

# 2020 INFECTIOUS DISEASE BOARD REVIEW

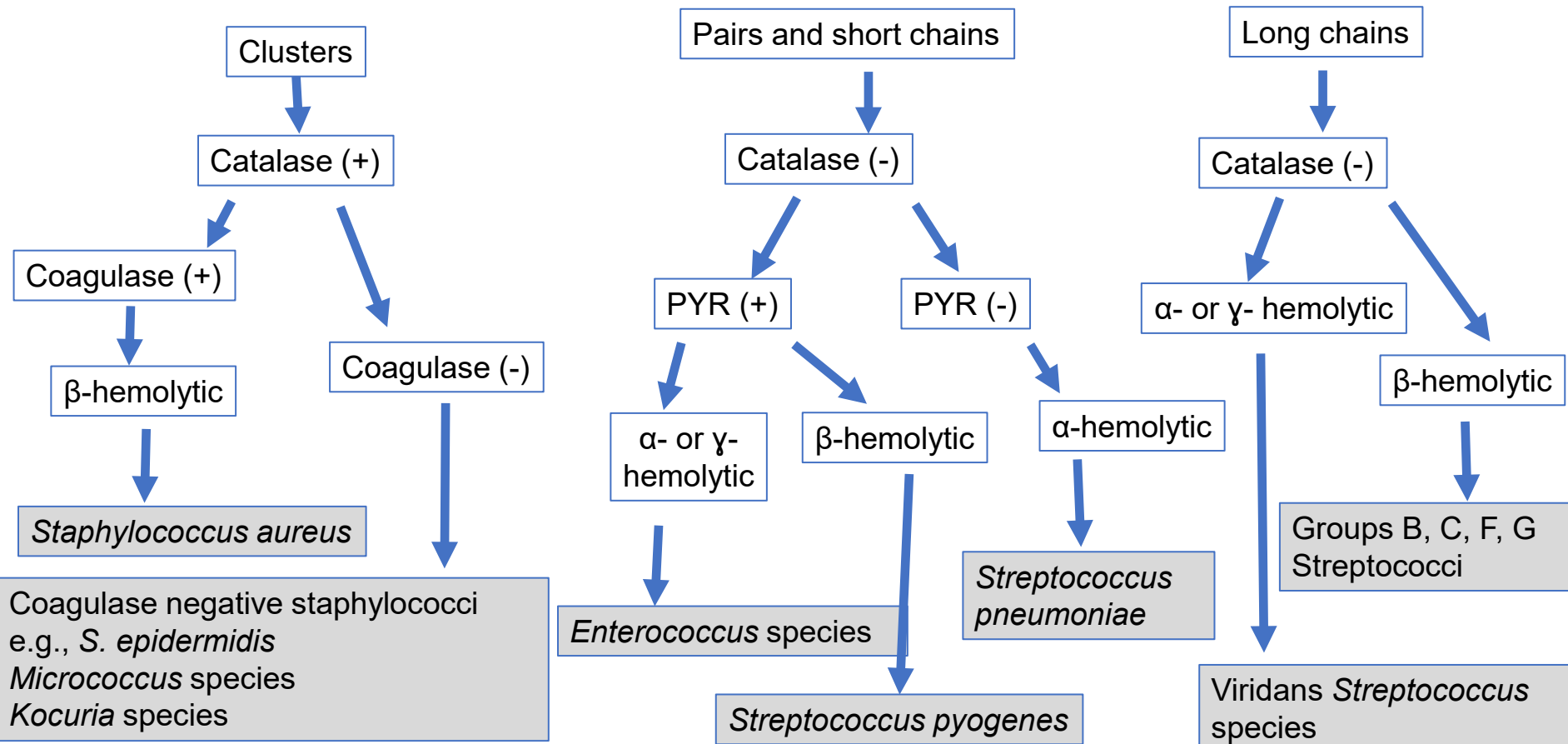


## Microbiology Primer

Robin Patel, MD  
Patrick R. Murray, PhD  
With input from Anna Lau, PhD

# **Bacteriology**

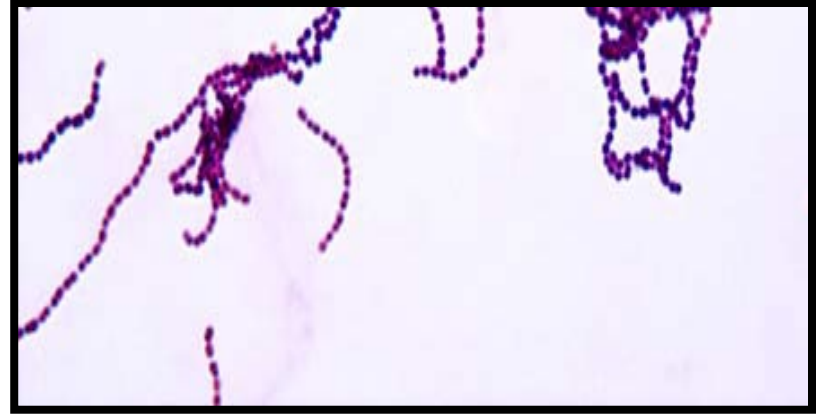
# Gram Positive Cocci



# Gram-Positive Cocci



Staphylococci



Streptococci

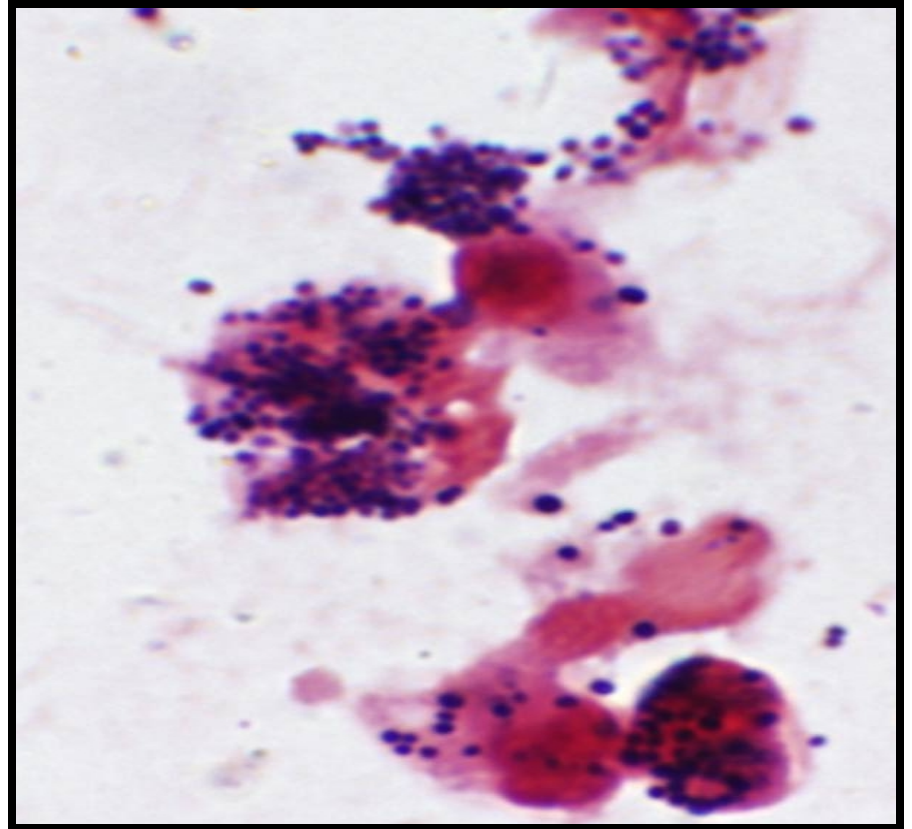


Enterococci

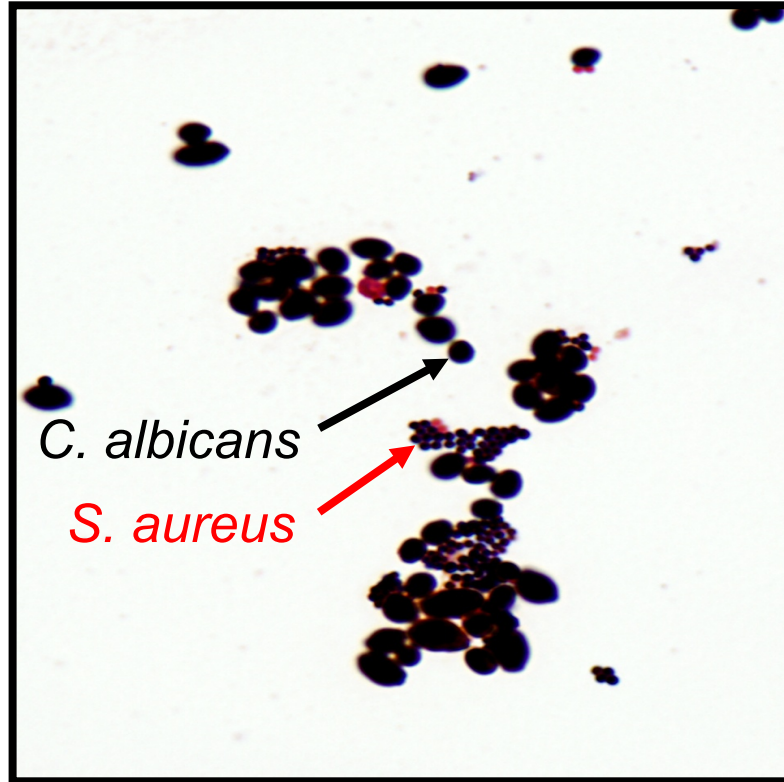


# ***Staphylococcus aureus***

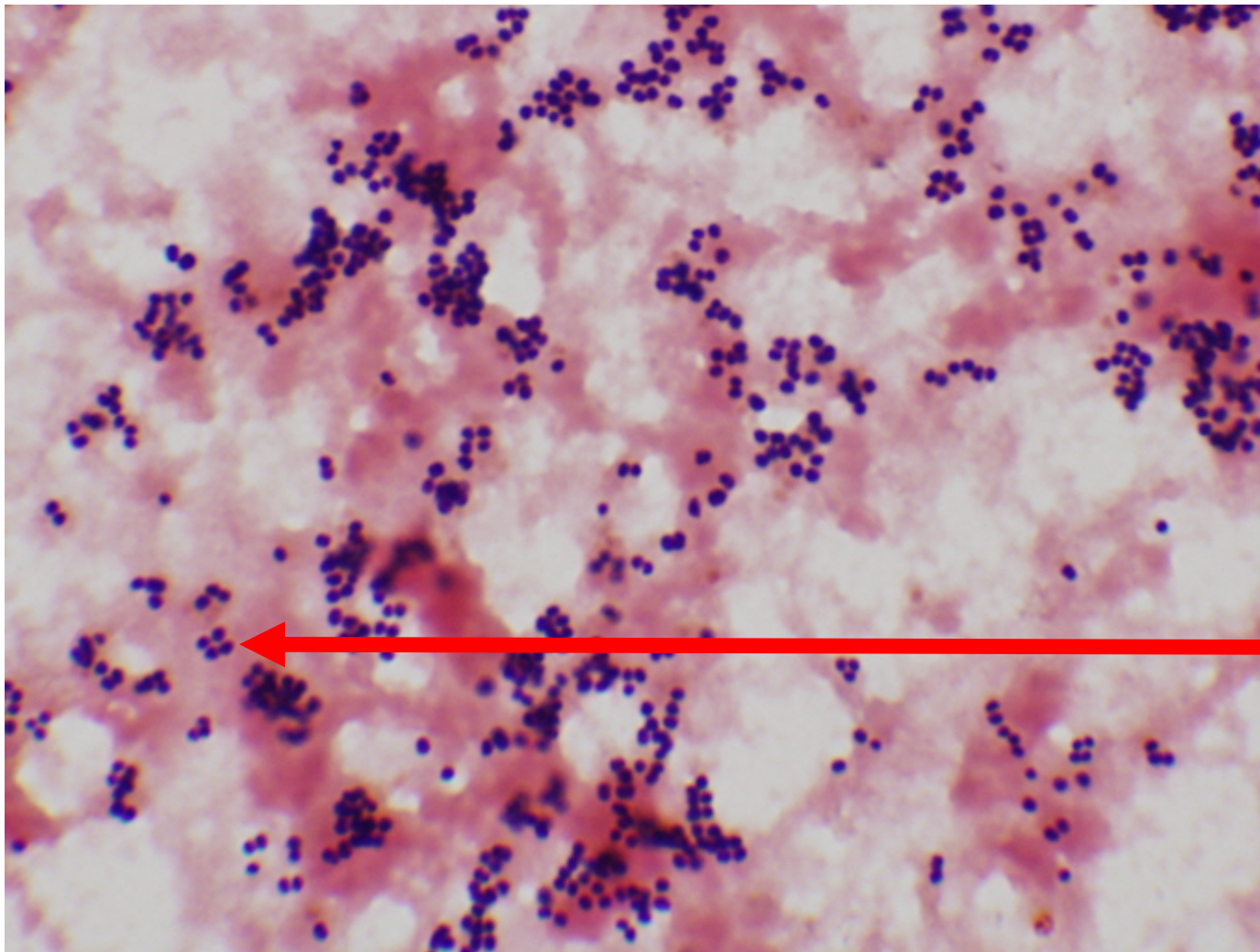
- In clinical specimens, *S. aureus* appears as pairs, small clusters, or within phagocytic cells



# Yeasts May Appear Gram-Positive



Note size difference between *Staphylococcus aureus* and *Candida albicans*



Gram-positive cocci in clusters indicate *Staphylococcus*, *Micrococcus* or *Kocuria* species (plus others)

Tetrads may indicate coagulase negative staphylococci or *Micrococcus* species

# Streptococcus species – $\beta$ -hemolytic

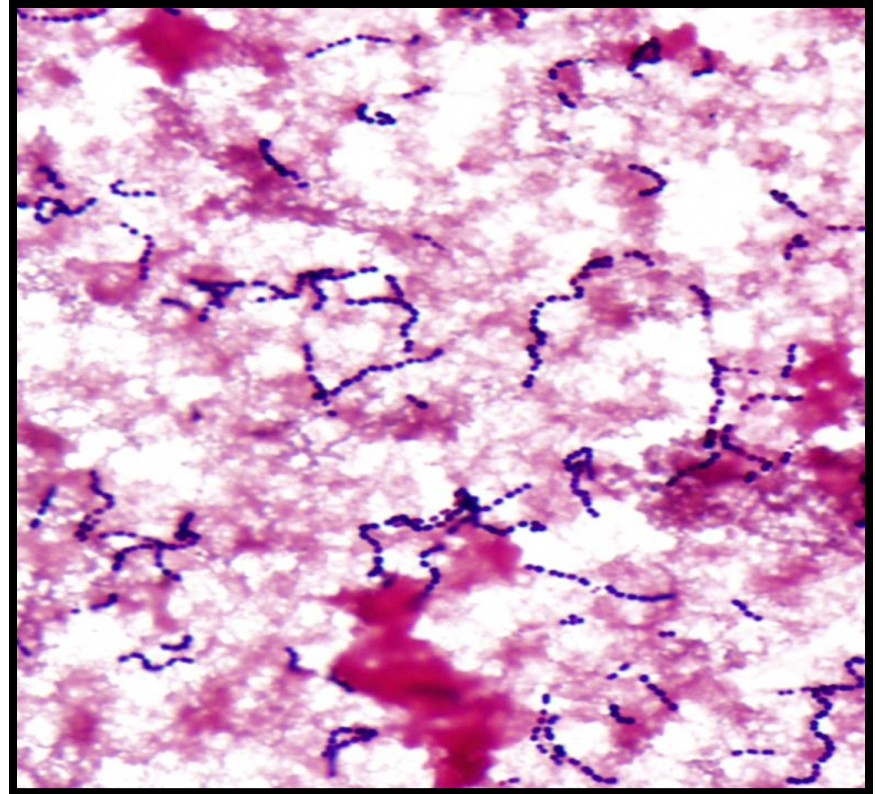
Species	Lancefield	Special features
<i>S. pyogenes</i>	A	Small colony Large zone of $\beta$ -hemolysis “String of pearls” Gram stain (round cells) PYR (+); Bacitracin susceptible
<i>S. agalactiae</i>	B	Large colony Small zone of $\beta$ -hemolysis CAMP test (+); hippurate hydrolysis (+) May be confused with <i>Listeria monocytogenes</i>
<i>S. dysgalactiae</i> subsp. <i>dysgalactiae</i>	C	
<i>S. dysgalactiae</i> subsp. <i>equisimilis</i>	C, G, (A, L)	
<i>S. anginosus</i> group	A, C, G, F	Can be $\alpha$ - or $\beta$ -hemolytic
<i>S. porcinus</i>	E, P, U, V	



# *Streptococcus pyogenes*

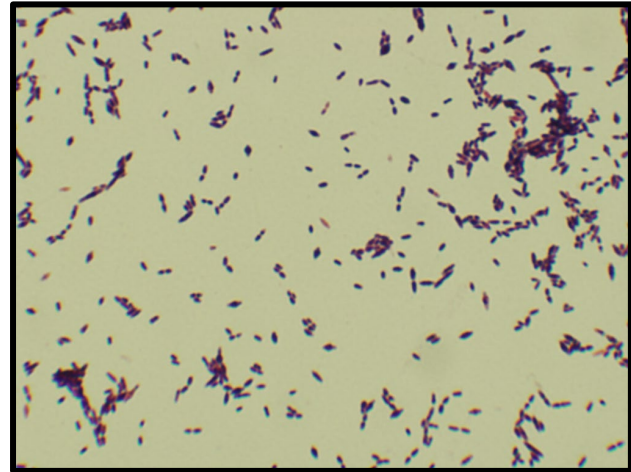
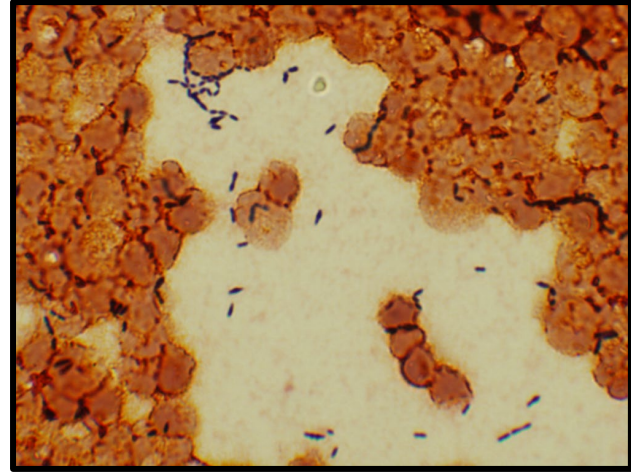
## Group A Streptococcus

- Forms long chains of round cells, described as “string of pearls”
- $\beta$ -hemolytic colonies
  - Only PYR (pyrrolidonyl arylamidase) positive *Streptococcus* species



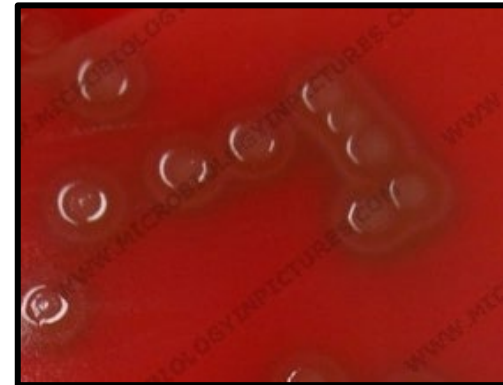
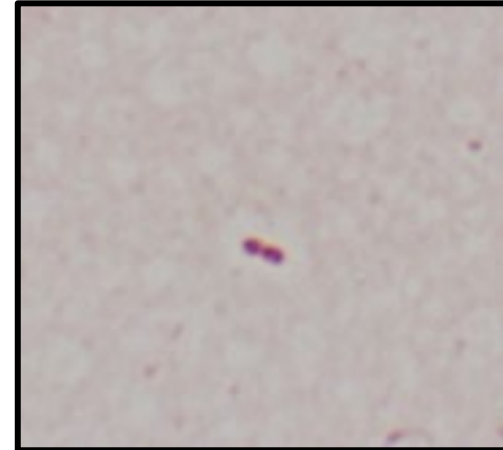
# Nutritionally Variant Streptococci

- Two genera
  - *Abiotrophia* species
  - *Granulicatella* species
- Require pyridoxal or cysteine to grow
- Will grow only on chocolate agar
- May be resistant to penicillin
  - *Granulicatella adiacens* may be ceftriaxone-resistant



# *Streptococcus pneumoniae*

- Member of *Streptococcus mitis* group
- Generally stains uniformly in clinical specimens but may decolorize, appearing Gram-negative in culture
- Individual cells appear elongated, arranged in pairs and short chains
- Refractile capsule
- Elongated, lancet shaped cells
- Doughnut-shaped colonies (raised edges, indented center)
- Cultured organisms rapidly identified by exposing cells to bile which “dissolves” bacteria (“bile soluble”)
  - Optochin susceptible



# Viridans group Streptococci

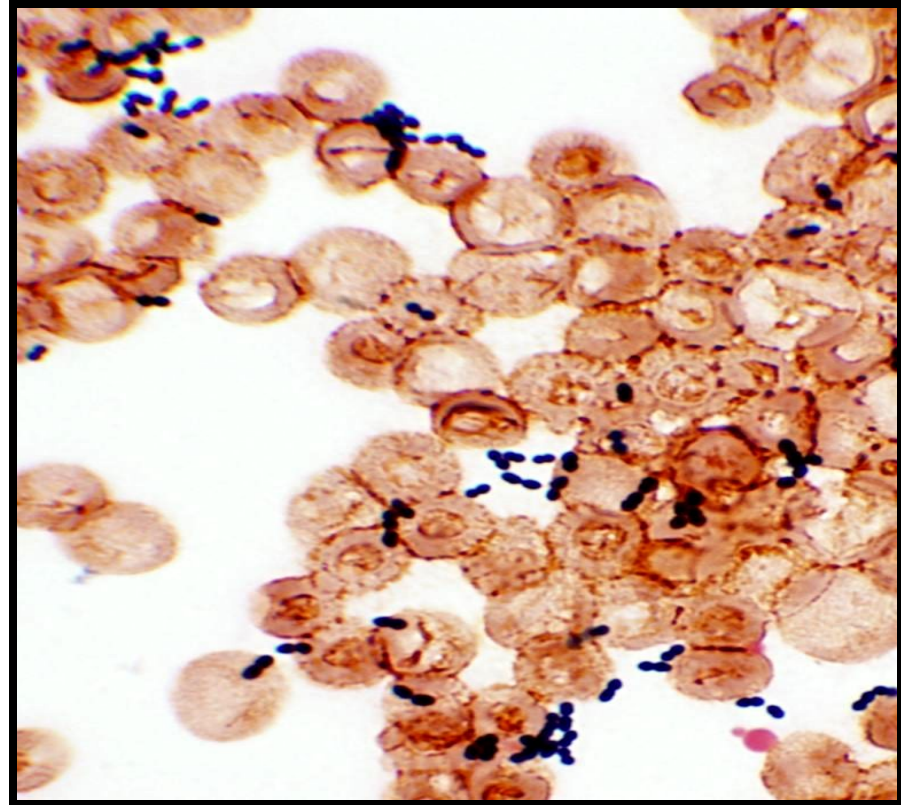
## >25 Species Divided into 5 groups

Group	Species
<i>mutans</i>	<i>S. mutans</i> <i>S. sobrinus</i>
<i>anginosus</i> ( <i>milleri</i> )	<i>S. anginosus</i> <i>S. constellatus</i> <i>S. intermedius</i>  <i>anginosus</i> group associated with abscesses
<i>mitis</i>	<i>S. mitis</i> <i>S. pneumoniae</i> <i>S. sanguinis</i> <i>S. gordonii</i> <i>S. oralis</i>
<i>salivarius</i>	<i>S. salivarius</i> <i>S. vestibularis</i>
<i>bovis</i>	<i>S. gallolyticus</i> subsp. <i>gallolyticus</i> (associated with gastrointestinal malignancies) <i>S. gallolyticus</i> subsp. <i>pasteurianus</i> (associated with neonatal meningitis) <i>S. gallolyticus</i> subsp. <i>macedonicus</i> <i>S. infantarius</i> subsp. <i>infantarius</i> and subsp. <i>coli</i> <i>S. alactolyticus</i> <i>S. equinus</i>

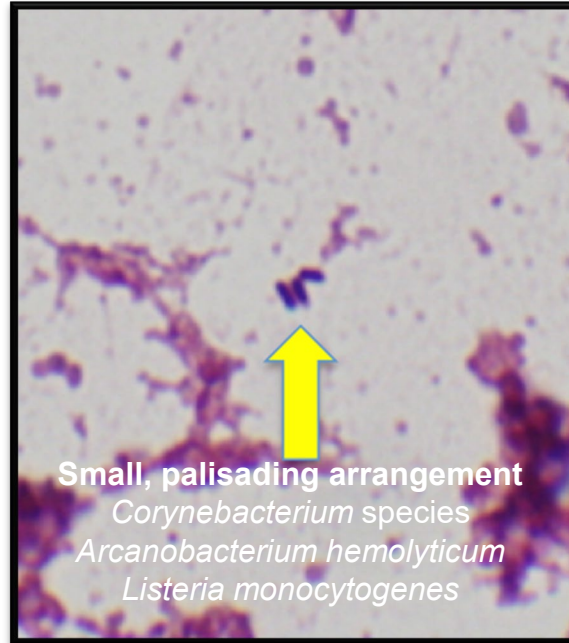


# Enterococci

- Elongated Gram-positive cocci in pairs & short chains
  - Cultured organisms may be identified by positive PYR (pyrrolidonyl arylamidase) test, bile esculin (+), grow in 6.5% NaCl, most grow at 45°C, motile
- Most important species
  - *E. faecium* (often vancomycin and ampicillin resistant)
  - *E. faecalis* (typically vancomycin and ampicillin susceptible)
  - *E. gallinarum*
  - *E. casseliflavus* (yellow)

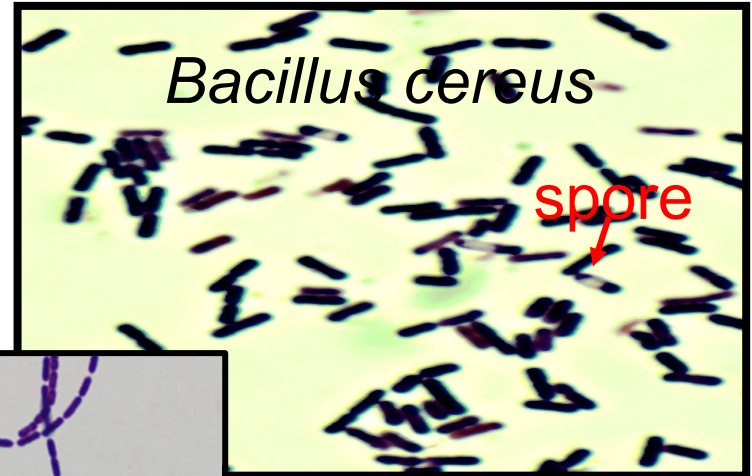


# Gram Positive Rods - Gram Stain Morphologies



# Spore-Forming Gram-Positive Rods

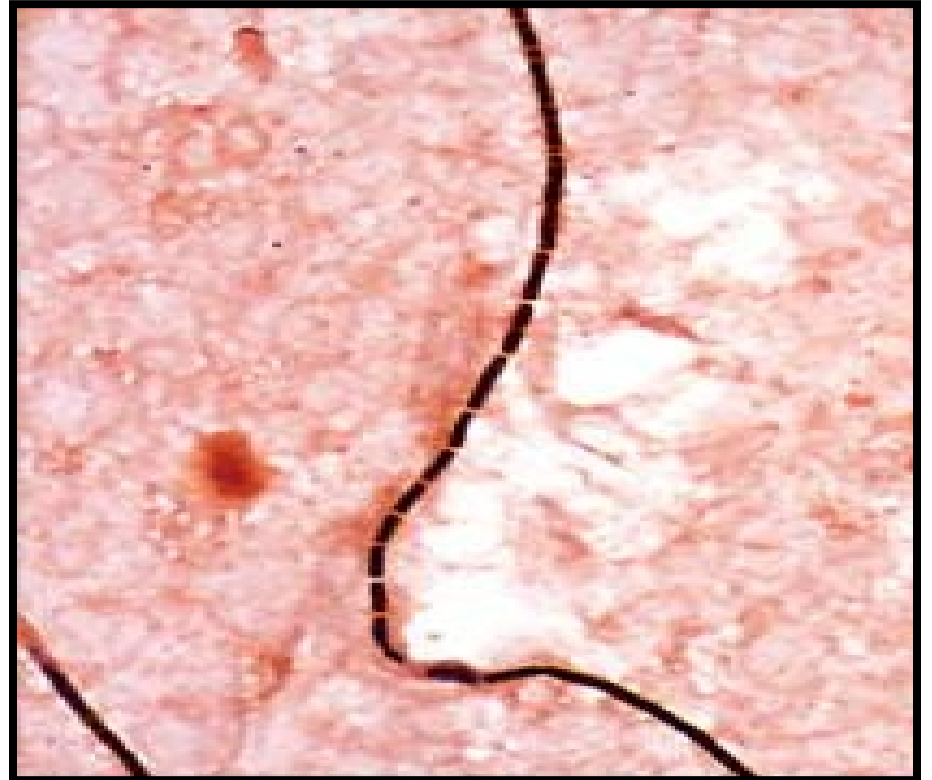
- Aerobes
  - *Bacillus* species
    - *B. anthracis* -  $\gamma$ -hemolytic & non-motile
    - *B. cereus* -  $\beta$ -hemolytic & motile
- Anaerobes
  - *Clostridium* species
    - *C. perfringens*
    - *C. septicum*
    - *C. difficile*
- Spores appear as clear areas



Note presence of spores indicated by cell clearing and/or swelling

# ***Bacillus anthracis***

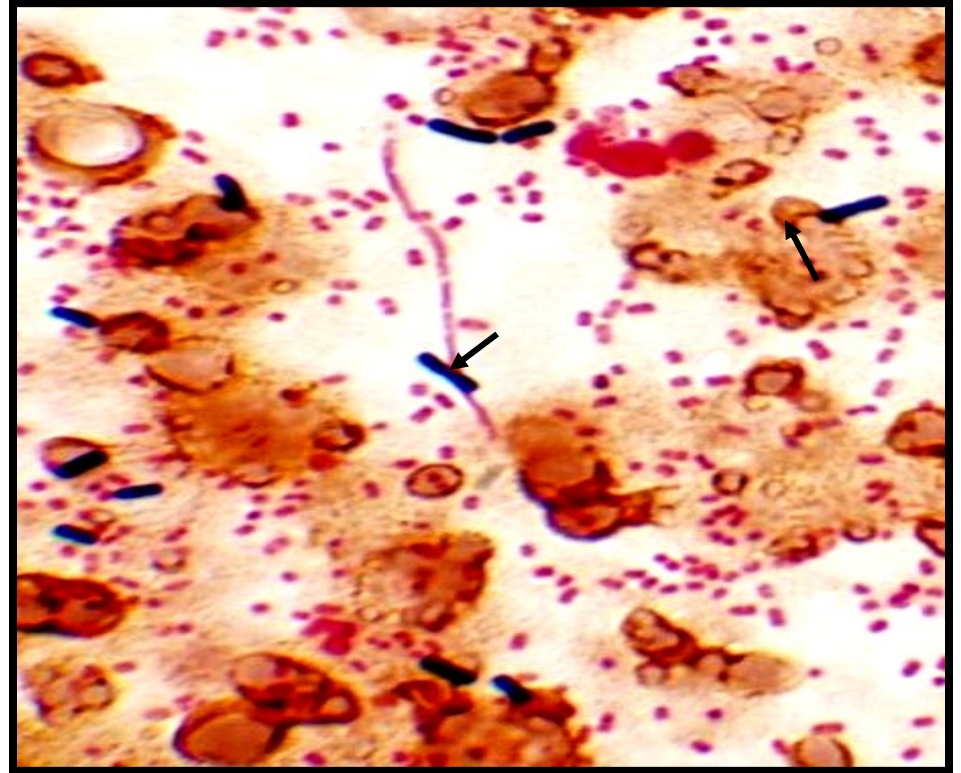
- Gram stain of *B. anthracis* in blood culture from bacteremic patient
  - Spores not seen in image
  - Bacteria form long chains





# *Clostridium perfringens*

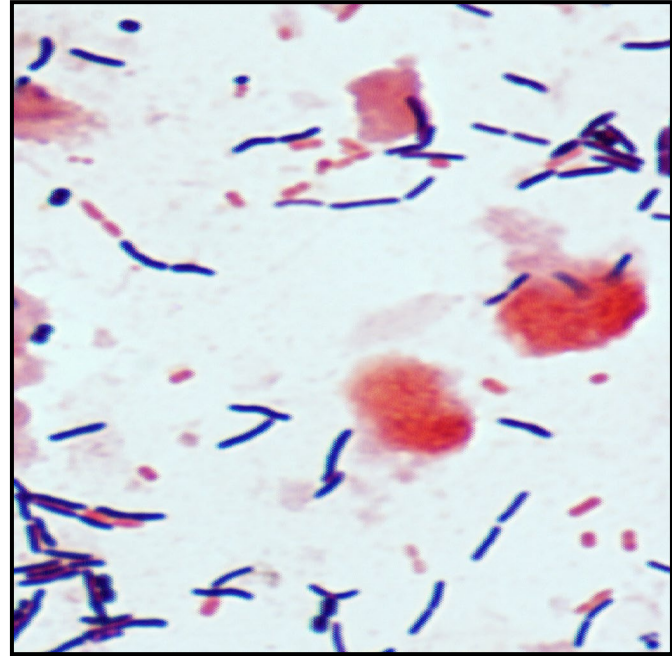
- Common species isolated from clinical specimens
- Spores almost never seen
- Rods “boxcar shaped” or rectangular
  - Larger than most bacteria



*C. perfringens* (arrows) in mixed culture with *Escherichia coli* and *Klebsiella pneumoniae*

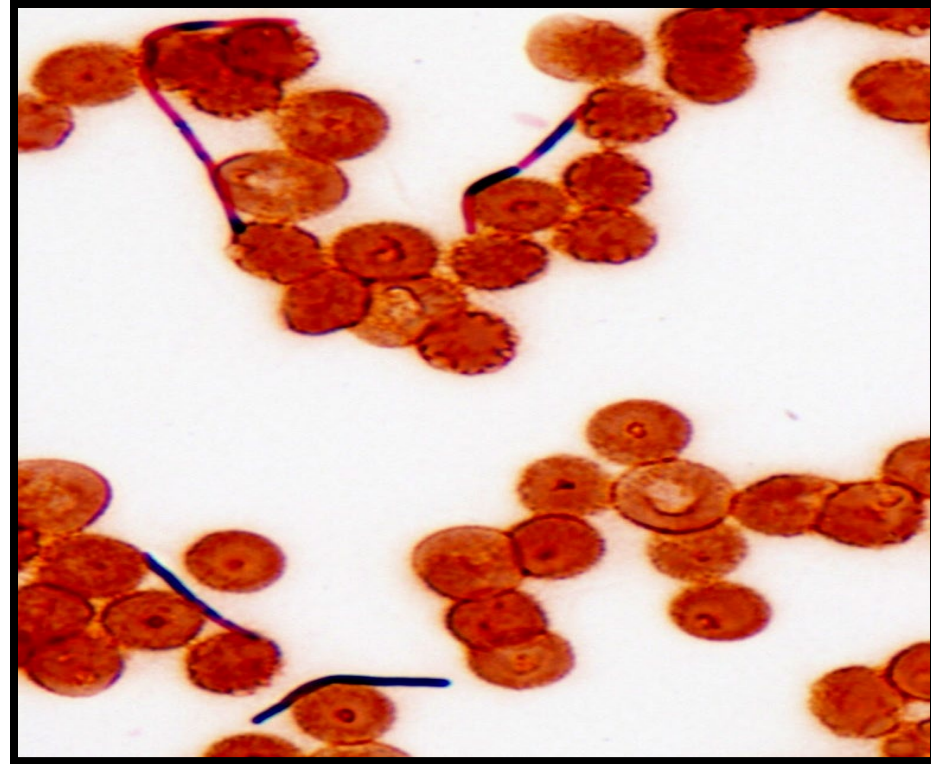
# *Clostridioides difficile*

- Culture not recommended
  - Takes days
  - Toxin production must be documented
- Diagnostic approaches
  - Glutamate dehydrogenase (GDH) antigen followed, if positive, by toxins A/B enzyme immunoassay (EIA) or cytotoxin assay (possible NAAT if negative)
  - Nucleic acid amplification test (NAAT)
- Toxins A/B EIA
  - Less sensitive than NAAT



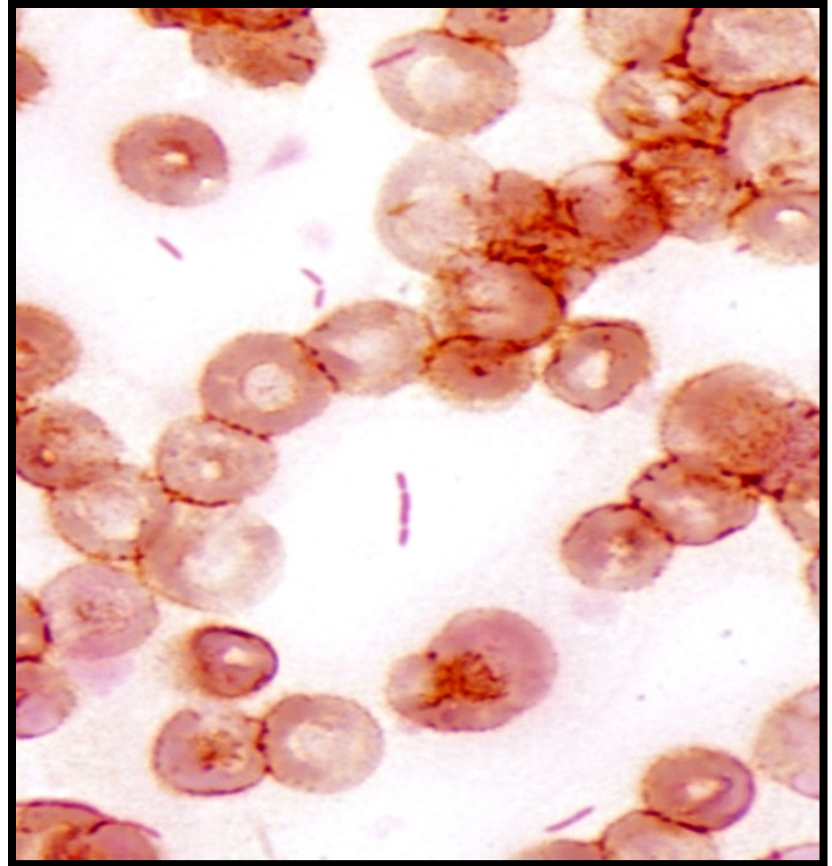
# *Clostridium septicum*

- Grows rapidly
- Forms elongated rods arranged in chains
- Forms spores in specimens and culture (not seen in image)
- Almost always clinically significant when isolated in blood cultures
  - Overwhelming sepsis originating from large intestine
  - Associated with colon cancer, hematologic malignancy



# *Clostridium tertium*

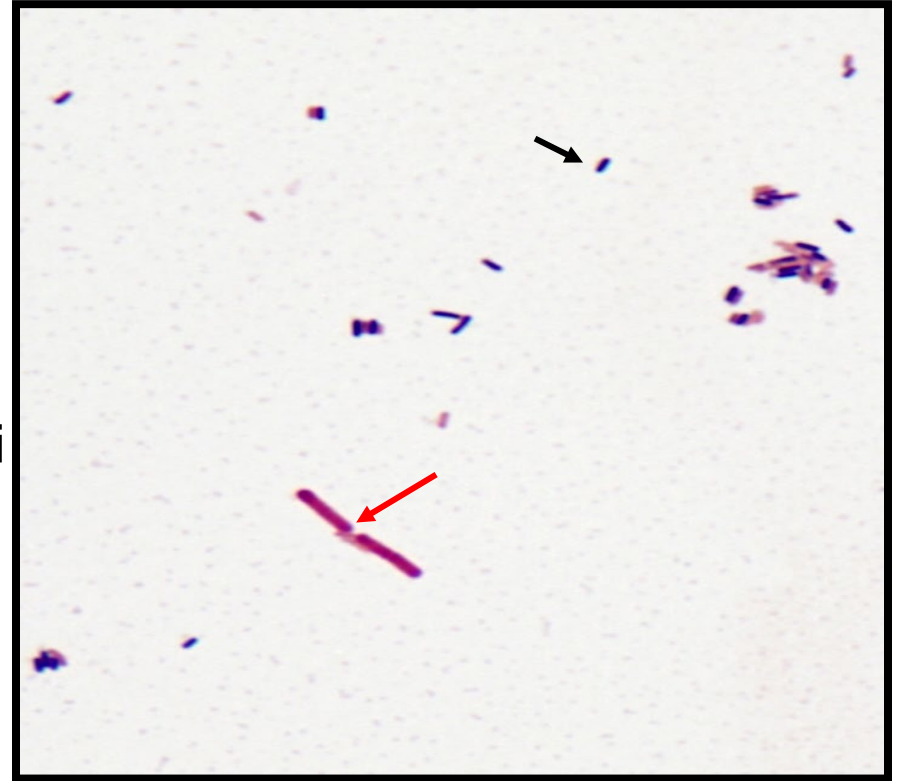
- Characteristically stains Gram-negative
  - Clue that this is not a Gram-negative bacterium → will not grow on MacConkey agar
- May grow aerobically but grows better anaerobically
- May form spores (not seen in image)





# *Listeria monocytogenes*

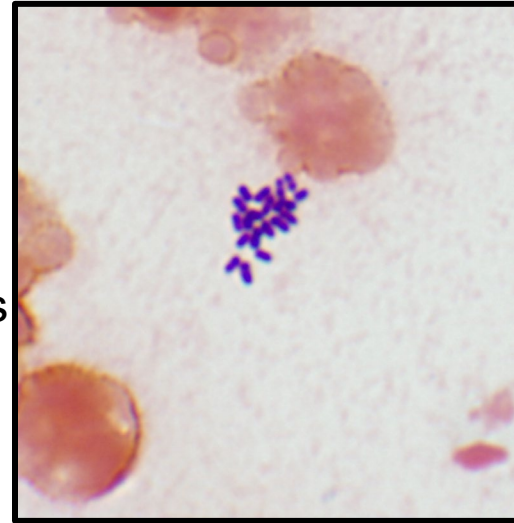
- Small, non-spore forming, Gram-positive rods
- Colonies weakly  $\beta$ -hemolytic
- Can grow at 4°C
- Tumbling motility
- May be mistaken for either streptococci or corynebacteria and discounted as contaminant
  - CAMP (+) and hippurate hydrolysis (+)
    - May be confused with *S. agalactiae*
- Grows slowly



Compare size *Listeria* cells (black arrow) to pair of Gram-negative rods (red arrow)

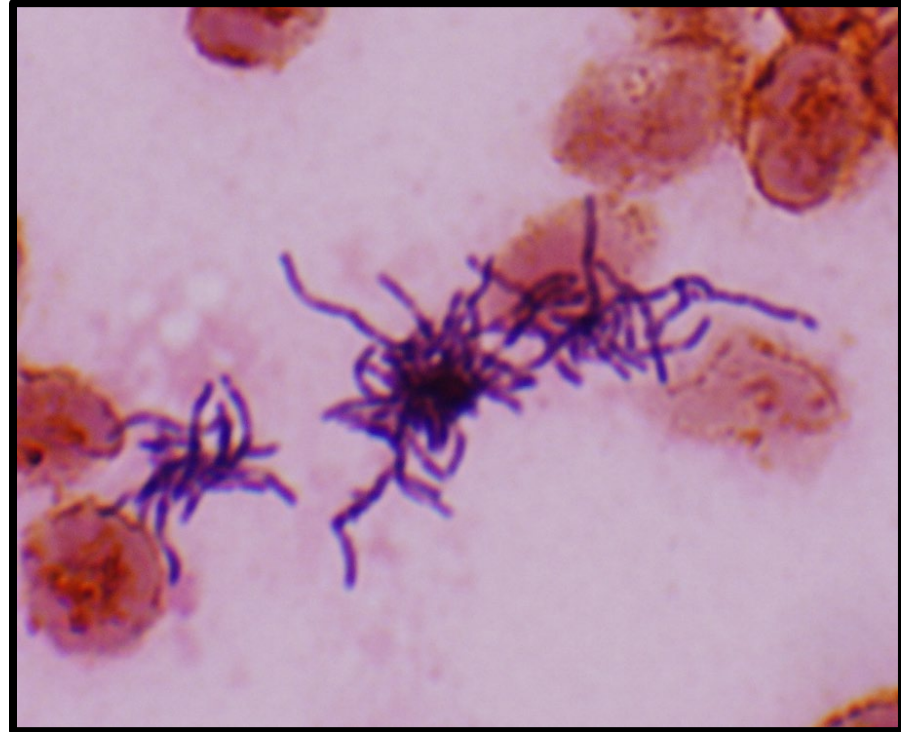
# ***Corynebacterium* species, *Arcanobacterium hemolyticum***

- *C. diphtheriae* pathogenic; other species less pathogenic
  - Clump or palisade
    - Aerobes in contrast to *Cutibacterium/Propionibacterium* species which are anaerobes
- *Arcanobacterium hemolyticum*
  - Catalase (-); distinguishes from *Corynebacterium* species
  - Presents like streptococcal pharyngitis
  - $\beta$  hemolytic
  - Negative on Lancefield grouping performed on  $\beta$ -hemolytic streptococci



# ***Cutibacterium* (formerly *Propionibacterium*) *acnes***

- **Common contaminant**
  - Occasional cause of PJI, endocarditis, CSF shunt infection
- **Anaerobic growth**
- **Slow growing – several days**
- **Forms clumps of Gram-positive rods**
  - Anaerobic counterpart to *Corynebacterium* species



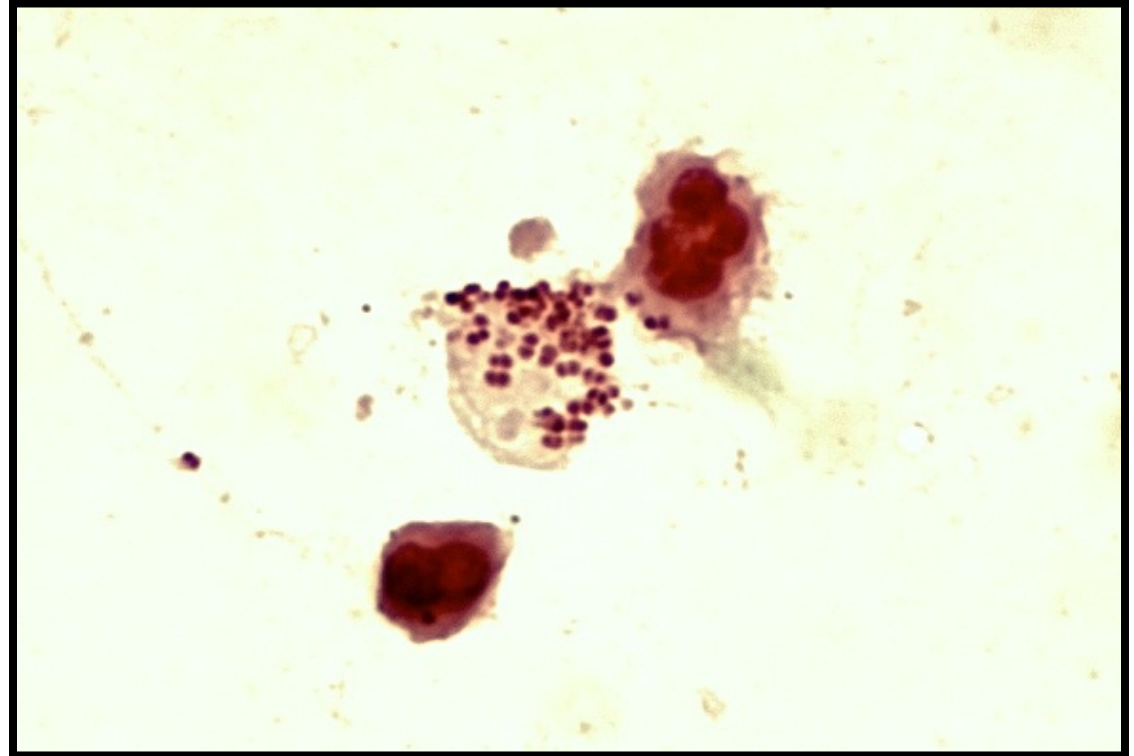
# Gram-Negative Cocci, Coccobacilli, and Rods

- Cocci
  - *Neisseria* species
  - *Moraxella catarrhalis*
- Coccobacilli
  - *Moraxella* species
  - *Acinetobacter* species
  - *Haemophilus* species
- Rods
  - *Enterobacteriaceae* (e.g., *Escherichia coli*, *Klebsiella* species)
  - *Pseudomonas* species, *Stenotrophomonas maltophilia*, *Burkholderia* species
  - Miscellaneous (e.g., *Bacteroides* species, *Fusobacterium* species)

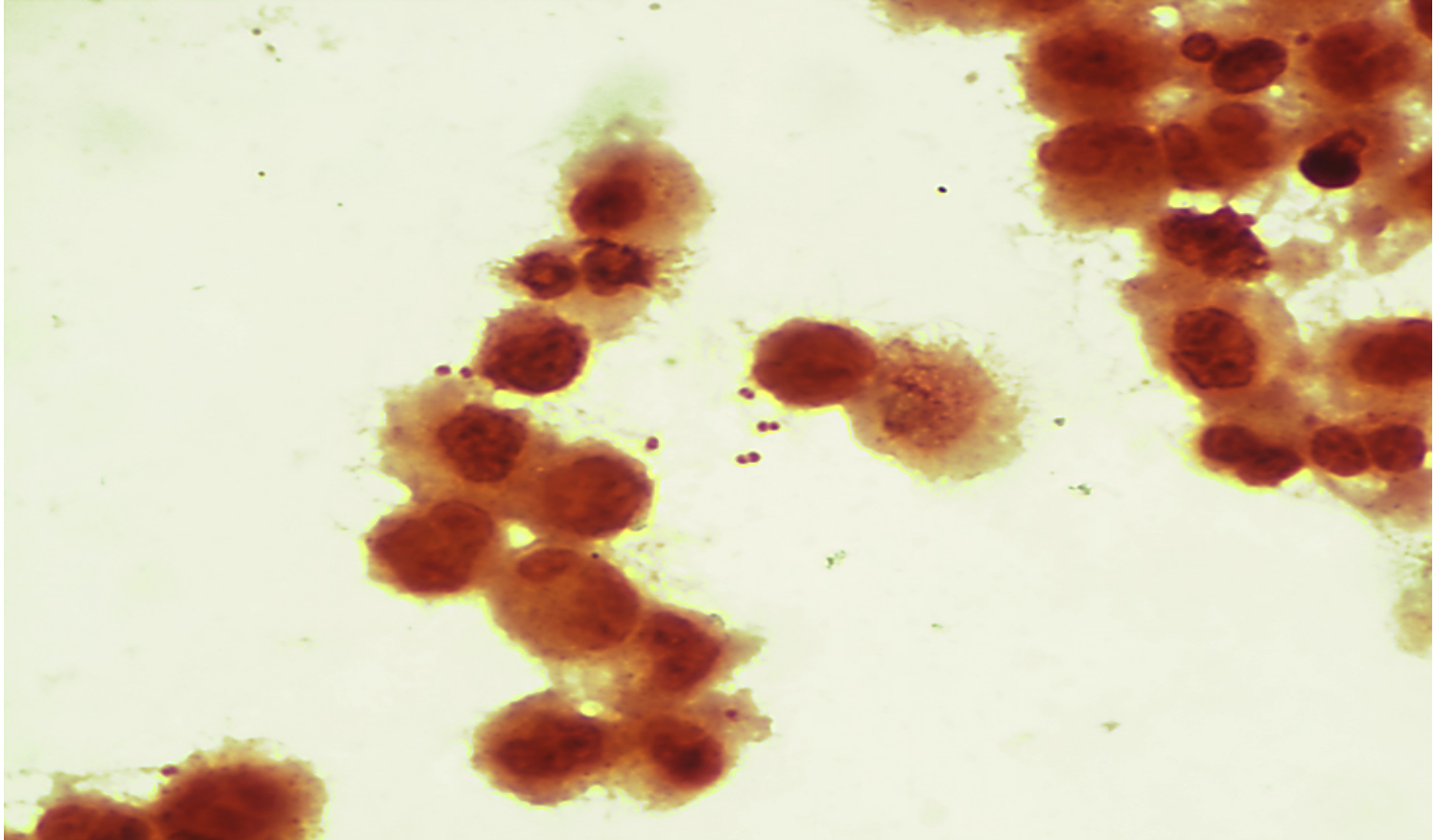
# ***Neisseria gonorrhoeae* and *Neisseria meningitidis***

## ***Neisseria gonorrhoeae***

- Typically arranged in pairs with adjoining sides flattened into shape resembling a “coffee bean”
- Gram-negative coccobacilli may be mistaken for *Neisseria* species
- Fastidious
- Modified Thayer Martin used for gonococcal culture
- No growth on MacConkey agar



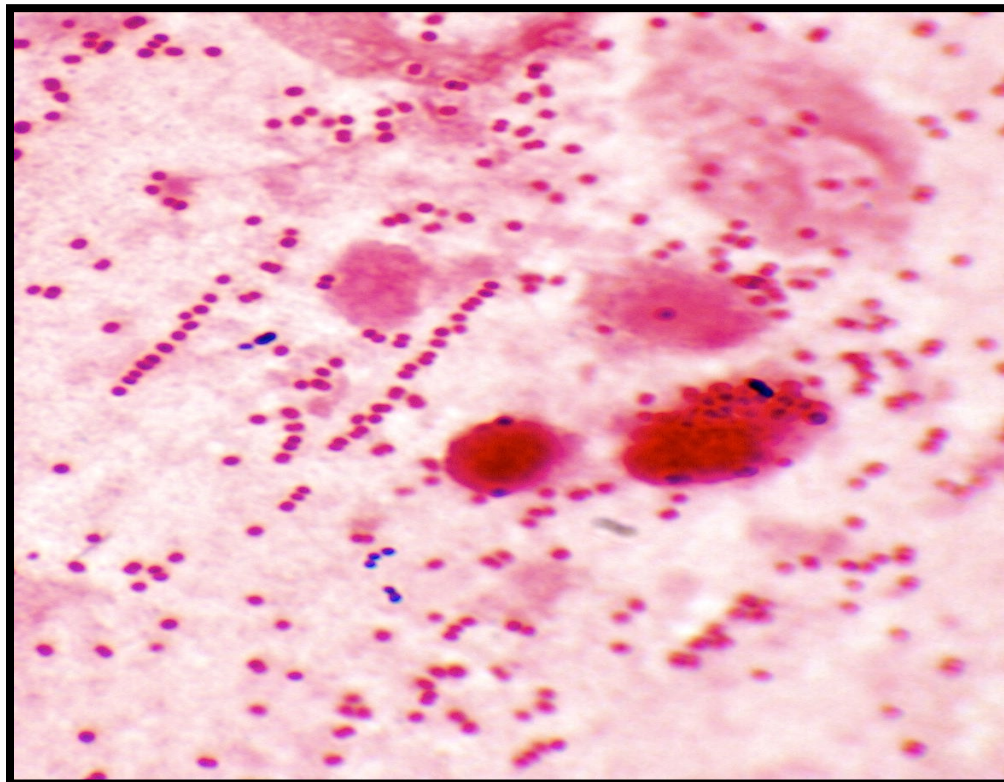
# ***Neisseria meningitidis***





# ***Moraxella catarrhalis***

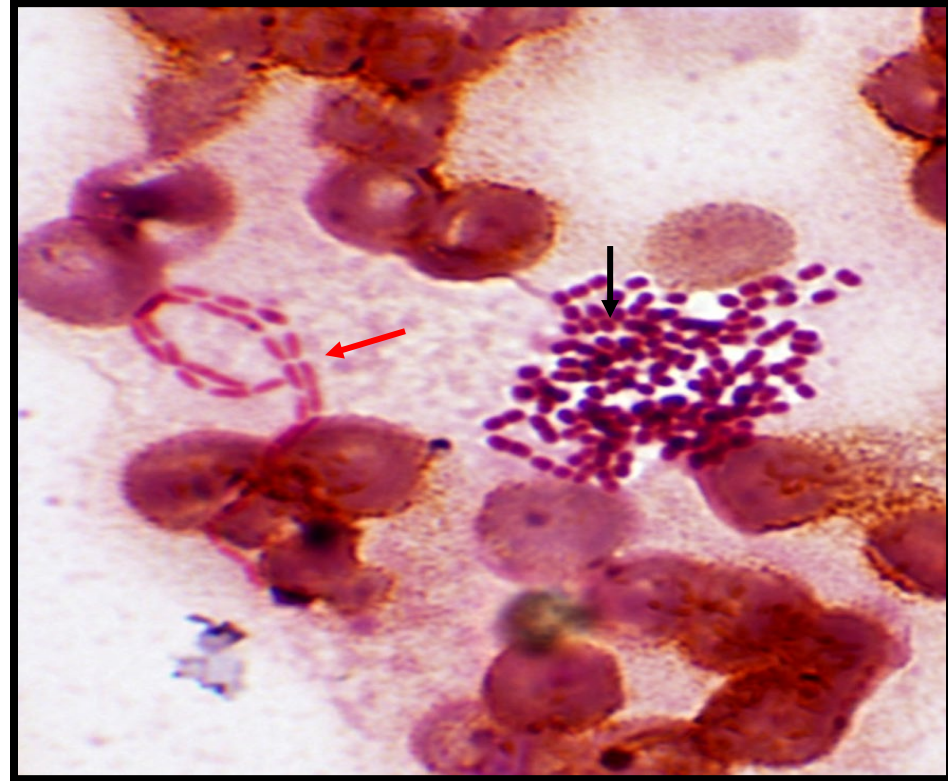
- *Moraxella* species are typically Gram-negative rods
- *M. catarrhalis* originally classified as a *Neisseria* species because its morphology resembles *Neisseria* species



***M. catarrhalis* in sputum from patient with pneumonia - note the large number of neutrophils**

# *Acinetobacter* species

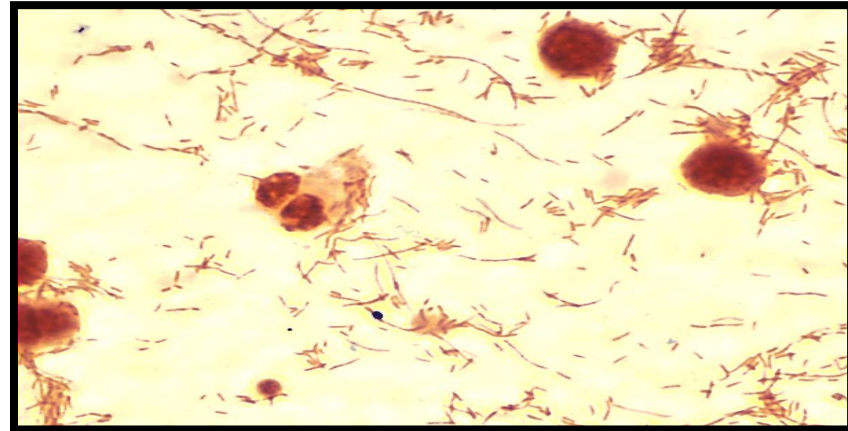
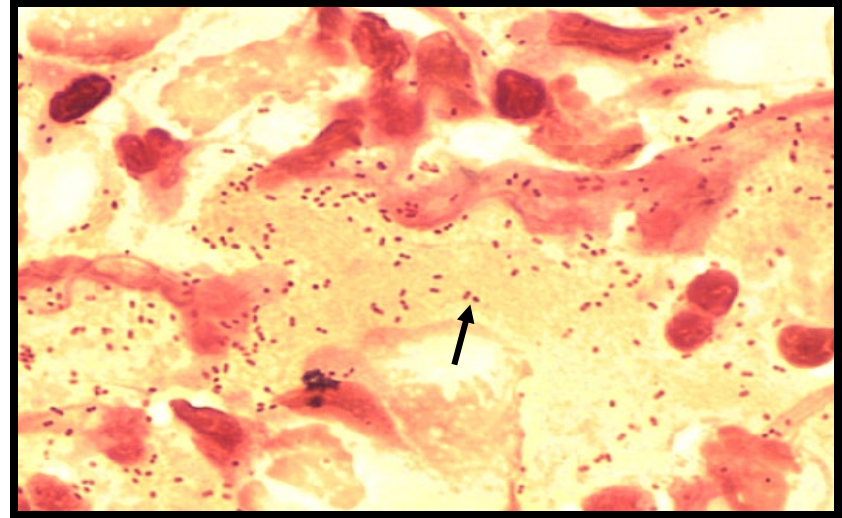
- Gram-negative coccobacilli that may retain crystal violet and resemble Gram-positive cocci in pairs (black arrow).
- Other bacterium shown is *Pseudomonas* species (red arrow), a more obvious Gram-negative rod
- *Acinetobacter* species are larger than *Neisseria* species; adjoining sides are not flattened





# *Haemophilus* species

- Very small Gram-negative rods or coccobacilli (top figure)
- Long pleomorphic forms (bottom figure) may be seen in patients receiving antibiotics
- Bottom figure is Gram stain of spinal fluid from child with *H. influenzae* meningitis
- *H. influenzae* requires X and V factors to grow; will only grow in chocolate agar or as satellite colonies on sheep blood agar - tiny colonies around  $\beta$  hemolytic colony (e.g., *E. coli*, *S. aureus*)



# Gram-Negative Rods

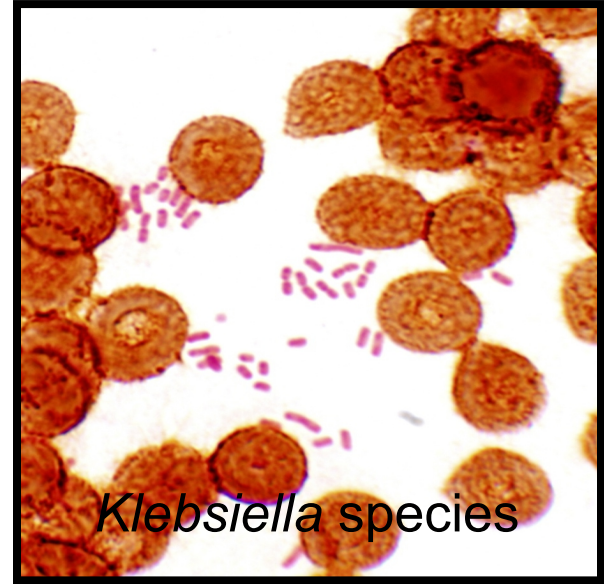
Subdivided here by spot oxidase test and growth on MacConkey agar\*

	Growth on MacConkey agar	
Oxidase	Yes	No
Positive	<i>Aeromonas</i> species <i>Vibrio</i> species <i>Burkholderia</i> species <i>Elizabethkingia meningoseptica</i> <i>Pseudomonas</i> species	<i>Aggregatibacter</i> species <i>Cardiobacterium</i> species <i>Eikenella corrodens</i> <i>Kingella kingae</i> <i>Pasteurella multocida</i> <i>Brucella</i> species
Negative	<i>Chromobacterium</i> species Enterobacterales <i>Acinetobacter</i> species <i>Stenotrophomonas maltophilia</i>	<i>Capnocytophaga canimorsus</i> <i>Capnocytophaga cynodegmi</i> <i>Streptobacillus moniliformis</i> <i>Francisella tularensis</i>

\*Selective for Gram negatives; differentiates lactose fermenters (pink colonies) from non-lactose fermenters (colorless colonies)

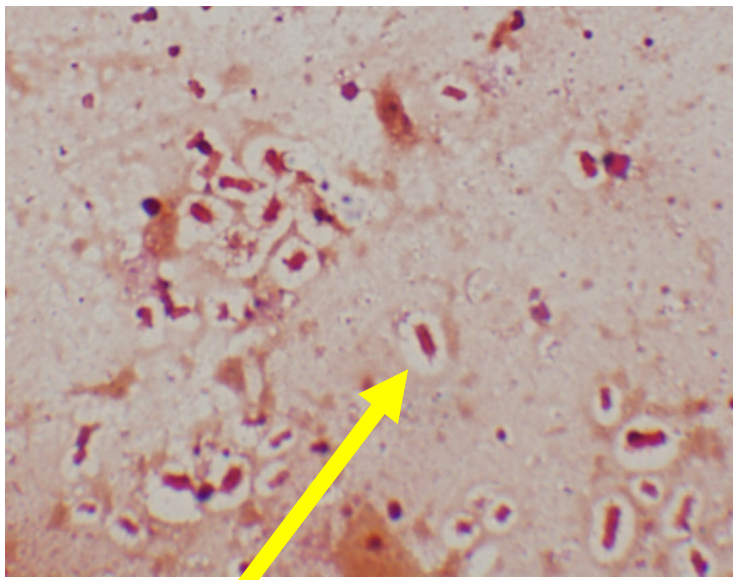
# Enterobacteriales

- >100 genera
- Glucose fermenters
- Oxidase negative
  - Exception (*Plesiomonas* species)
- Grow on MacConkey agar
- Relatively large Gram-negative rods
  - May stain more intensely at ends (“bipolar” staining)



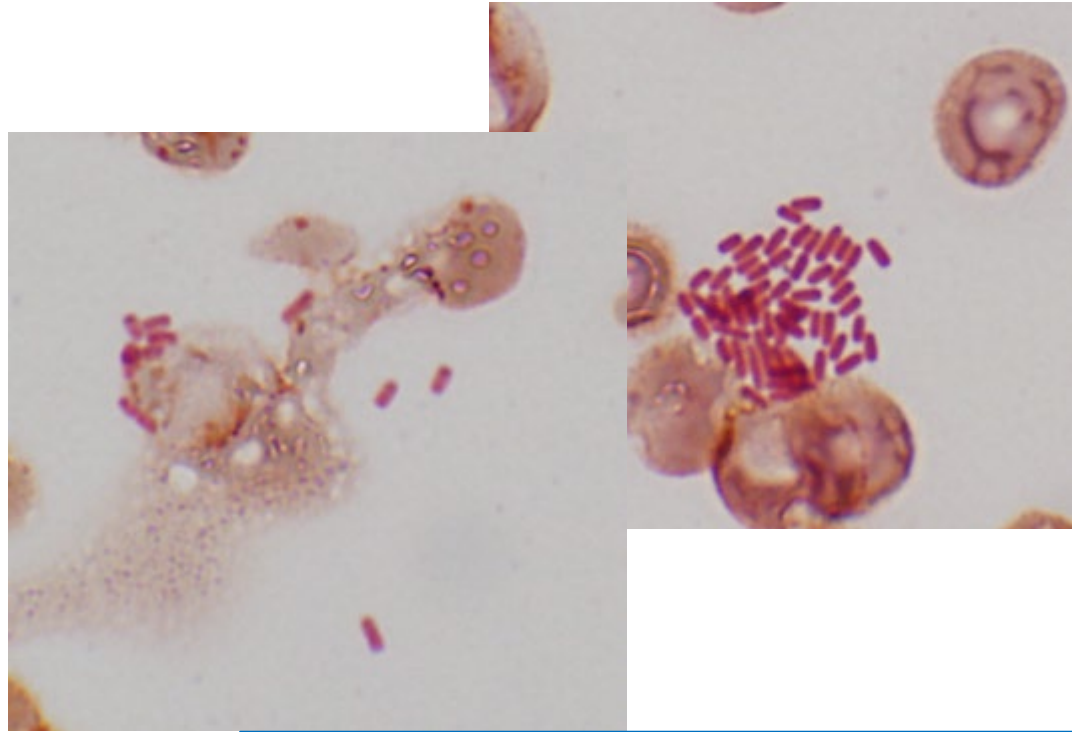
*Klebsiella* species

<u>Lactose fermentors</u> (pink colonies on MacConkey)	<u>Late lactose fermentors</u> (pink colonies at 48 h)	<u>Non-lactose fermentors</u> (colorless on MacConkey)
<i>Escherichia coli</i>	<i>Citrobacter</i> species	<i>Shigella</i> species
<i>Klebsiella</i> species ( <i>pneumoniae</i> , <i>oxytoca</i> , <i>aerogenes</i> )	<i>Serratia</i> species	<i>Salmonella</i> species
<i>Enterobacter</i> species ( <i>cloacae</i> complex)		<i>Yersinia</i> species
		<i>Proteus</i> species
		<i>Providencia</i> species



Example of encapsulated *K. pneumoniae*. Notice the clearing around the cells.

***K. pneumoniae* often has a "short, fat, chubby" appearance ± capsule**

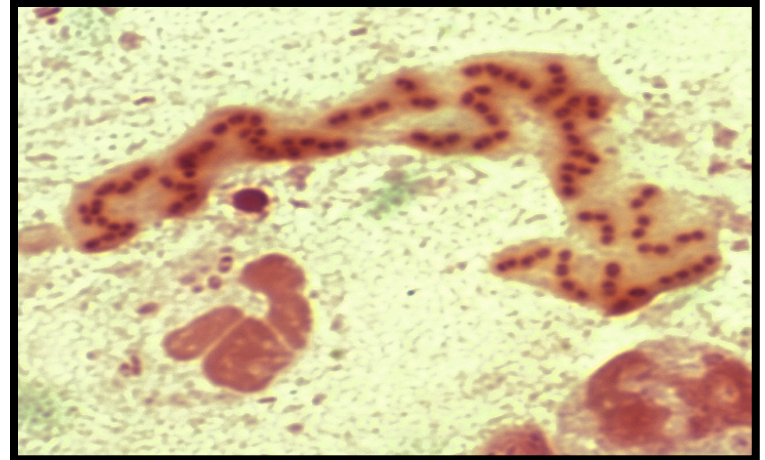
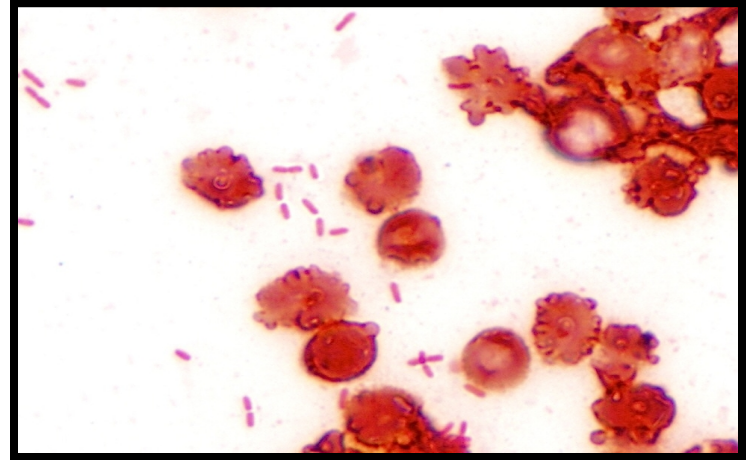


Gram stains of *E. cloacae* complex and *E. coli* from blood. Notice the **bipolar staining** (darkening at the ends) suggestive of the Enterobacterales



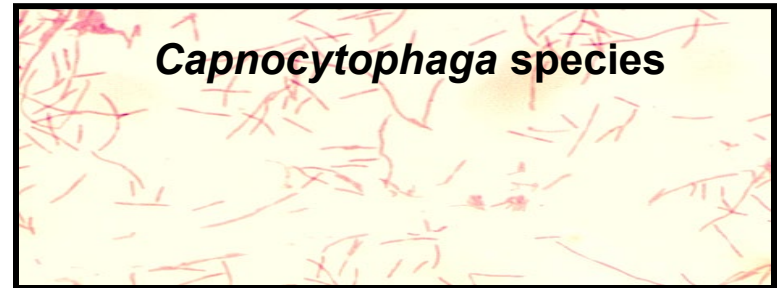
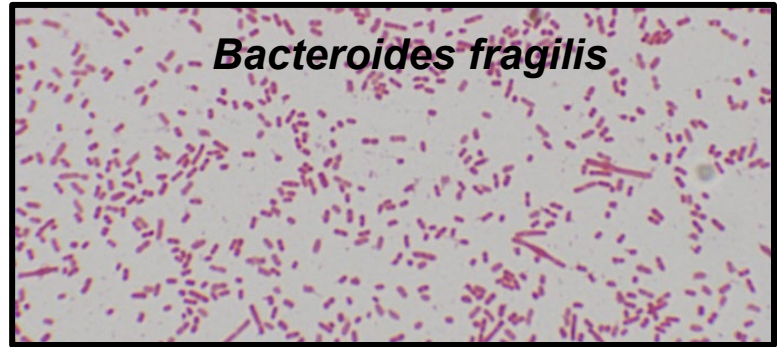
# Gram-Negative Rods, continued

- *Pseudomonas* species,  
*Burkholderia* species,  
*Stenotrophomonas maltophilia*
  - Small Gram-negative rods typically arranged singly or in pairs
  - *Pseudomonas aeruginosa* may be surrounded by extracellular matrix/capsule (bottom figure)
- Non-glucose, non-lactose fermenters
- *P. aeruginosa* smells like grapes



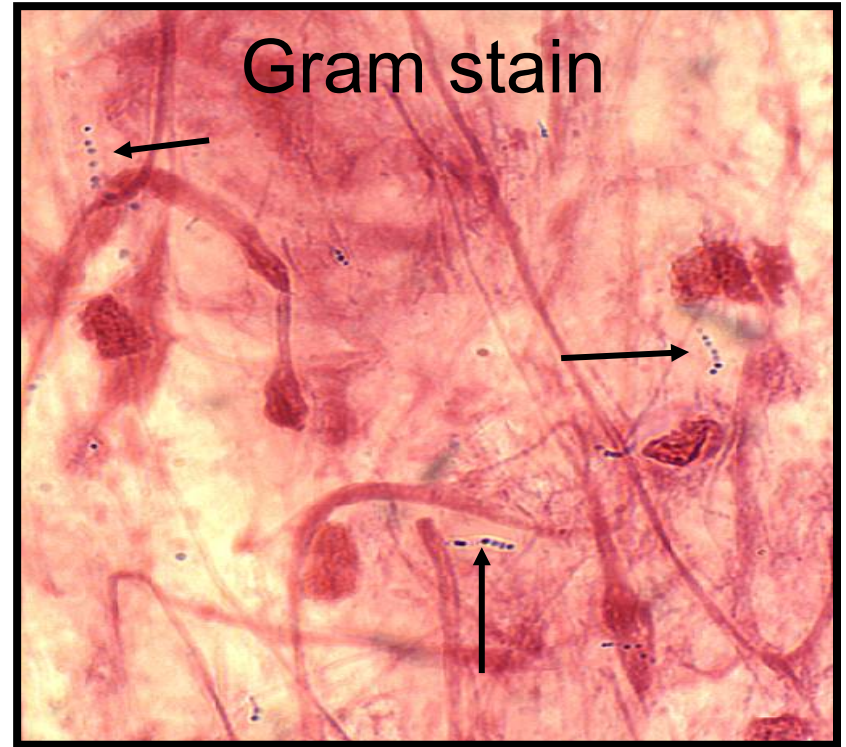
# Gram-Negative Rods, continued

- *Bacteroides fragilis*
  - Anaerobe
  - Pleomorphic Gram-negative rods - short and long
- Thin and long Gram-negative rods
  - Anaerobe
    - *Fusobacterium* species
  - Aerobe
    - *Capnocytophaga* species



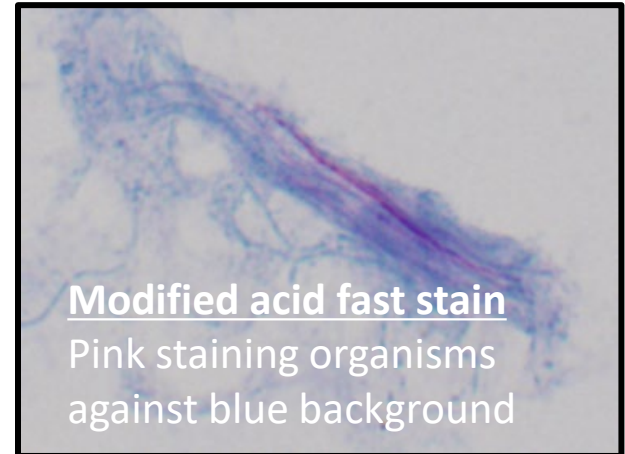
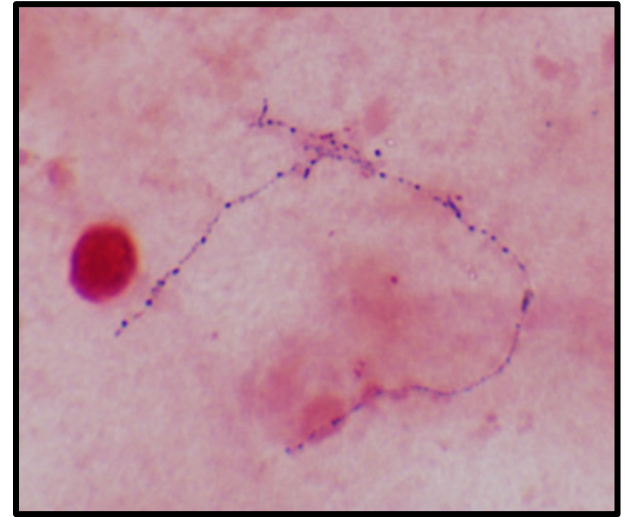
# *Mycobacterium* species

- Mycobacteria *may* retain crystal violet in Gram stain appearing as beaded rods (arrows)
- Rapid growers (growth within 7 days)
  - *M. chelonae*
  - *M. abscessus* group (includes *M. massiliensis*)
  - *M. mucogenicum*
  - *M. fortuitum*
- *M. avium* complex (growth 5-7 days)
- Slow growers (growth >10 days)
  - *M. tuberculosis* complex (includes *M. bovis*)
- Scotochromogens (colonies produce pigment without light exposure)
  - *M. gordonae*, *M. scrofulaceum*
- Photochromogens (colonies produce pigment upon exposure to light)
  - *M. marinum*, *M. kansasii*



# *Nocardia* species

- Gram stain - **thin, filamentous, branching forms**, stain irregularly
- No other acid-fast organisms form long, branching structures
- Differentiation from streptococci:
  - True branching of filaments
  - “Beads” do not touch and are irregularly distributed
- Modified acid fast stain



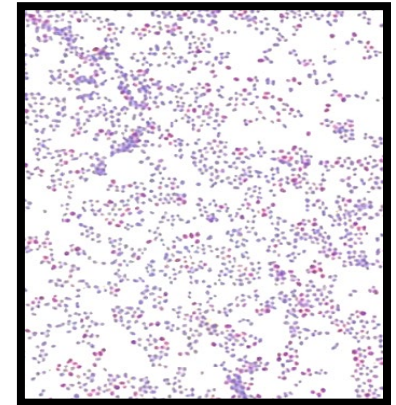


# *Rhodococcus* species – Gram Stain

- Retain crystal violet dye more uniformly than *Mycobacterium* or *Nocardia* species
- After growth for few hours, stain well with Gram stain and appear rod-like
- With longer incubation, bacteria assume coccoid form
- Most cells decolorize easily with weak acid-alcohol solution, so only a few cells appear “red” (acid fast)



4 hours



24 hours\*

\*Photo was prepared from colony of *Rhodococcus* species growing on media used for mycobacteria

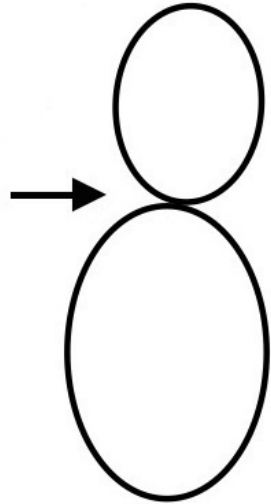
If colonies grown on the blood agar plates used in bacteriology, very few cells appear acid fast

# **Mycology**

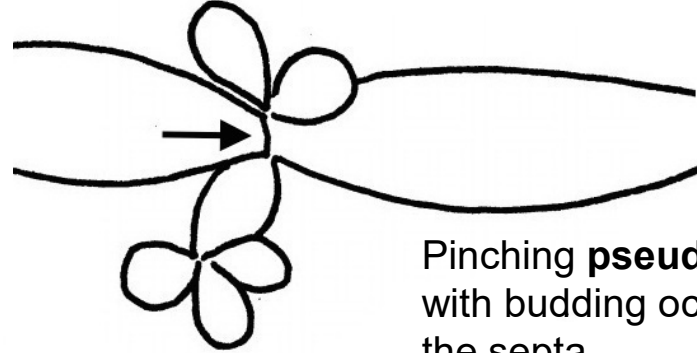
# Yeasts and Molds

- **Yeasts** (single cell organisms)
  - Some produce pseudohyphae (pinching & budding at septa)
  - *C. albicans* can produce pseudohyphae AND true hyphae
  - *Candida* species (e.g., *C. albicans*, *C. glabrata*, *C. tropicalis*, *C. parapsilosis*, *C. krusei*, *C. auris*)
  - *Cryptococcus* species (e.g., *C. neoformans*, *C. gattii*)
  - *Malassezia* species (e.g., *Malassezia furfur*)
  - *Pneumocystis jirovecii*
- **Molds** (multicellular organisms)
  - Septate (true hyphae; parallel walls, branching before or after the septa) *versus* aseptate
  - Hyaline (colony reverse cream/tan/white) *versus* dematiaceous (colony reverse black/brown; presence of melanin)
- **Dimorphic fungi** (yeast phase 35° C/mold phase 25° C)
  - *Histoplasma capsulatum*, *Blastomyces dermatitidis*, *Sporothrix schenckii*, *Coccidioides immitis/posadasii*

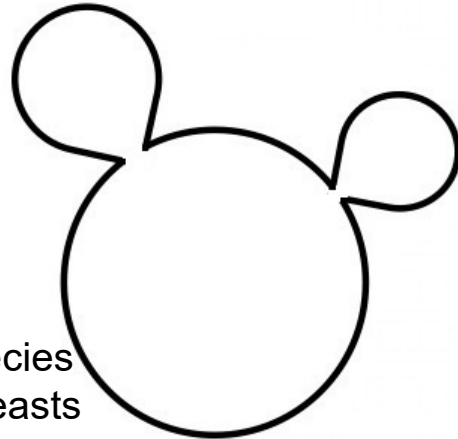
# Yeasts



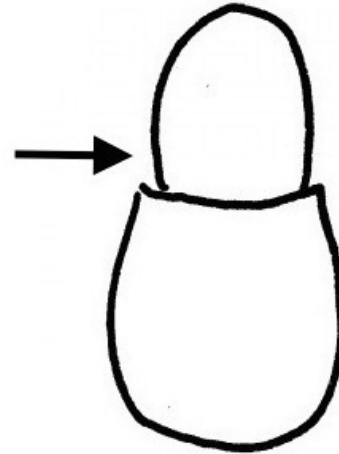
*Candida* species  
Budding **oval** yeasts



Pinching **pseudohyphae**  
with budding occurring at  
the septa



*Cryptococcus* species  
Budding **round** yeasts



*Malassezia* species  
with prominent  
**collarete** at point of  
budding

# *Candida* species

- Top: *Candida albicans* isolated in blood culture; note **yeast cells** and **pseudohyphae**
- Bottom: Yeast colonies with “feet” indicative of pseudohyphae production



Gram stain of budding yeasts



Yeast colonies with “feet”



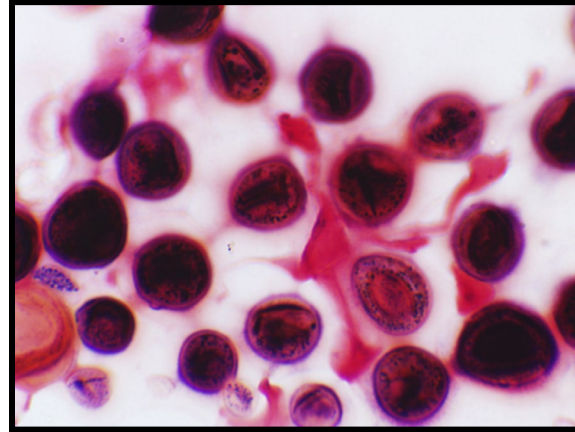
# Germ Tube Test

- Rapid test for identification of *Candida albicans*
  - *Candida dubliniensis* also germ tube positive
- Yeast cells inoculated in serum form “germ tubes” within 2 hours
- “Tube” is continuous extension from yeast cell; no septum exists
  - Distinguishes germ tubes from pseudohyphae



# *Cryptococcus neoformans/gattii*

- Cells suspended in India ink appear as different sized round cells with clear halo (capsule)
  - Inflammatory cells uniform size
- Budding may be seen
- India ink stain not commonly used today
  - Replaced with more sensitive cryptococcal antigen test



# *Malassezia* species

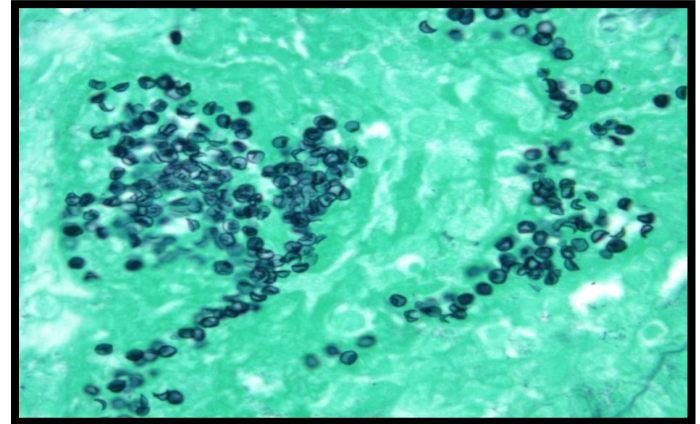
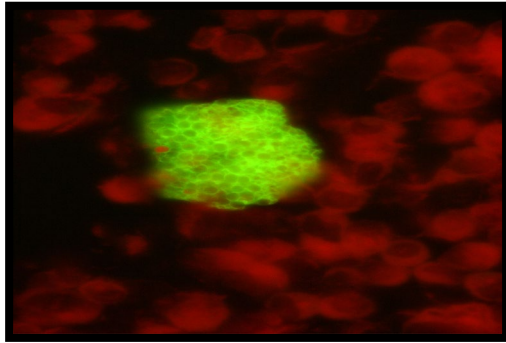
- Grow on skin surface
  - Pityriasis versicolor
  - Rarely, systemic infection
- Small yeast cells have prominent “collarette” that forms where daughter cells bud
- Short hyphal elements may be seen in skin scrapings (“spaghetti and meatballs”)
- *Malassezia furfur* culture requires supplementation of media with lipids (e.g., olive oil).



Fluorescent calciifluor white stain of *Malassezia furfur* showing prominent collarette (arrow)

# *Pneumocystis jirovecii*

- Formerly considered a parasite, now classified as a fungus
- Two developmental forms observed in human tissues
  - Trophozoite
  - Cyst



## **Direct Fluorescent Antibody Test**

- Fluorescein-conjugated monoclonal antibodies against *Pneumocystis* stain cyst wall green with contents being unstained
- Trophozoites appear as small polygons or spheres outlined in green

## **Methenamine Silver Stain**

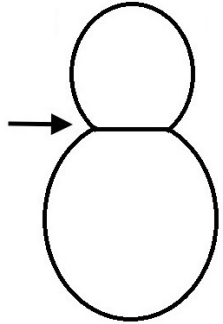
- Performed in surgical pathology
- Cysts stain brown-black
- Trophozoites do not stain

# Dimorphic Fungi

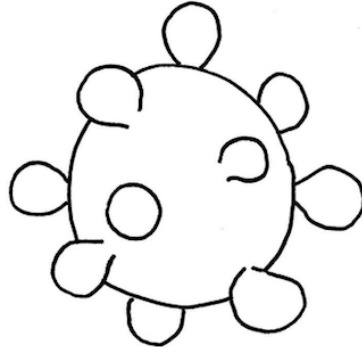
- Exist in two forms
  - Yeasts at body temperature (except *Coccidioides immitis/posadasii* which forms a spherule)
  - Molds at room temperature (or 30°C)
- Most commonly isolated dimorphic fungi, US:
  - *Histoplasma capsulatum*
  - *Blastomyces dermatitidis*
  - *Coccidioides immitis/posadasii*
  - *Sporothrix schenckii*



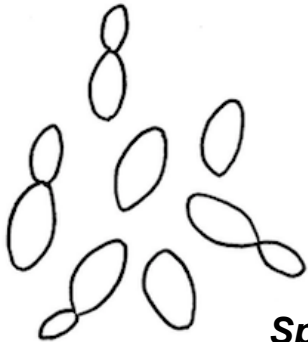
# Dimorphic Fungi (35°C)



**Blastomyces dermatitidis**  
Broad-based budding yeast

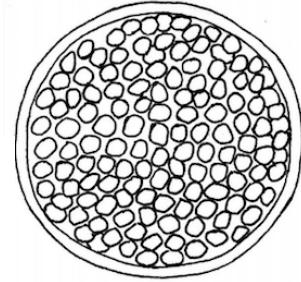


**Paracoccidioides brasiliensis**  
Mariner's wheel

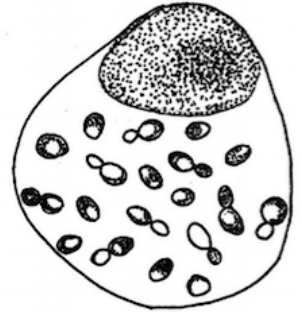


**Sporothrix schenckii**  
Elongated cigar-shaped yeasts

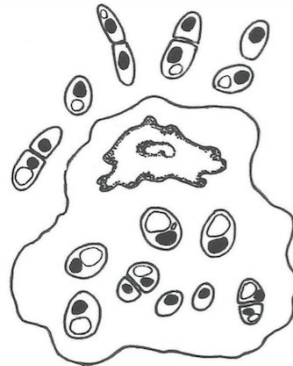
**Coccidioides immitis/posadasii**  
Spherules containing endospores



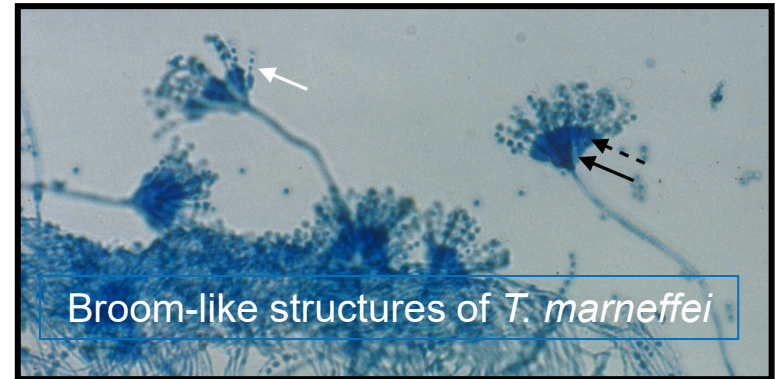
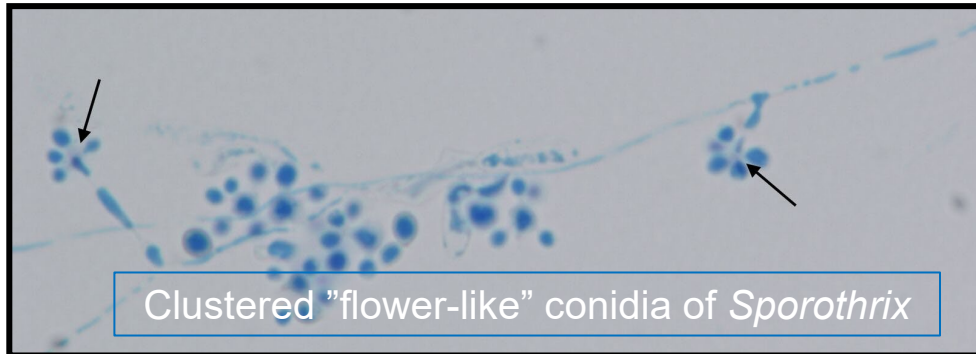
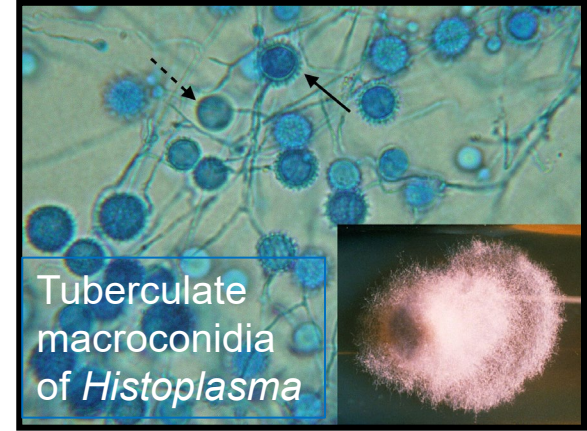
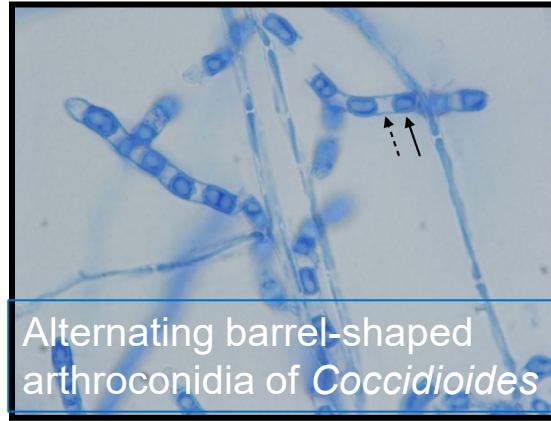
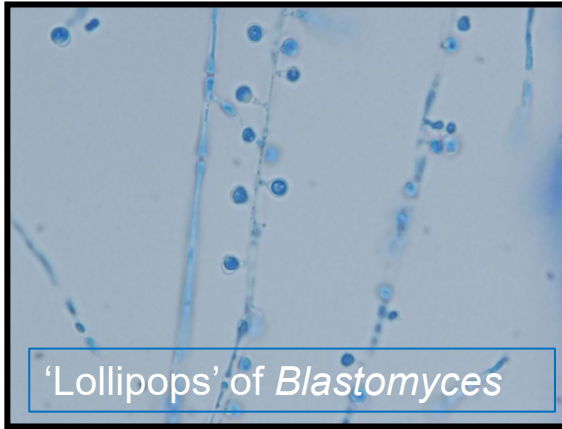
**Histoplasma capsulatum**  
Small, intracellular yeasts



**Talaromyces marneffeii**  
Elongated non-budding yeasts, divides by binary fission

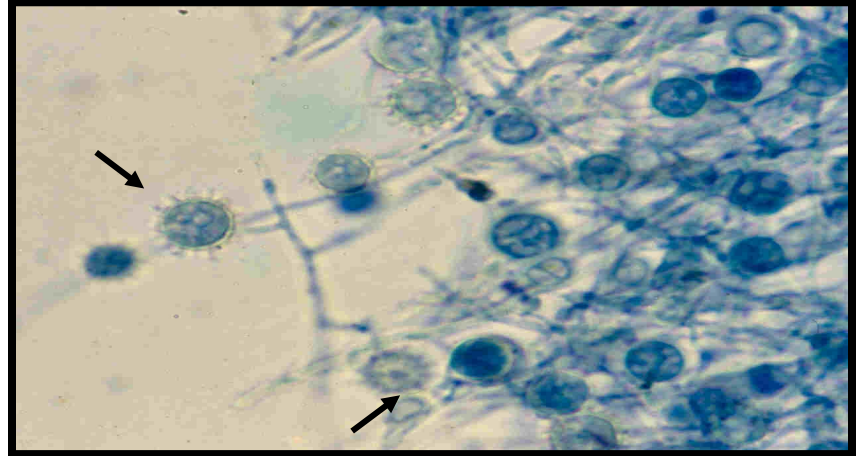
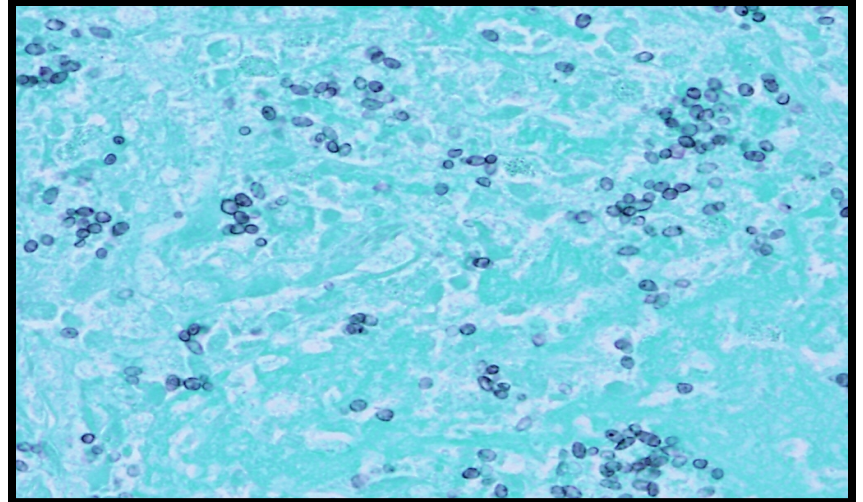


# Dimorphic Fungi (25°C)



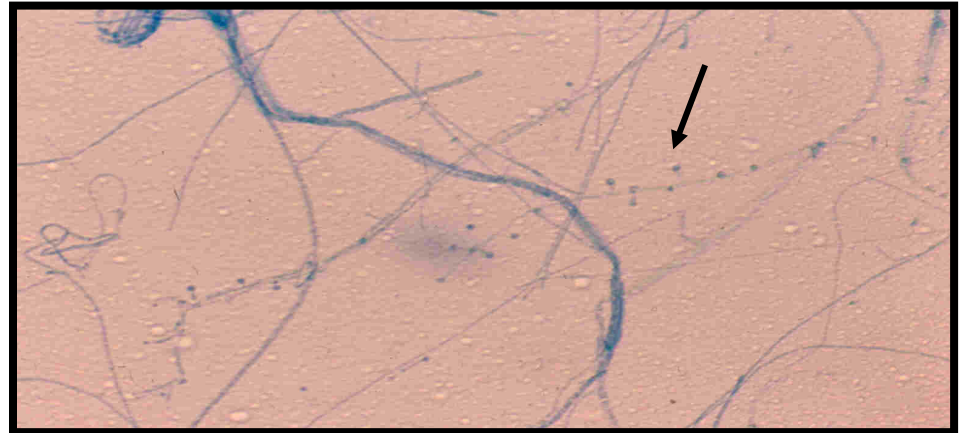
# *Histoplasma capsulatum*

- Forms small (2-4  $\mu\text{m}$ ) yeast cells in tissue (top, silver stain)
- Forms filamentous forms in culture (bottom, lactophenol cotton blue stain)
  - Note thin hyphae, microconidia, and large macroconidia (arrows) with knobby (tuberculate) surfaces



# ***Blastomyces dermatitidis***

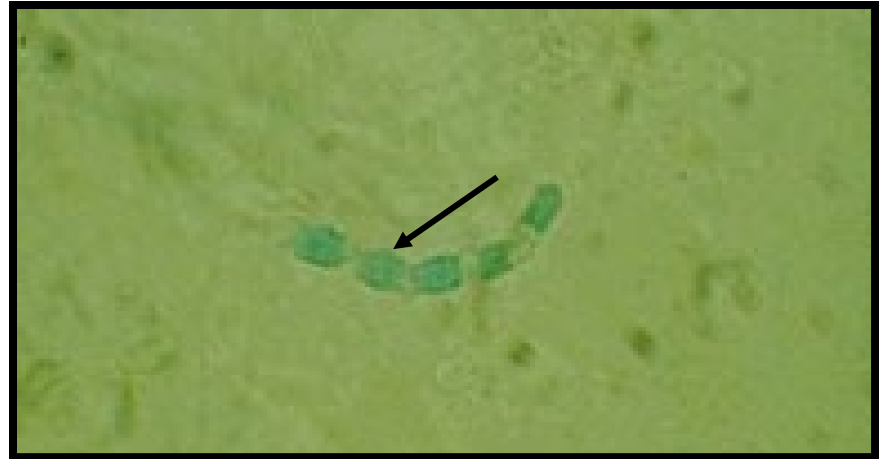
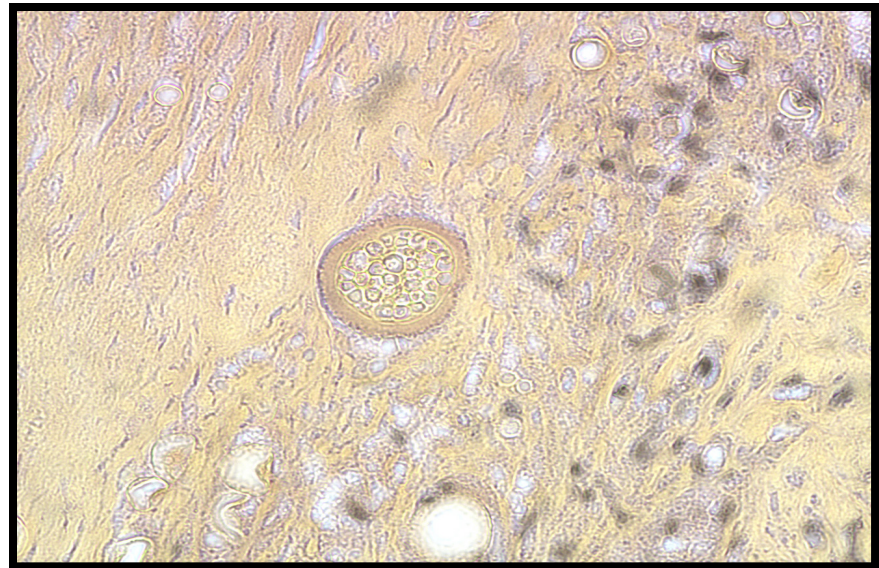
- Forms large (8-15  $\mu\text{m}$ ) yeast cells in tissue and hyphal forms in culture at ambient temperature
- Yeasts (top) have thick wall and form a broad base where daughter cell buds
- Mold form (bottom) has thin hyphae with numerous small microconidia attached to hyphae by thin branches (resembles lollipops)





# *Coccidioides* *immitis/posadasii*

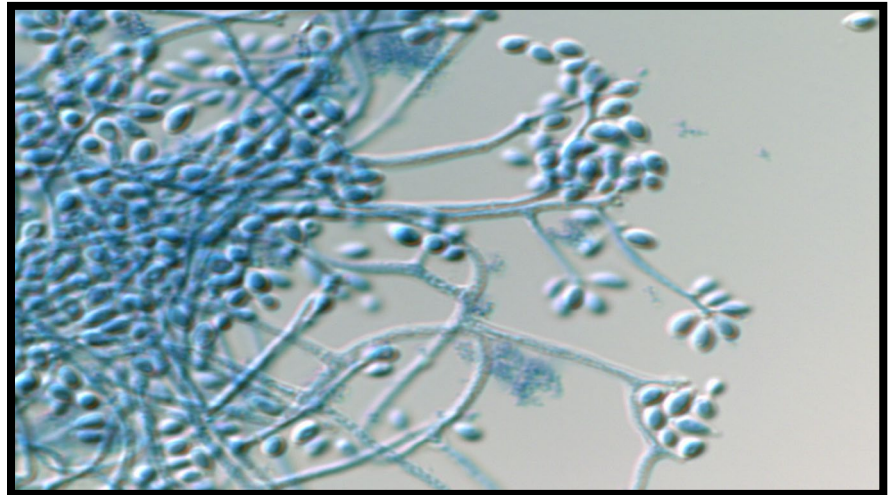
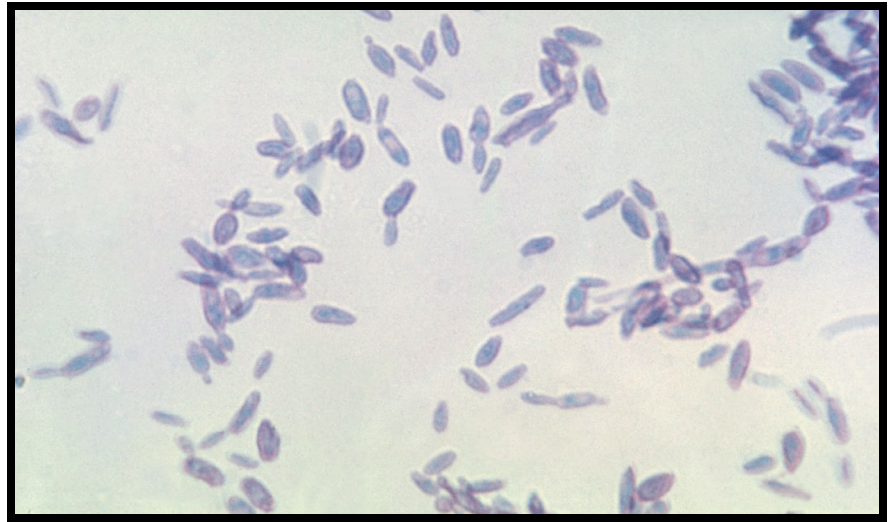
- Forms large (up to 120  $\mu\text{m}$ ), endospore filled “**spherules**” (top) in tissues
- Forms filamentous forms at room temperature (30°C)
- Barrel shaped, spore-like structures (**arthroconidia**) formed in alternate hyphal cells (bottom, arrow)





# *Sporothrix schenckii*

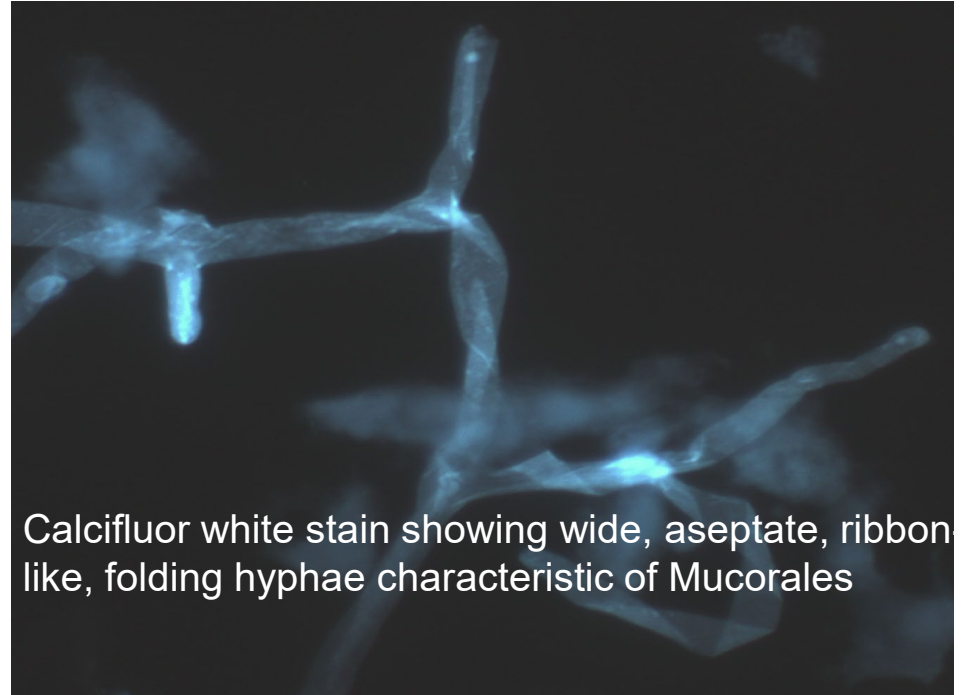
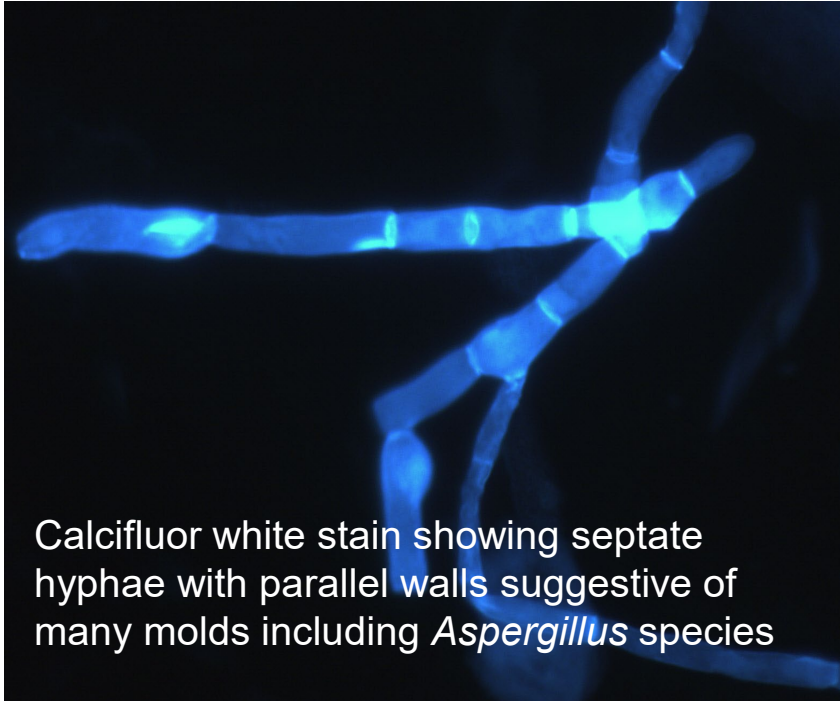
- Forms narrow based yeast cells in tissue (top)
- Forms delicate hyphae with cluster (“**flowerette**”) of conidia (spores) at end of narrow stalk (bottom)
- Conidia can be darkly pigmented



# Filamentous Fungi – Molds (Selected)

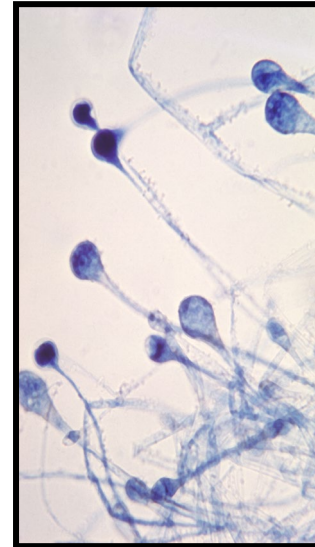
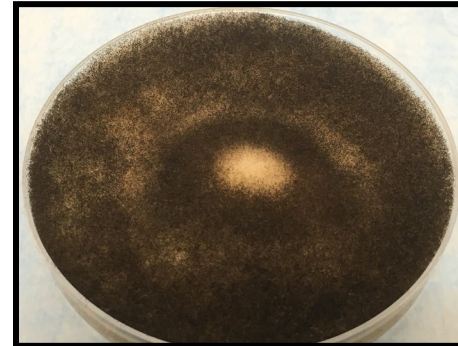
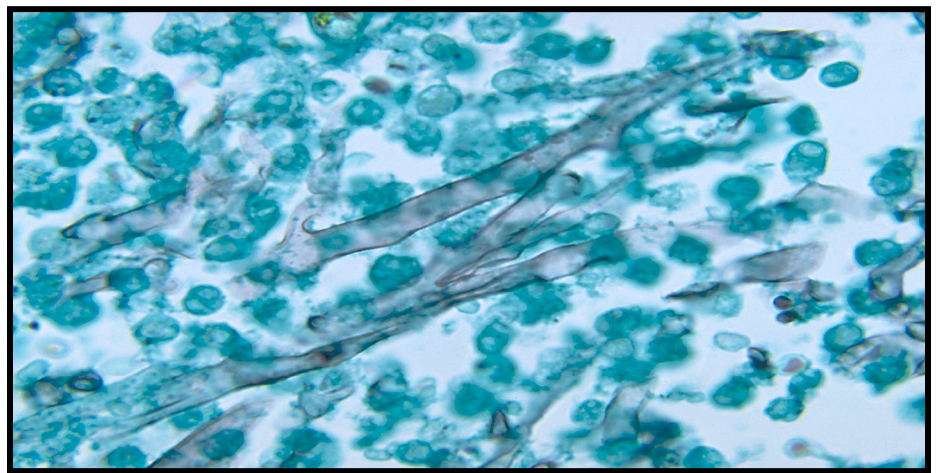
- Nonseptated molds (Mucormycetes - e.g., *Rhizopus* species, *Mucor* species, *Rhizomucor* species, *Lichtheimia* species, *Cunninghamella* species)
- Lightly colored or hyaline, septated molds
  - Opportunistic fungi (e.g., *Aspergillus* species, *Fusarium* species, *Paecilomyces* species, *Scopulariopsis* species, *Penicillium* species)
  - Dermatophytes (e.g., *Trichophyton* species, *Epidermophyton floccosum*, *Microsporum* species)
- Darkly pigmented (dematiaceous), septated molds (e.g., *Alternaria* species, *Bipolaris* species, *Curvularia* species, *Exophiala* species, *Exserohilum* species, *Scedosporium* species, *Cladosporium* species, *Phialophora* species, *Cladophialophora* species, *Ochrochonis* species)

# Septate *versus* Aseptate Hyphae



# Mucormycetes

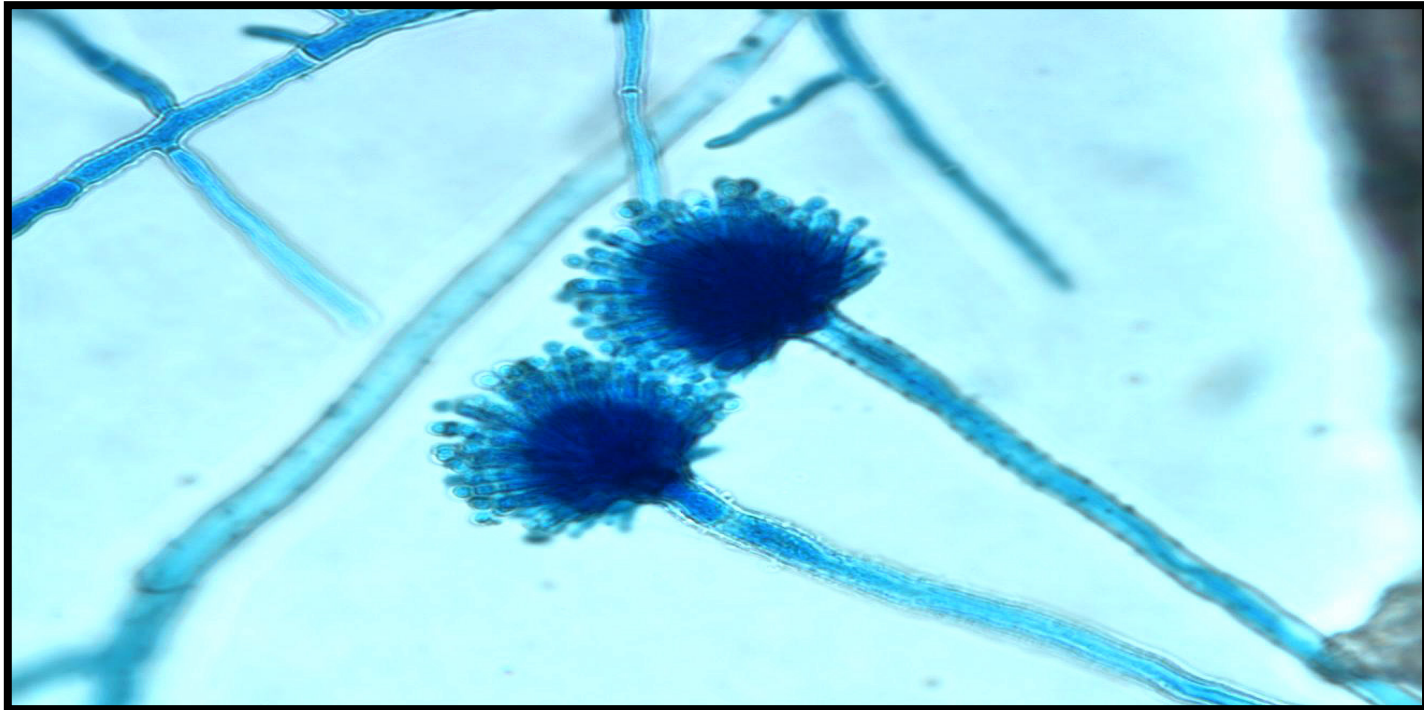
- No (or just a few) septae (divisions) within hyphae
- Grow quickly
  - “Lid lifters”, “cotton candy”
- In tissue (top; silver stain), hyphae appear broad and ribbon-like with irregular walls and irregular branching
- Bottom figures shows *Mucor* species in culture





# ***Aspergillus* species**

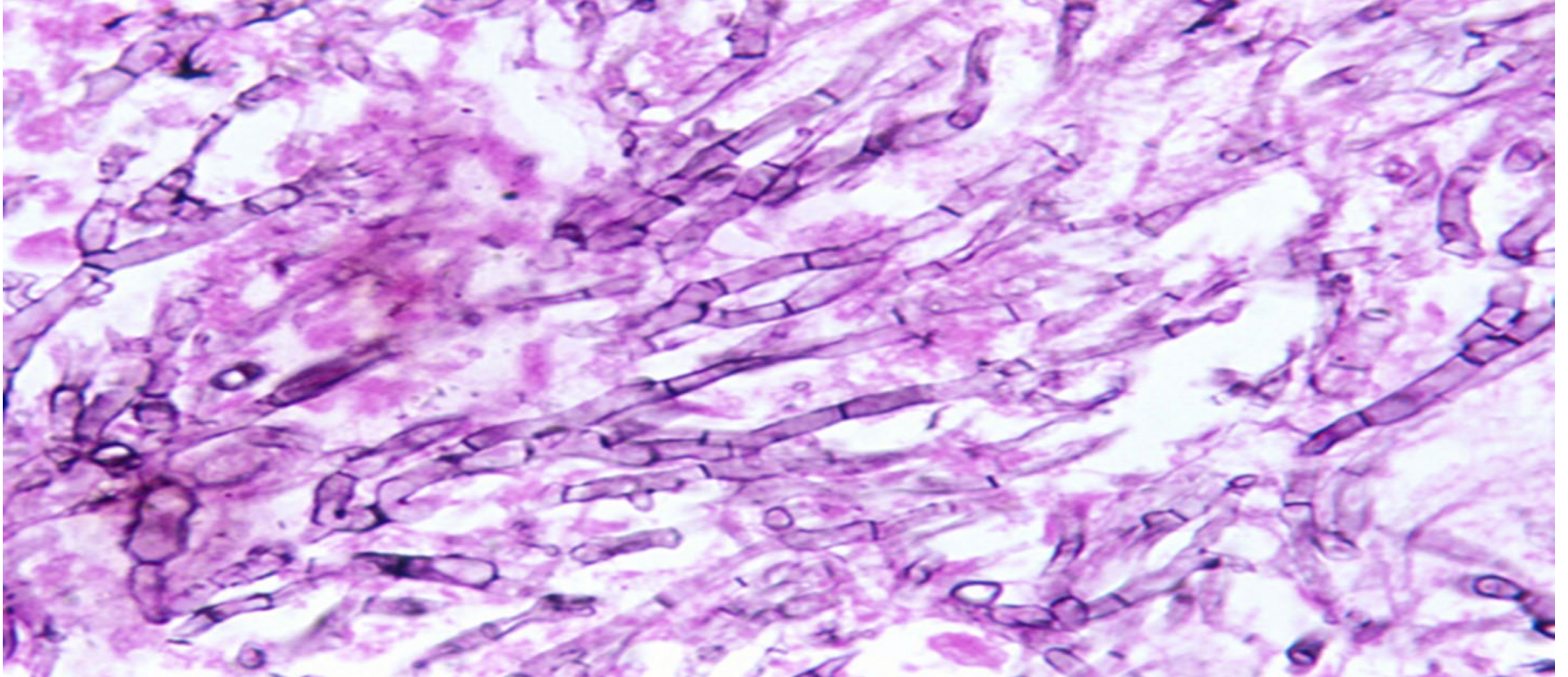
- In culture with characteristic fruiting bodies (bottom) – conidiophore covered with conidia



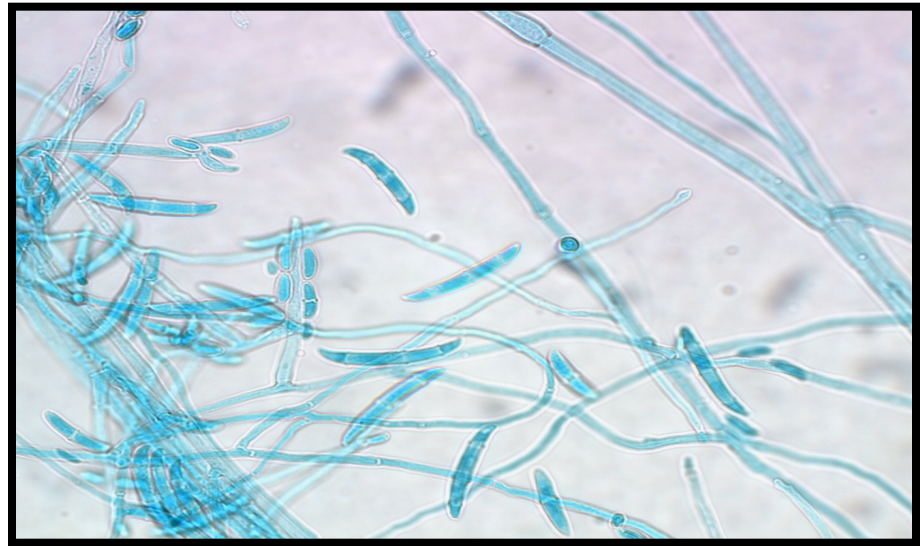
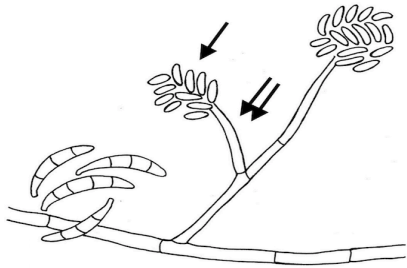


# ***Aspergillus* species**

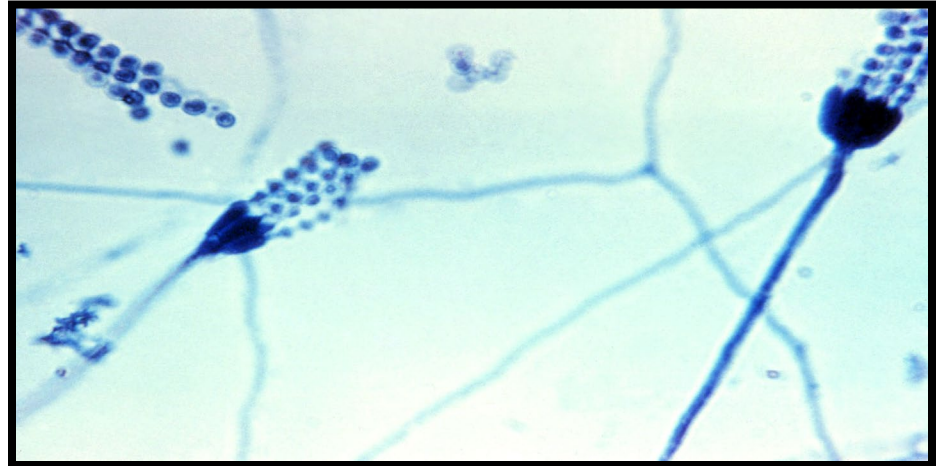
- In tissue - note uniform diameter and branching of septated hyphae [acute angle (45°) branching]



- Fusiform or sickle shaped multicelled macroconidia of *Fusarium* species (top)
  - Most likely mold to grow from blood culture bottles

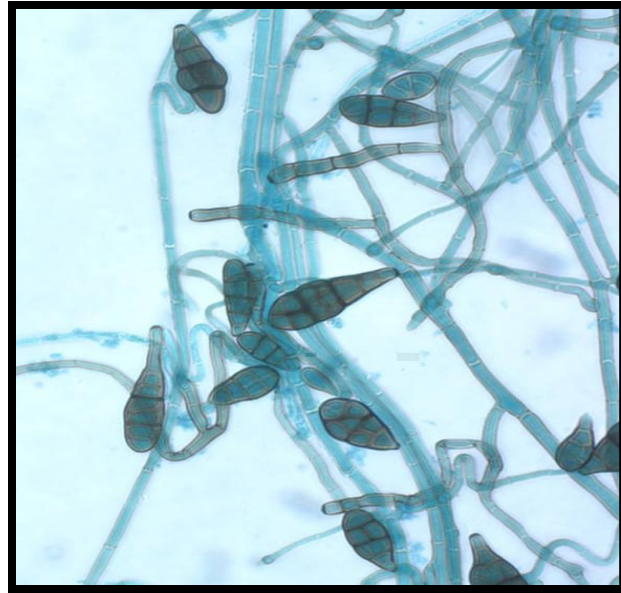


- Fruiting structures and thin hyphae of *Penicillium* species in culture (bottom)





# Dematiaceous Molds

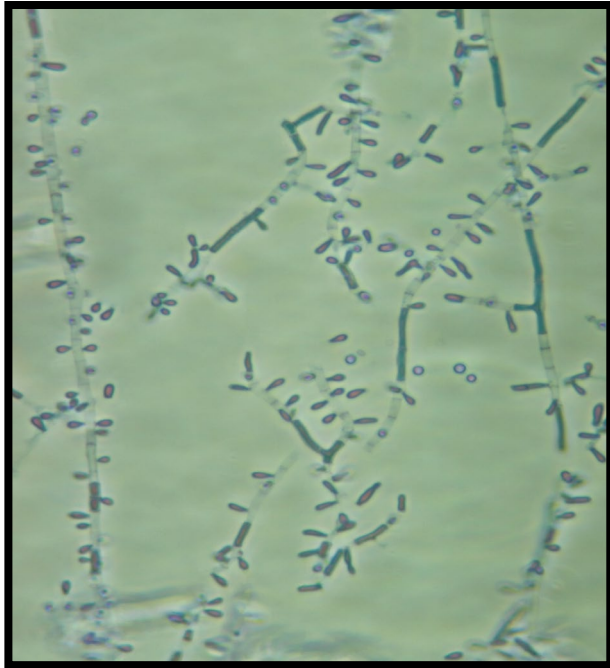


***Alternaria* species**

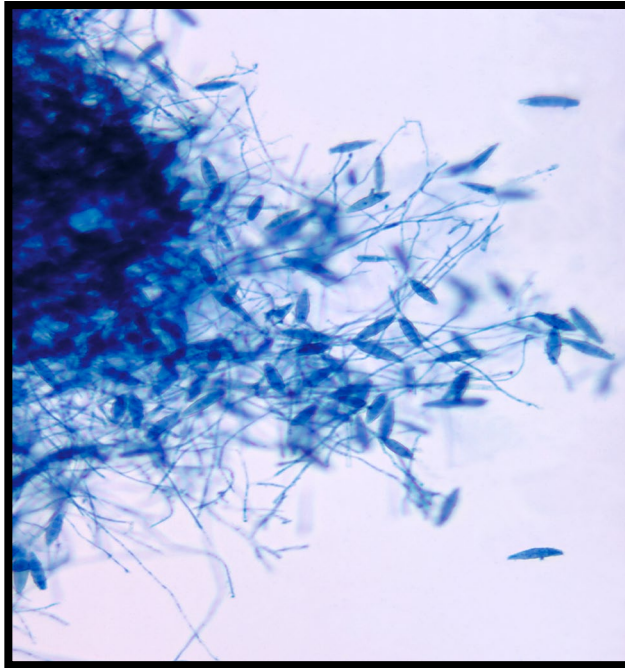
***Bipolaris* species**

***Curvularia* species**

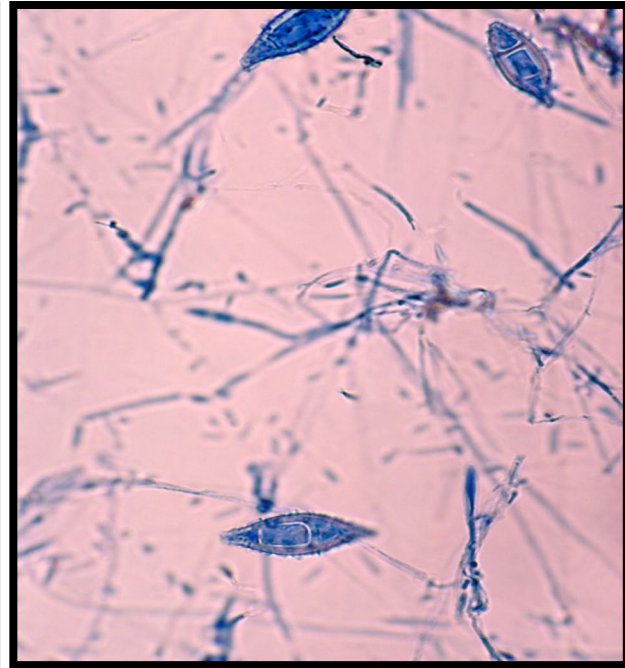
# Dermatophytes



***Trichophyton*  
species**



***Epidermophyton  
floccosum***



***Microsporum*  
species**

# Parasitology

- Protozoa
  - Amoebae (intestinal and free-living)
  - Flagellates and ciliates
  - Coccidia and microsporidia
  - *Plasmodium* species and *Babesia* species
  - *Leishmania* species and trypanosomes
- Helminths (worms)
  - Nematodes or roundworms
  - Trematodes or flukes (flatworms)
  - Cestodes or tapeworms



# Intestinal and Urogenital Amoebae, Flagellates, and Ciliates

## Parasite Amoebae

Pathogenic  
*Entamoeba histolytica*

Nonpathogenic  
*Entamoeba dispar*  
*Entamoeba hartmanni*  
*Entamoeba coli*  
*Endolimax nana*  
*Iodamoeba butschlii*  
*Entamoeba moskvoskii*  
*Entamoeba bangladeshi*

## Flagellates

*Giardia species*  
*Trichomonas vaginalis*  
*Dientamoeba fragilis* (?)  
(amoeboflagellate)

*Chilomastix mesnili*  
*Pentatrichomonas hominis*  
*Enteromonas*  
*Retortamonas*

## Ciliates

*Balantidium coli*

## Other

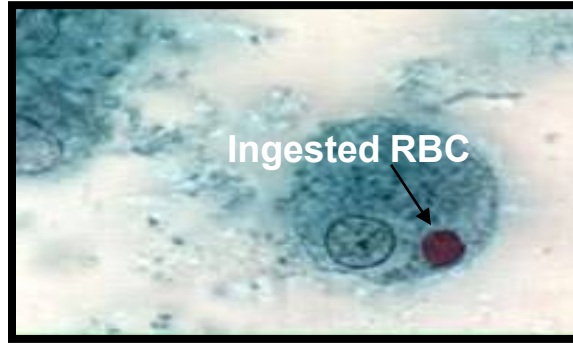
*Blastocystis hominis* (?)

# Amoebae

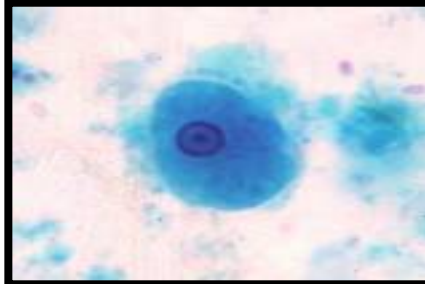
- *Entamoeba histolytica*, human pathogen
  - Nonpathogenic species
    - *E. dispar*, *E. hartmanni*, *E. nana*, *E. coli*, *E. bangladeshi*
- Most amoebae have two stages
  1. Trophozoite - actively replicating
  2. Cyst - dormant, stable (not destroyed by gastric acids, infectious)
- Detection and identification most amoebae by recognition of cyst or trophozoite forms in stool specimens
  - Exception *E. histolytica*
    - Antigen detection tests, NAATs, serology (for extraintestinal disease)
- *Blastocystis hominis* (not an amoeba) common, occasionally associated with disease

## Trophozoites

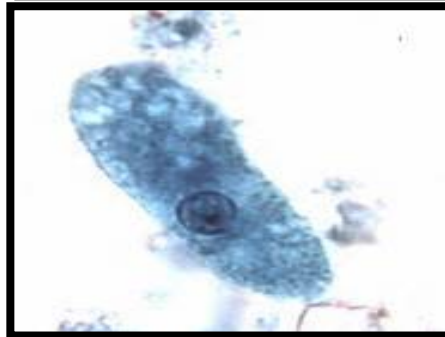
*Entamoeba histolytica*



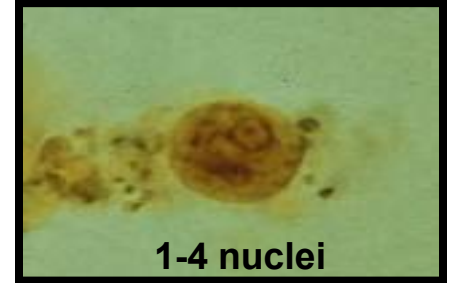
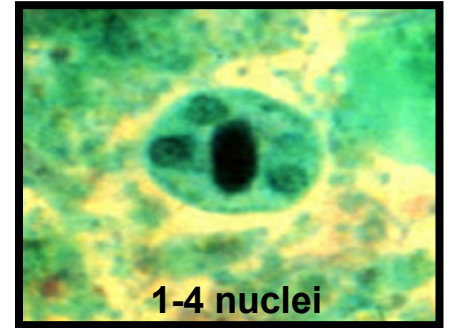
*Entamoeba hartmanni*



*Entamoeba coli*



## Cysts

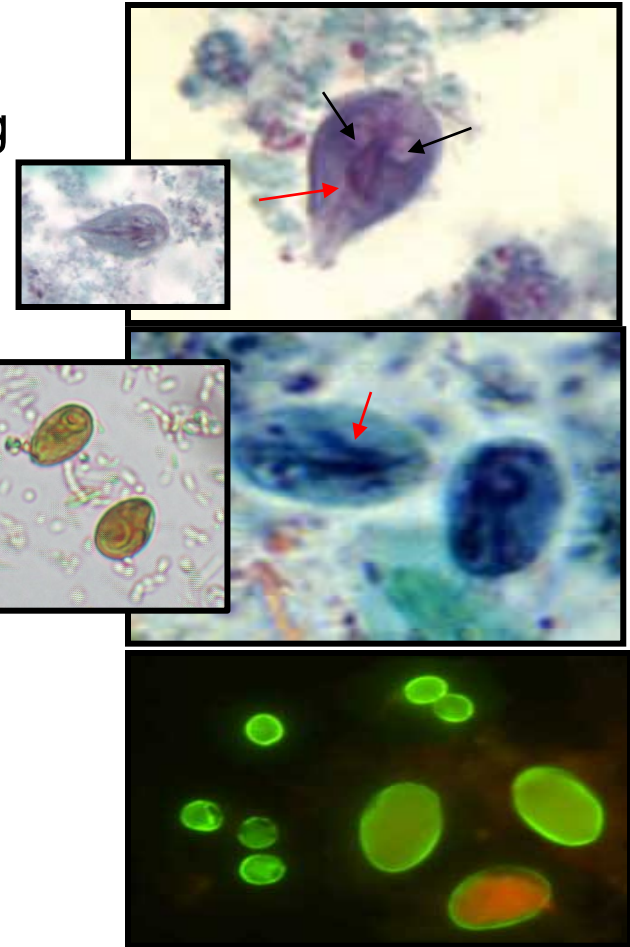


# Flagellates

- Human pathogens
  - *Giardia* species, *Trichomonas vaginalis*
- *Dientamoeba fragilis* “occasional” human pathogen - mostly insignificant colonization
- Urogenital specimen contaminated with fecal matter may contain nonpathogenic flagellates (e.g., *Pentatrichomonas*, *Enteromonas*, *Retortamonas*) which can be confused with *T. vaginalis*
- *Chilomastix mesnili* nonpathogenic flagellate occasionally found in fecal specimens

# *Giardia* species

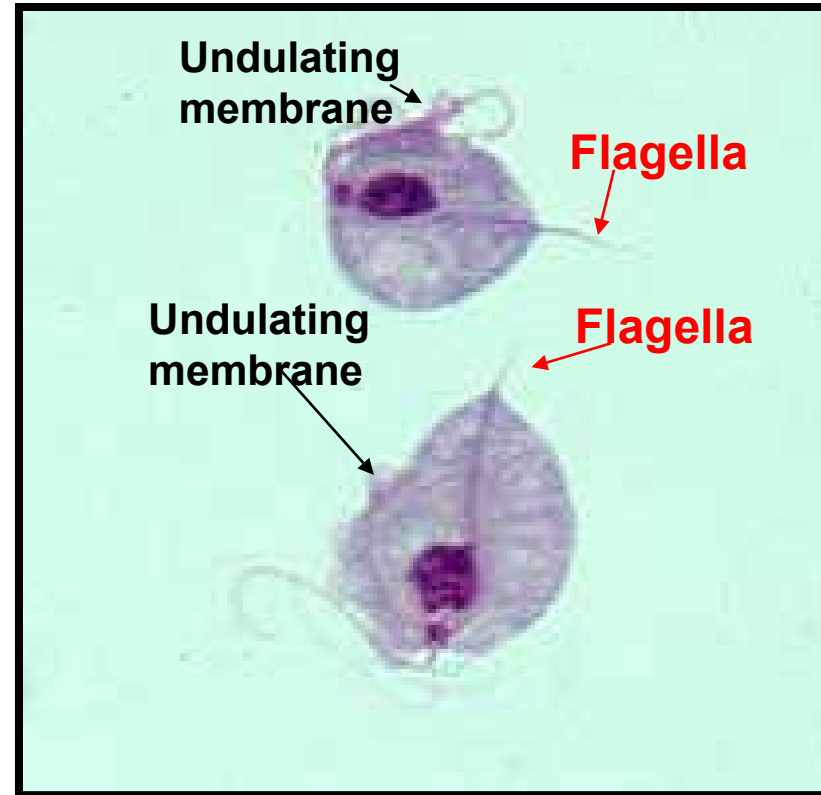
- Trophozoites (top) - pear-shaped with 2 nuclei (black arrows) within concave “sucking disk”; 2 curved rods (red arrow) lie below nuclei, 8 flagella
- Cysts (middle) - ovoid to ellipsoid; 4 nuclei present in mature cysts; central fibrils (red arrow)
- Immunoassays, direct fluorescent antibody tests (bottom), and NAATs more sensitively detect parasites in clinical specimens
  - DFA test shown uses a mixture of fluorescein-labeled antibodies to *Giardia* species (large oval cells) and *Cryptosporidium* species (small round cells)





# *Trichomonas vaginalis*

- Trophozoites only (no cyst stage)
  - Pear-shaped
- Diagnosis
  - Microscopy or culture
  - NAATs - most sensitive, specific test



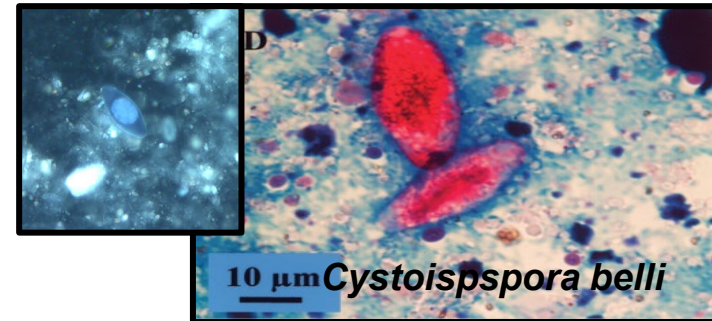
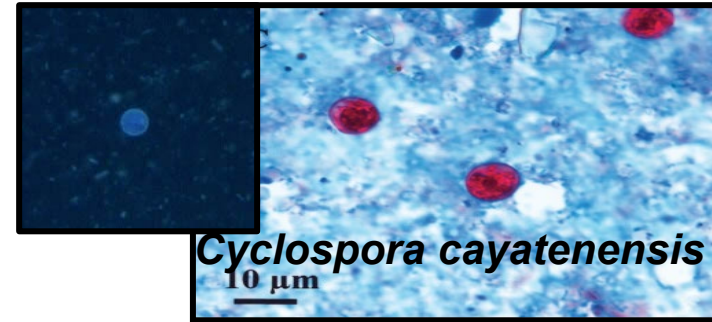
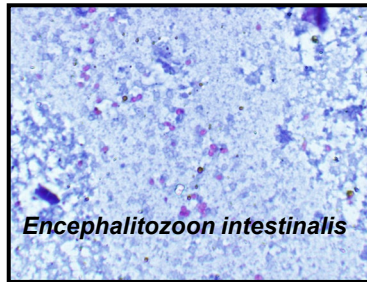
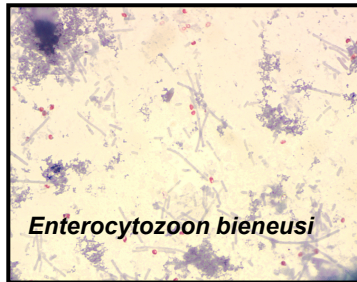
# *Balantidium coli*

- Only ciliated amoeba to cause human disease
- Trophozoite and cyst stages
- Very large



# Coccidia and Microsporidia

- Stain (weakly, non-uniformly) with acid-fast stains:
  1. *Cryptosporidium* species (4-6  $\mu\text{m}$ )
  2. *Cyclospora cayatenensis* (8-10  $\mu\text{m}$ )
  3. *Cystisosporea belli* (14x30  $\mu\text{m}$ )
- Most do not stain with traditional Ova and Parasite (O&P) stains (e.g., Trichrome stain).
- *C. cayatenensis* and *Cystisosporea belli* autofluoresce with ultraviolet light
- Microsporida (1-4  $\mu\text{m}$  – similar to bacteria) stain with chromotrope stains (e.g., strong Trichrome blue - shown); NAATs or electron microscopy



# Blood and Tissue Protozoa

- *Plasmodium* species
  - *P. falciparum*, *P. vivax*, *P. ovale*, *P. malariae*, *P. knowlesi*
- *Babesia* species
  - *B. microti*, *B. duncani*, MO-1, *B. divergens* (Europe)
- *Leishmania* species
  - *L. donovani*, *L. tropica*, *L. major*, etc.
- *Trypanosoma* species
  - *T. cruzi*, *T. brucei rhodesiense*, *T. brucei gambiense*
- Commonly detected by
  - Giemsa stain of blood
  - Hematoxylin and eosin (H&E) stain of tissues

# *Plasmodium* species

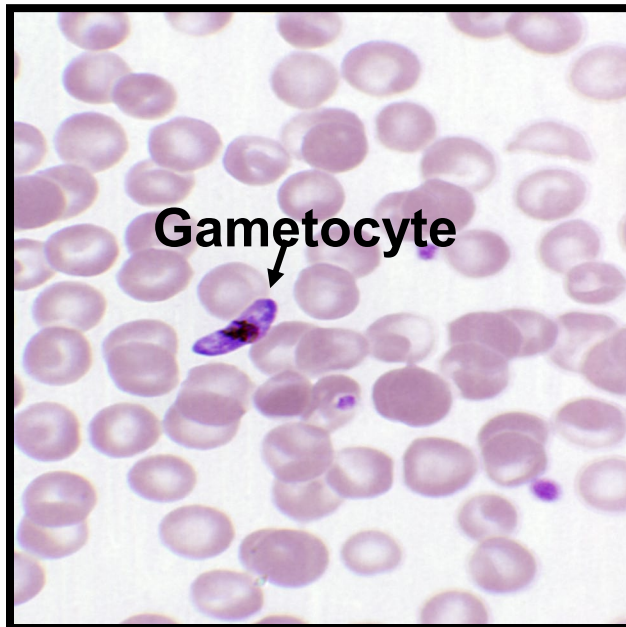
- Gold standard = thick and thin smears, **Giemsa stain**
- **Thick smear for increased sensitivity** (more blood volume examined because RBCs lysed)
- **Thin smear for organism species identification** (structures preserved in RBCs)

	<i>P. falciparum</i>	<i>P. vivax</i>	<i>P. ovale</i>	<i>P. malariae</i>	<i>P. knowlesi</i>
<b>RBCs</b>	Any	Young	Young	Old	Any
<b>RBC Size</b>	Normal	Bigger	Normal to Big	Small to Normal	Normal
<b>Stippling</b>		<b>Schuffners Dots</b>	<b>Schuffners Dots</b>		
<b>Stages</b>	<b>Rings and Gametocytes</b>	All	All	All	All
<b>Multiple Rings</b>	<b>Common</b>				Common
<b>Troph</b>	<b>Delicate; 2 chromatin dots; applique form</b>			<b>Band shape</b>	2 chromatin dots
<b>Schizonts</b>		12-24	8-12	<b>6-12 (rosette)</b>	10-16
<b>Gametocyte</b>	<b>Banana shaped</b>				

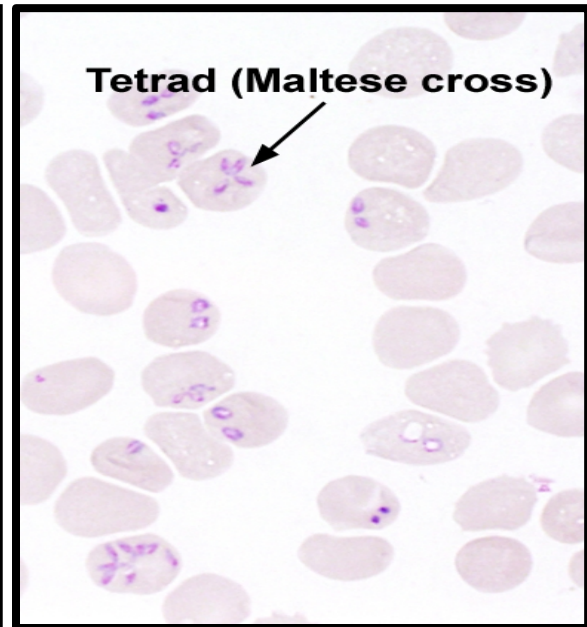




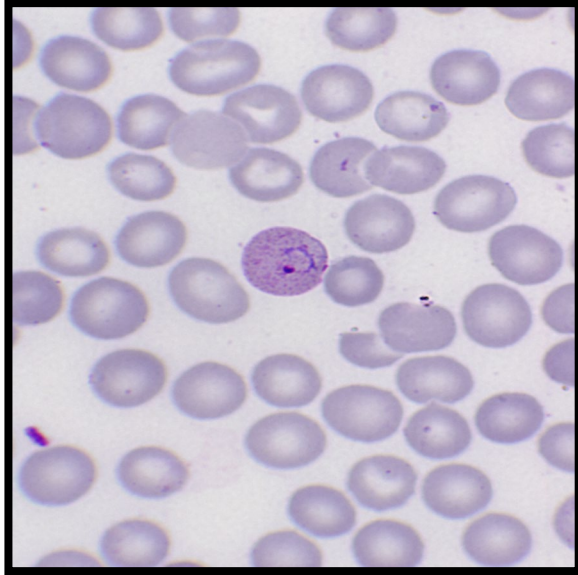
*P. falciparum*



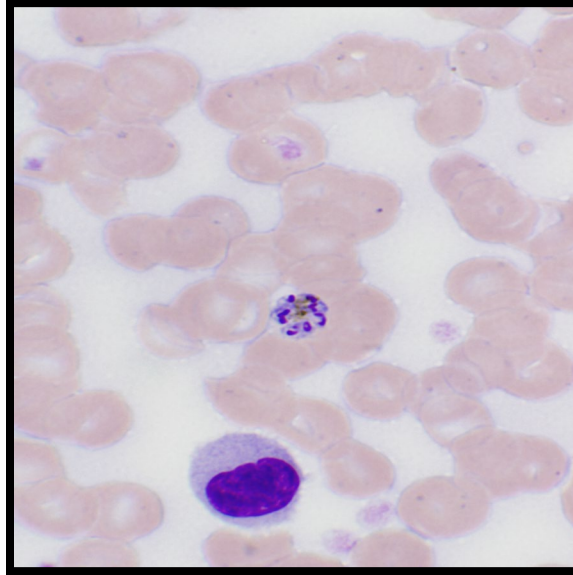
*P. falciparum*



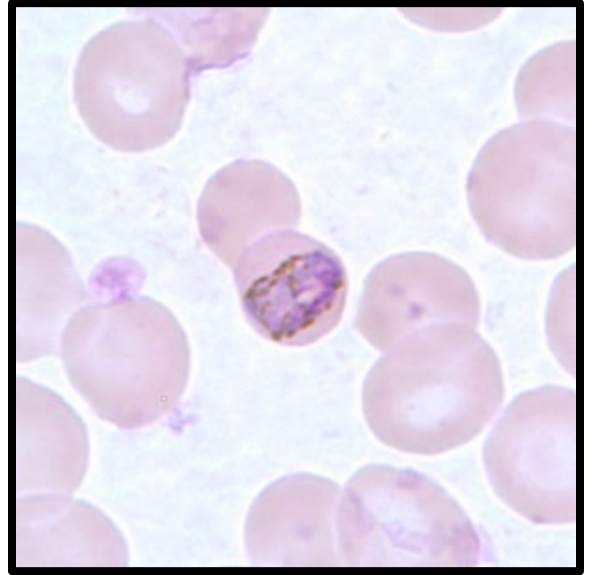
*Babesia microti*



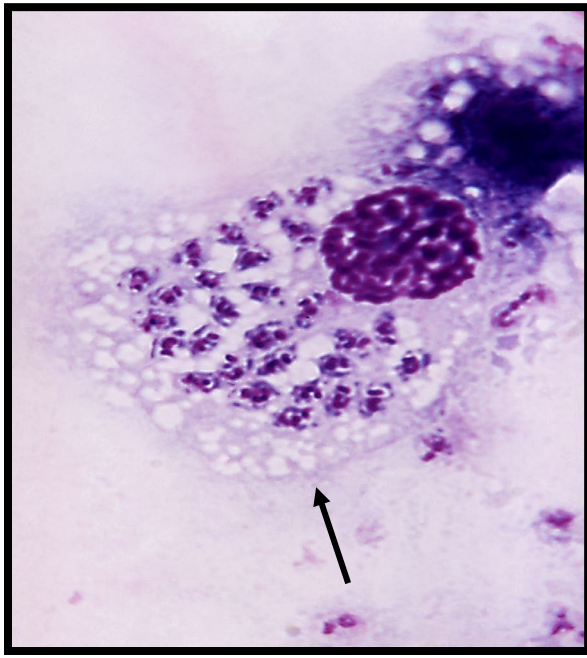
***P. vivax***  
**(amoeboid trophozoite)**



***P. malariae***  
**(“rosette” schizont)**

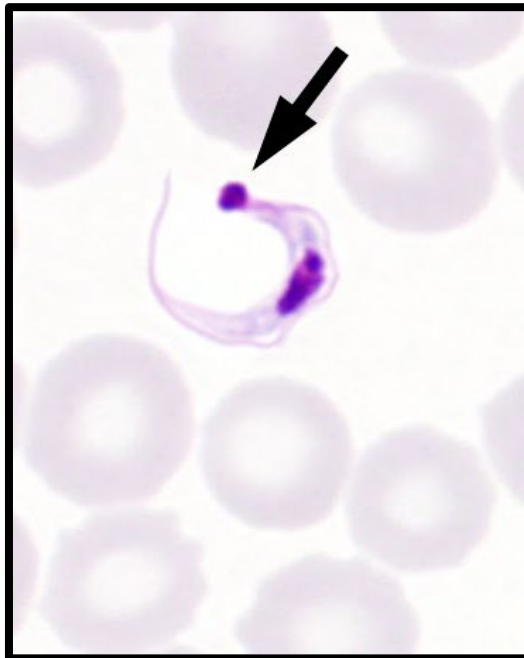


***P. malariae***  
**(band form trophozoite)**

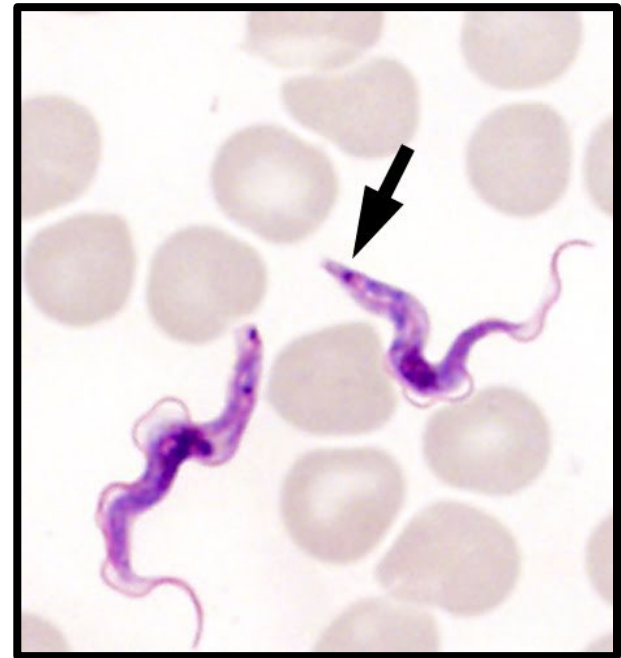


***Leishmania*  
amastigotes in tissue**

→ may be confused with *Histoplasma* –  
look for strongly staining kinetoplast



**“C” curve and large  
kinetoplast of  
*Trypanosoma cruzi*  
promastigote in  
peripheral blood**



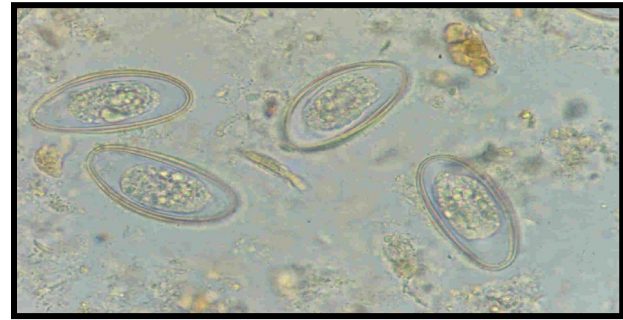
**“S” curve and small  
kinetoplast of  
*Trypanosoma brucei*  
promastigote in  
peripheral blood**

# Nematodes (Roundworms)

- **Intestinal nematodes**
  - *Enterobius vermicularis* (pinworm)
  - *Trichuris trichiura* (whipworm)
  - *Ascaris lumbricoides*
  - *Strongyloides stercoralis* (threadworm)
  - *Necator americanus*, *Ancylostoma duodenale* (hookworms)
- **Blood nematodes**
  - *Brugia malayi*, *Wuchereria bancrofti* (elephantiasis)
  - *Loa loa* (African eye worm)
  - *Onchocerca volvulus* (onchocerciasis or river blindness)
- **Tissue nematodes**
  - *Trichinella spiralis* (trichinosis)
  - *Toxocara canis* (visceral larva migrans)
  - *Ancylostoma caninum* (cutaneous larva migrans)

*Enterobius vermicularis* (pinworm) eggs

- Anal cellophane tape test
- Embryo surrounded by colorless shell, shell flattened on one side



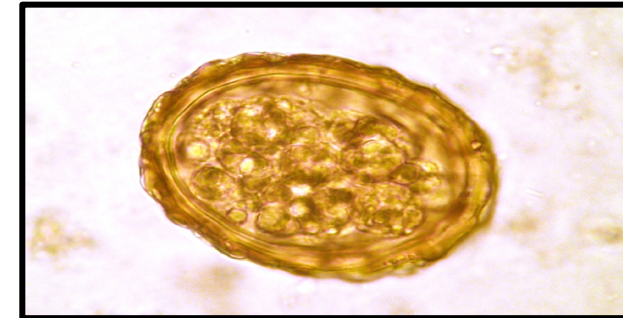
*Trichuris trichiura* (whipworm) egg

- Stool
- Naturally stained yellow-brown, with thick wall, clear polar plugs at each end



*Ascaris lumbricoides* egg

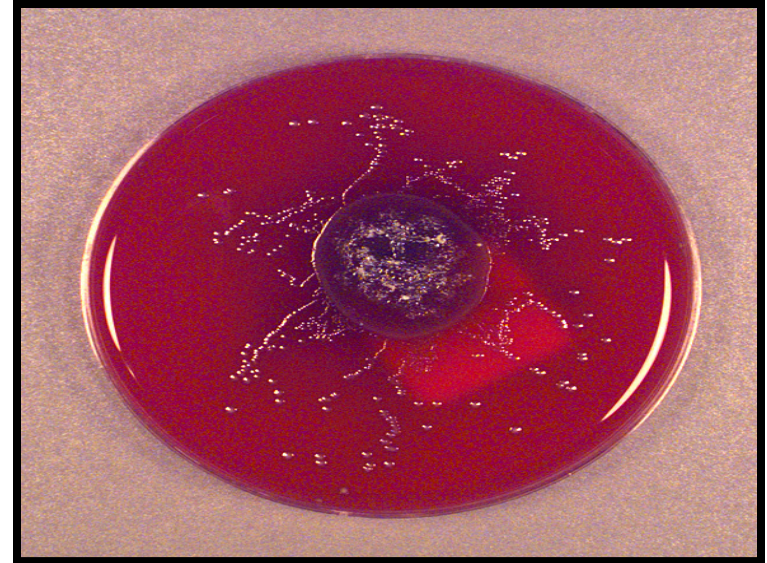
- Stool
- Either fertile (shown) or infertile
  - Infertile eggs larger and more oval shaped
- Yellow-brown color with thick, rough shell
- Adult worms occasionally passed in stool





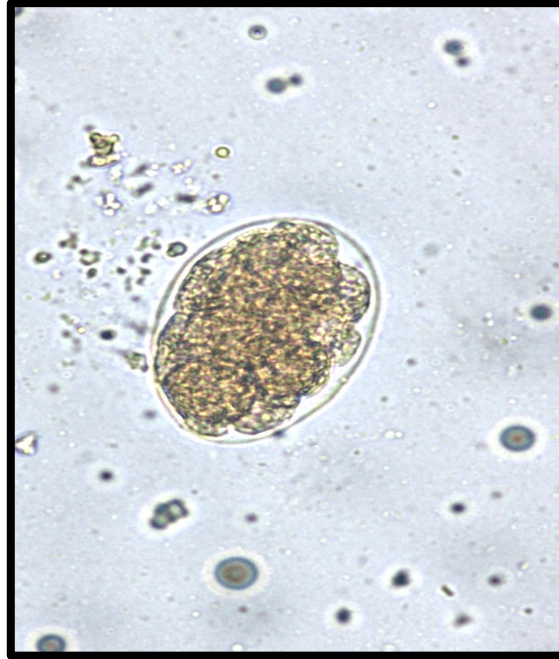
# *Strongyloides stercoralis*

- Larvae - two forms
  1. **Rhabditiform** (in stool)
  2. **Filariform** (infectious stage that develops in soil and occasionally in patient (leads to autoinfection))
- Larvae can be detected microscopically (top) or by placing feces on plate and detecting migrating larvae when they leave a trail of bacterial colonies (bottom)



# *Ancylostoma duodenale* (Old World) and *Necator americanus* (New World) Hookworms

- Eggs have thin, colorless shell surrounding developing larva
- If stool specimens left at room temperature, larvae can hatch and will resemble *Strongyloides* larvae

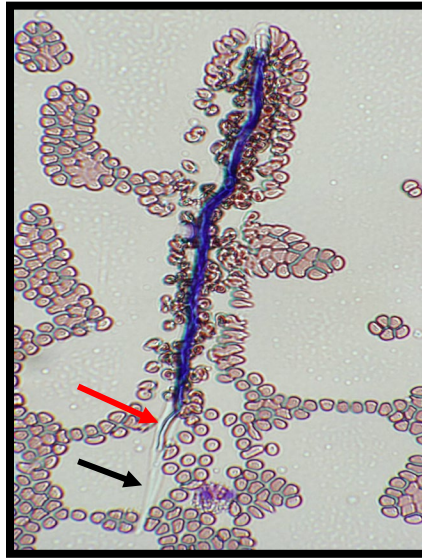


## *Wuchereria bancrofti*



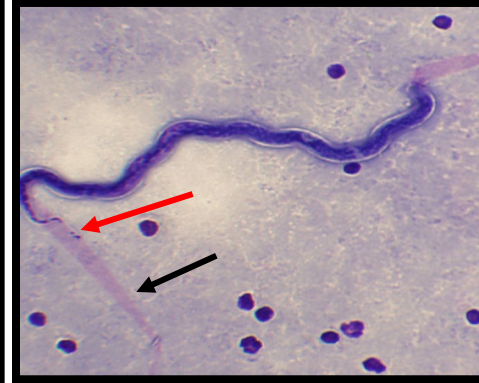
- Large microfilariae
- Transparent sheath
- Nuclei do not extend to tip of tail (red arrow)

## *Loa loa*



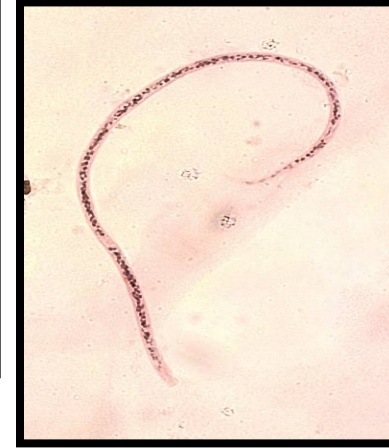
- Small microfilariae
- Sheath (black arrow)
- Nuclei extend to tip of tail (red arrow)

## *Brugia malayi*



- Small microfilariae
- Sheath (black arrow)
- Gap between last and next to last nuclei in tail (red arrow)

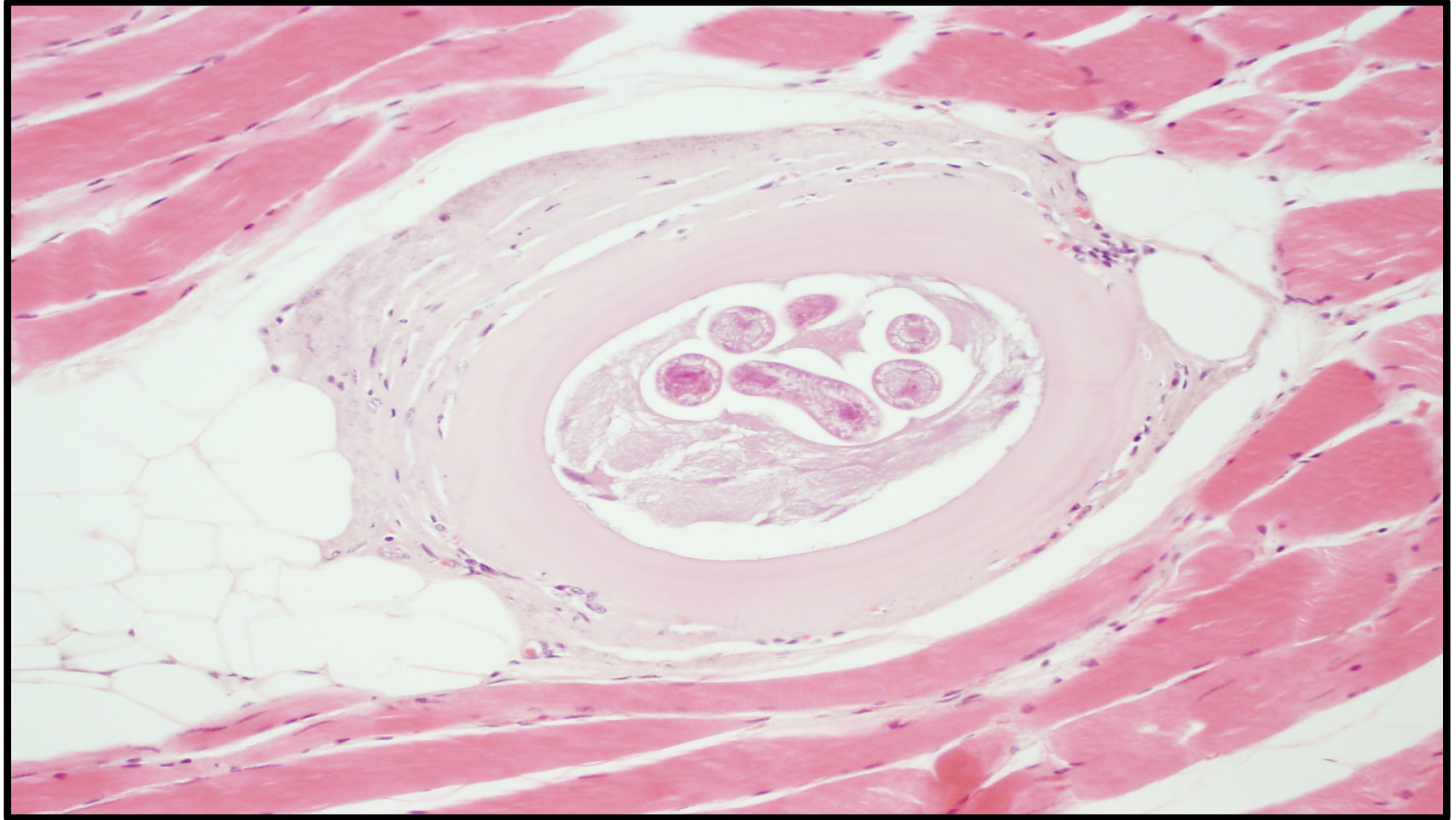
## *Onchocerca volvulus*



- Adults mature in subcutaneous nodules
- Microfilariae in skin
  - No sheath
  - Nuclei do not extend to tip of tail



# *Trichinella spiralis*



# Trematodes (Flatworms)

- **Intestinal trematode**
  - *Fasciolopsis buski*
- **Tissue trematodes**
  - *Fasciola hepatica*
  - *Clonorchis sinensis* (Chinese liver fluke)
  - *Paragonimus* species (lung fluke)
- **Schistosomes**
  - *Schistosoma mansoni*
  - *Schistosoma japonica*
  - *Schistosoma haematobium*



# Parasites with Opercula

*Clonorchis sinensis*



Acquired by ingestion of raw/undercooked fish

- Eggs
  - 17-30x13-18 $\mu$ m
  - Prominent operculum (black arrow) at small end
  - Knob (red arrow) opposite end

*Fasciola hepatica*



Acquired by ingestion of aquatic vegetation (e.g., watercress)

- Eggs
  - 130-150x63-90 $\mu$ m
  - Inconspicuous operculum

*Fasciolopsis buski*



Acquired by ingestion of aquatic vegetation (e.g., water chestnuts)

- Eggs
  - 130-140x80-85 $\mu$ m
  - Inconspicuous operculum

*Paragonimus species*



Acquired by eating raw or inadequately cooked crabs and crayfish

- Eggs
  - 80-120x45-70 $\mu$ m
  - Prominent operculum (arrow)

# ***Schistosoma* species**

Large, thin-shelled eggs with no operculum



*Schistosoma mansoni*  
(prominent lateral spine)



*Schistosoma hematobium*  
(prominent terminal spine)



*Schistosoma japonicum*  
(inconspicuous spine)

# Cestodes (Tapeworms)

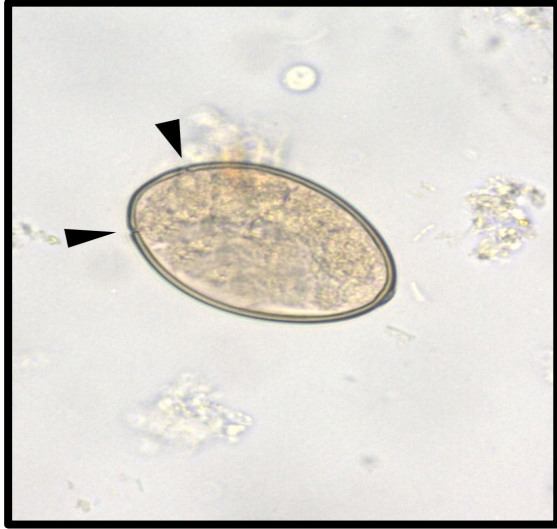
- **Intestinal cestodes**

- *Diphyllobothrium latum* (fish tapeworm)
- *Taenia solium* (pork tapeworm)
- *Taenia saginata* (beef tapeworm)
- *Hymenolepis nana* (dwarf tapeworm)
- *Hymenolepis diminuta* (rat tapeworm)
- *Dipylidium caninum* (dog tapeworm)

- **Tissue cestodes**

- *Echinococcus granulosus* (hydatid disease)
- *Taenia solium* (cysticercosis)

***Diphyllobothrium latum***



**Egg**

- Oval shape
- Operculum (arrowheads)
- Absence of hooks

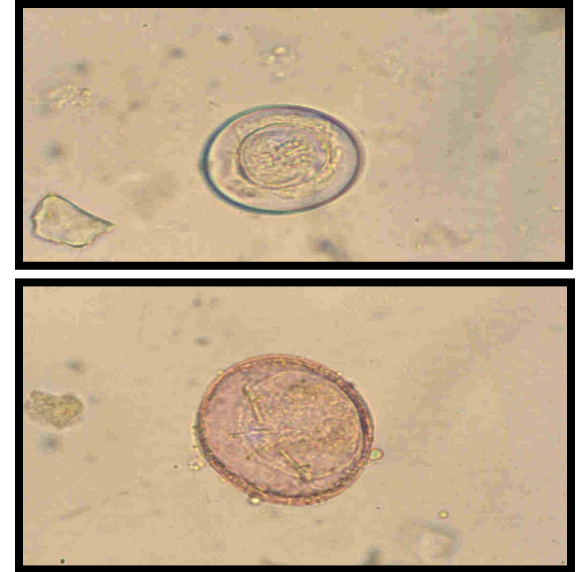
***Taenia saginata*,  
*Taenia solium***



**Eggs identical**

- Thick wall
- Outer membrane may or may not be present

***Hymenolepis nana*,  
*H. diminuta***



**Eggs**

- Hooks
- Outer membrane
- *H. nana* (top) eggs smaller than *H. diminuta* (bottom) eggs