

## Anti-inflammatory and toxicity studies of atranorin extracted from *Cladina kalbii* Ahti in rodents

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Atranorin (ATR) is the main compound from the lichen *Cladina kalbii* Ahti, which grows in the arid regions of northeastern Brazil. This study was conducted to evaluate the anti-inflammatory and toxicological properties of ATR. To evaluate anti-inflammatory properties, paw edema was induced by injecting 0.1 mL of carrageenan into the subplantar region of the right hind paw of rats, and leukocyte migration was induced by injection of 500 µL of carrageenan into the peritoneal cavity of mice. In addition, we determined ATR cytotoxicity in L929 cells by MTT assay and acute (5 g/kg-single dose) and subchronic (50 mg/kg-30 days) toxicity tests in Wistar rats. The results showed that ATR (100 mg/kg and 200 mg/kg) exhibited significant anti-inflammatory activity (paw edema and leukocyte migration). In the acute toxicity test, the animals showed hypoactivity and lethargy during the initial period (first 6 hours) and increase in total protein, total and indirect bilirubin, and alkaline phosphatase after 14 days in ATR-treated male rats. The subchronic toxicity test revealed increases in total protein, globulin, gamma-glutamyl transferase, alkaline phosphatase, and total and direct bilirubin in ATR-treated female rats. Histological analysis revealed no changes in the architecture and morphology of the organs. These results suggest that ATR has significant anti-inflammatory activity, with no significant acute and subchronic toxicity or cytotoxicity.

**Uniterms:** *Cladina kalbii*/pharmacognosy. Atranorin/anti-inflammatory activity. Atranorin/toxicity. Medicinal plants.

Atranorina (ATR) é o principal composto do líquen *Cladina kalbii* Ahti, que cresce em terras áridas do nordeste brasileiro. Este estudo foi realizado para avaliar as propriedades antiinflamatórias e toxicológicas da ATR. Para avaliar as propriedades antiinflamatórias, o edema de pata foi induzido, administrando-se 0,1 mL de carragenina na região subplantar da pata traseira direita e a migração leucocitária foi induzida pela injeção de 500 µL de carragenina no peritônio. Além disso, determinou-se a citotoxicidade da ATR, utilizando-se a linhagem celular L929, através do teste de MTT e dos testes de toxicidade aguda (5 g/kg - dose única) e subcrônica (50 mg/kg-30 dias) em ratos Wistar. Os resultados mostraram que nas doses de (100 mg/kg e 200 mg/kg) a ATR exibiu atividade antiinflamatória significativa nos ensaios de edema de pata e migração leucocitária. Nos testes de toxicidade aguda, os animais apresentaram hipoatividade e letargia no período inicial (primeiras 6 horas) e aumento das proteínas totais, bilirrubinas total e indireta e fosfatase alcalina depois de 14 dias nos machos tratados. Para o ensaio subcrônico, houve aumento das proteínas totais, gama-glutamil-transferase, fosfatase alcalina e bilirrubina total e direta nas fêmeas tratadas com ATR. Não foram encontradas alterações na arquitetura e morfologia das lâminas histológicas observadas. Esses resultados sugerem que a ATR apresenta atividade antiinflamatória significativa, sem apresentar significativa toxicidade aguda, subcrônica e citotoxicidade.

**Unitermos:** *Cladina kalbii*/farmacognosia. Atranorina/atividade antiinflamatória. Atranorina/toxicidade. Plantas medicinais.

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## INTRODUCTION

Lichens are some of the most important sources of biologically active compounds (Barnes *et al.*, 2000). They have a worldwide distribution and live in a symbiotic relationship with fungi (mycobiont), algae (phycobiont), and/or cyanobacteria (photobiont) (Honegger, 1991). This symbiosis provides physical protection, water, and mineral supply for both partners. They produce common intracellular constituents, including proteins, amino acids, polyols, carotenoids, polysaccharides, and vitamins. The secondary metabolites found in lichens are phenolics that accumulate either in the cortex or in the cell walls of medullary hyphae, and they are mainly acetyl-polymalonyl pathway derivatives (Honda, Vilegas, 1999). Furthermore, lichens are able to synthesize several metabolites, which are composed of aliphatic, cycloaliphatic, aromatic, and terpenic compounds (Huneck, 1999).

Polysaccharides, proteins, and secondary metabolites produced by lichens have attracted the attention of investigators due to their biological activities. For centuries, a number of the 17,000 known lichen species have been used by humans for medicinal purposes. In the northeastern region of Brazil, lichen species vary according to the environment, from the coastal zone (humid) to the inner region, where the climate is semi-arid (caatinga). In these habitats, several species are considered to be bioactive (Maia *et al.*, 2002; Pereira, 1996). Atranorin (ATR) (Figure 1) is the main compound from the lichen *Cladina kalbii* Ahti, which grows in the arid regions of northeastern Brazil. ATR is an important member of the depside group. The molecular structures of these depsides present aromatic esters with a methyl ester terminal group (Howell *et al.*, 2003).

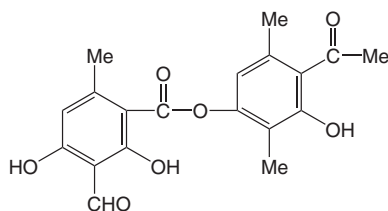


FIGURE 1 - Chemical structure of atranorin.

Studies on the pharmacological properties of *C. kalbii* and its constituents have revealed antinociceptive effects (Pereira, 1996; Maia *et al.*, 2002), antimicrobial action against *Staphylococcus aureus* and *Bacillus subtilis* (Falcão *et al.*, 2004; Ribeiro *et al.*, 2002), and photoprotection capacity (Fernandez *et al.*, 1998) as well as antioxidant activity (Hidalgo *et al.*, 1994). Additionally, a recent

report has demonstrated the antinociceptive property of ATR in mice (Melo, 2008; Siqueira *et al.*, 2010). Despite the widespread medicinal use of *C. kalbii* root extracts in Brazil and elsewhere, there are no published studies on their toxicological and anti-inflammatory properties.

This study aimed to evaluate the anti-inflammatory activity, cytotoxicity, and toxic effects of acute (single dose) and subchronic (30 days) oral administration of ATR extracted from *C. kalbii* in rodents.

## MATERIAL AND METHODS

### Lichen material

*C. kalbii* was collected in March 2007 in Itabaiana, State of Sergipe, Northeastern Brazil (10°44'S, 37°23'W), and identified by M.P. Marcelli (Botanical Institute of São Paulo-SP, Brazil). SP 393235.

### Extraction and isolation of ATR

ATR (C<sub>19</sub>H<sub>18</sub>O<sub>8</sub>) was isolated from the crude extract of the lichen *C. kalbii*. The air-dried parts (100 g) of *C. kalbii* were extracted with 150 mL of chloroform using a Soxhlet apparatus to isolate ATR. The crude extract was filtered and stored at 4 °C for 24 h for ATR precipitation.

### Animals

Male and female Wistar rats (150-200 g) and male Swiss mice (28-32 g) were obtained from Universidade Federal de Sergipe, Brazil. Toxicity tests were performed in male and female rats and anti-inflammatory tests were performed only in male rats or mice. The animals were kept under standard laboratory conditions (20±2 °C) and housed in polypropylene cages, with food and water *ad libitum*. All experimental protocols were approved by the Institutional Animal Care and Use Committee of Universidade Federal de Sergipe (CEPA/UFS 100/2006). All experiments were performed from 1:00 p.m. to 6:00 p.m.

### Anti-inflammatory study

#### *Carrageenan-induced hind paw edema in rats*

Acute hind paw edema was induced in male rats by injecting 0.1 mL of carrageenan (prepared as 1% suspension in sterile normal saline) into the subplantar region of the right hind paw (Winter *et al.*, 1962). ATR (50, 100, and 200 mg/kg, p.o.) was administered to three different groups, while two other groups served as negative and positive controls and received vehicle (0.9% saline with two drops

of 0.2% Tween 80, a solvent for ATR) and standard drug (aspirin, 300 mg/kg, p.o.), respectively. ATR and aspirin were administered 1 h prior to the injection of carrageenan. The rat pedal volume up to the ankle joint was measured using a plethysmometer (Model LE 7500 Panlab, Barcelona, Spain) at 3 h after the injection of carrageenan.

#### *Leukocyte migration to the peritoneal cavity*

Leukocyte migration was induced by injection of 500  $\mu$ L of carrageenan (500  $\mu$ g/cavity, i.p.) into the peritoneal cavity of male mice 1 h after administration of ATR (50, 100, and 200 mg/kg, p.o.), vehicle (0.9% saline with two drops of 0.2% Tween 80), or dexamethasone (2 mg/kg, s.c., n = 6) by a modification of the technique previously described by Bastos *et al.* (2007). Mice were euthanized by cervical dislocation 4 h after carrageenan injection. Shortly after, phosphate-buffered saline (PBS) containing EDTA (1 mM, i.p., 10 mL) was injected. Immediately after, the abdomen was briefly massaged for further fluid collection, which was centrifuged (2000 rpm, 5 min) at room temperature. The supernatant was disposed and 1 mL of PBS was added to the precipitate. A 10- $\mu$ L aliquot of this suspension was diluted in 200  $\mu$ L of Turk solution, and total cells were counted in a Neubauer chamber under optical microscopy. The results were expressed as the number of leukocytes/mL. The percentage of leukocyte inhibition (%LI) was calculated as follows:  $\%LI = (1 - T/C) \times 100$ , where T represents the test groups' leukocyte counts, and C represents the control groups' leukocyte counts.

### **Toxicological studies**

#### *Subchronic toxicity study*

Toxicological assays were performed with 20 male and 20 female rats, divided into 4 groups of 10 animals each (experimental and control groups of male and female rats) (BRASIL, 2004). For the subchronic study, daily doses of ATR (50 mg/kg) dissolved in saline with 0.2% Tween 80 (vehicle) were administered to the experimental groups over a 30-day period, whereas the controls received only saline.

At the end of the experimental period, and after a 12 h fast, the animals were anesthetized with thiopental (50 mg/kg) and blood (3-5 mL) was collected from the abdominal aorta. Subsequently, the animals were sacrificed (hemorrhagic shock) and a detailed study of the gross and microscopic features of internal organs as well as hematological and biochemical analyses of blood were carried out. The position, shape, size, texture, consistency, and color of internal organs (lungs, heart, liver, viscera, and kidneys) were macroscopically examined for any

signs of gross changes. These organs were then collected, weighed to determine relative organ weights, and fixed in 10% phosphate-buffered formalin solution for subsequent histological procedures.

#### *Acute toxicity study*

Toxicological assays were performed with 12 male and 12 female rats, divided into 4 groups of 6 animals each (experimental and control groups of male and female rats) (BRASIL, 2004). The animals received 5 g/kg of ATR by oral administration (gavage). ATR, dissolved in saline with 0.2% Tween 80 (vehicle), was administered to the experimental groups, whereas the controls received only saline.

Specific behaviors (sedation, reduced ambulation, response to touch, analgesia, and defecation) were observed and graded according to Almeida *et al.* (1999) at 1, 2, 3, and 4 h after gavage. Finally, the animals were monitored daily for 14 days to verify lethality. At the end of the experimental period, and after a 12 h fast, the animals were anesthetized with thiopental and blood was collected from the abdominal aorta. Subsequently, the animals were sacrificed (hemorrhagic shock) and a detailed study of the gross and microscopic features of internal organs as well as hematological and biochemical analyses of blood were carried out. The position, shape, size, texture, consistency, and color of internal organs (lungs, heart, liver, viscera, brain, and kidneys) were macroscopically examined for any signs of gross changes. These organs were then collected, weighed to determine relative organ weights, and fixed in 10% phosphate-buffered formalin solution for subsequent histological procedures.

### **Parameters**

#### *Hematological analysis*

Blood samples were collected into EDTA tubes. Measurements of erythrocytes, hemoglobin, hematocrit, leukocyte, neutrophil, lymphocyte, eosinophil, monocyte, basophil, atypical lymphocyte, and platelets were determined using Pentra-120 (ABX-HORIBA) automated equipment according to the method described by Morris and Davey (1999).

#### *Biochemical parameters*

Serum was separated from non-heparinized blood and assayed for serum urea, creatinine, total protein, globulin, albumin, aspartate aminotransferase (AST), alanine aminotransferase (ALT), gamma-glutamyl transferase (GGT), total bilirubin (TBIL), direct bilirubin (DBIL), alkaline phosphatase (ALP), lactate dehydrogenase (LDH), calcium, sodium, potassium, and uric acid. Biochemical

parameters were determined using Dade Behring Dimension RxL automated equipment.

### Tissue analysis

Formalin-fixed samples of internal organs were dehydrated, cleared, and embedded in paraffin according to standard histological procedures. Then, 5  $\mu$ -thick sections of paraffin-embedded tissues were obtained and stained with hematoxylin-eosin. Morphological analysis of the histological sections was performed by light microscopy in a blinded fashion, following a numerical protocol so that the pathologist was blinded to treatment groups until the end of the experiment. Morphometric analysis was used to compare mean kidney glomerular perimeter between female rats (control and test groups). The images were digitized using 5 histological fields for each case (200x magnification), employing an Olympus image capture system. The digital images were then processed using specific morphometry software (Image Tool 3.0).

### Determination of cytotoxicity

L929 cells (fibroblast cells) from the Rio de Janeiro Cell Bank, University of Rio de Janeiro, Rio de Janeiro, Brazil, were routinely grown in 150 cm<sup>2</sup> tissue culture flasks in DMEM supplemented with 1% (v/v) of an antibiotic solution containing 5 mg of penicillin, 5 mg of streptomycin and 10 mg of neomycin per mL, and 7.5% or 10.0% (v/v) heat-inactivated fetal bovine serum at 37°C under 5% CO<sub>2</sub>.

### MTT assay

Cell sensitivity to ATR was determined by standard spectrophotometric 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Cells were seeded at a density of 10<sup>5</sup> cells/well into 96-well plates and incubated for 24 h at 37°C in a 95% air, 5% CO<sub>2</sub> atmosphere. Then, 20  $\mu$ L of ATR at different concentrations in PBS was added to the culture plates for 24 h. After treatment, cells were rinsed once with PBS. Serum-free culture medium, without phenol red, was added back to all wells. Cells were then incubated for 4 h with MTT solution (5 mg/mL). The yellow tetrazolium salt was metabolized by viable cells to form purple formazan crystals. The crystals were solubilized overnight (12 h) in a mixture consisting of 20% sodium dodecyl sulfate (SDS) in HCl (0.01 M). The product was quantified spectrophotometrically by measuring absorbance at 570 nm using a microplate reader ( $\mu$ Quant™, BioTek Instruments Inc., USA). Cell viability was expressed as the percentage of viable cells in relation to the control cells.

### Statistical analysis

The Student *t* test was employed in the statistical analysis of the results (Origin 6.2 or GraphPad Prism 4.01). Data obtained from the anti-inflammatory study were evaluated by one-way analysis of variance (ANOVA), followed by Dunnett's or Fisher's tests. All values were expressed as back-transformed mean  $\pm$  standard deviation (SD). Differences between groups were considered statistically significant at  $p < 0.05$ .

## RESULTS AND DISCUSSION

### Anti-inflammatory study

The mean increase in paw edema volume was about 0.82 $\pm$ 0.17 mL in vehicle-treated rats. ATR (100 and 200 mg/kg, p.o.) significantly reduced ( $p < 0.05$ ) mean paw edema volume (29.3 and 32.9%, respectively), as compared with the control group, at 3 h after carrageenan injection. Similarly, aspirin (300 mg/kg, p.o.) showed highly significant ( $p < 0.001$ ) anti-inflammatory activity, with a 76.4% inhibition rate (Table I).

Several mediators are released by carrageenan in the rat paw. The initial phase of the response to carrageenan may be due to the release of histamine and serotonin, whereas kinins may play a role in the middle phase (Di Rosa, Sorrentino, 1968), and prostaglandins may be the most important mediators in the final 3-5 h of the post-carrageenan response (Vinegar *et al.*, 1969). Bugni *et al.* (2009) showed that part of the antinociceptive effect obtained with ATR may be due to inhibition of cyclooxygenase (COX) enzymes. Additionally, Siqueira *et al.* (2010) suggested that ATR modulates neurogenic and inflammatory pain in capsaicin- and formalin-induced orofacial pain test models through a naloxone-sensitive mechanism.

Figure 2 shows the inhibitory effect of ATR on carrageenan-induced leukocyte migration into the peritoneal cavity 4 h after stimulus at all doses (31.9, 35.9, and 42.5% at doses of 50, 100, and 200 mg/kg, respectively;  $p < 0.05$ ). The results obtained with the control group support the effect of ATR, since the vehicle showed no activity and the control drug (dexamethasone, 2 mg/kg, s.c.) inhibited leukocyte migration (92.2%,  $p < 0.001$ ), as compared with the control group.

Cell recruitment during inflammation depends on the orchestrated release of local mediators responsible for local vascular and tissue changes as well as for the recruitment of host defense cells (Luster *et al.*, 2005). Inflammation induced by carrageenan involves cell migration,



**TABLE I** - Effect of atranorin (ATR) or aspirin on carrageenan-induced hind paw edema in rats (n = 8)

Treatment	Dose (mg/kg)	Carrageenan-induced hind paw edema volume (mL) <sup>a</sup>	% inhibition
Vehicle	-	0.82 ± 0.17	-
ATR	50	0.77 ± 0.15	6.1
ATR	100	0.58 ± 0.11 <sup>b</sup>	29.3 <sup>d</sup>
ATR	200	0.55 ± 0.08 <sup>b</sup>	32.9 <sup>d</sup>
Aspirin	300	0.21 ± 0.05 <sup>c</sup>	76.4 <sup>e</sup>

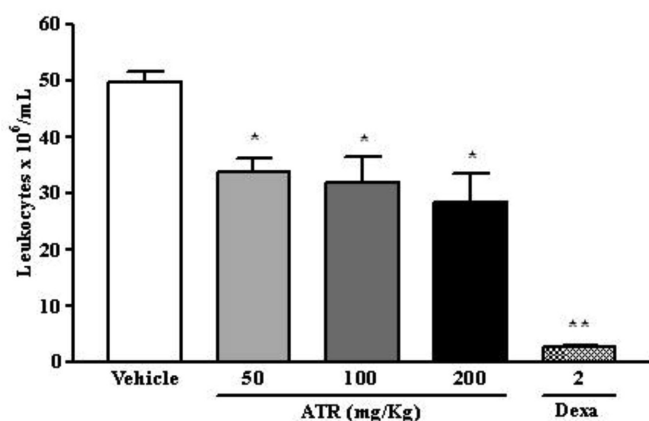
<sup>a</sup> Values expressed as mean ± standard error of the mean (SEM).

<sup>b</sup> P < 0.05 (one-way ANOVA and Dunnett's test), significantly different from control.

<sup>c</sup> P < 0.001 (one-way ANOVA and Dunnett's test), significantly different from control.

<sup>d</sup> P < 0.05 (Fisher's test), significantly different from control.

<sup>e</sup> P < 0.001 (Fisher's test), significantly different from control.



**FIGURE 2** - Effect of atranorin (ATR) on carrageenan-induced leukocyte migration into the peritoneal cavity in mice. Groups of mice were pretreated with vehicle (control group, 10 mL/kg, p.o.), dexamethasone (Dexa, 2 mg/kg, s.c.), or ATR at concentrations of 50, 100, and 200 mg/kg (p.o.) 60 min before carrageenan (500 µg/cavity, 500 µL, i.p.)-induced peritonitis. Cell counts were performed at 4 h after injection of carrageenan. Values are expressed as mean ± standard error of the mean (SEM). \*p < 0.05 and \*\*p < 0.001 when compared to control group. ANOVA followed by Dunnett's test (n = 6, per group).

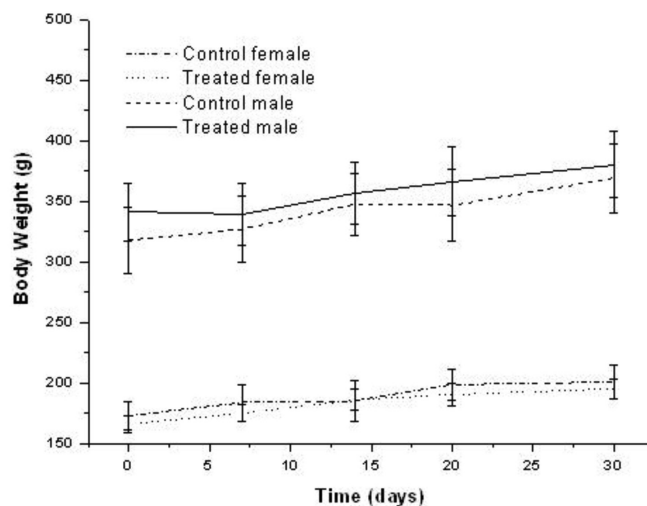
plasma exudation, and production of chemical mediators, such as nitric oxide, prostaglandin E2, interleukin (IL)-1β, IL-6, and tumor necrosis factor (TNF)-α (Salvemini *et al.*, 1996; Loram *et al.*, 2007). Those mediators are able to recruit leukocytes, such as neutrophils, as demonstrated in several experimental models.

Kumar and Muller (1999) demonstrated that ATR effectively inhibits the biosynthesis of leukotriene B4 in bovine polymorphonuclear leukocytes, which could also lead to an anti-inflammatory effect. Because ATR inhibited carrageenan-induced paw edema volume and leukocyte migration, it may be suggested that these activities are

related to inhibition of the synthesis of many inflammatory mediators, whose involvement in cell migration is well established.

### Subchronic toxicity study

Administration of ATR (50 mg/kg, p.o.) for 30 days did not modify the profile of weight gain when test and control groups were compared (Figure 3). In addition, food and water consumption for control and treated animals was kept within the established limits for this animal species.



**FIGURE 3** - Changes in rat body weight during 30-day atranorin (ATR) treatment (50 mg/kg). Each point represents mean ± standard deviation (SD) of n = 10.

There were no changes in the weight of lungs, heart, liver, and stomach (Table II) 30 days after subchronic treatment with 50-mg/kg ATR. The internal organs of both control and treated animals showed no unusual signs.

Moreover, both macro and microscopic analyses revealed normal findings. The organs of both control and treated groups were unremarkable and comparable to each sex. There was no further evidence of histopathological changes. The architecture and cell appearance of the examined internal organs were comparatively unremarkable in both groups and sexes. Only the female rats showed significant differences in the relative weight of the kidneys, as shown in Table II.

Thus, a morphometric analysis was performed in the kidney glomerular cells, which showed no significant differences when compared to test and control groups. Scientific literature on the renal histology and morphometry

of animals treated with lichens is still rare. The differences in kidney weights could suggest a renal reduction in the area of glomerular corpuscle and capsular space (Daniels, Hostetter, 1990). However, gross and microscopic pathologic examination of the kidneys detected no abnormalities. Also, the values observed for kidney weight are consistent with the limits considered normal for this animal species, according to Hossne *et al.* (1986).

Results of the hematological study are presented in Table III. The data show that hemoglobin, hematocrit, leukocyte, neutrophil, lymphocyte, eosinophil, monocyte, basophil, atypical lymphocyte, and platelet levels for the control rats were not significantly different from

**TABLE II** - Changes in organ weight of rats after subchronic oral treatment with atranorin (ATR) at a daily dose of 50 mg/kg for 30 days

Groups	Organ weight				
	Lung (g)	Heart (g)	Liver (g)	Kidneys (g)	Stomach (g)
CF	1.52 ± 0.21	0.81 ± 0.09	7.45 ± 0.85	1.91 ± 0.28	3.85 ± 0.99
TF	1.45 ± 0.13	0.77 ± 0.08	7.43 ± 0.50	1.54 ± 0.09*	3.95 ± 0.70
CM	2.39 ± 0.26	1.45 ± 0.20	9.97 ± 2.03	3.01 ± 0.31	8.84 ± 1.25
TM	2.54 ± 0.31	1.47 ± 0.20	9.98 ± 1.11	2.88 ± 0.22	9.36 ± 1.60

CF = control female; CM = control male; TF = treated female; TM = treated male.

Data are expressed as mean ± standard deviation (SD), n = 10.

\* Indicates statistical difference between control and ATR groups (p < 0.05).

**TABLE III** - Effect of 30-day oral administration of atranorin (ATR) on hematological parameters, considering sex interaction, in Wistar rats (n = 10)

Parameters	Groups					
	CF	TF	UFS Female Vivarium	CM	TM	UFS Male Vivarium
Erythrocytes (×10 <sup>6</sup> /mm <sup>3</sup> )	8.32 ± 0.17	8.14 ± 0.40	7.3 - 8.64	8.28 ± 0.58	7.74 ± 0.42*	6.7 - 8.6
Hemoglobin (g/dl)	14.10 ± 0.33	13.83 ± 0.52	13.2 - 15.1	15.94 ± 0.77	15.33 ± 0.56	12.8 - 15.9
Hematocrit (%)	42.93 ± 0.94	41.76 ± 1.86	39.1 - 48.5	45.55 ± 3.48	42.57 ± 3.15	39.1 - 48.7
Leukocyte (×10 <sup>3</sup> /mm <sup>3</sup> )	6.41 ± 1.00	7.07 ± 0.97	4.7 - 12.98	7.72 ± 1.43	7.25 ± 1.83	5.7 - 13.9
Neutrophil (%)	4.12 ± 1.24	4.11 ± 0.33	5.4 - 37.5	20.40 ± 6.83	18.33 ± 4.21	17.1 - 47.9
Lymphocyte (%)	94 ± 1.77	94.11 ± 0.60	57.9 - 90.0	65.90 ± 7.04	68.55 ± 3.20	48.7 - 78.1
Eosinophil (%)	0	0	0.2 - 4.5	0	0	0.3 - 4.0
Monocyte (%)	1.25 ± 0.46	1	0.6 - 7.9	13.7 ± 2.40	13.11 ± 1.69	1.0 - 6.5
Basophil (%)	0.5 ± 0.53	0.66 ± 0.50	0 - 0.8	0	0	0 - 0.8
Atypical lymphocyte (%)	0.12 ± 0.35	0.11 ± 0.33	0 - 1.0	0	0	0 - 1.0
Platelets (×10 <sup>3</sup> /mm <sup>3</sup> )	9627.5 ± 58.21	9940.00 ± 72.83	757 - 1476	7352 ± 20.13	7352 ± 25.84	837 - 1455

CF = control female; CM = control male; TF = treated female; TM = treated male; UFS = Universidade Federal de Sergipe, Northeastern Brazil.

Data are expressed as mean ± standard deviation (SD), n = 10.

\* Indicates statistical difference between control and ATR groups (p < 0.05).

those of ATR-treated rats during the study period (Table III). Only erythrocyte counts were significantly lower in treated males 30 days after treatment, as compared to the control group ( $p < 0.05$ ). However, these values could be considered normal because, according to Schalm (1986), the number of erythrocytes may range from 5.25 to 8.5 million in male rats. Furthermore, these values are also in agreement with the reference values for rats in the university vivarium (these reference values were determined in a recent survey, but such data are not yet published).

The increase in erythrocytes may be due to overproduction of hematopoietic regulatory elements, such as colony-stimulating factors, erythropoietin, and thrombopoietin, by stromal cells and macrophages in the bone marrow (Chang-Gue *et al.*, 2003), thus providing a local environment for hematopoiesis (Udut *et al.*, 2005). In addition, the fact that such a significant difference was not verified between the female groups supports the idea that this finding is merely contingent.

Plasma biochemical data are described in Table IV. No significant changes were observed in urea, creatinine, albumin, AST, ALT, LDH, calcium, sodium, potassium, and uric acid. Nevertheless, the study showed an increase in total protein and globulin, demonstrating that liver function was preserved (Motta, 2003). Furthermore, the increase in GGT, DBIL, and ALP revealed biliary cholestasis (Henry, 2008), probably due to an incipient inflammatory process.

The histopathological examination of selected organs (lungs, heart, liver, viscera, and kidneys) from treated and control animals showed normal architecture, suggesting that daily oral administration of ATR for 30 days caused no detrimental changes or morphological disturbances.

### Acute toxicity study

In the acute toxicity test, doses of 5 g/kg of ATR

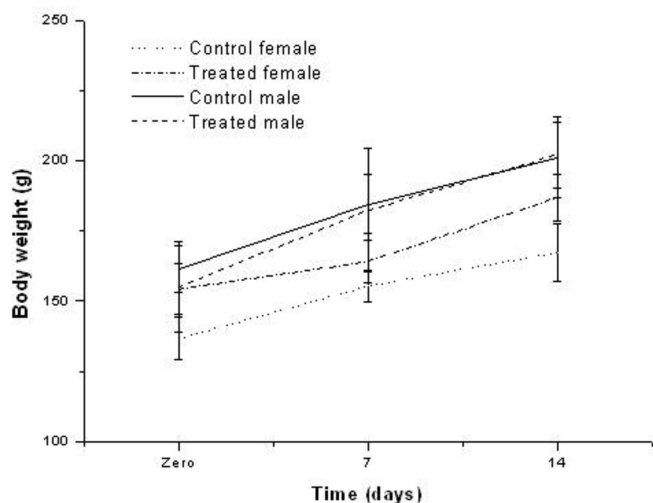
**TABLE IV** - Serum biochemical findings in rats treated with atranorin (ATR) at a daily dose of 50 mg/kg for 30 days

Parameters	Groups					
	CF	TF	UFS Female Vivarium	CM	TM	UFS Male Vivarium
Urea (mg/dL)	58.80 ± 33.40	43.70 ± 5.91	30 - 57	37.30 ± 4.16	33.70 ± 4.08	30 - 42
Creatinine (mg/dL)	0.40 ± 0.22	0.41 ± 0.09	0.4 - 0.7	0.40 ± 0.04	0.33 ± 0.11	0.44 - 0.64
Total protein (g/dL)	6.10 ± 0.40	6.43 ± 0.27*	6.1 - 7.4	5.38 ± 0.32	5.47 ± 0.22	5.4 - 6.6
Globulin (g/dL)	4.71 ± 0.39	5.12 ± 0.21*	3.1 - 4.3	4.36 ± 0.24	4.44 ± 0.16	2.7 - 3.5
Albumin (g/dL)	1.32 ± 0.05	1.31 ± 0.07	2.6 - 3.4	1.02 ± 0.10	1.03 ± 0.09	2.7 - 3.2
AST (U/L)	77.33 ± 11.21	68.33 ± 6.74	83 - 184	122.22 ± 45.46	100.74 ± 18.63	81 - 180
ALT (U/L)	63.75 ± 17.46	74.36 ± 7.07	26 - 60	70.87 ± 23.7	60.88 ± 9.31	36 - 58
GGT (U/L)	8.79 ± 1.39	12.11 ± 1.26*	--	8.91 ± 1.37	8.82 ± 0.91	--
TBil (mg/dL)	0.07 ± 0.02	0.12 ± 0.02*	0.07 - 0.09	0.25 ± 0.05	0.24 ± 0.03	0.07 - 0.08
DBil (mg/dL)	0.02 ± 0.01	0.05 ± 0.01*	0.01 - 0.03	0.17 ± 0.01	0.16 ± 0.01	0.01 - 0.03
IBil (mg/dL)	0.04 ± 0.02	0.06 ± 0.01*	0.01 - 0.01	0.5 ± 0.28	0.39 ± 0.28	0.01 - 0.01
ALP (U/L)	100.8 ± 46.84	156.8 ± 36.52*	63 - 138	89.27 ± 28.0	73.49 ± 7.04	79 - 196
LDH	305.1 ± 89.71	219 ± 116.29	--	1270.5 ± 14.84	979.00 ± 318.4	--
Ca <sup>2+</sup> (mEq/L)	9.83 ± 0.48	10.08 ± 0.48		10.42 ± 0.28	10.53 ± 0.35	--
Na <sup>+</sup> (mEq/L)	142.3 ± 10.78	145.55 ± 1.23	135 - 144	140.23 ± 1.31	140.00 ± 0.83	132 - 146
K <sup>+</sup> (mEq/L)	4.46 ± 1.12	4.31 ± 0.76	4.2 - 6.4	5.79 ± 1.55	5.81 ± 0.58	3.7 - 5.7
Uric acid (mg/dL)	1.44 ± 0.57	1.41 ± 0.80	0.9 - 2.0	2.49 ± 1.26	2.02 ± 0.54	0.9 - 2.0

ALP = alkaline phosphatase; ALT = alanine aminotransferase; AST = aspartate aminotransferase; CF = control female; CM = control male; DBil = direct bilirubin; GGT = gamma-glutamyl transferase; IBil = indirect bilirubin; LDH = lactate dehydrogenase; TBil = total bilirubin; TF = treated female; TM = treated male; UFS = Universidade Federal de Sergipe, Northeastern Brazil. Data are expressed as mean ± standard deviation (SD), n = 10.

\* Indicates statistical difference between control and ATR groups ( $p < 0.05$ ).

caused no deaths in rats during a 14-day observation period. The rats showed no signs of toxicity or changes in general behavior or any other physiological activities. Except for reduced locomotor activity ('hypoactivity') and lethargy, observed within the first 24 hours of ATR administration (at a single dose of 5 g/kg), no other signs of toxicity were noted in rats treated with oral ATR. Figure 4 shows a graphical representation of changes in body weight following the treatment of rats with ATR and saline. There was a significant increase ( $p < 0.05$ ) in body weight over the experimental period for all treatment groups.



**FIGURE 4** - Changes in rat body weight during atranorin (ATR) treatment (5 g/kg-single dose). Each point represents mean  $\pm$  standard deviation (SD) of  $n = 10$ .

As shown in Table V, organ weights (lung, heart, liver, viscera, and brain) of ATR-treated rats did not differ from those of the control group.

Hematological analysis in the acute study (Table VI) showed no significant differences in the parameters examined in control or treated animals of both sexes. At a dose of 5 g/kg (single dose), only the male group showed a significant decrease in lymphocyte (%), as compared with the corresponding control group. In contrast, there was no

significant change in any of the differential lymphocyte counts in female rats. Moreover, the difference noted in lymphocyte count between groups is within the reference values given by Mitruka and Rawnsley (1981), 65-84.5%, suggesting the absence of clinically relevant alterations.

The biochemical profile of treated and control rats is presented in Table VII. Acute oral administration of ATR (5 g/kg-single dose) only altered bilirubin levels, with no significant changes in serum urea, creatinine, globulin, AST, ALT, GGT, DHL, calcium, potassium, and uric acid. Because of a significant increase in TBIL and DBIL alone, it may be suggested that a reduction in conjugation rate occurs within the hepatocyte (Henry, 2008). Altered ALP and sodium levels in the male group were considered normal, since they are in agreement with the reference values for rats in the university vivarium. In addition, no abnormalities or further evidence of histopathological changes were found in any of control or treated rats of both sexes.

Gross pathologic examination of the tissues detected no abnormality in weight or appearance between control and experimental groups. Moreover, the organs of both groups were unremarkable and comparable to each sex. Microscopic analysis revealed that the architecture and cell appearance of the internal organs were comparatively unremarkable in both groups and sexes. These findings indicate that oral administration of purified ATR extracted from *C. kalbii* promotes no morphological changes in the vital organs of animals and suggest that this substance provides no toxicological effects.

### Determination of cytotoxicity

MTT is a water-soluble tetrazolium salt, which is converted to an insoluble purple formazan product by cleavage of the tetrazolium ring by succinate dehydrogenase activity within the mitochondria. The formazan product is impermeable to cell membranes and therefore accumulates in healthy cells (Fotakis, Timbrell, 2006).

ATR was evaluated in a mouse fibroblast cell line (L929) in order to examine its cytotoxic effects on normal

**TABLE V** - Changes in organ weight of rats after acute oral treatment with atranorin (ATR) at a daily dose of 5 g/kg (single dose)

Groups	Lung (g)	Heart (g)	Liver (g)	Kidneys (g)	Viscera (g)	Brain (g)
CF	1.28 $\pm$ 0.02	0.61 $\pm$ 0.03	5.75 $\pm$ 0.33	1.35 $\pm$ 0.02	4.45 $\pm$ 0.07	1.30 $\pm$ 0.12
TF	1.15 $\pm$ 0.12	0.77 $\pm$ 0.11	5.76 $\pm$ 0.21	1.46 $\pm$ 0.16	3.20 $\pm$ 0.94	1.37 $\pm$ 0.11
CM	1.52 $\pm$ 0.31	0.98 $\pm$ 0.07	7.95 $\pm$ 0.51	1.59 $\pm$ 0.01	4.66 $\pm$ 1.23	1.2 $\pm$ 0.43
TM	1.39 $\pm$ 0.02	0.96 $\pm$ 0.13	7.36 $\pm$ 1.63	2.01 $\pm$ 0.27	5.36 $\pm$ 3.85	1.66 $\pm$ 0.08

CF = control female; CM = control male; TF = treated female; TM = treated male.

Data are expressed as mean  $\pm$  standard deviation (SD),  $n = 6$ .



**TABLE VI** - Effect of oral administration of atranorin (ATR) (5 g/kg-single dose) on hematological parameters, considering sex interaction, in Wistar rats (n = 10)

Parameters	Groups					
	CF	TF	UFS Female Vivarium	CM	CT	UFS Male Vivarium
Erythrocytes ( $\times 10^6/\text{mm}^3$ )	7.12 $\pm$ 0.17	7.84 $\pm$ 0.54	7.3 - 8.64	8.28 $\pm$ 0.58	8.80 $\pm$ 0.39	6.7 - 8.6
Hemoglobin (g/dl)	15.3 $\pm$ 0.21	14.7 $\pm$ 0.37	13.2 - 15.1	15.94 $\pm$ 0.77	15.8 $\pm$ 0.61	12.8 - 15.9
Hematocrit (%)	40.65 $\pm$ 0.91	41.96 $\pm$ 1.26	39.1 - 48.5	45.55 $\pm$ 3.48	44.45 $\pm$ 1.37	39.1 - 48.7
Leukocyte ( $\times 10^3/\text{mm}^3$ )	7.10 $\pm$ 0.14	8.13 $\pm$ 0.30	4.7 - 12.98	7.72 $\pm$ 1.438	9.70 $\pm$ 1.43	5.7 - 13.9
Neutrophil (%)	12.5 $\pm$ 0.63	9.00 $\pm$ 1.14	5.4 - 37.5	20.4 $\pm$ 6.83	14 $\pm$ 5.65	17.1 - 47.9
Lymphocyte (%)	85.5 $\pm$ 4.94	88.66 $\pm$ 1.15	57.9 - 90.0	65.9 $\pm$ 7.04	84.02 $\pm$ 8.61*	48.7 - 78.1
Eosinophil (%)	1 $\pm$ 0	1 $\pm$ 0	0.2 - 4.5	1 $\pm$ 0	1 $\pm$ 0	0.3 - 4.0
Monocyte (%)	1 $\pm$ 0	2 $\pm$ 0	0.6 - 7.9	13.7 $\pm$ 2.40	12.75 $\pm$ 3.5	1.0 - 6.5
Basophil (%)	-	-	0 - 0.8	-	-	0 - 0.8
Atypical lymphocyte (%)	-	-	0 - 1.0	-	-	0 - 1.0
Platelets ( $\times 10^3/\text{mm}^3$ )	741.00 $\pm$ 12.72	905.00 $\pm$ 11.52	757 - 1476	735.20 $\pm$ 20.21	845.50 $\pm$ 15.87	837 - 1455

CF = control female; CM = control male; TF = treated female; TM = treated male; UFS = Universidade Federal de Sergipe, Northeastern Brazil.

Data are expressed as mean  $\pm$  standard deviation (SD), n = 6.

\* Indicates statistical difference between control and ATR groups ( $p < 0.05$ ).

**TABLE VII** - Serum biochemical findings in rats treated with atranorin (ATR) at a single dose (5 g/kg)

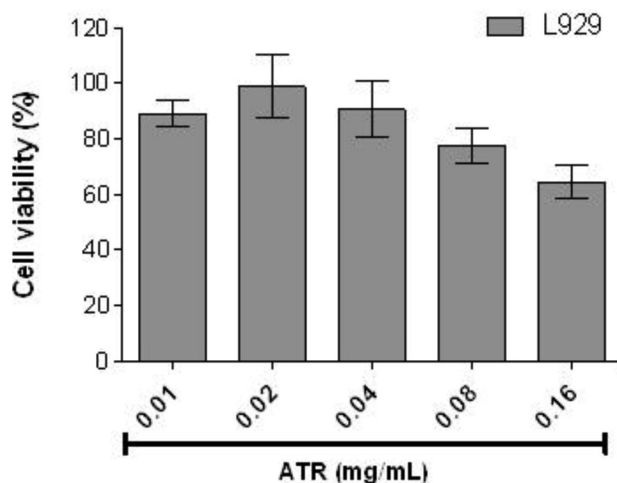
Parameters	Groups					
	FC	FT	UFS Female Vivarium	MC	MT	UFS Male Vivarium
Urea (mg/dL)	50.00 $\pm$ 12.72	35.66 $\pm$ 4.50	30 - 57	37.3 $\pm$ 4.16	37.25 $\pm$ 2.62	30 - 42
Creatinine (mg/dL)	0.51 $\pm$ 0.14	0.5 $\pm$ 0.10	0.4 - 0.7	0.4 $\pm$ 0.04	0.37 $\pm$ 0.05	0.44 - 0.64
Total Protein (g/dL)	6.00 $\pm$ 0.28	4.62 $\pm$ 3.19	6.1 - 7.4	5.38 $\pm$ 0.32	5.8 $\pm$ 0.31*	5.4 - 6.6
Globulin (g/dL)	4.85 $\pm$ 0.21	4.56 $\pm$ 0.46	3.1 - 4.3	4.36 $\pm$ 0.24	4.5 $\pm$ 0.31	2.7 - 3.5
Albumin (g/dL)	1.15 $\pm$ 0,07	1.35 $\pm$ 0.12	2.6 - 3.4	1.02 $\pm$ 0.10	1.3 $\pm$ 0,00*	2.7 - 3.2
AST (U/L)	110.00 $\pm$ 0.56	95.66 $\pm$ 10.21	83 - 184	122.71 $\pm$ 45.46	120.50 $\pm$ 32.46	81 - 180
ALT (U/L)	63.01 $\pm$ 0,03	55.66 $\pm$ 11.93	26 - 60	70.80 $\pm$ 23.73	77.50 $\pm$ 21.62	36 - 58
GGT (U/L)	9.51 $\pm$ 0.70	8.01 $\pm$ 1.73	--	8.90 $\pm$ 1.37	7.25 $\pm$ 1.70	--
TBil (mg/dL)	0.54 $\pm$ 0.03	0.56 $\pm$ 0.02	0.07 - 0.09	0.25 $\pm$ 0.05	0.46 $\pm$ 0.23*	0.07 - 0.08
DBil (mg/dL)	0.11 $\pm$ 0	0.06 $\pm$ 0.04	0.01 - 0.03	0.17 $\pm$ 0.01	0.05 $\pm$ 0.02*	0.01 - 0.03
IBil (mg/dL)	0.43 $\pm$ 0.03	0.5 $\pm$ 0.02	0.01 - 0.01	0.51 $\pm$ 0.28	0.41 $\pm$ 0.20	0.01 - 0.01
ALP (U/L)	91 $\pm$ 9.89	95.66 $\pm$ 21.59	63 - 138	89.2 $\pm$ 28.09	153.25 $\pm$ 15.26*	79 - 196
LDH (U/L)	772.00 $\pm$ 244.65	841.33 $\pm$ 371.81	--	1563.0 $\pm$ 188.0	1170.5 $\pm$ 225.5	--
Ca <sup>2+</sup> (mEq/L)	9.75 $\pm$ 0.21	10.51 $\pm$ 0.41	--	10.49 $\pm$ 0.28	10.52 $\pm$ 0.29	--
Na <sup>+</sup> (mEq/L)	135.50 $\pm$ 0.70	136.66 $\pm$ 0.57	132 - 146	140.2 $\pm$ 1.31	136.5 $\pm$ 1.00*	132 - 146
K <sup>+</sup> (mEq/L)	4,25 $\pm$ 0,21	5,8 $\pm$ 1,53	3.7 - 5.7	5,79 $\pm$ 1,55	5.35 $\pm$ 1.19	3.7 - 5.7
Uric acid (mg/dL)	1.4 $\pm$ 0.28	1.54 $\pm$ 0.68	1.2 - 2.5	2.49 $\pm$ 1.26	1.30 $\pm$ 0.43	0.9 - 2.0

ALP = alkaline phosphatase; ALT = alanine aminotransferase; AST = aspartate aminotransferase; CF = control female; CM = control male; DBil = direct bilirubin; GGT = gamma-glutamyl transferase; IBil = indirect bilirubin; LDH = lactate dehydrogenase; TBil = total bilirubin; TF = treated female; TM = treated male; UFS = Universidade Federal de Sergipe, Northeastern Brazil.

Data are expressed as mean  $\pm$  standard deviation (SD), n = 6.

\* Indicates statistical difference between control and ATR groups ( $p < 0.05$ ).

cells. ATR cytotoxicity on the growth of L929 cells is shown in Figure 5. Cell proliferation was analyzed 24 h after L929 cells had been cultured with ATR at 0.01, 0.02, 0.04, 0.08, and 0.16 mg/mL using MTT assay. ATR at concentrations of 0.01-0.16 mg/mL did not show cytotoxic effects on the L929 cell line. These results suggest that ATR has no cytotoxic effects at these concentrations. Melo *et al.* (2011) reported an effect of ATR on SH-SY5Y cultures, a neuroblastoma-derived catecholaminergic cell line.



**FIGURE 5** - Cytotoxic effect of atranorin (ATR) in L929 cell line. Each value represents the mean  $\pm$  standard deviation (SD). ATR was dissolved in 12.5% dimethylformamide in the middle of the cell. The cytotoxicity of the solvent of these two cell lines and L929 (fibroblast) was evaluated. The concentration of 12.5% in the cells was not toxic.

Different concentrations of ATR alone had no effect on cell viability, as assessed by MTT assay. When cells are treated with  $H_2O_2$ , there is a significant decrease in cell viability to 40% of control levels. Co-incubation with ATR protects SH-SY5Y cells against the cytotoxic effects of  $H_2O_2$ . Nonetheless, all ATR concentrations reversed the effect of  $H_2O_2$  (Melo *et al.*, 2011).

## CONCLUSION

In summary, data from the present study indicated that ATR has anti-inflammatory properties, but further studies are necessary to establish its correct mechanism of action. It is possible to speculate about the inhibition of the synthesis of many inflammatory mediators involved in cell migration. Additionally, there were no relevant signs or significant changes in hematological, biochemical, and histopathological parameters resulting from the long-term administration of ATR. It is therefore concluded that ATR, at the doses used in this study, produced no remarkable

toxic effects on rats and might be an important tool in the management and/or treatment of inflammatory disorders.

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