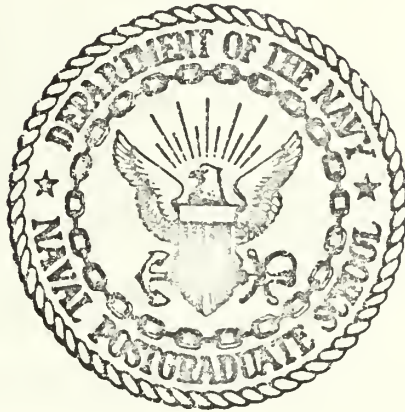


SUBTIDAL CONCRETE PILING FAUNA IN
MONTEREY HARBOR, CALIFORNIA

Winfield Donat

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THESIS

SUBTIDAL CONCRETE PILING FAUNA
IN
MONTEREY HARBOR, CALIFORNIA

by

Winfield Donat III

September, 1975

Thesis Advisor: Eugene C. Haderlie

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area increments. Wet biomass measurements were taken, the organisms were identified and an evaluation of species abundance in each sample was made. Data are given by a Table of Species with abundance in each sample, a Species List with comments on particular organisms, drawings representing the more prominent animals observed and in situ photographs of various piling animals.

Subtidal Concrete Piling Fauna
in
Monterey Harbor, California

by

Winfield Donat III
Lieutenant, United States Navy
A.B., University of North Carolina, 1967

Submitted in partial fulfillment of the
requirements for the degree of

MASTER OF SCIENCE IN OCEANOGRAPHY

from the
NAVAL POSTGRADUATE SCHOOL
September 1975

ABSTRACT

Piling organisms were scraped off one side of a concrete piling from the bottom to the low intertidal zone beneath Municipal Wharf No. 2 in Monterey Harbor, California. Sampling was performed at 0.5 m² surface area increments. Wet biomass measurements were taken, the organisms were identified and an evaluation of species abundance in each sample was made. Data are given by a Table of Species with abundance in each sample, a Species List with comments on particular organisms, drawings representing the more prominent animals observed and in situ photographs of various piling animals.

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DEDICATION

This thesis is dedicated to Winfield Donat, Jr., whose love and respect for coastal organisms provided me a legacy for which I shall be ever grateful.

I. INTRODUCTION

Biological fouling on surfaces exposed to the marine environment is a subject of much concern to the marine engineer. With increased emphasis on concrete construction an ideal substratum is thus provided for organism attachment often to the detriment of a structure's efficiency. While concrete pilings are relatively unaffected in their usefulness by attached biological growth, they do provide an excellent surface from which to collect foulers in the intertidal and subtidal zones. To date, no publications have been found which address these fouling communities to any degree of completeness on the west coast of the United States. Several short term studies have been published using various substrate materials in fouling racks in west coast harbors which involved removal of these materials from the water for observation.

The purpose of this study was to collect and identify organisms from a selected concrete piling--designated Piling "A"--under Municipal Wharf No. 2 in Monterey Harbor, California. Collections were made over the piling's length from the lowest part of the intertidal zone to the bottom 7.1 m below zero tide level. A separate study of the intertidal zone by Professor Eugene C. Haderlie, Oceanography Department, Naval Postgraduate School, Monterey, California, is presently being conducted on this and other nearby pilings. The piling was put in place during wharf construction in

1926 (Haderlie, 1968) and no unusual disturbance of it is known to have occurred which would have seriously affected the fouling community since that time.

Tabulation of organisms from the wharf piling community in Monterey Harbor will provide an index for future studies concerned with the effects of pollution or any expansion of harbor facilities which might be contemplated.

II. ENVIRONMENTAL CONSIDERATIONS

Monterey Municipal Wharf No. 2 is located in the southern end of Monterey Bay (Figure 1). It is supported by concrete pilings each approximately 2 m in circumference with an octagonal cross-section. Piling "A" is the easternmost concrete piling in a transverse row of 12 (Figure 2). This is the 25th such row from the distal end of the wharf and is approximately 300 m from the shore. It is 2 m inward from the edge of the wharf. A creosoted wooden fender piling is to the east of Piling "A" at the wharf's edge.

Water motion around the pilings is primarily from tidal fluctuation and wave action. The lowest tide for the year is approximately 0.5 m below zero tide level from standard tide tables. Piling fauna was collected to -0.6 m because intertidal studies previously collected down to this level. Wave heights were observed during the period of this study to be less than 1 meter.

Figure 3 shows the monthly average morning surface temperatures from 1 July, 1974, to 30 June, 1975, at the

study site. Two years' biweekly averages of daily morning surface salinity measurements in Figures 4 and 5 were the only data available for the wharf area.

Exposure of the transverse row of pilings to direct sunlight is limited primarily to Piling "A" and this is during early morning hours only.

Some scouring by the bottom sediment of the lowest half meter of the pilings is believed to exist. Mean grain sizes vary between 2ϕ and 3ϕ (diameters between 0.25 mm and 0.125 mm) in the area around Piling "A" (Dorman, 1968).

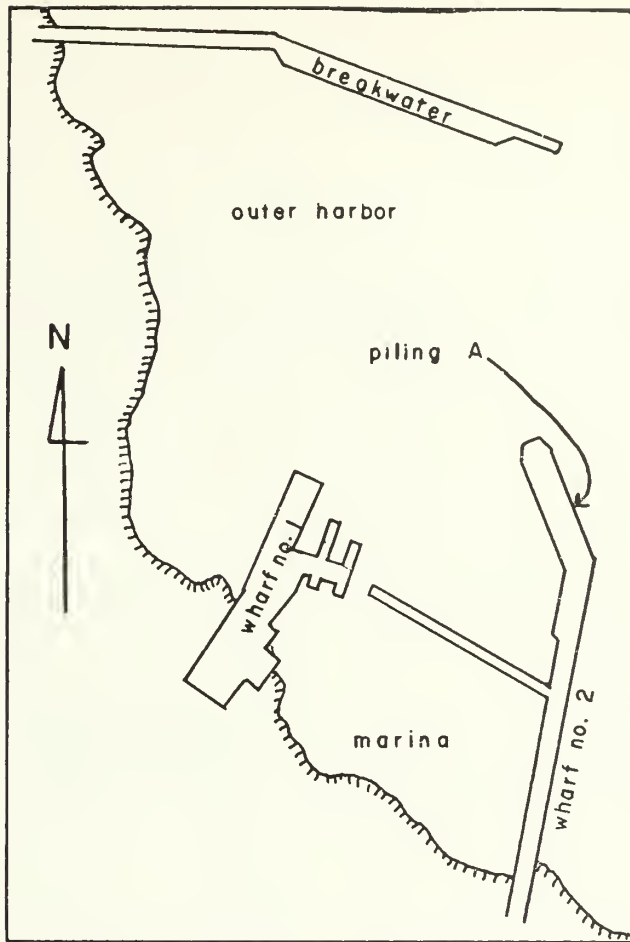


Figure 1. Monterey Harbor showing position of study area.

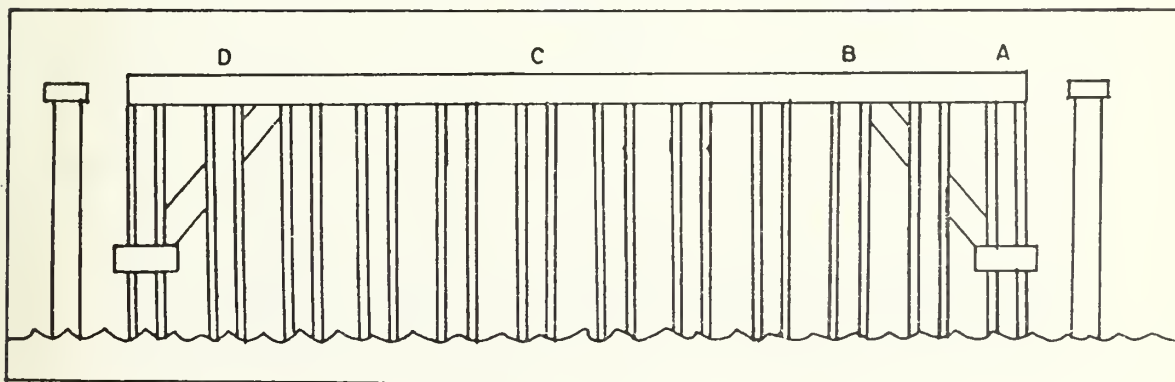


Figure 2. Plan of transverse row of pilings beneath Wharf No. 2 showing the relative positions of concrete Pilings "A", "B", "C" and "D".

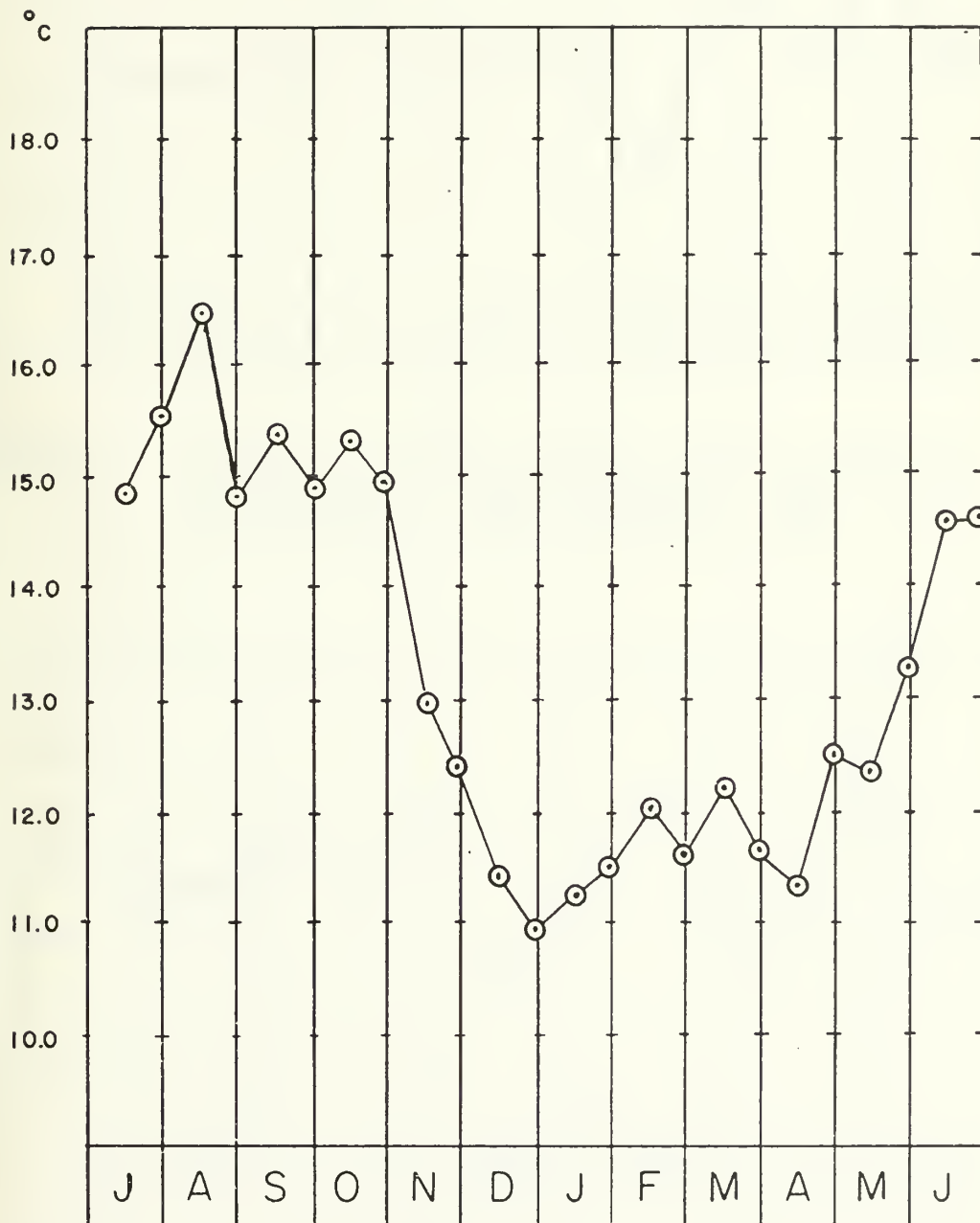


Figure 3. Biweekly morning surface temperature averages at Piling "A" from 1 July, 1974, to 30 June, 1975.

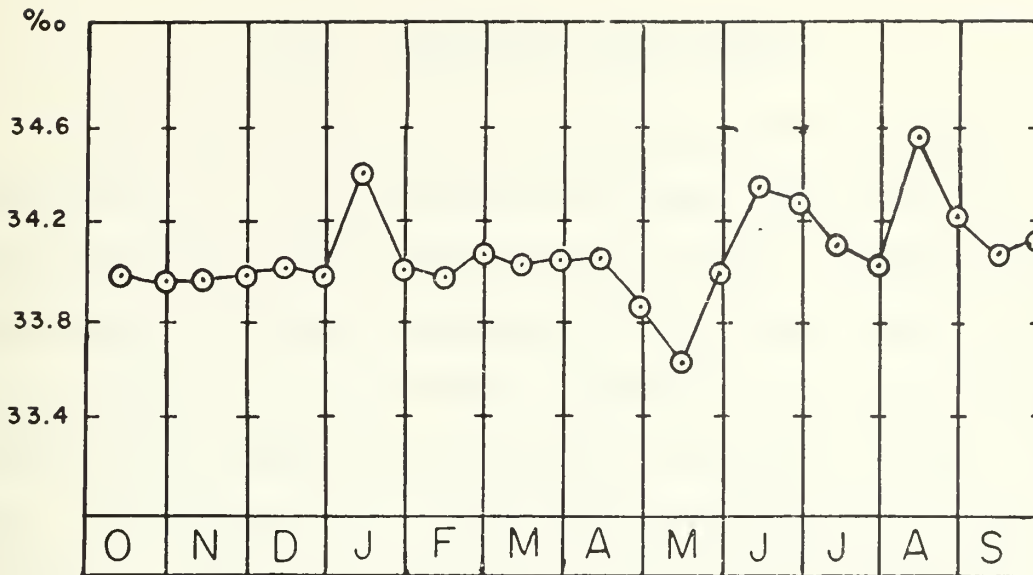


Figure 4. Biweekly morning surface salinity averages at Piling "A" from 1 October, 1966, to 30 September, 1967 (Haderlie, 1968).

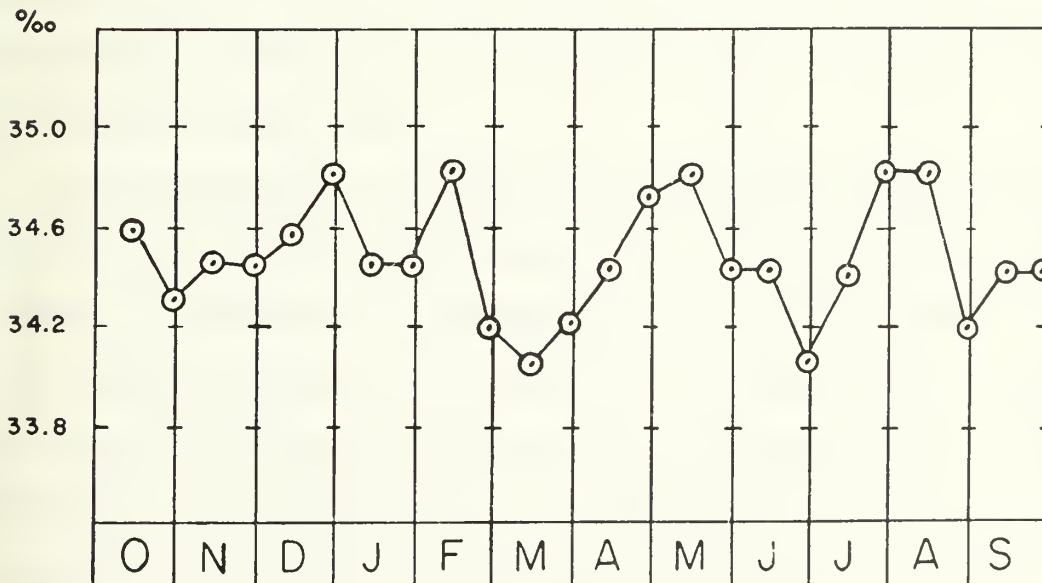


Figure 5. Biweekly morning surface salinity averages at Piling "A" from 1 October, 1967, to 30 September, 1968 (Haderlie, 1969).

III. EQUIPMENT AND METHODS

All dives were made with standard SCUBA equipment including a 3/8 inch neoprene wet suit. The dates and missions of the dives are listed in Table I. Eight dives were made to become acquainted with the collection problem, test collection methods and equipment, review the pilings to observe the organisms in situ and photograph Pilings "A", "B", "C" and "D" for a color slide collection of these pilings. The next 13 dives over the period 1 September, 1974, to 12 May, 1975, involved collection of biomass material from the southern half of Piling "A." The last dive was conducted to photograph with black and white film various segments of the pilings for this thesis and to note any difference between other pilings and what was known of Piling "A" from previous dives.

Photography was conducted using a Nikonos II underwater camera with a Nikkor wide-angle lens (1:3.5, f-28 mm) and a Subsea Products Mk 150 underwater flash attachment. A 0.61 m wooden dowel was secured to the base of the flash attachment for distance measurement to eliminate focusing problems. An area of about 0.25 m² was captured on film using this technique. Greater areas could not be photographed because of the significant extinction of the flash at greater distances from the subject material in the usually turbid water.

Table I

Dates of Dives and Missions

Date	Mission
29 March, 1974	Indoctrination dive.
5 April, 1974	Depth line measurement, piling review.
19 April, 1974	Prepare/test collecting stages, piling review.
4 May, 1974	Photography, piling review.
24 May, 1974	Photography, piling review.
31 May, 1974	Photography, piling review.
11 Agusut, 1974	Photography, sample collection.
16 August, 1974	Sample collection, test collection bag.
1 September, 1974	Material collection (-7.1 to -6.5 m).
13 September, 1974	Material collection (-6.5 to -6.0 m).
4 October, 1974	Material collection (-6.0 to -5.5 m).
21 December, 1974	Material collection (-5.5 to -5.0 m).
26 December, 1974	Material collection (-5.0 to -4.5 m).
13 January, 1975	Material collection (-4.5 to -4.0 m).
17 January, 1975	Material collection (-4.0 to -3.5 m).
19 February, 1975	Material collection (-3.5 to -3.0 m).
24 March, 1975	Material collection (-3.0 to -2.5 m).
1 April, 1975	Material collection (-2.5 to -2.0 m).
7 April, 1975	Material collection (-2.0 to -1.5 m).
14 April, 1975	Material collection (-1.5 to -1.0 m).
12 May, 1975	Material collection (-1.0 to -0.6 m).
17 July, 1975	Photography, observations.

Kodak High Speed Ektachrome (ASA-160) was used for the color slides with aperture settings between f-11 and f-12, shutter speed at 1/60 second and flash output at 150 watt-seconds. Kodak Tri-X (ASA 400) was used for the black and white pictures. Camera settings were held at 1/60 second and f-22 while the flash was set at 50, 100 and 150 watt-seconds. All film was processed by the N.P.S. Educational Media Department.

Collection of material beyond reach when standing on the bottom necessitated the use of two stages on which to stand or kneel (Figure 6). When the upper support line was secured above water to the piling or adjacent platform structure and the girdle was passed around the piling about one meter below the level of material to be collected, adequate stability was provided for working and yet allowed freedom of movement.

A collection bag was improvised from an old plankton net (Figure 7) $1\frac{1}{2}$ m long with a $\frac{1}{2}$ m diameter opening. A line around the piling kept the lower lip of the bag against the piling face and the upper lip was held at a slight angle to the piling. A slow sweeping motion of the hand down into the bag opening carried the falling material that was scraped loose into the bag with negligible loss. Upon completion, the bag was removed from the piling, tied just below the opening and taken to the surface.

Scraping was conducted at half meter intervals up the piling and over half the circumference (1 m) such that the samples were taken over 0.5 m^2 piling surface area. A

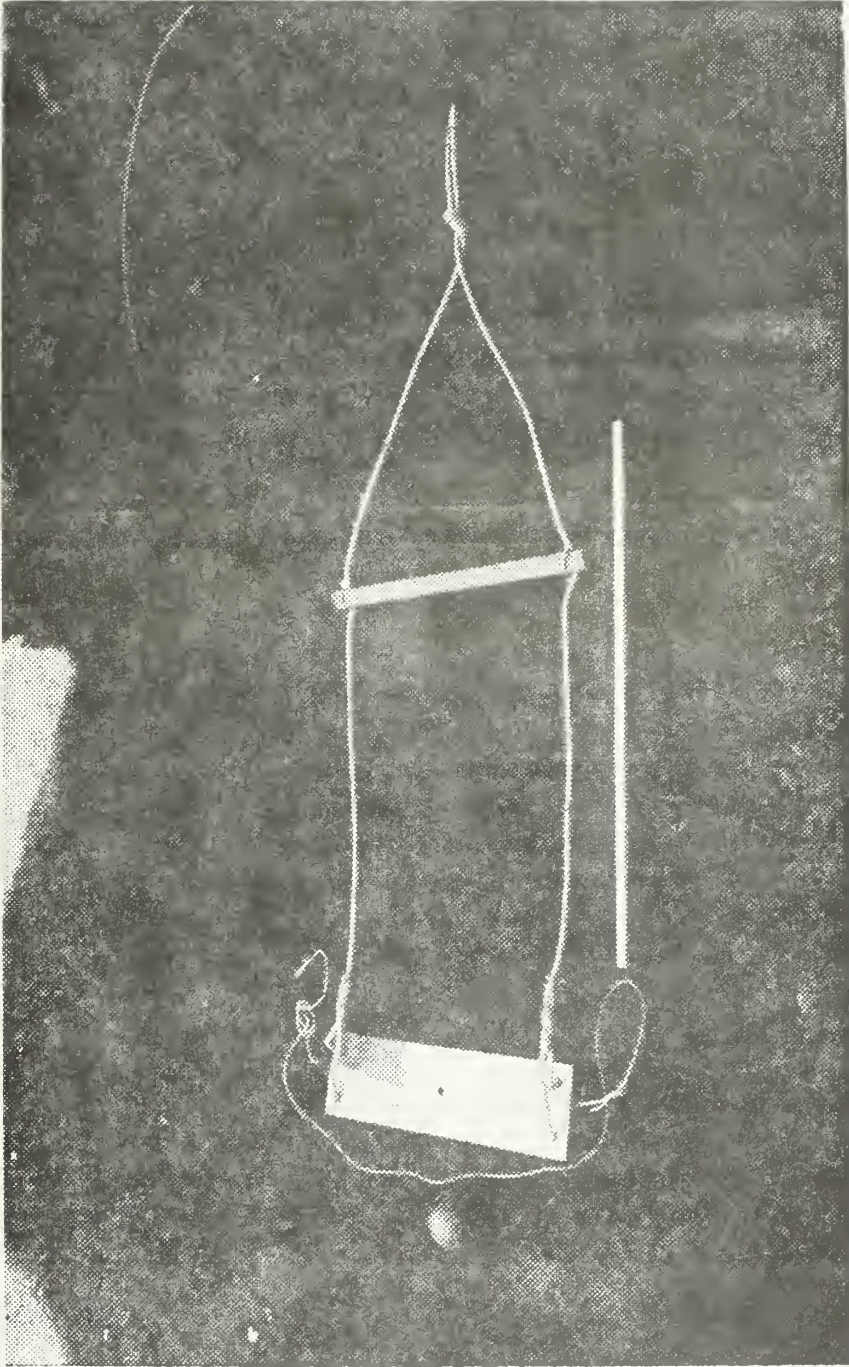


Figure 6. Underwater stage for collection of piling organisms.

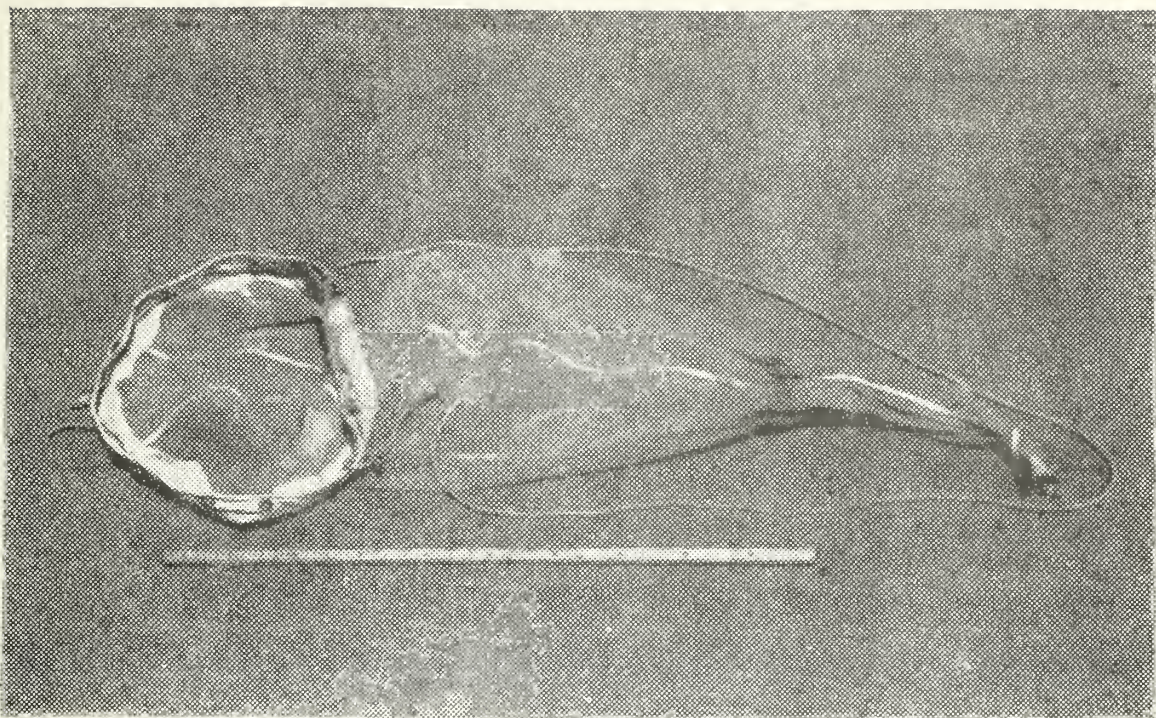


Figure 7. Collection bag made from an old plankton net.

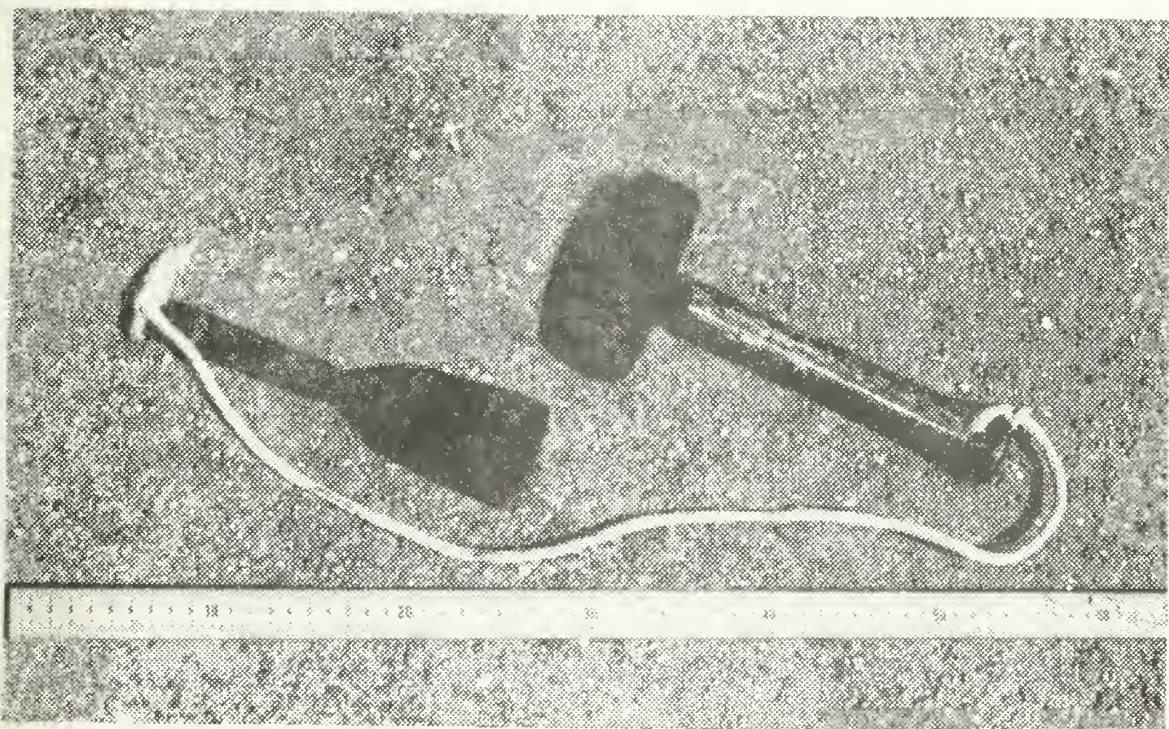


Figure 8. Chisel and sledge hammer used for scraping organisms from the piling.

homemade steel chisel 18 cm in length with a 8 cm blade (Figure 8) was used. A small 3½ pound sledge hammer was required to remove the large barnacles, rock oysters and the larger algal holdfasts.

While equipment was stowed after the dive, the bag was kept at the water's surface and not removed until just before leaving the wharf area. It was then taken directly to the lab and placed on a flat concrete surface outside in the shade for five minutes. This was the only effort made for free-water removal from the sample prior to mass measurement. Table II shows the masses for each half-meter increment. As much material as possible was placed in aquaria with running seawater. The remaining material was immersed in fresh seawater in porcelain pans. The water was drained and replaced twice a day until all material in each pan was observed and identified.

Initial separation of the material involved removal of as many of the plumose anemones Metridium senile as possible in that their acontia and probably discharged nematocysts fouled and killed many small organisms. Large epifauna such as the sea stars and large crabs were removed and classified, and, when present, ascidians (Aplidium solidum) were separated. The remaining organisms were identified during every available time thereafter. It was essential to review the material at once as the animals tended to die rapidly. Small clumps of material were separated under a dissecting scope and the animals contained therein were removed and

Table II

Wet Biomass Measurements per 0.5 m²

Depth (m)	Biomass (Kg)
-0.6 to -1.0	11.58
-1.0 to -1.5	6.81
-1.5 to -2.0	4.99
-2.0 to -2.5	4.77
-2.5 to -3.0	6.95
-3.0 to -3.5	8.63
-3.5 to -4.0	7.58
-4.0 to -4.5	3.75
-4.5 to -5.0	10.78
-5.0 to -5.5	5.83
-5.5 to -6.0	5.44
-6.0 to -6.5	3.55
-6.5 to -7.1	2.92

identified. As each new organism was observed, it was placed in a vial of 75% ethyl alcohol with a label.

The most useful invertebrate keys were The Intertidal Invertebrates of the Central California Coast (Light, 1954) and its revised edition, Light's Manual: Intertidal Invertebrates of the Central California Coast (Smith and Carlton, 1975). The Annelids were also keyed out in the two books by Hartman (1968, 1969) which give extensive descriptions of all known Polychaetes of California. For the Arthropods the barnacles collected were confirmed with the article by Henry (1942) while the review of the California marine Decapod Crustacea by Schmitt (1921) was referred to for the Eucarida. The publications by Keen (1971), Keen and Coan (1974) and McLean (1969) were used in conjunction with the Light's Manual for the Gastropods, Polyplacophorans and Bivalves while the MacFarland memoirs (1966) assisted in identifying the Oposthobranchs. The works of Osburn (1950, 1952, 1953) were used almost exclusively for Bryozoan classification and the Ophiuroids were confirmed in the May (1924) article. Finally, the algae were identified from Smith (1964) and with the assistance of Dr. I. A. Abbott of Hopkins Marine Station.

IV. PRESENTATION OF FINDINGS

The report of findings is presented in three parts: Appendix A.--Table of Species with their abundance at each depth interval of collection; Appendix B.--Species List with comments on some particular species and Appendix C.--Drawings over one meter intervals representing the more prevalent animals collected. Also, in situ photographs of piling organisms with comments on how fauna of the other pilings differed from Piling "A" are contained in this section.

The Table of Species is presented by phyla. Numbers in parentheses by the phyla represent the number of species identified. The column "Lg. Dim." tabulates the approximate largest dimensions of the organisms that were found. Measurements are given in mm. No lengths are given for Cliona celata (Porifera), Bowerbankia gracilis (Bryozoa), Tubulipora tuba (Bryozoa) and Phoronis vancouverensis (Phoronida) in that their colonial forms were not definitive in their extent. Sizes for other Bryozoans and Aplidium solidum (Chordata) represent the largest dimension of colonies found.

Numbers of each species found in the 0.5 m² collection samples are primarily given by quantitative symbols identified in Table III. Where the letter "P" was used the species were present in numbers that were difficult to count. The algae are also listed in this manner. The tabulation of

abundance probably represents too low a figure of those organisms actually present especially with the smaller animals but they depict the best estimate of the observer. The asterisks in the final column refer to comments made about the organisms in the Species List.

Piling "A" subtidal included 235 species of animals and seven species of algae. As shown by the histogram in Figure 9 there is a definite trend of increasing numbers of species present at shallower depths. The largest number occurred between -0.6 m and -1.5 m in association with the tubed Annelid Phyllochaetopterus prolifica the colonies of which form thick collars on pilings between the low intertidal and -1.5 m. On some pilings this collar extends as far as 0.4 m out from the concrete surface.

There is relatively little change in the numbers of species between -2.5 m and -6.0 m. It is in this intermediate range where extensive colonies of Phoronis vancouverensis cover a large area of the piling surface. The dense, intertwined tubes of this filter feeder allow very little circulation of water down into their colonies. When scraped from the piling, clouds of black, sulfur-reduced organic material underneath the colonies were released into the water. The small Annelid Caulleriella alata was particularly abundant among the Phoronis vancouverensis tubes.

Over the deepest half-meter of Piling "A" exposed areas of the concrete surface were noted. This is believed to be due primarily to the scouring of the piling by the fine grain bottom sands which are moved by tidal currents. The

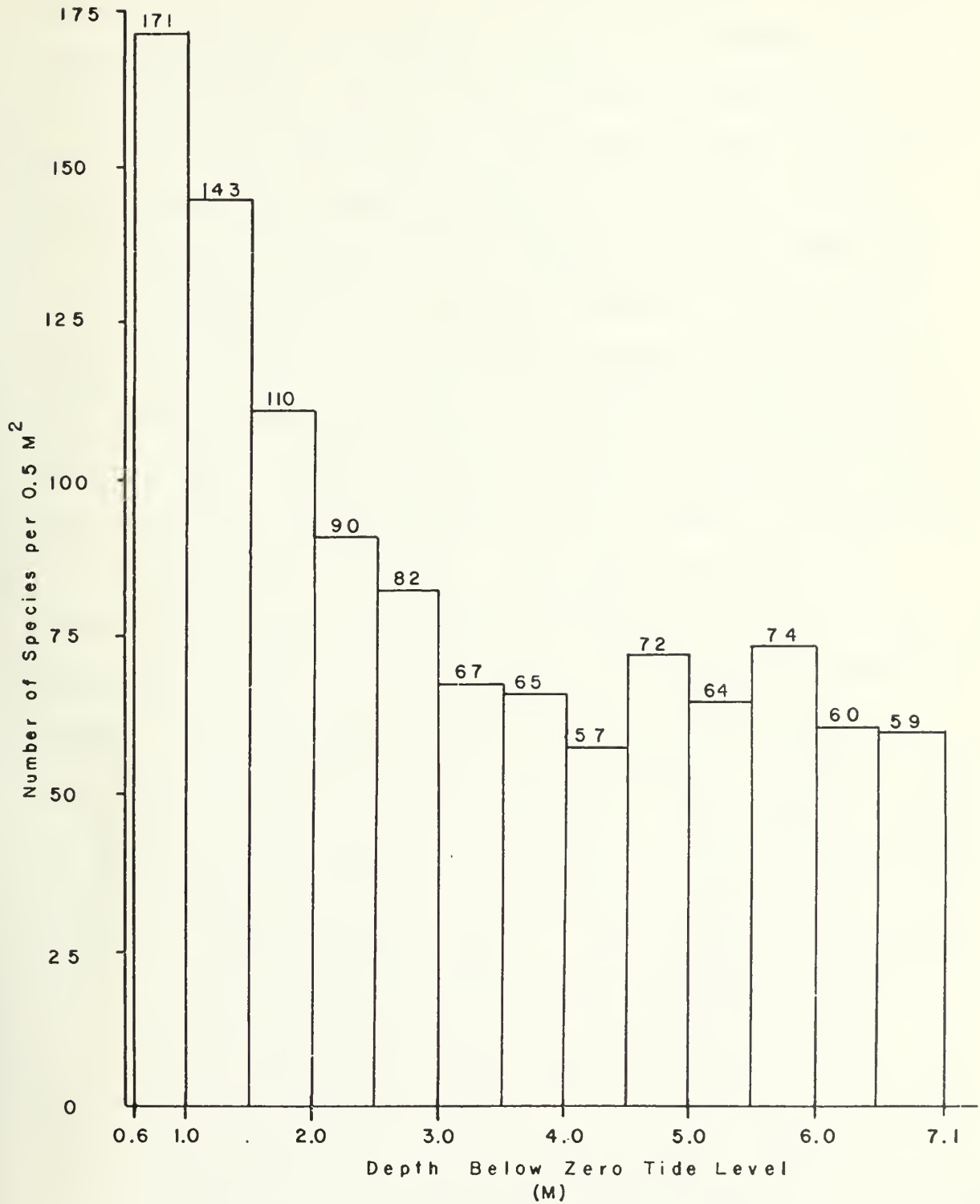


Figure 9. Histogram of numbers of species found per 0.5 m² area scraped at each half-meter collection level.

minute, calcareous tubes of Spirorbis spp. and the barnacle Balanus crenatus were numerous in this interval. The anemone Metridium senile, so prevalent at all other depths, were relatively sparse. At the base of the pilings the sea stars could possibly contribute to the reduced numbers of species in that they were often found on this and other pilings.

The Species List was also arranged by phyla with additional categories (i.e., classes, families, orders, etc.) given for clarity. The categories within each phylum and the species names follow the invertebrate key of Smith and Carlton (1975) since it was the latest publication available and since most species found on the pilings were identified or confirmed using this key. For those organisms not specifically identified using the Smith and Carlton key a notation is made in brackets after the organisms' specific names. The bracketed letters refer to the authors of publications used for their classification. An explanation of this notation is given in Table IV. The comments made on certain species refer to a description of their abundance or habitat, characteristics used in classification, frequency of occurrence, size and difficulties in classification.

Plates I through VI were drawn to give a pictorial representation of those organisms which were found from the bottom of Piling "A" to -0.1 m tidal level. Choices of the animals drawn on each plate were made primarily in accordance with their predominance (abundance and size). Actual drawings of species were made whenever possible; however, in some cases generalized forms were used since several

species have very similar morphological features. A key to the animals used in the plates follows the plates. Sizes of the animals were generally drawn larger with respect to the piling face area than in real life. Also, relative sizes of individual animals are not necessarily accurate.

On the last dive following collection of organisms from Piling "A" black-and-white photographs were taken and a general comparison was attempted between this piling and the others of the same transverse row, primarily Pilings "B", "C" and "D." An immediate observation pointing out dissimilar characteristics of the fouling fauna was that the low intertidal/subtidal biotic collar on Piling "A" was composed primarily of Phyllochaetopterus prolifica tube masses. Most other piling collars observed, however, were composed chiefly of stacked shells of living and dead Balanus nubilus. Figure 10 is an example of the latter.

Another obvious difference was that there were very few Metridium senile seen on any piling in the row studied other than Piling "A". Figure 11 illustrates these plumose anemones fully extended at -1.0 m on Piling "A". When they were present on the other pilings they occurred singularly and were much larger, some attaining a cross-sectional diameter of up to 10 cm.

The presence of Corynactis californica almost reciprocated that of Metridium senile in that none were found on Piling "A" but they were extremely plentiful on the other pilings. While plentiful, their distribution was patchy;



Figure 10. Biotic collar on a piling adjacent to Piling "B" extended from the low intertidal zone to shallow subtidal depths. Most of the mass is composed of Balanus nubilus shells.

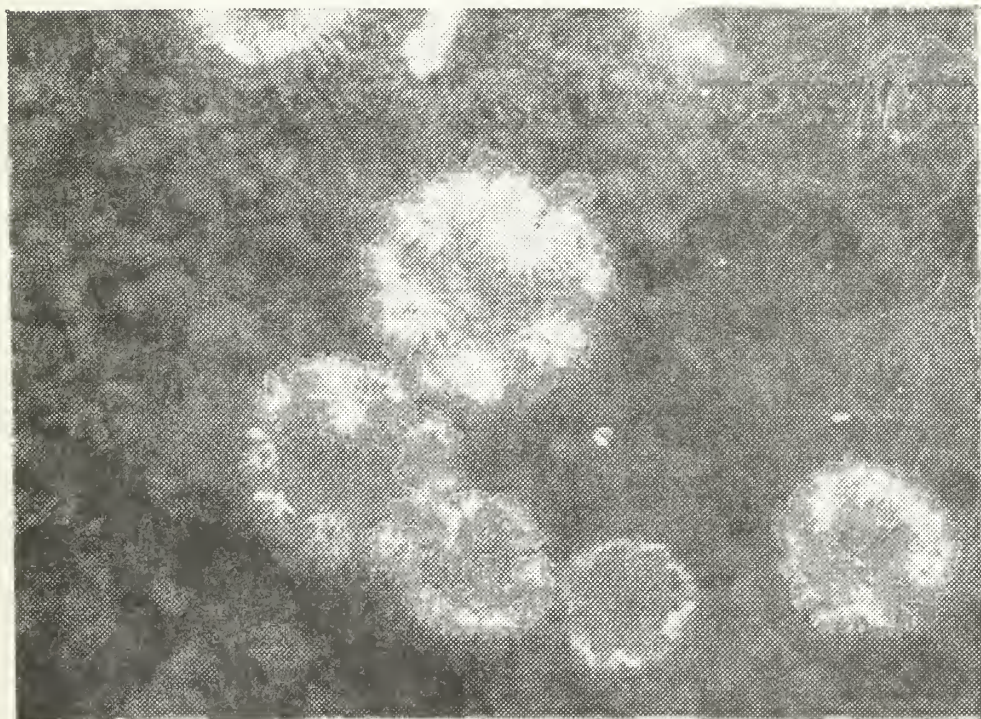


Figure 11. The plumose anemone Metridium senile attached to the thick Phyllochaetopterus prolifica collar at -1.0 m on Piling "A".

e.g., large portions of the pilings would be found void of them. This is probably due to their clonal nature (Smith and Carlton, 1975. p.93.) Various shades of red, purple and orange delineated different clones that lived in close proximity. It also appeared that their abundance was inversely proportional to depth where maximum numbers were reached in the low intertidal zone.

The larger green anemone Anthopleura xanthogrammica was observed infrequently at shallow depths (Figure 12) although most that were seen under the wharf were located well within the intertidal zone as shown just above the thick Balanus nubilus collar in Figure 10.

Phyllochaetopterus prolifica was much more prevalent at all depths on Pilings "B", "C" and "D" than recorded from Piling "A." This was particularly true on Piling "D" where large tube masses were noted from the intertidal zone to the bottom. In association with this, expansive coverage of Phoronis vancouverensis was not observed as found on Piling "A."

As reported for Piling "A" at the shallow subtidal depths these masses of Phyllochaetopterus prolifica harbored a great variety of organisms. While in situ observations did not allow a close scrutiny of the deeper tube masses on Pilings "B", "C" and "D" several animals absent from Piling "A" were readily observed. The Feather Duster Worm, Eudistylia spp., was commonly seen among the Phyllochaetopterus prolifica tubes. One was removed and identified as

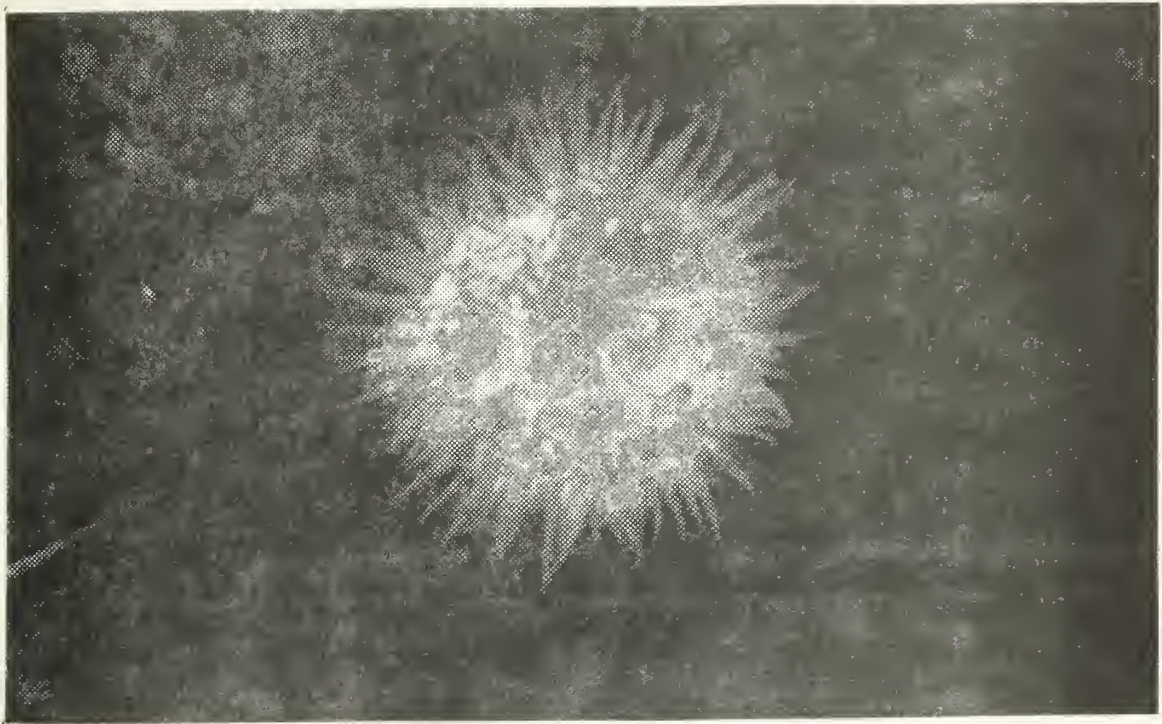


Figure 12. The large green anemone Anthopleura xanthogrammica at -1.5 m on Piling "B".

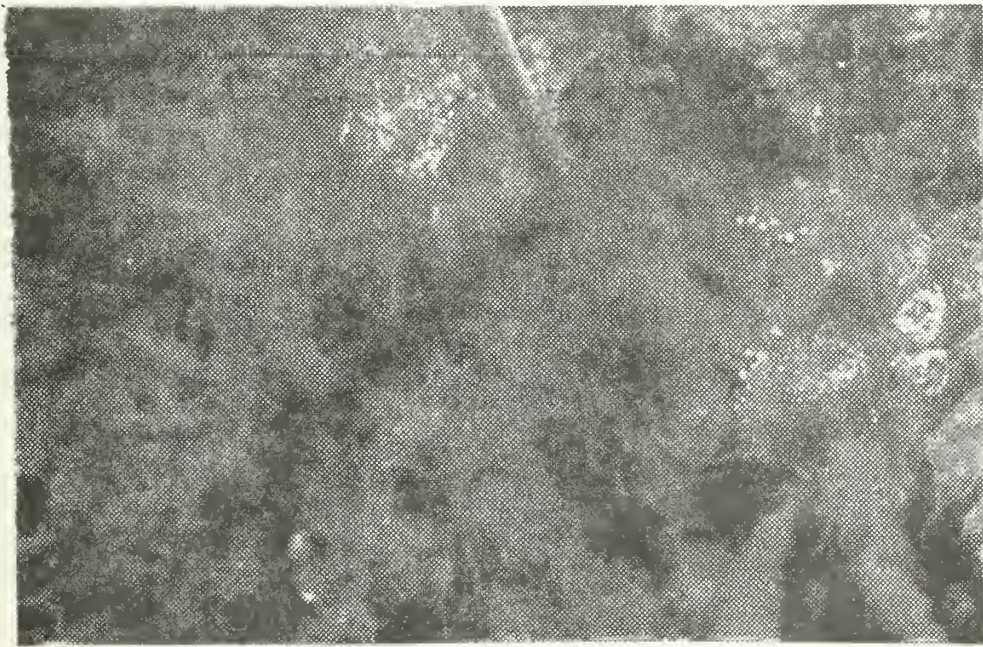


Figure 13. Extended tentacles of Eudistylia vancouveri can be seen just below the pointer tip. The tube is buried in dense Phyllochaetopterus prolifica tubes on Piling "D" at -4.0 m.

Eudistylia vancouveri (Kinberg, 1867); however, it is believed that E. polymorpha (Johnson, 1901) also exists on these pilings (see Blake's comments on E. vancouveri in the Light's Manual, 1975. p. 238). Figure 13 illustrates E. vancouveri with tentacles extended from its tube.

Two Holothuroid species were frequently encountered among these tube masses on Pilings "C" and "D." Only one small Eupentacta quinquesemita was found on Piling "A" but several larger specimens (to 10 cm) were taken from these two pilings. Figure 14 shows one specimen which was extracted from the Phyllochaetopterus prolifica tube masses and placed on these tubes for photographing. The larger sea cucumber Cucumaria miniata (Brandt, 1835) was also seen often nestling in close association with Eupentacta quinquesemita. While the bodies of both these species were completely concealed by the Phyllochaetopterus prolifica tubes their presence was indicated by their prominent respiratory trees extending beyond the tube masses. Stichopus californicus (Stimpson, 1857) has been considered a "characteristic" sea cucumber species on wharf pilings on the west coast (Ricketts and Calvin, 1968. p.369) but only one specimen has been observed (on Piling "C" at -3 m, 24 May, 1974) since dives first began.

There appeared to be little difference in the Balanus nubilus populations on these three pilings as compared with Piling "A." Both live and dead forms are found on all the pilings subtidally. Figure 15 displays a live B. nubilus



Figure 14. The small white sea cucumber Eupentacta quinquesemita at -3.0 m on Piling "C."

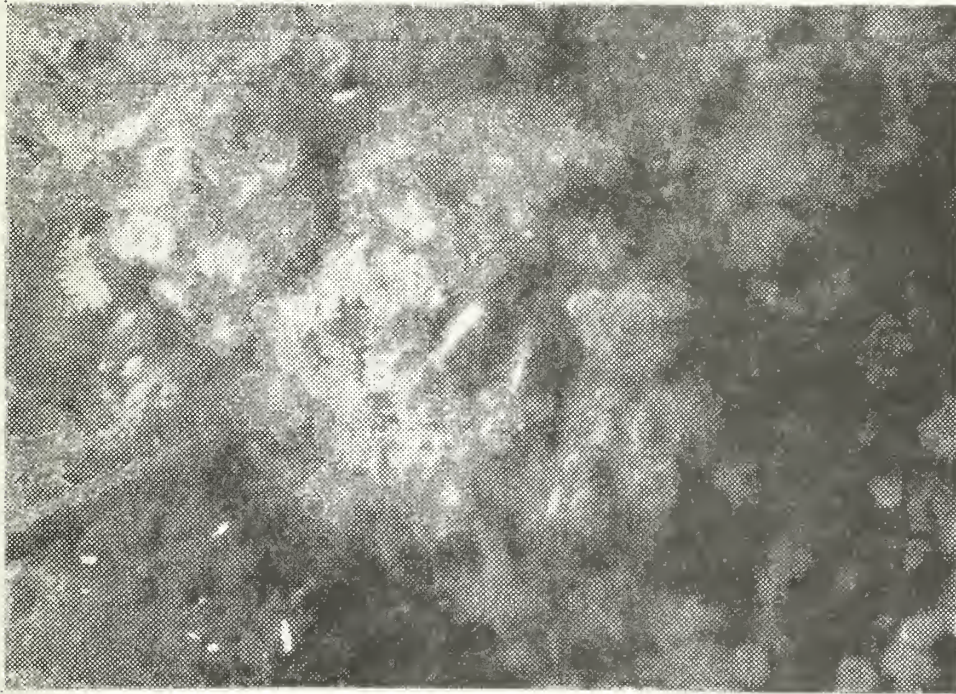


Figure 15. A live Balanus nubilus is located to the right of the pointer and is covered by Corynactis californica. The large horns of the terga can be seen protruding from the opening. The picture was taken at -5.0 m on Piling "D."

covered by Corynactis californica while Figure 16 shows a large shell again covered by C. californica and partially eroded away due to the action of Cliona celata.

The tunicates were not as well represented on Pilings "B", "C" and "D" by Aplidium solidum as on Piling "A" either in numbers of colonies or in colony size; however, Ascidia ceratodes (Figure 17) and Styela montereyensis were more abundant.

The concentration of the nudibranchs Aegires albo-punctatus and Hermisenda crassicornis appeared to be greater than that of Piling "A." Figure 18 shows two specimens of each of these two species together on Phyllochaetopterus prolifica tubes at -4.0 m on Piling "C."

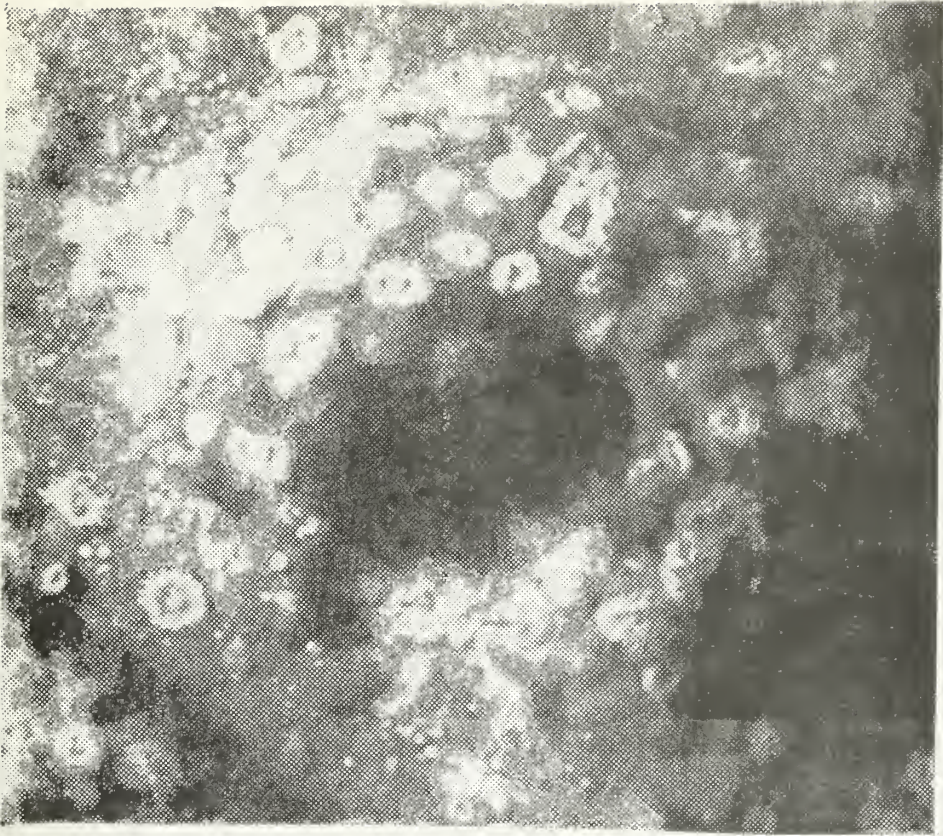


Figure 16. A large Balanus nubilus shell on Piling "B" at -5.0 m covered by Corynactis californica and partially eroded away by Cliona celata.

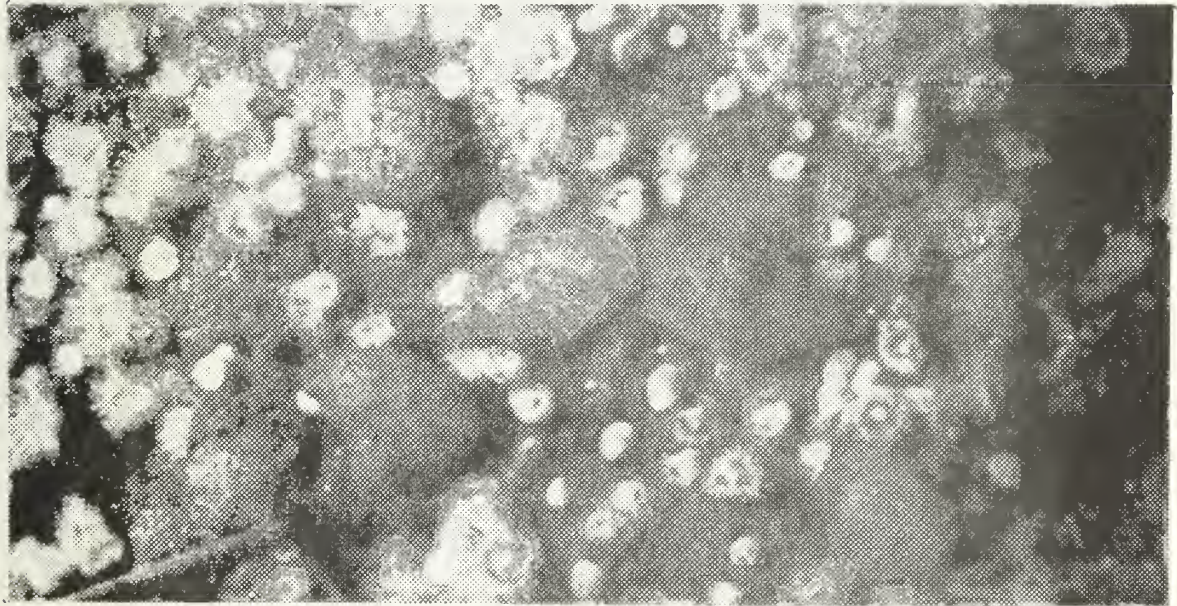


Figure 17. Seven Ascidia ceratodes attached to the concrete surface of Piling "B" at -2.5 m.



Figure 18. Two pairs of nudibranchs on Phyllochaetopterus prolifica tubes on Piling "C" at -4.0 m. Aegires albopunctatus are the white forms in the center. The light colored antennae of Hermisenda crassicornis can be seen to the upper right.

V. CONCLUSION AND COMMENTS

A concrete piling beneath Monterey Municipal Wharf No. 2 was chosen from which to collect subtidal fauna for classification. Material collected over a surface area of 0.5 m² constituted a single sample and thirteen such samples were taken sequentially beginning at the bottom of the piling and extending to the lowest limit of the intertidal zone. A total of 235 animal species was recorded (plus seven algal species) as well as an evaluation of their abundance in each sample.

The piling selected for study was dominated by Metridium senile which are common only on the easternmost concrete pilings. They along with dense colonies of Phoronis vancouverensis at intermediate depths and Phyllochaetopterus prolifica at shallow depths appear to establish unique environmental conditions on Piling "A" which were not realized on the more western pilings observed. The characteristics of increased numbers of species and increased numbers of individual organisms when approaching the intertidal zone were dramatically realized, particularly at depths less than -2.0 m.

Continued research in the study of these wharf pilings is needed to more adequately tabulate the various species present. While the eastern piling community is described

herein there appears to be sufficient difference between it and those communities to the west under the wharf to warrant a continuation of this type of work. Specific and more detailed analyses of a smaller piling surface area would produce more accuracy in the population numbers as well as allow concentration on environmental limiting factors present. Narrowing the scope still further, a study of the interrelationships of only a few organisms such as Balanus nubilus, Cliona celata and Dodecaceria fewkesi--is important for an adequate description of the dynamics of the piling community.

APPENDIX A Table of Species listed in accordance with depth below zero tide level.

Animals (235)	L.g. (mm)												-6.5 to -7.0m	See Species List	
	-0.6 to -1.0m	-1.0 to -1.5m	-1.5 to -2.0m	-2.0 to -2.5m	-2.5 to -3.0m	-3.0 to -3.5m	-3.5 to -4.0m	-4.0 to -4.5m	-4.5 to -5.0m	-5.0 to -5.5m	-5.5 to -6.0m	-6.0 to -6.5m			
<u>Protozoa</u> (2)															
<i>Gromia oviformis</i>		A	A	A	A	A	A	A	A	A	A	A	A	A	
<i>Haplophragmoides columbiensis</i>		A	A	A	A	A	A	A	A	A	A	A	A	A	
var. <i>evolutum</i>															
<u>Porifera</u> (5)															
<i>Cliona celata</i>		R(1)	R(1)	R(3)	R(2)	R(2)	R(2)	0(9)	R(5)	R(1)	0(11)	R(4)	R(2)	R(4)	R(4)
<i>Stelletta clarella</i>	20	X	X	X	X	X	X	X	X	R(1)	X	X	X	X	X
<i>Leucosolenia eleanor</i>	100	R(1)	R(5)	R(2)	0(6)	R(1)	R(2)	R(2)	X	R(1)	X	X	X	X	X
<i>Leucandra heathi</i>	60	R(1)	X	R(2)	R(5)	R(2)	R(1)	R(1)	X	X	X	X	X	X	X
<i>Leucilla nuttingi</i>	50	A	A	F	F	O	F	F	F	F	0	0	R(4)	0	R
<u>Coelenterata</u> (2)															
<i>Obelia</i> spp.	10	P	P	P	P	P	P	P	P	P	P	P	P	P	P
<i>Metridium senile</i>	100	A	A	A	A	A(665)	A	A	A	A	A	A	A(294)	A	F
<u>Platyhelminthes</u> (6)															
<i>Hoplana californica</i>	10	R(2)	R(1)	X	X	X	X	X	X	X	X	X	X	X	X
<i>Notoplana acticola</i>	20	0	R(4)	X	X	X	X	X	X	X	X	X	X	X	X
<i>Notoplana iniqueta</i>	25	A	F	0	R	X	X	X	X	X	X	X	X	X	X
<i>Stylochus atenculatus</i>	80	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X
<i>Stylochus tripartitus</i>	15	0	0	R(2)	X	X	X	X	X	X	X	X	X	X	X
<i>Eurylepta aurantiaca</i>	20	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	X
<u>Nemertea</u> (13)															
<i>Carinoma mutabilis</i>	30	F	R	X	X	X	X	X	X	X	X	X	X	X	X
<i>Tubulanus pellucidus</i>	15	0	0	X	X	X	X	X	X	X	X	X	X	X	X
<i>Tubulanus sexlineatus</i>	120	0	0	0	R(2)	R(2)	R(1)	R(1)	R(2)	R	R(1)	R	R	R(1)	R
<i>Baseodiscus punnettii</i>	80	R(1)	R(1)	X	X	X	X	X	X	X	X	X	X	X	X
<i>Cerebratulus californiensis</i>	100	A	F	R(1)	X	R(2)	X	X	X	X	X	X	X	X	X
<i>Lineus pictifrons</i>	50	R(4)	R(2)	X	X	X	X	X	X	X	X	X	X	X	X
<i>Lineus vegetus</i>	40	R	R	X	X	X	X	X	X	X	X	X	X	X	R(1)
<i>Micrura pardalis</i>	20	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X
<i>Micrura verrilli</i>	70	0	0	R(2)	X	X	R(1)	X	X	X	X	X	X	X	X
<i>Amphiporus bimaculatus</i>	60	A	F	0	0	0	X	X	X	X	X	X	X	X	X
<i>Amphiporus imparispinosus</i>	15	0	0	R	X	R(2)	X	X	X	X	X	X	0	X	R(3)
<i>Paranemertes peregrina</i>	30	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X
<i>Tetrastemma nigrifrons</i>	40	0	0	R(1)	0	X	X	X	X	X	X	X	R(1)	X	X
<u>Sipuncula</u> (1)															
<i>Phascolosoma agassizii</i>	120	A	A	0	0	0	F	X	X	0	0	F	0	0	0
<u>Annelida</u> (85)															
<i>Eunoe barbata</i>	20	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	X
<i>Halosydna brevisetosa</i>	100	A	A	A	A	A	F	F	F	F	A	A	F	A	0
<i>Lepidonotus squamatus</i>	5	R	X	X	X	X	X	X	X	X	X	X	X	X	X
<i>Thormora johnstoni</i>	15	R(1)	X	X	X	X	R	X	X	X	X	X	X	X	X

Animals (235) cont.	Lg. Dim. (mm)	-1.0 to -1.0m	-1.5 to -1.5m	-2.0 to -2.0m	-2.5 to -2.5m	-3.0 to -3.0m	-3.5 to -4.0m	-4.0 to -4.5m	-4.5 to -5.0m	-5.0 to -5.5m	-5.5 to -6.0m	-6.0 to -6.5m	-6.5 to -7.0m	See Species List
<u>Annelida (85) continued</u>														
<i>Peisidice aspera</i>	15	A	F	A	F	F	O	F	O	F	O	O	O	*
<i>Palaemonetes bellus</i>	20	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Anaitides groenlandica</i>	50	X	R	X	X	X	X	X	X	X	X	X	X	
<i>Anaitides madeirensis</i>	35	X	R	X	X	X	X	X	X	X	X	X	X	
<i>Anaitides mucosa</i>	15	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Anaitides williamsi</i>	25	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Eteone californica</i>	10	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Eulalia aviculisetata</i>	30	A	A	F	R	A	F	A	O	O	O	O	O	*
<i>Eulalia bilineata</i>	40	O	F	R	X	X	X	X	X	X	X	X	X	
<i>Eulalia viridis</i>	60	R(1)	X	X	X	X	X	X	X	X	X	X	X	*
<i>Eumida bifoliata</i>	20	A	A	O	R	R	R	R	X	X	X	X	X	*
<i>Genetyllis castanea</i>	15	F	O	X	X	X	X	X	X	X	X	X	X	*
<i>Notophyllum imbricatum</i>	100	X	R(1)	X	X	X	X	X	X	X	X	X	X	*
<i>Sige californiensis</i>	15	X	R	X	X	X	X	X	X	X	X	X	X	*
<i>Amphiduros pacificus</i>	20	F	O	R	R	R	R	R	X	X	X	X	X	*
<i>Ophiodromus pugettensis</i>	30	A	A	F	R	O	X	X	F	X	X	X	X	*
<i>Amblosyllis</i> sp.	15	O	X	X	X	X	X	X	X	X	X	X	X	*
<i>Autolytus varius</i>	30	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Eusyllis assimilis</i>	15	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Exogone</i> sp.	5	X	R(1)	X	X	X	X	X	X	X	X	X	X	*
<i>Haplosyllis spongicola</i>	40	F	A	X	X	R	X	X	X	X	X	X	X	*
<i>Odontosyllis phosphorea</i>	30	A	O	R	X	X	X	X	X	X	X	X	X	*
<i>Odontosyllis</i> sp.	20	A	F	O	X	X	X	X	X	X	X	X	X	*
<i>Pionosyllis gigantea</i>	15	X	R	X	X	X	X	X	X	X	X	X	X	*
<i>Syllis elongata</i>	35	A	O	R	O	X	X	X	X	X	X	X	X	*
<i>Syllis gracilis</i>	30	A	A	O	O	F	O	F	O	O	O	O	O	*
<i>Typosyllis aciculata</i>	10	F	R	O	X	X	X	X	X	X	X	X	X	*
<i>Typosyllis bella</i>	20	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Typosyllis fasciata</i>	20	O	O	R	R	O	X	X	X	X	X	X	X	*
<i>Typosyllis hyalina</i>	20	A	A	O	O	F	X	X	X	X	X	X	X	*
<i>Typosyllis pulchra</i>	20	R	O	X	X	X	X	X	X	X	X	X	X	*
<i>Neanthes caudata</i>	30	O	X	X	X	X	X	X	X	X	X	X	X	*
<i>Nereis eakini</i>	100	A	A	O	R	R	R	R	O	X	X	X	X	*
<i>Nereis latescens</i>	40	X	O	R	R	R	X	X	X	X	X	X	X	*
<i>Nereis natans</i>	15	R	X	X	X	X	X	X	X	X	X	X	X	*
<i>Nereis pelagica neongripes</i>	30	R(1)	X	X	X	X	X	X	X	X	X	X	X	*
<i>Nereis vexillosa</i>	20	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Nereis zonata</i>	60	O	R	X	X	X	X	X	X	X	X	X	X	*
<i>Platynereis bicanaliculata</i>	70	A	F	X	X	X	X	X	X	X	X	X	X	*
<i>Pseudonereis</i> sp.	20	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Palola paloloides</i>	>200	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Dorvillea monilloceras</i>	60	A	F	O	X	X	X	X	X	X	X	X	X	*
<i>Dorvillea rudolphi</i>	25	O	R	R	X	X	X	X	X	X	X	X	X	*
<i>Lumbrineris erecta</i>	200	A	F	F	O	F	A	F	A	F	O	O	O	*
<i>Lumbrineris tetraura</i>	100	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Lumbrineris zonata</i>	30	O	R	X	X	X	X	X	X	X	X	X	X	*
<i>Arabella iricolor</i>	300	A	F	X	X	X	X	X	X	X	X	X	X	*
<i>Drilonereis nuda</i>	150	X	X	X	X	X	X	X	X	X	X	X	X	*

Animals (235) cont.

Lg. Dim. (mm)	to																See Species List
	-1.0m	-1.5m	-2.0m	-2.5m	-3.0m	-3.5m	-4.0m	-4.5m	-5.0m	-5.5m	-6.0m	-6.5m	-7.0m				
Annelida (85) continued																	
100	R	X	O	X	X	X	X	X	X	X	X	X	X	X	X	X	*
20	X	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
100	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	*
15	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
20	O	R	X	R	R	R	R	R	R	R	R	R	R	R	R	R	*
20	O	R	X	R	R	R	R	R	R	R	R	R	R	R	R	R	*
40	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
12	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
8	R(2)	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	R(1)	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
30	R	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
50	F	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
60	O	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
25	A	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
20	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
30	O	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
20	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
15	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
30	A	A	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
40	X	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
25	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	X	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	*
12	R	O	R	R	R	R	R	R	R	R	R	R	R	R	R	R	*
8	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
15	X	O	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
100	O	F	O	F	O	F	O	F	O	F	O	F	O	F	O	F	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
50	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
140	R(2)	R(1)	O(8)	R(4)	O(5)	O(10)	O(10)	R(1)	F(13)	R(5)	O(6)	O(6)	R(4)	R(1)	A	*	
20	R(4)	X	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	X	*
20	R(1)	O	R(2)	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	*
10	A	A	O	F	X	X	X	X	X	X	X	X	X	X	X	X	*
7	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	O	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
10	X	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
5	A	A	A	X	R	X	X	X	X	X	X	X	X	X	X	X	*
Arthropoda (32)																	
50	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
140	R(2)	R(1)	O(8)	R(4)	O(5)	O(10)	O(10)	R(1)	F(13)	R(5)	O(6)	O(6)	R(4)	R(1)	A	*	
20	R(4)	X	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	X	*
20	R(1)	O	R(2)	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	*
10	A	A	O	F	X	X	X	X	X	X	X	X	X	X	X	X	*
7	R	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
10	O	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
10	X	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	*
45	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	*
5	A	A	A	X	R	X	X	X	X	X	X	X	X	X	X	X	*

Animals (235) cont.	Lg. Dim. (mm)	-0.6 to -1.0	-1.0 to -1.5	-1.5 to -2.0	-2.0 to -2.5	-2.5 to -3.0	-3.0 to -3.5	-3.5 to -4.0	-4.0 to -4.5	-4.5 to -5.0	-5.0 to -5.5	-5.5 to -6.0	-6.0 to -6.5	-6.5 to -7.0	See Species List
Arthropoda (32) continued															
<i>Tritella laevis</i>	5	A	A	A	A	A	A	A	A	A	A	F	F		
<i>Caprella verrucosa</i>	10	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	
<i>Heptacarpus taludicola</i>	30	F	O(8)	X	X	X	X	X	X	X	X	X	X	X	
<i>Heptacarpus taylori</i>	20	R(2)	R(1)	X	X	X	X	X	X	X	X	R(2)	X	X	
<i>Spirontocaris prionta</i>	30	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Alpheus dentipes</i>	20	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Bateus harfordi</i>	50	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Loxorhynchus crispatus</i>	150	O(6)	X	R(1)	X	X	X	R(1)	X	X	X	X	X	R(1)	*
<i>Mimulus foliatus</i>	20	R(3)	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Pugettia producta</i>	40	X	R(5)	X	X	X	X	X	X	X	X	X	X	X	
<i>Cancer antennarius</i>	70	A	F	R(5)	X	X	R(1)	X	R(3)	R(1)	X	X	X	X	
<i>Cancer jordani</i>	30	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Cancer sp.</i>	20	F	X	X	F	R(2)	X	X	X	X	X	X	X	X	
<i>Lophopanopeus bellus</i>	25	F	F	O(9)	X	X	R(1)	X	R(3)	R(2)	X	X	X	X	
<i>Pinnixa longipes</i>	12	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Pachycheles pubescens</i>	20	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Pachycheles rudis</i>	20	X	X	X	X	X	X	X	R(4)	X	X	X	X	X	
<i>Petrolisthes cinctipes</i>	15	X	X	R(1)	X	X	X	X	R(1)	X	X	X	X	X	
<i>Phoxichilidium femoratum</i>	10	R(3)	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Pycnogonum stearnsi</i>	10	A	A	R(3)	R(2)	R(1)	X	R(1)	R(1)	X	X	R(1)	R(2)	R(2)	
Mollusca (61)															
<i>Callistoichiton crassicostratus</i>	30	X	X	X	X	X	X	X	R(1)	X	X	X	X	X	*
<i>Lepidozona californiensis</i>	20	A	F	X	R(1)	X	X	X	R(2)	X	X	X	X	X	
<i>Lepidozona mertensii</i>	20	X	X	R(4)	X	X	X	X	X	X	R(1)	X	X	X	
<i>Mopalia ciliata</i>	40	X	X	X	X	X	X	X	X	X	X	X	X	R(1)	
<i>Diodora aspera</i>	15	X	X	X	X	X	X	X	R(2)	X	X	X	X	X	
<i>Megatebennis bimaculatus</i>	12	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Acmaea mitra</i>	8	X	R(1)	X	X	X	X	X	R(1)	X	X	X	X	X	
<i>Lacuna unifasciata</i>	5	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Alvinia compacta</i>	2	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Barleeia acuta</i>	3	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Barleeia californica</i>	2	R(4)	X	X	X	X	R(1)	X	X	X	X	X	X	X	*
<i>Truncatella californica</i>	4	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Caecum californicum</i>	2	A	A	R(1)	X	X	X	X	X	X	X	X	X	X	*
<i>Caecum dalli</i>	2	O	R(4)	X	X	X	X	X	X	X	X	X	X	X	*
<i>Fartulum occidentale</i>	2	A	A	X	X	X	X	X	X	X	X	X	X	X	
<i>Bittium attenuatum</i>	6	R(1)	R(2)	X	X	X	X	X	X	X	X	X	X	X	
<i>Crepidatella lingulata</i>	12	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	
<i>Polinices sp.</i>	1	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	*
<i>Amphissa versicolor</i>	8	A	A	O	X	X	X	R(1)	R(1)	X	X	X	X	X	
<i>Mitrella aurantiaca</i>	7	A	A	O	X	X	X	R(3)	R(1)	X	X	R(1)	R(1)	X	
<i>Mitrella carinata</i>	10	A	A	O(8)	R(4)	X	X	X	X	X	X	R(2)	R(2)	X	
<i>Mitrella tuberosa</i>	6	O	O	R(1)	R(1)	X	X	X	X	X	X	R(1)	R(1)	X	
<i>Fusinus luteopictus</i>	8	R	R	X	X	X	X	X	X	X	X	X	X	X	
<i>Turbonilla kelseyi</i>	4	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Acanthodoris brunnea</i>	15	X	X	X	X	X	X	X	X	X	X	X	X	X	
<i>Acanthodoris rhodoceras</i>	10	X	R(1)	X	X	X	X	X	X	X	X	X	X	R(2)	X

Animals (235) cont.	Lg. Dim. (mm)	-1.0 to -1.0m		-1.5 to -1.5m		-2.0 to -2.0m		-2.5 to -3.0m		-3.0 to -3.5m		-4.0 to -4.5m		-4.5 to -5.0m		-5.0 to -5.5m		-6.0 to -6.5m		-6.5 to -7.0m		See Species List
		P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	
<u>Bryozoa</u> (13) continued																						
Hippodiplosia insculpta	70	P	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Hippochoa hyalina	30	P	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Microporella californica	20	P	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
<u>Phoronida</u> (1)																						
Phoronis vancouverensis	-	P																				*
<u>Echinodermata</u> (10)																						
Strongylocentrotus spp.	15	A	R(4)	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
Dermasterias imbricata	300	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
Patiria miniata	170	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
Pisaster brevispinus	350	X	X	X	X	X	X	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	
Pisaster giganteus	200	X	X	X	X	X	X	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	
Pisaster ochraceus	200	X	X	X	X	X	X	X	X	R(1)	X	X	X	X	X	X	X	X	X	X	X	
Amphipholis squamata	40	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Ophiotheris papillosa	15	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Ophiothrix spiculata	50	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Eupentacta quinquesemita	10	R(1)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
<u>Chordata</u> (4)																						
Aplidium solidum	170	F	A	F(14)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	F(16)	*
Ascidia ceratodes	30	O	R(4)	R(1)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	R(5)	
Styela montereyensis	40	R(2)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
Styela truncata	10	R(3)	R(4)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
<u>Plants</u> (7)																						
<u>Phaeophyta</u> (4)																						
Laminaria spp.	150	X	P	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	*
Dictyonereis californicum	800	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Dictyonereis reticulata	100	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Macrocystis pyrifera	120	P	P	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
<u>Rhodophyta</u> (3)																						
Rhodomenia pacifica	30	X	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	
Polyneura latissima	20	X	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	
Pterosiphonia dendroidea	15	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	

Table III

Explanation of symbols used in the Table of Species for numbers of organisms collected in the 0.5 m² samples.

Symbol	Descriptive Term	Numbers
R	Rare	1 - 5
O	Occasional	6 - 10
F	Frequent	11 - 20
A	Abundant	> 20
P	Present	Undetermined
X	Absent	0

APPENDIX B Species List

ANIMALS

PROTOZOA: While many unicellular forms were seen and were known to have been collected, two species were large enough to be recognized easily under the dissecting scope.

SARCODINA

Gromia oviformis Dujardin, 1835.

FORAMINIFERA

Haplophragmoides columbiensis var. evolutum Cushman and McCulloch, 1939.

PORIFERA: The classification of sponges presents difficulty due to the varied structure of the spicules. Several encrusting forms were collected at intermediate depths but were not identified.

DEMOSPONGIAE

Cliona celata Grant, 1826. Found at all depths penetrating mostly dead Balanus nubilus shells. At -4.0 to -4.5 m, however, one large shell was taken alive while moderately infested by Cliona celata. The sponge had penetrated through the bases of the lateral parapets but had not bored through the inner calcareous mantle layer. Numbers tabulated represent the number of discrete shells in which the sponge was found. In some cases C. celata existed separately from the shells between the piling surface and outer layers of organic matter.

Stelletta clarella de Laubenfels, 1930. One found (-4.0 to -4.5 m) associated with Scrupocellaria californica and identified, in part, by the penetration of the surface spicules into the fingers when handled (Ricketts and Calvin, 1968. p. 106).

CALCAREA

Leucosolenia eleanor Urban, 1905. Numbers tabulated refer to the individual clusters of anastomosing tubes.

Leucandra heathi Urban, 1905. Rarely found; attached in Bryozoan colonies and with Phyllochaetopterus prolifica. All were less than 20 mm except one at 60 mm.

Leucilla nuttingi (Urban, 1902). Groups are tabulated. Typically, 4-10 individuals were found in close association in each group.

COELENTERATA

HYDROZOA

Obelia spp. Polyps were commonly encountered but species were difficult to count and classify. Large dense masses were not seen as are common in the intertidal zone.

ANTHOZOA

Metridium senile (Linnaeus, 1767). Extremely numerous throughout. Fewer numbers were particularly noted at the base of the piling and in the Phyllochaetopterus prolifica tubes at shallow depths. Attachment occurred on all substrates available.

PLATYHELMINTHES

TURBELLARIA (The only order present was POLYCLADIDA.)

Hoploplana californica Hyman, 1953.

Notoplana acticola (Boone, 1929).

Notoplana inquieta (Heath and McGregor, 1912). Most common flatworm observed. They were most abundant among the tube bases of Phyllochaetopterus prolifica.

Stylochus atentaculatus Hyman, 1953.

Stylochus tripartitus Hyman, 1953.

Eurylepta aurantiaca (Heath and McGregor, 1912).

NEMERTEA

ANOPLA

- Carinoma mutabilis Griffin, 1898.
Tubulanus pellucidus (Coe, 1895).
Tubulanus sexlineatus (Griffin, 1898).
Baseodiscus punnetti (Coe, 1904).
Cerebratulus californiensis Coe, 1905.
Lineus pictifrons Coe, 1904.
Lineus vegetus Coe, 1931.
Micrura pardalis Coe, 1905.
Micrura verrilli Coe, 1901.
Amphiporus bimaculatus Coe, 1901.
Amphiporus imparispinosus Griffin, 1898.
Paranemertes peregrina Coe, 1901.
Tetrastemma nigrifrons Coe, 1904.

SIPUNCULA

Phascolosoma agassizii Keferstein, 1867. Found in piling cracks, dead Balanus nubilus shells, Phyllochaetopterus prolifica colonies and dense masses of Phoronis vancouverensis.

ANNELIDA

POLYCHAETA (This was the most diverse group found and is presented here by families following Blake in the Light's Manual, 1975.)

POLYNOIDAE

Eunoe barbata Moore, 1910 /Ha, 1968/*. Found free-living among Phyllochaetopterus prolifica tubes.

*See Table IV at the end of the Species List.

Halosydna brevisetosa Kinberg, 1855. The most common scale worm present. Hartman (1968) describes four species of Halosydna, three of which (less H. latior Chamberlain, 1919.) appeared to exist on the piling but were difficult to differentiate. Blake in the Light's Manual (1975) equates the two most common species found on the piling--H. brevisetosa and H. johnsoni (Darboux, 1899). His scheme is being followed here. The other species from Hartman--H. tuberculifer Chamberlain, 1919-- is believed to exist on the piling but separation from H. brevisetosa is difficult.

Lepidonotus squamatus (Linnaeus, 1767.)

Thormora johnstoni (Kinberg, 1855) /Ha, 1968/.

PEISIDICIDAE

Peisidice aspera Johnson, 1897. Very common and occurring at all depths with Halosydna brevisetosa.

CRYSOPETALIDAE

Paleanotus bellis (Johnson, 1897).

PHYLLODOCIDAE

Anaitides groenlandica (Oersted, 1843).

Anaitides madeirensis (Langerhans, 1880)
/Ha, 1968/.

Anaitides mucosa (Oersted, 1843).

Anaitides williamsi Hartman, 1936.

Eteone californica Hartman, 1936.

Eulalia aviculisetata Hartman, 1936. The most common Phyllodocid found over the entire piling.

Eulalia bilineata (Johnston, 1840).

Eulalia viridis (Linnaeus, 1767).

Eumida bifoliata (Moore, 1909). Abundant in the Phyllochaetopterus prolifica masses. Two forms were noted differing only in the coloration of their peristomal segment and their location. The first with the peristomium the same color as posterior segments was found consistently on and among P. prolifica tubes; whereas, the second with an irridescent white coloration on the dorsum of the peristomium was found in otherwise empty tubes. Specimens of the latter sent to Professor James A. Blake, Pacific Marine Station, University of the Pacific, Dillon Beach, California, were classified as Eumida bifoliata.

Genetyllis castanea (Marenzeller, 1879).

Notophyllum imbricatum Moore, 1906 /Ha, 1968/.
One large specimen was collected.

Sige californiensis Chamberlin, 1919.

HESIONIDAE

Amphiduros pacificus Hartman, 1961 /Ha, 1968/.
Specimens collected were extremely fragile. Tentacular processes and dorsal cirri were easily broken off with handling or when the animal was placed in alcohol.

Ophiodromus pugettensis (Johnson, 1901). All specimens were free-living with none found in the ambulacral grooves of the asteroids collected (Light's Manual, 1975. p. 184-185; Hartman, 1968. p. 369).

SYLLIDAE

Amblosyllis sp. (= Amblyosyllis sp., Hartman, 1968. p. 397).

Autolytus varius Treadwell, 1914 /Ha, 1968/. The specimens collected were taken from inside an unknown encrusting sponge and agree favorably with the description by Hartman (1968, p. 403); however, they did not appear to have the nuchal epaulettes extending from the hind margin of the prostomium as depicted by Autolytus spp. by Blake in the Light's Manual (1975, p. 184).

Eusyllis assimilis Marenzeller, 1875 /Ha, 1965/.

Exogone sp. Specimen was too small to identify species.

Haplosyllis spongicola (Grube, 1855). Often seen with 10-15 posterior-most segments enlarged and purple from the contained ova. Occasionally, separate eye-spots and rudimentary tentacles would be seen on the first of these segments. Also, these segments were twice seen as separate organisms, freely functioning, with prostomium, eyes, tentacles (similar to Exogone spp.) and simple limbate setae dorsally positioned in the parapodia as well as a compliment of compound falsigers like those commonly found in the Syllids.

Odontosyllis phosphorea Moore, 1909.

Odontosyllis sp. This form is similar to O. phosphorea except in coloration. Rather than the dorsum having dark, transverse bands there are five distinct longitudinal bands dorsally between the two dorsal cirri. Specimens were sent to Professor Blake who stated that the various species of this genus have yet to be described for California and the specimens did not exactly agree with those species previously recorded from the west coast.

Pionosyllis gigantea Moore, 1908.

Syllis elongata (Johnson, 1901).

Syllis gracilis Grube, 1840.

Typosyllis aciculata Treadwell, 1945.

Typosyllis bella Chamberlin, 1919 /Ha, 1968/.

Typosyllis fasciata (Malmgren, 1867) /Ha, 1968/.

Typosyllis hyalina (Grube, 1863).

Typosyllis pulchra (Berkeley and Berkeley, 1938).

NEREIDAE

Neanthes caudata (della Chiaje, 1828) /Ha, 1968/.

Nereis eakini Hartman, 1936. The most common Nereid encountered. All specimens counted in this species had the same characteristics as described by Hartman (1968, p. 537) except that on area III of the maxillary ring specimens collected had about 20 paragnaths vice 4-6.

Nereis latescens Chamberlin, 1919.

Nereis natans Hartman, 1936 /Ha, 1968/. The specimens collected were similar to this species described by Hartman (1968, p. 545) but in addition the posterior dorsal cirri had 8-10 discrete bumps on the leading edges. Also, the body lengths were greater -15 mm vice 8 mm.

Nereis pelagica neonigripes Hartman, 1936.

Nereis vexillosa Grube, 1851. It was expected that more specimens would be found and at shallower depths than the two tabulated at -4.0 to -4.5 m (Hartman, 1968, p.551).

Nereis zonata Malmgren, 1867 /Ha, 1968/.

Platynereis bicanaliculata (Baird, 1863).

Pseudonereis sp. /Ha, 1968/. The genus was included in the key by Hartman (1968, p. 497) but no species was described.

EUNICIDAE

Palola paloloides (Moore, 1909) /Ha, 1968/. An extremely long species. Specimens taken were found in and among dead Balanus nubilus shells, bryozoans and Aplidium solidum.

DORVILLEIDAE

Dorvillea moniloceras (Moore, 1909). A very common species most abundant at shallow depths among Phyllochaetopterus prolifica tube masses.

Dorvillea rudolphi (delle Chiaje, 1828).

LUMBRINERIDAE

Lumbrineris erecta (Moore, 1904). The most common Lumbrinerid found at all depths but particularly abundant shallow in Phyllochaetopterus prolifica tube masses.

Lumbrineris tetraura (Schmarda, 1861).

Lumbrineris zonata (Johnson, 1901).

ARABELLIDAE

Arabella iricolor (Montagu, 1804). Very long specimens were collected at shallower depths. This species was conspicuously absent from mid-depths.

Drilonereis nuda Moore, 1909.

ORBINIDAE

Naineris dendritica (Kinberg, 1867).

SPIONIDAE (It was expected that more specimens of this family would be found than the several collected from the descriptions given by Blake in the Light's Manual, 1975, p. 208-216.)

Polydora spongicola Berkeley and Berkeley, 1950.

CHAETOPTERIDAE

Phyllochaetopterus prolifica Potts, 1914. The most commonly encountered Annelid. Between -0.6 and -1.5 m the entire surface of the piling was covered by this species. Their tube aggregates form the basis for the diversity of the biotic community at these shallow depths. Deeper than -1.5 m the species is generally abundant but significantly reduced in numbers.

CIRRATULIDAE

Caulleriella alata (Southern, 1914) /Ha, 1969/. Very common, particularly among the mats of Phoronis vancouverensis tubes. It is believed that other species of this genus exist but their small size and the physiological characteristics upon which species differentiation is based made classification difficult. Most specimens observed were smaller than the 15 mm listed.

Cirratulus cirratus (Muller, 1776).

Cirriformia luxuriosa (Moore, 1904).

Dodecaceria fewkesi Berkeley and Berkeley, 1954.
This was originally classified from Hartman (1968, p. 255) as D. concharum Oersted, 1943, from the shape of the distally excavate, thick spines imbedded in the posterior segments and from the description that this species is found penetrating into calcareous shells. The specimens collected were abundant in almost all the dead Balanus nubilus shells. Specimens were sent to Professor Blake who identified them as Dodecaceria fewkesi, however. At -4.0 to -4.5 m one large, live Balanus nubilus shell was found which had been penetrated to the inner mantle layer by Dodecaceria fewkesi.

SCALIBREGMIDAE

Onoscolex pacificus (Moore, 1909). Abundant among the basal portions of Phyllochaetopterus prolifica tubes only.

OPHELIIDAE

Polyophthalmus pictus (Dujardin, 1839).

CAPITELLIDAE

Capitella capitata (Fabricius, 1780).

SABELLARIIDAE (Both forms observed were small and found on Phyllochaetopterus prolifica tubes only.)

Sabellaria gracilis Hartman, 1944.

Sabellaria nanella Chamberlin, 1919.

TEREBELLIDAE (All species except Polycirrus spp. and Thelepus setosus were only found forming tubes with Phyllochaetopterus prolifica masses in shallow water.)

Amphitrite cirrata (Muller, 1776) /Ha, 1969/.

Eupolymnia crescentis Chamberlin, 1919.

Neoamphitrite robusta (Johnson, 1901).

Pista alata Moore, 1909 /Ha, 1969/.

Pista brevibranchiata Moore, 1923.

Pista pacifica Berkeley and Berkeley, 1942.

Polycirrus spp. A fairly common form. Blake in the Light's Manual (1975, p. 234-235) and Hartman (1969, p. 629) state that this genus is confused and the species are unclear.

Ramex californiensis Hartman, 1944.

Spinospaera oculata Hartman, 1944.

Thelepus crispus Johnson, 1901.

Thelepus hamatus Moore, 1905 /Ha, 1969/.

Thelepus setosus (Quatrefages, 1865) /Ha, 1969/.

SABELLIDAE

Chone mollis (Bush, 1904).

SERPULIDAE

Chitinopoma groenlandica (Morch, 1863) /Ha, 1969/.

Eupomatus gracilis Bush, 1904.

Serpula vermicularis Linnaeus, 1767. Small specimens were taken off Phyllochaetopterus prolifica tubes at shallow depths while the largest ones were attached to the exposed surface of Pododesmus cepio and inside dead Balanus nubilus shells at intermediate and deep depths.

Spirorbis borealis Daudin, 1800.

Spirorbis eximius Bush, 1904.

Spirorbis moerchi Levinsen, 1883.

Spirorbis spirillum (Linnaeus, 1758).

Spirorbis spp.

NOTE: The above Spirorbis species are listed after Blake in the Light's Manual (1975, p. 239-242). He states that the systematics of Spirorbis are not well known. Specimens of the above forms were sent to Dr. Phyllis Knight-Jones, University College of Swansea, Singleton Park, Swansea, Wales, as she is preparing a paper on the Spirorbids found between Alaska and Panama. She is considering the Spirorbids as a

family apart from the Serpulids and apparently assigning different generic and specific names to those listed above. Unfortunately, the complete key and paper are not due for publication "for a year or two." An added problem exists in the classification of these organisms with the occurrence in some of opposite spiraling of the calcareous tubes.

ARTHROPODA (The following list of Crustaceans--the only class present--is arranged by sub-family after the Light's Manual, 1975, p. 244-424.)

CRUSTACEA

CIRRIPEDIA (The dimensions tabulated are basal diameters.)

Balanus aquila Pilsbry, 1907.

Balanus crenatus Bruguiere, 1897. The most common barnacle present, most abundant attached to the piling at deepest depths and attached to Balanus nubilus, large algae, crabs, and Phyllochaetopterus prolifica tubes at shallower depths.

Balanus nubilus Darwin, 1854. The largest barnacle found was present at all depths attached directly to the pilings or smaller forms on larger ones of the same species. Only 25 per cent were collected alive. The shells of the rest were degraded to various stages by Cliona celata and Dodecaceria fewkesi. Whether or not these organisms actually caused the death of the barnacles could not be discerned. The shells provided a point of attachment for a number of other organisms including Bryozoans, Annelids, Metridium senile, sponges, Phoronis vancouverensis and tunicates.

Balanus tintinnabulum californicus Pilsbry, 1916. Only small forms were found. The specimen taken at -4.0 to -4.5 m was attached to the carapace of Loxorhynchus crispatus.

MALACOSTRACA

Idotea resecata Stimpson, 1857. Small forms were found on Macrocystis pyrifera holdfasts.

Jaeropsis dubia dubia Menzies, 1951.

Accedomoera vagor Barnard, 1969. Gammarid.

Atylus levidensus Barnard, 1956, Gammarid.

Corophium insidiosum Crawford, 1937. The most abundant Gammarid seen forming burrows in detritus.

Micropotopus sp. /Li/. Gammarid.

NOTE: The above four Gammarids represent the largest or most abundant species collected. It was obvious additional species were collected but they could not be identified due to size or systematic complexity.

Perotripus brevis (La Follette, 1915).

Deutella californica Mayer, 1890.

Tritella laevis Mayer, 1903.

Caprella verrucosa Boeck, 1871.

Heptacarpus paludicola Holmes, 1900.

Heptacarpus taylori (Stimpson, 1857).

Spirontocaris prionta (Stimpson, 1864).

Alpheus dentipes (Guerin, 1832) /Sch/.

Bateus harfordi (Kingsley, 1878).

Loxorhynchus crispatus Stimpson, 1857. The largest specimen taken had a mass of 182 grams (-4.0 to -4.5 m). The ones taken from shallower depths were all small--to 10 mm carapace length.

Mimulus foliatus Stimpson, 1860.

Pugettia producta (Randall, 1839).

Cancer antennarius Stimpson, 1856.

Cancer jordani Rathbun, 1900.

Cancer sp. This form was commonly found at shallower depths but could not be identified as to species. Dorsolateral carapace spines numbered 4-5. This is believed to be an immature form of one of the above Cancer species.

Lophopanopeus bellus (Stimpson, 1860).

Pinnixa longipes (Lockington, 1877).

Pachycheles pubescens Holmes, 1900.

Pachycheles rudis Stimpson, 1859.

Petrolisthes cinctipes (Randall, 1839).

PYCNOGONIDA

Phoxichilidium femoratum (Rathke, 1799).

Pycnogonum stearnsi Ives, 1892. This was the only species of Pycnogonum encountered. Hedgpeth in the Light's Manual (1975, p. 424) describes P. rickettsi Schmitt, 1934, as usually being associated with Metridium senile. Despite careful inspection of the numerous M. senile collected, it was not observed.

MOLLUSCA (Species are presented by class.)

POLYPLACOPHORA (The chitons appeared to be limited in their extent on the piling due to the lack of a relatively free, hard surface as a substrate for attachment. This was with the exception of Lepidozona californiensis which was often seen in shallow water clinging to Phyllochaetopterus prolifica tubes.)

Callistochiton crassicostatus Pilsbry, 1893.

Lepidozona californiensis Berry, 1931 /Mc/.

Lepidozona mertensii (Middendorff, 1846).

Mopalia ciliata (Sowerby, 1840).

GASTROPODA

Diodora aspera (Rathke, 1833).

Megatebennis bimaculatus (Dall, 1871).

Acmaea mitra Rathke, 1833.

Lacuna unifasciata Carpenter, 1857.

Alvinia compacta (Carpenter, 1864) /Mc/.

Barleeia acuta (Carpenter, 1864).

Barleeia californica Bartsch, 1920 /Mc/.

Truncatella californica Pfeiffer, 1857 /Mc/.
Specimens collected were complete and not truncated.

Caecum californicum Dall, 1885.

Caecum dalli Bartsch, 1920 /Mc/. Specimens taken had significantly more numerous and more closely spaced annular rings than C. californicum.

Fartulum occidentale Bartsch, 1920 /Mc/.

Bittium attenuatum Carpenter, 1864.

Crepidatella lingulata (Gould, 1846).

Polinices sp. This form was too small to determine the species.

Amphissa versicolor Dall, 1871.

Mitrella aurantiaca (Dall, 1871).

Mitrella carinata (Hinds, 1844).

Mitrella tuberosa (Carpenter, 1864).

Fusinus luteopictus (Dall, 1877).

Turbonilla kelseyi Dall and Bartsch, 1909 /Mc/.

OPISTHOBRANCHIA

Acanthodoris brunnea MacFarland, 1905.

Acanthodoris rhodoceras Cockerell and Eliot, 1905.

Aegires albopunctatus MacFarland, 1905.

Aeolidia papillosa (Linnaeus, 1761).

Anisodoris nobilis (MacFarland, 1905).

Archidoris montereyensis (Cooper, 1862).

Cadlina luteomarginata MacFarland, 1966.

Cadlina modesta MacFarland, 1966.

Carambe pacifica MacFarland and O'Donoghue, 1929.

Doriopsilla albopunctata (Cooper, 1863).
Hermisenda crassicornis (Eschscholtz, 1831).
Polycera atra MacFarland, 1905.
Trinchesia albocrusta (MacFarland, 1966).
Triopha carpenteri (Sterns, 1873).
Triopha maculata MacFarland, 1905.

BIVALVIA

Lima hemphilli Hertlein and Strong, 1946 /Ke/.
These beautiful, active file shells with very long annulated mantle tentacles were found among the bases of Phyllochaetopterus prolifica tubes.

Hinnites giganteus (Gray, 1825). The abundance of these rock scallops as tabulated is associated mostly with their juvenile forms which were about 10 mm long and were similar in appearance to Pectinidae genera. Only one adult was found at -1.5 to -2.0 m.

Pododesmus cepio (Gray, 1850). A large shell attached usually directly to the piling but several smaller forms were attached to Balanus nubilus shells.

Adula diegensis (Dall, 1911). Specimens taken were small and mostly attached to Phyllochaetopterus prolifica tubes.

Lithophaga plumula kelseyi Hertlein and Strong, 1946. While often found boring into calcareous substrata, the specimens found here were nestling in the recesses of dead Balanus nubilus shells.

Modiolus carpenteri Soot-Ryen, 1963.

Modiolus rectus (Conrad, 1837).

NOTE: No specimens of Modiolus capax (Conrad, 1837) were found though they are usually a common piling form (Coan and Carlton in the Light's Manual, 1975, p. 555).

Mytilus californianus Conrad, 1837.
Mytilus edulis Linnaeus, 1758.

Modiolus spp. Numerous small mussels were seen throughout the intermediate and shallower depths with periostrecal hairs present. It is believed some might be Mytilus edulis juveniles however.

Gregariella chenui (Recluz, 1842) /Ke/. This rather rare species was found deeply embedded in the bases of Phyllochaetopterus prolifica tubes.

Chama pellucida Broderip, 1835.

Lasaea spp. Numerous small forms believed to be associated with this genus were observed.

Kellia laperousii (Deshayes, 1839). Very common; found nestling in all material.

Protothaca staminea (Conrad, 1837). A beautiful white shell with cancellate sculpture. Some forms had prominent raised concentric lamellae.

Petricola tellimyialis (Carpenter, 1864) /Mc/. An extremely common species found most often associated with Bugula neritina, Scrupocellaria californica and Phyllochaetopterus prolifica.

Semele rupicola Dall, 1915.

Cryptomya californica (Conrad, 1837). Although normally burrowing into sand or mud, three specimens were taken from the pilings.

Hiatella arctica (Linnaeus, 1767). The most common bivalve observed. Specimens took on various distorted forms and some had prominent lamellae.

Penitella conradi Valenciennes, 1846. The only specimen observed was taken from its bored hole in a large, dead Balanus nubilus shell.

Entodesma saxicola (Baird, 1863).

Lyonsia californica Conrad, 1837. Normally found in mud substrates but here were nestled among Phyllochaetopterus prolifica tubes.

ECTOPROCTA (Species are listed according to orders. Individual zoaria could not be separated for counting and their presence or absence only is indicated.)

CTENOSTOMATA

Bowerbankia gracilis O'Donoghue, 1926. Most abundant at shallow depths covering the distal end of Phyllochaetopterus prolifica tubes.

Crisia maxima Robertson, 1910.

Crisulipora occidentalis Robertson, 1910.

Tubulipora tuba (Gabb and Horn, 1862).

CHEILOSTOMATA

Bugula neritina Linnaeus, 1758. A quite common form. Colonies were present in a wide variety of blue-purple-red hues.

Lyrula hippocrepis (Hincks, 1882). Found encrusting on hard substrates as well as enveloping Phyllochaetopterus prolifica tubes.

Membranipora membranacea (Linnaeus, 1767). Found only attached to brown algae.

Scrupocellaria californica Trask, 1857. It is possible that other species of this genus were present but S. californica appeared to be the most common form.

Celleporaria brunnea (Hincks, 1884).

Cryptosula pallasiana (Moll, 1803).

Hippodiplosia insculpta (Hincks, 1882).

Hippochoa hyalina (Linnaeus, 1758) /Os, 1952/.

Microporella californica (Busk, 1856).

PHORONIDA

Phoronis vancouverensis Pixell, 1912. One of the most abundant species of animals found, particularly between -1.5 and -6.0 m. The tubes form thick encrusting mats directly on the piling and establish an anoxic environment in and under them.

ECHINODERMATA (The species are listed by class.)

ECHINOIDEA

Strongylocentrotus spp. All specimens collected were too immature to determine whether they were S. purpuratus (Stimpson, 1857) or S. franciscanus (Agassiz, 1863).

ASTEROIDEA

Dermasterias imbricata (Grube, 1857).

Patiria miniata (Brandt, 1835). The one shallow water specimen was 12 mm across.

Pisaster brevispinus (Stimpson, 1857).

Pisaster giganteus (Stimpson, 1857).

Pisaster ochraceus (Brandt, 1835).

OPHIUROIDEA

Amphipholis squamata (delle Chiaje, 1829).

Ophiopteris papillosa (Lyman, 1875).

Ophiothrix spiculata Le Conte, 1851.

HOLOTHUZOIDEA

Eupentacta quinquesemita (Selenka, 1867).

CHORDATA (Specimens collected were all in the class Acidiacea.)

Aplidium solidum (Ritter and Forsyth, 1917). The larger colonies were taken between -1.5 to -3.0 m. The largest was 576 grams.

Ascidia ceratodes (Huntsman, 1912).

Styela montereyensis (Dall, 1872).

Styela truncata Ritter, 1901. Specimens collected were identified by Dr. D. P. Abbott, Hopkins Marine Station, Pacific Grove, California.

ALGAE

PHAEOPHYTA

Laminaria spp. It is believed a large specimen observed was L. andersonii Eaton, 1876. The holdfasts had branched haptera but the blade was whole and not incised (Smith, 1944, p. 137). Several small forms were found.

Dictyoneurum californicum Ruprecht, 1852.

Dictyoneuropsis reticulata (Saunders, 1895) Smith, 1942.

Macrocystis pyrifera (Linnaeus, 1771) Agardh, 1820.

RHODOPHYTA (Species were identified by Dr. I. A. Abbott, Hopkins Marine Station, Pacific Grove, California.)

Rhodomenia pacifica Kylin, 1931.

Polyneura latissima (Harvey, 1862) Kylin, 1924.

Pterosiphonia dendroidea (Montague, 1837) Falkenberg, 1901.

Table IV

Explanation of bracketed notation in the Species List identifying authors of publications in which observed organisms were classified.

Notation	Author
/Ha, 1968/	Hartman, 1968
/Ha, 1969/	Hartman, 1969
/Ke/	Keen, 1971
/Li/	Light, 1954
/Mc/	McLean, 1969
/Os, 1952/	Osburn, 1952
/Sch/	Schmitt, 1921

APPENDIX C. Drawings of Piling Animals with Key

Plate I

Piling "A" -6.0 m to -7.1 m

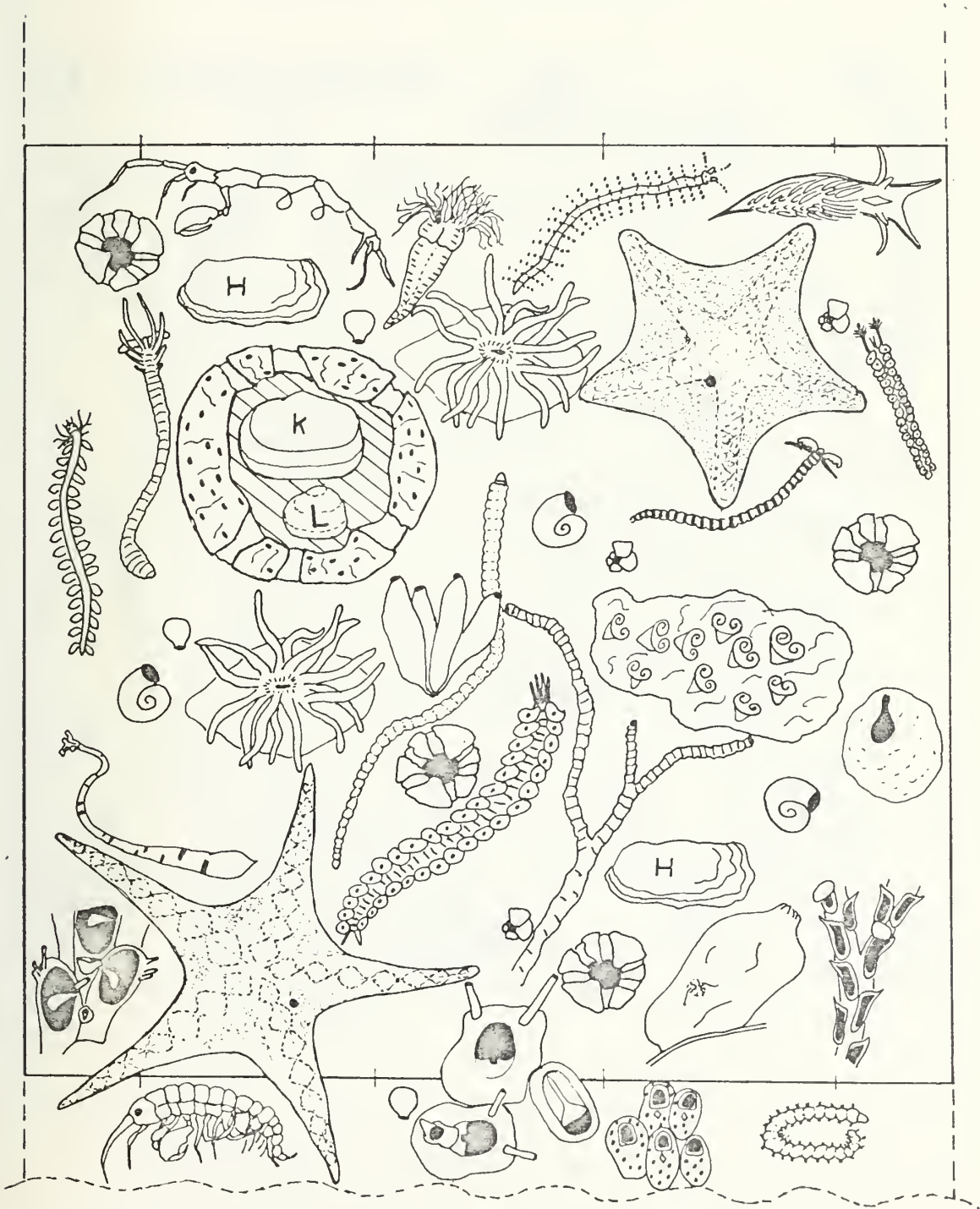


Plate II

Piling "A" -5.0 m to -6.0 m

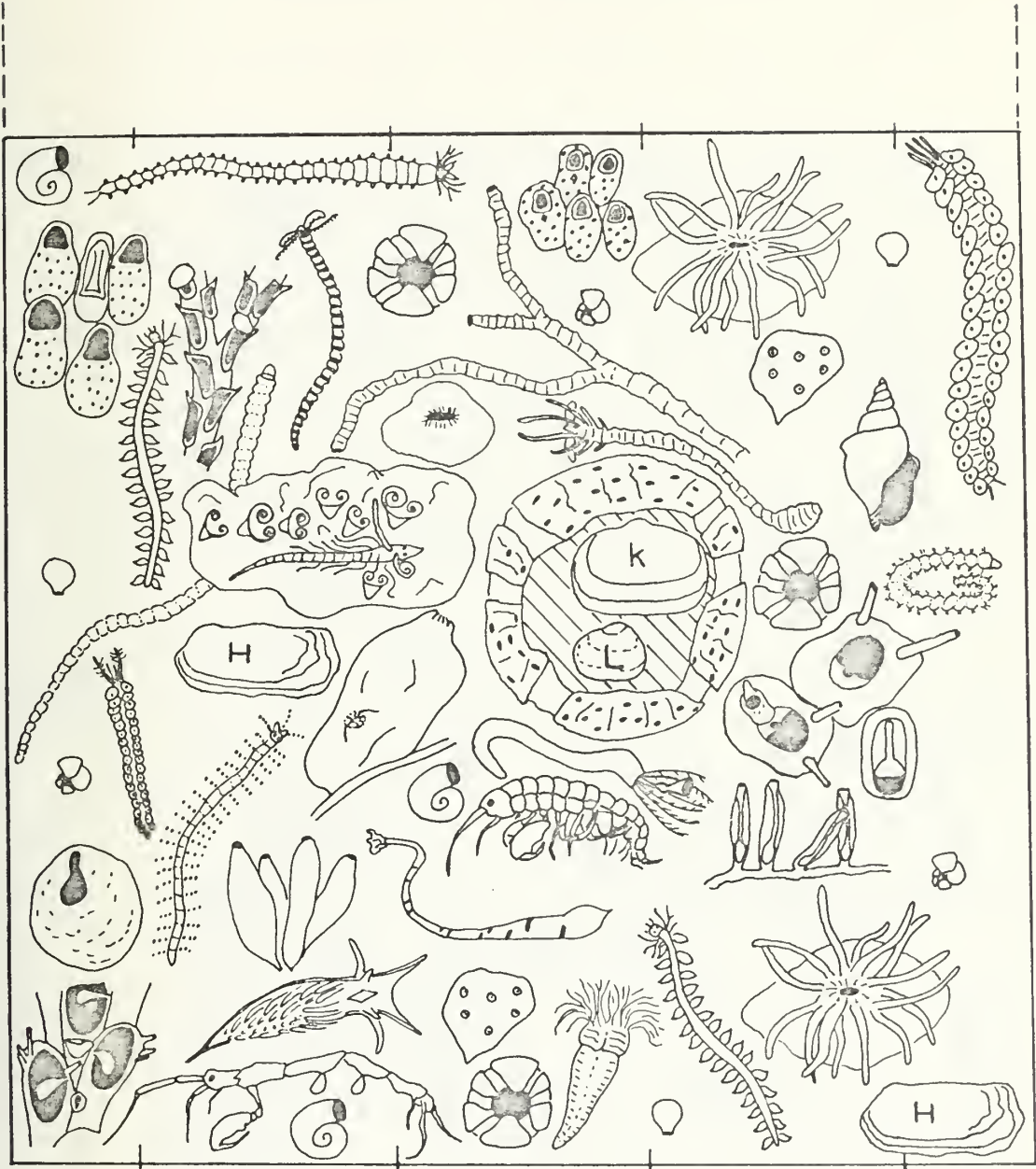


Plate III

Piling "A" -4.0 m to -5.0 m



Plate IV

Piling "A" -3.0 m to -4.0 m

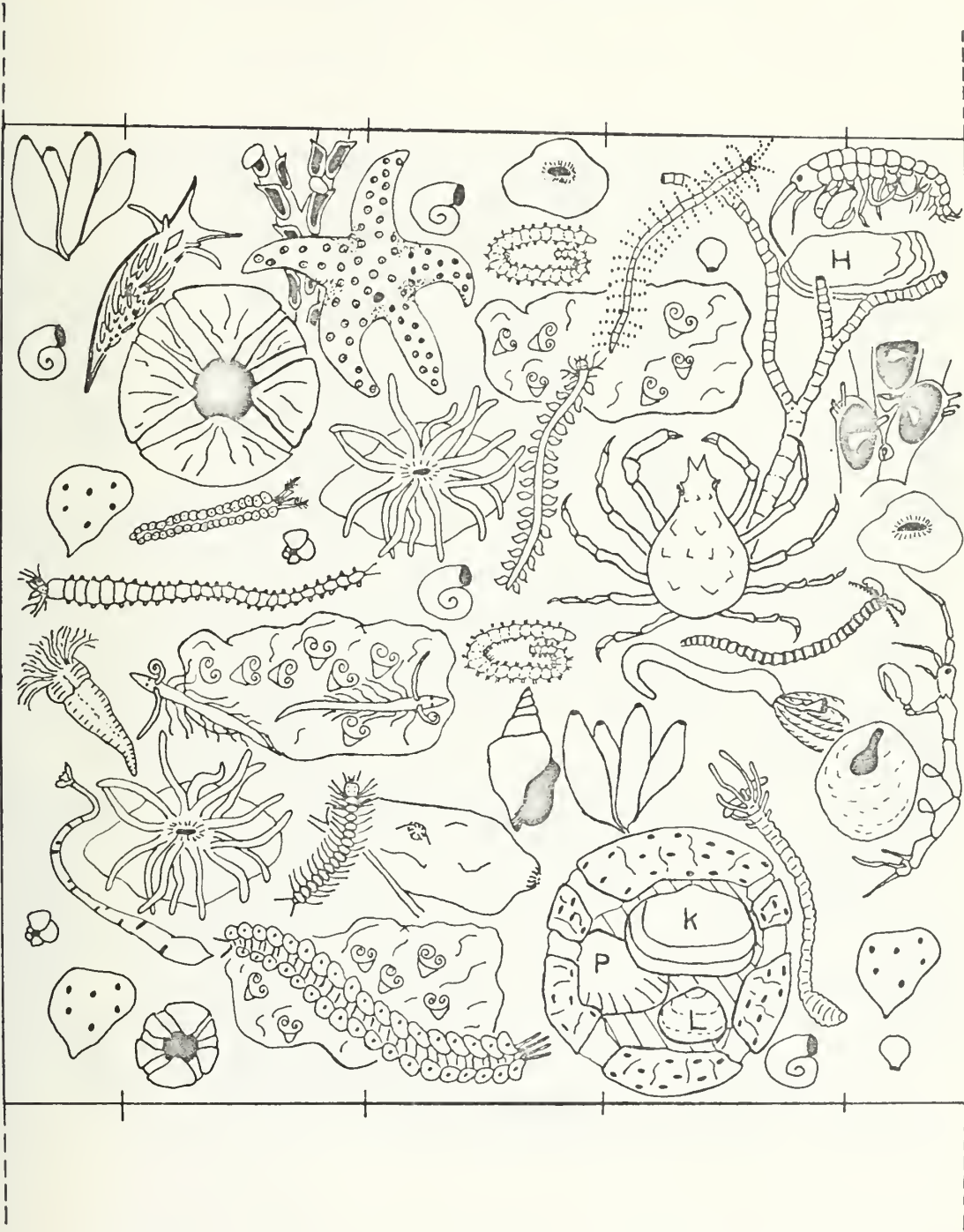


Plate V

Piling "A" -2.0 m to -3.0 m

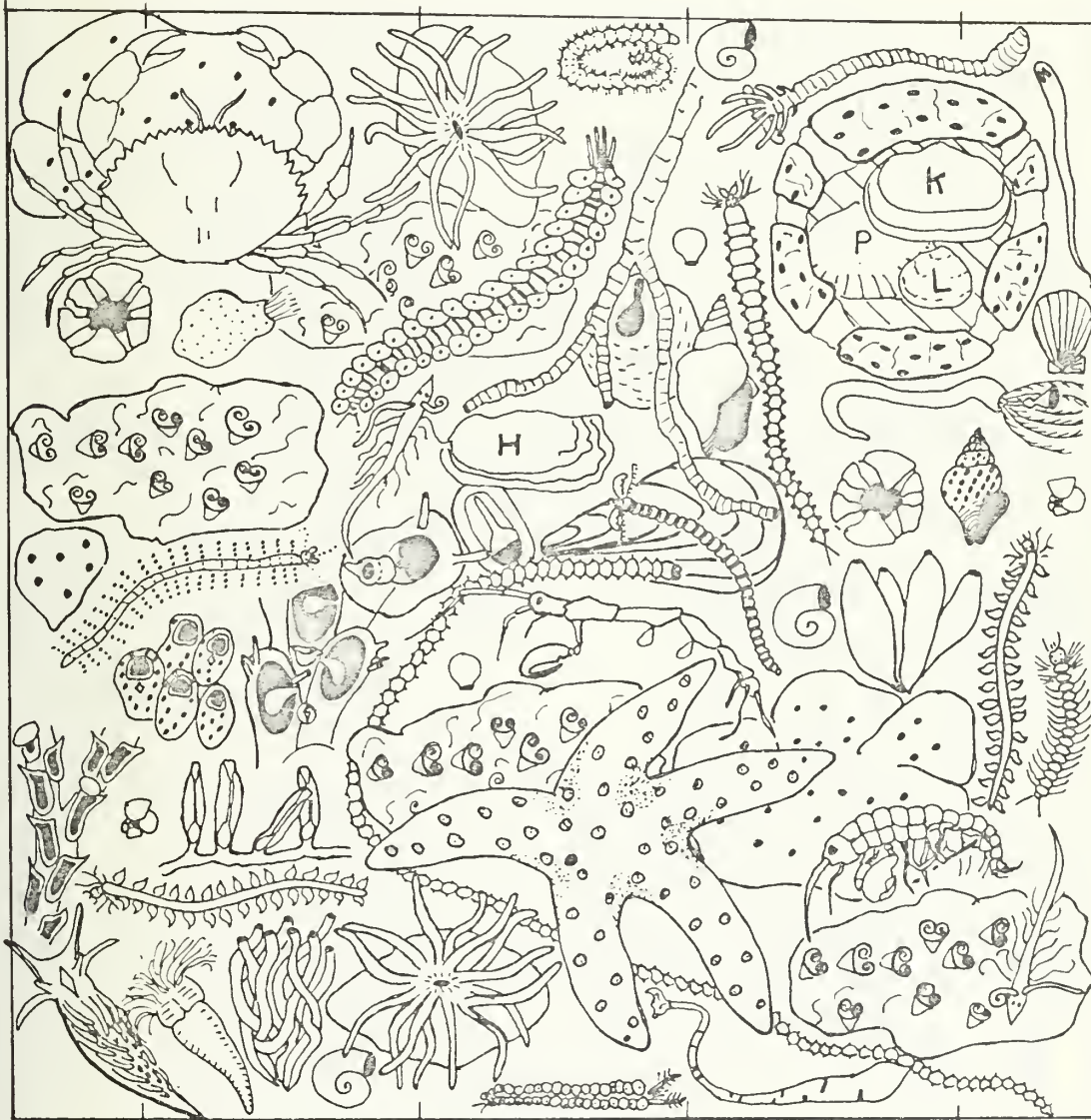


Plate VI

Piling "A" -1.0 m to -2.0 m



Key to Plates I - VI

Figures represent prominent species as listed. Others are composite forms of several species. Sources of the figures are given for those that are not original.

PROTOZOA



Gromia oviformis



Haplophragmoides columbiensis
var. evolutum

PORIFERA



Cliona celata (black spots
represent oscula protruding
from barnacle wall plate)



Leucosolenia eleanor

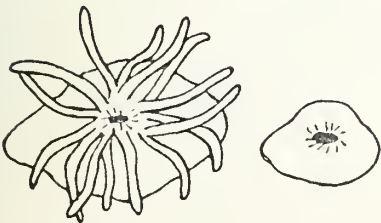


Leucandra heathi



Leucilla nuttingi

COELENTERATA



Metridium senile

PLATYHELMINTHES

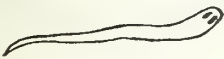


Turbellarian form

NEMERTEA



Cerebratulus californiensis



Amphiporus bimaculatus

SIPUNCULA



Phascolosoma agassizii

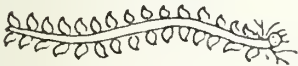
ANNELIDA



Halosydna brevisetosa



Peisidice aspera



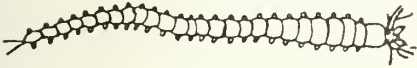
Phyllodocid form



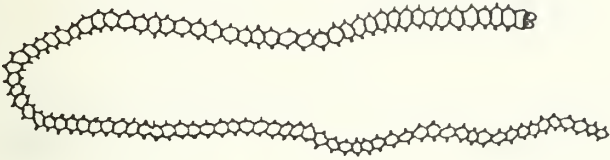
Ophiodromus pugettensis



Syllid form



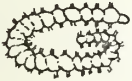
Nereid form



Palola paloloides



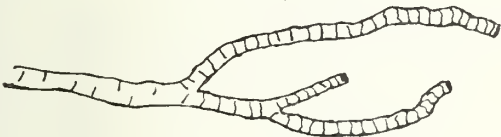
Dorvillea moniloceras



Lumbrinerid form



Arabella iricolor



Phyllochaetopterus prolifica
(tube)



Caulleriella alata



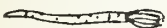
Dodecaceria fewkesi



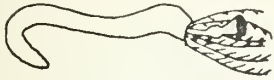
Polycirrus spp.



Spinosphaera oculata



Chone mollis



Serpula vermicularis

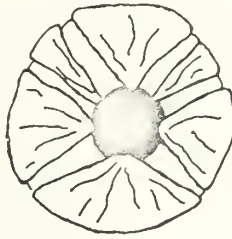
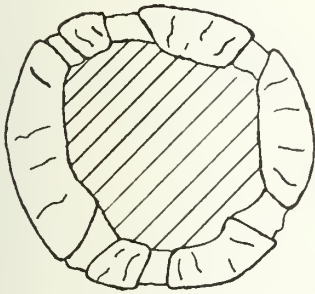


Spirorbis spp.

ARTHROPODA



Balanus crenatus



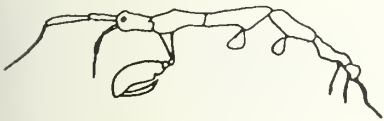
Balanus nubilus (dead and alive)



Jaeropus dubia dubia



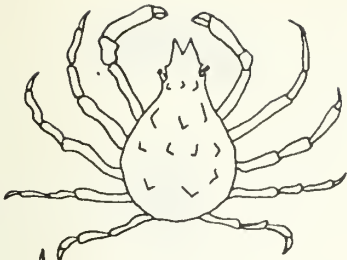
Gammarid form



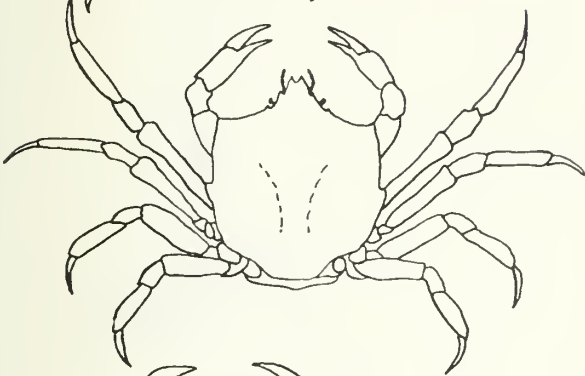
Caprellid form



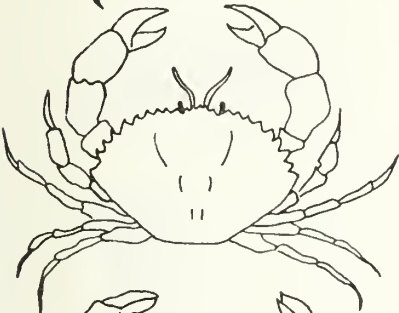
Caridean form



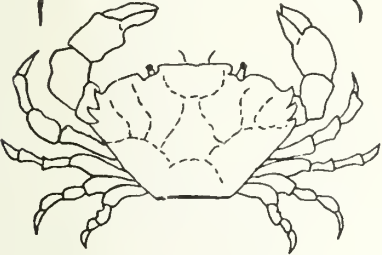
Loxorhynchus crispatus



Pugettia producta (after
Smith and Carlton, 1975,
p. 397.)



Cancer antennarius (after
Smith and Carlton, 1975,
p. 397.)

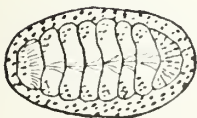


Lophopanopeus bellus



Pycnogonum stearnsi

MOLLUSCA



Lepidozona californiensis



Caecium californicum



Amphissa versicolor



Mitrella spp.



Hermissenda crassicornis



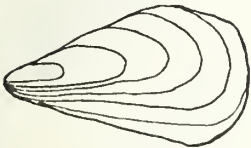
Trinchesia albocrusta



Hinnites giganteus (juvenile
STAGE)



Pododesmus cepio (view of
attached valve)



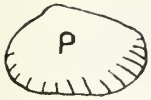
Mytilid form



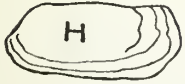
Lasaea spp.



Kellia laperousii



Petricola tellimyalis



Hiatella arctica

BRYOZOA



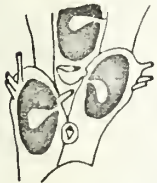
Bowerbankia gracilis



Bugula neritina



Lyrula hippocrepis



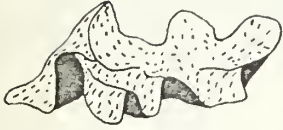
Scrupocellaria californica



Celleporaria brunnea

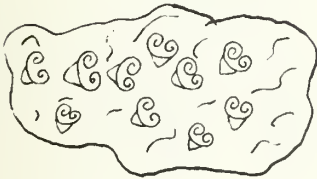


Cryptosula pallasiana



Hippodiplosia insculpta

PHORONIDA

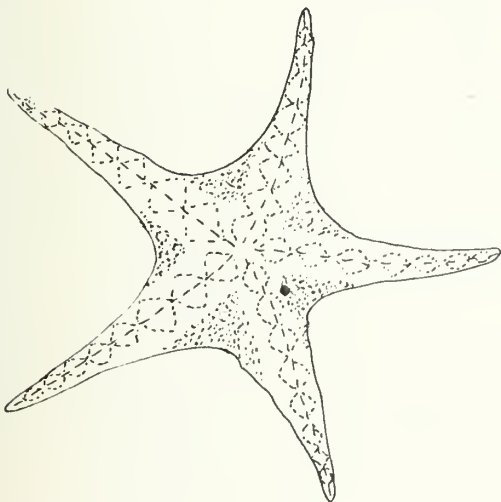


Phoronis vancouverensis

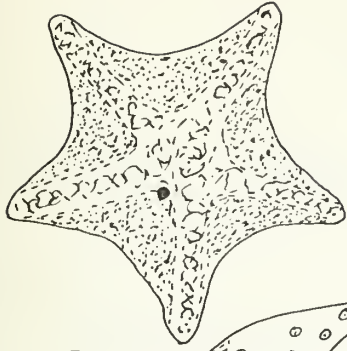
ECHINODERMATA



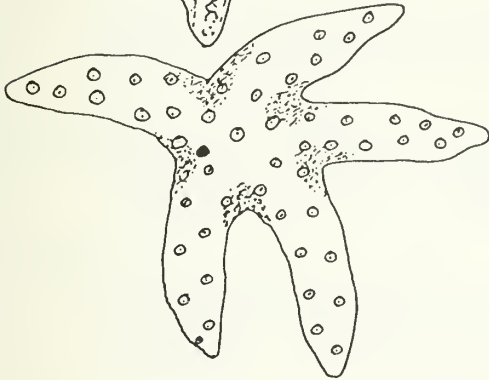
Strongylocentrotus spp.



Dermasterias imbricata

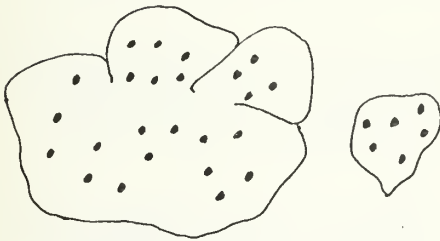


Patiria miniata



Pisaster giganteus

CHORDATA



Aplidium solidum



Ascidia ceratodes

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