  <br> Attolini, 2000 <br> Palola cf. viridis Gray, in Stair, 1847 <br> Lysidice ninetta Audouin \& Milne <br> Edwards, 1833 <br> Marphysa sanguinea (Montagu, 1807) <br> Marphysa cf. sanguinea (Montagu, 1807) <br> Marphysa cf. atlantica Kinberg, 1865 <br> Marphysa sp. | 1 (IBUFRJ 381) from Brazil, $20^{\circ} 40^{\prime} 26^{\prime} \mathrm{S} 37^{\circ} 42^{\prime} 36^{\prime} \mathrm{W}$ <br> 2 (IBUFRJ 382) from Brazil, $17^{\circ} 48^{\prime} 01^{\prime} \mathrm{S} 35^{\circ} 52^{\prime} 52^{\prime} \mathrm{W}$ <br> 1 (SMNH 28292) from Western Australia <br> 1 (ECOSUR Euni-2) from Mexico <br> 1 (IBUFRJ 383) from France, Dinard <br> 3 (IBUFRJ 377) from UK, Plymouth Sound <br> 1 (IBUFRJ 378) from Brazil, Paraná <br> 1 (IBUFRJ 379) from Brazil, Rio de Janeiro, $22^{\circ} 55^{\prime}$ S $44^{\circ} 10^{\prime} \mathrm{W}$ <br> 3 (IBUFRJ 380) from Brazil, Espírito Santo |

Table 4. Composite coding character matrix

|  | 1 | 1111111112 | 2222222223 | 3333333334 | 4444444445 | 555555555 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species/characters | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 123456789 |
| Amphinome rostrata | 0001?? ?311 | 1101100200 | 0-------- | -0-020214- | 4010000001 | ---0----0 |
| Oenone fulgida | 01000---11 | 2000--3001 | 1010-44000 | -11310100- | 0--0000000 | 110210101 |
|  |  |  |  |  |  | 4 |
| Lumbrineris fragilis | 00000---00 | ---0--3101 | 1121133100 | 010100100- | 0--0000000 | 0000----1 |
| Lumbrineris latreille | 01000---00 | ---0--2101 | 1121133100 | 010100200- | 0--0002000 | 1100----1 |
| Dorvillea sociabilis | 01000---01 | 1201104101 | 200?-? ? ? ? | -11011103- | 0--0001100 | 1100----1 |
|  | 2 |  |  |  |  |  |

Table 4. Continued

|  | 1 | 1111111112 | 2222222223 | 3333333334 | 4444444445 | 555555555 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species/characters | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 123456789 |
| Diopatra tridentata | 00002--011 | 0121121011 | 1221011101 | 11111?0021 | 3101010000 | 110210101 |
| Kinbergonuphis tenuis | 21002--111 | 0121121011 | 1221011101 | 11111?0021 | 2111010000 | 110210101 |
| Lysidice ninetta | 0000101-11 | 1110--1101 | 1221001101 | 0313120121 | 0--1001100 | 000200001 |
|  | 1 | 2 |  | 2 |  | 1111 |
| Palola brasiliensis | 0000101211 | 0111111111 | 1221022101 | 0212120220 | 1220001100 | 0100----1 |
|  | 112 | 2 |  |  |  |  |
| Palola cf. viridis | 0000102011 | 0111112111 | 1221022101 | 0212120220 | 1220001100 | 0100----1 |
| Marphysa sp. | 0100100111 | 0111111101 | 1221001101 | 1111110221 | 2211001100 | 000201001 |
|  | 1 | 2 |  |  |  |  |
| Marphysa sanguinea | 1000101111 | 0111112101 | 1221001101 | 1111110211 | 2211100010 | 000200001 |
|  | 1 |  |  |  |  | 11 |
| Marphysa cf. atlantica | 1010101111 | 0111112101 | 12210011?1 | 1111100211 | 2211101100 | 000201001 |
| Eunice aphroditois | 0100102111 | 0011011111 | 1221000111 | 1111110211 | 2111001100 | 000201000 |
|  | 11 | 11 |  | 2 |  | 11 |
| Eunice denticulata | 0100100111 | 0111011111 | 1221000111 | 1111110110 | 2211001100 | 000201001 |
|  | 2 | 12 |  | 2222 |  | 1 |
| Eunice cf. flavopicta | 0100102111 | 0111112111 | 1221000111 | 1111110110 | 2111001100 | 000201000 |
| Eunice norvegica | 0100102111 | 0111112111 | 1221001111 | 1111110221 | 2111001100 | 000201000 |
|  | 1 |  |  |  |  | 1 |
| Eunice cariboea | 0000100111 | 0111111111 | 1221001101 | 1111100221 | 0--1001100 | 000201000 |
|  | 11 | 2 |  |  |  | 1 |
|  | 2 |  |  |  |  |  |
| Eunice frauenfeldi | 0100101011 | 0111111111 | 1221001111 | 1111110221 | 2211001100 | 000201000 |
| Eunice cf. grubei | $010010 ? 011$ | ? 111112111 | 1221001111 | 1111120221 | 2011001100 | 010201000 |
| Eunice thomasiana | 0100101011 | 0211211111 | 1221001111 | 1111112121 | 2111001100 | 010201000 |
| Eunice dubitata | 0100101011 | 0111112111 | 1221001111 | 1111112121 | 2001001100 | 010201000 |
| Eunice cf. torquata | $\begin{aligned} & 0000101011 \\ & 1 \end{aligned}$ | 0211212111 | 1221001111 | 1111112121 | 2011001100 | 010201000 |
| Eunice fucata | 0100101011 | 0111111111 | 1221001111 | 1111110221 | 2101001100 | 010101100 |
|  | 2 | 2 |  |  |  |  |
| Eunice sebastiani | 0100101111 | 0?11?12111 | 1221001111 | 1111112111 | 2111001100 | 01011110? |
| Eunice harassii | 0100101011 | 0111111111 | 1221001111 | 1111110121 | 2111001100 | 110200100 |
|  | 12 | 2 |  | 22 |  |  |
| Eunice antennata | 0100101011 | 0211211111 | 1221011101 | 1111110021 | 2111001200 | 110300101 |
|  | 1 | 2 |  | 2 |  |  |
| Eunice rubra | 0100100011 | 0211211111 | 1221011101 | 1111110121 | 2111001200 | 110300101 |
|  | 111 |  |  | 22 |  | 1 |
|  | 2 |  |  |  |  |  |
| Eunice miurai | 0100101011 | 0211211111 | 1221011101 | 1111112121 | 2101001200 | 110300100 |
|  |  |  |  |  |  | 1 |
| Eunice websteri | 0000112011 | 0111111111 | 1221011101 | 1111110221 | 2001001100 | 110210101 |
|  | 11 |  |  |  |  |  |
| Eunice vittata | 0100112011 | 0111111111 | 1221011101 | 1111110221 | 2001001100 | 110310101 |
| Eunice atlantica | 0100110011 | 0111112111 | 1221011101 | 1111110221 | 2001001100 | 110300101 |
|  | 2 |  |  |  |  |  |
| Eunice unifrons | 0100112011 | 0111111111 | 1221011101 | 1111110221 | 2001001100 | 110300101 |
|  | 1 | 2 |  |  |  |  |
| Eunice pennata | 0100100011 | 0111112111 | 1221011101 | 1111110221 | 2001001100 | 110200101 |
| Eunice cf. semisegregata | 0100101011 | 0111112111 | 1221011111 | 1111110111 | 2001001100 | 110200101 |
|  | 1 |  |  | 22 |  |  |
| Eunice cf. atlantica | 0000100011 | 0111112111 | 1221011101 | 1111100221 | 2001001100 | 110300101 |
| Eunice stigmatura | 0100102011 | 0111112111 | 1221011101 | 1111110221 | 2011001100 | 110300101 |
|  | 1 |  |  |  |  |  |

[^1]programs treat polymorphic characters (Wiens, 2000); however, information about this variation is certainly useful for understanding, for example, allometries caused by growth and other kinds of variation.
Trees were rooted using the outgroup method. Characters were treated as unordered and equally weighted; polymorphic characters were coded as such. The tree search was heuristic, using the option TBR (tree bisection and reconnection) + TBR in NONA (Goloboff, 1999). Analyses started with a stepwise addition tree with taxa randomly added in 2000 replicates and 100 trees held on each step of the TBR. During the search branches were collapsed when the minimum branch length was zero.
Parsimony analyses results were summarized in strict, $50 \%$ majority rule and Adams consensus of all most parsimonious trees. Branch support was calculated using bootstrap (Felsenstein, 1985) (2000 replicates, ten search replicates and max TBR) and Bremer support (Bremer, 1988). We used Winclada (Nixon, 2002) and NONA (Goloboff, 1999) to construct the matrices, run parsimony and bootstrap analyses and examine trees. Bremer support was calculated using PaUP*4.0b10 (Swofford, 2001) and TreeRot (Sorenson, 1999). We used ACCTRAN optimization to examine character evolution, but only unambiguous optimized characters (those that have the same optimization under ACCTRAN or DELTRAN) are considered as support synapomorphies for clades.
In order to determine how much longer trees are when traditional taxonomically informal groups (Table 2), Eunice, and Eunicidae are constrained as monophyletic, we ran additional parsimony analyses (using the composite matrix and the same settings as described above) constraining these groups as monophyletic.

## DESCRIPTION OF CHARACTERS

Only composite characters are described here (see Appendix 1 for a list of reductive characters). Uninformative characters are underlined.

1. Shape of cross section of body: 0 , circular; 1 , dorsoventrally flattened; 2 , ventrally convex.
2. Body shape: 0, cylindrical, abruptly tapering anteriorly and posteriorly; 1, evenly tapering from anterior-median chaetigers.
3. Relative width and length of chaetigers: 0 , at most ten times wider than long; 1 , more than ten times wider than long.
4. Posterior extension of prostomium (caruncle): 0 , absent; 1, present.
5. Frontal lips (sensu Paxton, 1998): 0, frontal edge entire; 1, separated frontally by a narrow notch (Fig. 1A-C); 2, separated frontally by a distinct space (Fig. 1D).
6. Frontal lips separated by a narrow notch anteriorly: 0, round (Fig. 1A, B); 1, truncate (Fig. 1C). This character refers to the shape of the anterior end of the frontal lips in dorsal view of the specimen.
7. Frontal lips separated by a narrow notch, dorsally: 0 , inflated (Fig. 2A); 1, flat (Fig. 2B); 2, dimpled (Fig. 2C). This character refers to the dorsal shape of the frontal lip in frontal view of the specimen. Characters 6 and 7 are linked to character state 1 of character 5.
8. Arrangement of prostomial appendages: 0 , antennae and palps evenly spaced (Fig. 1C, D); 1, median antenna isolated by a gap from the other antennae and palps (Fig. 1A); 2, palps isolated by a gap from the antennae (Fig. 1B); 3, head appendages on conspicuously different regions of the prostomium, median antennae on the caruncle.
9. Median antennae: 0 , absent; 1 , present.
10. Lateral antennae: 0 , absent; 1 , present.
11. Antennal styles: 0 , digitiform, tapering, or clavate (Fig. 3A-C); 1, medially inflated (fusiform) (Fig. 3D); 2, button shaped (Fig. 3E). Fauchald (1992) distinguished digitiform, tapering, and clavate states; further study has shown that there is a continuous gradation among these 'states' making differentiation impossible. Hartman (1944) and Miura (1977) suggested that the different shapes of styles are of little taxonomic significance.


Figure 1. Schematic drawing of the dorsal view of the prostomium and anterior end of peristomium; circles and semicircles represent the arrangement of prostomial appendages (antennae and palps). A, round frontal lips separated by a narrow notch, median antennae isolated by a gap from the other antennae and palps. B, round frontal lips separated by a narrow notch, palps isolated by a gap from the antennae. C, truncate frontal lips separated by a narrow notch, antennae and palps evenly spaced. D, frontal lips separated by a distinct space, antennae and palps evenly spaced.


Figure 2. Schematic drawing of the frontal view of specimens that have frontal lips separated by a narrow notch. A, frontal lips dorsally inflated. B, frontal lips dorsally flat. C, frontal lips dorsally dimpled.


Figure 3. Schematic drawing of the shape of prostomial appendages. A, digitiform. B, tapering. C, clavate. D, fusiform. E , button shaped.
12. Articulations of antennal styles: 0 , absent; 1 , short or long cylinders (Fig. 4A); 2, moniliform (Fig. 4B). In many specimens the styles are strongly wrinkled and distinction between wrinkles and true articulations may be difficult (Day, 1967 e.g. Marphysa fragilis Treadwell, 1911; Eunice mucronata Moore, 1903 in Fauchald, 1992). Styles were considered truly articulated only if they had at least one complete groove visible on all sides of the style. When articulations were present along the whole style, scoring was based on the shape present in the basal-median region of the style.
13. Antennal ceratophore: 0 , absent; 1 , short, usually ring-shaped; 2, relatively long, usually articulated. Shape (character 11), articulations (character 12), and ceratophores (character 13) of median and lateral paired antennal styles are coded in the same character because the two kinds of antennae do not appear to be morphologically independent. The presence of both kinds of antennae is a clearly independent feature, but they are always of the same kind in these characters when both are present.


Figure 4. Schematic drawing of the shape the articulations of prostomial appendages. A, short or long cylinders. $B$, moniliform.
14. Palpal styles: 0 , absent; 1 , digitiform, tapering, or clavate (Fig. 3A-C). We considered the palps as the missing prostomial appendages in Lysidice ninetta Audouin \& Milne Edwards, 1833 and Oenone fulgida Savigny, 1818. Antennae and palps used to be considered as the same structures (e.g. Chamberlin, 1919; Treadwell, 1921), and the nature of the missing appendages in the genera Lysidice and Nematonereis, as well as in members of the family Oenonidae, was not a concern. Orrhage (1995) demonstrated that the innervation of the palps differed from that of the three antennae, thus corroborating an idea first put forward by Binard \& Jeener (1928) of separating the five head appendages in Eunice into three antennae and an outer pair of palps (called AI in Fauchald, 1992). In the genus Lysidice in which only three head appendages are present it is still not clear how these should be considered in relation to the appendages present in other eunicids. Here the lateral appendages are considered antennae, because of their placement between the eyes on the posterior region of the prostomium. However, Steiner (2000) considered the paired appendages in the specimen of $L$. ninetta she examined as palps, because of their position in front of the eyes.
15. Palpal styles articulations: 0 , absent; 1 , short or long cylinders (Fig. 4A); 2, moniliform (Fig. 4B).
16. Palpophores: 0, absent; 1, short, usually ringshaped; 2 , relatively long, usually articulated.
17. Peristomial ventrolateral lips: 0 , absent; 1 , set off by distinct grooves; 2, visible only as elevated surfaces; 3, scoop shaped; 4, medially short, about half as long as lateral side of the peristomium.
18. Peristomium: 0 , a single ring; 1 , two rings distinct at least dorsally; 2, reduced to region around the mouth.
19. Peristomial cirri: 0 , absent; 1 , present.
20. Mouth: 0 , extending to the ventral side of anterior chaetigers; 1 , restricted to peristomium.
21. Number of rows of maxillary plates: 0 , absent; 1 , one pair; 2, two or more pairs.
22. Calcium carbonate mineralzing jaws: 0 , absent; 1 , calcite; 2, aragonite.
23. Maxillary carrier: 0, ctenognath; 1, prionognath; 2, labidognath. We describe the shape of the maxillary carriers using the terminology generally used to group the jaws. This is because of both the debate about the ctenognath carrier (e.g. Paxton, 2004) and the consistent shape of the carriers in prionognaths and labidognaths (Colbath, 1989). Prionognath carriers are narrow, many times narrower than the posterior end of maxillae I (MxI), longer than the remaining maxillary apparatus anterior to it, not covered by muscles, and not attached to MxI. Labidognath carriers are wide, with the anterior end almost as wide as the posterior end of MxI , narrowing towards the posterior end, never longer than the remaining jaw apparatus, covered dorsally by a thin muscle layer, and attached to MxI.
24. MxI: 0 , dentate; 1 , fang shaped.
25. Fang shaped MxI: 0, facing each other; 1, facing dorsal side of the body.
26. Shape of MxIII: 0, dentate, front end part of distal arc with left MxIV (Fig. 5A-G); 1, dentate, at least in part located behind MxII (Fig. 5B); 2, edentate, behind MxII (Fig. 5D, E); 3, dentate, triangular; 4, dentate, with teeth arranged anterior to posterior, most anterior longest. Both Orensanz (1990) and Fauchald (1992) suggested that the shape of MxIII and its position in relation to other maxillae could be taxonomically informative.
27. Left MxIV: 0, wider than long with teeth on less than half of the plate, often only two teeth present (Fig. 5A); 1, wider than long with teeth on more than half of the plate, always more than two teeth present (Fig. 5B, C, F); 2, longer than wide often with just one tooth (Fig. 5D, E); 3, triangular, edentate, or with few teeth at the tip; 4, teeth decreasing in size from anterior to posterior, anteriormost tooth longest. The width and length of MxIV are defined here, respectively, as the axis in which it is attached to the body and the axis that it projects from the attachment into the lumen of the pharyngeal bulb.
28. MxV: 0, fang shaped; 1, plate like, a small tooth may be present.
29. MxVI: 0, absent; 1, present (Fig. 5A).
30. Right MxIII: 0, present; 1, absent.
31. Relative length of labidognath maxillary carrier: 0 , longer than half the length of MxI; 1, shorter than half the length of MxI.
32. Mandibles: 0, absent; 1, flat (Fig. 6A); 2, strongly curved, forming an open scoop, plates fused anteriorly,
anterior end strongly calcified (Fig. 6B, C); 3, curved, plates separated in a V shape at the anterior end, not strongly calcified (Fig. 6D, E). Although the mandibles of both Lysidice and Palola are curved, we prefer to consider them as different character states, because of the obvious differences between them, instead of assuming that the presence of the curved state is a homology. In addition to the differences described above, in Palola the strongly calcified anterior end is present beyond the organic matrix, whereas in Lysidice calcification closely follows the matrix.
33. Mandibles: 0, fused medially; 1, attached by ligaments.
34. Placement of the muscle fibre complex F1 + F2 (Desière, 1967) on the pharyngeal bulb: 0, absent; 1, posterior to mandible carriers (Fig. 7A); 2, above mandible carriers (Fig. 7B); 3, between mandible carriers (Fig. 7C).
35. Notopodia: 0, absent; 1, reduced to notopodial cirri; 2, complete chaetal lobe.
36. Neuropodia in mid-body distally: 0 , round; 1 , truncate; 2, pointed.
37. Neuropodial prechaetal lobes in mid-body: 0, absent; 1 , longer than acicular lobes; 2 , about as long as acicular lobes.
38. Neuropodial postchaetal lobes in mid-body: 0, longer than acicular lobes; 1 , about as long as acicular lobes; 2 , shorter than acicular lobes.
39. Ventral cirri: 0 , absent; 1 , tapering or digitiform in first few chaetigers, thereafter basally inflated; 2 , tapering or digitiform in anterior and posterior chaetigers, basally inflated in median chaetigers; 3, digitiform, bases not inflated anywhere along the body; 4, tapering, bases not inflated anywhere along the body. 40. Shape of the inflated region of ventral cirri: 0 , ventral ridge; 1 , scoop shaped, rounded.
41. Shape of the branchiae: 0 , absent; 1 , single filaments; 2, filaments linearly arranged; 3 , spiraled; 4 , bushy. In both pectinate and palmate branchiae filaments are linearly arranged on a stem. The difference between these shapes is in the relative length between the stem and filaments (Fauchald, 1992); this may be difficult to distinguish as an almost continuous variation between extremes may be present; consequently, character state 2 unites both pectinate and palmate branchiae.
42. Branchiae start: 0 , on chaetiger 3 ; 1 , between chaetigers 4 and 9; 2, start late (after chaetiger 10).
43. Branchial distribution: 0, mostly limited to the first third of the body (present on less than $55 \%$ of the body segments); 1 , in most of the body (present on more than $65 \%$ of the body); 2 , mostly limited to the second third of the body (present on less than $55 \%$ of the body segments). Percentages refer to number of chaetigers in which branchiae are continuously present in relation to the total number of chaetigers in the body.


Figure 5. Maxillae. A, Eunice denticulata Webster, 1884 dorsal view. B, C, Eunice rubra Grube, 1856 dorsal view, and detail of maxillae IV (MxIV) and MxV. D, E, Palola brasiliensis Zanol, Paiva \& Attolini, 2000 dorsal view, and detail of MxIII, MxIV, and MxV. F, G, Lysidice ninetta Audouin \& Milne Edwards, 1833 detail of MxIV and MxV, and dorsal view. MxIII1, front end of MxIII as part of the distal arc; MxIII2, MxIII at least in part located behind MxII; MxIII3, MxIII edentate behind MxII; MxIV1, MxIV with teeth in less than half of the plate; MxIV2, MxIV with teeth in more than half of the plate; MxIV3, MxIV longer than wide, most of the time just with one tooth; MxVI, maxillae VI; MxC1, maxillary carrier greater than half of MxI; MxC2, maxillary carrier smaller than half of MxI.
44. Narrow, long pectinate chaetae: 0 , absent; 1, present.
45. Spatula-shaped pectinate chaetae (short, wider than acicula with wide teeth): 0 , absent; 1, present (Fig. 8A, B). When present these are usually found in the middle and posterior regions of the body.
46. Pseudocompound falcigers or spinigers: 0, absent; 1, present in a limited number of anterior chaetigers.
47. Distal end of compound falcigers (present in most of the body): 0, absent; 1, bidentate; 2, multidentate.
48. Distal ends of compound falcigers in the $2.5 \%$ posteriormost chaetigers: 0 , absent; 1 , all bidentate; 2 , biand tridentate. Percentage refers to total number of chaetigers present.
49. Compound spinigers: 0 , absent; 1 , present.
50. Calcareous chaetae: 0 , absent; 1 , present.


Figure 6. Mandibles. A, Eunice rubra Grube, 1856 ventral view. B, C, Palola brasiliensis Zanol, Paiva \& Attolini, 2000 ventral, and dorsal views. D, E, Lysidice ninetta Audouin \& Milne Edwards, 1833 ventral, and dorsal views. MI, muscle insertion; OM, outline of the organic matrix. Scale bars $=1 \mathrm{~mm}$.


Figure 7. Pharyngeal bulb musculature, ventral view. A, Eunice cf. torquata Quatrefages, 1866. B, Palola brasiliensis Zanol, Paiva \& Attolini, 2000. C, Lysidice ninetta Audouin \& Milne Edwards, 1833. F1 + F2, muscle fibre complex F1 + F2 (Desière, 1967); MND, mandibles; MS, muscle; Mx, maxillae.
51. Color of neuropodial acicula: 0, dark (brown, dark brown or black); 1 , light (nearly clear or yellow).
52. Color pattern of neuropodial acicula: 0 , sheath and core with distinctly different colours; 1, sheath and core with similar colour.
53. Distal ends of neuropodial acicula: 0, blunt-tipped, conical or irregularly knobbed; 1 , hammer-headed or bidentate.
54. Distal ends of subacicular hooks: 0 , absent; 1 , falcate; 2 , bidentate; 3 , tridentate; 4 , spinigererous. Subacicular hooks are the ventralmost chaetae, usually not parallel to the remaining chaetae, but emerging at a more or less distinct angle to them, and inserted in the parapodium in front of the chaetal bundle.
55. Number of subacicular hooks per parapodium: 0 , always single, except for replacements; 1, at least paired. The presence of multiple subacicular hooks was not included as a separate state because the information we have in the present analysis is insufficient to distinguish them consistently; however, distinguishing paired from multiple subacicular hooks may be informative in a larger group of species. Some of the species with multiple subacicular hooks have them arranged in a fan; others have them in irregular bundles. This arrangement may also be an informative character and will be considered in a future analysis. 56. Color of subacicular hooks: 0, light (nearly clear or yellow); 1, dark (brown, dark brown, or black).


Figure 8. Marphysa cf. atlantica Kinberg, 1865. A, spatula-shaped pectinate chaetae. B, parapodium of chaetiger 109, left side. SPC, spatula-shaped pectinate chaetae.
57. Color pattern of subacicular hooks: 0 , sheath and core with distinctly different colours; 1 , sheath and core with similar colour.
58. Distribution of subacicular hooks: 0, present in every, or nearly every segment after first occurrence; 1 , may be missing irregularly, sometimes in many segments.
59. Ventral pygidial cirri: 0 , absent; 1, present.

Most polymorphic characters are soft-body features. This variability may be a result of size differences among specimens examined, but may also be caused by different fixation procedures or the condition of the preserved specimens (Treadwell, 1911; Hartman, 1944; Nonato \& Luna, 1970; Steiner et al., 2002). Polymorphism in characters like 53 and 54 are caused by variation along the body of the specimens.

## RESULTS

We concentrate on the results of the analyses of composite characters because these analyses yielded a more resolved strict consensus tree; the major differences between the results of the two matrices are caused by the ambiguous placement of Palola and Onuphidae.

## COMPOSITE ANALYSES

Unconstrained parsimony analyses yielded 200 most parsimonious trees (MPT) (length $=172$ steps; consistency index $=0.57$; retention index $=0.74$ ). The Eunicidae/Onuphidae clade and Onuphidae were both monophyletic, but Onuphidae was deeply nested within Eunicidae, making the later paraphyletic (Figs 9, 10A, B). The Eunicidae/Onuphidae clade (Fig. 9, clade 6) excluding Palola had low support, and a monophyletic Eunicidae was present in trees just one step longer than the unconstrained MPT. All putative exclusive synapomorphies for Eunicidae, such as dorsal lips separated by a narrow notch, and short ring-shaped ceratophores and palpophores, supported the whole Eunicidae/Onuphidae clade in unconstrained MPT (Fig. 9, clade 7) and could not be unambiguously optimized on trees constraining Eunicidae to be monophyletic. When we constrained Eunicidae as monophyletic, the strict consensus of the MPT showed almost no resolution within the family. In this constrained analysis the only eunicid clades present were the ones with bootstrap support higher than $50 \%$ in the unconstrained analysis, and the clade (Eunice norvegica (Linnaeus, 1767) (Eunice aphroditois (Pallas, 1788) (Eunice denticulata Webster, 1884; Eunice cf. flavopicta Izuka, 1912))).

Onuphidae was sister to Eunice websteri Fauchald, 1969 in most MPT and to clade 1 (Fig. 9) in the remaining trees. The clade in which Onuphidae is nested in most MPT (Fig. 10A, B) was supported by homoplastic characters; many of these are polymorphic in several species and may be inapplicable in Onuphidae. However, the clade ((Onuphidae, E. websteri) Eunice vittata (Chiaje, 1829)) is supported by having peristomial ventrolateral lips set off by distinct grooves and by the presence of at least two subacicular hooks per parapodium: characters that are generally present in Onuphidae.

The monophyly of Eunice and most of its taxonomically informal groups was rejected in the present results: they were paraphyletic in all MPT (Figs 9, 10A, B). The group bearing dark unidentate hooks (group D), represented by Eunice fucata Ehlers, 1887 and Eunice sebastiani Nonato, 1965, was the only informal subgroup monophyletic in some unconstrained MPT. The monophyly of all other informal groupings required the trees to be between three and eight steps longer than the MPT (Table 5). The dark coloured subacicular hooks had a unique origin, but the Eunice informal subgroup based on this character was paraphyletic because the genera Marphysa and Lysidice were nested within the same clade (Fig. 9, clade 4); species of these two genera also have dark coloured subacicular hooks, although some species may be polymorphic for this feature.


Figure 9. Strict consensus tree of 200 most parsimonious trees resulting from composite analyses. Tree statistics: length $=200$ steps; consistency index $=0.49$; retention index $=0.65$. Characters optimized on nodes under ACCTRAN, unambiguous characters in bold face and underlined. Characters used for the first time in this study and promising sources of phylogenetic signal are shaded in grey. White and black hashmarks represent homoplasious and non-homoplasious characters, respectively. Bold numerals below branches are Bremer support/Bootstrap values; single numerals refer to Bremer support. Numerals beside square brackets are clade numbers. *Eunicidae outgroup species; **Onuphidae species.


Figure 10. Consensuses of 200 most parsimonious trees resulting from composite analyses. A, $50 \%$ majority rule consensus, numerals below branches represent the percentage of the most parsimonious trees in which the clade is present. B, Adams consensus. *Eunicidae outgroup species; **Onuphidae species.

Two clades of Eunice species were present in all MPT. In one clade, Eunice antennata (Lamarck, 1818)/ Eunice rubra Grube, 1856/Eunice miurai CarreraParra \& Salazar-Vallejo, 1998, all species have yellow tridentate hooks, and compound falcigers of posteriormost chaetigers are bi- and tridentate. This clade was also present in all strict consensus trees resulting from constrained analyses in which its members had not been constrained to different clades. The other clade, clade 5 (Fig. 9), groups species in which MxIII is part of a distal arc with left MxIV and is placed side by side with right MxIV. The Eunice species of clade 5 were grouped in a similar clade even when Marphysa and Lysidice species were forced out of it, in the constrained analyses forcing Eunice to be monophyletic. $E$. aphroditois is the sister taxon to ( $E$. denticulata, $E$. cf. flavopicta) in all MPT of unconstrained and constrained analyses.

The basal polytomy in the clade Eunicidae/ Onuphidae in the strict consensus (Fig. 9) resulted from the 'wild-card' (Nixon \& Wheeler, 1992) behav-

Table 5. Summary of results of constrained analyses (see Table 2 for descriptions of groups). Unconstrained analyses resulted in 200 most parsimonious trees, length $=172$ steps. A, number of most parsimonious trees; B, difference in length (steps) from unconstrained most parsimonious trees

| Constrained clades | A | B |
| :--- | ---: | ---: |
| Eunicidae | 804 | +1 |
| Eunice | 24 | +6 |
| Eunice, yellow hooks | 962 | +5 |
| Eunice, dark hooks | 6 | +3 |
| A group | 163 | +7 |
| B group | 192 | +3 |
| C group | 100 | +3 |
| D group | 8 | $\pm 0$ |
| A1 group | 1170 | +3 |
| B2 group | 1560 | +5 |
| C1 group | 2 | +8 |
| C2 group |  | +3 |

iour in the MPT of Eunice cf. semisegregata Fauchald, 1969; Eunice pennata (Müller, 1776), Eunice cf. atlantica Kinberg, 1865, and clade 1, which have conflicting placement among the MPT because they lack synapomorphies to resolve their relationships either with each other or with other species.
Most of the MPT did not support the monophyly of Marphysa based on the traditional characters, i.e. the absence of peristomial cirri, combined with the presence of five prostomial appendages. The two Marphysa sister species were grouped based on the flattened shape of the body and the presence of spatula-shaped pectinate chaetae; characters not widely or clearly described in previous studies.

The genus Palola is the basalmost clade in the Eunicidae/Onuphidae clade because of its lack of subacicular hooks and pectinate chaetae, and because of the presence of maxillary carriers longer than half of MxI. Palola is the only eunicid genus included in the analyses not nested within Eunice.
see Figure 11 for consensus trees. A monophyletic Eunicidae with Onuphidae as its sister group is supported in some MPT (Fig. 11B). In the MPT in which Eunicidae is paraphyletic, Onuphidae has the same sister-group relationships as in the composite analyses. Palola is the basalmost eunicid when the Eunicidae is paraphyletic, and is nested within Eunice when Eunicidae is monophyletic. Palola is sister to Lysidice or E. cf. atlantica when nested within Eunice. Clade 5 (Fig. 9) of the composite analyses collapses in the strict consensus of the reductive analyses (Fig. 11A) because of the uncertain placement of Palola. When Palola is removed from the reductive analyses clade 5 (Fig. 9) is present in the strict consensus tree.

## DISCUSSION

The discussion concentrates on the results of the strict consensus tree of the composite analyses, and character numbers refer to composite coding.

## Reductive analyses

Parsimony analyses yielded 858 MPT (length $=177$ steps; consistency index $=0.56$; retention index $=0.73$ ),

## PHYLOGENETIC RELATIONSHIPS AND TAXONOMY

Eunice and its informal taxonomic groups were found to be paraphyletic in this study, supporting results


Figure 11. Consensuses of 858 most parsimonious trees resulting from reductive analyses. A, Strict consensus, numerals below branches are Bremer support/Bootstrap values; single numerals refer to Bremer support. B, $50 \%$ majority rule consensus, numerals below branches represent the percentage of the most parsimonious trees in which the clade is present. C, Adams consensus. *Eunicidae outgroup species; **Onuphidae species.
from previous phylogenetic analyses that included Eunice species (e.g. Struck et al., 2006). A consistent characteristic revealed for the first time in all trees resulting from the present analyses was the placement of E. aphroditois, type species of Eunice s.s., and $E$. antennata, type species of the now defunct Leodice, in different clades (Figs 9, 11). This implies that as currently defined the genus Eunice could be split into at least two monophyletic groups, one representing Eunice s.s. and the other Leodice, each with its original type species. However, analyses with more resolved results are necessary before one can justify dividing Eunice. In the current results the species composition of each of these monophyletic groups, their characterizing synapomorphies, and the monophyletic group placement of most of the included Eunice species cannot be established with confidence because of the lack of basal resolution and the low support of clades.

The ambiguous placement of species with yellow bidentate hooks and the unique origin of black bidentate hooks in the present results contrasted with the results in Fauchald (1992), in which the yellow-hook clade was the only consistent one in all trees, and species with black hooks had conflicting arrangements among all trees. The analyses in Fauchald (1992) included a broader taxonomic sample than the present one. However, his results were flawed in that the analyses did not find all MPT and did not include characters that can be shown to be important in resolving the present trees (e.g. characters 26, 29, 48, and 59).

Marphysa and Lysidice were grouped with Eunice cariboea Grube, 1856 within Eunice s.l. (Fig. 9, clade 3). The synapomorphies supporting this clade are either reversals to the Eunicidae/Onuphidae plesiomorphic states (characters 29 and 59) or reversals within this clade (character 19). If descriptions in the literature are accurate, the absence of MxVI (character 29) and the presence of the ventral pygidial cirri (character 59) may be more variable within clade 3 (Fig. 9) than is represented by our taxa sample. Orensanz (1975) described Marphysa as having either only dorsal or both dorsal and ventral pygidial cirri. Furthermore, both Aiyar (1931) and Treadwell (1921) mentioned the presence of MxVI in Marphysa and Lysidice. The synapomorphies corroborating this clade are likely to have been produced by the lack of understanding and under-representation of the variability described above, and/or by error induced by paedomorphic evolution (e.g. Wiens, Bonnet \& Chippindale, 2005; T. Struck, pers. comm.) as all the species in this clade have paedomorphic features. Juvenile traits in Eunice include absence of palps, as in Lysidice, absence of peristomial cirri, as in Marphysa and Lysidice, and branchiae either absent or
poorly developed, as in E. cariboea and Lysidice (e.g. Giangrande, 1989; Lu \& Fauchald, 1998; Nogueira, Steiner \& Amaral, 2001).

The paraphyletic Eunicidae present in all MPT yielded by the composite analyses, but present in just some of the MPT from the reductive analyses, makes the idea of a monophyletic Eunicidae even more problematic. The molecular phylogenies of the order Eunicida are inconsistent on this issue (Struck et al., 2002, 2006); however, the taxa sample of Onuphidae and Eunicidae in that study was not large enough to address the relationships either within or among them adequately. In the present phylogeny Onuphidae is always sister to species with yellow hooks. In contrast, in the 18 S rDNA phylogeny of the order Eunicida (Struck et al., 2006) Onuphidae is sister to a clade equivalent to the present dark-coloured subacicular hook one (Fig. 9, clade 4).

## Character evolution

The low resolution among Eunice species at the base of the tree was caused in part to the fact that many of the characters commonly used in Eunice taxonomy are homoplasies. Among others, these characters include the shape of articulation of prostomial appendages, and the branchial shape and distribution. Most exclusive synapomorphies within clade 6 (Fig. 9), other than the dark colour of the subacicular hooks, were characters used for the first time in this study, such as the jaw characters (e.g. characters 26, 27, and 29) and characters derived from close examination of chaetal variation along the body (e.g. characters 45 and 48). Both types of characters, as well as other features of the buccal apparatus, appear to be promising sources of phylogenetic signal. To date, however, they have been poorly documented and their variation has not been properly taken into account.

Miura's (1986) hypothesis of branchial distribution evolution and the evolution of the dentition of subaciular hooks could not be tested by our results. The basalmost polytomy in clade 6 (Fig. 9) biased the plesiomorphic state. However, it is clear that unidentate subacicular hooks were derived from bidentate ones (Fig. 12A) as suggested by Orensanz (1990). Both species that bear unidentate hooks, E.fucata and E. sebastiani, are nested within clade 5 (Fig. 9) in which the plesiomorphic state of subacicular hooks is bidentate (Fig. 12A).

The dark colour of subacicular hooks was derived once from the plesiomorphic light-colour condition and reversed to this condition twice independently (Fig. 12B). The reversals occurred in L. ninetta and Marphysa sanguinea (Montagu, 1807), which showed polymorphism for this character. The colour variation and pattern of colours of subacicular hooks
are not well understood, but they may vary with size of the specimen and with the position along the body of the segment examined (Day, 1967; Fauchald, 1992).

The colour and the dentition of subacicular hooks are considered important features in the taxonomy of the family Eunicidae, mainly in the genus Eunice (e.g. Hartman, 1944; Fauchald, 1992). Miura (1986) considered them conservative enough to suggest that they could be used to divide the genus Eunice in groups, perhaps corresponding to subgeneric- or generic-level taxa. However, based on the present results, these groups are paraphyletic and most similarities in hook colour and dentition are homoplasies.

## CONCLUSION

Eunice s.l. and its informal subgroups are paraphyletic. Eunice may consist of at least two monophyletic
groups, Eunice s.s. and Leodice. However, the diagnostic features of these groups, their species composition, and the monophyletic group placement of most other species of Eunice remains unresolved. This study provides the first analytical hypotheses of phylogeny for Eunice, and it is a step towards a better understanding of the phylogeny of Eunice and also of Eunicidae.

Additional information is needed in order to obtain a more stable hypothesis of phylogeny, and future analyses should focus on the whole family Eunicidae and include a wider sample of Onuphidae species. Besides the inclusion of more species in the analyses, other characters that might have phylogenetic importance should be added and molecular data considered. Some sources of probable informative morphological characters may be found in the ontogeny (Åkesson, 1967), internal anatomy (Müller, 2006), and electronic microscopy (e.g. Hayashi \& Yamane, 1994).


Figure 12. Evolution of subacicular hook. A, B, dentition (character 54) and colour (character 56). Characters optimized on the strict consensus of most parsimonious trees resulting from composite analyses, both characters are unambiguously optimized. *Eunicidae outgroup species; **Onuphidae species.

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## APPENDIX 1

## REDUCTIVE CHARACTERS: UNINFORMATIVE CHARACTERS UNDERLINED

1. Shape of cross section of body: 0 , circular; 1 , dorsoventrally flattened; 2 , ventrally convex.
2. Body shape: 0 , cylindrical, abruptly tapering anteriorly and posteriorly; 1 , evenly tapering from ante-rior-median chaetigers.
3. Relative width and length of chaetigers: 0 , at most ten times wider than long; 1, more than ten times wider than long.
4. Posterior extension of prostomium (caruncle): 0, absent; 1, present.
5. Frontal lips (sensu Paxton, 1998): 0, separated frontally by a narrow notch (Fig. 1A-C); 1, separated frontally by a distinct space (Fig. 1D); 2, frontal edge entire. 6. Frontal lips separated by a narrow notch anteriorly: 0, round (Fig. 1A, B); 1, truncate (Fig. 1C).
6. Frontal lips separated by a narrow notch, dorsally: 0, inflated (Fig. 2A); 1, flat (Fig. 2B); 2, dimpled (Fig. 2C).
7. Arrangement of prostomial appendages: 0, antennae and palps evenly spaced (Fig. 1C, D); 1, median antenna isolated by a gap from the other antennae and palps (Fig. 1A); 2, palps isolated by a gap from the antennae (Fig. 1B); 3, head appendages on conspicuously different regions of the prostomium, median antennae on the caruncle.
8. Median antennae: 0 , absent; 1 , present.
9. Antennal styles: 0 , digitiform, tapering, or clavate (Fig. 3A-C); 1, medially inflated (fusiform) (Fig. 3D); 2, button shaped (Fig. 3E).
10. Articulations of antennal styles: 0 , absent, 1, present.
11. Shape of articulations of antennal styles: 0 , short or long cylinders (Fig. 4A); 1, moniliform (Fig. 4B).
12. Antennal ceratophore: 0 , absent; 1, present.
13. Shape of antennal ceratophore: 0 , short, usually ring shaped; 1 , relatively long, usually articulated.
14. Lateral antennae: 0 , absent; 1 , present.
15. Palpal styles: 0 , absent; 1 , digitiform, tapering, or clavate (Fig. 3A-C).
16. Palpal styles articulations: 0 , absent; 1 , present.
17. Shape of palpal styles articulations: 0, short or long cylinders (Fig. 4A); 1, moniliform (Fig. 4B).
18. Palpophores: 0, absent; 1, present.
19. Shape of palpophores: 0, short, usually ring shaped; 1 , relatively long, usually articulated.
20. Peristomial ventrolateral lips: 0 , absent; 1 , present.
21. Shape of peristomial ventrolateral lips: 0 , set off by distinct grooves; 1, visible only as elevated surfaces; 2 , scoop shaped; 3 , medially short, about half as long as lateral side of the peristomium.
22. Peristomium: 0 , a single ring; 1 , two rings distinct at least dorsally; 2, reduced to region around the mouth.
23. Peristomial cirri: 0 , absent; 1 , present.
24. Mouth: 0 , extending to the ventral side of anterior chaetigers; 1 , restricted to peristomium.
25. Maxillary plates: 0 - absent; 1- present.
26. Number of rows of maxillary plates: 0 , one pair; 1 , two or more pairs.
27. Calcium carbonate mineralizing jaws: 0, absent; 1, present.
28. Calcium carbonate mineralizing jaws: 0 , calcite; 1 , aragonite.
29. Maxillary carrier: 0 , ctenognath; 1, prionognath; 2, labidognath.
30. MxI: 0 , dentate; 1 , fang shaped.
31. Fang-shaped MxI: 0 , facing each other; 1, facing dorsal side of the body.
32. Shape of MxIII: 0 , dentate, front end part of distal arc with left MxIV (Fig. 5A, G); 1, dentate, at least in part located behind MxII (Fig. 5B); 2, edentate, behind MxII (Fig. 5D, E); 3, dentate, triangular; 4, dentate, with teeth arranged anterior to posterior, most anterior longest.
33. Left MxIV: 0 , wider than long with teeth on less than half of the plate, often only two teeth present (Fig. 5A); 1, wider than long with teeth on more than half of the plate, always more than two teeth present (Fig. 5B, C, F); 2, longer than wide often with just one tooth (Fig. 5D, E); 3, triangular, edentate, or with few teeth at the tip; 4, teeth decreasing in size from anterior to posterior, anteriormost tooth longest.
34. MxV: 0 , fang shaped; 1, plate like, a small tooth may be present.
35. MxVI: 0, absent; 1, present (Fig. 5A).
36. Right MxIII: 0, present; 1, absent.
37. Relative length of labidognath maxillary carrier: 0 , longer than half the length of MxI; 1 , shorter than half the length of MxI.
38. Mandibles: 0 , absent; 1 , present.
39. Shape of mandibles: 0, flat (Fig. 6A); 1, strongly curved, forming an open scoop, plates fused anteriorly, anterior end strongly calcified (Fig. 6B, C); 2, curved, plates separated in a V shape at the anterior end, not strongly calcified (Fig. 6D, E).
40. Mandibles: 0, fused medially; 1, attached by ligaments.
41. Muscle fibre complex F1 + F2 (Desière, 1967) on the pharyngeal bulb: 0 , absent; 1 , present.
42. Placement of the muscle fibre complex F1 + F2 (Desière, 1967) on the pharyngeal bulb: 0 , posterior to mandible carriers (Fig. 7A); 1, above mandible carriers (Fig. 7B); 2, between mandible carriers (Fig. 7C).
43. Notopodia: 0, absent; 1, present.
44. Notopodia: 0 , reduced to notopodial cirri; 1 , complete chaetal lobe.
45. Neuropodia in mid-body distally: 0 , round; 1 , truncate; 2 , pointed.
46. Neuropodial prechaetal lobes in mid-body: 0, longer than acicular; 1 , about as long as acicular lobes; 2, lobes absent.
47. Neuropodial postchaetal lobes in mid-body: 0, longer than acicular lobes; 1 , about as long as acicular lobes; 2 , shorter than acicular lobes.
48. Ventral cirri: 0 , absent; 1 , present.
49. Shape of ventral cirri: 0 , tapering or digitiform in first few chaetigers, thereafter basally inflated; 1 , tapering or digitiform in anterior and posterior chaetigers, basally inflated in median chaetigers; 2 , digiti-
form, bases not inflated anywhere along the body; 3, tapering, bases not inflated anywhere along the body. 51. Shape of the inflated region of ventral cirri: 0 , ventral ridge; 1, scoop shaped, rounded.
50. Branchiae: 0 , absent; 1 , present.
51. Shape of the branchiae: 0 , single filaments; 1 , filaments linearly arranged; 2 , spiraled; 3 , bushy.
52. Branchiae start: 0 , on chaetiger $3 ; 1$, between chaetigers 4 and $9 ; 2$, start late (after chaetiger 10).
53. Branchial distribution: 0, mostly limited to the first third of the body (present on less than $55 \%$ of the body segments); 1 , in most of the body (present on more than $65 \%$ of the body); 2 , mostly limited to the second third of the body (present on less than $55 \%$ of the body segments).
54. Narrow, long pectinate chaetae: 0 , absent; 1 , present.
55. Spatula-shaped pectinate chaetae (short, wider than acicula with wide teeth): 0 , absent; 1 , present (Fig. 8A, B).
56. Pseudocompound falcigers or spinigers: 0 , absent; 1 , present in a limited number of anterior chaetigers. 59. Compound falcigers: 0 , absent; 1 , present.
57. Distal end of compound falcigers (present in most of the body): 0 , bidentate; 1 , multidentate.
58. Compound falcigers in the $2.5 \%$ posteriormost chaetigers: 0 , absent; 1 , present.
59. Distal end of compound falcigers in the $2.5 \%$ posteriormost chaetigers: 0 , all bidentate; 1 , bi- and tridentate.
60. Compound spinigers: 0 , absent; 1 , present.
61. Calcareous chaetae: 0 , absent; 1 , present.
62. Color of neuropodial acicula: 0 , dark (brown, dark brown, or black); 1 , light (nearly clear or yellow).
63. Color pattern of neuropodial acicula: 0 , sheath and core with distinctly different colours; 1 , sheath and core with similar colour.
64. Distal end of neuropodial acicula: 0 , blunt-tipped, conical, or irregularly knobbed; 1 , hammer-headed or bidentate.
65. Subacicular hooks: 0 , absent; 1 , present.
66. Distal end of subacicular hooks: 0 , falcate; 1 , bidentate; 2 , tridentate; 3 , spinigererous.
67. Number of subacicular hooks per parapodium: 0 , always single, except for replacements; 1 , at least paired.
68. Color of subacicular hooks: 0 , light (nearly clear or yellow); 1, dark (brown, dark brown, or black).
69. Color pattern of subacicular hooks: 0 , sheath and core with distinctly different colours; 1 , sheath and core with similar colour.
70. Distribution of subacicular hooks: 0 , present in every, or nearly every, segment after first occurrence; 1 , may be missing irregularly, sometimes in many segments.
71. Ventral pygidial cirri: 0 , absent; 1 , present.
APPENDIX 2
REDUCTIVE CODING CHARACTER MATRIX

|  | 1234567891 | 1111111112 | 2222222223 | 3333333334 | 4444444445 | 5555555556 | 6666666667 | 7777 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 0 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234 |
| Amphinome rostrata | 0001?? ?311 | 100-11100- | 0-2000---- | --------0- | -0-1101113 | -13010000- | 0-01---0-- | ---0 |
| Oenone fulgida | 01002---12 | 0-0-10---- | 12001100-1 | 0-44000-10 | 112100000- | -0---0000- | 0-00110111 | 0101 |
|  |  |  |  |  |  |  | 3 |  |
| Lumbrineris fragilis | 00002---0- | ----00---- | 1210110102 | 1133100010 | 0100-0000- | -0---0000- | 0-000000-- | ---1 |
| Lumbrineris latreille | 01002---0- | ----00---- | 1110110102 | 1133100010 | 0100-0100- | -0---00011 | 0-001100-- | --1 |
| Dorvillea sociabilis | 01002---00 | 110-11100- | 13101110-0 | ?-? ? ? ? ?-10 | 10-1010012 | -0---00010 | 10001100-- | ---1 |
|  | 2 |  |  |  |  |  |  |  |
| Diopatra tridentata | 00001--010 | 1011111011 | 1001110112 | 1011101110 | 11010?2011 | 122101010- | 0-00110111 | 0101 |
| Kinbergonuphis tenuis | 21001--110 | 1011111011 | 1001110112 | 1011101110 | 11010?2011 | 111111010- | 0-00110111 | 0101 |
| Lysidice ninetta | 0000001-11 | 101010---- | 1010110112 | 1001101012 | 1121022111 | 10---10010 | 1000000110 | 0001 |
|  | 1 |  | 1 |  |  |  | 11 | 11 |
| Palola brasiliensis | 0000001210 | 1010111010 | 1011110112 | 1022101011 | 1111022211 | 0102200010 | 10000100-- | ---1 |
|  | 112 |  | 1 |  |  |  |  |  |
| Palola cf. viridis | 0000002010 | 1010111010 | 1111110112 | 1022101011 | 1111022211 | 0102200010 | 10000100-- | ---1 |
| Marphysa sp. | 0100000110 | 1010111010 | 1010110112 | 1001101110 | 1101012211 | 1112110010 | 1000000110 | 1001 |
|  | 1 |  | 1 |  |  |  |  |  |
| Marphysa sanguinea | 1000001110 | 1010111010 | 1110110112 | 1001101110 | 1101012210 | 111211100- | 0-10000110 | 0001 |
|  | 1 |  |  |  |  |  |  | 11 |
| Marphysa cf. atlantica | 1010001110 | 1010111010 | 1110110112 | 10011?1110 | 1101002210 | 1112111010 | 1000000110 | 1001 |
| Eunice aphroditois | 0100002110 | 0010111010 | 1011110112 | 1000111110 | 1101012210 | 1111110010 | 1000000110 | 1000 |
|  | 11 | 1 |  |  | 1 |  | 1 |  |
| Eunice denticulata | 0100000110 | 1010111010 | 1011110112 | 1000111110 | 1101011110 | 0112110010 | 1000000110 | 1001 |
|  | 2 |  | 1 |  | 2221 |  | 1 |  |
| Eunice cf. flavopicta | 0100002110 | 1010111010 | 1111110112 | 1000111110 | 1101012110 | 0111110010 | 1000000110 | 1000 |
| Eunice norvegica | 0100002110 | 1010111010 | 1111110112 | 1001111110 | 1101012211 | 1111110010 | 1000000110 | $\begin{array}{r} 1000 \\ 1 \end{array}$ |
|  | 1 |  |  |  |  |  |  |  |
| Eunice cariboea | 0000000110 | 1010111010 | $\begin{aligned} & 1011110112 \\ & 1 \end{aligned}$ | $1001101110$ | $1101002211$ | $\text { 10-- - } 10010$ | $1000000110$ | 1000 |
|  | 11 |  |  |  |  |  |  | 1 |
|  | 2 |  |  |  |  |  |  |  |

APPENDIX 2 Continued

|  | 1234567891 | 111111112 | 2222222223 | 3333333334 | 4444444445 | 5555555556 | 666666667 | 7777 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 0 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234 |
| Eunice frauenfeldi | 0100001010 | 1010111010 | 1011110112 | 1001111110 | 1101012211 | 1112110010 | 1000000110 | 1000 |
| Eunice cf. grubei | 010000?01? | 1010111010 | 1111110112 | 1001111110 | 1101022211 | 1110110010 | 1000010110 | 1000 |
| Eunice thomasiana | 0100001010 | 1110111110 | 1011110112 | 1001111110 | 1101011111 | 1111110010 | 1000010110 | 1000 |
| Eunice dubitata | 0100001010 | 1010111010 | 1111110112 | 1001111110 | 1101011111 | 1110010010 | 1000010110 | 1000 |
| Eunice cf. torquata | $\begin{aligned} & 0000001010 \\ & 1 \end{aligned}$ | 1110111110 | 1111110112 | 1001111110 | 1101011111 | 1110110010 | 1000010110 | 1000 |
| Eunice fucata | $\begin{gathered} 0100001010 \\ 2 \end{gathered}$ | 1010111010 | $\begin{aligned} & 1011110112 \\ & 1 \end{aligned}$ | 1001111110 | 1101012211 | 1111010010 | 1000010100 | 1100 |
| Eunice sebastiani | 0100001110 | 1?1011??10 | 1111110112 | 1001111110 | 1101011110 | 1111110010 | 1000010101 | 110? |
| Eunice harassii | 0100001010 | 1010111010 | 1011110112 | 1001111110 | 1101011111 | 1111110010 | 1000110110 | 0100 |
|  | 12 |  | 1 |  | 22 |  |  |  |
| Eunice antennata | 0100001010 | 1110111110 | 1011110112 | 1011101110 | 1101012011 | 1111110010 | 1100110120 | 0101 |
|  | 1 |  | 1 |  | 2 |  |  |  |
| Eunice rubra | 0100000010 | 1110111110 | 1011110112 | 1011101110 | 1101011111 | 1111110010 | 1100110120 | 0101 |
|  | 111 |  |  |  | 22 |  | 1 |  |
|  | 2 |  |  |  |  |  |  |  |
| Eunice miurai | 0100001010 | 1110111110 | 1011110112 | 1011101110 | 1101011111 | 1111010010 | 1100110120 | 0100 |
|  |  |  |  |  |  |  | 1 |  |
| Eunice websteri | 0000012010 | 1010111010 | 1011110112 | 1011101110 | 1101012211 | 1110010010 | 1000110111 | 0101 |
|  | 11 |  |  |  |  |  |  |  |
| Eunice vittata | 0100012010 | 1010111010 | 1011110112 | 1011101110 | 1101012211 | 1110010010 | 1000110121 | 0101 |
| Eunice atlantica | 0100010010 | 1010111010 | 1111110112 | 1011101110 | 1101012211 | 1110010010 | 1000110120 | 0101 |
|  | 2 |  |  |  |  |  |  |  |
| Eunice unifrons | 0100012010 | 1010111010 | 1011110112 | 1011101110 | 1101012211 | 1110010010 | 1000110120 | 0101 |
|  | 1 |  | 1 |  |  |  |  |  |
| Eunice pennata | 0100000010 | 1010111010 | 1111110112 | 1011101110 | 1101012211 | 1110010010 | 1000110110 | 0101 |
| Eunice cf. semisegregata | 0100001010 | 1010111010 | 1111110112 | 1011111110 | 1101011110 | 1110010010 | 1000110110 | 0101 |
|  | 1 |  |  |  | 21 |  |  |  |
| Eunice cf. atlantica | 0000000010 | 1010111010 | 1111110112 | 1011101110 | 1101002211 | 1110010010 | 1000110120 | 0101 |
| Eunice stigmatura | 0100002010 | 1010111010 | 1111110112 | 1011101110 | 1101012211 | 1110110010 | 1000110120 | 0101 |
|  | 1 |  |  |  |  |  |  |  |


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