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Dermocystidium sp. (Mesomycetozoea: Dermocystidiaceae) primary ocular infection in a koi carp (Cyprinus carpio var. koi)

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Keywords:	Mesomycetozoea, Dermocystidium sp., koi carp, eye

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1	SHORT COMMUNICATION
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3	Dermocystidium sp. (Mesomycetozoea: Dermocystidiaceae) primary ocular infection in a koi
4	carp (Cyprinus carpio var. koi)
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14	Running head: Dermocystidium sp. ocular infection in a koi
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Dermocystidium spp. (Mesomycetozoea: Dermocystidiaceae) comprises several species able to 26 27 infect a wide range of marine and freshwater fish (Gozlan et al., 2014; Rowley et al., 2013). The taxonomic position of *Dermocystidium* is still under debate, even though Dyková and Lom (1992) 28 observed hyphae associated to *Dermocystidium koi* infection, suggesting its fungal nature. 29 Currently, the genus is classified in the superclass Mesomycetozoea which includes protists 30 "fungus-like" in the supergroup Opisthokonta (Gozlan et al., 2014). 31 The lesions caused by *Dermocystidium* spp. in fish are chronic and characterized by visible cysts or 32 nodules on skin and fins (Feist et al., 2004; Wildgoose, 1995; Zhang & Wang, 2005), but severe 33 gills and systemic infections causing mortality have been described in several species (Höglund et 34 al., 1997; Olson & Holt, 1995; Athanassopoulou, 1998; Landsberg & Paperna, 1992; Kirkbright et 35 al., 2016; Fujimoto et al., 2018). 36 The present study described a primary ocular infection by *Dermocystidium* sp. in a koi carp 37 (Cyprinus carpio var. koi). The koi carp was found dead by the owner of a private pond and sent 38 refrigerated to the laboratory. The fish presented an evident subconjunctival whitish exophytic 39 tissue, surrounded by reddish edges, that protruded over both ocular bulbs (Fig. 1a). The fish was 40 necropsied and tissues fixed in 10% buffered formalin. The ocular globes were then dissected under 41 stereomicroscope and smears of the exophytic tissue placed on slides, clarified with lactophenol or 42 43 stained with China ink, and examined under light microscope. For histopathology, tissues were routinely processed, sections were cut at 4 µm and stained with Haematoxylin and Eosin (H&E), 44 Periodic Acid Shiff (PAS) and Gomori. 45 46 At the stereomicroscope, the peribulbar tissue and anterior chamber presented numerous filiform 3mm-length white hyphal-like cysts (Fig. 1b). At the light microscope, the cysts were filled with 47 round variably sized (5.53-12.90 µm in diameter; mean=9.71; sd=1.99; n=80) mature spores 48 referable to *Dermocystidium* sp. admixed with developmental stages (>5 μm). The spores showed a 49 large refractile body or central vacuole and a peripheral cytoplasm with a nucleus and three or more 50 51 vacuoles (Fig. 1c).

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At histology, the peribulbar soft tissues until muscle and the choroid space of the posterior chamber, adjacent to the retina, were infiltrated by numerous elongated cysts (Fig. 2f-h). The cysts had a thick eosinophilic wall and contained numerous spores (Fig. 2h,i). Around the cysts, a granulomatous inflammatory reaction was visible (Fig. 2h). The iris and the ciliary body were also replete by numerous free mature spores. An intense mixed inflammatory infiltrate with multinucleated giant cells, vascular congestion and abundant granulation tissue were observed (Fig. 2g). Haemorrhagic and necrotic areas were observed. Some basophilic granular aggregates referable to bacteria were also found. Neither developmental stages of *Dermocystidium* sp. nor inflammatory reaction were seen in internal organs. PAS and Gomori stains helped to visualize the developmental stages (Fig. 1d,e) and exclude the presence of spores in other organs. In literature, only three descriptions of ocular infection by *Dermocystidium* sp. are reported in fish (Elkan, 1962; Molnar et al., 2008; Fujimoto et al., 2018). The present case is the first report of a primary ocular infection in a koi carp. The spore size and morphology are consistent with literature reports of *Dermocystidium koi* (Gjurcevic et al., 2008; Hoshina & Sahara, 1950). The life cycle of Dermocystidium spp. is partly unknown. The parasite produces motile zoospores which encyst in the host tissues, form sporonts by division of the plasmodia until subsequent maturation of spores (Lotman et al., 2000; Mahboub & Shaheen, 2020). In experimental trials, it has been demonstrated that fish can acquire the infection through intra-gastric and immersion routes. The pathogen cycle begins from the ingestion of spores through feeding on infected fish or contaminated feed or via penetration of spores into the gills (Mahboub & Shaheen, 2020). Considering the economic value of koi carps and the possible direct transmission of the infection in optimal temperatures condition, Dermocystidium spp. infection should be considered as an important disease also in ornamental species.

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Conflicts of interests

The authors declare to have no conflict of interests.

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Data availability

- The data that support the findings of this study are available from the corresponding author upon
- 81 request.

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References

- Athanassopoulou, F. (1998). Susceptibility of carp, roach and goldfish to a *Dermocystidium*-like
- 85 organism. Journal of Fish Biology, 53(5), 1137–1139. https://doi.org/10.1111/j.1095-
- 86 8649.1998.tb00470.x
- Byková, I., & Lom, J. (1992). New evidence of fungal nature of *Dermocystidium koi* Hoshina and
- 88 Sahara, 1950. *Journal of Applied Ichthyology*, 8(1–4), 180–185.
- 89 https://doi.org/10.1111/j.1439-0426.1992.tb00681.x
- 90 Elkan, E. (1962). Dermocystidium gasterostei n. sp., a parasite of Gasterosteus aculeatus L. and
- 91 *Gasterosteus pungitius* L. *Nature*, 196, 958–960. https://doi.org/10.1038/196958a0
- 92 Feist, S. W., Longshaw, M., Hurrell, R. H., & Mander, B. (2004). Observations of *Dermocystidium*
- 93 sp. infections in bullheads, Cottus gobio L., from a river in southern England. Journal of Fish
- 94 Diseases, 27(4), 225–231. https://doi.org/10.1111/j.1365-2761.2004.00535.x
- 95 Fujimoto, R. Y., Couto, M. V. S., Sousa, N. C., Diniz, D. G., Diniz, J. A. P., Madi, R. R., Martins
- 96 M. L., Eiras, J. C. (2018). *Dermocystidium* sp. infection in farmed hybrid fish *Colossoma*
- 97 macropomum × Piaractus brachypomus in Brazil. Journal of Fish Diseases, 41(3), 565–568.
- 98 https://doi.org/10.1111/jfd.12761
- 99 Gjurcevic, E., Bambir, S., Kozaric, Z., Kuzir, S., Gavrilovic, A., & Pašalic, I. (2008).
- Dermocystidium infection in common carp broodstock (Cyprinus carpio L.) from Croatia. In
- Bulletin of the European Association of Fish Pathologists, 28(6), 222-229.
- Gozlan, R. E., Marshall, W. L., Lilje, O., Jessop, C. N., Gleason, F. H., & Andreou, D. (2014).
- 103 Current ecological understanding of fungal-like pathogens of fish: What lies beneath?

- 104 Frontiers in Microbiology, 5(62), 1–16. https://doi.org/10.3389/fmicb.2014.00062
- Höglund, J., Alfjorden, A., & Nikkilä, T. (1997). Infection of juvenile salmon Salmo salar with a
- 106 Dermocystidium-like organism in Sweden. Diseases of Aquatic Organisms, 30(3), 171–176.
- 107 https://doi.org/10.3354/dao030171
- Hoshina, T., & Sahara, Y. (1950). A new Species of the Genus *Dermocystidium*, D. koi sp. nov.,
- Parasitic in *Cyprinus carpio* L. *Nippon Suisan Gakkaishi*, 15(12), 825–829.
- 110 https://doi.org/10.2331/suisan.15.825
- Kirkbright, D., Huber, P., Lillie, B. N., & Lumsden, J. S. (2016). *Dermocystidium*-like organism
- linked with a mortality event in yellow perch *Perca flavescens* (Mitchill) in Ontario, Canada.
- Journal of Fish Diseases, 39(5), 597–601. https://doi.org/10.1111/jfd.12379
- Landsberg, J., & Paperna, I. (1992). Systemic granuloma in goldfish caused by a *Dermocystidium*-
- like aetiological agent. Diseases of Aquatic Organisms, 13, 75–78.
- https://doi.org/10.3354/dao013075
- Lotman, K., Pekkarinen, M., & Kasesalu, J. (2000). Morphological observations on the life cycle of
- Dermocystidium cyprini Cervinka and Lom, 1974, parasitic in carps (Cyprinus carpio). Acta
- 119 *Protozoologica*, 39(2), 125–134.
- Mahboub, H. H., & Shaheen, A. (2020). Prevalence, diagnosis and experimental challenge of
- Dermocystidium sp. infection in Nile tilapia (Oreochromis niloticus) in Egypt. Aquaculture,
- 516, 7345562. https://doi.org/10.1016/j.aquaculture.2019.734556
- Molnar, K., Muller, T., Lefler, K. K., & Csorbai, B. (2008). *Dermocystidium* infection in the eye of
- crucian carp. Magyar Allatorvosok Lapja, 130(1), 53–56.
- Olson, Robert E., & Holt, R. A. (1995). The gill pathogen *Dermocystidium salmonis* in Oregon
- salmonids. Journal of Aquatic Animal Health, 7(2), 111–117. https://doi.org/10.1577/1548-
- 127 8667(1995)007<0111:TGPDSI>2.3.CO;2
- Rowley, J. J. L., Gleason, F. H., Andreou, D., Marshall, W. L., Lilje, O., & Gozlan, R. (2013).
- Impacts of mesomycetozoean parasites on amphibian and freshwater fish populations. *Fungal*

130	Biology Reviews, 27(3-4), 100-111. https://doi.org/10.1016/j.fbr.2013.09.002
131	Wildgoose, W. H. (1995). Dermocystidium koi found in skin lesions in koi carp (Cyprinus carpio).
132	The Veterinary Record, 137(13), 317–318. https://doi.org/10.1136/vr.137.13.317
133	Zhang, Q., & Wang, Z. (2005). Dermocystidium sp. infection in cultured juvenile southern catfish
134	Silurus meridionalis in China. Diseases of Aquatic Organisms, 65(3), 245-250.
135	https://doi.org/10.3354/dao065245
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137	Figure legends
138	Figure 1. (a) Koi carp showing an ocular exophytic whitish tissue. (b) Numerous filiform hyphal-
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140	spores morphologically referable to <i>Dermocystidium</i> sp. (bar=10 μm). The spores show a large
141	refractile body (RB) and a peripheral cytoplasm with a nucleus and vacuoles (inset, arrow). (d)
142	Gomori stain shows cytological details of the spores (bar=10 µm). (e) PAS stain evidences the
143	developmental stages of the parasite. Four zoospores are visible inside a spore (arrowhead) (bar=10
144	μm).

Figure 2. (f) The choroid space near the retina is infiltrated by numerous cysts (asterisk) (H&E, bar=500 μm). (g) The iris and the ciliary body are replete by numerous free mature spores (arrowhead). A severe inflammatory reaction and granulation tissue are evident. Note the multinucleated giant cells (inset) (H&E, bar=50 μm). (h) Around the cysts, a granulomatous inflammatory reaction characterized by a wall of epithelioid cells is visible (inset) (H&E, bar=100 μm). (i) The cysts show a thick eosinophilic wall and contain spores at different developmental stages (H&E, bar=50 μm).

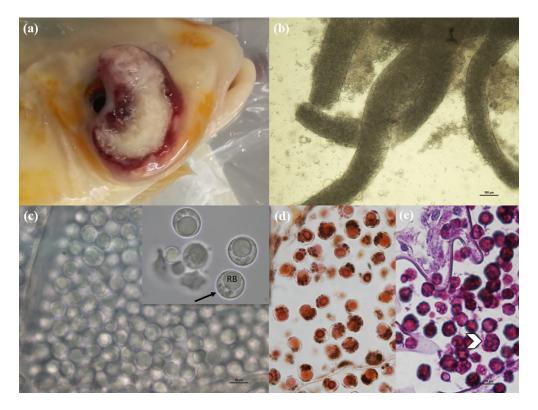


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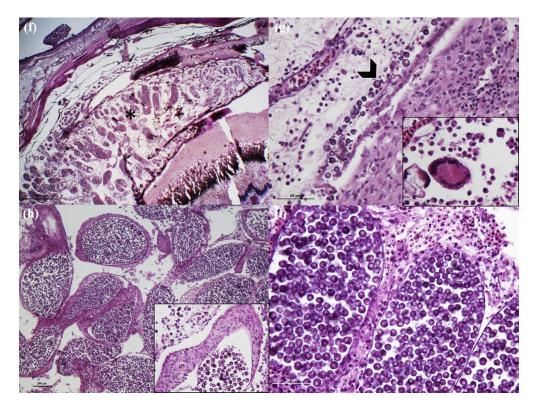


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