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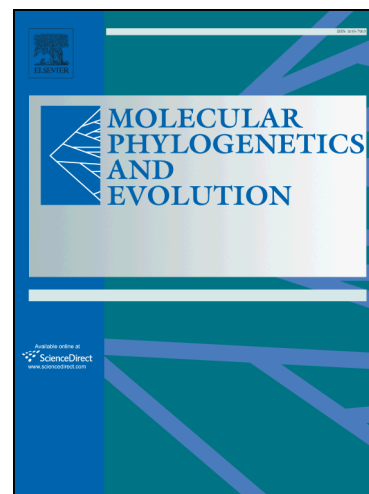
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**Myrteae phylogeny, calibration, biogeography and diversification patterns: increased understanding in the most species rich tribe of Myrtaceae.**

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#### ABSTRACT

Myrteae (c. 2500 species; 51 genera) is the largest tribe of Myrtaceae and an ecologically important groups of angiosperms in the Neotropics. Systematic relationships in Myrteae are complex, hindering conservation initiatives and jeopardizing evolutionary modelling. A well-supported and robust phylogenetic hypothesis was here targeted towards a comprehensive understanding of the relationships within the tribe. The resultant topology was used as a base for key evolutionary analyses such as age estimation, historical biogeography and diversification rate patterns. One nuclear (*ITS*) and seven chloroplast (*psbA-trnH*, *matK*, *ndhF*, *trnI-trnF*, *trnQ-rps16*, *rpl16* and *rpl32-trnL*) DNA regions for 115 taxa representing 46 out of the 51 genera in the tribe were accessed and analysed using maximum likelihood and Bayesian inference tools for phylogenetic reconstruction. Dates of diversification events were estimated and contrasted using two distinct fossil sets (macro and pollen) in BEAST. The subsequent dated phylogenies were compared and analysed for biogeographical patterns using BioGeoBEARS and diversification rates using BAMM. Myrteae phylogeny presents strong statistical support for three major clades within the tribe: Australasian group, *Myrtus* group and Main Neotropical Lineage. Dating results from calibration using macrofossil are an average of 20 million years older and show an early Paleocene origin of Myrteae, against a mid-Eocene one from the pollen fossil calibration. Biogeographic analysis shows the origin of Myrteae in Zealandia in both calibration approaches, followed by a widespread distribution throughout the still-linked Gondwana continents and diversification of Neotropical endemic lineages by later vicariance. Best configuration shift indicates three points of acceleration in diversification rates, all of them occurring in the Main Neotropical Lineage. Based on the reconstructed topology, several new taxonomic placements were recovered, including: the relative position of *Myrtus communis*, the placement of the *Blepharocalyx* group, the absence of generic endemism in the Caribbean, and the paraphyletism of the former *Pimenta* group. Distinct calibration approaches affect biogeography interpretation, increasing the number of necessary long distance dispersal events in the topology with older nodes. It is hypothesised that biological intrinsic factors such as modifications of embryo type and polyploidy might have played a role in accelerating shifts of diversification rates in Neotropical lineages. Future perspectives include formal subtribal classification, standardization of fossil calibration approaches and better links between diversification shifts and trait evolution.

Key words: *Eugenia*, evolution, *Myrcia*, *Myrtus*, *Psidium*, systematics.

#### Highlights:

- The present Myrteae phylogeny reports key differences among group relationships in comparison to previous published trees;
- Future nomenclatural changes are necessary in at least eight genera that proved to be either poly or paraphyletic;
- Three increases in diversification rates contribute to the high diversity of Myrteae in the Neotropics;
- Ancestral range estimation emphasizes the significance of higher latitude in the early diversification of the tribe;
- Strong evidence for long distance dispersal event in Myrteae is found only in *Rhodamnia* and *Eugenia* group.
- The only European Myrtaceae, *Myrtus communis*, is part of a mainly Neotropical clade.

## 1. Introduction

Myrtaceae is a large family of woody flowering plants represented by around 5500 accepted species, classified in 144 genera and 17 tribes (Wilson et al., 2005; Wilson, 2011; WCSP, 2016). Myrtaceae represents an old, mid-Cretaceous lineage within the order Myrtales (c. 85mya, Berger et al., 2016) and is characterized by a strong southern-hemisphere, Gondwanan distribution (Thornhill et al., 2015). Myrtaceae is an important floristic component in the areas where it is most species diverse, especially in the forests of Southeast Asia, Australia and South America (e.g. Johnson and Briggs, 1981, Kochummen et al., 1990, Oliveira-Filho and Fontes, 2000, Flora of Brazil, 2016). In Neotropical environments, all Myrtaceae diversity (excluding a single species from tribe Metrosidereae, *Metrosideros stipularis*, restricted to Chile, Pillon et al., 2015) is represented by a sole lineage: tribe Myrteae (Wilson et al., 2005, Lucas et al., 2007). Myrteae is the most diverse tribe within Myrtaceae both in number of species (c. 2500) and genera (51), representing half of the family's biodiversity (Wilson, 2011; WCSP, 2016). Myrteae species are ecologically important in many Neotropical environments due to the fleshy berries eaten by birds and mammals and the white generalist flowers that supply pollen and resources to a variety of bee species (Mori et al., 1983, NicLughadha and Proença, 1996, Gressler et al., 2006, see Figure 1). Due to its ecological importance, a growing interest has been addressed by researchers using Myrteae as a model group for evolutionary, ecological and conservation studies in Neotropical biomes (e.g. Murray-Smith et al., 2009, Lucas and Büniger, 2015, Staggemeier et al., 2015, Giaretta et al., 2015).

### 1.1 Myrteae systematics and diversity

A common barrier encountered by those wishing to study Myrteae is the problematic systematics of the group. The homogeneous morphology of flowers, fruits and vegetative characters between even distantly related Myrteae species makes taxonomy in the tribe a tiresome process even for specialists and until recently resulted in its neglect (McVaugh, 1968, Landrum and Kawasaki, 1997, Lucas et al., 2005). Recent phylogenetic systematic studies and taxonomic revision of individual clades within the tribe has improved the understanding of relationships and characterization of smaller groups (e.g. Landrum, 1981, Landrum, 1986, Proença, 1990, Grifo, 1992, Lucas et al., 2011, Murillo-A et al., 2012, Mazine et al., 2014, Staggemeier et al., 2015). However, narrower distributed genera not sampled at the molecular level until now remain phylogenetically unplaced. To place such taxa in a broader phylogenetic system is central to improve the understanding of relationships and evolution within this ecologically important tribe.

Although morphologically similar, Myrteae lineages have an uneven, heterogeneous distribution of biodiversity in terms of species per genus. Two thirds of the diversity of described species occurs in only two genera, *Eugenia* s.l. (sensu Mazine et al., 2014) and *Myrcia* s.l. (sensu Lucas et al., 2011), which are also two of the largest angiosperm genera (Frodin, 2004) with c. 1000 and 700 species, respectively (WCSP, 2016). Furthermore, these two genera have been consistently proved to be sister to species poor lineages in the tribe (Lucas et al., 2007, this study), increasing the extant diversity disparity between closely related clades.

### 1.2 Myrteae global geographic distribution

Although most extant biodiversity of Myrteae is restricted to the Neotropics, at least 15 genera (Wilson, 2011) and ca. 450 species are found in other continents. These are predominantly from Southeast Asia, Northeast Australia and the Pacific islands, including New Caledonia and New Zealand (Scott, 1978, Snow, 2000, Wilson, 2009, Snow et al., 2011, WCSP, 2016). A few species of *Eugenia* are also found in Africa, Madagascar and Mauritius (Van Wyk, 1982, van der Merwe et al.,

2005, Snow, 2008) and an additional genus, *Myrtus*, represents the only European/Northern African lineage (Lucas et al., 2007, Migliore et al., 2011). On the American continent, most species diversity is found in the rainforests and savannah of central and eastern Brazil, the Guiana shield and Caribbean (McVaugh, 1968, Mori et al., 1983, Oliveira-Filho and Fontes, 2000, Holst, 2003, Murray-Smith et al., 2009); less but still significant biodiversity is found in continental Central America and the low-land Amazon basin (Landrum, 1992, WCSP, 2016). Species diversity is relatively low in the subtropical and temperate areas of southern South-America (Patagonia) and the high altitude Andes, but these areas boast a significant array of endemic genera (e.g. *Ugni*, *Amomyrtus*, *Legrandia*, *Luma*; Landrum, 1981, 1986, 1988).

Previous phylogenetic analyses consistently showed *Myrtus* representing a sister clade to all of the extant Myrteae (Lucas et al., 2005, 2007, Biffin et al., 2010, Thornhill et al., 2015). In these studies, most Australasian genera also group in a distinct clade, sister to the that containing all Neotropical clades (Lucas et al., 2005, 2007). The relative position of these clades in the tribe, in addition to biogeographical analysis in a broader Myrtaceae context (Thornhill et al., 2015) shows that Australia represents the most likely ancestral range in the family and that Neotropical genera are likely a result from a more recent event of vicariance between Australia and South America, while the distribution of *Myrtus* is attributed either to a previous wider distribution of the tribe or to an old long distance dispersion and establishment (henceforward coined LDDE) event.

### 1.3 Study aims

Despite recent progress in understanding relationships within Myrteae using molecular tools (e.g. Lucas et al., 2011, Snow et al., 2011, Murillo-A et al., 2012, Mazine et al., 2014, Staggemeier et al., 2015, Santos et al., 2016), available studies have focused mainly on smaller clades and still lack complete generic sampling, ultimately preventing proper examination of relationships within the tribe. Improving taxonomic and DNA sampling when building phylogenetic trees is known to solve controversial relationships in plants (e.g. APG IV, 2016). Results from such improved phylogenies are key to elucidating systematic problems and also to detect consistent evolutionary patterns as low statistically supported and unbalanced phylogenetic trees may present unreliable branching patterns, branch lengths and substitution models, all of which are ultimately misleading when estimating dates or any other subsequent analysis. Improved phylogenetic resolution in Myrteae will allow more reliable systematic, biogeographic and evolutionary hypotheses of diversity in the tribe. Therefore, the aims of this study are to:

- 1) Develop a well-supported and robust phylogenetic chronogram for Myrteae including all main lineages (46 out of 51 genera and all main clades within large genera).
- 2) Propose a biogeographical hypothesis of evolution of the tribe allowing detection of variation (shifts) in ancestral geographical ranges within a global perspective.
- 3) Estimate diversification rate variation to understand the evolution of heterogeneous diversity among closely related lineages.

## 2. Methods

### 2.1 Taxonomic sampling

The selected sample includes a large range of lineages and geographical distributions within Myrteae. In the case of the mega-diverse genera *Myrcia s.l.* and *Eugenia s.l.*, at least one species was sampled from each informal group (soon to be recognized as formal sections, Mazine et al. in prep, Lucas et al. in prep.) in each genus, following the clade classifications of Lucas et al. (2011) for the nine *Myrcia s.l.* clades and Mazine et al. (2014) and Bunger et al. (2016) for the ten *Eugenia s.l.*



clades (clades 1 to 9 and section *Speciosae*). Fieldwork was conducted in Brazil, Jamaica, Costa Rica, Dominican Republic, New Caledonia, Singapore and Malaysia to collect missing taxa for DNA extraction. Samples were supplemented from the living collection of the Royal Botanic Gardens Kew (K). Duplicate vouchers were deposited in local herbaria and in the Kew herbarium.

The final sample comprises 115 terminals representing 114 species. These include 99 species representing 46 of the 51 genera of Myrteae, 16 genera more than the previous published sample (Lucas et al., 2007). *Blepharocalyx salicifolius* was sampled twice, due to inconsistent placement in past studies (Lucas et al., 2005, Lucas et al., 2007, Murillo-A et al., 2012, de-Carvalho, 2013). Fifteen species were chosen as outgroups based on previous phylogenetic works (Lucas et al., 2007, Biffin et al., 2010, Thornhill et al., 2015). These represent five tribes of Myrtaceae: Leptospermeae (*Leptospermum scoparium*, defined as the furthestmost outgroup in all analysis), Eucalypteae (*Eucalyptus perriniana*), Metrosidereae (*Metrosideros perforata*, *M. stipularis* and *M. nervulosa*), Tristanieae (*Xanthostemon compacta* and *X. montivaga*) and Syzygieae (*Syzygium jambos*, *S. mairei*, *S. gustavioides*, *S. buxifolium*, *S. paniculatum*, *S. amplifolium*, *S. muellerii* and *S. guineense*). Previous studies provide evidence that Metrosidereae, Syzygieae and Tristanieae are closely related to Myrteae (part of the BKMMST clade *sensu* Biffin et al., 2010). See Appendix for a full list of sampled species and vouchers.

## 2.2 Extraction and Sequencing

DNA extraction followed the CTAB extraction protocol for long term DNA storage (Doyle and Doyle, 1987, with modifications following Lucas et al., 2007, and Staggemeier et al., 2015). Approximately 200 milligrams of leaf tissue were used for each extraction. Eight DNA regions were selected for sequencing based on their informative quality evidenced in previous Myrtaceae studies (Lucas et al., 2005, Lucas et al., 2007, Snow et al., 2011, Murillo-A et al., 2012, Mazine et al., 2014, Staggemeier et al., 2015). These are the nuclear region ITS and seven chloroplast regions: *psbA-trnH*, *matK*, *ndhF*, *trnI-trnF*, *trnQ-rps16*, *rpl16* and *rpl32-trnL*. Sequencing was performed using traditional Sanger sequencing protocol, following Lucas et al. (2007). Information on primers and PCR conditions are available in Supplementary Material 1 and 2. Raw sequences were imported and assembled using Geneious (v. 9, Kearse et al., 2012). Resulting contigs were aligned separately for each region using Muscle (Edgar, 2004) implemented in Geneious and adjusted manually. A total of 535 new sequences were generated in this study. Sequences sourced from Genbank are listed in Appendix.

## 2.3 Phylogenetic Analysis

The seven chloroplast regions were concatenated resulting in a matrix of 6453 base pairs, hereafter referred to as the 'cpDNA dataset'. This and the 'nuclear dataset', including only the ITS region (916 base pairs), were used to run two independent Bayesian Inference (BI) phylogenetic analysis. The best evolutionary model was estimated prior to phylogenetic reconstruction using jModelTest 2 (Darriba et al., 2012). Estimation resulted in a best model of GTR gamma+inv for both nuclear and cpDNA datasets. Models were then implemented in MrBayes on XSEDE V. 3.2.6 (Ronquist and Huelsenbeck, 2003) executed in Cipres and run for 15,000,000 generations using default parameters. After visual comparison between phylogenies based on nuclear and cpDNA datasets separately (see section 3.1: *Phylogenetic tree analysis - Grouping and Main lineages*), both nuclear and cpDNA matrices were concatenated resulting in a final matrix of 7369 base pairs, hereafter referred to as the 'combined dataset'. For this matrix, Maximum Likelihood (ML) and BI were run independently to compare topologies and node support (bootstrap vs. posterior probabilities, respectively). For the ML analysis, the final concatenated alignment (available in

Supplementary Material 3) was converted into a simplified Nexus file in Mesquite v3.04 (Maddison and Maddison, 2015) and sourced as input to RAxML-HPC2 (Stamatakis, 2014) analysis implemented in Cipres (Miller et al., 2010). Outputs of all phylogenetic analysis were read using Figtree v1.4.2 (Rambaut, 2014).

#### 2.4 Fossil calibration and Dating

Dates of Myrteae diversification events are controversial. Myrtaceae and Myrteae phylogenies have been dated using fossil calibration and molecular clock approaches in at least seven previous studies (Sytsma et al., 2004, Biffin et al., 2010, Thornhill et al., 2012a, 2015, Murillo-A et al., 2016, Staggemeier et al., 2015, Berger et al., 2016 – see Supplementary Material 4). Except on the occasions where studies were conducted by the same research group, most obtain different dates for similar nodes, sometimes extremely (e.g. Berger et al. [2016] date the crown node of Myrteae at 18 million years old, whilst Murillo-A et al. [2016] date the same node at 92 million years old). The differences in dates appear partially related to phylogeny sample size and balance, but distinctly dependent on the fossils selected and their position in calibration analysis. Because phylogenetic node age is key to interpretation of historical biogeography, reliable fossil selection, calibration and dating analysis is critical; it is discouraging to realise that these decisions are so subjective and open to interpretation. In dating estimation using fossil calibration the standard protocol is to place the estimate minimum date of a fossil on the stem node of a related extant monophyletic taxa in the phylogeny (Forest, 2009). A survey of the oldest fossil records with affinity to Myrteae was conducted and a relatively good fossil record was found assigned to the tribe in the literature. Many fossil descriptions tentatively link them to modern genera (see Supplementary Material 5) however, in reality it is very difficult to identify individual Myrteae genera based on only a few morphological characters. For this reason, the safest approach is to choose the oldest fossil remains confidently described as any genus in Myrteae and place them in the deepest nodes of the tribe.

The oldest fossil records of Myrteae are represented by macrofossil from the upper Cretaceous of Antarctica and represent remains of wood (*Myrceugenelloxylon antarcticus*) and leaves (*Myrciophyllum santacruzensis*) that are similar to extant *Luma* and *Myrcia* respectively (Berry, 1939, Poole et al., 2003). Other wood and leaf fossils from the Paleocene at extreme southern latitudes show affinity in form and distribution to modern genera (e.g. Ragonese, 1980, Troncoso et al., 2002). The most popular fossil from this period used for calibration of Myrteae studies, however, is *Paleomyrtinae*, a fossil fruit with affinity to *Psidium* or *Mosiera* recorded far from any other Myrteae records, in Northern North America (Pigg et al., 1993). Recently, another Paleocene/Eocene macrofossil from the northern hemisphere was described and placed in Myrteae: *Myrtineoxylon maomingensis*, from China (Oskolski et al., 2013). This is stated to be similar to extant Australasian group genera (*sensu* Lucas et al., 2007). Macrofossils assigned to Myrteae found in Eocene deposits are also common and show similar distribution to modern Myrteae (see Supplementary Material 5).

Pollen fossil in Myrteae is, contrariwise, only found in more recent, mid-late Eocene deposits. Myrtaceae pollen fossil (represented by the genus *Myrtacedeites*) was recently reviewed by Thornhill and Macphail (2012) and even though these are found in deposits as old as the Cretaceous, only one species, *M. verrucosus*, shows morphology that undoubtedly places it as Myrteae. Myrteae pollen morphology is conservative (Thornhill et al., 2012b) and in this sense, *Myrtacedeites verrucosus* represents the most reliable fossil record for Myrteae. At least two varieties of *Myrtacedeites verrucosus* are found in late Eocene deposits of Australia, New Zealand, Patagonia and Panama, suggesting Myrteae was an already widespread and diverse group during

that period. *Myrtacedeites verrucosus* is not however, found in deposits of earlier periods (Thornhill and Macphail, 2012).

An important and antagonistic reasoning arises here; pollen fossil of Myrtaceae was recently reviewed and is found to be up to 90 million years old (Thornhill and Macphail, 2012), however, the morphotype that closely matches Myrteae only appears and apparently diversifies in mid Eocene deposits. Added to the hypothesis that pollen is usually the first structure to fossilize when an angiosperm group diversifies (Sauquet et al., 2012), it appears that Myrteae had not diversified before the mid Eocene. Alternatively, if identification of the late cretaceous and Paleocene macrofossils assigned to Myrteae are correct, then Myrteae has to be older than the dates showed by fossil pollen. Furthermore, it is not possible to combine pollen and macrofossil datasets in this case, because they would be placed on similar nodes or represent paradoxical calibration (e.g. if the fossil *Myrceogenia chubutenses* is used to calibrate the stem node of *Myrceogenia* at 66mya, the oldest *Myrtacedeites verrucosus* remains cannot be used to calibrate the whole of the Neotropical Myrteae at 37mya, because the first represents a shallower node in the phylogeny than the second). The solution adopted by this study is to compare two calibration approaches using two distinct fossil sets: a macrofossil set, based on the oldest fossil remains assigned to Myrteae in the literature; and a pollen fossil set, based on different records of *Myrtacedeites verrucosus* remains. The macrofossil approach referred to as Approach A, considered three fossil records: *Myrceogeneloxylon antarcticus*, the oldest fossil in Myrteae, was placed on the crown node of Myrteae calibrating it at 66 million years ago (mya). The following fossils were placed based on their geographical distribution: the crown of the Australasian group was calibrated at 41mya, based on the minimum age estimate of *Myrtineoxylon maomingensis*, a fossil remain from China with affinity to *Octamyrtus*. *Paleomyrtineae princetonensis* from the Paleocene was used to calibrate the crown node of the Myrtus group+Main Neotropical Lineage clade at 56mya, given its reported affinities to modern *Psidium* and *Mosiera* and its distribution closer to extant Neotropical Myrteae.

The second approach is referred to as Approach B and considers three distinct records of *Myrtacedeites verrucosus* (revised by Thornhill and Macphail, 2012) and additional secondary calibration points. The placement of the three remains of *M. verrucosus* was geographically based, following a similar protocol to that of Thornhill et al. (2012a). The oldest record of the pollen in the Neotropics (*Myrtacedeites verrucosus* from the mid-Eocene of Panama and Argentina) was placed on the crown node of the *Myrtus* group+Main Neotropical Lineage clade, calibrating it at 37mya. The oldest *Myrtacedeites verrucosus* recorded for Australia was placed on the crown node of the Australasian group, calibrating it at 35 mya. Finally, *Myrtacedeites verrucosus* remains found in New Zealand from 23mya was used to calibrate the crown node of the *Myrteola* group, the only clade currently found in New Zealand (Lucas et al., 2007, this study). Secondary calibration points from the broader Myrtaceae analysis of Thornhill et al. (2012a, 2015) were used to calibrate the crown of Myrteae at 41mya and the crown of the BKMMST clade (Myrteae + sister tribes, sensu Biffin et al., 2010) at 66 mya. In both approaches A and B, the root of the family was constrained to be no older than 85 mya (following Berger et al. 2016). A summary of the calibration points used and the rate parameters applied in Beast are summarized in Table 1. Both approaches A and B were used to produce dated phylogenies using a lognormal relaxed clock set for Birth-Death speciation and 50,000,000 generations in BEAST v.1.8.3. (Drummond et al., 2012). Two analyses were run for each approach, results were checked for convergence in Tracer v1.6.0 (Rambaut et al., 2013), burnin was selected as 0.1% of total trees and final chronograms (dated phylogenies) were visualised in Figtree v1.4.2 (Rambaut, 2014).

## 2.5 Historical Biogeography Inference

BioGeoBEARS (Matzke, 2013) implemented in R (R Core Team, 2016) was used to analyze ancestral geographical range variation over resulting chronograms (Approaches A and B). BioGeoBEARS allows implementation of a third free parameter “j” (founder event/jump speciation) that permits a daughter lineage to have a different area from the direct ancestor a feature that improves the log likelihood of resulting inferences of ancestral areas in comparison to a model with only two free parameters (e.g. dispersion/extinction only in Lagrange, Ree and Smith, 2008). BioGeoBEARS does not work well when many possible ancestral areas are implemented unless the maximum number of areas any species may occupy is reduced. Range area per terminal in the phylogeny was therefore coded in relation to species distributions, not genera. In this way, most terminals are restricted to single area. Area coding aimed to consider the current distribution of the group and historical geology and tectonics. The seven areas chosen were: (A) South America, (B) Central+North America (including the greater Antilles in the Caribbean), (C) Australia and New Guinea (referred to as Australia+NG), (D) New Caledonia and New Zealand (referred to as NCNZ, representing the Zealandia plate, Trewick et al., (2007)), (E) Africa (here including Madagascar), (F) Mediterranean Europe and (G) Southeast Asia (referred to as SEAsia). Distribution ranges, time slice matrices and values of area adjacency through time are available as Supplementary Material 6.

### 2.6 Diversification Rates Analysis

Configuration shifts in diversification rates were calculated using speciation/extinction model type analysis in BAMM (Rabosky et al., 2014). BAMM works with incomplete phylogenetic datasets and allows a certain degree of phylogenetic uncertainty (see BAMM documentation). Missing taxa per tip or clade in the phylogenetic tree was estimated using previously published works (Wilson et al., 2005, Wilson, 2011, Lucas et al., 2007, Lucas et al., 2011, Mazine et al., 2014, Staggemeier et al., 2015, Santos et al., 2016, WCSP, 2016). In the largest genera, *Myrcia* s.l. and *Eugenia* s.l., the numbers of species per clade was estimated by specific studies (Mazine et al., 2014) and unpublished data (Lucas et al., in prep, Faria Júnior, 2014, Bünge, 2015). Priors for the BAMM control file were generated using the dated phylogenetic tree input into the function *setBAMMpriors* in the package *BAMMtools* v2.5.2 implemented in R (R Core Team, 2016), estimating 2500 species in Myrteae. The control file was set for 100,000,000 generations and the analysis was run twice as recommended (see BAMM documentation), giving similar results. Resultant MCMC Log likelihoods were tested against generation number for convergence using the *coda* package implemented in R (R Core Team, 2016). All other outputs contained in the “*event\_data*” file were analysed using *BAMMtools* in R. A recent paper casted doubt in the reliability of results produced by BAMM (Moore et al., 2016), but the criticism concerning the priors used by the software were adjusted in the latest version (see BAMM documentation). Other problems cited by that study can be applied to most macroevolutionary methods (e.g. estimation of extinct clades) and in this sense BAMM was not considered better or worse than similar software. Priors and proportion of samples per clade are given in Supplementary Material 7.

## 3. Results

### 3.1 Phylogenetic tree analysis - Grouping and Main lineages

Phylogenetic analysis shows Myrteae to be a coherent, well defined group with >0.95 posterior probability and 100% bootstrap support in cpDNA, nuclear and combined datasets analyses (node A, Figure 2, Supplementary Materials 8 and 9). The next deepest node in the tribe’s phylogeny (node B, Figure 2) is poorly supported by all datasets while the two following nodes (nodes C and D, Figure 2) are recovered with strong posterior probability (>0.95) and high bootstrap support (>70) in the combined and cpDNA datasets. Four lineages result from divergences at these

four nodes (A, B, C and D). One of them represents a single, ungrouped monotypic genus (*Myrtastrum*) and the other three are here informally coined: the Australasian group, the *Myrtus* group and the Main Neotropical Lineage (colour coded in Figure 2 as orange, blue and green respectively).

The backbone of the Main Neotropical Lineage is poorly supported in all dataset analyses, but eight major clades with high bootstrap (>70) and/or posterior probability (>0.95) supports are recovered in the combined dataset and here informally named: the *Eugenia*, *Pimenta*, *Myrteola*, *Myrceugenia*, *Myrcia*, *Plinia*, *Blepharocalyx* and *Psidium* groups. These eight clades are also recognized with similar representing taxa and support in the cpDNA dataset analysis (Supplementary Material 8). The nuclear dataset analysis presents poor support for most of the deepest nodes in the phylogeny and is mostly non-informative to analyse relationship between and within these clades. The relationship between *Plinia* sp1 as sister to *Myrrhinium atropurpureum* is the only strongly supported arrangement in the nuclear dataset analysis that differs from the cpDNA and combined datasets (Supplementary Material 9). In the next sections, relationships within each of the ten clades (the eight clades within the Main Neotropical Lineage plus *Myrtus* and Australasian groups) and two ungrouped genera (*Myrtastrum* and *Amomyrtus*) are discussed based on the combined dataset (Figure 2). Diversity estimates per clade are taken from WCSP (2016) and Wilson (2011).

### 3.1.1 The Australasian group

The Australasian group (in orange, Figure 2) has similar configuration to the informal Australasian group *sensu* Lucas et al. (2007). It is positioned as sister to the *Myrtus* group+Main Neotropical lineage clade and includes species within the genera *Gossia*, *Uromyrtus*, *Rhodamnia*, *Austromyrtus*, *Decaspermum*, *Octamyrtus*, *Rhodomyrtus*, *Kanakomyrtus*, *Pilidiostigma* and *Archirhodomyrtus*. This lineage comprises genera restrictedly distributed in Southeast Asia, Australia and Pacific islands (Figure 3A) and an estimated c. 250 accepted species. Supports both from ML and BI analysis are high (>70 bootstrap and/or 0.95 posterior probability) for most internal nodes in the clade, except for the positions of *Austromyrtus*.

### 3.1.2 The *Myrtus* group

The *Myrtus* group (in blue, Figure 2) contains the only European genus *Myrtus* and three Neotropical genera: *Accara*, *Chamguava* and *Calycolpus*. This group is recovered in all molecular dataset analyses, although relationships within the group vary slightly depending on the dataset under examination and the type of phylogenetic analysis (ML or BI). The main distinction is the placement of *Accara* and *Myrtus* that swap positions between sister to the rest of the group or to *Chamguava*. The two species of *Calycolpus* always appear as a strong supported group. Based on these results, *Myrtus* group present a peculiar discontinuous distribution throughout Mediterranean and Neotropical areas (Figure 3B) and an estimated diversity of c. 20 species.

### 3.1.3 Main Neotropical lineage

The Main Neotropical Lineage (in green, Figure 2) presents eight well supported (PP >0.95, BS >70) clades: the *Blepharocalyx*, *Psidium*, *Pimenta*, *Myrteola*, *Myrceugenia*, *Plinia*, *Myrcia*, *Eugenia* groups. The latter five are very similar to the circumscription of Lucas et al. (2007). With the exception of the consistently well supported relationship between the *Plinia* and *Myrcia* groups, the relationship between these groups is poorly resolved within the Neotropical lineage. The *Blepharocalyx* group is endemic to the Neotropics (Figure 3C) and includes *Blepharocalyx salicifolius* and *B. eggersii*. *Blepharocalyx* is a genus of only four accepted species and future additions to the phylogeny may also place *Blepharocalyx myriophyllus* (the only unsampled *Blepharocalyx* species in

this study) in this group increasing diversity to three accepted species. Currently accepted *Blepharocalyx cruckshanksii* is nested in the *Myrceugenia* group. The *Psidium* group includes the genera *Mosiera*, *Myrrhynium*, *Psidium* and at least one species of the polyphyletic *Calyptrogenia* (*C. biflora*).

The *Pimenta* group includes the genera *Curitiba*, *Acca* (*A. sellowiana*), *Campomanesia*, *Legrandia*, *Pimenta* and at least one species of *Eugenia* (*Eugenia yumana*), nested within *Pimenta*. Taken in this sense, the group is endemic to the Neotropics (Figure 3C) and includes an estimated c. 50 species. The *Myrteola* group includes the genera *Lophomyrtus*, *Neomyrtus*, *Myrteola*, *Ugni* and *Lenwebbia*, and contains c. 15 species. This group presents an atypical geographical distribution within the tribe, with two genera (*Ugni* and *Myrteola*) endemic to Patagonia and the alpine biomes of South and Central America, one genus endemic to Australia (*Lenwebbia*) and two genera endemic to New Zealand (*Neomyrtus* and *Lophomyrtus*) (Figure 3D). The *Myrceugenia* group includes the genera *Luma*, *Myrceugenia* and one species of the polyphyletic *Blepharocalyx* (*B. cruckshanksii*); an estimated c. 50 species are assigned here. This group presents a somewhat restricted distribution to sub-temperate and subtropical biomes of South America, mainly Chile and Southern Brazil (Figure 3E). The *Plinia* group includes the genera *Plinia* (emerging paraphyletic), *Algrizea*, *Myrciaria*, *Siphoneugenia* and *Neomitranthes* and an estimated diversity of c. 120 species. The *Myrcia* group includes four genera: *Mitranthes*, *Myrcia*, *Marlierea* and *Calyptranthes*. This group is estimated to include around 700 species. Both *Plinia* and *Myrcia* groups are endemic to the Neotropics (Figure 3C). The *Eugenia* group includes the genera *Myrcianthes*, *Hottea*, *Pseudanamomis*, and *Calyptrogenia*. Clade 9 (sensu Mazine et al., 2014) appears polyphyletic in our analysis with all old world species (including *Eugenia roseopetiolata*, *E. reinwardtiana*, *E. bullata* and *E. paludosa*, here defined as clade 9a) appearing monophyletic in an unrelated, well supported clade. The *Eugenia* group is the most diverse and widespread group in Myrteae, with around 1000 species and a pantropical distribution (Figure 3F).

#### 3.1.4 Ungrouped genera: *Myrtastrum* and *Amomyrtus*

Two genera, *Myrtastrum* and *Amomyrtus*, appear ungrouped in the combined dataset. *Myrtastrum*, a monotypic genus endemic to New Caledonia (shown in orange, Figure 3D), appears either isolated as sister to all extant Myrteae in the combined and nuclear datasets, or as sister to *Myrtus* group+Main Neotropical lineage, in the cpDNA dataset analysis. *Amomyrtus*, a genus of two species endemic to Patagonia (shown in purple, Figure 3D), appears as sister to *Myrceugenia* group in both the cpDNA and combined dataset, though this relationship presents a poor support in the latter. This relationship is not supported by the nuclear dataset, where it appears as sister to *Legrandia*, again with a low support.

#### 3.2 Dating inference

Figure 4 contrasts results from calibration using the two fossil datasets (approaches A and B). Relationships between the *Eugenia*, *Pimenta* and *Myrteola* groups receive high statistical support (PP >0.95) in the chronograms compared to the lower support returned from the ML and BI analysis. Other aspects of the topology, including outgroup relationships, show discreet differences between chronograms where node support is low.

Because the macrofossil ages are older, approach A returns older dates for all nodes within Myrteae. In this analysis, the stem node of Myrteae (Figure 4A"a") is estimated as being from the late-Cretaceous (80.72 mya) and the crown node (Figure 4A"b") from the Cretaceous-Paleocene boundary (KT boundary, 65.55 mya). Approach A also suggests that the three major clades within

Myrteae (the Australasian group, *Myrtus* group and the Main Neotropical Lineage) split soon after initial Myrteae diversification, in the Paleocene and early-Eocene, between 63 mya and 53 mya (highlighted in Figure 4A). The diversification of all major clades within the Main Neotropical Lineage are estimated in this analysis to have taken place in the Eocene, between 52 and 39 mya. The oldest crown nodes in this analysis are: the Australasian group (59.05 mya), the *Eugenia* group (44.42 mya) and the *Pimenta* group (44.41 mya). The youngest crown nodes in this analysis are: the *Plinia* group (39.61 mya), the *Myrcia* group (39.19 mya) and the *Psidium* group (39.12 mya).

Myrteae pollen fossil is younger than the macrofossils and consequently ages estimated from this fossil set (approach B, Figure 4B) are younger than those from approach A. In this approach, the stem node of Myrteae (Figure 4B" a") is estimated from the late-Paleocene (58.96 mya) and the crown node (Figure 4B" b") dates to the mid-late Eocene (40.76 mya), around 25 mya younger than the same nodes in approach A. In approach B the three major clades within Myrteae (Australasian and *Myrtus* groups and the Main Neotropical Lineage) again split immediately after initial Myrteae diversification (highlighted in Figure 4B) but these events are estimated to have occurred between 40 mya and 35 mya, in the late Eocene. In this approach the diversification of all major clades within the Main Neotropical Lineage are estimated to have taken place between the late-Eocene and Oligocene. The oldest and youngest crown nodes in this analysis are similar to approach A but between 15 mya and 20 mya younger. The oldest groups in this analysis are: the Australasian group (36.88 mya), the *Pimenta* group (29.40 mya) and the *Eugenia* group (29.29 mya). The youngest crown nodes in this analysis are: the *Psidium* group (25.62 mya), the *Myrcia* group (25.58 mya) and the *Myrteola* group (23.39 mya). Median age estimates and 95% confidence intervals (CI) for diversification dates of the main nodes of both analysis are plotted and contrasted in Table 2.

### 3.3 Biogeographical patterns

BioGeoBEARS was applied to chronograms resulting from both calibration approaches (Figure 5). In each case results indicate a higher value of log likelihood for three parameters (DEC+j, LnL = -156.72 and LnL = -161.48 for approaches A and B respectively) in comparison to two parameters (DEC, LnL = -202.75 and LnL = -207.92 for approaches A and B respectively) showing jump speciation (i.e. dispersal between non-adjacent areas) as an important pattern in range variation of Myrteae. The most probable ancestral areas for the stem and crown nodes of Myrteae (Figure 5 "a", "b" respectively) is NCNZ in both analyses.

In the Australasian group the ancestral range of the crown node also has high probability of being NCNZ in both dating approaches but subsequent nodes show multiple shifts from NCNZ to Australia+NG and SEAsia and back to NCNZ. These shifts are estimated to date from the Eocene-Oligocene (shifts 2-7, Figure 5A) in approach A and from the Oligocene to late Miocene (shifts 2-7, Figure 5B) in approach B. The clade composed of the *Myrtus* group + Main Neotropical Lineage share a most likely ancestral area of South America for both approaches shifting from a previous NCNZ range (shift 1, Figure 5) during the Paleocene (approach A) or the late-Eocene (approach B). The estimate of ancestral range for the stem and crown node of the *Myrtus* group presents an important difference between approaches A and B. In approach A an early South American range shifts to Central+North America range during the late Paleocene (shift 8, Figure 5A) influenced by the distribution of *Chamguava* on the latter tectonic plate. This then shifts to the Mediterranean during the mid-Eocene for *Myrtus* (shift 9, Figure 5A) and to South America for *Calycolpus* and *Accara* in the late-Eocene to early-Oligocene (shifts 10 and 11, Figure 5A). In dating approach B, the crown node of the *Myrtus* group presents high probability of ancestral range in South America, shifting from there to the Mediterranean area during the late Oligocene for *Myrtus* (shift 8, Figure 5B) and to

Central+North America in the early Miocene for *Chamguava* (shift 9, Figure 5B). In the Main Neotropical Lineage the most likely areas of ancestral range for both Approaches A and B is South America. In approach A, nine shifts from South to Central+North America (shifts 12, 14, 16, 18, 19, 23, 25, 27, 29, Figure 5A) and seven shifts back to South America (shifts 13, 15, 16, 20, 24, 26, 28, Figure 5A) are detected in this lineage. These occurred during the Eocene-Oligocene time slice and are observed in all clades with the exceptions of the *Myrceugenia* and *Myrteola* groups. In approach B, the same nine shifts from South to Central+North America are detected in the same groups (shifts 10, 11, 12, 13, 14, 17, 18, 19, 23, Figure 5B). In approach B however, these shifts are no older than the early Miocene and no shifts back to South America are observed. Events of dispersion from the Neotropics (areas A and B) to the region of Australia+NG and NCNZ (areas C and D) are observed in the *Myrteola* and in *Eugenia* groups. In the *Myrteola* group this event is estimated in approach A to have occurred from South America to Australia+NG in the late Eocene (in *Lenwebbia*, shift 21, Figure 5A) and afterwards to NCNZ (in *Neomyrtus* + *Lophomyrtus*, shift 22, Figure 5A). In approach B, the same event is estimated to have occurred in the late Oligocene and with a higher probability for the route NCNZ to Australia+NG than the other way around (shifts 15 and 16, Figure 5B). The *Eugenia* group presents a more complex series of dispersion events. In both approaches A and B, a shift from the Central+North America region to NCNZ is observed in the common ancestor of the clade containing the Australasian and African species (shift 29 in Figure 5A and 20 in Figure 5B). This lineage subsequently disperses to Africa+Madagascar (represented by *Eugenia rosapetiolata*, shift 30 in Figure 5A and 21 in Figure 5B) and to Southeast Asia (represented by *Eugenia reinwardtiana*, shift 31 in Figure 5A and 22 in Figure 5B). Even though the geographic sequence of events in this *Eugenia* clade is the same, the estimated date for these dispersion events in approach A is the late Oligocene, while in approach B it is at least 10 million years later, in the Miocene.

### 3.4 Diversification Rate Shifts

Number of configuration shifts and log likelihood were higher than 1000 (significantly more than the recommended minimum of 200) after burnin for all BAMM analyses. Convergence between log likelihood and number of generations was observed in analysis with both callibrations (Approach A and B). The 95% credible set of rate shift configurations sampled with BAMM included 91 distinct shift configurations for approach A and 73 for approach B, of which the configurations with the highest probability included two or three shifts for both approaches. Posterior probability for a null model (i.e. no diversification rate shifts) was lower than could be estimated in both cases, therefore a Bayes factor was not calculated (see BAMM documentation). Thus, diversification rate heterogeneity is clear in the dataset. Mean phylorate through time is plotted for both chronograms in Figure 6. In both approaches, the best configuration shift indicates three points of increasing diversification rates, all of which occur in the Main Neotropical Lineage. The highest shift configuration probability shows three shifts towards acceleration of diversification rates positioned in similar branches in the two analyses: one in the common ancestor of most extant species of *Eugenia*, (Figure 6Aa, Ba), one in the crown node of *Psidium* (Figure 6Ab, Bb) and one in the common ancestor between *Plinia* and *Myrcia* groups (Figure 6Ac, Bc). In approach A, shifts in the *Eugenia* and *Plinia+Myrcia* groups occurred at the mid or late-Eocene, while that in *Psidium* occurred at the Oligocene/Miocene boundary. In approach B, both shifts in the *Eugenia* and *Plinia+Myrcia* groups occurred at the Oligocene, while the one in *Psidium* dates to the mid-Miocene. Due to its younger dating estimation, approach B presents higher diversification rates through the tribe than approach A.

## 4. Discussion

### 4.1 Systematic Implications



The phylogeny of Myrteae resulting from the combined dataset was reconstructed by a more informative molecular matrix and has considerably broader lineage sampling and higher statistical support in the deep nodes than those in previous works (e.g. Wilson et al., 2005, Lucas et al., 2005, Lucas et al., 2007, Murillo-A et al., 2012, Thornhill et al., 2015) and can be used to understand the systematics, evolution and ecology of the tribe more accurately. Low support in most branches from the nuclear database makes it difficult to evaluate potential incongruence between nuclear and cpDNA trees. There is not enough evidence to detect, for example, the role of ancient hybridization events in Myrteae history, usually noted by incongruence between these genomes (e.g. Soltis and Kuzoff, 1995). The only clear incongruence, the position of *Plinia* sp1 as sister to *Myrrhinium atropurpureum*, has to be investigated but may be an artefact of the sequencing process (e.g. contamination).

One of the main differences between this and previous phylogenetic hypotheses is the relative position of the three main lineages: the Australasian and *Myrtus* groups and the Main Neotropical Lineage. In the first phylogenetic works focused on the tribe (Lucas et al., 2005, Lucas et al., 2007), *Myrtus communis* appeared as the sister lineage to all extant Myrteae and the Australasian clade appeared sister to the equivalent Main Neotropical Lineage clade. With this broader sample however, it is evident that *Myrtus* forms part of a predominantly Neotropical lineage. Within the Main Neotropical lineage, novel subtribal relationships are the inclusion of the *Blepharocalyx* group, formally ungrouped (Lucas et al., 2005, 2007, Murillo-A et al., 2012) or placed next to *Pimenta* (de-Carvalho, 2013) and the position of *Algrizea*, previously unplaced (Lucas et al., 2007), within *Plinia* group (also shown but not discussed in Staggemeier et al., 2015). Another novelty is the division of the former *Pimenta* group genera (sensu Lucas et al., 2007) into two groups, the *Pimenta* group and the new *Psidium* group, and one ungrouped species *Amomyrtus luma*. The placement of *Amomyrtus luma* fluctuates, but the high support of the relationship between *Amomyrtus* and the *Myrceogenia* group in the cpDNA dataset, in addition to similar geographical distribution, might mean that this genus will be treated as *Myrceogenia* group in the future. Further analysis to better place this genus within Myrteae is desirable.

Genera that will require nomenclatural adjustment include: *Hottea*, *Pseudanamomis* (both nested inside *Eugenia*), *Calyptrogenia* (polyphyletic, with species nested in *Eugenia* and *Psidium*), *Mitranthes* (nested within *Myrcia* s.l.), *Eugenia* (polyphyletic, with at least one species nested in *Pimenta*) and *Plinia* (paraphyletic). *Blepharocalyx* is known to be polyphyletic since the first molecular works in the tribe, likely requiring the resurrection of the genus *Temu* for *Blepharocalyx cruckshanksii* (see Lucas et al., 2007). *Calyptrogenia biflora* is noted to strongly resemble the continental America species *Psidium amplexicaule* Pers., but formal synonymization is required. A further important result from this phylogenetic topology is that it seems that the Caribbean, previously considered home to four endemic genera, apparently has no generic endemism in Myrteae, as *Hottea*, *Calyptrogenia*, *Mitranthes*, and *Pseudanamomis* are all nested inside larger widespread genera.

Of the five here unsampled, accepted genera in Myrteae (based on Wilson 2011), *Meteromyrtus* has recently been shown to be nested in *Eugenia* (Wilson and Heslewood, 2016). The remaining four (*Myrtella* from New Guinea, Andean *Amomyrtella*, *Lithomyrtus* from Australia and *Stereocaryum* from New Caledonia) are still to be placed. These four genera present straight stamens in the bud, so based on this consistent morphological character it is likely that their positions will be other than within the *Myrcia*, *Plinia* or *Blepharocalyx* groups, in which stamens are consistently incurved (Vasconcelos et al., 2015). These results, in addition to the already proven

polyphyletism of the classical subtribal classification based on embryo morphology (Lucas et al., 2007) brings consistency to the current understanding of Myrteae and its classification.

#### 4.2 Comparative Dating analysis

Results from comparative fossil calibration show important distinctions between estimated crown node ages using different approaches. Thornhill et al. (2012a) also contrast macro and microfossil calibration in Myrtaceae, combining the two fossil sets in a third calibration analysis. The fossils selected in the study presented here however, had to be placed on the same nodes so a combined dataset was not possible. Since calibration was performed with fossils of different ages on similar nodes in each approach, the resulting date distinction is expected but it is useful to demonstrate subjectivity when choosing fossil placement and how this influences interpretation of dates. Even though dates stabilize towards shallower nodes, especially when considering confidence intervals, overlap between dates from approaches A and B is still low (see Figure 7).

Approach A, using only macrofossil data finds estimated dates similar to Sytsma et al. (2004) and Staggemeier et al. (2015), suggesting a first event of Myrteae diversification in the Paleocene. An estimated age near the KT boundary might link increased Myrteae species diversity to increased mammal and bird diversity following dinosaur extinction (Cracraft, 2001, Penny and Phillips, 2004). A preference of mammals and birds for fleshy berries may have provided a selective advantage over the capsular fruits of closely related tribes of Myrtaceae (Friis, 1987, Biffin et al., 2010). On the other hand, approach B finds a similar dates to Biffin et al. (2010) and Thornhill et al. (2012a), suggesting a first event of Myrteae diversification in the Eocene. In this approach, the explanation for the KT boundary above could be applied to the BKMSST clade (Myrteae and sister tribes, *sensu* Biffin et al., 2010) as this clade has other fleshy fruited Myrtaceae tribes and appears in approach B to date from the KT boundary (Thornhill et al., 2012a). In further support of approach B, the younger dates returned better explain the current distribution of Myrteae with less necessary LDDE events (see section below).

#### 4.3 Biogeographical inference

The biogeographical analyses presented here provides a hypothesis of how Myrteae acquired its present Pantropical geographical distribution. Thornhill et al. (2015) and Berger et al. (2016) using a smaller Myrteae sample, recovered Australia as the most likely ancestral area of early diversification for Myrtaceae. The present study infers NCNZ as the ancestral range of Myrteae, with high probability in both approaches A and B (Figure 5 "a", "b"). There is evidence, however, that large portions of Zealandia, including New Caledonia and New Zealand, were underwater between the Eocene and Oligocene (Gibbs, 2004), casting doubt on a potential NCNZ Eocene origin suggested by the more recent dates of approach B. Some hypothesis, however, indicate that other adjacent land portions of the Zealandia continent were above sea level when NCNZ was submerged; these neighbouring islands could have acted as refugia, preserving representative biodiversity in Zealandia from lineages that have since undergone extinction in other continents (e.g. Australia) even when NCNZ was submerged (e.g. Condamine et al., 2016). This pattern would explain the survival and present distribution of *Myrtastrum*, a monotypic genus endemic to New Caledonian and sister to the rest of Myrteae. Even though a possible NCNZ origin can be explained, the safest conclusion may be that Myrteae shows an eastern Gondwana ancestral area that today is represented by NCNZ and also Australia+NG. Reasons for this include the proximity of the Zealandia and Australian plate during that period (Trewick et al., 2007), the possibility that NCNZ species diversity observed today is a relict of more widespread lineages (as reasoned above) and the possibility that incomplete sampling of some deeper-node genera is biasing the analysis (*Gossia* and *Uromyrtus*, for instance are

also diverse in Australia+NG [WCSP, 2016] but area coding according to species distribution influenced the reconstruction towards NCNZ).

Approaches A and B show similar area shifts (numbered in Figure 5), but occurring during distinct time periods. The older age estimation of approach A causes it to present more area shifts (32 in comparison with 23 from approach B), perhaps due to area adjacencies of different time slices (see Supplementary Material 6). The dating divergences between approaches also affect the number of LDDE events necessary to explain the current distribution in Myrteae (see summary in Table 3). Although events of LDDE are an important process in angiosperm biogeography (Crisp et al., 2011), long transmarine diversification events are considered less likely than short distance dispersion and diversification by vicariance or continental population isolation (Howe and Smallwood, 1982). The first area shift recorded in both approaches A and B is the transition from NCNZ to South America from the stem to the crown node of the clade containing *Myrtus* group and the Main Neotropical Lineage (shift 1, Figure 5A,B). LDDE is unlikely here as until around 40 mya, South America was still linked to portions of eastern Gondwana, forming a single continent connected by Antarctica (McLoughlin, 2001). It is possible that, after initial diversification in eastern Gondwana, Myrteae became widespread throughout Antarctica and South America; there is evidence that global temperature was much warmer in the early Cenozoic (Huber et al., 1995) and that rainforest vegetation covered Antarctica until around 30 mya (Francis and Poole, 2002; Francis et al., 2008). Abundant Myrtaceae fossil records found at high latitudes in South America, southern Patagonia and nearby Antarctica (Supplementary Material 5, Eklund, 2003, Hayes et al., 2006, Francis et al., 2008) also provide evidence for this hypothesis. The scenario of a widespread Myrteae throughout these continents, followed by their late-Eocene disconnection (McLoughlin, 2001) and Miocene Antarctica glaciation (Kennett et al., 1975) with consequent vicariance between the Australasian group and *Myrtus* group+Main Neotropical Lineage on distinct sides of the globe is likely in both dating scenarios.

In the Australasian group, most area shifts between SE Asia, Australia+NG and NCNZ, in both approaches, occurred in a period range where proximity between these continents did not require LDDE events. The only exception is *Rhodamnia cinerea* that shifts from Australia+NG to SE Asia (shift 3, Figure 5A,B) in the Eocene to early Oligocene; this may only be explained by LDDE, given the distance between these areas in that period (McLoughlin, 2001). In both approaches A and B, there is evidence for a quick northerly vertical expansion into the whole of South America soon after initial diversification in that continent. In approach A, a series of shifts back and forth South America and Central+North America are observed occurring mostly from the early Eocene to the late Oligocene. Such area shifts, however, would require multiple LDDE events, because these two continents were too far apart during that period (McLoughlin, 2001). Similar area shifts in approach B are estimated to have occurred much more recently, mostly during the Miocene, when South and North America were closer together or connected by the Panama Isthmus (Montes et al., 2015) suggesting short distance dispersion events. The only exception is the diversification of *Myrcianthes fragrans* to the greater Antilles that would require an LDDE event in both approaches.

Based on past phylogenetic position and northern hemisphere distribution, past studies proposed that the current geographical range of *Myrtus* might be a relic from a much wider distribution of Myrteae (Berry, 1915, Thornhill et al., 2015). However, the highly supported sister relationship of *Myrtus* to exclusively Neotropical genera, including Central American *Chamguava*, provides evidence of vertical movement through the American continents towards the Mediterranean, perhaps by relatively short distance dispersal via what is today Greenland and northern Europe, under a warmer paleo-climatic regime (Zachos et al. 2001). Possible evidence for

this event is the presence of the *Paleomyrtineae* fossil from this period in North Dakota (Pigg et al., 1993). The diversification of the *Myrtus* group from South to Central+North America in the Paleocene as estimated by approach A (shift 8, Figure 5A) is possible without LDDE events due to the Nicoya island complex, which linked present day Ecuador and Central America during that period (Dengo, 1975, Gentry, 1982). In approach B, the shift between South America to Central+North America in the stem node of the *Myrtus* group is not recovered. In this approach, the estimated shift occurs from South America straight to Mediterranean Europe (shift 8, Figure 5B). Nevertheless, much later dates for this shift in this approach means that a similar route from South to Central+North America and Europe would be possible without LDDE events, because of the proximity of these continents in the Miocene. *Myrtus* genetic diversification varies however, from the east to west of its range (Migliore et al., 2011), not congruent with vertical movement through the American continent. This complex pattern requires future research.

Two clades (*Myrteola* and *Eugenia* groups) within the Main Neotropical Lineage also have representatives in Australia+NG, SE Asia and Africa, but these colonisation events likely occurred in different periods and by different processes. Antarctica remained habitable and in proximity to NCNZ and South America until the late Oligocene (Francis et al., 2008). In both approaches A and B (when considering upper confidence interval limits), the shift in ancestral area in the *Myrteola* group from South America to NCNZ and Australia+NG occurred before this bridge was severed by ice-sheet formation, suggesting the possibility of terrestrial migration or Antarctic colonization followed by vicariance, giving the *Myrteola* group a *Nothofagus*-like distribution (van Stenis, 1971, Swenson et al., 2001). Adaptations that may have allowed this group to achieve this range and survival in Antarctica until later than sister lineages even in colder climates, include their shrubby habit, winter seed dormancy (Smith-Ramirez et al., 1998) and likely frost resistant wood anatomy (Schmid and Baas, 1984), uncommon in other Myrteae (Lucas et al., 2007).

Due to stabilization of dates at the shallower nodes and considering the confidence intervals, Australasian and African *Eugenia* events of dispersion are estimated to have occurred at similar dates, around the late Oligocene-early Miocene, in both dating approaches. Considering an ancestral area of Central+North America for the clade and that Antarctica was already covered by ice-sheets and no longer habitable (Zachos et al., 1991, Ivany et al., 2006) at the Miocene, the only scenario possible to explain *Eugenia*'s current pantropical distribution is a series of LDDE events (similar to other plant groups such as *Psychotria*, Matzke, 2013, and Simaroubaceae, Clayton et al., 2009). The picture proposed by the results of biogeographic analysis is that this event was towards the east, from the Caribbean (in *Pseudanmomis*) colonizing first NCNZ, then Africa and lastly SE Asia, but a larger *Eugenia* sample from these regions may prove otherwise. Particular abilities of the *Eugenia* lineage that underwent long-distance dispersal, to cross marine boundaries, might explain why species of this group are also found in many islands of the Indian and Pacific oceans. Many (possibly all) South African species of *Eugenia* are cryptically dioecious, a character unrecorded for the genus out of Africa (van der Merwe et al., 2005, Vasconcelos pers. obs.). Dioecy is linked to small green or white flowers, generalistic pollination systems and to island floras where in extreme cases, such as Hawaii, over a quarter of the species can be dioecious (Bawa, 1980). It is possible that dioecy of extant South African *Eugenia* species is a legacy of island-hopping ancestors. Further research focused on innovative reproductive characteristics necessary for such dispersal, such as co-evolution with migratory birds, seed resistance and self-compatibility (Baker, 1955) will be necessary to better understand the unique distribution patterns of this group.

#### 4.4 Changes in diversification rates, key innovations and mega-diverse genera

This study demonstrates heterogeneity of diversification rates in Myrteae. Both dating approaches return similar results in this case: the three main accelerating shifts of diversification rates occurred in the Main Neotropical lineage. This explains why species diversity of the tribe in this continent is ten times higher than in the Old World (Lucas et al., 2007, WCSP 2016). In evolutionary biology, some of the most plausible explanations for changes in diversification rates are related to acquisition of new biological traits in the lineage (e.g. key-innovations, Donoghue, 2005). This is a reasonable hypothesis for Myrteae: differences in characters related to embryo morphology in *Myrcia*, *Plinia* and *Eugenia* have been proposed as adaptive advantages for these groups (Landrum, 1986, Landrum and Stevenson, 1986). The *Plinia* and *Eugenia* groups, with independent origins, present homogeneous cotyledons that have been related to seedling starch storage (Landrum, 1986) while *Myrcia* have leaf-like, well developed embryos that allow faster germination. These embryo forms are different from extant Myrteae that do not exhibit these specialisations.

The accelerating diversification rate shift in *Psidium* however, is less likely to be linked to the embryo as in this group it is similar to those found in the Australasian and *Pimenta* groups (Landrum and Stevenson, 1986). A possible explanation for the success of *Psidium* may be linked to cytogenetic events: *Psidium* is the Myrteae lineage with the highest documented cases of polyploidy (Costa et al., 2008), frequently associated with increased fitness (Wood et al., 2009, Madlung, 2013). The bony *Psidium* testa opening via an operculum (a synapomorphy of the genus) through which germination occurs (Landrum and Stevenson, 1986) may also be a factor, promoting mechanical seed dormancy conducive to success in seasonal environments. It is also notable that all invasive species of Myrteae are *Psidium* (Richardson and Rejmanek, 2011), showing adaptive features of this lineage that might be linked to its higher diversification rate.

## 5. Conclusions Remarks and Future directions

This work provides an up to date phylogeny to be used as a base for further systematic and modelling studies in Myrteae. The dating, biogeography and diversification patterns analyses clarify the evolutionary picture of the most diverse tribe in Myrtaceae, but also raise a number of avenues for future studies. These include, for instance: a better resolution for the relationships in the backbone of the main Neotropical lineage; nomenclatural changes in poly and paraphyletic genera; formalization of subtribal nomenclature; detailed biogeographical analysis of individual clades; the importance of high southern latitudes in early Myrteae diversification events; and better links between acceleration shifts in diversification rates and trait evolution. Results from the comparative dating approaches using macro and microfossil separately show how the choice of fossil set and placement interpretation affects all interpretation of subsequent evolutionary analysis. Calibration using pollen fossil evidence (approach B) requires less LDDE events to explain current Myrteae distribution. This, in addition to the reasoning provided in the section 2.4 (*Fossil calibration and Dating*), suggests that this dating approach is more reliable and should be preferred by future studies in Myrteae.

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## Tables

Table 1: Summary of two fossil sets and secondary calibration points selected to estimate diversification rates in Myrteae. Rate (normal or lognormal) is based on Beast parameters. For fossil reference see Supplementary Material 5.

Table 2: Median age estimations and 95% confidence intervals (CI) for dates of the main Myrteae nodes based on BEAST analysis.

Table 3: Summary of most likely events responsible for area shifts in Myrteae based on age period and confidence intervals. LDDE events were considered when distance between areas are recorded as 0.1 or 0.5 for the time slice (see Supplementary Material 6)

## Figure Captions

Figure 1: Biodiversity of Myrteae represented by the characteristic polystemonous white flowers (A - H) and fleshy, berry-like fruits (I - O). (A) *Accara elegans*; (B) *Calyptrogenia cuspidata*; (C) *Eugenia involucrata*; (D) *Archirhodomyrtus turbinata*; (E) *Luma apiculata*; (F) *Myrcia splendens*; (G) *Campomanesia adamantium*; (H) *Myrciaria floribunda*; (I) *Eugenia puniceifolia*; (J) *Hottea neibensis*; (K) *Myrcia sp1* (voucher T. Vasconcelos 307); (L) *Gossia clusioides*; (M) *Chamguava schippii*; (N) *Siphoneugenia densiflora* (O) *Myrtastrum rufopunctatum*. Size of reproductive structures varies between c. 0.5 to 3cm. Pictures by R. Aguilar (M) and T. Vasconcelos (all besides M).

Figure 2: Myrteae ML phylogenetic tree resulting from the combined dataset analysis. Bootstrap percentages greater than 50 are shown above branches; clades receiving posterior probabilities greater than 0.95 in equivalent BI analysis are indicated by thicker branches. Arrows indicate clades that were not recovered in BI analysis. \*Clade numbers *sensu* Mazine et al. (2014). \*\*Clade numbers *sensu* Lucas et al. (2011). ‘Spe’: section *Speciosae sensu* Büniger et al. (2016).

Figure 3: Global species distribution of Myrteae, as sourced from WCSP (2016).

Figure 4: Comparative dating analysis in Myrteae generated by Beast and based on two distinct fossil sets. (A) Calibration using macrofossil dataset (approach A). (B) Calibration using microfossil dataset (approach B). “a” and “b” indicate Myrteae stem and crown nodes respectively. Highlighted areas

show divergence between the three major clades (Australasian and *Myrtus* groups and the Main Neotropical lineage) in each calibration. Fossil placements used to calibrate each chronogram are marked with red arrows and refer to estimations presented in Table 1.

Figure 5: Biogeographic inference recovered from BioGeoBEARS analysis in phylogenies dated with (A) Macrofossil dataset ( $j=0.0574$ ;  $\text{LnL}=-156.72$ ), and (B) pollen fossil data set ( $j=0.055$ ;  $\text{LnL}=-161.48$ ). “a” and “b” represent Myrteae stem and crown node respectively. Range shifts are numerated above pie charts.

Figure 6: Phylorate showing the single best shift configuration recovered from BAMM in chronograms resulting from (A) macrofossil calibration and (B) pollen fossil calibration. Three accelerating shifts on diversification rates (marked by “a”, “b” and “c”) are detected in each case. Color coding (blue to red) is in scale of species per million years.

Figure 7: Graph comparing crown node ages of macrofossil calibration (orange) and pollen fossil calibration (blue). Bars show confidence intervals per node.

### Supplementary Material

S1: Primers.

S2: PCR conditions.

S3: Combined dataset alignment (nexus format).

S4: Previous studies with Myrteae dating estimates.

S5: Myrteae fossil survey.

S6: BioGeoBEARS files.

S7: Bamm files.

S8: BI phylogeny based on cpDNA dataset.

S9: BI phylogeny based on nuclear (ITS) dataset.

S10: Chronogram based on macrofossil calibration (newick format)

S11: Chronogram based on pollen fossil calibration (newick format)



|  |                    |                            |            |            |            |            |            |            |            |            |
|--|--------------------|----------------------------|------------|------------|------------|------------|------------|------------|------------|------------|
|  | 588                |                            |            |            |            |            |            |            |            |            |
| <i>Campomanesia adamantium</i> (Cambess.) O.Berg         | T. Vasconcelos 474 | Brazil (Minas Gerais)      | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Campomanesia velutina</i> (Cambess.) O.Berg           | T. Vasconcelos 507 | Brazil (Distrito Federal)  | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Chamguava schippii</i> (Standl.) Landrum              | D. Aguilar 9833    | Costa Rica                 | this study | this study | this study | this study | this study | this study | this study | this study |
| <i>Curitiba prismatica</i> (D.Legrand) Salywon & Landrum | D.F. Lima 551      | Brazil (Paraná)            | this study | this study | this study | this study | this study | this study | this study | this study |
| <i>Decaspermum fruticosum</i> J.R.Forst. & G.Forst       | T. Vasconcelos 730 | Malaysia (Sabah)           | this study | this study | this study | this study | this study |            |            | this study |
| <i>Decaspermum humile</i> (Sweet ex G.Don) A.J.Scott     | S. Belsham M82     | RGB Melbourne (cultivated) | AM234128   | this study | AY498780*  | AM489824   | this study |            | this study |            |
| <i>Decaspermum vitis-idaea</i> Stapf                     | T. Vasconcelos 729 | Malaysia (Sabah)           | this study |            | this study | this study | this study | this study | this study |            |
| <i>Eucalyptus perriniana</i> F.Muell. ex Rodway          | E. Lucas 283       | RBG Kew (cultivated)       | AM234139   | AM489985   | this study | AM489825   | this study | this study | this study | this study |
| <i>Eugenia acutata</i> Miq.                              | T. Vasconcelos 506 | Brazil (Distrito Federal)  | this study |            | this study | this study | this study | this study |            | this study |
| <i>Eugenia adenocalyx</i> DC.                            | A. Giaretta 1441   | Brazil (Roraima)           | this study |            | this study | this study | this study | this study |            | this study |
| <i>Eugenia angustissima</i> O.Berg                       | T. Vasconcelos 405 | Brazil (Goias)             | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Eugenia azurensis</i> O.Berg                          | J.E.Q. Faria 4186  | Brazil (Bahia)             | this study |            | this study | this study | this study | this study | this study |            |
| <i>Eugenia biflora</i> (L.) DC.                          | F.F. Mazine 1075   | Brazil                     | KJ187610   | this study | this study | KJ469659   |            |            | this study |            |
| <i>Eugenia brevistyla</i> D.Legrand                      | F.F. Mazine 993    | Brazil                     | KJ187614   |            | this study | KJ469663   |            |            | this study |            |
| <i>Eugenia bullata</i> Pancher ex Guillaumin             | T. Vasconcelos 608 | New Caledonia              | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Eugenia bunchonsiifolia</i> Nied.                     | T. Vasconcelos 466 | Brazil (Espírito Santo)    | this study |            | this study | this study | this study | this study |            | this study |
| <i>Eugenia involucrata</i> DC.                           | T. Vasconcelos 256 | Brazil (Distrito Federal)  | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Eugenia longiracemosa</i> Kiaersk.                    | T. Vasconcelos 310 | Brazil (Amazonas)          | this study |            | this study | this study | this study | this study |            | this study |
| <i>Eugenia monticola</i> (Sw.) DC.                       | T. Vasconcelos 566 | Dominican Republic         | this study | JQ588481*  | this study | this study | this study | this study | this study | this study |
| <i>Eugenia myrcianthes</i> Nied.                         | Savassi ESA 85681  | Brazil                     | KJ187652   | this study | AY498784   | KJ469702   | this study | this study |            | this study |
| <i>Eugenia paludosa</i> Pancher ex Brongn. & Gris        | T. Vasconcelos 646 | New Caledonia              | this study |            | this study | this study | this study | this study |            | this study |

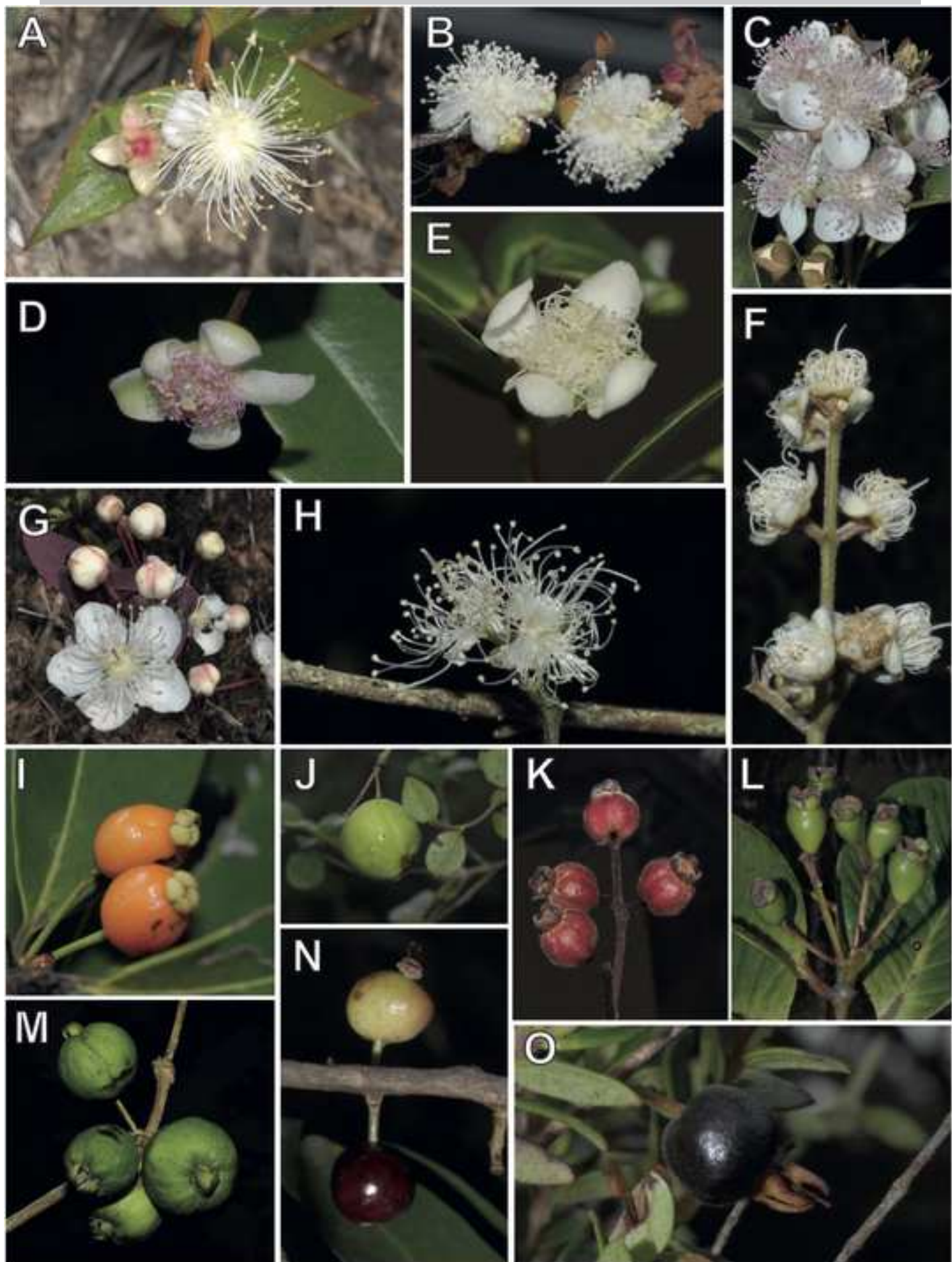
|   |                     |                            |            |           |            |            |            |            |            |            |
|---|---------------------|----------------------------|------------|-----------|------------|------------|------------|------------|------------|------------|
| <i>Eugenia paracatuana</i> O.Berg                           | P.O. Rosa 1399      | Brazil (Goias)             | this study |           |            | this study | this study | this study |            | this study |
| <i>Eugenia puniceifolia</i> (Kunth) DC.                     | F.F. Mazine 1065    | Brazil (Mato Grosso)       | this study |           | this study | AM489827*  |            |            | this study |            |
| <i>Eugenia reinwardtiana</i> (Blume) DC.                    | B. Holst 8870       | MSBG (cultivated)          | this study | KM894685* | this study |            | AY463131*  |            | this study |            |
| <i>Eugenia roseopetiolata</i> N.Snow & Cable                | T. Vasconcelos s.n. | RBG Kew (cultivated)       | this study |           | this study | this study | this study | this study | this study | this study |
| <i>Eugenia stipitata</i> McVaugh                            | T. Vasconcelos 677  | Singapore BG (cultivated)  | this study |           | this study | this study | this study | this study | this study |            |
| <i>Eugenia uniflora</i> L.                                  | E. Lucas 207        | RBG Kew (cultivated)       | AM234088   | AM489986  | this study | AM489828   | AF215627*  |            | KP722326   | KP722202   |
| <i>Eugenia yumana</i> Alain                                 | T. Vasconcelos      | Dominican Republic         | this study |           | this study | this study | this study | this study | this study | this study |
| <i>Gossia clusoides</i> (Brongn. & Gris) N.Snow             | J. Soewarto HB 14   | New Caledonia              | this study |           | this study | this study | this study | this study | this study | this study |
| <i>Hottea neibensis</i> Alain                               | T. Vasconcelos 590  | Dominican Republic         | this study |           | this study | this study | this study | this study | this study | this study |
| <i>Kanakomyrtus dawsoniana</i> N.Snow                       | T. Vasconcelos 639  | New Caledonia              | this study |           | this study | this study | this study | this study |            |            |
| <i>Legrandia concinna</i> (Phil.) Kausel                    | RBGE 1999-0656      | RBG Edinburgh (cultivated) | AM234072   | AM489990  | this study | AM489839   |            |            |            |            |
| <i>Lenwebbia prominens</i> N.Snow & Guymer                  | N. Snow 7463        | Australia (Queensland)     | this study | AY521538* |            | this study |            | this study |            |            |
| <i>Leptospermum scoparium</i> J.R.Forst. & G.Forst.         | E. Lucas 284        |                            | AM234142   | AM489991  | AM235423   | AM489840   | AM235459   |            | KF591267   |            |
| <i>Lophomyrtus obcordata</i> (Raoul) Burret                 | S. Belsham M41      | New Zealand                | AM234146   | AM489993  | this study | AM489842   | this study | this study |            |            |
| <i>Luma apiculata</i> (DC.) Burret                          | E. Lucas 208        | RBG Kew (cultivated)       | AM234101   | AM489995  | AY498795   | AM489843   | JN660959*  | this study | KP722331   | KP722209   |
| <i>Marlierea umbraticola</i> (Kunth) O.Berg                 | M.A.D. Souza s.n.   | Brazil (Amazonas)          | KP722392   |           | KP722470   | KP722300   | this study | this study | KP722350   | KP722246   |
| <i>Metrosideros nervulosa</i> C.Moore & F.Muell.            | (all from GenBank)  |                            | JF950784   | DQ088535  | AY498802   |            | DQ088395   |            | JF950929   |            |
| <i>Metrosideros perforata</i> (J.R.Forst. & G.Forst.) Druce | E. Lucas 209        | RBG Kew (cultivated)       | AM234141   | AM489998  | this study | AM489848   | this study | this study | this study |            |
| <i>Metrosideros stipularis</i> (Hook. & Arn.) Hook.f.       | (all from GenBank)  |                            | AM234071   | AF368222  |            | AM489884   |            |            |            |            |
| <i>Mitranthes clarendonensis</i> (Proctor) Proctor          | T. Vasconcelos 511  | Jamaica                    | this study |           | this study | this study | this study |            | this study | this study |
| <i>Mitranthes glabra</i> Proctor                            | E. Lucas 1224       | Jamaica                    | this study |           | this study | this study | this study | this study | this study | this study |
| <i>Mosiera longipes</i> (O.Berg) Small                      | Salywon 1183        | U.S.A. (Florida)           | this study |           | this study | this study | this study | this study | this study |            |

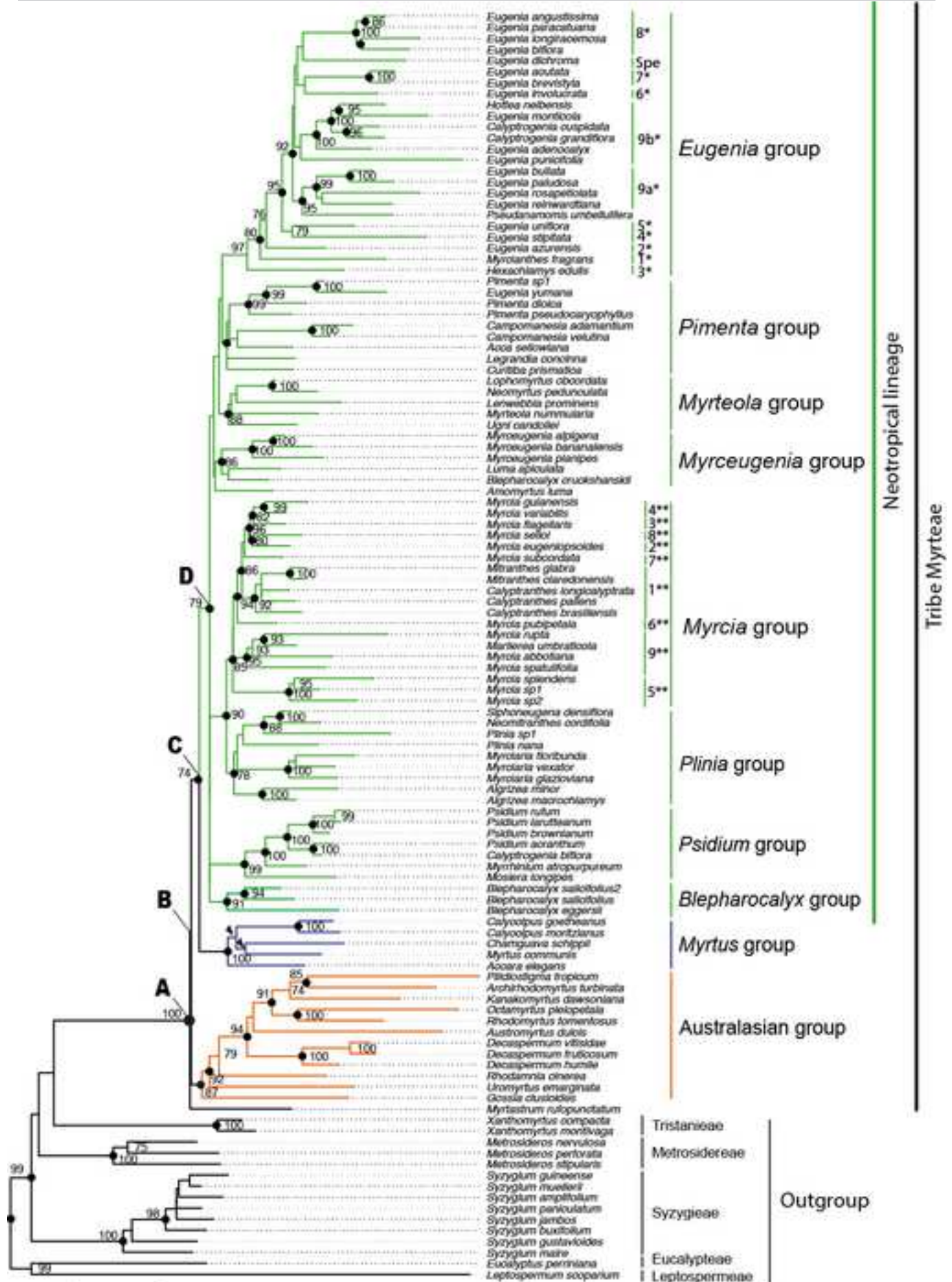


|  |                    |  |            |            |            |            |            |            |            |            |
|--|--------------------|--|------------|------------|------------|------------|------------|------------|------------|------------|
| <i>Myrceugenia alpigena</i> (DC.) Landrum                          | E. Lucas 167       | Brazil (Minas Gerais)                    | AM234098   | JN660991   | KP722441   | AM489854   | JN660941.  | this study | KP722376   | JN661090   |
| <i>Myrceugenia bananalensis</i> Bezerra & Landrum                  | J.E.Q. Faria 4049  | Brazil (Distrito Federal)                | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrceugenia planipes</i> (Hook. & Arn.) O.Berg                  | L. Landrum s.n.    | Chile                                    | this study | JN661027*  | this study | this study | this study | this study |            |            |
| <i>Myrcia abbotiana</i> (Urb.) Alain                               | T. Vasconcelos 571 | Dominican Republic                       | this study |            |            |            | this study | this study |            |            |
| <i>Myrcia rupta</i> M.L.Kawas. & B.Holst                           | T. Vasconcelos 311 | Brazil (Amazonas)                        | this study |            | this study | this study | this study | this study |            | this study |
| <i>Myrcia eugeniopsoides</i> (D.Legrand & Kausel) Mazine           | E. Lucas 61        | Brazil (Sao Paulo)                       | AM234107   | AM489996   | KP722429   | AM489845   | this study | this study | JN091327   | KP722205   |
| <i>Myrcia flagellaris</i> (D.Legrand) Sobral                       | E. Lucas 83        | Brazil (Sao Paulo)                       | AM234113   | AM489989   | KP722430   | AM489836   | this study | this study | JN091350   | KP722206   |
| <i>Myrcia guianensis</i> (Aubl.) DC.                               | Harley 50307       | Brazil                                   | JN091225   | this study | this study | this study | this study |            | JN091351   |            |
| <i>Myrcia pubipetala</i> Miq.                                      | E. Lucas 86        | Brazil (Sao Paulo)                       | AM234114   | AM490001   | KP722426   | AM489855   | this study | this study | JN091364   | KP722273.  |
| <i>Myrcia selloi</i> (Spreng.) N.Silveira                          | E. Lucas 110       | Brazil                                   | JN091240   | JN091315   | KP722436   | JN091431   | this study | this study | JN091371   | KP722212   |
| <i>Myrcia sp2</i>  | J.E.Q. Faria 4193  | Brazil (Bahia)                           | this study |            | this study | this study | this study | this study |            |            |
| <i>Myrcia sp1</i>  | T. Vasconcelos 307 | Brazil (Amazonas)                        | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrcia spathulifolia</i> Proença                                | J.E.Q. Faria 4214  | Brazil (Bahia)                           | this study |            | this study | this study | this study | this study |            | this study |
| <i>Myrcia splendens</i> (Sw.) DC.                                  | T. Vasconcelos 587 | Dominican Republic                       | this study |            | this study | this study | this study | this study | this study |            |
| <i>Myrcia subcordata</i> DC.                                       | M. Santos 586      | Brazil (Minas Gerais)                    | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrcianthes fragrans</i> (Sw.) McVaugh                          | B. Holst 8862      | Guyane                                   | KJ187655   | KJ772955   | AY498803*  | KJ469705   |            |            |            |            |
| <i>Myrciaria floribunda</i> (H.West ex Willd.) O.Berg              | T. Vasconcelos 388 | Brazil (Amazonas)                        | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrciaria glazioviana</i> (Kiaersk.) G.M.Barroso ex Sobral      | T. Vasconcelos 413 | Brazil (Bahia)                           | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrciaria vexator</i> McVaugh                                   | T. Vasconcelos 709 | Singapore BG (cultivated)                | this study | AY521544*  | this study | this study | this study | this study | this study | this study |
| <i>Myrrhinium atropurpureum</i> Schott in K.P.J.Sprengel           | Costa, I.R. 594    | Brazil (Rio de Janeiro)                  | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Myrtastrum rufopunctatum</i> (Pancher ex Brongn. & Gris) Burret | J. Soewarto HB 10  | New Caledonia RBG Edinburgh (cultivated) | this study | this study | this study | this study | this study | this study | this study | this study |
| <i>Myrteola nummularia</i> (Lam.) O.Berg                           | RBGE 1996-1096     | RBG Edinburgh (cultivated)               | AM234068   | AM490008   | this study | AM489871   | this study | this study | this study | this study |
| <i>Myrtus communis</i> L.  | E. Lucas 211       | RBG Kew (cultivated)                     | AM234149   | AM490009   | this study | AM489872   | JN660939*  | this study | KP722327   | KP722221   |
| <i>Neomitranthes cordifolia</i> (D.Legrand) D.Legrand              | Forster 1011       | Brazil                                   | AM489410   |            |            | AM489569   | this study | this study | JN091386   | this study |

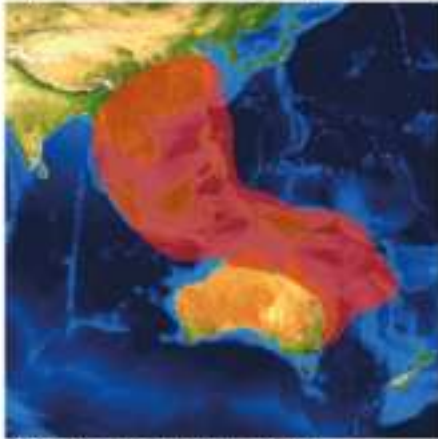
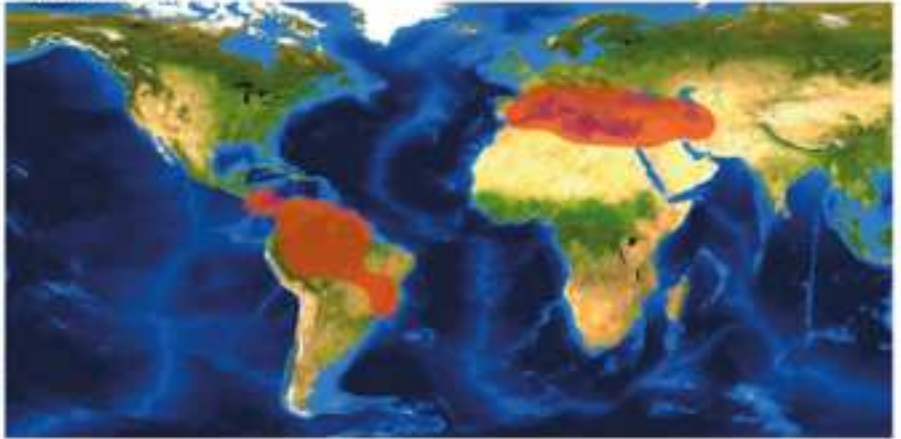
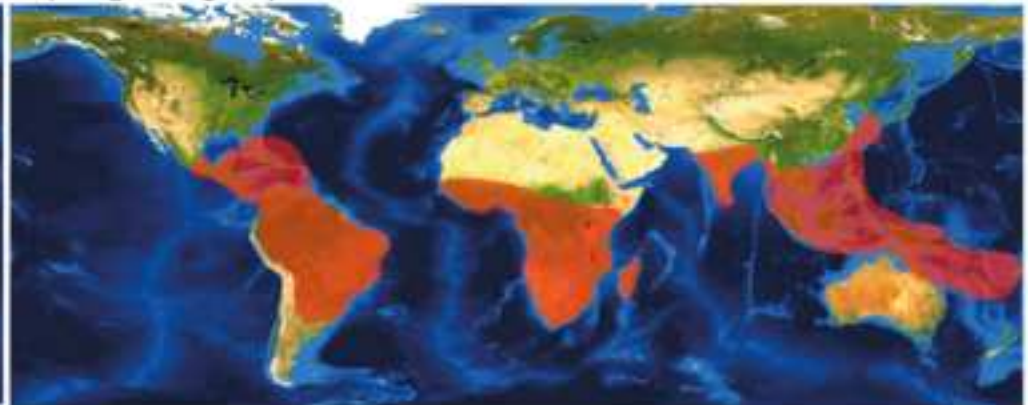
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|---|--------------------|---------------------------|------------|------------|------------|------------|------------|------------|------------|------------|
| <i>Neomyrtus pedunculata</i> (Hook.f.) Allan        | S. Belsham M42     | New Zealand               | AM234144   | AM490010   |            | AM490637   | this study |            |            |            |
| <i>Octamyrtus pleiopetala</i> Diels                 | R. Johns s.n.      | New Guinea                | AM234130   |            | this study | AM489873   | this study | this study | this study |            |
| <i>Pilidiostigma tropicum</i> L.S.Sm.               | Forster 27636      | Australia (Queensland)    | this study |            | this study | this study |            | this study |            | this study |
| <i>Pimenta dioica</i> (L.) Merr.                    | E. Lucas 212       | RBG Kew (cultivated)      | AM234081   | AM490011   | this study | AM489874   | this study | this study | this study |            |
| <i>Pimenta pseudocaryophyllus</i> (Gomes) Landrum   | E. Lucas 161       | Brazil                    | AM234083   | AM490013   | this study | AM489876   | this study | this study | this study | this study |
| <i>Pimenta</i> sp1                                  | T. Vasconcelos 576 | Dominican Republic        | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Plinia nana</i> Sobral                           | F.F. Mazine 662    | Brazil (Minas Gerais)     | this study |            |            | this study | this study | this study | this study | this study |
| <i>Plinia</i> sp1                                   | B. Holst 9482      | French Guiana             | this study |            | this study | this study | this study | this study | this study |            |
| <i>Pseudanmomis umbellulifera</i> (Kunth) Kausel    | T. Vasconcelos 572 | Dominican Republic        | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Psidium acranthum</i> Urb.                       | T. Vasconcelos 578 | Dominican Republic        | this study |            | this study | this study | this study | this study |            | this study |
| <i>Psidium brownianum</i> Mart. ex DC.              | T. Vasconcelos 465 | Brazil (Bahia)            | this study |            | this study | this study | this study | this study | this study | this study |
| <i>Psidium laruotteanum</i> Cambess.                | J.E.Q. Faria 2362  | Brazil (Bahia)            |            | this study | this study | this study | this study | this study | this study | this study |
| <i>Psidium rufum</i> Mart. ex DC.                   | J.E.Q. Faria 4270  | Brazil (Minas Gerais)     | this study |            | this study | this study | this study | this study | this study |            |
| <i>Rhodamnia cinerea</i> Jack                       | T. Vasconcelos 672 | Singapore                 | this study | KJ709064*  | this study | this study | this study | this study | this study | this study |
| <i>Rhodomyrtus tomentosa</i> (Aiton) Hassk          | T. Vasconcelos 678 | Singapore BG (cultivated) | this study | AF105093*  | this study | this study | this study | this study | this study | this study |
| <i>Siphoneugena densiflora</i> O.Berg               | F.F. Mazine 1050   | Brazil                    | AM489412   |            | KP722444   | AM489571   | this study | this study | JN091389   | KP722220   |
| <i>Syzygium amplifolium</i> L.M.Perry               | (all from GenBank) |                           | EF026620   | DQ088556   | DQ088381   |            | DQ088416   |            |            |            |
| <i>Syzygium buxifolium</i> Hook. & Arn.             | (all from GenBank) |                           | KP093045   | KP093852   | DQ088491   | KJ687225   | DQ088424   |            | AB817604   |            |
| <i>Syzygium guineense</i> (Willd.) DC.              | (all from GenBank) |                           | EF026628   | DQ088581   | DQ088500   |            | DQ088432   |            |            |            |
| <i>Syzygium gustavioides</i> (F.M.Bailey) B.Hyland  | (all from GenBank) |                           | AY187194   | DQ088582   | DQ088501   |            | DQ088433   |            |            |            |
| <i>Syzygium jambos</i> (L.) Alston in H.Trimen      | E. Lucas 214       | RBG Kew (cultivated)      | AM234135   | AM490017   | this study | AM489882   | DQ088434*  | this study | this study |            |
| <i>Syzygium muellerii</i> (Miq.) Miq.               | (all from GenBank) |                           | EF026634   | DQ088593   | DQ088511   |            | DQ088439   |            |            |            |
| <i>Syzygium maire</i> (A.Cunn.) Sykes & Garn.-Jones | NZFRI29089         | New Zealand               | KM064865   | KM065310   | DQ088508   | AM489883   | DQ088438   |            |            |            |
| <i>Syzygium oblatum</i> (Roxb.) Wall. ex            | (all from          |                           | KR532632   | AB924759   |            | KR532989   |            |            |            |            |

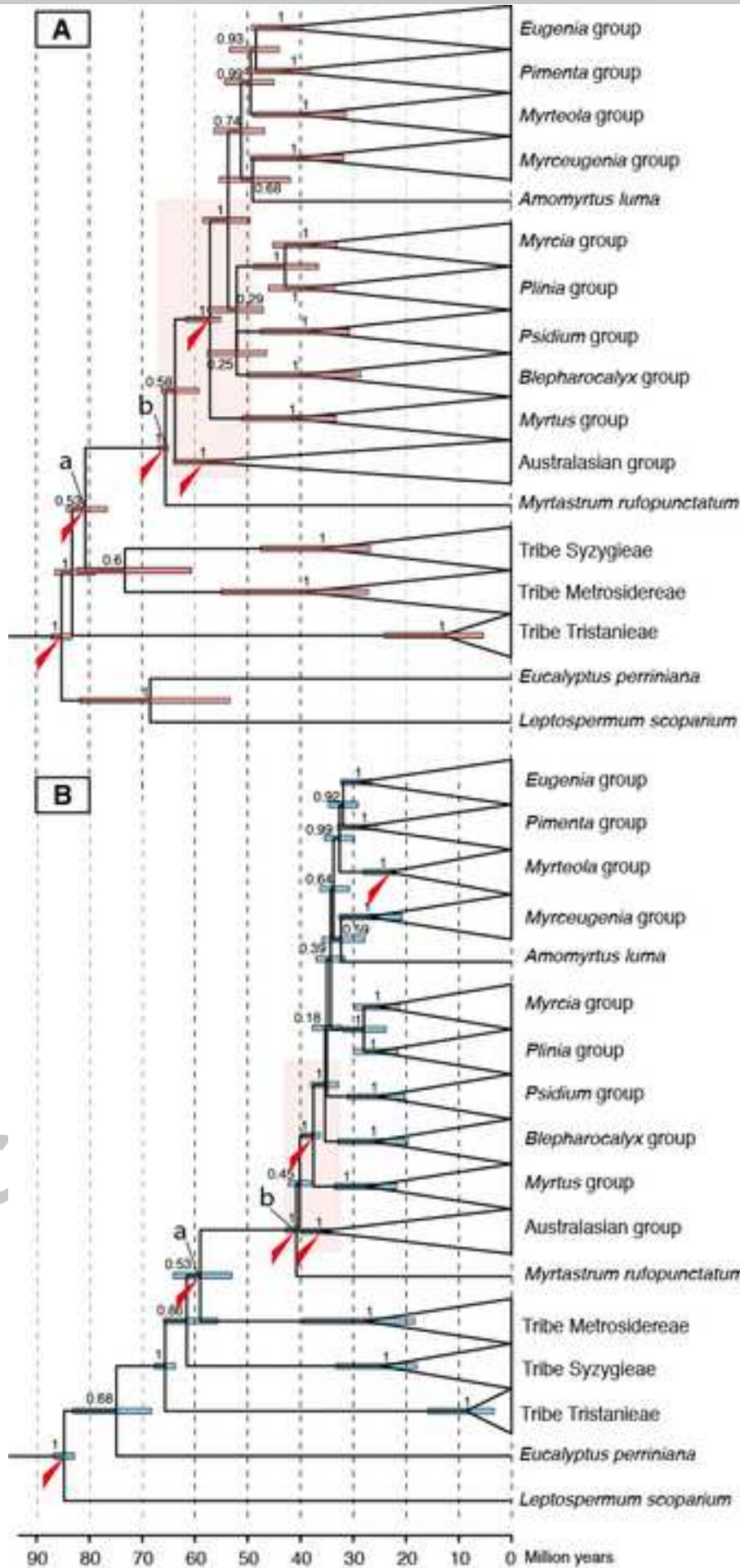
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|---|------------------------|----------------------|------------|------------|------------|------------|------------|------------|------------|------------|
| A.M.Cowan & Cowan   | GenBank)               |                      |            |            |            |            |            |            |            |            |
| <i>Syzygium paniculatum</i> Gaertn.                         | (all from<br>GenBank)  |                      | KM065112   | KM065271   | DQ088515   |            | DQ088441   |            |            |            |
| <i>Ugni candollei</i> (Barnéoud) O.Berg                     | T. Vasconcelos<br>s.n. | RBG Kew (cultivated) | this study | this study | this study | this study | this study | this study | this study | this study |
| <i>Uromyrtus emarginata</i> (Pancher ex<br>Baker f.) Burret | T. Vasconcelos<br>628  | New Caledonia        | this study | this study | this study | this study | this study | this study |            |            |
| <i>Xanthomyrtus compacta</i> (Ridl.) Diels                  | P. Edwards<br>4214A    | New Guinea           | AM234148   |            | this study | AM489887   | this study | this study | this study | this study |
| <i>Xanthomyrtus montivaga</i> A.J.Scott                     | E. Lucas 16            | New Guinea           | AM234147   |            | this study | AM489886   | this study | this study |            |            |





A) Australasian group

B) *Myrtus* groupC) *Myrcia*, *Psidium*, *Plinia*,  
*Pimenta*, *Blepharocalyx* groupsD) *Myrteola* groupE) *Myrceugenia* groupF) *Eugenia* group



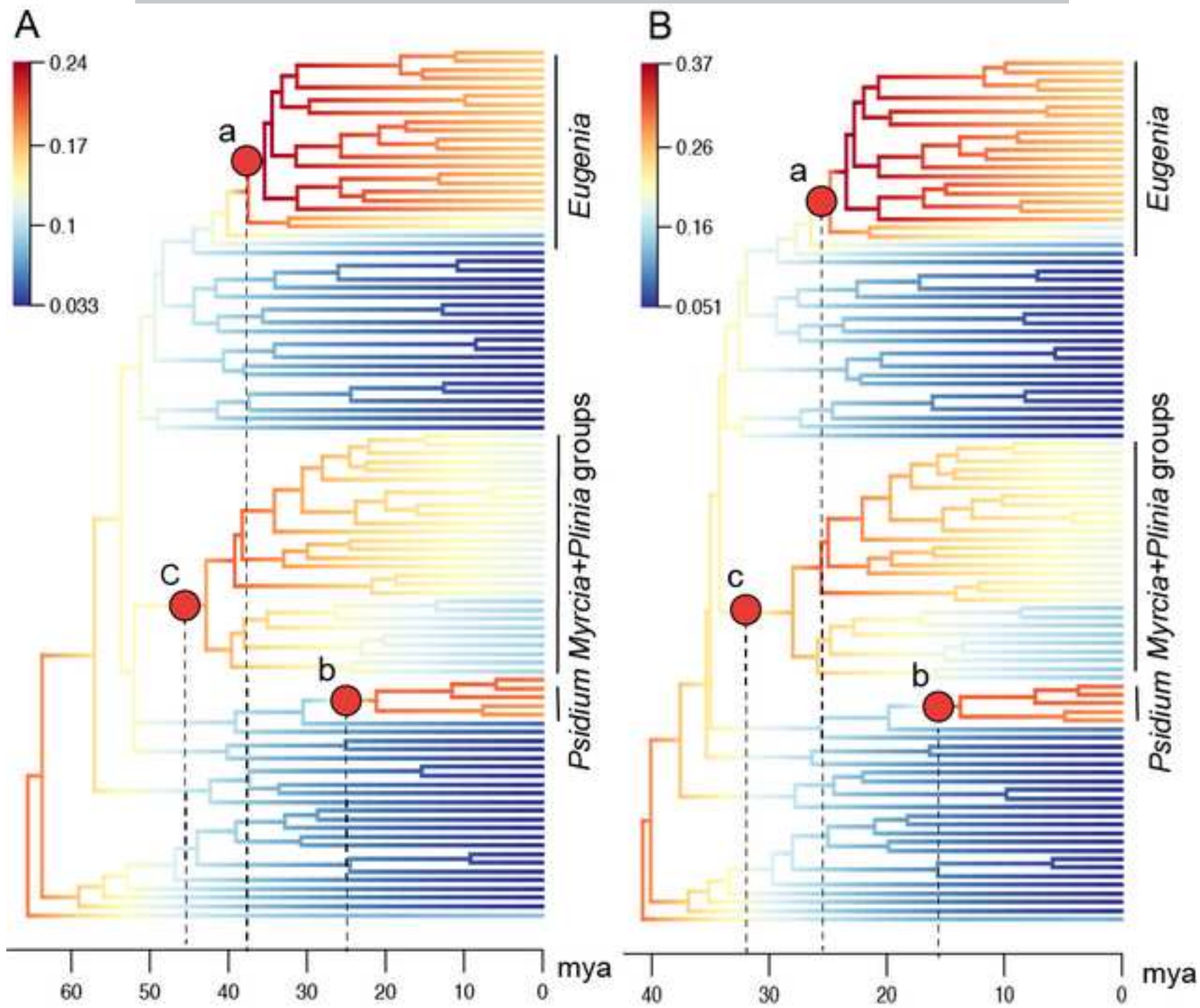
Tribe Myrteae

Tribe Myrteae

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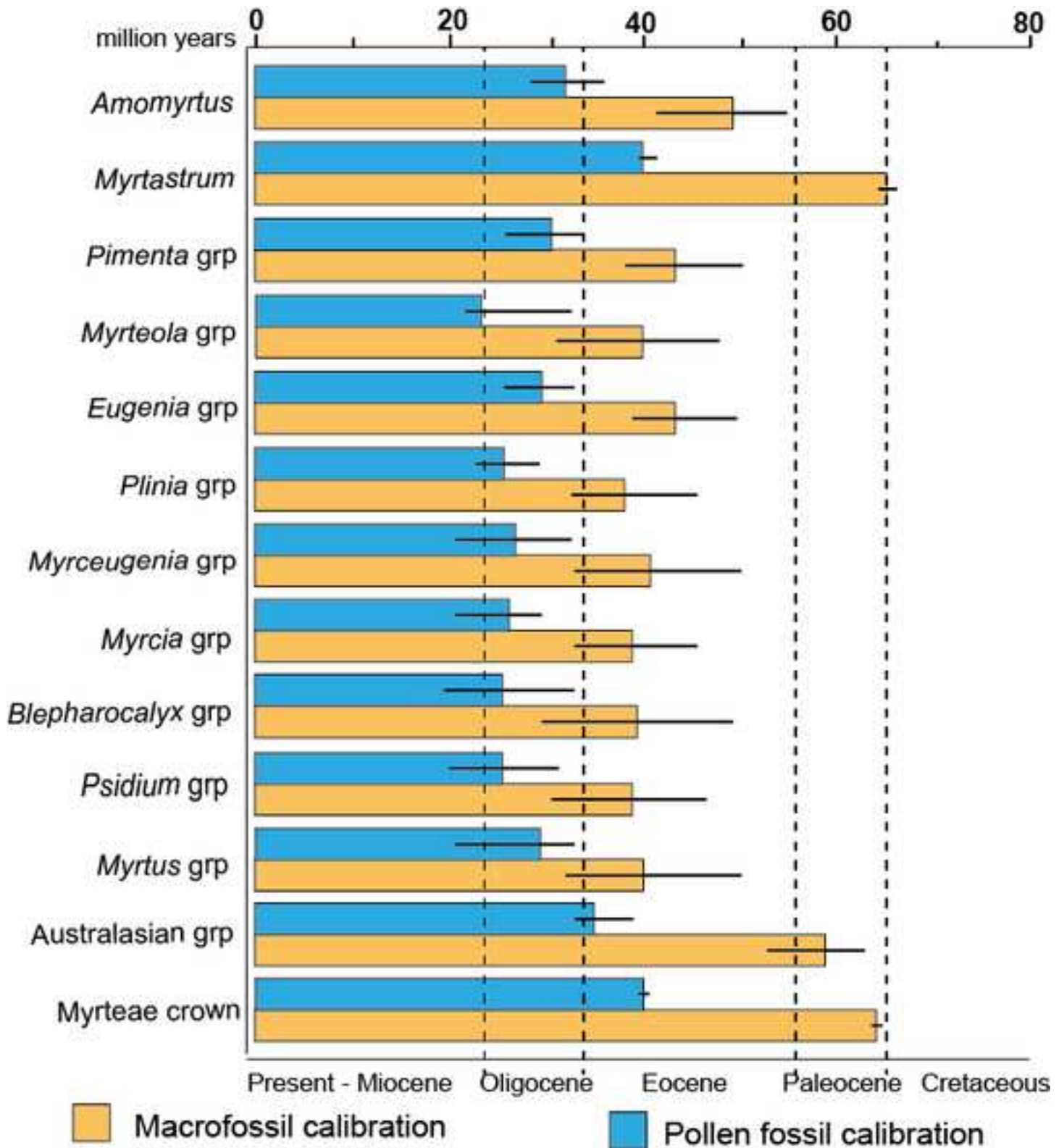


Table 1

| <b>Approach A: Macrofossil</b>                       | Node                      | Age (in million years ago) | Rate      |
|--|---------------------------|----------------------------|-----------|
| <i>Myrceugenelloxylon antarcticus</i>                | Myrteae crown             | 66 (late-Cretaceous)       | Lognormal |
| <i>Myrtineoxylon maomingensis</i>                    | Australasian group crown  | 40 (Mid-Eocene)            | Lognormal |
| <i>Paleomyrtinae princetonensis</i>                  | Neotropical lineage crown | 56 (late-Palaeocene)       | Lognormal |
| <b>Approach B – Pollen fossil</b>                    |                           |                            |           |
| Secondary calibration point – Thornhill et al. 2012  | Crown BKMST               | 63.1 (early-Paleocene)     | Normal    |
| Secondary calibration point – Thornhill et al. 2012  | Crown Myrteae             | 41 (early-Eocene)          | Normal    |
| <i>Myrtaceideites verrucosus</i> (Panama, Argentina) | Neotropical lineage crown | 37.2 (late-Eocene)         | Lognormal |
| <i>Myrtaceideites verrucosus</i> (Australia)         | Australasian group crown  | 35 (late-Eocene)           | Lognormal |
| <i>Myrtaceideites verrucosus</i> (New Zealand)       | Myrteola group crown      | 23 (late-Oligocene)        | Lognormal |
| <b>Both approaches:</b>                              |                           |                            |           |
| Secondary calibration point – Berger et al. (2016)   | Myrtaceae crown           | 85 (Cretaceous)            | Normal    |

Table 2

| Clade  | Approach A (Macrofossil) Age (95% HPD)<br>in million of years |                       | Approach B (Microfossil) Age (95% HPD)<br>in million of years |                       |
|--|---|-----------------------|---|-----------------------|
|  | Stem  | Crown                 | Stem  | Crow                  |
| Myrteae  | 80.72 (76.64 – 84.27)   | 65.55 (65.03 – 66.80) | 58.96 (53.00 – 64.07)   | 40.76 (40.03 – 42.76) |
| Australasian<br>Lineage<br>(Australasian<br>group) | 63.73 (59.25 – 66.24)   | 59.05 (52.80 – 63.96) | 40.09 (38.01 – 42.22)   | 36.88 (34.16 – 39.62) |
| <i>Myrtus</i> group                                | 57.09 (55.06 – 61.68)   | 42.34 (33.20 – 51.04) | 37.56 (36.27 – 39.73)   | 27.78 (21.80 – 33.60) |
| <i>Psidium</i> group                               | 52.03 (46.33 – 57.60)   | 39.12 (30.75 – 47.47) | 35.01 (32.34 – 37.70)   | 25.62 (20.14 – 31.07) |
| <i>Blepharocalyx</i><br>group                      | 52.03 (46.33 – 57.60)   | 40.15 (28.49 – 49.95) | 35.36 (32.80 – 38.03)   | 26.38 (19.64 – 32.90) |
| <i>Myrcia</i><br>supergroup                        | 42.85 (36.57 – 48.76)   | 39.19 (33.04 – 45.17) | 27.99 (23.83 – 31.98)   | 25.58 (21.32 – 29.73) |
| <i>Myrceugenia</i><br>group                        | 49.00 (41.84 – 55.34)   | 41.40 (31.72 – 49.42) | 32.32 (27.85 – 35.86)   | 27.33 (20.83 – 32.62) |
| <i>Plinia</i> group                                | 42.85 (36.57 – 48.76)   | 39.61 (33.35 – 46.00) | 27.99 (23.83 – 31.98)   | 25.86 (21.66 – 29.93) |
| <i>Eugenia</i><br>supergroup                       | 48.36 (44.01 – 53.22)   | 44.42 (39.58 – 49.17) | 31.93 (29.16 – 34.63)   | 29.29 (26.55 – 32.29) |

Table 3:

| Shift Number (Figure 5) | Approach A shifts   | Area shift                        | Age (CI 95%)          | Geological time | Likely nature of event inferred by period age                                   |
|-------------------------|---|-----------------------------------|-----------------------|-----------------|---|
| 1                       | Neotropical stem - crown  | NCNZ - South America              | 63.73 (59.25 - 66.24) | early-Paleocene | Land migration and vicariance   |
| 2                       | Australasian group - first shift to Australia                   | NCNZ - Australia+NG               | 55.93 (49.52 - 61.56) | early-Eocene    | Short distance dispersal and/or vicariance                                      |
| 3                       | Australasian group - <i>Rhodamnia</i>                           | Australia+NG - SE Asia            | 52.89 (46.14 - 58.78) | early-Eocene    | LDDE only   |
| 4                       | Australasian group - shift to Zealandia                         | Australia+NG - NCNZ               | 43.96 (37.16 - 50.39) | mid-Eocene      | Short distance dispersal and/or vicariance                                      |
| 5                       | Australasian group - second shift to Australia                  | NCNZ - Australia+NG               | 28.64 (20.27 - 36.84) | early-Oligocene | Short distance dispersal and/or vicariance                                      |
| 6                       | Australasian group - <i>Rhodomyrtus</i>                         | NCNZ - SE Asia                    | 30.76 (22.17 - 38.85) | early-Oligocene | LDDE, but lower CI limit also allows short distance dispersal and/or vicariance |
| 7                       | Australasian group - <i>Decaspermum</i>                         | Australia+NG - SE Asia            | 24.52 (15.79 - 33.66) | late-Oligocene  | LDDE, but lower CI limit also allows short distance dispersal and/or vicariance |
| 8                       | <i>Myrtus</i> group - North American shift                      | South America to Central+North Am | 57.08 (55.06 - 61.68) | late-Paleocene  | Short distance dispersal and/or vicariance                                      |
| 9                       | <i>Myrtus</i> group - <i>Myrtus</i>                             | South America to Mediterranean EU | 42.34 (33.19 - 51.04) | mid-Eocene      | Short distance dispersal and/or vicariance                                      |
| 10                      | <i>Myrtus</i> group - South America shift ( <i>Calycolpus</i> ) | Central+North Am to South America | 37.37 (28.58 - 46.19) | late-Eocene     | LDDE only   |
| 11                      | <i>Myrtus</i> group - South America shift ( <i>Accara</i> )     | Central+North Am to South America | 33.56 (24 - 42.78)    | early-Oligocene | LDDE only   |
| 12                      | <i>Psidium</i> group - stem                                     | South America to Central+North Am | 52.03 (46.33 - 57.6)  | early-Eocene    | LDDE, but upper CI limit also allows short distance dispersal and/or vicariance |

|    |  |  |                       |                 |  |
|----|--|--|-----------------------|-----------------|--|
| 13 | <i>Psidium</i> group - first shift to South America  | Central+North Am to South America      | 39.12 (30.75 - 47.47) | mid-Eocene      | LDDE only  |
| 14 | <i>Psidium</i> group - Caribbean <i>Psidium</i>      | South America to Central+North Am      | 30.5 (22.7 - 38.74)   | early-Oligocene | LDDE, but lower CI limit also allows short distance dispersal or vicariance  |
| 15 | <i>Psidium</i> group - second shift to South America | Central+North Am to South America      | 21.15 (14.66 - 28.9)  | early-Miocene   | Short distance dispersal or vicariance                                       |
| 16 | <i>Plinia</i> group - <i>Myrciaria</i>               | South America to Central+North Am      | 23.15 (15.89 - 31.29) | late-Oligocene  | LDDE, but lower CI limit also allows short distance dispersal or vicariance  |
| 17 | <i>Plinia</i> group - <i>Myrciaria</i>               | Central+North Am to South America      | 20.23 (12.97 - 28.33) | early-Miocene   | Short distance dispersal and/or vicariance                                   |
| 18 | <i>Myrcia</i> group - first North American shift     | South America to Central+North Am      | 32.98 (26.47 - 40.14) | early-Oligocene | LDDE only  |
| 19 | <i>Myrcia</i> group - shift to South America         | Central+North America to South America | 30.59 (22.72 - 37.25) | early-Oligocene | LDDE, but lower CI limit also allows short distance dispersal and vicariance |
| 20 | <i>Myrcia</i> group - second North American shift    | South America to Central+North Am      | 23.79 (16.89 - 30.79) | late-Oligocene  | LDDE, but lower CI limit also allows short distance dispersal and vicariance |
| 21 | <i>Myrteola</i> group - New Zealand                  | South America to NCNZ                  | 40.64 (31.28 - 48.68) | mid-Eocene      | Short distance dispersal and/or vicariance                                   |
| 22 | <i>Myrteola</i> group - Australia                    | NCNZ - Australia+NG                    | 34.14 (23.40 - 43.89) | late-Eocene     | Short distance dispersal and/or vicariance                                   |
| 23 | Pimenta group - North American shift                 | South America to Central+North Am      | 41.58 (34.48 - 48.24) | mid-Eocene      | LDDE only  |
| 24 | Pimenta group - Pimenta pseudocaryophyllus           | Central+North Am to South America      | 34.08 (26.07 - 41.98) | late-Eocene     | LDDE only  |
| 25 | Eugenia crown - Myrcianthes                          | South America to Central+North Am      | 44.42 (39.58 - 49.17) | mid-Eocene      | LDDE only  |
| 26 | Eugenia - shift back SA                              | Central+North Am to South America      | 42.01 (37.38 - 46.86) | mid-Eocene      | LDDE only  |
| 27 | Eugenia - shift Umbellatae caribbean                 | South America to Central+North Am      | 31.38 (26.55 - 36.41) | early-Oligocene | LDDE only  |
| 28 | Eugenia - shift Umbellatae back to SA                | Central+North Am to South America      | 25.7 (20.33 - 30.93)  | late-Oligocene  | LDDE, but lower CI limit also allows short distance                          |

|              |  |  |                        |                 | dispersal and vicariance   |
|--------------|--|--|------------------------|-----------------|--|
| 29           | Eugenia - Pseudanamomis                  | South America to Central+North Am            | 35.42 (31.02 - 39.08)  | late-Eocene     | LDDE only  |
| 30           | Eugenia - NCNZ                           | Central+North Am to NCNZ                     | 31.24 (25.69 - 36.73)  | early-Oligocene | LDDE only  |
| 31           | Eugenia - Africa                         | NCNZ to Africa                               | 25.72 (20.04 - 31.55)  | late-Oligocene  | LDDE only  |
| 32           | Eugenia - SA Asia                        | Africa to SE Asia                            | 22.75 (16.15 - 28.88)  | early-Miocene   | Land migration   |
| Shift Number | Approach A shifts                        | Nature and timing of tested geological event | Age (HPD 95% interval) | Geological time | Likely nature of event inferred by age                                       |
| 1            | Neotropical stem - crown                 | NCNZ to South America                        | 40.09 (38.01 - 42.21)  | late-Eocene     | Land migration and vicariance  |
| 2            | Australasian grp - first Australia shift | NCNZ to Australia+NG                         | 35.15 (31.99 - 38.61)  | late-Eocene     | Short distance dispersal and/or vicariance                                   |
| 3            | Australasian grp - Rhodamnia             | Australia+NG to SE Asia                      | 33.37 (29.81 - 36.96)  | early-Oligocene | LDDE only  |
| 4            | Australasian grp - shift to Zealandia    | Australia+NG to NCNZ                         | 25 (21.07 - 29)        | late-Oligocene  | Short distance dispersal and/or vicariance                                   |
| 5            | Australasian grp - Rhodomyrtus           | Australia+NG to SE Asia                      | 19.85 (14.64 - 24.64)  | early-Miocene   | Short distance dispersal and/or vicariance                                   |
| 6            | Australasian grp - Decaspermum           | Australia+NG to SE Asia                      | 5.87 (2.75 - 9.9)      | late-Miocene    | Short distance dispersal and/or vicariance                                   |
| 7            | Australasian grp - Pilidiostigma         | NCNZ to Australia+NG                         | 18.23 (13.35 - 23.15)  | early-Miocene   | LDDE, but upper CI limit also allows short distance dispersal and vicariance |
| 8            | Myrtus group - Myrtus                    | South America to Mediterranean EU            | 27.78 (21.79 - 33.60)  | late-Oligocene  | LDDE, but lower CI limit also allows short distance dispersal and vicariance |
| 9            | Myrtus group - Chamguava                 | South America to Central+North Am            | 22.03 (15.88 - 28.22)  | early-Miocene   | Short distance dispersal and/or vicariance                                   |
| 10           | Psidium group - Moseira                  | South America to Central+North Am            | 25.62 (20.14 - 31.07)  | late-Oligocene  | LDDE, but lower CI limit also allows short distance dispersal and vicariance |
| 11           | Psidium group - Caribbean Psidium        | South America to Central+North Am            | 13.73 (9.38 - 18.58)   | mid-Miocene     | Short distance dispersal and/or vicariance                                   |
| 12           | Plinia group - Myrciaria                 | South America to Central+North               | 13.55 (8.38 - 18.86)   | mid-Miocene     | Short distance dispersal   |

|    |  |                                   |                       |                |  |
|----|--|-----------------------------------|-----------------------|----------------|--|
|    |  | Am                                |                       |                | and/or vicariance  |
| 13 | Myrcia group - <i>M. abbotiana</i>           | South America to Central+North Am | 19.59 (14.70 - 24.39) | early-Miocene  | Short distance dispersal and/or vicariance                                   |
| 14 | Myrcia group - <i>Calyptranthes</i>          | South America to Central+North Am | 12.73 (8.27 - 17.35)  | mid-Miocene    | Short distance dispersal and/or vicariance                                   |
| 15 | Myrteola group - Australia                   | South America to Australia+NG     | 23.39 (22.04 - 28.02) | late-Oligocene | Land migration and vicariance  |
| 16 | Myrteola group - New Zealand                 | Australia+NG to NCNZ              | 20.45 (14.55 - 26.16) | early-Miocene  | LDDE, but upper CI limit also allows short distance dispersal and vicariance |
| 17 | Pimenta group - North American shift         | South America to Central+North Am | 22.52 (17.52 - 27.46) | early-Miocene  | Short distance dispersal and/or vicariance                                   |
| 18 | Eugenia - <i>Myrcianthes</i>                 | South America to Central+North Am | 27.72 (24.83 - 30.71) | late-Oligocene | LDDE only  |
| 19 | Eugenia - shift three - <i>Pseudanamomis</i> | South America to Central+North Am | 23.44 (21.88 - 27.99) | late-Oligocene | LDDE, but lower CI limit also allows short distance dispersal and vicariance |
| 20 | Eugenia - NCNZ                               | Central+North Am to NCNZ          | 20.69 (17.24 - 24.1)  | early-Miocene  | LDDE only  |
| 21 | Eugenia - Africa                             | NCNZ to Africa                    | 16.87 (12.07 - 20.43) | early-Miocene  | LDDE only  |
| 22 | Eugenia - SE Asia                            | Africa to SE Asia                 | 14.96 (10.82 - 19.06) | mid-Miocene    | Land migration   |
| 23 | Eugenia - shift two - <i>Umbellatae</i>      | South America to Central+North Am | 16.93 (13.58 - 20.36) | early-Miocene  | Short distance dispersal and/or vicariance                                   |



Graphical abstract



A